ISSN 1948-5204 (online)

World Journal of *Gastrointestinal Oncology*

World J Gastrointest Oncol 2016 June 15; 8(6): 481-525

Published by Baishideng Publishing Group Inc

WJGO World Journal of Gastrointestinal Oncology

A peer-reviewed, online, open-access journal of gastrointestinal oncology

Editorial Board

2016-2019

The *World Journal of Gastrointestinal Oncology* Editorial Board consists of 420 members, representing a team of worldwide experts in gastrointestinal oncology. They are from 42 countries, including Argentina (2), Australia (11), Austria (1), Belgium (5), Brazil (2), Canada (5), Chile (2), China (55), Czech Republic (1), Denmark (1), Finland (2), France (8), Germany (23), Greece (13), Hungary (2), India (9), Iran (3), Ireland (2), Israel (4), Italy (39), Japan (44), Kuwait (2), Mexico (1), Netherlands (8), New Zealand (2), Norway (1), Poland (3), Portugal (4), Romania (1), Saudi Arabia (1), Serbia (2), Singapore (4), South Korea (29), Spain (10), Sweden (5), Switzerland (2), Syria (1), Thailand (1), Turkey (6), United Arab Emirates(1), United Kingdom (12), and United States (90).

Hiroki Kuniyasu, *Kashihara*
Yukinori Kurokawa, *Osaka*
Chihaya Maesawa, *Iwate*
Yoshinori Marunaka, *Kyoto*
Osam Mazda, *Kyoto*
Shinichi Miyagawa, *Matumoto*
Toshiyuki Nakayama, *Fukuoka*
Masahiko Nishiyama, *Saitama*
Koji Oba, *Kyoto*
Masayuki Ohtsuka, *Chiba*
Tomoyuki Shibata, *Toyoake*
Mitsugi Shimoda, *Tochigi*
Haruhiko Sugimura, *Hamamatsu*
Tomomitsu Tahara, *Aichi*
Shinji Takai, *Takatsuki*
Satoru Takayama, *Aichi*
Akio Tomoda, *Tokyo*
Akihiko Tsuchida, *Tokyo*
Yasuo Tsuchiya, *Niigata*
Takuya Watanabe, *Niigata*
Toshiaki Watanabe, *Tokyo*
Hiroki Yamaue, *Wakayama*
Hiroshi Yasuda, *Kumamoto*
Yutaka Yonemura, *Oosaka*
Reigetsu Yoshikawa, *Osaka*

Kuwait

Fahd Al-Mulla, *Safat*
Salem Alshemmari, *Safat*

Mexico

O Arrieta, *Mexico City*

Netherlands

Elisabeth Bloemena, *Amsterdam*
JP De Boer, *Amsterdam*
Peter JK Kuppen, *Leiden*
Gerrit Albert Meijer, *Amsterdam*
Anya N Milne, *Utrecht*
M Muller, *Groningen*
Cornelis FM Sier, *Leiden*

New Zealand

Lynnette Robin Ferguson, *Auckland*
Jonathan Barnes Koea, *Auckland*

Norway

Kjetil Soreide, *Stavanger*

Poland

Andrzej Szkaradkiewicz, *Poznań*
Michal Tenderenda, *Polskiego*
Jerzy Wydmanski, *Gliwice*

Portugal

Celso Albuquerque Reis, *Oporto*
Lucio Lara Santos, *Porto*
Maria Raquel Campos Seruca, *Porto*
Manuel Rodrigues Teixeira, *Oporto*

Romania

Marius Raica, *Timisoara*

Saudi Arabia

Ragab Hani Donkol, *Abha*

Serbia

Milos M Bjelovic, *Belgrade*
Goran Z Stanojevic, *Nish*

Singapore

Peh Yean Cheah, *Singapore*
Si-Shen Feng, *Singapore*
Zhi-Wei Huang, *Singapore*
Qi Zeng, *Singapore*

South Korea

Seungmin Bang, *Seoul*
Daeho Cho, *Seoul*
Byung I Choi, *Seoul*
Hyun Cheol Chung, *Seoul*
Sang-Uk Han, *Suwon*
Jun-Hyeog Jang, *Inchon*
Seong Woo Jeon, *Taegu*
Dae Hwan Kang, *Inchon*
Gyeong Hoon Kang, *Seoul*
Dong Yi Kim, *Kwangju*
Jae J Kim, *Seoul*
Jin Cheon Kim, *Seoul*
Jong Gwang Kim, *Daegu*
Min Chan Kim, *Pusan*
Samyong Kim, *Daegu*
Tae IL Kim, *Seoul*
Young-Woo Kim, *Goyang-si*
Inchul Lee, *Seoul*
Jung Weon Lee, *Seoul*
Kyu Taek Lee, *Seoul*
Kyung Hee Lee, *Daegu*
Na Gyong Lee, *Seoul*
Suk Kyeong Lee, *Seoul*
Jong-Baeck Lim, *Seoul*
Young Joo Min, *Ulsan*
Sung-Soo Park, *Seoul*
Young Kee Shin, *Seoul*
Hee Jung Son, *Seoul*
Si Young Song, *Seoul*

Spain

Manuel Benito, *Madrid*
JI Casal, *Madrid*
Antoni Castells, *Barcelona*
E Folch-Puy, *Barcelona*
Jose JG Marin, *Salamanca*
Joan Maurel, *Barcelona*
Jose M Ramia, *Madrid*
Margarita Sanchez-Beato, *Madrid*
Laura Valle, *Barcelona*
Jesus Vioque, *Alacant*

Sweden

Nils Albiin, *Stockholm*
Samuel Lundin, *Goteborg*
Haile Mahteme, *Uppsala*
Richard Palmqvist, *Umea*
Ning Xu, *Lund*

Switzerland

Paul M Schneider, *Zurich*
Luigi Tornillo, *Basel*

Syria

Zuhir Alshehabi, *Lattakia*

Thailand

Sopit Wongkham, *Khon Kaen*

Turkey

Ugur Coskun, Ankara
Vedat Goral, Izmir
Yavuz Selim Sari, *YeniLevent*
Mesut Tez, *Ankara*
Murat H Yener, *Tekirdag*

United Arab Emirates

Riyad Bendardaf, *Sharjah*

United Kingdom

Runjan Chetty, *Glasgow*
Chris Deans, *Edinburgh*
Dipok Kumar Dhar, *London*
Giuseppe Garcea, *Leicester*
Oleg Gerasimenko, *Liverpool*
Neena Kalia, *Birmingham*
Anthony Maraveyas, *East Yorkshire*
Andrew Maw, *North Wales*
Kymberley Thorne, *Swansea*
Chris Tselepis, *Birmingham*

Nicholas FS Watson, *Nottingham*
Ling-Sen Wong, *Coventry*

United States

Shrikant Anant, *Oklahoma*
Seung Joon Baek, *Knoxville*
Jamie S Barkin, *Miami Beach*
H Bernstein, *Tucson*
Kimberly Maureen Brown, *Kansas City*
Weibiao Cao, *Providence*
Deliang Cao, *Springfield*
Chris N Conteas, *Los Angeles*
Pelayo Correa, *New Orleans*
JJ John Cullen, *Lowa*
James C Cusack, *Boston*
Ananya Das, *Scottsdale*
Juan Dominguez-Bendala, *Miami*
Wafik S El-Deiry, *Philadelphia*
Laura Elnitski, *Rockville*
Thomas Joseph Fahey, *New York*
James W Freeman, *San Antonio*
Bruce Joseph Giantonio, *Philadelphia*
Ajay Goel, *Dallas*
Karen Gould, *Omaha*
GA Nagana Gowda, *Lafayette*
Stephen Randolph Grobmyer, *Gainesville*
Young S Hahn, *Charlottesville*
John W Harmon, *Baltimore*
Paul J Higgins, *Albany*
Steven Norbit Hochwald, *Gainesville*
Su-Yun Huang, *Houston*
Qin Huang, *Duarte*
Jamal A Ibdah, *Columbia*
Yihong Jiang-Cao Kaufmann, *Little Rock*
Temitope Olubunmilayo Keku, *Chapel Hill*
Vijay P Khatri, *Sacramento*
Peter Sean Kozuch, *New York*
Sunil Krishnan, *Houston*
Robert R Langley, *Houston*
Otto Schiueh-Tzang Lin, *Seattle*
Ke-Bin Liu, *Augusta*
Rui-Hai Liu, *Ithaca*
Deryk Thomas Loo, *South San Francisco*
Andrew M Lowy, *La Jolla*
Bo Lu, *Nashville*
David M Lubman, *Ann Arbor*
James David Luketich, *Pittsburgh*
Ju-Hua Luo, *Morgantown*
Henry Thomson Lynch, *Omaha*
Shelli R McAlpine, *San Diego*
Ellen Darcy McPhail, *Rochester*
Anil Mishra, *Cincinnati*
Priyabrata Mukherjee, *Rochester*
Steffan Todd Nawrocki, *Memphis*
Shuji Ogino, *Boston*
Macaulay Onuigbo, *Eau Claire*
Philip Agop Philip, *Detriot*
Blase N Polite, *Chicago*
James A Radosevich, *Chicago*
Robert Raffaniello, *New York*
Jasti S Rao, *Peoria*
Srinevas Kadumpalli Reddy, *Durham*
Stephen H Safe, *Houston*
Muhammad Wasif Saif, *New Haven*
Prateek Sharma, *Kansas City*
Eric Tatsuo Shinohara, *Philadelphia*
Liviu Andrei Sicinschi, *New Orleans*
Pankaj K Singh, *Omaha*
Seema Singh, *Mobile*
William Small, *Chicago*
Sanjay Srivastava, *Amarillo*
Gloria H Su, *New York*
Sujha Subramanian, *Waltham*
David W Townsend, *Boothbay Harbor*
Asad Umar, *Rockville*
Ji-Ping Wang, *Buffalo*
Zheng-He Wang, *Cleveland*
Michael J Wargovich, *Charleston*
Neal W Wilkinson, *Iowa City*
Siu-Fun Wong, *Pomona*
Shen-Hong Wu, *New York*
Ke-Ping Xie, *Houston*
Dong Xie, *Los Angeles*
Hao-Dong Xu, *Rochester*
Xiao-Chun Xu, *Houston*
Zeng-Quan Yang, *Detroit*
Gary Y Yang, *Buffalo*
Wan-Cai Yang, *Chicago*
Zuo-Feng Zhang, *South Los Angeles*
Andrew X Zhu, *Boston*

WJGO

World Journal of Gastrointestinal Oncology

Contents

Monthly Volume 8 Number 6 June 15, 2016

TOPIC HIGHLIGHT

REVIEW

ORIGINAL ARTICLE

Case Control Study

Contents

World Journal of Gastrointestinal Oncology
Volume 8 Number 6 June 15, 2016

NAME OF JOURNAL
World Journal of Gastrointestinal Oncology

ISSN
ISSN 1948-5204 (online)

LAUNCH DATE
October 15, 2009

FREQUENCY
Monthly

Xiu-Xia Song, Vice Director
World Journal of Gastrointestinal Oncology
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: editorialoffice@wjgnet.com
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
http://www.wjgnet.com

PUBLISHER
Baishideng Publishing Group Inc
8226 Regency Drive,
Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
http://www.wjgnet.com

PUBLICATION DATE
June 15, 2016

INSTRUCTIONS TO AUTHORS
Full instructions are available online at http://www.wjgnet.com/bpg/g_info_20160116143427.htm

ONLINE SUBMISSION
http://www.wjgnet.com/esps/

World Journal of Gastrointestinal Oncology

Submit a Manuscript: http://www.wjgnet.com/esps/
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
DOI: 10.4251/wjgo.v8.i6.481

World J Gastrointest Oncol 2016 June 15; 8(6): 481-488
ISSN 1948-5204 (online)

TOPIC HIGHLIGHT

2016 Colorectal Cancer: Global view

Colorectal cancer in the young, many questions, few answers

Kemal I Deen, Hiroshi Silva, Raeed Deen, Pramodh C Chandrasinghe

Kemal I Deen, Consultant in Colon and Rectal Surgery, The Asiri Surgical Hospital, Colombo 11600, Sri Lanka

Hiroshi Silva, Pramodh C Chandrasinghe, The University of Kelaniya Medical School, Ragama 11600, Sri Lanka

Raeed Deen, The University of Sydney Medical School, Sydney NSW 2006, Australia

Author contributions: All authors equally contributed to this paper with conception and design, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

Conflict-of-interest statement: No potential conflicts of interest relevant to this article were reported.

Correspondence to: Kemal I Deen, MD, MS, FRCS, FACRSI, FNat, Ac Sci, Consultant in Colon and Rectal Surgery, The Asiri Surgical Hospital, No.21 Kirimandala Mawatha, Colombo 11600, Sri Lanka. kemaldeen4@gmail.com
Telephone: +94-777-746158

Received: January 4, 2016
Peer-review started: January 5, 2016
First decision: January 30, 2016
Revised: February 29, 2016
Accepted: March 14, 2016
Article in press: March 16, 2016
Published online: June 15, 2016

Abstract

At a time where the incidence of colorectal cancer, a disease predominantly of developed nations, is showing a decline in those 50 years of age and older, data from the West is showing a rising incidence of this cancer in young individuals. Central to this has been the 75% increase in rectal cancer incidence in the last four decades. Furthermore, predictive data based on mathematical modelling indicates a 124 percent rise in the incidence of rectal cancer by the year 2030 - a statistic that calls for collective global thought and action. While predominance of colorectal cancer (CRC) is likely to be in that part of the large bowel distal to the splenic flexure, which makes flexible sigmoidoscopic examination an ideal screening tool, the cost and benefit of mass screening in young people remain unknown. In countries where the incidence of young CRC is as high as 35% to 50%, the available data do not seem to indicate that the disease in young people is one of high red meat consuming nations only. Improvement in our understanding of genetic pathways in the aetiology of CRC, chiefly of the MSI, CIN and CIMP pathway, supports the notion that up to 30% of CRC is genetic, and may reflect a familial trait or environmentally induced changes. However, a number of other germline and somatic mutations, some of which remain unidentified, may play a role in the genesis of this cancer and stand in the way of a clear understanding of CRC in the young. Clinically, a proportion of young persons with CRC die early after curative surgery, presumably from aggressive tumour biology, compared with the majority in whom survival after operation will remain unchanged for five years or greater. The challenge in the future will be to determine, by genetic fingerprinting or otherwise, those at risk of developing CRC and the determinants of survival in those who develop CRC. Ultimately, prevention and early detection, just like for those over 50 years with CRC, will determine the outcome of CRC in young persons. At present, aside from those with an established familial tendency, there is no consensus on screening young persons who may be at risk. However, increasing awareness of this cancer in the young and the established benefit of prevention in older persons, must be a message that should be communicated with medical students,

primary health care personnel and first contact doctors. The latter constitutes a formidable challenge.

Key words: Colon cancer; Young age; Rectal cancer; Colorectal cancer; Young patients; Survival; Early onset

Core tip: This review of colorectal cancer in the young focuses on new data that reveal CRC to be more a left sided cancer than previously thought and the predicted rise by the year 2030. The article outlines the genetics of colorectal cancer (CRC) and discusses limitation in current knowledge in establishing a fingerprint for sporadic CRC. Aside from diet in its aetiology, luminal alkalinity and the colonic microbiome may be contributory and require further research. The review discusses the need for increased awareness of CRC in the young and the need for global consensus on screening young people at risk.

Deen KI, Silva H, Deen R, Chandrasinghe PC. Colorectal cancer in the young, many questions, few answers. *World J Gastrointest Oncol* 2016; 8(6): 481-488 Available from: URL: http://www.wjgnet.com/1948-5204/full/v8/i6/481.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i6.481

INTRODUCTION

Colorectal cancer (CRC) is now the fourth most common cause of cancer deaths, with 600000 deaths reported worldwide annually - about 8% of all cancer deaths[1,2]. It is the third most common cancer in men and the second most common cancer in women. The sporadic form, known to affect individuals in their fifth and sixth decades of life[3], arises from a pre-existing polyp which progresses to cancer through the adenoma-dysplasia-carcinoma sequence; a pathological process which, in general, takes five to ten years[4], and lends itself to prevention by screening[5,6]. CRC is a disease of developed nations, and screening by faecal occult blood testing and colonoscopy has stemmed its incidence in those over 50 years[6]. By contrast, CRC in the young, was a disease prevalent in the developing world[7-14] compared with Australia, New Zealand and the West, where its prevalence in young individuals was low[11,15,16]. However, more recently, there has been an increase in the number of reports of CRC in the young from the developed world[17-19]. This is of concern because the incidence of rectal cancer has risen by 75% in the last 40 years[20-22], contributing chiefly to the overall rise in cancer prevalence. Furthermore, this disease affects people in the prime of their life, and unlike cancer in older individuals, there is limited knowledge about the aetiology and pathogenesis of CRC in the young. The aim of this review is to present the current status of CRC in the young and to highlight areas for future research.

EPIDEMIOLOGY/PREVALENCE

Historically, CRC in young patients was highest in proportional prevalence from the Asian region. Studies have reported a high young cancer prevalence of 38% in Egypt[7], 18% in Turkey[8], 39% in India[9], 29% in Nepal[10], 23% from Saudi Arabia[11], 19.7% from Sri Lanka[12], 52% from a single institution in Pakistan[13] and 10.1% from Taiwan[14]. Most significantly, a recent study from the United States[19], where the authors evaluated the records of 393241 patients over a 15-years period, revealed an overall decline in CRC by 0.92% - the effect attributed to screening. While this was true for those over 50 years old with CRC, the study observed an alarming increase in CRC in those less than 50 years, specifically, in young patients less than 35 years. Using statistical modelling, the authors predicted an increase in colon cancer by 90% in patients aged 20 to 34 years and 27.7% in those 35 to 49 years old by the year 2030. For rectal cancer, the predicted percentage increase in cancer prevalence for these two age groups was 124.2% and 46% respectively. Gender based analysis of CRC in young patients revealed an equal prevalence in young men and women[22] contradicting the theory that female hormones are protective of colon and rectal cancer. Furthermore, a 1991 study of young patients in North America showed that the disease occurred in 34% more black men and 45% more black women compared with white Caucasian counterparts[23]. Most young patients did not report a family history of CRC; O'Connell *et al*[22] revealed that only 23% of young patients with CRC reported the presence of cancer in a family member.

FAMILY HISTORY

Contrary to previous knowledge, a current estimate of the proportion of CRC likely to have a major hereditary component is between 15% and 30%[24]. The common heritable syndromes in CRC are either familial adenomatous polyposis (FAP) or hereditary non-polyposis colorectal cancer (HNPCC)[25-27] known to be found in 2 to 5 percent of all patients with CRC. Familial adenomatous polyposis is defined by phenotype if an individual has multiple colonic polyps, usually over 100, in association with loss of the tumour suppressor gene -the adenomatous polyposis coli-APC gene-located on the long arm of chromosome 5 (5q21)[25]. Most FAP patients will develop CRC by age 40 years, while in a minority, cancer will manifest in the fifth decade or after, due to the presence of the attenuated *FAP* gene. In contrast to FAP, HNPCC, first described by Henry Lynch, is characterised by the presence of fewer colonic polyps or cancer that is indistinguishable from sporadic CRC. In both conditions, which are of autosomal dominant inheritance, family history is of prime importance. For HNPCC, an affected member or members of a family should have had either CRC (Lynch type 1-site specific) or other extra-intestinal cancers (Lynch type 2), in association with

an index patient with CRC. In the absence of definitive genetic testing, a detailed family history was essential and formed the core of the Amsterdam and Bethesda criteria to make a diagnosis of HNPCC[28,29]. Currently, we know that young patients with an underlying genetic syndrome are more likely to have a family history of cancer and present earlier compared with those with no known genetic syndrome, who presented with late stage metastatic disease[30]. Thus, family history must continue to remain an essential component of clinical evaluation in patients with CRC, while it is essential to note that up to 20 percent of patients with a germline mutation in the study reported by Mork *et al*[30] had no family history of CRC.

ANATOMIC DISTRIBUTION

Several studies have reported that CRC in the young is a condition mostly confined to the left colon and rectum; in a retrospective study of young patients, Leff *et al*[31] revealed that 65% of cancers were in the rectum and that 83% of all colon and rectal cancers were distal to the splenic flexure. Kumar *et al*[32] reported that CRC was confined to the left colon and rectum in 67% of their study population Furthermore, O'Connell *et al*[22] in a structured review of 55 studies comprising 6425 patients with young CRC, reported that cancer of the rectum was most frequent (54%). In the most recent publication of the Surveillance Epidemiology and End Result (SEER) study from the United States, dominance of cancer in the left colon and rectum was again mirrored[19].

PRESENTATION

Studies have shown that CRC in young patients presents with three cardinal features of rectal bleeding, abdominal pain and alteration in bowel habit - constipation, altered stool diameter, mucoid rectal discharge[33,34]. In general, CRC diagnosis in young patients was associated with a delay of approximately 6 mo[33]. Physician related delay in diagnosis was chiefly because of a lack of understanding and suspicion of this disease in the young, where symptoms in young patients were considered due to such benign causes as haemorrhoidal disease by first contact physicians and patients alike. Some other factors that may contribute to delay are patients' preference in seeking non-traditional methods of symptom relief, such as Ayurvedha and Chinese medical treatment, in Asia, and because practitioners of allopathic medicine fail to perform a focused rectal examination at the point of first contact. With current worldwide reports of increasing prevalence of young CRC, it is important that we offer young symptomatic patients flexible sigmoidoscopy early, after comprehensive clinical examination, including focused digital rectal examination.

PATHOLOGY

In young patients, CRC is likely to be found in those with a heritable syndrome[28-30] such as FAP and HNPCC. In the Lynch Syndrome, tumours have been known to be predominant in the proximal colon[35,36], but recent research revealed contradictory data where the most frequent site among early onset CRC patients was the distal colon[37]. Of these, between 40 and 60 percent were in the rectum[38,39]. In the WHO classification of tumours[40], HNPCC and sporadic CRC with microsatellite instability have been classified based on the site and microscopic criteria. These are (1) proximally located mucinous adenocarcinomas which are commonly well circumscribed and are moderate-to-well differentiated; (2) proximally located poorly differentiated adenocarcinomas which show failure of gland formation with malignant epithelium arranged in small clusters, irregular trabeculae or large aggregates in well circumscribed tumours; and (3) adenomas in HNPCC indicating features of high cancer risk including villous and high grade intraepithelial neoplasia which display good circumscription and present as polypoid growths, plaques, bulky masses or ulcers rather than diffuse growths or strictures[40]. In a single centre study, mucinous and signet-ring histological subtypes and poor to non-differentiated tumours were frequently seen among the young[38,41,42], and accounted for 41.5% of all tumours[38]. The incidence of tumour *in situ* (Tis) was lower in young patients compared with older patients and may indicate either failure of early detection or rapid progression from adenoma to carcinoma in the young compared with older patients[43]. Other features that suggest more aggressive tumour biology in the young compared with older patients are the higher percentages of patients with lymph node metastasis ($\geqslant$ 4 lymph nodes), distance metastasis and stage Ⅳ disease[41,42].

GENETICS

All colorectal cancers occur from genetic mutations, which are part of a familial syndrome, hereditary syndrome or as sporadic cancer[44]. Frequent among young patients are either FAP, variants of FAP or HNPCC. Historically, in the sporadic subtype, the origin of CRC was attributed to various common or rare genetic alterations that displayed variable penetrance, and remained largely unidentified[45]. It is now estimated that up to 30% of CRC may have a hereditary component, with identifiable genetic aberration, especially if cancer occurs in the young[23,24,30,46]. Next generation sequencing (NGS) is likely to further increase our knowledge of hitherto unidentified chromosome aberrations in association with cancer[47] resulting in such diagnoses as the Li-Fraumeni syndrome, Cowden's disease, Juvenile polyposis and Peutz-Jegher syndrome[46].

Different from germline mutations, somatic mutation, that may be spontaneous or follow contact with luminal carcinogens, may result in genetic alteration of a colonocyte in which control of apoptosis is lost in conjunction with a series of chromosomal changes that create microsatellite instability[43]. In fact, the aetiology and range of hitherto unidentified germline and early onset somatic mutations is likely to be more extensive than

previously understood, which makes our understanding of the pathology in young patients with sporadic cancer even more complex. Essential to our understanding of tumourigenesis is knowledge of preservation of DNA integrity in the intestinal epithelial cell; deep within the base of the intestinal crypt lies the colonocyte stem cell that is covered in a thick layer of mucus. Each stem cell is designed to replicate into a transit amplifier stem cell and an inert stem cell that remains in the protected crypt base, remote from contact with carcinogens that may be present in the lumen of large bowel, thus preserving its DNA intact. In health, upward migration of the amplifier cell will give rise to a functional colonocyte that will shed in 5 to 7 d by genetically determined apoptosis, controlled by the *p53* gene located on chromosome 17 and the mitogen-activated protein kinase pathway (MAPK)[43]. The MAPK pathway, of which KRAS and BRAF proteins are part, regulates cell proliferation, cell differentiation, cellular aging and apoptosis[48]. Programmed colonocyte death prevents the propagation of mutagenic change, and constitutes yet another strategy of preserving intestinal cell DNA integrity[43]. In the adenoma-carcinoma sequence, initialisation of neoplastic change occurs with silencing of the tumour suppressor genes located on chromosome 5 (*APC* gene), followed by serial changes in chromosome 17 (*p53* gene-mutated in colorectal cancer) and chromosome 18 (long arm deletion)[49]. Furthermore, simultaneous activation of the proto-oncogene K-Ras will lead to uncontrolled cell growth[49]. Hence, both germline mutations and somatic mutations may drive colorectal cancer in the young.

Currently, the genetic mechanisms that trigger CRC are grounded in three major pathways; chromosomal instability (CIN), microsatellite instability (MSI) and the cytosine-phosphate-guanine island methylator phenotype pathway (CIMP) pathway[50,51] - mechanisms that create genomic instability, which together with a process that will selectively support mutagenic driver cells, produce colorectal cancer. It is essential in our understanding of this process that none of these pathways is mutually exclusive. However, CIN aberrations, by far, constitute the most common pathway in the development of CRC[52].

CIN pathway

This describes the classical adenoma-dysplasia-carcinoma sequence in which it is thought that tumour formation is a result of progressive and sequential inactivation of tumour suppressor genes and, correspondingly, activation of tumour promoting oncogenes - mutation in the adenomatous polyposis coli (APC) gene being an important initial step in this pathway[52]. Likewise, it is known that mutation of the KRAS oncogene contributes to CIN-associated sporadic CRC in up to a half of such sporadic cancer[53]. Since RAS proteins control signaling in cell differentiation and apoptosis, disruption of such pathways will lead to neoplastic transformation. CIN-associated tumours comprise 75% to 80% of all tumours found in Western populations[54].

MSI pathway

It is known that formation of new strands of DNA may be interrupted by base pair mismatches, *i.e.,* mutations which may be either deletions or insertions. In health, the role of mismatch repair proteins is to bind, remove and repair the region of the mismatch error. In cells with malfunction of mismatch repair proteins, these mutations will tend to accumulate within areas of DNA coding called microsatellites. Such areas of microsatellite instability are the cause of sporadic CRC[55].

CIMP pathway

This pathway of CRC differs fundamentally from CIN and MSI, in that, it causes mutation and epigenetic silencing of genes that control the cell cycle outside the APC control system. This pathway is chiefly associated with a group of protein kinases known as BRAF proteins, and usually occurs due to promoter methylation and silencing of the mut-L homologue 1 gene (MLH-1- short arm of chromosome 3), resulting in microsatellite instability. CIMP associated cancer is frequently found in patients of older age, has a slight female preponderance and is associated with right sided colon cancer, similar to the Lynch syndrome. However, it is rare for patients with Lynch syndrome-associated CRC to have BRAF mutations, which helps differentiate Lynch syndrome associated CRC from sporadic CRC[56]. Thus, it becomes evident that no two colorectal cancers are likely to be the same, and that each will have its own unique characteristic genetic "fingerprint". It is also known that each cancer may have more than one of the aforementioned carcinogenic pathways[57,58], which makes genetic imprinting of sporadic CRC all that more challenging. Furthermore, since CIN and MSI associated CRC is known to respond differently to chemotherapeutic agents and impact on cancer related survival, to enable tumour specific personalized treatments, future standard pathological tumour work-up may have to include such genetic "fingerprinting".

RISK FACTORS

A historic study of tumour genesis in the colon shed light on the alkaline environment in the lumen of the colon which, combined with secondary bile acids, is a promoter of tumour formation[59]. N-nitroso compounds and ammonia, produced from bacterial action upon undigested protein products, and secondary bile acids alter the luminal environment, which affect colonocyte function and deplete oxygen levels in the colonic mucosa, thus favouring tumourigenesis. Furthermore, rapid urbanization with environmental pollution, lifestyle alterations such as reduction in physical activity and change in dietary patterns in young individuals[9,60], may have also contributed to the rising incidence of CRC, although this alone does not explain its disproportionate rise in incidence in previously

low incidence parts of the world[61].

SURVIVAL

Multiple studies of young patients with CRC from cancer registries have shown that, in young patients, 5-year survival did not differ from older patients despite a greater proportion of locally advanced cancer, regional lymph node involvement and less favourable histological types in the young[61-63]. Ruiz *et al*[64] showed an overall survival rate of 69.4% and 67.4% at 5 years for colon and rectal cancer respectively from a cancer registry database in Peru. Likewise, Parc *et al*[63], reporting survival data from the central South Korean cancer registry, revealed a 5-year survival of 66% for young patients with cancer of the proximal colon, 70% for patients with distal colon cancer and 66% in patients with rectal cancer. However, if young patients with CRC present with concomitant metastasis, or in the case of a small proportion of patients with unfavourable histological features (poorly differentiated cancer, signet ring cancer), survival may be poor[65]. Chan *et al*[66] have shown that survival in young patients with a poor prognosis is predictable, and that maximum survival in this group of young patients after surgical intervention is no more than 20 mo.

SCREENING

CRC screening guidelines currently recommend routine screening of individuals from the age of 50 years. The screening tests range from invasive procedures such as flexible sigmoidoscopy and colonoscopy, through imaging investigations such as virtual colonoscopy, to minimally invasive procedures such as faecal occult tests[67].

Although each test has its own different advantages and limitations, colonoscopy - widely regarded as the gold standard - has shown to decrease the incidence of CRC up to 80%. However, it is essential to note that colonoscopy is not a perfect test - studies have shown a miss rate of 6%-12% of adenomas > 1 cm and 5% for CRC[67]. Faecal occult tests have shown promise too; an example being, the faecal immunochemical test which has shown high rates of detection of prevalent CRC in an asymptomatic population[68].

With the rising incidence and mortality of CRC in young patients, effective screening methods must be able to detect these tumours early. Current guidelines suggest that individuals with a family history of CRC or adenomatous polyps, other than FAP, undergo screening earlier than at 50 years. That is, from the age of 40 or 10 years before the youngest cancer affected family member, while those with a family history of FAP undergo screening in adolescence[68]. Population based early-onset CRC screening has not been justified due to low prevalence, cost and potential adverse procedural outcomes outweighing the benefits[17]. To detect early onset CRC, suggestions have been to undertake routine screening from 40 years, instead of 50 years - however, decision analysis models have shown no significant life-year gains for this change[37].

To combat the rising incidence by screening of potential early onset CRC patients, awareness among physicians, primary healthcare workers and the lay public must increase. For the physician, this should begin at the stage of medical school by integration of preventive medicine and longitudinal cancer prevention modules into medical school curriculums - which have shown positive results[69], and will improve the future physician's ability to identify young individuals at high risk.

In terms of young patient awareness, it is imperative that young adults are aware of screening for early onset CRC. A study revealed that university students had very poor knowledge of CRC screening, indicating the necessity for early-onset cancer awareness campaigns[70]. Another feasible plan to improve screening rates is the employment of a well-trained lay cancer-screening navigator; this person's role would involve contacting individuals, discussing the importance of screening for CRC and implementing screening procedures such as faecal tests sent by mail. Although this was a feasible strategy for older patients aged 50 to 74 years[71], it has yet to be determined how effective this strategy would be in younger individuals.

To avoid low screening rates, patients' screening method preferences require consideration. Studies have shown faecal aversion to be one of the chief hindrances to screening participation, and a survey revealed that 78% of participants would prefer to provide a blood sample instead[72]. One such blood test to detect CRC, which requires further development, is the assessment of circulating methylated SEPT9 DNA, and although it is able to detect CRC in an asymptomatic individual, improved sensitivity is required for population screening[73]. A highly sensitive and specific blood test for CRC could very well become the gold standard in the future, and thereby decrease incidence and mortality rates.

CONCLUSION

An epidemic of colorectal cancer in young patients is imminent. Based on better understanding of genetic mechanisms, currently it is estimated that genetic predisposition to colorectal cancer is 30% of all CRC. The figure is likely to be higher in young patients if all young patients with CRC were to have genetic assessment by NGS testing. While the MSI, CIN and CIMP pathways have been isolated and well defined, a number of germline and somatic mutations in CRC are likely to manifest from widespread use of NGS, multiple panel genetic tests. Furthermore, multiple permutations of genetic alterations are likely to show up in individual CRCs, with overlap of previously known syndrome based genetic changes, which will make individual genetic fingerprinting of CRC more complex and perhaps the age of onset of CRC, that is, whether young or older, irrelevant. In lifestyle assessment, populations, such as in Egypt, where consumption of red meat is high seem to have similar proportions of young

patients with CRC compared with predominantly non-meat eating populations, such as is found in India, which further complicates the search for a common lifestyle aetiology. What is common across the world in lifestyle is the growing fast food industry and childhood obesity; more thought and research needs to focus on its contributory role. For the present, the majority of cases of CRC remains sporadic and of multifactorial origin: Diet and nutrition, obesity, the colonic microbiome, smoking, alcohol consumption and hitherto unknown germline or somatic mutation. The role of screening for CRC in young patients is not likely to follow a "one test fits all" policy until we have worldwide genetic data in this group of patients. At present, mass screening by flexible sigmoidoscopy is expensive and may yield low productive rates. However, better education of medical students, primary healthcare personnel and first contact doctors, about the benefit of prevention and early detection of CRC in the young is likely to improve early detection rates in young persons. Whether early detection influences lead-time in such young patients with cancer remains unresolved, as some studies have shown a clear cut-off in survival at around 2 years. It is a formidable challenge to fight the rising incidence and mortality in early onset CRC patients, an effort that will require global co-operation and consensus.

REFERENCES

1 **Ferlay J**, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010; **127**: 2893-2917 [PMID: 21351269 DOI: 10.1002/ijc.25516]

2 **American Cancer Society.** Global Cancer Facts and Figures. 2nd ed. Atlanta: American Cancer Society, 2011

3 **Siegel R**, Ward E, Brawley O, Jemal A. Cancer statistics, 2011: the impact of eliminating socioeconomic and racial disparities on premature cancer deaths. *CA Cancer J Clin* 2011; **61**: 212-236 [PMID: 21685461 DOI: 10.3322/caac.20121]

4 **Vogelstein B**, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM, Bos JL. Genetic alterations during colorectal-tumor development. *N Engl J Med* 1988; **319**: 525-532 [PMID: 2841597 DOI: 10.1056/NEJM198809013190901]

5 **Scholefield JH**, Moss S, Sufi F, Mangham CM, Hardcastle JD. Effect of faecal occult blood screening on mortality from colorectal cancer: results from a randomised controlled trial. *Gut* 2002; **50**: 840-844 [PMID: 12010887 DOI: 10.1136/gut.50.6.840]

6 **Bond JH**. Fecal occult blood test screening for colorectal cancer. *Gastrointest Endosc Clin N Am* 2002; **12**: 11-21 [PMID: 11916154]

7 **Abou-Zeid AA**, Khafagy W, Marzouk DM, Alaa A, Mostafa I, Ela MA. Colorectal cancer in Egypt. *Dis Colon Rectum* 2002; **45**: 1255-1260 [PMID: 12352245 DOI: 10.1007/s10350-004-6401-z]

8 **Alici S**, Aykan NF, Sakar B, Bulutlar G, Kaytan E, Topuz E. Colorectal cancer in young patients: characteristics and outcome. *Tohoku J Exp Med* 2003; **199**: 85-93 [PMID: 12705353 DOI: 10.1620/tjem.199.85]

9 **Gupta S**, Bhattacharya D, Acharya AN, Majumdar S, Ranjan P, Das S. Colorectal carcinoma in young adults: a retrospective study on Indian patients: 2000-2008. *Colorectal Dis* 2010; **12**: e182-e189 [PMID: 20128837 DOI: 10.1111/j.1463-1318.2010.02223.x]

10 **Singh Y**, Vaidya P, Hemandas AK, Singh KP, Khakurel M. Colorectal carcinoma in Nepalese young adults: presentation and outcome. *Gan To Kagaku Ryoho* 2002; **29** Suppl 1: 223-229 [PMID: 11890110]

11 **Isbister WH**. Colorectal cancer Below Age 40 in The Kingdom of Saudi Arabia. *Aust N Z J Surg* 1992; **62**: 468-472 [PMID: 1590715 DOI: 10.1111/j.1445-2197.1992.tb07227.x]

12 **de Silva MV**, Fernando MS, Fernando D. Comparison of some clinical and histological features of colorectal carcinoma occurring in patients below and above 40 years. *Ceylon Med J* 2000; **45**: 166-168 [PMID: 11293963 DOI: 10.4038/cmj.v45i4.6722]

13 **Amini AQ**, Samo KA, Memon AS. Colorectal cancer in younger population: our experience. *J Pak Med Assoc* 2013; **63**: 1275-1277 [PMID: 24392559]

14 **Han-Shiang C**. Curative resection of colorectal adenocarcinoma: multivariate analysis of 5-year follow-up. *World J Surg* 1999; **23**: 1301-1306 [PMID: 10552125 DOI: 10.1007/s002689900666]

15 **Adloff M**, Arnaud JP, Schloegel M, Thibaud D, Bergamaschi R. Colorectal cancer in patients under 40 years of age. *Dis Colon Rectum* 1986; **29**: 322-325 [PMID: 3009108 DOI: 10.1007/BF02554121]

16 **Keating J**, Yong D, Cutler G, Johnston J. Multidisciplinary treatment of colorectal cancer in New Zealand: survival rates from 1997-2002. *N Z Med J* 2006; **119**: U2238 [PMID: 16998579]

17 **Young JP**, Win AK, Rosty C, Flight I, Roder D, Young GP, Frank O, Suthers GK, Hewett PJ, Ruszkiewicz A, Hauben E, Adelstein BA, Parry S, Townsend A, Hardingham JE, Price TJ. Rising incidence of early-onset colorectal cancer in Australia over two decades: report and review. *J Gastroenterol Hepatol* 2015; **30**: 6-13 [PMID: 25251195 DOI: 10.1111/jgh.12792]

18 **Steliarova-Foucher E**, Stiller C, Kaatsch P, Berrino F, Coebergh JW, Lacour B, Parkin M. Geographical patterns and time trends of cancer incidence and survival among children and adolescents in Europe since the 1970s (the ACCISproject): an epidemiological study. *Lancet* 2004; **364**: 2097-2105 [PMID: 15589307 DOI: 10.1016/S0140-6736(04)17550-8]

19 **Bailey CE**, Hu CY, You YN, Bednarski BK, Rodriguez-Bigas MA, Skibber JM, Cantor SB, Chang GJ. Increasing disparities in the age-related incidences of colon and rectal cancers in the United States, 1975-2010. *JAMA Surg* 2015; **150**: 17-22 [PMID: 25372703 DOI: 10.1001/jamasurg.2014.1756]

20 **Merrill RM**, Anderson AE. Risk-adjusted colon and rectal cancer incidence rates in the United States. *Dis Colon Rectum* 2011; **54**: 1301-1306 [PMID: 21904146 DOI: 10.1097/DCR.0b013e3182242bd3]

21 **O'Connell JB**, Maggard MA, Liu JH, Etzioni DA, Livingston EH, Ko CY. Rates of colon and rectal cancers are increasing in young adults. *Am Surg* 2003; **69**: 866-872 [PMID: 14570365]

22 **O'Connell JB**, Maggard MA, Livingston EH, Yo CK. Colorectal cancer in the young. *Am J Surg* 2004; **187**: 343-348 [PMID: 15006562 DOI: 10.1016/j.amjsurg.2003.12.020]

23 **Griffin PM**, Liff JM, Greenberg RS, Clark WS. Adenocarcinomas of the colon and rectum in persons under 40 years old. A population-based study. *Gastroenterology* 1991; **100**: 1033-1040 [PMID: 2001800]

24 **Slattery ML**. Diet, lifestyle, and colon cancer. *Semin Gastrointest Dis* 2000; **11**: 142-146 [PMID: 10950460]

25 **Fearnhead NS**, Britton MP, Bodmer WF. The ABC of APC. *Hum Mol Genet* 2001; **10**: 721-733 [PMID: 11257105 DOI: 10.1093/hmg/10.7.721]

26 **Jass JR**. Hereditary Non-Polyposis Colorectal Cancer: the rise and fall of a confusing term. *World J Gastroenterol* 2006; **12**: 4943-4950 [PMID: 16937488]

27 **Lynch HT**, Lynch JF. Hereditary nonpolyposis colorectal cancer (Lynch syndromes I and II): a common genotype linked to oncogenes? *Med Hypotheses* 1985; **18**: 19-28 [PMID: 4069033 DOI: 10.1016/0306-9877(85) 90115-x]

28 **Vasen HF**, Watson P, Mecklin JP, Lynch HT. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. *Gastroenterology* 1999; **116**: 1453-1456 [PMID: 10348829 DOI: 10.1016/S0016-5085(99)70510-X]

29 **Umar A**, Boland CR, Terdiman JP, Syngal S, de la Chapelle A, Rüschoff J, Fishel R, Lindor NM, Burgart LJ, Hamelin R, Hamilton SR, Hiatt RA, Jass J, Lindblom A, Lynch HT, Peltomaki

P, Ramsey SD, Rodriguez-Bigas MA, Vasen HF, Hawk ET, Barrett JC, Freedman AN, Srivastava S. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. *J Natl Cancer Inst* 2004; **96**: 261-268 [PMID: 14970275 DOI: 10.1093/jnci/djh034]

30 **Mork ME**, You YN, Ying J, Bannon SA, Lynch PM, Rodriguez-Bigas MA, Vilar E. High Prevalence of Hereditary Cancer Syndromes in Adolescents and Young Adults With Colorectal Cancer. *J Clin Oncol* 2015; **33**: 3544-3549 [PMID: 26195711 DOI: 10.2015.61.4503]

31 **Leff DR**, Chen A, Roberts D, Grant K, Western C, Windsor AC, Cohen CR. Colorectal cancer in the young patient. *Am Surg* 2007; **73**: 42-47 [PMID: 17249455]

32 **Kumar RR**, King J, Holt A, Huynh R, Mittal R, Deen R, Kim J. Prevalence of left-sided colorectal cancer and benefit of flexible sigmoidoscopy: a county hospital experience. *Am Surg* 2007; **73**: 994-997 [PMID: 17983066]

33 **Dozois EJ**, Boardman LA, Suwanthanma W, Limburg PJ, Cima RR, Bakken JL, Vierkant RA, Aakre JA, Larson DW. Young-onset colorectal cancer in patients with no known genetic predisposition: can we increase early recognition and improve outcome? *Medicine* (Baltimore) 2008; **87**: 259-263 [PMID: 18794708 DOI: 10.1097/MD.0b013e3181881354]

34 **Taggarshe D**, Rehil N, Sharma S, Flynn JC, Damadi A. Colorectal cancer: are the "young" being overlooked? *Am J Surg* 2013; **205**: 312-316; discussion 316 [PMID: 23414955 DOI: 10.1016/j.amjsurg.2012.10.016]

35 **Lynch HT**, Lynch PM, Lanspa SJ, Snyder CL, Lynch JF, Boland CR. Review of the Lynch syndrome: history, molecular genetics, screening, differential diagnosis, and medicolegal ramifications. *Clin Genet* 2009; **76**: 1-18 [PMID: 19659756 DOI: 10.1111/j.1399-0004.2009.01230.x]

36 **da Silva FC**, de Oliveira LP, Santos EM, Nakagawa WT, Aguiar Junior S, Valentin MD, Rossi BM, de Oliveira Ferreira F. Frequency of extracolonic tumors in Brazilian families with Lynch syndrome: analysis of a hereditary colorectal cancer institutional registry. *Fam Cancer* 2010; **9**: 563-570 [PMID: 20697958 DOI: 10.1007/s10689-010-9373-2]

37 **Ahnen DJ**, Wade SW, Jones WF, Sifri R, Mendoza Silveiras J, Greenamyer J, Guiffre S, Axilbund J, Spiegel A, You YN. The increasing incidence of young-onset colorectal cancer: a call to action. *Mayo Clin Proc* 2014; **89**: 216-224 [PMID: 24393412 DOI: 10.1016/j.mayocp.2013.09.006]

38 **You YN**, Xing Y, Feig BW, Chang GJ, Cormier JN. Young-onset colorectal cancer: is it time to pay attention? *Arch Intern Med* 2012; **172**: 287-289 [PMID: 22157065 DOI: 10.1001/archinternmed.2011.602.]

39 **Zahir MN**, Azhar EM, Rafiq S, Ghias K, Shabbir-Moosajee M. Clinical features and outcome of sporadic colorectal carcinoma in young patients: a cross-sectional analysis from a developing country. *ISRN Oncol* 2014; **2014**: 461570 [PMID: 25006505 DOI: 10.1155/2014/461570]

40 **Hamilton SR,** Aaltonen LA. Pathology and genetics of tumours of the digestive system. Lyon: IARC press, 2000: 103-126

41 **Fu JF**, Huang YQ, Yang J, Yi CH, Chen HL, Zheng S. Clinical characteristics and prognosis of young patients with colorectal cancer in Eastern China. *World J Gastroenterol* 2013; **19**: 8078-8084 [PMID: 24307803 DOI: 10.3748/wjg.v19.i44.8078]

42 **Domergue J**, Ismail M, Astre C, Saint-Aubert B, Joyeux H, Solassol C, Pujol H. Colorectal carcinoma in patients younger than 40 years of age. Montpellier Cancer Institute experience with 78 patients. *Cancer* 1988; **61**: 835-840 [PMID: 3338041]

43 **Armaghany T**, Wilson JD, Chu Q, Mills G. Genetic alterations in colorectal cancer. *Gastrointest Cancer Res* 2012; **5**: 19-27 [PMID: 22574233]

44 **Stigliano V**, Sanchez-Mete L, Martayan A, Anti M. Early-onset colorectal cancer: a sporadic or inherited disease? *World J Gastroenterol* 2014; **20**: 12420-12430 [PMID: 25253942 DOI: 10.3748/wjg.v20.i35.12420]

45 **Taylor DP**, Burt RW, Williams MS, Haug PJ, Cannon-Albright LA. Population-based family history-specific risks for colorectal cancer: a constellation approach. *Gastroenterology* 2010; **138**: 877-885 [PMID: 19932107 DOI: 10.1053/j.gastro.2009.11.044]

46 **Stoffel EM**. Colorectal Cancer in Young Individuals: Opportunities for Prevention. *J Clin Oncol* 2015; **33**: 3525-3527 [PMID: 26371141 DOI: 10.1200/JCO.2015.61.4503]

47 **Frampton GM**, Fichtenholtz A, Otto GA, Wang K, Downing SR, He J, Schnall-Levin M, White J, Sanford EM, An P, Sun J, Juhn F, Brennan K, Iwanik K, Maillet A, Buell J, White E, Zhao M, Balasubramanian S, Terzic S, Richards T, Banning V, Garcia L, Mahoney K, Zwirko Z, Donahue A, Beltran H, Mosquera JM, Rubin MA, Dogan S, Hedvat CV, Berger MF, Pusztai L, Lechner M, Boshoff C, Jarosz M, Vietz C, Parker A, Miller VA, Ross JS, Curran J, Cronin MT, Stephens PJ, Lipson D, Yelensky R. Development and validation of a clinical cancer genomic profiling test based on massively parallel DNA sequencing. *Nat Biotechnol* 2013; **31**: 1023-1031 [PMID: 24142049 DOI: 10.1038/nbt.2696]

48 **Fang JY**, Richardson BC. The MAPK signaling pathways and colorectal cancer. *Lancet Oncol* 2005; **6**: 322- 327 [PMID: 15863380]

49 **Weinberg RA.** The Biology of Cancer. Baltimore, MD: Garland Science, 2006

50 **Grady WM**, Pritchard CC. Molecular alterations and biomarkers in colorectal cancer. *Toxicol Pathol* 2014; **42**: 124-139 [PMID: 24178577 DOI: 10.1177/0192623313505155]

51 **Lengauer C**, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. *Nature* 1998; **396**: 643-649 [PMID: 9872311]

52 **Powell SM**, Zilz N, Beazer-Barclay Y, Bryan TM, Hamilton SR, Thibodeau SN, Vogelstein B, Kinzler KW. APC mutations occur early during colorectal tumorigenesis. *Nature* 1992; **359**: 235-237 [PMID: 1528264]

53 **Tan C**, Du X. KRAS mutation testing in metastatic colorectal cancer. *World J Gastroenterol* 2012; **18**: 5171-5180 [PMID: 23066310 DOI: 10.3748/wjg.v18.i37.5171]

54 **Bardhan K**, Liu K. Epigenetics and colorectal cancer pathogenesis. *Cancers* (Basel) 2013; **5**: 676-713 [PMID: 24216997 DOI: 10.3390/cancers5020676]

55 **Geiersbach KB**, Samowitz WS. Microsatellite instability and colorectal cancer. *Arch Pathol Lab Med* 2011; **135**: 1269-1277 [PMID: 21970482 DOI: 10.5858/arpa.2011-0035-RA]

56 **Iacopetta B**, Li WQ, Grieu F, Ruszkiewicz A, Kawakami K. BRAF mutation and gene methylation frequencies of colorectal tumours with microsatellite instability increase markedly with patient age. *Gut* 2006; **55**: 1213-1214 [PMID: 16849360 DOI: 10.1136/gut.2006.095455]

57 **Kinzler KW**, Vogelstein B. Landscaping the cancer terrain. *Science* 1998; **280**: 1036-1037 [PMID: 9616081]

58 **Raskov H**, Pommergaard HC, Burcharth J, Rosenberg J. Colorectal carcinogenesis--update and perspectives. *World J Gastroenterol* 2014; **20**: 18151-18164 [PMID: 25561783 DOI: 10.3748/wjg.v20.i48.18151]

59 **Newmark HL**, Lupton JR. Determinants and consequences of colonic luminal pH: implications for colon cancer. *Nutr Cancer* 1990; **14**: 161-173 [PMID: 1964727 DOI: 10.1080/01635589009514091]

60 **Aune D**, Chan DS, Lau R, Vieira R, Greenwood DC, Kampman E, Norat T. Dietary fibre, whole grains, and risk of colorectal cancer: systematic review and dose-response meta-analysis of prospective studies. *BMJ* 2011; **343**: d6617 [PMID: 22074852 DOI: 10.1136/bmj.d6617]

61 **McKay A**, Donaleshen J, Helewa RM, Park J, Wirtzfeld D, Hochman D, Singh H, Turner D. Does young age influence the prognosis of colorectal cancer: a population-based analysis. *World J Surg Oncol* 2014; **12**: 370 [PMID: 25466394 DOI: 10.1186/1477-7819-12-370]

62 **Schellerer VS**, Merkel S, Schumann SC, Schlabrakowski A, Förtsch T, Schildberg C, Hohenberger W, Croner RS. Despite aggressive histopathology survival is not impaired in young patients with colorectal cancer: CRC in patients under 50 years of age. *Int J Colorectal Dis* 2012; **27**: 71-79 [PMID: 21881876 DOI:

10.1007/s00384-011-1291-8]
63 **Park HC**, Shin A, Kim BW, Jung KW, Won YJ, Oh JH, Jeong SY, Yu CS, Lee BH. Data on the characteristics and the survival of korean patients with colorectal cancer from the Korea central cancer registry. *Ann Coloproctol* 2013; **29**: 144-149 [PMID: 24032114 DOI: 10.3393/ac.2013.29.4.144]
64 **Ruiz R**, Taxa L, Casanova L, Ruiz E, Montenegro P. Clinicopathologic features and survival outcomes of colorectal cancer in young patients: Experience from a cancer institute in Peru. *Ann Oncol* 2015; **26**: 70-71 [DOI: 10.1093/annonc/mdv233.240]
65 **Wang MJ**, Ping J, Li Y, Adell G, Arbman G, Nodin B, Meng WJ, Zhang H, Yu YY, Wang C, Yang L, Zhou ZG, Sun XF. The prognostic factors and multiple biomarkers in young patients with colorectal cancer. *Sci Rep* 2015; **5**: 10645 [PMID: 26013439 DOI: 10.1038/srep10645]
66 **Chan KK**, Dassanayake B, Deen R, Wickramarachchi RE, Kumarage SK, Samita S, Deen KI. Young patients with colorectal cancer have poor survival in the first twenty months after operation and predictable survival in the medium and long-term: analysis of survival and prognostic markers. *World J Surg Oncol* 2010; **8**: 82 [PMID: 20840793 DOI: 10.1186/1477-7819-8-82]
67 **Geiger TM**, Ricciardi R. Screening options and recommendations for colorectal cancer. *Clin Colon Rectal Surg* 2009; **22**: 209-217 [PMID: 21037811 DOI: 10.1055/s-0029-1242460]
68 **Levin B**, Lieberman DA, McFarland B, Smith RA, Brooks D, Andrews KS, Dash C, Giardiello FM, Glick S, Levin TR, Pickhardt P, Rex DK, Thorson A, Winawer SJ. Screening and surveillance for the early detection of colorectal cancer and adenomatous polyps, 2008: a joint guideline from the American Cancer Society, the US Multi-Society Task Force on Colorectal Cancer, and the American College of Radiology. *CA Cancer J Clin* 2008; **58**: 130-160 [PMID: 18322143 DOI: 10.3322/CA.2007.0018]
69 **Geller AC**, Prout MN, Miller DR, Siegel B, Sun T, Ockene J, Koh HK. Evaluation of a cancer prevention and detection curriculum for medical students. *Prev Med* 2002; **35**: 78-86 [PMID: 12079444 DOI: 10.1006/pmed.2002.1044]
70 **Al-Naggar RA**, Bobryshev YV. Knowledge of colorectal cancer screening among young Malaysians. *Asian Pac J Cancer Prev* 2013; **14**: 1969-1974 [PMID: 23679301 DOI: 10.7314/APJCP.2013.14.3.1969]
71 **Liu G**, Perkins A. Using a lay cancer screening navigator to increase colorectal cancer screening rates. *J Am Board Fam Med* 2015; **28**: 280-282 [PMID: 25748770 DOI: 10.3122/jabfm.2015.02.140209]
72 **Osborne JM**, Wilson C, Moore V, Gregory T, Flight I, Young GP. Sample preference for colorectal cancer screening tests: Blood or stool? *Open J Preventat Med* 2012; **2**: 326-331 [DOI: 10.4236/ojpm.2012.23047]
73 **Church TR**, Wandell M, Lofton-Day C, Mongin SJ, Burger M, Payne SR, Castaños-Vélez E, Blumenstein BA, Rösch T, Osborn N, Snover D, Day RW, Ransohoff DF. Prospective evaluation of methylated SEPT9 in plasma for detection of asymptomatic colorectal cancer. *Gut* 2014; **63**: 317-325 [PMID: 23408352 DOI: 10.1136/gutjnl-2012-304149]

P- Reviewer: Martinez JD, Park JH, Yaeger R **S- Editor**: Qi Y
L- Editor: A **E- Editor**: Lu YJ

World Journal of Gastrointestinal Oncology

Submit a Manuscript: http://www.wjgnet.com/esps/
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
DOI: 10.4251/wjgo.v8.i6.489

World J Gastrointest Oncol 2016 June 15; 8(6): 489-497
ISSN 1948-5204 (online)

TOPIC HIGHLIGHT

2016 Gastric Cancer: Global view

On the road to standardization of D2 lymph node dissection in a European population of patients with gastric cancer

Roman Yarema, Giovanni de Manzoni, Taras Fetsych, Myron Ohorchak, Mykhailo Pliatsko, Maria Bencivenga

Roman Yarema, Taras Fetsych, Department of Oncology and Medical Radiology, Lviv National Medical University named after Danylo Halytskyi, 79010 Lviv, Ukraine

Giovanni de Manzoni, Maria Bencivenga, Department of Surgery, General and Upper G.I. Surgery Division, University of Verona, 37126 Verona, Italy

Myron Ohorchak, Department of Abdominal Surgery, Lviv Regional State Cancer Diagnostic and Therapeutic Center, 79000 Lviv, Ukraine

Mykhailo Pliatsko, Department of Endoscopic, Lviv Regional Clinical Diagnostic Center, 79010 Lviv, Ukraine

Author contributions: Yarema R performed the research and wrote the paper; de Manzoni G, Fetsych T, Ohorchak M, Pliatsko M and Bencivenga M contributed critical revision of the manuscript for important intellectual content.

Conflict-of-interest statement: No conflict of interest.

Correspondence to: Roman Yarema, MD, PhD, Department of Oncology and Medical Radiology, Lviv National Medical University named after Danylo Halytskyi, Pekarska str., 69, 79010 Lviv, Ukraine. yaremarom@rambler.ru
Telephone: +38-67-9406933
Fax: +38-32-2757632

Received: January 15, 2016
Peer-review started: January 18, 2016
First decision: February 22, 2016
Revised: March 24, 2016
Accepted: April 14, 2016

Article in press: April 18, 2016
Published online: June 15, 2016

Abstract

The amount of lymph node dissection (LD) required during surgical treatment of gastric cancer surgery has been quite controversial. In the 1970s and 1980s, Japanese surgeons developed a doctrine of aggressive preventive gastric cancer surgery that was based on extended (D2) LD volumes. The West has relatively lower incidence rates of gastric cancer, and in Europe and the United States the most common LD volume was D0-1. This eventually caused a scientific conflict between the Eastern and Western schools of surgical thought: Japanese surgeons determinedly used D2 LD in surgical practice, whereas European surgeons insisted on repetitive clinical trials in the European patient population. Today, however, one can observe the results of this complex evolution of views. The D2 LD is regarded as an unambiguous standard of gastric cancer surgical treatment in specialized European centers. Such a consensus of the Eastern and Western surgical schools became possible due to the longstanding scientific and practical search for methods that would help improve the results of gastric cancer surgeries using evidence-based medicine. Today, we can claim that D2 LD could improve the prognosis in European populations of patients with gastric cancer, but only when the surgical quality of LD execution is adequate.

Key words: Gastric cancer; D2 lymph node dissection; Evidence-based medicine; European patients; Regional lymph nodes

Core tip: The amount of lymph node dissection required

during surgical treatment of gastric cancer has been quite controversial. We can now claim that D2 lymph node dissection improves the prognosis in European populations with gastric cancer, but only when the surgical quality of the lymph node dissection execution is adequate.

Yarema R, de Manzoni G, Fetsych T, Ohorchak M, Pliatsko M, Bencivenga M. On the road to standardization of D2 lymph node dissection in a European population of patients with gastric cancer. *World J Gastrointest Oncol* 2016; 8(6): 489-497 Available from: URL: http://www.wjgnet.com/1948-5204/full/v8/i6/489.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i6.489

INTRODUCTION

Radical surgery for malignant tumors traditionally includes mandatory one-piece removal of regional lymph nodes (LNs). This approach was introduced over 100 years ago by an American surgeon, W.S. Halsted, and has been used to determine the extent of surgery in basic sites of neoplasia including tumors in the gastrointestinal tract. Despite its high clinical effectiveness and use as a standard treatment in Asia, extensive D2/D3 lymph node dissection (LD) has not been widely used in gastric cancer (GC) surgery in Europe and the Americas until recently.

Indeed until recently, European clinical recommendations for cancer treatment did not suggest D2 LD as a surgical standard of care[1]. The relevance of this issue is also evident when considering the surgical standard of Western randomized trials on multimodal treatment for GC. The MAGIC trial set the standard for combined treatment of GC in the European Union, and D2 LD was performed in only 42.5% of patients[2]. The US standard multimodal treatment for GC is based on the INT 0116 trial[3] in which an extended LD was performed in only 10% of patients. In a large-scale clinical trial on perioperative chemoradiotherapy effectiveness (the CRITICS trial; ongoing in Europe), the planned extension of LD is more limited than D2[4]. Thus, the issue of standardization in lymphadenectomy extension for GC in Western countries remains relevant.

DEFINITION AND LEVELS OF LYMPHNODAL DISSECTION IN GASTRIC CANCER

Lymphatic efflux from the stomach travels through a complex multidirectional network[5]. Lymph from different sections of the stomach is drained into the para-aortal LN collector through one of four routes: (1) left subdiaphragmatic *via* the LN in the circulation of the left lower diaphragmatic artery; (2) abdominal *via* the LN along the left gastric, splenic, and common hepatic arteries and the celiac trunk; (3) upper mesenteric that receives lymph from the subpyloric LNs and runs along the upper mesenteric artery; and (4) retropancreatic, which is associated with LNs of the hepatoduodenal ligament, upper mesenteric vessels and common hepatic artery. Both the left subdiaphragmatic and abdominal routes drain lymph from the upper third of the stomach. The lymphatic efflux from the gastric body drains primarily through the abdominal route, and lymph efflux from the distal stomach drains through abdominal, upper mesenteric and retropancreatic routes[6].

Metastases to regional LNs are diagnosed in 37%-65% of patients with tumors in the gastric corpus, in 44%-80% of patients with tumors in the proximal stomach, and in 50%-59% of patients with tumors in the distal stomach[7,8]. The involvement of regional LNs depends directly on the depth of primary tumor invasion. In intra- and sub-epithelial tumors, regional lymphogenous metastases are diagnosed in 0%-5.5% and 19%-31% of patients, respectively[7,9]. In muscle or subserosal layer invasions, regional LN involvement increases to 30%-62%; in serous membrane tumors, regional LN metastases are found in 74% of patients, and 90%-91% in cases with infiltration of adjacent organs[7].

The first one-piece tissue dissection of regional lymphogenous metastasis during the course of GC surgery was carried out in 1962 by Jinnai *et al*[10]. Since then, the concept of extended radical LD has become an essential stage in the strategy of GC surgical treatment in Japan. Research in the field of lymph node (LN) topography and extended clinical efficiency formed the basis of the first edition of "General Rules for the Gastric Cancer Study", which was published in the early 1960s under the auspices of the Japanese Research Society for Gastric Cancer[11]. The first English edition of these guidelines was published in Europe in 1995. Subsequently, research performed by the Japanese Gastric Cancer Association (JGCA) formed the basis for a second English edition based on the Japanese classification of gastric cancer by the JGCA[12] as well as Japanese gastric cancer treatment guidelines[13]. These guidelines describe the following groups of stomach LNs (Table 1, Figure 1).

According to the classification of gastric cancer by the JGCA (1998)[12], the stomach lymphatic system consists of three LN compartments. Each of these is a temporary barrier that prevents tumor cells from entering the lymphatic system. Grouping stomach lymph collectors into compartments created the basis for determining the gradation of category "N" at staging and a theoretical basis for the extension of LD according to tumor site as reported in the following table (Table 2)[12]. The LN groups 12b, p and above are classified as N3 - in the given classification-this is equivalent to distant metastases.

Of note, in the last version of tumor-node-metastasis (TNM) classification introduced by the Union for International Cancer Control (UICC)[14], category "N" is determined not by the topography but rather by the number of affected regional LNs. Accordingly, in the last version of JGCA guidelines (2011)[13], the extension of nodal dissection is defined according to the extension of

Table 1 The lymphatic system of the stomach[12]

LN groups	LN topography
№1	Right paracardiac LNs
№2	Left paracardiac LNs
№3	LNs along the lesser curvature
№4sa	LNs along the short gastric vessels
№4sb	LNs along the left gastroepiploic vessels
№4d	LNs along the right gastroepiploic vessels
№5	Suprapyloric LNs
№6	Infrapyloric LNs
№7	LNs along the left gastric artery
№8a	LNs along the common hepatic artery (anterosuperior group)
№9	LNs at the celiac trunk
№10	LNs at the splenic hilum
№11p	LNs along the proximal splenic artery
№11d	LNs along the distal splenic artery
№12a	LNs in the hepatoduodenal ligament (along the hepatic artery)
№12b	LNs in the hepatoduodenal ligament (along the bile duct)
№12p	LNs in the hepatoduodenal ligament (behind the portal vein)
№13	Retro-pancreaticoduodenal LNs
№14a	LNs along the superior mesenteric artery
№14v	LNs along the superior mesenteric vein
№15	LNs along the middle colic vessels
№16	Para-aortic LNs
№17	LNs on the anterior surface of the pancreatic head
№18	LNs along the inferior margin of the pancreas
№19	Infradiaphragmatic LNs
№20	LNs in the esophageal hiatus of the diaphragm

LNs: Lymph nodes.

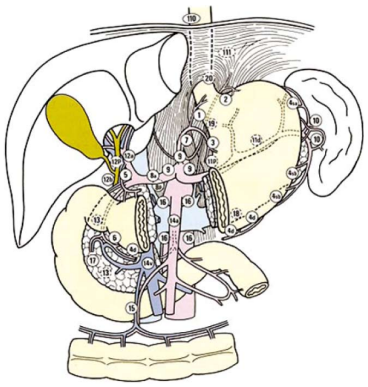

Figure 1 Topography of stomach lymph node groups[12].

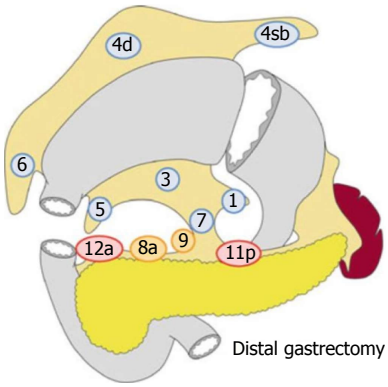

Figure 2 Lymph node dissection levels in distal subtotal gastrectomy[13].

gastric resection as reported in the following figures.

During distal subtotal gastrectomy, the lymph node dissection levels are as follows: (1) D0: LD in a volume less than D1; (2) D1: №1, 3, 4sb, 4d, 5, 6, 7; (3) D1 +: D1 plus №8a, 9; and (4) D2: D1 plus №8a, 9, 11p, 12a (Figure 2).

In gastrectomy, the LD levels are as follows: (1) D0: LD in a volume less than D1; (2) D1: №1-7; (3) D1 +: D1 plus №8a, 9, 11p; and (4) D2: D1 plus №8a, 9, 10, 11p, 11d, 12a (Figure 3).

Levels of LD in proximal subtotal gastrectomy: (1) D0: LD in a volume less than D1; (2) D1: №1, 2, 3a, 4sa, 4sb, 7; and (3) D1 +: D1 plus №8a, 9, 11p (Figure 4).

LD extended beyond these definitions are classified as D2 +. Their effectiveness remains controversial; therefore, they are currently not recommended for routine use in clinical practice[13].

Gastric cancer classification by JGCA (1998) has demonstrated its high efficiency in several clinical studies[5,15,16]. LN staging based on topography laid the grounds for JGCA's classification. These are considered anatomical in contrast to the rather mechanistic quantitative approach of the UICC classification. This allows for consideration of disease propagation and for more accurate prognosis. In support of this thesis, the correlated survival of patients with lesions of various LN groups has been studied patients with the same number of regional lymphogenous metastases, survival differed depending on the LN collectors in which lesions were located[17]. Thus, localization as well as the quantity of metastatically-affected regional LNs has a probable prognostic value. According to Y. Noguchi[18], in N0, LN lesion groups 1-6 (N1 according JGCA), LN lesion groups 7-12 (N2), and LN groups 13-16 (N3), the 5-year survival rate was 85%, 60%, 25% and 11%, respectively.

A significant advantage of the second JGCA gastric cancer classification in terms of practical application is its direct link with the volume of LD based on the staging principle of lymphogenous metastasis. Of note, the Japanese classification uses the term "regional lymph node". This is defined not only by the lymph node topography, but also by the site of the primary tumor in the stomach; the UICC classification does not provide this differentiation.

Another obvious advantage of the classification offered by JGCA[12] lies in the possibility of extrapolating data about the regional LN condition into the UICC classification. The reverse conversion is not possible; therefore, it is not possible to conduct a comparative analysis of retrospective studies in a different series.

Western pathologists and surgeons criticize the Japanese GC classification mainly because of its complexity and also because precision mapping is laborious in practice. However, the Eastern and Western GC classifications are finally approaching each other. This tendency can be observed in the latest edition of the

Table 2 Lymph node groups (compartments 1-3) by location of tumor

Location lymph node station	LMU/MUL MLU/UML	LD/L	LM/M/ML	MU/UM	U	E+
No. 1 rt paracardial	1	2	1	1	1	
No. 2 lt paracardial	1	M	3	1	1	
No. 3 lesser curvature	1	1	1	1	1	
No. 4sa short gastric	1	M	3	1	1	
No. 4sb lt gastroepiploic	1	3	1	1	1	
No. 4d rt gastroepiploic	1	1	1	1	2	
No. 5 suprapyloric	1	1	1	1	3	
No. 6 infrapyloric	1	1	1	1	3	
No. 7 lt gastric artery	2	2	2	2	2	
No. 8a ant comm hepatic	2	2	2	2	2	
No. 8b post comm hepatic	3	3	3	3	3	
No. 9 celiac artery	2	2	2	2	2	
No. 10 splenic hilum	2	M	3	2	2	
No. 11p proximal splenic	2	2	2	2	2	
No. 11d distal splenic	2	M	3	2	2	
No. 12a lt hepatoduodenal	2	2	2	2	3	
No. 12b,p post hepatoduod	3	3	3	3	3	
No. 13 retropancreatic	3	3	3	M	M	
No. 14v sup mesenteric v.	2	2	3	3	M	
No. 14a sup mesenteric a.	M	M	M	M	M	
No. 15 middle colic	M	M	M	M	M	
No. 16a1 aortic hiatus	M	M	M	M	M	
No. 16a2,b1 paraaortic, middle	3	3	3	3	3	
No. 16b2 paraaortic, caudal	M	M	M	M	M	
No. 17 ant pancreatic	M	M	M	M	M	
No. 18 inf pancreatic	M	M	M	M	M	
No. 19 infradiaphragmatic	3	M	M	3	3	2
No. 20 esophageal hiatus	3	M	M	3	3	1
No. 110 lower paraesophag	M	M	M	M	M	3
No. 111 supradiaphragmatic	M	M	M	M	M	3
No. 112 post mediastinal	M	M	M	M	M	3

M: Lymph nodes regarded as distant metastasis.

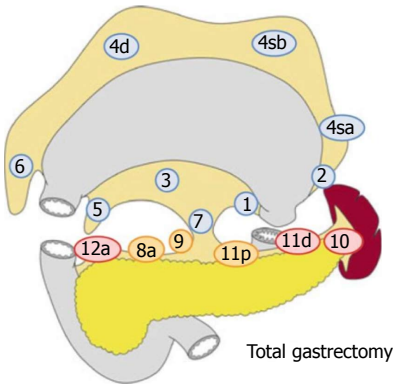

Figure 3 Lymph node dissection levels in gastrectomy[13].

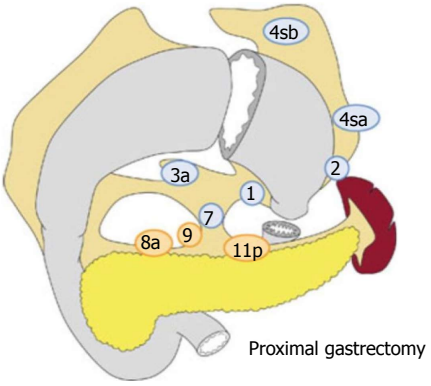

Figure 4 Lymph node dissection levels in proximal subtotal gastrectomy[13].

TNM UICC classification and the latest editions of the JCGA gastric cancer treatment guidelines[13,14].

DEBATE ON THE EXTENT OF LYMPHNODAL DISSECTION: EASTERN *VS* WESTERN POSITION

Results of a retrospective analysis of LD D2 were first published in Japan in 1970 by Mine *et al*[19]. The authors reported a slight increase in the survival rate among patients with pN0 and a probable increase in the 5-year survival rate from 10% to 21% in the group pN+. Similar results were reported in a study by Kodama *et al*[20], who indicated an increase in the 5-year survival rate from 33% to 58% in the entire group of patients.

In the 1970s and 1980s, Japanese surgeons developed a doctrine of aggressive preventive GC surgery based on the extended (D2) and super-extended (D3) LD volumes[21]. Concurrently, in Europe and the United States, the most common LD volume was D0-1. Due to the relatively lower GC incidence rates in the West,

European and American surgeons continued to reframe the ideology and master the techniques of extended interventions in GC cases until the end of the 1990s. This eventually caused a scientific conflict between the Eastern and Western schools of surgical thought. Japanese surgeons used D2 LD in surgical practice, whereas European surgeons insisted on repetitive clinical trials in the European patient population. They reasoned that certain biological differences in GC were present in the "Eastern" type[22].

One of the most significant publications from that time was a study of a European population of patients with GC by Pacelli *et al*[23]. The authors reported a probable increase in the 5-year survival rate from 30% (D1, LD) to 49% (D2, 3 LD) for patients with stage Ⅲ GC and from 50% to 65% in the entire group of patients.

Similar results were obtained by a group of German surgeons supervised by Siewert *et al*[24] during the course of a prospective multicentric trial of nearly 2500 patients. A probable increase in the survival rate was reported in patients with stages Ⅱ-ⅢA GC. However, in patients with pN2 (TNM UICC) or with extensive tumor invasion of the gastric serosa, D2 LD was not associated with increased survival.

Over time, researchers increasingly noted the low credibility of non-randomized studies. The results of the first randomized trials published by Dent *et al*[25] and Robertson *et al*[26] featured high rates of postoperative complications and mortality. However, the results did not provide high levels of credibility because of the small numbers of patients enrolled. The first large-scale randomized multicentric study of the efficacy of D2 LD in a population of European patients with GC was carried out in the 1990s.

This study, known as the Dutch trial[27], involved 1078 randomized patients and was organized by the Dutch Gastric Cancer Group. At the same time, the British MRS (Medical Research Society) carried out its own trial[28] with 400 randomized patients. The first results of these studies were preliminarily published in 1997 at the Second International Gastric Cancer Congress (IGCC) in Munich. However, the necessity of compliance with the full volume of D2 LD dramatically increased the frequency of splenectomies (up to 37% in the Dutch study and up to 65% in the British) and resections of the pancreas (30% in the Dutch study and 56% in the British) in all groups. These studies showed a dramatic increase in the number of postoperative complications after D2 LD (from 25% after performing D0-1 in the control group up to 43% in the Dutch trial and from 28% to 46% in the British trial). They also showed an increase in the postoperative mortality rate (from 4% to 10% in the Dutch trial and from 6.5% to 13% in the British trial)[27,28]. In the Eastern Asian series however, the rate of postoperative complications was 17%-21%[29,30]. The postoperative mortality rate after D2 LD in Eastern clinics was also significantly lower than in Europe-less than 2% in the Japanese nationwide registry[31] and less than 1%[30] or even zero[29] in specialized centers.

After a 5-year follow-up of European randomized studies, the expected increase in survival of D2 LD group was not achieved; the 5-year survival in the Dutch trial was 45% in group D1 LD and 47% in group D2 LD. In the British trial, it was 35% in group D1 LD and 33% in group D2 LD[32,33] (Figure 5).

Thus, the European oncology society preliminarily concluded that the extended LD volumes used in European GC patients were ineffective. This was based on evidence-based medicine and relied on the results of the two major Western randomized trials. However, a detailed analysis of this study and all potential reasons for the lack of a positive result were shown at the 1999 IGCC in Seoul. The summary of this analysis was later published in the *New England Journal of Medicine*[34]. Despite a good design and detailed statistical analysis, the study had some serious shortcomings that made the results ambiguous. These included:

The large number of participating surgical centers (about 80 clinics), which resulted in surgeons obtaining an insufficient amount of practical experience in the surgical procedures required for the study. For instance, some surgeons performed fewer than 5 D2 LD surgeries per year. This not only potentially affected the level of postoperative complications and mortality, but also led to a reduction in LN removal in the course of D2 LD and consequently to a reduction in radical surgeries[34].

There was a lack of surgery standardization (there were no clear criteria for splenectomy or spleen-saving dissection of the 10th LN group, instrumental or manual anastomosis, *etc.*).

Conversely, surgeons participating in the randomized trial in Taiwan performed a minimum of 80 D2 LD surgeries before the study began. The results of that study revealed a possible increase in survival rates when extended volumes of LD were performed[35].

The median number of LNs removed is an important indicator of LD quality. Significant geographic fluctuations of this indicator in the performance of D2 LD have now been established. There are diametrically polar indicators in European randomized trials. In the British study, the median number of removed LNs was 17[28]; in the Dutch study, the number was 30[32]. There were 25-26 LNs removed in the Western retrospective studies[36,37] and 54 LNs removed in Japanese specialized centers[30]. The minimum adequate number of LNs to be removed in gastric cancer surgeries-according to the requirements of TNM UICC (2009)[14]-is 15. This level of LD was provided in 86%[36] to 95%[37] of patients in the Western retrospective studies and in 100% of patients in the Japanese studies[30]. According to Siewert *et al*[24], the efficiency of LD execution can meet the standards of D2 only when a minimum of 26 LNs are removed.

The average frequency of metastatic lesions in LNs of group №10th (LNs of the splenic hilum) in various tumor sites in the stomach is 8.8%. Metastatic lesions in these LNs are likely to worsen the prognosis[38]. The application of splenectomy on principle including for LN dissection of the 10th group was not effective in patients

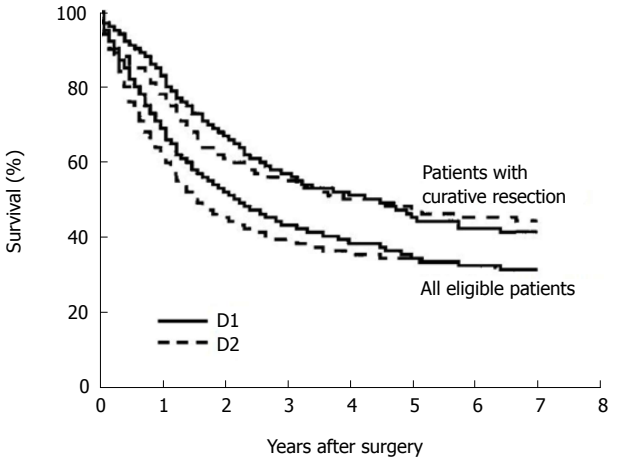

Figure 5 Patient survival in the Dutch trial[32].

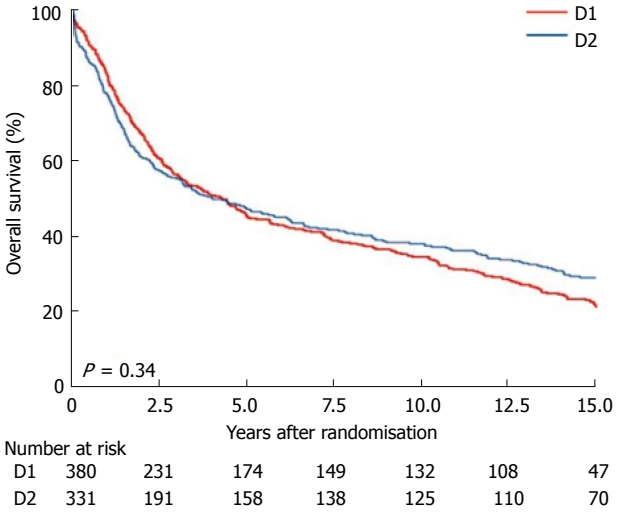

Figure 6 Survival of patients in the Dutch trial after a 15-year observation[43].

with GC until recently. A small study conducted in Korea by Yu *et al*[39] demonstrated a tendency toward increased survival after splenectomy; however, this result was not statistically significant. A meta-analysis conducted in 2009 by Yang *et al*[40] also confirmed an increase in the 5-year survival rate of patients with GC after splenectomy. According to other authors[38], unless the tumor has invaded the spleen, splenectomy is necessary only in case of LN lesions in group №4sa. Therefore, despite the fact that LN dissection of the 10th group is regulated by the JGCA guidelines (2011)[13], the role of splenectomy as a standard stage of D2 LD remains controversial. The answer to this question will likely be clarified soon after the publication of the results of a large randomized trial investigating the efficacy of splenectomy in Japanese patients with cancer of the upper third of the stomach (JCOG 0110 that began in Japan in 2002)[41].

Despite the previous pessimistic results, Hartgrink *et al*[42] conducted a second analysis of the "Dutch material" in 2001. They found a significant increase in survival in group D2 LD, especially in patients with metastases in LNs of the first stage of metastasis (N1 by JGCA). After 15 years of observation of patients during the Dutch trial, no significant difference in survival between groups under observation has not been noted. However, when the most controversial group of patients with splenectomies and resection of the pancreatic gland was excluded from the analysis, the 15-year survival rate increased dramatically from 22% in D1 LD to 35% in D2 LD (P = 0.006)[43] (Figure 6).

In 2013, the results of meta-analysis obtained by 12 randomized controlled major European trials on LD D2 effectiveness were published. These clearly proved the thesis concerning an increased risk of postoperative complications with D2 LD and the possible increase in survival only in the group that did not have splenectomy and resection of the pancreatic gland[44]. Therefore, in the latest European oncology guidelines, D2 LD is the standard surgical procedure but only in highly specialized centers with extensive experience in such surgeries as well as postoperative care[45].

According to the Japanese guidelines on the gastric cancer treatment issued by JGCA (2011)[13], the algorithm of surgical treatment in patients with GC is as follows (Figure 7).

The amount of LD required during surgical treatment of gastric cancer surgery has been quite controversial. Today, however, in light of evidence-based medicine, one can observe the results of this complex evolution of views: D2 LD is considered an unambiguous standard of GC surgical treatment in specialized centers according to national recommendations in Germany[46], the United Kingdom[47] and Italy[48] as well as mutual recommendations of the European Society of Medical Oncologists, Surgical Oncologists and Radiation Therapists (ESMO-ESSO-ESTRO)[45]. Such a consensus of the Eastern and Western surgical schools became possible due to the longstanding scientific and practical search for methods that would help improve the results of GC surgeries using evidence-based medicine[49]. In Western surgical terminology, D2 LD is now called a standard volume of intervention, whereas D2 + LD is an extended operation.

This debate into the effectiveness of extended (D2 + LD) interventions in GC cases remains open. A well-known clinical study conducted by Sasako *et al*[34] did not demonstrate an increase in survival after D2 + para-aortic LD for patients with resectable GC. However, many recent studies have demonstrated the possibility of increased survival after the application of extended LD in a selected group of patients with a high risk of metastasis in LNs of the N°16 station[50,51].

Furthermore, the effectiveness of laparoscopic D2 LD in GC cases remains undetermined. Today, clinical research is underway in the KLASS-2 trial, which aims to determine the effectiveness of such interventions. The impact of interventions with D1 +, D2 and D2 + LD on the risk of intraperitoneal progression of GC after

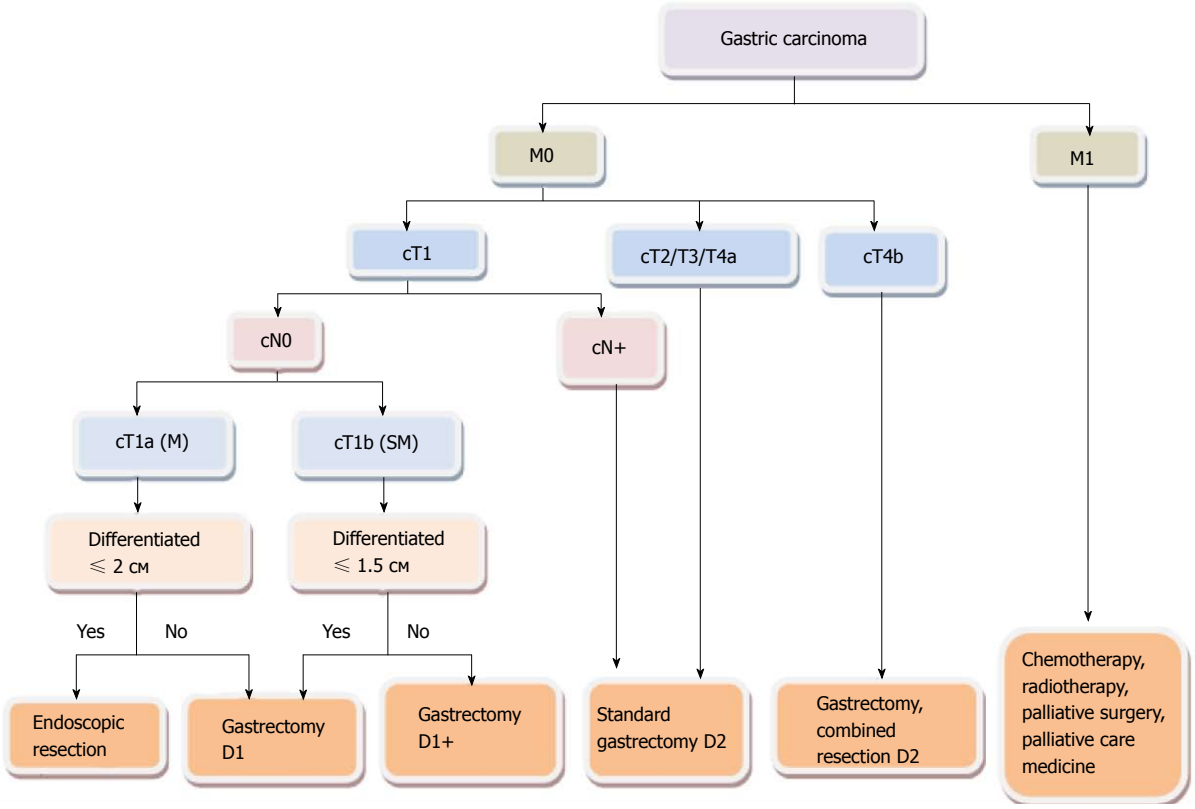

Figure 7 Algorithm of surgical treatment of patients with gastric cancer according to the guidelines provided by Japanese Gastric Cancer Association (2011)[13].

surgery[6] remains unknown.

CONCLUSION

The data show that D2 LD can improve the prognosis in European GC patients, but only when the surgical quality of LD execution is adequate. As part of the 10th IGCC in 2013 in Verona, Italy, the former president of *the European* Society of *Surgical Oncology*, Professor C. van de Velde, noted in his expert lecture that "the only way to improve the efficiency of surgical treatment of gastric cancer in Europe is to place patients in specialized surgical centers, provide training so that individual surgeons could specialize on the issue of LD D2 and an objective and permanent audit on quality of lymphadenectomy in each surgical center".

REFERENCES

1 **Okines A**, Verheij M, Allum W, Cunningham D, Cervantes A; ESMO Guidelines Working Group. Gastric cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2010; **21** Suppl 5: v50-v54 [PMID: 20555102 DOI: 10.1093/annonc/mdq164]

2 **Cunningham D**, Allum WH, Stenning SP, Thompson JN, Van de Velde CJ, Nicolson M, Scarffe JH, Lofts FJ, Falk SJ, Iveson TJ, Smith DB, Langley RE, Verma M, Weeden S, Chua YJ. Perioperative chemotherapy versus surgery alone for resectable gastroesophageal cancer. *N Engl J Med* 2006; **355**: 11-20 [PMID: 16822992 DOI: 10.1056/NEJMoa055531]

3 **Macdonald JS**, Smalley SR, Benedetti J, Hundahl SA, Estes NC, Stemmermann GN, Haller DG, Ajani JA, Gunderson LL, Jessup JM, Martenson JA. Chemoradiotherapy after surgery compared with surgery alone for adenocarcinoma of the stomach or gastroesophageal junction. *N Engl J Med* 2001; **345**: 725-730 [PMID: 11547741 DOI: 10.1056/NEJMoa010187]

4 **Dutch Colorectal Cancer Group**. Randomized phase III trial of adjuvant chemotherapy or chemoradiotherapy in resectable gastric cancer (CRITICS). In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available from: URL: https://clinicaltrials.gov/ct2/show/NCT00407186 NLM Identifier: NCT00407186

5 **Maruyama K**, Gunvén P, Okabayashi K, Sasako M, Kinoshita T. Lymph node metastases of gastric cancer. General pattern in 1931 patients. *Ann Surg* 1989; **210**: 596-602 [PMID: 2818028]

6 **de Manzoni G**, Roviello F, Siquini W. Surgery in the multimodal management of gastric cancer. Milan: Springer-Verlag Italia, 2012: 266

7 **Di Leo A**, Marrelli D, Roviello F, Bernini M, Minicozzi A, Giacopuzzi S, Pedrazzani C, Baiocchi LG, de Manzoni G. Lymph node involvement in gastric cancer for different tumor sites and T stage: Italian Research Group for Gastric Cancer (IRGGC) experience. *J Gastrointest Surg* 2007; **11**: 1146-1153 [PMID: 17576611 DOI: 10.1007/s11605-006-0062-2]

8 **Shen KH**, Wu CW, Lo SS, Hsieh MC, Hsia CY, Chiang SC, Lui WY. Factors correlated with number of metastatic lymph nodes in gastric cancer. *Am J Gastroenterol* 1999; **94**: 104-108 [PMID: 9934739 DOI: 10.1111/j.1572-0241.1999.00779.x]

9 **Gotoda T**, Yanagisawa A, Sasako M, Ono H, Nakanishi Y, Shimoda T, Kato Y. Incidence of lymph node metastasis from early gastric cancer: estimation with a large number of cases at two large centers. *Gastric Cancer* 2000; **3**: 219-225 [PMID: 11984739 DOI: 10.1007/PL00011720]

10 **Jinnai D**, Tanaka S. Technique of extended radical operation for gastric cancer. *Geka Chiryo* 1962; **7**: 316-324

11 **Japanese Research Society for Gastric Cancer**. The general rules for gastric cancer study. *Jpn J Surg* 1963; **16**: 121-123

12 **Japanese Gastric Cancer Association.** Japanese Classification of

Gastric Carcinoma - 2nd English Edition. *Gastric Cancer* 1998; **1**: 10-24 [PMID: 11957040 DOI: 10.1007/s101209800016]
13 **Japanese Gastric Cancer Association**. Japanese gastric cancer treatment guidelines 2010 (ver. 3). *Gastric Cancer* 2011; **14**: 113-123 [PMID: 21573742 DOI: 10.1007/s10120-011-0042-4]
14 **Sobin LH**, Gospodarowicz MK, Wittekind Ch. TNM classification of malignant tumors. 7th ed. Blackwell Publishing, 2010: 310
15 **de Manzoni G**, Verlato G, di Leo A, Guglielmi A, Laterza E, Ricci F, Cordiano C. Perigastric lymph node metastases in gastric cancer: comparison of different staging systems. *Gastric Cancer* 1999; **2**: 201-205 [PMID: 11957098 DOI: 10.1007/s101209900035]
16 **Maruyama K**, Sasako M, Kinoshita T, Sano T, Katai H. Surgical treatment for gastric cancer: the Japanese approach. *Semin Oncol* 1996; **23**: 360-368 [PMID: 8658220]
17 **Isozaki H**, Okajima K, Kawashima Y, Yamada S, Nakata E, Nishimura J, Ichinona T. Prognostic value of the number of metastatic lymph nodes in gastric cancer with radical surgery. *J Surg Oncol* 1993; **53**: 247-251 [PMID: 8341056 DOI: 10.1002/jso.2930530412]
18 **Noguchi Y**, Imada T, Matsumoto A, Coit DG, Brennan MF. Radical surgery for gastric cancer. A review of the Japanese experience. *Cancer* 1989; **64**: 2053-2062 [PMID: 2680049 DOI: 10.1002/1097-0142(19891115)64: 10<2053: : AID-CNCR2820641014>3.0.CO; 2-J]
19 **Mine M**, Majima S, Harada M, Etani S. End results of gastrectomy for gastric cancer: effect of extensive lymph node dissection. *Surgery* 1970; **68**: 753-758 [PMID: 5473423]
20 **Kodama Y**, Sugimachi K, Soejima K, Matsusaka T, Inokuchi K. Evaluation of extensive lymph node dissection for carcinoma of the stomach. *World J Surg* 1981; **5**: 241-248 [PMID: 7245793]
21 **Maeta M**, Yamashiro H, Saito H, Katano K, Kondo A, Tsujitani S, Ikeguchi M, Kaibara N. A prospective pilot study of extended (D3) and superextended para-aortic lymphadenectomy (D4) in patients with T3 or T4 gastric cancer managed by total gastrectomy. *Surgery* 1999; **125**: 325-331 [PMID: 10076618]
22 **Jatzko G**, Pertl A, Jagoditsch M. Chirurgische therapie und ergebnisse beim magenfrühkarzinom. *Chir Gastroenterol* 1999; **15**: 223-226 [DOI: 10.1159/000012561]
23 **Pacelli F**, Doglietto GB, Bellantone R, Alfieri S, Sgadari A, Crucitti F. Extensive versus limited lymph node dissection for gastric cancer: a comparative study of 320 patients. *Br J Surg* 1993; **80**: 1153-1156 [PMID: 8402119 DOI: 10.1002/bjs.1800800930]
24 **Siewert JR**, Böttcher K, Roder JD, Busch R, Hermanek P, Meyer HJ. Prognostic relevance of systematic lymph node dissection in gastric carcinoma. German Gastric Carcinoma Study Group. *Br J Surg* 1993; **80**: 1015-1018 [PMID: 8402053 DOI: 10.1002/bjs.1800800829]
25 **Dent DM**, Madden MV, Price SK. Randomized comparison of R1 and R2 gastrectomy for gastric carcinoma. *Br J Surg* 1988; **75**: 110-112 [PMID: 3349293 DOI: 10.1002/bjs.1800750206]
26 **Robertson CS**, Chung SC, Woods SD, Griffin SM, Raimes SA, Lau JT, Li AK. A prospective randomized trial comparing R1 subtotal gastrectomy with R3 total gastrectomy for antral cancer. *Ann Surg* 1994; **220**: 176-182 [PMID: 8053740]
27 **Bonenkamp JJ**, Songun I, Hermans J, Sasako M, Welvaart K, Plukker JT, van Elk P, Obertop H, Gouma DJ, Taat CW. Randomised comparison of morbidity after D1 and D2 dissection for gastric cancer in 996 Dutch patients. *Lancet* 1995; **345**: 745-748 [PMID: 7891484]
28 **Cuschieri A**, Fayers P, Fielding J, Craven J, Bancewicz J, Joypaul V, Cook P. Postoperative morbidity and mortality after D1 and D2 resections for gastric cancer: preliminary results of the MRC randomised controlled surgical trial. The Surgical Cooperative Group. *Lancet* 1996; **347**: 995-999 [PMID: 8606613]
29 **Wu CW**, Hsiung CA, Lo SS, Hsieh MC, Shia LT, Whang-Peng J. Randomized clinical trial of morbidity after D1 and D3 surgery for gastric cancer. *Br J Surg* 2004; **91**: 283-287 [PMID: 14991627 DOI: 10.1002/bjs.4433]
30 **Sano T**, Sasako M, Yamamoto S, Nashimoto A, Kurita A, Hiratsuka M, Tsujinaka T, Kinoshita T, Arai K, Yamamura Y, Okajima K. Gastric cancer surgery: morbidity and mortality results from a prospective randomized controlled trial comparing D2 and extended para-aortic lymphadenectomy--Japan Clinical Oncology Group study 9501. *J Clin Oncol* 2004; **22**: 2767-2773 [PMID: 15199090 DOI: 10.1200/JCO.2004.10.184]
31 **Fujii M**, Sasaki J, Nakajima T. State of the art in the treatment of gastric cancer: from the 71st Japanese Gastric Cancer Congress. *Gastric Cancer* 1999; **2**: 151-157 [PMID: 11957089 DOI: 10.1007/s101209900011]
32 **Bonenkamp JJ**, Hermans J, Sasako M, van de Velde CJ, Welvaart K, Songun I, Meyer S, Plukker JT, Van Elk P, Obertop H, Gouma DJ, van Lanschot JJ, Taat CW, de Graaf PW, von Meyenfeldt MF, Tilanus H. Extended lymph-node dissection for gastric cancer. *N Engl J Med* 1999; **340**: 908-914 [PMID: 10089184 DOI: 10.1056/NEJM199903253401202]
33 **Cuschieri A**, Weeden S, Fielding J, Bancewicz J, Craven J, Joypaul V, Sydes M, Fayers P. Patient survival after D1 and D2 resections for gastric cancer: long-term results of the MRC randomized surgical trial. Surgical Co-operative Group. *Br J Cancer* 1999; **79**: 1522-1530 [PMID: 10188901 DOI: 10.1038/sj.bjc.6690243]
34 **Sasako M**, Sano T, Yamamoto S, Kurokawa Y, Nashimoto A, Kurita A, Hiratsuka M, Tsujinaka T, Kinoshita T, Arai K, Yamamura Y, Okajima K. D2 lymphadenectomy alone or with para-aortic nodal dissection for gastric cancer. *N Engl J Med* 2008; **359**: 453-462 [PMID: 18669424 DOI: 10.1056/NEJMoa0707035]
35 **Wu CW**, Hsiung CA, Lo SS, Hsieh MC, Chen JH, Li AF, Lui WY, Whang-Peng J. Nodal dissection for patients with gastric cancer: a randomised controlled trial. *Lancet Oncol* 2006; **7**: 309-315 [PMID: 16574546 DOI: 10.1016/S1470-2045(06)70623-4]
36 **Smith BR**, Stabile BE. Aggressive D2 lymphadenectomy is required for accurate pathologic staging of gastric adenocarcinoma. *Am Surg* 2006; **72**: 849-852 [PMID: 17058719]
37 **de Manzoni G**, Verlato G, Guglielmi A, Laterza E, Genna M, Cordiano C. Prognostic significance of lymph node dissection in gastric cancer. *Br J Surg* 1996; **83**: 1604-1607 [PMID: 9014687 DOI: 10.1002/bjs.1800831137]
38 **Chen XL**, Yang K, Zhang WH, Chen XZ, Zhang B, Chen ZX, Chen JP, Zhou ZG, Hu JK. Metastasis, risk factors and prognostic significance of splenic hilar lymph nodes in gastric adenocarcinoma. *PLoS ONE* 2014; **9**: e99650 [PMID: 24915065 DOI: 10.1371/journal.pone.0099650]
39 **Yu W**, Choi GS, Chung HY. Randomized clinical trial of splenectomy versus splenic preservation in patients with proximal gastric cancer. *Br J Surg* 2006; **93**: 559-563 [PMID: 16607678 DOI: 10.1002/bjs.5353]
40 **Yang K**, Chen XZ, Hu JK, Zhang B, Chen ZX, Chen JP. Effectiveness and safety of splenectomy for gastric carcinoma: a meta-analysis. *World J Gastroenterol* 2009; **15**: 5352-5359 [PMID: 19908346 DOI: 10.3748/wjg.15.5352]
41 **Sano T**, Yamamoto S, Sasako M; Japan Clinical Oncology Group Study LCOG 0110-MF. Randomized controlled trial to evaluate splenectomy in total gastrectomy for proximal gastric carcinoma: Japan clinical oncology group study JCOG 0110-MF. *Jpn J Clin Oncol* 2002; **32**: 363-364 [PMID: 12417603 DOI: 10.1093/jjco/hyf085]
42 **Hartgrink HH**, van de Velde CJH; On behalf of the Dutch Gastric Cancer Group. Update of the Dutch D1 vs D2 gastric cancer trial. The 4th International Gastric Cancer Congress; 2001 April 29-May 2; New-York, USA
43 **Songun I**, Putter H, Kranenbarg EM, Sasako M, van de Velde CJ. Surgical treatment of gastric cancer: 15-year follow-up results of the randomised nationwide Dutch D1D2 trial. *Lancet Oncol* 2010; **11**: 439-449 [PMID: 20409751 DOI: 10.1016/S1470-2045(10)70070-X]
44 **Jiang L**, Yang KH, Guan QL, Zhao P, Chen Y, Tian JH. Survival and recurrence free benefits with different lymphadenectomy for resectable gastric cancer: a meta-analysis. *J Surg Oncol* 2013; **107**: 807-814 [PMID: 23512524 DOI: 10.1002/jso.23325]
45 **Waddell T**, Verheij M, Allum W, Cunningham D, Cervantes A, Arnold D. Gastric cancer: ESMO-ESSO-ESTRO Clinical Practice

Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2013; **24** Suppl 6: vi57-vi63 [PMID: 24078663 DOI: 10.1093/annonc/mdt344]

46 **Meyer HJ**, Hölscher AH, Lordick F, Messmann H, Mönig S, Schumacher C, Stahl M, Wilke H, Möhler M. [Current S3 guidelines on surgical treatment of gastric carcinoma]. *Chirurg* 2012; **83**: 31-37 [PMID: 22127381 DOI: 10.1007/s00104-011-2149-x]

47 **Allum WH**, Blazeby JM, Griffin SM, Cunningham D, Jankowski JA, Wong R; Association of Upper Gastrointestinal Surgeons of Great Britain and Ireland, the British Society of Gastroenterology and the British Association of Surgical Oncology. Guidelines for the management of oesophageal and gastric cancer. *Gut* 2011; **60**: 1449-1472 [PMID: 21705456 DOI: 10.1136/gut.2010.228254]

48 **De Manzoni G**, Baiocchi GL, Framarini M, De Giuli M, D'Ugo D, Marchet A, Nitti D, Marrelli D, Morgagni P, Rinnovati A, Rosati R, Roviello F, Allieta R, Berti S, Bracale U, Capelli P, Cavicchi A, Di Martino N, Donini A, Filippini A, Francioni G, Frascio M, Garofalo A, Giulini SM, Grassi GB, Innocenti P, Martino A, Mazzocconi G, Mazzola L, Montemurro S, Palasciano N, Pantuso G, Pernthaler H, Petri R, Piazza D, Sacco R, Sgroi G, Staudacher C, Testa M, Vallicelli C, Vettoretto N, Zingaretti C, Capussotti L, Morino M, Verdecchia GM. The SIC-GIRCG 2013 Consensus Conference on Gastric Cancer. *Updates Surg* 2014; **66**: 1-6 [PMID: 24523031 DOI: 10.1007/s13304-014-0248-1]

49 **Verlato G**, Giacopuzzi S, Bencivenga M, Morgagni P, De Manzoni G. Problems faced by evidence-based medicine in evaluating lymphadenectomy for gastric cancer. *World J Gastroenterol* 2014; **20**: 12883-12891 [PMID: 25278685 DOI: 10.3748/wjg.v20.i36.12883]

50 **Roviello F**, Pedrazzani C, Marrelli D, Di Leo A, Caruso S, Giacopuzzi S, Corso G, de Manzoni G. Super-extended (D3) lymphadenectomy in advanced gastric cancer. *Eur J Surg Oncol* 2010; **36**: 439-446 [PMID: 20392590 DOI: 10.1016/j.ejso.2010.03.008]

51 **de Manzoni G**, Di Leo A, Roviello F, Marrelli D, Giacopuzzi S, Minicozzi AM, Verlato G. Tumor site and perigastric nodal status are the most important predictors of para-aortic nodal involvement in advanced gastric cancer. *Ann Surg Oncol* 2011; **18**: 2273-2280 [PMID: 21286941 DOI: 10.1245/s10434-010-1547-5]

P- Reviewer: Gu GL, Inokuchi M, Park WS **S- Editor**: Gong ZM
L- Editor: A **E- Editor**: Lu YJ

World Journal of Gastrointestinal Oncology

Submit a Manuscript: http://www.wjgnet.com/esps/
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
DOI: 10.4251/wjgo.v8.i6.498

World J Gastrointest Oncol 2016 June 15; 8(6): 498-508
ISSN 1948-5204 (online)

REVIEW

Malignant biliary obstruction: From palliation to treatment

Brian R Boulay, Aleksandr Birg

Brian R Boulay, Division of Gastroenterology and Hepatology, Department of Medicine, University of Illinois Hospital and Health Sciences System, Chicago, IL 60612, United States

Aleksandr Birg, Department of Medicine, University of Illinois Hospital and Health Sciences System, Chicago, IL 60612, United States

Author contributions: All authors equally contributed to this paper with conception of the topic, literature review and analysis, drafting, critical revision and editing, and final approval of the final version.

Conflict-of-interest statement: No potential conflicts of interest or financial support.

Correspondence to: Brian R Boulay, MD, MPH, Division of Gastroenterology and Hepatology, Department of Medicine, University of Illinois Hospital and Health Sciences System, 840 S Wood Street, MC 716, Chicago, IL 60612, United States. bboulay@uic.edu
Telephone: +1-312-4131999
Fax: +1-312-4133798

Received: July 17, 2015
Peer-review started: July 19, 2015
First decision: September 17, 2015
Revised: March 14, 2016
Accepted: April 21, 2016
Article in press: April 23, 2016
Published online: June 15, 2016

Abstract

Malignant obstruction of the bile duct from cholangiocarcinoma, pancreatic adenocarcinoma, or other tumors is a common problem which may cause debilitating symptoms and increase the risk of subsequent surgery. The optimal treatment - including the decision whether to treat prior to resection - depends on the type of malignancy, as well as the stage of disease. Preoperative biliary drainage is generally discouraged due to the risk of infectious complications, though some situations may benefit. Patients who require neoadjuvant therapy will require decompression for the prolonged period until attempted surgical cure. For pancreatic cancer patients, self-expanding metallic stents are superior to plastic stents for achieving lasting decompression without stent occlusion. For cholangiocarcinoma patients, treatment with percutaneous methods or nasobiliary drainage may be superior to endoscopic stent placement, with less risk of infectious complications or failure. For patients of either malignancy who have advanced disease with palliative goals only, the choice of stent for endoscopic decompression depends on estimated survival, with plastic stents favored for survival of < 4 mo. New endoscopic techniques may actually extend stent patency and patient survival for these patients by achieving local control of the obstructing tumor. Both photodynamic therapy and radiofrequency ablation may play a role in extending survival of patients with malignant biliary obstruction.

Key words: Pancreatic neoplasms; Cholangiocarcinoma; Extrahepatic cholestasis; Stents; Catheter ablation

Core tip: Treatment of malignant biliary obstruction from cholangiocarcinoma or pancreatic cancer can be performed *via* endoscopic, percutaneous, or surgical means. The decision of when or how to achieve biliary decompression depends on the patient's condition, location of stricture, and stage of malignancy. Not all patients require biliary decompression, particularly with resectable tumors. Self-expanding metallic stents or plastic stents may be used for distal malignancy, depending on stage and prognosis. Stents, nasobiliary drainage, or percutaneous drains may

be used for hilar strictures. Endoscopic catheter-based therapies such as photodynamic therapy or radiofrequency ablation may prolong patient survival by achieving local tumor control.

Boulay BR, Birg A. Malignant biliary obstruction: From palliation to treatment. *World J Gastrointest Oncol* 2016; 8(6): 498-508 Available from: URL: http://www.wjgnet.com/1948-5204/full/v8/i6/498.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i6.498

INTRODUCTION

Obstruction of the extrahepatic bile ducts from a malignant process presents both a diagnostic and therapeutic challenge. It is a common problem, with as many of 70% of pancreatic cancer patients presenting with obstruction upon diagnosis[1]. Obstruction may serve as the initial sign of disease - such as in the classic presentation of painless jaundice in pancreatic ductal adenocarcinoma - or may occur during progression of malignancy once the diagnosis is established. The two most common malignant neoplasms known to occlude the bile ducts are pancreatic ductal adenocarcinoma and primary bile duct cancer (cholangiocarcinoma). Other causes of malignant biliary obstruction can include ampullary carcinoma, primary duodenal adenocarcinoma, pancreatic neuroendocrine tumors, or occlusion of the hepatic hilum due to lymphadenopathy at the porta hepatis (as seen in metastatic colon cancer or lymphoma). Of note, some premalignant lesions such as biliary papillomatosis may cause an obstructive picture similar to malignancy. Benign conditions such as autoimmune cholangiopathy must also be ruled out, so obtaining tissue *via* endoscopic retrograde cholangiography (ERCP) with brush biopsy or core biopsy, or endoscopic ultrasound with fine needle aspiration (FNA) is paramount[2]. Only once a firm diagnosis of malignancy is secured can the final choice of treatment be made.

Occlusion of the bile ducts may cause debilitating symptoms such as pruritus and malaise, and thus treatment is often recommended on that basis alone. This may come in the form of surgical resection if the patient presents with resectable disease. However, both pancreatic cancer and cholangiocarcinoma are notorious for presenting at an advanced stage in which immediate surgery is contraindicated. Treatment goals for these patients include downstaging of the tumor with chemoradiotherapy, or strictly palliative measures. Relief of biliary obstruction is recommended in either setting. Treatment of distal malignant biliary obstruction from pancreatic cancer is typically managed by an endoscopically placed single biliary prosthesis, whereas hilar strictures can be more challenging to manage due to the need to access the left and right systems of the biliary tree.

Within the past decade endoscopic techniques have been developed to treat tumor ingrowth into the bile duct with photodynamic therapy or radiofrequency ablation, and recent studies show promise in expanding the role of endoscopic treatment. While the primary role at this time is to provide biliary decompression and relieve jaundice, the ability to provide therapy for these tumors represents a major shift in the role of the endoscopist. This review will consider the options for management based on the location of obstruction, as well as the stage of the underlying malignancy.

THE EFFECT OF JAUNDICE

The decision whether to decompress obstructed bile ducts in a patient with resectable disease has traditionally been quite controversial. Jaundice has long been recognized as an important preoperative risk factor in the setting of malignancy[3,4]. Several mechanisms have been described through which jaundice exerts its negative effects. Jaundice is thought to impair cellular immunity, allowing tumor growth and metastasic progression if left untreated[5]. In addition, obstruction to flow of bile decreases its availability in the enteric system for the absorption of lipid-soluble vitamins, including vitamin K, leading to coagulopathy and increased surgical bleeding risk. Even additional administration of oral vitamin K may be inadequate to reach appropriate levels of coagulation in obstructive cases[6], which may further complicate any planned surgery. Bacterial and endotoxin translocation through the intestinal mucosa has also been demonstrated in jaundiced patients, making SIRS and sepsis a serious complication that can develop even prior to surgery[6]. Jaundice has also been shown to increase the risk of infection if not treated before surgery[7], as well as complications after surgery[8]. Thus, there is a theoretical benefit to biliary drainage for relief of jaundice prior to surgical resection of these tumors.

It is thought the benefit of preoperative biliary drainage (PBD) may vary depending on the level of obstruction and planned surgery: Distal biliary obstruction from pancreatic cancer or distal cholangiocarcinoma may be treated surgically without the need for preoperative decompression, while hilar obstruction may require decompression to improve surgical outcomes. The differences in strategy likely stem from the need for partial hepatic resection in the treatment of hilar tumors, which may benefit from preoperative decompression.

MALIGNANT DISTAL BILIARY OBSTRUCTION

As many as 70% of patients with newly diagnosed pancreatic cancer have some degree of biliary tract obstruction at the time of diagnosis. Decompression *via* endoscopic stent placement can palliate jaundice and pruritus for symptomatic relief[9]. Stent placement may also speed allow the patient to begin chemotherapy regimens by reducing the risk of chemotoxicity in a cholestatic liver[10]. Endoscopic stent placement into the common bile duct is a fairly routine procedure (technically successful in over 90% of cases) and

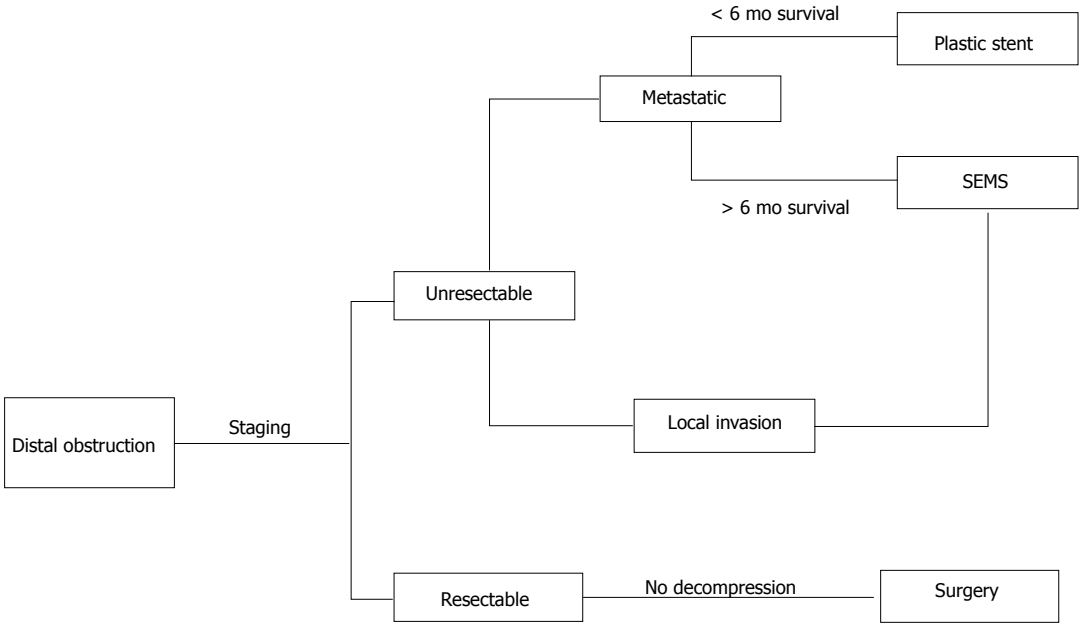

Figure 1 Algorithm for treatment of distal malignant biliary obstruction based on disease stage. Patients who are not candidates for ERCP with stent placement may undergo EUS-BD or percutaneous drainage. Adapted from Boulay BR, Parepally M. *World J Gastroenterol* 2014; **20**: 9345-9353. SEMS: Self-expanding metallic stent; ERCP: Endoscopic retrograde cholangiography; EUS-BD: Endoscopic ultrasound-guided biliary drainage.

has thus become the most common method of achieving biliary decompression[11]. The choice of plastic or metallic stents depends on factors such as cost-effectiveness, expected length of survival, and diagnostic certainty. Over the past decade the use of self-expanding metal stents (SEMS) has become more common for treatment of both benign and malignant biliary strictures. While surgeons initially discouraged use of SEMS for pancreatic cancer due to concerns of increasing the difficulty of resection, SEMS do not interfere with planned pancreaticoduodenectomy as long as the stent does not involve the hilum[12]. Thus the role for SEMS in treatment of obstruction from pancreatic cancer has grown in recent years.

Distal cholangiocarcinoma has a similar presentation, pattern of spread, and poor prognosis when compared to pancreatic ductal adenocarcinoma. At times the two diseases may be indistinguishable from each other upon initial presentation. Distal cholangiocarcinoma tends to infiltrate the adjacent pancreas, duodenum, and vasculature as well as nearby lymphatics, leading to locally advanced disease and eventually metastases. Thus, the same staging evaluation can be performed to assess for local invasion, with the goal of curative surgical resection when possible[13]. Once the stage is known, similar principles of biliary decompression are applied as in pancreatic ductal adenocarcinoma. The management is based on disease stage as depicted in Figure 1.

RESECTABLE DISEASE

Surgical resection is the definitive treatment for patients who present with early-stage pancreatic cancer[14]. PBD in resectable pancreatic cancer is not automatically recommended. Despite the beneficial effects of relieving jaundice, PBD has been associated with increased complications including various types of infections as well as pancreatic fistulas[15-17]. In 2010, van der Gaag *et al*[18] reported a randomized trial of 202 patients demonstrating that preoperative biliary drainage with stents was linked to increased complications compared to surgery alone in resectable pancreatic cancer. In this seminal study, rates of serious complications were 39% in the early-surgery group and 74% in the preoperative drainage group. Of note, the preoperative biliary drainage group waited 4 to 6 wk for surgery and were treated with plastic stents, both of which may have contributed to the poor performance of the biliary decompression group. However, based largely upon the experience noted by van der Gaag *et al*[18], preoperative biliary decompression for distal biliary obstruction is not recommended except to treat cholangitis or intractable pruritus.

LOCALLY ADVANCED DISEASE AND NEOADJUVANT THERAPY

Even those patients who undergo surgical resection of pancreatic cancer have poor long-term survival rates, so there is growing interest in the use of neoadjuvant therapy to boost outcomes, even for resectable pancreatic tumors[19]. Neoadjuvant chemoradiotherapy has also been used for locally advanced tumors to downstage them and permit eventual surgical resection. Biliary stents are placed prior to neoadjuvant chemoradiotherapy, with the expectation of remaining patent until the time of surgery. Unfortunately, this expectation has not always been met when plastic stents are used during the preoperative period.

Nuerous studies have now shown that SEMSs are preferable to plastic stents in patients undergoing neoadjuvant therapy[20-24]. Retrospective reviews of plastic stent performance during neoadjuvant therapy have demonstrated poor performance with the frequent need for unplanned stent exchange due to stent occlusion or cholangitis[20,25]. Adams *et al*[22] described a complication rate nearly 7 times higher with plastic stents, with a 3 times higher rate of hospitalization among a 52 patient cohort. SEMSs are clearly more expensive than plastic stents, but their lower occlusion rates (and thus fewer unplanned stent exchanges) make them a more cost effective choice for patients undergoing neoadjuvant chemoradiotherapy with planned surgical resection[26].

There are several choices of SEMS type for use in patients with malignant distal biliary obstruction undergoing neoadjuvant therapy or in palliative cases. When considering uncovered (USEMSs) or covered stents (CSEMSs), the difference in design leads to a trade-off in adverse events between tissue ingrowth in USEMS and stent migration in CSEMS. One recent meta-analysis concluded that CSEMSs afforded an average of 61 d longer patency than USEMSs in palliative cases, with the cost of an increased incidence in migration (RR 8.11)[27]. In contrast, a retrospective cohort study showed no difference in overall obstruction (CSEMSs 35% *vs* USEMSs 38%) among 749 patients, merely that the mechanisms of obstruction varied by stent design (tumor ingrowth *vs* debris)[28]. Partially covered SEMSs appear to have similar performance characteristics to fully covered SEMSs and USEMS[27,29].

Novel stent designs may further improve the performance of SEMS. A modified CSEMS with low axial force and uncovered flare ends has been developed with the goal of reducing stent migration, and when compared to USEMS had significantly longer patency (mean 219.3 d *vs* 166.9 d) and fewer unplanned procedures (23% *vs* 37%) compared to USEMSs[30]. Drug eluting stents have been designed in an attempt to improve SEMS prevent tumor ingrowth and stent occlusion[31]. An early multicenter prospective study using a paclitaxel-eluting stent did not show improved performance compared to conventional USEMS, though other stents are currently in development[32]. Anti-reflux stents have been developed to limit duodenal contents into the bile ducts and limit stent occlusion[23]. Initial experience with anti-reflux SEMS has yielded conflicting results, with one study showing long-term patency possibly exceeding conventional SEMS[33] while a smaller study showed a disappointing rate of early occlusion[34]. Further studies are needed to demonstrate the efficacy of anti-reflux and drug-eluting SEMS to determine their role in maintaining long-term stent patency.

Despite the wealth of data demonstrating the superiority of SEMS over plastic stents for malignant biliary obstruction, endoscopists may prefer to place a removable plastic stent if the diagnosis of malignancy is uncertain at that time of ERCP (such as in facilities where endoscopic ultrasound with FNA and rapid on-site evaluation by cytopathologists is not available). In this situation, benign conditions such as chronic pancreatitis or autoimmune cholangiopathy may be suspected as the etiology of the biliary stricture, and a removable stent is preferable. The use of a fully covered SEMS is ideal when suspicion of malignancy is high and life expectancy exceeds 4 mo.

PALLIATIVE DECOMPRESSION IN DISTAL OBSTRUCTION WITH METASTATIC DISEASE

In the setting of incurable pancreatic cancer, patients may often present with advanced disease and limited life expectancy. SEMS may not be cost-effective for these patients, since their main advantage of durable patency is not applicable. SEMSs cost 15-40 times more than most plastic stents, and are only cost effective if the patient survives > 4 mo[26]. Some authors have suggested that SEMSs should be used only in patients without distant metastases[35]. The presence of liver metastases has been shown to predict mortality in pancreatic cancer, though the data is sparse and generally used only to determine operative risk[36-38]. Although determination of life expectancy can be difficult and more research is needed to identify prognostic factors, endoscopists should be aware of the cost savings with plastic stents in patients with poor functional status and limited expected survival.

NON-ENDOSCOPIC BILIARY DRAINAGE IN MALIGNANT DISTAL OBSTRUCTION

In cases where ERCP fails or cannot be performed, percutaneous transhepatic biliary drainage (PTBD) has traditionally been used to create a tract for internal and external drainage. External drains have the potential downside of requiring emptying and flushing of the drain as well as routine drain exchange[39]. Recent trials of percutaneous SEMS placement have also demonstrated good safety and effectiveness[40-42]. An alternate second-line approach is endoscopic ultrasound-guided biliary drainage (EUS-BD), which has been increasingly shown to be both safe and effective when standard ERCP approaches fail. This technique can be used to achieve decompression *via* EUS-guided rendezvous procedure, EUS-guided choledochoduodenostomy, and EUS- guided hepatic gastrostomy[43]. Complications can include bile leak, bleeding, or pneumoperitoneum. EUS-BD remains technically complex and limited to high-volume expert centers[44].

Surgical biliary bypass remains an option, particularly when life expectancy exceeds 6 mo. Trials comparing surgical bypass to endoscopic therapy have shown similar mortality and fewer incidents of recurrent biliary obstruction when surgery was performed, though these

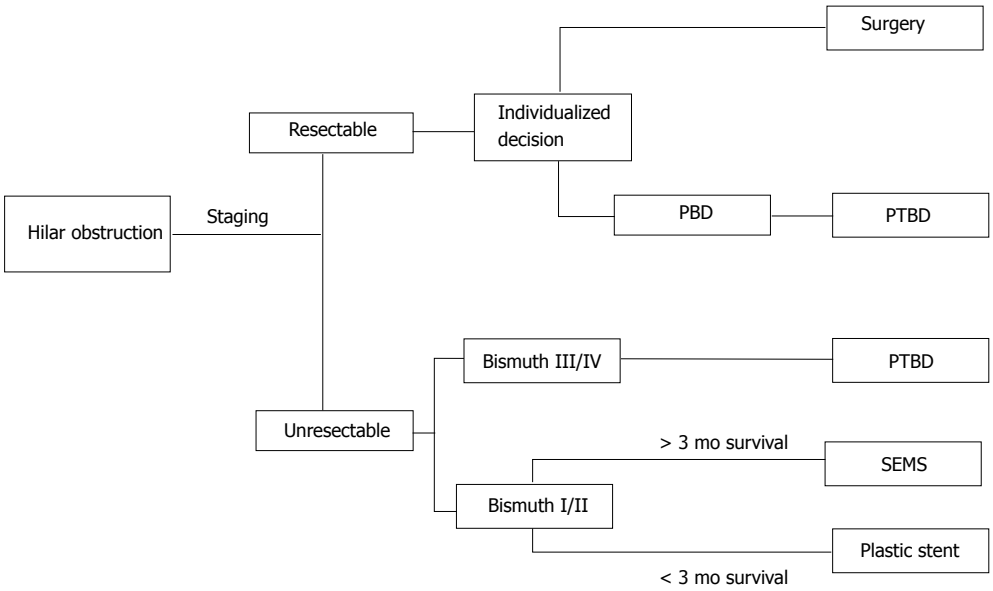

Figure 2 Algorithm for treatment of hilar malignant biliary obstruction. PBD is an individualized decision based on local expertise. PTBD: Percutaneous transhepatic biliary drainage; SEMS: Self-expanding metallic stent; PBD: Preoperative biliary drainage.

trials preceded the widespread use of SEMS and do not reflect current endoscopic practice[45,46]. Surgical bypass can also relieve biliary and gastric outlet obstruction at the same time *via* creation of a surgical gastrojejunostomy and biliary bypass. However, biliary and gastroduodenal obstruction can also be relieved endoscopically during the same procedure, with placement of a duodenal SEMS followed by an endoscopic approach to the papilla for ERCP guided biliary stent placement or even EUS-BD[47-50]. Thus, while ERCP is the preferred method for management of malignant distal biliary obstruction, a multidisciplinary team including interventional radiologists and surgeons is ideal for management of unusually difficult strictures when ERCP fails.

HILAR CHOLANGIOCARCINOMA: SURGICALLY TREATABLE DISEASE

Malignant obstruction of the extrahepatic bile ducts at the liver hilum can be much more difficult to treat, given the possible involvement of the left and right hepatic ducts and the need to decompress the left and right lobes individually. Complete resection is the only curative treatment for cholangiocarcinoma. Unfortunately, most patients will present with obstructive symptoms or frank jaundice later in the disease course[51]. Resection modalities vary depending on the location of the malignancy: Intrahepatic tumors are treated with hepatic resection while extrahepatic tumors can be classified as hilar cholangiocarcinoma (Klatskin tumor) or distal cholangiocarcinoma. The level of the cystic duct demarcates hilar *vs* distal tumors. Hilar cholangiocarcinoma may still require partial hepatic resection as part of definitive management[52]. In patients with Bismuth class 4 strictures involving the left and right hepatic duct, or vascular involvement of the hepatic artery or portal vein, surgical resection is contraindicated and neoadjuvant or palliative techniques are indicated[53]. Figure 2 depicts the overall management of hilar obstruction based on disease stage.

Several different imaging modalities can be used to determine the degree of proximal tumor extension. This is of critical importance in deciding on resectability and the optimal means of achieving biliary decompression. Computed tomography (CT) is the most commonly used type of imaging at the time of diagnosis, and multidetector CT (MDCT) has an accuracy of 86% in evaluating the ductal extent of hilar cholangiocarcinoma[54]. Magnetic resonance imaging (MRI) with magnetic resonance cholangiopancreatography (MRCP) provides detailed images of the biliary tree, though the presence of biliary stents can reduce the accuracy of staging[55]. The accuracy of MRI in determining hilar involvement is estimated at 89%, similar to that of MDCT. Both types of imaging also provide information regarding lymph nodes and direct invasion of nearby structures. In contrast, direct cholangiography by either ERCP or percutaneous means may not allow imaging of the entire biliary tree due to complete obstruction of some segments by tumor. Direct cholangiography is also invasive and presents risks of bleeding or infection which are avoidable with cross-sectional imaging. Thus, MDCT or MRCP are both acceptable initial imaging strategies for treatment planning, though the presence of stents may favor the use of MDCT.

When a patient with hilar cholangiocarcinoma presents with resectable disease, the treatment team must consider whether to pursue PBD. As with pancreatic cancer, preoperative biliary drainage has long been a debated topic, though most authorities advocate against it in the absence of cholangitis[56,57]. Drainage is recommended in

special situations such as cholangitis as well as patients with symptomatic jaundice (*e.g.*, pruritus) or renal failure[10]. In the absence of these factors, preoperative drainage has been previously discouraged on the basis that it confers no mortality benefit but may cause adverse events. Meta-analyses and systematic reviews of randomized controlled trials of preoperative biliary drainage have found increased complications and morbidity when PBD was performed, though overall mortality does not differ between the two groups[58,59].

Nonetheless, it is standard practice to perform preoperative biliary drainage in countries such as Japan and South Korea, and there is reason to believe it can be helpful in optimizing surgical outcomes with proper patient selection. One reason for this enthusiasm has been increased recognition of the flaws in earlier studies which argued against preoperative drainage. A large number of studies from 1980s to early 2000s did not have standardized timing to clear obstruction and jaundice, with some prolonged periods that contributed to stent occlusions and associated complications[60]. Even now, the optimal bilirubin level for surgery and optimal duration of PBD have not been established. The risk of drain occlusion or inflammatory change within the bile duct with prolonged drainage must be weighed against the risk of progressive obstruction on the outcome of liver resection during curative surgery. Thus preoperative biliary drainage is indicated when there will be a delay for surgery, such as in patients who undergo selective portal vein embolization in the setting of inadequate future liver remnant. In addition, the high-quality cholangiograms obtained in PBD can assist with treatment planning by delineating the extent of tumor involvement within the segmental bile ducts (though staging with MDCT or MRCP can provide adequate staging information without the risks of decompression).

It is possible that the benefit of PBD depends on the location of biliary obstruction within the hilum. Farges *et al*[61] performed a multicenter retrospective analysis of 366 patients undergoing extended right or left hepatectomy for hilar cholangiocarcinoma. PBD was not found to improve mortality overall, though a subgroup analysis revealed improved mortality in patients undergoing right hepatectomy (through reduced incidence of post-operative liver failure) while mortality was worse with left hepatectomy (through increased risk of sepsis)[61]. This intriguing data will require confirmation with additional studies but may help guide the controversial decision whether to perform PBD in the future.

HILAR CHOLANGIOCARCINOMA: TECHNIQUES FOR PREOPERATIVE BILIARY DRAINAGE

Currently three techniques are available for preoperative biliary drainage of hilar malignancy: PTBD, endoscopic nasobiliary drainage (ENBD), and endoscopic retrograde biliary drainage (ERBD) with stent placement. No randomized controlled trial has been performed to compare these techniques. For hilar tumors, ERBD can be technically challenging with the need to place multiple stents to drain obstructed biliary segments. The rate of complications is high, with reports showing morbidity of 25%-50% and mortality rates of 3%-5% with hilar tumors[62]. The complications are mainly due to cholangitis with stent failure or inadequate drainage[3,8].

ENBD has the advantage of providing drainage while allowing repeat cholangiography as needed prior to surgery with easy access to the drained intrahepatic segments[7]. Nasobiliary drainage is also a safe technique with fewer complications than PTBD[8]. However, nasobiliary tubes can be easily dislodged and may be poorly tolerated due to patient discomfort[63]. The use of multiple nasobiliary tubes to provide bilateral drainage for Bismuth Ⅳ hilar tumors has been performed but is technically demanding.

Percutaneous transhepatic biliary drainage has become the preferred method for preoperative biliary decompression in some centers. This technique is attractive due to its low complication rate when compared to endoscopic stent placement. Kloek *et al*[3] reviewed 101 patients who had undergone PBD and found 48% infection rate with ERBD, while PTBD was associated with only 9% infection rate. The chief problem with PTBD is the risk of tumor seeding along the drain tract, which is estimated at 5%-20%[7,8]. External transhepatic drains may cause patient discomfort and sometimes require additional oral intake of bile acid supplements, representing an uncomfortable nuisance to the patient[8]. Despite these drawbacks, the safety and comparative ease of drain placement (compared to ENBD) make PTBD a reasonable option depending on the local expertise of the treatment team.

HILAR CHOLANGIOCARCINOMA: PALLIATIVE THERAPY FOR MALIGNANT BILIARY OBSTRUCTION

While preoperative treatment tends to employ ENBD or PTBD, these therapies are less practical in the setting of unresectable disease. For patients with Bismuth Ⅰ or Ⅱ tumors, ERBD has similar performance to PTBD while being less invasive. However, patients with more advanced hilar obstruction (Bismuth Ⅲ or Ⅳ) are more difficult to palliate with biliary stents. A retrospective review of 126 patients with Bismuth Ⅲ-Ⅳ obstruction demonstrated higher success rates with PTBD over SEMS (93% *vs* 77%), though median survival was similar between the two groups[64]. Thus, PTBD is generally favored over endoscopic therapy even for palliation in advanced hilar obstruction.

The goal for palliative drainage is to relieve jaundice by draining an adequate liver volume (50% or more). This can be achieved with a single stent in Bismuth Ⅰ tumors, though the strategy with more advanced hilar disease is more complex. Drainage of > 50% of the liver volume may require more than one catheter or stent, though the right lobe of the liver takes up 50%-60% of the liver volume and successful drainage of the right lobe with a single stent may be sufficient to relieve

jaundice. The question of whether to place a single or multiple drains can be answered by volume assessment of the liver (volumetry) using cross-sectional imaging by CT or MRI. MRI imaging can be used to guide the placement of a single stent to access the dominant lobe and provide adequate drainage with a single stent[65].

Much like the treatment of distal obstruction with pancreatic malignancies, the past decade has seen increasing use of SEMS over plastic stents. There have been numerous studies that show significant advantages to metal stents *vs* plastic polyethylene stents[56]. The length of patency for SEMS compared to plastic is much higher (as much as 12 mo *vs* 3 mo), owing to the 8-10 mm diameter of SEMS compared to the narrower 7, 8.5 or 10 French plastic stents. Complications from occlusion or migration are less common, though SEMS are notably much more expensive than plastic stents[66]. Of course, patients with low performance status or advanced illness may not be expected to benefit from the prolonged duration of patency in SEMS; patients with life expectancy of < 3 mo would achieve the same benefit of palliation with lower costs using plastic stents. Cost analysis has demonstrated superiority of SEMS when patient survival is expected to exceed 3 mo[56,66-68]. As with pancreatic cancer patients, the treating endoscopist must consider expected length of survival when choosing an appropriate and cost-effective stent for palliation.

When bilateral or multisegmental stents are required for adequate biliary drainage, various techniques are available to place SEMS and take advantage of the prolonged duration of patency relative to plastic stents. Not all patients will require multiple SEMS; a meta-analysis by Sawas *et al*[69] shows no statistical difference in rate of failures or cholangitis between unilateral and bilateral SEMS placement for hilar tumors. Other retrospective data has favored the use of bilateral stents over unilateral stenting, with superior length of patency using multiple stents[70]. The two main techniques for use of multiple stents for hilar tumors are the "side by side" method, in which both stents are placed in parallel, or the "stent-in-stent" or "Y" method, in which the second stent is placed through the mesh interstices of the first stent with their distal ends overlapping[71]. There are no large studies indicating which method is superior in terms of technical or clinical success. Additional studies have looked at a 3-branch stent-in-stent to allow for better patency of stenting, with promising results but high degree of challenge for the endoscopist[72,73]. It is generally recommended that endoscopic therapy of advanced hilar strictures be performed by experienced endoscopists in a tertiary center with available backup by interventional radiologists and surgeons.

ENDOSCOPIC ADJUVANT TREATMENT OF BILIARY OBSTRUCTION: MOVING BEYOND STENTING

Patients with cholangiocarcinoma and pancreatic cancer continue to have notoriously poor prognoses when surgical cure is not an option. Meta-analyses have not shown significant improvement with standard chemoradiation regimens for biliary malignancies[74], likely due to late presentation and aggressive nature on presentation. However, two endoscopic therapies aimed at providing local control of malignant biliary obstruction have shown some promise in early studies.

Over the past decade the use of photodynamic therapy (PDT) has been studied for palliation of unresectable cholangiocarcinoma. This technique employs a photosensitizing molecule such as porfimer sodium which accumulates in tissue with rapid turnover such as malignant cells. After 48 h laser irradiation is then used to treat the tumor, leading to selective apoptosis within the tumor mass *via* generation of oxidative radicals. The application of oxygen and light can be performed through a cholangioscope for precise phototherapy administration to limit damage on normal tissue[75]. The first randomized controlled trial of PDT when compared to biliary stenting alone showed a dramatic increase in survival time from 98 d to 493 d[76]. Another RCT also showed median survival increased from 210 to 630 d[77]. Retrospective data also contributes to the body of information supporting increased survival and quality of life when PDT is used in additional to biliary stents as well as chemotherapy[78,79]. Side effects from phototherapy are mainly related to photosensitivity, requiring patients to avoid direct sunlight for 4-6 wk. In addition, the high cost of PDT may be a factor preventing its widespread use for local control of unresectable cholangiocarcinoma.

Radiofrequency ablation, previously used for colonic or esophageal malignancies as well as hepatocellular carcinoma, has also been increasingly studied for local treatment of biliary obstructive malignancies. Compared to PDT, it offers low cost and is technically simple to perform. RFA induces ablative necrosis and can be used to palliate known biliary malignancies by using a bipolar probe placed at the site of obstruction[75]. RFA can be performed percutaneously or *via* a catheter inserted *via* ERCP. Ablation uses 7-10 W bursts to create coagulative necrosis of the intraductal tumor mass and when performed *via* ERCP is followed by biliary stent placement[51,75]. Plastic stents are applied when future ablations are planned, while SEMS may be used when a single session is planned. The risk of adverse events is low but includes hemobilia and biliary fistulas. The body of literature supporting RFA for biliary malignancies is not as robust as that for PDT, consisting mostly of retrospective series[80]. A retrospective comparison by Strand *et al*[81] compared results in 48 patients (16 RFA, 32 PDT) which demonstrated similar median survival (9.6 mo in RFA, 7.5 mo in PDT). Future studies will be required to determine the optimal techniques for RFA, as well as the patient populations who are most likely to benefit. European studies have also investigated the use of RFA therapy to treat occlusion of SEMS without the need for additional stent placement[82].

The role of RFA in distal malignant biliary obstruction has

not been defined, though early experience is encouraging. In a retrospective study of 20 patients undergoing RFA of biliary strictures, 8 patients had distal obstruction due to pancreatic adenocarcinoma or intraductal papillary mucinous neoplasm. The study showed a median increase of 3.5 mm in bile duct diameter following RFA treatment, with maintenance of stent patency at 30 d[83]. Similarly, a registry of 69 patients who underwent RFA for malignant biliary obstruction included 19 patients with pancreatic cancer. Again, the median diameter of the bile duct improved following RFA treatment. Interestingly, the pancreatic cancer patients responded better to RFA than cholangiocarcinoma patients, with RR 1.8 for stricture improvement[84]. Other outcomes such as length of survival have not yet been studied in these patients. While further data is needed, including high-quality prospective data, the ability to achieve local control and prolong survival in these diseases using endoscopic therapies is an exciting prospect.

CONCLUSION

The management of malignant biliary obstruction requires consideration of several factors prior to the act of decompression *via* endoscopic or percutaneous means. The location of the stricture and underlying malignancy will affect the approach, as hilar stricture may be much more difficult to treat compared to simple strictures of the distal common bile duct. The stage of underlying malignancy also plays a role, as resectable disease may not typically require preoperative biliary drainage. For most cases, preoperative biliary drainage is discouraged due to the high incidence of infectious complications. In patients undergoing neoadjuvant therapy for locally advanced disease, SEMS appear to be the optimal approach for distal strictures while most data for hilar strictures appears to favor PTBD or ENBD. For palliation in advanced malignancy, SEMS are generally favored if life expectancy exceeds 3-4 mo, while plastic stents are favored in patients with particularly poor prognosis. The use of photodynamic therapy and radiofrequency ablation to achieve local control of these malignancies is an important step in prolonging survival, and presents an opportunity for endoscopists to have an increased role beyond stent placement in improving the quality and quantity of life for patients with cancer and biliary obstruction.

REFERENCES

1 **Kruse EJ**. Palliation in pancreatic cancer. *Surg Clin North Am* 2010; **90**: 355-364 [PMID: 20362791]

2 **Kassir R**, Barabino G, Bageacu S, Ferrari G, Abboud K, Dumas O, Peoc'h M, Porcheron J. Biliary papillomatosis in the common bile duct. *Endoscopy* 2013; **45** Suppl 2 UCTN: E197-E198 [PMID: 23832505 DOI: 10.1055/s-0033-1344163]

3 **Kloek JJ**, van der Gaag NA, Aziz Y, Rauws EA, van Delden OM, Lameris JS, Busch OR, Gouma DJ, van Gulik TM. Endoscopic and percutaneous preoperative biliary drainage in patients with suspected hilar cholangiocarcinoma. *J Gastrointest Surg* 2010; **14**: 119-125 [PMID: 19756881 DOI: 10.1007/s11605-009-1009-1]

4 **Sauvanet A**, Boher JM, Paye F, Bachellier P, Sa Cuhna A, Le Treut YP, Adham M, Mabrut JY, Chiche L, Delpero JR. Severe Jaundice Increases Early Severe Morbidity and Decreases Long-Term Survival after Pancreaticoduodenectomy for Pancreatic Adenocarcinoma. *J Am Coll Surg* 2015; **221**: 380-389 [PMID: 26206638 DOI: 10.1016/j.jamcollsurg.2015.03.058]

5 **Strasberg SM**, Gao F, Sanford D, Linehan DC, Hawkins WG, Fields R, Carpenter DH, Brunt EM, Phillips C. Jaundice: an important, poorly recognized risk factor for diminished survival in patients with adenocarcinoma of the head of the pancreas. *HPB* (Oxford) 2014; **16**: 150-156 [PMID: 23600768 DOI: 10.1111/hpb.12094]

6 **Papadopoulos V**, Filippou D, Manolis E, Mimidis K. Haemostasis impairment in patients with obstructive jaundice. *J Gastrointestin Liver Dis* 2007; **16**: 177-186 [PMID: 17592568]

7 **Nimura Y**. Preoperative biliary drainage before resection for cholangiocarcinoma (Pro). *HPB* (Oxford) 2008; **10**: 130-133 [PMID: 18773090 DOI: 10.1080/13651820801992666]

8 **Iacono C**, Ruzzenente A, Campagnaro T, Bortolasi L, Valdegamberi A, Guglielmi A. Role of preoperative biliary drainage in jaundiced patients who are candidates for pancreatoduodenectomy or hepatic resection: highlights and drawbacks. *Ann Surg* 2013; **257**: 191-204 [PMID: 23013805]

9 **Ballinger AB**, McHugh M, Catnach SM, Alstead EM, Clark ML. Symptom relief and quality of life after stenting for malignant bile duct obstruction. *Gut* 1994; **35**: 467-470 [PMID: 7513672]

10 **Conroy T**, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. *N Engl J Med* 2011; **364**: 1817-1825 [PMID: 21561347 DOI: 10.1056/NEJMoa1011923]

11 **Dumonceau JM**, Delhaye M, Tringali A, Dominguez-Munoz JE, Poley JW, Arvanitaki M, Costamagna G, Costea F, Devière J, Eisendrath P, Lakhtakia S, Reddy N, Fockens P, Ponchon T, Bruno M. Endoscopic treatment of chronic pancreatitis: European Society of Gastrointestinal Endoscopy (ESGE) Clinical Guideline. *Endoscopy* 2012; **44**: 784-800 [PMID: 22752888 DOI: 10.1055/s-0032-1309840]

12 **Lawrence C**, Howell DA, Conklin DE, Stefan AM, Martin RF. Delayed pancreaticoduodenectomy for cancer patients with prior ERCP-placed, nonforeshortening, self-expanding metal stents: a positive outcome. *Gastrointest Endosc* 2006; **63**: 804-807 [PMID: 16650542]

13 **Ercolani G**, Dazzi A, Giovinazzo F, Ruzzenente A, Bassi C, Guglielmi A, Scarpa A, D'Errico A, Pinna AD. Intrahepatic, perihilar and distal cholangiocarcinoma: Three different locations of the same tumor or three different tumors? *Eur J Surg Oncol* 2015; **41**: 1162-1169 [PMID: 26095704 DOI: 10.1016/j.ejso.2015.05.013]

14 **Wray CJ**, Ahmad SA, Matthews JB, Lowy AM. Surgery for pancreatic cancer: recent controversies and current practice. *Gastroenterology* 2005; **128**: 1626-1641 [PMID: 15887155]

15 **Sohn TA**, Yeo CJ, Cameron JL, Pitt HA, Lillemoe KD. Do preoperative biliary stents increase postpancreaticoduodenectomy complications? *J Gastrointest Surg* 2000; **4**: 258-267; discussion 267-268 [PMID: 10769088]

16 **Pisters PW**, Hudec WA, Hess KR, Lee JE, Vauthey JN, Lahoti S, Raijman I, Evans DB. Effect of preoperative biliary decompression on pancreaticoduodenectomy-associated morbidity in 300 consecutive patients. *Ann Surg* 2001; **234**: 47-55 [PMID: 11420482]

17 **Namias N**, Demoya M, Sleeman D, Reever CM, Raskin JB, Ginzburg E, Minhaj M, Pappas PA, Padron I, Levi JU. Risk of postoperative infection in patients with bactibilia undergoing surgery for obstructive jaundice. *Surg Infect* (Larchmt) 2005; **6**: 323-328 [PMID: 16201942]

18 **van der Gaag NA**, Rauws EA, van Eijck CH, Bruno MJ, van der Harst E, Kubben FJ, Gerritsen JJ, Greve JW, Gerhards MF, de Hingh IH, Klinkenbijl JH, Nio CY, de Castro SM, Busch OR, van

Gulik TM, Bossuyt PM, Gouma DJ. Preoperative biliary drainage for cancer of the head of the pancreas. *N Engl J Med* 2010; **362**: 129-137 [PMID: 20071702 DOI: 10.1056/NEJMoa0903230]

19 **Artinyan A**, Anaya DA, McKenzie S, Ellenhorn JD, Kim J. Neoadjuvant therapy is associated with improved survival in resectable pancreatic adenocarcinoma. *Cancer* 2011; **117**: 2044-2049 [PMID: 21523715 DOI: 10.1002/cncr.25763]

20 **Boulay BR**, Gardner TB, Gordon SR. Occlusion rate and complications of plastic biliary stent placement in patients undergoing neoadjuvant chemoradiotherapy for pancreatic cancer with malignant biliary obstruction. *J Clin Gastroenterol* 2010; **44**: 452-455 [PMID: 20179612 DOI: 10.1097/MCG.0b013e3181d2ef06]

21 **Wasan SM**, Ross WA, Staerkel GA, Lee JH. Use of expandable metallic biliary stents in resectable pancreatic cancer. *Am J Gastroenterol* 2005; **100**: 2056-2061 [PMID: 16128952]

22 **Adams MA**, Anderson MA, Myles JD, Khalatbari S, Scheiman JM. Self-expanding metal stents (SEMS) provide superior outcomes compared to plastic stents for pancreatic cancer patients undergoing neoadjuvant therapy. *J Gastrointest Oncol* 2012; **3**: 309-313 [PMID: 23205306 DOI: 10.3978/j.issn.2078-6891.2011.050]

23 **Misra SP**, Dwivedi M. Reflux of duodenal contents and cholangitis in patients undergoing self-expanding metal stent placement. *Gastrointest Endosc* 2009; **70**: 317-321 [PMID: 19539920 DOI: 10.1016/j.gie.2008.12.054]

24 **Boulay BR**. Biliary stents for pancreas cancer with obstruction: the problem with plastic. *J Gastrointest Oncol* 2012; **3**: 306-308 [PMID: 23205305 DOI: 10.3978/j.issn.2078-6891.2012.047]

25 **Ge PS**, Hamerski CM, Watson RR, Komanduri S, Cinnor BB, Bidari K, Klapman JB, Lin CL, Shah JN, Wani S, Donahue TR, Muthusamy VR. Plastic biliary stent patency in patients with locally advanced pancreatic adenocarcinoma receiving downstaging chemotherapy. *Gastrointest Endosc* 2015; **81**: 360-366 [PMID: 25442083 DOI: 10.1016/j.gie.2014.08.020]

26 **Moss AC**, Morris E, Leyden J, MacMathuna P. Do the benefits of metal stents justify the costs? A systematic review and meta-analysis of trials comparing endoscopic stents for malignant biliary obstruction. *Eur J Gastroenterol Hepatol* 2007; **19**: 1119-1124 [PMID: 17998839]

27 **Saleem A**, Leggett CL, Murad MH, Baron TH. Meta-analysis of randomized trials comparing the patency of covered and uncovered self-expandable metal stents for palliation of distal malignant bile duct obstruction. *Gastrointest Endosc* 2011; **74**: 321-327.e1-3 [PMID: 21683354 DOI: 10.1016/j.gie.2011.03.1249]

28 **Lee JH**, Krishna SG, Singh A, Ladha HS, Slack RS, Ramireddy S, Raju GS, Davila M, Ross WA. Comparison of the utility of covered metal stents versus uncovered metal stents in the management of malignant biliary strictures in 749 patients. *Gastrointest Endosc* 2013; **78**: 312-324 [PMID: 23591331 DOI: 10.1016/j.gie.2013.02.032]

29 **Telford JJ**, Carr-Locke DL, Baron TH, Poneros JM, Bounds BC, Kelsey PB, Schapiro RH, Huang CS, Lichtenstein DR, Jacobson BC, Saltzman JR, Thompson CC, Forcione DG, Gostout CJ, Brugge WR. A randomized trial comparing uncovered and partially covered self-expandable metal stents in the palliation of distal malignant biliary obstruction. *Gastrointest Endosc* 2010; **72**: 907-914 [PMID: 21034891 DOI: 10.1016/j.gie.2010.08.021]

30 **Kitano M**, Yamashita Y, Tanaka K, Konishi H, Yazumi S, Nakai Y, Nishiyama O, Uehara H, Mitoro A, Sanuki T, Takaoka M, Koshitani T, Arisaka Y, Shiba M, Hoki N, Sato H, Sasaki Y, Sato M, Hasegawa K, Kawabata H, Okabe Y, Mukai H. Covered self-expandable metal stents with an anti-migration system improve patency duration without increased complications compared with uncovered stents for distal biliary obstruction caused by pancreatic carcinoma: a randomized multicenter trial. *Am J Gastroenterol* 2013; **108**: 1713-1722 [PMID: 24042190 DOI: 10.1038/ajg.2013.305]

31 **Shah T**. Drug-eluting stents in malignant biliary obstruction: where do we stand? *Dig Dis Sci* 2013; **58**: 610-612 [PMID: 23250674 DOI: 10.1007/s10620-012-2507-7]

32 **Jang SI**, Kim JH, You JW, Rhee K, Lee SJ, Kim HG, Han J, Shin IH, Park SH, Lee DK. Efficacy of a metallic stent covered with a paclitaxel-incorporated membrane versus a covered metal stent for malignant biliary obstruction: a prospective comparative study. *Dig Dis Sci* 2013; **58**: 865-871 [PMID: 23179148 DOI: 10.1007/s10620-012-2472-1]

33 **Hu B**, Wang TT, Shi ZM, Wang SZ, Lu R, Pan YM, Huang H, Wang SP. A novel antireflux metal stent for the palliation of biliary malignancies: a pilot feasibility study (with video). *Gastrointest Endosc* 2011; **73**: 143-148 [PMID: 20970788 DOI: 10.1016/j.gie.2010.08.048]

34 **Kim DU**, Kwon CI, Kang DH, Ko KH, Hong SP. New antireflux self-expandable metal stent for malignant lower biliary obstruction: in vitro and in vivo preliminary study. *Dig Endosc* 2013; **25**: 60-66 [PMID: 23286258 DOI: 10.1111/j.1443-1661.2012.01324.x]

35 **Soderlund C**, Linder S. Covered metal versus plastic stents for malignant common bile duct stenosis: a prospective, randomized, controlled trial. *Gastrointest Endosc* 2006; **63**: 986-995 [PMID: 16733114]

36 **Inal A**, Kos FT, Algin E, Yildiz R, Berk V, Tugba Unek I, Colak D, Colak D, Kucukoner M, Tamer Elkiran E, Helvaci K, Geredeli C, Dane F, Balakan O, Ali Kaplan M, Gok Durnali A, Harputoglu H, Goksel G, Ozdemir N, Buyukberber S, Gumus M, Ozkan M, Benekli M, Isikdogan A. Prognostic factors in patients with advanced pancreatic cancer treated with gemcitabine alone or gemcitabine plus cisplatin: retrospective analysis of a multicenter study. *J BUON* 2012; **17**: 102-105 [PMID: 22517701]

37 **Zhang DX**, Dai YD, Yuan SX, Tao L. Prognostic factors in patients with pancreatic cancer. *Exp Ther Med* 2012; **3**: 423-432 [PMID: 22969906 DOI: 10.3892/etm.2011.412]

38 **Hartwig W**, Hackert T, Hinz U, Gluth A, Bergmann F, Strobel O, Büchler MW, Werner J. Pancreatic cancer surgery in the new millennium: better prediction of outcome. *Ann Surg* 2011; **254**: 311-319 [PMID: 21606835 DOI: 10.1097/SLA.0b013e31821fd334]

39 **Robson PC**, Heffernan N, Gonen M, Thornton R, Brody LA, Holmes R, Brown KT, Covey AM, Fleischer D, Getrajdman GI, Jarnagin W, Sofocleous C, Blumgart L, D'Angelica M. Prospective study of outcomes after percutaneous biliary drainage for malignant biliary obstruction. *Ann Surg Oncol* 2010; **17**: 2303-2311 [PMID: 20358300 DOI: 10.1245/s10434-010-1045-9]

40 **Piñol V**, Castells A, Bordas JM, Real MI, Llach J, Montañà X, Feu F, Navarro S. Percutaneous self-expanding metal stents versus endoscopic polyethylene endoprostheses for treating malignant biliary obstruction: randomized clinical trial. *Radiology* 2002; **225**: 27-34 [PMID: 12354980]

41 **Briggs CD**, Irving GR, Cresswell A, Peck R, Lee F, Peterson M, Cameron IC. Percutaneous transhepatic insertion of self-expanding short metal stents for biliary obstruction before resection of pancreatic or duodenal malignancy proves to be safe and effective. *Surg Endosc* 2010; **24**: 567-571 [PMID: 19609609 DOI: 10.1007/s00464-009-0598-9]

42 **Mahgerefteh S**, Hubert A, Klimov A, Bloom AI. Clinical Impact of Percutaneous Transhepatic Insertion of Metal Biliary Endoprostheses for Palliation of Jaundice and Facilitation of Chemotherapy. *Am J Clin Oncol* 2015; **38**: 489-494 [PMID: 24064748]

43 **Sarkaria S**, Lee HS, Gaidhane M, Kahaleh M. Advances in endoscopic ultrasound-guided biliary drainage: a comprehensive review. *Gut Liver* 2013; **7**: 129-136 [PMID: 23560147 DOI: 10.5009/gnl.2013.7.2.129]

44 **Shah JN**, Marson F, Weilert F, Bhat YM, Nguyen-Tang T, Shaw RE, Binmoeller KF. Single-operator, single-session EUS-guided anterograde cholangiopancreatography in failed ERCP or inaccessible papilla. *Gastrointest Endosc* 2012; **75**: 56-64 [PMID: 22018554 DOI: 10.1016/j.gie.2011.08.032]

45 **Glazer ES**, Hornbrook MC, Krouse RS. A meta-analysis of randomized trials: immediate stent placement vs. surgical bypass in the palliative management of malignant biliary obstruction. *J Pain Symptom Manage* 2014; **47**: 307-314 [PMID: 23830531 DOI: 10.1016/j.jpainsymman.2013.03.013]

46 **Moss AC**, Morris E, Leyden J, MacMathuna P. Malignant distal biliary obstruction: a systematic review and meta-analysis of endoscopic and surgical bypass results. *Cancer Treat Rev* 2007; **33**: 213-221 [PMID: 17157990]

47 **Kaw M**, Singh S, Gagneja H. Clinical outcome of simultaneous self-expandable metal stents for palliation of malignant biliary and duodenal obstruction. *Surg Endosc* 2003; **17**: 457-461 [PMID: 12404053]

48 **Mutignani M**, Tringali A, Shah SG, Perri V, Familiari P, Iacopini F, Spada C, Costamagna G. Combined endoscopic stent insertion in malignant biliary and duodenal obstruction. *Endoscopy* 2007; **39**: 440-447 [PMID: 17516351]

49 **Moon JH**, Choi HJ. Endoscopic double-metallic stenting for malignant biliary and duodenal obstructions. *J Hepatobiliary Pancreat Sci* 2011; **18**: 658-663 [PMID: 21655973 DOI: 10.1007/s00534-011-0409-2]

50 **Tonozuka R**, Itoi T, Sofuni A, Itokawa F, Moriyasu F. Endoscopic double stenting for the treatment of malignant biliary and duodenal obstruction due to pancreatic cancer. *Dig Endosc* 2013; **25** Suppl 2: 100-108 [PMID: 23617659 DOI: 10.1111/den.12063]

51 **Guler S**, Cimen S, Molinari M. Advances in loco-regional palliation of unresectable cholangiocarcinomas. *Ann Palliat Med* 2014; **3**: 65-74 [PMID: 25841505]

52 **Blechacz B**, Komuta M, Roskams T, Gores GJ. Clinical diagnosis and staging of cholangiocarcinoma. *Nat Rev Gastroenterol Hepatol* 2011; **8**: 512-522 [PMID: 21808282 DOI: 10.1038/nrgastro.2011.131]

53 **Khan SA**, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005; **366**: 1303-1314 [PMID: 16214602]

54 **Ruys AT**, van Beem BE, Engelbrecht MR, Bipat S, Stoker J, Van Gulik TM. Radiological staging in patients with hilar cholangiocarcinoma: a systematic review and meta-analysis. *Br J Radiol* 2012; **85**: 1255-1262 [PMID: 22919007 DOI: 10.1259/bjr/88405305]

55 **Choi JY**, Kim MJ, Lee JM, Kim KW, Lee JY, Han JK, Choi BI. Hilar cholangiocarcinoma: role of preoperative imaging with sonography, MDCT, MRI, and direct cholangiography. *AJR Am J Roentgenol* 2008; **191**: 1448-1457 [PMID: 18941084 DOI: 10.2214/AJR.07.3992]

56 **Khan SA**, Davidson BR, Goldin RD, Heaton N, Karani J, Pereira SP, Rosenberg WM, Tait P, Taylor-Robinson SD, Thillainayagam AV, Thomas HC, Wasan H. Guidelines for the diagnosis and treatment of cholangiocarcinoma: an update. *Gut* 2012; **61**: 1657-1669 [PMID: 22895392]

57 **Rerknimitr R**, Angsuwatcharakon P, Ratanachu-ek T, Khor CJ, Ponnudurai R, Moon JH, Seo DW, Pantongrag-Brown L, Sangchan A, Pisespongsa P, Akaraviputh T, Reddy ND, Maydeo A, Itoi T, Pausawasdi N, Punamiya S, Attasaranya S, Devereaux B, Ramchandani M, Goh KL. Asia-Pacific consensus recommendations for endoscopic and interventional management of hilar cholangiocarcinoma. *J Gastroenterol Hepatol* 2013; **28**: 593-607 [PMID: 23350673 DOI: 10.1111/jgh.12128]

58 **Liu F**, Li Y, Wei Y, Li B. Preoperative biliary drainage before resection for hilar cholangiocarcinoma: whether or not? A systematic review. *Dig Dis Sci* 2011; **56**: 663-672 [PMID: 20635143 DOI: 10.1007/s10620-010-1338-7]

59 **Fang Y**, Gurusamy KS, Wang Q, Davidson BR, Lin H, Xie X, Wang C. Meta-analysis of randomized clinical trials on safety and efficacy of biliary drainage before surgery for obstructive jaundice. *Br J Surg* 2013; **100**: 1589-1596 [PMID: 24264780 DOI: 10.1002/bjs.9260]

60 **Wiggers JK**, Coelen RJ, Rauws EA, van Delden OM, van Eijck CH, de Jonge J, Porte RJ, Buis CI, Dejong CH, Molenaar IQ, Besselink MG, Busch OR, Dijkgraaf MG, van Gulik TM. Preoperative endoscopic versus percutaneous transhepatic biliary drainage in potentially resectable perihilar cholangiocarcinoma (DRAINAGE trial): design and rationale of a randomized controlled trial. *BMC Gastroenterol* 2015; **15**: 20 [PMID: 25887103 DOI: 10.1186/s12876-015-0251-0]

61 **Farges O**, Regimbeau JM, Fuks D, Le Treut YP, Cherqui D, Bachellier P, Mabrut JY, Adham M, Pruvot FR, Gigot JF. Multicentre European study of preoperative biliary drainage for hilar cholangiocarcinoma. *Br J Surg* 2013; **100**: 274-283 [PMID: 23124720]

62 **Park do H**, Kim MH, Choi JS, Lee SS, Seo DW, Kim JH, Han J, Kim JC, Choi EK, Lee SK. Covered versus uncovered wallstent for malignant extrahepatic biliary obstruction: a cohort comparative analysis. *Clin Gastroenterol Hepatol* 2006; **4**: 790-796 [PMID: 16716757]

63 **Kawakami H**, Kuwatani M, Onodera M, Haba S, Eto K, Ehira N, Yamato H, Kudo T, Tanaka E, Hirano S, Kondo S, Asaka M. Endoscopic nasobiliary drainage is the most suitable preoperative biliary drainage method in the management of patients with hilar cholangiocarcinoma. *J Gastroenterol* 2011; **46**: 242-248 [PMID: 20700608 DOI: 10.1007/s00535-010-0298-1]

64 **Paik WH**, Park YS, Hwang JH, Lee SH, Yoon CJ, Kang SG, Lee JK, Ryu JK, Kim YT, Yoon YB. Palliative treatment with self-expandable metallic stents in patients with advanced type III or IV hilar cholangiocarcinoma: a percutaneous versus endoscopic approach. *Gastrointest Endosc* 2009; **69**: 55-62 [PMID: 18657806 DOI: 10.1016/j.gie.2008.04.005]

65 **Hintze RE**, Abou-Rebyeh H, Adler A, Veltzke-Schlieker W, Felix R, Wiedenmann B. Magnetic resonance cholangiopancreatography-guided unilateral endoscopic stent placement for Klatskin tumors. *Gastrointest Endosc* 2001; **53**: 40-46 [PMID: 11154487]

66 **Strom TJ**, Klapman JB, Springett GM, Meredith KL, Hoffe SE, Choi J, Hodul P, Malafa MP, Shridhar R. Comparative long-term outcomes of upfront resected pancreatic cancer after preoperative biliary drainage. *Surg Endosc* 2015; **29**: 3273-3281 [PMID: 25631110 DOI: 10.1007/s00464-015-4075-3]

67 **Sangchan A**, Kongkasame W, Pugkhem A, Jenwitheesuk K, Mairiang P. Efficacy of metal and plastic stents in unresectable complex hilar cholangiocarcinoma: a randomized controlled trial. *Gastrointest Endosc* 2012; **76**: 93-99 [PMID: 22595446 DOI: 10.1016/j.gie.2012.02.048]

68 **Sangchan A**, Chaiyakunapruk N, Supakankunti S, Pugkhem A, Mairiang P. Cost utility analysis of endoscopic biliary stent in unresectable hilar cholangiocarcinoma: decision analytic modeling approach. *Hepatogastroenterology* 2014; **61**: 1175-1181 [PMID: 25513063]

69 **Sawas T**, Al Halabi S, Parsi MA, Vargo JJ. Self-expandable metal stents versus plastic stents for malignant biliary obstruction: a meta-analysis. *Gastrointest Endosc* 2015; **82**: 256-267.e7 [PMID: 25982849 DOI: 10.1016/j.gie.2015.03.1980]

70 **Liberato MJ**, Canena JM. Endoscopic stenting for hilar cholangiocarcinoma: efficacy of unilateral and bilateral placement of plastic and metal stents in a retrospective review of 480 patients. *BMC Gastroenterol* 2012; **12**: 103 [PMID: 22873816 DOI: 10.1186/1471-230X-12-103]

71 **Dumas R**, Demuth N, Buckley M, Peten EP, Manos T, Demarquay JF, Hastier P, Caroli-Bosc FX, Rampal P, Delmont JP. Endoscopic bilateral metal stent placement for malignant hilar stenoses: identification of optimal technique. *Gastrointest Endosc* 2000; **51**: 334-338 [PMID: 10699784]

72 **Uchida D**, Kato H, Muro S, Noma Y, Yamamoto N, Horiguchi S, Harada R, Tsutsumi K, Kawamoto H, Okada H, Yamamoto K. Efficacy of Endoscopic Over 3-branched Partial Stent-in-Stent Drainage Using Self-expandable Metallic Stents in Patients With Unresectable Hilar Biliary Carcinoma. *J Clin Gastroenterol* 2015; **49**: 529-536 [PMID: 25159682 DOI: 10.1097/MCG.0000000000000213]

73 **Kawamoto H**, Tsutsumi K, Fujii M, Harada R, Kato H, Hirao K, Kurihara N, Nakanishi T, Mizuno O, Ishida E, Ogawa T, Fukatsu H, Sakaguchi K. Endoscopic 3-branched partial stent-in-stent deployment of metallic stents in high-grade malignant hilar biliary stricture (with videos). *Gastrointest Endosc* 2007; **66**: 1030-1037 [PMID: 17963891]

74 **Horgan AM**, Amir E, Walter T, Knox JJ. Adjuvant therapy in the treatment of biliary tract cancer: a systematic review and meta-

analysis. *J Clin Oncol* 2012; **30**: 1934-1940 [PMID: 22529261 DOI: 10.1200/JCO.2011.40.5381]

75 **Patel J**, Rizk N, Kahaleh M. Role of photodynamic therapy and intraductal radiofrequency ablation in cholangiocarcinoma. *Best Pract Res Clin Gastroenterol* 2015; **29**: 309-318 [PMID: 25966430 DOI: 10.1016/j.bpg.2015.02.008]

76 **Ortner ME**, Caca K, Berr F, Liebetruth J, Mansmann U, Huster D, Voderholzer W, Schachschal G, Mössner J, Lochs H. Successful photodynamic therapy for nonresectable cholangiocarcinoma: a randomized prospective study. *Gastroenterology* 2003; **125**: 1355-1363 [PMID: 14598251]

77 **Zoepf T**, Jakobs R, Arnold JC, Apel D, Riemann JF. Palliation of nonresectable bile duct cancer: improved survival after photodynamic therapy. *Am J Gastroenterol* 2005; **100**: 2426-2430 [PMID: 16279895]

78 **Hong MJ**, Cheon YK, Lee EJ, Lee TY, Shim CS. Long-term outcome of photodynamic therapy with systemic chemotherapy compared to photodynamic therapy alone in patients with advanced hilar cholangiocarcinoma. *Gut Liver* 2014; **8**: 318-323 [PMID: 24827630 DOI: 10.5009/gnl.2014.8.3.318]

79 **Park do H**, Lee SS, Park SE, Lee JL, Choi JH, Choi HJ, Jang JW, Kim HJ, Eum JB, Seo DW, Lee SK, Kim MH, Lee JB. Randomised phase II trial of photodynamic therapy plus oral fluoropyrimidine, S-1, versus photodynamic therapy alone for unresectable hilar cholangiocarcinoma. *Eur J Cancer* 2014; **50**: 1259-1268 [PMID: 24485665 DOI: 10.1016/j.ejca.2014.01.008]

80 **Dolak W**, Schreiber F, Schwaighofer H, Gschwantler M, Plieschnegger W, Ziachehabi A, Mayer A, Kramer L, Kopecky A, Schrutka-Kölbl C, Wolkersdörfer G, Madl C, Berr F, Trauner M, Püspök A. Endoscopic radiofrequency ablation for malignant biliary obstruction: a nationwide retrospective study of 84 consecutive applications. *Surg Endosc* 2014; **28**: 854-860 [PMID: 24196547 DOI: 10.1007/s00464-013-3232-9]

81 **Strand DS**, Cosgrove ND, Patrie JT, Cox DG, Bauer TW, Adams RB, Mann JA, Sauer BG, Shami VM, Wang AY. ERCP-directed radiofrequency ablation and photodynamic therapy are associated with comparable survival in the treatment of unresectable cholangiocarcinoma. *Gastrointest Endosc* 2014; **80**: 794-804 [PMID: 24836747 DOI: 10.1016/j.gie.2014.02.1030]

82 **Pai M**, Valek V, Tomas A, Doros A, Quaretti P, Golfieri R, Mosconi C, Habib N. Percutaneous intraductal radiofrequency ablation for clearance of occluded metal stent in malignant biliary obstruction: feasibility and early results. *Cardiovasc Intervent Radiol* 2014; **37**: 235-240 [PMID: 23842684 DOI: 10.1007/s00270-013-0688-x]

83 **Figueroa-Barojas P**, Bakhru MR, Habib NA, Ellen K, Millman J, Jamal-Kabani A, Gaidhane M, Kahaleh M. Safety and efficacy of radiofrequency ablation in the management of unresectable bile duct and pancreatic cancer: a novel palliation technique. *J Oncol* 2013; **2013**: 910897 [PMID: 23690775 DOI: 10.1155/2013/910897]

84 **Sharaiha RZ**, Sethi A, Weaver KR, Gonda TA, Shah RJ, Fukami N, Kedia P, Kumta NA, Clavo CM, Saunders MD, Cerecedo-Rodriguez J, Barojas PF, Widmer JL, Gaidhane M, Brugge WR, Kahaleh M. Impact of Radiofrequency Ablation on Malignant Biliary Strictures: Results of a Collaborative Registry. *Dig Dis Sci* 2015; **60**: 2164-2169 [PMID: 25701319 DOI: 10.1007/s10620-015-3558-3]

P- Reviewer: Espinel J, Kassir R, Li W, Mercado MA, Pinho R
S- Editor: Gong XM **L- Editor**: A **E- Editor**: Lu YJ

World Journal of Gastrointestinal Oncology

Submit a Manuscript: http://www.wjgnet.com/esps/
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
DOI: 10.4251/wjgo.v8.i6.509

World J Gastrointest Oncol 2016 June 15; 8(6): 509-519
ISSN 1948-5204 (online)

REVIEW

Genetic risks and familial associations of small bowel carcinoma

Santosh Shenoy

Santosh Shenoy, Department of Surgery, KCVA, University of Missouri at Kansas City, Kansas, MO 64128, United States

Author contributions: Shenoy S collected the data, wrote, and revised the manuscript.

Conflict-of-interest statement: There is no conflict of interest, no funding or financial disclosures.

Correspondence to: Santosh Shenoy, MD, FACS, Department of Surgery, KCVA, University of Missouri at Kansas City, 4801 E Linwood Blvd, Kansas, MO 64128, United States. shenoy2009@hotmail.com
Telephone: +1-816-8614700-55431
Fax: +1-816-9224609

Received: December 23, 2015
Peer-review started: December 24, 2015
First decision: January 30, 2016
Revised: February 2, 2016
Accepted: March 14, 2016
Article in press: March 16, 2016
Published online: June 15, 2016

Abstract

Adenocarcinoma of small intestines (SBA) is a relatively rare malignancy with poor outcomes due to delayed diagnosis. Fifty percent of patients have metastases on presentation and therefore early detection and treatment offers the best long term outcomes. Certain genetic polyposis syndromes and familial diseases are associated with increased risks for SBA. These include familial adenomatous polyposis (FAP), Lynch syndromes (LS), Juvenile polyposis syndrome, Peutz-Jeghers syndrome, Crohn's disease (CD) and celiac disease. Mutations in *APC* gene, Mismatch repair genes, *STK11* gene, and *SMAD4* gene have been implicated for the genetic diseases respectively. While there are no specific inherited genetic mutations for CD, genome-wide association studies have established over 140 loci associated with CD. CpG island mutations with defects in mismatch repair genes have been identified in celiac disease. Significant diagnostic advances have occurred in the past decade and intuitively, it would seem beneficial to use these advanced modalities for surveillance of these patients. At present it is debatable and no clear data exists to support this approach except for established guidelines to diagnose duodenal polyps in FAP, and LS. Here we discuss the genetic alterations, cancer risks, signaling mechanisms and briefly touch the surveillance modalities available for these genetic and clinical syndromes. English language articles from PubMed/Medline and Embase was searched were collected using the phrases "small-bowel adenocarcinoma, genetics, surveillance, familial adenomatous polyposis, lynch syndromes, Peutz-Jeghers syndrome, juvenile polyposis syndrome, CD and celiac disease". Figures, tables and schematic diagram to illustrate pathways are included in the review.

Key words: Small intestinal adenocarcinoma; Genetic risks; Mutations; Signaling pathways; Surveillance

Core tip: Adenocarcinoma of small intestine (SBA) is a relatively rare malignancy with poor outcomes due to delayed diagnosis. Certain genetic and familial diseases are associated with increased risks for SBA. These include Familial adenomatous polyposis, lynch syndromes, juvenile polyposis syndrome, Peutz-Jeghers syndrome, Crohn's disease and celiac disease. We discuss the clinical implications of

this aggressive cancer focusing on the genetic and familial associations, signaling mechanisms and available diagnostic modalities for surveillance.

Shenoy S. Genetic risks and familial associations of small bowel carcinoma. *World J Gastrointest Oncol* 2016; 8(6): 509-519 Available from: URL: http://www.wjgnet.com/1948-5204/full/v8/i6/509.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i6.509

INTRODUCTION

Small intestine comprises majority of the anatomical length and absorptive surface of gastrointestinal (GI) tract but accounts for less than five percent of GI tract malignancies[1]. According to the seer's database an estimated 9410 new small bowel adenocarcinoma (SBA) cases and 1260 deaths may have occurred in the United States in 2015[2].

Certain genetic syndromes and familial diseases are associated with SBA (Table 1). These are a heterogeneous group of familial polyposis and non-polyposis syndromes, inflammatory bowel diseases, and autoimmune diseases with distinct epidemiology, genetics, clinical presentation, treatment strategies, surveillance and outcomes.

These groups with inherent risk for both small bowel and colorectal cancers (CRC) have established surveillance recommendations for CRC but there are no clear guidelines for surveillance of small bowel cancers. Significant diagnostic advances have occurred in the past decade and patients may benefit for small bowel surveillance using these diagnostic modalities.

FAMILIAL ADENOMATOUS POLYPOSIS

Familial adenomatous polyposis (FAP) is an autosomal dominant genetic disorder affecting approximately 1:10000 newborns caused by mutation of the *APC* gene on the long arm of chromosome 5. Multiple polyps of the colon and rectum are pathognomic of FAP. Polyps could be sessile or pedunculated and histology's may vary from tubular to villous adenoma. Most patients develop polyps by second decade and if untreated colon malignancy by the fourth decade (15% of gene carriers by age 10 years, 75% by 20 years, and 90% by 30 years)[3].

The incidence of small intestinal cancers in FAP is not clear however the adenoma-carcinoma sequence for development of cancer is well established[3-6]. In addition these patients are predisposed to multiple small bowel adenomatous polyps usually in the duodenum and periampullary region[7,8].

The pathogenesis of these polyps is due to dysregulation of the canonical Wnt/β-catenin pathway. The APC protein is a tumor suppressor, involved in cell adhesion, transduction and transcription, cell cycle control, maintenance of fidelity of chromosomal segregation and apoptosis. As part of a scaffolding protein complex, it is a negative regulator of Wnt signaling pathway (Figure 1).

In the absence of Wnt signaling, cytosolic β-catenin that is not bound by cell-cell adherens junction is transferred to the degradation complex consisting of the proteins APC, axin, casein kinase 1 (CK1) and glycogen synthase kinase 3 (GSK3). CK1 and GSK3 phosphorylate and prime the unbound β-catenin targeting it for ubiquitination, and leads it to proteasome to be digested. This prevents translocation and accumulation of β-catenin into the nucleus.

Normally nuclear translocation of β-catenin leads to the expression of genes such as c-Myc and Wnt target genes: Promoting cell growth, division, proliferation and differentiation. It also regulates cell-cell adhesion and is important for tissue formation[4,5].

More than 700 mutations of *APC* gene have been identified with the classic and attenuated types of FAP. *APC* gene mutation leads to production of truncated, nonfunctional version of this protein. This truncated APC protein fails to suppress the canonical Wnt/β-catenin pathways even in the absence of Wnt signaling, results in unopposed translocation of β-catenin in the nucleus and stimulates transcription of c-Myc and other Wnt target genes that leads to the formation of polyps and predispose to cancers[4,5]. In addition APC also interacts with microtubules, loss of APC may lead to mitotic spindle defects, leading to chromosome abnormalities when cells divide.

While colorectal polyps and cancer remains the primary tumors in FAP and advanced surgical techniques have reduced mortality from colorectal carcinoma, the leading second primary malignancy in these individuals is duodenal and small bowel carcinoma. The prevalence of duodenal adenomas is 50%-90% and these patients carry a relative risk of 330 for adenocarcinoma or up to 5% lifetime risk. The risk is highest in periampullary adenomas[9]. The adenoma formation is not restricted to the duodenum but also noted in jejunum and ileum in 50%-75% of the patients. Studies using video capsule endoscopy (VCE) and balloon-assisted enteroscopy (BAE) confirm the presence of jejunal and ileal polyps frequently in FAP, especially with extensive duodenal polyposis[10-12].

The increased risk for SBA appears to correlate between the severity of duodenal polyposis and presence of jejunal polyps[10-13]. Scattered case series, report an association of marked duodenal polyposis, with higher stages of the disease on diagnosis and worse prognosis[7]. Spigelman in 1989 developed an endoscopic scoring system (stage 1-4) to describe the severity of duodenal polyps in FAP. Predictors include the number, size, histology and the degree of dysplasia[8]. The risk of progression to adenocarcinoma is associated with the size and histology of these polyps: 8.3% risk for sub centimeter polyps to 30% for polyps greater than 2 cm. Tubular adenoma carries a risk of (14%), increases to (23%) for tubulo-villous adenoma, and (36%) for villous

Table 1 Genetic risks and familial associations of small bowel carcinoma

Syndrome	Mode of inheritance	Mutated/associated gene	Relative risk (95%CI)	Lifetime risk for SBA	Polyps/pathway
FAP[7,29]	Autosomal dominant (AD)	APC	330 (132-681)	3%-5%	Adenoma-carcinoma
HNPCC/ LS[13,14,29]	AD	MMR (MSH2, MSH6, MLH1 ,PMS2)	291 (71-681)	1%-4%	Adenoma-carcinoma
PJS[31,36,37]	AD	STK11	500 (220-1306)	1.7%-13%	Hamartoma, adenoma-Ca
JPS[38,41]	AD	BMPR1A, SMAD4	Unknown	Unknown	Hamartoma, adenoma-Ca
Crohn's disease	Unknown (genome wide studies have associated 140 loci)	Unknown	30-60[44-49] (15-609)	2.2% after/25 yr	Dysplasia-carcinoma
Celiac disease	Association with HLA-DQ2,HLA-DQ8	Unknown	60-80[61-63] (7-240)	< 1%	Adenoma-carcinoma

FAP: Familial adenomatous polyposis; APC: Adenomatous polyposis coli; HNPCC: Hereditary nonpolyposis colorectal cancer; MMR: Mismatch repair gene; LS: Lynch syndrome; PJS: Peutz-Jeghers syndrome; STK11: Serine threonine kinase; JPS: Juvenile polyposis syndrome; BMPRIA: Bone morphogenetic protein receptor, type IA; SMAD4: Mothers against decapentaplegic homolog; HLA: Human leukocyte antigen complex.

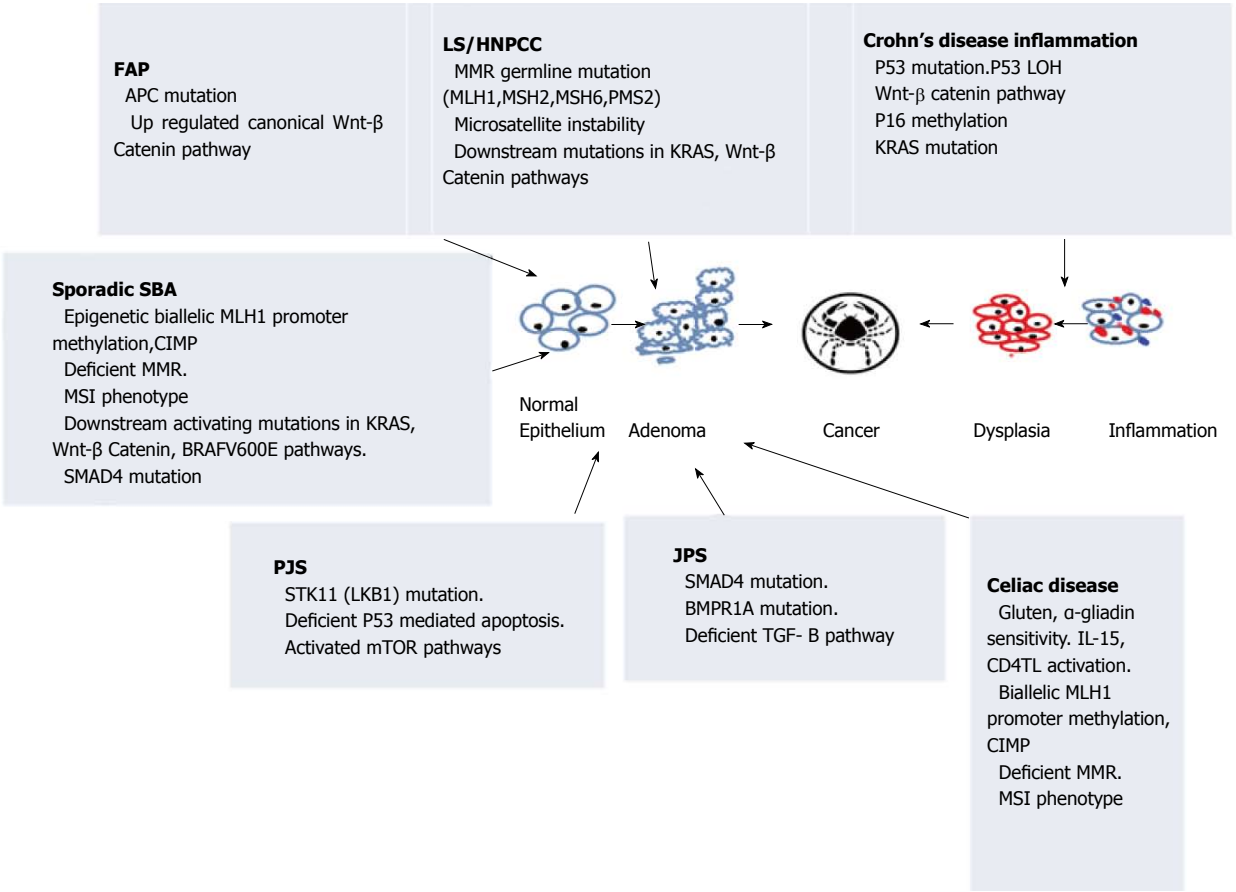

Figure 1 Schematic drawing of genetic and molecular pathways predisposing to small bowel carcinoma. Wnt: Wingless-type MMTV integration site family; KRAS: Kirsten rat sarcoma viral oncogene homolog; LOH: Loss of heterozygosity; CIMP: CpG island methylator phenotype; MSI: Microsatellite instability; BRAFV600E: V-raf murine sarcoma viral oncogenes homolog B; mTOR: Mammalian target of rapamycin; TGF-β: Transforming growth factor β; IL: Interleukin; CD4TL: CD4 T-lymphocytes.

adenomas[1,14,15]. The significance of small bowel polyps beyond the duodenum is not defined given the fact that up to 44% of patients with FAP develop extensive (stage 4) duodenal polyps with aging but overall incidence of cancer is less than 5%[13,16].

Due to this reason gastroduodenal surveillance with endoscopy is generally limited for duodenal polyps[7]. The exact age and interval to begin surveillance upper endoscopy is still debatable, some authors recommend annual endoscopy starting after colonic polyps are diagnosed or as early as 15 years of age[17] while other authors suggest starting at age 25 years and interval

Figure 2 65-year-old male with familial adenomatous polyposis, previous total proctocolectomy 35 years ago with ileostomy adenocarcinoma: Polypoid growth at the ileostomy orifice.

based on severity as suggested by Spigelman grading system[13,18].

Based on the existing data there are no recommendations or guidelines for surveillance of small bowel beyond the duodenum in FAP. Further research is required to identify what patients with FAP are at an increased risk for small bowel carcinoma[13,19].

One unique subset of patients is FAP with ileostomy and ileoanal pouch carcinoma. Currently prophylactic restorative proctocolectomy with ileoanal pouch anastomosis (IPAA) is the preferred operation in FAP. Previously total proctocolectomy with end ileostomy was the operation most often performed for FAP[20,21]. These patients with functioning ileostomies have an inherent risk for development of ileostomy adenocarcinoma (Figure 2). Adenomas frequently form in 35% of ileoanal pouches, examined in FAP who underwent restorative proctocolectomy[22]. The risk of developing adenomas increases with the longevity of these functioning ileostomies. The estimated risks at 5, 10 and 15 years were 7%, 35%, 75% respectively. This predilection to form adenomas, may progress to adenocarcinoma. Positive immunostaining of β-catenin, p53 and frequent occurrence of KRAS mutations suggests adenoma-carcinoma sequence similar to colorectal cancers[23]. The current recommendations for these patients is periodic clinical and endoscopic examination of their stomas and pouches with biopsies of any suspicious lesions[21,24].

LYNCH SYNDROME

Lynch syndromes (LS) are an autosomal dominant genetic disorder with germline mutations of mismatch repair genes (MMR): MLH1, MSH2, MSH6 and PMS2. MLH1 and MSH2 mutation variants represent about 90% of families with LS; MSH6 variants in another 7%-10% and PMS2 mutation in less than 5%. Germline deletions in EPCAM (epithelial cell adhesion molecule) inactivate MSH2 in a small subset (< 1%) of patients with LS[17].

Affected individuals carry the risk for colorectal, endometrial and ovary, genitourinary tract, stomach, hepatobiliary, pancreas and small bowel cancers (Figure 3).

The pathogenesis of these tumors involves microsatellites, which are short stretches of DNA with repetitive sequences of nucleotides and are susceptible to acquiring errors when *MMR* gene function is impaired. *MMR* genes present on different chromosomes coordinate the activities of other proteins such as DNA polymerase that maintain the fidelity of DNA replication and genomic integrity. MMR system encode for proteins that form DNA MMR complexes. These correct small insertions or deletions that may occur during somatic division. Thus MMR system proofreads and repairs defects that were overlooked by DNA polymerase.

Cancerous cells with defective *MMR* gene function exhibit microsatellite instability. This refers to an inconsistent number of microsatellite nucleotide repeats when compared to normal tissue. This phenotype with a markedly high rate of mutations involving cell-cycle regulation increases the risk of malignancy (Figure 1)[17].

Immunochemistry of the tumor samples are used to detect the absence of the protein products of mismatch repair genes. These gene products function as dimers: MSH2 protein may complex with MSH6 or MSH3 protein, and MLH1 protein complexes with PMS2 or PMS1 protein. MSH6 and PMS2 proteins are unstable when unpaired. A pathogenic variant in MSH2 typically results in loss of expression of the proteins MSH2/MSH6 and a germline pathogenic variant in MLH1 results in loss of expression of the proteins MLH1/PMS2. Germline pathogenic variants in MSH6 and PMS2 typically do not result in loss of MSH2 or MLH1 expression because these proteins are still present in other pairings.

LS accounts for 3% to 5% of all CRC[25] and it is the commonest inherited colon cancer syndrome. The average age of malignancy in LS is 44 years, *vs* 64 years in sporadic CRC[3,17].

The risk factors for SBA in LS patient's increase with age, beginning at 40 years and a tenfold rise by the age of 60[26]. Compared to sporadic SBA in general population, patients with LS present a decade earlier. About 10% of patients develop cancers before the age of 30[27]. The lifetime risk for SBA is estimated as 1%-4% and is greater than 100 fold risk compared to general population[13,14,28].

In 30%-70% patients with LS small bowel cancer may be the primary malignancy to manifest[29]. The incidence appears higher in MLH1, and MSH2 carriers compared to MSH6[13,29]. Further regional variations between various registries have been noted for the incidence of small bowel cancer. For instance Finnish and French (HNPCC/LS) patients have lesser incidence of small bowel cancer compared to Dutch (HNPCC/LS) patients[28,29].

Most data from series of patients point to adenoma-carcinoma sequence comparable to colorectal neoplasia. Molecular data as described earlier indicate accumulation of mutations as an inciting event in the development of small bowel cancers similar to colorectal cancers. Some authors recommend that patients presenting with SBA routinely undergo analysis of the MMR phenotype

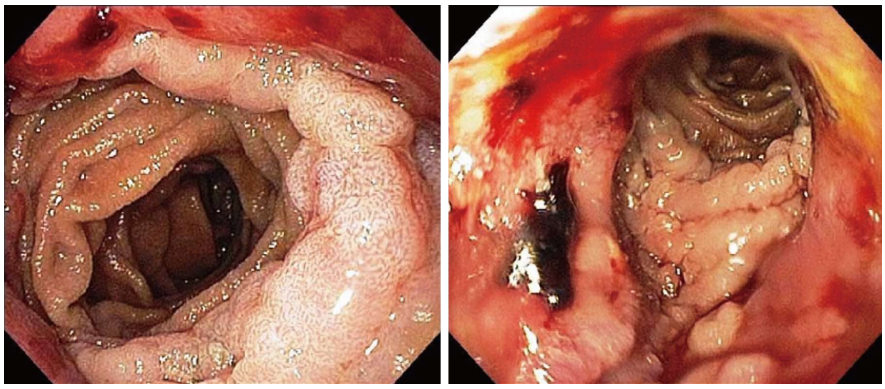

Figure 3 Sixty-nine-year-old male with family history of Lynch syndrome, jejunal adenocarcinoma, viewed on small bowel enteroscopy.

and screened for LS[13,28,29]. This is especially true for histological findings of mucinous tumors infiltrated with lymphocytes and pushing tumor border suggestive of MSI phenotype in 75% patients[29]. There are implications in choosing adjuvant chemotherapy regimen in this phenotype, as cancers deficient in MMR proteins may be resistant to 5-FU based chemotherapy[30].

Upper endoscopic surveillance is recommended over the age of 30 years for gastric and duodenal polyps however at present there are no guidelines for small bowel cancer surveillance in LS[13,14,17,26,28].

PEUTZ-JEGHERS SYNDROME

Peutz-Jeghers syndrome (PJS) is an autosomal dominant condition with mutation in the serine threonine kinase 11 (*STK11*) genes on the short arm of chromosome 19. The incidence of PJS is reported to be 1 in 50000 to 1 in 200000 live births. PJS is characterized by melanin spots on the buccal mucosa and predilection to form multiple gastrointestinal hamartomas and polyps. These are scattered throughout the small bowel, predominantly in the jejunum and ileum.

The *STK11* gene (also called *LKB1*) encodes for enzyme serine/threonine kinase 11[31]. STK11 is a tumor suppressor gene and associates with TP53 to regulate TP53-dependent apoptosis pathways[32]. It also has a role in cell polarity, cell metabolism and energy homeostasis[33]. Inactivation of STK11 is an early event in the development of hamartoma and adenocarcinoma. In addition to loss of STK11 function and altered TP53 expression, adenocarcinomas in PJS also demonstrate loss of heterozygosity (LOH) in 17p and 18q. These deletions are associated with an increased tendency of disease dissemination in colorectal cancer. STK11 also exerts its inhibitory effects by phosphorylating and activating 14 protein kinases, all related to the AMP-activated protein kinases (AMPK)[33]. AMPK is an evolutionally conserved serine threonine kinase and its activation by STK11 leads to upregulation of signaling through the TSC (Tuberous sclerosis) complex. This in turn negatively regulates mTOR pathways. Loss of STK11 activity leads to increased mTOR activity and characterized by an increased risk of malignancy (Figure 1)[31,33,34].

Hamartomatous and adenoma polyps are scattered throughout the small bowel, predominantly in the jejunum and ileum. Patients with PJS are predisposed to multiple GI tract and non GI tract malignancies which include breast, ovaries, testicular, pancreas, esophagus, stomach and non-small cell lung cancers[34,35].

SBA has been known to occur in PJS. Meta-analysis of SBA in PJS compared to general population indicates a relative risk of 520[36]. The life time incidence for adenocarcinoma is 1.7%-13% and rises rapidly in elderly[36,37]. Adenocarcinoma originates from both adenomas and hamartomas. Intraepithelial neoplasia is observed in the hamartoma lesions[29,36,37]. Due to the rarity of this condition, current surveillance protocols are not evidence-based. Endoscopies are performed more often to detect polyps which may pose a risk for intussusception, obstruction rather than cancers. Routine screening is recommended, beginning at age 18 with every 2-3 year interval[31,35,36]. Recent study suggests surveillance with VCE beginning at the age 8 years and performed every three years if polyps are detected at initial examination. With a negative initial exam, surveillance should recommence at 18 years[31].

JUVENILE POLYPOSIS SYNDROME

Juvenile polyposis syndrome (JPS) is an autosomal dominant disorder which is characterized by multiple hamartomatous polyps of the gastrointestinal tract and is the most common hamartomatous polyp syndromes with prevalence estimated to be between 1 in 16000 to 1 in 100000[3,38]. Juvenile refers to the sporadic inflammatory hamartomatous polyps of childhood, rather than the age of onset. Most affected individuals have some polyps by age 20 years[3]. Most are benign polyps, but malignant transformation may occur resulting in increased lifetime risk for colon (10%-40%) and stomach (21%) cancers and less commonly involving the small bowel and pancreas. The lifetime risk of SBA has been difficult to estimate due to the rarity of the disease and is also reduced by screening polypectomies. Malignant transformation occurs through traditional adenoma to cancer transformation sequence. Multiple genetic alterations similar to colorectal neoplasia also play a role in neoplastic transformation of juvenile polyps[39].

Two genes, SMAD4, BMPR1A, have been implicated in

the pathogenesis of polyps in JPS. They encode proteins for either, transforming growth factor-β (TGF-β) or bone morphogenetic protein (BMP) signaling pathways. The *SMAD* gene on chromosome 18q21.1, adjacent to DCC (deleted in colon cancer) is a part of the TGF-β signal transduction pathway. SMAD4 proteins transmit TGF-β related growth-suppressing signals from cell membranes to nucleus mediating growth inhibition and apoptosis. The SMAD4 protein serves both as a transcription factor and as a tumor suppressor[3]. More than 60 mutations in the *SMAD4* gene have been implicated in JPS. This results in the production of a truncated, nonfunctional protein thereby preventing transmission of TGF-β growth suppressing signals from the cell surface to the nucleus (Figure 1) leading to unregulated cell growth and susceptibility to polyp formation in JPS.

Mutations in BMPR1A on chromosome 10 are found in 20% to 25% of individuals with JPS[40]. BMPR1A is a serine-threonine kinase (STK) type Ⅰ receptor of the TGF-β superfamily, which when activated leads to phosphorylation of SMAD4 proteins. Mutations result in abnormal BMPR1A protein which cannot bind to ligands in the TGF-β pathway and interferes with the activation of the SMAD protein complex[41].

Given the rarity of this disease there is no data on the incidence, relative risks, or life time risks of SBA and at present no guidelines exist for surveillance. Some authors do recommend upper endoscopy every 3-5 years from age 15, and repeated annually if polyps are diagnosed[42].

CROHN'S DISEASE

Crohn's disease (CD) is an autoimmune inflammatory bowel disease affecting the GI tract with predilection for small intestine. The prevalence in North America ranges from 26.0 to 198.5 cases per 100000 persons. The incidence rates range from 3.1 to 14.6 cases per 100000 people per year[43]. CD is characterized by transmural granulomatous inflammation of the small bowel in a discontinuous fashion and a tendency to form stenosis, strictures and fistulae. Adenocarcinoma of small intestines is a rare complication of CD with meta-analysis showing relative risks reported to be between 30 as 60 (95%CI: 15.9-60.9) compared to the general population[44-48] and cumulative risk of 2.2% after 25 years of regional ileitis[48,49]. The risk increases with chronicity of the disease, young age of onset, male sex, distal small bowel disease with strictures and fistulae.

CD results from abnormal mucosal immune response to environmental factors in genetically susceptible hosts. The granulomatous inflammation comprises of aggregates of macrophages, lymphocytes, plasma cells, and multi-nucleated giant cells that are formed in response to the release of inflammatory cytokines such as tumor necrosis factor[15,50]. Etiologies in the pathogenesis of inflammatory bowel disease include genetic susceptibility, environmental, microbial factors and their interaction with intestinal epithelial cells and components of innate and adaptive immune system. Genetic susceptibility is confirmed with higher prevalence in monozygotic twins and the familial clustering of the disease. A meta-analysis of six twin studies with a combined set of 112 MZ and 196 DZ twin pairs reported concordance rates of 30.3% and 3.6% respectively[51]. Since 2006, genome-wide association studies have established over 140 loci associated with CD risk, however the significance and the contribution to the disease risk remains to be defined[52].

The GI tract is continuously exposed to commensal internal flora and also pathogenic organisms and other environmental antigens. The integrity of the mucosal barrier is maintained by tight junctions occurring between adjacent epithelial cells and the relative impermeability of the apical villous epithelium which serves as an important function in the innate immune system. Complementing these are other cells such as Paneth cells which secrete antimicrobial substances such as, lysozymes, cysteine-rich defensins, and IgA and goblet cells which secretes mucus[15,53]. These and other intrinsic defense mechanisms in the intestinal mucosa dilute, limit the adherence and invasion of commensal and pathogenic microorganisms and antigens. Alteration of this barrier leads to abnormal immune response by the effector lymphocytes and other proinflammatory cytokines leading to a state of chronic intestinal inflammation and its sequelae. Inflammatory cytokines produced by the immune system includes interleukins, chemokines, growth factors, and extracellular proteases. They interact with cell surface receptors and subsequently target genes which influence clonal neoplastic proliferation, angiogenesis and invasion through the basement membrane. In addition, excessive formation of reactive oxygen and nitrogen free radicals are potentially damaging to DNA and the integrity of cell surface membranes[15].

Adenocarcinoma in CD is seen in the effected segments of the bowel which suggests inflammation-dysplasia-carcinoma sequence[45,48,54,55]. Genetic alterations occur, which transform dysplastic mucosa to carcinoma. The prevalence of *MSI*, *APC*, *DCC* gene mutations are low, one study however showed 43% of patients with adenocarcinoma in CD carry K-RAS mutations, and overexpression of *p53* gene product in 71% of Crohn's associated carcinoma[54]. Overexpression of p53 is helpful to elucidate transformation from inflammation to dysplasia as inflammation does not overexpress p53[55]. A mutational analyses of multiple areas of intestine from ten patients with CD and intestinal cancer, mutations in KRAS, CDKN2A (p16), and TP53 that were observed in tumor cells was also present in non-tumor, and both nondysplastic and dysplastic epithelium suggestive of a field defect in CD[56].

Another study on 41 patients with CD and small bowel cancer showed dysplasia association in 50% of the patients suggesting an inflammation-dysplasia-adenocarcinoma sequence in CD-related SBA, similar to what is observed in chronic colitis-related colorectal cancer (Figure 1)[55,56]. The rarity of adenocarcinoma in CD makes mutation studies difficult. Perhaps analysis in multinational pooled data may reveal more information.

Symptoms highly suspicious for adenocarcinoma are development of a new small bowel stricture refractory to steroids or maximal medical management or a long standing quiescent disease with newly diagnosed small bowel obstruction. These warrant attention without delay. Compared to adenocarcinoma arising *de novo*, adenocarcinoma in CD present at a median age of 48 years, is more common in males, ileum as most common site and mucinous signet ring cell is more frequently seen[57]. Early diagnosis and small bowel resection offers the best success for long term survival. Unfortunately majority of adenocarcinoma are diagnosed on post-operative specimens of resected bowel with metastatic nodal disease noted in 50% and distant metastases in 40% of patients. At present however there are no surveillance guidelines to detect SBA in patients with CD however study investigating the benefit of endoscopic surveillance of the small bowel lesions greater than 10 years duration is in progress[55].

CELIAC DISEASE

Celiac disease is a chronic inflammatory autoimmune small intestinal disorder due to gluten sensitivity, an antigen in wheat, barley, rye and malt. It occurs in adults and children and effects 1% of the population. Celiac disease is associated with both human leukocyte antigen (HLA) and non-HLA genes and with other immune disorders, notably juvenile diabetes and thyroid disease. It is genetically associated with individuals positive for human leukocyte antigen-DQ2 or DQ 8. Familial aggregation is noted with 70% concordance in monozygotic twins[58]. α-gliadin; a component of gluten is a 33 amino acid peptide sequence and is resistant to degradation by the proteases in the human intestines. Immune response to gliadin promotes inflammatory reaction in the small bowel. Infiltration of the lamina propria and the epithelium with chronic inflammatory cells (predominantly CD4 lymphocytes) triggers a cascade releasing cytokines, interferon-γ, interleukin-15 and metalloproteinases resulting in destruction of enterocytes, crypt hyperplasia and villous atrophy[59,60].

Patients with celiac disease have an increased risk for enteropathy associated lymphomas as well as adeno-carcinoma of the small intestine compared to the general population[59,61]. Given the rarity of celiac disease and adenocarcinoma the true incidence is difficult to ascertain, however the reported relative risk is increased between 60-80 compared to the general population[61-63]. Most commonly seen in jejunum, the natural history seems to follow the adenoma-carcinoma sequence as seen in colorectal neoplasms. Small bowel mucosa in celiac disease does not show any premalignant field defect or dysplasia in mucosa adjacent to the adenocarcinoma. However the mechanism for formation of adenomas in celiac disease has not yet been elucidated[64].

Recent molecular studies have shown that celiac disease associated adenocarcinomas in the elderly are characterized by high level of CpG island methylation (CIMP), MLH1 inactivation, microsatellite instability (MSI) and defect in the MMR pathways (Figure 1)[65-67]. Methylation of CpG sites within the promoters of genes can lead to their silencing. This feature is found in a number of human cancers. Similar to LS as described earlier, celiac disease should be considered in the differential diagnosis in patients presenting with sporadic SBA, in the elderly, especially with MSI positivity[65-67]. These sporadic and celiac associated tumors however show CIMP (CpG island methylator phenotype) and BRAFV600E hotspot mutations that serve to distinguish them from LS cases.

The risk for adenocarcinoma rises in longstanding, untreated celiac disease. Symptoms of celiac disease diagnosed in children and treated with gluten free diet often improve. This may create a false notion of having overcome the disease, with resurgence later in life. Development of new symptoms of weight loss, abdominal pain, anemia, blood loss, and fever in patients who were on a gluten free diet should raise suspicion of neoplastic transformation and should be thoroughly evaluated[59]. At present there are no guidelines for small bowel surveillance for adenocarcinoma or lymphoma in asymptomatic patients with celiac disease.

SURVEILLANCE MODALITIES FOR SMALL BOWEL

The manifestations of small bowel malignancy are generally nonspecific and often diagnosed in advanced stages. Fifty percent of patients have metastases at diagnosis. Mean duration of symptoms before diagnosis is 10 mo[68]. Diagnosis is often made with a combination of diagnostic tests which includes both endoscopy and radiography. Considerable advances have occurred in endoscopic techniques with introduction of capsule endoscopy and balloon assisted endoscopy. Also advances in both computed tomography (CT) and magnetic resonance imaging (MRI) enterography and enteroclysis are playing an increasing role in evaluation of small bowel diseases.

Endoscopy

Esophagogastroduodenal (EGD) endoscopy with front and side viewing camera is the standard diagnostic procedure and is accurate in identifying, biopsy of lesions proximal to the ligament of Trietz. Push enteroscopy can visualize the duodenum, proximal jejunum while balloon assisted enteroscopy (BAE) can visualize the entire small bowel (Figure 3). However the latter techniques are time consuming, technically challenging and often requires deep sedation or general anesthesia[69]. BAE encompasses both single and double balloon techniques and can be performed through the oral or anal route. A complete small bowel examination can be accomplished in up to 80% of the patients. It carries the advantage of ability to perform endoscopic interventions such as biopsy, polypectomy and marking the lesion[69-71]. A fewer studies

utilizing BAE techniques have confirmed the presence of small bowel polyps in patients with FAP[10,71,72].

Video capsule endoscopy

Video capsule endoscopy (VCE) has become one of the most important investigational tools for small bowel mucosal evaluation. Due to ease of the procedure it has become a first line tool to detect small bowel abnormalities in non-obstructed patients for evaluation of small intestinal diseases such as occult GI bleeding, suspected CD, celiac disease, small bowel tumors, and motility disorders[73]. Most VCE studies show the presence of small bowel polyps ranging 50%-87% in patients with FAP[11,12] and there are a few case series suggesting the role of VCE in LS[74,75]. A study comparing VCE to MRI showed the advantage of VCE to detect smaller polyps. Polyps larger than 15 mm were detected equally in both groups, whereas smaller polyps were seen much more often with capsule endoscopy. Polyps that were smaller than 5 mm were exclusively seen with capsule endoscopy. However, location of the detected polyps and determination of their exact sizes was more accurate by MRI[76,77].

Drawback for VCU include capsule retention, missed polyps < 1 cm, especially duodenal polyps (due to rapid transit)[73,78]. Using combination of VCE and subsequent BAE for endoscopic intervention offers an ideal method of surveillance and treatment in these polyposis syndromes, avoiding a laparotomy. The value of such approach is yet to be demonstrated[13].

CT and MRI enterography and enteroclysis

Advances in temporal and spatial resolution offered by CT scan and MRI scan with newer enteric agents used to distend the small bowel have replaced barium radiography as the preferred diagnostic tests. Both CT and MRI scan provide details of the bowel wall and the mesentery and the surrounding viscera. Enterography entails using oral contrast while for enteroclysis a nasojejunal tube need to be inserted to deliver the contrast. Enteroclysis provides better bowel distension offers improved mucosal details. MRI enteroclysis has been shown to be a more dynamic and sensitive than CT enteroclysis for mucosal details. These are due to better soft tissue contrast that is achieved with MRI[79,80]. A study on 150 patients with MRI enteroclysis showed sensitivity, specificity of 86% and 98% respectively[81]. A recent study compared VCE to MRI enteroclysis with results showing higher specificity of MRI images in detecting small bowel lesions[82]. The authors attributed this to the distension of the small bowel with enteroclysis and a three dimensional views compared to a unidirectional view of the VCE. Secondly MRI enteroclysis may be beneficial in stenosis or strictures in small bowel disease as the risk of capsule retention are eliminated.

CONCLUSION

Certain genetic and familial diseases are associated with increased risks for SBA. The pathogenesis and molecular mechanisms for some of these syndromes are described and the risk varies according to the types of polyps and polyposis syndromes. Although the overall incidence of SBA is low the prognosis remains dismal due to nonspecific symptoms and often a delay in diagnosis. Intuitively it would seem that use of surveillance modalities may benefit these patients at higher risk for SBA. At present it is debatable and there is no data to support this approach except for established guidelines to diagnose duodenal polyps in FAP, and LS. Further research, perhaps multi-institutional study is warranted focusing on identifying patients who are at risk for small intestinal adenocarcinoma and on optimal surveillance strategies.

REFERENCES

1 **Shenoy S.** Primary small-bowel malignancy: update in tumor biology, markers, and management strategies. *J Gastrointest Cancer* 2014; **45**: 421-430 [PMID: 25339426 DOI: 10.1007/s12029-014-9658-z]

2 Surveillance, Epidemiology, and End Results (SEER) Program. SEER*Stat Database: Statistics at a Glance: Small Intestine Cancer. [accessed 2015 Oct 15]. Available From: URL: http://www.seer.cancer.gov

3 **Mishra N**, Hall J. Identification of patients at risk for hereditary colorectal cancer. *Clin Colon Rectal Surg* 2012; **25**: 67-82 [PMID: 23730221 DOI: 10.1055/s-0032-1313777]

4 **Fearnhead NS**, Britton MP, Bodmer WF. The ABC of APC. *Hum Mol Genet* 2001; **10**: 721-733 [PMID: 11257105 DOI: 10.1093/hmg/10.7.721]

5 **Näthke IS**. The adenomatous polyposis coli protein: the Achilles heel of the gut epithelium. *Annu Rev Cell Dev Biol* 2004; **20**: 337-366 [PMID: 15473844 DOI: 10.1146/annurev.cellbio.20.012103.094541]

6 **Galiatsatos P**, Foulkes WD. Familial adenomatous polyposis. *Am J Gastroenterol* 2006; **101**: 385-398 [PMID: 16454848 DOI: 10.1111/j.1572-0241.2006.00375.x]

7 **Offerhaus GJ**, Giardiello FM, Krush AJ, Booker SV, Tersmette AC, Kelley NC, Hamilton SR. The risk of upper gastrointestinal cancer in familial adenomatous polyposis. *Gastroenterology* 1992; **102**: 1980-1982 [PMID: 1316858]

8 **Spigelman AD**, Williams CB, Talbot IC, Domizio P, Phillips RK. Upper gastrointestinal cancer in patients with familial adenomatous polyposis. *Lancet* 1989; **2**: 783-785 [PMID: 2571019 DOI: 10.1016/S0140-6736(89)90840-4]

9 **Kadmon M**, Tandara A, Herfarth C. Duodenal adenomatosis in familial adenomatous polyposis coli. A review of the literature and results from the Heidelberg Polyposis Register. *Int J Colorectal Dis* 2001; **16**: 63-75 [PMID: 11355321 DOI: 10.1007/s003840100290]

10 **Matsumoto T**, Esaki M, Yanaru-Fujisawa R, Moriyama T, Yada S, Nakamura S, Yao T, Iida M. Small-intestinal involvement in familial adenomatous polyposis: evaluation by double-balloon endoscopy and intraoperative enteroscopy. *Gastrointest Endosc* 2008; **68**: 911-919 [PMID: 18561922 DOI: 10.1016/j.gie.2008.02.067]

11 **Schulmann K**, Hollerbach S, Kraus K, Willert J, Vogel T, Möslein G, Pox C, Reiser M, Reinacher-Schick A, Schmiegel W. Feasibility and diagnostic utility of video capsule endoscopy for the detection of small bowel polyps in patients with hereditary polyposis syndromes. *Am J Gastroenterol* 2005; **100**: 27-37 [PMID: 15654777 DOI: 10.1111/j.1572-0241.2005.40102.x]

12 **Plum N**, May A, Manner H, Ell C. Small-bowel diagnosis in patients with familial adenomatous polyposis: comparison of push enteroscopy, capsule endoscopy, ileoscopy, and enteroclysis. *Z Gastroenterol* 2009; **47**: 339-346 [PMID: 19358059 DOI: 10.1055/s-2008-1027984]

13 **Koornstra JJ**. Small bowel endoscopy in familial adenomatous polyposis and Lynch syndrome. *Best Pract Res Clin Gastroenterol*

2012; **26**: 359-368 [PMID: 22704577 DOI: 10.1016/j.bpg.2012.01.022]
14 **Raghav K**, Overman MJ. Small bowel adenocarcinomas--existing evidence and evolving paradigms. *Nat Rev Clin Oncol* 2013; **10**: 534-544 [PMID: 23897080 DOI: 10.1038/nrclinonc.2013.132]
15 **Schottenfeld D**, Beebe-Dimmer JL, Vigneau FD. The epidemiology and pathogenesis of neoplasia in the small intestine. *Ann Epidemiol* 2009; **19**: 58-69 [PMID: 19064190 DOI: 10.1016/j.annepidem.2008.10.004]
16 **Mathus-Vliegen EM**, Boparai KS, Dekker E, van Geloven N. Progression of duodenal adenomatosis in familial adenomatous polyposis: due to ageing of subjects and advances in technology. *Fam Cancer* 2011; **10**: 491-499 [PMID: 21416262 DOI: 10.1007/s10689-011-9433-2]
17 **Lynch HT**, Lynch PM, Lanspa SJ, Snyder CL, Lynch JF, Boland CR. Review of the Lynch syndrome: history, molecular genetics, screening, differential diagnosis, and medicolegal ramifications. *Clin Genet* 2009; **76**: 1-18 [PMID: 19659756 DOI: 10.1111/j.1399-0004.2009.01230.x]
18 **Vasen HF**, Möslein G, Alonso A, Aretz S, Bernstein I, Bertario L, Blanco I, Bülow S, Burn J, Capella G, Colas C, Engel C, Frayling I, Friedl W, Hes FJ, Hodgson S, Järvinen H, Mecklin JP, Møller P, Myrhøi T, Nagengast FM, Parc Y, Phillips R, Clark SK, de Leon MP, Renkonen-Sinisalo L, Sampson JR, Stormorken A, Tejpar S, Thomas HJ, Wijnen J. Guidelines for the clinical management of familial adenomatous polyposis (FAP). *Gut* 2008; **57**: 704-713 [PMID: 18194984 DOI: 10.1136/gut.2007.136127]
19 **Alderlieste YA**, Rauws EA, Mathus-Vliegen EM, Fockens P, Dekker E. Prospective enteroscopic evaluation of jejunal polyposis in patients with familial adenomatous polyposis and advanced duodenal polyposis. *Fam Cancer* 2013; **12**: 51-56 [PMID: 23054214 DOI: 10.1007/s10689-012-9571-1]
20 **Church J**. Familial adenomatous polyposis. *Surg Oncol Clin N Am* 2009; **18**: 585-598 [PMID: 19793567 DOI: 10.1016/j.soc.2009.07.002]
21 **Shenoy S**, Cassim R. Ileostomy adenocarcinoma associated with familial adenomatous polyposis (FAP): new problem in old disease. *Int J Colorectal Dis* 2009; **24**: 1475-1476 [PMID: 19488768 DOI: 10.1007/s00384-009-0739-6]
22 **Parc YR**, Olschwang S, Desaint B, Schmitt G, Parc RG, Tiret E. Familial adenomatous polyposis: prevalence of adenomas in the ileal pouch after restorative proctocolectomy. *Ann Surg* 2001; **233**: 360-364 [PMID: 11224623 DOI: 10.1097/00000658-200103000-00009]
23 **Hata K**, Watanabe T, Kawamura YJ, Ishigami H, Kanazawa T, Tada T, Zhao B, Koketsu S, Nagawa H. K-ras mutation and loss of heterozygosity at 17p with beta-catenin accumulation in intramucosal carcinoma of the ileostomy in familial adenomatous polyposis: a case report. *Dig Dis Sci* 2003; **48**: 2310-2314 [PMID: 14714618 DOI: 10.1023/B: DDAS.0000007868.52339.22]
24 **Quah HM**, Samad A, Maw A. Ileostomy carcinomas a review: the latent risk after colectomy for ulcerative colitis and familial adenomatous polyposis. *Colorectal Dis* 2005; **7**: 538-544 [PMID: 16232232 DOI: 10.1111/j.1463-1318.2005.00807.x]
25 **Hampel H**, Frankel WL, Martin E, Arnold M, Khanduja K, Kuebler P, Clendenning M, Sotamaa K, Prior T, Westman JA, Panescu J, Fix D, Lockman J, LaJeunesse J, Comeras I, de la Chapelle A. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. *J Clin Oncol* 2008; **26**: 5783-5788 [PMID: 18809606 DOI: 10.1200/JCO.2008.17.5950]
26 **ten Kate GL**, Kleibeuker JH, Nagengast FM, Craanen M, Cats A, Menko FH, Vasen HF. Is surveillance of the small bowel indicated for Lynch syndrome families? *Gut* 2007; **56**: 1198-1201 [PMID: 17409122 DOI: 10.1136/gut.2006.118299]
27 **Cheung DY**, Choi MG. Current advance in small bowel tumors. *Clin Endosc* 2011; **44**: 13-21 [PMID: 22741107 DOI: 10.5946/ce.2011.44.1.13]
28 **Aparicio T**, Zaanan A, Svrcek M, Laurent-Puig P, Carrere N, Manfredi S, Locher C, Afchain P. Small bowel adenocarcinoma: epidemiology, risk factors, diagnosis and treatment. *Dig Liver Dis* 2014; **46**: 97-104 [PMID: 23796552 DOI: 10.1016/j.dld.2013.04.013]
29 **Schulmann K**, Brasch FE, Kunstmann E, Engel C, Pagenstecher C, Vogelsang H, Krüger S, Vogel T, Knaebel HP, Rüschoff J, Hahn SA, Knebel-Doeberitz MV, Moeslein G, Meltzer SJ, Schackert HK, Tympner C, Mangold E, Schmiegel W. HNPCC-associated small bowel cancer: clinical and molecular characteristics. *Gastroenterology* 2005; **128**: 590-599 [PMID: 15765394 DOI: 10.1053/j.gastro.2004.12.051]
30 **Sargent DJ**, Marsoni S, Monges G, Thibodeau SN, Labianca R, Hamilton SR, French AJ, Kabat B, Foster NR, Torri V, Ribic C, Grothey A, Moore M, Zaniboni A, Seitz JF, Sinicrope F, Gallinger S. Defective mismatch repair as a predictive marker for lack of efficacy of fluorouracil-based adjuvant therapy in colon cancer. *J Clin Oncol* 2010; **28**: 3219-3226 [PMID: 20498393 DOI: 10.1200/JCO.2009.27.1825]
31 **Beggs AD**, Latchford AR, Vasen HF, Moslein G, Alonso A, Aretz S, Bertario L, Blanco I, Bülow S, Burn J, Capella G, Colas C, Friedl W, Møller P, Hes FJ, Järvinen H, Mecklin JP, Nagengast FM, Parc Y, Phillips RK, Hyer W, Ponz de Leon M, Renkonen-Sinisalo L, Sampson JR, Stormorken A, Tejpar S, Thomas HJ, Wijnen JT, Clark SK, Hodgson SV. Peutz-Jeghers syndrome: a systematic review and recommendations for management. *Gut* 2010; **59**: 975-986 [PMID: 20581245 DOI: 10.1136/gut.2009.198499]
32 **Karuman P**, Gozani O, Odze RD, Zhou XC, Zhu H, Shaw R, Brien TP, Bozzuto CD, Ooi D, Cantley LC, Yuan J. The Peutz-Jegher gene product LKB1 is a mediator of p53-dependent cell death. *Mol Cell* 2001; **7**: 1307-1319 [PMID: 11430832 DOI: 10.1016/S1097-2765(01)00258-1]
33 **Alessi DR**, Sakamoto K, Bayascas JR. LKB1-dependent signaling pathways. *Annu Rev Biochem* 2006; **75**: 137-163 [PMID: 16756488 DOI: 10.1146/annurev.biochem.75.103004.142702]
34 **Gruber SB**, Entius MM, Petersen GM, Laken SJ, Longo PA, Boyer R, Levin AM, Mujumdar UJ, Trent JM, Kinzler KW, Vogelstein B, Hamilton SR, Polymeropoulos MH, Offerhaus GJ, Giardiello FM. Pathogenesis of adenocarcinoma in Peutz-Jeghers syndrome. *Cancer Res* 1998; **58**: 5267-5270 [PMID: 9850045]
35 **Giardiello FM**, Trimbath JD. Peutz-Jeghers syndrome and management recommendations. *Clin Gastroenterol Hepatol* 2006; **4**: 408-415 [PMID: 16616343 DOI: 10.1016/j.cgh.2005.11.005]
36 **Giardiello FM**, Brensinger JD, Tersmette AC, Goodman SN, Petersen GM, Booker SV, Cruz-Correa M, Offerhaus JA. Very high risk of cancer in familial Peutz-Jeghers syndrome. *Gastroenterology* 2000; **119**: 1447-1453 [PMID: 11113065 DOI: 10.1053/gast.2000.20228]
37 **Hearle N**, Schumacher V, Menko FH, Olschwang S, Boardman LA, Gille JJ, Keller JJ, Westerman AM, Scott RJ, Lim W, Trimbath JD, Giardiello FM, Gruber SB, Offerhaus GJ, de Rooij FW, Wilson JH, Hansmann A, Möslein G, Royer-Pokora B, Vogel T, Phillips RK, Spigelman AD, Houlston RS. Frequency and spectrum of cancers in the Peutz-Jeghers syndrome. *Clin Cancer Res* 2006; **12**: 3209-3215 [PMID: 16707622 DOI: 10.1158/1078-0432.CCR-06-0083]
38 **Chow E**, Macrae F. A review of juvenile polyposis syndrome. *J Gastroenterol Hepatol* 2005; **20**: 1634-1640 [PMID: 16246179 DOI: 10.1111/j.1440-1746.2005.03865.x]
39 **Wu TT**, Rezai B, Rashid A, Luce MC, Cayouette MC, Kim C, Sani N, Mishra L, Moskaluk CA, Yardley JH, Hamilton SR. Genetic alterations and epithelial dysplasia in juvenile polyposis syndrome and sporadic juvenile polyps. *Am J Pathol* 1997; **150**: 939-947 [PMID: 9060832]
40 **Howe JR**, Sayed MG, Ahmed AF, Ringold J, Larsen-Haidle J, Merg A, Mitros FA, Vaccaro CA, Petersen GM, Giardiello FM, Tinley ST, Aaltonen LA, Lynch HT. The prevalence of MADH4 and BMPR1A mutations in juvenile polyposis and absence of BMPR2, BMPR1B, and ACVR1 mutations. *J Med Genet* 2004; **41**: 484-491 [PMID: 15235019 DOI: 10.1136/jmg.2004.018598]
41 **Sayed MG**, Ahmed AF, Ringold JR, Anderson ME, Bair JL, Mitros FA, Lynch HT, Tinley ST, Petersen GM, Giardiello FM, Vogelstein B, Howe JR. Germline SMAD4 or BMPR1A mutations and phenotype of juvenile polyposis. *Ann Surg Oncol* 2002; **9**: 901-906 [PMID: 12417513 DOI: 10.1007/BF02557528]

42 **Lynch HT**, Drescher K, Knezetic J, Lanspa S. Genetics, biomarkers, hereditary cancer syndrome diagnosis, heterogeneity and treatment: a review. *Curr Treat Options Oncol* 2014; **15**: 429-442 [PMID: 24827900 DOI: 10.1007/s11864-014-0293-5]

43 **Loftus EV**, Schoenfeld P, Sandborn WJ. The epidemiology and natural history of Crohn's disease in population-based patient cohorts from North America: a systematic review. *Aliment Pharmacol Ther* 2002; **16**: 51-60 [PMID: 11856078 DOI: 10.1046/j.1365-2036.2002.01140.x]

44 **Canavan C**, Abrams KR, Mayberry J. Meta-analysis: colorectal and small bowel cancer risk in patients with Crohn's disease. *Aliment Pharmacol Ther* 2006; **23**: 1097-1104 [PMID: 16611269 DOI: 10.1111/j.1365-2036.2006.02854.x]

45 **Feldstein RC**, Sood S, Katz S. Small bowel adenocarcinoma in Crohn's disease. *Inflamm Bowel Dis* 2008; **14**: 1154-1157 [PMID: 18275076 DOI: 10.1002/ibd.20393]

46 **von Roon AC**, Reese G, Teare J, Constantinides V, Darzi AW, Tekkis PP. The risk of cancer in patients with Crohn's disease. *Dis Colon Rectum* 2007; **50**: 839-855 [PMID: 17308939 DOI: 10.1007/s10350-006-0848-z]

47 **Shaukat A**, Virnig DJ, Howard D, Sitaraman SV, Liff JM, Lederle FA. Crohn's disease and small bowel adenocarcinoma: a population-based case-control study. *Cancer Epidemiol Biomarkers Prev* 2011; **20**: 1120-1123 [PMID: 21467236 DOI: 10.1158/1055-9965.EPI-10-1281]

48 **Friedman S**. Cancer in Crohn's disease. *Gastroenterol Clin North Am* 2006; **35**: 621-639 [PMID: 16952744 DOI: 10.1016/j.gtc.2006.07.008]

49 **Dossett LA**, White LM, Welch DC, Herline AJ, Muldoon RL, Schwartz DA, Wise PE. Small bowel adenocarcinoma complicating Crohn's disease: case series and review of the literature. *Am Surg* 2007; **73**: 1181-1187 [PMID: 18092659]

50 **Braun J**, Wei B. Body traffic: ecology, genetics, and immunity in inflammatory bowel disease. *Annu Rev Pathol* 2007; **2**: 401-429 [PMID: 18039105 DOI: 10.1146/annurev.pathol.1.110304.100128]

51 **Brant SR**. Update on the heritability of inflammatory bowel disease: the importance of twin studies. *Inflamm Bowel Dis* 2011; **17**: 1-5 [PMID: 20629102 DOI: 10.1002/ibd.21385]

52 **Liu JZ**, Anderson CA. Genetic studies of Crohn's disease: past, present and future. *Best Pract Res Clin Gastroenterol* 2014; **28**: 373-386 [PMID: 24913378 DOI: 10.1016/j.bpg.2014.04.009]

53 **Wehkamp J**, Stange EF. Paneth cells and the innate immune response. *Curr Opin Gastroenterol* 2006; **22**: 644-650 [PMID: 17053443 DOI: 10.1097/01.mog.0000245541.95408.86]

54 **Rashid A**, Hamilton SR. Genetic alterations in sporadic and Crohn's-associated adenocarcinomas of the small intestine. *Gastroenterology* 1997; **113**: 127-135 [PMID: 9207270 DOI: 10.1016/S0016-5085(97)70087-8]

55 **Svrcek M**, Piton G, Cosnes J, Beaugerie L, Vermeire S, Geboes K, Lemoine A, Cervera P, El-Murr N, Dumont S, Scriva A, Lascols O, Ardizzone S, Fociani P, Savoye G, Le Pessot F, Novacek G, Wrba F, Colombel JF, Leteurtre E, Bouhnik Y, Cazals-Hatem D, Cadiot G, Diebold MD, Rahier JF, Delos M, Fléjou JF, Carbonnel F. Small bowel adenocarcinomas complicating Crohn's disease are associated with dysplasia: a pathological and molecular study. *Inflamm Bowel Dis* 2014; **20**: 1584-1592 [PMID: 25029614 DOI: 10.1097/MIB.0000000000000112]

56 **Galandiuk S**, Rodriguez-Justo M, Jeffery R, Nicholson AM, Cheng Y, Oukrif D, Elia G, Leedham SJ, McDonald SA, Wright NA, Graham TA. Field cancerization in the intestinal epithelium of patients with Crohn's ileocolitis. *Gastroenterology* 2012; **142**: 855-864.e8 [PMID: 22178590 DOI: 10.1053/j.gastro.2011.12.004]

57 **Widmar M**, Greenstein AJ, Sachar DB, Harpaz N, Bauer JJ, Greenstein AJ. Small bowel adenocarcinoma in Crohn's disease. *J Gastrointest Surg* 2011; **15**: 797-802 [PMID: 21336499 DOI: 10.1007/s11605-011-1441-x]

58 **Murray JA**. The widening spectrum of celiac disease. *Am J Clin Nutr* 1999; **69**: 354-365 [PMID: 10075317]

59 **Green PH**, Cellier C. Celiac disease. *N Engl J Med* 2007; **357**: 1731-1743 [PMID: 17960014 DOI: 10.1056/NEJMra071600]

60 **Sollid LM**, Jabri B. Triggers and drivers of autoimmunity: lessons from coeliac disease. *Nat Rev Immunol* 2013; **13**: 294-302 [PMID: 23493116 DOI: 10.1038/nri3407]

61 **Green PH**, Fleischauer AT, Bhagat G, Goyal R, Jabri B, Neugut AI. Risk of malignancy in patients with celiac disease. *Am J Med* 2003; **115**: 191-195 [PMID: 12935825 DOI: 10.1016/S0002-9343(03)00302-4]

62 **Green PHR SN**, Panagi SG, Goldstein SL, Mcmahon DJ, Absan H, Neugut AI. Characteristics of adult celiac disease in the USA: results of a national survey. *Am J Gastroenterol* 2001; **96**: 126-131 [PMID: 11197241 DOI: 10.1111/j.1572-0241.2001.03462.x]

63 **Peters U**, Askling J, Gridley G, Ekbom A, Linet M. Causes of death in patients with celiac disease in a population-based Swedish cohort. *Arch Intern Med* 2003; **163**: 1566-1572 [PMID: 12860579 DOI: 10.1001/archinte.163.13.1566]

64 **Rampertab SD**, Forde KA, Green PH. Small bowel neoplasia in coeliac disease. *Gut* 2003; **52**: 1211-1214 [PMID: 12865284 DOI: 10.1136/gut.52.8.1211]

65 **Bergmann F**, Singh S, Michel S, Kahlert C, Schirmacher P, Helmke B, Von Knebel Doeberitz M, Kloor M, Bläker H. Small bowel adenocarcinomas in celiac disease follow the CIM-MSI pathway. *Oncol Rep* 2010; **24**: 1535-1539 [PMID: 21042749]

66 **Diosdado B**, Buffart TE, Watkins R, Carvalho B, Ylstra B, Tijssen M, Bolijn AS, Lewis F, Maude K, Verbeke C, Nagtegaal ID, Grabsch H, Mulder CJ, Quirke P, Howdle P, Meijer GA. High-resolution array comparative genomic hybridization in sporadic and celiac disease-related small bowel adenocarcinomas. *Clin Cancer Res* 2010; **16**: 1391-1401 [PMID: 20179237 DOI: 10.1158/1078-0432.CCR-09-1773]

67 **Potter DD**, Murray JA, Donohue JH, Burgart LJ, Nagorney DM, van Heerden JA, Plevak MF, Zinsmeister AR, Thibodeau SN. The role of defective mismatch repair in small bowel adenocarcinoma in celiac disease. *Cancer Res* 2004; **64**: 7073-7077 [PMID: 15466202 DOI: 10.1158/0008-5472.CAN-04-1096]

68 **Talamonti MS**, Goetz LH, Rao S, Joehl RJ. Primary cancers of the small bowel: analysis of prognostic factors and results of surgical management. *Arch Surg* 2002; **137**: 564-570; discussion 570-571 [PMID: 11982470 DOI: 10.1001/archsurg.137.5.564]

69 **Fry LC**, Vormbrock K, Olano C, Mönkemüller K. Small-bowel endoscopy. *Endoscopy* 2011; **43**: 978-984 [PMID: 22057762 DOI: 10.1055/s-0031-1291422]

70 **Yano T**, Yamamoto H. Current state of double balloon endoscopy: the latest approach to small intestinal diseases. *J Gastroenterol Hepatol* 2009; **24**: 185-192 [PMID: 19215331 DOI: 10.1111/j.1440-1746.2008.05773.x]

71 **Riccioni ME**, Urgesi R, Cianci R, Spada C, Nista EC, Costamagna G. Single-balloon push-and-pull enteroscopy system: does it work? A single-center, 3-year experience. *Surg Endosc* 2011; **25**: 3050-3056 [PMID: 21487872 DOI: 10.1007/s00464-011-1669-2]

72 **Mönkemüller K**, Fry LC, Ebert M, Bellutti M, Venerito M, Knippig C, Rickes S, Muschke P, Röcken C, Malfertheiner P. Feasibility of double-balloon enteroscopy-assisted chromoendoscopy of the small bowel in patients with familial adenomatous polyposis. *Endoscopy* 2007; **39**: 52-57 [PMID: 17252461 DOI: 10.1055/s-2006-945116]

73 **Eliakim R**. Video capsule endoscopy of the small bowel. *Curr Opin Gastroenterol* 2013; **29**: 133-139 [PMID: 23221650 DOI: 10.1097/MOG.0b013e32835bdc03]

74 **Baichi MM**, Arifuddin RM, Mantry PS. Metachronous small bowel adenocarcinomas detected by capsule endoscopy in a patient with hereditary nonpolyposis colorectal cancer. *Dig Dis Sci* 2007; **52**: 1134-1136 [PMID: 17342393 DOI: 10.1007/s10620-006-9395-7]

75 **Saurin JC**, Pilleul F, Soussan EB, Manière T, D'Halluin PN, Gaudric M, Cellier C, Heresbach D, Gaudin JL. Small-bowel capsule endoscopy diagnoses early and advanced neoplasms in asymptomatic patients with Lynch syndrome. *Endoscopy* 2010; **42**: 1057-1062 [PMID: 20821360 DOI: 10.1055/s-0030-1255742]

76 **Caspari R**, von Falkenhausen M, Krautmacher C, Schild H, Heller J, Sauerbruch T. Comparison of capsule endoscopy and magnetic resonance imaging for the detection of polyps of the small intestine in patients with familial adenomatous polyposis or with Peutz-

Jeghers' syndrome. *Endoscopy* 2004; **36**: 1054-1059 [PMID: 15578294 DOI: 10.1055/s-2004-826041]

77 **Tescher P**, Macrae FA, Speer T, Stella D, Gibson R, Tye-Din JA, Srivatsa G, Jones IT, Marion K. Surveillance of FAP: a prospective blinded comparison of capsule endoscopy and other GI imaging to detect small bowel polyps. *Hered Cancer Clin Pract* 2010; **8**: 3 [PMID: 20361877 DOI: 10.1186/1897-4287-8-3]

78 **Ross A**, Mehdizadeh S, Tokar J, Leighton JA, Kamal A, Chen A, Schembre D, Chen G, Binmoeller K, Kozarek R, Waxman I, Dye C, Gerson L, Harrison ME, Haluszka O, Lo S, Semrad C. Double balloon enteroscopy detects small bowel mass lesions missed by capsule endoscopy. *Dig Dis Sci* 2008; **53**: 2140-2143 [PMID: 18270840 DOI: 10.1007/s10620-007-0110-0]

79 **Masselli G**, Gualdi G. CT and MR enterography in evaluating small bowel diseases: when to use which modality? *Abdom Imaging* 2013; **38**: 249-259 [PMID: 23011551 DOI: 10.1007/s00261-012-9961-8]

80 **Maglinte DD**, Sandrasegaran K, Lappas JC, Chiorean M. CT Enteroclysis. *Radiology* 2007; **245**: 661-671 [PMID: 18024448 DOI: 10.1148/radiol.2453060798]

81 **Masselli G**, Polettini E, Casciani E, Bertini L, Vecchioli A, Gualdi G. Small-bowel neoplasms: prospective evaluation of MR enteroclysis. *Radiology* 2009; **251**: 743-750 [PMID: 19304922 DOI: 10.1148/radiol.2513081819]

82 **Van Weyenberg SJ**, Bouman K, Jacobs MA, Halloran BP, Van der Peet DL, Mulder CJ, Van Kuijk C, Van Waesberghe JH. Comparison of MR enteroclysis with video capsule endoscopy in the investigation of small-intestinal disease. *Abdom Imaging* 2013; **38**: 42-51 [PMID: 22527155 DOI: 10.1007/s00261-012-9892-4]

P- Reviewer: Fujimori S, Fujino Y, Sipahi AM **S- Editor**: Gong ZM
L- Editor: A **E- Editor**: Lu YJ

World Journal of Gastrointestinal Oncology

Submit a Manuscript: http://www.wjgnet.com/esps/
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
DOI: 10.4251/wjgo.v8.i6.520

World J Gastrointest Oncol 2016 June 15; 8(6): 520-525
ISSN 1948-5204 (online)

ORIGINAL ARTICLE

Case Control Study

Genetic polymorphisms of *interleukin 1β* gene and sporadic pancreatic neuroendocrine tumors susceptibility

Dimitrios Karakaxas, Anna Sioziou, Gerasimos Aravantinos, Ahmet Coker, Ioannis S Papanikolaou, Theodoros Liakakos, Christos Dervenis, Maria Gazouli

Dimitrios Karakaxas, Christos Dervenis, Department of General Surgery, Agia Olga Hospital, 14233 Athens, Greece

Anna Sioziou, Maria Gazouli, Department of Basic Medical Sciences, Laboratory of Biology, Medical School, National and Kapodistrian University of Athens, 11527 Athens, Greece

Gerasimos Aravantinos, Second Department of Medical Oncology, Agioi Anargiroi Cancer Hospital, Kifisia, 14564 Athens, Greece

Ahmet Coker, Department of General Surgery, Ege University School of Medicine, 35040 Izmir, Turkey

Ioannis S Papanikolaou, Hepatogastroenterology Unit, 2nd Department of Internal Medicine and Research Unit, Attikon University General Hospital, Medical School, National and Kapodistrian University of Athens, 12462 Athens, Greece

Theodoros Liakakos, 1st Department of Surgery, Laiko Athens General Hospital, Medical School, National and Kapodistrian University of Athens, 11527 Athens, Greece

Author contributions: Karakaxas D, Aravantinos G, Coker A, Papanikolaou IS, Liakakos T and Dervenis C collected the samples and the patients data; Karakaxas D, Sioziou A and Gazouli M performed the experiments; Liakakos T, Dervenis C and Gazouli M designed the study, wrote and corrected the manuscript.

Supported by Hellenic Society of Medical Oncology, No. 5839/08-04-2015.

Institutional review board statement: The study was approved by the ethics committee of Attikon Hospital.

Informed consent statement: All patients gave informed consent.

Conflict-of-interest statement: All authors do not have any conflict-of interest to declare.

Data sharing statement: Technical appendix, statistical code, and dataset available from the corresponding author at mgazouli@med.uoa.gr.

Correspondence to: Maria Gazouli, PhD, Assistant Professor of Molecular Biology, Department of Basic Medical Sciences, Laboratory of Biology, Medical School, National and Kapodistrian University of Athens, Michalakopoulou 176, 11527 Athens, Greece. mgazouli@med.uoa.gr
Telephone: +30-210-7462231
Fax: +30-210-7462231

Received: February 11, 2016
Peer-review started: February 14, 2016
First decision: March 1, 2016
Revised: March 1, 2016
Accepted: March 17, 2016
Article in press: March 17, 2016
Published online: June 15, 2016

Abstract

AIM: To evaluate the association between the interleukin 1β (IL-1β) polymorphisms and the pancreatic neuroendocrine tumor (pNET) development.

METHODS: A case-control study was conducted analyzing IL-1β polymorphisms using germline DNA collected in a population-based case-control study of pancreatic cancer (51 pNET cases, 85 pancreatic ductal adenocarcinoma cases, 19 intraductal papillary mucinous neoplasm and 98 healthy controls).

RESULTS: The distribution of genotypes for the -511

C/T polymorphism in the pNET patient groups showed significant difference compared to the control group. It is known that the carriers of the IL-1β -511T allele have increased concentrations of IL-1β. The -511 CT and TT high-expression genotypes were over-represented in pNET patients.

CONCLUSION: The findings of this study suggested a possible role of IL-1β -511 C/T genotypes in the pathogenesis of pNETs since the presence of the IL-1β -511 CT and TT genotypes and the T allele was associated with an increased risk of pNET only.

Key words: Interleukin 1β; Neuroendocrine tumors; Pancreas

Core tip: Pancreatic neuroendocrine tumors (pNETs) are a heterogeneous group of rare neoplasms derived from pancreatic endocrine cells and have significantly different tumor biology and present better prognosis compared with tumors of the exocrine pancreas, like pancreatic adenocarcinomas. It is widely accepted that chronic inflammation contributes to pathogenesis of many pancreatic diseases, including pancreatic carcinogenesis. Interleukin 1β (IL-1β) is a highly active pro-inflammatory cytokine with multiple biological effects, such as directing cancer cells to either neuroendocrine differentiation or to development of adenocarcinoma. The purpose of the study was to evaluate the association between the IL-1β polymorphisms and the pNET development.

Karakaxas D, Sioziou A, Aravantinos G, Coker A, Papanikolaou IS, Liakakos T, Dervenis C, Gazouli M. Genetic polymorphisms of *interleukin 1β* gene and sporadic pancreatic neuroendocrine tumors susceptibility. *World J Gastrointest Oncol* 2016; 8(6): 520-525 Available from: URL: http://www.wjgnet.com/1948-5204/full/v8/i6/520.htm DOI: http://dx.doi.org/10.4251/wjgo.v8.i6.520

INTRODUCTION

Pancreatic neuroendocrine tumors (pNETs) are a heterogeneous group of rare neoplasms derived from pancreatic endocrine cells[1-5]. The annual incidence of pNETs is estimated to be approximately 3.65 per 100000 individuals in the United States and occur sporadically or may be associated with genetic syndromes such as multiple endocrine neoplasia type 1 (MEN-1), von Hippel-Lindau syndrome (VHL), von Recklinghausen disease (neurofibromatosis NF-1), and tuberous sclerosis complex (TSC)[6-8].

PNETs are mainly considered functionally inactive tumors, but when related with hormone or peptide overproduction, such as insulin, gastrin, glucagon, vasoactive intestinal polypeptide (VIP) and somatostatin they are responsible for many characteristic clinical syndromes, with insulinoma being the most common PNETs are usually asymptomatic[9,10], have significantly different tumor biology, and present better prognosis compared with tumors of the exocrine pancreas, like pancreatic adenocarcinomas (PDACs)[11].

The molecular basis of pNETs pathogenesis is poorly characterized but several recent reports have been conducted in order to clarify their etiology[12].

It is widely accepted that chronic inflammation contributes to pathogenesis of many pancreatic diseases, including pancreatic carcinogenesis[13,14]. However, the exact mechanism by which chronic inflammation promotes carcinogenesis is still unknown. During carcinogenesis the host-mediated anti-tumor activity is suppressed, whereas pro-inflammatory events support tumor growth, angiogenesis, invasion and metastasis[15]. The inflammatory response is mediated by cytokines, which are glycoproteins or soluble proteins and their role in cancer immunity and carcinogenesis has been well established[16-18].

Neuroendocrine tumors express various cytokines and growth-factors. Several pro-inflammatory cytokines have been found in pNETs tissue suggesting their involvement in pNET development[19-21]. Additionally, numerous studies suggested that gastroenteropancreatic-NETs occur more frequently in the environment of chronic inflammation[22-24]. Thus, cytokines such as interleukin 1 (IL-1) poses an important role in neuroendocrine tumors since direct cancer cells to either neuroendocrine differentiation or to development of adenocarcinoma, while exogenously added IL-1 results in a decrease of chromogranin A (CgA) and simultaneous increase in carcinoembryonic antigen (CEA) secretion[25].

IL-1β is a highly active pro-inflammatory cytokine with multiple biological effects[26]. IL-1β protein levels are related to the intensity of the inflammatory response, and regarding to pancreas, IL-1β is implicated in cancer progression, especially tumor invasiveness, metastasis and angiogenesis[27,28].

The *IL-1β* gene is located in the IL1 cluster on chromosome 2q and several single nucleotide polymorphisms (SNPs) of this gene influence the regulation of its expression and function have been studied[29-32]. There are two SNPs in the proximal promoter region of the *IL-1β* gene, -511 C/T and +3954 T/C, which both have been correlated with gastrointestinal cancers, such as gastric, hepatocellular cancer (HCC) and pancreatic cancer[33-36]. Recently, Cigrovski Berković *et al*[37] reported that the *IL-1β* -511 SNP contributes to the pNET susceptibility.

We conducted a case-control study to analyze *IL-1β* polymorphisms as risk factors for pNETs using germline DNA collected in a population-based case-control study of pancreatic cancer [51 pNET cases, 85 PDAC cases, 19 intraductal papillary mucinous neoplasm (IPMN) and 98 healthy controls] conducted in the Athens, Greece and Izmir, Turkey areas.

MATERIALS AND METHODS

Patients

The case-control study included 51 pNET cases (22

Table 1 Characteristics of the patients and controls *n* (%)

Characteristic	PDAC	pNET	IPMN	Controls
Total number	85	51	19	98
Mean age (yr)	59.12	56.31	57.91	58.9
Gender				
Male	51 (60)	20 (39.2)	11 (57.9)	74 (75.5)
Female	34 (40)	31 (60.8)	8 (42.1)	24 (24.5)
Tumor stage				
Ⅰ	13 (15.3)			
Ⅱ	36 (42.4)			
Ⅲ	33 (38.8)			
Ⅳ	3 (3.5)			
G stage				
G1		35 (68.6)		
G2		14 (27.5)		
G3		2 (3.9)		
Tumor location				
Head	64 (75.3)	19 (37.3)	7 (36.8)	
Body and tail	21 (24.7)	32 (62.7)	12 (63.2)	
Differentiation status				
Well	10 (11.8)			
Moderate	39 (45.9)			
Poor	36 (42.3)			

pNET: Pancreatic neuroendocrine tumor; PDAC: Pancreatic adenocarcinoma; IPMN: Intraductal papillary mucinous neoplasm.

nonfunctional and 29 functional), 85 PDAC cases, 19 IPMN and 98 healthy controls (Table 1). None of the cases had a history of chronic pancreatitis. For subsequent analysis, we excluded cases and controls with known genetic syndromes (*e.g.*, MEN1, MEN2, VHL or TSC). Controls were healthy blood donors with no evidence of inflammation. The diagnosis in all cases was established by standard procedures and confirmed histopathologically either from operatively resected tumors or biopsy tissues, in cases of unresectable tumors. Before commencement of the study, the Ethical committee at the participating centers approved the recruitment protocols. All participants were informed regarding the study, and their written consent was provided.

Genotyping

Genomic DNAs were isolated from peripheral ethylenediaminetetraacetic acid-treated blood of patients and healthy controls using the NucleoSpin Blood Kit (Macherey-Nagel, Germany). The *IL-1β* -511 C/T (rs16944) polymorphism was detected by PCR-RFLP using the set of primers: 5′-TGGCATTGATCTGGTTCATC-3′ and 5′-GTTTAGGAATCTTCCCACTT-3′. The 35 cycles of PCR were carried out at 94 ℃ for 5 min, 94 ℃ for 1 min, 58 ℃ for 40 s and 72 ℃ for 1 min and the final cycle of 72 ℃ for 5 min. Amplified PCR products were digested with *Ava*I for 2 h at 37 ℃. The fragments of 189- and 116-bp revealed homozygosity for the C allele, and 305-bp indicated homozygosity for the T allele. The +3954 C/T (rs 1143634) polymorphism was detected with the 5′-TCAGGTGTCCTCGAAGAAATCAAA-3′ and 5′-GGTTTTTTGCTGTGAGTCCC-3′ set of primers and the cycling parameters for that was 94 ℃ for 5 min, 94 ℃ for 45 s, 56 ℃ for 45 s and 72 ℃ for 45 s and the final cycle of 72 ℃ for 5 min. After 35 cycles the PCR product were digested for 2 h at 65 ℃ with *Taq*I. The fragments of 97- and 85-bp revealed homozygosity for the C allele and on the other hand 182-bp fragments showed homozygosity for the T allele.

Statistical analysis

Genotype frequencies were compared with the χ^2 with Yate's correction using S-Plus (v. 6.2, Insightful, Seattle, WA). Odds ratios (ORs) and 95%CIs were obtained with GraphPad (v. 3.00, GraphPad Software, San Diego, CA). The *P* values are all two-sided, and *P* values of < 0.05 were considered to be significant. Hardy-Weinberg equilibrium was verified by calculating the expected frequencies and numbers and was tested separately in patients and in controls using the goodness-of-fit χ^2 test. Haplotype analysis was performed using the http://bioinfo.iconcologia.net/SNPstats software.

RESULTS

The clinicopathological characteristics of the studied population are summarized in Table 1. The genotype frequencies of the *IL-1β* -511 C/T and +3954 C/T polymorphisms between PDAC, pNET, IPMN patients and controls are given in Table 2. All genotype distributions were in Hardy-Weinberg equilibrium. The distribution of genotypes for the -511 C/T polymorphism in the pNET patient groups only showed significant difference compared to the control group. It is known that the carriers of the *IL-1β* -511T allele have increased concentrations of IL-1β[38]. The -511 CT and TT high-expression genotypes were over-represented in pNET patients (Table 2). However, the presence of the +3954T

Table 2 Genotype and allele frequencies of the interleukin 1β -511 C/T and +3954 C/T polymorphisms in pancreatic adenocarcinoma, pancreatic neuroendocrine tumor and intraductal papillary mucinous neoplasm patients and controls

	Controls (n = 98)	PDAC (n = 85)	P; OR (95%CI)	pNET (n = 51)	P; OR (95%CI)	IPMN (n = 19)	P; OR (95%CI)
-511 C/T							
CC	44	35	1	13	1	6	1
CT	47	44	0.64; 1.18 (0.64-2.16)	31	0.04; 2.23 (1.04-4.81)	10	0.59; 1.56 (0.52-4.65)
TT	7	6	1; 1.08 (0.33-3.49)	7	0.04; 3.95 (1.13-13.84)	3	0.16; 3.14 (0.64-15.56)
CT + TT	54	50	0.37; 1.36 (0.75-2.44)	38	0.02; 2.38 (1.13-5.02)	13	0.32; 1.76 (0.62-5.03)
C allele	135	114	1	57	1	22	1
T allele	61	56	0.74; 1.09 (0.7-1.69)	45	0.03; 1.75 (1.07-2.86)	16	0.19; 1.61 (0.79-3.28)
+3954 C/T							
CC	45	50	1	33	1	8	1
CT	44	28	0.08; 0.57 (0.31-1.07)	16	0.07; 0.49 (0.24-1.03)	10	0.79; 1.28 (0.46-3.54)
TT	9	7	0.59; 0.7 (0.24-2.04)	2	0.19; 0.3 (0.06-1.49)	1	1; 0.62 (0.07-5.64)
CT + TT	53	35	0.1; 0.59 (0.33-1.07)	18	0.04; 0.46 (0.23-0.93)	11	0.81; 1.18 (0.43-3.15)
C allele	134	128	1	82	1	26	1
T allele	62	42	0.16; 0.71 (0.45-1.12)	20	0.03; 0.53 (0.29-0.94)	12	0.85; 1.07 (0.51-2.25)

pNET: Pancreatic neuroendocrine tumor; PDAC: Pancreatic adenocarcinoma; IPMN: Intraductal papillary mucinous neoplasm.

allele seems to have a protective role in the pNET development since it is found to be over-represented in healthy controls. The haplotype analysis did not reveal any significant association. No significant association was found between genotypes, haplotypes, and clinicopathological data of the patients.

DISCUSSION

PNETs are a rare, heterogeneous group of neuroendocrine tumors. They usually have a better prognosis than the PDACs. The cause of these tumors is not fully understood, but differential expression of proinflammatory cytokines were found in pNET tissues[19-21]. The findings of this study suggested a possible role of *IL-1β* -511 C/T genotypes in the pathogenesis of pNETs since the presence of the *IL-1β* -511 CT and TT genotypes and the T allele was associated with an increased risk of pNET only. None significant correlation was found with PDAC and IPMN cases. Although Barber *et al*[36], reported that the +3954 C/T polymorphism of the *IL-1β* gene predisposes to pancreatic cancer; our findings did not reveal any significant association. Additionally, they are partly in agreement with the findings of Cigrovski Berkovic *et al*[37], which suggest that there is an association between the *IL-1β* -511 C/T genotype and the susceptibility to pNET, especially functional pNETs. In our study we did not find any haplotype combination to be statistically associated with the susceptibility to pNETs, neither PDAC nor IMPN cases, but we observed that the +3954T allele is over-represented among healthy controls compared to pNET cases suggesting that this allele might have a protective role in pNET development.

Carcinogenesis in the gastrointestinal tract and pancreas is often associated with chronic inflammation[39-42]. It is known that the carriers of the -511T allele associated with high IL-1β serum levels[38], and in different type of cancers IL-1β levels correlate with inflammation, worse prognosis and carcinoembrional antigen (CEA) levels, a well-known biomarker of tumor exocrine differentiation[25,43].

Our previous results suggested that *TNF-α* -1031 polymorphism is associated with the development of pNET and IPMN[41], and several studies supported that pro-inflammatory cytokines were detected in pNET tissues signifying their etiological involvement[19,44]. Taken these into consideration future studies in larger populations are needed to elucidate the role of cytokines and inflammatory pathway in the sporadic pNET development.

COMMENTS

Background

Carcinogenesis in the gastrointestinal tract and pancreas is often associated with chronic inflammation. The study provides evidence of a role of interleukin 1β (*IL-1β*) -511 C/T genotypes in the pathogenesis of pancreatic neuroendocrine tumors (pNETs).

Research frontiers

PNETs are a rare, heterogeneous group of neuroendocrine tumors. They usually have a better prognosis than the pancreatic adenocarcinomas. The cause of these tumors is not fully understood, but differential expression of proinflammatory cytokines were found in pNET tissues. Identifying genetic factors associated basically with pNET incidence may help in the primary prevention of pNET across the globe.

Innovations and breakthroughs

The study suggested a possible role of *IL-1β* -511 C/T genotypes in the pathogenesis of pNETs since the presence of the *IL-1β* -511 CT and TT genotypes and the T allele was associated with an increased risk of pNET only.

Applications

The study contributes to elucidate the role of cytokines and inflammatory pathway in the sporadic pNET development.

Terminology

PNETs: Pancreatic neuroendocrine tumors; PDACs: Pancreatic adenocarcinomas; IPMN: Intraductal papillary mucinous neoplasm.

Peer-review

This is an interesting study that looks at IL-1β as a potential inflammatory

cytokine stimulus for tumour formation in pNETs. While chronic inflammation is known to contribute to carcinogenesis, in the pancreas, this is peculiar to PDAC where association with chronic pancreatitis is not uncommon.

REFERENCES

1 **Vortmeyer AO**, Huang S, Lubensky I, Zhuang Z. Non-islet origin of pancreatic islet cell tumors. *J Clin Endocrinol Metab* 2004; **89**: 1934-1938 [PMID: 15070966 DOI: 10.1210/jc.2003.031575]

2 **Yalcin S**, Oyan B, Bayraktar Y. Current medical treatment of pancreatic neuroendocrine tumors. *Hepatogastroenterology* 2007; **54**: 278-284 [PMID: 17419276 DOI: 10.3390/cancers2031419]

3 **Ehehalt F**, Saeger HD, Schmidt CM, Grützmann R. Neuroendocrine tumors of the pancreas. *Oncologist* 2009; **14**: 456-467 [PMID: 19411317 DOI: 10.1634/theoncologist.2008-0259]

4 **Krampitz GW**, Norton JA. Pancreatic neuroendocrine tumors. *Curr Probl Surg* 2013; **50**: 509-545 [PMID: 24206780 DOI: 10.1067/j.cpsurg.2013.08.001]

5 **Rindi G**, Wiedenmann B. Neuroendocrine neoplasms of the gut and pancreas: new insights. *Nat Rev Endocrinol* 2012; **8**: 54-64 [PMID: 21808296 DOI: 10.1038/nrendo.2011.120]

6 **Lawrence B**, Gustafsson BI, Chan A, Svejda B, Kidd M, Modlin IM. The epidemiology of gastroenteropancreatic neuroendocrine tumors. *Endocrinol Metab Clin North Am* 2011; **40**: 1-18, vii [PMID: 21349409 DOI: 10.1016/j.ecl.2010.12.005]

7 **Zikusoka MN**, Kidd M, Eick G, Latich I, Modlin IM. The molecular genetics of gastroenteropancreatic neuroendocrine tumors. *Cancer* 2005; **104**: 2292-2309 [PMID: 16258976 DOI: 10.1002/cncr.21451]

8 **Klöppel G**, Perren A, Heitz PU. The gastroenteropancreatic neuroendocrine cell system and its tumors: the WHO classification. *Ann N Y Acad Sci* 2004; **1014**: 13-27 [PMID: 15153416 DOI: 10.1196/annals.1294.002]

9 **Yao JC**, Eisner MP, Leary C, Dagohoy C, Phan A, Rashid A, Hassan M, Evans DB. Population-based study of islet cell carcinoma. *Ann Surg Oncol* 2007; **14**: 3492-3500 [PMID: 17896148 DOI: 10.1245/s10434-007-9566-6]

10 **Metz DC**, Jensen RT. Gastrointestinal neuroendocrine tumors: pancreatic endocrine tumors. *Gastroenterology* 2008; **135**: 1469-1492 [PMID: 18703061 DOI: 10.1053/j.gastro.2008.05.047]

11 **Fesinmeyer MD**, Austin MA, Li CI, De Roos AJ, Bowen DJ. Differences in survival by histologic type of pancreatic cancer. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 1766-1773 [PMID: 16030115 DOI: 10.1158/1055-9965.EPI-05-0120]

12 **Grötzinger C**. Tumour biology of gastroenteropancreatic neuroendocrine tumours. *Neuroendocrinology* 2004; **80** Suppl 1: 8-11 [PMID: 15477708 DOI: 10.1159/000080732]

13 **Farrow B**, Evers BM. Inflammation and the development of pancreatic cancer. *Surg Oncol* 2002; **10**: 153-169 [PMID: 12020670 DOI: 10.1016/S0960-7404(02)00015-4]

14 **Algül H**, Treiber M, Lesina M, Schmid RM. Mechanisms of disease: chronic inflammation and cancer in the pancreas--a potential role for pancreatic stellate cells? *Nat Clin Pract Gastroenterol Hepatol* 2007; **4**: 454-462 [PMID: 17667994 DOI: 10.1038/ncpgasthep0881]

15 **de Visser KE**, Eichten A, Coussens LM. Paradoxical roles of the immune system during cancer development. *Nat Rev Cancer* 2006; **6**: 24-37 [PMID: 16397525 DOI: 10.1038/nrc1782]

16 **Smyth MJ**, Cretney E, Kershaw MH, Hayakawa Y. Cytokines in cancer immunity and immunotherapy. *Immunol Rev* 2004; **202**: 275-293 [PMID: 15546400 DOI: 10.1111/j.0105-2896.2004.00199.x]

17 **Seruga B**, Zhang H, Bernstein LJ, Tannock IF. Cytokines and their relationship to the symptoms and outcome of cancer. *Nat Rev Cancer* 2008; **8**: 887-899 [PMID: 18846100 DOI: 10.1038/nrc2507]

18 **Błogowski W**, Deskur A, Budkowska M, Sałata D, Madej-Michniewicz A, Dąbkowski K, Dołęgowska B, Starzyńska T. Selected cytokines in patients with pancreatic cancer: a preliminary report. *PLoS One* 2014; **9**: e97613 [PMID: 24849506 DOI: 10.1371/journal.pone.0097613]

19 **Massironi S**, Sciola V, Peracchi M, Ciafardini C, Spampatti MP, Conte D. Neuroendocrine tumors of the gastro-entero-pancreatic system. *World J Gastroenterol* 2008; **14**: 5377-5384 [PMID: 18803349 DOI: 10.3748/wjg.14.5377]

20 **Franko J**, Feng W, Yip L, Genovese E, Moser AJ. Non-functional neuroendocrine carcinoma of the pancreas: incidence, tumor biology, and outcomes in 2,158 patients. *J Gastrointest Surg* 2010; **14**: 541-548 [PMID: 19997980 DOI: 10.1007/s11605-009-1115-0]

21 **Chan AO**, Kim SG, Bedeir A, Issa JP, Hamilton SR, Rashid A. CpG island methylation in carcinoid and pancreatic endocrine tumors. *Oncogene* 2003; **22**: 924-934 [PMID: 12584572 DOI: 10.1038/sj.onc.1206123]

22 **Klöppel G**, Clemens A. The biological relevance of gastric neuroendocrine tumors. *Yale J Biol Med* 1996; **69**: 69-74 [PMID: 9041691]

23 **Le Marc'hadour F**, Bost F, Peoc'h M, Roux JJ, Pasquier D, Pasquier B. Carcinoid tumour complicating inflammatory bowel disease. A study of two cases with review of the literature. *Pathol Res Pract* 1994; **190**: 1185-1192; discussion 1185-1192 [PMID: 7792207 DOI: 10.1016/S0344-0338(11)80445-0]

24 **Cigrovski Berkovic M**, Cacev T, Catela Ivkovic T, Zjacic-Rotkvic V, Kapitanovic S. New insights into the role of chronic inflammation and cytokines in the etiopathogenesis of gastroenteropancreatic neuroendocrine tumors. *Neuroendocrinology* 2014; **99**: 75-84 [PMID: 24686050 DOI: 10.1159/000362339]

25 **Abdul M**, Hoosein N. Relationship of the interleukin-1 system with neuroendocrine and exocrine markers in human colon cancer cell lines. *Cytokine* 2002; **18**: 86-91 [PMID: 12096923 DOI: 10.1006/cyto.2001.1019]

26 **Dinarello CA**. Biologic basis for interleukin-1 in disease. *Blood* 1996; **87**: 2095-2147 [PMID: 8630372]

27 **Corbett JA**, Sweetland MA, Wang JL, Lancaster JR, McDaniel ML. Nitric oxide mediates cytokine-induced inhibition of insulin secretion by human islets of Langerhans. *Proc Natl Acad Sci USA* 1993; **90**: 1731-1735 [PMID: 8383325 DOI: 10.1073/pnas.90.5.1731]

28 **Apte RN**, Dotan S, Elkabets M, White MR, Reich E, Carmi Y, Song X, Dvozkin T, Krelin Y, Voronov E. The involvement of IL-1 in tumorigenesis, tumor invasiveness, metastasis and tumor-host interactions. *Cancer Metastasis Rev* 2006; **25**: 387-408 [PMID: 17043764 DOI: 10.1007/s10555-006-9004-4]

29 **di Giovine FS**, Takhsh E, Blakemore AI, Duff GW. Single base polymorphism at -511 in the human interleukin-1 beta gene (IL1 beta). *Hum Mol Genet* 1992; **1**: 450 [PMID: 1301918 DOI: 10.1093/hmg/1.6.450]

30 **Pociot F**, Mølvig J, Wogensen L, Worsaae H, Nerup J. A TaqI polymorphism in the human interleukin-1 beta (IL-1 beta) gene correlates with IL-1 beta secretion in vitro. *Eur J Clin Invest* 1992; **22**: 396-402 [PMID: 1353022 DOI: 10.1111/j.1365-2362.1992.tb01480.x]

31 **Guasch JF**, Bertina RM, Reitsma PH. Five novel intragenic dimorphisms in the human interleukin-1 genes combine to high informativity. *Cytokine* 1996; **8**: 598-602 [PMID: 8894434 DOI: 10.1006/cyto.1996.0080]

32 **Chen H**, Wilkins LM, Aziz N, Cannings C, Wyllie DH, Bingle C, Rogus J, Beck JD, Offenbacher S, Cork MJ, Rafie-Kolpin M, Hsieh CM, Kornman KS, Duff GW. Single nucleotide polymorphisms in the human interleukin-1B gene affect transcription according to haplotype context. *Hum Mol Genet* 2006; **15**: 519-529 [PMID: 16399797 DOI: 10.1093/hmg/ddi469]

33 **Haukim N**, Bidwell JL, Smith AJ, Keen LJ, Gallagher G, Kimberly R, Huizinga T, McDermott MF, Oksenberg J, McNicholl J, Pociot F, Hardt C, D'Alfonso S. Cytokine gene polymorphism in human disease: on-line databases, supplement 2. *Genes Immun* 2002; **3**: 313-330 [PMID: 12209358 DOI: 10.1038/sj.gene.6363881]

34 **El-Omar EM**, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, Herrera J, Lissowska J, Yuan CC, Rothman N, Lanyon G, Martin M, Fraumeni JF, Rabkin CS. Interleukin-1 polymorphisms associated with increased risk of gastric

cancer. *Nature* 2000; **404**: 398-402 [PMID: 10746728 DOI: 10.1038/35006081]

35 **Wang Y**, Kato N, Hoshida Y, Yoshida H, Taniguchi H, Goto T, Moriyama M, Otsuka M, Shiina S, Shiratori Y, Ito Y, Omata M. Interleukin-1beta gene polymorphisms associated with hepatocellular carcinoma in hepatitis C virus infection. *Hepatology* 2003; **37**: 65-71 [PMID: 12500190 DOI: 10.1053/jhep.2003.50017]

36 **Barber MD**, Powell JJ, Lynch SF, Fearon KC, Ross JA. A polymorphism of the interleukin-1 beta gene influences survival in pancreatic cancer. *Br J Cancer* 2000; **83**: 1443-1447 [PMID: 11076651 DOI: 10.1054/bjoc.2000.1479]

37 **Cigrovski Berković M**, Catela Ivković T, Marout J, Zjačić-Rotkvić V, Kapitanović S. Interleukin 1β gene single-nucleotide polymorphisms and susceptibility to pancreatic neuroendocrine tumors. *DNA Cell Biol* 2012; **31**: 531-536 [PMID: 21988351 DOI: 10.1089/dna.2011.1317]

38 **Chourasia D**, Achyut BR, Tripathi S, Mittal B, Mittal RD, Ghoshal UC. Genotypic and functional roles of IL-1B and IL-1RN on the risk of gastroesophageal reflux disease: the presence of IL-1B-511*T/IL-1RN*1 (T1) haplotype may protect against the disease. *Am J Gastroenterol* 2009; **104**: 2704-2713 [PMID: 19603010 DOI: 10.1038/ajg.2009.382]

39 **Landi S**, Moreno V, Gioia-Patricola L, Guino E, Navarro M, de Oca J, Capella G, Canzian F. Association of common polymorphisms in inflammatory genes interleukin (IL)6, IL8, tumor necrosis factor alpha, NFKB1, and peroxisome proliferator-activated receptor gamma with colorectal cancer. *Cancer Res* 2003; **63**: 3560-3566 [PMID: 12839942]

40 **Theodoropoulos G**, Papaconstantinou I, Felekouras E, Nikiteas N, Karakitsos P, Panoussopoulos D, Lazaris ACh, Patsouris E, Bramis J, Gazouli M. Relation between common polymorphisms in genes related to inflammatory response and colorectal cancer. *World J Gastroenterol* 2006; **12**: 5037-5043 [PMID: 16937502 DOI: 10.3748/wjg.v12.i31.5037]

41 **Lin WW**, Karin M. A cytokine-mediated link between innate immunity, inflammation, and cancer. *J Clin Invest* 2007; **117**: 1175-1183 [PMID: 17476347 DOI: 10.1172/JCI31537]

42 **Karakaxas D**, Gazouli M, Coker A, Agalianos C, Papanikolaou IS, Patapis P, Liakakos T, Dervenis C. Genetic polymorphisms of inflammatory response gene TNF-α and its influence on sporadic pancreatic neuroendocrine tumors predisposition risk. *Med Oncol* 2014; **31**: 241 [PMID: 25213764 DOI: 10.1007/s12032-014-0241-z]

43 **Deans DA**, Wigmore SJ, Gilmour H, Paterson-Brown S, Ross JA, Fearon KC. Elevated tumour interleukin-1beta is associated with systemic inflammation: A marker of reduced survival in gastro-oesophageal cancer. *Br J Cancer* 2006; **95**: 1568-1575 [PMID: 17088911 DOI: 10.1038/sj.bjc.6603446]

44 **Berković M**, Cacev T, Zjacić-Rotkvić V, Kapitanović S. TNF-alpha promoter single nucleotide polymorphisms in gastroenteropancreatic neuroendocrine tumors. *Neuroendocrinology* 2006; **84**: 346-352 [PMID: 17164537 DOI: 10.1159/000097988]

P- Reviewer: Guo XZ, Ooi LLPJ **S- Editor**: Ji FF **L- Editor**: A
E- Editor: Lu YJ

Published by **Baishideng Publishing Group Inc**
8226 Regency Drive, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
http://www.wjgnet.com