

World Journal of *Clinical Pediatrics*

World J Clin Pediatr 2013 November 8; 2(4): 31-76



Editorial Board

2012-2016

The *World Journal of Clinical Pediatrics* Editorial Board consists of 247 members, representing a team of worldwide experts in pediatrics. They are from 43 countries, including Argentina (1), Australia (7), Austria (4), Belgium (2), Brazil (4), Canada (7), Chile (2), China (22), Denmark (2), Egypt (10), Finland (1), France (5), Germany (4), Greece (8), India (14), Iran (5), Israel (7), Italy (22), Japan (6), Mexico (2), Netherlands (2), New Zealand (1), Nigeria (3), Norway (1), Pakistan (2), Poland (2), Portugal (1), Russia (2), Saudi Arabia (2), Serbia (2), Singapore (3), Slovenia (1), South Africa (2), South Korea (2), Spain (5), Sweden (4), Switzerland (1), Thailand (2), Tunisia (1), Turkey (18), United Arab Emirates (1), United Kingdom (11), United States (43).

EDITOR-IN-CHIEF

Eduardo H Garin, *Gainesville*

GUEST EDITORIAL BOARD MEMBERS

Hsiao-Wen Chen, *Taipei*
Ming-Ren Chen, *Taipei*
Mu-Kuan Chen, *Changhua*
Ching-Chi Chi, *Chiayi*
Hung-Chih Lin, *Taichung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Alcides Richard Troncoso, *Buenos Aires*



Australia

Garry Inglis, *Herston*
Jagat Kanwar, *Victoria*
Katherine Kedzierska, *Parkville*
Eline Suzanne Klaassens, *Brisbane*
Sam S Mehr, *Sydney*
Jing Sun, *Brisbane*
Cuong Duy Tran, *Adelaide*



Austria

Gerhard Cvirn, *Graz*
Claudia Elisabeth Gundacker, *Vienna*
Bernhard Resch, *Graz*
Amulya K Saxena, *Graz*



Belgium

Karel Allegaert, *Leuven*

Yvan Vandenplas, *Brussels*



Brazil

Rejane Correa Marques, *Rio de Janeiro*
Priscila Krauss Pereira, *Rio de Janeiro*
Maria L Seidl-de-Moura, *Rio de Janeiro*
Sandra Elisabete Vieira, *São Paulo*



Canada

Helen SL Chan, *Toronto*
Ediriweera Desapriya, *Vancouver*
Eleftherios P Diamandis, *Toronto*
Ran D Goldman, *Vancouver*
Manjula Gowrishankar, *Edmonton*
Prakesh S Shah, *Toronto*
Pia Wintermark, *Montreal*



Chile

René Mauricio Barria, *Valdivia*
Irene Morales Bozo, *Santiago*



China

Yu-Zuo Bai, *Shenyang*
Xiao-Ming Ben, *Nanjing*
Kwong-Leung Chan, *Hong Kong*
Xian-Hui He, *Guangzhou*
Jian Hu, *Harbin*
Xi-Tai Huang, *Tianjin*
Huang-Xian Ju, *Nanjing*
Ren Lai, *Kunming*
Li Liu, *Xi'an*

Xue-Qun Luo, *Guangzhou*

Ai-Guo Ren, *Beijing*
Chiu-Lai Shan, *Hong Kong*
Yuk Him Tam, *Hong Kong*
Jin-Xing Wang, *Jinan*
Jun-Jun Wang, *Beijing*
Long-Jiang Zhang, *Nanjing*
Yi-Hua Zhou, *Nanjing*



Denmark

Jesper Bo Nielsen, *Odense*
Ole D Wolthers, *Randers*



Egypt

Mosaad Abdel-Aziz, *Cairo*
Hesham E Abdel-Hady, *Mansoura*
Mohammed Al-Biltagi, *Tanta*
Mohammad MS Al-Hagggar, *Mansoura*
Ashraf MAB Bakr, *Mansoura*
Badr Eldin Mostafa, *Cairo*
Rania Refaat, *Cairo*
Omar Mamdouh Shaaban, *Assiut*
Maysaa El Sayed Zaki, *Mansoura*
Magdy Mohamed Zedan, *Mansoura*



Finland

Bright Ibeabughichi Nwaru, *Tampere*



France

Philippe Georgel, *Strasbourg*
Grill Jacques, *Villejuif*

Manuel Lopez, *Saint Etienne*
Georgios Stamatias, *Issy-les-Moulineaux*
Didier Vieau, *Villeneuve d'Ascq*



Germany

Yeong-Hoon Choi, *Cologne*
Carl Friedrich Classen, *Rostock*
Stephan Immenschuh, *Hannover*
Ales Janda, *Freiburg im Breisgau*



Greece

Michael B Anthracopoulos, *Rion-Patras*
Savas Grigoriadis, *Thessaloniki*
Vasiliki-Maria Iliadou, *Thessaloniki*
Theofilos M Kolettis, *Ioannina*
Ariadne Malamitsi-Puchner, *Athens*
Dimitrios Papandreou, *Thessaloniki*
Kostas N Priftis, *Athens*
Ioannis Michael Vlastos, *Heraklion*



India

Amit Agrawal, *Ambala*
Sameer Bakhshi, *New Delhi*
Atmaram H Bandivdekar, *Mumbai*
Sandeep Bansal, *Chandigarh*
Sriparna Basu, *Varanasi*
Ashu Seith Bhalla, *New Delhi*
Sushil Kumar Kabra, *New Delhi*
Praveen Kumar, *New Delhi*
Kaushal Kishor Prasad, *Chandigarh*
Yogesh Kumar Sarin, *New Delhi*
Kushaljit Singh Sodhi, *Chandigarh*
Raveenthiran V Venkatachalam, *Tamilnadu*
B Viswanatha, *Bangalore*
Syed Ahmed Zaki, *Mumbai*



Iran

Mehdi Bakhshae, *Mashhad*
Maria Cheraghi, *Ahvaz*
Mehran Karimi, *Shiraz*
Samileh Noorbakhsh, *Tehran*
Firoozeh Sajedi, *Tehran*



Israel

Shraga Aviner, *Ashkelon*
Aviva Fattal-Valevski, *Ramat Aviv*
Rafael Gorodischer, *Omer*
Gil Klinger, *Petah Tikva*
Asher Ornoy, *Jerusalem*
Giora Pillar, *Haifa*
Yehuda Shoenfeld, *Tel-Hashomer*



Italy

Roberto Antonucci, *Cagliari*
CarloV Bellieni, *Siena*
Silvana Cicala, *Naples*
Sandro Contini, *Parma*

Enrico Stefano Corazziari, *Rome*
Vincenzo Cuomo, *Rome*
Vassilios Fanos, *Cagliari*
Filippo Festini, *Florence*
Irene Figa-Talamanca, *Rome*
Dario Galante, *Foggia*
Fabio Grizzi, *Milan*
Alessandro Inserra, *Rome*
Achille Iolascon, *Naples*
Cantinotti Massimiliano, *Pietranta*
Ornella Milanese, *Padova*
Giovanni Nigro, *L'Aquila*
Giuseppe Rizzo, *Rome*
Claudio Romano, *Messina*
Mario Santinami, *Milano*
Gianluca Terrin, *Rome*
Alberto Tommasini, *Trieste*
Giovanni Vento, *Rome*



Japan

Ryo Aeba, *Tokyo*
Kazunari Kaneko, *Osaka*
Hideaki Senzaki, *Saitama*
Kohichiro Tsuji, *Tokyo*
Toru Watanabe, *Niigata*
Takayuki Yamamoto, *Yokkaichi*



Mexico

Fernando Guerrero-Romero, *Durango*
Mara Medeiros, *Mexico*



Netherlands

Jacobus Burggraaf, *Leiden*
Paul Eduard Sijens, *Groningen*



New Zealand

Simon James Thornley, *Auckland*



Nigeria

Akeem Olawale Lasisi, *Ibadan*
Tinuade Adetutu Ogunlesi, *Sagamu*
Joseph Ubini Ese Onakewhor, *Benin*



Norway

Lars T Fadnes, *Bergen*



Pakistan

Niloufer Sultan Ali, *Karachi*
Shakila Zaman, *Lahore*



Poland

Piotr Czuderna, *Gdansk*
Joseph Prandota, *Wroclaw*



Portugal

Alexandre M Carmo, *Porto*



Russia

Perepelitsa S Alexandrovna, *Kaliningrad*
Vorsanova Svetlana, *Moscow*



Saudi Arabia

Naser Labib Rezk, *Riyadh*
Amna Rehana Siddiqui, *Riyadh*



Serbia

Bjelakovic Borisav Bojko, *Nis*
Mirela Erić, *Novi Sad*



Singapore

Quak Seng Hock, *Singapore*
Anselm CW Lee, *Singapore*
Alvin Soon Tiong Lim, *Singapore*



Slovenia

Rok Orel, *Ljubljana*



South Africa

David Kenneth Stones, *Free State*
Eric Oghenerioborue Udjo, *Pretoria*



South Korea

Byung-Ho Choe, *Daegu*
Dong-Hee Lee, *Seoul*



Spain

Pilar Codoñer-Franch, *Valencia*
Claudio Golfier, *Barcelona*
Pablo Menendez, *Andalucía*
Juan F Martínez-Lage Sánchez, *Murcia*
Juan Antonio Tovar, *Madrid*



Sweden

Moustapha Hassan, *Stockholm*
Maria Christina Jenmalm, *Linköping*
Sandra Kleinau, *Uppsala*
Birgitta Lindberg, *Luleå*



Switzerland

Ulf Kessler, *Bern*

**Thailand**

Surasak Sangkhathat, *Hat Yai*
Viroj Wiwanitkit, *Bangkok*

**Tunisia**

John C Anyanwu, *Tunis Belvedere*

**Turkey**

Sinem Akgül, *Ankara*
Ayse Tuba Altug, *Ankara*
Suna Asilsoy, *Seyhan-Adana*
Ozgu Aydogdu, *Izmir*
Kadir Babaoglu, *Kocaeli*
Aksoy Berna, *Kocaeli*
Murat Biteker, *Istanbul*
Merih Çetinkaya, *Istanbul*
Aynur Emine Cicekcibasi, *Konya*
Elvan Caglar Citak, *Mersin*
Cem Dane, *Istanbul*
Mintaze Kerem Günel, *Ankara*
Ahmet Güzel, *Samsun*
Salih Kavukcu, *Izmir*
Fethullah Kenar, *Denizli*
Selim Kurtoglu, *Kayseri*
Turker Ozyigit, *Istanbul*
Yalçın Tüzün, *Istanbul*

**United Arab Emirates**

Iradj Amirlak, *Al Ain*

**United Kingdom**

Keith Collard, *Plymouth*
ASahib El-Radhi, *London*
Edzard Ernst, *Exeter*
Mohammad K Hajihosseini, *Norwich*
Tain-Yen Hsia, *London*
Claudio Nicoletti, *Norwich*
Cordula Margaret Stover, *Leicester*
Alastair Gordon Sutcliffe, *London*
Adrian Graham Thomas, *Manchester*
Richard Trompeter, *London*
Petros V Vlastarakos, *Stevenage*

**United States**

Stephen C Aronoff, *Philadelphia*
Hossam M Ashour, *Detroit*
Paul Ashwood, *Sacramento*
David C Bellinger, *Boston*
Vineet Bhandari, *New Haven*
FR Breijo-Marquez, *Boston*
Itzhak Brook, *Washington*
Patrick D Brophy, *Iowa*
Lavjay Butani, *Sacramento*

Archana Chatterjee, *Omaha*
Lisa M Cleveland, *San Antonio*
Shri R Deshpande, *Atlanta*
Michael Morgan Dowling, *Dallas*
Abdulrahman M El-Sayed, *New York*
Donald N Forthal, *Irvine*
Gregory Kane Friedman, *Birmingham*
Kenneth William Gow, *Seattle*
Dorothy I Bulas, *Washington*
Christopher L Coe, *Madison*
Elias Jabbour, *Houston*
Michael Van Doren Johnston, *Baltimore*
Ram V Kalpatthi, *Gainesville*
Stephen S Kim, *Annandale*
Edward Yungjae Lee, *Annandale*
Jing Lin, *New York*
Jorge Lopez, *Gainesville*
Aurelia Meloni-Ehrig, *Gainesville*
Murielle Mimeault, *Omaha*
Natan Noviski, *Omaha*
Michael David Seckeler, *Charlottesville*
Chetan Chandulal Shah, *Little Rock*
Mohamed Tarek M Shata, *Cincinnati*
Tsz-Yin So, *Greensboro*
Dennis Charles Stevens, *Sioux Falls*
Ru-Jeng Teng, *Milwaukee*
Rajan Wadhawan, *St Petersburg*
Hongjun Wang, *St Charleston*
Marie Wang, *Menlo Park*
Richard Wang, *Atlanta*
Wladimir Wertelecki, *Annandale*
Shu Wu, *Miami*
Fadi Xu, *Albuquerque*

Contents

Quarterly Volume 2 Number 4 November 8, 2013

EDITORIAL	31	Indoor smoke and prenatal and childhood growth: The role of (gestational) age <i>Ghosh R</i>
REVIEW	36	Echocardiography in children with Down syndrome <i>Al-Biltagi M</i>
	46	Bacterial colonization and intestinal mucosal barrier development <i>Huang XZ, Zhu LB, Li ZR, Lin J</i>
MINIREVIEWS	54	Imaging evaluation of hemoptysis in children <i>Singh D, Bhalla AS, Thotton Veedu P, Arora A</i>
BRIEF ARTICLE	65	Tramadol use in pediatric sickle cell disease patients with vaso-occlusive crisis <i>Borgerding MP, Absher RK, So TY</i>
	70	Pediatric vs adult pulmonary tuberculosis: A retrospective computed tomography study <i>Thotton Veedu P, Bhalla AS, Vishnubhatla S, Kabra SK, Arora A, Singh D, Gupta AK</i>

APPENDIX I-V Instructions to authors

ABOUT COVER Editorial Board Member of *World Journal of Clinical Pediatrics*, Mohammed A Al-Biltagi, MD, PhD, Associate Professor of Paediatrics, Paediatric Department, Faculty of Medicine, Tanta University, El Bahr Str, Tanta 31527, Egypt

AIM AND SCOPE *World Journal of Clinical Pediatrics (World J Clin Pediatr, WJCP, online ISSN 2219-2808, DOI: 10.5409)* is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJCP covers a variety of clinical medical topics, including fetal diseases, inborn, newborn diseases, infant diseases, genetic diseases, diagnostic imaging, endoscopy, and evidence-based medicine and epidemiology. Priority publication will be given to articles concerning diagnosis and treatment of pediatric diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to *WJCP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

INDEXING/ABSTRACTING *World Journal of Clinical Pediatrics* is now indexed in Digital Object Identifier.

FLYLEAF I-III Editorial Board

EDITORS FOR THIS ISSUE Responsible Assistant Editor: *Xin-Xin Che* Responsible Science Editor: *Huan-Huan Zhai*
 Responsible Electronic Editor: *Ya-Jing Lu*
 Proofing Editor-in-Chief: *Lian-Sheng Ma*

NAME OF JOURNAL
World Journal of Clinical Pediatrics

ISSN
 ISSN 2219-2808 (online)

LAUNCH DATE
 June 8, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Eduardo H Garin, MD, Professor, Department of Pediatrics, University of Florida, 1600 SW Archer Road, HD214, Gainesville, FL 32610, United States

EDITORIAL OFFICE
 Jin-Lei Wang, Director
 Xiu-Xia Song, Vice Director
World Journal of Clinical Pediatrics

Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
 Telephone: +86-10-85381891
 Fax: +86-10-85381893
 E-mail: wjcp@wjnet.com
<http://www.wjnet.com>

PUBLISHER
 Baishideng Publishing Group Co., Limited
 Flat C, 23/F, Lucky Plaza,
 315-321 Lockhart Road, Wan Chai,
 Hong Kong, China
 Fax: +852-6555-7188
 Telephone: +852-3177-9906
 E-mail: bpgoffice@wjnet.com
<http://www.wjnet.com>

PUBLICATION DATE
 November 8, 2013

COPYRIGHT
 © 2013 Baishideng. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjnet.com/2219-2808/g_info_20100722180909.htm.

ONLINE SUBMISSION
<http://www.wjnet.com/esp/>

Indoor smoke and prenatal and childhood growth: The role of (gestational) age

Rakesh Ghosh

Rakesh Ghosh, Division of Environmental Health, University of Southern California, Los Angeles, CA 90032, United States

Author contributions: Ghosh R solely contributed to this paper.

Correspondence to: Rakesh Ghosh, Division of Environmental Health, University of Southern California, Room 213, 2001 N Soto St., Los Angeles, CA 90032,

United States. rakeshgh@usc.edu

Telephone: +1-323-442-8272 Fax: +1-323-442-3272

Received: February 18, 2013 Revised: July 1, 2013

Accepted: July 9, 2013

Published online: November 8, 2013

Abstract

Growth at birth and during infancy predicts several outcomes in the immediate future as well as in the long term. Weight and height are commonly used surrogates of growth, however, infants and young children are constantly growing unlike adults. Hence, weight and height alone are insufficient measures of growth if the time component is not associated with them. Recent studies have investigated the relationship between indoor air pollution and growth using height and weight. In this commentary, I have argued using a directed acyclic graph, that a causal association between indoor pollution exposure and growth at birth cannot be established unless birth weight is adjusted for gestational age. Furthermore, to make any causal inference between growth during the first few years of life and indoor exposure, in addition to age standardization, studies must also account for fetal growth to discount any continuation of prenatal effects, which may be in the causal pathway. A careful consideration is warranted from future studies investigating these relationships.

© 2013 Baishideng. All rights reserved.

Key words: Biofuel; Coal smoke; Wood smoke; Birth weight; Fetal growth; Height

Core tip: Prenatal and early childhood estimators of growth, such as birth weight, height *etc.* by themselves are inadequate measures for inter-individual comparison, unless accompanied by gestational or chronologic age. The existing evidence points toward an association between indoor air pollution and growth, however few considered age. In order to establish a causal relationship it is imperative to consider age adjusted growth.

Ghosh R. Indoor smoke and prenatal and childhood growth: The role of (gestational) age. *World J Clin Pediatr* 2013; 2(4): 31-35 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/31.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.31>

INTRODUCTION

Reduced growth prenatally and after birth, which is associated with childhood mortality and morbidity^[1] and also with chronic diseases during adulthood^[2], is considered a major problem in many developing countries. Coincidentally, indoor air pollution is also high in these countries^[3]. More than half of the world population, predominantly from developing countries use some form of biofuel. A few studies have presented evidence that indoor air pollution may be one of the hitherto unknown factors associated with reduced prenatal and early childhood growth. This is a commentary on the studies mostly from the last decade, focusing on residual confounding that may arise if (gestational) age is not accounted for. There are many other sources of bias that pose similar threats to internal validity, which are not the focus of this commentary.

PRENATAL GROWTH

Prenatal exposure to indoor biofuel smoke and birth weight has now been investigated in several populations.

A Zimbabwean study^[4] reported maternal exposure to combustion smoke from wood, dung or straw was associated with infants born with 175 g ($P < 0.01$) lower mean birth weight than those who resided in homes that used gas or electricity for cooking. In spite of the significant association, owing to the cross-sectional nature, it is difficult to establish if exposure preceded outcome. Furthermore, the association with birth weight may not be construed to be with prenatal growth, as it was not adjusted for gestational age.

A study on Guatemalan infants^[5] reported 63 g (95%CI: 1-126) reduction in birth weight for those exposed to wood smoke compared to infants from households that used electricity or gas. This cross-sectional study accounted for some important variables but not gestational age and the possibility of exposure misclassification exists. This was one of the earliest studies and the magnitude of the point estimate is on the lower side compared to the estimates of other studies conducted subsequently.

Another Guatemalan study^[6] reported 89 g (95%CI: -27-+204) higher birth weight and the association with low birth weight (LBW) was 0.74 (95%CI: 0.33-1.66) for infants born in families that used a chimney stove compared to infants born in families that used open fire. The study, though not sufficiently powered, provides higher quality of evidence owing to the longitudinal design but did not adjust for gestational age.

A population-based longitudinal study^[7] from India, designed from a randomized trial reported a 105 g (95%CI: 140-69) reduction of birth weight and a RR of 1.49 (95%CI: 1.25-1.77) for LBW amongst those prenatally exposed to wood or dung smoke compared to those unexposed. The estimate was adjusted for a range of covariates including secondhand tobacco smoke (SHS) and a surrogate of socioeconomic status (SES). This study provides stronger evidence because of the design, availability of information on and adjustment for potential confounders as well as for the power to detect a significant difference in the exposed and unexposed group.

Another population based retrospective cohort study^[8] from Pakistan reported adjusted OR of 1.64 (95%CI: 1.10-2.35) for LBW and -82 g (95%CI: -170-+9) reduction in birth weight (continuous) among those exposed to wood smoke compared to natural gas combustion exposure. The study accounted for a wide range of potential confounders directly, including tobacco use, maternal BMI *etc.* and others indirectly, using propensity scores. The association with LBW was significant, while the association with birth weight was not, even though, birth weight as a continuous variable would have more variations (than the dichotomy of LBW), and hence lend more power to the model, assuming both models had same number of observations. There was a degree of misclassification in growth when LBW (without gestational age) was considered because all LBW infants were not due to growth retardation. This dichotomy and

hence the misclassification was absent in the continuous birth weight model and may have been the cause for the statistically non-significant result. Interestingly, the study from India^[7] reported a RR of 1.21 (95%CI: 1.11-1.31) for small for gestational age (SGA, < 10 percentile of birth weight-for-gestational age), which was less than half of the LBW estimate (RR = 1.49) from the same study. SGA reflects retarded prenatal growth more appropriately than LBW or birth weight because it takes into account gestational age. It is appreciated that accurate gestational age measurement is a challenge in settings in which these studies were conducted.

A meta-analysis^[9] pooled all the estimates from the above studies (and one more), to summarize information and increase power. However, is it prudent to pool together estimates from cross-sectional and longitudinal studies? Perhaps, a non-significant test of heterogeneity is an argument in favor. If one digs deeper, even the two longitudinal studies, the models from which the LBW/birth weight estimates were used for pooling^[6,7], did not have a single covariate in common. Furthermore, it included two estimates from the Indian cohort study^[7] one for term LBW and the other for preterm LBW. The original article did not anywhere associate gestational age with the LBW estimate (RR = 1.49); presumably the authors performed additional stratified analyses but the estimates (term LBW in the meta-analysis and all LBW in the original paper) are strikingly similar. The comparison in the meta-analysis is essentially between term and preterm LBW-while the former is certainly due to intrauterine growth retardation, the latter is due to early parturition and may or may not be from growth retardation? A careful approach with particular attention to study characteristics and their differences, beyond any statistical test, should be considered before choosing the meta-analytic approach, to avoid spurious results.

POSTNATAL GROWTH

A few studies also investigated exposure to indoor smoke and early childhood growth. About two decades ago the first study^[10] provided evidence of an adverse association between exposure to smoke produced from gas burning during cooking and height at 10 years, a 3.3 cm reduction ($P < 0.03$) in the exposed compared to the unexposed. The cross-sectional study on Kuwaiti and European children was a modest attempt to explore the relationship by accounting for important determinants like SHS, SES, ethnicity *etc.* It acknowledged that there may have been selection bias because fewer Kuwaiti families participated and amongst those who did, a larger proportion used gas. The authors did not standardize height, which made the comparison amongst participants untenable unless they were all of 10 years when height was measured.

A seven country study^[11] using data from national surveys investigated exposure to biofuel smoke (wood/straw/dung *vs* electricity/gas/biogas/kerosene) and child's height-

for-age and reported -0.13 (95%CI: -0.19--0.07) SD units reduction in a multilevel analyses. The analytical strategy and the wide range of covariates (including population differences) make this important evidence except the fact that owing to the cross-sectional nature temporal precedence of exposure cannot be established. In a categorical analysis it reported an OR of 1.27 (95%CI: 1.02-1.59) for severe stunting, defined as $-3 \text{ SD} \leq Z < -2 \text{ SD}$, amongst exposed vs unexposed. Another study^[12] using data from Indian national family health survey reported larger association for severe stunting, OR of 1.90 (95%CI: 1.49-2.42) for those using biofuels compared to those who used cleaner fuels (definition similar to the previous one). Amongst the strength was the large number of covariates used but it did not account for multistage clustered sampling and there may be some unaccounted variance in the models affecting the significance.

Two longitudinal studies provide evidence of the association between exposure to biofuel generated smoke and reduced height-for-age. The study from India^[7] investigated growth at 6 mo and reported RR of 1.45 (95%CI: 1.20-1.75) for underweight ($< -2 \text{ SD}$, weight-for-age) for those exposed to wood and dung smoke compared to those unexposed and RR of 1.30 (95%CI: 1.06-1.60) for stunting ($< -2 \text{ SD}$, height-for-age) in the same comparison groups. As stated above, this is one of the high quality evidence we have so far. A second longitudinal study^[13] measured continuous height-for-age of children 36 mo old in Caucasian children from the Czech Republic and reported 1.3 cm reduction in height of 36 mo old children exposed to coal smoke compared to those who were unexposed. Retarded growth (from biofuels exposure or something else) may be initiated prenatally; such a condition needs to be accounted if we are to estimate the magnitude of a true causal association with early childhood growth. The Czech study adjusted for growth at birth using birth weight-for-gestational age-and-sex thus discounting for any such conditions initiated prenatally.

The currently available studies are heterogeneous-one longitudinal study^[13] reported continuous Z-scores while another^[7] reported both continuous Z-scores and dichotomous stunting. Furthermore, the definition of stunting was different from those used in the cross-sectional studies.

CONCLUSION

The underlying question is - can exposure to indoor biofuel smoke impede normal growth before and after birth? Residual confounding is an important issue that needs to be addressed to answer this question. In case of prenatal growth without the adjustment for gestational age none of the evidence can be concluded as causal. Additionally, maternal tobacco smoking^[14] and household ETS^[15] exposure are also established confounders. ETS adjustment may appear to be conservative because some of the constituents of tobacco smoke and indoor biofuel smoke are similar (*e.g.*, PAH, nitrogen oxides, PM_{2.5}), it is still a robust approach to eliminate the possibility of type

1 error and to estimate the true magnitude of the effect. The converse argument against the idea could be that this may lead to over-adjustment, masking or attenuating a true association.

Childhood height and weight change relatively rapidly with inherent age and sex differences. Standardization for age and sex using reference populations is therefore necessary to make the results generalizable. Additionally, in the developing countries malnourishment is an important factor for retarded childhood growth and studies investigating indoor air pollution and postnatal growth in these settings should also account for factors (*viz.* SES and morbidity) that cause malnourishment. Interestingly, all but one childhood growth study used age-standardized measures^[10], which is pertinent. If standardization by age is imperative for growth after birth, why is it not for prenatal growth, when it is well known that a week or even days of gestational age in the third trimester make a difference to birth weight? It will be inappropriate to establish causality between indoor air pollution exposure and prenatal growth, using birth weight (or LBW) without gestational age, which, at best, is a crude surrogate of growth.

Another key issue for studies on childhood growth is accounting for growth at birth. It is critical to differentiate reduction in growth due to the exogenous exposures (secondhand smoke or indoor air pollution) during childhood from that which is simply a continuation of a retarded trajectory initiated prenatally. A retarded trajectory initiated prenatally may be due to similar exposures accrued over the prenatal period or may be from other causes, *e.g.*, malnourished mother. Either way, it is important to make this distinction for an appropriate assessment of the magnitude of the association between indoor air pollution exposure and postnatal growth.

The key issues presented in the two paragraphs above can also be described using a Directed Acyclic Graph (DAG) shown in Figure 1. Age and gestational age are determinants of pre and postnatal growth, respectively and according to the definition of confounder these two variables need to be connected to exposure and outcome either through a directed or a backdoor path. Aside from the fact that higher age prolongs exposure, age or gestational age is not connected to indoor air pollution and the outcomes through any directed or unblocked backdoor paths in the DAG (Figure 1). However, age is absolutely necessary to measure growth when weight or height is used as the outcome. For example, an infant with 2800 g birth weight born at 42 wk does not have the same growth as another infant with the same weight but born at 37 wk. Therefore, it should not be mistaken that absence of any directed or backdoor path between age and exposure/outcome justifies its exclusion from consideration in the relationship. On the contrary, weight or height as an outcome is an insufficient measure of growth without age. Additionally, the DAG shows that postnatal growth has a backdoor path connecting prenatal growth and indoor air pollution, which suggests adjustment of prenatal growth is imperative in any model

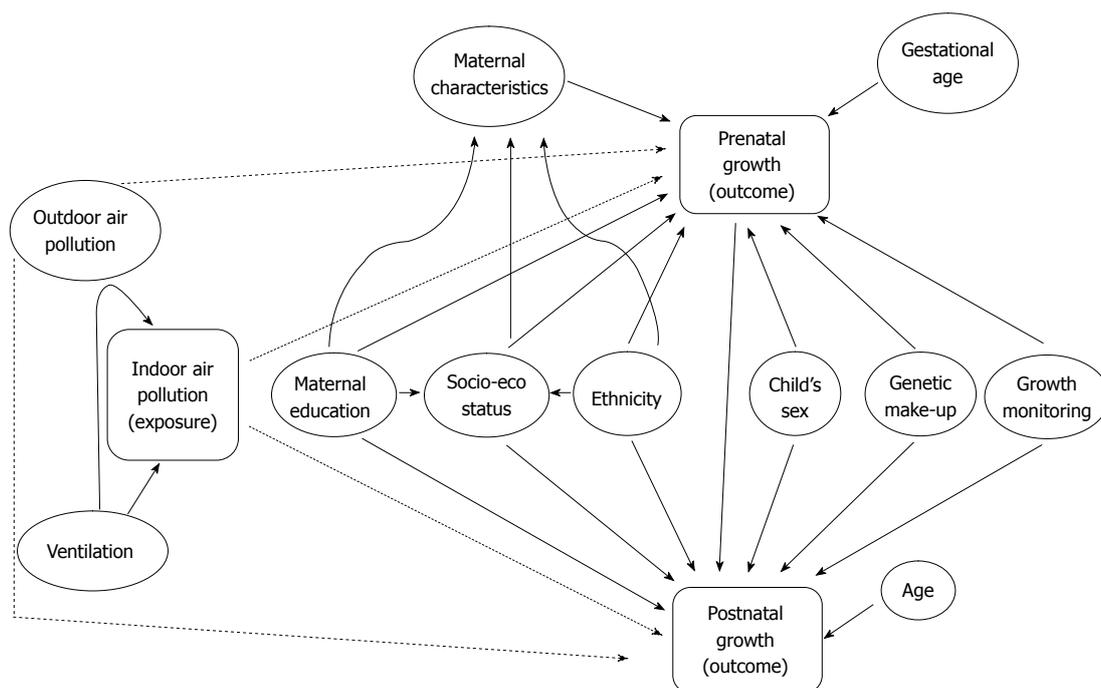


Figure 1 A directed acyclic graph depicting the relationship between indoor air and prenatal and early childhood growth.

associating indoor air pollution with postnatal growth.

The other variable that deserves a mention from the DAG (Figure 1) is outdoor air pollution. The causality of the relationship between outdoor air pollution and growth is yet to be ascertained. Meanwhile, outdoor air pollution will influence indoor air and vice versa if the windows and doors are kept open for long duration. This would apparently suggest that outdoor air pollution should be adjusted while investigating the relationship between indoor air pollution and growth. However, if outdoor and indoor air has the same pollutants and under the assumption of causal relationships, adjustment for one to investigate the relationship with the other, would be taking out the very association one is interested to find.

To conclude, the evidence point towards a potential adverse association between indoor air pollution exposure and growth, and encourages further well-designed investigation with adequate power, addressing the limitations of the current ones, to estimate the true magnitude. The last trimester and early childhood is marked by steady growth, it is therefore important to adjust for gestational age or chronologic age, respectively, to eliminate any differences in growth due to age before an adverse impact can be assessed. An exercise of procuring these datasets to perform a standardized analysis can be the next step forward.

REFERENCES

1 **McCormick MC.** The contribution of low birth weight to infant mortality and childhood morbidity. *N Engl J Med* 1985; **312**: 82-90 [PMID: 3880598 DOI: 10.1056/

NEJM198501103120204]
 2 **Whincup PH,** Kaye SJ, Owen CG, Huxley R, Cook DG, Anazawa S, Barrett-Connor E, Bhargava SK, Birgisdottir BE, Carlsson S, de Rooij SR, Dyck RF, Eriksson JG, Falkner B, Fall C, Forsén T, Grill V, Gudnason V, Hulman S, Hyppönen E, Jeffreys M, Lawlor DA, Leon DA, Minami J, Mishra G, Osmond C, Power C, Rich-Edwards JW, Roseboom TJ, Sachdev HS, Syddall H, Thorsdottir I, Vanhala M, Wadsworth M, Yarbrough DE. Birth weight and risk of type 2 diabetes: a systematic review. *JAMA* 2008; **300**: 2886-2897 [PMID: 19109117 DOI: 10.1001/jama.2008.886]
 3 **Bruce N,** Perez-Padilla R, Albalak R. Indoor air pollution in developing countries: a major environmental and public health challenge. *Bull World Health Organ* 2000; **78**: 1078-1092 [PMID: 11019457]
 4 **Mishra V,** Dai X, Smith KR, Mika L. Maternal exposure to biomass smoke and reduced birth weight in Zimbabwe. *Ann Epidemiol* 2004; **14**: 740-747 [PMID: 15519895 DOI: 10.1016/j.annepidem.2004.01.009]
 5 **Boy E,** Bruce N, Delgado H. Birth weight and exposure to kitchen wood smoke during pregnancy in rural Guatemala. *Environ Health Perspect* 2002; **110**: 109-114 [PMID: 11781172 DOI: 10.1289/ehp.02110109]
 6 **Thompson LM,** Bruce N, Eskenazi B, Diaz A, Pope D, Smith KR. Impact of reduced maternal exposures to wood smoke from an introduced chimney stove on newborn birth weight in rural Guatemala. *Environ Health Perspect* 2011; **119**: 1489-1494 [PMID: 21652290 DOI: 10.1289/ehp.1002928]
 7 **Tielsch JM,** Katz J, Thulasiraj RD, Coles CL, Sheeladevi S, Yanik EL, Rahmathullah L. Exposure to indoor biomass fuel and tobacco smoke and risk of adverse reproductive outcomes, mortality, respiratory morbidity and growth among newborn infants in south India. *Int J Epidemiol* 2009; **38**: 1351-1363 [PMID: 19759098 DOI: 10.1093/ije/dyp286]
 8 **Siddiqui AR,** Gold EB, Yang X, Lee K, Brown KH, Bhutta ZA. Prenatal exposure to wood fuel smoke and low birth weight. *Environ Health Perspect* 2008; **116**: 543-549 [PMID: 18414641 DOI: 10.1289/ehp.10782]
 9 **Pope DP,** Mishra V, Thompson L, Siddiqui AR, Rehfuess

- EA, Weber M, Bruce NG. Risk of low birth weight and still-birth associated with indoor air pollution from solid fuel use in developing countries. *Epidemiol Rev* 2010; **32**: 70-81 [PMID: 20378629 DOI: 10.1093/epirev/mxq005]
- 10 **Jedrychowski W**, Khogali M, Elkarim MA. Height and lung function in preadolescent children of Kuwaitis and European origin: a pilot survey on health effects of gas cooking in the Middle East. *Arch Environ Health* 1991; **46**: 361-365 [PMID: 1772261]
- 11 **Kyu HH**, Georgiades K, Boyle MH. Maternal smoking, bio-fuel smoke exposure and child height-for-age in seven developing countries. *Int J Epidemiol* 2009; **38**: 1342-1350 [PMID: 19622677 DOI: 10.1093/ije/dyp253]
- 12 **Mishra V**, Retherford RD. Does biofuel smoke contribute to anaemia and stunting in early childhood? *Int J Epidemiol* 2007; **36**: 117-129 [PMID: 17085456 DOI: 10.1093/ije/dyl234]
- 13 **Ghosh R**, Amirian E, Dostal M, Sram RJ, Hertz-Picciotto I. Indoor coal use and early childhood growth. *Arch Pediatr Adolesc Med* 2011; **165**: 492-497 [PMID: 21300646]
- 14 **Roquer JM**, Figueras J, Botet F, Jiménez R. Influence on fetal growth of exposure to tobacco smoke during pregnancy. *Acta Paediatr* 1995; **84**: 118-121 [PMID: 7756793 DOI: 10.1111/j.1651-2227.1995.tb13592.x]
- 15 **Salmasi G**, Grady R, Jones J, McDonald SD. Environmental tobacco smoke exposure and perinatal outcomes: a systematic review and meta-analyses. *Acta Obstet Gynecol Scand* 2010; **89**: 423-441 [PMID: 20085532 DOI: 10.3109/00016340903505748]

P- Reviewers: Cáceres DD, Simkhovich B **S- Editor:** Wen LL
L- Editor: A **E- Editor:** Lu YJ



Echocardiography in children with Down syndrome

Mohammed A Al-Biltagi

Mohammed A Al-Biltagi, Paediatric Department, Faculty of Medicine, Tanta University, Tanta 31527, Egypt
Author contributions: Al-Biltagi M solely contributed to this paper.

Correspondence to: Mohammed A Al-Biltagi, MD, PhD, Associate Professor of Paediatrics, Paediatric Department, Faculty of Medicine, Tanta University, El Bahr Str, Tanta 31527, Egypt. mbelrem@hotmail.com

Telephone: +97-33-9545472 Fax: +20-40-2213543

Received: May 2, 2013 Revised: June 5, 2013

Accepted: June 19, 2013

Published online: November 8, 2013

Echocardiographic examination is recommended for all neonates with DS in the first month of life, before any cardiac surgery, to follow up after cardiac surgery and for serial evaluation of pulmonary hypertension. It is also indicated before involvement in non-cardiac major surgery and before involvement in physical exercise.

Al-Biltagi M. Echocardiography in children with Down syndrome. *World J Clin Pediatr* 2013; 2(4): 36-45 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/36.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.36>

Abstract

Congenital heart disease is a common problem in children with Down syndrome (DS). Echocardiography plays an important role in the detection of both structural and functional abnormalities in this group of patients. Fetal echocardiography can help in the early recognition of DS by detecting soft markers of DS, but its main role is to define the exact nature of the suspected cardiac problem in the fetus. Postnatal echocardiography is mandatory in the first month of life for all neonates with DS. It is also indicated before any cardiac surgery and for serial follow-up after cardiac surgery. In this article, we discuss the types and mechanism of cardiac abnormalities in DS children and the role of both fetal and postnatal echocardiography in the detection of these abnormalities.

© 2013 Baishideng. All rights reserved.

Key words: Fetal; Echocardiography; Congenital heart disease; Down syndrome

Core tip: Cardiac affection is a common issue in the Down syndrome (DS) population, in the form of both congenital and acquired heart disorders. Echocardiography plays an important role in the detection of such disorders. Fetal echocardiography can detect cardiac disorders as early as 10-12 wk of gestation.

INTRODUCTION

Down syndrome (DS) is an autosomal trisomy 21 and is one of the most frequently occurring chromosomal abnormalities. DS occurs once in every 600 to 800 live births and is frequently associated with congenital heart disease (CHD). The incidence of CHDs increases from 0.8% in the general population to approximately 40%-65% in patients with DS. At the same time, children with DS comprise approximately 10% of all children with CHD. Such malformations include all structural and functional cardiac defects present at birth, even if discovered later in life. These malformations can be single or multiple and usually lead to significant implications for the children and their families. These children may develop congestive heart failure, pulmonary vascular disease, pneumonia, or failure to thrive. CHDs are the most common cause of death in children with DS during the first two years of life^[1,2].

Atrioventricular septal defects (AVSD; with or without other CHD) and ventricular septal defects (VSD; with or without other CHD) have both been reported as the most common congenital heart defects and make up approximately 45% and 35% of CHD associated with DS, respectively. Additionally, isolated secundum atrial septal defect (ASD), isolated persistent patent ductus arteriosus (PDA), and an isolated Fallout of Tetralogy

(TOF) individually comprise approximately 8%, 7%, and 4% of CHD associated with DS, respectively. The remaining 1% of CHDs found in patients with DS includes arch abnormalities (aortic coarctation, right aortic arch, aberrant right subclavian artery). DS tends to be associated with the more severe forms of endocardial cushion defect, while the inlet VSD is common in trisomy 21. Several cardiac lesions seen in the non-DS population are rarely if ever found in individuals with DS, *e.g.*, heterotaxy, aortic coarctation or transposition of the great arteries. These differences between the types of CHD in DS and non-DS populations could suggest the impact of a third copy of a gene or genes on chromosome 21 on only specific developmental points^[3,4].

There is an increased prevalence of persistent pulmonary hypertension of the neonate (PPHN) in children with DS. These children have an increased risk of developing PPHN even in the absence of structural heart disease and should be followed until resolution of the pulmonary hypertension^[5]. Weijerman *et al*^[6], 2010 observed a significantly increased and elevated incidence of PPHN in neonates with DS (5.2%) compared to the general population. Another rare presentation in neonates with DS is the association of hypertrophic cardiomyopathy with complete atrioventricular canal defect in an infant with trisomy 21^[7].

While a defect of the atrioventricular canal remains the most common heart malformation in children with DS, the type of associated CHD may be affected by various factors. For example, the parents' consanguinity status could affect the pattern of CHDs. Al-Jarallah^[8], 2009 documented a slightly higher frequency of CHD in a sample of DS children from a Saudi population with a high consanguineous marriage rate. That study found that VSD was the most frequently detected CHD with the predominance of left-right shunt lesions and the relative rarity of cyanotic and complex CHD in this DS population. However, Chéhab *et al*^[9] previously showed that the risk of congenital cardiac anomalies in children with DS was not associated with the parents' consanguinity; instead, having a maternal age above 32 years was more associated with a lesser occurrence of congenital cardiac anomalies in children with DS.

Ethnicity and sex are additional factors that appear related to the type and frequency of CHD in the DS population. In a study conducted in the United States of America, Freeman *et al*^[10] showed that atrioventricular septal defects had the most significant sex and ethnic differences, with twice as many females affected and with twice as many blacks and half as many Hispanics affected compared to whites. In the Saudi population with DS, VSD was the most common (33.3%) followed by AVSD (22.8%), ASD (21.1%), patent ductus arteriosus (14%) and tetralogy of Fallot (11%). In a Turkish sample, the most common single defect was AVSD (34.2%), followed by second ASD (16.7%) and VSD (16.5%)^[11,12]. On the other hand, PDA was the most common cardiac malformation observed in

Guatemalan children with DS, followed by VSD, ASD and then AVSD^[13]. The most common cardiac malformations in Mexican children with DS were ASD, VSD and PDA, while the AVSDs were less common than the other malformations^[14].

Children with DS may also have dysfunctional autonomic cardiac regulation even in the absence of concomitant congenital heart disease, which may be manifested mainly in a reduced heart rate response to excitatory stimuli, including arousal from sleep. This is especially noted if accompanied by sleep-disordered breathing (SDB). O'Driscoll *et al*^[15] described a compromised acute cardio-respiratory response and dampened sympathetic response to SDB in children with DS and SDB compared to typically developed children with SDB. This altered response may be due to inadequate sympathetic activation or blunted vagal withdrawal and could reflect autonomic dysfunction in children with DS that may place them at increased risk for cardiovascular complications, such as pulmonary hypertension.

DS children with CHD have a greater predisposition to develop irreversible pulmonary arterial hypertension (PAH). The increased incidence of pulmonary hypertension in DS children could be a result of additional related problems, such as an upper airway obstruction, pulmonary hypoplasia, structural lung disease, thinner media of the pulmonary arterioles, abnormal pulmonary vasculature growth, alveolar hypoventilation, recurrent pulmonary infection or gastro-esophageal reflux^[16]. It has been suggested that PAH may develop earlier and may have a more violent course in patients with Down's syndrome, carrying with it a high risk of morbidity in a relatively young patient^[17,18].

MECHANISM OF CHD IN CHILDREN WITH DS

Heart development in humans is complex and starts very early, from the third to eight weeks of gestation. Development begins with a primitive tube that beats at 25 d of gestation and ends in the four-chamber heart. Many steps occur after the formation of the primitive heart tube, including looping, cell migration, cell transition, and septation events^[19]. The development of CHD is a multifactorial condition and is affected by a series of molecular signaling pathways and morphological events. In children with DS, it is postulated that variations in gene dosage of chromosome 21, environmental factors and genetic modifications not linked to chromosome 21 contribute to the development of CHD^[20].

Down's syndrome is most commonly caused by the presence of an extra copy of all or part of human chromosome 21 (Hsa21). The extra set of the approximately 200-300 genes on the chromosome leads to the many abnormalities associated with this condition. Due to triplication of Hsa21, there is a 1.5-fold increase in the expression of some, if not all, of these genes present in the

extra copy. However, all of these genes do not necessarily exhibit a straightforward 1.5-fold increase in expression, and only 30% of *Hsa21* genes are significantly over-expressed. Gene expression may be regulated by dosage compensation, which means that only a subset of these genes will exhibit the expected 50% increase in expression^[21]. Genetic imbalance caused by the presence of an extra copy of chromosome 21 will seriously disrupt one or more developmental pathways. This imbalance could also induce also an interaction between *Hsa21* genes and other disomic genes with relatively subtle or massively disruptive effects on genes located on chromosomes other than 21^[22]. These effects could be mediated through modulation of transcription factors, chromatin remodeling proteins, and related molecules or other targets. Thus, the imbalance-induced dysregulation of pathways involved in heart development may cause the cardiac defects observed in DS^[23].

The DS critical region (DSCR) is located on the long (q) arm of chromosome 21 and contains a number of genes that are thought to be responsible for some, if not all, of the features of DS. These genes may include the DS critical region 1 gene, or DSCR1, on chromosome 21q11.2; DSCR2 on chromosome 21q22.3; DSCR3 on chromosome 21q22.2; DSCR4 on chromosome 21q22.2; and DSCR5 in the chromosome region 21q22.1-q22.2. DS critical region 1, also known as Calcipressin-1, *Adapt78*, myocyte-enriched calcineurin-interacting protein 1 or regulator of calcineurin 1, is a 252 amino acid protein that belongs to the RCAN family (regulators of calcineurin) and exists as 4 alternatively spliced isoforms. DSCR1 is abundantly expressed in skeletal muscle, as well as in the brain and heart, and is thought to influence cardiac and nervous system development^[24]. DSCR1 is possibly part of a signal transduction pathway involving both the heart and brain and is implicated in cardiac valve formation and in the inhibition of cardiac hypertrophy^[25]. Overexpression of DSCR1 may be involved in the pathogenesis of DS, in particular mental retardation or cardiac defects^[26]. Barlow *et al*^[27] had previously mapped the DS-CHD region in humans to a 5.27-Mb chromosomal segment containing 82 genes, and Korbelt *et al*^[28] had narrowed this segment to a 2.82-Mb critical region likely involved in a DS-CHD endocardial cushion defect.

The presence of specific gene variants or modifiers could, in addition to trisomy 21, further increase the susceptibility to cardiac defects. One such genetic modifier is cysteine-rich with epidermal growth factor (EGF)-like domains (CRELD1), initially identified as a candidate for the AVSD2 locus. CRELD1 belongs to an epidermal growth factor-like family and encodes a cell surface protein that likely functions as a cell adhesion molecule. CRELD1 encodes a novel cell adhesion molecule that is expressed during cardiac cushion development. Missense mutations in CRELD1 have been found in DS patients with AVSD. CRELD1 (3p25.1) is one of the well-studied non-chromosome 21 loci variations that may predispose one to a heart defect. Other genetic modifiers have also

been shown to affect heart development. For example, somatic mutations in basic helix-loop-helix (bHLH) transcription factor *HEY2* (gridlock) have been identified in CHD in people with DS but not in euploid populations with heart defects^[29-32].

Environmental factors interact with the trisomic genome and may modify the occurrence of associated CHD in children with DS. Maternal cigarette smoking, for instance, has been associated with ASVD, TOF, and ASD but not with VSD^[33]. However, Khoury and Erickson observed an inverse relationship between maternal alcohol use and the presence of VSD in children with DS. Maternal folic acid supplementation may be associated with a reduced risk for CHD. Individuals with DS are at a high risk for CHD and have been shown to have abnormal folate metabolism^[34]. Bean *et al*^[35] found that a lack of maternal folic acid supplementation was more frequent among infants with DS and AVSD or ASD than among infants with DS and no heart defect or with VSD.

FETAL ECHOCARDIOGRAPHY

Fetal echocardiography is considered in cases of suspected DS. Such instances include the observation of a fetal nuchal translucency measurement of 3.5 mm or greater in the first trimester, the presence of a single umbilical artery or following an abnormal or incomplete cardiac evaluation on an anatomic scan, 4-chamber study. Fetal echocardiography can identify fetal cardiac structures as early as 10-12 wk of gestation using vaginal probes with high-resolution transducers, while conventional trans-abdominal echocardiography can detect fetal cardiac anomalies by 16-18 wk of gestation. The optimum period in which to perform a screening examination is at 20-22 wk because, at that time, the fetal cardiac structures can be defined more clearly with ultrasound screening in more than 90% of cases. Fetal echocardiographic examination can be difficult because of fetal physiology, the effect on flow across defects and valves, the inability to see the fetus for orientation reference, and an inability to examine the fetus for clinical findings. Likewise, a detailed heart evaluation can be very difficult during the third trimester because of acoustic shadowing, as in cases of maternal obesity or prone fetal position^[36].

Fetal echocardiography can assist in the early recognition of DS by detecting soft syndrome markers, but its main role is to define the exact nature of the cardiac anomaly suspected in the fetus. Such an assessment helps both the parents and the treating physician make the most informed decisions. Fetal echocardiography can provide the possibility of pregnancy termination in cases of severe malformations and of treating in utero the potentially treatable and less severe disorders, *e.g.*, fetal supraventricular tachycardia^[37]. In addition, fetal echocardiography can identify babies with complex congenital heart diseases that need delivery in a special tertiary care level center equipped with a Neonatal Intensive Care Unit so that during the transition from pre- to post-natal

life, the baby does not face periods of hypoxia or acidosis and can be given immediate care^[38].

Soft syndrome markers are ultrasound findings that are considered abnormal and whose presence increases the risk for underlying fetal aneuploidy and congenital heart diseases. These markers are nonspecific and could also be present in fetuses without abnormalities. They are often transient and can be readily detected during the second trimester^[39].

Echogenic intracardiac foci (EICF) are examples of these soft markers that have been associated with DS, as well as with trisomy 13. They are a common finding seen in approximately 4% of obstetric sonograms, and their incidence can vary with ethnicity. The lowest rates of EICF are seen in black populations, while the highest rates occur among Asian patients. These foci are of no hemodynamic or other short or long term clinical or functional significance, but their importance arises from being a possible marker of a chromosomal abnormality^[40]. These foci are discrete areas of increased echogenicity in the region of papillary muscles. They may be single or multiple and usually present in the left ventricle (88% of cases) but occasionally present in the right ventricle (5%) and may be bilateral in approximately 7% of cases. The right-sided, biventricular, multiple, or significantly obvious EICF are associated with a higher risk for fetal aneuploidy compared with the more common single, left ventricular EICF^[41,42].

These foci may result from the aggregation of chordal tissues that have failed to fenestrate completely, the enhancement of abnormal tissue, or from a collection of fibrous tissue with increased echogenicity. They may also represent true microcalcifications within the cardiac muscle^[43]. The larger size of the left ventricular papillary muscle and the larger masses of chordae tissue, the more likely is to see echogenic foci in this area^[44].

A finding of EICF is subjective. Its detection depends on a variety of factors, including the resolution of the sonographic equipment, technique, thoroughness of the examination, and the sonographer's experience^[45]. For proper visualization and grading of EICF, an appropriate transducer frequency (≤ 5 MHz) and an appropriate gain setting should be used. These foci can be diagnosed on the standard 4-chamber view of the fetal heart. Fetal position is also important because intracardiac foci are best visualized when the cardiac apex is oriented toward the transducer. The foci echogenicities are graded according to their comparison to the surrounding bones, especially the thoracic spine. In grade 1, the echogenicity is less than that of the thoracic spine and the EICF image will be lost before that of the thoracic spine. Grade 2 suggests that the echogenicity is representative of bone and that the EICF and thoracic spine images disappear at the same gain setting. In grade 3, the echogenicity of the EICF is greater than that of bone, and the thoracic spine image will be lost before that of the EICF^[46].

The foci should be differentiated from other cardiac

conditions that can be misdiagnosed with these foci. These conditions include intra-cardiac tumors (rhabdomyomas, teratomas, fibromas, hemangiomas), ventricular thrombi (especially if adherent to the papillary muscles in the left ventricle), dystrophic valves, air in the chambers from fetal demise, endocardial fibroelastosis that is usually multiple and along the endocardial surface, idiopathic infantile arterial calcification, viral infections or metabolic disorders^[47].

The presence of an aberrant right subclavian artery (ARSA) is another potential new soft marker that is more commonly seen in fetuses with trisomy 21 and other chromosomal defects than in normal fetuses. An ARSA has been found postnatally in 1%-2% of normal individuals (from neonates to adulthood) at autopsy, but its incidence is increased in cases of DS, with figures ranging between 2.9% and 37.0%^[48]. Although it can be considered a weak marker, the second trimester diagnosis of an ARSA should prompt a detailed search for additional "soft markers" and potential structural defects^[49].

Once DS is suspected, fetal echocardiography should be performed to detect associated structural cardiac abnormalities. A cardiac anomaly can be classified according to its detectability by fetal echocardiography and its severity. With regard to detectability, the structural cardiac abnormalities are classified as detectable or undetectable. Detectable cardiac anomalies are those that can be identified during routine antenatal assessment incorporating a four-chamber view of the heart at approximately eighteen week' gestation. These abnormalities usually produce marked cardiomegaly, an obvious abnormality of the atrioventricular connection, or a disparity between the sizes of the atria or ventricles or both. In undetectable abnormalities, the four-chamber view fails to identify them as types of major malformations affecting the left or right heart outflows. Ventricular septal defect, pulmonary stenosis, aortic stenosis, and coarctation of the aorta are considered among the possible undetectable abnormalities^[57].

With regard to severity, structural cardiac abnormalities are classified into "complex," "significant," and "minor" heart disease. Complex heart disease occurs when there is an atretic or hypoplastic valve or cardiac chamber, which may include a hypoplastic left heart, Truncus arteriosus, pulmonary atresia with ventricular septal defect, or a double outlet ventricle. Heart diseases are considered significant when four chambers of normal or increased size and four valves are present but require intervention, *e.g.*, Ebstein/tricuspid valve dysplasia, significant complete AVSD, large VSD, partial AVSD, ASD, PDA, severe aortic stenosis, severe pulmonary stenosis (PS), transposition of the great arteries (TGA), coarctation of the aorta, total anomalous pulmonary venous connection, or TOF. Cardiac abnormalities are considered minor when no treatment is required, such as in cases with small VSD or relatively mild aortic or pulmonary valve stenosis. If there are multiple cardiac abnormalities, the disorder is

classified according to the most severe diagnosis. There are two exceptions from this classification: the endocardial fibroelastosis, which is classified as “complex”, and complete AVSD, which is classified as “significant”^[50].

There is a further classification according to severity suggested by Choi *et al*^[51], who identified 5 classes of fetal echocardiography results: normal, minor abnormalities, simple cardiac anomalies, moderate cardiac anomalies, and complex cardiac anomalies. Minor abnormalities are those that do not require any interference, such as PFO, isolate peripheral PS, abnormally looking aortic arch, simple right aortic arch, tortuous ductus without obstruction, and mild right ventricular dilation with or without tricuspid regurgitation (at most mild regurgitation). Simple cardiac anomalies are defined as a simple defect or a defect completely correctable by medical treatment, such as VSD, ASD, and possibly CoA. Moderate cardiac anomalies include defects that are surgically correctable with a low risk for reoperation, such as TOF, CoA, AVSD, and complete TGA. Complex cardiac anomalies are defined as defects correctable with surgery but that carry a high risk for sequelae or defects potentially suitable for a Fontan procedure, such as a double outlet right ventricle, TGA with PS, critical PS, and other Fontan candidates (pulmonary atresia with intact ventricular septum, functional single ventricle, hypoplastic left heart syndrome). There is an additional classification according to surgical risk that has the disadvantages of having great variability between institutions and the need to change the risk stratification in accordance with future advancements in surgical treatment^[51,52].

For optimal views of the heart, it is best to direct the fetal cardiac apex toward the anterior maternal wall. Optimization of the sonographic images could be achieved by appropriate adjustment of technical settings, such as acoustic focus, frequency selection, signal gain, image magnification, temporal resolution, harmonic imaging, and Doppler-related parameters. Accurate interpretation of obtained echocardiographic images will depend on a firm understanding of the anatomy of the fetal heart, either to diagnose a congenital abnormality or to confirm normality. According to guidelines from the American Institute for Ultrasound in Medicine, fetal echocardiography should include imaging of the aortic arch, ductal arch, four-chamber view, inferior vena cava, left ventricular outflow tract (LVOT), right ventricular outflow tract (RVOT), short-axis views (“low” for ventricles and “high” for outflow tracts), superior vena cava, and three-vessel and trachea views^[53].

For fetuses with major CHD, a full diagnosis requires a sequential segmental approach, similar to postnatal practice^[54]. The first step taken in studying the fetal heart is to definitely recognize the “Situs.” *In situs solitus*, the inferior vena cava (IVC) is anterior and to the right of the aorta, the abdominal aorta is posterior and to the left of the spinal cord, the gastric air bubble is on the left side and the liver is on the right. *In situs inversus*, there is a mirror image pattern, with the aorta to the right of

the IVC, and *in situs ambiguous*, the aorta and IVC are located on the same side of the spine in right isomerism, while the aorta is centrally located with an interrupted IVC in left isomerism^[55]. For determination of the cardiac position and axis, the heart is normally located within the left chest, with the apex pointing to the left (levocardia). In dextrocardia, the heart is located within the right chest with the apex pointing to the right, while in mesocardia the heart is centrally located with the apex pointing anteriorly. Dextroposition should be distinguished from dextrocardia by the normal left-sided axis of the heart despite being displaced to the right due to extracardiac reasons (for example, diaphragmatic hernia, congenital pulmonary cystic adenomatoid malformation, pleural effusion, *etc.*)^[38,56].

The four-chamber view (Figure 1A and B) is a key view of the fetal heart. It is effective in prenatal cardiac screening and can detect 43%-96% of fetal anomalies. It has the advantage of using fetal ribs as external reference points to ensure that the sonographer has “cut” the thorax in the appropriate plane. In a correct four-chamber view, there should be the appearance of a single rib around the fetal thorax. The four-chamber view is situated in a more horizontal plane than in the postnatal period because of the large liver. It can show the two atria and ventricles along with atrioventricular (AV) valves (mitral and tricuspid) and septa (interventricular and interatrial)^[57]. The detectability of CHDs by fetal echocardiography increases if the outflow tracts are examined as well as the four-chamber view, but appreciation of abnormalities in the outflow tracts is more challenging than in the four-chamber view^[58]. “Extended basic views” of LVOT and RVOT increase the sensitivity of anomaly detection. Alternatively, a comprehensive set of five short-axis projections can be acquired. The LVOT view can be obtained by tilting the transducer 45° from the four-chamber view perpendicular to the septum. This view can show the aorta originating from the LVOT with its main great vessels of the head and neck appearing distally. The RVOT view can be obtained by further rotation in the same direction and by gentle rocking the transducer from the LVOT view. In this view, the pulmonary artery can be seen rising from the RV and dividing into its main branches. The left pulmonary artery and ductus arteriosus can be observed, and the ascending aorta is seen centrally, wrapped by the RV and PA. A comprehensive set of five short-axis projections can also be obtained by cranial or caudal angulation of the ultrasound probe from the four-chamber view with the ventricular septum parallel to the ultrasound beam and the fetal spine^[54]. The pulmonary artery and RVOT can be visualized in the most cephalad short-axis view as they course around the aorta. Despite being small, the fetal pulmonary branches bifurcation can still be identified. The ductus arteriosus can likewise be identified and traced into the descending aorta. With caudal and more horizontal angulation from this plane, the ventricles and their respective AV valves can be observed. These views are better for the detection of conotruncal

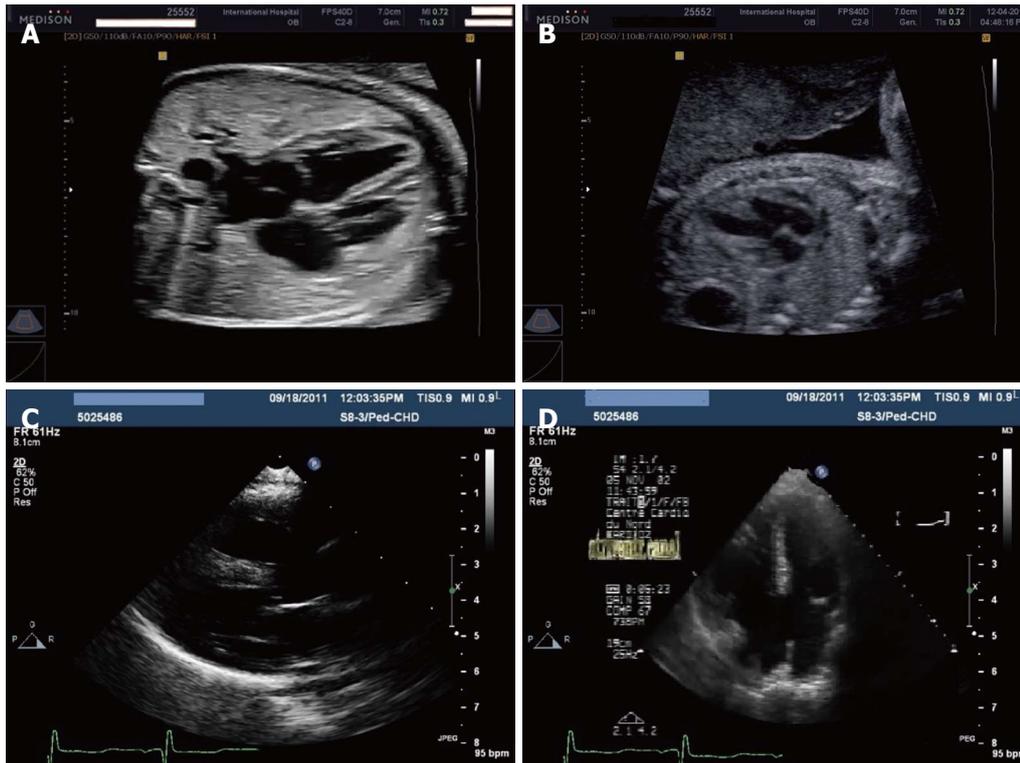


Figure 1 Ultrasonography. A: Normal 4-chamber view by fetal echocardiography at 26 wk gestation; B: Four-chamber view by fetal echocardiography at 22 wk gestation showing ventricular septal defects; C: Left-parasternal long-axis view in an infant with Down syndrome and complete atrioventricular (AV) canal and pulmonary hypertension; D: Apical 4-chamber view in an infant with Down syndrome and complete AV canal and pulmonary hypertension.

abnormalities that could otherwise be missed in more routine views. However, a specific diagnosis should not be made from a single plane.

The left and right fetal ventricles are nearly the same size, and the diameter of the pulmonary artery is typically larger than the aorta by approximately 10%. Ventricular volumes can be measured in 2-D mode using the Simpson rule. Other important measures include the RV/LV ratio, LV wall thickness, septal wall thickness, left atrial dimension, PA diameter, and aortic root diameter. These measures should be plotted against gestational age, which can be determined by measurement of the biparietal diameter or fetal length. These measures are helpful in the diagnosis of ventricular hypoplasia (either left or right) and cardiomegaly due to various congenital abnormalities, including pericardial effusion, aneurysms, cardiomyopathies, or tumors. Fetal heart motion, heart rate, wall thickness, chamber size, and motion of the valves or myocardium can be easily traced by M-mode, which can provide information on wall thickness and ventricular shortening fraction. The fetal long axis function may provide additional insight into endocardial function, which is most usefully in the detection of early ischemic changes before the development of sonographically detectable endocardial fibroelastosis. Moreover, color Doppler can be used to detect vascular flow through cardiac chambers, vascular structures, and septal defects^[59-63].

Clur *et al*^[64] showed that cardiac functions in trisomy

21 fetuses were abnormal irrespective of the presence of CHD. Evidence for cardiac loading (increased preload and afterload) and LV systolic (in the first trimester) and later diastolic dysfunction was observed. The authors showed significant reduction of tricuspid valve (TV) A-wave velocity and aortic valve peak velocity in trisomy 21 compared with normal fetuses. In addition, they also identified significant increases in the TV-E/A ratio and the ductus venosus-pulsatility index for veins and decreased pulmonary valve peak velocity. Moreover, the authors observed some evidence of left ventricular (LV) systolic dysfunction, such as a reduction of stroke volume (SV) and an increased myocardial performance index. They also found significant reduction of the mitral valve A-wave peak velocity and E/TVI ratio after 14 wk of gestation in the trisomy 21 fetuses with normal hearts compared to the controls with increased nuchal translucency thickness.

A complete AV canal can be more easily visualized in a 4-chamber view than an ASD primum, so that diagnosis of ASD primum type should be considered whenever a defect is noted in the portion of the atrial septum near the AV valves (septum primum). Opening the AV valve during diastole makes a large complete AV canal more obvious on the 4-chamber view, while during systole, the atrial and ventricular septal defects can be clearly seen above and below the closed AV valves, rendering its diagnosis possible as early as the late first trimester. Color

flow Doppler can show mixing of flow in the area of the septum primum defect, the dysplastic AV valves, and the ventricular septal defect. It can also show a lack of separation between the right and left ventricular inflow tracts in diastole, producing a classic “H” configuration. Color flow Doppler is also helpful in detecting and evaluating the degree of dysplastic AV insufficiency that may be severe enough to produce fetal heart failure with ascites^[65].

TRANSTHORACIC ECHOCARDIOGRAPHY

Although it has been recommended that infants with trisomy should have an echocardiogram in the first month of life, the value of routine neonatal echocardiography in this population is still in debate. Nevertheless, physical examination alone is not sufficient to identify cardiovascular anomalies in neonates with DS. In the newborn with DS, the potential benefits of early diagnosis, in the context of physical examination findings, should be considered in determining whether an echocardiogram should be performed during the neonatal period. Echocardiography should be obtained in all children with DS before proceeding with surgery^[66,67].

Echocardiographic examination can provide thorough real-time, non-invasive anatomic and functional information at relatively low cost. In neonates, the echocardiographic windows are more easily obtained and clearer than at any other age because of the reduced interference by lung tissue, which is impenetrable to ultrasound, and because the heart and the great vessels are nearer the probe. Echocardiographic examination should be conducted systematically with the classic standard views [left parasternal, apical, subcostal and suprasternal (Figure 1C and D)] and completed with Doppler ultrasound (color Doppler, pulsed Doppler and continuous wave Doppler). Trans-thoracic echocardiographic examination can usually detect cardiac defects in most cases. It can also determine the level of intra-cardiac shunting, along with its degree and direction, which can be confirmed by saline contrast injection. One mL of saline/blood mixture (which provides a greater intensity and more prolonged effect than the use of agitated saline only) is rapidly injected into a peripheral vein while capturing a four-chamber view of the heart. The simultaneous appearance of bright echoes from air bubbles in the fluid in the right ventricle and left atrium documents right-to-left atrial shunting as the air bubbles produced by hand agitation are too large to cross the pulmonary vascular bed, thereby predominantly aiding visualization of the right heart. The injection of the fluids into a vein that drains into the inferior vena cava could produce better results^[68,69].

Echocardiography can also evaluate cardiac functions. For example, left ventricular systolic function can be evaluated by measuring left ventricular ejection fraction, fraction shortening, SV, stroke index, cardiac output and cardiac index. Left ventricular diastolic function can be evaluated by measuring the left ventricular inflow velocity pattern or

trans-mitral flow velocity pattern [the early diastolic filling velocity (E-wave) is normally higher than the peak atrial filling velocity (A-wave)], the pulmonary venous flow velocity pattern (pulmonary venous flow velocity pattern usually consists of the antegrade flow during ventricular systole (S-waves: S and S₂), antegrade flow during early ventricular diastole (D wave), and retrograde flow), the flow propagation velocity (Vp) during the rapid filling period and the peak early diastolic velocity of the mitral annulus (Ea, E_a). The global function of LV can be measured using the Tei index. The Tei index is the first comprehensive index of cardiac functions that covers both systolic and diastolic functions, and it deteriorates and improves with either systolic or diastolic dysfunction^[70].

The quantitative assessment of RV size and function is often difficult because of the complex geometric anatomy, anatomical position under the sternum and the heavily trabeculated chamber with poor endocardial definition. However, 2-D echocardiography can easily obtain valuable information about RV size and function. Right ventricular dilatation, compared to the LV size, is a sign of RV dysfunction. Additionally, abnormal motion of the interventricular septum and the eccentricity index estimate RV pressure overload. The Tei index and the tricuspid annular plane systolic excursion both correlate well with RV function^[71].

Echocardiography can also evaluate the presence and severity of pulmonary arterial hypertension, which is relatively more common in DS neonates than in non-DS neonates. Doppler echocardiography allows estimation of both systolic and diastolic pulmonary artery pressure (PAP). The systolic PAP can be estimated by measuring the tricuspid regurgitation jet that represents the right ventricular-right atrial pressure gradient. Both PAP and RV systolic pressure are nearly equal in the absence of stenosis of the RV outflow tract, which is the reason that systolic PAP is commonly estimated by techniques that measure RV systolic pressure^[72].

Additionally, echocardiography is a useful tool for following up cases and for evaluating treatment outcome. Likewise, it has a further role even in the absence of congenital heart disease. Echocardiography can detect the presence of pericardial effusion in children with DS and hypothyroidism. A number of case studies have described the presence of pericardial effusion in this group of patients. Anah *et al*^[73], for instance, described the presence of a complex association of DS-hypothyroidism-pericardial effusion.

Echocardiography may also reveal impaired cardiac function even in the absence of congenital or structural cardiac defects. A number of studies have documented impaired cardiac functions in patients with DS. They recommended echocardiographic examination before involving patients with DS in surgery or physical exercise, even in the absence of structural cardiac diseases^[74,75].

Recommendation

Echocardiographic examination is recommended in chil-

dren with DS in the following situations: (1) in the first month of life for all neonates with DS; (2) before any cardiac surgery; (3) follow-up after cardiac surgery; (4) serial evaluation of pulmonary hypertension; (5) before involvement in major non-cardiac surgery; and (6) before involvement in physical exercise.

CONCLUSION

Echocardiography plays an important role in the detection of both structural and functional abnormalities in children with DS. Fetal echocardiography can help in the early recognition of DS by detecting soft markers of DS; however, its main role is to define the exact nature of the cardiac problem suspected in the fetus. Postnatal echocardiography is recommended in the first month of life for all neonates with DS. It is also indicated before any cardiac surgery and for serial follow-up after cardiac surgery.

REFERENCES

- Hoffman JI, Kaplan S. The incidence of congenital heart disease. *J Am Coll Cardiol* 2002; **39**: 1890-1900 [PMID: 12084585 DOI: 10.1016/S0735-1097(02)01886-7]
- Salih A. Congenital heart disease in down syndrome: experience of kurdistan of Iraq. *Duhok Med J* 2011; **5**: 24-33
- Seale A, Shinebourne EA. Cardiac problems in Down syndrome. *Curr Pediatr* 2004; **14**: 33-38 [DOI: 10.1016/j.cupe.2003.09.005]
- Freeman SB, Taft LF, Dooley KJ, Allran K, Sherman SL, Hasold TJ, Khoury MJ, Saker DM. Population-based study of congenital heart defects in Down syndrome. *Am J Med Genet* 1998; **80**: 213-217 [PMID: 9843040]
- Shah PS, Hellmann J, Adatia I. Clinical characteristics and follow up of Down syndrome infants without congenital heart disease who presented with persistent pulmonary hypertension of newborn. *J Perinat Med* 2004; **32**: 168-170 [PMID: 15085894 DOI: 10.1515/JPM.2004.030]
- Weijerman ME, van Furth AM, van der Mooren MD, van Weissenbruch MM, Rammeloo L, Broers CJ, Gemke RJ. Prevalence of congenital heart defects and persistent pulmonary hypertension of the neonate with Down syndrome. *Eur J Pediatr* 2010; **169**: 1195-1199 [PMID: 20411274 DOI: 10.1007/s00431-010-1200-0]
- Eidem BW, Jones C, Cetta F. Unusual association of hypertrophic cardiomyopathy with complete atrioventricular canal defect and Down syndrome. *Tex Heart Inst J* 2000; **27**: 289-291 [PMID: 11093415]
- Al-Jarallah AS. Down's syndrome and the pattern of congenital heart disease in a community with high parental consanguinity. *Med Sci Monit* 2009; **15**: CR409-CR412 [PMID: 19644417]
- Chéhab G, Chokor I, Fakhouri H, Hage G, Saliba Z, El-Rassi I. [Congenital heart disease, maternal age and parental consanguinity in children with Down's syndrome]. *J Med Liban* 2007; **55**: 133-137 [PMID: 17966733]
- Freeman SB, Bean LH, Allen EG, Tinker SW, Locke AE, Druschel C, Hobbs CA, Romitti PA, Royle MH, Torfs CP, Dooley KJ, Sherman SL. Ethnicity, sex, and the incidence of congenital heart defects: a report from the National Down Syndrome Project. *Genet Med* 2008; **10**: 173-180 [PMID: 18344706 DOI: 10.1097/GIM.0b013e3181634867]
- Abbag FI. Congenital heart diseases and other major anomalies in patients with Down syndrome. *Saudi Med J* 2006; **27**: 219-222 [PMID: 16501680]
- Nisli K, Oner N, Candan S, Kayserili H, Tansel T, Tireli E, Karaman B, Omeroglu RE, Dindar A, Aydogan U, Başaran S, Ertugrul T. Congenital heart disease in children with Down's syndrome: Turkish experience of 13 years. *Acta Cardiol* 2008; **63**: 585-589 [PMID: 19014001]
- Vida VL, Barnoya J, Larrazabal LA, Gaitan G, de Maria Garcia F, Castañeda AR. Congenital cardiac disease in children with Down's syndrome in Guatemala. *Cardiol Young* 2005; **15**: 286-290 [PMID: 15865831]
- de Rubens Figueroa J, del Pozzo Magaña B, Pablos Hach JL, Calderón Jiménez C, Castrejón Urbina R. [Heart malformations in children with Down syndrome]. *Rev Esp Cardiol* 2003; **56**: 894-899 [PMID: 14519277]
- O'Driscoll DM, Horne RS, Davey MJ, Hope SA, Anderson V, Trinder J, Walker AM, Nixon GM. Cardiac and sympathetic activation are reduced in children with Down syndrome and sleep disordered breathing. *Sleep* 2012; **35**: 1269-1275 [PMID: 22942505 DOI: 10.5665/sleep.2084]
- King P, Tulloh R. Management of pulmonary hypertension and Down syndrome. *Int J Clin Pract Suppl* 2011; (174): 8-13 [PMID: 22171818 DOI: 10.1111/j.1742-1241.2011.02823.x]
- Banjar HH. Down's syndrome and pulmonary arterial hypertension. *PVRI Review* 2009; **1**: 213-216 [DOI: 10.4103/0974-6013.57752]
- Fudge JC, Li S, Jagers J, O'Brien SM, Peterson ED, Jacobs JP, Welke KF, Jacobs ML, Li JS, Pasquali SK. Congenital heart surgery outcomes in Down syndrome: analysis of a national clinical database. *Pediatrics* 2010; **126**: 315-322 [PMID: 20624800 DOI: 10.1542/peds.2009-3245]
- Moorman A, Webb S, Brown NA, Lamers W, Anderson RH. Development of the heart: (1) formation of the cardiac chambers and arterial trunks. *Heart* 2003; **89**: 806-814 [PMID: 12807866 DOI: 10.1136/heart.89.7.806]
- Aanhaanen WT, Moorman AF, Christoffels VM. Origin and development of the atrioventricular myocardial lineage: insight into the development of accessory pathways. *Birth Defects Res A Clin Mol Teratol* 2011; **91**: 565-577 [PMID: 21630423 DOI: 10.1002/bdra.20826]
- Ait Yahya-Graison E, Aubert J, Dauphinot L, Rivals I, Prieur M, Golfier G, Rossier J, Personnaz L, Creau N, Bléhaut H, Robin S, Delabar JM, Potier MC. Classification of human chromosome 21 gene-expression variations in Down syndrome: impact on disease phenotypes. *Am J Hum Genet* 2007; **81**: 475-491 [PMID: 17701894 DOI: 10.1086/520000]
- Prandini P, Deutsch S, Lyle R, Gagnebin M, Delucinge Vivier C, Delorenzi M, Gehrig C, Descombes P, Sherman S, Dagna Bricarelli F, Baldo C, Novelli A, Dallapiccola B, Antonarakis SE. Natural gene-expression variation in Down syndrome modulates the outcome of gene-dosage imbalance. *Am J Hum Genet* 2007; **81**: 252-263 [PMID: 17668376 DOI: 10.1086/519248]
- Ripoll C, Rivals I, Ait Yahya-Graison E, Dauphinot L, Paly E, Mircher C, Ravel A, Grattau Y, Bléhaut H, Mégarbane A, Dembour G, de Fréminville B, Touraine R, Créau N, Potier MC, Delabar JM. Molecular signatures of cardiac defects in Down syndrome lymphoblastoid cell lines suggest altered cilium and Hedgehog pathways. *PLoS One* 2012; **7**: e41616 [PMID: 22912673 DOI: 10.1371/journal.pone.0041616]
- Casas C, Martínez S, Pritchard MA, Fuentes JJ, Nadal M, Guimerà J, Arbonés M, Flórez J, Soriano E, Estivill X, Alcántara S. Dscr1, a novel endogenous inhibitor of calcineurin signaling, is expressed in the primitive ventricle of the heart and during neurogenesis. *Mech Dev* 2001; **101**: 289-292 [PMID: 11231093]
- Lange AW, Molkenin JD, Yutzey KE. DSCR1 gene expression is dependent on NFATc1 during cardiac valve formation and colocalizes with anomalous organ development in trisomy 16 mice. *Dev Biol* 2004; **266**: 346-360 [PMID: 14738882]

- DOI: 10.1016/j.ydbio]
- 26 **Fuentes JJ**, Pritchard MA, Planas AM, Bosch A, Ferrer I, Estivill X. A new human gene from the Down syndrome critical region encodes a proline-rich protein highly expressed in fetal brain and heart. *Hum Mol Genet* 1995; **4**: 1935-1944 [PMID: 8595418]
 - 27 **Barlow GM**, Chen XN, Shi ZY, Lyons GE, Kurnit DM, Celle L, Spinner NB, Zackai E, Pettenati MJ, Van Riper AJ, Veke-mans MJ, Mjaatvedt CH, Korenberg JR. Down syndrome congenital heart disease: a narrowed region and a candidate gene. *Genet Med* 2001; **3**: 91-101 [PMID: 11280955 DOI: 10.1097/00125817-200103000-00002]
 - 28 **Korbel JO**, Tirosh-Wagner T, Urban AE, Chen XN, Kasowski M, Dai L, Grubert F, Erdman C, Gao MC, Lange K, Sobel EM, Barlow GM, Aylsworth AS, Carpenter NJ, Clark RD, Cohen MY, Doran E, Falik-Zaccai T, Lewin SO, Lott IT, McGillivray BC, Moeschler JB, Pettenati MJ, Puschel SM, Rao KW, Shaffer LG, Shohat M, Van Riper AJ, Warburton D, Weissman S, Gerstein MB, Snyder M, Korenberg JR. The genetic architecture of Down syndrome phenotypes revealed by high-resolution analysis of human segmental trisomies. *Proc Natl Acad Sci USA* 2009; **106**: 12031-12036 [PMID: 19597142 DOI: 10.1073/pnas.0813248106]
 - 29 **Maslen CL**, Babcock D, Robinson SW, Bean LJ, Dooley KJ, Willour VL, Sherman SL. CRELD1 mutations contribute to the occurrence of cardiac atrioventricular septal defects in Down syndrome. *Am J Med Genet A* 2006; **140**: 2501-2505 [PMID: 17036335 DOI: 10.1002/ajmg.a.31494]
 - 30 **Maslen CL**. Molecular genetics of atrioventricular septal defects. *Curr Opin Cardiol* 2004; **19**: 205-210 [PMID: 15096951 DOI: 10.1097/00001573-200405000-00003]
 - 31 **Rupp PA**, Fouad GT, Egelston CA, Reifsteck CA, Olson SB, Knosp WM, Glanville RW, Thornburg KL, Robinson SW, Maslen CL. Identification, genomic organization and mRNA expression of CRELD1, the founding member of a unique family of matricellular proteins. *Gene* 2002; **293**: 47-57 [PMID: 12137942 DOI: 10.1016/S0378-1119(02)00696-0]
 - 32 **Zatyka M**, Priestley M, Ladusans EJ, Fryer AE, Mason J, Latif F, Maher ER. Analysis of CRELD1 as a candidate 3p25 atrioventricular septal defect locus (AVSD2). *Clin Genet* 2005; **67**: 526-528 [PMID: 15857420 DOI: 10.1111/j.1399-0004.2005.00435.x]
 - 33 **Torfs CP**, Christianson RE. Maternal risk factors and major associated defects in infants with Down syndrome. *Epidemiology* 1999; **10**: 264-270 [PMID: 10230836]
 - 34 **Khoury MJ**, Erickson JD. Can maternal risk factors influence the presence of major birth defects in infants with Down syndrome? *Am J Med Genet* 1992; **43**: 1016-1022 [PMID: 1415327]
 - 35 **Bean LJ**, Allen EG, Tinker SW, Hollis ND, Locke AE, Druschel C, Hobbs CA, O'Leary L, Romitti PA, Royle MH, Torfs CP, Dooley KJ, Freeman SB, Sherman SL. Lack of maternal folic acid supplementation is associated with heart defects in Down syndrome: a report from the National Down Syndrome Project. *Birth Defects Res A Clin Mol Teratol* 2011; **91**: 885-893 [PMID: 21987466 DOI: 10.1002/bdra.22848]
 - 36 **Johnson B**, Simpson LL. Screening for congenital heart disease: a move toward earlier echocardiography. *Am J Perinatol* 2007; **24**: 449-456 [PMID: 17992711]
 - 37 **Abu-Harb M**, Wyllie J, Hey E, Richmond S, Wren C. Antenatal diagnosis of congenital heart disease and Down's syndrome: the potential effect on the practice of paediatric cardiology. *Br Heart J* 1995; **74**: 192-198 [PMID: 7547001 DOI: 10.1136/hrt.74.2.192]
 - 38 **Caserta L**, Ruggeri Z, D'Emidio L, Coco C, Cignini P, Girgenti A, Mangiafico L, Giorlandino C. Two-dimensional fetal echocardiography: where we are. *J Prenat Med* 2008; **2**: 31-35 [PMID: 22439025]
 - 39 **Nicolaidis KH**, Snijders RJ, Gosden CM, Berry C, Campbell S. Ultrasonographically detectable markers of fetal chromosomal abnormalities. *Lancet* 1992; **340**: 704-707 [PMID: 1355807]
 - 40 **Shipp TD**, Bromley B, Lieberman E, Benacerraf BR. The frequency of the detection of fetal echogenic intracardiac foci with respect to maternal race. *Ultrasound Obstet Gynecol* 2000; **15**: 460-462 [PMID: 11005111 DOI: 10.1046/j.1469-0705.2000.00138.x]
 - 41 **Wax JR**, Mather J, Steinfeld JD, Ingardia CJ. Fetal intracardiac echogenic foci: current understanding and clinical significance. *Obstet Gynecol Surv* 2000; **55**: 303-311 [PMID: 10804537]
 - 42 **Wax JR**, Royer D, Mather J, Chen C, Aponte-García A, Steinfeld JD, Ingardia CJ. A preliminary study of sonographic grading of fetal intracardiac echogenic foci: feasibility, reliability and association with aneuploidy. *Ultrasound Obstet Gynecol* 2000; **16**: 123-127 [PMID: 11117080 DOI: 10.1046/j.1469-0705.2000.00206.x]
 - 43 **Bronshtein M**, Jakobi P, Ofir C. Multiple fetal intracardiac echogenic foci: not always a benign sonographic finding. *Prenat Diagn* 1996; **16**: 131-135 [PMID: 8650123 DOI: 10.1002/(SICI)1097-0223(199602)16:]
 - 44 **Petrikovsky BM**, Challenger M, Wyse LJ. Natural history of echogenic foci within ventricles of the fetal heart. *Ultrasound Obstet Gynecol* 1995; **5**: 92-94 [PMID: 7719874 DOI: 10.1046/j.1469-0705.1995.05020092.x]
 - 45 **Nyberg DA**, Souter VL, El-Bastawissi A, Young S, Luthhardt F, Luthy DA. Isolated sonographic markers for detection of fetal Down syndrome in the second trimester of pregnancy. *J Ultrasound Med* 2001; **20**: 1053-1063 [PMID: 11587012]
 - 46 **Wax JR**, Cartin A, Pinette MG, Blackstone J, Michaud J, Byers S, Boutin N. Sonographic grading of fetal intracardiac echogenic foci in a population at low risk of aneuploidy. *J Clin Ultrasound* 2003; **31**: 31-38 [PMID: 12478650 DOI: 10.1002/jcu.10127]
 - 47 **Roberts DJ**, Genest D. Cardiac histologic pathology characteristic of trisomies 13 and 21. *Hum Pathol* 1992; **23**: 1130-1140 [PMID: 1398642]
 - 48 **Willruth AM**, Dwinger N, Ritgen J, Stressig R, Geipel A, Gembruch U, Berg C. Fetal aberrant right subclavian artery (ARSA) - a potential new soft marker in the genetic scan? *Ultraschall Med* 2012; **33**: E114-E118 [PMID: 21614745 DOI: 10.1055/s-0029-1245935]
 - 49 **Paladini D**, Sglavo G, Pastore G, Masucci A, D'Armiento MR, Nappi C. Aberrant right subclavian artery: incidence and correlation with other markers of Down syndrome in second-trimester fetuses. *Ultrasound Obstet Gynecol* 2012; **39**: 191-195 [PMID: 21793087 DOI: 10.1002/uog.10053]
 - 50 **Abu-Harb M**, Hey E, Wren C. Death in infancy from unrecognised congenital heart disease. *Arch Dis Child* 1994; **71**: 3-7 [PMID: 8067789 DOI: 10.1136/adc.71.1.3]
 - 51 **Choi EY**, Lee CH, Yoon MJ, Han ES, Hong JS, Jung YS, Choi JY. Impact of fetal diagnosis of congenital heart disease on parents. *Korean J Pediatr* 2006; **49**: 1073-1078 [DOI: 10.3345/kjp.2006.49.10.1073]
 - 52 **Cha S**, Kim GB, Kwon BS, Bae EJ, Noh CI, Lim HG, Kim WH, Lee JR, Kim YJ, Choi JY. Recent trends in indications of fetal echocardiography and postnatal outcomes in fetuses diagnosed as congenital heart disease. *Korean Circ J* 2012; **42**: 839-844 [PMID: 23323122 DOI: 10.4070/kcj.2012.42.12.839]
 - 53 American Institute for Ultrasound in Medicine (AIUM). AIUM practice guideline for the performance of fetal echocardiography. Laurel, MD: AIUM; 2010. Cited 2010-10-05. Available from: URL: <http://www.smfm.org/attachedfiles/fetalEchoaiumsmfm.pdf>
 - 54 **Simpson JM**. Impact of fetal echocardiography. *Ann Pediatr Cardiol* 2009; **2**: 41-50 [PMID: 20300268 DOI: 10.4103/0974-2069.52806]
 - 55 **Carvalho JS**, Ho SY, Shinebourne EA. Sequential segmental analysis in complex fetal cardiac abnormalities: a logical

- approach to diagnosis. *Ultrasound Obstet Gynecol* 2005; **26**: 105-111 [PMID: 16041685 DOI: 10.1002/uog.1970]
- 56 **Gagnon A**, Wilson RD, Allen VM, Audibert F, Blight C, Brock JA, Désilets VA, Johnson JA, Langlois S, Murphy-Kaulbeck L, Wyatt P. Evaluation of prenatally diagnosed structural congenital anomalies. *J Obstet Gynaecol Can* 2009; **31**: 875-881, 882-889 [PMID: 19941713]
- 57 **Silverman NH**, Golbus MS. Echocardiographic techniques for assessing normal and abnormal fetal cardiac anatomy. *J Am Coll Cardiol* 1985; **5**: 20S-29S [PMID: 3880776 DOI: 10.1016/S0735-1097(85)80140-6]
- 58 **Lee W**, Allan L, Carvalho JS, Chaoui R, Copel J, Devore G, Hecher K, Munoz H, Nelson T, Paladini D, Yagel S. ISUOG consensus statement: what constitutes a fetal echocardiogram? *Ultrasound Obstet Gynecol* 2008; **32**: 239-242 [PMID: 18663769 DOI: 10.1002/uog.6115]
- 59 **Allan L**. Technique of fetal echocardiography. *Pediatr Cardiol* 2004; **25**: 223-233 [PMID: 15360115 DOI: 10.1007/s00246-003-0588-y]
- 60 **Naderi S**, McGahan JP. A primer for fetal cardiac imaging: a stepwise approach for 2-dimensional imaging. *Ultrasound Q* 2008; **24**: 195-206 [PMID: 18776813 DOI: 10.1097/RUQ.0b013e3181862b84]
- 61 **Allan LD**, Joseph MC, Boyd EG, Campbell S, Tynan M. M-mode echocardiography in the developing human fetus. *Br Heart J* 1982; **47**: 573-583 [PMID: 7082505 DOI: 10.1136/hrt.47.6.573]
- 62 **Stamm ER**, Drose JA. The fetal heart. In: Rumack CA, Wilson SR, Charboneau WJ, editors. *Diagnostic ultrasound*, 2nd ed. St. Louis, MO: Mosby, 1998: 1123-1159
- 63 **Gardiner HM**. Fetal echocardiography: 20 years of progress. *Heart* 2001; **86** Suppl 2: II12-II22 [PMID: 11709530 DOI: 10.1136/heart.86]
- 64 **Clur SA**, Oude Rengerink K, Ottenkamp J, Bilardo CM. Cardiac function in trisomy 21 fetuses. *Ultrasound Obstet Gynecol* 2011; **37**: 163-171 [PMID: 20814928 DOI: 10.1002/uog.8819]
- 65 **Craig B**. Atrioventricular septal defect: from fetus to adult. *Heart* 2006; **92**: 1879-1885 [PMID: 17105897 DOI: 10.1136/hrt.2006.093344]
- 66 **Ghaffar S**, Lemler MS, Fixler DE, Ramaciotti C. Trisomy 21 and congenital heart disease: effect of timing of initial echocardiogram. *Clin Pediatr (Phila)* 2005; **44**: 39-42 [PMID: 15678229 DOI: 10.1177/000992280504400104]
- 67 **McElhinney DB**, Straka M, Goldmuntz E, Zackai EH. Correlation between abnormal cardiac physical examination and echocardiographic findings in neonates with Down syndrome. *Am J Med Genet* 2002; **113**: 238-241 [PMID: 12439890 DOI: 10.1002/ajmg.10803]
- 68 **Bull MJ**. Health supervision for children with Down syndrome. *Pediatrics* 2011; **128**: 393-406 [PMID: 21788214 DOI: 10.1542/peds.2011-1605]
- 69 **Fineman JR**, Heymann MA, Morin FC. Fetal and postnatal circulations: pulmonary and persistent pulmonary hypertension of the newborn. In: Allen HD, Gutgesell HP, Clarck EB, Driscoll DJ, editors. *Moss and Adams' heart disease in infants, children and adolescents, including the fetus and young adult*. Philadelphia, USA: Lippincott Williams & Wilkins, 2001: 41-63
- 70 The Terminology and Diagnostic Criteria Committee of The Japan Society of Ultrasonics in Medicine. Standard measurement of cardiac function indexes. *J Med Ultrasonics* 2006; **33**: 123-127 [DOI: 10.1007/s10396-006-0100-4]
- 71 **Cho YK**, Ma JS. Right ventricular failure in congenital heart disease. *Korean J Pediatr* 2013; **56**: 101-106 [PMID: 23559970 DOI: 10.3345/kjp.2013.56.3.101]
- 72 **Suesawolal M**, Cleary JP, Chang AC. Advances in diagnosis and treatment of pulmonary arterial hypertension in neonates and children with congenital heart disease. *World J Pediatr* 2010; **6**: 13-31 [PMID: 20143207 DOI: 10.1007/s12519-010-0002-9]
- 73 **Anah MU**, Ansa VO, Etiuma AU, Udoh EE, Inej EO, Asindi AA. Recurrent pericardial effusion associated with hypothyroidism in Down Syndrome: a case report. *West Afr J Med* 2011; **30**: 210-213 [PMID: 22120489]
- 74 **Russo MG**, Pacileo G, Marino B, Pisacane C, Calabrò P, Ammirati A, Calabrò R. Echocardiographic evaluation of left ventricular systolic function in the Down syndrome. *Am J Cardiol* 1998; **81**: 1215-1217 [PMID: 9604950]
- 75 **Al-Biltagi M**, Serag AR, Hefidah MM, Mabrouk MM. Evaluation of cardiac functions with Doppler echocardiography in children with Down syndrome and anatomically normal heart. *Cardiol Young* 2013; **23**: 174-180 [PMID: 22717046 DOI: 10.1017/S1047951112000613]

P- Reviewers: Breijo-Marquez FR, Pavlovic M
S- Editor: Gou SX **L- Editor:** A **E- Editor:** Lu YJ



Bacterial colonization and intestinal mucosal barrier development

Xiao-Zhong Huang, Li-Bin Zhu, Zhong-Rong Li, Jing Lin

Xiao-Zhong Huang, Li-Bin Zhu, Zhong-Rong Li, Jing Lin, Yuying Children's Hospital of Wenzhou Medical University, Wenzhou 325027, Zhejiang Province, China

Jing Lin, Division of Newborn Medicine, Department of Pediatrics, Kravis Children's Hospital of the Icahn School of Medicine at Mount Sinai, New York, NY 10029-6574, United States

Author contributions: Huang XZ wrote the first draft; Zhu LB and Li ZR contributed some sections of the first draft; Lin J initiated the project and finalized the manuscript.

Supported by In part by Zhejiang Provincial Natural Science Foundation, No. LY12H04005 and LY13H040011

Correspondence to: Jing Lin, MD, Division of Newborn Medicine, Department of Pediatrics, Kravis Children's Hospital of the Icahn School of Medicine at Mount Sinai, Box 1508, One Gustave L. Levy Place, New York, NY 10029-6574, United States. jing.lin@mssm.edu

Telephone: +1-212-2416186 Fax: +1-212-5345207

Received: July 3, 2013 Revised: August 7, 2013

Accepted: September 18, 2013

Published online: November 8, 2013

Abstract

The intestinal tract is colonized soon after birth with a variety of ingested environmental and maternal microflora. This process is influenced by many factors including mode of delivery, diet, environment, and the use of antibiotics. Normal intestinal microflora provides protection against infection, ensures tolerance to foods, and contributes to nutrient digestion and energy harvest. In addition, enteral feeding and colonization with the normal commensal flora are necessary for the maintenance of intestinal barrier function and play a vital role in the regulation of intestinal barrier function. Intestinal commensal microorganisms also provide signals that foster normal immune system development and influence the ensuing immune responses. There is increasingly recognition that alterations of the microbial gut flora and associated changes in intestinal barrier function may be related to certain diseases of the gastrointestinal tract. This review summarizes recent advances in un-

derstanding the complex ecosystem of intestinal microbiota and its role in regulating intestinal barrier function and a few common pediatric diseases. Disruption in the establishment of a stable normal gut microflora may contribute to the pathogenesis of diseases including inflammatory bowel disease, nosocomial infection, and neonatal necrotizing enterocolitis.

© 2013 Baishideng. All rights reserved.

Key words: Bacterial colonization; Intestinal barrier; Intestinal microflora; Microbiota; Neonatal necrotizing enterocolitis; Nosocomial infection; Premature infants; Short chain fatty acids

Core tip: This review summarizes recent advances in understanding the complex ecosystem of intestinal microbiota and its role in regulating intestinal barrier function and a few common pediatric diseases. There is increasingly recognition that the stimulation of initial intestinal microbial colonization is important for proper maturation of the innate immune system and continued regulation and maintenance of intestinal barrier function. Disruption of the establishment of a stable normal gut microflora may contribute to the pathogenesis of diseases including inflammatory bowel disease, nosocomial infection, and neonatal necrotizing enterocolitis in premature infants.

Huang XZ, Zhu LB, Li ZR, Lin J. Bacterial colonization and intestinal mucosal barrier development. *World J Clin Pediatr* 2013; 2(4): 46-53 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/46.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.46>

INTRODUCTION

The human gut is home to a large collection of micro-

organisms, the composition of which varies along the intestinal tract. The important role of normal microbial intestinal colonization in human health has gained increased recognition over the past several decades. The gut microflora, which is composed of approximately 10^{14} bacteria, or approximately 10 times the number of body cells, is now considered as a functional human organ^[1]. New studies such as the Human Microbiome Project are revealing how the gut microflora manipulates and complements physiology in ways that are important for the host^[2-4]. The normal gut microflora provides protection against infection, educates the immune system, ensures tolerance to foods, and contributes to nutrient digestion and energy harvest. In addition to these important functions, normal microbial colonization of the intestine is important in the induction of the host innate response and plays a vital role in the regulation and maintenance of intestinal barrier function. Disruption in the establishment of a stable normal gut microflora may be associated with or even contribute to the pathogenesis of diseases including inflammatory bowel disease (IBD), nosocomial infection, and neonatal necrotizing enterocolitis (NEC)^[5]. This review summarizes recent advances in understanding the complex ecosystem of the gut microflora and the roles of gut microflora in regulating intestinal barrier function as well as a few common pediatric diseases which may be related to an altered gut microbiota.

ESTABLISHMENT OF NORMAL INTESTINAL MICROBIAL COLONIZATION IS ESSENTIAL FOR THE POSTNATAL INTESTINAL BARRIER MATURATION

Before birth, the intestine is sterile. The intestinal tract becomes colonized soon after birth with a variety of ingested environmental and maternal microorganisms. This process is influenced by many factors including mode of delivery, diet, environment, and the use of antibiotics^[6,7]. For example, a breast-fed full-term infant normally has an intestinal microbiota in which bifidobacteria predominate over potentially harmful bacteria, whereas in formula-fed infants, enterococci, bacteroides and clostridia predominate^[8]. In premature infants, the immature intestinal mucosa is even more sensitive to gut colonizing bacteria. Host defenses can be improved by feeding the breast milk which helps the immature intestinal mucosal immune system to develop and respond appropriately to the highly variable bacterial colonization^[8].

The dense communities of bacteria in the intestine are separated from body tissues by a monolayer of intestinal epithelial cells. Therefore, normal intestinal function depends on the establishment and maintenance of the mucosal epithelial barrier which prevents the invasion of host tissues by resident bacteria. The assembly of the multiple components of the intestinal barrier is initiated during fetal development and continues during early postnatal life; thus the intestinal barrier has not completely developed

soon after birth, particularly in preterm infants^[9]. Several studies indicate that the normal bacterial colonization process may be important for postnatal intestinal barrier development. By using a newborn piglet model, Kansagra *et al.*^[10] demonstrated that intestinal barrier function was significantly less developed in full term newborn piglets receiving total parental nutrition compared to those receiving enteral nutrition. Even in the mature intestine, lack of enteral nutrition is associated with loss of intestinal epithelial barrier function which can lead to bacterial translocation and subsequent sepsis^[11]. In a rodent model, replacing enteral nutrition with parenteral nutrition can lead to bacterial translocation from the gut^[11,12]. Total parenteral nutrition significantly increases intestinal permeability, which can be ameliorated by enteral feeding and especially with a fiber enhanced diet^[13]. All of these results suggest that enteral feeding and colonization with the normal commensal flora are necessary for the maintenance of intestinal barrier function.

Recent studies have demonstrated that certain commensal bacteria increase intestinal epithelial cell survival by inhibiting the activation of the epithelial cell proapoptotic pathway associated with pathogenic bacteria^[14]. The intestinal commensal flora is also involved in maintenance of barrier function by inducing increased epithelial cell proliferation and enhancing intestinal epithelial integrity, through translocation of the tight junction proteins and up-regulation of genes involved in desmosome maintenance^[15,16]. Fermentation products of commensal bacterial have been shown to enhance the intestinal barrier function by facilitating the assembly of tight junctions through the activation of AMP-activated protein kinase^[17]. On the other hand, the deletion of all detectable commensal flora in the gut by a four-week course of orally administration of vancomycin, neomycin, metronidazole, and ampicillin leads to a more severe intestinal mucosal injury in a dextran sulfate sodium induced mouse colitis model^[18]. The importance of normal bacterial colonization in the development and maintenance of the intestinal barrier is further supported by the observations that the gastrointestinal tract gene expression profile and intestinal barrier development are altered by early treatment with broad-spectrum antibiotics or withholding enteral feeding^[19].

INTESTINAL MICROFLORA STIMULATES THE MATURATION OF THE MUCOSAL INNATE IMMUNE SYSTEM

The colonization with normal gut microflora protects against infection from pathogenic bacteria. This long-known but poorly understood protection provided by commensal flora against pathogens is commonly referred to as colonization resistance^[20-24]. Several mechanisms have been proposed to explain colonization resistance including a direct competition for nutrients, prevention of access to adherence sites, limitation of pathogen proliferation

through production of inhibitory substances or conditions, and stimulation of host natural immune defenses^[21,25].

Before birth, the fetus is protected from microbial exposure, and postnatal stimulation by initial intestinal microbial colonization is important for the proper maturation of the innate immune system. Exposure to immunostimulatory microbial constituents may trigger activation of the infant's mucosal innate immune system as shown recently in the gnotobiotic (germ-free) mouse model^[26-28]. For example, when compared to mice with normal intestinal microbial colonization, gnotobiotic mice need 30% more calories to maintain their body weight, exhibit an enhanced susceptibility to infection with enteropathogenic bacteria, and have an immature immune system^[28].

Remarkably, intestinal mucosal homeostasis is maintained despite the large surface area continuously being exposed to different bacterial species^[29,30]. To face the menace represented by intimate contact with a huge concentration of bacteria, the intestinal epithelium has evolved into a highly regulated barrier that can recruit immune cells of hematopoietic origin and produce mucus and a diverse arsenal of antimicrobial proteins that directly kill or inhibit the growth of microorganisms^[30,31]. The intestinal epithelium comprises several cell lineages. Enterocytes constitute the most abundant epithelial cell type, and secrete several antimicrobial proteins. Paneth cells are unique to the small intestine and secrete abundant quantities of antimicrobial proteins, such as α -defensins. Finally, goblet cells secrete mucin glycoproteins and trefoil factors that assemble to form a thick mucus layer overlying the epithelium^[32].

The development of the gut immune system is initiated before birth by a genetic program that drives the formation of Peyer's patches and mesenteric lymph nodes, but its postnatal maturation depends on the establishment of a balanced indigenous microbiota^[28]. Intestinal commensal microorganisms provide signals that foster normal immune system development and influence the ensuing immune responses^[33]. Signals delivered by these commensal microorganisms drive the development of isolated lymphoid follicles, stimulate maturation of Peyer's patches and initiate the migration of IgA producing plasma cells and mature T lymphocytes into the mucosa^[33-35]. Among the first colonizing organisms evident in the intestine of newborn infants are strains of *Escherichia coli* (*E. coli*) derived from the maternal gastrointestinal tract^[36-38]. It has been demonstrated in animal models that commensal *E. coli* strains can inhibit invasive *E. coli* O157:H7 growth in the intestine^[39]. These studies prove that the gut commensal microflora clearly have important effects on the development of normal immunity.

INTERACTION BETWEEN INTESTINAL MICROFLORA AND IMMUNE HOMEOSTASIS

The intestinal microflora regulates immune homeostasis

through many different mechanisms. Gut epithelia actively sense enteric bacteria and play an essential role in maintaining host-microbial homeostasis at the mucosal interface. Many innate immune responses are regulated by Toll-like receptors (TLRs), a conserved family of innate immune receptors that recognize microbial-derived molecules, including lipopolysaccharide, lipoprotein, RNA, and methylated DNA. Experiments in mice demonstrate that the beneficial effects of commensal bacteria are mediated *via* TLRs^[40]. Intestinal epithelial cells and mucosal immune cells express pattern-recognition-receptors such as TLRs, enabling them to respond to distinctive microbial-associated molecular patterns^[41]. TLRs are therefore critical for the specific detection of microbe-associated patterns, allowing differentially regulated responses to commensal versus pathogenic flora. The recognition of commensal bacterial-derived molecules by TLRs represents a critical component of the symbiosis between the host and indigenous microflora and is important for protection against gut injury and associated mortality^[42]. Deregulated interactions between commensal bacteria and TLRs have been reported to promote chronic inflammation and tissue damage, such as that seen in IBD^[43]. TLRs may also direct expression of the MyD88-dependent antimicrobial response. Paneth cell-intrinsic MyD88 activation limits translocation and dissemination of microbes across the mucosal barrier, while having little impact on luminal microbial numbers^[44]. These results highlight the essential role of TLR-dependent pathways in compartmentalization of enteric commensal bacteria^[45].

TLR-dependent signals mediate important regenerative signals to maintain intestinal mucosal integrity^[46]. TLR-mediated protection may work through both constitutive and damage-induced production of protective factors by TLRs expressed on colonic epithelium^[40]. Stimulation of TLRs results in activation of multiple signaling cascades that control expression of a wide range of innate immune response genes^[47,48]. Recent evidence indicates a role for CpG DNA, the bacterial agonist of TLR9, in mediating some of the beneficial effects of probiotics in the intestine, and more generally, to modulate the immuno-physiological status of the gut^[49]. Furthermore, TLR2 has been demonstrated to be responsible for the effects of *Bacteroides fragilis*. This bacterium possesses an unusual capsular polysaccharide A that exerts potent immunoregulatory effects and can dampen intestinal inflammation in several models of colitis^[50,51]. Systemic administration of flagellin, a bacterial protein that stimulates TLR5 protects mice from infection with vancomycin-resistant enterococcus (VRE)^[52].

Another immune regulatory effect of commensal bacteria involves the inhibition of the nuclear factor- κ B (NF- κ B) pathway through the stabilization of I γ B α ^[53]. I γ B α is a central inhibitor of the NF- κ B pathway, which acts by retaining the inactive NF- κ B dimers in the cytosol. Most pro-inflammatory signals trigger the NF- κ B pathway by inducing the phosphorylation of I γ B α , which targets the molecule for degradation by the

ubiquitin/proteasome system^[54]. Incubation of epithelial cells with nonpathogenic *Salmonella* has been shown to induce the accumulation of I γ B α through the down-regulation of the protein's ubiquitination^[53]. Similarly, it has been recently reported that the probiotic bacteria *Lactobacillus casei* also inhibits the NF- κ B pathway by targeting the degradation of I γ B α ^[55].

The specialized intestinal epithelial cells are capable of secreting proteins into the lumen of the intestinal tract which enhance epithelial barrier function and/or interact with the bacterial flora resident in the intestine. Goblet cells are highly polarized exocrine epithelia that secrete proteins apically into the lumen of the small and large intestine through the release of secretory granules. One particular class of glycoproteins produced by goblet cells, known as mucins, forms a viscoelastic protective gel that covers the intestinal epithelium^[56]. Another class of secretory peptides, now designated as trefoil factors, is also normally produced by goblet cells and is important for the maintenance and repair of the gut mucosal barrier^[57]. Mucins interact with trefoil factors and perform a defensive role during establishment of the intestinal barrier. Mucin oligosaccharides influence the bacterial milieu of the intestinal tract by enhancing the ability of certain bacteria to colonize the intestinal tract while inhibiting the adherence of others^[58]. Mucins are also a direct source of carbohydrates and peptides that can promote the growth of bacteria^[59]. To further enhance the symbiotic relationship between gut bacteria on the host, bacteria can alter mucus synthesis, secretion, and chemical composition^[60]. Changes in mucin profile in response to bacterial colonization suggest a potential role as a protective mechanism against pathogenic invasion of the intestinal mucosa during early development^[61].

The mucosal barrier is also reinforced by secretory immunoglobulin A (sIgA) and sIgM generated through external translocation of locally produced dimeric IgA and pentameric IgM. The dimeric IgA and pentameric IgM, containing a small polypeptide called joining chain to form sIgA and sIgM, can be actively exported by secretory epithelia. This external transport is mediated by the polymeric Ig receptor (pIgR), also known as membrane secretory component (SC)^[62]. Notably, pIgR/SC knock-out mice that lack secretory IgA and IgM antibodies exhibit reduced epithelial barrier function with aberrant mucosal leakiness^[63]. sIgA and sIgM form the first line of defense against pathogens as well as other potentially dangerous agents. Therefore, secretory immunity is of great importance for the intestinal epithelial barrier. In newborn infants, only a few IgA-secreting cells circulate in the blood. However this number is remarkably increased after 1 mo of age mainly due to the progressive microbial stimulation of gut-associated lymphoid tissues^[64]. A much faster establishment of secretory immunity is often seen in infants from developing countries where there is exposure to a heavy microbial load, and an associated lower incidence of atopy^[65]. Altogether, the secretory immune system is critical for the mucosal barrier

function and the intestinal epithelial barrier maturation depends on exposure to microbial factors from the environment and the indigenous microbiota.

DISEASES ASSOCIATED WITH AN ALTERED INTESTINAL MICROFLORA AND ABNORMAL BARRIER FUNCTION

Alterations of the microbial gut flora and changes in intestinal barrier function are associated with certain diseases of the gastrointestinal tract. There is growing evidence that changes of the intestinal flora composition may play a pathogenic role in IBD, nosocomial infection, and NEC^[66-68]. It has been proposed that a genetic predisposition causes IBD patients to have a deregulated immune response against harmless antigens derived from intestinal commensal bacteria and changes of the intestinal flora composition have been described in patients with IBD^[66]. Several studies found an enhanced number of Proteobacteria and Actinobacteria but decreased numbers of Firmicutes (particularly the species *Faecalibacterium prausnitzii*) in stool samples of IBD patients as compared to healthy controls^[69,70]. In biopsies of pediatric IBD patients, higher numbers of mucosa-associated aerobic and facultative-anaerobic bacteria were found, whereas bacterial species of the normal anaerobic intestinal flora such as *Bifidobacteria* were reduced^[71,72].

It is well recognized that nosocomial infection is frequently a consequence of gut derived organisms. The infections with highly antibiotic-resistant bacteria are usually acquired during hospitalizations. Destruction of the normal flora by antibiotic administration disinhibits antibiotic-resistant members of these bacterial families, leading to their expansion to very high densities^[73]. Reintroduction of a diverse intestinal microbiota to densely VRE-colonized mice eliminates VRE from the intestinal tract^[74]. Characterization of the fecal microbiota of patients undergoing allogeneic hematopoietic stem cell transplantation demonstrated that intestinal colonization with *Barnesiella* confers resistance to intestinal domination and bloodstream infections with VRE^[74]. Furthermore, there is an increased incidence of septic complications in patients receiving parenteral as opposed to enteral nutrition and this, in some cases, is due to alterations in intestinal barrier function predisposing to bacterial translocation^[75].

In premature infants, colonization with abnormal gut flora increases the risk of hospital acquired infection or late-onset sepsis (LOS)^[76]. Prolonged use of broad spectrum antibiotics, reduced bowel motility, immature epithelial host defenses, lack of enteral feeding, and parenteral nutrition are common risk factors for an altered microbial gut flora and abnormal mucosal barrier function. The possible subsequent bacterial translocation from the gastrointestinal tract may be an important pathway initiating LOS in premature infants^[76]. Mai *et al.*^[77] analyzed stool samples that had been prospectively

collected from ten preterm infants with LOS and from 18 matched controls. A 16S rRNA based approach was utilized to compare microbiota diversity and identify specific bacterial signatures that differed in their prevalence between cases and controls. They found that the types and distributions of bacteria that initially colonize the intestine in premature infants differ in those with LOS compared to uninfected control babies. Therefore, it was proposed that a distortion in normal microbiota composition, and not an enrichment of potential pathogens, is associated with LOS in preterm infants. This may suggest that administration of probiotics may protect high-risk neonates and infants from developing sepsis. However, currently there is no clinical evidence regarding the usefulness of probiotics or prebiotics for the prevention of nosocomial sepsis in preterm infants^[78].

Failure of the postnatal developmental of the intestinal barrier in the immature intestine plays an important role in the pathogenesis of NEC, a devastating disease seen mainly in preterm infants^[68,79,80]. A major component of the pathophysiology of NEC is the nature of the interaction of bacteria with the premature gut. The pattern of bacterial colonization in the intestine of the premature neonate is quite different from that of the healthy full-term infant. Infants requiring intensive care acquire intestinal organisms slowly, and the establishment of bifidobacteria flora is retarded. A delayed bacterial colonization of the gut with a limited number of bacterial species tends to be virulent. Indeed, several clinical observational studies have shown that the duration of antibiotic exposure including prenatal exposure to antibiotics is associated with an increased risk of NEC in preterm infants^[81-83]. Therefore, an aberrant colonization of the bowel of the premature infant has been proposed to contribute to the development of NEC^[84]. By using non-culture-based microbial analyses of feces, Wang *et al.*^[85] studied fecal samples of ten preterm infants with NEC and ten matched controls and found that patients with NEC had less bacterial diversity and an increased abundance of γ -proteobacteria in the stools. Similar findings were presented in another study by Mai *et al.*^[86], in which one of the bacterial signatures detected more frequently in NEC patients matched closest to γ -proteobacteria. These observations suggest that abnormal patterns of microbiota contribute to the cause of NEC. However, a study by Mshildadze *et al.*^[87] using the same technology demonstrated that the overall microbial profiles in patients with NEC were not different from those of control infants. Thus to date, molecular methods have not clarified the bacterial pathogenesis of NEC.

None of the clinical studies to date has been able to fulfill Koch's postulate linking NEC to a particular pathogen. Nevertheless, we proposed a hypothesis that excessive production/accumulation of short chain fatty acids (SCFAs) due to bacterial fermentation of undigested formula or abnormal bacterial colonization contributes to the pathogenesis of NEC^[88,89]. There is substantial indirect clinical evidence to support the theory that bacte-

rial fermentation is involved in the development of NEC in premature infants^[90]. Further, in two separate studies, increased breath hydrogen excretion (an indicator of bacterial fermentation and an indirect measurement of SCFAs production) was found in NEC patients even prior to the onset of clinical symptoms^[91,92]. The well-known finding of pneumatosis intestinalis (gas in the bowel wall) in NEC patients is also thought to be secondary to hydrogen gas produced by bacterial fermentation^[93]. Recent reports of several cases of premature infants who developed NEC after they were fed SimplyThick[®], a xanthan gum-based thickener used in the management of dysphagia, is another example^[94,95]. Increased production of hydrogen and SCFAs as the consequence of accumulation of luminal carbohydrates and fecal bacteria fermentation of xanthan gum were proposed as the main mechanisms for NEC^[94]. Both probiotics and prebiotics have been proposed to promote a healthy gut microbiota in human, and oral probiotics have been proven to be effective in reducing the incidence of NEC in premature infants in several clinical trials^[96-98]. On the other hand, there is no evidence showing that prebiotics can effectively reduce the incidence of infection or NEC in premature infants. Therefore, there is insufficient evidence to recommend the use of oligosaccharides as prebiotics in formula for premature infants since these may prove to be unsafe.

In summary, this review summarizes recent advances in understanding the complex ecosystem of intestinal microbiota and its role in regulating intestinal barrier function and a few common pediatric diseases. There is increasingly recognition that the stimulation of initial intestinal microbial colonization is important for proper maturation of the innate immune system and continued regulation and maintenance of intestinal barrier function. Disruption of the establishment of a stable normal gut microflora may contribute to the pathogenesis of diseases including IBD, nosocomial infection, and NEC in premature infants.

ACKNOWLEDGEMENTS

The authors wish to thank Drs. Ian Holzman and Robert Green for their critical review and comments.

REFERENCES

- 1 **Guarino A**, Wudy A, Basile F, Ruberto E, Buccigrossi V. Composition and roles of intestinal microbiota in children. *J Matern Fetal Neonatal Med* 2012; **25** Suppl 1: 63-66 [PMID: 22348506 DOI: 10.3109/14767058.2012.663231]
- 2 Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. *Nature* 2012; **486**: 207-214 [PMID: 22699609 DOI: 10.1038/nature11234]
- 3 **Dave M**, Higgins PD, Middha S, Rioux KP. The human gut microbiome: current knowledge, challenges, and future directions. *Transl Res* 2012; **160**: 246-257 [PMID: 22683238 DOI: 10.1016/j.trsl.2012.05.003]
- 4 **Buccigrossi V**, Nicastro E, Guarino A. Functions of intestinal microflora in children. *Curr Opin Gastroenterol* 2013; **29**: 31-38 [PMID: 23196853 DOI: 10.1097/MOG.0b013e32835a3500]
- 5 **Murgas Torrazza R**, Neu J. The developing intestinal micro-

- biome and its relationship to health and disease in the neonate. *J Perinatol* 2011; **31** Suppl 1: S29-S34 [PMID: 21448201 DOI: 10.1038/jp.2010.172]
- 6 **Dominguez-Bello MG**, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, Knight R. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* 2010; **107**: 11971-11975 [PMID: 20566857 DOI: 10.1073/pnas.1002601107]
 - 7 **Koenig JE**, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, Angenent LT, Ley RE. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci USA* 2011; **108** Suppl 1: 4578-4585 [PMID: 20668239 DOI: 10.1073/pnas.1000081107]
 - 8 **Cilieborg MS**, Boye M, Sangild PT. Bacterial colonization and gut development in preterm neonates. *Early Hum Dev* 2012; **88** Suppl 1: S41-S49 [PMID: 22284985 DOI: 10.1016/j.earlhumdev.2011.12.027]
 - 9 **Walker WA**. Development of the intestinal mucosal barrier. *J Pediatr Gastroenterol Nutr* 2002; **34** Suppl 1: S33-S39 [PMID: 12082386]
 - 10 **Kansagra K**, Stoll B, Rognerud C, Niinikoski H, Ou CN, Harvey R, Burrin D. Total parenteral nutrition adversely affects gut barrier function in neonatal piglets. *Am J Physiol Gastrointest Liver Physiol* 2003; **285**: G1162-G1170 [PMID: 12969831]
 - 11 **Alverdy JC**, Aoys E, Moss GS. Total parenteral nutrition promotes bacterial translocation from the gut. *Surgery* 1988; **104**: 185-190 [PMID: 3135625]
 - 12 **Deitch EA**, Xu D, Naruhn MB, Deitch DC, Lu Q, Marino AA. Elemental diet and IV-TPN-induced bacterial translocation is associated with loss of intestinal mucosal barrier function against bacteria. *Ann Surg* 1995; **221**: 299-307 [PMID: 7717784]
 - 13 **Mosenthal AC**, Xu D, Deitch EA. Elemental and intravenous total parenteral nutrition diet-induced gut barrier failure is intestinal site specific and can be prevented by feeding nonfermentable fiber. *Crit Care Med* 2002; **30**: 396-402 [PMID: 11889319]
 - 14 **Ohland CL**, Macnaughton WK. Probiotic bacteria and intestinal epithelial barrier function. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G807-G819 [PMID: 20299599 DOI: 10.1152/ajpgi.00243.2009]
 - 15 **Ashida H**, Ogawa M, Kim M, Mimuro H, Sasakawa C. Bacteria and host interactions in the gut epithelial barrier. *Nat Chem Biol* 2012; **8**: 36-45 [PMID: 22173358 DOI: 10.1038/nchembio.741]
 - 16 **Sharma R**, Young C, Neu J. Molecular modulation of intestinal epithelial barrier: contribution of microbiota. *J Biomed Biotechnol* 2010; **2010**: 305879 [PMID: 20150966 DOI: 10.1155/2010/305879]
 - 17 **Peng L**, Li ZR, Green RS, Holzman IR, Lin J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J Nutr* 2009; **139**: 1619-1625 [PMID: 19625695 DOI: 10.3945/jn.109.104638]
 - 18 **Rakoff-Nahoum S**, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* 2004; **118**: 229-241 [PMID: 15260992]
 - 19 **Schumann A**, Nutten S, Donnicola D, Comelli EM, Mansourian R, Cherbut C, Corthesy-Theulaz I, Garcia-Rodenas C. Neonatal antibiotic treatment alters gastrointestinal tract developmental gene expression and intestinal barrier transcriptome. *Physiol Genomics* 2005; **23**: 235-245 [PMID: 16131529]
 - 20 **Rolfe RD**. Interactions among microorganisms of the indigenous intestinal flora and their influence on the host. *Rev Infect Dis* 1984; **6** Suppl 1: S73-S79 [PMID: 6372040]
 - 21 **Stecher B**, Hardt WD. Mechanisms controlling pathogen colonization of the gut. *Curr Opin Microbiol* 2011; **14**: 82-91 [PMID: 21036098 DOI: 10.1016/j.mib.2010.10.003]
 - 22 **Vollaard EJ**, Clasener HA. Colonization resistance. *Antimicrob Agents Chemother* 1994; **38**: 409-414 [PMID: 8203832]
 - 23 **Reid G**, Howard J, Gan BS. Can bacterial interference prevent infection? *Trends Microbiol* 2001; **9**: 424-428 [PMID: 11553454]
 - 24 **Liévin-Le Moal V**, Servin AL. The front line of enteric host defense against unwelcome intrusion of harmful microorganisms: mucins, antimicrobial peptides, and microbiota. *Clin Microbiol Rev* 2006; **19**: 315-337 [PMID: 16614252]
 - 25 **Marteau P**, Seksik P, Lepage P, Doré J. Cellular and physiological effects of probiotics and prebiotics. *Mini Rev Med Chem* 2004; **4**: 889-896 [PMID: 15544550]
 - 26 **Tourneur E**, Chassin C. Neonatal immune adaptation of the gut and its role during infections. *Clin Dev Immunol* 2013; **2013**: 270301 [PMID: 23737810 DOI: 10.1155/2013/270301]
 - 27 **Lotz M**, Gütle D, Walther S, Ménard S, Bogdan C, Hornef MW. Postnatal acquisition of endotoxin tolerance in intestinal epithelial cells. *J Exp Med* 2006; **203**: 973-984 [PMID: 16606665]
 - 28 **Hooper LV**. Do symbiotic bacteria subvert host immunity? *Nat Rev Microbiol* 2009; **7**: 367-374 [PMID: 19369952 DOI: 10.1038/nrmicro2114]
 - 29 **Hayashi H**, Sakamoto M, Benno Y. Phylogenetic analysis of the human gut microbiota using 16S rDNA clone libraries and strictly anaerobic culture-based methods. *Microbiol Immunol* 2002; **46**: 535-548 [PMID: 12363017]
 - 30 **Ley RE**, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell* 2006; **124**: 837-848 [PMID: 16497592]
 - 31 **Sonnenburg JL**, Angenent LT, Gordon JI. Getting a grip on things: how do communities of bacterial symbionts become established in our intestine? *Nat Immunol* 2004; **5**: 569-573 [PMID: 15164016]
 - 32 **Gallo RL**, Hooper LV. Epithelial antimicrobial defence of the skin and intestine. *Nat Rev Immunol* 2012; **12**: 503-516 [PMID: 22728527 DOI: 10.1038/nri3228]
 - 33 **Hooper LV**, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science* 2012; **336**: 1268-1273 [PMID: 22674334 DOI: 10.1126/science.1223490]
 - 34 **Guy-Grand D**, Griscelli C, Vassalli P. The gut-associated lymphoid system: nature and properties of the large dividing cells. *Eur J Immunol* 1974; **4**: 435-443 [PMID: 4213445]
 - 35 **Macpherson AJ**, Harris NL. Interactions between commensal intestinal bacteria and the immune system. *Nat Rev Immunol* 2004; **4**: 478-485 [PMID: 15173836]
 - 36 **Hooper LV**. Bacterial contributions to mammalian gut development. *Trends Microbiol* 2004; **12**: 129-134 [PMID: 15001189]
 - 37 **Mshvildadze M**, Neu J, Mai V. Intestinal microbiota development in the premature neonate: establishment of a lasting commensal relationship? *Nutr Rev* 2008; **66**: 658-663 [PMID: 19019028 DOI: 10.1111/j.1753-4887.2008.00119.x]
 - 38 **Conroy ME**, Shi HN, Walker WA. The long-term health effects of neonatal microbial flora. *Curr Opin Allergy Clin Immunol* 2009; **9**: 197-201 [PMID: 19398905 DOI: 10.1097/ACI.0b013e32832b3f1d]
 - 39 **Leatham MP**, Banerjee S, Autieri SM, Mercado-Lubo R, Conway T, Cohen PS. Precolonized human commensal *Escherichia coli* strains serve as a barrier to *E. coli* O157: H7 growth in the streptomycin-treated mouse intestine. *Infect Immun* 2009; **77**: 2876-2886 [PMID: 19364832 DOI: 10.1128/IAI.00059-09]
 - 40 **Rakoff-Nahoum S**, Hao L, Medzhitov R. Role of toll-like receptors in spontaneous commensal-dependent colitis. *Immunity* 2006; **25**: 319-329 [PMID: 16879997]
 - 41 **Artis D**. Epithelial-cell recognition of commensal bacteria and maintenance of immune homeostasis in the gut. *Nat Rev Immunol* 2008; **8**: 411-420 [PMID: 18469830 DOI: 10.1038/nri2316]
 - 42 **Sirard JC**, Bayardo M, Didierlaurent A. Pathogen-specific

- TLR signaling in mucosa: mutual contribution of microbial TLR agonists and virulence factors. *Eur J Immunol* 2006; **36**: 260-263 [PMID: 16453385]
- 43 **Barbara G**, Stanghellini V, Brandi G, Cremon C, Di Nardo G, De Giorgio R, Corinaldesi R. Interactions between commensal bacteria and gut sensorimotor function in health and disease. *Am J Gastroenterol* 2005; **100**: 2560-2568 [PMID: 16279914]
- 44 **Vaishnav S**, Behrendt CL, Ismail AS, Eckmann L, Hooper LV. Paneth cells directly sense gut commensals and maintain homeostasis at the intestinal host-microbial interface. *Proc Natl Acad Sci USA* 2008; **105**: 20858-20863 [PMID: 19075245 DOI: 10.1073/pnas.0808723105]
- 45 **Slack E**, Hapfelmeier S, Stecher B, Velykoredko Y, Stoel M, Lawson MA, Geuking MB, Beutler B, Tedder TF, Hardt WD, Bercik P, Verdu EF, McCoy KD, Macpherson AJ. Innate and adaptive immunity cooperate flexibly to maintain host-microbiota mutualism. *Science* 2009; **325**: 617-620 [PMID: 19644121 DOI: 10.1126/science.1172747]
- 46 **Pull SL**, Doherty JM, Mills JC, Gordon JL, Stappenbeck TS. Activated macrophages are an adaptive element of the colonic epithelial progenitor niche necessary for regenerative responses to injury. *Proc Natl Acad Sci USA* 2005; **102**: 99-104 [PMID: 15615857]
- 47 **Medzhitov R**. Recognition of microorganisms and activation of the immune response. *Nature* 2007; **449**: 819-826 [PMID: 17943118]
- 48 **Beutler BA**. TLRs and innate immunity. *Blood* 2009; **113**: 1399-1407 [PMID: 18757776 DOI: 10.1182/blood-2008-07-019307]
- 49 **Watson JL**, McKay DM. The immunophysiological impact of bacterial CpG DNA on the gut. *Clin Chim Acta* 2006; **364**: 1-11 [PMID: 16153626]
- 50 **Mazmanian SK**, Round JL, Kasper DL. A microbial symbiosis factor prevents intestinal inflammatory disease. *Nature* 2008; **453**: 620-625 [PMID: 18509436 DOI: 10.1038/nature07008]
- 51 **Round JL**, Mazmanian SK. Inducible Foxp3⁺ regulatory T-cell development by a commensal bacterium of the intestinal microbiota. *Proc Natl Acad Sci USA* 2010; **107**: 12204-12209 [PMID: 20566854 DOI: 10.1073/pnas.0909122107]
- 52 **Kinnebrew MA**, Ubeda C, Zenewicz LA, Smith N, Flavell RA, Pamer EG. Bacterial flagellin stimulates Toll-like receptor 5-dependent defense against vancomycin-resistant *Enterococcus* infection. *J Infect Dis* 2010; **201**: 534-543 [PMID: 20064069 DOI: 10.1086/650203]
- 53 **Neish AS**, Gewirtz AT, Zeng H, Young AN, Hobert ME, Karmali V, Rao AS, Madara JL. Prokaryotic regulation of epithelial responses by inhibition of IkappaB-alpha ubiquitination. *Science* 2000; **289**: 1560-1563 [PMID: 10968793]
- 54 **Neish AS**, Naumann M. Microbial-induced immunomodulation by targeting the NF- κ B system. *Trends Microbiol* 2011; **19**: 596-605 [PMID: 21955402 DOI: 10.1016/j.tim.2011.08.004]
- 55 **Tien MT**, Girardin SE, Regnault B, Le Bourhis L, Dillies MA, Coppée JY, Bourdet-Sicard R, Sansonetti PJ, Pédrón T. Anti-inflammatory effect of *Lactobacillus casei* on Shigella-infected human intestinal epithelial cells. *J Immunol* 2006; **176**: 1228-1237 [PMID: 16394013]
- 56 **Specian RD**, Oliver MG. Functional biology of intestinal goblet cells. *Am J Physiol* 1991; **260**: C183-C193 [PMID: 1996606]
- 57 **Yu K**, Jiang SF, Lin MF, Wu JB, Lin J. Extraction and purification of biologically active intestinal trefoil factor from human meconium. *Lab Invest* 2004; **84**: 390-392 [PMID: 14767484]
- 58 **Belley A**, Keller K, Göttke M, Chadee K. Intestinal mucins in colonization and host defense against pathogens. *Am J Trop Med Hyg* 1999; **60**: 10-15 [PMID: 10344672]
- 59 **Falk PG**, Hooper LV, Midtvedt T, Gordon JL. Creating and maintaining the gastrointestinal ecosystem: what we know and need to know from gnotobiology. *Microbiol Mol Biol Rev* 1998; **62**: 1157-1170 [PMID: 9841668]
- 60 **Deplancke B**, Gaskins HR. Microbial modulation of innate defense: goblet cells and the intestinal mucus layer. *Am J Clin Nutr* 2001; **73**: 1131S-1141S [PMID: 11393191]
- 61 **Forder RE**, Howarth GS, Tivey DR, Hughes RJ. Bacterial modulation of small intestinal goblet cells and mucin composition during early posthatch development of poultry. *Poult Sci* 2007; **86**: 2396-2403 [PMID: 17954591]
- 62 **Brandtzaeg P**. Update on mucosal immunoglobulin A in gastrointestinal disease. *Curr Opin Gastroenterol* 2010; **26**: 554-563 [PMID: 20693891 DOI: 10.1097/MOG.0b013e32833dccc8]
- 63 **Johansen FE**, Pekna M, Norderhaug IN, Haneberg B, Hietala MA, Krajci P, Betsholtz C, Brandtzaeg P. Absence of epithelial immunoglobulin A transport, with increased mucosal leakiness, in polymeric immunoglobulin receptor/secretory component-deficient mice. *J Exp Med* 1999; **190**: 915-922 [PMID: 10510081]
- 64 **Nahmias A**, Stoll B, Hale E, Ibegbu C, Keyserling H, Innis-Whitehouse W, Holmes R, Spira T, Czerkinsky C, Lee F. IgA-secreting cells in the blood of premature and term infants: normal development and effect of intrauterine infections. *Adv Exp Med Biol* 1991; **310**: 59-69 [PMID: 1809028]
- 65 **Mellander L**, Carlsson B, Jalil F, Söderström T, Hanson LA. Secretory IgA antibody response against *Escherichia coli* antigens in infants in relation to exposure. *J Pediatr* 1985; **107**: 430-433 [PMID: 2411902]
- 66 **Sartor RB**. Genetics and environmental interactions shape the intestinal microbiome to promote inflammatory bowel disease versus mucosal homeostasis. *Gastroenterology* 2010; **139**: 1816-1819 [PMID: 21029802 DOI: 10.1053/j.gastro.2010.10.036]
- 67 **Papoff P**, Ceccarelli G, d'Ettoire G, Cerasaro C, Caresta E, Midulla F, Moretti C. Gut microbial translocation in critically ill children and effects of supplementation with pre- and probiotics. *Int J Microbiol* 2012; **2012**: 151393 [PMID: 22934115 DOI: 10.1155/2012/151393]
- 68 **Torrazza RM**, Neu J. The altered gut microbiome and necrotizing enterocolitis. *Clin Perinatol* 2013; **40**: 93-108 [PMID: 23415266 DOI: 10.1016/j.clp.2012.12.009]
- 69 **Mylonaki M**, Rayment NB, Rampton DS, Hudspith BN, Brostoff J. Molecular characterization of rectal mucosa-associated bacterial flora in inflammatory bowel disease. *Inflamm Bowel Dis* 2005; **11**: 481-487 [PMID: 15867588]
- 70 **Sokol H**, Seksik P, Furet JP, Firmesse O, Nion-Larmurier I, Beaugerie L, Cosnes J, Corthier G, Marteau P, Doré J. Low counts of *Faecalibacterium prausnitzii* in colitis microbiota. *Inflamm Bowel Dis* 2009; **15**: 1183-1189 [PMID: 19235886 DOI: 10.1002/ibd.20903]
- 71 **Skowitz A**, Jacobi M, Frick JS, Richter M, Rusch K, Köhler H. Microbiota in pediatric inflammatory bowel disease. *J Pediatr* 2010; **157**: 240-244.e1 [PMID: 20400104 DOI: 10.1016/j.jpeds.2010.02.046]
- 72 **Conte MP**, Schippa S, Zamboni I, Penta M, Chiarini F, Seganti L, Osborn J, Falconieri P, Borrelli O, Cucchiara S. Gut-associated bacterial microbiota in paediatric patients with inflammatory bowel disease. *Gut* 2006; **55**: 1760-1767 [PMID: 16648155]
- 73 **Ubeda C**, Taur Y, Jenq RR, Equinda MJ, Son T, Samstein M, Viale A, Socci ND, van den Brink MR, Kamboj M, Pamer EG. Vancomycin-resistant *Enterococcus* domination of intestinal microbiota is enabled by antibiotic treatment in mice and precedes bloodstream invasion in humans. *J Clin Invest* 2010; **120**: 4332-4341 [PMID: 21099116 DOI: 10.1172/JCI43918]
- 74 **Ubeda C**, Bucci V, Caballero S, Djukovic A, Toussaint NC, Equinda M, Lipuma L, Ling L, Gobourne A, No D, Taur Y, Jenq RR, van den Brink MR, Xavier JB, Pamer EG. Intestinal microbiota containing *Barnesiella* species cures vancomycin-resistant *Enterococcus faecium* colonization. *Infect Immun* 2013; **81**: 965-973 [PMID: 23319552 DOI: 10.1128/IAI.01197-12]

- 75 **MacFie J**, O'Boyle C, Mitchell CJ, Buckley PM, Johnstone D, Sudworth P. Gut origin of sepsis: a prospective study investigating associations between bacterial translocation, gastric microflora, and septic morbidity. *Gut* 1999; **45**: 223-228 [PMID: 10403734]
- 76 **Sherman MP**. New concepts of microbial translocation in the neonatal intestine: mechanisms and prevention. *Clin Perinatol* 2010; **37**: 565-579 [PMID: 20813271 DOI: 10.1016/j.clp.2010.05.006]
- 77 **Mai V**, Torrazza RM, Ukhanova M, Wang X, Sun Y, Li N, Shuster J, Sharma R, Hudak ML, Neu J. Distortions in development of intestinal microbiota associated with late onset sepsis in preterm infants. *PLoS One* 2013; **8**: e52876 [PMID: 23341915 DOI: 10.1371/journal.pone.0052876]
- 78 **Nair V**, Soraisham AS. Probiotics and prebiotics: role in prevention of nosocomial sepsis in preterm infants. *Int J Pediatr* 2013; **2013**: 874726 [PMID: 23401695 DOI: 10.1155/2013/874726]
- 79 **Carlisle EM**, Morowitz MJ. The intestinal microbiome and necrotizing enterocolitis. *Curr Opin Pediatr* 2013; **25**: 382-387 [PMID: 23657248 DOI: 10.1097/MOP.0b013e3283600e91]
- 80 **Gibbs K**, Lin J, Holzman IR. Necrotizing enterocolitis: the state of the science. *Indian J Pediatr* 2007; **74**: 67-72 [PMID: 17264459]
- 81 **Cotten CM**, Taylor S, Stoll B, Goldberg RN, Hansen NI, Sanchez PJ, Ambalavanan N, Benjamin DK. Prolonged duration of initial empirical antibiotic treatment is associated with increased rates of necrotizing enterocolitis and death for extremely low birth weight infants. *Pediatrics* 2009; **123**: 58-66 [PMID: 19117861 DOI: 10.1542/peds.2007-3423]
- 82 **Alexander VN**, Northrup V, Bizzarro MJ. Antibiotic exposure in the newborn intensive care unit and the risk of necrotizing enterocolitis. *J Pediatr* 2011; **159**: 392-397 [PMID: 21489560 DOI: 10.1016/j.jpeds.2011.02.035]
- 83 **Weintraub AS**, Ferrara L, Deluca L, Moshier E, Green RS, Oakman E, Lee MJ, Rand L. Antenatal antibiotic exposure in preterm infants with necrotizing enterocolitis. *J Perinatol* 2012; **32**: 705-709 [PMID: 22157626 DOI: 10.1038/jp.2011.180]
- 84 **Neu J**, Walker WA. Necrotizing enterocolitis. *N Engl J Med* 2011; **364**: 255-264 [PMID: 21247316 DOI: 10.1056/NEJMra1005408]
- 85 **Wang Y**, Hoening JD, Malin KJ, Qamar S, Petrof EO, Sun J, Antonopoulos DA, Chang EB, Claud EC. 16S rRNA gene-based analysis of fecal microbiota from preterm infants with and without necrotizing enterocolitis. *ISME J* 2009; **3**: 944-954 [PMID: 19369970]
- 86 **Mai V**, Young CM, Ukhanova M, Wang X, Sun Y, Casella G, Theriaque D, Li N, Sharma R, Hudak M, Neu J. Fecal microbiota in premature infants prior to necrotizing enterocolitis. *PLoS One* 2011; **6**: e20647 [PMID: 21674011 DOI: 10.1371/journal.pone.0020647]
- 87 **Mshvildadze M**, Neu J, Shuster J, Theriaque D, Li N, Mai V. Intestinal microbial ecology in premature infants assessed with non-culture-based techniques. *J Pediatr* 2010; **156**: 20-25 [PMID: 19783002 DOI: 10.1016/j.jpeds.2009.06.063]
- 88 **Lin J**. Too much short chain fatty acids cause neonatal necrotizing enterocolitis. *Med Hypotheses* 2004; **62**: 291-293 [PMID: 14962641]
- 89 **Lin J**. Effects of short chain fatty acids on the intestinal barrier. *Curr Nutr Food Sci* 2013; **9**: 93-98
- 90 **Clark DA**, Thompson JE, Weiner LB, McMillan JA, Schneider AJ, Rokahr JE. Necrotizing enterocolitis: intraluminal biochemistry in human neonates and a rabbit model. *Pediatr Res* 1985; **19**: 919-921 [PMID: 4047761]
- 91 **Garstin WI**, Boston VE. Sequential assay of expired breath hydrogen as a means of predicting necrotizing enterocolitis in susceptible infants. *J Pediatr Surg* 1987; **22**: 208-210 [PMID: 3559860]
- 92 **Cheu HW**, Brown DR, Rowe MI. Breath hydrogen excretion as a screening test for the early diagnosis of necrotizing enterocolitis. *Am J Dis Child* 1989; **143**: 156-159 [PMID: 2492749]
- 93 **Engel RR**, Virning NL, Hunt CE, Levitt MD. Origin of mural gas in necrotizing enterocolitis. *Pediatr Res* 1973; **7**: 292 (abstract)
- 94 **Woods CW**, Oliver T, Lewis K, Yang Q. Development of necrotizing enterocolitis in premature infants receiving thickened feeds using SimplyThick®. *J Perinatol* 2012; **32**: 150-152 [PMID: 22289705 DOI: 10.1038/jp.2011.105]
- 95 **Beal J**, Silverman B, Bellant J, Young TE, Klontz K. Late onset necrotizing enterocolitis in infants following use of a xanthan gum-containing thickening agent. *J Pediatr* 2012; **161**: 354-356 [PMID: 22575248 DOI: 10.1016/j.jpeds.2012.03.054]
- 96 **Bin-Nun A**, Bromiker R, Wilschanski M, Kaplan M, Rudensky B, Caplan M, Hammerman C. Oral probiotics prevent necrotizing enterocolitis in very low birth weight neonates. *J Pediatr* 2005; **147**: 192-196 [PMID: 16126048]
- 97 **Lin HC**, Hsu CH, Chen HL, Chung MY, Hsu JF, Lien RI, Tsao LY, Chen CH, Su BH. Oral probiotics prevent necrotizing enterocolitis in very low birth weight preterm infants: a multicenter, randomized, controlled trial. *Pediatrics* 2008; **122**: 693-700 [PMID: 18829790 DOI: 10.1542/peds.2007-3007]
- 98 **Deshpande G**, Rao S, Patole S, Bulsara M. Updated meta-analysis of probiotics for preventing necrotizing enterocolitis in preterm neonates. *Pediatrics* 2010; **125**: 921-930 [PMID: 20403939 DOI: 10.1542/peds.2009-1301]

P- Reviewers: Choe BH, Urganci N **S- Editor:** Gou SX

L- Editor: A **E- Editor:** Wang CH



Imaging evaluation of hemoptysis in children

Divya Singh, Ashu Seith Bhalla, Prasad Thotton Veedu, Arundeep Arora

Divya Singh, Ashu Seith Bhalla, Prasad Thotton Veedu, Arundeep Arora, Department of Radiodiagnosis, All India Institute of Medical Sciences, New Delhi 110029, India

Author contributions: Singh D, Arora A and Thotton Veedu P completed the initial literature survey; Singh D and Bhalla AS drafted the manuscript; all authors read and approved the final manuscript.

Correspondence to: Ashu Seith Bhalla, MD, MAMS, Additional Professor, Department of Radiodiagnosis, All India Institute of Medical Sciences, Ansari Nagar, New Delhi 110029, India. ashubhalla1@yahoo.com

Telephone: +91-11-26594925 Fax: +91-98-68398805

Received: May 28, 2013 Revised: September 4, 2013

Accepted: October 16, 2013

Published online: November 8, 2013

Abstract

Hemoptysis is an uncommon but distressing symptom in children. It poses a diagnostic challenge as it is difficult to elicit a clear history and perform thorough physical examination in a child. The cause of hemoptysis in children can vary with the child's age. It can range from infection, milk protein allergy and congenital heart disease in early childhood, to vasculitis, bronchial tumor and bronchiectasis in older children. Acute lower respiratory tract infections are the most common cause of pediatric hemoptysis. The objective of imaging is to identify the source of bleeding, underlying primary cause, and serve as a roadmap for invasive procedures. Hemoptysis originates primarily from the bronchial arteries. The imaging modalities available for the diagnostic evaluation of hemoptysis include chest radiography, multi-detector computed tomography (MDCT), magnetic resonance imaging (MRI) and catheter angiography. Chest radiography is the initial screening tool. It can help in lateralizing the bleeding with high degree of accuracy and can detect several parenchymal and pleural abnormalities. However, it may be normal in up to 30% cases. MDCT is a rapid, non-invasive multiplanar imaging modality. It aids in evaluation of hemoptysis by depiction of underlying disease, assessment of

consequences of hemorrhage and provides panoramic view of the thoracic vasculature. The various structures which need to be assessed carefully include the pulmonary parenchyma, tracheobronchial tree, pulmonary arteries, bronchial arteries and non-bronchial systemic arteries. Since the use of MDCT entails radiation exposure, optimal low dose protocols should be used so as to keep radiation dose as low as reasonably achievable. MRI and catheter angiography have limited application.

© 2013 Baishideng. All rights reserved.

Key words: Hemoptysis; Lower respiratory tract infection; Bronchiectasis; Cystic fibrosis; Foreign body

Core tip: Hemoptysis is a cause of immense concern to the child, the family and the pediatrician. Thorough history and physical examination is necessary to ascertain its presence, which is particularly challenging in the pediatric population. Imaging has an important role in identifying the source of bleeding and its underlying cause. Acute lower respiratory tract infections are the most common cause of pediatric hemoptysis. The imaging modalities include chest radiography, multi-detector computed tomography (MDCT), magnetic resonance imaging (MRI) and catheter angiography. MDCT is a rapid multiplanar imaging modality which should be used judiciously to keep radiation dose to a minimum. MRI and catheter angiography have selected application.

Singh D, Bhalla AS, Thotton Veedu P, Arora A, Imaging evaluation of hemoptysis in children. *World J Clin Pediatr* 2013; 2(4): 54-64 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/54.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.54>

INTRODUCTION

Hemoptysis is defined as expectoration of blood or blood tinged-sputum due to bleeding from the respiratory tract^[1]. Massive hemoptysis is termed as blood loss > 8

Table 1 Causes of hemoptysis in children

Acute lower respiratory tract infections
Bacterial
Viral
Fungal
Parasitic
Bronchiectasis
Aspiration
Cystic fibrosis
Ciliary dyskinesia
Post-infectious
Congenital heart diseases
Eisenmenger syndrome
Aplasia/hypoplasia of pulmonary artery or veins
Primary pulmonary hypertension
Pulmonary artery narrowing
Infectious
Inflammatory
Pulmonary thromboembolism
Pulmonary arteriovenous malformation
Alveolar hemorrhage syndrome
Idiopathic
Associated with rheumatologic disease
Pulmonary-renal syndrome
Neoplasms
Bronchial carcinoid
Bronchial adenoma
Metastatic
Foreign body
Trauma
Cryptogenic

mL/kg in 24 h^[2]. The life-threatening element in massive hemoptysis is asphyxiation due to flooding of the airways by blood. Hence, securing the airway needs immediate attention. Hemoptysis in lesser amounts poses a diagnostic challenge in pediatric patients as it may initially remain unnoticed because children tend to swallow their sputum and are unable to provide a clear history or undergo thorough physical examination. It is a cause of immense concern to the child, the parents and the pediatrician. After confirming the presence of hemoptysis, the next step is to establish the cause, so that an appropriate treatment regimen can be adopted. The spectrum of causes of hemoptysis in children is considerably different from that of the adults. Imaging has a pivotal role in evaluation of hemoptysis. There are various modalities which can be resorted to, namely, conventional radiography, multi-detector computed tomography (MDCT), magnetic resonance imaging (MRI) and in certain cases, catheter angiography which can also fulfill a therapeutic purpose. The advent of MDCT has paved the way for non-invasive multi-dimensional visualization of the thoracic vasculature, tracheobronchial tree and pulmonary parenchyma. This is of tremendous value as it can obviate the need for invasive bronchoscopic procedure with its attendant complications. The following sections illustrate the etiology, pathogenesis and role of imaging in hemoptysis.

ETIOPATHOGENESIS

There are several causes of hemoptysis in children (Table 1).

The common causes are acute lower respiratory tract infections, bronchiectasis (due to cystic fibrosis, aspiration, ciliary dyskinesias, post infectious), congenital heart disease and foreign body aspiration. Of these, acute lower respiratory tract infections may constitute upto 40% of cases^[3]. The etiology also varies with the child's age. Sim *et al*^[4] observed that infection, Heiner syndrome (milk protein allergy) and congenital heart disease were the major causes in early childhood; while during late childhood, vasculitis, bronchial tumor and bronchiectasis were far more prevalent.

The lungs receive dual blood supply; one from the high pressure bronchial arteries, the other from the pulmonary arteries with relatively lower pressure. The pulmonary arteries account for 99% of the arterial blood supply to the lungs and take part in gas exchange while the bronchial arteries provide nourishment to the supporting structures of the airways and form the vasa vasora of the pulmonary arteries. The bronchial vasculature is in close proximity to the pulmonary arteries at the level of the vasa vasorum where the two systems are connected by thin-walled anastomoses between the systemic and pulmonary capillaries^[5,6]. Pulmonary vascular obstructive disorders (congenital heart disease, vasculitis, embolism) open up these anastomoses in regions of the lung that are deprived of their pulmonary arterial blood flow. This subjects these fragile vessels to increased systemic arterial pressure and can cause hemoptysis by rupturing into the alveoli or bronchial airways.

In the setting of tracheobronchial infection, there is inflammation of the airways. As a result, they become congested and friable, which increases their susceptibility to bleed. Chronic inflammation (as in bronchiectasis) can lead to increase in systemic arterial flow due to release of angiogenic growth factors which lead to neovascularization and formation of "leaky" vessels prone to rupture. Approximately 5% of patients with cystic fibrosis can present with massive hemoptysis^[7]. This is due to hypertrophy of bronchial arteries along with the presence of multiple bronchopulmonary anastomoses. Foreign body aspiration causes hemoptysis due to mechanical trauma or due to associated intense inflammation incited by organic matter. Pulmonary hemosiderosis is an uncommon but significant cause of hemoptysis in children. It is mostly idiopathic; however, it may be associated with an allergy to cow's milk (Heiner syndrome)^[8]. Although rare in children, endobronchial or pulmonary parenchymal tumors (carcinoid, bronchial adenoma) may cause significant hemorrhage.

Imaging evaluation of hemoptysis

The aim of initial diagnostic evaluation is to identify the immediate source of bleeding along with determination of the primary cause of hemoptysis. Traditionally, the diagnostic algorithms in acute setting have been based on various combinations of conventional radiography, chest computed tomography (CT) and thoracic aortography. MDCT has now made comprehensive visualization of

thoracic anatomy possible. It provides high-resolution images of the thoracic and upper abdominal vasculature which aids in diagnosis and also provides a roadmap prior to any intervention. CT findings can forewarn the endoscopist about the presence of peribronchial or endoluminal aneurysms. MRI does not have a role in the acute setting. However, it may serve as a problem-solving tool in certain situations.

Conventional radiography

Chest radiography serves as a valuable screening modality. It can help in lateralizing the bleeding with high degree of accuracy and can detect several parenchymal and pleural abnormalities. The commonly used views include frontal and, in some cases, lateral. The utility of lateral radiographs is in case of presence of a radio-opaque foreign body on frontal view, when it can determine if it is in the trachea or esophagus. Lynch *et al*^[9] observed that addition of a lateral radiograph in children with pneumonia did not improve diagnostic accuracy. Common findings include presence of focal infiltrates which may suggest infection. Unilateral air-trapping with hyperinflation can give a clue towards presence of an unsuspected tracheobronchial foreign body. A radio-opaque foreign body may be seen. Ancillary findings include “tram-track” appearance of bronchiectasis; pulmonary nodules, lymphadenopathy, pleural effusion; cardiomegaly; and vascular redistribution due to pulmonary venous obstruction. Approximately one-third of chest radiographs may be normal in children with hemoptysis. A tracheobronchial source of bleed may eventually be identified in about half of these cases^[10]. Therefore, additional follow-up testing is recommended in patients with hemoptysis in whom the underlying cause is not detected by initial radiography.

MDCT

The role of MDCT in evaluation of hemoptysis includes: (1) depiction of underlying disease; (2) assessment of consequences of hemorrhage which may be a cause of clinical concern or may conceal the underlying abnormalities; and (3) panoramic visualization of the thoracic vasculature by various reconstruction techniques.

Technique

CT technique involves acquisition of multiple sections from the base of the neck to the level of the renal arteries (L2 level). Optimal enhancement of the pulmonary and systemic arteries is achieved by administration of 2 mL/kg body weight of iodinated non-ionic contrast media containing 300 mg I/mL at a rate of 4 mL/s *via* a wide gauge cannula. The scan should be started during the phase of peak systemic arterial enhancement (scanning delay of 18 s or a threshold of 100 HU in the descending aorta with bolus tracking). Images should be acquired with thin collimation and with the table movement adjusted to allow wide volume coverage during a single breath-hold. Radiation exposure is a significant

concern in the pediatric population. Hence, the exposure parameters and kilovoltage need to be adjusted according to the patient's weight so as to minimize radiation dose with optimal image quality.

Data processing and interpretation

Since a large volume of data is acquired, the images are best interpreted at the scanner console or remote workstation. The soft-tissue structures and lung parenchyma can be assessed adequately in axial sections of 5 mm thickness with mediastinal and lung window settings, respectively (Figure 1A and B). High resolution CT images allow detailed evaluation of the pulmonary parenchyma. The tracheobronchial airway can be evaluated on thinner sections and reformatted images.

Two-dimensional maximum intensity projection images (Figure 1C) in the coronal/oblique and sagittal planes can demonstrate the tortuous course of the bronchial arteries from the descending thoracic aorta to the lungs. Intercostal and internal mammary arteries are best visualized in the coronal planes while the inferior phrenic arteries and celiac axis branches are demonstrated well in axial images. Three-dimensional volumetric and shaded-surface-display images not only depict the abnormal vessel, but also illustrate its relationship with the surrounding structures, thus providing a preview of the internal anatomy.

Minimum Intensity Projection can be used to generate images of the central airways (Figure 1D) and demonstrate areas of air-trapping within the lungs. These can provide valuable perspective in defining a lesion prior to any intervention. Therefore, a host of reconstructed images need to be analyzed for a thorough CT assessment of hemoptysis.

STEPWISE STRUCTURAL ASSESSMENT

Lung parenchyma

The pulmonary parenchyma should be evaluated for presence of bronchiectasis, consolidation and ground-glass opacity. The site of hemorrhage can be localized on the basis of presence of fluid density material in the segmental and lobar bronchi and ground-glass opacity with hazy consolidation which represents alveolar hemorrhage. Acute hemorrhage can mask the underlying pathology. Blood clots can also simulate more ominous entities like masses.

Tracheobronchial tree

This should be evaluated for presence of any stenosis which may be due to intraluminal (foreign body, neoplasm) or extraluminal (lymphadenopathy, fibrosing mediastinitis) causes. Multiplanar reformatted (MPR) images are accurate in detection of lesions, depiction of degree of narrowing, distal visualization and calculation of distance of the lesion from the carina in selected locations^[11].

Pulmonary arteries

The pulmonary arteries should be analyzed for any narrowing due to extrinsic or intrinsic causes. The presence

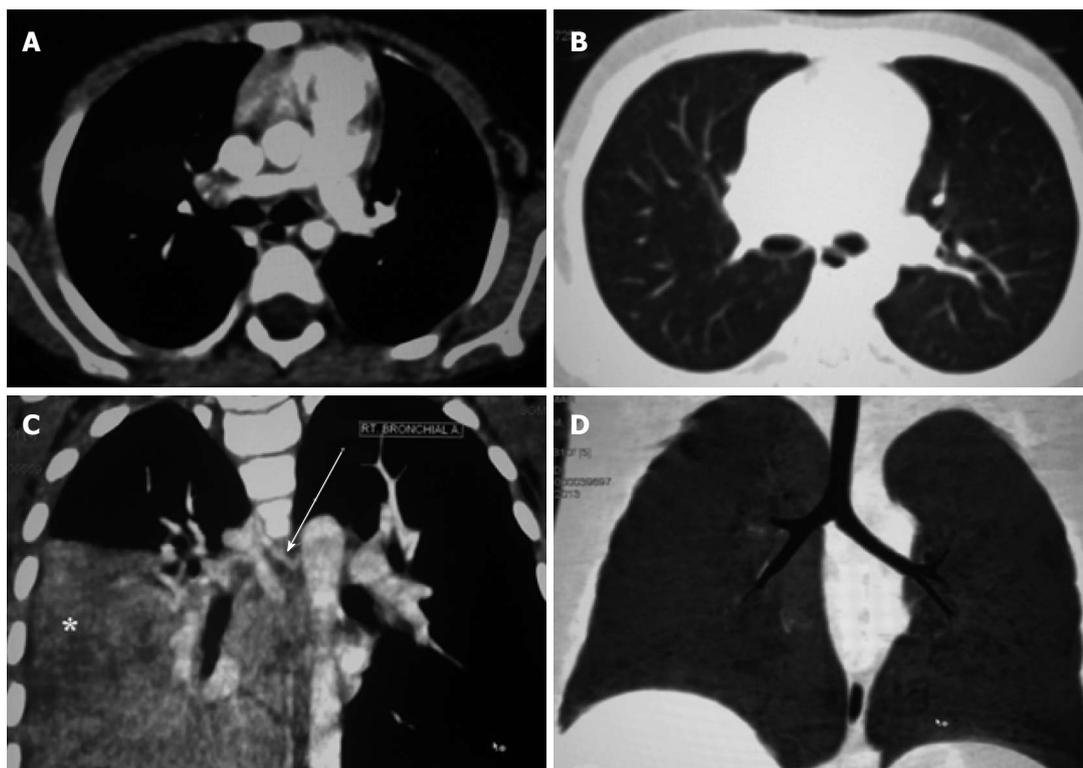


Figure 1 Multi-detector computed tomography image interpretation. Axial computed tomography image showing mediastinal window (A) and lung window (B). Coronal maximum intensity projection image (C) demonstrating the origin and proximal portion of the right bronchial artery (arrow). There is consolidation in the right lower lobe (asterisk). Coronal Minimum Intensity Projection image (D) delineating the central airways.

of accompanying subpleural areas of enhancement can represent lung infarcts. The pulmonary arteries can also show dilatation (Rasmussen aneurysm) and pulmonary arteriovenous malformations.

Bronchial arteries

Hemoptysis originates from bronchial arteries in 95% of cases^[12]. A bronchial artery diameter of more than 2 mm is considered abnormal^[13]. In 70% of individuals, bronchial arteries arise from the descending thoracic aorta between T5 and T6 levels. There are usually one or two bronchial arteries supplying each lung, arising independently or from a common trunk. They are visualized as a cluster of enhancing nodules in the posterior mediastinum just below the level of the aortic arch on axial images. Active bleeding can rarely be detected on CT. Anomalous bronchial arteries are defined as arteries which originate outside the T5-T6 level. Their most common site of origin is the concavity of the aortic arch^[14].

Non-bronchial systemic arteries

Non-bronchial systemic arteries can arise from the branches of brachiocephalic arteries, subclavian arteries, axillary, internal mammary and infradiaphragmatic branches from the inferior phrenic artery and celiac axis^[15,16]. On CT, these are seen as dilated tortuous arteries that are not parallel to the bronchi. The presence of pleural thickening greater than 3 mm with enhancing arteries within the extrapleural fat is a pointer of presence of these vessels^[17].

MRI

MRI does not have any utility in imaging evaluation of acute hemoptysis. Since it has superior soft-tissue resolution, it is excellent in the evaluation of the mediastinum and hilum in the non-emergent setting. It provides less information about the lung parenchyma. It may be used to demonstrate arteriovenous malformations and congenital anomalies of the pulmonary arteries and delineate the nature of mediastinal soft-tissue in fibrosing mediastinitis. With the introduction of hyperpolarized nuclei like ^3He and ^{129}Xe , the horizon of MRI is likely to expand from limited utility in evaluating the pulmonary parenchyma to evaluation of pulmonary structure, function and metabolism with a high sensitivity^[18]. Ventilation and dynamic imaging in patients with asthma and cystic fibrosis have shown regional patterns of obstruction and ventilation defects in these individuals. Further knowledge can go a long way in the early diagnosis, monitoring disease progression and evaluation of response to treatment^[19-21].

COMMON CAUSES OF PEDIATRIC HEMOPTYSIS

Acute lower respiratory tract infections

Tracheobronchitis, pneumonia and lung abscess can lead to hemoptysis. The infective process may be bacterial, viral, fungal or parasitic in origin. Although tuberculosis is a significant cause of adult hemoptysis, very few cases have been reported in the pediatric literature^[22]. Chest radio-

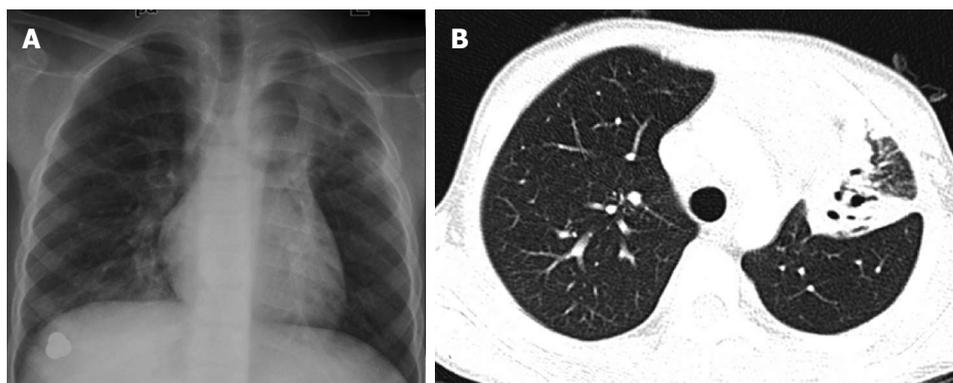


Figure 2 Chest radiograph (A) and axial computed tomography image (B) showing consolidation with cavitation in the left upper lobe.

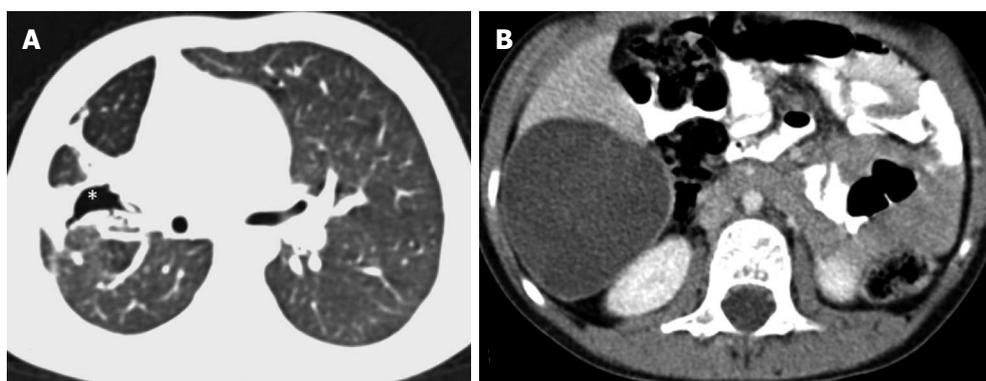


Figure 3 A seven-year-old girl with ruptured pulmonary hydatid cyst. Axial computed tomography image (A) showing the ruptured cyst with air (asterisk) in the right upper lobe along with surrounding consolidation. Axial section of the abdomen (B) shows an unruptured cyst in the segment VI of the liver.

graphs can show pulmonary infiltrates, hyperinflation and cavity with or without air-fluid level (Figure 2A). There may be associated pleural effusion and lymphadenopathy. CT findings can be in the form of consolidation, ground-glass opacity, interstitial thickening, air-trapping, cavity with shaggy walls and air-fluid level, pleural effusion and mediastinal or hilar lymphadenopathy (Figure 2B). CT can also demonstrate complications like empyema (thick enhancing visceral and parietal pleura, “split pleura” sign), bronchopleural fistula, *etc.*

Parasitic cysts (*Echinococcus*) can cause hemoptysis by rupturing into the airway. These may be seen as fluid density lesions with a smooth wall and air foci due to communication with the adjacent bronchus (Figure 3A). Detached membranes and daughter cysts can be visualized within the cyst. Concomitant cysts may be seen in other organs, most commonly in the liver (Figure 3B).

The most commonly implicated fungus is *Aspergillus*. It can have a varied spectrum of presentation, namely aspergilloma, allergic bronchopulmonary aspergillosis (ABPA), semi-invasive aspergillosis, airway or angioinvasive aspergillosis^[23,24]. Aspergilloma is the saprophytic colonization of a pre-existing cavity by the fungus and is typically seen as an opacity within a cavity producing the “air-crescent” sign. It is mobile and can show postural change in position. ABPA is a manifestation of type I and III hypersensitivity reaction to the organ-

ism and presents as central bronchiectasis with mucous plugged bronchi producing ‘finger-in-glove’ appearance with upper lobe preponderance on radiograph. The mucous plugs have a high-density on CT (Figure 4). There may be centrilobular nodules with “tree-in-bud” appearance. Some patients can also have associated allergic fungal sinonasal disease. Invasive aspergillosis is encountered in immunocompromised patients. Invasive airway disease presents as peribronchial areas of consolidation and multiple branching centrilobular nodules on CT^[25]. Nodules with surrounding ground-glass opacity (halo sign) or pleural-based, wedge shaped areas of consolidation (Figure 5) are the hallmark of angioinvasive aspergillosis^[26].

Bronchiectasis

Bronchiectasis can occur secondary to aspiration, infections, cystic fibrosis and ciliary dyskinesias. On chest radiographs, it manifests as “tram-track”, parallel line opacities, ring opacities and tubular structures (Figure 6A). However, chest radiographs are insensitive for detecting mild to moderate disease. CT (Figure 6B) has a higher sensitivity and on CT imaging, bronchiectasis is characterized by the absence of normal bronchial tapering, the presence of visible bronchi in the peripheral 1 cm of the lung and a bronchoarterial ratio more than 1 (signet ring sign). The etiology can be narrowed by considering the anatomic location and distribution of pathology.

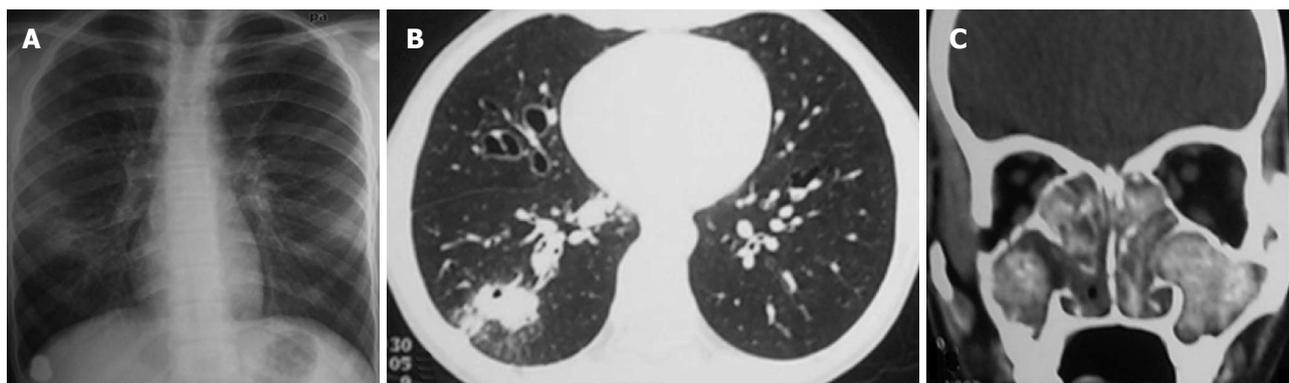


Figure 4 A 12-year-old girl with allergic bronchopulmonary aspergillosis. Frontal chest radiograph (A) and axial computed tomography (CT) image (B) showing tubular opacities with consolidation in the right lung suggestive of mucocoeles along with cystic bronchiectasis in bilateral lungs. Coronal CT image (C) of the patient showing evidence of bilateral allergic fungal rhinosinusitis.

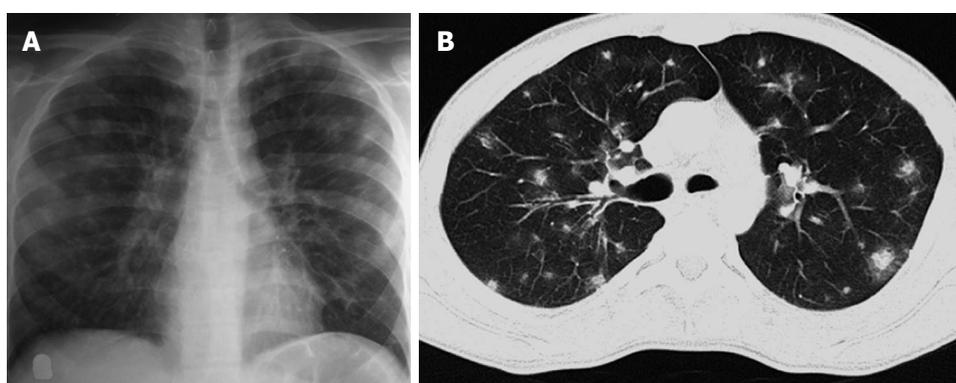


Figure 5 A 17-year-old boy with acute lymphocytic leukemia along with angioinvasive aspergillosis. Chest radiograph (A) showing multiple fluffy nodules in bilateral lung fields. High resolution CT image (B) of the same patient shows multiple nodules with surrounding ground glass opacity (halo sign).

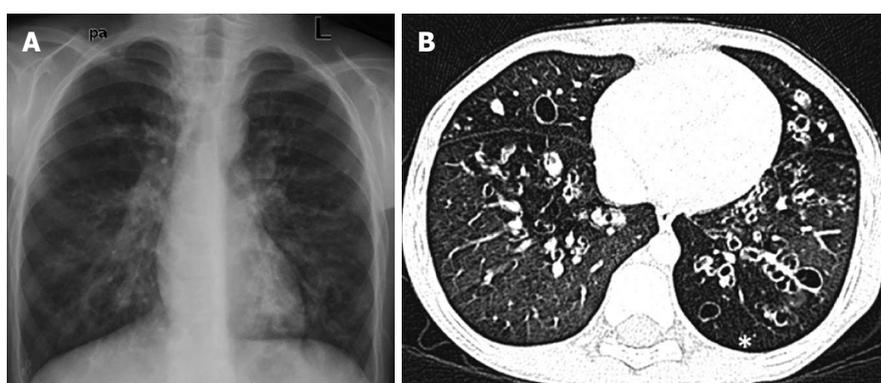


Figure 6 A 10-year-old boy with post-infectious bronchiectasis. Frontal chest radiograph (A) showing multiple cystic lucencies and tubular opacities in both lungs. Chest high resolution CT image (B) shows multiple areas of cystic bronchiectasis with associated air trapping (asterisk).

Aspiration tends to involve the lower lobes (right > left). Cystic fibrosis shows lung hyperinflation and interstitial infiltrates with upper lobe preponderance (Figure 7). Bronchiectasis due to ciliary dyskinesias has a lower lobe predisposition^[27].

Congenital heart disease

Hemoptysis can occur in patients with congenital heart diseases associated with pulmonary artery or venous stenosis or atresia. This is attributed to hemorrhage from enlarged, tortuous aorto-pulmonary collateral arteries and thrombotic

lesions in the small pulmonary arteries^[28]. Chest radiography may show cardiomegaly with abnormality in cardiac silhouette and a small hilum. An abnormal vascular channel parallel to the right cardiac border (scimitar vein) can be seen in pulmonary venobar hypoplasia^[29]. There may be associated pulmonary volume loss. MDCT is the modality of choice to demonstrate the site and extent of pulmonary artery narrowing and delineate anomalous pulmonary venous drainage (Figure 8). It exquisitely depicts the various aorto-pulmonary collaterals. Other associated cardiac anomalies can also be evaluated^[30].

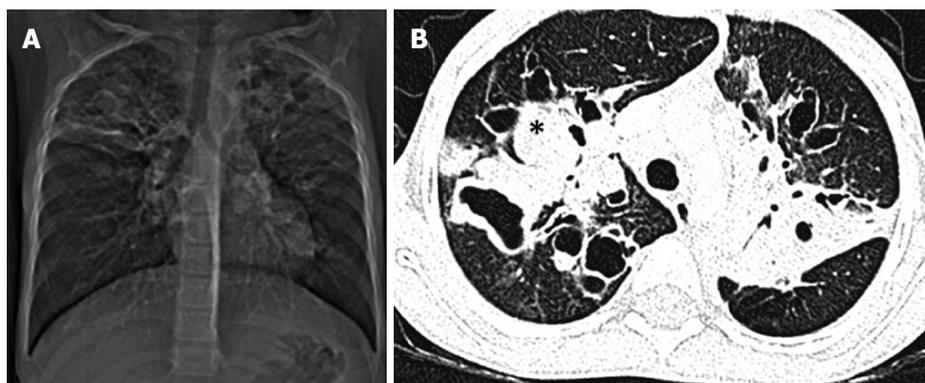


Figure 7 A 7-year-old boy with cystic fibrosis. Computed tomography (CT) scout image (A) and axial CT chest image (B) showing bilateral upper lobe bronchiectasis with bronchocele formation (asterisk) due to mucous plugging and sparing of lower zones.

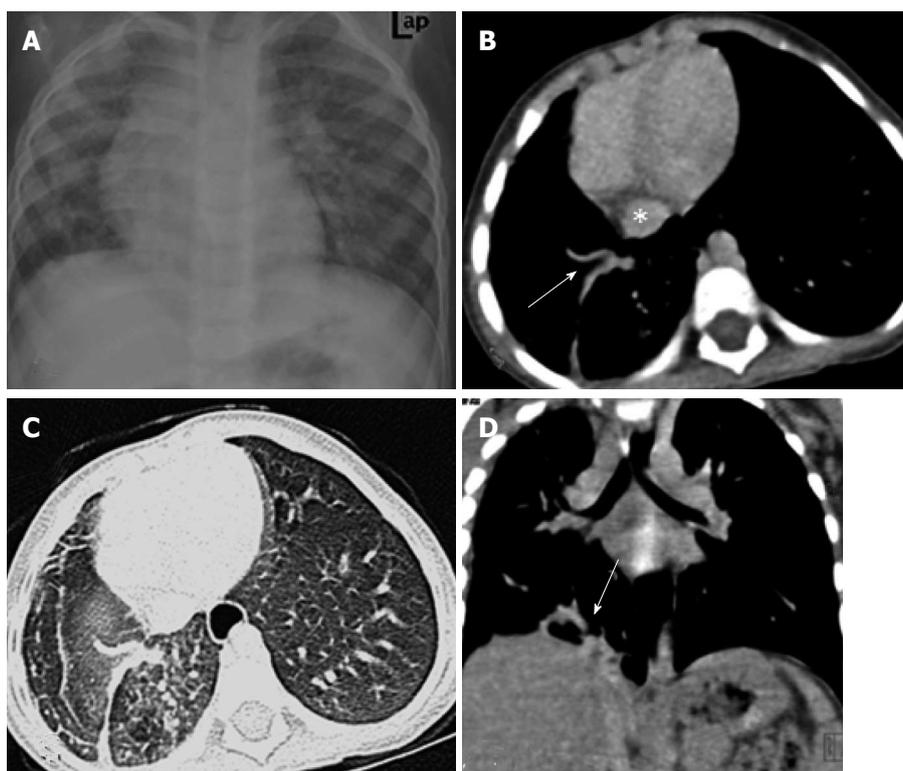


Figure 8 Pulmonary venolobar hypoplasia. Chest radiograph (A) shows volume loss of the right hemithorax with ipsilateral mediastinal shift. Contrast-enhanced computed tomography images (B-D) showing anomalous right inferior pulmonary vein (arrows) coursing inferiorly towards the inferior vena cava (asterisk).

Pulmonary artery narrowing

Chronic pulmonary artery narrowing can occur due to a variety of causes like infections, inflammation and thromboembolism^[31]. Infections are the most common cause. Narrowing of the pulmonary artery can be caused in the setting of infection by mediastinal lymphadenopathy or fibrosis. Fibrosis may be focal or diffuse. CT finding of focal fibrosis is a calcified soft-tissue mass in the paratracheal and hilar location. It can occur secondary to tuberculosis in the developing countries and histoplasmosis in United States. The diffuse form manifests as an infiltrative, non-calcified soft-tissue mass extending into multiple mediastinal compartments. It can be associated with autoimmune disorders, drugs, or be idiopathic^[32]. Pulmonary artery narrowing in these cases leads to pulmonary hypoperfusion and consequent bronchial artery hypertrophy while can lead to hemoptysis of varying severity. CT pulmonary angiography is the investigation of choice in this condition as it elucidates pulmonary artery

narrowing and bronchial/systemic artery hypertrophy^[33].

Pulmonary arteriovenous malformations

Pulmonary arteriovenous malformations (PAVM) are direct communication between the branches of the pulmonary artery and veins without capillary bed. There is a strong association between PAVM and hereditary hemorrhagic telangiectasia^[34]. Chest radiography is an important tool for diagnosis and follow-up. Classic findings of PAVM are a round or oval well-defined mass, frequently lobulated, ranging in size from 1-5 cm. Two-thirds of these are located in the lower lobe. A connecting vessel may be seen radiating from the hilum. MDCT can identify the PAVM and connecting vessels more accurately (Figure 9). PAVMs have rapid blood flow and hence produce a low intensity signal on MRI. Catheter angiography remains the gold standard in diagnosis of PAVM as it defines the angio-architecture which is necessary before therapeutic embolization or surgical resection.

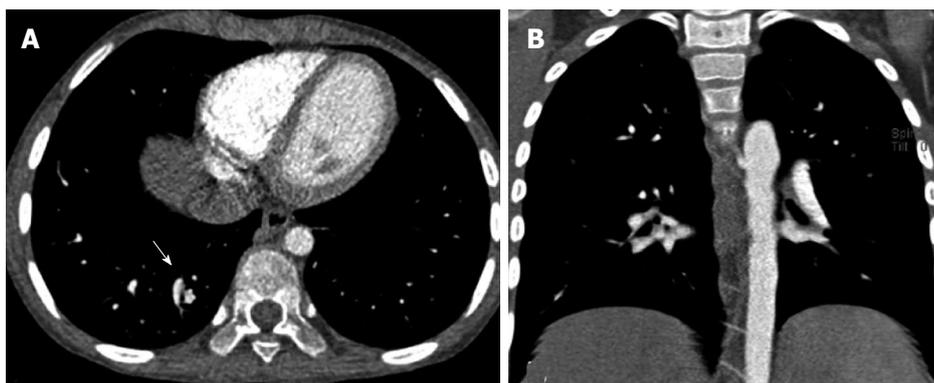


Figure 9 Pulmonary arteriovenous malformation. Axial (A) and coronal (B) computed tomography images showing abnormal communication between branches of the pulmonary artery and vein in right lower lobe (arrows).

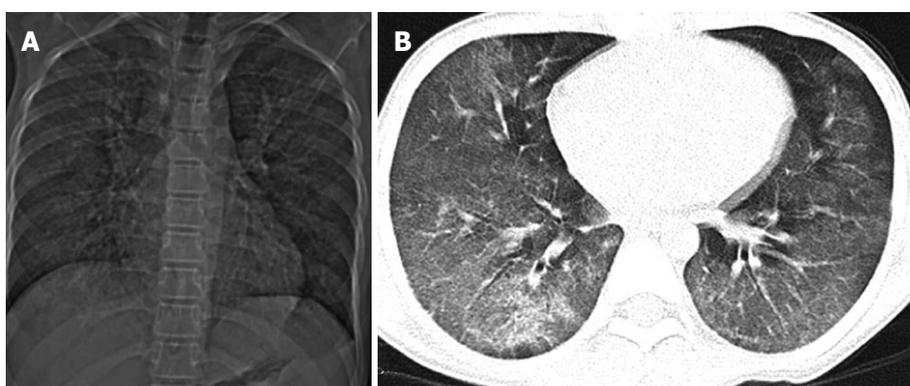


Figure 10 A 15-year-old boy with idiopathic pulmonary hemosiderosis. Scout computed tomography (CT) image (A) and axial CT image (B) showing diffuse ground glass opacity in bilateral lungs.

Idiopathic pulmonary hemosiderosis

Idiopathic pulmonary hemosiderosis (IPH) is a rare pulmonary disorder which manifests as a triad of hemoptysis, anemia and diffuse parenchymal infiltrates on chest radiographs^[35]. Diagnosis is confirmed by detection of hemosiderin-laden macrophages in broncho-alveolar lavage fluid, sputum or gastric aspirate. Secondary hemosiderosis is associated with systemic vasculitis, bleeding disorders and cardiac disease. Imaging findings are non-specific and need to be correlated with clinical and laboratory data to arrive at a diagnosis of IPH. Chest radiographs may reveal symmetric diffuse or patchy alveolar shadows sparing lung apices, which can clear on follow-up imaging. CT can show diffuse or patchy ground-glass opacity (Figure 10). There can be interstitial thickening in some cases^[36].

Foreign body

Foreign body aspiration can be a cause of hemoptysis primarily in patients less than 3 years of age. The aspirated foreign body can be visualized on a radiograph if it is radio-opaque. Associated radiographic findings include non-specific infiltrates, atelectasis, areas of hyperinflation, parenchymal consolidation or bronchiectasis (Figure 11A). Chest radiographs can be normal in 30% cases.

MPR and endoluminal virtual bronchoscopic images derived from MDCT can delineate the shape, location and volume of a foreign body. It can reveal associated pulmonary parenchymal changes (Figure 11B and C). Thus, imaging can help the surgeon plan the bronchoscopy for safe removal of foreign body^[37].

Neoplasm

Bronchial neoplasms are a rare cause of hemoptysis (Figure 12). Bronchial carcinoid tumors are the most frequent primary pulmonary neoplasms of childhood. The lesion can be central or peripheral. Radiological findings include hilar or perihilar masses with lobulated margins and associated obstructive changes (atelectasis, consolidation, bronchocele or hyperinflation)^[38]. On CT, carcinoid is seen as a well-defined, centrally located mass that narrows or deforms the airway and contains diffuse or punctuate calcification. It shows intense homogenous contrast enhancement. However, all carcinoids do not enhance. There can be associated pulmonary obstructive changes and mediastinal/hilar lymphadenopathy^[39].

CONCLUSION

Hemoptysis is a distressing symptom for the child, the

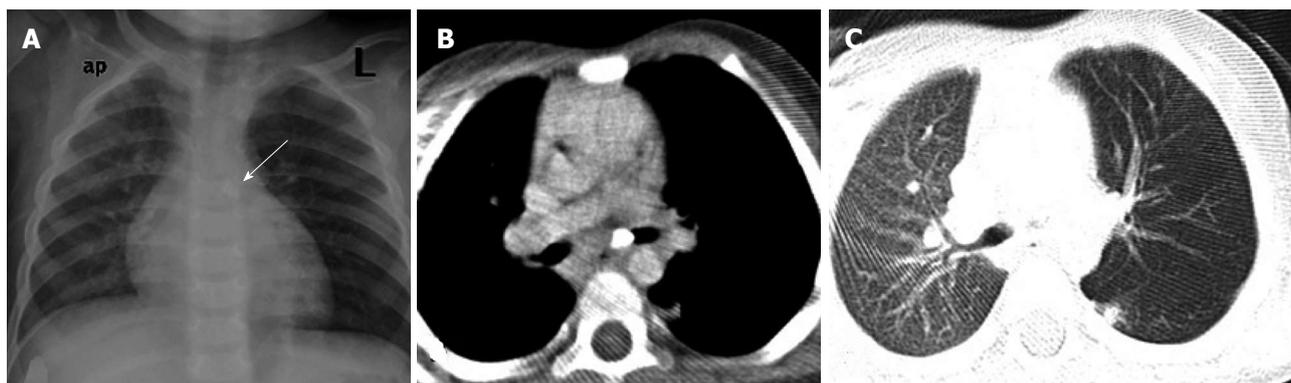


Figure 11 Foreign body aspiration. Chest radiograph (A) shows a radio-opaque foreign body in the left main bronchus (arrow) with hyperinflation of the left lung. Axial computed tomography images (B, C) delineate the morphology of the foreign body in the left main bronchus causing luminal compromise. There is associated air trapping in the left lung with patchy consolidation in the apical segment of the lower lobe.



Figure 12 Bronchial carcinoid. Scout computed tomography (CT) image (A) revealing non-visualization of the right main bronchus with volume loss and opacification of the right hemithorax along with bronchiectasis in right lower zone. Axial CT image (B) shows a mass in the right lung with mediastinal infiltration. Coronal Minimum Intensity Projection image (C) shows the outline of the mass projecting in the right main bronchus along with bronchiectasis in the right lower lobe.

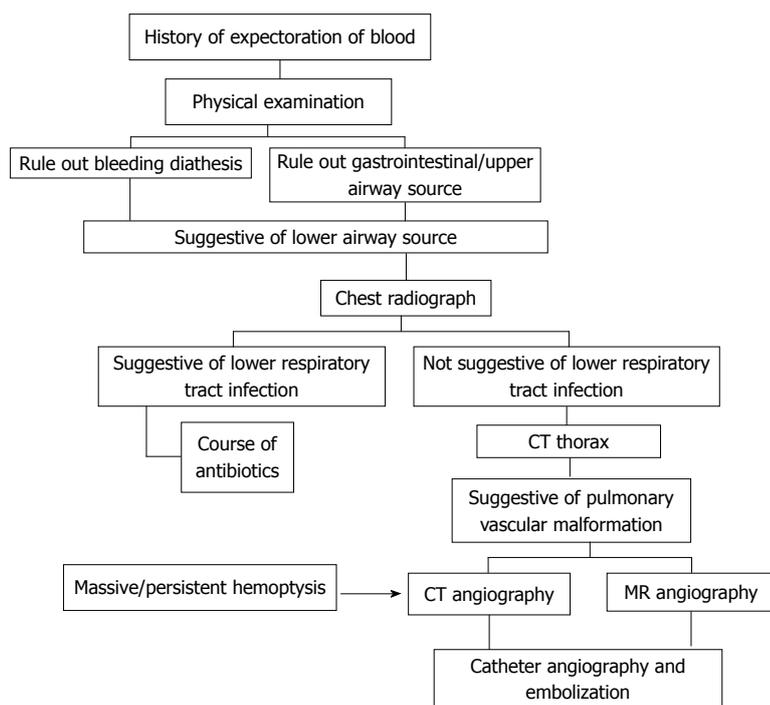


Figure 13 Flowchart depicting approach to a child presenting with hemoptysis. MR: Magnetic resonance; CT: Computed tomography.

family and the pediatrician. It poses a diagnostic challenge. Once the presence of hemoptysis has been ascertained, one needs to identify the source of bleeding and primary underlying cause. Acute lower respiratory tract infections are the most common cause of pediatric hemoptysis. The imaging modalities available for the work-up of hemoptysis include chest radiography, MDCT, MRI and catheter angiography. Chest radiographs may be normal in 30% cases. MDCT is a rapid, non-invasive multiplanar imaging modality which should be tailored to keep radiation dose to a minimum for optimal evaluation of hemoptysis in pediatric patients. MRI and catheter angiography have selected application. The use of the various imaging tools available is determined by the clinical presentation and the possible etiology (Figure 13). Maximum diagnostic and therapeutic benefit can be attained by the judicious use of imaging modalities in a child presenting with hemoptysis.

REFERENCES

- 1 **Fraser RS**, Pare P, Pare PD. Hemoptysis. In: Fraser RS, Pare P, Pare PD. Diseases of the chest. Philadelphia, Pa: Saunders, 1988: 394–396
- 2 **Knott-Craig CJ**, Oosthuizen JG, Rossouw G, Joubert JR, Barnard PM. Management and prognosis of massive hemoptysis. Recent experience with 120 patients. *J Thorac Cardiovasc Surg* 1993; **105**: 394-397 [PMID: 8445918]
- 3 **Turcios NL**, Vega M. The child with hemoptysis. *Hosp Pract (Off Ed)* 1987; **22**: 214, 217-218 [PMID: 3116012]
- 4 **Sim J**, Kim H, Lee H, Ahn K, Lee SI. Etiology of hemoptysis in children: a single institutional series of 40 cases. *Allergy Asthma Immunol Res* 2009; **1**: 41-44 [PMID: 20224669 DOI: 10.4168/aaair.2009.1.1.41]
- 5 **Pump KK**. The bronchial arteries and their anastomoses in the human lung. *Dis Chest* 1963; **43**: 245-255 [PMID: 13972526 DOI: 10.1378/chest.43.3.245]
- 6 **Deffebach ME**, Charan NB, Lakshminarayan S, Butler J. The bronchial circulation. Small, but a vital attribute of the lung. *Am Rev Respir Dis* 1987; **135**: 463-481 [PMID: 3544986]
- 7 **Stern RC**, Wood RE, Boat TF, Matthews LW, Tucker AS, Doershuk CF. Treatment and prognosis of massive hemoptysis in cystic fibrosis. *Am Rev Respir Dis* 1978; **117**: 825-828 [PMID: 655488]
- 8 **Dearborn DG**. Pulmonary hemorrhage in infants and children. *Curr Opin Pediatr* 1997; **9**: 219-224 [PMID: 9229159 DOI: 10.1097/00008480-199706000-00005]
- 9 **Lynch T**, Gouin S, Larson C, Patenaude Y. Does the lateral chest radiograph help pediatric emergency physicians diagnose pneumonia? A randomized clinical trial. *Acad Emerg Med* 2004; **11**: 625-629 [PMID: 15175199]
- 10 **Stankiewicz JA**, Puczynski M, Lynch JM. Embolization in the treatment of massive hemoptysis in patients with cystic fibrosis. *Ear Nose Throat J* 1985; **64**: 180-184 [PMID: 3996265]
- 11 **Sundarakumar DK**, Bhalla AS, Sharma R, Hari S, Guleria R, Khilnani GC. Multidetector CT evaluation of central airways stenoses: Comparison of virtual bronchoscopy, minimal-inversion projection, and multiplanar reformatted images. *Indian J Radiol Imaging* 2011; **21**: 191-194 [PMID: 22013293 DOI: 10.4103/0971-3026.85366]
- 12 **Ferebee SH**, Mount FW. Chemotherapy of tuberculosis, progress and promise. *Public Health Rep* 1957; **72**: 412-420 [PMID: 13432111 DOI: 10.1148/rg.226015180]
- 13 **Furuse M**, Saito K, Kunieda E, Aihara T, Touei H, Ohara T, Fukushima K. Bronchial arteries: CT demonstration with arteriographic correlation. *Radiology* 1987; **162**: 393-398 [PMID: 3797652]
- 14 **Cauldwell EW**, Siekert RG. The bronchial arteries; an anatomic study of 150 human cadavers. *Surg Gynecol Obstet* 1948; **86**: 395-412 [PMID: 18905113]
- 15 **Do KH**, Goo JM, Im JG, Kim KW, Chung JW, Park JH. Systemic arterial supply to the lungs in adults: spiral CT findings. *Radiographics* 2001; **21**: 387-402 [PMID: 11259703]
- 16 **Swanson KL**, Johnson CM, Prakash UB, McKusick MA, Andrews JC, Stanson AW. Bronchial artery embolization: experience with 54 patients. *Chest* 2002; **121**: 789-795 [PMID: 11888961 DOI: 10.1378/chest.121.3.789]
- 17 **Yoon W**, Kim YH, Kim JK, Kim YC, Park JG, Kang HK. Massive hemoptysis: prediction of nonbronchial systemic arterial supply with chest CT. *Radiology* 2003; **227**: 232-238 [PMID: 12601194 DOI: 10.1148/radiol.2271020324]
- 18 **Emami K**, Stephen M, Kadlecsek S, Cadman RV, Ishii M, Rizi RR. Quantitative assessment of lung using hyperpolarized magnetic resonance imaging. *Proc Am Thorac Soc* 2009; **6**: 431-438 [PMID: 19687215 DOI: 10.1513/pats.200902-008AW]
- 19 **Panth S**, Fain S, Holmes J, Fuller S, Korosec F, Grist T. Assessment of lung ventilation, gas trapping and pulmonary perfusion in patients with asthma during inhaled corticosteroid withdrawal. Proceedings of the 12th Annual Meeting of ISMRM, Kyoto, Japan, 2004 (Abstract 764)
- 20 **McMahon CJ**, Dodd JD, Hill C, Woodhouse N, Wild JM, Fischele S, Gallagher CG, Skehan SJ, van Beek EJ, Masterson JB. Hyperpolarized 3helium magnetic resonance ventilation imaging of the lung in cystic fibrosis: comparison with high resolution CT and spirometry. *Eur Radiol* 2006; **16**: 2483-2490 [PMID: 16871384 DOI: 10.1007/s00330-006-0311-5]
- 21 **Koumellis P**, van Beek EJ, Woodhouse N, Fischele S, Swift AJ, Paley MN, Hill C, Taylor CJ, Wild JM. Quantitative analysis of regional airways obstruction using dynamic hyperpolarized 3He MRI-preliminary results in children with cystic fibrosis. *J Magn Reson Imaging* 2005; **22**: 420-426 [PMID: 16104046 DOI: 10.1002/jmri.20402]
- 22 **Wong KS**, Wang CR, Lin TY. Hemoptysis in children. *Changcheng Yixue Zazhi* 1998; **21**: 57-62 [PMID: 9607265]
- 23 **Aquino SL**, Kee ST, Warnock ML, Gamsu G. Pulmonary aspergillosis: imaging findings with pathologic correlation. *AJR Am J Roentgenol* 1994; **163**: 811-815 [PMID: 8092014]
- 24 **Logan PM**, Müller NL. CT manifestations of pulmonary aspergillosis. *Crit Rev Diagn Imaging* 1996; **37**: 1-37 [PMID: 8744521]
- 25 **Logan PM**, Primack SL, Miller RR, Müller NL. Invasive aspergillosis of the airways: radiographic, CT, and pathologic findings. *Radiology* 1994; **193**: 383-388 [PMID: 7972747]
- 26 **Franquet T**, Müller NL, Giménez A, Gueembe P, de La Torre J, Bagué S. Spectrum of pulmonary aspergillosis: histologic, clinical, and radiologic findings. *Radiographics* 2001; **21**: 825-837 [PMID: 11452056]
- 27 **Cantin L**, Bankier AA, Eisenberg RL. Bronchiectasis. *AJR Am J Roentgenol* 2009; **193**: W158-W171 [PMID: 19696251 DOI: 10.2214/AJR.09.3053]
- 28 **Haroutunian LM**, Neill CA. Pulmonary complications of congenital heart disease: hemoptysis. *Am Heart J* 1972; **84**: 540-559 [PMID: 4672656 DOI: 10.1016/0002-8703(72)90479-6]
- 29 **Ferguson EC**, Krishnamurthy R, Oldham SA. Classic imaging signs of congenital cardiovascular abnormalities. *Radiographics* 2007; **27**: 1323-1334 [PMID: 17848694 DOI: 10.1148/rg.275065148]
- 30 **Gilkeson RC**, Ciancibello L, Zahka K. Pictorial essay. Multidetector CT evaluation of congenital heart disease in pediatric and adult patients. *AJR Am J Roentgenol* 2003; **180**: 973-980 [PMID: 12646439 DOI: 10.2214/ajr.180.4.1800973]
- 31 **Castañer E**, Gallardo X, Rimola J, Pallardó Y, Mata JM, Perendreu J, Martin C, Gil D. Congenital and acquired pulmonary artery anomalies in the adult: radiologic overview. *Radiographics* 2006; **26**: 349-371 [PMID: 16549603 DOI: 10.1148/rg.262055092]

- 32 **Rossi SE**, McAdams HP, Rosado-de-Christenson ML, Franks TJ, Galvin JR. Fibrosing mediastinitis. *Radiographics* 2001; **21**: 737-757 [PMID: 11353121]
- 33 **Bhalla AS**, Gupta P, Mukund A, Kabra SK, Kumar A. Pulmonary artery narrowing: A less known cause for massive hemoptysis. *Oman Med J* 2013; **28**: 43-46 [DOI: 10.5001/omj.2013.43]
- 34 **Khurshid I**, Downie GH. Pulmonary arteriovenous malformation. *Postgrad Med J* 2002; **78**: 191-197 [PMID: 11930021 DOI: 10.1136/pmj.78.918.191]
- 35 **Rezkalla MA**, Simmons JL. Idiopathic pulmonary hemosiderosis and alveolar hemorrhage syndrome: case report and review of the literature. *S D J Med* 1995; **48**: 79-85 [PMID: 7740300]
- 36 **Kabra SK**, Bhargava S, Lodha R, Satyavani A, Walia M. Idiopathic pulmonary hemosiderosis: clinical profile and follow up of 26 children. *Indian Pediatr* 2007; **44**: 333-338 [PMID: 17536132]
- 37 **Bai W**, Zhou X, Gao X, Shao C, Califano JA, Ha PK. Value of chest CT in the diagnosis and management of tracheo-bronchial foreign bodies. *Pediatr Int* 2011; **53**: 515-518 [PMID: 21129123 DOI: 10.1111/j.1442-200X.2010.03299.x]
- 38 **Nessi R**, Basso Ricci P, Basso Ricci S, Bosco M, Blanc M, Uslenghi C. Bronchial carcinoid tumors: radiologic observations in 49 cases. *J Thorac Imaging* 1991; **6**: 47-53 [PMID: 1649924]
- 39 **Jeung MY**, Gasser B, Gangi A, Charneau D, Ducroq X, Kessler R, Quoix E, Roy C. Bronchial carcinoid tumors of the thorax: spectrum of radiologic findings. *Radiographics* 2002; **22**: 351-365 [PMID: 11896225]

P- Reviewers: Gow KW, Sijens PE **S- Editor:** Zhai HH
L- Editor: Wang TQ **E- Editor:** Lu YJ



Tramadol use in pediatric sickle cell disease patients with vaso-occlusive crisis

Mary P Borgerding, Randall K Absher, Tsz-Yin So

Mary P Borgerding, Randall K Absher, Department of Pharmacy, Wesley Long Hospital, Greensboro, NC 27401, United States
Tsz-Yin So, Department of Pharmacy, Moses H Cone Memorial Hospital, Greensboro, NC 27401, United States

Author contributions: Borgerding MP performed the majority of the research; Absher RK helped with the statistical analysis of the data; So TY helped with the design of the study and edited the manuscript.

Correspondence to: Tsz-Yin So, PhD, BCPS, Department of Pharmacy, Moses H. Cone Memorial Hospital, 1200 N. Elm St., Greensboro, NC 27401, United States. jeremy.so@conehealth.com
Telephone: +1-336-8327287

Received: March 28, 2013 Revised: May 30, 2013

Accepted: June 28, 2013

Published online: November 8, 2013

Abstract

AIM: To evaluate whether the addition of scheduled oral tramadol to intravenous morphine and intravenous ketorolac reduces morphine requirements.

METHODS: This single-centered, Institutional Review Board-approved, retrospective study at Moses Cone Memorial Hospital included pediatric patients who were ≥ 2 years old with vaso-occlusive crisis (VOC) caused by sickle cell disease (SCD), were on morphine patient-controlled analgesia (PCA), and had scheduled oral tramadol added to their standard pain regimen. The study population was admitted between March 2008 and March 2011. The data was collected from electronic records and included age, weight, morphine use, tramadol use, hemoglobin, pain scores, number of days on PCA, length of hospital stay, respiratory rate, and polyethylene glycol use. Thirty patients were analyzed as independent admissions and seven patients as paired admissions.

RESULTS: Eighteen pediatric SCD patients with VOC received morphine PCA and intravenous ketorolac and

twelve patients received morphine PCA and intravenous ketorolac and scheduled oral tramadol. Baseline characteristics were similar between both groups with the exception of the average weight, which was greater in the tramadol group than in the morphine group. The average morphine requirements in patients with and without the use of tramadol were similar, both for the independent admissions [0.58 mg/kg per day vs 0.65 mg/kg per day ($P = 0.31$)] and the paired admissions [0.71 mg/kg per day vs 0.77 mg/kg per day ($P = 0.5$)]. The daily polyethylene glycol requirement was less in the tramadol group for both the independent [0.5 g/kg per day vs 0.6 g/kg per day ($P = 0.64$)] and paired admissions analyses [and 0.41 g/kg per day vs 0.55 g/kg per day ($P = 0.67$)].

CONCLUSION: The addition of scheduled tramadol in patients receiving concomitant morphine and ketorolac demonstrates a trend toward decreased morphine and polyethylene glycol use.

© 2013 Baishideng. All rights reserved.

Key words: Pediatrics; Sickle cell; Tramadol; Morphine; Vaso-occlusive crisis

Core tip: A small clinical study has shown that balanced analgesia using intravenous morphine, intravenous ketorolac, and intravenous tramadol followed by erythrocytapheresis was effective, as shown by pain relief and significant improvement in mood and sleep, in seven sickle cell disease patients aged three to twenty-eight years who presented with vaso-occlusive crisis. The objective of this study is to evaluate whether the addition of scheduled oral tramadol to intravenous morphine plus intravenous ketorolac provides adequate pain relief, and reduces morphine requirements, adverse effects, length of patient-controlled analgesia therapy, and length of hospital stay.

Borgerding MP, Absher RK, So TY. Tramadol use in pediatric

sickle cell disease patients with vaso-occlusive crisis. *World J Clin Pediatr* 2013; 2(4): 65-69 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/65.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.65>

INTRODUCTION

One of the major causes of hospitalization for patients with sickle cell disease (SCD) is vaso-occlusive crises (VOC). The hallmark characteristics of VOC include organ damage and pain due to the presence of dense red blood cells. The pain is generated through multiple pathways including somatic, neuropathic, and vascular mechanisms^[1-4].

Balanced analgesia is a strategy based on the co-administration of drugs with different pharmacological mechanisms in order to control pain at different origins, improve the efficacy of treatment, and reduce adverse effects of each drug^[5]. The administration of ketorolac and tramadol has been proven as a form of effective balanced analgesia, particularly for post-operative pain and pain caused by trauma^[6]. Tramadol and its active metabolite (M1) work by binding to the mu-opioid receptors in the central nervous system (CNS) and inhibiting the reuptake of norepinephrine and serotonin, causing inhibition of the ascending pain pathway and altering the perception of and response to pain^[7]. Tramadol is clinically known to have a better safety profile than the other major opioids, causing less respiratory depression and constipation.

In 2005, de Franceschi *et al.*^[8] published a study evaluating balanced analgesia using intravenous (*iv*) morphine, *iv* ketorolac, and *iv* tramadol followed by erythrocytapheresis in seven SCD patients aged three to twenty-eight years who presented with VOC. The co-administration of tramadol and ketorolac was effective in all VOC, as shown by pain relief and significant improvement in mood and sleep. The use of erythrocytapheresis, which is not available at our hospital, Moses Cone Memorial Hospital (MCMH), likely contributed to pain relief in this study.

At MCMH pediatric patients who presented prior to August 2010 with VOC caused by SCD were routinely prescribed *iv* morphine and *iv* ketorolac for pain control. However, because of the high morphine requirement in this patient population, severe respiratory depression and constipation can occur. After August 2010, pediatric physicians began adding scheduled oral (*po*) tramadol to the standard regimen of *iv* morphine patient-controlled analgesia (PCA) and *iv* ketorolac in an attempt to reduce narcotic-induced side effects. The objective of this retrospective study is to evaluate whether the addition of scheduled *po* tramadol to *iv* morphine and *iv* ketorolac reduces morphine requirements, provides adequate pain relief, decreases length of hospital stay, and reduces severe respiratory depression, severe constipation, and length of PCA therapy.

MATERIALS AND METHODS

This single-centered, Institutional Review Board (IRB)-approved, retrospective study included pediatric patients who were ≥ 2 years old with VOC caused by SCD, were on PCA morphine and *iv* ketorolac and had scheduled *po* tramadol added to their regimen. Tramadol was dosed at 1-2 mg/kg per dose *po* every four to 6 h (max: 400 mg/d and 100 mg/dose) and ketorolac at 0.5 mg/kg per dose *iv* every 6 h (Max: 30 mg/dose). Morphine PCA orders included a basal rate, intermittent dose, lockout interval, and a 1-hour and 4-hour limit. Using the International Classification of Diseases (ICD)-9 code for SCD, all patients < 21 years old who were admitted between March 2008 and March 2011 were included in this retrospective review. Patients were excluded from the review if they did not have a diagnosis of VOC or did not receive morphine PCA. The data was collected from electronic records and included age, weight, morphine use, tramadol use, hemoglobin, pain scores, number of days on PCA, length of hospital stay, respiratory rate, and polyethylene glycol (PEG) use. All patients were analyzed as independent admissions. Additionally, patients with multiple admissions during the study period (at least one with morphine only and one with both morphine and tramadol) were analyzed as paired admissions, acting as their own controls.

The primary outcome of this study was average daily morphine requirement. Secondary outcomes included average pain scores, respiratory rate, PEG dose, length of PCA therapy and number of days in the hospital.

All patients were analyzed as independent admissions using the Wilcoxon Rank Sum test. Patients who had multiple admissions, one with tramadol use and one without were also analyzed as paired admissions using the Wilcoxon Signed Rank test. The statistical analysis was completed using Stata, Version 10.1 (Cary, NC).

RESULTS

Between March 2008 and March 2011 eighteen pediatric SCD patients with VOC received morphine PCA and *iv* ketorolac and twelve patients received morphine PCA plus *iv* ketorolac and scheduled *po* tramadol. Baseline characteristics were similar between both groups with the exception of the average weight, which was greater in the tramadol group than in the morphine group because the latter group had a younger sample (Table 1).

Average morphine requirements with and without tramadol were similar in the independent admission analysis [0.58 mg/kg per day and 0.65 mg/kg per day, respectively ($P = 0.31$)]. Average morphine requirements with and without tramadol were also similar in the paired admissions analysis [0.71 mg/kg per day and 0.77 mg/kg per day, respectively ($P = 0.5$)]. Contradictory to what was expected, pain scores were higher when tramadol was added to the pain regimen for both the independent admissions (6.75 *vs* 5) and the paired admissions (6.5 *vs* 5.5). The daily

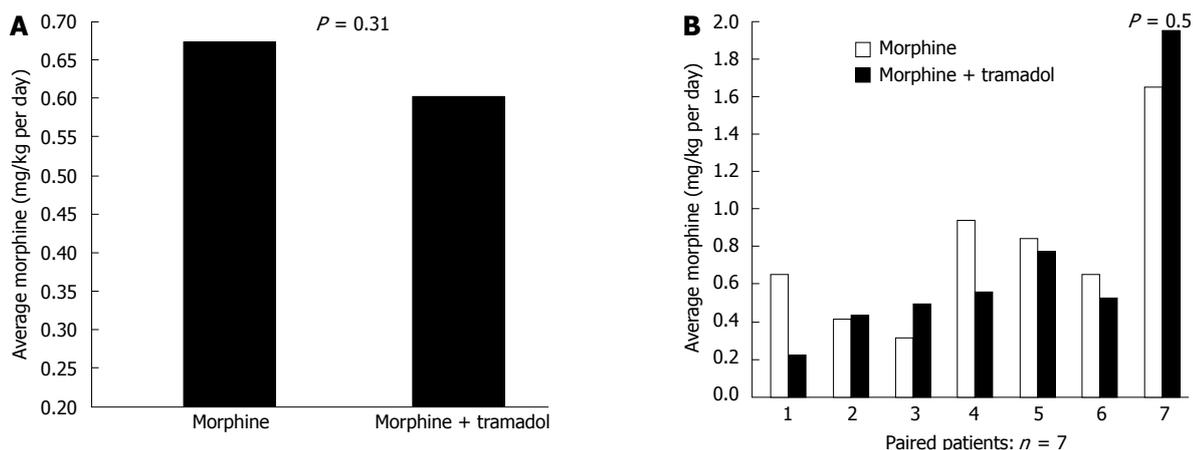


Figure 1 Average morphine requirement for independent admissions (A) and average morphine requirement for paired admissions (B).

Table 1 Baseline characteristics		
	Morphine	Morphine + tramadol
Sample size	18	12
Age (yr), Mean (range)	11.5 (3-20)	13.4 (7-20)
Gender		
Male	9	6
Female	9	6
Weight (kg), mean (range)	38.7 (11.6-77.4)	52 (27.0-77.4) ^a
Hemoglobin (mg/dL), mean (range)	9.0 (6.0-11.9)	9.4 (6.5-12.0)

^a $P < 0.05$ vs morphine.

polyethylene glycol requirement was less in the tramadol group for both the independent and paired admissions analyses (0.5 g/kg per day vs 0.6 g/kg per day and 0.41 g/kg per day vs 0.55 g/kg per day, respectively) but neither difference was statistically significant ($P = 0.64$ and 0.67 , respectively). The paired admissions analysis demonstrated a greater difference in PEG requirements which, while not statistically significant, may provide a more accurate comparison. Furthermore, there was no difference in the length of stay, number of days on PCA, or respiratory rate between groups in either analysis (Tables 2 and 3).

DISCUSSION

The addition of scheduled oral tramadol in patients receiving concomitant morphine and ketorolac did result in numerically lower average daily morphine requirements (Figure 1A) and polyethylene glycol use; however, differences in these endpoints were not statistically significant (Table 2). In the paired admissions analysis, four of the six patients who received less than 1 mg/kg per day of morphine used less morphine when tramadol was added to their regimen (Figure 1B); however, pain scores did not correlate with the decreased morphine requirement.

The lack of correlation between pain scores and morphine requirement may be due to the subjectivity of pain. After completion of the study, a pediatric psychiatrist

spoke with several patients in the study and found that many patients did not understand how to use the visual analogue pain scale or the numeric scale to rate their pain. When asked to rate their pain as red, yellow or green (red corresponding to the most pain and green the least), patients gave more accurate representations of their true pain level. Extensive education on rating pain would be required to provide a more precise representation of pain relief with and without the use of tramadol.

There were several limitations to this study. The retrospective nature of the study forced us to rely on electronic nursing documentation for all of our data collection, which may have resulted in some inaccurately charted data. Frequently, bowel movements were not documented. We therefore measured average daily polyethylene glycol use to assess constipation. Additionally, we were unable to stratify patients based on their basal morphine PCA rate and determine how much of their daily morphine requirement was due to demand dosing, as this information was not documented electronically. Patients with multiple admissions for VOC can develop tolerance to narcotics, resulting in an increased morphine basal rate requirement. Documentation of this may have provided a more accurate assessment of which analgesic regimen provided better pain control and fewer narcotic-induced side effects.

VOC most commonly involves the back, legs, knees, arms, chest and abdomen. The location of the vaso-occlusive crisis has a significant impact on the intensity of pain and the ability to control that pain; however, this study did not stratify patients based on VOC location or disease severity^[9]. Additionally, the disease severity has interpatient and inpatient variability making it more difficult to compare patients. Also, one patient in this study had drug-seeking behavior, which may have skewed the results causing increased average morphine requirement and pain scores.

A larger, controlled study would be more likely to determine statistical difference in the primary and secondary endpoints. Pediatric physicians at MCMH no longer routinely prescribe tramadol in this population but con-

Table 2 Independent admissions statistical analysis

	Morphine (<i>n</i> = 18)	Morphine + tramadol (<i>n</i> = 12)	<i>P</i> -value ¹
Morphine requirement (mg/kg per day), mean ± SD	0.65 ± 0.35	0.58 ± 0.48	0.31
Pain score, median (range)	5 (0-8)	6.75 (3-9)	0.07
PCA duration (d), median (range)	4.5 (0-13)	5 (3-14)	0.33
LOS (d), median (range)	7 (4-14)	7.5 (3-17)	0.85
Respiratory rate, mean (range)	21 (14-29)	19 (16-23)	0.28
Polyethylene glycol dose (gm/kg per day), mean (range)	0.6 (0-1.66)	0.5 (0-1.12)	0.64

¹*P*-value calculated using Wilcoxon Rank Sum test. PCA: Patient-controlled analgesia; LOS: Length of stay.

Table 3 Paired admissions statistical analysis (*n* = 7)

	Morphine	Morphine + tramadol	<i>P</i> -value ¹
Morphine requirement (mg/kg per day), mean ± SD	0.77 ± 0.44	0.71 ± 0.57	0.50
Pain score, median (range)	5.5 (2-8)	6.5 (3-8)	0.27
PCA duration (d), median (range)	6 (4-13)	7 (3-10)	0.35
LOS (d), median (range)	7 (5-14)	9 (3-12)	0.61
Respiratory rate, mean (range)	19 (14-20)	19 (17-23)	0.87
Polyethylene glycol dose (gm/kg per day), mean (range)	0.55 (0.28-1.1)	0.41 (0.28-0.98)	0.67

¹*P*-value calculated using Wilcoxon Rank Sum test. PCA: Patient-controlled analgesia; LOS: Length of stay.

tinue to use it in patients who have clinically shown improved pain control with tramadol in the past. Tramadol use is appropriate in this population as it has proven safe, usually causing no additional side effects and potentially providing some benefit in controlling pain and reducing narcotic-induced constipation. If tramadol continues to be clinically beneficial for pain control in other patients in this population, it may be possible to review our primary and secondary endpoints on a larger scale to determine any true differences.

ACKNOWLEDGEMENTS

The authors would like to thank Dr. Ola Akintemi, Dr. Jackie Roh, Dr. Michelle Turner, and Dr. Alison Grimsley for their effort in evaluating the research. Without their help, this work would not have been accomplished.

COMMENTS

Background

Pain is a hallmark of vaso-occlusive crisis (VOC) caused by sickle-cell disease (SCD). Intravenous (*iv*) morphine plus *iv* ketorolac is generally the combination of choice for VOC in SCD patients at Moses Cone Memorial Hospital (MCMH); however, morphine can cause severe respiratory depression and constipation. The objective of this study is to evaluate whether the addition of scheduled oral tramadol to *iv* morphine plus *iv* ketorolac provides adequate pain relief, and reduces morphine requirements, adverse effects, length of PCA therapy, and length of hospital stay.

Research frontiers

In 2005, de Franceschi *et al* published a study evaluating balanced analgesia using *iv* morphine, *iv* ketorolac, and *iv* tramadol followed by erythrocytapheresis in seven SCD patients aged three to twenty-eight who presented with VOC. The co-administration of tramadol and ketorolac was effective in all VOC, as shown by pain relief and significant improvement in mood and sleep.

Innovations and breakthroughs

This is the first article evaluating the use of oral tramadol in addition to *iv* morphine and ketorolac on whether this combination provides adequate pain relief, and reduces morphine requirements, adverse effects, length of PCA therapy, and length of hospital stay.

Applications

The study results suggest that the addition of scheduled oral tramadol in patients receiving concomitant morphine and ketorolac demonstrates a trend toward decreased morphine and polyethylene glycol use.

Terminology

Erythrocytapheresis is a process in which red blood cells are extracted from the whole blood.

Peer review

This is a very good clinical study in which the authors analyzed the efficacy of adding oral tramadol to usual pain regimen used for sickle cell pain crisis.

REFERENCES

- 1 Platt OS, Thorington BD, Brambilla DJ, Milner PF, Rosse WF, Vichinsky E, Kinney TR. Pain in sickle cell disease. Rates and risk factors. *N Engl J Med* 1991; **325**: 11-16 [PMID: 1710777 DOI: 10.1056/NEJM199107043250103]
- 2 Cluster S, Vichinsky EP. Managing sickle cell disease. *BMJ* 2003; **327**: 1151-1155 [PMID: 14615343 DOI: 10.1136/bmj.327.7424.1151]
- 3 Rees DC, Olujuhunbe AD, Parker NE, Stephens AD, Telfer P, Wright J. Guidelines for the management of the acute painful crisis in sickle cell disease. *Br J Haematol* 2003; **120**: 744-752 [PMID: 12614204 DOI: 10.1046/j.1365-2141.2003.04193.x]
- 4 Benjamin L. Nature and treatment of the acute painful episodes in sickle cell disease. In: Steinberg MH, Forget BG, Higgs DR, Nagel RL, editors. Disorders of hemoglobin: genetics, pathophysiology and clinical management. Cambridge. United Kingdom: Cambridge University Press, 2001: 671-710
- 5 Kehlet H, Dahl JB. The value of "multimodal" or "balanced analgesia" in postoperative pain treatment. *Anesth Analg* 1993; **77**: 1048-1056 [PMID: 8105724 DOI: 10.1213/0000539-1

- 99311000-00030]
- 6 **Scott LJ**, Perry CM. Tramadol: a review of its use in perioperative pain. *Drugs* 2000; **60**: 139-176 [PMID: 10929933 DOI: 10.2165/00003495-200060010-00008]
- 7 Lexicomp 2011: Tramadol. [Lexi-Comp ONLINE web site]. Available from: URL: http://www.uptodate.com.libproxy.lib.unc.edu/contents/tramadol-pediatric-drug-information?detectedLanguage=en&source=search_result&search=tramadol&selectedTitle=2%7E86&provider=noPro
- 8 **de Franceschi L**, Finco G, Vassanelli A, Zaia B, Ischia S, Corrocher R. A pilot study on the efficacy of ketorolac plus tramadol infusion combined with erythrocytapheresis in the management of acute severe vaso-occlusive crises and sickle cell pain. *Haematologica* 2004; **89**: 1389-1391 [PMID: 15531461]
- 9 **Yale SH**, Nagib N, Guthrie T. Approach to the vaso-occlusive crisis in adults with sickle cell disease. *Am Fam Physician* 2000; **61**: 1349-1356, 1349-1356 [PMID: 10735342]

P- Reviewer: Al-Haggag M **S- Editor:** Wen LL
L- Editor: A **E- Editor:** Lu YJ



Pediatric vs adult pulmonary tuberculosis: A retrospective computed tomography study

Prasad Thotton Veedu, Ashu Seith Bhalla, Sreenivas Vishnubhatla, Sushil Kumar Kabra, Arundeeep Arora, Divya Singh, Arun Kumar Gupta

Prasad Thotton Veedu, Ashu Seith Bhalla, Arundeeep Arora, Divya Singh, Arun Kumar Gupta, Department of Radiodiagnosis, All India Institute of Medical Sciences, New Delhi 110029, India

Sreenivas Vishnubhatla, Department of Biostatistics, All India Institute of Medical Sciences, New Delhi 110029, India

Sushil Kumar Kabra, Department of Pediatrics, All India Institute of Medical Sciences, New Delhi 110029, India

Author contributions: Thotton Veedu P, Bhalla AS and Gupta AK were responsible for performing the radiological investigation and involved in image and data analysis; Kabra SK performed the clinical evaluation of the patients; Vishnubhatla S performed the statistical analysis; Arora A and Singh D were involved in analysis of imaging and manuscript preparation.

Correspondence to: Ashu Seith Bhalla, Additional Professor, Department of Radiodiagnosis, All India Institute of Medical Sciences, Ansari Nagar, New Delhi 110029, India. ashubhalla1@yahoo.com

Telephone: +91-11-265949258 Fax: +91-11-26588641

Received: July 28, 2013 Revised: September 25, 2013

Accepted: October 15, 2013

Published online: November 8, 2013

Abstract

AIM: To compare the manifestations of chest tuberculosis (TB) in pediatric and adult patients based on contrast enhanced computed tomography of chest.

METHODS: This was a retrospective study consisting of 152 patients of chest TB including 48 children and 104 adults who had undergone contrast enhanced computed tomography of chest prior to treatment. The patterns and severity of parenchymal, mediastinal and pleural manifestations were analyzed and compared among different age groups.

RESULTS: Parenchymal changes observed include consolidation, air space nodules, miliary TB, cavita-

tion, bronchiectasis and fibrosis and these were noted in 60% of children, 71% of adolescents and 76.9% of adults. These changes were more common in right upper lobe in all age groups. There was no significant difference in the frequency of these changes (except nodules) in different age groups. Centrilobular nodules were seen less commonly in children less than 10 years ($P = 0.028$). Pleural effusion was noted in 28 (18.42%) patients and pericardial effusion in 8 (5.3%) patients. No significant difference in the serosal involvement is seen among children and adults. Mediastinal adenopathy was seen 70% of children, 76.3% adolescents and 76.9% of adults and paratracheal nodes were seen most frequently. Nodes had similar features (except matting) among all age groups. Matting of nodes was seen more commonly in children ($P = 0.014$).

CONCLUSION: Pediatric chest tuberculosis can have severe parenchymal lesions and nodal involvement similar to adults. The destructive lung changes observed in children needs immediate attention in view of the longer life span they have and hence in formulating optimal treatment strategies.

© 2013 Baishideng. All rights reserved.

Key words: Tuberculosis; Pulmonary; Primary tuberculosis; Children; Computed tomography

Core tip: Primary tuberculosis in children was traditionally thought to be distinct from reactivation tuberculosis in terms of location, pattern and severity. On the contrary, aggressive forms of pulmonary tuberculosis akin to adult forms are increasingly seen in pediatric clinical practice especially in adolescents. Our study revealed that similar to older patients, children with tuberculosis are equally prone to develop significant destructive lung changes with severe sequelae. Having longer life expectancy the impact is much more severe in children.

Moreover, the cavitating lesions with high bacterial load make them highly infective and pose an important threat to community health.

Thotton Veedu P, Bhalla AS, Vishnubhatla S, Kabra SK, Arora A, Singh D, Gupta AK. Pediatric vs adult pulmonary tuberculosis; a retrospective CT study. *World J Clin Pediatr* 2013; 2(4): 70-76 Available from: URL: <http://www.wjgnet.com/2219-2808/full/v2/i4/70.htm> DOI: <http://dx.doi.org/10.5409/wjcp.v2.i4.70>

INTRODUCTION

Pulmonary tuberculosis (TB) is a common lung infection worldwide with higher prevalence in developing countries. It continues to be a major medical and social problem with high morbidity and mortality. TB is second only to human immunodeficiency virus (HIV)/acquired immune deficiency syndrome (AIDS) as the greatest killer worldwide due to a single infectious agent. In 2011, 8.7 million people contracted tuberculosis out of which 1.4 million died from TB. About half a million children (0-14 years) fell ill with TB, and 64000 children died from the disease in 2011^[1]. The annual risk of tuberculous infection in children in developing countries is 2%-5%. About 8%-20 % of deaths due to tuberculosis occur in children^[2,3].

Tuberculosis in children is mostly related to primary infection and earlier studies stated that it present with various forms of relatively less aggressive primary tuberculosis^[4]. Traditionally it was thought that manifestations of primary tuberculosis are distinct from reactivation tuberculosis in terms of location and pattern^[5,6]. The most common form of pediatric TB, the classical primary complex consists of a focal parenchymal lesion typically in mid-lower zones with enlarged draining hilar/paratracheal node. Other presentations of primary TB include miliary TB^[7], exudative pleuritis and tracheo-bronchial TB^[8,9]. However contrary to the common notion, aggressive forms of pulmonary tuberculosis akin to adult forms are increasingly seen in pediatric clinical practice especially in adolescents^[10-13]. A thorough review of available literature did not reveal any comparative studies with computed tomography (CT) scan in pediatric and adult tuberculosis. Our retrospective study is aimed to compare the pulmonary manifestations of TB in pediatric and adult population.

MATERIALS AND METHODS

In this retrospective study we analyzed CT records of 152 patients of pulmonary tuberculosis who underwent CT scans from November 2010 to January 2013. The diagnosis of pulmonary tuberculosis was based on clinical/and radiographic/and pathologic criteria. Patients were presented with clinical symptoms such as cough for more than two weeks, fever, weight loss, hemoptysis or anorexia. Along with clinical features two out of four of the fol-

lowing criteria had to be met: (1) A positive mantoux test; (2) History of contact with a sputum positive patient of tuberculosis; (3) Radiographic findings of mycobacterium tuberculosis such as primary complex, miliary disease, cavitory lesion, or hilar adenopathy; and (4) Isolation of AFB from sputum, gastric aspirate or broncho-alveolar lavage, lymph node aspirate^[14-19]. Immunocompromised patients were excluded from the study. Informed consent and clearance from the local ethical committee was not required due to the retrospective nature of the study.

CT scans were performed either on Somatom Sensation 40 (Siemens, Erlangen, Germany) or Somatom Definition Flash (Siemens healthcare, Forchheim, Germany). Images were acquired after administering intravenous nonionic iodinated contrast [Iomeron 400 (Iomeprol, Bracco, Milano, Italy), Iohexol 300 (Omnipaque, GE Health care, Ireland)] which was injected by hand with an average delay of 50-70 s and thus providing venous phase images. Adult patients were given 60 mL of contrast and pediatric dose was calculated according to the body weight not exceeding 2 mL/kg.

Scanning was performed in adults with a collimation of 1.2 mm, a pitch of 1.4:1, a 512 × 512 matrix, field view of 38 cm, 120 kVp, and 100 mAs. In children images were acquired with a smaller field of view with 120 kVp and variable mAs (on Somatom Sensation 40) and variable kVp and mAs (on Somatom Definition Flash) according to the body thickness by tube current modulation.

Image reconstruction

After acquisition images were reconstructed in lung and mediastinal windows. Lung window images were reconstructed using sharp kernel (B60f) and a wide window width of 1500 HU with centre at -600 HU. Mediastinal window images are reconstructed using smooth kernel (B30f) and window width of 400 HU with centre at 40 HU. For both lung and mediastinal windows images were reconstructed with a thickness of 5mm in adults and 3 mm in children. HRCT images were reconstructed in lung window settings using ultra-sharp kernel (U80f) with section thickness of 1.5 mm. For HRCT reconstruction interslice gap was 10 mm in adults and 8 mm in children.

Image analysis

Images were reviewed by two radiologists, with 15 years and 7 years of experience in thoracic imaging and interpretations made by consensus. Images were qualitatively analyzed for the presence of parenchymal changes and lymph nodal involvement. Parenchymal changes such as consolidation, centrilobular nodules, miliary nodules, bronchiectasis, cavitation and fibrosis were assessed. For analyzing the zonal predominance and bulkiness of the disease both lungs were divided into upper, mid and lower zones. Lung field from apex to carina as upper zone, carina to the level of inferior pulmonary veins as mid zone and below as lower zone. Distribution of

abnormalities and total number of zones involved as a measurement of bulkiness of the disease were assessed. Mediastinal and hilar lymph nodes were assessed for the size, necrosis, matting and calcification. Other findings like pleural and pericardial effusion were noted.

Statistical analysis

For descriptive statistics the study population was divided into three groups; group A: children (less than or equal to 10 years), group B: adolescents (11-18 years) and Group C: adults (above 18 years). The incidence, pattern and severity of parenchymal changes and nodal involvement were compared among the groups. They were also divided as below and above 10 years; and below and above 18 years for the determination of statistical significance. Statistical analysis was done using stata statistical software (version 12.1). χ^2 test and Fisher's exact test were used for analysis. A *P* value of less than 0.05 was considered as significant.

RESULTS

The study group included 80 males and 72 females patients ranging in age from 3 mo to 96 years (mean 30 years). They were comprised of 10 children, 38 adolescents and 104 adults. Incidence and pattern of parenchymal changes and lymph nodal involvement were analyzed and compared among different groups (Figure 1).

We compared the parenchymal changes among pediatric, adolescent and adult patients. Parenchymal lesions were noted in 6 (60%) children, 27 (71%) adolescents and 80 (76.9%) adults (Table 1). Parenchymal changes were more common in right upper followed by right middle zone in all age groups. Higher incidence of changes noted in upper and middle zones than lower zones. Left lower zone is least commonly involved in patients older than 10 years. Among the patients with parenchymal disease, multiple zones (≥ 3) were seen to be involved in 3 (30%) children, 16 (42.1%) adolescents and 45 (56.25%) adult patients (Table 1). The average number of zones involved in children, adolescent and adult patients were 2.83, 3.33 and 3.32 respectively.

Among the 152 patients we studied, consolidation was found in 54 (35.53%), centrilobular nodules in 93 (61.18%), bronchiectasis in 26 (17.11%), miliary in 4 (2.63%), fibrosis in 22 (14.47%) and cavitation in 36 (23.84%) patients. There was no statistically significant difference in the incidence of consolidation, miliary nodules, bronchiectasis, cavitation and fibrosis among different age groups studied. Centrilobular nodules were seen less commonly in children (*P* = 0.028) (Table 1).

Mediastinal lymphadenopathy was seen in 7 (70%), 29 (76.3%) and 74 (71.2%) respectively in children, adolescents and adult patients (Table 1). Among the mediastinal lymph nodes right paratracheal is the most commonly involved followed by subcarinal in all age groups. Involvement of multiple nodal groups (≥ 2) was seen in 28 (58%) of younger patients (≤ 18 years) and 51 (49%) older pa-

tients. In less than 10 years category all the 7 (70%) children with significant lymphadenopathy had involvement of multiple nodal groups. Involvement of multiple nodes (≥ 2 groups) was more commonly seen in children less than 10 years. The average number of nodal groups involved in children, adolescent and adult patients were 3.57, 2.52 and 2.23 respectively. Lymph node matting was seen more commonly in children (Table 1).

In 4 (40%) patients below 10 years lymphadenopathy was the only finding. Similarly 11 (29%) patients between 10 to 18 years and 24 (23%) patients above 18 years had only lymphadenopathy.

Pleural effusion was noted in 28 (18.42%) patients and 13 of them showed loculation. Pericardial effusion was present in 8 (5.3%) patients. No significant difference in the serosal involvement is seen among children and adults (Table 1).

DISCUSSION

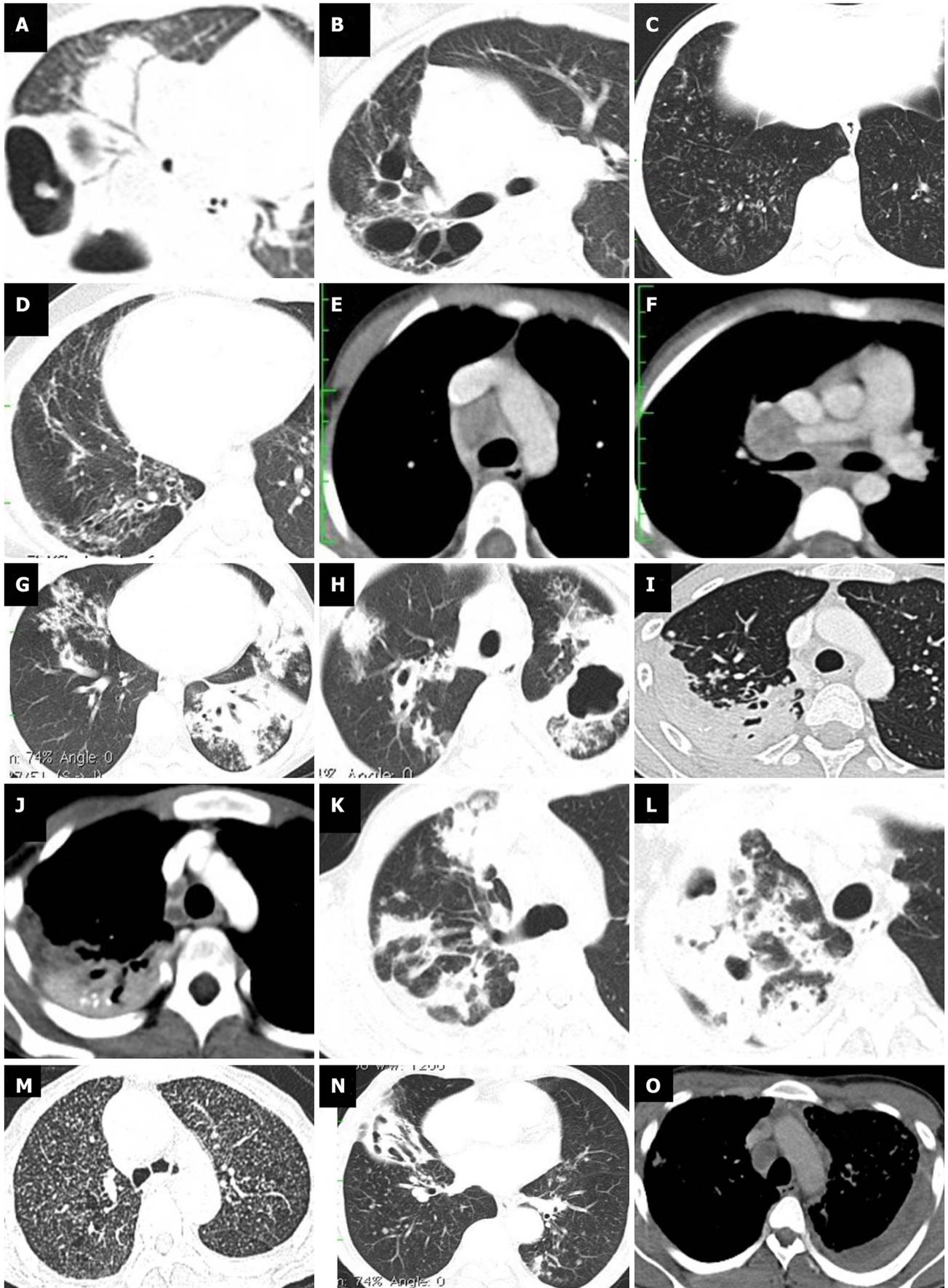
Tuberculosis continues to be a major medical, social and economic problem worldwide with its high morbidity and mortality. Children constitute about 5%-10% of patients suffering from tuberculosis worldwide. World Health Organization (WHO) data shows that in the year 2011 itself about half a million children fell ill with tuberculosis and about 64000 died from the disease^[1].

The pathologic form of pulmonary tuberculosis is classically classified as primary and post primary or secondary tuberculosis and is depended on the sensitivity of infected host. Primary tuberculosis presenting with hilar/paratracheal adenopathy with or without focal parenchymal changes in mid-lower zones is thought to be predominant form of tuberculosis in children. However contrary to the traditional belief incidence of adult form of TB is increasingly seen in pediatric patients in terms of location and severity. Previous authors have already questioned the validity of the terminologies (primary and secondary tuberculosis) in recent literature^[10,11].

Our study included 152 patients including 48 children (below 18 years). Previous studies on pediatric tuberculosis used varying age criteria for children (Table 2). We also have separately analyzed data of children below 10 years and adolescents (11-18 years).

Mediastinal lymph nodes

Mediastinal lymphadenopathy is the predominant finding in primary tuberculosis either alone or in association with lung lesions^[20]. Our study showed mediastinal adenopathy in 7 (70%), 29 (76.3%) and 74 (71.2%) respectively in children, adolescents and adult patients. Compared to previous studies slightly lower incidence of adenopathy is observed in our study (Table 2). Although there was no significant difference in the incidence of adenopathy in different age groups, there was a definite trend in the extensiveness of nodal involvement. Involvement of multiple nodal groups was seen significantly more common in children. Mediastinal adenopathy was the only finding in



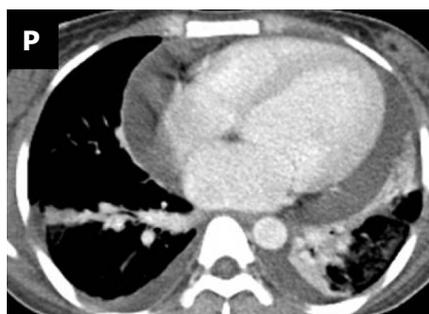


Figure 1 Computed tomography findings in children (less than 10 years), adolescents (10-18 years), and adults (above 18 years) with chest tuberculosis. A-F: Children; A: Parenchymal consolidation in right lung with adjacent thick walled cavities; B: Multiple cavities with adjacent fibrosis in right upper lobe; C: Tiny low density centrilobular nodules seen diffusely in both lungs; D: Fibrobronchiectatic changes in right middle and lower lobes; E-F: Enlarged low attenuating necrotic nodes in right paratracheal, and right hilar location respectively; G-J: Adolescents; G: Parenchymal consolidation with centrilobular nodules in left lung, nodules in right middle lobe; H: Thick walled cavities with surrounding consolidation and air space nodules in both upper lobes; I-J: Pleural based fibrotic changes with calcification in right upper lobe with necrotic right paratracheal node; K-P: Adults; K: Multifocal consolidation in right upper lobe; L: Thick walled cavities in right upper lobe; M: Miliary TB- multiple tiny nodules distributed randomly in both lungs; N: Fibrobronchiectasis in right middle lobe with nodules in left lower lobe; O: Necrotic right paratracheal node with empyema in left side; P: Pericardial and bilateral pleural effusion.

Table 1 Incidence of lung and nodal involvement, Parenchymal changes, Mediastinal lymph nodal involvement and Pleural and pericardial effusion in the patients *n* (%)

		Children (<i>n</i> = 10)	Adolescents (<i>n</i> = 38)	Adults (<i>n</i> = 104)
Incidence of lung and nodal involvement	Parenchymal Lesions	6 (60)	27 (71)	80 (76.9)
	Mediastinal nodes	7 (70)	29 (76.3)	74 (71.2)
Zonal distribution of parenchymal changes	Right upper	5 (50)	19 (50)	58 (55.77)
	Left upper	2 (20)	15 (39.47)	45 (43.27)
	Right middle	4 (40)	19 (50)	54 (51.92)
	Left middle	1 (10)	16 (42.11)	43 (41.35)
	Right lower	3 (30)	12 (31.58)	35 (33.65)
	Left lower	2 (20)	9 (23.68)	30 (28.85)
Pattern of parenchymal changes	Consolidation	3 (30)	16 (42.11)	35 (33.65)
	Centrilobular nodules ^a	2 (20)	24 (63.16)	67 (64.42)
	Miliary nodules	1 (10)	0 (0.00)	3 (2.88)
	Bronchiectasis	1 (10)	8 (21.05)	17 (16.35)
	Fibrosis	1 (10)	7 (18.42)	14 (13.46)
Lymph nodal distribution	Cavitation	3 (30)	10 (26.32)	23 (22.33)
	Paratracheal	7 (70)	22 (57.59)	57 (54.81)
	Precarinal	6 (60)	9 (23.68)	27 (25.96)
	Subcarinal	7 (70)	17 (44.74)	40 (38.46)
	Hilar	5 (50)	13 (34.21)	27 (25.96)
Characteristics of lymph nodes	AP window	0 (0)	9 (23.68)	12 (11.54)
	Lymphadenopathy	7 (70)	21 (55.26)	50 (48.08)
	Necrosis	6 (60)	17 (44.74)	43 (41.35)
	Matting ^a	5 (50)	11 (28.95)	16 (15.38)
Pleural and pericardial effusion in the patients	Calcification	4 (40)	10 (26.32)	25 (24.04)
	Pleural effusion	2 (20)	5 (13.16)	21 (20.19)
	Pleural loculation	1 (10)	2 (5.3)	10 (9.6)
	Pericardial effusion	0 (0)	1 (2.63)	7 (6.73)

^a*P* < 0.05.

31.3% of children (group A and B) against 23% of older patients (group C). This is different from data of Kim *et al*^[21] and Khatami *et al*^[22] who showed isolated nodal involvement in 7% and 10% respectively. Paratracheal followed by subcarinal were the most common locations of nodal involvement in all the groups and is similar to a recent data from Andronikou *et al*^[23] and Mukund *et al*^[24].

Necrotic nodes characterized by inhomogeneous

attenuation or low attenuation centre and enhancing peripheral rim were seen in 48% of children (group A and B) and 41% of adults (group C)^[25]. Necrotic nodes were shown in 65% of children in a study by Mukund *et al*^[24] which is comparable. Matted nodes were seen in 50% of young children (group A) against 19% of older patients (groups B and C) (*P* = 0.02). Nodal calcification was seen in 40%, 26.3% and 24% of children, adolescents and

Table 2 Previous studies on imaging in pediatric chest tuberculosis

Ref.	Age (yr)	Mod	No	Cons	Nodu	Mil	Cavity	Bects	Node	Plefn
Leung <i>et al</i> ^[4]	< 16	X-ray	191	69%	NS	NS	NS	NS	92%	6%
Kim <i>et al</i> ^[21]	< 14	CT	41	49%	29%	17%	7%	NS	83%	17%
Khatami <i>et al</i> ^[22]	< 15	X ray	30	43.30%	NS	NS	NS	NS	90%	6.70%
Koh <i>et al</i> ^[10]	15-19	X-ray	90	25%	96%	NS	45%	0	2%	0
Mukund <i>et al</i> ^[24]	< 17	CT	91	NS	NS	NS	NS	NS	96.70%	NS
Our study	< 18	CT	48	39.60%	54%	2%	27%	18.80%	75%	14.60%

Mod: Imaging modality used; No: Number of cases; Cons: Consolidation; Nodu: Centrilobular nodules; Mil: Miliary nodules; Bects: bronchiectasis; Node: Mediastinal adenopathy; Plefn: Pleural effusion; NS: Not specified.

adults respectively. Nodal calcification was seen in 12 % and 28.4% of children in studies by Kim *et al*^[21] and Mukund *et al*^[24] respectively. Hence apart from higher incidence of matting seen in young children no other significant difference in nodal characteristics is seen in different age groups.

Parenchymal changes

The typical parenchymal change in primary tuberculosis is focal consolidation (Ghon's focus) seen classically in mid and lower zones. Cavitation, fibrosis and bronchiectasis are not commonly seen in primary tuberculosis. In our study, right upper zone was most commonly involved in all age groups and is comparable to the data by Koh *et al*^[10] in which upper zone predominance is seen in 49% of patients. 68.75% of children (group A and B) showed lung parenchymal changes of tuberculosis against 76.9% of older patients (group C). Cavitation was seen in 30% of young children and 26% adolescents. Cavitating pneumonia was seen in 7% of children in the study by Kim *et al*^[21] and 45% of adolescents by Koh *et al*^[10]. The recognition of cavitating disease in children is important because the presence of cavities correlate with organism load, drug resistance, treatment outcome and infectivity^[11]. Bronchiectasis was seen in 18.8 % children (group A and B) against 16.4% in adults (Table 1).

There was no significant statistical difference in the incidence of consolidation, miliary disease, bronchiectasis, fibrosis and cavitation among different age groups. Centrilobular nodules were less commonly seen in children. When we compared the incidence of nodules in children (group A) against older patients (group B and C) significantly lower incidence is noted in young children ($P = 0.014$).

Our study showed comparable frequency of pleural effusion, empyema and pericardial effusion in all age groups. It showed 20% and 13% incidence of pleural effusion in children and adolescents (Table 1). Kim *et al*^[21] observed pleural effusion in 17% of children with tuberculosis.

To conclude, children with pulmonary tuberculosis are equally prone to develop significant destructive changes in the lung with severe sequelae similar to older patients. The impact is much more severe in cases of children because they have longer life expectancy. Moreover,

the cavitating lesions with high bacterial load make these children highly infective and pose an important hazard to community health, than previously thought. The similar location and aggressiveness of parenchymal changes observed in children, blurs the boundary defining primary and reactivation tuberculosis. Hence the need for revision of these terminologies demands urgent attention.

Limitations

First, our study included only those patients who had undergone CT scan at a tertiary institution and hence may not represent the exact patient population at the primary care level. However as all the patients in different age groups had been filtered in the same way the comparisons should remain valid. Secondly, we had only ten patients in below 10 years category and hence further studies including more number of children will help in strengthening the observations.

COMMENTS

Background

Tuberculosis is a contagious lung disease caused by mycobacterium tuberculosis. It is widely prevalent in developing countries and is a major threat to community health. Tuberculosis in children is traditionally believed to be less severe and less contagious as compared to adult disease. However recent evidences suggest that the distinction is far from truth and children are presenting with severe disease in presentation and outcome. This retrospective study is done to compare the disease characteristics based on imaging [computed tomography (CT) of chest] among children and adults who were presented to a tertiary care centre.

Research frontiers

Tuberculosis in children can present in various forms ranging from subclinical infection to destructive parenchymal disease or extensive miliary disease. Early recognition of children having severe disease is essential in the management of disease and in prevention of spread. Although chest radiography is the primary imaging modality in chest tuberculosis, CT is advisable in children presenting with atypical and severe manifestations. Studies on this background are essential in planning and modifying the strategies in the management of pediatric tuberculosis.

Innovations and breakthroughs

Present study has shown that the disease can strike children with similar aggressiveness as in adults and can very much be a source of infection in the community. Hence it is important to take adequate measures to prevent disease spread along with optimal treatment planning.

Applications

Early recognition of severe manifestations of pediatric chest tuberculosis is beneficial in optimal treatment planning. Despite the higher radiation exposure

involved CT of chest is the best investigation to get an accurate estimate of the disease in atypical and complicated cases.

Terminology

CT stands for CT which is a radiological investigation using X-rays to produce tomographic images of the body. In the chest using different post-processing techniques CT provides much greater information than radiography.

Peer review

The study of pediatric vs adult pulmonary tuberculosis is very good. The study highlights the incidence of severe manifestations in pediatric chest tuberculosis, the early detection of which is beneficial in optimal treatment planning.

REFERENCES

- 1 WHO Tuberculosis Fact sheet N°104. Reviewed on February 2013 Available from: URL: <http://www.who.int/mediacentre/factsheets/fs104/en/index.html>. Accessed June 15, 2013
- 2 **Kabra SK**, Lodha R, Seth V. Some current concepts on childhood tuberculosis. *Indian J Med Res* 2004; **120**: 387-397 [PMID: 15520488]
- 3 **Walls T**, Shingadia D. Global epidemiology of paediatric tuberculosis. *J Infect* 2004; **48**: 13-22 [PMID: 14667788 DOI: 10.1016/S0163-4453(03)00121-X]
- 4 **Leung AN**, Müller NL, Pineda PR, FitzGerald JM. Primary tuberculosis in childhood: radiographic manifestations. *Radiology* 1992; **182**: 87-91 [PMID: 1727316]
- 5 Diagnostic Standards and Classification of Tuberculosis in Adults and Children. This official statement of the American Thoracic Society and the Centers for Disease Control and Prevention was adopted by the ATS Board of Directors, July 1999. This statement was endorsed by the Council of the Infectious Disease Society of America, September 1999. *Am J Respir Crit Care Med* 2000; **161**: 1376-1395 [PMID: 10764337 DOI: 10.1164/ajrccm.161.4.16141]
- 6 **Fonseca-Santos J**. Tuberculosis in children. *Eur J Radiol* 2005; **55**: 202-208 [PMID: 15950420 DOI: 10.1016/j.ejrad.2005.05.001]
- 7 **Hussey G**, Chisholm T, Kibel M. Miliary tuberculosis in children: a review of 94 cases. *Pediatr Infect Dis J* 1991; **10**: 832-836 [PMID: 1749696 DOI: 10.1097/00006454-199111000-00008]
- 8 **Milković D**, Richter D, Zorčić-Letoja I, Raos M, Koncul I. Chest radiography findings in primary pulmonary tuberculosis in children. *Coll Antropol* 2005; **29**: 271-276 [PMID: 16117335]
- 9 **Van Dyck P**, Vanhoenacker FM, Van den Brande P, De Schepper AM. Imaging of pulmonary tuberculosis. *Eur Radiol* 2003; **13**: 1771-1785 [PMID: 12942281 DOI: 10.1007/s00330-002-1612-y]
- 10 **Koh WJ**, Jeong YJ, Kwon OJ, Kim HJ, Cho EH, Lew WJ, Lee KS. Chest radiographic findings in primary pulmonary tuberculosis: observations from high school outbreaks. *Korean J Radiol* 2010; **11**: 612-617 [PMID: 21076586 DOI: 10.3348/kjr.2010.11.6.612]
- 11 **Marais BJ**, Parker SK, Verver S, van Rie A, Warren RM. Primary and postprimary or reactivation tuberculosis: time to revise confusing terminology? *AJR Am J Roentgenol* 2009; **192**: W198; author reply W199-W200 [PMID: 19304682]
- 12 **Marais BJ**, Gie RP, Hesselning AH, Beyers N. Adult-type pulmonary tuberculosis in children 10-14 years of age. *Pediatr Infect Dis J* 2005; **24**: 743-744 [PMID: 16094237 DOI: 10.1097/01.inf.0000173305.04212.09]
- 13 **Agrons GA**, Markowitz RI, Kramer SS. Pulmonary tuberculosis in children. *Semin Roentgenol* 1993; **28**: 158-172 [PMID: 8516692]
- 14 **Escreet BC**, Cowie RL. Criteria for the diagnosis of pulmonary tuberculosis. *S Afr Med J* 1983; **63**: 850-854 [PMID: 6857401]
- 15 **Campbell IA**, Bah-Sow O. Pulmonary tuberculosis: diagnosis and treatment. *BMJ* 2006; **332**: 1194-1197 [PMID: 16709993 DOI: 10.1136/bmj.332.7551.1194]
- 16 **Vallejo JG**, Ong LT, Starke JR. Clinical features, diagnosis, and treatment of tuberculosis in infants. *Pediatrics* 1994; **94**: 1-7 [PMID: 8008511]
- 17 **Newton SM**, Brent AJ, Anderson S, Whittaker E, Kampmann B. Paediatric tuberculosis. *Lancet Infect Dis* 2008; **8**: 498-510 [PMID: 18652996 DOI: 10.1016/S1473-3099(08)70182-8]
- 18 **Shingadia D**, Novelli V. Diagnosis and treatment of tuberculosis in children. *Lancet Infect Dis* 2003; **3**: 624-632 [PMID: 14522261 DOI: 10.1016/S1473-3099(03)00771-0]
- 19 **Marais BJ**, Pai M. Recent advances in the diagnosis of childhood tuberculosis. *Arch Dis Child* 2007; **92**: 446-452 [PMID: 17449528 DOI: 10.1136/adc.2006.104976]
- 20 **McAdams HP**, Erasmus J, Winter JA. Radiologic manifestations of pulmonary tuberculosis. *Radiol Clin North Am* 1995; **33**: 655-678 [PMID: 7610237]
- 21 **Kim WS**, Moon WK, Kim IO, Lee HJ, Im JG, Yeon KM, Han MC. Pulmonary tuberculosis in children: evaluation with CT. *AJR Am J Roentgenol* 1997; **168**: 1005-1009 [PMID: 9124105 DOI: 10.2214/ajr.168.4.9124105]
- 22 **Khatami A**, Sabouri S, Ghoroubi J, Rassouli N, Abdollah GF. Radiological Findings of Pulmonary tuberculosis in infants and young children. *Iran J Radiol* 2008; **5**: 231-234
- 23 **Andronikou S**, Joseph E, Lucas S, Brachmeyer S, Du Toit G, Zar H, Swingler G. CT scanning for the detection of tuberculous mediastinal and hilar lymphadenopathy in children. *Pediatr Radiol* 2004; **34**: 232-236 [PMID: 14710313 DOI: 10.1007/s00247-003-1117-0]
- 24 **Mukund A**, Khurana R, Bhalla AS, Gupta AK, Kabra SK. CT patterns of nodal disease in pediatric chest tuberculosis. *World J Radiol* 2011; **3**: 17-23 [PMID: 21286491 DOI: 10.4329/wjr.v3.i1.17]
- 25 **Moon WK**, Im JG, Yeon KM, Han MC. Mediastinal tuberculous lymphadenitis: CT findings of active and inactive disease. *AJR Am J Roentgenol* 1998; **170**: 715-718 [PMID: 9490959 DOI: 10.2214/ajr.170.3.9490959]

P- Reviewers: Abdel-Aziz M, Al-Hagggar M
S- Editor: Qi Y L- Editor: A E- Editor: Lu YJ



GENERAL INFORMATION

World Journal of Clinical Pediatrics (*World J Clin Pediatr*, *WJCP*, online ISSN 2219-2808, DOI: 10.5409) is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

Aim and scope

WJCP covers a variety of clinical medical topics, including fetal diseases, inborn, newborn diseases, infant diseases, genetic diseases, diagnostic imaging, endoscopy, and evidence-based medicine and epidemiology. The current columns of *WJCP* include editorial, frontier, diagnostic advances, therapeutics advances, field of vision, mini-reviews, review, topic highlight, medical ethics, original articles, case report, clinical case conference (Clinicopathological conference), and autobiography. Priority publication will be given to articles concerning diagnosis and treatment of pediatric diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to *WJCP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

WJCP is edited and published by Baishideng Publishing Group (BPG). BPG has a strong professional editorial team composed of science editors, language editors and electronic editors. BPG currently publishes 42 OA clinical medical journals, including 41 in English, has a total of 15 471 editorial board members or peer reviewers, and is a world first-class publisher.

Columns

The columns in the issues of *WJCP* will include: (1) Editorial: The editorial board members are invited to make comments on an important topic in their field in terms of its current research status and future directions to lead the development of this discipline; (2) Frontier: The editorial board members are invited to select a highly cited cutting-edge original paper of his/her own to summarize major findings, the problems that have been resolved and remain to be resolved, and future research directions to help readers understand his/her important academic point of view and future research directions in the field; (3) Diagnostic Advances: The editorial board members are invited to write high-quality diagnostic advances in their field to improve the diagnostic skills of readers. The topic covers general clinical diagnosis, differential diagnosis, pathological diagnosis, laboratory diagnosis, imaging diagnosis, endoscopic diagnosis, biotechnological diagnosis, functional diagnosis, and physical diagnosis; (4) Therapeutics Advances: The editorial board members are invited to write high-quality therapeutic advances in their field to help improve the therapeutic skills of readers. The topic covers medication therapy, psychotherapy, physical therapy, replacement therapy, interventional therapy, minimally invasive therapy, endoscopic therapy, transplantation therapy, and surgical therapy; (5) Field of Vision: The editorial board members are invited to write commentaries on classic articles, hot topic articles, or latest articles to keep readers at the forefront of research and increase their levels of clinical research. Classic articles refer to papers that are

included in Web of Knowledge and have received a large number of citations (ranking in the top 1%) after being published for more than years, reflecting the quality and impact of papers. Hot topic articles refer to papers that are included in Web of Knowledge and have received a large number of citations after being published for no more than 2 years, reflecting cutting-edge trends in scientific research. Latest articles refer to the latest published high-quality papers that are included in PubMed, reflecting the latest research trends. These commentary articles should focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions. Basic information about the article to be commented (including authors, article title, journal name, year, volume, and inclusive page numbers; (6) Minireviews: The editorial board members are invited to write short reviews on recent advances and trends in research of molecular biology, genomics, and related cutting-edge technologies to provide readers with the latest knowledge and help improve their diagnostic and therapeutic skills; (7) Review: To make a systematic review to focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions; (8) Topic Highlight: The editorial board members are invited to write a series of articles (7-10 articles) to comment and discuss a hot topic to help improve the diagnostic and therapeutic skills of readers; (9) Medical Ethics: The editorial board members are invited to write articles about medical ethics to increase readers' knowledge of medical ethics. The topic covers international ethics guidelines, animal studies, clinical trials, organ transplantation, etc.; (10) Clinical Case Conference or Clinicopathological Conference: The editorial board members are invited to contribute high-quality clinical case conference; (11) Original Articles: To report innovative and original findings in clinical pediatrics; (12) Brief Articles: To briefly report the novel and innovative findings in clinical pediatrics; (13) Meta-Analysis: Covers the systematic review, mixed-treatment comparison, meta-regression, and overview of reviews, in order to summarize a given quantitative effect, e.g., the clinical effectiveness and safety of clinical treatments by combining data from two or more randomized controlled trials, thereby providing more precise and externally valid estimates than those which would stem from each individual dataset if analyzed separately from the others; (14) Case Report: To report a rare or typical case; (15) Letters to the Editor: To discuss and make reply to the contributions published in *WJCP*, or to introduce and comment on a controversial issue of general interest; (16) Book Reviews: To introduce and comment on quality monographs of clinical pediatrics; and (17) Autobiography: The editorial board members are invited to write their autobiography to provide readers with stories of success or failure in their scientific research career. The topic covers their basic personal information and information about when they started doing research work, where and how they did research work, what they have achieved, and their lessons from success or failure.

Name of journal

World Journal of Clinical Pediatrics

ISSN

ISSN 2219-2808 (online)

Launch date

June 8, 2012

Instructions to authors

Frequency

Quarterly

Editor-in-Chief

Eduardo H Garin, MD, Professor, Department of Pediatrics, University of Florida, 1600 SW Archer Road. HD214, Gainesville, FL 32610, United States

Editorial office

Jin-Lei Wang, Director
Xiu-Xia Song, Vice Director
World Journal of Clinical Pediatrics
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: wjcp@wjnet.com
<http://www.wjnet.com>

Publisher

Baishideng Publishing Group Co., Limited
Flat C, 23/F, Lucky Plaza, 315-321 Lockhart Road,
Wan Chai, Hong Kong, China
Telephone: +852-58042046
Fax: +852-31158812
E-mail: bpgoffice@wjnet.com
<http://www.wjnet.com>

Production center

Beijing Baishideng BioMed Scientific Co., Limited
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381892
Fax: +86-10-85381893

Representative office

USA Office
8226 Regency Drive,
Pleasanton, CA 94588-3144, United States

Instructions to authors

Full instructions are available online at http://www.wjnet.com/2219-2808/g_info_20100722180909.htm.

Indexed and Abstracted in

Digital Object Identifier.

SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Redit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJCP* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now

available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at: <http://www.wjgnet.com/esps/>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/2219-2808/g_info_20100722180909.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjcp@wjgnet.com, or by telephone: +86-10-85381892. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g., Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province,

country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g., Telephone: +86-10-85381892 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision on acceptance is made only when at least two experts recommend publication of an article. All peer-reviewers are acknowledged on Express Submission and Peer-review System website.

Abstract

There are unstructured abstracts (no less than 200 words) and structured abstracts. The specific requirements for structured abstracts are as follows:

An informative, structured abstract should accompany each manuscript. Abstracts of original contributions should be structured into the following sections: AIM (no more than 20 words; Only the purpose of the study should be included. Please write the Aim in the form of "To investigate/study/..."), METHODS (no less than 140 words for Original Articles; and no less than 80 words for Brief Articles), RESULTS (no less than 150 words for Original Articles and no less than 120 words for Brief Articles; You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g., 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$), and CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Core tip

Please write a summary of less than 100 words to outline the most innovative and important arguments and core contents in your paper to attract readers.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both.

Illustrations

Figures should be numbered as 1, 2, 3, etc., and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ... etc. It is our principle to publish high resolution-figures for the E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, etc., and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any

Instructions to authors

footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a*P* < 0.05, ^b*P* < 0.01 should be noted (*P* > 0.05 should not be noted). If there are other series of *P* values, ^c*P* < 0.05 and ^d*P* < 0.01 are used. A third series of *P* values can be expressed as ^e*P* < 0.05 and ^f*P* < 0.01. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, etc., in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g., PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic

effect of Jianpi Yishen decoction in treatment of Pixu-diarthra. *Shijie Huaren Xiaobua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498.

2002 Aug 1

Statistical dataWrite as mean \pm SD or mean \pm SE.**Statistical expression**

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h; blood glucose concentration, *c* (glucose) 6.4 \pm 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantum numbers can be found at: http://www.wjgnet.com/2219-2808/g_info_20100725073806.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, *etc.*

Biology: *H. pylori*, *E. coli*, *etc.*

Examples for paper writing

All types of articles' writing style and requirement will be found in the link: <http://www.wjgnet.com/esps/NavigationInfo.aspx?id=15>

RESUBMISSION OF THE REVISED MANUSCRIPTS

Authors must revise their manuscript carefully according to the

revision policies of Baishideng Publishing Group Co., Limited. The revised version, along with the signed copyright transfer agreement, responses to the reviewers, and English language Grade A certificate (for non-native speakers of English), should be submitted to the online system *via* the link contained in the e-mail sent by the editor. If you have any questions about the revision, please send e-mail to esps@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2219-2808/g_info_20100725073726.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2219-2808/g_info_20100725073445.htm.

Proof of financial support

For papers supported by a foundation, authors should provide a copy of the approval document and serial number of the foundation.

STATEMENT ABOUT ANONYMOUS PUBLICATION OF THE PEER REVIEWERS' COMMENTS

In order to increase the quality of peer review, push authors to carefully revise their manuscripts based on the peer reviewers' comments, and promote academic interactions among peer reviewers, authors and readers, we decide to anonymously publish the reviewers' comments and author's responses at the same time the manuscript is published online.

PUBLICATION FEE

WJCP is an international, peer-reviewed, OA online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium and format, provided the original work is properly cited. The use is non-commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 600 USD per article. All invited articles are published free of charge.



百世登

Baishideng®

Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza, 315-321 Lockhart Road,

Wan Chai, Hong Kong, China

Fax: +852-31158812

Telephone: +852-58042046

E-mail: bpgoffice@wjgnet.com

<http://www.wjgnet.com>

