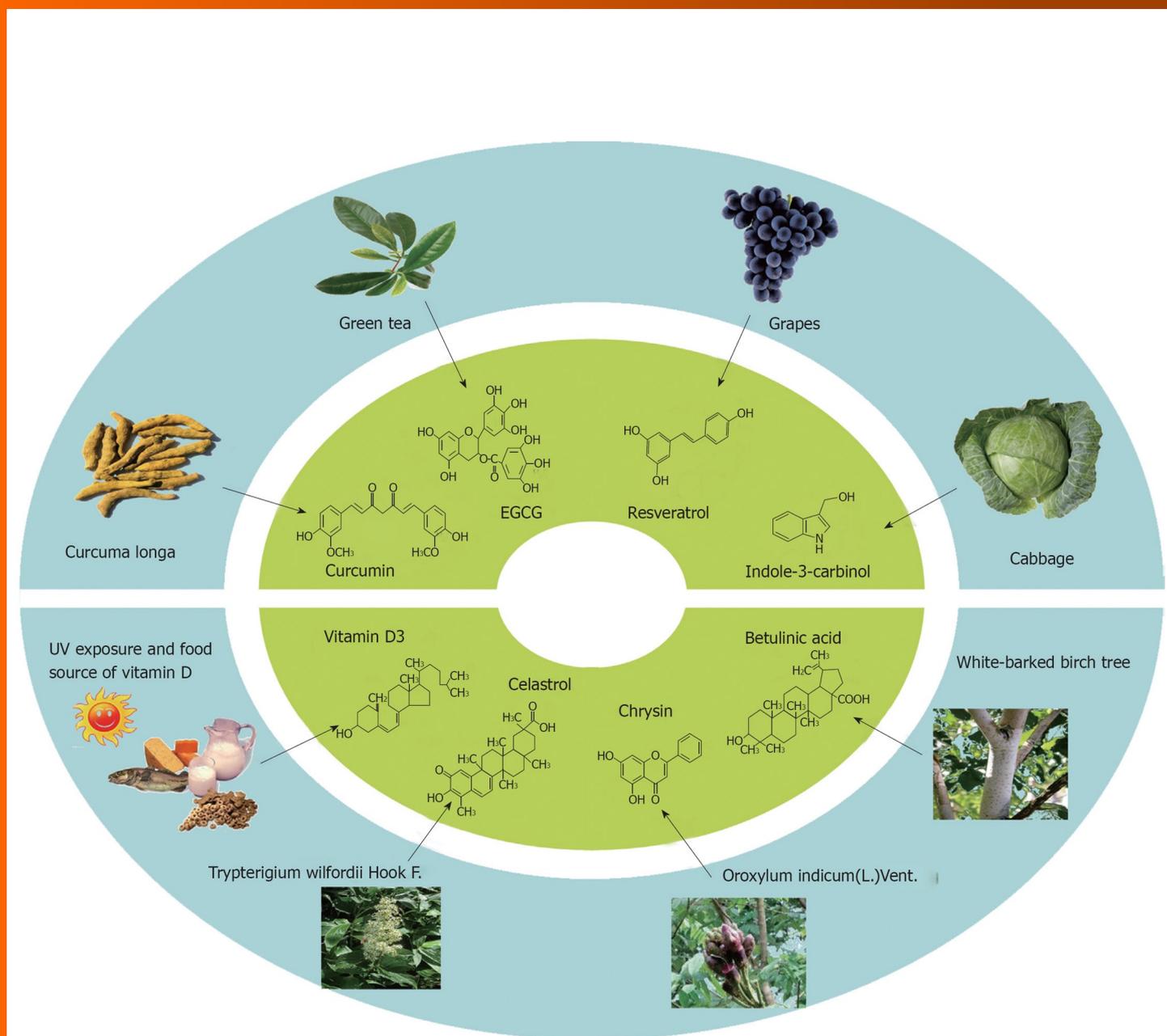


World Journal of *Experimental Medicine*

World J Exp Med 2012 June 20; 2(3): 37-64



Editorial Board

2011-2015

The *World Journal of Experimental Medicine* Editorial Board consists of 104 members, representing a team of worldwide experts in experimental medicine. They are from 30 countries, including Argentina (3), Australia (4), Belgium (2), Brazil (1), Canada (2), China (11), Czech Republic (1), Denmark (2), France (4), Germany (3), Greece (3), India (4), Ireland (1), Israel (1), Italy (7), Japan (6), Lebanon (1), Malaysia (2), Mexico (1), Norway (1), Saudi Arabia (1), Singapore (1), Slovenia (1), South Korea (5), Spain (3), Sweden (3), Switzerland (1), Turkey (2), United Kingdom (2), and United States (25).

EDITOR-IN-CHIEF

De-Ling Kong, *Tianjin*
Atsushi Mizoguchi, *Boston*
Baohong Zhang, *Greenville*

GUEST EDITORIAL BOARD MEMBERS

Nan-Shan Chang, *Tainan*
Kow-Tong Chen, *Tainan*
Chih-Ping Hsu, *Hsin-Chu*
Hung-Jen Liu, *Taichung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Beatriz Basso, *Córdoba*
Cristina Ester Carnovale, *Rosario*
Angel Catalá, *La Plata*



Australia

Filip Braet, *Sydney*
Xianlan Cui, *Launceston*
Xiao-Jun Du, *Melbourne*
HS Nagaraja, *Queensland*



Belgium

Olivier Bruyere, *Liege*
Nathalie Cools, *Edegem*



Brazil

Niels Olsen Saraiva Câmara, *Sao Paulo*



Canada

Alfonso Iorio, *Hamilton*
Xiaoyan Jiang, *Vancouver*



China

Long Chen, *Nanjing*
Heng-Mi Cui, *Nanjing*
Volodymyr Dvornyk, *Hong Kong*
Jian-Xin Gao, *Shanghai*
Chun-Yan Ji, *Jinan*
Yang-Fu Jiang, *Chengdu*



Czech Republic

Jan Bernardy, *Brno*



Denmark

Shan Gao, *Aarhus*
Per Hildebrandt, *Frederiksberg*



France

Nadia Alfaidy, *Grenoble*
Abdel Aouacheria, *Lyon*
Jean-Marc Cavaillon, *Paris*
Jean-Marc Egly, *Illkirch*



Germany

Sorin Armeanu-Ebinger, *Tübingen*
Magali Cucchiari, *Homburg*

Mohamed Hassan, *Duesseldorf*



Greece

Effie K Basdra, *Athens*
Maria Dalamaga, *Athens*
Moses Elisaf, *Ioannina*



India

Malay Chatterjee, *Kolkata*
Vijay Chauthaiwale, *Ahmedabad*
Bibhu Ranjan Das, *Mumbai*
Satya N Das, *New Delhi*



Ireland

Steven G Gray, *Dublin*



Israel

Elena Feinstein, *Ness Ziona*



Italy

Alessandro Busca, *Turin*
Giovanni Di Salvo, *Naples*
Francesco Dieli, *Palermo*
Amalia Forte, *Naples*
Umberto Galderisi, *Naples*
Gabriele Grassi, *Trieste*
Fabio Grizzi, *Rozzano*



Japan

Winn Aung, *Chiba*

Hiroshi Fukazawa, *Mito*
Hideaki Hara, *Gifu*
Toshio Hattori, *Sendai*
Atsushi Hosui, *Suita*
Peng Huang, *Okayama*



Lebanon

Hala Gali-Muhtasib, *Beirut*



Malaysia

Gam Lay Harn, *Penang*
Kamsiah Jaarin, *Kuala Lumpur*



Mexico

Javier Camacho, *Mexico*



Norway

Brynjar Foss, *Stavanger*



Saudi Arabia

Mostafa M El-Naggar, *Jazan*



Singapore

Ivy Ho, *Singapore*



Slovenia

Damjan Glavac, *Ljubljana*



South Korea

Dalwoong Choi, *Seoul*
Kang-Yell Choi, *Seoul*
Joo-hun Ha, *Seoul*
Eui-Bae Jeung, *Cheongju*
Chang-Duk Jun, *Gwangju*



Spain

Isabel Andia, *Bilbao*
Javier Arias-Diaz, *Madrid*
Vicente Felipo, *Valencia*



Sweden

Karl O Fagerstrom, *Kagerod*
Robert Hahn, *Tullinge*
Susanne Jacobsson, *Örebro*



Switzerland

Florian Bihl, *Bellinzona*



Turkey

Ali Kudret Adiloglu, *Ankara*

Mutay Aslan, *Antalya*



United Kingdom

Dominique Bonnet, *London*
David Gilham, *Manchester*



United States

Anshu Agrawal, *Irvine*
Mikhail Alexeyev, *Mobile*
Robert J Amato, *Houston*
Raymond T Bartus, *San Diego*
Ajay Singh Behl, *Minneapolis*
Fabian Benencia, *Athens*
Arun Bhunia, *West Lafayette*
Ramireddy Bommireddy, *Tucson*
Michael Borchers, *Cincinnati*
Alexander Bukreyev, *Galveston*
Lu Cai, *Louisville*
Arvind Chhabra, *Farmington*
Yingzi Cong, *Galveston*
Akram Da'darah, *North Grafton*
Liutao Du, *Los Angeles*
Nejat Düzgüneş, *San Francisco*
Charles E Egwuagu, *Bethesda*
Lianchun Fan, *Indianapolis*
Bingliang Fang, *Houston*
Seshu K Gudlavalleti, *Omaha*
Diane M Harper, *Leawood*
Mohamed I Husseiny, *Los Angeles*
Miroslaw Janowski, *Baltimore*



Contents

Bimonthly Volume 2 Number 3 June 20, 2012

EDITORIAL	37	Potential for a pluripotent adult stem cell treatment for acute radiation sickness <i>Rodgerson DO, Reidenberg BE, Harris AG, Pecora AL</i>
REVIEW	45	Natural compounds as anticancer agents: Experimental evidence <i>Wang J, Jiang YF</i>
BRIEF ARTICLE	58	Indirect calorimetry in obese female subjects: Factors influencing the resting metabolic rate <i>Hagedorn T, Poggiogalle E, Savina C, Coletti C, Paolini M, Scavone L, Neri B, Donini LM</i>

ACKNOWLEDGMENTS I Acknowledgments to reviewers of *World Journal of Experimental Medicine*

APPENDIX I Meetings
I-V Instructions to authors

ABOUT COVER Wang J, Jiang YF.
Natural compounds as anticancer agents: Experimental evidence.
World J Exp Med 2012; 2(3): 45-57
<http://www.wjgnet.com/2220-315X/full/v2/i3/45.htm>

AIM AND SCOPE *World Journal of Experimental Medicine (World J Exp Med, WJEM, online ISSN 2220-315X, DOI: 10.5493)* is a bimonthly peer-reviewed, online, open-access, journal supported by an editorial board consisting of 104 experts in experimental medicine from 30 countries.

The aim of *WJEM* is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of experimental medicine. *WJEM* covers topics concerning clinical laboratory medicine, biochemical examination (applied and basic research in laboratory automation and information system, biochemical methodology, and biochemical diagnostics), clinical microbiology, immunodiagnosics (laboratory diagnosis of infectious diseases, tumor markers and their application, laboratory diagnosis of autoimmune diseases, and immunotechnology), clinical laboratory management (laboratory quality control and management, traceability and calibration, information management system and laboratory automation, and laboratory biosafety management), and experimental medicine-related traditional medicine, and integrated Chinese and Western medicine. The journal also publishes original articles and reviews that report the results of experimental medicine-related applied and basic research in fields such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

FLYLEAF I-II Editorial Board

EDITORS FOR THIS ISSUE Responsible Assistant Editor: Yuan Zhou Responsible Science Editor: Jin-Lei Wang
Responsible Electronic Editor: Xiao-Mei Zheng Proofing Editorial Office Director: Jin-Lei Wang
Proofing Editor-in-Chief: Lian-Sheng Ma

NAME OF JOURNAL
World Journal of Experimental Medicine

ISSN
ISSN 2220-315X (online)

LAUNCH DATE
December 20, 2011

FREQUENCY
Bimonthly

EDITING
Editorial Board of *World Journal of Experimental Medicine*
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: wjem@wjgnet.com
<http://www.wjgnet.com>

EDITOR-IN-CHIEF
De-Ling Kong, PhD, Professor, Institute of Molecular Biology, Nankai University, Tianjin 300071, China

Atsushi Mizoguchi, MD, PhD, Associate Professor in Pathology, Harvard Medical School, Molecular Pathology Unit, Massachusetts General Hospital, CNY149-6024, 13th Steert, Charlestown, MA 02114, United States

Baohong Zhang, PhD, Assistant Professor of Biology, Department of Biology, East Carolina University, Greenville, NC 27858, United States

EDITORIAL OFFICE
Jian-Xia Cheng, Director
Jin-Lei Wang, Vice Director
World Journal of Experimental Medicine
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: wjem@wjgnet.com
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Co., Limited
Room 1701, 17/F, Henan Building,
No.90 Jaffe Road, Wanchai, Hong Kong, China
Fax: +852-31158812

Telephone: +852-58042046
E-mail: bpg@baishideng.com
<http://www.wjgnet.com>

PUBLICATION DATE
June 20, 2012

COPYRIGHT
© 2012 Baishideng. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

INSTRUCTIONS TO AUTHORS
Full instructions are available online at http://www.wjgnet.com/2220-315x/g_info_20100722180909.htm.

ONLINE SUBMISSION
<http://www.wjgnet.com/esp/>

Potential for a pluripotent adult stem cell treatment for acute radiation sickness

Denis O Rodgerson, Bruce E Reidenberg, Alan G Harris, Andrew L Pecora

Denis O Rodgerson, Alan G Harris, Andrew L Pecora, NeoStem, Inc., New York, NY 10170, United States

Bruce E Reidenberg, Department of Pharmacology, Weill Medical College of Cornell University, New York, NY 10170, United States

Andrew L Pecora, The Cancer Center, Hackensack University Medical Center, Hackensack, NJ 07601, United States

Author contributions: All authors contributed equally to this paper.

Correspondence to: Denis O Rodgerson, PhD, Director of Stem Cell Science, NeoStem, Inc., 420 Lexington Avenue, Suite 450, New York, NY 10170,

United States. drodgerson@neostem.com

Telephone: +1-818-3261233 Fax: +1-646-5147787

Received: March 28, 2012 Revised: June 7, 2012

Accepted: June 15, 2012

Published online: June 20, 2012

Abstract

Accidental radiation exposure and the threat of deliberate radiation exposure have been in the news and are a public health concern. Experience with acute radiation sickness has been gathered from atomic blast survivors of Hiroshima and Nagasaki and from civilian nuclear accidents as well as experience gained during the development of radiation therapy for cancer. This paper reviews the medical treatment reports relevant to acute radiation sickness among the survivors of atomic weapons at Hiroshima and Nagasaki, among the victims of Chernobyl, and the two cases described so far from the Fukushima Dai-Ichi disaster. The data supporting the use of hematopoietic stem cell transplantation and the new efforts to expand stem cell populations *ex vivo* for infusion to treat bone marrow failure are reviewed. Hematopoietic stem cells derived from bone marrow or blood have a broad ability to repair and replace radiation induced damaged blood and immune cell production and may promote blood vessel formation and tissue repair. Additionally, a constituent of bone marrow-derived, adult pluripotent stem cells, very small embryonic like stem cells, are highly resistant to ioniz-

ing radiation and appear capable of regenerating radiation damaged tissue including skin, gut and lung.

© 2012 Baishideng. All rights reserved.

Key words: Nuclear accident; Acute radiation syndrome; Radiological casualties; Stem cell transplantation; Cellular therapy; Emergency response; Ionizing radiation injury; Hematopoietic rescue; Pluripotent stem cells; Induced pluripotent stem cells; Mesenchymal stem cells; Very small embryonic-like stem cells; Mobilizing agents

Peer reviewer: Magali Cucchiari, PhD, Assistant Professor, Molecular Biology, Center of Experimental Orthopaedics, Saarland University Medical Center, Kirrbergerstr. Bldg 37, D-66421 Homburg/Saar, Germany

Rodgerson DO, Reidenberg BE, Harris AG, Pecora AL. Potential for a pluripotent adult stem cell treatment for acute radiation sickness. *World J Exp Med* 2012; 2(3): 37-44 Available from: URL: <http://www.wjgnet.com/2220-315X/full/v2/i3/37.htm> DOI: <http://dx.doi.org/10.5493/wjem.v2.i3.37>

INTRODUCTION

Accidental radiation exposure and the threat of deliberate radiation exposure have been in the news and are a public health concern. This paper will describe the state of the art of stem cell treatment of acute radiation sickness. Acute radiation sickness is defined as “a combination of clinical syndromes occurring in stages during hours to weeks after exposure as injury to various tissues and organs is expressed”^[1]. Experience with acute radiation sickness has been gathered from atomic blast survivors of Hiroshima and Nagasaki and from civilian nuclear accidents as well as experience gained during the development of radiation therapy for cancer. Based on these sources, an approximate dose threshold for each target organ (Table 1) and a time course of illness can be estimated (Table 2).

Note that bone marrow failure (infection, hemorrhage) is not the exclusive cause of death. High dose radiation can kill with cerebral edema and enteritis and pneumonitis, independent of infection. These syndromes are unlikely to be treatable with hematopoietic stem cell transplantation. Pluripotent (ability to differentiate into all three germ layers) stem cells with potential to regenerate multiple tissue types would add an important benefit for the treatment of acute radiation syndrome. Very small embryonic-like stem cells that can be obtained from adults in autologous cell-dose quantities offer such an advantage and are discussed in more detail latter in this paper.

The standard of care is described in the United States Armed Forces Radiobiology Research Institute's "Medical Treatment of Radiological Casualties"^[2]. There are many complexities to caring for patients after radiation exposure. Patients who have been exposed to an explosion may have life-threatening injury not related to radiation exposure. Patients may be externally or internally contaminated with radioactive particles. Rapid and effective decontamination can prevent serious sequelae including bone marrow failure. Another complication is radiation induced emesis which can be dehydrating and limit the utility of orally administered countermeasures. Medical countermeasures for radiation exposure can be classified into 3 groups^[3]: (1) Radioprotectants prevent radiation damage to cells (e.g., amifostine); (2) Radiation mitigators limit radiation damage (e.g., pentoxifylline); and (3) Radionuclide eliminators enhance excretion of radionuclides (e.g., Prussian Blue).

The aspects of acute radiation sickness for which hematopoietic stem cell transplantation is appropriate is amelioration of bone marrow suppression and immune suppression and tissue damage repair. This would fit the classification of "radiation mitigators" because it limits damage that has already occurred. For purposes of describing the value of hematopoietic stem cell transplantation in acute radiation sickness, patients who received between 2 Gy -10 Gy are recommended to be treated with white blood cell supporting cytokines, either G-CSF (filgrastim, peg-filgrastim) or GM-CSF (sargramostim). Cytokines are unlikely to be clinically useful in most cases where exposure exceeds 4 Gy. Patients for whom CSFs are unsuccessful are candidates for hematopoietic stem cell transplantation. The published data on the success of bone marrow transplantation following non-therapeutic radiation exposure include the experience of 13 Chernobyl victims described in the next section. In total reports from 58 people exposed to radiation in excess of 5 Gy, half of whom had an allogeneic transplant, revealed that only three of 29 patients transplanted were alive at one year post exposure. Deaths occurred due to the development of graft-*vs*-host disease and other complications unique to allogeneic transplant that could be avoided if autologous bone marrow or blood-derived stem cells were collected and stored before the exposure and used in place of allogeneic cells.

CURRENT INFORMATION ON ACCIDENTAL (CIVILIAN) OR DELIBERATE (MILITARY, TERRORISM) RADIATION EXPOSURE

In addition to approximately 20 civilian and 60 military nuclear accidents, there have been 3 major nuclear accidents as of June 2011: Three Mile Island in the United States, Chernobyl in the former Soviet Union and Fukushima Dai-Ichi in Japan. In these accidents, it is very difficult to quantify the amount of radiation released, but some information is available on acute radiation sickness following the accidents. The Three Mile Island accident during which a portion of the nuclear fuel melted down in a TMI-2 reactor, but did not breach the containment walls, occurred on March 28, 1979. The widespread perception of great danger from this accident was based on expert's concern that the containment vessel might explode, widely distributing radioactive material. In fact, the containment vessel maintained integrity. Even though increased radiation levels were detected inside the plant and at least 50 workers were exposed, no acute radiation sickness from this accident has been reported^[4,5].

The Chernobyl accident on April 26, 1986 was much more serious with significant public health consequences. The difficulty in verifying documentation and medical records of the government of the Soviet Union makes an assessment of the extent of acute radiation sickness due to the Chernobyl accident impossible to reconstruct. Nevertheless, a comprehensive review of available information was published by the New York Academy of Sciences in November 2009^[6]. The lowest estimate of acute mortality from the Chernobyl disaster is 9000 victims^[7]. Soviet physicians reported on 13 bone marrow transplantations for acute radiation sickness due to exposure at Chernobyl. Twelve of 13 patients had skin injuries resembling burns from 20%-100% body surface area in addition to decreasing white blood cell counts. Four of 8 patients with non-HLA identical donors received T-cell depleted bone marrow transplants. Only 2 of the transplant recipients survived to the 3 year follow up. The deaths reported were not attributed to prolonged neutropenia/infection or to thrombocytopenia/bleeding. Interestingly, two of the transplant recipients had evidence of transient engraftment with donor cells followed by recovery of autologous bone marrow^[8].

Most recently, on March 11, 2011, a magnitude 9 earthquake followed by a tsunami estimated at 14 meters high, destroyed part of TEPCOs Fukushima Dai-ichi nuclear power plant and resulted in several explosions. The International Atomic Energy Commission Briefing disclosed Fukushima prefecture received 1.5 microSv/h on March 31 over a natural background of 0.1 microSv/h^[9]. Two workers were reported to have received radiation burns to ankles when wading in contaminated water^[10]. These are the only two cases of acute radiation sickness

Table 1 Approximate threshold doses of conventionally fractionated therapeutic radiation for clinically detrimental nonstochastic effects in various tissues

Organ	Injury at 5 yr	Threshold dose (sv) ¹	Irradiation field (area)
Fetus	Death	2	Whole
Bone marrow	Hypoplasia	2	Whole
Ovary	Permanent sterility	2-3	Whole
Lens	Cataract	5	Whole
Testes	Permanent sterility	5-15	Whole
Cartilage, child	Arrested growth	10	Whole
Breast, child	Hypoplasia	10	5 cm ²
Bone, child	Arrested growth	20	10 cm ²
Bone marrow	Hypoplasia, fibrosis	20	Localized
Muscle, child	Hypoplasia	20-30	Whole
Kidney	Nephrosclerosis	23	Whole
Lymph nodes	Atrophy	33-45	-
Liver	Liver failure, ascites	35	Whole
Lung	Pneumonitis, fibrosis	40	Lobe
Heart	Pericarditis, pancarditis	40	Whole
Stomach, small intestine, colon	Ulcer, perforation	45	100 cm ²
Thyroid	Hypothyroidism	45	Whole
Pituitary	Hypopituitarism	45	Whole
Lymphatics	Sclerosis	50	-
Central nervous system (brain)	Necrosis	50	Whole
Spinal cord	Necrosis, transection	50	5 cm ²
Salivary glands	Xerostomia	50	50 cm ²
Cornea	Keratitis	50	Whole
Capillaries	Telangiectasis, fibrosis	50-60	-
Breast, adult	Atrophy, necrosis	> 50	Whole
Rectum	Ulcer, stricture	55	100 cm ²
Skin	Ulcer, severe fibrosis	55	100 cm ²
Eye	Panophthalmitis, hemorrhage	55	Whole
Oral mucosa	Ulcer, severe fibrosis	60	50 cm ²
Esophagus	Ulcer, stricture	60	75 cm ²
Cartilage, adult	Necrosis	60	Whole
Urinary bladder	Ulcer, contracture	60	Whole
Bone, adult	Necrosis, fracture	60	10 cm ²
Ear (inner)	Deafness	> 60	Whole
Adrenal	Hypoadrenalism	> 60	Whole
Vagina	Ulcer, fistula	90	5 cm
Muscle, adult	Atrophy	> 100	Whole
Uterus	Necrosis, perforation	> 100	Whole

¹Dose causing effect in 1% to 5% of exposed persons. Modified from^[49,50].

reported to date. In October 2011, a consensus document was published that includes additional individual case reports from sparsely documented historical civilian accidental exposures with the caveat that information from those reports was insufficient to guide future therapy^[11].

From a perspective on military use of nuclear weapons, the acute radiation sickness due to use of atomic weapons on Hiroshima and Nagasaki during World War Two has been reviewed^[12]. Survey of 1216 survivors of the blast in Hiroshima, sheltered in a building, revealed that 451 died on the first day and 201 died in the succeeding 2 mo, presumably from the hematopoietic component of acute radiation syndrome. Since transplantation had not been developed, there are no data on bone marrow transplant or stem cell treatment of acute radiation sickness after weapons discharge. It is the mortality figures from the Hiroshima and Nagasaki bombs that form the basis of military mathematical models to predict acute radiation sickness following nuclear weapons discharge.

The United States Health and Human Services' Office of Preparedness and Emergency Operations has made public the scenarios being used to prepare the United States. Two of these scenarios (#1 and #11) include nuclear weapons. These scenarios are being used to plan public health resource prioritizations and can be applied to estimate the number of patients who would potentially benefit from hematopoietic stem cell transplantation. National Planning Scenario #1 envisions a 1 KT nuclear detonation^[13]. Col. Jarrett published an estimate of a 4 × 3 km oval that would receive 4 Gy from a 1 kT nuclear detonation^[1]. Utilizing published population densities (New York City = 4500 people/km² and San Francisco = 5400 people/km²), this area (9.4 km²) would represent between 42 000 and 50 000 victims. National Planning Scenario #11 envisions a Radioactivity Dispersal Device ("Dirty Bomb") which would produce a "no entry" zone (> 1 Gy exposure) of 500 m in diameter (0.2 km²). Utilizing the same published population densities as above, this would represent between 900 and 1080 victims^[14]. These estimates demonstrate

Table 2 Symptoms, therapy and prognosis of whole body ionizing radiation injury

	0-1 Sv	1-2 Sv	2-6 Sv	6-10 Sv	10-20 Sv	> 50 Sv
Therapeutic needs	None	Observation	Specific treatment	Possible treatment	Palliative	Palliative
Vomiting	None	5%-50%	> 3 Gy, 100%	100%	100%	100%
Time to nausea, vomiting	-	3 h	2 h	1 h	30 min	< 30 min
Main locus of injury	None	Lymphocytes	Bone marrow	Bone marrow	Small bowel	Brain
Symptoms and signs	-	Moderate leukopenia, epilation	Leukopenia, hemorrhage, epilation	Leukopenia, hemorrhage, epilation	Diarrhea, fever, electrolyte imbalance	Ataxia, coma, convulsions
Critical period	-	-	4-6 wk	4-6 wk	5-14 d	1-4 h
Therapy	Re-assurance	Observation	Transfusion of granulocytes, platelets	Transfusion, antibiotics, bone marrow transplantation	Fluids and salts, possible bone marrow transplantation	Palliative
Prognosis	Excellent	Excellent	Guarded	Guarded	Poor	Hopeless
Lethality	None	None	0%-80%	80%-100%	100%	100%
Time of death	-	-	2 mo	1-2 mo	2 wk	1-2 d
Cause of death	-	-	Infection, hemorrhage	Hemorrhage, infection, pneumonitis	Enteritis, infection	Cerebral edema

Modified from^[51].

that even “small” events in a crowded environment may create enormous demands on the local medical system, and would probably exceed the capabilities of almost all facilities.

As discussed earlier, currently available treatment for radiation exposures of greater than 1 Gy are palliative. Hematopoietic stem cell transplantation to rescue patients for whom cytokine therapy failed has several limitations. The primary limitation is that the donor pool is limited by the need for at least partial HLA matching. As an example, the United States National Marrow Donor Program reports among 9 million donors, only 650 000 (7%) are African American, making bone marrow matching for African Americans difficult^[13]. Similar problems probably exist for other under-represented ethnic groups. Once the hematopoietic transplant has engrafted, there is continuous need for immunosuppression. In addition to the risks of life-threatening infection during titration of immunosuppressant medication, some of these medications have dose limiting acute and chronic toxicity independent of graft-*vs*-host disease^[16].

Autologous hematopoietic stem cell treatment would solve the problems of immunosuppression and graft-*vs*-host disease. If people at risk were to receive G-CSF mobilized cells collected prior to exposure than there would be sufficient cells available to prevent the profound cytopenia and immune suppression that follows exposure to 4 Gy or more of radiation. In addition, a small volume of bone marrow (100-200 mL) collected prior to exposure and then expanded *ex vivo* post exposure, may also be sufficient to reconstitute hematopoiesis and immune function. The major concern is whether hematopoietic stem cells capable of re-constituting the bone marrow could be expanded *ex vivo* from 100-200 mL, before bone marrow suppression became life-threatening. Clinical studies using marrow, mobilized blood and cord blood have demonstrated the feasibility of doing so^[17-19]. Harvesting hematopoietic stem cells from damaged marrow is being done

with complex protocols in cancer treatment. However, there are many reports of protocols failing to mobilize hematopoietic stem cells sufficient for reconstitutive use. For example, fludarabine exposure in adults with follicular lymphoma predicted a poor hematopoietic stem cell harvest evidenced by > 5 d apheresis requirement^[20]. In a retrospective analysis of 204 patients, Ford *et al*^[21] calculated that platinum based drugs and etoposide exposure were most highly correlated with poor hematopoietic stem cell mobilization as reflected by the absence of CD34+ cells on the first day that the white blood cell count was greater than 500. Stem cell mobilization was reported successful in only 12 of 20 (60%) patients with chronic lymphocytic leukemia^[22]. These results suggest that chemotherapy treatment at a minimum impairs hematopoietic stem cell mobilization. In addition, a new study confirms expectations, that age between 65-69 years impairs hematopoietic stem cell mobilization relative to younger patients with the same disease^[23]. In contrast, hematopoietic stem cell harvest in children is not limited by mobilization, but by scaling factors in extracorporeal volumes and anticoagulation necessary for the apheresis machine and vascular access for sufficient flow^[24]. No reports of hematopoietic stem cell harvest from pregnant women could be found on PubMed search. Ford *et al*^[21] did not find any correlation of poor hematopoietic stem cell mobilization with prior radiotherapy which gives these authors hope that victims of acute radiation sickness could have their hematopoietic stem cells successfully harvested. However, radiation damage may result in long term issues such as myelodysplasia and leukemia so use of cells previously stored and not exposed to radiation would appear optimal.

NEW PROCEDURES ON THE HORIZON

New mobilizing agents

New mobilizing agents are being developed to replace the colony stimulating factors. The hematopoietic stem

cell harvesting described above utilized G-CSF (filgrastim or peg-filgrastim) and/or GM-CSF (sargramostim) as mobilizing agents. The newer agent, Mozobil® (perixaflor) was approved in the United States in 2009 and acts by reversibly binding CXCR4 and inhibiting CXCR4/CXCL12 anchoring^[25]. Mobilization with Mozobil® (perixaflor) increased successful 4 d apheresis harvesting from 88% (136/154 patients) with G-CSF (filgrastim) alone to 95% (141/148 patients) with both G-CSF (filgrastim) and Mozobil® (perixaflor)^[25]. Natalizumab is an antibody in development as a mobilizing agent that binds to VCAM-1 and interferes with VCAM-1/VLA-4 anchoring^[26].

Though un-glycosylated thrombopoietin combined with G-CSF was effective at mobilizing hematopoietic stem cells, the risk of developing autoimmune thrombocytopenia led to the cessation of development of un-glycosylated thrombopoietin^[27]. The effect of thrombopoietin agonists on animal models of radiation induced thrombopenia are in progress for the peptide Nplate (romiplostim) and orally dosed small molecule Promecta (eltrombopag) and full length, glycosylated, recombinant human thrombopoietin^[27].

Burdelya *et al*^[28] reported mouse radio-protective activity from a *Salmonella enterica* flagellin derivative given 1 h after radiation exposure and rhesus monkey protection when given 45 min prior to radiation exposure. Its putative mechanism of action is *via* toll like receptor 5 (TLR5) to nuclear factor- κ B (NF- κ B) signaling to multiple cytokines including G-CSF. This product is under active development by Cleveland Biolabs, Inc., (Buffalo, NY).

Parathyroid hormone (PTH) appears to mobilize stem cells to peripheral blood in mice with a distinct mechanism from G-CSF^[26]. In a Phase 1 study in patients with at least one failed peripheral stem cell harvest attempt, the combination of PTH (teraparitide) for days 1-14 and G-CSF (filgrastim) for days 10-14 prior to apheresis resulted in 9/20 (45%) patients meeting pre-specified mobilization criteria. The authors note that this level of success was also seen in second mobilization attempts with a combination of filgrastim (G-CSF) and sargramostim (GM-CSF). A spontaneous observation study of patients with primary hyperparathyroidism showed CD45+/CD34+/c-kit+ and CD45+/CD34+/CXCR4+ bone marrow progenitor cells were increased relative to matched controls. Interestingly, the primary hyperparathyroidism patients had lower G-CSF dosing than controls, while stem cell factor and erythropoietin were not different between groups^[29]. Another spontaneous observation study evaluated hemodialysis patients with varying levels of secondary hypo- and hyper-parathyroidism. In patients with high PTH, circulating hematopoietic stem cells were higher than controls or patients with normal PTH levels. In patients with low PTH, circulating stem cell numbers were lower than patients with normal or elevated PTH^[30].

Lastly, α -tocopherol succinate is being explored as a single dose inducer of endogenous G-CSF equivalent to a multiday course of G-CSF (filgrastim)^[31].

STEM CELL THERAPY

Induced pluripotent stem cells

Another source of cells to reconstitute radiation damaged bone marrow would be adult induced pluripotent stem cells (iPS). Recent laboratory work shows that fibroblasts can be induced to become pluripotent stem cells without using retroviruses, only using mRNA for key transforming factors^[32]. Human iPS maintain a flat colony morphology in the laboratory when maintained with basic Fibroblast Growth Factor. Induction of iPS into hematopoietic stem cells covering all three lineages (myelopoietic, erythropoietic and thrombopoietic) has not yet been described. Current laboratory confirmation of pluripotency is the ability of iPS to form teratomas *in vitro* and *in vivo*. Though this is a large potential clinical problem, progress continues to be made and a recent report describes reprogramming skin fibroblasts from a patient with thalassemia (single gene mutation)^[33]. Laboratory size colonies appeared in 3 wk following transfection with an engineered retrovirus and were induced to form "hemaglobinized" (HbF producing) colonies in another 2 wk using specially prepared growth media. No description of resulting myeloid or thrombopoietic cells were offered^[33]. In addition, the "retro-differentiation" process currently requires approximately 100 adult cells to create 1 induced pluripotent stem cell (1% efficient) and the cells so produced exhibit early senescence^[34]. Recently Zhao *et al*^[35] have shown that iPS can be rejected due to abnormal gene expression in some iPS cells which can induce a T-cell-dependent immune response. These authors recommend that the immunogenicity of autologous cells should be carefully evaluated before these cells are considered for therapeutic purposes. So as of this writing, iPS are not on the near-horizon for clinical use in acute radiation sickness. Barriers that will need to be overcome to use iPS to treat acute radiation sickness include: production of all three bone marrow lineages from iPS; managing the risk of using oncogene sequences for induction or the identification of alternate induction techniques; improving the speed of hematopoietic stem cell production to a timeframe consistent with rescuing the patient from bone marrow failure; eliminating the risk of rejection; and scaling up production for mass casualties.

Mesenchymal stem cells

A commercial product of human mesenchymal stem cells (MSC) prepared from multiple bone marrow donors (Prochymal®) is being developed for acute Graft-vs-Host Disease in bone marrow transplants. Since one lineage of MSC matures into bone marrow stromal cells, MSC have been considered as a supportive treatment for primary engrafting of bone marrow transplants. The initial publication in 2007 that human MSC home to radiation-damaged tissue in mice provided evidence of the potential for restorative therapy outside of the bone marrow^[36]. Hu *et al*^[37] demonstrated that MSC rescued

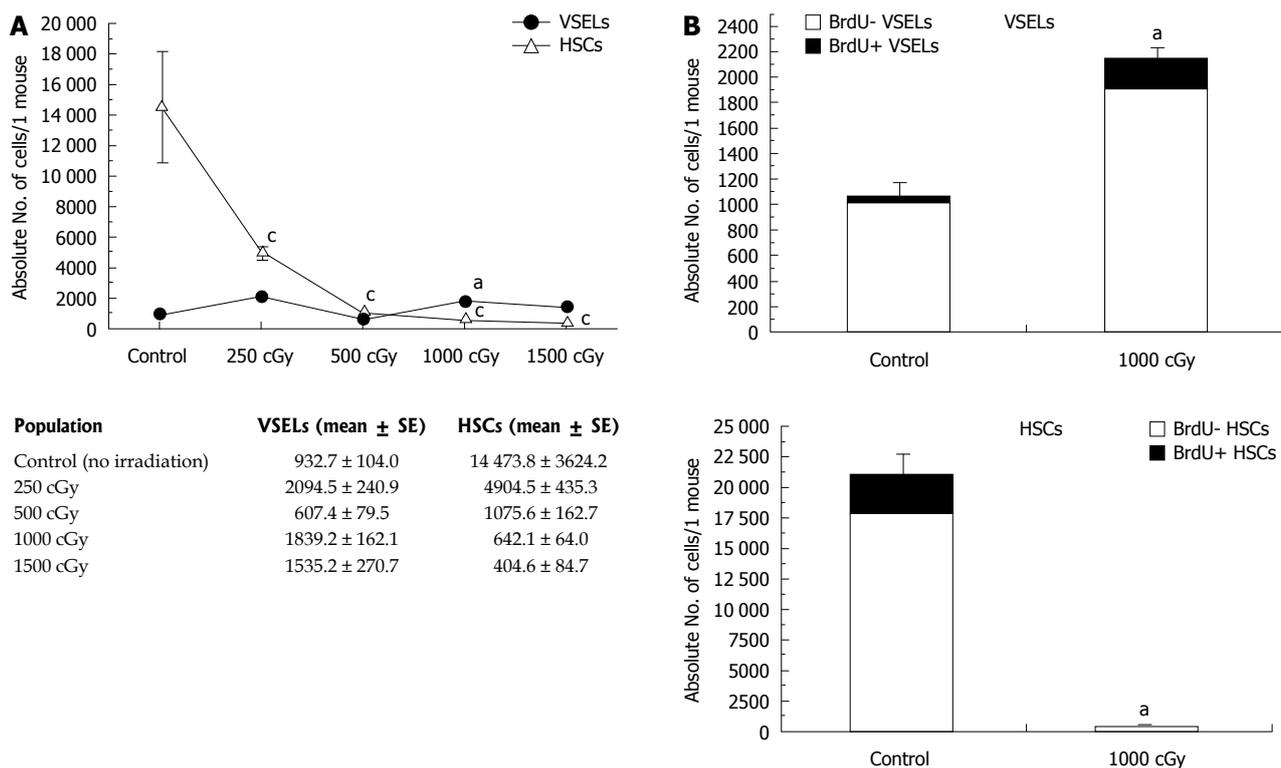


Figure 1 Resistance of very small embryonic-like stem cells to γ -radiation. The content of very small embryonic-like stem cells (VSELs) and HSCs was evaluated in murine BM following whole-body irradiation with different doses of γ -radiation (250, 500, 1000 and 1500 cGy) when compared to control (no irradiation). A: Absolute numbers of VSELs (Sca-1+/Lin-/CD45-) and HSCs (Sca-1+/Lin-/CD45+) in BM after 4 d post-irradiation. The table presents mean numbers of VSELs and HSCs per mouse (mean \pm SE). ^a $P < 0.05$ vs control (VSELs); ^c $P < 0.05$ vs control (HSCs); B: Absolute numbers of VSELs and HSCs incorporating BrdU following whole-body irradiation. Data are presented as mean absolute numbers of VSELs and HSCs per mouse (mean \pm SE) (From Ref. [43], with permission). ^a $P < 0.05$ vs Control.

fatally irradiated mice. Lange *et al.*^[38] extended the findings to show that MSCs effect rescue in fatally irradiated mice by anti-inflammatory and “hematopoietic stem cell niche modulating” effects such that endogenous hematopoiesis recovers. Thus, it is possible that MSCs may indeed have a beneficial effect in acute radiation sickness, however it is credible that this effect is due to paracrine, trophic and endogenous stem cell recruitment, rather than a regenerative capability. As the work of Heider *et al.*^[39] would suggest, bone marrow derived pluripotent stem cells give rise to MSCs and thus are supportive of the role of MSCs in hematopoiesis, but MSCs themselves are not regenerative.

In this setting, Osiris Therapeutics and Genzyme Corporation are collaborating to develop Prochymal[®] for the treatment of acute radiation sickness. Current clinical trials in steroid refractory graft-*vs*-host disease, where patients have received significant radiation exposure to treat their underlying disease, will provide a human model on which to base animal studies of Acute radiation sickness^[40].

Myeloid progenitor cell product

Though not a stem cell product, CLT008, being developed by Cellerant Therapeutics (San Carlos, CA), is a multi-person sourced human cell population derived from donor bone marrow. This product is reported to

have capability to mature into monocytes, neutrophils and red blood cells. Though it does not have the potential to mature into T and B lymphocytes, it is being considered as a temporary therapy until the host marrow recovers^[41].

Very small embryonic-like cells

Very small embryonic-like cells (VSELs) are pluripotent and present in many tissues and circulate in peripheral blood. The properties and therapeutic potentials of VSELs have been recently reviewed^[42]. VSELs can differentiate into multiple cell types *in vitro* and in mice^[43]. Kassmer *et al.*^[44] recently reported that bone marrow derived stem cells that were not hematopoietic were able to differentiate into type 2 pneumocytes in fatally irradiated mice. These small bone marrow cells were reported to be identical to VSELs (personal communication, DS Krause, 2010). VSELs have been documented to be present during routine bone marrow or hematopoietic stem cell harvesting^[45]. Ratajczak and colleagues report that in addition to hematopoietic stem cells, VSELs are mobilized during burn injury^[46]. This adds important information to the mobilization of VSELs during acute myocardial infarct and stroke in humans^[47,48]. So it is likely that some spontaneous mobilization of VSELs will be happening during acute radiation sickness. Murine VSELs are highly radiation-resistant relative to a general population of hematopoietic stem cells, tolerating 1 Gy of γ radiation

and retaining *ex vivo* pluripotent differentiating activity (Figure 1)^[43]. Also important is that the *ex vivo* expansion of VSELs requires only 5-10 d in culture^[43]. Barriers that will need to be overcome to use VSELs to treat acute radiation sickness include: confirming radio-resistant characteristics of VSELs in humans; confirming that VSEL expansion using growth media doesn't activate oncogenes; and scaling up production for mass casualties.

CONCLUSION

Bone Marrow reconstitution as a partial treatment for acute radiation sickness has developed significantly since bone marrow transplantation was utilized for Chernobyl disaster victims in 1986. Use of autologous bone marrow or mobilized and harvested hematopoietic stem cells should eliminate the risk of graft-vs-host disease. The potential of autologous sourced stem cells is being evaluated now. Autologous cell sources include induced hematopoietic stem cells, induced pluripotent stem cells from adult differentiated tissue, MSC from bone marrow, myeloid progenitor cells from bone marrow, and VSEL stem cells from peripheral blood. Autologous human VSELs are emerging as fully functional stem cells that not only have wide-ranging regenerative competence, but have the critically important attribute of radiation resistance. The ultimate goal will be utilizing autologous, expanded stem cell infusions that would reconstitute many of the tissues damaged by radiation exposure.

REFERENCES

- 1 **Jarrett DG**. Medical aspects of ionizing radiation weapons. *Mil Med* 2001; **166**: 6-8
- 2 **Armed Forces Radiobiology Research Institute**. Medical management of radiological casualties. Available from: URL: <http://www.usuhs.mil/afri/outreach/pdf/3edmmrhandbook.pdf>
- 3 **Koenig KL**, Goans RE, Hatchett RJ, Mettler FA, Schumacher TA, Noji EK, Jarrett DG. Medical treatment of radiological casualties: current concepts. *Ann Emerg Med* 2005; **45**: 643-652
- 4 **Corey GR**. A brief review of the accident at Three Mile Island. IAEA Expert Report. Available from: URL: <http://www.iaea.org/Publications/Magazines/Bulletin/Bull215/21502795459.pdf>
- 5 **National Museum of American History**. Three mile island: The inside story. Available from: URL: <http://americanhistory.si.edu/tmi/tmi04.htm>
- 6 **Nesterenko AB**, Nesterenko VB, Yablokov AV. Consequences of the chernobyl catastrophe for public health. *Ann N Y Acad Sci* 2009; **1181**: 31-220
- 7 **Nesterenko AV**, Nesterenko AB, Yablokov AV. The difficult truth about chernobyl. *Ann N Y Acad Sci* 2009; **1181**: 1-3
- 8 **Baranov A**, Gale RP, Guskova A, Piatkin E, Selidovkin G, Muravyova L, Champlin RE, Danilova N, Yevseeva L, Petrosyan L. Bone marrow transplantation after the Chernobyl nuclear accident. *N Engl J Med* 1989; **321**: 205-212
- 9 **International Atomic Energy Agency**. Fukushima Nuclear Accident: Radiological Monitoring and Consequences. June 2, 2011. Available from: URL: <http://www.iaea.org/news-center/news/tsunamiupdate01.html/>
- 10 **Jolly D**, Tabuchi H, Bradsher K. Tainted water at two reactors increases alarm for Japanese. The New York Times, March 27, 2011: A1
- 11 **Dainiak N**, Gent RN, Carr Z, Schneider R, Bader J, Buglova E, Chao N, Coleman CN, Ganser A, Gorin C, Hauer-Jensen M, Huff LA, Lillis-Hearne P, Maekawa K, Nemhauser J, Powles R, Schünemann H, Shapiro A, Stenke L, Valverde N, Weinstock D, White D, Albanese J, Meineke V. First global consensus for evidence-based management of the hematopoietic syndrome resulting from exposure to ionizing radiation. *Disaster Med Public Health Prep* 2011; **5**: 202-212
- 12 **Solomon F** and Marsdon RQ. The Medical Implications of Nuclear War. Institute of Medicine, United States National Academy of Sciences. Available from: URL: http://www.nap.edu/openbook.php?record_id=940&page=R1
- 13 **United States Department of Health and Human Services**. ASBR Playbook. Available from: URL: <http://www.phe.gov/Preparedness/planning/playbooks/stateandlocal/nuclear/Pages/default.aspx>
- 14 **Demographia World Urban Areas**. July 2012. Available from: URL: <http://www.demographia.com/db-worldua.pdf>
- 15 **Be The Match Press Release**. July 9, 2010. The NMDP Partners with the Congressional Black Caucus Foundation for African American Bone Marrow Awareness Month to Increase Awareness about Bone Marrow and Cord Blood Donation. Available from: URL: http://marrow.org/News/News_Releases/2010/African_American_Marrow_Month.aspx
- 16 **Koh LP**, Chen CS, Tai BC, Hwang WY, Tan LK, Ng HY, Linn YC, Koh MB, Goh YT, Tan B, Lim S, Lee YM, Tan KW, Liu TC, Ng HJ, Loh YS, Mow BM, Tan DC, Tan PH. Impact of postgrafting immunosuppressive regimens on nonrelapse mortality and survival after nonmyeloablative allogeneic hematopoietic stem cell transplant using the fludarabine and low-dose total-body irradiation 200-cGy. *Biol Blood Marrow Transplant* 2007; **13**: 790-805
- 17 **Pecora AL**, Stiff P, Jennis A, Goldberg S, Rosenbluth R, Price P, Goltry KL, Douville J, Armstrong RD, Smith AK, Preti RA. Prompt and durable engraftment in two older adult patients with high risk chronic myelogenous leukemia (CML) using ex vivo expanded and unmanipulated unrelated umbilical cord blood. *Bone Marrow Transplant* 2000; **25**: 797-799
- 18 **Pecora AL**, Stiff P, LeMaistre CF, Bayer R, Bachier C, Goldberg SL, Parthasarathy M, Jennis AA, Smith AK, Douville J, Chen B, Armstrong RD, Mandalam RK, Preti R. A phase II trial evaluating the safety and effectiveness of the Aastrom-Replicell system for augmentation of low-dose blood stem cell transplantation. *Bone Marrow Transplant* 2001; **28**: 295-303
- 19 **Stiff P**, Chen B, Franklin W, Oldenberg D, Hsi E, Bayer R, Shpall E, Douville J, Mandalam R, Malhotra D, Muller T, Armstrong RD, Smith A. Autologous transplantation of ex vivo expanded bone marrow cells grown from small aliquots after high-dose chemotherapy for breast cancer. *Blood* 2000; **95**: 2169-2174
- 20 **Waterman J**, Rybicki L, Bolwell B, Copelan E, Pohlman B, Sweetenham J, Dean R, Sobecks R, Andresen S, Kalaycio M. Fludarabine as a risk factor for poor stem cell harvest, treatment-related MDS and AML in follicular lymphoma patients after autologous hematopoietic cell transplantation. *Bone Marrow Transplant* 2012; **47**: 488-493
- 21 **Ford CD**, Green W, Warenski S, Petersen FB. Effect of prior chemotherapy on hematopoietic stem cell mobilization. *Bone Marrow Transplant* 2004; **33**: 901-905
- 22 **Leupin N**, Schuller JC, Solenthaler M, Heim D, Rovo A, Beretta K, Gregor M, Bargetzi MJ, Brauchli P, Himmelmann A, Hanselmann S, Zenhäusern R. Efficacy of rituximab and cladribine in patients with chronic lymphocytic leukemia and feasibility of stem cell mobilization: a prospective multicenter phase II trial (protocol SAKK 34/02). *Leuk Lymphoma* 2010; **51**: 613-619
- 23 **Roncon S**, Barbosa IL, Campilho F, Lopes SM, Campos A, Carvalhais A. Mobilization and collection of peripheral

- blood stem cells in multiple myeloma patients older than 65 years. *Transplant Proc* 2011; **43**: 244-246
- 24 **Moog R.** Peripheral blood stem cell collection in children: Management, techniques and safety. *Transfus Apher Sci* 2010; **43**: 203-205
- 25 Mozobil (plerixafor) US Package Insert. Genzyme Corporation 2010. Available from: URL: <http://www.genzyme.com/contact-us.aspx>
- 26 **Brunner S, Zaruba MM, Huber B, David R, Vallaster M, Assmann G, Mueller-Hoecker J, Franz WM.** Parathyroid hormone effectively induces mobilization of progenitor cells without depletion of bone marrow. *Exp Hematol* 2008; **36**: 1157-1166
- 27 **DiCarlo AL, Poncz M, Cassatt DR, Shah JR, Czarniecki CW, Maidment BW.** Medical countermeasures for platelet regeneration after radiation exposure. Report of a workshop and guided discussion sponsored by the National Institute of Allergy and Infectious Diseases, Bethesda, MD, March 22-23, 2010. *Radiat Res* 2011; **176**: e0001-e0015
- 28 **Burdelya LG, Krivokrysenko VI, Tallant TC, Strom E, Gleiberman AS, Gupta D, Kurnasov OV, Fort FL, Osterman AL, Didonato JA, Feinstein E, Gudkov AV.** An agonist of toll-like receptor 5 has radioprotective activity in mouse and primate models. *Science* 2008; **320**: 226-230
- 29 **Brunner S, Theiss HD, Murr A, Negele T, Franz WM.** Primary hyperparathyroidism is associated with increased circulating bone marrow-derived progenitor cells. *Am J Physiol Endocrinol Metab* 2007; **293**: E1670-E1675
- 30 **Coppolino G, Bolignano D, De Paola L, Giulino C, Mannella A, Riccio M, Mascaro MA, Lombardi G, Fuiano G, Lombardi L, Buemi M.** Parathyroid hormone and mobilization of circulating bone marrow-derived cells in uremic patients. *J Investig Med* 2011; **59**: 823-828
- 31 **Singh VK, Brown DS, Kao TC.** Alpha-tocopherol succinate protects mice from gamma-radiation by induction of granulocyte-colony stimulating factor. *Int J Radiat Biol* 2010; **86**: 12-21
- 32 **Okita K, Yamanaka S.** Induced pluripotent stem cells: opportunities and challenges. *Philos Trans R Soc Lond B Biol Sci* 2011; **366**: 2198-2207
- 33 **Ye L, Chang JC, Lin C, Sun X, Yu J, Kan YW.** Induced pluripotent stem cells offer new approach to therapy in thalassemia and sickle cell anemia and option in prenatal diagnosis in genetic diseases. *Proc Natl Acad Sci USA* 2009; **106**: 9826-9830
- 34 **Feng Q, Lu SJ, Klimanskaya I, Gomes I, Kim D, Chung Y, Honig GR, Kim KS, Lanza R.** Hemangioblastic derivatives from human induced pluripotent stem cells exhibit limited expansion and early senescence. *Stem Cells* 2010; **28**: 704-712
- 35 **Zhao T, Zhang ZN, Rong Z, Xu Y.** Immunogenicity of induced pluripotent stem cells. *Nature* 2011; **474**: 212-215
- 36 **Mouisseddine M, François S, Semont A, Sache A, Allenet B, Mathieu N, Frick J, Thierry D, Chapel A.** Human mesenchymal stem cells home specifically to radiation-injured tissues in a non-obese diabetes/severe combined immunodeficiency mouse model. *Br J Radiol* 2007; **80 Spec No 1**: S49-S55
- 37 **Hu KX, Sun QY, Guo M, Ai HS.** The radiation protection and therapy effects of mesenchymal stem cells in mice with acute radiation injury. *Br J Radiol* 2010; **83**: 52-58
- 38 **Lange C, Brunswig-Spickenheier B, Cappallo-Obermann H, Eggert K, Gehling UM, Rudolph C, Schlegelberger B, Cornils K, Zustin J, Spiess AN, Zander AR.** Radiation rescue: mesenchymal stromal cells protect from lethal irradiation. *PLoS One* 2011; **6**: e14486
- 39 **Heider A.** Murine bone marrow derived very small embryonic-like (VSEL) stem cells give rise to mesenchymal stromal cells. Proc, ISSCR. Annual Meeting, Toronto, Canada, June 16-20, 2011. Abstract 2519
- 40 http://www.osiristx.com/prod_ars.php and <http://www.osiristx.com/clinical.php>
- 41 http://www.cellera.com/tech_clt008_rad.htm
- 42 **Rodgerson DO, Harris AG.** A comparison of stem cells for therapeutic use. *Stem Cell Rev* 2011; **7**: 782-796
- 43 **Ratajczak J, Wysoczynski M, Zuba-Surma E, Wan W, Kucia M, Yoder MC, Ratajczak MZ.** Adult murine bone marrow-derived very small embryonic-like stem cells differentiate into the hematopoietic lineage after coculture over OP9 stromal cells. *Exp Hematol* 2011; **39**: 225-237
- 44 **Kassmer SH, Bruscia E, Zhang PX, Krause DS.** Bone marrow derived lung epithelial cells are derived predominantly from nonhematopoietic cells. *Stem Cells* 2012; **3**: 491-499
- 45 **Sovalat H, Scrofani M, Eidenschenk A, Pasquet S, Rimelen V, Hénon P.** Identification and isolation from either adult human bone marrow or G-CSF-mobilized peripheral blood of CD34(+)/CD133(+)/CXCR4(+)/ Lin(-)CD45(-) cells, featuring morphological, molecular, and phenotypic characteristics of very small embryonic-like (VSEL) stem cells. *Exp Hematol* 2011; **39**: 495-505
- 46 **Drukala J, Paczkowska E, Kucia M, Młyńska E, Krajewski A, Machaliński B, Madeja Z, Ratajczak MZ.** Stem cells, including a population of very small embryonic-like stem cells, are mobilized into peripheral blood in patients after skin burn injury. *Stem Cell Rev* 2012; **8**: 184-194
- 47 **Wojakowski W, Tendera M, Kucia M, Zuba-Surma E, Paczkowska E, Ciosek J, Hałasa M, Król M, Kazmierski M, Buszman P, Ochała A, Ratajczak J, Machaliński B, Ratajczak MZ.** Mobilization of bone marrow-derived Oct-4+ SSEA-4+ very small embryonic-like stem cells in patients with acute myocardial infarction. *J Am Coll Cardiol* 2009; **53**: 1-9
- 48 **Paczkowska E, Kucia M, Koziarska D, Halasa M, Safranow K, Masiuk M, Karbicka A, Nowik M, Nowacki P, Ratajczak MZ, Machalinski B.** Clinical evidence that very small embryonic-like stem cells are mobilized into peripheral blood in patients after stroke. *Stroke* 2009; **40**: 1237-1244
- 49 **Rubin P, Casarett GW.** A direction for clinical radiation pathology: The tolerance dose. In: Vaeth JM, editor. *Frontiers of Radiation Therapy and Oncology*. Basel: Karger, 1971: 1-16
- 50 ICRP, 1984. Nonstochastic effects of ionizing radiation. ICRP Publication 41. *Ann. ICRP* 14 (3)
- 51 **Phillips TL.** Radiation injury. In: Wyngaarden JB, Smith LH Jr, Bennett JC, editors. *Cecil Textbook of Medicine*, 19th ed. Philadelphia: WB Saunders, 1992: 2354

S- Editor Li JY L- Editor A E- Editor Zheng XM

Natural compounds as anticancer agents: Experimental evidence

Jiao Wang, Yang-Fu Jiang

Jiao Wang, School of Basic Medicine, Chengdu University of Traditional Chinese Medicine, Chengdu 610075, Sichuan Province, China

Yang-Fu Jiang, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

Author contributions: Wang J and Jiang YF equally contributed to this paper.

Supported by National Natural Science Foundation of China, No. 81001587

Correspondence to: Yang-Fu Jiang, Professor, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China. jyangfu@scu.edu.cn

Telephone: +86-28-85164044 Fax: +86-28-85164045

Received: February 21, 2012 Revised: June 6, 2012

Accepted: June 15, 2012

Published online: June 20, 2012

© 2012 Baishideng. All rights reserved.

Key words: Cancer; Chemoprevention; Natural agents; (-)-Epigallocatechin-3-gallate; Resveratrol; Curcumin; Vitamin D; Indole-3-carbinol

Peer reviewer: Umberto Galderisi, PhD, Associate Professor of Molecular Biology, Department Experimental Medicine, Second University of Naples, Via L. De Crecchio 7, 80138 Napoli, Italy

Wang J, Jiang YF. Natural compounds as anticancer agents: Experimental evidence. *World J Exp Med* 2012; 2(3): 45-57 Available from: URL: <http://www.wjgnet.com/2220-315X/full/v2/i3/45.htm> DOI: <http://dx.doi.org/10.5493/wjem.v2.i3.45>

Abstract

Cancer prevention research has drawn much attention worldwide. It is believed that some types of cancer can be prevented by following a healthy life style. Cancer chemoprevention by either natural or synthetic agents is a promising route towards lowering cancer incidence. In recent years, the concept of cancer chemoprevention has evolved greatly. Experimental studies in animal models demonstrate that the reversal or suppression of premalignant lesions by chemopreventive agents is achievable. Natural occurring agents such as dietary phytochemicals, tea polyphenols and resveratrol show chemopreventive activity in animal models. Moreover, clinical trials for testing the safety and efficacy of a variety of natural agents in preventing or treating human malignancy have been ongoing. Here, we summarize experimental data on the chemopreventive or tumor suppressive effects of several natural compounds including curcumin, (-)-epigallocatechin-3-gallate, resveratrol, indole-3-carbinol, and vitamin D.

INTRODUCTION

Cancer is common disease that limits lifespan. Many factors including life style, genetic variation, virus infection and chronic inflammation may affect the susceptibility to cancer. In the past decades, both the diagnosis and treatment of malignant tumors are improving. In addition to traditional treatments such as chemotherapy and radiotherapy, molecular targeted therapy is emerging as a promising trend for cancer therapeutics. For those who are at high risk for cancer, chemoprevention may be an alternative intervention to inhibit or delay carcinogenesis. While a number of chemotherapeutic agents have been administered in the clinic for many years, there is still long way to go for chemopreventive agents to be safely and effectively administered in human populations. The identification of chemopreventive targets and biomarkers that can help monitoring their effectiveness are a huge challenge. In addition to synthetic compounds, many natural products have been found to be able to inhibit carcinogenesis, at least in animal models. There are many ongoing clinical trails to test the safety and efficacy of natural agents in preventing or treating cancer (Table 1). It is highly likely that natural agents can be used for cancer prevention

without recognizable adverse effects. Here, we highlight the experimental evidence concerning some natural agents that exhibit protective effect against cancer. The sources of some natural compounds are shown in Figure 1.

CURCUMIN

Curcumin, a polyphenolic molecule isolated from the roots (rhizomes) of the plant *Curcuma longa*, is a promising compound for cancer chemoprevention and therapy^[1,2]. Although *Curcuma longa* and its chemical components have been used in Chinese and Hindu medicine for thousands of years, curcumin has attracted much attention in recent decades because of its anticancer activity. The beneficial effects of curcumin include anti-oxidant, anti-inflammatory, anti-proliferative, and anti-angiogenic properties^[1,3]. Many preclinical studies of curcumin have shown anti-carcinogenic and therapeutic effects in various tumor cell lines and xenograft models. The anticancer efficacy of curcumin is well established in a range of animal cancer models including those associated with colon, breast, pancreas, lung, kidney, bladder, blood and skin^[1]. It is worth mentioning that curcumin has the ability to kill cancer cells selectively without apparent toxicity to nonmalignant cells, a good property for a cancer-preventive candidate^[4]. Extensive toxicological screening and preclinical investigation showed minimal adverse effects of curcumin administration in mice, rats, dogs, and monkeys^[5]. Phase I and phase II clinical trials have already demonstrated the safety of curcumin even at high doses (8-12 g/d) over several months. Adverse events were mainly nausea and diarrhea^[6-9].

Regardless of its excellent safety profile, the poor solubility and low bioavailability of curcumin are obstacles to therapeutic drug development. Data on the pharmacokinetics, metabolites, and systemic bioavailability of curcumin in rodents and humans show that curcumin is poorly absorbed, rapidly metabolized, and may have limited systemic bioavailability^[10,11]. In fact, after oral administration of curcumin, very low concentrations of curcumin or corresponding metabolites are found in patient serum and tissues outside the gastrointestinal tract^[8,12,13]. Compared to the effective concentrations *in vitro* (5-50 $\mu\text{mol/L}$), the poor absorption and bioavailability of curcumin suggest that its anticancer effects may be limited *in vivo*. Some approaches to improving the bioavailability of curcumin have been investigated, including the combination with adjuvants, the use of chemical analogues and novel delivery methods^[14,15].

It is interesting that studies in animals still show curcumin is an effective agent for several cancer models, in spite of its limited bioavailability. It is not clear whether this efficacy comes from unmeasured curcumin metabolites, or for other unknown indirect effects. The pharmacodynamic data in humans is still limited. Currently, several phase I and phase II clinical trials are ongoing to investigate the benefits of curcumin as a chemopreventive and chemotherapeutic agent in a variety of cancers,

including multiple myeloma, pancreatic cancer, breast cancer and colon cancer (www.clinicaltrials.gov). According to reported data from these ongoing studies, some results are promising. Results from a phase I clinical study of twenty-five patients with various pre-malignant or high-risk lesions suggested that oral curcumin may have chemopreventive effects on these lesions (histological improvement)^[8]. In another study from the Cleveland Clinic in Florida, five patients with familial adenomatous polyposis were treated with a combination of curcumin and quercetin three times a day for a mean duration of 6 mo. The numbers and sizes of polyps were reduced in all patients compared to baseline values^[16]. A recent phase II clinical trial reported by Carroll and colleagues investigated curcumin's potential activities for prevention of colorectal neoplasia in smokers with aberrant crypt foci (ACF). The data showed a significant reduction of ACF number by curcumin at the 4-g dose level and indicated that curcumin may have cancer-preventive effects in the setting of early pre-invasive neoplastic lesions. Interpretation of this study, however, was limited by the remaining controversy around ACF as a biomarker of colon carcinogenesis, nonrandomization and lack of placebo group^[17].

Curcumin efficacy in the treatment of human pancreatic cancer has been reported in a phase II clinical trial in patients with advanced disease. Patients received 8 g curcumin by mouth daily until disease progression, with restaging every 2 mo. Twenty-one of twenty-five patients were evaluable for response. Low concentrations of curcumin were able to elicit a biological effect by down-regulating the expression of nuclear factor κB (NF- κB), cyclooxygenase (COX)-2 and phosphorylated signal transducer and activator of transcription 3 in peripheral blood mononuclear cells derived from patients^[9]. A further phase II clinical studies suggested that a combination of gemcitabine and curcumin is a feasible treatment for patients with pancreatic cancer^[18]. Bayet-Robert *et al.*^[19] treated 14 advanced and metastatic breast cancer patients with a combination of curcumin and docetaxel. The study demonstrated that the combination therapy decreased the vascular endothelial growth factor (VEGF) levels and showed encouraging efficacy. Hopefully, there will soon be more data to demonstrate the anticancer effects of curcumin, especially measurements which confirm the mechanism-based molecular targets which are really implicated *in vivo*.

(-)-EPIGALLOCATECHIN-3-GALLATE

Tea, from the plant *Camellia sinensis*, is one of the most popular beverages consumed worldwide. The most abundant chemical compound in green tea is catechins, which include (-)-epigallocatechin-3-gallate (EGCG), (-)-epigallocatechin, (-)-epicatechin-3-gallate and (-)-epicatechin. Among these, EGCG accounts for more than 50% of the total catechins, and appears to be the most effective and best-studied constituent of green tea^[20].

EGCG holds considerable promise for chemoprevention according to epidemiological, cell culture, animal

Table 1 Selected ongoing clinical trials with natural anti-cancer compounds

Agent	Phase	Status	Trial type	Conditions/cancer type	Combination	Trial No.
Curcumin	Phase II	Recruiting	Prevention	Familial adenomatous polyposis		NCT00641147
	Phase I	Completed	Prevention	Healthy		NCT00027495
	Phase I	Recruiting	Treatment	Advanced osteosarcoma		NCT00689195
	Phase II	Recruiting	Treatment	Advanced pancreatic cancer		NCT00094445
Tea polyphenols	Phase II	Recruiting	Prevention	Postmenopausal women with high breast density		NCT00917735
	Phase II	Recruiting	Prevention	Tobacco use disorder		NCT00611650
	Phase I	Recruiting	Prevention	Premalignant lesions of the head and neck	Erlotinib	NCT01116336
	Phase I	Recruiting	Treatment	Small cell lung carcinoma		NCT01317953
	Phase II	Recruiting	Treatment	Multiple myeloma and plasma cell neoplasm		NCT00942422
	Phase II	Completed	Treatment	Bladder cancer		NCT00088946
Indole-3-carbinol/ 3,3-diindolylmethane	Phase II	Completed	Prevention	Healthy men at risk for prostate cancer progression		NCT00579332
	Phase III	Recruiting	Prevention	Women with a BRCA1 mutation		NCT01022333
	Phase I	Completed	Treatment	Prostate cancer	Radical prostatectomy	NCT00450229
	Phase II	Recruiting	Treatment	Breast cancer		NCT01391689
Vitamin D	Phase II	Completed	Observation of the relationship between vitamin D level and thyroid cancer	Thyroid cancer		NCT00719615
	Phase II	Recruiting	Prevention	Postmenopausal women at high risk for breast cancer		NCT00859651
	Phase III	Enrolling by invitation	Prevention	Adenomatous colon polyps	Calcium	NCT00399607
	Phase I	Recruiting	Treatment	Metastatic melanoma	Temozolomide	NCT00301067
	Phase II	Completed	Treatment	Metastatic or locally advanced pancreatic cancer	Docetaxel	NCT00238199
	Phase IV	Completed	Prevention of osteoporosis	Postmenopausal breast cancer survivors	Risedronate; calcium	NCT00567606

Data from the United States National Institutes of Health, <http://www.clinicaltrials.gov/>.

and clinical studies. EGCG has been shown to cause growth inhibition and apoptosis in a number of human cancer cell lines *in vitro* and inhibit tumor incidence and multiplicity in animal models, such as liver, colon, prostate, pancreas, mammary glands, lung and skin cancer models^[21]. The mechanisms underlying EGCG anticancer effects include anti-oxidant activities, modification of carcinogen metabolism, prevention of DNA damage, induction of cell cycle arrest and apoptosis, inhibition of metastasis, proteasome inhibition and modulation of multiple signal transduction pathways [epidermal growth factor receptor, human epidermal growth factor receptor 2, VEGF receptor, insulin-like growth factor (IGF)1R, phosphoinositide 3-kinase/AKT, mitogen-activated protein kinase and NF- κ B signaling]^[22]. Abundant evidence from animal cancer models has demonstrated the strong chemopreventive effects of EGCG. As early as 1987, Yoshizawa *et al.*^[23] reported that the application of EGCG suppressed 7,12-dimethylbenz[a]anthracene (DMBA) plus teleocidin-initiated carcinogenesis. EGCG significantly decreased tumor incidence and burden per mouse compared with controls. The subsequent studies showed EGCG to have a broad spectrum against carcinogens. In the animal model, EGCG effectively inhibited

(4-methylnitro-samino)-1-(3-pyridyl)-1-butanone (NNK) and benzo(a)pyrene-induced lung cancer, azoxymethane-induced ACF and colon tumors, NNK and diethylnitrosamine-induced liver tumors, UV-induced skin cancer, and N-butyl-N-(4-hydroxybutyl)-nitrosamine-induced urinary bladder tumors^[24].

Despite the impressive anti-tumor effect of EGCG in animal, the available epidemiological evidence on tea consumption and cancer prevention in humans has not yielded conclusive results. The inconsistent results of epidemiological studies were probably due to various confounding factors. The quantity and quality of the tea consumed will definitely affect the outcome of epidemiological studies. In addition, the effect of caffeine in tea, large intersubject and intrasubject variability could be additional contributing factors to the inconsistency^[24]. Furthermore, the concentrations of EGCG in plasma and tissue after simply drinking green tea is low compared to the effective concentrations used in cell culture experiments (10-100 μ mol/L)^[20]. To minimize the confounding effects, more potent tools were used in later well-designed clinical intervention studies, including a defined green tea catechin (GTC) extract, EGCG-enriched fractions such as Polyphenone E (the EGCG content about

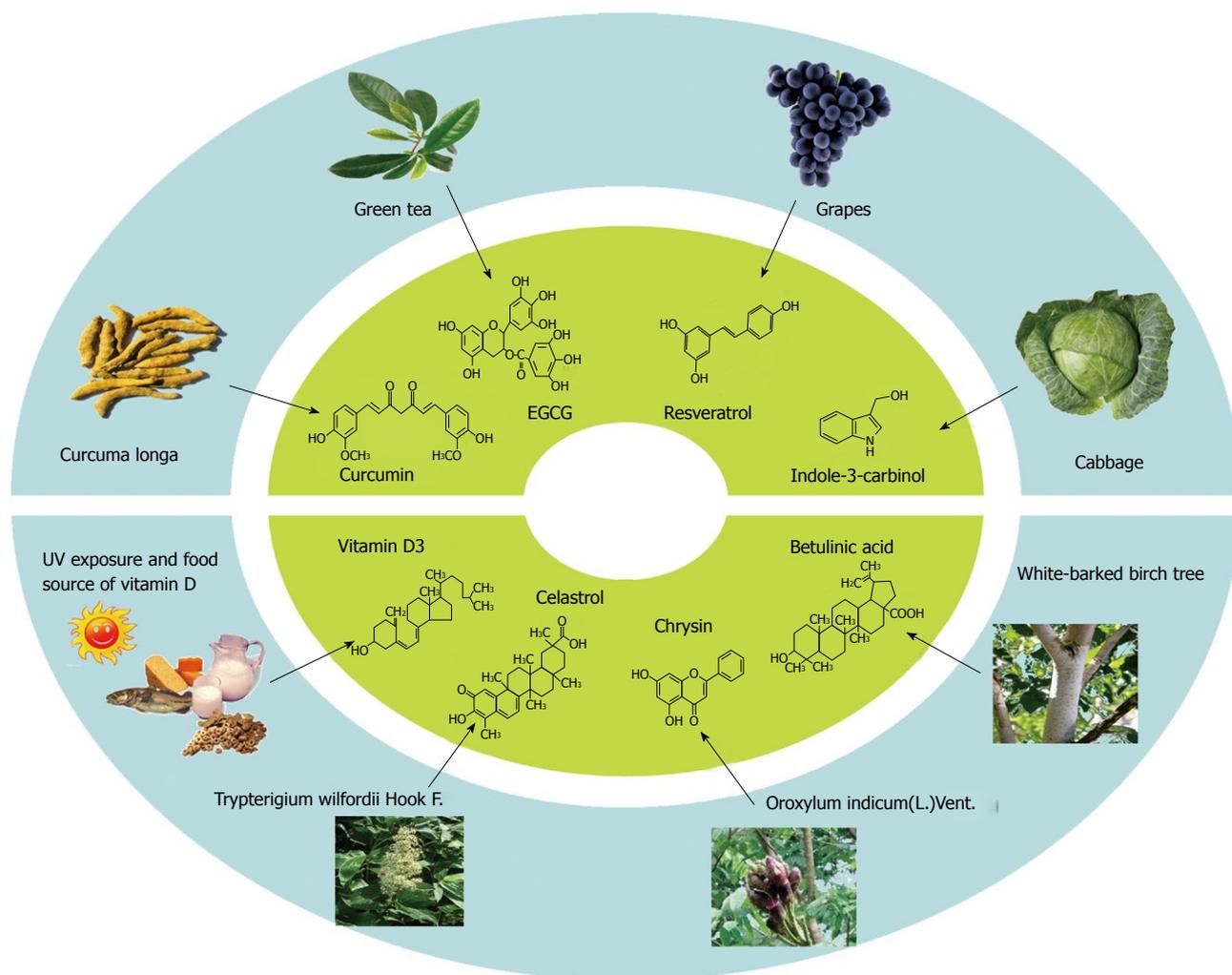


Figure 1 The sources of natural anti-cancer agents, including curcumin, (-)-epigallocatechin-3-gallate, resveratrol, indole-3-carbinol, and vitamin D, chrysin, celastrol and betulinic acid.

70%) and highly purified EGCG provided by a pharmaceutical company.

Several studies of the systemic bioavailability of orally administered catechins in human volunteers have been conducted. Chow *et al.*^[25,26] conducted several studies to examine the safety, tolerability, and pharmacokinetic properties of single and multiple dose administration of EGCG and Polyphenon E from 200 to 800 mg. Their studies showed that the oral bioavailability of tea polyphenols in humans was low. Oral administration of EGCG and Polyphenon E at the same dose level (based on EGCG content) resulted in similar plasma EGCG levels. However, the repeated administration of 800 mg of green tea polyphenols once daily for 4 wk resulted in a 60% increase in the systemic availability of free EGCG, which may be due to inhibition of presystemic elimination of this catechin. The majority of clinical studies demonstrate the safety and limited side effects of EGCG. However, a recent review suggested a causal association between green tea and liver damage. This hepatotoxicity may be attributed to EGCG or its metabolites which, under particular conditions related to the patient's metabo-

lism, can induce oxidative stress in the liver^[27].

Some data are currently available from EGCG chemoprevention and chemotherapy trials, which offer us more details of EGCG action in the human body. Ahn and coworkers reported that oral treatment of polyphenon E or purified EGCG, 200 mg daily for 12 wk, was effective in patients with human papilloma virus (HPV)-infected cervical lesions^[28]. A pilot study conducted in Japan, investigated the effect of green tea extract (GTE) on metachronous colorectal adenomas. Oral administration of GTE, 1.5 g/d for 12 mo, in addition to a tea drinking life-style, showed efficacy in preventing the incidence of metachronous adenoma in patients 1 year postpolypectomy. The incidence of metachronous colorectal adenomas at end-point colonoscopy was 31% (20 of 65) in the control group and 15% (9 of 60) in the GTE group^[29]. Another encouraging clinical trial investigated possible prostate cancer chemoprevention with oral GTCs. Sixty volunteers with high-grade prostate intraepithelial neoplasia received either 600 mg of GTCs or placebo daily for 12 mo. The primary end point was prevalence of prostate cancer. After 1 year of follow-up, only 3% of

the patients in the treatment group developed prostate cancer, compared with 30% in the placebo group^[30]. In a 2-year follow-up, despite the high drop-out rate (57% in the placebo-arm and 55% in the GTCs-arm), three further cancer diagnoses appeared. One prostate cancer was diagnosed among 13 GTC-treated patients and 2 among 9 placebo-treated patients. Overall, treatment with GTCs led to almost 80% reduction in prostate cancer diagnosis. These results suggest that the inhibition of prostate cancer progression after 1 year of GTCs administration was long-lasting and no adverse effect was associated with the treatment^[31].

The results from clinical trials with EGCG are not all positive. A phase I study performed in 49 patients with various tumours reported no major antitumor responses when using GTE at the maximum-tolerated dose of 4.2 g/m² once daily or 1.0 g/m² three times day^[32]. In a Phase II study, green tea showed minimal anti-neoplastic activity, as defined by a decline in prostate specific antigen (PSA) levels, among 42 patients with androgen independent prostate carcinoma. Only a single patient manifested a 50% decrease in PSA level from baseline and this response was not sustained beyond 2 mo. Green tea toxicity occurred in 69% of patients and included nausea, emesis, insomnia, fatigue, diarrhea, abdominal pain, and confusion^[33]. On the other hand, other therapeutic trials with EGCG have shown promising results. A recent phase II clinical trial demonstrated the effects of short-term supplementation with Polyphenon E on serum biomarkers in prostate cancer patients. Twenty-six men were given daily doses of Polyphenon E (total of 800 mg EGCG) before radical prostatectomy (average drug administration of 6 wk). Polyphenon E administration significantly reduced serum levels of hepatocyte growth factor, VEGF, PSA, IGF-I, IGF binding protein (IGFBP)-3, and the IGF-I/IGFBP-3 ratio with no adverse effects on liver function. These findings support a potential role for Polyphenon E in the treatment or prevention of prostate cancer^[34]. Shanafelt *et al.*^[35] reported that EGCG induced apoptotic cell death in the leukemic B-cells from a majority of patients with chronic lymphocytic leukemia (CLL), and four patients with low grade B-cell malignancies developed positive responses shortly after self-initiating EGCG therapy by oral ingestion of EGCG containing products. Based on this evidence, the same group conducted a phase I trial to define the clinical benefit of Polyphenon E. Thirty-three previously untreated patients with asymptomatic Rai stage 0 to II CLL received Polyphenon E treatment. Declines in absolute lymphocyte count (ALC) and/or lymphadenopathy were observed in the majority of patients. One patient achieved partial remission and more than 50% of study patients attained a sustained decline in ALC of 20% and a 50% reduction in lymphadenopathy at some point during treatment. No differences in response were observed based on IgVH, ZAP-70, or CD38 gene mutation status except for trisomy 12^[36]. Furthermore, this research group is conducting an ongoing phase II trial to evaluate

efficacy of Polyphenon E at 2000 mg dose, twice a day in patients with asymptomatic Rai stage 0 to II CLL (www.clinicaltrials.gov).

While EGCG alone is active in suppressing cancer, combination of EGCG with other agents may be more promising. EGCG reportedly exhibits synergistic effects with other anti-cancer drugs, such as curcumin, chrysin, tamoxifen, etoposide, 5-fluorouracil, temozolomide, taxane, erlotinib^[37-43]. Consequently, several clinical trials of EGCG in combination with other drugs for cancer treatment are now ongoing. On the other hand, recent studies indicate that EGCG may be able to block the therapeutic efficacy of some anticancer agents such as bortezomib and other boronic acid-based proteasome inhibitors. This should be highly relevant for clinical considerations^[44,45].

RESVERATROL

Resveratrol, a polyphenol, was first isolated in 1940 as an ingredient of the roots of white hellebore (*Veratrum grandiflorum* O.Loes) and has since been found in various food sources including red wine, grapes, mulberries, peanuts^[46,47]. Resveratrol was identified in 1963 as the active constituent of the roots of *Polygonum cuspidatum*, a plant used in Chinese and Japanese traditional medicine^[46]. Jang *et al.*^[48] reported the ability of resveratrol to inhibit carcinogenesis at multiple stages, including initiation, promotion and progression. Subsequent studies demonstrated the strong chemopreventive efficacy of resveratrol in many different animal models of carcinogenesis. Oral or local application of resveratrol in mice or rats significantly reduced DMBA-initiated and 12-O-tetradecanoylphorbol-13-acetate-promoted skin tumors, suppressed DMBA-induced mammary carcinogenesis, inhibited 1,2-dimethylhydrazine-induced colon carcinogenesis and N-nitrosomethylbenzylamine-induced esophageal tumors. Overall, the majority of these studies strongly support the chemopreventive effect of resveratrol, although there are exceptions in which a lack of *in vivo* benefit has been observed^[49-53].

Besides its chemopreventive effects, extensive study over the past decade has suggested that resveratrol might be a promising candidate for cancer therapy by interfering with many signaling pathways that regulate cell proliferation, apoptosis, inflammation, angiogenesis and metastasis^[46,47]. It suppresses the proliferation in a wide variety of human tumor cells *in vitro* and in xenograft models. Resveratrol was also reported to exhibit synergistic chemopreventive effects with other anti-cancer drugs, such as cisplatin, doxorubicin and vinorelbine^[54-56]. However, a recent study indicated that resveratrol can significantly attenuate the efficacy of paclitaxel's anticancer actions in certain human breast cancer cell lines both *in vitro* and *in vivo*^[57], suggesting that concomitant use of resveratrol with paclitaxel may be detrimental in certain types of human cancers. More preclinical and clinical testing of the potential benefits and risks of using resveratrol as an anticancer adjuvant in cancer patients is warranted.

The bioavailability and the pharmacokinetics of resveratrol have been studied in experimental animals and humans. These studies showed that resveratrol was rapidly absorbed after oral intake, and rapidly metabolized to glucuronide and sulphate conjugates which result in the low concentrations of resveratrol observed in plasma^[47]. The low bioavailability led to uncertainty over whether oral resveratrol can reach the bioactive concentrations in target tissues. The limited data from research about the tissue distribution of resveratrol and its metabolites offers us some clues. Data from mice demonstrated the significant accumulation of resveratrol in the intestine, stomach, liver, kidney and bile^[58,59]. In a clinical study, a level of resveratrol was found in colon tissue in excess of that required for activity *in vitro*, which supported the colon as a target organ for oral resveratrol in humans^[60].

A wide range of doses of resveratrol (0.1-1500 mg/kg) was used in animal studies with various efficacy and low toxicity, although more human studies are needed to establish the relevant dose for human use. To date, limited data was obtained from human studies performed with resveratrol, and it is difficult to compare the results concerning the safety and tolerability of resveratrol because of variations in conditions of administration (e.g., pure resveratrol formulation, or other non-pure resveratrol samples in various matrices). It was generally agreed by the expert panel of Resveratrol 2010 that at least some portion of the population is likely take 1-2 mg of resveratrol per day from dietary sources and at this amount is almost certain to be safe for chronic consumption^[61].

Two clinical studies in healthy volunteers investigated the cancer-preventive effect of resveratrol through examination of related biomarkers. A phase I study carried out in forty healthy volunteers showed that ingesting a range of doses of resveratrol (0.5, 1.0, 2.5 or 5.0 g daily) for 29 d caused a decrease in circulating IGF-1 and IGFBP-3, respectively, compared to pre-dosing values. At the 2.5 g dose level, the decrease was most marked. The observed decrease in circulating IGF-1 and IGFBP-3 may contribute to cancer chemopreventive activity. Several other potential markers of activity were also investigated in blood samples from the volunteers. Resveratrol neither significantly affected circulating levels of prostaglandin E-2 (PGE-2), reflecting perturbation of the arachidonic acid cascade, nor influenced leukocyte levels of the malondialdehyde-DNA adduct M1dG, reflecting DNA oxidation^[62]. Consistent with the evidence *in vitro* and in animal models, another clinical study performed in healthy volunteers showed that resveratrol intervention inhibited the phenotypic indices of CYP3A4, CYP2D6, and CYP2C9 and induced the phenotypic index of 1A2. In addition, in individuals with low baseline GST- π levels and UGT1A1 activity, intervention was associated with a significant increase in enzyme activity. Modulation of enzyme systems involved in carcinogen activation and detoxification could be one of the mechanisms responsible for the cancer preventive effect of resveratrol. However,

such activities may also alter the pharmacokinetics of other drugs. Therefore, the authors suggested that further clinical studies should consider evaluation of lower doses of resveratrol to minimize adverse metabolic drug interactions^[63].

Based on the evidence from animal studies which showed oral administration of resveratrol can efficiently induced apoptosis in colon cancer with high levels achievable in local tissue, most of the clinical therapeutic trials have focused on investigating the effects of resveratrol on colon cancer. The first reported clinical trial of resveratrol in patients with colon cancer was conducted to assess the effects of a low dose of a plant-derived resveratrol formulation and resveratrol-containing freeze-dried grape powder (GP) on biomarkers related to the Wnt pathway, a key signaling pathway activated in over 85% of colon cancers^[64]. Eight patients received 14 d of treatment until the day prior to surgery for colon cancer resection. Resveratrol and GP had significant activity in inhibiting Wnt targets on normal colonic mucosa, such as cyclin D1 and axin II. However, GP treatment increased the expression of some Wnt target genes in colon cancer, including myc and cyclin D1. Resveratrol may have more clinical utility for colon cancer prevention rather than for treatment of established colon cancer^[64]. In another clinical study, twenty patients with histologically confirmed colorectal cancer consumed 8 daily doses of resveratrol at 0.5 or 1.0 g prior to surgical resection. Resveratrol was found to be well tolerated. With respect to its activity in target tissues, resveratrol slightly inhibited cell proliferation in colorectal cancer tissue after ingestion, as assessed by Ki-67 immunostaining^[60]. A recent report of a phase I, randomised, double-blind clinical trial, described the effects of SRT501 (micronized resveratrol) in patients with colorectal cancer and hepatic metastases^[65]. Cleaved caspase-3, a marker of apoptosis, was significantly increased by 39% in malignant hepatic tissue following SRT501 treatment, compared to tissue from placebo-treated patients. However, SRT501 failed to change the levels of several other biomarkers associated with cell survival and apoptosis in plasma or in tumor tissues, including PGE-2, VEGF, IGF-1, Ki67, phospho-Akt (ser473), Akt1, phospho-GSK3, GSK3, phospho-extracellular signal-regulated kinase (ERK), ERK, phospho-JNK, JNK, β -catenin, survivin, BCL2, Bax or PARP^[65].

Cellular senescence is an anticancer mechanism that our organism may implement to arrest cancer cells. The arrest of senescence and inhibition of cancer growth appear to be two antagonistic activities. However, it is well documented that resveratrol has both anti-senescence and anti-cancer activities, indicating that it has complex roles in preventing ageing and carcinogenesis. In addition, the numerous formulations of resveratrol used in clinical research and its potential interactions with other drugs make it difficult to recommend an optimal dosage for clinical usage. Long-term clinical trials are needed to validate the anti-cancer effect of resveratrol when used as a drug or as food supplement.

INDOLE-3-CARBINOL

Indole-3-carbinol (I3C), an indole compound, is naturally found in many plants, particularly in cruciferous vegetables such as broccoli, cabbage, cauliflower, Brussels sprouts, and bok choy^[66,67]. Glucobrassicin, a major component of cruciferous vegetables, is hydrolyzed in acidic conditions to give I3C^[68]. I3C is chemically unstable in aqueous and gastric acidic environments, such as those encountered under cell culture conditions and the acidic environment of the stomach *in vivo*. In acidic conditions, I3C is rapidly converted to numerous condensation products, of which 3,3-diindolylmethane (DIM) is the most active and effective metabolite^[66,69]. The effect of I3C *in vivo* might, at least in part, be attributable to the formation of DIM.

To date, I3C and its metabolite, DIM, have been demonstrated in numerous epidemiological and preclinical studies to possess cancer preventive properties. *In vitro* studies demonstrated that both I3C and DIM inhibit growth of most types of hormone-dependent and -independent cancer cells (breast, prostate, liver, lung, colon, cervix, and ovarian cancers)^[66,70-72]. In addition, in *in vivo* studies, I3C and DIM have been shown to have pronounced chemopreventive effects against growth of both spontaneous and chemically induced cancers in various animal models^[73-77]. The anti-cancer properties of I3C is attributable to its ability to modulate multiple signaling pathways which control DNA repair, hormonal regulation, inflammation, cell division and growth, apoptosis, angiogenesis, and multiple drug resistance^[66,78,79]. I3C has been shown to induce phase 1 and phase 2 enzymes that metabolize carcinogens, prevent carcinogen-DNA adduct formation, regulate several nuclear transcription factors [such as estrogen receptor (ER), aryl hydrocarbon receptors (AhR, Sp1 and NF- κ B)], modulate anti-apoptotic and pro-apoptotic factors, repress extracellular matrix-degrading proteases, and reverse the process of epithelial mesenchymal transition *via* regulation of key miRNAs^[66,67,80-82]. Among these multiple mechanisms, the most important effect of I3C and DIM is modulation of estrogen metabolism. I3C and DIM has received special attention as an effective chemopreventive agent against hormone-dependent cancers such as breast, cervical and prostate cancers, for the most part, due to its ability to negatively regulate ER α signaling and alter cytochrome P450-mediated estrogen metabolism^[83,84]. Despite the low affinity of I3C for ER α , it significantly inhibits ER α activity thereby diminishing estrogen-mediated cellular and biochemical effects in estrogen-responsive cells and tissues^[85]. I3C could also induce ER protein ubiquitination and degradation in a process requiring AhR, which binds to a wide range of ligands including DIM^[86,87]. In addition, a recent study has identified DIM as a ligand-independent activator of ER β , a molecule associated with anti-proliferative activity in breast cancer cells^[88]. Moreover, I3C and DIM may reverse the metabolism of estradiol to a more beneficial pathway, thus reducing levels of toxic

16 α -OHE₁ and increasing levels of protective 2-OHE₁, which correlates with reduced risk of breast cancer and other cancers including cervical and prostate cancer^[89].

Several studies have been conducted to detect the pharmacokinetics of I3C. Upon oral administration of 250 mg/kg I3C to female CD-1 mice, I3C and its acid condensation products were absorbed and distributed systemically into a number of well-perfused tissues^[90]. In contrast, in human testing, no I3C was found in the plasma after giving a single dose of up to 1200 mg or multiple-doses at 400 mg administered twice daily for 4 wk, and DIM was the only detectable I3C-derived compound in plasma^[91]. These results support the concept that I3C may serve as the prodrug rather than the actual therapeutic agent. Most clinical data on I3C and DIM indicate a good safety profile and only minor adverse effects. Rosen *et al.*^[92] reported a long term clinical study using I3C for the treatment of recurrent respiratory papillomatosis. Among 11 patients having a complete response to I3C, the average number of months on I3C was 50.2 mo, and no immediate or long-term side effects were found. However, I3C also has been found to promote cancer of the liver in rats, raising some doubt its use^[93,94].

Both I3C and DIM have already undergone human clinical trials, most of them focused on investigating effects on hormone responsive cancers, including cervical dysplasia, breast cancer, vulvar intraepithelial neoplasia (VIN), and prostate cancer. Similar to the effects observed in animal models, several studies showed that I3C and DIM strongly affect estradiol metabolism in healthy humans^[95,96]. They were able to induce the activity of cytochrome P450 isoenzyme CYP1A2, and increase the ratio of 2-OHE₁:16 α -OHE₁. A similar beneficial shift in estrogen metabolites was also observed in early studies on women with increased risk for breast cancer^[97-99] and a more detailed study on postmenopausal women with a history of early-stage breast cancer^[100]. Based on the preclinical evidence which indicated that I3C and DIM may offer benefit for diseases caused by HPV, two studies explored the effect on cervical intraepithelial neoplasia (CIN). Bell and colleagues used I3C administered orally to treat women for CIN. Thirty patients with biopsy-proven CIN II-III were randomized to receive placebo or 200 or 400 mg/d I3C for 12 wk. There was a statistically significant regression of CIN in patients treated with oral I3C compared with placebo. In addition, the 2/16 α -hydroxyestrone ratio changed in a dose-dependent fashion^[101]. In another randomized clinical trial, oral DIM at a dose of 2 mg/kg per day for 12 wk was well tolerated with no significant toxicity and clinically significant improvement was demonstrated in patients with grade 2 or 3 CIN^[102]. Naik *et al.*^[103] reported the results of a randomized phase II trial of I3C in the treatment of VIN. In this study, 12 women were randomized to receive 200 or 400 mg/d of I3C, following histological confirmation of high-grade VIN. I3C administration led to a significant reduction in symptoms, lesion size, and severity as well as significant improvement in estrogen metabolism.

Recently, in a phase I clinical trial, Rajoria *et al.*^[104] recommended DIM as an anti-estrogenic dietary supplement to help reduce the risk of developing thyroid proliferative disease based on the fact that DIM was detected in thyroid tissue and that DIM supplementation significantly improved estrogen metabolism.

VITAMIN D

In addition to its originally identified role on calcium homeostasis and bone metabolism, vitamin D is being recognized as a steroid hormone which exerts a wide range of biological activities related to various clinical disorders including cancer^[105,106]. Vitamin D₂ is mainly produced by the irradiation of yeast or plant ergosterol^[106]. In humans, vitamin D₃ (cholecalciferol) is synthesized naturally in skin cell by exposing to ultraviolet B radiation in sunlight, the major source of vitamin D for most humans. In the skin, 7-dehydrocholesterol, a cholesterol precursor is converted to vitamin D₃. Vitamin D₃ is hydroxylated to 25-hydroxyvitamin D₃ (25-OH-D₃) in the liver by 25-hydroxylase and further converted to 1 α ,25-dihydroxycholecalciferol (1,25-(OH)₂D₃, calcitriol) by 1 α -hydroxylase in the kidney and other tissues. Calcitriol is mainly catabolized by 24-hydroxylase (CYP24A1) to 1 α ,24,25-(OH)₂D₃, removing its bioactivity^[107].

Most of the anticancer effects of 1,25-(OH)₂D₃ are mediated through binding to its specific receptor, the vitamin D receptor (VDR). In the cell nucleus, 1,25-(OH)₂D₃ binds to VDR, which subsequently heterodimerizes with another nuclear receptor, the retinoid X receptor^[106,108]. The heterodimer binds to vitamin D responsive element in target genes and initiates the regulation of specific genes, including those involved in the regulation of cell growth, differentiation, apoptosis, and inflammation, the key mechanisms underlying the development and progression of cancer^[109-112]. 1,25-(OH)₂D₃ has been shown to have significant anticancer effect on prostate, colon, breast, lung, liver, skin and pancreatic cancer cells, which express VDR^[105,112]. In addition, through a so-called non-genomic mechanism, 1,25-(OH)₂D₃ may have rapid effects on cellular functions^[113,114].

Substantial experimental studies, *in vitro* and in animals, showed the significant antitumor action of vitamin D and thoroughly investigated mechanisms at cellular and molecular levels^[115-117]. Furthermore, a large number of epidemiological studies explored the relationships between cancer incidence and geographic location, ultraviolet irradiation, and circulating levels of 25-OH-D₃^[118-125]. Despite abundant experimental evidence in support of an inverse association between vitamin D status and cancer risk, the available epidemiological evidence has not demonstrated consistently positive results to date. With some exceptions, most epidemiological studies have reported that populations in areas with low UV exposure have an increased risk of various types of cancers such as prostate, colon and breast^[119-125].

Serum 25-OH-D₃ level is the most widely used indi-

cator of vitamin D status in relation to other vitamin D metabolites^[126]. The result of a large population-based case-control study supported by the Long Island Breast Cancer Study Project showed that plasma 25-OH-D₃ level was inversely associated with breast cancer risk in a concentration-dependent fashion. Women with circulating 25-OH-D₃ above 40 ng/mL had approximately a 40% reduction in breast cancer risk compared with those who were vitamin D deficient^[127]. A large population-based case-control study from Germany reported similar findings^[128]. However, results from prospective studies did not support an association between vitamin D status and breast cancer. The inconsistent results also extend to studies of other types of cancer^[129]. A recent meta-analysis suggested that, in well-fed populations, an inverse relationship between serum 25-OH-D₃ levels and colorectal cancer existed (2630 cases in 9 studies), while no association was found for breast (6175 cases in 10 studies) and prostate cancer (3956 cases in 11 studies)^[130].

Excessive vitamin D intake was associated with additional toxic effects, such as hyperphosphatemia and hypercalciuria with clinical symptoms including nausea and vomiting, dehydration, muscle weakness, lethargy and confusion. The toxicity generally occurred when the daily dose exceeds 10 000 IU of vitamin D on a chronic basis^[131,132]. In this respect, several vitamin D analogs were synthesized with an attempt to minimize these side effects^[117,133]. In addition, recent investigations have followed the approach of intermittent administration of calcitriol in very high doses which elicits its antiproliferative effects with only transient hypercalcemia^[134,135]. There are still many unresolved questions regarding the maximum tolerated dose, optimal biologic dose and the optimal schedule for available vitamin D formulations.

A number of cancer intervention trials in humans have been conducted using vitamin D and its metabolites, as well as analogs alone or in combination with other anticancer drugs for the prevention or treatment of various cancers, especially prostate cancer, breast cancer and colorectal cancer. Similar to the results of the epidemiological studies, the outcomes of the clinical trials are inconclusive. The largest number of clinical studies have attempted to assess the efficacy of vitamin D in prostate cancer patients. Several phase II studies with calcitriol in combination with various chemotherapies showed a decrease in prostate-specific antigen levels. Additionally, in a large randomized, double-blinded, phase II trial (called ASCENT I) in patients with advanced prostate cancer ($n = 250$), the administration of a high dose (45 mg) of calcitriol (DN101) in combination with docetaxel caused a significant improvement in overall survival while there were no changes in the PSA response. Unfortunately, the 900 patient phase III study (ASCENT II) was stopped early because of inferior survival in the DN101 group, thus failing to confirm the improved survival seen in the ASCENT I trial^[136]. In a recent phase II study, high-dose intravenous calcitriol at a dose of 74 μ g weekly in combination with dexamethasone failed to produce a

clinical or PSA response in men with advanced prostate cancer^[137]. As regards breast cancer, the large intervention trial in 36 282 postmenopausal women conducted by the Women's Health Initiative showed calcium plus vitamin D supplementation for an average of 7 years did not reduce the incidence of invasive breast cancer compared with placebo^[138]. In contrast, another randomized study of vitamin D supplementation (cholecalciferol) indicated that diagnosis of all invasive cancers including breast cancer and colon cancer was substantially reduced in the vitamin D replacement group^[139]. For colorectal cancer, Fedirko *et al.*^[140] conducted a double-blind, 2 × 2 factorial clinical trial to test the effects of calcium and vitamin D₃ alone or in combination on markers of apoptosis in the normal colorectal mucosa. Bax expression along the full length of the crypts increased by 56% in the vitamin D group (*vs* placebo). However, another randomized trial indicated that calcium with vitamin D supplementation had no detectable effect on the incidence of colorectal cancer^[141].

OTHER PROMISING NATURAL AGENTS

Besides the natural agents mentioned above, there are other natural compounds with chemopreventive and chemotherapeutic potential. Chrysin, a natural flavonoid found in many plants, honey and propolis, possesses strong anti-inflammatory and anti-oxidant activity, and exhibits anti-cancer activity against leukemia, malignant glioma, breast carcinoma, cervical cancer, prostate cancer, lung cancer and colon cancer^[142]. Chrysin may inhibit cell proliferation and induce apoptosis by cell cycle arrest, inactivation of Akt signaling, activation of caspases, suppression of COX-2 and NF- κ B activation, and inhibition of proteasome activity^[142,143]. Chrysin has also been shown to reverse multidrug resistance of cancer cells *via* inhibition of P-glycoprotein and breast cancer resistance protein (BCRP/ABCG2)^[144,145]. However, there are few clinical study of the activity of chrysin against human cancers. Celastrol, a quinone methide triterpene, is the major active compound derived from the root of *Tripterygium wilfordii* Hook F. (the Chinese Thunder of God Vine), and generally used for the treatment of inflammatory and auto-immune diseases^[146]. Celastrol has attracted considerable attention recently, for its potential anti-cancer effects, with a broad spectrum of activity against multiple cancer types in both cell culture and animal models. Several molecular mechanisms of celastrol have been identified, including inhibition of IKK-NF- κ B signaling, disruption of the Cdc37/Hsp90 interaction, proteasome inhibition and heat shock response activation^[147,148]. However, no systematic clinical trials in human subjects have been carried out with celastrol to date. Betulinic acid is a natural product that is present in a variety of plants, especially the white-barked birch tree. The molecule is a member of the triterpene family of compounds that exhibit a variety of biological activities, especially, potent anti-HIV-1 and antitumor properties^[149,150]. This triterpene

has been found to inhibit the proliferation of a variety tumor cells and suppress tumor growth in animal studies. Numerous molecular targets for betulinic acid have been reported including enzymes (kinases, aminopeptidase N, acetyl-CoA acetyltransferase, topoisomerase I / II), the proteasome, NF- κ B and cell cycle regulation^[150,151]. In future studies, more anticancer mechanisms of betulinic acid will be delineated and clinical studies are needed to confirm the therapeutic value and exact anticancer target of betulinic acid in the human body.

REFERENCES

- 1 **Epstein J**, Sanderson IR, Macdonald TT. Curcumin as a therapeutic agent: the evidence from in vitro, animal and human studies. *Br J Nutr* 2010; **103**: 1545-1557
- 2 **Jurenka JS**. Anti-inflammatory properties of curcumin, a major constituent of *Curcuma longa*: a review of preclinical and clinical research. *Altern Med Rev* 2009; **14**: 141-153
- 3 **Wilken R**, Veena MS, Wang MB, Srivatsan ES. Curcumin: A review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma. *Mol Cancer* 2011; **10**: 12
- 4 **Ravindran J**, Prasad S, Aggarwal BB. Curcumin and cancer cells: how many ways can curry kill tumor cells selectively? *AAPS J* 2009; **11**: 495-510
- 5 Clinical development plan: curcumin. *J Cell Biochem Suppl* 1996; **26**: 72-85
- 6 **Sharma RA**, McLelland HR, Hill KA, Ireson CR, Euden SA, Manson MM, Pirmohamed M, Marnett LJ, Gescher AJ, Steward WP. Pharmacodynamic and pharmacokinetic study of oral *Curcuma* extract in patients with colorectal cancer. *Clin Cancer Res* 2001; **7**: 1894-1900
- 7 **Sharma RA**, Euden SA, Platten SL, Cooke DN, Shafayat A, Hewitt HR, Marczylo TH, Morgan B, Hemingway D, Plummer SM, Pirmohamed M, Gescher AJ, Steward WP. Phase I clinical trial of oral curcumin: biomarkers of systemic activity and compliance. *Clin Cancer Res* 2004; **10**: 6847-6854
- 8 **Cheng AL**, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, Lin JT, Lin BR, Ming-Shiang W, Yu HS, Jee SH, Chen GS, Chen TM, Chen CA, Lai MK, Pu YS, Pan MH, Wang YJ, Tsai CC, Hsieh CY. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res* 2001; **21**: 2895-2900
- 9 **Dhillon N**, Aggarwal BB, Newman RA, Wolff RA, Kunnumakkara AB, Abbruzzese JL, Ng CS, Badmaev V, Kurzrock R. Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clin Cancer Res* 2008; **14**: 4491-4499
- 10 **Asai A**, Miyazawa T. Occurrence of orally administered curcuminoid as glucuronide and glucuronide/sulfate conjugates in rat plasma. *Life Sci* 2000; **67**: 2785-2793
- 11 **Ireson C**, Orr S, Jones DJ, Verschoyle R, Lim CK, Luo JL, Howells L, Plummer S, Jukes R, Williams M, Steward WP, Gescher A. Characterization of metabolites of the chemopreventive agent curcumin in human and rat hepatocytes and in the rat in vivo, and evaluation of their ability to inhibit phorbol ester-induced prostaglandin E₂ production. *Cancer Res* 2001; **61**: 1058-1064
- 12 **Garcea G**, Jones DJ, Singh R, Dennison AR, Farmer PB, Sharma RA, Steward WP, Gescher AJ, Berry DP. Detection of curcumin and its metabolites in hepatic tissue and portal blood of patients following oral administration. *Br J Cancer* 2004; **90**: 1011-1015
- 13 **Garcea G**, Berry DP, Jones DJ, Singh R, Dennison AR, Farmer PB, Sharma RA, Steward WP, Gescher AJ. Consumption of the putative chemopreventive agent curcumin by cancer patients: assessment of curcumin levels in the colorectum and

- their pharmacodynamic consequences. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 120-125
- 14 **Padhye S**, Chavan D, Pandey S, Deshpande J, Swamy KV, Sarkar FH. Perspectives on chemopreventive and therapeutic potential of curcumin analogs in medicinal chemistry. *Mini Rev Med Chem* 2010; **10**: 372-387
 - 15 **Sarkar FH**, Li Y, Wang Z, Padhye S. Lesson learned from nature for the development of novel anti-cancer agents: implication of isoflavone, curcumin, and their synthetic analogs. *Curr Pharm Des* 2010; **16**: 1801-1812
 - 16 **Cruz-Correa M**, Shoskes DA, Sanchez P, Zhao R, Hyland LM, Wexner SD, Giardiello FM. Combination treatment with curcumin and quercetin of adenomas in familial adenomatous polyposis. *Clin Gastroenterol Hepatol* 2006; **4**: 1035-1038
 - 17 **Carroll RE**, Benya RV, Turgeon DK, Vareed S, Neuman M, Rodriguez L, Kakarala M, Carpenter PM, McLaren C, Meyskens FL, Brenner DE. Phase IIa clinical trial of curcumin for the prevention of colorectal neoplasia. *Cancer Prev Res (Phila)* 2011; **4**: 354-364
 - 18 **Kanai M**, Yoshimura K, Asada M, Imaizumi A, Suzuki C, Matsumoto S, Nishimura T, Mori Y, Masui T, Kawaguchi Y, Yanagihara K, Yazumi S, Chiba T, Guha S, Aggarwal BB. A phase I/II study of gemcitabine-based chemotherapy plus curcumin for patients with gemcitabine-resistant pancreatic cancer. *Cancer Chemother Pharmacol* 2011; **68**: 157-164
 - 19 **Bayet-Robert M**, Kwiatkowski F, Leheurteur M, Gachon F, Planchat E, Abrial C, Mouret-Reynier MA, Durando X, Barthomeuf C, Chollet P. Phase I dose escalation trial of docetaxel plus curcumin in patients with advanced and metastatic breast cancer. *Cancer Biol Ther* 2010; **9**: 8-14
 - 20 **Nagle DG**, Ferreira D, Zhou YD. Epigallocatechin-3-gallate (EGCG): chemical and biomedical perspectives. *Phytochemistry* 2006; **67**: 1849-1855
 - 21 **Khan N**, Mukhtar H. Cancer and metastasis: prevention and treatment by green tea. *Cancer Metastasis Rev* 2010; **29**: 435-445
 - 22 **Khan N**, Mukhtar H. Multitargeted therapy of cancer by green tea polyphenols. *Cancer Lett* 2008; **269**: 269-280
 - 23 **Yoshizawa S**, Horiuchi T, Fujiki H, Yoshida T, Okuda T, Sugimura T. Antitumor promoting activity of (-)-epigallocatechin gallate, the main constituent of "Tannin" in green tea. *Phytother Res* 1987; **1**: 44-47
 - 24 **Yang CS**, Ju J, Lu G, Xiao H, Hao X, Sang S, Lambert JD. Cancer prevention by tea and tea polyphenols. *Asia Pac J Clin Nutr* 2008; **17 Suppl 1**: 245-248
 - 25 **Chow HH**, Cai Y, Alberts DS, Hakim I, Dorr R, Shahi F, Crowell JA, Yang CS, Hara Y. Phase I pharmacokinetic study of tea polyphenols following single-dose administration of epigallocatechin gallate and polyphenon E. *Cancer Epidemiol Biomarkers Prev* 2001; **10**: 53-58
 - 26 **Chow HH**, Cai Y, Hakim IA, Crowell JA, Shahi F, Brooks CA, Dorr RT, Hara Y, Alberts DS. Pharmacokinetics and safety of green tea polyphenols after multiple-dose administration of epigallocatechin gallate and polyphenon E in healthy individuals. *Clin Cancer Res* 2003; **9**: 3312-3319
 - 27 **Mazzanti G**, Menniti-Ippolito F, Moro PA, Cassetti F, Raschetti R, Santuccio C, Mastrangelo S. Hepatotoxicity from green tea: a review of the literature and two unpublished cases. *Eur J Clin Pharmacol* 2009; **65**: 331-341
 - 28 **Ahn WS**, Yoo J, Huh SW, Kim CK, Lee JM, Namkoong SE, Bae SM, Lee IP. Protective effects of green tea extracts (polyphenon E and EGCG) on human cervical lesions. *Eur J Cancer Prev* 2003; **12**: 383-390
 - 29 **Shimizu M**, Fukutomi Y, Ninomiya M, Nagura K, Kato T, Araki H, Sukanuma M, Fujiki H, Moriwaki H. Green tea extracts for the prevention of metachronous colorectal adenomas: a pilot study. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 3020-3025
 - 30 **Bettuzzi S**, Brausi M, Rizzi F, Castagnetti G, Peracchia G, Corti A. Chemoprevention of human prostate cancer by oral administration of green tea catechins in volunteers with high-grade prostate intraepithelial neoplasia: a preliminary report from a one-year proof-of-principle study. *Cancer Res* 2006; **66**: 1234-1240
 - 31 **Brausi M**, Rizzi F, Bettuzzi S. Chemoprevention of human prostate cancer by green tea catechins: two years later. A follow-up update. *Eur Urol* 2008; **54**: 472-473
 - 32 **Pisters KM**, Newman RA, Coldman B, Shin DM, Khuri FR, Hong WK, Glisson BS, Lee JS. Phase I trial of oral green tea extract in adult patients with solid tumors. *J Clin Oncol* 2001; **19**: 1830-1838
 - 33 **Jatoi A**, Ellison N, Burch PA, Sloan JA, Dakhil SR, Novotny P, Tan W, Fitch TR, Rowland KM, Young CY, Flynn PJ. A phase II trial of green tea in the treatment of patients with androgen independent metastatic prostate carcinoma. *Cancer* 2003; **97**: 1442-1446
 - 34 **McLarty J**, Bigelow RL, Smith M, Elmajian D, Ankem M, Cardelli JA. Tea polyphenols decrease serum levels of prostate-specific antigen, hepatocyte growth factor, and vascular endothelial growth factor in prostate cancer patients and inhibit production of hepatocyte growth factor and vascular endothelial growth factor in vitro. *Cancer Prev Res (Phila)* 2009; **2**: 673-682
 - 35 **Shanafelt TD**, Lee YK, Call TG, Nowakowski GS, Dingli D, Zent CS, Kay NE. Clinical effects of oral green tea extracts in four patients with low grade B-cell malignancies. *Leuk Res* 2006; **30**: 707-712
 - 36 **Shanafelt TD**, Call TG, Zent CS, LaPlant B, Bowen DA, Roos M, Secreto CR, Ghosh AK, Kabat BF, Lee MJ, Yang CS, Jelinek DF, Erlichman C, Kay NE. Phase I trial of daily oral Polyphenon E in patients with asymptomatic Rai stage 0 to II chronic lymphocytic leukemia. *J Clin Oncol* 2009; **27**: 3808-3814
 - 37 **Somers-Edgar TJ**, Scandlyn MJ, Stuart EC, Le Nedelec MJ, Valentine SP, Rosengren RJ. The combination of epigallocatechin gallate and curcumin suppresses ER alpha-breast cancer cell growth in vitro and in vivo. *Int J Cancer* 2008; **122**: 1966-1971
 - 38 **Sun X**, Huo X, Luo T, Li M, Yin Y, Jiang Y. The anticancer flavonoid chrysin induces the unfolded protein response in hepatoma cells. *J Cell Mol Med* 2011; **15**: 2389-2398
 - 39 **Sartippour MR**, Pietras R, Marquez-Garban DC, Chen HW, Heber D, Henning SM, Sartippour G, Zhang L, Lu M, Weinberg O, Rao JY, Brooks MN. The combination of green tea and tamoxifen is effective against breast cancer. *Carcinogenesis* 2006; **27**: 2424-2433
 - 40 **Ermakova SP**, Kang BS, Choi BY, Choi HS, Schuster TF, Ma WY, Bode AM, Dong Z. (-)-Epigallocatechin gallate overcomes resistance to etoposide-induced cell death by targeting the molecular chaperone glucose-regulated protein 78. *Cancer Res* 2006; **66**: 9260-9269
 - 41 **Pyrko P**, Schönthal AH, Hofman FM, Chen TC, Lee AS. The unfolded protein response regulator GRP78/BiP as a novel target for increasing chemosensitivity in malignant gliomas. *Cancer Res* 2007; **67**: 9809-9816
 - 42 **Stearns ME**, Wang M. Synergistic Effects of the Green Tea Extract Epigallocatechin-3-gallate and Taxane in Eradication of Malignant Human Prostate Tumors. *Transl Oncol* 2011; **4**: 147-156
 - 43 **Zhang X**, Zhang H, Tighiouart M, Lee JE, Shin HJ, Khuri FR, Yang CS, Chen ZG, Shin DM. Synergistic inhibition of head and neck tumor growth by green tea (-)-epigallocatechin-3-gallate and EGFR tyrosine kinase inhibitor. *Int J Cancer* 2008; **123**: 1005-1014
 - 44 **Shah JJ**, Kuhn DJ, Orłowski RZ. Bortezomib and EGCG: no green tea for you? *Blood* 2009; **113**: 5695-5696
 - 45 **Golden EB**, Lam PY, Kardosh A, Gaffney KJ, Cadenas E, Louie SG, Petasis NA, Chen TC, Schönthal AH. Green tea polyphenols block the anticancer effects of bortezomib and other boronic acid-based proteasome inhibitors. *Blood* 2009; **113**: 5927-5937
 - 46 **Signorelli P**, Ghidoni R. Resveratrol as an anticancer nutri-

- ent: molecular basis, open questions and promises. *J Nutr Biochem* 2005; **16**: 449-466
- 47 **Shankar S**, Singh G, Srivastava RK. Chemoprevention by resveratrol: molecular mechanisms and therapeutic potential. *Front Biosci* 2007; **12**: 4839-4854
- 48 **Jang M**, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, Fong HH, Farnsworth NR, Kinghorn AD, Mehta RG, Moon RC, Pezzuto JM. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997; **275**: 218-220
- 49 **Kalra N**, Roy P, Prasad S, Shukla Y. Resveratrol induces apoptosis involving mitochondrial pathways in mouse skin tumorigenesis. *Life Sci* 2008; **82**: 348-358
- 50 **Whitsett T**, Carpenter M, Lamartiniere CA. Resveratrol, but not EGCG, in the diet suppresses DMBA-induced mammary cancer in rats. *J Carcinog* 2006; **5**: 15
- 51 **Sengottuvelan M**, Deeptha K, Nalini N. Resveratrol attenuates 1,2-dimethylhydrazine (DMH) induced glycoconjugate abnormalities during various stages of colon carcinogenesis. *Phytother Res* 2009; **23**: 1154-1158
- 52 **Li ZG**, Hong T, Shimada Y, Komoto I, Kawabe A, Ding Y, Kaganoi J, Hashimoto Y, Imamura M. Suppression of N-nitrosomethylbenzylamine (NMBA)-induced esophageal tumorigenesis in F344 rats by resveratrol. *Carcinogenesis* 2002; **23**: 1531-1536
- 53 **Bove K**, Lincoln DW, Tsan MF. Effect of resveratrol on growth of 4T1 breast cancer cells in vitro and in vivo. *Biochem Biophys Res Commun* 2002; **291**: 1001-1005
- 54 **Rezk YA**, Balulad SS, Keller RS, Bennett JA. Use of resveratrol to improve the effectiveness of cisplatin and doxorubicin: study in human gynecologic cancer cell lines and in rodent heart. *Am J Obstet Gynecol* 2006; **194**: e23-e26
- 55 **Scifo C**, Milasi A, Guarnera A, Sinatra F, Renis M. Resveratrol and propolis extract: an insight into the morphological and molecular changes induced in DU145 cells. *Oncol Res* 2006; **15**: 409-421
- 56 **Harikumar KB**, Kunnumakkara AB, Sethi G, Diagaradjane P, Anand P, Pandey MK, Gelovani J, Krishnan S, Guha S, Aggarwal BB. Resveratrol, a multitargeted agent, can enhance antitumor activity of gemcitabine in vitro and in orthotopic mouse model of human pancreatic cancer. *Int J Cancer* 2010; **127**: 257-268
- 57 **Fukui M**, Yamabe N, Zhu BT. Resveratrol attenuates the anticancer efficacy of paclitaxel in human breast cancer cells in vitro and in vivo. *Eur J Cancer* 2010; **46**: 1882-1891
- 58 **Sale S**, Verschoyle RD, Boocock D, Jones DJ, Wilsher N, Ruparelia KC, Potter GA, Farmer PB, Steward WP, Gescher AJ. Pharmacokinetics in mice and growth-inhibitory properties of the putative cancer chemopreventive agent resveratrol and the synthetic analogue trans 3,4,5,4'-tetramethoxystilbene. *Br J Cancer* 2004; **90**: 736-744
- 59 **Vitrac X**, Desmoulière A, Brouillaud B, Krisa S, Deffieux G, Barthe N, Rosenbaum J, Mérillon JM. Distribution of [¹⁴C]-trans-resveratrol, a cancer chemopreventive polyphenol, in mouse tissues after oral administration. *Life Sci* 2003; **72**: 2219-2233
- 60 **Patel KR**, Brown VA, Jones DJ, Britton RG, Hemingway D, Miller AS, West KP, Booth TD, Perloff M, Crowell JA, Brenner DE, Steward WP, Gescher AJ, Brown K. Clinical pharmacology of resveratrol and its metabolites in colorectal cancer patients. *Cancer Res* 2010; **70**: 7392-7399
- 61 **Smoliga JM**, Vang O, Baur JA. Challenges of translating basic research into therapeutics: resveratrol as an example. *J Gerontol A Biol Sci Med Sci* 2012; **67**: 158-167
- 62 **Brown VA**, Patel KR, Viskaduraki M, Crowell JA, Perloff M, Booth TD, Vasilinin G, Sen A, Schinas AM, Piccirilli G, Brown K, Steward WP, Gescher AJ, Brenner DE. Repeat dose study of the cancer chemopreventive agent resveratrol in healthy volunteers: safety, pharmacokinetics, and effect on the insulin-like growth factor axis. *Cancer Res* 2010; **70**: 9003-9011
- 63 **Chow HH**, Garland LL, Hsu CH, Vining DR, Chew WM, Miller JA, Perloff M, Crowell JA, Alberts DS. Resveratrol modulates drug- and carcinogen-metabolizing enzymes in a healthy volunteer study. *Cancer Prev Res (Phila)* 2010; **3**: 1168-1175
- 64 **Nguyen AV**, Martinez M, Stamos MJ, Moyer MP, Planutis K, Hope C, Holcombe RF. Results of a phase I pilot clinical trial examining the effect of plant-derived resveratrol and grape powder on Wnt pathway target gene expression in colonic mucosa and colon cancer. *Cancer Manag Res* 2009; **1**: 25-37
- 65 **Howells LM**, Berry DP, Elliott PJ, Jacobson EW, Hoffmann E, Hegarty B, Brown K, Steward WP, Gescher AJ. Phase I randomized, double-blind pilot study of micronized resveratrol (SRT501) in patients with hepatic metastases--safety, pharmacokinetics, and pharmacodynamics. *Cancer Prev Res (Phila)* 2011; **4**: 1419-1425
- 66 **Aggarwal BB**, Ichikawa H. Molecular targets and anticancer potential of indole-3-carbinol and its derivatives. *Cell Cycle* 2005; **4**: 1201-1215
- 67 **Kim YS**, Milner JA. Targets for indole-3-carbinol in cancer prevention. *J Nutr Biochem* 2005; **16**: 65-73
- 68 **Verhoeven DT**, Verhagen H, Goldbohm RA, van den Brandt PA, van Poppel G. A review of mechanisms underlying anticarcinogenicity by brassica vegetables. *Chem Biol Interact* 1997; **103**: 79-129
- 69 **Grose KR**, Bjeldanes LF. Oligomerization of indole-3-carbinol in aqueous acid. *Chem Res Toxicol* 1992; **5**: 188-193
- 70 **Kim EJ**, Park SY, Shin HK, Kwon DY, Surh YJ, Park JH. Activation of caspase-8 contributes to 3,3'-diindolylmethane-induced apoptosis in colon cancer cells. *J Nutr* 2007; **137**: 31-36
- 71 **Kandala PK**, Srivastava SK. Activation of checkpoint kinase 2 by 3,3'-diindolylmethane is required for causing G2/M cell cycle arrest in human ovarian cancer cells. *Mol Pharmacol* 2010; **78**: 297-309
- 72 **Ahmad A**, Kong D, Sarkar SH, Wang Z, Banerjee S, Sarkar FH. Inactivation of uPA and its receptor uPAR by 3,3'-diindolylmethane (DIM) leads to the inhibition of prostate cancer cell growth and migration. *J Cell Biochem* 2009; **107**: 516-527
- 73 **Bradlow HL**, Michnovicz J, Telang NT, Osborne MP. Effects of dietary indole-3-carbinol on estradiol metabolism and spontaneous mammary tumors in mice. *Carcinogenesis* 1991; **12**: 1571-1574
- 74 **Kojima T**, Tanaka T, Mori H. Chemoprevention of spontaneous endometrial cancer in female Donryu rats by dietary indole-3-carbinol. *Cancer Res* 1994; **54**: 1446-1449
- 75 **Oganesian A**, Hendricks JD, Williams DE. Long term dietary indole-3-carbinol inhibits diethylnitrosamine-initiated hepatocarcinogenesis in the infant mouse model. *Cancer Lett* 1997; **118**: 87-94
- 76 **Kim YH**, Kwon HS, Kim DH, Shin EK, Kang YH, Park JH, Shin HK, Kim JK. 3,3'-diindolylmethane attenuates colonic inflammation and tumorigenesis in mice. *Inflamm Bowel Dis* 2009; **15**: 1164-1173
- 77 **Kassie F**, Kalscheuer S, Matise I, Ma L, Melkamu T, Upadhyaya P, Hecht SS. Inhibition of vinyl carbamate-induced pulmonary adenocarcinoma by indole-3-carbinol and myoinositol in A/J mice. *Carcinogenesis* 2010; **31**: 239-245
- 78 **Weng JR**, Tsai CH, Kulp SK, Wang D, Lin CH, Yang HC, Ma Y, Sargeant A, Chiu CF, Tsai MH, Chen CS. A potent indole-3-carbinol derived antitumor agent with pleiotropic effects on multiple signaling pathways in prostate cancer cells. *Cancer Res* 2007; **67**: 7815-7824
- 79 **Kunimasa K**, Kobayashi T, Kaji K, Ohta T. Antiangiogenic effects of indole-3-carbinol and 3,3'-diindolylmethane are associated with their differential regulation of ERK1/2 and Akt in tube-forming HUVEC. *J Nutr* 2010; **140**: 1-6
- 80 **Marconett CN**, Sundar SN, Poindexter KM, Stueve TR, Bjeldanes LF, Firestone GL. Indole-3-carbinol triggers aryl hydrocarbon receptor-dependent estrogen receptor (ER)alpha pro-

- tein degradation in breast cancer cells disrupting an ERalpha-GATA3 transcriptional cross-regulatory loop. *Mol Biol Cell* 2010; **21**: 1166-1177
- 81 **Hung WC**, Chang HC. Indole-3-carbinol inhibits Sp1-induced matrix metalloproteinase-2 expression to attenuate migration and invasion of breast cancer cells. *J Agric Food Chem* 2009; **57**: 76-82
- 82 **Li Y**, VandenBoom TG, Kong D, Wang Z, Ali S, Philip PA, Sarkar FH. Up-regulation of miR-200 and let-7 by natural agents leads to the reversal of epithelial-to-mesenchymal transition in gemcitabine-resistant pancreatic cancer cells. *Cancer Res* 2009; **69**: 6704-6712
- 83 **Auborn KJ**, Fan S, Rosen EM, Goodwin L, Chandraskaren A, Williams DE, Chen D, Carter TH. Indole-3-carbinol is a negative regulator of estrogen. *J Nutr* 2003; **133**: 2470S-2475S
- 84 **Meng Q**, Yuan F, Goldberg ID, Rosen EM, Auborn K, Fan S. Indole-3-carbinol is a negative regulator of estrogen receptor-alpha signaling in human tumor cells. *J Nutr* 2000; **130**: 2927-2931
- 85 **Ashok BT**, Chen YG, Liu X, Garikapaty VP, Sepowitz R, Tschorn J, Roy K, Mittelman A, Tiwari RK. Multiple molecular targets of indole-3-carbinol, a chemopreventive anti-estrogen in breast cancer. *Eur J Cancer Prev* 2002; **11 Suppl 2**: S86-S93
- 86 **Okino ST**, Pookot D, Basak S, Dahiya R. Toxic and chemopreventive ligands preferentially activate distinct aryl hydrocarbon receptor pathways: implications for cancer prevention. *Cancer Prev Res (Phila)* 2009; **2**: 251-256
- 87 **Wang TT**, Milner MJ, Milner JA, Kim YS. Estrogen receptor alpha as a target for indole-3-carbinol. *J Nutr Biochem* 2006; **17**: 659-664
- 88 **Vivar OI**, Saunier EF, Leitman DC, Firestone GL, Bjeldanes LF. Selective activation of estrogen receptor-beta target genes by 3,3'-diindolylmethane. *Endocrinology* 2010; **151**: 1662-1667
- 89 **Czapski J**. Cancer preventing properties of cruciferous vegetables. *Veg Crops Res Bull* 2009; **70**: 5-18
- 90 **Andernton MJ**, Manson MM, Verschoyle RD, Gescher A, Lamb JH, Farmer PB, Steward WP, Williams ML. Pharmacokinetics and tissue disposition of indole-3-carbinol and its acid condensation products after oral administration to mice. *Clin Cancer Res* 2004; **10**: 5233-5241
- 91 **Reed GA**, Arneson DW, Putnam WC, Smith HJ, Gray JC, Sullivan DK, Mayo MS, Crowell JA, Hurwitz A. Single-dose and multiple-dose administration of indole-3-carbinol to women: pharmacokinetics based on 3,3'-diindolylmethane. *Cancer Epidemiol Biomarkers Prev* 2006; **15**: 2477-2481
- 92 **Rosen CA**, Bryson PC. Indole-3-carbinol for recurrent respiratory papillomatosis: long-term results. *J Voice* 2004; **18**: 248-253
- 93 **Kim DJ**, Lee KK, Han BS, Ahn B, Bae JH, Jang JJ. Biphasic modifying effect of indole-3-carbinol on diethylnitrosamine-induced preneoplastic glutathione S-transferase placental form-positive liver cell foci in Sprague-Dawley rats. *Jpn J Cancer Res* 1994; **85**: 578-583
- 94 **Stoner G**, Casto B, Ralston S, Roebuck B, Pereira C, Bailey G. Development of a multi-organ rat model for evaluating chemopreventive agents: efficacy of indole-3-carbinol. *Carcinogenesis* 2002; **23**: 265-272
- 95 **Michnovicz JJ**, Bradlow HL. Altered estrogen metabolism and excretion in humans following consumption of indole-3-carbinol. *Nutr Cancer* 1991; **16**: 59-66
- 96 **Michnovicz JJ**, Adlercreutz H, Bradlow HL. Changes in levels of urinary estrogen metabolites after oral indole-3-carbinol treatment in humans. *J Natl Cancer Inst* 1997; **89**: 718-723
- 97 **Wong GY**, Bradlow L, Sepkovic D, Mehl S, Mailman J, Osborne MP. Dose-ranging study of indole-3-carbinol for breast cancer prevention. *J Cell Biochem Suppl* 1997; **28-29**: 111-116
- 98 **Bradlow HL**, Michnovicz JJ, Halper M, Miller DG, Wong GY, Osborne MP. Long-term responses of women to indole-3-carbinol or a high fiber diet. *Cancer Epidemiol Biomarkers Prev* 1994; **3**: 591-595
- 99 **Reed GA**, Peterson KS, Smith HJ, Gray JC, Sullivan DK, Mayo MS, Crowell JA, Hurwitz A. A phase I study of indole-3-carbinol in women: tolerability and effects. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 1953-1960
- 100 **Dalessandri KM**, Firestone GL, Fitch MD, Bradlow HL, Bjeldanes LF. Pilot study: effect of 3,3'-diindolylmethane supplements on urinary hormone metabolites in postmenopausal women with a history of early-stage breast cancer. *Nutr Cancer* 2004; **50**: 161-167
- 101 **Bell MC**, Crowley-Nowick P, Bradlow HL, Sepkovic DW, Schmidt-Grimminger D, Howell P, Mayeaux EJ, Tucker A, Turbat-Herrera EA, Mathis JM. Placebo-controlled trial of indole-3-carbinol in the treatment of CIN. *Gynecol Oncol* 2000; **78**: 123-129
- 102 **Del Priore G**, Gudipudi DK, Montemarano N, Restivo AM, Malanowska-Stega J, Arslan AA. Oral diindolylmethane (DIM): pilot evaluation of a nonsurgical treatment for cervical dysplasia. *Gynecol Oncol* 2010; **116**: 464-467
- 103 **Naik R**, Nixon S, Lopes A, Godfrey K, Hatem MH, Monaghan JM. A randomized phase II trial of indole-3-carbinol in the treatment of vulvar intraepithelial neoplasia. *Int J Gynecol Cancer* 2006; **16**: 786-790
- 104 **Rajoria S**, Suriano R, Parmar PS, Wilson YL, Megwalu U, Moscatello A, Bradlow HL, Sepkovic DW, Geliebter J, Schantz SP, Tiwari RK. 3,3'-diindolylmethane modulates estrogen metabolism in patients with thyroid proliferative disease: a pilot study. *Thyroid* 2011; **21**: 299-304
- 105 **Holick MF**. Vitamin D: its role in cancer prevention and treatment. *Prog Biophys Mol Biol* 2006; **92**: 49-59
- 106 **Rosen CJ**. Clinical practice. Vitamin D insufficiency. *N Engl J Med* 2011; **364**: 248-254
- 107 **Anaizi N**. Rediscovering vitamin D. *Libyan J Med* 2010; **5**:
- 108 **Plum LA**, DeLuca HF. The functional metabolism and molecular biology of vitamin D action. *Clin Rev Bone Miner Metab* 2009; **7**: 20-41
- 109 **Ingraham BA**, Bragdon B, Nohe A. Molecular basis of the potential of vitamin D to prevent cancer. *Curr Med Res Opin* 2008; **24**: 139-149
- 110 **Krishnan AV**, Feldman D. Molecular pathways mediating the anti-inflammatory effects of calcitriol: implications for prostate cancer chemoprevention and treatment. *Endocr Relat Cancer* 2010; **17**: R19-R38
- 111 **Chung I**, Han G, Seshadri M, Gillard BM, Yu WD, Foster BA, Trump DL, Johnson CS. Role of vitamin D receptor in the antiproliferative effects of calcitriol in tumor-derived endothelial cells and tumor angiogenesis in vivo. *Cancer Res* 2009; **69**: 967-975
- 112 **Norman AW**, Bouillon R. Vitamin D nutritional policy needs a vision for the future. *Exp Biol Med (Maywood)* 2010; **235**: 1034-1045
- 113 **Fleet JC**. Rapid, membrane-initiated actions of 1,25 dihydroxyvitamin D: what are they and what do they mean? *J Nutr* 2004; **134**: 3215-3218
- 114 **Norman AW**. Minireview: vitamin D receptor: new assignments for an already busy receptor. *Endocrinology* 2006; **147**: 5542-5548
- 115 **Ordonez-Moran P**, Larriba MJ, Pendas-Franco N, Aguilera O, Gonzalez-Sancho JM, Munoz A. Vitamin D and cancer: an update of in vitro and in vivo data. *Front Biosci* 2005; **10**: 2723-2749
- 116 **Bouillon R**, Carmeliet G, Verlinden L, van Etten E, Verstuyf A, Luderer HF, Lieben L, Mathieu C, Demay M. Vitamin D and human health: lessons from vitamin D receptor null mice. *Endocr Rev* 2008; **29**: 726-776
- 117 **Masuda S**, Jones G. Promise of vitamin D analogues in the treatment of hyperproliferative conditions. *Mol Cancer Ther* 2006; **5**: 797-808
- 118 **Garland CF**, Garland FC, Gorham ED, Lipkin M, Newmark H, Mohr SB, Holick MF. The role of vitamin D in cancer pre-

- vention. *Am J Public Health* 2006; **96**: 252-261
- 119 **Grant WB**. Geographic variation of prostate cancer mortality rates in the United States: Implications for prostate cancer risk related to vitamin D. *Int J Cancer* 2004; **111**: 470-471; author reply 472
- 120 **Waltz P**, Chodick G. International comparisons of prostate cancer mortality rates with dietary practices and sunlight levels. *Urol Oncol* 2007; **25**: 85
- 121 **Waltz P**, Chodick G. Assessment of ecological regression in the study of colon, breast, ovary, non-Hodgkin's lymphoma, or prostate cancer and residential UV. *Eur J Cancer Prev* 2008; **17**: 279-286
- 122 **Gupta D**, Lammersfeld CA, Trukova K, Lis CG. Vitamin D and prostate cancer risk: a review of the epidemiological literature. *Prostate Cancer Prostatic Dis* 2009; **12**: 215-226
- 123 **John EM**, Koo J, Schwartz GG. Sun exposure and prostate cancer risk: evidence for a protective effect of early-life exposure. *Cancer Epidemiol Biomarkers Prev* 2007; **16**: 1283-1286
- 124 **Cui Y**, Rohan TE. Vitamin D, calcium, and breast cancer risk: a review. *Cancer Epidemiol Biomarkers Prev* 2006; **15**: 1427-1437
- 125 **Spina C**, Tangpricha V, Yao M, Zhou W, Wolfe MM, Maehr H, Uskokovic M, Adorini L, Holick MF. Colon cancer and solar ultraviolet B radiation and prevention and treatment of colon cancer in mice with vitamin D and its Gemini analogs. *J Steroid Biochem Mol Biol* 2005; **97**: 111-120
- 126 **Heaney RP**. The Vitamin D requirement in health and disease. *J Steroid Biochem Mol Biol* 2005; **97**: 13-19
- 127 **Crew KD**, Gammon MD, Steck SE, Hershman DL, Cremers S, Dworakowski E, Shane E, Terry MB, Desai M, Teitelbaum SL, Neugut AI, Santella RM. Association between plasma 25-hydroxyvitamin D and breast cancer risk. *Cancer Prev Res (Phila)* 2009; **2**: 598-604
- 128 **Abbas S**, Linseisen J, Slinger T, Kropp S, Mutschelknauss EJ, Flesch-Janys D, Chang-Claude J. Serum 25-hydroxyvitamin D and risk of post-menopausal breast cancer—results of a large case-control study. *Carcinogenesis* 2008; **29**: 93-99
- 129 **Freedman DM**, Chang SC, Falk RT, Purdue MP, Huang WY, McCarty CA, Hollis BW, Graubard BI, Berg CD, Ziegler RG. Serum levels of vitamin D metabolites and breast cancer risk in the prostate, lung, colorectal, and ovarian cancer screening trial. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 889-894
- 130 **Gandini S**, Boniol M, Haukka J, Byrnes G, Cox B, Sneyd MJ, Mullie P, Autier P. Meta-analysis of observational studies of serum 25-hydroxyvitamin D levels and colorectal, breast and prostate cancer and colorectal adenoma. *Int J Cancer* 2011; **128**: 1414-1424
- 131 **Hathcock JN**, Shao A, Vieth R, Heaney R. Risk assessment for vitamin D. *Am J Clin Nutr* 2007; **85**: 6-18
- 132 **Jones G**. Pharmacokinetics of vitamin D toxicity. *Am J Clin Nutr* 2008; **88**: 582S-586S
- 133 **Ma Y**, Khalifa B, Yee YK, Lu J, Memezawa A, Savkur RS, Yamamoto Y, Chintalacheruvu SR, Yamaoka K, Stayrook KR, Bramlett KS, Zeng QQ, Chandrasekhar S, Yu XP, Linebarger JH, Iturria SJ, Burris TP, Kato S, Chin WW, Nagpal S. Identification and characterization of noncalcemic, tissue-selective, nonsteroidal vitamin D receptor modulators. *J Clin Invest* 2006; **116**: 892-904
- 134 **Beer TM**, Lemmon D, Lowe BA, Henner WD. High-dose weekly oral calcitriol in patients with a rising PSA after prostatectomy or radiation for prostate carcinoma. *Cancer* 2003; **97**: 1217-1224
- 135 **Trump DL**, Potter DM, Muindi J, Brufsky A, Johnson CS. Phase II trial of high-dose, intermittent calcitriol (1,25 dihydroxyvitamin D3) and dexamethasone in androgen-independent prostate cancer. *Cancer* 2006; **106**: 2136-2142
- 136 **Scher HI**, Jia X, Chi K, de Wit R, Berry WR, Albers P, Henick B, Waterhouse D, Ruether DJ, Rosen PJ, Meluch AA, Nordquist LT, Venner PM, Heidenreich A, Chu L, Heller G. Randomized, open-label phase III trial of docetaxel plus high-dose calcitriol versus docetaxel plus prednisone for patients with castration-resistant prostate cancer. *J Clin Oncol* 2011; **29**: 2191-2198
- 137 **Chadha MK**, Tian L, Mashtare T, Payne V, Silliman C, Levine E, Wong M, Johnson C, Trump DL. Phase 2 trial of weekly intravenous 1,25 dihydroxy cholecalciferol (calcitriol) in combination with dexamethasone for castration-resistant prostate cancer. *Cancer* 2010; **116**: 2132-2139
- 138 **Chlebowski RT**, Johnson KC, Kooperberg C, Pettinger M, Wactawski-Wende J, Rohan T, Rossouw J, Lane D, O'Sullivan MJ, Yasmeen S, Hiatt RA, Shikany JM, Vitolins M, Khandekar J, Hubbell FA. Calcium plus vitamin D supplementation and the risk of breast cancer. *J Natl Cancer Inst* 2008; **100**: 1581-1591
- 139 **Lappe JM**, Travers-Gustafson D, Davies KM, Recker RR, Heaney RP. Vitamin D and calcium supplementation reduces cancer risk: results of a randomized trial. *Am J Clin Nutr* 2007; **85**: 1586-1591
- 140 **Fedirko V**, Bostick RM, Flanders WD, Long Q, Shaukat A, Rutherford RE, Daniel CR, Cohen V, Dash C. Effects of vitamin D and calcium supplementation on markers of apoptosis in normal colon mucosa: a randomized, double-blind, placebo-controlled clinical trial. *Cancer Prev Res (Phila)* 2009; **2**: 213-223
- 141 **Wactawski-Wende J**, Kotchen JM, Anderson GL, Assaf AR, Brunner RL, O'Sullivan MJ, Margolis KL, Ockene JK, Phillips L, Pottern L, Prentice RL, Robbins J, Rohan TE, Sarto GE, Sharma S, Stefanick ML, Van Horn L, Wallace RB, Whitlock E, Bassford T, Beresford SA, Black HR, Bonds DE, Brzyski RG, Caan B, Chlebowski RT, Cochran B, Garland C, Gass M, Hays J, Heiss G, Hendrix SL, Howard BV, Hsia J, Hubbell FA, Jackson RD, Johnson KC, Judd H, Kooperberg CL, Kuller LH, LaCroix AZ, Lane DS, Langer RD, Lasser NL, Lewis CE, Limacher MC, Manson JE. Calcium plus vitamin D supplementation and the risk of colorectal cancer. *N Engl J Med* 2006; **354**: 684-696
- 142 **Khoo BY**, Chua SL, Balam P. Apoptotic effects of chrysin in human cancer cell lines. *Int J Mol Sci* 2010; **11**: 2188-2199
- 143 **Bonfili L**, Cecarini V, Amici M, Cuccioloni M, Angeletti M, Keller JN, Eleuteri AM. Natural polyphenols as proteasome modulators and their role as anti-cancer compounds. *FEBS J* 2008; **275**: 5512-5526
- 144 **Bansal T**, Jaggi M, Khar RK, Talegaonkar S. Emerging significance of flavonoids as P-glycoprotein inhibitors in cancer chemotherapy. *J Pharm Pharm Sci* 2009; **12**: 46-78
- 145 **Wang X**, Morris ME. Effects of the flavonoid chrysin on nitrofurantoin pharmacokinetics in rats: potential involvement of ABCG2. *Drug Metab Dispos* 2007; **35**: 268-274
- 146 **Liu Z**, Ma L, Zhou GB. The main anticancer bullets of the Chinese medicinal herb, thunder god vine. *Molecules* 2011; **16**: 5283-5297
- 147 **Salminen A**, Lehtonen M, Paimela T, Kaarniranta K. Celastrol: Molecular targets of Thunder God Vine. *Biochem Biophys Res Commun* 2010; **394**: 439-442
- 148 **Kannaiyan R**, Shanmugam MK, Sethi G. Molecular targets of celastrol derived from Thunder of God Vine: potential role in the treatment of inflammatory disorders and cancer. *Cancer Lett* 2011; **303**: 9-20
- 149 **Aiken C**, Chen CH. Betulinic acid derivatives as HIV-1 antivirals. *Trends Mol Med* 2005; **11**: 31-36
- 150 **Fulda S**, Kroemer G. Targeting mitochondrial apoptosis by betulinic acid in human cancers. *Drug Discov Today* 2009; **14**: 885-890
- 151 **Mullauer FB**, Kessler JH, Medema JP. Betulinic acid, a natural compound with potent anticancer effects. *Anticancer Drugs* 2010; **21**: 215-227

S- Editor Li JY L- Editor Hughes D E- Editor Zheng XM

Indirect calorimetry in obese female subjects: Factors influencing the resting metabolic rate

Theresa Hagedorn, Eleonora Poggiogalle, Claudia Savina, Cecilia Coletti, Maddalena Paolini, Luciano Scavone, Barbara Neri, Lorenzo Maria Donini

Theresa Hagedorn, Claudia Savina, Cecilia Coletti, Maddalena Paolini, Luciano Scavone, Rehabilitation Clinical Institute "Villa delle Querce", Nemi, 00040 Rome, Italy

Eleonora Poggiogalle, Barbara Neri, Lorenzo Maria Donini, Department of Experimental Medicine, Medical Physiopathology, Food Science and Endocrinology Section, Food Science and Human Nutrition Research Unit, Sapienza University of Rome, 00185 Rome, Italy

Author contributions: Hagedorn T and Donini LM contributed equally to this work; Hagedorn T and Savina C designed the research; Hagedorn T, Savina C, Coletti C and Paolini M performed the research; Scavone L and Neri B analyzed the data; Poggiogalle E and Donini LM wrote the paper.

Correspondence to: Lorenzo Maria Donini, Professor, Department of Experimental Medicine, Medical Physiopathology, Food Science and Endocrinology Section, Food Science and Human Nutrition Research Unit, Sapienza University of Rome, P.le Aldo Moro, 5, 00185 Rome, Italy. lorenzomaria.donini@uniroma1.it
Telephone: +39-6-49690216 Fax: +39-6-49910699

Received: March 28, 2012 Revised: May 29, 2012

Accepted: June 1, 2012

Published online: June 20, 2012

Abstract

AIM: To evaluate selected factors influencing resting energy expenditure (REE) in obese female subjects.

METHODS: Seventy seven 61 obese Caucasian women [mean age of 52.93 ± 13.45 years, and mean body mass index (BMI) of 41.78 ± 11.54 kg/m²] were enrolled; measurements of resting metabolic rate (RMR) by a ventilated, open-circuit system, indirect calorimeter were performed after an overnight fast. Body composition as well as medications, physical parameters, blood samples, disease pattern, and smoking were considered.

RESULTS: RMR was significantly associated with body weight ($r = 0.732$, $P < 0.001$), body height ($r = 0.401$,

$P = 0.008$), BMI ($r = 0.504$, $P < 0.001$), waist circumference ($r = 0.602$, $P < 0.001$), mid-upper arm circumference ($r = 0.417$, $P = 0.006$), mid-upper arm muscle circumference ($r = 0.344$, $P = 0.028$), total body water ($r = 0.339$, $P = 0.035$), body temperature ($r = 0.409$, $P = 0.007$), smoking ($P = 0.031$), serum T₄ levels ($r = 0.331$, $P = 0.036$), obstructive sleep apnoea syndrome (OSAS; $P = 0.023$), impaired glucose tolerance (IGT; $P = 0.017$) and impaired glycaemic status, including hyperinsulinism, IGT and diabetes mellitus ($P = 0.003$).

CONCLUSION: Future research should be prompted to optimize the procedure of indirect calorimetry to achieve clinical benefits in obese subjects.

© 2012 Baishideng. All rights reserved.

Key words: Indirect calorimetry; Obesity; Resting metabolic rate; Resting energy expenditure

Peer reviewer: Moses Elisaf, Professor of Medicine, University of Ioannina, Medical School, Department of Internal Medicine, 451 10 Ioannina, Greece

Hagedorn T, Poggiogalle E, Savina C, Coletti C, Paolini M, Scavone L, Neri B, Donini LM. Indirect calorimetry in obese female subjects: Factors influencing the resting metabolic rate. *World J Exp Med* 2012; 2(3): 58-64 Available from: URL: <http://www.wjgnet.com/2220-315X/full/v2/i3/58.htm> DOI: <http://dx.doi.org/10.5493/wjem.v2.i3.58>

INTRODUCTION

An imbalance between energy expenditure (EE) and energy intake (EI) underlies the accumulation of exceeding body fat (BF) in obese subjects^[1,2]. Moreover, adiposity represents a common soil at the origin of different disorders such as type 2 diabetes mellitus (T2DM), hyperten-

sion, dyslipidemia, coronary heart disease (CHD), metabolic syndrome (MetS), sleep apnoea, as well as cancer^[3].

In overweight and obese patients, the exact assessment of basal metabolism plays a pivotal role in order to tailor a balanced nutritional support. Predictive equations have been originally developed from data collected in normal-weight individuals^[4,5]; even if they are considered to be a rapid and easy indirect method of resting metabolic rate (RMR) definition, they could not be thoroughly appropriate for obese patients, as a lack of correspondence between predicted values and real metabolic rate has been described in this subset of subjects^[6].

The direct measurement of RMR should be performed when the clear-cut definition of energy requirements is needed to address correctly dietary interventions^[7]. Indirect calorimetry (IC) is based on the evaluation of O₂ consumption (V_{O₂}) and CO₂ production (V_{CO₂}), and it is the most common method currently used with this purpose^[8].

Several factors have been shown to influence RMR measurement by IC. FFM, gender, and age may affect RMR, whereas the role of other factors, such as T2DM, ethnicity, menstrual cycle, hypertension, thyroid function, and smoking, remains to be univocally clarified^[9,10].

The aim of this study was the analysis of selected factors potentially affecting the RMR for a better understanding of determinants of RMR, providing evidence that would be helpful in obesity management and prevention of obesity-related comorbidities, optimizing weight loss strategies.

MATERIALS AND METHODS

Subjects

Sixty-one obese Caucasian women (mean age: 52.93 ± 13.45 years, and mean BMI: 41.78 ± 11.54 kg/m²), were enrolled in the study. Secondary diseases and medications at the time of the recruitment were not considered as exclusion criteria. After the IC performance, 19 women were excluded because the steady state criterion was not achieved. Hence, a sample of 42 female obese subjects was considered.

Study design

Examinations were performed under thermo-neutral conditions, from 8 to 10 o'clock in the morning^[11]. Participants had to remain in a 12-h fasting state before the examination, refraining from any heavy physical activity in the same period, abstaining from smoking in the preceding 2 h, and without ingesting coffee or water during 4 h prior to examination^[12].

RMR measurement

RMR was defined for each subject using an open-circuit indirect computerized calorimeter (stable IC: Quark RMR Cosmed), equipped with a canopy. Measurements were performed along a 15-min period preceded by a 10-min rest. Thermo-neutral conditions in a darkened room were

chosen, leaving subjects in a comfortable environment. Subjects were asked not to move and not to talk during the test performance. Heart rate (HR) was monitored. Excluding the preliminary 5 min of each measurement, the subsequent 10 min were considered for the evaluation of data, expressed in kJ/d; also the steady state criterion was established (5-min stable period of < 10% variation in measured V_{O₂} and V_{CO₂}). O₂ consumption and CO₂ production were standardized for temperature, barometric pressure and humidity^[13].

Anthropometric parameters

Body weight (BW) was measured to an accuracy of 0.1 kg through a standard column body scale. Body height (BH) was determined using a rigid stadiometer to an accuracy of 0.5 cm. BMI was calculated as BW/(BH)². Waist circumference (WC) was gathered using a standard measuring tape, to an accuracy of 0.1 cm. Mid arm circumference (MAC) was gathered using a standard measuring tape, on the non-dominant arm midway between the shoulder and elbow, to an accuracy of 0.1 cm.

Body composition

Bioimpedance analysis was performed on the right body side^[14] using a bioimpedance analyser AKERN Bioresearch SRL, Pontassieve, FL, Italy. Skinfold measurements: fat mass (FM) was estimated from the sum of four skinfolds (SF): tricipital (TSF), bicipital, subscapular, and suprailiac, measured on the non-dominant body side. Each measurement was repeated for three times, and the mean value was calculated to reduce the variability in the performance (Harpenden Caliper User manual). Body density (BD) was assessed using the Durnin and Womersley sex- and age- adjusted linear regression equation; from BD, FM was calculated by the Siri equation^[15].

Physical parameters

Measurements of blood pressure (BP) were performed on the left upper arm, to an accuracy of 5 mmHg. The body temperature (BT) was measured in the armpit, using a standard clinical thermometer.

Laboratory data/biochemistry

Blood samples were taken from each subject after an overnight fast. The following biochemical parameters were assayed: glucose, insulin, triglycerides (TG), total cholesterol (TC), LDL-cholesterol, HDL-cholesterol, thyroid stimulating hormone (TSH), thyroxin (T₄), albumin, and pre-albumin.

Laboratory tests were performed using a COBAS-MIRA analyser and a Cell-Dyn 1700 Analyser (Abbott), at the Laboratory of the "Villa delle Querce" Clinical Rehabilitation Institute. Serum concentrations of the biological indices were determined by routine methods with conventional commercial kits (ABX, Rome, Italy).

Disorders, diseases, medications and lifestyle habits

These parameters encompassed: T2DM (defined as fast-

Table 1 Anthropometric characteristics, clinical parameters and body composition

Variable	mean ± SD
Age (yr)	52.72 ± 13.96
BW ¹ (kg)	102.97 ± 26.55
BH ¹ (cm)	158.41 ± 6.71
BMI ¹ (kg/m ²)	41.35 ± 11.37
WC ¹ (cm)	115.44 ± 22.08
HR ¹ (bpm)	66.61 ± 11.03
SBP ¹ (mmHg)	127.32 ± 18.06
DBP ¹ (mmHg)	78.04 ± 8.58
CB ¹ (cm)	39.01 ± 8.23
CMB ¹ (cm)	29.31 ± 5.27
FM ^{1,2} (%)	42.86 ± 10.06
FM ^{1,3} (%)	45.34 ± 28.07
FFM ^{1,3} (%)	54.66 ± 28.07
MM ^{1,3} (%)	32.16 ± 25.71
BCM ^{1,3} (%)	44.49 ± 8.40
BCM ¹ -Index ³	8.91 ± 2.43
TBW ^{1,3} (%)	41.06 ± 20.42
ECW ^{1,3} (%)	52.56 ± 5.37
ICW ^{1,3} (%)	47.44 ± 5.37
RMR ¹ (kcal/d)	1537.91 ± 281.76

Descriptive data in obese women. ¹See the full text for abbreviations of subjects' anthropometric parameters; ²Estimated by Harpenden caliper; ³Estimated by BIA. BW: Body weight; BH: Body height; BMI: Body mass index; WC: Waist circumference; HR: Heart rate; FM: Fat mass; RMR: Resting metabolic rate.

ing glucose levels ≥ 126 mg/dL), impaired glucose tolerance (IGT) (fasting glucose levels ranging from 110 to 126 mg/dL), hyperinsulinism, obstructive sleep apnoea syndrome (OSAS), hypothyroidism/dysthyroidism, dyslipidemia, hypertension (BP > 130/85 mmHg), depression. Metformin, levothyroxine, anti-depressants and anti-hypertensive agents were included. Smoking habit (more than ten cigarettes per day), and the menstrual state (regular/irregular menses and postmenopausal period) were also taken into account.

Statistical analysis

Baseline data and subjects' characteristics were evaluated and collected. Statistical analysis was performed using Excel 2007 and SPSS 10.0 for Windows. Parameters were expressed by mean values and by standard deviation (SD) indicating the minimal and maximal extremity of each range of values. The outcome dependent variable, the RMR, was expressed in kcal/d. Independent variables were expressed as continuous or as categorical variables.

Pearson's correlation coefficient was used to correlate the means of the RMR with parameters. Linear regression models, analysis of variance (ANOVA), and independent *t* test were used. Differences and correlations were considered statistically significant when *P* value was < 0.05.

RESULTS

A total of sixty-one women (mean age of 52.93 ± 13.45 years, and mean BMI of 41.78 ± 11.54 kg/m²), were

Table 2 Parameters influencing resting metabolic rate (descriptive data)

Variable	Subjects (%)
T2DM	33.3
IGT	18.8
Hyperinsulinism	26.1
OSAS	17.4
Dysthyroidism	31.3
Dyslipidemia	13.0
Hypertension	65.2
Depression	27.5
Metformin	36.2
Antidepressants	18.8
Antihypertensive drugs	10.1
Levothyroxine	31.9
Smoking habit	43.8
Postmenopausal state	0

T2DM: Type 2 diabetes mellitus; IGT: Impaired glucose tolerance; OSAS: Obstructive sleep apnoea syndrome.

Table 3 Correlation analysis among subjects' anthropometric parameters, body composition and resting metabolic rate (kJ/d) (n = 42)

Variables	Correlation (r)	<i>P</i> ⁴
Age (yr)	0.006	0.969
BW ¹ (kg)	0.732	< 0.001
BH ¹ (cm)	0.401	0.008
BMI ¹ (kg/m ²)	0.504	< 0.001
WC ¹ (cm)	0.602	< 0.001
MAC ¹ (cm)	0.417	0.006
MAMC ¹ (cm)	0.344	0.028
FM ^{1,2} and FFM ^{1,2} (%)	0.169	0.290
FM ^{1,3} and FFM ^{1,3} (%)	0.113	0.485
MM ^{1,3} (%)	0.105	0.522
BCM ^{1,3} (%)	0.033	0.837
BCM ¹ -Index ³	0.120	0.458
TBW ^{1,3} (%)	0.339	0.035
ECW ^{1,3} and ICW ^{1,3} (%)	0.029	0.865

¹See the full text for abbreviations of subjects' anthropometric parameters; ²Estimated by Harpenden caliper; ³Estimated by BIA. BW: Body weight; ⁴Differences and correlations were considered statistically significant when *P* value was < 0.05. BH: Body height; BMI: Body mass index; WC: Waist circumference; FM: Fat mass.

studied. Descriptive data, anthropometric characteristics, and selected parameters are shown in Tables 1 and 2, respectively. Just forty two of the sixty one women achieved the steady state criterion. Thus, for further evaluation, 42 obese women (mean age 52.72 ± 13.96 years and mean BMI 41.35 ± 11.37 kg/m²) were considered only.

Factors influencing the RMR

Anthropometric parameters influencing the RMR:

The following anthropometric parameters: BW (*r* = 0.732, *P* < 0.001), BMI (*r* = 0.504, *P* < 0.001), WC (*r* = 0.602, *P* < 0.001), MAC (*r* = 0.417, *P* = 0.006), BH (*r* = 0.401, *P* = 0.008), MAMC (*r* = 0.344, *P* = 0.028), and TBW (*r* = 0.339, *P* = 0.035), showed a statistically significant correlation with RMR (Table 3).

Table 4 Correlation of subjects' physical characteristics with resting metabolic rate (*n* = 42)

Variables	Correlation (<i>r</i>)	<i>P</i> ¹
Blood pressure (mmHg) systolic/diastolic	0.048/0.035	0.759/0.825
Heart rate (b/min)	0.296	0.050
SpO ₂ (%)	0.158	0.318
Body temperature (°C)	0.409	0.007

¹Differences and correlations were considered statistically significant when *P* value was < 0.05.

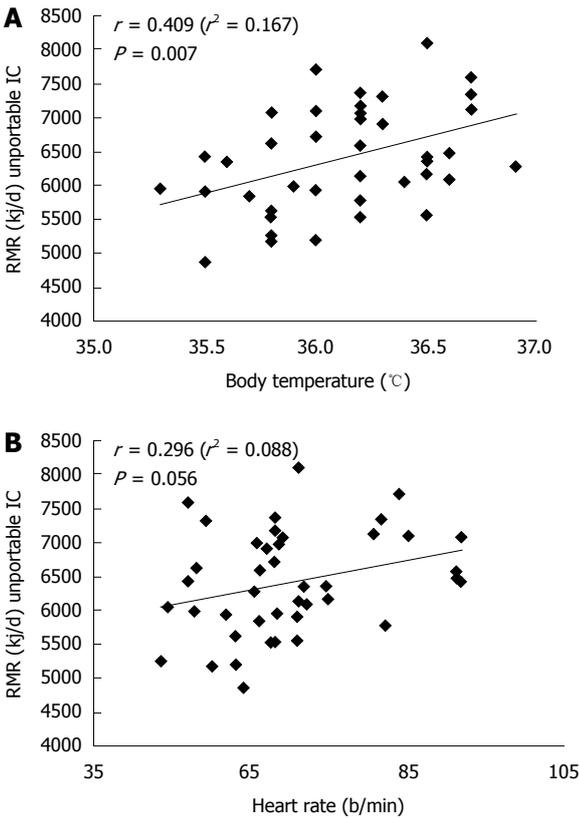


Figure 1 Correlation analysis between resting metabolic rate and body temperature (A) and heart rate (B). RMR: Resting metabolic rate.

Physical parameters influencing the RMR: BP, HR, SpO₂, and BT were considered (Table 4). No significant correlation was found for BP ($r = 0.048/0.035$, for systolic and diastolic BP, respectively) and SpO₂ ($r = 0.158$) indicating a lack of influence of these parameters on the measured RMR. 29 (69%) of the total subjects were treated with anti-hypertensive agents, likely accounting for normal BP values. On the other hand, correlations were obtained with HR ($r = 0.296$, $P = 0.050$; Figure 1B: $P < 0.05$), and a significant association was found with BT ($r = 0.409$, $P = 0.007$; Figure 1A). As shown in Figure 1A ($P < 0.05$), BT was positively correlated with RMR.

A slight positive correlation between RMR and HR was described, showing a trend toward a higher estimated RMR consistently with an increase of HR, but without statistically significant results.

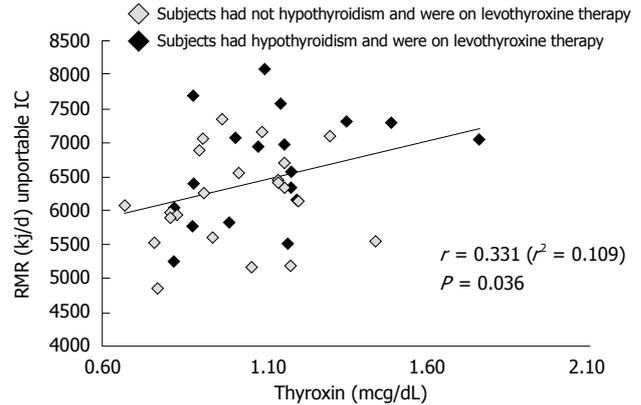


Figure 2 Correlation analysis between resting metabolic rate and thyroxin levels. RMR: Resting metabolic rate.

Blood parameters influencing the RMR: In Table 3 correlation analysis among RMR and serum TC, TG, LDL-cholesterol, HDL-cholesterol, glucose, insulin, TSH, T₄, albumin and pre-albumin levels is shown. Just one significant positive association has been reported between RMR and serum T₄ concentrations ($r = 0.331$, $P = 0.036$) (Figure 2: $P < 0.05$); as illustrated in Figure 2, a significant positive correlation has been found between RMR and T₄ concentrations, even considering progressively increasing serum T₄ levels within the normal range of values. Eighteen subjects had hypothyroidism and were on levothyroxine therapy (Figure 2).

Other factors influencing the RMR: A correlation analysis among RMR and disorders, diseases, medications, and smoking was performed. A total of 13 parameters were included: type 2 diabetes mellitus (T2DM- including patients on metformin therapy), IGT, hyperinsulinism, thyroid function (hypothyroidism and dysthyroidism), dyslipidemia, hypertension (including subjects on hypertensive therapy), OSAS, depression, anxiety, menstrual cycle or postmenopausal state, antidepressant medications, metformin, levothyroxine, and smoking. A significant relationship was demonstrated between RMR and smoking ($P = 0.031$), OSAS ($P = 0.023$), IGT ($P = 0.005$), and impaired glycemic status- including IGT, T2DM, and hyperinsulinism ($P = 0.003$).

Environmental conditions influencing the RMR: Data provided by correlation analysis excluded any significant association among RMR and environmental variables (room temperature: $r = 0.118$, $P = 0.456$; room humidity: $r = 0.076$, $P = 0.634$, and atmospheric air pressure: $r = 0.018$, $P = 0.910$).

DISCUSSION

In the present study we selected and considered anthropometric and physical characteristics, blood parameters, as well as environmental conditions, to ascertain their impact on the RMR of obese female subjects.

As expected, BW and BH were significantly positively correlated with RMR, as well as BMI did, confirming the important role of BW and BH in modulating the basal metabolic rate, as these variables were already taken into account in early studies for the estimation of RMR.

WC, MAC and MAMC were found to be significantly positively associated with RMR. WC is a strong predictor for visceral adipose tissue, and it is currently and universally accepted as an independent cardiovascular risk marker for obese people^[16]. Just few studies exploring and assessing the relationship between WC and RMR exist^[17,18]. The correlation of MAC and MAMC with RMR could be explained by a higher accumulation of FM in the mid-upper arm in obese subjects, which is proportional to the accumulation of adipose tissue in other parts of the body.

BW as a significant predictor of increased RMR, and the higher percentage of FM and FFM in obese subjects, may lead also to a higher MAC and MAMC, hence to a higher RMR.

Several studies reported that RMR decreases parallel aging, because of the loss of FFM and the increase in FM, since body mass and BC are the major predictors of EE^[1]. In the present study no significant correlation was found among FM, FFM, BCM, MM, BCMI and RMR. FM was expected to contribute slightly to RMR, thus no significant correlation was obtained, and it is supported by previous evidence^[19]. The influence of FM on RMR has not been univocally assessed, results being inconsistent across studies, even if it seems to play only a minor additional role.

FFM alone is well known to be related to RMR^[20] though in the present study no correlation emerged, nor with values obtained by anthropometry neither by BIA, whereas a positive association was found between RMR and TBW. Special considerations should be highlighted when body composition of obese subjects is determined by BIA. In obese and overweight individuals the higher hydration of FFM is responsible for an overestimation of FFM and underestimation of FM. Additionally, the body build of obese subjects, especially abdominal obesity, concurs to the mentioned impairment of the results provided by the BIA^[21]. Surprisingly in the present study TBW was significantly positively correlated with RMR, while FFM and FM were not. Even if BIA could not be completely reliable in our sample of obese participants, an increased total body water content mirroring a higher FFM may explain the positive association with RMR, that is notably higher as FFM increases.

Hence, the finding of a positive association between RMR and TBW, in absence of the same relation linking RMR and FFM, could be likely justified by the limited accuracy of BIA for FFM and FM assessment in such an altered hydration status that is obesity.

Otherwise, we evaluated FFM as a unique compartment, while growing evidence recognizes that FFM is energetically heterogeneous because it encompasses organs and tissues having a different metabolic rate^[22].

Age did not show any correlation with RMR. Basal metabolism was shown to drop by 1%-2% per decade over the age of 20 to 75 years^[23], and that aging process is accompanied by the apparent replacement of a certain proportion of MM by a gain in FM^[24]. Actually, a potential reason for the inconsistency of our findings may be that mean age of our participants was lower than age of subjects in prior studies reporting an association between age and RMR^[25].

Evaluating physical characteristics and the environmental conditions of the examination room, significant results were obtained only regarding BT. No significant results were obtained by the correlation analysis of RMR with BP, HR, SpO₂, MET, room humidity and atmospheric air pressure.

No significant correlation with RMR was found for the selected blood parameters, save for serum T₄ levels. The other blood parameters did not show any significant effect on the RMR, but it is likely that a detection of significantly abnormal values was difficult, since most subjects were on pharmacological therapy to maintain blood values in the normal range.

A wide spectrum of evidence supports the involvement of thyroid hormone in mechanisms responsible for thermogenesis, affecting EE and basal metabolic rate^[26].

Resting energy expenditure (REE) has been shown to be very sensitive to modulation by thyroid hormone^[27].

In the present study higher T₄ concentrations were associated with higher RMR. Although thyroid hormone concentration was within the normal range, a significant effect of T₄ concentration on RMR was however detectable. 19 subjects in the present study had hypothyroidism with no significant effect on metabolism, likely justified by levothyroxine chronic supplementation.

The effect of smoking status on EE has been at the centre of a number of studies attempting to assess the relationship linking the smoking attitude or its cessation to BW fluctuations^[28,29]. The acute or short-term effect of cigarette smoking or nicotine leads to changes in metabolism that imply an increase in total EE or in RMR in few studies^[30,31], despite this hypothesis remains to be confirmed. Moreover, smoking effect has been found to be different in obese smokers when compared to lean smokers^[32]. In our study, smokers were all obese, and their RMR was increased by 10.4% than non-smokers' RMR. Mechanisms other than the direct influence on energy and metabolism may account for these results, as nicotine can affect appetite and sympathetic nervous system action.

An aspect that strengthens our finding is that the positive correlation between smoking and RMR exists even after patients refrained from smoking at least 2 h before the examination, eliminating the very short-term nicotine effects.

Disorders and diseases as well as medicines did not show any significant correlation with RMR, save OSAS, IGT, and impaired glycaemic status (including IGT, T2DM, and hyperinsulinism), that were significantly correlated with RMR.

In agreement with prior studies, we found that RMR was increased in OSAS obese subjects when compared to obese women without OSAS^[33,34].

OSAS patients are usually obese, and abnormalities in intrathoracic pressure lead to an increased work of breathing, as well as frequent arousals and increased sympathetic activity, that may account for the increased EE^[33,34].

Additionally, we found that RMR was significantly different in obese female subjects with an impaired glycemic status, when compared to their normoglycemic counterparts.

We considered subgroups including patients with IGT, diabetes mellitus, and hyperinsulinism, respectively, and subgroup analysis showed that RMR was significantly increased in patients with IGT or in whom with hyperinsulinism, whereas only a trend toward higher values of RMR was observed in patients with diabetes mellitus; previous studies had demonstrated an increased RMR in diabetic patients, and several mechanisms were implicated to explain the increased EE, such as the activation of energy-consuming metabolic processes- gluconeogenesis and other substrate cycles- and an increased sympathetic nervous system activity. We have to stress that in the subgroup of type 2 diabetic patients, those who were on metformin therapy were not considered separately, and it may account for the slight increase in RMR in the whole subgroup. Moreover, no significant difference has emerged from the comparison of RMR between diabetic subjects treated with metformin and non-treated patients. Metformin is responsible for improved glycemic control, thus it may avoid overcoming metabolic adaptations of RMR that typically occur in T2DM^[35]. Therefore, the significant association between increased RMR and IGT and the tendency toward a higher RMR in type 2 diabetic patients could be due to the common soil of insulin resistance underlying the different aspects of glucose abnormalities; on the other hand, just a little body of evidence is available at present investigating the relationship between insulin resistance and RMR^[36].

Definitive conclusions may not be drawn. First, sample size was relatively small. Moreover, we studied a group of white female subjects exclusively, and influence of both gender and ethnicity on RMR has been widely recognized.

It is noteworthy that 33 of the 77 subjects, originally entering the study, failed to achieve the steady state criterion for IC measurements, hence they were subsequently excluded.

In stable, spontaneously breathing patients, like those evaluated in the present study, anxiety or hyperventilation have been frequently addressed to impact the achievement of the steady state^[37]. The failure in satisfying the steady state criterion could be attributable to the assumption of a very stringent definition of steady state: less than 10% for changes in $\dot{V}O_2$ and $\dot{V}CO_2$ over a period of 5 min. In existing literature less rigorous steady state criteria were applied, even if a weaker strength of correlation

measurements with the 24-h REE was obtained when establishing a $\dot{V}O_2/\dot{V}CO_2$ change by $\leq 15\%$ or 20% . We adopted a more stringent definition of steady state in order to confer a higher accuracy to the results. We do not know if obesity may represent a limitation for the satisfaction of a very strict steady state criterion, and similar data are lacking for obese population at present.

In conclusion, the association among several anthropometric, physical and environmental factors, as well as disorders, diseases, medications, smoking habit and RMR was explored. Although significant results emerged from the correlation analysis, showing that RMR was positively related to BW, BH, BMI, WC, MAC, MAMC, TBW, BT, serum T₄ levels, levothyroxine and smoking, the reason for the significance was not always thoroughly clear, and it could be related to coincidental association within and between parameters, considering that RMR can range widely within a group of people.

The precise identification of factors influencing the RMR is not only an experimental concern, but it represents a real clinical challenge. IC is an important method allowing an individualized obese patient care. Defining factors interfering with RMR will be useful to maximize the beneficial effects deriving from a tailored therapeutic approach. Moreover it would prompt further research in order to address and to improve obesity management.

COMMENTS

Background

Obesity is spreading worldwide in an epidemic fashion. Research should be prompted to increase knowledge about mechanisms underlying metabolism regulation in order to optimize tools and potential care interventions.

Research frontiers

Currently technology offers tools that could be useful in a personalized approach to the management of obesity. Indirect calorimetry (IC) could be a technological resource to improve the tailored treatment of obese subjects.

Innovations and breakthroughs

Because of the larger life expectancy and the presence of comorbidities linked both to the ageing process and to obesity itself, a variety of factors potentially affecting the resting metabolic rate (RMR) should be taken into account when considering obese subjects than their lean and healthy counterparts.

Applications

By understanding how resting energy expenditure (REE) is modulated in obese subjects, this study may represent a future strategy addressed to a technologically supported treatment and a more precise dietary intervention in this subset of patients.

Terminology

IC is a common method to assess exactly the REE, based on respiratory gas exchange, allowing defining thoroughly the characteristics of nutritional interventions.

Peer review

The authors examined selected factors potentially influencing the RMR in obese women. REE was evaluated using IC. The results suggest that this experimental approach could represent an interesting method for an individualized obese patient care.

REFERENCES

- 1 **Leitzmann C.** Nutrition ecology: origin and definition. *Forum Nutr* 2003; **56**: 220-221
- 2 **Greenberg AS, Obin MS.** Obesity and the role of adipose

- tissue in inflammation and metabolism. *Am J Clin Nutr* 2006; **83**: 461S-465S
- 3 **Pi-Sunyer FX.** The obesity epidemic: pathophysiology and consequences of obesity. *Obes Res* 2002; **10** Suppl 2: 97S-104S
 - 4 **Reeves MM, Capra S.** Predicting energy requirements in the clinical setting: are current methods evidence based? *Nutr Rev* 2003; **61**: 143-151
 - 5 **Haugen HA, Chan LN, Li F.** Indirect calorimetry: a practical guide for clinicians. *Nutr Clin Pract* 2007; **22**: 377-388
 - 6 **Livingston EH, Kohlstadt I.** Simplified resting metabolic rate-predicting formulas for normal-sized and obese individuals. *Obes Res* 2005; **13**: 1255-1262
 - 7 **Compher C, Frankenfield D, Keim N, Roth-Yousey L.** Best practice methods to apply to measurement of resting metabolic rate in adults: a systematic review. *J Am Diet Assoc* 2006; **106**: 881-903
 - 8 **Levine JA.** Measurement of energy expenditure. *Public Health Nutr* 2005; **8**: 1123-1132
 - 9 **Horgan GW, Stubbs J.** Predicting basal metabolic rate in the obese is difficult. *Eur J Clin Nutr* 2003; **57**: 335-340
 - 10 **Solomon SJ, Kurzer MS, Calloway DH.** Menstrual cycle and basal metabolic rate in women. *Am J Clin Nutr* 1982; **36**: 611-616
 - 11 **Nieman DC, Trone GA, Austin MD.** A new handheld device for measuring resting metabolic rate and oxygen consumption. *J Am Diet Assoc* 2003; **103**: 588-592
 - 12 **Haugen HA, Melanson EL, Tran ZV, Kearney JT, Hill JO.** Variability of measured resting metabolic rate. *Am J Clin Nutr* 2003; **78**: 1141-1145
 - 13 **Severinghaus JW.** Water vapor calibration errors in some capnometers: respiratory conventions misunderstood by manufacturers? *Anesthesiology* 1989; **70**: 996-998
 - 14 **Chumlea WC, Guo SS.** Bioelectrical impedance and body composition: present status and future directions. *Nutr Rev* 1994; **52**: 123-131
 - 15 **Durnin JV, Womersley J.** Body fat assessed from total body density and its estimation from skinfold thickness: measurements on 481 men and women aged from 16 to 72 years. *Br J Nutr* 1974; **32**: 77-97
 - 16 **Janssen I, Katzmarzyk PT, Ross R.** Duration of overweight and metabolic health risk in American men and women. *Ann Epidemiol* 2004; **14**: 585-591
 - 17 **Armellini F, Robbi R, Zamboni M, Todesco T, Castelli S, Bosello O.** Resting metabolic rate, body-fat distribution, and visceral fat in obese women. *Am J Clin Nutr* 1992; **56**: 981-987
 - 18 **den Besten C, Vansant G, Weststrate JA, Deurenberg P.** Resting metabolic rate and diet-induced thermogenesis in abdominal and gluteal-femoral obese women before and after weight reduction. *Am J Clin Nutr* 1988; **47**: 840-847
 - 19 **Usui C, Takahashi E, Gando Y, Sanada K, Oka J, Miyachi M, Tabata I, Higuchi M.** Relationship between blood adipocytokines and resting energy expenditure in young and elderly women. *J Nutr Sci Vitaminol (Tokyo)* 2007; **53**: 529-535
 - 20 **Leibel RL, Rosenbaum M, Hirsch J.** Changes in energy expenditure resulting from altered body weight. *N Engl J Med* 1995; **332**: 621-628
 - 21 **Coppini LZ, Waitzberg DL, Campos AC.** Limitations and validation of bioelectrical impedance analysis in morbidly obese patients. *Curr Opin Clin Nutr Metab Care* 2005; **8**: 329-332
 - 22 **Javed F, He Q, Davidson LE, Thornton JC, Albu J, Boxt L, Krasnow N, Elia M, Kang P, Heshka S, Gallagher D.** Brain and high metabolic rate organ mass: contributions to resting energy expenditure beyond fat-free mass. *Am J Clin Nutr* 2010; **91**: 907-912
 - 23 **Keys A, Taylor HL, Grande F.** Basal metabolism and age of adult man. *Metabolism* 1973; **22**: 579-587
 - 24 **Lizzer S, Bedogni G, Lafortuna CL, Marazzi N, Busti C, Galli R, De Col A, Agosti F, Sartorio A.** Relationship between basal metabolic rate, gender, age, and body composition in 8,780 white obese subjects. *Obesity (Silver Spring)* 2010; **18**: 71-78
 - 25 **Manini TM, Everhart JE, Anton SD, Schoeller DA, Cummings SR, Mackey DC, Delmonico MJ, Bauer DC, Simonsick EM, Colbert LH, Visser M, Tylavsky F, Newman AB, Harris TB.** Activity energy expenditure and change in body composition in late life. *Am J Clin Nutr* 2009; **90**: 1336-1342
 - 26 **Kim B.** Thyroid hormone as a determinant of energy expenditure and the basal metabolic rate. *Thyroid* 2008; **18**: 141-144
 - 27 **Tagliaferri M, Berselli ME, Calò G, Minocci A, Savia G, Petroni ML, Viberti GC, Liuzzi A.** Subclinical hypothyroidism in obese patients: relation to resting energy expenditure, serum leptin, body composition, and lipid profile. *Obes Res* 2001; **9**: 196-201
 - 28 **Chiolero A, Faeh D, Paccaud F, Cornuz J.** Consequences of smoking for body weight, body fat distribution, and insulin resistance. *Am J Clin Nutr* 2008; **87**: 801-809
 - 29 **Bradley DP, Johnson LA, Zhang Z, Subar AF, Troiano RP, Schatzkin A, Schoeller DA.** Effect of smoking status on total energy expenditure. *Nutr Metab (Lond)* 2010; **7**: 81
 - 30 **Perkins KA, Epstein LH, Stiller RL, Marks BL, Jacob RG.** Acute effects of nicotine on resting metabolic rate in cigarette smokers. *Am J Clin Nutr* 1989; **50**: 545-550
 - 31 **Hofstetter A, Schutz Y, Jéquier E, Wahren J.** Increased 24-hour energy expenditure in cigarette smokers. *N Engl J Med* 1986; **314**: 79-82
 - 32 **Audrain JE, Klesges RC, Klesges LM.** Relationship between obesity and the metabolic effects of smoking in women. *Health Psychol* 1995; **14**: 116-123
 - 33 **Lin CC, Chang KC, Lee KS.** Effects of treatment by laser-assisted uvuloplasty on sleep energy expenditure in obstructive sleep apnea patients. *Metabolism* 2002; **51**: 622-627
 - 34 **Ryan CF, Love LL, Buckley PA.** Energy expenditure in obstructive sleep apnea. *Sleep* 1995; **18**: 180-187
 - 35 **Nawata K, Sohmiya M, Kawaguchi M, Nishiki M, Kato Y.** Increased resting metabolic rate in patients with type 2 diabetes mellitus accompanied by advanced diabetic nephropathy. *Metabolism* 2004; **53**: 1395-1398
 - 36 **De Luis DA, Aller R, Izaola O.** Resting energy expenditure and insulin resistance in obese patients, differences in women and men. *Eur Rev Med Pharmacol Sci* 2006; **10**: 285-289
 - 37 **da Rocha EE, Alves VG, da Fonseca RB.** Indirect calorimetry: methodology, instruments and clinical application. *Curr Opin Clin Nutr Metab Care* 2006; **9**: 247-256

S- Editor Li JY L- Editor A E- Editor Zheng XM

Acknowledgments to reviewers of World Journal of Experimental Medicine

We acknowledge our sincere thanks to our reviewers. Many reviewers have contributed their expertise and time to the peer review, a critical process to ensure the quality of our World Series Journals. Both the editors of the journals and authors of the manuscripts submitted to the journals are grateful to the following reviewers for reviewing the articles (either published or rejected) over the past period of time.

Jan Bernardy, MVD, PhD, Assistant Professor, Swine Clinic, Veterinary Faculty, Palackého str No. 1, 61300 Brno, Czech Republic

Nadia Alfaidy, PhD, iRTSV-Biology of Cancer and infection, INSERM U1036, 17 rue des Martyrs, 38054 Grenoble, France

Sorin Armeanu-Ebinger, Professor, Department of Pediatric Surgery, Children's Hospital, University of Tuebingen, Hoppe-Seyler-Str.3, 72076 Tübingen, Germany

Effie K Basdra, DMD, Associate Professor, Department of Biological Chemistry, University of Athens Medical School, 75 M. Asias street – Goudi, GR-11527 Athens, Greece

Arun Bhunia, BVSc, PhD, Professor of Molecular Food Microbiology, Department of Food Science, Department of Comparative Pathobiology, Purdue University, 745 Agriculture Mall Dr., West Lafayette, IN 47907, United States

Steven G Gray, PhD, Translational Cancer Research Group, Department of Clinical Medicine, Trinity Centre for Health Sciences, Rm 2.103, Institute of Molecular Medicine, St James's Hospital, Dublin 8, Ireland

Winn Aung, MBBS, PhD, Diagnostic Imaging Program, Molecular Imaging Center, National Institute of Radiological Sciences, 4-9-1 Anagawa, Inage-ku, Chiba 263-8555, Japan

Gam Lay Harn, PhD, Associate Professor, School of Pharmaceutical Sciences, Universiti Sains Malaysia, Minden 11800, Penang, Malaysia

Dalwoong Choi, Associate Professor, Department of Environmental Health, College of Health Sciences, Korea University, JungLeung-3-Dong, SungBook-Gu, Seoul 136-703, South Korea

Karl O Fagerstrom, PhD, Associate Professor, Fagerstrom Consulting, Jordkull 3670, Kagerod 26878, Sweden

Ali Kudret Adiloglu, MD, Associate Professor, S.B. Ankara Egitim ve Arastirma Hastanesi, Mikrobiyoloji Laboratuvarı Klinik left, Ulucanlar Cd., 19. Sok. No: 16/6 Israil Evleri 06500, Emek, Ankara, Turkey

Anshu Agrawal, PhD, Associate Adjunct Professor, C-240A, Med/Sci-I, Division of Basic and Clinical Immunology, Department of Medicine, University of California, Irvine, CA 92697, United States

Events Calendar 2012

January 15-20, 2012
Fungal Pathogens: From Basic
Biology to Drug
Santa Fe, NM, United States

January 20-20, 2012
Exploiting Bacteriophages for
Bioscience, Biotechnology and
Medicine
London, United Kingdom

January 22-27, 2012
Biology of Spirochetes
Ventura, CA, United States

February 7-12, 2012
Gene Silencing by Small RNAs
Vancouver, British Columbia,
Canada

March 4 -10, 2012
Malaria Experimental Genetics
Hinnton, Cambridge,
United Kingdom

March 12-13, 2012
2nd Annual International
Conference on Bioinformatics and
Computational Biology Special
Track: Stem Cell Research
Singapore

March 12-13, 2012
BICB 2012: 2nd Annual International
Conference on Bioinformatics and
Computational Biology (updated)
Global Science and Technology
Forum
Thailand

March 18-21, 2012
Annual Conference of the
Association for General and Applied
Microbiology
Tubingen, Germany

March 31-April 3, 2012
22nd European Congress of Clinical
Microbiology and Infectious
Diseases ECCMID
London, United Kingdom

April 2-4, 2012
Electron transfer at the microbe-
mineral interface
Norwich, United Kingdom

April 18, 2012
6th Broadening Microbiology
Horizons in Biomedical Science
Meeting
Stratford-Upon-Avon,
United Kingdom

May 6-12, 2012
4th ASM Conference on Prokaryotic

Cell Biology and Development
Montreal, Canada

May 7-19, 2012
Bioinformatics and comparative
genomes analyses
Napoli, Italy

May 8 - 10, 2012
Exploring Human Host-Microbiome
Interactions in Health and Disease
Cambridge, United Kingdom

May 30-31, 2012
European Lab Automation
Hamburg, Germany

June 3-8, 2012
Anaerobes in Health and Disease;
How to Isolate, Identify and Look
for Resistance in a Cost-Effective
Way
Szeged, Hungary

June 16-21, 2012
Gene transcription in yeast
Girona, Spain

June 21-22, 2012
Swiss Joint Annual Meeting
St. Gallen, Switzerland

July 22-27, 2012
15th International Conference on

Experimental Mechanics
University of Porto
Portugal

July 29-August 2, 2012
XV IS-MPMI Kyoto 2012.
International Congress on Molecular
Plant-Microbe Interactions
Kyoto, Japan

August 18-22, 2012
The 30th World Congress of
Biomedical Laboratory Science
Berlin, Germany

August 25-September 1, 2012
Update on Indications, Interactions
and Complications in the Use of
Pharmaceuticals
Honolulu, Hawaii, United States

September 7-9, 2012
International Congress of Maritime
Medicine
Odessa, Ukraine

October 14-24, 2012
Medical Ethics and Legal Medicine
Miami, Florida, United States

December 1-6, 2012
Qatar Health 2012
Doha, Qatar

GENERAL INFORMATION

World Journal of Experimental Medicine (*World J Exp Med*, *WJEM*, online ISSN 2220-315X, DOI: 10.5493) is a bimonthly peer-reviewed, online, open-access (OA), journal supported by an editorial board consisting of 104 experts in experimental medicine from 30 countries.

The biggest advantage of the OA model is that it provides free, full-text articles in PDF and other formats for experts and the public without registration, which eliminates the obstacle that traditional journals possess and usually delays the speed of the propagation and communication of scientific research results. The open access model has been proven to be a true approach that may achieve the ultimate goal of the journals, i.e. the maximization of the value to the readers, authors and society.

Maximization of personal benefits

The role of academic journals is to exhibit the scientific levels of a country, a university, a center, a department, and even a scientist, and build an important bridge for communication between scientists and the public. As we all know, the significance of the publication of scientific articles lies not only in disseminating and communicating innovative scientific achievements and academic views, as well as promoting the application of scientific achievements, but also in formally recognizing the "priority" and "copyright" of innovative achievements published, as well as evaluating research performance and academic levels. So, to realize these desired attributes of *WJEM* and create a well-recognized journal, the following four types of personal benefits should be maximized. The maximization of personal benefits refers to the pursuit of the maximum personal benefits in a well-considered optimal manner without violation of the laws, ethical rules and the benefits of others. (1) Maximization of the benefits of editorial board members: The primary task of editorial board members is to give a peer review of an unpublished scientific article via online office system to evaluate its innovativeness, scientific and practical values and determine whether it should be published or not. During peer review, editorial board members can also obtain cutting-edge information in that field at first hand. As leaders in their field, they have priority to be invited to write articles and publish commentary articles. We will put peer reviewers' names and affiliations along with the article they reviewed in the journal to acknowledge their contribution; (2) Maximization of the benefits of authors: Since *WJEM* is an OA journal, readers around the world can immediately download and read, free of charge, high-quality, peer-reviewed articles from *WJEM* official website, thereby realizing the goals and significance of the communication between authors and peers as well as public reading; (3) Maximization of the benefits of readers: Readers can read or use, free of charge, high-quality peer-reviewed articles without any limits, and cite the arguments, viewpoints, concepts, theories, methods, results, conclusion or facts and data of pertinent literature so as to validate the innovativeness, scientific and practical values of their own research achievements, thus ensuring that their articles have novel arguments or viewpoints, solid evidence and correct conclusion; and (4) Maximization of the benefits of employees: It is an iron law that a first-class journal is unable to exist without first-class editors, and only first-class editors can create a first-class academic journal. We insist on strengthening our team cultivation and construction so that every employee, in an open, fair and transparent environment, could contribute their wisdom to edit and publish high-quality articles,

thereby realizing the maximization of the personal benefits of editorial board members, authors and readers, and yielding the greatest social and economic benefits.

Aims and scope

WJEM aims to rapidly report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of experimental medicine. *WJEM* covers topics concerning clinical laboratory medicine (applied and basic research in hematology, body fluid examination, cytomorphology, genetic diagnosis of hematological disorders, thrombosis and hemostasis, and blood typing and transfusion), biochemical examination (applied and basic research in laboratory automation and information system, biochemical methodology, and biochemical diagnostics), clinical microbiology (microbiological laboratory quality control and management; microbiological specimen collection and its influencing factors; conventional, automatic or molecular detection of clinical microorganisms; monitoring of bacterial and fungal drug resistance, drug resistance mechanisms, and rational application of antibiotics; monitoring and control of nosocomial infections), immunodiagnostics (laboratory diagnosis of infectious diseases, tumor markers and their application, laboratory diagnosis of autoimmune diseases, and immunotechnology), clinical laboratory management (laboratory quality control and management, traceability and calibration, information management system and laboratory automation, and laboratory biosafety management), and experimental medicine-related traditional medicine, and integrated Chinese and Western medicine. The journal also publishes original articles and reviews that report the results of experimental medicine-related applied and basic research in fields such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

Columns

The columns in the issues of *WJEM* will include: (1) Editorial: To introduce and comment on the substantial advance and its importance in the fast-developing areas; (2) Frontier: To review the most representative achievements and comment on the current research status in the important fields, and propose directions for the future research; (3) Topic Highlight: This column consists of three formats, including (A) 10 invited review articles on a hot topic, (B) a commentary on common issues of this hot topic, and (C) a commentary on the 10 individual articles; (4) Observation: To update the development of old and new questions, highlight unsolved problems, and provide strategies on how to solve the questions; (5) Guidelines for Clinical Practice: To provide guidelines for clinical diagnosis and treatment; (6) Review: To systemically review the most representative progress and unsolved problems in the major scientific disciplines, comment on the current research status, and make suggestions on the future work; (7) Original Articles: To originally report the innovative and valuable findings in experimental medicine; (8) Brief Articles: To briefly report the novel and innovative findings in experimental medicine; (9) Case Report: To report a rare or typical case; (10) Letters to the Editor: To discuss and make reply to the contributions published in *WJEM*, or to introduce and comment on a controversial issue of general interest; (11) Book Reviews: To introduce and comment on quality monographs of experimental medicine; and (12) Guidelines: To introduce consensus and guidelines reached by international and national academic authorities worldwide on the research in experimental medicine.

Instructions to authors

Name of journal

World Journal of Experimental Medicine

ISSN

ISSN 2220-315X (online)

Editor-in-Chief

De-Ling Kong, PhD, Professor, Institute of Molecular Biology, Nankai University, Tianjin 300071, China

Atsushi Mizoguchi, MD, PhD, Associate Professor in Pathology, Harvard Medical School, Molecular Pathology Unit, Massachusetts General Hospital, CNY149-6024, 13th Steert, Charlestown, MA 02114, United States

Baohong Zhang, PhD, Assistant Professor of Biology, Department of Biology, East Carolina University, Greenville, NC 27858, United States

Editorial Office

World Journal of Experimental Medicine

Editorial Department: Room 903, Building D,
Ocean International Center,
No. 62 Dongsihuan Zhonglu,
Chaoyang District, Beijing 100025, China
E-mail: wjem@wjnet.com
<http://www.wjnet.com>
Telephone: +86-10-85381891
Fax: +86-10-85381893

Indexed and Abstracted in

Digital Object Identifier.

Published by

Baishideng Publishing Group Co., Limited

SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJEM* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names

of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submis-

sion System at: <http://www.wjgnet.com/esps/>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/2220-315x/g_info_20100722180909.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjem@wjgnet.com, or by telephone: +86-10-85381891. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g. Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g. Telephone: +86-10-85381892 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision for acceptance is made only when at least two experts recommend an article for publication. Reviewers for accepted manuscripts are

acknowledged in each manuscript, and reviewers of articles which were not accepted will be acknowledged at the end of each issue. To ensure the quality of the articles published in *WJR*, reviewers of accepted manuscripts will be announced by publishing the name, title/position and institution of the reviewer in the footnote accompanying the printed article. For example, reviewers: Professor Jing-Yuan Fang, Shanghai Institute of Digestive Disease, Shanghai, Affiliated Renji Hospital, Medical Faculty, Shanghai Jiaotong University, Shanghai, China; Professor Xin-Wei Han, Department of Radiology, The First Affiliated Hospital, Zhengzhou University, Zhengzhou, Henan Province, China; and Professor Anren Kuang, Department of Nuclear Medicine, Huaxi Hospital, Sichuan University, Chengdu, Sichuan Province, China.

Abstract

There are unstructured abstracts (no more than 256 words) and structured abstracts (no more than 480). The specific requirements for structured abstracts are as follows:

An informative, structured abstracts of no more than 480 words should accompany each manuscript. Abstracts for original contributions should be structured into the following sections. AIM (no more than 20 words): Only the purpose should be included. Please write the aim as the form of "To investigate/study/...; MATERIALS AND METHODS (no more than 140 words); RESULTS (no more than 294 words): You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g. 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$; CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both. The main text format of these sections, editorial, topic highlight, case report, letters to the editors, can be found at: http://www.wjgnet.com/2220-315x/g_info_20100725072755.htm.

Illustrations

Figures should be numbered as 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Figures should be either Photoshop or Illustrator files (in tiff, eps, jpeg formats) at high-resolution. Examples can be found at: <http://www.wjgnet.com/1007-9327/13/4520.pdf>; <http://www.wjgnet.com/1007-9327/13/4554.pdf>; <http://www.wjgnet.com/1007-9327/13/4891.pdf>; <http://www.wjgnet.com/1007-9327/13/4986.pdf>; <http://www.wjgnet.com/1007-9327/13/4498.pdf>. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ... *etc.* It is our principle to publish high resolution-figures for the printed and E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any

Instructions to authors

footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a*P* < 0.05, ^b*P* < 0.01 should be noted (*P* > 0.05 should not be noted). If there are other series of *P* values, ^c*P* < 0.05 and ^d*P* < 0.01 are used. A third series of *P* values can be expressed as ^e*P* < 0.05 and ^f*P* < 0.01. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, etc., in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g. PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaobua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS-A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. Emerg Infect Dis serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h, blood glucose concentration, *c* (glucose) 6.4 \pm 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23243641.

The format for how to accurately write common units and quantum numbers can be found at: http://www.wjgnet.com/2220-315x/g_info_20100725073806.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, *etc.*

Biology: *H. pylori*, *E. coli*, *etc.*

Examples for paper writing

Editorial: http://www.wjgnet.com/2220-315x/g_info_20100725071851.htm

Frontier: http://www.wjgnet.com/2220-315x/g_info_20100725071932.htm

Topic highlight: http://www.wjgnet.com/2220-315x/g_info_20100725072121.htm

Observation: http://www.wjgnet.com/2220-315x/g_info_20100725072232.htm

Guidelines for basic research: http://www.wjgnet.com/2220-315x/g_info_20100725072344.htm

Guidelines for clinical practice: http://www.wjgnet.com/2220-315x/g_info_20100725072543.htm

Review: http://www.wjgnet.com/2220-315x/g_info_20100725072656.htm

Original articles: http://www.wjgnet.com/2220-315x/g_info_20100725072755.htm

Brief articles: http://www.wjgnet.com/2220-315x/g_info_20100725072920.htm

Case report: http://www.wjgnet.com/2220-315x/g_info_20100725073015.htm

Letters to the editor: http://www.wjgnet.com/2220-315x/g_info_20100725073136.htm

Book reviews: http://www.wjgnet.com/2220-315x/g_info_20100725073214.htm

Guidelines: http://www.wjgnet.com/2220-315x/g_info_20100725073300.htm

SUBMISSION OF THE REVISED MANUSCRIPTS AFTER ACCEPTED

Please revise your article according to the revision policies of *WJEM*. The revised version including manuscript and high-resolution image figures (if any) should be re-submitted online (<http://www.wjgnet.com/2220-315xoffice/>). The author should send the copyright transfer letter, responses to the reviewers, English language Grade B certificate (for non-native speakers of English) and final manuscript checklist to wjem@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A or B.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2220-315x/g_info_20100725073726.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2220-315x/g_info_20100725073445.htm.

Proof of financial support

For paper supported by a foundation, authors should provide a copy of the document and serial number of the foundation.

Links to documents related to the manuscript

WJEM will be initiating a platform to promote dynamic interactions between the editors, peer reviewers, readers and authors. After a manuscript is published online, links to the PDF version of the submitted manuscript, the peer-reviewers' report and the revised manuscript will be put on-line. Readers can make comments on the peer reviewer's report, authors' responses to peer reviewers, and the revised manuscript. We hope that authors will benefit from this feedback and be able to revise the manuscript accordingly in a timely manner.

Science news releases

Authors of accepted manuscripts are suggested to write a science news item to promote their articles. The news will be released rapidly at EurekAlert/AAAS (<http://www.eurekalert.org>). The title for news items should be less than 90 characters; the summary should be less than 75 words; and main body less than 500 words. Science news items should be lawful, ethical, and strictly based on your original content with an attractive title and interesting pictures.

Publication fee

WJEM is an international, peer-reviewed, OA, online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 600 USD per article. Editorial, topic highlights, book reviews and letters to the editor are published free of charge.