

# World Journal of *Hypertension*

*World J Hypertens* 2016 February 23; 6(1): 1-65



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*World Journal of Hypertension* is currently no indexing/abstracting.

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**NAME OF JOURNAL**  
*World Journal of Hypertension*

**ISSN**  
ISSN 2220-3168 (online)

**LAUNCH DATE**  
December 23, 2011

**FREQUENCY**  
Quarterly

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**PUBLISHER**  
Baishideng Publishing Group Inc  
8226 Regency Drive,  
Pleasanton, CA 94588, USA  
Telephone: +1-925-223-8242  
Fax: +1-925-223-8243  
E-mail: [bpgoffice@wjnet.com](mailto:bpgoffice@wjnet.com)  
Help desk: <http://www.wjnet.com/esps/helpdesk.aspx>  
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**PUBLICATION DATE**  
February 23, 2016

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## Regression of cardiovascular remodeling in hypertension: Novel relevant mechanisms

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**Supported by** Comisión Nacional de Investigación Científica y Tecnológica (CONICYT, Chile) grants FONDECYT 1121060 (to JEJ and MPO), FONDEF D11I1122 (to MPO and JEJ) and FONDAP 15130011 (to MPO).

**Conflict-of-interest statement:** The authors declare no conflicts of interest regarding this manuscript.

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Received: June 16, 2015  
 Peer-review started: June 18, 2015  
 First decision: July 31, 2015  
 Revised: November 16, 2015  
 Accepted: December 3, 2015  
 Article in press: December 4, 2015  
 Published online: February 23, 2016

### Abstract

Asymptomatic organ damage due to progressive kidney damage, cardiac hypertrophy and remodeling put hypertensive patients at high risk for developing heart and renal failure, myocardial infarction and stroke. Current antihypertensive treatment normalizes high blood pressure, partially reverses organ damage, and reduces the incidence of heart and renal failure. Activation of the renin-angiotensin system (RAS) is a primary mechanism of progressive organ damage and, specifically, a major cause of both renal and cardiovascular fibrosis. Currently, inhibition of the RAS system [mainly with angiotensin I converting enzyme inhibitors or angiotensin II (Ang II) receptor antagonists] is the most effective antihypertensive strategy for normalizing blood pressure and preventing target organ damage. However, residual organ damage and consequently high risk for cardiovascular events and renal failure still persist. Accordingly, in hypertension, it is relevant to develop new therapeutic perspectives, beyond reducing blood pressure to further prevent/reduce target organ damage by acting on pathways that trigger and maintain cardiovascular and renal remodeling. We review here relevant novel mechanisms of target organ damage in hypertension, their role and evidence in prevention/regression of cardiovascular remodeling and their possible clinical impact as well. Specifically, we focus on the signaling pathway RhoA/Rho kinase, on the impact of the vasodilatory peptides from the RAS and some insights on the role of estrogens and myocardial chymase in cardiovascular hypertensive remodeling.

**Key words:** Remodeling; Hypertrophy; Rho kinase; Myosin phosphatase target subunit 1; Angiotensin; Angiotensin1-9; Chymase; Angiotensin1-7

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**Core tip:** Antihypertensive treatment normalizes high



blood pressure, partially reverses organ damage, and reduces the incidence of heart and renal failure. However, residual organ damage and high risk for cardiovascular events still persist. We review here novel relevant mechanisms of cardiovascular damage in hypertension, their role and evidence in prevention/regression of cardiovascular remodeling and their possible clinical impact. We focus on the signaling pathway RhoA/Rho kinase, on the impact of the vasodilatory peptides from the renin angiotensin system and some insights on the role of estrogens and myocardial chymase in cardiovascular remodeling due to hypertension.

Jalil JE, Ocaranza MP. Regression of cardiovascular remodeling in hypertension: Novel relevant mechanisms. *World J Hypertens* 2016; 6(1): 1-17 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/1.htm> DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.1>

## INTRODUCTION

Target organ damage in hypertension causing asymptomatic renal dysfunction, atrial size enlargement along with cardiac hypertrophy and remodeling place hypertensive subjects at a very high risk condition to develop cardiac failure, progressive kidney disease, myocardial infarction and stroke as well. Up-to-date antihypertensive drug therapy reduces to normality elevated arterial blood pressure, does revert organ damage to some extent, and diminishes the occurrence of cardiac and renal disease. However, the clinical impact of all antihypertensive drug classes is not substantially different among most clinical outcomes when the blood pressure effect is equivalent<sup>[1]</sup>. Besides, permanent stimulation of the renin-angiotensin axis is a fundamental process of continuing damage to the target organs and a main cause of fibrosis both in the kidney and also in the myocardium. Current pharmacological blockade of the renin-angiotensin axis (primarily with inhibitors of angiotensin I converting enzyme (ACE) or by blocking the angiotensin II receptor) is a most effective antihypertensive strategy for normalizing high blood pressure and for preventing continuing end organ damage<sup>[2]</sup>. However, both residual damage in the target organs and consequently, a condition of high hazard for experiencing major clinical events still persist.

Thus, in hypertension, it is most crucial the development of new therapeutic viewpoints, further than only reducing blood pressure to better prevent/decrease target organ damage by aiming to paths triggering and maintaining cardiovascular remodeling and also in the kidney<sup>[2]</sup>. Our purpose is to review here three novel mechanisms of target organ damage in hypertension, their role and evidence on regression of cardiovascular remodeling and their possible clinical impact as well. Specifically, we will concentrate on the signaling pathway RhoA/Rho kinase, on the impact of

**Table 1 Rho kinase downstream target proteins, some signaling pathways and cellular functions<sup>[6]</sup>**

ROCK downstream target protein	Signaling pathway	Function
Myosin binding subunit of MLC/MYPT1	MYPT1/MLC	Stress fiber formation
MLC2	MYPT1/MLC2	Mediates calcium sensitization and thereby enhances and sustains contraction in the vascular bed
LIM kinase/cofilin Ezrin/radixin/moesin Adducin	LIM kinase/cofilin	Stress fiber formation

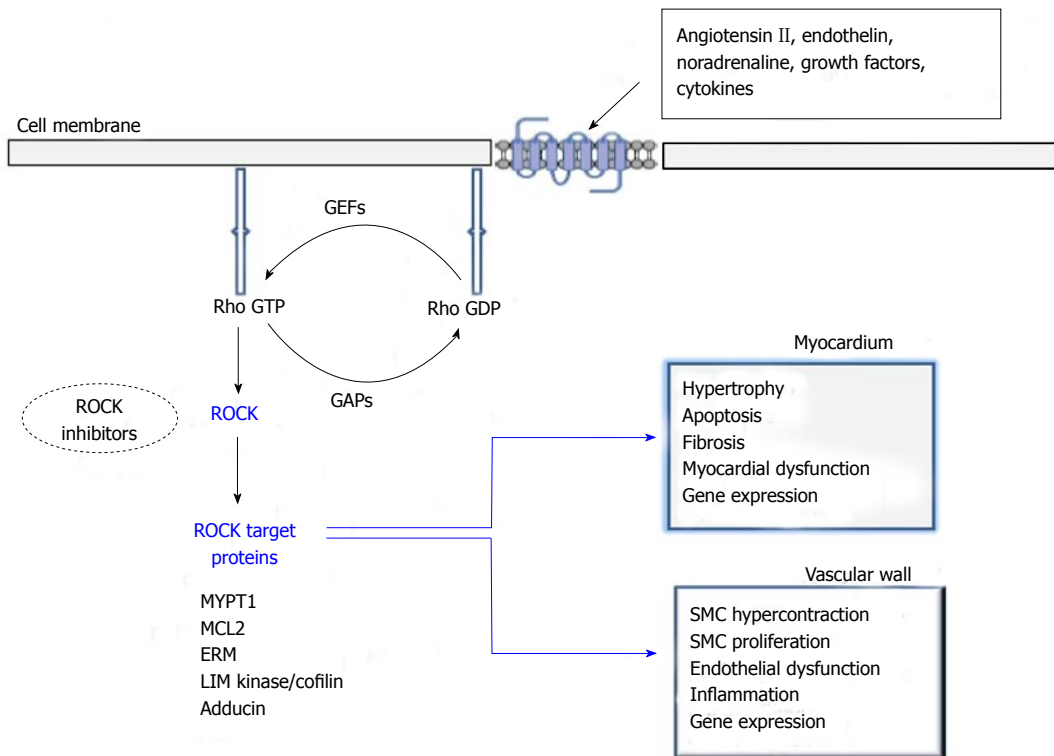
ROCK: Rho kinase; MLC: Myosin light chain; MYPT1: Myosin phosphatase target subunit 1.

the vasodilatory peptides from the renin angiotensin system and on the role of estrogens and the myocardial chymase-angiotensin II pathway in cardiovascular hypertensive remodeling. Interestingly, the 3 aforementioned mechanisms interact strongly with the renin angiotensin system at the cardiovascular level.

## RHO KINASE, HYPERTENSION AND CARDIOVASCULAR REMODELING

The small protein Rho (a guanosine triphosphatase) and its target Rho kinase (ROCK), have important functions in blood pressure modulation, by regulating smooth muscle contraction and additionally in cardiovascular remodeling. Agonists of receptors coupled to the G protein in the cell membrane (such as endothelin, angiotensin II, or noradrenalin), growth factors and cytokines activate Rho<sup>[3-6]</sup> (Figure 1). Some actomyosin-associated proteins, such as myosin light chain (MLC) phosphatase, myosin light chain 2, LIM-kinase, ezrin radixin-moesin and adducin are considered physiological ROCK substrates<sup>[7-9]</sup> (Table 1 and Figure 1). In non-hypertensive people, activation of the classical renin angiotensin system induced by low-salt diet does increase RhoA-ROCK signaling and does stimulate activation of the RhoA guanine exchange factor Arhgef1, which is implicated in vascular tone regulation and in hypertension induced by angiotensin II) in circulating mononuclear cells<sup>[10]</sup>. Immediately after Rho activation, this small protein is translocated to the cell membrane where it phosphorylates and activates ROCK (Figure 1), controlling in this way several cellular functions (Table 2), the majority of them related to remodeling. Activated ROCK does phosphorylate MLC phosphatase, which becomes inhibited. This cascade does stimulate tonic contraction of the smooth muscle within the vessels, development of stress fibers, and also cell migration. Thus, activation of both Rho and ROCK has significant effects on numerous cardiovascular diseases<sup>[4-6,11,12]</sup>, especially in hypertension<sup>[13]</sup>.

Administration of ROCK inhibitors reduces blood



**Figure 1** Rho kinase activation and downstream effects on cardiovascular remodeling in hypertension (as well as in cardiovascular disease). GEFs: Guanine nucleotide exchange factors; GAPs: GTPase activating proteins; GTP: Guanosine-triphosphate; GDP: Guanosine-diphosphate; MYPT1: Myosin binding subunit of myosin light chain phosphatase 1; MLC2: Myosin light chain 2; ERM: Ezrin/radixin/moesin; SMC: Smooth muscle cell. Adapted from references<sup>[2,47,144,145]</sup>.

**Table 2** Cellular functions controlled by the RhoA/Rho kinase pathway<sup>[7-9]</sup>

Cytoskeletal dynamics and actin organization (formation of stress fibers and focal adhesion complexes)
Cell contraction
Adhesion
Morphology
Motility
Transcriptional regulation (pro remodeling genes)
Cell proliferation and cytokinesis
Differentiation
Apoptosis
Insulin-stimulated insulin receptor substrate-1 phosphorylation
Development

pressure effectively in the rat with spontaneous hypertension (SHR)<sup>[14-17]</sup>, in the deoxycorticosterone acetate (DOCA) salt hypertensive model in rats<sup>[14,15,18]</sup>, renal hypertensive rodent<sup>[14]</sup>, L-NAME hypertensive rats<sup>[19,20]</sup> and in also in normotensive rats<sup>[14,15,17,20]</sup> which indicates that blood pressure fall by inhibitors of Rho kinase does not depend on the mechanism of hypertension<sup>[21]</sup>. Furthermore, the ROCK intracellular signaling cascade is activated in human hypertension<sup>[22,23]</sup> where elevated ROCK activity appears to be a consequence derived from up-regulation of the renin angiotensin system and also from higher levels of reactive oxygen species (ROS)<sup>[8,9]</sup>. ROCK inhibition decreases smooth muscle contractility by reducing MLC phosphorylation in the smooth muscle cell and by

enhancing endothelial function through reestablishing eNOS activity and NO production<sup>[8,14,18,24-28]</sup>.

Cardiovascular inflammation and remodeling are also reduced by ROCK inhibition<sup>[8]</sup> through: (1) suppressing the levels of cytokines and adhesion molecules such as plasminogen activator inhibitor-1 (PAI-1), monocyte chemoattractant protein 1 (MCP-1) and the transforming growth factor 1 in endothelial and in smooth muscle cells<sup>[18,24,29,30]</sup>; (2) by inhibiting in endothelial cells ROS production through down-regulation of NADPH oxidase<sup>[24,31,32]</sup>; (3) by reducing in smooth muscle cells secretion of cyclophilin A<sup>[33]</sup>; and also (4) by augmenting the levels of angiotensin 1-9<sup>[18]</sup>. Moreover, ROCK inhibitors delivered in the brainstem reduce blood pressure and sympathetic nerve in hypertensive rodents<sup>[34,35]</sup>.

In hypertension, there are experimental data available on the significant role of ROCK activation on developing myocardial hypertrophy, remodeling and ventricular dysfunction. In the rodent with salt-sensitive hypertension, cardiac hypertrophy was importantly reduced by using Y-27632, a specific ROCK inhibitor<sup>[36]</sup>. In this experimental model, upregulated *RhoA*, *ROCK* gene expression and phosphorylated MLC in the stage with hypertrophy were also inhibited by ROCK inhibition<sup>[36]</sup>. Besides, fasudil attenuated cardiac fibrosis possibly throughout inhibition of inflammatory cells myocardial infiltration in hypertensive rats<sup>[37]</sup>. Additionally, activated ROCK in the aorta observed in rats with genetically determined elevated levels of

angiotensin converting enzyme and the peptide angiotensin II, is reduced by Fasudil administration causing reduced gene expression that stimulate vascular remodeling (like transforming growth factor 1, PAI-1 and MCP-1) and also enhances oxidative species in the vasculature<sup>[24]</sup>.

Long-term inhibition of ROCK using fasudil ameliorated diastolic cardiac failure in the Dahl hypertensive rat<sup>[38]</sup>. Besides, in rats with left ventricular hypertrophy (LVH) due to pressure overload, inhibition of ROCK with GSK-576371 recovered LV chamber geometry, improved diastolic function and reduced myocardial fibrosis<sup>[39]</sup> and recently, long term treatment of DOCA-salt and *N*<sup>ω</sup>-nitro-L-arginine methyl ester (LNAME) hypertensive rats with the more potent ROCK inhibitor SAR407899 reduced hypertension and cardiac and renal remodeling in a dose-dependent way in both models<sup>[19]</sup>. Interestingly, in DOCA hypertensive rats, blood pressure reduction and protective effects on hypertensive organ damage of SAR407899 were superior compared to amlodipine and also to ramipril<sup>[19]</sup> and hearts of hypertensive DOCA or LNAME animals treated with SAR407899 had significantly better systolic left ventricular (LV) function (measured as heart power *in vitro*). Additionally, endothelial-dependent relaxation was significantly and dose-dependently improved after long-term treatment with SAR407899<sup>[19]</sup>. An important amelioration of myocardial interstitial fibrosis and expression of collagen genes and of CD3 and CD68 (markers of infiltrating macrophages and leukocytes) in both models was observed, possibly explained by the relevant Rho kinase function in cellular migration and cytokinesis through cytoskeleton modulation<sup>[19]</sup>.

In more advanced heart disease secondary to hypertension, and evident impairment in cardiac function (both diastolic and systolic), it is very likely that ROCK activity levels be rather similar to Rho kinase activation levels observed in cardiac failure secondary to different mechanisms and its inhibition could produce in this situation similar benefits. In the mouse overexpressing *Gαq*, deletion of *ROCK1* gene did prevent LV chamber dilatation and improved cardiac contractility<sup>[40]</sup>. Furthermore, in cardiomyocytes in culture, ROCK activation up-regulates Bax *via* p53 to induce apoptosis<sup>[41]</sup>. In the transgenic mouse that overexpress the isoform MYPT2, activation of myosin phosphatase induced LV function decline and remodeling, probably by reducing calcium sensitivity, along with impairing the myofibrillar organization, which is the original report about the functions of both MYPT2 and myosin phosphatase, and the consequences of *in-vivo* cardiac MLC phosphorylation<sup>[42]</sup>.

### **Assessing Rho kinase activation in human circulating leukocytes, a possible marker of cardiovascular remodeling and risk**

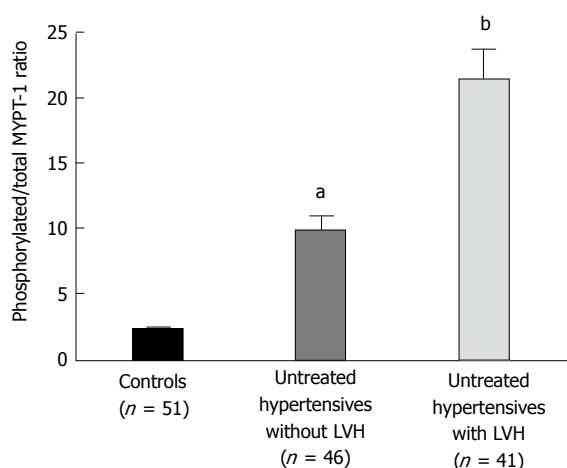
In people diagnosed with metabolic syndrome (MetS), Liu *et al*<sup>[43]</sup> reported for the first time significantly increased ROCK activity by 31% through the

measurement of ROCK phosphorylation in circulating leukocytes using the approach of determining myosin binding subunit phosphorylation (MBS). At the same time they observed that plasma concentrations of high-sensitivity C-reactive protein were substantially higher and that circulating levels of adiponectin were significantly lower in MetS subjects as compared with control subjects. Additionally, in this population they found that increased ROCK activation was significantly related with body mass, waist circumference, fasting glucose, high-sensitivity C reactive protein, and additionally with triglyceride levels<sup>[43]</sup>. In this clinical study the probability of increased ROCK activity was considerably increased with the amount of MetS components. Experimental findings indicate that insulin resistance promoted by ROCK is implicated in myocardial damage in rats with MetS and that this action of Rho kinase is probably through the IRS-1-PI3-kinase-protein kinase B (Akt) signaling cascade<sup>[44]</sup>. In humans, ROCK activation in leukocytes is also enhanced by smoking and does predict endothelial dysfunction<sup>[45]</sup>.

In circulating leukocytes, Hata *et al*<sup>[46]</sup> measured the activity of Rho kinase by assessing the relation amid phosphorylated myosin-binding subunit (p-MBS) on myosin light chain phosphatase to the total MBS and also the change on the blood flow in the forearm (FBF) as a pharmacological action of the distinctive Rho kinase inhibitor fasudil using strain-gauge plethysmography in control subjects and also in subjects diagnosed with a cardiovascular illness. Compared to healthy subjects, they found that leukocyte p-MBS/total-MBS ratio was substantially higher (by 90%) in the diagnosed patients<sup>[46]</sup>. Besides, they found that the characteristic inhibitor of ROCK fasudil increased FBF by 300% only in their patients with cardiovascular disease, but this was not the case not in the healthy control group<sup>[46]</sup>. Moreover, they found an important relationship between leukocyte p-MBS/total-MBS and maximal FBF induced by fasudil in the group with cardiovascular disease ( $r = 0.59$ ), not in the healthy subjects.

Lately, we have evaluated the level of ROCK activation in leukocytes obtained from venous blood, by quantifying the relationship of phospho to total MLC phosphatase 1 (known as MYPT1-P/T) as a potential remodeling marker in untreated hypertensive patients (HT), in HT patients with LVH or with type II diabetes mellitus receiving specific treatment and also in patients with congestive cardiac failure and LV systolic reduced function<sup>[47-49]</sup>.

In a recent clinical follow up trial with the aim to determine the correlation amid the observed activity of ROCK and a first main cardiovascular event along with hospitalization rates for congestive cardiac failure, the levels of ROCK activity were determined in leukocytes by the technique of Western blot in more than 600 subjects who undertook a health-screening examination<sup>[50]</sup>. After a median period of 42 mo of follow-up, 29 deaths were registered (10 of them because of cardiovascular causes), 2 of them were



**Figure 2** Comparative levels of Rho kinase activity in circulating leukocytes (determined as phosphorylated/total MYPT-1 ratio) in healthy normotensive controls, in untreated hypertensive patients without LVH and in untreated hypertensive patients with left ventricular hypertrophy (Data shown as mean  $\pm$  SEM). <sup>a</sup> $P < 0.01$  vs Controls; <sup>b</sup> $P < 0.01$  vs untreated hypertensive patients without left ventricular hypertrophy (after significant ANOVA, respectively). Adapted, with permission from reference<sup>[47]</sup>. LVH: Left ventricular hypertrophy.

diagnosed with a myocardial infarction, in 20 of them at least one revascularization procedure was performed, 15 developed a stroke, and 17 required hospitalization for congestive cardiac failure<sup>[50]</sup>. In the above-mentioned study, once the adjustment for several confounding variables (such as age, gender, known risk factors and other relevant predictors of cardiovascular illness) was performed, ROCK activity remained as a robust independent indicator of a first main cardiovascular event (the hazard ratio was 2.19), of death as a consequence of cardiovascular disease (hazard ratio, 2.57), cerebrovascular accident (hazard ratio, 2.14), and the clinical necessity for revascularization (reported hazard ratio was 2.68)<sup>[50]</sup>. The authors concluded that ROCK activity levels determined in circulating leukocytes may be a new marker of cardiovascular events and propose that its inhibition may be a novel therapeutical approach to achieve effective prevention of cardiovascular disease<sup>[50]</sup>.

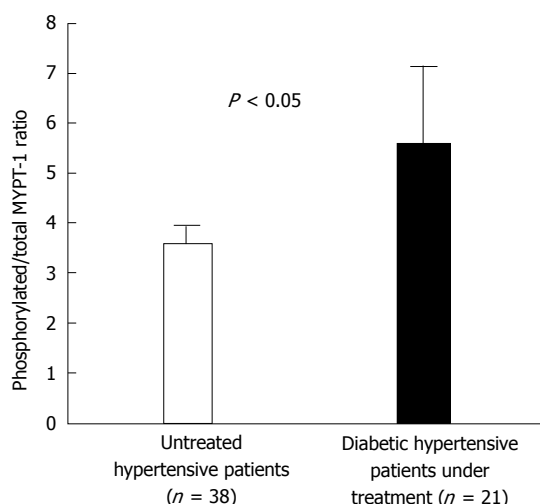
**Rho kinase activation in subjects with essential hypertension:** In a case-control study with the goal of comparing ROCK activation levels in subjects diagnosed with hypertension against a healthy normotensive control group in regard to the existence of LVH assessed by 2D-echocardiography we measured LV mass and dimensions in addition to LV performance and ROCK activation levels in leukocytes from venous blood (by MYPT1-p/t levels through Western blot)<sup>[47]</sup>. Compared to non-hypertensive controls, MYPT1-p/t was considerably higher by 450% in the group lacking LVH and by 900% in the group with confirmed LVH by 2D-echocardiography (Figure 2). In contrast with the hypertensive subjects without LVH, MYPT1-p/t was considerable higher (by 200%) in the subjects with

hypertension along with echocardiographic confirmed LVH<sup>[47]</sup>. Additionally, in the hypertensive subjects with evidence of eccentric LVH had an MYPT1-p/t relationship remarkably higher (by 400%) compared to hypertensive subjects and no eccentric LVH. Patients having an E/e' ratio measured in the transmitral diastolic flow  $\geq 15$  showed a substantially higher MYPT1-p/t relationship (26%) as related to the levels in those subjects with a smaller E/e' ratio. This study concluded that ROCK activation levels determined in leukocytes from venous blood are significantly raised up in hypertensive subjects with definite cardiac hypertrophy compared with HT patients without LVH. ROCK activation is additionally increased when eccentric hypertrophy is present. Therefore, in subjects with essential hypertension, ROCK phosphorylation/activation determined in leukocytes from venous blood is correlated to pathological myocardial remodeling and could contribute as one indicator of LVH<sup>[47]</sup>.

Similarly, Hata *et al.*<sup>[51]</sup> recently observed significantly higher Rho kinase activation levels in circulating leukocytes in subjects with essential hypertension when contrasted to healthy individuals by 37%. Besides, in these hypertensive patients under antihypertensive treatment, ROCK activity levels were substantially lowered in patients using calcium channel antagonists compared to patients receiving as antihypertensive treatment inhibitors of the renin-angiotensin-aldosterone axis, thiazides, or  $\beta$ -blockers<sup>[51]</sup>. These observations suggest that increased Rho kinase activity associated to hypertension may cause activation of leukocytes along with leukocyte infiltration into the vessel wall, which favors atherosclerosis progression<sup>[51]</sup> (as well as remodeling) and suggest the possible clinical relevance of determining in this context the degrees of Rho kinase activation. In a randomized clinical study with the aim of assessing the impact of the specific aldosterone receptor blocker eplerenone, on the endothelial function determined by flow mediated dilatation (FMD) and on Rho kinase activation as well, determined in leukocytes obtained from venous blood in subjects with essential hypertension, 60 patients were received eplerenone, the antagonist of calcium channels nifedipine, or losartan for 48 wk<sup>[52]</sup>. They observed that FMD was increased and leukocyte Rho kinase activity was reduced with eplerenone, whereas nifedipine reduced Rho kinase activity but did not modify FMD<sup>[52]</sup>. In the aforementioned clinical trial losartan augmented FMD but did not modify Rho kinase activity. In this clinical study, both the blood pressure reducing effect and also the vasodilation levels induced by nitroglycerin were similar with the different three antihypertensive drugs throughout the follow-up<sup>[52]</sup>.

Rho kinase activity is importantly increased in subjects with very high cardiovascular risk and remodeling hazard, as is frequently observed in subjects with the combined diagnosis of both type 2 diabetes mellitus along with essential hypertension. In a cross sectional clinical study carried through comparing three groups





**Figure 3** Rho kinase activity in circulating leukocytes (determined as phosphorylated/total MYPT-1 ratio) in untreated hypertensive patients (white bar, mean age 48 years, mean BP 121 mmHg) and in hypertensive diabetic patients under antihypertensive and anti-diabetic pharmacological treatment (black bar, mean age 51, mean BP 111 mmHg). Data shown as mean  $\pm$  SEM, adapted with permission from reference<sup>[48]</sup>.

of subjects<sup>[48]</sup>: Essential hypertensive patients under no medical treatment, patients with both hypertension and type II diabetes under treatment (with similar degrees of left cardiac mass) and normotensive control subjects, in the patients having the two aforementioned medical conditions, increased ROCK activation (determined in venous blood leukocytes) was found as compared to hypertensive subjects not getting pharmacological antihypertensive treatment<sup>[48]</sup> (Figure 3). In this clinical study, in the diabetic hypertensive patients compared to both non-diabetic hypertensives and to normotensive controls, increased levels of oxidative stress were found<sup>[48]</sup>. These findings were correlated with reduced arterial compliance and could help to explain the unfavorable vascular remodeling that is commonly detected in hypertensive plus diabetic patients receiving treatment<sup>[48]</sup>.

In a very recent prospective clinical trial aimed to evaluate the effect of the specific ROCK inhibitor fasudil on diastolic LV function parameters observed in a group of individuals diagnosed with type 2 diabetes presenting with preserved systolic performance, 250 patients with the established clinical diagnosis (62% of them hypertensives), were allocated to receive the ROCK inhibitor (14 d, 30 mg *iv* twice per day) or to placebo<sup>[53]</sup>. As planned, echocardiographic parameters were determined before and after 1 mo receiving this treatment. In relationship with the group that was randomized to placebo, in the subjects that were randomized to fasudil, an important reduction in both blood pressure (diastolic) and in echocardiographic late diastolic transmitral flow was observed<sup>[53]</sup>. In the aforementioned clinical study, deceleration time, relaxation time (isovolumic), peak early annular diastolic velocity ( $e'$ ), peak of late diastolic annular velocity, as well as the E to  $e'$  ratio also showed an important

recovery by fasudil administration for one month<sup>[53]</sup>. Moreover, the Em to Am ratio, both the relaxation (isovolumic) and deceleration times, along with E to  $e'$  ratio values observed after receiving treatment with fasudil in the subjects with baseline diminished left ventricular chamber relaxation diverged significantly from what was noticed in the patients presenting with normal left ventricular chamber relaxation. Accordingly, clinical ROCK inhibition by using fasudil improved for the first time left ventricular chamber diastolic function parameters in diabetic patients (the majority of them hypertensives) and normal systolic performance<sup>[53]</sup>.

### **Rho kinase increased signaling in subjects with progressive cardiac remodeling: the case of congestive cardiac failure with systolic function decline:**

Rho kinase activity is markedly augmented in those subjects with established cardiac failure (CF) due to systolic dysfunction. In a cross sectional study comparing control healthy subjects with patients with chronic and clinically stable CF due to systolic dysfunction under optimal medical treatment, we observed that Rho kinase activation (determined in circulating leukocytes as the MYPT1-P/T ratio) was increased by 100-fold and that it was inversely related with ejection fraction<sup>[49]</sup>. Interestingly, in those patients with CF with LV diameter  $\geq 60$  mm MYPT1-P/T was significantly more elevated than in the CF subjects with LV diameter  $< 60$  mm. Thus, ROCK activity is markedly augmented in patients with stable chronic CF receiving optimal medical treatment, and Rho kinase signaling is robustly related to pathologic LV chamber remodeling and to systolic function decline as well<sup>[49]</sup>. Another clinical study examined whether ROCK activation (determined in venous blood leukocytes) is increased in congestive CF and how it is related with the clinical prognosis in 170 patients admitted with this clinical condition<sup>[54]</sup>. Patients were prospectively followed up for  $14.4 \pm 7.2$  mo or up to the event of cardiac death. Observed Rho kinase signaling in the patients with congestive CF was significantly higher than that of two control groups. The protein concentrations of both Rho kinase isoforms (ROCK1 and ROCK2) as well as the measured activation of the up-river Rho kinase cascade GTPase RhoA in the congestive CF patients were significantly higher than what was observed in both control groups<sup>[54]</sup>. Dyspnea at rest, reduced left ventricular systolic function and impaired renal function were all independent factors predicting Rho kinase activity levels at baseline in the subjects with congestive CF<sup>[54]</sup>. By combining Rho kinase activity with N terminal pro brain natriuretic peptide (NT-proBNP) an incremental value in the estimate of long-term mortality (when compared with only the NT-proBNP measurement) was observed. Thus, Rho kinase activity is elevated in these patients with extreme myocardial remodeling, it is also associated with higher mortality and it might be an additional biomarker to congestive CF risk assessment<sup>[54]</sup>. In a clinical study to

evaluate whether Rho kinase activity in venous blood leukocytes is elevated in subjects presenting with an established acute coronary syndrome and if Rho kinase activation does predict long-term cardiovascular events, 188 patients with ACS and 61 control subjects were evaluated<sup>[55]</sup>. The authors found significantly increased ROCK activity in the two clinical groups (myocardial infarction and unstable angina) when it was compared to control subjects. Besides, patients with both elevated NT-proBNP and Rho kinase activity on admission had a five-fold hazard of a major cardiovascular outcome in relation to the observed hazard in those subjects with low NT-proBNP and low ROCK activity<sup>[55]</sup>. Their main conclusion was that by combining Rho kinase activity and NT-proBNP levels a subset of acute coronary syndrome patients at particularly high risk might be identified<sup>[55]</sup>.

Altogether, these findings strongly suggest that ROCK activation in circulating human leukocytes is directly related to pathological cardiovascular remodeling, from early target organ damage in hypertension to extreme cardiovascular disease. Besides, as this measurement possibly mirrors remodeling it has prognosis value for disease progression, clinical events and conceivably for target organ damage/disease prevention and regression.

## IMPACT OF THE VASODILATORY PEPTIDES FROM THE RENIN ANGIOTENSIN SYSTEM ON MECHANISMS IN HYPERTENSION AND CARDIOVASCULAR REMODELING

Remodeling of the cardiovascular structures does occur as a response, not only to modifications in blood arterial pressure or flow but also to variations in the neural and hormonal milieus, where the renin-angiotensin-aldosterone axis exerts a major influence<sup>[56]</sup>. The aforementioned neurohormonal system, one of the oldest phylogenetically hormonal systems, is most recognized because of its fundamental role in regulating hydromineral and cardiovascular homeostasis<sup>[56,57]</sup>. The renin-angiotensin-aldosterone axis is a fundamental element of CV physiology having a main pathophysiological role by regulating vascular tone, blood pressure, sodium and potassium balance and vascular responses to both injury and inflammation<sup>[58]</sup>. Long lasting activation of the renin angiotensin system, throughout both the octapeptide angiotensin II and the mineralocorticoid hormone aldosterone, causes hypertension and at the same time does stimulate prohypertrophy, proinflammation, prothrombosis, and atherogenesis pathways strongly linked with hypertensive organ damage.

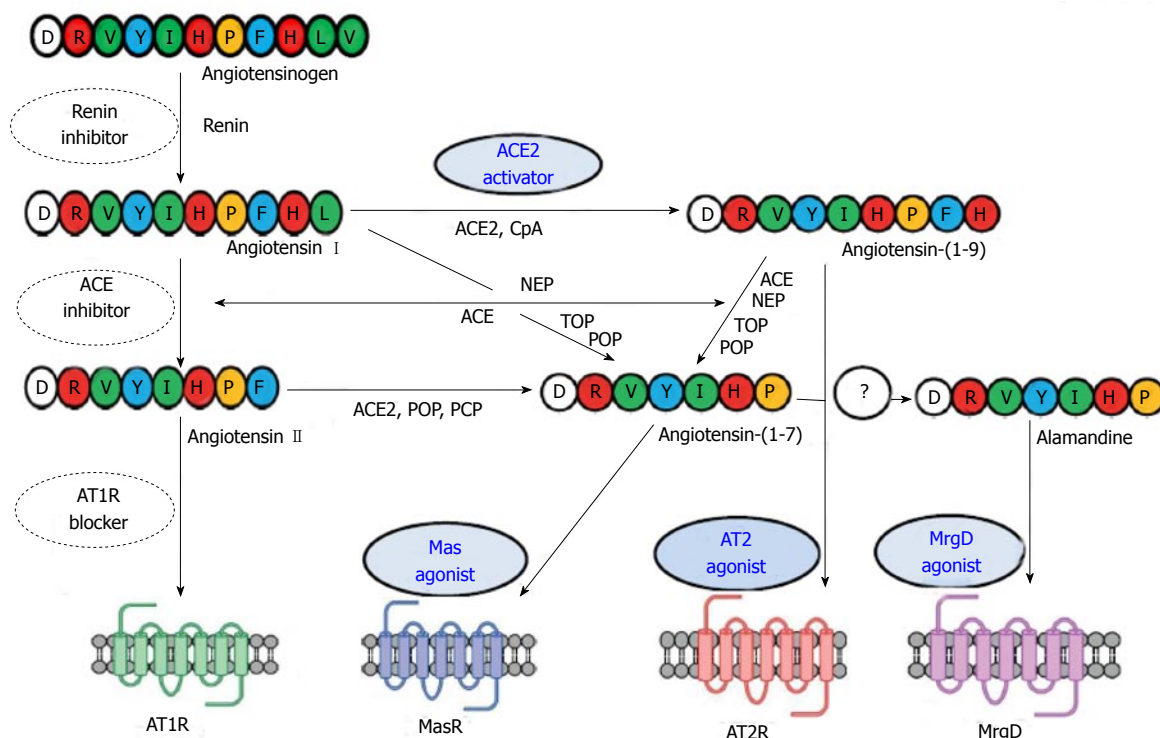
During a long period, a main research focus has been production and signaling of Angiotensin II, highlighting both the ACE and renin regarding its

production. With the precise description of both main angiotensin II receptors 1 (AT1R) and 2 (AT2R), most of the issues related to the biochemistry, pharmacology, and physiology of the renin-angiotensin-aldosterone system seemed to be resolved<sup>[57,59]</sup>. Extensive studies of the RAAS for a long time, were mainly focused on four classes of drugs targeting the renin-angiotensin-aldosterone system at different levels: Angiotensin-converting enzyme inhibitors (known as -prils), Angs receptor blockers (-sartans), renin inhibitors (-kirens) and mineralocorticoid receptor antagonists (Figure 4). All of them are cornerstones in the treatment of HT. However, there are more recent discoveries extending our understanding of important properties of the renin-angiotensin-aldosterone system. In the most recent decades, increasing evidence has been accumulated indicating an exceeding complexity of the RAAS. Among some novel discoveries is the finding and location of the physiologically important aions of Ang-(1-7)<sup>[60]</sup>, Ang-(1-9)<sup>[61]</sup> and alamandine<sup>[62]</sup> acting through identifiable tissue receptors<sup>[60,62,63]</sup> (Figure 4).

### **New vasodilatory peptides in the RAAS and hypertensive remodeling**

The therapeutic effectiveness of "classic" blocking the RAAS for treating HT and also related cardiovascular illness has been extensively well established. However, in the most recent decades growing facts and data indicate that the roles and biological functions of the RAAS go beyond the effects initially described. Currently, the amount of biologically relevant end-products of the RAAS is even now increasing, which raises new possibilities to attack through this axis cardiovascular disease, and specifically hypertensive CV remodeling. These facts are particularly accurate for the peptides Ang (1-7) along with Ang (1-9) and lately, for alamandine (Figure 4), and in most of the situations these peptides display biological effects opposing to Angiotensin II.

**Ang-(1-7):** Production of the active molecule Ang-(1-7) mostly depends on the cleavage of the octapeptide Ang II which is performed by ACE2 (Figure 4). Moreover, Ang-(1-7) can be also formed through hydrolysis of the decapeptide Ang I performed by other peptidases such as prolyl-endopeptidase, neutral-endo-peptidase (NEP), and timeth-oligopeptidase which cleave the Pro<sup>7</sup>-Phe<sup>8</sup> bond to remove the final three amino acids. The function of the prolyl endopeptidase (PEP)<sup>[64]</sup>, oligopeptidase (TOP) and of NEP<sup>[65]</sup> in the enzymatic hydrolysis from Ang I onto Ang-(1-7) depends on tissue distribution and substrate availability of the enzymes. Neprilysin behaves as an especially active enzyme which has been primarily located in the vessel endothelial cells while the thimet oligopeptidase enzymatic activity is relevant in the cleavage toward Ang-(1-7) occurring within the smooth muscle vascular cells<sup>[66,67]</sup>. Ang-(1-7) is also formed from Ang II by ACE2<sup>[68]</sup>. The level of Ang-(1-7) is regulated by the action of ACE which



**Figure 4** Schematic view of the current renin-angiotensin system and sites of possible therapeutic interventions in hypertension and cardiovascular remodeling. Letters in blue: Receptor agonists or enzyme activators; surrounded by interrupted lines: Enzyme/receptor inhibitors. AT1R: Angiotensin II receptor type 1; AT2R: Angiotensin II receptor type 2; ACE: Angiotensin converting enzyme I; ACE2: Angiotensin converting enzyme 2; MrgDR: Mas-related G protein-coupled receptor; NEP: Neutral endopeptidase; TOP: Thimet oligopeptidase; POP: Prolyl oligopeptidase; PCP: Prolyl carboxypeptidase. Adapted and modified from reference<sup>[146]</sup>.

hydrolyzes Ang-(1-7) to Ang-(1-5)<sup>[65]</sup> (Figure 4).

Ang-(1-7) produces its biological effects and acts throughout its own receptor, the Mas receptor (MasR), one of the membrane receptors coupled to G protein<sup>[8]</sup>. This receptor mediates the current known actions of this heptapeptide, as most of them can be prevented by the specific blocker, D-Ala<sup>7</sup>-Ang-(1-7) (A779)<sup>[8]</sup>. The Ang-(1-7) observed effects acting throughout the MasR in the CV system consistently include vasodilation, antihypertrophy, antiarrhythmogenesis, antifibrogenesis and antithrombogenesis<sup>[69-71]</sup>.

**Angiotensin (1-9):** The first observations about Ang-(1-9) (Figure 4) showed a rapid appearance of leucine after the injection of radiolabeled Ang I into dog renal and pulmonary arteries<sup>[72,73]</sup>, by the enzymatic action of a carboxypeptidase that was breaking down Ang I forming des-Leu<sup>10</sup> Ang I. It is possible to generate Ang-(1-9) from Ang I through the effect produced by several enzymes (carboxypeptidase-type), including cathepsin A and ACE2<sup>[74-76]</sup>, although at a relatively slow rate compared to the making of Ang-(1-7) starting from Ang II<sup>[77]</sup>. Moreover, it was observed that an inhibitor of ACE2 doesn't have an effect on Ang-(1-9) formation, but benzylsuccinate, a Cx<sup>A</sup> inhibitor, does stop the formation of Ang-(1-9) and rises the levels of Ang I in heart membranes<sup>[78]</sup>. Alternatively, it is possible to cleave Ang-(1-9) to Ang-(1-7) by the ACE carboxypeptidase or by the effects of other enzymes including prolyl

endopeptidase, NEP and thimetoligopeptidase 1 (TOD)<sup>[79,80]</sup>. Recently, it was found that Ang-(1-9) has the capability to be hydrolyzed into the peptide Ang-(2-9) by aminopeptidase A<sup>[81]</sup> (Figure 4).

Initially Ang-(1-9) was considered as a biologically non active peptide, operating indirectly by competing with Ang I for the ACE active site, and therefore reducing Ang II levels while increasing Ang-(1-7)'s<sup>[76,77,82]</sup>. However, increasing evidence has confirmed that Ang-(1-9) does work as a molecule with relevant cardiovascular effects both *in vitro* in addition to *in vivo*, through the AT2R<sup>[61,63,83,84]</sup>.

**Alamandine:** Recently a new member of RAAS has been discovered, the heptapeptide Ala-Arg-Val-Tyr-Ile-His-Pro, known as alamandine<sup>[62]</sup> (Figure 4). By using mass spectrometry, alamandine was identified as a chemical product of a catalytic hydrolysis of Ang A, an octapeptide, by ACE2<sup>[62]</sup>. Alamandine is composed by a sequence of amino acids which is extremely similar to Ang-(1-7). Both peptides diverge only in one amino acid residue (alanine in place of an aspartate residue) located at the amine-terminus. Alamandine may be also synthesized by decarboxylation of the N-terminal aspartate amino acid residue from Ang-(1-7). The enzyme which is in charge of the ultimate reaction remains unknown<sup>[62]</sup>. However, Alamandine degradation is not yet completely understood. Nevertheless, it is possible that aminopeptidases have a most relevant

role, since the subtraction of Ala1 could conduct to form Ang-(2-7), deemed as a non-active molecule, although it shows inhibitory activity of ACE<sup>[85]</sup>. Other Ang-(1-7)-degrading enzymes, such as NEP or neprilysin, may also participate, given alamandine's similarity to Ang-(1-7)<sup>[86]</sup> (Figure 4).

Although Alamandine is rather similar to the peptide Ang-(1-7) and the biochemical effects of both molecules look to be close, alamandine acts through a different receptor, the mas-related G-protein coupled receptor D (MrgD)<sup>[62]</sup>. Alamandine produces endothelial-dependent vasodilation in rat and mice aortic rings<sup>[62]</sup>. It has recently been observed that oral delivery of alamandine (by including it within HP- $\beta$  cyclodextrin), produced similar effects to those already observed for Ang-(1-7) such as a long-term antihypertensive effect in SHR and a main reduction of cardiac deposition levels of collagens I and III as well as fibronectin in isoproterenol-treated rats<sup>[62]</sup>.

#### Target receptors for the second RAAS arm

**Mas receptor and AT2R:** the two receptors, the AT2R and the Mas receptor, are G protein coupled receptors (GPCRs) (Figure 4) with their conventional seven transmembrane domains<sup>[60,68]</sup>. Interestingly, even though their signaling mechanisms are quite unusual for GPCRs and not completely understood, again, major resemblances have been observed. For both receptors, signaling by activating phosphatases, particularly the Src homology 2 domain-containing protein tyrosine phosphatase (SHP)-1 and SHP-2, seems to be crucial<sup>[87-90]</sup>. In both situations, it has been found that phosphatase activation does interfere, in an inhibitory way, with kinase driven signaling cascades, producing inflammation or hypertrophy involving molecules like the mitogen-activated protein kinases or the nuclear factor  $\kappa$ B (NF- $\kappa$ B)<sup>[91,92]</sup>. Another shared signaling mechanism of importance is the augmented NO synthesis and consequent increase of cyclic guanosine monophosphate (cGMP) levels, which does mediate the vasodilation effects of both receptors<sup>[93-95]</sup>. Furthermore, these two receptors are able to develop dimers with the AT1R which results in a functional inhibition of this latter one<sup>[96,97]</sup>.

**The MrgD:** MrgD belongs to the GPCRs family and it is associated to the MasR<sup>[98]</sup>. The MrgD is located in the myocardium in addition to the vessels wall<sup>[62]</sup>. Shinohara *et al.*<sup>[99]</sup> found that a small amino acid,  $\beta$ -alanine could internalize the MrgD, and thus be able to induce intracellular influx of calcium and to inhibit production of cAMP in Chinese hamster ovary cultured cells (CHO) that expressed rat, mouse, or human MrgD<sup>[99]</sup>. The effect in calcium influx can be understood by the connection of the MrgD with the G-protein  $\alpha$  subunit (Gq), and the cAMP suppression does suggest an interaction concerning the MrgD with the inhibitory G protein (Gi)<sup>[99]</sup>. Furthermore, MrgD activation through  $\beta$ -alanine also suppressed KCNQ/M-type potassium chan-

nels, and in this way also increased neuron excitability by means of the Gq and phospholipase C (PLC) pathways<sup>[100]</sup>. In addition, the incubation of alamandine with CHO cells that are transfected with MrgD induces significant NO release<sup>[62]</sup>.

It has been proposed that this MrgD receptor is related to pain sensation<sup>[101]</sup>, sensitiveness to thermic and mechanical stimulation<sup>[102]</sup>, and tumorigenic activity<sup>[103]</sup>. It was also observed that MRGD is able to transduce intracellular signaling of Ang-(1-7)<sup>[104]</sup>.

#### Clinical approach to the Ang-(1-7)-MasR, Ang-(1-9)-AT2R and Alamandine-MrgD axis in hypertension and cardiovascular remodeling

**Ang-(1-7)-MasR Axis:** ACE inhibitors (ACEI) and Ang II receptor blockers (ARB) can affect in part the ACE2-Ang-(1-7) system. Experimental studies in myocardial infarcted rats showed that chronic administration of enalapril prevented myocardial hypertrophy and contractile dysfunction in addition to increased ACE2 activity in plasma and in the ventricular wall<sup>[105]</sup>.

ACEI increase the Ang I levels, which are hydrolyzed to produce Ang-(1-7), through the actions of both ACE2 and NEP. The arterial pressure reduction effects due to ACEI are also associated to increased excretion of Ang-(1-7), an observation reported in urine collected from subjects with essential hypertension receiving the ACEI captopril during 6 mo<sup>[106]</sup>. It's well recognized that ACEI are able to diminish excretion of urinary protein in subjects with established type 2 diabetes mellitus<sup>[107]</sup>, and interestingly, in the ACE2 knock-out mice the proteinuric blocking effect of ACEI disappears<sup>[108]</sup>. At this respect, ARBs may be here markedly effective because elevated Ang II levels as a consequence of them will promote Ang-(1-7) production<sup>[109]</sup>. Additionally, the low affinity binding of Ang-(1-7) to the AT1 receptor may allow this peptide to work as an antagonist in the presence of Ang II<sup>[110]</sup>. In this regard, normotensive rodents with high ACE and Ang II along with low NEP activity<sup>[111]</sup> and Ang-(1-7) concentration<sup>[112]</sup> (genetically determined) showed a higher hypertensive response (chronic) after renovascular hypertension induction<sup>[113]</sup>. Besides, the inverse correlation observed among the amounts of both Ang II in addition to Ang-(1-7) in the aforementioned rodents, determined increased cardiac fibrous tissue deposition after isoproterenol administration<sup>[114]</sup> and also ROCK activation in the aortic wall as well as stimulation of genes that promote vascular remodeling (such as the monocyte chemoattractant protein 1 gene, the transforming growth factor 1 gene, and PAI gene)<sup>[24]</sup> and also higher oxidative stress levels in the vessels wall in normotensive rodents<sup>[115]</sup>.

In humans, similar relationships have been observed<sup>[116,117]</sup>. Particularly, in hypertensive patients having the DD-ACE genotype (with increased ACE levels), the Ang-(1-7) blood levels were reduced by 4 fold as compared to those observed in patients having the II-ACE genotype (and consequently lower ACE levels)<sup>[116]</sup>. In these subjects we reported an important effect of the



I/D ACE genotype on circulating NEP enzymatic activity in addition to an interactive effect amid the I/D ACE genotype status and the hypertensive condition<sup>[117]</sup>.

By blocking the classic ACE-Ang II-AT1R axis a well-recognized and effective anti-hypertensive and antiproteinuric treatment is obtained. More recently, a few patients have received activators of the ACE2-Ang-(1-7)-Mas receptor pathway, which can be separated in two main types: (1) those compounds that augment the enzymatic activity of ACE2 and will impact the system by increasing Ang II inactivation of<sup>[118]</sup> and (2) those molecules that increase Ang-(1-7) production and are particularly oriented to stimulate the MasR<sup>[119]</sup>. At this time, in the case of ACE2, little molecules have been developed which activate ACE2<sup>[120]</sup>. In rats with SHR, a leading ACE2 activator compound (XNT) diminishes BP and does recover ventricular function<sup>[121]</sup>. The recombinant human ACE2 has been also developed as a different attempt to use the possible therapeutic capabilities of ACE2. At this respect, it has recently been observed that rhACE2 administration attenuates diabetic kidney damage through a mechanism involving both Ang II reduction and Ang-(1-7) increasing signaling<sup>[122]</sup>. AVE 0991 is the first synthetic compound (non-peptide) developed in order to stimulate the MasR<sup>[123]</sup>. This molecule is an orally active MasR agonist that imitates the consequences of administering Ang-(1-7) on the kidney, the vessels, and on the heart as well<sup>[124,125]</sup>. AVE0991 does considerably prevent organ damage in SHR and also in rats with hypertension induced by L-NAME by preserving cardiac contractility, avoiding hypertension, and by reducing urinary protein excretion<sup>[123]</sup>. Two new designed peptides, CGEN-856 as well as CGEN-857, target the other activator of GPCR, and also show high specificity for the Mas receptor<sup>[126]</sup>.

**The Ang-(1-9)-AT2R axis:** The first observations regarding the biological actions of ACE2 and Ang-(1-9) counter-regulating the ACE/Ang II axis were made by Ocaranza *et al.*<sup>[105]</sup> (Figure 4). In myocardial infarction (MI) rats, down regulation of circulating and cardiac ACE2 enzymatic activity is observed in the chronic phase of LV dysfunction and this effect is precluded by enalapril<sup>[105]</sup>. When rats with MI or with the sham procedure received the ACEI enalapril for 2 mo, Ang-(1-9) levels in plasma were increased significantly but Ang-(1-7) levels were not modified<sup>[105]</sup>. Thus, by taking into account these observations, it was proposed that Ang-(1-9) rather than Ang-(1-7) acts as a counter-regulator of Ang II in this model of heart failure<sup>[105]</sup>.

Ang (1-9) does regulate cardiac hypertrophy both *in vivo* in addition to *in vitro*<sup>[61,63]</sup>. In rats with MI that received vehicle, enalapril, or candesartan during 8 wk, Ang (1-9) did prevent myocardial hypertrophy and increased plasma Ang-(1-9) circulating levels by several fold<sup>[61]</sup>. Besides, in those experiments, Ang-(1-9) plasma levels correlated inversely with several markers of cardiac hypertrophy, even by adjusting for reduction of blood pressure<sup>[61]</sup>. This observed action was very

specific, since no correlation was found between cardiac hypertrophy with Ang-(1-7), nor with Ang II neither with the bradykinin levels. In other experiments, chronic treatment with Ang-(1-9) to rats with MI by using osmotic pumps diminished circulating levels of Ang II and enzymatic activity of the ACE and also to prevented myocardial hypertrophy<sup>[61]</sup>. Since there are available *in vitro* data showing that Ang-(1-9) incubation with ACE does generate Ang-(1-7)<sup>[76]</sup>, and this peptide negatively regulates hypertrophy<sup>[127,128]</sup>, the blocker of the Mas receptor A779 was utilized in order to assess whether Ang-(1-7) could intervene in the actions of Ang-(1-9). Even though A779 did augment blood levels of Ang-(1-7) by almost 3 fold, this specific blocker did not alter the Ang-(1-9) suppression effect on cardiomyocyte (CM) hypertrophy secondary to MI<sup>[61]</sup>. In experiments using cultured CM incubated with noradrenaline, IGF-1<sup>[61]</sup> or Ang II<sup>[63]</sup>, Ang-(1-9) prevented hypertrophy of cardiac cells and this action was mediated through the AT2R<sup>[63]</sup>. In addition, by the same AT2R mediated mechanism, Ang-(1-9) treatment did alleviate streptozotocin (STZ) induced cardiomyopathy dose dependently and did attenuate cardiac dysfunction in rats with diabetes induced by STZ<sup>[129]</sup>.

Recently, it has been described that long term treatment with Ang-(1-9) significantly reduced HT and hypertensive cardiovascular damage in two experimental models: the Ang II infusion model and the Goldblatt model (2K-1C) as well<sup>[84]</sup>. In these experiments, Ang-(1-9) also blunted the modifications in LV systolic function (ejection fraction) in both hypertensive models, without having an effect in the control rats<sup>[84]</sup>. Co-administration of Ang-(1-9) together with A779 did not modify the antihypertensive capability of Ang-(1-9) but PD123319, a specific AT2R antagonist, did entirely abolish the favorable effect of Ang-(1-9) on hypertension and on cardiovascular remodeling<sup>[84]</sup>. In cultured cardiac rat fibroblasts, we have recently observed that Ang-(1-9) was able to reduce fibroblast proliferation promoted throughout Ang II and also collagen content with no effects on differentiation of fibroblasts onto myofibroblasts<sup>[84]</sup>. The biological effects of Ang-(1-9) on hypertensive CV remodeling were corroborated in the rat with spontaneous hypertension which is stroke-prone (SHRSP)<sup>[130]</sup>. Those facts demonstrate that activation of the AT2R produced by Ang-(1-9) has a significant myocardial antifibrotic effect that may be associated to a direct effect on cardiac fibroblasts. This preclinical findings suggest a possible clinical approach for cardiovascular complications from hypertension, by stimulating the AT2R using Ang-(1-9) and obtaining in this way antihypertrophic and antifibrotic protective effects.

The AT2R stimulation activates among other mechanisms the NO-cGMP dependent pathway<sup>[131]</sup>. This happens through direct or indirect effects *via* bradykinins or by augmented activity or expression of endothelial NOS<sup>[131]</sup>. Additionally, AT2R activation might be able to induce relaxation by inverse regulation of the

Rho kinase pathway in the vascular wall<sup>[132]</sup>.

The release of endothelial vasodilators in response to Ang-(1-9) may be a mechanism involved in the beneficial consequences of Ang-(1-9) observed in hypertensive rodents. In *ex vivo* resistance arteries from Ang II treated rats, it has been observed that Ang-(1-9) does preserve relaxation induced by Ach (which is dependent from endothelium)<sup>[84]</sup>. Ang-(1-9) did also augment the concentration of eNOS mRNA in the aortic wall, which is associated to higher plasma concentrations of nitrate. These observed effects of Ang-(1-9) were completely inhibited by using PD123319, which is coherent with the concept that Ang-(1-9) does increase bioavailability of NO through a mechanism mediated by the AT2R<sup>[84]</sup>. In keeping with these results by blocking Enos a significantly increased contractile response to Phe in aortic rings of SHRSP chronically treated with Ang-(1-9) infusion has been found<sup>[130]</sup>. Furthermore, Ang-(1-9) does stimulate secretion of ANP without modifying the atrial contractility<sup>[83]</sup> and this observed effect of Ang-(1-9) is attenuated by using an AT2R antagonist but not when using AT1R nor MasR pharmacologic antagonism. Furthermore, by using inhibitors of phosphatidylinositol 3-kinase (PI3K), nitric oxide synthase (NOS), Akt, or soluble guanylyl cyclase, Ang-(1-9)-induced ANP secretion is blocked. The above-mentioned observations consistently suggest that the Ang-(1-9) peptide does stimulate secretion of ANP throughout the AT2R-PI3K-Akt-NO-cGMP cascade<sup>[83]</sup>. The release of arachidonic acid - another potent vasodilator - may be also actively implicated here, in addition to the NO<sup>[133]</sup>.

Regulation of Ang-(1-9) by Rho kinase was assessed by Ocaranza and coworkers for first time in hypertensive DOCA-salt rats, by inhibiting Rho-kinase with fasudil<sup>[18]</sup>. In the above mentioned experimental model, it was noticed that over expression of genes promoting cardiovascular remodeling such as transforming growth factor 1, PAI-1 and the MCP-1 molecule were lower by using the specific Rho kinase inhibitor, whereas both ACE2 enzymatic activity and blood levels of Ang-(1-9) were substantially increased. Remarkably, the changes observed in ACE, ACE2 and in the levels of Ang-(1-9) were clearly observed in both experimental groups receiving fasudil (the sham group and the DOCA group with hypertension)<sup>[18]</sup>. Thus, this new action of Rho kinase inhibition on ACE2 (gene expression/enzymatic activity) in addition to lowering Ang-(1-9) levels might also conduce to its salutary effects in HT, atherosclerosis disease, and CV remodeling.

AT2R agonists could represent a new class of drugs aimed to preclude and reverse hypertensive CV remodeling. In isolated conductance and resistance vessels, several investigators have shown that CGP42112A (a peptide agonist)<sup>[134,135]</sup> and more recently the agonist C21<sup>[136]</sup> induce vasorelaxation, consistent with the concept of AT2R opposing to the AT1R. A recent study assessed AT2R-stimulation with C21 on post-MI cardiac function<sup>[137]</sup>. Treatment using C21 began

24 h after MI and it was administered during 7 d. The aforementioned treatment with C21 diminished the scar size and this reduction was due to a favorable effect of C21 on myocardial remodeling post MI that also did improve systolic and diastolic cardiac dysfunction. Besides, C21 treatment diminished the content of inflammatory cytokines (IL-1b, IL-2, IL-6) and pro-apoptotic markers (caspase-3, Fas-ligand). Moreover, both the monocyte chemoattractant-protein 1 and levels of myeloperoxidase, a biomarker of oxidative stress, were significantly decreased by C21<sup>[137]</sup>. Therefore, AT2R stimulation (direct) did improve cardiac function following an experimental MI throughout both anti-inflammatory and anti-oxidant mechanisms and by a more favorable scar remodeling.

Amelioration of inflammation seems to be a fundamental mechanism of action of AT2R-agonism<sup>[138]</sup>. It has been recently observed that AT2R-stimulation with C21 inhibits NF-κB activity and subsequent synthesis of interleukin 6 and other cytokines that promote inflammation by activating tyrosine-phosphatases, serine/threonine-phosphatases and also CYP2C/2J enzymes leading in this way to increased 11, 12-EET synthesis<sup>[91]</sup>. 11,12-EET has been shown by Node *et al.*<sup>[139]</sup> in 1999 to have anti-inflammatory characteristics by inhibiting NF-κB. The fact that the described action of C21 on IL-6 promoter activity was similar in strength to the effect of hydrocortisone administered at an equivalent dose indicates that the AT2R could be clinically useful as a beneficial target in cardiovascular disease, and also in inflammatory clinical conditions<sup>[140]</sup>; an hypothesis which needs to be tested in future experiments.

**Alamandine-MrgD axis:** Alamandine produces vasorelaxation in phenylephrine-contracted aortic rings and when microinjected into central areas critically involved in BP control, such as caudal ventrolateral and rostral ventrolateral medulla. Alamandine produces a decrease vs increase in BP in the former vs latter, respectively, revealing that it acts locally and centrally in a rather similar manner with respect to Ang-(1-7) (Figure 4). Additionally, oral alamandine administration produces a longstanding antihypertensive effect in SHR rats<sup>[62]</sup>, suggesting a therapeutic potential for the conditions with underlying cardiovascular remodeling. Furthermore, alamandine has direct effects on remodeling by diminishing cardiac deposition of collagen as well as fibronectin in the rat model of myocardial fibrosis induced by isoproterenol<sup>[62]</sup>. Since alamandine is a very new molecule within the RAS, currently there are scant data about its participation in disease, although it has been observed elevated plasma alamandine concentrations in subjects with nephropathy, suggesting that alamandine may be involved in some pathophysiological conditions<sup>[62]</sup>.

The identification of alamandine and its MrgDR contributes to provide novel insights to the knowledge of the RAS pathophysiology and also opens new

possibilities in order to develop therapeutic approaches oriented to prevent/treat CV remodeling, particularly in hypertension.

## POSSIBLE ROLE OF ESTROGENS IN PREVENTING HYPERTENSIVE CARDIAC HYPERTROPHY THROUGH MYOCARDIAL CHYMASE AND ANGIOTENSIN II

Since cardiovascular disease in females is consistently increasing, it becomes relevant to better know the connections among age, gender, and cardiovascular health status in a more precise way. With the assumption that estrogens do prevent pathological cardiac remodeling secondary to pressure overload throughout inhibition of mast cell chymase release, Li *et al.*<sup>[141]</sup> recently performed thoracic aortic constriction in intact and in ovariectomized rats (an experimental model resembling postmenopausal hypertension). Three days previously to the aortic constriction surgery, ovariectomized rats began to receive 17 $\beta$ -Estradiol, an inhibitor of the chymase (or a mast cell stabilizer)<sup>[141]</sup>. Their main findings were that both density and degranulation of the mast cells, circulating chymase levels - able to hydrolyze angiotensin I onto the octapeptide angiotensin II - and cardiac transforming growth factor-1 were augmented due to the constriction procedure in ovariectomized rats and that replacement therapy with estrogens significantly diminished the cardiac levels of increased chymase, both degranulation and density of the mast cells, circulating chymase levels and active transforming growth factor-1 in the myocardium as well<sup>[141]</sup>. They also observed in this experimental model that estrogens did prevent myocardial hypertrophy and fibrosis<sup>[141]</sup> (which is rather similar to cardiac failure with normal systolic performance commonly associated to hypertensive diastolic dysfunction)<sup>[142]</sup>. By using the above mentioned experimental model they concluded that the mast cell derived chymase release, which is able to be inhibited by estrogens, is responsible for myocardial protection in the case of adverse cardiac remodeling resulting from transverse aortic constriction<sup>[141]</sup>.

Novel observations derived from experimental models that emulate the cardiovascular phenotype of women having reduced estrogens levels (such in early ovarian failure or in postmenopause), including the ovariectomized congenic mRen2. Lewis rat and transverse thoracic aortic constriction in ovariectomized rats models provide substantial data in the sense that estrogens actively modulate the tissue renin-angiotensin-aldosterone system, and signaling intracellular pathways related to NOS, in some measure through the G protein-coupled receptor 30 (in the membrane), additionally identified as the G protein-coupled estrogen receptor 1<sup>[143]</sup> which might also be connected to the pro remodeling Rho A/Rho kinase signaling pathway<sup>[142]</sup>.

Current limitations and challenges: More potent ROCK inhibition is a main challenge at this time even though recent preclinical evidence of newer inhibitors in this regard is available<sup>[15,19]</sup>. As a consequence, clinical studies with ROCK inhibitors in hypertension will follow. In the field of the vasodilatory peptides from the RAS, Ang 1-9 is effective as antihypertensive and anti-remodeling. However, human data are necessary as well as pharmacodynamic information and means for appropriate delivery. With relationship to the implications of the estrogens-myocardial chymase interaction, from our point of view, more preclinical data are required since the number of studies is small.

## CONCLUSION

The discussed evidences in this review about the three aforementioned novel mechanisms of hypertensive myocardial remodeling: the Rho kinase intracellular signaling pathway, the vasodilatory peptides from the RAS and the estrogens-myocardial chymase interaction, open new therapeutic opportunities to effectively get better quality of life, reduce/avoid hypertensive cardiovascular remodeling and residual hypertensive risk.

## REFERENCES

- 1 **Thomopoulos C**, Parati G, Zanchetti A. Effects of blood pressure-lowering on outcome incidence in hypertension: 5. Head-to-head comparisons of various classes of antihypertensive drugs - overview and meta-analyses. *J Hypertens* 2015; **33**: 1321-1341 [PMID: 26039526 DOI: 10.1097/HJH.0000000000000614]
- 2 **Jalil JE**, Jalil R, Ocaranza MP. Combined rho kinase and renin-angiotensin system inhibition: a new therapeutic perspective for renal and cardiovascular remodeling. *Hypertens Res* 2010; **33**: 883-885 [PMID: 20664549 DOI: 10.1038/hr.2010.129]
- 3 **Nakagawa O**, Fujisawa K, Ishizaki T, Saito Y, Nakao K, Narumiya S. ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serine/threonine kinase in mice. *FEBS Lett* 1996; **392**: 189-193 [PMID: 8772201 DOI: 10.1016/0014-5793(96)00811-3]
- 4 **Shimokawa H**, Takeshita A. Rho-kinase is an important therapeutic target in cardiovascular medicine. *Arterioscler Thromb Vasc Biol* 2005; **25**: 1767-1775 [PMID: 16002741 DOI: 10.1161/01.ATV.0000176193.83629.c8]
- 5 **Jalil J**, Lavandero S, Chiong M, Ocaranza MP. [Rho/Rho kinase signal transduction pathway in cardiovascular disease and cardiovascular remodeling]. *Rev Esp Cardiol* 2005; **58**: 951-961 [PMID: 16053829 DOI: 10.1157/13078132]
- 6 **Shimokawa H**, Rashid M. Development of Rho-kinase inhibitors for cardiovascular medicine. *Trends Pharmacol Sci* 2007; **28**: 296-302 [PMID: 17482681 DOI: 10.1016/j.tips.2007.04.006]
- 7 **Watanabe H**, Iino K, Ito H. Rho-kinase in leukocytes. An emerging biomarker for heart failure. *Circ J* 2013; **77**: 2471-2472 [PMID: 23912838 DOI: 10.1253/circj.CJ-13-0935]
- 8 **Shi J**, Wei L. Rho kinases in cardiovascular physiology and pathophysiology: the effect of fasudil. *J Cardiovasc Pharmacol* 2013; **62**: 341-354 [PMID: 23921309 DOI: 10.1097/FJC.0b013e3182a3718f]
- 9 **Julian L**, Olson MF. Rho-associated coiled-coil containing kinases (ROCK): structure, regulation, and functions. *Small GTPases* 2014; **5**: e29846 [PMID: 25010901 DOI: 10.4161/sgtp.29846]
- 10 **Carbone ML**, Brégeon J, Devos N, Chadeuf G, Blanchard A, Azizi M, Pacaud P, Jeunemaître X, Loirand G. Angiotensin II activates the RhoA exchange factor Arhgef1 in humans. *Hypertension*



- 2015; **65**: 1273-1278 [PMID: 25870189 DOI: 10.1161/HYPERTENSIONAHA.114.05065]
- 11 **Dong M**, Yan BP, Liao JK, Lam YY, Yip GW, Yu CM. Rho-kinase inhibition: a novel therapeutic target for the treatment of cardiovascular diseases. *Drug Discov Today* 2010; **15**: 622-629 [PMID: 20601092 DOI: 10.1016/j.drudis.2010.06.011]
- 12 **Zhou Q**, Gensch C, Liao JK. Rho-associated coiled-coil-forming kinases (ROCKs): potential targets for the treatment of atherosclerosis and vascular disease. *Trends Pharmacol Sci* 2011; **32**: 167-173 [PMID: 21242007 DOI: 10.1016/j.tips.2010.12.006]
- 13 **Loirand G**, Pacaud P. Involvement of Rho GTPases and their regulators in the pathogenesis of hypertension. *Small GTPases* 2014; **5**: 1-10 [PMID: 25496262 DOI: 10.4161/sgtp.28846]
- 14 **Uehata M**, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature* 1997; **389**: 990-994 [PMID: 9353125]
- 15 **Löhn M**, Plettenburg O, Ivashchenko Y, Kannt A, Hofmeister A, Kadereit D, Schaefer M, Linz W, Kohlmann M, Herbert JM, Janiak P, O'Connor SE, Ruetten H. Pharmacological characterization of SAR407899, a novel rho-kinase inhibitor. *Hypertension* 2009; **54**: 676-683 [PMID: 19597037 DOI: 10.1161/HYPERTENSIONAHA.109.134353]
- 16 **Sehon CA**, Wang GZ, Viet AQ, Goodman KB, Dowdell SE, Elkins PA, Semus SF, Evans C, Jolivet LJ, Kirkpatrick RB, Dul E, Khandekar SS, Yi T, Wright LL, Smith GK, Behm DJ, Bentley R, Doe CP, Hu E, Lee D. Potent, selective and orally bioavailable dihydropyrimidine inhibitors of Rho kinase (ROCK1) as potential therapeutic agents for cardiovascular diseases. *J Med Chem* 2008; **51**: 6631-6634 [PMID: 18842034 DOI: 10.1021/jm8005096]
- 17 **Kast R**, Schirok H, Figueroa-Pérez S, Mittendorf J, Gnath MJ, Apeler H, Lenz J, Franz JK, Knorr A, Hütter J, Lobell M, Zimmermann K, Münter K, Augstein KH, Ehmke H, Stasch JP. Cardiovascular effects of a novel potent and highly selective azaindole-based inhibitor of Rho-kinase. *Br J Pharmacol* 2007; **152**: 1070-1080 [PMID: 17934515]
- 18 **Ocaranza MP**, Rivera P, Novoa U, Pinto M, González L, Chiong M, Lavandero S, Jalil JE. Rho kinase inhibition activates the homologous angiotensin-converting enzyme-angiotensin-(1-9) axis in experimental hypertension. *J Hypertens* 2011; **29**: 706-715 [PMID: 21330937 DOI: 10.1097/HJH.0b013e3283440665]
- 19 **Löhn M**, Plettenburg O, Kannt A, Kohlmann M, Hofmeister A, Kadereit D, Monecke P, Schiffer A, Schulte A, Ruetten H, Ivashchenko Y. End-organ protection in hypertension by the novel and selective Rho-kinase inhibitor, SAR407899. *World J Cardiol* 2015; **7**: 31-42 [PMID: 25632317 DOI: 10.4330/wjc.v7.i1.31]
- 20 **Dhaliwal JS**, Casey DB, Greco AJ, Badejo AM, Gallen TB, Murthy SN, Nossaman BD, Hyman AL, Kadowitz PJ. Rho kinase and Ca<sup>2+</sup> entry mediate increased pulmonary and systemic vascular resistance in L-NAME-treated rats. *Am J Physiol Lung Cell Mol Physiol* 2007; **293**: L1306-L1313 [PMID: 17766587]
- 21 **Grisk O**. Potential benefits of rho-kinase inhibition in arterial hypertension. *Curr Hypertens Rep* 2013; **15**: 506-513 [PMID: 23852615 DOI: 10.1007/s11906-013-0373-0]
- 22 **Masumoto A**, Hirooka Y, Shimokawa H, Hironaga K, Setoguchi S, Takeshita A. Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. *Hypertension* 2001; **38**: 1307-1310 [PMID: 11751708]
- 23 **Smith CJ**, Santhanam L, Alexander LM. Rho-Kinase activity and cutaneous vasoconstriction is upregulated in essential hypertensive humans. *Microvasc Res* 2013; **87**: 58-64 [PMID: 23481864 DOI: 10.1016/j.mvr.2013.02.005]
- 24 **Rivera P**, Ocaranza MP, Lavandero S, Jalil JE. Rho kinase activation and gene expression related to vascular remodeling in normotensive rats with high angiotensin I converting enzyme levels. *Hypertension* 2007; **50**: 792-798 [PMID: 17785632 DOI: 10.1161/HYPERTENSIONAHA.107.095117]
- 25 **Seko T**, Ito M, Kureishi Y, Okamoto R, Moriki N, Onishi K, Isaka N, Hartshorne DJ, Nakano T. Activation of RhoA and inhibition of myosin phosphatase as important components in hypertension in vascular smooth muscle. *Circ Res* 2003; **92**: 411-418 [PMID: 12600888 DOI: 10.1161/01.RES.0000059987.90200.44]
- 26 **Tsounapi P**, Saito M, Kitatani K, Dimitriadis F, Ohmasa F, Shimizu S, Kinoshita Y, Takenaka A, Satoh K. Fasudil improves the endothelial dysfunction in the aorta of spontaneously hypertensive rats. *Eur J Pharmacol* 2012; **691**: 182-189 [PMID: 22819709 DOI: 10.1016/j.ejphar.2012.07.016]
- 27 **Hassona MD**, Abouelnaga ZA, Elnakish MT, Awad MM, Alhaj M, Goldschmidt-Clermont PJ, Hassanain H. Vascular hypertrophy-associated hypertension of profilin1 transgenic mouse model leads to functional remodeling of peripheral arteries. *Am J Physiol Heart Circ Physiol* 2010; **298**: H2112-H2120 [PMID: 20400688 DOI: 10.1152/ajpheart.00016.2010]
- 28 **Chan CK**, Mak JC, Man RY, Vanhoutte PM. Rho kinase inhibitors prevent endothelium-dependent contractions in the rat aorta. *J Pharmacol Exp Ther* 2009; **329**: 820-826 [PMID: 19193928 DOI: 10.1124/jpet.108.148247]
- 29 **Takeda K**, Ichiki T, Tokunou T, Iino N, Fujii S, Kitabatake A, Shimokawa H, Takeshita A. Critical role of Rho-kinase and MEK/ERK pathways for angiotensin II-induced plasminogen activator inhibitor type-1 gene expression. *Arterioscler Thromb Vasc Biol* 2001; **21**: 868-873 [PMID: 11348889 DOI: 10.1161/01.ATV.21.5.868]
- 30 **Rikitake Y**, Liao JK. Rho-kinase mediates hyperglycemia-induced plasminogen activator inhibitor-1 expression in vascular endothelial cells. *Circulation* 2005; **111**: 3261-3268 [PMID: 15956119 DOI: 10.1161/CIRCULATIONAHA.105.534024]
- 31 **Sun Q**, Yue P, Ying Z, Cardounel AJ, Brook RD, Devlin R, Hwang JS, Zweier JL, Chen LC, Rajagopalan S. Air pollution exposure potentiates hypertension through reactive oxygen species-mediated activation of Rho/ROCK. *Arterioscler Thromb Vasc Biol* 2008; **28**: 1760-1766 [PMID: 18599801 DOI: 10.1161/ATVBAHA.108.166967]
- 32 **Higashi M**, Shimokawa H, Hattori T, Hiroki J, Mukai Y, Morikawa K, Ichiki T, Takahashi S, Takeshita A. Long-term inhibition of Rho-kinase suppresses angiotensin II-induced cardiovascular hypertrophy in rats in vivo: effect on endothelial NAD(P)H oxidase system. *Circ Res* 2003; **93**: 767-775 [PMID: 14500337]
- 33 **Satoh K**, Fukumoto Y, Shimokawa H. Rho-kinase: important new therapeutic target in cardiovascular diseases. *Am J Physiol Heart Circ Physiol* 2011; **301**: H287-H296 [PMID: 21622831 DOI: 10.1152/ajpheart.00327.2011]
- 34 **Ito K**, Hirooka Y, Sakai K, Kishi T, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in brain stem contributes to blood pressure regulation via sympathetic nervous system: possible involvement in neural mechanisms of hypertension. *Circ Res* 2003; **92**: 1337-1343 [PMID: 12791705]
- 35 **Ito K**, Hirooka Y, Kishi T, Kimura Y, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in the brainstem contributes to hypertension caused by chronic nitric oxide synthase inhibition. *Hypertension* 2004; **43**: 156-162 [PMID: 14732730]
- 36 **Mita S**, Kobayashi N, Yoshida K, Nakano S, Matsuoka H. Cardioprotective mechanisms of Rho-kinase inhibition associated with eNOS and oxidative stress-LOX-1 pathway in Dahl salt-sensitive hypertensive rats. *J Hypertens* 2005; **23**: 87-96 [PMID: 15643129]
- 37 **Ishimaru K**, Ueno H, Kagitani S, Takabayashi D, Takata M, Inoue H. Fasudil attenuates myocardial fibrosis in association with inhibition of monocyte/macrophage infiltration in the heart of DOCA/salt hypertensive rats. *J Cardiovasc Pharmacol* 2007; **50**: 187-194 [PMID: 17703135]
- 38 **Fukui S**, Fukumoto Y, Suzuki J, Saji K, Nawata J, Tawara S, Shinozaki T, Kagaya Y, Shimokawa H. Long-term inhibition of Rho-kinase ameliorates diastolic heart failure in hypertensive rats. *J Cardiovasc Pharmacol* 2008; **51**: 317-326 [PMID: 18356698 DOI: 10.1097/FJC.0b013e31816533b7]
- 39 **Phrommintikul A**, Tran L, Kompa A, Wang B, Adrahtas A, Cantwell D, Kelly DJ, Krum H. Effects of a Rho kinase inhibitor on pressure overload induced cardiac hypertrophy and



- associated diastolic dysfunction. *Am J Physiol Heart Circ Physiol* 2008; **294**: H1804-H1814 [PMID: 18245565 DOI: 10.1152/ajpheart.01078.2007]
- 40 **Shi J**, Zhang YW, Summers LJ, Dorn GW, Wei L. Disruption of ROCK1 gene attenuates cardiac dilation and improves contractile function in pathological cardiac hypertrophy. *J Mol Cell Cardiol* 2008; **44**: 551-560 [PMID: 18178218 DOI: 10.1016/j.yjmcc.2007.11.018]
- 41 **Del Re DP**, Miyamoto S, Brown JH. RhoA/Rho kinase up-regulate Bax to activate a mitochondrial death pathway and induce cardiomyocyte apoptosis. *J Biol Chem* 2007; **282**: 8069-8078 [PMID: 17234627]
- 42 **Mizutani H**, Okamoto R, Moriki N, Konishi K, Taniguchi M, Fujita S, Dohi K, Onishi K, Suzuki N, Satoh S, Makino N, Itoh T, Hartshorne DJ, Ito M. Overexpression of myosin phosphatase reduces Ca(2+) sensitivity of contraction and impairs cardiac function. *Circ J* 2010; **74**: 120-128 [PMID: 19966500]
- 43 **Liu PY**, Chen JH, Lin LJ, Liao JK. Increased Rho kinase activity in a Taiwanese population with metabolic syndrome. *J Am Coll Cardiol* 2007; **49**: 1619-1624 [PMID: 17433952]
- 44 **Li CB**, Li XX, Chen YG, Gao HQ, Bao CM, Liu XQ, Bu PL, Zhang J, Zhang Y, Ji XP. Myocardial remodeling in rats with metabolic syndrome: role of Rho-kinase mediated insulin resistance. *Acta Biochim Pol* 2012; **59**: 249-254 [PMID: 22693684]
- 45 **Hidaka T**, Hata T, Soga J, Fujii Y, Idei N, Fujimura N, Kihara Y, Noma K, Liao JK, Higashi Y. Increased leukocyte rho kinase (ROCK) activity and endothelial dysfunction in cigarette smokers. *Hypertens Res* 2010; **33**: 354-359 [PMID: 20139919 DOI: 10.1038/hr.2010.3]
- 46 **Hata T**, Goto C, Soga J, Hidaka T, Fujii Y, Idei N, Fujimura N, Maruhashi T, Mikami S, Kihara Y, Chayama K, Noma K, Liao JK, Higashi Y. Measurement of Rho-associated kinase (ROCK) activity in humans: validity of leukocyte p-MBS/t-MBS in comparison with vascular response to fasudil. *Atherosclerosis* 2011; **214**: 117-121 [PMID: 21035804 DOI: 10.1016/j.atherosclerosis.2010.10.005]
- 47 **Gabrielli L**, Winter JL, Godoy I, McNab P, Padilla I, Cordova S, Rigotti P, Novoa U, Mora I, García L, Ocaranza MP, Jalil JE. Increased rho-kinase activity in hypertensive patients with left ventricular hypertrophy. *Am J Hypertens* 2014; **27**: 838-845 [PMID: 24363278 DOI: 10.1093/ajh/hpt234]
- 48 **Gabrielli L**, Berkovitz A, Mora I, Novoa U, Godoy I, MacNab P, Córdova S, Padilla I, Rigotti P, García L, Lavandero S, Ocaranza MP, Jalil J. Increased Rho-kinase activity in peripheral leukocytes is associated to oxidative stress and decreased compliance of the arterial wall in diabetic hypertensive patients. *Rev Chil Cardiol* 2010; **30**: 34-41
- 49 **Ocaranza MP**, Gabrielli L, Mora I, García L, McNab P, Godoy I, Braun S, Córdova S, Castro P, Novoa U, Chiong M, Lavandero S, Jalil JE. Markedly increased Rho-kinase activity in circulating leukocytes in patients with chronic heart failure. *Am Heart J* 2011; **161**: 931-937 [PMID: 21570525]
- 50 **Kajikawa M**, Noma K, Maruhashi T, Mikami S, Iwamoto Y, Iwamoto A, Matsumoto T, Hidaka T, Kihara Y, Chayama K, Nakashima A, Goto C, Liao JK, Higashi Y. Rho-associated kinase activity is a predictor of cardiovascular outcomes. *Hypertension* 2014; **63**: 856-864 [PMID: 24379190 DOI: 10.1161/HYPERTENSIONAHA.113.02296]
- 51 **Hata T**, Soga J, Hidaka T, Idei N, Fujii Y, Fujimura N, Mikami S, Maruhashi T, Kihara Y, Chayama K, Kato H, Noma K, Liao JK, Higashi Y. Calcium channel blocker and Rho-associated kinase activity in patients with hypertension. *J Hypertens* 2011; **29**: 373-379 [PMID: 21063203 DOI: 10.1097/HJH.0b013e328340902d]
- 52 **Fujimura N**, Noma K, Hata T, Soga J, Hidaka T, Idei N, Fujii Y, Mikami S, Maruhashi T, Iwamoto Y, Kihara Y, Chayama K, Kato H, Liao JK, Higashi Y. Mineralocorticoid receptor blocker eplerenone improves endothelial function and inhibits Rho-associated kinase activity in patients with hypertension. *Clin Pharmacol Ther* 2012; **91**: 289-297 [PMID: 22205191 DOI: 10.1038/clpt.2011.227]
- 53 **Guo R**, Su Y, Yan J, Sun H, Wu J, Liu W, Xu Y. Fasudil improves short-term echocardiographic parameters of diastolic function in patients with type 2 diabetes with preserved left ventricular ejection fraction: a pilot study. *Heart Vessels* 2015; **30**: 89-97 [PMID: 24390764 DOI: 10.1007/s00380-013-0458-3]
- 54 **Dong M**, Liao JK, Fang F, Lee AP, Yan BP, Liu M, Yu CM. Increased Rho kinase activity in congestive heart failure. *Eur J Heart Fail* 2012; **14**: 965-973 [PMID: 22588320 DOI: 10.1093/eurjhf/hfs068]
- 55 **Dong M**, Liao JK, Yan B, Li R, Zhang M, Yu CM. A combination of increased Rho kinase activity and N-terminal pro-B-type natriuretic peptide predicts worse cardiovascular outcome in patients with acute coronary syndrome. *Int J Cardiol* 2013; **167**: 2813-2819 [PMID: 22921817 DOI: 10.1016/j.ijcard.2012.07.007]
- 56 **Intengan HD**, Schiffrin EL. Structure and mechanical properties of resistance arteries in hypertension: role of adhesion molecules and extracellular matrix determinants. *Hypertension* 2000; **36**: 312-318 [PMID: 10988257 DOI: 10.1161/01.HYP.36.3.312]
- 57 **Unger T**. The angiotensin type 2 receptor: variations on an enigmatic theme. *J Hypertens* 1999; **17**: 1775-1786 [PMID: 10703869 DOI: 10.1097/00004872-199917121-00001]
- 58 **Ferrario CM**. Role of angiotensin II in cardiovascular disease therapeutic implications of more than a century of research. *J Renin Angiotensin Aldosterone Syst* 2006; **7**: 3-14 [PMID: 17083068 DOI: 10.3317/jraas.2006.003]
- 59 **de Gasparo M**, Catt KJ, Inagami T, Wright JW, Unger T. International union of pharmacology. XXIII. The angiotensin II receptors. *Pharmacol Rev* 2000; **52**: 415-472 [PMID: 10977869]
- 60 **Santos RA**, Simoes e Silva AC, Maric C, Silva DM, Machado RP, de Buhr I, Heringer-Walther S, Pinheiro SV, Lopes MT, Bader M, Mendes EP, Lemos VS, Campagnole-Santos MJ, Schultheiss HP, Speth R, Walther T. Angiotensin-(1-7) is an endogenous ligand for the G protein-coupled receptor Mas. *Proc Natl Acad Sci USA* 2003; **100**: 8258-8263 [PMID: 12829792 DOI: 10.1073/pnas.1432869100]
- 61 **Ocaranza MP**, Lavandero S, Jalil JE, Moya J, Pinto M, Novoa U, Apablaza F, Gonzalez L, Hernandez C, Varas M, Lopez R, Godoy I, Verdejo H, Chiong M. Angiotensin-(1-9) regulates cardiac hypertrophy in vivo and in vitro. *J Hypertens* 2010; **28**: 1054-1064 [PMID: 20411619 DOI: 10.1097/HJH.0b013e328335d291]
- 62 **Lautner RQ**, Villela DC, Fraga-Silva RA, Silva N, Verano-Braga T, Costa-Fraga F, Jankowski J, Jankowski V, Sousa F, Alzamora A, Soares E, Barbosa C, Kjeldsen S, Oliveira A, Braga J, Savergnini S, Maia G, Peluso AB, Passos-Silva D, Ferreira A, Alves F, Martins A, Raizada M, Paula R, Motta-Santos D, Klempin F, Pimenta A, Alenina N, Sinisterra R, Bader M, Campagnole-Santos MJ, Santos RA. Discovery and characterization of alamandine: a novel component of the renin-angiotensin system. *Circ Res* 2013; **112**: 1104-1111 [PMID: 23446738 DOI: 10.1161/CIRCRESAHA.113.301077]
- 63 **Flores-Muñoz M**, Smith NJ, Haggerty C, Milligan G, Nicklin SA. Angiotensin1-9 antagonises pro-hypertrophic signalling in cardiomyocytes via the angiotensin type 2 receptor. *J Physiol* 2011; **589**: 939-951 [PMID: 21173078 DOI: 10.1113/jphysiol.2010.203075]
- 64 **Welches WR**, Santos RA, Chappell MC, Brosnihan KB, Greene LJ, Ferrario CM. Evidence that prolyl endopeptidase participates in the processing of brain angiotensin. *J Hypertens* 1991; **9**: 631-638 [PMID: 1653799 DOI: 10.1097/00004872-199107000-00008]
- 65 **Chappell MC**, Pirro NT, Sykes A, Ferrario CM. Metabolism of angiotensin-(1-7) by angiotensin-converting enzyme. *Hypertension* 1998; **31**: 362-367 [PMID: 9453329 DOI: 10.1161/01.HYP.31.1.362]
- 66 **Welches WR**, Brosnihan KB, Ferrario CM. A comparison of the properties and enzymatic activities of three angiotensin processing enzymes: angiotensin converting enzyme, prolyl endopeptidase and neutral endopeptidase 24.11. *Life Sci* 1993; **52**: 1461-1480 [PMID: 8387132 DOI: 10.1016/0024-3205(93)90108-F]
- 67 **Ferrario CM**, Chappell MC. A new myocardial conversion of angiotensin I. *Curr Opin Cardiol* 1994; **9**: 520-526 [PMID: 7987030 DOI: 10.1097/00001573-199409000-00004]
- 68 **Tipnis SR**, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A human homolog of angiotensin-converting enzyme. Cloning and

- functional expression as a captopril-insensitive carboxypeptidase. *J Biol Chem* 2000; **275**: 33238-33243 [PMID: 10924499 DOI: 10.1074/jbc.M002615200]
- 69 **Ferreira AJ**, Santos RA. Cardiovascular actions of angiotensin-(1-7). *Braz J Med Biol Res* 2005; **38**: 499-507 [PMID: 15962175 DOI: 10.1590/S0100-879X2005000400003]
- 70 **Santos RA**, Ferreira AJ, Simões E Silva AC. Recent advances in the angiotensin-converting enzyme 2-angiotensin(1-7)-Mas axis. *Exp Physiol* 2008; **93**: 519-527 [PMID: 18310257 DOI: 10.1113/expphysiol.2008.042002]
- 71 **Giani JF**, Mayer MA, Muñoz MC, Silberman EA, Höcht C, Taira CA, Gironacci MM, Turyn D, Dominici FP. Chronic infusion of angiotensin-(1-7) improves insulin resistance and hypertension induced by a high-fructose diet in rats. *Am J Physiol Endocrinol Metab* 2009; **296**: E262-E271 [PMID: 19001546 DOI: 10.1152/ajpendo.90678.2008]
- 72 **Opafil S**, Sanders CA, Haber E. In-vivo and in-vitro conversion of angiotensin I to angiotensin II in dog blood. *Circ Res* 1970; **26**: 591-599 [PMID: 4315298 DOI: 10.1161/01.RES.26.5.591]
- 73 **Opafil S**, Tregear GW, Koerner T, Barnes BA, Haber E. Mechanism of pulmonary conversion of angiotensin I to angiotensin II in the dog. *Circ Res* 1971; **29**: 682-690 [PMID: 4331484]
- 74 **Jackman HL**, Massad MG, Sekosan M, Tan F, Brovkovich V, Marcic BM, Erdős EG. Angiotensin 1-9 and 1-7 release in human heart: role of cathepsin A. *Hypertension* 2002; **39**: 976-981 [PMID: 12019279]
- 75 **Kokkonen JO**, Saarinen J, Kovanen PT. Regulation of local angiotensin II formation in the human heart in the presence of interstitial fluid. Inhibition of chymase by protease inhibitors of interstitial fluid and of angiotensin-converting enzyme by Ang-(1-9) formed by heart carboxypeptidase A-like activity. *Circulation* 1997; **95**: 1455-1463 [PMID: 9118513]
- 76 **Donoghue M**, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, Donovan M, Woolf B, Robison K, Jeyaseelan R, Breitbart RE, Acton S. A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. *Circ Res* 2000; **87**: E1-E9 [PMID: 10969042]
- 77 **Rice GI**, Thomas DA, Grant PJ, Turner AJ, Hooper NM. Evaluation of angiotensin-converting enzyme (ACE), its homologue ACE2 and neprilysin in angiotensin peptide metabolism. *Biochem J* 2004; **383**: 45-51 [PMID: 15283675]
- 78 **Garabelli PJ**, Modrall JG, Penninger JM, Ferrario CM, Chappell MC. Distinct roles for angiotensin-converting enzyme 2 and carboxypeptidase A in the processing of angiotensins within the murine heart. *Exp Physiol* 2008; **93**: 613-621 [PMID: 18356559]
- 79 **Keidar S**, Kaplan M, Gamliel-Lazarovich A. ACE2 of the heart: From angiotensin I to angiotensin (1-7). *Cardiovasc Res* 2007; **73**: 463-469 [PMID: 17049503]
- 80 **Trask AJ**, Averill DB, Ganten D, Chappell MC, Ferrario CM. Primary role of angiotensin-converting enzyme-2 in cardiac production of angiotensin-(1-7) in transgenic Ren-2 hypertensive rats. *Am J Physiol Heart Circ Physiol* 2007; **292**: H3019-H3024 [PMID: 17308000]
- 81 **Schwacke JH**, Spainhour JC, Ierardi JL, Chaves JM, Arthur JM, Janech MG, Velez JC. Network modeling reveals steps in angiotensin peptide processing. *Hypertension* 2013; **61**: 690-700 [PMID: 23283355]
- 82 **Drummer OH**, Kourtis S, Johnson H. Effect of chronic enalapril treatment on enzymes responsible for the catabolism of angiotensin I and formation of angiotensin II. *Biochem Pharmacol* 1990; **39**: 513-518 [PMID: 2407244]
- 83 **Cha SA**, Park BM, Gao S, Kim SH. Stimulation of ANP by angiotensin-(1-9) via the angiotensin type 2 receptor. *Life Sci* 2013; **93**: 934-940 [PMID: 24177599]
- 84 **Ocaranza MP**, Moya J, Barrientos V, Alzamora R, Hevia D, Morales C, Pinto M, Escudero N, García L, Novoa U, Ayala P, Diaz-Araya G, Godoy I, Chiong M, Lavandero S, Jalil JE, Michea L. Angiotensin-(1-9) reverses experimental hypertension and cardiovascular damage by inhibition of the angiotensin converting enzyme/Ang II axis. *J Hypertens* 2014; **32**: 771-783 [PMID: 24463937]
- 85 **Paula RD**, Lima CV, Britto RR, Campagnole-Santos MJ, Khosla MC, Santos RA. Potentiation of the hypotensive effect of bradykinin by angiotensin-(1-7)-related peptides. *Peptides* 1999; **20**: 493-500 [PMID: 10458520]
- 86 **Etelvino GM**, Peluso AA, Santos RA. New components of the renin-angiotensin system: alamandine and the MAS-related G protein-coupled receptor D. *Curr Hypertens Rep* 2014; **16**: 433 [PMID: 24760442]
- 87 **Bedeck K**, Elbaz N, Sutren M, Masson M, Susini C, Strosberg AD, Nahmias C. Angiotensin II type 2 receptors mediate inhibition of mitogen-activated protein kinase cascade and functional activation of SHP-1 tyrosine phosphatase. *Biochem J* 1997; **325** (Pt 2): 449-454 [PMID: 9230127]
- 88 **Gava E**, Samad-Zadeh A, Zimpelmann J, Bahramifard N, Kitten GT, Santos RA, Touyz RM, Burns KD. Angiotensin-(1-7) activates a tyrosine phosphatase and inhibits glucose-induced signalling in proximal tubular cells. *Nephrol Dial Transplant* 2009; **24**: 1766-1773 [PMID: 19144997]
- 89 **Noet S**, Nahmias C. Signal transduction from the angiotensin II AT2 receptor. *Trends Endocrinol Metab* 2000; **11**: 1-6 [PMID: 10652498]
- 90 **Sampaio WO**, Henrique de Castro C, Santos RA, Schifffrin EL, Touyz RM. Angiotensin-(1-7) counterregulates angiotensin II signaling in human endothelial cells. *Hypertension* 2007; **50**: 1093-1098 [PMID: 17984366]
- 91 **Rompe F**, Artuc M, Hallberg A, Alterman M, Ströder K, Thöne-Reineke C, Reichenbach A, Schacherl J, Dahlöf B, Bader M, Alenina N, Schwaninger M, Zuberbier T, Funke-Kaiser H, Schmidt C, Schunck WH, Unger T, Steckelings UM. Direct angiotensin II type 2 receptor stimulation acts anti-inflammatory through epoxyeicosatrienoic acid and inhibition of nuclear factor kappaB. *Hypertension* 2010; **55**: 924-931 [PMID: 20157051]
- 92 **El-Hashim AZ**, Renno WM, Raghupathy R, Abdou HT, Akhtar S, Benter IF. Angiotensin-(1-7) inhibits allergic inflammation, via the MAS1 receptor, through suppression of ERK1/2- and NF-κB-dependent pathways. *Br J Pharmacol* 2012; **166**: 1964-1976 [PMID: 22339213]
- 93 **Gohlke P**, Pees C, Unger T. AT2 receptor stimulation increases aortic cyclic GMP in SHRSP by a kinin-dependent mechanism. *Hypertension* 1998; **31**: 349-355 [PMID: 9453327]
- 94 **Heitsch H**, Brovkovich S, Malinski T, Wiemer G. Angiotensin-(1-7)-Stimulated Nitric Oxide and Superoxide Release From Endothelial Cells. *Hypertension* 2001; **37**: 72-76 [PMID: 11208759]
- 95 **Stegbauer J**, Potthoff SA, Quack I, Mergia E, Clasen T, Friedrich S, Vonend O, Woznowski M, Königshausen E, Sellin L, Rump LC. Chronic treatment with angiotensin-(1-7) improves renal endothelial dysfunction in apolipoproteinE-deficient mice. *Br J Pharmacol* 2011; **163**: 974-983 [PMID: 21371005]
- 96 **Abdalla S**, Lother H, Abdel-tawab AM, Qitterer U. The angiotensin II AT2 receptor is an AT1 receptor antagonist. *J Biol Chem* 2001; **276**: 39721-39726 [PMID: 11507095]
- 97 **Kostenis E**, Milligan G, Christopoulos A, Sanchez-Ferrer CF, Heringer-Walther S, Sexton PM, Gembardt F, Kellett E, Martini L, Vanderheyden P, Schultheiss HP, Walther T. G-protein-coupled receptor Mas is a physiological antagonist of the angiotensin II type 1 receptor. *Circulation* 2005; **111**: 1806-1813 [PMID: 15809376]
- 98 **Milasta S**, Pediani J, Appelbe S, Trim S, Wyatt M, Cox P, Fidock M, Milligan G. Interactions between the Mas-related receptors MrgD and MrgE alter signalling and trafficking of MrgD. *Mol Pharmacol* 2006; **69**: 479-491 [PMID: 16282220]
- 99 **Shinohara T**, Harada M, Ogi K, Maruyama M, Fujii R, Tanaka H, Fukusumi S, Komatsu H, Hosoya M, Noguchi Y, Watanabe T, Moriya T, Itoh Y, Hinuma S. Identification of a G protein-coupled receptor specifically responsive to beta-alanine. *J Biol Chem* 2004; **279**: 23559-23564 [PMID: 15037633]
- 100 **Crozier RA**, Ajit SK, Kaftan EJ, Pausch MH. MrgD activation inhibits KCNQ/M-currents and contributes to enhanced neuronal excitability. *J Neurosci* 2007; **27**: 4492-4496 [PMID: 17442834]
- 101 **Dong X**, Han S, Zylka MJ, Simon MI, Anderson DJ. A diverse

- family of GPCRs expressed in specific subsets of nociceptive sensory neurons. *Cell* 2001; **106**: 619-632 [PMID: 11551509]
- 102 **Rau KK**, McIlwraith SL, Wang H, Lawson JJ, Jankowski MP, Zylka MJ, Anderson DJ, Koerber HR. Mrgprd enhances excitability in specific populations of cutaneous murine polymodal nociceptors. *J Neurosci* 2009; **29**: 8612-8619 [PMID: 19571152]
- 103 **Nishimura S**, Uno M, Kaneta Y, Fukuchi K, Nishigohri H, Hasegawa J, Komori H, Takeda S, Enomoto K, Nara F, Agatsuma T. MRGD, a MAS-related G-protein coupled receptor, promotes tumorigenesis and is highly expressed in lung cancer. *PLoS One* 2012; **7**: e38618 [PMID: 22715397]
- 104 **Gembardt F**, Grajewski S, Vahl M, Schultheiss HP, Walther T. Angiotensin metabolites can stimulate receptors of the Mas-related genes family. *Mol Cell Biochem* 2008; **319**: 115-123 [PMID: 18636314]
- 105 **Ocaranza MP**, Godoy I, Jalil JE, Varas M, Collantes P, Pinto M, Roman M, Ramirez C, Copaja M, Diaz-Araya G, Castro P, Lavandero S. Enalapril attenuates downregulation of Angiotensin-converting enzyme 2 in the late phase of ventricular dysfunction in myocardial infarcted rat. *Hypertension* 2006; **48**: 572-578 [PMID: 16908757]
- 106 **Luque M**, Martin P, Martell N, Fernandez C, Brosnihan KB, Ferrario CM. Effects of captopril related to increased levels of prostacyclin and angiotensin-(1-7) in essential hypertension. *J Hypertens* 1996; **14**: 799-805 [PMID: 8793704]
- 107 **Jennings DL**, Kalus JS, Coleman CI, Manierski C, Yee J. Combination therapy with an ACE inhibitor and an angiotensin receptor blocker for diabetic nephropathy: a meta-analysis. *Diabet Med* 2007; **24**: 486-493 [PMID: 17367311]
- 108 **Tikellis C**, Bialkowski K, Pete J, Sheehy K, Su Q, Johnston C, Cooper ME, Thomas MC. ACE2 deficiency modifies renoprotection afforded by ACE inhibition in experimental diabetes. *Diabetes* 2008; **57**: 1018-1025 [PMID: 18235039]
- 109 **Igase M**, Kohara K, Nagai T, Miki T, Ferrario CM. Increased expression of angiotensin converting enzyme 2 in conjunction with reduction of neointima by angiotensin II type 1 receptor blockade. *Hypertens Res* 2008; **31**: 553-559 [PMID: 18497476]
- 110 **Bosnyak S**, Jones ES, Christopoulos A, Aguilar MI, Thomas WG, Widdop RE. Relative affinity of angiotensin peptides and novel ligands at AT1 and AT2 receptors. *Clin Sci (Lond)* 2011; **121**: 297-303 [PMID: 21542804]
- 111 **Oliveri C**, Ocaranza MP, Campos X, Lavandero S, Jalil JE. Angiotensin I-converting enzyme modulates neutral endopeptidase activity in the rat. *Hypertension* 2001; **38**: 650-654 [PMID: 11566949]
- 112 **Ocaranza MP**, Palomera C, Román M, Bargetto J, Lavandero S, Jalil JE. Effect of hypertension on angiotensin-(1-7) levels in rats with different angiotensin-I converting enzyme polymorphism. *Life Sci* 2006; **78**: 1535-1542 [PMID: 16229862]
- 113 **Ocaranza MP**, Piddo AM, Faúndez P, Lavandero S, Jalil JE. Angiotensin I-converting enzyme gene polymorphism influences chronic hypertensive response in the rat Goldblatt model. *J Hypertens* 2002; **20**: 413-420 [PMID: 11875308]
- 114 **Ocaranza MP**, Diaz-Araya G, Carreño JE, Muñoz D, Riveros JP, Jalil JE, Lavandero S. Polymorphism in gene coding for ACE determines different development of myocardial fibrosis in rats. *Am J Physiol Heart Circ Physiol* 2004; **286**: H498-H506 [PMID: 14527934]
- 115 **Jalil JE**, Pérez A, Ocaranza MP, Bargetto J, Galaz A, Lavandero S. Increased aortic NADPH oxidase activity in rats with genetically high angiotensin-converting enzyme levels. *Hypertension* 2005; **46**: 1362-1367 [PMID: 16230508]
- 116 **Jalil JE**, Palomera C, Ocaranza MP, Godoy I, Román M, Chiong M, Lavandero S. Levels of plasma angiotensin-(1-7) in patients with hypertension who have the angiotensin-I-converting enzyme deletion/deletion genotype. *Am J Cardiol* 2003; **92**: 749-751 [PMID: 12972127]
- 117 **Jalil JE**, Ocaranza MP, Oliveri C, Córdova S, Godoy I, Chamorro G, Braun S, Fardella C, Michel JB, Lavandero S. Neutral endopeptidase and angiotensin I converting enzyme insertion/deletion gene polymorphism in humans. *J Hum Hypertens* 2004; **18**: 119-125 [PMID: 14730327]
- 118 **Ferrario CM**. ACE2: more of Ang-(1-7) or less Ang II? *Curr Opin Nephrol Hypertens* 2011; **20**: 1-6 [PMID: 21045683]
- 119 **Grace JA**, Klein S, Herath CB, Granzow M, Schierwagen R, Masing N, Walther T, Sauerbruch T, Burrell LM, Angus PW, Trebicka J. Activation of the MAS receptor by angiotensin-(1-7) in the renin-angiotensin system mediates mesenteric vasodilatation in cirrhosis. *Gastroenterology* 2013; **145**: 874-884.e5 [PMID: 23796456]
- 120 **Hernández Prada JA**, Ferreira AJ, Katovich MJ, Shenoy V, Qi Y, Santos RA, Castellano RK, Lampkins AJ, Gubala V, Ostrov DA, Raizada MK. Structure-based identification of small-molecule angiotensin-converting enzyme 2 activators as novel antihypertensive agents. *Hypertension* 2008; **51**: 1312-1317 [PMID: 18391097]
- 121 **Ferreira AJ**, Shenoy V, Qi Y, Fraga-Silva RA, Santos RA, Katovich MJ, Raizada MK. Angiotensin-converting enzyme 2 activation protects against hypertension-induced cardiac fibrosis involving extracellular signal-regulated kinases. *Exp Physiol* 2011; **96**: 287-294 [PMID: 21148624]
- 122 **Oudit GY**, Liu GC, Zhong J, Basu R, Chow FL, Zhou J, Loibner H, Janzek E, Schuster M, Penninger JM, Herzenberg AM, Kassiri Z, Scholey JW. Human recombinant ACE2 reduces the progression of diabetic nephropathy. *Diabetes* 2010; **59**: 529-538 [PMID: 19934006]
- 123 **Wiemer G**, Dobrucki LW, Louka FR, Malinski T, Heitsch H. AVE 0991, a nonpeptide mimic of the effects of angiotensin-(1-7) on the endothelium. *Hypertension* 2002; **40**: 847-852 [PMID: 12468568]
- 124 **Santos RA**, Frézard F, Ferreira AJ. Angiotensin-(1-7): blood, heart, and blood vessels. *Curr Med Chem Cardiovasc Hematol Agents* 2005; **3**: 383-391 [PMID: 16250869]
- 125 **Pinheiro SV**, Simões e Silva AC, Sampaio WO, de Paula RD, Mendes EP, Bontempo ED, Pesquero JB, Walther T, Alenina N, Bader M, Bleich M, Santos RA. Nonpeptide AVE 0991 is an angiotensin-(1-7) receptor Mas agonist in the mouse kidney. *Hypertension* 2004; **44**: 490-496 [PMID: 15326087]
- 126 **Shemesh R**, Toporik A, Levine Z, Hecht I, Rotman G, Wool A, Dahary D, Gofer E, Kliger Y, Soffer MA, Rosenberg A, Eshel D, Cohen Y. Discovery and validation of novel peptide agonists for G-protein-coupled receptors. *J Biol Chem* 2008; **283**: 34643-34649 [PMID: 18854305]
- 127 **Tallant EA**, Ferrario CM, Gallagher PE. Angiotensin-(1-7) inhibits growth of cardiac myocytes through activation of the mas receptor. *Am J Physiol Heart Circ Physiol* 2005; **289**: H1560-H1566 [PMID: 15951342]
- 128 **Grobe JL**, Mecca AP, Lingis M, Shenoy V, Bolton TA, Machado JM, Speth RC, Raizada MK, Katovich MJ. Prevention of angiotensin II-induced cardiac remodeling by angiotensin-(1-7). *Am J Physiol Heart Circ Physiol* 2007; **292**: H736-H742 [PMID: 17098828]
- 129 **Zheng H**, Pu SY, Fan XF, Li XS, Zhang Y, Yuan J, Zhang YF, Yang JL. Treatment with angiotensin-(1-9) alleviates the cardiomyopathy in streptozotocin-induced diabetic rats. *Biochem Pharmacol* 2015; **95**: 38-45 [PMID: 25801006]
- 130 **Flores-Munoz M**, Work LM, Douglas K, Denby L, Dominiczak AF, Graham D, Nicklin SA. Angiotensin-(1-9) attenuates cardiac fibrosis in the stroke-prone spontaneously hypertensive rat via the angiotensin type 2 receptor. *Hypertension* 2012; **59**: 300-307 [PMID: 22184331]
- 131 **Abadir PM**, Carey RM, Siragy HM. Angiotensin AT2 receptors directly stimulate renal nitric oxide in bradykinin B2-receptor-null mice. *Hypertension* 2003; **42**: 600-604 [PMID: 12953015]
- 132 **Savoia C**, Tabet F, Yao G, Schiffrin EL, Touyz RM. Negative regulation of RhoA/Rho kinase by angiotensin II type 2 receptor in vascular smooth muscle cells: role in angiotensin II-induced vasodilation in stroke-prone spontaneously hypertensive rats. *J Hypertens* 2005; **23**: 1037-1045 [PMID: 15834290]
- 133 **Chen Z**, Tan F, Erdős EG, Deddish PA. Hydrolysis of angiotensin peptides by human angiotensin I-converting enzyme and the resensitization of B2 kinin receptors. *Hypertension* 2005; **46**: 1368-1373 [PMID: 16246972]
- 134 **Henrion D**, Kubis N, Lévy BI. Physiological and pathophysiological functions of the AT(2) subtype receptor of angiotensin

- II: from large arteries to the microcirculation. *Hypertension* 2001; **38**: 1150-1157 [PMID: 11711513]
- 135 **Widdop RE**, Matrougui K, Levy BI, Henrion D. AT2 receptor-mediated relaxation is preserved after long-term AT1 receptor blockade. *Hypertension* 2002; **40**: 516-520 [PMID: 12364356]
- 136 **Bosnyak S**, Welungoda IK, Hallberg A, Alterman M, Widdop RE, Jones ES. Stimulation of angiotensin AT2 receptors by the non-peptide agonist, Compound 21, evokes vasodepressor effects in conscious spontaneously hypertensive rats. *Br J Pharmacol* 2010; **159**: 709-716 [PMID: 20128808]
- 137 **Kaschina E**, Grzesiak A, Li J, Foryst-Ludwig A, Timm M, Rompe F, Sommerfeld M, Kemnitz UR, Curato C, Namsolleck P, Tschöpe C, Hallberg A, Alterman M, Hucko T, Paetsch I, Dietrich T, Schnackenburg B, Graf K, Dahlöf B, Kintscher U, Unger T, Steckelings UM. Angiotensin II type 2 receptor stimulation: a novel option of therapeutic interference with the renin-angiotensin system in myocardial infarction? *Circulation* 2008; **118**: 2523-2532 [PMID: 19029468]
- 138 **Rompe F**, Unger T, Steckelings UM. The angiotensin AT2 receptor in inflammation. *Drug News Perspect* 2010; **23**: 104-111 [PMID: 20369075]
- 139 **Node K**, Huo Y, Ruan X, Yang B, Spiecker M, Ley K, Zeldin DC, Liao JK. Anti-inflammatory properties of cytochrome P450 epoxygenase-derived eicosanoids. *Science* 1999; **285**: 1276-1279 [PMID: 10455056]
- 140 **Walters PE**, Gaspari TA, Widdop RE. Angiotensin-(1-7) acts as a vasodepressor agent via angiotensin II type 2 receptors in conscious rats. *Hypertension* 2005; **45**: 960-966 [PMID: 15767466]
- 141 **Li J**, Jubair S, Janicki JS. Estrogen inhibits mast cell chymase release to prevent pressure overload-induced adverse cardiac remodeling. *Hypertension* 2015; **65**: 328-334 [PMID: 25403608 DOI: 10.1161/HYPERTENSIONAHA.114.04238]
- 142 **Jalil JE**, Ocaranza MP. Estrogens and myocardial chymase: new insights into pathological hypertrophy and remodeling. *Hypertension* 2015; **65**: 271-272 [PMID: 25403603 DOI: 10.1161/HYPERTENSIONAHA.114.04375]
- 143 **Zhao Z**, Wang H, Jessup JA, Lindsey SH, Chappell MC, Groban L. Role of estrogen in diastolic dysfunction. *Am J Physiol Heart Circ Physiol* 2014; **306**: H628-H640 [PMID: 24414072 DOI: 10.1152/ajpheart.00859.2013]
- 144 **Guan R**, Xu X, Chen M, Hu H, Ge H, Wen S, Zhou S, Pi R. Advances in the studies of roles of Rho/Rho-kinase in diseases and the development of its inhibitors. *Eur J Med Chem* 2013; **70**: 613-622 [PMID: 24211637 DOI: 10.1016/j.ejmech.2013.10.048]
- 145 **Surma M**, Wei L, Shi J. Rho kinase as a therapeutic target in cardiovascular disease. *Future Cardiol* 2011; **7**: 657-671 [PMID: 21929346 DOI: 10.2217/fca.11.51]
- 146 **Mendoza-Torres E**, Oyarzún A, Mondaca-Ruff D, Azocar A, Castro PF, Jalil JE, Chiong M, Lavandero S, Ocaranza MP. ACE2 and vasoactive peptides: novel players in cardiovascular/renal remodeling and hypertension. *Ther Adv Cardiovasc Dis* 2015; **9**: 217-237 [PMID: 26275770]

**P- Reviewer:** Kosmas Ioannis P, Masafumi T **S- Editor:** Qiu S  
**L- Editor:** A **E- Editor:** Wu HL





## RhoA signaling and blood pressure: The consequence of failing to “Tone it Down”

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**Author contributions:** Bai X, Dee R and Mangum KD contributed equally to this work; all authors contributed to literature review and analysis, drafting and critical revision and editing, and approval of the final version.

**Supported by** the National Heart, Lung, and Blood Institute, National Institutes of Health to Taylor JM, Nos. HL-081844 and HL-071054; and the Muscular Dystrophy Association to Taylor JM, No. MDA255577.

**Conflict-of-interest statement:** No conflicts of interest.

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**Received:** September 30, 2015  
**Peer-review started:** October 8, 2015  
**First decision:** November 6, 2015  
**Revised:** November 24, 2015  
**Accepted:** January 21, 2016  
**Article in press:** January 22, 2016  
**Published online:** February 23, 2016

### Abstract

Uncontrolled high blood pressure is a major risk factor

for heart attack, stroke, and kidney failure and contributes to an estimated 25% of deaths worldwide. Despite numerous treatment options, estimates project that reasonable blood pressure (BP) control is achieved in only about half of hypertensive patients. Improvements in the detection and management of hypertension will undoubtedly be accomplished through a better understanding of the complex etiology of this disease and a more comprehensive inventory of the genes and genetic variants that influence BP regulation. Recent studies (primarily in pre-clinical models) indicate that the small GTPase RhoA and its downstream target, Rho kinase, play an important role in regulating BP homeostasis. Herein, we summarize the underlying mechanisms and highlight signaling pathways and regulators that impart tight spatial-temporal control of RhoA activity. We also discuss known allelic variations in the RhoA pathway and consider how these polymorphisms may affect genetic risk for hypertension and its clinical manifestations. Finally, we summarize the current (albeit limited) clinical data on the efficacy of targeting the RhoA pathway in hypertensive patients.

**Key words:** Hypertension; Blood pressure; RhoA; Smooth muscle contraction; Guanine nucleotide exchange factor; GTPase activating protein; Polymorphisms

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**Core tip:** Studies (primarily in pre-clinical models) indicate that the small GTPase RhoA and its downstream target, Rho kinase, play an important role in regulating blood pressure homeostasis. Herein, we summarize the underlying mechanisms and highlight signaling pathways and regulators that impart tight spatial-temporal control of RhoA activity. We also discuss known allelic variations in the RhoA pathway and consider how these polymorphisms may affect genetic risk for hypertension and its clinical manifestations. Finally, we summarize the current (albeit limited) clinical data on the efficacy of

targeting the RhoA pathway in hypertensive patients.

Bai X, Dee R, Mangum KD, Mack CP, Taylor JM. RhoA signaling and blood pressure: The consequence of failing to “Tone it Down”. *World J Hypertens* 2016; 6(1): 18-35 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/18.htm> DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.18>

## INTRODUCTION

Although hypertension is a major risk factor for stroke, myocardial infarction, and kidney failure and contributes to over 350000 deaths annually in the United States<sup>[1]</sup>, we know surprisingly little about its development or the mechanisms by which it promotes cardiovascular disease. A number of antihypertensive drugs are available, but regimens are usually chosen empirically and multiple drugs that target different organ systems are frequently required for effective treatment. One reason for these difficulties is that blood pressure (BP) is a complex trait that is regulated by many organ systems and a large number of humoral factors. Thus, a better understanding of the molecular and genetic mechanisms that control BP under normal and pathologic conditions should lead to novel drug targets and/or to personalized therapies that are more effective and less toxic. Recent advances suggest that RhoA signaling plays a role in the development human hypertension. The focus of this review will be: (1) to highlight the mechanisms underlying RhoA-dependent regulation of BP; (2) to discuss how allelic variations in the RhoA signaling pathway affect genetic risk for hypertension and its clinical manifestations; and (3) to summarize the current (albeit limited) clinical data on the efficacy of targeting this pathway in hypertensive patients.

As a critical regulator of the actin cytoskeleton and acto-myosin contractility, the small GTPase, RhoA, regulates a variety of cellular processes including force development, endocytosis, exocytosis, adhesion, migration, proliferation, and differentiation<sup>[2]</sup>. Like all GTPases, RhoA is regulated by guanosine triphosphate (GTP) binding and cycles between the active GTP-bound form and the inactive GDP-bound form. When GTP-bound, RhoA interacts with a variety of effector molecules that mediate its effects on the actin cytoskeleton including the Rho-associated coiled-coil domain containing protein kinases (ROCK I and II), the diaphanous-related formins (mDia1 and mDia2), protein kinase N, citron kinase, rhotekin, and rhotekin. With respect to regulation of BP, Rho kinases are arguably the most important effectors as evidenced by the findings that increased ROCK activity has been observed in spontaneously hypertensive rats and some hypertensive patient populations<sup>[3,4]</sup> and ROCK inhibitors like Y-27632, Fasudil, and SAR407899 have been shown to reduce BP

in hypertensive animal models and patients<sup>[5]</sup>.

## RHOA SIGNALING AND BP REGULATION

BP homeostasis is tightly controlled by many organ systems and humoral factors that regulate peripheral vascular resistance, sodium and water balance, and cardiac output. Below we summarize the role RhoA signaling in the regulation of BP highlighting recent findings that implicate this pathway in the development of hypertension.

### *RhoA and arteriole tone*

Vascular resistance is a major determinant of BP and is controlled, in large part, by smooth muscle cell (SMC) contraction within small peripheral arterioles<sup>[6-10]</sup>. Excitation-contraction coupling in SMC is mediated by the  $\text{Ca}^{2+}$ -dependent activation of myosin light chain kinase (MLCK), and SMC tension is directly proportional to myosin light chain (MLC) phosphorylation as this enables myosins molecular interaction with actin<sup>[11,12]</sup>. Interestingly, besides promoting an increase in intracellular  $\text{Ca}^{2+}$ , many G protein-coupled receptor (GPCR)-coupled contractile agonists including angiotensin II (AngII), norepinephrine, and endothelin-1 (ET1) also stimulate RhoA activity in SMC and in intact arteries<sup>[3,4,13]</sup>. Active RhoA leads to Rho-kinase (ROCK)-dependent inhibition of myosin phosphatase and results in elevated MLCK activity and enhanced sensitization to  $\text{Ca}^{2+}$ <sup>[3,14-16]</sup>. Importantly, several studies in animal models and patients (described in further detail below) indicate that RhoA-dependent pathways are involved in the increased vascular resistance associated with hypertension<sup>[3-5,13,17]</sup>.

Active RhoA also induces *de novo* formation of actin filaments that are necessary for force development and SMC contraction. Rho-dependent actin remodeling occurs by both ROCK-dependent and independent processes. The Rho effectors mDia 1 and 2 directly catalyze actin polymerization in cooperation with the actin binding protein, profilin, whereas ROCK stimulates actin polymerization by inhibiting the disassembly of actin polymers through LIM-kinase-dependent inhibition of cofilin (ROCK activates LIM-kinase 1 and LIM-kinase 2 by phosphorylation at threonine 508 or 505 respectively within the activation loop<sup>[18-22]</sup>). ROCK also phosphorylates ERM proteins which enhances their tethering to integral plasma membrane proteins and promotes actin filament stabilization<sup>[23]</sup>.

Recent studies indicate that RhoA signaling also controls SMC contractile gene expression by regulating the nuclear translocation of the Myocardin-related transcription factors (MRTF-A and MRTF-B). Under conditions of low RhoA activity, monomeric (G)-actin binding to the MRTF N-terminus masks a nuclear localization sequence resulting in cytoplasmic sequestration of these serum response factor co-factors. The fall in cytoplasmic G-actin levels that occurs upon RhoA-mediated actin polymerization promotes MRTF

nuclear accumulation and promotes the expression of SM  $\alpha$ -actin, SM  $\gamma$ -actin, SM myosin heavy chain, calponin, and SM22<sup>[24,25]</sup>. Thus, not only does RhoA control SMC contractility, but it also regulates the levels of the SMC-specific contractile proteins that support this function. Moreover, elevated RhoA in endothelial cells impairs endothelial cell-mediated vasorelaxation as it decreases availability of the potent vasodilator, nitric oxide by reducing both eNOS expression and activity<sup>[26-30]</sup>. In sum, signaling through RhoA enhances Ca<sup>2+</sup> sensitivity, promotes actin remodeling and induces expression of contractile proteins and these responses are necessary for maintaining sustained SMC contractility and elevated vessel tone (Figure 1).

### **RhoA and kidney function**

The kidneys play a major role in regulating BP by controlling sodium excretion and blood volume. In addition, since the kidneys are highly perfused organs receiving up to 25% of total cardiac output, increased contractility of renal arterioles can significantly increase total peripheral vascular resistance. In most vascular beds, arteriolar tone is controlled by autonomic innervation and circulating hormones. However, in pre-glomerular afferent arterioles, increased kidney perfusion (manifesting as increased renal BP) stimulates SMC contraction through the tubuloglomerular feedback and myogenic responses (see<sup>[31]</sup> for review). The former mechanism is mediated by increased glomerular filtration and NaCl delivery from the loop of Henle to the macula densa (MD), a cluster of epithelial cells located at the junction between the distal convoluted tubule and the end of the thick ascending limb and adjacent to the abluminal SMCs of the afferent arterioles. Increased NaCl uptake by MD cells results in secretion of ATP and adenosine which stimulate afferent arteriole SMC contraction *via* P2Y4/P2Y6 and A2 GPCRs, respectively. The myogenic response is mediated by the activation of stretch-sensitive cation channels. Together these mechanisms stabilize renal blood flow to protect the sensitive glomerular capillaries from flow-induced trauma. Importantly, afferent arterioles express RhoA, ROCK I and II<sup>[32]</sup>, and several studies have convincingly demonstrated that the Rho/Rho kinase pathway influences both of these feedback mechanisms in response to increased kidney perfusion<sup>[33-37]</sup>. The requirement of RhoA is likely due, at least in part, to its necessity for P2Y4/P2Y6 and A2 receptor-dependent contractility. Indeed, ATP (*via* P2Y4/Y6) and adenosine (*via* A2) stimulate RhoA activity in SMC and their pressor responses were prevented by pretreatment with the Rho-kinase inhibitor, Y-27632<sup>[32]</sup>.

Interestingly, recent evidence indicates that RhoA may play an additional role in other cell types within the kidney to impact volume homeostasis. In particular, RhoA activity in tubular epithelial cells can regulate sodium reabsorption and excretion primarily by altering the density and location of epithelial sodium channels (ENaCs) and the sodium-hydrogen exchanger

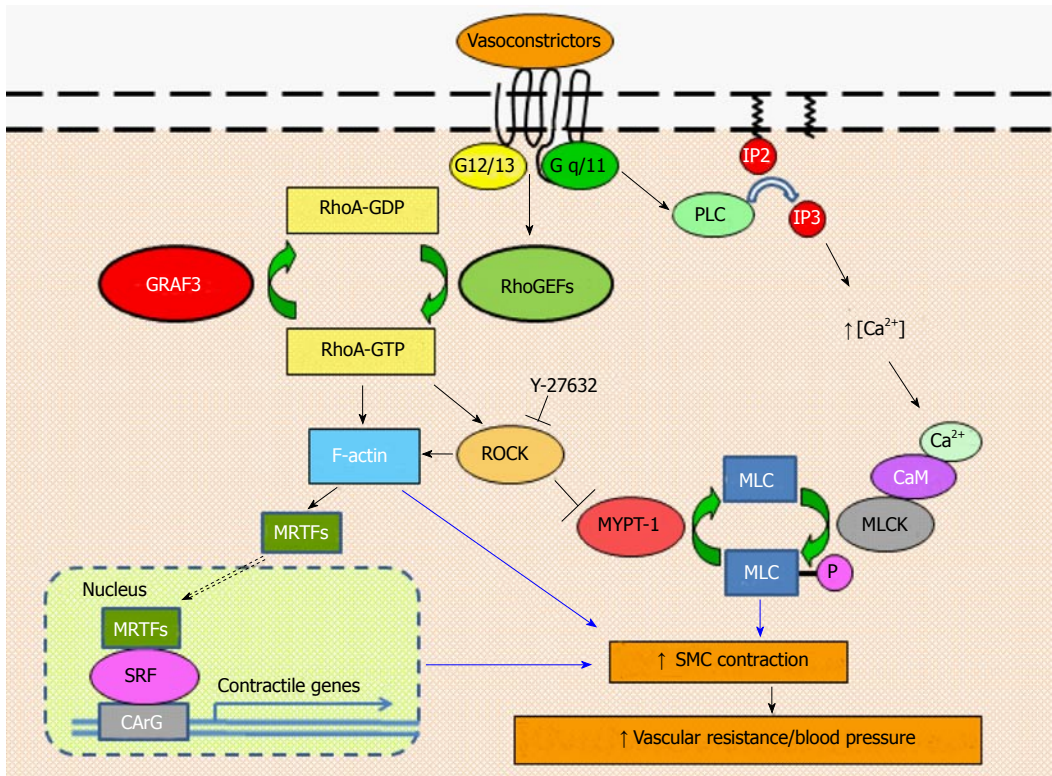
(NHE3)<sup>[38]</sup>. *In vitro* studies in cultured epithelial cells indicated that the Na<sup>+</sup> current through ENaCs was significantly increased by expression of wildtype or constitutively active RhoA (G14V) and suppressed by expression of dominant negative RhoA (T19N). The changes in current correlated with alterations in the density of ENaCs at the PM<sup>[39]</sup> and mechanistic studies determined that RhoA signaling was essential for intracellular vesicle mediated transport of ENaCs to the apical cell surface<sup>[40,41]</sup>. RhoA signaling also regulates the activity and subcellular localization of NHE3, a key regulator of sodium absorption in the proximal convoluted tubule. NHE3 associates with ezrin and cortical actin filaments at the plasma membrane and treatment with either the RhoA inhibitor, diphtheria toxin toxin B, or Y-27632 disrupted these interactions and promoted the internalization of NHE3 to sub-membrane compartments<sup>[42,43]</sup>. Moreover, Nishiki *et al.*<sup>[44]</sup> showed that spontaneously hypertensive rats exhibited elevated NHE3 activity and a exaggerated level of Na<sup>+</sup> reabsorption when compared to normotensive controls and that Na<sup>+</sup> reabsorption was normalized by treatment of the hypertensive animals with Y27632.

### **RhoA in the central and peripheral nervous system**

The central nervous system (CNS) constantly assesses pressure levels in the vasculature and makes necessary signaling adjustments to prevent BP variability. The main mechanism by which the CNS monitors BP is through a rapid negative feedback loop termed the baroreceptor reflex. Baroreceptors are sensory neurons located primarily in the aortic arch and carotid sinuses that continuously respond to pressure-induced stretching of the vessels in which they reside. Impulses from baroreceptors are relayed *via* glossopharyngeal and vagus nerves to the nucleus tractus solitarius (NTS) in the brainstem<sup>[45]</sup>, which in turn relays the signal to the rostral ventrolateral medulla<sup>[46]</sup> and increases or decreases parasympathetic and sympathetic stimulation to the heart and vessels accordingly. Interestingly, the CNS component of this feedback loop has been shown to be dependent on RhoA/Rho-kinase signaling. Rho-kinase inhibitors microinjected directly into the NTS or infection of this structure with an adenovirus expressing a dominant-inhibitory form of Rho-kinase reduces sympathetic nerve activity, heart rate, and BP in normotensive rats and these effects are even more pronounced in spontaneously hypertensive rats<sup>[47,48]</sup>. Moreover, infusing the ROCK inhibitor, Y27632, into the neural cistern attenuated the BP increase that resulted from AII infusion into the same area of the brainstem<sup>[49]</sup>.

While RhoA's effects on the CNS are clear, a heretofore understudied area in this field is the extent to which RhoA regulates neurotransmitter release from the perivascular nerves which are known to play a major role in the control of resistance arteriole tone. While it has long been known that RhoGTPases have an important and conserved function in mediating neuronal





**Figure 1** Schematic summarizing RhoA-dependent regulation of vascular smooth muscle contraction and blood pressure homeostasis. Excitation-contraction coupling in smooth muscle cell (SMC) is mediated by the  $\text{Ca}^{2+}$ -dependent activation of myosin light chain kinase (MLCK), and SMC tension is directly proportional to myosin light chain (MLC) phosphorylation (p) as this enables myosin's molecular interaction with actin. SMC contractility is also regulated by GPCR-coupled contractile agonist-mediated activation of the small GTPase RhoA. Downstream activation of Rho kinase (ROCK) inhibits myosin phosphatase target subunit 1 (MYPT-1), and results in increased levels of pMLC to promote smooth muscle contraction. RhoA also stimulates G-actin polymerization to filamentous actin (F-actin). Actin polymerization increases SMC tension and stimulates myocardin-related transcription factor (MRTFs) nuclear translocation which promotes SRF-dependent transcription of contractile genes. RhoGAPs (such as GRAF3) and RhoGEFs dynamically regulate RhoA activity to achieve blood pressure balance.

survival and death and that tight spatiotemporal control of RhoA is necessary for appropriate neuronal development (neurite outgrowth, growth cone dynamics) and regeneration, to our knowledge no studies have explored the consequence of Rho-kinase inhibition on peripheral nerve structure or function. Future studies to this end are warranted, because some studies in cells and invertebrate model systems indicate that Rho/Rho kinase signaling may limit the release of sympathetic (contractile) agents and promote the release of parasympathetic relaxation factors from motor neurons. For example, Yamaguchi *et al.*<sup>[50]</sup> found that  $\text{Ga}_{12/13}$ -mediated activation of RhoA/ROCK inhibited  $\text{Ca}^{2+}$  dependent exocytosis of the contractile neurotransmitter dopamine in PC12 cells. In support of these studies, an activating mutation in ArhGEF10, a RhoGEF highly expressed in the peripheral nervous system, was identified in patients who exhibited slowed nerve conduction velocities<sup>[51,52]</sup>. On the other hand, Hiley *et al.*<sup>[53]</sup> reported that release of the relaxation neurotransmitter, acetylcholine from cholinergic motor neurons in *C. elegans*, required the regulators of G protein signaling (RGS)-RhoGEF dependent activation of Rho A. Thus, it is formally possible that inhibition of RhoA in peripheral nerves could lead to an increase total peripheral resistance and BP. This concept requires

further exploration if RhoA/ROCK inhibitors are to be considered as future anti-hypertensive therapies.

### RhoA in the myocardium

Several studies have shown that RhoA signaling has direct effects on cardiac function that increase cardiac output and BP. Transgenic mice that overexpressed either  $\text{GDI}\alpha$  or dominant negative RhoA exhibited conduction defects and cardiomyocytes isolated from these mice exhibited decreased L-type  $\text{Ca}^{2+}$  channel currents that likely contributed to the decreased contractility observed *in vivo*<sup>[54,55]</sup>. Vlasblom *et al.*<sup>[56]</sup> showed that treatment of neonatal ventricular cardiomyocytes with Y27632 reduced the expression and activity of the sarcoplasmic reticulum  $\text{Ca}^{2+}$  ATPase, SERCA2a, thereby limiting the amount available for  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release in the next cardiac cycle. In addition, RhoA-dependent pathways have been shown to be critical for phosphorylation and sensitization of cardiac troponin T complex to intracellular  $\text{Ca}^{2+}$  levels<sup>[57]</sup>. Moreover, while not initially thought to be a major mechanism for modulating cardiac contractility, it is becoming clear that cardiac MLC phosphorylation can enhance muscle contractility by increasing  $\text{Ca}^{2+}$  sensitivity<sup>[58]</sup> and that MLC phosphatase is a target for Rho kinase-dependent inhibition in the myocardium (like



in SMC). Indeed, Lauriol *et al.*<sup>[59]</sup> showed that cardiac-restricted deletion of RhoA led to decreased contractility and this effect was correlated with decreased MLC activity. Other similarities between RhoA signaling in cardiomyocytes and SMC include the ability of RhoA-mediated signals to promote differentiation/maturation by promoting the expression of contractile genes<sup>[60]</sup>.

## CONTROL OF RHOA GTPASE ACTIVITY

Rho proteins act as molecular switches that cycle between an inactive GDP-bound form and an active GTP-bound form and this cycle is under the direct control of three groups of regulatory proteins. Guanine dissociation inhibitors (GDIs) sequester RhoA into an inactive cytoplasmic fraction, guanine nucleotide exchange factors (GEFs) activate RhoA by facilitating exchange of GDP for GTP, and GTPase activating proteins (GAPs) promote RhoA's intrinsic GTPase activity to hydrolyze GTP to GDP and efficiently turn off (or tone down) RhoA-dependent signaling. The GEF and GAP protein families are quite large and structurally diverse and it is likely that additional differences in expression patterns and post translational modification allow for tissue-specific and tight spatio-temporal control of RhoA activity. The following section summarizes the known mechanisms for controlling RhoA activity in SMC.

### RhoGEFs and BP control

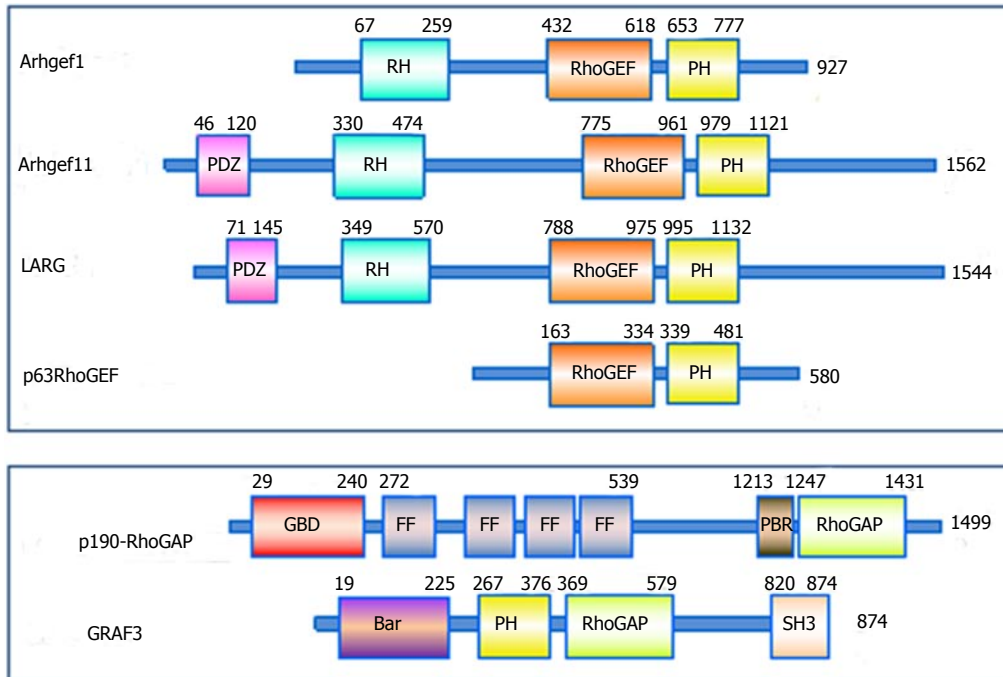
GEFs activate small GTPases by increasing the GDP dissociation rate by several orders of magnitude which in turn promotes GTP-binding since GTP is in an approximately 10:1 molar excess to GDP in mammalian cells<sup>[61]</sup>. To date, over 24 different Rho-selective GEFs have been identified. The common functional domain of RhoGEFs is the Dbl homology (DH) domain (also referred to as the RhoGEF domain), which typically serves as both the catalytic site and the major binding interface for RhoA (Figure 2). A pleckstrin homology (PH) domain is almost always found downstream of the DH domain and this unit serves to facilitate membrane binding and cooperates with DH domains to fully activate RhoA<sup>[62]</sup>. Other common functional domains include the RGS domain that binds large G-proteins to couple the GPCR and Rho signaling pathways and the Postsynaptic Density 95, disk large, Zona occludens-1 (PDZ) domain that binds to Plexin-B1 and Lysophosphatidic acid (LPA) receptor to transmit Semaphorin 4D (57) and LPA signals<sup>[63]</sup>, respectively.

The major contractile agonists that stimulate RhoA activity in SMC include AII, phenylephrine (PE), ET1, and thromboxane A<sub>2</sub>. The GPCRs for these ligands couple to various G $\alpha$  subunits, including G $\alpha_{12/13}$  and G $\alpha_{q/11}$ , but each has distinct Gprotein coupling properties. For example, PE signals almost exclusively through G $\alpha_q$  and thromboxane through G $\alpha_{12/13}$  while other agonist-receptor interactions lead to more promiscuous G protein activation. These heterotrimeric GTPases in turn either

directly or indirectly activate a range of RhoGEFs. Therefore, it is likely that each agonist could stimulate somewhat overlapping but distinct set of RhoGEFs to enable fine tuning of the extent and duration of Ca<sup>2+</sup> sensitization and thus engender precise spatial and temporal control of vessel tone.

RGS family of RhoGEFs (LARG, p115RhoGEF, and PDZRhoGEF)<sup>[64]</sup> has received a lot of attention in the BP field because these proteins can interact with (and be directly activated by) G $\alpha_{12}$  and G $\alpha_{13}$ <sup>[65]</sup>. Indeed, activation of RhoA by many of the aforementioned contractile agonists is mediated, at least in part, by these RhoGEFs. However, studies performed in mice lacking the  $\alpha$  subunits of G $\alpha_{q/11}$  or G $\alpha_{12/13}$  in smooth muscle convincingly showed that activation of these RhoGEFs are not simply due to direct binding to G $\alpha_{12/13}$ <sup>[13]</sup>. In fact G $\alpha_{12/13}$  depletion did not affect the pressor effects of AII, or PE, and only modestly reduced the pressor effects of ET1. Instead, depletion of G $\alpha_{q/11}$  completely abrogated PE-induced pressor responses and dramatically attenuated responses to AII. Subsequent studies from Guilluy *et al.*<sup>[13]</sup> identified p115RhoGEF (p115) as the critical GEF that mediates AII-dependent RhoA activity in SMC and small arterioles and showed that smooth muscle specific deletion of p115 rendered mice resistant to AII-dependent hypertension. Interestingly, their mechanistic studies confirmed that p115 activation did not require G $\alpha_{12}$ /G $\alpha_{13}$ , but instead, was governed by G $\alpha_q$ -mediated, Janus tyrosine kinase-dependent phosphorylation of Tyr738 in the PH domain. Importantly, phosphorylation mimetic and deficient variants at Tyr738 elevated and reduced p115's GEF activity, respectively<sup>[13]</sup>. As discussed in further detail below, phosphorylation-dependent activation of RhoGEFs has since emerged as a critical regulatory pathway. However, it should also be noted that AII-dependent activation of RhoA in SMC likely involves additional pathways as AII signaling in SMC has been linked to inhibition of p190Rho GTPase activating protein (see below<sup>[66]</sup>); upregulation of LARG<sup>[67]</sup>, upregulation of PDZ-RhoGEF<sup>[68]</sup>, and G $\alpha_q$ /Ca<sup>2+</sup>/proline-rich tyrosine kinase 2 (PYK2) tyrosine kinase mediated phosphorylation/activation of PDZ-RhoGEF<sup>[69]</sup>. Indeed, Ying *et al.*<sup>[69]</sup> showed that Ca<sup>2+</sup>/PYK2-dependent activation of PDZ-RhoGEF was necessary for maximal AII induced RhoA activation.

Interestingly, p115 mutant mice also exhibited a partial reduction in DOCA/salt-induced hypertension but had normal basal BP and normal pressor responses to ET1 and PE; agents that also act through G $\alpha_q$ -dependent signaling pathways. Future studies are necessary to determine how PE and ET1/G $\alpha_q$  signals differ from those induced by AII. However, recent studies by the Somlyo laboratory shed some light on this phenomena as they found that there is functional overlap between p115 and LARG. Indeed using genetic mouse models, they found that while the time it took to reach maximal contraction was increased in SMC-



**Figure 2 Multi-domain architecture of RhoGEF and RhoGAP proteins known to regulate smooth muscle cell phenotype.** The catalytic domain of RhoGEFs is termed a Dbl homology (DH) domain, which serves as the major binding interface with Rho GTPases and catalyzes the dissociation of GDP from the GTPase. Pleckstrin homology (PH) domains are almost always downstream of the DH domain and these units cooperate to fully activate the GTPase. Other functional domains contained in specific RhoGEFs include the RH (Regulators of G protein Signaling Homology) domain and PDZ (Postsynaptic density 95, disk large, zona occludens-1) domain. RhoGAPs are also multi-domain containing proteins. The RhoGAP domain facilitates GTP hydrolysis and inhibits RhoA activity while other domains can regulate RhoGAP targeting and function. For example, BAR (Bin/amphiphysin/Rvs), PH, or polybasic region (PBR) domains direct lipid binding and promote membrane localization. Other domains are involved in protein-protein interactions such as GTP-binding domain (GBD), diphenylalanine motifs (FF) and the SH3 (SRC Homology 3) domains. The amino acid numbers are shown above each protein are based on the human orthologs (<http://www.ncbi.nlm.nih.gov/>).

specific double knockouts, maximal contraction of smooth muscle from PDZ-RhoGEF and LARG double knockdown tissues was similar to that of the single mutants<sup>[70]</sup>. Thus it is formally possible that AII (but not ET1 and PE) induced hypertension is blocked by p115 knockout because AII has a relatively reduced capacity to activate LARG. Interestingly, Medlin *et al.*<sup>[71]</sup> identified the vasoconstrictor agonist sphingosine-1-phosphate (S1P) as a potent activator of LARG in cultured vascular SMC. While this pathway has not yet been confirmed *in vivo* the finding that LARG knockout mice are resistant to salt-induced hypertension<sup>[4]</sup> which leads to volume overload-induced stretching of vessels is consistent with the thesis that LARG may regulate RhoA activity and SMC contractility in response to mechanical forces<sup>[72]</sup>.

Besides activation of the RGS GEFs, several studies have linked  $G_{\alpha q/11}$ -dependent activation of RhoA to the Trio family of RhoGEFs (Trio, Duet, and p63-RhoGEF)<sup>[73,74]</sup>. P63-RhoGEF is highly expressed in arterial smooth muscle and it has recently been shown to be important for the early phase of AII-dependent vessel contractility<sup>[75]</sup> and for maximal pressor response to other vasoconstrictors such as PE and ET1 that act through  $G_{\alpha q/11}$ <sup>[76]</sup>. Moreover, another non-RGS RhoGEF termed lymphoid blast crisis (Lbc) has been shown to be critical for serotonin-dependent activation of RhoA and contractility in vascular SMC<sup>[77]</sup>. In summary, specific vasoconstrictors can lead to activation of distinct

but overlapping sets of RhoGEFs (each with different activation kinetics, catalytic activities, and subcellular locales). A question that warrants further studies is to what extent GEFs might also govern control over the activation of specific RhoA effector subsets by various agonists.

Like p115, PDZRhoGEF and LARG are also activated by tyrosine phosphorylation. Focal adhesion kinase (FAK) as well as its related family member, PYK2 phosphorylate PDZRhoGEF<sup>[69,78]</sup>, while LARG was shown to be activated by FAK<sup>[78]</sup> and Tec<sup>[64,69]</sup>. Future identification of the specific sites of phosphorylation will aid in determining if there is a conserved mechanism by which these modifications promote GEF activity.

Control of RhoGEF expression is another important means of regulating RhoA activity in the vasculature. Because p115-RhoGEF, PDZ-RhoGEF, and LARG each play a role in BP regulation in rodents, it is no surprise that these GEFs are expressed in both conductance and resistance arteries of rats and mice<sup>[4,68,69,79]</sup>. P63RhoGEF is also abundant in the peripheral vasculature<sup>[80]</sup>. Interestingly, the expression of many of these RhoGEFs fluctuates as BP changes, suggesting that dynamic regulation of their expression is important for BP control. The most comprehensive study performed to date revealed that expression of each of the five RhoGEFs linked to Rho-A dependent vasoconstriction (p115, LARG, PDZ-RHOGEF, p63 RhoGEF, and Lbc)

are all down-regulated in cultured mesenteric artery SMC following treatment with AII for 48 h. Moreover, treatment with the Rho Kinase inhibitor fasudil prevented the AII-mediated suppression of p115, LARG and PDZ-RhoGEF indicating that RGSrhoGEF expression is governed, at least in part, by negative-feedback signaling through the Rho/Rho kinase cascade. A similar decrease in RGSrhoGEF expression was observed in mesenteric arteries from rats treated with AII for 14 d<sup>[80]</sup>. Whether RhoGEF expression is altered in or contributes to hypertension in animal models is less clear. Ying *et al.*<sup>[81]</sup> reported that aortic expression of all 3 RGSrhoGEFs was higher in aortas from 12 wk old SHR than in normotensive rats. Similarly, a comprehensive microarray analysis revealed that LARG expression was upregulated in DOCA-salt hypertensive mice<sup>[82]</sup>. In contrast, Hilgers *et al.*<sup>[68]</sup> reported that mesenteric arteries from 14 d AII-treated rats exhibited decreased mRNA levels, but increased protein levels of PDZ RhoGEF. Thus, while it is clear that GEF expression is dynamic, the extent to which elevated expression of these factors contributes to the induction of hypertension and reduced expression to BP normalization is currently unresolved and requires further study.

### RhoGAPs and BP control

GAPs inhibit Rho signaling by enhancing the intrinsic ability of Rho to hydrolyze GTP<sup>[83,84]</sup>. More than 70 RhoGAPs have been identified in eukaryotes that can be divided into 23 subfamilies<sup>[85]</sup>. Like the RhoGEFs, RhoGAPs are typically large multi-domain containing proteins and their diverse structures allow for dynamic and selective inhibition of small GTPase signaling (see Figure 2). Several Rho-selective GAPs including p190RhoGAP, ArhGAP1, Myr5, GRAF1, and GRAF3 have been shown to regulate RhoA in cultured vascular SMC<sup>[86,87]</sup>.

To our knowledge, GRAF3 is the only RhoGAP that has been implicated in the regulation blood pressure. The founding member of the GRAF (GTPase regulator associated with FAK-1) family, was originally identified by our group<sup>[88-90]</sup> by screening an embryonic  $\lambda$ gt11 expression library for proteins that interacted with the carboxyl-terminal domain of FAK<sup>[88]</sup>. The GRAF family's three members are defined by an N-terminal BAR (Bin/amphiphysin/Rvs) domain, a phosphatidylserine (PS)-binding PH domain, a central Rho-GAP domain, a serine/proline rich domain, and a C-terminal SH3 domain (Figure 1A). The GRAF1 SH3 domain was shown to specifically bind to a proline-rich region in the carboxy terminus of FAK and this protein-protein interaction was important for directing GRAF1 to the actin cytoskeleton<sup>[88]</sup>. GRAF1 is expressed predominantly in the brain and striated muscle (cardiac and skeletal), and our studies in GRAF1-depleted *Xenopus* and mice revealed that GRAF1-dependent inhibition of RhoA activity promoted mammalian muscle growth

by facilitating myoblast fusion and injury repair<sup>[90-93]</sup>. GRAF2 is more ubiquitously expressed<sup>[94]</sup> and could partially compensate for the loss of GRAF1 during myotube formation supporting at least some functional redundancy within this family<sup>[92]</sup>. Evolutionarily, GRAF3 is the youngest family member and is the most recently annotated. Interestingly, our genome wide analyses of chromatin structure in primary human SMC suggested that this gene was regulated in a smooth muscle-specific fashion. Indeed, we found that GRAF3 was highly and selectively expressed in SMC with particularly high expression in resistance vessels<sup>[87]</sup>. After validating that GRAF3 functioned as a RhoA-specific GAP in these cells, we considered the possibility that GRAF3 might control BP homeostasis. Importantly, we found that homozygous GRAF3 knockdown mice showed a consistent and significant elevation in systolic, diastolic, and mean arterial BP (+ 20-30 mmHg). The observation that heterozygous GRAF3 knockdown mice exhibited a 15 mmHg increase in BP strongly supports a dose-dependent relationship between GRAF3 expression and BP. GRAF3-deficient mice exhibited significantly elevated pressor responses following treatment with AII, ET1, or PE and these effects were inhibited by treatment of GRAF3 deficient mice with Y-27632. Accordingly, RhoA activity and myosin light chain phosphorylation were elevated in GRAF3-depleted SMC *in vitro* and *in vivo*<sup>[87]</sup>.

The remarkable SMC-selective expression pattern of this Rho-selective GAP when coupled with the ability of ROCK inhibition to normalize the hypertensive phenotype of GRAF3-deficient animals strongly supports a model in which GRAF3 plays a major role in regulating BP homeostasis by limiting RhoA-mediated SMC contractility in resistance vessels<sup>[87]</sup>. Interestingly, as discussed in further detail below, a large GWAS revealed that polymorphisms in the GRAF3 gene contribute to BP variation in humans<sup>[95,96]</sup>. Thus future studies that strive to determine the mechanisms that control variations in GRAF3 expression and/or activity will likely lead to important insights into how to better control BP in the general population.

p190RhoGAP may also play a role in limiting RhoA-dependent arterial tone. p190RhoGAP contains an amino-terminal GTP-binding domain, a large middle domain with multiple protein-protein interaction motifs (diphenylalanine, FF motifs) a polybasic region, and a carboxy-terminal GAP domain<sup>[97]</sup>. Knockdown of p190RhoGAP in SMC by siRNA increased RhoA/Rock activity<sup>[98]</sup>, and several studies have shown that p190RhoGAP is activated by phosphorylation of Y1105 by cAbl and Src tyrosine kinases<sup>[66,99]</sup>. p190RhoGAP is a substrate for the tyrosine phosphatase, SHP-2, and SHP-2-dependent dephosphorylation of p190RhoGAP was shown to be important for the initial burst in RhoA activity in SMCs treated AII and ET1<sup>[98]</sup>. Interestingly, ROCK-dependent phosphorylation at Ser1150 attenuated p190RhoGAP activity creating a positive feedback loop for further RhoA activation<sup>[86]</sup>. Pho-

sphorylation of several C-terminal residues by ERK also suppresses p190RhoGAP activity during focal adhesion formation<sup>[100]</sup>. Finally, although not yet shown in SMC, p190RhoGAP has also been shown to be regulated by phospholipid binding<sup>[101]</sup>. Additional studies will be necessary to determine if p190RhoGAP plays an important role in BP regulation *in vivo*.

### Regulation of GDIs

GDIs bind to GDP-bound GTPases and inhibit GDP dissociation. GDI binding also limits translocation of GTPases to the membrane effectively "locking" them in the inactive state. Indeed, studies have shown that GDIs can inhibit RhoA dependent  $\text{Ca}^{2+}$  sensitization in SMCs treated with  $\alpha$ -adrenergic and muscarinic agonists<sup>[102]</sup>. However, the extent to which RhoGDIs regulate BP or RhoA activity *in vivo* is unclear. One study showed that RhoGDI $\alpha$  knock out mice displayed a salt-dependent increase in BP, but this effect was attributed to an increase in Rac1 activity in the kidney<sup>[103]</sup>. However, since SMC-specific Rac1 knockout mice were hypertensive and exhibited increased RhoA activity, it will be important to measure RhoA and Rac1 activity in SMC in RhoGDI $\alpha$  knock out mice<sup>[104]</sup>. RhoGDIs have been shown to bind to and regulate RhoGEFs and RhoGAPs<sup>[105]</sup>, an effect that could indirectly influence RhoA activity and vessel tone. Thus, additional studies will be needed to assess RhoGDIs' role in RhoA dependent blood pressure regulation.

### Direct regulation of RhoA

Additional control of RhoA signaling may be imparted by mechanisms that alter RhoA protein levels and/or alter functional post-translational modifications. Notably, protein ubiquitination followed by proteasome-dependent degradation is a major means of fine-tuning protein levels and Chen *et al.*<sup>[106]</sup> reported that RhoA is a direct target of the Rho-BTB/Cullin-3 E3 ubiquitin ligase degradation pathway. Interestingly, the Sigmund laboratory found that Cullin-3 regulated vascular smooth muscle function and arterial BP through a RhoA/Rho-kinase pathway. Moreover, they found that a human hypertension-associated mutation in Cullin-3 in which exon 9 is deleted led to decreased Cullin-3 activity and reduced ubiquitin-mediated RhoA degradation<sup>[107,108]</sup> (see below for further discussion of these and other genetic variants that influence RhoA signaling and human hypertension). Ubiquitination-dependent regulation of RhoA is also catalyzed by a distinct E3 ubiquitin ligase termed SMAD ubiquitin regulatory factor (Smurf1)<sup>[109,110]</sup>. Interestingly, Smurf1-dependent degradation of RhoA in endothelial cells has been linked to the development of cerebral cavernous malformation (CCM) a disease that is accompanied by hyperpermeable blood vessels in the brain. CCM results from the homozygous inactivating mutations in one of three *ccm* genes. Crose *et al.*<sup>[109]</sup> demonstrated that *ccm2* bound directly to Smurf1 and that this interaction

regulated RhoA degradation, likely explaining the common biochemical defect of elevated RhoA/ROCK signaling and increased permeability observed in *ccm* mutant endothelial cells<sup>[109,110]</sup>. Whether this class of E3 ligases regulates RhoA in levels in SMC is currently unknown.

Signaling *via* nitric oxide and reactive oxygen species may add another level of spatial/temporal control of RhoA signaling in the vasculature. Levels of ROS increase in the vasculature under a number of pathological conditions including hypertension, and ROS-mediated activation of RhoA has been demonstrated in vascular smooth muscle<sup>[81]</sup>. Interestingly, Aghajanian *et al.*<sup>[111]</sup> have demonstrated that ROS can mediate direct activation of RhoA by reversible oxidation of reactive cysteines C16/C19 and that this acts in a similar fashion to GEFs in that it leads to nucleotide displacement and increased GTP binding. Because oxidation of RhoA does not impair RhoGEF binding it is possible that ROS-dependent oxidation might prime RhoA for vasoconstrictor-dependent activation. In contrast, the reactive vasodilator, NO has been shown to inhibit RhoA activation *via* post-translational modification. For example, treatment of SMC with the pharmacological NO donor, PAPA-NONOate promoted RhoA S-nitrosylation that reduced GTP binding and therefore inactivated RhoA<sup>[112]</sup>. NO signaling may also limit RhoA activity in SMC by promoting cGMP-dependent phosphorylation of RhoA on sites that attenuate membrane targeting (and activation) of RhoA<sup>[113,114]</sup>. However, the role that such post-translational modifications play *in vivo* has yet to be explored.

In summary, RhoA activity in SMC can be dynamically regulated by transcriptional and post-translational mechanisms that alter RhoA protein, its activators, and its inhibitors. Collectively these mechanisms play an important role in precise spatial-temporal control of vessel tone and BP homeostasis. Importantly, while several RhoAGEFs have been shown to be necessary for development of vasoconstrictor-induced hypertension, our recent results in GRAF3-depleted mice demonstrated for the first time that GAP-dependent control of RhoA activity in SMC contributes to the maintenance of basal BP<sup>[87]</sup>.

## GENETIC REGULATION OF THE RHOA PATHWAY IN HUMAN HYPERTENSION

Hypertension is a devastating disease associated with significant morbidity and mortality due to detrimental pressure-related effects on the kidneys, heart, lungs, brain, and peripheral vasculature. Hypertension affects roughly 80 million people (approximately 32.6% of adults) in the United States alone and was predicted to be primarily responsible for 25% of deaths worldwide in 2010<sup>[115]</sup>. Despite the fact that nearly 70 drugs (from 15 distinct classes of compounds) are approved



for treatment of hypertension in the United States, estimates project that reasonable BP control is achieved in only about half of hypertensive patients. This reality coupled with recent projections that the incidence of hypertension will increase to about 41% in the United States by 2030, indicate the urgent need for better screening and treatment modalities<sup>[116]</sup>. Improvements in the detection and management of hypertension will undoubtedly be accomplished through a better understanding of the complex etiology of this disease.

One way to better predict patient response to therapy is to gain a more comprehensive understanding of the genes and genetic variants that influence BP regulation. Recent projections indicate that up to 60% of BP variation can be explained by genetic factors, but that no single gene exerts a principal effect. Thus, BP is considered to have a complex non-Mendelian mode of inheritance. Indeed a combination of classic positional cloning strategies in families with numerous affected members combined with more recent population-based GWAS studies have led to the identification of 25 rare mutations and 53 SNPs that are predicted to contribute to BP control<sup>[117]</sup>. The aim of this section of is to highlight variants that impinge on the expression or activity of members of the RhoA signaling axis.

### ***RhoA-related forms of monogenic hypertension***

Virtually all known cases of monogenic hypertension are associated with volume expansion resulting from mutations in genes involved in renal salt handling or hormones that affect mineralocorticoid activity. However, although hypertensive patients with Gordon's Syndrome (pseudohypoaldosteronism type IIE) present with salt handling abnormalities, the high BP in these patients is caused by an autosomal dominant mutation in the Cullin-3 gene (see above). Interestingly, this E3 ligase helps target RhoA for proteosomal degradation and *in vitro* studies indicate that increased RhoA/ROCK signaling in vascular SMC may also play a role in Gordon's Syndrome patients<sup>[118]</sup>. Exclusion of exon 9 abrogates the Cullin-3 dependent interactions between RhoBTB and the E3 ligase and as RhoBTB serves as a chaperone to recruit RhoA to this degradation complex, expression of exon 9-deficient Cullin-3 leads to aberrant RhoA accumulation<sup>[107,108]</sup>.

### ***SNP/EQTls in RhoA-signaling molecules***

Because Rho kinases are major RhoA effector proteins and because both animal and human studies have shown that treatment with Rho-kinase inhibiting compounds lowers BP, a number of case-controlled studies were designed to determine if genetic variants in these genes might influence the development of human hypertension. One group examined the effect of ROCK2 genetic variations on BP in 168 pairs of mono- and dizygotic twins. In this study, four variants were identified in ROCK2, the most notable of which was a nonsynonymous SNP in exon 10 that resulted

in a substitution of Thr with Asn at amino acid 431. Importantly, the Asn substitution was associated with increased systemic vascular resistance and BP and was predicted to account for 3%-5% of the BP variance between these patients<sup>[119]</sup>. Another study in which 18 tag SNPs within the ROCK2 locus were genotyped in 586 normotensive controls and 607 hypertensive Caucasian patients identified a haplotype defined by four SNPs (rs965665, rs10178332, rs6755196, rs10929732) that was recessively associated with a lower risk of hypertension ( $P = 0.003$ ). However, a subsequent study in a separate population of 1344 Chinese patients with coronary artery disease and hypertension and 1267 ethnically and geographically matched controls did not find an association between this haplotype and either BP or cardiovascular disease<sup>[120,121]</sup>. Thus, future studies are necessary to determine the relevance of these SNPs with respect to BP control in the general population.

Recent studies have implicated artery stiffness in the pathology of HTN and this parameter has been shown to be a valuable predictor of end organ failure<sup>[122-126]</sup>. Decreased vessel compliance elevates the mechanical load on the myocardium but also increases peripheral pulse-pressure in the microvasculature resulting in tissue damage in high flow organs such as the brain and kidneys. Until very recently, increased vascular stiffness during aging or the development of HTN was thought to result from changes in extracellular matrix content and composition (*i.e.*, elastin degradation, collagen deposition, *etc.*). However, new studies suggest that the intrinsic mechanical properties of VSMC (including RhoA-dependent formation of force-generating actin filaments, and increased cell adhesion to the extracellular matrix) may also play a role<sup>[127,128]</sup>. Notably, Liao *et al.*<sup>[129]</sup> identified two SNPs in ROCK2 that were in complete linkage disequilibrium and associated with arterial stiffness in 1483 un-selected patients from a Chinese population in Taiwan. Subsequent, *in vitro* studies revealed that both SNPs were functional. One SNP, rs978906, affected ROCK2 expression by interfering with microRNA(miR)-1183 binding to its 3' UTR, while the other, rs9808232, which was located in a protein-coding region, increased ROCK2 activity<sup>[129]</sup>.

As noted above, S1P is a major upstream activator of RhoA in SMC and has vasoconstrictive effects *in vivo*<sup>[71,130]</sup>. Interestingly, Fenger *et al.*<sup>[131,132]</sup> assessed the significance of 353 genetic variants contained within exons of genes in the metabolic sphingolipid network. Of these SNPs, 34 and 40 haplotypes were associated with changes in diastolic or systolic pressures respectively in their 2556 subjects. They found that while the BP effects could not be explained by any single gene, several 2-gene interaction pairs were highly correlated with BP variations. S1P is generated from ceramide in a process that involves two critical enzymes ceramidase (ASAH1) and sphingosine 1- kinase (SPHK1) and the most significant of the 2-gene interactions identified were contained in these genes<sup>[131,132]</sup>, further

supporting a role for RhoA signaling in the development of hypertension. It is likely that future gene interaction studies such as these will provide a powerful approach to both predict hypertension risk and possibly inform treatment options.

In the past decade, many GWAS studies have identified common genetic variations in coding and non-coding genomic regions that vary between individuals and are associated with changes in BP and several of these variants occur in genes linked to the Rho signaling cascade. Notably, one GWAS study that used hypertension as a dichotomous trait identified eight loci associated with BP, and two of these variants were located in RhoA-related genes. One of the target genes was the aforementioned RhoBTB1 which functions with the Cullin-3 complex to maintain low RhoA levels<sup>[107,118]</sup>. Another SNP was found at the rhotekin-2 (RHTKN) locus. Although rhotekin was one of the first identified RhoA effector molecules (it has high affinity to Rho-GTP and is widely used in pull down assays for activated RhoA<sup>[133]</sup>), how Rhotekin functions at a cellular level is still unclear. Nonetheless this association is provocative and clearly indicates that future studies are warranted. Two separate GWAS for BP variation and hypertension have identified significant association signals in the RhoA-interacting protein, plekstrin homology domain containing family A member 7 (PLEKHA7)<sup>[134,135]</sup>. PLEKHA7 is highly expressed in the kidney and heart and localizes on the cytoplasmic surface of adherens junctions, where it interacts with junctional proteins cingulin and paracingulin to regulate the activity of Rho family GTPases, including RhoA<sup>[136]</sup>. While the functional SNP(s) have yet to be identified, the finding that PLEKHA7 is required for the development of salt-induced hypertension *in vivo*, highlights the functional importance of this RhoA-interacting protein in BP regulation<sup>[137]</sup>.

Finally, two separate GWAS for BP and cardiovascular disease endpoints identified a novel BP associated locus containing two SNPs in perfect linkage disequilibrium (rs633185 and rs604723) within *GRAF3* gene (*ArhGAP42*). Both SNPs were associated with a significant reduction in BP with each copy of the minor allele<sup>[95,96]</sup>. Of extreme importance, as noted above, we reported that mice in which *GRAF3* was depleted developed significant hypertension that was RhoA-dependent<sup>[87]</sup>. Interestingly, the BP locus falls within the first intron of the *GRAF3* gene, indicating that one or both SNPs may affect expression of this Rho-GAP and result in altered SMC contractility. Indeed, data within the Genotype-Tissue Expression database indicated that *GRAF3* RNA levels in tibial artery samples were 3-fold higher in patients homozygous for the minor T allele compared to patients homozygous for the major C allele ( $P < 1.5 \times 10^{-10}$ ,<sup>[138]</sup>). Moreover, using allele-specific quantitative RT PCR on RNA isolated from human aortic SMC heterozygous at rs604723, we found that the minor T allele was associated with a significant increase in mRNA expression (Mangum and Mack, personal

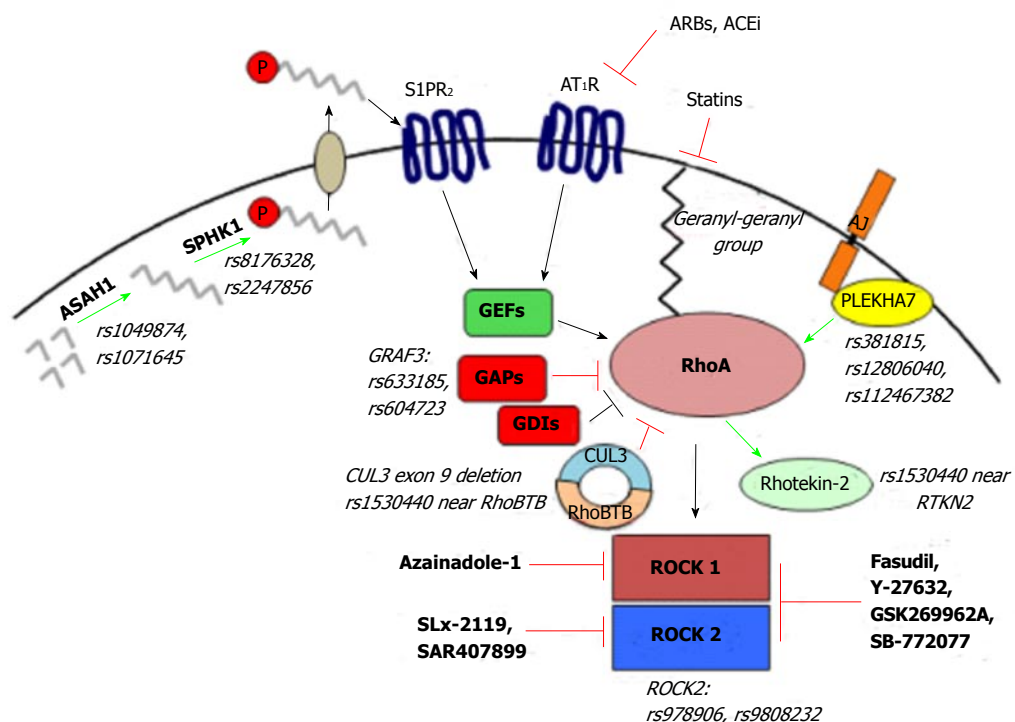
communication) and we identified a novel cis element by which this allele upregulates *GRAF3* transcription. To our knowledge, this is only the second functional SNP identified in a GWAS study that has been linked to a causal gene and pathway (the first being rs5068 located within *NPPA/B*<sup>[117]</sup>).

Collectively, these studies will likely have important implications in the future diagnosis and treatment of hypertension. For example, patients predicted to exhibit aberrantly high levels of RhoA signaling may respond better to anti-hypertensive regimens directly targeting vessel tone, compared to those that target blood volume. Moreover, they reveal that the RhoA signaling axis may provide highly selective targets for the treatment of human hypertension and related cardiovascular sequela.

## PHARMACOLOGICAL REGULATION OF RHOA AND RHO-DEPENDENT PATHWAYS

Despite the importance of RhoA signaling in the development of hypertension, few treatments are currently available that target this signaling axis. However, some commonly used anti-hypertensives may interfere with RhoA signaling (Figure 3). For example, since RhoA-dependent regulation of vascular tone is a major contributor to AII-mediated increases in BP<sup>[13,139]</sup>, the highly utilized class of anti-hypertensives that target AII (*i.e.*, ACE inhibitors and AII receptor blockers) may exert some of their BP lowering effects by reducing RhoA activation. Moreover, although used to treat high cholesterol, HMG-CoA reductase inhibitors such as simvastatin and atorvastatin also have anti-hypertensive properties<sup>[140]</sup> and their BP lowering effects have been attributed to their ability to block RhoA signaling. RhoA is known to be modified by covalent attachment of a geranylgeranyl isoprenyl to a C-terminal Cys, and this modification (which is blocked by simvastatin treatment) is required for membrane localization and activation of RhoA<sup>[141]</sup>.

While not yet included in standard of care treatment for hypertension, several pharmacologic agents have been developed for inhibiting Rho kinases. In general, kinases make good drug targets due to the relative ease of targeting specific molecules to the ATP-binding pockets of these enzymes. To date, most of the Rho kinase inhibitors utilized in animal studies and clinical trials target the ATP-binding pockets of both ROCK isoforms<sup>[142-144]</sup>. Although not clinically used in the United States, studies abroad provide compelling evidence for the use of this therapeutic approach for BP control. One particularly effective ROCK inhibitor, fasudil, is currently used in Japan to treat cerebral vasospasm and clinical trials determined that fasudil was also effective in decreasing peripheral vascular resistance in hypertensive patients<sup>[5]</sup>. However, despite their wide use in cells and animal disease models, neither



**Figure 3 Pharmacologic and genetic regulation of the RhoA signaling axis.** Schematic indicating the sites of action of pharmacological inhibitors (bold) of RhoA signaling molecules. Polymorphisms (SNPs/eQTLs) that could influence RhoA signaling are also shown. AJ: Adherens junction; A2R: Angiotensin type II receptor; ARBs: Angiotensin receptor blockers; ACEIs: Angiotensin converting enzyme inhibitors; ASAH1: Acid ceramidase; SPHK1: Sphingosine kinase 1.

fasudil nor Y-27632 exhibit suitable specificity for a therapeutic as they can inhibit the activity of several other kinases including PKC, PKA, and MLCK, at higher concentrations<sup>[145,146]</sup>. These compounds also suffer from having short half-lives, which is a highly undesirable attribute of a drug designed to treat a longstanding disease<sup>[147]</sup>. Thus, there is great need for development of additional potent, yet specific, ROCK inhibitors that can be safely used in patients<sup>[148]</sup>. While a few such compounds have been developed recently with such attributes<sup>[149-153]</sup>, whether any these compounds exhibit the necessary selectivity and pharmacogenetic profiles required for BP management in patients requires further study. Moving forward, given the importance of RhoGEFs and RhoGAPs in the control of SM contractility and BP, we believe that it will be possible to engineer clinically-relevant small molecule regulators of these enzymes that could be used to develop new and effective anti-hypertensive therapies.

## CONCLUSION: FUTURE POSSIBILITIES FOR PERSONALIZED TREATMENT OF HYPERTENSION

Current anti-hypertensive therapy is often empirically based and involves multiple drug regimens<sup>[154,155]</sup>—an approach that is moderately effective at best as it frequently contributes to unwanted side effects and intolerance or non-adherence to medication. Accordingly, more effective and specific anti-hyper-

tensive agents are necessary. Moreover, based on the fact that BP is a highly variable trait among individuals, a better understanding of the genetic mechanisms regulating this disease is critical for a more personalized treatment plan for patients. Given the numerous regulatory and counter-regulatory mechanisms modulating the RhoA axis, this central axis provides an excellent opportunity for identifying genetic biomarkers that correlate with different levels of hypertensive risk and drug responses. Indeed, genetic variations in both upstream activators and downstream mediators of RhoA have been linked to BP regulation (Figure 3). Screening for such variants could potentially be used to tailor more effective individualized treatments. For example, one study showed that the BP lowering effects the ACE inhibitors or the angiotensin receptor blockers were more pronounced in patients carrying a GG genotype at the -391 RGS2 (Regulators of G-protein signaling 2) locus when compared to responses in GC or CC genotype carriers—while no differences were observed in the responses to calcium channel antagonists<sup>[156]</sup>. Although RGS2 is known to couple to ATR1, the underlying mechanism by which these polymorphisms lead to altered sensitivity is currently unknown. Genetic differences in pharmacogenetics also play a role in response to anti-hypertensive agents, for example polymorphisms in the gene were associated with reduced BP-lowering effects of the  $\beta$ AR-blocker atenolol<sup>[157]</sup>. Whether any of the aforementioned Rho-signaling SNPs influence specific responses to or bio-availability of anti-hypertensive treatments remains

a critical unexplored question. The clinical utility of targeting the RhoA pathway should also be further explored.

## REFERENCES

- 1 **Go AS**, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Blaha MJ, Dai S, Ford ES, Fox CS, Franco S, Fullerton HJ, Gillespie C, Hailpern SM, Heit JA, Howard VJ, Huffman MD, Judd SE, Kissela BM, Kittner SJ, Lackland DT, Lichtman JH, Lisabeth LD, Mackey RH, Magid DJ, Marcus GM, Marelli A, Matchar DB, McGuire DK, Mohler ER, Moy CS, Mussolino ME, Neumar RW, Nichol G, Pandey DK, Paynter NP, Reeves MJ, Sorlie PD, Stein J, Towfighi A, Turan TN, Virani SS, Wong ND, Woo D, Turner MB. Heart disease and stroke statistics--2014 update: a report from the American Heart Association. *Circulation* 2014; **129**: e28-e292 [PMID: 24352519 DOI: 10.1161/01.cir.0000441139.02102.80]
- 2 **Heasman SJ**, Ridley AJ. Mammalian Rho GTPases: new insights into their functions from in vivo studies. *Nat Rev Mol Cell Biol* 2008; **9**: 690-701 [PMID: 18719708 DOI: 10.1038/nrm2476]
- 3 **Loirand G**, Pacaud P. The role of Rho protein signaling in hypertension. *Nat Rev Cardiol* 2010; **7**: 637-647 [PMID: 20808285 DOI: 10.1038/nrcardio.2010.136]
- 4 **Wirth A**, Benyó Z, Lukasova M, Leutgeb B, Wettschurek N, Gorbey S, Orsy P, Horváth B, Maser-Gluth C, Greiner E, Lemmer B, Schütz G, Gutkind JS, Offermanns S. G12-G13-LARG-mediated signaling in vascular smooth muscle is required for salt-induced hypertension. *Nat Med* 2008; **14**: 64-68 [PMID: 18084302 DOI: 10.1038/nm1666]
- 5 **Masumoto A**, Hirooka Y, Shimokawa H, Hironaga K, Setoguchi S, Takeshita A. Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. *Hypertension* 2001; **38**: 1307-1310 [PMID: 11751708 DOI: 10.1161/hy1201.096541]
- 6 **Cowley AW**. The genetic dissection of essential hypertension. *Nat Rev Genet* 2006; **7**: 829-840 [PMID: 17033627 DOI: 10.1038/nrg1967]
- 7 **Davis MJ**, Wu X, Nurkiewicz TR, Kawasaki J, Davis GE, Hill MA, Meininger GA. Integrins and mechanotransduction of the vascular myogenic response. *Am J Physiol Heart Circ Physiol* 2001; **280**: H1427-H1433 [PMID: 11247750]
- 8 **Davis MJ**, Hill MA. Signaling mechanisms underlying the vascular myogenic response. *Physiol Rev* 1999; **79**: 387-423 [PMID: 10221985]
- 9 **Hall JE**. The kidney, hypertension, and obesity. *Hypertension* 2003; **41**: 625-633 [PMID: 12623970 DOI: 10.1161/01.HYP.0000052314.95497.78]
- 10 **Lifton RP**, Gharavi AG, Geller DS. Molecular mechanisms of human hypertension. *Cell* 2001; **104**: 545-556 [PMID: 11239411 DOI: 10.1016/S0092-8674(01)00241-0]
- 11 **Etienne-Manneville S**, Hall A. Rho GTPases in cell biology. *Nature* 2002; **420**: 629-635 [PMID: 12478284 DOI: 10.1038/nature01148]
- 12 **Budzyn K**, Marley PD, Sobey CG. Targeting Rho and Rho-kinase in the treatment of cardiovascular disease. *Trends Pharmacol Sci* 2006; **27**: 97-104 [PMID: 16376997 DOI: 10.1016/j.tips.2005.12.002]
- 13 **Guilluy C**, Brégeon J, Toumaniantz G, Rolli-Derkinderen M, Retailleau K, Loufrani L, Henrion D, Scalbert E, Bril A, Torres RM, Offermanns S, Pacaud P, Loirand G. The Rho exchange factor Arhgef1 mediates the effects of angiotensin II on vascular tone and blood pressure. *Nat Med* 2010; **16**: 183-190 [PMID: 20098430 DOI: 10.1038/nm2079]
- 14 **Amano M**, Ito M, Kimura K, Fukata Y, Chihara K, Nakano T, Matsuura Y, Kaibuchi K. Phosphorylation and activation of myosin by Rho-associated kinase (Rho-kinase). *J Biol Chem* 1996; **271**: 20246-20249 [PMID: 8702756 DOI: 10.1074/jbc.271.34.20246]
- 15 **Kimura K**, Ito M, Amano M, Chihara K, Fukata Y, Nakafuku M, Yamamori B, Feng J, Nakano T, Okawa K, Iwamatsu A, Kaibuchi K. Regulation of myosin phosphatase by Rho and Rho-associated kinase (Rho-kinase). *Science* 1996; **273**: 245-248 [PMID: 8662509 DOI: 10.1126/science.273.5272.245]
- 16 **Mueller BK**, Mack H, Teusch N. Rho kinase, a promising drug target for neurological disorders. *Nat Rev Drug Discov* 2005; **4**: 387-398 [PMID: 15864268 DOI: 10.1038/nrd1719]
- 17 **Guilluy C**, Eddahibi S, Agard C, Guignabert C, Izikki M, Tu L, Savale L, Humbert M, Fadel E, Adnot S, Loirand G, Pacaud P. RhoA and Rho kinase activation in human pulmonary hypertension: role of 5-HT signaling. *Am J Respir Crit Care Med* 2009; **179**: 1151-1158 [PMID: 19299501 DOI: 10.1164/rccm.200805-691OC]
- 18 **Ohashi K**, Nagata K, Maekawa M, Ishizaki T, Narumiya S, Mizuno K. Rho-associated kinase ROCK activates LIM-kinase 1 by phosphorylation at threonine 508 within the activation loop. *J Biol Chem* 2000; **275**: 3577-3582 [PMID: 10652353 DOI: 10.1074/jbc.275.5.3577]
- 19 **Sumi T**, Matsumoto K, Nakamura T. Specific activation of LIM kinase 2 via phosphorylation of threonine 505 by ROCK, a Rho-dependent protein kinase. *J Biol Chem* 2001; **276**: 670-676 [PMID: 11018042 DOI: 10.1074/jbc.M007074200]
- 20 **Wirth A**. Rho kinase and hypertension. *Biochim Biophys Acta* 2010; **1802**: 1276-1284 [PMID: 20460153 DOI: 10.1016/j.bbdis.2010.05.002]
- 21 **Yang N**, Higuchi O, Ohashi K, Nagata K, Wada A, Kangawa K, Nishida E, Mizuno K. Cofilin phosphorylation by LIM-kinase 1 and its role in Rac-mediated actin reorganization. *Nature* 1998; **393**: 809-812 [PMID: 9655398]
- 22 **Vardoulis L**, Moustakas A, Stournaras C. LIM-kinase 2 and cofilin phosphorylation mediate actin cytoskeleton reorganization induced by transforming growth factor-beta. *J Biol Chem* 2005; **280**: 11448-11457 [PMID: 15647284 DOI: 10.1074/jbc.M402651200]
- 23 **Matsui T**, Maeda M, Doi Y, Yonemura S, Amano M, Kaibuchi K, Tsukita S, Tsukita S. Rho-kinase phosphorylates COOH-terminal threonines of ezrin/radixin/moesin (ERM) proteins and regulates their head-to-tail association. *J Cell Biol* 1998; **140**: 647-657 [PMID: 9456324 DOI: 10.1083/jcb.140.3.647]
- 24 **Lockman K**, Hinson JS, Medlin MD, Morris D, Taylor JM, Mack CP. Sphingosine 1-phosphate stimulates smooth muscle cell differentiation and proliferation by activating separate serum response factor co-factors. *J Biol Chem* 2004; **279**: 42422-42430 [PMID: 15292266 DOI: 10.1074/jbc.M405432200]
- 25 **Olson EN**, Nordheim A. Linking actin dynamics and gene transcription to drive cellular motile functions. *Nat Rev Mol Cell Biol* 2010; **11**: 353-365 [PMID: 20414257 DOI: 10.1038/nrm2890]
- 26 **Laufs U**, Liao JK. Post-transcriptional regulation of endothelial nitric oxide synthase mRNA stability by Rho GTPase. *J Biol Chem* 1998; **273**: 24266-24271 [PMID: 9727051 DOI: 10.1074/jbc.273.37.24266]
- 27 **Zhou W**, Negash S, Liu J, Raj JU. Modulation of pulmonary vascular smooth muscle cell phenotype in hypoxia: role of cGMP-dependent protein kinase and myocardin. *Am J Physiol Lung Cell Mol Physiol* 2009; **296**: L780-L789 [PMID: 19251841 DOI: 10.1152/ajplung.90295.2008]
- 28 **Zhou Q**, Liao JK. Rho kinase: an important mediator of atherosclerosis and vascular disease. *Curr Pharm Des* 2009; **15**: 3108-3115 [PMID: 19754385 DOI: 10.2174/138161209789057986]
- 29 **Wolfrum S**, Dendorfer A, Rikitake Y, Stalker TJ, Gong Y, Scalia R, Dominiak P, Liao JK. Inhibition of Rho-kinase leads to rapid activation of phosphatidylinositol 3-kinase/protein kinase Akt and cardiovascular protection. *Arterioscler Thromb Vasc Biol* 2004; **24**: 1842-1847 [PMID: 15319269 DOI: 10.1161/01.ATV.0000142813.33538.82]
- 30 **Ming XF**, Barandier C, Viswambharan H, Kwak BR, Mach F, Mazzolai L, Hayoz D, Ruffieux J, Rusconi S, Montani JP, Yang Z. Thrombin stimulates human endothelial arginase enzymatic activity via RhoA/ROCK pathway: implications for atherosclerotic endothelial dysfunction. *Circulation* 2004; **110**: 3708-3714 [PMID: 15569838 DOI: 10.1161/01.CIR.0000142867.26182.32]
- 31 **Carlström M**, Wilcox CS, Arendshorst WJ. Renal autoregulation in health and disease. *Physiol Rev* 2015; **95**: 405-511 [PMID: 25834230 DOI: 10.1152/physrev.00042.2012]



- 32 **Inscho EW**. ATP, P2 receptors and the renal microcirculation. *Purinergic Signal* 2009; **5**: 447-460 [PMID: 19294530 DOI: 10.1007/s11302-009-9147-1]
- 33 **Yano H**, Hayashi K, Momiyama T, Saga H, Haruna M, Sobue K. Transcriptional regulation of the chicken caldesmon gene. Activation of gizzard-type caldesmon promoter requires a CArG box-like motif. *J Biol Chem* 1995; **270**: 23661-23666 [PMID: 7559534 DOI: 10.1074/jbc.270.40.23661]
- 34 **Homma K**, Hayashi K, Wakino S, Tokuyama H, Kanda T, Tatematsu S, Hasegawa K, Fujishima S, Hori S, Saruta T, Itoh H. Rho-kinase contributes to pressure-induced constriction of renal microvessels. *Keio J Med* 2014; **63**: 1-12 [PMID: 24429483 DOI: 10.2302/kjm.2013-0001-OA]
- 35 **Roos MH**, van Rodijnen WF, van Lambalgen AA, ter Wee PM, Tangelder GJ. Renal microvascular constriction to membrane depolarization and other stimuli: pivotal role for rho-kinase. *Pflugers Arch* 2006; **452**: 471-477 [PMID: 16523358 DOI: 10.1007/s00424-006-0053-x]
- 36 **Shi Y**, Wang X, Chon KH, Cupples WA. Tubuloglomerular feedback-dependent modulation of renal myogenic autoregulation by nitric oxide. *Am J Physiol Regul Integr Comp Physiol* 2006; **290**: R982-R991 [PMID: 16293681 DOI: 10.1152/ajpregu.00346.2005]
- 37 **Nakamura A**, Hayashi K, Ozawa Y, Fujiwara K, Okubo K, Kanda T, Wakino S, Saruta T. Vessel- and vasoconstrictor-dependent role of rho/rho-kinase in renal microvascular tone. *J Vasc Res* 2003; **40**: 244-251 [PMID: 12902637 DOI: 10.1159/000071888]
- 38 **Loirand G**, Pacaud P. Involvement of Rho GTPases and their regulators in the pathogenesis of hypertension. *Small GTPases* 2014; **5**: 1-10 [PMID: 25496262 DOI: 10.4161/sgtp.28846]
- 39 **Staruschenko A**, Nichols A, Medina JL, Camacho P, Zheleznova NN, Stockand JD. Rho small GTPases activate the epithelial Na<sup>(+)</sup> channel. *J Biol Chem* 2004; **279**: 49989-49994 [PMID: 15448132 DOI: 10.1074/jbc.M409812200]
- 40 **Pochynyuk O**, Medina J, Gamber N, Genth H, Stockand JD, Staruschenko A. Rapid translocation and insertion of the epithelial Na<sup>+</sup> channel in response to RhoA signaling. *J Biol Chem* 2006; **281**: 26520-26527 [PMID: 16829523 DOI: 10.1074/jbc.M603716200]
- 41 **Karpushev AV**, Ilatovskaya DV, Pavlov TS, Negulyaev YA, Staruschenko A. Intact cytoskeleton is required for small G protein dependent activation of the epithelial Na<sup>+</sup> channel. *PLoS One* 2010; **5**: e8827 [PMID: 20098689 DOI: 10.1371/journal.pone.0008827]
- 42 **Hayashi H**, Szász K, Coady-Osberg N, Furuya W, Bretscher AP, Orlowski J, Grinstein S. Inhibition and redistribution of NHE3, the apical Na<sup>+</sup>/H<sup>+</sup> exchanger, by Clostridium difficile toxin B. *J Gen Physiol* 2004; **123**: 491-504 [PMID: 15078917 DOI: 10.1085/jgp.200308979]
- 43 **Szász K**, Kurashima K, Kapus A, Paulsen A, Kaibuchi K, Grinstein S, Orlowski J. RhoA and rho kinase regulate the epithelial Na<sup>+</sup>/H<sup>+</sup> exchanger NHE3. Role of myosin light chain phosphorylation. *J Biol Chem* 2000; **275**: 28599-28606 [PMID: 10893221 DOI: 10.1074/jbc.M001193200]
- 44 **Nishiki K**, Tsuruoka S, Kawaguchi A, Sugimoto K, Schwartz GJ, Suzuki M, Imai M, Fujimura A. Inhibition of Rho-kinase reduces renal Na-H exchanger activity and causes natriuresis in rat. *J Pharmacol Exp Ther* 2003; **304**: 723-728 [PMID: 12538827 DOI: 10.1124/jpet.102.041871]
- 45 **Struthers AD**, Dollery CT. Central nervous system mechanisms in blood pressure control. *Eur J Clin Pharmacol* 1985; **28** Suppl: 3-11 [PMID: 2865146 DOI: 10.1007/BF00543703]
- 46 **Pilowsky PM**, Goodchild AK. Baroreceptor reflex pathways and neurotransmitters: 10 years on. *J Hypertens* 2002; **20**: 1675-1688 [PMID: 12195099]
- 47 **Ito K**, Hirooka Y, Sakai K, Kishi T, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in brain stem contributes to blood pressure regulation via sympathetic nervous system: possible involvement in neural mechanisms of hypertension. *Circ Res* 2003; **92**: 1337-1343 [PMID: 12791705 DOI: 10.1161/01.RES.0000079941.59846.D4]
- 48 **Ito K**, Hirooka Y, Kimura Y, Shimokawa H, Takeshita A. Effects of hydroxyfasudil administered to the nucleus tractus solitarius on blood pressure and heart rate in spontaneously hypertensive rats. *Clin Exp Hypertens* 2005; **27**: 269-277 [PMID: 15835390 DOI: 10.1081/CEH-200048876]
- 49 **Sagara Y**, Hirooka Y, Nozoe M, Ito K, Kimura Y, Sunagawa K. Pressor response induced by central angiotensin II is mediated by activation of Rho/Rho-kinase pathway via AT1 receptors. *J Hypertens* 2007; **25**: 399-406 [PMID: 17211247 DOI: 10.1097/HJH.0b013e328010b87f]
- 50 **Yamaguchi Y**, Katoh H, Yasui H, Aoki J, Nakamura K, Negishi M. Galpha(12) and galpha(13) inhibit Ca(2+)-dependent exocytosis through Rho/Rho-associated kinase-dependent pathway. *J Neurochem* 2000; **75**: 708-717 [PMID: 10899946 DOI: 10.1046/j.1471-4159.2000.0750708.x]
- 51 **Salu KJ**, Bosmans JM, Huang Y, Hendriks M, Verhoeven M, Levels A, Cooper S, De Scheerder IK, Vrints CJ, Bult H. Effects of cytochalasin D-eluting stents on intimal hyperplasia in a porcine coronary artery model. *Cardiovasc Res* 2006; **69**: 536-544 [PMID: 16386237 DOI: 10.1016/j.cardiores.2005.11.012]
- 52 **Noma K**, Goto C, Nishioka K, Jitsuiki D, Umemura T, Ueda K, Kimura M, Nakagawa K, Oshima T, Chayama K, Yoshizumi M, Liao JK, Higashi Y. Roles of rho-associated kinase and oxidative stress in the pathogenesis of aortic stiffness. *J Am Coll Cardiol* 2007; **49**: 698-705 [PMID: 17291936 DOI: 10.1016/j.jacc.2006.06.082]
- 53 **Hiley E**, McMullan R, Nurrish SJ. The Galpha12-RGS RhoGEF-RhoA signalling pathway regulates neurotransmitter release in C. elegans. *EMBO J* 2006; **25**: 5884-5895 [PMID: 17139250 DOI: 10.1038/sj.emboj.7601458]
- 54 **Wei L**, Taffet GE, Khoury DS, Bo J, Li Y, Yatani A, Delaughter MC, Kleivitsky R, Hewett TE, Robbins J, Michael LH, Schneider MD, Entman ML, Schwartz RJ. Disruption of Rho signaling results in progressive atrioventricular conduction defects while ventricular function remains preserved. *FASEB J* 2004; **18**: 857-859 [PMID: 15033930 DOI: 10.1096/fj.03-0664fj]
- 55 **Yatani A**, Irie K, Otani T, Abdellatif M, Wei L. RhoA GTPase regulates L-type Ca<sup>2+</sup> currents in cardiac myocytes. *Am J Physiol Heart Circ Physiol* 2005; **288**: H650-H659 [PMID: 15471984 DOI: 10.1152/ajpheart.00268.2004]
- 56 **Vlasblom R**, Muller A, Beckers CM, van Nieuw Amerongen GP, Zuidwijk MJ, van Hardeveld C, Paulus WJ, Simonides WS. RhoA-ROCK signaling is involved in contraction-mediated inhibition of SERCA2a expression in cardiomyocytes. *Pflugers Arch* 2009; **458**: 785-793 [PMID: 19294414 DOI: 10.1007/s00424-009-0659-x]
- 57 **Vahebi S**, Kobayashi T, Warren CM, de Tombe PP, Solaro RJ. Functional effects of rho-kinase-dependent phosphorylation of specific sites on cardiac troponin. *Circ Res* 2005; **96**: 740-747 [PMID: 15774859 DOI: 10.1161/01.RES.0000162457.56568.7d]
- 58 **Draeger A**, Stelzer EH, Herzog M, Small JV. Unique geometry of actin-membrane anchorage sites in avian gizzard smooth muscle cells. *J Cell Sci* 1989; **94** (Pt 4): 703-711 [PMID: 2630565]
- 59 **Lauriol J**, Keith K, Jaffré F, Couvillon A, Saci A, Goonasekera SA, McCarthy JR, Kessinger CW, Wang J, Ke Q, Kang PM, Molkentin JD, Carpenter C, Kontaridis MI. RhoA signaling in cardiomyocytes protects against stress-induced heart failure but facilitates cardiac fibrosis. *Sci Signal* 2014; **7**: ra100 [PMID: 25336613 DOI: 10.1126/scisignal.2005262]
- 60 **Stepanova OV**, Chadin AV, Masiutin AG, Kulikova TG, Gurin IaV, Sergeeva IA, Shirinski VP. Rho-associated protein kinase is involved in establishing the contractile phenotype of cardiomyocytes. *Biofizika* 2010; **55**: 880-885 [PMID: 21033356]
- 61 **Bos JL**, Rehmann H, Wittinghofer A. GEFs and GAPs: critical elements in the control of small G proteins. *Cell* 2007; **129**: 865-877 [PMID: 17540168 DOI: 10.1016/j.cell.2007.05.018]
- 62 **Aittaleb M**, Boguth CA, and Tesmer JJG. Structure and Function of Heterotrimeric G Protein-Regulated Rho Guanine Nucleotide Exchange Factors. *Mol Pharmacol* 2010; **77**: 111-125 [PMID: 19880753 DOI: 10.1124/mol.109.061234]
- 63 **Yamada T**, Ohoka Y, Kogo M, Inagaki S. Physical and functional

- interactions of the lysophosphatidic acid receptors with PDZ domain-containing Rho guanine nucleotide exchange factors (RhoGEFs). *J Biol Chem* 2005; **280**: 19358-19363 [PMID: 15755723 DOI: 10.1074/jbc.M414561200]
- 64 **Suzuki N**, Nakamura S, Mano H, Kozasa T. Galpha 12 activates Rho GTPase through tyrosine-phosphorylated leukemia-associated RhoGEF. *Proc Natl Acad Sci USA* 2003; **100**: 733-738 [PMID: 12515866 DOI: 10.1073/pnas.0234057100]
  - 65 **Fukuhara S**, Chikumi H, Gutkind JS. RGS-containing RhoGEFs: the missing link between transforming G proteins and Rho? *Oncogene* 2001; **20**: 1661-1668 [PMID: 11313914]
  - 66 **Bregeon J**, Loirand G, Pacaud P, Rolli-Derkinderen M. Angiotensin II induces RhoA activation through SHP2-dependent dephosphorylation of the RhoGAP p190A in vascular smooth muscle cells. *Am J Physiol Cell Physiol* 2009; **297**: C1062-C1070 [PMID: 19692654 DOI: 10.1152/ajpcell.00174.2009]
  - 67 **Ying Z**, Jin L, Palmer T, Webb RC. Angiotensin II up-regulates the leukemia-associated Rho guanine nucleotide exchange factor (RhoGEF), a regulator of G protein signaling domain-containing RhoGEF, in vascular smooth muscle cells. *Mol Pharmacol* 2006; **69**: 932-940 [PMID: 16354763 DOI: 10.1124/mol.105.017830]
  - 68 **Hilgers RH**, Todd J, Webb RC. Increased PDZ-RhoGEF/RhoA/Rho kinase signaling in small mesenteric arteries of angiotensin II-induced hypertensive rats. *J Hypertens* 2007; **25**: 1687-1697 [PMID: 17620967 DOI: 10.1097/HJH.0b013e32816f778d]
  - 69 **Ying Z**, Giachini FR, Tostes RC, Webb RC. PYK2/PDZ-RhoGEF links Ca<sup>2+</sup> signaling to RhoA. *Arterioscler Thromb Vasc Biol* 2009; **29**: 1657-1663 [PMID: 19759375 DOI: 10.1161/ATVBAHA.109.190892]
  - 70 **Artamonov MV**, Momotani K, Stevenson A, Trentham DR, Derewenda U, Derewenda ZS, Read PW, Gutkind JS, Somlyo AV. Agonist-induced Ca<sup>2+</sup> sensitization in smooth muscle: redundancy of Rho guanine nucleotide exchange factors (RhoGEFs) and response kinetics, a caged compound study. *J Biol Chem* 2013; **288**: 34030-34040 [PMID: 24106280 DOI: 10.1074/jbc.M113.514596]
  - 71 **Medlin MD**, Staus DP, Dubash AD, Taylor JM, Mack CP. Sphingosine 1-phosphate receptor 2 signals through leukemia-associated RhoGEF (LARG), to promote smooth muscle cell differentiation. *Arterioscler Thromb Vasc Biol* 2010; **30**: 1779-1786 [PMID: 20702813 DOI: 10.1161/ATVBAHA.110.209395]
  - 72 **Guilluy C**, Swaminathan V, Garcia-Mata R, O'Brien ET, Superfine R, Burrige K. The Rho GEFs LARG and GEF-H1 regulate the mechanical response to force on integrins. *Nat Cell Biol* 2011; **13**: 722-727 [PMID: 21572419 DOI: 10.1038/ncb2254]
  - 73 **Rojas RJ**, Yohe ME, Gershburg S, Kawano T, Kozasa T, Sondek J. Galphq directly activates p63RhoGEF and Trio via a conserved extension of the Dbl homology-associated pleckstrin homology domain. *J Biol Chem* 2007; **282**: 29201-29210 [PMID: 17606614 DOI: 10.1074/jbc.M703458200]
  - 74 **Lutz S**, Shankaranarayanan A, Coco C, Ridilla M, Nance MR, Vettel C, Baltus D, Evelyn CR, Neubig RR, Wieland T, Tesmer JJ. Structure of Galphq-p63RhoGEF-RhoA complex reveals a pathway for the activation of RhoA by GPCRs. *Science* 2007; **318**: 1923-1927 [PMID: 18096806 DOI: 10.1126/science.1147554]
  - 75 **Wuertz CM**, Lorincz A, Vettel C, Thomas MA, Wieland T, Lutz S. p63RhoGEF--a key mediator of angiotensin II-dependent signaling and processes in vascular smooth muscle cells. *FASEB J* 2010; **24**: 4865-4876 [PMID: 20739613 DOI: 10.1096/fj.10-155499]
  - 76 **Momotani K**, Artamonov MV, Utepbergenov D, Derewenda U, Derewenda ZS, Somlyo AV. p63RhoGEF couples Ga(q/11)-mediated signaling to Ca<sup>2+</sup> sensitization of vascular smooth muscle contractility. *Circ Res* 2011; **109**: 993-1002 [PMID: 21885830 DOI: 10.1161/CIRCRESAHA.111.248898]
  - 77 **Bear MD**, Li M, Liu Y, Giel-Moloney MA, Fanburg BL, Toksoz D. The Lbc Rho guanine nucleotide exchange factor  $\alpha$ -catulin axis functions in serotonin-induced vascular smooth muscle cell mitogenesis and RhoA/ROCK activation. *J Biol Chem* 2010; **285**: 32919-32926 [PMID: 20696764 DOI: 10.1074/jbc.M109.062513]
  - 78 **Chikumi H**, Fukuhara S, Gutkind JS. Regulation of G protein-linked guanine nucleotide exchange factors for Rho, PDZ-RhoGEF, and LARG by tyrosine phosphorylation: evidence of a role for focal adhesion kinase. *J Biol Chem* 2002; **277**: 12463-12473 [PMID: 11799111 DOI: 10.1074/jbc.M108504200]
  - 79 **Jin L**, Ying Z, Hilgers RH, Yin J, Zhao X, Imig JD, Webb RC. Increased RhoA/Rho-kinase signaling mediates spontaneous tone in aorta from angiotensin II-induced hypertensive rats. *J Pharmacol Exp Ther* 2006; **318**: 288-295 [PMID: 16569756 DOI: 10.1124/jpet.105.100735]
  - 80 **Cario-Toumaniantz C**, Ferland-McCollough D, Chadeuf G, Toumaniantz G, Rodriguez M, Galizzi JP, Lockhart B, Bril A, Scalbert E, Loirand G, Pacaud P. RhoA guanine exchange factor expression profile in arteries: evidence for a Rho kinase-dependent negative feedback in angiotensin II-dependent hypertension. *Am J Physiol Cell Physiol* 2012; **302**: C1394-C1404 [PMID: 22322975 DOI: 10.1152/ajpcell.00423.2011]
  - 81 **Ying Z**, Jin L, Dorrance AM, Webb RC. Increased expression of mRNA for regulator of G protein signaling domain-containing Rho guanine nucleotide exchange factors in aorta from stroke-prone spontaneously hypertensive rats. *Am J Hypertens* 2004; **17**: 981-985 [PMID: 15485764 DOI: 10.1016/j.amjhyper.2004.05.006]
  - 82 **Benter IF**, Canatan H, Benboubetra M, Yousif MH, Akhtar S. Global upregulation of gene expression associated with renal dysfunction in DOCA-salt-induced hypertensive rats occurs via signaling cascades involving epidermal growth factor receptor: a microarray analysis. *Vascul Pharmacol* 2009; **51**: 101-109 [PMID: 19410658 DOI: 10.1016/j.vph.2009.04.004]
  - 83 **Puetz S**, Lubomirov LT, Pfitzer G. Regulation of smooth muscle contraction by small GTPases. *Physiology* (Bethesda) 2009; **24**: 342-356 [PMID: 19996365 DOI: 10.1152/physiol.00023.2009]
  - 84 **Rittinger K**, Walker PA, Eccleston JF, Smerdon SJ, Gambin SJ. Structure at 1.65 Å of RhoA and its GTPase-activating protein in complex with a transition-state analogue. *Nature* 1997; **389**: 758-762 [PMID: 9338791]
  - 85 **Tcherkezian J**, Lamarche-Vane N. Current knowledge of the large RhoGAP family of proteins. *Biol Cell* 2007; **99**: 67-86 [PMID: 17222083 DOI: 10.1042/BC20060086]
  - 86 **Mori K**, Amano M, Takefuji M, Kato K, Morita Y, Nishioka T, Matsuura Y, Murohara T, Kaibuchi K. Rho-kinase contributes to sustained RhoA activation through phosphorylation of p190A RhoGAP. *J Biol Chem* 2009; **284**: 5067-5076 [PMID: 19103606 DOI: 10.1074/jbc.M806853200]
  - 87 **Bai X**, Lenhart KC, Bird KE, Suen AA, Rojas M, Kakoki M, Li F, Smithies O, Mack CP, Taylor JM. The smooth muscle-selective RhoGAP GRAF3 is a critical regulator of vascular tone and hypertension. *Nat Commun* 2013; **4**: 2910 [PMID: 24335996 DOI: 10.1038/ncomms3910]
  - 88 **Hildebrand JD**, Taylor JM, Parsons JT. An SH3 domain-containing GTPase-activating protein for Rho and Cdc42 associates with focal adhesion kinase. *Mol Cell Biol* 1996; **16**: 3169-3178 [PMID: 8649427 DOI: 10.1128/MCB.16.6.3169]
  - 89 **Taylor JM**, Macklem MM, Parsons JT. Cytoskeletal changes induced by GRAF, the GTPase regulator associated with focal adhesion kinase, are mediated by Rho. *J Cell Sci* 1999; **112** (Pt 2): 231-242 [PMID: 9858476]
  - 90 **Taylor JM**, Hildebrand JD, Mack CP, Cox ME, Parsons JT. Characterization of graf, the GTPase-activating protein for rho associated with focal adhesion kinase. Phosphorylation and possible regulation by mitogen-activated protein kinase. *J Biol Chem* 1998; **273**: 8063-8070 [PMID: 9525907 DOI: 10.1074/jbc.273.14.8063]
  - 91 **Doherty JT**, Lenhart KC, Cameron MV, Mack CP, Conlon FL, Taylor JM. Skeletal muscle differentiation and fusion are regulated by the BAR-containing Rho-GTPase-activating protein (Rho-GAP), GRAF1. *J Biol Chem* 2011; **286**: 25903-25921 [PMID: 21622574 DOI: 10.1074/jbc.M111.243030]
  - 92 **Lenhart KC**, Becherer AL, Li J, Xiao X, McNally EM, Mack CP, Taylor JM. GRAF1 promotes ferlin-dependent myoblast fusion. *Dev Biol* 2014; **393**: 298-311 [PMID: 25019370 DOI: 10.1016/j.ydbio.2014.06.025]

- 93 **Lenhart KC**, O'Neill TJ, Cheng Z, Dee R, Demonbreun AR, Li J, Xiao X, McNally EM, Mack CP, Taylor JM. GRAF1 deficiency blunts sarcolemmal injury repair and exacerbates cardiac and skeletal muscle pathology in dystrophin-deficient mice. *Skelet Muscle* 2015; **5**: 27 [PMID: 26301073 DOI: 10.1186/s13395-015-0054-6]
- 94 **Ren XR**, Du QS, Huang YZ, Ao SZ, Mei L, Xiong WC. Regulation of CDC42 GTPase by proline-rich tyrosine kinase 2 interacting with PSGAP, a novel pleckstrin homology and Src homology 3 domain containing rhoGAP protein. *J Cell Biol* 2001; **152**: 971-984 [PMID: 11238453 DOI: 10.1083/jcb.152.5.971]
- 95 **Ehret GB**, Munroe PB, Rice KM, Bochud M, Johnson AD, Chasman DI, Smith AV, Tobin MD, Verwoert GC, Hwang SJ, Pihur V, Vollenweider P, O'Reilly PF, Amin N, Bragg-Gresham JL, Teumer A, Glazer NL, Launer L, Zhao JH, Aulchenko Y, Heath S, Söber S, Parsa A, Luan J, Arora P, Dehghan A, Zhang F, Lucas G, Hicks AA, Jackson AU, Peden JF, Tanaka T, Wild SH, Rudan I, Igl W, Milaneschi Y, Parker AN, Fava C, Chambers JC, Fox ER, Kumari M, Go MJ, van der Harst P, Kao WH, Sjögren M, Vinay DG, Alexander M, Tabara Y, Shaw-Hawkins S, Whincup PH, Liu Y, Shi G, Kuusisto J, Tayo B, Seielstad M, Sim X, Nguyen KD, Lehtimäki T, Matullo G, Wu Y, Gaunt TR, Onland-Moret NC, Cooper MN, Platou CG, Org E, Hardy R, Dahgam S, Palmen J, Vitart V, Braund PS, Kuznetsova T, Uitterwaal CS, Adeyemo A, Palmas W, Campbell H, Ludwig B, Tomaszewski M, Tzoulaki I, Palmer ND, Aspelund T, Garcia M, Chang YP, O'Connell JR, Steinle NI, Grobbee DE, Arking DE, Kardina SL, Morrison AC, Hernandez D, Najjar S, McArdle WL, Hadley D, Brown MJ, Connell JM, Hingorani AD, Day IN, Lawlor DA, Beilby JP, Lawrence RW, Clarke R, Hopewell JC, Ongen H, Dreisbach AW, Li Y, Young JH, Bis JC, Kähönen M, Viikari J, Adair LS, Lee NR, Chen MH, Olden M, Pattaro C, Bolton JA, Köttgen A, Bergmann S, Mooser V, Chaturvedi N, Frayling TM, Islam M, Jafar TH, Erdmann J, Kulkarni SR, Bornstein SR, Grässler J, Groop L, Voight BF, Kettunen J, Howard P, Taylor A, Guarrera S, Ricceri F, Emilsson V, Plump A, Barroso I, Khaw KT, Weder AB, Hunt SC, Sun YV, Bergman RN, Collins FS, Bonnycastle LL, Scott LJ, Stringham HM, Peltonen L, Perola M, Vartiainen E, Brand SM, Staessen JA, Wang TJ, Burton PR, Soler Artigas M, Dong Y, Snieder H, Wang X, Zhu H, Lohman KK, Rudock ME, Heckbert SR, Smith NL, Wiggins KL, Doumatey A, Shriner D, Veldre G, Viigimaa M, Kinra S, Prabhakaran D, Tripathy V, Langefeld CD, Rosengren A, Thelle DS, Corsi AM, Singleton A, Forrester T, Hilton G, McKenzie CA, Salako T, Iwai N, Kita Y, Ogihara T, Ohkubo T, Okamura T, Ueshima H, Umemura S, Eyheramendy S, Meitinger T, Wichmann HE, Cho YS, Kim HL, Lee JY, Scott J, Sehmi JS, Zhang W, Hedblad B, Nilsson P, Smith GD, Wong A, Narisu N, Stančáková A, Raffel LJ, Yao J, Kathiresan S, O'Donnell CJ, Schwartz SM, Ikrum MA, Longstreth WT, Mosley TH, Seshadri S, Shrine NR, Wain LV, Morken MA, Swift AJ, Laitinen J, Prokopenko I, Zitting P, Cooper JA, Humphries SE, Danesh J, Rasheed A, Goel A, Hamsten A, Watkins H, Bakker SJ, van Gilst WH, Janipalli CS, Mani KR, Yajnik CS, Hofman A, Mattace-Raso FU, Oostra BA, Demirkan A, Isaacs A, Rivadeneira F, Lakatta EG, Orru M, Scuteri A, Ala-Korpela M, Kangas AJ, Lyytikäinen LP, Soininen P, Tukiainen T, Würtz P, Ong RT, Dörr M, Kroemer HK, Völker U, Völzke H, Galan P, Hercberg S, Lathrop M, Zelenika D, Deloukas P, Mangino M, Spector TD, Zhai G, Meschia JF, Nalls MA, Sharma P, Terzic J, Kumar MV, Denniff M, Zukowska-Szczekowska E, Wagenknecht LE, Fowkes FG, Charchar FJ, Schwarz PE, Hayward C, Guo X, Rotimi C, Bots ML, Brand E, Samani NJ, Polasek O, Talmud PJ, Nyberg F, Kuh D, Laan M, Hveem K, Palmer LJ, van der Schouw YT, Casas JP, Mohlke KL, Vineis P, Raitakari O, Ganesh SK, Wong TY, Tai ES, Cooper RS, Laakso M, Rao DC, Harris TB, Morris RW, Dominiczak AF, Kivimäki M, Marmot MG, Miki T, Saleheen D, Chandak GR, Coresh J, Navis G, Salomaa V, Han BG, Zhu X, Kooner JS, Melander O, Ridker PM, Bandinelli S, Gyllenstein UB, Wright AF, Wilson JF, Ferrucci L, Farrall M, Tuomilehto J, Pramstaller PP, Elosua R, Soranzo N, Sijbrands EJ, Altshuler D, Loos RJ, Shuldiner AR, Gieger C, Meneton P, Uitterlinden AG, Wareham NJ, Gudnason V, Rotter JJ, Rettig R, Uda M, Strachan DP, Witteman JC, Hartikainen AL, Beckmann JS, Boerwinkle E, Vasan RS, Boehnke M, Larson MG, Järvelin MR, Psaty BM, Abecasis GR, Chakravarti A, Elliott P, van Duijn CM, Newton-Cheh C, Levy D, Caulfield MJ, Johnson T. Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. *Nature* 2011; **478**: 103-109 [PMID: 21909115 DOI: 10.1038/nature10405]
- 96 **Wain LV**, Verwoert GC, O'Reilly PF, Shi G, Johnson T, Johnson AD, Bochud M, Rice KM, Henneman P, Smith AV, Ehret GB, Amin N, Larson MG, Mooser V, Hadley D, Dörr M, Bis JC, Aspelund T, Esko T, Janssens AC, Zhao JH, Heath S, Laan M, Fu J, Pistis G, Luan J, Arora P, Lucas G, Pirastu N, Pichler I, Jackson AU, Webster RJ, Zhang F, Peden JF, Schmidt H, Tanaka T, Campbell H, Igl W, Milaneschi Y, Hottenga JJ, Vitart V, Chasman DI, Trompet S, Bragg-Gresham JL, Alizadeh BZ, Chambers JC, Guo X, Lehtimäki T, Kühnel B, Lopez LM, Polašek O, Boban M, Nelson CP, Morrison AC, Pihur V, Ganesh SK, Hofman A, Kundu S, Mattace-Raso FU, Rivadeneira F, Sijbrands EJ, Uitterlinden AG, Hwang SJ, Vasan RS, Wang TJ, Bergmann S, Vollenweider P, Waebler G, Laitinen J, Pouta A, Zitting P, McArdle WL, Kroemer HK, Völker U, Völzke H, Glazer NL, Taylor KD, Harris TB, Alavere H, Haller T, Keis A, Tammesoo ML, Aulchenko Y, Barroso I, Khaw KT, Galan P, Hercberg S, Lathrop M, Eyheramendy S, Org E, Söber S, Lu X, Nolte IM, Penninx BW, Corre T, Masciullo C, Sala C, Groop L, Voight BF, Melander O, O'Donnell CJ, Salomaa V, d'Adamo AP, Fabretto A, Faletra F, Ulivi S, Del Greco F, Facheris M, Collins FS, Bergman RN, Beilby JP, Hung J, Musk AW, Mangino M, Shin SY, Soranzo N, Watkins H, Goel A, Hamsten A, Gider P, Loitfelder M, Zeginigg M, Hernandez D, Najjar SS, Navarro P, Wild SH, Corsi AM, Singleton A, de Geus EJ, Willemsen G, Parker AN, Rose LM, Buckley B, Stott D, Orru M, Uda M, van der Klauw MM, Zhang W, Li X, Scott J, Chen YD, Burke GL, Kähönen M, Viikari J, Döring A, Meitinger T, Davies G, Starr JM, Emilsson V, Plump A, Lindeman JH, Hoen PA, König IR, Felix JF, Clarke R, Hopewell JC, Ongen H, Breteler M, DeBette S, Destefano AL, Fornage M, Mitchell GF, Smith NL, Holm H, Stefansson K, Thorleifsson G, Thorsteinsdottir U, Samani NJ, Preuss M, Rudan I, Hayward C, Deary IJ, Wichmann HE, Raitakari OT, Palmas W, Kooner JS, Stolk RP, Jukema JW, Wright AF, Boomsma DI, Bandinelli S, Gyllenstein UB, Wilson JF, Ferrucci L, Schmidt R, Farrall M, Spector TD, Palmer LJ, Tuomilehto J, Pfeuffer A, Gasparini P, Siscovick D, Altshuler D, Loos RJ, Toniolo D, Snieder H, Gieger C, Meneton P, Wareham NJ, Oostra BA, Metspalu A, Launer L, Rettig R, Strachan DP, Beckmann JS, Witteman JC, Erdmann J, van Dijk KW, Boerwinkle E, Boehnke M, Ridker PM, Jarvelin MR, Chakravarti A, Abecasis GR, Gudnason V, Newton-Cheh C, Levy D, Munroe PB, Psaty BM, Caulfield MJ, Rao DC, Tobin MD, Elliott P, van Duijn CM. Genome-wide association study identifies six new loci influencing pulse pressure and mean arterial pressure. *Nat Genet* 2011; **43**: 1005-1011 [PMID: 21909110 DOI: 10.1038/ng.922]
- 97 **Jiang W**, Sordella R, Chen GC, Hakre S, Roy AL, Settleman J. An FF domain-dependent protein interaction mediates a signaling pathway for growth factor-induced gene expression. *Mol Cell* 2005; **17**: 23-35 [PMID: 15629714 DOI: 10.1016/j.molcel.2004.11.024]
- 98 **Kimura K**, Eguchi S. Angiotensin II type-1 receptor regulates RhoA and Rho-kinase/ROCK activation via multiple mechanisms. Focus on "Angiotensin II induces RhoA activation through SHP2-dependent dephosphorylation of the RhoGAP p190A in vascular smooth muscle cells". *Am J Physiol Cell Physiol* 2009; **297**: C1059-C1061 [PMID: 19741194 DOI: 10.1152/ajpcell.00399.2009]
- 99 **Haskell MD**, Nickles AL, Agati JM, Su L, Dukes BD, Parsons SJ. Phosphorylation of p190 on Tyr1105 by c-Src is necessary but not sufficient for EGF-induced actin disassembly in C3H10T1/2 fibroblasts. *J Cell Sci* 2001; **114**: 1699-1708 [PMID: 11309200]
- 100 **Pullikuth AK**, Catling AD. Extracellular signal-regulated kinase promotes Rho-dependent focal adhesion formation by suppressing p190A RhoGAP. *Mol Cell Biol* 2010; **30**: 3233-3248 [PMID:



- 20439493 DOI: 10.1128/MCB.01178-09]
- 101 **Lévay M**, Bartos B, Ligeti E. p190RhoGAP has cellular RacGAP activity regulated by a polybasic region. *Cell Signal* 2013; **25**: 1388-1394 [PMID: 23499677 DOI: 10.1016/j.cellsig.2013.03.004]
- 102 **Gong MC**, Gorenne I, Read P, Jia T, Nakamoto RK, Somlyo AV, Somlyo AP. Regulation by GDI of RhoA/Rho-kinase-induced Ca<sup>2+</sup> sensitization of smooth muscle myosin II. *Am J Physiol Cell Physiol* 2001; **281**: C257-C269 [PMID: 11401849]
- 103 **Shibata S**, Mu S, Kawarazaki H, Muraoka K, Ishizawa K, Yoshida S, Kawarazaki W, Takeuchi M, Ayuzawa N, Miyoshi J, Takai Y, Ishikawa A, Shimosawa T, Ando K, Nagase M, Fujita T. Rac1 GTPase in rodent kidneys is essential for salt-sensitive hypertension via a mineralocorticoid receptor-dependent pathway. *J Clin Invest* 2011; **121**: 3233-3243 [PMID: 21765214 DOI: 10.1172/JCI41324]
- 104 **Li J**, Rohaila S, Gelber N, Rutka J, Sabah N, Gladstone RA, Wei C, Hu P, Kharbanda RK, Redington AN. MicroRNA-144 is a circulating effector of remote ischemic preconditioning. *Basic Res Cardiol* 2014; **109**: 423 [PMID: 25060662 DOI: 10.1007/s00395-014-0423-z]
- 105 **Artamonov MV**, Jin L, Franke AS, Momotani K, Ho R, Dong XR, Majesky MW, Somlyo AV. Signaling pathways that control rho kinase activity maintain the embryonic epicardial progenitor state. *J Biol Chem* 2015; **290**: 10353-10367 [PMID: 25733666 DOI: 10.1074/jbc.M114.613190]
- 106 **Chen W**, Chu Y, Zhu D, Yan C, Liu J, Ji K, Gao P. Perivascular gene transfer of dominant-negative N19RhoA attenuates neointimal formation via inhibition of TGF-beta1-Smad2 signaling in rats after carotid artery balloon injury. *Biochem Biophys Res Commun* 2009; **389**: 217-223 [PMID: 19706289 DOI: 10.1016/j.bbrc.2009.08.104]
- 107 **Boyden LM**, Choi M, Choate KA, Nelson-Williams CJ, Farhi A, Toka HR, Tikhonova IR, Bjornson R, Mane SM, Colussi G, Lebel M, Gordon RD, Semmekrot BA, Poujol A, Välimäki MJ, De Ferrari ME, Sanjad SA, Gutkin M, Karet FE, Tucci JR, Stockigt JR, Keppler-Noreuil KM, Porter CC, Anand SK, Whiteford ML, Davis ID, Dewar SB, Bettinelli A, Fadowski JJ, Belsha CW, Hunley TE, Nelson RD, Trachtman H, Cole TR, Pinski M, Bockenhauer D, Shenoy M, Vaidyanathan P, Foreman JW, Rasoulpour M, Thameem F, Al-Shahrouri HZ, Radhakrishnan J, Gharavi AG, Goilav B, Lifton RP. Mutations in kelch-like 3 and cullin 3 cause hypertension and electrolyte abnormalities. *Nature* 2012; **482**: 98-102 [PMID: 22266938 DOI: 10.1038/nature10814]
- 108 **Ibeawuchi SR**, Agbor LN, Quelle FW, Sigmund CD. Hypertension-causing Mutations in Cullin3 Protein Impair RhoA Protein Ubiquitination and Augment the Association with Substrate Adaptors. *J Biol Chem* 2015; **290**: 19208-19217 [PMID: 26100637 DOI: 10.1074/jbc.M115.645358]
- 109 **Croce LE**, Hilder TL, Sciaky N, Johnson GL. Cerebral cavernous malformation 2 protein promotes smad ubiquitin regulatory factor 1-mediated RhoA degradation in endothelial cells. *J Biol Chem* 2009; **284**: 13301-13305 [PMID: 19318350 DOI: 10.1074/jbc.C900009200]
- 110 **Richardson BT**, Dibble CF, Borikova AL, Johnson GL. Cerebral cavernous malformation is a vascular disease associated with activated RhoA signaling. *Biol Chem* 2013; **394**: 35-42 [PMID: 23096573 DOI: 10.1515/hsz-2012-0243]
- 111 **Aghajanian A**, Wittchen ES, Campbell SL, Burrige K. Direct activation of RhoA by reactive oxygen species requires a redox-sensitive motif. *PLoS One* 2009; **4**: e8045 [PMID: 19956681 DOI: 10.1371/journal.pone.0008045]
- 112 **Zuckerbraun BS**, Stoyanovsky DA, Sengupta R, Shapiro RA, Ozanich BA, Rao J, Barbato JE, Tzeng E. Nitric oxide-induced inhibition of smooth muscle cell proliferation involves S-nitrosation and inactivation of RhoA. *Am J Physiol Cell Physiol* 2007; **292**: C824-C831 [PMID: 16914531 DOI: 10.1152/ajpcell.00592.2005]
- 113 **Sandu OA**, Ito M, Begum N. Selected contribution: insulin utilizes NO/cGMP pathway to activate myosin phosphatase via Rho inhibition in vascular smooth muscle. *J Appl Physiol* (1985) 2001; **91**: 1475-1482 [PMID: 11509551]
- 114 **Sauzeau V**, Le Jeune H, Cario-Toumaniantz C, Smolenski A, Lohmann SM, Bertoglio J, Chardin P, Pacaud P, Loirand G. Cyclic GMP-dependent protein kinase signaling pathway inhibits RhoA-induced Ca<sup>2+</sup> sensitization of contraction in vascular smooth muscle. *J Biol Chem* 2000; **275**: 21722-21729 [PMID: 10783386 DOI: 10.1074/jbc.M000753200]
- 115 **Carretero OA**, Oparil S. Essential hypertension. Part I: definition and etiology. *Circulation* 2000; **101**: 329-335 [PMID: 10645931 DOI: 10.1161/01.CIR.101.3.329]
- 116 **Mozaffarian D**, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M, de Ferranti S, Després JP, Fullerton HJ, Howard VJ, Huffman MD, Judd SE, Kissela BM, Lackland DT, Lichtman JH, Lisabeth LD, Liu S, Mackey RH, Matchar DB, McGuire DK, Mohler ER, Moy CS, Muntner P, Mussolino ME, Nasir K, Neumar RW, Nichol G, Palaniappan L, Pandey DK, Reeves MJ, Rodriguez CJ, Sorlie PD, Stein J, Towfighi A, Turan TN, Virani SS, Willey JZ, Woo D, Yeh RW, Turner MB. Heart disease and stroke statistics--2015 update: a report from the American Heart Association. *Circulation* 2015; **131**: e29-322 [PMID: 25520374 DOI: 10.1161/CIR.000000000000152]
- 117 **Padmanabhan S**, Caulfield M, Dominiczak AF. Genetic and molecular aspects of hypertension. *Circ Res* 2015; **116**: 937-959 [PMID: 25767282 DOI: 10.1161/CIRCRESAHA.116.303647]
- 118 **Pelham CJ**, Ketsawatsomkron P, Groh S, Grobe JL, de Lange WJ, Ibeawuchi SR, Keen HL, Weatherford ET, Faraci FM, Sigmund CD. Cullin-3 regulates vascular smooth muscle function and arterial blood pressure via PPARγ and RhoA/Rho-kinase. *Cell Metab* 2012; **16**: 462-472 [PMID: 23040068 DOI: 10.1016/j.cmet.2012.08.011]
- 119 **Seasholtz TM**, Wessel J, Rao F, Rana BK, Khandrika S, Kennedy BP, Lillie EO, Ziegler MG, Smith DW, Schork NJ, Brown JH, O'Connor DT. Rho kinase polymorphism influences blood pressure and systemic vascular resistance in human twins: role of heredity. *Hypertension* 2006; **47**: 937-947 [PMID: 16585408 DOI: 10.1161/01.HYP.0000217364.45622.f0]
- 120 **Rankinen T**, Church T, Rice T, Markward N, Blair SN, Bouchard C. A major haplotype block at the rho-associated kinase 2 locus is associated with a lower risk of hypertension in a recessive manner: the HYPGENE study. *Hypertens Res* 2008; **31**: 1651-1657 [PMID: 18971541 DOI: 10.1291/hypres.31.1651]
- 121 **Liu L**, Cao Y, Cui G, Li Z, Sun J, Zhang L, Chen C, Wang Y, Wang P, Ding H, Wang DW. Association analysis of polymorphisms in ROCK2 with cardiovascular disease in a Chinese population. *PLoS One* 2013; **8**: e53905 [PMID: 23326532 DOI: 10.1371/journal.pone.0053905]
- 122 **Hashimoto J**, Ito S. Some mechanical aspects of arterial aging: physiological overview based on pulse wave analysis. *Ther Adv Cardiovasc Dis* 2009; **3**: 367-378 [PMID: 19574288 DOI: 10.1177/1753944709338942]
- 123 **Williams B**. Evaluating interventions to reduce central aortic pressure, arterial stiffness and morbidity--mortality. *J Hypertens* 2012; **30** Suppl: S13-S18 [PMID: 23124100 DOI: 10.1097/HJH.0b013e328353e523]
- 124 **Luft FC**. Molecular mechanisms of arterial stiffness: new insights. *J Am Soc Hypertens* 2012; **6**: 436-438 [PMID: 23199674 DOI: 10.1016/j.jash.2012.10.004]
- 125 **Moody WE**, Edwards NC, Chue CD, Ferro CJ, Townend JN. Arterial disease in chronic kidney disease. *Heart* 2013; **99**: 365-372 [PMID: 23118349 DOI: 10.1136/heartjnl-2012-302818]
- 126 **Safar ME**, Nilsson PM. Pulsatile hemodynamics and cardiovascular risk factors in very old patients: background, sex aspects and implications. *J Hypertens* 2013; **31**: 848-857 [PMID: 23449020 DOI: 10.1097/HJH.0b013e328353e5b9]
- 127 **Galmiche G**, Labat C, Mericskay M, Aissa KA, Blanc J, Retailleau K, Bourhim M, Coletti D, Loufrani L, Gao-Li J, Feil R, Challande P, Henrion D, Decaux JF, Regnault V, Lacolley P, Li Z. Inactivation of serum response factor contributes to decrease vascular muscular tone and arterial stiffness in mice. *Circ Res* 2013; **112**: 1035-1045 [PMID: 23426017 DOI: 10.1161/CIRCRESAHA.113.301076]
- 128 **Qiu H**, Zhu Y, Sun Z, Trzeciakowski JP, Gansner M, Depre C, Resuello RR, Natividad FF, Hunter WC, Genin GM, Elson EL, Vatner DE, Meininger GA, Vatner SF. Short communication:



- vascular smooth muscle cell stiffness as a mechanism for increased aortic stiffness with aging. *Circ Res* 2010; **107**: 615-619 [PMID: 20634486 DOI: 10.1161/CIRCRESAHA.110.221846]
- 129 **Liao YC**, Liu PY, Lin HF, Lin WY, Liao JK, Juo SH. Two functional polymorphisms of ROCK2 enhance arterial stiffening through inhibiting its activity and expression. *J Mol Cell Cardiol* 2015; **79**: 180-186 [PMID: 25481646 DOI: 10.1016/j.yjmcc.2014.11.023]
  - 130 **Mendelson K**, Evans T, Hla T. Sphingosine 1-phosphate signalling. *Development* 2014; **141**: 5-9 [PMID: 24346695 DOI: 10.1242/dev.094805]
  - 131 **Fenger M**, Linneberg A, Jeppesen J. Network-based analysis of the sphingolipid metabolism in hypertension. *Front Genet* 2015; **6**: 84 [PMID: 25788903 DOI: 10.3389/fgene.2015.00084]
  - 132 **Fenger M**, Linneberg A, Jørgensen T, Madsbad S, Søbye K, Eugen-Olsen J, Jeppesen J. Genetics of the ceramide/sphingosine-1-phosphate rheostat in blood pressure regulation and hypertension. *BMC Genet* 2011; **12**: 44 [PMID: 21569466 DOI: 10.1186/1471-2156-12-44]
  - 133 **Ren XD**, Kiosses WB, Schwartz MA. Regulation of the small GTP-binding protein Rho by cell adhesion and the cytoskeleton. *EMBO J* 1999; **18**: 578-585 [PMID: 9927417 DOI: 10.1093/emboj/18.3.578]
  - 134 **Levy D**, Ehret GB, Rice K, Verwoert GC, Launer LJ, Dehghan A, Glazer NL, Morrison AC, Johnson AD, Aspelund T, Aulchenko Y, Lumley T, Köttgen A, Vasan RS, Rivadeneira F, Eiriksdottir G, Guo X, Arking DE, Mitchell GF, Mattace-Raso FU, Smith AV, Taylor K, Scharpf RB, Hwang SJ, Sijbrands EJ, Bis J, Harris TB, Ganesh SK, O'Donnell CJ, Hofman A, Rotter JI, Coresh J, Benjamin EJ, Uitterlinden AG, Heiss G, Fox CS, Witteman JC, Boerwinkle E, Wang TJ, Gudnason V, Larson MG, Chakravarti A, Psaty BM, van Duijn CM. Genome-wide association study of blood pressure and hypertension. *Nat Genet* 2009; **41**: 677-687 [PMID: 19430479 DOI: 10.1038/ng.384]
  - 135 **Lin Y**, Lai X, Chen B, Xu Y, Huang B, Chen Z, Zhu S, Yao J, Jiang Q, Huang H, Wen J, Chen G. Genetic variations in CYP17A1, CACNB2 and PLEKHA7 are associated with blood pressure and/or hypertension in She ethnic minority of China. *Atherosclerosis* 2011; **219**: 709-714 [PMID: 21963141 DOI: 10.1016/j.atherosclerosis.2011.09.006]
  - 136 **Citi S**, Pulimeno P, Paschoud S. Cingulin, paracingulin, and PLEKHA7: signaling and cytoskeletal adaptors at the apical junctional complex. *Ann N Y Acad Sci* 2012; **1257**: 125-132 [PMID: 22671598 DOI: 10.1111/j.1749-6632.2012.06506.x]
  - 137 **Endres BT**, Priestley JR, Palygin O, Flister MJ, Hoffman MJ, Weinberg BD, Grzybowski M, Lombard JH, Staruschenko A, Moreno C, Jacob HJ, Geurts AM. Mutation of Plekha7 attenuates salt-sensitive hypertension in the rat. *Proc Natl Acad Sci USA* 2014; **111**: 12817-12822 [PMID: 25136115 DOI: 10.1073/pnas.1410745111]
  - 138 **GTE Consortium**. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science* 2015; **348**: 648-660 [PMID: 25954001 DOI: 10.1126/science.1262110]
  - 139 **Carbone ML**, Brégeon J, Devos N, Chadeuf G, Blanchard A, Azizi M, Pacaud P, Jeunemaître X, Loirand G. Angiotensin II activates the RhoA exchange factor Arhgef1 in humans. *Hypertension* 2015; **65**: 1273-1278 [PMID: 25870189 DOI: 10.1161/HYPERTENSIONAHA.114.05065]
  - 140 **Kanaki AI**, Sarafidis PA, Georgianos PI, Kanavos K, Tziolas IM, Zebekakis PE, Lasaridis AN. Effects of low-dose atorvastatin on arterial stiffness and central aortic pressure augmentation in patients with hypertension and hypercholesterolemia. *Am J Hypertens* 2013; **26**: 608-616 [PMID: 23449607 DOI: 10.1093/ajh/hps098]
  - 141 **Brandes RP**. Statin-mediated inhibition of Rho: only to get more NO? *Circ Res* 2005; **96**: 927-929 [PMID: 15890979 DOI: 10.1161/01.RES.0000168040.70096.2a]
  - 142 **Olson MF**. Applications for ROCK kinase inhibition. *Curr Opin Cell Biol* 2008; **20**: 242-248 [PMID: 18282695 DOI: 10.1016/j.ceb.2008.01.002]
  - 143 **Davies SP**, Reddy H, Caivano M, Cohen P. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem J* 2000; **351**: 95-105 [PMID: 10998351 DOI: 10.1042/bj3510095]
  - 144 **Liao JK**, Seto M, Noma K. Rho kinase (ROCK) inhibitors. *J Cardiovasc Pharmacol* 2007; **50**: 17-24 [PMID: 17666911 DOI: 10.1097/FJC.0b013e318070d1bd]
  - 145 **Asano M**, Nomura Y. Comparison of inhibitory effects of Y-27632, a Rho kinase inhibitor, in strips of small and large mesenteric arteries from spontaneously hypertensive and normotensive Wistar-Kyoto rats. *Hypertens Res* 2003; **26**: 97-106 [PMID: 12661918 DOI: 10.1291/hypres.26.97]
  - 146 **Uehata M**, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature* 1997; **389**: 990-994 [PMID: 9353125 DOI: 10.1038/40187]
  - 147 **Surma M**, Wei L, Shi J. Rho kinase as a therapeutic target in cardiovascular disease. *Future Cardiol* 2011; **7**: 657-671 [PMID: 21929346 DOI: 10.2217/fca.11.51]
  - 148 **Satoh K**, Fukumoto Y, Shimokawa H. Rho-kinase: important new therapeutic target in cardiovascular diseases. *Am J Physiol Heart Circ Physiol* 2011; **301**: H287-H296 [PMID: 21622831 DOI: 10.1152/ajpheart.00327.2011]
  - 149 **Dhaliwal JS**, Badejo AM, Casey DB, Murthy SN, Kadowitz PJ. Analysis of pulmonary vasodilator responses to SB-772077-B [4-(7-((3-amino-1-pyrrolidinyl)carbonyl)-1-ethyl-1H-imidazo(4,5-c)pyridin-2-yl)-1,2,5-oxadiazol-3-amine], a novel aminofurazan-based Rho kinase inhibitor. *J Pharmacol Exp Ther* 2009; **330**: 334-341 [PMID: 19369577 DOI: 10.1124/jpet.109.151449]
  - 150 **Doe C**, Bentley R, Behm DJ, Lafferty R, Stavenger R, Jung D, Bamford M, Panchal T, Grygielko E, Wright LL, Smith GK, Chen Z, Webb C, Khandekar S, Yi T, Kirkpatrick R, Dul E, Jolivet L, Marino JP, Willette R, Lee D, Hu E. Novel Rho kinase inhibitors with anti-inflammatory and vasodilatory activities. *J Pharmacol Exp Ther* 2007; **320**: 89-98 [PMID: 17018693 DOI: 10.1124/jpet.106.110635]
  - 151 **Boerma M**, Fu Q, Wang J, Loose DS, Bartolozzi A, Ellis JL, McGonigle S, Paradise E, Sweetnam P, Fink LM, Vozenin-Brotons MC, Hauer-Jensen M. Comparative gene expression profiling in three primary human cell lines after treatment with a novel inhibitor of Rho kinase or atorvastatin. *Blood Coagul Fibrinolysis* 2008; **19**: 709-718 [PMID: 18832915 DOI: 10.1097/MBC.0b013e32830b2891]
  - 152 **Löhn M**, Plettenburg O, Kannt A, Kohlmann M, Hofmeister A, Kadereit D, Monecke P, Schiffer A, Schulte A, Ruetten H, Ivashchenko Y. End-organ protection in hypertension by the novel and selective Rho-kinase inhibitor, SAR407899. *World J Cardiol* 2015; **7**: 31-42 [PMID: 25632317 DOI: 10.4330/wjc.v7.i1.31]
  - 153 **Löhn M**, Plettenburg O, Ivashchenko Y, Kannt A, Hofmeister A, Kadereit D, Schaefer M, Linz W, Kohlmann M, Herbert JM, Janiak P, O'Connor SE, Ruetten H. Pharmacological characterization of SAR407899, a novel rho-kinase inhibitor. *Hypertension* 2009; **54**: 676-683 [PMID: 19597037 DOI: 10.1161/HYPERTENSIONAHA.109.134353]
  - 154 **Carretero OA**, Oparil S. Essential hypertension : part II: treatment. *Circulation* 2000; **101**: 446-453 [PMID: 10653838 DOI: 10.1161/01.CIR.101.4.446]
  - 155 **Epstein M**, Bakris G. Newer approaches to antihypertensive therapy. Use of fixed-dose combination therapy. *Arch Intern Med* 1996; **156**: 1969-1978 [PMID: 8823150 DOI: 10.1001/archinte.199.6.00440160081011]
  - 156 **He F**, Luo J, Zhang Z, Luo Z, Fan L, He Y, Wen J, Zhu D, Gao J, Wang Y, Qian Y, Zhou H, Chen X, Zhang W. The RGS2 (-391, C > G) genetic variation correlates to antihypertensive drug responses in Chinese patients with essential hypertension. *PLoS One* 2015; **10**: e0121483 [PMID: 25849301 DOI: 10.1371/journal.pone.0121483]
  - 157 **Vandell AG**, Lobmeyer MT, Gawronski BE, Langae TY, Gong

Y, Gums JG, Beitelshes AL, Turner ST, Chapman AB, Cooper-DeHoff RM, Bailey KR, Boerwinkle E, Pepine CJ, Liggett SB, Johnson JA. G protein receptor kinase 4 polymorphisms:  $\beta$ -blocker

pharmacogenetics and treatment-related outcomes in hypertension. *Hypertension* 2012; **60**: 957-964 [PMID: 22949529 DOI: 10.1161/HYPERTENSIONAHA.112.198721]

**P- Reviewer:** Park JB, Stourmaras C, Wang J **S- Editor:** Wang JL  
**L- Editor:** A **E- Editor:** Wu HL



## Place of baroreceptor activation therapy in the treatment of resistant hypertension

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**Supported by** FORICA (The Foundation for Advanced Research in Hypertension and Cardiovascular diseases); the University of Padua (to GPR); HORIZON 2020; and COST-ADMIRE.

**Conflict-of-interest statement:** Authors declare no conflict of interests for this article.

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Received: September 1, 2015  
 Peer-review started: September 8, 2015  
 First decision: September 29, 2015  
 Revised: October 7, 2015  
 Accepted: December 9, 2015  
 Article in press: December 10, 2015  
 Published online: February 23, 2016

### Abstract

This mini review describes the development of the therapeutic concept of baroreceptor stimulation over the

last fifty years alongside the more recent introduction of it for the treatment of drug - resistant hypertension. The pros and cons of this strategy of treatment over renal sympathetic denervation are also discussed in the light of the results of the studies done in the last decade.

**Key words:** Resistant hypertension; Treatment; Baroreceptor stimulation; Renal denervation

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**Core tip:** We herein describe the development of the therapeutic concept of baroreceptor stimulation for the treatment of drug-resistant hypertension. The ups and downs of this treatment strategy are discussed in the light of the results of the studies done in the last decade.

Rossi GP, Azzolini M. Place of baroreceptor activation therapy in the treatment of resistant hypertension. *World J Hypertens* 2016; 6(1): 36-40 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/36.htm> DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.36>

### INTRODUCTION

Baroreceptor activation blunts sympathetic activity and enhances vagal tone, thus rebalancing the neural output to the heart, the vessels, and the kidney in favour of the latter. This treatment strategy, currently defined as baroreceptor activation therapy (BAT), was originally conceived 50 years ago with the ultimate aim of lowering blood pressure (BP): Proof-of-concept studies were first published by Bilgutay *et al*<sup>[1]</sup> and then by Torresani *et al*<sup>[2]</sup>. They could nicely document the achievement of a marked reduction in BP in patients who were resistant to the (few) drugs available at

**Table 1** Therapeutic strategies in patients with resistant hypertension

Recommendations	Class of recommendation <sup>1</sup>	Level of evidence <sup>2</sup>
In resistant hypertensive patients it is recommended that physicians check whether the drugs included in the existing multiple drug regimen have any BP lowering effect, and withdraw them if their effect is absent or minimal	I	C
Mineralocorticoid receptor antagonists, amiloride, and the alpha-1-blocker doxazosin should be considered, if no contraindications exist	IIa	B
In case of ineffectiveness of drug treatment invasive procedures such as renal denervation and baroreceptor stimulation may be considered	IIb	C
Until more evidence is available on the long-term efficacy and safety of renal denervation and baroreceptor stimulation, it is recommended that these procedures remain in the hands of experienced operators and diagnosis and follow-up restricted to hypertension centers	I	C
It is recommended that the invasive approaches are considered only for truly resistant hypertensive patients, with clinic values $\geq 160$ mmHg SBP or $\geq 110$ mmHg DBP and with BP elevation confirmed by ABPM	I	C

Class of recommendation<sup>1</sup>: Class I: Evidence and/or general agreement that a given treatment or procedure is beneficial, useful, effective; Class II: Conflicting evidence and/or a divergence of opinion about the usefulness/efficacy of the given treatment or procedure; Class IIa: Weight of evidence/opinion is in favor of usefulness/efficacy; Class IIb: Usefulness/efficacy is less well established by evidence/opinion. Level of evidence<sup>2</sup>: Level of evidence B: Data derived from a single randomized clinical trial or large non-randomized studies; Level of evidence C: Consensus of opinion of the experts and/or small studies, retrospective studies, registries. ABPM: Ambulatory blood pressure monitoring; BP: Blood pressure; DBP: Diastolic blood pressure; SBP: Systolic blood pressure.

that time. Unfortunately the delivering of this therapy involved an invasive process and, moreover, stimulation was achieved by means of an external device. Accordingly, it turned out to be unpractical out of the hospital. For these reasons, alongside the development of multiple effective BP-lowering medications, BAT was rapidly abandoned.

It took more than forty years for this simple physiological principle to find again its way to clinical application, thanks to the development of implantable devices. The latter were shown to unequivocally lower BP in the DEBUT study<sup>[3]</sup>. Furthermore, in the Heart Rate Variability sub-study, they were also reported to improve the sympatho-vagal balance<sup>[3]</sup>. At that time it became readily evident that the BAT not only lowered BP and sympathetic nerve activity to the muscles<sup>[4]</sup>, but was also effective in regressing left ventricular (LV) hypertrophy, in improving LV geometry, and in decreasing BP all around the clock<sup>[3]</sup>.

In June 2010, when the first international meeting on Resistant Hypertension was held in Padua, Italy<sup>[5]</sup>, it seemed, therefore, that BAT was emerging as the “front runner” in the therapeutic armamentarium for the then resurfacing problem of resistant hypertension.

### **Strong competition in the therapeutic armamentarium**

At about the same time the publication of the first study on renal denervation<sup>[6]</sup> and, immediately after, of the Simplicity HTN-1<sup>[7]</sup> introduced another strong competitor for BAT in the race for the best treatment of drug-resistant hypertension: Percutaneous renal denervation. This treatment modality, which is undoubtedly less invasive than BAT, was shown to be effective in decreasing BP in carefully selected cases.

In June 2013, based on the evidence provided in the Simplicity HTN-2 trial that was published thereafter<sup>[8]</sup>, the ESC/ESH Guidelines released statements concerning

the use of BAT or renal denervation (Table 1)<sup>[9]</sup>. With a class IIb level of recommendation C these guidelines affirmed that invasive procedures, such as denervation and baroreceptor stimulation, “may be considered” in case of ineffectiveness of drug treatment<sup>[9]</sup>. In other words, it was explicitly acknowledged that the evidence supporting usefulness/efficacy of these therapies was not well established by evidence/opinion (Class IIb), and that the level of evidence depended on consensus of experts, and/or small studies, retrospective studies and/or registries (Level of evidence C).

Of further importance, with a class I level C the ESC/ESH guidelines recommended that “these procedures remain in the hands of experienced operators and diagnosis and follow-up restricted to hypertension centers”. It was further emphasized that these invasive approaches were to be “considered only for truly resistant hypertension”.

In March 2014, the scenario changed again with the publication of the first randomized single-blinded, sham-controlled study on renal denervation, the Simplicity HTN-3. While conclusively proving the safety of renal denervation in experienced hands, this study failed on one of its primary endpoints, *e.g.*, the demonstration that renal denervation was more effective than a “sham” procedure in effectively lowering BP<sup>[10]</sup>. After the publication of this study, the results of a smaller multi-center French study, the DENERHTN<sup>[11]</sup>, added further fuel to the debate. According to this study, when given on top of a carefully planned stepped pharmacologic treatment, renal denervation provided almost identical BP values as placebo at 6 mo follow-up, although the fall of BP, both systolic and diastolic, was greater in the renal denervation arm. However, this was apparently only because the patients in this group had slightly higher baseline values, suggesting the possibility of a “regression toward the mean” effect<sup>[11]</sup>, and raising



questions about the effectiveness of renal denervation in lowering BP. According to the plot depicting the individual BP changes, some patients did show a marked BP reduction and others showed no BP fall whatsoever.

At about the same time, the Prague study<sup>[12]</sup> also found no evidence for superiority of renal denervation over medical therapy. In this study the patients were randomized to either a multidrug treatment comprising the mineralocorticoid antagonist spironolactone, or to renal denervation. The BP values at 6 mo follow-up were similar in the renal denervation and medical treatment arm. While the long-term results of this study are still awaited, it seems fair to say that it remains to be conclusively proven that renal denervation entails an all-round treatment for all patients with truly resistant hypertension. A head-to-head comparison of BAT and renal denervation is shown in Table 2.

### The challenges of proving effectiveness of treatments for resistant hypertension

There is no question that the diagnosis of true drug-resistant hypertension is a difficult one, particularly out of tertiary referral centers, *e.g.*, centers where all diagnostic tools cannot be available<sup>[12]</sup>. Of note, in the patients initially selected at the French specialized centers that participated in the DENEHRT study, about 50% were excluded because of secondary HT, a rate that is remarkably higher than commonly perceived. This observation suggests that out of the hypertension referral centers under detection of secondary forms of hypertension is a diffuse phenomenon, which recognizes several causes. Nevertheless, the high exclusion rate due to secondary hypertension in the French Study indicates that, in keeping with the guidelines, any invasive procedures for the treatment of HT should be restricted to the centers that are competent in reliably rule out secondary forms of HT. This is not an easy task in patients who are resistant to drug treatment for a very simple reason: these patients are, by definition, on multiple drugs; moreover, they can carry multiple conditions that affect renin-angiotensin-aldosterone system. Thereby, the measurements of renin and aldosterone, the two key biomarkers for the identification of the most common forms of secondary HT, *e.g.*, primary aldosteronism and renovascular hypertension, can be markedly biased. In most cases this problem can hamper the diagnosis.

### The current place of BAT

Nowadays BAT devices allow switching on and off the implanted device; therefore, it provides the ideal within-patient design for assessment of the effect on BP. By such strategy the BP-lowering efficacy of BAT has been proven beyond any doubts<sup>[13]</sup>. Moreover, these features also allowed the demonstration that the effects of BAT are reversible and reproducible over time. Of much importance for the patients with resistant hypertension, who had a BP that was not controlled after renal

**Table 2 Similarities and differences between renal denervation and baroreceptor activation therapy**

Features	Renal denervation	BAT
SNS	↓	↓
Invasive	Yes	Yes
Safe	Yes <sup>1</sup>	Yes <sup>1</sup>
BP short-term	↓	↓
BP long-term	?	?
Side	Bilateral	Monolateral
Evidence of success	Delay	Immediate
Reversible	No	Yes
Heart rate effect	↓	↓↓
Metabolic effect	Yes	?
Need for follow-up	Yes	Yes
Logistics <sup>2</sup>	+	+++
Costs	+	+++

SNS: Sympathetic nervous system; BP: Blood pressure; BAT: Baroreceptor activation therapy; <sup>1</sup>Complications rate < 3%; <sup>2</sup>Interventional Radiology *vs* Vascular surgery.

denervation<sup>[14]</sup>, BAT was shown to work well, albeit only in small subsets of patients.

Finally, BAT can be used in patients who have contraindications to renal denervation because of unsuitable renal anatomy, previous renal endovascular treatment and/or impaired glomerular filtration rate. These advantages have to be weighed against its invasiveness, which however, has been diminished by the development of the smaller 2<sup>nd</sup> generation devices, and, more importantly, by the demonstration that unilateral BAT is not inferior to bilateral BAT at least for lowering BP<sup>[13]</sup>. Decreased invasiveness of the implantation, alongside improved experience of the surgeon and the medical team, will likely result into fewer complications and shorter hospital stay, thus decreasing the costs and increasing the acceptance of the BAT.

### Technical aspects: Implantation technique

The system for delivering BAT consists of a carotid sinus lead and a pulse generator. Implantation of the pulse generator is generally performed by a vascular surgeon experienced in a subcutaneous infra-clavicular chest wall pocket, in the fashion of a pacemaker. Electrode implantation is also performed at the same time by a vascular surgeon experienced in carotid artery revascularization by surgical exposure of the carotid sinus through a transverse cervicotomy over the carotid bifurcation. The sinus region is then mapped by temporarily placing the electrode in various locations and applying electrical stimulation to determine the location with the greatest sensitivity to BAT. Sensitivity is measured by observing the hemodynamic changes, *e.g.*, reduction of heart rate and/or BP, associated with increased parasympathetic and/or decreased sympathetic nerve traffic. The electrode is then affixed to the sinus, while the opposite end of the lead is brought to the pulse generator pocket by means of a subcutaneous tunnel<sup>[15]</sup>.

BAT dose is up-titrated over a series of follow-up

visits, much like medications are up-titrated; therapy is initiated at a moderate level in the absence of side effects, then therapy levels are up-titrated as long as the patient can tolerate it, with the objective of achieving full BP lowering at around 3 mo. Because the electrode-baroreceptor interface is unique to each patient, there is no standard dose of the therapy; the focus is therefore to tailor BAT to each individual patient to achieving a therapeutic dose in the absence of side effects<sup>[15]</sup>.

### Disadvantages and limitations of BAT

From what presented thus far, it might seem that BAT is the ideal treatment for all patients, but this is likely not the case, in that, as for all new techniques the initial studies aimed at proving the concept, should be followed by larger prospective multicentre studies in order to prove the effectiveness in the long-term control of arterial hypertension and, moreover, in the prevention of cardiovascular events and the improvement of survival. Overall, the main limitations of BAT entails its logistics requirements: The invasive nature of the procedure, the need for a vascular surgery unit, for general anesthesia, and of an outpatient clinic for periodical follow-up visits in order to check and replace generator battery, and/or timely determine if a possible device failure occurred. Finally, the costs, definitely higher than those of renal denervation, render BAT a therapeutic option to be reserved only for few very well selected patients.

## CONCLUSION

Available accumulating data indicate that BAT is effective and safe. However, patients are required to follow the precautions that are mandatory for all those with implantable devices, and to stay in contact with the hypertension center for regular check-up and monitoring of the battery status. While these disadvantages can be easily overcome with proper logistic arrangements, whether BAT can reduce CV events in the long run is the key question that could only be answered with a large international multicenter study. What control group would be suitable and ethically acceptable to this end is a critical issue that also needs to be addressed.

## ACKNOWLEDGMENTS

We thank the Foundation for Advanced Research In Hypertension and Cardiovascular diseases (www.forica.it).

## REFERENCES

- 1 **Bilgutay AM**, Lillehei CW. Treatment of hypertension with an implantable electronic device. *JAMA* 1965; **191**: 649-653 [PMID: 14242423 DOI: 10.1001/jama.1965.03080080039010]
- 2 **Torresani J**, Heuillet G, Monties JR, Baille Y, Jouve A, Garcia M. Severe arterial hypertension. Treatment by stimulation of the carotid sinus nerve. *Arch Mal Coeur Vaiss* 1967; **60**: 1032-1040 [PMID: 4963450]

- 3 **Wustmann K**, Kucera JP, Scheffers I, Mohaupt M, Kroon AA, de Leeuw PW, Schmidli J, Allemann Y, Delacretaz E. Effects of chronic baroreceptor stimulation on the autonomic cardiovascular regulation in patients with drug-resistant arterial hypertension. *Hypertension* 2009; **54**: 530-536 [PMID: 19620513 DOI: 10.1161/HYPERTENSIONAHA.109.134023]
- 4 **Heusser K**, Tank J, Engeli S, Diedrich A, Menne J, Eckert S, Peters T, Sweep FC, Haller H, Pichlmaier AM, Luft FC, Jordan J. Carotid baroreceptor stimulation, sympathetic activity, baroreflex function, and blood pressure in hypertensive patients. *Hypertension* 2010; **55**: 619-626 [PMID: 20101001 DOI: 10.1161/HYPERTENSIONAHA.109.140665]
- 5 **Rossi GP**, Pessina AC. Resistant Arterial Hypertension. Padova: CLEUP Sc, 2010: 12-15
- 6 **Schlaich MP**, Sobotka PA, Krum H, Lambert E, Esler MD. Renal sympathetic-nerve ablation for uncontrolled hypertension. *N Engl J Med* 2009; **361**: 932-934 [PMID: 19710497 DOI: 10.1056/NEJMc0904179]
- 7 **Krum H**, Schlaich M, Whitbourn R, Sobotka PA, Sadowski J, Bartus K, Kapelak B, Walton A, Sievert H, Thambar S, Abraham WT, Esler M. Catheter-based renal sympathetic denervation for resistant hypertension: a multicentre safety and proof-of-principle cohort study. *Lancet* 2009; **373**: 1275-1281 [PMID: 19332353 DOI: 10.1016/S0140-6736(09)60566-3]
- 8 **Esler MD**, Krum H, Sobotka PA, Schlaich MP, Schmieder RE, Böhm M. Renal sympathetic denervation in patients with treatment-resistant hypertension (The Symplicity HTN-2 Trial): a randomised controlled trial. *Lancet* 2010; **376**: 1903-1909 [PMID: 21093036 DOI: 10.1016/S0140-6736(10)62039-9]
- 9 **Mancia G**, Fagard R, Narkiewicz K, Redán J, Zanchetti A, Böhm M, Christiaens T, Cifkova R, De Backer G, Dominiczak A, Galderisi M, Grobbee DE, Jaarsma T, Kirchhof P, Kjeldsen SE, Laurent S, Manolis AJ, Nilsson PM, Ruilope LM, Schmieder RE, Sirnes PA, Sleight P, Viigimaa M, Waeber B, Zannad F. 2013 Practice guidelines for the management of arterial hypertension of the European Society of Hypertension (ESH) and the European Society of Cardiology (ESC): ESH/ESC Task Force for the Management of Arterial Hypertension. *J Hypertens* 2013; **31**: 1925-1938 [PMID: 24107724 DOI: 10.1097/HJH.0b013e328364ca4c]
- 10 **Bhatt DL**, Kandzari DE, O'Neill WW, D'Agostino R, Flack JM, Katzen BT, Leon MB, Liu M, Mauri L, Negoita M, Cohen SA, Oparil S, Rocha-Singh K, Townsend RR, Bakris GL. A controlled trial of renal denervation for resistant hypertension. *N Engl J Med* 2014; **370**: 1393-1401 [PMID: 24678939 DOI: 10.1056/NEJMoA1402670]
- 11 **Azizi M**, Sapoval M, Gosse P, Monge M, Bobrie G, Delsart P, Midulla M, Mounier-Véhier C, Courand PY, Lantelme P, Denolle T, Dourmap-Collas C, Trillaud H, Pereira H, Plouin PF, Chatellier G. Optimum and stepped care standardised antihypertensive treatment with or without renal denervation for resistant hypertension (DENERHTN): a multicentre, open-label, randomised controlled trial. *Lancet* 2015; **385**: 1957-1965 [PMID: 25631070 DOI: 10.1016/S0140-6736(14)61942-5]
- 12 **Rosa J**, Widimský P, Toušek P, Petrák O, Čurila K, Waldauf P, Bednář F, Zelinka T, Holaj R, Štrauch B, Šomlóová Z, Táborský M, Václavík J, Kociánová E, Branny M, Nykl I, Jiravský O, Widimský J. Randomized comparison of renal denervation versus intensified pharmacotherapy including spironolactone in true-resistant hypertension: six-month results from the Prague-15 study. *Hypertension* 2015; **65**: 407-413 [PMID: 25421981 DOI: 10.1161/HYPERTENSIONAHA.114.04019]
- 13 **de Leeuw PW**, Alnima T, Lovett E, Sica D, Bisognano J, Haller H, Kroon AA. Bilateral or unilateral stimulation for baroreflex activation therapy. *Hypertension* 2015; **65**: 187-192 [PMID: 25331845 DOI: 10.1161/HYPERTENSIONAHA.114.04492]
- 14 **Hoppe UC**, Brandt MC, Wachter R, Beige J, Rump LC, Kroon AA, Cates AW, Lovett EG, Haller H. Minimally invasive system for baroreflex activation therapy chronically lowers blood pressure with pacemaker-like safety profile: results from the Barostim neo

trial. *J Am Soc Hypertens* 2012; **6**: 270-276 [PMID: 22694986 DOI: 10.1016/j.jash.2012.04.004]

- 15 **Abraham WT**, Zile MR, Weaver FA, Butter C, Ducharme A, Halbach M, Klug D, Lovett EG, Müller-Ehmsen J, Schafer JE,

Senni M, Swarup V, Wachter R, Little WC. Baroreflex Activation Therapy for the Treatment of Heart Failure With a Reduced Ejection Fraction. *JACC Heart Fail* 2015; **3**: 487-496 [PMID: 25982108 DOI: 10.1016/j.jchf.2015.02.006]

**P- Reviewer:** Chello M, Kietzmann T, Kosmas Ioannis P, Ramiro S, Salles GF, Tzu-Hung C, Tan XR **S- Editor:** Qiu S  
**L- Editor:** A **E- Editor:** Wu HL



## Roles of catecholamine related polymorphisms in hypertension

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Author contributions: Orun O solely contributed to this paper.

Conflict-of-interest statement: None.

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Received: August 21, 2015  
 Peer-review started: August 24, 2015  
 First decision: September 30, 2015  
 Revised: November 2, 2015  
 Accepted: December 1, 2015  
 Article in press: December 2, 2015  
 Published online: February 23, 2016

### Abstract

The objective of this review is to summarize current data obtained so far in catecholamine-essential hypertension (EH) relationships on a genetic basis. As the major elements driving the sympathetic system's actions, catecholamines modulate a variety of physiological processes and mutations related to the system. This could generate serious disorders, such as severe mental illnesses, stress-induced disorders, or impaired

control of blood pressure or motor pathways. EH is idiopathic, and the genetic basis of its causes and substantial interindividual discrepancies in response to different types of treatments are the focus of interest. Susceptibility to disease or efficacy of treatments are thought to reflect genomic variabilities among individuals. Therefore, outlining the available knowledge in functional genetic polymorphisms linked to EH will make the picture clearer and will help to establish future prospects in the field.

**Key words:** Single nucleotide polymorphism; Catecholamine; Adrenergic receptor; Dopamine receptor; Hypertension; Epinephrine; Norepinephrine

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**Core tip:** Catecholamines are the major elements of sympathetic system's actions, therefore they also act as important regulators of blood pressure. Polymorphism studies require a tedious approach since there are inconsistencies among the studies due to different ethnical origins, subject size and self discrepancies among individuals. Nevertheless, there are many promising findings and still more fields to investigate. Especially role of genes involved in the biosynthesis and metabolism of catecholamines were relatively missing. This review summarizes the current knowledge about catecholamine-related polymorphisms on the basis of development, prognosis and drug response of essential hypertension and aims to improve better assessment of the disease.

Orun O. Roles of catecholamine related polymorphisms in hypertension. *World J Hypertens* 2016; 6(1): 41-52 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/41.htm>  
 DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.41>



## INTRODUCTION

Catecholamines are the class of molecules containing a catechol ring, which consists of a 2-hydroxyl-attached benzene ring, together with an amine chain. Involving epinephrine (or adrenaline), norepinephrine (or noradrenaline), and dopamine, this class of molecules serves to regulate both metabolic and neural mechanisms in the body and act as important targets for a large group of pharmacological agents.

Adrenaline is a hormone synthesized and released by the adrenal medulla into the bloodstream. Its concentration in plasma can rapidly rise up several-fold under a physical or mental tension, and when a sufficiently high concentration is achieved, it can trigger noradrenaline release from adrenergic terminals, an action indirectly affecting neuronal transmission. Noradrenaline, on the other hand, acts as a neurotransmitter rather than a hormone, though it shares very similar chemical structure with adrenaline, except a methyl group. It is released by neurons in the brain and, similar to adrenaline, it acts through alpha or beta adrenergic receptors. Dopamine is the metabolic precursor of noradrenaline and adrenaline, and it also acts as a transmitter/neuromodulator in the central nervous system through dopaminergic receptors.

As a crucial element of drug actions, catecholamine polymorphisms became a focus of interest in various disorders. Hypertension, one of the most common disease worldwide especially among elderly people, is characterized by high blood pressure (BP) and heart rate, two parameters effectively connected to sympathetic denervation. It is a complex disorder with polygenic and environmental determinants. In the majority of cases, it is idiopathic and there is no clear indication of the source. Therefore, searches for hypertension-related genes, mutations, and polymorphisms will assist in the gene-therapeutic approaches and design of target-based therapies. There are more than 50 genes identified so far through association studies, with the reservation of publication bias from selectivity of positive results and limited genotype-phenotype relation analysis<sup>[1]</sup>.

Since the sympathetic nervous system (SNS) is one of the major mechanisms in the rapid regulation and maintenance of BP, it has been hypothesized that the SNS could have a substantial role in the development of essential hypertension (EH). Catecholamines are the mediators of SNS response, and their release from chromaffin cells or ganglionic neuronal ends would affect myocardiocytes; vascular smooth muscle contraction; blood flow through renal, coronary, and cerebral systems; systolic and diastolic BP, etc.

This review will briefly discuss the contribution of genetic polymorphisms in EH by combining known catecholamine-related polymorphisms with the anticipated metabolic and neurologic targets in the regulation of hypertension and will try to accomplish its significance in applications using a combinatorial approach with

the available clinical data on that issue. The elements of the SNS, parasympathetic nervous system, and other BP regulators directly or indirectly correlated with catecholamine action and hormonal regulation of BP and heart rate will be overviewed in this perspective based on the current knowledge of the polymorphisms of the relevant elements. It is believed that this compilation will help to integrate current accumulated knowledge on the field, provide a preliminary perspective for the design of future studies, and increase our understanding of the genetic basis of catecholaminergic system components in this one of the most prevalent and complexly structured disorder.

## RECEPTOR POLYMORPHISMS

One study has indicated that the long arm of human chromosome 5 contains a cluster of genes presumably involved in BP regulation<sup>[2]</sup>. This region contains genes encoding the alpha1B ( $\alpha1B$ ) and beta2-adrenergic ( $\beta2$ ) receptors and dopamine receptor type 1A (D1A). The study was conducted with young Caucasians, and showed that this genome region has a significant association with systolic BP. Further studies involved other receptor types as well, with a prominence in  $\beta2$ -adrenoceptors, which will be discussed in detail later.

## ALPHA-ADRENERGIC RECEPTORS

The  $\alpha$ -adrenergic receptors ( $\alpha$ -AR) are classified into two-subtypes:  $\alpha1$  and  $\alpha2$ . They are G-protein-coupled receptors, and they activate second messenger systems through the activation of G-proteins ( $G_q$  or  $G_{i/o}$ ).  $\alpha1$  mediates vasoconstriction and plays an important role in the regulation of vascular tone, while  $\alpha2$  serves to regulate noradrenaline release from presynaptic terminals.

### ADRA1

The human  $\alpha1A$ -AR is the predominant  $\alpha1$ -AR subtype in vascular smooth muscle, the heart, and the liver. Considering its role in smooth muscle contraction, an early study has investigated the role of a previously determined polymorphism in  $\alpha1A$ -AR, Arg492Cys, in normotensive and hypertensive black and white American individuals and determined the allele frequency distribution. Arg492 was found to be significantly higher in African-Americans with respect to Caucasians, but the frequency of the variant Cys492 was similar in normotensive and hypertensive individuals<sup>[3]</sup>.

Another  $\alpha1A$ -AR polymorphism, Arg347Cys, was examined in a large sample of the Brazilian population (a total of 1568 individuals were involved in the study)<sup>[4]</sup>. In this study, the Cys/Cys genotype was found to be significantly associated with hypertension ( $P = 0.06$ ). Moreover, the response to daily treatment with nifedipine, an anti-hypertensive agent, was found

to be related to the same polymorphism in a Chinese population<sup>[5]</sup>. The later study noted that patients carrying the Cys347 allele of the  $\alpha$ 1A-adrenoceptor gene (ADRA1A) had a greater systolic BP reduction than did those carrying two Arg347 alleles of the  $\alpha$ 1A-adrenoceptor gene ( $32.5 \pm 14.0$  mmHg vs  $27.3 \pm 15.5$  mmHg, respectively,  $P = 0.006$ ) after daily treatment with an oral dose of 30 mg nifedipine gastrointestinal therapeutic system for 16 d, however, diastolic BP reduction was not associated with the Arg347Cys polymorphism. In addition, no significant associations were observed between BP reduction and two other polymorphisms (Arg16Gly and Gln27Glu) of the beta2-adrenoceptor ( $\beta$ 2-AR) gene.

When the  $\alpha$ 1B-adrenoceptor gene (ADRA1B) was examined for possible polymorphisms, it was found that polymorphisms of this gene are much rare than expected considering its close location to the genes of  $\beta$ 2-AR and dopamine receptors (DR)<sup>[2,6]</sup>. An amino acid addition at position 368 (368Arg) and a substitution (Arg371Gly) were investigated in a small population of 24 male patients with uncomplicated EH (12 Caucasians, 12 African-Americans) and 21 male normotensive (NT), first-degree relatives of the study group (12 Caucasians, 9 African-Americans)<sup>[6]</sup>. The study was unable to detect a relationship between these polymorphisms and BP levels or response to phenylephrine, an alpha-agonist used as a decongestant.

The role of the  $\alpha$ 1D-AR subtype in hypertension development was investigated in mice through a salt-induced hypertension model. The study suggested that  $\alpha$ 1D-AR plays an important role in developing a high BP in response to dietary salt-loading, and that agents having selective  $\alpha$ 1D-AR antagonism could have significant therapeutic potential in the treatment of hypertension<sup>[7]</sup>. To our knowledge, there are no studies reporting an association between ADRA1B or ADRA1D gene variants and hypertension.

A recent genome-wide study has strengthened the role of the adrenergic alpha1 receptor (ADRA1) pathway in hypertension and BP regulation. The ADRA1 pathway showed a strong association with diastolic BP ( $P_{\text{path}} < 0.0007$ ) and hypertension ( $P_{\text{path}} < 0.0009$ ) than systolic BP ( $P_{\text{path}} < 0.06$ ). This pathway consisted of genes involved in adrenaline and noradrenaline synthesis, in vascular smooth muscle cell signal transduction leading to intracellular calcium release, and in major regulatory proteins. The study especially stratified the association of  $\alpha$ 1B-AR (ADRA1B) and the phenylethanolamine N-methyl transferase (PNMT) gene, the enzyme that catalyzes conversion of norepinephrine to epinephrine by the transfer of a methyl group<sup>[8]</sup>. The paper, however, emphasized the fact that neither of the remaining pathways utilizing the PNMT reached pathway significance, nor did the removal of ADRA1 receptor genes affect observed ADRA1 pathway significance, suggesting that none of the elements could be self-sufficient mediators for the observed associations.

## ADRA2

The  $\alpha$ 2A-ARs are mainly involved in neurotransmitter release from sympathetic nerves. They are found on pre- and post-synaptic neurons of the central and peripheral nervous systems and blood vessels, and their involvement in BP regulation has been reported by various studies<sup>[9-11]</sup>. Yet, studies investigating the relationship between hypertensivity and different polymorphic sites mostly indicate a lack of association in various ethnic populations.

The  $\alpha$ 2A-ARs act through the  $G_i/G_o$  family of G-proteins, and they help to regulate a wide range of physiologic functions, including vascular, cardiac, and metabolic systems, as well as the central and peripheral nervous systems. Agonist binding to receptors causes the receptor to couple with related G-proteins, which in turn initiates effector responses like the inhibition of adenylyl cyclase or the activation of phospholipase C. Pre-synaptic activation of  $\alpha$ 2-adrenoceptors in sympathetic nerve endings and noradrenergic neurons leads to inhibition of norepinephrine release. Central nervous system activation of post-synaptic  $\alpha$ 2-adrenoceptors inhibits sympathetic activity, which results in hypotension and bradycardia, as well as sedation. Therefore,  $\alpha$ 2 agonists could be potent antihypertensive agents. Higher doses of  $\alpha$ 2-AR agonists, on the other hand, activate smooth muscle receptors in the arterial resistance vessels and could produce hypertension<sup>[12]</sup>.

The BP and other responses to  $\alpha$ 2-AR agonists and antagonists can show high variability among individuals depending on the population pool. Like other signaling systems, variations can involve different elements through the signaling pathway, like G-proteins or effector enzymes, which will be discussed later. On the receptor side, a single nucleotide polymorphism (SNP) of  $\alpha$ 2-AR, which results in Asn-to-Lys substitution at amino acid 251 of the third intracellular loop (position 753), was identified in a study conducted by Small *et al*<sup>[13]</sup>. Subsequently, a total of 376 individuals (125 + 99 NT, 75 + 77 HT for Caucasians and African-Americans, respectively) were genotyped for this locus. The frequency of Lys-251 was 10-fold greater in African-Americans than in Caucasians, but was not associated with EH. Since the third intracellular loop forms the main site of G-protein interaction, the functional role of this substitution was also examined in a cell expression system. There were no detectable changes in ligand binding and basal function, but [<sup>35</sup>S] GTPgammaS binding was 40% greater in Lys251 form. The findings implicated that this small replacement represented a gain of agonist-promoted function with enhanced inhibition of adenylyl cyclase, activation of MAP kinase signaling, or stimulation of phospholipase C/inositol phosphate pathways.

Based on these observations, it can be said that  $\alpha$ 1A-AR polymorphisms R347C and R492C significantly contributed to BP regulation. There is an ongoing research related to the other SNPs in  $\alpha$ 1A as well as

$\alpha$ 1B and  $\alpha$ 1D subtypes and further investigations are needed to accurately assess their roles in hypertension.

## **$\beta$ -ADRENERGIC RECEPTORS**

The  $\beta$ -adrenergic receptors couple to either  $G_s$  or  $G_i$  (heterotrimeric stimulatory and inhibitory G-proteins) proteins.  $\beta_1$ -ARs are the predominant type in the sympathetic control of heart rate and myocardial contraction. Protein kinase A, activated through the  $\beta_1$ -AR  $\rightarrow G_s \rightarrow$  adenylate cyclase (AC)  $\rightarrow$  cyclic adenosine monophosphate (cAMP) pathway, phosphorylates a set of regulatory proteins in cardiac excitation-contraction coupling, such as L-type  $Ca^{2+}$  channels or SERCA proteins.  $\beta_2$ -ARs cause smooth muscle relaxation and bronchodilation. Defective  $\beta_2$ -mediated vasodilation could result in both increased arterial resistance and reduced venous compliance.  $\beta$ -ARs are effectively used as targets to exogenously administered inhibitory agents, known as  $\beta$ -blockers. The  $\beta_3$  receptor, a relatively novel subtype, is mostly found in brown adipose tissue and plays role in the enhancement of lipolysis in this tissue, and is also responsible for thermogenesis in skeletal muscles.

### **ADRB1**

There are many SNPs identified in the gene of  $\beta$ -ARs corresponding to different parts in structure<sup>[14]</sup>. Functional SNPs related to BP regulation were Ser49Gly at the N-terminus and Arg389Gly at the C-terminus of  $\beta_1$ -AR. Three genetic polymorphisms, one of which belongs to the  $\beta_1$ -AR, were investigated in Japanese hypertensive subjects by Shioji *et al*<sup>[15]</sup>. The polymorphisms alpha-adducin (ADD1/Gly460Trp),  $\beta_1$ -adrenoreceptor (ADRB1/Arg389Gly), and G-protein  $\beta_3$  subunit (GNB3/C825T) were screened in 867 males and 1013 females. The ADRB1/R389G polymorphism and hypertensive status in male subjects were close to the significance ( $P = 0.0702$ ). ADD1/G460W polymorphism was associated with hypertension in female subjects, and the GNB3/C825T polymorphism was not associated with hypertensive status in either male or female subjects. None of the polymorphisms was significantly effective on the disease. The relationship between the two polymorphisms (Ser49Gly and Arg389Gly) and BP or heart rate was also tested in a small group of patients (101 subjects) with EH and left ventricular hypertrophy treated with  $\beta_1$ -AR blocker atenolol for 12 wk. Though reduction in heart rate was greater in Gly49 patients compared to the Ser/Ser genotype, there was no significant effect detected on heart rate and BP<sup>[16]</sup>.

### **ADRB2**

At 1998, Timmermann *et al*<sup>[17]</sup> reported four intragenic variants at the promoter region and N-terminus of the  $\beta_2$ -AR in a study involving the offspring of 23 hypertensive and 22 normotensive European families. These mutations were a C $\rightarrow$ T substitution at -47 in the 5' cistron causing Arg $\rightarrow$ Cys exchange, a T $\rightarrow$ C

substitution at -20, and an A $\rightarrow$ G substitution at +46, resulting in Arg $\rightarrow$ Gly exchange of amino acid at 16 and a C $\rightarrow$ G substitution leading to Gln $\rightarrow$ Glu exchange at amino acid 27. All variants were found to be in linkage disequilibrium, but in particular the position -47 variant was significantly higher in frequency in the offspring of hypertensive parents, and Arg16Gly at +46 was significantly associated with parental hypertension and higher BP in this sample pool. Later studies further supported this relationship, mostly focused on the Arg16Gly and Gln27Glu substitutions, which introduce a change on the extracellular part of the receptor; however, as the data accumulated from then, so did contradictory findings. In order to compare study outcomes, a summary of cumulative data according to the ethnicity, study size, and allele distributions for  $\beta_2$ -AR is introduced in Table 1<sup>[18-30]</sup>.

Studies were also conducted to determine if  $\beta_2$ -polymorphisms have an effect on response to antihypertensive reagents. Benazepril is an angiotensin-converting enzyme (ACE) inhibitor used primarily in the treatment of hypertension, congestive heart failure, and heart attacks. A study investigated the role of Arg16Gly polymorphism on systolic and diastolic BP s (SBP and DBP) before and after a 15-d benazepril treatment in a Chinese population that consisted of a total of 931 hypertensive subjects, and showed that ADRB2 R16G polymorphism may play an important role in DBP response to benazepril treatment<sup>[31]</sup>.

### **ADRB3**

The most widely studied  $\beta_3$ -AR polymorphism is the missense mutation at position 64, which replaces tryptophane at this position with arginine. This polymorphism was found to be linked with high body mass index and obesity<sup>[32-34]</sup>. A white population (German) with type 2 diabetes carrying the Arg variant had higher BP and was more hypertensive, though they admitted to intense antihypertensive treatment<sup>[35]</sup>. Likewise, a similar study conducted in a large unselected Southern Italian population involving 979 patients showed that carriers of the Trp64Arg genotype were more often in the upper one-third of abdominal adiposity and were more hypertensive than the Trp64Trp homozygotes<sup>[36]</sup>.

Several polymorphisms previously reported as risk factors in elevated BP and hypertension-ADRA1B, ADRA2A, ADRB1, and ADRB2-were examined in relation to systolic and diastolic BP s and heart rate, both at rest and in response to stress, by McCaffery *et al*<sup>[37]</sup>. Subjects (350 normotensive individuals) of European-American origin were analyzed for their BP s and adrenergic receptor variants at seven sites. At position 1165 of the ADRB1 gene (Gly386Arg), G allele carriers showed higher systolic and diastolic BPs compared to homozygotes for the C allele. In addition, the AA genotype at position 145 of the gene (Ser49Gly) was found to be associated with SBP and DBP. At position 46 of the ADRB2 gene (Arg16Gly), GG homozygotes had

**Table 1** List of recent studies on  $\beta$ 2-adrenergic receptor polymorphisms, blood pressure and hypertension

SNP	Ref.	Ethnicity	Sample size (HT/NT)	Association/significance	Parameter
Arg16Gly	Kotanko <i>et al</i> <sup>[18]</sup> 1997	African Caribbeans	136/81	Yes	Hypertension
Arg16Gly	Gratze <i>et al</i> <sup>[19]</sup> 1999	Austrian Caucasians	57 NT	Yes	Blood pressure regulation
Gln27Glu Arg16Gly	Candy <i>et al</i> <sup>[20]</sup> 2000	Black South African	192/123	No	Hypertension, blood pressure Left ventricular mass
Gln27Glu Arg16Gly	Bray <i>et al</i> <sup>[21]</sup> 2000	Non-hispanic whites	589 families (> 2000)	Yes	Hypertension systolic, diastolic and mean arterial pressure
Gln27Glu Arg16Gly	Jia <i>et al</i> <sup>[22]</sup> 2000	Caucasians	298/298	No	Hypertension
Gln27Glu Arg16Gly	Xie <i>et al</i> <sup>[23]</sup> 2000	Black or white Americans	356/307	No	Hypertension
Arg16Gly	Herrmann <i>et al</i> <sup>[24]</sup> 2000	Black or white Americans	243	No	Hypertension
T-47C Gln27Glu Arg16Gly	Kato <i>et al</i> <sup>[25]</sup> 2001	Japanese	842/633	No	Hypertension
T-47C Gln27Glu Arg16Gly	Ranade <i>et al</i> <sup>[26]</sup> 2001	Chinese	> 800/> 800	Yes (only for Arg16Gly)	Hypertension
Gln27Glu Arg16Gly Thr164Ile	Tomaszewski <i>et al</i> <sup>[27]</sup> 2002	European (Polish)	638	No	Hypertension
Gln27Glu Arg16Gly Thr164Ile	Pereira <i>et al</i> <sup>[28]</sup> 2003	Brasilian	1576	Yes	Hypertension, blood pressure
Gln27Glu Arg16Gly	Galletti <i>et al</i> <sup>[29]</sup> 2004	Non-selected group-middle aged men	405 HT 563 overweight	No	Hypertension, overweight
T-47C Gln27Glu Arg16Gly	Ge <i>et al</i> <sup>[30]</sup> 2005	Han Chinese	503/504	Yes	Hypertension

SNP: Single nucleotide polymorphism; HT: Hypertensive ; NT: Normotensive.

higher resting DBP and AG heterozygotes had lower SBP than other genotypes.

Considering the close relationship between obesity and high BP, polymorphisms of  $\beta$ 2 and  $\beta$ 3 were investigated in a group of Japanese subjects (1121 men), selected as overweight or obese but not having diabetes mellitus or hypertension. The findings of the study demonstrated that the Arg64 allele of  $\beta$ 3 and the Gly16 allele of  $\beta$ 2 could have an indicatory role to predict weight gain-induced BP elevation in obese subjects<sup>[38]</sup>. Similarly, a total of 437 Chinese subjects, including 149 obese hypertensive patients and 139 non-obese essential hypertensive patients, were genotyped to investigate the association between Trp64Arg, Arg16Gly, and Gln27Glu polymorphisms and the susceptibility to obesity and hypertension in a Chinese population. The data revealed that the frequencies of  $\beta$ 3-AR 64Arg and  $\beta$ 2-AR 27Glu were significantly higher in obese hypertensive patients than in the non-obese hypertensive population<sup>[39]</sup>.

The distribution of  $\beta$ -receptor polymorphisms was also determined in hypertension-related complications,

such as left ventricular hypertrophy and arterial stiffness. In a group of 300 patients, pulse wave velocity and hyperemia was found to be associated with Ser49Gly of  $\beta$ 1-AR, while left ventricular hypertrophy was more related to Glu27Gln  $\beta$ 2 polymorphism, suggesting that these two polymorphisms have an effect on the development of arterial stiffness and left ventricular hypertrophy in EH<sup>[40]</sup>.

BP response to the  $\beta$ -blocker atenolol administered 50 mg twice a day was examined in association with hypertension-related and closely linked SNPs of the  $\beta$ 1-adrenergic receptor (Ser49Gly and Arg389Gly) and the  $\beta$ 2-adrenergic receptor (Cys19Arg, Gly16Arg and Gln27Glu), together with G-protein  $\beta$ 3-subunit (A3882C, G5249A and C825T) in EH patients. None of the SNPs were found to be associated with EH, except GNB3 SNPs and BP responses in females<sup>[41]</sup>.

$\beta$ -AR mutations are preeminent in BP regulation and EH compared to other adrenergic receptor subtypes.  $\beta$ -blockers are the well known medications in cardiovascular disorders. A large family of antagonists, such as oxprenolol or pindolol, are in current use to



block or suppress epinephrine or norepinephrine-mediated actions of the sympathetic system. These lower the heart rate, the force of contraction and reduce the BP. Therefore, it is no surprise that primary antihypertensive effects of adrenergic receptors belong to the  $\beta$ -AR family. Arg16Gly and Gln27Glu are likely to be potential genetic factors to consider and worthy of attention. Positive associations were reported in large scale Chinese cohorts, but studies conducted in other populations are rather inconsistent and should be supported with further analyses.

## DOPAMINE RECEPTORS

Dopamine is a neurotransmitter with a variety of roles, majorly in the brain, but also throughout the body. In the brain, it mediates reward-motivated reactions, and helps to produce coordinated motor output, neuro-endocrine regulation, etc. Thus, several important diseases, like Parkinson's or schizophrenia, are highly interfered with dopamine activity. Outside the brain, it acts as a vasodilator in blood vessels, and in kidneys it controls renal sodium excretion.

Dopamine receptors are classified into two families: D1-like (includes D1 and D5) and D2-like (includes D2, D3, and D4). Both D1- and D2-like receptors mainly exist in the central nervous system, as well as on the smooth muscle of renal arteries, the juxtaglomerular apparatus, and the tubules of the kidney and cardiopulmonary system. Like adrenergic receptors, they are G-protein-coupled receptors with seven transmembrane domains. Both members of the D1-family, D1 and D5, could interact with stimulatory  $G_s$ , but coupling with other members of G-proteins can be different for each subtype. For example, D1 can also interact with  $G_o$ , participating in the regulation of ion channels like  $Ca^{2+}$ ,  $K^+$ , and  $Na^+$ , while D5 can couple to  $G_i$  members.

The activation of AC through  $G_s$  will cause activation of protein kinase A, which in turn will phosphorylate target proteins. In kidney proximal tubules, phosphorylation of two proteins by PKA, the  $Na^+$ -H $^+$  exchanger (NHE), and  $Na^+$ -K $^+$  ATPase (NKA) will inhibit their activation and affect sodium transport across tubules.

The relationship between salt intake and the development of hypertension, together with the renal functions of the dopaminergic system, brought dopaminergic receptor polymorphisms into attention in hypertension research. In 2000, Sato *et al*<sup>[42]</sup> screened 131 Japanese EH subjects for the A-48G polymorphic site in the DRD1 gene and showed that EH patients carrying the G allele had a higher diastolic BP in general. Later, the allele frequencies of two SNPs, A-48G and G-94A, were determined in a larger cohort, consisting of 493 hypertensive Caucasian subjects. In contrast to the study involving Japanese patients, this study was unable to show any correlation with hypertension in this population, reflecting the role of ethnicity in polymorphism-related secondary effects<sup>[43]</sup>.

When renal clearance of sodium was taken into

consideration, the DRD1 polymorphisms A-48G, G-94A, and C-800T were shown to have an effect in the reabsorption of sodium, especially from distal tubules. In a multivariate association analysis, it was shown that DRD1-94GG homozygotes had lower reabsorption rates. The transmission of the DRD1 AGC haplotype was found to be associated with lower systolic and diastolic BP in a family-based analysis<sup>[44]</sup>. In a small Turkish cohort involving 101 EH patients run by our group, we were not able to obtain such a correlation with the same SNPs (A-48G and G-94A), suggesting that further analysis is required to clear the picture<sup>[45]</sup>.

Dopamine D2 receptors act through  $G_i$  proteins, and their inhibitory action on AC reduces the noradrenaline release from sympathetic nerve terminals. Rosmond *et al*<sup>[46]</sup> examined a common polymorphism in the coding region, an NcoI site in exon 6 (position 1128) in relation to BP and personality disorders. They found that NcoI site polymorphism of DRD2 is associated with BP, and that the TT genotype was significantly more frequent in hypertensive subjects (284 randomly selected 51-year-old Swedish men) compared to controls.

Obviously, one big gap to be filled is the relation of dopaminergic system variants with hypertension. So far, just a few SNPs have been explored and shown to be associated with EH. Effects are mostly through salt transport in the renal tubules and A-48G in DRD1, in particular, have confirmative data. There are not many available data concerning dopaminergic receptor subtype polymorphisms and in view of its role in the SNS, this field should be considered more extensively in future studies.

## OTHER FACTORS

As mentioned previously, catecholamines are the major contributors of SNS actions and the catecholaminergic system is an essential component for the performance of SS activities. There are many enzymes involved in the biosynthesis of catecholamines, which occurs in the chromaffin cells of the adrenal medulla and post-ganglionic fibers of the SNS (Figure 1). The removal of secreted molecules requires mainly actions of two enzymes, monoamine oxidase and catechol-O-methyltransferase (COMT), but also other downstream enzymes, like aldehyde dehydrogenase or aldehyde reductase (Figures 2 and 3). Synthesized catecholamines are stored in vesicles, where stabilization of the vesicle core requires other supportive peptides, like chromogranin<sup>[47]</sup>. There are also presynaptic transporters that help to remove released molecules from the synaptic cleft back to the presynaptic terminal; these transporters are the targets for drugs of abuse<sup>[48]</sup>. To our knowledge, there is no report affirming the role of transporter polymorphisms in BP levels or hypertension development, except some preliminary studies suggesting a predisposition<sup>[49]</sup>.

In action, catecholamines act through their receptors and start signal transduction. As mentioned above,

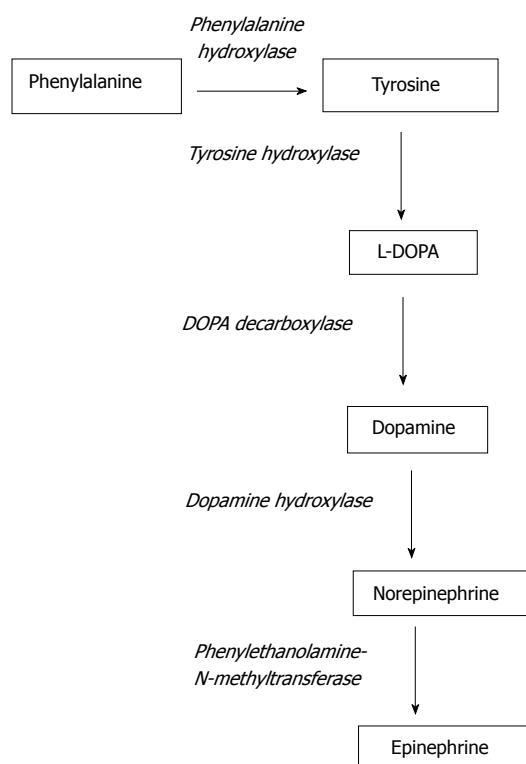


Figure 1 Outline of catecholamine biosynthesis.

catecholamine receptors belong to the GPCR family, and they couple with heterotrimeric G-proteins, finally ending with the activation of protein kinases. There are many proteins taking place on these signal transduction pathways, reflecting the complexity and limited power of association studies. In this large frame of action, the effects of polymorphisms related to the system are too rich to cover in full perspective. In the context of BP and hypertension, however, some important factors are presented by several groups as strong candidates worthy of mention, and will be summarized briefly in the rest of the manuscript.

## BIOSYNTHESIS-RELATED FACTORS

The first step in the biosynthesis of dopamine is the formation of L-DOPA, a dopamine precursor, from the amino acid tyrosine by the enzyme tyrosine hydroxylase (TH). TH is the rate-limiting enzyme in catecholamine synthesis. Recent studies have indicated that several polymorphisms of the *TH* gene contribute to BP regulation. Two SNPs at the promoter region of the gene, C-824T and A-581G, were found to be strongly associated with higher BP under stress<sup>[50]</sup>. In 2010, it was shown that these replacements seriously alter TH promoter activity<sup>[51,52]</sup>. In accordance with this, Nielsen *et al*<sup>[53]</sup> reported that the -824T allele increased the relative risk of hypertension by 45%<sup>[47]</sup>. A study involving 1266 hypertensive subjects searched for the effect of C-824T of TH in hypertension, in addition to the two loci of chromogranin A (CHGA). CHGA is a

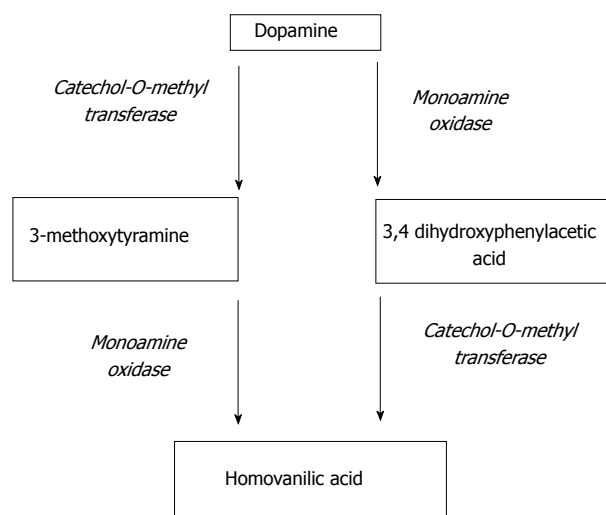


Figure 2 Enzymatic degradation process of dopamine.

peptide located in storage vesicles, and early reports have displayed a strong association between 3'-UTR (C +87T) and EH<sup>[54,55]</sup>. CHGA polymorphism predicted the risk of developing hypertensive kidney disease in African-Americans. Homozygosity for the minor alleles at T-1014C, T-988G, and G-462A at the promoter region of CGHA exhibited lower stress-induced BP elevations<sup>[56]</sup>.

Another SNP localized to the dopamine hydroxylase promoter, C-970T, was also found to be related with the risk of developing hypertension<sup>[57]</sup>. More recently, genetic variations of the *PNMT* gene in relation to hypertension were reported in several studies. The distribution of two SNPs, G-367A (rs3764351) and G-161A (rs876493), together with their haplotypes, was screened in 316 pairs of HT and NT patients. Two SNPs' AA haplotypes were found to be less common in hypertensives and therefore suggested to be correlated with the decreased risk of EH in the Han Chinese population<sup>[58]</sup>.

## METABOLISM-RELATED FACTORS

The degradation of secreted catecholamines to prevent prolonged stimulation of SNS is very important for the modulation of physiological processes, involving BP and related cardiac functions. Renalase, a novel flavin adenine nucleotide-dependent amine oxidase, is secreted by the kidneys; it helps to reduce the circulating catecholamine concentration. Eight selected SNPs of the renalase gene were genotyped in 503 cases, and three SNPs - rs2576178, rs2296545, and rs2114406 - showed significant associations with EH<sup>[59]</sup>. The frequency of allele A for rs2576178 in patients with hypertensive and concomitant coronary heart disease was markedly higher. Similarly, the frequency of the C allele of rs2296545 was higher in hypertensives, showing that both genotypes may be contributing to the development of hypertension and chronic heart

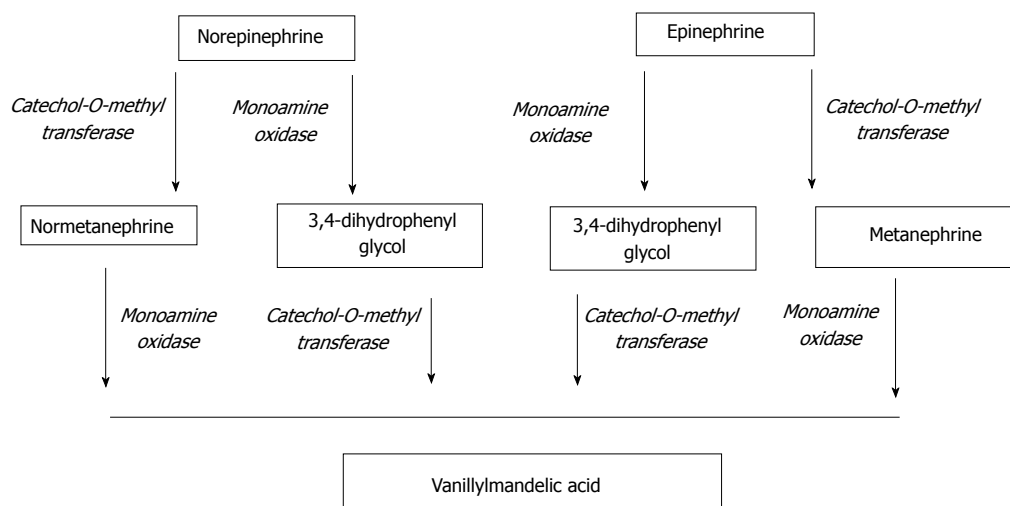


Figure 3 Enzymatic degradation process of epinephrine and norepinephrine.

disease<sup>[60]</sup>.

There were inconsistent results with respect to the *COMT* gene variant *Val158Met* and hypertension. A study on a Japanese population, involving 735 men, showed that the Met allele is associated with higher BP and higher prevalence of hypertension in Japanese men<sup>[61]</sup>. Another study on a Chinese population, including 215 hypertensive patients, did not detect such a relationship<sup>[62]</sup>. In the 1995-1997 Word-Trøndelag Health Study (HUNT) group involving 2591 individuals, the Val/Val genotype was found to be more frequent among individuals with hypertension<sup>[63]</sup>.

## SIGNAL TRANSDUCTION

A single-base substitution of C825T in exon 10 of the gene encoding the G protein  $\beta 3$  subunit of heterotrimeric Gi proteins (GNB3) was found to be associated with hypertension<sup>[64]</sup>. This polymorphism leads to alternative splicing of exon 9 and has been associated with enhanced Gi signaling and ion transport<sup>[64,65]</sup>.

Enhanced G-protein-mediated signaling resulting from the truncated C825T form of the G-protein  $\beta$ -subunit may cause high BP as a result of increased NHE activity in tubules, elevated calcium concentration in the cytoplasm, and increased contractility<sup>[66]</sup>.

A large accumulation of studies is present in the literature, strongly supporting the role of C825T in hypertension<sup>[64,67-70]</sup>. Although the majority of studies has shown an association between the 825T allele and hypertension, there are some contradicting reports, especially in subjects of African and Asian origin, again emphasizing the importance of ethnic origin. Two studies performed by our group also showed that the frequency of the 825T-allele was higher in hypertensive subjects compared to that of controls, and that the difference was statistically significant<sup>[71,72]</sup>.

Recent studies report that the effect of the dopaminergic system in hypertension is mostly due to the

impairment of the intrarenal dopaminergic system, and one of the major players is the GRK4, a serine/threonine G-protein receptor kinase, which initiates a desensitization process of the receptor and prevents constitutive activity. Functional polymorphisms of this protein could enhance GRK4 activity, which will reduce dopamine receptor transduction. Increased GRK4 activity also increases angiotensin II AT1 receptor activity that is associated with EH<sup>[73]</sup>.

In a small cohort involving 100 EH patients, three gene variants of GRK4 (R65L, A142V, and A486V) were found to be associated with antihypertensive treatment responses<sup>[74]</sup>. The responses of homozygous double variants of 65L and 142V in particular were much less than the other variants. In another study consisting of 168 Caucasian EH patients, the V allele of the A486V variant was shown to be associated with hypertension and systolic BP<sup>[75]</sup>. A much larger cohort consisting of 934 whites and African-Americans (44.2%) was also investigated for three proteins of GRK4; it was determined that the 65L allele had a significant effect on systolic BP<sup>[76]</sup>.

Overall, among the synthesis, degradation and signal transduction pathways, there are several replacements possibly involved in the development and progression of hypertension. The most established and well-characterized of these is the C825T mutation in the heterotrimeric G-protein  $\beta$ -subunit. Tyrosine hydroxylase, the primary enzyme for the synthesis of catecholamines, and GRK4, an enzyme with a vital role in salt-transport through regulation of dopamine receptor activity, are conspicuous factors in the assessment of the disease.

## CONCLUSION

In the new guidelines released by the European Society of Hypertension (ESH) and the European Society of Cardiology (ESC), hypertension have been re-

evaluated in the context of its role as a risk factor for cardiovascular diseases<sup>[77,78]</sup>. Among the previously defined or recently added parameters, individualized therapy approaches are a major concern to estimate overall risk for the patient for the determination and application of the most appropriate treatment and drug regimen<sup>[79]</sup>. In this perspective, genetic factors need to be well-characterized since they are important contributors of individualized risk assessments.

Regulation of BP could be enhanced and related cardiovascular damage could be reduced if predictors are properly stratified.

As stated above, this review has been restricted to the genetic polymorphisms determined in the catecholamine pathways in relation to BP regulation and hypertension. The selected works contained mostly either positive association studies, unique studies, or a few rare reports in the field of interest. Most of the works in the field are relatively novel, and there is a great number of vacancies to be filled out.

Polymorphism studies always have drawbacks causing them to have inconsistent results, such as ethnicity, sample power, sex, polygenetic factors or linkage effects, and, in the case of drug response studies, periods and consistencies of applied treatments, the reliability of control groups, etc. Nevertheless, as will be recognized from the aforementioned reports, the findings are quite remarkable, and a number of studies coincide closely in the outcomes; several variants are highly promising in their potential as predictive markers to estimate the susceptibility of patients to developing hypertension or negative responses to anti-hypertensive drug treatments. As genome-wide association studies are added up, more reliable predictions and their clinical relevance will be achievable, leading the way to more appropriate risk assessments.

## REFERENCES

- 1 **Morris BJ**, Benjafield AV, Lin RC. Essential hypertension: genes and dreams. *Clin Chem Lab Med* 2003; **41**: 834-844 [PMID: 12940506 DOI: 10.1515/CCLM.2003.127]
- 2 **Krushkal J**, Xiong M, Ferrell R, Sing CF, Turner ST, Boerwinkle E. Linkage and association of adrenergic and dopamine receptor genes in the distal portion of the long arm of chromosome 5 with systolic blood pressure variation. *Hum Mol Genet* 1998; **7**: 1379-1383 [PMID: 9700190 DOI: 10.1093/hmg/7.9.1379]
- 3 **Xie HG**, Kim RB, Stein CM, Gainer JV, Brown NJ, Wood AJ. Alpha1A-adrenergic receptor polymorphism: association with ethnicity but not essential hypertension. *Pharmacogenetics* 1999; **9**: 651-656 [PMID: 10591546]
- 4 **Freitas SR**, Pereira AC, Floriano MS, Mill JG, Krieger JE. Association of alpha1A-adrenergic receptor polymorphism and blood pressure phenotypes in the Brazilian population. *BMC Cardiovasc Disord* 2008; **8**: 40 [PMID: 19105822 DOI: 10.1186/1471-2261-8-40]
- 5 **Zhang Y**, Hong X, Liu H, Huo Y, Xu X. Arg347Cys polymorphism of alpha1A-adrenoceptor gene is associated with blood pressure response to nifedipine GITS in Chinese hypertensive patients. *J Hum Genet* 2009; **54**: 360-364 [PMID: 19444285 DOI: 10.1038/jhg.2009.42]
- 6 **Büscher R**, Herrmann V, Ring KM, Kailasam MT, O'Connor DT, Parmer RJ, Insel PA. Variability in phenylephrine response and essential hypertension: a search for human alpha(1B)-adrenergic receptor polymorphisms. *J Pharmacol Exp Ther* 1999; **291**: 793-798 [PMID: 10525102]
- 7 **Tanoue A**, Koba M, Miyawaki S, Koshimizu TA, Hosoda C, Oshikawa S, Tsujimoto G. Role of the alpha1D-adrenergic receptor in the development of salt-induced hypertension. *Hypertension* 2002; **40**: 101-106 [PMID: 12105146 DOI: 10.1161/01.HYP.0000022062.70639.1C]
- 8 **Reder NP**, Tayo BO, Salako B, Ogunniyi A, Adeyemo A, Rotimi C, Cooper RS. Adrenergic alpha-1 pathway is associated with hypertension among Nigerians in a pathway-focused analysis. *PLoS One* 2012; **7**: e37145 [PMID: 22615923 DOI: 10.1371/journal.pone.0037145]
- 9 **Gavras I**, Manolis AJ, Gavras H. The alpha2-adrenergic receptors in hypertension and heart failure: experimental and clinical studies. *J Hypertens* 2001; **19**: 2115-2124 [PMID: 11725152]
- 10 **Freeman K**, Farrow S, Schmaier A, Freedman R, Schork T, Lockette W. Genetic polymorphism of the alpha 2-adrenergic receptor is associated with increased platelet aggregation, baroreceptor sensitivity, and salt excretion in normotensive humans. *Am J Hypertens* 1995; **8**: 863-869 [PMID: 8541000 DOI: 10.1016/0895-7061(95)00155-I]
- 11 **Lockette W**, Ghosh S, Farrow S, MacKenzie S, Baker S, Miles P, Schork A, Cadaret L. Alpha 2-adrenergic receptor gene polymorphism and hypertension in blacks. *Am J Hypertens* 1995; **8**: 390-394 [PMID: 7619352 DOI: 10.1016/0895-7061(95)00024-J]
- 12 **Ma D**, Rajakumaraswamy N, Maze M. alpha2-Adrenoceptor agonists: shedding light on neuroprotection? *Br Med Bull* 2004; **71**: 77-92 [PMID: 15684247 DOI: 10.1093/bmb/ldh036]
- 13 **Small KM**, Forbes SL, Brown KM, Liggett SB. An asn to lys polymorphism in the third intracellular loop of the human alpha 2A-adrenergic receptor imparts enhanced agonist-promoted Gi coupling. *J Biol Chem* 2000; **275**: 38518-38523 [PMID: 10948191 DOI: 10.1074/jbc.M004550200]
- 14 **Taylor MR**, Bristow MR. The emerging pharmacogenomics of the beta-adrenergic receptors. *Congest Heart Fail* 2004; **10**: 281-288 [PMID: 15591842]
- 15 **Shioji K**, Kokubo Y, Mannami T, Inamoto N, Morisaki H, Mino Y, Tagoi N, Yasui N, Iwami N. Association between hypertension and the alpha-adducin, beta1-adrenoreceptor, and G-protein beta3 subunit genes in the Japanese population; the Suita study. *Hypertens Res* 2004; **27**: 31-37 [PMID: 15055253 DOI: 10.1291/hypres.27.31]
- 16 **Karlsson J**, Lind L, Hallberg P, Michaëlsson K, Kurland L, Kahan T, Malmqvist K, Ohman KP, Nyström F, Melhus H. Beta1-adrenergic receptor gene polymorphisms and response to beta1-adrenergic receptor blockade in patients with essential hypertension. *Clin Cardiol* 2004; **27**: 347-350 [PMID: 15237695]
- 17 **Timmermann B**, Mo R, Luft FC, Gerdts E, Busjahn A, Omvik P, Li GH, Schuster H, Wienker TF, Hoehe MR, Lund-Johansen P. Beta-2 adrenoceptor genetic variation is associated with genetic predisposition to essential hypertension: The Bergen Blood Pressure Study. *Kidney Int* 1998; **53**: 1455-1460 [PMID: 9607174 DOI: 10.1046/j.1523-1755.1998.00926.x]
- 18 **Kotanko P**, Binder A, Tasker J, DeFreitas P, Kamdar S, Clark AJ, Skrabal F, Caulfield M. Essential hypertension in African Caribbeans associates with a variant of the beta2-adrenoceptor. *Hypertension* 1997; **30**: 773-776 [PMID: 9336371 DOI: 10.1161/01.HYP.30.4.773]
- 19 **Gratz G**, Fortin J, Labugger R, Binder A, Kotanko P, Timmermann B, Luft FC, Hoehe MR, Skrabal F. beta-2 Adrenergic receptor variants affect resting blood pressure and agonist-induced vasodilation in young adult Caucasians. *Hypertension* 1999; **33**: 1425-1430 [PMID: 10373227 DOI: 10.1161/01.HYP.33.6.1425]
- 20 **Candy G**, Samani N, Norton G, Woodiwiss A, Radevski I, Wheatley A, Cockcroft J, Hall IP. Association analysis of beta2 adrenoceptor polymorphisms with hypertension in a Black African population. *J Hypertens* 2000; **18**: 167-172 [PMID: 10694184]
- 21 **Bray MS**, Krushkal J, Li L, Ferrell R, Kardias S, Sing CF, Turner ST, Boerwinkle E. Positional genomic analysis identifies the beta(2)-adrenergic receptor gene as a susceptibility locus for human hypertension. *Circulation* 2000; **101**: 2877-2882 [PMID: 10869257]



- DOI: 10.1161/01.CIR.101.25.2877]
- 22 **Jia H**, Sharma P, Hopper R, Dickerson C, Lloyd DD, Brown MJ. beta2-adrenoceptor gene polymorphisms and blood pressure variations in East Anglian Caucasians. *J Hypertens* 2000; **18**: 687-693 [PMID: 10872552]
  - 23 **Xie HG**, Stein CM, Kim RB, Gainer JV, Sofowora G, Dishy V, Brown NJ, Goree RE, Haines JL, Wood AJ. Human beta2-adrenergic receptor polymorphisms: no association with essential hypertension in black or white Americans. *Clin Pharmacol Ther* 2000; **67**: 670-675 [PMID: 10872649 DOI: 10.1067/mcp.2000.106293]
  - 24 **Herrmann V**, Büscher R, Go MM, Ring KM, Hofer JK, Kailasam MT, O'Connor DT, Parmer RJ, Insel PA. Beta2-adrenergic receptor polymorphisms at codon 16, cardiovascular phenotypes and essential hypertension in whites and African Americans. *Am J Hypertens* 2000; **13**: 1021-1026 [PMID: 10981553 DOI: 10.1016/S0895-7061(00)01188-2]
  - 25 **Kato N**, Sugiyama T, Morita H, Kurihara H, Sato T, Yamori Y, Yazaki Y. Association analysis of beta(2)-adrenergic receptor polymorphisms with hypertension in Japanese. *Hypertension* 2001; **37**: 286-292 [PMID: 11230287 DOI: 10.1161/01.HYP.37.2.286]
  - 26 **Ranade K**, Shue WH, Hung YJ, Hsuing CA, Chiang FT, Pesich R, Hebert J, Olivier M, Chen YD, Pratt R, Olshen R, Curb D, Botstein D, Risch N, Cox DR. The glycine allele of a glycine/arginine polymorphism in the beta2-adrenergic receptor gene is associated with essential hypertension in a population of Chinese origin. *Am J Hypertens* 2001; **14**: 1196-1200 [PMID: 11775126 DOI: 10.1016/S0895-7061(01)02213-0]
  - 27 **Tomaszewski M**, Brain NJ, Charchar FJ, Wang WY, Lacka B, Padmanabahn S, Clark JS, Anderson NH, Edwards HV, Zukowska-Szczechowska E, Grzeszczak W, Dominiczak AF. Essential hypertension and beta2-adrenergic receptor gene: linkage and association analysis. *Hypertension* 2002; **40**: 286-291 [PMID: 12215468 DOI: 10.1161/01.HYP.0000029105.21202.FE]
  - 28 **Pereira AC**, Floriano MS, Mota GF, Cunha RS, Herkenhoff FL, Mill JG, Krieger JE. Beta2 adrenoceptor functional gene variants, obesity, and blood pressure level interactions in the general population. *Hypertension* 2003; **42**: 685-692 [PMID: 12900437 DOI: 10.1161/01.HYP.0000085648.65419.17]
  - 29 **Galletti F**, Iacone R, Ragone E, Russo O, Della Valle E, Siani A, Barba G, Farinara E, Strazzullo V, Strazzullo P. Lack of association between polymorphism in the beta2-adrenergic receptor gene, hypertension, and obesity in the Olivetti heart study. *Am J Hypertens* 2004; **17**: 718-720 [PMID: 15323067 DOI: 10.1016/j.amjhyper.2004.04.012]
  - 30 **Ge D**, Huang J, He J, Li B, Duan X, Chen R, Gu D. beta2-Adrenergic receptor gene variations associated with stage-2 hypertension in northern Han Chinese. *Ann Hum Genet* 2005; **69**: 36-44 [PMID: 15638826 DOI: 10.1046/j.1529-8817.2003.00093.x]
  - 31 **Huang G**, Xing H, Hao K, Peng S, Wu D, Guang W, Huang A, Hong X, Wang Y, Feng Y, Zhang Y, Li J, Chen C, Wang B, Zhang X, Li D, Yu Y, Liu J, Zhu G, Huo Y, Chen D, Hou Y, Wang X, Xu X, Niu T, Xu X. Beta2 adrenergic receptor gene Arg16Gly polymorphism is associated with therapeutic efficacy of benazepril on essential hypertension in Chinese. *Clin Exp Hypertens* 2004; **26**: 581-592 [PMID: 15554460]
  - 32 **Kurabayashi T**, Carey DG, Morrison NA. The beta 3-adrenergic receptor gene Trp64Arg mutation is overrepresented in obese women. Effects on weight, BMI, abdominal fat, blood pressure, and reproductive history in an elderly Australian population. *Diabetes* 1996; **45**: 1358-1363 [PMID: 8826971 DOI: 10.2337/diab.45.10.1358]
  - 33 **Yoshioka K**, Yoshida T, Sakane N, Umekawa T, Takahashi T, Sakai Y, Kondo M. Association of Trp64Arg mutation of the beta 3-adrenergic receptor gene with NIDDM, current and maximal body mass index. *Diabetologia* 1996; **39**: 1410-1411 [PMID: 8933016]
  - 34 **Yoshida T**, Sakane N. Association between beta3-adrenoreceptor polymorphism with obesity and diabetes in Japan. *Intern Med* 1999; **38**: 207-209 [PMID: 10225689]
  - 35 **Ringel J**, Kreutz R, Distler A, Sharma AM. The Trp64Arg polymorphism of the beta3-adrenergic receptor gene is associated with hypertension in men with type 2 diabetes mellitus. *Am J Hypertens* 2000; **13**: 1027-1031 [PMID: 10981554 DOI: 10.1016/S0895-7061(00)00290-9]
  - 36 **Strazzullo P**, Iacone R, Siani A, Cappuccio FP, Russo O, Barba G, Barbato A, D'Elia L, Trevisan M, Farinara E. Relationship of the Trp64Arg polymorphism of the beta3-adrenoceptor gene to central adiposity and high blood pressure: interaction with age. Cross-sectional and longitudinal findings of the Olivetti Prospective Heart Study. *J Hypertens* 2001; **19**: 399-406 [PMID: 11288809]
  - 37 **McCaffery JM**, Pogue-Geile MF, Ferrell RE, Petro N, Manuck SB. Variability within alpha- and beta-adrenoreceptor genes as a predictor of cardiovascular function at rest and in response to mental challenge. *J Hypertens* 2002; **20**: 1105-1114 [PMID: 1203679]
  - 38 **Kawaguchi H**, Masuo K, Katsuya T, Sugimoto K, Rakugi H, Ogihara T, Tuck ML. beta2- and beta3-Adrenoceptor polymorphisms relate to subsequent weight gain and blood pressure elevation in obese normotensive individuals. *Hypertens Res* 2006; **29**: 951-959 [PMID: 17378367 DOI: 10.1291/hypres.29.951]
  - 39 **Mo W**, Zhang GG, Yang TL, Dai XP, Li HH, Zeng H, Liu J, Tan YM, Zhou HH, Liu ZQ. The genetic polymorphisms of beta3-adrenergic receptor (AR) Trp64Arg and beta2-AR Gln27Glu are associated with obesity in Chinese male hypertensive patients. *Clin Chem Lab Med* 2007; **45**: 493-498 [PMID: 17439327 DOI: 10.1515/CCLM.2007.089]
  - 40 **Yuan M**, Ohishi M, Ito N, Sugimoto K, Takagi T, Terai M, Katsuya T, Rakugi H, Wu Z, Ogihara T. Genetic influences of beta-adrenoceptor polymorphisms on arterial functional changes and cardiac remodeling in hypertensive patients. *Hypertens Res* 2006; **29**: 875-881 [PMID: 17345787 DOI: 10.1291/hypres.29.875]
  - 41 **Filigheddu F**, Argiolas G, Degortes S, Zaninello R, Frau F, Pitzoi S, Bulla E, Bulla P, Troffa C, Glorioso N. Haplotypes of the adrenergic system predict the blood pressure response to beta-blockers in women with essential hypertension. *Pharmacogenomics* 2010; **11**: 319-325 [PMID: 20235788 DOI: 10.2217/pgs.09.158]
  - 42 **Sato M**, Soma M, Nakayama T, Kanmatsuse K. Dopamine D1 receptor gene polymorphism is associated with essential hypertension. *Hypertension* 2000; **36**: 183-186 [PMID: 10948075 DOI: 10.1161/01.HYP.36.2.183]
  - 43 **Beige J**, Bellmann A, Sharma AM, Gessner R. Ethnic origin determines the impact of genetic variants in dopamine receptor gene (DRD1) concerning essential hypertension. *Am J Hypertens* 2004; **17**: 1184-1187 [PMID: 15607627 DOI: 10.1016/j.amjhyper.2004.07.013]
  - 44 **Staessen JA**, Kuznetsova T, Zhang H, Maillard M, Bochud M, Hasenkamp S, Westerkamp J, Richart T, Thijs L, Li X, Brand-Herrmann SM, Burnier M, Brand E. Blood pressure and renal sodium handling in relation to genetic variation in the DRD1 promoter and GRK4. *Hypertension* 2008; **51**: 1643-1650 [PMID: 18413491 DOI: 10.1161/HYPERTENSIONAHA.107.109611]
  - 45 **Orun O**, Nacar C, Cabadak H, Tiber PM, Doğan Y, Güneşel Ö, Fak AS, Kan B. Investigation of the association between dopamine D1 receptor gene polymorphisms and essential hypertension in a group of Turkish subjects. *Clin Exp Hypertens* 2011; **33**: 418-421 [PMID: 21797797 DOI: 10.3109/10641963.2011.561898]
  - 46 **Rosmond R**, Rankinen T, Chagnon M, Pérusse L, Chagnon YC, Bouchard C, Björntorp P. Polymorphism in exon 6 of the dopamine D(2) receptor gene (DRD2) is associated with elevated blood pressure and personality disorders in men. *J Hum Hypertens* 2001; **15**: 553-558 [PMID: 11494094]
  - 47 **Currie G**, Freel EM, Perry CG, Dominiczak AF. Disorders of blood pressure regulation-role of catecholamine biosynthesis, release, and metabolism. *Curr Hypertens Rep* 2012; **14**: 38-45 [PMID: 22068338 DOI: 10.1007/s11906-011-0239-2]
  - 48 **Lorang D**, Amara SG, Simerly RB. Cell-type-specific expression of catecholamine transporters in the rat brain. *J Neurosci* 1994; **14**: 4903-4914 [PMID: 8046459]
  - 49 **Ksiazek P**, Buraczynska K, Buraczynska M. Norepinephrine transporter gene (NET) polymorphism in patients with type 2 diabetes. *Kidney Blood Press Res* 2006; **29**: 338-343 [PMID: 17124432 DOI: 10.1159/000097356]
  - 50 **Rao F**, Wen G, Gayen JR, Das M, Vaingankar SM, Rana BK,

- Mahata M, Kennedy BP, Salem RM, Stridsberg M, Abel K, Smith DW, Eskin E, Schork NJ, Hamilton BA, Ziegler MG, Mahata SK, O'Connor DT. Catecholamine release-inhibitory peptide catestatin (chromogranin A(352-372)): naturally occurring amino acid variant Gly364Ser causes profound changes in human autonomic activity and alters risk for hypertension. *Circulation* 2007; **115**: 2271-2281 [PMID: 17438154 DOI: 10.1161/CIRCULATIONAHA.106.628859]
- 51 **Zhang K**, Zhang L, Rao F, Brar B, Rodriguez-Flores JL, Taupenot L, O'Connor DT. Human tyrosine hydroxylase natural genetic variation: delineation of functional transcriptional control motifs disrupted in the proximal promoter. *Circ Cardiovasc Genet* 2010; **3**: 187-198 [PMID: 20124442 DOI: 10.1161/CIRCGENETICS.109.904813]
- 52 **Rao F**, Zhang K, Zhang L, Rana BK, Wessel J, Fung MM, Rodriguez-Flores JL, Taupenot L, Ziegler MG, O'Connor DT. Human tyrosine hydroxylase natural allelic variation: influence on autonomic function and hypertension. *Cell Mol Neurobiol* 2010; **30**: 1391-1394 [PMID: 20571875 DOI: 10.1007/s10571-010-9535-7]
- 53 **Nielsen SJ**, Jeppesen J, Torp-Pedersen C, Hansen TW, Linneberg A, Fenger M. Tyrosine hydroxylase polymorphism (C-824T) and hypertension: a population-based study. *Am J Hypertens* 2010; **23**: 1306-1311 [PMID: 20706199 DOI: 10.1038/ajh.2010.165]
- 54 **Chen Y**, Rao F, Rodriguez-Flores JL, Mahapatra NR, Mahata M, Wen G, Salem RM, Shih PA, Das M, Schork NJ, Ziegler MG, Hamilton BA, Mahata SK, O'Connor DT. Common genetic variants in the chromogranin A promoter alter autonomic activity and blood pressure. *Kidney Int* 2008; **74**: 115-125 [PMID: 18432188 DOI: 10.1038/ki.2008.113]
- 55 **Salem RM**, Cadman PE, Chen Y, Rao F, Wen G, Hamilton BA, Rana BK, Smith DW, Stridsberg M, Ward HJ, Mahata M, Mahata SK, Bowden DW, Hicks PJ, Freedman BI, Schork NJ, O'Connor DT. Chromogranin A polymorphisms are associated with hypertensive renal disease. *J Am Soc Nephrol* 2008; **19**: 600-614 [PMID: 18235090 DOI: 10.1681/ASN.2007070754]
- 56 **Kang SW**. Adrenergic genetic mechanisms in hypertension and hypertensive kidney disease. *Electrolyte Blood Press* 2013; **11**: 24-28 [PMID: 23946762 DOI: 10.5049/EBP.2013.11.1.24]
- 57 **Chen Y**, Rao F, Rodriguez-Flores JL, Mahata M, Fung MM, Stridsberg M, Vaingankar SM, Wen G, Salem RM, Das M, Cockburn MG, Schork NJ, Ziegler MG, Hamilton BA, Mahata SK, Taupenot L, O'Connor DT. Naturally occurring human genetic variation in the 3'-untranslated region of the secretory protein chromogranin A is associated with autonomic blood pressure regulation and hypertension in a sex-dependent fashion. *J Am Coll Cardiol* 2008; **52**: 1468-1481 [PMID: 19017515 DOI: 10.1016/j.jacc.2008.07.047]
- 58 **Huang C**, Zhang S, Hu K, Ma Q, Yang T. Phenylethanolamine N-methyltransferase gene promoter haplotypes and risk of essential hypertension. *Am J Hypertens* 2011; **24**: 1222-1226 [PMID: 21866188 DOI: 10.1038/ajh.2011.124]
- 59 **Zhao Q**, Fan Z, He J, Chen S, Li H, Zhang P, Wang L, Hu D, Huang J, Qiang B, Gu D. Renalase gene is a novel susceptibility gene for essential hypertension: a two-stage association study in northern Han Chinese population. *J Mol Med (Berl)* 2007; **85**: 877-885 [PMID: 17216203]
- 60 **Li X**, Jiang W, Li L, Huang R, Yang Q, Yang Y, Hong Y, Tang X. Renalase gene polymorphism in patients with hypertension and concomitant coronary heart disease. *Kidney Blood Press Res* 2014; **39**: 9-16 [PMID: 24821235 DOI: 10.1159/000355771]
- 61 **Htun NC**, Miyaki K, Song Y, Ikeda S, Shimbo T, Muramatsu M. Association of the catechol-O-methyl transferase gene Val158Met polymorphism with blood pressure and prevalence of hypertension: interaction with dietary energy intake. *Am J Hypertens* 2011; **24**: 1022-1026 [PMID: 21776034 DOI: 10.1038/ajh.2011.93]
- 62 **Zhou H**, Yang X, Liu Z, Yang T. Association of catechol-O-methyltransferase Val 108/158 Met polymorphism with essential hypertension. *Zhongnan Daxue Xuebao Yixueban* 2014; **39**: 790-796 [PMID: 25202947 DOI: 10.3969/j.issn.1672-7347.2014.08.006]
- 63 **Hagen K**, Pettersen E, Stovner LJ, Skorpen F, Holmen J, Zwart JA. High systolic blood pressure is associated with Val/Val genotype in the catechol-o-methyltransferase gene. The Nord-Trøndelag Health Study (HUNT). *Am J Hypertens* 2007; **20**: 21-26 [PMID: 17198907 DOI: 10.1016/j.amjhyper.2006.05.023]
- 64 **Siffert W**, Roskopf D, Siffert G, Busch S, Moritz A, Erbel R, Sharma AM, Ritz E, Wichmann HE, Jakobs KH, Horsthemke B. Association of a human G-protein beta3 subunit variant with hypertension. *Nat Genet* 1998; **18**: 45-48 [PMID: 9425898 DOI: 10.1038/ng0198-45]
- 65 **Pietruck F**, Moritz A, Montemurro M, Sell A, Busch S, Roskopf D, Virchow S, Esche H, Brockmeyer N, Jakobs KH, Siffert W. Selectively enhanced cellular signaling by Gi proteins in essential hypertension. G alpha i2, G alpha i3, G beta 1, and G beta 2 are not mutated. *Circ Res* 1996; **79**: 974-983 [PMID: 8888689 DOI: 10.1161/01.RES.79.5.974]
- 66 **Iiri T**, Bourne HR. G proteins propel surprise. *Nat Genet* 1998; **18**: 8-10 [PMID: 9425887 DOI: 10.1038/ng0198-8]
- 67 **Benjafield AV**, Jeyasingam CL, Nyholt DR, Griffiths LR, Morris BJ. G-protein beta3 subunit gene (GNB3) variant in causation of essential hypertension. *Hypertension* 1998; **32**: 1094-1097 [PMID: 9856980 DOI: 10.1161/01.HYP.32.6.1094]
- 68 **Beige J**, Hohenbleicher H, Distler A, Sharma AM. G-Protein beta3 subunit C825T variant and ambulatory blood pressure in essential hypertension. *Hypertension* 1999; **33**: 1049-1051 [PMID: 10205246 DOI: 10.1161/01.HYP.33.4.1049]
- 69 **Hengstenberg C**, Schunkert H, Mayer B, Döring A, Löwel H, Hense HW, Fischer M, Riegger GA, Holmer SR. Association between a polymorphism in the G protein beta3 subunit gene (GNB3) with arterial hypertension but not with myocardial infarction. *Cardiovasc Res* 2001; **49**: 820-827 [PMID: 11230982 DOI: 10.1016/S0008-6363(00)00292-3]
- 70 **Schunkert H**, Hense HW, Döring A, Riegger GA, Siffert W. Association between a polymorphism in the G protein beta3 subunit gene and lower renin and elevated diastolic blood pressure levels. *Hypertension* 1998; **32**: 510-513 [PMID: 9740618 DOI: 10.1161/01.HYP.32.3.510]
- 71 **Cabadak H**, Orun O, Nacar C, Dogan Y, Guneyel O, Fak AS, Kan B. The role of G protein  $\beta 3$  subunit polymorphisms C825T, C1429T, and G5177A in Turkish subjects with essential hypertension. *Clin Exp Hypertens* 2011; **33**: 202-208 [PMID: 21473734 DOI: 10.3109/10641963.2010.531855]
- 72 **Mega Tiber P**, Orun O, Kocakaya O, Cabadak H, Nacar C, Fak AS, Kan B. Association of Agt, Gnb3 and Enos gene polymorphisms with cardiovascular parameters in a Turkish population with essential hypertension. *Exp Clin Cardiol* 2014; **20**: 4606-4612
- 73 **Harris RC**. Abnormalities in renal dopamine signaling and hypertension: the role of GRK4. *Curr Opin Nephrol Hypertens* 2012; **21**: 61-65 [PMID: 22123211 DOI: 10.1097/MNH.0b013e32834de2cb]
- 74 **Muskalla AM**, Suter PM, Saur M, Nowak A, Hersberger M, Krayenbuehl PA. G-protein receptor kinase 4 polymorphism and response to antihypertensive therapy. *Clin Chem* 2014; **60**: 1543-1548 [PMID: 25301854 DOI: 10.1373/clinchem.2014.226605]
- 75 **Speirs HJ**, Katyk K, Kumar NN, Benjafield AV, Wang WY, Morris BJ. Association of G-protein-coupled receptor kinase 4 haplotypes, but not HSD3B1 or PTP1B polymorphisms, with essential hypertension. *J Hypertens* 2004; **22**: 931-936 [PMID: 15097232 DOI: 10.1097/01.hjh.0000098298.36684.3f]
- 76 **Zhu H**, Lu Y, Wang X, Treiber FA, Harshfield GA, Snieder H, Dong Y. The G protein-coupled receptor kinase 4 gene affects blood pressure in young normotensive twins. *Am J Hypertens* 2006; **19**: 61-66 [PMID: 16461192]
- 77 **Mancia G**, De Backer G, Dominiczak A, Cifkova R, Fagard R, Germano G, Grassi G, Heagerty AM, Kjeldsen SE, Laurent S, Narkiewicz K, Ruilope L, Rynkiewicz A, Schmieder RE, Struijker Boudier HA, Zanchetti A, Vahanian A, Camm J, De Caterina R, Dean V, Dickstein K, Filippatos G, Funck-Brentano C, Hellemans I, Kristensen SD, McGregor K, Sechtem U, Silber S, Tendera M, Widimsky P, Zamorano JL, Kjeldsen SE, Erdine S, Narkiewicz K, Kiowski W, Agabiti-Rosei E, Ambrosioni E, Cifkova R, Dominiczak A, Fagard R, Heagerty AM, Laurent S, Lindholm LH, Mancia G, Manolis A, Nilsson PM, Redon J, Schmieder RE,

- Struijker-Boudier HA, Viigimaa M, Filippatos G, Adamopoulos S, Agabiti-Rosei E, Ambrosioni E, Bertomeu V, Clement D, Erdine S, Farsang C, Gaita D, Kiowski W, Lip G, Mallion JM, Manolis AJ, Nilsson PM, O'Brien E, Ponikowski P, Redon J, Ruschitzka F, Tamargo J, van Zwieten P, Viigimaa M, Waeber B, Williams B, Zamorano JL. 2007 Guidelines for the management of arterial hypertension: The Task Force for the Management of Arterial Hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). *Eur Heart J* 2007; **28**: 1462-1536 [PMID: 17562668 DOI: 10.1093/eurheartj/ehm236]
- 78 **Mancia G**, Fagard R, Narkiewicz K, Redon J, Zanchetti A, Böhm M, Christiaens T, Cifkova R, De Backer G, Dominiczak A, Galderisi M, Grobbee DE, Jaarsma T, Kirchhof P, Kjeldsen SE, Laurent S, Manolis AJ, Nilsson PM, Ruilope LM, Schmieder RE, Sirnes PA, Sleight P, Viigimaa M, Waeber B, Zannad F, Redon J, Dominiczak A, Narkiewicz K, Nilsson PM, Burnier M, Viigimaa M, Ambrosioni E, Caulfield M, Coca A, Olsen MH, Schmieder RE, Tsioufis C, van de Borne P, Zamorano JL, Achenbach S, Baumgartner H, Bax JJ, Bueno H, Dean V, Deaton C, Erol C, Fagard R, Ferrari R, Hasdai D, Hoes AW, Kirchhof P, Knuuti J, Kolh P, Lancellotti P, Linhart A, Nihoyannopoulos P, Piepoli MF, Ponikowski P, Sirnes PA, Tamargo JL, Tendera M, Torbicki A, Wijns W, Windecker S, Clement DL, Coca A, Gillebert TC, Tendera M, Rosei EA, Ambrosioni E, Anker SD, Bauersachs J, Hitij JB, Caulfield M, De Buyzere M, De Geest S, Derumeaux GA, Erdine S, Farsang C, Funck-Brentano C, Gerc V, Germano G, Gielen S, Haller H, Hoes AW, Jordan J, Kahan T, Komajda M, Lovic D, Mahrholdt H, Olsen MH, Ostergren J, Parati G, Perk J, Polonia J, Popescu BA, Reiner Z, Rydén L, Sirenko Y, Stanton A, Struijker-Boudier H, Tsioufis C, van de Borne P, Vlachopoulos C, Volpe M, Wood DA. 2013 ESH/ESC guidelines for the management of arterial hypertension: the Task Force for the Management of Arterial Hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). *Eur Heart J* 2013; **34**: 2159-2219 [PMID: 23771844 DOI: 10.1093/eurheartj/ehs151]
- 79 **Scicchitano P**, Gesualdo M, Carbonara S, Palmiero P, Nazzaro P, Zito A, Ricci G, de Gennaro L, Caldarola P, Cortese F, Ciccone MM. What's New and What Gaps in 2013 European Guidelines for the Management of Arterial Hypertension: A Reappraisal. *Cardiol Angiol: An Int J* 2015; **3**: 181-191 [DOI: 10.9734/CA/2015/17967]

**P- Reviewer:** Ahmed MN, Ciccone MM **S- Editor:** Ji FF  
**L- Editor:** A **E- Editor:** Wu HL



## Management of hypertension: Current state of the art and challenges

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Conflict-of-interest statement: The authors declare that they no conflict of interest.

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Received: September 30, 2015  
Peer-review started: October 8, 2015  
First decision: November 30, 2015  
Revised: December 4, 2015  
Accepted: January 16, 2016  
Article in press: January 18, 2016  
Published online: February 23, 2016

### Abstract

Hypertension is a major modifiable cardiovascular risk factor. Hypertension is also recognized as the most

important risk factor for global disease burden. It is well established that a sustained reduction in blood pressure by drugs reduces the incidence of cardiovascular morbidity and mortality. In recent years, studies and new guidelines published for the management of hypertension. Awareness, treatment and control of hypertension are very poor, despite the new guidelines. We highlighted the management of hypertension in the light of current literature.

**Key words:** Hypertension; Therapy; Blood pressure; Cardiovascular risk factor

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**Core tip:** Hypertension is a major modifiable cardiovascular risk factor. It is well established that a sustained reduction in blood pressure by drugs reduces the incidence of cardiovascular morbidity and mortality. There are several types of drugs that can be used in the management of hypertension. But, the ideal treatment strategy remains uncertain for such a common and treatable disease. In recent years, studies and new guidelines were published addressing management of hypertension. Despite new guidelines, awareness, treatment and control of hypertension are very poor. We highlighted the management of hypertension in the light of current literature.

Turgut F, Yaprak M, Abdel-Rahman E. Management of hypertension: Current state of the art and challenges. *World J Hypertens* 2016; 6(1): 53-59 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/53.htm> DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.53>

### INTRODUCTION

Hypertension, a major contributor to cardiovascular



complications and premature death, is a modifiable cardiovascular risk factor<sup>[1]</sup>. Several studies have demonstrated that high blood pressure (BP) has a strong positive association with cardiovascular morbidity and mortality<sup>[2,3]</sup>. Hypertension is remarkably common across the world and its prevalence is strongly influenced by age and lifestyle factors<sup>[4,5]</sup>. Management of hypertension is especially important as hypertension is well recognized as the most important risk factor for global disease burden.

It is well established that treatment of hypertension reduces the risk of cardiovascular morbidity and mortality<sup>[6,7]</sup>. In contrast, untreated or poorly controlled hypertension is associated with permanent morbidity and mortality. The ultimate goal of antihypertensive therapy is the reduction of cardiovascular morbidity and mortality. There are several types of drugs that can be used in the management of hypertension. Yet, the ideal treatment strategy remains uncertain for such a common and treatable condition. There are new evidences regarding the management of hypertension. More recently, the European Society of Hypertension (ESH) and the European Society of Cardiology (ESC) published joint hypertension guidelines in 2013<sup>[8]</sup>. The panel members who were appointed to the Eighth Joint National Committee (JNC) also published the 2014 JNC report<sup>[9]</sup>. While these were in agreement on many points with previous guidelines, there were some important differences. This review highlights the management of hypertension in the light of current literature.

## NON-PHARMACOLOGICAL THERAPY

### *Lifestyle changes*

All guidelines recommend that the management of hypertension should start with life style modification<sup>[8-10]</sup>. Several lifestyle interventions have been shown to reduce BP<sup>[11,12]</sup>. Beside reducing high BP, these strategies are beneficial in managing most of the other cardiovascular risk factors<sup>[13]</sup>. Lifestyle changes recommended by the current guidelines include several interventions and combination of all these interventions. This has not changed as compared to previous guidelines (Table 1). It is generally believed that BP lowering effect of lifestyle modification is equivalent to drug monotherapy and can also delay drug therapy in patients with stage 1 hypertension.

### *Renal nerve denervation*

The sympathetic nervous system seems to play an important role in resistant hypertension<sup>[14]</sup>. Two clinical trials (Symplicity HTN 1 and Symplicity HTN 2) have shown the efficacy of renal sympathetic denervation with a post-procedure decline of 27/17 mmHg at 12 mo and 32/12 mmHg at 6 mo, respectively, with few minor adverse events<sup>[15,16]</sup>. Most recently, results of Symplicity HTN-3 (Renal Denervation in Patients with Uncontrolled Hypertension) trial showed no further reduction in office or ambulatory BP after 1-year follow up<sup>[17]</sup>. It seems

**Table 1 Recommended lifestyle modifications for the management of hypertension**

Weight loss (in obese or overweight patients)
Salt reduction
Regular exercise
Moderation of alcohol consumption
Smoking cessation
Increased consumption of vegetables, fruits and low-fat dairy products

that renal denervation is safe but has no superior BP lowering effects compared with adjustment of drug treatment<sup>[18]</sup>. In contrast, another more recent study, The Renal Denervation for Hypertension trial, showed that renal denervation plus standardized stepped-care antihypertensive therapy decreases BP more than the same standardized stepped-care antihypertensive therapy alone at 6 mo in patients with well-defined resistant hypertension<sup>[19]</sup>. So far, conflicting BP lowering effects of renal denervation have been reported. Thus, further studies are needed to reinforce renal denervation as a treatment modality for hypertension.

## PHARMACOLOGICAL THERAPY

Despite the non-pharmacological intervention, if BP is still above target, drug therapy should be initiated. There are five major classes of antihypertensive drugs: angiotensin converting enzyme (ACE) inhibitors, angiotensin receptor blockers (ARBs), diuretics, calcium channel blockers and  $\beta$ -blockers. In general, these drugs rarely have serious side effects when appropriately initiated and adequately monitored. We will not focus on the safety profile of these drugs, as it is beyond the scope of this review. According to current ESH/ESC hypertension guideline, grade 1 hypertensive patients with low/moderate cardiovascular risk can initially be treated with monotherapy<sup>[8]</sup>. On the other hand, JNC-8 panel based their recommendation based on the age of the patients. They recommended initiating therapy to lower BP at systolic BP  $\geq 150$  mmHg and diastolic BP  $\geq 90$  mmHg for patients aged  $\geq 60$  years, and systolic BP  $\geq 140$  mmHg and diastolic BP  $\geq 90$  mmHg for patients aged  $< 60$  years<sup>[9]</sup>.

## MONOTHERAPY

After a long waiting time, recently JNC-8 report (recommendations from only randomized controlled trials) was published. In the same line with 2013 ESH/ESC and National Institute for Health and Clinical Excellence (NICE) hypertension guideline, JNC-8 no longer recommends only thiazide-type diuretics as the initial therapy in most patients. As initial therapy, a thiazide-type diuretic, calcium channel blocker, ACE inhibitor and ARB can be started for uncomplicated hypertension<sup>[9]</sup>. The 2014 JNC report dismissed  $\beta$ -blockers as first-line therapy. Along the same line, the NICE clinical guideline did not recommend the first-

line use of diuretics and  $\beta$ -blockers<sup>[20]</sup>. Nevertheless, the 2013 ESH/ESC hypertension guidelines kept all 5 major classes of drugs in their recommendations as first-line regimens because of their opinions that the main benefits of antihypertensive therapy are due to lowering BP *per se* and largely independent of the drugs employed<sup>[8]</sup>.

BP control seems to be more important than a specific agent used to achieve that control. In a recent metaanalysis of 18 trials of 23215 Asian patients, a 10 mmHg reduction in systolic BP was associated with a 39.5% reduction in composite cardiovascular endpoints, and a 30% reduction in stroke, regardless of drug class<sup>[21]</sup>. Similarly, in a more recent metaanalysis of trials comparing the renin angiotensin aldosterone system (RAS) inhibitors vs other antihypertensive drugs as first-line therapy in patients with primary hypertension, all-cause mortality was similar between these drugs<sup>[22]</sup>. Still, the choice of drugs may be influenced by other factors as age, ethnicity/race, and other clinical characteristics. Trials in special patient groups (patients with diabetes, coronary artery disease, chronic kidney disease and proteinuria) have proposed that a specific drug group or combinations of certain drugs might be superior to others<sup>[8-10]</sup>. Thus, patients with special conditions should be considered to start with an appropriate drug based on their comorbidities (Table 2). We think that most hypertensive patients have comorbidities and initiating antihypertensive therapy generally requires compelling indications to select a specific drug group. Otherwise, in absence of comorbidity, it appears that the mere control of BP is more important than the class of antihypertensive drug being used.

## RAS BLOCKERS

Based on a large body of evidence, RAS blockers have been used to decrease the incidence of end-organ damage and cardiovascular mortality<sup>[23-25]</sup>. ACE inhibitors and ARBs can be considered first-line therapy in the management of hypertension, particularly in patients with diabetes mellitus. However, more recent studies showed that ACE inhibitors and ARBs do not have similar effects on cardiovascular outcomes and total mortality.

A metaanalysis of 20 clinical trials involving 158998 patients examined the effect of ACE inhibitors and ARBs in patients with hypertension<sup>[26]</sup>. ACE inhibitors significantly reduced all-cause mortality (HR = 0.90; 95%CI: 0.84-0.97;  $P = 0.04$ ) whereas ARBs did not (HR = 0.99; 95%CI: 0.94-1.04;  $P = 0.683$ ). In a metaanalysis evaluating the effects of ACE inhibitors and ARBs on all-cause mortality, cardiovascular events and deaths in patients with diabetes mellitus, ACE inhibitors reduced all-cause mortality (RR = 0.87; 95%CI: 0.78-0.98), cardiovascular mortality (RR = 0.83; 95%CI: 0.70-0.99), and major cardiovascular events (RR = 0.86; 95%CI: 0.77-0.95), whereas ARBs

**Table 2** Drugs to be preferred in patients with special conditions

Comorbidity	First-line therapy
Ischemic heart disease	$\beta$ -blocker (unless contraindicated) Long-acting calcium channel blocker ACE inhibitors (ARBs if ACE inhibitors not tolerated)
Heart failure	ACE inhibitors (ARBs if ACE inhibitors not tolerated) $\beta$ -blockers Aldosterone antagonists
Diabetes	ACE inhibitors (ARBs if ACE inhibitors not tolerated) $\beta$ -blockers Calcium channel blockers
Chronic kidney disease	ACE inhibitors or ARBs Loop diuretics rather than a thiazide diuretic (or combination)
Stroke	Diuretic + ACE inhibitors
Asymptomatic organ damage	
Left ventricular hypertrophy	ACE inhibitors, ARBs, Calcium channel blockers
Proteinuria	ACE inhibitors, ARBs

ACE: Angiotensin converting enzyme; ARBs: Angiotensin II receptor blockers.

had no benefits on these outcomes<sup>[27]</sup>. In another metaanalysis that included nine randomized controlled trials, no difference was found in total mortality or cardiovascular outcomes for ARBs as compared with ACE inhibitors<sup>[28]</sup>. According to these evidences, we can conclude that while ACE inhibitors can be used as a first-line therapy, ARBs are preferred for patients who have adverse reactions to ACE inhibitors although there is no agreement among the guidelines.

## DIURETICS

Thiazide and thiazide like diuretics (*e.g.*, indapamide, chlorthalidone) remain essential in the management of hypertension. The JNC-7 recommended that thiazide diuretics should be the preferred drugs in most hypertensive patients, either alone or combined with other classes of drugs<sup>[10]</sup>. Although it is well known that thiazide-type diuretics are effective in reducing BP and preventing cardiovascular disease in hypertensive subjects, it is not clear whether all drugs in this class are equally safe and effective. Recently, the choice of diuretics has emerged as a controversial issue with some evidence favoring the long-acting agent, chlorthalidone, in preference to hydrochlorothiazide. A recent retrospective observational cohort analysis from the Multiple Risk Factor Intervention Trial data set compared the effects of chlorthalidone vs hydrochlorothiazide on cardiovascular event rates<sup>[29]</sup>. Chlorthalidone treatment was associated with significantly fewer cardiovascular events; lower systolic BP, potassium, and total and low-density lipoprotein cholesterol levels; and significantly higher uric acid levels compared with hydrochlorothiazide.

A large observational study with up to five years of follow up reported head-to-head comparative data

on the effects of newly prescribed chlorthalidone vs hydrochlorothiazide on cardiovascular and safety outcomes in elderly patients<sup>[30]</sup>. Chlorthalidone was not associated with fewer adverse cardiovascular events or deaths than hydrochlorothiazide in elderly patients; however, it was associated with a greater incidence of electrolyte abnormalities, particularly hypokalemia.

In a recent meta-analysis of 14 trials comparing head to head thiazide-like and thiazide-type diuretics, systolic BP reduction was greater with chlorthalidone and indapamide without more adverse effects<sup>[31]</sup>. These data suggest using chlorthalidone as preferred thiazide type diuretic for the management of hypertension. On the other hand, hydrochlorothiazide has a dose related BP-lowering effect and greater effect on systolic BP than diastolic BP, thus lowering pulse pressure more than other antihypertensive drugs<sup>[32]</sup>. We believe that it is too early to reach a final conclusion, as there are no randomized trials that directly compare cardiovascular outcomes in hypertensive patients treated with thiazide-type diuretics vs thiazide-like diuretics.

Mineralocorticoid receptors have been shown to play important roles in the pathogenesis of hypertension and hypertension-related cardiovascular outcomes<sup>[33-35]</sup>. Recent studies have implicated that aldosterone excess as an important pathophysiologic factor in a large fraction of patients with resistant hypertension<sup>[36]</sup>. Spironolactone can be tried in patients with resistant hypertension requiring three or more drugs to achieve BP control unless contraindicated<sup>[20]</sup>. Eplerenone may be used as an alternative in patients who experience hormonally related side effects with spironolactone.

We conclude that diuretics remain as leading agents in the management of hypertension. Based on the available data, thiazide-like diuretics (such as chlorthalidone, 12.5 to 25 mg/d) may be preferred to thiazide type diuretics. Moreover, when BP cannot be controlled with other drugs, combining thiazide-like diuretics with ACE inhibitors or ARBs are usually very effective. Combining diuretics with aldosterone antagonists may also be worthwhile in special patient population.

## CALCIUM CHANNEL BLOCKERS

Calcium channel blockers have potent BP-lowering effects and have been the most widely used antihypertensive drugs. Several studies have showed that calcium channel blockers had efficacy not only in lowering BP but also in reducing cardiovascular morbidity and mortality in patients with hypertension<sup>[37]</sup>. In a recent meta-analysis of 31 randomized controlled trials, calcium channel blockers reduced stroke more than either placebo (OR = 0.68; 95%CI: 0.61-0.75) or  $\beta$ -blockers (OR = 0.79; 95%CI: 0.72-0.87), but was not different from ACE inhibitors and diuretics<sup>[38]</sup>. Another Cochrane meta-analysis of randomized trials comparing first-line calcium channel blockers with other antihypertensive classes did not find difference among

calcium channel blockers, ACE inhibitors or ARBs in terms of all-cause mortality, however, it provided evidence supporting the use of calcium channel blockers over  $\beta$ -blockers in terms of total cardiovascular events, stroke and cardiovascular mortality<sup>[39]</sup>. Calcium channel blockers are broadly classified into two groups as dihydropyridine and non-dihydropyridine groups. Non-dihydropyridine calcium channel blockers are more negatively chronotropic and inotropic than the dihydropyridine subclasses, and are generally not recommended to use as first-line therapy in the management of hypertension. The NICE guidelines recommend particularly calcium channel blockers as first-line therapy in hypertensive patients aged over 55 years<sup>[20]</sup>. We conclude that calcium channel blockers may be used as initial first-line therapy particularly in hypertensive patients without compelling co-morbidities or as a component of combination therapy.

## $\beta$ -BLOCKERS

Whether  $\beta$ -blockers should be placed as first-line therapy in the management of hypertension is probably the most controversial issue among major guidelines. Some do not recommend  $\beta$ -blockers as first-line therapy for hypertension<sup>[9,20]</sup>. But, the 2013 ESH/ESC guidelines continued to recommend  $\beta$ -blockers as one of the first-line anti-hypertensive drugs<sup>[8]</sup>. On the other hand, the 2014 NICE hypertension guidelines put  $\beta$ -blockers as step 4 drugs.  $\beta$ -blockers can be used as additional therapy to further lower BP, but they may have a special benefit in preventing recurrent coronary artery disease<sup>[7]</sup>.

The class of  $\beta$ -blockers is heterogeneous, and all the drugs in this class may not be the same<sup>[40]</sup>. Atenolol, metoprolol, carvedilol and nebivolol have different properties in terms of efficacy and side effects. But a recent meta-analysis comparing atenolol and non-atenolol  $\beta$ -blockers found that  $\beta$ -blockers had similar effect on cardiovascular end points in hypertensive patients without compelling indications<sup>[41]</sup>. Only, in the elderly (> 60 years), atenolol was inferior to the other drugs in reducing stroke. We conclude that while  $\beta$ -blockers remain the standard of care for patients with coronary artery disease, particularly after acute myocardial infarction<sup>[42]</sup>, their role in the management of hypertension without coronary artery disease remains controversial.

## COMBINATION THERAPY

Combination therapy may have benefit patients through multiple and potentially complementary pharmacologic mechanisms of action. Thus, combining drugs with different classes may be more effective than titrating dose of a single agent. Initiating treatment with a drug combination rather than a single agent is increasingly utilized as a therapeutic strategy. According to the current guidelines, a large majority of patients

require simultaneous administration of two or more antihypertensive drugs to reach the target BP<sup>[8-10]</sup>. Specifically in patients with stage 2 hypertension, it appears that early combination therapy may lower BP to targets sooner.

### Choice of combination therapy

A diuretic,  $\beta$ -blocker, calcium channel blocker and ACE inhibitors (or ARBs) can be combined in the management of hypertension. Amongst the various combinations of antihypertensive drugs, it is generally considered that combining an ACE inhibitor or ARB with a diuretic produces fully additive BP reduction<sup>[43]</sup>. Combination of antihypertensive agents as initial therapy in stage 2 hypertension can lead to markedly improved BP control in patients as compared with mono-therapeutic regimens<sup>[10,44]</sup>.

Recently, it was noted that the addition of an ACE inhibitor or ARB to a dihydropyridine calcium-channel blocker is increasingly being used<sup>[45]</sup>. The only trial (ACCOMPLISH) comparing ACE inhibitor-calcium channel blocker combination and ACE inhibitor-diuretic combination found significant superiority of ACE inhibitor-calcium channel blocker over diuretic combination<sup>[46]</sup>. This combination may also reduce the incidence and severity of edema caused by calcium channel blocker. The combination of ACE inhibitor (or ARB) in addition to diuretic or calcium channel blocker may be used as initial combination therapy. But, it appears that calcium channel blockers are better than diuretics as a component in combination therapy.

### Combination of RAS blockers

Currently, the combination of an ACE inhibitor and ARB is not recommended<sup>[9]</sup>. Recent studies showed that combination therapy did not prove to be superior to the use of an ACE inhibitor or ARB alone in reducing the primary or secondary outcomes. Previously, in the Ongoing Telmisartan Alone and in Combination with Ramipril Global Endpoint Trial, a randomized study of combination therapy vs monotherapy in persons at increased cardiovascular risk, no cardiovascular or renal benefits were observed with combination therapy<sup>[47,48]</sup>. In a recent randomized, controlled study, combination therapy with ACE inhibitors and ARBs provided no benefit to major primary and secondary outcomes in patients with diabetic nephropathy<sup>[49]</sup>. Furthermore, patients receiving combination therapy had an excess occurrence of hyperkalemia and acute kidney injury. The addition of direct renin inhibitor, aliskiren, to standard therapy with ACE inhibitors or ARBs in patients with type 2 diabetes did not reduce cardiovascular or renal outcomes as compared with placebo. On the contrary, the combination therapy resulted in an increased number of adverse events<sup>[50]</sup>.

## CONCLUSION

In conclusion, based on the available data, the amount

of BP reduction rather than the choice of antihypertensive drug are the major determinant of reduction in cardiovascular risk in patients with hypertension. But, some hypertensive patients may have compelling indication for a specific antihypertensive drug, which may offer particular benefit independent of BP control. Successful treatment requires identification and reversal of lifestyle factors contributing to treatment resistance; diagnosis and appropriate treatment of secondary causes of hypertension; and use of effective combination regimens. Combination therapy may be necessary in the majority of the patients with hypertension, and current guidelines recommend routine initiation of a combination in patients with stage 2 hypertension. ACE inhibitors, ARBs and diuretics including aldosterone antagonists can result in clinically significant alterations of serum electrolytes and kidney function. Thus, after the initiation of these agents, a chemistry profile should be obtained.

## REFERENCES

- 1 **Cupples LA**, D'Agostino RB. Some risk factors related to the annual incidence of cardiovascular disease and death using pooled repeated biennial measurements: Framingham Heart Study, 30-year followup. The Framingham Study: An Epidemiological Investigation of Cardiovascular Disease. In: Kannel WB, Wolf PA, Garrison RJ. Washington DC: Government Printing Office, 1987: 9-20
- 2 **van den Hoogen PC**, Feskens EJ, Nagelkerke NJ, Menotti A, Nissinen A, Kromhout D. The relation between blood pressure and mortality due to coronary heart disease among men in different parts of the world. Seven Countries Study Research Group. *N Engl J Med* 2000; **342**: 1-8 [PMID: 10620642 DOI: 10.1056/NEJM200001063420101]
- 3 **Stamler J**, Stamler R, Neaton JD. Blood pressure, systolic and diastolic, and cardiovascular risks. US population data. *Arch Intern Med* 1993; **153**: 598-615 [PMID: 8439223 DOI: 10.1001/archinte.1993.00410050036006]
- 4 **Turgut F**, Yesil Y, Balogun RA, Abdel-Rahman EM. Hypertension in the elderly: unique challenges and management. *Clin Geriatr Med* 2013; **29**: 593-609 [PMID: 23849010 DOI: 10.1016/j.cger.2013.05.002]
- 5 **Go AS**, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Blaha MJ, Dai S, Ford ES, Fox CS, Franco S, Fullerton HJ, Gillespie C, Hailpern SM, Heit JA, Howard VJ, Huffman MD, Judd SE, Kissela BM, Kittner SJ, Lackland DT, Lichtman JH, Lisabeth LD, Mackey RH, Magid DJ, Marcus GM, Marelli A, Matchar DB, McGuire DK, Mohler ER, Moy CS, Mussolino ME, Neumar RW, Nichol G, Pandey DK, Paynter NP, Reeves MJ, Sorlie PD, Stein J, Towfighi A, Turan TN, Virani SS, Wong ND, Woo D, Turner MB. Heart disease and stroke statistics--2014 update: a report from the American Heart Association. *Circulation* 2014; **129**: e28-e292 [PMID: 24352519 DOI: 10.1161/01.cir.0000441139.02102.80]
- 6 **Neal B**, MacMahon S, Chapman N. Effects of ACE inhibitors, calcium antagonists, and other blood-pressure-lowering drugs: results of prospectively designed overviews of randomised trials. Blood Pressure Lowering Treatment Trialists' Collaboration. *Lancet* 2000; **356**: 1955-1964 [PMID: 11130523 DOI: 10.1016/S0140-6736(00)03307-9]
- 7 **Law MR**, Morris JK, Wald NJ. Use of blood pressure lowering drugs in the prevention of cardiovascular disease: meta-analysis of 147 randomised trials in the context of expectations from prospective epidemiological studies. *BMJ* 2009; **338**: b1665 [PMID: 19454737 DOI: 10.1136/bmj.b1665]
- 8 **Mancia G**, Fagard R, Narkiewicz K, Redán J, Zanchetti A, Böhm



- M, Christiaens T, Cifkova R, De Backer G, Dominiczak A, Galderisi M, Grobbee DE, Jaarsma T, Kirchhof P, Kjeldsen SE, Laurent S, Manolis AJ, Nilsson PM, Ruilope LM, Schmieder RE, Sirnes PA, Sleight P, Viigimaa M, Waeber B, Zannad F. 2013 Practice guidelines for the management of arterial hypertension of the European Society of Hypertension (ESH) and the European Society of Cardiology (ESC): ESH/ESC Task Force for the Management of Arterial Hypertension. *J Hypertens* 2013; **31**: 1925-1938 [PMID: 24107724 DOI: 10.1097/HJH.0b013e328364ca4c]
- 9 **James PA**, Oparil S, Carter BL, Cushman WC, Dennison-Himmelfarb C, Handler J, Lackland DT, LeFevre ML, MacKenzie TD, Oggedge O, Smith SC, Svetkey LP, Taler SJ, Townsend RR, Wright JT, Narva AS, Ortiz E. 2014 evidence-based guideline for the management of high blood pressure in adults: report from the panel members appointed to the Eighth Joint National Committee (JNC 8). *JAMA* 2014; **311**: 507-520 [PMID: 24352797 DOI: 10.1001/jama.2013.284427]
  - 10 **Chobanian AV**, Bakris GL, Black HR, Cushman WC, Green LA, Izzo JL, Jones DW, Materson BJ, Oparil S, Wright JT, Roccella EJ. The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. *JAMA* 2003; **289**: 2560-2572 [PMID: 12748199 DOI: 10.1001/jama.289.19.2560]
  - 11 **Sacks FM**, Svetkey LP, Vollmer WM, Appel LJ, Bray GA, Harsha D, Obarzanek E, Conlin PR, Miller ER, Simons-Morton DG, Karanja N, Lin PH. Effects on blood pressure of reduced dietary sodium and the Dietary Approaches to Stop Hypertension (DASH) diet. DASH-Sodium Collaborative Research Group. *N Engl J Med* 2001; **344**: 3-10 [PMID: 11136953 DOI: 10.1056/NEJM200101043440101]
  - 12 **Núñez-Córdoba JM**, Valencia-Serrano F, Toledo E, Alonso A, Martínez-González MA. The Mediterranean diet and incidence of hypertension: the Seguimiento Universidad de Navarra (SUN) Study. *Am J Epidemiol* 2009; **169**: 339-346 [PMID: 19037007 DOI: 10.1093/aje/kwn335]
  - 13 **Rees K**, Hartley L, Flowers N, Clarke A, Hooper L, Thorogood M, Stranges S. 'Mediterranean' dietary pattern for the primary prevention of cardiovascular disease. *Cochrane Database Syst Rev* 2013; **8**: CD009825 [PMID: 23939686 DOI: 10.1002/14651858.CD009825.pub2]
  - 14 **Nathan S**, Bakris GL. The future of renal denervation in resistant hypertension. *Curr Hypertens Rep* 2014; **16**: 494 [PMID: 25304105 DOI: 10.1007/s11906-014-0494-0]
  - 15 **Esler MD**, Krum H, Sobotka PA, Schlaich MP, Schmieder RE, Böhm M. Renal sympathetic denervation in patients with treatment-resistant hypertension (The Symplicity HTN-2 Trial): a randomised controlled trial. *Lancet* 2010; **376**: 1903-1909 [PMID: 21093036 DOI: 10.1016/S0140-6736(10)62039-9]
  - 16 **Krum H**, Schlaich M, Whitbourn R, Sobotka PA, Sadowski J, Bartus K, Kapelak B, Walton A, Sievert H, Thammar S, Abraham WT, Esler M. Catheter-based renal sympathetic denervation for resistant hypertension: a multicentre safety and proof-of-principle cohort study. *Lancet* 2009; **373**: 1275-1281 [PMID: 19332353 DOI: 10.1016/S0140-6736(09)60566-3]
  - 17 **Bakris GL**, Townsend RR, Flack JM, Brar S, Cohen SA, D'Agostino R, Kandzari DE, Katzen BT, Leon MB, Mauri L, Negoita M, O'Neill WW, Oparil S, Rocha-Singh K, Bhatt DL. 12-month blood pressure results of catheter-based renal artery denervation for resistant hypertension: the SYMPPLICITY HTN-3 trial. *J Am Coll Cardiol* 2015; **65**: 1314-1321 [PMID: 25835443 DOI: 10.1016/j.jacc.2015.01.037]
  - 18 **Fadi Elmula FE**, Hoffmann P, Larstorp AC, Fossum E, Brekke M, Kjeldsen SE, Gjønnæss E, Hjörnholm U, Kjaer VN, Rostrop M, Os I, Stenehjem A, Høieggan A. Adjusted drug treatment is superior to renal sympathetic denervation in patients with true treatment-resistant hypertension. *Hypertension* 2014; **63**: 991-999 [PMID: 24591332 DOI: 10.1161/HYPERTENSIONAHA.114.03246]
  - 19 **Azizi M**, Sapoval M, Gosse P, Monge M, Bobrie G, Delsart P, Midulla M, Mounier-Véhier C, Courand PY, Lantelme P, Denolle T, Dourmap-Collas C, Trillaud H, Pereira H, Plouin PF, Chatellier G. Optimum and stepped care standardised antihypertensive treatment with or without renal denervation for resistant hypertension (DENERHTN): a multicentre, open-label, randomised controlled trial. *Lancet* 2015; **385**: 1957-1965 [PMID: 25631070 DOI: 10.1016/S0140-6736(14)61942-5]
  - 20 Hypertension in adults: diagnosis and management. 2011-08. Available from: URL: <http://www.nice.org.uk/guidance/cg127/chapter/key-priorities-for-implementation#choosing-antihypertensive-drug-treatment>
  - 21 **Yano Y**, Briasoulis A, Bakris GL, Hoshida S, Wang JG, Shimada K, Kario K. Effects of antihypertensive treatment in Asian populations: a meta-analysis of prospective randomized controlled studies (CARDiovascular protectionN group in Asia: CARNA). *J Am Soc Hypertens* 2014; **8**: 103-116 [PMID: 24157055 DOI: 10.1016/j.jash.2013.09.002]
  - 22 **Xue H**, Lu Z, Tang WL, Pang LW, Wang GM, Wong GW, Wright JM. First-line drugs inhibiting the renin angiotensin system versus other first-line antihypertensive drug classes for hypertension. *Cochrane Database Syst Rev* 2015; **1**: CD008170 [PMID: 25577154 DOI: 10.1002/14651858.CD008170.pub2]
  - 23 **Brenner BM**, Cooper ME, de Zeeuw D, Keane WF, Mitch WE, Parving HH, Remuzzi G, Snapinn SM, Zhang Z, Shahinfar S. Effects of losartan on renal and cardiovascular outcomes in patients with type 2 diabetes and nephropathy. *N Engl J Med* 2001; **345**: 861-869 [PMID: 11565518 DOI: 10.1056/NEJMoa011161]
  - 24 **Ruggenenti P**, Perna A, Remuzzi G. ACE inhibitors to prevent end-stage renal disease: when to start and why possibly never to stop: a post hoc analysis of the REIN trial results. Ramipril Efficacy in Nephropathy. *J Am Soc Nephrol* 2001; **12**: 2832-2837 [PMID: 11729254]
  - 25 **Turgut F**, Balogun RA, Abdel-Rahman EM. Renin-angiotensin-aldosterone system blockade effects on the kidney in the elderly: benefits and limitations. *Clin J Am Soc Nephrol* 2010; **5**: 1330-1339 [PMID: 20498247 DOI: 10.2215/CJN.08611209]
  - 26 **van Vark LC**, Bertrand M, Akkerhuis KM, Brugts JJ, Fox K, Mourad JJ, Boersma E. Angiotensin-converting enzyme inhibitors reduce mortality in hypertension: a meta-analysis of randomized clinical trials of renin-angiotensin-aldosterone system inhibitors involving 158,998 patients. *Eur Heart J* 2012; **33**: 2088-2097 [PMID: 22511654 DOI: 10.1093/eurheartj/ehs075]
  - 27 **Cheng J**, Zhang W, Zhang X, Han F, Li X, He X, Li Q, Chen J. Effect of angiotensin-converting enzyme inhibitors and angiotensin II receptor blockers on all-cause mortality, cardiovascular deaths, and cardiovascular events in patients with diabetes mellitus: a meta-analysis. *JAMA Intern Med* 2014; **174**: 773-785 [PMID: 24687000 DOI: 10.1001/jamainternmed.2014.348]
  - 28 **Li EC**, Heran BS, Wright JM. Angiotensin converting enzyme (ACE) inhibitors versus angiotensin receptor blockers for primary hypertension. *Cochrane Database Syst Rev* 2014; **8**: CD009096 [PMID: 25148386 DOI: 10.1002/14651858.CD009096.pub2]
  - 29 **Dorsch MP**, Gillespie BW, Erickson SR, Bleske BE, Weder AB. Chlorthalidone reduces cardiovascular events compared with hydrochlorothiazide: a retrospective cohort analysis. *Hypertension* 2011; **57**: 689-694 [PMID: 21383313 DOI: 10.1161/HYPERTENSIONAHA.110.161505]
  - 30 **Dhalla IA**, Gomes T, Yao Z, Nagge J, Persaud N, Hellings C, Mamdani MM, Juurlink DN. Chlorthalidone versus hydrochlorothiazide for the treatment of hypertension in older adults: a population-based cohort study. *Ann Intern Med* 2013; **158**: 447-455 [PMID: 23552325 DOI: 10.7326/0003-4819-158-6-201303190-00004]
  - 31 **Roush GC**, Ernst ME, Kostis JB, Tandon S, Sica DA. Head-to-head comparisons of hydrochlorothiazide with indapamide and chlorthalidone: antihypertensive and metabolic effects. *Hypertension* 2015; **65**: 1041-1046 [PMID: 25733245 DOI: 10.1161/HYPERTENSIONAHA.114.05021]
  - 32 **Musini VM**, Nazer M, Bassett K, Wright JM. Blood pressure-lowering efficacy of monotherapy with thiazide diuretics for primary hypertension. *Cochrane Database Syst Rev* 2014; **5**: CD003824 [PMID: 24869750 DOI: 10.1002/14651858.CD003824.pub2]

- 33 **Calhoun DA**, Jones D, Textor S, Goff DC, Murphy TP, Toto RD, White A, Cushman WC, White W, Sica D, Ferdinand K, Giles TD, Falkner B, Carey RM. Resistant hypertension: diagnosis, evaluation, and treatment. A scientific statement from the American Heart Association Professional Education Committee of the Council for High Blood Pressure Research. *Hypertension* 2008; **51**: 1403-1419 [PMID: 18391085 DOI: 10.1161/HYPERTENSIONAHA.108.189141]
- 34 **Maron BA**, Leopold JA. Aldosterone receptor antagonists: effective but often forgotten. *Circulation* 2010; **121**: 934-939 [PMID: 20177008 DOI: 10.1161/CIRCULATIONAHA.109.895235]
- 35 **Vasan RS**, Evans JC, Larson MG, Wilson PW, Meigs JB, Rifai N, Benjamin EJ, Levy D. Serum aldosterone and the incidence of hypertension in nonhypertensive persons. *N Engl J Med* 2004; **351**: 33-41 [PMID: 15229305 DOI: 10.1056/NEJMoa033263]
- 36 **Byrd JB**, Brook RD. A critical review of the evidence supporting aldosterone in the etiology and its blockade in the treatment of obesity-associated hypertension. *J Hum Hypertens* 2014; **28**: 3-9 [PMID: 23698003 DOI: 10.1038/jhh.2013.42]
- 37 **Staessen JA**, Fagard R, Thijs L, Celis H, Arabidze GG, Birkenhäger WH, Bulpitt CJ, de Leeuw PW, Dolly CT, Fletcher AE, Forette F, Leonetti G, Nachev C, O'Brien ET, Rosenfeld J, Rodicio JL, Tuomilehto J, Zanchetti A. Randomised double-blind comparison of placebo and active treatment for older patients with isolated systolic hypertension. The Systolic Hypertension in Europe (Syst-Eur) Trial Investigators. *Lancet* 1997; **350**: 757-764 [PMID: 9297994 DOI: 10.1016/S0140-6736(97)05381-6]
- 38 **Chen GJ**, Yang MS. The effects of calcium channel blockers in the prevention of stroke in adults with hypertension: a meta-analysis of data from 273,543 participants in 31 randomized controlled trials. *PLoS One* 2013; **8**: e57854 [PMID: 23483932 DOI: 10.1371/journal.pone.0057854]
- 39 **Chen N**, Zhou M, Yang M, Guo J, Zhu C, Yang J, Wang Y, Yang X, He L. Calcium channel blockers versus other classes of drugs for hypertension. *Cochrane Database Syst Rev* 2010; **(8)**: CD003654 [PMID: 20687074 DOI: 10.1002/14651858.CD003654.pub4]
- 40 **Poirier L**, Tobe SW. Contemporary use of  $\beta$ -blockers: clinical relevance of subclassification. *Can J Cardiol* 2014; **30**: S9-S15 [PMID: 24684855 DOI: 10.1016/j.cjca.2013.12.001]
- 41 **Kuyper LM**, Khan NA. Atenolol vs nonatenolol  $\beta$ -blockers for the treatment of hypertension: a meta-analysis. *Can J Cardiol* 2014; **30**: S47-S53 [PMID: 24750981 DOI: 10.1016/j.cjca.2014.01.006]
- 42 **Bangalore S**, Steg G, Deedwania P, Crowley K, Eagle KA, Goto S, Ohman EM, Cannon CP, Smith SC, Zeymer U, Hoffman EB, Messerli FH, Bhatt DL.  $\beta$ -Blocker use and clinical outcomes in stable outpatients with and without coronary artery disease. *JAMA* 2012; **308**: 1340-1349 [PMID: 23032550 DOI: 10.1001/jama.2012.12559]
- 43 **Suzuki H**, Shimada K, Fujiwara K. Antihypertensive effectiveness of combination therapy with losartan/hydrochlorothiazide for 'real world' management of isolated systolic hypertension. *Ther Adv Cardiovasc Dis* 2015; **9**: 10-18 [PMID: 25367171 DOI: 10.1177/1753944714558244]
- 44 **Khanna A**, Lefkowitz L, White WB. Evaluation of recent fixed-dose combination therapies in the management of hypertension. *Curr Opin Nephrol Hypertens* 2008; **17**: 477-483 [PMID: 18695388 DOI: 10.1097/MNH.0b013e3283069d72]
- 45 **Makani H**, Bangalore S, Romero J, Wever-Pinzon O, Messerli FH. Effect of renin-angiotensin system blockade on calcium channel blocker-associated peripheral edema. *Am J Med* 2011; **124**: 128-135 [PMID: 21295192 DOI: 10.1016/j.amjmed.2010.08.007]
- 46 **Jamerson K**, Weber MA, Bakris GL, Dahlöf B, Pitt B, Shi V, Hester A, Gupta J, Gatlin M, Velazquez EJ. Benazepril plus amlodipine or hydrochlorothiazide for hypertension in high-risk patients. *N Engl J Med* 2008; **359**: 2417-2428 [PMID: 19052124 DOI: 10.1056/NEJMoa0806182]
- 47 **Mann JF**, Schmieder RE, McQueen M, Dyal L, Schumacher H, Pogue J, Wang X, Maggioni A, Budaj A, Chaitiraphan S, Dickstein K, Keltai M, Metsärinne K, Oto A, Parkhomenko A, Piegas LS, Svendsen TL, Teo KK, Yusuf S. Renal outcomes with telmisartan, ramipril, or both, in people at high vascular risk (the ONTARGET study): a multicentre, randomised, double-blind, controlled trial. *Lancet* 2008; **372**: 547-553 [PMID: 18707986 DOI: 10.1016/S0140-6736(08)61236-2]
- 48 **Yusuf S**, Teo KK, Pogue J, Dyal L, Copland I, Schumacher H, Dagenais G, Sleight P, Anderson C. Telmisartan, ramipril, or both in patients at high risk for vascular events. *N Engl J Med* 2008; **358**: 1547-1559 [PMID: 18378520 DOI: 10.1056/NEJMoa0801317]
- 49 **Fried LF**, Emanuele N, Zhang JH, Brophy M, Conner TA, Duckworth W, Leehey DJ, McCullough PA, O'Connor T, Palevsky PM, Reilly RF, Seliger SL, Warren SR, Watnick S, Peduzzi P, Guarino P. Combined angiotensin inhibition for the treatment of diabetic nephropathy. *N Engl J Med* 2013; **369**: 1892-1903 [PMID: 24206457 DOI: 10.1056/NEJMoa1303154]
- 50 **Parving HH**, Brenner BM, McMurray JJ, de Zeeuw D, Haffner SM, Solomon SD, Chaturvedi N, Persson F, Desai AS, Nicolaidis M, Richard A, Xiang Z, Brunel P, Pfeffer MA. Cardiorenal end points in a trial of aliskiren for type 2 diabetes. *N Engl J Med* 2012; **367**: 2204-2213 [PMID: 23121378 DOI: 10.1056/NEJMoa1208799]

**P-Reviewer:** Cheng TH, Salles G, Sanchez R, Tan XR, Zhao D

**S-Editor:** Qiu S **L-Editor:** A **E-Editor:** Wu HL



## Renal venous hypertension

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**Author contributions:** All authors equally contributed to this paper with conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

**Conflict-of-interest statement:** The authors declare no conflicts of interest regarding this manuscript.

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Received: August 3, 2015

Peer-review started: August 3, 2015

First decision: September 29, 2015

Revised: October 30, 2015

Accepted: December 29, 2015

Article in press: December 31, 2015

Published online: February 23, 2016

examination, diagnosis can be made with ultrasound, computed tomography, or magnetic resonance conventional venography. Symptoms and haemodynamic significance of the compression determine the ideal treatment method. This review of the literature discusses normal and pathological developmental aspects of renocaval venous segment and related circulatory disorders, summarizes congenital and acquired changes in left renal vein and their impact on development of renal venous hypertension. Also will be discussed surgical tactics of portosystemic shunting and their potential effects on renal hemodynamics.

**Key words:** Renal venous hypertension; Nutcracker syndrome; Kidney; Portal hypertension; Splenorenal shunts

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**Core tip:** Renal venous hypertension characterized by the presence of left renal vein dilatation, varicocele and hematuria. Being a rare cause of hematuria its etiology is diverse but of precise characteristics. Diagnosis is not easy and treatment requires ruling out its precise etiology and considering the intensity of the compression phenomenon because of interventionist attitudes have important implications and are not risk free.

Aliev MM, Yuldashev RZ, Adilova GS, Dekhqonboev AA. Renal venous hypertension. *World J Hypertens* 2016; 6(1): 60-65  
 Available from: URL: <http://www.wjgnet.com/2220-3168/full/v6/i1/60.htm> DOI: <http://dx.doi.org/10.5494/wjh.v6.i1.60>

## Abstract

Renal venous hypertension usually seen in young, otherwise healthy individuals and can lead to significant overall morbidity. Aside from clinical findings and physical

## INTRODUCTION

Renal venous hypertension (RVH) - venous insufficiency caused by inadequate drainage of blood through the renal vein<sup>[1]</sup>. There are two main reasons in the development of the RVH: Structural abnormalities of

renocaval segment; acquired changes of the left renal vein.

### **Structural anomalies of renocaval segment and their clinical importance in the development of the RVH**

Development of renocaval segment (inferior vena cava, renal, gonadal, adrenal and lumbar veins) - is a complex process in which there is consecutive regression and shifting of three venous structures, such as posterior cardinal, supracardinal and subcardinal veins<sup>[2-5]</sup>. Inferior vena cava (IVC) and its branches formed from different embryological structures; their segments formed from all three systems mentioned above. The formation of these veins can be impaired at any stage of development<sup>[2,3]</sup>.

Among the developmental abnormalities of the IVC clinical significance in development of the RVH matters - left-sided IVC, with this type of abnormality abdominal aorta compresses IVC at the site of their contact, thus will cause congestion in left renal vein (LRV) and recurrent left sided hematuria<sup>[6,7]</sup>.

Typical abnormality of the right renal vein is anomaly of their quantities, which is due to the fact, that the right renal vein embryogenesis does not undergo significant transformations. Essentially, the significance of these abnormalities in development of RVH negligible<sup>[2,3,8]</sup>.

Clinically significant abnormalities often observed in the LRV, which related to its development. For example, retention of both limbs of the left portion of circumaortic venous ring leads to the formation of the circumaortic LRV, which occurs in 1%-17% of cases according to different authors<sup>[2,3,5,8-13]</sup>. In this type of anomaly there are pre-aortic and retroaortic limbs. In this situation, the pre-aortic limb usually receives the adrenal, gonadal, and phrenic veins; the retro-aortic limb receives the lumbar and the hemiazygous veins. The retroaortic limb passes obliquely and downward to reach the inferior vena cava at a lower level<sup>[8-13]</sup>. The clinical significance of this anomaly is that the impeded outflow from the retroaortic limb leads to congestive venous hypertension and increased blood flow in pre-aortic limb<sup>[8,9,11-13]</sup>. Knowledge of this anomaly is important for the angiographer performing renal and/or adrenal venography. In addition, it is of surgical importance when a left renal transplant and/or splenorenal shunt are considered. As for splenorenal shunt operations, opinions are contradictory. For example, some authors<sup>[14]</sup> recommend to perform splenorenal shunt, without the risk of RVH, while according to other researchers<sup>[15,16]</sup> the connection of splenic vein to retroaortic limb leads not only to inadequate drainage portal system with recurrent bleedings, but also the risk of development of RVH.

Another type of abnormality is retroaortic LRV (single or multiple). The retroaortic type of LRV occurs in approximately 2-6.6/cent<sup>[17-22]</sup>. When the ventral limb atrophies a retroaortic renal vein occurs. In this transformation, there are anatomical prerequisites for disorders of venous hemodynamics - the emergence of

congestive venous hypertension, clinically manifested by proteinuria or hematuria and the development of the secondary varicocele<sup>[1,5,11,19,23]</sup>. Performing splenorenal shunt in this type of LRV is not advisable, since drainage of a large amount of blood from the portal system leads to RVH one hand and recurrent bleeding from gastroesophageal varices on the others<sup>[15]</sup>.

### **Acquired changes of the LRV**

Human body has anatomical preconditions, which may cause significant haemodynamic alterations that may lead to clinical symptoms and significant associated morbidity. The clinical manifestations of this predisposition is nutcracker syndrome<sup>[24]</sup>. In view of the insufficiency of symptoms during the first decade of life, specified condition practically have not described in pediatric patients, in most cases classified as associated finding. The nutcracker syndrome refers to compression of the LRV between the superior mesenteric artery and abdominal aorta. Obstruction of LRV occasionally causes clinically significant venous hypertension resulting in unexplained left flank, gross haematuria, with formation of periureteric and gonadal varices and varicocele in relatively young and previously healthy patients<sup>[25-29]</sup>. Other possible symptoms include pelvic congestion, chronic pediatric fatigue syndrome and orthostatic proteinuria<sup>[30-38]</sup>.

Other rare acquired causes of RVH includes renal vein thrombosis, organic renal vein stenosis and arteriovenous fistula<sup>[39-41]</sup>.

Well known that LRV mostly used in performing various types portosystemic shunts for portal hypertension. Issues related to presence of RVH in patients who underwent portosystemic shunting recent years draw increasing attention of researchers<sup>[15,16,42,43]</sup>. The data about the state of the left kidney after portosystemic shunting operation are very controversial. For example, some authors argue that performing end-to-end splenorenal shunt provides the venous drainage from the portal system to IVC without renal dysfunction<sup>[44-48]</sup>. But according to other data<sup>[14-16,42,43]</sup>, impeded outflow of the left renal vein leads to not only venous hypertensive nephropathy, but can be cause of insufficiency of created anastomosis and therefore unsatisfactory results of surgical treatment. Furthermore, impeded outflow of LRV results in venous hypertension with the formation of intra- and extrarenal collaterals and/or the development of gonadal vein reflux resulting retrograde flow and has been implicated in the development of varicocele or ovaricocele<sup>[49]</sup>. According to experimentally induced extrahepatic portal hypertension<sup>[50-54]</sup> shunting end renal vein to side splenic vein (renosplenic) after ligation of the LRV lateral to the adrenolumbar tributary, leads to haemorrhagic necrosis of the left kidney. Thus, the ureteric, lumbar and pericapsular collaterals cannot adequately drain the left kidney. Ligation of the LRV on the medial side of the adrenolumbar tributary maintained a patent left renal vein in all cases<sup>[50,52,53]</sup>.



Practical experience has shown that performing splenorenal anastomosis with ligation of the LRV proximal to the confluence of the adrenal vein - in one third of cases causes decreasing of renal function (according to the excretory urography), renal infarction, hematuria and proteinuria<sup>[52,53]</sup>.

In addition, in the pathogenesis of the RVH renal arterial blood flow is essential<sup>[55]</sup>. High pressure in the renal artery in systemic arterial hypertension increases tone of sympathetic-adrenal system, which causes vasoconstriction in the cortex and increases medullary blood flow. Autoregulation mechanisms lead to increasing pressure in the renal venous system, which are the anatomical and functional characteristics of the vascular bed of the kidney. The diversity of intrarenal arteriovenous shunts, venous network ensures acceptance of a large amount of blood in the face of increasing its arterial delivery - this is the pathogenesis of RVH in systemic arterial hypertension<sup>[55]</sup>. On the other hand, congenital or acquired arteriovenous fistula leads to the restructuring of angioarchitectonics of kidneys and in this case pressure in renal veins increases due to shunting of blood through the abnormal arteriovenous communications. The blood from the arterial bed drains to venous rout bringing extraordinary pressure to the veins. Thus, developed the renal venous hypertension<sup>[55,56]</sup>.

### **Diagnostics of RVH**

In the evaluation of renal hemodynamics, intravascular pressure indicators are most important. Retrograde left renal venography and measurement of the pressure gradient between the left renal vein and the IVC are procedures of choice for diagnosing RVH. Normally, this gradient is determined in a horizontal position from a healthy child was equal to  $0.13 \pm 0.02$  kPa, with individual variations  $0.33 \pm 0.05$  kPa<sup>[57,58]</sup>. A number of studies indicated that the anomalies of the LRV (usually circumaortic and retroaortic LRV), the pressure gradient increases significantly (up to 0.86 kPa). However, these techniques are invasive and use of such invasive examinations is generally deemed imprudent in children, and non-invasive imaging studies are preferable. Recently progressive development of non-invasive imaging techniques led to that Doppler ultrasound (US) has become the method of choice in the diagnosis of RVH. During the last decade, increased the number of publications describing different ultrasound descriptions of renocaval segment anomalies<sup>[1,27-29,34-36,57-60]</sup>. Also in details described intrarenal arteriovenous shunts<sup>[61-63]</sup>. Recent publications dedicated in most cases for nutcracker syndrome<sup>[27-29,34-36,57-60]</sup>. Kim *et al.*<sup>[58]</sup> suggested that a ratio of the AP diameter, and peak velocity (PV) between the hilar and aortomesenteric portions of the LRV of greater than 5.0 could be used as the cut-off level for the diagnosis of nutcracker syndrome with a sensitivity of 80% and a specificity of 94%. However, it has not yet been confirmed whether these criteria can be applied to children with clinically

suspected nutcracker syndrome. In addition, detection of collateral veins around the left renal vein at color Doppler US is a reliable criterion for the diagnosis of nutcracker phenomenon<sup>[27]</sup>. However, the LRV flow patterns and collateral vein formations associated with nutcracker phenomenon depend on the degree and stage of the phenomenon<sup>[58]</sup>. In patients with early nutcracker phenomenon, LRV distention and high pressure gradients exist before collateral veins develop. Moreover, in patients with collateral veins, the presence of a distended left renal vein and hypertension of the left renal vein indicate that the nutcracker phenomenon is noncompensatory<sup>[58]</sup>.

Regardless of the incidence angle, the resistances in the renal artery can be evaluated by measuring the resistive index and pulsatility index if the vessel is identified by colour Doppler. Increasing these rates in some cases may be indirect evidence of the venous outflow disturbances from LRV<sup>[15,16]</sup>.

Recently, non-invasive methods such as computed tomography (CT) and magnetic resonance imaging (MRI) have been used in the diagnosis of nutcracker syndrome<sup>[10,12,41,64-66]</sup>. Performing of the study for our opinion, more appropriate to carry out not only for diagnostic purposes of RVH but also to assess the topographic anatomy course of renocaval segment and their relative position to the vessels of the v. porta and abdominal aorta in the planning of vascular surgery in the retroperitoneal space.

### **The clinical manifestations of RVH**

The clinical presentation of RVH include the development of collateral blood flow and symptoms of renal function disorders<sup>[1,5,28,29,67]</sup>. The increased venous pressure within the renal circulation promotes the development of collaterals of the renal pelvis, and this plexus of abnormal hypertensive veins causes microhematuria or gross hematuria, orthostatic proteinuria<sup>[6,19,30-38]</sup>. Other possible symptoms include left flank pain, left-sided varicocele, pelvic congestion, chronic pediatric fatigue syndrome, and gastrointestinal symptoms<sup>[1,43,67]</sup>.

Performing various types of splenorenal shunts using abnormally developed LRV due to portal hypertension can become a reason of unsatisfactory results with recurrent bleeding from gastroesophageal varices<sup>[14-16,42,43,68]</sup>. In addition, shunting the large amounts of blood from portal vein and its tributaries to abnormally developed LRV manifests as clinical signs of renal venous hypertension<sup>[14,16,69]</sup>.

Different therapeutic methodologies have been used in treatment of RVH. In general, moderate manifestations may be controlled with conservative methods<sup>[70]</sup>. Nearly all surgical approaches aim to relieve the LRV outflow obstruction<sup>[70-85]</sup>. Surgical modalities including autotransplantation of the left kidney, LRV bypass with graft interposition and reanastomosis to the IVC anteriorly has been performed with satisfying results<sup>[73-75]</sup>. Renal autotransplantation may offer maximal efficiency in terms of normalizing renal venous

circulation. In more severe cases with hematuria, significant stenosis of LRV, varicocele, left flank pain and pressure gradient more than 1.33 kPa preferable intervention on LRV. Lot of evidence of the efficacy of endovascular interventions - methods of stenting and balloon angioplasty<sup>[76-85]</sup>. Initially performed *via* a transperitoneal approach, an external stent can be wrapped around the renal vein to prevent its compression by the mesoaortic clamp. The procedure has now also been performed by laparoscopic surgery. External and internal stenting procedures by either minimally invasive or endovascular approaches are promising treatment options. However, the risk of erosion of adjacent structures and dislodgment of the stent has not been defined yet.

However, surgical treatment methods have certain disadvantages. Thus, venous vascular suture can be considered as a potential source of thrombosis<sup>[72,83]</sup>. Postoperative complications may even lead to nephrectomy<sup>[84]</sup>. Even traditionally performed safe operations intravascular stents placement - can have few complications<sup>[79-82]</sup>.

## CONCLUSION

There are reasonable basis for research on the status of renocaval segment for modern pediatric surgeons, urologists, specialists concerned in portal hypertension, liver kidney transplant surgeons. The presence of RVH should be considered on the basis of a thorough clinical examination in patients with hematuria, left flank pain, varicocele, and symptoms of pelvic venous congestion. Dilatation of LRV and its tributaries, anomalies, additional communications observed on ultrasonography, computed tomography CT, or MRI should alert the physician to consider the diagnosis. If the symptoms merit, in particular if cystoscopy demonstrates left ureteral hematuria, selective left renal venography with pullback determination of renocaval pressure gradient is the diagnostic test of choice and should be performed in all patients. At the same time, complexity of revealing the causes of RVH with above mentioned methods, it is feasible to study the role of arterial blood, not only because of their lack of data, but also well-known factors associated with abnormal blood supply, and it is widely performed operations of decompression of the portal system through the LRV. Despite numerous studies, reasonableness of performing various types of splenorenal shunts in portal hypertension with prerequisites for RVH remains debatable. Finally, it is not enough studied phenomenon of nutcracker syndrome after surgical and congenital splenorenal shunts.

## REFERENCES

- 1 **Mendizábal S**, Román E, Serrano A, Berbel O, Simón J. Left renal vein hypertension syndrome. *Nefrología* 2005; **25**: 141-146 [PMID: 15912650]

- 2 **Chuang VP**, Mena CE, Hoskins PA. Congenital anomalies of the inferior vena cava. Review of embryogenesis and presentation of a simplified classification. *Br J Radiol* 1974; **47**: 206-213 [PMID: 4824552]
- 3 **Chuang VP**, Mena CE, Hoskins PA. Congenital anomalies of the left renal vein: angiographic consideration. *Br J Radiol* 1974; **47**: 214-218 [PMID: 4824553]
- 4 **Valentine RJ**, Dougald C, Gillivray M, Charles Blankenship Gary G. Wew Variations in the anatomic relationship of the left renal vein to the left renal artery at the aorta. *Clin Anat* 1990; **3**: 249-255 [DOI: 10.1002/ca.980030402]
- 5 **Turgut HB**, Bircan MK, Hatipoğlu ES, Doğruyol S. Congenital anomalies of left renal vein and its clinical importance: a case report and review of literature. *Clin Anat* 1996; **9**: 133-135 [PMID: 8720788 DOI: 10.1002/(SICI)1098-2353(1996)9]
- 6 **Lopatkin NA**, Morozov AV, Lopatkina LN. Essential renal haemorrhages. *Eur Urol* 1978; **4**: 115-119 [PMID: 631152]
- 7 **Giordano JM**, Trout HH. Anomalies of the inferior vena cava. *J Vasc Surg* 1986; **3**: 924-928 [PMID: 3520027 DOI: 10.1067/mva.1986.avs0030924]
- 8 **Beckmann CF**, Abrams HL. Renal venography: anatomy, technique, applications, analysis of 132 venograms, and a review of the literature. *Cardiovasc Intervent Radiol* 1980; **3**: 45-70 [PMID: 6989497 DOI: 10.1007/BF02551962]
- 9 **Senecail B**, Bobeuf J, Forlodou P, Nonent M. Two rare anomalies of the left renal vein. *Surg Radiol Anat* 2003; **25**: 465-467 [PMID: 13680187 DOI: 10.1007/s00276-003-0164-4]
- 10 **Tatar I**, Tore HG, Celik HH, Karcaaltincaba M. Retroaortic and circumaortic left renal veins with their CT findings and review of the literature. *Anat* 2008; **2**: 72-76 [DOI: 10.2399/ana.08.072]
- 11 **Mathews R**, Smith PA, Fishman EK, Marshall FF. Anomalies of the inferior vena cava and renal veins: embryologic and surgical considerations. *Urology* 1999; **53**: 873-880 [PMID: 10223477 DOI: 10.1016/S0090-4295(99)00007-2]
- 12 **Trigaux JP**, Vandroogenbroek S, De Wispelaere JF, Lacrosse M, Jamart J. Congenital anomalies of the inferior vena cava and left renal vein: evaluation with spiral CT. *J Vasc Interv Radiol* 1998; **9**: 339-345 [PMID: 9540920 DOI: 10.1016/S1051-0443(98)70278-7]
- 13 **Satyapal KS**, Kalideen JM, Haffjee AA, Singh B, Robbs JV. Left renal vein variations. *Surg Radiol Anat* 1999; **21**: 77-81 [PMID: 10370998 DOI: 10.1007/BF01635058]
- 14 **Ferzauli AN**, Razumovskiy AY, Vodolazov YA. Renal venous hemodynamics after portosystemic shunting in children with extrahepatic portal hypertension. *Annals Surg* 1997; **1**: 12-16
- 15 **Aliiev MM**, Yuldashev RZ, Adilova GS, Yusupaileva GA. Renal blood flow before and after portosystemic shunt in children with portal hypertension. *Pediatr Surg Int* 2014; **30**: 295-299 [PMID: 24448909 DOI: 10.1007/s00383-014-3463-4]
- 16 **Aliiev MM**, Yuldashev RZ, Adilova GS. Portosystemic shunts and its influence on renal hemodynamics in patients with portal hypertension. *Pediatr Surg* 2015; **3**: 3-9
- 17 **Davis CJ**, Lundberg GD. Retroaortic left renal vein, a relatively frequent anomaly. *Am J Clin Pathol* 1968; **50**: 700-703 [PMID: 4881947]
- 18 **Thomas TV**. Surgical implications of retroaortic left renal vein. *Arch Surg* 1970; **100**: 738-740 [PMID: 5444497 DOI: 10.1001/archsurg.1970.01340240076016]
- 19 **Gibo M**, Onitsuka H. Retroaortic left renal vein with renal vein hypertension causing hematuria. *Clin Imaging* 1998; **22**: 422-424 [PMID: 9876912]
- 20 **Karkos CD**, Bruce IA, Thomson GJ, Lambert ME. Retroaortic left renal vein and its implications in abdominal aortic surgery. *Ann Vasc Surg* 2001; **15**: 703-708 [PMID: 11769156 DOI: 10.1007/s10016-001-0022-y]
- 21 **Minniti S**, Visentini S, Procacci C. Congenital anomalies of the venae cavae: embryological origin, imaging features and report of three new variants. *Eur Radiol* 2002; **12**: 2040-2055 [PMID: 12136323 DOI: 10.1007/s00330-001-1241-x]
- 22 **Lee SE**, Park DS, Chung SY, Lee YT. Retroaortic renal vein. *Korean J Urol* 2002; **43**: 84-86

- 23 **Nam JK**, Park SW, Lee SD, Chung MK. The clinical significance of a retroaortic left renal vein. *Korean J Urol* 2010; **51**: 276-280 [PMID: 20428432 DOI: 10.4111/kju.2010.51.4.276]
- 24 **Grant JCB**. Methods of Anatomy. Baltimore: Williams & Wilkins, 1937: 158
- 25 **Sansilvestri MP**, Rupin A, Badier-Commander C. Chronic venous insufficiency: dysregulation of collagen synthesis. *Angiology* 2003; **54**: 13-18 [DOI: 10.1177/000331970305400103]
- 26 **Braedel HU**, Steffens J, Ziegler M, Polsky MS, Platt ML. A possible ontogenic etiology for idiopathic left varicocele. *J Urol* 1994; **151**: 62-66 [PMID: 8254834]
- 27 **Takebayashi S**, Ueki T, Ikeda N, Fujikawa A. Aosis of the nutcracker syndrome with color Doppler sonography: correlation with flow patterns on retrograde left renal venography. *AJR* 1999; **172**: 39-43
- 28 **Takemura T**, Iwasa H, Yamamoto S, Hino S, Fukushima K, Isokawa S, Okada M, Yoshioka K. Clinical and radiological features in four adolescents with nutcracker syndrome. *Pediatr Nephrol* 2000; **14**: 1002-1005 [PMID: 10975316 DOI: 10.1007/s004670050062]
- 29 **Rudloff U**, Holmes RJ, Prem JT, Faust GR, Moldwin R, Siegel D. Mesoaortic compression of the left renal vein (nutcracker syndrome): case reports and review of the literature. *Ann Vasc Surg* 2006; **20**: 120-129 [PMID: 16374539 DOI: 10.1007/s10016-005-5016-8]
- 30 **Lau X**, Lo R, Chan FL, Wong KK. The posterior «nutcracker»: hematuria secondary to retroaortic left renal vein. *Urology* 1986; **28**: 437-439 [DOI: 10.1016/0090-4295(86)90085-3]
- 31 **Shaper KR**, Jackson M, Williams G. The nutcracker syndrome: an uncommon cause of haematuria. *Br J Urol* 1994; **74**: 144-146 [DOI: 10.1111/j.1464-410X.1994.tb16575.x]
- 32 **Challenger RJ**, Scott Dougherty W, Flisak ME, Flanagan RG. Left renal vein hypertension as a cause of persistent gross hematuria. *Urology* 1996; **48**: 468-472 [DOI: 10.1016/S0090-4295(96)00205-1]
- 33 **Gibo M**, Onitsuka H. Retroaortic left renal vein with renal vein hypertension causing hematuria. *Clin Imaging* 1998; **22**: 422-424 [DOI: 10.1016/S0899-7071(98)00067-9]
- 34 **Park SJ**, Lim JW, Cho BS, Yoon TY, Oh JH. Nutcracker syndrome in children with orthostatic proteinuria: diagnosis on the basis of Doppler sonography. *J Ultrasound Med* 2002; **21**: 39-45; quiz 46 [PMID: 11794401]
- 35 **Cheon JE**, Kim WS, Kim IO, Kim SH, Yeon KM, Ha IS, Cheong HI, Choi Y. Nutcracker syndrome in children with gross haematuria: Doppler sonographic evaluation of the left renal vein. *Pediatr Radiol* 2006; **36**: 682-686 [PMID: 16770671 DOI: 10.1007/s00247-006-0145-y]
- 36 **Ekim M**, Bakkaloglu SA, Tümer N, Sanlidilek U, Salih M. Orthostatic proteinuria as a result of venous compression (nutcracker phenomenon)--a hypothesis testable with modern imaging techniques. *Nephrol Dial Transplant* 1999; **14**: 826-827 [PMID: 10328451 DOI: 10.1093/ndt/14.4.826]
- 37 **Cho BS**, Choi YM, Kang HH, Park SJ, Lim JW, Yoon TY. Diagnosis of nut-cracker phenomenon using renal Doppler ultrasound in orthostatic proteinuria. *Nephrol Dial Transplant* 2001; **16**: 1620-1625 [PMID: 11477164 DOI: 10.1093/ndt/16.8.1620]
- 38 **Faizan MK**, Finn LS, Paladin AM, McDonald RA. A 14-year-old girl with recumbent proteinuria. *Pediatr Nephrol* 2002; **17**: 379-381 [PMID: 12042899 DOI: 10.1007/s00467-002-0822-5]
- 39 **Brill PW**, Mitty HA, Strauss L. Renal Vein Thrombosis: A Cause of Intrarenal Calcification in the Newborn. *Pediatr Radiol* 1977; **6**: 172-175 [DOI: 10.1007/bf00972111]
- 40 **Calligaro KD**, Dougherty MJ. Renal artery aneurysms and arteriovenous fistulae. In: Rutherford RB. Vascular surgery, 5th ed. Philadelphia: Saunders, 2000: 1697-1706
- 41 **Gayer G**, Zissin R, Rimon U, Guranda L, Apter S, Hertz M. Vascular lesions of the renal sinus. *Emerg Radiol* 2003; **10**: 135-141 [PMID: 15290501 DOI: 10.1007/s10140-003-0301-8]
- 42 **Polyaev YA**, Sukhov MN, Garbuzov RV, Drozdov AV, Mylnikov AA. Angiographic diagnosis of pathology associated with extrahepatic portal hypertension in children and its impact on surgical approach. *Detskaya bolnitsa* 2010; **1**: 9-15
- 43 **Sukhov MN**, Polyaev YA, Drozdov AV. Varicose veins of the small pelvis and varicocele in children with extrahepatic portal hypertension. *Detskaya bolnitsa* 2011; **1**: 13-18
- 44 **Simon JS**, Brown AA, Ross HB. Ligation of the left renal vein in splenorenal anastomosis without impairment of renal function. *Br J Surg* 1972; **59**: 170-173 [PMID: 5014516 DOI: 10.1002/bjs.1800590303]
- 45 **Cohen D**, Stephen M. Control of bleeding in extrahepatic portal hypertension - the reverse splenorenal shunt and portal-azygos disconnection Aust. *Paediatr J* 1984; **20**: 147-150 [DOI: 10.1111/j.1440-1754.1984.tb00065.x]
- 46 **Lee SG**, Moon DB, Ahn CS, Kim KH, Hwang S, Park KM, Ha TY, Ko GY, Sung KB, Song GW, Jung DH, Moon KM, Kim BS, Cho YP. Ligation of left renal vein for large spontaneous splenorenal shunt to prevent portal flow steal in adult living donor liver transplantation. *Transpl Int* 2007; **20**: 45-50 [PMID: 17181652 DOI: 10.1111/j.1432-2277.2006.00392.x]
- 47 **Samson RH**, Lepore MR, Showalter DP, Nair DG, Lanoue JB. Long-term safety of left renal vein division and ligation to expedite complex abdominal aortic surgery. *J Vasc Surg* 2009; **50**: 500-504; discussion 504 [PMID: 19595540 DOI: 10.1016/j.jvs.2009.04.041]
- 48 **Slater RR**, Jabbour N, Abbass AA, Patil V, Hundley J, Kazimi M, Kim D, Yoshida A, Abouljoud M. Left renal vein ligation: a technique to mitigate low portal flow from splenic vein siphon during liver transplantation. *Am J Transplant* 2011; **11**: 1743-1747 [PMID: 21668639 DOI: 10.1111/j.1600-6143.2011.03578.x]
- 49 **Schulte-Baukloh H**, Kämmer J, Felfe R, Stürzebecher B, Knispel HH. Surgery is inadvisable: massive varicocele due to portal hypertension. *Int J Urol* 2005; **12**: 852-854 [PMID: 16201987 DOI: 10.1111/j.1442-2042.2005.01170.x]
- 50 **Threefoot SA**, Cabrera-Gil C, Pearson Jr JE. Collateral circulation, renal function and histology after experimental obstruction of renal veins. *Chest* 1970; **58**: 249-260 [DOI: 10.1378/chest.58.3.249]
- 51 **Darewicz J**, Cylwik B, Gruszecki W. Effect of clamping of the renal vein in dogs on certain biochemical and histopathological changes. *Int Urol Nephrol* 1976; **8**: 271-276 [PMID: 14081 DOI: 10.1007/BF02082090]
- 52 **Solomon MJ**, Stening MR, Hargrave JC. The fate of the left kidney after end renosplenic shunt in experimentally induced extrahepatic portal hypertension. *Aust N Z J Surg* 1990; **60**: 545-548 [PMID: 2357180 DOI: 10.1111/j.1445-2197.1990.tb07423.x]
- 53 **Effeney DJ**, Stoney RJ. Wylie's atlas of vascular surgery. Venous disease and miscellaneous arteriopathies. Philadelphia: JB Lippincott Company Philadelphia, 1993: 38-54
- 54 **Satyapal KS**, Kalideen JM. The renal veins in the human cadaveric fetus: their importance as contributors to collateral flow. *J Anat* 1995; **186** (Pt 2): 405-409 [PMID: 7649840]
- 55 **Lopatkin NA**, Morozov AV, Jitnikova LN. Stenosis of renal vein. *Meditsina* 1984; **1**: 12-21
- 56 **Matsell DG**, Jones DP, Boulden TF, Burton EM, Baum SL, Tonkin IL. Arteriovenous fistula after biopsy of renal transplant kidney: diagnosis and treatment. *Pediatr Nephrol* 1992; **6**: 562-564 [PMID: 1482648 DOI: 10.1007/BF00866508]
- 57 **Carl P**, Stark L, Ouzoun N, Reindl P. Venous pressure in idiopathic varicocele. *Eur Urol* 1993; **24**: 214-220 [PMID: 8375442]
- 58 **Kim WS**, Cheon JE, Kim IO, Kim SH, Yeon KM, Kim KM, Choi H. Hemodynamic investigation of the left renal vein in pediatric varicocele: Doppler US, venography, and pressure measurements. *Radiology* 2006; **241**: 228-234 [PMID: 16908673 DOI: 10.1148/radiol.2411050271]
- 59 **Butros SR**, Liu R, Oliveira GR, Ganguli S, Kalva S. Venous compression syndromes: clinical features, imaging findings and management. *Br J Radiol* 2013; **86**: 20130284 [PMID: 23908347 DOI: 10.1259/bjr.20130284]
- 60 **Mallat F**, Hmida W, Othmen MB, Mosbah F. Mixed nutcracker syndrome with left renal vein duplication: A severe and exceptional presentation in an 18-year-old boy. *Urol Ann* 2015; **7**: 244-247 [PMID: 25836700 DOI: 10.4103/0974-7796.150494]
- 61 **Derchi LE**, Martinoli C, Pontremoli R. Postbiopsy arteriovenous fistulas of the native kidneys diagnosed by Doppler US. *Eur. Radiol* 1993; **3**: 186-189 [DOI: 10.1007/BF00169798]
- 62 **Deane C**, Cowan N, Giles J. Arteriovenous fistulas in renal

- transplants: color Doppler ultrasound observations *Urol. Radiol* 1992; **13**: 211-217
- 63 **Ahn J**, Cohen HI. Post renal biopsy complication: perinephric hematoma and arteriovenous fistula *J. Ultrasound Med* 1995; **14**: 327-328
  - 64 **Zerin JM**, Hernandez RJ, Sedman AB, Kelsch RC. "Dilatation" of the left renal vein on computed tomography in children: a normal variant. *Pediatr Radiol* 1991; **21**: 267-269 [PMID: 1870922 DOI: 10.1007/BF02018620]
  - 65 **Kim KW**, Cho JY, Kim SH, Yoon JH, Kim DS, Chung JW, Park JH. Diagnostic value of computed tomographic findings of nutcracker syndrome: correlation with renal venography and renocaval pressure gradients. *Eur J Radiol* 2011; **80**: 648-654 [PMID: 20869828]
  - 66 **Wong HI**, Chen MC, Wu CS, Fu KA, Lin CH, Weng MJ, Liang HL, Pan HB. The usefulness of fast-spin-echo T2-weighted MR imaging in Nutcracker syndrome: a case report. *Korean J Radiol* 2010; **11**: 373-377 [PMID: 20461194 DOI: 10.3348/kjr.2010.11.3.373]
  - 67 **Gupta R**, Gupta A, Aggarwal N. Variations of gonadal veins: embryological prospective and clinical significance. *J Clin Diagn Res* 2015; **9**: AC08-AC10 [PMID: 25859438 DOI: 10.7860/JCDR/2015/9493.5578]
  - 68 **Satran L**, Amplatz K, Wolfson JJ, Leonarid AS, Sharp HL. Abnormal left kidney following splenorenal shunt. *Amer Roentgen* 1969; **106**: 92-96
  - 69 **Yuldashev RZ**, Aliev MM, Adilova GS. Impact of high portal pressure on renal hemodynamics in children with portal hypertension. *J Hepat* 2015; **62** Suppl 2: S820-821 [DOI: 10.1016/S0168-8278(15)31432-x]
  - 70 **Menard MT**. Nutcracker syndrome: when should it be treated and how? *Perspect Vasc Surg Endovasc Ther* 2009; **21**: 117-124 [PMID: 19703821 DOI: 10.1177/1531003509338402]
  - 71 **Lin WQ**, Huang HF, Li M, Wang ZG, Chen JH, He XL, Zhou JQ. Diagnosis and therapy of the nutcracker phenomenon: long-term follow-up. *Zhonghua Waike Zazhi* 2003; **41**: 889-892 [PMID: 14728826]
  - 72 **Wang L**, Yi L, Yang L, Liu Z, Rao J, Liu L, Yang J. Diagnosis and surgical treatment of nutcracker syndrome: a single-center experience. *Urology* 2009; **73**: 871-876 [PMID: 19193424 DOI: 10.1016/j.urology.2008.11.043]
  - 73 **Thompson PN**, Farling RC, III, Chang BB. A case of nutcracker syndrome: treatment by mesoaoartic transposition. *J Vasc Surg* 1992; **16**: 663-665 [DOI: 10.1016/0741-5214(92)90176-9]
  - 74 **Chuang CK**, Chu SH, Lai PC. The nutcracker syndrome managed by autotransplantation. *J Urol* 1997; **157**: 1833-1834 [DOI: 10.1097/00005392-199705000-00083]
  - 75 **Marone EM**, Psacharopulo D, Kahlberg A, Coppi G, Chiesa R. Surgical treatment of posterior nutcracker syndrome. *J Vasc Surg* 2011; **54**: 844-847 [PMID: 21458199 DOI: 10.1016/j.jvs.2011.01.038]
  - 76 **Hohenfellner M**, D'Elia G, Hampel C, Dahms S, Thuroff JW. Transposition of the left renal vein for treatment of the nutcracker phenomenon: long-term follow-up. *Urology* 2002; **59**: 354-357 [DOI: 10.1016/S0090-4295(01)01537-0]
  - 77 **Barnes RW**, Fleisher HL 3rd, Redman JF, Smith JW, Harshfield DL, Ferris EJ. Mesoaoartic compression of the left renal vein (the so-called nutcracker syndrome): repair by a new stenting procedure. *J Vasc Surg* 1988; **8**: 415-421 [DOI: 10.1067/mva.1988.avs0080415]
  - 78 **Neste MG**, Narasimham DL, Belcher KK. Endovascular stent placement as a treatment for renal venous hypertension. *J Vasc Interv Radiol* 1996; **7**: 859-861 [DOI: 10.1016/S1051-0443(96)70861-8]
  - 79 **Segawa N**, Azuma H, Iwamoto Y. Expandable metallic stent placement for nutcracker phenomenon. *Urology* 1999; **53**: 631-633 [DOI: 10.1016/S0090-4295(98)00355-0]
  - 80 **Park YB**, Lim SH, Ahn JH. Nutcracker syndrome: intravascular stenting approach. *Nephrol Dial Trans* 2000; **15**: 99-101 [DOI: 10.1093/ndt/15.1.99]
  - 81 **Takahashi Y**, Sano A, Matsuo M. An effective "transluminal balloon angioplasty" therapy for pediatric chronic fatigue syndrome with nutcracker phenomenon. *Clin Nephrol* 2000; **53**: 77-78 [PMID: 10661488]
  - 82 **Zhang H**, Zhang N., Li M. Treatment of six cases of left renal nutcracker phenomenon: surgery and endografting. *Chin. Med J* 2003; **116**: 1782-1784
  - 83 **d'Archambeau O**, Maes M, De Schepper AM. The pelvic congestion syndrome: role of the "nutcracker phenomenon" and results of endovascular treatment. *JBR-BTR* 2004; **87**: 1-8 [PMID: 15055326]
  - 84 **Wang X**, Zhang Y, Li C, Zhang H. Results of endovascular treatment for patients with nutcracker syndrome. *J Vasc Surg* 2012; **56**: 142-148 [PMID: 22575480 DOI: 10.1016/j.jvs.2012.01.007]
  - 85 **Yih ND**, Chyen LH, Cunli Y, Jaywantraj PS, Isip AB, Anil SA. Renosplenic shunting in the nutcracker phenomenon: a discussion and paradigm shift in options? A novel approach to treating nutcracker syndrome. *Int J Angiol* 2014; **23**: 71-76 [PMID: 24627622 DOI: 10.1055/s-0033-1348883]

**P- Reviewer:** Cheng TH, Tan XR    **S- Editor:** Qiu S    **L- Editor:** A  
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