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Polycystins and mechanotransduction: From physiology to disease

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Abstract

Polycystins are key mechanosensor proteins able to respond to mechanical forces of external or internal origin. They are widely expressed in primary cilium and plasma membrane of several cell types including kidney, vascular endothelial and smooth muscle cells,

osteoblasts and cardiac myocytes modulating their physiology. Interaction of polycystins with diverse ion channels, cell-cell and cell-extracellular matrix junctional proteins implicates them in the regulation of cell structure, mechanical force transmission and mechanotransduction. Their intracellular localization in endoplasmic reticulum further regulates subcellular trafficking and calcium homeostasis, finely-tuning overall cellular mechanosensitivity. Aberrant expression or genetic alterations of polycystins lead to severe structural and mechanosensing abnormalities including cyst formation, deregulated flow sensing, aneurysms, defective bone development and cancer progression, highlighting their vital role in human physiology.

Key words: Polycystins; Mechanotransduction; Kidney; Endothelium; Osteoblasts; Cancer

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Core tip: Polycystins are key regulators of mechanosensation in several cell types including kidney, vascular endothelial and smooth muscle cells, osteoblasts and cardiac myocytes. Their expression in primary cilium, plasma membrane and endoplasmic reticulum, along with their ability to interact with diverse ion channels, cell-cell and cell-extracellular matrix junctional proteins renders polycystins as essential regulators of overall cellular mechanoreponse. Abnormal expression or genetic defects of polycystins result in severe structural and mechanosensing faults including cyst formation, deregulated flow sensing, aneurysms, defective bone development and cancer progression, highlighting their crucial role in human physiology.

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INTRODUCTION

Cellular mechanosensitivity plays fundamental role in cell viability and function, tissue development and maintenance of organs. Most cell types are able to respond to mechanical forces which provide them with a means of actively sensing and responding to mechanical properties such as topography and rigidity of the environment. Mechanical forces can be of external (such as acceleration, gravity, touch, stretch, sound) or of internal origin (breathing, fluid flow, blood pressure, osmotic pressure, heart contraction or any membrane deformation) and can vary from modest to high intensity depending on the cell type^[1].

Mechanosensitivity constitutes a three-step process. It starts with detection of a mechanical stimulus by a cellular component, followed by mechanotransduction that converts the mechanical signal into a biophysical or biochemical signal, ending with the mechanoreponse during which signal sensation and transduction integrate over space and time^[2].

Several mechanosensation models have been proposed based on the nature of the mechanosensor proteins^[2-4]. Transmembrane proteins sense mechanical stimuli through changes in tension in their surrounding lipid bilayer ("bilayer tension model"^[5]). Proteins involved in cell adhesion and maintenance of cell structure sense mechanical tension through binding with structural components that can transmit force from the intracellular or extracellular side or both ("Tethered protein model"). In this model interaction between the mechanosensor and proteins of the cell-cell junctions, extracellular matrix (ECM), focal adhesion points, microtubules or actin cytoskeleton has been reported. The way a protein responds to the applied force can also differ. A force applied to a mechanosensing protein can unfold it and expose cryptic peptides that can activate intracellular pathways and mechanotransduction^[6] ("protein unfolding model"). Mechanical forces either exert a direct effect on the intrinsic activity of the mechanosensor proteins such as ion-channel gating, enzyme activity or ligand-receptor interactions ("mechanosensitive protein activity model"), or an indirect effect by activating a non-mechanosensitive protein leading to mechanotransduction ("adjacent mechanosensitive protein model"). The indirect activation can be through ligand release or through protein-protein interaction. All these models can also work in concert in order to form mechanosensitive complexes.

Mechanosensation is widely distributed in cellular compartments and involves the interaction of several protein complexes including adherent junctions, desmosomes, integrins, focal adhesion points, receptors and actin microtubules. Most of these proteins are connected to signaling pathways that involve cytosolic molecules, calcium signaling or transcription factors^[7]. Their expression should match within time and space to the sensory function of the mechanosensor organ, while removal of the protein should annul the sensory

response. Mutations that change protein function can modify the mechanosensing ability of the organ or cell. A heterologous expression of the mechanosensor protein in another cell type should lead to a mechanical response.

Interestingly, the protein family of polycystins has been shown to physically interact with most mechanosensing protein complexes mentioned above. Polycystins are implicated in renal flow sensing^[8], vascular pressure and flow mechanosensation^[9-11], blood-brain barrier mechanical injury^[12], nodal flow sensing^[13], skeletal development and osteoblast differentiation^[14,15] as well as cancer progression^[16].

POLYCYSTINS - STRUCTURE AND LOCALIZATION

Polycystins are large integral proteins, broadly expressed in human tissues including kidneys, blood vessels, heart, liver, pancreas, bone and brain. They are found localized in the primary cilium, at the plasma membrane and at endoplasmic reticulum (ER) where they associate and interact with numerous partners^[17].

Polycystin 1 (PC1, 460 kDa) consists of 11 transmembrane segments, a short intracellular C-terminal region (200 amino acids) and an extracellular N-terminal part (3000 amino acids) which contains several protein motifs. These include a G protein-coupled receptor proteolytic site, two cysteine-flanked leucine-rich repeats, sixteen Ig-like domains and a C-lectin domain. The terminal intracellular C-region contains a coiled-coil domain (CC) as well as a G protein-binding site (G).

PC1 is found localized at the primary cilium and at the plasma membrane being involved in interactions between proteins and between proteins-carbohydrates. Scientific data support interaction of PC1 with many proteins localized at focal adhesion points, adherens junctions and desmosomes^[18].

Polycystin 2 (PC2, TRPP2, 110 kDa) is composed of six transmembrane segments, an intracellular N-terminus which contains a ciliary sorting motif and an intracellular C-terminus with a calcium-binding EF domain, an ER retention domain and a CC domain. PC2 is located in the ER^[19] while its translocation to the plasma membrane has been reported to require the presence of PC1^[20].

PC2 belongs to the transient receptor potential (TRP) channel family proteins. It has been shown to interact with cytoskeletal proteins as well as other mechanosensitive ion channels in different cells, including potassium-selective stretch-activated potassium channels and non-selective cationic SAC channels.

PC1 and PC2 may interact through their CC domains located in the cytoplasmic C-termini forming an ion channel complex, as well as with many other partners in various subcellular localizations^[20,21]. They are considered as important regulators of calcium homeostasis by affecting the resting cytosolic calcium

concentration, decreasing sarcoplasmic reticulum (SR) Ca²⁺-ATPase (SERCA2a) expression and inhibiting the passive leakage of Ca²⁺ from the ER^[22].

Since the original studies that identified polycystins and their gene mutations as a causative link to autosomal dominant polycystic kidney disease (ADPKD), considerable progress has been made in revealing the physiological functions of these proteins in multiple tissues, such as lung, kidney, cardiovascular, brain and bone^[14].

POLYCYSTINS - MECHANOTRANSDUCTION IN THE KIDNEY

In the kidney, polycystins are detected at the cilia of renal epithelial cells^[23]. PC1 though its extracellular domain is functions as mechanosensor detecting urine flow. Activation of PC1 leads to mechanotransduction by opening the PC2 channel allowing calcium entry and triggering intracellular calcium release in the ER through inositol 1,4,5-trisphosphate (IP3) or ryanodine receptors^[8,24]. The mechanical properties of PC1 can be changed by osmolytes such as sorbitol or urea (a major urine component) and modulate mechanosensation^[25]. Furthermore, at the primary cilium flow detection by PC1/PC2 complex induces proteolytic cleavage of the intracellular PC1 C-terminus (34 kDa fragment, called CTT), activation of signaling pathways [mammalian target of rapamycin (mTOR), Janus kinase/signal transducers and activators of transcription (JAK/STAT), Wntless-Int (Wnt)] and gene expression changes in order to get a mechanoreponse^[26,27]. PC2 does not seem to need the presence of PC1 for its channel activity but it rather forms a heteromeric channel with TRP channel subfamily c member 1 (TRPC1)^[27].

Flow sensing by primary cilium has been associated with increased intracellular calcium concentration being lost in PC1 or PC2-deficient cells and it has been proposed to result in cyst formation in polycystic kidneys^[28].

Loss-of-function *Pkd1* or *Pkd2* gene mutations encoding PC1 and PC2, are responsible for ADPKD, the most common kidney disease, affecting almost 1 in 1000 individuals^[29]. ADPKD clinical phenotype involves cysts presence in the kidney, pancreas, and liver along with severe cardiovascular defects. Arterial hypertension and intracranial aneurysms are often associated with this multisystem disease.

POLYCYSTINS - MECHANOTRANSDUCTION IN VASCULAR TISSUES

PC1 and PC2 expression has been observed in the plasma membrane and primary cilium of endothelial

cells, proposed to transmit extracellular shear stress^[30]. Shear stress-induced activation of PC2 has been demonstrated to increase the biosynthesis of intracellular NO leading to smooth muscle dilatation and flow-induced vascular relaxation^[31].

In agreement, a previous study from our group using partial carotid stenosis to induce low shear stress *in vivo*, has shown upregulation of PC1 and PC2 in endothelium at the low shear stress area^[11], implicating both proteins in blood flow alterations sensing. Since low shear stress conditions have been associated with atherosclerotic plaque development, a role of polycystins in atherosclerosis is possible.

Polycystins have been shown to interact with the two major calcium-release channels, IP3 receptors in epithelial cells and ryanodine receptors in cardiomyocytes. PC1 interacts with the IP3 receptors to reduce calcium levels^[32]. Similarly, in the heart, PC2 interacts with the ryanodine receptor RyR2 *via* its C-terminus to modulate release of Ca²⁺ from the SR stores^[33].

Notably, PC2 can form a channel with TRPC1, being activated in response to mechanical damage of blood-brain barrier endothelial cells by promoting Ca²⁺ influx and formation of actin stress fibers^[12].

Finally, in vascular smooth muscle cells PC2 has been implicated in sensing pressure volume and in mesenteric and cerebral arteries in sensing myogenic tone^[19,34].

POLYCYSTINS - MECHANOTRANSDUCTION IN OSTEOBLASTIC LINEAGE CELLS

In osteoblasts, the polycystin-primary cilia signaling complex has been attributed a mechanosensory role that regulates skeletogenesis and bone formation. Evaluation of the skeletal phenotype of *Pkd1*-deficient mice revealed PC1 implication in bone development and in the regulation of osteoblast function through intracellular calcium-dependent control of Runx2 expression. Furthermore, abnormal bone development and osteopenia was observed upon loss of *Pkd1* function in mice due to impaired osteoblast differentiation^[14].

Another study of a mouse model with midpalatal suture expansion demonstrated that proliferation and differentiation of periosteal osteochondroprogenitor cells that were mechanically stimulated requires *Pkd1*^[35]. This is in concert with a recent study from our group, exploring PC1 involvement in mechanical load (stretching)-induced signaling pathways in human pre-osteoblasts. In this study, PC1 was revealed as a major mechanosensor molecule in osteoblasts that modulates their differentiation and gene transcription through the calcineurin/nuclear factor of activated T-cells signaling pathway, thus controlling bone formation^[15].

POLYCYSTINS INTERACT WITH CELL ADHESION AND CYTOSKELETAL PROTEINS TO TUNE OVERALL CELLULAR MECHANOSENSITIVITY

Polycystins ability to form multiprotein complexes with components of adherens junctions, focal adhesions, desmosomes and cytoskeleton suggests their critical role in regulating cell-ECM as well as cell-cell interactions.

Strong homophilic interactions between the Ig-like domain of PC1 with other PC1 and Ig domain-containing proteins of neighboring cells have been detected in the Madin-Darby canine kidney cells indicating their role in intercellular adhesion^[30]. In cultures of renal epithelial cells, PC1 localizes at lateral cell junctions, involved in cell-cell interactions^[30]. In addition, PC1 has been demonstrated to bind to focal adhesion proteins including integrins, pp125 focal adhesion kinase (pp125FAK), vinculin and paxillin which are required for cell adhesion to the ECM^[36,37]. Of note, some of these associations have been lost in ADPKD epithelial cells^[36].

In aortic smooth muscle cells, PC1 interacts with PC2 at the dense plaques which are involved in the interaction between the cytoskeleton, plasma membrane and ECM^[38]. Regarding the components of adherens junctions, PC1 associates with E-cadherin and α -, β - and γ -catenins in a multiprotein complex required for maintenance of tissue structure and function^[39,40]. In ADPKD renal epithelial cells, disruption of this multiprotein complex has been observed leading to depletion of PC1 and E-cadherin from the plasma membrane.

In desmosomes, the most abundant cell-cell junctions, PC1 has been found to directly bind through its cytoplasmic tail the CC motif of the intermediate filament proteins cytokeratins 8 and 18, desmin and vimentin^[41]. In ADPKD cysts, these desmosomal elements are severely disoriented^[42].

Finally, cytoskeletal proteins such as actin-binding proteins, cardiac troponin and tropomyosin have been directly associated with PC2 channel activity upon stimulation by osmotic or hydrostatic pressure^[43,44]. These numerous partners indicate that polycystins play a central role in the way cells adapt to their mechanical environment.

POLYCYSTINS - IMPLICATION IN CANCER PROGRESSION

All the aforementioned protein interactions that affect cell-cell and cell-ECM communication have been previously associated with cancer. Moreover, mechanical signals have been shown to regulate cancer cell interactions influencing decisive steps of invasion and metastasis. Recently, we have demonstrated the implication of PC1/PC2 in colorectal cancer progression.

Overexpression of polycystins was associated with aggressive colorectal cancer phenotype *in vitro*. Clinical analyses revealed a correlation of elevated PC1 expression with poor recurrence-free survival, while aberrant PC2 levels was correlated with poor overall survival^[16]. Several studies have also reported a connection between cancer proliferation, migration and metastasis with alterations in ion channels expression, and particularly with changes in TRP channel proteins^[45]. More specifically, TRP channel subfamily M member 8 and TRP protein homologue (TRP6) were found highly expressed in prostate cancer where they correlate with histological grade. TRP channel subfamily M member 7 is implicated in proliferation and growth of breast cancer and head and neck tumor cells while TRP channel subfamily V member 4 and TRPC1 were associated with glioma growth^[45]. Since some of these channels are known PC2 partners, it is highly likely the implication of polycystins in these malignancies and is currently under investigation.

CONCLUSION

Polycystins are envisioned as polymodal cellular sensors, critical regulators of cell structure integrity, cell communication, force transmission and subcellular trafficking in a broad range of cell types. Aberrant or defective expression of these proteins leads to abnormalities in calcium homeostasis and mechanosensation in major organs contributing to severe pathological conditions including ADPKD and cancer.

Future research should focus in elucidation of the mechanisms involved to integrate the information that arises from polycystin complexes into cellular functions. *In vitro* studies are needed to determine the kind of stimuli that trigger polycystins activation and the way that mechanical stimuli and ligand binding is sensed by PKD complexes. Are polycystins directly activated by mechanical stimuli or indirectly *via* activation of another protein partner? Animal models are required to define the functional consequences of PKD dysfunction in different cell types. How critical are polycystin-induced calcium signaling and enzymatic cascades for cellular growth and differentiation? Is there a specific role for multiple PKD complexes present at different locations in a single cell? How polycystins expression varies in different types of malignancy? Understanding the molecular mechanisms that underpin polycystins functions is urgently needed to identify novel and effective therapeutic schemes for the affected organs in the future.

REFERENCES

- 1 **Ernstrom GG**, Chalfie M. Genetics of sensory mechanotransduction. *Annu Rev Genet* 2002; **36**: 411-453 [PMID: 12429699 DOI: 10.1146/annurev.genet.36.061802.101708]
- 2 **Vogel V**, Sheetz M. Local force and geometry sensing regulate cell functions. *Nat Rev Mol Cell Biol* 2006; **7**: 265-275 [PMID:

- 16607289 DOI: 10.1038/nrm1890]
- 3 **Nilius B**, Honoré E. Sensing pressure with ion channels. *Trends Neurosci* 2012; **35**: 477-486 [PMID: 22622029 DOI: 10.1016/j.tins.2012.04.002]
 - 4 **Brierley SM**. Molecular basis of mechanosensitivity. *Auton Neurosci* 2010; **153**: 58-68 [PMID: 19683967 DOI: 10.1016/j.autneu.2009.07.017]
 - 5 **Yoshimura K**, Sokabe M. Mechanosensitivity of ion channels based on protein-lipid interactions. *J R Soc Interface* 2010; **7** Suppl 3: S307-S320 [PMID: 20356872 DOI: 10.1098/rsif.2010.0095]
 - 6 **Gao M**, Craig D, Vogel V, Schulten K. Identifying unfolding intermediates of FN-III(10) by steered molecular dynamics. *J Mol Biol* 2002; **323**: 939-950 [PMID: 12417205 DOI: 10.1016/S0022-2836(02)01001-X]
 - 7 **Bukoreshtliiev NV**, Haase K, Pelling AE. Mechanical cues in cellular signalling and communication. *Cell Tissue Res* 2013; **352**: 77-94 [PMID: 23224763 DOI: 10.1007/s00441]
 - 8 **Nauli SM**, Alenghat FJ, Luo Y, Williams E, Vassilev P, Li X, Elia AE, Lu W, Brown EM, Quinn SJ, Ingber DE, Zhou J. Polycystins 1 and 2 mediate mechanosensation in the primary cilium of kidney cells. *Nat Genet* 2003; **33**: 129-137 [PMID: 12514735 DOI: 10.1038/ng1076]
 - 9 **Sharif-Naeini R**, Folgering JH, Bichet D, Duprat F, Lauritzen I, Arhatte M, Jodar M, Dedman A, Chatelain FC, Schulte U, Retailleau K, Loufrani L, Patel A, Sachs F, Delmas P, Peters DJ, Honoré E. Polycystin-1 and -2 dosage regulates pressure sensing. *Cell* 2009; **139**: 587-596 [PMID: 19879844 DOI: 10.1016/j.cell.2009.08.045]
 - 10 **Nauli SM**, Kawanabe Y, Kaminski JJ, Pearce WJ, Ingber DE, Zhou J. Endothelial cilia are fluid shear sensors that regulate calcium signaling and nitric oxide production through polycystin-1. *Circulation* 2008; **117**: 1161-1171 [PMID: 18285569 DOI: 10.1161/CIRCULATIONAHA.107.710111]
 - 11 **Varela A**, Katsiboulas M, Tousoulis D, Politi K, Papaioannou TG, Vaina S, Davos CH, Piperi C, Stefanadis C, Basdra EK, Papavassiliou AG. Early shear stress signaling on vascular endothelium by a modified partial carotid ligation model. *Int J Cardiol* 2011; **152**: 413-416 [PMID: 21917331 DOI: 10.1016/j.ijcard.2011.08.051]
 - 12 **Berrout J**, Jin M, O'Neil RG. Critical role of TRPP2 and TRPC1 channels in stretch-induced injury of blood-brain barrier endothelial cells. *Brain Res* 2012; **1436**: 1-12 [PMID: 22192412 DOI: 10.1016/j.brainres.2011.11.044]
 - 13 **McGrath J**, Somlo S, Makova S, Tian X, Brueckner M. Two populations of node monocilia initiate left-right asymmetry in the mouse. *Cell* 2003; **114**: 61-73 [PMID: 12859898 DOI: 10.1016/S0092-8674(03)00511-7]
 - 14 **Xiao ZS**, Quarles LD. Role of the polycystin-primary cilia complex in bone development and mechanosensing. *Ann N Y Acad Sci* 2010; **1192**: 410-421 [PMID: 20392267 DOI: 10.1111/j.1749-6632.2009.05239.x]
 - 15 **Dalagiorougou G**, Piperi C, Georgopoulou U, Adamopoulos C, Basdra EK, Papavassiliou AG. Mechanical stimulation of polycystin-1 induces human osteoblastic gene expression via potentiation of the calcineurin/NFAT signaling axis. *Cell Mol Life Sci* 2013; **70**: 167-180 [PMID: 23014991 DOI: 10.1007/s00018-012-1164-5]
 - 16 **Gargalionis AN**, Korkolopoulou P, Farmaki E, Piperi C, Dalagiorougou G, Adamopoulos C, Levidou G, Saetta A, Fragkou P, Tsioli P, Kiaris H, Zizi-Serbetzoglou A, Karavokyros I, Papavassiliou KA, Tsavaris N, Patsouris E, Basdra EK, Papavassiliou AG. Polycystin-1 and polycystin-2 are involved in the acquisition of aggressive phenotypes in colorectal cancer. *Int J Cancer* 2015; **136**: 1515-1527 [PMID: 25123959 DOI: 10.1002/ijc.29140]
 - 17 **Retailleau K**, Duprat F. Polycystins and partners: proposed role in mechanosensitivity. *J Physiol* 2014; **592**: 2453-2471 [PMID: 24687583 DOI: 10.1113/jphysiol.2014.271346]
 - 18 **Palsson R**, Sharma CP, Kim K, McLaughlin M, Brown D, Arnaut MA. Characterization and cell distribution of polycystin, the product of autosomal dominant polycystic kidney disease gene 1. *Mol Med* 1996; **2**: 702-711 [PMID: 8972485]
 - 19 **Koulen P**, Cai Y, Geng L, Maeda Y, Nishimura S, Witzgall R, Ehrlich BE, Somlo S. Polycystin-2 is an intracellular calcium release channel. *Nat Cell Biol* 2002; **4**: 191-197 [PMID: 11854751 DOI: 10.1038/ncb754]
 - 20 **Hanaoka K**, Qian F, Boletta A, Bhunia AK, Piontek K, Tsiokas L, Sukhatme VP, Guggino WB, Germino GG. Co-assembly of polycystin-1 and -2 produces unique cation-permeable currents. *Nature* 2000; **408**: 990-994 [PMID: 11140688 DOI: 10.1038/35050128]
 - 21 **Qian F**, Boletta A, Bhunia AK, Xu H, Liu L, Ahrabi AK, Watnick TJ, Zhou F, Germino GG. Cleavage of polycystin-1 requires the receptor for egg jelly domain and is disrupted by human autosomal-dominant polycystic kidney disease 1-associated mutations. *Proc Natl Acad Sci USA* 2002; **99**: 16981-16986 [PMID: 12482949 DOI: 10.1073/pnas.252484899]
 - 22 **Mekahli D**, Sammels E, Luyten T, Welkenhuyzen K, van den Heuvel LP, Levtchenko EN, Gijsbers R, Bultynck G, Parys JB, De Smedt H, Missiaen L. Polycystin-1 and polycystin-2 are both required to amplify inositol-trisphosphate-induced Ca²⁺ release. *Cell Calcium* 2012; **51**: 452-458 [PMID: 22456092 DOI: 10.1016/j.ceca.2012.03.002]
 - 23 **Pazour GJ**, San Agustin JT, Folliot JA, Rosenbaum JL, Witman GB. Polycystin-2 localizes to kidney cilia and the ciliary level is elevated in orpk mice with polycystic kidney disease. *Curr Biol* 2002; **12**: R378-R380 [PMID: 12062067 DOI: 10.1016/S0960-9822(02)00877-1]
 - 24 **Jin X**, Mohieldin AM, Muntean BS, Green JA, Shah JV, Mykytyn K, Nauli SM. Cilioplasm is a cellular compartment for calcium signaling in response to mechanical and chemical stimuli. *Cell Mol Life Sci* 2014; **71**: 2165-2178 [PMID: 24104765 DOI: 10.1007/s00018-013-1483-1]
 - 25 **Ma L**, Xu M, Oberhauser AF. Naturally occurring osmolytes modulate the nanomechanical properties of polycystic kidney disease domains. *J Biol Chem* 2010; **285**: 38438-38443 [PMID: 20937836 DOI: 10.1074/jbc.M110.183913]
 - 26 **Low SH**, Vasanth S, Larson CH, Mukherjee S, Sharma N, Kinter MT, Kane ME, Obara T, Weimbs T. Polycystin-1, STAT6, and P100 function in a pathway that transduces ciliary mechanosensation and is activated in polycystic kidney disease. *Dev Cell* 2006; **10**: 57-69 [PMID: 16399078 DOI: 10.1016/j.devcel.2005.12.005]
 - 27 **Merrick D**, Chapin H, Baggs JE, Yu Z, Somlo S, Sun Z, Hogensch JB, Caplan MJ. The γ -secretase cleavage product of polycystin-1 regulates TCF and CHOP-mediated transcriptional activation through a p300-dependent mechanism. *Dev Cell* 2012; **22**: 197-210 [PMID: 22178500 DOI: 10.1016/j.devcel.2011.10.028]
 - 28 **Kotsis F**, Boehlke C, Kuehn EW. The ciliary flow sensor and polycystic kidney disease. *Nephrol Dial Transplant* 2013; **28**: 518-526 [PMID: 23314319 DOI: 10.1093/ndt/gfs524]
 - 29 **Bichet D**, Peters D, Patel AJ, Delmas P, Honoré E. Cardiovascular polycystins: insights from autosomal dominant polycystic kidney disease and transgenic animal models. *Trends Cardiovasc Med* 2006; **16**: 292-298 [PMID: 17055386 DOI: 10.1016/j.tcm.2006.07.002]
 - 30 **Ibraghimov-Beskrovnaya O**, Dackowski WR, Foggensteiner L, Coleman N, Thiru S, Petry LR, Burn TC, Connors TD, Van Raay T, Bradley J, Qian F, Onuchic LF, Watnick TJ, Piontek K, Hakim RM, Landes GM, Germino GG, Sandford R, Klinger KW. Polycystin: in vitro synthesis, in vivo tissue expression, and subcellular localization identifies a large membrane-associated protein. *Proc Natl Acad Sci USA* 1997; **94**: 6397-6402 [PMID: 9177229 DOI: 10.1073/pnas.94.12.6397]
 - 31 **AbouAlaiwi WA**, Takahashi M, Mell BR, Jones TJ, Ratnam S, Kolb RJ, Nauli SM. Ciliary polycystin-2 is a mechanosensitive calcium channel involved in nitric oxide signaling cascades. *Circ Res* 2009; **104**: 860-869 [PMID: 19265036 DOI: 10.1161/CIRCRESAHA.108.192765]
 - 32 **Li Y**, Santos NG, Yu S, Woodward OM, Qian F, Guggino WB. Polycystin-1 interacts with inositol 1,4,5-trisphosphate receptor to modulate intracellular Ca²⁺ signaling with implications for polycystic kidney disease. *J Biol Chem* 2009; **284**: 36431-36441 [PMID: 19854836 DOI: 10.1074/jbc.M109.068916]

- 33 **Anyatonwu GI**, Estrada M, Tian X, Somlo S, Ehrlich BE. Regulation of ryanodine receptor-dependent calcium signaling by polycystin-2. *Proc Natl Acad Sci USA* 2007; **104**: 6454-6459 [PMID: 17404231 DOI: 10.1073/pnas.0610324104]
- 34 **Narayanan D**, Bulley S, Leo MD, Burris SK, Gabrick KS, Boop FA, Jaggar JH. Smooth muscle cell transient receptor potential polycystin-2 (TRPP2) channels contribute to the myogenic response in cerebral arteries. *J Physiol* 2013; **591**: 5031-5046 [PMID: 23858011 DOI: 10.1113/jphysiol.2013.258319]
- 35 **Hou B**, Kolpakova-Hart E, Fukai N, Wu K, Olsen BR. The polycystic kidney disease 1 (Pkd1) gene is required for the responses of osteochondroprogenitor cells to midpalatal suture expansion in mice. *Bone* 2009; **44**: 1121-1133 [PMID: 19264154 DOI: 10.1016/j.bone.2009.02.018]
- 36 **Wilson PD**, Geng L, Li X, Burrow CR. The PKD1 gene product, "polycystin-1," is a tyrosine-phosphorylated protein that colocalizes with alpha2beta1-integrin in focal clusters in adherent renal epithelia. *Lab Invest* 1999; **79**: 1311-1323 [PMID: 10532593]
- 37 **Weston BS**, Bagn eris C, Price RG, Stirling JL. The polycystin-1 C-type lectin domain binds carbohydrate in a calcium-dependent manner, and interacts with extracellular matrix proteins in vitro. *Biochim Biophys Acta* 2001; **1536**: 161-176 [PMID: 11406351 DOI: 10.1016/S0925-4439(01)00046-1]
- 38 **Qian Q**, Li M, Cai Y, Ward CJ, Somlo S, Harris PC, Torres VE. Analysis of the polycystins in aortic vascular smooth muscle cells. *J Am Soc Nephrol* 2003; **14**: 2280-2287 [PMID: 12937304 DOI: 10.1097/01.ASN.0000080185.38113.A3]
- 39 **Huan Y**, van Adelsberg J. Polycystin-1, the PKD1 gene product, is in a complex containing E-cadherin and the catenins. *J Clin Invest* 1999; **104**: 1459-1468 [PMID: 10562308 DOI: 10.1172/JCI51111]
- 40 **Geng L**, Burrow CR, Li HP, Wilson PD. Modification of the composition of polycystin-1 multiprotein complexes by calcium and tyrosine phosphorylation. *Biochim Biophys Acta* 2000; **1535**: 21-35 [PMID: 11113628 DOI: 10.1016/S0925-4439(00)00079-X]
- 41 **Xu GM**, Sikaneta T, Sullivan BM, Zhang Q, Andreucci M, Stehle T, Drummond I, Arnaout MA. Polycystin-1 interacts with intermediate filaments. *J Biol Chem* 2001; **276**: 46544-46552 [PMID: 11581269 DOI: 10.1074/jbc.M107828200]
- 42 **Russo RJ**, Husson H, Joly D, Bukanov NO, Patey N, Knebelmann B, Ibraghimov-Beskrovnaya O. Impaired formation of desmosomal junctions in ADPKD epithelia. *Histochem Cell Biol* 2005; **124**: 487-497 [PMID: 16187067 DOI: 10.1007/s00418-005-0055-3]
- 43 **Montalbetti N**, Li Q, Gonz alez-Perrett S, Semprine J, Chen XZ, Cantiello HF. Effect of hydro-osmotic pressure on polycystin-2 channel function in the human syncytiotrophoblast. *Pflugers Arch* 2005; **451**: 294-303 [PMID: 16025301 DOI: 10.1007/s00424-005-1458-7]
- 44 **Li Q**, Dai Y, Guo L, Liu Y, Hao C, Wu G, Basora N, Michalak M, Chen XZ. Polycystin-2 associates with tropomyosin-1, an actin microfilament component. *J Mol Biol* 2003; **325**: 949-962 [PMID: 12527301 DOI: 10.1016/S0022-2836(02)01333-5]
- 45 **Chen J**, Luan Y, Yu R, Zhang Z, Zhang J, Wang W. Transient receptor potential (TRP) channels, promising potential diagnostic and therapeutic tools for cancer. *Biosci Trends* 2014; **8**: 1-10 [PMID: 24647107 DOI: 10.5582/bst.8.1]

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Endoplasmic reticulum stress-mediated pathways to both apoptosis and autophagy: Significance for melanoma treatment

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Abstract

Melanoma is the most aggressive form of skin cancer. Disrupted intracellular signaling pathways are responsible for melanoma's extraordinary resistance to current chemotherapeutic modalities. The pathophysiologic basis for resistance to both chemo- and radiation therapy is rooted in altered genetic and epigenetic mechanisms that, in turn, result in the impairing of cell death machinery and/or excessive activation of cell growth and survival-dependent pathways. Although most current melanoma therapies target mitochondrial dysregulation, there is increasing evidence that endoplasmic reticulum (ER) stress-associated pathways play a role in the potentiation, initiation and maintenance of cell death machinery and autophagy. This review focuses on the reliability of ER-associated pathways as therapeutic targets for melanoma treatment.

Key words: Melanoma; Endoplasmic reticulum; Apoptosis; Autophagy; Signaling pathways; Chemotherapy

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Core tip: This editorial describes the clinical validity of the endoplasmic reticulum (ER) as therapeutic target

for melanoma treatment. In addition, we highlight in this review the mechanistic role of ER stress in the modulation of both apoptosis and autophagy-associated pathways. Drugs that perturb ER function may represent an alternative approach for melanoma treatment. This paper reviews the previous and current published studies on the reliability of ER-associated pathways as therapeutic targets for melanoma.

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INTRODUCTION

Although melanoma accounts for less than 5% of all skin cancers, it exhibits the highest mortality rate of all cutaneous tumors and its incidence is rapidly increasing^[1]. The high mortality rate is the result of the propensity of metastatic dissemination throughout the body^[2], and the development of resistance mechanisms that permit melanoma to evade normal immune surveillance mechanisms and the anti-tumor effects of chemotherapy^[3]. Early detection and surgical excision of early stage disease offers the best hope of cure in patients with primary melanoma^[4]. Even with new targeted therapies, the prognosis for advanced metastatic malignant melanoma is poor^[5]. The available options for patients with advanced malignant melanoma patients provide limited therapeutic benefit with successful treatments often being measured in months of increased survival rather than years^[6-8]. The potential to develop resistance mechanisms that counteract drug-induced apoptosis and evade host immunological responses is particularly devastating^[9]. Accordingly, the replacement of single agent chemotherapy with targeted therapies is revolutionizing systemic therapy^[10]. Besides the mechanistic role of mitochondrial damage-dependent pathways in the modulation of anti-cancer agent-induced apoptosis of tumor cells, anti-cancer agents can also improve killing efficiency *via* endoplasmic reticulum (ER) stress-dependent pathways^[11-13]. While autophagy-mediated tumor death in response to anti-cancers is clinically relevant, these anti-cancer agents can also induce autophagy-mediated cytoprotective mechanisms^[12,13], a pattern of tumor resistance to chemotherapy.

Metastatic melanoma demonstrates particularly poor response rates to single chemotherapeutic agents^[14,15]. For instance, dacarbazine (DTIC) demonstrates no impact on survival, though it is considered to be one of the most effective agents that is used as standard therapy for the treatment of metastatic melanoma^[16,17]. Other anticancer agents such as cisplatin, carmustine

and the vinca alkaloids (*e.g.*, vindesine and vinblastine) fail to show any therapeutic advantage over DTIC^[18], though several combination chemotherapy regimens demonstrate a modest increased response rate^[19].

Melanoma's resistance to therapy is the results of an upregulation in pro-survival factors, which potentiate tumor maintenance and progression^[20]. One of these factors is the inducible transcription factor NF- κ B that is responsible for the regulation of the expression of genes related to apoptosis^[21]. It is also, central to the development of tumor resistance to alkylating agents such as DTIC^[22-24]. Accordingly, the inhibition of NF- κ B pathway may improve the cytotoxic efficacy of alkylating agent-based therapy. To that end, preclinical studies *in vitro* and *in vivo* using human melanoma tumor models revealed that the therapeutic efficiency of DTIC or temozolomide is enhanced with the addition of the proteasome inhibitor, bortezomib^[25,26].

Traditional mono- or multi-chemotherapy regimens are also associated with the development of significant adverse effects^[27,28]. The development of new tumor types in these patients is attributed to the molecular action of the anticancer agents leading to the induction and/or destruction of aberrant signaling pathways.

The molecular action of chemotherapy in tumor cells is commonly associated with phenotypic alterations including cell death and survival-dependent mechanisms including apoptosis and autophagy^[12,13].

Apoptosis and autophagy occur in normal cells. These are essential physiological mechanisms required for the maintenance of organismal and cellular homeostasis^[29]. Current information about autophagy in melanoma focuses on autophagosome formation and/or autolysosome degradation in response to a variety of therapeutic agents using melanoma derived cell lines^[13,30,31]. Chemotherapy induction of autophagy serves to protect melanoma cells from intended chemotherapy-induced apoptosis. In fact, the induction of autophagy following the treatment of melanoma cells with bortezomib reduces bortezomib-induced apoptosis^[13]. Similarly, the induction of autophagy by esomeprazole, a proton pump inhibitor, blocks melanoma cell death^[32]. Based on this preclinical evidence, the modulation of autophagy-associated pathways offers a promising treatment strategy to increase treatment efficiency by overcoming melanoma resistance to chemotherapy.

The involvement of ER stress in the modulation of apoptotic mechanisms leading to melanoma cell death has been reported in several studies^[12,13,33]. This may result from the induction of BH3 proteins such as Noxa and Puma leading to the inhibition of Bcl-2 localization at the ER membrane, alterations in the distribution of the calcium flux which produce ER stress^[13,34].

Although ER stress and autophagy are capable of modulating each other in tumor tissues, their specific function is thought to be tumor type and stage-dependent^[34-36]. The clinical potential of ER stress and/or autophagy-associated pathways as therapeutic

target for melanoma treatment has been reported in several studies^[37-39]. For example, BRAF wild type (wt) melanoma is more sensitive to ER stress-based therapies than melanoma with hyperactivating BRAF mutations^[40]. The frequency of BRAF mutation seems to be associated with elevated levels of autophagy in melanoma. Accordingly, ER stress-induced apoptosis of melanoma cells harboring oncogenic BRAF is lower than those observed in BRAF wt melanoma cells^[40-42]. Inhibition of autophagy is a good strategy to sensitize BRAF wt melanoma cells to ER stress-mediated apoptosis. In addition, the development of anti-cancer agents based on the enhancement or suppression of these processes may be relevant therapeutic strategies^[38,43,44].

Tumor resistance or response to available therapeutic modalities depends on the balance between apoptosis and autophagy-associated mechanisms^[45,46]. Although the development of the most available therapeutic approaches focuses on the excessive activation of mitochondrial dysregulation-dependent pathways leading to apoptosis, there is increasing evidence that ER stress-associated pathways represent an important therapeutic target for melanoma treatment^[13,47]. Thus, the development of anti-cancer agents with ability to trigger the intrinsic activation of ER stress/unfolded protein response (UPR)-associated pathways may offer a novel therapeutic strategy for tumor treatment. UPR is mediated in response to the enhancement of protein synthesis through the activation of mitogen-activated protein kinase kinase/extracellular signal-regulated kinase (MEK/ERK) pathway that, in turn, induces cell proliferation, a mechanism that can block ER stress-induced apoptosis^[48]. Thus, ER stress-dependent pathways have been proposed to represent a new therapeutic target for melanoma treatment^[10,49]. Accordingly, the inhibition of oncogenic BRAF (V600E) and/or MEK-attenuated activation of inositol-requiring enzyme 1 (IRE1) and activating transcription factor 6 (ATF6) signaling of the UPR in melanoma cells may sensitize melanoma cells to apoptosis. Our work focuses on the reliability of ER stress-dependent pathways as a therapeutic target for melanoma treatment.

FUNCTION OF ER IN NORMAL AND TUMOR CELLS

ER is a network of tubules and flattened sacs comprising rough and smooth regions that differ in their structure and function^[50]. The rough ER is characterized by the existence of ribosomes attached to the cytoplasmic side of the membrane, whereas the smooth ER lack these ribosomes^[50]. ER plays a crucial role in normal cellular functioning, by processing of post-translational modification and folding of secretory and membrane proteins. These secretory and membrane proteins are synthesized along the membrane of the rough ER and subsequently are passed onto the

Golgi apparatus, where they undergo further post-translational modifications by the attachment of lipid and glucose moieties in a lipidation and glycosylation-dependent manner, respectively^[51]. The ability of ER to correctly fold nascent proteins depends on chaperone proteins that, under normal physiological condition, are in excess in the ER lumen^[52]. The function of most chaperone proteins is known to be Ca²⁺-dependent^[53]. ER contains a high concentration of Ca²⁺ and is the only cellular organelle that plays an essential role in intracellular Ca²⁺ homeostasis^[54]. Thus, the escalation of intracellular calcium into the cytoplasm is a signal for pathophysiological alteration of the cells. This pathological phenomenon results from ER stress in response to externally physical or chemical stressors, such as radiation and toxins^[55].

ER function is critical for the regulation of many aspects of cell physiology, such as vesicle trafficking, lipid and membrane biogenesis as well as protein targeting and secretion. Normal and tumor cells react rapidly to ER stress *via* mechanisms mediated by a set of ER stress-associated pathways. The regulation of these pathways is thought to be the consequence of the perturbations in ER function, such as the accumulation of unfolded or misfolded proteins, as well as the accumulation of ER lipid, glycolipid imbalances, or alteration in the ionic or redox conditions in the lumen of ER^[56,57]. Three distinct signaling pathways have been identified as ER stress-dependent pathways, namely protein kinase RNA-like endoplasmic reticulum kinase (PERK), ATF6, and IRE1 pathways. The primary purpose of these pathways is implicated to promote cell survival by mechanisms mediated through the reduction of the misfolded protein^[58]. Figure 1 outlines the ER stress-associated pathways in normal and tumor cells.

INDUCTION OF ER STRESS-ASSOCIATED PATHWAYS BY ANTI-CANCER AGENTS

Dysregulation of ER homeostasis is a primary pathophysiological mechanism responsible for the initiation of an ER stress response that leads to the development of a number of human diseases including cancer^[59]. The induction of ER stress by anti-cancer agents and other stimuli has been reported in several studies. The anti-cancer agent's bortezomib, vinblastine and taxol trigger ER stress in melanoma cells^[13,60,61]. Similarly, caffeic acid phenethyl ester, the BH3 mimetic obatoclax and the Abbott Compound ABT-737 have been reported to induce ER stress in melanoma^[33,62]. Interestingly, the induction of ER stress in melanoma cells by these agents is correlated with the deregulation of ER stress associated pathways including eukaryotic translation initiation factor 2 α (eIF2 α) and PERK.

ER stress induced activation of PERK leads to the phosphorylation of the eIF2 α that inhibits the translation and subsequently triggers cell cycle arrest^[63]. CHOP (C/EBP homology protein) is downstream of PERK-

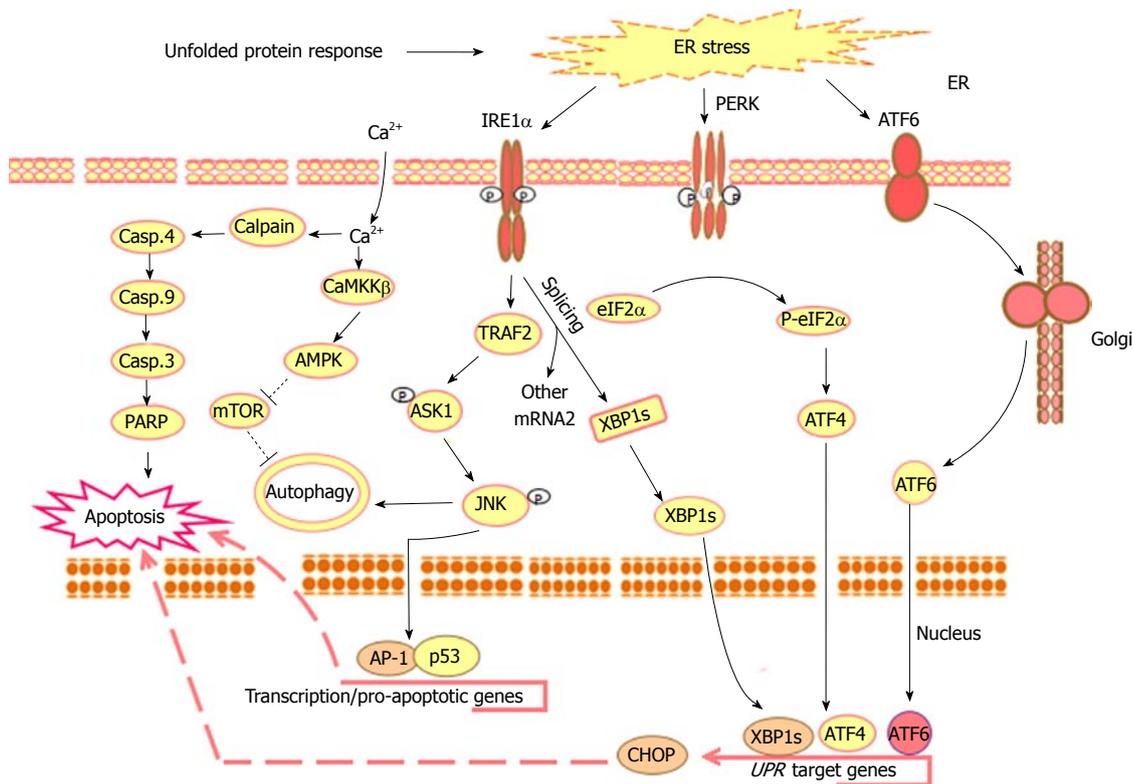


Figure 1 Outline of the main unfolded protein response-mediated mechanisms in response to the molecular action of anti-cancer agents. Upon the accumulation of misfolded proteins in the ER lumen, chaperones, the ER stress sensors PERK, IRE1 α and ATF6 become active. The phosphorylation of PERK allows it to assemble in a homo-dimer to form an active form that, in turn, results in the phosphorylation of eukaryotic initiation factor2 α (eIF2 α) to initiate UPR downstream response leading to reduction of the protein overload to ER by the suppression of the translation and activation of ATF4 together with ER stress associated transcription factors such as, CHOP, PERK, IRE1 and ATF6. Activated PERK phosphorylates the translation initiation factor eIF2 α to decrease the protein synthesis and enhance stress-inducible messages, such as ATF4. During ER stress the ATF6 traffics to the Golgi apparatus, where it is cleaved by S1P/S2P proteases. The cleavage of ATF6 from the Golgi membrane facilitates its localization to the nucleus, where it enhances the transcriptional up-regulation of UPR target genes leading to apoptosis. Whereas, activated IRE1 α is implicated into several functions: One of these functions is essential to drive the splice mechanism of the XBP-1 mRNA to allow the translation of mature XBP-1 protein that, in turn, functions as a transcription factor to promote the transcription of UPR target genes such as CHOP leading to apoptosis; the other function of the activated IRE1 α is to recruit TRAF2 that subsequently mediates the phosphorylation of ASK1 and the activation of its downstream JNK leading to the activation of the transcription factors AP-1 and p53 that are essential for the transcription of the pro-apoptotic genes and genes implicated in the processes of autophagosome formation and autophagy. UPR results also in intracellular calcium release leading to cell death *via* Calpain/Caspase-4, Caspase-9, caspase-3 and PARP axis or autophagy *via* CaMKK β /AMPK axis leading to the inhibition of mTOR and subsequently autophagy. ER: Endoplasmic reticulum; PERK: Protein kinase RNA-like endoplasmic reticulum kinase; IRE1 α : Inositol-requiring protein 1; ATF6: Activating transcription factor 6; UPR: Unfolded protein response; XBP-1: X-box binding protein 1; CHOP: CCAAT/enhancer-binding protein (C/EBP) homologous protein; TRAF2: TNF receptor-associated factor 2; ASK1: Apoptosis signal regulating kinase 1; JNK: C-jun-N-terminal kinase; PARP: Poly (ADP-ribose) polymerase; CaMKK β : Calmodulin-dependent protein kinase kinase- β ; AMPK: AMP-activated protein kinase; mTOR: Mammalian target of rapamycin.

eIF2 α -ATF4 and involved in the regulation of the apoptotic proteins of the Bcl-2 family members^[64]. ER stress-induced activation of IRE1 α is responsible for the regulation of the transcription factor XBP1^[65]. Once ER stress is initiated, the conversion of unspliced XBP1 mRNA to mature mRNA is mediated permitting the translation and further modification of this protein to operate as an active transcription factor^[66]. The activation of the transcription factor XBP1 is essential for the induction of the transcription of ER-related genes that, in turn, mediate the disposal of unfolded proteins^[67]. Although this panel of responses is mainly implicated in restoring ER homeostasis, sustained ER stress is essential for the promotion of apoptosis^[68,69]. More importantly, it has been demonstrated that ER stress-associated pathways are involved in the modulation of the anti-cancer agent-induced apoptosis of tumor cells, particularly, in melanoma^[13]. In recent

years, we and others uncovered the mechanistic role of ER stress-associated pathways such as PERK-ATF4-CHOP/Bim and IRE1 α -ASK1-JNK-AP-1/HSF1-HSP70, in the modulation of anti-cancer agent-induced apoptosis of melanoma cells^[12,13]. More importantly, we demonstrated that Noxa-induced ER stress triggers apoptosis of melanoma cells *via* mechanism mediated by ASK1-JNK/p38 axis^[58,70]. Also, apoptosis related protein-2 (APR-2)-induced ER-stress drives apoptosis of melanoma cells *via* mechanism mediated by three parallel pathways, namely IRE1 α /tumor necrosis factor receptor-associated factor 2 (TRAF2)-ASK1-JNK/Cytochrome c/caspase-9/caspase-3/PARP, Calpain-caspase-4-/caspase-9/caspase-3/PARP, and PERK-ATF4-CHOP/Bim^[58]. Furthermore, bortezomib/vinblastine-induced ER stress in melanoma cells is essential for the induction of cell survival *via* autophagy-dependent pathways including, IRE1 α -ASK1-JNK-AP-1/HSF1-

HSP70 axis. More importantly, in our laboratory, we demonstrated that the inhibition of IRE1 α -ASK1-JNK-AP-1/HSF1-HSP70 pathways synergistically enhance bortezomib or vinblastine-induced apoptosis of melanoma cells^[12,13].

ER stress-mediated pathways to apoptosis in melanoma

It is established that the primary function of ER stress is to restore normal ER homeostasis and to engage cytoprotective mechanisms to counteract or mediate both intra- and extracellular-induced alterations^[71]. Therefore, if the induced ER stress is strong or persistent, the ER enhanced dysfunction becomes irreversible and consequently triggers cell death machinery to initiate apoptosis. Thus, the destruction of the ER stress-dependent pathways that are essential for the modulation of the cytoprotective machinery by small molecule-inhibitors would be expected to trigger apoptosis of tumor cells. In addition, the enhancement of key components leading to excessive activation of apoptotic pathways, such as the mammalian IRE1 α , could impact the regulation of kinases such as ASK1^[62]. The activation of the pro-apoptotic kinase ASK1, the upstream kinase of the JNK pathway, is essential for the regulation of ER stress-induced apoptosis of melanoma in response to chemotherapeutic agents such as vinblastine^[12], as well as in response to pro-apoptotic proteins such as the BH- only proteins such as Noxa^[70] and APR-2^[58]. Unlike various tumor types, particularly, those undergoing prolonged ER stress, the ER stress-dependent pathways such as IRE1 α and ATF6 are persistent in melanoma cells^[72]. Thus, it is expected that constitutive activation of both IRE1 α and ATF6 would be associated with the development of melanoma resistance to anti-cancer agents^[72]. Accordingly, the destruction of IRE1 α and/or ATF6 signaling pathways has been reported to trigger apoptosis *via* mechanism mediated by PERK pathway^[73]. The role of the PERK pathway in the modulation of ER stress-induced apoptosis has been demonstrated in various tumor types including melanoma *via* mechanism mediated by the BH3-only protein Bim^[58,73].

Although the UPR is established as a cyto- protective response, excessive and/or persistent activation of ER stress-associated pathways can also trigger apoptosis^[74]. However, the mechanism whereby UPR switches from the cyto- protection to apoptosis is thought to be the consequence of the attenuation of IRE1 α and/or ATF6 activities^[72,75]. The resistance of melanoma cells to most anti-cancer therapies during the course of anti-cancer-induced ER stress is attributed to the fact that the melanoma cells have adapted to ER stress. Although the molecular mechanisms that describe the contribution of ER stress in melanoma survival has been established, several studies revealed that the resistance of melanoma cells to ER stress-induced apoptosis results from the prolonged activation of the IRE1 α and ATF6 pathways that, in turn, lead to the attenuation of the PERK signaling pathway^[72].

Accordingly, the knockdown of IRE1 α or ATF6 sensitizes melanoma cells to ER stress-induced apoptosis^[33]. To that end, the destruction of the IRE1 α /XBP-1 pathway along ER stress is expected to overcome melanoma resistance to ER stress inducers. The involvement of IRE1 α in the activation of PI3K/Akt pathway together with the induction of Mcl-1 expression has been suggested to play an essential role in the modulation ER stress-induced survival of melanoma cells^[76]. ATF6 is involved in the transcriptional regulation of both GRP78 and XBP-1 and thereby plays an important role in melanoma resistance to ER stress-induced apoptosis^[77]. In conclusion, the differential response of various tumor types to PERK activation seems to rest on cellular factors and/or cell growth and survival pathways-dependent activation. Although the importance of IRE1/XBP-1 axis in tumor growth and survival has been established^[78,79], its mechanistic role in the promotion of the XBP1 splicing processes and the subsequent effect on the components of the downstream signaling pathway have not been well characterized. More importantly, the activation of IRE1 kinase has been reported to be essential for the activation of c-Jun-N-terminal kinase, JNK and NF- κ B pathways besides its role in the modulation of the induced unfolded protein response^[79,80]. Upon the induction of ER stress, IRE1 kinase becomes capable of recruiting TRAF2. This results in the activation of both JNK and NF- κ B pathways^[81]. The mechanisms, involved in the modulation of ER stress are outlined in Figure 1.

ER stress-mediated pathways to autophagy in melanoma

Autophagy is a highly conserved degradation pathway that is responsible for the elimination of damaged cellular components. This process is implicated in several physiological and pathological processes leading to cell survival or cell death, and is characterized by the formation of double membrane autophagosomes^[82]. The functional role of autophagy in melanoma has been reported in the context of *in vitro* analysis of chemotherapy-mediated effects in established melanoma cells^[12,13]. We recently demonstrated that the induction of autophagic machinery in response to bortezomib protects melanoma cells from bortezomib-induced apoptosis^[13]. Also, the induction of autophagy in response to the treatment with esomeprazole, a proton pump inhibitor, plays an essential role in the delay of melanoma cell death^[32]. More importantly, the inhibition of ER stress-induced autophagy by the knockdown of Mcl-1, heat shock protein 70 (HSP70) or the inhibition of Bcl-2 potentiates bortezomib-induced apoptosis of melanoma cells^[13,83]. Also, the combination of autophagy inhibitors 3-MA, bafilomycin A1 (BafA1) or LY294002 with the antiproliferative agents such as sanguinolite has the potential to reduce melanoma cell viability^[84]. Orlotti *et al*^[85] addressed the essential role of autophagy in the protection of melanoma cells from G-quadruplex ligand-induced-ER stress associated with DNA damage. Also, G-quadruplex ligand-induced autophagy has been suggested to be the consequence

of Ataxia Telangiectasia mutated-dependent DNA damage response as well as the transactivation of the cyclin-dependent kinase inhibitor 1A^[84]. Although its mechanistic role in tumor survival and resistance to treatment with chemo-and radiotherapy has been established, autophagy can also enhance the killing efficiency of chemotherapy-based treatments in various tumor types including melanoma^[86]. In recent years, autophagic cell death, also known as type II apoptosis, gained more attention, as a potential therapeutic target for tumor treatment. Soares *et al.*^[87], demonstrated that the combination of CI-IB-MECA inhibitor and paclitaxel can induce mTOR-dependent autophagic cell death, as well as caspase-dependent and/or independent apoptosis in melanoma cells. In addition, the potential of the micro-tubule poison, JG-03-14, to cause cytotoxic effects in melanoma cells both *in vitro* and *in vivo* via autophagy-dependent mechanism has been approved^[88]. Thus, chemotherapeutic agents, whose cytotoxicity is mediated by autophagy-dependent mechanisms are considered to be suitable therapeutic approaches, particularly for tumors conferring resistance to anti-cancer agents-induced apoptosis. In addition, the identification of ER stress-associated pathways as a link between BRAF signaling and cytoprotective autophagy provides a potential therapeutic target for melanoma treatment^[46]. Anti-cancer agents-induced autophagy is mostly resistant to several kinase inhibitors, particularly, those targeting the link between autophagic machinery and PI3K/AKT/mTOR pathway^[89,90].

The common genetic alterations leading to the development of malignant melanoma are widely established to be the consequence of the activating mutations in *NRAS* and *BRAF* proto-oncogenes^[91,92]. Also, genome-wide mutation detection in melanoma derived cell lines and primary tumors revealed significant alterations in the *BRAF* gene^[93]. The most identified mutations were found to affect a single residue (V600E) that is located in the kinase activation domain of BRAF^[94,95]. The importance of BRAF mutation is attributed to the potential role of RAF serine/threonine kinases, the most important key signaling components in the RAS pathways^[96]. The clinical relevance of BRAF in melanoma is based on its mechanistic role in the activation of melanocytes in cAMP-dependent pathway in response to α -melanocyte-stimulating hormone-mediated activation of melanocortin receptor 1^[97]. Accordingly, the mutation in the *BRAF* gene with its consequent impact on melanoma development and progression has gained increasing attention as a therapeutic target in melanoma. The development of a broad-spectrum of kinase inhibitors confirmed the clinical relevance of the inhibition of BRAF as an efficient therapeutic strategy for melanoma treatment. These kinase inhibitors have demonstrated the ability to inhibit BRAF, mutant BRAFV600E, and CRAF^[98]. The most potent BRAF inhibitors, vemurafenib and dabrafenib, have demonstrated antitumor activity for advanced melanoma in phase III trials, particularly in patients with BRAF mutations^[99]. Also, MEK inhibitors, such as

trametinib, showed significant antitumor activity in melanoma patients with a V600 BRAF mutation^[100]. Other MEK inhibitors, such as Binimetinib, exhibit antitumor activity in patients with advanced melanoma, who demonstrate NRAS mutation^[101]. Most importantly, the combination of BRAF inhibitors such as dabrafenib with MEK inhibitors such as trametinib have enhanced therapeutic benefits when compared with the response rate to dabrafenib alone^[102]. Despite the demonstration of therapeutic progress by both BRAF and MEK inhibitors, most patients with metastatic melanoma fail to achieve a clinical cure^[103]. The development of more effective therapeutics for advanced metastatic melanoma requires a direct evaluation of novel and innovative therapies. The roles of the Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR pathways in controlling growth and the implications for sensitivity to treatment of melanoma are outlined in Figure 2.

While MAP kinase pathways modulate autophagy-associated cell death^[104], accumulated evidence demonstrates that autophagy also plays a role in the promotion of tumor resistance and survival via MAP kinase pathway-dependent mechanisms^[13,105-107]. Specifically, the induction of cytoprotective autophagy counteracts MAP kinase-mediated pathways to apoptosis in response to chemotherapy-based treatments^[13,108]. The presence of autophagosomes in tumor cells undergoing apoptosis in response to the treatment with chemotherapy is evidence for the ability of tumor cells to evade the cytotoxicity via autophagy-dependent pathways^[109]. Thus, the inhibition of autophagic machinery induced by chemotherapeutics, such as bortezomib, may prove to be an effective therapeutic strategy^[13]. In addition, these pathways may play a role in ER stress suppression of the anti-tumor efficiency of vemurafenib, dabrafenib and trametinib in melanoma patients harboring activating NRAS or BRAF mutations (Figure 2).

Anti-cancer agents affecting ER stress-associated pathways to apoptosis of melanoma

There are a number of United States Food and Drug Administration-approved anti-cancer agents that influence key components of ER stress-dependent pathways. For example, the ruthenium-derived compounds trigger the expression of ER stress proteins such as, Bip, XBP1, PDI, and CHOP leading to tumor growth inhibition or cell death^[110,111]. Also, the anti-cancer agent 2-Hydroxyoleic acid triggers ER stress and autophagy in various human glioma cell lines^[112]. Furthermore, the inhibition of the proteasome system with bortezomib overcomes resistance in a variety of tumors via mechanisms mediated by the accumulation of misfolded proteins that overwhelm the ER-associated degradation pathway that produce ER stress^[113]. This mechanism is well described in multiple myeloma (MM) cells that constitutively express ER stress-associated survival factors that are essential for propagation and maintenance of MM cells^[114,115]. Thus, proteasome inhibitors induce apoptosis in MM because the UPR is

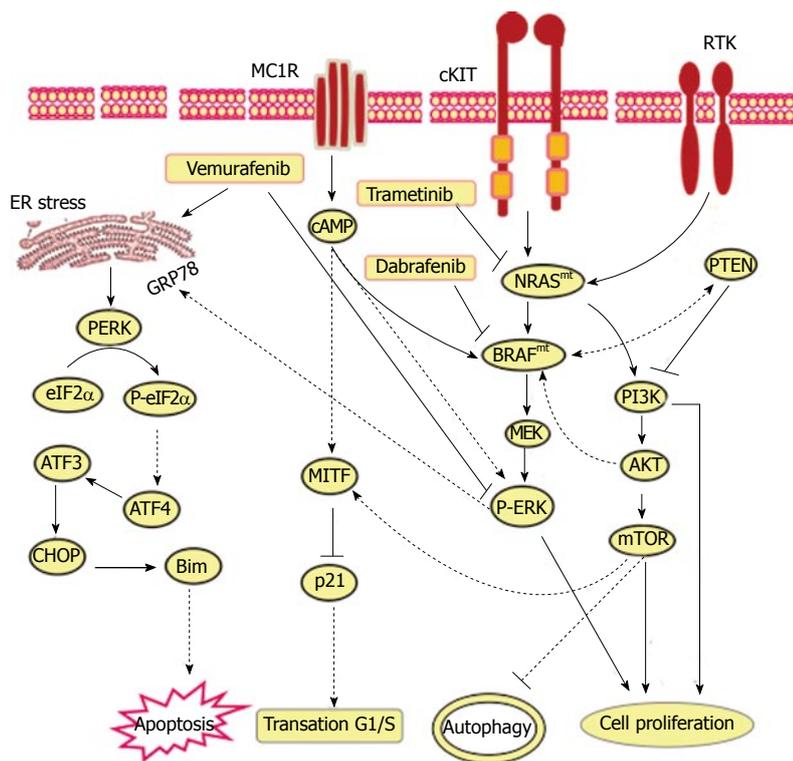


Figure 2 Proposed models for the mechanistic role of endoplasmic reticulum stress in the modulation of the anti-tumor efficiency of vemurafenib, dabrafenib and trametinib in melanoma patients harboring activating neuroblastoma RAS viral (v-ras) oncogene homolog or rapidly accelerated fibrosarcoma murine sarcoma viral (v-raf) oncogene homolog B mutations. The main function of MC1R, TRK on melanoma cell is to transmit extracellular signaling that is essential for the activation of RAS-RAF-MEK-ERK and PI3K-AKT-mTOR pathways to mediate various cellular functions including melanoma initiation, progression and resistance. Thus, the exposure of melanoma (BRAF^{mt} and NRAS^{mt}) cells to either Vemurafenib, Dabrafenib or Trametinib, respectively, results in inhibition of MC1R, cKIT and TRK-mediated activation of RAS-RAF-MEK-ERK and PI3K-AKT-mTOR pathways. As consequence the inhibition of RAS-RAF-MEK-ERK and PI3K-AKT-mTOR pathways results in the execution melanoma cell death via mechanism-mediated by ER stress-dependent pathways including PERK-eIF2α-ATF4-ATF3-CHOP-Bim pathway. NRAS: Neuroblastoma Rat Sarcoma viral (v-ras) oncogene homolog; BRAF: Rapidly Accelerated Fibrosarcoma murine sarcoma viral (v-raf) oncogene homolog B; MC1R: Melanocortin 1 receptor; TRK: CKIT and receptor tyrosine kinase; PI3K: Phosphatidylinositol-4,5-bisphosphate 3-kinase; AKT: Protein kinase B.

unable to mediate the degradation of the misfolded proteins^[116]. In fact, compared to other cell lines, MM cells are the most sensitive to proteasome inhibitors-induced apoptosis *via* mechanism mediated by the activation of UPR-associated pathways including PERK and ATF4, and the pro-apoptotic target, CHOP^[117].

The involvement of ER stress in the modulation of melanoma cell death in response to the treatment with anti-cancer agents has been studied extensively. For example, Syed *et al*^[118], demonstrated that fisetin-induced apoptosis of melanoma cells is mediated by ER stress-associated pathways such as IRE1α, XBP1s, ATF4 and GRP78^[119]. Also, the small molecule inhibitor honokiol, a potent anti-tumorigenic compound, has been shown to trigger apoptosis of melanoma cells *via* a mechanism mediated by the binding of honokiol to the unfolded ATPase domain of GRP78 leading to the induction of ER stress and pro-apoptotic associated pathways. Beck *et al*^[120], addressed an important role for ER stress-associated pathways in the modulation of the anti-cancer agents. For example, in patients with BRAFV600E-mutated melanoma vemurafenib-induced apoptosis is associated with increased levels of the spliced isoform of the transcription factor, XBP1, a marker for the induction of ER stress, and with increased phosphorylation of the translation initiation factor eIF2α. Also, ER stressors such as diallyl trisulfide play a role in the sensitization of melanoma cells to death receptor- induced apoptosis^[121]. Moreover, the role of ER stress-associated pathways in the modulation of the anti-tumor activity of the natural marine compound, 11-dehydrosinulariolide has been demonstrated^[122]. Interestingly, the 11-dehydrosinulariolide compound

was found to trigger apoptosis of melanoma cells *via* mechanism-mediated by both PERK/eIF2α/ATF4/CHOP and ATF6/CHOP pathways^[122]. In another study, Hiscutt *et al*^[123], demonstrated that knockdown of the X-linked inhibitor of apoptosis protein (XIAP) enhances both fenretinide and bortezomib-induced apoptosis of metastatic melanoma cells *via* ER stress-mediated pathways. Also, melanoma under ER stress shows more susceptibility to obatoclax-induced apoptosis^[124]. Moreover, the role of ER stress-associated signaling pathways, GRP78, ATF6, IRE1α, and PERK/eIF2α has been reported to be essential for docetaxel-induced apoptosis of melanoma^[125]. More importantly, it has been suggested that the constitutively activated MEK/ERK pathway results in resistance of melanoma cells to ER stress-induced apoptosis. Accordingly, Jiang *et al*^[48], demonstrated that the inhibition of MEK by U0126 inhibitor or by the knockdown of MEK1 by its specific siRNA sensitizes melanoma cells to tunicamycin- or thapsigargin-induced apoptosis. Also, the induction of ER stress by Tunicamycin can sensitize human melanoma cells to tumor necrosis factor-related apoptosis in response to ligand-induced apoptosis^[126].

CONCLUSION

Although it has been demonstrated that ER stress-dependent pathways play a significant role in the regulation of tumor initiation and resistance, it is more difficult to confirm the hypothesis that ER is a valid therapeutic target for tumor treatment. The induction of UPR is a cellular mechanism that reduces or prevents the cytotoxic effect of anti-cancer treatment.

Accordingly, the destruction of key UPR components should provide an effective therapeutic strategy for melanoma treatment. Moreover, a functional analysis of UPR-mediated pathways, particularly those which are essential for cell survival or cell death, may help to identify key molecules of the aberrant pathways whose excessive activation and/or inhibition may overcome melanoma resistance to standard treatments. In addition, gaining an understanding of the molecular mechanisms of UPR may provide insight into the development of therapeutic strategies such as the development of small molecule inhibitors to control melanoma through the modulation of UPR signaling. Just as most current melanoma therapies were developed following a functional analysis of their ability to trigger mitochondrial dysregulation, ER stress-dependent pathways could provide new therapeutic targets designed to effect key components of aberrant signaling pathways.

REFERENCES

- 1 **Chinembiri TN**, du Plessis LH, Gerber M, Hamman JH, du Plessis J. Review of natural compounds for potential skin cancer treatment. *Molecules* 2014; **19**: 11679-11721 [PMID: 25102117 DOI: 10.3390/molecules190811679]
- 2 **Murray C**, D'Intino Y, MacCormick R, Nassar B, Walsh N. Melanosis in association with metastatic malignant melanoma: report of a case and a unifying concept of pathogenesis. *Am J Dermatopathol* 1999; **21**: 28-30 [PMID: 10027522 DOI: 10.1097/0000372-199902000-00006]
- 3 **Čeović R**, Smolković N, Pašić A, Kostović K, Hrsan D. Multiple basal cell carcinomas of lower legs with stasis dermatitis: a therapeutic challenge. *Acta Dermatovenerol Croat* 2012; **20**: 191-196 [PMID: 23069306]
- 4 **Essner R**. Surgical treatment of malignant melanoma. *Surg Clin North Am* 2003; **83**: 109-156 [PMID: 12691453 DOI: 10.1016/S0039-6109(02)00205-0]
- 5 **Gipponi M**, Solari N, Giovinazzo D, Queirolo P, Bertoglio S, Villa G, Gualco M, Bleidl D, Cafiero F. The role of sentinel lymph node biopsy in patients with local recurrence or in-transit metastasis of melanoma. *Anticancer Res* 2014; **34**: 3197-3203 [PMID: 24922694]
- 6 **Larkin J**, Del Vecchio M, Ascierto PA, Krajsova I, Schachter J, Neyns B, Espinosa E, Garbe C, Sileni VC, Gogas H, Miller WH, Mandalà M, Hossers GA, Arance A, Queirolo P, Hauschild A, Brown MP, Mitchell L, Veronesi L, Blank CU. Vemurafenib in patients with BRAF(V600) mutated metastatic melanoma: an open-label, multicentre, safety study. *Lancet Oncol* 2014; **15**: 436-444 [PMID: 24582505 DOI: 10.1016/S1470-2045(14)70051-8]
- 7 **Bercolet A**, Arance A, Lopez Martin JA, Soriano V, Muñoz E, Alonso L, Espinosa E, Lopez Criado P, Valdivia J, Martin Algarra S. Ipilimumab for advanced melanoma: experience from the Spanish Expanded Access Program. *Melanoma Res* 2014; **24**: 577-583 [PMID: 25046550 DOI: 10.1097/CMR.000000000000108]
- 8 **Culos KA**, Cuellar S. Novel targets in the treatment of advanced melanoma: new first-line treatment options. *Ann Pharmacother* 2013; **47**: 519-526 [PMID: 23548648 DOI: 10.1345/aph.1R614]
- 9 **Güven K**, Kittler H, Wolff K, Pehamberger H. Cisplatin and carboplatin combination as second-line chemotherapy in dacarbazine-resistant melanoma patients. *Melanoma Res* 2001; **11**: 411-415 [PMID: 11479430 DOI: 10.1097/00008390-200108000-00012]
- 10 **Martin S**, Hill DS, Paton JC, Paton AW, Birch-Machin MA, Lovat PE, Redfern CP. Targeting GRP78 to enhance melanoma cell death. *Pigment Cell Melanoma Res* 2010; **23**: 675-682 [PMID: 20546536 DOI: 10.1111/j.1755-148X.2010.00731.x]
- 11 **El-Khattouti A**, Selimovic D, Haikel Y, Megahed M, Gomez CR, Hassan M. Identification and analysis of CD133(+) melanoma stem-like cells conferring resistance to taxol: An insight into the mechanisms of their resistance and response. *Cancer Lett* 2014; **343**: 123-133 [PMID: 24080340 DOI: 10.1016/j.canlet.2013.09.024]
- 12 **Selimovic D**, Badura HE, El-Khattouti A, Soell M, Porzig BB, Spernger A, Ghanjati F, Santourlidis S, Haikel Y, Hassan M. Vinblastine-induced apoptosis of melanoma cells is mediated by Ras homologous A protein (Rho A) via mitochondrial and non-mitochondrial-dependent mechanisms. *Apoptosis* 2013; **18**: 980-997 [PMID: 23564313 DOI: 10.1007/s10495-013-0844-4]
- 13 **Selimovic D**, Porzig BB, El-Khattouti A, Badura HE, Ahmad M, Ghanjati F, Santourlidis S, Haikel Y, Hassan M. Bortezomib/ proteasome inhibitor triggers both apoptosis and autophagy-dependent pathways in melanoma cells. *Cell Signal* 2013; **25**: 308-318 [PMID: 23079083 DOI: 10.5493/wjem.v2.i2.7]
- 14 **Bajetta E**, Del Vecchio M, Vitali M, Martinetti A, Ferrari L, Queirolo P, Sertoli MR, Cainelli T, Cellerino R, Cascinelli N. A feasibility study using polychemotherapy (cisplatin + vindesine + dacarbazine) plus interferon-alpha or monochemotherapy with dacarbazine plus interferon-alpha in metastatic melanoma. *Tumori* 2001; **87**: 219-222 [PMID: 11693798]
- 15 **Meckbach D**, Keim U, Richter S, Leiter U, Eigentler TK, Bauer J, Pflugfelder A, Büttner P, Garbe C, Weide B. BRAF-V600 mutations have no prognostic impact in stage IV melanoma patients treated with monochemotherapy. *PLoS One* 2014; **9**: e89218 [PMID: 24586605 DOI: 10.1371/journal.pone.0089218]
- 16 **Lee CK**, Jung M, Choi HJ, Kim HR, Kim HS, Roh MR, Ahn JB, Chung HC, Heo SJ, Rha SY, Shin SJ. Results of a Phase II Study to Evaluate the Efficacy of Docetaxel and Carboplatin in Metastatic Malignant Melanoma Patients Who Failed First-Line Therapy Containing Dacarbazine. *Cancer Res Treat* 2015; **47**: 781-789 [PMID: 25687848 DOI: 10.4143/crt.2014.261]
- 17 **Linardou H**, Pentheroudakis G, Vathalitis I, Gogas H, Pectasides D, Makatsoris T, Fountzilias G, Bafaloukos D. Predictive biomarkers to chemotherapy in patients with advanced melanoma receiving the combination of cisplatin--vinblastine--temozolomide (PVT) as first-line treatment: a study of the Hellenic Cooperative Oncology Group (HECOG). *Anticancer Res* 2015; **35**: 1105-1113 [PMID: 25667500]
- 18 **Lens MB**, Eisen TG. Systemic chemotherapy in the treatment of malignant melanoma. *Expert Opin Pharmacother* 2003; **4**: 2205-2211 [PMID: 14640919 DOI: 10.1517/14656566.4.12.2205]
- 19 **Queirolo P**, Picasso V, Spagnolo F. Combined BRAF and MEK inhibition for the treatment of BRAF-mutated metastatic melanoma. *Cancer Treat Rev* 2015; **41**: 519-526 [PMID: 25944484 DOI: 10.1016/j.ctrv.2015.04.010]
- 20 **Thu YM**, Su Y, Yang J, Splittgerber R, Na S, Boyd A, Mosse C, Simons C, Richmond A. NF-κB inducing kinase (NIK) modulates melanoma tumorigenesis by regulating expression of pro-survival factors through the β-catenin pathway. *Oncogene* 2012; **31**: 2580-2592 [PMID: 21963849 DOI: 10.1038/onc.2011.427]
- 21 **Kim A**, Son M, Kim KI, Yang Y, Song EY, Lee HG, Lim JS. Elevation of intracellular cyclic AMP inhibits NF-κB-mediated thymosin beta4 expression in melanoma cells. *Exp Cell Res* 2009; **315**: 3325-3335 [PMID: 19500569 DOI: 10.1016/j.yexcr.2009.05.024]
- 22 **Sims JT**, Ganguly SS, Bennett H, Friend JW, Tepe J, Plattner R. Imatinib reverses doxorubicin resistance by affecting activation of STAT3-dependent NF-κB and HSP27/p38/AKT pathways and by inhibiting ABCB1. *PLoS One* 2013; **8**: e55509 [PMID: 23383209 DOI: 10.1371/journal.pone.0055509]
- 23 **Levati L**, Ruffini F, Muzi A, Umezawa K, Graziani G, D'Atri S, Lacial PM. Placenta growth factor induces melanoma resistance to temozolomide through a mechanism that involves the activation of the transcription factor NF-κB. *Int J Oncol* 2011; **38**: 241-247 [PMID: 21109946 DOI: 10.3892/ijo.00000844]
- 24 **Lev DC**, Ruiz M, Mills L, McGary EC, Price JE, Bar-Eli M. Dacarbazine causes transcriptional up-regulation of interleukin 8 and vascular endothelial growth factor in melanoma cells: a possible escape mechanism from chemotherapy. *Mol Cancer Ther* 2003; **2**: 753-763 [PMID: 12939465]
- 25 **Poklepovic A**, Youssefian LE, Winning M, Birdsell CA, Crosby NA, Ramakrishnan V, Ernstoff MS, Roberts JD. Phase I trial of

- bortezomib and dacarbazine in melanoma and soft tissue sarcoma. *Invest New Drugs* 2013; **31**: 937-942 [PMID: 23315028 DOI: 10.1007/s10637-012-9913-8]
- 26 **Su Y**, Amiri KI, Horton LW, Yu Y, Ayers GD, Koehler E, Kelley MC, Puzanov I, Richmond A, Sosman JA. A phase I trial of bortezomib with temozolomide in patients with advanced melanoma: toxicities, antitumor effects, and modulation of therapeutic targets. *Clin Cancer Res* 2010; **16**: 348-357 [PMID: 20028756 DOI: 10.1158/1078-0432.CCR-09-2087]
- 27 **Macdonald JB**, Macdonald B, Golitz LE, LoRusso P, Sekulic A. Cutaneous adverse effects of targeted therapies: Part II: Inhibitors of intracellular molecular signaling pathways. *J Am Acad Dermatol* 2015; **72**: 221-236; quiz 237-238 [PMID: 25592339 DOI: 10.1016/j.jaad.2014.07.033]
- 28 **Kreuter A**, van Eijk T, Lehmann P, Fischer M, Horn T, Assaf C, Schley G, Herbst R, Kellner I, Weisbrich C, Hyun J, Wieland U, Schlaak M, Rübber A, Lommel K. Electrochemotherapy in advanced skin tumors and cutaneous metastases - a retrospective multicenter analysis. *J Dtsch Dermatol Ges* 2015; **13**: 308-315 [PMID: 25819239 DOI: 10.1111/ddg.12583]
- 29 **Chu SC**, Hsieh YS, Yu CC, Lai YY, Chen PN. Thymoquinone induces cell death in human squamous carcinoma cells via caspase activation-dependent apoptosis and LC3-II activation-dependent autophagy. *PLoS One* 2014; **9**: e101579 [PMID: 25000169 DOI: 10.1371/journal.pone.0101579]
- 30 **Wengrod J**, Wang D, Weiss S, Zhong H, Osman I, Gardner LB. Phosphorylation of eIF2 α triggered by mTORC1 inhibition and PP6C activation is required for autophagy and is aberrant in PP6C-mutated melanoma. *Sci Signal* 2015; **8**: ra27 [PMID: 25759478 DOI: 10.1126/scisignal.aaa0899]
- 31 **Dong H**, Tian L, Li R, Pei C, Fu Y, Dong X, Xia F, Wang C, Li W, Guo X, Gu C, Li B, Liu A, Ren H, Wang C, Xu H. IFN γ -induced Irgm1 promotes tumorigenesis of melanoma via dual regulation of apoptosis and Bif-1-dependent autophagy. *Oncogene* 2015; **34**: 5363-5371 [PMID: 25619828 DOI: 10.1038/ncr.2014.459]
- 32 **Marino ML**, Fais S, Djavaheri-Mergny M, Villa A, Meschini S, Lozupone F, Venturi G, Della Mina P, Patingre S, Rivoltini L, Codogno P, De Milito A. Proton pump inhibition induces autophagy as a survival mechanism following oxidative stress in human melanoma cells. *Cell Death Dis* 2010; **1**: e87 [PMID: 21368860 DOI: 10.1038/cddis.2010.67]
- 33 **Wroblewski D**, Jiang CC, Croft A, Farrelly ML, Zhang XD, Hersey P. OBATOCLAX and ABT-737 induce ER stress responses in human melanoma cells that limit induction of apoptosis. *PLoS One* 2013; **8**: e84073 [PMID: 24367627 DOI: 10.1371/journal.pone.0084073]
- 34 **Huang GJ**, Deng JS, Huang SS, Wang SY, Chang YS, Kuo YH. Bioassay guided isolation and identification of anti-inflammatory active compounds from the root of *Ficus formosana*. *J Agric Food Chem* 2013; **61**: 11008-11015 [PMID: 24200240 DOI: 10.1021/jf4033766]
- 35 **Wang X**, Ma M, Teng J, Zhang J, Zhou S, Zhang Y, Wu E, Ding X. Chronic exposure to cerebrospinal fluid of multiple system atrophy in neuroblastoma and glioblastoma cells induces cytotoxicity via ER stress and autophagy activation. *Oncotarget* 2015; **6**: 13278-13294 [PMID: 25965819]
- 36 **Shen Y**, Yang J, Zhao J, Xiao C, Xu C, Xiang Y. The switch from ER stress-induced apoptosis to autophagy via ROS-mediated JNK/p62 signals: A survival mechanism in methotrexate-resistant choriocarcinoma cells. *Exp Cell Res* 2015; **334**: 207-218 [PMID: 25912909 DOI: 10.1016/j.yexcr.2015.04.010]
- 37 **Bertrand L**, Toborek M. Dysregulation of Endoplasmic Reticulum Stress and Autophagic Responses by the Antiretroviral Drug Efavirenz. *Mol Pharmacol* 2015; **88**: 304-315 [PMID: 25987489 DOI: 10.1124/mol.115.098590]
- 38 **Liu H**, He Z, Simon HU. Targeting autophagy as a potential therapeutic approach for melanoma therapy. *Semin Cancer Biol* 2013; **23**: 352-360 [PMID: 23831275 DOI: 10.1016/j.semcancer.2013.06.008]
- 39 **Fecher LA**, Amaravadi RK, Schuchter LM, Flaherty KT. Drug targeting of oncogenic pathways in melanoma. *Hematol Oncol Clin North Am* 2009; **23**: 599-618, x [PMID: 19464605 DOI: 10.1016/j.hoc.2009.03.004]
- 40 **Roulstone V**, Pedersen M, Kyula J, Mansfield D, Khan AA, McEntee G, Wilkinson M, Karapanagiotou E, Coffey M, Marais R, Jebar A, Errington-Mais F, Melcher A, Vile R, Pandha H, McLaughlin M, Harrington KJ. BRAF- and MEK-Targeted Small Molecule Inhibitors Exert Enhanced Antimelanoma Effects in Combination With Oncolytic Reovirus Through ER Stress. *Mol Ther* 2015; **23**: 931-942 [PMID: 25619724 DOI: 10.1038/mt.2015.15]
- 41 **Goodall ML**, Wang T, Martin KR, Kortus MG, Kauffman AL, Trent JM, Gately S, MacKeigan JP. Development of potent autophagy inhibitors that sensitize oncogenic BRAF V600E mutant melanoma tumor cells to vemurafenib. *Autophagy* 2014; **10**: 1120-1136 [PMID: 24879157 DOI: 10.4161/auto.28594]
- 42 **Ho AL**, Musi E, Ambrosini G, Nair JS, Deraje Vasudeva S, de Stanchina E, Schwartz GK. Impact of combined mTOR and MEK inhibition in uveal melanoma is driven by tumor genotype. *PLoS One* 2012; **7**: e40439 [PMID: 22808163 DOI: 10.1371/journal.pone.0040439]
- 43 **Chatterjee SJ**, Pandey S. Chemo-resistant melanoma sensitized by tamoxifen to low dose curcumin treatment through induction of apoptosis and autophagy. *Cancer Biol Ther* 2011; **11**: 216-228 [PMID: 21088500 DOI: 10.4161/cbt.11.2.13798]
- 44 **Chatterjee S**, Willis N, Locks SM, Mott JH, Kelly CG. Dosimetric and radiobiological comparison of helical tomotherapy, forward-planned intensity-modulated radiotherapy and two-phase conformal plans for radical radiotherapy treatment of head and neck squamous cell carcinomas. *Br J Radiol* 2011; **84**: 1083-1090 [PMID: 22101580 DOI: 10.1259/bjr/53812025]
- 45 **Mohana-Kumaran N**, Hill DS, Allen JD, Haass NK. Targeting the intrinsic apoptosis pathway as a strategy for melanoma therapy. *Pigment Cell Melanoma Res* 2014; **27**: 525-539 [PMID: 24655414 DOI: 10.1111/pcmr.12242]
- 46 **Ma XH**, Piao SF, Dey S, McAfee Q, Karakousis G, Villanueva J, Hart LS, Levi S, Hu J, Zhang G, Lazova R, Klump V, Pawelek JM, Xu X, Xu W, Schuchter LM, Davies MA, Herlyn M, Winkler J, Koumenis C, Amaravadi RK. Targeting ER stress-induced autophagy overcomes BRAF inhibitor resistance in melanoma. *J Clin Invest* 2014; **124**: 1406-1417 [PMID: 24569374 DOI: 10.1172/JCI70454]
- 47 **Hawrock ST**, Carew JS, Pino MS, Highshaw RA, Dunner K, Nuang P, Abbruzzese JL, McConkey DJ. Bortezomib sensitizes pancreatic cancer cells to endoplasmic reticulum stress-mediated apoptosis. *Cancer Res* 2005; **65**: 11658-11666 [PMID: 16357177 DOI: 10.4161/cbt.11.2.13798]
- 48 **Jiang CC**, Chen LH, Gillespie S, Wang YF, Kiejda KA, Zhang XD, Hersey P. Inhibition of MEK sensitizes human melanoma cells to endoplasmic reticulum stress-induced apoptosis. *Cancer Res* 2007; **67**: 9750-9761 [PMID: 17942905 DOI: 10.1158/0008-5472.CAN-07-2047]
- 49 **Bakhshi J**, Weinstein L, Poksay KS, Nishinaga B, Bredesen DE, Rao RV. Coupling endoplasmic reticulum stress to the cell death program in mouse melanoma cells: effect of curcumin. *Apoptosis* 2008; **13**: 904-914 [PMID: 18493855 DOI: 10.1007/s10495-008-0221-x]
- 50 **Park SH**, Blackstone C. Further assembly required: construction and dynamics of the endoplasmic reticulum network. *EMBO Rep* 2010; **11**: 515-521 [PMID: 20559323 DOI: 10.1038/embor.2010.92]
- 51 **Chen X**, Karnovsky A, Sans MD, Andrews PC, Williams JA. Molecular characterization of the endoplasmic reticulum: insights from proteomic studies. *Proteomics* 2010; **10**: 4040-4052 [PMID: 21080494 DOI: 10.1002/pmic.201000234]
- 52 **Voisine C**, Pedersen JS, Morimoto RI. Chaperone networks: tipping the balance in protein folding diseases. *Neurobiol Dis* 2010; **40**: 12-20 [PMID: 20472062 DOI: 10.1016/j.nbd.2010.05.007]
- 53 **Luo B**, Lee AS. The critical roles of endoplasmic reticulum chaperones and unfolded protein response in tumorigenesis and anticancer therapies. *Oncogene* 2013; **32**: 805-818 [PMID: 22508478 DOI: 10.1038/ncr.2012.130]

- 54 **Stutzmann GE**, Mattson MP. Endoplasmic reticulum Ca(2+) handling in excitable cells in health and disease. *Pharmacol Rev* 2011; **63**: 700-727 [PMID: 21737534 DOI: 10.1124/pr.110.003814]
- 55 **Bravo R**, Parra V, Gatica D, Rodriguez AE, Torrealba N, Paredes F, Wang ZV, Zorzano A, Hill JA, Jaimovich E, Quest AF, Lavandero S. Endoplasmic reticulum and the unfolded protein response: dynamics and metabolic integration. *Int Rev Cell Mol Biol* 2013; **301**: 215-290 [PMID: 23317820 DOI: 10.1016/B978-0-12-407704-1.00005-1]
- 56 **Timmins JM**, Ozcan L, Seimon TA, Li G, Malagelada C, Backs J, Backs T, Bassel-Duby R, Olson EN, Anderson ME, Tabas I. Calcium/calmodulin-dependent protein kinase II links ER stress with Fas and mitochondrial apoptosis pathways. *J Clin Invest* 2009; **119**: 2925-2941 [PMID: 19741297 DOI: 10.1172/JCI38857]
- 57 **Kadowaki H**, Nishitoh H. Signaling pathways from the endoplasmic reticulum and their roles in disease. *Genes (Basel)* 2013; **4**: 306-333 [PMID: 24705207 DOI: 10.3390/genes4030306]
- 58 **Selimovic D**, Ahmad M, El-Khattouti A, Hannig M, Haikel Y, Hassan M. Apoptosis-related protein-2 triggers melanoma cell death by a mechanism including both endoplasmic reticulum stress and mitochondrial dysregulation. *Carcinogenesis* 2011; **32**: 1268-1278 [PMID: 21693538 DOI: 10.1093/carcin/bgr112]
- 59 **Bi K**, Nishihara K, Machleidt T, Hermanson S, Wang J, Sakamuru S, Huang R, Xia M. Identification of known drugs targeting the endoplasmic reticulum stress response. *Anal Bioanal Chem* 2015; **407**: 5343-5351 [PMID: 25925857]
- 60 **Shi Y**, Yang Y, Hoang B, Bardeleben C, Holmes B, Gera J, Lichtenstein A. Therapeutic potential of targeting IRES-dependent c-myc translation in multiple myeloma cells during ER stress. *Oncogene* 2015 May 11; Epub ahead of print [PMID: 25961916 DOI: 10.1007/s00216-015-8694-2]
- 61 **Selimovic D**, Hassan M, Haikel Y, Hengge UR. Taxol-induced mitochondrial stress in melanoma cells is mediated by activation of c-Jun N-terminal kinase (JNK) and p38 pathways via uncoupling protein 2. *Cell Signal* 2008; **20**: 311-322 [PMID: 18068334 DOI: 10.1016/j.cellsig.2007.10.015]
- 62 **El-Khattouti A**, Sheehan NT, Monico J, Drummond HA, Haikel Y, Brodell RT, Megahed M, Hassan M. CD133⁺ melanoma subpopulation acquired resistance to caffeic acid phenethyl ester-induced apoptosis is attributed to the elevated expression of ABCB5: significance for melanoma treatment. *Cancer Lett* 2015; **357**: 83-104 [PMID: 25449786 DOI: 10.1016/j.canlet.2014.10.043]
- 63 **Suga N**, Gao E, Zhang Y, Ma X, Olsen JF. The corticofugal system for hearing: recent progress. *Proc Natl Acad Sci USA* 2000; **97**: 11807-11814 [PMID: 11050213 DOI: 10.1073/pnas.97.22.11807]
- 64 **Galehdar Z**, Swan P, Fuerth B, Callaghan SM, Park DS, Cregan SP. Neuronal apoptosis induced by endoplasmic reticulum stress is regulated by ATF4-CHOP-mediated induction of the Bcl-2 homology 3-only member PUMA. *J Neurosci* 2010; **30**: 16938-16948 [PMID: 21159964 DOI: 10.1523/JNEUROSCI.1598-10.2010]
- 65 **Tohmonda T**, Yoda M, Mizuochi H, Morioka H, Matsumoto M, Urano F, Toyama Y, Horiuchi K. The IRE1 α -XBP1 pathway positively regulates parathyroid hormone (PTH)/PTH-related peptide receptor expression and is involved in pth-induced osteoclastogenesis. *J Biol Chem* 2013; **288**: 1691-1695 [PMID: 23235147 DOI: 10.1074/jbc.C112.424606]
- 66 **Back SH**, Lee K, Vink E, Kaufman RJ. Cytoplasmic IRE1 α -mediated XBP1 mRNA splicing in the absence of nuclear processing and endoplasmic reticulum stress. *J Biol Chem* 2006; **281**: 18691-18706 [PMID: 16644724 DOI: 10.1074/jbc.M602030200]
- 67 **Vaeteewoottacharn K**, Kariya R, Matsuda K, Taura M, Wongkham C, Wongkham S, Okada S. Perturbation of proteasome function by bortezomib leading to ER stress-induced apoptotic cell death in cholangiocarcinoma. *J Cancer Res Clin Oncol* 2013; **139**: 1551-1562 [PMID: 23877657 DOI: 10.1007/s00432-013-1473-6]
- 68 **Allagnat F**, Christulia F, Ortis F, Pirot P, Lortz S, Lenzen S, Eizirik DL, Cardozo AK. Sustained production of spliced X-box binding protein 1 (XBP1) induces pancreatic beta cell dysfunction and apoptosis. *Diabetologia* 2010; **53**: 1120-1130 [PMID: 20349222 DOI: 10.1007/s00125-010-1699-7]
- 69 **Prasanthi JR**, Larson T, Schommer J, Ghribi O. Silencing GADD153/CHOP gene expression protects against Alzheimer's disease-like pathology induced by 27-hydroxycholesterol in rabbit hippocampus. *PLoS One* 2011; **6**: e26420 [PMID: 22046282 DOI: 10.1371/journal.pone.0026420]
- 70 **Hassan M**, Alaoui A, Feyen O, Mirmohammadsadegh A, Essmann F, Tannappel A, Gulbins E, Schulze-Osthoff K, Hengge UR. The BH3-only member Noxa causes apoptosis in melanoma cells by multiple pathways. *Oncogene* 2008; **27**: 4557-4568 [PMID: 18408751 DOI: 10.1038/onc.2008.90]
- 71 **Xi H**, Barredo JC, Merchan JR, Lampidis TJ. Endoplasmic reticulum stress induced by 2-deoxyglucose but not glucose starvation activates AMPK through CaMKK β leading to autophagy. *Biochem Pharmacol* 2013; **85**: 1463-1477 [PMID: 23500541 DOI: 10.1016/j.bcp.2013.02.037]
- 72 **Tay KH**, Luan Q, Croft A, Jiang CC, Jin L, Zhang XD, Tseng HY. Sustained IRE1 and ATF6 signaling is important for survival of melanoma cells undergoing ER stress. *Cell Signal* 2014; **26**: 287-294 [PMID: 24240056 DOI: 10.1016/j.cellsig.2013.11.008]
- 73 **Croft A**, Tay KH, Boyd SC, Guo ST, Jiang CC, Lai F, Tseng HY, Jin L, Rizos H, Hersey P, Zhang XD. Oncogenic activation of MEK/ERK primes melanoma cells for adaptation to endoplasmic reticulum stress. *J Invest Dermatol* 2014; **134**: 488-497 [PMID: 23921951 DOI: 10.1038/jid.2013.325]
- 74 **Huang TT**, Lin HC, Chen CC, Lu CC, Wei CF, Wu TS, Liu FG, Lai HC. Resveratrol induces apoptosis of human nasopharyngeal carcinoma cells via activation of multiple apoptotic pathways. *J Cell Physiol* 2011; **226**: 720-728 [PMID: 20717957 DOI: 10.1002/jcp.22391]
- 75 **Han X**, Zhou J, Zhang P, Song F, Jiang R, Li M, Xia F, Guo FJ. IRE1 α dissociates with BiP and inhibits ER stress-mediated apoptosis in cartilage development. *Cell Signal* 2013; **25**: 2136-2146 [PMID: 23816533 DOI: 10.1016/j.cellsig.2013.06.011]
- 76 **Kim HS**, Kim TJ, Yoo YM. Melatonin combined with endoplasmic reticulum stress induces cell death via the PI3K/Akt/mTOR pathway in B16F10 melanoma cells. *PLoS One* 2014; **9**: e92627 [PMID: 24647338 DOI: 10.1371/journal.pone.0092627]
- 77 **Hu MC**, Gong HY, Lin GH, Hu SY, Chen MH, Huang SJ, Liao CF, Wu JL. XBP-1, a key regulator of unfolded protein response, activates transcription of IGF1 and Akt phosphorylation in zebrafish embryonic cell line. *Biochem Biophys Res Commun* 2007; **359**: 778-783 [PMID: 17560942 DOI: 10.1016/j.bbrc.2007.05.183]
- 78 **Thorpe JA**, Schwarze SR. IRE1 α controls cyclin A1 expression and promotes cell proliferation through XBP-1. *Cell Stress Chaperones* 2010; **15**: 497-508 [PMID: 20013084 DOI: 10.1007/s12192-009-0163-4]
- 79 **Guichard C**, Peduzzi E, Fay M, Marie JC, Braut-Boucher F, Daniel F, Grodet A, Gougerot-Pocidallo MA, Chastre E, Kotelevets L, Lizard G, Vandewalle A, Driss F, Ogier-Denis E. Dihydroxyphenylethanol induces apoptosis by activating serine/threonine protein phosphatase PP2A and promotes the endoplasmic reticulum stress response in human colon carcinoma cells. *Carcinogenesis* 2006; **27**: 1812-1827 [PMID: 16524888 DOI: 10.1093/carcin/bgl009]
- 80 **Zhang H**, Nakajima S, Kato H, Gu L, Yoshitomi T, Nagai K, Shinmori H, Kokubo S, Kitamura M. Selective, potent blockade of the IRE1 and ATF6 pathways by 4-phenylbutyric acid analogues. *Br J Pharmacol* 2013; **170**: 822-834 [PMID: 23869584 DOI: 10.1111/bph.12306]
- 81 **Zhang C**, Kawauchi J, Adachi MT, Hashimoto Y, Oshiro S, Aso T, Kitajima S. Activation of JNK and transcriptional repressor ATF3/LRF1 through the IRE1/TRAF2 pathway is implicated in human vascular endothelial cell death by homocysteine. *Biochem Biophys Res Commun* 2001; **289**: 718-724 [PMID: 11726207 DOI: 10.1006/bbrc.2001.6044]
- 82 **Law BY**, Chan WK, Xu SW, Wang JR, Bai LP, Liu L, Wong VK. Natural small-molecule enhancers of autophagy induce autophagic cell death in apoptosis-defective cells. *Sci Rep* 2014; **4**: 5510 [PMID: 24981420 DOI: 10.1038/srep05510]
- 83 **Armstrong JL**, Corazzari M, Martin S, Pagliarini V, Falasca L, Hill DS, Ellis N, Al Sabah S, Redfern CP, Fimia GM, Piacentini

- M, Lovat PE. Oncogenic B-RAF signaling in melanoma impairs the therapeutic advantage of autophagy inhibition. *Clin Cancer Res* 2011; **17**: 2216-2226 [PMID: 21270111 DOI: 10.1158/1078-0432.CCR-10-3003]
- 84 **Xie Z**, Xie Y, Xu Y, Zhou H, Xu W, Dong Q. Bafilomycin A1 inhibits autophagy and induces apoptosis in MG63 osteosarcoma cells. *Mol Med Rep* 2014; **10**: 1103-1107 [PMID: 24890793 DOI: 10.3892/mmr.2014.2281]
- 85 **Orlotti NI**, Cimino-Reale G, Borghini E, Pennati M, Sissi C, Perrone F, Palumbo M, Daidone MG, Folini M, Zaffaroni N. Autophagy acts as a safeguard mechanism against G-quadruplex ligand-mediated DNA damage. *Autophagy* 2012; **8**: 1185-1196 [PMID: 22627293 DOI: 10.4161/auto.20519]
- 86 **Rebecca VW**, Massaro RR, Fedorenko IV, Sondak VK, Anderson AR, Kim E, Amaravadi RK, Maria-Engler SS, Messina JL, Gibney GT, Kudchadkar RR, Smalley KS. Inhibition of autophagy enhances the effects of the AKT inhibitor MK-2206 when combined with paclitaxel and carboplatin in BRAF wild-type melanoma. *Pigment Cell Melanoma Res* 2014; **27**: 465-478 [PMID: 24490764 DOI: 10.1111/pcmr.12227]
- 87 **Soares AS**, Costa VM, Diniz C, Fresco P. Combination of CI-IB-MECA with paclitaxel is a highly effective cytotoxic therapy causing mTOR-dependent autophagy and mitotic catastrophe on human melanoma cells. *J Cancer Res Clin Oncol* 2014; **140**: 921-935 [PMID: 24659394 DOI: 10.1007/s00432-014-1645-z]
- 88 **Biggers JW**, Nguyen T, Di X, Gupton JT, Henderson SC, Emery SM, Alotaibi M, White KL, Brown R, Almenara J, Gewirtz DA. Autophagy, cell death and sustained senescence arrest in B16/F10 melanoma cells and HCT-116 colon carcinoma cells in response to the novel microtubule poison, JG-03-14. *Cancer Chemother Pharmacol* 2013; **71**: 441-455 [PMID: 23178952 DOI: 10.1007/s00280-012-2024-6]
- 89 **Zhang L**, Wang H, Xu J, Zhu J, Ding K. Inhibition of cathepsin S induces autophagy and apoptosis in human glioblastoma cell lines through ROS-mediated PI3K/AKT/mTOR/p70S6K and JNK signaling pathways. *Toxicol Lett* 2014; **228**: 248-259 [PMID: 24875536 DOI: 10.1016/j.toxlet.2014.05.015]
- 90 **Xie X**, White EP, Mehnert JM. Coordinate autophagy and mTOR pathway inhibition enhances cell death in melanoma. *PLoS One* 2013; **8**: e55096 [PMID: 23383069 DOI: 10.1371/journal.pone.0055096]
- 91 **Platz A**, Egyhazi S, Ringborg U, Hansson J. Human cutaneous melanoma; a review of NRAS and BRAF mutation frequencies in relation to histogenetic subclass and body site. *Mol Oncol* 2008; **1**: 395-405 [PMID: 19383313 DOI: 10.1016/j.molonc.2007.12.003]
- 92 **Tsao H**, Goel V, Wu H, Yang G, Haluska FG. Genetic interaction between NRAS and BRAF mutations and PTEN/MMAC1 inactivation in melanoma. *J Invest Dermatol* 2004; **122**: 337-341 [PMID: 15009714 DOI: 10.1046/j.0022-202X.2004.22243.x]
- 93 **Sensi M**, Nicolini G, Petti C, Bersani I, Lozupone F, Molla A, Vegetti C, Nonaka D, Mortarini R, Parmiani G, Fais S, Anichini A. Mutually exclusive NRASQ61R and BRAFV600E mutations at the single-cell level in the same human melanoma. *Oncogene* 2006; **25**: 3357-3364 [PMID: 16462768 DOI: 10.1038/sj.onc.1209379]
- 94 **Eisenhardt AE**, Olbrich H, Röring M, Janzarik W, Anh TN, Cin H, Remke M, Witt H, Korshunov A, Pfister SM, Omran H, Brummer T. Functional characterization of a BRAF insertion mutant associated with pilocytic astrocytoma. *Int J Cancer* 2011; **129**: 2297-2303 [PMID: 21190184 DOI: 10.1002/ijc.25893]
- 95 **Maldonado JL**, Fridlyand J, Patel H, Jain AN, Busam K, Kageshita T, Ono T, Albertson DG, Pinkel D, Bastian BC. Determinants of BRAF mutations in primary melanomas. *J Natl Cancer Inst* 2003; **95**: 1878-1890 [PMID: 14679157 DOI: 10.1093/jnci/djg123]
- 96 **Deichmann M**, Thome M, Benner A, Kirschner M, Hassanzadeh J, Kurzen H. Preponderance of the oncogenic V599E and V599K mutations in B-raf kinase domain is enhanced in melanoma cutaneous/subcutaneous metastases. *BMC Cancer* 2005; **5**: 58 [PMID: 15935100 DOI: 10.1186/1471-2407-5-58]
- 97 **Dumaz N**, Hayward R, Martin J, Ogilvie L, Hedley D, Curtin JA, Bastian BC, Springer C, Marais R. In melanoma, RAS mutations are accompanied by switching signaling from BRAF to CRAF and disrupted cyclic AMP signaling. *Cancer Res* 2006; **66**: 9483-9491 [PMID: 17018604 DOI: 10.1158/0008-5472.CAN-05-4227]
- 98 **Qin J**, Xin H, Nickoloff BJ. Specifically targeting ERK1 or ERK2 kills melanoma cells. *J Transl Med* 2012; **10**: 15 [PMID: 22277029 DOI: 10.1186/1479-5876-10-15]
- 99 **Peters S**, Bouchaab H, Zimmermann S, Bucher M, Gaide O, Letovanec I, Homicsko K, Michielin O. Dramatic response of vemurafenib-induced cutaneous lesions upon switch to dual BRAF/MEK inhibition in a metastatic melanoma patient. *Melanoma Res* 2014; **24**: 496-500 [PMID: 25185693 DOI: 10.1097/CMR.0000000000000055]
- 100 **Anforth R**, Carlos G, Clements A, Kefford R, Fernandez-Peñas P. Cutaneous adverse events in patients treated with BRAF inhibitor-based therapies for metastatic melanoma for longer than 52 weeks. *Br J Dermatol* 2015; **172**: 239-243 [PMID: 25040674 DOI: 10.1111/bjd.13200]
- 101 **Urner-Bloch U**, Urner M, Stieger P, Galliker N, Winterton N, Zübel A, Moutouh-de Parseval L, Dummer R, Goldinger SM. Transient MEK inhibitor-associated retinopathy in metastatic melanoma. *Ann Oncol* 2014; **25**: 1437-1441 [PMID: 24864047 DOI: 10.1093/annonc/mdu169]
- 102 **Uribe P**, Anforth RM, Kefford RF, Fernandez-Peñas P. Acneiform eruption in a patient with metastatic melanoma after ceasing combination dabrafenib/trametinib therapy. *Melanoma Res* 2014; **24**: 501-503 [PMID: 24922191 DOI: 10.1097/CMR.0000000000000096]
- 103 **Okwan-Duodu D**, Pollack BP, Lawson D, Khan MK. Role of radiation therapy as immune activator in the era of modern immunotherapy for metastatic malignant melanoma. *Am J Clin Oncol* 2015; **38**: 119-125 [PMID: 23648438 DOI: 10.1097/COC.0b013e3182940dc3]
- 104 **Maddodi N**, Huang W, Havighurst T, Kim K, Longley BJ, Setaluri V. Induction of autophagy and inhibition of melanoma growth in vitro and in vivo by hyperactivation of oncogenic BRAF. *J Invest Dermatol* 2010; **130**: 1657-1667 [PMID: 20182446 DOI: 10.1038/jid.2010.26]
- 105 **Stefanis L**, Larsen KE, Rideout HJ, Sulzer D, Greene LA. Expression of A53T mutant but not wild-type alpha-synuclein in PC12 cells induces alterations of the ubiquitin-dependent degradation system, loss of dopamine release, and autophagic cell death. *J Neurosci* 2001; **21**: 9549-9560 [PMID: 11739566 DOI: 10.1046/j.1471-4159.2001.00474.x]
- 106 **Lv S**, Wang X, Zhang N, Sun M, Qi W, Li Y, Yang Q. Autophagy facilitates the development of resistance to the tumor necrosis factor superfamily member TRAIL in breast cancer. *Int J Oncol* 2015; **46**: 1286-1294 [PMID: 25572822 DOI: 10.3892/ijo.2014.2812]
- 107 **He H**, Zang LH, Feng YS, Chen LX, Kang N, Tashiro S, Onodera S, Qiu F, Ikejima T. Physalin A induces apoptosis via p53-Noxa-mediated ROS generation, and autophagy plays a protective role against apoptosis through p38-NF-κB survival pathway in A375-S2 cells. *J Ethnopharmacol* 2013; **148**: 544-555 [PMID: 23684722 DOI: 10.1016/j.jep.2013.04.051]
- 108 **Escalante AM**, McGrath RT, Karolak MR, Dorr RT, Lynch RM, Landowski TH. Preventing the autophagic survival response by inhibition of calpain enhances the cytotoxic activity of bortezomib in vitro and in vivo. *Cancer Chemother Pharmacol* 2013; **71**: 1567-1576 [PMID: 23572175 DOI: 10.1007/s00280-013-2156-3]
- 109 **Zalckvar E**, Berissi H, Mizrachy L, Idelchuk Y, Koren I, Eisenstein M, Sabanay H, Pinkas-Kramarski R, Kimchi A. DAP-kinase-mediated phosphorylation on the BH3 domain of beclin 1 promotes dissociation of beclin 1 from Bcl-XL and induction of autophagy. *EMBO Rep* 2009; **10**: 285-292 [PMID: 19180116 DOI: 10.1038/embor.2008.246]
- 110 **Meng X**, Leyva ML, Jenny M, Gross I, Benosman S, Fricker B, Harlepp S, Hébraud P, Boos A, Wlosik P, Bischoff P, Sirlin C, Pfeffer M, Loeffler JP, Gaidon C. A ruthenium-containing organometallic compound reduces tumor growth through induction of the endoplasmic reticulum stress gene CHOP. *Cancer Res* 2009; **69**: 5458-5466 [PMID: 19549908 DOI: 10.1158/0008-5472.CAN-08-4408]
- 111 **Gaidon C**, Jeannequin P, Bischoff P, Pfeffer M, Sirlin C, Loeffler JP. Ruthenium (II)-derived organometallic compounds induce

- cytostatic and cytotoxic effects on mammalian cancer cell lines through p53-dependent and p53-independent mechanisms. *J Pharmacol Exp Ther* 2005; **315**: 1403-1411 [PMID: 16169939]
- 112 **Marcilla-Etxenike A**, Martín ML, Noguera-Salvà MA, García-Verdugo JM, Soriano-Navarro M, Dey I, Escribá PV, Busquets X. 2-Hydroxyoleic acid induces ER stress and autophagy in various human glioma cell lines. *PLoS One* 2012; **7**: e48235 [PMID: 23133576 DOI: 10.1371/journal.pone.0048235]
- 113 **Brem GJ**, Mylonas I, Brüning A. Eeyarestatin causes cervical cancer cell sensitization to bortezomib treatment by augmenting ER stress and CHOP expression. *Gynecol Oncol* 2013; **128**: 383-390 [PMID: 23107612 DOI: 10.1016/j.ygyno.2012.10.021]
- 114 **Cea M**, Cagnetta A, Fulciniti M, Tai YT, Hideshima T, Chauhan D, Roccaro A, Sacco A, Calimeri T, Cottini F, Jakubikova J, Kong SY, Patrone F, Nencioni A, Gobbi M, Richardson P, Munshi N, Anderson KC. Targeting NAD⁺ salvage pathway induces autophagy in multiple myeloma cells via mTORC1 and extracellular signal-regulated kinase (ERK1/2) inhibition. *Blood* 2012; **120**: 3519-3529 [PMID: 22955917 DOI: 10.1182/blood-2012-03-416776]
- 115 **Manni S**, Brancalion A, Mandato E, Tubi LQ, Colpo A, Pizzi M, Cappellesso R, Zaffino F, Di Maggio SA, Cabrelle A, Marino F, Zambello R, Trentin L, Adami F, Gurrieri C, Semenzato G, Piazza F. Protein kinase CK2 inhibition down modulates the NF- κ B and STAT3 survival pathways, enhances the cellular proteotoxic stress and synergistically boosts the cytotoxic effect of bortezomib on multiple myeloma and mantle cell lymphoma cells. *PLoS One* 2013; **8**: e75280 [PMID: 24086494 DOI: 10.1371/journal.pone.0075280]
- 116 **Obeng EA**, Carlson LM, Gutman DM, Harrington WJ, Lee KP, Boise LH. Proteasome inhibitors induce a terminal unfolded protein response in multiple myeloma cells. *Blood* 2006; **107**: 4907-4916 [PMID: 16507771 DOI: 10.1182/blood-2005-08-3531]
- 117 **Jiang Q**, Li F, Shi K, Wu P, An J, Yang Y, Xu C. Involvement of p38 in signal switching from autophagy to apoptosis via the PERK/eIF2 α /ATF4 axis in selenite-treated NB4 cells. *Cell Death Dis* 2014; **5**: e1270 [PMID: 24874742 DOI: 10.1038/cddis.2014.200]
- 118 **Syed DN**, Lall RK, Chamcheu JC, Haidar O, Mukhtar H. Involvement of ER stress and activation of apoptotic pathways in fisetin induced cytotoxicity in human melanoma. *Arch Biochem Biophys* 2014; **563**: 108-117 [PMID: 25016296 DOI: 10.1016/j.abb.2014.06.034]
- 119 **Martin S**, Lamb HK, Brady C, Lefkove B, Bonner MY, Thompson P, Lovat PE, Arbiser JL, Hawkins AR, Redfern CP. Inducing apoptosis of cancer cells using small-molecule plant compounds that bind to GRP78. *Br J Cancer* 2013; **109**: 433-443 [PMID: 23807168 DOI: 10.1038/bjc.2013.325]
- 120 **Beck D**, Niessner H, Smalley KS, Flaherty K, Paraiso KH, Busch C, Sinnberg T, Vasseur S, Iovanna JL, Drießen S, Stork B, Wesselborg S, Schaller M, Biedermann T, Bauer J, Lasithiotakis K, Weide B, Eberle J, Schitteck B, Schadendorf D, Garbe C, Kulms D, Meier F. Vemurafenib potently induces endoplasmic reticulum stress-mediated apoptosis in BRAFV600E melanoma cells. *Sci Signal* 2013; **6**: ra7 [PMID: 23362240 DOI: 10.1126/scisignal.2003057]
- 121 **Murai M**, Inoue T, Suzuki-Karasaki M, Ochiai T, Ra C, Nishida S, Suzuki-Karasaki Y. Diallyl trisulfide sensitizes human melanoma cells to TRAIL-induced cell death by promoting endoplasmic reticulum-mediated apoptosis. *Int J Oncol* 2012; **41**: 2029-2037 [PMID: 23064375 DOI: 10.3892/ijo.2012.1656]
- 122 **Su TR**, Tsai FJ, Lin JJ, Huang HH, Chiu CC, Su JH, Yang YT, Chen JY, Wong BS, Wu YJ. Induction of apoptosis by 11-dehydrosinulariolide via mitochondrial dysregulation and ER stress pathways in human melanoma cells. *Mar Drugs* 2012; **10**: 1883-1898 [PMID: 23015779 DOI: 10.3390/md10081883]
- 123 **Hiscutt EL**, Hill DS, Martin S, Kerr R, Harbottle A, Birch-Machin M, Redfern CP, Fulda S, Armstrong JL, Lovat PE. Targeting X-linked inhibitor of apoptosis protein to increase the efficacy of endoplasmic reticulum stress-induced apoptosis for melanoma therapy. *J Invest Dermatol* 2010; **130**: 2250-2258 [PMID: 20520630 DOI: 10.1038/jid.2010.146]
- 124 **Jiang CC**, Wroblewski D, Yang F, Hersey P, Zhang XD. Human melanoma cells under endoplasmic reticulum stress are more susceptible to apoptosis induced by the BH3 mimetic obatoclax. *Neoplasia* 2009; **11**: 945-955 [PMID: 19724688 DOI: 10.1593/neo.09692]
- 125 **Mhaidat NM**, Thorne R, Zhang XD, Hersey P. Involvement of endoplasmic reticulum stress in Docetaxel-induced JNK-dependent apoptosis of human melanoma. *Apoptosis* 2008; **13**: 1505-1512 [PMID: 18989785 DOI: 10.1007/s10495-008-0276-8]
- 126 **Jiang CC**, Chen LH, Gillespie S, Kiejda KA, Mhaidat N, Wang YF, Thorne R, Zhang XD, Hersey P. Tunicamycin sensitizes human melanoma cells to tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis by up-regulation of TRAIL-R2 via the unfolded protein response. *Cancer Res* 2007; **67**: 5880-5888 [PMID: 17575157 DOI: 10.1158/0008-5472.CAN-07-0213]

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Enzymatic antioxidant system in vascular inflammation and coronary artery disease

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Abstract

In biological systems there is a balance between the production and neutralization of reactive oxygen species (ROS). This balance is maintained by the presence of natural antioxidants and antioxidant enzymes such

as superoxide dismutase (SOD), catalase and glutathione peroxidase. The enhancement of lipid peroxidation or the decrease of antioxidant protection present in metabolic diseases or bad lifestyle can induce endothelial dysfunction and atherosclerosis. Clinical studies have shown that oxidative stress can increase ROS reducing the formation of antioxidant defences, especially in subjects with coronary artery disease (CAD). Some observation indicated that in the early stages of the disease there is a homeostatic up-regulation of the antioxidant enzyme system in response to increased free radicals to prevent vascular damage. As soon as free radicals get to chronically elevated levels, this compensation ceases. Therefore, SOD and the other enzymes may represent a good therapeutic target against ROS, but they are not useful markers for the diagnosis of CAD. In conclusion antioxidant enzymes are reduced in presence of metabolic disease and CAD. However the existence of genes that promote their enzymatic activity could contribute to create new drugs for the treatment of damage caused by metabolic diseases or lifestyle that increases the plasma ROS levels.

Key words: Superoxide dismutase; Catalase; Glutathione peroxidase; Antioxidant enzyme; Coronary artery disease; Reactive oxygen species; Vascular inflammation

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Core tip: This review shows that antioxidant enzymes are very important factors for the prevention and treatment of atherosclerotic disease, but more studies are required to understand whether they can be used as markers for diagnosis of coronary artery disease. The presence of polymorphic genes that increases the activity and expression of these enzymes can be considered important for the development of new therapeutic strategies. In our opinion further efforts should be directed especially on this last point, in

order to find new therapies to increase the function of antioxidant enzymes in metabolic disease or other risk factors.

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INTRODUCTION

Oxygen has played a crucial role in the evolution^[1-2] inducing the aerobic organisms to develop an adaptation to its toxicity by the presence of antioxidant systems. Oxygen is always metabolized to produce oxygen derived free radicals^[3] (superoxide $O_2^{\cdot-}$, hydroxyl OH^{\cdot}) and non-radical (hydrogen peroxide H_2O_2) all termed reactive oxygen species (ROS). In physiological condition there is a balance between the production and neutralization of ROS^[4]. Small amount of ROS are constantly generated^[5] and may often be useful for the immune system^[6] and defense against microorganisms^[5]. Conversely, high doses of ROS determine oxidative stress responsible for serious metabolic dysfunctions and damage to biological macromolecules^[7]. The enhancement of lipid peroxidation or a decrease in antioxidant protection can frequently induce the reaction with the nucleophilic centers of the DNA, RNA and proteins leading to irreversible damage such as cytotoxicity, mutagenicity and carcinogenicity. For instance, intracellular $O_2^{\cdot-}$, hydroxyl radical (OH^{\cdot}) and H_2O_2 play an important role in endothelial dysfunction, hypertension and atherosclerosis, inducing the expression of ICAM-1 and monocyte adhesion in endothelial cells^[8,9]. To minimize the damage caused by free radicals the organism utilize enzymatic and non-enzymatic antioxidant systems. Of the first group are the superoxide dismutase (SOD), glutathione peroxidase (GPX), catalase (CAT), glutathione (GSH), while the second group consists of vitamin A, ascorbic acid (vitamin C) and alpha-tocopherol (vitamin E)^[10]. The fact that the activity of SOD is more intense in humans compared to other species could explain the longevity of our species suggesting that humans have better protection against ROS^[11,12]. The imbalance between pro-oxidant and antioxidant systems can occur by an overproduction of ROS, (as the radical $O_2^{\cdot-}$, or OH^{\cdot}), or because of the drastic reduction of antioxidant systems. Among the main sources of generation of ROS are the mitochondrial electron transport chain^[13], the system of NADPH oxidase and nitric oxide synthase (Figure 1). It has been known for years that NAD(P)H oxidase is a major source of superoxide in vascular tissue^[14,15] and in cardiac cells^[16].



And it has been demonstrated that its activity is increased by angiotensin II^[17], thrombin, platelet-derived growth factor (PDGF), tumor necrosis factor- α (TNF- α) and lactosylceramide^[18-20].

ENZYMATIC ANTIOXIDANT SYSTEM

SOD converts the highly reactive radical $O_2^{\cdot-}$ to the less reactive radical H_2O_2 , which in turn can be destroyed by CAT or GPX, protecting the dehydratase (dehydratase hydroxyacid, aconitase, 6 phosphogluconate dehydratase, fumarase A and B). In humans, there are three forms of SOD: cytosolic (Cu, Zn-SOD), mitochondrial (Mn-SOD) and extracellular (EC-SOD)^[21]. The respiratory chain in the mitochondria is the major source of oxygen radicals. Mn-SOD is of primary importance in removing $O_2^{\cdot-}$ ^[22] and is essential for life. Cu, Zn-SOD seems to play an important role in the first line of antioxidant defense catalyzing the dismutation of $O_2^{\cdot-}$ radicals to form H_2O_2 and molecular oxygen, however, knock-outs experiments have shown that it is not essential for life^[23]. EC-SOD is a tetrameric glycoprotein containing zinc and copper that has a high affinity for heparin. In mammalian tissues it is regulated by cytokines^[24].

CAT consists of 4 ferriprotoporphyrin groups per molecule, is known as the most efficient enzyme since it is never saturated by the presence of H_2O_2 ^[7]. CAT reacts with H_2O_2 and with proton donors (ROOH) producing H_2O . CAT protects the cells from the production of H_2O_2 playing an important role in the acquisition of tolerance to oxidative stress as an adaptive response of the cells^[25].

GPX also catalyzes the reduction of a variety of hydroperoxides using GSH (ROOH and H_2O_2). The cells that contain low levels of GPX are much more susceptible to the toxicity of compounds such as adriamycin which produces hydroperoxides^[26] and seems important as a line of defense against peroxynitrite.

ASSOCIATION BETWEEN ANTIOXIDANT ENZYME AND VASCULAR DISEASE

The increased oxidative stress is associated with the pathogenesis of coronary artery disease (CAD)^[27,28]. Clinical studies have shown that oxidative stress can increase ROS reducing the formation of antioxidant defenses^[27,28]. Some authors have demonstrated that the reduction of activity of antioxidant enzymes such as CAT, SOD and GPX facilitates the oxidative aggression to the cells, especially in subjects with CAD^[29]. The study showed that in the early stages of CAD, SOD and CAT levels increased to protect and prevent lipid peroxidation whereas they decreased significantly with the worsening of the disease^[29]. These observations indicate that in the early stages of the disease there is a homeostatic up-regulation involving the antioxidant

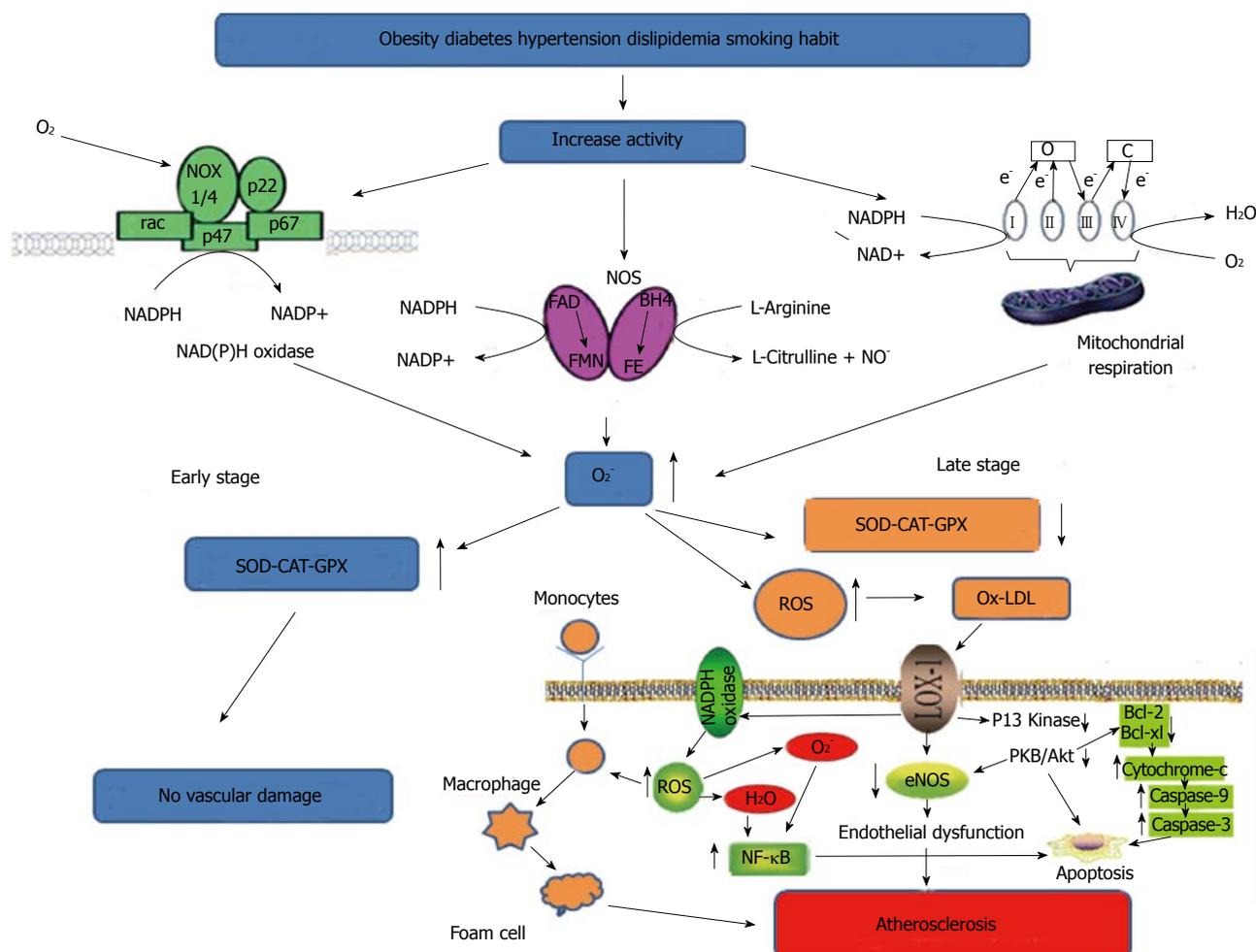


Figure 1 Biochemical events that favor the increase of reactive oxygen species. In the early stages of CAD, ROS do not cause damage due to the presence of an enzymatic compensatory mechanism. In late stage this mechanism is saturated and no longer allows an efficient defense, so that other biochemical events lead to vascular damage. ROS: Reactive oxygen species; SOD: Superoxide dismutase; CAT: Catalase; GPX: Glutathione peroxidase; CAD: Coronary artery disease; NF-κB: Nuclear factor-κB; NADPH: Nicotinamide adenine dinucleotide phosphate-oxidase; LOX-1: Lectin-like oxidized low-density lipoprotein receptor-1; eNOS: Endothelial nitric oxide synthase; FAD: Flavin adenine dinucleotide; BH4: Tetrahydrobiopterin; FE: Heme iron; FMN: Flavin mononucleotide.

enzyme system in response to increased free radicals to prevent vascular damage. In later stages of the disease, when free radicals get to chronically elevated levels, this mechanism, that has reached the saturation, suddenly crashes showing a reduction of antioxidant enzyme activities. In our study^[30] in which we examined a population of CAD patients with different numbers of affected vessels, we did not find a significant difference in SOD levels even if higher values were observed in patients with three or four numbers of injured vessels (Table 1). We can speculate that in presence of high oxygen free radical levels, the compensatory response is only partially related to the activity or expression of the EC-SOD enzyme linked to the damaged artery. However, our data do not disagree with the data of Gupta *et al.*^[29] concerning the patients in the late stages of the disease, when the vascular damage is already present; in fact they did not find any difference in SOD and CAT activity. The difference is mainly in the fact that the authors have also examined patients in the early stages of CAD, when the antioxidant enzyme system was

not yet saturated. Therefore, EC-SOD do not appear a useful marker for the diagnosis of CAD or to stratify the patients population^[30] (Table 1), but it may represent a good therapeutic target against ROS in CAD. In another study, Kotur-Stevuljevic *et al.*^[31] found that the enzymatic activity of erythrocyte SOD in patients with CAD was significantly decreased comparing to healthy volunteers. A significant difference was also evident among patients with stenosis less than 50% compared to those with stenosis higher than 50%. However, the activity of SOD in patients with stenosis less than 50% was not significantly different from the control group, showing that also a compensatory mechanism did not exist in this population. This means that the homeostatic enzyme response occurs when the vascular damage has not yet been produced. This phase could represent an important time point for therapeutic treatments that stimulate the enzyme system for a longer time or with natural scavenger to massively reduce the presence of ROS produced by metabolic diseases or from unhealthy lifestyle. From the clinical point of view

Table 1 Inflammatory parameters, and serum levels of extracellular superoxide dismutase and free radicals in a group of patients with coronary artery disease and healthy volunteers drawn from the study of Lubrano *et al*^[30]

Number of injured vessels	IL-6 (pg/mL)	TNF- α (pg/mL)	CRP (mg/dL)	Peroxy radicals (UC)	EC-SOD (U/mL)
Controls	1.05 \pm 0.2	2.5 \pm 1.1	0.15 \pm 0.03	197 \pm 15.5	2.91 \pm 0.4
1	2.7 \pm 0.7	1.05 \pm 0.3	0.83 \pm 0.7	241 \pm 30.7	2.9 \pm 0.4
2	2.6 \pm 1.7	1.34 \pm 0.2	1.6 \pm 0.1	246 \pm 12.5	2.7 \pm 0.6
3	3.3 \pm 1 ^a	0.98 \pm 0.4	1.1 \pm 0.7	272 \pm 20.2	3.8 \pm 0.7
4	3.5 \pm 0.6 ^a	3.9 \pm 1.5	1.4 \pm 1.1	273 \pm 30.5	5.1 \pm 1.3

^aP < 0.05. TNF- α : Tumor necrosis factor- α ; EC-SOD: Extracellular superoxide dismutase; CAD: Coronary artery disease; CRP: C-reactive protein.

it would be important to combine the analysis of SOD, CAT, GPX with markers of vascular inflammation such as cytokines and C-reactive protein in cases where it is already established the presence of risk factors such as hypertension, diabetes or hypercholesterolemia, but in the absence of plaque formation. The intimal thickening, effect induced by endothelial activation, is an early event to consider. At this stage the cells begin to produce inflammatory cytokines that attract monocytes, adhesion molecules, receptors for oxidized lipoproteins and a massive increase of ROS. Of course, more studies are needed to determine the precise role of these enzymes in protecting the arteries from ROS damage, in order to clarify whether they can be inserted in both the prevention and the treatment of atherosclerotic disease.

INFLUENCE OF THE MAIN RISK FACTORS IN THE MODULATION OF ANTIOXIDANT ENZYME

Obesity has been considered an important factor in causing various health problems, especially in vascular disease^[32]. It has been observed that the adipose tissue secretes adipokines, responsible for the production of ROS, and independent factors, for the generation of systemic oxidative stress^[33]. The persistence of obesity implies an increase of inflammatory cytokines and an excessive consumption of oxygen, which generates free radicals in the respiratory chain coupled to oxidative phosphorylation in mitochondria. In the long term, the accumulation of fat may deplete the sources of antioxidants and significantly decrease the activity of enzymes such as SOD, CAT and GPX and the presence of non-enzymatic factors such as vitamin E, vitamin C and β -carotene^[33].

Even in the case of diabetes there is convincing experimental evidence and clinical trials that have demonstrated that the onset of the disease is closely associated with oxidative stress^[34,35]. Potential sources of ROS in diabetes can be justified by the increase in glucose oxidation and by the changes of redox balance through a cascade of ROS generated by mitochondria. This process has been associated with the onset of type 1 diabetes (DM1) caused by pancreatic beta cell apoptosis, and the onset of type 2 diabetes (DM2) caused by insulin resistance^[36]. Some authors

demonstrated that high glucose levels could stimulate cytochrome P450 activity by excessive nicotinamide adenine dinucleotide phosphate-oxidase (NADPH) produced by glucose metabolism^[37]. In addition, ketosis, a hallmark of DM1, seems to increase the production of oxygen radicals in this patients^[38]. The reduced enzymatic activity of CAT, SOD, GSH-PX, and glutathione reductase (GSH-Rx), as well as high levels of thiobarbituric acid (TBARS), an indirect measure of the production of ROS, that seem to be consistently high in diabetes^[39] are important indices for interpreting the extent of the disease. Recent studies have shown that the levels of SOD and glutathione S-transferase activities were significantly lower in patients with T2DM compared to healthy subjects^[40]. It is known that the use of vitamin E as a dietary supplement for patients with CAD entails a significant benefit in reducing the symptoms of angina pectoris^[41]. In diabetic rats the beneficial effect of vitamin E showed the delay of onset of coronary atherosclerosis compared to untreated. The slowing of the development of the disease was due to a reduction in oxidative stress, and not secondary to a decrease in the glucose or cholesterol in plasma, for the fact that the respective plasma concentrations remained unchanged in the diabetic mice supplemented with vitamin E^[42]. Other studies have supported these results, in fact, it was observed that a triple antioxidant therapy (Vitamin E, lipoic acid, and vitamin C) in diabetic volunteers attenuated oxidative stress reducing the formation of methemoglobin *in vitro* and in glycated hemoglobin *in vivo*^[43]. Numerous clinical studies have shown a decrease in EC-SOD in African Americans with hypertension, in patients with vasospastic angina, calcific aortic stenosis and in patients with DM2, compared with control subjects^[44,45]. Furthermore, it was observed that the standard dietary treatment for type 2 diabetic patients produces an increase of the SOD and GPX activity^[46].

It is evident that the mechanism, that renders low-density lipoprotein a good substrate for the production of foam cells and the initiation of atherosclerotic events, is their oxidative modification^[28]. As for other conditions, some investigators have shown that the overexpression of antioxidant enzymes can slow the progression of atherosclerosis^[47]. Tests carried out on animals have found that the use of natural antioxidants supplements leads to the increase of enzyme activities^[48], whereas

the intake of excessive dietary lipids and therefore an excess of energy and cholesterol has a negative influence on antioxidant enzymes. A negative correlation between dietary cholesterol and the markers of antioxidant enzyme activity, including CAT and GPX, was observed. The authors have shown a significant improvement in erythrocyte antioxidant capacity, as increased activity for SOD, CAT and GPX in children with hypercholesterolemia who have followed a diet with reduced saturated fat and introduction of several fatty acids for 6 mo^[49].

CLINICAL IMPLICATIONS

Increased expression of CuZn-SOD (SOD1) protects muscle cells from oxidative damage. It was observed that overexpression of *SOD1* gene inhibits the DNA binding activity of activator protein-1 and NF- κ B. Interesting prospects are given by the fact that the substitution of valine with alanine has been shown to induce an increase of 30%-40% in the activity Mn-SOD in the mitochondria with consequent reduction of the risk of CAD and acute myocardial infarction^[50]. Even the overexpression of GPX reduces oxidation of the phospholipids, the formation of hydroperoxides of cholesterol, as well as pro-inflammatory lipid peroxides generated by LPO and COX, reducing oxidative stress and vascular atherosclerosis progression. From these observations we conclude that the antioxidant enzyme system is inversely associated with a high-fat diet, and as previously described, the increase in vitamin E, vitamin C, and β -carotene is associated with the strengthening of SOD, therefore the feeding is an important factor in the prevention and treatment of oxidative damage caused by ROS. In the near future it will be possible to study also the genetic polymorphism. The existence of a gene that promotes the enzymatic activity of SOD can contribute to create new drugs for the prevention of damage caused by metabolic diseases or lifestyle that increases the plasma levels of ROS. We believe that further studies should be performed to determine if there is a mechanism of compensation of the antioxidant enzyme system induced by the presence of ROS, and in this case to understand when it begins and what is its intensity. This fact is not very clear from previous studies, because if on one hand it seems to develop before vascular lesion, on the other hand it has never been observed in the presence of metabolic diseases, when the vascular damage has not yet happened.

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REFERENCES

1 **Falkowski PG.** Evolution. Tracing oxygen's imprint on earth's

metabolic evolution. *Science* 2006; **311**: 1724-1725 [PMID: 16556831 DOI: 10.1126/science.1125937]

2 **Farrugia G, Balzan R.** Oxidative stress and programmed cell death in yeast. *Front Oncol* 2012; **2**: 64 [PMID: 22737670 DOI: 10.3389/fonc.2012.00064]

3 **Raymond J, Segrè D.** The effect of oxygen on biochemical networks and the evolution of complex life. *Science* 2006; **311**: 1764-1767 [PMID: 16556842 DOI: 10.1126/science.1118439]

4 **Turrens JF, Boveris A.** Generation of superoxide anion by the NADH dehydrogenase of bovine heart mitochondria. *Biochem J* 1980; **191**: 421-427 [PMID: 6263247 DOI: 10.1042/bj1910421]

5 **Lee YJ, Galoforo SS, Berns CM, Chen JC, Davis BH, Sim JE, Corry PM, Spitz DR.** Glucose deprivation-induced cytotoxicity and alterations in mitogen-activated protein kinase activation are mediated by oxidative stress in multidrug-resistant human breast carcinoma cells. *J Biol Chem* 1998; **273**: 5294-5299 [PMID: 9478987 DOI: 10.1074/jbc.273.9.5294]

6 **Sun J, Chen Y, Li M, Ge Z.** Role of antioxidant enzymes on ionizing radiation resistance. *Free Radic Biol Med* 1998; **24**: 586-593 [PMID: 9559871 DOI: 10.1016/S0891-5849(97)00291-8]

7 **Lledias F, Rangel P, Hansberg W.** Oxidation of catalase by singlet oxygen. *J Biol Chem* 1998; **273**: 10630-10637 [PMID: 9553125 DOI: 10.1074/jbc.273.17.10630]

8 **Bradley JR, Johnson DR, Pober JS.** Four different classes of inhibitors of receptor-mediated endocytosis decrease tumor necrosis factor-induced gene expression in human endothelial cells. *J Immunol* 1993; **150**: 5544-5555 [PMID: 8390537]

9 **Sellak H, Franzini E, Hakim J, Pasquier C.** Reactive oxygen species rapidly increase endothelial ICAM-1 ability to bind neutrophils without detectable upregulation. *Blood* 1994; **83**: 2669-2677 [PMID: 7513210]

10 **Mataix J, Quiles JL, Huertas JR, Battino M, Mañas M.** Tissue specific interactions of exercise, dietary fatty acids, and vitamin E in lipid peroxidation. *Free Radic Biol Med* 1998; **24**: 511-521 [PMID: 9580480 DOI: 10.1016/S0891-5849(97)00288-8]

11 **Tolmasoff JM, Ono T, Cutler RG.** Superoxide dismutase: correlation with life-span and specific metabolic rate in primate species. *Proc Natl Acad Sci USA* 1980; **77**: 2777-2781 [PMID: 6771758 DOI: 10.1073/pnas.77.5.2777]

12 **Lopez-Torres M, Perez-Campo R, Rojas C, Cadenas S, Barja G.** Maximum life span in vertebrates: relationship with liver antioxidant enzymes, glutathione system, ascorbate, urate, sensitivity to peroxidation, true malondialdehyde, in vivo H₂O₂, and basal and maximum aerobic capacity. *Mech Ageing Dev* 1993; **70**: 177-199 [PMID: 8246633 DOI: 10.1016/0047-6374(93)90047-U]

13 **Narayanan D, Xi Q, Pfeffer LM, Jaggar JH.** Mitochondria control functional CaV1.2 expression in smooth muscle cells of cerebral arteries. *Circ Res* 2010; **107**: 631-641 [PMID: 20616314 DOI: 10.1161/CIRCRESAHA.110.224345]

14 **Rajagopalan S, Kurz S, Münzel T, Tarpey M, Freeman BA, Griending KK, Harrison DG.** Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation. Contribution to alterations of vasomotor tone. *J Clin Invest* 1996; **97**: 1916-1923 [PMID: 8621776 DOI: 10.1172/JCI118623]

15 **Pagano PJ, Ito Y, Tornheim K, Gallop PM, Tauber AI, Cohen RA.** An NADPH oxidase superoxide-generating system in the rabbit aorta. *Am J Physiol* 1995; **268**: H2274-H2280 [PMID: 7611477]

16 **Mohazzab-H KM, Kaminski PM, Wolin MS.** Lactate and PO₂ modulate superoxide anion production in bovine cardiac myocytes: potential role of NADH oxidase. *Circulation* 1997; **96**: 614-620 [PMID: 9244234 DOI: 10.1161/01.CIR.96.2.614]

17 **Griending KK, Minieri CA, Ollerenshaw JD, Alexander RW.** Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res* 1994; **74**: 1141-1148 [PMID: 8187280 DOI: 10.1161/01.RES.74.6.1141]

18 **De Keulenaer GW, Alexander RW, Ushio-Fukai M, Ishizaka N, Griending KK.** Tumour necrosis factor alpha activates a p22phox-based NADH oxidase in vascular smooth muscle. *Biochem J* 1998; **329** (Pt 3): 653-657 [PMID: 9445395 DOI: 10.1042/bj3290653]

- 19 **Marumo T**, Schini-Kerth VB, Fisslthaler B, Busse R. Platelet-derived growth factor-stimulated superoxide anion production modulates activation of transcription factor NF-kappaB and expression of monocyte chemoattractant protein 1 in human aortic smooth muscle cells. *Circulation* 1997; **96**: 2361-2367 [PMID: 9337212 DOI: 10.1161/01.CIR.96.7.2361]
- 20 **Patterson C**, Ruef J, Madamanchi NR, Barry-Lane P, Hu Z, Horaist C, Ballinger CA, Brasier AR, Bode C, Runge MS. Stimulation of a vascular smooth muscle cell NAD(P)H oxidase by thrombin. Evidence that p47(phox) may participate in forming this oxidase in vitro and in vivo. *J Biol Chem* 1999; **274**: 19814-19822 [PMID: 10391925 DOI: 10.1074/jbc.274.28.19814]
- 21 **Majima HJ**, Oberley TD, Furukawa K, Mattson MP, Yen HC, Szweda LI, St Clair DK. Prevention of mitochondrial injury by manganese superoxide dismutase reveals a primary mechanism for alkaline-induced cell death. *J Biol Chem* 1998; **273**: 8217-8224 [PMID: 9525927 DOI: 10.1074/jbc.273.14.8217]
- 22 **Guan Y**, Hickey MJ, Borgstahl GE, Hallewell RA, Lepock JR, O'Connor D, Hsieh Y, Nick HS, Silverman DN, Tainer JA. Crystal structure of Y34F mutant human mitochondrial manganese superoxide dismutase and the functional role of tyrosine 34. *Biochemistry* 1998; **37**: 4722-4730 [PMID: 9537987 DOI: 10.1021/bi972394I]
- 23 **Reaume AG**, Elliott JL, Hoffman EK, Kowall NW, Ferrante RJ, Siwek DF, Wilcox HM, Flood DG, Beal MF, Brown RH, Scott RW, Snider WD. Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nat Genet* 1996; **13**: 43-47 [PMID: 8673102 DOI: 10.1038/ng0596-43]
- 24 **Buschfort C**, Muller MR, Seeber S, Rajewsky MF, Thomale J. DNA excision repair profiles of normal and leukemic human lymphocytes: functional analysis at the single-cell level. *Cancer Res* 1997; **57**: 651-658 [PMID: 9044842]
- 25 **Hunt CR**, Sim JE, Sullivan SJ, Featherstone T, Golden W, Von Kapp-Herr C, Hock RA, Gomez RA, Parsian AJ, Spitz DR. Genomic instability and catalase gene amplification induced by chronic exposure to oxidative stress. *Cancer Res* 1998; **58**: 3986-3992 [PMID: 9731512]
- 26 **Taylor SD**, Davenport LD, Speranza MJ, Mullenbach GT, Lynch RE. Glutathione peroxidase protects cultured mammalian cells from the toxicity of adriamycin and paraquat. *Arch Biochem Biophys* 1993; **305**: 600-605 [PMID: 8373199 DOI: 10.1006/abbi.1993.1467]
- 27 **Antoniades C**, Tousoulis D, Tentolouris C, Toutouzias P, Stefanadis C. Oxidative stress, antioxidant vitamins, and atherosclerosis. From basic research to clinical practice. *Herz* 2003; **28**: 628-638 [PMID: 14689123 DOI: 10.1007/s00059-003-2417-8]
- 28 **Stocker R**, Keaney JF. Role of oxidative modifications in atherosclerosis. *Physiol Rev* 2004; **84**: 1381-1478 [PMID: 15383655 DOI: 10.1152/physrev.00047.2003]
- 29 **Gupta S**, Sodhi S, Mahajan V. Correlation of antioxidants with lipid peroxidation and lipid profile in patients suffering from coronary artery disease. *Expert Opin Ther Targets* 2009; **13**: 889-894 [PMID: 19606928]
- 30 **Lubrano V**, Di Cecco P, Zucchelli GC. Role of superoxide dismutase in vascular inflammation and in coronary artery disease. *Clin Exp Med* 2006; **6**: 84-88 [PMID: 16820996 DOI: 10.1007/s10238-006-0100-0]
- 31 **Kotur-Stevuljevic J**, Memon L, Stefanovic A, Spasic S, Spasojevic-Kalimanovska V, Bogavac-Stanojevic N, Kalimanovska-Ostric D, Jelić-Ivanovic Z, Zunic G. Correlation of oxidative stress parameters and inflammatory markers in coronary artery disease patients. *Clin Biochem* 2007; **40**: 181-187 [PMID: 17070511 DOI: 10.1016/j.clinbiochem.2006.09.007]
- 32 **Lastra G**, Manrique CM, Hayden MR. The role of beta-cell dysfunction in the cardiometabolic syndrome. *J Cardiometab Syndr* 2006; **1**: 41-46 [PMID: 17675900 DOI: 10.1111/j.0197-3118.2006.05458.x]
- 33 **Fernández-Sánchez A**, Madrigal-Santillán E, Bautista M, Esquivel-Soto J, Morales-González A, Esquivel-Chirino C, Durante-Montiel I, Sánchez-Rivera G, Valadez-Vega C, Morales-González JA. Inflammation, oxidative stress, and obesity. *Int J Mol Sci* 2011; **12**: 3117-3132 [PMID: 21686173 DOI: 10.3390/ijms12053117]
- 34 **Rösen P**, Nawroth PP, King G, Möller W, Tritschler HJ, Packer L. The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes Metab Res Rev* 2001; **17**: 189-212 [PMID: 11424232 DOI: 10.1002/dmrr.196]
- 35 **Sayed MR**, Iman MM, Dawlat AS. Biochemical changes in experimental diabetes before and after treatment with mangifera indica and psidium guava extracts. *Int J Pharm Biomed Sci* 2011; **2**: 29-41
- 36 **West IC**. Radicals and oxidative stress in diabetes. *Diabet Med* 2000; **17**: 171-180 [PMID: 10784220 DOI: 10.1046/j.1464-5491.2000.00259.x]
- 37 **Jain SK**. Hyperglycemia can cause membrane lipid peroxidation and osmotic fragility in human red blood cells. *J Biol Chem* 1989; **264**: 21340-21345 [PMID: 2592379]
- 38 **Jain SK**, Kannan K, Lim G. Ketosis (acetoacetate) can generate oxygen radicals and cause increased lipid peroxidation and growth inhibition in human endothelial cells. *Free Radic Biol Med* 1998; **25**: 1083-1088 [PMID: 9870562 DOI: 10.1016/S0891-5849(98)00140-3]
- 39 **Johansen JS**, Harris AK, Rychly DJ, Ergul A. Oxidative stress and the use of antioxidants in diabetes: linking basic science to clinical practice. *Cardiovasc Diabetol* 2005; **4**: 5 [PMID: 15862133 DOI: 10.1186/1475-2840-4-5]
- 40 **Verma S**, Sagar N, Vats P, Shukla KN, Abbas M, Banerjee M. Antioxidant enzyme levels as markers for type 2 diabetes mellitus. *Int J Bioassays* 2013; **2**: 685-690
- 41 **Shute EV**. Proposed study of vitamin E therapy. *Can Med Assoc J* 1972; **106**: 1057 [PMID: 5032131]
- 42 **Otero P**, Bonet B, Herrera E, Rabano A. Development of atherosclerosis in the diabetic BALB/c mice. Prevention with Vitamin E administration. *Atherosclerosis* 2005; **182**: 259-265 [PMID: 16159598 DOI: 10.1016/j.atherosclerosis.2005.02.024]
- 43 **Coleman MD**, Fernandes S, Khanderia L. A preliminary evaluation of a novel method to monitor a triple antioxidant combination (vitamins E, C and α -lipoic acid) in diabetic volunteers using in vitro methaemoglobin formation. *Environ Toxicol Pharmacol* 2003; **14**: 69-75 [PMID: 21782664 DOI: 10.1016/S1382-6689(03)00027-9]
- 44 **Yamashita K**, Takahiro K, Kamezaki F, Adachi T, Tasaki H. Decreased plasma extracellular superoxide dismutase level in patients with vasospastic angina. *Atherosclerosis* 2007; **191**: 147-152 [PMID: 16584734 DOI: 10.1016/j.atherosclerosis.2006.03.008]
- 45 **Liao M**, Liu Z, Bao J, Zhao Z, Hu J, Feng X, Feng R, Lu Q, Mei Z, Liu Y, Wu Q, Jing Z. A proteomic study of the aortic media in human thoracic aortic dissection: implication for oxidative stress. *J Thorac Cardiovasc Surg* 2008; **136**: 65-72, 72.e1-3 [PMID: 18603055 DOI: 10.1016/j.jtcvs.2007.11.017]
- 46 **Sekeroğlu MR**, Sahin H, Dülger H, Algün E. The effect of dietary treatment on erythrocyte lipid peroxidation, superoxide dismutase, glutathione peroxidase, and serum lipid peroxidation in patients with type 2 diabetes mellitus. *Clin Biochem* 2000; **33**: 669-674 [PMID: 11166015 DOI: 10.1016/S0009-9120(00)00190-9]
- 47 **Blankenberg S**, Rupprecht HJ, Bickel C, Torzewski M, Hafner G, Tiret L, Smieja M, Cambien F, Meyer J, Lackner KJ. Glutathione peroxidase 1 activity and cardiovascular events in patients with coronary artery disease. *N Engl J Med* 2003; **349**: 1605-1613 [PMID: 14573732 DOI: 10.1056/NEJMoa030535]
- 48 **Shih CK**, Chang JH, Yang SH, Chou TW, Cheng HH. beta-Carotene and canthaxanthin alter the pro-oxidation and antioxidation balance in rats fed a high-cholesterol and high-fat diet. *Br J Nutr* 2008; **99**: 59-66 [PMID: 17640418 DOI: 10.1017/S0007114507781497]
- 49 **Codoñer-Franch P**, Bataller Alberola A, Domingo Camarasa JV, Escribano Moya MC, Valls Bellés V. Influence of dietary lipids on the erythrocyte antioxidant status of hypercholesterolaemic children. *Eur J Pediatr* 2009; **168**: 321-327 [PMID: 18548274 DOI: 10.1007/s00431-008-0762-6]

50 **Fujimoto H**, Taguchi J, Imai Y, Ayabe S, Hashimoto H, Kobayashi H, Ogasawara K, Aizawa T, Yamakado M, Nagai R, Ohno M. Manganese superoxide dismutase polymorphism affects the

oxidized low-density lipoprotein-induced apoptosis of macrophages and coronary artery disease. *Eur Heart J* 2008; **29**: 1267-1274 [PMID: 17967822 DOI: 10.1093/eurheartj/ehm500]

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Disease control by regulation of P-glycoprotein on lymphocytes in patients with rheumatoid arthritis

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Abstract

The main purpose of treatment of rheumatoid arthritis (RA) with disease modifying antirheumatic drugs (DMARDs) is to control activation of lymphocytes, although some patients do not respond adequately to such treatment. Among various mechanisms of multidrug resistance, P-glycoprotein (P-gp), a member of ATP-binding cassette transporters, causes drug-resistance by efflux of intracellular drugs. Certain stimuli, such as tumor necrosis factor- α , activate lymphocytes and induce P-gp expression on lymphocytes, as evident in active RA. Studies from our laboratories showed spontaneous nuclear accumulation of human Y-box-binding protein-1, a multidrug resistance 1 transcription factor, in unstimulated lymphocytes, and surface overexpression of P-gp on peripheral lymphocytes of RA patients with high disease activity. The significant correlation between P-gp expression level and RA disease activity is associated with active efflux of drugs from the lymphocyte cytoplasm and in drug-resistance. However, the use of biological agents that reduce P-gp expression as well as P-gp antagonists (*e.g.*, cyclosporine) can successfully reduce the efflux of corticosteroids from lymphocytes *in vitro*, suggesting that both types of drugs can be used to overcome drug-resistance and improve clinical outcome. We conclude that lymphocytes activated by various stimuli in RA patients with highly active disease acquire P-gp-mediated multidrug resistance against corticosteroids and probably some DMARDs, which are substrates of P-gp. Inhibition/reduction of P-gp could overcome such drug resistance. Expression of P-gp on lymphocytes is a promising marker of drug resistance and a suitable therapeutic target to prevent drug resistance in patients with active RA.

Key words: *Multidrug resistance 1* gene; P-glycoprotein; Lymphocytes; Disease activity; Rheumatoid arthritis

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Core tip: In patients with refractory rheumatoid arthritis (RA) and high disease activity, overexpression of P-glycoprotein (P-gp) on lymphocytes can cause resistance to anti-rheumatic drugs through efflux of intracellular drugs from these cells. Lymphocytes activated by various stimuli, including tumor necrosis factor- α in RA patients apparently acquire P-gp-mediated multidrug resistance against certain anti-rheumatic drugs, which are substrates of P-gp. The use of biological agents that reduce P-gp expression as well as P-gp antagonists can successfully reduce the efflux of drugs from lymphocytes, suggesting that they can be used to overcome drug-resistance and improve clinical outcome.

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INTRODUCTION

Rheumatoid arthritis (RA) is manifested by inflammatory and chronic destruction of multiple joints with occasional systemic organ complications based on immune abnormality^[1]. Poor control of RA is associated with severe painful disability and impairments at work and life. The strategic treatment to control immune-mediated synovial inflammation, joint destruction and extra-organ manifestation is by early intervention with synthetic or biological disease modifying anti-rheumatic drugs (DMARDs). Early treatment of RA with DMARDs can result in prevention of joint destruction and a better long-term outcome^[2]. DMARDs commonly target lymphocytes and the cytokines produced by these cells, which play an important role in the pathogenesis of RA^[3]. However, we often encounter RA patients who are refractory to these DMARDs and fail in the control of high disease activity^[4]. Thus, overcoming activated lymphocytes involved in drug-resistance is an important goal of the treatment in some refractory RA patients. P-glycoprotein (P-gp) is a member of ATP-binding cassette transporters and is induced on the cell membrane by certain stimuli. P-gp transports multiple drugs from the cytoplasm to the cell exterior, resulting in the development of drug resistance. Here, we discuss the importance of P-gp on activated lymphocytes and its relevance to multidrug-resistance and the potential for treatments targeting P-gp on lymphocytes to overcome drug-resistance in refractory patients with RA.

MECHANISMS OF DRUG RESISTANCE MEDIATED BY P-GP

P-gp is encoded by the multidrug resistance-1

Table 1 Relation of P-glycoprotein with disease modifying antirheumatic drugs and immunosuppressants

Drug	Pharmacological substrates of P-glycoprotein	Competitive inhibitor of P-glycoprotein
Corticosteroids	Yes	No
Cyclosporine	Yes	Yes
Tacrolimus	Yes	Yes
Methotrexate	No	No
Leflunomide	No	No
Hydroxychloroquine	Yes	Yes
Sulfasalazine	Yes	Unknown
D-penicillamine	Yes	Unknown
Colchicine	Yes	No
Cyclophosphamide	No	No
Azathioprine	No	No

(MDR-1)^[5-7], a member of the ATP-binding cassette transporter superfamily of genes. P-gp is recognized by structurally diverse, hydrophobic/amphiphilic substrates, ranging from 300 to 2000 Da, catches these substrates like a "vacuum cleaner" during passing through the cell membrane, and pumps them out of the cells in a manner dependent on the energy of ATP hydrolysis. Therefore, Corticosteroids, certain immunosuppressants and DMARDs, including antimalarial drugs, are extruded from lymphocytes with overexpression of P-gp, which leads to reductions in the concentrations of these drugs in cytoplasm and failure of their intracellular effects (Table 1)^[8-13]. Indeed, P-gp-mediated efflux of corticosteroids from lymphocytes can result in low cytoplasmic corticosteroid concentrations and development of corticosteroid resistance in systemic lupus erythematosus^[14]. Thus, excessive excretion of the drugs from P-gp-overexpressing lymphocytes can be involved in the drug-resistance often observed in patients with RA.

THE REGULATION OF P-GP EXPRESSION ON LYMPHOCYTES

Endothelial cells of the blood brain barrier and various epithelial cells show congenial expression of P-gp for protection of cells from toxic substances. In contrast, P-gp expression on normal resting lymphocytes is marginal but can be induced by certain stimuli^[15-17]. Overexpression of P-gp appears to be closely associated with the nuclear localization of human Y-box-binding protein-1 (YB-1) in various tumors^[18]. YB-1, which is a member of the DNA/RNA-binding protein family containing a cold-shock domain, is activated in response to various genotoxic stimuli and drives the transcription of *MDR-1* gene^[18]. We have demonstrated that lymphocytes can be activated by various stimuli, such as cytokines and extracellular matrix to induce P-gp expression on lymphocytes, based on the following sequence of events; activation and translocation of YB-1 by IL-2, tumor necrosis factor- α (TNF- α) (Figure 1A) and fragmented hyaluronan, transcriptional activation

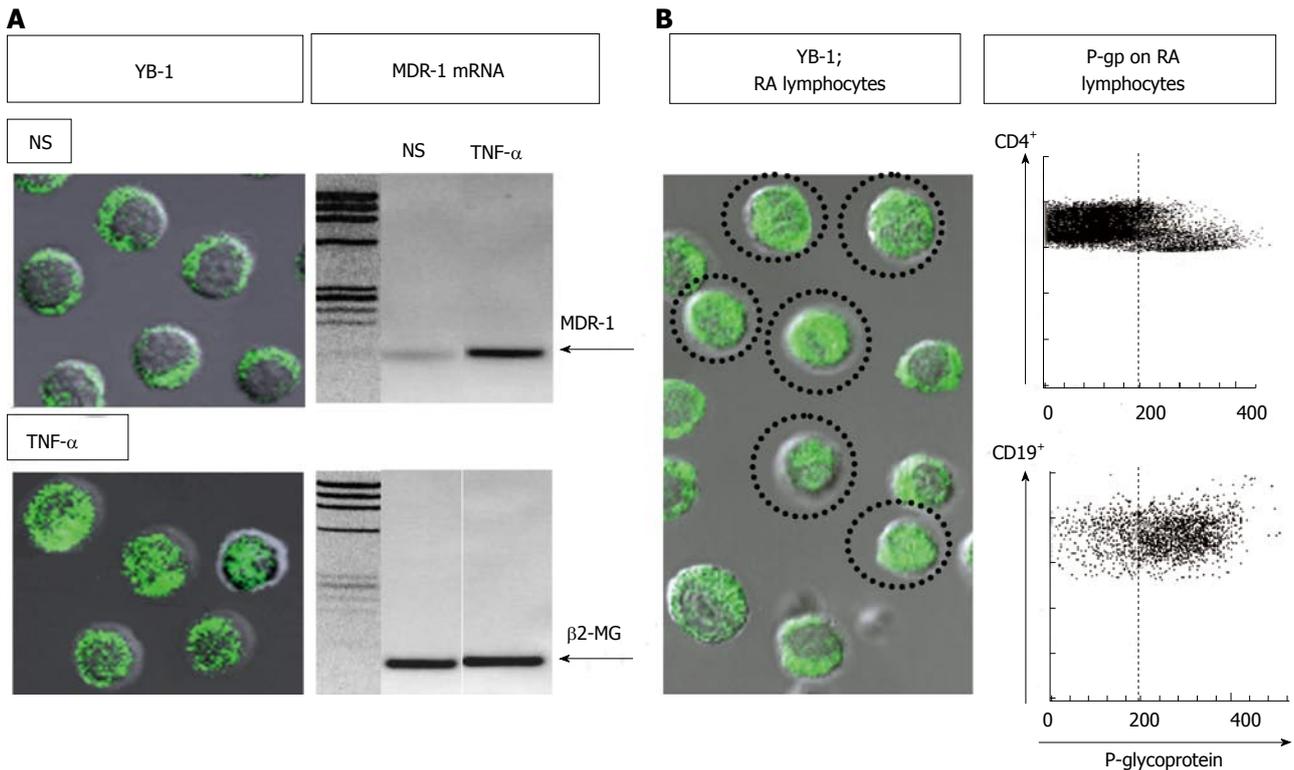


Figure 1 Up-regulation of nuclear translocation of Y-box-binding protein-1, transcription of multidrug resistance 1 in lymphocytes, and P-glycoprotein expression on lymphocytes. A: Left: Immunostaining and confocal microscopy analysis of Y-box-binding protein-1 (YB-1) in 1×10^5 of peripheral blood mononuclear cells (PBMCs). YB-1 was expressed in the cytoplasm of all non-stimulated PBMCs (NS). In contrast, nuclear translocation of YB-1 was induced in 30% or more of PBMCs incubated with 10 ng/mL of tumor necrosis factor- α (TNF- α). Immunostaining for YB-1 using a specific antibody (Ab) against YB-1^[19] with FITC-conjugated anti-rabbit IgG Ab (BD Biosciences Pharmingen). Confocal analysis of YB-1 using a LSM 5 Pascal invert Laser Scan Microscope (Carl Zeiss Microscope Systems, Germany). Magnification, $\times 600$; Right: Multidrug resistance-1 (MDR-1) mRNA expression was examined by RT-PCR using total RNA extracted from PBMCs incubated with 10 ng/mL of TNF- α or no stimulation (NS). The primer sequences were as follows: human $\beta 2$ -microglobulin forward 5'-ACCCCTGAAAAGATGA-3', reverse 5'-ATCTTCAAACCTCCATGATG-3'; human MDR-1 forward 5'-CCCATCATTGCAATAGCAGG-3', reverse 5'-GTTCAAAATTCTGCTCCTGA-3'. Amplified products were electrophoresed with Marker 4 (Nippon Gene, Tokyo) on 3% agarose gels; B: Spontaneous nuclear translocation of YB-1 and P-glycoprotein (P-gp) expression on lymphocytes from a typical patient with active rheumatoid arthritis (RA). Left: Immunostaining and confocal microscopy analysis of YB-1 in 1×10^5 of PBMCs. YB-1 was expressed in the nuclei of a proportion of unstimulated PBMCs (encircled cells). Magnification, $\times 600$; Right: P-gp expression on CD4⁺ and CD19⁺ peripheral blood lymphocytes. The dotted line represents the gate set to discriminate negative from positive stained cells as determined by control FITC-conjugated anti-mouse IgG Ab. The specific antibodies for staining and flow cytometric analysis were as follows: staining for P-gp using MRK16 (a specific monoclonal Ab against P-gp; Kyowa Medex, Tokyo) with FITC-conjugated goat anti-mouse IgG Ab (BD Biosciences Pharmingen), cy-chrome-conjugated CD4 monoclonal Ab, cy-chrome-conjugated CD19 monoclonal Ab (BD Biosciences Pharmingen).

of MDR-1 by activated YB-1, P-gp expression on the cell surface membrane of lymphocytes, expelling added dexamethasone from lymphocytes, leading to a fall in intracellular dexamethasone concentration^[16,17]. Serum and synovial concentrations of IL-2 are high in patients with active RA^[19,20]. TNF- α is a clinically validated pathogenic factor in inflammatory erosive arthritis in RA and is pivotal target for directed biologic intervention^[3,21-23]. Fragmented hyaluronan is increased in the RA synovium and synovial fluid^[24,25]. The enhanced production of fragmented hyaluronan is due to increased digestion of native hyaluronan, which is increased in inflammatory foci like synovitis by inflammatory cytokines including IL-1 β and TNF- α ^[26] and by oxygen-derived free radicals^[24,25]. Overexpression of P-gp on lymphocytes induced by these stimuli^[15,16], which also play important roles in the disease activity of RA, parallels the activation of lymphocytes. P-gp expression is preferentially high on CD69-expressing lymphocytes, which is a well-defined marker of early activation of lymphocytes^[16,17]. Actually, lymphocytes

in highly active RA patients, *i.e.*, pathologically active lymphocytes, show accumulation of YB-1 in the nuclei and overexpression of P-gp on the cell surface membrane (Figure 1B). Thus, lymphocyte activation by certain stimuli induces YB-1 activation followed by P-gp overexpression, resulting in acquisition of multidrug resistance mediated by P-gp (Figure 2A and B, and Table 1). Therefore, the presence of active lymphocytes that have acquired P-gp-mediated multidrug resistance are probably involved in the failure of disease control in patients with active RA.

CLINICAL VALIDATION OF THE RELATIONSHIP BETWEEN P-GP-EXPRESSING RA LYMPHOCYTES AND DRUG RESISTANCE

The expression level of P-gp is significantly high on most peripheral CD4⁺ T cells and CD19⁺ B cells in RA

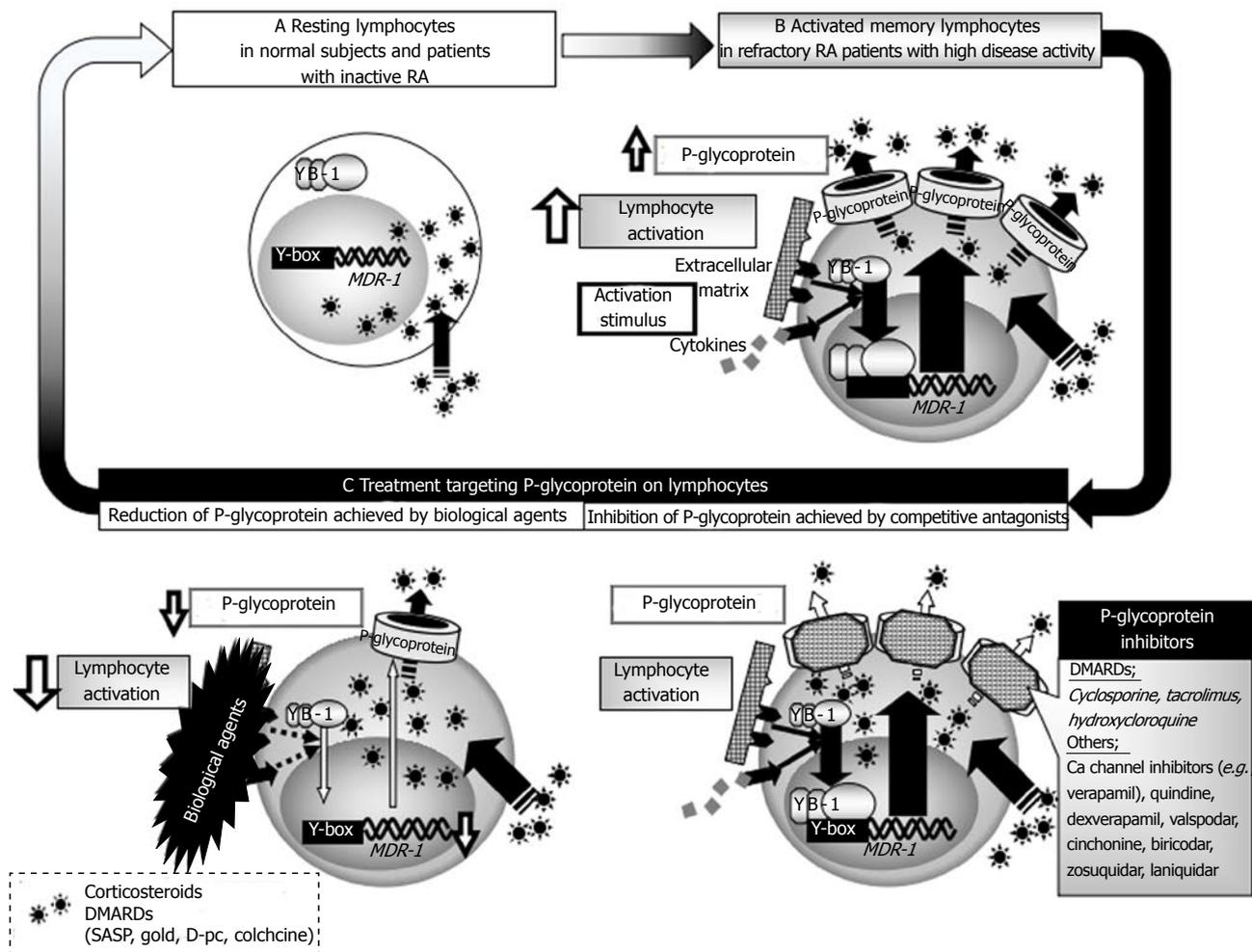


Figure 2 Schematic diagram of the relevance of P-glycoprotein to drug resistance in rheumatoid arthritis. A: Y-box-binding protein-1 is located in the cytoplasm of lymphocytes and P-glycoprotein (P-gp) is only marginally expressed on normal lymphocytes of normal subjects and patients with inactive RA; B: In patients with highly active rheumatoid arthritis (RA), various stimuli induce P-gp expression on lymphocytes, which leads to active efflux of drugs from lymphocytes, resulting in drug-unresponsiveness and failure to control disease activity; C: Reduction of P-gp achieved by intensive immunosuppressive therapy and inhibition of P-gp by competitive antagonists, such as cyclosporine, could overcome P-gp-related drug-resistance in patients with highly active RA. DMARDs: Disease modifying antirheumatic drugs; MDR-1: Multidrug resistance 1.

patients, but marginal in normal subjects. Evidence indicates that P-gp expression level on lymphocytes characteristically correlates significantly with RA disease activity, estimated by the disease activity score (DAS) 28-3^[27], and that its levels on CD4⁺ T cells and CD19⁺ B cells are markedly increased in patients with inadequate response to one DMARD treatment with corticosteroids or to at least two DMARDs for at least 2 years, compared with responders and normal volunteers (Figure 3).

To elucidate the relationship between P-gp expression and P-gp-mediated efflux of intracellular drugs *in vitro*, intracellular and extracellular concentrations of dexamethasone, a representative substrate of P-gp (Table 1), were determined by the C/M ratio, an index of intracellular concentration of dexamethasone (C) and extracellular concentration of dexamethasone in the conditioned medium (M). Using this method, we analyzed *in vitro* peripheral blood lymphocytes from RA patients with highly active disease (DAS 28-3 > 5.1). Significantly low intracellular dexamethasone levels were found in P-gp-overexpressing lymphocytes, compared

with those from RA patients with mild to moderate active disease (DAS 28-3 < 5.1) and normal volunteers. The above findings indicate that overexpression of P-gp on activated lymphocytes leads to active efflux of certain intracellular drugs, substrates of P-gp, from the cells, and results in the development of drug resistance and failure of the disease control in highly active RA (Figure 2B). Taken together, it is possible that treatment that targets P-gp on lymphocytes could overcome drug-resistance in refractory patients with RA. Treatment modalities that target P-gp are classified into those that reduce P-gp expression and others that inhibit P-gp function.

BIOLOGICAL AGENTS CAN OVERCOME P-GP-MEDIATED DRUG RESISTANCE IN PATIENTS WITH REFRACTORY RHEUMATOID ARTHRITIS

TNF- α is a central player in the inflammatory process

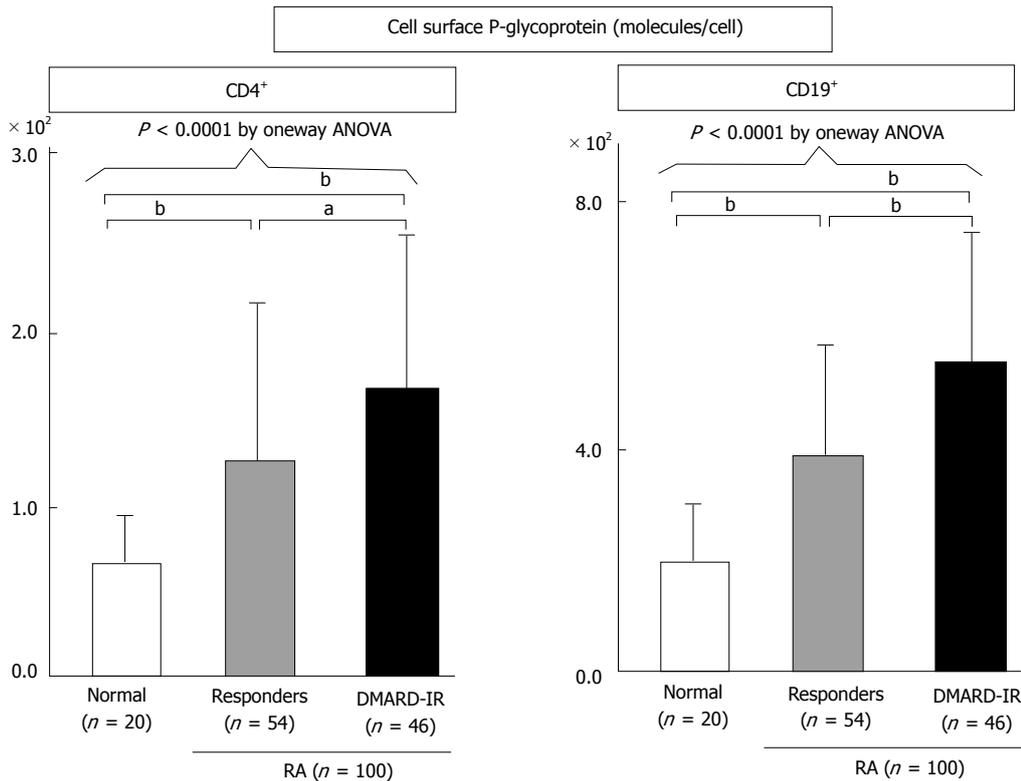


Figure 3 Expression of P-glycoprotein on lymphocytes from patients with refractory rheumatoid arthritis, as determined by flow cytometry. P-glycoprotein (P-gp) expression on CD4⁺ or CD19⁺ peripheral blood lymphocytes from 20 normal volunteers (open bar) and 100 RA patients [responders, hatched bars; with inadequate response to DMARDs (DMARD-IR), closed bars]. The specific antibodies for staining and flow cytometric analysis were as follows: staining for P-gp using MRK16 [a specific monoclonal antibody (Ab) against P-gp; Kyowa Medex, Tokyo] with FITC-conjugated goat anti-mouse IgG Ab (BD Biosciences Pharmingen), cy-chrome-conjugated CD4 monoclonal Ab, cy-chrome-conjugated CD19 monoclonal Ab (BD Biosciences Pharmingen). Data represent the number of molecules expressed per cell, calculated using standard QIFIKIT beads. Values are mean \pm SD of independent experiments. One-way ANOVA were used to compare data between groups. ^a $P < 0.05$, ^b $P < 0.01$, by multiple comparison. DMARD: Disease modifying antirheumatic drug; RA: Rheumatoid arthritis.

of RA, as it exacerbates erosive synovitis and enhances disease activity, and is thus an excellent molecular target for directed biologic intervention^[3,21-23]. Infliximab and etanercept, two highly effective antagonists of TNF- α , have revolutionized treatment strategies for RA. In one study, infliximab improved clinical disease activity in intractable RA through significant reduction of P-gp expression levels on CD4⁺ T cells and CD19⁺ B cells, which were otherwise uncontrolled by treatment with MTX^[27]. In another study published by our group, etanercept significantly reduced CD69 and P-gp expression on CD4⁺ T cells and CD19⁺ B cells in each of 11 patients with intractable RA, including two who experienced secondary loss of infliximab efficacy and eight who did not use MTX^[17]. The above two studies demonstrated that treatment with TNF- α antagonists can improve RA disease activity within two weeks^[17,27], and allow tapering or withdrawal of steroid therapy^[17]. These effects were associated with recovery of intracellular dexamethasone levels in lymphocytes accompanied by falls in P-gp levels in these cells^[17,27]. The results imply that intensive treatment with TNF- α antagonists reduces P-gp expression and results in rescue of DMARD and steroid concentrations in the cytoplasm of lymphocytes.

TNF antagonists act extracellularly and inhibit lymphocyte activation without being affected by

P-gp^[17]. Inhibition of lymphocyte activation by TNF- α antagonists probably suppresses YB-1-driven transcriptional activation of MDR-1 and P-gp expression on lymphocytes, and excretion of multiple drugs from the cytoplasm to the extracellular compartment. Therefore, inhibition of lymphocyte activation by TNF- α antagonists can probably thwart P-gp-mediated treatment resistance in refractory patients with RA (Figure 2C).

Translated clinically, when treatment with DMARDs and corticosteroids fails to control high disease activity in RA patients, administration of DMARDs or immunosuppressants that escape from P-gp-orchestrated excretion (Table 1), is a better treatment option. Furthermore, treatment with biological agents including TNF- α antagonists, should be initiated.

P-GP COMPETITORS REGULATE P-GP-MEDIATED DRUG RESISTANCE IN PATIENTS WITH REFRACTORY RHEUMATOID ARTHRITIS

The immunosuppressants cyclosporine and tacrolimus act as calcineurin inhibitors and inhibit cytokine production promoted by NF-AT, as well as act as P-gp-

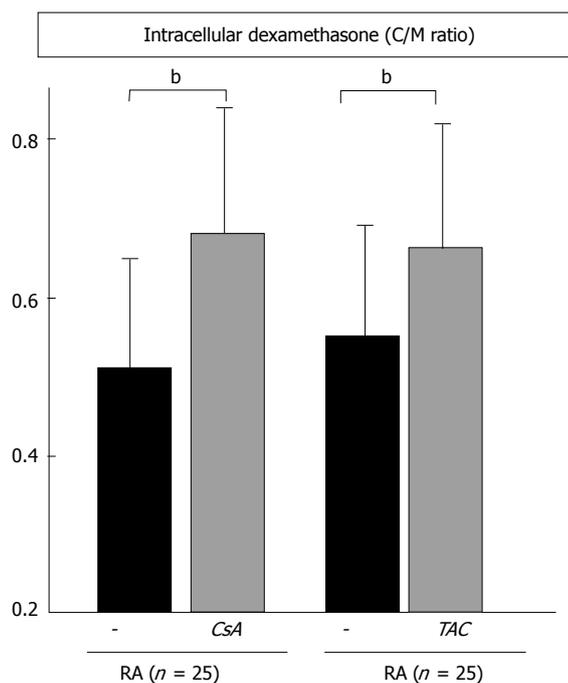


Figure 4 Inhibition of P-glycoprotein-related dexamethasone efflux by cyclosporine and tacrolimus. The C/M ratio in peripheral blood mononuclear cells of 25 rheumatoid arthritis (RA) patients was measured in the absence or presence of 100 ng/mL of cyclosporine (CsA), or 10 ng/mL of tacrolimus (TAC). Data are mean \pm SD. ^b $P < 0.01$, by the paired *t* test.

competitive inhibitors. Recent therapeutic intervention studies have investigated the efficacy of P-gp competitors, such as cyclosporine and its derivatives, in overcoming P-gp-mediated treatment resistance^[28,29]. We have demonstrated that two P-gp competitive immunosuppressants, cyclosporine and tacrolimus, inhibited dexamethasone excretion from IL-2-activated lymphocytes, and that this action was concentration-dependent. In addition, treatment with these competitors resulted in recovery of intracellular dexamethasone concentrations in IL-2-activated lymphocytes by doses lower than trough levels measured clinically when these agents are used as calcineurin inhibitors^[15]. Actually, the addition of low doses of tacrolimus and cyclosporine to lymphocytes harvested from highly active RA patients, resulted in recovery of intra-lymphocytes dexamethasone concentrations (Figure 4).

In our clinical trial, we experienced refractory patients with high disease activity who were treated with low-dose cyclosporine or tacrolimus and showed significant improvement of clinical features within two weeks, which was not accompanied by falls in P-gp expression levels on lymphocytes^[30]. One representative case was a 56-year-old woman with P-gp-overexpressing lymphocytes. Despite treatment with MTX, sulphasalazine, and D-penicillamine (D-pc), the RA disease activity flared several times and the joints destruction rapidly progressed during a period of two years. Low-dose cyclosporine therapy (serum cyclosporine concentration < 100 ng/mL), added to MTX with D-pc, markedly improved RA disease activity and

normalized CRP and MMP-3 within two months. This outcome suggests that cyclosporine and tacrolimus, administered at low doses and over a relatively short period of time than necessary for NFAT inhibition, competitively inhibit elimination of intracellular drugs through P-gp on lymphocytes without simultaneously reducing P-gp expression, and can thus be used to overcome drug resistance and result in improvement of clinical features. We propose that competitors of P-gp, including cyclosporine, tacrolimus and hydroxychloroquine (Table 1), are potentially effective therapies for RA patients with high disease activity who are refractory to treatments with conventional DMARDs (Figure 2C).

CONCLUSION

Pharmacotherapy is the main form of treatment of RA; therefore, drug resistance induced by activated lymphocytes is a potentially serious challenge in the clinical management of RA. P-gp overexpression on activated pathogenic lymphocytes leads to the development of P-gp-mediated multidrug resistance. Therefore, treatments that target P-gp and preferentially reduce P-gp-mediated drug-resistance, can overcome drug-resistance. Taken together, we propose that the level of P-gp expressed on peripheral lymphocytes measured in RA patients is a useful marker for P-gp-mediated drug resistance and might help in the selection of more effective treatment, including treatment that reduce P-gp expression (such as biological agents) and inhibit P-gp function (such as P-gp competitors).

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REFERENCES

- 1 Smolen JS, Aletaha D. The assessment of disease activity in rheumatoid arthritis. *Clin Exp Rheumatol* 2010; **28**: S18-S27 [PMID: 20576221]
- 2 Smolen JS, Landewé R, Breedveld FC, Buch M, Burmester G, Dougados M, Emery P, Gaujoux-Viala C, Gossec L, Nam J, Ramiro S, Winthrop K, de Wit M, Aletaha D, Betteridge N, Bijlsma JW, Boers M, Buttgerit F, Combe B, Cutolo M, Damjanov N, Hazes JM, Kouloumas M, Kvien TK, Mariette X, Pavelka K, van Riel PL, Rubbert-Roth A, Scholte-Voshaar M, Scott DL, Sokka-Isler T, Wong JB, van der Heijde D. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2013 update. *Ann Rheum Dis* 2014; **73**: 492-509 [PMID: 24161836 DOI: 10.1136/annrheumdis-2013-204573]
- 3 Choy EH, Panayi GS. Cytokine pathways and joint inflammation in rheumatoid arthritis. *N Engl J Med* 2001; **344**: 907-916 [PMID: 11259725 DOI: 10.1056/NEJM200103223441207]
- 4 Jorgensen C, Maillefert JF. Multidrug resistances genes in rheumatology. Is their role immunological or pharmacological? *Joint Bone Spine* 2000; **67**: 8-10 [PMID: 10773963]
- 5 Kuwano M, Uchiumi T, Hayakawa H, Ono M, Wada M, Izumi H, Kohno K. The basic and clinical implications of ABC transporters, Y-box-binding protein-1 (YB-1) and angiogenesis-related factors in

- human malignancies. *Cancer Sci* 2003; **94**: 9-14 [PMID: 12708467 DOI: 10.1111/j.1349-7006.2003.tb01344.x]
- 6 **Leonard GD**, Polgar O, Bates SE. ABC transporters and inhibitors: new targets, new agents. *Curr Opin Investig Drugs* 2002; **3**: 1652-1659 [PMID: 12476969]
 - 7 **McKeegan KS**, Borges-Walmsley MI, Walmsley AR. The structure and function of drug pumps: an update. *Trends Microbiol* 2003; **11**: 21-29 [PMID: 12526851 DOI: 10.1016/S0966-842X(02)00010-0]
 - 8 **Jansen G**, Scheper RJ, Dijkmans BA. Multidrug resistance proteins in rheumatoid arthritis, role in disease-modifying antirheumatic drug efficacy and inflammatory processes: an overview. *Scand J Rheumatol* 2003; **32**: 325-336 [PMID: 15080263 DOI: 10.1080/03009740310004333]
 - 9 **Ueda K**. ABC proteins protect the human body and maintain optimal health. *Biosci Biotechnol Biochem* 2011; **75**: 401-409 [PMID: 21389634]
 - 10 **Llorente L**, Richaud-Patin Y, Diaz-Borjón A, Alvarado de la Barrera C, Jakez-Ocampo J, de la Fuente H, Gonzalez-Amaro R, Diaz-Jouanen E. Multidrug resistance-1 (MDR-1) in rheumatic autoimmune disorders. Part I: Increased P-glycoprotein activity in lymphocytes from rheumatoid arthritis patients might influence disease outcome. *Joint Bone Spine* 2000; **67**: 30-39 [PMID: 10773966]
 - 11 **List AF**, Kopecky KJ, Willman CL, Head DR, Slovak ML, Douer D, Dakhil SR, Appelbaum FR. Cyclosporine inhibition of P-glycoprotein in chronic myeloid leukemia blast phase. *Blood* 2002; **100**: 1910-1912 [PMID: 12176916]
 - 12 **Pariante CM**. The role of multi-drug resistance p-glycoprotein in glucocorticoid function: studies in animals and relevance in humans. *Eur J Pharmacol* 2008; **583**: 263-271 [PMID: 18275949 DOI: 10.1016/j.ejphar.2007.11.067]
 - 13 **van der Heijden JW**, Oerlemans R, Tak PP, Assaraf YG, Kraan MC, Scheffer GL, van der Laken CJ, Lems WF, Scheper RJ, Dijkmans BA, Jansen G. Involvement of breast cancer resistance protein expression on rheumatoid arthritis synovial tissue macrophages in resistance to methotrexate and leflunomide. *Arthritis Rheum* 2009; **60**: 669-677 [PMID: 19248091 DOI: 10.1002/art.24354]
 - 14 **Tsujiura S**, Saito K, Nakayamada S, Nakano K, Tanaka Y. Clinical relevance of the expression of P-glycoprotein on peripheral blood lymphocytes to steroid resistance in patients with systemic lupus erythematosus. *Arthritis Rheum* 2005; **52**: 1676-1683 [PMID: 15934077 DOI: 10.1002/art.21032]
 - 15 **Tsujiura S**, Saito K, Nakayamada S, Nakano K, Tsukada J, Kohno K, Tanaka Y. Transcriptional regulation of multidrug resistance-1 gene by interleukin-2 in lymphocytes. *Genes Cells* 2004; **9**: 1265-1273 [PMID: 15569157 DOI: 10.1111/j.1365-2443.2004.00803.x]
 - 16 **Tsujiura S**, Saito K, Kohno K, Tanaka Y. Fragmented hyaluronan induces transcriptional up-regulation of the multidrug resistance-1 gene in CD4+ T cells. *J Biol Chem* 2006; **281**: 38089-38097 [PMID: 17038319 DOI: 10.1074/jbc.M601030200]
 - 17 **Tsujiura S**, Saito K, Nakayamada S, Tanaka Y. Etanercept overcomes P-glycoprotein-induced drug resistance in lymphocytes of patients with intractable rheumatoid arthritis. *Mod Rheumatol* 2010; **20**: 139-146 [PMID: 19915943 DOI: 10.3109/s10165-009-0247-0]
 - 18 **Kuwano M**, Oda Y, Izumi H, Yang SJ, Uchiumi T, Iwamoto Y, Toi M, Fujii T, Yamana H, Kinoshita H, Kamura T, Tsuneyoshi M, Yasumoto K, Kohno K. The role of nuclear Y-box binding protein 1 as a global marker in drug resistance. *Mol Cancer Ther* 2004; **3**: 1485-1492 [PMID: 15542787]
 - 19 **Kuroda T**, Tanabe N, Sakatsume M, Nozawa S, Mitsuka T, Ishikawa H, Tohyama CT, Nakazono K, Murasawa A, Nakano M, Gejyo F. Interleukin-2 levels are elevated in the bone marrow serum of patients with mutilans-type rheumatoid arthritis. *Clin Rheumatol* 2002; **21**: 23-27 [PMID: 11954879 DOI: 10.1007/s100670200006]
 - 20 **Camilleri JP**, Amos N, Williams BD, Emery P, Williams LA, Jessop JD. Serum soluble interleukin 2 receptor levels and radiological progression in early rheumatoid arthritis. *J Rheumatol* 2001; **28**: 2576-2578 [PMID: 11764199]
 - 21 **Smolen JS**, Aletaha D, Redlich K. The pathogenesis of rheumatoid arthritis: new insights from old clinical data? *Nat Rev Rheumatol* 2012; **8**: 235-243 [PMID: 22410633 DOI: 10.1038/nrrheum.2012.23]
 - 22 **Matsuno H**, Yudoh K, Katayama R, Nakazawa F, Uzuki M, Sawai T, Yonezawa T, Saeki Y, Panayi GS, Pitzalis C, Kimura T. The role of TNF-alpha in the pathogenesis of inflammation and joint destruction in rheumatoid arthritis (RA): a study using a human RA/SCID mouse chimera. *Rheumatology (Oxford)* 2002; **41**: 329-337 [PMID: 11934972 DOI: 10.1093/rheumatology/41.3.329]
 - 23 **Tanaka Y**. Intensive treatment and treatment holiday of TNF-inhibitors in rheumatoid arthritis. *Curr Opin Rheumatol* 2012; **24**: 319-326 [PMID: 22388646 DOI: 10.1097/BOR.0b013e3283524e4c]
 - 24 **Balazs EA**, Watson D, Duff IF, Roseman S. Hyaluronic acid in synovial fluid. I. Molecular parameters of hyaluronic acid in normal and arthritis human fluids. *Arthritis Rheum* 1967; **10**: 357-376 [PMID: 6046018 DOI: 10.1002/art.1780100407]
 - 25 **Dahl LB**, Dahl IM, Engström-Laurent A, Granath K. Concentration and molecular weight of sodium hyaluronate in synovial fluid from patients with rheumatoid arthritis and other arthropathies. *Ann Rheum Dis* 1985; **44**: 817-822 [PMID: 4083937 DOI: 10.1136/ard.44.12.817]
 - 26 **Wells AF**, Klareskog L, Lindblad S, Laurent TC. Correlation between increased hyaluronan localized in arthritic synovium and the presence of proliferating cells. A role for macrophage-derived factors. *Arthritis Rheum* 1992; **35**: 391-396 [PMID: 1567487 DOI: 10.1002/art.1780350405]
 - 27 **Tsujiura S**, Saito K, Nawata M, Nakayamada S, Tanaka Y. Overcoming drug resistance induced by P-glycoprotein on lymphocytes in patients with refractory rheumatoid arthritis. *Ann Rheum Dis* 2008; **67**: 380-388 [PMID: 17660216 DOI: 10.1136/ard.2007.070821]
 - 28 **O'Brien MM**, Lacayo NJ, Lum BL, Kshirsagar S, Buck S, Ravindranath Y, Bernstein M, Weinstein H, Chang MN, Arceci RJ, Sikic BI, Dahl GV. Phase I study of valspodar (PSC-833) with mitoxantrone and etoposide in refractory and relapsed pediatric acute leukemia: a report from the Children's Oncology Group. *Pediatr Blood Cancer* 2010; **54**: 694-702 [PMID: 20209646 DOI: 10.1002/pbc.22366]
 - 29 **Morjani H**, Madoulet C. Immunosuppressors as multidrug resistance reversal agents. *Methods Mol Biol* 2010; **596**: 433-446 [PMID: 19949935 DOI: 10.1007/978-1-60761-416-6_19]
 - 30 **Suzuki K**, Saito K, Tsujiura S, Nakayamada S, Yamaoka K, Sawamukai N, Iwata S, Nawata M, Nakano K, Tanaka Y. Tacrolimus, a calcineurin inhibitor, overcomes treatment unresponsiveness mediated by P-glycoprotein on lymphocytes in refractory rheumatoid arthritis. *J Rheumatol* 2010; **37**: 512-520 [PMID: 20080907 DOI: 10.3899/jrheum.090048]

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Retrospective Study

Association of insulin resistance with serum ferritin and aminotransferases-iron hypothesis

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Institutional review board statement: The National Center for Health Statistics of the Centers for Disease Control and Prevention conducted the NHANES III in the United States from 1988 through 1994. This survey was designed to assess the health and nutrition status of a large representative sample in the United States. The survey and data collection was approved by the NHANES Institutional Review Board (IRB).

Informed consent statement: Documented consent was obtained from participants of the NHANES III at the participation of survey.

Conflict-of-interest statement: We have no financial relationships to disclose.

Data sharing statement: No additional data are available.

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Abstract

AIM: To investigate the relationship of iron indices with diabetes mellitus (DM) in those without hemochromatosis.

METHODS: This cross-sectional study examined data collected during the Third National Health and Nutrition Examination Survey (NHANES III). Only those who fasted properly and were not anemic with transferrin saturation < 45% were included ($n = 6849$). Insulin sensitivity and beta cell function were calculated from fasting glucose and insulin concentrations. Indices of iron metabolism were examined in the presence or absence of DM. We examined the relationship of insulin sensitivity and beta cell function with serum ferritin concentration. The influence of C-reactive protein and liver enzymes was also investigated.

RESULTS: Serum ferritin concentration was significantly higher in diabetic subjects ($P = 0.0001$ to < 0.000001). The difference remained significant after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement ($P = 0.03$ to < 0.000001). In those who did not take insulin, serum ferritin concentration was negatively associated with insulin sensitivity ($P = 0.05$ to 0.00001), but not with beta cell function. The alanine aminotransferase was correlated with serum ferritin concentration ($P = 0.02$ to < 0.000001) but not with insulin sensitivity, suggesting the role of the liver in iron-associated insulin resistance.

CONCLUSION: As most of diabetes is type 2 diabetes and insulin resistance is a cardinal feature of type 2 diabetes, disordered iron metabolism could play a role in the pathogenesis of insulin resistance and type 2 diabetes through its effect on liver function.

Key words: Diabetes mellitus; Insulin sensitivity; Beta cell function; Ferritin; Liver

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Core tip: Hemochromatosis and excess iron load has been implicated to play a role in the pathogenesis of diabetes mellitus. Serum ferritin concentration was significantly higher in diabetic subjects. Serum ferritin concentration was negatively associated with insulin sensitivity, but not with beta cell function. The association of alanine aminotransferase correlated with serum ferritin concentration, but not insulin sensitivity, suggesting the role of the liver in iron-associated insulin resistance. Disordered iron metabolism could play a role in the pathogenesis of insulin resistance and type 2 diabetes mellitus through its effect on liver function.

Huang J, Karnchanasorn R, Ou H-Y, Feng W, Chuang L-M, Chiu KC, Samoa R. Association of insulin resistance with serum ferritin and aminotransferases-iron hypothesis. *World J Exp Med* 2015; 5(4): 232-243 Available from: URL: <http://www.wjgnet.com>

INTRODUCTION

Diabetes mellitus (DM) is a common manifestation (53%-80%) of hereditary hemochromatosis^[1], which is an autosomal recessive disorder caused by mutations in a gene designated HFE (OMIM: 235200). A mutation, C282Y, was detected in 83% of the patients, while it was only found in 3.2% of control chromosomes^[2]. However, the allelic frequencies of C282Y mutation were similar between diabetic and control groups (6.3% vs 5.5%) in a large population from the United Kingdom^[3]. Meta-analysis of published studies showed no evidence for over-representation of this allele in patients with type 2 diabetes^[3]. Therefore, the C282Y mutation does not play a role in the pathogenesis of type 2 diabetes. Nevertheless, the role of iron metabolism in the pathogenesis of diabetes in the general population has been suggested in many cross-sectional studies^[4-7]. Furthermore, a nested case-control study suggested a potential interaction between the HFE genotypes and heme iron in relation to the risk of type 2 diabetes^[8].

In hereditary hemochromatosis, both insulin resistance and impaired insulin secretion have been suggested to play a role in its pathogenesis^[9]. The role of insulin resistance in patients with secondary hemochromatosis from thalassemia major has been reported, while an additional defect in beta cell secretion cannot be excluded^[10]. The association of serum ferritin concentration and insulin resistance has been reported in various liver diseases^[11,12]. Furthermore, the underlying mechanism of iron-associated abnormal glucose homeostasis in the general population is not well understood.

To examine the role of iron in the pathogenesis of diabetes, we investigated the iron indices and the relative influence of an inflammatory marker and liver enzymes on glucose homeostasis in a nationally representative survey, third National Health and Nutrition Examination Survey (NHANES III).

MATERIALS AND METHODS

Ethics statement

The National Center for Health Statistics of the Centers for Disease Control and Prevention conducted the NHANES III in the United States from 1988 through 1994. This survey was designed to assess the health and nutrition status of a large representative sample in the United States. The survey and data collection was approved by the NHANES Institutional Review Board and documented consent was obtained from participants. Analysis of de-identified data from the survey is exempt from the federal regulations for the protection of human research participants. Only de-identified data from the survey was used in this study.

Study design and study sample

Detailed descriptions of the survey and the analytical methods of various assays have been published^[13] and are also available at its website (<http://www.cdc.gov/nchs/about/major/nhanes/datalink.htm#NHANESIII>).

Race and ethnicity were self-reported by the participants. NHANES III was designed to provide reliable information from three major racial/ethnic groups: Non-Hispanic whites (NHW), non-Hispanic blacks (NHB), and Hispanics. The 4th group was excluded from this analysis for its small sample size and for encompassing a heterogeneous racial/ethnic group. There were 15021 subjects who had serum ferritin, fasting glucose and insulin concentration measured. Proper fasting is required to define diabetes status and to calculate insulin sensitivity and beta cell function from the fasting samples^[14,15]. Only those who fasted for ≥ 8 h and ≤ 16 h were included ($n = 7701$). We excluded 180 subjects with hemoglobin < 11 g/dL, which is frequently associated with iron deficiency and falsely low HbA_{1c}. Since hemochromatosis is an established cause of diabetes, those with transferrin saturation ≥ 45 were also excluded^[16] ($n = 672$), which identified 98% of iron-overloaded subjects^[17].

Ascertainment of DM

Diabetes was defined as a fasting glucose concentration ≥ 126 mg/dL (7.0 mmol/L) or a 2-h postchallenged glucose concentration ≥ 200 mg/dL (11.1 mmol/L)^[14]. Without the 2-h postchallenged glucose concentration, the diagnosis of diabetes is frequently missed in those with elevated 2-h postchallenged glucose concentrations and normal fasting glucose concentrations^[18]. However, only 3010 subjects had 2-h postchallenged glucose concentration measured. Their HbA_{1c} was very well correlated with 2-h postchallenged glucose concentration ($r = 0.7558$, $P < 0.000001$) and 2-h postchallenged glucose concentration of 200 mg/dL (11.1 mmol/L) was equivalent to HbA_{1c} of 6.3%. Therefore, we also defined diabetes in those with HbA_{1c} $\geq 6.3\%$.

Calculation of beta cell function and insulin sensitivity

Beta cell function (%B) and insulin sensitivity (%S) were calculated based on the homeostasis model assessment (HOMA)^[15,19].

$$\%B = (20 \times \text{fasting insulin concentration in mU/L}) / (\text{fasting glucose concentration in mmol/L} - 3.5).$$

$$\%S = 22.5 / (\text{fasting insulin concentration in mU/L} \times \text{fasting glucose concentration in mmol/L}).$$

Those with fasting glucose concentration < 3.5 mmol/L were excluded from analysis, since they had negative %B ($n = 9$). %B and %S obtained from the HOMA had been shown to correlate very well with the measured beta cell function and insulin sensitivity from various methods^[15,20-22]. A quantitative insulin

sensitivity check index (QUICKI), which had been shown to correlate with the measured insulin sensitivity by hyperinsulinemic euglycemic clamp very well^[23], was also used.

$$\text{QUICKI} = 1 / [\log_{10}(\text{fasting insulin concentration in mU/L}) + \log_{10}(\text{fasting glucose concentration in mg/dL})].$$

All of these methods have been validated in both non-diabetic subjects and diabetic subjects who did not take insulin^[21-23]. Those who took insulin were excluded from these analyses ($n = 51$).

Statistical analysis

General descriptive variables were expressed as means \pm SD. Since gender and ethnicity could potentially affect both iron metabolism and glucose homeostasis, the data were analyzed separately by gender and ethnic groups. Continuous variables were compared using two-tail Student *t* test between two groups or Analysis of Variance for more than two groups. Continuous data were expressed as means with 95%CI. Analysis of variance was used to examine the influence of covariates [age and body mass index (BMI)] on continuous variables between two groups. Least square regression analysis was used to investigate the relationship between two continuous variables. The influence of covariates (age, BMI, alcohol consumption, and mineral/iron intake) was also accounted for least square regression analysis. To further assess the association of serum ferritin concentration with estimated beta function and insulin sensitivity indices as well as the association of liver aminotransferases and C-reactive protein (CRP) with serum ferritin concentration and estimated insulin sensitivity indices, we also examine the trend across the quintile of serum ferritin concentration, liver aminotransferases and CRP. The comparisons were also adjusted for age, BMI, alcohol consumption, and mineral/iron intake. All the analyses were conducted in SYSTAT 11, Systat Software, Inc., Point Richmond, California, United States. A *P* value less than 0.05 was considered significant.

RESULTS**Study populations**

The clinical features of the studied subjects were shown by gender and ethnic groups in Table 1. Based on previously published upper reference ranges^[24], in male participants, 8.2% had elevated aspartate aminotransferase (AST > 37 U/L) and 9.6% had elevated alanine aminotransferase (ALT > 40 U/L) and in female participants, 7.7% had elevated AST (> 37 U/L) and 7.1% had elevated ALT (> 37 U/L).

Comparison of indices of iron metabolism in the presence or absence of diabetes

Iron, total iron binding capacity (TIBC), transferrin saturation, and serum ferritin concentration were

Table 1 Clinical features of studied subjects

	Non-Hispanic whites		Non-Hispanic blacks		Hispanics	
	Male	Female	Male	Female	Male	Female
<i>n</i>	1373	1602	896	1057	957	964
Age (yr)	55 ± 19	54 ± 20	43 ± 17	42 ± 16	42 ± 17	41 ± 17
Systolic blood pressure (mmHg)	130 ± 18	125 ± 21	128 ± 18	122 ± 20	125 ± 17	120 ± 20
Diastolic blood pressure (mmHg)	76 ± 10	72 ± 9	79 ± 11	74 ± 11	76 ± 10	71 ± 10
Body mass index (kg/m ²)	26.75 ± 4.63	26.39 ± 5.80	26.74 ± 5.34	29.07 ± 7.22	27.10 ± 4.64	28.26 ± 5.97
Transferrin saturation (%)	27.93 ± 8.39	24.58 ± 8.78	26.22 ± 8.11	21.75 ± 8.51	28.23 ± 8.59	22.95 ± 8.98
Ferritin (mcg/L)	176 ± 142	95 ± 104	215 ± 170	93 ± 111	169 ± 142	67 ± 95
Aspartate aminotransferase (U/L)	22 ± 8	19 ± 8	25 ± 17	22 ± 8	27 ± 16	23 ± 19
Alanine aminotransferase (U/L)	18 ± 11	14 ± 10	19 ± 13	18 ± 11	28 ± 24	21 ± 23
Gamma glutamyl transferase (U/L)	35 ± 55	21 ± 20	46 ± 59	35 ± 55	46 ± 53	30 ± 31
C-reactive protein (mg/dL)	0.43 ± 0.76	0.45 ± 0.62	0.48 ± 0.79	0.53 ± 0.65	0.44 ± 0.84	0.53 ± 0.80

Data presented mean ± SD.

Table 2 Comparison of serum indices of iron by the presence or absence of diabetes mellitus

	Non-Hispanic whites				Non-Hispanic blacks				Hispanics			
	Male		Female		Male		Female		Male		Female	
	DM	Non-DM	DM	Non-DM	DM	Non-DM	DM	Non-DM	DM	Non-DM	DM	Non-DM
<i>n</i>	166	1207	137	1465	98	798	106	951	115	842	132	832
Iron (mcg/dL)												
Mean	93	95	83	87	82	87	70	75	90	98	82	84
(95%CI)	(89, 97)	(94, 97)	(78, 88)	(86, 89)	(77, 87)	(85, 89)	(64, 75)	(74, 77)	(84, 97)	(96, 100)	(76, 87)	(82, 86)
<i>P</i> ¹		NS		NS		NS		NS		0.01		NS
<i>P</i> ²		NS		NS		NS		NS		NS		NS
<i>P</i> ³		NS		NS		NS		NS		NS		NS
Total iron binding capacity (mcg/dL)												
Mean	351	344	359	360	333	334	342	353	353	354	362	377
(95%CI)	(343, 359)	(341, 346)	(350, 368)	(357, 363)	(322, 344)	(330, 337)	(331, 352)	(349, 356)	(343, 363)	(351, 358)	(351, 373)	(372, 381)
<i>P</i> ¹		NS		NS		NS		NS		NS		0.01
<i>P</i> ²		0.002		NS		NS		NS		NS		NS
<i>P</i> ³		NS		NS		NS		NS		NS		NS
Transferrin saturation (%)												
Mean	27	28	23	25	25	26	21	22	26	28	23	23
(95%CI)	(26, 28)	(28, 29)	(22, 25)	(24, 25)	(23, 26)	(26, 27)	(19, 22)	(21, 22)	(24, 28)	(27, 28)	(22, 25)	(22, 23)
<i>P</i> ¹		NS		NS		NS		NS		0.03		NS
<i>P</i> ²		NS		NS		NS		NS		NS		NS
<i>P</i> ³		NS		NS		NS		NS		NS		NS
Ferritin (mcg/L)												
Mean	228	169	152	84	282	206	167	86	230	150	138	56
(95%CI)	(200, 256)	(161, 177)	(129, 175)	(79, 89)	(237, 327)	(195, 218)	(139, 195)	(79, 92)	(190, 270)	(142, 159)	(104, 171)	(52, 60)
<i>P</i> ¹		< 0.000001		< 0.000001		0.00003		< 0.000001		< 0.000001		< 0.000001
<i>P</i> ²		0.0001		0.000003		0.02		0.0001		0.00001		< 0.000001
<i>P</i> ³		0.0002		0.000003		0.03		0.0001		0.000004		< 0.000001

Data presented mean with 95%CI. To convert values for iron and total iron binding capacity to mol/L, multiply by 0.1791. ¹*P* values for unadjusted comparison; ²*P* values for comparison after adjustment for age and body mass index; ³*P* values for comparison after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement. NS: Not significant (*P* > 0.05); DM: Diabetes mellitus; NDM: Non-diabetic mellitus.

compared between those with or without diabetes (Table 2). No consistent results were observed for iron, TIBC, and transferrin saturation, while serum ferritin concentration was markedly higher in diabetic than in non-diabetic subjects in all six groups. Diabetic subjects were older than non-diabetic subjects by 14-16 years (*P* < 0.000001) and were also more obese than non-diabetic subjects per BMI by 2.40-4.81 kg/m² (*P* < 0.000001). The difference in ferritin concentration between diabetic and non-diabetic subjects remained significant after adjustment of age, BMI, alcohol intake, and mineral/iron supplement. Thus, diabetes was

associated with elevated serum ferritin concentration.

Association of serum ferritin concentration with beta cell function and insulin sensitivity

Diabetes results from an imbalance between beta cell function and insulin sensitivity. Thus, serum ferritin concentration could potentially be associated with either beta cell function, insulin sensitivity, or both. No association between %B and serum ferritin concentration was found after adjustment for both age and BMI (Table 3). In contrast, serum ferritin concentration was negatively associated with %S and

Table 3 Estimated beta cell function and insulin sensitivity indices by quintile of serum ferritin concentrations

	Quintile 1	Quintile 2	Quintile 3	Quintile 4	Quintile 5	P ¹	P ²	P ³
Beta cell function by the homeostasis model assessment (%B) ⁴								
Non-Hispanic white males	103 (96, 110)	98 (92, 105)	97 (90, 104)	106 (94, 118)	113 (102, 124)	NS	NS	NS
Non-Hispanic white females	117 (110, 124)	115 (108, 122)	109 (103, 114)	111 (104, 119)	110 (102, 118)	NS	NS	NS
Non-Hispanic black males	113 (104, 122)	118 (107, 130)	112 (103, 120)	144 (107, 181)	117 (106, 129)	NS	NS	NS
Non-Hispanic black females	177 (156, 197)	178 (161, 196)	151 (137, 164)	135 (121, 150)	135 (123, 147)	0.000001	NS	NS
Hispanic males	104 (91, 118)	112 (101, 124)	106 (97, 114)	123 (106, 141)	128 (116, 140)	0.004	NS	NS
Hispanic females	152 (140, 165)	150 (135, 164)	140 (129, 150)	136 (124, 147)	130 (119, 141)	0.003	NS	NS
Insulin sensitivity by the homeostasis model assessment (%S) ⁵								
Non-Hispanic white males	0.503 (0.472, 0.535)	0.549 (0.513, 0.585)	0.518 (0.484, 0.553)	0.494 (0.463, 0.525)	0.421 (0.388, 0.455)	0.00005	NS	0.05
Non-Hispanic white females	0.643 (0.610, 0.677)	0.603 (0.570, 0.637)	0.574 (0.539, 0.609)	0.555 (0.519, 0.591)	0.455 (0.420, 0.491)	< 0.000001	0.04	0.02
Non-Hispanic black males	0.562 (0.516, 0.608)	0.553 (0.504, 0.601)	0.559 (0.512, 0.605)	0.555 (0.498, 0.611)	0.48 (0.426, 0.535)	0.04	NS	0.05
Non-Hispanic black females	0.510 (0.476, 0.545)	0.479 (0.437, 0.522)	0.453 (0.421, 0.485)	0.458 (0.422, 0.495)	0.379 (0.343, 0.414)	0.000001	0.02	0.007
Hispanic males	0.538 (0.495, 0.580)	0.533 (0.489, 0.578)	0.527 (0.472, 0.583)	0.524 (0.460, 0.588)	0.338 (0.307, 0.370)	< 0.000001	0.01	0.007
Hispanic females	0.523 (0.482, 0.563)	0.535 (0.484, 0.587)	0.464 (0.426, 0.502)	0.444 (0.403, 0.485)	0.321 (0.289, 0.352)	< 0.000001	0.005	0.003
Insulin sensitivity by the simple QUICKI ⁶								
Non-Hispanic white males	0.339 (0.336, 0.342)	0.343 (0.340, 0.346)	0.340 (0.336, 0.343)	0.337 (0.334, 0.341)	0.327 (0.323, 0.331)	< 0.000001	0.008	0.002
Non-Hispanic white females	0.352 (0.349, 0.355)	0.348 (0.345, 0.351)	0.345 (0.342, 0.348)	0.343 (0.340, 0.346)	0.33 (0.327, 0.334)	< 0.000001	0.00005	0.00001
Non-Hispanic black males	0.344 (0.339, 0.348)	0.342 (0.338, 0.347)	0.343 (0.339, 0.348)	0.341 (0.335, 0.346)	0.333 (0.328, 0.338)	0.003	0.03	0.01
Non-Hispanic black females	0.340 (0.336, 0.343)	0.335 (0.331, 0.339)	0.333 (0.329, 0.337)	0.333 (0.329, 0.337)	0.322 (0.317, 0.326)	< 0.000001	0.0006	0.0001
Hispanic males	0.341 (0.336, 0.345)	0.34 (0.336, 0.345)	0.339 (0.335, 0.344)	0.337 (0.332, 0.342)	0.318 (0.314, 0.322)	< 0.000001	0.0001	0.00001
Hispanic females	0.340 (0.336, 0.344)	0.340 (0.336, 0.345)	0.334 (0.330, 0.338)	0.330 (0.326, 0.335)	0.315 (0.310, 0.319)	< 0.000001	0.00003	0.00002

Data presented mean with 95%CI. ¹P values for trend, unadjusted; ²P values for trend, after adjustment for age and body mass index; ³P values for trend, after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement; ⁴%B = (20 × fasting insulin concentration in mU/L) / (fasting glucose concentration in mmol/L - 3.5). Those with negative %B were excluded from analysis; ⁵%S = 22.5/(fasting insulin concentration in mU/L × fasting glucose concentration in mmol/L); ⁶Quicki = 1/[log(fasting glucose concentration in mg/dL) + log(fasting insulin concentration in mU/L)]. NS: Not significant; QUICKI: Quantitative insulin sensitivity check index.

QUICKI in all six groups. This relationship persisted after adjustment for age, BMI, alcohol consumption, and mineral/iron supplement. Therefore, we concluded that ferritin concentration was negatively associated with insulin sensitivity.

Role of inflammation in association between serum ferritin concentration and insulin sensitivity

In addition to reflection of the body iron store, serum ferritin is also an acute reactant. To explore the role of inflammation on the observed correlation, we examined the relationship of these indices with a marker of inflammation, CRP. No consistent association of CRP with either %S or QUICKI was observed (Table 4). A positive association between CRP and serum ferritin concentration was observed in NHW (both males and females), non-Hispanic black females, and Hispanic

females, but not in non-Hispanic black. The associations remained unchanged after adjustment for age, BMI, alcohol intake, and mineral/iron supplement. Therefore, inflammation could not provide a uniform explanation for the underlying mechanism of the observed correlation between serum ferritin concentration and insulin sensitivity.

Role of the liver in association between serum ferritin concentration and insulin sensitivity

Elevated liver enzymes could result from iron deposition in the liver. To examine the role of the liver in the association of serum ferritin concentration with insulin sensitivity, we investigated the association of liver enzymes with serum ferritin concentration and insulin sensitivity. We focused on AST or SGOT, gamma glutamyl transpeptidase (GGT), and ALT or SGPT.

Table 4 Serum ferritin concentration and estimated insulin sensitivity indices by quintile of inflammatory marker - C-reactive protein

	Quintile 1	Quintile 2	Quintile 3	Quintile 4	Quintile 5	P ¹	P ²	P ³
Serum ferritin concentration (mcg/L)								
Non-Hispanic white males	142 (130, 155)	157 (144, 170)	179 (164, 195)	201 (180, 222)	201 (181, 221)	< 0.000001	0.000002	0.000005
Non-Hispanic white females	19 (18, 20)	49 (48, 50)	120 (116, 125)	152 (134, 171)	108 (95, 120)	0.000003	0.00003	0.00003
Non-Hispanic black males	186 (163, 209)	187 (167, 207)	267 (236, 298)	227 (202, 253)	254 (281, 228)	NS	NS	NS
Non-Hispanic black females	90 (74, 106)	79 (66, 91)	77 (65, 89)	102 (87, 118)	115 (98, 132)	0.002	0.0009	0.001
Hispanic males	150 (134, 166)	157 (135, 179)	161 (137, 185)	157 (136, 178)	172 (154, 191)	NS	NS	NS
Hispanic females	52 (44, 60)	56 (48, 64)	63 (53, 72)	72 (60, 84)	92 (68, 115)	0.00001	0.007	0.006
Insulin sensitivity by the homeostasis model assessment (%S) ⁴								
Non-Hispanic white males	0.541 (0.505, 0.578)	0.531 (0.500, 0.562)	0.546 (0.509, 0.582)	0.459 (0.425, 0.494)	0.412 (0.383, 0.440)	< 0.000001	0.05	0.03
Non-Hispanic white females	0.684 (0.651, 0.718)	0.627 (0.592, 0.661)	0.596 (0.558, 0.634)	0.494 (0.461, 0.526)	0.422 (0.392, 0.452)	< 0.000001	0.004	0.004
Non-Hispanic black males	0.562 (0.517, 0.607)	0.572 (0.526, 0.618)	0.561 (0.499, 0.623)	0.549 (0.488, 0.611)	0.478 (0.426, 0.530)	0.05	NS	NS
Non-Hispanic black females	0.524 (0.489, 0.558)	0.524 (0.487, 0.560)	0.497 (0.464, 0.530)	0.404 (0.369, 0.438)	0.327 (0.288, 0.365)	< 0.000001	0.004	0.002
Hispanic males	0.558 (0.507, 0.608)	0.484 (0.438, 0.530)	0.525 (0.472, 0.578)	0.513 (0.460, 0.566)	0.372 (0.334, 0.411)	0.00001	NS	NS
Hispanic females	0.538 (0.495, 0.582)	0.53 (0.481, 0.579)	0.471 (0.430, 0.513)	0.392 (0.355, 0.429)	0.353 (0.319, 0.387)	< 0.000001	NS	NS
Insulin sensitivity by the simple QUICKI ⁵								
Non-Hispanic white males	0.342 (0.339, 0.346)	0.342 (0.339, 0.345)	0.342 (0.339, 0.346)	0.332 (0.328, 0.336)	0.328 (0.325, 0.331)	< 0.000001	0.01	0.008
Non-Hispanic white females	0.356 (0.354, 0.359)	0.351 (0.348, 0.354)	0.347 (0.343, 0.350)	0.336 (0.333, 0.340)	0.328 (0.324, 0.331)	< 0.000001	0.004	0.005
Non-Hispanic black males	0.345 (0.341, 0.349)	0.345 (0.340, 0.349)	0.341 (0.336, 0.346)	0.34 (0.334, 0.345)	0.332 (0.327, 0.338)	0.01	NS	NS
Non-Hispanic black females	0.341 (0.337, 0.345)	0.341 (0.337, 0.345)	0.339 (0.335, 0.342)	0.327 (0.323, 0.331)	0.314 (0.310, 0.319)	< 0.000001	0.000002	0.000001
Hispanic males	0.342 (0.338, 0.347)	0.335 (0.331, 0.340)	0.338 (0.333, 0.343)	0.338 (0.333, 0.342)	0.321 (0.317, 0.326)	< 0.000001	NS	NS
Hispanic females	0.342 (0.338, 0.346)	0.34 (0.336, 0.344)	0.334 (0.329, 0.338)	0.325 (0.321, 0.329)	0.319 (0.314, 0.323)	< 0.000001	0.02	0.03

Data presented mean with 95%CI. ¹P values for trend, unadjusted; ²P values for trend, after adjustment for age and body mass index; ³P values for trend, after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement; ⁴%S = 22.5/(fasting insulin concentration in mU/L × fasting glucose concentration in mmol/L); ⁵Quicki = 1/[log(fasting glucose concentration in mg/dL) + log(fasting insulin concentration in mU/L)]. NS: Not significant; QUICKI: Quantitative insulin sensitivity check index.

The association of AST with %S and QUICKI was only noted in Hispanic males, but not in other 5 groups (Table 5). Adjustment for age, BMI, alcohol consumption, and mineral/iron supplement had no impact on the results. In contrast, a very close association was noted between AST and ferritin concentration in all 6 groups. Since AST is present in many tissues, including heart, skeletal muscle, kidney, and brain, we could not exclude the role of the liver based on no association between AST and insulin sensitivity in some groups.

GGT is a very sensitive indicator of hepatobiliary diseases and is found predominately throughout the hepatobiliary system, but also in other tissues. It was negatively associated with %S and QUICKI in all 6 groups (Table 6). The association remained after adjustment for age, BMI, alcohol consumption, and mineral/iron supplement. It was positively associated with serum ferritin concentration after adjustment for

age and BMI. Therefore, the relationship of GGT with insulin sensitivity and serum ferritin concentration could provide a mechanistic insight of the liver in the association between insulin sensitivity and serum ferritin concentration.

The primary source of ALT is the liver. It was negatively associated with both %S and QUICKI in all six groups, and remained highly significant regardless of the adjustment for age, BMI, alcohol consumption, and mineral/iron supplement (Table 7). This relationship indicated an association of insulin resistance and liver diseases. Serum ferritin concentration was positively associated with ALT and also this association remained significant regardless of adjustment for age, BMI, alcohol consumption, and mineral/iron supplement. Since ALT is an indicator of liver diseases, a positive association between ALT and serum ferritin concentration suggests that increased iron deposition

Table 5 Serum ferritin concentration and estimated insulin sensitivity indices by quintile of aspartate aminotransferase

	Quintile 1	Quintile 2	Quintile 3	Quintile 4	Quintile 5	P ¹	P ²	P ³
Serum ferritin concentration(mcg/L)								
Non-Hispanic white males	148 (134, 162)	154 (140, 167)	180 (165, 196)	170 (155, 186)	228 (205, 251)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	58 (51, 65)	61 (55, 68)	85 (76, 94)	102 (91, 112)	140 (122, 158)	0.0004	0.02	0.02
Non-Hispanic black males	175 (155, 196)	203 (178, 228)	197 (173, 222)	227 (203, 251)	267 (236, 298)	0.05	0.003	0.004
Non-Hispanic black females	84 (71, 96)	77 (66, 88)	80 (67, 92)	95 (81, 109)	127 (106, 149)	0.00001	0.004	0.005
Hispanic males	139 (124, 153)	134 (118, 151)	149 (134, 163)	155 (132, 177)	223 (194, 251)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	47 (39, 56)	49 (42, 55)	59 (50, 68)	79 (57, 101)	101 (86, 115)	< 0.000001	0.00001	0.00002
Insulin sensitivity by the homeostasis model assessment (%S) ⁴								
Non-Hispanic white males	0.491 (0.457, 0.526)	0.517 (0.484, 0.550)	0.497 (0.466, 0.528)	0.506 (0.471, 0.541)	0.478 (0.441, 0.514)	NS	NS	NS
Non-Hispanic white females	0.575 (0.544, 0.607)	0.597 (0.559, 0.630)	0.591 (0.554, 0.628)	0.555 (0.521, 0.588)	0.506 (0.470, 0.543)	0.02	NS	NS
Non-Hispanic black males	0.512 (0.465, 0.559)	0.531 (0.479, 0.584)	0.528 (0.486, 0.569)	0.557 (0.508, 0.605)	0.561 (0.499, 0.623)	NS	NS	NS
Non-Hispanic black females	0.447 (0.410, 0.484)	0.452 (0.418, 0.487)	0.472 (0.437, 0.506)	0.461 (0.424, 0.499)	0.443 (0.401, 0.485)	NS	NS	NS
Hispanic males	0.501 (0.460, 0.542)	0.55 (0.495, 0.604)	0.524 (0.473, 0.574)	0.477 (0.424, 0.530)	0.404 (0.359, 0.449)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	0.472 (0.430, 0.514)	0.48 (0.443, 0.516)	0.468 (0.430, 0.507)	0.49 (0.442, 0.539)	0.374 (0.330, 0.418)	0.006	NS	NS
Insulin sensitivity by the simple QUICKI ⁵								
Non-Hispanic white males	0.337 (0.333, 0.340)	0.34 (0.337, 0.343)	0.338 (0.335, 0.341)	0.338 (0.334, 0.342)	0.334 (0.330, 0.338)	NS	NS	NS
Non-Hispanic white females	0.346 (0.343, 0.349)	0.347 (0.344, 0.350)	0.347 (0.343, 0.350)	0.343 (0.340, 0.346)	0.336 (0.332, 0.340)	0.009	NS	NS
Non-Hispanic black males	0.337 (0.332, 0.342)	0.339 (0.334, 0.344)	0.341 (0.337, 0.345)	0.343 (0.338, 0.348)	0.3417 (0.336, 0.346)	NS	NS	NS
Non-Hispanic black females	0.331 (0.327, 0.335)	0.332 (0.328, 0.336)	0.335 (0.332, 0.339)	0.333 (0.329, 0.337)	0.33 (0.326, 0.334)	NS	NS	NS
Hispanic males	0.337 (0.333, 0.342)	0.342 (0.337, 0.346)	0.338 (0.333, 0.343)	0.333 (0.328, 0.338)	0.325 (0.320, 0.329)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	0.334 (0.329, 0.338)	0.336 (0.332, 0.340)	0.334 (0.330, 0.338)	0.335 (0.331, 0.340)	0.32 (0.315, 0.325)	0.0001	NS	NS

Data presented mean with 95%CI. ¹P values for trend, unadjusted; ²P values for trend, after adjustment for age and body mass index; ³P values for trend, after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement; ⁴%S = 22.5/(fasting insulin concentration in mU/L × fasting glucose concentration in mmol/L); ⁵Quicki = 1/[log(fasting glucose concentration in mg/dL) + log(fasting insulin concentration in mU/L)]. NS: Not significant; QUICKI: Quantitative insulin sensitivity check index.

in the liver is associated with liver dysfunction. Furthermore, a positive association between ALT and serum ferritin concentration and a negative association between ALT and insulin sensitivity suggests a negative association of serum ferritin concentration with insulin sensitivity as we had observed.

DISCUSSION

To examine the role of iron metabolism in diabetes, the indices of iron metabolism were compared in patients with and without diabetes. We found that subjects with diabetes had a higher serum ferritin concentration than those without diabetes. To explore the underlying pathophysiology, we observed that serum ferritin concentration was negatively associated with insulin sensitivity (%S and QUICKI), but not with beta cell function. Therefore, a high serum ferritin concentration

is associated with insulin resistance and is a risk factor for DM.

Since ferritin contains the second largest pool of iron in the body next to hemoglobin^[16], serum ferritin concentration closely correlates with total body iron stores, mainly in the liver^[26]. In this study, serum ferritin concentration is negatively associated with insulin sensitivity, suggesting an association of insulin resistance with total body iron stores. However, it is well known that ferritin is also an acute phase reactant^[26]. To further explore this issue, we examined the correlation of CRP, an inflammatory marker^[27], with insulin sensitivity and serum ferritin concentration. Without a consistent result across 3 ethnic/racial groups and both genders, we concluded that in this study, no consistent relationship of CRP with either insulin sensitivity or serum ferritin concentration was observed in all 6 groups. Furthermore, in this population only

Table 6 Serum ferritin concentration and estimated insulin sensitivity indices by quintile of gamma glutamyl transferase

	Quintile 1	Quintile 2	Quintile 3	Quintile 4	Quintile 5	P ¹	P ²	P ³
Serum ferritin concentration (mcg/L)								
Non-Hispanic white males	141 (125, 157)	146 (131,161)	174 (156, 191)	179 (162, 196)	227 (204, 250)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	55 (48, 62)	70 (61, 78)	95 (81, 110)	92 (77, 107)	137 (121, 154)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic black males	165 (144, 187)	191 (163, 219)	221 (195, 247)	220 (194, 246)	298 (263, 333)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic black females	60 (45, 74)	79 (68, 91)	84 (67, 100)	97 (83, 112)	141 (119, 163)	< 0.000001	< 0.000001	< 0.000001
Hispanic males	105 (93, 117)	137 (121, 153)	152 (132, 172)	173 (144, 202)	240 (208, 272)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	38 (30, 47)	38 (32, 44)	59 (47, 70)	82 (70, 93)	109 (90, 129)	< 0.000001	< 0.000001	< 0.000001
Insulin sensitivity by the homeostasis model assessment (%S) ⁴								
Non-Hispanic white males	0.622 (0.584, 0.659)	0.521 (0.486, 0.556)	0.5061 (0.465, 0.546)	0.422 (0.387, 0.458)	0.402 (0.367, 0.436)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	0.732 (0.695, 0.769)	0.648 (0.609, 0.686)	0.587 (0.547, 0.626)	0.48 (0.444, 0.516)	0.397 (0.361, 0.433)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic black males	0.607 (0.560, 0.654)	0.588 (0.522, 0.653)	0.537 (0.491, 0.583)	0.489 (0.429, 0.548)	0.462 (0.404, 0.520)	0.0001	NS	0.05
Non-Hispanic black females	0.554 (0.515, 0.594)	0.454 (0.419, 0.489)	0.45 (0.409, 0.491)	0.407 (0.372, 0.442)	0.383 (0.332, 0.434)	< 0.000001	0.0002	0.00002
Hispanic males	0.67 (0.625, 0.715)	0.562 (0.486, 0.638)	0.423 (0.383, 0.463)	0.439 (0.376, 0.501)	0.359 (0.299, 0.419)	< 0.000001	0.00005	0.000005
Hispanic females	0.623 (0.569, 0.677)	0.494 (0.448, 0.539)	0.424 (0.381, 0.466)	0.338 (0.301, 0.375)	0.343 (0.298, 0.389)	< 0.000001	0.000002	0.000003
Insulin sensitivity by the simple QUICKI ⁵								
Non-Hispanic white males	0.351 (0.348, 0.354)	0.341 (0.338, 0.344)	0.339 (0.335, 0.342)	0.328 (0.324, 0.332)	0.325 (0.321, 0.330)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	0.361 (0.358, 0.364)	0.353 (0.350, 0.356)	0.347 (0.344, 0.350)	0.335 (0.332, 0.339)	0.323 (0.319, 0.327)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic black males	0.349 (0.345, 0.353)	0.344 (0.338, 0.350)	0.342 (0.337, 0.346)	0.334 (0.328, 0.339)	0.331 (0.325, 0.337)	< 0.000001	0.01	0.004
Non-Hispanic black females	0.345 (0.341, 0.348)	0.334 (0.330, 0.338)	0.332 (0.328, 0.337)	0.327 (0.323, 0.332)	0.321 (0.315, 0.326)	< 0.000001	< 0.000001	< 0.000001
Hispanic males	0.356 (0.352, 0.360)	0.341 (0.335, 0.347)	0.330 (0.325, 0.334)	0.329 (0.323, 0.334)	0.318 (0.312, 0.324)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	0.351 (0.346, 0.355)	0.338 (0.334, 0.342)	0.329 (0.325, 0.334)	0.318 (0.313, 0.323)	0.316 (0.311, 0.322)	< 0.000001	< 0.000001	< 0.000001

Data presented mean with 95%CI. ¹P values for trend, unadjusted; ²P values for trend, after adjustment for age and body mass index; ³P values for trend, after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement; ⁴%S = 22.5/(fasting insulin concentration in mU/L × fasting glucose concentration in mmol/L); ⁵Quicki = 1/[log(fasting glucose concentration in mg/dL) + log(fasting insulin concentration in mU/L)]. NS: Not significant; QUICKI: Quantitative insulin sensitivity check index.

1.28% (range: 0.64% in Hispanic males to 1.95% in non-Hispanic black males) of the participants had an elevated CRP ≥ 3 mg/L, which is the 90th percentile for healthy young adults^[28]. After exclusion of those with a CRP ≥ 3 mg/L, association of CRP with insulin sensitivity was only observed in NHW females ($P = 0.006$ for %S and $P = 0.00002$ for QUICKI) and NHB females ($P = 0.03$ for %S and $P = 0.0001$ for QUICKI) and correlation of CRP with ferritin concentration was only observed in NHW females ($P = 0.005$), NHB females ($P = 0.001$), and MA females ($P = 0.01$), after adjustment for age and BMI. Therefore, it is very unlikely that inflammation is the underlying mechanism for the observed negative association between serum ferritin concentration and insulin sensitivity.

Next we examined the role of the liver on the observed association between insulin sensitivity and serum ferritin concentration. Among these three liver

enzymes, AST is the least specific liver maker and ALT is the most liver specific marker. All three liver enzymes were correlated with serum ferritin concentration very well, suggesting a closed association between elevated serum ferritin and liver dysfunction. However, we observed the different strengths of the correlation of insulin sensitivity across three liver enzymes. Among them, AST is the least specific for the liver diseases, the association between AST and insulin sensitivity only observed in Hispanic male group. In contrast, a negative association of ALT and GGT with insulin sensitivity was observed in all 6 groups. The negative correlation of liver enzymes with insulin sensitivity indicates the role of hepatic dysfunction in insulin resistance. Therefore, these observations imply the role of iron-associated elevated ALT and GGT in the pathogenesis of insulin resistance. The role of the liver in the pathogenesis of DM is well-established^[29,30]. Furthermore, the relation-

Table 7 Serum ferritin concentration and estimated insulin sensitivity indices by quintile of alanine aminotransferase

	Quintile 1	Quintile 2	Quintile 3	Quintile 4	Quintile 5	P ¹	P ²	P ³
Serum ferritin concentration (mcg/L)								
Non-Hispanic white males	147 (132, 162)	169 (153, 185)	168 (152, 183)	177 (159, 194)	220 (200, 241)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	71 (63, 79)	71 (61, 82)	86 (77, 95)	95 (84, 107)	123 (107, 139)	0.05	0.02	0.02
Non-Hispanic black males	168 (147, 189)	187 (165, 210)	201 (177, 225)	247 (219, 275)	267 (238, 296)	0.01	0.00005	0.00005
Non-Hispanic black females	80 (67, 92)	79 (68, 89)	87 (73, 100)	98 (81, 114)	120 (100, 140)	0.00003	0.000004	0.000005
Hispanic males	128 (113, 143)	128 (116, 140)	142 (126, 157)	172 (144, 200)	229 (206, 253)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	42 (35, 49)	70 (48, 93)	57 (49, 65)	64 (55, 73)	101 (87, 116)	< 0.000001	0.000009	0.000009
Insulin sensitivity by the homeostasis model assessment (%S) ⁴								
Non-Hispanic white males	0.551 (0.517, 0.584)	0.544 (0.511, 0.578)	0.512 (0.476, 0.549)	0.483 (0.448, 0.518)	0.399 (0.370, 0.429)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	0.634 (0.602, 0.666)	0.628 (0.591, 0.664)	0.583 (0.546, 0.622)	0.55 (0.519, 0.582)	0.429 (0.395, 0.463)	0.00002	0.004	0.002
Non-Hispanic black males	0.63 (0.576, 0.684)	0.576 (0.522, 0.630)	0.564 (0.514, 0.615)	0.478 (0.423, 0.524)	0.44 (0.397, 0.482)	0.00002	0.003	0.003
Non-Hispanic black females	0.527 (0.490, 0.563)	0.498 (0.462, 0.534)	0.462 (0.424, 0.500)	0.427 (0.389, 0.465)	0.363 (0.330, 0.396)	< 0.000001	< 0.000001	< 0.000001
Hispanic males	0.635 (0.569, 0.700)	0.57 (0.525, 0.615)	0.48 (0.430, 0.530)	0.448 (0.410, 0.486)	0.324 (0.295, 0.352)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	0.55 (0.509, 0.591)	0.528 (0.484, 0.571)	0.468 (0.427, 0.509)	0.409 (0.365, 0.452)	0.331 (0.295, 0.368)	< 0.000001	0.000003	0.000002
Insulin sensitivity by the simple QUICKI ⁵								
Non-Hispanic white males	0.344 (0.341, 0.347)	0.343 (0.339, 0.346)	0.339 (0.335, 0.342)	0.335 (0.332, 0.339)	0.326 (0.322, 0.329)	< 0.000001	< 0.000001	< 0.000001
Non-Hispanic white females	0.352 (0.349, 0.355)	0.35 (0.347, 0.345)	0.346 (0.342, 0.349)	0.343 (0.340, 0.346)	0.327 (0.324, 0.331)	0.000001	0.0005	0.0002
Non-Hispanic black males	0.349 (0.344, 0.354)	0.344 (0.339, 0.349)	0.343 (0.338, 0.348)	0.335 (0.330, 0.339)	0.33 (0.326, 0.335)	0.00005	0.01	0.009
Non-Hispanic black females	0.341 (0.337, 0.345)	0.338 (0.334, 0.342)	0.333 (0.329, 0.337)	0.33 (0.326, 0.334)	0.321 (0.316, 0.325)	< 0.000001	< 0.000001	< 0.000001
Hispanic males	0.348 (0.343, 0.353)	0.345 (0.341, 0.349)	0.333 (0.328, 0.338)	0.332 (0.328, 0.336)	0.317 (0.313, 0.321)	< 0.000001	< 0.000001	< 0.000001
Hispanic females	0.343 (0.339, 0.347)	0.34 (0.336, 0.344)	0.334 (0.330, 0.338)	0.327 (0.323, 0.331)	0.315 (0.311, 0.320)	< 0.000001	< 0.000001	< 0.000001

Data presented mean with 95%CI. ¹P values for trend, unadjusted; ²P values for trend, after adjustment for age and body mass index; ³P values for trend, after adjustment for age, body mass index, alcohol consumption, and mineral/iron supplement; ⁴%S = 22.5/(fasting insulin concentration in mU/L × fasting glucose concentration in mmol/L); ⁵Quicki = 1/[log(fasting glucose concentration in mg/dL) + log(fasting insulin concentration in mU/L)]. NS: Not significant; QUICKI: Quantitative insulin sensitivity check index.

ship of elevated ALT concentration with diabetes and insulin resistance has been reported^[31]. In Pima Indians, elevated ALT was associated with hepatic insulin resistance but not with whole body insulin resistance or beta cell function^[32]. Insulin sensitivity obtained from the HOMA as used in this study, has been demonstrated to be correlated with hepatic insulin sensitivity^[33]. Thus from the observations in this study, iron could play a role in hepatic insulin resistance.

The role of serum ferritin concentration in diabetes^[4] has been examined in this population. Elevated ferritin concentration has been reported to be associated with an increased risk of diabetes, but the role of inflammation could not be excluded^[4]. Elevated serum ferritin concentration also has been reported to be associated with insulin resistance^[34]. In the present study, we confirmed the association of serum ferritin concentration with DM and insulin sensitivity assessed by both %S

and QUICKI. Although the association of inflammation and insulin resistance has been demonstrated in this population^[35], we demonstrated that the inflammatory hypothesis is not likely the underlying mechanism of the reported associations in this study. In addition, from the observed associations of ALT and GGT with serum ferritin concentration and insulin sensitivity, we provided the evidence suggesting that the role of liver in the pathogenesis of iron-associated insulin resistance.

Iron-induced oxidant stress has proposed to play a key role in iron-mediated tissue damage^[36-38]. Although the molecular events of iron-mediated tissue damage have not been fully elucidated, mitochondria are the targets of iron-mediated damage and iron may be preferentially toxic to cells with high mitochondrial activity^[39], such as hepatocytes and pancreatic beta cells. Impaired mitochondrial activity has been observed in the insulin-resistant offspring of patients with type

2 diabetes^[40] and mitochondrial defect can also lead to the metabolic syndrome^[41]. Therefore, iron-induced oxidative stress with mitochondrial dysfunction could be one of the underlying mechanisms in these metabolic disorders.

Because of the cross-sectional nature of the study, a temporal relationship and the biological basis of the association between serum ferritin concentration and these metabolic disorders could not be established. However, our observations have some bearing on the plausible mechanisms. Furthermore, the caustic role of iron in these processes is suggested by interventional studies. In patients with clinical evidence of non-alcoholic fatty liver disease, quantitative phlebotomy induced iron depletion to a level of near-iron deficiency results in a 40%-55% improvement of both fasting and glucose-stimulated plasma insulin concentrations and near-normalization of ALT^[42]. Quantitative phlebotomy also leads to improvement in insulin sensitivity in a group of subjects with high-ferritin type 2 diabetes^[43]. Therefore, iron could be the culprit of these conditions.

The current sample set did provide enough information to distinguish type 1 and type 2 diabetes. However, as 95% of diabetes is type 2 diabetes and insulin resistance is a cardinal feature of type 2 diabetes, the current study is most applicable to type 2 diabetes. Our observations are consistent with the published results^[44-46] with some new clinical implications. In this study, even without clinical evidence of iron overload, iron could be associated with liver damage and insulin resistance. A clinical trial of quantitative phlebotomy in the subjects with elevated ferritin concentration is warranted to test this hypothesis. Once it is demonstrated, quantitative phlebotomy could be recommended for those patients with DM or insulin resistance, who also have elevated serum ferritin concentration. Although the underlying molecular mechanism of the association remains to be elucidated, our observations imply that the liver could play a role in iron-associated insulin resistance.

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COMMENTS

Background

Liver has been noted to play a role in the pathogenesis of type 2 diabetes. Hemochromatosis and excess iron load has been implicated to play a role in the pathogenesis of type 2 diabetes. Iron metabolism could a role in the pathogenesis of type 2 diabetes.

Research frontiers

Diabetes mellitus is a common manifestation of hemochromatosis. Although the common mutation of *HFE* is not associated with type 2 diabetes, the iron metabolism could play a role in the pathogenesis of diabetes. In hemochromatosis, both insulin resistance and impaired insulin secretion have

been suggested to play a role in its pathogenesis. However, the underlying mechanism of iron-associated abnormal glucose homeostasis in the general population is not well understood.

Innovations and breakthroughs

In this study, the authors found that subjects with diabetes had a higher serum ferritin concentration than those without diabetes. The authors also observed that serum ferritin concentration was negatively associated with insulin sensitivity (%S and QUICKI), but not with beta cell function. Therefore, a high serum ferritin concentration is associated with insulin resistance and is a risk factor for diabetes mellitus. The authors further demonstrated that the inflammatory hypothesis is not likely the underlying mechanism of the reported associations in this study. In addition, from the observed associations of alanine aminotransferase (ALT) and gamma glutamyl transpeptidase with serum ferritin concentration and insulin sensitivity, the authors provided the evidence suggesting that the role of liver in the pathogenesis of iron-associated insulin resistance. Iron-induced oxidant stress has proposed to play a key role in iron-mediated tissue damage and iron-induced oxidative stress with mitochondrial dysfunction could be one of the underlying mechanisms in these metabolic disorders.

Applications

As in patients with clinical evidence of non-alcoholic fatty liver disease, quantitative phlebotomy induced iron depletion to a level of near-iron deficiency results in a 40%-55% improvement of both fasting and glucose-stimulated plasma insulin concentrations and near-normalization of ALT and quantitative phlebotomy also leads to improvement in insulin sensitivity in a group of subjects with high-ferritin type 2 diabetes, a clinical trial of quantitative phlebotomy in the subjects with elevated ferritin concentration is warranted to test this hypothesis. Once it is demonstrated, quantitative phlebotomy could be recommended for those patients with diabetes mellitus or insulin resistance, who also have elevated serum ferritin concentration.

Terminology

Ferritin: Ferritin is a protein in the body that binds to iron; most of the iron stored in the body is bound to ferritin. In humans, it acts as a buffer against iron deficiency and iron overload. Serum ferritin concentration closely correlates with total body iron stores, mainly in the liver.

Peer-review

This is an interesting study. The authors have evaluated the relationship of iron with diabetes mellitus (DM) and concluded that disordered iron metabolism could play a role in the pathogenesis of insulin resistance and DM through its effect on liver function.

REFERENCES

- 1 **Witte DL**, Crosby WH, Edwards CQ, Fairbanks VF, Mitros FA. Practice guideline development task force of the College of American Pathologists. Hereditary hemochromatosis. *Clin Chim Acta* 1996; **245**: 139-200 [PMID: 8867884 DOI: 10.1016/0009-8981(95)06212-2]
- 2 **Feder JN**, Penny DM, Irrinki A, Lee VK, Lebrón JA, Watson N, Tsuchihashi Z, Sigal E, Bjorkman PJ, Schatzman RC. The hemochromatosis gene product complexes with the transferrin receptor and lowers its affinity for ligand binding. *Proc Natl Acad Sci USA* 1998; **95**: 1472-1477 [PMID: 9465039 DOI: 10.1073/pnas.95.4.1472]
- 3 **Halsall DJ**, McFarlane I, Luan J, Cox TM, Wareham NJ. Typical type 2 diabetes mellitus and HFE gene mutations: a population-based case - control study. *Hum Mol Genet* 2003; **12**: 1361-1365 [PMID: 12783844 DOI: 10.1093/hmg/ddg149]
- 4 **Ford ES**, Cogswell ME. Diabetes and serum ferritin concentration among U.S. adults. *Diabetes Care* 1999; **22**: 1978-1983 [PMID: 10587829 DOI: 10.2337/diacare.22.12.1978]
- 5 **Haap M**, Fritsche A, Mensing HJ, Häring HU, Stumvoll M. Association of high serum ferritin concentration with glucose intolerance and insulin resistance in healthy people. *Ann Intern Med* 2003; **139**: 869-871 [PMID: 14623634 DOI: 10.7326/0003-4819-139

- 9-10-200311180-00029]
- 6 **Jiang R**, Manson JE, Meigs JB, Ma J, Rifai N, Hu FB. Body iron stores in relation to risk of type 2 diabetes in apparently healthy women. *JAMA* 2004; **291**: 711-717 [PMID: 14871914 DOI: 10.1001/jama.291.6.711]
 - 7 **Tuomainen TP**, Nyssönen K, Salonen R, Tervahauta A, Korpela H, Lakka T, Kaplan GA, Salonen JT. Body iron stores are associated with serum insulin and blood glucose concentrations. Population study in 1,013 eastern Finnish men. *Diabetes Care* 1997; **20**: 426-428 [PMID: 9051399 DOI: 10.2337/diacare.20.3.426]
 - 8 **Qi L**, Meigs J, Manson JE, Ma J, Hunter D, Rifai N, Hu FB. HFE genetic variability, body iron stores, and the risk of type 2 diabetes in U.S. women. *Diabetes* 2005; **54**: 3567-3572 [PMID: 16306377 DOI: 10.2337/diabetes.54.12.3567]
 - 9 **Yaouanq JM**. Diabetes and haemochromatosis: current concepts, management and prevention. *Diabete Metab* 1995; **21**: 319-329 [PMID: 8586148]
 - 10 **Cario H**, Holl RW, Debatin KM, Kohne E. Insulin sensitivity and beta-cell secretion in thalassaemia major with secondary haemochromatosis: assessment by oral glucose tolerance test. *Eur J Pediatr* 2003; **162**: 139-146 [PMID: 12655415]
 - 11 **Furutani M**, Nakashima T, Sumida Y, Hirohama A, Yoh T, Kakisaka Y, Mitsuyoshi H, Senmaru H, Okanoue T. Insulin resistance/beta-cell function and serum ferritin level in non-diabetic patients with hepatitis C virus infection. *Liver Int* 2003; **23**: 294-299 [PMID: 12895270 DOI: 10.1034/j.1600-0676.2003.00841.x]
 - 12 **Hsiao TJ**, Chen JC, Wang JD. Insulin resistance and ferritin as major determinants of nonalcoholic fatty liver disease in apparently healthy obese patients. *Int J Obes Relat Metab Disord* 2004; **28**: 167-172 [PMID: 14610526]
 - 13 Plan and operation of the Third National Health and Nutrition Examination Survey, 1988-94. Series 1: programs and collection procedures. *Vital Health Stat 1* 1994; **(32)**: 1-407 [PMID: 7975354]
 - 14 **American Diabetes Association**. Diagnosis and classification of diabetes mellitus. *Diabetes Care* 2004; **27** Suppl 1: S5-S10 [PMID: 14693921 DOI: 10.2337/diacare.27.2007.S5]
 - 15 **Matthews DR**, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; **28**: 412-419 [PMID: 3899825 DOI: 10.1007/bf00280883]
 - 16 **Pietrangelo A**. Hereditary hemochromatosis--a new look at an old disease. *N Engl J Med* 2004; **350**: 2383-2397 [PMID: 15175440 DOI: 10.1056/NEJMra031573]
 - 17 **McLaren CE**, McLachlan GJ, Halliday JW, Webb SI, Leggett BA, Jazwinska EC, Crawford DH, Gordeuk VR, McLaren GD, Powell LW. Distribution of transferrin saturation in an Australian population: relevance to the early diagnosis of hemochromatosis. *Gastroenterology* 1998; **114**: 543-549 [PMID: 9496946 DOI: 10.1016/s0016-5085(98)70538-4]
 - 18 **Barr RG**, Nathan DM, Meigs JB, Singer DE. Tests of glycemia for the diagnosis of type 2 diabetes mellitus. *Ann Intern Med* 2002; **137**: 263-272 [PMID: 12186517 DOI: 10.7326/0003-4819-137-4-200208200-00011]
 - 19 **Levy JC**, Matthews DR, Hermans MP. Correct homeostasis model assessment (HOMA) evaluation uses the computer program. *Diabetes Care* 1998; **21**: 2191-2192 [PMID: 9839117 DOI: 10.2337/diacare.21.12.2191]
 - 20 **Bonora E**, Targher G, Alberiche M, Bonadonna RC, Saggiani F, Zenere MB, Monauni T, Muggeo M. Homeostasis model assessment closely mirrors the glucose clamp technique in the assessment of insulin sensitivity: studies in subjects with various degrees of glucose tolerance and insulin sensitivity. *Diabetes Care* 2000; **23**: 57-63 [PMID: 10857969 DOI: 10.2337/diacare.23.1.57]
 - 21 **Hermans MP**, Levy JC, Morris RJ, Turner RC. Comparison of insulin sensitivity tests across a range of glucose tolerance from normal to diabetes. *Diabetologia* 1999; **42**: 678-687 [PMID: 10382587 DOI: 10.1007/s001250051215]
 - 22 **Hermans MP**, Levy JC, Morris RJ, Turner RC. Comparison of tests of beta-cell function across a range of glucose tolerance from normal to diabetes. *Diabetes* 1999; **48**: 1779-1786 [PMID: 10480608 DOI: 10.2337/diabetes.48.9.1779]
 - 23 **Katz A**, Nambi SS, Mather K, Baron AD, Follmann DA, Sullivan G, Quon MJ. Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans. *J Clin Endocrinol Metab* 2000; **85**: 2402-2410 [PMID: 10902785 DOI: 10.1210/jcem.85.7.6661]
 - 24 **Clark JM**, Brancati FL, Diehl AM. The prevalence and etiology of elevated aminotransferase levels in the United States. *Am J Gastroenterol* 2003; **98**: 960-967 [PMID: 12809815 DOI: 10.1111/j.1572-0241.2003.07486.x]
 - 25 **Cook JD**. Clinical evaluation of iron deficiency. *Semin Hematol* 1982; **19**: 6-18 [PMID: 6763340]
 - 26 **Hulthén L**, Lindstedt G, Lundberg PA, Hallberg L. Effect of a mild infection on serum ferritin concentration--clinical and epidemiological implications. *Eur J Clin Nutr* 1998; **52**: 376-379 [PMID: 9630391 DOI: 10.1038/sj.ejcn.1600573]
 - 27 **Pepys MB**, Hirschfield GM. C-reactive protein: a critical update. *J Clin Invest* 2003; **111**: 1805-1812 [PMID: 12813013 DOI: 10.1172/jci18921]
 - 28 **Shine B**, de Beer FC, Pepys MB. Solid phase radioimmunoassays for human C-reactive protein. *Clin Chim Acta* 1981; **117**: 13-23 [PMID: 7333010 DOI: 10.1016/0009-8981(81)90005-x]
 - 29 **Cherrington AD**. Banting Lecture 1997. Control of glucose uptake and release by the liver in vivo. *Diabetes* 1999; **48**: 1198-1214 [PMID: 10331429 DOI: 10.2337/diabetes.48.5.1198]
 - 30 **DeFronzo RA**. Lilly lecture 1987. The triumvirate: beta-cell, muscle, liver. A collusion responsible for NIDDM. *Diabetes* 1988; **37**: 667-687 [PMID: 3289989 DOI: 10.2337/diab.37.6.667]
 - 31 **Hanley AJ**, Williams K, Festa A, Wagenknecht LE, D'Agostino RB, Kempf J, Zinman B, Haffner SM. Elevations in markers of liver injury and risk of type 2 diabetes: the insulin resistance atherosclerosis study. *Diabetes* 2004; **53**: 2623-2632 [PMID: 15448093 DOI: 10.2337/diabetes.53.10.2623]
 - 32 **Vozarova B**, Stefan N, Lindsay RS, Saremi A, Pratley RE, Bogardus C, Tataranni PA. High alanine aminotransferase is associated with decreased hepatic insulin sensitivity and predicts the development of type 2 diabetes. *Diabetes* 2002; **51**: 1889-1895 [PMID: 12031978 DOI: 10.2337/diabetes.51.6.1889]
 - 33 **Matsuda M**, DeFronzo RA. Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care* 1999; **22**: 1462-1470 [PMID: 10480510 DOI: 10.2337/diacare.22.9.1462]
 - 34 **Jehn M**, Clark JM, Guallar E. Serum ferritin and risk of the metabolic syndrome in U.S. adults. *Diabetes Care* 2004; **27**: 2422-2428 [PMID: 15451911 DOI: 10.2337/diacare.27.10.2422]
 - 35 **Chen J**, Wildman RP, Hamm LL, Muntner P, Reynolds K, Whelton PK, He J. Association between inflammation and insulin resistance in U.S. nondiabetic adults: results from the Third National Health and Nutrition Examination Survey. *Diabetes Care* 2004; **27**: 2960-2965 [PMID: 15562214 DOI: 10.2337/diacare.27.12.2960]
 - 36 **Reddy MB**, Clark L. Iron, oxidative stress, and disease risk. *Nutr Rev* 2004; **62**: 120-124 [PMID: 15098859 DOI: 10.1301/nr.2004.mar.120-124]
 - 37 **Pietrangelo A**. Iron-induced oxidant stress in alcoholic liver fibrogenesis. *Alcohol* 2003; **30**: 121-129 [PMID: 12957296 DOI: 10.1016/s0741-8329(03)00126-5]
 - 38 **Tuomainen TP**, Diczfalusy U, Kaikkonen J, Nyssönen K, Salonen JT. Serum ferritin concentration is associated with plasma levels of cholesterol oxidation products in man. *Free Radic Biol Med* 2003; **35**: 922-928 [PMID: 14556856 DOI: 10.1016/s0891-5849(03)00433-7]
 - 39 **Eaton JW**, Qian M. Molecular bases of cellular iron toxicity. *Free Radic Biol Med* 2002; **32**: 833-840 [PMID: 11978485 DOI: 10.1016/s0891-5849(02)00772-4]
 - 40 **Petersen KF**, Dufour S, Befroy D, Garcia R, Shulman GI. Impaired mitochondrial activity in the insulin-resistant offspring of patients with type 2 diabetes. *N Engl J Med* 2004; **350**: 664-671 [PMID: 14960743 DOI: 10.1056/NEJMoa031314]
 - 41 **Wilson FH**, Hariri A, Farhi A, Zhao H, Petersen KF, Toka HR,

- Nelson-Williams C, Raja KM, Kashgarian M, Shulman GI, Scheinman SJ, Lifton RP. A cluster of metabolic defects caused by mutation in a mitochondrial tRNA. *Science* 2004; **306**: 1190-1194 [PMID: 15498972 DOI: 10.1126/science.1102521]
- 42 **Facchini FS**, Hua NW, Stoohs RA. Effect of iron depletion in carbohydrate-intolerant patients with clinical evidence of nonalcoholic fatty liver disease. *Gastroenterology* 2002; **122**: 931-939 [PMID: 11910345 DOI: 10.1053/gast.2002.32403]
- 43 **Fernández-Real JM**, Peñarroja G, Castro A, García-Bragado F, Hernández-Aguado I, Ricart W. Blood letting in high-ferritin type 2 diabetes: effects on insulin sensitivity and beta-cell function. *Diabetes* 2002; **51**: 1000-1004 [PMID: 11916918 DOI: 10.2337/diabetes.51.4.1000]
- 44 **Shi Z**, Hu X, Yuan B, Pan X, Meyer HE, Holmboe-Ottesen G. Association between serum ferritin, hemoglobin, iron intake, and diabetes in adults in Jiangsu, China. *Diabetes Care* 2006; **29**: 1878-1883 [PMID: 16873796 DOI: 10.2337/dc06-0327]
- 45 **Fumeron F**, Péan F, Driss F, Balkau B, Tichet J, Marre M, Grandchamp B. Ferritin and transferrin are both predictive of the onset of hyperglycemia in men and women over 3 years: the data from an epidemiological study on the Insulin Resistance Syndrome (DESIR) study. *Diabetes Care* 2006; **29**: 2090-2094 [PMID: 16936158 DOI: 10.2337/dc06-0093]
- 46 **Choi KM**, Lee KW, Kim HY, Seo JA, Kim SG, Kim NH, Choi DS, Baik SH. Association among serum ferritin, alanine aminotransferase levels, and metabolic syndrome in Korean postmenopausal women. *Metabolism* 2005; **54**: 1510-1514 [PMID: 16253641 DOI: 10.1016/j.metabol.2005.05.018]

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How reliable is online diffusion of medical information targeting patients and families?

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Abstract

AIM: To determine whether online diffusion of the "Ten Warning Signs of Primary Immunodeficiency Diseases (PID)" adheres to accepted scientific standards.

METHODS: We analyzed how reproducible is online diffusion of a unique instrument, the "Ten Warning Signs of PID", created by the Jeffrey Modell Foundation (JMF), by Google-assisted searches among highly visited sites from professional, academic and scientific organizations; governmental agencies; and patient support/advocacy organizations. We examined the diffusion, consistency of use and adequate referencing of this instrument. Where applicable, variant versions of the instrument were examined for changes in factual content that would have practical impact on physicians or on patients and their families.

RESULTS: Among the first 100 sites identified by Google search, 85 faithfully reproduced the JMF model, and correctly referenced to its source. By contrast, the other 15 also referenced the JMF source but presented one or more changes in content relative to their purported model and therefore represent uncontrolled variants, of unknown origin. Discrepancies identified in the latter included changes in factual content of the original JMF list (C), as well as removal (R) and introduction (I) of novel signs (Table 2), all made without reference to any scientific publications that might account for the drastic changes in factual content. Factual changes include changes in

the number of infectious episodes considered necessary to raise suspicion of PID, as well as the inclusion of various medical conditions not mentioned in the original. Together, these changes will affect the way physicians use the instrument to consult or to inform patients, and the way patients and families think about the need for specialist consultation in view of a possible PID diagnosis.

CONCLUSION: The retrieved adaptations and variants, which significantly depart from the original instrument, raise concerns about standards for scientific information provided online to physicians, patients and families.

Key words: Information technology and human health; Expert consultation online; Online medical information; Warning signs; Infection; Diagnosis

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Core tip: We analyzed how reproducible is online diffusion of a unique instrument, the "Ten Warning Signs of Primary Immunodeficiency Diseases", to define whether information made available to physicians, families and support/advocacy groups through the internet adheres to accepted scientific standards. The results show that this instrument is diffused through many sites, but the actual scientific contents often depart substantially from their purported model. This raises concerns about the quality of scientific information provided online on medical matters, and on the need for corrective mechanisms in an age when the public is increasingly dependent on the internet as primary source of knowledge.

Xavier-Elsas P, Bastos SE, Gaspar-Elsas MIC. How reliable is online diffusion of medical information targeting patients and families? *World J Exp Med* 2015; 5(4): 244-250 Available from: URL: <http://www.wjgnet.com/2220-315X/full/v5/i4/244.htm> DOI: <http://dx.doi.org/10.5493/wjem.v5.i4.244>

INTRODUCTION

The personal computer and access to the Internet have affected medicine as rarely seen with previous technological advances, due to the large-scale inclusion of patients, families and support/advocacy groups as a highly motivated audience. Emerging online resources for communication, diffusion and education in the medical sciences profoundly changed the way physicians and patients view the possible diagnosis of uncommon, chronic disease, and the prospects for long and complex evaluation procedures. The Internet is now a major source of information on all aspects of medical science, which is consulted routinely by doctors worldwide. Many laymen also consult the Internet about rare, incapacitating diseases, and make

decisions on the basis of factual information retrieved through Google and similar search tools. For families, this often means deciding for, or against, further search for specialty physicians and reference centers. Hence, online information must be consistent in itself, and based on best standards for scientific evidence, both in its initial presentation and in subsequent revised versions (whenever these are prompted by advances in medical knowledge). Deviance from this course will likely increase the risk of misinforming the public, entailing unpredictable risks for patients and their families.

The field of primary immunodeficiency diseases (PID) is a rapidly evolving area of medical science in which the advantages of diffusing medical information online are easy to grasp. PID^[1] are a group of chronic, complex diseases^[2] which poses a unique public health problem, because: (1) as a group, they are rare, but present worldwide; (2) they are biologically complex and very heterogeneous^[3,4], but nevertheless share a set of major outcomes (increased susceptibility to repeated and/or opportunistic infections, autoimmunity and malignancy) which predispose to increased mortality and/or severe complications and sequelae^[3]; (3) they arise in congenital defects of the immune system, which often lead to clinical deficiencies in infancy and childhood, and unless corrected through costly and technically complex procedures, may have a major impact on the growth and development, as well as socialization and schooling of the child^[1-4]; and (4) they have a genetic basis that accounts for family clustering of cases, thereby making accurate diagnosis and increased awareness among health professionals indispensable for adequate family counseling^[1-4].

In short, although PID patients must be managed by highly specialized professional teams, they mostly come from environments which lack specialists, and further lack the information about PID that would prompt families and primary care physicians to seek for specialists' counsel. Together, this creates the need for mechanisms to bring together the demographically rare, geographically dispersed and clinically heterogeneous patient population, on the one hand, and the institutionally rare, geographically concentrated and technologically demanding specialist population, on the other hand. This task can be made easier by letting the patients, their families and their primary care physicians know that they need the type of specialized care which is not available everywhere, but at specific tertiary institutions to which they can be referred. This challenge has been met by the Jeffrey Modell Foundation (JMF) through the elaboration of a list of Ten Warning Signs of Primary Immunodeficiency (hereafter referred to as "the JMF list"), to be diffused among patients and their families, primary care physicians and the general public, to raise awareness that certain clinical manifestations reflect an underlying PID that needs diagnostic workout. The creation of the JMF list has merit in itself, but its usefulness depends on whether it is scientifically accurate and correctly referenced, wherever it is found

online. Because this well-structured and authoritative text provides an ideal probe to examine whether medical information can be propagated online risk-free, we addressed here the online diffusion of the scientific information it embodies, as well as the emergence of variants that may appear in this process. The results show that the “Ten warning signs of primary immune deficiency” are quite suitable for an object-lesson on the fate of scientific information in the Cyberage.

MATERIALS AND METHODS

Study design

Initial evaluation of the frequency, extent and types of variation among online sites in the contents and referencing of the JMF list was carried out in the 100 most accessed web pages identified by Google with search term Ten warning signs of Primary Immune Deficiency (search date April 23, 2012). This was done to approximate as much as possible the most frequent situation, namely that in which someone tries, for the first time, to learn something about the Ten Warning Signs of PID online. To further assess the diffusion, accuracy of citation and consistency of use of the JMF list online, we examined in detail sites from: (1) professional, academic and scientific organizations, including, but not limited to, International Union of Immunological Societies, European Society of Immunodeficiencies, Sociedade Brasileira de Imunologia, Associação Brasileira de Alergia e Imunopatologia; (2) governmental agencies, including CDC and NIH; and (3) patient support organizations, including JMF and BRAGID (Grupo Brasileiro de Imunodeficiência).

Definition of terms

For the purposes of this study: (1) the concept is defined here as any list of Warning signs of PID, to be diffused among the nonspecialist public, regardless of whether it derives from the original JMF list; (2) the application of this Concept refers to the creation and use of any list of Warning signs of PID, which could be retrieved in our search; (3) referencing refers to the acknowledgement, by any given site applying the Concept, that it is aware of the JMF site (usually embodied in a link to the JMF site), or that JMF is considered a valid source of information on PID, regardless of whether the Concept as development in the site are actually related to the JMF list (*i.e.*, correct referencing) or not (incorrect referencing). Referencing, correct or incorrect, demonstrates that the site applying the Concept derives a measure of scientific credibility from acknowledging JMF as the referred source; and (4) variation refers to any departure in structure, language, or factual content (including quantitative information), from the original JMF list, which is accessible at the JMF site (www.jmfworld.com).

On the other hand, any variant list retrievable by the search tool could also originate in a completely independent source, unrelated to JMF, and in this

case would represent an independent application of the Concept. If such a hypothetical alternative source were properly referenced, the application would not be included in our study as an undue variation of the JMF model. In our study, however (see results and discussion), both preserved and variant versions of the JMF list, some of the latter strikingly different from the JMF original, were all referenced to the JMF site.

Identification and classification of variants

Variation was considered significant if: (1) information contained in the original was removed (R), or changed (C) in factual/quantitative content so as to be of practical consequence; (2) information not contained in the original was introduced (I); and (3) information contained in the original was rearranged (displaced in its original context), or generalized (absorbed into a more comprehensive class not explicitly mentioned in the original) so as to change (C) the interpretation of the conveyed message. Deletion or addition of signs represents extreme examples of removal and introduction, respectively. A significant variant may differ from the standard in one or more signs, and therefore one significant change is sufficient to define a variant. On the other hand, preservation (P) of the original content is defined as complete identity of content between any given variant and the JMF list, and therefore precludes any significant change of content. Accordingly, less significant, or nonsignificant, variation, such as a change in language that did not affect the factual information, or a change in the order in which the warning signs were listed, was also found in our study, but was not further discussed below, to help us focus on discussing consequential issues.

The frequency of variation is defined as the ratio between the number of variant versions and the number of total versions sampled. The extent of variation (*i.e.*, the degree to which any given version differs from the purported original) found, as well as the types of variation detected (removal, introduction, change of content), can be found in the accompanying lists of sites which faithfully reproduce the JMF list (Table 1) and of variations found in those that do not (Table 2). For the extreme forms of introduction, which result in addition of novel signs, amounting to a drastic change in content, we further recorded the number of novel signs added to the original JMF material.

RESULTS

Preservation/change of the original message among highly accessed pages. Although numerous online information sources nominally refer to the JMF list, not every site consulted shows complete P of JMF original contents. The majority of pages faithfully reproduce the original information (Table 1), and are representative of adequate online sources of medical information (out of 48, 10 were from pharmaceutical companies; 21 from patient support groups focusing on severe and

Table 1 Identification of the sites with complete preservation of contents relative to the Jeffrey Modell Foundation list among the top 100 accessed by Google

<p> www.jmfworld.com/ http://www.info4pi.org http://www.pia.org.uk/publications/10_signs_of_pia http://www.nichd.nih.gov/health/topics/primary_immunodeficiency.cfm www.cisociety.com/files/what-is-PID.html www.hizenra.com/docs/Hizenra_10_warning_signs_PIDD.pdf http://www.justmommies.com/forums/f325-choosing-not-to-vaccinate/2004706-10-warning-signs-primary-immunodeficiency-psa-conjunction-cdc.html http://pediatrics.about.com/od/primaryimmunodeficiency/Primary_Immunodeficiency.htm https://www.mygardian.com/gardian/.../resources.html; http://www.allergyclinic.co.nz/guides/74.html http://smr.newswire.ca/en/canadian-immunodeficiency-society/april-29-world-primary-immunodeficiency-day http://www.gammagardliquid.com/professional-resources/order-tools-and-resources.html http://www.treatingpi.com/primary-immunodeficiency-faq.aspx http://www.uhhospitals.org/rainbowchildren/ourservices/allergyimmunology/tabid/3016/patientresources.aspx http://smr.newswire.ca/en/canadian-immunodeficiency-society/april-29-world-primary-immunodeficiency-day http://www.justmommies.com/forums/f325-choosing-not-to-vaccinate/2004706-10-warning-signs-primary-immunodeficiency-psa-conjunction-cdc.html http://www.baxterbiotherapeutics.com/us/us_t_immunoglobulin.html https://webforms.baxterbioscience.com/immune/order.jspa http://allergistmommy.blogspot.com/2011/07/always-sick-could-it-be-immune.html http://journals.lww.com/pidj/Citation/2011/10000/Clinical_Features_That_Identify_Children_With.6.aspx https://www.wellpartner.com/SpecialtyRxFaqs.aspx www.uhhospitals.org/.../immunodeficiency.pdf http://www.chw.org/display/PPF/DocID/36622/router.asp http://www.biorxiv.net/brochures/10-Warning-Signs-of-PI.pdf http://healthlibrary.stanford.edu/resources/bodysystems/immune_pid.html http://www.pia.org.uk/gp_info.html http://allergycases.org/2009/08/primary-immunodeficiency-disorders-pidd.html http://www.savanna.k12.ok.us/Nurse/illustrated_poster_eng%20illustrated.pdf http://www.isitpid.com/news_detail.cfm?e=7 http://ainotes.wikispaces.com/Immunodeficiency+Warning+Signs http://nihrecord.od.nih.gov/newsletters/10_31_2000/story02.htm http://www.allergyclinic.co.nz/whatsnew.html http://www.babycenter.com/204_most-warning-signs-for-kids-8217-immune-disorders-are-off-ba_10349792.bc http://www.gammaked.com/media/10_Warning_Signs_of_PI.pdf http://www.disabled-world.com/disability/awareness/primary-immunodeficiency.php https://plus.google.com/110859855629071891085/posts/5ijl3nVYayp http://www.ipopi.org/uploads/JMF%20Advocacy%20Toolkit_2011.pdf http://books.google.com.br/books?id=H7GVhb27mo4C&pg=PA42&lpg=PA42&dq=10+Warning+Signs+of+Primary+Immunodeficiency&source=bl&ts=pz0rX9QK2E&sig=wKp2GGabmE0g7q_GRYB-vpwTt3g&hl=pt-BR&ei=iqOLTqCCBuXm0QGUrVX6BA&sa=X&oi=book_result&ct=result&resnum=5&ved=0CDgQ6AEwBDhQ#v=onepage&q=10%20Warning%20Signs%20of%20Primary%20Immunodeficiency&f=false http://ezinearticles.com/comment.php?10-Warning-Signs-of-Primary-Immunodeficiency&id=1297957 http://dallasallergy.net/services/immunodeficiency https://webforms.baxterbioscience.com/ggliquid/orderNew.jspa http://www.doaj.org/doaj?func=abstract&id=467945 http://www.rare-diseases.eu/2010/IMG/Media/Czerniawska-PID%20Poster_Jeffrey%20Modell%20Foundation.pdf </p>
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chronic conditions; 4 mirrored the JMF site; 5 were from hospitals; 1 from an elementary school; 2 from NIH; 1 from an university; 1 from a scientific society; 3 reviewed a book on PID). Nevertheless, we found a sizeable minority (15 out of the 100 most highly accessed sites identified by Google) that displayed lists departing substantially from their purported model (Table 2). Discrepancies identified in the latter included changes in factual content of the original JMF list (C), as well as removal (R) and introduction (I) of novel signs (Table 2), all made without reference to any scientific publications that might account for the drastic changes in factual content.

Incorrect referencing in variant pages

Importantly, 14 of these 15 variant lists directly refer to the original JMF (direct links), or to sites with links

to JMF (indirect links), as their source, and therefore represents undisputable examples of incorrect referencing, an unacceptable practice by current standards of scientific communication. Importantly, the variant versions include some sponsored by professional societies, governmental regulatory and research agencies, and respected biomedical research institutes, which thereby lend authority to an online resource that fails to meet standards of citation and referencing.

Major departures from the JMF list

Among the top 100 pages identified by Google, 3 formed a set with similar content, which departed strikingly from the JMF list. This included 1 page associated with the Mayo Clinic (<http://www.mayoclinic.com/health/primary-immunodeficiency/DS01006>), and 2 pages associated with www.livestrong.com (www.livestrong.com).

Table 2 Variant versions of the Jeffrey Modell Foundation list in the first 100 pages retrieved by Google

Site consulted	10 warning signs of primary immunodeficiency of the Jeffrey Modell Foundation										I		JMF	
	#1	#2	#3	#4	#5	#6	#7	#8	#9	#10	Y/N	n	Direct link ¹	Indirect link
a	C	P	P	P	P	P	C	P	C	P	N	0	N	N
b	C	P	P	R	P	P	C	P	C	P	N	0	Y	N
c	C	C	C	R	P	C	R	R	R	C	Y	4	N	Y
d	C	P	P	P	P	P	C	P	C	P	N	0	Y	N
e	C	P	P	P	P	P	C	P	C	P	N	0	Y	N
f	C	P	P	P	P	P	C	P	C	P	N	0	Y	N
g	C	C	P	P	P	P	C	R	C	P	Y	1	N	Y
h	C	P	R	P	P	P	C	P	C	P	Y	4	Y	N
i	C	P	P	P	P	P	C	P	C	P	N	0	N	Y
j	C	P	R	P	P	P	C	P	R	P	Y	4	Y	N
k	C	P	P	P	P	P	C	P	C	P	N	0	Y	N
l	C	P	P	P	P	P	C	P	C	P	N	0	N	Y
m	C	P	P	P	P	P	C	P	C	P	N	0	Y	N
n	C	P	P	P	P	P	C	P	C	P	N	0	N	Y
o	C	P	P	P	P	P	C	P	C	P	N	0	Y	N

¹<http://www.info4pi.org/aboutPI/index.cfm?section=aboutPI&content=warningsigns>; ²<http://www.stopgettingssick.com/template.cfm-1685>; ³<http://www.aafp.org/afp/2003/1115/p2011.html>; ⁴<http://www.livestrong.com/article/71656-top-ten-warning-signs-primary/>; ⁵<http://docphyl.com/index.php?x=pages/pimmunodeficiency>; ⁶<http://ezinearticles.com/?10-Warning-Signs-of-Primary-Immunodeficiency&id=1297957>; ⁷http://www.momlogic.com/resources/primary_immunodeficiency_diseases.php; ⁸<http://www.irishhealth.com/article.html?id=3927>; ⁹http://www.aefat.es/docs/inmunodefprim_aafp.pdf; ¹⁰www.bpl.co.uk/...primary-immunodeficiency/39; ¹¹<http://www.aafp.org/afp/2003/1115/p2001.html>; ¹²<http://www.insidetoronto.com/insidetoronto/article/57439>; ¹³<http://www.allergyconsumerreview.com/sinus-infection.html>; ¹⁴<http://www.aacijournal.com/content/7/S1/S11>; ¹⁵thegunnybag.files.wordpress.com/.../primary-im; ¹⁶http://www.nichd.nih.gov/health/topics/primary_immunodeficiency.cfm. P: Preserved; R: Removed; I: Introduced; C: Changed content; n: Number of novel signs introduced. Search verified 04/23/2012.

livestrong.com/article/71656-top-ten-warning-signs-primary/; and www.livestrong.com/article/253355-10-signs-of-primary-immunodeficiency/), which presents itself as based on the Mayo Clinic. In this particular example, the key message was profoundly altered, but presented no support of scientific evidence to justify these changes. It also included a series of explanatory notes in language accessible to the nonmedical public, which represents a further departure from the language in which the JMF original is cast, of obvious consequence for the impact on families and patients, regardless of whether it is scientifically accurate. This specific example, which is of undeniable relevance, given the high number of accesses, as well as the high scientific profile of the sponsoring institution, and the wider audience presumably intended, illustrates the hazards of uncontrolled diffusion of medical information online.

Variation is not unsequential

Scientific information was changed in some variant versions so as to have the potential for undesirable consequences in medical practice. A clear example follows: The "Four or more new ear infections within 1 year" of the JMF list were changed into "Eight or more new ear infections within 1 year" in several sites (The American Academy of Family Physicians; University of Wisconsin; the <http://www.stopgettingssick.com/template.cfm-1685> patient support site; The Australasian Society of Clinical Immunology and Allergy/ASCIA). In the Mayo Clinic-related sites mentioned above, the same original content was generalized to "Frequent/recurrent infections". In the first case, a

patient who would previously be considered worthy of evaluation for PID (according to the JMF list) must now suffer twice as many ear infections before qualifying as a candidate for PID evaluation. In the second case, the criteria became so broad as to lose their usefulness, for the definition would equally well apply to the numerous people living in conditions of poor hygiene and sanitation worldwide, who have no PID. The fate of the patient will therefore hinge on which version of the list has been consulted by the physician, an issue with broader implications for the rights of patients to appropriate medical attention.

A further example of generalization is provided by the Mayo Clinic, which introduced in their list of warning signs the following: inflammation and infection in internal organs, including the liver; autoimmune disorders, such as Lupus erythematosus, arthritis or type I diabetes; hematological disorders, such as low platelet counts and anemias; digestive problems, including cramps, loss of appetite, nausea and diarrhea; and delayed growth and development. Considering the high frequency with which any of the preceding occurs in the absence of PID, it is open to question whether such comprehensive enumeration of "warning signs for PID" will benefit patients and improve the physician's capabilities to deal with PID.

Local adaptations - is PID the same everywhere?

A different, but related, finding points to the adaptation of the original model to regional peculiarities. A very good example of adaptation is offered by BRAGID (Brazilian Group for Immune Deficiency), a patient-support organization. Although the BRAGID site (www.

imunopediatria.org.br) is not within the top 100 sites identified by Google, and therefore was not included in Table 2, it is here analyzed in detail because it plays a very important role in Brazil, where other sources of information on PID are scarce. The list accessed through BRAGID includes: "repeated intestinal infections, chronic diarrhea, severe asthma, collagen (autoimmune) diseases, adverse effects of BCG (tuberculosis) vaccination, mycobacterial infections, and a clinical phenotype suggestive of immune deficiency" (an odd item in a list directed to a public supposed to know little about immune deficiency and even less about the associated clinical phenotypes).

This adaptation likely reflects an effort to increase the usefulness of the concept to a developing country, where exposure to the mycobacteria that causes tuberculosis remains very important. The reference to adverse effects of BCG vaccination is equally appropriate to this context, since this is not a universally enforced practice, but is very relevant to Brazil as well as to PID patients^[5-7]. However, the spectrum of clinical conditions enumerated includes repeated intestinal infections, chronic diarrhea, severe asthma and autoimmune diseases. This is less likely to help, as in a developing country such as Brazil infectious pathogens and environmental pollutants are major contributing factors for intestinal infection, chronic diarrhea, severe asthma, and even autoimmune manifestations in endemic diseases, which far outweigh their possible contribution to PID detection and management. As an effort of regional adaptation is to be commended, when justified by local factors, one should never omit the fact that this is an evolution from the original concept of signs of PID; by consequence, referencing to a well-established model without recording that it is a modification counters standard practice.

DISCUSSION

Anonymous expertise in an age of evidence-based medicine

It is surprising that factual content that informs medical decisions and medical education could undergo such uncontrolled variation online, especially in an age that stresses the importance of rigorous scientific documentation for sound medical practice. The fate of the JMF list online should raise concern in clinicians and scientists alike, because this unexplained variance in factual content counters basic rules in scientific communication, as significant changes in content were made without justification or references, and no responsibility was taken for them. Through incorrect referencing to the original from which they depart, variant lists are further misleading both physicians and patients as to the scientific basis of their content, and evading accountability for the possible untoward consequences of disseminating inaccurate information. Such attitude is common in many dimensions of the Cyberspace, since any private citizen with access to

the Internet can upload content for all to view and download elsewhere, no matter how idiosyncratic or absurd this content may be. In domains other than medical science, the cover of anonymity also facilitates the dissemination of opinions that would probably be held back, if the author could easily be made accountable for them. Curiously, this sharply contrasts with the traditional environment in medical science, where authors typically identify themselves to the public, and highlight their professional qualifications and experience when delivering expert opinion, while referees tend to be covered by anonymity, although they act as guardians of scientific standards of originality, veracity and accountability. In an age of democratic online publishing, the provision of arbitrary, anonymous views as innocent "variations" of somebody else's better-known work does not further the cause of evidence-based medicine.

Academic freedom, scientific accountability and public trust

The JMF list of warning signs, like any scientific document, is perfectable and even open to challenge, a particularly timely issue, as illustrated by recent publications^[8-10]. This work does not address the validity of the original list, but to show that it did not undergo online diffusion without a significant amount of distortion, adaptation or modification. Our intent is to highlight the potential presented by the Internet for distortion or corruption of factual information when nobody is paying attention to what is being diffused, and to who takes responsibility for the modifications.

While we concede that online medical information should be subject to no censorship, it is not sound science to advance a dissenting opinion which has neither an identifiable author nor a peer-reviewed reference. Academic freedom allows anyone to create his/her own list of warning signs of PID, or to modify an existing one if it is understood to be outdated. In the field of immunological diseases, which includes PID, a time-honored classification by Gell and Coombs has recently been revised in-depth, on the basis of extensive scientific evidence^[11]. The latter revision was certainly not an anonymous document, nor lacked references to back it. By the same token, those willing to do so for lists of PID signs should take responsibility for the changes made, in addition to acknowledging their indebtedness to the original they improved upon, rather than attributing their own views to the JMF through incorrect referencing. The confidence of the public in medical information diffused online could only increase if unjustified changes to this information were reported to the original source, thereby promoting sound practices and scientific accountability.

We hope this examination of a strictly defined issue (variations of content in a single piece of medical information), which is highly appropriate as an object for study, but relevant to a relatively small number of people worldwide, will prompt others, better qualified

than ourselves, to extend our initial probing to broader areas, where issues may be manifold and potentially relevant to public health in a larger scale. Among the most interesting and timely subjects for such a broadened examination, we would suggest online resources focusing on other diseases such as AIDS and tuberculosis, which, like PID, involve issues of increased susceptibility to infection, but have far greater social impact, due to the number of affected subjects worldwide.

COMMENTS

Background

Information technology and the Internet have profoundly changed the way physicians, patients, families and support/advocacy groups access and handle information on diseases, diagnoses, and novel/experimental treatments, but it remains unclear whether this revolutionary change adheres to accepted scientific standards of reliability, accuracy and referencing. The authors analyzed how reproducible is online diffusion of a unique instrument, the "Ten Warning Signs of primary immunodeficiency diseases (PID)". This was done because: (1) PID are a large and heterogeneous group of diseases, with related but distinct mechanisms, variable clinical presentations and complex therapeutic choices, requiring specialized diagnostic tools and expert counseling, not necessarily available in general hospitals and clinics; (2) the rarity of most varieties of PID, and the geographical concentration of the diagnostic and therapeutic resources, highlight the practical value of bringing together PID specialists and prospective patients through information diffused online; and (3) this instrument is clearly and concisely written, providing accurate, well-organized and useful information, therefore unreliable diffusion of its contents would have, in practice, a negative effect on the quality of information provided online.

Research frontiers

Although most people wouldn't think of the personal computer and the internet as advanced medical technology, they are fully integrated into medical practice in most countries, and the public, including doctors, trusts them to make important decisions. Nevertheless, there is no systematic follow-up of the contents of medical information pieces propagated in the internet. This is difficult to do in most cases, because large pieces of information on the same subjects would have to be compared, and there are many pieces to compare. In the authors' case, this examination is made easy, because the authors followed a very small piece of factual information up, which is well-structured and divided in numbered statements, so that comparison between the original and any variants is facilitated. This approach allowed the authors to examine, for the first time, the issue of uncontrolled fates for medical information online, and to evaluate the extent of the resulting distortions.

Innovations and breakthroughs

What is novel in the study is that, by examining the diffusion, consistency of use and adequate referencing of this instrument, among highly visited sites from professional, academic and scientific organizations, governmental agencies and patient support/advocacy organizations, the authors could show that the actual contents displayed often depart substantially from their purported model, in form and factual content, including unjustified eliminations and additions. In short, in a sizable proportion of cases, information is deleted, distorted or corrupted, but these changes go unnoticed or unreported.

Applications

The study shows that, by the appropriate choice of a piece of information to be monitored, it is possible to determine how well it adheres to accepted

standards of scientific communication, which is central to its credibility and practical usefulness. This strategy should be applicable to other areas of Medicine affecting large numbers of people, as for instance those concerned by tuberculosis, AIDS or the benefits and risks of vaccination.

Terminology

PID are a large number of different diseases of variable clinical presentation and severity, and demanding specialized diagnosis and management which is not available everywhere. Due to their similarities in mechanisms and manifestations, PID as a group is associated with a constellation of clinical signs and symptoms, mostly due to a higher frequency of infection. The "Ten Warning Signs of PID" is short list of such signs and symptoms, intended to help families to decide whether or not a given patient should be evaluated for PID.

Peer-review

The authors have picked a usual yet interesting and important question regarding the diffusion of medical information in the cyberspace.

REFERENCES

- 1 **Notarangelo LD**, Fischer A, Geha RS, Casanova JL, Chapel H, Conley ME, Cunningham-Rundles C, Etzioni A, Hammartröm L, Nonoyama S, Ochs HD, Puck J, Roifman C, Seger R, Wedgwood J. Primary immunodeficiencies: 2009 update. *J Allergy Clin Immunol* 2009; **124**: 1161-1178 [PMID: 20004777 DOI: 10.1016/j.jaci.2009.10.013]
- 2 **Bonilla FA**, Bernstein IL, Khan DA, Ballas ZK, Chinen J, Frank MM, Kobrynski LJ, Levinson AI, Mazer B, Nelson RP, Orange JS, Routes JM, Shearer WT, Sorensen RU. Practice parameter for the diagnosis and management of primary immunodeficiency. *Ann Allergy Asthma Immunol* 2005; **94**: S1-63 [PMID: 15945566]
- 3 **Savides C**, Shaker M. More than just infections: an update on primary immune deficiencies. *Curr Opin Pediatr* 2010; **22**: 647-654 [PMID: 20683329 DOI: 10.1097/MOP.0b013e32833dd28b]
- 4 **Turvey SE**, Bonilla FA, Junker AK. Primary immunodeficiency diseases: a practical guide for clinicians. *Postgrad Med J* 2009; **85**: 660-666 [PMID: 20075404 DOI: 10.1136/pgmj.2009.080630]
- 5 **Dara M**, Acosta CD, Rusovich V, Zellweger JP, Centis R, Migliori GB. Bacille Calmette-Guérin vaccination: the current situation in Europe. *Eur Respir J* 2014; **43**: 24-35 [PMID: 24381321 DOI: 10.1183/09031936.00113413]
- 6 **Haile M**, Källénus G. Recent developments in tuberculosis vaccines. *Curr Opin Infect Dis* 2005; **18**: 211-215 [PMID: 15864097]
- 7 **Hesseling AC**, Cotton MF, Fordham von Reyn C, Graham SM, Gie RP, Hussey GD. Consensus statement on the revised World Health Organization recommendations for BCG vaccination in HIV-infected infants. *Int J Tuberc Lung Dis* 2008; **12**: 1376-1379 [PMID: 19017445]
- 8 **Arkwright PD**, Gennery AR. Ten warning signs of primary immunodeficiency: a new paradigm is needed for the 21st century. *Ann N Y Acad Sci* 2011; **1238**: 7-14 [PMID: 22129048 DOI: 10.1111/j.1749-6632.2011.06206.x]
- 9 **Subbarayan A**, Colarusso G, Hughes SM, Gennery AR, Slatter M, Cant AJ, Arkwright PD. Clinical features that identify children with primary immunodeficiency diseases. *Pediatrics* 2011; **127**: 810-816 [PMID: 21482601 DOI: 10.1542/peds.2010-3680]
- 10 **MacGinnitie A**, Aloï F, Mishra S. Clinical characteristics of pediatric patients evaluated for primary immunodeficiency. *Pediatr Allergy Immunol* 2011; **22**: 671-675 [PMID: 21449988 DOI: 10.1111/j.1399-3038.2011.01167.x]
- 11 **Descotes J**, Choquet-Kastylevsky G. Gell and Coombs's classification: is it still valid? *Toxicology* 2001; **158**: 43-49 [PMID: 11164991]

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