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Treatment options for alcoholic and non-alcoholic fatty liver disease: A review

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Abstract

Alcoholic liver disease (ALD) and non-alcoholic fatty liver disease (NAFLD) are serious health problems worldwide. These two diseases have similar pathological spectra, ranging from simple steatosis to hepatitis to cirrhosis and hepatocellular carcinoma. Although most people with excessive alcohol or calorie intake display abnormal fat accumulation in the liver (simple steatosis), a small percentage develops progressive liver disease. Despite extensive research on understanding the pathophysiology of both these diseases there are still no targeted therapies available. The treatment for ALD remains as it was 50 years ago: abstinence, nutritional support and corticosteroids (or pentoxifylline as an alternative if steroids are contraindicated). As for NAFLD, the treatment modality is mainly directed toward weight loss and co-morbidity management. Therefore, new pathophysiology directed therapies are urgently needed. However, the involvement of several inter-related pathways in the pathogenesis of these diseases suggests that a single therapeutic agent is unlikely to be an effective treatment strategy. Hence, a combination therapy towards multiple targets would eventually be required. In this review, we delineate the treatment options in ALD and NAFLD, including various new targeted therapies that are currently under investigation. We hope that soon we will be having an effective multi-therapeutic regimen for each disease.

Key words: Alcoholic liver disease; Non-alcoholic fatty

liver disease; Treatment options; Glucocorticoids; Liver transplantation

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Core tip: Alcoholic liver disease (ALD) and non-alcoholic fatty liver disease (NAFLD) are serious health problems worldwide. Despite extensive research on understanding the pathophysiology of both these diseases there are still no targeted therapies available. In this review, we delineate the treatment options in ALD and NAFLD, including various new targeted therapies that are currently under investigation.

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INTRODUCTION

Alcoholic liver disease (ALD) and non-alcoholic fatty liver disease (NAFLD) are serious health issues whose incidences are on the rise with each passing decade. Alcohol is responsible for approximately 4% of all deaths annually and 5% of all disabilities worldwide^[1]. The Centers for Disease Control and Prevention in 2013 have estimated that in the United States acute deaths from alcohol attributable causes have outnumbered deaths from chronic diseases (44000 to 35000) (<http://apps.nccd.cdc.gov/ardi/homepage.aspx>). Motor vehicle accidents have been considered the leading cause of acute death from alcohol-attributable injuries. While the incidence and prevalence of NAFLD is on the rise with each passing decade. Presently, 25%-35% and 5%-15% of the general population of Western and Asian countries, respectively, are affected by this disease^[2]. This proportion is even higher in people with type 2 diabetes (60%-70%), and in those who are obese or morbidly obese (75%-92%) compared to the general population^[3-5]. The prevalence of obesity in the United States has increased from 10% to 60% of the total population in the last three decades and is considered to be one of the main factors for the increasing prevalence of NAFLD^[6].

The risk factors for both these diseases are well known. Patients with ALD consume an excessive amount of alcohol while NAFLD patients are usually obese; have insulin resistance and/or metabolic syndrome. Available data from various studies show that NAFLD may be the hepatic manifestation of metabolic syndrome^[7]. The spectrum of both diseases ranges from benign steatosis to hepatitis to cirrhosis and hepatocellular carcinoma. Most patients with NAFLD or ALD have hepatic steatosis which is usually asymptomatic; only 20%-35% of these

patients' progress to steatohepatitis or cirrhosis^[8]. The role of genetic polymorphism, mainly the patatin-like phospholipase domain containing 3 (*PNPLA3*) gene (rs738409 variant), has also recently been shown to be a risk factor for progression to advanced liver disease in both NAFLD and ALD that could explain why only a subset of patients who are chronic alcohol abusers or have high caloric intake present with progressive liver injury. Evidence from recent studies has shown this variability may be related to *PNPLA3* variant (rs738409). The single nucleotide polymorphism rs738409 variant within the *PNPLA3* causes a substitution of methionine for isoleucine at position 148. The GG phenotype of this *PNPLA3* variant, rs738409, predicts a greater risk of progression to cirrhosis and HCC than the GC and CC phenotype which have shown to have a smaller risk for progression^[9-12]. Despite an increased understanding of the pathophysiology and risk factors for ALD and NAFLD, we still do not have an appropriate therapeutic regimen for either disease.

The treatment options of ALD have not changed in the last four decades, and abstinence is still the cornerstone of treatment. This is supported by nutrition therapy and steroids^[13,14]. Unfortunately, alcoholic hepatitis, which is the most serious manifestation of ALD, has a short term mortality of up to 50% in patients who are unresponsive to corticosteroid treatment^[15]. Furthermore, limited treatment options are available for patients who are steroid non-responders or have contraindications to steroid usage (upper gastrointestinal bleed, impaired renal functions and sepsis). While the treatment for NAFLD is mainly directed toward attenuating the risk factor such as gradual weight loss by lifestyle modification with a focus on nutrition and exercise^[16,17], other therapies utilizing insulin sensitizers (thiazolidinediones) and antioxidants (vitamin E) also have been found to be useful. However, their long-term safety and adverse effects have not been rigorously evaluated.

Thus, effective and safe therapeutic regimens are needed for these liver diseases. In this review, we present the current therapies as well as upcoming potential new approaches and treatment strategies for both diseases.

ALD TREATMENT

General management

For the last 50 years, abstinence has remained the primary therapy for ALD treatment. However, serious symptoms develop with the abrupt cessation of alcohol. Treating the alcohol withdrawal syndrome is thus extremely important and requires administration of fluid, calories, vitamins and minerals. Unstable patients need to be admitted to a critical care unit and airway protection is often required in patients with hepatic encephalopathy. Table 1 summarizes the treatment options and potential new options for ALD

Table 1 Treatment options for alcoholic liver disease and alcoholic steatohepatitis

General management
Abstinence
Nutritional support
Glucocorticosteroids
Pentoxifylline
Anti-TNF therapy
Antioxidants
Liver transplantation
Potential new therapies
Probiotics and antibiotics
S-adenosylmethionine
Betaine
Targeting various chemokines and interleukins
Endocannabinoids antagonists
Osteopontin inhibition
Stem cell therapy

and ASH (alcoholic steatohepatitis).

Alcohol withdrawal syndrome: This syndrome is characterized by symptoms that occur 6-24 h after abrupt cessation of alcohol in patients who drink consistently and excessively. Long acting benzodiazepines like chlordiazepoxide or diazepam are administered for prevention of seizures while intermediate acting benzodiazepines like lorazepam are recommended in withdrawal patients who are elderly or have had recent head trauma or liver or respiratory failure^[18]. Antiepileptic like carbamazepine can also be used as a benzodiazepine substitute for preventing seizures. Antipsychotics like haloperidol can be used if patients have excess agitation or psychotic symptoms^[18]. Alcoholics are usually malnourished and deficient in vitamins, especially vitamin B1 (thiamine), thus putting them at risk of developing Wernicke encephalopathy, so all such patients should be given thiamine^[19]. Parenteral thiamine is preferred over oral thiamine because in addition to impaired gastrointestinal absorption in alcoholics, oral thiamine has poor bioavailability that does not allow for attaining a sufficient concentration in cerebrospinal fluid. However, parenteral thiamine has a short half-life, thus multiple high-dosing is required to achieve sufficient concentration for active and passive diffusion through the blood brain barrier^[20-22]. An IV dose of 500 mg (three times daily for two consecutive days) is recommended, followed by 500 mg of IV or IM thiamine for five more days if a response to the therapy is seen^[23-25]. Another dosing regimen is 500 mg of IV thiamine (three times daily for 2-3 d), followed by 250 mg of IV thiamine for the next 2-3 d. The intravenous regimen is then followed by oral thiamine treatment indefinitely^[26,27]. Thiamine should be given before fluids containing glucose to prevent neurological damage.

Abstinence: The first step towards treatment requires the patient to accept the fact that they are

dependent and addicted to alcohol. Abstinence can resolve alcoholic fatty liver disease and can improve the survival rate of cirrhotic or decompensated liver failure patients. Thus, motivating the patients to abstain from alcohol and follow the proper treatment regime are major steps. However, preventing relapse in such patients has always been a big challenge. Patients can participate in Alcoholics Anonymous group meetings for self-control and motivation. Psychological support by an addiction specialist can also help in maintaining sobriety. Recognizing and treating any associated psychiatric conditions can be helpful in such patients^[28]. Pharmacotherapy also helps in maintaining sobriety; drugs like naltrexone and acamprosate assist in reducing alcohol intake in heavy drinkers^[29,30]. Topiramate has found to be effective in decreasing craving and withdrawal symptoms in alcoholics^[31]. Disulfiram, an acetaldehyde dehydrogenase inhibitor, is also being used. It causes accumulation of serum acetaldehyde, which produces unpleasant sensations of nausea, vomiting, abdominal pain and dizziness. Such sensations deter patients from consuming alcohol^[32]. Baclofen, a γ -amino butyric acid agonist, has also been found effective in promoting abstinence^[33]. Most of these drugs (disulfiram, acamprosate, naltrexone, topiramate and baclofen), are used only for treatment of alcohol dependence^[34] but are not FDA approved for ALD treatment. Of these, only baclofen has been studied for its safety and efficacy in clinical trials that compared placebo and baclofen treatment for ALD patients. In this trial, baclofen (10 mg or 20 mg twice daily) was more effective than placebo in maintaining abstinence. A 53% reduction in the number of drinks per day in the 10 mg baclofen group ($P < 0.0001$) and a 68% reduction in the number of drinks per day in the 20 mg baclofen group was reported, thus demonstrating a dose response effect of baclofen^[35]. Naltrexone and disulfiram should be avoided in patients with liver problems because of hepatotoxicity^[36]. Moreover, naltrexone should also be avoided in patients with renal failure due to its active tubular secretion^[37]. Acamprosate, topiramate and baclofen were found to be safe in such patients^[33,38,39]. Another drug which can help to maintain abstinence and reduce craving is metadoxine (MTD). Although not available in the United States, MTD is approved for use in several European countries. Apart from sustaining abstinence, oral MTB is also useful in acute ethanol intoxication since it is rapidly absorbed and augments alcohol metabolism by enhancing acetaldehyde dehydrogenase activity^[39]. In a clinical trial with ASH patients, MTX improved liver function tests in 1 mo compared to the placebo^[40]. In another trial, improved 3-6 mo survival of severely ill ASH patients who received the combination therapy with MTX compared to those that received monotherapy with either steroids or pentoxifylline (PTX) was observed. The survival rate was higher of the MTD-treated groups at

3 mo (PTX + MTD 59.4% vs PTX 33.3%, $P = 0.04$; steroids + MTD 68.6% vs steroids 20%, $P = 0.0001$) and at 6 mo (PTX + MTD 50% vs PTX 18.2%, $P = 0.01$; steroids + MTD 48.6% vs steroids 20%, $P = 0.003$) than in the non-MTD treated groups. Furthermore, the patients receiving MTD maintained greater abstinence than those not on MTD (74.5% vs 59.4%, $P = 0.02$)^[41]. Smoking and obesity are independent risk factor for the progression of ALD^[42,43]. Hence, lifestyle modifications like weight loss and smoking cessation are also helpful. Hepatitis C virus (HCV) is another independent risk factor for ALD progression due to synergistic deleterious effects of both agents in promoting liver injury and HCC development^[44,45]. The main mechanisms of this effect are that both alcohol and HCV alter cellular immunity, increase free radical oxidative damage, and in the case of alcohol exposure, promote replication of HCV. Thus, the combination often result in the presence of advanced liver disease with severe histological features at a much younger age and a decreased survival^[46,47]. It has been reported that alcoholic patients with HCV infection have a 30 fold increased risk of getting cirrhosis^[48] and two to eight fold increased risk of all-cause mortality compared with those without the HCV infection^[49,50]. Thus, all ALD patients should be screened for HCV before starting treatment and all HCV patients should be advised to stop or reduce alcohol consumption^[51].

Nutritional support: Most patients with ALD are malnourished, and disease severity often correlates with the degree of malnutrition^[52]. Most of the complications of ALD are strongly associated with protein calorie malnutrition^[53]. Thus, nutrition support is one of the important steps in ALD treatment. Vitamins (like folate, vitamin B6, vitamin B12^[54,55], vitamin A and thiamine^[56] and minerals (like selenium, zinc, copper, and magnesium) are often found to be altered in ALD and some believe that these alterations play a role in initiation and progression of liver injury^[57]. Especially, zinc levels are decreased in ALD patients and in animal models, and its supplementation has been shown to improve ALD^[58]. A major study has also shown that enteral nutrition reduces infectious complications and improves 1-year mortality in such patients^[59,60].

The American College of Gastroenterology and the American Association for the Study of Liver Diseases guidelines recommend 1.2 to 1.5 g/kg per d of protein intake and 35 to 40 kcal/kg per d of body weight for energy intake in patients with ALD^[61]. This type of malnourished patient is often predisposed to infections so empiric antibiotic treatment is also advised.

Glucocorticosteroids: There have been various clinical trials on the use of corticosteroids for treating ALD patients^[62-64]. Despite mixed outcomes, corticosteroids are overall beneficial for survival of these patients. Unfortunately, 40% of patients are unresponsive to corticosteroid with virtually no other treatment options.

Hence, new target oriented therapies are critically required for the management of such patients^[15].

A meta-analysis which pooled data from 3 randomized control trials, found that patients with modified DF ≥ 32 or MELD score ≥ 21 treated with prednisolone at 40 mg/d for 28 d and then a tapered dose over 2-4 wk (Class I, level A), conferred a 28-d survival benefit of glucocorticoids (85%) vs placebo (65%), with mortality decreasing from 35% in controls to 15% in patients on steroids^[64]. Early changes in bilirubin levels (at day 7 of treatment) and the Lille score were used to predict the prognosis following steroid administration^[65]. A Lille's score greater than 0.45 on the 7th d after initiation of the treatment indicated that the patient was unresponsive to steroid therapy and predicted a lower survival rate of 25% at 6 mo. Recently this score has been re-classified as complete responders (score ≤ 0.16), partial responders (score 0.16-0.56), and null responders (score ≥ 0.56), and is associated with the 28-d survival rate of 91%, 79% and 53%, respectively, with $P < 0.0001$ ^[13]. Steroids have been found to have a significant beneficial effect in complete and partial responders but not in null responders, hence discontinuation of steroid therapy is recommended for non-responders^[66]. In addition to non-responders, steroids are generally avoided in patients with active infection, gastrointestinal bleeding, chronic hepatitis B virus infection or hepatorenal syndrome (HRS) because of adverse effects in these patient populations^[67]. Steroids are relatively contraindicated in severe AH patients with coexistent sepsis. Thus, such patients may be treated with second line drug PTX^[68]. Patients should also be screened for any infection before starting steroids and for infective complications while on steroids. Occurrence of sepsis and infective complications while the patient is on steroids is a poor prognostic sign^[69]. It has been reported that patients infected after initiation of steroids had a significant lower 2-mo survival than patients with no infection (46.4% \pm 6.9% vs 77.5% \pm 3.2%, $P < 0.00001$). Thus, it is very important to differentiate infection at admission from that which occurs after starting the steroid treatment, as survival rates differ significantly. Overall, infection was more common in steroid null responders than responders^[70].

PTX: Steroids are generally used as the first line of treatment in severe alcoholic hepatitis patients with DF ≥ 32 , except in those with renal failure or HRS or contraindication to steroids^[71]. PTX (400 mg 3 times per day for 28 d) is a substitute in such cases (Class I, level B). PTX decreases pro-inflammatory cytokines like TNF- α , has anti-fibrotic properties^[72] and confers a mortality benefit by reducing the incidence of HRS^[73]. A pilot study in ASH patients using PTX demonstrated reduced mortality and HRS incidence when compared to patients given a placebo^[74]. These findings were later confirmed in a double-blind placebo controlled trial, where PTX decreased the 28-d mortality compared to

placebo (24.5% vs 46%). Also, 50% of those who died in the PTX group developed HRS, while 91.7% who died in the placebo group developed HRS, confirming that PTX reduces the incidence of HRS in such patients^[75]. A study in ASH patients comparing PTX and prednisolone have shown a better survival rate of 35.29% in the PTX group vs 14.71% on steroids. This reduced mortality was presumably because of a decrease in incidence of HRS and gastrointestinal bleeding in the PTX group. However, this study was underpowered^[76]. To date, no other study has shown any additional survival benefit of the combined PTX and corticosteroid treatment^[77,78]. A recently conducted randomized, multicenter, double-blind trial (STOPAH) across 65 hospitals in the United Kingdom that recruited more than a thousand patients revealed no impact of PTX on survival or disease progression in severe non-alcoholic steatohepatitis (NASH) patients in comparison to placebo^[79,80]. However, because of a lack of other treatment options, PTX is being used in some centers.

Anti-TNF therapy: Intestinal gut permeability is increased in chronic alcoholics that promotes the translocation of gut luminal antigens especially endotoxin to reach the liver and enhance TNF- α production^[81]. TNF- α has been found to correlate with disease severity in severe alcoholic hepatitis patients^[82], and also play a vital role in alcohol induced liver injury in various animal models of alcoholic liver injury^[83]. Further, mice deficient in TNF receptor 1 do not develop liver injury when administered alcohol^[83]. Based on these considerations, various human studies were undertaken using anti-TNF therapy. While initial studies were found to be promising, the results could not be duplicated in larger clinical trials. A large randomized controlled trial comparing prednisolone alone with a combination of prednisolone and infliximab had to be stopped before completion because of an increase in infection rate in the prednisolone and infliximab combination group^[84]. Further, patients had to be screened for tuberculosis and nocardia infection prior to participation in the study, thus limiting its clinical utility^[85].

Antioxidants: Alcohol causes oxidative stress by increasing reactive oxygen species (ROS), and decreasing endogenous antioxidant levels^[86]. But to date, all trials examining antioxidants (such as lecithin, β -carotene, vitamin C, vitamin E, allopurinol, desferrioxamine, and N-acetylcysteine) either alone or in combination with steroids have been disappointing^[87,88].

Liver transplantation: Liver transplantation remains the definitive therapy for end stage decompensated cirrhosis due to ALD. Severe alcoholic hepatitis patients nonresponsive to steroids have a 3 mo mortality rate of 70% and with HRS the mortality rate is $\geq 90\%$ unless the patients get liver transplantation^[89,90]. At

present, there are very few options for treating severe alcoholic hepatitis patients who are non-responsive to steroids and have a Lille score > 0.56 . Thus, liver transplantation remains the only hope for such patients, but the issue of transplantation in alcoholics has always remained controversial. Concerns include the risk of recidivism, poor compliance with postoperative care, and ALD being a self-inflicted disease^[91]. Recidivism following transplantation is a major challenge, which occurs at a rate of 10%-50%^[92,93]. A meta-analysis reviewing factors responsible for recidivism found 3 major variables: a poor social support system, a family history of alcohol abuse/dependence and pre-transplant abstinence of 6 mo or less^[94]. Thus, we need a multidisciplinary approach including the presence of an Alcohol Addiction Unit which can significantly contribute in reducing alcohol relapse after transplantation. Also, there should be psychological evaluation for any mental illness to determine patient suitability for transplantation.

Most transplant programs require the patients to undergo a 6-mo period of abstinence prior to transplantation^[95]. Studies over the years have provided data both for and against the 6-mo abstinence rule. One report suggested that the 6-mo period of abstinence would allow the liver to recover with medical treatment and possibly there would be no need for transplantation^[96]. Another study revealed that some recovery in liver function can take place within 3 mo of abstinence while many patients may die during the 6 mo of waiting period. This led to the suggestion of possibly reducing the period of abstinence to 3 mo^[97]. Yet another study has also challenged the 6-mo abstinence rule by showing beneficial effects of early liver transplantation in steroid non-responding severe alcoholic hepatitis patients. In this study patients (with Lille score of 0.88) after 13 d of being unresponsive to steroids were put on the transplant list and it was found that the 6-mo survival rate was higher in patients who received early transplantation than those who did not (77% vs 23%, $P < 0.001$)^[98].

However, patients who have received liver transplantation show a high incidence of de novo cancer^[99,100], lymphoproliferative disorder and skin cancer. In some cases, squamous cell carcinoma of the oropharynx or esophagus has also been detected, likely due to the cumulative effects of smoking and post-transplant immunosuppressive drugs. Also, liver transplantation due to ALD is associated with a high rate of cardiovascular complications^[101].

Potential new therapeutic options in ALD

Advances in basic science have helped to gain better insights into the pathophysiology of ALD that have provided new treatment options as discussed below.

Role of probiotics and antibiotics: Healthy intestinal flora is critically important for our well-being.

An alcohol-induced change in the gut microflora plays a major role in the pathogenesis of alcoholic hepatitis. Equally important in liver disease progression is the alcohol-induced increased intestinal permeability that allows for the gut luminal antigens, including endotoxin/LPS (component of the cell wall of gram negative bacteria), to reach the liver and promote the synthesis and secretion of several inflammatory cytokines^[102]. Various studies have proposed the use of probiotics in restoring the normal bowel flora in patients with ALD^[103]. In a study performed on patients with ALD it was shown that using probiotics (*Bifidobacterium* or *Lactobacillus*) for 4 wk enhances and normalizes neutrophil phagocytic capacity and helps in reducing endotoxin-driven elevation in cytokine levels^[104]. A similar study revealed significant improvement in AST, ALT and γ -glutamyl transferase levels in ALD patients administered probiotics (*Bifidobacterium* or *Lactobacillus*) for 5 d^[105]. Rifaximin, a biochemical derivative of Rifamycin, the drug for hepatic encephalopathy, given for 28 d in a clinical trial decreased systemic endotoxin levels^[106]. Indeed, blood LPS levels help in predicting response to steroids and mortality of alcoholic hepatitis patients^[107]. Thus, modifying the gut microbe flora by probiotics and antibiotics could be a potential therapeutic approach for treating ALD which is being actively pursued.

Role of S-adenosylmethionine and betaine:

S-adenosylmethionine (SAM) is a key methyl donor that is involved in many methylation reactions critical for normal liver function. SAM also acts as an antioxidant by activating the pathway for GSH synthesis. Decreased SAM levels have been reported in ALD patients; thus, elevating SAM levels could be a potential therapy. Various animal studies have shown liver injury can be reversed by preventing a decrease in SAM levels^[108]. Also, SAM administration decreases oxidative stress and hepatic stellate cell activation^[109]. A randomized controlled trial using SAM or placebo for 2 years in alcohol cirrhotic patients found that the mortality and liver transplantation rates were higher in the placebo group than in the SAM group (29% vs 12%)^[110]. Thus, there is need for long-term, high quality trials in the future to establish its effectiveness. Along the same line as SAM, betaine treatment has been very effective in improving liver injury in various animal models^[111,112]. By remethylation homocysteine to generate methionine, betaine not only removes the toxic metabolites homocysteine and S-adenosylhomocysteine, but also generates SAM and normalizes the methylation potential^[113]. Betaine is hepato-protective and prevents alcohol-induced steatosis, oxidative stress, apoptosis and abnormal protein accumulation^[111,112], and breakdown of sulphur containing amino acid^[114]. Clinical trials using betaine should be conducted.

Role of targeting various chemokines and

interleukins: Chemokines play a pivotal role in the pathogenesis of alcoholic hepatitis. Studies have shown that various chemokines and their subfamily members, including CXCL5, CXCL6, CXCL10 and CCL20, are notably high in ASH livers compared to normal control livers and higher levels correlate with worse prognosis and outcomes^[115,116]. Of these, CCL20 is the most elevated chemokine in ASH livers that attracts lymphocytes, monocytes, Th17 (Helper T17) cells, and dendritic cells. The consequent production of more chemokines and inflammatory mediators ultimately causes heavy neutrophilic infiltration and liver damage^[117,118]. Additional studies in the future are required to determine if targeting CCL20 and other chemokines can be an effective and safe therapeutic approach for ALD patients.

IL-8 is one of the most important chemoattractant of neutrophils, which further causes hepatic infiltration as well as increased portal pressure^[115]. A higher level of IL-8 in alcoholic hepatitis patients is associated with worse prognosis^[115]. A therapeutic approach towards counteracting IL-8 levels should be considered as it will decrease neutrophil infiltration of the liver and prevent progressive liver damage.

IL-22 plays a critical role in bacterial infections and tissue repair. It is a part of the IL-10 family which decreases the production of various pro-inflammatory cytokines^[119]. IL-22 has been found to have anti-apoptotic, antimicrobial, antioxidant and anti-steatotic effects, thus it can be used as a therapeutic option in ALD patients. It has been found that levels of T helper cells producing IL-22 correlate with improvement in alcoholic hepatitis patients^[120]. Recombinant IL-22 administration showed improvement of liver injury in ethanol-fed mice^[121] and in an animal model of acute hepatitis while blocking the IL-22 receptor led to worsening of the disease^[122]. Thus, up regulating IL-22 levels can be a potential therapy for ALD.

IL-17 increases chemotaxis of neutrophils and various other chemokines and its levels are found to be increased in alcoholic hepatitis^[123]. Secukinumab, an anti-IL-17 monoclonal antibody has shown favorable results in clinical trials of rheumatoid arthritis, psoriasis and uveitis^[124]. Up until now, no study has been done in patients with liver disease using this monoclonal antibody, which can be a potential therapy.

Role of endocannabinoids: Endocannabinoids signalling through cannabinoid receptors, CB-1 and CB-2, has been implicated in the pathogenesis of ALD^[125]. Studies using animal models of alcoholic liver injury revealed that CB1-deficient mice are resistant, whereas CB2-deficient mice are more susceptible to fatty liver damage^[126,127]. These findings suggested that therapy targeting CB1 and CB2 receptors should be utilized as an alternative for the management of ALD.

Role of osteopontin: There is substantial evidence

Table 2 Treatment options for non-alcoholic fatty liver disease and non-alcoholic steatohepatitis

Lifestyle changes	Weight loss
	Dietary changes
	Exercise
Insulin sensitizers	Thiazolidinedione's
	Metformin
Lipid lowering agents	Statins
	Ezetimibe
Hepatoprotective agents	UDCA
Antioxidants	Vitamin E
Incretin analogues	GLP-1 agonists
	DPP-IV inhibitors
Anti-inflammatory agents	PTX
Others	Probiotics
	Angiotensin receptor blockers
	Endocannabinoid antagonists
	Bariatric surgery
	Liver transplantation
Potential new therapeutic options	Caspases inhibitors
	ASK1 inhibitors
	p38 MAPK inhibitors
	PPAR- alpha and delta agonists
	FXR agonists
	NOX-1/4 inhibitors
	Galectin-3 antagonists
	Acetyl CoA carboxylase inhibitors
	FGF-21 and FGF-19 analogues
	CCR2 and CCR5 inhibitors
	SCD-1 inhibitors
	Lysyl oxidase-like 2 inhibitors
	Sirtuins

GLP-1: Glucagon-like peptide 1; DPP-IV: Dipeptidyl-peptidase IV; PTX: Pentoxifylline; PPAR: Peroxisome proliferator activated receptor; NOX: NADPH oxidase; FGF-21: Fibroblast growth factor 21; SCD1: Stearoyl coenzyme A desaturase 1.

suggesting that osteopontin (OPN) plays a notable role in wound healing in response to injury in many organs^[128]. It is an extracellular matrix protein with pro-fibrogenic properties, and is found to be highly expressed in alcoholic hepatitis patients^[129]. One study demonstrated attenuation of alcohol mediated liver disease in mice lacking OPN^[130]. More studies should be conducted to assess OPN as a potential therapeutic target.

Stem cell therapy: Hematopoietic stem cell transplantation is an evolving field. Though limited research has been performed in this area, it could be a promising therapeutic approach in the future. Recent studies have suggested that stem cell transplantation may reduce liver inflammation and improve fibrosis in patients with liver cirrhosis^[131]. Mesenchymal stem cells (MSC) directly inhibit the activation of hepatic stellate cells and may also induce apoptosis of hepatic stellate cells^[132]. They have also been reported to stimulate proliferation of endogenous hepatocytes^[133,134]. A pilot study performed on 12 patients with ALD to assess the regenerative capacity of the liver after infusion of bone marrow derived-MSC through the hepatic artery showed improvement in the histological grading with

an overall decrease in TGF- β , type 1 collagen and smooth muscle actin^[135]. A significant improvement in the Child-Pugh score and albumin levels was noted in another similar study on 9 cirrhotic patients given bone marrow derived stem cells *via* the portal vein^[136]. Liver function was also reported to be better after stem cell therapy in cirrhotic patients^[137]. Results of these studies are encouraging and stem cell therapy could serve as a potential breakthrough treatment for ALD. However, the benefits and safety of stem cells should be examined in a large sized RCT.

NAFLD TREATMENT

Like ALD there is no effective treatment to date for NAFLD. In the absence of a proven effective therapy, we must follow a multi-disciplinary approach in NAFLD treatment, where a combination of drugs and factors are taken into consideration to counter multiple pathological risk factors involved in NAFLD. These are summarized in Table 2 and are further discussed below.

Weight loss, dietary modification and changes in lifestyle

Treatment is mainly directed towards weight loss and risk factor reduction, as most patients are obese or have metabolic syndrome^[138]. A weight loss of 3%-5% reduces steatosis while a $\geq 5\%$ -7% drop in weight has been shown to resolve NASH. Greater reductions in weight (*i.e.*, $\geq 10\%$) may also improve hepatic fibrosis. Weight loss is mainly due to diet modification and exercise. However, the shortcoming of this approach is the lack of adherence and non-compliance with time^[139]. Various studies have shown the benefit of weight loss in NAFLD^[140]. Dietary modification also plays a key role since a carbohydrate-rich diet, especially with high fructose, is the major cause of obesity, insulin resistance and NAFLD development^[141]. Thus, sugar consumption should be kept below 10% of total caloric intake in a day and a fructose-rich diet should be avoided in such patients. Food rich in omega-3 fatty acid should be included and those rich in saturated fat and omega-6 fatty acid should be excluded from the diet^[142]. An omega-3 fatty acid rich diet promotes fatty acid oxidation and decreases fatty acid synthesis, thus improving the lipid profile. Fish and fish oil consumption should be promoted as they are rich in omega-3 fatty acid^[143]. Thus, diet and moderate exercise are preferred methods of natural weight loss. A study also revealed that a combination of diet changes and exercise lowered ALT levels better in NAFLD patients than insulin sensitizers or other hypoglycaemic drugs^[144]. Weight loss is also beneficial as it improves the cardiovascular risk profile^[145]. Nevertheless, it should be noted that weight loss should be gradual, as very rapid weight loss has been associated with a worsening of steatohepatitis and an increased risk for liver failure^[146] and gallstones^[147].

Apart from natural weight loss, drugs like Orlistat and Sibutramine are also being used for controlling weight. Orlistat is a lipase inhibitor, that prevents fat absorption in the liver and intestine, thus causing weight loss. Sibutramine on the other hand is a serotonin reuptake antagonist which suppresses appetite. Both agents have shown to reduce serum transaminase levels and hepatic steatosis^[148,149].

Insulin sensitizers

Since NAFLD is closely associated with obesity and metabolic syndrome, and both conditions cause insulin resistance, treatment strategies invariably include agents which enhance insulin sensitivity.

Thiazolidinedione: Thiazolidinediones (TZDs) are peroxisome proliferator activated receptor (PPAR)- γ agonists, which improve hepatic and peripheral insulin sensitivity^[150] via increasing plasma adiponectin levels^[151]. In addition, adiponectin is also shown to have anti-fibrotic and anti-inflammatory properties. Thus, multiple factors involved in pathogenesis of NAFLD such as high insulin resistance, low adiponectin levels and high pro-inflammatory cytokines are all targeted by these drugs. First generation TZDs (troglitazone) have shown improvement in steatohepatitis but had to be stopped due to hepatotoxicity^[152]. However, it paved the way for second generation TZDs (rosiglitazone and pioglitazone) which are not hepatotoxic and showed improvement in insulin resistance, hepatic steatosis and aminotransferases levels^[153,154]. A long-term therapy with second generation TZDs may be required as their benefits tend to reverse on discontinuation; however long term therapy is associated with various adverse effects like congestive heart failure, weight gain, peripheral oedema, anaemia and osteoporosis^[155,156]. Also, it has been found that TZD therapy alone without nutrition and lifestyle changes is often not effective^[154]. Thus, we need additional studies on a larger population with a combination of other drugs to find safe and efficacious treatment options.

Metformin: Metformin, a hypoglycaemic drug, is used for treatment of type 2 diabetes mellitus. Metformin improves hepatic and peripheral insulin resistance by decreasing hepatic gluconeogenesis, lipogenesis and glucose reabsorption from the gut and increasing fatty acid oxidation^[157]. While metformin does not cause weight gain as TZDs, it can cause some minor gastrointestinal adverse effects and sometimes lactic acidosis is seen in patients with renal impairment. Various studies have documented an improved insulin sensitivity, cholesterol and aminotransferase levels in NASH patients on metformin but the results are mixed when assessing biopsy-guided improvement in steatosis and NASH activity score (NAS)^[158]. Thus, while its effectiveness as a monotherapy is debatable,

metformin could be a part of a multi-therapeutic regimen for the management of NAFLD patients.

Lipid lowering agents

NAFLD is often associated with obesity and metabolic syndrome which is characterized by hypercholesterolemia and hypertriglyceridemia. Therefore, the use of lipid lowering agents could be beneficial. While, clofibrate did not show any beneficial effect on the liver tests or the histological scores^[159], gemfibrozil showed improvement in ALT levels in NAFLD patients compared to the placebo^[160]. Statins have also been tried but have shown variable effects. Nevertheless, lipid-lowering agents should be given as most NAFLD patients are hyperlipidemic and thus have a high risk of developing cardiovascular issues.

Ezetimibe, a drug that inhibits the reabsorption of lipids from the intestine, reduces serum TNF- α levels^[161], hepatic lipid content and ALT levels in a mouse model of NAFLD^[162]. Human studies for this drug are awaited.

UDCA

This drug has hepatoprotective properties and has been studied in various clinical trials for NAFLD treatment. Initial small studies revealed an improvement in liver enzyme levels and hepatic steatosis^[159], but a subsequent RCT showed no improvement in liver histology or aminotransferases^[163]. Thus, UDCA is not approved as a monotherapy but is part of a drug combination regime in various trials on NAFLD in progress.

Vitamin E

ROS generation plays an important part in the progression of NASH^[164]. Vitamin E and C decrease oxidative stress and thus have been evaluated in patients with NASH. Various clinical trials with vitamin E have revealed an improvement in liver test functions and reduction in oxidative stress markers but significantly less improvement in the histological grading of the disease has been noted^[165,166]. A recent trial using a combination of vitamins E and C for 6 mo showed that these were no better than placebo for treating patients with NASH^[167]. One study with a three arm trial involving placebo, UDCA and Vitamin E/UDCA combination showed improvement in histology only in the Vitamin E/UDCA combination arm^[168]. Another trial comparing a combination of pioglitazone and vitamin E with vitamin E alone over a period of 6 mo showed a decrease in serum ALT in both groups, but a significant histological improvement was only seen in the combination group^[169]. A meta-analysis involving high-dose vitamin E supplementation has shown an increase in all-cause mortality and cardiovascular deaths, thus decreasing the enthusiasm for vitamin E therapy^[170].

Incretin analogues

Glucagon-like peptide 1 agonists: Glucagon-like peptide 1 (GLP-1) is an incretin hormone that is

produced by intestinal mucosa L cells. GLP-1 has a short half-life, as it is rapidly degraded by dipeptidyl-peptidase IV (DPP-IV). GLP-1 agonists are resistant to DPP-IV and are useful since they lower blood glucose levels by decreasing glucagon secretion, delay gastric emptying and stimulate pancreatic β cells to increase insulin secretion. Furthermore, these agonists have a central appetite suppressive effect and promote weight loss which are favorable outcomes for obese NAFLD/NASH patients^[171]. In an obese mouse model, it has shown to improve insulin sensitivity and reduce hepatic steatosis^[172].

In various clinical trials, liraglutide has proven to be an effective therapeutic drug for type 2 diabetics producing a good glycemic control and significant weight loss in such patients. Since diabetes is an important component of metabolic syndrome and associated NAFLD development, the effective glycemic control and weight loss makes liraglutide a suitable therapeutic option for NAFLD^[173]. In a phase 2 clinical trial study (LEAN study) with 52 NASH subjects using liraglutide compared to placebo, 39% of patients using liraglutide vs 9% using placebo attained the primary endpoint (histological resolution of NASH without worsening of fibrosis). Two (9%) of 23 patients in the liraglutide group vs eight (36%) of 22 patients in the placebo group had progression of fibrosis. The trial was designed using A'Hern's single-group method, which required eight (38%) of 21 successes in the liraglutide group for the effect of liraglutide to be considered clinically significant. The liraglutide treatment group has also shown improved insulin sensitivity, reduced hepatic glucose production and lipogenesis (ClinicalTrials.gov-NCT01237119). Thus, liraglutide was safe, well-tolerated, and led to histological resolution of NASH, warranting longer term studies in such patients^[174,175].

DPP-IV inhibitor: DPP-IV inactivates both incretin hormones (GIP, GLP-1), therefore DPP-IV inhibitors are used in the treatment of type 2 diabetes^[176]. NASH patients exhibit higher DPP-IV expression^[177]. A cross sectional study on type 2 diabetics without any evident liver disease and NAFLD patients revealed a strong positive correlation of serum DPP-IV activity and insulin resistance with liver enzymes only in NAFLD patients^[178]. Serum DPP-IV activity was not increased in the type 2 diabetics with no evidence of liver disease. This led the authors to postulate that the increased serum DPP-IV reported in earlier studies in type 2 diabetics may have been due to some un-diagnosed liver disease and that the excess DPP-IV found in the serum of NAFLD patients is of hepatic origin. They further suggested that serum DPP-IV should be considered as a potential liver disease biomarker^[178]. This supposition was also corroborated by another study which analyzed human liver biopsy specimens and showed a strong correlation of DPP-IV expression with stages of

fatty liver and NASH^[179].

Furthermore, a DPP-IV inhibitor like sitagliptin, decreases hepatic steatosis and serum transaminases levels when given to diabetic NAFLD patients^[180,181]. In a recent randomized, double-blind, placebo controlled study, sitagliptin was shown to be safe but no more effective than placebo in improving hepatic steatosis and fibrosis in NAFLD patients. However, in comparison to sitagliptin, an increase in hyaluronic acid levels and increase in FIBROSpect II index (measure of liver fibrosis) was reported in the placebo arm^[182]. Another RCT comparing sitagliptin to placebo also revealed no improvement in fibrosis score or NAS after 24 wk of therapy, but reported that sitagliptin increased adiponectin and decreased γ -glutamyl transferase levels^[183]. There have been only a few clinical trials with sitagliptin till date. However, despite the lack of convincing evidence, the efficacy of sitagliptin in improving liver fibrosis in NAFLD cannot be ruled out. This is because not only were the trials underpowered but were possibly not long enough to evaluate its effectiveness. Hence, sitagliptin effect should be assessed in clinical trials of longer duration with larger number of enrolled patients with NAFLD/NASH.

PTX

PTX can be of potential benefit in NAFLD due to its effects on reducing free radical oxidative stress, TNF- α levels, and potential anti-fibrotic properties^[184]. In some trials, PTX has shown improvement in steatosis, lobular inflammation and ballooning degeneration in comparison to baseline, but improvement was not clinically significant when compared to placebo^[185]. In a small RCT on NASH patients, 400 mg PTX given three times per day for a period of 1 year decreased hepatic steatosis, inflammation and NAS by ≥ 2 points and modestly reduced fibrosis^[186]. This favorable response was due to a reduction in free-radical-mediated lipid peroxidation^[187]. In two recent small RCT evaluating the role of PTX has also shown beneficial effect by improving liver enzymes and histology in NAFLD patients^[188,189]. In a recent meta-analysis it was found that only PTX and OCA improve fibrosis in NASH patients^[190]. Therefore, further studies are warranted to determine its role in NAFLD/NASH treatment.

Others

Probiotics: Like alcoholic patients, NAFLD patients also exhibit gut bacterial overgrowth, enhanced gut permeability and increased paracellular leakage of gut luminal antigens, factors that promote NASH development. Thus, probiotics can be a therapeutic option for NASH patients^[191,192]. In a RCT, improvement in liver enzymes was noted in NAFLD patients on *Lactobacillus bulgaricus* and *Streptococcus thermophilus* treatment compared to placebo^[193]. In another study, patients randomized to a combination of *Bifidobacterium longum* with fructo-oligosaccharides plus lifestyle

modification (diet and exercise) or lifestyle modification alone for 24 wk^[194], showed a significant decrease in steatosis, TNF- α , AST and NAS in the combination treatment group. Thus, probiotics could also be a part of a combination therapy for NAFLD patients.

Angiotensin receptor blockers: NAFLD is often associated with metabolic syndrome and hypertension is an important component of metabolic syndrome. Thus, angiotensin receptor blockers can be a part of combination therapy regimen of NAFLD. A small pilot study of patients with NASH showed improvements in necro-inflammation and fibrosis with losartan (an angiotensin II receptor antagonist) treatment^[195]. Larger studies are required to explore their potential in the management of NAFLD.

Endocannabinoid antagonists: CB1 and CB2 are two receptors which mediate endocannabinoid (EC) activity. The CB1 receptor is mainly expressed in the brain and liver, while CB2 is mainly expressed in the immune cells. These receptors are found to be upregulated in various liver diseases^[196]. Anandamide, a highly potent endogenous agonist, has been shown to promote diet-induced obesity and hepatic steatosis in mice *via* acting on the CB-1 receptors.^[197] Conversely, CB-1 knockout or rimonabant (CB-1 receptor antagonist)-treated high-fat diet fed mice have less steatosis and weight gain than controls^[198]. However, while rimonabant was effective in promoting weight loss of obese patients in many clinical trials, it also caused intolerable adverse effects like depression, anxiety and increased suicidal tendencies that has led to its discontinuation for routine use. Thus, novel cannabinoid type 1 receptor blockers with selectivity for peripheral receptors are required which can have favorable metabolic benefits but decreased psychiatric adverse effects^[199,200].

Bariatric surgery: Steady weight loss with exercise and lifestyle modification has been found to increase insulin sensitivity and improve liver histology of NAFLD patients. But the rapid weight loss induced by bariatric surgery increases the risk of developing hepatic failure especially in the cirrhotic patients^[201,202]. Bariatric surgery is mostly done in non-cirrhotic NAFLD patients who are morbidly obese. It is, however, not recommended as a primary mode of treatment in such patients as there is still a risk of developing liver failure postoperatively.

Liver transplantation: NAFLD patients with end-stage decompensated liver disease should be considered for liver transplantation. But this is not a permanent cure as NAFLD has been shown to recur in post-transplant liver^[203]. This is because transplantation does not correct the multifactorial pathway alteration(s) responsible for NAFLD/NASH development. Therefore, the goals of therapy before and after transplant should be always

towards weight management, proper diet consumption and adequate control of glucose and lipids.

Potential new therapeutic options in NAFLD

With advancement in the field of technology, especially bioinformatics and biogenetics, new therapies are currently being tried for managing NASH, some of which are reviewed below.

Caspase inhibition/emricasan: Caspases are enzymes which are required for completion of various apoptotic pathways and for stimulation of various cytokines and therefore, can be a potential therapeutic target. Various animal studies in the past have supported this approach^[204,205]. Emricasan, a pan-caspase protease inhibitor, has been shown to inhibit apoptosis, inflammation and fibrosis in a preclinical model of NASH. A preliminary report of a phase II clinical trial showed significantly decreased serum ALT and cCK18 levels in NAFLD patients^[206]. The therapeutic effects of this drug have also been examined in various other fibrotic liver diseases where it has been shown to reduce hepatic venous pressure gradient (HVPG). A phase II trial on NASH patients with fibrosis (ClinicalTrials.gov Identifier: NCT02686762) is ongoing to evaluate the efficacy of emricasan (10 mg and 100 mg/d for 72 wk) to improve fibrosis without worsening of NASH (primary endpoint) and to assess histological improvement or resolution of NASH (secondary endpoint).

ASK1 inhibitors/ASK1-I: Apoptosis signal regulating kinase 1/ASK1 is a MAP3 kinase (mitogen activated protein 3 kinase) which induces apoptosis and fibrosis when activated by stimuli like hyperglycaemia, TGF- β and ROS. This enzyme has been shown to be activated in patients with NASH. GS 4997, a first-in-class, oral small molecule ASK1 inhibitor, given to animals with established NASH showed a significant reduction in hepatic steatosis, fibrosis, body weight, fasting blood glucose, insulin resistance, lipogenesis, cholesterol biosynthesis, plasma AST/ALT levels, and soluble/insoluble collagen and many metabolic parameters of NASH^[207-209]. GS-4997 is currently being investigated in a phase II clinical trial of patients with NASH (ClinicalTrials.gov Identifier: NCT02466516).

p38 MAPK inhibitors: Chronic inflammation is one risk factor that contributes to progression of NAFLD. p38 mitogen activated kinases (p38 MAPK) is a stress kinase whose activation has been shown to promote inflammation^[210-212]. In mammals, four p38 MAPK isoforms have been identified: p38a, b, c and d. p38 MAPK isoforms -c and -d have recently been shown to contribute to the development of steatosis and NASH in various models of NAFLD by regulating T-cell activation, neutrophil recruitment and macrophage production of TNF- α ^[213,214]. Studies have shown higher liver expression of p38 protein in obese individuals

with steatosis. Thus, deletion of p38 α and δ in the myeloid cells prevents neutrophil migration to the liver, protecting these animals against diet induced steatosis and inflammation^[215]. Therefore, p38 MAPK can be an effective potential target for NAFLD therapy.

PPAR- α and δ agonists (Elafibranor): PPAR- α is mainly expressed in liver and is principally involved in lipid metabolism, while PPAR- δ is found in various tissues of the body and is involved in fatty acid oxidation and insulin sensitivity. In various animal models, PPAR has been shown to be hepato-protective *via* its effect on decreasing lipid accumulation, inflammation and fibrosis^[216-218]. In a RCT (clinicaltrials.gov NCT01694849), a daily dose of 80 or 120 mg Elafibranor or placebo was given to non-cirrhotic NASH patients for 52 wk^[219]. The primary endpoint of this study (i.e. resolution of NASH without worsening of fibrosis), was not met. However, it was found that patients with an initial NAS of ≥ 4 on 120 mg/d of drug showed significant improvement in hepatic inflammation. Nevertheless, Elafibranor efficacy was described as sub-optimal. Another study is in phase 3 clinical trial to clarify Elafibranor's (GFT505) effectiveness (ClinicalTrials.gov: NCT02704403) to improve the histological grade and reduce all-cause mortality and liver-related outcomes in patients with NASH and fibrosis.

Farnesoid X receptor/FXR agonists (Obeticholic acid): Obeticholic acid (OCA) is a Farnesoid X receptor agonist. It is a synthetic derivative of natural bile acid chenodeoxycholic acid (CDCA), with potency 100 times more than CDCA. Farnesoid X receptor is a nuclear hormone receptor which regulates bile, cholesterol, glucose and lipid metabolism^[220,221]. These receptors act *via* multiple pathways; they inhibit hepatic lipogenesis, gluconeogenesis, glycogenolysis and maintain cholesterol balance and improve insulin sensitivity^[222,223]. In various animal models, OCA has shown anti-inflammatory and anti-fibrotic properties and also improves insulin resistance and hepatic steatosis^[224]. In an animal model, OCA was shown to reduce hepatic inflammation and fibrosis and also decreased intrahepatic vascular resistance and improved portal hypertension^[225]. Also in an animal model with advanced cirrhosis, treatment with OCA was shown to reduce gut bacterial translocation from 78.3% to 33.3% ($P < 0.01$) indicating its effect in maintaining intestinal barrier integrity. Thus, it can be used as an option to prevent bacterial infection in such patients^[226]. In a small pilot trial of diabetic patients with NAFLD, it was shown to decrease weight and serum γ -glutamyl transferase levels as well as an improvement in liver fibrosis^[227]. The multicenter trial (FLINT trial: NCT01265498) showed a decrease in NAS, an improvement in hepatic steatosis, and a small decrease in liver fibrosis in non-cirrhotic NAFLD patients on a daily dose of 25 mg OCA compared to

placebo^[228]. A phase 3, double blind RCT multicenter study is ongoing to evaluate the safety and efficacy of OCA in NASH patients (ClinicalTrials.gov Identifier: NCT02548351). The effect of OCA on liver histology in non-cirrhotic NASH patients with stage 2 or 3 fibrosis will be compared to placebo. 2065 patients are randomized in 1:1:1 to receive 10 mg OCA, 25 mg OCA or placebo. An interim analysis is to be done at 18 mo and the study is expected to end in 6 years. However, an increase in total cholesterol and triglycerides with a decrease in high density lipoprotein was also seen in the OCA group when compared to placebo^[228]. Two phase I studies conducted in healthy individuals given OCA for 14-20 d also reported decreased HDL and increased LDL cholesterol, regardless of the dose of OCA (5, 10 or 25 mg daily)^[229,230]. These pro-atherogenic effects can be a concern for NAFLD patients that already have a high risk for cardiovascular adverse events because of dyslipidemia. Therefore, combination therapies with FXR agonist and agents that prevent atherosclerosis are warranted. Apart from OCA, various other FXR agonists such as GW4064, PX20606, GS-9674 and INT-767 are being tested. GW4064, PX20606 and GS-9674 are synthetic non-steroidal FXR agonists. INT-767 is a dual agonist for FXR and TGR5 (the transmembrane G-protein bile acid receptor) while BAR502 is a dual agonist for FXR and GPBAR1 receptors. In various animal models, these agonists have been shown to improve NASH histological features, steatosis and fibrosis^[231-234]. Thus, clinical trials are anticipated for these agents as well.

NOX-1/4 inhibitors: NADPH oxidase (NOX), is an enzyme which catalyzes the production of ROS^[235]. In various animal models these enzymes are expressed on hepatic stellate cells and promote liver fibrosis and inflammation^[236]. In a murine model, NOX 1/4 inhibitor (GKT137831) has been found to decrease ROS production and fibrotic gene expression, thus decreasing liver inflammation and fibrosis^[235]. Therefore, these agents can have a beneficial effect in decreasing liver fibrosis in NASH patients but require further studies.

Galectin-3 antagonists: Galectins are proteins that bind to terminal galactose residues on glycoproteins^[237]. They are usually expressed in immune cells and are at very low levels in the body but their levels are increased during inflammation and fibrosis^[238,239]. Galectin-3 knockout mice show reduced hepatic fibrosis after liver injury. GR-MD-02, a galectin-3 inhibitor, has shown a decrease fibrosis, hepatic steatosis and collagen deposition in various animal models with NASH^[240]. A Phase II clinical trial for evaluation of the safety and efficacy of GR-MD-02 for the treatment of liver fibrosis and associated portal hypertension in patients with NASH cirrhosis is currently underway (ClinicalTrials.gov Identifier: NCT02462967). This study has enrolled

subjects with portal hypertension and biopsy proven NASH cirrhosis (excluding subjects with medium and large varices and those with decompensated cirrhosis). The expected primary completion date is October 2017 while the study is expected to complete in February 2018.

Acetyl CoA carboxylase inhibitor: Malonyl coenzyme A plays a key role in fatty acid metabolism that maintains a balance between lipogenesis and lipid oxidation^[241]. It promotes fatty acid synthesis, and inhibits β -oxidation of lipids. Malonyl CoA is generated from acetyl CoA and the key enzyme regulating this process is acetyl CoA carboxylase (ACC). Therefore, inhibiting ACC prevents fatty acid synthesis and promotes its oxidation. In a murine model of NAFLD, inhibition of ACC has been shown to decrease hepatic steatosis, lipogenesis and increase insulin sensitivity and fatty acid oxidation^[241]. Chronic administration of ND-630 (ACC isozyme 1 and 2 inhibitor) to diet-induced obese rats and Zucker diabetic fatty rats caused a reduction in hepatic steatosis, lowered haemoglobin A1C (0.9% reduction) and improved insulin sensitivity^[242]. Also in a crossover, randomized, double-blind trial, administration of a single dose of NDI-010976 (a highly potent and selective inhibitor of both ACC1 and ACC2) to overweight/obese subjects inhibited *de novo* lipogenesis in a dose dependent manner^[243]. Together, all these results suggest its usefulness in treating metabolic syndrome, type 2 diabetes mellitus, and fatty liver disease. Thus, large long term clinical trials in humans are needed.

FGF-21 and FGF-19 analogues: FGF-21 (fibroblast growth factor 21) is a hormone which is secreted mainly from the liver. It is a starvation-induced peptide hormone with pleiotropic effects whose levels are mainly increased during fasting^[244,245]. While FGF-21 concentrations are elevated in human subjects with NAFLD, a lack of FGF-21 worsened the metabolic disorders in an animal model of NASH^[246]. Conversely, treatment with FGF-21 analogue (BMS-986036) was found to improve insulin sensitivity, hepatic steatosis and decrease lipogenesis^[247]. In another animal model of NASH, LY240531 (a FGF-21 variant) was shown to increase fatty acid oxidation by enhancing hepatic mitochondrial oxygen consumption. Also, various inflammatory markers and AST and ALT levels were reduced, suggesting an attenuation of liver injury^[248]. BMS-986036 is currently being evaluated in a phase II trial of NASH patients (ClinicalTrials.gov Identifier: NCT02413372).

FXR activation in terminal ileum by bile acid promotes FGF-19 secretion which, in turn, decreases bile acid synthesis and gluconeogenesis^[245]. It also results in the activation of the FGFR4 receptor which has a proliferative impact on hepatocytes, thus raising the potential for tumorigenesis^[249]. NGM-282, a variant of FGF-19 has been shown to decrease bile acid synthesis

and gluconeogenesis without having a tumorigenic effect^[245]. In a preliminary pre-clinical study, NGM-282 was shown to improve hepatic steatosis and histological features of NASH in an animal model^[250].

CCR2 and CCR5 inhibitor (cenicriviroc): CCR2 and CCR5 are chemokine receptors which are mainly expressed in various immune cells like monocytes, macrophages, Kupffer cells, natural killer cells, T cells and stimulate hepatic stellate cells thus promoting fibrosis. These receptors can be inhibited by cenicriviroc (CVC) which is an inhibitor of the CCR2 and CCR5 receptors. CVC has been shown to decrease fibrosis and inflammation in various animal models of diet-induced NASH or substance-induced NASH^[251-254]. There is an ongoing trial (ClinicalTrials.gov Identifier: NCT02217475) with CVC to examine its efficacy in NASH patients with fibrosis. It will compare shorter vs longer CVC treatment and assess correlations between decreased inflammation and fibrosis^[255].

SCD-1 inhibitors (aramchol): Aramchol is a synthetic lipid molecule which decreases hepatic fat accumulation by decreasing lipogenesis and increasing fatty acid oxidation by inhibiting stearoyl coenzyme A desaturase 1 (SCD1) enzyme^[256]. This drug was found to decrease liver fat content significantly in 60 NAFLD patients who were given 100 or 300 mg of this drug daily for 3 mo; the effect of the drug on fibrosis was not determined^[256]. A phase II clinical trial of this drug is ongoing on NASH patients with fibrosis (ClinicalTrials.gov Identifier: NCT02279524).

Lysyl oxidase-like 2 inhibitor (sintuzumab): Lysyl oxidase-like 2 inhibitor is an enzyme which causes cross linkage of collagen, thus preventing its degradation^[257]. This enzyme has been found to promote fibrosis in liver diseases of various etiologies. A monoclonal antibody (sintuzumab) to this enzyme has been studied in various animal models and has shown to decrease fibrosis^[258]. Two big trials are ongoing to examine the efficacy of this drug in decreasing fibrosis and preventing progression to cirrhosis in such patients (ClinicalTrials.gov Identifier: NCT01672866 and NCT01672879).

Sirtuins: Sirtuins (SIRT) are information regulator proteins. There are various types of SIRT found in mammals. SIRT-1, a member of this family of proteins, has anti-inflammatory effects and increases insulin secretion and sensitivity^[259]. A decreased liver expression of SIRT-1 was observed in an animal model of NAFLD^[260]. Since SIRT-1 activator (resveratrol) was shown to improve hepatic steatosis and insulin sensitivity^[261], SIRT-1 could be a potential target for treatment of NAFLD patients' in future clinical studies.

Betaine: A phase II clinical trial of betaine in patients with a clinical diagnosis of NAFLD is close to completion.

This trial (ClinicalTrials.gov Identifier: NCT03073343) will evaluate the effect of two doses of oral betaine in reducing ALT levels in NAFLD patients with and without diabetes.

CONCLUSION

Both ALD and NAFLD are chronic liver diseases with similar spectrums from simple steatosis to cirrhosis with basic differences only in their etiology. Despite understanding much of the pathophysiology of both diseases, there is still no effective treatment for either disease. The treatment for ALD basically relies on alcohol abstinence, nutritional support, lifestyle modifications, steroids and symptomatic treatment of complications of cirrhosis while for NAFLD, the focus of treatment is on weight loss, exercise and the use of insulin sensitizers. Removal of the cause would be the most efficient way of treating both diseases. However, the involvement of several inter-related pathways in the pathogenesis of these diseases indicates that a single therapeutic agent is unlikely to be an effective treatment strategy. Hence, a combination therapy towards multiple targets would eventually be required. Future areas of research also include the safety, efficacy, and ethical considerations of liver transplant in severe ASH for patients who are not responding to medical therapy. Various new target-oriented therapies are under investigation for both diseases and hopefully soon we will be having an effective multi-therapeutic regimen for each disease.

REFERENCES

- 1 **Rehm J**, Mathers C, Popova S, Thavorncharoensap M, Teerawattananon Y, Patra J. Global burden of disease and injury and economic cost attributable to alcohol use and alcohol-use disorders. *Lancet* 2009; **373**: 2223-2233 [PMID: 19560604 DOI: 10.1016/S0140-6736(09)60746-7]
- 2 **Younossi ZM**, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016; **64**: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- 3 **Adams LA**, Angulo P, Lindor KD. Nonalcoholic fatty liver disease. *CMAJ* 2005; **172**: 899-905 [PMID: 15795412 DOI: 10.1503/cmaj.045232]
- 4 **Ballestri S**, Zona S, Targher G, Romagnoli D, Baldelli E, Nascimbeni F, Roverato A, Guaraldi G, Lonardo A. Nonalcoholic fatty liver disease is associated with an almost twofold increased risk of incident type 2 diabetes and metabolic syndrome. Evidence from a systematic review and meta-analysis. *J Gastroenterol Hepatol* 2016; **31**: 936-944 [PMID: 26667191 DOI: 10.1111/jgh.13264]
- 5 **Lonardo A**, Ballestri S, Guaraldi G, Nascimbeni F, Romagnoli D, Zona S, Targher G. Fatty liver is associated with an increased risk of diabetes and cardiovascular disease - Evidence from three different disease models: NAFLD, HCV and HIV. *World J Gastroenterol* 2016; **22**: 9674-9693 [PMID: 27956792 DOI: 10.3748/wjg.v22.i44.9674]
- 6 **Flegal KM**, Carroll MD, Ogden CL, Curtin LR. Prevalence and trends in obesity among US adults, 1999-2008. *JAMA* 2010; **303**: 235-241 [PMID: 20071471 DOI: 10.1001/jama.2009.2014]
- 7 **Rowell RJ**, Anstee QM. An overview of the genetics, mechanisms and management of NAFLD and ALD. *Clin Med (Lond)* 2015; **15** Suppl 6: s77-s82 [PMID: 26634687 DOI: 10.7861/clinmedicine.15-6-s77]
- 8 **Poynard T**, Mathurin P, Lai CL, Guyader D, Poupon R, Tainturier MH, Myers RP, Muntenau M, Ratzu V, Manns M, Vogel A, Capron F, Chedid A, Bedossa P; PANFIBROSIS Group. A comparison of fibrosis progression in chronic liver diseases. *J Hepatol* 2003; **38**: 257-265 [PMID: 12586290]
- 9 **Anstee QM**, Day CP. The Genetics of Nonalcoholic Fatty Liver Disease: Spotlight on PNPLA3 and TM6SF2. *Semin Liver Dis* 2015; **35**: 270-290 [PMID: 26378644 DOI: 10.1055/s-0035-1562947]
- 10 **Chamorro AJ**, Torres JL, Mirón-Canelo JA, González-Sarmiento R, Laso FJ, Marcos M. Systematic review with meta-analysis: the I148M variant of patatin-like phospholipase domain-containing 3 gene (PNPLA3) is significantly associated with alcoholic liver cirrhosis. *Aliment Pharmacol Ther* 2014; **40**: 571-581 [PMID: 25060292 DOI: 10.1111/apt.12890]
- 11 **Sookoian S**, Pirola CJ. Meta-analysis of the influence of I148M variant of patatin-like phospholipase domain containing 3 gene (PNPLA3) on the susceptibility and histological severity of nonalcoholic fatty liver disease. *Hepatology* 2011; **53**: 1883-1894 [PMID: 21381068 DOI: 10.1002/hep.24283]
- 12 **Trépo E**, Nahon P, Bontempi G, Valenti L, Falletti E, Nischalke HD, Hamza S, Corradini SG, Burza MA, Guyot E, Donati B, Spengler U, Hillon P, Toniutto P, Henrion J, Franchimont D, Devière J, Mathurin P, Moreno C, Romeo S, Deltenre P. Association between the PNPLA3 (rs738409 C>G) variant and hepatocellular carcinoma: Evidence from a meta-analysis of individual participant data. *Hepatology* 2014; **59**: 2170-2177 [PMID: 24114809 DOI: 10.1002/hep.26767]
- 13 **Mathurin P**, O'Grady J, Carithers RL, Phillips M, Louvet A, Mendenhall CL, Ramond MJ, Naveau S, Maddrey WC, Morgan TR. Corticosteroids improve short-term survival in patients with severe alcoholic hepatitis: meta-analysis of individual patient data. *Gut* 2011; **60**: 255-260 [PMID: 20940288 DOI: 10.1136/gut.2010.224097]
- 14 **Fialla AD**, Israelsen M, Hamberg O, Krag A, Gluud LL. Nutritional therapy in cirrhosis or alcoholic hepatitis: a systematic review and meta-analysis. *Liver Int* 2015; **35**: 2072-2078 [PMID: 25645300 DOI: 10.1111/liv.12798]
- 15 **Lucey MR**, Mathurin P, Morgan TR. Alcoholic hepatitis. *N Engl J Med* 2009; **360**: 2758-2769 [PMID: 19553649 DOI: 10.1056/NEJMra0805786]
- 16 **Musso G**, Gambino R, Cassader M, Pagano G. A meta-analysis of randomized trials for the treatment of nonalcoholic fatty liver disease. *Hepatology* 2010; **52**: 79-104 [PMID: 20578268 DOI: 10.1002/hep.23623]
- 17 **Chalasani N**, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, Charlton M, Sanyal AJ; American Association for the Study of Liver Diseases; American College of Gastroenterology; American Gastroenterological Association. The diagnosis and management of non-alcoholic fatty liver disease: Practice guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. *Am J Gastroenterol* 2012; **107**: 811-826 [PMID: 22641309 DOI: 10.1038/ajg.2012.128]
- 18 **Mayo-Smith MF**, Beecher LH, Fischer TL, Gorelick DA, Guillaume JL, Hill A, Jara G, Kasser C, Melbourne J; Working Group on the Management of Alcohol Withdrawal Delirium, Practice Guidelines Committee, American Society of Addiction Medicine. Management of alcohol withdrawal delirium. An evidence-based practice guideline. *Arch Intern Med* 2004; **164**: 1405-1412 [PMID: 15249349 DOI: 10.1001/archinte.164.13.1405]
- 19 **Day E**, Benthall P, Callaghan R, Kuruvilla T, George S. Thiamine for Wernicke-Korsakoff Syndrome in people at risk from alcohol abuse. *Cochrane Database Syst Rev* 2004; **(1)**: CD004033 [PMID: 14974055 DOI: 10.1002/14651858.CD004033.pub2]
- 20 **Donnino MW**, Vega J, Miller J, Walsh M. Myths and misconceptions of Wernicke's encephalopathy: what every emergency physician should know. *Ann Emerg Med* 2007; **50**: 715-721 [PMID: 17681641 DOI: 10.1016/j.annemergmed.2007.02.

- 007]
- 21 **Thomson AD.** Mechanisms of vitamin deficiency in chronic alcohol misusers and the development of the Wernicke-Korsakoff syndrome. *Alcohol Alcohol Suppl* 2000; **35**: 2-7 [PMID: 11304071]
 - 22 **Isenberg-Grzeda E, Kutner HE, Nicolson SE.** Wernicke-Korsakoff-syndrome: under-recognized and under-treated. *Psychosomatics* 2012; **53**: 507-516 [PMID: 23157990 DOI: 10.1016/j.psych.2012.04.008]
 - 23 **Cook CC, Hallwood PM, Thomson AD.** B Vitamin deficiency and neuropsychiatric syndromes in alcohol misuse. *Alcohol Alcohol* 1998; **33**: 317-336 [PMID: 9719389]
 - 24 **Galvin R, Bråthen G, Ivashynka A, Hillbom M, Tanasescu R, Leone MA; EFNS.** EFNS guidelines for diagnosis, therapy and prevention of Wernicke encephalopathy. *Eur J Neurol* 2010; **17**: 1408-1418 [PMID: 20642790 DOI: 10.1111/j.1468-1331.2010.03153.x]
 - 25 **Alcohol-Use Disorders: Diagnosis, Assessment and Management of Harmful Drinking and Alcohol Dependence.** Leicester (UK), 2011
 - 26 **Nishimoto A, Usery J, Winton JC, Twilla J.** High-dose Parenteral Thiamine in Treatment of Wernicke's Encephalopathy: Case Series and Review of the Literature. *In Vivo* 2017; **31**: 121-124 [PMID: 28064230 DOI: 10.21873/invivo.11034]
 - 27 **Thomson AD, Guerrini I, Marshall EJ.** The evolution and treatment of Korsakoff's syndrome: out of sight, out of mind? *Neuropsychol Rev* 2012; **22**: 81-92 [PMID: 22569770 DOI: 10.1007/s11065-012-9196-z]
 - 28 **Moos RH, King MJ, Patterson MA.** Outcomes of residential treatment of substance abuse in hospital- and community-based programs. *Psychiatr Serv* 1996; **47**: 68-74 [PMID: 8925349 DOI: 10.1176/ps.47.1.68]
 - 29 **Roozen HG, de Waart R, van der Windt DA, van den Brink W, de Jong CA, Kerkhof AJ.** A systematic review of the effectiveness of naltrexone in the maintenance treatment of opioid and alcohol dependence. *Eur Neuropsychopharmacol* 2006; **16**: 311-323 [PMID: 16361086 DOI: 10.1016/j.euroneuro.2005.11.001]
 - 30 **Mason BJ, Leher P.** Acamprosate for alcohol dependence: a sex-specific meta-analysis based on individual patient data. *Alcohol Clin Exp Res* 2012; **36**: 497-508 [PMID: 21895717 DOI: 10.1111/j.1530-0277.2011.01616.x]
 - 31 **Kenna GA, Lomastro TL, Schiesl A, Leggio L, Swift RM.** Review of topiramate: an antiepileptic for the treatment of alcohol dependence. *Curr Drug Abuse Rev* 2009; **2**: 135-142 [PMID: 19630744]
 - 32 **Fuller RK, Branchey L, Brightwell DR, Derman RM, Emrick CD, Iber FL, James KE, Lacoursiere RB, Lee KK, Lowenstam I.** Disulfiram treatment of alcoholism. A Veterans Administration cooperative study. *JAMA* 1986; **256**: 1449-1455 [PMID: 3528541]
 - 33 **Addolorato G, Leggio L, Ferrulli A, Cardone S, Vonghia L, Mirijello A, Abenavoli L, D'Angelo C, Caputo F, Zambon A, Haber PS, Gasbarrini G.** Effectiveness and safety of baclofen for maintenance of alcohol abstinence in alcohol-dependent patients with liver cirrhosis: randomised, double-blind controlled study. *Lancet* 2007; **370**: 1915-1922 [PMID: 18068515 DOI: 10.1016/S0140-6736(07)61814-5]
 - 34 **Addolorato G, Mirijello A, Leggio L, Ferrulli A, Landolfi R.** Management of alcohol dependence in patients with liver disease. *CNS Drugs* 2013; **27**: 287-299 [PMID: 23456576 DOI: 10.1007/s40263-013-0043-4]
 - 35 **Addolorato G, Leggio L, Ferrulli A, Cardone S, Bedogni G, Caputo F, Gasbarrini G, Landolfi R; Baclofen Study Group.** Dose-response effect of baclofen in reducing daily alcohol intake in alcohol dependence: secondary analysis of a randomized, double-blind, placebo-controlled trial. *Alcohol Alcohol* 2011; **46**: 312-317 [PMID: 21414953 DOI: 10.1093/alcacr/agr017]
 - 36 **Addolorato G, Russell M, Albano E, Haber PS, Wands JR, Leggio L.** Understanding and treating patients with alcoholic cirrhosis: an update. *Alcohol Clin Exp Res* 2009; **33**: 1136-1144 [PMID: 19389182 DOI: 10.1111/j.1530-0277.2009.00956.x]
 - 37 **Kershenovich D, Corona DL, Kershenovich R, Gutierrez-Reyes G.** Management of alcoholic liver disease: an update. *Alcohol Clin Exp Res* 2011; **35**: 804-805 [PMID: 21284670 DOI: 10.1111/j.1530-0277.2010.01402.x]
 - 38 **Haass-Koffler CL, Leggio L, Kenna GA.** Pharmacological approaches to reducing craving in patients with alcohol use disorders. *CNS Drugs* 2014; **28**: 343-360 [PMID: 24573997 DOI: 10.1007/s40263-014-0149-3]
 - 39 **Vuittonet CL, Halse M, Leggio L, Fricchione SB, Brickley M, Haass-Koffler CL, Tavares T, Swift RM, Kenna GA.** Pharmacotherapy for alcoholic patients with alcoholic liver disease. *Am J Health Syst Pharm* 2014; **71**: 1265-1276 [PMID: 25027533 DOI: 10.2146/ajhp140028]
 - 40 **Caballería J, Parés A, Brú C, Mercader J, García Plaza A, Caballería L, Clemente G, Rodrigo L, Rodés J.** Metadoxine accelerates fatty liver recovery in alcoholic patients: results of a randomized double-blind, placebo-control trial. Spanish Group for the Study of Alcoholic Fatty Liver. *J Hepatol* 1998; **28**: 54-60 [PMID: 9537864]
 - 41 **Higuera-de la Tijera F, Servín-Caamaño AI, Serralde-Zúñiga AE, Cruz-Herrera J, Pérez-Torres E, Abdo-Francis JM, Salas-Gordillo F, Pérez-Hernández JL.** Metadoxine improves the three- and six-month survival rates in patients with severe alcoholic hepatitis. *World J Gastroenterol* 2015; **21**: 4975-4985 [PMID: 25945012 DOI: 10.3748/wjg.v21.i16.4975]
 - 42 **Naveau S, Giraud V, Borotto E, Aubert A, Capron F, Chaput JC.** Excess weight risk factor for alcoholic liver disease. *Hepatology* 1997; **25**: 108-111 [PMID: 8985274 DOI: 10.1002/hep.510250120]
 - 43 **Klatsky AL, Armstrong MA.** Alcohol, smoking, coffee, and cirrhosis. *Am J Epidemiol* 1992; **136**: 1248-1257 [PMID: 1476147]
 - 44 **Donato F, Tagger A, Chiesa R, Ribero ML, Tomasoni V, Fasola M, Gelatti U, Portera G, Boffetta P, Nardi G.** Hepatitis B and C virus infection, alcohol drinking, and hepatocellular carcinoma: a case-control study in Italy. Brescia HCC Study. *Hepatology* 1997; **26**: 579-584 [PMID: 9303486 DOI: 10.1002/hep.510260308]
 - 45 **Hutchinson SJ, Bird SM, Goldberg DJ.** Influence of alcohol on the progression of hepatitis C virus infection: a meta-analysis. *Clin Gastroenterol Hepatol* 2005; **3**: 1150-1159 [PMID: 16271348]
 - 46 **Befrits R, Hedman M, Blomquist L, Allander T, Grillner L, Kinnman N, Rubio C, Hultcrantz R.** Chronic hepatitis C in alcoholic patients: prevalence, genotypes, and correlation to liver disease. *Scand J Gastroenterol* 1995; **30**: 1113-1118 [PMID: 8578173]
 - 47 **Chen CM, Yoon YH, Yi HY, Lucas DL.** Alcohol and hepatitis C mortality among males and females in the United States: a life table analysis. *Alcohol Clin Exp Res* 2007; **31**: 285-292 [PMID: 17250621 DOI: 10.1111/j.1530-0277.2006.00304.x]
 - 48 **Harris DR, Gonin R, Alter HJ, Wright EC, Buskell ZJ, Hollinger FB, Seeff LB; National Heart, Lung, and Blood Institute Study Group.** The relationship of acute transfusion-associated hepatitis to the development of cirrhosis in the presence of alcohol abuse. *Ann Intern Med* 2001; **134**: 120-124 [PMID: 11177315]
 - 49 **Henry JA, Moloney C, Rivas C, Goldin RD.** Increase in alcohol related deaths: is hepatitis C a factor? *J Clin Pathol* 2002; **55**: 704-707 [PMID: 12195003]
 - 50 **Tsui JI, Pletcher MJ, Vittinghoff E, Seal K, Gonzales R.** Hepatitis C and hospital outcomes in patients admitted with alcohol-related problems. *J Hepatol* 2006; **44**: 262-266 [PMID: 16226823 DOI: 10.1016/j.jhep.2005.07.027]
 - 51 **Novo-Veleiro I, Alvela-Suárez L, Chamorro AJ, González-Sarmiento R, Laso FJ, Marcos M.** Alcoholic liver disease and hepatitis C virus infection. *World J Gastroenterol* 2016; **22**: 1411-1420 [PMID: 26819510 DOI: 10.3748/wjg.v22.i4.1411]
 - 52 **Mendenhall C, Roselle GA, Gartside P, Moritz T.** Relationship of protein calorie malnutrition to alcoholic liver disease: a reexamination of data from two Veterans Administration Cooperative Studies. *Alcohol Clin Exp Res* 1995; **19**: 635-641 [PMID: 7573786]
 - 53 **Huisman EJ, Trip EJ, Siersema PD, van Hoek B, van Erpecum KJ.** Protein energy malnutrition predicts complications in liver cirrhosis. *Eur J Gastroenterol Hepatol* 2011; **23**: 982-989 [PMID:

- 21971339 DOI: 10.1097/MEG.0b013e32834aa4bb]
- 54 **Fragasso A**, Mannarella C, Ciancio A, Scarciolla O, Nuzzolese N, Clemente R, Vitullo E, Sacco A. Holotranscobalamin is a useful marker of vitamin B12 deficiency in alcoholics. *ScientificWorldJournal* 2012; **2012**: 128182 [PMID: 22481895 DOI: 10.1100/2012/128182]
 - 55 **Rossi RE**, Conte D, Massironi S. Diagnosis and treatment of nutritional deficiencies in alcoholic liver disease: Overview of available evidence and open issues. *Dig Liver Dis* 2015; **47**: 819-825 [PMID: 26164399 DOI: 10.1016/j.dld.2015.05.021]
 - 56 **McClain CJ**, Barve SS, Barve A, Marsano L. Alcoholic liver disease and malnutrition. *Alcohol Clin Exp Res* 2011; **35**: 815-820 [PMID: 21284673 DOI: 10.1111/j.1530-0277.2010.01405.x]
 - 57 **Halsted CH**. Nutrition and alcoholic liver disease. *Semin Liver Dis* 2004; **24**: 289-304 [PMID: 15349806 DOI: 10.1055/s-2004-832941]
 - 58 **Kang YJ**, Zhou Z. Zinc prevention and treatment of alcoholic liver disease. *Mol Aspects Med* 2005; **26**: 391-404 [PMID: 16099027 DOI: 10.1016/j.mam.2005.07.002]
 - 59 **Henkel AS**, Buchman AL. Nutritional support in patients with chronic liver disease. *Nat Clin Pract Gastroenterol Hepatol* 2006; **3**: 202-209 [PMID: 16582962 DOI: 10.1038/ncpgasthep0443]
 - 60 **Cabr   E**, Rodr  guez-Iglesias P, Caballer   J, Quer JC, S  nchez-Lombr  a JL, Par  s A, Papo M, Planas R, Gassull MA. Short- and long-term outcome of severe alcohol-induced hepatitis treated with steroids or enteral nutrition: a multicenter randomized trial. *Hepatology* 2000; **32**: 36-42 [PMID: 10869286 DOI: 10.1053/jhep.2000.8627]
 - 61 **Plauth M**, Cabr   E, Riggio O, Assis-Camilo M, Pirlich M, Kondrup J, DGEM (German Society for Nutritional Medicine), Ferenci P, Holm E, Vom Dahl S, M  ller MJ, Nolte W; ESPEN (European Society for Parenteral and Enteral Nutrition). ESPEN Guidelines on Enteral Nutrition: Liver disease. *Clin Nutr* 2006; **25**: 285-294 [PMID: 16707194 DOI: 10.1016/j.clnu.2006.01.018]
 - 62 **Forrest E**, Mellor J, Stanton L, Bowers M, Ryder P, Austin A, Day C, Gleeson D, O'Grady J, Masson S, McCune A, Patch D, Richardson P, Roderick P, Ryder S, Wright M, Thursz M. Steroids or pentoxifylline for alcoholic hepatitis (STOPAH): study protocol for a randomised controlled trial. *Trials* 2013; **14**: 262 [PMID: 23958271 DOI: 10.1186/1745-6215-14-262]
 - 63 **Rambaldi A**, Saconato HH, Christensen E, Thorlund K, Wetterslev J, Gluud C. Systematic review: glucocorticosteroids for alcoholic hepatitis--a Cochrane Hepato-Biliary Group systematic review with meta-analyses and trial sequential analyses of randomized clinical trials. *Aliment Pharmacol Ther* 2008; **27**: 1167-1178 [PMID: 18363896 DOI: 10.1111/j.1365-2036.2008.03685.x]
 - 64 **Mathurin P**, Mendenhall CL, Carithers RL Jr, Ramond MJ, Maddrey WC, Garstide P, Rueff B, Naveau S, Chaput JC, Poynard T. Corticosteroids improve short-term survival in patients with severe alcoholic hepatitis (AH): individual data analysis of the last three randomized placebo controlled double blind trials of corticosteroids in severe AH. *J Hepatol* 2002; **36**: 480-487 [PMID: 11943418]
 - 65 **Mathurin P**, Abdelnour M, Ramond MJ, Carbonell N, Fartoux L, Serfaty L, Valla D, Poupon R, Chaput JC, Naveau S. Early change in bilirubin levels is an important prognostic factor in severe alcoholic hepatitis treated with prednisolone. *Hepatology* 2003; **38**: 1363-1369 [PMID: 14647046 DOI: 10.1016/j.hep.2003.09.038]
 - 66 **Louvet A**, Naveau S, Abdelnour M, Ramond MJ, Diaz E, Fartoux L, Dharancy S, Texier F, Hollebecque A, Serfaty L, Boleslawski E, Deltenre P, Canva V, Pruvot FR, Mathurin P. The Lille model: a new tool for therapeutic strategy in patients with severe alcoholic hepatitis treated with steroids. *Hepatology* 2007; **45**: 1348-1354 [PMID: 17518367 DOI: 10.1002/hep.21607]
 - 67 **Depew W**, Boyer T, Omata M, Redeker A, Reynolds T. Double-blind controlled trial of prednisolone therapy in patients with severe acute alcoholic hepatitis and spontaneous encephalopathy. *Gastroenterology* 1980; **78**: 524-529 [PMID: 6985881]
 - 68 **Singal AK**, Walia I, Singal A, Soloway RD. Corticosteroids and pentoxifylline for the treatment of alcoholic hepatitis: Current status. *World J Hepatol* 2011; **3**: 205-210 [PMID: 21954408 DOI: 10.4254/wjh.v3.i8.205]
 - 69 **Saberi B**, Dadabhai AS, Jang YY, Gurakar A, Mezey E. Current Management of Alcoholic Hepatitis and Future Therapies. *J Clin Transl Hepatol* 2016; **4**: 113-122 [PMID: 27350941 DOI: 10.14218/JCTH.2016.00006]
 - 70 **Louvet A**, Wartel F, Castel H, Dharancy S, Hollebecque A, Canva-Delcambre V, Deltenre P, Mathurin P. Infection in patients with severe alcoholic hepatitis treated with steroids: early response to therapy is the key factor. *Gastroenterology* 2009; **137**: 541-548 [PMID: 19445945 DOI: 10.1053/j.gastro.2009.04.062]
 - 71 **O'Shea RS**, Dasarathy S, McCullough AJ; Practice Guideline Committee of the American Association for the Study of Liver Diseases; Practice Parameters Committee of the American College of Gastroenterology. Alcoholic liver disease. *Hepatology* 2010; **51**: 307-328 [PMID: 20034030 DOI: 10.1002/hep.23258]
 - 72 **Raetsch C**, Jia JD, Boigk G, Bauer M, Hahn EG, Riecken EO, Schuppan D. Pentoxifylline downregulates profibrogenic cytokines and procollagen I expression in rat secondary biliary fibrosis. *Gut* 2002; **50**: 241-247 [PMID: 11788567]
 - 73 **Assimakopoulos SF**, Thomopoulos KC, Labropoulou-Karatza C. Pentoxifylline: a first line treatment option for severe alcoholic hepatitis and hepatorenal syndrome? *World J Gastroenterol* 2009; **15**: 3194-3195 [PMID: 19575503 DOI: 10.3748/wjg.15.3194]
 - 74 **Runyon BA**, Antillon MR. Ascitic fluid pH and lactate: insensitive and nonspecific tests in detecting ascitic fluid infection. *Hepatology* 1991; **13**: 929-935 [PMID: 2029997]
 - 75 **Akriviadis E**, Botla R, Briggs W, Han S, Reynolds T, Shakil O. Pentoxifylline improves short-term survival in severe acute alcoholic hepatitis: a double-blind, placebo-controlled trial. *Gastroenterology* 2000; **119**: 1637-1648 [PMID: 11113085]
 - 76 **De BK**, Gangopadhyay S, Dutta D, Bakshi SD, Pani A, Ghosh P. Pentoxifylline versus prednisolone for severe alcoholic hepatitis: a randomized controlled trial. *World J Gastroenterol* 2009; **15**: 1613-1619 [PMID: 19340904 DOI: 10.3748/wjg.15.1613]
 - 77 **Mathurin P**, Louvet A, Duhamel A, Nahon P, Carbonell N, Boursier J, Anty R, Diaz E, Thabut D, Moirand R, Lebre D, Moreno C, Talbodec N, Paupard T, Naveau S, Silvain C, Pageaux GP, Sobesky R, Canva-Delcambre V, Dharancy S, Salleron J, Dao T. Prednisolone with vs without pentoxifylline and survival of patients with severe alcoholic hepatitis: a randomized clinical trial. *JAMA* 2013; **310**: 1033-1041 [PMID: 24026598 DOI: 10.1001/jama.2013.276300]
 - 78 **Sidhu SS**, Goyal O, Singla P, Gupta D, Sood A, Chhina RS, Soni RK. Corticosteroid plus pentoxifylline is not better than corticosteroid alone for improving survival in severe alcoholic hepatitis (COPE trial). *Dig Dis Sci* 2012; **57**: 1664-1671 [PMID: 22388710 DOI: 10.1007/s10620-012-2097-4]
 - 79 **Im GY**, Lucey MR. Practical Concerns and Controversies in the Management of Alcoholic Hepatitis. *Gastroenterol Hepatol (N Y)* 2016; **12**: 478-489 [PMID: 27917083]
 - 80 **Thursz MR**, Richardson P, Allison M, Austin A, Bowers M, Day CP, Downs N, Gleeson D, MacGilchrist A, Grant A, Hood S, Masson S, McCune A, Mellor J, O'Grady J, Patch D, Ratcliffe I, Roderick P, Stanton L, Vergis N, Wright M, Ryder S, Forrest EH; STOPAH Trial. Prednisolone or pentoxifylline for alcoholic hepatitis. *N Engl J Med* 2015; **372**: 1619-1628 [PMID: 25901427 DOI: 10.1056/NEJMoa1412278]
 - 81 **McClain CJ**, Hill DB, Barve SS. Infliximab and prednisolone: too much of a good thing? *Hepatology* 2004; **39**: 1488-1490 [PMID: 15185287 DOI: 10.1002/hep.20267]
 - 82 **Felver ME**, Mezey E, McGuire M, Mitchell MC, Herlong HF, Veech GA, Veech RL. Plasma tumor necrosis factor alpha predicts decreased long-term survival in severe alcoholic hepatitis. *Alcohol Clin Exp Res* 1990; **14**: 255-259 [PMID: 2190492]
 - 83 **Yin M**, Wheeler MD, Kono H, Bradford BU, Gallucci RM, Luster MI, Thurman RG. Essential role of tumor necrosis factor alpha in alcohol-induced liver injury in mice. *Gastroenterology* 1999; **117**: 942-952 [PMID: 10500078]
 - 84 **Naveau S**, Chollet-Martin S, Dharancy S, Mathurin P, Jouet P,

- Piquet MA, Davion T, Oberti F, Broët P, Emilie D, Foie-Alcool group of the Association Française pour l'Etude du Foie. A double-blind randomized controlled trial of infliximab associated with prednisolone in acute alcoholic hepatitis. *Hepatology* 2004; **39**: 1390-1397 [PMID: 15122768 DOI: 10.1002/hep.20206]
- 85 Ali T, Kaitha S, Mahmood S, Ftesi A, Stone J, Bronze MS. Clinical use of anti-TNF therapy and increased risk of infections. *Drug Healthc Patient Saf* 2013; **5**: 79-99 [PMID: 23569399 DOI: 10.2147/DHPS.S28801]
 - 86 Dey A, Cederbaum AI. Alcohol and oxidative liver injury. *Hepatology* 2006; **43**: S63-S74 [PMID: 16447273 DOI: 10.1002/hep.20957]
 - 87 Stewart S, Prince M, Bassendine M, Hudson M, James O, Jones D, Record C, Day CP. A randomized trial of antioxidant therapy alone or with corticosteroids in acute alcoholic hepatitis. *J Hepatol* 2007; **47**: 277-283 [PMID: 17532088 DOI: 10.1016/j.jhep.2007.03.027]
 - 88 Nguyen TH, Jacobs P, Hanrahan A, Fraser-Lee N, Wong W, Lee B, Ohinmaa A. Health care costs of persons with newly diagnosed hepatitis C virus: a population-based, observational study. *J Viral Hepat* 2008; **15**: 634-640 [PMID: 18435719 DOI: 10.1111/j.1365-2893.2008.00985.x]
 - 89 Testino G, Sumberaz A, Borro P. Comment to "liver transplantation for patients with alcoholic liver disease: an open question". *Dig Liver Dis* 2013; **45**: 80-81 [PMID: 22770950 DOI: 10.1016/j.dld.2012.06.003]
 - 90 Testino G, Ferro C, Sumberaz A, Messa P, Morelli N, Guadagni B, Ardizzone G, Valente U. Type-2 hepatorenal syndrome and refractory ascites: role of transjugular intrahepatic portosystemic stent-shunt in eighteen patients with advanced cirrhosis awaiting orthotopic liver transplantation. *Hepatogastroenterology* 2003; **50**: 1753-1755 [PMID: 14696397]
 - 91 Tan HH, Virmani S, Martin P. Controversies in the management of alcoholic liver disease. *Mt Sinai J Med* 2009; **76**: 484-498 [PMID: 19787655 DOI: 10.1002/msj.20135]
 - 92 Burra P, Mioni D, Cecchetto A, Cillo U, Zanusi G, Fagioli S, Naccarato R, Martinez D. Histological features after liver transplantation in alcoholic cirrhotics. *J Hepatol* 2001; **34**: 716-722 [PMID: 11434618]
 - 93 Pageaux GP, Bismuth M, Perny P, Costes V, Jaber S, Possoz P, Fabre JM, Navarro F, Blanc P, Domergue J, Eledjam JJ, Larrey D. Alcohol relapse after liver transplantation for alcoholic liver disease: does it matter? *J Hepatol* 2003; **38**: 629-634 [PMID: 12713874]
 - 94 Dew MA, DiMartini AF, Steel J, De Vito Dabbs A, Myaskovsky L, Unruh M, Greenhouse J. Meta-analysis of risk for relapse to substance use after transplantation of the liver or other solid organs. *Liver Transpl* 2008; **14**: 159-172 [PMID: 18236389 DOI: 10.1002/lt.21278]
 - 95 Testino G, Burra P, Bonino F, Piani F, Sumberaz A, Peressutti R, Giannelli Castiglione A, Patussi V, Fanucchi T, Ancarani O, De Cerce G, Iannini AT, Greco G, Mosti A, Durante M, Babocchi P, Quartini M, Mioni D, Aricò S, Basile A, Leone S, Lozer F, Scafato E, Borro P, Group of Italian Regions. Acute alcoholic hepatitis, end stage alcoholic liver disease and liver transplantation: an Italian position statement. *World J Gastroenterol* 2014; **20**: 14642-14651 [PMID: 25356027 DOI: 10.3748/wjg.v20.i40.14642]
 - 96 Lucey MR, Brown KA, Everson GT, Fung JJ, Gish R, Keeffe EB, Kneteman NM, Lake JR, Martin P, McDiarmid SV, Rakela J, Shiffman ML, So SK, Wiesner RH. Minimal criteria for placement of adults on the liver transplant waiting list: a report of a national conference organized by the American Society of Transplant Physicians and the American Association for the Study of Liver Diseases. *Liver Transpl Surg* 1997; **3**: 628-637 [PMID: 9404965]
 - 97 Veldt BJ, Lainé F, Guillygomarc'h A, Lauvin L, Boudjema K, Messner M, Brissot P, Deugnier Y, Moirand R. Indication of liver transplantation in severe alcoholic liver cirrhosis: quantitative evaluation and optimal timing. *J Hepatol* 2002; **36**: 93-98 [PMID: 11804670]
 - 98 Mathurin P, Moreno C, Samuel D, Dumortier J, Salleron J, Durand F, Castel H, Duhamel A, Pageaux GP, Leroy V, Dharancy S, Louvet A, Boleslawski E, Lucidi V, Gustot T, Francoz C, Letoublon C, Castaing D, Belghiti J, Donckier V, Pruvot FR, Duclos-Vallée JC. Early liver transplantation for severe alcoholic hepatitis. *N Engl J Med* 2011; **365**: 1790-1800 [PMID: 22070476 DOI: 10.1056/NEJMoa1105703]
 - 99 Herrero JI. De novo malignancies following liver transplantation: impact and recommendations. *Liver Transpl* 2009; **15** Suppl 2: S90-S94 [PMID: 19877025 DOI: 10.1002/lt.21898]
 - 100 Dumortier J, Guillaud O, Adham M, Boucaud C, Delafosse B, Bouffard Y, Paliard P, Scoazec JY, Boillot O. Negative impact of de novo malignancies rather than alcohol relapse on survival after liver transplantation for alcoholic cirrhosis: a retrospective analysis of 305 patients in a single center. *Am J Gastroenterol* 2007; **102**: 1032-1041 [PMID: 17313502 DOI: 10.1111/j.1572-0241.2007.01079.x]
 - 101 Burra P, Senzolo M, Adam R, Delvart V, Karam V, Germani G, Neuberger J, ELITA; ELTR Liver Transplant Centers. Liver transplantation for alcoholic liver disease in Europe: a study from the ELTR (European Liver Transplant Registry). *Am J Transplant* 2010; **10**: 138-148 [PMID: 19951276 DOI: 10.1111/j.1600-6143.2009.02869.x]
 - 102 Seo YS, Shah VH. The role of gut-liver axis in the pathogenesis of liver cirrhosis and portal hypertension. *Clin Mol Hepatol* 2012; **18**: 337-346 [PMID: 23323248 DOI: 10.3350/cmh.2012.18.4.337]
 - 103 Zhao HY, Wang HJ, Lu Z, Xu SZ. Intestinal microflora in patients with liver cirrhosis. *Chin J Dig Dis* 2004; **5**: 64-67 [PMID: 15612659]
 - 104 Stadlbauer V, Mookerjee RP, Hodges S, Wright GA, Davies NA, Jalan R. Effect of probiotic treatment on deranged neutrophil function and cytokine responses in patients with compensated alcoholic cirrhosis. *J Hepatol* 2008; **48**: 945-951 [PMID: 18433921 DOI: 10.1016/j.jhep.2008.02.015]
 - 105 Kirpich IA, Solovieva NV, Leikhter SN, Shidakova NA, Lebedeva OV, Sidorov PI, Bazhukova TA, Soloviev AG, Barve SS, McClain CJ, Cave M. Probiotics restore bowel flora and improve liver enzymes in human alcohol-induced liver injury: a pilot study. *Alcohol* 2008; **42**: 675-682 [PMID: 19038698 DOI: 10.1016/j.alcohol.2008.08.006]
 - 106 Vlachogiannakos J, Saveriadis AS, Viazis N, Theodoropoulos I, Foudoulis K, Manolakopoulos S, Raptis S, Karamanolis DG. Intestinal decontamination improves liver haemodynamics in patients with alcohol-related decompensated cirrhosis. *Aliment Pharmacol Ther* 2009; **29**: 992-999 [PMID: 19210289 DOI: 10.1111/j.1365-2036.2009.03958.x]
 - 107 Michelena J, Altamirano J, Abalde JG, Affò S, Morales-Ibanez O, Sancho-Bru P, Dominguez M, García-Pagán JC, Fernández J, Arroyo V, Ginès P, Louvet A, Mathurin P, Mehal WZ, Caballeria J, Bataller R. Systemic inflammatory response and serum lipopolysaccharide levels predict multiple organ failure and death in alcoholic hepatitis. *Hepatology* 2015; **62**: 762-772 [PMID: 25761863 DOI: 10.1002/hep.27779]
 - 108 Lieber CS. S-Adenosyl-L-methionine and alcoholic liver disease in animal models: implications for early intervention in human beings. *Alcohol* 2002; **27**: 173-177 [PMID: 12163146]
 - 109 Karaa A, Thompson KJ, McKillop IH, Clemens MG, Schrum LW. S-adenosyl-L-methionine attenuates oxidative stress and hepatic stellate cell activation in an ethanol-LPS-induced fibrotic rat model. *Shock* 2008; **30**: 197-205 [PMID: 18180699 DOI: 10.1097/shk.0b013e318160f417]
 - 110 Rambaldi A, Glud C. S-adenosyl-L-methionine for alcoholic liver diseases. *Cochrane Database Syst Rev* 2006; **(2)**: CD002235 [PMID: 16625556 DOI: 10.1002/14651858.CD002235.pub2]
 - 111 Kharbanda KK. Alcoholic liver disease and methionine metabolism. *Semin Liver Dis* 2009; **29**: 155-165 [PMID: 19387915 DOI: 10.1055/s-0029-1214371]
 - 112 Purohit V, Abdelmalek MF, Barve S, Benevenga NJ, Halsted CH, Kaplowitz N, Kharbanda KK, Liu QY, Lu SC, McClain CJ, Swanson C, Zakhari S. Role of S-adenosylmethionine, folate, and betaine in the treatment of alcoholic liver disease: summary of a symposium. *Am J Clin Nutr* 2007; **86**: 14-24 [PMID: 17616758]

- 113 **Kharbanda KK**, Mailliard ME, Baldwin CR, Beckenhauer HC, Sorrell MF, Tuma DJ. Betaine attenuates alcoholic steatosis by restoring phosphatidylcholine generation via the phosphatidylethanolamine methyltransferase pathway. *J Hepatol* 2007; **46**: 314-321 [PMID: 17156888 DOI: 10.1016/j.jhep.2006.08.024]
- 114 **Kim SJ**, Jung YS, Kwon DY, Kim YC. Alleviation of acute ethanol-induced liver injury and impaired metabolomics of S-containing substances by betaine supplementation. *Biochem Biophys Res Commun* 2008; **368**: 893-898 [PMID: 18267108 DOI: 10.1016/j.bbrc.2008.02.003]
- 115 **Dominguez M**, Miquel R, Colmenero J, Moreno M, García-Pagán JC, Bosch J, Arroyo V, Ginès P, Caballería J, Bataller R. Hepatic expression of CXC chemokines predicts portal hypertension and survival in patients with alcoholic hepatitis. *Gastroenterology* 2009; **136**: 1639-1650 [PMID: 19208360 DOI: 10.1053/j.gastro.2009.01.056]
- 116 **Gao B**, Xu M. Chemokines and alcoholic hepatitis: are chemokines good therapeutic targets? *Gut* 2014; **63**: 1683-1684 [PMID: 24515805 DOI: 10.1136/gutjnl-2013-306603]
- 117 **Affò S**, Morales-Ibanez O, Rodrigo-Torres D, Altamirano J, Blaya D, Dapito DH, Millán C, Coll M, Caviglia JM, Arroyo V, Caballería J, Schwabe RF, Ginès P, Bataller R, Sancho-Bru P. CCL20 mediates lipopolysaccharide induced liver injury and is a potential driver of inflammation and fibrosis in alcoholic hepatitis. *Gut* 2014; **63**: 1782-1792 [PMID: 24415562 DOI: 10.1136/gutjnl-2013-306098]
- 118 **Leake I**. Alcoholic hepatitis: potential role of cytokine CCL20 in alcoholic hepatitis. *Nat Rev Gastroenterol Hepatol* 2014; **11**: 76 [PMID: 24473363 DOI: 10.1038/nrgastro.2014.9]
- 119 **Horiguchi N**, Wang L, Mukhopadhyay P, Park O, Jeong WI, Lafdil F, Osei-Hyiaman D, Moh A, Fu XY, Pacher P, Kunos G, Gao B. Cell type-dependent pro- and anti-inflammatory role of signal transducer and activator of transcription 3 in alcoholic liver injury. *Gastroenterology* 2008; **134**: 1148-1158 [PMID: 18395093 DOI: 10.1053/j.gastro.2008.01.016]
- 120 **Støy S**, Sandahl TD, Dige AK, Agnholt J, Rasmussen TK, Grønbaek H, Deleuran B, Vilstrup H. Highest frequencies of interleukin-22-producing T helper cells in alcoholic hepatitis patients with a favourable short-term course. *PLoS One* 2013; **8**: e55101 [PMID: 23372820 DOI: 10.1371/journal.pone.0055101]
- 121 **Ki SH**, Park O, Zheng M, Morales-Ibanez O, Kolls JK, Bataller R, Gao B. Interleukin-22 treatment ameliorates alcoholic liver injury in a murine model of chronic-binge ethanol feeding: role of signal transducer and activator of transcription 3. *Hepatology* 2010; **52**: 1291-1300 [PMID: 20842630 DOI: 10.1002/hep.23837]
- 122 **Radaeva S**, Sun R, Pan HN, Hong F, Gao B. Interleukin 22 (IL-22) plays a protective role in T cell-mediated murine hepatitis: IL-22 is a survival factor for hepatocytes via STAT3 activation. *Hepatology* 2004; **39**: 1332-1342 [PMID: 15122762 DOI: 10.1002/hep.20184]
- 123 **Lemmers A**, Moreno C, Gustot T, Maréchal R, Degré D, Demetter P, de Nadai P, Geerts A, Quertinmont E, Vercruysse V, Le Moine O, Devière J. The interleukin-17 pathway is involved in human alcoholic liver disease. *Hepatology* 2009; **49**: 646-657 [PMID: 19177575 DOI: 10.1002/hep.22680]
- 124 **Hueber W**, Patel DD, Dryja T, Wright AM, Koroleva I, Bruin G, Antoni C, Draelos Z, Gold MH; Psoriasis Study Group, Durez P, Tak PP, Gomez-Reino JJ, Rheumatoid Arthritis Study Group, Foster CS, Kim RY, Samson CM, Falk NS, Chu DS, Callanan D, Nguyen QD, Uveitis Study Group, Rose K, Haider A, Di Padova F. Effects of AIN457, a fully human antibody to interleukin-17A, on psoriasis, rheumatoid arthritis, and uveitis. *Sci Transl Med* 2010; **2**: 52ra72 [PMID: 20926833 DOI: 10.1126/scitranslmed.3001107]
- 125 **Tam J**, Liu J, Mukhopadhyay B, Cinar R, Godlewski G, Kunos G. Endocannabinoids in liver disease. *Hepatology* 2011; **53**: 346-355 [PMID: 21254182 DOI: 10.1002/hep.24077]
- 126 **Jeong WI**, Osei-Hyiaman D, Park O, Liu J, Bátkai S, Mukhopadhyay P, Horiguchi N, Harvey-White J, Marsicano G, Lutz B, Gao B, Kunos G. Paracrine activation of hepatic CB1 receptors by stellate cell-derived endocannabinoids mediates alcoholic fatty liver. *Cell Metab* 2008; **7**: 227-235 [PMID: 18316028 DOI: 10.1016/j.cmet.2007.12.007]
- 127 **Louvet A**, Teixeira-Clerc F, Chobert MN, Deveaux V, Pavoiné C, Zimmer A, Pecker F, Mallat A, Lotersztajn S. Cannabinoid CB2 receptors protect against alcoholic liver disease by regulating Kupffer cell polarization in mice. *Hepatology* 2011; **54**: 1217-1226 [PMID: 21735467 DOI: 10.1002/hep.24524]
- 128 **Wang KX**, Denhardt DT. Osteopontin: role in immune regulation and stress responses. *Cytokine Growth Factor Rev* 2008; **19**: 333-345 [PMID: 18952487 DOI: 10.1016/j.cytogfr.2008.08.001]
- 129 **Morales-Ibanez O**, Domínguez M, Ki SH, Marcos M, Chaves JF, Nguyen-Khac E, Houchi H, Affò S, Sancho-Bru P, Altamirano J, Michelena J, García-Pagán JC, Abalde JG, Arroyo V, Caballería J, Laso FJ, Gao B, Bataller R. Human and experimental evidence supporting a role for osteopontin in alcoholic hepatitis. *Hepatology* 2013; **58**: 1742-1756 [PMID: 23729174 DOI: 10.1002/hep.26521]
- 130 **Altamirano J**, Bataller R. Alcoholic liver disease: pathogenesis and new targets for therapy. *Nat Rev Gastroenterol Hepatol* 2011; **8**: 491-501 [PMID: 21826088 DOI: 10.1038/nrgastro.2011.134]
- 131 **Zhang Z**, Wang FS. Stem cell therapies for liver failure and cirrhosis. *J Hepatol* 2013; **59**: 183-185 [PMID: 23353868 DOI: 10.1016/j.jhep.2013.01.018]
- 132 **Akiyama K**, Chen C, Wang D, Xu X, Qu C, Yamaza T, Cai T, Chen W, Sun L, Shi S. Mesenchymal-stem-cell-induced immunoregulation involves FAS-ligand-/FAS-mediated T cell apoptosis. *Cell Stem Cell* 2012; **10**: 544-555 [PMID: 22542159 DOI: 10.1016/j.stem.2012.03.007]
- 133 **Aurich H**, Sgodda M, Kaltwasser P, Vetter M, Weise A, Liehr T, Brulport M, Hengstler JG, Dollinger MM, Fleig WE, Christ B. Hepatocyte differentiation of mesenchymal stem cells from human adipose tissue in vitro promotes hepatic integration in vivo. *Gut* 2009; **58**: 570-581 [PMID: 19022918 DOI: 10.1136/gut.2008.154880]
- 134 **Kuo TK**, Hung SP, Chuang CH, Chen CT, Shih YR, Fang SC, Yang VW, Lee OK. Stem cell therapy for liver disease: parameters governing the success of using bone marrow mesenchymal stem cells. *Gastroenterology* 2008; **134**: 2111-2121, 2121.e1-2121.e3 [PMID: 18455168 DOI: 10.1053/j.gastro.2008.03.015]
- 135 **Jang YO**, Kim YJ, Baik SK, Kim MY, Eom YW, Cho MY, Park HJ, Park SY, Kim BR, Kim JW, Soo Kim H, Kwon SO, Choi EH, Kim YM. Histological improvement following administration of autologous bone marrow-derived mesenchymal stem cells for alcoholic cirrhosis: a pilot study. *Liver Int* 2014; **34**: 33-41 [PMID: 23782511 DOI: 10.1111/liv.12218]
- 136 **Terai S**, Ishikawa T, Omori K, Aoyama K, Marumoto Y, Urata Y, Yokoyama Y, Uchida K, Yamasaki T, Fujii Y, Okita K, Sakaida I. Improved liver function in patients with liver cirrhosis after autologous bone marrow cell infusion therapy. *Stem Cells* 2006; **24**: 2292-2298 [PMID: 16778155 DOI: 10.1634/stemcells.2005-0542]
- 137 **Ismail A**, Fouad O, Abdelnasser A, Chowdhury A, Selim A. Stem cell therapy improves the outcome of liver resection in cirrhotics. *J Gastrointest Cancer* 2010; **41**: 17-23 [PMID: 20012230 DOI: 10.1007/s12029-009-9092-9]
- 138 **Harrison SA**, Day CP. Benefits of lifestyle modification in NAFLD. *Gut* 2007; **56**: 1760-1769 [PMID: 17911352 DOI: 10.1136/gut.2006.112094]
- 139 **Hannah WN Jr**, Harrison SA. Lifestyle and Dietary Interventions in the Management of Nonalcoholic Fatty Liver Disease. *Dig Dis Sci* 2016; **61**: 1365-1374 [PMID: 27052013 DOI: 10.1007/s10620-016-4153-y]
- 140 **Tiig H**, Moschen A. Weight loss: cornerstone in the treatment of non-alcoholic fatty liver disease. *Minerva Gastroenterol Dietol* 2010; **56**: 159-167 [PMID: 20485253]
- 141 **Ouyang X**, Cirillo P, Sautin Y, McCall S, Bruchette JL, Diehl AM, Johnson RJ, Abdelmalek MF. Fructose consumption as a risk factor for non-alcoholic fatty liver disease. *J Hepatol* 2008; **48**: 993-999 [PMID: 18395287 DOI: 10.1016/j.jhep.2008.02.011]
- 142 **Simopoulos AP**. Dietary omega-3 fatty acid deficiency and high fructose intake in the development of metabolic syndrome, brain metabolic abnormalities, and non-alcoholic fatty liver disease.

- Nutrients* 2013; **5**: 2901-2923 [PMID: 23896654 DOI: 10.3390/nu5082901]
- 143 **Al-Gayyar MM**, Shams ME, Barakat EA. Fish oil improves lipid metabolism and ameliorates inflammation in patients with metabolic syndrome: impact of nonalcoholic fatty liver disease. *Pharm Biol* 2012; **50**: 297-303 [PMID: 22103753 DOI: 10.3109/13880209.2011.604088]
 - 144 **Akyüz F**, Demir K, Özdiş S, Aksoy N, Poturoğlu S, İbrişim D, Kaymakoglu S, Beşikçi F, Boztaş G, Cakaloğlu Y, Mungan Z, Cevikbaş U, Oktan A. The effects of rosiglitazone, metformin, and diet with exercise in nonalcoholic fatty liver disease. *Dig Dis Sci* 2007; **52**: 2359-2367 [PMID: 17429734 DOI: 10.1007/s10620-006-9145-x]
 - 145 **Balkestein EJ**, van Aggel-Leijssen DP, van Baak MA, Struijker-Boudier HA, Van Bortel LM. The effect of weight loss with or without exercise training on large artery compliance in healthy obese men. *J Hypertens* 1999; **17**: 1831-1835 [PMID: 10703876]
 - 146 **Luyckx FH**, Desai C, Thiry A, Dewé W, Scheen AJ, Gielen JE, Lefebvre PJ. Liver abnormalities in severely obese subjects: effect of drastic weight loss after gastropasty. *Int J Obes Relat Metab Disord* 1998; **22**: 222-226 [PMID: 9539189]
 - 147 **Weinsier RL**, Wilson LJ, Lee J. Medically safe rate of weight loss for the treatment of obesity: a guideline based on risk of gallstone formation. *Am J Med* 1995; **98**: 115-117 [PMID: 7847427 DOI: 10.1016/S0002-9343(99)80394-5]
 - 148 **Zelber-Sagi S**, Kessler A, Brazowsky E, Webb M, Lurie Y, Santo M, Leshno M, Blendis L, Halpern Z, Oren R. A double-blind randomized placebo-controlled trial of orlistat for the treatment of nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 2006; **4**: 639-644 [PMID: 16630771 DOI: 10.1016/j.cgh.2006.02.004]
 - 149 **Sabuncu T**, Nazligül Y, Karaoglanoglu M, Ucar E, Kilic FB. The effects of sibutramine and orlistat on the ultrasonographic findings, insulin resistance and liver enzyme levels in obese patients with non-alcoholic steatohepatitis. *Rom J Gastroenterol* 2003; **12**: 189-192 [PMID: 14502318]
 - 150 **Yki-Järvinen H**. Thiazolidinediones. *N Engl J Med* 2004; **351**: 1106-1118 [PMID: 15356308 DOI: 10.1056/NEJMra041001]
 - 151 **Bajaj M**, Suraamornkul S, Piper P, Hardies LJ, Glass L, Cersosimo E, Pratipanawatr T, Miyazaki Y, DeFronzo RA. Decreased plasma adiponectin concentrations are closely related to hepatic fat content and hepatic insulin resistance in pioglitazone-treated type 2 diabetic patients. *J Clin Endocrinol Metab* 2004; **89**: 200-206 [PMID: 14715850 DOI: 10.1210/jc.2003-031315]
 - 152 **Caldwell SH**, Hespdenheide EE, Redick JA, Iezzoni JC, Battle EH, Sheppard BL. A pilot study of a thiazolidinedione, troglitazone, in nonalcoholic steatohepatitis. *Am J Gastroenterol* 2001; **96**: 519-525 [PMID: 11232700 DOI: 10.1111/j.1572-0241.2001.03553.x]
 - 153 **Aithal GP**, Thomas JA, Kaye PV, Lawson A, Ryder SD, Spendlove I, Austin AS, Freeman JG, Morgan L, Webber J. Randomized, placebo-controlled trial of pioglitazone in nondiabetic subjects with nonalcoholic steatohepatitis. *Gastroenterology* 2008; **135**: 1176-1184 [PMID: 18718471 DOI: 10.1053/j.gastro.2008.06.047]
 - 154 **Ratzu V**, Giral P, Jacqueminet S, Charlotte F, Hartemann-Heurtier A, Serfaty L, Podevin P, Lacorte JM, Bernhardt C, Bruckert E, Grimaldi A, Poynard T, LIDO Study Group. Rosiglitazone for nonalcoholic steatohepatitis: one-year results of the randomized placebo-controlled Fatty Liver Improvement with Rosiglitazone Therapy (FLIRT) Trial. *Gastroenterology* 2008; **135**: 100-110 [PMID: 18503774 DOI: 10.1053/j.gastro.2008.03.078]
 - 155 **Lutchman G**, Modi A, Kleiner DE, Promrat K, Heller T, Ghany M, Borg B, Loomba R, Liang TJ, Premkumar A, Hoofnagle JH. The effects of discontinuing pioglitazone in patients with nonalcoholic steatohepatitis. *Hepatology* 2007; **46**: 424-429 [PMID: 17559148 DOI: 10.1002/hep.21661]
 - 156 **Juurlink DN**, Gomes T, Lipscombe LL, Austin PC, Hux JE, Mamdani MM. Adverse cardiovascular events during treatment with pioglitazone and rosiglitazone: population based cohort study. *BMJ* 2009; **339**: b2942 [PMID: 19690342 DOI: 10.1136/bmj.b2942]
 - 157 **Loomba R**, Lutchman G, Kleiner DE, Ricks M, Feld JJ, Borg BB, Modi A, Nagabhyru P, Sumner AE, Liang TJ, Hoofnagle JH. Clinical trial: pilot study of metformin for the treatment of non-alcoholic steatohepatitis. *Aliment Pharmacol Ther* 2009; **29**: 172-182 [PMID: 18945255 DOI: 10.1111/j.1365-2036.2008.03869.x]
 - 158 **Haukeland JW**, Konopski Z, Eggesbø HB, von Volkmann HL, Raschpichler G, Bjørø K, Haaland T, Løberg EM, Birkeland K. Metformin in patients with non-alcoholic fatty liver disease: a randomized, controlled trial. *Scand J Gastroenterol* 2009; **44**: 853-860 [PMID: 19811343 DOI: 10.1080/00365520902845268]
 - 159 **Laurin J**, Lindor KD, Crippin JS, Gossard A, Gores GJ, Ludwig J, Rakela J, McGill DB. Ursodeoxycholic acid or clofibrate in the treatment of non-alcohol-induced steatohepatitis: a pilot study. *Hepatology* 1996; **23**: 1464-1467 [PMID: 8675165 DOI: 10.1002/hep.510230624]
 - 160 **Basaranoglu M**, Acbay O, Sonsuz A. A controlled trial of gemfibrozil in the treatment of patients with nonalcoholic steatohepatitis. *J Hepatol* 1999; **31**: 384 [PMID: 10453959]
 - 161 **Assy N**, Grozovski M, Bersudsky I, Szvalb S, Hussein O. Effect of insulin-sensitizing agents in combination with ezetimibe, and valsartan in rats with non-alcoholic fatty liver disease. *World J Gastroenterol* 2006; **12**: 4369-4376 [PMID: 16865780 DOI: 10.3748/wjg.v12.i27.4369]
 - 162 **Zheng S**, Hoos L, Cook J, Tetzloff G, Davis H Jr, van Heek M, Hwa JJ. Ezetimibe improves high fat and cholesterol diet-induced non-alcoholic fatty liver disease in mice. *Eur J Pharmacol* 2008; **584**: 118-124 [PMID: 18329014 DOI: 10.1016/j.ejphar.2008.01.045]
 - 163 **Lindor KD**, Kowdley KV, Heathcote EJ, Harrison ME, Jorgensen R, Angulo P, Lymp JF, Burgart L, Colin P. Ursodeoxycholic acid for treatment of nonalcoholic steatohepatitis: results of a randomized trial. *Hepatology* 2004; **39**: 770-778 [PMID: 14999696 DOI: 10.1002/hep.20092]
 - 164 **Farrell GC**, Larter CZ. Nonalcoholic fatty liver disease: from steatosis to cirrhosis. *Hepatology* 2006; **43**: S99-S112 [PMID: 16447287 DOI: 10.1002/hep.20973]
 - 165 **Harrison SA**, Torgerson S, Hayashi P, Ward J, Schenker S. Vitamin E and vitamin C treatment improves fibrosis in patients with nonalcoholic steatohepatitis. *Am J Gastroenterol* 2003; **98**: 2485-2490 [PMID: 14638353 DOI: 10.1111/j.1572-0241.2003.08699.x]
 - 166 **Kugelmas M**, Hill DB, Vivian B, Marsano L, McClain CJ. Cytokines and NASH: a pilot study of the effects of lifestyle modification and vitamin E. *Hepatology* 2003; **38**: 413-419 [PMID: 12883485 DOI: 10.1053/jhep.2003.50316]
 - 167 **Adams LA**, Angulo P. Vitamins E and C for the treatment of NASH: duplication of results but lack of demonstration of efficacy. *Am J Gastroenterol* 2003; **98**: 2348-2350 [PMID: 14638333 DOI: 10.1111/j.1572-0241.2003.08695.x]
 - 168 **Dufour JF**, Oneta CM, Gonvers JJ, Bihl F, Cerny A, Cereda JM, Zala JF, Helbling B, Steuierwald M, Zimmermann A, Swiss Association for the Study of the Liver. Randomized placebo-controlled trial of ursodeoxycholic acid with vitamin e in nonalcoholic steatohepatitis. *Clin Gastroenterol Hepatol* 2006; **4**: 1537-1543 [PMID: 17162245 DOI: 10.1016/j.cgh.2006.09.025]
 - 169 **Sanyal AJ**, Mofrad PS, Contos MJ, Sargeant C, Luketic VA, Sterling RK, Stravitz RT, Shiffman ML, Clore J, Mills AS. A pilot study of vitamin E versus vitamin E and pioglitazone for the treatment of nonalcoholic steatohepatitis. *Clin Gastroenterol Hepatol* 2004; **2**: 1107-1115 [PMID: 15625656]
 - 170 **Miller ER 3rd**, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, Guallar E. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Ann Intern Med* 2005; **142**: 37-46 [PMID: 15537682]
 - 171 **Tushuizen ME**, Bunck MC, Pouwels PJ, van Waesberghe JH, Diamant M, Heine RJ. Incretin mimetics as a novel therapeutic option for hepatic steatosis. *Liver Int* 2006; **26**: 1015-1017 [PMID: 16953843 DOI: 10.1111/j.1478-3231.2006.01315.x]
 - 172 **Ding X**, Saxena NK, Lin S, Gupta NA, Anania FA. Exendin-4, a glucagon-like protein-1 (GLP-1) receptor agonist, reverses hepatic steatosis in ob/ob mice. *Hepatology* 2006; **43**: 173-181 [PMID:

- 16374859 DOI: 10.1002/hep.21006]
- 173 **Ostawal A**, Mocevic E, Kragh N, Xu W. Clinical Effectiveness of Liraglutide in Type 2 Diabetes Treatment in the Real-World Setting: A Systematic Literature Review. *Diabetes Ther* 2016; **7**: 411-438 [PMID: 27350545 DOI: 10.1007/s13300-016-0180-0]
 - 174 **Armstrong MJ**, Gaunt P, Aithal GP, Barton D, Hull D, Parker R, Hazlehurst JM, Guo K; LEAN trial team, Abouda G, Aldersley MA, Stocken D, Gough SC, Tomlinson JW, Brown RM, Hübscher SG, Newsome PN. Liraglutide safety and efficacy in patients with non-alcoholic steatohepatitis (LEAN): a multicentre, double-blind, randomised, placebo-controlled phase 2 study. *Lancet* 2016; **387**: 679-690 [PMID: 26608256 DOI: 10.1016/S0140-6736(15)00803-X]
 - 175 **Armstrong MJ**, Hull D, Guo K, Barton D, Hazlehurst JM, Gathercole LL, Nasiri M, Yu J, Gough SC, Newsome PN, Tomlinson JW. Glucagon-like peptide 1 decreases lipotoxicity in non-alcoholic steatohepatitis. *J Hepatol* 2016; **64**: 399-408 [PMID: 26394161 DOI: 10.1016/j.jhep.2015.08.038]
 - 176 **Drucker DJ**. Dipeptidyl peptidase-4 inhibition and the treatment of type 2 diabetes: preclinical biology and mechanisms of action. *Diabetes Care* 2007; **30**: 1335-1343 [PMID: 17337495 DOI: 10.2337/dc07-0228]
 - 177 **Balaban YH**, Korkusuz P, Simsek H, Gokcan H, Gedikoglu G, Pinar A, Hascelik G, Asan E, Hamaloglu E, Tatar G. Dipeptidyl peptidase IV (DDP IV) in NASH patients. *Ann Hepatol* 2007; **6**: 242-250 [PMID: 18007554]
 - 178 **Firneisz G**, Varga T, Lengyel G, Fehér J, Ghyczy D, Wichmann B, Selmei L, Tulassay Z, Rácz K, Somogyi A. Serum dipeptidyl peptidase-4 activity in insulin resistant patients with non-alcoholic fatty liver disease: a novel liver disease biomarker. *PLoS One* 2010; **5**: e12226 [PMID: 20805868 DOI: 10.1371/journal.pone.0012226]
 - 179 **Baumeier C**, Saussenthaler S, Kammel A, Jähner M, Schlüter L, Hesse D, Canouil M, Lobbens S, Caiazzo R, Raverdy V, Pattou F, Nilsson E, Pihlajamäki J, Ling C, Froguel P, Schürmann A, Schwenk RW. Hepatic DPP4 DNA Methylation Associates With Fatty Liver. *Diabetes* 2017; **66**: 25-35 [PMID: 27999105 DOI: 10.2337/db15-1716]
 - 180 **Iwasaki T**, Yoneda M, Inamori M, Shirakawa J, Higurashi T, Maeda S, Terauchi Y, Nakajima A. Sitagliptin as a novel treatment agent for non-alcoholic Fatty liver disease patients with type 2 diabetes mellitus. *Hepato gastroenterology* 2011; **58**: 2103-2105 [PMID: 22024083 DOI: 10.5754/hge11263]
 - 181 **Itou M**, Kawaguchi T, Taniguchi E, Oriishi T, Sata M. Dipeptidyl Peptidase IV Inhibitor Improves Insulin Resistance and Steatosis in a Refractory Nonalcoholic Fatty Liver Disease Patient: A Case Report. *Case Rep Gastroenterol* 2012; **6**: 538-544 [PMID: 22949894 DOI: 10.1159/000341510]
 - 182 **Cui J**, Philo L, Nguyen P, Hofflich H, Hernandez C, Bettencourt R, Richards L, Salotti J, Bhatt A, Hooker J, Haufe W, Hooker C, Brenner DA, Sirlin CB, Loomba R. Sitagliptin vs. placebo for non-alcoholic fatty liver disease: A randomized controlled trial. *J Hepatol* 2016; **65**: 369-376 [PMID: 27151177 DOI: 10.1016/j.jhep.2016.04.021]
 - 183 **Joy TR**, McKenzie CA, Tirona RG, Summers K, Seney S, Chakrabarti S, Malhotra N, Beaton MD. Sitagliptin in patients with non-alcoholic steatohepatitis: A randomized, placebo-controlled trial. *World J Gastroenterol* 2017; **23**: 141-150 [PMID: 28104990 DOI: 10.3748/wjg.v23.i1.141]
 - 184 **Duman DG**, Ozdemir F, Birben E, Keskin O, Ekşioğlu-Demiralp E, Celikel C, Kalayci O, Kalayci C. Effects of pentoxifylline on TNF- α production by peripheral blood mononuclear cells in patients with nonalcoholic steatohepatitis. *Dig Dis Sci* 2007; **52**: 2520-2524 [PMID: 17436095 DOI: 10.1007/s10620-006-9723-y]
 - 185 **Van Wagner LB**, Koppe SW, Brunt EM, Gottstein J, Gardikiotes K, Green RM, Rinella ME. Pentoxifylline for the treatment of non-alcoholic steatohepatitis: a randomized controlled trial. *Ann Hepatol* 2011; **10**: 277-286 [PMID: 21677329]
 - 186 **Zein CO**, Yerian LM, Gogate P, Lopez R, Kirwan JP, Feldstein AE, McCullough AJ. Pentoxifylline improves nonalcoholic steatohepatitis: a randomized placebo-controlled trial. *Hepatology* 2011; **54**: 1610-1619 [PMID: 21748765 DOI: 10.1002/hep.24544]
 - 187 **Zein CO**, Lopez R, Fu X, Kirwan JP, Yerian LM, McCullough AJ, Hazen SL, Feldstein AE. Pentoxifylline decreases oxidized lipid products in nonalcoholic steatohepatitis: new evidence on the potential therapeutic mechanism. *Hepatology* 2012; **56**: 1291-1299 [PMID: 22505276 DOI: 10.1002/hep.25778]
 - 188 **Du J**, Ma YY, Yu CH, Li YM. Effects of pentoxifylline on nonalcoholic fatty liver disease: a meta-analysis. *World J Gastroenterol* 2014; **20**: 569-577 [PMID: 24574727 DOI: 10.3748/wjg.v20.i2.569]
 - 189 **Zeng T**, Zhang CL, Zhao XL, Xie KQ. Pentoxifylline for the treatment of nonalcoholic fatty liver disease: a meta-analysis of randomized double-blind, placebo-controlled studies. *Eur J Gastroenterol Hepatol* 2014; **26**: 646-653 [PMID: 24743504 DOI: 10.1097/MEG.000000000000068]
 - 190 **Singh S**, Khera R, Allen AM, Murad MH, Loomba R. Comparative effectiveness of pharmacological interventions for nonalcoholic steatohepatitis: A systematic review and network meta-analysis. *Hepatology* 2015; **62**: 1417-1432 [PMID: 26189925 DOI: 10.1002/hep.27999]
 - 191 **Solga SF**, Diehl AM. Non-alcoholic fatty liver disease: lumen-liver interactions and possible role for probiotics. *J Hepatol* 2003; **38**: 681-687 [PMID: 12713883]
 - 192 **Miele L**, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, Mascianà R, Forgione A, Gabrieli ML, Perotti G, Vecchio FM, Rapaccini G, Gasbarrini G, Day CP, Grieco A. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* 2009; **49**: 1877-1887 [PMID: 19291785 DOI: 10.1002/hep.22848]
 - 193 **Aller R**, De Luis DA, Izaola O, Conde R, Gonzalez Sagrado M, Primo D, De La Fuente B, Gonzalez J. Effect of a probiotic on liver aminotransferases in nonalcoholic fatty liver disease patients: a double blind randomized clinical trial. *Eur Rev Med Pharmacol Sci* 2011; **15**: 1090-1095 [PMID: 22013734]
 - 194 **Malaguarnera M**, Vacante M, Antic T, Giordano M, Chisari G, Acquaviva R, Mastrojeni S, Malaguarnera G, Mistretta A, Li Volti G, Galvano F. Bifidobacterium longum with fructo-oligosaccharides in patients with non alcoholic steatohepatitis. *Dig Dis Sci* 2012; **57**: 545-553 [PMID: 21901256 DOI: 10.1007/s10620-011-1887-4]
 - 195 **Yokohama S**, Yoneda M, Haneda M, Okamoto S, Okada M, Aso K, Hasegawa T, Tokusashi Y, Miyokawa N, Nakamura K. Therapeutic efficacy of an angiotensin II receptor antagonist in patients with nonalcoholic steatohepatitis. *Hepatology* 2004; **40**: 1222-1225 [PMID: 15382153 DOI: 10.1002/hep.20420]
 - 196 **Mallat A**, Lotersztajn S. Endocannabinoids and liver disease. I. Endocannabinoids and their receptors in the liver. *Am J Physiol Gastrointest Liver Physiol* 2008; **294**: G9-G12 [PMID: 17975129 DOI: 10.1152/ajpgi.00467.2007]
 - 197 **Osei-Hyiaman D**, DePetrillo M, Pacher P, Liu J, Radaeva S, Bátkai S, Harvey-White J, Mackie K, Offertáler L, Wang L, Kunos G. Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. *J Clin Invest* 2005; **115**: 1298-1305 [PMID: 15864349 DOI: 10.1172/JCI23057]
 - 198 **Gary-Bobo M**, Elachouri G, Gallas JF, Janiak P, Marini P, Ravinet-Trillou C, Chabbert M, Cruccioli N, Pfersdorff C, Roque C, Arnone M, Croci T, Soubrié P, Oury-Donat F, Maffrand JP, Scatton B, Lacheretz F, Le Fur G, Herbert JM, Bensaid M. Rimonabant reduces obesity-associated hepatic steatosis and features of metabolic syndrome in obese Zucker fa/fa rats. *Hepatology* 2007; **46**: 122-129 [PMID: 17526015 DOI: 10.1002/hep.21641]
 - 199 **Rimonabant**: suicide and depression. Depression and suicidal tendencies are about twice as frequent with rimonabant as with placebo. *Prescrire Int* 2007; **16**: 250 [PMID: 18092422]
 - 200 **Christopoulou FD**, Kiortsis DN. An overview of the metabolic effects of rimonabant in randomized controlled trials: potential for other cannabinoid 1 receptor blockers in obesity. *J Clin Pharm Ther* 2011; **36**: 10-18 [PMID: 21198716 DOI: 10.1111/j.1365-2710.2010.01164.x]

- 201 **Stephen S**, Baranova A, Younossi ZM. Nonalcoholic fatty liver disease and bariatric surgery. *Expert Rev Gastroenterol Hepatol* 2012; **6**: 163-171 [PMID: 22375522 DOI: 10.1586/egh.11.97]
- 202 **Grimm IS**, Schindler W, Haluszka O. Steatohepatitis and fatal hepatic failure after biliopancreatic diversion. *Am J Gastroenterol* 1992; **87**: 775-779 [PMID: 1590319]
- 203 **Charlton M**, Kasparova P, Weston S, Lindor K, Maor-Kendler Y, Wiesner RH, Rosen CB, Batts KP. Frequency of nonalcoholic steatohepatitis as a cause of advanced liver disease. *Liver Transpl* 2001; **7**: 608-614 [PMID: 11460228 DOI: 10.1053/jlts.2001.25453]
- 204 **Anstee QM**, Concas D, Kudo H, Levene A, Pollard J, Charlton P, Thomas HC, Thursz MR, Goldin RD. Impact of pan-caspase inhibition in animal models of established steatosis and non-alcoholic steatohepatitis. *J Hepatol* 2010; **53**: 542-550 [PMID: 20557969 DOI: 10.1016/j.jhep.2010.03.016]
- 205 **Witek RP**, Stone WC, Karaca FG, Syn WK, Pereira TA, Agboola KM, Omenetti A, Jung Y, Teaberry V, Choi SS, Guy CD, Pollard J, Charlton P, Diehl AM. Pan-caspase inhibitor VX-166 reduces fibrosis in an animal model of nonalcoholic steatohepatitis. *Hepatology* 2009; **50**: 1421-1430 [PMID: 19676126 DOI: 10.1002/hep.23167]
- 206 **Shiffman M**, Freilich B, Vuppalandhi R, Watt K, Burgess G, Morris M, Sheedy B, Schiff E. LP37: A placebo-controlled, multicenter, double-blind, randomised trial of emricasan in subjects with non-alcoholic fatty liver disease (NAFLD) and raised transaminases. *J Hepatol* 2015; **62**: S282 [DOI: 10.1016/S0168-8278(15)30191-4]
- 207 **Karnik S**, Charlton M, Popov Y, Goodman ZD, Nash M, Sulfab M, Barry V, Huntzicker EG, French D, Li K, Decaris M, Emson C, Turner S, Breckenridge D, Tumas D. Pharmacological inhibition of apoptosis signal-regulating kinase 1 (ASK1) in a murine model of NASH with pre-existing disease blocks fibrosis, steatosis, and insulin resistance. *Hepatology* 2014; **60**: 570A
- 208 **Karnik S**, Charlton MR, Li L, Nash M, Sulfab M, Newstrom D, Huntzicker EG, French D, Goodman ZD, Shafizadeh T, Watkins S, Breckenridge D, Tumas D. Efficacy of an ASK1 Inhibitor to Reduce Fibrosis and Steatosis in a Murine Model of NASH is Associated with Normalization of Lipids and Hepatic Gene Expression and a Reduction in Serum Biomarkers of Inflammation and Fibrosis. *Hepatology* 2015; **62**: 877A
- 209 **Xiang M**, Wang PX, Wang AB, Zhang XJ, Zhang Y, Zhang P, Mei FH, Chen MH, Li H. Targeting hepatic TRAF1-ASK1 signaling to improve inflammation, insulin resistance, and hepatic steatosis. *J Hepatol* 2016; **64**: 1365-1377 [PMID: 26860405 DOI: 10.1016/j.jhep.2016.02.002]
- 210 **Sabio G**, Davis RJ. cJun NH2-terminal kinase 1 (JNK1): roles in metabolic regulation of insulin resistance. *Trends Biochem Sci* 2010; **35**: 490-496 [PMID: 20452774 DOI: 10.1016/j.tibs.2010.04.004]
- 211 **Sabin G**, Davis RJ. TNF and MAP kinase signalling pathways. *Semin Immunol* 2014; **26**: 237-245 [PMID: 24647229 DOI: 10.1016/j.smim.2014.02.009]
- 212 **Sabio G**, Kennedy NJ, Cavanagh-Kyros J, Jung DY, Ko HJ, Ong H, Barrett T, Kim JK, Davis RJ. Role of muscle c-Jun NH2-terminal kinase 1 in obesity-induced insulin resistance. *Mol Cell Biol* 2010; **30**: 106-115 [PMID: 19841069 DOI: 10.1128/MCB.01162-09]
- 213 **González-Terán B**, Cortés JR, Manieri E, Matesanz N, Verdugo Á, Rodríguez ME, González-Rodríguez Á, Valverde ÁM, Martín P, Davis RJ, Sabio G. Eukaryotic elongation factor 2 controls TNF- α translation in LPS-induced hepatitis. *J Clin Invest* 2013; **123**: 164-178 [PMID: 23202732 DOI: 10.1172/JCI65124]
- 214 **Risco A**, del Fresno C, Mambol A, Alsina-Beauchamp D, MacKenzie KF, Yang HT, Barber DF, Morcelle C, Arthur JS, Ley SC, Ardavin C, Cuenda A. p38 γ and p38 δ kinases regulate the Toll-like receptor 4 (TLR4)-induced cytokine production by controlling ERK1/2 protein kinase pathway activation. *Proc Natl Acad Sci USA* 2012; **109**: 11200-11205 [PMID: 22733747 DOI: 10.1073/pnas.1207290109]
- 215 **González-Terán B**, Matesanz N, Nikolic I, Verdugo MA, Sreeramkumar V, Hernández-Cosido L, Mora A, Crainiciuc G, Sáiz ML, Bernardo E, Leiva-Vega L, Rodríguez E, Bondía V, Torres JL, Perez-Sieira S, Ortega L, Cuenda A, Sanchez-Madrid F, Nogueiras R, Hidalgo A, Marcos M, Sabio G. p38 γ and p38 δ reprogram liver metabolism by modulating neutrophil infiltration. *EMBO J* 2016; **35**: 536-552 [PMID: 26843485 DOI: 10.15252/embj.201591857]
- 216 **Staels B**, Rubenstrunk A, Noel B, Rigou G, Delataille P, Millatt LJ, Baron M, Lucas A, Tailleux A, Hum DW, Ratzu V, Cariou B, Hanf R. Hepatoprotective effects of the dual peroxisome proliferator-activated receptor α/δ agonist, GFT505, in rodent models of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. *Hepatology* 2013; **58**: 1941-1952 [PMID: 23703580 DOI: 10.1002/hep.26461]
- 217 **Montagner A**, Polizzi A, Fouché E, Ducheix S, Lippi Y, Lasserre F, Barquissau V, Régnier M, Lukowicz C, Benhamed F, Iroz A, Bertrand-Michel J, Al Saati T, Cano P, Mselli-Lakhal L, Mithieux G, Rajas F, Lagarrigue S, Pineau T, Loiseau N, Postic C, Langin D, Wahli W, Guillou H. Liver PPAR α is crucial for whole-body fatty acid homeostasis and is protective against NAFLD. *Gut* 2016; **65**: 1202-1214 [PMID: 26838599 DOI: 10.1136/gutjnl-2015-310798]
- 218 **Piccinin E**, Moschetta A. Hepatic-specific PPAR α -FGF21 action in NAFLD. *Gut* 2016; **65**: 1075-1076 [PMID: 26992428 DOI: 10.1136/gutjnl-2016-311408]
- 219 **Ratzu V**, Harrison SA, Francque S, Bedossa P, Leheret P, Serfaty L, Romero-Gomez M, Boursier J, Abdelmalek M, Caldwell S, Drenth J, Anstee QM, Hum D, Hanf R, Roudot A, Megnier S, Staels B, Sanyal A; GOLDEN-505 Investigator Study Group. Elafibranor, an Agonist of the Peroxisome Proliferator-Activated Receptor- α and - δ , Induces Resolution of Nonalcoholic Steatohepatitis Without Fibrosis Worsening. *Gastroenterology* 2016; **150**: 1147-1159.e5 [PMID: 26874076 DOI: 10.1053/j.gastro.2016.01.038]
- 220 **Pellicciari R**, Costantino G, Camaioni E, Sadeghpour BM, Entrena A, Willson TM, Fiorucci S, Clerici C, Gioiello A. Bile acid derivatives as ligands of the farnesoid X receptor. Synthesis, evaluation, and structure-activity relationship of a series of body and side chain modified analogues of chenodeoxycholic acid. *J Med Chem* 2004; **47**: 4559-4569 [PMID: 15317466 DOI: 10.1021/jm049904b]
- 221 **Ballestri S**, Nascimbeni F, Romagnoli D, Baldelli E, Lonardo A. The Role of Nuclear Receptors in the Pathophysiology, Natural Course, and Drug Treatment of NAFLD in Humans. *Adv Ther* 2016; **33**: 291-319 [PMID: 26921205 DOI: 10.1007/s12325-016-0306-9]
- 222 **Fuchs M**. Non-alcoholic Fatty liver disease: the bile Acid-activated farnesoid x receptor as an emerging treatment target. *J Lipids* 2012; **2012**: 934396 [PMID: 22187656 DOI: 10.1155/2012/934396]
- 223 **Makri E**, Cholongitas E, Tziomalos K. Emerging role of obeticholic acid in the management of nonalcoholic fatty liver disease. *World J Gastroenterol* 2016; **22**: 9039-9043 [PMID: 27895393 DOI: 10.3748/wjg.v22.i41.9039]
- 224 **Ali AH**, Carey EJ, Lindor KD. Recent advances in the development of farnesoid X receptor agonists. *Ann Transl Med* 2015; **3**: 5 [PMID: 25705637 DOI: 10.3978/j.issn.2305-5839.2014.12.06]
- 225 **Verbeke L**, Mannaerts I, Schierwagen R, Govaere O, Klein S, Vander Elst I, Windmolders P, Farre R, Wenes M, Mazzone M, Nevens F, van Grunsven LA, Trebicka J, Laleman W. FXR agonist obeticholic acid reduces hepatic inflammation and fibrosis in a rat model of toxic cirrhosis. *Sci Rep* 2016; **6**: 33453 [PMID: 27634375 DOI: 10.1038/srep33453]
- 226 **Úbeda M**, Lario M, Muñoz L, Borrero MJ, Rodríguez-Serrano M, Sánchez-Díaz AM, Del Campo R, Lledó L, Pastor Ó, García-Bermejo L, Díaz D, Álvarez-Mon M, Albillos A. Obeticholic acid reduces bacterial translocation and inhibits intestinal inflammation in cirrhotic rats. *J Hepatol* 2016; **64**: 1049-1057 [PMID: 26723896 DOI: 10.1016/j.jhep.2015.12.010]
- 227 **Mudaliar S**, Henry RR, Sanyal AJ, Morrow L, Marschall HU, Kipnes M, Adorini L, Sciacca CI, Clopton P, Castellote E, Dillon P, Pruzanski M, Shapiro D. Efficacy and safety of the farnesoid X receptor agonist obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver disease. *Gastroenterology* 2013; **145**: 574-582.e1 [PMID: 23727264 DOI: 10.1053/j.gastro.2013.05.042]
- 228 **Neuschwander-Tetri BA**, Loomba R, Sanyal AJ, Lavine JE,

- Van Natta ML, Abdelmalek MF, Chalasani N, Dasarathy S, Diehl AM, Hameed B, Kowdley KV, McCullough A, Terrault N, Clark JM, Tonascia J, Brunt EM, Kleiner DE, Doo E; NASH Clinical Research Network. Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a multicentre, randomised, placebo-controlled trial. *Lancet* 2015; **385**: 956-965 [PMID: 25468160 DOI: 10.1016/S0140-6736(14)61933-4]
- 229 **Pencek R**, Marmon T, Roth JD, Liberman A, Hooshmand-Rad R, Young MA. Effects of obeticholic acid on lipoprotein metabolism in healthy volunteers. *Diabetes Obes Metab* 2016; **18**: 936-940 [PMID: 27109453 DOI: 10.1111/dom.12681]
- 230 **Ghosh Laskar M**, Eriksson M, Rudling M, Angelin B. Treatment with the natural FXR agonist chenodeoxycholic acid reduces clearance of plasma LDL whilst decreasing circulating PCSK9, lipoprotein(a) and apolipoprotein C-III. *J Intern Med* 2017; **281**: 575-585 [PMID: 28145001 DOI: 10.1111/joim.12594]
- 231 **Carino A**, Cipriani S, Marchianò S, Biagioli M, Santorelli C, Donini A, Zampella A, Monti MC, Fiorucci S. BAR502, a dual FXR and GPBAR1 agonist, promotes browning of white adipose tissue and reverses liver steatosis and fibrosis. *Sci Rep* 2017; **7**: 42801 [PMID: 28202906 DOI: 10.1038/srep42801]
- 232 **Gege C**, Kinzel O, Steeneck C, Schulz A, Kremoser C. Knocking on FXR's door: the "hammerhead"-structure series of FXR agonists - amphiphilic isoxazoles with potent in vitro and in vivo activities. *Curr Top Med Chem* 2014; **14**: 2143-2158 [PMID: 25388536]
- 233 **Haga S**, Yimin, Ozaki M. Relevance of FXR-p62/SQSTM1 pathway for survival and protection of mouse hepatocytes and liver, especially with steatosis. *BMC Gastroenterol* 2017; **17**: 9 [PMID: 28086800 DOI: 10.1186/s12876-016-0568-3]
- 234 **Schwabl P**, Hambruch E, Seeland BA, Hayden H, Wagner M, Gams L, Strobel B, Schubert TL, Riedl F, Mitteregger D, Burnet M, Starlinger P, Oberhuber G, Deuschle U, Rohr-Udilova N, Podesser BK, Peck-Radosavljevic M, Reiberger T, Kremoser C, Trauner M. The FXR agonist PX20606 ameliorates portal hypertension by targeting vascular remodelling and sinusoidal dysfunction. *J Hepatol* 2017; **66**: 724-733 [PMID: 27993716 DOI: 10.1016/j.jhep.2016.12.005]
- 235 **Aoyama T**, Paik YH, Watanabe S, Laleu B, Gaggini F, Fioraso-Cartier L, Molango S, Heitz F, Merlot C, Szyndralewicz C, Page P, Brenner DA. Nicotinamide adenine dinucleotide phosphate oxidase in experimental liver fibrosis: GKT137831 as a novel potential therapeutic agent. *Hepatology* 2012; **56**: 2316-2327 [PMID: 22806357 DOI: 10.1002/hep.25938]
- 236 **Paik YH**, Iwaisako K, Seki E, Inokuchi S, Schnabl B, Osterreicher CH, Kisseleva T, Brenner DA. The nicotinamide adenine dinucleotide phosphate oxidase (NOX) homologues NOX1 and NOX2/gp91(phox) mediate hepatic fibrosis in mice. *Hepatology* 2011; **53**: 1730-1741 [PMID: 21384410 DOI: 10.1002/hep.24281]
- 237 **Yang RY**, Rabinovich GA, Liu FT. Galectins: structure, function and therapeutic potential. *Expert Rev Mol Med* 2008; **10**: e17 [PMID: 18549522 DOI: 10.1017/S1462399408000719]
- 238 **Di Lella S**, Sundblad V, Cerliani JP, Guardia CM, Estrin DA, Vasta GR, Rabinovich GA. When galectins recognize glycans: from biochemistry to physiology and back again. *Biochemistry* 2011; **50**: 7842-7857 [PMID: 21848324 DOI: 10.1021/bi201121m]
- 239 **Henderson NC**, Sethi T. The regulation of inflammation by galectin-3. *Immunol Rev* 2009; **230**: 160-171 [PMID: 19594635 DOI: 10.1111/j.1600-065X.2009.00794.x]
- 240 **Traber PG**, Zomer E. Therapy of experimental NASH and fibrosis with galectin inhibitors. *PLoS One* 2013; **8**: e83481 [PMID: 24367597 DOI: 10.1371/journal.pone.0083481]
- 241 **Foster DW**. Malonyl-CoA: the regulator of fatty acid synthesis and oxidation. *J Clin Invest* 2012; **122**: 1958-1959 [PMID: 22833869]
- 242 **Harriman G**, Greenwood J, Bhat S, Huang X, Wang R, Paul D, Tong L, Saha AK, Westlin WF, Kapeller R, Harwood HJ Jr. Acetyl-CoA carboxylase inhibition by ND-630 reduces hepatic steatosis, improves insulin sensitivity, and modulates dyslipidemia in rats. *Proc Natl Acad Sci USA* 2016; **113**: E1796-E1805 [PMID: 26976583 DOI: 10.1073/pnas.1520686113]
- 243 **Stiede K**, Miao W, Blanchette HS, Beysen C, Harriman G, Harwood HJ Jr, Kelley H, Kapeller R, Schmalbach T, Westlin WF. Acetyl-coenzyme A carboxylase inhibition reduces de novo lipogenesis in overweight male subjects: A randomized, double-blind, crossover study. *Hepatology* 2017; **66**: 324-334 [PMID: 28470676 DOI: 10.1002/hep.29246]
- 244 **Inagaki T**. Research Perspectives on the Regulation and Physiological Functions of FGF21 and its Association with NAFLD. *Front Endocrinol (Lausanne)* 2015; **6**: 147 [PMID: 26441837 DOI: 10.3389/fendo.2015.00147]
- 245 **Nies VJ**, Sancar G, Liu W, van Zutphen T, Struik D, Yu RT, Atkins AR, Evans RM, Jonker JW, Downes MR. Fibroblast Growth Factor Signaling in Metabolic Regulation. *Front Endocrinol (Lausanne)* 2016; **6**: 193 [PMID: 26834701 DOI: 10.3389/fendo.2015.00193]
- 246 **Liu X**, Zhang P, Martin RC, Cui G, Wang G, Tan Y, Cai L, Lv G, Li Y. Lack of fibroblast growth factor 21 accelerates metabolic liver injury characterized by steatohepatitis in mice. *Am J Cancer Res* 2016; **6**: 1011-1025 [PMID: 27293995]
- 247 **Mu J**, Pinkstaff J, Li Z, Skidmore L, Li N, Myler H, Dallas-Yang Q, Putnam AM, Yao J, Bussell S, Wu M, Norman TC, Rodriguez CG, Kimmel B, Metzger JM, Manibusan A, Lee D, Zaller DM, Zhang BB, DiMarchi RD, Berger JP, Axelrod DW. FGF21 analogs of sustained action enabled by orthogonal biosynthesis demonstrate enhanced antidiabetic pharmacology in rodents. *Diabetes* 2012; **61**: 505-512 [PMID: 22210323 DOI: 10.2337/db11-0838]
- 248 **Lee JH**, Kang YE, Chang JY, Park KC, Kim HW, Kim JT, Kim HJ, Yi HS, Shong M, Chung HK, Kim KS. An engineered FGF21 variant, LY2405319, can prevent non-alcoholic steatohepatitis by enhancing hepatic mitochondrial function. *Am J Transl Res* 2016; **8**: 4750-4763 [PMID: 27904677]
- 249 **Wu X**, Ge H, Lemon B, Vonderfecht S, Weiszmann J, Hecht R, Gupte J, Hager T, Wang Z, Lindberg R, Li Y. FGF19-induced hepatocyte proliferation is mediated through FGFR4 activation. *J Biol Chem* 2010; **285**: 5165-5170 [PMID: 20018895 DOI: 10.1074/jbc.M109.068783]
- 250 **Luo J**, Ko B, To C, Ling L, Rossi S, DePaoli A, Tian H. P0932: Treatment with NGM282 significantly improves liver histopathology in a mouse model of non-alcoholic steatohepatitis (NASH). *J Hepatol* 2015; **62**: S694 [DOI: 10.1016/S0168-8278(15)31133-8]
- 251 **Hong F**, Chou HI, Friedman SL. Significant Anti-Fibrotic Activity of Cenicriviroc, A Dual CCR2/CCR5 Antagonist, in a Rat Model of Thioacetamide-Induced Liver Fibrosis and Cirrhosis. *Hepatology* 2013
- 252 **Lefebvre DR**, Hashiguchi T, Jenkins H, Nabhan A, Yoneyama H, Friedman S, Wolfgang GH. Anti-Fibrotic and Anti-Inflammatory Activity of the Dual CCR2 and CCR5 Antagonist Cenicriviroc in a Mouse Model of NASH. *Hepatology* 2013; **58**: 221A-222A
- 253 **Lefebvre E**, Moyle G, Reshef R, Richman LP, Thompson M, Hong F, Chou HL, Hashiguchi T, Plato C, Poulin D, Richards T, Yoneyama H, Jenkins H, Wolfgang G, Friedman SL. Antifibrotic Effects of the Dual CCR2/CCR5 Antagonist Cenicriviroc in Animal Models of Liver and Kidney Fibrosis. *PLoS One* 2016; **11**: e0158156 [PMID: 27347680 DOI: 10.1371/journal.pone.0158156]
- 254 **Seki E**, de Minicis S, Inokuchi S, Taura K, Miyai K, van Rooijen N, Schwabe RF, Brenner DA. CCR2 promotes hepatic fibrosis in mice. *Hepatology* 2009; **50**: 185-197 [PMID: 19441102 DOI: 10.1002/hep.22952]
- 255 **Friedman S**, Sanyal A, Goodman Z, Lefebvre E, Gottwald M, Fischer L, Ratzliff V. Efficacy and safety study of cenicriviroc for the treatment of non-alcoholic steatohepatitis in adult subjects with liver fibrosis: CENTAUR Phase 2b study design. *Contemp Clin Trials* 2016; **47**: 356-365 [PMID: 26944023 DOI: 10.1016/j.cct.2016.02.012]
- 256 **Safadi R**, Konikoff FM, Mahamid M, Zelber-Sagi S, Halpern M, Gilat T, Oren R; FLORA Group. The fatty acid-bile acid conjugate Aramchol reduces liver fat content in patients with nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 2014; **12**: 2085-2091.e1 [PMID: 24815326 DOI: 10.1016/j.cgh.2014.04.038]
- 257 **Moon HJ**, Finney J, Ronnebaum T, Mure M. Human lysyl oxidase-like 2. *Bioorg Chem* 2014; **57**: 231-241 [PMID: 25146937]

DOI: 10.1016/j.bioorg.2014.07.003]

- 258 **Barry-Hamilton V**, Spangler R, Marshall D, McCauley S, Rodriguez HM, Oyasu M, Mikels A, Vaysberg M, Ghermazien H, Wai C, Garcia CA, Velayo AC, Jorgensen B, Biermann D, Tsai D, Green J, Zaffryar-Eilot S, Holzer A, Ogg S, Thai D, Neufeld G, Van Vlasselaer P, Smith V. Allosteric inhibition of lysyl oxidase-like-2 impedes the development of a pathologic microenvironment. *Nat Med* 2010; **16**: 1009-1017 [PMID: 20818376 DOI: 10.1038/nm.2208]
- 259 **Morris BJ**. Seven sirtuins for seven deadly diseases of aging. *Free Radic Biol Med* 2013; **56**: 133-171 [PMID: 23104101 DOI: 10.1016/j.freeradbiomed.2012.10.525]
- 260 **Colak Y**, Ozturk O, Senates E, Tuncer I, Yorulmaz E, Adali G, Doganay L, Enc FY. SIRT1 as a potential therapeutic target for treatment of nonalcoholic fatty liver disease. *Med Sci Monit* 2011; **17**: HY5-HY9 [PMID: 21525818]
- 261 **Li L**, Hai J, Li Z, Zhang Y, Peng H, Li K, Weng X. Resveratrol modulates autophagy and NF- κ B activity in a murine model for treating non-alcoholic fatty liver disease. *Food Chem Toxicol* 2014; **63**: 166-173 [PMID: 23978414 DOI: 10.1016/j.fct.2013.08.036]

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Nonalcoholic fatty liver disease: Evolving paradigms

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Abstract

In the last years new evidence has accumulated on nonalcoholic fatty liver disease (NAFLD) challenging the paradigms that had been holding the scene over the previous 30 years. NAFLD has such an epidemic prevalence as to make it impossible to screen general population looking for NAFLD cases. Conversely, focusing on those cohorts of individuals exposed to the highest risk of NAFLD could be a more rational approach. NAFLD, which can be diagnosed with either non-invasive strategies or through liver biopsy, is a pathogenically complex and clinically heterogeneous disease. The existence of metabolic as opposed to genetic-associated disease, notably including "lean NAFLD" has recently been recognized. Moreover, NAFLD is a systemic condition, featuring metabolic, cardiovascular and (hepatic/extra-hepatic) cancer risk. Among the clinico-laboratory features of NAFLD we discuss hyperuricemia, insulin resistance, atherosclerosis, gallstones, psoriasis and selected endocrine derangements. NAFLD is a precursor of type 2 diabetes (T2D) and metabolic syndrome and progressive liver disease develops in T2D patients in whom the course of disease is worsened by NAFLD. Finally, lifestyle changes and drug treatment options to be implemented in the individual patient are also critically discussed. In conclusion, this review emphasizes the new concepts on clinical and pathogenic heterogeneity of NAFLD, a systemic disorder with a multifactorial pathogenesis and protean clinical manifestations. It is highly prevalent in certain cohorts

of individuals who are thus potentially amenable to selective screening strategies, intensive follow-up schedules for early identification of liver-related and extrahepatic complications and in whom earlier and more aggressive treatment schedules should be carried out whenever possible.

Key words: Nonalcoholic fatty liver disease; Biomarkers; Clinical correlates; Diagnosis; Epidemiology; Genetics; Liver histology; Management; Metabolic Syndrome; Pathogenesis; Screening; Type 2 diabetes

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Core tip: Nonalcoholic fatty liver disease (NAFLD) is a pandemic disease. Recent evidence highlights new concepts in clinical and pathogenic heterogeneity of NAFLD, a systemic disorder with a multifactorial pathogenesis and protean clinical manifestations. Other than the classical obese phenotype of NAFLD, a lean though metabolically abnormal variant has been recognized. Simple steatosis is no more considered a benign condition; insulin resistance is necessary but not sufficient for the disease progression, and NAFLD is not only a mere hepatic manifestation of metabolic syndrome, but may forerun the development of metabolic syndrome and cardio-renal complications. Several non-invasive diagnostic tests are now available and new drug treatment options are coming.

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INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) features excess intrahepatic ectopic triglyceride deposition^[1] in patients who are free of competing etiologies of liver disease^[2]. It is the most frequent liver disease and is associated with a wide spectrum of hepatic disorders ranging from simple steatosis to nonalcoholic steatohepatitis (NASH), cirrhosis and hepatocellular carcinoma (HCC)^[3]. Moreover, frequent co-morbidities of NAFLD include specific cardio-renal-metabolic conditions and increased hepatic/extrahepatic cancer risk^[4].

NAFLD is a pandemic disease worldwide, which has been paralleling the ongoing epidemics of obesity, type 2 diabetes (T2D), and metabolic syndrome (MetS)^[4]. The prevalence of NAFLD approaches 25%-30% in the Europe and United States general populations; but this figure surges to 80%-90% in selected cohorts of dysmetabolic individuals^[5-7]. Patients with NAFLD

have an increased risk of premature cardiovascular as well as of liver-related mortality^[8]. Of concern, up to 50% of cases NAFLD-HCC may occur in the absence of cirrhosis, a circumstance which will often worsen the outcome^[9,10].

Knowledge accumulated since 1980, when Ludwig named and described NASH^[11], has challenged most of the paradigms that had been holding the scene over the previous 30 years. For example, NAFLD is now considered a multifaceted complex systemic condition^[12,13] which exhibits a sexual dimorphism^[14] and follows a variable hepatic and extra-hepatic course^[15,16]. In addition, recent evidence suggests that simple steatosis may progress to NASH and advanced liver fibrosis^[17] challenging the previous dogma that "steatosis is a benign condition".

Insulin resistance (IR) is often associated with NAFLD but it is "necessary but not sufficient" for the disease to progress^[18]. Moreover, NAFLD may occur in lean individuals^[19,20] and it is not only a mere "hepatic manifestation of MetS", but may forerun the development of MetS^[21,22]. NAFLD is associated with secretion of diabetogenic hepatokines and inflammatory biomarkers that increase the risk of incident T2D by adversely affecting glucose homeostasis^[23]. Of concern, NAFLD and T2D are linked by a "vicious circle" which carries to accelerated worsening of liver disease^[24], more difficult metabolic control and earlier appearance of T2D complications^[25,26].

The historical "two-hit" pathogenic theory^[27] has been replaced by a multi-factorial model, which more incisively recapitulates the complexity of the pathogenesis of the disease^[28] by emphasizing the myriads of pathways leading to the same hepatic phenotype^[29]. A better definition of the role played by genetics in the development of NAFLD^[30] has led to the notion that, probably, two different NAFLD types exist: NAFLD associated with and NAFLD dissociated from MetS, and that each may differentially affect cardio-metabolic risk^[31,32]. In addition, the declining gradient of risk of NAFLD in Hispanic patient populations compared to non-Hispanic whites and African Americans may potentially be accounted for by polymorphisms in genes that control metabolism^[33]. The impressive amount of data implicating gut microbiota in the development of NAFLD may well illustrate the role of diet in increasing NAFLD risk^[34-36] and significantly impact our understanding of NAFLD pathogenesis and treatment^[4].

Over time, the diagnostic approach has also undergone substantial changes. Non-invasive physical and chemical biomarkers are now available^[37]. Liver biopsy is reserved to selected NAFLD cases in clinical practice and is more extensively performed and repeated to evaluate novel drug agents^[3,4,38,39]. Finally, new histological classifications have recently been proposed^[40] and the pros and cons of each system must be carefully evaluated^[41].

Within the complex frame of pathogenic and clinical heterogeneity of different NAFLD populations

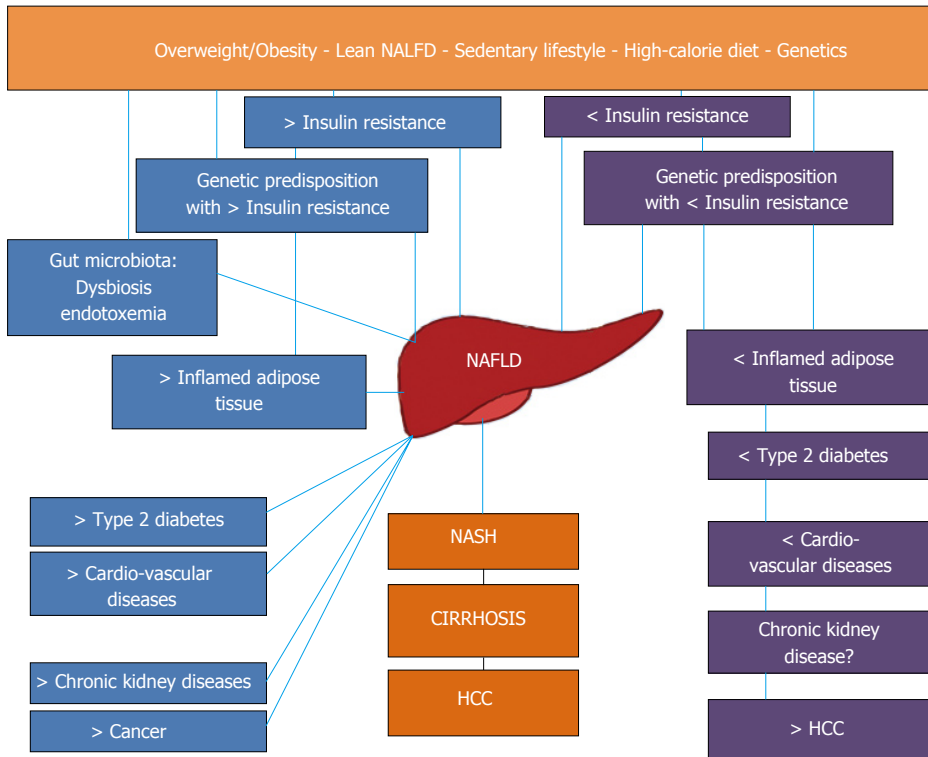


Figure 1 Nonalcoholic fatty liver disease as a pathogenically and clinically heterogeneous condition. Schematic representation of the pathogenic and clinical heterogeneity of different NAFLD populations. Left: “Metabolic” NAFLD is associated with adipose tissue dysfunction and IR and may progress towards hepatic and extrahepatic complications. Right: “Genetic” NAFLD seems to be disconnected from adipose tissue dysfunction and IR, is associated with an increased risk of liver disease progression but is probably spared from extrahepatic complications. NAFLD: Nonalcoholic fatty liver disease; NASH: Nonalcoholic steatohepatitis; HCC: Hepatocellular carcinoma.

synthesized in Figure 1, this review aims at disseminating the most updated paradigms on NAFLD and the possible implications for both clinical practice and research purposes.

NAFLD: TO SCREEN OR NOT TO SCREEN. PRINCIPLES OF EPIDEMIOLOGY

An ongoing debate regards whether any screening campaigns should be ever conducted to identify NAFLD individuals^[38,42]. Those supporting screening would argue that NAFLD patients are prone to excess liver-related morbidity/mortality and cardiovascular risk and thus in need of intensive follow-up schedules; those against would reply that there is no point in identifying NAFLD, a condition for which no licensed drug treatment schedule is available. At any rate, given the overwhelming prevalence of NAFLD in the general population worldwide, any sensitive approach should best be conducted based on our understanding of which specific cohorts of individuals are exposed to the highest risk of disease.

In the general population, the prevalence of NAFLD has been reported to widely range from 6.3% to 51% related to the different population/ethnicity evaluated as well as to the diagnostic methods utilized to assess the amount of intrahepatic fat content^[43].

In particular, the diagnostic technique used to capture cases of NAFLD has undoubtedly affected the results. For example, liver enzymes are far less sensitive than ultrasonographic scanning; in its turn, MR spectroscopy is the most sensitive although this technique is expensive and not universally available^[3]. Based on meta-analytic data, Younossi *et al.*^[6] have reported that, in Europe and in United States, the overall prevalence of NAFLD in the general population approximates 25%. However, there are areas of the world, such as south America and middle East, wherein the average prevalence peaks > 30%; conversely, Africa has the minimum prevalence of NAFLD worldwide^[6]. These impressive data clearly imply that no society worldwide can invest so many resources as to screen the general population for NAFLD. It is of interest, therefore, that modifiers of risk might assist health authorities as well as practicing physician in identifying individuals who are particularly prone to developing NAFLD. For example: male gender, middle age and Hispanic ethnicity, all reinforce the risk of NAFLD and, conversely, young women of Afro-American descent have the lowest risk of this disease^[7]. Further to the above physiological modifiers, a set of metabolic modifiers have also been identified. They include arterial hypertension and the MetS (whose presence doubles the prevalence of NAFLD up to 50% of individuals); obesity, dyslipidemia and T2D^[7]. In

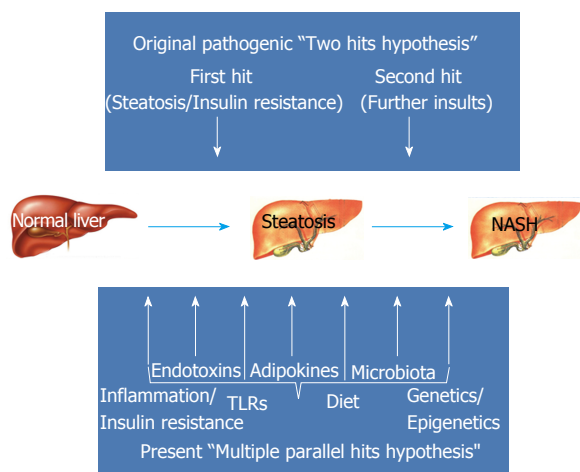


Figure 2 Nonalcoholic fatty liver disease pathogenesis: from two to multiple parallel hits. Schematic representation of the increasing grade of complexity gained in moving from earlier theories^[27] to more updated pathogenic paradigms^[28]. Top: Former "two hits" hypothesis: steatosis, the first 'hit', sensitizes the liver to the second "hits": oxidative stress, endotoxin, ATP depletion and adduct formation^[44]. Bottom: The "multiple hits" hypothesis considers multiple insults acting together on genetically predisposed subjects to induce NAFLD and provides a more accurate explanation of NAFLD pathogenesis. Such hits include IR, hormones secreted from the adipose tissue, nutritional factors, gut microbiota and genetic and epigenetic factors^[28]. NAFLD: Nonalcoholic fatty liver disease; TLRs: Toll-like receptors.

the general population, women are protected from developing NAFLD owing to their hormonal profile and/or set of chromosomes. Conversely, women with T2D are exposed to the same risk of NAFLD as men, indicating that T2D abrogates such a gender-related protection^[7,14].

PATHOGENESIS OF NAFLD: FROM TWO TO MULTIPLE HITS

In recent years, it has increasingly become evident that what had initially been called the NAFLD "two hit hypothesis", in which IR acted as the "first hit" by inducing lipid accumulation in the hepatocytes and increasing the vulnerability of the liver to further insults, referred to as the "second hit", that, in their turn, promoted hepatic injury, inflammation and fibrosis, is probably over-simplistic^[27,44]. Human NAFLD is rather a multi-factorial, non-communicable disease resulting from a complex interaction between multiple environmental and metabolic "hits" and a predisposing genetic background^[45] (Figure 2). A "multiple parallel hits hypothesis" seems more appropriate to recapitulate the complexity of NAFLD pathogenesis^[28], which results from numerous events originating within liver, adipose tissue, gastrointestinal tract and the muscle^[45].

An unhealthy lifestyle, characterized by sedentariness and high-calorie diet, is crucial for NAFLD development and progression. The unbalance between caloric intake and expenditure determines the ex-

pansion of fat depots, which become inflamed and insulin resistant, and release increased amounts of free fatty acids into the bloodstream, leading to ectopic fat accumulation in the liver, skeletal muscles and pancreas^[46]. The hepatic lipid content mainly derives from the increased lipolysis of triglycerides in the adipose tissue and, to a lesser extent, from dietary fats and sugars, and from de novo lipogenesis^[47]. Moreover, the dysfunctional adipose tissue overproduces pro-inflammatory cytokines, such as leptin and resistin, whereas the release of anti-inflammatory adipokines, such as adiponectin, is reduced, so further enhancing NAFLD progression^[48]. The role of intra-hepatocytic accumulation of specific toxic lipids, *via* lipotoxicity and inflammation, has emerged as a key player in the pathogenesis of NASH and its systemic complications^[49]. Saturated fatty acids have been found elevated in NASH patients and induce inflammation and hepatocyte apoptosis through activation of JNK/endoplasmic reticulum stress and oxidative stress/mitochondrial dysfunction^[50]. The accumulation of diacylglycerol and ceramide in the liver impairs hepatic insulin signaling, fuels gluconeogenesis and promotes the development of persisting hyperglycemia and, eventually, T2D^[51].

Unhealthy lifestyle also results in dysbiosis, *i.e.* quantitative and qualitative changes of gut microbiota composition^[52]. In NAFLD an increased Firmicutes/Bacteroidetes ratio and changes in metagenomic-based functional aspects of gut microbiota have all been described^[53]. Gut microbiota may contribute to the development and progression of NAFLD by triggering (both directly or *via* end-products of bacterial metabolism) different signaling pathways, by increasing the efficiency of caloric extraction from the food, and by inducing translocation of bacterial products *via* increased gut permeability^[54-57]. Circulating pathogen-associated molecular pattern (PAMPs) and damage-associated molecular patterns (DAMPs) interact with the family of pattern recognition receptors (toll-like receptors, TLRs) within the liver and induce several pro-inflammatory pathways, including over-expression of cytokines/chemokines, production of reactive oxygen species and activation of the inflammasome^[58,59]. Of note, activation of Nod-like receptor protein 3 inflammasome, upregulated by TLRs in response to the presence of PAMPs and DAMPs, has been associated with a novel cell death mechanism, referred to as hepatocyte pyroptosis^[60].

Several nuclear receptors with their molecular cascades are promising pharmacological targets for the treatment of NAFLD, given their pivotal role in the regulation of energy homeostasis and metabolic pathways. Owing to the development of specific targeted drugs, peroxisome proliferator-activated receptors (PPARs), farnesoid X receptor (FXR) and liver X receptors (LXRs), now deserve full attention. The PPARs superfamily includes PPAR- α , PPAR- β/δ

and PPAR- γ . PPAR- α , mainly expressed in liver and muscle, modulates the rates of fatty acids catabolism, lipogenesis and ketone body synthesis by acting as a nutrient sensor. PPAR- γ , abundantly expressed in the adipose tissue, promotes adipocyte differentiation and storage of triglycerides, and regulates glucose homeostasis. PPAR- β/δ , universally expressed in all organ tissues, regulates glucose and lipoprotein metabolism and exerts an anti-inflammatory role. FXR, mainly expressed in the liver and gut, acts as a sensor of bile acids and mediates the signaling effects exerted by bile acid on glucose and lipid metabolism. LXRs serve as lipid sensors in the liver and participate in regulating the metabolism of cholesterol and fatty acids. The role of nuclear receptors in NAFLD pathophysiology has been comprehensively reviewed elsewhere^[15].

The role of skeletal muscle in the pathogenesis of NAFLD is a matter of increasing research^[23]. Intra-myocellular lipid storage is an early step of ectopic fat accumulation and systemic IR, leads to increased delivery of glucose to the liver, where it becomes substrate for hepatic de novo lipogenesis, and has been associated with NAFLD^[61,62]. Accordingly, sarcopenia, a pathological condition featured by generalized loss of skeletal mass and strength, has recently been proposed as an additional key factor in the pathogenesis of NAFLD^[63-65]. The myokines, such as irisin, which is produced in response to physical activity and exerts several beneficial metabolic effects, may partly account for these associations^[66,67].

NAFLD AS A HETEROGENEOUS DISEASE ENTITY: THE ROLE OF GENETICS

NAFLD is a complex disorder with a variable natural history. Subtle inter-patient genetic variation and environmental factors combine to determine variation in disease progression^[68]. Several clinical features are clues to genetics playing a major role in NAFLD. First, up to a quarter of the general population are at risk of progressive disease; however only a minority will experience associated liver-related morbidity^[69]. Second, both the development and the progression of disease tend to cluster as distinct traits among "NAFLD families"^[7,70]. Third, among non-diabetics, NAFLD will typically affect men more often than women^[7] which, further to hormonal variations, may mirror genetic factors. Fourth, although NAFLD is strictly linked with obesity^[7] not all obese subjects will develop NAFLD and, more importantly, NAFLD can also be found in non-obese individuals^[20]. On these grounds, in the last few years scientific attention has focused on NAFLD genetic features.

Role of *PNPLA3*, *TM6SF2* and *MBOAT7*

Among all analyzed genes patatin-like phospholipase domain-containing protein 3 (*PNPLA3*, also called

adiponutrin) and trans-membrane 6 super family 2 (*TM6SF2*) are deemed to play the most significant role. More recently, membrane bound O-acyltransferase domain-containing 7 gene (*MBOAT7*) has also gained attention. They are all involved in the process of intra-hepatic accumulation of triglycerides.

PNPLA3 gene encodes for a 481-aminoacid protein expressed in the liver and adipose tissues, with function of triglyceride hydrolase and acetyl-CoA-independent transacylase, suggesting a double role of catabolic lipase activity and anabolic lipogenic activity^[71-73]. The nonsynonymous variant rs738409 of *PNPLA3* is a genetic polymorphism characterized by the substitution of isoleucine to methionine at position 148 (I148M). Studies consistently show a strong association between I148M variant and hepatocellular triglycerides accumulation^[74-76]. I148M variant is a loss of function allele that determines an altered hepatic lipid metabolism, eventually leading to hepatic droplet lipid accumulation, lower hepatic VLDL secretion and lower adiponectin production^[77]. Adiponectin is a protein with anti-inflammatory activity and onco-suppressive properties; lower circulating levels of adiponectin can account for the propensity of I148M variant to the progression from NAFLD to NASH and the increased risk of HCC^[77,78].

TM6SF2, also known as KIAA 1926, is a gene at the 19p13.11 locus, with unknown biological function that encodes a protein high expressed in the small intestine, kidney and liver. It is involved in the pathophysiology of NAFLD by modifying hepatic lipid secretion; a decreased hepatic expression of this protein is associated with an increased size and number of hepatocytic lipid droplets^[79]. In particular, studies have shown that a variant of *TM6SF2*, rs58542926 c.449 C>T (E167K), is associated with NAFLD evolution, also with dyslipidemia and cardiovascular risk. The minor variant allele (T) is linked with a reduction of the excretion of hepatic VLDL and a lower level of circulating triglyceride and LDL-cholesterol. Conversely, C-allele is associated with a higher hepatic excretion of VLDL, serum triglyceride and cholesterol. Thus, the minor allele exhibits a higher risk of advanced hepatic disease, though protecting from metabolic and cardiovascular disease^[80]. Conversely, increased metabolic risk is associated with C-allele^[81]. Morris *et al.*^[82], based on their large meta-analysis have hypothesized a correlation with T-allele variant and risk developing T2D, but this theory remains to be confirmed^[82]. Recently, it has been shown that *TM6SF2* (E167K) gene variant is associated with an altered lipidomic profile featuring deficiency of polyunsaturated phosphatidylcholine and excess of polyunsaturated free fatty acids in the liver. This lipidomic profile leads to decreased concentrations of circulating VLDL and increased liver fat content^[83]. Moreover, the same polymorphism has been found to modulate postprandial lipid metabolism, nutrient oxidation and glucose homeostasis in response to dietary fat^[84]. These findings

are compatible with the above reported notion that carriers of TM6SF2 polymorphisms may be specifically exposed to liver-related as opposed to vascular manifestations of NAFLD.

Finally, it has been recently shown that the rs641738 C>T variant in MBOAT7 gene, which is involved in phosphatidylinositol acyl-chain remodeling, increases the risk of developing NAFLD; inflammation and fibrosis^[85,86].

Lean NAFLD

As previously reported in this review, genetic factors play a major role in development and progression of NAFLD. Such a role of genetics is further reinforced when lean NAFLD patients are taken into consideration. It is well-known that NAFLD is often found in obese and dysmetabolic patients. Accordingly, the "Lean NAFLD" phenotype has attracted interest among researchers in as much as it features selectively expanded visceral adiposity though preserving a normal body mass index and, sometimes, even a normal waist circumference. Worryingly, these individuals develop IR and atherogenic dyslipidemia similarly to obese patients and have, therefore, in the past been alluded to as "metabolically obese, normal weight"^[87]. Lean NAFLD cases have been described among different ethnic population, mainly among Asians, who tend to develop the metabolic complications of obesity for BMI values inferior to those of Caucasians^[88,89]. The mechanism(s) through which lean NAFLD develop(s) is/are poorly characterized and the disease goes often under-recognized. Though the role of genetic factors is likely to be more substantial in "lean NAFLD" than that found in "obese NAFLD"^[90], a recent statistically sophisticated study has suggested that both lean and obese NAFLD patients share cardio-metabolic risk commonalities and that the former, despite normal body weight, nonetheless have excess of abdominal adiposity and other MetS features^[19]. Accordingly, also lean NAFLD patients may benefit from increased physical activity which is effective in reducing visceral fat^[91]. Of note, the findings of a very recent study confirm that NAFLD development and progression derive from a complex interplay between genetic and environmental factor, by clearly demonstrating the presence of a synergy between adiposity and the genetic risk of NAFLD conferred by multiple loci^[92].

DIAGNOSTIC STRATEGIES

Diagnosis and staging of NAFLD fully rely on liver biopsy. Histological examination of liver tissue specimens is the gold-standard for quantitating steatosis, diagnosing NASH and staging fibrosis. The last of these three tasks bears major prognostic significance given that fibrosis is associated with long-term liver-related and, probably, cardiovascular outcomes^[93-95]. Over the past two decades, different criteria have been suggested for scoring and staging histological

lesions, and different definitions of NASH have been used^[40,96-99]. Besides discrepancies and subjectivity in interpretation, liver biopsy has additional limitations such as cost, invasiveness and sampling variability which limit the feasibility of this procedure in all the patients with suspected NAFLD and have called for the development of alternative non-invasive diagnostic and staging procedures. Despite some intrinsic and specific drawbacks^[100,101], these non-invasive tools, especially if used in combination or sequentially, may help in restricting the number of and assist in a better selection of potential candidates to liver biopsy^[102].

Histological classification: what's new?

Steatosis, namely a minimal threshold of 5% of hepatocytes containing fat droplets in biopsy specimen, is a prerequisite for the diagnosis of NAFLD. The histological definition of adult NASH is based on a combination of three elementary lesions, *i.e.*, steatosis, hepatocellular ballooning and lobular inflammation^[41,97].

Four main pathologic classifications of NAFLD have been proposed: Matteoni's, Brunt's, Kleiner's classification with the NAFLD Activity Score (NAS) and the recent Fatty Liver Inhibition of Progression (FLIP) algorithm with the Steatosis, Activity, and Fibrosis (SAF) score^[40,96-99].

In 2005 the NASH Clinical Research Network designed and validated a histological feature scoring system that addressed the full spectrum of lesions of NAFLD. Currently, NAS is the most widely used histological classification for NAFLD/NASH and its use is recommended for defining and quantifying disease activity in clinical trials^[98]. NAS, ranging from 0 to 8, is derived from the sum of steatosis (0-3), ballooning (0-2) and lobular inflammation (0-3). However, at least two out of its three components show a high variability among pathologists; and data increasingly suggest that NAS is poorly correlated with metabolic risk factors^[103], is unable to predict fibrosis progression^[104] and to prognosticate liver-related and overall mortality in NAFLD patients^[93,94,105].

More recently, Bedossa and the FLIP Consortium created a simple histological algorithm (FLIP algorithm) based on a scoring system, the SAF score (Steatosis, Activity, Fibrosis) intended for pathologists to reliably diagnose NASH, assess disease severity and limit interobserver variation^[40,99]. The SAF score separately assesses steatosis, activity and fibrosis. Activity, which ranges from 0 to 4, is derived from the combination of the semi-quantitative values of hepatocellular ballooning (0-2) and lobular inflammation (0-2). NASH according to the FLIP algorithm is diagnosed when steatosis, ballooning and lobular inflammation are all scored ≥ 1 . The main discrepancy between NAS and SAF scores is that SAF does not include steatosis in the activity score, owing to the questionable role of steatosis *per se* as a marker of disease progression and prognosis. Moreover, fibrosis stage, the strongest

Table 1 Main features and differences between Nonalcoholic Steatohepatitis Activity score and Steatosis, Activity, and Fibrosis score

	NAS	SAF
Scoring system	Steatosis + Lobular Inflammation + Ballooning: 0-8	Steatosis, Activity (Lobular Inflammation + Ballooning), Fibrosis: S ₀₋₃ , A ₀₋₄ , F ₀₋₄
Details of scoring	<p>Steatosis</p> <p>0: < 5%</p> <p>1: 5%-33%</p> <p>2: 34%-66%</p> <p>3: > 67%</p> <p>Lobular inflammation</p> <p>0:00</p> <p>1: < 2/20X</p> <p>2: 2-4/20X</p> <p>3: > 4/20X</p> <p>Ballooning (number of ballooned hepatocytes)</p> <p>0: None</p> <p>1: Few</p> <p>2: Many</p> <p>Fibrosis stage</p> <p>F0: None</p> <p>F1a: zone 3 perisinusoidal, delicate</p> <p>F1b: perisinusoidal, dense</p> <p>F1c: portal only</p> <p>F2: 1a or 1b + portal</p> <p>F3: bridging</p> <p>F4: cirrhosis</p>	<p>Steatosis</p> <p>0: < 5%</p> <p>1: 5%-33%</p> <p>2: 34%-66%</p> <p>3: > 67%</p> <p>Lobular inflammation</p> <p>0:00</p> <p>1: < 2/20X</p> <p>2: ≥ 2/20X</p> <p>Ballooning (size and shape of hepatocytes)</p> <p>0: normal hepatocytes</p> <p>1: clusters, reticulated cytoplasm</p> <p>2: enlarged hepatocytes</p> <p>Fibrosis stage</p> <p>F0: 0</p> <p>F1: zone 3 perisinusoidal (all), or portal only</p> <p>F2: zone 3 + portal</p> <p>F3: bridging</p> <p>F4: cirrhosis</p>

NAS: NASH activity score; SAF: Steatosis, Activity, and Fibrosis score; NASH: Nonalcoholic steatohepatitis.

predictor of outcomes in chronic liver disease, was not included in the NAS. This novel histological classification has shown excellent applicability and reproducibility among pathologists for the assessment of liver lesions in morbidly obese patients and in NAFLD patients with MetS^[40,99]. However, it will have to undergo further validation in clinical practice. Table 1 shows the main features of NAS and SAF scores and highlights the main differences between these scoring systems.

Role of imaging techniques

Imaging methods are the most commonly used non-invasive tools for the diagnosis of steatosis. The main advantages of ultrasonography are its safety, low cost, wide availability and the overall scanning of abdominal organs. Traditionally, ultrasonography has been considered a technique with a low sensitivity in identifying fatty liver infiltration when less than 30% of hepatocytes are steatotic. Of note, a recent study has demonstrated that ultrasonography, especially when implemented with standardized measurements and semi-quantitative scores, is sensitive for an amount of steatosis as low as 10%^[106] and may predict metabolic derangements and liver histology changes^[106-108]. Further studies are necessary to confirm these novel findings. The main limitations of ultrasonography are its inability in differentiating steatosis from fibrosis, the issues with morbid obese individuals, and its operator and machinery dependency.

The controlled attenuation parameter (CAP) is a new promising screening method that measures ultrasound attenuation in the liver using signals obtained

by the Vibration Controlled Transient Elastometry (TE) (Fibroscan®). Interestingly, CAP appears to be more sensitive in detecting lesser degrees of steatosis compared to other widely available imaging methods. Moreover, being coupled with liver stiffness measurements, it has the advantage of simultaneously estimating liver fibrosis^[108-110]. However, the underlying disease, BMI and T2D may affect the findings and thus must be taken into consideration in the interpretation of CAP results^[111].

Over the last 10 years, several ultrasound elastographic techniques have been implemented, including TE, acoustic radiation force impulse (ARFI) elastography, 2D shear-wave elastography and real-time strain elastography. TE, is the most studied elastographic method for the estimation of liver fibrosis in chronic liver disease; in NAFLD, TE can be used to confidently rule out severe fibrosis and cirrhosis (with a nearly 90% negative predictive value) and for monitoring disease progression^[11,112]. Of note, the amount of steatosis has emerged as a significant factor affecting the performance of liver stiffness measurements with TE^[112,113]. For this reason, it has been suggested that taking into account CAP values may improve the prediction of liver fibrosis by TE^[113].

Magnetic resonance imaging techniques, and particularly magnetic resonance spectroscopy, are able to accurately quantify liver fat content; the advantages include high sensitivity for assessing small amounts of intrahepatic triglycerides and sampling a large liver volume^[114]. More recently, magnetic resonance elastography and multiparametric magnetic

resonance offer the possibility of performing a thorough characterization of liver tissue by assessing steatosis, fibrosis and haemosiderosis at the same time^[115,116], and have been shown to be able to predict clinical outcomes in patients with various chronic liver disease, notably including NAFLD^[117]. Cost and limited availability, however, restrict their use to research purposes.

Performance and limitations of biomarkers

Several serum biomarker panels have been developed for predicting steatosis, diagnosing NASH and estimating fibrosis^[118] which are, in theory, the preferred diagnostic tool for large-scale screening and epidemiological studies on NAFLD^[4,38]. For example, Bedogni's Fatty Liver Index (FLI) has been largely used to capture steatosis in epidemiological studies^[119]. The strength of FLI as a surrogate marker comes from the demonstration that it can predict clinical outcomes related to the MetS, such as risk of T2D, accelerated atherosclerosis and cardiovascular risk, as well as hepatic, cardiovascular and cancer-related mortality^[120,121]. While clinically relevant, these associations might be conveyed independently of steatosis *per se* since FLI incorporates variables that are part of the MetS phenotype^[101]. In fact, recent studies have found only a limited correlation between FLI and the amount of liver fat *per se*, as defined by liver biopsy or magnetic resonance spectroscopy^[101,122,123]. As for the FLI, other steatosis biomarkers, such as the NAFLD liver fat score, the Hepatic Steatosis Index and the Lipid Accumulation Product, generally show good performance in identifying steatosis, though they are unable to predict liver fat content and are confounded by hepatic histological changes other than steatosis, mainly necro-inflammation and fibrosis^[101,123].

With regard to the possibility of non-invasively differentiating NASH from simple steatosis, several biomarkers (acute-phase proteins, cytokines, markers of oxidative stress and apoptosis, as is the case of cytokeratin-18 fragments) and multiple complex models have been investigated, but the majority of them lack external validation, have only been tested among selected populations and have been inconsistent in their performance of detecting NASH^[102,118].

Several simple non-invasive fibrosis scoring systems, notably including NAFLD Fibrosis Score and Fib-4 can, however, reliably exclude advanced fibrosis in a high proportion of NAFLD patients, allowing a more targeted use of liver biopsy^[124]. Their use in clinical practice is recommended by updated hepatological guidelines^[4,38], and they reliably predict liver-related and cardiovascular complications and death in NAFLD patients^[125-127]. However, they were principally developed and validated in patients aged < 65 years of age, hence their cut-offs need to be adapted in the elderly^[128]. It has been shown that the combination or the sequential use of different scoring systems or different non-invasive techniques (TE plus biomarker)

improve the diagnostic accuracy in detecting severe liver fibrosis, thus further reducing the need for liver biopsy^[129,130].

Recently, several miRNAs^[131], epigenetic biomarkers^[132], lipidomic^[133,134], metabolomics^[135] and proteomic^[136] profiles have been found differently expressed in NAFLD patients and could be used as surrogate markers of progressive liver injury. Probably, a multi-domain, multi-component approach which integrates variables from a diverse set of data sources deriving from phenotypic, genomic, lipidomic, metabolomic and proteomic domains will be the avenue for future investigations in the noninvasive assessment of NAFLD^[136,137].

NAFLD: CLINICO-LABORATORY FEATURES

Usually asymptomatic, NAFLD is a phenotypically polymorphic disease which, owing to its systemic nature, has a variable clinical presentation, a multitude of potentially associated disease and a rich spectrum of laboratory features^[4,12,138]. Understanding this remarkable phenotypic polymorphism is key in guiding the diagnostic process in the individual patient.

Most patients will seek medical advice on the grounds of otherwise inexplicably deranged liver tests/ultrasonographic findings compatible with fatty changes. Collectively, these laboratory/ultrasonographic abnormalities tend to be often discovered either during routine medical check-up performed by asymptomatic individuals or by patients who have nonspecific and causally unrelated complaints^[139,140].

Insulin resistance, hyperuricemia and atherogenic dyslipidemia

The association of NAFLD with IR and its close connection with the MetS was independently reported by three unrelated groups of investigators as early as 1999^[141-143]. In particular, Marchesini *et al.*^[141] by comparing 46 patients with NAFLD to 92 age and sex-matched healthy controls with the homeostasis model assessment method found that NAFLD was associated with IR and hyperinsulinemia even in lean subjects with normal glucose tolerance. Consistently, Cortez-Cortez-Pinto *et al.*^[142] in 30 patients with NAFLD found that 80% were either obese or dyslipidemic; 50% had arterial hypertension and 33% T2D; moreover, glucose metabolism was altered in 69% and hyperinsulinemia and hyperleptinemia were common. Along the same line, Lonardo, based on his systematic search of the available literature, concluded that fatty liver most commonly affected middle-aged men with obesity, altered glucose metabolism, hyperlipidemia, and hypertension^[143].

Over time, these pioneer studies suggesting an intimate connection of NAFLD with IR and the MetS have extensively been confirmed and carried further.

For example Ballestri *et al.*^[144] have recently shown that HOMA-IR, a widely used index of IR, along with serum uric acid concentrations, is an independent predictor of both steatosis and ballooning; waist circumference predicts both ballooning and hepatic fibrosis^[144]. Moreover, the same group of investigators, also found that baseline NAFLD predicts the development of incident T2D and MetS over a median 5-year follow-up^[22]. Taken collectively, these findings strongly support the notion that NAFLD should no longer be considered a mere "hepatic manifestation of the MetS" but rather, and more appropriately, both a precursor and a consequence of the MetS^[21,31,145-147].

Lonardo *et al.*^[139] were first in reporting that both fasting insulin (a sensitive marker of IR) and uric acid were independent predictors of NAFLD. These findings have been independently confirmed by several studies^[148-152], including two meta-analytic reviews^[153,154]. Of interest, although serum uric acid concentrations are associated with indices of IR, both parameters are independently associated with NASH, at least in children and adolescents^[155], suggesting that each probably carries an independent contribution to the development of progressive liver disease.

NAFLD is typically characterized by atherogenic dyslipidemia featuring larger and triglyceride over-enriched circulating very-low-density lipoproteins (VLDLs), small dense low-density lipoproteins (LDLs) and low and dysfunctional high-density lipoproteins (HDLs). Of note, when NAFLD progresses to severe fibrosis and cirrhosis, dyslipidemia will apparently improve, probably owing to failing hepatic synthetic capacity^[144,156-159].

Liver tests, parameters of iron metabolism and non-organ specific autoantibodies

Liver tests are typically normal or mildly elevated in NAFLD. Gamma-glutamyltransferase (GGT) may be slightly elevated and is increasingly recognized as a marker of metabolic disturbances and cardiovascular risk^[160]. Aminotransferases do not identify progressive disease, given that patients with normal liver enzymes are spared neither from NASH nor significant fibrosis^[161].

The diagnostic process followed to investigate a suspect liver disease may include the determination of parameters of iron metabolism and non-organ specific autoantibodies (NOSA). Elevated serum ferritin, closely associated with hepatic iron deposition, is common in NAFLD and is strongly correlated with IR, more advanced disease and increased mortality^[162-164]. Whether therapeutic strategies aimed at correcting iron metabolism may be beneficial in NAFLD remains controversial.

It is important to highlight that the positivity of NOSA, in the appropriate clinical and epidemiological context, is compatible with NAFLD. Loria *et al.*^[165], in their pilot study, were first in reporting that NOSA

positivity in NAFLD was more prevalent than in the general population and that high-titer ANA was associated with IR. Bringing this line of research further, the clinical research network, in a multicenter study including a total of 864 NAFLD patients found that NOSA are frequently positive in NAFLD in the absence of autoimmune hepatitis and their occurrence is not associated with more advanced histological features^[166].

NAFLD AS A MULTISYSTEM DISEASE: CARDIO-METABOLIC AND CANCER RISK

As early as 1995, was it first suggested that NAFLD was a systemic condition with a specific cardio-metabolic involvement^[167], a notion which is now universally accepted^[12,168].

In particular, our understanding of the close and mutual relationship of NAFLD with the MetS has evolved and the bidirectional relationship among the two conditions is now largely acknowledged (Figure 3)^[4,31,146,147]. IR, in its turn, will drive the course of liver disease to progressively fibrosing liver disease culminating with cirrhosis, but also promote the development of T2D^[169]. NAFLD almost doubles the risk of incident T2D^[22]. However, NAFLD patients usually die of extra-hepatic causes, frequently for cardiovascular diseases, which underpins the importance to early diagnose and aggressively treat CVD risk factors, as extensively discussed elsewhere^[170]. A meta-analysis of 27 cross-sectional studies has shown a robust association between NAFLD and several markers of subclinical atherosclerosis burden, such as an increased risk, independent of classical CVD risk factors, of coronary artery calcification, carotid intima-media thickness, impaired flow-mediated vasodilatation and arterial stiffness^[171]. Moreover, studies have shown that NAFLD is also associated with increased risk of atrial fibrillation and aortic valve sclerosis^[172-175].

While cardiovascular mortality is the leading cause of death in NAFLD patients, malignancies, mainly affecting the gastrointestinal tract (liver, colon, esophagus, stomach, and pancreas) and extra-intestinal sites (kidney in men, and breast in women) rank second^[176,177]. The association between NAFLD and colorectal cancer (CRC) has been investigated: some studies have shown a higher prevalence of CRC in NAFLD population than patients without, and this association is increased in presence of NASH^[178].

NAFLD AND T2D: A DANGEROUS COMBINATION

T2D accounts for more than 90% of those 415 million people worldwide who live with diabetes; T2D may lead to micro-macrovascular complications that cause profound sufferings and put a heavy burden on health-care systems worldwide^[179].

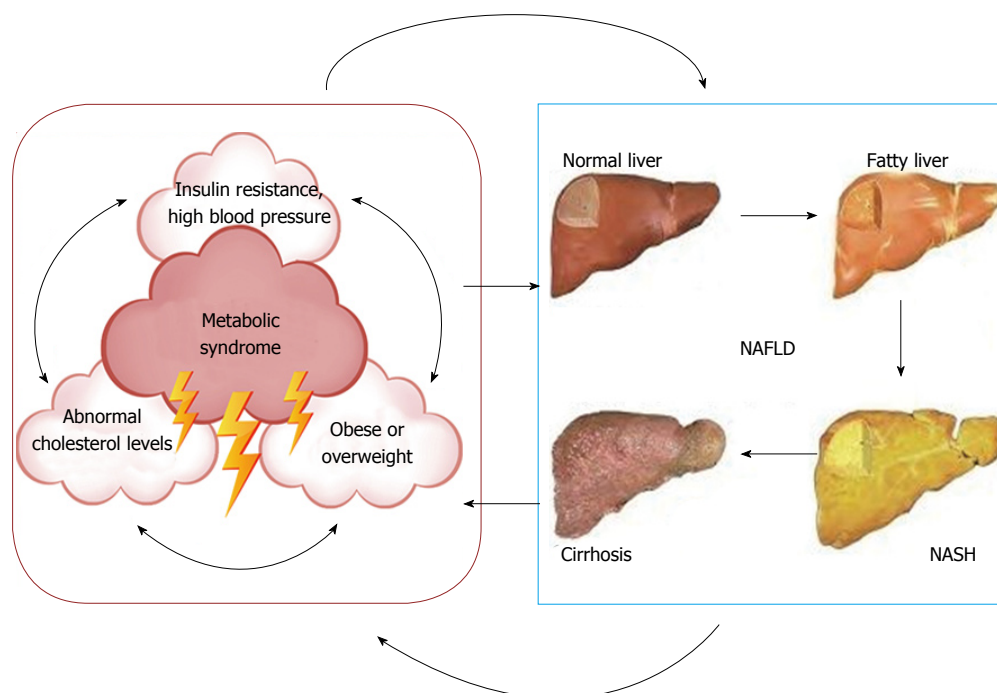


Figure 3 The closed loop nonalcoholic fatty liver disease-metabolic syndrome circuit. Schematic representation illustrating the mutual cause-and-effect relationship of NAFLD with the Metabolic syndrome^[4,31,146,147]. NAFLD: Nonalcoholic fatty liver disease; NASH: Nonalcoholic steatohepatitis.

The common soil and the mutual relationship

The prevalence of NAFLD is increased among those with T2D and T2D rescinds the typical gender dimorphism of NAFLD^[7,180,181]. Conversely, NAFLD is associated with a 2-to-5-fold risk of developing T2D after adjustment for several metabolic confounders^[22,169,182].

Common features in the epidemiology, risk factors and natural history of NAFLD and T2D can be identified: overweight and obesity^[183-185]; a hypercaloric diet coupled with a sedentary behavior leading to weight gain^[186,187]; IR and MetS components, including combined dyslipidemia^[147,188].

As previously highlighted, NAFLD is not only a consequence, but also a cause of T2D and MetS generating a "vicious circle" and worsens the course of T2D^[21,24,31,189]. It is mandatory to interrupt this circle and in this regard, it is encouraging that even transient remission/improvement of NAFLD is independently associated with reduced incidence of T2D^[190,191].

The consequences of the association

T2D and NAFLD interact in enhancing the risk of promoting endothelial dysfunction, atherosclerosis, cardiovascular diseases, chronic kidney disease and advanced retinopathy^[21,168,192,193]. The coexistence of T2D and NAFLD predicts the development of hepatic fibrosis^[194-198]. Treatment of NAFLD, by affecting those pathogenic factors linking fatty liver with atherosclerosis (hence the so called "atherogenic liver") was predicted to be able to reduce cardiovascular risk a few years ago^[199]. Consistent with such a prediction, a recent study reported that reduced CIMT progression

in the entire cohort of 92 patients was independently associated with decreased liver fat content assessed with magnetic resonance spectroscopy^[200].

T2D and NAFLD are also associated with an increased incidence and mortality from several cancer types and, similarly, mortality for malignancies is the second cause of mortality in patients with NAFL^[178,201-203]. Similar to NAFLD, T2D enhances the risk of developing HCC and may aggravates HCC outcomes^[204-206].

NAFLD in those with T2D will follow a worse prognosis *via* hastened progression to NASH, liver cirrhosis and its complications and increased risk of developing NAFLD-HCC^[43,207-210]. Specularly, concurrent NAFLD will often worsen the course of T2D^[211].

Diagnostic issues

An early diagnosis and treatment of NAFLD may potentially increase life expectancy of T2D patients^[209]. Conversely, in subjects with NAFLD, periodical screening for T2D is encouraged, given that poor metabolic control will lead to progressive liver disease which, in its turn, is associated with enhanced cardiometabolic risk^[38,193,212].

NAFLD: A LARGE SPECTRUM OF CLINICAL ASSOCIATIONS

An impressively heterogeneous spectrum of clinical conditions have been associated with NAFLD spanning from carotid atherosclerosis through polycystic ovarian syndrome^[23]. Some will be shortly reviewed here; others have recently been covered in detail

elsewhere^[4,213,214].

Atherosclerosis and gallstones

A pioneer study by Lonardo *et al.*^[215] evaluated the triangular association of NAFLD, gallstones and atherosclerosis. A decade later, we now know that the most severe NAFLD forms are strongly associated with excess cardiovascular mortality, probably as a direct/indirect result of hepatic fibrosis^[193,212]. Concerning the NAFLD-gallstone duo, a systematic review and meta-analysis conducted in 12 observational studies (9 cross-sectional studies, 2 cohort studies and 1 case-control study) found that gallstones are significantly associated with NAFLD^[216]. Further studies should be conducted regarding the underlying mechanism of this association. However, it is now clear that bile acids are no longer “a detergent which facilitates the absorption of fatty foodstuffs” such as it was deemed in the past; but rather, they must be regarded as signaling molecules, which coordinately regulate metabolism and inflammation acting not only in entero-hepatic tissues, but in peripheral organs as well^[217].

Psoriasis

This line of research was almost anecdotally inaugurated by a report of three cases from the POLI.ST.E.N.A study^[218]. Several systematic studies have subsequently confirmed this novel association and a recent systematic review and meta-analysis supports an association between psoriasis and NAFLD suggesting that screening for NAFLD may be warranted among those with psoriasis^[219].

Endocrine derangements

Again, this topic was originally initiated by pioneer clinical observations. First, it was reported that endocrine derangements may eventually be conducive to NAFLD as a result of hormones being master regulators of body fat distribution and cell metabolism^[220]. Next, endocrine pathways specifically leading to NASH were identified^[221]. Among these, hypothyroidism has been associated to the whole NAFLD spectrum, steatosis to NASH-HCC^[156, 221-224].

A large Rotterdam population-based survey, aimed at investigating the association between variations in thyroid function and NAFLD, prospectively, in 9419 adults with a median follow-up time of 10 years; the presence of fibrosis in those with NAFLD was assessed with TE^[225]. Data have shown that, compared to euthyroid individuals, hypothyroid ones were exposed to a higher risk of NAFLD and of clinically relevant fibrosis; consistently, on the opposite side of the spectrum, hyperthyroidism was associated with a protection from developing NAFLD^[225]. These findings may potentially disclose novel treatment strategies.

and physical exercise are the mainstay of treatment of NAFLD, a condition no drugs are specifically licensed for^[4,38].

Diet

A 3%-5% weight loss will improve steatosis; however, 5%-7% reductions are necessary to decrease hepatic inflammatory changes and 7%-10% to obtain remission of NAFLD/NASH and regression of fibrosis. Calorie restriction *per se* rather than specific diets are deemed to be beneficial^[226]. In particular, despite emphasis is usually put in restricting dietary carbohydrates, moderate-carbohydrate diets do not induce a more profound reduction in liver enzymes (a non-validated surrogate index of NAFLD severity) than low/moderate-fat diets^[227].

Physical exercise

Exercise effectively reduces visceral and perhaps liver fat. A recent meta-analysis found that aerobic exercise but not resistance training exercise was effective in reducing visceral fat in overweight/obese adults with T2D^[228]. The benefits of exercise in reducing intrahepatic triglyceride content will typically occur with minimal or no body weight loss^[227,229], suggesting that diet and physical exercise will improve NAFLD and should be associated whenever possible given that they act *via* different pathways. Indeed, diet combined with exercise is associated with improvement in NAFLD activity score^[227].

Drugs

Drug treatment (better if within the frame of a clinical trial) should be reserved to those NASH individuals who do not respond to lifestyle changes. Drugs used to treat concurrent metabolic disorders (e.g., statins and antidiabetics) should be differentiated from drugs aimed at treating NASH *per se*.

Statins, glitazones and liraglutide belong to the first group. Our appreciation of the role of statins in hepatology has undergone a dramatic evolution from under-prescription based on fears of hepatotoxicity^[230] to their identification as potentially protectors from the full spectrum of liver damage in NASH, notably including liver fibrosis and HCC^[231-238].

As regards the oral antidiabetic agents glitazones, a meta-analysis of five eligible randomized controlled trials showed that these drugs improved neither fibrosis nor IR; however, lobular inflammation decreased in NASH patients who received TZD treatment^[239]. However, a more recent meta-analytic review was specifically aimed at investigating the role of thiazolidinediones in advanced hepatic fibrosis in those with NASH analyzed 8 RCTs (5 evaluating pioglitazone and 3 evaluating rosiglitazone) enrolling 516 patients with biopsy-proven NASH for a duration of 6 to 24 mo. Among all studies combined, thiazolidinedione therapy was associated with improved advanced fibrosis, fibrosis of any stage

NAFLD: PRINCIPLES OF TREATMENT

It is widely accepted that lifestyle changes, namely diet

Table 2 Drugs useful in treating nonalcoholic steatohepatitis - A neck-to-neck comparison of findings from two different meta-analytic studies

	Sawangjit <i>et al</i> ^[244]	Singh <i>et al</i> ^[243]
Fibrosis	OCA ¹ improves fibrosis. PTX ² , TZDs plus Metformin ² , TZDs plus Losartan ² might potentially be effective.	PTX ¹ and OCA ¹ improve fibrosis
Resolution of NASH	TZDs ¹ and Vitamin E ¹ use is associated with resolution of NASH	Vitamin E ¹ , TZDs ¹ , and OCA ¹ improve ballooning degeneration
NAS	PTX ¹ , OCA ¹ , vitamin E ¹ , and TZDs ¹ improve NAS	
Steatosis		PTX ¹ , Vitamin E ² , TZD ¹ and OCA ² significantly improve steatosis
Lobular inflammation		PTX ¹ , TZDs ¹ and OCA ² significantly improve lobular Inflammation

¹High quality of evidence; ²Moderate quality of evidence. NASH: Nonalcoholic steatohepatitis; NAS: NASH activity score; OCA: Obeticholic acid; PTX: Pentoxifylline; TZDs: Thiazolidinediones.

and NASH resolution. Analyses restricted to RCTs enrolling patients without diabetes yielded similar results. All effects were accounted for by pioglitazone use^[240]. Weight gain and lower limb edema occurred more frequently with thiazolidinedione therapy^[240]. These novel data indicate that pioglitazone improves advanced fibrosis in NASH, even in patients without diabetes.

A phase 2 randomized study conducted on 52 patients (26 per arm) tested the efficacy of liraglutide (1.8 mg/daily *via* subcutaneous route) vs placebo. The use of liraglutide was associated with a 39% NASH resolution rate vs 9% in the placebo arm (RR = 4.3, 95%CI: 1.0-17.7, *P* = 0.019). This study was funded by the manufacturers of liraglutide^[241].

A few drugs, such as vitamin E and obeticholic acid (OCA) have specifically been tested against NASH. Based on a meta-analysis of five RCTs vitamin E significantly reduced liver enzymes (AST, ALT, ALP); steatosis, inflammation and hepatocellular ballooning compared to the control group. Vitamin E treatment in adult NASH was also associated with reduction of fibrosis^[242]. Concerns have been raised, however, on the long-term safety of Vitamin E and the lack of data in patients with T2D and cirrhosis. A network meta-analysis found that pentoxifylline and OCA improve fibrosis, and vitamin E, TZDs, and OCA improve ballooning degeneration in patients with NASH^[243]. A subsequent network meta-analysis substantially confirmed these findings by showing that OCA significantly improves fibrosis, while vitamin E and TZDs result in a significant increase in NASH resolution^[244]. Table 2 compares the findings from the above two meta-analytic studies^[243,244].

Thinking out of the box. Innovative treatment strategies

Further to the above discussed notions on the relationship of the entire NAFLD spectrum with endocrine and metabolic derangements, a line of innovative treatment options can be envisaged. For example, administration of thyroid receptor agonists (GC-1 and KB2115) improves NAFLD in a ob/ob mouse model and a xantine oxidase inhibitor (febuxostat) antagonizes the development of disease in a rodent NASH model^[245,246].

Also our understanding of the role of nuclear receptors in the pathogenesis and natural course of NAFLD may lead to novel treatments^[15]. The above-mentioned OCA exerts its effects by agonizing FXR. Elafibranor, a dual PPAR- α/δ agonist, has been shown to resolve NASH without fibrosis worsening and to improve the cardiometabolic profile in a recent phase 2 randomized trial of patients with NASH^[247]. Oltipraz is a synthetic inhibitor of LXR- α whose antisteatotic properties have previously been shown in a mouse model. A recent phase 2 randomized controlled trial found that, in patients with NAFLD, a 24-wk course of oltipraz significantly reduced intrahepatic fat content assessed with magnetic resonance spectroscopy in a dose-dependent manner; however IR, liver enzymes, lipids and cytokines were unaltered^[248].

CONCLUSION

NAFLD is an emerging multifaceted systemic disease with a heavy epidemiological burden. Further to hepatic risks, NAFLD also affects cardiovascular, metabolic, renal and endocrine systems as well as cancer development with a significant hepatic and extra-hepatic impact on morbidity and mortality^[4]. The bi-directional relationship between NAFLD and MetS-T2D is an important emerging topic underpinning the mutual influence and the need to interrupt the *vicious circle* associating liver and dysmetabolism. However, we now know that IR is a necessary though not sufficient requisite of the development of NAFLD and that gender, genetic background and microbiota also play a major role^[4,14]. It is anticipated that pathogenic complexity and multi-factoriality will have to be translated into more finely tailored diagnostic and treatment strategies. This implies that "ad hoc" studies are necessary to more precisely identify the individual target populations to be addressed. Presently, lifestyle changes effected *via* dietary restrictions and physical exercising remain the only recommended treatment of NAFLD; in all overweight/obese patients, weight loss is mandatory^[4]. However, given that only a small minority of such individuals will succeed in maintaining their ideal body weight, and that no currently marketed

drugs are specifically licensed for NASH, data of ongoing studies are eagerly awaited to offer these numerous and heterogeneous patients effective and safe tailored drug treatment options.

REFERENCES

- Petäjä EM, Yki-Järvinen H. Definitions of Normal Liver Fat and the Association of Insulin Sensitivity with Acquired and Genetic NAFLD-A Systematic Review. *Int J Mol Sci* 2016; **17**: pii: E633 [PMID: 27128911 DOI: 10.3390/ijms17050633]
- Loria P, Adinolfi LE, Bellentani S, Bugianesi E, Grieco A, Fargion S, Gasbarrini A, Loguercio C, Lonardo A, Marchesini G, Marra F, Persico M, Prati D, Baroni GS; NAFLD Expert Committee of the Associazione Italiana per lo studio del Fegato. Practice guidelines for the diagnosis and management of nonalcoholic fatty liver disease. A decalogue from the Italian Association for the Study of the Liver (AISF) Expert Committee. *Dig Liver Dis* 2010; **42**: 272-282 [PMID: 20171943 DOI: 10.1016/j.dld.2010.01.021]
- Nascimbeni F, Pais R, Bellentani S, Day CP, Ratziu V, Loria P, Lonardo A. From NAFLD in clinical practice to answers from guidelines. *J Hepatol* 2013; **59**: 859-871 [PMID: 23751754 DOI: 10.1016/j.jhep.2013.05.044]
- Italian Association for the Study of the Liver (AISF). AISF position paper on nonalcoholic fatty liver disease (NAFLD): Updates and future directions. *Dig Liver Dis* 2017; **49**: 471-483 [PMID: 28215516 DOI: 10.1016/j.dld.2017.01.147]
- Bellentani S. The epidemiology of non-alcoholic fatty liver disease. *Liver Int* 2017; **37** Suppl 1: 81-84 [PMID: 28052624 DOI: 10.1111/liv.13299]
- Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Metanalytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016; **64**: 73-84 [PMID: 26707365 DOI: 10.1002/hep.28431]
- Non-alcoholic Fatty Liver Disease Study Group, Lonardo A, Bellentani S, Argo CK, Ballestri S, Byrne CD, Caldwell SH, Cortez-Pinto H, Grieco A, Machado MV, Miele L, Targher G. Epidemiological modifiers of non-alcoholic fatty liver disease: Focus on high-risk groups. *Dig Liver Dis* 2015; **47**: 997-1006 [PMID: 26454786 DOI: 10.1016/j.dld.2015.08.004]
- Calzadilla Bertot L, Adams LA. The Natural Course of Non-Alcoholic Fatty Liver Disease. *Int J Mol Sci* 2016; **17**: pii: E774 [PMID: 27213358 DOI: 10.3390/ijms17050774]
- Giannini EG, Marabotto E, Savarino V, Trevisani F, di Nolfo MA, Del Poggio P, Benvegnù L, Farinati F, Zoli M, Borzio F, Catuelli E, Chiaramonte M; Italian Liver Cancer (ITALICA) Group. Hepatocellular carcinoma in patients with cryptogenic cirrhosis. *Clin Gastroenterol Hepatol* 2009; **7**: 580-585 [PMID: 19418607 DOI: 10.1016/j.cgh.2009.01.001]
- Piscaglia F, Svegliati-Baroni G, Barchetti A, Pecorelli A, Marinelli S, Tiribelli C, Bellentani S; HCC-NAFLD Italian Study Group. Clinical patterns of hepatocellular carcinoma in nonalcoholic fatty liver disease: A multicenter prospective study. *Hepatology* 2016; **63**: 827-838 [PMID: 26599351 DOI: 10.1002/hep.28368]
- Ludwig J, Viggiano TR, McGill DB, Oh BJ. Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. *Mayo Clin Proc* 1980; **55**: 434-438 [PMID: 7382552]
- Petta S, Valenti L, Bugianesi E, Targher G, Bellentani S, Bonino F; Special Interest Group on Personalised Hepatology of the Italian Association for the Study of the Liver (AISF); Special Interest Group on Personalised Hepatology of Italian Association for Study of Liver AISF. A "systems medicine" approach to the study of non-alcoholic fatty liver disease. *Dig Liver Dis* 2016; **48**: 333-342 [PMID: 26698409 DOI: 10.1016/j.dld.2015.10.027]
- Hardy T, Oakley F, Anstee QM, Day CP. Nonalcoholic Fatty Liver Disease: Pathogenesis and Disease Spectrum. *Annu Rev Pathol* 2016; **11**: 451-496 [PMID: 26980160 DOI: 10.1146/annurev-pathol-012615-044224]
- Ballestri S, Nascimbeni F, Baldelli E, Marrazzo A, Romagnoli D, Lonardo A. NAFLD as a Sexual Dimorphic Disease: Role of Gender and Reproductive Status in the Development and Progression of Nonalcoholic Fatty Liver Disease and Inherent Cardiovascular Risk. *Adv Ther* 2017; **34**: 1291-1326 [PMID: 28526997 DOI: 10.1007/s12325-017-0556-1]
- Ballestri S, Nascimbeni F, Romagnoli D, Baldelli E, Lonardo A. The Role of Nuclear Receptors in the Pathophysiology, Natural Course, and Drug Treatment of NAFLD in Humans. *Adv Ther* 2016; **33**: 291-319 [PMID: 26921205 DOI: 10.1007/s12325-016-0306-9]
- Lonardo A, Sookoian S, Chonchol M, Loria P, Targher G. Cardiovascular and systemic risk in nonalcoholic fatty liver disease - atherosclerosis as a major player in the natural course of NAFLD. *Curr Pharm Des* 2013; **19**: 5177-5192 [PMID: 23432668]
- Singh S, Allen AM, Wang Z, Prokop LJ, Murad MH, Loomba R. Fibrosis progression in nonalcoholic fatty liver vs nonalcoholic steatohepatitis: a systematic review and meta-analysis of paired-biopsy studies. *Clin Gastroenterol Hepatol* 2015; **13**: 643-654.e1-e9; quiz e39-e40 [PMID: 24768810 DOI: 10.1016/j.cgh.2014.04.014]
- Lonardo A, Bellentani S, Ratziu V, Loria P. Insulin resistance in nonalcoholic steatohepatitis: necessary but not sufficient - death of a dogma from analysis of therapeutic studies? *Expert Rev Gastroenterol Hepatol* 2011; **5**: 279-289 [PMID: 21476922 DOI: 10.1586/egh.11.19]
- Sookoian S, Pirola CJ. Systematic review with meta-analysis: risk factors for non-alcoholic fatty liver disease suggest a shared altered metabolic and cardiovascular profile between lean and obese patients. *Aliment Pharmacol Ther* 2017; **46**: 85-95 [PMID: 28464369 DOI: 10.1111/apt.14112]
- Kim D, Kim WR. Nonobese Fatty Liver Disease. *Clin Gastroenterol Hepatol* 2017; **15**: 474-485 [PMID: 27581063 DOI: 10.1016/j.cgh.2016.08.028]
- Lonardo A, Ballestri S, Marchesini G, Angulo P, Loria P. Nonalcoholic fatty liver disease: a precursor of the metabolic syndrome. *Dig Liver Dis* 2015; **47**: 181-190 [PMID: 25739820 DOI: 10.1016/j.dld.2014.09.020]
- Ballestri S, Zona S, Targher G, Romagnoli D, Baldelli E, Nascimbeni F, Roverato A, Guaraldi G, Lonardo A. Nonalcoholic fatty liver disease is associated with an almost twofold increased risk of incident type 2 diabetes and metabolic syndrome. Evidence from a systematic review and meta-analysis. *J Gastroenterol Hepatol* 2016; **31**: 936-944 [PMID: 26667191 DOI: 10.1111/jgh.13264]
- Ballestri S, Nascimbeni F, Romagnoli D, Baldelli E, Targher G, Lonardo A. Type 2 Diabetes in Non-Alcoholic Fatty Liver Disease and Hepatitis C Virus Infection--Liver: The "Musketeer" in the Spotlight. *Int J Mol Sci* 2016; **17**: 355 [PMID: 27005620 DOI: 10.3390/ijms17030355]
- Loria P, Lonardo A, Anania F. Liver and diabetes. A vicious circle. *Hepatol Res* 2013; **43**: 51-64 [PMID: 23332087 DOI: 10.1111/j.1872-034X.2012.01031.x]
- Valenti L, Bugianesi E, Pajvani U, Targher G. Nonalcoholic fatty liver disease: cause or consequence of type 2 diabetes? *Liver Int* 2016; **36**: 1563-1579 [PMID: 27276701 DOI: 10.1111/liv.13185]
- Adams LA, Anstee QM, Tilg H, Targher G. Non-alcoholic fatty liver disease and its relationship with cardiovascular disease and other extrahepatic diseases. *Gut* 2017; **66**: 1138-1153 [PMID: 28314735 DOI: 10.1136/gutjnl-2017-313884]
- Day CP, James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology* 1998; **114**: 842-845 [PMID: 9547102]
- Tilg H, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836-1846 [PMID: 21038418 DOI: 10.1002/hep.24001]
- Larrain S, Rinella ME. A myriad of pathways to NASH. *Clin Liver Dis* 2012; **16**: 525-548 [PMID: 22824479 DOI: 10.1016/j.cld.2012.05.009]
- Dongiovanni P, Valenti L. Genetics of nonalcoholic fatty liver disease. *Metabolism* 2016; **65**: 1026-1037 [PMID: 26409295 DOI: 10.1016/j.metabol.2016.05.009]

- 10.1016/j.metabol.2015.08.018]
- 31 **Yki-Järvinen H.** Non-alcoholic fatty liver disease as a cause and a consequence of metabolic syndrome. *Lancet Diabetes Endocrinol* 2014; **2**: 901-910 [PMID: 24731669 DOI: 10.1016/S2213-8587(14)70032-4]
- 32 **Lonardo A,** Nascimbeni F, Ponz de Leon M. Nonalcoholic fatty liver disease and COPD: is it time to cross the diaphragm? *Eur Respir J* 2017; **49**: pii: 1700546 [PMID: 28596428 DOI: 10.1183/13993003.00546-2017]
- 33 **Kalia HS,** Gaglio PJ. The Prevalence and Pathobiology of Nonalcoholic Fatty Liver Disease in Patients of Different Races or Ethnicities. *Clin Liver Dis* 2016; **20**: 215-224 [PMID: 27063265 DOI: 10.1016/j.cld.2015.10.005]
- 34 **Betrapally NS,** Gillevet PM, Bajaj JS. Changes in the Intestinal Microbiome and Alcoholic and Nonalcoholic Liver Diseases: Causes or Effects? *Gastroenterology* 2016; **150**: 1745-1755.e3 [PMID: 26948887 DOI: 10.1053/j.gastro.2016.02.073]
- 35 **Zhu L,** Baker RD, Zhu R, Baker SS. Gut microbiota produce alcohol and contribute to NAFLD. *Gut* 2016; **65**: 1232 [PMID: 26984853 DOI: 10.1136/gutjnl-2016-311571]
- 36 **Marchesi JR,** Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, Quraishi MN, Kinross J, Smidt H, Tuohy KM, Thomas LV, Zoetendal EG, Hart A. The gut microbiota and host health: a new clinical frontier. *Gut* 2016; **65**: 330-339 [PMID: 26338727 DOI: 10.1136/gutjnl-2015-309990]
- 37 **Boursier J,** Vergnol J, Guillet A, Hiriart JB, Lannes A, Le Bail B, Michalak S, Chermak F, Bertrais S, Foucher J, Oberti F, Charbonnier M, Fouchard-Hubert I, Rousselet MC, Calès P, de Lédinghen V. Diagnostic accuracy and prognostic significance of blood fibrosis tests and liver stiffness measurement by FibroScan in non-alcoholic fatty liver disease. *J Hepatol* 2016; **65**: 570-578 [PMID: 27151181 DOI: 10.1016/j.jhep.2016.04.023]
- 38 **European Association for the Study of the Liver (EASL);** European Association for the Study of Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. *J Hepatol* 2016; **64**: 1388-1402 [PMID: 27062661 DOI: 10.1016/j.jhep.2015.11.004]
- 39 **Bugianesi E,** Rosso C, Cortez-Pinto H. How to diagnose NAFLD in 2016. *J Hepatol* 2016; **65**: 643-644 [PMID: 27401791 DOI: 10.1016/j.jhep.2016.05.038]
- 40 **Bedossa P;** FLIP Pathology Consortium. Utility and appropriateness of the fatty liver inhibition of progression (FLIP) algorithm and steatosis, activity, and fibrosis (SAF) score in the evaluation of biopsies of nonalcoholic fatty liver disease. *Hepatology* 2014; **60**: 565-575 [PMID: 24753132 DOI: 10.1002/hep.27173]
- 41 **Brunt EM.** Nonalcoholic Fatty Liver Disease: Pros and Cons of Histologic Systems of Evaluation. *Int J Mol Sci* 2016; **17**: pii: E97 [PMID: 26771611 DOI: 10.3390/ijms17010097]
- 42 **Chalasani N,** Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, Charlton M, Sanyal AJ; American Gastroenterological Association; American Association for the Study of Liver Diseases; American College of Gastroenterology. The diagnosis and management of non-alcoholic fatty liver disease: practice guideline by the American Gastroenterological Association, American Association for the Study of Liver Diseases, and American College of Gastroenterology. *Gastroenterology* 2012; **142**: 1592-1609 [PMID: 22656328 DOI: 10.1053/j.gastro.2012.04.001]
- 43 **Vernon G,** Baranova A, Younossi ZM. Systematic review: the epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. *Aliment Pharmacol Ther* 2011; **34**: 274-285 [PMID: 21623852 DOI: 10.1111/j.1365-2036.2011.04724.x]
- 44 **Day CP,** Saksena S. Non-alcoholic steatohepatitis: definitions and pathogenesis. *J Gastroenterol Hepatol* 2002; **17** Suppl 3: S377-S384 [PMID: 12472967]
- 45 **Caligiuri A,** Gentilini A, Marra F. Molecular Pathogenesis of NASH. *Int J Mol Sci* 2016; **17**: pii: E1575 [PMID: 27657051 DOI: 10.3390/ijms17091575]
- 46 **Shulman GI.** Ectopic fat in insulin resistance, dyslipidemia, and cardiometabolic disease. *N Engl J Med* 2014; **371**: 1131-1141 [PMID: 25229917 DOI: 10.1056/NEJMr1011035]
- 47 **Donnelly KL,** Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J Clin Invest* 2005; **115**: 1343-1351 [PMID: 15864352 DOI: 10.1172/JCI23621]
- 48 **Polyzos SA,** Toulis KA, Goulis DG, Zavos C, Kountouras J. Serum total adiponectin in nonalcoholic fatty liver disease: a systematic review and meta-analysis. *Metabolism* 2011; **60**: 313-326 [PMID: 21040935 DOI: 10.1016/j.metabol.2010.09.003]
- 49 **Neuschwander-Tetri BA.** Hepatic lipotoxicity and the pathogenesis of nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. *Hepatology* 2010; **52**: 774-788 [PMID: 20683968 DOI: 10.1002/hep.23719]
- 50 **Leamy AK,** Egnatchik RA, Young JD. Molecular mechanisms and the role of saturated fatty acids in the progression of non-alcoholic fatty liver disease. *Prog Lipid Res* 2013; **52**: 165-174 [PMID: 23178552 DOI: 10.1016/j.plipres.2012.10.004]
- 51 **Seppälä-Lindroos A,** Vehkavaara S, Häkkinen AM, Goto T, Westerbacka J, Sovijärvi A, Halavaara J, Yki-Järvinen H. Fat accumulation in the liver is associated with defects in insulin suppression of glucose production and serum free fatty acids independent of obesity in normal men. *J Clin Endocrinol Metab* 2002; **87**: 3023-3028 [PMID: 12107194 DOI: 10.1210/jcem.87.7.8638]
- 52 **Mehal WZ.** The Gordian Knot of dysbiosis, obesity and NAFLD. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 637-644 [PMID: 23958600 DOI: 10.1038/nrgastro.2013.146]
- 53 **Tilg H,** Kaser A. Gut microbiome, obesity, and metabolic dysfunction. *J Clin Invest* 2011; **121**: 2126-2132 [PMID: 21633181 DOI: 10.1172/JCI58109]
- 54 **Turnbaugh PJ,** Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JL. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006; **444**: 1027-1031 [PMID: 17183312 DOI: 10.1038/nature05414]
- 55 **Canli PD,** Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, Burcelin R. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* 2008; **57**: 1470-1481 [PMID: 18305141 DOI: 10.2337/db07-1403]
- 56 **Tremaroli V,** Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature* 2012; **489**: 242-249 [PMID: 22972297 DOI: 10.1038/nature11552]
- 57 **Pendyala S,** Walker JM, Holt PR. A high-fat diet is associated with endotoxemia that originates from the gut. *Gastroenterology* 2012; **142**: 1100-1101.e2 [PMID: 22326433 DOI: 10.1053/j.gastro.2012.01.034]
- 58 **Roh YS,** Seki E. Toll-like receptors in alcoholic liver disease, non-alcoholic steatohepatitis and carcinogenesis. *J Gastroenterol Hepatol* 2013; **28** Suppl 1: 38-42 [PMID: 23855294 DOI: 10.1111/jgh.12019]
- 59 **Miura K,** Ohnishi H. Role of gut microbiota and Toll-like receptors in nonalcoholic fatty liver disease. *World J Gastroenterol* 2014; **20**: 7381-7391 [PMID: 24966608 DOI: 10.3748/wjg.v20.i23.7381]
- 60 **Wree A,** Marra F. The inflammasome in liver disease. *J Hepatol* 2016; **65**: 1055-1056 [PMID: 27660175 DOI: 10.1016/j.jhep.2016.07.002]
- 61 **Samuel VT,** Shulman GI. The pathogenesis of insulin resistance: integrating signaling pathways and substrate flux. *J Clin Invest* 2016; **126**: 12-22 [PMID: 26727229 DOI: 10.1172/JCI77812]
- 62 **Kitajima Y,** Hyogo H, Sumida Y, Eguchi Y, Ono N, Kuwashiro T, Tanaka K, Takahashi H, Mizuta T, Ozaki I, Eguchi T, Kimura Y, Fujimoto K, Anzai K; Japan Nonalcoholic Fatty Liver Disease Study Group (JSG-NAFLD). Severity of non-alcoholic steatohepatitis is associated with substitution of adipose tissue in skeletal muscle. *J Gastroenterol Hepatol* 2013; **28**: 1507-1514 [PMID: 23577962 DOI: 10.1111/jgh.12227]

- 63 **Lee YH**, Jung KS, Kim SU, Yoon HJ, Yun YJ, Lee BW, Kang ES, Han KH, Lee HC, Cha BS. Sarcopaenia is associated with NAFLD independently of obesity and insulin resistance: Nationwide surveys (KNHANES 2008-2011). *J Hepatol* 2015; **63**: 486-493 [PMID: 25772036 DOI: 10.1016/j.jhep.2015.02.051]
- 64 **Koo BK**, Kim D, Joo SK, Kim JH, Chang MS, Kim BG, Lee KL, Kim W. Sarcopenia is an independent risk factor for non-alcoholic steatohepatitis and significant fibrosis. *J Hepatol* 2017; **66**: 123-131 [PMID: 27599824 DOI: 10.1016/j.jhep.2016.08.019]
- 65 **Petta S**, Ciminnisi S, Di Marco V, Cabibi D, Cammà C, Licata A, Marchesini G, Craxi A. Sarcopenia is associated with severe liver fibrosis in patients with non-alcoholic fatty liver disease. *Aliment Pharmacol Ther* 2017; **45**: 510-518 [PMID: 28028821 DOI: 10.1111/apt.13889]
- 66 **Zhang HJ**, Zhang XF, Ma ZM, Pan LL, Chen Z, Han HW, Han CK, Zhuang XJ, Lu Y, Li XJ, Yang SY, Li XY. Irisin is inversely associated with intrahepatic triglyceride contents in obese adults. *J Hepatol* 2013; **59**: 557-562 [PMID: 23665283 DOI: 10.1016/j.jhep.2013.04.030]
- 67 **Arias-Loste MT**, Ranchal I, Romero-Gómez M, Crespo J. Irisin, a link among fatty liver disease, physical inactivity and insulin resistance. *Int J Mol Sci* 2014; **15**: 23163-23178 [PMID: 25514415 DOI: 10.3390/ijms151223163]
- 68 **Liu YL**, Reeves HL, Burt AD, Tiniakos D, McPherson S, Leathart JB, Allison ME, Alexander GJ, Piguet AC, Anty R, Donaldson P, Aithal GP, Francque S, Van Gaal L, Clement K, Ratzliff V, Dufour JF, Day CP, Daly AK, Anstee QM. TM6SF2 rs58542926 influences hepatic fibrosis progression in patients with non-alcoholic fatty liver disease. *Nat Commun* 2014; **5**: 4309 [PMID: 24978903 DOI: 10.1038/ncomms5309]
- 69 **Anstee QM**, Day CP. The Genetics of Nonalcoholic Fatty Liver Disease: Spotlight on PNPLA3 and TM6SF2. *Semin Liver Dis* 2015; **35**: 270-290 [PMID: 26378644 DOI: 10.1055/s-0035-1562947]
- 70 **Bhadoria AS**, Kedarisetty CK, Bihari C, Kumar G, Jindal A, Bhardwaj A, Shasthry V, Vyas T, Benjamin J, Sharma S, Sharma MK, Sarin SK. Impact of family history of metabolic traits on severity of non-alcoholic steatohepatitis related cirrhosis: A cross-sectional study. *Liver Int* 2017; **37**: 1397-1404 [PMID: 28231412 DOI: 10.1111/liv.13396]
- 71 **Wilson PA**, Gardner SD, Lambie NM, Commans SA, Crowther DJ. Characterization of the human patatin-like phospholipase family. *J Lipid Res* 2006; **47**: 1940-1949 [PMID: 16799181 DOI: 10.1194/jlr.M600185-JLR200]
- 72 **Naik A**, Košir R, Rozman D. Genomic aspects of NAFLD pathogenesis. *Genomics* 2013; **102**: 84-95 [PMID: 23545492 DOI: 10.1016/j.ygeno.2013.03.007]
- 73 **Browning JD**. Common genetic variants and nonalcoholic Fatty liver disease. *Clin Gastroenterol Hepatol* 2013; **11**: 1191-1193 [PMID: 23707460 DOI: 10.1016/j.cgh.2013.05.013]
- 74 **Kotronen A**, Johansson LE, Johansson LM, Roos C, Westerbacka J, Hamsten A, Bergholm R, Arkkila P, Arola J, Kiviluoto T, Fisher RM, Ehrenborg E, Orho-Melander M, Ridderstråle M, Groop L, Yki-Järvinen H. A common variant in PNPLA3, which encodes adiponutrin, is associated with liver fat content in humans. *Diabetologia* 2009; **52**: 1056-1060 [PMID: 19224197 DOI: 10.1007/s00125-009-1285-z]
- 75 **Hernaez R**, McLean J, Lazo M, Brancati FL, Hirschhorn JN, Borecki IB, Harris TB; Genetics of Obesity-Related Liver Disease (GOLD) Consortium, Nguyen T, Kamel IR, Bonekamp S, Eberhardt MS, Clark JM, Kao WH, Speliotes EK. Association between variants in or near PNPLA3, GCKR, and PPP1R3B with ultrasound-defined steatosis based on data from the third National Health and Nutrition Examination Survey. *Clin Gastroenterol Hepatol* 2013; **11**: 1183-1190.e2 [PMID: 23416328 DOI: 10.1016/j.cgh.2013.02.011]
- 76 **Takaki A**, Kawai D, Yamamoto K. Molecular mechanisms and new treatment strategies for non-alcoholic steatohepatitis (NASH). *Int J Mol Sci* 2014; **15**: 7352-7379 [PMID: 24786095 DOI: 10.3390/ijms15057352]
- 77 **Severson TJ**, Besur S, Bonkovsky HL. Genetic factors that affect nonalcoholic fatty liver disease: A systematic clinical review. *World J Gastroenterol* 2016; **22**: 6742-6756 [PMID: 27547017 DOI: 10.3748/wjg.v22.i29.6742]
- 78 **Trépo E**, Nahon P, Bontempi G, Valenti L, Falletti E, Nischalke HD, Hamza S, Corradini SG, Burza MA, Guyot E, Donati B, Spengler U, Hillon P, Toniutto P, Henrion J, Franchimont D, Devière J, Mathurin P, Moreno C, Romeo S, Deltenre P. Association between the PNPLA3 (rs738409 C>G) variant and hepatocellular carcinoma: Evidence from a meta-analysis of individual participant data. *Hepatology* 2014; **59**: 2170-2177 [PMID: 24114809 DOI: 10.1002/hep.26767]
- 79 **Kozlitina J**, Smagris E, Stender S, Nordestgaard BG, Zhou HH, Tybjaerg-Hansen A, Vogt TF, Hobbs HH, Cohen JC. Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet* 2014; **46**: 352-356 [PMID: 24531328 DOI: 10.1038/ng.2901]
- 80 **Dongiovanni P**, Petta S, Maglio C, Fracanzani AL, Pipitone R, Mozzi E, Motta BM, Kaminska D, Rametta R, Grimaudo S, Pelusi S, Montalcini T, Alisi A, Maggioni M, Kärjä V, Borén J, Käkälä P, Di Marco V, Xing C, Nobili V, Dallapiccola B, Craxi A, Pihlajamäki J, Fargion S, Sjöström L, Carlsson LM, Romeo S, Valenti L. Transmembrane 6 superfamily member 2 gene variant disentangles nonalcoholic steatohepatitis from cardiovascular disease. *Hepatology* 2015; **61**: 506-514 [PMID: 25251399 DOI: 10.1002/hep.27490]
- 81 **Holmen OL**, Zhang H, Fan Y, Hovelson DH, Schmidt EM, Zhou W, Guo Y, Zhang J, Langhammer A, Løchen ML, Ganesh SK, Vatten L, Skorpén F, Dalen H, Zhang J, Pennathur S, Chen J, Platou C, Mathiesen EB, Wilsgaard T, Njølstad I, Boehnke M, Chen YE, Abecasis GR, Hveem K, Willer CJ. Systematic evaluation of coding variation identifies a candidate causal variant in TM6SF2 influencing total cholesterol and myocardial infarction risk. *Nat Genet* 2014; **46**: 345-351 [PMID: 24633158 DOI: 10.1038/ng.2926]
- 82 **Morris AP**, Voight BF, Teslovich TM, Ferreira T, Segrè AV, Steinthorsdottir V, Strawbridge RJ, Khan H, Grallert H, Mahajan A, Prokopenko I, Kang HM, Dina C, Esko T, Fraser RM, Kanoni S, Kumar A, Lagou V, Langenberg C, Luan J, Lindgren CM, Müller-Nurasyid M, Pechlivanis S, Rayner NW, Scott LJ, Wiltshire S, Yengo L, Kinnunen L, Rossin EJ, Raychaudhuri S, Johnson AD, Dimas AS, Loos RJ, Vedantam S, Chen H, Florez JC, Fox C, Liu CT, Rybin D, Couper DJ, Kao WH, Li M, Cornelis MC, Kraft P, Sun Q, van Dam RM, Stringham HM, Chines PS, Fischer K, Fontanillas P, Holmen OL, Hunt SE, Jackson AU, Kong A, Lawrence R, Meyer J, Perry JR, Platou CG, Potter S, Rehnberg E, Robertson N, Sivapalaratnam S, Stančáková A, Stirrups K, Thorleifsson G, Tikkanen E, Wood AR, Almgren P, Atalay M, Benediktsson R, Bonnycastle LL, Burtt N, Carey J, Charpentier G, Crenshaw AT, Doney AS, Dorkhan M, Edkins S, Emilsson V, Eury E, Forsen T, Gertow K, Gigante B, Grant GB, Groves CJ, Guiducci C, Herder C, Hreidarsson AB, Hui J, James A, Jonsson A, Rathmann W, Klopp N, Kravic J, Krjutskov K, Langford C, Leander K, Lindholm E, Lobbens S, Männistö S, Mirza G, Mühleisen TW, Musk B, Parkin M, Rallidis L, Saramies J, Sennblad B, Shah S, Sigurdsson G, Silveira A, Steinbach G, Thorand B, Trakalo J, Veglia F, Wennauer R, Winckler W, Zabaneh D, Campbell H, van Duijn C, Uitterlinden AG, Hofman A, Sijbrands E, Abecasis GR, Owen KR, Zeggini E, Trip MD, Forouhi NG, Syvänen AC, Eriksson JG, Peltonen L, Nöthen MM, Balkau B, Palmer CN, Lyssenko V, Tuomi T, Isomaa B, Hunter DJ, Qi L; Wellcome Trust Case Control Consortium; Meta-Analyses of Glucose and Insulin-related traits Consortium (MAGIC) Investigators; Genetic Investigation of ANthropometric Traits (GIANT) Consortium; Asian Genetic Epidemiology Network-Type 2 Diabetes (AGEN-T2D) Consortium; South Asian Type 2 Diabetes (SAT2D) Consortium, Shuldiner AR, Roden M, Barroso I, Wilsgaard T, Beilby J, Hovingh K, Price JF, Wilson JF, Rauramaa R, Lakka TA, Lind L, Dedoussis G, Njølstad I, Pedersen NL, Khaw KT, Wareham NJ, Keinanen-Kiukkaanniemi SM, Saaristo TE,

- Korpi-Hyövähti E, Saltevo J, Laakso M, Kuusisto J, Metspalu A, Collins FS, Mohlke KL, Bergman RN, Tuomilehto J, Boehm BO, Gieger C, Hveem K, Cauchi S, Froguel P, Baldassarre D, Tremoli E, Humphries SE, Saleheen D, Danesh J, Ingelsson E, Ripatti S, Salomaa V, Erbel R, Jöckel KH, Moebus S, Peters A, Illig T, de Faire U, Hamsten A, Morris AD, Donnelly PJ, Frayling TM, Hattersley AT, Boerwinkle E, Melander O, Kathiresan S, Nilsson PM, Deloukas P, Thorsteinsdottir U, Groop LC, Stefansson K, Hu F, Pankow JS, Dupuis J, Meigs JB, Altshuler D, Boehnke M, McCarthy MI; DIAbetes Genetics Replication And Meta-analysis (DIAGRAM) Consortium. Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet* 2012; **44**: 981-990 [PMID: 22885922 DOI: 10.1038/ng.2383]
- 83 **Luukkonen PK**, Zhou Y, Nidhina Haridas PA, Dwivedi OP, Hyötyläinen T, Ali A, Juuti A, Leivonen M, Tukiainen T, Ahonen L, Scott E, Palmer JM, Arola J, Orho-Melander M, Vikman P, Anstee QM, Olkkonen VM, Orešič M, Groop L, Yki-Järvinen H. Impaired hepatic lipid synthesis from polyunsaturated fatty acids in TM6SF2 E167K variant carriers with NAFLD. *J Hepatol* 2017; **67**: 128-136 [PMID: 28235613 DOI: 10.1016/j.jhep.2017.02.014]
- 84 **Musso G**, Cipolla U, Cassader M, Pinach S, Saba F, De Micheli F, Paschetta E, Bongiovanni D, Framarin L, Leone N, Berrutti M, Rosina F, Corvisieri S, Molinaro F, Sircana A, Gambino R. TM6SF2 rs58542926 variant affects postprandial lipoprotein metabolism and glucose homeostasis in NAFLD. *J Lipid Res* 2017; **58**: 1221-1229 [PMID: 28242789 DOI: 10.1194/jlr.M075028]
- 85 **Mancina RM**, Dongiovanni P, Petta S, Pingitore P, Meroni M, Rametta R, Borén J, Montalcini T, Pujia A, Wiklund O, Hindy G, Spagnuolo R, Motta BM, Pipitone RM, Craxi A, Fargion S, Nobili V, Käkälä P, Kärjä V, Männistö V, Pihlajamäki J, Reilly DF, Castro-Perez J, Kozlitina J, Valenti L, Romeo S. The MBOAT7-TMC4 Variant rs641738 Increases Risk of Nonalcoholic Fatty Liver Disease in Individuals of European Descent. *Gastroenterology* 2016; **150**: 1219-1230.e6 [PMID: 26850495 DOI: 10.1053/j.gastro.2016.01.032]
- 86 **Luukkonen PK**, Zhou Y, Hyötyläinen T, Leivonen M, Arola J, Orho-Melander M, Orešič M, Yki-Järvinen H. The MBOAT7 variant rs641738 alters hepatic phosphatidylinositols and increases severity of non-alcoholic fatty liver disease in humans. *J Hepatol* 2016; **65**: 1263-1265 [PMID: 27520876 DOI: 10.1016/j.jhep.2016.07.045]
- 87 **Conus F**, Rabasa-Lhoret R, Péronnet F. Characteristics of metabolically obese normal-weight (MONW) subjects. *Appl Physiol Nutr Metab* 2007; **32**: 4-12 [PMID: 17332780 DOI: 10.1139/H07-926]
- 88 **Deurenberg-Yap M**, Deurenberg P. Is a re-evaluation of WHO body mass index cut-off values needed? The case of Asians in Singapore. *Nutr Rev* 2003; **61**: S80-S87 [PMID: 12828197]
- 89 **WHO Expert Consultation**. Appropriate body-mass index for Asian populations and its implications for policy and intervention strategies. *Lancet* 2004; **363**: 157-163 [PMID: 14726171 DOI: 10.1016/S0140-6736(03)15268-3]
- 90 **Wei JL**, Leung JC, Loong TC, Wong GL, Yeung DK, Chan RS, Chan HL, Chim AM, Woo J, Chu WC, Wong VW. Prevalence and Severity of Nonalcoholic Fatty Liver Disease in Non-Obese Patients: A Population Study Using Proton-Magnetic Resonance Spectroscopy. *Am J Gastroenterol* 2015; **110**: 1306-1314; quiz 1315 [PMID: 26215532 DOI: 10.1038/ajg.2015.235]
- 91 **Wattacheril J**, Sanyal AJ. Lean NAFLD: An Underrecognized Outlier. *Curr Hepatol Rep* 2016; **15**: 134-139 [PMID: 27668144 DOI: 10.1007/s11901-016-0302-1]
- 92 **Stender S**, Kozlitina J, Nordestgaard BG, Tybjaerg-Hansen A, Hobbs HH, Cohen JC. Adiposity amplifies the genetic risk of fatty liver disease conferred by multiple loci. *Nat Genet* 2017; **49**: 842-847 [PMID: 28436986 DOI: 10.1038/ng.3855]
- 93 **Angulo P**, Kleiner DE, Dam-Larsen S, Adams LA, Bjornsson ES, Charatcharoenwitthaya P, Mills PR, Keach JC, Lafferty HD, Stahler A, Haflidadottir S, Bendtsen F. Liver Fibrosis, but No Other Histologic Features, Is Associated With Long-term Outcomes of Patients With Nonalcoholic Fatty Liver Disease. *Gastroenterology* 2015; **149**: 389-397.e10 [PMID: 25935633 DOI: 10.1053/j.gastro.2015.04.043]
- 94 **Ekstedt M**, Hagström H, Nasr P, Fredrikson M, Stål P, Kechagias S, Hultcrantz R. Fibrosis stage is the strongest predictor for disease-specific mortality in NAFLD after up to 33 years of follow-up. *Hepatology* 2015; **61**: 1547-1554 [PMID: 25125077 DOI: 10.1002/hep.27368]
- 95 **Dulai PS**, Singh S, Patel J, Soni M, Prokop LJ, Younossi Z, Sebastiani G, Ekstedt M, Hagstrom H, Nasr P, Stal P, Wong VW, Kechagias S, Hultcrantz R, Loomba R. Increased risk of mortality by fibrosis stage in nonalcoholic fatty liver disease: Systematic review and meta-analysis. *Hepatology* 2017; **65**: 1557-1565 [PMID: 28130788 DOI: 10.1002/hep.29085]
- 96 **Matteoni CA**, Younossi ZM, Gramlich T, Boparai N, Liu YC, McCullough AJ. Nonalcoholic fatty liver disease: a spectrum of clinical and pathological severity. *Gastroenterology* 1999; **116**: 1413-1419 [PMID: 10348825]
- 97 **Brunt EM**, Janney CG, Di Bisceglie AM, Neuschwander-Tetri BA, Bacon BR. Nonalcoholic steatohepatitis: a proposal for grading and staging the histological lesions. *Am J Gastroenterol* 1999; **94**: 2467-2474 [PMID: 10484010 DOI: 10.1111/j.1572-0241.1999.01377.x]
- 98 **Kleiner DE**, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu YC, Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal AJ; Nonalcoholic Steatohepatitis Clinical Research Network. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313-1321 [PMID: 15915461 DOI: 10.1002/hep.20701]
- 99 **Bedossa P**, Poitou C, Veyrie N, Bouillot JL, Basdevant A, Paradis V, Tordjman J, Clement K. Histopathological algorithm and scoring system for evaluation of liver lesions in morbidly obese patients. *Hepatology* 2012; **56**: 1751-1759 [PMID: 22707395 DOI: 10.1002/hep.25889]
- 100 **Nascimbeni F**, Lebray P, Fedchuk L, Oliveira CP, Alvares-da-Silva MR, Varault A, Ingiliz P, Ngo Y, de Torres M, Munteanu M, Poynard T, Ratzu V; LIDO Study Group. Significant variations in elastometry measurements made within short-term in patients with chronic liver diseases. *Clin Gastroenterol Hepatol* 2015; **13**: 763-771.e1-e6 [PMID: 25086193 DOI: 10.1016/j.cgh.2014.07.037]
- 101 **Fedchuk L**, Nascimbeni F, Pais R, Charlotte F, Housset C, Ratzu V; LIDO Study Group. Performance and limitations of steatosis biomarkers in patients with nonalcoholic fatty liver disease. *Aliment Pharmacol Ther* 2014; **40**: 1209-1222 [PMID: 25267215 DOI: 10.1111/apt.12963]
- 102 **Bedossa P**, Patel K. Biopsy and Noninvasive Methods to Assess Progression of Nonalcoholic Fatty Liver Disease. *Gastroenterology* 2016; **150**: 1811-1822.e4 [PMID: 27003601 DOI: 10.1053/j.gastro.2016.03.008]
- 103 **Brunt EM**, Kleiner DE, Wilson LA, Belt P, Neuschwander-Tetri BA; NASH Clinical Research Network (CRN). Nonalcoholic fatty liver disease (NAFLD) activity score and the histopathologic diagnosis in NAFLD: distinct clinicopathologic meanings. *Hepatology* 2011; **53**: 810-820 [PMID: 21319198 DOI: 10.1002/hep.24127]
- 104 **Ekstedt M**, Franzén LE, Mathiesen UL, Kechagias S. Low clinical relevance of the nonalcoholic fatty liver disease activity score (NAS) in predicting fibrosis progression. *Scand J Gastroenterol* 2012; **47**: 108-115 [PMID: 22126450 DOI: 10.3109/00365521.2011.634024]
- 105 **Younossi ZM**, Stepanova M, Rafiq N, Makhlof H, Younoszai Z, Agrawal R, Goodman Z. Pathologic criteria for nonalcoholic steatohepatitis: interprotocol agreement and ability to predict liver-related mortality. *Hepatology* 2011; **53**: 1874-1882 [PMID: 21360720 DOI: 10.1002/hep.24268]
- 106 **Ballestri S**, Nascimbeni F, Baldelli E, Marrazzo A, Romagnoli D, Targher G, Lonardo A. Ultrasonographic fatty liver indicator detects mild steatosis and correlates with metabolic/histological parameters in various liver diseases. *Metabolism* 2017; **72**: 57-65

- [PMID: 28641784 DOI: 10.1016/j.metabol.2017.04.003]
- 107 **Ballestri S**, Lonardo A, Romagnoli D, Carulli L, Losi L, Day CP, Loria P. Ultrasonographic fatty liver indicator, a novel score which rules out NASH and is correlated with metabolic parameters in NAFLD. *Liver Int* 2012; **32**: 1242-1252 [PMID: 22520641 DOI: 10.1111/j.1478-3231.2012.02804.x]
 - 108 **Ballestri S**, Romagnoli D, Nascimbeni F, Francica G, Lonardo A. Role of ultrasound in the diagnosis and treatment of nonalcoholic fatty liver disease and its complications. *Expert Rev Gastroenterol Hepatol* 2015; **9**: 603-627 [PMID: 25694178 DOI: 10.1586/17474124.2015.1007955]
 - 109 **Myers RP**, Pollett A, Kirsch R, Pomier-Layrargues G, Beaton M, Levstik M, Duarte-Rojo A, Wong D, Crotty P, Elkasab M. Controlled Attenuation Parameter (CAP): a noninvasive method for the detection of hepatic steatosis based on transient elastography. *Liver Int* 2012; **32**: 902-910 [PMID: 22435761 DOI: 10.1111/j.1478-3231.2012.02781.x]
 - 110 **de Lédinghen V**, Vergniol J, Capdepon M, Chermak F, Hiriart JB, Cassinotto C, Merrouche W, Foucher J, Brigitte le B. Controlled attenuation parameter (CAP) for the diagnosis of steatosis: a prospective study of 5323 examinations. *J Hepatol* 2014; **60**: 1026-1031 [PMID: 24378529 DOI: 10.1016/j.jhep.2013.12.018]
 - 111 **Karlas T**, Petroff D, Sasso M, Fan JG, Mi YQ, de Lédinghen V, Kumar M, Lupsor-Platon M, Han KH, Cardoso AC, Ferraioli G, Chan WK, Wong VW, Myers RP, Chayama K, Friedrich-Rust M, Beaugrand M, Shen F, Hiriart JB, Sarin SK, Badea R, Jung KS, Marcellin P, Filice C, Mahadeva S, Wong GL, Crotty P, Masaki K, Bojunga J, Bedossa P, Keim V, Wiegand J. Individual patient data meta-analysis of controlled attenuation parameter (CAP) technology for assessing steatosis. *J Hepatol* 2017; **66**: 1022-1030 [PMID: 28039099 DOI: 10.1016/j.jhep.2016.12.022]
 - 112 **Petta S**, Maida M, Macaluso FS, Di Marco V, Cammà C, Cabibi D, Craxi A. The severity of steatosis influences liver stiffness measurement in patients with nonalcoholic fatty liver disease. *Hepatology* 2015; **62**: 1101-1110 [PMID: 25991038 DOI: 10.1002/hep.27844]
 - 113 **Petta S**, Wong VW, Cammà C, Hiriart JB, Wong GL, Marra F, Vergniol J, Chan AW, Di Marco V, Merrouche W, Chan HL, Barbara M, Le-Bail B, Arena U, Craxi A, de Lédinghen V. Improved noninvasive prediction of liver fibrosis by liver stiffness measurement in patients with nonalcoholic fatty liver disease accounting for controlled attenuation parameter values. *Hepatology* 2017; **65**: 1145-1155 [PMID: 27639088 DOI: 10.1002/hep.28843]
 - 114 **Nasr P**, Forsgren MF, Ignatova S, Dahlström N, Cedersund G, Leinhard OD, Norén B, Ekstedt M, Lundberg P, Kechagias S. Using a 3% Proton Density Fat Fraction as a Cut-Off Value Increases Sensitivity of Detection of Hepatic Steatosis, Based on Results From Histopathology Analysis. *Gastroenterology* 2017; **153**: 53-55.e7 [PMID: 28286210 DOI: 10.1053/j.gastro.2017.03.005]
 - 115 **Banerjee R**, Pavlides M, Tunnicliffe EM, Piechnik SK, Sarania N, Philips R, Collier JD, Booth JC, Schneider JE, Wang LM, Delaney DW, Fleming KA, Robson MD, Barnes E, Neubauer S. Multiparametric magnetic resonance for the non-invasive diagnosis of liver disease. *J Hepatol* 2014; **60**: 69-77 [PMID: 24036007 DOI: 10.1016/j.jhep.2013.09.002]
 - 116 **Park CC**, Nguyen P, Hernandez C, Bettencourt R, Ramirez K, Fortney L, Hooker J, Sy E, Savides MT, Alquirash MH, Valasek MA, Rizo E, Richards L, Brenner D, Sirlin CB, Loomba R. Magnetic Resonance Elastography vs Transient Elastography in Detection of Fibrosis and Noninvasive Measurement of Steatosis in Patients With Biopsy-Proven Nonalcoholic Fatty Liver Disease. *Gastroenterology* 2017; **152**: 598-607.e2 [PMID: 27911262 DOI: 10.1053/j.gastro.2016.10.026]
 - 117 **Pavlides M**, Banerjee R, Sellwood J, Kelly CJ, Robson MD, Booth JC, Collier J, Neubauer S, Barnes E. Multiparametric magnetic resonance imaging predicts clinical outcomes in patients with chronic liver disease. *J Hepatol* 2016; **64**: 308-315 [PMID: 26471505 DOI: 10.1016/j.jhep.2015.10.009]
 - 118 **Machado MV**, Cortez-Pinto H. Non-invasive diagnosis of non-alcoholic fatty liver disease. A critical appraisal. *J Hepatol* 2013; **58**: 1007-1019 [PMID: 23183525 DOI: 10.1016/j.jhep.2012.11.021]
 - 119 **Bedogni G**, Bellentani S, Miglioli L, Masutti F, Passalacqua M, Castiglione A, Tiribelli C. The Fatty Liver Index: a simple and accurate predictor of hepatic steatosis in the general population. *BMC Gastroenterol* 2006; **6**: 33 [PMID: 17081293 DOI: 10.1186/1471-230X-6-33]
 - 120 **Gastaldelli A**, Kozakova M, Højlund K, Flyvbjerg A, Favuzzi A, Mitrakou A, Balkau B; RISC Investigators. Fatty liver is associated with insulin resistance, risk of coronary heart disease, and early atherosclerosis in a large European population. *Hepatology* 2009; **49**: 1537-1544 [PMID: 19291789 DOI: 10.1002/hep.22845]
 - 121 **Calori G**, Lattuada G, Ragogna F, Garancini MP, Crosignani P, Villa M, Bosi E, Ruotolo G, Piemonti L, Perseghin G. Fatty liver index and mortality: the Cremona study in the 15th year of follow-up. *Hepatology* 2011; **54**: 145-152 [PMID: 21488080 DOI: 10.1002/hep.24356]
 - 122 **Guin B**, Crevisy-Girod E, Binquet C, Duvillard L, Masson D, Lepage C, Hamza S, Krausé D, Verges B, Minello A, Cercueil JP, Hillon P, Petit JM. Prediction for steatosis in type-2 diabetes: clinico-biological markers versus 1H-MR spectroscopy. *Eur Radiol* 2012; **22**: 855-863 [PMID: 22101800 DOI: 10.1007/s00330-011-2326-9]
 - 123 **Cuthbertson DJ**, Weickert MO, Lythgoe D, Sprung VS, Dobson R, Shoaiee-Moradie F, Umpleby M, Pfeiffer AF, Thomas EL, Bell JD, Jones H, Kemp GJ. External validation of the fatty liver index and lipid accumulation product indices, using 1H-magnetic resonance spectroscopy, to identify hepatic steatosis in healthy controls and obese, insulin-resistant individuals. *Eur J Endocrinol* 2014; **171**: 561-569 [PMID: 25298375 DOI: 10.1530/EJE-14-0112]
 - 124 **McPherson S**, Stewart SF, Henderson E, Burt AD, Day CP. Simple non-invasive fibrosis scoring systems can reliably exclude advanced fibrosis in patients with non-alcoholic fatty liver disease. *Gut* 2010; **59**: 1265-1269 [PMID: 20801772 DOI: 10.1136/gut.2010.216077]
 - 125 **Angulo P**, Bugianesi E, Björnsson ES, Charatcharoenwithaya P, Mills PR, Barrera F, Haflidadottir S, Day CP, George J. Simple noninvasive systems predict long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* 2013; **145**: 782-789.e4 [PMID: 23860502 DOI: 10.1053/j.gastro.2013.06.057]
 - 126 **Kim D**, Kim WR, Kim HJ, Therneau TM. Association between noninvasive fibrosis markers and mortality among adults with nonalcoholic fatty liver disease in the United States. *Hepatology* 2013; **57**: 1357-1365 [PMID: 23175136 DOI: 10.1002/hep.26156]
 - 127 **Perazzo H**, Munteanu M, Ngo Y, Lebray P, Seurat N, Rutka F, Couteau M, Jacqueminet S, Giral P, Monneret D, Imbert-Bismut F, Ratzu V, Hartemann-Huertier A, Housset C, Poynard T; FLIP Consortium. Prognostic value of liver fibrosis and steatosis biomarkers in type-2 diabetes and dyslipidaemia. *Aliment Pharmacol Ther* 2014; **40**: 1081-1093 [PMID: 25186086 DOI: 10.1111/apt.12946]
 - 128 **McPherson S**, Hardy T, Dufour JF, Petta S, Romero-Gomez M, Allison M, Oliveira CP, Francque S, Van Gaal L, Schattenberg JM, Tiniakos D, Burt A, Bugianesi E, Ratzu V, Day CP, Anstee QM. Age as a Confounding Factor for the Accurate Non-Invasive Diagnosis of Advanced NAFLD Fibrosis. *Am J Gastroenterol* 2017; **112**: 740-751 [PMID: 27725647 DOI: 10.1038/ajg.2016.453]
 - 129 **Demir M**, Lang S, Nierhoff D, Drebber U, Hardt A, Wedemeyer I, Schulte S, Quasdorff M, Goesser T, Töx U, Steffen HM. Stepwise combination of simple noninvasive fibrosis scoring systems increases diagnostic accuracy in nonalcoholic fatty liver disease. *J Clin Gastroenterol* 2013; **47**: 719-726 [PMID: 23442837 DOI: 10.1097/MCG.0b013e3182819a89]
 - 130 **Petta S**, Vanni E, Bugianesi E, Di Marco V, Cammà C, Cabibi D, Mezzabotta L, Craxi A. The combination of liver stiffness measurement and NAFLD fibrosis score improves the noninvasive diagnostic accuracy for severe liver fibrosis in patients with nonalcoholic fatty liver disease. *Liver Int* 2015; **35**: 1566-1573 [PMID: 24798049 DOI: 10.1111/liv.12584]
 - 131 **Panera N**, Gnani D, Crudele A, Ceccarelli S, Nobili V, Alisi A. MicroRNAs as controlled systems and controllers in non-alcoholic

- fatty liver disease. *World J Gastroenterol* 2014; **20**: 15079-15086 [PMID: 25386056 DOI: 10.3748/wjg.v20.i41.15079]
- 132 **Murphy SK**, Yang H, Moylan CA, Pang H, Dellinger A, Abdelmalek MF, Garrett ME, Ashley-Koch A, Suzuki A, Tillmann HL, Hauser MA, Diehl AM. Relationship between methylome and transcriptome in patients with nonalcoholic fatty liver disease. *Gastroenterology* 2013; **145**: 1076-1087 [PMID: 23916847 DOI: 10.1053/j.gastro.2013.07.047]
 - 133 **Puri P**, Wiest MM, Cheung O, Mirshahi F, Sargeant C, Min HK, Contos MJ, Sterling RK, Fuchs M, Zhou H, Watkins SM, Sanyal AJ. The plasma lipidomic signature of nonalcoholic steatohepatitis. *Hepatology* 2009; **50**: 1827-1838 [PMID: 19937697 DOI: 10.1002/hep.23229]
 - 134 **Anjani K**, Lhomme M, Sokolovska N, Poitou C, Aron-Wisniewsky J, Bouillot JL, Lesnik P, Bedossa P, Kontush A, Clement K, Dugail I, Tordjman J. Circulating phospholipid profiling identifies portal contribution to NASH signature in obesity. *J Hepatol* 2015; **62**: 905-912 [PMID: 25450212 DOI: 10.1016/j.jhep.2014.11.002]
 - 135 **Alonso C**, Fernández-Ramos D, Varela-Rey M, Martínez-Arranz I, Navasa N, Van Liempd SM, Lavín Trueba JL, Mayo R, Ilisso CP, de Juan VG, Iruarizaga-Lejarreta M, de la Cruz-Villar L, Mincholé I, Robinson A, Crespo J, Martín-Duce A, Romero-Gómez M, Sann H, Platon J, Van Eyk J, Aspichueta P, Noureddin M, Falcón-Pérez JM, Anguita J, Aransay AM, Martínez-Chantar ML, Lu SC, Mato JM. Metabolomic Identification of Subtypes of Nonalcoholic Steatohepatitis. *Gastroenterology* 2017; **152**: 1449-1461.e7 [PMID: 28132890 DOI: 10.1053/j.gastro.2017.01.015]
 - 136 **Wood GC**, Chu X, Argyropoulos G, Benotti P, Rolston D, Mirshahi T, Petrick A, Gabrielson J, Carey DJ, DiStefano JK, Still CD, Gerhard GS. A multi-component classifier for nonalcoholic fatty liver disease (NAFLD) based on genomic, proteomic, and phenomic data domains. *Sci Rep* 2017; **7**: 43238 [PMID: 28266614 DOI: 10.1038/srep43238]
 - 137 **Kotronen A**, Peltonen M, Hakkarainen A, Sevestianova K, Bergholm R, Johansson LM, Lundbom N, Rissanen A, Ridderstråle M, Groop L, Orho-Melander M, Yki-Järvinen H. Prediction of non-alcoholic fatty liver disease and liver fat using metabolic and genetic factors. *Gastroenterology* 2009; **137**: 865-872 [PMID: 19524579 DOI: 10.1053/j.gastro.2009.06.005]
 - 138 **Lonardo A**, Bagni A, Tarugi P, Loria P. The wide spectrum of steatohepatitis: a report of four cases and a review of the literature. *Eur J Gastroenterol Hepatol* 2004; **16**: 1043-1050 [PMID: 15371930]
 - 139 **Lonardo A**, Loria P, Leonardi F, Borsatti A, Neri P, Pulvirenti M, Verrone AM, Bagni A, Bertolotti M, Ganazzi D, Carulli N; POLI.STE.N.A. Study Group. Policentrica Steatosi Epatica Non Alcolica. Fasting insulin and uric acid levels but not indices of iron metabolism are independent predictors of non-alcoholic fatty liver disease. A case-control study. *Dig Liver Dis* 2002; **34**: 204-211 [PMID: 11990393]
 - 140 **Nascimbeni F**, Loria P, Ratziv V. Non-alcoholic fatty liver disease: diagnosis and investigation. *Dig Dis* 2014; **32**: 586-596 [PMID: 25034292 DOI: 10.1159/000360510]
 - 141 **Marchesini G**, Brizi M, Morselli-Labate AM, Bianchi G, Bugianesi E, McCullough AJ, Forlani G, Melchionda N. Association of nonalcoholic fatty liver disease with insulin resistance. *Am J Med* 1999; **107**: 450-455 [PMID: 10569299]
 - 142 **Cortez-Pinto H**, Camilo ME, Baptista A, De Oliveira AG, De Moura MC. Non-alcoholic fatty liver: another feature of the metabolic syndrome? *Clin Nutr* 1999; **18**: 353-358 [PMID: 10634920 DOI: 10.1054/clnu.1999.0047]
 - 143 **Lonardo A**. Fatty liver and nonalcoholic steatohepatitis. Where do we stand and where are we going? *Dig Dis* 1999; **17**: 80-89 [PMID: 10545713 DOI: 10.1016/S0261-5614(99)80015-6]
 - 144 **Ballestri S**, Nascimbeni F, Romagnoli D, Lonardo A. The independent predictors of non-alcoholic steatohepatitis and its individual histological features.: Insulin resistance, serum uric acid, metabolic syndrome, alanine aminotransferase and serum total cholesterol are a clue to pathogenesis and candidate targets for treatment. *Hepatol Res* 2016; **46**: 1074-1087 [PMID: 26785389 DOI: 10.1111/hepr.12656]
 - 145 **Vanni E**, Bugianesi E, Kotronen A, De Minicis S, Yki-Järvinen H, Svegliati-Baroni G. From the metabolic syndrome to NAFLD or vice versa? *Dig Liver Dis* 2010; **42**: 320-330 [PMID: 20207596 DOI: 10.1016/j.dld.2010.01.016]
 - 146 **Ma J**, Hwang SJ, Pedley A, Massaro JM, Hoffmann U, Chung RT, Benjamin EJ, Levy D, Fox CS, Long MT. Bi-directional analysis between fatty liver and cardiovascular disease risk factors. *J Hepatol* 2017; **66**: 390-397 [PMID: 27729222 DOI: 10.1016/j.jhep.2016.09.022]
 - 147 **Zhang Y**, Zhang T, Zhang C, Tang F, Zhong N, Li H, Song X, Lin H, Liu Y, Xue F. Identification of reciprocal causality between non-alcoholic fatty liver disease and metabolic syndrome by a simplified Bayesian network in a Chinese population. *BMJ Open* 2015; **5**: e008204 [PMID: 26395497 DOI: 10.1136/bmjopen-2015-008204]
 - 148 **Liu CQ**, He CM, Chen N, Wang D, Shi X, Liu Y, Zeng X, Yan B, Liu S, Yang S, Li X, Li X, Li Z. Serum uric acid is independently and linearly associated with risk of nonalcoholic fatty liver disease in obese Chinese adults. *Sci Rep* 2016; **6**: 38605 [PMID: 27924915 DOI: 10.1038/srep38605]
 - 149 **Liu J**, Xu C, Ying L, Zang S, Zhuang Z, Lv H, Yang W, Luo Y, Ma X, Wang L, Xun Y, Ye D, Shi J. Relationship of serum uric acid level with non-alcoholic fatty liver disease and its inflammation progression in non-obese adults. *Hepatol Res* 2017; **47**: E104-E112 [PMID: 27172177 DOI: 10.1111/hepr.12734]
 - 150 **Zhao CC**, Wang AP, Li LX, Li TT, Chen MY, Zhu Y, Yu TP, Bao YQ, Jia WP. Urine uric acid excretion is associated with nonalcoholic fatty liver disease in patients with type 2 diabetes. *J Diabetes Complications* 2016; **30**: 1074-1080 [PMID: 27161518 DOI: 10.1016/j.jdiacomp.2016.04.017]
 - 151 **Ozelik F**, Yiginer O. The relationship between serum uric acid levels and the major risk factors for the development of nonalcoholic fatty liver disease. *Liver Int* 2016; **36**: 768-769 [PMID: 26790569 DOI: 10.1111/liv.13063]
 - 152 **Lombardi R**, Pisano G, Fargion S. Role of Serum Uric Acid and Ferritin in the Development and Progression of NAFLD. *Int J Mol Sci* 2016; **17**: 548 [PMID: 27077854 DOI: 10.3390/ijms17040548]
 - 153 **Zhou Y**, Wei F, Fan Y. High serum uric acid and risk of nonalcoholic fatty liver disease: A systematic review and meta-analysis. *Clin Biochem* 2016; **49**: 636-642 [PMID: 26738417 DOI: 10.1016/j.clinbiochem.2015.12.010]
 - 154 **Jaruvongvanich V**, Ahuja W, Wirunsawanya K, Wijarnpreecha K, Ungprasert P. Hyperuricemia is associated with nonalcoholic fatty liver disease activity score in patients with nonalcoholic fatty liver disease: a systematic review and meta-analysis. *Eur J Gastroenterol Hepatol* 2017; **29**: 1031-1035 [PMID: 28639970 DOI: 10.1097/MEG.0000000000000931]
 - 155 **Mosca A**, Nobili V, De Vito R, Crudele A, Scorletti E, Villani A, Alisi A, Byrne CD. Serum uric acid concentrations and fructose consumption are independently associated with NASH in children and adolescents. *J Hepatol* 2017; **66**: 1031-1036 [PMID: 28214020 DOI: 10.1016/j.jhep.2016.12.025]
 - 156 **Carulli L**, Ballestri S, Lonardo A, Lami F, Violi E, Losi L, Bonilauri L, Verrone AM, Odoardi MR, Scaglioni F, Bertolotti M, Loria P. Is nonalcoholic steatohepatitis associated with a high-threshold-normal thyroid stimulating hormone level and lower cholesterol levels? *Intern Emerg Med* 2013; **8**: 297-305 [PMID: 21559749 DOI: 10.1007/s11739-011-0609-4]
 - 157 **Siddiqui MS**, Fuchs M, Idowu MO, Luketic VA, Boyett S, Sargeant C, Stravitz RT, Puri P, Matherly S, Sterling RK, Contos M, Sanyal AJ. Severity of nonalcoholic fatty liver disease and progression to cirrhosis are associated with atherogenic lipoprotein profile. *Clin Gastroenterol Hepatol* 2015; **13**: 1000-1008.e3 [PMID: 25311381 DOI: 10.1016/j.cgh.2014.10.008]
 - 158 **Jiang ZG**, Tapper EB, Connelly MA, Pimentel CF, Feldbrügge L, Kim M, Krawczyk S, Afzhal N, Robson SC, Herman MA, Otvos JD, Mukamal KJ, Lai M. Steatohepatitis and liver fibrosis are predicted by the characteristics of very low density lipoprotein in nonalcoholic fatty liver disease. *Liver Int* 2016; **36**: 1213-1220

- [PMID: 26815314 DOI: 10.1111/liv.13076]
- 159 **Lucero D**, Miksztowicz V, Gualano G, Longo C, Landeira G, Álvarez E, Zago V, Brites F, Berg G, Fassio E, Schreier L. Nonalcoholic fatty liver disease associated with metabolic syndrome: Influence of liver fibrosis stages on characteristics of very low-density lipoproteins. *Clin Chim Acta* 2017; **473**: 1-8 [PMID: 28802640 DOI: 10.1016/j.cca.2017.08.006]
 - 160 **Lonardo A**, Romagnoli D. Gamma glutamyl transferase: A novel cardiovascular outfit for an old liver test. *Indian J Med Res* 2016; **143**: 4-7 [PMID: 26997005 DOI: 10.4103/0971-5916.178574]
 - 161 **Fracanzani AL**, Valenti L, Bugianesi E, Andreoletti M, Colli A, Vanni E, Bertelli C, Fatta E, Bignamini D, Marchesini G, Fargion S. Risk of severe liver disease in nonalcoholic fatty liver disease with normal aminotransferase levels: a role for insulin resistance and diabetes. *Hepatology* 2008; **48**: 792-798 [PMID: 18752331 DOI: 10.1002/hep.22429]
 - 162 **Kowdley KV**, Belt P, Wilson LA, Yeh MM, Neuschwander-Tetri BA, Chalasani N, Sanyal AJ, Nelson JE, NASH Clinical Research Network. Serum ferritin is an independent predictor of histologic severity and advanced fibrosis in patients with nonalcoholic fatty liver disease. *Hepatology* 2012; **55**: 77-85 [PMID: 21953442 DOI: 10.1002/hep.24706]
 - 163 **Ryan JD**, Armitage AE, Cobbolt JF, Banerjee R, Borsani O, Dongiovanni P, Neubauer S, Morovat R, Wang LM, Pasricha SR, Fargion S, Collier J, Barnes E, Drakesmith H, Valenti L, Pavlides M. Hepatic iron is the major determinant of serum ferritin in NAFLD patients. *Liver Int* 2017; Epub ahead of print [PMID: 28679028 DOI: 10.1111/liv.13513]
 - 164 **Hagström H**, Nasr P, Bottai M, Ekstedt M, Kechagias S, Hultcrantz R, Stål P. Elevated serum ferritin is associated with increased mortality in non-alcoholic fatty liver disease after 16 years of follow-up. *Liver Int* 2016; **36**: 1688-1695 [PMID: 27064133 DOI: 10.1111/liv.13144]
 - 165 **Loria P**, Lonardo A, Leonardi F, Fontana C, Carulli L, Verrone AM, Borsatti A, Bertolotti M, Cassani F, Bagni A, Muratori P, Ganazzi D, Bianchi FB, Carulli N. Non-organ-specific autoantibodies in nonalcoholic fatty liver disease: prevalence and correlates. *Dig Dis Sci* 2003; **48**: 2173-2181 [PMID: 14705824]
 - 166 **Vuppalanchi R**, Gould RJ, Wilson LA, Unalp-Arida A, Cummings OW, Chalasani N, Kowdley KV, Nonalcoholic Steatohepatitis Clinical Research Network (NASH CRN). Clinical significance of serum autoantibodies in patients with NAFLD: results from the nonalcoholic steatohepatitis clinical research network. *Hepatol Int* 2012; **6**: 379-385 [PMID: 21557024 DOI: 10.1007/s12072-011-9277-8]
 - 167 **Lonardo A**, Bellini M, Tondelli E, Frazzoni M, Grisendi A, Pulvirenti M, Della Casa G. Nonalcoholic steatohepatitis and the "bright liver syndrome": should a recently expanded clinical entity be further expanded? *Am J Gastroenterol* 1995; **90**: 2072-2074 [PMID: 7485040]
 - 168 **Byrne CD**, Targher G. NAFLD: a multisystem disease. *J Hepatol* 2015; **62**: S47-S64 [PMID: 25920090 DOI: 10.1016/j.jhep.2014.12.012]
 - 169 **Anstee QM**, Targher G, Day CP. Progression of NAFLD to diabetes mellitus, cardiovascular disease or cirrhosis. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 330-344 [PMID: 23507799 DOI: 10.1038/nrgastro.2013.41]
 - 170 **Lonardo A**, Ballestri S, Targher G, Loria P. Diagnosis and management of cardiovascular risk in nonalcoholic fatty liver disease. *Expert Rev Gastroenterol Hepatol* 2015; **9**: 629-650 [PMID: 25327387 DOI: 10.1586/17474124.2015.965143]
 - 171 **Oni ET**, Agatston AS, Blaha MJ, Fialkow J, Cury R, Sposito A, Erbel R, Blankstein R, Feldman T, Al-Mallah MH, Santos RD, Budoff MJ, Nasir K. A systematic review: burden and severity of subclinical cardiovascular disease among those with nonalcoholic fatty liver; should we care? *Atherosclerosis* 2013; **230**: 258-267 [PMID: 24075754 DOI: 10.1016/j.atherosclerosis.2013.07.052]
 - 172 **Targher G**, Mantovani A, Pichiri I, Rigolon R, Dauriz M, Zoppini G, Morani G, Vassanelli C, Bonora E. Non-alcoholic fatty liver disease is associated with an increased prevalence of atrial fibrillation in hospitalized patients with type 2 diabetes. *Clin Sci (Lond)* 2013; **125**: 301-309 [PMID: 23596966 DOI: 10.1042/CS20130036]
 - 173 **Targher G**, Valbusa F, Bonapace S, Bertolini L, Zenari L, Rodella S, Zoppini G, Mantovani W, Barbieri E, Byrne CD. Non-alcoholic fatty liver disease is associated with an increased incidence of atrial fibrillation in patients with type 2 diabetes. *PLoS One* 2013; **8**: e57183 [PMID: 23451184 DOI: 10.1371/journal.pone.0057183]
 - 174 **Otto CM**, Prendergast B. Aortic-valve stenosis--from patients at risk to severe valve obstruction. *N Engl J Med* 2014; **371**: 744-756 [PMID: 25140960 DOI: 10.1056/NEJMra1313875]
 - 175 **Rossi A**, Targher G, Zoppini G, Ciccoira M, Bonapace S, Negri C, Stoico V, Faggiano P, Vassanelli C, Bonora E. Aortic and mitral annular calcifications are predictive of all-cause and cardiovascular mortality in patients with type 2 diabetes. *Diabetes Care* 2012; **35**: 1781-1786 [PMID: 22699285 DOI: 10.2337/dc12-0134]
 - 176 **Angulo P**. Long-term mortality in nonalcoholic fatty liver disease: is liver histology of any prognostic significance? *Hepatology* 2010; **51**: 373-375 [PMID: 20101746 DOI: 10.1002/hep.23521]
 - 177 **Tilg H**, Moschen AR. Mechanisms behind the link between obesity and gastrointestinal cancers. *Best Pract Res Clin Gastroenterol* 2014; **28**: 599-610 [PMID: 25194178 DOI: 10.1016/j.bpg.2014.07.006]
 - 178 **Sanna C**, Rosso C, Marietti M, Bugianesi E. Non-Alcoholic Fatty Liver Disease and Extra-Hepatic Cancers. *Int J Mol Sci* 2016; **17**: pii: E717 [PMID: 27187365 DOI: 10.3390/ijms17050717]
 - 179 **Chatterjee S**, Khunti K, Davies MJ. Type 2 diabetes. *Lancet* 2017; **389**: 2239-2251 [PMID: 28190580 DOI: 10.1016/S0140-6736(17)30058-2]
 - 180 **Loomba R**, Abraham M, Unalp A, Wilson L, Lavine J, Doo E, Bass NM; Nonalcoholic Steatohepatitis Clinical Research Network. Association between diabetes, family history of diabetes, and risk of nonalcoholic steatohepatitis and fibrosis. *Hepatology* 2012; **56**: 943-951 [PMID: 22505194 DOI: 10.1002/hep.25772]
 - 181 **Targher G**, Marchesini G, Byrne CD. Risk of type 2 diabetes in patients with non-alcoholic fatty liver disease: Causal association or epiphenomenon? *Diabetes Metab* 2016; **42**: 142-156 [PMID: 27142870 DOI: 10.1016/j.diabet.2016.04.002]
 - 182 **Armstrong MJ**, Adams LA, Canbay A, Syn WK. Extrahepatic complications of nonalcoholic fatty liver disease. *Hepatology* 2014; **59**: 1174-1197 [PMID: 24002776 DOI: 10.1002/hep.26717]
 - 183 **Logue J**, Walker JJ, Leese G, Lindsay R, McKnight J, Morris A, Philip S, Wild S, Sattar N; Scottish Diabetes Research Network Epidemiology Group. Association between BMI measured within a year after diagnosis of type 2 diabetes and mortality. *Diabetes Care* 2013; **36**: 887-893 [PMID: 23139375 DOI: 10.2337/dc12-0944]
 - 184 **Seo DC**, Choe S, Torabi MR. Is waist circumference $\geq 102/88$ cm better than body mass index ≥ 30 to predict hypertension and diabetes development regardless of gender, age group, and race/ethnicity? Meta-analysis. *Prev Med* 2017; **97**: 100-108 [PMID: 28137662 DOI: 10.1016/j.ypmed.2017.01.012]
 - 185 **Camilleri M**, Malhi H, Acosta A. Gastrointestinal Complications of Obesity. *Gastroenterology* 2017; **152**: 1656-1670 [PMID: 28192107 DOI: 10.1053/j.gastro.2016.12.052]
 - 186 **Lionetti L**, Mollica MP, Lombardi A, Cavaliere G, Gifuni G, Barletta A. From chronic overnutrition to insulin resistance: the role of fat-storing capacity and inflammation. *Nutr Metab Cardiovasc Dis* 2009; **19**: 146-152 [PMID: 19171470 DOI: 10.1016/j.numecd.2008.10.010]
 - 187 **Boden G**, Homko C, Barrero CA, Stein TP, Chen X, Cheung P, Fecchio C, Koller S, Merali S. Excessive caloric intake acutely causes oxidative stress, GLUT4 carbonylation, and insulin resistance in healthy men. *Sci Transl Med* 2015; **7**: 304re7 [PMID: 26355033 DOI: 10.1126/scitranslmed.aac4765]
 - 188 **Bugianesi E**, McCullough AJ, Marchesini G. Insulin resistance: a metabolic pathway to chronic liver disease. *Hepatology* 2005; **42**: 987-1000 [PMID: 16250043 DOI: 10.1002/hep.20920]
 - 189 **Loria P**, Marchesini G, Nascimbeni F, Ballestri S, Maurantonio M, Carubbi F, Ratzu V, Lonardo A. Cardiovascular risk, lipidemic phenotype and steatosis. A comparative analysis of cirrhotic and

- non-cirrhotic liver disease due to varying etiology. *Atherosclerosis* 2014; **232**: 99-109 [PMID: 24401223 DOI: 10.1016/j.atherosclerosis.2013.10.030]
- 190 **Yamazaki H**, Tsuboya T, Tsuji K, Dohke M, Maguchi H. Independent Association Between Improvement of Nonalcoholic Fatty Liver Disease and Reduced Incidence of Type 2 Diabetes. *Diabetes Care* 2015; **38**: 1673-1679 [PMID: 26156527 DOI: 10.2337/dc15-0140]
- 191 **Fukuda T**, Hamaguchi M, Kojima T, Mitsuhashi K, Hashimoto Y, Ohbora A, Kato T, Nakamura N, Fukui M. Transient remission of nonalcoholic fatty liver disease decreases the risk of incident type 2 diabetes mellitus in Japanese men. *Eur J Gastroenterol Hepatol* 2016; **28**: 1443-1449 [PMID: 27603300 DOI: 10.1097/MEG.0000000000000736]
- 192 **Strazzullo P**, D'Elia L, Cairella G, Garbagnati F, Cappuccio FP, Scafili L. Excess body weight and incidence of stroke: meta-analysis of prospective studies with 2 million participants. *Stroke* 2010; **41**: e418-e426 [PMID: 20299666 DOI: 10.1161/STROKEAHA.109.576967]
- 193 **Targher G**, Byrne CD, Lonardo A, Zoppini G, Barbui C. Non-alcoholic fatty liver disease and risk of incident cardiovascular disease: A meta-analysis. *J Hepatol* 2016; **65**: 589-600 [PMID: 27212244 DOI: 10.1016/j.jhep.2016.05.013]
- 194 **Koehler EM**, Plompen EP, Schouten JN, Hansen BE, Darwish Murad S, Taimr P, Leebeek FW, Hofman A, Stricker BH, Castera L, Janssen HL. Presence of diabetes mellitus and steatosis is associated with liver stiffness in a general population: The Rotterdam study. *Hepatology* 2016; **63**: 138-147 [PMID: 26171685 DOI: 10.1002/hep.27981]
- 195 **Shima T**, Uto H, Ueki K, Takamura T, Kohgo Y, Kawata S, Yasui K, Park H, Nakamura N, Nakatou T, Tanaka N, Umemura A, Mizuno M, Tanaka J, Okanoue T. Clinicopathological features of liver injury in patients with type 2 diabetes mellitus and comparative study of histologically proven nonalcoholic fatty liver diseases with or without type 2 diabetes mellitus. *J Gastroenterol* 2013; **48**: 515-525 [PMID: 22911170 DOI: 10.1007/s00535-012-0653-5]
- 196 **Pais R**, Charlotte F, Fedchuk L, Bedossa P, Lebray P, Poynard T, Ratzin V; LIDO Study Group. A systematic review of follow-up biopsies reveals disease progression in patients with non-alcoholic fatty liver. *J Hepatol* 2013; **59**: 550-556 [PMID: 23665288 DOI: 10.1016/j.jhep.2013.04.027]
- 197 **Nakahara T**, Hyogo H, Yoneda M, Sumida Y, Eguchi Y, Fujii H, Ono M, Kawaguchi T, Imajo K, Aikata H, Tanaka S, Kanemasa K, Fujimoto K, Anzai K, Saibara T, Sata M, Nakajima A, Itoh Y, Chayama K, Okanoue T; Japan Study Group of Nonalcoholic Fatty Liver Disease. Type 2 diabetes mellitus is associated with the fibrosis severity in patients with nonalcoholic fatty liver disease in a large retrospective cohort of Japanese patients. *J Gastroenterol* 2014; **49**: 1477-1484 [PMID: 24277052 DOI: 10.1007/s00535-013-0911-1]
- 198 **McPherson S**, Hardy T, Henderson E, Burt AD, Day CP, Anstee QM. Evidence of NAFLD progression from steatosis to fibrosing-steatohepatitis using paired biopsies: implications for prognosis and clinical management. *J Hepatol* 2015; **62**: 1148-1155 [PMID: 25477264 DOI: 10.1016/j.jhep.2014.11.034]
- 199 **Maurantonio M**, Ballestri S, Odoardi MR, Lonardo A, Loria P. Treatment of atherogenic liver based on the pathogenesis of nonalcoholic fatty liver disease: a novel approach to reduce cardiovascular risk? *Arch Med Res* 2011; **42**: 337-353 [PMID: 21843565 DOI: 10.1016/j.arcmed.2011.08.004]
- 200 **Bhatia L**, Scorletti F, Curzen N, Clough GF, Calder PC, Byrne CD. Improvement in non-alcoholic fatty liver disease severity is associated with a reduction in carotid intima-media thickness progression. *Atherosclerosis* 2016; **246**: 13-20 [PMID: 26748347 DOI: 10.1016/j.atherosclerosis.2015.12.028]
- 201 **Rao Kondapally Seshasai S**, Kaptoge S, Thompson A, Di Angelantonio E, Gao P, Sarwar N, Whincup PH, Mukamal KJ, Gillum RF, Holme I, Njolstad I, Fletcher A, Nilsson P, Lewington S, Collins R, Gudnason V, Thompson SG, Sattar N, Selvin E, Hu FB, Danesh J; Emerging Risk Factors Collaboration. Diabetes mellitus, fasting glucose, and risk of cause-specific death. *N Engl J Med* 2011; **364**: 829-841 [PMID: 21366474 DOI: 10.1056/NEJMoa1008862]
- 202 **Ong JP**, Pitts A, Younossi ZM. Increased overall mortality and liver-related mortality in non-alcoholic fatty liver disease. *J Hepatol* 2008; **49**: 608-612 [PMID: 18682312 DOI: 10.1016/j.jhep.2008.06.018]
- 203 **Adams LA**, Harmsen S, St Sauver JL, Charatcharoenwithaya P, Enders FB, Therneau T, Angulo P. Nonalcoholic fatty liver disease increases risk of death among patients with diabetes: a community-based cohort study. *Am J Gastroenterol* 2010; **105**: 1567-1573 [PMID: 20145609 DOI: 10.1038/ajg.2010.18]
- 204 **Bugianesi E**, Leone N, Vanni E, Marchesini G, Brunello F, Carucci P, Musso A, De Paolis P, Capussotti L, Salizzoni M, Rizzetto M. Expanding the natural history of nonalcoholic steatohepatitis: from cryptogenic cirrhosis to hepatocellular carcinoma. *Gastroenterology* 2002; **123**: 134-140 [PMID: 12105842]
- 205 **Younossi ZM**, Otgonsuren M, Henry L, Venkatesan C, Mishra A, Erario M, Hunt S. Association of nonalcoholic fatty liver disease (NAFLD) with hepatocellular carcinoma (HCC) in the United States from 2004 to 2009. *Hepatology* 2015; **62**: 1723-1730 [PMID: 26274335 DOI: 10.1002/hep.28123]
- 206 **Rinaldi L**, Nascimbeni F, Giordano M, Masetti C, Guerrero B, Amelia A, Fascione MC, Ballestri S, Romagnoli D, Zampino R, Nevola R, Baldelli E, Iuliano N, Rosato V, Lonardo A, Adinolfi LE. Clinical features and natural history of cryptogenic cirrhosis compared to hepatitis C virus-related cirrhosis. *World J Gastroenterol* 2017; **23**: 1458-1468 [PMID: 28293093 DOI: 10.3748/wjg.v23.i8.1458]
- 207 **Ortiz-Lopez C**, Lomonaco R, Orsak B, Finch J, Chang Z, Kochunov VG, Hardies J, Cusi K. Prevalence of prediabetes and diabetes and metabolic profile of patients with nonalcoholic fatty liver disease (NAFLD). *Diabetes Care* 2012; **35**: 873-878 [PMID: 22374640 DOI: 10.2337/dc11-1849]
- 208 **Hamaguchi E**, Takamura T, Sakurai M, Mizukoshi E, Zen Y, Takeshita Y, Kurita S, Arai K, Yamashita T, Sasaki M, Nakanuma Y, Kaneko S. Histological course of nonalcoholic fatty liver disease in Japanese patients: tight glycemic control, rather than weight reduction, ameliorates liver fibrosis. *Diabetes Care* 2010; **33**: 284-286 [PMID: 19880582 DOI: 10.2337/dc09-0148]
- 209 **Zoppini G**, Fedeli U, Gennaro N, Saugo M, Targher G, Bonora E. Mortality from chronic liver diseases in diabetes. *Am J Gastroenterol* 2014; **109**: 1020-1025 [PMID: 24890439 DOI: 10.1038/ajg.2014.132]
- 210 **Davila JA**. Diabetes and hepatocellular carcinoma: what role does diabetes have in the presence of other known risk factors? *Am J Gastroenterol* 2010; **105**: 632-634 [PMID: 20203644 DOI: 10.1038/ajg.2009.715]
- 211 **Giorda C**, Forlani G, Manti R, Mazzella N, De Cosmo S, Rossi MC, Nicolucci A, Russo G, Di Bartolo P, Ceriello A, Guida P; AMD-Annals Study Group. Occurrence over time and regression of nonalcoholic fatty liver disease in type 2 diabetes. *Diabetes Metab Res Rev* 2017; **33** [PMID: 28032449 DOI: 10.1002/dmrr.2878]
- 212 **Lonardo A**, Sookoian S, Pirola CJ, Targher G. Non-alcoholic fatty liver disease and risk of cardiovascular disease. *Metabolism* 2016; **65**: 1136-1150 [PMID: 26477269 DOI: 10.1016/j.metabol.2015.09.017]
- 213 **Targher G**, Rossini M, Lonardo A. Evidence that non-alcoholic fatty liver disease and polycystic ovary syndrome are associated by necessity rather than chance: a novel hepato-ovarian axis? *Endocrine* 2016; **51**: 211-221 [PMID: 26024975 DOI: 10.1007/s12020-015-0640-8]
- 214 **Targher G**, Lonardo A, Rossini M. Nonalcoholic fatty liver disease and decreased bone mineral density: is there a link? *J Endocrinol Invest* 2015; **38**: 817-825 [PMID: 26003827 DOI: 10.1007/s40618-015-0315-6]
- 215 **Lonardo A**, Lombardini S, Scaglioni F, Ballestri S, Verrone AM, Bertolotti M, Carulli L, Ganazzi D, Carulli N, Loria P. Fatty liver, carotid disease and gallstones: a study of age-related associations.

- World J Gastroenterol* 2006; **12**: 5826-5833 [PMID: 17007049 DOI: 10.3748/wjg.v12.i36.5826]
- 216 **Jaruvongvanich V**, Sanguankeo A, Upala S. Significant Association Between Gallstone Disease and Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-Analysis. *Dig Dis Sci* 2016; **61**: 2389-2396 [PMID: 26993825 DOI: 10.1007/s10620-016-4125-2]
 - 217 **Chávez-Talavera O**, Tailleux A, Lefebvre P, Staels B. Bile Acid Control of Metabolism and Inflammation in Obesity, Type 2 Diabetes, Dyslipidemia, and Nonalcoholic Fatty Liver Disease. *Gastroenterology* 2017; **152**: 1679-1694.e3 [PMID: 28214524 DOI: 10.1053/j.gastro.2017.01.055]
 - 218 **Lonardo A**, Loria P, Carulli N. Concurrent non-alcoholic steatohepatitis and psoriasis. Report of three cases from the POLI. S.T.E.N.A. study. *Dig Liver Dis* 2001; **33**: 86-87 [PMID: 11303985]
 - 219 **Candia R**, Ruiz A, Torres-Robles R, Chávez-Tapia N, Méndez-Sánchez N, Arrese M. Risk of non-alcoholic fatty liver disease in patients with psoriasis: a systematic review and meta-analysis. *J Eur Acad Dermatol Venerol* 2015; **29**: 656-662 [PMID: 25418531 DOI: 10.1111/jdv.12847]
 - 220 **Lonardo A**, Carani C, Carulli N, Loria P. 'Endocrine NAFLD' a hormonocentric perspective of nonalcoholic fatty liver disease pathogenesis. *J Hepatol* 2006; **44**: 1196-1207 [PMID: 16618516 DOI: 10.1016/j.jhep.2006.03.005]
 - 221 **Loria P**, Carulli L, Bertolotti M, Lonardo A. Endocrine and liver interaction: the role of endocrine pathways in NASH. *Nat Rev Gastroenterol Hepatol* 2009; **6**: 236-247 [PMID: 19347015 DOI: 10.1038/nrgastro.2009.33]
 - 222 **Chi HC**, Chen SL, Tsai CY, Chuang WY, Huang YH, Tsai MM, Wu SM, Sun CP, Yeh CT, Lin KH. Thyroid hormone suppresses hepatocarcinogenesis via DAPK2 and SQSTM1-dependent selective autophagy. *Autophagy* 2016; **12**: 2271-2285 [PMID: 27653365 DOI: 10.1080/15548627.2016.1230583]
 - 223 **Perra A**, Plateroti M, Columbano A. T3/TRs axis in hepatocellular carcinoma: new concepts for an old pair. *Endocr Relat Cancer* 2016; **23**: R353-R369 [PMID: 27353037 DOI: 10.1530/ERC-16-0152]
 - 224 **Frau C**, Loi R, Petrelli A, Perra A, Menegon S, Kowalik MA, Pinna S, Leoni VP, Fornari F, Gramantieri L, Ledda-Columbano GM, Giordano S, Columbano A. Local hypothyroidism favors the progression of preneoplastic lesions to hepatocellular carcinoma in rats. *Hepatology* 2015; **61**: 249-259 [PMID: 25156012 DOI: 10.1002/hep.27399]
 - 225 **Bano A**, Chaker L, Plompen EP, Hofman A, Dehghan A, Franco OH, Janssen HL, Darwish Murad S, Peeters RP. Thyroid Function and the Risk of Nonalcoholic Fatty Liver Disease: The Rotterdam Study. *J Clin Endocrinol Metab* 2016; **101**: 3204-3211 [PMID: 27270473 DOI: 10.1210/jc.2016-1300]
 - 226 **Hannah WN Jr**, Harrison SA. Effect of Weight Loss, Diet, Exercise, and Bariatric Surgery on Nonalcoholic Fatty Liver Disease. *Clin Liver Dis* 2016; **20**: 339-350 [PMID: 27063273 DOI: 10.1016/j.cld.2015.10.008]
 - 227 **Katsagoni CN**, Georgoulis M, Papatheodoridis GV, Panagiotakos DB, Kontogianni MD. Effects of lifestyle interventions on clinical characteristics of patients with non-alcoholic fatty liver disease: A meta-analysis. *Metabolism* 2017; **68**: 119-132 [PMID: 28183444 DOI: 10.1016/j.metabol.2016.12.006]
 - 228 **Sabag A**, Way KL, Keating SE, Sultana RN, O'Connor HT, Baker MK, Chuter VH, George J, Johnson NA. Exercise and ectopic fat in type 2 diabetes: A systematic review and meta-analysis. *Diabetes Metab* 2017; **43**: 195-210 [PMID: 28162956 DOI: 10.1016/j.diabet.2016.12.006]
 - 229 **Keating SE**, George J, Johnson NA. The benefits of exercise for patients with non-alcoholic fatty liver disease. *Expert Rev Gastroenterol Hepatol* 2015; **9**: 1247-1250 [PMID: 26289101 DOI: 10.1586/17474124.2015.1075392]
 - 230 **Demyen M**, Alkhaloufi K, Pyrsopoulos NT. Lipid-lowering agents and hepatotoxicity. *Clin Liver Dis* 2013; **17**: 699-714, x [PMID: 24099026 DOI: 10.1016/j.cld.2013.07.016]
 - 231 **Dongiovanni P**, Petta S, Mannisto V, Mancina RM, Pipitone R, Karja V, Maggioni M, Kakela P, Wiklund O, Mozzi E, Grimaudo S, Kaminska D, Rametta R, Craxi A, Fargion S, Nobili V, Romeo S, Pihlajamaki J, Valenti L. Statin use and non-alcoholic steatohepatitis in at risk individuals. *J Hepatol* 2015; **63**: 705-712 [PMID: 25980762 DOI: 10.1016/j.jhep.2015.05.006]
 - 232 **Nascimbeni F**, Aron-Wisniewsky J, Pais R, Tordjman J, Poitou C, Charlotte F, Bedossa P, Poynard T, Clément K, Ratziu V, LIDO study Group. Statins, antidiabetic medications and liver histology in patients with diabetes with non-alcoholic fatty liver disease. *BMJ Open Gastroenterol* 2016; **3**: e000075 [PMID: 27110380 DOI: 10.1136/bmjgast-2015-000075]
 - 233 **Zhou YY**, Zhu GQ, Wang Y, Zheng JN, Ruan LY, Cheng Z, Hu B, Fu SW, Zheng MH. Systematic review with network meta-analysis: statins and risk of hepatocellular carcinoma. *Oncotarget* 2016; **7**: 21753-21762 [PMID: 26943041 DOI: 10.18632/oncotarget.7832]
 - 234 **Simon TG**, Bonilla H, Yan P, Chung RT, Butt AA. Atorvastatin and fluvastatin are associated with dose-dependent reductions in cirrhosis and hepatocellular carcinoma, among patients with hepatitis C virus: Results from ERCHIVES. *Hepatology* 2016; **64**: 47-57 [PMID: 26891205 DOI: 10.1002/hep.28506]
 - 235 **Lonardo A**, Loria P. Potential for statins in the chemoprevention and management of hepatocellular carcinoma. *J Gastroenterol Hepatol* 2012; **27**: 1654-1664 [PMID: 22849701 DOI: 10.1111/j.1440-1746.2012.07232.x]
 - 236 **Björkhem-Bergman L**, Backheden M, Söderberg Löfdal K. Statin treatment reduces the risk of hepatocellular carcinoma but not colon cancer-results from a nationwide case-control study in Sweden. *Pharmacoepidemiol Drug Saf* 2014; **23**: 1101-1106 [PMID: 25074765 DOI: 10.1002/pds.3685]
 - 237 **McGlynn KA**, Divine GW, Sahasrabudhe VV, Engel LS, VanSlooten A, Wells K, Yood MU, Alford SH. Statin use and risk of hepatocellular carcinoma in a U.S. population. *Cancer Epidemiol* 2014; **38**: 523-527 [PMID: 25113938 DOI: 10.1016/j.canep.2014.06.009]
 - 238 **Singh S**, Singh PP, Roberts LR, Sanchez W. Chemopreventive strategies in hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol* 2014; **11**: 45-54 [PMID: 23938452 DOI: 10.1038/nrgastro.2013.143]
 - 239 **He L**, Liu X, Wang L, Yang Z. Thiazolidinediones for nonalcoholic steatohepatitis: A meta-analysis of randomized clinical trials. *Medicine (Baltimore)* 2016; **95**: e4947 [PMID: 27759627 DOI: 10.1097/MD.0000000000004947]
 - 240 **Musso G**, Cassader M, Paschetta E, Gambino R. Thiazolidinediones and Advanced Liver Fibrosis in Nonalcoholic Steatohepatitis: A Meta-analysis. *JAMA Intern Med* 2017; **177**: 633-640 [PMID: 28241279 DOI: 10.1001/jamainternmed.2016.9607]
 - 241 **Armstrong MJ**, Gaunt P, Aithal GP, Barton D, Hull D, Parker R, Hazlehurst JM, Guo K; LEAN trial team, Abouda G, Aldersley MA, Stocken D, Gough SC, Tomlinson JW, Brown RM, Hübscher SG, Newsome PN. Liraglutide safety and efficacy in patients with non-alcoholic steatohepatitis (LEAN): a multicentre, double-blind, randomised, placebo-controlled phase 2 study. *Lancet* 2016; **387**: 679-690 [PMID: 26608256 DOI: 10.1016/S0140-6736(15)00803-X]
 - 242 **Sato K**, Goshō M, Yamamoto T, Kobayashi Y, Ishii N, Ohashi T, Nakade Y, Ito K, Fukuzawa Y, Yoneda M. Vitamin E has a beneficial effect on nonalcoholic fatty liver disease: a meta-analysis of randomized controlled trials. *Nutrition* 2015; **31**: 923-930 [PMID: 26059365 DOI: 10.1016/j.nut.2014.11.018]
 - 243 **Singh S**, Khara R, Allen AM, Murad MH, Loomba R. Comparative effectiveness of pharmacological interventions for nonalcoholic steatohepatitis: A systematic review and network meta-analysis. *Hepatology* 2015; **62**: 1417-1432 [PMID: 26189925 DOI: 10.1002/hep.27999]
 - 244 **Sawangjit R**, Chongmelaxme B, Phisalprapa P, Saokaew S, Thakkinstian A, Kowdley KV, Chaiyakunapruk N. Comparative efficacy of interventions on nonalcoholic fatty liver disease (NAFLD): A PRISMA-compliant systematic review and network meta-analysis. *Medicine (Baltimore)* 2016; **95**: e4529 [PMID: 27099026 DOI: 10.1016/j.cld.2013.07.016]

- 27512874 DOI: 10.1097/MD.0000000000004529]
- 245 **Martagón AJ**, Lin JZ, Cimini SL, Webb P, Phillips KJ. The amelioration of hepatic steatosis by thyroid hormone receptor agonists is insufficient to restore insulin sensitivity in ob/ob mice. *PLoS One* 2015; **10**: e0122987 [PMID: 25849936 DOI: 10.1371/journal.pone.0122987]
- 246 **Nakatsu Y**, Seno Y, Kushiya A, Sakoda H, Fujishiro M, Katasako A, Mori K, Matsunaga Y, Fukushima T, Kanaoka R, Yamamotoya T, Kamata H, Asano T. The xanthine oxidase inhibitor febuxostat suppresses development of nonalcoholic steatohepatitis in a rodent model. *Am J Physiol Gastrointest Liver Physiol* 2015; **309**: G42-G51 [PMID: 25999428 DOI: 10.1152/ajpgi.00443.2014]
- 247 **Ratzliff V**, Harrison SA, Francque S, Bedossa P, Leher P, Serfaty L, Romero-Gomez M, Boursier J, Abdelmalek M, Caldwell S, Drenth J, Anstee QM, Hum D, Hanf R, Roudot A, Megnier S, Staels B, Sanyal A; GOLDEN-505 Investigator Study Group. Elafibranor, an Agonist of the Peroxisome Proliferator-Activated Receptor- α and - δ , Induces Resolution of Nonalcoholic Steatohepatitis Without Fibrosis Worsening. *Gastroenterology* 2016; **150**: 1147-1159.e5 [PMID: 26874076 DOI: 10.1053/j.gastro.2016.01.038]
- 248 **Kim W**, Kim BG, Lee JS, Lee CK, Yeon JE, Chang MS, Kim JH, Kim H, Yi S, Lee J, Cho JY, Kim SG, Lee JH, Kim YJ. Randomised clinical trial: the efficacy and safety of oltipraz, a liver X receptor alpha-inhibitory dithiolethione in patients with non-alcoholic fatty liver disease. *Aliment Pharmacol Ther* 2017; **45**: 1073-1083 [PMID: 28225186 DOI: 10.1111/apt.13981]

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New therapeutic perspectives in irritable bowel syndrome: Targeting low-grade inflammation, immuno-neuroendocrine axis, motility, secretion and beyond

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Abstract

Irritable bowel syndrome (IBS) is a chronic, recurring, and remitting functional disorder of the gastrointestinal tract characterized by abdominal pain, distention, and changes in bowel habits. Although there are several drugs for IBS, effective and approved treatments for one or more of the symptoms for various IBS subtypes are needed. Improved understanding of pathophysiological mechanisms such as the role of impaired bile acid metabolism, neurohormonal regulation, immune

dysfunction, the epithelial barrier and the secretory properties of the gut has led to advancements in the treatment of IBS. With regards to therapies for restoring intestinal permeability, multiple studies with prebiotics and probiotics are ongoing, even if to date their efficacy has been limited. In parallel, much progress has been made in targeting low-grade inflammation, especially through the introduction of drugs such as mesalazine and rifaximin, even if a better knowledge of the mechanisms underlying the low-grade inflammation in IBS may allow the design of clinical trials that test the efficacy and safety of such drugs. This literature review aims to summarize the findings related to new and investigational therapeutic agents for IBS, most recently developed in preclinical as well as Phase 1 and Phase 2 clinical studies.

Key words: Therapy; Low grade inflammation; Motility; Secretion; Irritable bowel syndrome; Immunoendocrine axis

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Core tip: Irritable bowel syndrome (IBS) is a chronic, recurring, and remitting functional disorder of the gastrointestinal tract characterized by abdominal pain, distention, and changes in bowel habits. Despite there are several drugs for IBS, effective and approved treatments for one or more of the symptoms for various IBS subtypes are needed. The understanding of pathophysiological mechanisms such as the role of impaired bile acid metabolism, neurohormonal regulation, immune dysfunction, the epithelial barrier and secretory properties of the gut has led to advancements in the treatment of IBS. This literature review aims to summarize the findings relating the new and investigational therapeutic agents for IBS, most recently developed in preclinical as well as Phase 1 and Phase 2 clinical studies.

Sinagra E, Morreale GC, Mohammadian G, Fusco G, Guarnotta V, Tomasello G, Cappello F, Rossi F, Amvrosiadis G, Raimondo D. New therapeutic perspectives in irritable bowel syndrome: Targeting low-grade inflammation, immuno-neuroendocrine axis, motility, secretion and beyond. *World J Gastroenterol* 2017; 23(36): 6593-6627 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6593.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6593>

INTRODUCTION

Irritable bowel syndrome (IBS) is a chronic, recurring, and remitting functional disorder of the gastrointestinal (GI) tract characterized by abdominal pain, distention, and changes in bowel habits that do not have a known structural or anatomical explanation^[1].

IBS is a global problem, with anywhere from 5% to 15% of the general population showing symptoms that would satisfy a definition of IBS^[2-4]. IBS considerably affects quality of life and imposes a profound burden on patients, physicians and the health-care system^[5]. For example, the IBIS-C study recently assessed the socio-economic burden of moderate-to-severe IBS with constipation in six European countries (France, Germany, Italy, Spain, Sweden and the United Kingdom), showing that IBS represents a main cause of absenteeism in the workplace^[6].

Regarding the sex-related prevalence of IBS, in Western countries, the prevalence of IBS in women outnumbers that in men by 2:1^[7,8], and within the patient population who have consultations with primary care physicians, women outnumber men by 3:1^[7,9]. Finally, in tertiary care settings, the number of women with IBS is 4 to 5 times higher than the number of men^[7-10].

According to Rome III, IBS is defined based on the presence of: recurrent abdominal pain or discomfort at least 3 d/mo in the past 3 mo associated with two or more of the following: (1) improvement with defecation; (2) onset associated with a change in frequency of stool; and (3) onset associated with a change in form (appearance) of stool.

These criteria should be fulfilled for the past 3 mo with symptom onset at least 6 mo before diagnosis^[11]. Recently, the Rome IV criteria implemented the knowledge accumulated since Rome III was published almost ten years ago.

According to Rome IV, IBS is defined on the basis of the presence of: Recurrent abdominal pain, on average, at least 1 d per week in the last 3 mo, associated with 2 or more of the following criteria: (1) related to defecation; (2) associated with a change in frequency of stool; and (3) associated with a change in form (appearance) of stool. These criteria should be fulfilled for the last 3 mo with symptom onset at least 6 mo before diagnosis^[12].

In contrast to the Rome III criteria, the term discomfort has been deleted from the last definition and from subsequent diagnostic criteria because not all languages have the term "discomfort". This word has different meanings in different languages, which can result in ambiguity with patients^[12]. Furthermore, the last definition implies a change in the frequency of abdominal pain, highlighting that patients should have symptoms of abdominal pain at least 1 d per week during the past 3 mo^[12]. Finally, the sentence "improvement with defecation" was substituted in the current definition by "related to defecation", as a large subset of IBS patients do not have an improvement in abdominal pain with defecation but instead complain of worsening^[12].

According to the Rome IV criteria, IBS is subtyped according to the predominant bowel habit as follows:

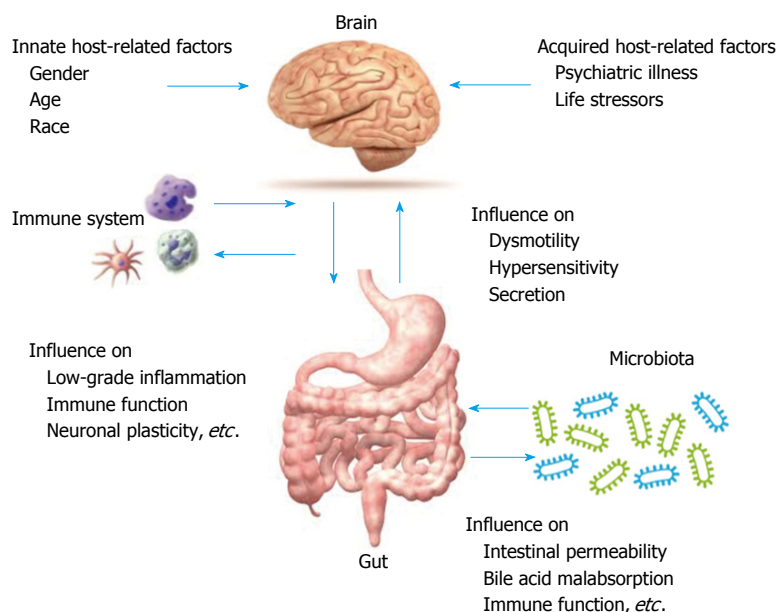


Figure 1 Complex interplay between low-grade inflammation, immuno-neuroendocrine axis, and microbiota. Brain (influenced by multiple innate and acquired factors) and gut interact bidirectionally to shape the clinical phenotype of irritable bowel syndrome (IBS). This bi-directional pathway acts not only on gastrointestinal motility, visceral sensitivity and secretion; however, the influence of both the immune system and microbiota modulates several functions that could create the definitive clinical phenotype of IBS.

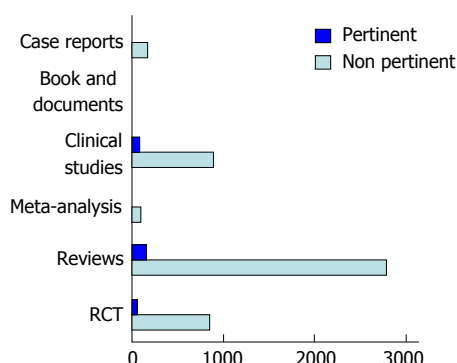


Figure 2 Literature findings on the relationship between irritable bowel syndrome and inflammation ($n = 317$). RCT: Randomized controlled trials.

IBS with constipation (IBS-C), IBS with diarrhoea (IBS-D), mixed type (IBS-M), and unclassified (IBS-U)^[12]. The definition of bowel habit type is based on the patient's description of the stool form by referring to the Bristol Stool Scale^[13]. Furthermore, IBS patients can be grouped into sporadic (nonspecific) and post-infectious (PI-IBS)/inflammatory bowel disease (IBD)-associated (IBD-IBS)^[14,15].

Although there are several drugs for IBS in the pipeline, there is a continuous need for effective and approved treatments for one or more of the symptoms of IBS subtypes^[16-18]. The understanding of pathophysiological mechanisms such as the role of altered bile acid metabolism, neurohormonal regulation, immune dysfunction, the epithelial barrier and secretory properties of the gut has led to progress in the treatment options of IBS (Figure 1)^[18,19].

This literature review aims to summarize the

findings relating the new and investigational therapeutic agents for IBS most recently developed in preclinical as well as Phase 1 and Phase 2 clinical studies.

MATERIALS AND METHODS

We carried out a bibliographic search in MEDLINE for the period January 1966 to December 2016 and focused on identifying publications describing the new therapeutic pharmacological approaches in IBS. Information was also obtained from abstracts and the latest results found in the Clinicaltrial.gov database. The keywords used were: irritable bowel syndrome, inflammation, immunoendocrine axis, intestinal permeability, IBS-C, IBS-D, therapy. The inclusion criteria to select articles were based on design (systematic reviews, meta-analysis, clinical trials, and experimental studies on animals) and population (adult patients > 18 years of age). We excluded articles not relevant for this topic.

According to the abovementioned criteria, 5127 studies were found and 4810 studies were excluded because they were not relevant for this topic (Figure 2).

LOW-GRADE INFLAMMATION IN IRRITABLE BOWEL SYNDROME

Recently, the scientific community has focused its attention on the pivotal role of low-grade mucosal inflammation in IBS, considering evidence showing that some patients with IBS have an increased number of inflammatory cells in the colonic and ileal mucosa, with regard to control patients^[20].

In fact, the intestinal mucosa harbours a florid immune system that can be regarded as “physiologically inflamed”^[20,21]. Thus, low-grade inflammation, which likely plays a multifactorial role in IBS pathophysiology, can only be evaluated using quantitative assessments^[20-22].

The available data^[23,24] on low-grade inflammation in IBS patients is often expressed as average numbers and are mainly focused on IBS-D. Thus, it is unclear whether this event occurs only in selected subsets of IBS patients^[25].

Therefore, IBS could be considered a micro-organic disease, where there is an increased number of mucosal immunocytes (*i.e.*, mast cells, eosinophils, and T cells) in adult and paediatric patients. Several precipitating factors have been claimed, including food allergy, abnormal microbiota, bile acid malabsorption, and increased intestinal permeability^[26]. The magnitude of the inflammatory response is several-fold less than that seen in acute inflammation in inflammatory bowel disease. The above-reported evidence provides a rationale to evaluate the efficacy of intestinal anti-inflammatory therapies in patients with IBS that we will touch upon in the next section.

ANTI-INFLAMMATORY THERAPIES IN IRRITABLE BOWEL SYNDROME

Corticosteroids

In the study performed by Dunlop *et al.*^[24], twenty-nine patients with post-infectious irritable bowel syndrome underwent a randomized, double-blind, placebo-controlled trial of 3 wk of oral prednisolone, 30 mg/d. Mucosal enterochromaffin cells, T lymphocytes and mast cells were evaluated in rectal biopsies before and after treatment, and bowel symptoms were reported in a daily diary. In this study, enterochromaffin cell counts did not change significantly after either prednisolone or placebo. Although lamina propria T-lymphocyte counts decreased significantly after prednisolone, but not after placebo, this was not linked with any significant treatment-related improvement in abdominal pain, diarrhoea, frequency or urgency^[24].

Antibiotics

Rifaximin is a rifamycin derivative that acts by inhibiting bacterial ribonucleic acid (RNA) synthesis. It is virtually unabsorbed after oral administration, so it is used mainly to treat local dysfunctions within the gastrointestinal tract^[27].

The Food and Drug Administration (FDA) initially approved rifaximin to treat traveller's diarrhoea caused by *Escherichia coli* and to prevent the recurrence of hepatic encephalopathy.

Successively, the FDA approved rifaximin in IBS-D “naïve” patients at a dose of 550 mg three times a

day for 14 d as well as in patients experiencing a recurrence of symptoms.

Rifaximin improves IBS symptoms through a variety of mechanisms directed at the gastrointestinal tract. In fact, much evidence from animal experiments shows that rifaximin either improves or maintains microbiota diversity and bacterial composition in IBS, reduces intestinal cytokine inflammation, provides gut-barrier protection preventing attachment and internalization of coliforms and pathogens with reduced epithelial cell inflammation and pathogen-induced inflammatory response, and reduces visceral hyperalgesia^[28].

In a combined analysis of two separate Phase 3 trials (TARGET 1 and 2), a 14-d course of rifaximin 550 mg three times daily in IBS-D patients significantly increased the percentage of relief of global IBS symptoms and improved IBS-related distention and abdominal pain, discomfort, and loose or watery stools compared with placebo for up to 10 wk post-treatment^[29,30].

Successively, TARGET 3 was performed to test the safety and efficacy of a repeated treatment with rifaximin in patients experiencing a recurrence of IBS symptoms. In this study, the percentage of responders during the 18-wk follow-up (in terms of pain and stool consistency improvements) to randomized repeat treatment was significantly greater with rifaximin vs placebo^[31]. The safety profile of rifaximin in patients with IBS-D was generally similar to that observed with placebo^[30].

In fact, constipation was only reported in 1 (0.3%) patient in the rifaximin group and 3 (1.0%) patients in the placebo group. Only one patient in each treatment group suspended the drug. One case of *Clostridium difficile* infection occurred (in a patient who had been off of rifaximin for several weeks but was receiving a concomitant systemic antibiotic)^[30,31].

In conclusion, these trials show that a 2-wk course of rifaximin could improve IBS-D-related symptoms, and in the case of persistence of symptoms, retreatment may ameliorate abdominal pain and stool consistency with possible improvements in bloating and stool urgency in some patients. While patients were retreated within an 18-wk period of follow-up in the study, it is still unclear as to when and how often treatment should be given. In addition, the identification of those patients who might likely respond to rifaximin remains to be investigated.

Recently, Ghoshal *et al.*^[32] evaluated symptom resolution among IBS patients with or without small intestinal bacterial overgrowth (SIBO) on norfloxacin treatment and its efficacy in obtaining negative SIBO test results as compared with placebo. In this study, 80 IBS patients (Rome III) were evaluated for SIBO by gut aspirate culture. Patients with a colony count ≥ 10 CFU/mL and those without SIBO were separately randomized to 800 mg/d norfloxacin for 10 d or placebo. The global symptom score (blind), Rome III

criteria, aspirate culture, and glucose hydrogen breath test were assessed before and 1 mo after treatment, and patients were followed up for 6 mo. Although norfloxacin was more effective at decreasing the symptom score at 1 mo among patients with compared with those without SIBO but not placebo, the scores were comparable at 6 mo. Symptoms more often resolved to turn Rome III negative in SIBO patients treated with norfloxacin compared with placebo at 1 mo. Patients without SIBO and a colony count of 10 CFU/mL responded more than those with a colony count less than 10 CFU/mL^[32].

Mast cell stabilizers

Since mast cell activation was thought to be involved in visceral hypersensitivity, a study was undertaken by Klooker *et al.*^[33] to evaluate the effect of ketotifen, a mast cell stabilizer, on rectal sensitivity and symptoms in patients with IBS. In this case-control study, 60 patients with IBS underwent a barostat study to assess rectal sensitivity before and after 8 wk of treatment. After the initial barostat, patients were randomised to receive ketotifen or placebo. Ketotifen increased the threshold for discomfort in patients with IBS and visceral hypersensitivity but not placebo. This effect was not observed in normosensitive patients with IBS. Ketotifen significantly reduced abdominal pain and other IBS symptoms and improved quality of life. However, whether this effect was secondary to the mast cell stabilising properties of ketotifen or H1 receptor antagonism remains a topic of future research^[33].

Successively, Lobo *et al.*^[34] showed a clinical Benefit of Disodium Cromoglycate (DSCG) in IBS in a double-blind, placebo-controlled clinical assay with prolonged (6 mo) oral administration of DSCG (DSCG), since it induces mast cell-mediated recovery of the healthy-like innate immunity gene expression profile in the jejunal mucosa^[34].

Finally, since histamine sensitizes the nociceptor transient reporter potential channel V1 (TRPV1) and has been observed to play role in visceral hypersensitivity in animals, Wouters *et al.*^[35,36] investigated the role of ebastine, an antagonist of histamine receptor H1 (HRH1), in reducing symptoms of patients in a randomized placebo-controlled trial. After a 2-wk run-in period, subjects were enrolled randomly to groups given either the HRH1 antagonist ebastine or placebo for 12 wk. Rectal biopsy specimens were collected, barostat studies were performed, and symptoms were recorded (using the validated gastrointestinal symptom rating scale) before and after the 12-wk period. Patients were followed up for a further 2 wk. The primary end point of the study was the evaluation of ebastine efficacy on the symptom score evoked by rectal distension. Compared with the placebo group, patients treated with ebastine had reduced visceral hypersensitivity, increased symptom relief, and reduced

abdominal pain scores^[35,36].

Mesalazine

The therapeutic potential of aminosalicylates, whose benefits in chronic inflammatory bowel diseases are well known, has been focused on as a potential cure for IBS^[37,38].

The largest studies on mesalazine in IBS have been conducted by Barbara and Lam. Barbara *et al.*^[39] conducted a phase 3, multicentre, tertiary setting, randomised, double-blind placebo-controlled trial in patients with Rome III-confirmed IBS. Patients were randomly assigned to either 800 mg mesalazine or placebo three times daily for 12 wk and were followed for an additional 12 wk. The primary efficacy endpoint was satisfactory relief of abdominal pain/discomfort for at least half of the weeks of the treatment period. The secondary endpoint was satisfactory relief of overall IBS symptoms. The responder patients were 68.6% in the mesalazine group vs 67.4% in the placebo group. However, with the 75% rule or > 75% rule, there was a higher percentage of responders in the mesalazine group than placebo of 11.6% and 5.9%, respectively, although these differences were not significant. For the key secondary endpoint, in the mesalazine group, overall symptom improvement was observed and a significant difference of 15.1% vs placebo with the > 75% rule was reached. The authors concluded that mesalazine treatment was not superior to placebo on the study primary endpoint, but a subgroup of patients with IBS had a sustained therapy response and benefits from mesalazine therapy^[39].

On the other hand, Lam *et al.*^[40] conducted a double-blind, randomised placebo-controlled trial of 2 g mesalazine twice daily compared with placebo for 3 mo in Rome III criteria patients with IBS-D. The authors compared the mesalazine and placebo effects on stool frequency as the primary endpoint and secondarily assessed the effect of mesalazine on abdominal pain, stool consistency, urgency and satisfactory relief of IBS symptoms. In total, 136 IBS-D patients (82 female, 54 male) were enrolled; 10 patients withdrew from each group. The intention to treat analysis showed that the mean daily stool frequency during weeks 11 and 12 was 2.8 (SD 1.2) in the mesalazine group and 2.7 (SD 1.9) in the placebo group, with a group difference of 0.1. The authors concluded that mesalazine did not ameliorate abdominal pain, stool consistency or percentage with satisfactory relief compared with placebo during the last 2 weeks' follow-up. However a post hoc analysis in 13 post-infectious patients with IBS tended to show benefit, even though this finding needs to be confirmed in larger studies^[40].

A point of weakness of these studies is that the use of endpoints for response may be easily met by patients in the placebo arm, resulting in placebo response rates of almost 70% for satisfactory relief

Table 1 Summary of the literature findings about anti-inflammatory therapies in irritable bowel syndrome

Drug	Ref.	No. of patients	Study design	Outcome
Corticosteroids (prednisolone)	Dunlop <i>et al</i> ^[24]	29 patients with post-infectious irritable bowel syndrome	Randomized, double-blind, placebo-controlled trial of 3 wk of oral prednisolone, 30 mg/d	Not associated with any significant treatment-related improvement in abdominal pain, diarrhoea, frequency or urgency
Antibiotics (Rifaximin)	Pimentel <i>et al</i> ^[29] Target 1 e 2	623 IBS patients in TARGET 1 and 637 IBS in TARGET 2	Phase 3 trials, 14 d with rifaximin 550 mg 3 times daily	Significantly increased the percentage of relief of global IBS symptoms and improved IBS-related bloating and abdominal pain, discomfort, and loose or watery stools, with regard to placebo for up to 10 wk post-treatment
Antibiotics (norfloxacin)	Ghoshal <i>et al</i> ^[32]	80 IBS patients evaluate for SIBO	Randomized, double-blind, placebo-controlled trial; patients were randomized to 800 mg/d norfloxacin for 10 d or placebo	Although norfloxacin was more effective at reducing the symptom score at 1 mo among patients with compared with those without SIBO but not placebo, the scores were comparable at 6 mo. Symptoms more often resolved to turn Rome III negative in SIBO patients treated with norfloxacin compared with placebo at 1 mo
Mast cell stabilizers (Ketotifen)	Klooker <i>et al</i> ^[33]	60 IBS patients	Case Control study; abarostat study to assess rectal sensitivity before and after 8 wk of treatment and, after the initial barostat, patients were randomised to receive ketotifen or placebo	Ketotifen but not placebo increased the threshold for discomfort in patients with IBS with visceral hypersensitivity, but this effect was not observed in normosensitive patients with IBS. Ketotifen significantly decreased abdominal pain and other IBS symptoms and improved quality of life
Mast cells stabilizers (DSCG)	Lobo <i>et al</i> ^[34]		Randomized, double-blind, placebo-controlled trial; with prolonged (6 mo) oral administration of DSCG	Induces Mast Cell-Mediated Recovery of Healthy-Like Innate Immunity Genes Expression Profile in the Jejunal Mucosa
Mast cells stabilizers (ebastin)	Wouters <i>et al</i> ^[35]	65 IBS patients	Double-blind placebo-controlled trial, after 2-wk run-in period, subjects were assigned randomly to groups ebastine (20 mg/d; <i>n</i> = 28) or placebo (<i>n</i> = 27) for 12 wk	Compared with subjects given placebo, those given ebastine had reduced visceral hypersensitivity, increased symptom relief, and reduced abdominal pain scores
Mesalazine	Barbara <i>et al</i> ^[39]	185 patients with IBS	A phase 3, multicentre, tertiary setting, randomised, double-blind, placebo-controlled trial in patients with Rome III confirmed IBS. Patients were randomly assigned to either mesalazine, 800 mg, or placebo, three times daily for 12 wk, and were followed for additional 12 wk	Mesalazine treatment was not superior than placebo on the study primary endpoint, but a subgroup of patients with IBS showed a sustained therapy response and benefits from a mesalazine therapy
	Lam <i>et al</i> ^[40]	136 patients with IBS-D	A double-blind, randomised placebo-controlled trial of 2 g mesalazine twice daily compared with placebo for 3 mo	The authors concluded that mesalazine did not improve abdominal pain, stool consistency or percentage with satisfactory relief compared with placebo during the last 2 weeks' follow-up, however a post hoc analysis in 13 post-infectious patients with IBS appeared to show benefit but this needs confirmation in a larger group ^[40]

IBS : Irritable bowel syndrome.

of abdominal pain or discomfort and > 60% for satisfactory relief of overall IBS symptoms in the trial performed by Barbara *et al*^[39] and in > 40% for satisfactory relief of IBS symptoms in the trial performed by Min *et al*^[41].

It may have therefore have been preferable to use a once daily dosing schedule in both trials in order to reduce the placebo response rates, thus increasing the likelihood of detecting a statistically significant difference between mesalazine and placebo.

Based on this evidence, it is necessary that further studies prove the efficacy of mesalazine for IBS. Studies aimed at evaluating the role of aminosali-

cylates and other potential anti-inflammatory treatment options, including probiotics, non-absorbable antibiotics, histamine receptor antagonists and protease inhibitors on IBS symptoms or pathophysiology are now warranted^[39].

Table 1 sums up the literature findings about anti-inflammatory therapies in irritable bowel syndrome.

INTESTINAL PERMEABILITY IN IRRITABLE BOWEL SYNDROME

An increase in intestinal permeability can be seen in

many conditions, such as infectious gastroenteritis and irritable bowel disease^[42]. The intestinal barrier has long been a focus of gastroenterological research^[43] and its role in IBS has been discussed in many studies. Most studies show an increase in intestinal permeability of patients with IBS-D and post-infectious IBS (PI-IBS)^[43-46].

Among the first to describe intestinal permeability in patients with PI-IBS were Spiller *et al.*^[47], who detected an increased lactulose/mannitol ratio in the urine of IBS patients compared to healthy controls.

Marshall *et al.*^[48] also described an increase in permeability of patients with IBS after an outbreak of bacterial gastroenteritis but could not show a difference in permeability between PI-IBS and non PI-IBS.

There are genetic risk factors for developing PI-IBS and CDH1, which codes for E-cadherin, a tight junction (TJ) protein that is involved in the epithelial barrier function of the gut^[49], hence suggesting the pathophysiological mechanism through which some patients experience increased permeability.

The mechanism of increased permeability in patients with IBS is suggested to involve tight junction dysfunction or involvement of the adherence proteins^[44]. Among factors that could influence permeability is stress. Male soldiers were evaluated in a prospective study during and after combat training with an increase in physiological and psychological stress. Their training induced an increase in gastrointestinal symptoms and alteration in the permeability of the gut barrier^[50]. Since stress has been suggested to be one of the pathophysiological factors involved in developing IBS, this mechanism could explain the reason for the gastrointestinal symptoms^[51].

Another factor that has been evaluated is the intraluminal content of patients with IBS, where faecal supernatants from patients have increased the colonic permeability in mice^[52,53].

Both intracellular [zonula occludens (ZO)-1, ZO-2, and ZO-3, and cingulin] and surface-membrane proteins [occludin, claudins, and junctional adhesion molecules (JAM)] are the main components of TJ^[26,54]. Adherens junctions are mainly made up of e-cadherin, catenin, and actin filaments^[26,55].

Inflammation has also been described to be a factor in increasing intestinal permeability, not only in inflammatory bowel disease^[43] but also in IBS, where the increase in mast cells and the mediators increased the effects on the intercellular junctions^[44].

Finally, other factors, such as hormonal and neuro-hormonal pathways, nutritional factors, ethanol consumption and several drugs (nonsteroidal anti-inflammatory drugs, methotrexate, tacrolimus, protonic pump inhibitors), could affect the intestinal barrier, a factor that needs to be further evaluated^[26]. The knowledge of affection of the intestinal permeability in IBS patients will help in the development of new therapies in order

to restore the gut barrier, a topic we will touch upon in the next section.

Table 2 sums up the literature findings about therapies restoring intestinal permeability in irritable bowel syndrome.

THERAPIES RESTORING INTESTINAL PERMEABILITY IN IRRITABLE BOWEL SYNDROME

Probiotics

The human intestinal microbiota represents one of the densest, biodiverse, and rapidly evolving bacterial ecosystems. The intestinal microbiome, that is, its collective genome, is an adaptive entity that varies with diet, lifestyle and environment, providing a further metabolic flexibility to the human super organism and functional traits that humans have not evolved on their own^[56]. Therefore, the potential of manipulating the gut microbiota in these disorders is assessed^[57].

The mechanisms through which probiotics alter the intestinal microbial flora could be direct, changing the bacterial macroenvironment of the lumen, or indirect, through the stimulation of the immune system and the improvement of mucosal function, for example, by modulating the invasion and adherence of the epithelial cells of the gut by pathogenic bacteria, thus normalizing gut permeability^[58-60]. The use of probiotics in patients with IBS seems to be effective in achieving improvement in the global IBS symptoms^[61,62], but how it affects the intestinal permeability is less evaluated in humans^[46]. Most studies have shown that altering the intraluminal content affects the barrier functions of the gut, and studies on rodent models of IBS have shown different data^[63].

Despite the growing interest of the scientific community in research in the field of probiotics, the interpretation of the scientific literature on the value of these preparations' results is difficult due to the wide variability in the species, strains and doses employed in the preparations as well as the low methodological quality of the available trials, often due to the poor design and the small sample size.

Several meta-analysis have been published on this topic^[64-66], all concluding that probiotics might be efficacious in IBS, but the actual benefit and the most effective species and strains are uncertain.

In the meta-analysis performed by Ford *et al.*^[61], including forty-three randomized controlled trials, probiotics showed beneficial effects on global IBS, abdominal pain, bloating, and flatulence scores^[61]. Probiotics appeared to be successful in chronic idiopathic constipation (CIC), but there were only two randomized controlled trials, and again, since trials for probiotics are few in number, no specific conclusions could be obtained^[61].

Table 2 Summary of the literature findings about therapies restoring intestinal permeability in irritable bowel syndrome

Drug	Ref.	No. of patients	Study design	Outcome
Probiotics	Ford <i>et al</i> ^[61]	Forty-three RCTs were eligible for inclusion	Metanalysis	Probiotics had beneficial effects on global IBS, abdominal pain, bloating, and flatulence scores. Data for prebiotics and synbiotics in IBS were sparse. Probiotics appeared to have beneficial effects in CIC (mean increase in number of stools per week = 1.49; 95%CI: 1.02-1.96), but there were only two RCTs. Synbiotics also appeared beneficial (RR of failure to respond to therapy = 0.78; 95%CI: 0.67-0.92). Again, trials for prebiotics were few in number, and no definite conclusions could be drawn
	Mazurak <i>et al</i> ^[67]	Fifty-six papers	Metanalysis	The heterogeneity of the studies of probiotics in IBS questions the value of meta-analyses and the use of different bacterial strains and different mixtures of these strains, as well as different dosages, are the main contributors to this heterogeneity
Glutamine	Akobeng <i>et al</i> ^[73]	Two randomized trial	Cochrane analysis	Not significant difference in the permeability and no effect in the clinical remission
Larazotide acetate	Leffler <i>et al</i> ^[78]	342 adults with celiac disease who had been on a gluten free diet (GFD) for 12 mo or longer and maintained their current GFD during the study	Randomized, double-blind, placebo-controlled study assessed larazotide acetate 0.5, 1, or 2 mg 3 times daily	Reduce signs and symptoms in celiac disease patients on a GFD better than a GFD alone

In the last updated meta-analysis performed by Mazurak *et al*^[67], including fifty-six papers (twenty-seven studies using multi-species bacterial preparations and twenty-nine using single-strain probiotics), they analysed the efficacy of probiotics regarding patients included, treatment duration, probiotic dosage, and outcome measures. According to the authors, the heterogeneity of the studies of probiotics in IBS impairs the value of meta-analyses. The use of different bacterial strains and different mixtures of these strains, as well as different dosages, may be the main factors contributing to this heterogeneity^[67]. Currently, there is limited evidence for the efficacy of a small number of single-strain probiotics in IBS (mostly bifidobacteria), and this evidence leads to the performance of trials with inclusion and exclusion criteria closely following the European Medicines Agency (EMA) and the Food and Drug Administration (FDA) guidelines for clinical trials in IBS^[68,69], including the definition of minimal severity for inclusion, global primary endpoints, and adequate secondary end-points (pain, bloating, and a clinically meaningful responder definition). Such trials should include at least 8 wk of therapy, an adequate follow-up period and restriction to one of the different IBS subtypes^[67].

Glutamine

Glutamine is one of the compounds that has been investigated as a treatment of conditions with leaky gut. It has been shown to regulate the protein turnover in enterocytes of pig^[70], reduce intestinal permeability in intestinal cell cultures and maintain transepithelial resistance^[71]. Glutamine has also been shown to

maintain the integrity of the intestinal barrier in critically ill patients by reducing the incidence of infections^[72]. Glutamine treatment in patients with Crohn's disease was recently reviewed in a Cochrane analysis^[73]. In this review, only two randomized controls were included, and neither showed a significant difference in the permeability and neither had any effect on clinical remission.

Glutamine treatment in patients with IBS is less examined. Glutamine synthetase expression is lower in the small bowel and colonic mucosa of patients with IBS-D with increased intestinal permeability^[74]. Therefore, one recent pilot study on IBS-D showed that with a higher glutamine concentration, Claudin-1 expression increases, thus improving the permeability^[75]. However, further studies are needed for using glutamine as a supplement treatment for IBS.

Larazotide acetate

Larazotide acetate (LA) is a tight-junction regulator peptide preventing the opening of intestinal epithelial TJ^[76]. The safety, tolerance and pharmacokinetics of LA were studied in a randomized double-blind placebo-controlled study conducted on celiac disease subjects challenged with gluten^[76,77].

Recently, in a multicentre, randomized, double-blind placebo-controlled study, LA at doses of 0.5, 1, or 2 mg 3 times daily was evaluated to relieve ongoing symptoms in 342 adults with celiac disease who had been on a gluten-free diet (GFD) for 12 mo or longer and maintained their current GFD during the study. A 0.5 mg dose of Larazotide acetate appeared to reduce signs and symptoms in celiac disease patients on a

GFD better than a GFD alone. Although the results were mixed, this study resulted in the successful use of a novel therapeutic agent targeting tight junction regulation in those patients with CeD who are symptomatic despite a GFD^[78].

Therefore, the modulation of the tight junction could represent a paradigm shift in the treatment of immune mediated and inflammatory diseases (Celiac Disease, IBD, IBS, *etc.*).

THERAPEUTIC TARGETS IN CONSTIPATION-PREDOMINANT IRRITABLE BOWEL SYNDROME

Constipation-predominant irritable bowel syndrome (IBS-C) is a frequent disorder and represents one of the main causes of ambulatory visits. Abdominal pain and discomfort characterize IBS-C, making it different from chronic idiopathic constipation^[79].

It is now well known that treatment focusing only on bowel transit does not provide complete relief to patients with IBS-C. A global evaluation of the pathophysiology of IBS-C has led to the use of sensory end points like complete spontaneous bowel movements and the FDA combined end point (abdominal pain and complete spontaneous bowel movements) in clinical trials^[79].

For example, new information on the mechanisms underlying pain sensation in chronic visceral hypersensitivity as well as insights into the mechanism of action of new drugs targeting abdominal pain in IBS have recently been obtained by preclinical experiments in rodent models^[80]. A number of drugs that we will touch upon in the next section are actually in development.

Linacotide

Linacotide (MD-1100 acetate) is a novel orally active 14-amino acid peptide of the guanylin family of cyclic guanosine monophosphate (cGMP)-regulating guanylate cyclase-C (GC-C) agonists. It has been approved by the FDA and by the EMA for the treatment of moderate to severe IBS-C in adults. Its action is focused on the increase of fluid secretion, favouring gastrointestinal transit, and has GC-C-mediated analgesic effects^[81].

It is recommended at a dose of 290 µg orally once a day before meals. Linacotide is converted to an active metabolite (MM-419447) that has the same pharmacodynamics and pharmacokinetics as the parent drug.

In 2007, Andresen *et al.*^[82] investigated the effect of 5 d of linacotide on transit and bowel function in 36 women with IBS-C according to Rome II criteria randomized in a 1:1:1 fashion for placebo, linacotide 100 µg, and linacotide 1000 µg.

Patients with slow colonic transit or slower transit than the mean for healthy controls were studied for 5 d at baseline and 5 d during the treatment. Patients collected all the information regarding gastric, small

bowel, and colonic transit by scintigraphy and bowel function using stool diaries, which included Bristol Stool Form Scale (BSFS) scores for stool consistency, ease of stool passage scores, and completeness of evacuation.

Linacotide did not show any effect on gastric emptying or colonic filling. It did show a significant effect on ascending colon emptying t_{1/2} times ($P = 0.015$) and on overall total colonic transit times at 48 h ($P = 0.02$) at the 1000 µg dose ($P = 0.004$), but not at the 100 µg dose, as well as on increased stool frequency, decreased stool consistency, improved ease of passage, and acceleration of time to first bowel movement ($P < 0.001$)^[82].

In 2010, Johnston *et al.*^[83] investigated the efficacy and safety of 12 wk of linacotide at a daily dose range of 75-600 µg in a phase IIb randomized double-blind parallel-group multicentre placebo-controlled trial conducted on 420 patients with IBS-C (female patients = 92%). Patients had to meet Rome II criteria, with fewer than three spontaneous bowel movements (SBMs) per week, and straining, lumpy/hard stools, or sensation of incomplete evacuation more than 25% of the time for at least 12 wk in the 12 mo preceding study entry. The primary endpoint was a change in the number of complete spontaneous bowel movements (CSBMs). Secondary endpoints were the effect on individual symptoms, quality of life (QOL), the number of patients who were CSBM responders (at least three CSBMs/wk and an increase of one CSBM from baseline for 75% of the study duration), and global relief responders (symptoms being somewhat, considerably, or completely relieved for 100% of the study duration or completely relieved for 50% of the study duration).

For the 75, 150, 300 and 600 µg linacotide doses, the mean change in CSBMs per week was 2.90, 2.49, 3.61 and 2.68, respectively ($P < 0.01$), and the percentage of patients who were CSBM responders was 25%, 19.5%, 32% and 24%, respectively. Patients treated with linacotide showed an adequate relief response (33%-51% vs 22%) and a global relief response (44%-55% vs 29%) compared to placebo. All doses of linacotide significantly improved bowel habits, including frequency of short bowel movements (SBMs) ($P \leq 0.001$) and CSBMs ($P \leq 0.01$), severity of straining ($P \leq 0.001$), stool consistency ($P \leq 0.001$), and abdominal pain scores ($P \leq 0.05$), than placebo. Abdominal discomfort, bloating, and global IBS-C measures were also improved for all doses except for the 75 µg (abdominal discomfort) and 150 µg (bloating) doses. The linacotide effect was observed at the first week and lasted throughout the 12 wk of treatment.

The approval of linacotide for IBS-C was based on two randomized double-blind placebo-controlled phase III trials similar in study design, end points, and patient demographics^[84-86].

Primary end points included both the FDA-recommended combined primary end point and a more rigorous combined primary end point that required

even more CSBM responses for 9 of 12 wk. Secondary end points included patient-reported abdominal pain, discomfort, and bloating; straining severity; and weekly SBM and CSBM frequency and stool consistency. The first phase III trial included 804 adults with IBS-C who were randomized 1:1 to receive linaclotide 290 lg or placebo daily for 26 wk, with change-from-baseline end points measured at 12 and 26 wk^[84]. Attrition rates were 18.5% at 12 wk and 25.6% at 26 wk. At 26 wk, the majority of patients withdrew from the study due to adverse events in the linaclotide arm (10.2%) and perceived lack of efficacy in the placebo arm (8.2%). Over 12 wk, the FDA combined primary end point was achieved by 33.7% of patients receiving linaclotide compared with 13.9% of patients receiving placebo ($P < 0.0001$).

Linaclotide was also superior to placebo in the more rigorous investigator-defined combined primary end point that was reached by 12.7% of linaclotide-treated patients vs 3.0% of placebo-treated patients ($P < 0.0001$). At 26 wk, 32.4% of patients receiving linaclotide and 13.2% of patients receiving placebo ($P < 0.0001$) reached the FDA combined primary end point.

Improvements in all secondary end points occurred in the linaclotide group at weeks 12 and 26. The second phase III trial of linaclotide was composed by a 12-wk treatment phase followed by a 4-wk randomized withdrawal phase^[86]. A total of 803 adults with IBS-C were randomized to receive linaclotide 290 lg or placebo once/d for 12 wk. Approximately 78% of patients completed the entire 16-wk study, and most of the patients who suspended the study did so due to adverse events in the linaclotide arm (7.9%).

In the 12-wk active treatment phase, linaclotide demonstrated statistically significant improvements in all primary and secondary efficacy end points compared with placebo. Approximately one-third (33.6%) of patients receiving linaclotide fulfilled both components of the FDA end point compared with 21% of patients receiving placebo ($P < 0.0001$). Statistically significant improvements were observed also in abdominal pain, discomfort, and bloating in linaclotide-treated patients, with a mean reduction of about 2 points from baseline (on an 11-point scale) compared with reductions of 1.1 with placebo ($P < 0.0001$ for each measure).

In the linaclotide arm, an improvement in severity of straining, constipation, and stool consistency was observed compared with the placebo arm (all $P < 0.0001$).

Linaclotide caused diarrhoea, abdominal pain, flatulence, headache, viral gastroenteritis, and abdominal distension as adverse events. Diarrhoea, the most common, occurred in less than 20% of patients, probably due to increased fluid secretion and accelerated colonic transit^[84,86].

In Johnston *et al.*^[83] phase IIb dose-ranging trial, diarrhoea of mild to moderate severity was the primary

dose-dependent adverse effect observed. It occurred in 11.4%, 12.2%, 16.5% and 18.0% of patients in the 75, 150, 300 and 600 µg linaclotide dose groups, respectively, compared with 1.2% in the placebo group. Dehydration or electrolyte disturbances were not found, although one instance of faecal impaction occurred^[83]. In the studies by Rao *et al.*^[86] and Chey *et al.*^[84], 4.5%-5.7% of the linaclotide-treated patients and 0.2%-0.3% of the placebo group discontinued the study due to diarrhoea.

In a phase III clinical trial in IBS-C, patients experienced adverse events more in the linaclotide 290-lg group (65.4%) than in the placebo (56.6%, $P < 0.05$) group^[87]. In another IBS-C phase III trial, adverse effects in the linaclotide group were reported at a similar rate to placebo (56.2% vs 53.0%, $P = 0.39$)^[88]. Adverse events were reported by 60.5% of patients receiving linaclotide 145 lg, 55.7% of patients receiving linaclotide 290 lg, and 52.1% of patients receiving placebo^[89].

In phase III clinical trials in patients with IBSC, diarrhoea was the most frequently reported adverse event, occurring in 19.5%-19.7% of patients in the linaclotide groups compared with 2.5%-3.5% of patients receiving placebo ($P < 0.0001$).

In randomized trials, linaclotide at 145 µg/d was best tolerated with improvement in CSBM/Wand symptoms in patients with CIC. Patients with IBS-C best responded to the 290-µg daily dose^[84,85]. Linaclotide appeared to be very well tolerated.

Linaclotide is approved for the treatment of IBS-C in both male and female adults at a dosage of 290 lg once/d and for the treatment of CIC at a dosage of 145 lg once/d. The medication should be taken 30 min prior to breakfast. Renal or hepatic impairment is unlikely to affect the metabolism or clearance of linaclotide or its metabolite due to its low systemic exposure.

In conclusion, linaclotide can represent a targeted approach that addresses the complexity of symptoms associated with the syndrome. Linaclotide has been reported to safely improve IBS-C abdominal pain severity, bowel movement quality, and bowel movement frequency as well as key symptoms of abdominal fullness, bloating, and discomfort, with associated improvements in QOL. Based on the United States FDA and the EMA, linaclotide fulfils the recommended endpoints with a number needed to treat (NNT) ranging from 4.39 to 7.9. It is effective and can be associated with diarrhoea as the most common adverse effect leading to suspension of the medication in approximately 5% of patients. According to recent clinical evidence, linaclotide should be considered for patients with IBS-C due to its effect on abdominal pain and bowel symptom improvement.

Plecanatide

Plecanatide is a 16-amino acid GC-C agonist currently used in phase III clinical trials for CIC and phase II

trials for IBS-C^[90]. Plecanatide mimics the endogenous agonist of the GC-C receptor in the intestinal tract. Like that of uroguanylin, plecanatide's actions are pH-dependent, with the most favorable efficacy in the acidic environment of the duodenum. Similar to linaclotide, plecanatide luminally activates the GC-C receptor on gastrointestinal mucosal epithelial cells, leading to intracellular secretory and extracellular anti-nociceptive effects *via* a cGMP-mediated second messenger pathway^[91]. A phase III randomized double-blind trial in 951 patients with CIC treated with 0.3, 1 or 3 mg plecanatide or placebo once/d for 12 wk was conducted^[92]. The primary end points were weekly (more than three CSBMs/wk and an increase of more than one CSBM/wk from baseline) or an overall study response (weekly response for 9 of 12 wk, including 3 of the last 4 wk to ensure durability of response). The percentage of overall responders was significantly higher in the plecanatide 3 mg group compared with placebo (19% vs 10.7%, $P = 0.009$). Weekly responder rates were also significantly higher in plecanatide 3 mg than placebo for weeks 1-12. Patients treated with 3 mg showed an improvement in stool frequency, consistency, straining, and quality of life compared with placebo. Data for other plecanatide doses were not shown.

Plecanatide potentially has low risk of adverse cardiovascular effects, as its systemic absorption is very low. According to the phase I study for evaluation of the safety and tolerability of plecanatide in humans^[93], no measurable systemic absorption was observed at any doses of oral plecanatide. Plecanatide was safe and well tolerated up to the highest dose. Diarrhoea was the most prevalent side effect, but its frequency did not statistically significantly differ between placebo and plecanatide, and appeared not to be dose-related in the plecanatide-treated subjects. Other gastrointestinal events were nausea, abdominal discomfort and pain, and vomiting. In a Phase II dose escalation trial involving a total of 84 chronic constipation patients recruited with modified Rome III criteria, 14 d of plecanatide therapy improved stool frequency, stool consistency, straining and overall relief of chronic constipation symptoms. To confirm the safety and efficacy of plecanatide, two Phase III trials (NCT01982240 and SP304203-00) have been planned. In the United States and Canada, the Phase III trial NCT01982240 was initiated in November 2013 with adult chronic constipation patients and was expected to be completed in February 2015^[94].

Prucalopride

Prucalopride is authorized in several countries (not in the United States) for women with CIC unresponsive to laxatives^[95]. As a very highly selective 5-HT₄ agonist, prucalopride has no measurable affinity for other receptors. In safety evaluation tests, prucalopride

showed no h ERG (human ether-à-go-go-related gene) channel inhibitory activity. It is not arrhythmogenic, and it promotes colonic motility^[96].

At dosages of 2 mg and 4 mg per day, this drug produced a low incidence of QT interval prolongation. Even up to 20 mg per day (10-fold higher than the recommended dosage), prucalopride displayed no clinically relevant effects on cardiovascular parameters in healthy volunteers. Prucalopride improved stool frequency and consistency, and it dose-dependently enhanced colonic transit in healthy controls or chronic constipation patients with no negative impact on gastric emptying or small bowel transit^[97]. The patients' quality of life was significantly improved by prucalopride treatment.

In three pivotal trials, prucalopride showed a good efficacy in increasing CSBMs per week and in improving perceived disease severity and quality of life in patients with CC. A study conducted on 620 patients with CC treated with 2 or 4 mg of prucalopride for 12 wk showed that it increased one or more CSBMs per week compared to the control group^[98-100]. In another trial conducted on 713 patients with CC, 2 or 4 mg of prucalopride increased the frequency to three or more CSBMs per week and improved evacuation completeness, perceived disease severity, and quality of life^[101]. In another study conducted on patients 65 years or older with CIC, prucalopride at a dose of 1 mg for 4 wk did not cause any changes in an electrocardiogram or corrected QT (QTc) interval, showing its safety for the treatment of CIC in the elderly^[97,102]. A study conducted on Asian subjects with CIC reported similar efficacy and safety as that observed in Western populations^[103]. In a pooled analysis of the study with Asian subjects and the three pivotal trials, increased stool frequency of approximately three or more CSBMs per week was observed in Asian (34% vs 11%, $P < 0.001$) and non-Asian (24.6% vs 10.6%, $P < 0.001$) women. Prucalopride was shown to be safe and well tolerated^[104], improving CIC abdominal symptoms such as abdominal discomfort, bloating, straining, and painful bowel movements^[105]. Another study conducted on a small number of patients showed the efficacy of prucalopride not only in the treatment of slow transit constipation but also of obstructed defecation and IBS-C^[106].

In a recent analysis, Camilleri *et al.*^[106] evaluated the efficacy of prucalopride using the data from six phase 3 and 4 multicentre double-blind randomized placebo-controlled parallel-group trials performed across three continents.

Over the 12-wk treatment period, prucalopride-treated patients consistently achieved a mean of 3 CSBMs/wk compared to placebo with the treatment response observed in the individual trials^[98-100,102]. On the other hand, the SPD555-401 trial was the only trial that failed to demonstrate a statistically significant effect of

prucalopride on this primary endpoint after both 12 and 24 wk of treatment, without any plausible explanation of this lack of efficacy^[107,108]. In the current study, no differences were found between men and women, although over time, a difference in the response rate has been reported. This could be related to differences in demographics (other than gender) and disease characteristics at baseline or to intrinsic differences in responsiveness to prucalopride between men and women. Furthermore, prucalopride was significantly more effective than placebo, as demonstrated by many secondary endpoints, including improvements in PAC-SYM (Patient Assessment of Constipation Symptoms) and PAC-QOL (Patient Assessment of Constipation Quality of Life) scores and rescue medication use. An exploratory efficacy analysis showed that prucalopride treatment was effective even in patients with very severe CIC and those with no SBMs at baseline.

In the current integrated analyses, the NNT with prucalopride used to achieve the primary efficacy endpoint in one patient was 8.8 (95%CI: 7.1-11.6). In a meta-analysis of data from three trials of linaclotide in patients with CIC, the NNT for the primary endpoint of these trials (3 SCBMs/wk and an increase of 1 SCBM/wk, for 75 % of weeks) was 7 (95%CI: 5-8)^[109]. Prucalopride has a favorable safety and tolerability profile^[110]. Notably, no cardiovascular safety signals were observed. Indeed, the mean QT interval corrected according to Bazett's formula (QTcB) and the mean QT interval corrected according to Fridericia's formula (QTcF) were both/470 ms. A potential limitation of this integrated analysis is moderate heterogeneity ($I^2 = 56\%$) due to a deviation of the results of one of the six trials compared to the others.

Prucalopride was well absorbed from the gastrointestinal tract, with an absolute oral bioavailability of more than 90%. Its main elimination route was *via* theurine (60%-70% excreted unchanged in the urine). Because prucalopride has a low level of metabolism by liver, its pharmacokinetics is unlikely to be altered by hepatic impairment, and no CYP3A4 drug interactions are anticipated. In Europe, 2 mg of prucalopride has been approved for the treatment of chronic constipation in women who have no adequate response to laxatives^[111].

Headache (in 25%-30%), nausea (12%-25%), abdominal pain (16%-23%), and diarrhoea (12%-19%) were observed as adverse events.

Recently, a randomized trial compared prucalopride with Macrogol/PEG 3350 plus electrolytes in patients with CIC. Prucalopride showed a non-inferiority for the primary outcome, even though PEG showed a superiority in improving gastrointestinal transit, stool frequency, and number of spontaneous bowel movements^[112]. Although no studies have yet evaluated the efficacy of prucalopride in IBS-C, it is expected that it may also be efficacious for the disease symptoms.

However, the worsening of abdominal pain may limit its use in clinical practice.

YKP10811

YKP10811 is a novel substituted benzamide derivative, small molecule with high binding affinity to the 5-HT₄ receptor^[113]. In cellular functional assays conducted with the 5-HT₄ receptor, YKP10811 showed weak agonist activity that was dose dependent and reproducible. These results indicated that YKP10811 acts as a partial agonist of the 5-HT₄ receptor. YKP10811 did not show any significant off-target binding to any other receptors, enzymes, or serotonin-receptor subtypes at 1 mmol/L, except for binding to the 5-HT_{2A} receptor and the 5-HT_{2B} receptor. Thus, YKP10811 has 120-fold and 6-fold lower affinity, respectively, for 5-HT_{2A} and 5-HT_{2B} receptors than for 5-HT₄. In cellular functional assays, YKP10811 showed antagonist activity at the 5-HT_{2B} receptor with a median inhibitory concentration. In rats, YKP10811 accelerated colonic transit by 37% at a dose as low as 1 mg/kg. In dogs, 0.3 mg/kg YKP10811 accelerated colonic transit by 45.5% at 2 h after dosing. The accelerated colonic transit in dogs was associated with significantly increased colon contractions and defecation. YKP10811 significantly reduced visceral hypersensitivity in multiple pain models in rats. In a phase I double-blind randomized 9-d placebo-controlled multiple-ascending dose study in healthy volunteers at doses of 5, 15, 30 and 45 mg once daily, YKP10811 was well tolerated with minimal side effects. In a single-center randomized parallel-group double-blind placebo-controlled study^[114] in patients with functional constipation, YKP10811 enhanced gastrointestinal and colonic transit and improved bowel function during an 8-d treatment trial. The effect of YKP10811 on colonic transit was mirrored by improvements in softer stool consistency and faster time to first bowel movement, suggesting that YKP10811 has encouraging effects on these clinical end points. In addition to pharmacodynamic effects in patients with functional constipation, improvements in bowel functions are validated and measurable end points recommended for the treatment of functional constipation^[115]. These findings suggest that YKP10811 may be a potential new medication for the treatment of functional constipation. YKP10811 had a robust effect on accelerating, by 30% to 40%, colonic emptying when compared with placebo. Ascending colon emptying has been reported to have the greatest contribution to overall colonic transit^[116] because the ascending and transverse colon constitute the "reservoir" or storage regions of the human colon^[117]. Among the other 5-HT₄-receptor agonists previously studied with the same method, 4 mg prucalopride and 30 and 50 mg velusetrag^[118] also accelerated AC emptying. Emptying of the proximal colon correlates linearly with faecal weight^[119], which largely reflects

stool water content, and as expected based on prior studies, the overall colonic transit was correlated linearly with stool consistency, with less significant association with the number of bowel movements per day. The results also showed the dual action (agonist/antagonist) of YKP10811 seen in *in vitro* studies. YKP10811 facilitated the electrical field stimulation-induced neurogenic twitch of guinea pig ileum at lower concentrations. This type of dual action (agonist/antagonist) of YKP10811 under the same assay conditions was also shown in the peristaltic reflex test, with an EC₅₀ of 0.5 mmol/L and an IC₅₀ of 21 mmol/L. There is a significant gap in concentration ranges (> 40-fold difference) for stimulatory vs inhibitory effects of YKP10811 *in vitro* (unpublished data; SK Life Science, Inc). Two participants, 1 receiving placebo and 1 receiving 20 mg YKP10811, had prolonged QTc (> 470 ms). Both participants discontinued the study on the advice of the investigators, even though the QTc prolongation was minimal (functions in patients with functional constipation). Thus, YKP10811 is likely to be of benefit to patients with functional constipation without rectal evacuation disorders. The safety and efficacy of this novel agent should be studied in larger multicentre clinical trials. With further studies, the current data suggest that YKP10811 would expand the therapeutic options beyond the recently approved secretagogue medications for the treatment of functional constipation, lubiprostone and linaclotide^[120,121].

YKP10811 was reported to be safe and tolerable in healthy volunteers. Except for a Phase II clinical trial in C-IBS patients (NCT02082457)^[122], there were only two registered Phase II trials that evaluated the efficacy and safety of YKP10811 in comparison with placebo in subjects with CIC (NCT015 23184, NCT01989234)^[123,124]. Collectively, 420 eligible subjects were enrolled to be treated with different doses of YKP10811 or placebo once daily for 8 d and 12 wk in two trials. The results have not been completed for reporting yet. This drug is pending to pass Phases II and III of clinical trials, expected in 2016.

Renzapride

Renzapride (a novel benzamide substitute) is a full agonist for the 5HT₄ receptor and an antagonist to 5HT_{2b} and 5HT₃ receptors. It can accelerate the gastrointestinal tract transit and motility stimulating the 5HT₄ and 5HT_{2b} receptors^[125], and it appears to be a promising therapeutic agent for constipation, which is predominant in IBS patients. It is safe and has only a few adverse effects^[126,127]. Several clinical trials have been performed to evaluate its potential efficacy in IBS patients, confirming that renzapride does not cause cardiac arrhythmias in clinical doses, unlike cisapride^[126,127]. It is excreted renally and is not metabolized by cytochrome P450 enzymes.

Thus, no drug interactions *via* affecting cytochrome P450 enzymes have been reported^[125,127]. Renzapride stimulates colonic transit and reduces transit time and pain in IBS patients due to its prokinetic property, providing a benefit in those patients with constipation^[128]. In addition, a dose-dependent efficacy of this drug has been demonstrated^[126]. In a phase II study of 46 women with IBS-C, renzapride at a dose of 4 mg q.d. favoured colonic transit and increased ascending colon emptying compared to placebo^[129]. A large multicentre European trial confirmed the effects of 4 mg renzapride q.d. in the improvement of frequency of bowel movements and stool consistency in IBS-C^[130]. Much pharmacodynamic data support renzapride's prokinetic effects. As for the prior European study, statistically significant differences in the frequency of bowel movements and stool consistency in favour of renzapride 4 mg q.d. were relatively small. In addition, renzapride did not improve the feelings of completeness of bowel movements or the amount of straining. Several systematic reviews have shown the efficacy of 5HT receptor modulators in IBS patients. In 2009, Ford *et al.*^[131] conducted a meta-analysis by reviewing placebo-controlled clinical trials up to 2008 on the efficacy of known 5-HT₃ antagonists and 5-HT₄ agonists in IBS. They observed that renzapride and cisapride were not more effective than placebo in IBS patients.

Other investigators also evaluated the efficacy of combined 5HT₃ antagonists/5HT₄ agonists (cisapride and renzapride) in IBS patients^[132] and observed that 1 and 2 mg of renzapride was ineffective in relieving IBS symptoms, supporting the results obtained by Ford. However, these authors showed that 4 mg of renzapride was significantly more effective than placebo.

Recently, a meta-analysis^[133] from randomized placebo-controlled clinical trials, including 2528 C-IBS, non-C-IBS, and non-D-IBS patients according to the Rome criteria, was performed. The study confirmed that renzapride had no significant effects in relieving symptoms in IBS patients compared to placebo. To reach a convincing conclusion on the effectiveness of renzapride, a clinical trial compared with placebo was performed. Renzapride at a dose of 4 mg was compared to placebo for 5 wk or less and more than 5 wk. Although the differences were not statistically significant, the results were clinically important and significant for both treatment durations. Therefore, these results could be considered for renzapride 4 mg, while more trials are necessary to determine the effectiveness of this novel drug more precisely. As regards adverse effects, no statistically significant differences between renzapride and placebo were found, except for diarrhoea occurrence, which was higher in patients treated with renzapride. In addition, renzapride caused more withdrawals due to adverse effects and/

or low efficacy in patients. One of the limitations of this meta-analysis was the evaluation of trials with different patient inclusion criteria (age, sex, lifestyle and compliance). In addition, the trials evaluated had different durations of treatment and endpoints. The treatment durations ranged from 2 wk^[134] to 12 wk^[135]. To avoid heterogeneity, patients were divided into two groups according to treatment duration and time of reporting the results (5 wk or less and more than 5 wk), although there were few data in each group. The safety data from these phase III studies indicated that renzapride was generally well tolerated, even though ischaemic colitis was reported in the long-term study in 3 patients. However, evaluating the total of patients treated with renzapride during the study, the overall rate of ischaemic colitis appeared comparable with that reported for other 5-HT₃ receptor antagonists^[136]. In conclusion, renzapride is not only superior to placebo in relieving IBS symptoms (abdominal pain and discomfort), but it also causes increased diarrhoea occurrence compared with placebo and appears to be associated with many drop-outs. Therefore, this drug might be a cost burden to patients, without any advantages in efficacy. Indeed, during the trial, no improvements in frequency of bowel movements, straining, or completeness of evacuation were observed in patients treated with renzapride. Taken together, these data suggest that renzapride is unlikely to provide clinically meaningful improvement in IBS symptoms.

Velusetrag

Velusetrag is an orally administered available 5-HT₄ agonist developed by Theravance. The binding affinity of this drug for the 5-HT₄ receptor is more than 500-fold that of other 5-HT receptor subtypes^[137]. The major metabolite detected in plasma after oral velusetrag is THRX-830449, which is a full agonist and is approximately equipotent to velusetrag. Metabolism occurs through the CYP3A4 system. In healthy subjects^[138], at steady state, the THRX-830449 to velusetrag AUC ratio is approximately 0.5 following once-daily dosing of velusetrag (15 mg).

Increased smooth muscle contractility of the antrum, fundus, duodenum and jejunum was observed in velusetrag-treated dogs^[139]. Velusetrag increased guinea-pig colonic transit and produced dose-dependent relaxation of the rat esophagus^[140]. Relief of constipation using velusetrag was also confirmed in chronic constipation patients^[141].

Velusetrag was approximately 6- or 86-fold more potent than cisapride or mosapride after intravenous dosing and 9- or 18-fold more potent than tegaserod or cisapride, respectively, after intraduodenal administration^[141].

Its low risk for cardiovascular events has been confirmed in an *in vitro* investigation demonstrating no effect on hERG channel conductance^[140]. In a preclinical

study that compared *in vivo* activity of velusetrag vs tegaserod in guinea pig, the subcutaneous administration of velusetrag increased colonic transit more than tegaserod did. Velusetrag was more potent than tegaserod when orally administered in a dog GI smooth muscle contractility model^[142]. Velusetrag exhibited an acceptable oral bioavailability in rats and dogs^[140], while the systemic effect of the drug was increased by an increase in the administered dose in healthy volunteers^[143-145]. Both single (up to 70 mg) and multiple (up to 50 mg, for 2 wk) dosing of velusetrag in healthy subjects showed a dose-dependent effect on GI motility^[145].

There have been two Phase II clinical trials to evaluate the clinical efficacy of this drug. In one of these two studies^[138], 60 healthy volunteers were randomly assigned, in double-blind fashion, to placebo or 5, 15, 30 or 50 mg velusetrag, with transit measurements after single and 6-d dosing.

The GI transit was evaluated in a randomized double-blind placebo-controlled study conducted on 60 healthy subjects randomly assigned to receive velusetrag at a dose of 5, 15, 30 or 50 mg or placebo either as a single dose or for 6 d^[137]. Velusetrag at single dose (30 and 50 mg) favoured colonic transit, evaluated by colonic filling at 6 h and geometric center at 24 h, while this effect was not observed in patients treated with placebo. Similarly, velusetrag at multiple doses (15-50 mg doses) favoured gastric emptying compared with placebo ($P = 0.002$). In this study by Manini *et al.*^[115], an improvement of stool frequency and consistency by velusetrag in a subset of 11 patients with chronic constipation was also reported. Pharmacokinetic evaluations demonstrated a similar profile in healthy and CIC subjects^[143]. Velusetrag was well tolerated in the Phase I study when administered in single and repeated doses in healthy subjects. In the Phase I clinical trial, the most commonly reported adverse event was diarrhoea, which is expected because of velusetrag's mechanism of action^[141,143]. In a Phase II randomized, double-blind, placebo-controlled trial, the efficacy and safety of velusetrag were compared with placebo in 401 subjects with CC. SBM frequency, CSBM and other associated symptoms with CIC were significantly improved compared with placebo in patients who received velusetrag for 4 wk. The most effective dose was 15 mg once daily. Most of the adverse events, such as diarrhoea, headache, nausea and vomiting, were mild to moderate. These adverse events were common in the first days of treatment with the dose of 50 mg once daily. The number of withdrawals due to adverse events was 18 vs 1 for the velusetrag- and placebo-treated subjects, respectively. The number of withdrawals were 4, 3 and 11 in the 15-, 30- and 50-mg treated groups, respectively. However, the medicine was well tolerated with no cardiac complications^[146]. Another Phase II study of

velusetrag in 401 patients with chronic constipation treated for 4 wk showed that there were significant treatment effects on the average daily number of bowel movements compared with placebo^[147].

The most common adverse effects of velusetrag were those frequently associated with 5-HT₄ agonists, including diarrhoea, headache and nausea. These dose-dependent adverse effects were mild to moderate and usually occurred within the initial days of dosing. Clinically relevant doses of velusetrag in animals or humans did not generate severe side effects on blood pressure, heart rate or electrocardiogram. In isolated porcine or canine coronary arteries, velusetrag showed no contractile activity^[148]. In the randomized, double-blind, placebo-controlled study in 60 healthy subjects, there was no significant treatment effect on heart rate recorded by ECG after treatment for the prior 5–6 d. In this study, there were also no serious adverse events, and predictable GI effects such as diarrhoea and altered bowel movement were the main adverse events recorded^[140]. These results suggest that velusetrag appears to be well tolerated. Further careful clinical studies will be required to further evaluate the safety and tolerability of this drug.

Naronapride

Naronapride (ATI-7505)^[149] is a benzamide 5-HT₄ receptor agonist that activates 5-HT₄ receptors but has almost no actions on the other 5-HT subtypes. The design of ATI-7505 was based on the prototypical agent, cisapride. However, unlike cisapride, which is a mixture of (3R, 4S) and (3S, 4R) isomers of substituted piperidine-based scaffolds, ATI-7505 is the pure (3S, 4R) isomer. ATI-7505, with its (R)-quinuclidinyl moiety, is metabolized by ubiquitous carboxyl esterases to a single metabolite, ATI-7500.

This potent and selective 5-HT₄ receptor agonist showed different pharmacodynamic and pharmacokinetic properties from previous nonselective 5-HT₄ agonists. Hydrolytic esterase metabolism, unlike oxidative CYP450 metabolism, is a large-capacity metabolic system that can easily handle therapeutic amounts of xenobiotics. This large-capacity system implies that other drugs metabolized by esterases are not expected to induce drug-drug interactions of ATI-7505 with other drugs. There is also no interaction with drugs metabolized by a different enzymatic system, such as CYP450^[150]. Naronapride is not metabolized by CYP450 enzymes, and thus, less drug-drug interaction occurs.

A thorough QT study showed that naronapride had no obvious effect on cardiac repolarization at either therapeutic or supratherapeutic doses. The structure of naronapride is similar to that of cisapride, but it is more selective than cisapride and thus interacts minimally with hERG channels and 5-HT₃ receptors^[150]. ATI-7500, the main metabolite of naronapride, is 100-fold less

active than the parent drug. Unlike prucalopride and velusetrag, neither naronapride nor ATI-7500 can pass the blood-brain barrier, therefore reducing the incidence of side effects. This new benzamide exhibited GI prokinetic effects, stimulated colonic transit and reduced stool consistency in healthy male and female subjects^[151]. One Phase II randomized double-blind placebo-controlled dose definition study evaluated several doses of orally administered naronapride (20, 40, 80 and 120 mg twice a day) in 210 patients with CC. This study evaluated the clinical effects of 9 days' treatment with three doses of ATI-7505 at 3, 10 and 20 mg on GI and colonic transit using a validated scintigraphic method. There were borderline effects on gastric emptying at half-time. However, ATI-7505 stimulated colonic transit at 24 h and ascending colonic emptying. There was looser stool consistency as measured by the Bristol stool form scale with the 10- and 20-mg t.i.d. doses. This finding suggests that ATI-7505 appears to have prokinetic properties in both stomach and colon in healthy subjects and, particularly, in the colon. Further clinical trials of larger numbers of patients with functional gastrointestinal disorders, such as patients with CIC, are required to evaluate clinical efficacy.

The inhibition of the delayed rectifier K⁺ current in response to ATI-7505 in patch-clamped HEK293 (human embryo kidney) cells transfected with the human IKr channel is very weak, suggesting that there would be an adequate safety window between activity in the GI tract and potential cardiac toxicity. In addition, the primary metabolite ATI-7500 is 100-fold less active than the parent drug at the 5-HT₄ receptor and, as with ATI-7505, has no detectable HERG channel inhibitory activity at concentrations up to 100 µmol/L. Preliminary data on intensive cardiac safety monitoring suggest that ATI-7505 is safe as regards the cardiac profile^[149].

The most common drug-related adverse events were headache, diarrhoea, nausea and vomiting. Headache and abdominal pain were reported more frequently by the maximum dose of naronapride^[152].

Chenodeoxycholic acid

Chenodeoxycholic acid (CDCA) is a bile acid that can induce colonic electrolyte secretion by acting on the membrane-bound bile acid GPBA receptor (TGR5) on enterocytes, subsequently leading to the stimulation of cAMP generation and electrogenic chloride secretion. Supplementation with specific bile acid analogues or by using drugs that inhibit ileal bile acid reabsorption may benefit constipation patients.

Oral chenodeoxycholic acid at doses of 750–1000 mg/d can increase bowel movements, decrease stool consistency, and reduce the time to defecation in IBS-C^[153].

They were previously used for the dissolution of

gallstones, and they are known to favour diarrhoea at high doses in healthy controls and constipation patients^[154]. The effects of CDCA on gastrointestinal and colonic function have been evaluated in healthy volunteers and patients with irritable bowel syndrome with constipation. In a randomized controlled trial, 500 mg and 1000 mg CDCA given to 60 healthy volunteers for 4 d led to dose-dependent acceleration of colonic transit. In addition, significant increases in stool frequency, decreases in stool consistency, and improvements in ease of stool passage were reported with CDCA^[155]. In a double-blind placebo-controlled study, Rao *et al.*^[153] demonstrated that sodium chenodeoxycholate^[156] stimulated colonic transit and improved bowel function in 36 women with irritable bowel syndrome with constipation. Increased stool frequency, greater ease of stool passage and looser stool consistency were observed in patients treated with sodium chenodeoxycholate 500 mg or 1000 mg for 4 d as compared with controls. Unfortunately, over 40% of sodium chenodeoxycholate-treated patients had light abdominal cramping or pain. Whether these side effects could be mitigated at a lower dose remains to be determined.

Elobixibat

Elobixibat is an orally administered available potent inhibitor of ileal bile acid transporter with minimal systemic exposure^[157]. Elobixibat (A3309) reduces bile acid enterohepatic recirculation and upregulates bile acid synthesis as measured by serum C4 levels. It also depletes liver cholesterol and reduces serum LDL^[158], thus increasing the delivery of bile acids to the proximal colon, which in turn increased fluid secretion, colonic motility and stool frequency, and it improved stool consistency and relieved constipation-related symptoms in chronic idiopathic constipation patients^[159,160].

In a phase I trial, elobixibat stimulated colonic transit in a dose-dependent way. In a randomized phase II trial, elobixibat at doses of 15 and 20 mg/d showed an improvement of stool consistency and of stool passage, increased the number of SBMs and reduced straining in female patients with CIC^[161]. In a dose-finding randomized trial, elobixibat increased C4, reduced LDL cholesterol, increased colonic transit from 3 to 1.9 d and increased the number of SBM and CSBM/wk in patients with CIC compared to placebo. The treatment with elobixibat also resulted in an improvement of bloating severity, but no effects on abdominal pain or discomfort were reported^[157]. The well-tolerated doses were 5-10 mg, with a discontinuation rate during the phase IIb trial of 13%, rising to 23% for the 15 mg group. Fifty-four percent of patients developed adverse events, such as abdominal cramps, relieved by defecation, and diarrhoea. However, the side effects were not different from those of the placebo group^[162]. In a large randomized trial

conducted on patients with CIC, the 10- and 15-mg doses increased SBMs and reduced the time to SBM (12 h with the 10-mg dose, 7 h with the 15-mg dose and 24 h with the placebo). In patients treated with elobixibat, an increased spontaneous laxation within 24 h was observed compared with placebo (75 % on 15 mg/d and 45 % on placebo).

The side effects of elobixibat are mainly gastrointestinal tract-related. Although higher dosages of elobixibat caused abdominal pain and diarrhoea more frequently, no severe adverse effects occurred in the Phase I and Phase II clinical trials. The Phase III clinical trials are ongoing to determine the best tolerated dose and to examine the effects of long-term administration.

Complete spontaneous bowel movements per week increased in a dose-dependent way. An improvement of stool consistency and bloating was observed at the 10- and 15-mg doses. Side effects such as abdominal pain and diarrhoea were also dose-dependent, notably for the 15-mg dose^[157].

Elobixibat is a promising anti-constipation drug. However, there are no studies in cancer or in OIC (opioid induced constipation) patients. Due to its prokinetic activity, elobixibat is not recommended in patients with mechanical bowel obstruction.

According to the results of Phase II trials in chronic idiopathic constipation patients, elobixibat was safe and generally well tolerated, even at a dose up to 20 mg per day.

As illustrated by elobixibat, the advantages of IBAT inhibitors may be especially attractive, which may boost research on other IBAT inhibitors, such as SC-435, S-8921 and S-0960^[163-165].

Lubiprostone

Lubiprostone, a first-in-class drug for the treatment of chronic idiopathic constipation and irritable bowel syndrome in adult women with constipation is believed to be a highly selective locally acting activator of CIC-2 channels^[166]. Lubiprostone can tautomerize between the inactive form I and the active form II^[167]. Lubiprostone acts mainly by activating specific type-2 chloride channels (CIC-2) on the apical membrane of the enterocytes^[166] that are involved in ion and fluid transport across the epithelial membrane. Once channels are opened, chloride enters the enterocyte in the basal membrane through the action of Na-K-2Cl active cotransporters. This mechanism results in an electrochemical gradient favouring chloride secretion. It leads to an overall concentration-dependent raise in intestinal fluid secretion without any impairment on serum sodium and potassium levels. These mechanisms explain how lubiprostone increases the number of colonic spontaneous bowel movements per week. However, lubiprostone efficacy on the abdominal pain score is only partially known

and needs further investigation. Lubiprostone also activates a prostaglandin receptor (EP4), which in turn activates cystic fibrosis conductance regulators (CFTR)^[168]. The activation of EP4 receptors favours colonic smooth muscle and gastric longitudinal muscle *via* vagal nerve endings^[169]. Lubiprostone changes mucin, which improves the gut microbiome, creating an anti-inflammatory environment^[170]. Unlike linaclotide, lubiprostone does not increase pain thresholds^[171].

Lubiprostone pharmacokinetics is not impaired by renal failure. However, great adverse events with a standard dose of lubiprostone can result in cases of mild-to-moderate hepatic impairment (Child-Pugh class A and B), which increases the lubiprostone metabolite M3. Thus, in cases of liver impairment, a reduction of lubiprostone starting doses is required. Lubiprostone metabolism does not involve cytochromes. The catabolism is mediated by carbonyl reductase in the stomach and jejunum^[172]. Lubiprostone is unlikely to have major drug interactions.

In healthy subjects, a reduction in gastric emptying, an increase in gastric fasting volume, a reduction in maximum tolerated gastric volumes and a stimulation of small bowel and colon transit was observed with lubiprostone at a dose of 24 µg twice daily^[173]. However, the effects on gastric motility may mask the nausea side effect.

Two 12-wk double-blind randomized multicentre placebo-controlled phase III clinical trials^[174] and one 36-wk open-label extension study^[175] contributed to the FDA's approval of lubiprostone for the treatment of IBSC in women. A total of 1171 patients were randomized 2:1 to receive either 8 µg lubiprostone or placebo twice/d. The primary end point of each study was the evaluation of response rate, measured by patient-reported improvements from baseline in IBS-C symptoms. As secondary end points, monthly responder rates, changes from baseline in SBM frequency, stool consistency, straining, distention, abdominal pain/discomfort (each measured on a 5-point Likert scale) and change in health-related quality of life were evaluated.

The discontinuation rate in both studies was 24%, firstly due to withdrawal of consent and secondly to adverse events and perceived lack of efficacy. Lubiprostone was superior in the primary end point compared to placebo (17.9% vs 10.1%, $P = 0.001$). Patients treated with lubiprostone reported more improvements in all secondary end points than placebo. Lubiprostone was associated with a more significant improvement in abdominal pain/discomfort than placebo from baseline to month 2 (0.43 vs 0.35, $P = 0.039$) and month 3 (0.45 vs 0.36, $P = 0.028$).

Lubiprostone significantly changed the mean SBM frequency from baseline to month 1 compared with placebo, even though the numerical data were not included.

Two 4-wk phase III randomized double-blind placebo-controlled multicentre clinical trials were conducted on a total of 479 patients to evaluate the short-term efficacy and safety of lubiprostone in patients with CIC with identical study designs and primary end points. After a 2-wk baseline period, eligible subjects received 24 lg lubiprostone or placebo twice/d.

The number of patient-reported SBMs, defined as any BM occurring 24 h or longer after the use of an alternative drug used to relieve constipation (rescue medication), during the first week of treatment was the primary end point of each study^[176,177]. Lubiprostone was associated with a statistically higher frequency of SBMs during the first week of treatment than placebo.

Improvements in other secondary end points, such as stool consistency, straining, and constipation severity were also observed in patients treated with lubiprostone compared with placebo in all 4 wk in both studies. However, significant improvement in abdominal distention and discomfort compared with placebo was not observed in either study.

As regards side effects, a similar percentage of patients reporting at least one treatment-related adverse event for IBS-C was observed in the lubiprostone (50%) and placebo (51%) groups. The most common side effects were gastrointestinal (19% with lubiprostone vs 14% with placebo). Serious adverse events were similar between the two groups (1%). Nausea was the most frequent treatment-related event^[11,12], although it may be reduced by administering lubiprostone with meals.

A unique adverse effect occurred with the initial dose. In rare cases, acute transit dyspnea and ischemic colitis were observed^[176,178].

Caution in the use of lubiprostone should be used for infants of breastfeeding mothers due to the risk of diarrhoea^[179]. Limited data are available on the lubiprostone effects in paediatric patients, and further, larger studies are required. In an open-label 4-wk clinical trial conducted on paediatric patients with CIC (mean age 10.2 years)^[179], lubiprostone was efficacious and well tolerated at daily doses of 12-48 lg. The recommended dose of lubiprostone for the treatment of CIC in both adult men and women is 24 lg twice/d, while for the treatment of IBSC in adult women, it is 8 lg twice/d. Due to its minimal systemic absorption and its metabolism through a cytochrome P450-independent pathway, lubiprostone-drug interactions are unlikely, even though *in vitro* studies have suggested that methadone may decrease the efficacy of lubiprostone by reducing chloride channel type 2 activation^[179].

In patients with moderate or severe hepatic impairment (Child-Pugh class B or C), a dose reduction might be suggested, while in patients with renal failure, no dosage adjustment is recommended.

A small percentage (8%-13%) of patients over 65 years were included in clinical trials with lubiprostone. The safety profile was similar in elderly and younger patients, even though, due to the limited number of patients over 65 years, no differences in clinical response were observed^[179], and further studies are needed.

Like linaclotide, lubiprostone is contraindicated in mechanical bowel obstruction. To confirm the indications in the treatment of IBS-C in adults, more and larger trials are required. Due to the chronic nature of IBS-C and CIC, post-marketing studies are necessary to confirm the long-term efficacy and safety of lubiprostone. All randomized clinical trials were of limited duration (12-26 wk). However, in open-label extension studies, a safety over 52 wk was demonstrated.

Despite the efficacy, the side effects (e.g., nausea, abdominal pain) and the high cost may limit the use of lubiprostone.

Tenapanor

Tenapanor, also known as AZD1722 or RDX5791, is a first-in-class orally available inhibitor of NHE3 that is minimally absorbed in the gastrointestinal tract—this constitutes a significant therapeutic benefit, as it may act on the drug target^[180,181]. Consequently, tenapanor increases intestinal Na⁺ contents, which leads to an increase in intestinal fluid volume and accelerates the whole GI transit, as shown in rats. Moreover, tenapanor inhibits the absorption of phosphorus, which is independent of typical phosphorus transporters in the intestines, namely, sodium-dependent phosphate transport protein 2B (NaPi2b) and Na (+)-dependent phosphate transporter (PiT1). Tenapanor is stable at room temperature and is formulated into tablets ranging from 1 to 50 mg. Absorption, distribution, metabolism and excretion (ADME) studies have revealed that tenapanor is minimally absorbed and metabolized. For example, experiments in rats showed 92.2% ± 1.6% recovery of tenapanor in faeces upon oral administration^[182]. In humans, the inactive metabolites of tenapanor were found in plasma, but they were only approximately 9% of the parent compound. In pharmacokinetic studies, tenapanor was observed at relatively low concentrations in plasma (average < 3 ng/mL) of rats and dogs, but only sporadically (29/76 and 0/92, respectively).

Oral administration of tenapanor (at doses of 0.1 and 3 mg/kg) produced a dose-dependent increase in faecal water content and stool consistency in rats. The effect of tenapanor at a dose of 50 mg/kg twice daily on stool form was assessed in cynomolgus monkeys. The animals were observed for 4 d before treatment. Soft or watery stools were observed in monkeys on tenapanor treatment, and stool consistency was normalized on day 6 of the experiment. Under physiological conditions, tenapanor given orally at doses of 3, 10, 30 and 50 mg/kg did not affect visceral

sensitivity or the changes in intestinal volumes induced by colorectal distension in comparison with the control and tegaserod-treated (5 mg/kg administered per os) groups^[183]. However, tenapanor (30 and 50 mg/kg) had a dose-dependent antinociceptive effect in the acute restraint stress-induced intestinal hypersensitivity to colorectal distension. The antinociceptive potential of tenapanor was comparable with that of the tegaserod-treated group.

The safety and tolerance of tenapanor were assessed in a randomized, double-blind, placebo-controlled study^[184,185]. Eighty healthy volunteers were included in the study (male and female). Tenapanor was given orally at the doses ranging from 10 to 900 mg (as a single administration) and for 7 consecutive days at doses ranging from 3 to 100 mg to assess the safety of tenapanor administration. Tenapanor was also beneficial for the percent of days with a spontaneous bowel movement. Finally, no serious side effects were observed, and there were very few adverse events^[186]. Phase II a In a II a double-blind randomized placebo-controlled study on 181 patients with IBS-C^[187], tenapanor was given orally at doses of 10, 30 and 100 mg once daily for 4 consecutive weeks with 2 wk follow-up. The primary end point (change in complete spontaneous bowel movements from baseline to week 4) was not met in this study, and the incidence of diarrhoea was comparable with that of the placebo group. However, an improvement in bloating and abdominal pain was noted in IBS-C patients. In Phase II b In a II b randomized double-blind placebo-controlled multicentre study, 371 IBS-C patients were divided into four groups: placebo and tenapanor (5, 20 and 50 mg) treated twice daily for 12 wk with 4 wk follow-up. The primary efficacy end point was met in 60.7% of the tenapanor-treated group (at a dose of 50 mg) vs 33.7% of the placebo-treated group. The overall responder was met in 50.0% of the tenapanor-treated group (50 mg) vs 23.6% for placebo (after 12 wk). After 12 wk, adequate relief in IBS-C symptoms was observed in 63.1% of the tenapanor-treated group (50 mg twice daily) vs 39.3% in placebo. The effectiveness of tenapanor therapy was maintained during entire time of the clinical study. The treatment satisfaction patient scale questionnaire showed that tenapanor-treated (50 mg) IBS-C patients were quite or very satisfied (65% vs 38% for the placebo-treated group). The drug was well tolerated in all groups, and no serious adverse effects were noted. The most common adverse effect was diarrhoea in the tenapanor-treated group (50 mg twice daily), reported in 11.2% of IBS-C patients vs 0% in placebo. Safety and tolerability In the preclinical studies in rats, tenapanor did not influence gastric emptying^[187].

Piromelatine

Melatonin is engaged in the regulation of gastrointestinal motility and sensation. When administered

orally in pharmacological doses, it has shown beneficial effects on abdominal pain in IBS patients without any effects on sleep disturbances^[188]. It was also shown that oral melatonin significantly stimulated colonic transit time in healthy subjects, and it may be a promising option for future research on the agents modulating bowel motility^[189]. Melatonin synthesized in the enteroendocrine cells of the intestinal mucosa reaches the liver *via* the portal vein^[190]. Melatonin is a potent accelerator of duodenal mucosal bicarbonate secretion, which neutralizes the acid content of the stomach in the duodenum, and it seems to be engaged in the acid-induced stimulation of the secretion^[191]. Melatonin protects the gastrointestinal mucosa due to an antioxidant action, a decrease in secretion of hydrochloric acid, stimulation of the immune system, promotion of epithelial regeneration, and increased microcirculation^[192,193].

Recently, it was shown that patients with IBS had significantly lower 6-SMLT (6-sulphatoxymelatonin)/creatinine level compared with healthy controls^[194]. The lack of statistical difference in 6-SMLT/creatinine levels between the constipation and diarrhoea groups is difficult to explain. In some patients, the symptoms could be recurrent, or there could be some subjects with mixed (IBS-M) or unsubtyped (IBS-U) IBS. This study's results agree with those obtained by Bultman^[195] and Lu *et al.*^[196] who performed the study on female patients with IBS and found decreased salivary melatonin and urine 6-SMLT level compared to non-IBS volunteers. Low melatonin levels were observed in women with eating disorders. Low melatonin concentrations have been associated with increased depressive symptoms, such as sadness, bodily discomfort, inner tension, difficulties in attention concentration and pain.

Serotonin, an endogenous amine and the precursor of melatonin, synthesized and released from enteroendocrine cells of the gastrointestinal mucosa is thought to play an important role in the pathogenesis of IBS^[197]. Antagonists of the serotonin 5-HT₃ receptor are beneficial in patients with IBS-D, whereas the partial agonist of the serotonin 5-HT₄ receptor (tegaserod) alleviates symptoms of IBS with constipation, especially in females. The role of melatonin as the regulator of circadian and seasonal rhythmicity has been established^[198,199]. Patients with functional disorders of the gastrointestinal tract also had sleep disorders, and some of them suffered from increased neural excitability and anxiety^[199,200]. There were speculations concerning a possible role of melatonin in functional dyspepsia (FD), particularly ulcer-like dyspepsia. In two types of FD, one with epigastric pain and another with postprandial distress syndrome, the melatonin level is varied, and different dyspeptic symptoms may be related to differences in melatonin secretion. Sleep disturbances are common in patients with IBS and are among the most important extraintestinal symptoms,

markedly affecting quality of life and psychosocial well-being^[201]. In a double-blind placebo-controlled study, Camilleri *et al.*^[202] showed that melatonin improves abdominal pain in IBS patients with sleep disturbances. Currently, conventional treatment for irritable bowel syndrome is quite unsatisfactory. Despite multiple therapeutic interventions, no long-term effect has been achieved. On the other hand, up to 80% of patients with IBS treated with hypnotherapy showed an improvement of their symptoms^[203]. These observations emphasize the possible role of melatonin in the pathogenesis of irritable bowel syndrome and in its therapy^[204,205].

Daikenchuto

Daikenchuto (TU-100), a traditional Japanese drug (Kampo medicine), is indicated in the treatment of adhesive bowel obstruction^[206,207]. TU-100 is a mixture of extract powders from dried Japanese pepper, processed ginger, ginseng radix, and maltose powder. In many trials, the TU-100 prokinetic effect has been demonstrated to be useful in treating GI hypomotility^[208]. Studies conducted on postoperative patients after gastrointestinal surgery showed that TU-100 prevented postoperative ileus, but little is known about the TU-100 effects in patients who did not undergo major gastrointestinal surgery^[209]. Iturrino *et al.*^[210] performed a randomized controlled trial to evaluate the effects of oral TU-100, 2.5 g t.d.s. or 5 g t.d.s. compared to placebo t.d.s. on gastrointestinal and colonic transit, rectal compliance and sensation thresholds, anal sphincter pressures and bowel function in women with functional constipation. In this study, there were no significant effects on gastrointestinal and colonic transit, rectal compliance, anal sphincter pressures, recto-anal pressure difference, or rectal sensation thresholds. The highest dose was associated with lower rectal sensation thresholds for first sensation and gas. There were no treatment effects on psychosensory symptoms, stool frequency, stool consistency or quality of life^[211].

On the other hand, Manabe *et al.*^[211] reported that TU-100 provided a clinically significant promotility effect in small bowel and ascending colon transit in healthy subjects. TU-100 is quite safe and well tolerated and is a potential treatment for IBS-C and functional constipation^[210].

Recently, however, Acosta *et al.*^[212] did not report any significant effects of TU-100 on rectal sensation ratings, sensation thresholds, rectal fasting or postprandial tone, rectal compliance, bowel function, abdominal pain or bloating scores, or IBS quality of life. Further randomized controlled trials in patients with IBS-C or functional constipation using both clinical and validated biomarkers are required.

DA-6886

DA-6886, a gastrointestinal prokinetic benzamide

derivative, is a novel 5-HT₄ receptor agonist. Experimental studies showed that it may represent a highly potent and selective 5-HT₄ receptor agonist to stimulate colonic transit in mice, having a favorable safety profile in patients with IBS-C and chronic constipation^[213]. Currently, a phase I dose block-randomized double-blind placebo-controlled single/multiple dosing dose escalation clinical trial with an open-labelled food effect is being conducted to evaluate the safety and pharmacokinetics of single-dose DA-6866 in healthy male subjects^[214].

Table 3 sums up the literature findings about irritable bowel syndrome-C therapies.

THERAPEUTIC TARGET IN DIARRHOEA-PREDOMINANT IRRITABLE BOWEL SYNDROME

To date, the treatment options for IBS-D are limited and frequently unsuccessful. However, the incidence of IBS-D is currently increasing, thus causing a heavy economic burden both for patients and health care systems worldwide. As for IBS-C, a complete understanding of IBS-C pathophysiology has favoured the use of sensory end points such as complete spontaneous bowel movements and the FDA combined end point (abdominal pain and complete spontaneous bowel movements) in clinical trials^[79].

Furthermore, also in the setting of IBS-D, pre-clinical studies in rodents have recently improved the understanding of the mechanisms underlying the alterations in gastrointestinal motility, sensitivity and secretion. A number of drugs that we will touch upon in the next section are actually in development.

Ramosetron

Ramosetron is a potent and selective 5-HT₃ receptor antagonist. 5-HT₃ receptors can be widely found in the central and peripheral nervous system^[215]. Intraluminal stimuli favour the release of 5-HT from enterochromaffin cells located in the mucosa^[215]. When secreted, 5-HT can activate 5-HT₃ receptors located on intrinsic primary afferent neurons with submucosal terminals. Thus, the peristaltic reflex and intestinal secretion can occur^[215,216]. 5-HT also activates 5-HT₃ and 5-HT₄ receptors located on primary afferent neurons of both splanchnic and vagal fibres, which are involved in sensory and motor responses^[217].

In experimental studies, corticotrophin-releasing hormone (CRH) exogenously administered or released from the central nervous system by stress peripherally activates the release of 5-HT, which in turn promotes defecation through the 5-HT₃ receptor. Ramosetron decreased defecation by CRH in a dose-dependent way^[218-219].

The first 5-HT₃ receptor antagonist to be introduced

was Alosetron, which has been demonstrated to be effective in the treatment of female patients with IBS-D^[220]. However, due to serious gastrointestinal events (ischemic colitis and severe constipation), it is still only available in the United States and is indicated for women with severe D-IBS refractory to conventional therapy.

Ramosetron was first tested by Lee *et al.*^[221] in a multicentre randomized open-label trial on 343 men with IBS-D. Patients were randomized to a 4-wk treatment of ramosetron 5 mg once daily or mebeverine 135 mg three times daily. An improvement in abdominal pain/discomfort and bowel habits in the ramosetron and mebeverine groups was observed during the treatment period. A significant reduction in abdominal pain/discomfort and urgency, stool form score, and stool frequency severity scores in both treatment arms was reported compared with the baseline.

Adverse events were observed in 7% and 4% of patients treated with ramosetron and mebeverine, respectively, even though no statistically significant differences were reported. Additionally, all the side effects were mild or moderate^[221].

Successively, Fukudo *et al.*^[222] performed a randomized double-blind placebo-controlled trial to determine whether ramosetron reduces diarrhoea in 296 male outpatients with IBS-D. Patients were treated with 5 mg of oral ramosetron ($n = 147$) or placebo ($n = 149$) once daily for 12 wk after a 1-wk baseline period. The primary end point was increased stool consistency in the first month. Secondary end points were the relief of overall IBS symptoms and the improvement of IBS-related quality of life. In the first month, patients on ramosetron treatment (74, 50.3%) showed an improvement of stool consistency compared to placebo (29, 19.6%) ($P < 0.001$). In patients treated with ramosetron, the monthly relief of overall IBS symptoms and IBS-related quality of life was demonstrated compared with placebo. Safety was evaluated in all 296 patients, with side effects occurring in 46.9% and 51.7% of ramosetron and placebo patients, respectively. All constipation and hard stools experienced in the ramosetron group that were related to the pharmacologic actions of ramosetron were classified as mild and resolved early without using rescue drugs^[222].

In another randomized double-blind placebo-controlled trial performed by Fukudo *et al.*^[223] on 576 female outpatients with IBS-D, patients were given either 2.5 µg ramosetron or placebo once daily for 12 wk. Patients treated with ramosetron reported global improvement, increased stool consistency, a significant decrease in abdominal pain and discomfort and significant improvement in QOL compared with placebo. Of the patients tested with ramosetron, 11.0% complained of constipation^[223]. Successively, in a phase III open-label uncontrolled long-term safety

Table 3 Summary of the literature findings about irritable bowel syndrome-C therapies

Drug	Ref.	No. of patients	Study design	Outcome
Linaclotide	Andresen <i>et al</i> ^[82]	36 women with IBS-C	Phase II a randomized, double-blind, placebo-controlled trial. Patients were randomized in a 1:1:1 fashion to placebo, linaclotide 100 µg, and linaclotide 1000 µg and was evaluated the effect of 5 d	No treatment effects were seen for gastric emptying or colonic filling with linaclotide. Significant treatment effects were found for ascending colon emptying t _{1/2} times ($P = 0.015$) and overall total colonic transit times at 48 h ($P = 0.02$), for the 1000 µg dose ($P = 0.004$) but not the 100 µg dose, as well as overall treatment effects on increased stool frequency, decreased stool consistency, improved ease of passage, and acceleration of time to first bowel movement ($P < 0.001$) ^[82]
	Johnston <i>et al</i> ^[83]	420 patients with IBS-C	Phase II b randomized, double-blind, parallel-group, multicenter, placebo-controlled trial evaluate 12 wk of linaclotide at a daily dose range of 75-600 µg	Compared with placebo, all doses of linaclotide significantly improved bowel habits, including frequency of SBMs and CSBMs, severity of straining, stool consistency, as well as abdominal pain scores. Abdominal discomfort, bloating, and global IBS-C measures were also improved, for all doses except for the 75 µg (abdominal discomfort) and 150 µg dose (bloating). Effects were present for the first week, and sustained throughout the 12 wk of treatment
	Chey <i>et al</i> ^[84]	804 adults with IBS-C	Phase III trials randomized, double-blind, placebo-controlled to receive linaclotide 290 lg or placebo daily for 26 wk, with change-from-baseline end points measured at 12 and 26 wk	Over 12 wk, the FDA combined primary end point was achieved by 33.7% of patients receiving linaclotide compared with 13.9% of patients receiving placebo ($P < 0.0001$)
	Videlock <i>et al</i> ^[109]	7 trials of linaclotide in patients with IBS-C or CC	A meta-analysis from MEDLINE, EMBASE, and the Cochrane central register of controlled trials were searched for randomized, placebo-controlled trials	The NNT for the primary endpoint of these trials (3 SCBMs/wk and an increase of C1 SCBM/wk, for 75% of weeks) was 7 (95%CI: 5-8)
	Rao <i>et al</i> ^[86]	803 adults with IBS-C	Phase III trials randomized, double-blind, placebo-controlled to receive linaclotide 290 lg or placebo once/d for 12 wk	Linaclotide demonstrated statistically significant improvements in all primary and secondary efficacy end points compared with placebo Severity of straining, constipation, and stool consistency also improved in the linaclotide group compared with the placebo group
Plecanatide	Miner <i>et al</i> ^[92]	951 patients with CIC	Phase III, randomized, double-blind trial, received plecanatide 0.3, 1 or 3 mg, or placebo once/d for 12 wk	The proportion of overall responders was significantly greater with plecanatide 3 mg compared with placebo (19% vs 10.7%, $P = 0.009$); weekly responder rates were also significantly greater for plecanatide 3 mg than placebo for weeks 1-12. Improvements in stool frequency, consistency, straining, and quality of life were also noted with the 3-mg dose vs placebo. Data for other plecanatide doses were not reported
Prucalopride	Quigley <i>et al</i> ^[88]	620 patients with CC	A double-blind, placebo-controlled study. Patients receiving 2 or 4 mg of prucalopride for 12 wk	Increased one or more CSBMs per week compared to patients in the control group
	Camilleri <i>et al</i> ^[98]	713 patients with CC	A double-blind, placebo-controlled study. Patients receiving 2 or 4 mg of prucalopride for 12 wk	Increased frequency of three or more CSBMs per week, and improved evacuation completeness, perceived disease severity, and quality of life
	Müller-Lissner <i>et al</i> ^[97]	Elderly patients aged 65 years and older with CC	A double-blind, placebo-controlled study. Patients receiving 2 or 4 mg of prucalopride for 12 wk	No changes in electrocardiogram or corrected QT (QTc) interval were reported, indicating its safety for the treatment of CC in the elderly
	Ke <i>et al</i> ^[102]	4 Randomized, Placebo-controlled Studies	A Pooled Analysis	Safe and well-tolerated It was also effective in improving the abdominal symptoms of CC such as abdominal discomfort, bloating, straining, and painful bowel movements
YKP10811	Shin <i>et al</i> ^[114]	55 patients	A single-center, randomized, parallel-group, double-blinded, placebo-controlled study were assigned randomly to groups given YKP10811 10 mg ($n = 15$), 20 mg ($n = 16$), 30 mg ($n = 15$), or placebo ($n = 11$) daily for 8 d	Enhanced gastrointestinal and colonic transit and improved bowel function during an 8-d treatment trial. In general, the 10-mg and 20-mg doses were the most effective in accelerating colonic transit. No serious adverse events were observed
Renzapride	Camilleri <i>et al</i> ^[101]	46 women with IBS-C	In a phase II study	Renzapride 4 mg q.d. accelerated colonic transit and increased ascending colon emptying vs placebo
	George <i>et al</i> ^[130]	510 patients	Multicentre, randomized, placebo-controlled, double-blind study men and women were randomized to placebo or renzapride (1, 2 or 4 mg/d) for 12 wk	4 mg renzapride q.d. in terms of improving frequency of bowel movements and stool consistency

	Ford <i>et al</i> ^[131]	29 RCTs were eligible for inclusion	Meta-analysis of placebo-controlled clinical	Renzapride and cisapride were not more effective than placebo in IBS patients
	Mozaffari <i>et al</i> ^[133]	2528 C-IBS and non C-, non D-IBS patients	Meta-analysis from randomized placebo-controlled clinical trials	Renzapride has no significant advantage over placebo in relieving symptoms in IBS patients
Velusetrag	Manini <i>et al</i> ^[115]	60 healthy volunteers	Phase II clinical trials, pt were randomly assigned, in double-blind fashion, to placebo, 5, 15, 30 or 50 mg velusetrag, with transit measurements after single and 6-d dosing	Single doses of velusetrag (30 and 50 mg), but not placebo, accelerated colonic transit, as measured by colonic filling at 6 h and geometric center at 24 h
	Goldberg <i>et al</i> ^[140]	401 subjects with CC	In a Phase II randomized, double-blind, placebo-controlled trial	Short bowel movement (SBM) frequency, complete SBM and other associated symptoms with CC were significantly improved in comparison with placebo in patients who received velusetrag for 4 wk
Naronapride	Dennis <i>et al</i> ^[149]	210 patients with CC.	Phase II, randomized, double-blind, placebo-controlled, dose definition study (orally 20, 40, 80 and 120 mg twice a day) to evaluate the clinical effects of 9 days'	There were borderline effects on gastric emptying at half-time; however, ATI-7505 accelerated colonic transit at 24 h and ascending colonic emptying
Chenodeoxycolic acid	Odunsi-Shiyanbade <i>et al</i> ^[155]	60 healthy volunteers	Randomized controlled trial, CDCA 500 mg and 1000 mg given for 4 d	Significant increases in stool frequency, decreases in stool consistency, and improvements in ease of stool passage were reported with CDCA
	Rao <i>et al</i> ^[153]	36 female patients	Double-blind placebo-controlled study	Accelerated colonic transit and improved bowel function
Elobixibat	Simrén <i>et al</i> ^[162]	30 patients	Dose-finding randomized trial five dose-levels (range: 0.1-10 mg/d) or to placebo	Increased C4, reduced LDL cholesterol and increased colonic transit from 3 to 1.9 d, and increased the number of SBM and CSBM/W in patients with CIC
	Chey <i>et al</i> ^[157]	190 patients	Were randomized to 5, 10, or 15 mg A3309 or placebo once daily. 8-wk, multicenter, randomized, double-blind, placebo-controlled, parallel group, phase II b study,	A3309 increased stool frequency and improved constipation-related symptoms in CIC; effects were maintained over 8 wk of treatment
Lubiprostone	Drossman <i>et al</i> ^[173]	1171 patients in total	Two double-blind, randomized, multicenter, placebo-controlled phase III clinical trials, a 12-wk randomized 2:1 to receive either lubiprostone 8 lg or matching placebo twice/d with food; a 36-wk open-label extension study	Lubiprostone was superior to placebo in the primary end point of overall responders, greater improvements in all secondary outcome measures compared with placebo
	Johanson <i>et al</i> ^[175]	479 patients in total	Two 4-wk phase III, randomized, double-blind, placebo-controlled, multicenter clinical trials	Patients treated with lubiprostone had a statistical higher frequency of SBMs during the first week of treatment compared with placebo
	Barish <i>et al</i> ^[182]	127 pediatric patients with CIC (mean age 10.2 yr)	An open-label 4-wk clinical trial	Demonstrated that lubiprostone was efficacious and well tolerated at total daily doses of 12-48 lg
AZD1722	Rosenbaum <i>et al</i> ^[185]	181 patients with IBS-C	Phase II a In a II a double-blind, randomized placebo-controlled study, Tenapanor was given orally at the doses of 10, 30 and 100 mg once daily for 4 consecutive weeks with 2 wk follow-up	The primary end point [change in complete spontaneous bowel movements (CSBM) from baseline to week 4] was not met in this study and the incidence of diarrhea was comparable with placebo group. However, improvement in bloating and abdominal pain was noted in IBS-C patients
	Rosenbaum <i>et al</i> ^[186]	371 IBS-C patients	Phase II b In a II b randomized, double-blind, placebo-controlled, multicenter study	The overall responder was met in 50.0% of tenapanor-treated group (50 mg) <i>vs</i> 23.6% for placebo (after 12 wk). After 12 wk, adequate relief in IBS-C symptoms was observed in 63.1% of tenapanor-treated group (50 mg twice daily) <i>vs</i> 39.3% in placebo
	Rosenbaum <i>et al</i> ^[187]	356 patients	A double-blind, placebocontrolled, randomized phase 2b trial 12-wk dose-ranging study evaluating tenapanor 5 mg, 20 mg or 50 mg b.i.d. <i>vs</i> placebo (1/2)	Tenapanor 50 mg b.i.d. significantly improved CSBM responder rate (primary endpoint) compared with placebo in patients with IBS-C. Tenapanor 50 mg b.i.d. also improved key secondary endpoints compared with placebo, including overall responder rate, abdominal pain responder rate and stool frequency. In addition, improvements were observed in several exploratory endpoints addressing a range of symptoms in patients with IBS-C. Tenapanor was generally well tolerated and had minimal systemic availability. Tenapanor shows promise as a future treatment option for patients with IBS-C

Neu p11 (piromalatine)	Camilleri <i>et al</i> ^[202]	40 IBS patients	In double blind placebo controlled study were randomly assigned to receive either melatonin 3 mg (<i>n</i> = 20) or matching placebo (<i>n</i> = 20) at bedtime for two weeks	Melatonin 3 mg at bedtime for two weeks significantly attenuated abdominal pain and reduced rectal pain sensitivity without improvements in sleep disturbance or psychological distress. The findings suggest that the beneficial effects of melatonin On abdominal pain in IBS patients with sleep disturbances are independent of its action on sleep disturbances or psychological profiles
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IBS : Irritable bowel syndrome.

trial, the long-term safety and efficacy of 2.5 and 5 µg of ramosetron treatment in women with IBS-D was reported. However, the authors of the study concluded that “clinicians should be aware that one-fifth of women with IBS-D receiving ramosetron may suffer from constipation during treatment”^[224].

Finally, a recent systematic review and meta-analysis performed to explore of the safety and efficacy of ramosetron both in male and female patients with IBS-D concluded that ramosetron is efficacious in both male and female patients with diarrhoea-predominant IBS, even if large-scale studies are needed to assess its effects on different ethnicities^[225].

LX-1031

LX-1031 is a tryptophan hydroxylase (TPH) inhibitor that reduces peripheral serotonin production. It is indicated for conditions characterized by excess 5-HT expression such as IBS-D and, possibly, carcinoid diarrhoea. The blocking of excess 5-HT effects is well established^[226]. Previously, pharmacological attempts aimed at inhibiting 5-HT synthesis, such as parachlorophenylalanine, but the central adverse effects due to the inhibition of brain 5-HT synthesis with consequent affective disorders blocked their use^[227].

LX-1031 does not cross the blood-brain barrier and, thus, does not pose risk of depression^[228].

In a phase II multicentre randomized double-blind placebo-controlled proof-of-concept study^[229] performed on 155 patients, the subjects were assigned randomly in a double-blind fashion to 1 of 2 doses of LX1031 (250 mg 4 times/d or 1000 mg 4 times/d) or placebo taken daily during the 28-d treatment period^[229]. Patients treated with LX1031 at the dose of 1000 mg significantly improved the primary efficacy end point, namely, the relief of IBS pain and discomfort, compared with placebo at week 1. No significant improvements were observed at weeks 2, 3 or 4. Adverse effects reported were generally mild, self-limited, and evenly distributed across the placebo and both LX1031 treatment arms^[229]. The relationship between clinical improvement and reduction in serotonin synthesis shown in this study supports LX1031's proposed mechanism of action in IBS and thus supports serotonin synthesis inhibition in the GI tract as a novel therapeutic strategy for the treatment of IBS-D^[229].

ASP-7147

Activation of the Bombesin-2 receptor may be involved in the regulation of gastrointestinal motility and intestinal secretion. ASP7147 is a novel small molecule Bombesin-2 receptor antagonist that reduces motility and intestinal secretions. Indeed, ASP 7147-mediated inhibition of this receptor may improve symptoms in patients with IBS-D. In the RCT performed by Lembo *et al*^[230] on 64 patients during a 4-wk study, ASP7147 showed promise as a safe and effective new therapy for both men and women with IBS-D, demonstrating improvement in multiple symptoms of IBS-D. The persistence of treatment effect suggests the possibility of retained efficacy with less-frequent dosing in follow-on trials^[230].

JNJ-27018966

JNJ-27018966 is a dual µ-opioid agonist and δ-opioid receptor antagonist that has been shown to have benefits in patients with IBS-D^[231]. A randomized controlled double-blind study compared JNJ-27018966, at doses of 25, 100 and 200 mg twice daily, to placebo in 807 patients with IBS-D. Diarrhoea and pain were significantly improved in patients treated with JNJ-27018966 at the doses of 25 and 200 mg twice-daily compared to placebo (12, 13.8 and 5.7%, respectively, *P* < 0.05 for both comparisons to placebo)^[232].

ROSE-010

ROSE-010 is a glucagon-like peptide 1 analogue that decreases gastric emptying and motility^[1]. Hellström *et al*^[233] conducted a randomized crossover placebo-controlled trial on 160 patients with IBS and associated abdominal pain treated with ROSE-010 100 µg once daily, 300 µg once daily or placebo. ROSE-010 was associated with a twofold greater response to abdominal pain compared to placebo (*P* < 0.05 for all comparisons) and significantly higher patient-reported satisfaction (*P* < 0.05). The most frequent treatment-related side effect was nausea, which was experienced by 19, 37 and 0% of ROSE-010 100 µg, ROSE-010 300 µg and placebo, respectively^[233-234].

AST-120

AST-120, also known as kremezin, is an orally administered intestinal sorbent that has been reported to slow

chronic kidney disease (CKD) progression and to delay the initiation of dialysis by reducing the levels of renal toxins or their precursors in the gastrointestinal (GI) tract^[235,236].

It has been shown that AST 120 exerts its properties in IBS by acting on intestinal permeability, reduction of visceral sensitivity and alteration of gut motility^[236]. In a randomized double-blind controlled study conducted on 115 non-constipation-related IBS patients, AST-120 at a dose of 2 g three times daily significantly improved the percentage of patients with at least a 50% decrease in the number of days with abdominal pain compared to placebo (26.8% vs 10.2%, respectively). Additionally, AST-120 significantly improved bloating and stool consistency compared to placebo. The safety profile of AST-120 was similar to that of placebo^[237].

Ibodutant

Antagonists of NK2 receptors have been suggested to modulate gastrointestinal chemical-induced impaired motility and stress-induced impaired bowel habits in humans, as recent phase 2 clinical trials have reported^[238]. In a randomized double-blind controlled trial conducted on 559 IBS-D patients, ibodutant, a neurokinin-2 receptor antagonist, significantly improved abdominal pain, overall symptoms and quality of life compared to placebo. Ibodutant at doses of 1, 3 or 10 mg once daily showed superiority over placebo, with the 10 mg once daily dose being the most effective and women showing a better response than men^[239,240]. Considering the limited number of effective available therapeutic options for IBS-D, ibodutant may become an important and safe treatment option, depending on whether ongoing phase 3 studies will confirm the efficacy observed in phase 2 studies^[238].

Asimadoline

Asimadoline, a kappa-opioid receptor agonist, acts peripherally, inducing analgesic and antidiarrheal effects^[241]. Action in the central nervous system is not required for asimadoline efficacy in the treatment of IBS. Asimadoline reduces sensation in response to colonic distension at subnoxious pressures in healthy subjects and in IBS patients without impairment of colonic compliance. Asimadoline decreased the appetite and enhanced the postprandial gastric volume (in healthy women). However, there were no significant effects on gastrointestinal transit, colonic compliance, fasting or postprandial colonic tone. In a clinical trial conducted on 40 patients with functional dyspepsia (according to Rome II criteria), asimadoline did not significantly impair appetite or symptoms over 8 wk. However, asimadoline at a dose of 0.5 mg significantly reduced the appetite in patients, with higher postprandial fullness scores and daily postprandial fullness severity (over 8 wk). Patients treated with asimadoline at a

dose of 1.0 mg showed borderline significant effects. In a clinical trial conducted on patients with IBS, the average pain 2 h post-on-demand treatment with asimadoline was not significantly decreased. At the post hoc analyses, asimadoline was demonstrated to be effective in mixed IBS^[242]. Successively, in a randomized controlled double-blind trial conducted on 596 patients with IBS-D, asimadoline at doses of 0.15, 0.5 and 1 mg twice daily was compared to placebo. Patients treated with asimadoline at a dose of 0.5 mg twice daily had a twofold significant improvement in the total number of months with adequate relief of IBS pain, pain scores, urgency and frequency^[243,244].

Colesevelam

Bile acids have several physiologic functions and are actively reabsorbed (up to 95%) in the terminal ileum^[245,246]. Disruption of the enterohepatic circulation of bile acids due to ileal disease (inflammatory bowel diseases) or idiopathic bile acid malabsorption is responsible for chronic diarrhoea^[246]. Faecal concentrations of bile acids in IBS-D or functional diarrhoea are unknown. While earlier studies suggested up-regulation of the ileal active transporter^[247] as a result of chronic loss of bile acids (which may reduce the bile acids reaching the colon), other data suggest increased delivery to the colon may occur if the ileal reabsorptive capacity for bile acids is exceeded^[246].

Odunsi-Shiyanbade *et al.*^[155] showed in 12 IBS-D patients that colesevelam modestly affected overall colonic transit (in patients treated with colesevelam, the emptying of the ascending colon was approximately 4 h longer compared to placebo). Furthermore, colesevelam favoured stool passage and somewhat firmer stool consistency. No effects on mucosal permeability or safety were found^[246]. Successively, Camilleri *et al.*^[248] performed a 10-d single-center unblinded single-dose trial on the effects of colesevelam in 12 IBS-D patients. They demonstrated that colesevelam accelerates the delivery of BAs to stool, while improving stool consistency. It also stimulates hepatic BA synthesis, avoiding steatorrhea in patients with IBS-D. The overall effects are due to luminal BA sequestration by colesevelam^[248]. All the abovementioned studies suggest that there is an opportunity to diagnose and specifically treat the cause of symptoms in IBS-D.

Solifenacin

Solifenacin is a muscarinic type 3 receptor antagonist recommended in the treatment of overactive bladder (OAB) in adults^[249]. Since 1967, M3 receptor antagonists such as mepenzolate bromide have been used in Japan as modulating agents of gastrointestinal motility. However, no clinical trials had been designed to evaluate the efficacy for IBS defined under the modern Rome criteria. Given the high rate of comorbidity between IBS

and OAB^[250], and considering that solifenacin acts on bowel dysfunction similarly to darifenacin, a selective M3 receptor antagonist with equivalent potencies, solifenacin was evaluated on symptomatic relief in 20 IBS-D patients in an open-label trial^[249]. After a 2-wk observation period, solifenacin was administered for 6 wk. Later, solifenacin was suspended, and ramosetron, a serotonin 3 receptor antagonist, was given for 4 wk. Solifenacin was not inferior to ramosetron in the treatment of IBS with diarrhoea^[249].

The results of this study suggested the potential therapeutic application of solifenacin in the treatment of IBS-D. However, the possible placebo effect could not be excluded. Therefore, further placebo-controlled parallel group studies are required to confirm the efficacy of solifenacin^[249].

Tiropamide

Tiropamide, a derivative of tyrosine, has a spasmolytic effect on the intestine, decreasing Ca^{2+} release into intestinal smooth muscle^[251-253]. In a double-blind placebo-controlled randomized trial performed by Lee *et al.*^[251], tiropamide was associated with an improvement of total symptom scores for 4 wk compared with 3 wk in the placebo group. In addition, only patients treated with tiropamide improved abdominal pain at week 4^[254].

Lee *et al.*^[251] successively performed a multicentre, randomized, non-inferiority trial involving 287 patients with IBS randomly assigned to either tiropamide 100 mg or octylonium 20 mg t.i.d. (means 3 times a day) for 4 wk^[1]. The visual analogue scale (VAS) scores of abdominal pain at week 4 were significantly reduced in both tiropamide and octylonium groups, even though the change from baseline was similar in the 2 groups^[251]. In both groups, abdominal pain and discomfort assessed using VAS scores, diaries and IBS-QoL were improved, and no differences in the changes from baseline were observed. Side effects were similar in both groups. No severe side effects involving either drug were observed^[251].

Despite the useful results of the abovementioned study, further studies are required to elucidate tiropamide's pharmacodynamic and pharmacokinetic properties and its mechanism of action on the intestine^[251].

Eluxadoline

Eluxadoline is a μ - and κ -opioid receptor agonist and δ -opioid receptor antagonist. Its action is directed to the enteric nervous system, with slight side effects in the central nervous system. Its use was approved by the United States Food and Drug Administration on May 2015^[255-257].

Patients with IBS-D receiving eluxadoline (100 mg twice daily) in a phase II dose-ranging study had greater efficacy compared with patients receiving placebo after 12 wk^[258]. Eluxadoline improved the

number of daily bowel movements and decreased the episodes of urgency and incontinence experienced by patients during the 3-mo treatment period^[258]. Eluxadoline had an overall favorable safety profile, with nausea, abdominal pain, vomiting, and constipation the most commonly reported AEs^[258].

Subsequently, in two large Phase 3 trials (IBS-3001 and IBS-3002), the efficacy of eluxadoline in patients with IBS-D was shown^[259].

Finally, Cash *et al.*^[260] reported pooled safety and tolerability data from Phase 2 and 3 clinical studies for the approved doses of eluxadoline: 75 and 100 mg. The authors demonstrated that constipation and nausea were the most common adverse events^[260]. Consistent with the known adverse effects of opioid agonists, clinically apparent sphincter of Oddi spasm events were observed in eluxadoline-treated patients without a gallbladder. The majority of these cases were observed in patients on the higher dose of eluxadoline, thus suggesting a possible association^[260].

Table 4 sums up the literature findings about irritable bowel syndrome-D therapies.

CONCLUSION

IBS currently remains a field of intense therapeutic research, in which most of the aforementioned studies focus on stool-pattern-specific subcategories of patients with this condition. Multiple further drugs are also under evaluation. Among these, alpha galactosidase (AG) was shown to reduce meteorism associated with black bean ingestion, even though it is unknown whether it may have a benefit on IBS^[261]. However, in a subsequent study performed by Hillilä *et al.*^[262], no evidence to support the use of AG routinely in IBS patients was found.

With regards to therapies restoring intestinal permeability, multiple studies with prebiotics and probiotics^[263] are ongoing, even if to date their efficacy has been limited. In parallel, much progress has been made in targeting low-grade inflammation, especially through the introduction of drugs such as mesalazine and rifaximin, even if a better knowledge of the mechanisms underlying the low-grade inflammation in IBS may support the design of clinical trials aimed at evaluating the efficacy and safety of such drugs.

On the other hand, the non-pharmacological treatment of IBS is often viewed as attractive. Faecal microbiota transfer, dietary interventions, holistic and integrative medicine approaches currently represent possible future therapeutic alternatives in this setting.

In conclusion, long-term studies and comparative studies with pharmacotherapy, as well as elucidation of the underlying mechanisms of action, are still needed to find the correct algorithm to manage IBS patients.

Table 4 Summary of the literature findings about irritable bowel syndrome-D therapies

Drug	Ref.	No. of pt	Study design	Outcomes
Ramosetron	Lee <i>et al</i> ^[213]	343 male pt	A multicenter, randomized, open-label trial male patients with IBS-D; pt were randomized to either a 4-wk treatment of ramosetron 5 mg once daily ,or a 4-wk treatment of mebeverine 135 mg three times daily	Global IBS symptoms, abdominal pain/discomfort and abnormal bowel habits in the ramosetron and mebeverine groups significantly increased during the treatment period. The severity scores of abdominal pain/discomfort and urgency, the stool form score, and the stool frequency in both treatment arms were significantly reduced, compared with the baselines
	Fukudo <i>et al</i> ^[222]	296 male pt	A randomized, double-blind, placebo-controlled trial in male patients with IBS-D Patients were given 5 mg oral ramosetron (<i>n</i> = 147) or placebo (<i>n</i> = 149) once daily for 12 wk after a 1-wk baseline period	Improving stool consistency in the first month. The ramosetron group had significantly higher monthly rates of relief of overall IBS symptoms and IBS-related quality of life than the placebo group. Adverse events occurring in 46.9% and 51.7% of ramosetron and placebo patients, respectively
	Fukudo <i>et al</i> ^[223]	576 female pt	A randomized, double-blind, placebo-controlled trial. The subjects received either 2.5 µg ramosetron or placebo once daily for 12 wk.	Global improvement, an increased stool consistency a significant reductions in abdominal pain and discomfort and greater improvement in QOL compared with placebo
Lx1031	Brown <i>et al</i> ^[229]	155 patients	A phase- II multicenter, randomized, double-blind, placebo-controlled, the subjects were assigned randomly in a double-blind fashion to 1 of 2 doses of LX1031 (250 mg 4 times/d or 1000 mg 4 times/d) or placebo, taken daily during the 28-d treatment period	Improved significantly in patients given 1000 mg LX1031 compared with those given placebo, at week 1, together with nonsignificant improvements at weeks 2, 3 and 4. Adverse Effects reported were generally mild, self-limited, and evenly distributed across the placebo and both LX1031 treatment arms
ASP-7147	Lembo <i>et al</i> ^[230]	64 patients	RCT performed on during a 4-wk	Demonstrating improvement in multiple symptoms of IBS-D. The persistence of treatment effect suggests the possibility of retained efficacy with less frequent dosing in follow-on trials
JNJ-27018966	[232]	807 patients	A randomized, controlled, double-blind study, 25, 100, and 200 mg twice daily to placebo	The composite of diarrhea and pain was significantly improved in the JNJ-27018966 25 and 200 mg twice-daily groups compared to placebo
ROSE-010	Hellström <i>et al</i> ^[233]		A randomized placebo-controlled trial. Patients were randomized to ROSE-010 100 µg once daily, 300 µg once daily or placebo	Treatment with ROSE-010 resulted in a two fold greater response to abdominal pain compared to placebo and significantly greater patient-reported satisfaction with ROSE-010. The most common treatment-related adverse effect was nausea
AST-120	Tack <i>et al</i> ^[95]	160 patients 115 non-constipation-related IBS patients	A randomized, double-blind, controlled study	AST-120 2 g three times daily significantly improved the proportion of patients with at least a 50% reduction in the number of days with abdominal pain compared to placebo. AST-120 resulted in significantly improved bloating and numerically improved stool consistency compared to placebo. The safety profile AST-120 was similar to placebo
Ibodontant	Trinkley <i>et al</i> ^[231]	559 IBS-D patients	A randomized, double-blind, controlled trial	Improved abdominal pain, satisfactory relief of overall symptoms, and quality of life compared to placebo. All three doses of ibodontant (1, 3, 10 mg once daily) were superior to placebo, but 10 mg once daily was most effective and females responded better than males
Asimadoline	Trinkley <i>et al</i> ^[231]	596 IBS-D patients	A randomized, controlled, double-blind trial compared asimadoline 0.15, 0.5 and 1 mg twice daily to placebo	Asimadoline 0.5 mg twice daily significantly improved by two fold the total number of months with adequate relief of IBS pain, pain scores, urgency and frequency
Colestevlam	Odunsi-Shiyanbade <i>et al</i> ^[155]	12 IBS-D patients	Single center trial	Colestevlam modestly affected overall colonic transit (emptying of the ascending colon took an average 4 h longer in patients given colestevlam compared to placebo). Furthermore, colestevlam was associated with greater ease of stool passage and somewhat firmer stool consistency. No effects on mucosal permeability or safety were identified
	Camilleri <i>et al</i> ^[240]	12 IBS-D patients	A 10-d single-center, unblinded, single-dose trial	Colestevlam increases delivery of BAs to stool while improving stool consistency, and increases hepatic BA synthesis, avoiding steatorrhea in patients with IBS-D

Solinefacin	Fukushima <i>et al</i> ^[249]	20 IBS-D patients	An open-label trial. After a 2-wk observation period, all participants received solifenacin for 6 wk. Subsequently, the administration of solifenacin was discontinued and ramosetron, a serotonin 3 receptor antagonist, was administered for 4 wk	The efficacy of solifenacin in the treatment of IBS with diarrhea was not inferior to that of ramosetron
Tiropamide	Lee <i>et al</i> ^[251]	287 patients	A multicenter, randomized, non-inferiority Patients randomly allocated to either tiropamide 100 mg or octylonium 20 mg t.i.d (means 3 times a day) for 4 wk	Tiropamide led to symptom improvement in terms of total symptom scores for 4 wk, compared with 3 wk in the placebo group; in addition, at week 4 abdominal pain was only improved in the tiropamide group. The incidence of adverse events was similar in the 2 groups, and no severe adverse events involving either drug were observed

IBS : Irritable bowel syndrome.

REFERENCES

- Sinagra E, Romano C, Cottone M. Psychopharmacological treatment and psychological interventions in irritable bowel syndrome. *Gastroenterol Res Pract* 2012; **2012**: 486067 [PMID: 22956940 DOI: 10.1155/2012/486067]
- Lovell RM, Ford AC. Global prevalence of, and risk factors for, irritable bowel syndrome: a meta-analysis. *Clin Gastroenterol Hepatol* 2012; **10**: 712-721
- Quigley EM, Abdel-Hamid H, Barbara G, Bhatia SJ, Boeckxstaens G, De Giorgio R, Delvaux M, Drossman DA, Foxx-Orenstein AE, Guarner F, Gwee KA, Harris LA, Hungin AP, Hunt RH, Kellow JE, Khalif IL, Kruis W, Lindberg G, Olano C, Moraes-Filho JP, Schiller LR, Schmulson M, Simrén M, Tzeuton C. A global perspective on irritable bowel syndrome: a consensus statement of the World Gastroenterology Organisation Summit Task Force on Irritable Bowel Syndrome. *J Clin Gastroenterol* 2012; **46**: 356-366 [PMID: 22499071 DOI: 10.1097/MCG.0b013e318247157c]
- Ford AC, Moayyedi P, Lacy BE, Lembo AJ, Saito YA, Schiller LR, Soffer EE, Spiegel BM, Quigley EM; Task Force on the Management of Functional Bowel Disorders. American College of Gastroenterology monograph on the management of irritable bowel syndrome and chronic idiopathic constipation. *Am J Gastroenterol* 2014; **109** Suppl 1: S2-S26; quiz S27 [PMID: 25091148 DOI: 10.1038/ajg.2014.187]
- Enck P, Aziz Q, Barbara G, Farmer AD, Fukudo S, Mayer EA, Niesler B, Quigley EM, Rajilić-Stojanović M, Schemann M, Schwille-Kiuntke J, Simren M, Zipfel S, Spiller RC. Irritable bowel syndrome. *Nat Rev Dis Primers* 2016; **2**: 16014 [PMID: 27159638 DOI: 10.1038/nrdp.2016.14]
- Layer P, Andresen V, Diemert S, Mackinnon J, Bertsch J, Fortea J, Tack J. Economic Burden and Quality of Life of Moderate-To-Severe Irritable Bowel Syndrome With Constipation (Ibs-C) In Germany: Results From The Ibis-C Study. *Value Health* 2015; **18**: A624 [PMID: 26533504 DOI: 10.1016/j.jval.2015.09.2193]
- Mulak A, Taché Y. Sex difference in irritable bowel syndrome: do gonadal hormones play a role? *Gastroenterol Pol* 2010; **17**: 89-97 [PMID: 25435761]
- Heitkemper M, Jarrett M, Bond EF, Chang L. Impact of sex and gender on irritable bowel syndrome. *Biol Res Nurs* 2003; **5**: 56-65 [PMID: 12886671]
- Longstreth GF, Wolde-Tsadik G. Irritable bowel-type symptoms in HMO examinees. Prevalence, demographics, and clinical correlates. *Dig Dis Sci* 1993; **38**: 1581-1589 [PMID: 8359067]
- Toner BB, Akman D. Gender role and irritable bowel syndrome: literature review and hypothesis. *Am J Gastroenterol* 2000; **95**: 11-16 [PMID: 10638553]
- Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorders. *Gastroenterology* 2006; **130**: 1480-1491 [PMID: 16678561]
- Mearin F, Lacy BE, Chang L, Chey WD, Lembo AJ, Simren M, Spiller R. Bowel Disorders. *Gastroenterology* 2016; Epub ahead of print [PMID: 27144627 DOI: 10.1053/j.gastro.2016.02.031]
- Lewis SJ, Heaton KW. Stool form scale as a useful guide to intestinal transit time. *Scand J Gastroenterol* 1997; **32**: 920-924 [PMID: 9299672]
- El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Irritable bowel syndrome: diagnosis, pathogenesis and treatment options. New York: Nova Science Publishers, Inc., 2012
- El-Salhy M, Gundersen D, Gilja OH, Hatlebakk JG, Hausken T. Is irritable bowel syndrome an organic disorder? *World J Gastroenterol* 2014; **20**: 384-400 [PMID: 24574708 DOI: 10.3748/wjg.v20.i2.384]
- Dupont HL. Review article: evidence for the role of gut microbiota in irritable bowel syndrome and its potential influence on therapeutic targets. *Aliment Pharmacol Ther* 2014; **39**: 1033-1042 [DOI: 10.1111/apt.12728.46]
- Spiller R, Garsed K. Postinfectious irritable bowel syndrome. *Gastroenterology* 2009; **136**: 1979-88 [DOI: 10.1053/j.gastro.2009.02.074.5]
- Matricon J, Meleine M, Gelot A, Piche T, Dapigny M, Muller E, Ardid D. Review article: Associations between immune activation, intestinal permeability and the irritable bowel syndrome. *Aliment Pharmacol Ther* 2012; **36**: 1009-1031 [PMID: 23066886 DOI: 10.1111/Apt.12080]
- Wadhwa A, Camilleri M, Grover M. New and Investigational Agents for Irritable Bowel Syndrome. *Curr Gastroenterol Rep* 2015; **17**: 46 [PMID: 26446557 DOI: 10.1007/s11894-015-0473-x]
- Barbara G, De Giorgio R, Stanghellini V, Cremon C, Corinaldesi R. A role for inflammation in irritable bowel syndrome? *Gut* 2002; **51** Suppl 1: i41-44 [PMID 12077063]
- Lee E, Schiller LR, Fordtran JS. Quantification of colonic lamina propria cells by means of a morphometric point-counting method. *Gastroenterology* 1988; **94**: 409-418 [PMID: 3335315]
- Salzmann JL, Peltier-Koch F, Bloch F, Petite JP, Camilleri JP. Morphometric study of colonic biopsies: a new method of estimating inflammatory diseases. *Lab Invest* 1989; **60**: 847-851 [PMID: 2733385]
- Barbara G, Cremon C, Carini G, Bellacosa L, Zecchi L, De Giorgio R, Corinaldesi R, Stanghellini V. The immune system in irritable bowel syndrome. *J Neurogastroenterol Motil* 2011; **17**: 349-359 [PMID: 22148103 DOI: 10.5056/jnm.2011.17.4.349]
- Dunlop SP, Jenkins D, Neal KR, Naesdal J, Borgaonker M, Collins SM, Spiller RC. Randomized, double-blind, placebo-controlled trial of prednisolone in post-infectious irritable bowel syndrome. *Aliment Pharmacol Ther* 2003; **18**: 77-84 [PMID: 12848628]
- Sinagra E, Pompei G, Tomasello G, Cappello F, Morreale GC, Amvrosiadis G, Rossi F, Lo Monte AI, Rizzo AG, Raimondo D. Inflammation in irritable bowel syndrome: Myth or new treatment target? *World J Gastroenterol* 2016; **22**: 2242-2255 [PMID: 26900287 DOI: 10.3748/wjg.v22.i7.2242]
- Barbara G, Feinle-Bisset C, Ghoshal UC, Quigley EM, Santos J, Vanner S, Vergnolle N, Zoetendal EG. The Intestinal Microenvironment and Functional Gastrointestinal Disorders. *Gastroenterology* 2016; Epub ahead of print [PMID: 27144620]

- DOI: 10.1053/j.gastro.2016.02.028]
- 27 **Gillis JC**, Brogden RN. Rifaximin. A review of its antibacterial activity, pharmacokinetic properties and therapeutic potential in conditions mediated by gastrointestinal bacteria. *Drugs* 1995; **49**: 467-484 [PMID: 7774516]
 - 28 **DuPont HL**. Review article: the antimicrobial effects of rifaximin on the gut microbiota. *Aliment Pharmacol Ther* 2016; **43** Suppl 1: 3-10 [PMID: 26618921 DOI: 10.1111/apt.13434]
 - 29 **Pimentel M**, Lembo A, Chey WD, Zakko S, Ringel Y, Yu J, Mareya SM, Shaw AL, Bortey E, Forbes WP; TARGET Study Group. Rifaximin therapy for patients with irritable bowel syndrome without constipation. *N Engl J Med* 2011; **364**: 22-32 [PMID: 21208106 DOI: 10.1056/NEJMoa1004409]
 - 30 **Lacy BE**. Diagnosis and treatment of diarrhea-predominant irritable bowel syndrome. *Int J Gen Med* 2016; **9**: 7-17 [PMID: 26929659 DOI: 10.2147/IJGM.S93698]
 - 31 **Lembo A**, Pimentel M, Rao SS, Schoenfeld P, Cash B, Weinstock LB, Golden PL, Paterson C, Bortey E, Forbes WP. Efficacy and safety of repeat treatment with rifaximin for diarrhea-predominant irritable bowel syndrome (IBS-D): results of the TARGET 3 study. Presented at: American College of Gastroenterology (ACG) 2014 Annual Scientific Meeting; October 17-22; 2014; Philadelphia, PA
 - 32 **Ghoshal UC**, Srivastava D, Misra A, Ghoshal U. A proof-of-concept study showing antibiotics to be more effective in irritable bowel syndrome with than without small-intestinal bacterial overgrowth: a randomized, double-blind, placebo-controlled trial. *Eur J Gastroenterol Hepatol* 2016; **28**: 281-289 [PMID: 26731696 DOI: 10.1097/MEG.0000000000000557]
 - 33 **Klooker TK**, Braak B, Koopman KE, Welting O, Wouters MM, van der Heide S, Schemann M, Bischoff SC, van den Wijngaard RM, Boeckxstaens GE. The mast cell stabiliser ketotifen decreases visceral hypersensitivity and improves intestinal symptoms in patients with irritable bowel syndrome. *Gut* 2010; **59**: 1213-1221 [PMID: 20650926 DOI: 10.1136/gut.2010.213108]
 - 34 **Lobo B**, Pigrau M, Martinez C, González-Castro AM, Guilarte M, de torres I, Salvo-Romero E, Rodiño-Janeiro BK, Fortea M, Cotoner CA, Azpiroz F, Vicario M, Santos J. Clinical Benefit and Intestinal Mucosal Transcriptome Modulation After Long-Term Mast Cell Stabilization With Oral Disodium Cromoglycate in Diarrhea-Predominant Irritable Bowel Syndrome (IBS-D) Patients. *Gastroenterology* 2015; **148**: S-494 [DOI: 10.1016/S0016-5085(09)60139-6]
 - 35 **Wouters MM**, Balemans D, Van Wanrooy S, Dooley J, Cibert-Goton V, Alpizar YA, Valdez-Morales EE, Nasser Y, Van Veldhoven PP, Vanbrabant W, Van der Merwe S, Mols R, Ghesquière B, Cirillo C, Kortekaas I, Carmeliet P, Peetermans WE, Vermeire S, Rutgeerts P, Augustijns P, Hellings PW, Belmans A, Vanner S, Bulmer DC, Talavera K, Vanden Berghe P, Liston A, Boeckxstaens GE. Histamine Receptor H1-Mediated Sensitization of TRPV1 Mediates Visceral Hypersensitivity and Symptoms in Patients With Irritable Bowel Syndrome. *Gastroenterology* 2016; **150**: 875-877. e9 [PMID: 26752109 DOI: 10.1053/j.gastro.2015.12.034]
 - 36 **Wouters MM**, Vicario M, Santos J. The role of mast cells in functional GI disorders. *Gut* 2016; **65**: 155-168 [PMID: 26194403 DOI: 10.1136/gutjnl-2015-309151]
 - 37 **Dorofeyev AE**, Kiriyan EA, Vasilenko IV, Rassokhina OA, Elin AF. Clinical, endoscopic and morphological efficacy of mesalazine in patients with irritable bowel syndrome. *Clin Exp Gastroenterol* 2011; **4**: 141-153 [PMID: 21753896 DOI: 10.2147/CEG.S18381]
 - 38 **Barbara G**, Stanghellini V, Cremon C, De Giorgio R, Fronzoni L, Serra M, Corinaldesi R. Aminosalicilates and other anti-inflammatory compounds for irritable bowel syndrome. *Dig Dis* 2009; **27** Suppl 1: 115-121 [PMID: 20203507 DOI: 10.1159/000268131]
 - 39 **Barbara G**, Cremon C, Annesse V, Basilisco G, Bazzoli F, Bellini M, Benedetti A, Benini L, Bossa F, Buldrini P, Cicala M, Cuomo R, Germanà B, Molteni P, Neri M, Rodi M, Saggiaro A, Scribano ML, Vecchi M, Zoli G, Corinaldesi R, Stanghellini V. Randomised controlled trial of mesalazine in IBS. *Gut* 2016; **65**: 82-90 [PMID: 25533646 DOI: 10.1136/gutjnl-2014-308188]
 - 40 **Lam C**, Tan W, Leighton M, Hastings M, Lingaya M, Falcone Y, Zhou X, Xu L, Whorwell P, Walls AF, Zaitoun A, Montgomery A, Spiller RC. Efficacy and mode of action of mesalazine in the treatment of diarrhoea-predominant irritable bowel syndrome (IBS-D): a multicentre, parallel-group, randomised placebo-controlled trial. Southampton (UK): NIHR Journals Library; 2015
 - 41 **Min T**, Ford AC. Efficacy of mesalazine in IBS. *Gut* 2016; **65**: 187-188 [PMID: 25873641 DOI: 10.1136/gutjnl-2015-309593]
 - 42 **Hollander D**, Vadheim CM, Brettholz E, Petersen GM, Delahunty T, Rotter JJ. Increased intestinal permeability in patients with Crohn's disease and their relatives. A possible etiologic factor. *Ann Intern Med* 1986; **105**: 883-885 [PMID: 3777713]
 - 43 **Scaldeferri F**, Pizzoferrato M, Gerardi V, Lopetuso L, Gasbarrini A. The gut barrier: new acquisitions and therapeutic approaches. *J Clin Gastroenterol* 2012; **46** Suppl: S12-S17 [PMID: 22955350 DOI: 10.1097/MCG.0b013e31826ae849]
 - 44 **Öhman L**, Törnblom H, Simrén M. Crosstalk at the mucosal border: importance of the gut microenvironment in IBS. *Nat Rev Gastroenterol Hepatol* 2015; **12**: 36-49 [PMID: 25446728 DOI: 10.1038/nrgastro.2014.200]
 - 45 **Camilleri M**, Gorman H. Intestinal permeability and irritable bowel syndrome. *Neurogastroenterol Motil* 2007; **19**: 545-552 [PMID: 17593135]
 - 46 **Barbara G**, Zecchi L, Barbaro R, Cremon C, Bellacosa L, Marcellini M, De Giorgio R, Corinaldesi R, Stanghellini V. Mucosal permeability and immune activation as potential therapeutic targets of probiotics in irritable bowel syndrome. *J Clin Gastroenterol* 2012; **46** Suppl: S52-S55 [PMID: 22955358 DOI: 10.1097/MCG.0b013e318264e918]
 - 47 **Spiller RC**, Jenkins D, Thornley JP, Hebden JM, Wright T, Skinner M, Neal KR. Increased rectal mucosal enteroendocrine cells, T lymphocytes, and increased gut permeability following acute Campylobacter enteritis and in post-dysenteric irritable bowel syndrome. *Gut* 2000; **47**: 804-811 [PMID: 11076879]
 - 48 **Marshall JK**, Thabane M, Garg AX, Clark W, Meddings J, Collins SM; WEL Investigators. Intestinal permeability in patients with irritable bowel syndrome after a waterborne outbreak of acute gastroenteritis in Walkerton, Ontario. *Aliment Pharmacol Ther* 2004; **20**: 1317-1322 [PMID: 15606393]
 - 49 **Villani AC**, Lemire M, Thabane M, Belisle A, Geneau G, Garg AX, Clark WF, Moayyedi P, Collins SM, Franchimont D, Marshall JK. Genetic risk factors for post-infectious irritable bowel syndrome following a waterborne outbreak of gastroenteritis. *Gastroenterology* 2010; **138**: 1502-1513 [PMID: 20044998 DOI: 10.1053/j.gastro.2009.12.049]
 - 50 **Phua LC**, Wilder-Smith CH, Tan YM, Gopalakrishnan T, Wong RK, Li X, Kan ME, Lu J, Keshavarzian A, Chan EC. Gastrointestinal Symptoms and Altered Intestinal Permeability Induced by Combat Training Are Associated with Distinct Metabotypic Changes. *J Proteome Res* 2015; **14**: 4734-4742 [PMID: 26506213 DOI: 10.1021/acs.jproteome.5b00603]
 - 51 **Moloney RD**, Johnson AC, O'Mahony SM, Dinan TG, Greenwood-Van Meerveld B, Cryan JF. Stress and the Microbiota-Gut-Brain Axis in Visceral Pain: Relevance to Irritable Bowel Syndrome. *CNS Neurosci Ther* 2016; **22**: 102-117 [PMID: 26662472 DOI: 10.1111/cns.12490]
 - 52 **Annaházi A**, Ferrier L, Bézirard V, Lévêque M, Eutamène H, Ait-Belgnaoui A, Coëffier M, Ducrotté P, Róka R, Inczeff O, Gecse K, Rosztóczy A, Molnár T, Ringel-Kulka T, Ringel Y, Piche T, Theodorou V, Wittmann T, Bueno L. Luminal cysteine-proteases degrade colonic tight junction structure and are responsible for abdominal pain in constipation-predominant IBS. *Am J Gastroenterol* 2013; **108**: 1322-1331 [PMID: 23711626 DOI: 10.1038/ajg.2013.152]
 - 53 **Gecse K**, Róka R, Ferrier L, Leveque M, Eutamène H, Cartier C, Ait-Belgnaoui A, Rosztóczy A, Izbéki F, Fioramonti J, Wittmann T, Bueno L. Increased faecal serine protease activity in diarrhoeic IBS patients: a colonic lumenal factor impairing colonic permeability and sensitivity. *Gut* 2008; **57**: 591-599 [PMID: 18194983 DOI: 10.1136/gut.2007.140210]

- 54 **Turner JR.** Intestinal mucosal barrier function in health and disease. *Nat Rev Immunol* 2009; **9**: 799-809 [PMID: 19855405 DOI: 10.1038/nri2653]
- 55 **Günzel D, Yu AS.** Claudins and the modulation of tight junction permeability. *Physiol Rev* 2013; **93**: 525-569 [PMID: 23589827 DOI: 10.1152/physrev.00019.2012]
- 56 **Ash C, Mueller K.** Manipulating the Microbiota. *Science* 2016; **352**: 530-531 [PMID: 27126033 DOI: 10.1126/science.352.6285.530]
- 57 **Marchesi JR, Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, Quraishi MN, Kinross J, Smidt H, Tuohy KM, Thomas LV, Zoetendal EG, Hart A.** The gut microbiota and host health: a new clinical frontier. *Gut* 2016; **65**: 330-339 [PMID: 26338727 DOI: 10.1136/gutjnl-2015-309990]
- 58 **Borowiec AM, Fedorak RN.** The role of probiotics in management of irritable bowel syndrome. *Curr Gastroenterol Rep* 2007; **9**: 393-400 [PMID: 17991340]
- 59 **Madsen K, Cornish A, Soper P, McKaigney C, Jijon H, Yachimec C, Doyle J, Jewell L, De Simone C.** Probiotic bacteria enhance murine and human intestinal epithelial barrier function. *Gastroenterology* 2001; **121**: 580-591 [PMID: 11522742]
- 60 **Zeng J, Li YQ, Zuo XL, Zhen YB, Yang J, Liu CH.** Clinical trial: effect of active lactic acid bacteria on mucosal barrier function in patients with diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 2008; **28**: 994-1002 [PMID: 18671775 DOI: 10.1111/j.1365-2036.2008.03818.x]
- 61 **Ford AC, Quigley EM, Lacy BE, Lembo AJ, Saito YA, Schiller LR, Soffer EE, Spiegel BM, Moayyedi P.** Efficacy of prebiotics, probiotics, and synbiotics in irritable bowel syndrome and chronic idiopathic constipation: systematic review and meta-analysis. *Am J Gastroenterol* 2014; **109**: 1547-1561; quiz 1546, 1562 [PMID: 25070051 DOI: 10.1038/ajg.2014.202]
- 62 **Distruitti E, Monaldi L, Ricci P, Fiorucci S.** Gut microbiota role in irritable bowel syndrome: New therapeutic strategies. *World J Gastroenterol* 2016; **22**: 2219-2241 [PMID: 26900286 DOI: 10.3748/wjg.v22.i7.2219]
- 63 **Ohland CL, Macnaughton WK.** Probiotic bacteria and intestinal epithelial barrier function. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G807-G819 [PMID: 20299599 DOI: 10.1152/ajpgi.00243.2009]
- 64 **Nikfar S, Rahimi R, Rahimi F, Derakhshani S, Abdollahi M.** Efficacy of probiotics in irritable bowel syndrome: a meta-analysis of randomized, controlled trials. *Dis Colon Rectum* 2008; **51**: 1775-1780 [PMID: 18465170 DOI: 10.1007/s10350-008-9335-z]
- 65 **Hoveyda N, Heneghan C, Mahtani KR, Perera R, Roberts N, Glasziou P.** A systematic review and meta-analysis: probiotics in the treatment of irritable bowel syndrome. *BMC Gastroenterol* 2009; **9**: 15 [PMID: 19220890 DOI: 10.1186/1471-230X-9-15]
- 66 **Moayyedi P, Ford AC, Talley NJ, Cremonini F, Foxx-Orenstein AE, Brandt LJ, Quigley EM.** The efficacy of probiotics in the treatment of irritable bowel syndrome: a systematic review. *Gut* 2010; **59**: 325-332 [PMID: 19091823 DOI: 10.1136/gut.2008.167270]
- 67 **Mazurak N, Broelz E, Storr M, Enck P.** Probiotic Therapy of the Irritable Bowel Syndrome: Why Is the Evidence Still Poor and What Can Be Done About It? *J Neurogastroenterol Motil* 2015; **21**: 471-485 [PMID: 26351253 DOI: 10.5056/jnm15071]
- 68 **Guidance for Industry.** Irritable bowel syndrome-clinical evaluation of drugs for treatment: Food and Drug Administration (FDA), Center for Drug Evaluation and Research, May 2012
- 69 **Guideline on the evaluation of medicinal products for the treatment of irritable bowel syndrome.** CPMP/EWP/785/97 Rev. 1: European Medicines Agency (EMA), Committee for Medicinal Products for Human use, Sept 2014
- 70 **Xi P, Jiang Z, Dai Z, Li X, Yao K, Zheng C, Lin Y, Wang J, Wu G.** Regulation of protein turnover by L-glutamine in porcine intestinal epithelial cells. *J Nutr Biochem* 2012; **23**: 1012-1017 [PMID: 22000664 DOI: 10.1016/j.jnutbio.2011.05.009]
- 71 **Rapin JR, Wiernsperger N.** Possible links between intestinal permeability and food processing: A potential therapeutic niche for glutamine. *Clinics (Sao Paulo)* 2010; **65**: 635-643 [PMID: 20613941 DOI: 10.1590/S1807-59322010000600012]
- 72 **De-Souza DA, Greene LJ.** Intestinal permeability and systemic infections in critically ill patients: effect of glutamine. *Crit Care Med* 2005; **33**: 1125-1135 [PMID: 15891348]
- 73 **Akobeng AK, Elawad M, Gordon M.** Glutamine for induction of remission in Crohn's disease. *Cochrane Database Syst Rev* 2016; **2**: CD007348 [PMID: 26853855 DOI: 10.1002/14651858.CD007348.pub2]
- 74 **Zhou Q, Souba WW, Croce CM, Verne GN.** MicroRNA-29a regulates intestinal membrane permeability in patients with irritable bowel syndrome. *Gut* 2010; **59**: 775-784 [PMID: 19951903 DOI: 10.1136/gut.2009.181834]
- 75 **Bertrand J, Ghouzali I, Guérin C, Bôle-Feysot C, Gouteux M, Déchelotte P, Ducrotté P, Coëffier M.** Glutamine Restores Tight Junction Protein Claudin-1 Expression in Colonic Mucosa of Patients With Diarrhea-Predominant Irritable Bowel Syndrome. *JPEN J Parenter Enter Nutr* 2016; **40**: 1170-1176 [PMID: 25972430]
- 76 **Gayathri D, Rashmi BS.** Development of Celiac Disease; Pathogenesis and Strategies to Control: A Molecular Approach. *J Nutr Food Sci* 2014; **4**: 310 [DOI: 10.4172/2155-9600.1000310]
- 77 **Paterson BM, Lammers KM, Arrieta MC, Fasano A, Meddings JB.** The safety, tolerance, pharmacokinetic and pharmacodynamic effects of single doses of AT-1001 in coeliac disease subjects: a proof of concept study. *Aliment Pharmacol Ther* 2007; **26**: 757-766 [PMID: 17697209]
- 78 **Leffler DA, Kelly CP, Green PH, Fedorak RN, DiMarino A, Perrow W, Rasmussen H, Wang C, Bercik P, Bachir NM, Murray JA.** Larazotide acetate for persistent symptoms of celiac disease despite a gluten-free diet: a randomized controlled trial. *Gastroenterology* 2015; **148**: 1311-1319.e6 [PMID: 25683116 DOI: 10.1053/j.gastro.2015.02.008]
- 79 **Nusrat S, Miner PB Jr.** New pharmacological treatment options for irritable bowel syndrome with constipation. *Expert Opin Emerg Drugs* 2015; **20**: 625-636 [PMID: 26548544 DOI: 10.1517/14728214.2015.1105215]
- 80 **Blackshaw LA, Brierley SM.** Emerging receptor target in the pharmacotherapy of irritable bowel syndrome with constipation. *Expert Rev Gastroenterol Hepatol* 2013; **7**: 15-19 [PMID: 23859756 DOI: 10.1586/17474124.2013.820045]
- 81 **Sood R, Ford AC.** Linaclotide: new mechanisms and new promise for treatment in constipation and irritable bowel syndrome. *Ther Adv Chronic Dis* 2013; **4**: 268-276 [PMID: 24179669 DOI: 10.1177/2040622313500110]
- 82 **Andresen V, Camilleri M, Busciglio IA, Grudell A, Burton D, McKinzie S, Foxx-Orenstein A, Kurtz CB, Sharma V, Johnston JM, Currie MG, Zinsmeister AR.** Effect of 5 days linaclotide on transit and bowel function in females with constipation-predominant irritable bowel syndrome. *Gastroenterology* 2007; **133**: 761-768 [PMID: 17854590]
- 83 **Johnston JM, Kurtz CB, Macdougall JE, Lavins BJ, Currie MG, Fitch DA, O'Dea C, Baird M, Lembo AJ.** Linaclotide improves abdominal pain and bowel habits in a phase IIb study of patients with irritable bowel syndrome with constipation. *Gastroenterology* 2010; **139**: 1877-1886.e2 [PMID: 20801122 DOI: 10.1053/j.gastro.2010.08.041]
- 84 **Chey WD, Lembo AJ, Lavins BJ, Shiff SJ, Kurtz CB, Currie MG, MacDougall JE, Jia XD, Shao JZ, Fitch DA, Baird MJ, Schneider HA, Johnston JM.** Linaclotide for irritable bowel syndrome with constipation: a 26-week, randomized, double-blind, placebo-controlled trial to evaluate efficacy and safety. *Am J Gastroenterol* 2012; **107**: 1702-1712 [PMID: 22986437]
- 85 **Chang L, Lembo AJ, Lavins BJ, Shiff SJ, Hao X, Chickering JG, Jia XD, Currie MG, Kurtz CB, Johnston JM.** The impact of abdominal pain on global measures in patients with chronic idiopathic constipation, before and after treatment with linaclotide: a pooled analysis of two randomised, double-blind, placebo-controlled, phase 3 trials. *Aliment Pharmacol Ther* 2014; **40**: 1302-1312 [PMID: 25312449 DOI: 10.1111/apt.12985]
- 86 **Rao S, Lembo AJ, Shiff SJ, Lavins BJ, Currie MG, Jia XD, Shi K, MacDougall JE, Shao JZ, Eng P, Fox SM, Schneider HA, Kurtz**

- CB, Johnston JM. A 12-week, randomized, controlled trial with a 4-week randomized withdrawal period to evaluate the efficacy and safety of linaclotide in irritable bowel syndrome with constipation. *Am J Gastroenterol* 2012; **107**: 1714-1724; quiz p.1725 [PMID: 22986440]
- 87 **Busby RW**, Bryant AP, Bartolini WP, Cordero EA, Hannig G, Kessler MM, Mahajan-Miklos S, Pierce CM, Solinga RM, Sun LJ, Tobin JV, Kurtz CB, Currie MG. Linaclotide, through activation of guanylate cyclase C, acts locally in the gastrointestinal tract to elicit enhanced intestinal secretion and transit. *Eur J Pharmacol* 2010; **649**: 328-335 [PMID: 20863829 DOI: 10.1016/j.ejphar.2010.09.019]
 - 88 **Quigley EM**, Tack J, Chey WD, Rao SS, Forste J, Falques M, Diaz C, Shiff SJ, Currie MG, Johnston JM. Randomised clinical trials: linaclotide phase 3 studies in IBS-C - a prespecified further analysis based on European Medicines Agency-specified endpoints. *Aliment Pharmacol Ther* 2013; **37**: 49-61 [PMID: 23116208 DOI: 10.1111/apt.12123]
 - 89 **Lembo AJ**, Schneier HA, Shiff SJ, Kurtz CB, MacDougall JE, Jia XD, Shao JZ, Lavins BJ, Currie MG, Fitch DA, Jeglinski BI, Eng P, Fox SM, Johnston JM. Two randomized trials of linaclotide for chronic constipation. *N Engl J Med* 2011; **365**: 527-536 [PMID: 21830967 DOI: 10.1056/NEJMoa1010863]
 - 90 ClinicalTrials.gov Registry, 2014. Available from: URL: <http://www.clinicaltrials.gov>
 - 91 **Shailubhai K**. Therapeutic applications of guanylate cyclase-C receptor agonists. *Curr Opin Drug Discov Devel* 2002; **5**: 261-268 [PMID: 11926132]
 - 92 **Miner P**, Surowitz R, Fogel R, Koltun W, Drossman DA, Camilleri M, Mangel A, Barrow L, Jacob GS, Shailubhai K. Plecanatide, a novel guanylate cyclase-C (GC-C) receptor agonist, is efficacious and safe in patients with chronic idiopathic constipation (CIC): results from a 951 patient, 12-week, multi-center trial[abstract]. *Gastroenterology* 2013; **144**: S163
 - 93 **Shailubhai K**, Comiskey S, Foss JA, Feng R, Barrow L, Comer GM, Jacob GS. Plecanatide, an oral guanylate cyclase C agonist acting locally in the gastrointestinal tract, is safe and well-tolerated in single doses. *Dig Dis Sci* 2013; **58**: 2580-2586 [PMID: 23625291 DOI: 10.1007/s10620-013-2684-z]
 - 94 **Shailubhai K**, Barrow L, Talluto C, Comiskey S, Foss J, Feng R. Plecanatide, a guanylate cyclase C agonist improves bowel habits and symptoms associated with chronic constipation in a phase II a clinical study. *Am J Gastroenterol* 2011; **106**: S502
 - 95 **Tack J**, Quigley E, Camilleri M, Vandeplasse L, Kerstens R. Efficacy and safety of oral prucalopride in women with chronic constipation in whom laxatives have failed: an integrated analysis. *United European Gastroenterol J* 2013; **1**: 48-59 [PMID: 24917940 DOI: 10.1177/2050640612474651]
 - 96 **Keating GM**. Prucalopride: a review of its use in the management of chronic constipation. *Drugs* 2013; **73**: 1935-1950 [PMID: 24194435 DOI: 10.1007/s40265-013-0140-1]
 - 97 **Müller-Lissner S**, Rykx A, Kerstens R, Vandeplasse L. A double-blind, placebo-controlled study of prucalopride in elderly patients with chronic constipation. *Neurogastroenterol Motil* 2010; **22**: 991-998, e255 [PMID: 20529205 DOI: 10.1111/j.1365-2982.2010.01533.x]
 - 98 **Camilleri M**, Kerstens R, Rykx A, Vandeplasse L. A placebo-controlled trial of prucalopride for severe chronic constipation. *N Engl J Med* 2008; **358**: 2344-2354 [PMID: 18509121 DOI: 10.1056/NEJMoa0800670]
 - 99 **Tack J**, van Outryve M, Beyens G, Kerstens R, Vandeplasse L. Prucalopride (Resolor) in the treatment of severe chronic constipation in patients dissatisfied with laxatives. *Gut* 2009; **58**: 357-365 [PMID: 18987031 DOI: 10.1136/gut.2008.162404]
 - 100 **Quigley EM**, Vandeplasse L, Kerstens R, Ausma J. Clinical trial: the efficacy, impact on quality of life, and safety and tolerability of prucalopride in severe chronic constipation--a 12-week, randomized, double-blind, placebo-controlled study. *Aliment Pharmacol Ther* 2009; **29**: 315-328 [PMID: 19035970 DOI: 10.1111/j.1365-2036.2008.03884.x]
 - 101 **Camilleri M**, Beyens G, Kerstens R, Robinson P, Vandeplasse L. Safety assessment of prucalopride in elderly patients with constipation: a double-blind, placebo-controlled study. *Neurogastroenterol Motil* 2009; **21**: 1256-e117 [PMID: 19751247 DOI: 10.1111/j.1365-2982.2009.01398.x]
 - 102 **Ke M**, Zou D, Yuan Y, Li Y, Lin L, Hao J, Hou X, Kim HJ. Prucalopride in the treatment of chronic constipation in patients from the Asia-Pacific region: a randomized, double-blind, placebo-controlled study. *Neurogastroenterol Motil* 2012; **24**: 999-e541 [PMID: 22882724 DOI: 10.1111/j.1365-2982.2012.01983.x]
 - 103 **Ke M**, Tack J, Quigley EM, Zou D, Choi SC, Leelakusolvong S, Liu A, Kim J. Effect of Prucalopride in the Treatment of Chronic Constipation in Asian and Non-Asian Women: A Pooled Analysis of 4 Randomized, Placebo-controlled Studies. *J Neurogastroenterol Motil* 2014; **20**: 458-468 [PMID: 25273116 DOI: 10.5056/jnm14029]
 - 104 **Tack J**, Stanghellini V, Dubois D, Joseph A, Vandeplasse L, Kerstens R. Effect of prucalopride on symptoms of chronic constipation. *Neurogastroenterol Motil* 2014; **26**: 21-27 [PMID: 24106924 DOI: 10.1111/nmo.12217]
 - 105 **Jadav AM**, McMullin CM, Smith J, Chapple K, Brown SR. The association between prucalopride efficacy and constipation type. *Tech Coloproctol* 2013; **17**: 555-559 [PMID: 23703575 DOI: 10.1007/s10151-013-1017-8]
 - 106 **Camilleri M**, Piessevaux H, Yiannakou Y, Tack J, Kerstens R, Quigley EM, Ke M, Da Silva S, Levine A. Efficacy and Safety of Prucalopride in Chronic Constipation: An Integrated Analysis of Six Randomized, Controlled Clinical Trials. *Dig Dis Sci* 2016; **61**: 2357-2372 [PMID: 27056037 DOI: 10.1007/s10620-016-4147-9]
 - 107 **Piessevaux H**, Corazziari E, Rey E, Simren M, Wiechowska-Kozłowska A, Kerstens R, Cools M, Barrett K, Levine A. A randomized, double-blind, placebo-controlled trial to evaluate the efficacy, safety, and tolerability of long-term treatment with prucalopride. *Neurogastroenterol Motil* 2015; **27**: 805-815 [PMID: 25808103 DOI: 10.1111/nmo.12553]
 - 108 **Yiannakou Y**, Piessevaux H, Bouchoucha M, Schiefke I, Filip R, Gabalec L, Dina I, Stephenson D, Kerstens R, Etherson K, Levine A. A randomized, double-blind, placebo-controlled, phase 3 trial to evaluate the efficacy, safety, and tolerability of prucalopride in men with chronic constipation. *Am J Gastroenterol* 2015; **110**: 741-748 [PMID: 25869393 DOI: 10.1038/ajg.2015.115]
 - 109 **Videlock EJ**, Cheng V, Cremonini F. Effects of linaclotide in patients with irritable bowel syndrome with constipation or chronic constipation: a meta-analysis. *Clin Gastroenterol Hepatol* 2013; **11**: 1084-1092.e3; quiz e68 [PMID: 23644388 DOI: 10.1016/j.cgh.2013.04.032]
 - 110 **Mendzelevski B**, Ausma J, Chanter DO, Robinson P, Kerstens R, Vandeplasse L, Camm J. Assessment of the cardiac safety of prucalopride in healthy volunteers: a randomized, double-blind, placebo- and positive-controlled thorough QT study. *Br J Clin Pharmacol* 2012; **73**: 203-209 [PMID: 21848574 DOI: 10.1111/j.1365-2125.2011.04088.x]
 - 111 **Tack J**, Camilleri M, Chang L, Chey WD, Galligan JJ, Lacy BE, Müller-Lissner S, Quigley EM, Schuurkes J, De Maeyer JH, Stanghellini V. Systematic review: cardiovascular safety profile of 5-HT₄ agonists developed for gastrointestinal disorders. *Aliment Pharmacol Ther* 2012; **35**: 745-767 [PMID: 22356640 DOI: 10.1111/j.1365-2036.2012.05011.x]
 - 112 **Cinca R**, Chera D, Gruss HJ, Halphen M. Randomised clinical trial: macrogol/PEG 3350+electrolytes versus prucalopride in the treatment of chronic constipation -- a comparison in a controlled environment. *Aliment Pharmacol Ther* 2013; **37**: 876-886 [PMID: 23480216 DOI: 10.1111/apt.12278]
 - 113 **SK biopharmaceuticals company**. YKP10811 (chronic constipations). BIO_USA-Poster_11_YKP_1081
 - 114 **Shin A**, Acosta A, Camilleri M, Boldingh A, Burton D, Ryks M, Rhoten D, Zinsmeister AR. A randomized trial of 5-hydroxytryptamine₄-receptor agonist, YKP10811, on colonic transit and bowel function in functional constipation. *Clin Gastroenterol Hepatol* 2015; **13**: 701-708.e1 [PMID: 25148765]

- DOI: 10.1016/j.cgh.2014.08.012]
- 115 **Manini ML**, Camilleri M, Goldberg M, Sweetser S, McKinzie S, Burton D, Wong S, Kitt MM, Li YP, Zinsmeister AR. Effects of Velusetrag (TD-5108) on gastrointestinal transit and bowel function in health and pharmacokinetics in health and constipation. *Neurogastroenterol Motil* 2010; **22**: 42-49, e7-e8 [PMID: 19691492 DOI: 10.1111/j.1365-2982.2009.01378.x]
 - 116 **CDER**. US Department of Health and Human Services, Food and Drug Administration Center for Drug Evaluation and Research (CDER). Guidance for industry. Irritable bowel syndrome-clinical evaluation of drugs for treatment. Accessed April 1, 2014
 - 117 **Manabe N**, Cremonini F, Camilleri M, Sandborn WJ, Burton DD. Effects of bisacodyl on ascending colon emptying and overall colonic transit in healthy volunteers. *Aliment Pharmacol Ther* 2009; **30**: 930-936 [PMID: 19678812 DOI: 10.1111/j.1365-2036.2009.04118.x]
 - 118 **Bouras EP**, Camilleri M, Burton DD, Thomforde G, McKinzie S, Zinsmeister AR. Prucalopride accelerates gastrointestinal and colonic transit in patients with constipation without a rectal evacuation disorder. *Gastroenterology* 2001; **120**: 354-360 [PMID: 11159875]
 - 119 **Vassallo M**, Camilleri M, Phillips SF, Brown ML, Chapman NJ, Thomforde GM. Transit through the proximal colon influences stool weight in the irritable bowel syndrome. *Gastroenterology* 1992; **102**: 102-108 [PMID: 1727743]
 - 120 **Mozaffari S**, Didari T, Nikfar S, Abdollahi M. Phase II drugs under clinical investigation for the treatment of chronic constipation. *Expert Opin Investig Drugs* 2014; **23**: 1485-1497 [PMID: 24960333 DOI: 10.1517/13543784.2014.932770]
 - 121 **Rao SS**, Quigley EM, Shiff SJ, Lavins BJ, Kurtz CB, MacDougall JE, Currie MG, Johnston JM. Effect of linaclotide on severe abdominal symptoms in patients with irritable bowel syndrome with constipation. *Clin Gastroenterol Hepatol* 2014; **12**: 616-623 [PMID: 24075889 DOI: 10.1016/j.cgh.2013.09.022]
 - 122 SK Chemicals Co., Ltd. Efficacy and safety of ykp10811 in subjects with irritable bowel syndrome with constipation. 2014
 - 123 SK Life Science. A Phase 2 study to evaluate pharmacodynamics of ykp10811 in patients with chronic or functional constipation. 2012
 - 124 SK Life Science. A multicenter, double-blind, randomized, placebo-controlled, 12-week, dose-range-finding trial of ykp10811 capsules administered once daily to subjects with chronic idiopathic constipation. 2013
 - 125 **Meyers NL**, Hickling RI. Pharmacology and metabolism of renzapride: a novel therapeutic agent for the potential treatment of irritable bowel syndrome. *Drugs R D* 2008; **9**: 37-63 [PMID: 18095752]
 - 126 **Meyers NL**, Hickling RI. The cardiovascular safety profile of renzapride, a novel treatment for irritable bowel syndrome. *J Int Med Res* 2007; **35**: 848-866 [PMID: 18034998]
 - 127 **Tack J**, Middleton SJ, Horne MC, Piessevaux H, Bloor JS, Meyers NL, Palmer RM. Pilot study of the efficacy of renzapride on gastrointestinal motility and symptoms in patients with constipation-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 2006; **23**: 1655-1665 [PMID: 16696817]
 - 128 **Scarpellini E**, Tack J. Renzapride: a new drug for the treatment of constipation in the irritable bowel syndrome. *Expert Opin Investig Drugs* 2008; **17**: 1663-1670 [PMID: 18922103 DOI: 10.1517/13543784.17.11.1663]
 - 129 **Camilleri M**, McKinzie S, Fox J, Foxx-Orenstein A, Burton D, Thomforde G, Baxter K, Zinsmeister AR. Effect of renzapride on transit in constipation-predominant irritable bowel syndrome. *Clin Gastroenterol Hepatol* 2004; **2**: 895-904 [PMID: 15476153]
 - 130 **George AM**, Meyers NL, Hickling RI. Clinical trial: renzapride therapy for constipation-predominant irritable bowel syndrome--multicentre, randomized, placebo-controlled, double-blind study in primary healthcare setting. *Aliment Pharmacol Ther* 2008; **27**: 830-837 [PMID: 18284648 DOI: 10.1111/j.1365-2036.2008.03649.x]
 - 131 **Ford AC**, Brandt LJ, Young C, Chey WD, Foxx-Orenstein AE, Moayyedi P. Efficacy of 5-HT₃ antagonists and 5-HT₄ agonists in irritable bowel syndrome: systematic review and meta-analysis. *Am J Gastroenterol* 2009; **104**: 1831-1843; quiz 1844 [PMID: 19471254 DOI: 10.1038/ajg.2009.223]
 - 132 **Ervin CM**, Mangel AW. Clinical trials in irritable bowel syndrome: a review. *Rev Recent Clin Trials* 2013; **8**: 9-22 [PMID: 23130604]
 - 133 **Mozaffari S**, Nikfar S, Abdollahi M. Efficacy and tolerability of renzapride in irritable bowel syndrome: a meta-analysis of randomized, controlled clinical trials including 2528 patients. *Arch Med Sci* 2014; **10**: 10-18 [PMID: 24701208 DOI: 10.5114/aoms.2014.40729]
 - 134 **He WR**, Zhang FC, Liang LX. Mixed 5-HT₃ antagonists/5-HT₄ agonists for irritable bowel syndrome: a systematic review. *World Chin J Digestol* 2011; **19**: 3277-3283
 - 135 **Lembo AJ**, Cremonini F, Meyers N, Hickling R. Clinical trial: renzapride treatment of women with irritable bowel syndrome and constipation - a double-blind, randomized, placebo-controlled, study. *Aliment Pharmacol Ther* 2010; **31**: 979-990 [PMID: 20163375 DOI: 10.1111/j.1365-2036.2010.04265.x]
 - 136 **Chang L**, Chey WD, Harris L, Olden K, Surawicz C, Schoenfeld P. Incidence of ischemic colitis and serious complications of constipation among patients using alosetron: systematic review of clinical trials and post-marketing surveillance data. *Am J Gastroenterol* 2006; **101**: 1069-1079 [PMID: 16606352]
 - 137 **Jiang C**, Xu Q, Wen X, Sun H. Current developments in pharmacological therapeutics for chronic constipation. *Acta Pharm Sin B* 2015; **5**: 300-309 [PMID: 26579459 DOI: 10.1016/j.actph.2015.05.006]
 - 138 **Buchwald P**, Bodor N. Recent advances in the design and development of soft drugs. *Pharmazie* 2014; **69**: 403-413 [PMID: 24974571]
 - 139 **Smith JA**, Beattie DT, Marquess D, Shaw JP, Vickery RG, Humphrey PP. The in vitro pharmacological profile of TD-5108, a selective 5-HT₄ receptor agonist with high intrinsic activity. *Naunyn Schmiedeberg's Arch Pharmacol* 2008; **378**: 125-137 [PMID: 18415081 DOI: 10.1007/s00210-008-0282-y]
 - 140 **Goldberg M**, Li YP, Johanson JF, Mangel AW, Kitt M, Beattie DT, Kersey K, Daniels O. Clinical trial: the efficacy and tolerability of velusetrag, a selective 5-HT₄ agonist with high intrinsic activity, in chronic idiopathic constipation - a 4-week, randomized, double-blind, placebo-controlled, dose-response study. *Aliment Pharmacol Ther* 2010; **32**: 1102-1112 [PMID: 21039672 DOI: 10.1111/j.1365-2036.2010.04456.x]
 - 141 **Beattie DT**, Armstrong SR, Shaw JP, Marquess D, Sandlund C, Smith JA, Taylor JA, Humphrey PP. The in vivo gastrointestinal activity of TD-5108, a selective 5-HT₄ receptor agonist with high intrinsic activity. *Naunyn Schmiedeberg's Arch Pharmacol* 2008; **378**: 139-147 [PMID: 18408918 DOI: 10.1007/s00210-008-0281-z]
 - 142 **Beattie DT**, Higgins DL, Ero MP, Amagasa SM, Vickery RG, Kersey K, Hopkins A, Smith JA. An in vitro investigation of the cardiovascular effects of the 5-HT₄ receptor selective agonists, velusetrag and TD-8954. *Vascul Pharmacol* 2013; **58**: 150-156 [PMID: 23201772 DOI: 10.1016/j.vph.2012.11.002]
 - 143 **Shaw JP**, Beattie D, Cheong SK. Preclinical Pharmacokinetics of TD-5108, a selective, high intrinsic activity and orally bioavailable 5-HT₄ receptor agonist. AAPS Ann Meet Expos. Presented at the annual meeting of the American Association of Pharmaceutical Scientists 2007; **9**: 2422
 - 144 **Wong SL**, Goldberg MR, Shaw J, Lanni C, Ganju J, Ballow CH, Kittal MM. In healthy subjects, TD-5108, a selective high intrinsic activity 5-HT₄ receptor agonist, shows dose-proportional pharmacokinetics and exhibits a profile consistent with once-daily dosing. *Gastroenterology* 2007; **132**: A374
 - 145 **Goldberg MR**, Wong SL, Ganju J, Li YP, Ballow CH, Kitt MM. TD-5108, a selective 5-HT₄ agonist with high intrinsic activity, shows immediate and sustained prokinetic activity in healthy subjects. *Gastroenterology* 2007; **132**: A60
 - 146 **Nee J**, Feuerstein JD. Review: Prucalopride, velusetrag, bisacodyl, and sodium picosulfate improve chronic idiopathic constipation. *Ann Intern Med* 2016; **165**: JC41 [PMID: 27750301 DOI: 10.7326/

- ACPJC-2016-165-8-041]
- 147 **Goldberg MR**, Li YP, Pitzer K, Johanson JF, Mangel AW, Kitt MM. TD-5108, a selective 5-HT₄ agonist, is consistently better than placebo regardless of response definition in patients with chronic constipation. *Gastroenterology* 2008; **134**: A545
 - 148 **Beattie DT**, Zamora F, Armstrong SR, Pulido-Rios T, Humphrey PP. Tegaserod, but not TD-5108, has effects in porcine and canine isolated coronary arteries. Proceedings of the British Pharmacological Society 2007, Dec; Brighton, UK. org/abstracts/Vol5Issue2abst138P.pdfS
 - 149 **Dennis D**, Palme M, Irwin I, Druzgala P, Teichman S. ATI-7505 is a novel, selective 5-HT₄ receptor agonist that causes gastrointestinal prokinetic activity in dogs. *Gastroenterology* 2004; **126** Suppl2: A641
 - 150 **Camilleri M**, Vazquez-Roque MI, Burton D, Ford T, McKinzie S, Zinsmeister AR, Druzgala P. Pharmacodynamic effects of a novel prokinetic 5-HT receptor agonist, ATI-7505, in humans. *Neurogastroenterol Motil* 2007; **19**: 30-38 [PMID: 17187586]
 - 151 **Shin A**, Camilleri M, Kolar G, Erwin P, West CP, Murad MH. Systematic review with meta-analysis: highly selective 5-HT₄ agonists (prucalopride, velusetrag or naronapride) in chronic constipation. *Aliment Pharmacol Ther* 2014; **39**: 239-253 [PMID: 24308797 DOI: 10.1111/apt.12571]
 - 152 **Palme M**, Milner PG, Ellis DJ, Marmon T, Canafax DM. A novel gastrointestinal prokinetic, ATI-7505, increased spontaneous bowel movements (SBMs) in a phase II, randomized, placebo-controlled study of patients with chronic idiopathic constipation (CIC). *Gastroenterology* 2010; **138**: S128-S129
 - 153 **Rao AS**, Wong BS, Camilleri M, Odunsi-Shiyanbade ST, McKinzie S, Ryks M, Burton D, Carlson P, Lamsam J, Singh R, Zinsmeister AR. Chenodeoxycholate in females with irritable bowel syndrome-constipation: a pharmacodynamic and pharmacogenetic analysis. *Gastroenterology* 2010; **139**: 1549-1558, 1558.e1 [PMID: 20691689 DOI: 10.1053/j.gastro.2010.07.052]
 - 154 **Bazzoli F**, Malavolti M, Petronelli A, Barbara L, Roda E. Treatment of constipation with chenodeoxycholic acid. *J Int Med Res* 1983; **11**: 120-123 [PMID: 6852359]
 - 155 **Odunsi-Shiyanbade ST**, Camilleri M, McKinzie S, Burton D, Carlson P, Busciglio IA, Lamsam J, Singh R, Zinsmeister AR. Effects of chenodeoxycholate and a bile acid sequestrant, colestevlam, on intestinal transit and bowel function. *Clin Gastroenterol Hepatol* 2010; **8**: 159-165 [PMID: 19879973 DOI: 10.1016/j.cgh.2009.10.020]
 - 156 **Bellini M**, Gambaccini D, Salvadori S, Tosetti C, Urbano MT, Costa F, Monicelli P, Mumolo MG, Ricchiuti A, De Bortoli N, Marchi S. Management of chronic constipation in general practice. *Tech Coloproctol* 2014; **18**: 543-549 [PMID: 24272606 DOI: 10.1007/s10151-013-1093-9]
 - 157 **Chey WD**, Camilleri M, Chang L, Rikner L, Graffner H. A randomized placebo-controlled phase IIb trial of A3309, a bile acid transporter inhibitor, for chronic idiopathic constipation. *Am J Gastroenterol* 2011; **106**: 1803-1812 [PMID: 21606974 DOI: 10.1038/ajg.2011.162]
 - 158 **Acosta A**, Camilleri M. Elobixibat and its potential role in chronic idiopathic constipation. *Therap Adv Gastroenterol* 2014; **7**: 167-175 [PMID: 25057297 DOI: 10.1177/1756283X14528269]
 - 159 **Wong BS**, Camilleri M. Elobixibat for the treatment of constipation. *Expert Opin Investig Drugs* 2013; **22**: 277-284 [PMID: 23215781 DOI: 10.1517/13543784.2013.753056]
 - 160 **Maneerattanaporn M**, Chey WD. Targeting bile acids in the treatment of constipation. *Expert Rev Gastroenterol Hepatol* 2011; **5**: 657-659 [PMID: 22017692 DOI: 10.1586/egh.11.63]
 - 161 **Wong BS**, Camilleri M, McKinzie S, Burton D, Graffner H, Zinsmeister AR. Effects of A3309, an ileal bile acid transporter inhibitor, on colonic transit and symptoms in females with functional constipation. *Am J Gastroenterol* 2011; **106**: 2154-2164 [PMID: 21876564 DOI: 10.1038/ajg.2011.285]
 - 162 **Simrén M**, Bajor A, Gillberg PG, Rudling M, Abrahamsson H. Randomised clinical trial: The ileal bile acid transporter inhibitor A3309 vs. placebo in patients with chronic idiopathic constipation - a double-blind study. *Aliment Pharmacol Ther* 2011; **34**: 41-50 [PMID: 21545606 DOI: 10.1111/j.1365-2036.2011.04675.x]
 - 163 **Sakamoto S**, Kusuhara H, Miyata K, Shimaoka H, Kanazu T, Matsuo Y, Nomura K, Okamura N, Hara S, Horie K, Baba T, Sugiyama Y. Glucuronidation converting methyl 1-(3,4-dimethoxyphenyl)-3-(3-ethylvaleryl)-4-hydroxy-6,7,8-trimethoxy-2-naphthoate (S-8921) to a potent apical sodium-dependent bile acid transporter inhibitor, resulting in a hypocholesterolemic action. *J Pharmacol Exp Ther* 2007; **322**: 610-618 [PMID: 17470645]
 - 164 **Li H**, Xu G, Shang Q, Pan L, Shefer S, Batta AK, Bollineni J, Tint GS, Keller BT, Salen G. Inhibition of ileal bile acid transport lowers plasma cholesterol levels by inactivating hepatic farnesoid X receptor and stimulating cholesterol 7 α -hydroxylase. *Metabolism* 2004; **53**: 927-932 [PMID: 15254889]
 - 165 **West KL**, Zern TL, Butteiger DN, Keller BT, Fernandez ML. SC-435, an ileal apical sodium co-dependent bile acid transporter (ASBT) inhibitor lowers plasma cholesterol and reduces atherosclerosis in guinea pigs. *Atherosclerosis* 2003; **171**: 201-210 [PMID: 14644388]
 - 166 **Cuppoletti J**, Malinowska DH, Tewari KP, Li QJ, Sherry AM, Patchen ML, Ueno R. SPI-0211 activates T84 cell chloride transport and recombinant human CIC-2 chloride currents. *Am J Physiol Cell Physiol* 2004; **287**: C1173-C1183 [PMID: 15213059]
 - 167 **Lacy BE**, Levy LC. Lubiprostone: a chloride channel activator. *J Clin Gastroenterol* 2007; **41**: 345-351 [PMID: 17413599]
 - 168 **Ao M**, Venkatasubramanian J, Boonkaewwan C, Ganesan N, Syed A, Benya RV, Rao MC. Lubiprostone activates Cl⁻ secretion via cAMP signaling and increases membrane CFTR in the human colon carcinoma cell line, T84. *Dig Dis Sci* 2011; **56**: 339-351 [PMID: 21140215 DOI: 10.1007/s10620-010-1495-8]
 - 169 **Bassil AK**, Borman RA, Jarvie EM, McArthur-Wilson RJ, Thangiah R, Sung EZ, Lee K, Sanger GJ. Activation of prostaglandin EP receptors by lubiprostone in rat and human stomach and colon. *Br J Pharmacol* 2008; **154**: 126-135 [PMID: 18332851 DOI: 10.1038/bjp.2008.84]
 - 170 **Musch MW**, Wang Y, Claud EC, Chang EB. Lubiprostone decreases mouse colonic inner mucus layer thickness and alters intestinal microbiota. *Dig Dis Sci* 2013; **58**: 668-677 [PMID: 23329012 DOI: 10.1007/s10620-012-2509-5]
 - 171 **Whitehead WE**, Palsson OS, Gangarosa L, Turner M, Tucker J. Lubiprostone does not influence visceral pain thresholds in patients with irritable bowel syndrome. *Neurogastroenterol Motil* 2011; **23**: 944-e400 [PMID: 21914041 DOI: 10.1111/j.1365-2982.2011.01776.x]
 - 172 **Raschi E**, De Ponti F. Lubiprostone: pharmacokinetic, pharmacodynamic, safety and regulatory aspects in the treatment of constipation-predominant irritable bowel syndrome. *Expert Opin Drug Metab Toxicol* 2014; **10**: 293-305 [PMID: 24387275 DOI: 10.1517/17425255.2013.876410]
 - 173 **Drossman DA**, Chey WD, Johanson JF, Fass R, Scott C, Panas R, Ueno R. Clinical trial: lubiprostone in patients with constipation-associated irritable bowel syndrome--results of two randomized, placebo-controlled studies. *Aliment Pharmacol Ther* 2009; **29**: 329-341 [PMID: 19006537 DOI: 10.1111/j.1365-2036.2008.03881.x]
 - 174 **Chey WD**, Drossman DA, Johanson JF, Scott C, Panas RM, Ueno R. Safety and patient outcomes with lubiprostone for up to 52 weeks in patients with irritable bowel syndrome with constipation. *Aliment Pharmacol Ther* 2012; **35**: 587-599 [PMID: 22251419 DOI: 10.1111/j.1365-2036.2011.04983.x]
 - 175 **Johanson JF**, Morton D, Geenen J, Ueno R. Multicenter, 4-week, double-blind, randomized, placebo-controlled trial of lubiprostone, a locally-acting type-2 chloride channel activator, in patients with chronic constipation. *Am J Gastroenterol* 2008; **103**: 170-177 [PMID: 17916109]
 - 176 **Barish CF**, Drossman D, Johanson JF, Ueno R. Efficacy and safety of lubiprostone in patients with chronic constipation. *Dig Dis Sci* 2010; **55**: 1090-1097 [PMID: 20012484 DOI: 10.1007/s10620-009-1068-x]
 - 177 **Spierings EL**, Rauck R, Brewer R, Marcuard S, Vallejo R. Long-Term Safety and Efficacy of Lubiprostone in Opioid-induced Constipation in Patients with Chronic Noncancer Pain. *Pain*

- Pract* 2015; Epub ahead of print [PMID: 26328775 DOI: 10.1111/papr.12347]
- 178 **Takeda**. Amitiza (lubiprostone) package insert. Deerfield, IL; 2013
 - 179 **Hyman PE**, Di Lorenzo C, Prestridge LL, Youssef NN, Ueno R. Lubiprostone for the treatment of functional constipation in children. *J Pediatr Gastroenterol Nutr* 2014; **58**: 283-291 [PMID: 24048162 DOI: 10.1097/MPG.0000000000000176]
 - 180 **Bell N**, Carreras C, Charmot D, Chen T, Leadbetter M, Jacobs J, Lewis J. Compounds and Methods for Inhibiting NHE-Mediated Antiport in the treatment of Disorders Associated with Fluid Retention or Salt Overload and Gastrointestinal Tract Disorders. World Intellectual Property Organization; WO2014029984; 2014
 - 181 **Charmot D**. Non-systemic drugs: a critical review. *Curr Pharm Des* 2012; **18**: 1434-1445 [PMID: 22300258]
 - 182 **Spencer AG**, Jacobs JW, Leadbetter MR, Carreras CW, Du X, Bell N, Koo-McCoy S, Kohler JN, Labonté E, Rosenbaum DP, Navre M, Charmot D. RDX5791, a First-in-Class Minimally Systemic NHE3 Inhibitor in Clinical Development for CIC and IBSC, Increases Intestinal Sodium Leading to Enhanced Intestinal Fluid Volume and Transit. Proceedings of the Drug Disease Week 2011, Chicago. *Gastroenterology* 2011; **140**: S-99
 - 183 **Spencer AG**, Labonté ED, Rosenbaum DP, Plato CF, Carreras CW, Leadbetter MR, Kozuka K, Kohler J, Koo-McCoy S, He L, Bell N, Tabora J, Joly KM, Navre M, Jacobs JW, Charmot D. Intestinal inhibition of the Na⁺/H⁺ exchanger 3 prevents cardiorenal damage in rats and inhibits Na⁺ uptake in humans. *Sci Transl Med* 2014; **6**: 227ra36 [PMID: 24622516 DOI: 10.1126/scitranslmed.3007790]
 - 184 **Eutamene E**, Charmot D, Navre M, Bueno L. Visceral Antinociceptive Effects of RDX5791, a First-in-Class Minimally Systemic NHE3 Inhibitor on Stress-Induced Colorectal Hypersensitivity to Distension in Rats. Proceedings of the DDW meeting 2011, Chicago IL. *Gastroenterology*; 2015; **140**: S-57-8
 - 185 **Rosenbaum DP**, Spencer AG, Jacobs J, Charmot D. The safety, tolerability, systemic exposure, and effect on bowel habits of single and multiple doses of the intestinal sodium re-uptake inhibitor RDX5791 in Normal Healthy Volunteers. Proceeding of the ACG meeting 2011, Washington DC. *Am J Gastroenterol* 2011; **106**: S504
 - 186 **Rosenbaum DP**. Safety, tolerability, pharmacokinetics and pharmacodynamic of AZD1722 in healthy male and female Japanese subjects. ClinicalTrials.gov Identifier: NCT02176252. 2011, 2015
 - 187 **Rosenbaum DP**. The efficacy of AZD1722 in Constipation Predominant Irritable Bowel Syndrome (IBS-C). ClinicalTrials.gov Identifier: NCT01923428. 2014
 - 188 **Song GH**, Leng PH, Gwee KA, Mochhala SM, Ho KY. Melatonin improves abdominal pain in irritable bowel syndrome patients who have sleep disturbances: a randomised, double blind, placebo controlled study. *Gut* 2005; **54**: 1402-1407 [PMID: 15914575]
 - 189 **Lu WZ**, Song GH, Gwee KA, Ho KY. The effects of melatonin on colonic transit time in normal controls and IBS patients. *Dig Dis Sci* 2009; **54**: 1087-1093 [PMID: 18720001 DOI: 10.1007/s10620-008-0463-z]
 - 190 **Lembo A**, Camilleri M. Chronic constipation. *N Engl J Med* 2003; **349**: 1360-1368 [PMID: 14523145]
 - 191 **Isolauri E**, Kalliomäki M, Laitinen K, Salminen S. Modulation of the maturing gut barrier and microbiota: a novel target in allergic disease. *Curr Pharm Des* 2008; **14**: 1368-1375 [PMID: 18537659]
 - 192 **Brzozowski T**, Konturek PC, Konturek SJ, Pajdo R, Bielanski W, Brzozowska I, Stachura J, Hahn EG. The role of melatonin and L-tryptophan in prevention of acute gastric lesions induced by stress, ethanol, ischemia, and aspirin. *J Pineal Res* 1997; **23**: 79-89 [PMID: 9392446]
 - 193 **Guerrero JM**, Reiter RJ. Melatonin-immune system relationships. *Curr Top Med Chem* 2002; **2**: 167-179 [PMID: 11899099]
 - 194 **Radwan P**, Skrzydło-Radomska B, Radwan-Kwiatk K, Burak-Czapiuk B, Strzemecka J. Is melatonin involved in the irritable bowel syndrome? *J Physiol Pharmacol* 2009; **60** Suppl 3: 67-70 [PMID: 19996484]
 - 195 **Bultman SJ**. Emerging roles of the microbiome in cancer. *Carcinogenesis* 2014; **35**: 249-255 [PMID: 24302613 DOI: 10.1093/carcin/bgt392]
 - 196 **Lu WZ**, Ho KY. Irritable bowel syndrome patients have decreased salivary melatonin and urine 6-hydroxymelatonin levels compared with healthy controls. *Gut* 2003; **52**: 821
 - 197 **Sun SX**, Dibonaventura M, Purayidathil FW, Wagner JS, Dabbous O, Mody R. Impact of chronic constipation on health-related quality of life, work productivity, and healthcare resource use: an analysis of the National Health and Wellness Survey. *Dig Dis Sci* 2011; **56**: 2688-2695 [PMID: 21380761 DOI: 10.1007/s10620-011-1639-5]
 - 198 **Chey WD**. Tegaserod and other serotonergic agents: what is the evidence? *Rev Gastroenterol Disord* 2003; **3** Suppl 2: S35-S40 [PMID: 12776001]
 - 199 **Thompson WG**, Longstreth GF, Drossman DA, Heaton KW, Irvine EJ, Müller-Lissner SA. Functional bowel disorders and functional abdominal pain. *Gut* 1999; **45** Suppl 2: II43-II47 [PMID: 10457044]
 - 200 **Song KH**. Practical Methods to Assess Chronic Constipation. *J Neurogastroenterol Motil* 2015; **21**: 307-308 [PMID: 26130627 DOI: 10.5056/jnm15100]
 - 201 **Houghton LA**, Heyman DJ, Whorwell PJ. Symptomatology, quality of life and economic features of irritable bowel syndrome-the effect of hypnotherapy. *Aliment Pharmacol Ther* 1996; **10**: 91-95 [PMID: 8871448]
 - 202 **Camilleri M**, Drossman DA, Becker G, Webster LR, Davies AN, Mawe GM. Emerging treatments in neurogastroenterology: a multidisciplinary working group consensus statement on opioid-induced constipation. *Neurogastroenterol Motil* 2014; **26**: 1386-1395 [PMID: 25164154 DOI: 10.1111/nmo.12417]
 - 203 **Nelson AD**, Camilleri M. Chronic opioid induced constipation in patients with nonmalignant pain: challenges and opportunities. *Therap Adv Gastroenterol* 2015; **8**: 206-220 [PMID: 26136838 DOI: 10.1177/1756283X15578608]
 - 204 **Thor PJ**, Krolczyk G, Gil K, Zurowski D, Nowak L. Melatonin and serotonin effects on gastrointestinal motility. *J Physiol Pharmacol* 2007; **58** Suppl 6: 97-103 [PMID: 18212403]
 - 205 **Bubenik GA**. Thirty four years since the discovery of gastrointestinal melatonin. *J Physiol Pharmacol* 2008; **59** Suppl 2: 33-51 [PMID: 18812627]
 - 206 **Furukawa Y**, Shiga Y, Hanyu N, Hashimoto Y, Mukai H, Nishikawa K. [Effect of Chinese herbal medicine on gastrointestinal motility and bowel obstruction]. *Jpn J Gastroenterol Surg* 1995; **28**: 956-960
 - 207 **Itoh T**, Yamakawa J, Mai M, Yamaguchi N, Kanda T. The effect of the herbal medicine dai-kenchu-to on post-operative ileus. *J Int Med Res* 2002; **30**: 428-432 [PMID: 12235926]
 - 208 **Takeda T**, Kamiura S, Kimura T. Effectiveness of the herbal medicine daikenchuto for radiation-induced enteritis. *J Altern Complement Med* 2008; **14**: 753-755 [PMID: 18637762 DOI: 10.1089/acm.2007.0748]
 - 209 **Endo S**, Nishida T, Nishikawa K, Nakajima K, Hasegawa J, Kitagawa T, Ito T, Matsuda H. Dai-kenchu-to, a Chinese herbal medicine, improves stasis of patients with total gastrectomy and jejunal pouch interposition. *Am J Surg* 2006; **192**: 9-13 [PMID: 16769267]
 - 210 **Iturrino J**, Camilleri M, Wong BS, Linker Nord SJ, Burton D, Zinsmeister AR. Randomised clinical trial: the effects of daikenchuto, TU-100, on gastrointestinal and colonic transit, anorectal and bowel function in female patients with functional constipation. *Aliment Pharmacol Ther* 2013; **37**: 776-785 [PMID: 23451764 DOI: 10.1111/apt.12264]
 - 211 **Manabe N**, Camilleri M, Rao A, Wong BS, Burton D, Busciglio I, Zinsmeister AR, Haruma K. Effect of daikenchuto (TU-100) on gastrointestinal and colonic transit in humans. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G970-G975 [PMID: 20378829 DOI: 10.1152/ajpgi.00043.2010]
 - 212 **Acosta A**, Camilleri M, Linker-Nord S, Busciglio I, Iturrino J, Szarka LA, Zinsmeister AR. A Pilot Study of the Effect of Daikenchuto on Rectal Sensation in Patients with Irritable Bowel

- Syndrome. *J Neurogastroenterol Motil* 2016; **22**: 69-77 [PMID: 26486374 DOI: 10.5056/jnm15120]
- 213 **Lee MJ**, Cho KH, Park HM, Sung HJ, Choi S, Im W. Pharmacological profile of DA-6886, a novel 5-HT₄ receptor agonist to accelerate colonic motor activity in mice. *Eur J Pharmacol* 2014; **735**: 115-122 [PMID: 24769304 DOI: 10.1016/j.ejphar.2014.03.061]
 - 214 Available from: URL: <https://clinicaltrials.gov/ct2/show/NCT01633723>
 - 215 **Gershon MD**. The enteric nervous system: a second brain. *Hosp Pract* (1995) 1999; **34**: 31-32, 35-38, 41-2 passim [PMID: 10418549]
 - 216 **Cooke HJ**. Neurotransmitters in neuronal reflexes regulating intestinal secretion. *Ann N Y Acad Sci* 2000; **915**: 77-80 [PMID: 11193603]
 - 217 **Crowell MD**. Role of serotonin in the pathophysiology of the irritable bowel syndrome. *Br J Pharmacol* 2004; **141**: 1285-1293 [PMID: 15100164]
 - 218 **Miyata K**, Ito H, Fukudo S. Involvement of the 5-HT₃ receptor in CRH-induce defecation in rats. *Am J Physiol* 1998; **274**: G827-G831 [PMID: 9612262]
 - 219 **Funatsu T**, Takeuchi A, Hirata T, Keto Y, Akuzawa S, Sasamata M. Effect of ramosetron on conditioned emotional stress-induced colonic dysfunction as a model of irritable bowel syndrome in rats. *Eur J Pharmacol* 2007; **573**: 190-195 [PMID: 17658508]
 - 220 **Camilleri M**, Northcutt AR, Kong S, Dukes GE, McSorley D, Mangel AW. Efficacy and safety of alosetron in women with irritable bowel syndrome: a randomised, placebo-controlled trial. *Lancet* 2000; **355**: 1035-1040 [PMID: 10744088]
 - 221 **Lee KJ**, Kim NY, Kwon JK, Huh KC, Lee OY, Lee JS, Choi SC, Sohn CI, Myung SJ, Park HJ, Choi MK, Bak YT, Rhee PL. Efficacy of ramosetron in the treatment of male patients with irritable bowel syndrome with diarrhea: a multicenter, randomized clinical trial, compared with mebeverine. *Neurogastroenterol Motil* 2011; **23**: 1098-1104 [PMID: 21920001 DOI: 10.1111/j.1365-2982.2011.01771.x]
 - 222 **Fukudo S**, Ida M, Akiho H, Nakashima Y, Matsueda K. Effect of ramosetron on stool consistency in male patients with irritable bowel syndrome with diarrhea. *Clin Gastroenterol Hepatol* 2014; **12**: 953-959.e4 [PMID: 24315882 DOI: 10.1016/j.cgh.2013.11.024]
 - 223 **Fukudo S**, Kinoshita Y, Okumura T, Ida M, Akiho H, Nakashima Y, Nishida A, Haruma K. Ramosetron Reduces Symptoms of Irritable Bowel Syndrome With Diarrhea and Improves Quality of Life in Women. *Gastroenterology* 2016; **150**: 358-366.e8 [PMID: 26551550 DOI: 10.1053/j.gastro.2015.10.047]
 - 224 **Fukudo S**, Kinoshita Y, Okumura T, Ida M, Hayashi K, Akiho H, Nakashima Y, Haruma K. Effect of ramosetron in female patients with irritable bowel syndrome with diarrhea: a phase III long-term study. *J Gastroenterol* 2016; **51**: 874-882 [PMID: 26800997 DOI: 10.1007/s00535-016-1165-5]
 - 225 **Gupta N**, Garg SK, Gupta R, Mahajan S, Sule S. Safety and Efficacy of Ramosetron in Men and Women With IBS-D: Systematic Review and Meta-Analysis. *Gastroenterology* 2016; **150**: Pages S1-S1271
 - 226 Serotonin now: clinical implications of inhibiting its synthesis with para-chlorophenylalanine. *Ann Intern Med* 1970; **73**: 607-630 [PMID: 4319081]
 - 227 **Camilleri M**, Bueno L, Andresen V, De Ponti F, Choi MG, Lembo A. Pharmacological, Pharmacokinetic, and Pharmacogenomic Aspects of Functional Gastrointestinal Disorders. *Gastroenterology* 2016; Epub ahead of print [PMID: 27144621 DOI: 10.1053/j.gastro.2016.02.029]
 - 228 **Camilleri M**. LX-1031, a tryptophan 5-hydroxylase inhibitor, and its potential in chronic diarrhea associated with increased serotonin. *Neurogastroenterol Motil* 2011; **23**: 193-200 [PMID: 21159063 DOI: 10.1111/j.1365-2982.2010.01643.x]
 - 229 **Brown PM**, Drossman DA, Wood AJ, Cline GA, Frazier KS, Jackson JI, Bronner J, Freiman J, Zambrowicz B, Sands A, Gershon MD. The tryptophan hydroxylase inhibitor LX1031 shows clinical benefit in patients with nonconstipating irritable bowel syndrome. *Gastroenterology* 2011; **141**: 507-516 [PMID: 21684281 DOI: 10.1053/j.gastro.2011.05.005]
 - 230 **Lembo A**, Huber J, Schinagl RM, Waters SJ, Harris MS. *Gastroenterology* 2015; **148**: S-69
 - 231 **Trinkley KE**, Nahata MC. Medication management of irritable bowel syndrome. *Digestion* 2014; **89**: 253-267 [PMID: 24992947 DOI: 10.1159/000362405]
 - 232 Lazard Capital Markets Annual Healthcare Conference. Furiex Pharmaceuticals. Available from: URL: [http://files.shareholder.com/downloads/ABEA-4H9PM3/0x0x550458/a546b8d0-d614-4136-9514-3c960c74649f/Lazard Capital Markets Annual Healthcare Conference Presentation](http://files.shareholder.com/downloads/ABEA-4H9PM3/0x0x550458/a546b8d0-d614-4136-9514-3c960c74649f/Lazard_Capital_Markets_Annual_Healthcare_Conference_Presentation). Accessed November 11, 2013.
 - 233 **Hellström PM**, Hein J, Bytzer P, Björnsson E, Kristensen J, Schambye H. Clinical trial: the glucagon-like peptide-1 analogue ROSE-010 for management of acute pain in patients with irritable bowel syndrome: a randomized, placebo-controlled, double-blind study. *Aliment Pharmacol Ther* 2009; **29**: 198-206 [PMID: 18945254 DOI: 10.1111/j.1365-2036.2008.03870.x]
 - 234 **Li ZY**, Zhang N, Wen S, Zhang J, Sun XL, Fan XM, Sun YH. Decreased glucagon-like peptide-1 correlates with abdominal pain in patients with constipation-predominant irritable bowel syndrome. *Clin Res Hepatol Gastroenterol* 2017; Epub ahead of print [PMID: 28215540 DOI: 10.1016/j.clinre.2016.12.007]
 - 235 **Konishi K**, Nakano S, Tsuda S, Nakagawa A, Kigoshi T, Koya D. AST-120 (Kremezin) initiated in early stage chronic kidney disease stunts the progression of renal dysfunction in type 2 diabetic subjects. *Diabetes Res Clin Pract* 2008; **81**: 310-315 [PMID: 18550198 DOI: 10.1016/j.diabres.2008.04.024]
 - 236 **Mosińska P**, Storr M, Fichna J. The role of AST-120 and protein-bound uremic toxins in irritable bowel syndrome: a therapeutic perspective. *Therap Adv Gastroenterol* 2015; **8**: 278-284 [PMID: 26327918 DOI: 10.1177/1756283X15587866]
 - 237 **Tack JF**, Miner PB Jr, Fischer L, Harris MS. Randomised clinical trial: the safety and efficacy of AST-120 in non-constipating irritable bowel syndrome - a double-blind, placebo-controlled study. *Aliment Pharmacol Ther* 2011; **34**: 868-877 [PMID: 21883322 DOI: 10.1111/j.1365-2036.2011.04818.x]
 - 238 **Corsetti M**, Akyuz F, Tack J. Targeting tachykinin receptors for the treatment of functional gastrointestinal disorders with a focus on irritable bowel syndrome. *Neurogastroenterol Motil* 2015; **27**: 1354-1370 [PMID: 26088804 DOI: 10.1111/nmo.12616]
 - 239 Available from: URL: <https://clinicaltrials.gov/ct2/show/NCT00761007>
 - 240 **Camilleri M**, Boeckstaens G. Dietary and pharmacological treatment of abdominal pain in IBS. *Gut* 2017; **66**: 966-974 [PMID: 28232472 DOI: 10.1136/gutjnl-2016-313425]
 - 241 **Mangel AW**, Hicks GA. Asimadoline and its potential for the treatment of diarrhea-predominant irritable bowel syndrome: a review. *Clin Exp Gastroenterol* 2012; **5**: 1-10 [PMID: 22346361 DOI: 10.2147/CEG.S23274]
 - 242 **Camilleri M**. Novel pharmacology: asimadoline, a kappa-opioid agonist, and visceral sensation. *Neurogastroenterol Motil* 2008; **20**: 971-979 [PMID: 18715494 DOI: 10.1111/j.1365-2982.2008.01183.x]
 - 243 **Mangel AW**, Bornstein JD, Hamm LR, Buda J, Wang J, Irish W, Urso D. Clinical trial: asimadoline in the treatment of patients with irritable bowel syndrome. *Aliment Pharmacol Ther* 2008; **28**: 239-249 [PMID: 18466359 DOI: 10.1111/j.1365-2036.2008.03730.x]
 - 244 **Foxx-Orenstein AE**. New and emerging therapies for the treatment of irritable bowel syndrome: an update for gastroenterologists. *Therap Adv Gastroenterol* 2016; **9**: 354-375 [PMID: 27134665 DOI: 10.1177/1756283X16633050]
 - 245 **Mottacki N**, Simrén M, Bajor A. Review article: bile acid diarrhoea - pathogenesis, diagnosis and management. *Aliment Pharmacol Ther* 2016; **43**: 884-898 [PMID: 26913381 DOI: 10.1111/apt.13570]
 - 246 **Hofmann AF**. The continuing importance of bile acids in liver and intestinal disease. *Arch Intern Med* 1999; **159**: 2647-2658 [PMID: 10597755]
 - 247 **van Tilburg AJ**, de Rooij FW, van Blankenstein M, van den

- Berg JW, Bosman-Jacobs EP. Na dependent bile acid transport in the ileum: the balance between diarrhea and constipation. *Gastroenterology* 1990; **98**: 25-32 [PMID: 2293590]
- 248 **Camilleri M**, Acosta A, Busciglio I, Boldingh A, Dyer RB, Zinsmeister AR, Lueke A, Gray A, Donato LJ. Effect of colesvelam on faecal bile acids and bowel functions in diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 2015; **41**: 438-448 [PMID: 25594801 DOI: 10.1111/apt.13065]
- 249 **Fukushima Y**, Suzuki H, Matsuzaki J, Kiyosue A, Hibi T. Efficacy of Solifenacin on Irritable Bowel Syndrome With Diarrhea: Open-label Prospective Pilot Trial. *J Neurogastroenterol Motil* 2012; **18**: 317-323 [PMID: 22837880 DOI: 10.5056/jnm.2012.18.3.317]
- 250 **Matsuzaki J**, Suzuki H, Fukushima Y, Hirata K, Fukuhara S, Okada S, Hibi T. High frequency of overlap between functional dyspepsia and overactive bladder. *Neurogastroenterol Motil* 2012; **24**: 821-827 [PMID: 22616664 DOI: 10.1111/j.1365-2982.2012.01939.x]
- 251 **Lee KN**, Lee OY, Choi MG, Sohn CI, Huh KC, Park KS, Kwon JG, Kim N, Rhee PL, Myung SJ, Lee JS, Lee KJ, Park H, Lee YC, Choi SC, Jung HK, Jee SR, Choi CH, Kim GH, Park MI, Sung IK. Efficacy and Safety of Tiropramide in the Treatment of Patients With Irritable Bowel Syndrome: A Multicenter, Randomized, Double-blind, Non-inferiority Trial, Compared With Octylonium. *J Neurogastroenterol Motil* 2014; **20**: 113-121 [PMID: 24466452 DOI: 10.5056/jnm.2014.20.1.113]
- 252 **Takayanagi I**, Hisayama T, Iwase M, Sakuma N, Nagai H. Pharmacological properties of tiropamide, an antispasmodic drug. *Gen Pharmacol* 1989; **20**: 335-339 [PMID: 2744399]
- 253 **Urano T**, Shirane M, Wada K, Tsunematsu R, Nagahamaya K, Matsuoka Y, Sunagane N, Kubota K. Possible mechanisms of action of the antispasmodic agent tiropamide in the isolated detrusor from rats. *Jpn J Pharmacol* 1992; **60**: 275-280 [PMID: 1337131]
- 254 **Park SH**, Jang CH, Han JY, Choi MG, Choi GY, Chung IS, Chung KW, Sun HS, Kim BS. Double blind clinical trial of tiropamide in irritable bowel syndrome. *Korean J Gastroenterol* 1993; **25**: 877-883
- 255 **Sobolewska-Włodarczyk A**, Włodarczyk M, Storr M, Fichna J. Clinical potential of eluxadoline in the treatment of diarrhea-predominant irritable bowel syndrome. *Ther Clin Risk Manag* 2016; **12**: 771-775 [PMID: 27257381 DOI: 10.2147/TCRM.S83722]
- 256 FDA approves two therapies to treat IBS-D. Available from: URL: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm448328.htm>.
- 257 Actavis announces FDA acceptance for filing of NDA for eluxadoline. Available from: URL: <http://www.prnewswire.com/news-releases/actavis-announces-fda-acceptance-for-filing-of-nda-for-eluxadoline-273557591.html>.
- 258 **Dove LS**, Lembo A, Randall CW, Fogel R, Andrae D, Davenport JM, McIntyre G, Almenoff JS, Covington PS. Eluxadoline benefits patients with irritable bowel syndrome with diarrhea in a phase 2 study. *Gastroenterology* 2013; **145**: 329-338.e1 [PMID: 23583433 DOI: 10.1053/j.gastro.2013.04.006]
- 259 **Lembo AJ**, Lacy BE, Zuckerman MJ, Schey R, Dove LS, Andrae DA, Davenport JM, McIntyre G, Lopez R, Turner L, Covington PS. Eluxadoline for Irritable Bowel Syndrome with Diarrhea. *N Engl J Med* 2016; **374**: 242-253 [PMID: 26789872 DOI: 10.1056/NEJMoa1505180]
- 260 **Cash BD**, Lacy BE, Schoenfeld PS, Dove LS, Covington PS. Safety of Eluxadoline in Patients with Irritable Bowel Syndrome with Diarrhea. *Am J Gastroenterol* 2017; **112**: 365-374 [PMID: 27922029 DOI: 10.1038/ajg.2016.542]
- 261 **Ganiats TG**, Norcross WA, Halverson AL, Burford PA, Palinkas LA. Does Beano prevent gas? A double-blind crossover study of oral alpha-galactosidase to treat dietary oligosaccharide intolerance. *J Fam Pract* 1994; **39**: 441-445 [PMID: 7964541]
- 262 **Hillilä M**, Färkkilä MA, Sipponen T, Rajala J, Koskenpato J. Does oral α -galactosidase relieve irritable bowel symptoms? *Scand J Gastroenterol* 2016; **51**: 16-21 [PMID: 26133538 DOI: 10.3109/00365521.2015.1063156]
- 263 **Sinagra E**, Tomasello G, Cappello F, Leone A, Cottone M, Bellavia M, Rossi F, Facella T, Damiani P, Zeenny MN, Damiani F, Abruzzo A, Damiano G, Palumbo VD, Cocchi M, Jurjus A, Spinelli G, Lo Monte AI, Raimondo D. Probiotics, prebiotics and symbiotics in inflammatory bowel diseases: state-of-the-art and new insights. *J Biol Regul Homeost Agents* 2013; **27**: 919-933 [PMID: 24382173]

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Glucocorticosteroid therapy in inflammatory bowel diseases: From clinical practice to molecular biology

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Abstract

Inflammatory bowel diseases (IBDs), such as ulcerative colitis and Crohn's disease, are chronic pathologies associated with a deregulated immune response in the intestinal mucosa, and they are triggered by environmental factors in genetically susceptible individuals. Exogenous glucocorticoids (GCs) are widely used as anti-inflammatory therapy in IBDs. In the past, patients with moderate or severe states of inflammation received GCs as a first line therapy with an important effectiveness in terms of reduction of the disease activity and the induction of remission. However, this treatment often results in detrimental side effects. This downside drove the development of second generation GCs and more precise (non-systemic) drug-delivery methods. Recent clinical trials show that most of these new treatments have similar effectiveness to first generation GCs with fewer adverse effects. The remaining challenge in successful treatment of IBDs

concerns the refractoriness and dependency that some patients encounter during GCs treatment. A deeper understanding of the molecular mechanisms underlying GC response is key to personalizing drug choice for IBDs patients to optimize their response to treatment. In this review, we examine the clinical characteristics of treatment with GCs, followed by an in depth analysis of the proposed molecular mechanisms involved in its resistance and dependence associated with IBDs. This thorough analysis of current clinical and biomedical literature may help guide physicians in determining a course of treatment for IBDs patients and identifies important areas needing further study.

Key words: Inflammatory bowel diseases; Ulcerative colitis; Crohn's disease; Glucocorticoid dependence; Glucocorticoid resistance

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Core tip: Glucocorticoids (GCs) are widely used in patients with Inflammatory Bowel Diseases who have moderate or severe disease activity; however, some of them do not respond to treatment or become dependent. Knowledge of both the clinical approach of GCs treatment as well as the molecular basis underlying their effects will help physicians prescribe drugs that will lead to better outcomes for patients.

Dubois-Camacho K, Ottum PA, Franco-Muñoz D, De la Fuente M, Torres-Riquelme A, Díaz-Jiménez D, Olivares-Morales M, Astudillo G, Quera R, Hermoso MA. Glucocorticosteroid therapy in inflammatory bowel diseases: From clinical practice to molecular biology. *World J Gastroenterol* 2017; 23(36): 6628-6638 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6628.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6628>

INTRODUCTION

Inflammatory bowel diseases (IBDs), including ulcerative colitis (UC) and Crohn's disease (CD), are multifactorial disorders comprised of both environmental and genetic factors. Even though they are different disease entities, UC and CD are grouped under the generic term of IBDs given their similar activity/remission stages, chronicity, immunological pathophysiology and uncertain etiology^[1-3]. The prevalence of UC is 505 per 100000 people in Europe and 249 per 100000 people in North America, and CD is 322 per 100000 people in Europe and 319 per 100000 people in North America^[4]. An increased incidence has been reported in some South American and mid-eastern Asian countries such as Chile^[5], Brazil^[6], Israel^[7], and Malaysia^[8]. This tendency might be related to the industrialization phenomenon and the

influence of environmental risk factors^[4].

IBDS: A MULTIFACTORIAL COMPLEX DISORDER

Various genetic and environmental factors have been associated with development of IBDs. Polymorphisms in a vast number of genes which impair functions such as lymphocyte activation, autophagy, pathogen sensing, stress response, antigen presentation, and chemotaxis, among others have been described^[9]. The presence of such genetic variants may contribute to an imbalance in the immune response and increased predisposition to IBDs^[10,11]. Moreover, environmental factors, such as diet, smoking, appendectomy, breast-feeding behavior in childhood, vitamin D deficiency and infections, play key roles in inflammatory manifestations, and have been described as risk factors for IBDs^[12,13]. Furthermore, intestinal microbiome composition differs in IBDs patients compared to healthy subjects: IBD patients have a more predominant *Proteobacteria* population and decreased *Firmicutes* and *Bacteroidetes* populations^[14,15]. Environmental factors might be involved in creating such imbalance in microbiome composition, and this may give rise to a constant inflammatory antigenic stimulus in the gut that causes chronic inflammation in genetically susceptible individuals^[14].

These multifactorial components involved in the origin and progression of IBDs, has prevented the development of a specific treatment applicable to all patients. Current treatment options seek to diminish inflammation in order to control symptoms and keep the patient in a state of remission or symptom improvement. For example, aminosalicylates (5-asa derivatives) are used to treat mild disease, while immunosuppressants such as azathioprine/6-mercaptopurine and methotrexate, and biological therapy such as anti-TNF antibodies or anti-integrin are prescribed for moderate to severe disease^[16]. Finally, glucocorticoids (GCs) are used to achieve, but not to maintain, remission in patients with moderate to severe activity^[17].

GC THERAPY IN IBDs: ADMINISTRATION AND EFFECTIVENESS

The first study that demonstrated the usefulness of GCs (cortisone) in controlling severe UC attacks was published in 1955^[18]. Since the 70s, the first generation of GCs (prednisone, methylprednisolone, hydrocortisone) has been used to induce clinical remission in IBDs patients. However, significant adverse effects have led to the development of second generation GCs (budesonide, budesonide MMX, Bedolmethasone dipropionate)^[19-21]. A characteristic of GC treatments and their use is summarized in Table 1.

Table 1 Glucocorticoids of first and second generation and his medical uses

Glucocorticoids	Component	Indications	Ref.
1 st generation	Prednisone	Moderate to Severe cases	[3,25,27]
	Methyl-Prednisolone	of IBDs	[26]
2 nd generation	Hydrocortisone	Short duration of treatment	[29]
	Budesonide	Moderate CD cases	[32-34]
	Budesonide MMX	Mild to moderate UC cases	[36,37]
	Beclomethasone dipropionate	Topical administration	[40,41]
	Erythrocyte - Mediated Delivery of Dexamethasone	In research for long term treatments	[42]

IBDs: Inflammatory bowel diseases; CD: Crohn's disease; UC: Ulcerative colitis.

First generation GCs: more effective than 5-asa and immunomodulatory treatment with significant secondary effects

Several studies have compared the efficacy and safety of traditional GCs to other treatments by measuring the rates of remission and adverse side effects. Two independent placebo-controlled double-blind randomized trials showed that 47% of active CD patients treated with prednisone (1 mg/kg, ranging from 40-60 mg/d) achieved clinical remission, defined as CDAI score < 150 (for a review on clinical and endoscopy scoring see references^[22-24]), compared to 26% of patients receiving placebo after 4 mo of treatment. Neither sulfasalazine (0.5-1 gr/15 kg) nor azathioprine (2-2.5 mg/kg per day) achieved significant effectiveness^[25]. Later, a placebo-controlled study showed that high doses of methylprednisolone (12-48 mg/d), sulfasalazine (3 gr/d), or both in active CD patients, were significantly effective compared to the placebo, and patients treated with methylprednisolone (alone or in combination with sulfasalazine) showed faster improvement compared to sulfasalazine^[26].

Despite the effectiveness of GCs demonstrated in patients with severe CD, a prospective multicenter study comprised of patients with active CD on prednisone treatment (1 mg/kg per day) concluded that while 92% achieved clinical remission, only 29% of them reached endoscopic remission^[27]. The presence of stenosis or fistula secondary to CD which occurs in patients who do not reach endoscopic remission, indicates a poor response to GC therapy and requires surgical intervention^[27]. This suggests that GCs are efficient in improving the inflammatory response but other strategies are needed to increase mucosal healing.

Regarding UC, systemic GCs are the first choice therapy in cases of severe colitis^[3]. In acute and critical UC, 76% of the patients treated with prednisolone (20 mg/d), plus a nightly rectal drip of hydrocortisone succinate sodium (100 mg in solution), presented clinical remission vs 52% of patients who were treated with sulfasalazine (8 gr/d the first week, 4 gr/d the second week)^[28] demonstrating the clinical efficacy of this treatment. Likewise, two consecutive trials in UC patients with left side colitis determined that oral prednisone (1 mg/kg per day) was effective in severe stages of the disease compared with calcium lactate

(1-3 gr; placebo), and prednisone treatment led to clinical remission more quickly and more often than those treated with salazopyrin (4 gr/d; a sulfasalazine derivate) and topic hydrocortisone treatments (100 mg/150 mL of sterile water administered as an enema)^[29].

Despite the effectiveness of first generation GCs, they are characterized by several serious side effects that limit their long-term use. Among these, some of the most important are: metabolic (altered distribution of fat, Cushing's face or steroid-induced diabetes), eye-related (cataracts and glaucoma), dermatological (bruising and urticarial), gastrointestinal (gastric ulcer and gastrointestinal bleeding), and musculoskeletal side effects (osteopenia to osteoporosis). Additionally, adverse effects on the central nervous system have been described (insomnia, anxiety, hyperactivity, psychotic processes), along with hypertension, hypothalamic-pituitary-adrenal axis suppression and increased susceptibility to infections due to immunosuppression^[19]. However, its use in IBDs patients has not be associated with atherosclerosis progression^[30]. Finally, there is evidence that GC use increases the likelihood of sepsis of gastrointestinal origin; therefore it should be prescribed with caution in these patients^[31].

Second generation GCs: high anti-inflammatory activity and fewer side effects.

The previously described serious side-effects related to first-generation GCs drove the development of second-generation steroids, which maximize the amount of corticosteroid locally available for distal ileum and proximal colon and minimize systemic bioavailability^[3,26]. These new formulations of GCs with limited absorption include budesonide, budesonide MMX (multimatrix delivery system MMX) and beclomethasone dipropionate^[3,19].

Budesonide, similar effectiveness with fewer adverse effects: Budesonide undergoes hepatic inactivation before reaching the systemic circulation (first-pass metabolism) which reduces its bioavailability^[3]. Various placebo-controlled clinical studies have provided evidence about the efficacy of budesonide in IBDs patients. A multicenter randomized, placebo-controlled, double-blind clinical

trial was conducted in CD patients who received three different budesonide doses (3, 9 and 15 mg/d)^[32]. After 8 wk of treatment, 51% of those receiving 9 mg budesonide progressed to remission in CD patients with ileum and proximal colon disease activity. Thus, this dose was effective and safe but depended on disease location. A randomized, double-blind, double-dummy controlled trial conducted in patients with mild to moderately active CD demonstrated that budesonide (9 mg/d for 2 mo) was as effective as prednisone (40 mg/d for the first 2 wk, 30 mg/d the third week, and gradually tapered to 5 mg/wk at the end of the study lasting 2 mo) in patients with terminal ileum, cecum, and/or ascending colon inflammation^[33].

Subsequently, a multicenter double-blind, randomized trial was conducted in CD patients with moderate activity in the distal ileum and ascending colon. More patients that received budesonide (4.5 mg twice daily, or 9 mg once daily) achieved clinical remission compared to placebo 2 wk after beginning treatment, and this difference was statistically significant^[34]. In addition, a pooled analysis of trials in CD patients found significantly fewer GC-related adverse events compared to patients treated with conventional corticosteroids (RR = 0.64, 95%CI: 0.54-0.76)^[35].

An improved oral formulation of budesonide uses the colonic delivery technology Multi-Matrix System (MMX) to extend drug release in the colon (Budesonide MMX). Due to this characteristic, it is indicated for the treatment of UC patients that do not respond to standard doses of salicylates. A placebo-controlled study demonstrated that a higher proportion of UC patients with mild to moderate activity treated with budesonide MMX (9 mg), reached clinical remission and symptom resolution compared to placebo. In contrast, a lower dose of budesonide MMX (6 mg) and mesalamine, another salicylate treatment (2.4 gr/d), did not significantly improve either parameter^[36]. Furthermore, the higher dose of Budesonide MMX was efficient in inducing remission in patients with mild to moderate UC disease who did not achieve remission with 5-asa treatments such as mesalamine or sulfasalazine^[36,37]. Currently, physicians prescribe Budesonide MMX in UC patients that do not respond to traditional maintenance therapy before treating with azathioprine/6-mercaptopurin.

In 2015, data from 5 clinical studies, including double-blind, randomized, and open label studies, showed that rates of adverse effects were similar between budesonide MMX 9 mg and 6 mg (54.5% and 60.6% respectively) and placebo (50.5%) in patients with mild to moderate UC^[38]. Furthermore, the open label studies showed less frequency of adverse effects with budesonide MMX (3 mg or 9 mg) compared to placebo. The most common adverse effects in these studies were headache, nausea and urinary tract infection demonstrating that second generation GCs not only have fewer side effects, but those effects

are less severe compared to first generation GCs side effects.

Beclomethasone dipropionate, a second generation option with low systemic activity:

Beclomethasone dipropionate (BDP) is a second-generation corticosteroid with topical effects and minimal systemic activity. It is administered as a pro-drug and is partially metabolized in the lower gastrointestinal tract^[39]. The effectiveness and tolerability of BDP vs budesonide have been evaluated in CD patients whose clinical characteristics did not include complications such as stenosis or fistulas. A study showed that BDP (10 mg/d) was less effective than budesonide (9 mg/d) in CD patients treated for 2 mo. They concluded that this was due to the pharmacokinetic properties of budesonide^[40].

A study was conducted to evaluate the efficacy and safety of BDP enemas vs prednisone enemas in active distal UC after 1 mo of treatment. They determined that clinical and endoscopic remission occurred in 29% of patients who received BDP and in 25% of patients who had standard topical GC treatment, and both groups experienced few adverse effects^[41]. This evidence leads to the conclusion that BDP has similar efficacy to traditional GCs.

GC delivery technology, the next step to avoid side effects

Novel drug delivery systems capable of slowly releasing drugs into the bloodstream at a low concentration such as the erythrocyte-mediated drug delivery have been proposed. The membrane characteristics of autologous erythrocytes render them ideal drug carriers because their permeability allows for the diffusion of small molecules. This has been demonstrated using dexamethasone 21-phosphate (Dex 21-P), a biologically inactive compound^[42]. Once encapsulated, an enzyme resident in the red blood cell dephosphorylates Dex 21-P to form the corresponding active metabolite, dexamethasone (Dex), which is then released into the bloodstream by passive diffusion through the cytoplasmic membrane of erythrocytes^[42].

A study recruited 40 patients with mild to moderate UC, who had not responded to mesalamine and randomly assigned them to one of the following three treatment groups: Dex 21-P encapsulated into erythrocytes (DEE) delivered *via* two infusions 14 d apart, oral prednisolone infusions (0.5 mg/kg for 2 wk by 6 mg/wk tapering), and placebo. The group of DEE and oral prednisolone achieved a higher rate of clinical and endoscopic remission (75%) after 2 mo compared to placebo^[42]. DEE-treated patients showed no adverse effects associated with GCs, in contrast to 80% of prednisone group, in whom acne, hirsutism and weight gain were reported. This delivery tool is an attractive choice for patients with IBDs to avoid most of GCs' adverse effects.

Resistance and dependence, an unresolved issue for GC treatment

More than half of IBDs patients on GC therapy respond to treatment, approximately 28% have a partial response, and 19% are non-responders (GC-resistant). However, about 20% of IBDs patients on long-term GC therapy become dependent^[43,44]. Therefore, clarifying this GC resistance/dependency is crucial to making appropriate decisions regarding patient treatment. GC resistance or refractoriness is the inability of GCs to exert their effects on target tissues, thus limiting the efficacy of the therapy. In IBDs, GC resistance is defined as the persistence of an active manifestation of the pathology, despite having received standard treatment of 0.75 mg/kg per day for 4 or more weeks^[44].

On the other hand, GC dependency is the need for GCs to maintain remission. Patients are considered GC-dependent if they fail to taper to steroid doses below 10 mg within 4 mo (starting dose 0.75-1 mg/kg oral prednisone-equivalent) or if they relapse within 3 mo after the discontinuation of GC treatment^[44]. Usually, long-term treatment with GCs is associated with its resistance or dependency and a more aggressive clinical phenotype in IBD patients^[45].

MOLECULAR BASIS OF GLUCOCORTICOID TREATMENT

Exogenous GCs are highly lipophilic compounds, making them widely bioavailable. Similar to endogenous cortisol, GCs are primarily transported in the bloodstream bound to corticosteroid-binding globulin and, to a lesser extent, to albumin^[46]. GCs have the ability to passively diffuse through cell membranes and interact with the glucocorticoid receptor (GR), a member of the nuclear receptor superfamily of ligand-dependent transcription factors^[47].

Molecular characteristics of GR and its impact on Immune system

GR is a cytosolic protein with a molecular weight of 94 kDa encoded by the *NR3C1* gene (5q31.3). It contains nine exons that mainly code for two transcripts formed by alternative splicing: the α and β isoforms^[48]. In addition to GR β , three less-well-characterized isoforms have been reported: GR γ , GR-A and GR-P^[49]. Moreover, GR α and GR β can also undergo alternative translation initiation in exon 2, which generates eight additional GR isoforms with truncated N-terminals giving them distinct properties^[49].

The amino terminal region of GR α contains a ligand-independent transactivation domain (AF-1), a highly conserved central DNA binding domain (DBD), and a hinge segment. The carboxyl-terminal region contains the ligand-binding domain, which includes an AF-2 region that interacts with co-regulators in a ligand-dependent manner^[50]. The GR β is a shorter protein which differs from GR α in its C-terminal

domain and antagonizes the activity of GR α ^[51]. After GC binding, the receptors undergo conformational changes and expose the DBD, interact with chromatin, and regulate gene expression^[50].

The GR-GC complex can inhibit proinflammatory proteins such as nuclear factor κ B and AP-1 through protein-protein interactions. These molecular mechanisms down-regulate the expression of proinflammatory cytokines and chemokines, such as IL-1 α , IL-1 β and IL-8. In addition, GCs up-regulate the expression of other cytokines that suppress the production of inflammatory mediators: such as transforming growth factor- β 3 (TGF- β) and IL-10, increasing its anti-inflammatory function. They also inhibit T and B lymphocyte proliferation, and promote a tolerant macrophage profile (M2) (Figure 1)^[46,52,53].

Molecular basis of GC resistance, a clue for personalized treatment

The molecular mechanisms of GC resistance in IBDs have been associated with changes in the GR isoform levels, polymorphisms in *NR3C1* or genes involved in GC bioavailability, and impaired cytokine production^[46,48,54,55]. The most important genes associated to GC resistance are summarized in Table 2. Changes in levels of GR α and β isoforms have been associated with GC resistance. The α -isoform is the most well-studied protein, but only increased levels of the β -isoform have been associated to GC resistance in clinical trials in UC, asthma, nasal polyposis, cancer and chronic lymphocytic leukemia^[53]. The GR β protein is located primarily in the nucleus and is found mostly in T lymphocytes, macrophages, neutrophils, eosinophils, and peripheral blood mononuclear cells; however, it has also been reported in brain, lung, and heart tissue^[56,57]. It has been proposed that GR β may act as a dominant negative to GR α , since GR β does not bind to GCs, but rather it interacts with glucocorticoid element response (GRE) causing GC resistance. This mechanism alters GR α signaling independent of GR α -GR β heterodimer formation^[58]. An *in vitro* study showed that overexpression of GR β in a colonocyte cell model induces a vast deregulation of gene expression without Dex treatment, and a proportion of these genes have been shown to be altered in IBDs patients^[59]. Thus, GR β may directly change transcriptional gene activity of proinflammatory molecules. This effect is independent of GR α function and may promote the GC-resistance.

Furthermore, GR β can bind to RU-486, a GR α antagonist, in such a way as to change the cellular location of this GR α , and alter its ability to regulate gene expression^[60]. In addition, it has previously been shown that the expression of GR β can be transcriptionally activated in regulatory T lymphocytes and neutrophils by proinflammatory cytokines^[61]. Hence, GR β expression may affect the immune response of these cells promoting inflammation and non-response to GC^[55,57,62].

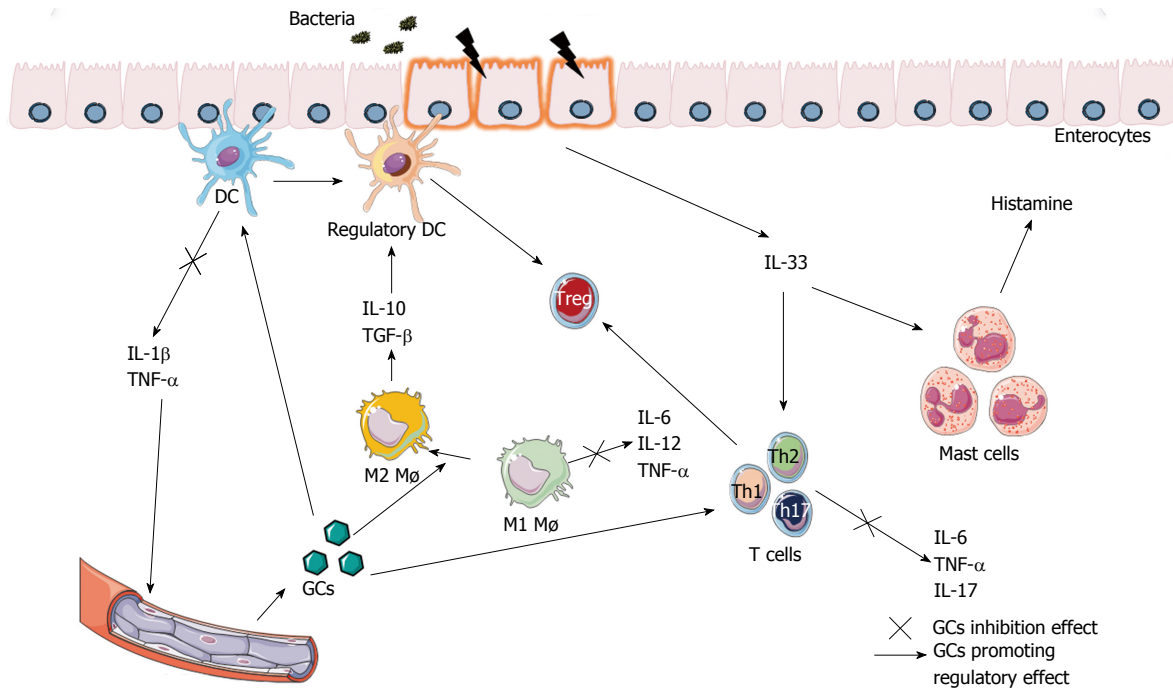


Figure 1 Immunologic dynamics and the responses triggered by glucocorticoids in inflammatory bowel diseases. During activation of the immune response in inflammatory bowel diseases (IBDs), the invasion of bacteria and the cytokine cascade triggered by the disruption of the epithelial barrier prompt an inflammatory response in the intestinal mucosa. Among the different cells that participate in the pathology of IBDs, the most important are represented in this diagram. When bacterial organisms interact with dendritic cells (DC), or the intestinal epithelium is disrupted due to an inflammatory response, the production of proinflammatory cytokines and the release of DAMPs (Damaged- Associated Molecular Patterns) such as TNF- α and IL-33, respectively, is elicited. This leads to the infiltration of immune cells such as monocytes which differentiate into proinflammatory M1 macrophages (M ϕ) and lymphocytes T (T cells), along with mast cell activation. In the presence of glucocorticoids (GCs), the infiltrating monocytes differentiate into a regulatory M2 profile M ϕ and CD103⁺ DCs. These tolerogenic cells produce anti-inflammatory cytokines, skew the infiltrating naive T cells toward a T regulatory phenotype, and control the inflammatory response. Also, the action of GCs blocks the production of proinflammatory cytokines by the M1 M ϕ , T cells and DCs, leading to the healing of the intestinal mucosa. Figure 1 was produced using Servier Medical Art from <http://smart.servier.com>. TGF- β : Transforming growth factor- β ; IL: Interleukin.

Table 2 Molecular highlights on responses to steroidal treatment

Gene/protein	Function	Molecular alteration or genetic variant	Effect on GC response	Ref.
NR3C1	GC receptor	Increased expression of the isoform rs6189 rs6190 rs6195	Block signaling of the isoform/GC resistance Altered transactivation of GC receptor/GC resistance	[58] [54,63]
MDR1	Export of drugs from the cell	rs41423247 rs1045642 rs1128503 rs1045642 rs1800896	Enhanced sensitivity to GC Alternative splicing of receptor/GC resistance Decreased MDR1 levels/GC resistance in Brazilian CD patients Possible misfolding/GC dependence in Chinese CD patients	[65] [54,63] [68] [75-77]
IL-10	Anti-inflammatory cytokine		Lowered production of IL-10/GC dependence in UC and CD patients	[45]
EBF3	Transcription factor	Decreased levels of EBF3	More severely inflamed phenotype/GC dependence	[72]
PAR2	G protein coupled receptor	Methylation of gene promoter	GC resistance/dependence in UC patients.	[82]

UC: Ulcerative colitis; CD: Crohn's disease; GCs: Glucocorticoids.

In 2009, a study was conducted to measure the expression of GR β in intestinal mucosa of IBD patients that were refractory to treatment with GCs^[55]. High levels of the GR α isoform were found in both GC-resistant and responsive patients; however, GC-resistant patients expressed higher levels of GR β , in comparison with the GC-responder group. In the intestinal mucosa, the cells that primarily expressed

the GR β receptor were CD4⁺ and CD8⁺ T lymphocytes, macrophages and B cells. Fibroblasts and vascular endothelial cells expressed GR β in a minor proportion. These results suggest that a higher ratio of GR β /GR α in inflammatory mononuclear cells disturbs the effects of GCs resulting in refractory outcomes.

Single nucleotide polymorphisms (SNPs) in *NR3C1* gene, have been associated with GC resistance^[54],

and these SNPs may result in different effects on the receptor structure and its signal transduction^[54]. The most-studied SNPs that could induce a refractory phenotype in IBDs patients are: ER22/23EK (rs6189 and rs6190), N363S (rs6195), and BcII (rs41423247)^[54]. The ER22/23EK is produced by a change in the amino acid residues 22 and 23 in the GR, where two glutamic acids (E) are replaced by an arginine (R) in position 22, and a lysine (K) in position 23^[63]. These SNPs affect the transactivation process in *in vitro* assays, where the receptor is less likely to change its conformation and induce gene transcription in the presence of GCs^[64]. The SNP N363S is in codon 363 of exon 2 and results in an amino acid change from asparagine (N) to serine (S). This SNP was reported to be associated with enhanced sensitivity to GCs, as was demonstrated by a Dex suppression test^[65]. The BcII SNP was identified as a cytosine-guanine substitution 646 nucleotides downstream from exon 2, yielding 2.2 and 3.9 kb long fragments^[63], which could suggest alternative splicing. A meta-analysis that included a mostly Caucasian population, analyzed the association of each of these 3 SNPs with GC resistance in IBDs patients^[54]. Although no association was found, the analysis of the presence of all three SNPs, (or combinations of two SNPs) as a polygenic additive effect, could show risk associations of these variants with GC resistance.

Like the GR, the P-glycoprotein (P-gp) has also been associated with GC resistance in IBDs. P-gp is a trans-membrane glycoprotein with a molecular weight of 170 kDa encoded by the *MDR1* gene, which is located on the long arm of chromosome 7. P-gp is responsible for the absorption, distribution, metabolism and excretion of various drugs including GC^[66]. A relationship between increased P-gp levels in circulating lymphocytes and intestinal epithelial cells with GC-resistant IBDs patients has been reported^[67]. Polymorphisms in the *MDR1* gene have been shown to alter P-gp function, which in turn modifies the response to GCs^[67]. A case-control study in a Brazilian population demonstrated that SNP C3435T (rs1045642), located in exon 26 of *MDR1* was associated with GC resistance in CD but not in UC patients^[68].

Additionally, higher expression of proinflammatory cytokines has been associated with GC resistance. Overexpression of IL-6 and IL-8 has been associated with non-responsiveness to GC treatment^[69]. In pediatric patients with UC, a similar correlation was found between the expression levels of IL-6 and unresponsiveness to GC^[70]. In this study, the patients who did not respond to GC had higher levels of IL-6, compared to responders. Given the pleiotropic nature of IL-6, this correlation could reflect a cause-effect relationship between the signaling of the GC receptor and the expression of proinflammatory cytokines, but the real reason for the failure to respond remains unclear^[70].

Similarly, in CD a correlation between cytokine production and resistance to GC was found. A clinical

study showed that in intestinal mucosa from GC-resistant patients, the rate of apoptosis of T and B cells, presence of caspase-3 and IL-10 production were diminished compared to healthy and GC-responsive individuals^[71]. These findings suggest a strong relationship between cytokine signaling and the inhibition of the GR effect in inflammatory diseases.

Analyses of gene expression using microarrays have been conducted to associate gene deregulation with GC resistance from colonoscopic biopsies of IBDs patients. *ATG16L1*, a gene related to the autophagy process was down-regulated in GC-resistant patients^[72]. A low expression of the ATG16L1 protein has been associated with altered secretion of mucus by Paneth cells^[73]. Furthermore, inhibition of autophagy led to Dex-resistance in a lymphoma cell line and in a mouse model^[74]. This evidence indicates the importance of autophagy in the GC-resistant phenotype.

GC dependence: another genetic issue

Contrary to GC refractoriness, the molecular mechanisms behind GC dependence are poorly studied. There is evidence that associates *MDR1* SNPs (rs1128503 and rs1045642) with GC-dependent CD patients in a Chinese population^[75]. Both SNPs correspond to a change from cytosine (C) to thymine (T) and are in coding regions of the gene (exon 12 and 26, respectively). Based on computational protein analysis, these SNPs, as haplotypes, seem to affect protein folding which could alter P-gp levels^[76], but this mechanism has not been experimentally demonstrated. Nevertheless, the rs1045642 - CC was frequently found in individuals with significantly higher P-gp levels in plasma along with increased protein activity (measured by rhodamine efflux in CD56⁺ natural killer cells), compared to carriers of the TT genotype^[77]. This study suggests that increased GC transportation out of cells might drive a decreased GC response leading to the dependent phenotype^[75].

SNPs in the *IL-10* gene have been associated with GC dependency as well. The SNP -1082 A/G (rs1800896) is located in the promoter sequence of the *IL-10* gene, and a case control study of this variant showed an association with GC dependence in UC and CD patients^[45]. This SNP was also related to a lower production of IL-10 in IBDs patients. Together, this evidence suggests that intrinsically inferior levels of IL-10 may generate GC dependency because exogenous GCs would be necessary to reach the anti-inflammatory IL-10 content necessary to achieve sustained remission. Other mechanisms, similar to those of refractoriness, such as altered expression of GR that impair intracellular signaling, have been proposed^[78].

Downregulation of EBF3 was related to GC dependence in IBD patients^[72]. The transcription factor EBF3 is a downstream target of SMAD2/3, which are components of the TGF- β signaling pathway^[79]. This anti-inflammatory mechanism might be impaired in the context of low EBF3 expression^[80]. Moreover,

EBF3 down-regulation has been associated with a low rate of apoptosis and increased cell proliferation^[81]. This evidence suggests that decreased EBF3 levels might allow for a more severely inflamed phenotype making necessary a continual GC administration to maintain remission in IBD patients and driving their dependence.

Epigenetics knowledge in GCs dependence/resistance in IBDs

Epigenetic changes may also cause GC dependence/resistance. In the context of IBDs, higher methylation of the *Protease-Activated Receptor 2 (PAR2)* gene, has been associated with steroid-dependent and resistance phenotypes in UC patients^[82]. PAR2 is a G protein coupled receptor that is involved in pro-inflammatory responses and play a role in IBDs pathogenesis^[82]. However, its specific role in GCs dependence/resistance has not been described.

Various studies relate the epigenetics changes such as hypermethylation of promoter genes with severe disease and risk of neoplasia^[83-85]. There is evidence that epigenetic shifts can occur in the early stages of embryonic development in genes known as metastable epialleles, and these events can appear, as shown in rat and murine models, by environmental factors such as stress^[86-88]. However, more analyses are needed to clarify the role of epigenetics in controlling the molecular mechanisms that underlie dependence or resistant to GCs in IBDs patients.

CONCLUSION

It has been over half a century since the first use of GC in IBD, yet amazingly, it currently remains a widely used therapeutic tool, specifically in disease exacerbations. The latest pharmacological applications, known as second-generation steroids, have contributed to the reduction of adverse reactions, however, increased effectiveness would need to be demonstrated in order for them to replace traditional GC therapy.

Despite findings showing positive GC responses, some patients remain resistant or become dependent to this drug. Deepening our understanding of the molecular basis of these undesired effects will help us generate novel, personalized GC therapies, depending on a patient's genetic background and cytokine profiles.

REFERENCES

- 1 **Ananthakrishnan AN.** Environmental risk factors for inflammatory bowel diseases: a review. *Dig Dis Sci* 2015; **60**: 290-298 [PMID: 25204669 DOI: 10.1007/s10620-014-3350-9]
- 2 **Rietdijk ST, D'Haens GR.** Recent developments in the treatment of inflammatory bowel disease. *J Dig Dis* 2013; **14**: 282-287 [PMID: 23419117 DOI: 10.1111/1751-2980.12048]
- 3 **De Cassan C, Fiorino G, Danese S.** Second-generation corticosteroids for the treatment of Crohn's disease and ulcerative colitis: more effective and less side effects? *Dig Dis* 2012; **30**: 368-375 [PMID: 22796798 DOI: 10.1159/000338128]
- 4 **Molodecky NA, Soon IS, Rabi DM, Ghali WA, Ferris M, Chernoff G, Benchimol EI, Panaccione R, Ghosh S, Barkema HW, Kaplan GG.** Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology* 2012; **142**: 46-54 [PMID: 22001864 DOI: 10.1053/j.gastro.2011.10.001]
- 5 **Simian D, Fluxá D, Flores L, Lubascher J, Ibáñez P, Figueroa C, Kronberg U, Acuña R, Moreno M, Quera R.** Inflammatory bowel disease: A descriptive study of 716 local Chilean patients. *World J Gastroenterol* 2016; **22**: 5267-5275 [PMID: 27298570 DOI: 10.3748/wjg.v22.i22.5267]
- 6 **Victoria CR, Sassak LY, Nunes HR.** Incidence and prevalence rates of inflammatory bowel diseases, in midwestern of São Paulo State, Brazil. *Arq Gastroenterol* 2009; **46**: 20-25 [PMID: 19466305 DOI: 10.1590/S0004-28032009000100009]
- 7 **Abu Freha N, Schwartz D, Elkrinawi J, Ben Yakov G, Abu Tailakh M, Munteanu D, Abu Ganim A, Fich A.** Inflammatory bowel disease among Bedouin Arabs in southern Israel: urbanization and increasing prevalence rates. *Eur J Gastroenterol Hepatol* 2015; **27**: 230-234 [PMID: 25563139 DOI: 10.1097/MEG.0000000000000263]
- 8 **Hilmi I, Jaya F, Chua A, Heng WC, Singh H, Goh KL.** A first study on the incidence and prevalence of IBD in Malaysia--results from the Kinta Valley IBD Epidemiology Study. *J Crohns Colitis* 2015; **9**: 404-409 [PMID: 25744112 DOI: 10.1093/ecco-jcc/jjv039]
- 9 **Khor B, Gardet A, Xavier RJ.** Genetics and pathogenesis of inflammatory bowel disease. *Nature* 2011; **474**: 307-317 [PMID: 21677747 DOI: 10.1038/nature10209]
- 10 **Glocker EO, Kotlarz D, Boztug K, Gertz EM, Schäffer AA, Noyan F, Perro M, Diestelhorst J, Allroth A, Murugan D, Hätscher N, Pfeifer D, Sykora KW, Sauer M, Kreipe H, Lacher M, Nustede R, Woellner C, Baumann U, Salzer U, Koletzko S, Shah N, Segal AW, Sauerbrey A, Buderus S, Snapper SB, Grimbacher B, Klein C.** Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. *N Engl J Med* 2009; **361**: 2033-2045 [PMID: 19890111 DOI: 10.1056/NEJMoa0907206]
- 11 **Di Meglio P, Di Cesare A, Laggner U, Chu CC, Napolitano L, Villanova F, Tosi I, Capon F, Trembath RC, Peris K, Nestle FO.** The IL23R R381Q gene variant protects against immune-mediated diseases by impairing IL-23-induced Th17 effector response in humans. *PLoS One* 2011; **6**: e17160 [PMID: 21364948 DOI: 10.1371/journal.pone.0017160]
- 12 **Pappa HM, Langereis EJ, Grand RJ, Gordon CM.** Prevalence and risk factors for hypovitaminosis D in young patients with inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2011; **53**: 361-364 [PMID: 21613964 DOI: 10.1097/MPG.0b013e3182250b3e]
- 13 **Loftus EV.** Clinical epidemiology of inflammatory bowel disease: Incidence, prevalence, and environmental influences. *Gastroenterology* 2004; **126**: 1504-1517 [PMID: 15168363 DOI: 10.1053/j.gastro.2004.01.063]
- 14 **Sartor RB, Mazmanian SK.** Intestinal Microbes in Inflammatory Bowel Diseases. *Am J Gastroenterol Suppl* 2012; **1**: 15-21 [DOI: 10.1038/ajgsup.2012.4]
- 15 **Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR.** Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci USA* 2007; **104**: 13780-13785 [PMID: 17699621 DOI: 10.1073/pnas.0706625104]
- 16 **Leitner GC, Vogelsang H.** Pharmacological- and non-pharmacological therapeutic approaches in inflammatory bowel disease in adults. *World J Gastrointest Pharmacol Ther* 2016; **7**: 5-20 [PMID: 26855808 DOI: 10.4292/wjgpt.v7.i1.5]
- 17 **Kozuch PL, Hanauer SB.** Treatment of inflammatory bowel disease: a review of medical therapy. *World J Gastroenterol* 2008; **14**: 354-377 [PMID: 18200659 DOI: 10.3748/wjg.14.354]
- 18 **Truelove SC, Witts LJ.** Cortisone in ulcerative colitis; final report on a therapeutic trial. *Br Med J* 1955; **2**: 1041-1048 [PMID: 13260656 DOI: 10.1159/000199983]
- 19 **Ford AC, Bernstein CN, Khan KJ, Abreu MT, Marshall JK, Talley NJ, Moayyedi P.** Glucocorticosteroid therapy in inflammatory

- bowel disease: systematic review and meta-analysis. *Am J Gastroenterol* 2011; **106**: 590-599; quiz 600 [PMID: 21407179 DOI: 10.1038/ajg.2011.70]
- 20 **Danese S**, Hart A, Dignass A, Louis E, D'Haens G, Dotan I, Rogler G, D'Agay L, Iannacone C, Peyrin-Biroulet L. Effectiveness of budesonide MMX (Cortiment) for the treatment of mild-to-moderate active ulcerative colitis: study protocol for a prospective multicentre observational cohort study. *BMJ Open Gastroenterol* 2016; **3**: e000092 [PMID: 27239329 DOI: 10.1136/bmjgast-2016-000092]
 - 21 **Sherlock ME**, MacDonald JK, Griffiths AM, Steinhart AH, Seow CH. Oral budesonide for induction of remission in ulcerative colitis. *Cochrane Database Syst Rev* 2015; **(10)**: CD007698 [PMID: 26497719 DOI: 10.1002/14651858.CD007698.pub3]
 - 22 **Paine ER**. Colonoscopic evaluation in ulcerative colitis. *Gastroenterol Rep (Oxf)* 2014; **2**: 161-168 [PMID: 24879406 DOI: 10.1093/gastro/gou028]
 - 23 **Best WR**, Beckett JM, Singleton JW, Kern F. Development of a Crohn's disease activity index. National Cooperative Crohn's Disease Study. *Gastroenterology* 1976; **70**: 439-444 [PMID: 1248701]
 - 24 **Harvey RF**, Bradshaw JM. A simple index of Crohn's-disease activity. *Lancet* 1980; **1**: 514 [PMID: 6102236 DOI: 10.1016/S0140-6736(80)92767-1]
 - 25 **Summers RW**, Switz DM, Sessions JT, Beckett JM, Best WR, Kern F, Singleton JW. National Cooperative Crohn's Disease Study: results of drug treatment. *Gastroenterology* 1979; **77**: 847-869 [PMID: 38176]
 - 26 **Malchow H**, Ewe K, Brandes JW, Goebell H, Ehms H, Sommer H, Jesdinsky H. European Cooperative Crohn's Disease Study (ECCDS): results of drug treatment. *Gastroenterology* 1984; **86**: 249-266 [PMID: 6140202 DOI: 10.1016/S0022-3468(84)80296-1]
 - 27 **Modigliani R**, Mary JY, Simon JF, Cortot A, Soule JC, Gendre JP, Rene E. Picture of attacks of Crohn's evolution on prednisolone. Groupe d'Etude Thérapeutique des Affections Inflammatoires Digestives. *Gastroenterology* 1990; **98**: 811-818 [PMID: 2179031 DOI: 10.1016/0016-5085(90)90002-I]
 - 28 **Truelove SC**, Watkinson G, Draper G. Comparison of corticosteroid and sulphasalazine therapy in ulcerative colitis. *Br Med J* 1962; **2**: 1708-1711 [PMID: 13994348 DOI: 10.1136/bmj.2.5321.1708]
 - 29 **Lennard-jones JE**, Longmore AJ, Newell AC, Wilson CW, Jones FA. An assessment of prednisone, salazopyrin, and topical hydrocortisone hemisuccinate used as out-patient treatment for ulcerative colitis. *Gut* 1960; **1**: 217-222 [PMID: 13760840 DOI: 10.1136/gut.1.3.217]
 - 30 **Principi M**, Mastroiardo M, Scicchitano P, Gesualdo M, Sassara M, Guida P, Bucci A, Zito A, Caputo P, Albano F, Ierardi E, Di Leo A, Ciccone MM. Endothelial function and cardiovascular risk in active inflammatory bowel diseases. *J Crohns Colitis* 2013; **7**: e427-e433 [PMID: 23473915 DOI: 10.1016/j.crohns.2013.02.001]
 - 31 **Ho GT**, Chiam P, Drummond H, Loane J, Arnott ID, Satsangi J. The efficacy of corticosteroid therapy in inflammatory bowel disease: analysis of a 5-year UK inception cohort. *Aliment Pharmacol Ther* 2006; **24**: 319-330 [PMID: 16842459 DOI: 10.1111/j.1365-2036.2006.02974.x]
 - 32 **Greenberg GR**, Feagan BG, Martin F, Sutherland LR, Thomson AB, Williams CN, Nilsson LG, Persson T. Oral budesonide for active Crohn's disease. Canadian Inflammatory Bowel Disease Study Group. *N Engl J Med* 1994; **331**: 836-841 [PMID: 8078529 DOI: 10.1056/NEJM199409293311303]
 - 33 **Bar-Meir S**, Chowers Y, Lavy A, Abramovitch D, Sternberg A, Leichtmann G, Reshef R, Odes S, Moshkovitz M, Bruck R, Eliakim R, Maoz E, Mittmann U. Budesonide versus prednisone in the treatment of active Crohn's disease. The Israeli Budesonide Study Group. *Gastroenterology* 1998; **115**: 835-840 [PMID: 9753485 DOI: 10.1016/S0016-5085(98)70254-9]
 - 34 **Tremaine WJ**, Hanauer SB, Katz S, Winston BD, Levine JG, Persson T, Persson A. Budesonide CIR capsules (once or twice daily divided-dose) in active Crohn's disease: a randomized placebo-controlled study in the United States. *Am J Gastroenterol* 2002; **97**: 1748-1754 [PMID: 12135030 DOI: 10.1111/j.1572-0241.2002.05835.x]
 - 35 **Rezaie A**, Kuenzig ME, Benchimol EI, Griffiths AM, Otley AR, Steinhart AH, Kaplan GG, Seow CH. Budesonide for induction of remission in Crohn's disease. *Cochrane Database Syst Rev* 2015; **(6)**: CD000296 [PMID: 26039678 DOI: 10.1002/14651858.CD000296.pub4]
 - 36 **Sandborn WJ**, Travis S, Moro L, Jones R, Gaultier T, Bagin R, Huang M, Yeung P, Ballard ED. Once-daily budesonide MMX® extended-release tablets induce remission in patients with mild to moderate ulcerative colitis: results from the CORE I study. *Gastroenterology* 2012; **143**: 1218-1226.e1-e2 [PMID: 22892337 DOI: 10.1053/j.gastro.2012.08.003]
 - 37 **Danese S**, Siegel CA, Peyrin-Biroulet L. Review article: integrating budesonide-MMX into treatment algorithms for mild-to-moderate ulcerative colitis. *Aliment Pharmacol Ther* 2014; **39**: 1095-1103 [PMID: 24641622 DOI: 10.1111/apt.12712]
 - 38 **Lichtenstein GR**, Travis S, Danese S, D'Haens G, Moro L, Jones R, Huang M, Ballard ED, Bagin R, Hardiman Y, Collazo R, Sandborn WJ. Budesonide MMX for the Induction of Remission of Mild to Moderate Ulcerative Colitis: A Pooled Safety Analysis. *J Crohns Colitis* 2015; **9**: 738-746 [PMID: 26094251 DOI: 10.1093/ecco-jcc/jjv101]
 - 39 **Prantera C**, Marconi S. Glucocorticosteroids in the treatment of inflammatory bowel disease and approaches to minimizing systemic activity. *Therap Adv Gastroenterol* 2013; **6**: 137-156 [PMID: 23503968 DOI: 10.1177/1756283X12473675]
 - 40 **Tursi A**, Giorgetti GM, Brandimarte G, Elisei W, Aiello F. Beclomethasone dipropionate for the treatment of mild-to-moderate Crohn's disease: an open-label, budesonide-controlled, randomized study. *Med Sci Monit* 2006; **12**: PI29-PI32 [PMID: 16733496]
 - 41 **Campieri M**, Cottone M, Miglio F, Manenti F, Astegiano M, D'Arienzo A, Manguso F, D'Albasio G, Bonanomi A, Galeazzi R, Orlando A, Castiglione GN, Gionchetti P. Beclomethasone dipropionate enemas versus prednisolone sodium phosphate enemas in the treatment of distal ulcerative colitis. *Aliment Pharmacol Ther* 1998; **12**: 361-366 [PMID: 9690726 DOI: 10.1046/j.1365-2036.1998.00299.x]
 - 42 **Bossa F**, Latiano A, Rossi L, Magnani M, Palmieri O, Dallapiccola B, Serafini S, Damonte G, De Santo E, Andriulli A, Annesse V. Erythrocyte-mediated delivery of dexamethasone in patients with mild-to-moderate ulcerative colitis, refractory to mesalamine: a randomized, controlled study. *Am J Gastroenterol* 2008; **103**: 2509-2516 [PMID: 18721243 DOI: 10.1111/j.1572-0241.2008.02103.x]
 - 43 **Faubion WA**, Loftus EV, Harmsen WS, Zinsmeister AR, Sandborn WJ. The natural history of corticosteroid therapy for inflammatory bowel disease: a population-based study. *Gastroenterology* 2001; **121**: 255-260 [PMID: 11487534 DOI: 10.1053/gast.2001.26279]
 - 44 **Manz M**, Vavricka SR, Wanner R, Lakatos PL, Rogler G, Frei P, Safroneeva E, Schoepfer AM. Therapy of steroid-resistant inflammatory bowel disease. *Digestion* 2012; **86** Suppl 1: 11-15 [PMID: 23051721 DOI: 10.1159/000341952]
 - 45 **Castro-Santos P**, Suarez A, López-Rivas L, Mozo L, Gutierrez C. TNFalpha and IL-10 gene polymorphisms in inflammatory bowel disease. Association of -1082 AA low producer IL-10 genotype with steroid dependency. *Am J Gastroenterol* 2006; **101**: 1039-1047 [PMID: 16573780 DOI: 10.1111/j.1572-0241.2006.00501.x]
 - 46 **De Iudicibus S**, Franca R, Martelossi S, Ventura A, Decorti G. Molecular mechanism of glucocorticoid resistance in inflammatory bowel disease. *World J Gastroenterol* 2011; **17**: 1095-1108 [PMID: 21448414 DOI: 10.3748/wjg.v17.i9.1095]
 - 47 **Lu NZ**, Wardell SE, Burnstein KL, Defranco D, Fuller PJ, Giguere V, Hochberg RB, McKay L, Renoir JM, Weigel NL, Wilson EM, McDonnell DP, Cidlowski JA. International Union of Pharmacology. LXV. The pharmacology and classification of the nuclear receptor superfamily: glucocorticoid, mineralocorticoid, progesterone, and androgen receptors. *Pharmacol Rev* 2006; **58**:

- 782-797 [PMID: 17132855 DOI: 10.1124/pr.58.4.9]
- 48 **Hollenberg SM**, Weinberger C, Ong ES, Cerelli G, Oro A, Lebo R, Thompson EB, Rosenfeld MG, Evans RM. Primary structure and expression of a functional human glucocorticoid receptor cDNA. *Nature* 1985; **318**: 635-641 [PMID: 2867473 DOI: 10.1038/318635a0]
 - 49 **Oakley RH**, Cidlowski JA. Cellular processing of the glucocorticoid receptor gene and protein: new mechanisms for generating tissue-specific actions of glucocorticoids. *J Biol Chem* 2011; **286**: 3177-3184 [PMID: 21149445 DOI: 10.1074/jbc.R110.179325]
 - 50 **Chrousos GP**, Kino T. Intracellular glucocorticoid signaling: a formerly simple system turns stochastic. *Sci STKE* 2005; **2005**: pe48 [PMID: 16204701 DOI: 10.1126/stke.3042005pe48]
 - 51 **Kino T**, Su YA, Chrousos GP. Human glucocorticoid receptor isoform beta: recent understanding of its potential implications in physiology and pathophysiology. *Cell Mol Life Sci* 2009; **66**: 3435-3448 [PMID: 19633971 DOI: 10.1007/s00018-009-0098-z]
 - 52 **Almawi WY**, Melemedjian OK. Molecular mechanisms of glucocorticoid antiproliferative effects: antagonism of transcription factor activity by glucocorticoid receptor. *J Leukoc Biol* 2002; **71**: 9-15 [PMID: 11781376]
 - 53 **Gross KL**, Cidlowski JA. Tissue-specific glucocorticoid action: a family affair. *Trends Endocrinol Metab* 2008; **19**: 331-339 [PMID: 18805703 DOI: 10.1016/j.tem.2008.07.009]
 - 54 **Chen HL**, Li LR. Glucocorticoid receptor gene polymorphisms and glucocorticoid resistance in inflammatory bowel disease: a meta-analysis. *Dig Dis Sci* 2012; **57**: 3065-3075 [PMID: 22752665 DOI: 10.1007/s10620-012-2293-2]
 - 55 **Fujishima S**, Takeda H, Kawata S, Yamakawa M. The relationship between the expression of the glucocorticoid receptor in biopsied colonic mucosa and the glucocorticoid responsiveness of ulcerative colitis patients. *Clin Immunol* 2009; **133**: 208-217 [PMID: 19646928 DOI: 10.1016/j.clim.2009.07.006]
 - 56 **Oakley RH**, Webster JC, Sar M, Parker CR, Cidlowski JA. Expression and subcellular distribution of the beta-isoform of the human glucocorticoid receptor. *Endocrinology* 1997; **138**: 5028-5038 [PMID: 9348235 DOI: 10.1210/endo.138.11.5501]
 - 57 **Pujols L**, Mullol J, Roca-Ferrer J, Torrego A, Xaubet A, Cidlowski JA, Picado C. Expression of glucocorticoid receptor alpha- and beta-isoforms in human cells and tissues. *Am J Physiol Cell Physiol* 2002; **283**: C1324-C1331 [PMID: 12225995 DOI: 10.1152/ajpcell.00363.2001]
 - 58 **Oakley RH**, Sar M, Cidlowski JA. The human glucocorticoid receptor beta isoform. Expression, biochemical properties, and putative function. *J Biol Chem* 1996; **271**: 9550-9559 [PMID: 8621628 DOI: 10.1074/jbc.271.16.9550]
 - 59 **Nagy Z**, Acs B, Butz H, Feldman K, Marta A, Szabo PM, Baghy K, Pazmany T, Racz K, Liko I, Patocs A. Overexpression of GRβ in colonic mucosal cell line partly reflects altered gene expression in colonic mucosa of patients with inflammatory bowel disease. *J Steroid Biochem Mol Biol* 2016; **155**: 76-84 [PMID: 26480216 DOI: 10.1016/j.jsbmb.2015.10.006]
 - 60 **Lewis-Tuffin LJ**, Jewell CM, Bienstock RJ, Collins JB, Cidlowski JA. Human glucocorticoid receptor beta binds RU-486 and is transcriptionally active. *Mol Cell Biol* 2007; **27**: 2266-2282 [PMID: 17242213 DOI: 10.1128/MCB.01439-06]
 - 61 **Webster JC**, Oakley RH, Jewell CM, Cidlowski JA. Proinflammatory cytokines regulate human glucocorticoid receptor gene expression and lead to the accumulation of the dominant negative beta isoform: a mechanism for the generation of glucocorticoid resistance. *Proc Natl Acad Sci U S A* 2001; **98**: 6865-6870 [PMID: 11381138 DOI: 10.1073/pnas.121455098]
 - 62 **Boivin MA**, Ye D, Kennedy JC, Al-Sadi R, Shepela C, Ma TY. Mechanism of glucocorticoid regulation of the intestinal tight junction barrier. *Am J Physiol Gastrointest Liver Physiol* 2007; **292**: G590-G598 [PMID: 17068119 DOI: 10.1152/ajpgi.00252.2006]
 - 63 **van Rossum EFC**, Lamberts SWJ. Polymorphisms in the glucocorticoid receptor gene and their associations with metabolic parameters and body composition. *Recent Prog Horm Res* 2004; **59**: 333-57 [PMID: 14749509 DOI: 10.1210/rp.59.1.333]
 - 64 **Russcher H**, Smit P, van den Akker EL, van Rossum EF, Brinkmann AO, de Jong FH, Lamberts SW, Koper JW. Two polymorphisms in the glucocorticoid receptor gene directly affect glucocorticoid-regulated gene expression. *J Clin Endocrinol Metab* 2005; **90**: 5804-5810 [PMID: 16030164 DOI: 10.1210/jc.2005-0646]
 - 65 **Marti A**, Ochoa MC, Sánchez-Villegas A, Martínez JA, Martínez-González MA, Hebebrand J, Hinney A, Vedder H. Meta-analysis on the effect of the N363S polymorphism of the glucocorticoid receptor gene (GRL) on human obesity. *BMC Med Genet* 2006; **7**: 50 [PMID: 16725041 DOI: 10.1186/1471-2350-7-50]
 - 66 **Farrell RJ**, Menconi MJ, Keates AC, Kelly CP. P-glycoprotein-170 inhibition significantly reduces cortisol and ciclosporin efflux from human intestinal epithelial cells and T lymphocytes. *Aliment Pharmacol Ther* 2002; **16**: 1021-1031 [PMID: 11966513 DOI: 10.1046/j.1365-2036.2002.01238.x]
 - 67 **Hoffmeyer S**, Burk O, von Richter O, Arnold HP, Brockmüller J, Johné A, Cascorbi I, Gerloff T, Roots I, Eichelbaum M, Brinkmann U. Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo. *Proc Natl Acad Sci USA* 2000; **97**: 3473-3478 [PMID: 10716719 DOI: 10.1073/pnas.050585397]
 - 68 **Carvalho AT**, Frôes RS, Esberard BC, Santos JC, Rapozo DC, Grinman AB, Simão TA, Nicolau Neto P, Luiz RR, Carneiro AJ, Souza HS, Ribeiro-Pinto LF. Multidrug resistance 1 gene polymorphisms may determine Crohn's disease behavior in patients from Rio de Janeiro. *Clinics (Sao Paulo)* 2014; **69**: 327-334 [PMID: 24838898 DOI: 10.6061/clinics/2014(05)06]
 - 69 **Ishiguro Y**. Mucosal proinflammatory cytokine production correlates with endoscopic activity of ulcerative colitis. *J Gastroenterol* 1999; **34**: 66-74 [PMID: 10204613 DOI: 10.1007/s005350050218]
 - 70 **Wine E**, Mack DR, Hyams J, Otley AR, Markowitz J, Crandall WV, Leleiko N, Muise AM, Griffiths AM, Turner D. Interleukin-6 is associated with steroid resistance and reflects disease activity in severe pediatric ulcerative colitis. *J Crohns Colitis* 2013; **7**: 916-922 [PMID: 23339932 DOI: 10.1016/j.crohns.2012.12.012]
 - 71 **Santaolalla R**, Mañé J, Pedrosa E, Lorén V, Fernández-Bañares F, Mallolas J, Carrasco A, Salas A, Rosinach M, Forné M, Espinós JC, Loras C, Donovan M, Puig P, Mañosa M, Gassull MA, Viver JM, Esteve M. Apoptosis resistance of mucosal lymphocytes and IL-10 deficiency in patients with steroid-refractory Crohn's disease. *Inflamm Bowel Dis* 2011; **17**: 1490-1500 [PMID: 21674705 DOI: 10.1002/ibd.21507]
 - 72 **Montero-Meléndez T**, Llor X, García-Planella E, Perretti M, Suárez A. Identification of novel predictor classifiers for inflammatory bowel disease by gene expression profiling. *PLoS One* 2013; **8**: e76235 [PMID: 24155895 DOI: 10.1371/journal.pone.0076235]
 - 73 **Cadwell K**, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, Kishi C, Kc W, Carrero JA, Hunt S, Stone CD, Brunt EM, Xavier RJ, Sleekman BP, Li E, Mizushima N, Stappenbeck TS, Virgin HW. A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal Paneth cells. *Nature* 2008; **456**: 259-263 [PMID: 18849966 DOI: 10.1038/nature07416]
 - 74 **Jiang L**, Xu L, Xie J, Li S, Guan Y, Zhang Y, Hou Z, Guo T, Shu X, Wang C, Fan W, Si Y, Yang Y, Kang Z, Fang M, Liu Q. Inhibition of autophagy overcomes glucocorticoid resistance in lymphoid malignant cells. *Cancer Biol Ther* 2015; **16**: 466-476 [PMID: 25778879 DOI: 10.1080/15384047.2015.1016658]
 - 75 **Yang QF**, Chen BL, Zhang QS, Zhu ZH, Hu B, He Y, Gao X, Wang YM, Hu PJ, Chen MH, Zeng ZR. Contribution of MDR1 gene polymorphisms on IBD predisposition and response to glucocorticoids in IBD in a Chinese population. *J Dig Dis* 2015; **16**: 22-30 [PMID: 25346426 DOI: 10.1111/1751-2980.12205]
 - 76 **Fung KL**, Gottesman MM. A synonymous polymorphism in a common MDR1 (ABCB1) haplotype shapes protein function. *Biochim Biophys Acta* 2009; **1794**: 860-871 [PMID: 19285158]

- DOI: 10.1016/j.bbapap.2009.02.014]
- 77 **Hitzl M**, Drescher S, van der Kuip H, Schäffeler E, Fischer J, Schwab M, Eichelbaum M, Fromm MF. The C3435T mutation in the human MDR1 gene is associated with altered efflux of the P-glycoprotein substrate rhodamine 123 from CD56+ natural killer cells. *Pharmacogenetics* 2001; **11**: 293-298 [PMID: 11434506 DOI: 10.1097/00008571-200106000-00003]
 - 78 **Chikanza IC**, Kozaci D, Chernajovsky Y. The molecular and cellular basis of corticosteroid resistance. *J Endocrinol* 2003; **179**: 301-310 [PMID: 14656201 DOI: 10.1677/joe.0.1790301]
 - 79 **Bond HM**, Mesuraca M, Amodio N, Mega T, Agosti V, Fanello D, Pelaggi D, Bullinger L, Grieco M, Moore MA, Venuta S, Morrone G. Early hematopoietic zinc finger protein-zinc finger protein 521: a candidate regulator of diverse immature cells. *Int J Biochem Cell Biol* 2008; **40**: 848-854 [PMID: 17543573 DOI: 10.1016/j.biocel.2007.04.006]
 - 80 **Park SH**, Kim SK, Choe JY, Moon Y, An S, Park MJ, Kim DS. Hypermethylation of EBF3 and IRX1 genes in synovial fibroblasts of patients with rheumatoid arthritis. *Mol Cells* 2013; **35**: 298-304 [PMID: 23456299 DOI: 10.1007/s10059-013-2302-0]
 - 81 **Zhao LY**, Niu Y, Santiago A, Liu J, Albert SH, Robertson KD, Liao D. An EBF3-mediated transcriptional program that induces cell cycle arrest and apoptosis. *Cancer Res* 2006; **66**: 9445-9452 [PMID: 17018599 DOI: 10.1158/0008-5472.CAN-06-1713]
 - 82 **Tahara T**, Shibata T, Nakamura M, Yamashita H, Yoshioka D, Okubo M, Maruyama N, Kamano T, Kamiya Y, Fujita H, Nakagawa Y, Nagasaka M, Iwata M, Takahama K, Watanabe M, Nakano H, Hirata I, Arisawa T. Promoter methylation of protease-activated receptor (PAR2) is associated with severe clinical phenotypes of ulcerative colitis (UC). *Clin Exp Med* 2009; **9**: 125-130 [PMID: 19184329 DOI: 10.1007/s10238-008-0025-x]
 - 83 **Kuester D**, Guenther T, Biesold S, Hartmann A, Bataille F, Ruemmele P, Peters B, Meyer F, Schubert D, Bohr UR, Malfertheiner P, Lippert H, Silver AR, Roessner A, Schneider-Stock R. Aberrant methylation of DAPK in long-standing ulcerative colitis and ulcerative colitis-associated carcinoma. *Pathol Res Pract* 2010; **206**: 616-624 [PMID: 20630662 DOI: 10.1016/j.prp.2010.05.004]
 - 84 **Balasa A**, Gathungu G, Kisfali P, Smith EO, Cho JH, Melegh B, Kellermayer R. Assessment of DNA methylation at the interferon regulatory factor 5 (IRF5) promoter region in inflammatory bowel diseases. *Int J Colorectal Dis* 2010; **25**: 553-556 [PMID: 20127100 DOI: 10.1007/s00384-010-0874-0]
 - 85 **Sato F**, Harpaz N, Shibata D, Xu Y, Yin J, Mori Y, Zou TT, Wang S, Desai K, Leytin A, Selaru FM, Abraham JM, Meltzer SJ. Hypermethylation of the p14(ARF) gene in ulcerative colitis-associated colorectal carcinogenesis. *Cancer Res* 2002; **62**: 1148-1151 [PMID: 11861396]
 - 86 **Harris RA**, Nagy-Szakal D, Kellermayer R. Human metastable epiallele candidates link to common disorders. *Epigenetics* 2013; **8**: 157-163 [PMID: 23321599 DOI: 10.4161/epi.23438]
 - 87 **Weinhouse C**, Anderson OS, Jones TR, Kim J, Liberman SA, Nahar MS, Rozek LS, Jirtle RL, Dolinoy DC. An expression microarray approach for the identification of metastable epialleles in the mouse genome. *Epigenetics* 2011; **6**: 1105-1113 [PMID: 21829099 DOI: 10.4161/epi.6.9.17103]
 - 88 **Weaver IC**, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, Dymov S, Szyf M, Meaney MJ. Epigenetic programming by maternal behavior. *Nat Neurosci* 2004; **7**: 847-854 [PMID: 15220929 DOI: 10.1038/nn1276]

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Basic Study

Attenuation of MET-mediated migration and invasion in hepatocellular carcinoma cells by SOCS1

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Abstract

AIM

To investigate the role of suppressor of cytokine signaling 1 (SOCS1) in regulating MET-mediated invasive potential of hepatocellular carcinoma (HCC) cells.

METHODS

Stable derivatives of mouse (Hepa1-6) and human (hep3B, HepG2) HCC cell lines expressing SOCS1 or control vector were evaluated for their ability to migrate towards hepatocyte growth factor (HGF) in the transwell migration assay, invade extracellular matrix in response to HGF stimulation in a 3-D invasion assay by confocal microscopy, and to undergo anchorage-independent proliferation in semisolid agar. Following intravenous and intrasplenic inoculation into NOD.scid. gamma mice, the ability of Hepa cells to form orthotopic tumors was evaluated. Following HGF stimulation of Hepa and Hep3B cells, expression of proteins implicated in epithelial-to-mesenchymal transition was evaluated by western blot and qRT-PCR.

RESULTS

SOCS1 expression in mouse and human HCC cells inhibited HGF-induced migration through matrigel. In the

3-D invasion assay, HGF stimulation induced invasion of HCC cells across type-I collagen matrix, and SOCS1 expression significantly reduced the depth of invasion. SOCS1 expression also reduced the number and size of colonies formed by anchorage-independent growth in semisolid agar. Following intravenous inoculation, control Hepa cell formed large tumor nodules that obliterated the liver whereas the SOCS1-expressing Hepa cells formed significantly smaller nodules. Tumors formed by SOCS1-expressing cells showed reduced phosphorylation of STAT3 and ERK that was accompanied by reduced levels of MET protein expression. HGF stimulated Hepa cells expressing SOCS1 showed increased expression of E-cadherin and decreased expression of EGR1, SNAI1 and ZEB1. Comparable results were obtained with Hep3B cells. SOCS1 expressing HCC cells also showed reduced levels of EGR1 and SNAI1 transcripts.

CONCLUSION

Our findings indicate that loss of SOCS1-dependent control over epithelial-to-mesenchymal transition may contribute to MET-mediated migration, invasion and metastatic growth of HCC.

Key words: Migration; Invasion; MET; Hepatocellular carcinoma; Suppressor of cytokine signaling 1

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Core tip: The suppressor of cytokine signaling 1 (*SOCS1*) gene is frequently repressed in primary hepatocellular carcinoma (HCC) specimens, and mice lacking *SOCS1* are highly susceptible to experimental HCC. The tumor suppressor functions of *SOCS1* in HCC are not yet fully understood. We have shown that *SOCS1* regulates hepatocyte growth factor signaling *via* the MET receptor in HCC cells and inhibits their growth. In this study, we characterize *SOCS1* as a regulator of MET-mediated migration and invasion of HCC cells. We propose that *SOCS1* gene expression status could be exploited as a selection marker for precision therapies targeting MET in HCC.

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INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most prevalent cancers and a leading cause of worldwide cancer mortality^[1]. HCC develops slowly over two to three decades and most cases present advanced disease at the time of diagnosis. Treatment options for HCC are constrained by the extent of disease, and

are very limited and less effective in patients with advanced disease. Therefore, reducing HCC-associated mortality is critically dependent on the development of new treatment methods targeting molecular signaling pathways that promote HCC pathogenesis and diagnostic tools to facilitate targeted therapies^[2-4].

Invasive intrahepatic dissemination is a key factor in malignant growth of HCC and its poor prognosis^[5,6]. Up to 65% of HCC patients also present extrahepatic metastasis at autopsy^[7-9]. HCC can also metastasize to stomach *via* direct invasion^[10]. Detachment from the tumor mass and invasion of the extracellular matrix and the basement membrane are important steps in tumor cell invasion and metastasis. Cancer cells gain these abilities through epithelial-mesenchymal transition (EMT), a developmental genetic program that is crucial for embryogenesis and wound healing response^[11,12]. A wide spectrum of paracrine (from the tumor stroma) and autocrine cytokines and growth factors elicit and modulate the EMT program^[12]. Even though TGFβ is the most important inducer of EMT, growth factor receptor tyrosine kinase (RTK) signaling induced by hepatocyte growth factor (HGF), epidermal growth factor and platelet-derived growth factor can also activate the EMT program in carcinomas^[12,13].

The HGF receptor c-MET is overexpressed in many human cancers including HCC^[14]. MET not only promotes neoplastic growth of HCC cells but also facilitates tumor metastasis by promoting EMT^[14-16]. Recent studies have implicated the microRNAs miR-148a and miR-449a in regulating EMT in HCC cells by targeting the MET receptor^[17,18]. Previously, we have shown that suppressor of cytokine signaling 1 (*SOCS1*) is an important regulator of HGF signaling hepatocytes. *SOCS1* deficiency in primary mouse hepatocytes increases MET signaling and cell proliferation, whereas stable expression of *SOCS1* in human HCC cells attenuates HGF-induced cell growth^[19,20]. We have also shown that *SOCS1* binds to the MET receptor and promotes its ubiquitination and proteasomal degradation^[20]. The *SOCS1* gene is frequently repressed in HCC, and *Socs1*-deficient mice show high susceptibility to experimental HCC with larger and more numerous tumor nodules^[21-23], suggesting a role for *SOCS1* in controlling MET-mediated tumor cell invasion.

In this study, we examined the role of *SOCS1* in MET-mediated migration and invasion of HCC cells and their growth following delivery to the liver. Our findings show that *SOCS1* inhibits the invasive potential of HCC cells and reduces orthotopic tumor growth that could result at least partly from modulating the expression of proteins implicated in EMT.

MATERIALS AND METHODS

Cell lines, antibodies and reagents

HCC cell lines of mouse (Hepa1-6; Hepa) and human origin (Hep3B, HepG2) were obtained from ATCC

and grown in Dulbecco's modified Eagles medium (DMEM) containing 10% fetal bovine serum (FBS, from Sigma). Cells stably expressing full-length HA-SOCS1 through lentiviral transduction have been previously described^[20]. Recombinant mouse and human HGF were from Peprotech (Rocky Hill, NJ). Phospho-ERK (T202/Y204; #9106), phospho-STAT3 (Y705; #9131), E-cadherin (24E10), ZO-1 (D7D12), N-cadherin (D4R1H), vimentin (D21H3), EGR1 (44D5), SNAI1 (C15D3) and ZEB1 (D80D3) antibodies were from Cell Signaling Technology (Beverly, MA). Antibodies (Ab) against total ERK (sc-93, sc-153), STAT3 (sc-483) and MET (sc-161) were from Santa Cruz Biotechnology (Santa Cruz, CA). Mouse mAb against β -actin (A4700) was from Sigma Aldrich (Oakville, ON, Canada). Secondary antibodies were from Jackson Immuno-research Laboratories Inc. (Cedarlane, Burlington, ON, United States). Calcein was from Calbiochem (San Diego, CA, United States).

Orthotopic tumor growth

Male NOD.*scid.gamma* (NSG) mice (8-12 wk old) purchased from the Jackson Labs (Bar Harbor, ME, United States) were used to evaluate tumor growth *in vivo* under protocols approved by the Université de Sherbrooke ethical committee on animal care and use. To evaluate the growth of hepatoma cells in the liver, cells were injected *via* intravenous or intrasplenic/portal route. For intravenous inoculation, 10^6 Hepa-vector or Hepa-SOCS1 cells in 100 μ L volume were injected *via* the caudal vein. For intrasplenic inoculation, mice were anesthetized with ketamine (10 mg/kg) and the spleen was exposed through a small abdominal incision^[24]. Tumor cells (10^6 cells in 100 μ L) were injected into the spleen and mice were splenectomized 2 min later. Tumor nodules in the liver were examined 20 d later when the animals began to show distress. The images of hematoxylin and eosin (H and E) -stained sections of the liver were acquired using Nanozoomer Slide Scanner and analyzed by the Nanozoomer Digital Pathology (NDP) software (Hamamatsu Photonics, Japan).

Soft agar colony formation

After adding the bottom layer of 0.6% agar in 2 \times DMEM containing 20% FBS to 100 mm Petri dishes, 5000 HCC cells suspended in 0.3% agar in 2 \times DMEM containing 10% were added FBS as the top layer. Cells were fed every 3-5 d by overlying fresh 500 μ L of DMEM containing 10% FBS. After 3 wk culture, colonies were stained with 1 mg/mL methylthiazol tetrazolium for 3 h, photographed and counted using the NIH ImageJ software.

Transwell migration assay

Migration of cells was assessed using transwells (Corning) with 8 μ mol/L pore polycarbonated inserts, coated with growth factor-reduced matrigel (BD Pharmingen Biosciences, Palo Alto, CA, United States). The upper chamber contained 2 $\times 10^4$ cells and the lower

chamber contained 0.7 mL of complete medium with or without HGF. Migration through the membrane was determined after 24 h of incubation at 37 °C. Cells remaining on the top side of the transwell membrane were removed using a cotton swab, and cells migrated to bottom were stained with 0.5% crystal violet and examined by microscopy. Cells in five random fields of the lower side of the membrane were counted. Data represent means \pm SEM of three independent experiments.

3-dimensional invasion assay

The wells of a 96-well microtiter plate was covered with 50 μ L of 1% agarose containing 10% FBS as the bottom layer and overlaid with 50 μ L of fibrillar type-I collagen (3 mg/mL) (R and D Systems, Minneapolis, MN, United States). Control and SOCS1-expressing HCC cells were serum starved overnight and layered on the top (20000 cells/well in 100 μ L volume) in the presence or absence of HGF. After 48 h incubation, the cells were labeled with CellTrace™ Calcein Green, AM (Invitrogen, CA, United States) for 1 h, washed with PBS, fixed with 3% glutaraldehyde and confocal images were acquired using the FV1000 Olympus confocal microscope. The collagen matrix was scanned along the Z-axis at incremental 5 μ m depths in order to reconstruct 3-D images. The images were analyzed to assess the depth of cell migration, which is expressed as ratio of the fluorescence intensity at each 5 μ m layer over the fluorescence intensity of the non-invaded cells at the top 5 μ m layer.

Western blot

Lysates of liver tissues harboring the tumor nodules were prepared in a buffer containing 150 mmol/L NaCl, 50 mmol/L Tris-HCl, 1 mmol/L EDTA, pH 8.0 and protease and phosphatase inhibitor cocktails (Roche, Indianapolis, IN) for 2 min using the bead mill MM 400 (Retsch, Hann, Germany). After adding equal volume of the same buffer containing 0.2% SDS, 1% Triton X-100 and 1% sodium deoxycholate, the lysates were kept on a shaker at 4 °C for 30 min. Following centrifugation at 15000 *g* for 20 min, the supernatant was collected and protein concentration determined using the RC-DC Protein Assay Kit (Bio-Rad, Mississauga, ON, United States). Cultured cells were lysed directly in 1 \times SDS-PAGE sample buffer. Aliquots of 30-50 μ g proteins were analyzed by western blot. Secondary antibodies and enhanced chemiluminescence reagents were from GE Healthcare Life Sciences (Pittsburg, PA, United States). Western blot images were captured by the VersaDOC 5000 imaging system (Bio-Rad). Densitometric quantification of the western blot bands was carried out using the NIH ImageJ 1.62 software.

Real-time RT-PCR

For RT-PCR analysis, total RNA was isolated from 1 $\times 10^6$ cells, either unstimulated or stimulated with HGF for the indicated periods of time, using RiboZol™ (AMRESCO,

Solon, OH, United States). After verifying the RNA quality by UV absorption, the first complementary strand was made from 1 µg total RNA using QuantiTect® reverse transcription kit (Qiagen). The primers for gene expression analysis are as follows: Mouse *Egr1* (NM_007913): AGCGCCTTCAATCCTCAAG and TTTGGC TGGGATAACTCGTC; *Snai1* (NM_011427): GTGAAGAGA TACCAAGTCCAG and AAGATGCCAGCGAGGATG; Human *EGR1* (NM_001964): TGTCACCAACTCCTTCAGC and TCCTGTCCTTAAAGTCTCTTGTC; *SNAI1*: TCTAGGCCCTG GCTGCTACAA and ACATCTGAGTGGGTCTGGAGGTG; housekeeping genes: mouse *Gapdh* (NM_008084.3): ATGACATCAAGAAGGTGGTGAA and GTCTTACTCCTTGG AGGCCATGT; Human *GAPDH* (NM_002046.5): GATGA CATCAAGAAGGTGGTGAA and GTCTTACTCCTTGGAGG CCATGT. All primers showed more than 90% efficiency with a single melting curve in the MyQ15® cyclor (Bio-Rad, Mississauga, ON, United States). Expression levels of the housekeeping gene were used to calculate fold induction of the specific genes modulated by the presence or absence of SOCS1.

Statistical analysis

Statistical significance of the differences in cell proliferation, tumor growth and protein expression was evaluated by student's *t*-test.

RESULTS

Previously we have shown that SOCS1 inhibited HGF-induced MET signaling, cell proliferation and migration in mouse (Hepa) and human (Hep3B, HepG2, SNU-423) HCC cell lines^[19,20]. Here, we investigated whether SOCS1 also inhibits HGF-induced invasion of the extracellular matrix in some of these cell lines. At the concentration previously found to be effective in inducing cell proliferation and migration^[19], HGF induced cell scattering in murine Hepa cells and human Hep3B cells, which was attenuated by SOCS1 (Figure 1A). In a trans-well migration assay, Hepa cells expressing the control vector showed marked ability to invade the matrigel in response to HGF, whereas the SOCS1-expressing cells showed significantly reduced ability (Figure 1B). Similar reduction of HGF-induced matrix invasion by SOCS1 was also observed with the human HCC cell line Hep3B (Figure 1C).

Next, we used a 3-D matrix invasion assay using collagen as matrix, where the depth of invasion was evaluated by confocal microscopy and quantified (Figure 2A and B). HGF stimulation increased the ability of Hepa, Hep3B and HepG2 cells to invade the collagen matrix, and this invasive potential was markedly pronounced in Hep3B and HepG2 cells, as indicated by the maximum depth of invasion (Figure 2A and C). In all the three cell lines, SOCS1 reduced the HGF-induced invasive potential (Figure 2A and C). Even though the SOCS1-mediated inhibition of HGF-induced invasion

was only marginal in Hepa cells, Hep3B-SOCS1 and HepG2-SOCS1 cells showed significant reduction in HGF-induced invasive potential (Figure 2D). These results indicated that SOCS1 is an important regulator of MET signaling that promotes cell migration and invasion.

Tumor dissemination requires not only the ability to invade but also the ability to survive and undergo anchorage-independent proliferation. Therefore, we compared the ability of control and SOCS1-expressing HCC cells to form colonies in semi-solid agar. As shown in Figure 3A, Hepa-SOCS1 cells form fewer and smaller colonies than Hepa-V cells. Similar reduction of colony forming ability was also observed in Hep3B-SOCS1 and HepG2-SOCS1 cells compared to controls (Figure 3B).

To evaluate whether SOCS1 attenuates invasive tumor growth within the liver, we delivered control and SOCS1-expressing HCC cells *via* parenteral inoculation to the liver. For this purpose, we used murine Hepa1-6 cells, which can utilize endogenous HGF and other mouse growth factors better than human hepatoma cells^[25]. Following intravenous injection, the control cells formed numerous orthotopic tumor nodules that obliterated the liver, whereas Hepa-SOCS1 cells induced fewer nodules (Figure 4A). The lungs of these mice did not show macroscopically visible tumor, suggesting that Hepa cells preferentially colonized the liver. We also delivered the HCC cells to the liver *via* intrasplenic route through the portal circulation. In the latter setting, Hepa-vector cells obliterated the liver with numerous macroscopically visible nodules, whereas Hepa-SOCS1 cells formed markedly smaller nodules (Figure 4B and C). Orthotopic tumors formed by Hepa-SOCS1 showed markedly diminished phosphorylation of STAT3 and ERK (Figure 4D), presumably through SOCS1-mediated inhibition of HGF and other cytokine and growth factor signaling^[19,26,27]. Moreover, the SOCS1 expressing tumors showed significantly reduced levels of MET expression (Figure 4E), in agreement with our previous report that SOCS1 targets the activated MET receptor to the proteasomal degradation machinery^[20]. These results indicated that SOCS1 inhibits the invasive growth potential of HCC cells and that at least part of this invasive growth could result from attenuation of HGF-induced MET signaling.

To understand the mechanisms underlying SOCS1-mediated attenuation of the invasive growth of HCC cells, we evaluated the expression of proteins implicated in EMT that promote tumor cell spreading following HGF stimulation^[28-30]. Even though HGF stimulation did not appreciably affect the expression of E-Cadherin in control or SOCS1-expressing Hepa and Hep3B cells, the SOCS1-expressing cells showed increased basal expression of E-Cadherin (Figure 5A). SOCS1-expressing Hepa cells but not Hep3B cells showed discernible increase in ZO-1 expression, whereas the expression of N-Cadherin was not affected by SOCS1 in both cell

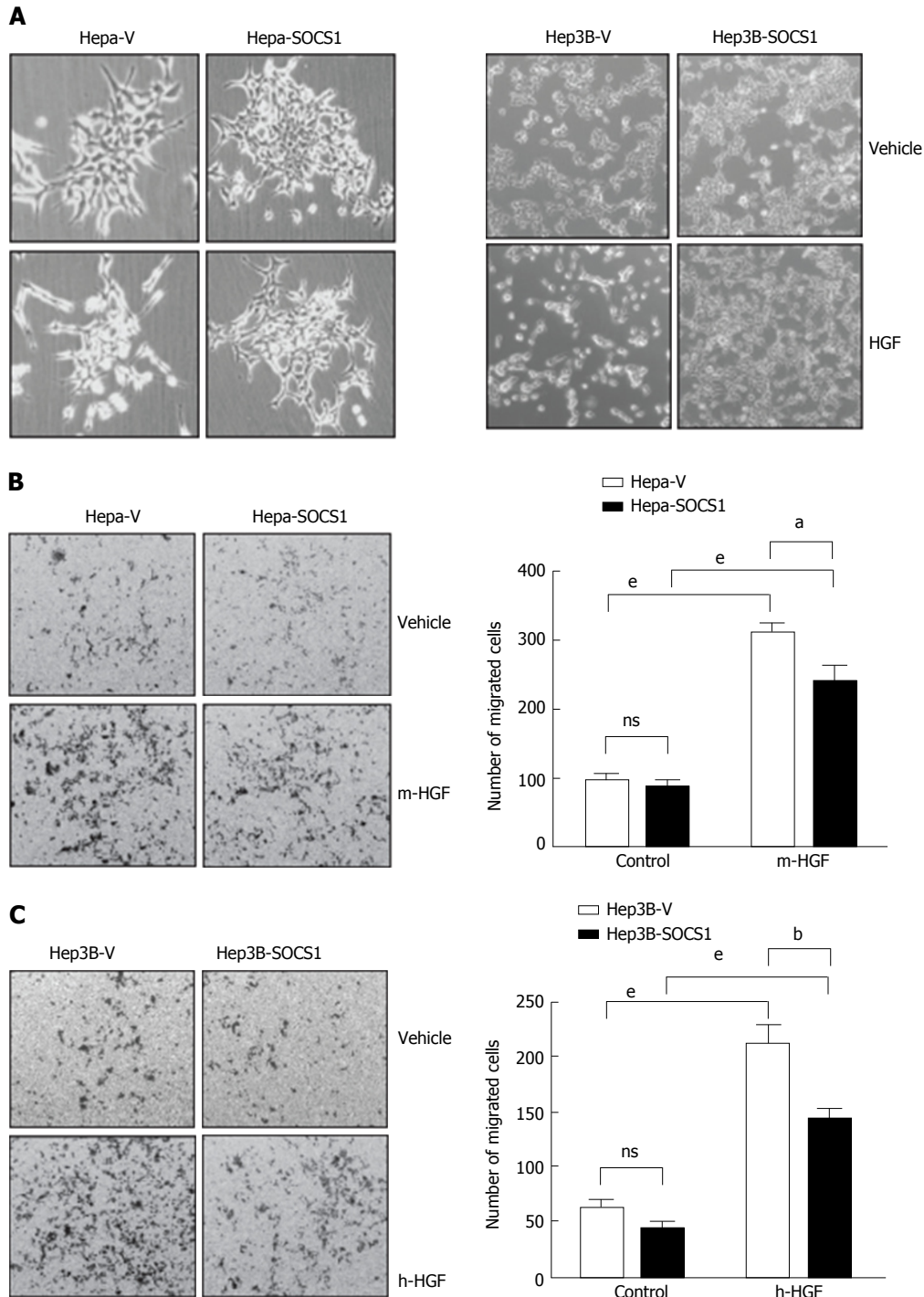


Figure 1 Suppressor of cytokine signaling 1 attenuates hepatocyte growth factor-induced scattering and migration of hepatocellular carcinoma cells. **A:** Hepa and Hep3B cells expressing SOCS1 (Hepa-SOCS1, Hep3B-SOCS1) or control vector (Hepa-V, Hep3B-V) were grown to less than 50% confluent state, starved in serum-free medium for 16 h and exposed to murine or human HGF (HGF, 25 ng/mL), respectively. After 48 h, the culture plates were examined by phase contrast microscopy to visualize HGF-induced cell scattering; **B:** Hepa-SOCS1 and Hepa-V cells were seeded onto Matrigel-coated trans-well migration chambers, and migration towards m-HGF was evaluated after 48 h. Representative images of cells that had migrated across the membrane and the number of migrated cells in five random fields from three separate experiments (mean \pm SE) are shown; **C:** Migration of Hep3B cells expressing SOCS1 (Hep3B-SOCS1) or control vector (Hepa-V) towards human HGF (h-HGF, 10 ng/mL). ^a $P < 0.05$, ^b $P < 0.01$, ^c $P < 0.001$. SOCS1: Suppressor of cytokine signaling 1.

lines. Analysis of gene expression showed marked reduction in the induction *Egr1* and *Snai1* genes in Hepa-SOCS1 cells, and *EGR1* and *SNAI2* genes in Hep3B-SOCS1 cells (Figure 5B). Western blot analysis showed reduced expression of EGR1, SNAI1 and

ZEB1 in Hepa-SOCS1 cells compared to Hepa-V cells, whereas only SNAI1 showed appreciable reduction in Hep3B cells (Figure 5A). These findings suggest that SOCS1 may modulate diverse components of the EMT signaling machinery during MET-mediated invasion of

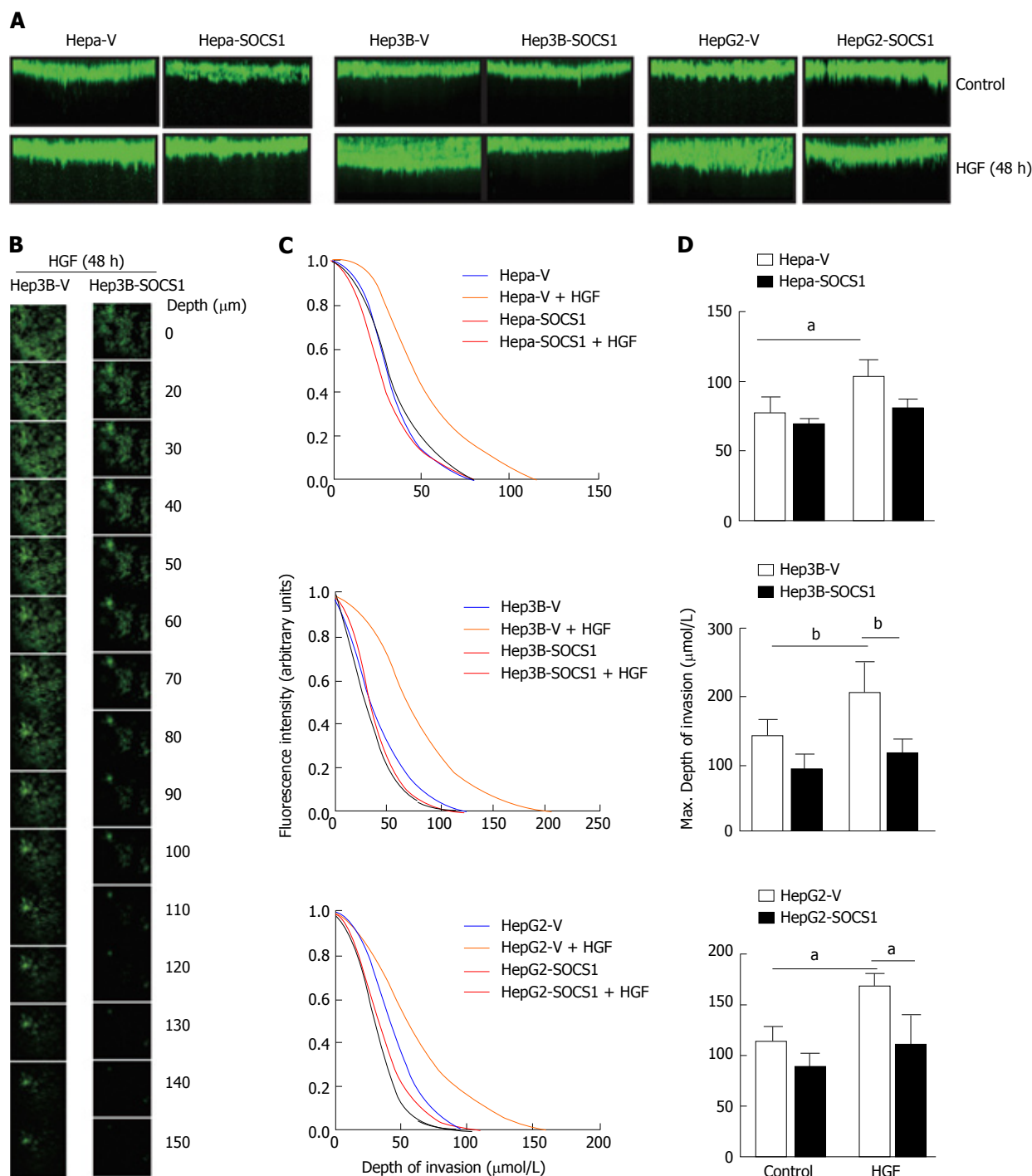


Figure 2 Inhibition of hepatocyte growth factor-induced invasion of collagen matrix by suppressor of cytokine signaling 1 in hepatocellular carcinoma cells. Hepa-V and Hepa-SOCS1, Hep3B-V and Hep3B-SOCS1, and HepG2-V and HepG2-SOCS1 cell lines were evaluated for their ability to invade collagen matrix in a 3-D invasion assay in the presence or absence of mouse HGF (25 ng/mL for Hepa cells) or human HGF (30 ng/mL for Hep3B, HepG2 cells). After 48 h, cells were fluorescently labeled with calcein Green, fixed and examined by confocal microscopy. A: The reconstructed cross-sectional images of cell migration, representative of two independent experiments, are shown; B: Representative images of Hep3B-V and Hep3B-SOCS1 cells migrated to different planes of the collagen-agarose matrix along the z-axis; C: Efficiency of migration (ratio of the fluorescence intensity at each 5 μm layer over the fluorescence intensity of the non-invaded cells at the top 5 μm); D: Maximum depth of invasion by control and SOCS1-expressing cells in the absence or presence of HGF. ^a $P < 0.05$, ^b $P < 0.01$. SOCS1: Suppressor of cytokine signaling 1.

HCC cells.

DISCUSSION

The *SOCS1* gene is frequently repressed in HCC by promoter CpG methylation^[21,31,32]. Recent meta-

analyses of published works on DNA methylation status of several HCC-associated genes have shown that *SOCS1* gene promoter hypermethylation is an important risk factor and predictive biomarker for HCC^[33,34]. Consistent with the role of *SOCS1* as a putative tumor suppressor in HCC, others and we

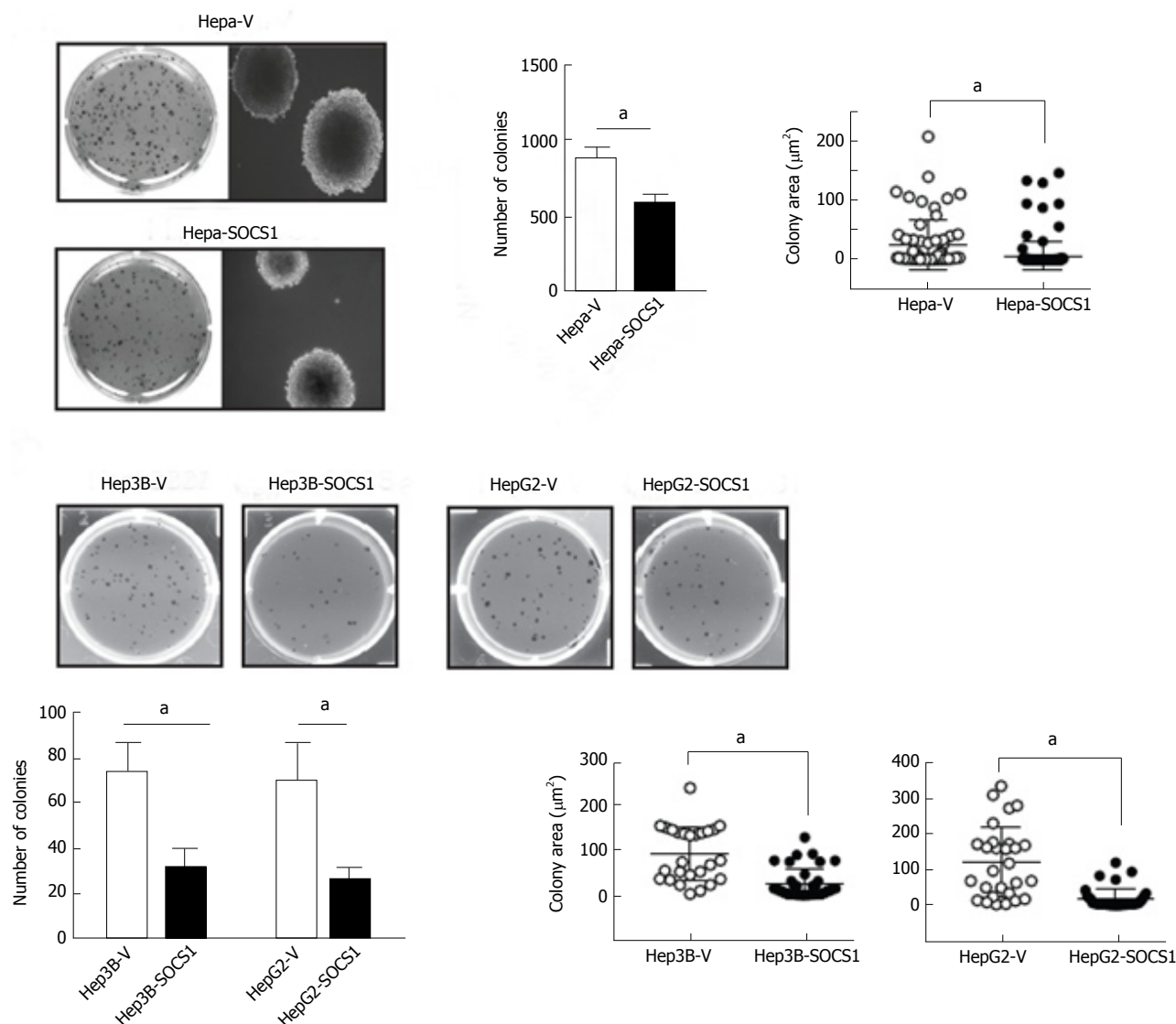


Figure 3 Reduced anchorage-independent proliferation of hepatocellular carcinoma cells expressing suppressor of cytokine signaling 1. SOCS1-expressing and control mouse Hepa (A) and human Hep3B and HepG2 (B) HCC cell lines were evaluated for their ability to form colonies in soft agar. After 3 wk culture, the colonies were photographed, and the number and size of colonies (area; mean \pm SE) were evaluated from triplicates of two separate experiments. ^a $P < 0.05$. SOCS1: Suppressor of cytokine signaling 1.

have shown that loss of *Socs1* gene expression in mice increases susceptibility to HCC induction^[22,23]. The loss of SOCS1 expression might contribute to HCC progression *via* multiple mechanisms. We have shown that SOCS1 is an important regulator of oncogenic MET RTK signaling and suppresses the oncogenic potential of P21CIP in hepatocytes^[19,20,23]. These reports suggested that the loss of SOCS1 expression in HCC by epigenetic mechanisms could promote cancer cell survival, migration and proliferation. The findings of the present study indicate that the loss of SOCS1-dependent regulation of HGF/MET signaling may also contribute to invasive growth of HCC cells through, at least partly, *via* promoting EMT.

Aberrant MET signaling that facilitate tumor progression and metastasis occurs in many cancers mainly *via* receptor overexpression, gene amplification or through the loss of MET-targeting microRNAs, although activating mutations and increased autocrine HGF

stimulation can also amplify MET signaling^[14]. While gain-of-function mutations and gene amplification of MET are rare in HCC, MET overexpression has been frequently reported^[14]. The elevated MET expression would not only lead to ligand-independent activation but also confer increased responsiveness to des-gamma-carboxy prothrombin, which is highly expressed in HCC^[35]. The molecular basis of MET overexpression in HCC has not been elucidated yet^[14]. The high frequency of SOCS1 promoter methylation in HCC^[33,34] and the ability of SOCS1 to promote MET degradation by proteasomes^[20] suggest that MET overexpression in HCC could result at least partly from the loss of SOCS1 mediated control of MET expression.

The MET signaling pathway is an important mediator of EMT in HCC progression *via* intra- and extra- hepatic metastases^[36,37]. EMT is mediated *via* induction of transcription factors (*SNAI1*, *SNAI2*, *ZEB1*, *ZEB2*, *TWIST1*), which repress genes coding for proteins

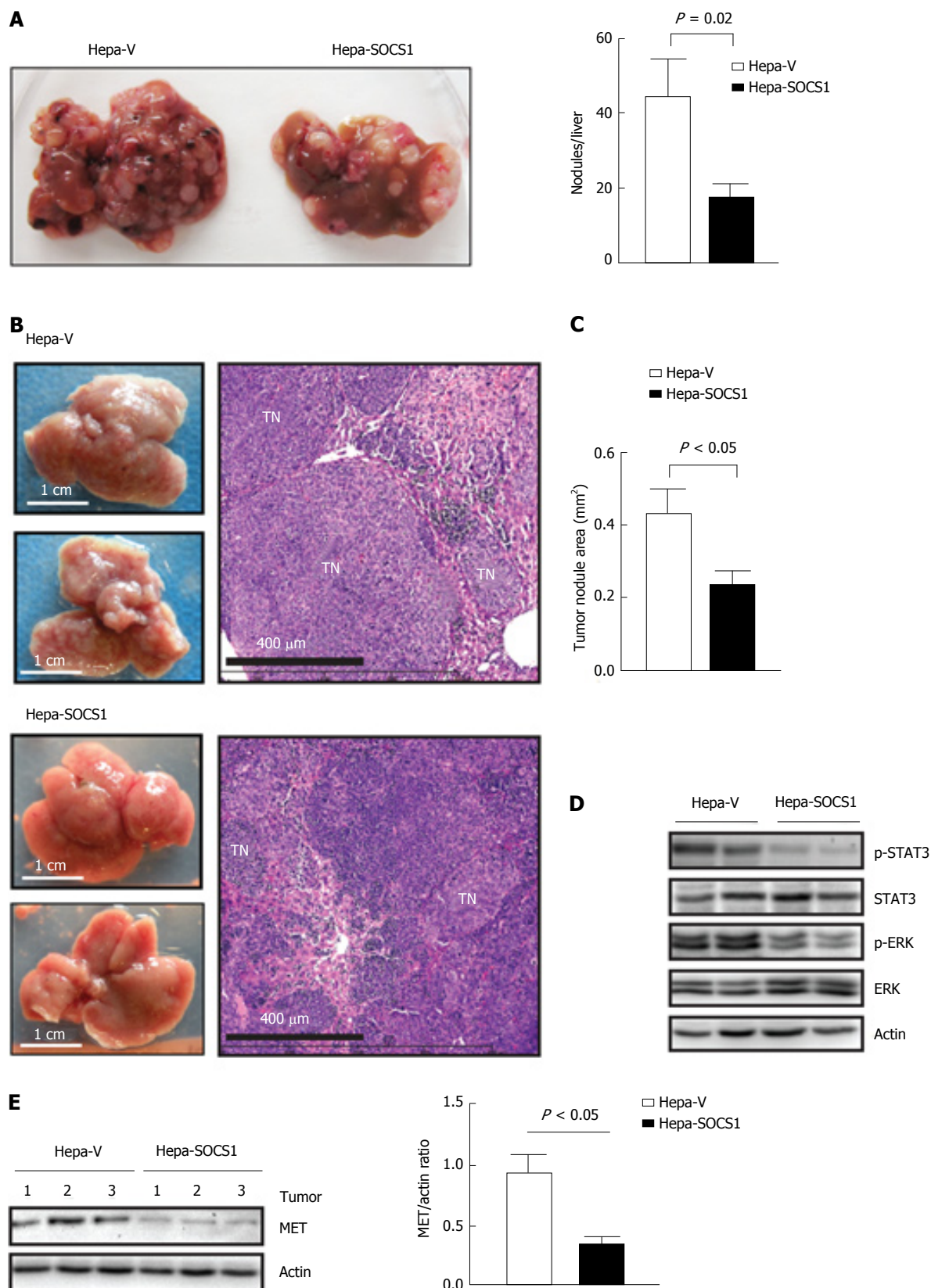


Figure 4 Suppressor of cytokine signaling 1 attenuates invasive growth of orthotopic Hepa cell tumors. A: Hepa-V and Hepa-SOCS1 cells were injected via intravenous route into NOD.*scid.gamma* (NSG) mice and the livers were examined macroscopically after 20 d. Representative images (left) and quantification of the liver tumor nodules (right) from four mice per group are shown; B-D: Hepa-vector and Hepa-SOCS1 were injected via intrasplenic route into NSG mice (1×10^6 cells per mouse; $n = 6$ per group from two separate experiments) and the mice were splenectomized. After 20 d, the liver tissues were examined macroscopically for tumor nodules. The dorsal and ventral views of representative livers (B, left) and H and E-stained sections of a representative liver for each group (B, right) are shown; C: For each group, the areas of thirty random tumor nodules from three different mice were measured digitally using the NDP software and compared by Student's paired *t* test; D: Western blot evaluation of STAT3 and ERK phosphorylation in the tumors from 2 mice per group; E: Western blot evaluation of MET expression and its quantification in the tumors from 3 mice per group. SOCS1: Suppressor of cytokine signaling 1.

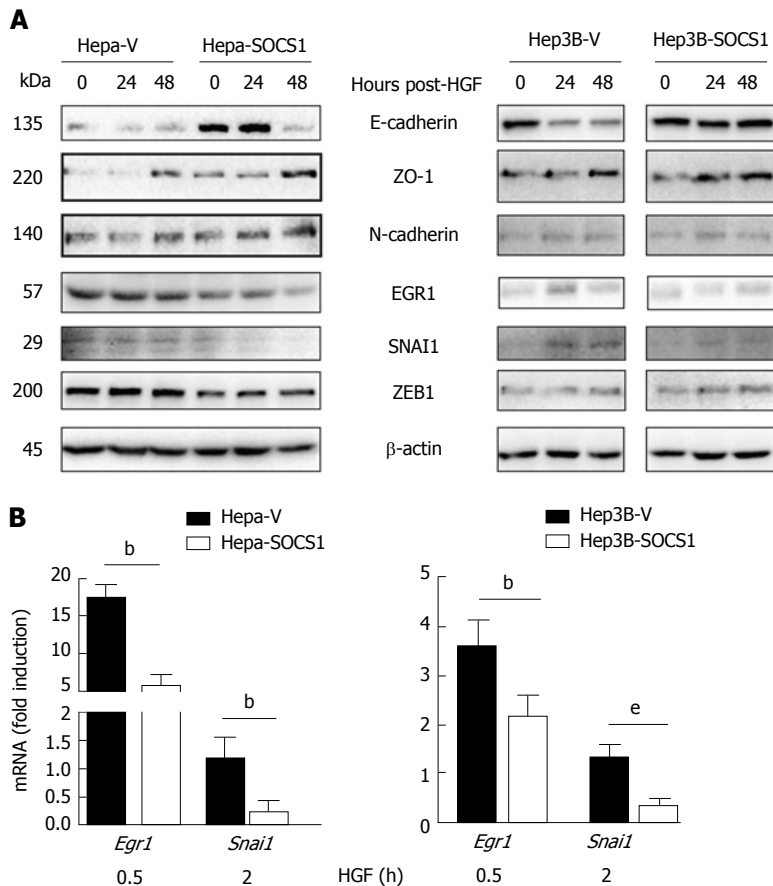


Figure 5 Effect of suppressor of cytokine signaling 1 on the expression of proteins modulated during epithelial-mesenchymal transition in hepatocyte growth factor-stimulated hepatocellular carcinoma cells. **A:** Hepa-V and Hepa-SOCS1, and Hep3B-V and Hep3B-SOCS1 were cultured in low-serum medium (0.25%) overnight and stimulated or not with m-HGF or h-HGF for 24 or 48 h. Unstimulated and HGF-stimulated cells were lysed in SDS sample buffer and analyzed by western blot for the indicated proteins. Representative data from two experiments are shown; **B:** Cells were stimulated with HGF for 30 min or 2 h and the expression of *Egr1* and *Snai1* genes was evaluated by qRT-PCR. Fold-induction relative to the housekeeping gene *Gapdh* was evaluated from triplicates of two separate experiments (mean \pm SE). ^b $P < 0.01$, ^e $P < 0.001$. SOCS1: Suppressor of cytokine signaling 1.

that maintain epithelial cell integrity^[12,13]. During EMT, progressive loss of epithelial markers (E-cadherin, occludin, ZO-1) and acquisition of mesenchymal markers (N-cadherin, vimentin) cause tumor cells to disrupt intercellular junctions, become motile, remodel cell-matrix interactions and become invasive. However, these changes may not be absolute and intermediate phenotypes may represent partial EMT^[37]. Reduced expression of E-cadherin occurs in 50%-60% of primary HCC that correlates with increased *SNAI1* expression and poor prognosis^[38]. Similar changes were reported in human and murine HCC cell lines with high invasive potential^[36,38]. Indeed, elevated MET expression has been associated with mesenchymal phenotype characterized by reduced expression of E-Cadherin and increased expression of the *CDH1* gene repressor *ZEB2* in human HCC cell lines MHCC97-L and MHCC97-H that display increased metastatic potential^[16]. Similar findings have been reported in murine HCC cell lines^[15]. A murine HCC line-derived circulating tumor cells isolated following intrahepatic implantation displayed elevated MET expression with reduced levels of E-Cadherin, implicating the EMT process in HCC dissemination^[39]. Our findings that

SOCS1 expression reduces HGF-induced invasive growth in HCC cells that is accompanied by increased expression of E-cadherin and discernibly reduced expression of *EGR1*, *SNAI1* and *ZEB1*, as well as the markedly diminished orthotopic growth of SOCS1-expressing HCC cells associated with reduced MET expression support the idea that SOCS1 is an important regulator of MET-mediated cancer progression *via* EMT. Thus, our findings add SOCS1 to the growing list of endogenous regulators of MET-mediated EMT, namely miR-148a, miR449a, SENP1^[17,30,40].

Molecular mechanisms underlying MET-mediated EMT are not yet fully elucidated. In HepG2 human HCC cells, HGF-induced cell scattering requires ERK activation, leading to induction of *EGR1* (Early growth response factor), a transcriptional activator of *SNAI1*, which represses *CDH1* encoding E-cadherin^[28]. The HGF-induced morphological and molecular changes of EMT in HCC cells could be reversed by blocking ERK activation using sorafenib that targets RTKs, or by U0126 that inhibits the ERK upstream kinase MEK1^[29]. Given that the MET signaling pathways are implicated in multiple aspects of cancer progression, attenuating MET signaling will also disrupt MET-mediated EMT^[37].

Several therapeutics targeting the MET signaling pathway including multikinase inhibitors, selective MET inhibitors, antibodies targeting HGF or MET, agents that neutralize HGF and molecules that disrupt MET signaling^[14]. Given the possible emergence of drug resistance to agents that target signaling pathways, and that MET can be activated by alternate ligands such as des-gamma-carboxy prothrombin and heparin^[35,41], alternate strategies are needed to effectively target MET-mediated HCC invasion and metastasis. In this context, anti-MET Ab such as DN-30 that promotes MET degradation^[42] and strategies to disrupt MET receptor expression^[14] could represent promising approaches. However, this would require identification of HCC patients who would benefit from aggressive MET-targeting therapy^[14]. Although MET overexpression could be a direct marker, clinical grade MET Ab are still under development^[43]. Given that SOCS1 is an important regulator of MET protein expression, and that loss of SOCS1 expression predominantly occur by promoter methylation in HCC, assessing SOCS1 methylation status could be a useful approach to identify HCC patients who would benefit from MET-targeting therapies.

In conclusion, our findings show that SOCS1 attenuates migration and invasion properties of HCC cells at least partly *via* modulation of MET-mediated EMT, and controls invasive tumor growth. Based on these findings, we propose that the *SOCS1* gene methylation and expression in primary HCC may serve as useful markers to identify patients for targeted MET therapy.

COMMENTS

Background

Invasive intrahepatic dissemination is a key factor in malignant growth of hepatocellular carcinoma (HCC) and its poor prognosis. HCC patients also present extrahepatic metastasis that may occur *via* direct invasion. Cancer cells gain invasive potential through epithelial-mesenchymal transition (EMT) induced by cytokines and growth factors that stimulate receptor tyrosine kinases (RTK). An important RTK implicated in EMT is the HGF receptor MET. Suppressor of cytokine signaling 1 (SOCS1) is an important regulator of MET signaling in hepatocytes.

Research frontiers

Emerging data implicate EMT in cancer growth and metastasis in HCC and many other cancers. Among the strategies to target EMT, the MET signaling pathway holds promise. The present study investigates the role of SOCS1 in regulating MET signaling involved in the invasive tumor growth of HCC.

Innovations and breakthroughs

This study shows that SOCS1 inhibits the invasive growth of HCC cells *in vitro* and *in vivo*, and adds SOCS1 to the growing list of endogenous regulators of MET-mediated EMT.

Applications

SOCS1 expression in HCC biopsies could be a useful biomarker of invasive and metastatic potential to identify patients suitable for treatment with MET-targeting therapies. Development of therapeutics to restore SOCS1 expression in cancer cells could be another approach to attenuate the invasive potential and metastatic growth of HCC.

Terminology

SOCS1 is a negative feedback regulator of cytokine and growth factor signaling. SOCS1 inhibits HGF-induced MET signaling pathways that promote invasive growth, which can be assessed *in vitro* using a 3-dimensional invasion assay and *in vivo* by orthotopic tumor growth.

Peer-review

In this study, the authors investigated the role of SOCS1 in the invasion ability of HCC. Extending the findings of their previous work, the authors show that SOCS1 inhibits the invasive growth of HCC cells by targeting MET signaling.

REFERENCES

- 1 Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; **65**: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 2 El-Serag HB, Marrero JA, Rudolph L, Reddy KR. Diagnosis and treatment of hepatocellular carcinoma. *Gastroenterology* 2008; **134**: 1752-1763 [PMID: 18471552 DOI: 10.1053/j.gastro.2008.02.090]
- 3 Lachenmayer A, Alsinet C, Chang CY, Llovet JM. Molecular approaches to treatment of hepatocellular carcinoma. *Dig Liver Dis* 2010; **42** Suppl 3: S264-S272 [PMID: 20547313]
- 4 Llovet JM, Villanueva A, Lachenmayer A, Finn RS. Advances in targeted therapies for hepatocellular carcinoma in the genomic era. *Nat Rev Clin Oncol* 2015; **12**: 408-424 [PMID: 26054909 DOI: 10.1038/nrcclinonc.2015.103]
- 5 Tang ZY. Hepatocellular carcinoma--cause, treatment and metastasis. *World J Gastroenterol* 2001; **7**: 445-454 [PMID: 11819809 DOI: 10.3748/wjg.v7.i4.445]
- 6 Hu L, Lau SH, Tzang CH, Wen JM, Wang W, Xie D, Huang M, Wang Y, Wu MC, Huang JF, Zeng WF, Sham JS, Yang M, Guan XY. Association of Vimentin overexpression and hepatocellular carcinoma metastasis. *Oncogene* 2004; **23**: 298-302 [PMID: 14647434]
- 7 Nakashima T, Okuda K, Kojiro M, Jimi A, Yamaguchi R, Sakamoto K, Ikari T. Pathology of hepatocellular carcinoma in Japan. 232 Consecutive cases autopsied in ten years. *Cancer* 1983; **51**: 863-877 [PMID: 6295617]
- 8 Kummer S, Shafi NQ. Metastatic hepatocellular carcinoma. *Clin Oncol (R Coll Radiol)* 2003; **15**: 288-294 [PMID: 12924460]
- 9 Kim HS, Shin JW, Kim GY, Kim YM, Cha HJ, Jeong YK, Jeong ID, Bang SJ, Kim DH, Park NH. Metastasis of hepatocellular carcinoma to the small bowel manifested by intussusception. *World J Gastroenterol* 2006; **12**: 1969-1971 [PMID: 16610010 DOI: 10.3748/wjg.v12.i12.1969]
- 10 Korkolis DP, Aggeli C, Plataniotis GD, Gontikakis E, Zerbinis H, Papantoniou N, Xinopoulos D, Apostolikas N, Vassilopoulos PP. Successful en bloc resection of primary hepatocellular carcinoma directly invading the stomach and pancreas. *World J Gastroenterol* 2009; **15**: 1134-1137 [PMID: 19266609 DOI: 10.3748/wjg.15.1134]
- 11 Polyak K, Weinberg RA. Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer* 2009; **9**: 265-273 [PMID: 19262571 DOI: 10.1038/nrc2620]
- 12 Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest* 2009; **119**: 1420-1428 [PMID: 19487818 DOI: 10.1172/JCI39104]
- 13 Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* 2014; **15**: 178-196 [PMID: 24556840 DOI: 10.1038/nrm3758]
- 14 Giordano S, Columbano A. Met as a therapeutic target in HCC: facts and hopes. *J Hepatol* 2014; **60**: 442-452 [PMID: 24045150 DOI: 10.1016/j.jhep.2013.09.009]
- 15 Ogunwobi OO, Liu C. Hepatocyte growth factor upregulation promotes carcinogenesis and epithelial-mesenchymal transition in hepatocellular carcinoma via Akt and COX-2 pathways. *Clin Exp Metastasis* 2011; **28**: 721-731 [PMID: 21744257 DOI: 10.1007/s10585-011-9404-x]
- 16 You H, Ding W, Dang H, Jiang Y, Rountree CB. c-Met represents a potential therapeutic target for personalized treatment in

- hepatocellular carcinoma. *Hepatology* 2011; **54**: 879-889 [PMID: 21618573 DOI: 10.1002/hep.24450]
- 17 **Zhang JP**, Zeng C, Xu L, Gong J, Fang JH, Zhuang SM. MicroRNA-148a suppresses the epithelial-mesenchymal transition and metastasis of hepatoma cells by targeting Met/Snail signaling. *Oncogene* 2014; **33**: 4069-4076 [PMID: 24013226 DOI: 10.1038/onc.2013.369]
 - 18 **Chen SP**, Liu BX, Xu J, Pei XF, Liao YJ, Yuan F, Zheng F. MiR-449a suppresses the epithelial-mesenchymal transition and metastasis of hepatocellular carcinoma by multiple targets. *BMC Cancer* 2015; **15**: 706 [PMID: 26471185 DOI: 10.1186/s12885-015-1738-3]
 - 19 **Gui Y**, Yeganeh M, Ramanathan S, Leblanc C, Pomerleau V, Ferbeyre G, Saucier C, Ilangumaran S. SOCS1 controls liver regeneration by regulating HGF signaling in hepatocytes. *J Hepatol* 2011; **55**: 1300-1308 [PMID: 21703184]
 - 20 **Gui Y**, Yeganeh M, Donates YC, Tobelaim WS, Chababi W, Mayhue M, Yoshimura A, Ramanathan S, Saucier C, Ilangumaran S. Regulation of MET receptor tyrosine kinase signaling by suppressor of cytokine signaling 1 in hepatocellular carcinoma. *Oncogene* 2015; **34**: 5718-5728 [PMID: 25728680 DOI: 10.1038/onc.2015.20]
 - 21 **Yoshikawa H**, Matsubara K, Qian GS, Jackson P, Groopman JD, Manning JE, Harris CC, Herman JG. SOCS-1, a negative regulator of the JAK/STAT pathway, is silenced by methylation in human hepatocellular carcinoma and shows growth-suppression activity. *Nat Genet* 2001; **28**: 29-35 [PMID: 11326271]
 - 22 **Yoshida T**, Ogata H, Kamio M, Joo A, Shiraishi H, Tokunaga Y, Sata M, Nagai H, Yoshimura A. SOCS1 is a suppressor of liver fibrosis and hepatitis-induced carcinogenesis. *J Exp Med* 2004; **199**: 1701-1707 [PMID: 15197228]
 - 23 **Yeganeh M**, Gui Y, Kandhi R, Bobbala D, Tobelaim WS, Saucier C, Yoshimura A, Ferbeyre G, Ramanathan S, Ilangumaran S. Suppressor of cytokine signaling 1-dependent regulation of the expression and oncogenic functions of p21(CIP1/WAF1) in the liver. *Oncogene* 2016; **35**: 4200-4211 [PMID: 26725321 DOI: 10.1038/onc.2015.485]
 - 24 **Bernier J**, Chababi W, Pomerleau V, Saucier C. Oncogenic engagement of the Met receptor is sufficient to evoke angiogenic, tumorigenic, and metastatic activities in rat intestinal epithelial cells. *Am J Physiol Gastrointest Liver Physiol* 2010; **299**: G677-G686 [PMID: 20539003 DOI: 10.1152/ajpgi.00315.2009]
 - 25 **Francone TD**, Landmann RG, Chen CT, Sun MY, Kuntz EJ, Zeng Z, Dematteo RP, Paty PB, Weiser MR. Novel xenograft model expressing human hepatocyte growth factor shows ligand-dependent growth of c-Met-expressing tumors. *Mol Cancer Ther* 2007; **6**: 1460-1466 [PMID: 17431125 DOI: 10.1158/1535-7163.MCT-06-0466]
 - 26 **Kazi JU**, Kabir NN, Flores-Morales A, Rönstrand L. SOCS proteins in regulation of receptor tyrosine kinase signaling. *Cell Mol Life Sci* 2014; **71**: 3297-3310 [PMID: 24705897 DOI: 10.1007/s00018-014-1619-y]
 - 27 **Trengove MC**, Ward AC. SOCS proteins in development and disease. *Am J Clin Exp Immunol* 2013; **2**: 1-29 [PMID: 23885323]
 - 28 **Grotegut S**, von Schweinitz D, Christofori G, Lehenbre F. Hepatocyte growth factor induces cell scattering through MAPK/Egr-1-mediated upregulation of Snail. *EMBO J* 2006; **25**: 3534-3545 [PMID: 16858414 DOI: 10.1038/sj.emboj.7601213]
 - 29 **Nagai T**, Arao T, Furuta K, Sakai K, Kudo K, Kaneda H, Tamura D, Aomatsu K, Kimura H, Fujita Y, Matsumoto K, Saijo N, Kudo M, Nishio K. Sorafenib inhibits the hepatocyte growth factor-mediated epithelial mesenchymal transition in hepatocellular carcinoma. *Mol Cancer Ther* 2011; **10**: 169-177 [PMID: 21220499 DOI: 10.1158/1535-7163.MCT-10-0544]
 - 30 **Zhang W**, Sun H, Shi X, Wang H, Cui C, Xiao F, Wu C, Guo X, Wang L. SENP1 regulates hepatocyte growth factor-induced migration and epithelial-mesenchymal transition of hepatocellular carcinoma. *Tumour Biol* 2016; **37**: 7741-7748 [PMID: 26695141 DOI: 10.1007/s13277-015-4406-y]
 - 31 **Yang B**, Guo M, Herman JG, Clark DP. Aberrant promoter methylation profiles of tumor suppressor genes in hepatocellular carcinoma. *Am J Pathol* 2003; **163**: 1101-1107 [PMID: 12937151]
 - 32 **Nomoto S**, Kinoshita T, Kato K, Otani S, Kasuya H, Takeda S, Kanazumi N, Sugimoto H, Nakao A. Hypermethylation of multiple genes as clonal markers in multicentric hepatocellular carcinoma. *Br J Cancer* 2007; **97**: 1260-1265 [PMID: 17968429 DOI: 10.1038/sj.bjc.6604016]
 - 33 **Zhang C**, Li J, Huang T, Duan S, Dai D, Jiang D, Sui X, Li D, Chen Y, Ding F, Huang C, Chen G, Wang K. Meta-analysis of DNA methylation biomarkers in hepatocellular carcinoma. *Oncotarget* 2016; **7**: 81255-81267 [PMID: 27835605 DOI: 10.18632/oncotarget.13221]
 - 34 **Liu M**, Cui LH, Li CC, Zhang L. Association of APC, GSTP1 and SOCS1 promoter methylation with the risk of hepatocellular carcinoma: a meta-analysis. *Eur J Cancer Prev* 2015; **24**: 470-483 [PMID: 25853848 DOI: 10.1097/CEJ.0000000000000121]
 - 35 **Suzuki M**, Shiraha H, Fujikawa T, Takaoka N, Ueda N, Nakanishi Y, Koike K, Takaki A, Shiratori Y. Des-gamma-carboxy prothrombin is a potential autologous growth factor for hepatocellular carcinoma. *J Biol Chem* 2005; **280**: 6409-6415 [PMID: 15582995 DOI: 10.1074/jbc.M406714200]
 - 36 **Ding W**, You H, Dang H, LeBlanc F, Galicia V, Lu SC, Stiles B, Rountree CB. Epithelial-to-mesenchymal transition of murine liver tumor cells promotes invasion. *Hepatology* 2010; **52**: 945-953 [PMID: 20564331 DOI: 10.1002/hep.23748]
 - 37 **Giannelli G**, Koudelkova P, Dituri F, Mikulits W. Role of epithelial to mesenchymal transition in hepatocellular carcinoma. *J Hepatol* 2016; **65**: 798-808 [PMID: 27212245 DOI: 10.1016/j.jhep.2016.05.007]
 - 38 **Yang MH**, Chen CL, Chau GY, Chiou SH, Su CW, Chou TY, Peng WL, Wu JC. Comprehensive analysis of the independent effect of twist and snail in promoting metastasis of hepatocellular carcinoma. *Hepatology* 2009; **50**: 1464-1474 [PMID: 19821482 DOI: 10.1002/hep.23221]
 - 39 **Ogunwobi OO**, Puszyk W, Dong HJ, Liu C. Epigenetic upregulation of HGF and c-Met drives metastasis in hepatocellular carcinoma. *PLoS One* 2013; **8**: e63765 [PMID: 23723997 DOI: 10.1371/journal.pone.0063765]
 - 40 **Ozen E**, Gozukizil A, Erdal E, Uren A, Bottaro DP, Atabey N. Heparin inhibits Hepatocyte Growth Factor induced motility and invasion of hepatocellular carcinoma cells through early growth response protein 1. *PLoS One* 2012; **7**: e42717 [PMID: 22912725 DOI: 10.1371/journal.pone.0042717]
 - 41 **İşcan E**, Güneş A, Korhan P, Yılmaz Y, Erdal E, Atabey N. The regulatory role of heparin on c-Met signaling in hepatocellular carcinoma cells. *J Cell Commun Signal* 2017; **11**: 155-166 [PMID: 27975162 DOI: 10.1007/s12079-016-0368-0]
 - 42 **Pacchiana G**, Chiriaco C, Stella MC, Petronzelli F, De Santis R, Galluzzo M, Carminati P, Comoglio PM, Michieli P, Vigna E. Monovalency unleashes the full therapeutic potential of the DN-30 anti-Met antibody. *J Biol Chem* 2010; **285**: 36149-36157 [PMID: 20833723 DOI: 10.1074/jbc.M110.134031]
 - 43 **Knudsen BS**, Zhao P, Resau J, Cottingham S, Gherardi E, Xu E, Berghuis B, Daugherty J, Grabinski T, Toro J, Giambernardi T, Skinner RS, Gross M, Hudson E, Kort E, Lengyel E, Ventura A, West RA, Xie Q, Hay R, Vande Woude G, Cao B. A novel multipurpose monoclonal antibody for evaluating human c-Met expression in preclinical and clinical settings. *Appl Immunohistochem Mol Morphol* 2009; **17**: 57-67 [PMID: 18815565 DOI: 10.1097/PAI.0b013e3181816ae2]

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Basic Study

Protective effects of oral glutathione on fasting-induced intestinal atrophy through oxidative stress

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Abstract

AIM

To determine whether oral glutathione (GSH) administration can alleviate the effects of fasting-induced intestinal atrophy in the small intestinal mucosa.

METHODS

Rats were divided into eight groups. One group was fed *ad libitum*, another was fed *ad libitum* and received oral GSH, and six groups were administered saline (SA) or GSH orally during fasting. Mucosal height, apoptosis, and cell proliferation in the jejunum were histologically evaluated. iNOS protein expression (by immunohistochemistry), nitrite levels (by high performance liquid chromatography, as a measure of NO production), 8-hydroxydeoxyguanosine formation (by ELISA, indicating ROS levels), glutathione/oxidized glutathione (GSH/GSSG) ratio (by enzymatic colorimetric detection), and γ -glutamyl transpeptidase (*Ggt1*) mRNA levels in the jejunum (by semi-quantitative RT-PCR) were also estimated.

RESULTS

Oral GSH administration was demonstrated to drastically reduce fasting-induced intestinal atrophy in the jejunum. In particular, jejunal mucosal height was enhanced in GSH-treated animals compared to SA-treated animals [527.2 ± 6.9 for 50 mg/kg GSH, 567.6 ± 5.4 for 500 mg/kg GSH *vs* 483.1 ± 4.9 (μm), $P < 0.01$ at 72 h]. This effect was consistent with decreasing changes in GSH-treated animals compared to SA-treated animals for iNOS protein staining [0.337 ± 0.016 for 50 mg/kg GSH, 0.317 ± 0.017 for 500 mg/kg GSH *vs* 0.430 ± 0.023 (area of staining part/area of tissue), $P < 0.01$ at 72 h] and NO [2.99 ± 0.29 for 50 mg/kg GSH, 2.88 ± 0.19 for 500 mg/kg GSH *vs* 5.34 ± 0.35 (nmol/g tissue), $P < 0.01$ at 72 h] and ROS [3.92 ± 0.46 for 50 mg/kg GSH, 4.58 ± 0.29 for 500 mg/kg GSH *vs* 6.42 ± 0.52 (8-OHdG pg/ μg DNA), $P < 0.01$, $P < 0.05$ at 72 h, respectively] levels as apoptosis mediators in the jejunum. Furthermore, oral GSH administration attenuated cell proliferation decreases in the fasting jejunum [182.5 ± 1.9 for 500 mg/kg GSH *vs* 155.8 ± 3.4 (5-BrdU positive cells/10 crypts), $P < 0.01$ at 72 h]. Notably, both GSH concentration and Ggt1 mRNA expression in the jejunum were also attenuated in rats following oral administration of GSH during fasting as compared with fasting alone [0.45 ± 0.12 *vs* 0.97 ± 0.06 (nmol/mg tissue), $P < 0.01$; 1.01 ± 0.11 *vs* 2.79 ± 0.39 (Ggt1 mRNA/Gapdh mRNA), $P < 0.01$ for 500 mg/kg GSH at 48 h, respectively].

CONCLUSION

Oral GSH administration during fasting enhances jejunal regenerative potential to minimize intestinal mucosal atrophy by diminishing fasting-mediated ROS generation and enterocyte apoptosis and enhancing cell proliferation.

Key words: Intestinal atrophy; Glutathione; Apoptosis; Cell proliferation; Inducible nitric oxide synthase

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Core tip: We have previously demonstrated that the intestinal mucosal atrophy consequent to fasting is due in large part to increased apoptosis in jejunal villi and decreased cell proliferation in jejunal crypts, with concomitant increased reactive oxygen species (ROS) and NO production and decreased glutathione (GSH). Here we demonstrate protection against fasting-induced intestinal mucosal atrophy by minimizing ROS induction in the intestinal mucosa through supplemental oral administration of an antioxidant such as GSH during fasting in rats. In particular, oral GSH administration during fasting enhances jejunal regenerative potential by diminishing enterocyte apoptosis and enhancing cell proliferation.

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induced intestinal atrophy through oxidative stress. *World J Gastroenterol* 2017; 23(36): 6650-6664 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6650.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6650>

INTRODUCTION

Fasting induces small intestinal mucosal atrophy including increased epithelial permeability and compromised tight junctions, which can lead to bacterial translocation, particularly in patients receiving a prolonged course of total parenteral nutrition (TPN)^[1,2]. Many surgeons involved in nutritional support therapy desire the identification of bioactive substances that may be efficacious in protecting against intestinal injury during long-term TPN administration, particularly with respect to intestinal barrier function and subsequent septic complications. Fasting and other states of malnutrition are associated with increased reactive oxygen species (ROS) formation, which has been implicated in the loss of intestinal mucosal structure and function under conditions of inflammation, injury, and shock^[3,4]. In addition, these fasting states are also accompanied by depletion of the critical antioxidant glutathione (GSH), which functions to eliminate induced ROS in the intestinal mucosa^[5-7]. Therefore, GSH is the most prevalent and important low-molecular-weight thiol in mammalian tissues^[8]. As GSH deficiency in tissues is associated with increased oxidative stress, aging, and increased risk of numerous chronic diseases as well as fasting^[9], the maintenance of tissue levels of GSH is critical for maintaining health and preventing disease and age-related biological insults.

GSH sources in the intestinal mucosa include intracellular synthesis, biliary supply, and dietary intake. The intestinal lumen receives a large quantity of hepatic GSH from biliary secretion^[10] and dietary GSH from fresh fruits, vegetables, and many types of meat^[11]. As the only enzyme of the γ -glutamyl cycle located on the outer surface of the plasma membrane, γ -glutamyl transpeptidase (GGT) plays a key role in GSH homeostasis by breaking down extracellular GSH and providing cysteine, the rate-limiting substrate for intracellular *de novo* GSH synthesis^[12]. The two intracellular enzymes, γ -glutamylcysteine synthetase and GSH synthetase, catalyze intracellular GSH synthesis from glutamate, cysteine, and glycine, which are translocated into cells by amino acid transporters on the surface of the plasma membrane following the resolution of extracellular GSH by GGT. Intracellular GSH plays an important role in antioxidant defense and the regulation of pathways essential for *in vivo* homeostasis through its catalysis by glutathione S-transferase (GST) and glutathione peroxidase^[9]. Studies of oral GSH supplementation in humans and laboratory animals have shown that the enhancement of intestinal mucosal GSH levels by oral GSH

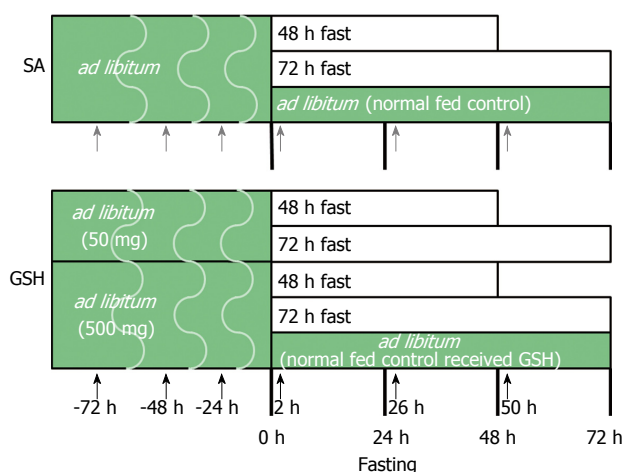


Figure 1 Experimental design. Rats were divided into eight groups, one of which was fed *ad libitum*, another of which was fed *ad libitum* and received oral glutathione (GSH), and six of which were administered saline (SA) or GSH orally prior to and after fasting. The large block arrows represent treatment with SA (closed gray arrow) and GSH (closed block arrow).

supplementation under conditions in which intracellular GSH status is compromised (*i.e.*, GSH deficiency-related biological insults) can restore tissue GSH and promote ROS metabolism^[10,13,14]. Thus, it has been described that orally administered GSH acts as backup for GSH-deficient tissue.

As an alternative, the ability to control extracellular redox provides another possible role for intestinal luminal GSH. Recent studies have shown that the thiol/disulfide redox status in the extracellular compartment regulates the ROS that are generated primarily in the intracellular compartment and serve an important function in the activation of proteins and up-regulation of antioxidant and detoxification systems^[15-17]. Furthermore, many proteins including transporters, receptors, and enzymes present on the surface of the plasma membrane and located in extracellular fluids contain cysteine and methionine residues that are subject to oxidation. Accordingly, these proteins respond to variations in the extracellular thiol/disulfide redox environment, which can alter their activity^[18]. Notably, recent demonstrations that treatment with GSH leads to a significant increase in the number of free sulfhydryls on the cell surface have led to the conclusion that extracellular GSH modulates the interaction between bacteria and epithelial respiratory cells and inhibits bacterial invasion into these cells^[19].

We previously suggested that fasting for 2-3 d causes intestinal mucosal atrophy resulting from increased apoptosis in jejunal villi, nitric oxide (NO) and ROS production following elevated inducible nitric oxide synthase (iNOS) expression, and decreased cell proliferation in jejunal crypts^[20]. Conversely, these apoptosis mediators are all suppressed by treatment with aminoguanidine, a selective iNOS inhibitor, with consequent mucosal recovery, suggesting that iNOS, which induces the production of ROS, is likely to

constitute a significant upstream factor promoting fasting-induced apoptosis in enterocytes. Furthermore, refeeding repaired fasting-induced jejunal atrophy by inhibiting these apoptosis mediators, including ROS, and also showed an association between ROS inhibition and increased cell proliferation for decreasing intestinal mucosal atrophy^[21]. Therefore, we hypothesized that protection against intestinal mucosal atrophy might be provided through the elimination of induced ROS in the intestinal mucosa by introducing an antioxidant such as GSH during a fasting period in rats.

Therefore, the objective of the present study was to investigate the effects of oral GSH administration on the manifestation of fasting-induced intestinal atrophy through the reduction of oxidative stress, with a particular focus on the possible participation of intracellular GSH levels and GGT expression on the cell surface as parameters for *de novo* intracellular GSH synthesis.

MATERIALS AND METHODS

Animals and experimental design

The experimental protocol and design were approved by the Institutional Animal Care and Use Committee at the Life Science Center of Josai University. Male Wistar rats at 9 wk of age were purchased from SLC (Shizuoka, Japan) and housed in wire-bottomed cages. The rats were placed in a room illuminated from 7:00 am to 7:00 pm (12:12-h light:dark cycle). The animals were allowed free access to deionized water and standard rat chow (CE-2, CLEA Japan) until the study began. At 10 wk of age, 59 rats were randomly divided into eight groups (Figure 1). Each group received saline (SA) or GSH (50 or 500 mg/kg b.w./d) using oral gavage needles at 24, 48, and 72 h before fasting, including a normally fed control group and a normally fed control that received GSH, with the following fasting durations in each group: (1) SA + *ad libitum* (normal fed control); (2) SA + 48-h fast; (3) SA + 72-h fast; (4) 500 mg/kg GSH + *ad libitum* (normal fed control received GSH); (5) 50 mg/kg GSH + 48-h fast; (6) 50 mg/kg GSH + 72-h fast; (7) 500 mg/kg GSH + 48-h fast; and (8) 500 mg/kg GSH + 72-h fast. All rats received SA or GSH by oral gavage for 2, 26, and 50 h after fasting. The GSH (Setria® reduced glutathione) used in this work was provided by Kyowa Hakko Bio Co., Ltd. The rats were weighed daily.

Collection of intestinal mucosa

After fasting, the rats were anesthetized and then euthanized by exsanguination. The entire small intestine was carefully removed and placed on ice. Then, 10 cm of tissue from the oral (duodenum) side was removed and the remainder of the intestine was divided into two segments: proximal (jejunum) and distal (ileum). The segments used in analyses comprised the jejunum from 3 to 5 cm distal to the duodenum^[22]. Samples with a

length of approximately 3 cm were fixed in 10% neutral buffered formalin for measurement of mucosal height and immunohistochemistry. The remaining segments were snap-frozen in liquid nitrogen and stored at -80 °C.

Histopathological analysis of mucosal height and apoptotic index

Fixed tissue samples were embedded in paraffin and sectioned (2-3 µm thickness) prior to being stained with hematoxylin and eosin (H&E). Mucosal height (villous height plus crypt depth) was measured using a microscope (BX41; Olympus, Tokyo, Japan) and a digital camera system (Penguin 150CL; Pixera, San Jose, CA, United States). Mucosal height was measured for at least 30 villi per animal.

To detect enterocyte apoptosis in the jejunum, apoptotic index (AI) using conventional light microscopy of H&E-stained specimens was used. We followed previously published methods^[20,23]. In brief, jejunal sections as used above for histopathological analysis were examined in a blinded manner for the typical attributes of apoptotic cells. We assessed 50 villus-crypt columns per rat. For each column, the number and position of apoptotic cells and the total number of cells were recorded. To account for the effects of fasting and GSH treatment on apoptosis, the average number of apoptotic cells in villi and crypts was determined along with AI. To identify the locations of apoptosis along the villi and crypts, AI distribution curves were constructed on the basis of group means, in which cell position vs AI was plotted at each position. Here, AI was defined as the total number of apoptotic cells at each cell position and is expressed as the percentage of the total number of cells counted at that cell position. Cell position 1 was set as the cell at the crypt-villus junction and the cell at the base of the crypt column for villus and crypt data, respectively.

Immunohistochemical assessment of iNOS expression and 5-bromo-2'-deoxyuridine (5-BrdU)-positive cells

Immunohistochemical staining was performed using a rabbit anti-iNOS polyclonal antibody^[24]. The specimens were dewaxed and treated for antigen retrieval by boiling in 10 mmol/L citrate buffer (pH 6.0)^[25]. After being washed with PBS, the specimens were incubated in 6% hydrogen peroxide and nonspecific binding was blocked with a 20% goat serum solution in PBS. Specimens were subsequently incubated with an anti-iNOS primary antibody (1:100; BD Transduction Laboratories, Lexington, KY, United States), except for the control sections, for which no primary antibody was used. A biotinylated goat anti-rabbit IgG (1:200; Vector Laboratories, Burlingame, CA, United States) was used as a secondary antibody. The sections were then treated using the VECTASTAIN Elite ABC Kit (Vector Laboratories), and reaction products were detected using color development with diaminobenzidine. Finally, the sections were counterstained with hematoxylin

and examined under a light microscope and a digital camera system. For each rat, 5 clearly dyed sections were chosen randomly and 5 random fields for each section were assessed (at 40 × magnification). The content of iNOS was quantitatively measured based on the average optical density using a digital camera and ImageJ software (National Institutes of Health, Bethesda, Maryland, United States)^[26].

To assess cell proliferation in the crypt, the cell proliferation index was determined using conventional light microscopy of specimens immunohistochemically stained for 5-BrdU^[27]. Rats were intraperitoneally injected with 100 mg/kg 5-BrdU prior to euthanasia. After paraffin embedding and sectioning, tissue sections were dewaxed and immersed in 3% hydrogen peroxide-methanol solution. The specimens were washed with PBS and denatured in 2 N hydrochloric acid. Following further PBS washing, the specimens were immersed in 0.1 mol/L boric acid buffer (pH 8.5), incubated with 20 µg/mL proteinase K at 37 °C, and then the reaction was terminated with PBS containing bovine serum albumin. The sections were incubated with a mouse anti-5-BrdU monoclonal antibody (1:50; Chemicon International, Billerica, MA, United States), with the exception of control samples, for which the primary antibody was omitted. A biotinylated goat anti-mouse IgG (1:200; Vector Laboratories) was used as a secondary antibody. The sections were then treated using the VECTASTAIN Elite ABC Kit (Vector Laboratories) and antibody binding was detected by color development after addition of diaminobenzidine. Finally, the sections were counterstained with hematoxylin and examined under a light microscope with a digital camera system. The number of labeled cells in at least 10 well-oriented longitudinal crypts was determined for each rat. Results are expressed as the number of 5-BrdU-labeled cells per crypt.

Jejunal nitrite concentrations

Nitrite concentrations in the jejunum were measured using a dedicated high-performance liquid chromatography (HPLC) system (ENO-20; EiCom, Kyoto, Japan)^[28]. Jejunal segments were homogenized with an equal volume of methanol and centrifuged for deproteinization at 12000 g at 4 °C for 5 min. The samples were then applied to the HPLC system. The nitrites and nitrates were separated using a reverse-phase column (NO-PAK; EiCom), after which nitrate was reduced to nitrite in a reduction column packed with copperized cadmium (NO-RED; EiCom). These nitrites were then mixed with the Griess reagent in a reaction coil and the change in absorbance was monitored at 540 nm.

DNA oxidation analysis

Oxidative stress in the jejunum was evaluated by quantifying 8-hydroxydeoxyguanosine (8-OHdG) present in DNA^[29]. 8-OHdG is a product of oxidative

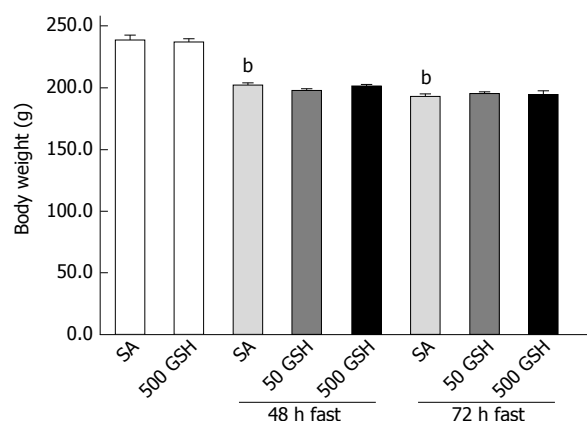


Figure 2 Effects of fasting and glutathione treatment on body weight. Fasting caused gradual decreases in body weight in both SA- and GSH-treated groups. There was no difference in weight loss between the GSH treatments and the respective SA-treated groups in each fasting period. Values represent the mean \pm SE. ^b $P < 0.01$ vs the normally fed control. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

DNA damage following specific enzymatic cleavage after 8-hydroxylation of the guanine base and is thought to constitute a marker of oxidative DNA damage, reflecting the DNA repair rate^[30]. Jejunal DNA was purified using the DNA Extractor TIS Kit (Wako Pure Chemical Industries Ltd., Osaka, Japan)^[31]. DNA samples were hydrolyzed to nucleosides by sequential incubation with 6 U of nuclease P1 (Wako) followed by 2 U of alkaline phosphatase (Wako). Hydrolysates were filtered through a VIVASPIN 500 MWCO 10000 filter (Sartorius Stedim Biotech, Gottingen, Germany). The levels of 8-OHdG in the filtered samples were determined using an ELISA kit (Japan Institute for the Control of Aging, Shizuoka, Japan).

Jejunal GSH and GSSG concentrations

GSH and GSSG concentrations were measured by using a GSSG/GSH Quantification Kit (Dojindo Molecular Technologies, Inc., Rockville, MD, United States). Jejunal segments were homogenized with 5% sodium sulfosalicylate (Wako) and centrifuged to remove proteins. For each sample, the supernatant was added to a well to which coenzyme working solution and enzyme working solution had been previously added, and incubated at room temperature. Then, the substrate working solution was added to the well and it was incubated at room temperature. The absorbance of samples and the GSH standard was measured at 405 nm using a microplate reader (TECAN GENios, Grodig, Austria). For measurement of GSSG, the supernatant was treated with masking solution and then added to a well as described above. The absorbance of samples and standard GSSG was measured at 405 nm.

Analysis of *Ggt1* mRNA expression by semi-quantitative reverse transcription-PCR

Total RNA was purified using the TaKaRa RNAiso Reagent (TaKaRa Bio, Kusatsu, Japan). Reverse transcription (RT)-

PCR was performed with total RNA using the RNA PCR kit (AMV) Version 3.0 (TaKaRa Bio): 1 cycle at 42 °C for 30 min, 99 °C for 5 min, and 5 °C for 5 min for reverse transcription, and 30 cycles at 94 °C for 30 s, 60 °C for 30 s, and 72 °C for 1 min for PCR. The following primer pairs (synthesized by TaKaRa Bio) were used: *Ggt1* forward primer, 5'-ACCACTCACCCAACCGCCTAC-3'; *Ggt1* reverse, 5'-ATCCGAACCTTGCCGTCCTT-3' (product size: 317 bp)^[12]. Expression of target mRNAs was measured relative to that of glyceraldehyde 3-phosphate dehydrogenase (*Gapdh*), which was determined using the following primers: *Gapdh* forward, 5'-GGCACAGTCAAGGCTGAGAATG-3'; *Gapdh* reverse, 5'-ATGGTGGTGAAGACGCCAGTA-3' (143 bp, TaKaRa Bio ID: RA015380). A portion of each PCR mixture was electrophoresed on a 2% agarose gel in Tris-borate-EDTA buffer and DNA bands were visualized using ethidium bromide staining. PCR product intensity was measured using the Gene Genius Bioimaging System (Syngene, Cambridge, United Kingdom).

Statistical analysis

Statistical analyses were performed using SPSS ver. 22 for Windows. All values are expressed as the mean \pm SE. One-way analysis of variance (ANOVA) followed by a Bonferroni multiple-comparison test was used for analyzing the statistical difference between the fasting periods in SA- or GSH-treated groups. Two-way ANOVA followed by a Bonferroni multiple-comparison test was used for analyzing the statistical difference between the SA- and GSH-treated groups for the fasting periods. Statistical significance was accepted at $P < 0.05$. The statistical analysis of this study was performed by Uchida H, who acquired biostatistics expertise during his training in public health.

RESULTS

Body weight changes

Figure 2 shows body weight changes in SA-treated and GSH-treated rats. Decreases in body weight in both groups were observed along with fasting. Rats fasted for 48 and 72 h with SA treatment showed approximately 15% ($P < 0.01$) and 19% ($P < 0.01$) body weight loss, respectively, compared with normally fed controls. There was no difference in weight loss between the GSH treatments and the respective SA-treated groups at each fasting period.

Histological characterization of jejunal mucosal atrophy

Jejunal mucosal atrophy was assessed by jejunal mucosal height. Decreased jejunal mucosal height in both SA- and GSH-treated rats was observed along with fasting (Figure 3). Rats fasted for 48 and 72 h with SA treatment showed a significant decrease ($P < 0.01$) in mucosal height compared with normally fed controls. However, significant differences ($P < 0.01$) in jejunal mucosal height were found between the SA-

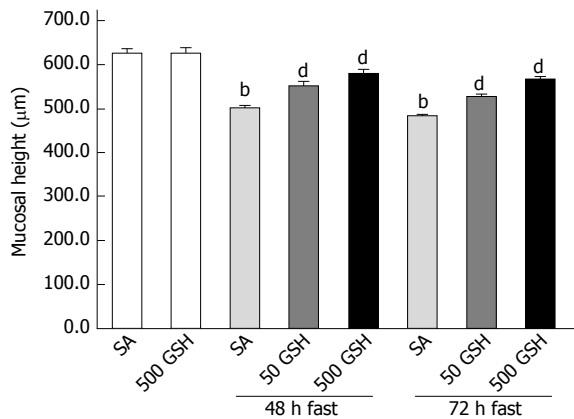


Figure 3 Effects of fasting and glutathione treatment on jejunal mucosal atrophy. Jejunal mucosal atrophy was assessed by jejunal mucosal height. Values represent the mean \pm SE. ^a $P < 0.01$ vs the normally fed control. ^b $P < 0.01$ vs the respective SA-treated group in each fasting period. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

and GSH-treated rats with 48 and 72 h of fasting.

iNOS protein expression in the jejunum

As shown in Figure 4A, iNOS protein staining was localized almost exclusively in the mucosal epithelial monolayer in the 48- and 72-h fasting periods compared with the normally fed controls, and a decreased level of staining occurred with GSH treatment. Quantitative measurement using the average optical density revealed that this expression significantly increased following fasting for 48 and 72 h with SA treatment ($P < 0.01$), whereas GSH treatments significantly decreased the fasting-induced enhancement of jejunal iNOS protein expression ($P < 0.05$ vs SA at 48 h fasting, $P < 0.01$ vs SA at 72 h fasting; Figure 4B).

Nitrite level in the jejunum

We next measured the levels of nitrite, which is a stable oxidation product of endogenous NO that is induced by iNOS. In SA-treated groups, fasting significantly increased the jejunal nitrite concentration with 48 and 72 h of fasting compared with the normally fed controls ($P < 0.01$; Figure 5). Conversely, GSH treatments significantly decreased the fasting-induced enhancement of the jejunal nitrite concentration compared with the respective SA-treated group for each fasting period ($P < 0.01$).

8-OHdG level in the jejunum

As we previously determined that fasting causes jejunal apoptosis *via* ROS production and induction of NO following increased iNOS expression, in the present study we measured levels of iNOS expression, nitrite (indicating NO production), and 8-OHdG (as a marker of ROS presence). Consistent with the abovementioned changes in iNOS expression, fasting increased intestinal nitrite levels, whereas the degree of increase was significantly reduced by GSH treatment. Furthermore,

the elevated jejunal 8-OHdG levels observed after fasting were significantly diminished by both 48 h and 72 h of GSH treatment ($P < 0.01$ for 50 mg/kg GSH vs SA with 72 h of fasting, $P < 0.05$ for 500 mg/kg GSH vs SA with 72 h of fasting; Figure 6).

Evaluation of enterocyte apoptosis

Apoptosis was determined by histomorphometry, which is preferable to terminal deoxynucleotidyl transferase dUTP nick-end labeling for quantitative assessment. Using histomorphometric assessment of jejunal cells, we evaluated the contribution of reduced apoptosis to the recovery from fasting-induced mucosal atrophy mediated by oral GSH treatment. The representative apoptosis changes as visualized by conventional light microscopy of H&E-stained specimens are shown in Figure 7A. AI distribution profiles in the villi and the crypt are also shown in Figure 7B. From the AI distribution profiles, increased apoptosis in the lower half of the villus (cell positions 1 to 40) was evident with 48 and 72 h of fasting in the SA-treated groups, and this increase was diminished by GSH treatment. In the SA-treated groups, fasting significantly increased jejunal villus AI with 48 and 72 h of fasting compared with that in the normally fed controls ($P < 0.01$; Table 1). GSH treatment significantly decreased the fasting-induced enhancement of villus AI compared with that in the respective SA-treated group for each fasting period ($P < 0.05$ for 50 mg/kg GSH vs SA with 48 or 72 h of fasting, $P < 0.01$ for 500 mg/kg GSH vs SA with 48 or 72 h of fasting). From the AI distribution profiles, increased apoptosis in the lower two-thirds of crypts (cell positions 1 to 20) was evident with 48 and 72 h of fasting in the SA-treated groups (Figure 7B), and this effect was diminished by GSH treatment. In the SA-treated groups, fasting significantly increased jejunal crypt AIs with 48 and 72 h of fasting compared with the normally fed controls ($P < 0.01$; Table 1). GSH treatments significantly ameliorated the fasting-induced increase in crypt AI compared with the respective SA-treated group for each fasting period ($P < 0.01$).

Evaluation of cell proliferation

GSH is known to exhibit an important function related to the regulation of cell proliferation by protecting against the damaging effects of ROS^[32]. To evaluate the effect of oral GSH administration on the reduced cell proliferation in fasting-induced jejunal mucosal atrophy, we assessed 5-BrdU incorporation, a proliferation indicator, in the jejunum. Although cell proliferation decreased with 48 and 72 h of fasting compared with that in the normally fed controls ($P < 0.01$; Figure 8), animals treated with GSH exhibited significantly higher levels of jejunal cell proliferation compared with those in the respective SA-treated group for each fasting period ($P < 0.01$ for 50 mg/kg GSH vs SA at 48 h fasting, and for 500 mg/kg GSH vs

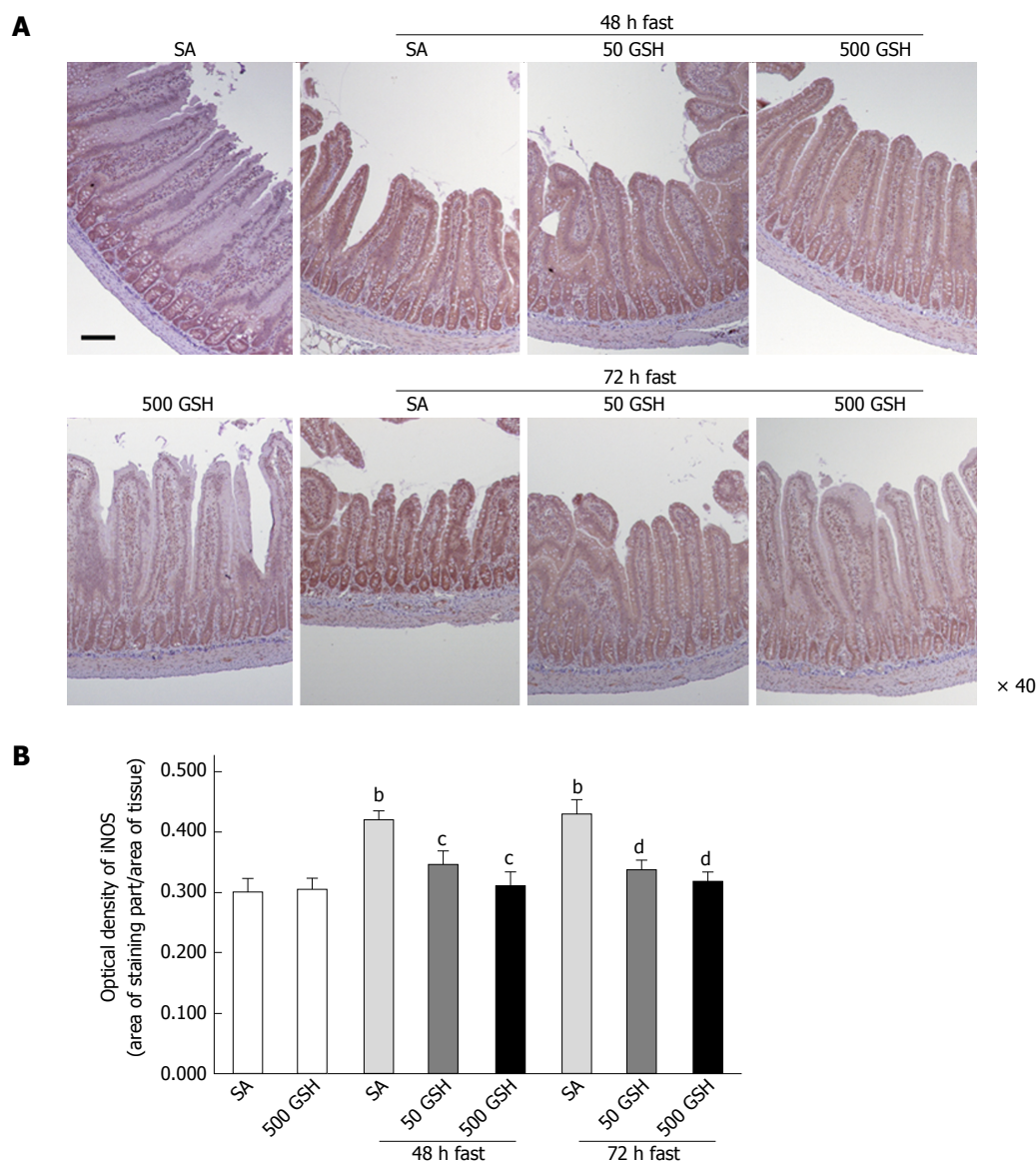


Figure 4 Effects of fasting and glutathione treatment on inducible nitric oxide synthase protein expression in the jejunum. A: Light micrographs of immunohistochemical staining for iNOS. In SA-treated groups, iNOS protein staining was localized almost exclusively in the mucosal epithelial monolayer with 48 and 72 h of fasting compared with the normally fed control. Decreased staining occurred with GSH treatment. Bar = 100 μ m. B: Optical density of iNOS protein. The content of iNOS protein was quantitatively measured by averaging the optical density. Values represent the mean \pm SE. ^b $P < 0.01$ vs normally fed controls. ^d $P < 0.01$, ^c $P < 0.05$ vs the respective SA group in each fasting period. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline; iNOS: Inducible nitric oxide synthase.

SA at 48 and 72 h fasting).

GSH and GSSG levels in the jejunum

GSH is one of the most important scavengers of ROS; additionally, GSH and oxidized GSH (*i.e.*, GSSG) are used as markers of oxidative stress^[33]. To evaluate GSH redox balance and GSH levels in the jejunum consequent to oral GSH administration during fasting, we assessed GSH and GSSG levels in the jejunum. Fasting significantly decreased the GSH concentration in the jejunum ($P < 0.01$; Figure 9A). GSH treatment significantly further decreased the fasting-induced decreases in the jejunal GSH concentration for each fasting period ($P < 0.01$). Conversely, GSSG concentration in the jejunum was similar to that of normally fed controls except for the 500 mg/kg GSH treatment

group with *ad libitum* feeding (Figure 9B).

Ggt1 mRNA expression in the jejunum

Fasting significantly increased *Ggt1* expression in the jejunum at 48 h of fasting ($P < 0.05$) (Figure 10) whereas GSH treatment significantly reduced *Ggt1* elevation at 48 h of fasting ($P < 0.05$, $P < 0.01$ for 50 mg/kg GSH, 500 mg/kg GSH vs SA at 48 h of fasting, respectively). A similar tendency was obtained for the 72-h fasting group, although this difference did not reach statistical significance.

DISCUSSION

Previous reports have suggested the involvement of GSH in fasting-induced intestinal mucosal atrophy; in

Table 1 Enterocyte apoptosis of the jejunal villus and crypt evaluated by conventional light microscopy of hematoxylin and eosin-stained specimens

	SA	500 GSH	48 h fast				72 h fast	
			SA	50 GSH	500 GSH	SA	50 GSH	500 GSH
Villus								
Cells per villus column, <i>n</i>	70 ± 1	69 ± 1	56 ± 2 ^b	64 ± 1 ^d	67 ± 2 ^d	56 ± 2 ^b	63 ± 1 ^d	65 ± 2 ^d
Apoptotic cells per villus column, <i>n</i>	0.06 ± 0.01	0.09 ± 0.01	0.20 ± 0.03 ^b	0.12 ± 0.01 ^d	0.11 ± 0.01 ^d	0.23 ± 0.03 ^b	0.18 ± 0.01	0.14 ± 0.02 ^c
Apoptotic index, %	0.09 ± 0.01	0.12 ± 0.01	0.38 ± 0.06 ^b	0.19 ± 0.02 ^c	0.18 ± 0.02 ^d	0.42 ± 0.06 ^b	0.29 ± 0.02 ^c	0.21 ± 0.02 ^d
Crypt								
Cells per villus column, <i>n</i>	28 ± 0.2	27 ± 0.4	22 ± 0.4 ^b	25 ± 0.3 ^d	27 ± 0.4 ^d	22 ± 0.3 ^b	23 ± 0.4 ^c	26 ± 0.3 ^d
Apoptotic cells per villus column, <i>n</i>	0.06 ± 0.01	0.06 ± 0.01	0.18 ± 0.01 ^b	0.08 ± 0.01 ^d	0.09 ± 0.01 ^d	0.17 ± 0.02 ^b	0.10 ± 0.01 ^d	0.11 ± 0.01 ^d
Apoptotic index, %	0.21 ± 0.04	0.22 ± 0.02	0.81 ± 0.05 ^b	0.32 ± 0.03 ^d	0.33 ± 0.03 ^d	0.78 ± 0.10 ^b	0.41 ± 0.06 ^d	0.40 ± 0.05 ^d

Apoptosis was measured as in Figure 7A. 7-8 rats were tested in each group. Values represent the mean ± SE. ^b*P* < 0.01 vs the normally fed control; ^d*P* < 0.01 or ^c*P* < 0.05 vs the SA-treated group in each fasting period. GSH: Glutathione; SA: Saline.

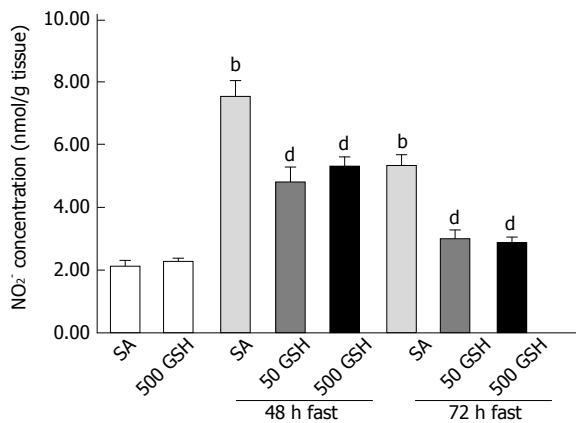


Figure 5 Effects of fasting and glutathione treatment on nitrite concentration in the jejunum. Nitrite concentrations were measured by HPLC using postcolumn derivatization with Griess reagent. Values represent the mean ± SE. ^b*P* < 0.01 vs the SA-treated group. ^d*P* < 0.01 vs the respective SA-treated group in each fasting period. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

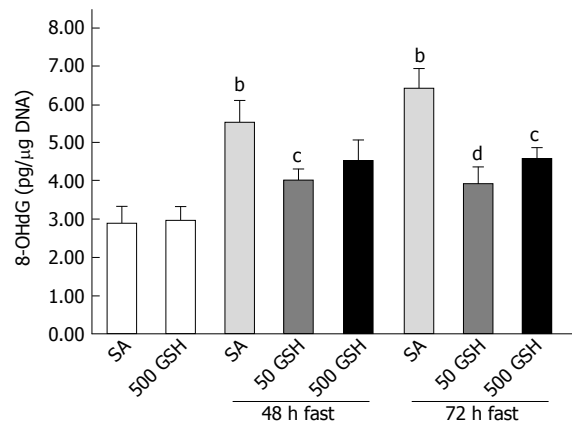


Figure 6 Effects of fasting and glutathione treatment on DNA oxidative damage in the jejunum. DNA oxidative damage was assessed by the level of 8-hydroxydeoxyguanosine (8-OHdG) in the jejunum. Values represent the mean ± SE. ^b*P* < 0.01 vs the normally fed control. ^c*P* < 0.01, ^d*P* < 0.05 vs the SA-treated group with 48 and 72 h of fasting. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

addition, fasting has been associated with increased ROS formation and depletion of the critical antioxidant GSH in the intestine^[4,6,34-36]. However, the roles of GSH in intestinal mucosal recovery from this condition as mediated by oral GSH administration have not yet been described. The objective of the present study was to investigate the effects of oral GSH administration on the development of fasting-induced intestinal atrophy, with a particular focus on the possible participation of intracellular GSH level and GGT expression on the cell surface as potential parameters of intracellular *de novo* GSH synthesis.

During fasting, the intestinal lumen receives very little or no GSH from two of the three primary sources, *i.e.*, biliary secretion^[10,37] and dietary intake^[11]. Therefore, the overall GSH levels of the intestinal mucosa decrease during fasting because the remaining GSH source, intracellular GSH, is primarily synthesized from GSH in the intestinal lumen.

There is growing evidence that dysfunctional

GSH homeostasis is involved in the etiology of several diseases. The previously reported conditions associated with GSH depletion include liver disease^[38], immune disorders^[39], neurodegenerative disease^[40], cardiovascular disease^[41], pulmonary disease^[42], arthritis, diabetes^[43], and the aging process itself^[44]. Thus, oral supplementation with GSH to increase the depressed GSH level has been studied extensively as a potential means to prevent these diseases by countering the negative effects of oxidative stress, which is one of their underlying causes. Several studies have shown that GSH supplementation in laboratory animals is effective, with benefits including enhancement of immune function^[45] and protection against carcinogenesis^[46]. In addition, daily consumption of GSH supplements in humans was effective at increasing body compartment stores of GSH, which may be of importance as the maintenance of tissue levels of GSH might be critical for maintaining health and preventing disease as well as age-related biological insults^[47]. Thus, it has been

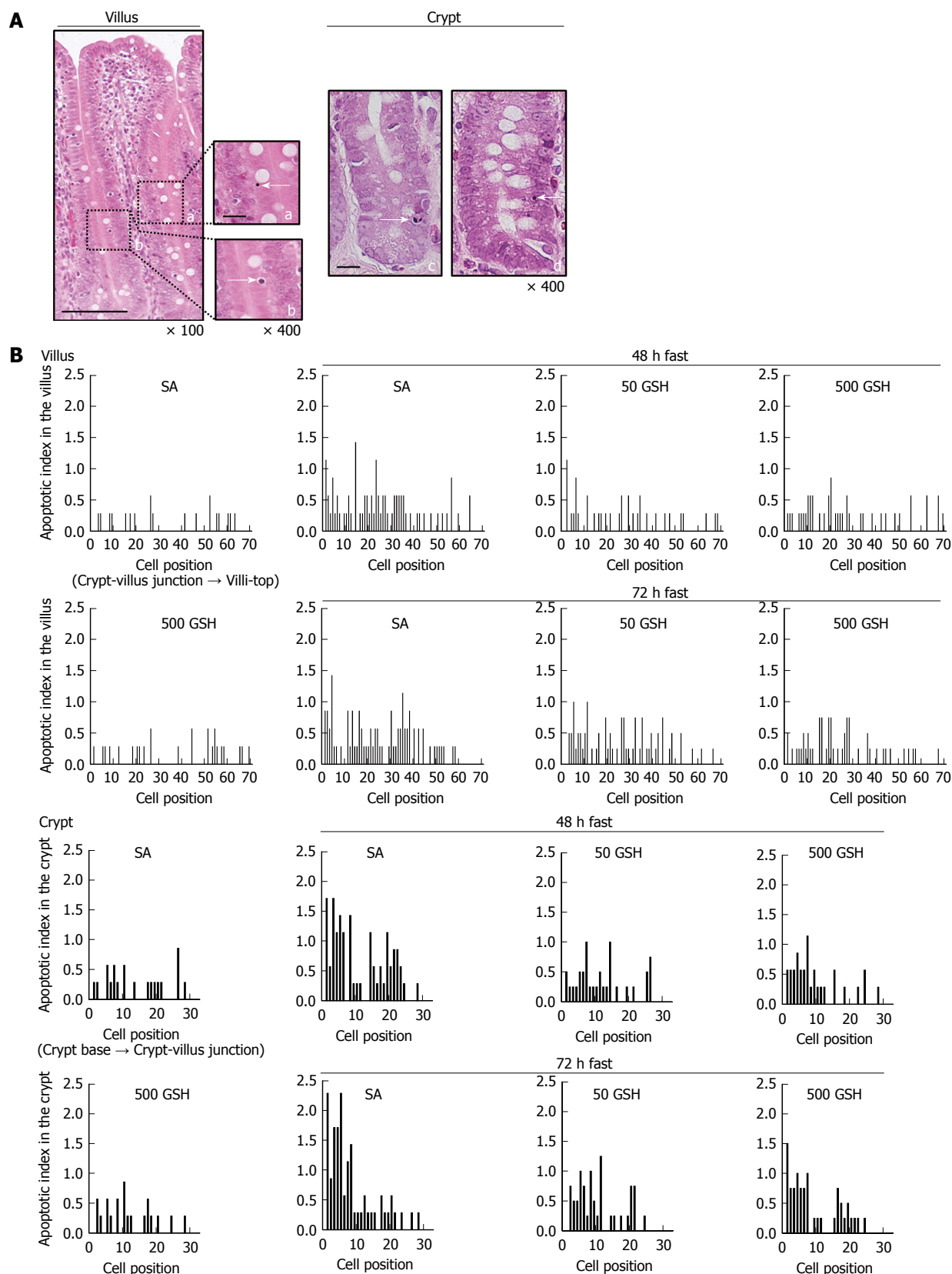


Figure 7 Effects of fasting and glutathione treatment on apoptotic index in the jejunal villus and crypt. Representative apoptotic changes determined by conventional light microscopy of hematoxylin and eosin (HE)-stained sections of the jejunal mucosa are shown in A. Left side: A jejunal villus from a 72-h fasted rat with 500 mg/kg GSH treatment. Apoptotic cells in the villus are indicated by an arrow showing an apoptotic corpuscle (a) and condensed chromatin (b). Bar = 100 μ m (low magnification). Bar = 20 μ m (high magnification). Right side: Jejunal crypts from 48- and 72-h fasted rats. Apoptotic cells in the crypt are indicated by an arrow showing an intensely eosinophilic cytoplasm and nuclear fragmentation (c) and condensed chromatin (d). Bar = 20 μ m. AI distribution curves in the villus and the crypt are shown in B. AI is defined as the total number of apoptotic cells at each cell position and is expressed as a percentage of the total number of cells counted at that cell position. Cell position 1 is defined as the cell at the crypt-villus junction and the cell at the base of the crypt column for the villus and crypt data, respectively. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

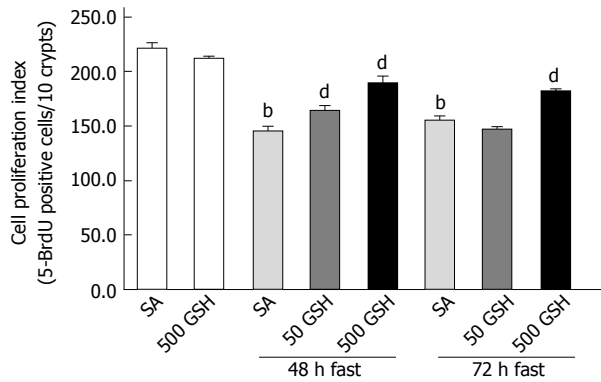


Figure 8 Effects of fasting and glutathione treatment on cell proliferation in the jejunum. Cell proliferation in the jejunal crypt was histologically assessed by 5-bromo-2'-deoxyuridine (5-BrdU) incorporation. The fraction of 5-BrdU-positive cells was expressed as the cell proliferation index (5-BrdU-positive cells/10 crypts). Values represent the mean \pm SE. ^b $P < 0.01$ vs the normally fed control. ^d $P < 0.01$ vs the respective SA-treated group in each fasting period. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline.

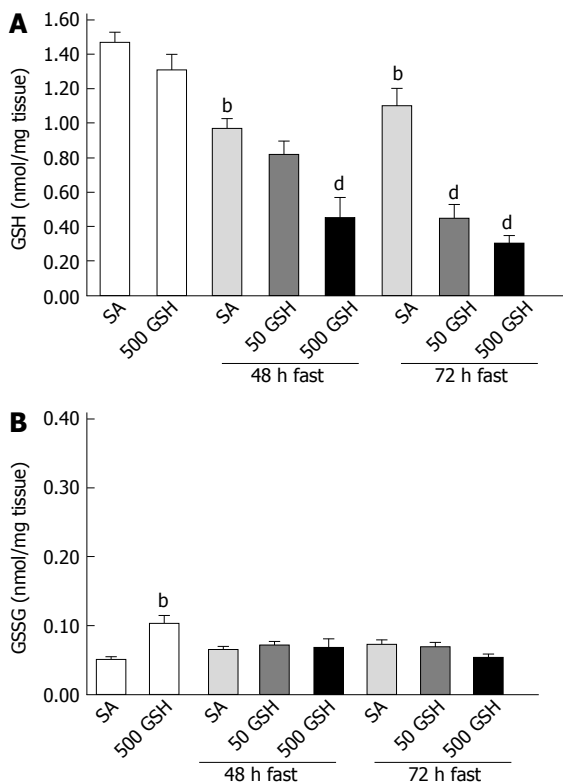


Figure 9 Effects of fasting and glutathione treatment on glutathione and oxidized glutathione concentration in the jejunum. GSH and GSSG concentration in the jejunum was determined by measuring the absorption derived from a colorimetric reaction with DTNB [5, 5'-dithiobis (2-nitrobenzoic acid)] coupled with the enzymatic recycling system. A: GSH concentration in the jejunum; B: GSSG concentration in the jejunum. Values are the mean \pm SE. ^b $P < 0.01$ vs the normally fed control. ^d $P < 0.01$ vs the respective SA-treated group in each fasting period. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline; GSSG: Oxidized glutathione.

suggested that orally administered GSH may act as a backup for intracellular GSH for GSH-deficient tissue. Therefore, the enhancement of intestinal mucosal GSH levels by oral GSH supplementation under conditions

of intracellular GSH deficiency-related biological insults such as fasting may restore tissue GSH and promote ROS metabolism, thus facilitating the recovery from intestinal mucosal atrophy.

Our previous studies suggested that fasting-induced intestinal mucosal atrophy could be alleviated by directly inhibiting the fasting-mediated NO production by iNOS and subsequent ROS enhancement associated with increased jejunal apoptosis *via*, e.g., iNOS inhibitor treatment^[20] or refeeding^[21]. Notably, an association was also observed between inhibition of ROS and increasing cell proliferation^[21]. The intestinal epithelial layer is uniquely organized for rapid self-renewal, which is achieved by the well-regulated processes of crypt-to-villus apoptosis and crypt stem cell proliferation. The intestinal epithelium sits at the interface between the intestinal mucosa and the intestinal lumen, and as such is prone to oxidative damage induced by luminal and intracellular oxidants. A previous study^[16] reviewed the GSH/GSSG redox mechanism, which modulates intestinal cell transition through cell proliferation, differentiation, or apoptosis and can govern the regenerative potential of the intestinal mucosa. Therefore, we hypothesized that protection against intestinal mucosal atrophy could be provided by controlling apoptosis and cell proliferation through the elimination of induced ROS in the intestinal mucosa by orally administering an antioxidant such as GSH during fasting in rats.

Here, we found that there was no difference in weight loss between fasting alone and oral GSH treatment during fasting (Figure 2) with the GSH levels used for this study. Conversely, jejunal mucosal atrophy was significantly increased by fasting but was remedied by oral GSH treatment during fasting (Figure 3), with significantly greater improvement resulting from the high vs low GSH dose. Furthermore, because the etiology of fasting-induced intestinal atrophy was shown to comprise ROS genesis from the NO produced by iNOS^[20,21], we measured iNOS protein expression in the jejunum. We identified an inverse relationship between intestinal mucosal atrophy and iNOS protein expression (primarily in the mucosal epithelial monolayer) associated with oral GSH treatment during fasting, most particularly with high levels of GSH administration (Figure 4). This is supported by our previous observations that iNOS inhibitor treatment during fasting and refeeding after fasting decreases fasting-induced atrophy and suppresses iNOS expression in the jejunum.

Oral GSH treatment during fasting inhibited the fasting-mediated generation of the apoptosis mediators NO and ROS concomitant with decreased iNOS expression (Figures 5 and 6). To determine the importance of apoptotic changes in the jejunum for intestinal recovery by oral GSH treatment (Figure 7A), we performed a histomorphometric assessment including AI score (Table 1) and distribution profiles

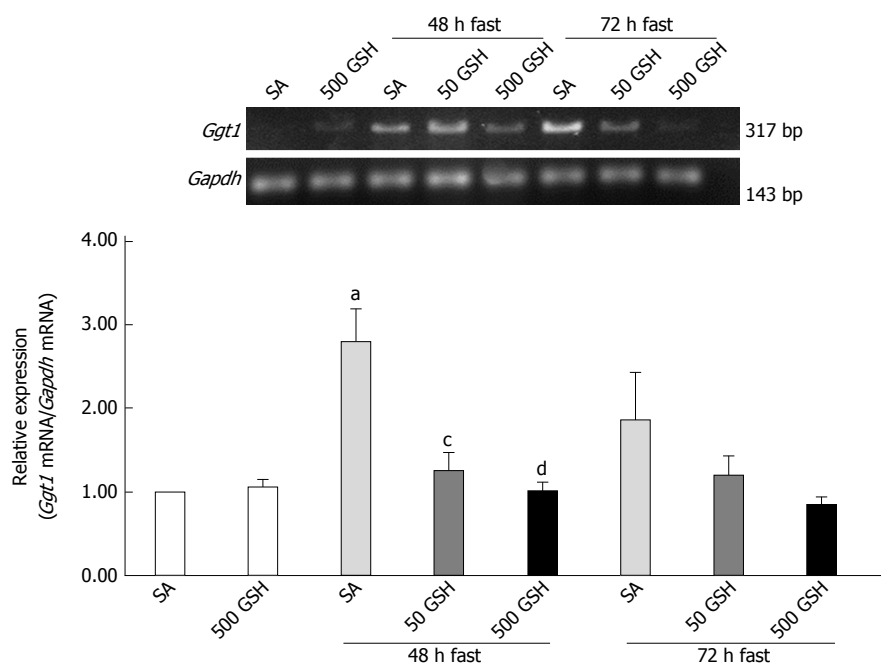


Figure 10 Effects of fasting and glutathione treatment on *Ggt1* mRNA expression in the jejunum. GGT is the only enzyme of the gamma-glutamyl cycle located on the outer surface of plasma membrane and plays key roles in GSH homeostasis by breaking down extracellular GSH. Values are the mean \pm SE. ^a $P < 0.05$ vs the normally fed control. ^b $P < 0.01$, ^c $P < 0.05$ vs the SA-treated group with 48 h of fasting. 7-8 rats were tested in each group. GSH: Glutathione; SA: Saline; GSSG: Oxidized glutathione.

(Figure 7B). The increased AI recorded in the villi and crypts after fasting decreased with oral GSH treatment (Table 1). Specifically, the lower half of the apoptotic enterocytes distributed across the entire jejunal villi and the apoptotic cells primarily located in the lower two-thirds of the crypt (Figure 7B) were particularly affected by oral GSH treatment during fasting. These results were broadly consistent with changes in iNOS, NO, and ROS as apoptosis mediators, suggesting that oral GSH administration during fasting may inhibit enterocyte apoptosis by diminishing the ROS in the jejunum generated by fasting.

GSH treatment also was found to alleviate the decreased jejunal cell proliferation mediated by fasting (Figure 8), with significantly greater improvement in fasting-induced jejunal mucosal atrophy resulting from high vs low dose GSH. These observations are consistent with the role of GSH in cell proliferation through protection against damage from ROS^[16,32]. Together, these findings indicate that oral GSH administration leads to less fasting-mediated enhancement and therefore decreased overall ROS levels in the jejunal mucosa, thereby functioning to increase cell proliferation. This suggests that oral GSH administration may provide regenerative potential to the mucosa during fasting.

Notably, although our findings replicated prior results^[6,34-36] that fasting significantly decreased jejunal GSH concentrations (Figure 9A), we found that the concentrations were further significantly decreased by oral GSH supplementation during fasting, with a greater effect from high vs low GSH dose. Conversely,

the jejunal concentrations of the GSH oxidation product GSSG were not changed by any feeding regime (Figure 9B). Furthermore, oral GSH treatment during fasting significantly decreased the fasting-induced ROS levels in the jejunum (Figure 6), but we consider it unlikely that this effect depends on antioxidant activity given the depressed GSH concentration in the jejunum. These results demonstrate that oral GSH administration during fasting did not contribute to the supply of GSH in the jejunum, and an increase in GSH oxidation was not observed, suggesting that oral GSH administration during fasting does not provide a backup for the jejunum-depleted GSH.

The synthesis of intracellular *de novo* GSH, which is critical for ROS removal and maintenance of GSH homeostasis, relies upon the enzyme activity of the plasma membrane protein GGT to break down extracellular GSH^[12]. In the present study, we demonstrated that oral GSH treatment significantly ameliorated the increased jejunal *Ggt1* mRNA expression observed following 48 h of fasting (Figure 10). In comparison, Jonas *et al.*^[4] indicated that fasting for 72 h did not change *Ggt1* mRNA levels in the ileum, although this was determined *via* ribonuclease protection assay rather than by semi-quantitative RT-PCR as used in the present study. The different findings may also be a consequence of the disparity in fasting duration. Notably, Ogasawara *et al.*^[34] showed that starvation causes a striking decrease of GGT activity in the jejunum, which is consistent with our finding of fasting-induced increases in *Ggt1* mRNA levels, as the mRNA expression of GGT might be upregulated by the

decline of GGT activity^[12]. Furthermore, Zhang *et al.*^[12] described that *Ggt1* mRNA expression is increased during oxidative stress, which was considered to constitute an adaptation to such adverse conditions. Similarly, we found that fasting increased jejunal ROS levels and up-regulated *Ggt1* mRNA expression in the jejunum in this study. However, considering that oral GSH treatment during fasting significantly decreased both jejunal GSH concentration and *Ggt1* mRNA expression, the supplemented GSH in the intestinal lumen may not be involved in intracellular *de novo* GSH synthesis.

The ability of GSH to control extracellular redox provides another possible role for intestinal luminal GSH during fasting, as the present study demonstrated that intracellular GSH levels of the jejunum mucosa were not increased by intestinal luminal GSH administration during fasting, although the GSH administration attenuated the fasting-induced ROS in the intestinal mucosa. Reversible redox reactions of intracellular thiol/disulfide pairs such as GSH/GSSG regulate diverse biological processes *in vivo*, including signaling for apoptosis and cell proliferation, enzyme catalysis, gene expression, and molecular folding and trafficking^[16-18,48,49]. Most studies have focused on the major intracellular thiols such as GSH^[50,51]; however, relatively little is known regarding the effects of changes in extracellular thiol/disulfide pairs. Biliary GSH is an important intestinal luminal source of GSH and attenuates ROS of the intestinal mucosa by increasing the GSH levels of the intestinal mucosa. However, it has been suggested that biliary GSH controls the status of the intestinal lumen and contributes to protection of the intestinal mucosa^[10]. The GSH of the intestinal lumen may act on the intestinal mucosa *via* both of these two pathways, and it is important to consider the control of the redox status of the intestinal lumen. In addition, the roles of extracellular (*i.e.*, intestinal luminal) GSH in the recovery from fasting-induced intestinal mucosal atrophy enabled by oral GSH treatment have not yet been described.

A few studies have examined the ability of the extracellular thiol/disulfide redox state to regulate apoptosis and cell proliferation. Circu *et al.*^[16] demonstrated that the luminal/extracellular redox environment is determined by the cysteine/cystine (Cys/CySS) redox pair with contributions from the GSH system, with the majority of Cys in the intestinal lumen originating from GGT enzymatic hydrolysis of GSH obtained from dietary intake and biliary supply. Intestinal cell proliferation and apoptosis are associated with quantitative changes in the redox potential (Eh) of the extracellular GSH/GSSG or Cys/CySS redox pair. In the present study, the changes in apoptosis and cell proliferation in the intestinal mucosa resulting from oral GSH administration during fasting may derive from Eh changes mediated by GSH and Cys in the intestinal lumen, which in turn may ultimately

control signaling proteins, enzyme catalysis, and gene expression. For example, Devadas *et al.*^[52] linked ROS formation with the progression of Fas-induced apoptosis *via* the protective effects of extracellular GSH, which are ascribed to its known capacity as an antioxidant. Specifically, extracellular GSH was deduced to inhibit Fas-mediated apoptosis following ROS reduction in cells because Fas crosslinking induced rapid generation of ROS well before the appearance of characteristic apoptotic changes. Furthermore, Fujise *et al.*^[22] suggested that fasting-induced apoptosis in the rat small intestine occurred *via* a receptor-mediated type I apoptotic pathway including induced expression of FasL, Fas, and TNFR1. Selleri *et al.*^[53] and Viard-Leveugle *et al.*^[54] provided evidence that iNOS expression and NO production are involved in the mechanism of FasL upregulation and Fas-mediated apoptosis. The iNOS-NO-ROS-FasL pathway represents a potential link between the apoptosis and intestinal atrophy observed in the fasting.

In the present study, fasting-induced intestinal mucosal atrophy resulting from increased apoptosis was caused by increased production of NO and ROS as apoptosis mediators following elevation of iNOS expression. Because the iNOS-NO-ROS-FasL pathway is part of the apoptotic mechanism in the intestinal mucosa atrophy occurring after fasting^[22,52-54], oral GSH treatment during fasting might inhibit Fas-mediated apoptosis following reduction of ROS levels in the jejunal mucosa resulting from Eh changes mediated by GSH and Cys in the intestinal lumen. Additionally, because iNOS protein expression was also induced by ROS produced from NO in fasting-induced intestinal mucosal atrophy in a previous study^[20], the decrease in ROS levels in the intestinal mucosa reduces iNOS protein expression and NO production. Moreover, Jonas *et al.*^[55] showed that the extracellular thiol/disulfide redox state modulates cell proliferation and that this system interacted with growth factor signaling in a human colon carcinoma cell line. Thus, the change in extracellular thiol/disulfide redox state in response to peptide growth factors indicated an interaction of growth factor-activated pathways and thiol/disulfide metabolism during intestinal cell proliferation.

Therefore, oral GSH administration during fasting may regulate the redox state of the intestinal lumen and consequently may both relieve Fas-mediated apoptosis and increase growth factor-mediated cell proliferation by ROS removal in jejunal epithelial cells (Figure 11). However, because our present study focused on intracellular GSH, further studies focusing on extracellular GSH are required to confirm this model. In particular, a study utilizing measurements of GSH/GSSG and Cys/CySS levels in the intestinal lumen along with Fas and growth factor signaling in the jejunum may be necessary to identify the mechanism underlying the recovery of the intestinal mucosal atrophy mediated by oral GSH administration during fasting.

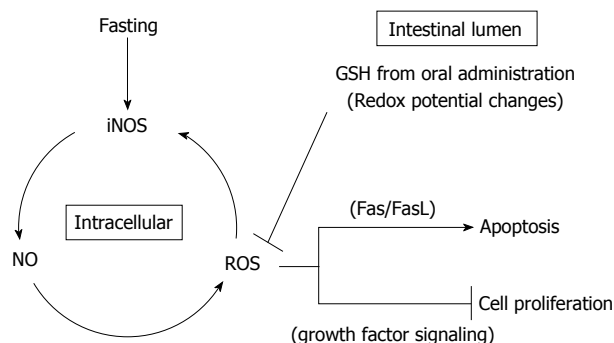


Figure 11 Schematic diagram of protective effects of glutathione in the intestinal lumen against fasting-induced intestinal atrophy, mediated through oxidative stress. The schematic diagram depicts a possible role of intestinal lumen redox status in the regulation of jejunal mucosa apoptosis and cell proliferation in fasting-induced intestinal atrophy, mediated through oxidative stress. Fasting causes increased production of NO and ROS as apoptosis mediators following elevation of iNOS expression. The changes in apoptosis and cell proliferation in the intestinal mucosa resulting from oral GSH administration during fasting may derive from intracellular ROS removal by redox potential changes mediated by GSH and Cys (originating from enzymatic hydrolysis of GSH) in the intestinal lumen. Intracellular ROS removal is considered to inhibit Fas-mediated apoptosis and increase growth factor-mediated cell proliferation. GSH: Glutathione; ROS: Reactive oxygen species.

In conclusion, oral GSH administration during fasting may provide regenerative potential to the jejunal mucosa through control of enterocyte apoptosis and cell proliferation. The GSH in the intestinal lumen provided through oral administration during fasting may not be involved in intracellular *de novo* GSH but may be useful to prevent intestinal mucosal atrophy by diminishing the levels of ROS in the jejunum generated by fasting.

COMMENTS

Background

Fasting induces small intestinal mucosal atrophy, which can lead to bacterial translocation, particularly in patients receiving a prolonged course of total parenteral nutrition (TPN), and is associated with increased reactive oxygen species (ROS) formation. This fasting state is also accompanied by depletion of the critical antioxidant glutathione (GSH), which functions to eliminate induced ROS in the intestinal mucosa. As GSH deficiency in tissues is associated with increased oxidative stress and fasting, the maintenance of tissue levels of GSH is critical for protection of small intestinal mucosa.

Research frontiers

The roles of GSH in intestinal mucosal recovery from fasting as mediated by oral GSH administration have not been investigated.

Innovations and breakthroughs

GSH treatment in the intestinal lumen could be a promising strategy to improve clinical outcome in patients receiving a prolonged course of TPN.

Applications

Oral GSH administration during fasting enhances jejunal regenerative potential to minimize intestinal mucosal atrophy by diminishing fasting-mediated ROS generation and enterocyte apoptosis and enhancing cell proliferation.

Peer-review

The manuscript by Uchida *et al* describes effects of oral GSH administration on fasting-induced intestinal atrophy in the small intestinal mucosa. The authors

found that oral GSH administration of rats during fasting significantly enhanced jejunal regenerative potential to minimize mucosal atrophy. This reduction of intestinal atrophy correlated with correction of histopathological outcomes and many biochemical parameters. The authors' conclusion that oral GSH improves intestinal atrophy is supported by convincing results.

REFERENCES

- 1 Ziegler TR. Molecular mechanisms of intestinal injury, repair, and growth. In: Takala J, Rombeau JL (eds). *Gut Dysfunction in Critical Illness*. New York: Springer-Verlag, 1996: 25-52
- 2 Shaw D, Gohil K, Basson MD. Intestinal mucosal atrophy and adaptation. *World J Gastroenterol* 2012; **18**: 6357-6375 [PMID: 23197881 DOI: 10.3748/wjg.v18.i44.6357]
- 3 Darmon N, Pélissier MA, Heyman M, Albrecht R, Desjeux JF. Oxidative stress may contribute to the intestinal dysfunction of weanling rats fed a low protein diet. *J Nutr* 1993; **123**: 1068-1075 [PMID: 8505667]
- 4 Jonas CR, Farrell CL, Scully S, Eli A, Estivariz CF, Gu LH, Jones DP, Ziegler TR. Enteral nutrition and keratinocyte growth factor regulate expression of glutathione-related enzyme messenger RNAs in rat intestine. *J Parenter Enteral Nutr* 2000; **24**: 67-75 [PMID: 10772185 DOI: 10.1177/014860710002400267]
- 5 Yu BP. Cellular defenses against damage from reactive oxygen species. *Physiol Rev* 1994; **74**: 139-162 [PMID: 8295932]
- 6 Jonas CR, Estivariz CF, Jones DP, Gu LH, Wallace TM, Diaz EE, Pascal RR, Cotsonis GA, Ziegler TR. Keratinocyte growth factor enhances glutathione redox state in rat intestinal mucosa during nutritional repletion. *J Nutr* 1999; **129**: 1278-1284 [PMID: 10395587]
- 7 Aw TY. Intestinal glutathione: determinant of mucosal peroxide transport, metabolism, and oxidative susceptibility. *Toxicol Appl Pharmacol* 2005; **204**: 320-328 [PMID: 15845421 DOI: 10.1016/j.taap.2004.11.016]
- 8 Franco R, Schoneveld OJ, Pappa A, Panayiotidis MI. The central role of glutathione in the pathophysiology of human diseases. *Arch Physiol Biochem* 2007; **113**: 234-258 [PMID: 18158646 DOI: 10.1080/13813450701661198]
- 9 Wu G, Fang YZ, Yang S, Lupton JR, Turner ND. Glutathione metabolism and its implications for health. *J Nutr* 2004; **134**: 489-492 [PMID: 14988435]
- 10 Aw TY. Biliary glutathione promotes the mucosal metabolism of luminal peroxidized lipids by rat small intestine in vivo. *J Clin Invest* 1994; **94**: 1218-1225 [PMID: 8083363 DOI: 10.1172/JCI117439]
- 11 Jones DP, Coates RJ, Flagg EW, Eley JW, Block G, Greenberg RS, Gunter EW, Jackson B. Glutathione in foods listed in the National Cancer Institute's Health Habits and History Food Frequency Questionnaire. *Nutr Cancer* 1992; **17**: 57-75 [PMID: 1574445 DOI: 10.1080/01635589209514173]
- 12 Zhang H, Forman HJ, Choi J. Gamma-glutamyl transpeptidase in glutathione biosynthesis. *Methods Enzymol* 2005; **401**: 468-483 [PMID: 16399403 DOI: 10.1016/S0076-6879(05)01028-1]
- 13 Aw TY, Williams MW. Intestinal absorption and lymphatic transport of peroxidized lipids in rats: effect of exogenous GSH. *Am J Physiol* 1992; **263**: G665-G672 [PMID: 1443140]
- 14 Schmitt B, Vicenzi M, Garrel C, Denis FM. Effects of N-acetylcysteine, oral glutathione (GSH) and a novel sublingual form of GSH on oxidative stress markers: A comparative crossover study. *Redox Biol* 2015; **6**: 198-205 [PMID: 26262996 DOI: 10.1016/j.redox.2015.07.012]
- 15 Imhoff BR, Hansen JM. Extracellular redox status regulates Nrf2 activation through mitochondrial reactive oxygen species. *Biochem J* 2009; **424**: 491-500 [PMID: 19778293 DOI: 10.1042/BJ20091286]
- 16 Circu ML, Aw TY. Intestinal redox biology and oxidative stress. *Semin Cell Dev Biol* 2012; **23**: 729-737 [PMID: 22484611 DOI: 10.1016/j.semcdb.2012.03.014]
- 17 Pérez S, Taléns-Visconti R, Rius-Pérez S, Finamor I, Sastre J. Redox signaling in the gastrointestinal tract. *Free Radic Biol Med*

- 2017; **104**: 75-103 [PMID: 28062361 DOI: 10.1016/j.freeradbiomed.2016.12.048]
- 18 **Moriarty-Craige SE**, Jones DP. Extracellular thiols and thiol/disulfide redox in metabolism. *Annu Rev Nutr* 2004; **24**: 481-509 [PMID: 15189129 DOI: 10.1146/annurev.nutr.24.012003.132208]
 - 19 **D'Orazio M**, Pacello F, Battistoni A. Extracellular glutathione decreases the ability of Burkholderia cenocepacia to penetrate into epithelial cells and to induce an inflammatory response. *PLoS One* 2012; **7**: e47550 [PMID: 23094061 DOI: 10.1371/journal.pone.0047550]
 - 20 **Ito J**, Uchida H, Yokote T, Ohtake K, Kobayashi J. Fasting-induced intestinal apoptosis is mediated by inducible nitric oxide synthase and interferon- γ in rat. *Am J Physiol Gastrointest Liver Physiol* 2010; **298**: G916-G926 [PMID: 20378828 DOI: 10.1152/ajpgi.00429.2009]
 - 21 **Ito J**, Uchida H, Machida N, Ohtake K, Saito Y, Kobayashi J. Inducible and neuronal nitric oxide synthases exert contrasting effects during rat intestinal recovery following fasting. *Exp Biol Med* (Maywood) 2017; **242**: 762-772 [PMID: 28195513 DOI: 10.1177/1535370217694434]
 - 22 **Fujise T**, Iwakiri R, Wu B, Amemori S, Kakimoto T, Yokoyama F, Sakata Y, Tsunada S, Fujimoto K. Apoptotic pathway in the rat small intestinal mucosa is different between fasting and ischemia-reperfusion. *Am J Physiol Gastrointest Liver Physiol* 2006; **291**: G110-G116 [PMID: 16574989 DOI: 10.1152/ajpgi.00393.2005]
 - 23 **Dahly EM**, Guo Z, Ney DM. Alterations in enterocyte proliferation and apoptosis accompany TPN-induced mucosal hypoplasia and IGF-I-induced hyperplasia in rats. *J Nutr* 2002; **132**: 2010-2014 [PMID: 12097684]
 - 24 **Morin MJ**, Karr SM, Faris RA, Gruppiso PA. Developmental variability in expression and regulation of inducible nitric oxide synthase in rat intestine. *Am J Physiol Gastrointest Liver Physiol* 2001; **281**: G552-G559 [PMID: 11447036]
 - 25 **Shi SR**, Chaiwun B, Young L, Cote RJ, Taylor CR. Antigen retrieval technique utilizing citrate buffer or urea solution for immunohistochemical demonstration of androgen receptor in formalin-fixed paraffin sections. *J Histochem Cytochem* 1993; **41**: 1599-1604 [PMID: 7691930 DOI: 10.1177/41.11.7691930]
 - 26 **Lu H**, Zhu B, Xue XD. Role of neuronal nitric oxide synthase and inducible nitric oxide synthase in intestinal injury in neonatal rats. *World J Gastroenterol* 2006; **12**: 4364-4368 [PMID: 16865779 DOI: 10.3748/wjg.v12.i27.4364]
 - 27 **Tang Y**, Swartz-Basile D, Swietlicki EA, Yi L, Rubin DC, Levin MS. Bax is required for resection-induced changes in apoptosis, proliferation, and members of the extrinsic cell death pathways. *Gastroenterology* 2004; **126**: 220-230 [PMID: 14699502]
 - 28 **Ohtake K**, Ishiyama Y, Uchida H, Muraki E, Kobayashi J. Dietary nitrite inhibits early glomerular injury in streptozotocin-induced diabetic nephropathy in rats. *Nitric Oxide* 2007; **17**: 75-81 [PMID: 17681477 DOI: 10.1016/j.niox.2007.06.004]
 - 29 **Inoue S**, Kawanishi S. Oxidative DNA damage induced by simultaneous generation of nitric oxide and superoxide. *FEBS Lett* 1995; **371**: 86-88 [PMID: 7664890]
 - 30 **Shigenaga MK**, Gimeno CJ, Ames BN. Urinary 8-hydroxy-2'-deoxyguanosine as a biological marker of in vivo oxidative DNA damage. *Proc Natl Acad Sci USA* 1989; **86**: 9697-9701 [PMID: 2602371]
 - 31 **Saito S**, Yamauchi H, Hasui Y, Kurashige J, Ochi H, Yoshida K. Quantitative determination of urinary 8-hydroxydeoxyguanosine (8-OH-dg) by using ELISA. *Res Commun Mol Pathol Pharmacol* 2000; **107**: 39-44 [PMID: 11334369]
 - 32 **Filomeni G**, Rotilio G, Ciriolo MR. Cell signalling and the glutathione redox system. *Biochem Pharmacol* 2002; **64**: 1057-1064 [PMID: 12213605]
 - 33 **Zitka O**, Skalickova S, Gumulec J, Masarik M, Adam V, Hubalek J, Trnkova L, Kruseova J, Eckschlagner T, Kizek R. Redox status expressed as GSH:GSSG ratio as a marker for oxidative stress in paediatric tumour patients. *Oncol Lett* 2012; **4**: 1247-1253 [PMID: 23205122 DOI: 10.3892/ol.2012.931]
 - 34 **Ogasawara T**, Ohnhaus EE, Hoensch HP. Glutathione and its related enzymes in the small intestinal mucosa of rats: effects of starvation and diet. *Res Exp Med (Berl)* 1989; **189**: 195-204 [PMID: 2568668]
 - 35 **Viña J**, Perez C, Furukawa T, Palacin M, Viña JR. Effect of oral glutathione on hepatic glutathione levels in rats and mice. *Br J Nutr* 1989; **62**: 683-691 [PMID: 2605158]
 - 36 **Kelly FJ**. Glutathione content of the small intestine: regulation and function. *Br J Nutr* 1993; **69**: 589-596 [PMID: 8490011]
 - 37 **Takahashi I**, Kern MK, Dodds WJ, Hogan WJ, Layman RD, Ammon HV. Fasting and postprandial hepatic bile flow in unanesthetized opossums. *Am J Physiol* 1990; **259**: G745-G752 [PMID: 2240217]
 - 38 **Loguercio C**, Taranto D, Vitale LM, Beneduce F, Del Vecchio Blanco C. Effect of liver cirrhosis and age on the glutathione concentration in the plasma, erythrocytes, and gastric mucosa of man. *Free Radic Biol Med* 1996; **20**: 483-488 [PMID: 8720922]
 - 39 **Herzenberg LA**, De Rosa SC, Dubs JG, Roederer M, Anderson MT, Ela SW, Deresinski SC, Herzenberg LA. Glutathione deficiency is associated with impaired survival in HIV disease. *Proc Natl Acad Sci USA* 1997; **94**: 1967-1972 [PMID: 9050888]
 - 40 **Smeyne M**, Smeyne RJ. Glutathione metabolism and Parkinson's disease. *Free Radic Biol Med* 2013; **62**: 13-25 [PMID: 23665395 DOI: 10.1016/j.freeradbiomed.2013.05.001]
 - 41 **Usal A**, Acartürk E, Yüregir GT, Unlüktür I, Demirci C, Kurt HI, Birand A. Decreased glutathione levels in acute myocardial infarction. *Jpn Heart J* 1996; **37**: 177-182 [PMID: 8676544]
 - 42 **Gul M**, Kutay FZ, Temocin S, Hanninen O. Cellular and clinical implications of glutathione. *Indian J Exp Biol* 2000; **38**: 625-634 [PMID: 11215303]
 - 43 **Nuttall SL**, Martin U, Sinclair AJ, Kendall MJ. Glutathione: in sickness and in health. *Lancet* 1998; **351**: 645-646 [PMID: 9500325]
 - 44 **Viña J**, Sastre J, Anton V, Bruseghini L, Esteras A, Asensi M. Effect of aging on glutathione metabolism. Protection by antioxidants. *EXS* 1992; **62**: 136-144 [PMID: 1450581]
 - 45 **Furukawa T**, Meydani SN, Blumberg JB. Reversal of age-associated decline in immune responsiveness by dietary glutathione supplementation in mice. *Mech Ageing Dev* 1987; **38**: 107-117 [PMID: 3600048]
 - 46 **Schwartz JL**, Shklar G. Glutathione inhibits experimental oral carcinogenesis, p53 expression, and angiogenesis. *Nutr Cancer* 1996; **26**: 229-236 [PMID: 8875560 DOI: 10.1080/01635589609514479]
 - 47 **Richie JP Jr**, Nichenametla S, Neidig W, Calcagnotto A, Haley JS, Schell TD, Muscat JE. Randomized controlled trial of oral glutathione supplementation on body stores of glutathione. *Eur J Nutr* 2015; **54**: 251-263 [PMID: 24791752 DOI: 10.1007/s00394-014-0706-z]
 - 48 **Kamata H**, Hirata H. Redox regulation of cellular signalling. *Cell Signal* 1999; **11**: 1-14 [PMID: 10206339]
 - 49 **Allen RG**, Tresini M. Oxidative stress and gene regulation. *Free Radic Biol Med* 2000; **28**: 463-499 [PMID: 10699758]
 - 50 **Gilbert HF**. Molecular and cellular aspects of thiol-disulfide exchange. *Adv Enzymol Relat Areas Mol Biol* 1990; **63**: 69-172 [PMID: 2407068]
 - 51 **Sato N**, Iwata S, Nakamura K, Hori T, Mori K, Yodoi J. Thiol-mediated redox regulation of apoptosis. Possible roles of cellular thiols other than glutathione in T cell apoptosis. *J Immunol* 1995; **154**: 3194-3203 [PMID: 7897207]
 - 52 **Devadas S**, Hinshaw JA, Zaritskaya L, Williams MS. Fas-stimulated generation of reactive oxygen species or exogenous oxidative stress sensitize cells to Fas-mediated apoptosis. *Free Radic Biol Med* 2003; **35**: 648-661 [PMID: 12957657]
 - 53 **Selleri C**, Sato T, Raiola AM, Rotoli B, Young NS, Maciejewski JP. Induction of nitric oxide synthase is involved in the mechanism of Fas-mediated apoptosis in haemopoietic cells. *Br J Haematol* 1997; **99**: 481-489 [PMID: 9401054 DOI: 10.1046/j.1365-2141.1996.4323240.x]
 - 54 **Viard-Leveugle I**, Gaide O, Jankovic D, Feldmeyer L, Kerl K, Pickard C, Roques S, Friedmann PS, Contassot E, French

LE. TNF- α and IFN- γ are potential inducers of Fas-mediated keratinocyte apoptosis through activation of inducible nitric oxide synthase in toxic epidermal necrolysis. *J Invest Dermatol* 2013; **133**: 489-498 [PMID: 22992806 DOI: 10.1038/jid.2012.330]

55 **Jonas CR**, Ziegler TR, Gu LH, Jones DP. Extracellular thiol/disulfide redox state affects proliferation rate in a human colon carcinoma (Caco2) cell line. *Free Radic Biol Med* 2002; **33**: 1499-1506 [PMID: 12446207]

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Basic Study

Faecal and mucosal microbiota in patients with functional gastrointestinal disorders: Correlation with toll-like receptor 2/toll-like receptor 4 expression

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Abstract

AIM

To investigate the intestinal luminal microbiota (LM) and mucosa-associated microbiota (MAM) in Chinese patients with functional gastrointestinal disorders (FGIDs) and examine the association between these communities and the expression of toll-like receptor (TLR) 2 and TLR4.

METHODS

Thirty-two Chinese subjects who suffered from symptoms of FGIDs, as confirmed by gastroenterologists, were enrolled in this study. Fresh faecal samples and descending colonic mucosal biopsies were collected from the subjects before (faecal) and during (mucosal)

flexible colonoscopy. For analysis of the samples, we performed high-throughput sequencing of the V3-V4 region of the *16S rRNA* gene and reverse transcription (RT)-PCR to detect the expression of colonic TLR2 and TLR4. Differences in the stool and mucosal microbiota were examined and a correlation network analysis was performed.

RESULTS

The microbiota of faecal samples was significantly more diverse and richer than that of the mucosal samples, and the LM and MAM populations differed significantly. TLR2 expression showed a significant positive correlation with TLR4 expression. In the MAM samples, the genera *Faecalibacterium* and *Ruminococcus*, which belong to the family Ruminococcaceae, were inversely correlated with TLR4 expression ($r = -0.45817$, $P = 0.0083$ and $r = -0.5306$, $P = 0.0018$, respectively). *Granulicatella*, which belongs to Carnobacteriaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely correlated with TLR2 expression ($r = -0.5573$, $P = 0.0010$ and $r = -0.5435$, $P = 0.0013$, respectively). In the LM samples, examination at phylum, class, or order level revealed no correlation with TLR4 expression. *Faecalibacterium*, which belongs to Ruminococcaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely correlated with TLR2 expression ($r = -0.5743$, $P = 0.0058$ and $r = -0.3905$, $P = 0.0271$, respectively).

CONCLUSION

Microbial compositions of LM and MAM in Chinese patients with FGIDs are different. Expression of TLRs may be affected by the type of bacteria that are present in the gut.

Key words: Gastrointestinal microbiota; *16S rRNA* gene; Toll-like receptors; High-throughput sequencing

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Core tip: To explore which bacteria regulate the expression of toll-like receptors (TLRs) and thereby affect intestinal functions, we performed high-throughput pyrosequencing of the bacterial *16S rRNA* gene, compared the microbial communities in the faeces and mucosa of Chinese patients with functional gastrointestinal disorders, and studied their association with the expression of colonic mucosal TLR2 and TLR4. Samples of luminal microbiota were different from those of mucosa-associated microbiota (MAM), and MAM samples were closely associated with TLR2/4 expression. The abundance of *Faecalibacterium* and *Ruminococcus* was lower in patients with gut disease, while the expression of TLRs is higher than in healthy controls. The presence of *Faecalibacterium* and *Ruminococcus* was significantly and negatively correlated with TLR4 expression, suggesting that these two bacteria, which colonize on the colonic mucosa, play a key role in gut diseases by regulating mucosal

TLR4 expression. *Granulicatella*, which belongs to Carnobacteriaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely correlated with TLR2 expression. Because the two genera contained not only pathogenic species but also probiotic species, it will be important to establish a better understanding of the relationship between TLRs and bacterial strains in future studies.

Dong LN, Wang JP, Liu P, Yang YF, Feng J, Han Y. Faecal and mucosal microbiota in patients with functional gastrointestinal disorders: Correlation with toll-like receptor 2/toll-like receptor 4 expression. *World J Gastroenterol* 2017; 23(36): 6665-6673 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6665.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6665>

INTRODUCTION

The intestinal microbiota is a complex community of bacteria, archaea, viruses, and eukarya. A wide variety of bacterial species in the gastrointestinal tract exerts numerous effects on the host and influences a variety of gastrointestinal functions^[1]. Faecal samples (representing the luminal niche) are examined in most studies of the intestinal microbiota because their collection is simple. However, recent studies have shown that the microbial compositions of the luminal microbiota (LM) and the mucosa-associated microbiota (MAM) differ, suggesting that these two distinct microbial populations play different roles within the intestinal microbiota ecosystem^[2]. LM is present in the whole intestine, whereas MAM represents a special niche. Because MAM is in close contact with the host, it may play a more prominent role in the intestine, whereas LM may play a key role in metabolic activities and nutrient harvest^[3]. Toll-like receptors (TLRs) are pattern recognition receptors expressed by various cells in the gastrointestinal tract. TLRs detect conserved microbial products and play a central role in the activation of innate and adaptive immune pathways. TLR2 and TLR4 are two of the best characterized TLRs that respond to microbial membrane components. A number of studies have examined the role of TLR signalling in microbiota-induced chronic inflammation and immunopathology^[4]. The microbiota may directly interact with the TLRs and regulate gut immune responses^[5].

In some intestinal diseases, such as inflammatory bowel disease (IBD) or irritable bowel syndrome (IBS), the expression of TLR2 and TLR4 is increased and the dysbiosis is observed. However, limited data are available on the correlation between the intestinal microbiota and TLRs in humans. To explore which bacteria regulate the expression of TLRs and thereby affect intestinal functions, we performed high-throughput pyrosequencing of the bacterial *16S rRNA*

gene, compared the microbial communities in the faeces and mucosa of Chinese patients with functional gastrointestinal disorders (FGIDs), and studied their association with the expression of colonic mucosal TLR2 and TLR4.

MATERIALS AND METHODS

Study subjects

Thirty-two Chinese subjects aged between 18 and 65 years, who were diagnosed with FGIDs, were recruited at the Department of Gastroenterology, Shanxi Provincial People's Hospital from 2013 to 2014. None of the subjects enrolled in the study had taken corticosteroids, opioids, probiotic and prebiotic supplements or antibiotics in the 6 mo preceding the study, no one had systemic comorbidity, and no one had a history of excessive alcohol intake (> 20 alcoholic drinks per week). Patients with a prior history of gastrointestinal surgery or intestinal organic disease were excluded. All subjects provided signed informed consent before participation. The study was performed in accordance with the principles of the Declaration of Helsinki and the study protocol was approved by the Ethics Committee of Shanxi Provincial People's Hospital, China.

Stool sample processing and DNA extraction

Faecal samples were collected at home before bowel preparation, frozen immediately at -20 °C, and transported within 6 h to the study centre, where they were stored at -80 °C until analysis. Bacterial DNA was extracted from the faecal samples with the QIAamp DNA Stool Mini Kit (Qiagen, Hilden, Germany), according to the manufacturer's protocol. The DNA concentrations were quantified with an Eppendorf BioSpectrometer (Eppendorf, Hamburg, Germany).

Mucosal sample collection and genomic DNA extraction

Colonic mucosal biopsies were collected from each subject during an unsedated flexible colonoscopy. Colonic mucosal biopsies were taken from the descending colon. After removal from the colon, the biopsies were immediately frozen at -20 °C and transported within 6 h to the study centre, where they were stored at -80 °C until analysis. Total genomic DNA was extracted from the colon samples with the QIAamp DNA Mini Kit (Qiagen), according to the manufacturer's instructions. The concentration and purity of the genomic DNA were measured with an Eppendorf BioSpectrometer.

PCR amplification of the V3-V4 region of bacterial 16S rRNA gene and Illumina sequencing

The bacterial genomic DNA was used as the template for amplification of the V3-V4 hypervariable region of the 16S rRNA gene with the forward primer 5'-GACTA

CHVGGGTATCTAATCC-3' and the reverse primer 5'-CC TACGGGNGGCWGCAG-3'.

Bioinformatics analysis

Pairs of reads from the original DNA fragments were merged using FLASH, which was designed in case the original DNA fragments were shorter than twice the read length. The sequencing reads of each sample were given a unique barcode and analysed with the Quantitative Insights Into Microbial Ecology (QIIME) software and the UPARSE pipeline. In brief, the reads were filtered with the QIIME quality filters using the default settings for Illumina processing and the operational taxonomic units (OTUs) were selected using the UPARSE pipeline. The samples were sequenced on an Illumina MiSeq Benchtop Sequencer and the bioinformatic analysis was performed by Genesky Biotechnologies Inc. (Shanghai, China).

The size of the bacterial groups is expressed as percentage of the total bacteria.

Quantitative real-time polymerase chain reaction

Total mucosal RNA was extracted from the colonic biopsies using the TaKaRa MiniBEST Universal RNA Extraction Kit (TaKaRa, Japan), according to the manufacturer's instructions. After reverse transcription with PrimeScript Reverse Transcriptase Mix (TaKaRa), the expression of *TLR2* and *TLR4* genes was determined by quantitative real-time polymerase chain reaction (qPCR) and SYBR Green technology on a Bio-Rad CFX96 Q-PCR instrument (Bio-Rad, United States) in duplicate. The specific primers for TLR2 were: 5'-TGATGCTGCCATTCTCATTG-3' (forward) and 5'-CGC AGCTCTCAGATTTACCC-3' (reverse); and for TLR4: 5'-CAGGGCTTTTCTGAGTCGTC-3' (forward) and 5'-TG AGCAGTCGTGCTGGTATC-3' (reverse). Each amplification reaction was run in duplicate in a final volume of 20 µL. All qPCR amplifications were optimized and performed in 0.2 mL 96-well plates with the following cycling program: initial denaturation at 95 °C for 10 min, followed by 40 cycles of 95 °C for 15 s and 60 °C for 30 s. Negative controls lacking the template DNA were included in triplicate. The relative amount of each mRNA was normalized to *GAPDH* (forward: 5'-CCATCAATGACCCCTTCATTG-3', reverse: 5'-CTTGAC GGTGCCATGGAATT-3') and data were analysed using the $2^{-\Delta\Delta CT}$ method.

Statistical analysis

All statistical analyses were performed with SPSS 22.0 for Windows (SPSS Inc., United States). To determine the statistical differences between the two groups, we used the independent *t* test and the Mann-Whitney test. Correlations were determined with Spearman's correlation test. Heat maps were generated by R-packages for TLR2 and TLR4 as environmental factors, to reveal the relationship between the diversity

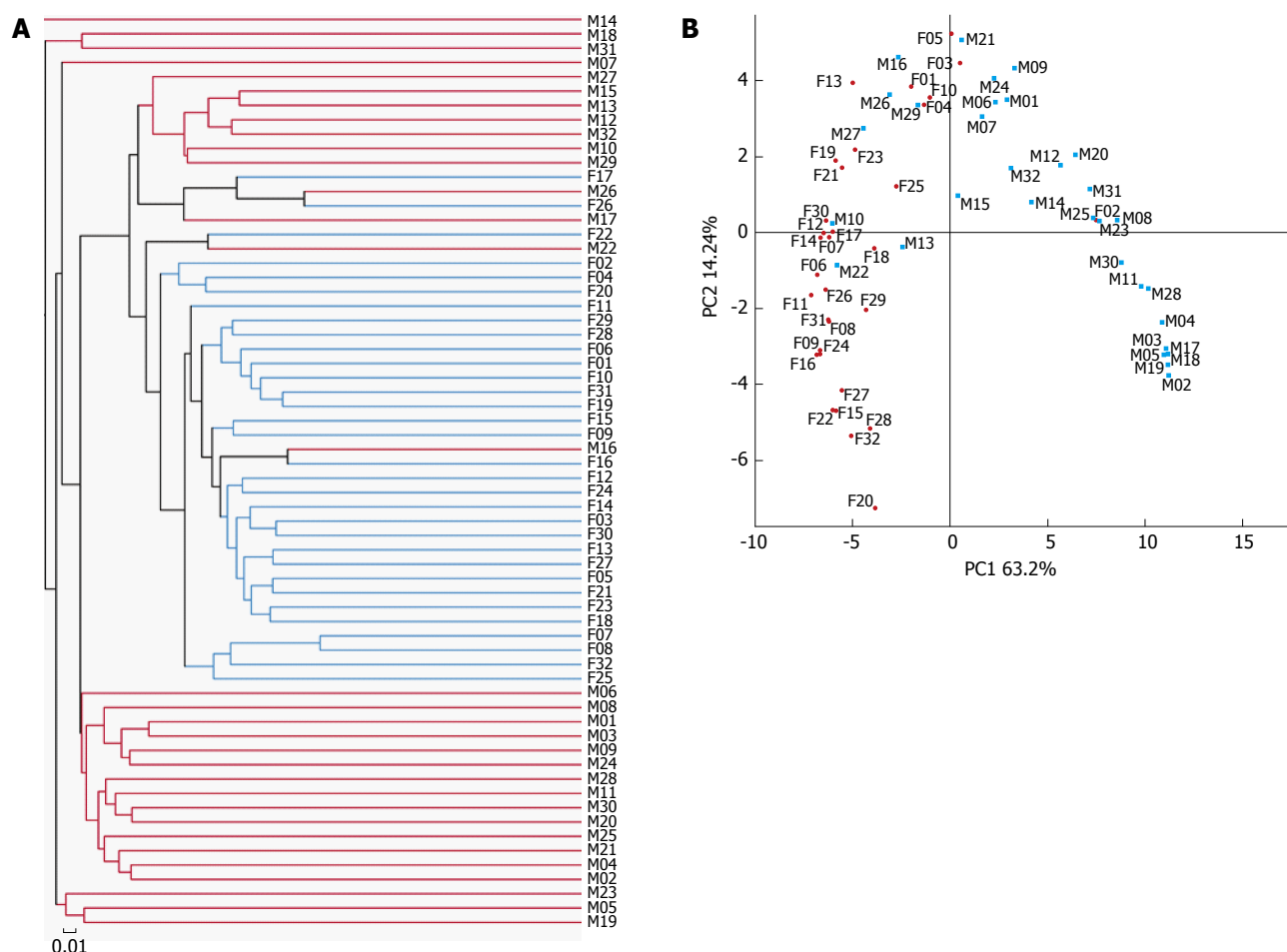


Figure 1 16S *rRNA* gene surveys reveal a clear separation between luminal microbiota and mucosa-associated microbiota. A: Dendrogram obtained from the complete linkage hierarchical clustering of the stool and mucosa samples based on the total operational taxonomic units; B: Principal coordinate analysis plot based on the weighted UniFrac metric.

Table 1 Comparison of the richness and diversity of mucosa-associated microbiota and luminal microbiota

	ACE	Chao	Shannon	Simpson
MAM	160.72 ± 51.62	151.56 ± 48.63	2.29 ± 1.24	0.33 ± 0.32
LM	195.50 ± 42.04	188.65 ± 42.70	2.901 ± 0.54	0.13 ± 0.07
<i>P</i> value	0.004 ^b	0.002 ^b	0.012 ^a	0.001 ^b

^a*P* < 0.05, ^b*P* < 0.01, MAM *vs* LM. MAM: Mucosa-associated microbiota; LM: Luminal microbiota.

and environmental factors based on the Pearson's correlation analyses.

RESULTS

Study population

We investigated 64 samples from 32 subjects with FGIDs [14 with diarrhoea-predominant IBS, 14 with functional dyspepsia (FD), and 4 with other diseases]. All subjects provided both a faecal sample and a colonic mucosal sample. The study population consisted of 50% females, and had a mean age of 49 years (range, 20-65 years) and a mean body mass

index of 23.24 kg/m² (range, 20-26 kg/m²).

Characteristics of the pyrosequencing results

We obtained a total of 4351929 raw reads and 3545053 reads remained after filtering. The sequencing analysis of the 64 samples identified 1026 OTUs. The rarefaction curves had a tendency to approach a saturation plateau, indicating that the number of samples in this study was sufficient. The same tendency was found in the Shannon-Wiener curves, indicating that the number of 16S *rRNA* gene sequences in the database was very abundant and reflected the vast majority of the microbiota. The gut faecal samples showed significantly more diversity and richness of the microbiota than the mucosal samples (Table 1) and samples of LM and MAM were significantly different (Figure 1A and B).

Significant differences between samples of LM and MAM were identified for almost all populated phyla. Bacteroidetes (44.7%) and Firmicutes (42.2%) were the most represented phyla in LM samples, followed by Proteobacteria (8.5%), whereas Proteobacteria (56.6%) was the most represented phylum in MAM samples, followed by Firmicutes (20.2%) and Bacteroidetes (12.7%) (*P* < 0.05; Figure 2).

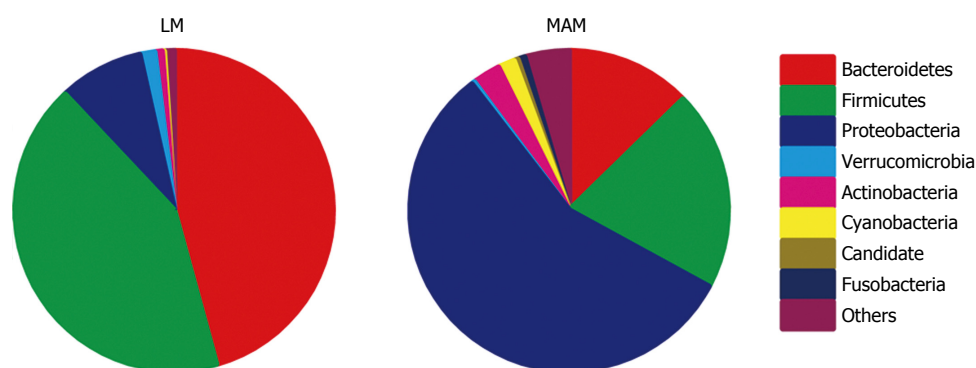


Figure 2 Relative abundance of bacterial genera in the stool and mucosa samples. Statistically significant differences were observed for all genera ($P < 0.05$). LM: Luminal microbiota; MAM: Mucosa-associated microbiota.

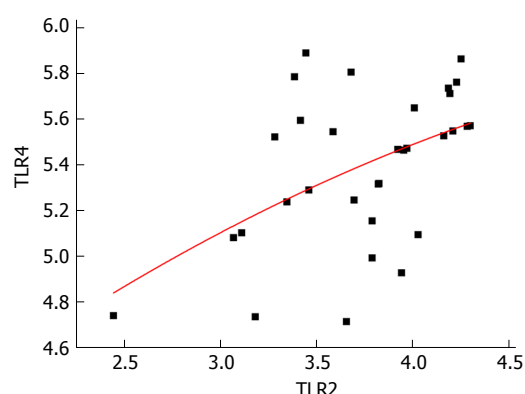


Figure 3 The expression of toll-like receptor 2 is significantly and positively correlated with toll-like receptor 4 expression ($r = 0.492$, $P = 0.004$). TLR: Toll-like receptor.

Examination at the genus level showed that the relative abundance of *Escherichia-Shigella*, *Streptococcus*, *Clostridium sensu stricto*1, *Sphingomonas*, *Acinetobacter*, *Brevundimonas*, and *Enhydrobacter* was significantly greater in samples of MAM than in LM samples, whereas abundance of *Bacteroides*, *Faecalibacterium*, *Incertain sedis*, *Subdoligranulum*, *Pseudobutyryvibrio*, *Megasphaera*, *Parasutterella*, *Akkermansia*, *Alistipes*, and *Lachnospira* was significantly lower in samples of MAM than in LM samples ($P < 0.05$).

Expression characteristics of mucosal TLR2 and TLR4

We measured the mRNA expression of TLR2 (3.7394 ± 0.43514) and TLR4 (5.3866 ± 0.33556) in all samples and expression of TLR2 was significantly and positively correlated with that of TLR4 ($r = 0.492$, $P = 0.004$) (Figure 3).

Correlation between gut microbiota and TLR4 expression

Regardless of phylum, class or order level, no bacterium was correlated with expression of TLR4 in LM samples.

At the phylum level, Firmicutes was inversely and significantly correlated with TLR4 expression in MAM samples ($r = -0.4676$, $P = 0.0070$). At the class level, Clostridia, which belongs to Firmicutes, was inversely

and significantly correlated with TLR4 expression in MAM samples ($r = -0.3913$, $P = 0.0268$). At the order level among the class Clostridia, the order Clostridia was inversely and significantly correlated with TLR4 expression ($r = -0.3906$, $P = 0.0271$), as well as the families Defluviitaleaceae ($r = -0.4227$, $P = 0.0159$), Peptostreptococcaceae ($r = -0.3611$, $P = 0.0422$), and Ruminococcaceae ($r = -0.4740$, $P = 0.0061$), which belong to the order Clostridia. The genera *Faecalibacterium* and *Ruminococcus*, which belong to the family Ruminococcaceae, were also inversely and significantly correlated with TLR4 expression ($r = -0.45817$, $P = 0.0083$ and $r = -0.5306$, $P = 0.0018$, respectively) (Figure 4).

Correlation between gut microbiota and TLR2 expression

The presence of bacteria in both LM and MAM samples was correlated with TLR2 expression. However, the taxa of microbiota was very different in these two sample populations.

In LM samples, Clostridia and Bacilli at the class level were inversely and significantly correlated with TLR2 expression ($r = -0.4238$, $P = 0.0156$ and $r = -0.4482$, $P = 0.0101$, respectively), as well as Clostridiales at the order level, which belongs to Clostridia, and Lactobacillales, which belongs to Bacilli ($r = -0.4482$, $P = 0.0156$ and $r = -0.4207$, $P = 0.0165$, respectively). At the family level, Ruminococcaceae, which belongs to Clostridiales, and Streptococcaceae, which belongs to Lactobacillales, were inversely and significantly correlated with TLR2 expression ($r = -0.4437$, $P = 0.0110$ and $r = -0.3839$, $P = 0.0300$, respectively). And at the genus level, *Faecalibacterium*, which belongs to Ruminococcaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely and significantly correlated with TLR2 expression ($r = -0.5743$, $P = 0.0058$ and $r = -0.3905$, $P = 0.0271$, respectively) (Figure 4).

In MAM samples, Bacilli at the class level was inversely and significantly correlated with TLR2 expression ($r = -0.4676$, $P = 0.0070$), as well as Lactobacillales at the order level, which belongs to

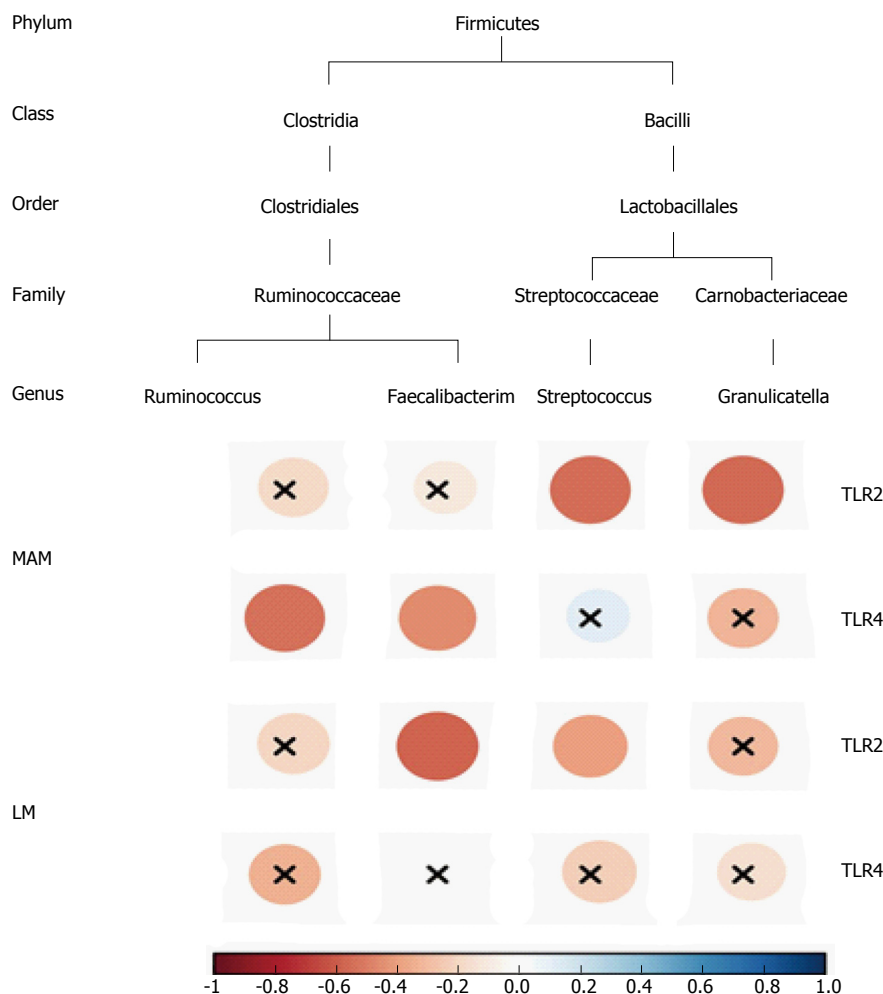


Figure 4 Correlogram showing the Pearson's correlation between key genera and toll-like receptors in both faecal and mucosa samples. Blue circles designate a positive correlation, while red ones designate a negative correlation. X means no significant result according to the significance level of 0.05. TLR: Toll-like receptor; MAM: Mucosa-associated microbiota; LM: Luminal microbiota.

Bacilli ($r = -0.4574$, $P = 0.0085$). At the family level, Carnobacteriaceae and Streptococcaceae, which belong to Lactobacillales, were inversely and significantly correlated with TLR2 expression ($r = -0.5554$, $P = 0.0010$ and $r = -0.5445$, $P = 0.0013$, respectively). And at the genus level, *Granulicatella*, which belongs to Carnobacteriaceae, and *Streptococcus*, which belongs to Streptococcaceae, were correlated with TLR2 expression ($r = -0.5573$, $P = 0.0010$ and $r = -0.5435$, $P = 0.0013$, respectively) (Figure 4).

DISCUSSION

Deep sequencing showed that more than 95% of the sequences in all stool and mucosal samples belonged to the three most popular bacterial phyla, consisting of Firmicutes, Bacteroidetes, and Proteobacteria. This is consistent with the findings of previous studies^[6], which showed that these phyla account for the majority of the gut microbiota in both stool and mucosal samples. The faecal samples displayed significantly greater bacterial diversity and richness than the mucosal samples, as in the study

of Ringel *et al.*^[2]. Comparing the proportions of the dominant bacterial taxa in the faecal and mucosal samples revealed significant differences. In this study, Proteobacteria was the predominant phylum in MAM samples. This is different from other reports and might reflect geographical differences^[7]. It is well known that the Chinese diet and genetics are very different from those in Western countries and these factors markedly influence the gut microbiota. Furthermore, MAM samples were taken from a distinct location in the colon, whereas LM samples displayed the whole intestinal microbiota. Therefore, MAM samples may reflect a stronger relationship with the host.

A correlation analysis was performed between the two bacterial populations at the phylum, class, order, family, and genus levels and the expression of TLR2 and TLR4. The results showed that although the Proteobacteria was the predominant phylum in MAM samples, the bacteria that correlated strongest with expression of TLRs were part of the phylum Firmicutes. The taxa that showed a significant correlation was selected and studied from the phylum to genus level. Finally, *Faecalibacterium*, *Ruminococcaceae*,

Streptococcus and *Granulicatella* genera were the bacteria that correlated strongest with TLRs expression. Although a positive correlation existed between the expression of TLR2 and TLR4, the types of bacteria associated with TLR4 expression were very different from those related to the expression of TLR2. However, this was in accordance with their distinct microbial compositions. The bacteria that correlated with TLR4 expression were only found in MAM samples. Bacteria that correlated with TLR2 expression originated from both LM and MAM. Both TLR4 and TLR2 expression were negatively correlated with MAM, however, TLR2 expression was mainly associated with bacteria belonging to the Bacilli class, whereas TLR4 expression was mainly associated with bacteria belonging to the Clostridia class.

The enteric commensal bacteria of the genus *Faecalibacterium*, which belongs to the group Clostridium, exert an anti-inflammatory effect^[8]. In the present study, *Faecalibacterium* from LM samples was negatively correlated with the expression of TLR2 and *Faecalibacterium* from MAM samples was negatively correlated with the expression of TLR4. *Faecalibacterium* richness was reduced in patients with IBS and IBD^[9,10]. TLR signalling, activated by pathogens, is involved in the pathogenesis of several infectious and inflammatory diseases. Imbalanced relationships among the environment, genetics, and host immunity may promote aberrant TLR signalling, thereby contributing on a large scale to acute and chronic intestinal inflammatory processes, such as IBD, colitis, and colorectal cancer^[11]. In our previous study, the expression of TLR2 and TLR4 was increased in diarrhoea-predominant IBS patients and decrease of Clostridia was strongly associated with higher TLR2 and TLR4 expression^[12]. In an animal study, *Faecalibacterium prausnitzii* and its supernatant (SN) had beneficial effects on intestinal epithelial barrier impairment in a chronic low-grade inflammation model^[13]. Round *et al.*^[14] showed that unlike pathogens whose TLR ligands trigger inflammation, some commensal bacteria exploit the TLR pathway to actively suppress immune reactions. The anti-inflammatory effect of *Faecalibacterium* may be involved in the upregulation of the TLRs expression. *Ruminococcus* is another common bacterium that is decreased in patients with IBD or IBS. In addition, a reduction in the relative abundance of potentially immunomodulatory gut bacteria including *Ruminococcus* is associated with exaggerated inflammatory cytokine responses to TLR ligands and subsequent development of IgE-associated eczema^[15]. In the present study, the decrease of *Faecalibacterium* and *Ruminococcus* in MAM samples was strongly associated with higher TLR4 expression, suggesting that the decrease of mucosal *Faecalibacterium* and *Ruminococcus* induced an increase in expression of TLR4 mRNA, which plays an important role in the causation of immune related gut diseases including IBD and IBS.

Streptococcus is a genus of (spherical) coccus Gram-positive bacteria belonging to the phylum Firmicutes phylum and the order Lactobacillales. Some streptococcal species, e.g., *S. pneumoniae*, are pathogenic. Tomlinson *et al.*^[16] suggested that leukocyte responses to bacterial lipoproteins are required for TLR2- and IL-1R-associated kinase-4-mediated inflammatory responses to *S. pneumoniae*. However, many other streptococcal species, e.g., *S. thermophilus*, are not pathogenic^[17] and are part of the commensal human microbiota of the mouth, skin, intestine, and upper respiratory tract. *S. thermophilus* is considered a probiotic and is a key strain of VSL#3, which is considered a therapeutic immunomodulatory tool^[18]. Streptococcaceae was negatively correlated with TLR2 expression in samples of both MAM and LM in our study. Although there is no direct evidence of inhibition of TLRs by Streptococcaceae, some beneficial streptococcal species may affect disease processes via the TLR2 pathway. Future research will explore which species belonging to Streptococcaceae can influence the expression of TLR2. *Granulicatella* is part of the normal body flora and is often difficult to culture in traditional media. A few reports mention that it can induce an infection similar to Streptococci^[19].

The composition of LM samples was different than that of MAM; MAM samples showed a stronger correlation with the expression of TLR2 and TLR4. The abundance of *Faecalibacterium* and *Ruminococcus* was lower in IBD and IBS, while TLRs expression was higher in patients than in healthy controls. *Faecalibacterium* and *Ruminococcus* were significantly and negatively correlated with TLR4 expression, suggesting that these two bacteria colonizing on the colonic mucosa play a key role in the pathogenesis of gut diseases by regulating the expression of mucosal TLR4. *Granulicatella*, which belongs to Carnobacteriaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely correlated with TLR2 expression.

One limitation of our study was the small biopsy from the colonic endoscopy and multiple sampling might cause bleeding, therefore we were not able to measure the expression of proteins. The other limitation of our study was the scattered distribution of the correlation between species levels, which might be explained by the restriction of pyrosequencing of species levels, therefore we did not report the results. It will be important to establish a better understanding of the relationship between TLRs and bacteria strains in future studies. Probiotics play an important role in the treatment and prevention of diseases. Further studies will have to verify the relationship between TLRs and bacteria^[20,21].

COMMENTS

Background

Toll-like receptors (TLRs) detect conserved microbial products and play a

central role in the activation of innate and adaptive immune pathways. TLR2 and TLR4 are two of the best characterized TLRs that respond to microbial membrane components. Faecal samples (representing the luminal niche) were used in most studies on intestinal microbiota because their collection is easy. However, more recent studies revealed distinct microbial composition between luminal microbiota (LM) and mucosa-associated microbiota (MAM). To explore which bacteria may regulate the expression of TLRs and thereby affect intestinal functions, we performed high-throughput pyrosequencing of the bacterial 16S rRNA gene, compared the microbial communities in the faeces and mucosa of Chinese patients with functional gastrointestinal disorders, and studied their association with the expression of colonic mucosal TLR2 and TLR4. LM differed from MAM; MAM showed a stronger association with the expression of TLR2 and TLR4.

Research frontiers

Recent advances in the understanding of intestinal immunology suggest that functional gastrointestinal disorders (FGIDs) may not all be 'functional', as considered for decades. This view was largely developed by the absence of active inflammation. Faecal microbiota and MAM play a vital role in gut immunity. It will be important to establish a better understanding of the relationship between TLRs and bacteria strains in FGIDs.

Innovations and breakthroughs

In some intestinal diseases, such as inflammatory bowel disease or irritable bowel syndrome, the expression of TLR2 and TLR4 is increased and dysbiosis is observed. However, limited data are available on the correlation between the intestinal microbiota and TLRs in humans. In this study, the abundance of *Faecalibacterium* and *Ruminococcus* was lower in many gut diseases, while the TLRs expression was higher than in healthy controls. *Faecalibacterium* and *Ruminococcus* were significantly and negatively correlated with TLR4 expression, suggesting that these two bacteria colonizing on the colonic mucosa play a key role in gut diseases by regulating the expression of mucosal TLR4. *Granulicatella*, which belongs to Carnobacteriaceae, and *Streptococcus*, which belongs to Streptococcaceae, were inversely correlated with TLR2.

Applications

Probiotics play an important role in the treatment and prevention of diseases. Further studies will have to verify the relationship between inflammatory proteins and bacteria. The bacteria may be used as probiotics in the treatment of diseases by regulating immunity.

Terminology

According to the literature, they described the microbiota from faecal samples as LM.

Peer-review

This is an interesting paper that investigated the intestinal LM and the MAM in Chinese patients with FGIDs and examined the association between these communities and the expression of TLR2 and TLR4. The manuscript is well written.

REFERENCES

- Kataoka K. The intestinal microbiota and its role in human health and disease. *J Med Invest* 2016; **63**: 27-37 [PMID: 27040049 DOI: 10.2152/jmi.63.27]
- Ringel Y, Maharshak N, Ringel-Kulka T, Wolber EA, Sartor RB, Carroll IM. High throughput sequencing reveals distinct microbial populations within the mucosal and luminal niches in healthy individuals. *Gut Microbes* 2015; **6**: 173-181 [PMID: 25915459 DOI: 10.1080/19490976.2015.1044711]
- Sundin J, Rangel I, Fuentes S, Heikamp-de Jong I, Hultgren-Hörnquist E, de Vos WM, Brummer RJ. Altered faecal and mucosal microbial composition in post-infectious irritable bowel syndrome patients correlates with mucosal lymphocyte phenotypes and psychological distress. *Aliment Pharmacol Ther* 2015; **41**: 342-351 [PMID: 25521822 DOI: 10.1111/apt.13055]
- Kramer CD, Genco CA. Microbiota, Immune Subversion, and Chronic Inflammation. *Front Immunol* 2017; **8**: 255 [PMID: 28348558 DOI: 10.3389/fimmu.2017.00255]
- de Kivit S, Tobin MC, Forsyth CB, Keshavarzian A, Landay AL. Regulation of Intestinal Immune Responses through TLR Activation: Implications for Pro- and Prebiotics. *Front Immunol* 2014; **5**: 60 [PMID: 24600450 DOI: 10.3389/fimmu.2014.00060]
- Thursby E, Juge N. Introduction to the human gut microbiota. *Biochem J* 2017; **474**: 1823-1836 [PMID: 28512250 DOI: 10.1042/BCJ20160510]
- Escobar JS, Klotz B, Valdes BE, Agudelo GM. The gut microbiota of Colombians differs from that of Americans, Europeans and Asians. *BMC Microbiol* 2014; **14**: 311 [PMID: 25495462 DOI: 10.1186/s12866-014-0311-6]
- Hornef MW, Pabst O. Real friends: *Faecalibacterium prausnitzii* supports mucosal immune homeostasis. *Gut* 2016; **65**: 365-367 [PMID: 26531718 DOI: 10.1136/gutjnl-2015-310027]
- Machiels K, Joossens M, Sabino J, De Preter V, Arijis I, Eeckhaut V, Ballet V, Claes K, Van Immerseel F, Verbeke K, Ferrante M, Verhaegen J, Rutgeerts P, Vermeire S. A decrease of the butyrate-producing species *Roseburia hominis* and *Faecalibacterium prausnitzii* defines dysbiosis in patients with ulcerative colitis. *Gut* 2014; **63**: 1275-1283 [PMID: 24021287 DOI: 10.1136/gutjnl-2013-304833]
- Lopez-Siles M, Martinez-Medina M, Abellà C, Busquets D, Sabat-Mir M, Duncan SH, Aldeguer X, Flint HJ, Garcia-Gil LJ. Mucosa-associated *Faecalibacterium prausnitzii* phylotype richness is reduced in patients with inflammatory bowel disease. *Appl Environ Microbiol* 2015; **81**: 7582-7592 [PMID: 26296733 DOI: 10.1128/AEM.02006-15]
- Frosali S, Pagliari D, Gambassi G, Landolfi R, Pandolfi F, Cianci R. How the Intricate Interaction among Toll-Like Receptors, Microbiota, and Intestinal Immunity Can Influence Gastrointestinal Pathology. *J Immunol Res* 2015; **2015**: 489821 [PMID: 26090491 DOI: 10.1155/2015/489821]
- Guo WT, Liu P, Dong LN, Wang JP. [The correlation study between the changes of intestinal mucosa predominant bacteria and Toll-like receptor 2, Toll-like receptor 4 gene expressions in diarrhea-predominant irritable bowel syndrome patients]. *Zhonghua Neike Zazhi* 2016; **55**: 541-543 [PMID: 27373290 DOI: 10.3760/cma.j.issn.0578-1426.2016.07.011]
- Martín R, Miquel S, Chain F, Natividad JM, Jury J, Lu J, Sokol H, Theodorou V, Bercik P, Verdu EF, Langella P, Bermúdez-Humarán LG. *Faecalibacterium prausnitzii* prevents physiological damages in a chronic low-grade inflammation murine model. *BMC Microbiol* 2015; **15**: 67 [PMID: 25888448 DOI: 10.1186/s12866-015-0400-1]
- Round JL, Lee SM, Li J, Tran G, Jabri B, Chatila TA, Mazmanian SK. The Toll-like receptor 2 pathway establishes colonization by a commensal of the human microbiota. *Science* 2011; **332**: 974-977 [PMID: 21512004 DOI: 10.1126/science.1206095]
- West CE, Rydén P, Lundin D, Engstrand L, Tulic MK, Prescott SL. Gut microbiome and innate immune response patterns in IgE-associated eczema. *Clin Exp Allergy* 2015; **45**: 1419-1429 [PMID: 25944283 DOI: 10.1111/cea.12566]
- Tomlinson G, Chimalapati S, Pollard T, Lapp T, Cohen J, Camberlein E, Stafford S, Periseleris J, Aldridge C, Vollmer W, Picard C, Casanova JL, Noursadeghi M, Brown J. TLR-mediated inflammatory responses to *Streptococcus pneumoniae* are highly dependent on surface expression of bacterial lipoproteins. *J Immunol* 2014; **193**: 3736-3745 [PMID: 25172490 DOI: 10.4049/jimmunol.1401413]
- Marcial G, Villena J, Faller G, Hensel A, de Valdéz GF. Exopolysaccharide-producing *Streptococcus thermophilus* CRL1190 reduces the inflammatory response caused by *Helicobacter pylori*. *Benef Microbes* 2017; **8**: 451-461 [PMID: 28504579 DOI: 10.3920/BM2016.0186]
- Ekmekci I, von Klitzing E, Fiebigler U, Neumann C, Bacher P, Scheffold A, Bereswill S, Heimesaat MM. The Probiotic Compound VSL#3 Modulates Mucosal, Peripheral, and Systemic

- Immunity Following Murine Broad-Spectrum Antibiotic Treatment. *Front Cell Infect Microbiol* 2017; **7**: 167 [PMID: 28529928 DOI: 10.3389/fcimb.2017.00167]
- 19 **York J**, Fisahn C, Chapman J. Vertebral Osteomyelitis Due to *Granulicatella Adiacens*, a Nutritionally Variant Streptococci. *Cureus* 2016; **8**: e808 [PMID: 27800289]
- 20 **Uccello M**, Malaguarnera G, Basile F, D'agata V, Malaguarnera M, Bertino G, Vacante M, Drago F, Biondi A. Potential role of probiotics on colorectal cancer prevention. *BMC Surg* 2012; **12** Suppl 1: S35 [PMID: 23173670 DOI: 10.1186/1471-2482-12-S1-S35]
- 21 **Sánchez B**, Delgado S, Blanco-Míguez A, Lourenço A, Gueimonde M, Margolles A. Probiotics, gut microbiota, and their influence on host health and disease. *Mol Nutr Food Res* 2017; **61**: [PMID: 27500859 DOI: 10.1002/mnfr.201600240]

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Case Control Study

Genetic biomarkers for hepatocellular cancer risk in a caucasian population

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Abstract

AIM

To uncover novel genetic markers that could contribute to predicting hepatocellular carcinoma (HCC) susceptibility in Caucasians.

METHODS

The present retrospective case-control study compared genotype frequencies between a cohort of HCC cases and two, independent, HCC-free, age/sex-matched control groups. The HCC cohort comprised 192 homogeneous patients that had undergone orthotopic liver transplantation. The first control group comprised 167 patients that were matched to the HCC cohort for the percentage of hepatitis B (HBV) and/or hepatitis C (HCV) infections. A second control group included 192 virus-free, healthy individuals that were used to evaluate the generalizability of the identified predictive markers. All cases and controls were Caucasian. The three study populations were characterized with a panel of 31 markers derived from 21 genes that encoded key proteins involved in hepatocarcinogenesis-related pathways. The study end-point was to assess the association between genetic variants and HCC onset.

RESULTS

Five genetic markers were identified as risk factors for HCC in high-risk patients infected with HBV/HCV. According to a dominant model, reduced HCC risk was associated with three polymorphisms: *ERCC1* rs3212986 (OR = 0.46, 95%CI: 0.30-0.71, $P = 0.0005$), *GST-P1* rs1138272 (OR = 0.41, 95%CI: 0.21-0.81, $P = 0.0097$), and *CYP17A1* rs743572 (OR = 0.50, 95%CI: 0.31-0.79, $P = 0.0032$). Conversely, according to a recessive model, increased HCC risk was associated with two polymorphisms: *XRCC3* rs1799794 (OR = 3.70, 95%CI: 1.02-13.39, $P = 0.0461$) and *ABCB1* rs1128503 (OR = 2.06, 95%CI: 1.18-3.61, $P = 0.0111$). These associations remained significant in a subgroup analysis, where patients were stratified according to viral status (HBV- or HCV-positive serology). Two variants exhibited a serology-specific effect: *ABCB1* rs1128503 (OR = 4.18, 95%CI: 1.55-11.29, $P = 0.0048$) showed an effect in the HBV-positive subgroup; and *ERCC1* rs3212986 (OR = 0.33, 95%CI: 0.18-0.60, $P = 0.0003$) showed an effect in the HCV-positive subgroup. Among the five markers identified, *ERCC1* rs3212986 (OR = 0.43, $P < 0.0001$) and *CYP17A1* rs743572 (OR = 0.73, $P = 0.0310$) had a different distribution in patients with HCC compared to healthy individuals. With a recursive partitioning approach, we also demonstrated that significant gene-gene interactions between *ERCC1* rs3212986, *CYP17A1* rs743572, *GST-P1* rs1138272, and the previously described *UGT1A7**3 predictive marker, played a role in the complex trait of HCC susceptibility.

CONCLUSION

We identified five polymorphisms and interactions that contributed crucially to predicting HCC risk. These findings represented an important step towards improving HCC diagnosis and management.

Key words: ERCC1; XRCC3; GST-P1; CYP17A1; MDR-1; Polymorphisms; Hepatocellular carcinoma risk; Early diagnosis; Antiviral therapy; Hepatitis B viral/hepatitis C viral

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Core tip: It is a great challenge to define new biomarkers of hepatocellular carcinoma (HCC) risk for HCC management. This work identified five genetic markers in key pathways linked to hepatocarcinogenesis (*ERCC1* rs3212986, *GSTP1* rs1138272, *CYP17A1* rs743572, *XRCC3* rs1799794, and *ABCB1* rs1128503), which could predict individual HCC susceptibility, particularly in a high-risk [hepatitis B viral/hepatitis C viral (HCV)-infected] Caucasian population. We also identified potential gene-gene interactions that should be included in the definition of HCC risk. These findings could contribute to improved HCC surveillance, early cancer diagnoses, and potential curative therapies. For patients with HCV, these markers could be considered selection criteria for the personalized use of recently developed, highly expensive anti-viral therapies.

De Mattia E, Cecchin E, Polesel J, Bignucolo A, Roncato R, Lupo F, Crovatto M, Buonadonna A, Tiribelli C, Toffoli G. Genetic biomarkers for hepatocellular cancer risk in a caucasian population. *World J Gastroenterol* 2017; 23(36): 6674-6684 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6674.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6674>

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide, with a particularly high prevalence in some areas of Asia and Africa. HCC represents the second leading cause of cancer-related deaths^[1]. Although liver cancer is less frequent in Western developed countries, recent data indicate that, due to dissemination of hepatitis B (HBV) and C (HCV) viral infections, the HCC incidence is also dramatically rising, in both the United States and Europe^[1,2].

Unlike most malignancies, the major risk factors for HCC development are well-defined. These risk factors are chronic HCV and HBV infections, liver cirrhosis, heavy alcohol intake, tobacco smoking, exposure to environmental and dietary carcinogens (*i.e.*, aflatoxin B1), genetic and metabolic liver disease (*i.e.*, hereditary hemochromatosis, non-alcoholic steatohepatitis), and other conditions capable of inducing liver damage^[2]. Moreover, the prevalence of HCC increases with age and male sex^[1-3].

In addition to environmental factors, recent insights into the biology of HCC have demonstrated that accumulations of genetic and epigenetic abnormalities can also play an essential role in liver carcinogenesis. In particular, data derived from candidate-gene investigations and, more recently, genome-wide associations studies have highlighted the possible role that genetic variants might play as significant determinants of HCC susceptibility^[4-6]. Additionally, host genetic factors were shown to affect the clinical course of HBV or HCV infections, because they

influence the individual's predisposition to disease progression and hepatocarcinogenesis^[7,8]. However, the currently available studies are highly heterogeneous in study design, sample size, ethnicity, and clinical-demographic features of the investigated population; these differences make it difficult to draw solid conclusions^[4,6]. Moreover, the majority of case-control investigations were performed with Asian populations; thus, replication studies with different ethnicities are warranted^[4-8].

It is of great clinical interest to identify genetic traits that can modify liver carcinogenesis, because they can potentially be used to improve preventive, diagnostic, and therapeutic strategies in HCC management^[3,9,10]. To date, the majority of patients with HCC are diagnosed at an advanced tumor stage, which precludes potentially curative therapies, including orthotopic liver transplantation (OLT), surgical resection, and local ablation^[3,9,10]. The HCC surveillance programs that could lead to early cancer detection and effective treatment are currently based on highly heterogeneous scoring systems that require costly measures^[3,9,10]. The identification of additional molecular biomarkers predictive of individual HCC susceptibility, particularly in a high-risk population (*i.e.*, HBV/HCV-positive), could be important in improving current guidelines for genetics-based patient screening and preventive strategies. In some countries, including the United States and Europe, HCV-related HCC accounts for most liver cancer incidents^[11]. Consequently, an increased understanding of the genetic contribution to the clinical course of viral infection could be essential in optimizing the use of newly developed, but highly expensive, direct acting antiviral (DAA) treatments^[12,13].

We previously reported that the low-activity alleles, *UGT1A1**28, *UGT1A7**3, *UGT1A9**22, and related haplotypes may play a protective role against HCC development in Caucasian populations. These alleles positively modulate the serum levels of beneficial molecules, like bilirubin^[14]. The present study aimed to extend genetic analyses by identifying additional molecular markers that could be integrated into the HCC risk stratification algorithms used in liver cancer prevention strategies. To that end, we analyzed a set of potential candidate HCC-risk markers with low penetrance. These candidate markers were based on functionally relevant polymorphic variants in genes that encoded proteins involved in key pathways underlying carcinogenesis^[6,15-17]. The pathways we investigated were: (1) membrane transporters that play an essential role in xenobiotic defense and prevent the entry of toxic environmental compounds that might damage liver tissue; (2) DNA synthesis and repair mechanisms that modulate the cell's capacity to respond to DNA damage induced by toxic agents, and thus, these mechanisms affect the accumulation of mutations in DNA; (3) detoxification systems that facilitate the elimination of detrimental endo/exogenous compounds; and (4) systems that defend

against oxidative stress and related cellular damage, which can play a crucial role in the development and progression of HCC. We designed a retrospective case-control study and enrolled a homogeneous group of patients with HCC that had received OLT and two age/sex-matched control groups (a group positive for HBV and/or HCV and a healthy population). We aimed to identify novel genetic markers that could contribute to the determination of individual HCC risk, particularly in Caucasian populations.

MATERIALS AND METHODS

Study population

We studied 192 patients with histologically confirmed HCC that had undergone OLTs between May 1991 and March 2006 at the Liver Transplantation Center of the S. Giovanni Battista Hospital (Turin, Italy). All patients were infected with HBV and/or HCV and had a cirrhotic liver. The first control group included 167 subjects with HBV and/or HCV infections, but no evidence of cancer (including HCC). These subjects were followed up for hepatitis infections between November 2009 and September 2010 at the Department of Laboratory Medicine, S. Maria degli Angeli Hospital (Pordenone, Italy). This control group was matched to the HCC group to ensure equivalent percentages of patients with HBV and/or HCV and similar distributions of gender and age (in quinquennia), when possible.

The second control group included 192 healthy subjects that were matched to patients with HCC for gender and age (in quinquennia). For this control group, individuals under 70 years of age were randomly selected from a pool of blood donors that visited the Centro di Riferimento Oncologico National Cancer Institute in Aviano, Italy. This control group also included individuals aged ≥ 70 years that were cancer-free without HBV/HCV infections. These individuals had been approached during occasional blood testing at the S. Maria degli Angeli Hospital, and they consented to provide a 5-mL blood sample for research purposes.

All cases and controls were Italian Caucasian individuals, and they all provided written informed consent for genetic analysis and for inclusion in the study. The study protocol conformed to the ethics guidelines of the 1975 Declaration of Helsinki, and it was approved by the Institutional Review Board of each participating institution (Ethics Committee of Azienda Ospedaliera Universitaria, S. Giovanni Battista, Torino).

Polymorphism selection and genetic analyses

Genomic DNA was extracted from peripheral blood with the High Pure PCR Template Preparation Kit (Roche Diagnostics GmbH, Mannheim, Germany). The samples underwent standard proteinase K digestion and successive template purification. The genomic DNA solution was stored at 4 °C. Positive and negative control samples were included in each analysis.

We considered a set of 31 molecular markers that

Table 1 Distribution of hepatocellular carcinoma patients, hepatitis B/hepatitis C -positive individuals and healthy subjects according to socio-demographic and clinical characteristics *n* (%)

Characteristics	HCC group	HBV/HCV group	Healthy group
Sex			
Men	166 (86.5)	141 (84.4)	166 (86.5)
Women	26 (13.5)	26 (15.6)	26 (13.5)
Age (yr)			
< 55	85 (44.3)	85 (50.9)	96 (50.0)
55-59	60 (31.3)	19 (11.4)	35 (18.2)
≥ 60	47 (24.5)	63 (37.7)	61 (31.8)
Hepatitis infection			
None	0 (0.0)	0 (0.0)	192 (100.0)
HBV+ and HCV-	74 (38.5)	74 (44.3)	0 (0.0)
HBV- and HCV+	109 (56.8)	88 (52.7)	0 (0.0)
HBV+ and HCV+	9 (4.7)	5 (3.0)	0 (0.0)
Years of orthotopic liver transplantation or resection			
1991-1995	27 (14.1)		
1996-2000	60 (31.3)		
2001-2006	102 (54.7)		
Number of nodes			
1	110 (57.3)		
≥ 2	72 (37.5)		
Unknown	10 (5.2)		
Maximal dimension of nodes (cm)			
≤ 2	79 (41.2)		
2.1 to ≤ 3	67 (34.9)		
> 3	37 (19.3)		
Unknown	9 (4.7)		

HCC: Hepatocellular carcinoma; HBV: Hepatitis B viral; HCV: Hepatitis C viral.

represented functionally relevant polymorphisms (PubMed-MEDLINE search) in genes that encoded key proteins involved in hepatocarcinogenesis-related pathways. In particular, the biological processes evaluated (and related genes) were: membrane transport (*ABCB1*, *ABCC2*), DNA synthesis and repair mechanisms (*MTHFR*, *TYMS*, *ERCC1*, *ERCC2*, *XRCC1*, *XRCC3*, *APE1*, *hMLH1*, *hMSH2*, *hOOG1*), detoxifying systems (*CYP3A4*, *CYP3A5*, *CYP17A1*, *CYP2D6*), and oxidative stress response (*GST-M1*, *GST-T1*, *GST-P1*; *GSTA1*, *SOD2*).

All the genes, variants, and assays applied are listed in Supplementary Table 1. For pyrosequencing, we used PSQ96MA (Qiagen, Hilden, Germany). PCR amplifications were performed in an Eppendorf Mastercycler gradient, with TaqGold DNA Polymerase (AB Applied Biosystems, Foster City, CA, United States). The details of the pyrosequencing assays, primer sequences, and PCR conditions are available upon request. Pre-designed TaqMan single-nucleotide polymorphism genotyping assays were conducted with the Applera TaqMan Universal Master mix on an ABI 7900HT Real-time PCR system (AB Applied Biosystems, Foster City, CA, United States), according to the manufacturer's instructions for optimal allelic discrimination. All commercial TaqMan assays were purchased from the Applied Biosystems website (www.appliedbiosystems.com). Detailed protocols for

genotyping methods, based on gel electrophoresis and enzymatic digestion, are available upon request.

Statistical analysis

In this study, we assessed associations between genetic markers and the individual risk of developing HCC. In healthy controls, deviation from Hardy-Weinberg equilibrium was tested for each polymorphism with the χ^2 test, and no deviation was found ($P < 0.05$).

The odds ratio (OR) and 95%CI were estimated with unconditional logistic regression, which was controlled for matching variables. We also investigated three genetic models (dominant, recessive, and additive) for associations, and we reported the most statistically significant one, based on the Wald χ^2 -test. Statistical significance was set at $P < 0.05$ (two-sided). The analyses were performed with SAS 9.2, and a statistical review of the study was performed by a biomedical statistician.

Recursive partitioning was performed to elucidate high-order relationships among genetic factors, when stratifying patients between HCC cases and HBV/HCV group. The analysis was carried out with the five genetic variants that were identified in the present study as significant predictors of HCC risk and with the previously described *UGT1A* markers (*i.e.*, *UGT1A1*28*, *UGT1A9*22*, *UGT1A7*3*)^[14].

RESULTS

Patient characteristics

The demographic, clinical, and serological characteristics of the 192 patients with HCC and the two control groups are reported in Table 1. The predominant gender was male (86.5% of patients with HCC, 84.4% of patients in the HBV/HCV group, and 86.5% of healthy subjects). The median ages at the time of enrolment were 56, 54 and 55 years for the HCC, HBV/HCV, and healthy groups, respectively. The HCC cohort was clinicopathologically homogeneous; all patients fulfilled the surgical intervention criteria (*i.e.*, T1/T2 primary tumor stage, evaluated with the TNM scale; well or moderately poorly differentiated grade at diagnosis; and no detectable vascular invasion).

Genetic polymorphisms and HCC risk

The average genotype call rates were 0.98 (range: 0.92-1.00), 1.00 (range: 0.99-1.00), and 0.99 (range: 0.95-1.00) for the HCC, HBV/HCV, and healthy groups, respectively. All case and control samples were included in the study, because they all reached the fixed call rate threshold of 90%.

HCC vs HBV/HCV group: Considering that chronic HBV and/or HCV infections represent a major risk factor for HCC development, a case/control analysis was initially performed to compare the genotype frequency distribution between patients with HCC and individuals

Table 2 Distribution of gene polymorphisms in patients with hepatocellular carcinoma, in hepatitis B/hepatitis C -positive patients, and in healthy controls *n* (%)

Gene	SNP	Genotype	HCC group	HBV/HCV group	Healthy group
<i>ERCC1</i>	rs3212986	GG	126 (67.7)	82 (49.1)	91 (47.4)
		GT	44 (23.7)	78 (46.7)	81 (42.2)
		TT	16 (8.6)	7 (4.2)	20 (10.4)
<i>XRCC3</i>	rs1799794	AA	128 (66.7)	108 (65.1)	137 (71.7)
		AG	52 (27.1)	55 (33.1)	49 (25.7)
		GG	12 (6.3)	3 (1.8)	5 (2.6)
<i>GST-P1</i>	rs1138272	CC	172 (92.5)	139 (83.2)	182 (94.8)
		CT	13 (7.0)	27 (16.2)	10 (5.2)
		TT	1 (0.5)	1 (0.6)	0 (0.0)
<i>CYP17A1</i>	rs743572	TT	72 (38.1)	39 (23.4)	54 (28.4)
		TC	84 (44.4)	88 (52.7)	91 (47.9)
		CC	33 (17.5)	40 (24.0)	45 (23.7)
<i>ABCB1</i>	rs1128503	CC	54 (28.1)	52 (31.1)	58 (30.2)
		CT	92 (47.9)	93 (55.7)	95 (49.5)
		TT	46 (24.0)	22 (13.2)	39 (20.3)

HCC: Hepatocellular carcinoma; SNP: Single-nucleotide polymorphism; HBV: Hepatitis B viral; HCV: Hepatitis C viral.

without HCC, but with matched HBV/HCV infections. The frequency distributions of variant genotypes were significantly different between cases and controls (Tables 2 and 3) for five polymorphisms. No Hardy-Weinberg disequilibrium among healthy controls was detected ($P > 0.05$), which suggested that it was a representative sampling of the investigated population. The observed genotype frequency distribution in healthy controls was consistent with data published in the literature for a Caucasian population (www.ncbi.nlm.nih.gov/snp).

Specifically, *ERCC1* rs3212986 (OR = 0.46, $P = 0.0005$), *GST-P1* rs1138272 (OR = 0.41, $P = 0.0097$), and *CYP17A1* rs743572 (OR = 0.50, $P = 0.0032$) markers were observed more frequently in the HBV/HCV group than in the HCC group, and they were significantly associated with reduced liver cancer risk, according to the dominant model. In contrast, the frequency of *XRCC3* rs1799794 (OR = 3.70; $P = 0.0461$) and *ABCB1* rs1128503 (OR = 2.06; $P = 0.0111$) markers were higher in the HCC group than in the HBV/HCV group, and they were associated with increased HCC risk, according to the recessive model (Table 3).

Given the molecular and clinicopathological differences between HBV- and HCV- related hepatocarcinogenesis^[18], a similar analysis was also carried out in subgroups, divided according to viral status (Table 4). In all cases, the ORs calculated for the HBV- and HCV-positive subgroups were concordant with those obtained for the entire combined group. The association was significant in both sub-groups for *CYP17A1* rs743572 (OR = 0.45, $P = 0.0315$ and OR = 0.51, $P = 0.0384$ in HBV-positive and HCV-positive subgroups, respectively). In contrast, *ABCB1* rs1128503 (OR = 4.18, $P = 0.0048$, HBV-positive subgroup) and *ERCC1* rs3212986 (OR = 0.33, $P = 0.0003$, HCV-positive

subgroup) showed significant associations in only one sub-group.

HCC vs healthy group: We also conducted a case case/control analysis of the genotype frequency distribution in patients with HCC compared to individuals without HCC or virus infections to evaluate the extent of the predictive value of the selected five markers (Table 3). Two markers were significantly associated with HCC risk. Specifically, *ERCC1* rs3212986 (OR = 0.43, $P < 0.0001$) and *CYP17A1* rs743572 (OR = 0.73, $P = 0.0310$) were associated with reduced HCC risk, according to the dominant and additive models, respectively.

Classification and regression tree analysis: The Classification and Regression Tree (CART) method allowed us to identify 6 subgroups of subjects according to genotype features, each with a different probability of HCC occurrence. Three of the terminal nodes exhibited high-probability (59%-76%), two exhibited intermediate-probability (36% and 40%), and one exhibited low-probability (6%) of developing HCC (Figure 1). The ORs and 95%CI were calculated for each high/intermediate-probability group, with respect to the reference low-probability group. The group with the highest percentage of HCC cases (group 5) was associated with an approximately 13-fold higher HCC risk compared to the reference low-probability group. The first node of the tree represented the *ERCC1* rs3212986 variant, which significantly interacted with *CYP17A1* rs743572, *GST-P1* rs1138272, and the *UGT1A7**3 marker in the generation of the tree (Figure 1).

DISCUSSION

HCC is a global healthcare problem. It is one of the most common malignancies worldwide, with an increasing incidence in Western developed countries. It represents the second cause of death attributable to cancer^[1,2]. The genetic origin of HCC has been a focus of research in the past few years. Some studies found that genetic variants could be predictive of the clinical course of HBV or HCV and an individual's predisposition to developing liver cancer^[4-8]. However, those studies were performed mainly in Asian populations; thus, few data are available on Caucasian populations. The ethnicity of the population being studied is a crucial factor in case-control studies, because polymorphism frequency varies greatly with geographical origin. This association could give rise to ethnicity-specific phenotypic effects^[4,14]. The present study revealed some novel genetic markers that could be used as early diagnosis indicators for liver cancer susceptibility in Caucasians. The availability of these markers could improve HCC risk stratification algorithms and surveillance programs.

Table 3 Odds ratios and corresponding 95%CI for hepatocellular carcinoma compared to hepatitis B/hepatitis C infected patients and blood donors according to genetic polymorphisms

Gene	SNP	Allelic change	HCC vs HBV/HCV			HCC vs Healthy		
			Mod	OR (95%CI) ¹	P value	Mod	OR (95%CI) ²	P value
<i>ERCC1</i>	rs3212986	G>T	Dom	0.46 (0.30-0.71)	0.0005 ^b	Dom	0.43 (0.28-0.65)	< 0.0001 ^b
<i>XRCC3</i>	rs1799794	A>G	Rec	3.70 (1.02-13.39)	0.0461 ^a	Rec	2.44 (0.84-7.08)	0.1003
<i>GST-P1</i>	rs1138272	C>T	Dom	0.41 (0.21-0.81)	0.0097 ^b	Add	1.58 (0.70-3.57)	0.2699
<i>CYP17A1</i>	rs743572	T>C	Dom	0.50 (0.31-0.79)	0.0032 ^b	Add	0.73 (0.55-0.97)	0.0310 ^a
<i>ABCB1</i>	rs1128503	C>T	Rec	2.06 (1.18-3.61)	0.0111 ^a	Dom	1.22 (0.75-1.98)	0.4194

¹Estimated through logistic regression model, adjusted for sex, age and HBV and/or HCV infection; ²Estimated through logistic regression model, adjusted for sex and age. ^aP < 0.05, ^bP < 0.01. HCC: Hepatocellular carcinoma; SNP: Single-nucleotide polymorphism; HBV: Hepatitis B viral; HCV: Hepatitis C viral.

Table 4 Odds ratios and corresponding 95%CI for hepatocellular carcinoma compared to hepatitis B/hepatitis C infected patients according to the viral status

Gene	SNP	Allelic change	Mod	HCC vs HBV/HCV			
				HBV-positive		HCV-positive	
				OR (95%CI) ¹	P value	OR (95%CI) ¹	P value
<i>ERCC1</i>	rs3212986	G>T	Dom	0.59 (0.30-1.15)	0.1231	0.33 (0.18-0.60)	0.0003 ^b
<i>XRCC3</i>	rs1799794	A>G	Rec	2.23 (0.39-12.82)	0.3695	6.73 (0.82-55.03)	0.0753
<i>GST-P1</i>	rs1138272	C>T	Dom	0.40 (0.14-1.15)	0.0881	0.41 (0.16-1.02)	0.0558
<i>CYP17A1</i>	rs743572	T>C	Dom	0.45 (0.22-0.93)	0.0315 ^a	0.51 (0.27-0.97)	0.0384 ^a
<i>ABCB1</i>	rs1128503	C>T	Rec	4.18 (1.55-11.29)	0.0048 ^b	1.49 (0.71-3.11)	0.2893

¹Estimated through logistic regression model, adjusted for sex, age and HBV and/or HCV infection. ^aP < 0.05, ^bP < 0.01. HCC: Hepatocellular carcinoma; SNP: Single-nucleotide polymorphism; HBV: Hepatitis B viral; HCV: Hepatitis C viral.

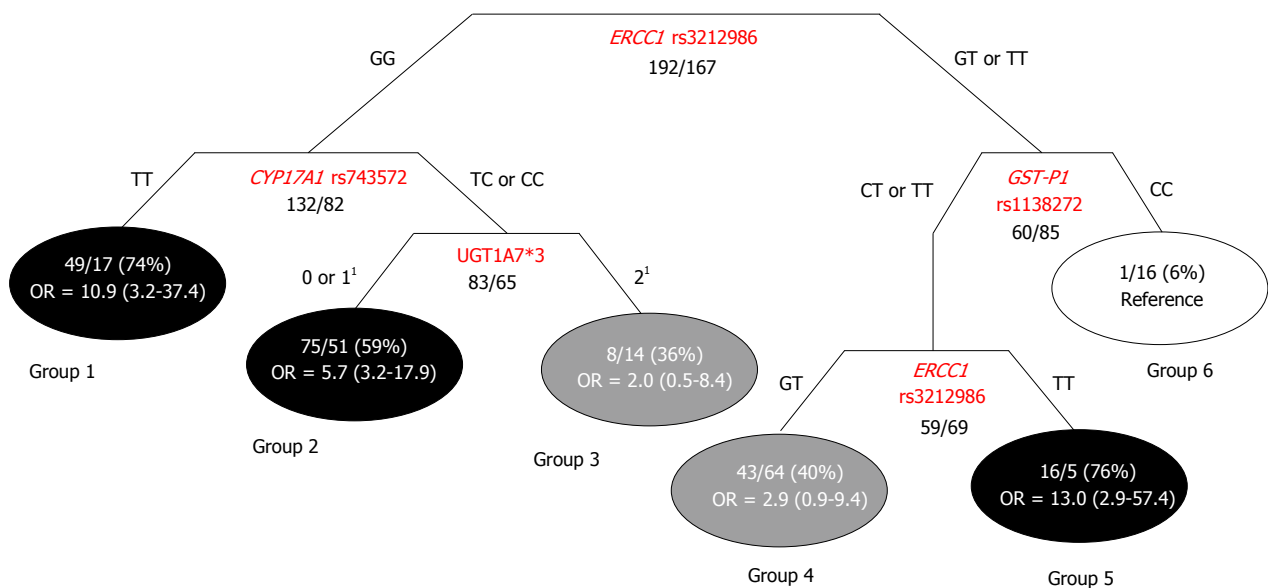


Figure 1 Classification and regression tree representation of the markers combination significantly predictive of hepatocellular carcinoma risk. Fractions indicate the number of HCC cases patients vs number of HBV/HCV infected patients (percentage of cases in parenthesis). Black circles represent terminal nodes with high probability to develop hepatocellular carcinoma (HCC) (ratio $\geq 70\%$); gray circles represent terminal nodes with intermediate probability to develop HCC ($30 \leq$ ratio $< 70\%$); white circles represent terminal nodes with low probability to develop HCC (ratio $< 30\%$). Odds ratios (OR) and 95%CI were calculated for each group in respect to the reference group (lower HCC risk) through logistic regression model, adjusted for sex, age and viral status. ¹Number of alleles carried by the patient. CART: Classification and regression tree; HCC: Hepatocellular carcinoma; HBV: Hepatitis B viral; HCV: Hepatitis C viral.

The main result of this work was the identification of five polymorphisms (*ERCC1* rs3212986, *GST-P1* rs1138272, *CYP17A1* rs743572, *XRCC3* rs1799794, and *ABCB1* rs1128503) that could identify sub-populations of patients among high-risk Caucasian individuals (HBV/

HCV-positive), which had differential predispositions for HCC development. The *ABCB1* rs1128503 and *ERCC1* rs3212986 variants displayed different effects, based on the viral status of the patient (HBV- or HCV-positive serology). This result was consistent with the

molecular and clinical-pathological differences between HBV- and HCV-related hepatocarcinogenesis^[18]. Two polymorphisms (*ERCC1* rs3212986, and *CYP17A1* rs743572) could also predict HCC risk among the healthy Caucasian population. A CART analysis provided evidence of gene-gene interactions between *ERCC1* rs3212986, *CYP17A1* rs743572, *GST-P1* rs1138272, and the previously described *UGT1A7**3 marker. These interactions defined subgroups of patients with genetic combinations that could further increase the risk of HCC.

The *ERCC1* rs3212986 (8092G>T) polymorphism was demonstrated to be a strong protective factor against liver cancer development in both high-risk (HCV/HBV-positive) and healthy individuals. Furthermore, in an analysis of subgroups with different viral statuses, a particularly significant effect was detected in the HCV-positive subgroup.

ERCC1 is one of several key rate-limiting enzymes in the nucleotide excision repair (NER) pathway. The NER mechanism of the DNA repair system maintains genomic integrity by removing inter-strand DNA crosslinks, and thus, it influences the cell's sensitivity to DNA-adducting carcinogens^[19]. A NER-related reduction in DNA repair capacity has been associated with some types of tumors (*i.e.*, lung, head and neck, prostate, glioma, ovarian cancers). Accordingly, *ERCC1* gene variants were shown to influence susceptibility to cancer^[20,21]. However, few studies have investigated the implication of the NER biological pathway and *ERCC1* polymorphisms, including rs3212986^[20,21], in hepatocarcinogenesis, and no data are available in Caucasian populations. The *ERCC1* rs3212986 variant, located in the 3' untranslated region (3'UTR) of the gene, is thought to increase transcript stability, although further biochemical studies are required to clarify the effect of this polymorphism on gene transcription and protein translation^[22,23]. Based on our data, we inferred that the rs3212986 variant allele was associated with elevated *ERCC1* expression and/or activity, which could lead to enhanced protection against the accumulation of mutagenic DNA lesions, and consequently, against hepatocarcinogenesis. However, we could not exclude that this marker might also be closely linked to other functionally relevant genetic variants^[21] that might alter *ERCC* activity and/or HCC onset.

The *XRCC3* rs1799794 (4541A>G) marker was associated with elevated HCC susceptibility in a high-risk (HCV/HBV-positive) population. *XRCC3* is another DNA repair protein that participates in the DNA double-strand break/recombination repair machinery. *XRCC3* was shown to be involved in the pathogenesis of many cancers, including HCC^[24,25]. Reduced *XRCC3* activity was associated with significantly elevated levels of bulky DNA adducts^[26]. Furthermore, epidemiological evidence, derived mainly from studies in an Asian population, have shown that *XRCC3* contributed to the repair of DNA damage induced by environmental toxins, such as aflatoxin B1, and that this effect

depended on viral status and ethnicity^[25,27]. The *XRCC3* rs1799794 variant, located in a regulatory region in the 5'UTR of the gene, was predicted to have a potential effect on protein expression levels^[28]. The results of the present study suggested that this polymorphism could reduce the ability to repair DNA adducts, which would contribute to the accumulation of mutagenic lesions, and thus, increase the risk of HCC onset. However, the exact molecular mechanism of the rs1799794 variant is currently unknown, and we could not exclude a linkage with another functional marker^[29].

The *GST-P1* rs1138272 (341C>T) polymorphism was found to have a protective effect on liver cancer development in a high-risk HCV/HBV-positive population. *GST-P1* belongs to the *GST* multigene family. It is a phase II enzyme involved in the inactivation of electrophilic xenobiotics by conjugation with glutathione^[30], which facilitates the excretion of molecules that are potentially toxic to the liver^[31]. This enzyme also contributes to protecting liver cells from damage related to reactive oxygen species generated, for instance, by chronic alcohol and tobacco abuse^[32]. Several studies have clearly demonstrated that silencing *GST-P1* expression by promoter methylation was positively correlated with the incidence of HCC^[31] and with virus-related hepatocarcinogenesis^[33]. In contrast, in meta-analyses, *GST-P1* genetic polymorphisms (*i.e.*, Ile105Val, rs1695) were not associated with HCC risk^[34,35] although the few data available were insufficient for drawing any definite conclusions. The less investigated *GST-P1* rs1138272 polymorphism is a missense variant (Val114Ala) that lies in the electron binding, active domain of the protein. This variant was reported to affect substrate specificity by altering its ability to distinguish between planar and non-planar substrates^[36,37]. Our results suggested that this functional change in *GST-P1* activity may result in a lower risk of HCC development; further clinical experimental studies are required to clarify the role of the rs1138272 polymorphism in the response of *GST-P1* to environmental xenobiotics and oxidative stress, particularly in a virus-infected liver.

The *CYP17A1* rs743572 (-34T>C) polymorphism was found to be a strong protective factor against HCC development in both the high-risk HCV/HBV-positive group and healthy individuals. *CYP17A1* is an enzyme that plays a critical role in the biosynthesis of steroid hormones by catalyzing both 17 α -hydroxylase and 17, 20-lyase activities^[38]. Some data derived from animal models and human epidemiologic studies have demonstrated that estrogens play an essential role in hepatocarcinogenesis; however, the exact mechanism underlying the effect of estrogen levels on HCC development is currently controversial^[39]. It was initially suggested that the *CYP17A1* rs743572 variant, located in the 5'UTR of the gene, served as a new Sp1-type (CCACC box) binding site, which provided an additional promoter, and thus, it could increase estrogen biosynthesis^[40]. However, subsequent studies could not replicate the phenotypic impact of this

variant on CYP17A1 mRNA expression or estrogen levels^[41]. Moreover, the involvement of the rs743572 marker on HCC development was not well defined^[42,43]. From the present data, it could be speculated that the rs743572 variant might contribute to changing CYP17A1 activity, and its dependent steroid levels, to provide a protective effect against liver cancer development. Estrogen is known to participate in various biological functions. For example, an increase in sex hormone levels was suggested to suppress the inflammatory processes mediated by pro-inflammatory cytokines^[44,45]. Additional work will be required to clarify the functional significance of the rs743572 marker and the molecular mechanism of CYP17A1 in hepatocellular carcinogenesis.

Finally, *ABCB1* rs1128503 (1236C>T) was associated with elevated HCC susceptibility in a high-risk (HCV/HBV-positive) population. Furthermore, an analysis of subgroups with different viral statuses showed that a particularly significant effect was detected in the HBV-positive subgroup. The *ABCB1* gene encodes MDR-1, also known as P-glycoprotein (P-gp). MDR-1 belongs to the ATP-binding cassette (ABC) transporter superfamily^[46,47]. This membrane efflux pump is widely expressed in various organs, including the liver, and it is physiologically involved in the protection of normal tissue from environmental and endogenous toxins^[46-48]. MDR1 is implicated in hepatic clearance of oxidative and other genotoxic products generated by chronic inflammation induced by HCC risk factors. Thus, it contributes to preventing the initiation of hepatocarcinogenesis^[49,50]. Many studies have described MDR1 over-expression in various tumor tissues, including liver cancer^[51]; thus, this membrane carrier and its functional genetic variants are good candidates for influencing liver susceptibility to HCC. *ABCB1* rs1128503 is a synonymous variant (Gly412Gly) of a residue located in the cytoplasmic domain of the transporter. It is reported to affect mRNA stability^[52], but the exact phenotypic impact of this variant on protein expression/activity remains to be defined^[53,54]. Several studies, performed in Asian populations, have investigated associations between *ABCB1* polymorphisms (including rs1128503) and the risk of developing HCC. A recent meta-analysis^[55] showed that polymorphic alleles of the *ABCB1* gene significantly increased HCC risk. In analyses of subgroups with different types of variations, both synonymous and non-synonymous polymorphisms were associated with elevated liver cancer susceptibility. In particular, synonymous variants (*i.e.*, rs1128503) were suggested to be correlated with changes in transcriptional and translational processes. In addition, genetic markers located in the cytoplasmic domain of the transporter, like rs1128503, were shown to have the greatest predictive power for HCC risk. These data, although obtained in Asian cohorts, were consistent with the result of the present study, which provided the first evidence of a role of the *ABCB1* rs1128503 marker in

HCC risk in a Caucasian population.

Due to the multifactorial nature of hepatocarcinogenesis, an exploratory CART analysis was applied to investigate interactions between the genetic factors identified as predictors of liver cancer susceptibility in the high-risk population (HBV/HCV-infected group). The polymorphism in the gene of the DNA repair system (*i.e.*, *ERCC1* rs3212986) was demonstrated to significantly interact with variants of genes that encoded phase I (*CYP17A1* rs74357) and II (*GST-P1* rs1138272, *UGT1A7*3*) metabolic enzymes; these interactions were important in defining individual HCC risk. The CART analysis generated a distinctive clustering of subjects, according to different probabilities of developing liver cancer. These data pointed out that previously acknowledged predictive markers, such as the *UGT1A*3* polymorphism^[14], should be considered in association with novel identified genetic variants for properly defining the complex trait of HCC susceptibility. In particular, *ERCC1* rs3212986 was shown to be the most relevant polymorphism for HCC risk stratification, because it appeared twice in the generated tree. This marker significantly interacted with three additional genetic variants (*CYP17A1* rs74357, *GST-P1* rs1138272, and *UGT1A7*3*) in defining individual susceptibility. The analysis of gene-gene interactions and the network shape uncovered specific genetic-context effects that could not be detected in evaluations of single markers. These findings highlighted the importance of combining genetic markers to obtain a better representation of the biological cooperation among multiple pathways in hepatocarcinogenesis.

The present study had some limitations. First, our case-cohort comprised a highly specific group of patients with HCC (patients that received OLT). These patients had definite clinical features that could limit the generalizability of the present findings. The validity of the present results was also limited to Caucasian individuals; the results remain to be verified in other populations, like Asians or Africans. Second, our work did not evaluate other confounding etiological factors, such as smoking and alcohol intake. However, the use of a control group matched to HCC cases for HBV/HCV viral status permitted us to control for this crucial confounding variable. Third, the exact functional and phenotypic properties of some of the genetic markers that we identified as predictors of HCC risk have not been well established. Moreover, we could not exclude any linkages with other functionally relevant genetic variants. Finally, considering the retrospective and exploratory nature of our case-control analysis, the results obtained should be interpreted with caution. Our findings will require independent validation in future prospective, large-scale epidemiological investigations that are performed in well-defined, homogeneous populations.

The validation of the present findings could have important clinical implications for improving HCC

management. The novel identified genetic markers and their gene-gene interactions, in combination with other well-known serum biomarkers (e.g., alphafetoprotein) and diagnostic imaging (e.g., ultrasonography)^[9], could contribute to improving the HCC risk stratification algorithms and surveillance programs, particularly for patients at risk. Those improvements might facilitate early cancer detection and might lead to potentially curative therapies. Furthermore, an enhanced understanding of the genetic factors associated with the evolution of HCV infection could be essential for the optimized use of the recently developed, but highly expensive, anti-HCV DAAs^[12,13]. Thus, improved screening could assist the health care system in containing costs and ensuring the most clinically appropriate treatments.

The present study identified five variants in genes that encode DNA repair system proteins (*i.e.*, ERCC1, XRCC3), phase I (*i.e.*, CYP17A1) and II (*i.e.*, GST-P1) enzymes, and a membrane transporter (*i.e.*, MDR-1). Our data suggested that these genes might play a role in predicting HCC susceptibility in a high-risk (HBV-HCV infected) population and in a healthy Caucasian population. The CART analysis pointed out the potential value of studying of gene-gene interactions for the stratification of individuals at different risks of developing HCC. Although validation is required, these preliminary results might have important clinical implications in the management of HCC prevention and patient care.

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COMMENTS

Background

Hepatocellular carcinoma (HCC) represents a global health problem, due to its high incidence and mortality rates. Although the etiologies of this neoplasia are well-defined, the majority of patients are diagnosed at advanced-stage disease, which excludes them from receiving a potentially curative therapy. Hence, they urgently need the definition of new biomarkers of HCC susceptibility that can permit early cancer detection, particularly for individuals in a high-risk population [hepatitis B (HBV)/hepatitis C (HCV)-positive]. Moreover, in developed countries (*i.e.*, United States and Europe), HCV-related HCC can account for most liver cancers; therefore, increasing our knowledge of the genetic contribution to the clinical course of viral infections could be essential in optimizing the use of the newly developed, but highly expensive, direct acting antiviral treatments.

Research frontiers

Accumulating data derived from candidate-gene investigations and genome-wide association studies have highlighted the possible role of genetic variants as significant predictors of HCC susceptibility and as determinants of the clinical course of HBV or HCV infections. However, the high heterogeneity among published works has prevented drawing definite conclusions. Furthermore, the majority of case-control investigations were performed in Asian populations; thus, replication studies with different ethnicities are warranted. Consequently, there is a critical need to intensify research efforts in identifying effective molecular biomarkers predictive of individual HCC risk in a homogeneous, well-defined cohort study, with particular attention to the (currently) poorly

investigated Caucasian population.

Innovations and breakthroughs

This work uncovered five markers in genes that encode key proteins involved in pathways underlying hepatocarcinogenesis (*ERCC1*-rs3212986, *GSTP1*-rs1138272, *CYP17A1*-rs743572, *XRCC3*-rs1799794, and *ABCB1*-rs112850). These markers were predictive of individual HCC susceptibility in a high-risk Caucasian population (HBV/HCV-positive). Two variants also exhibited serology-specific effects when a subgroup analysis was performed in patients stratified according to HBV or HCV positive serology. That analysis showed that *ABCB1* rs1128503 specifically affected the HBV-positive subgroup and *ERCC1* rs3212986 specifically affected the HCV-positive subgroup. Among the five markers identified, *ERCC1* rs3212986 and *CYP17A1* rs743572 retained predictive value for HCC risk among healthy individuals. In addition, they found gene-gene interactions between *ERCC1* rs3212986, *CYP17A1* rs743572, *GST-P1* rs1138272, and the previously described *UGT1A7**3 marker, which were included in the definition of the complex trait of HCC susceptibility.

Applications

The definition of genetic traits that confer susceptibility to liver carcinogenesis is of potential clinical interest. This information can facilitate improvements in preventive, diagnostic, and therapeutic strategies for HCC management. If validated, these markers and their interactions should be considered for improving HCC risk stratification algorithms and surveillance programs. Their inclusion in screening could facilitate early cancer detection and might lead to potentially curative therapies. For patients with HCV infections, the biomarkers we discovered could also contribute to optimizing the use of recently developed, but highly expensive, anti-viral therapies. These findings address a great challenge in HCC management and might help enhance care for patients with HCC.

Terminology

Polymorphism: a variation in the DNA sequence that occurs in a population at a frequency of 1% or higher; CART: classification and regression tree analysis; this is a statistical method, based on recursive partitioning, that is applied to elucidate potential high-order gene-gene and gene-environmental relationships among discriminating factors that contribute to a specific endpoint; ERCC1: a key rate-limiting enzyme that acts in the nucleotide excision repair pathway. This pathway represents a DNA repair mechanism designed to maintain genomic integrity by removing inter-strand DNA crosslinks; XRCC3: a DNA repair protein that participates in DNA double-strand break/recombination repair machinery; GST-P1: a phase II enzyme involved in the inactivation of electrophilic xenobiotics; it acts by conjugating with glutathione, which facilitates excretion of xenobiotics from the body; CYP17A1: an enzyme that plays a critical role in the biosynthesis of steroid hormones; MDR-1 (encoded by *ABCB1*): A widely expressed membrane efflux pump, physiologically involved in the protection of normal tissue from environmental and endogenous toxins.

Peer-review

The aim of this study was to find novel genetic markers that could contribute in predicting HCC susceptibility in Caucasians. The authors identified five genetic markers in key pathways linked to hepatocarcinogenesis (*ERCC1*-rs3212986, *GSTP1*-rs1138272, *CYP17A1*-rs743572, *XRCC3*-rs1799794, *ABCB1*-rs112850) that could predict the individual HCC susceptibility, especially in high-risk (HBV/HCV-infected) Caucasians population. The article provides the novel informations that could contribute to improving HCC surveillance, allowing early cancer diagnosis and possible curative therapy.

REFERENCES

- 1 Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; **65**: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 2 McGlynn KA, Petrick JL, London WT. Global epidemiology of hepatocellular carcinoma: an emphasis on demographic and regional variability. *Clin Liver Dis* 2015; **19**: 223-238 [PMID: 25921660 DOI: 10.1016/j.cld.2015.01.001]
- 3 Galun D, Basaric D, Zuvella M, Bulajic P, Bogdanovic A, Bidzic

- N, Milicevic M. Hepatocellular carcinoma: From clinical practice to evidence-based treatment protocols. *World J Hepatol* 2015; **7**: 2274-2291 [PMID: 26380652 DOI: 10.4254/wjh.v7.i20.2274]
- 4 **Jin F**, Xiong WJ, Jing JC, Feng Z, Qu LS, Shen XZ. Evaluation of the association studies of single nucleotide polymorphisms and hepatocellular carcinoma: a systematic review. *J Cancer Res Clin Oncol* 2011; **137**: 1095-1104 [PMID: 21240526 DOI: 10.1007/s00432-010-0970-0]
 - 5 **Miki D**, Ochi H, Hayes CN, Aikata H, Chayama K. Hepatocellular carcinoma: towards personalized medicine. *Cancer Sci* 2012; **103**: 846-850 [PMID: 22339805 DOI: 10.1111/j.1349-7006.2012.02242.x]
 - 6 **Nahon P**, Zucman-Rossi J. Single nucleotide polymorphisms and risk of hepatocellular carcinoma in cirrhosis. *J Hepatol* 2012; **57**: 663-674 [PMID: 22609306 DOI: 10.1016/j.jhep.2012.02.035]
 - 7 **Matsuura K**, Isogawa M, Tanaka Y. Host genetic variants influencing the clinical course of hepatitis B virus infection. *J Med Virol* 2016; **88**: 371-379 [PMID: 26255971 DOI: 10.1002/jmv.24350]
 - 8 **Matsuura K**, Tanaka Y. Host genetic variants influencing the clinical course of hepatitis C virus infection. *J Med Virol* 2016; **88**: 185-195 [PMID: 26211651 DOI: 10.1002/jmv.24334]
 - 9 **Marquardt JU**, Nguyen-Tat M, Galle PR, Wörns MA. Surveillance of Hepatocellular Carcinoma and Diagnostic Algorithms in Patients with Liver Cirrhosis. *Visc Med* 2016; **32**: 110-115 [PMID: 27413728 DOI: 10.1159/000445407]
 - 10 **Mazzanti R**, Arena U, Tassi R. Hepatocellular carcinoma: Where are we? *World J Exp Med* 2016; **6**: 21-36 [PMID: 26929917 DOI: 10.5493/wjem.v6.i1.21]
 - 11 **Wang CH**, Wey KC, Mo LR, Chang KK, Lin RC, Kuo JJ. Current trends and recent advances in diagnosis, therapy, and prevention of hepatocellular carcinoma. *Asian Pac J Cancer Prev* 2015; **16**: 3595-3604 [PMID: 25987009]
 - 12 **Luhnen M**, Waffenschmidt S, Gerber-Grote A, Hanke G. Health Economic Evaluations of Sofosbuvir for Treatment of Chronic Hepatitis C: a Systematic Review. *Appl Health Econ Health Policy* 2016; **14**: 527-543 [PMID: 27329481 DOI: 10.1007/s40258-016-0253-2]
 - 13 **Roderburg C**, Tacke F, Trautwein C. Antiviral Therapy in Patients with Viral Hepatitis and Hepatocellular Carcinoma: Indications and Prognosis. *Visc Med* 2016; **32**: 121-126 [PMID: 27413730 DOI: 10.1159/000444990]
 - 14 **De Mattia E**, Cecchin E, Polesel J, Lupo F, Tiribelli C, Crovatto M, Buonadonna A, Toffoli G. UGT1A polymorphisms as genetic biomarkers for hepatocellular carcinoma risk in Caucasian population. *Liver Int* 2017; **37**: 1345-1353 [PMID: 28294511 DOI: 10.1111/liv.13411]
 - 15 **Cecchin E**, Russo A, Corona G, Campagnutta E, Martella L, Boiocchi M, Toffoli G. UGT1A1*28 polymorphism in ovarian cancer patients. *Oncol Rep* 2004; **12**: 457-462 [PMID: 15254716]
 - 16 **Cecchin E**, Russo A, Campagnutta E, Martella L, Toffoli G. Lack of association of CYP1 B1*3 polymorphism and ovarian cancer in a Caucasian population. *Int J Biol Markers* 2004; **19**: 160-163 [PMID: 15255550]
 - 17 **Toffoli G**, Rossi D, Gaidano G, Cecchin E, Boiocchi M, Carbone A. Methylentetrahydrofolate reductase genotype in diffuse large B-cell lymphomas with and without hypermethylation of the DNA repair gene O6-methylguanine DNA methyltransferase. *Int J Biol Markers* 2003; **18**: 218-221 [PMID: 14535593]
 - 18 **Sukowati CH**, El-Khobar KE, Ie SI, Anfusio B, Muljono DH, Tiribelli C. Significance of hepatitis virus infection in the oncogenic initiation of hepatocellular carcinoma. *World J Gastroenterol* 2016; **22**: 1497-1512 [PMID: 26819517 DOI: 10.3748/wjg.v22.i4.1497]
 - 19 **Li C**, Wang LE, Wei Q. DNA repair phenotype and cancer susceptibility--a mini review. *Int J Cancer* 2009; **124**: 999-1007 [PMID: 19065660 DOI: 10.1002/ijc.24126]
 - 20 **Li Y**, Ou C, Shu H, Zhao H, Zhu B. The ERCC1-4533/8092, TNF- α 238/308 polymorphisms and the risk of hepatocellular carcinoma in Guangxi Zhuang populations of China: Case-control study. *Medicine (Baltimore)* 2016; **95**: e5217 [PMID: 27858866 DOI: 10.1097/MD.00000000000005217]
 - 21 **Wang B**, Xu Q, Yang HW, Sun LP, Yuan Y. The association of six polymorphisms of five genes involved in three steps of nucleotide excision repair pathways with hepatocellular cancer risk. *Oncotarget* 2016; **7**: 20357-20367 [PMID: 26967386 DOI: 10.18632/oncotarget.7952]
 - 22 **Rulli E**, Marabese M, Piva S, Bonomi L, Caiola E, Ganzinelli M. DNA repair gene polymorphisms in non-small-cell lung cancer patients treated with first-line platinum-containing chemotherapy. *Tumori* 2016; **102**: 367-375 [PMID: 27396427 DOI: 10.5301/tj.5000526]
 - 23 **Zhu J**, Hua RX, Jiang J, Zhao LQ, Sun X, Luan J, Lang Y, Sun Y, Shang K, Peng S, Ma J. Association studies of ERCC1 polymorphisms with lung cancer susceptibility: a systematic review and meta-analysis. *PLoS One* 2014; **9**: e97616 [PMID: 24841208 DOI: 10.1371/journal.pone.0097616]
 - 24 **Ji RB**, Qian YS, Hu AR, Hu YR. DNA repair gene XRCC3 T241M polymorphism and susceptibility to hepatocellular carcinoma in a Chinese population: a meta-analysis. *Genet Mol Res* 2015; **14**: 15988-15996 [PMID: 26662391 DOI: 10.4238/2015.December.7.11]
 - 25 **Zeng X**, Liu S, Yu H, Ji L, Li L, Huang J, Bai H, Qiu X. DNA repair capacity, DNA-strand break repair gene polymorphisms, and the incidence of hepatocellular carcinoma in southwestern Guangxi of China. *DNA Cell Biol* 2012; **31**: 1384-1391 [PMID: 22691054 DOI: 10.1089/dna.2012.1646]
 - 26 **Matullo G**, Palli D, Peluso M, Guarrera S, Carturan S, Celentano E, Krogh V, Munnia A, Tumino R, Polidoro S, Piazza A, Vineis P. XRCC1, XRCC3, XPD gene polymorphisms, smoking and (32)P-DNA adducts in a sample of healthy subjects. *Carcinogenesis* 2001; **22**: 1437-1445 [PMID: 11532866]
 - 27 **Yao JG**, Huang XY, Long XD. Interaction of DNA repair gene polymorphisms and aflatoxin B1 in the risk of hepatocellular carcinoma. *Int J Clin Exp Pathol* 2014; **7**: 6231-6244 [PMID: 25337275]
 - 28 **Fachal L**, Gómez-Caamaño A, Peleteiro P, Carballo A, Calvo-Crespo P, Sánchez-García M, Lobato-Busto R, Carracedo A, Vega A. Association of a XRCC3 polymorphism and rectum mean dose with the risk of acute radio-induced gastrointestinal toxicity in prostate cancer patients. *Radiother Oncol* 2012; **105**: 321-328 [PMID: 23075580 DOI: 10.1016/j.radonc.2012.09.013]
 - 29 **Smolkova B**, Dusinska M, Hemminki K. NBN and XRCC3 genetic variants in childhood acute lymphoblastic leukaemia. *Cancer Epidemiol* 2014; **38**: 563-568 [PMID: 25176580 DOI: 10.1016/j.canep.2014.08.002]
 - 30 **Watson MA**, Stewart RK, Smith GB, Massey TE, Bell DA. Human glutathione S-transferase P1 polymorphisms: relationship to lung tissue enzyme activity and population frequency distribution. *Carcinogenesis* 1998; **19**: 275-280 [PMID: 9498276]
 - 31 **Tian M**, Zhao B, Zhang J, Martin FL, Huang Q, Liu L, Shen H. Association of environmental benzo[a]pyrene exposure and DNA methylation alterations in hepatocellular carcinoma: A Chinese case-control study. *Sci Total Environ* 2016; **541**: 1243-1252 [PMID: 26476064 DOI: 10.1016/j.scitotenv.2015.10.003]
 - 32 **Mansoori AA**, Jain SK. Molecular Links between Alcohol and Tobacco Induced DNA Damage, Gene Polymorphisms and Pathophysiological Consequences: A Systematic Review of Hepatic Carcinogenesis. *Asian Pac J Cancer Prev* 2015; **16**: 4803-4812 [PMID: 26163595]
 - 33 **Kiran M**, Chawla YK, Kaur J. Methylation profiling of tumor suppressor genes and oncogenes in hepatitis virus-related hepatocellular carcinoma in northern India. *Cancer Genet Cytogenet* 2009; **195**: 112-119 [PMID: 19963110 DOI: 10.1016/j.cancergencyto.2009.06.021]
 - 34 **Chen J**, Ma L, Peng NF, Wang SJ, Li LQ. A meta-analysis of the relationship between glutathione S-transferases gene polymorphism and hepatocellular carcinoma in Asian population. *Mol Biol Rep* 2012; **39**: 10383-10393 [PMID: 23053942 DOI: 10.1007/s11033-012-1917-0]
 - 35 **White DL**, Li D, Nurgalieva Z, El-Serag HB. Genetic variants of glutathione S-transferase as possible risk factors for hepatocellular

- carcinoma: a HuGE systematic review and meta-analysis. *Am J Epidemiol* 2008; **167**: 377-389 [PMID: 18065725 DOI: 10.1093/aje/kwm315]
- 36 **Ali-Osman F**, Akande O, Antoun G, Mao JX, Buolamwini J. Molecular cloning, characterization, and expression in *Escherichia coli* of full-length cDNAs of three human glutathione S-transferase Pi gene variants. Evidence for differential catalytic activity of the encoded proteins. *J Biol Chem* 1997; **272**: 10004-10012 [PMID: 9092542]
 - 37 **Ji X**, Blaszczyk J, Xiao B, O'Donnell R, Hu X, Herzog C, Singh SV, Zimniak P. Structure and function of residue 104 and water molecules in the xenobiotic substrate-binding site in human glutathione S-transferase P1-1. *Biochemistry* 1999; **38**: 10231-10238 [PMID: 10441116 DOI: 10.1021/bi990668u]
 - 38 **Goldstone JV**, Sundaramoorthy M, Zhao B, Waterman MR, Stegeman JJ, Lamb DC. Genetic and structural analyses of cytochrome P450 hydroxylases in sex hormone biosynthesis: Sequential origin and subsequent coevolution. *Mol Phylogenet Evol* 2016; **94**: 676-687 [PMID: 26432395 DOI: 10.1016/j.ympev.2015.09.012]
 - 39 **Baldissera VD**, Alves AF, Almeida S, Porowski M, Giovenardi M. Hepatocellular carcinoma and estrogen receptors: Polymorphisms and isoforms relations and implications. *Med Hypotheses* 2016; **86**: 67-70 [PMID: 26804600 DOI: 10.1016/j.mehy.2015.11.030]
 - 40 **Carey AH**, Waterworth D, Patel K, White D, Little J, Novelli P, Franks S, Williamson R. Polycystic ovaries and premature male pattern baldness are associated with one allele of the steroid metabolism gene CYP17. *Hum Mol Genet* 1994; **3**: 1873-1876 [PMID: 7849715]
 - 41 **Xu J**, Lin X, Zhu H, Zhang Z, Yang B. Genetic variation of the CYP17 and susceptibility to endometrial cancer: a meta-analysis. *Mol Biol Rep* 2013; **40**: 5085-5091 [PMID: 23649771 DOI: 10.1007/s11033-013-2609-0]
 - 42 **Rossi L**, Leverì M, Gritti C, De Silvestri A, Zavaglia C, Sonzogni L, Silvestri L, Civardi E, Mondelli MU, Silini EM. Genetic polymorphisms of steroid hormone metabolizing enzymes and risk of liver cancer in hepatitis C-infected patients. *J Hepatol* 2003; **39**: 564-570 [PMID: 12971967]
 - 43 **Yuan X**, Zhou G, Zhai Y, Xie W, Cui Y, Cao J, Zhi L, Zhang H, Yang H, Zhang X, Qiu W, Peng Y, Zhang X, Yu L, Xia X, He F. Lack of association between the functional polymorphisms in the estrogen-metabolizing genes and risk for hepatocellular carcinoma. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 3621-3627 [PMID: 19064581 DOI: 10.1158/1055-9965.EPI-08-0742]
 - 44 **Hartwell HJ**, Petrosky KY, Fox JG, Horseman ND, Rogers AB. Prolactin prevents hepatocellular carcinoma by restricting innate immune activation of c-Myc in mice. *Proc Natl Acad Sci USA* 2014; **111**: 11455-11460 [PMID: 25049387 DOI: 10.1073/pnas.1404267111]
 - 45 **Montella M**, D'Arena G, Crispo A, Capunzo M, Nocerino F, Grimaldi M, Barbieri A, D'Ursi AM, Tecce MF, Amore A, Galdiero M, Ciliberto G, Giudice A. Role of Sex Hormones in the Development and Progression of Hepatitis B Virus-Associated Hepatocellular Carcinoma. *Int J Endocrinol* 2015; **2015**: 854530 [PMID: 26491442 DOI: 10.1155/2015/854530]
 - 46 **Choudhuri S**, Klaassen CD. Structure, function, expression, genomic organization, and single nucleotide polymorphisms of human ABCB1 (MDR1), ABCC (MRP), and ABCG2 (BCRP) efflux transporters. *Int J Toxicol* 2006; **25**: 231-259 [PMID: 16815813 DOI: 10.1080/10915810600746023]
 - 47 **Zhou SF**. Structure, function and regulation of P-glycoprotein and its clinical relevance in drug disposition. *Xenobiotica* 2008; **38**: 802-832 [PMID: 18668431 DOI: 10.1080/00498250701867889]
 - 48 **Leslie EM**, Deeley RG, Cole SP. Multidrug resistance proteins: role of P-glycoprotein, MRP1, MRP2, and BCRP (ABCG2) in tissue defense. *Toxicol Appl Pharmacol* 2005; **204**: 216-237 [PMID: 15845415 DOI: 10.1016/j.taap.2004.10.012]
 - 49 **Aleksandrova K**, Boeing H, Nöthlings U, Jenab M, Fedirko V, Kaaks R, Lukanova A, Trichopoulou A, Trichopoulos D, Boffetta P, Trepo E, Westphal S, Duarte-Salles T, Stepien M, Overvad K, Tjønneland A, Halkjaer J, Boutron-Ruault MC, Dossus L, Racine A, Lagiou P, Bamia C, Benetou V, Agnoli C, Palli D, Panico S, Tumino R, Vineis P, Bueno-de-Mesquita B, Peeters PH, Gram IT, Lund E, Weiderpass E, Quirós JR, Agudo A, Sánchez MJ, Gavrila D, Barricarte A, Dorronsoro M, Ohlsson B, Lindkvist B, Johansson A, Sund M, Khaw KT, Wareham N, Travis RC, Riboli E, Pischon T. Inflammatory and metabolic biomarkers and risk of liver and biliary tract cancer. *Hepatology* 2014; **60**: 858-871 [PMID: 24443059 DOI: 10.1002/hep.27016]
 - 50 **Sun B**, Karin M. Obesity, inflammation, and liver cancer. *J Hepatol* 2012; **56**: 704-713 [PMID: 22120206 DOI: 10.1016/j.jhep.2011.09.020]
 - 51 **Baldissera VD**, de Mattos AA, Coral GP, de Araujo FB, Marroni CA, de Mello Brandão AB, Ott Fontes PR, Schmidt Cerski CT, Hartmann AA, Kretzmann Filho NA. Evaluation of the C3435T polymorphism in the MDR1 gene in patients with hepatocellular carcinoma. *Ann Hepatol* 2012; **11**: 899-906 [PMID: 23109454 DOI: 10.1016/j.drudis.2007.12.010]
 - 52 **Frittitta L**, Ercolino T, Bozzali M, Argiolas A, Graci S, Santagati MG, Spampinato D, Di Paola R, Cisternino C, Tassi V, Vigneri R, Pizzuti A, Trischitta V. A cluster of three single nucleotide polymorphisms in the 3'-untranslated region of human glycoprotein PC-1 gene stabilizes PC-1 mRNA and is associated with increased PC-1 protein content and insulin resistance-related abnormalities. *Diabetes* 2001; **50**: 1952-1955 [PMID: 11473061]
 - 53 **Fung KL**, Gottesman MM. A synonymous polymorphism in a common MDR1 (ABCB1) haplotype shapes protein function. *Biochim Biophys Acta* 2009; **1794**: 860-871 [PMID: 19285158 DOI: 10.1016/j.bbapap.2009.02.014]
 - 54 **Haufroid V**. Genetic polymorphisms of ATP-binding cassette transporters ABCB1 and ABCC2 and their impact on drug disposition. *Curr Drug Targets* 2011; **12**: 631-646 [PMID: 21039333]
 - 55 **Wang ZC**, Liu LZ, Liu XY, Hu JJ, Wu YN, Shi JY, Yang LX, Duan M, Wang XY, Zhou J, Fan J, Gao Q. Genetic polymorphisms of the multidrug resistance 1 gene MDR1 and the risk of hepatocellular carcinoma. *Tumour Biol* 2015; **36**: 7007-7015 [PMID: 25861753 DOI: 10.1007/s13277-015-3407-1]

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Retrospective Cohort Study

Prognostic value of lymphovascular invasion in Bismuth-Corlette type IV hilar cholangiocarcinoma

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Abstract

AIM

To assess the prognostic value of lymphovascular invasion (LVI) in Bismuth-Corlette type IV hilar cholangiocarcinoma (HC) patients.

METHODS

A retrospective analysis was performed on 142 consecutively recruited type IV HC patients undergoing radical resection with at least 5 years of follow-up. Survival analysis was performed by the Kaplan-Meier method, and the association between the clinicopathologic variables and survival was evaluated by log-rank test. Multivariate analysis was adopted to identify the independent prognostic factors for overall survival (OS) and disease-free survival (DFS). Multiple logistic regression analysis was performed to determine the association between LVI and potential variables.

RESULTS

LVI was confirmed histopathologically in 29 (20.4%) patients. Multivariate analysis showed that positive resection margin (HR = 6.255, 95%CI: 3.485-11.229, $P < 0.001$), N1 stage (HR = 2.902, 95%CI: 1.132-7.439, $P = 0.027$), tumor size > 30 mm (HR = 1.942, 95%CI: 1.176-3.209, $P = 0.010$) and LVI positivity (HR = 2.799, 95%CI: 1.588-4.935, $P < 0.001$) were adverse

prognostic factors for DFS. The independent risk factors for OS were positive resection margin (HR = 6.776, 95%CI: 3.988-11.479, $P < 0.001$), N1 stage (HR = 2.827, 95%CI: 1.243-6.429, $P = 0.013$), tumor size > 30 mm (HR = 1.739, 95%CI: 1.101-2.745, $P = 0.018$) and LVI positivity (HR = 2.908, 95%CI: 1.712-4.938, $P < 0.001$). LVI was associated with N1 stage and tumor size > 30 mm. Multiple logistic regression analysis indicated that N1 stage (HR = 3.312, 95%CI: 1.338-8.198, $P = 0.026$) and tumor size > 30 mm (HR = 3.258, 95%CI: 1.288-8.236, $P = 0.013$) were associated with LVI.

CONCLUSION

LVI is associated with N1 stage and tumor size > 30 mm and adversely influences DFS and OS in type IV HC patients.

Key words: Bismuth-Corlette classification; Disease-free survival; Lymphovascular invasion; Overall survival; Hilar cholangiocarcinoma

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Core tip: Previous studies have reported that lymphovascular invasion (LVI) provokes an adverse impact on the long-term survival of several malignancies, including breast, gastric, and esophageal carcinoma, among many others. However, the correlation between LVI and hilar cholangiocarcinoma remains unclear. In our study, LVI was found to be an independent risk factor for overall survival and disease-free survival in Bismuth-Corlette type IV hilar cholangiocarcinoma patients. To our knowledge, this report indicates for the first time that LVI is an adverse predictor of long-term survival in the setting of type IV hilar cholangiocarcinoma.

Li B, Xiong XZ, Zhou Y, Wu SJ, You Z, Lu J, Cheng NS. Prognostic value of lymphovascular invasion in Bismuth-Corlette type IV hilar cholangiocarcinoma. *World J Gastroenterol* 2017; 23(36): 6685-6693 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6685.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6685>

INTRODUCTION

Hilar cholangiocarcinoma (HC), also known as Klatskin tumor, is a neoplasia arising from the biliary epithelium at the common hepatic duct bifurcation and may extend to the intrahepatic biliary tree and liver^[1,2]. The only clinical approach that is considered to provide patients with an opportunity for a curative outcome and importantly long-term survival is radical surgical resection^[2-6]. The Bismuth-Corlette classification is the most widely used preoperative system of evaluation that can predict resectability of the lesion and assist

in the design of an appropriate surgical approach. A Bismuth type IV lesion is defined as a tumor that can invade the secondary biliary radicals of both hepatic ducts. During the past decades, accompanied liver resection has been increasingly but gradually recognized as the mainstay of surgical approaches for targeting a Bismuth type IV tumor^[7-9], however, there are relatively few studies that have reported in any detail the factors that might affect long-term survival of type IV HC patients. The over-arching aim of the current study was to identify prognostic factors for long-term survival of patients following radical surgery for type IV HC patients, especially in the setting of lymphovascular invasion (LVI).

MATERIALS AND METHODS

Patient selection

One hundred and forty-two consecutive patients who underwent radical resection for a pathological diagnosis of type IV HC at the West China Hospital between January 2000 and February 2012 were enrolled in this study and then reviewed retrospectively. The inclusion criteria included the following: (1) patients who were confirmed with Bismuth type IV HC by pathological examination; and (2) patients who had undergone radical resection (R0 and R1 resection). The exclusion criteria included the following: (1) patients with gallbladder or intrahepatic cholangiocarcinoma extending to the hilum; (2) presence of a recurrent or metastatic tumor; and (3) R2 resection.

Preoperative workup

Preoperative assessment consisted of acquiring a medical history, physical examination, laboratory tests and radiographic analyses. All patients were evaluated by contrast-enhanced ultrasound, contrast-enhanced computed tomography or magnetic resonance cholangiography with magnetic resonance cholangiopancreatography with the intention of determining the Bismuth type, the location and extent of the tumor. Biliary drainage, including endoscopic retrograde cholangiopancreatography (ENBD) and percutaneous transhepatic cholangiodrainage (PTCD), was applied in the setting of patients presenting with obstructive jaundice that exceeded 85 $\mu\text{mol/L}$ total bilirubin.

Surgical characteristics of the patients

Based on the relative location and extent of the tumor, different types of resection were performed, which included extrahepatic bile duct resection and *en bloc* resection of the caudate lobe combined with left hemihepatectomy, right hemihepatectomy and trisectionectomy. In addition, standard regional lymph node dissection should be performed. However, under conditions where tumor metastases to the distant lymph nodes was confirmed during surgery, the surgical intervention was abandoned. According to

the American Joint Committee on Cancer (AJCC, 7th edition), the location of regional lymph nodes was defined as follows: along the common bile duct, cystic duct, portal vein and proper hepatic artery^[10]. Vascular resection and reconstruction was only performed when vessels could not be detached from the tumor.

Pathological examination

The pathological evidence of cancer was determined by examination of paraffin sections. All included Bismuth type IV HC cases were histopathologically confirmed by an experienced pathologist. The presence of tumor emboli within peritumoural endothelial lined spaces was defined as LVI. Resection margin was defined as ductal (*i.e.*, proximal and distal ducts) and with evidence of radial margins. The radial margin was defined as the vertical margin between the tumor edge and dissected periductal structures (*e.g.*, liver parenchyma, blood vessels and adjacent fat tissues). An R0 resection was defined as the presence of a microscopically tumor-free resection margin. An R1 resection was defined as microscopic evidence of tumor tissue at the resection margin, and an R2 resection was defined as macroscopic evidence of tumor tissue at the resection margin. In this study, radical resection was defined as an R0 and R1 resection, a negative resection margin indicated R0 resection, and a positive resection margin indicated an R1 resection.

Follow-up

Whether or not chemotherapy and radiotherapy can benefit HC patients was controversial. None of the patients received postoperative routine chemotherapy or radiotherapy. All enrolled patients had routine follow-ups every 3 mo in the first year and every 6 mo subsequently until at least 5 years after the surgery. The tumor markers [serum levels of carbohydrate antigen 19-9 (CA19-9) and carcinoembryonic antigen], liver functions and ultrasonography were conducted. If there was a suspicion of recurrence, contrast-enhanced computed tomography or magnetic resonance imaging was further performed. Tumor recurrence was diagnosed on the basis of the combined findings of typical radiological appearance, quantification of CA19-9 levels, and clinical presentation. The date of the first suspicious radiological finding was recorded as the date of initial disease recurrence.

Statistical analysis

Patient data were retrospectively collected and statistical analyses were performed with SPSS version 19.0 (SPSS Inc. Chicago, IL, United States). Survival was described using the Kaplan-Meier method and differences between subgroups were reviewed by the log-rank test. Multivariate analysis for prognostic factors was performed using a Cox proportional hazards model to analyze variables whose *P*-value was less than 0.1 in the univariate analysis. Multiple

logistical regression analysis was performed to determine the association between LVI and potential variables. Two-sided *P* values < 0.05 were considered statistically significant.

RESULTS

Patients' characteristics and operative outcomes

Patients' characteristics and operative outcomes are shown in Table 1. Altogether 142 patients had a radical resection for type IV HC, including 75 men and 67 women with a median age of 59 years (range: 23-78). Pre-operative biliary drainage was carried out in 105 of the 123 obstructive jaundice (total bilirubin > 85 μ mol/L) patients, wherein 71 patients underwent PCTD and 34 patients underwent ENBD. Preoperative portal vein embolization was performed in 6 patients.

Radical resection included extrahepatic bile duct resection and *en bloc* resection of the caudate lobe combined with left hemihepatectomy (*n* = 73, 51.4%), extended left hemihepatectomy (*n* = 5, 3.5%), left trisectionectomy (*n* = 6, 4.2%), right hemihepatectomy (*n* = 51, 35.9%), extended right hemihepatectomy (*n* = 5, 3.5%), and right trisectionectomy (*n* = 2, 1.4%). Regional lymph node dissection was conventionally performed. The R0 resection rate was 75.6%.

Clinicopathological variables influencing DFS and OS

As shown in Table 2, potential factors that might influence the DFS and OS were analyzed. Univariate analysis demonstrated that age (*P* = 0.039), preoperative ALB (*P* = 0.005), resection margin (*P* < 0.001), histologic grade (*P* = 0.023), T stage (*P* = 0.004), N stage (*P* < 0.001), AJCC stage (*P* < 0.001), LVI (*P* < 0.001), tumor size (*P* < 0.001), portal vein invasion (*P* = 0.003) and hepatic artery invasion (*P* = 0.008) significantly influenced DFS. By contrast, patient gender, preoperative CA19-9, surgical method, perineural invasion and transfusion did not significantly influence DFS. Preoperative ALB (*P* = 0.009), resection margin (*P* < 0.001), histologic grade (*P* = 0.026), T stage (*P* = 0.001), N stage (*P* < 0.001), AJCC stage (*P* < 0.001), LVI (*P* < 0.001), tumor size (*P* < 0.001), portal vein invasion (*P* = 0.002) and hepatic artery invasion (*P* = 0.013), but not patient age, gender, preoperative CA19-9, surgical methods, perineural invasion or transfusion, significantly influenced the OS. Multivariate analysis indicated that positive resection margin, higher N stage, tumor size > 30 mm and LVI were adverse factors that influenced DFS and OS.

Association between LVI and N stage and tumor size

LVI was confirmed histologically in 29 (20.4%) of the 142 patients. Table 3 shows the association of LVI with tumor size, N stage and AJCC stage. On univariate analysis, N stage (*P* < 0.001) and tumor size (*P* = 0.001), but not AJCC stage, were significantly associated with LVI. Multivariate analysis using a

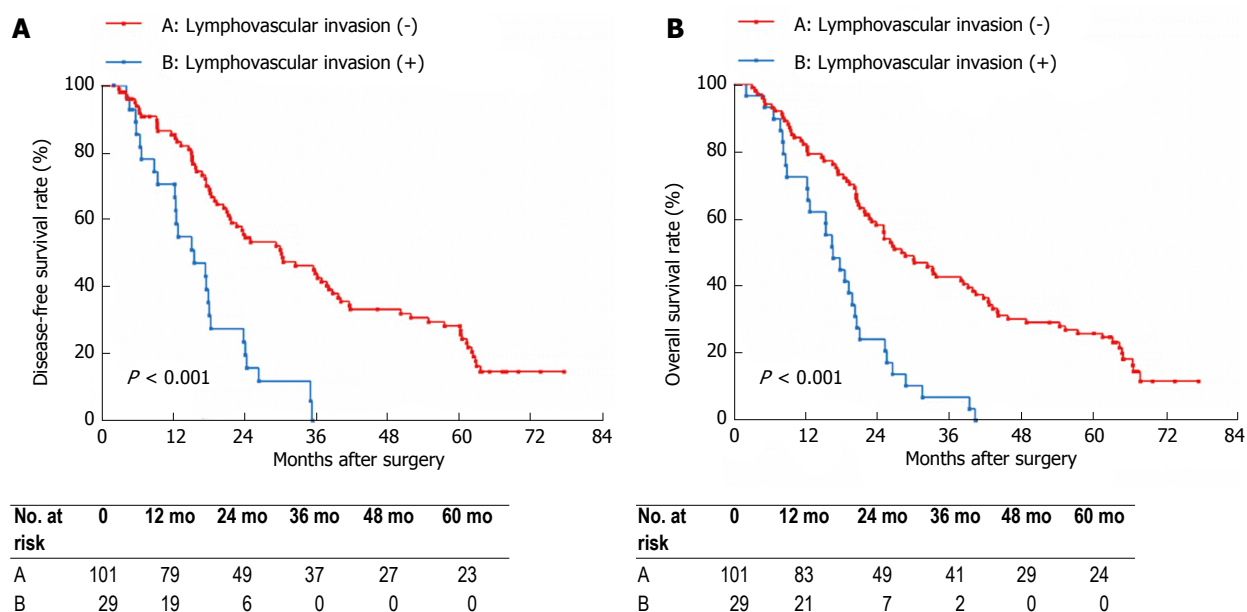


Figure 1 Disease-free survival and overall survival based on lymphovascular invasion in type IV hilar cholangiocarcinoma patients with radical resection. A: The 5-yr DFS rate in the LVI negative group was significantly higher than that in the LVI positive group; B: The 5-yr OS rate in the negative group was significantly higher than that in the LVI positive group. LVI: Lymphovascular invasion; DFS: Disease-free survival; OS: Overall survival.

Table 1 Demographics and operative outcomes

Characteristic	n (%)
Age (yr) ¹	59 (23-78)
Gender	
Male	75 (52.8)
Female	67 (47.2)
Albumin level (g/L) ²	36.74 ± 4.93
CA19-9	
> 200	82 (57.7)
< 200	60 (42.3)
Radiological examination	
Contrast-enhanced US	8 (5.6)
Contrast-enhanced CT	43 (30.3)
MRI + MRCP	91 (64.1)
Operative time (min) ²	429.47 ± 134.19
Blood loss (ml) ¹	600 (180-4000)
Transfusion	76 (53.5)
R0 resection	107 (75.4)
Differentiation degree	
Well differentiated	14 (9.9)
Moderately differentiated	90 (63.4)
Poorly differentiated	38 (26.8)
Perineural invasion positive	69 (48.6)
N stage	
N0	89 (62.7)
N1	53 (37.3)
Tumor diameter (mm) ¹	30 (12-55)
Hospital stay ¹	19 (5-115)

¹Parameters are presented as median and range; ²Parameters are presented as mean ± SD.

logistic regression model indicated that N1 stage ($P = 0.026$) and tumor size > 30 mm were significant factors that were associated with LVI in Bismuth type IV HC. DFS and OS based on LVI status are shown in Figure 1. The 5-year DFS rate in the LVI negative group was significantly higher than that in the LVI

positive group (22.8% vs 0%, $P < 0.001$) as was the OS (23.8% vs 0%, $P < 0.001$). Based on the LVI status, N0 stage patients and patients with tumor size ≤ 30 mm were divided into two subgroups, respectively. In the N0 stage subgroups, the 5-year DFS rate in the LVI negative group was significantly higher than that in the LVI positive group (31.0% vs 0%, $P = 0.002$), and the 5-year OS rate in the LVI negative group was also significantly higher than that in the LVI positive group (32.4% vs 0%, $P = 0.001$). In the tumor size ≤ 30 mm subgroups, the 5-year DFS rate in the LVI negative group was significantly higher than that in the LVI positive group (31.8% vs 0%, $P = 0.003$), and the 5-year OS rate in the LVI negative group was significantly higher than that in the LVI positive group (34.3% vs 0%, $P = 0.006$).

DISCUSSION

LVI is recognized as a dismal prognostic factor for OS in patients with breast cancer, colorectal cancer, and esophageal cancer^[11-14]. However, no studies have yet been published on whether LVI affects the prognosis of Bismuth type IV HC. Thus, the current study was undertaken to clarify the significance of LVI in type IV HC patients who had a radical resection. LVI is defined as the involvement of arterial vessels, venules and lymphatic channels^[11], but it is histologically difficult to distinguish, and the American Joint Committee on Cancer/Union Internationale Contre Cancer staging guidelines use the term lymphovascular to refer to those structures^[11]. Furthermore, LVI can be confirmed specifically on HE-stained specimens^[13]. In our series, LVI was confirmed histopathologically in 29 patients.

It is well recognized that resection margin status is

Table 2 Potential prognostic factors for disease-free survival and overall survival after radical resection for type IV hilar cholangiocarcinoma patients excluding operative mortality *n* (%)

Variable	Disease-free survival				Overall survival			
	5-yr survival	Univariate analysis		P value	5-yr survival	Univariate analysis		P value
		P value	Multivariate analysis HR (95%CI)			P value	Multivariate analysis HR (95%CI)	
Gender		0.682				0.965		
Male	13 (18.8)				14 (20.3)			
Female	10 (16.4)				10 (16.4)			
Age (yr)		0.039	1.291 (0.680-2.453)	0.435		0.230		
< 65	19 (19.2)				20 (20.2)			
> 65	4 (12.9)				4 (12.9)			
CA19-9		0.287				0.301		
< 200	13 (23.2)				14 (25)			
> 200	10 (13.5)				10 (13.5)			
ALB (g/L)		0.005	0.669 (0.413-1.085)	0.103		0.009	0.760 (0.492-1.173)	0.215
< 35	3 (7.1)				3 (7.1)			
> 35	21 (23.9)				20 (22.7)			
Surgical method		0.847				0.684		
Left-sided hepatectomy	13 (15.1)				13 (15.1)			
Right-sided hepatectomy	10 (17.2)				11 (19.0)			
Resection margin		< 0.001	6.255 (3.485-11.229)	< 0.001		< 0.001	6.776 (3.988-11.479)	< 0.001
Positive	0				0			
Negative	23 (23.7)				24 (24.7)			
Histologic grade		0.023	1.594 (0.994-2.554)	0.053		0.026	1.294 (0.830-2.017)	0.256
Well/moderate	20 (21.5)				21 (22.6)			
poor					3 (8.1)			
Perineural invasion		0.211				0.417		
Present	14 (22.6)				15 (24.2)			
Absent	9 (13.2)				9 (13.2)			
T stage		0.004	1.582 (0.390-6.415)	0.521		0.001	2.399 (0.734-7.836)	0.147
T1/2	22 (22)				21 (21)			
T3/4	2 (6.7)				2 (6.7)			
N stage		< 0.001	2.902 (1.132-7.439)	0.027		< 0.001	2.827 (1.243-6.429)	0.013
N0	22 (27.2)				23 (28.4)			
N1	1 (2.0)				1 (2.0)			
AJCC stage		< 0.001	0.673 (0.289-1.567)	0.358		< 0.001	0.844 (0.351-2.028)	0.704
Stage I / II	20 (32.8)				21 (34.4)			
Stage III/IV	3 (4.9)				3 (4.9)			
Lymphovascular invasion		< 0.001	2.799 (1.588-4.935)	< 0.001		< 0.001	2.908 (1.712-4.938)	< 0.001
Present	0					0.000		
Absent	23 (22.8)				24 (24.8)			
Tumor size (mm)		< 0.001	1.942 (1.176-3.209)	0.010		< 0.001	1.739 (1.101-2.745)	0.018
≤ 30	15 (20)				16 (21.3)			
> 30	8 (14.5)				8 (14.5)			
Portal vein invasion		0.003	1.759 (0.534-5.800)	0.353		0.002	1.130 (0.408-3.127)	0.815
Present	1 (4.2)				1 (4.2)			
Absent	22 (20.8)				23 (21.7)			
Hepatic invasion		0.008	1.499 (0.612-3.668)	0.376		0.013	1.196 (0.526-2.719)	0.669
Present	1 (9.1)	1 (9.1)			1 (9.1)			
Absent	22 (18.5)	22 (18.5)			23 (19.3)			
Transfusion		0.445				0.199		
Yes	11 (16.4)				11 (16.4)			
No	12 (19.0)				13 (20.6)			

the most important factor affecting long-term survival outcomes of HC patients^[14,15], and positive resection margin remains a major dismal prognostic factor for type IV HC patients. Furthermore, since type IV HC had been considered unresectable^[16], radical resection performed in such patients is technically challenging. Bracingly, in our center the R0 resection rate is 75.4%. Regional lymph node involvement represents another important dismal prognostic factor in HC patients who had undergone radical resection^[17,18]. Concordantly, the multivariate analysis showed that lymph node

metastasis was an adverse factor affecting DFS and OS in type IV HC. Furthermore, DeOliveira *et al.*^[19] proposed a new staging system, in which tumor size > 30 mm was defined as the T3 stage. The choice of 30 mm as a cutoff value for T3 is based on increasing evidence that the smaller the tumor, the better the prognosis^[20,21]. Our results indicated that tumor size > 30 mm had an unfavorable impact on both DFS and OS. More importantly, we found that LVI is a significant adverse prognostic factor influencing DFS and OS in multivariate analysis. To our knowledge, no other

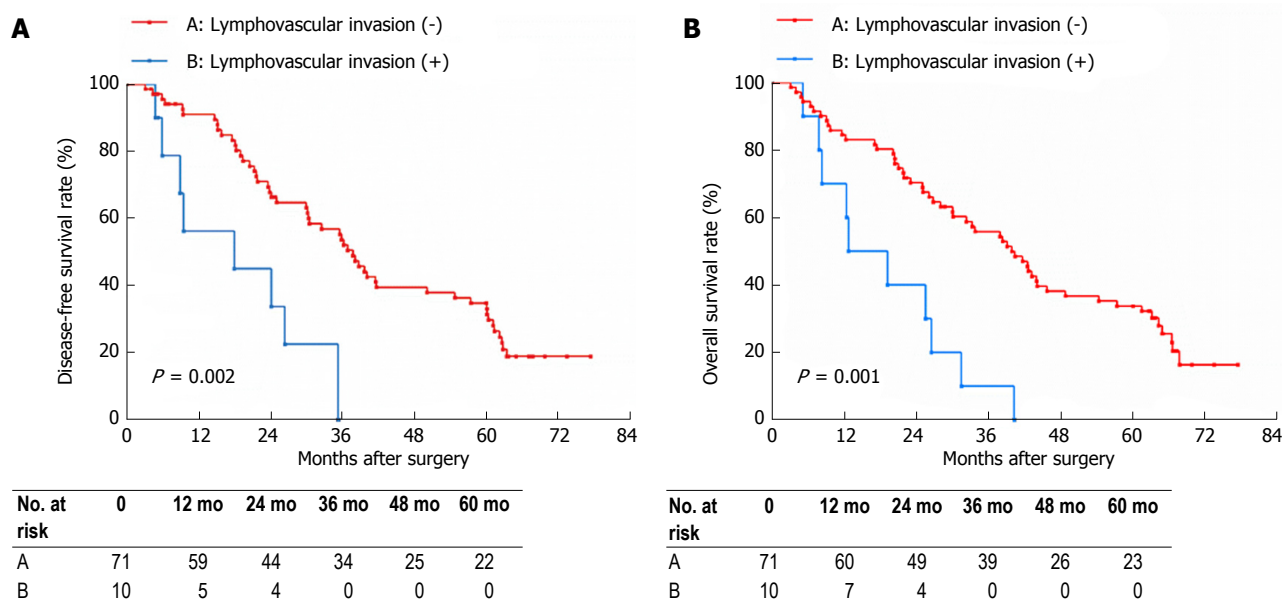


Figure 2 Disease-free survival and overall survival based on lymphovascular invasion in type IV hilar cholangiocarcinoma patients without lymph node metastasis. A: The 5-yr DFS rate in the LVI negative group was significantly higher than that in the LVI positive group; B: The 5-yr OS rate in the negative group was significantly higher than that in the LVI positive group. LVI: Lymphovascular invasion; DFS: Disease-free survival; OS: Overall survival.

reports have shown the correlation between LVI and the prognosis of type IV HC.

Previous studies suggested that LVI may interact with other adverse risk factors, which then have a dismal impact on the OS of esophageal cancer patients^[22,23]. Lee *et al*^[24] found that the presence of LVI correlated with the presence of lymph node metastasis in patients with gastric carcinoma. For different types of cholangiocarcinoma, the effect of LVI on prognosis was also different. Kim *et al*^[25] reported that LVI did not influence the survival of patients with distal cholangiocarcinoma. Fisher *et al*^[26] reported that LVI had an adverse influence on survival of patients with intrahepatic cholangiocarcinoma. Both of studies found that LVI was associated with lymph node metastasis^[25,26]. In our analysis, N1 stage was a significant factor that was associated with LVI in type IV HC patients. It is generally known that tumor cells and tumor stromal cells (such as macrophages and thrombocytes) can produce pro-lymphangiogenic factors, which increase lymphovascular density in and around the tumors^[27,28], mainly peritumoral regions. Lymph node metastases often occurred in tumors lacking intratumoral functional lymphatics, suggesting that functional lymphatics at the peritumoral regions are the route of lymphatic dissemination. Increased peritumoral lymphovascular density is considered to increase the flow of lymphatic fluid and provide an opportunity for invasive tumor cells to access the lymphatic vessels^[29]. This may elucidate our results that LVI is closely related to lymph node metastasis. Moreover, we speculated that LVI may be the precursor of lymph node metastasis. Thus, we divided patients without lymph nodes metastases into LVI positive and LVI negative subgroups, and found that the 5-year

DFS and OS rates in the LVI negative group were significantly higher than those in the LVI positive group (Figure 2A and B). This result indicates that LVI is an admirable prognostic predictor for patients with type IV HC when lymph node metastasis is absent.

Additionally, tumor size > 30 mm was another significant factor correlated with LVI in our series. Tumor size is recognized as a staging basis for many malignant tumors, including thyroid carcinoma, breast carcinoma, and liver carcinoma among others. Gurleyik *et al*^[30] reported that LVI correlated with tumor size, and the rate of LVI positive increased with tumor size in patients with breast carcinoma. The significant correlation between LVI and tumor size could be explained through two potential aspects: (1) as the tumor size increases, the peritumoral areas increase. Thus, the tumor is endowed with the potential to make contact with an increasing lymphovascularity, and thus the possibility of LVI increases; and (2) tumor size is proportional to the time of growth: the larger the tumor, the greater the duration of time that the tumor can continue developing and growing in size. During a relatively longer growth time, a tumor has increasing opportunities to develop LVI. In our study, patients with tumor size ≤ 30 mm were also divided into LVI positive and LVI negative subgroups. Here, the 5-year DFS and OS rates in the LVI negative group were significantly higher than those found in the LVI positive group (Figure 3A and B). This result indicates that LVI is also an excellent prognostic predictor in type IV HC patients with smaller tumor size.

Some limitations of the study should also be taken into account when interpreting the results. First, our study was retrospective with inherent limitations in its design. Thus, some clinical bias was inevitable. Next, LVI

Table 3 Logistic regression analysis for factors associated with lymphovascular invasion

Variable	Lymphovascular invasion (+), <i>n</i> = 29	Lymphovascular invasion (-), <i>n</i> = 113	Univariate analysis (<i>P</i> value)	OR (95%CI)	Multivariate analysis (<i>P</i> value)
AJCC stage			0.063	0.223 (0.026-1.927)	0.173
I / II	8	27			
III / IV	21	86			
Tumor diameter (mm)			0.001	3.258 (1.288-8.236)	0.013
≤ 30	10	77			
> 30	19	36			
N stage			< 0.001	3.312 (1.338-8.198)	0.026
N0	10	79			
N1	19	34			

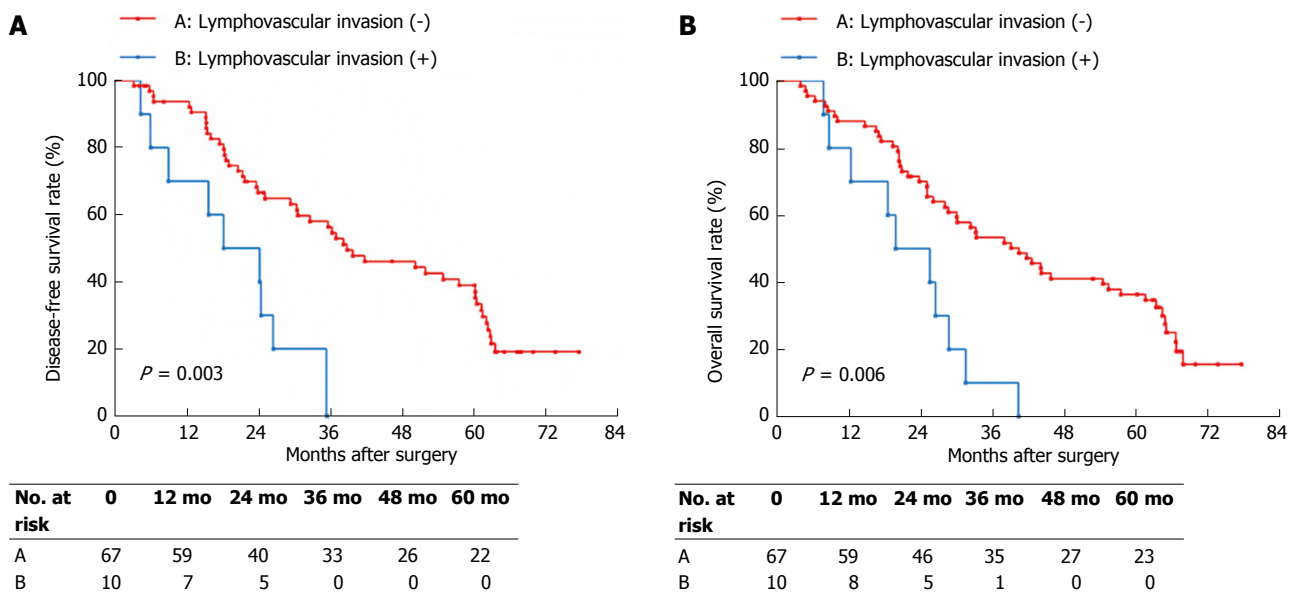


Figure 3 Disease-free survival and overall survival based on lymphovascular invasion in type IV hilar cholangiocarcinoma patients with tumor size ≤ 30 mm. A: The 5-yr DFS rate in the LVI negative group was significantly higher than that in the LVI positive group; B: The 5-yr OS rate in the negative group was significantly higher than that in the LVI positive group. LVI: Lymphovascular invasion; DFS: Disease-free survival; OS: Overall survival.

was confirmed by hematoxylin and eosin (HE) staining alone without application of an immunohistochemical staining with D2-40 antibody, which may improve the detection rate of LVI^[31]. Third, N1 stage and tumor size > 30 mm might serve as potential confounding factors that could affect the association between LVI and the eventual prognosis. Finally, we did not carry out preclinical medical experiments to elaborate the specific molecular mechanism that could play a key role in the capacity of LVI to affect the prognosis of type IV HC patients. Future research should take into account this topic with a greater sample size. Prospective studies, even randomized controlled trials, are also urgently needed. Of course, the specific molecular mechanism responsible for LVI affecting the prognosis of type IV HC patients will be determined by empirical research.

In conclusion, the presence of LVI may be regarded as an indicator of biologically aggressive behavior, metastatic ability, and regional and systemic risk of metastasizing the primary malignancy. LVI is associated with N1 stage and tumor size > 30 mm and imparts an adverse influence on OS and DFS in type IV HC patients

who received radical resection.

COMMENTS

Background

Despite advances in surgical techniques and that resection rates for Bismuth type IV hilar cholangiocarcinoma (HC) continue to increase, the prognosis of patients with type IV HC remains unsatisfactory. The reasons for this remain unclear and seem to be complex and multifactorial. Lymphovascular invasion (LVI) is associated with a poorer prognosis in patients with various malignancies. This study sought to investigate whether LVI could predict type IV HC prognosis.

Research frontiers

This study estimates prognostic factors that might be associated with overall survival (OS) and disease-free survival (DFS) after radical resection in type IV HC patients. To our knowledge, this study represents the first clinical insight indicating that LVI is associated with the prognosis of type IV HC.

Innovations and breakthroughs

This findings confirmed that R1 resection, N1 stage, presence of LVI and tumor size > 30 mm were adverse prognostic predictors for type IV HC patients after radical resection. Further, LVI was found to be associated with N1 stage and tumor size > 30 mm and adversely influence OS and DFS.

Applications

Observations from the present study might formally indicate novel factors for predicting post-surgical survival in HC. Moreover, LVI might present a potentially novel target for developing anti-cancer strategies.

Terminology

HC is a neoplasia arising from the biliary epithelium at the common hepatic duct bifurcation that might extend to the intrahepatic biliary tree and liver. Bismuth-Corlette classification is the most commonly used HC typing system, which is often used by surgeons to develop preliminary surgical protocols.

Peer-review

This is a retrospective study evaluating the effect of LVI on the prognosis of Bismuth-Corlette type IV HC. The authors concluded that LVI had an adverse influence on the prognosis of patients with Bismuth-Corlette type IV HC. This manuscript is very interesting and well written.

REFERENCES

- Xiong J, Nunes QM, Huang W, Wei A, Ke N, Mai G, Liu X, Hu W. Major hepatectomy in Bismuth types I and II hilar cholangiocarcinoma. *J Surg Res* 2015; **194**: 194-201 [PMID: 25454973 DOI: 10.1016/j.jss.2014.10.029]
- DeOliveira ML, Cunningham SC, Cameron JL, Kamangar F, Winter JM, Lillemoe KD, Choti MA, Yeo CJ, Schulick RD. Cholangiocarcinoma: thirty-one-year experience with 564 patients at a single institution. *Ann Surg* 2007; **245**: 755-762 [PMID: 17457168 DOI: 10.1097/01.sla.0000251366.62632.d3]
- Ji GW, Zhu FP, Wang K, Jiao CY, Shao ZC, Li XC. Clinical Implications of Biliary Confluence Pattern for Bismuth-Corlette Type IV Hilar Cholangiocarcinoma Applied to Hemihepatectomy. *J Gastrointest Surg* 2017; **21**: 666-675 [PMID: 28168674 DOI: 10.1007/s11605-017-3377-2]
- Kow AW, Wook CD, Song SC, Kim WS, Kim MJ, Park HJ, Heo JS, Choi SH. Role of caudate lobectomy in type III A and III B hilar cholangiocarcinoma: a 15-year experience in a tertiary institution. *World J Surg* 2012; **36**: 1112-1121 [PMID: 22374541 DOI: 10.1007/s00268-012-1497-0]
- Furusawa N, Kobayashi A, Yokoyama T, Shimizu A, Motoyama H, Miyagawa S. Surgical treatment of 144 cases of hilar cholangiocarcinoma without liver-related mortality. *World J Surg* 2014; **38**: 1164-1176 [PMID: 24305942 DOI: 10.1007/s00268-013-2394-x]
- Nagino M, Ebata T, Yokoyama Y, Igami T, Sugawara G, Takahashi Y, Nimura Y. Evolution of surgical treatment for perihilar cholangiocarcinoma: a single-center 34-year review of 574 consecutive resections. *Ann Surg* 2013; **258**: 129-140 [PMID: 23059502 DOI: 10.1097/SLA.0b013e3182708b57]
- Govil S, Reddy MS, Rela M. Surgical resection techniques for locally advanced hilar cholangiocarcinoma. *Langenbecks Arch Surg* 2014; **399**: 707-716 [PMID: 24893723 DOI: 10.1007/s00423-014-1216-4]
- Ito F, Cho CS, Rikkers LF, Weber SM. Hilar cholangiocarcinoma: current management. *Ann Surg* 2009; **250**: 210-218 [PMID: 19638920 DOI: 10.1097/SLA.0b013e3181afe0ab]
- Xiang S, Lau WY, Chen XP. Hilar cholangiocarcinoma: controversies on the extent of surgical resection aiming at cure. *Int J Colorectal Dis* 2015; **30**: 159-171 [PMID: 25376337 DOI: 10.1007/s00384-014-2063-z]
- Groot Koerkamp B, Wiggers JK, Allen PJ, Busch OR, D'Angelica MI, DeMatteo RP, Fong Y, Gonen M, Gouma DJ, Kingham TP, van Gulik TM, Jarnagin WR. American Joint Committee on Cancer staging for resected perihilar cholangiocarcinoma: a comparison of the 6th and 7th editions. *HPB (Oxford)* 2014; **16**: 1074-1082 [PMID: 25267346 DOI: 10.1111/hpb.12320]
- Hoda SA, Hoda RS, Merlin S, Shamonki J, Rivera M. Issues relating to lymphovascular invasion in breast carcinoma. *Adv Anat Pathol* 2006; **13**: 308-315 [PMID: 17075296 DOI: 10.1097/01.pap.0000213048.69564.26]
- Lee JH, Jang HS, Kim JG, Cho HM, Shim BY, Oh ST, Yoon SC, Kim YS, Choi BO, Kim SH. Lymphovascular invasion is a significant prognosticator in rectal cancer patients who receive preoperative chemoradiotherapy followed by total mesorectal excision. *Ann Surg Oncol* 2012; **19**: 1213-1221 [PMID: 21935746 DOI: 10.1245/s10434-011-2062-z]
- Lagarde SM, Phillips AW, Navidi M, Disep B, Immanuel A, Griffin SM. The presence of lymphovascular and perineural infiltration after neoadjuvant therapy and oesophagectomy identifies patients at high risk for recurrence. *Br J Cancer* 2015; **113**: 1427-1433 [PMID: 26554656 DOI: 10.1038/bjc.2015.354]
- Natsume S, Ebata T, Yokoyama Y, Igami T, Sugawara G, Shimoyama Y, Nagino M. Clinical significance of left trisectionectomy for perihilar cholangiocarcinoma: an appraisal and comparison with left hepatectomy. *Ann Surg* 2012; **255**: 754-762 [PMID: 22367444 DOI: 10.1097/SLA.0b013e31824a8d82]
- Nuzzo G, Giuliani F, Ardito F, Giovannini I, Aldrighetti L, Belli G, Bresadola F, Calise F, Dalla Valle R, D'Amico DF, Gennari L, Giuliani SM, Guglielmi A, Jovine E, Pellicci R, Pernthaler H, Pinna AD, Puleo S, Torzilli G, Capussotti L; Italian Chapter of the International Hepato-Pancreato-Biliary Association, Cillo U, Ercolani G, Ferrucci M, Mastrangelo L, Portolani N, Pulitanò C, Ribero D, Ruzzenente A, Scuderi V, Federico B. Improvement in perioperative and long-term outcome after surgical treatment of hilar cholangiocarcinoma: results of an Italian multicenter analysis of 440 patients. *Arch Surg* 2012; **147**: 26-34 [PMID: 22250108 DOI: 10.1001/archsurg.2011.771]
- Croome KP, Rosen CB, Heimbach JK, Nagorney DM. Is Liver Transplantation Appropriate for Patients with Potentially Resectable De Novo Hilar Cholangiocarcinoma? *J Am Coll Surg* 2015; **221**: 130-139 [PMID: 25872685 DOI: 10.1016/j.jamcollsurg.2015.01.064]
- Guglielmi A, Ruzzenente A, Campagnaro T, Pachera S, Conci S, Valdegamberi A, Sandri M, Iacono C. Prognostic significance of lymph node ratio after resection of peri-hilar cholangiocarcinoma. *HPB (Oxford)* 2011; **13**: 240-245 [PMID: 21418129 DOI: 10.1111/j.1477-2574.2010.00277.x]
- Giuliani F, Ardito F, Guglielmi A, Aldrighetti L, Ferrero A, Calise F, Giuliani SM, Jovine E, Breccia C, De Rose AM, Pinna AD, Nuzzo G. Association of Lymph Node Status With Survival in Patients After Liver Resection for Hilar Cholangiocarcinoma in an Italian Multicenter Analysis. *JAMA Surg* 2016; **151**: 916-922 [PMID: 27556741 DOI: 10.1001/jamasurg.2016.1769]
- Deoliveira ML, Schulick RD, Nimura Y, Rosen C, Gores G, Neuhaus P, Clavien PA. New staging system and a registry for perihilar cholangiocarcinoma. *Hepatology* 2011; **53**: 1363-1371 [PMID: 21480336 DOI: 10.1002/hep.24227]
- Wang ST, Shen SL, Peng BG, Hua YP, Chen B, Kuang M, Li SQ, He Q, Liang LJ. Combined vascular resection and analysis of prognostic factors for hilar cholangiocarcinoma. *Hepatobiliary Pancreat Dis Int* 2015; **14**: 626-632 [PMID: 26663011]
- Hu HJ, Mao H, Shrestha A, Tan YQ, Ma WJ, Yang Q, Wang JK, Cheng NS, Li FY. Prognostic factors and long-term outcomes of hilar cholangiocarcinoma: A single-institution experience in China. *World J Gastroenterol* 2016; **22**: 2601-2610 [PMID: 26937148 DOI: 10.3748/wjg.v22.i8.2601]
- Wang S, Chen X, Fan J, Lu L. Prognostic Significance of Lymphovascular Invasion for Thoracic Esophageal Squamous Cell Carcinoma. *Ann Surg Oncol* 2016; **23**: 4101-4109 [PMID: 27436201 DOI: 10.1245/s10434-016-5416-8]
- Chen WH, Huang YL, Chao YK, Yeh CJ, Chang HK, Tseng CK, Liu YH. Prognostic significance of lymphovascular invasion in patients with esophageal squamous cell carcinoma treated with neoadjuvant chemoradiotherapy. *Ann Surg Oncol* 2015; **22**: 338-343 [PMID: 25023545 DOI: 10.1245/s10434-014-3881-5]
- Lee K, Park DJ, Choe G, Kim HH, Kim WH, Lee HS. Increased intratumoral lymphatic vessel density correlates with lymph node metastasis in early gastric carcinoma. *Ann Surg Oncol* 2010; **17**: 73-80 [PMID: 19777179 DOI: 10.1245/s10434-009-0707-y]
- Kim HJ, Kim CY, Hur YH, Koh YS, Kim JC, Kim HJ, Cho CK.

- Prognostic factors for survival after curative resection of distal cholangiocarcinoma: perineural invasion and lymphovascular invasion. *Surg Today* 2014; **44**: 1879-1886 [PMID: 24535697 DOI: 10.1007/s00595-014-0846-z]
- 26 **Fisher SB**, Patel SH, Kooby DA, Weber S, Bloomston M, Cho C, Hatzaras I, Schmidt C, Winslow E, Staley CA 3rd, Maithel SK. Lymphovascular and perineural invasion as selection criteria for adjuvant therapy in intrahepatic cholangiocarcinoma: a multi-institution analysis. *HPB (Oxford)* 2012; **14**: 514-522 [PMID: 22762399 DOI: 10.1111/j.1477-2574.2012.00489.x]
 - 27 **Schlereth SL**, Refaian N, Iden S, Cursiefen C, Heindl LM. Impact of the prolymphangiogenic crosstalk in the tumor microenvironment on lymphatic cancer metastasis. *Biomed Res Int* 2014; **2014**: 639058 [PMID: 25254213 DOI: 10.1155/2014/639058]
 - 28 **Rofstad EK**, Huang R, Galappathi K, Andersen LM, Wegner CS, Hauge A, Gaustad JV, Simonsen TG. Functional intratumoral lymphatics in patient-derived xenograft models of squamous cell carcinoma of the uterine cervix: implications for lymph node metastasis. *Oncotarget* 2016; **7**: 56986-56997 [PMID: 27486768 DOI: 10.18632/oncotarget.10931]
 - 29 **Harrell MI**, Iritani BM, Ruddell A. Tumor-induced sentinel lymph node lymphangiogenesis and increased lymph flow precede melanoma metastasis. *Am J Pathol* 2007; **170**: 774-786 [PMID: 17255343 DOI: 10.2353/ajpath.2007.060761]
 - 30 **Gurleyik G**, Gurleyik E, Aker F, Aktekin A, Emir S, Gungor O, Saglam A. Lymphovascular invasion, as a prognostic marker in patients with invasive breast cancer. *Acta Chir Belg* 2007; **107**: 284-287 [PMID: 17685254]
 - 31 **Weber SK**, Sauerwald A, Pölcher M, Braun M, Debald M, Serce NB, Kuhn W, Brunagel-Walgenbach G, Rudlowski C. Detection of lymphovascular invasion by D2-40 (podoplanin) immunoexpression in endometrial cancer. *Int J Gynecol Cancer* 2012; **22**: 1442-1448 [PMID: 22964524 DOI: 10.1097/IGC.0b013e318269139b]

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Retrospective Study

Gastrointestinal symptom prevalence depends on disease duration and gastrointestinal region in type 2 diabetes mellitus

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Abstract

AIM

To unravel relationships between gastrointestinal (GI) symptoms impairing quality of life (QOL) and clinical profiles of diabetes mellitus (DM) patients.

METHODS

We enrolled 134 outpatients with type 2 DM. Mean

age was 64.7 years, mean body mass index was 24.7 kg/m², mean glycated hemoglobin was 7.1%, and mean DM duration was 13.7 years. GI symptom-related QOL was determined using the Izumo scale, based on five factors, *i.e.*, heartburn, gastralgia, postprandial fullness, constipation and diarrhea. The sum of scores obtained for the three questions in each domain was calculated, and subjects with a score of 5 or higher were considered to be symptomatic with impaired QOL. JMP Clinical version 5.0 was used for all statistical analyses.

RESULTS

Lower abdominal symptoms were found to be more frequent than those affecting the upper abdomen. Diabetic duration and medications showed associations with GI symptoms. We identified differences in peak prevalences of the five symptoms. Gastralgia ($P = 0.02$ *vs* 10-14 years) and total GI symptoms ($P = 0.01$ and $P = 0.02$ *vs* 5-9 years and 10-14 years, respectively) peaked at a diabetes duration of 15-19 years. Heartburn ($P = 0.004$) and postprandial fullness ($P = 0.03$) tended to increase with disease duration. Constipation and diarrhea showed bimodal peaks, with the first early and the second late (*e.g.*, $P = 0.03$ at 15-19 years *vs* 10-14 years for diarrhea) in the disease course. Finally, GI symptoms showed clustering that reflected the region of the GI tract affected, *i.e.*, constipation and diarrhea had similar frequencies ($P < 0.0001$).

CONCLUSION

Our study highlights the importance of questioning patients about QOL impairment due to abdominal symptoms, especially in the early and the late periods of diabetes.

Key words: Gastrointestinal symptoms; Questionnaire survey; Disease duration; Type 2 diabetes; Quality of life; Gastrointestinal tract regions

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Core tip: We describe the results of a questionnaire survey of 134 type 2 diabetes mellitus outpatients experiencing gastrointestinal symptoms. The novel finding is that symptom frequencies differed among disease durations and according to affected gastrointestinal regions. Lower abdominal symptoms not only manifested during the late but also in the early stage of diabetes when there were no organ complications related to this disease. Our study highlights the importance of not underestimating gastrointestinal symptoms and of questioning patients about quality of life impairment due to abdominal symptoms, especially in both the early period and after a diabetes duration of 10 or even 15 years.

Fujishiro M, Kushiyaama A, Yamazaki H, Kaneko S, Koketsu Y, Yamamotoya T, Kikuchi T, Sakoda H, Suzuki R, Kadowaki T.

Gastrointestinal symptom prevalence depends on disease duration and gastrointestinal region in type 2 diabetes mellitus. *World J Gastroenterol* 2017; 23(36): 6694-6704 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6694.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6694>

INTRODUCTION

The dramatic increase in the incidence of type (T)2 diabetes mellitus (DM) over the last century has become a major public health concern worldwide^[1]. Both micro- and macrovascular complications of diabetes damage a variety of organs, leading to various symptoms, and impair quality of life (QOL) in patients with T2DM.

Among these symptoms, those affecting the gastrointestinal (GI) tract occur frequently in patients with diabetes^[2]. Various GI symptoms are associated with this disease^[3-5], especially diabetic neuropathy which develops with longstanding diabetes^[6,7]. GI hyper- and hypofunction lead to delayed gastric or esophageal emptying, diabetic gastroparesis, constipation and diarrhea^[6,8,9]. Other GI-related disorders and symptoms are also known to accompany diabetes, such as obesity with gastroesophageal reflux disease^[10,11], and aging with gastric mucosal injury^[12].

GI disorders in diabetes have thus far been poorly characterized, despite their high frequencies in both T1DM and T2DM patients^[13]. Hyperglycemia-associated neuropathy was long considered to be a major mechanism underlying the pathogenesis of GI symptoms, probably *via* oxidative stress and inflammation, as with other microangiopathies. Levels of enteric hormones, such as incretin-related peptides [*e.g.*, glucagon-like peptide-1 (GLP-1), GLP-2, pancreatic polypeptide (PPY)]^[14] and serotonin^[15], and enteric neurotransmitters, such as vasoactive intestinal peptide (VIP)^[16], are altered in patients with DM, and affect GI motility and neural fiber growth. Furthermore, smooth muscle cells, interstitial cells of Cajal^[17], gut microbiota^[18] and intestinal stem cells^[19], may be altered in DM and these changes are potentially related to GI symptoms. The relevance of these mechanisms in the development of human GI symptoms in T2DM remains uncertain.

As for the pathophysiological mechanisms underlying the development of complications, hyperglycemia and the resultant glycation products^[20], oxidative stress^[21] and inflammation^[22], are the main candidates. GI symptoms in DM are associated with both poor glycemic control and diabetic complications, as shown in a large number of subjects^[23], although we found no relationships between GI symptoms and glycemic control, probably due to the long and varied disease durations of our subjects.

There has been little systematic and comprehensive research focusing on GI conditions in patients with

T2DM. The Izumo Scale, a validated and useful tool for QOL assessment, is a scale for assessing GI symptoms^[13,14]. Applying this scale, we investigated actual GI symptoms employing a questionnaire and determined whether the symptoms identified are related to the clinical profiles of patients with T2DM.

MATERIALS AND METHODS

Patients

Patients who visited the department of diabetes and metabolic diseases in our hospital, including those with T1DM and T2DM, from December 2011 to March 2014, were enrolled in this study. We asked 200 consecutive patients to answer the questionnaire. Of the 170 patients who gave informed consent and responded to the questionnaire, data from 134 were analyzed after exclusion of 31 non-T2DM patients and 5 with a past history of GI tract surgery. This protocol was approved by our institutional ethics review committees (approval number 3643). Patient QOL was assessed by asking the subjects if they suffered from GI symptoms and then scoring the symptoms according to the Izumo scale, as described below.

Patient profiles, including diabetic microangiopathy, were collected from medical records. We considered subjects to have "diabetic autonomic neuropathy (DAN)" if they had any autonomic signs or symptoms. Distal symmetric polyneuropathy (DSP) includes symmetric neurosensory signs and symptoms, such as tingling, numbness or cramps in the legs, based on the attending physician's assessment. Diabetic retinopathy (DR) includes non-proliferative DR diagnosed by the presence of microaneurysms and retinal hemorrhages^[24], and all DR cases were confirmed by an ophthalmologist. Diabetic nephropathy was clinically diagnosed by attending physicians based on microalbuminuria or overt proteinuria with no evidence of other kidney or urological disease^[25].

Izumo Scale

The Izumo Scale was developed and validated by Furuta *et al.*^[26]. The Izumo Scale is a self-administered questionnaire designed to assess the effects of abdominal symptoms on QOL and includes 15 items in five domains with three items in each domain: heartburn (questions 1-3), gastralgia (questions 4-6), postprandial fullness (questions 7-9), constipation (questions 10-12), and diarrhea (questions 13-15)^[27]. Each question is rated on a 6-point Likert scale from 0 to 5, with higher values indicating more severe symptoms. This scale has been shown to have good internal consistency, reproducibility, as well as good correlations with the visual analogue scale of abdominal symptoms and the Gastrointestinal Symptoms Rating Scale (GSRS), and is thus widely utilized in Japan^[28-31]. The sum of scores obtained for the three questions in each domain was calculated, and subjects with a score of 5 or higher were considered to be symptomatic

with impaired QOL^[30]. When the five symptoms were classified by GI region, heartburn was considered to involve the esophagus, gastralgia and postprandial fullness the upper GI tract, and constipation and diarrhea the lower GI tract.

Statistical analysis

JMP Clinical version 5.0 (SAS Institute, Cary, NC, United States) was used for all statistical analyses. ANOVA was used to compare scores among different groups. Frequencies were compared using Fisher's exact test. Trend comparisons for frequencies among disease duration, age, body mass index (BMI), and glycated hemoglobin (HbA1c) were performed using a Cochran-Armitage test. Hierarchical cluster analysis was performed to assess the relationships among GI symptoms. All *P* values are two-sided, and *P* < 0.05 was considered to indicate a statistically significant difference.

RESULTS

Characteristics of the enrolled patients

The characteristics of enrolled patients are listed in Table 1. We enrolled 134 patients with T2DM (87 males and 47 females, mean age: 64.7 years, range: 29-88 years). Mean diabetes duration was 13.7 (0.3-33.0) years, BMI was 24.7 (16.5-42.5) kg/m² and HbA1c was 7.1 (5.2-11.6)%. As to incidences of diabetic microangiopathy, 32 patients (24%) had DAN, 20 (15%) had DSP, 31 (23%) had DR and 64 (48%) had nephropathy. As to antidiabetic drugs, 8 patients (6%) received no medications, 34 (25%) used various forms of insulin including 22 (16%) also taking oral antidiabetic drugs (OADs), the details of which are presented in Table 1.

Eighty-six (64%) patients received OAD only, including 63 (47%) with one OAD, 30 (22%) with 2 OADs, 22 (16%) with three OADs, and 11 (8%) with four or more OADs. In detail, 38 patients (28%) used α -glucosidase inhibitors, 67 (50%) used biguanides, 26 (19%) used thiazolidinediones, 54 (40%) used sulfonylureas, 3 (2%) used glinides and 53 (40%) used dipeptidyl peptidase-4 (DPP4) inhibitors, at various daily doses, as indicated in Table 1. Six patients (4%) used GLP-1 agonists, 3 of whom were also taking an OAD. Sixty-two patients (46%) were receiving diet therapy with daily intakes ranging from 1200-1800 kcal (21-29 kcal/IBW) and, in 51 patients (38%), salt was restricted to less than 6 g/d.

As to antithrombotic or anti-inflammatory agents, 29 patients (22%) used antiplatelet agents, 7 (5%) used anticoagulants, 8 (6%) used steroids or non-steroidal anti-inflammatory drugs and 7 (5%) used other antithrombotic agents such as prostaglandin E1 derivatives, prostaglandin I2 derivatives, or ethyl esters of eicosapentaenoic acid. As to GI agents, 29 patients (22%) used antacids, 13 (10%) used mucosal protectives, 11 (8%) used antimicrobials, and 18

Table 1 Characteristics of the study population

Variable	n (%)	mean (95%CI)
Sex, male/female	87/47 (65/35)	
Age of 65 yr or older	72 (54)	64.7 (62.8-66.6)
BMI of 25 kg/m ² or more	56 (42)	24.7 (24.0-25.4)
HbA1c of 7% or more	69 (51)	7.1 (6.9-7.3)
Duration of diabetes, yr		13.7 (12.4-14.9)
≤ 4	12 (9)	
5-9	27 (20)	
10-14	35 (26)	
15-19	31 (23)	
≥ 20	29 (22)	
Incidence of diabetic microangiopathy		
Autonomic neuropathy	32 (24)	
Distal symmetric polyneuropathy	20 (15)	
Retinopathy	31 (23)	
Nephropathy	64 (48)	
Use of antidiabetic drugs		
None	8 (6)	
Insulins	34 (25)	
Insulins with OAD	22 (16)	
OAD only	86 (64)	
1 OAD	63 (47)	
2 OAD	30 (22)	
3 OAD	22 (16)	
≥ 4 OAD	11 (8)	
GLP-1	6 (4)	
GLP-1 with OAD	3 (2)	
Insulins	34 (25)	
Range of injections per day	1-5	
Range of total daily insulin doses, IU	2-114	
Insulin glargine	12	
Insulin detemir	7	
Human NPH insulin	14	
Insulin aspart	2	
Insulin glulisine	4	
Insulin lispro	2	
Premixed human insulin 30/70	9	
Biphasic insulin aspart 30/70	1	
α-Glucosidase inhibitors	38 (28)	
Acarbose (150/200/300 mg)	3 (1/1/1)	
Miglitol (50/75/100/150 mg)	22 (1/12/18)	
Voglibose (0.6/0.9 mg)	13 (5/8)	
Biguanides	67 (50)	
Buformin 150 mg	1	
Metformin (500/750/1000/1500/2250 mg)	66 (12/20/14/19/1)	
Thiazolidinediones	26 (19)	
Pioglitazone (15/30 mg)	26 (18/8)	
Sulfonylureas	54 (40)	
Glimepiride (0.5/1/2/3/4 mg)	26 (10/10/2/1/3/)	
Gliclazide (20/40/120 mg)	21 (15/5/1)	
Glibenclamide (1.25/2.5/5 mg)	7 (1/4/2)	
Glinides	3 (2)	
Repaglinide 0.75 mg	1	
Mitiglinide (10/30 mg)	2 (1/1)	
Dipeptidyl peptidase-4 inhibitors	53 (40)	
Alogliptin 25 mg	12	
Sitagliptin (50/100 mg)	17 (15/2)	
Vildagliptin (50/100 mg)	24 (5/19)	
Glucagon-like peptide-1 agonists	6 (4)	
Liraglutide (0.6/0.9 mg)	4 (1/3)	
Exenatide 20 mg	2	
Dietary information		
Receiving dietary therapy	62 (46)	
With salt restriction to < 6 g per day	51 (38)	
Range of total energy intake, kcal	1200-1800	
Range of total energy intake per IBW, kcal/IBW	21-29	
Use of antithrombotic or anti-inflammatory agents		
Antiplatelet agents	29 (22)	
Anticoagulants	7 (5)	

Steroids/NSAIDs	8 (6)
Others ¹	7 (5)
Use of GI agents	
Antacids	29 (22)
Mucosal protectives	13 (10)
Antimicrobials	11 (8)
GI stimulants	18 (13)
Anti-diarrheals	0 (0)
Baseline blood parameters, mean \pm SD	
WBC, $\times 1000/\mu\text{L}$	6.1 \pm 1.9
RBC, $\times 10000/\mu\text{L}$	439 \pm 78.1
Hb, g/dL	13.4 \pm 2.4
Hct, %	40.8 \pm 6.9
Plt, $\times 10000/\mu\text{L}$	21.9 \pm 6.6
TP, g/dL	7.1 \pm 0.5
Alb, g/dL	3.8 \pm 0.3
AST(GOT), U/L	24.4 \pm 13.4
ALT(GPT), U/L	24.5 \pm 17.7
γ -GTP, U/L	34.1 \pm 24.3
CK, U/L	113 \pm 63.6
T-Cho, mg/dL	180 \pm 39.2
HDL-C, mg/dL	60.3 \pm 18.3
TG, mg/dL	128 \pm 67.2
cLDL, mg/dL	95.4 \pm 26.2
BUN, mg/dL	17.6 \pm 11.9
Cre, mg/dL	0.9 \pm 0.8
eGFR, mL/min per 1.73 m ²	66.6 \pm 22.8
UA, mg/dL	5.5 \pm 1.4

¹Others' includes other antithrombotic agents such as prostaglandin E1 derivatives, prostaglandin I2 derivatives, or ethyl esters of eicosapentaenoic acid. Data are expressed as means with the 95%CI in parentheses, with numbers for each sex or numbers with the percentage of the total in parentheses. BMI: Body mass index; GI: Gastrointestinal; HbA1c: Hemoglobin A1c; IBW: Ideal body weight; NPH: Neutral protamine Hagedorn; NSAIDs: Nonsteroidal anti-inflammatory drugs; OAD: Oral antidiabetic drug.

(13%) used GI stimulants, but none were taking anti-diarrheal agents. Baseline blood parameter findings were unremarkable.

GI symptoms and diabetes duration

In the QOL assessment employing the Izumo Scale, the scores of total GI symptoms peaked during the 15- to 19-year diabetes duration period (Figure 1). When GI symptoms were analyzed separately, lower GI symptoms were found to be most frequent. Prevalence patterns differed between the esophagus/upper GI tract and lower GI tract regions. Scores for heartburn ($P = 0.004$) and postprandial fullness ($P = 0.03$) consistently and significantly rose as disease duration increased (Figure 2A).

Scores for gastralgia, like total GI symptoms, peaked during the 15- to 19-year diabetes duration period. Interestingly, only the scores for constipation and diarrhea showed bimodal peaks, the first in the early period of diabetes and the second late in the disease course. Age showed a similar but weaker relationship to diabetes duration. There were no apparent relationships of GI symptoms with age or BMI, but postprandial fullness, alone among the five symptoms, was slightly associated with high HbA1c levels (Figure 2B-D).

Relationships of GI symptoms with diabetic complications and medications

Next, we examined whether there were any relation-

ships between GI symptoms and patient characteristics such as diabetic microangiopathy and the drugs administered (Figure 3A-D). None of the GI symptoms showed significant relationships with any of the forms of diabetic microangiopathy (Figure 3A), though symptoms tended to be more frequent in patients with than in those without DAN and/or DSP. As shown in Figure 3B, the rate of DPP4 inhibitor use was lower in patients suffering from constipation ($P = 0.04$). Insulin therapy and oral hypoglycemic agents other than DPP4 inhibitors were not related to GI symptoms. The numbers of patients using glinides or GLP-1 agonists were too small to allow statistically meaningful evaluation. As shown in Figure 3C, of the 29 patients taking antiplatelet agents, 38% ($n = 11$) had constipation ($P = 0.046$) and 62% ($n = 18$) had diarrhea ($P = 0.03$). Of the 18 patients using GI stimulants, 44% ($n = 8$) had constipation ($P = 0.03$) (Figure 3D).

We next investigated the effect of diabetic duration on constipation, excluding users of DPP4 inhibitors, antiplatelet agents or GI stimulants. We focused on the association of diabetes duration with constipation (Figure 4A), and on that with diarrhea, except in antiplatelet agent users (Figure 4B). When users of these medications were excluded, the early and late peaks in constipation and diarrhea remained.

Clustering and overlap of GI symptoms

A cluster analysis tree is presented in Figure 5. Gastralgia and postprandial fullness are more similar to

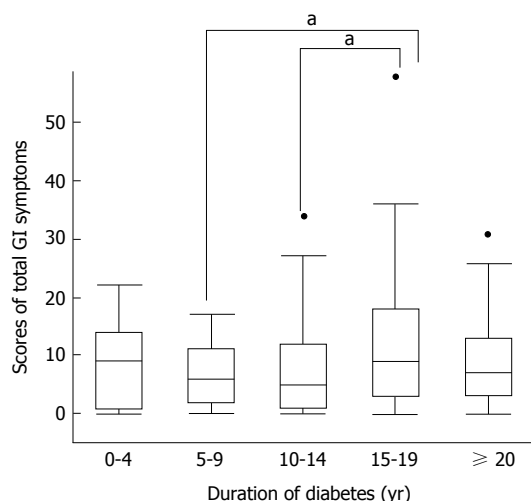


Figure 1 Distribution of the sum of scores of symptoms related to all GI areas by duration of diabetes. ^a $P < 0.05$, significantly different. Sums of symptom scores peaked during the 15- to 19-yr diabetes duration period, compared by Student's *t*-tests for paired periods ($P = 0.01$ and $P = 0.02$ when compared with 5-9 yr and 10-14 yr disease durations, respectively).

each other than to constipation and diarrhea. Heartburn is closer to upper than to lower GI symptoms. The relationship between constipation and diarrhea was found to be closer than the other associations identified in this study. The overlaps between two different symptoms on the Izumo Scale are shown in Table 2. Significant differences, determined by Fisher's exact test, between the presence and absence of two distinct symptoms, were demonstrated.

As in the cluster analysis, heartburn was associated with gastralgia ($P = 0.03$) or postprandial fullness ($P = 0.002$). Gastralgia and postprandial fullness ($P < 0.0001$) were strongly related to each other, as were constipation and diarrhea ($P < 0.0001$). Postprandial fullness was found to be related to all GI tract regions, showing modest interactions with both constipation ($P = 0.004$) and diarrhea ($P = 0.001$). Furthermore, all 31 patients with constipation also had episodes of diarrhea, while only 52.5% of those complaining mainly of diarrhea also had constipation (Table 2).

DISCUSSION

We investigated whether outpatients with T2DM actually have GI symptoms and, if present, the clinical severity of such symptoms. However, patients do not usually complain of GI symptoms. Overall, 10%-20% of adults have functional GI disorders^[32-35]. In patients with diabetes, symptoms involving all portions of the GI tract are reportedly more common than in the general population^[36]. However, we found that total symptom scores did not increase linearly with disease duration. In fact, rates peaked during different periods, according to GI regions. Lower abdominal symptoms were prominent early in the disease course, while diabetic complications were still mild, as well as in the late stage.

Significant confounding factors include aging, glycemic control and BMI reduction^[5,8-10,37]. Japanese patients have become increasingly obese over the past 20 years and those in the younger generation have poor glycemic control at the diagnosis of T2DM, independent of hyperglycemic symptoms^[38]. Therefore, symptom prevalence may change in the future.

We found no significant relationships between GI symptoms and either DAN or DSP. There might, however, be a detection bias or low sensitivity for DAN and DSP, since a previous report noted that symptoms tend to be related to lifestyle factors, rather than to either glycemic control or peripheral neuropathy^[36]. Another study found no association of GI symptoms with either diabetic neuropathy or psychiatric illnesses^[39]. Early symptoms might well be related to psychiatric illnesses and/or lifestyle factors but we did not obtain data pertaining to such factors in the present study.

Among anti-hyperglycemic medications, only the relationship between DPP4 inhibitors and constipation was significant. Constipation is a well-known side effect of these drugs^[40], and physicians often discontinue or do not start DPP4 inhibitors due to this adverse effect^[41]. Oral medications are generally prescribed to patients with poor glycemic control^[42] and are frequently used to manage longstanding diabetes^[43]. Administration of anti-platelet agents was associated with a variety of symptoms, as expected. These agents were still preferentially used despite apparently being harmful to the upper, middle and lower GI tracts^[44,45], since the treatment of cardiovascular diseases has the highest priority. However, the bimodal peaks for constipation and diarrhea late and early in the course of diabetes were maintained and were independent of medication use.

The mechanisms underlying rapid development of lower GI complications, such as diarrhea and constipation, are unknown. In our cluster analysis, the affected GI tract region was found to be an important factor. Furthermore, the mechanisms underlying the bimodal early and late disturbances are suggested to not be the same. Early diabetic neuropathy is reportedly multifactorial, being triggered by impairment of insulin signaling, abnormal blood flow, and oxidative stress including N-acetylglucosamine, activation of protein kinase C, activation of the polyol sugar pathway and glucose autooxidation, as well as non-enzymatic protein glycation^[46]. Insulin growth factors (IGFs) possess multiple neurotrophic functions, including neuronal survival, neurite outgrowth and regeneration. C-peptide also exerts IGF-like activity, such that endogenous proinsulin production might be related to the pathogenesis of GI symptoms^[47]. One possible explanation of mechanistic differences between early lower GI symptoms and upper GI symptoms might involve cellular composition. The lower GI tract contains colonic stem cells and their functions are disrupted in states of diabetic enteropathy^[19].

Table 2 Overlapping of gastrointestinal symptoms

Overlapping symptom	Heartburn, <i>n</i> = 9	Gastralgia, <i>n</i> = 11	Postprandial fullness, <i>n</i> = 17	Constipation, <i>n</i> = 31	Diarrhea, <i>n</i> = 59
Heartburn	-	3/11 ^a	5/17 ^b	5/31	6/59
Gastralgia	3/9 ^a	-	8/17 ^b	4/31	7/59
Postprandial fullness	5/9 ^b	8/11 ^a	-	9/31 ^b	14/59 ^b
Constipation	5/9	4/11	9/17 ^b	-	31/59 ^b
Diarrhea	6/9	7/11	13/17 ^b	31/31 ^b	-

^a*P* < 0.05 or ^b*P* < 0.01, significantly different.

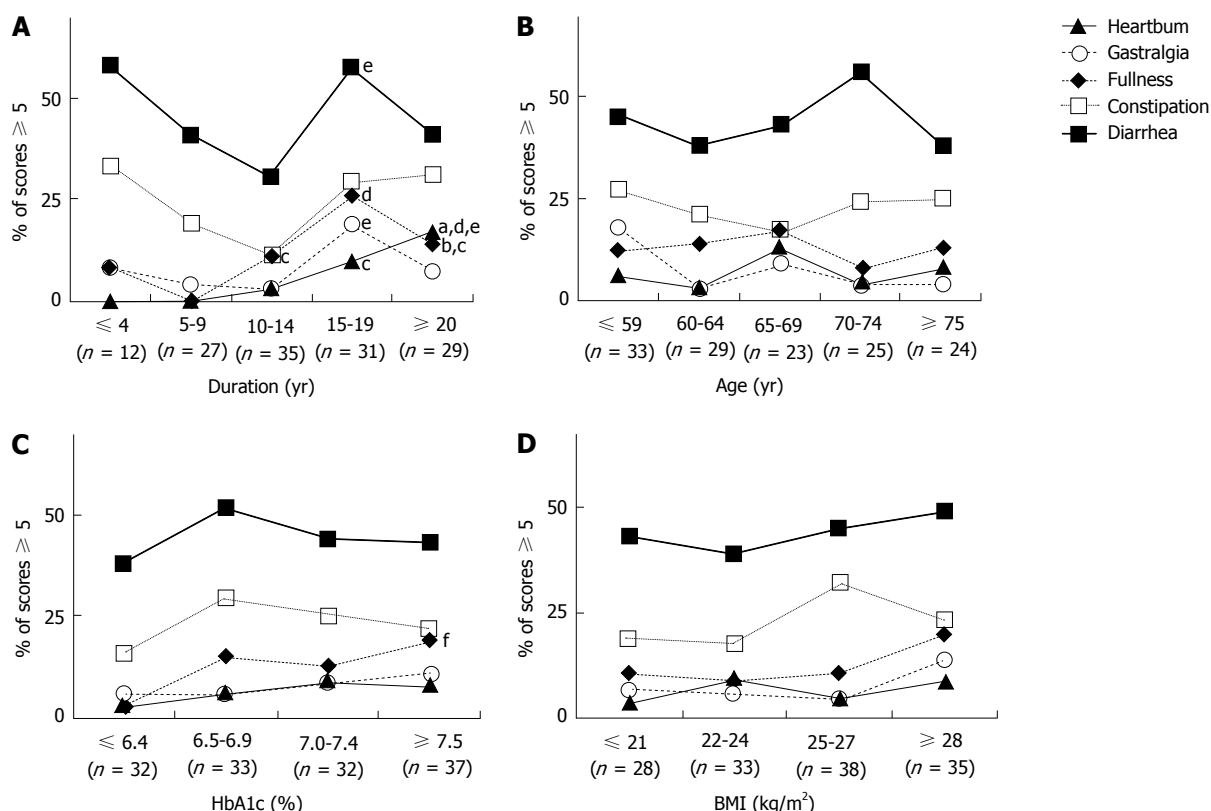


Figure 2 Relationships of abdominal symptoms with duration of diabetes, age, body mass index and hemoglobin A1c. The lines show the percentages of subjects with a score of 5 or higher, for each GI region, among those considered to be sufficiently symptomatic to impair quality of life. Comparisons among groups with five disease duration periods (A), five age ranges (B), four HbA1c levels (C) and four BMI levels (D) were performed using a Cochran-Armitage-trend-test. ^a*P* < 0.05 or ^b*P* < 0.01, significantly different. As shown in the figure (A), scores for heartburn (*P* = 0.004) and postprandial fullness (*P* = 0.029) consistently and significantly increased with disease duration. Student's *t*-tests for paired samples were performed to compare percentages of subjects with a score of 5 or higher, for each GI region, among five durations of diabetes (A), five age ranges (B), four HbA1c levels (C) and four BMI levels (D). ^c*P* < 0.05 vs 5-9 years, ^d*P* < 0.01 vs 5-9 years, or ^e*P* < 0.05 vs 10-14 years when compared with other disease duration periods and ^f*P* < 0.05 vs 6.4% or less which indicate significant differences. As shown in the figure (A), percentages of subjects with a score of 5 or higher for all symptoms except constipation in the disease duration period of 15-19 years were significantly higher than those in the 5-9 years and 10-14 years periods; *P* = 0.048 for heartburn, *P* = 0.024 for gastralgia, *P* = 0.008 for fullness and *P* = 0.03 for diarrhea. Likewise, percentages of subjects with a score of 5 or higher for heartburn and fullness in the disease duration period of 20 years or more were significantly higher than those in the 5-9 years and 10-14 years periods; *P* = 0.01 and *P* = 0.04 for heartburn and *P* = 0.02 for fullness. Furthermore, the percentage of subjects with a score of 5 or higher for fullness in the disease duration period of 10-14 yr was significantly higher than that in the 5-9 years period (*P* = 0.029). As shown in the figure (C), the percentage of subjects with a score of 5 or higher for fullness in those with HbA1c levels of 7.5% or above was significantly higher than that in those whose HbA1c was 6.4% or less (*P* = 0.030). BMI: Body mass index; HbA1c: Hemoglobin A1c.

Lower GI tract symptoms were found to be most frequent and cluster analysis demonstrated these symptoms to be related more to the affected region of the GI tract than to its functions, such as acid secretion and motility. In fact, a high concordance between constipation and diarrhea is unusual in general populations^[48]. Upper and lower GI symptoms reportedly overlap and early satiety more frequently

overlaps with constipation than diarrhea in general populations^[49,50].

This study has several limitations. First, it is difficult to determine the causes of symptoms with a cross-sectional study design. There is diagnostic uncertainty regarding the GI disorders studied, because the reporting of symptoms was mainly subjective. Endoscopy can be used to exclude organic diseases with

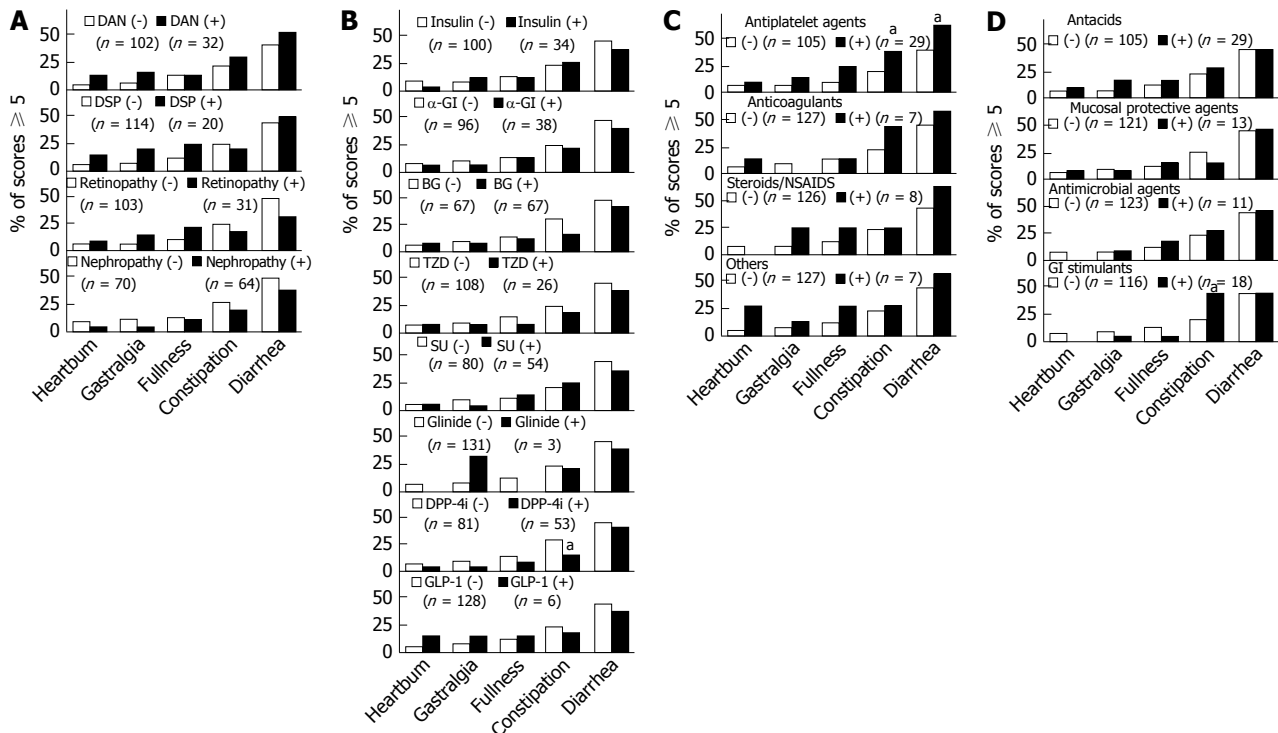


Figure 3 Relationships between various parameters and gastrointestinal symptoms involving each gastrointestinal region ("Heartburn" for esophagus, "Gastralgia" and "Postprandial Fullness" for upper abdomen, and "Constipation" and "Diarrhea" for lower abdomen). Bars: White indicates the absence and black the presence of each parameter. The percentages of subjects with a score of 5 or higher, for each GI region, considered to be sufficiently symptomatic to cause impaired quality of life, are shown. ^a $P < 0.05$, significant difference between presence and absence of each parameter, as analyzed by Fisher's exact test. A: Comparison between subjects with and without diabetic microangiopathy. The indicated percentages were obtained by dividing the number of subjects with a score of 5 or higher by the total number of patients in the same group (with or without diabetic microangiopathy); B: Comparisons between groups of patients with and without antidiabetic agent administration; C: Comparisons between groups of patients with and without antithrombotic or anti-inflammatory agents. 'Others' includes other antithrombotic agents such as prostaglandin E1 derivatives, prostaglandin I2 derivatives, or ethyl esters of eicosapentaenoic acid; D: Comparisons between groups of patients with and without GI agents. α GI: Alpha-glucosidase inhibitors; DAN: Diabetic autonomic neuropathy; DPP-4i: Dipeptidyl-peptidase 4 inhibitor; DSP: Distal symmetric polyneuropathy; GI: Gastrointestinal; GLP-1: Glucagon-like peptide-1; NSAIDs: Nonsteroidal anti-inflammatory drugs; SU: Sulphonyl urea; TZD: Thiazolidinedione.

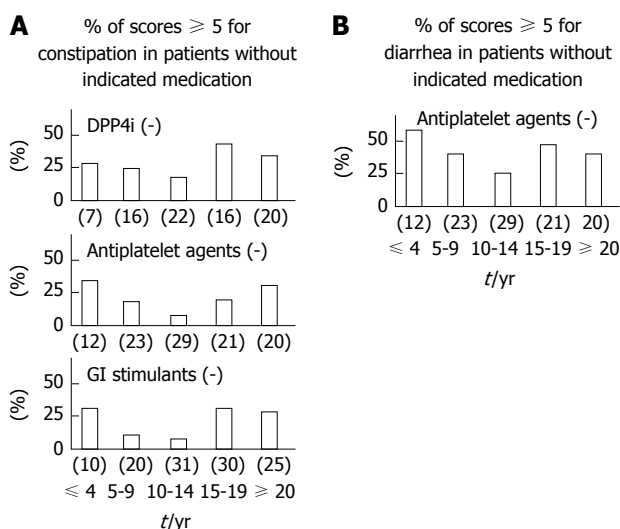


Figure 4 Relationships between various medications, which demonstrated significance as shown in Figure 3, and diabetes duration in terms of the presence of related gastrointestinal symptoms. White bars show the percentages of subjects with a score of 5 or higher for the indicated GI symptoms (A: Constipation, B: Diarrhea) without use of the indicated medications. Numbers under each bar within brackets are the number of patients not using the indicated medications. DPP-4i: Dipeptidyl-peptidase 4 inhibitor; GI: Gastrointestinal.

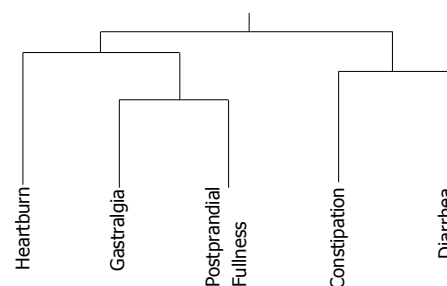


Figure 5 Cluster analysis was conducted for various gastrointestinal symptoms. The treeplot shows relationships among the five symptoms. Lengths of branches between symptoms indicate symptom similarity.

symptoms similar to those of diabetic GI disorders (e.g., inflammatory bowel disease, colonic polyposis, and celiac disease)^[51]. Few therapeutic or diagnostic approaches are available to assess the GI disorders possibly associated with diabetes. Organic diseases not requiring surgery and functional disorders could not be precisely distinguished because we did not perform upper and lower GI endoscopy or gastric emptying scintigraphy at the time of this study, since it was retrospective. Our results might have low sensitivity due to the small sample sizes in some groups.

In conclusion, our study highlights the importance of questioning patients about QOL impairment due to abdominal symptoms, especially lower GI symptoms in the early period and both lower and upper GI symptoms in the late period of diabetes. The heterogeneous nature of the underlying pathophysiological mechanisms underlying GI symptoms, including medication usage, should be taken into consideration when managing patients with T2DM.

COMMENTS

Background

Type 2 diabetes mellitus (T2DM) incidence is dramatically increasing, and both micro- and macrovascular complications of diabetes lead to various symptoms and impair quality of life (QOL) in T2DM patients. Among these symptoms, those affecting the gastrointestinal (GI) tract are frequent in T2DM patients. GI disorders in diabetes have been poorly characterized. Hyperglycemia-associated neuropathy was considered to be related to GI symptoms, probably via oxidative stress and inflammation. Levels of enteric hormones are altered in patients with DM, affecting both GI motility and neural fiber growth. Furthermore, smooth muscle cells, interstitial cells of Cajal, gut microbiota and intestinal stem cells may also be altered in DM and these changes might be related to GI symptoms. The relevance of these mechanisms in human GI symptoms affecting T2DM patients have yet to be clarified. The Izumo Scale, a validated and useful tool for QOL assessment, is a scale for assessing GI symptoms. Applying this scale, we investigated actual GI symptoms employing a questionnaire and determined whether the symptoms identified are related to the clinical profiles of patients with T2DM.

Research frontiers

This study highlights the importance of not underestimating gastrointestinal symptoms and of questioning patients about QOL impairment due to abdominal symptoms, especially in both the early period and after a diabetes duration of 10 or even 15 years.

Innovations and breakthroughs

The novel finding of this study is that symptom frequencies differed among disease durations and according to affected gastrointestinal regions. Lower abdominal symptoms not only manifested during the late but also in the early stage of diabetes when there were no organ complications related to this disease. This results are apparently inconsistent with the previously suggested mechanisms of GI symptoms related to DM, especially diabetic neuropathy which develops with longstanding diabetes.

Applications

This study highlights the importance of questioning patients about QOL impairment due to abdominal symptoms, especially lower GI symptoms in the early period and both lower and upper GI symptoms in the late period of diabetes, while underscoring the need for systematic and comprehensive research focusing on GI conditions in patients with T2DM.

Terminology

Izumo Scale: This self-administered questionnaire designed to assess the effects of abdominal symptoms on QOL includes 15 items in five domains with three items in each domain: heartburn, gastralgia, postprandial fullness, constipation, and diarrhea. This scale was developed and validated by Furuta *et al.* Each question is rated on a 6-point Likert scale from 0 to 5, with higher values indicating more severe symptoms.

Peer-review

Considering the paucity of data available in the literature on this topic, this study may reinforce the clinical relevance of assessing intestinal disorders in diabetes and increase the interest of the scientific community in this topic. Overall, this report describes relevant novel findings that may be further addressed and

confirmed with detailed experimental studies.

REFERENCES

- 1 **Guariguata L**, Whiting DR, Hambleton I, Beagley J, Linnenkamp U, Shaw JE. Global estimates of diabetes prevalence for 2013 and projections for 2035. *Diabetes Res Clin Pract* 2014; **103**: 137-149 [PMID: 24630390 DOI: 10.1016/j.diabres.2013.11.002]
- 2 **Talley NJ**, Young L, Bytzer P, Hammer J, Leemon M, Jones M, Horowitz M. Impact of chronic gastrointestinal symptoms in diabetes mellitus on health-related quality of life. *Am J Gastroenterol* 2001; **96**: 71-76 [PMID: 11197290 DOI: 10.1111/j.1572-0241.2001.03350.x]
- 3 **Phillips LK**, Deane AM, Jones KL, Rayner CK, Horowitz M. Gastric emptying and glycaemia in health and diabetes mellitus. *Nat Rev Endocrinol* 2015; **11**: 112-128 [PMID: 25421372 DOI: 10.1038/nrendo.2014.202]
- 4 **Bharucha AE**, Batey-Schaefer B, Cleary PA, Murray JA, Cowie C, Lorenzi G, Driscoll M, Harth J, Larkin M, Christofi M, Bayless M, Wimmergren N, Herman W, Whitehouse F, Jones K, Kruger D, Martin C, Ziegler G, Zinsmeister AR, Nathan DM. Delayed Gastric Emptying Is Associated With Early and Long-term Hyperglycemia in Type 1 Diabetes Mellitus. *Gastroenterology* 2015; **149**: 330-339 [PMID: 25980755 DOI: 10.1053/j.gastro.2015.05.007]
- 5 **Ko GT**, Chan WB, Chan JC, Tsang LW, Cockram CS. Gastrointestinal symptoms in Chinese patients with Type 2 diabetes mellitus. *Diabet Med* 1999; **16**: 670-674 [PMID: 10477212]
- 6 **Azpiroz F**, Malagelada C. Diabetic neuropathy in the gut: pathogenesis and diagnosis. *Diabetologia* 2016; **59**: 404-408 [PMID: 26643877 DOI: 10.1007/s00125-015-3831-1]
- 7 **Vinik AI**, Maser RE, Mitchell BD, Freeman R. Diabetic autonomic neuropathy. *Diabetes Care* 2003; **26**: 1553-1579 [PMID: 12716821]
- 8 **Horowitz M**, Harding PE, Maddox AF, Wishart JM, Akkermans LM, Chatterton BE, Shearman DJ. Gastric and oesophageal emptying in patients with type 2 (non-insulin-dependent) diabetes mellitus. *Diabetologia* 1989; **32**: 151-159 [PMID: 2753246]
- 9 **Chang J**, Rayner CK, Jones KL, Horowitz M. Diabetic gastroparesis-backwards and forwards. *J Gastroenterol Hepatol* 2011; **26** Suppl 1: 46-57 [PMID: 21199514 DOI: 10.1111/j.1440-1746.2010.06573.x]
- 10 **Akyüz F**, Uyanikoglu A, Ermis F, Arıcı S, Akyüz Ü, Baran B, Pınarbası B, Gul N. Gastroesophageal reflux in asymptomatic obese subjects: An esophageal impedance-pH study. *World J Gastroenterol* 2015; **21**: 3030-3034 [PMID: 25780302 DOI: 10.3748/wjg.v21.i10.3030]
- 11 **Matsuura B**, Nunoi H, Miyake T, Hiasa Y, Onji M. Obesity and gastrointestinal liver disorders in Japan. *J Gastroenterol Hepatol* 2013; **28** Suppl 4: 48-53 [PMID: 24251704 DOI: 10.1111/jgh.12238]
- 12 **Tarnawski AS**, Ahluwalia A, Jones MK. Increased susceptibility of aging gastric mucosa to injury: the mechanisms and clinical implications. *World J Gastroenterol* 2014; **20**: 4467-4482 [PMID: 24782600 DOI: 10.3748/wjg.v20.i16.4467]
- 13 **D'Addio F**, Fiorina P. Type 1 Diabetes and Dysfunctional Intestinal Homeostasis. *Trends Endocrinol Metab* 2016; **27**: 493-503 [PMID: 27185326 DOI: 10.1016/j.tem.2016.04.005]
- 14 **Latorre R**, Sternini C, De Giorgio R, Greenwood-Van Meerveld B. Enteroendocrine cells: a review of their role in brain-gut communication. *Neurogastroenterol Motil* 2016; **28**: 620-630 [PMID: 26691223 DOI: 10.1111/nmo.12754]
- 15 **Mawe GM**, Hoffman JM. Serotonin signalling in the gut-functions, dysfunctions and therapeutic targets. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 473-486 [PMID: 23797870 DOI: 10.1038/nrgastro.2013.105]
- 16 **Adeghate E**, Ponery AS, Sharma AK, El-Sharkawy T, Donáth T. Diabetes mellitus is associated with a decrease in vasoactive intestinal polypeptide content of gastrointestinal tract of rat. *Arch Physiol Biochem* 2001; **109**: 246-251 [PMID: 11880929 DOI: 10.1076/apab.109.3.246.11587]
- 17 **Ördög T**. Interstitial cells of Cajal in diabetic gastroenteropathy.

- Neurogastroenterol Motil* 2008; **20**: 8-18 [PMID: 18173559 DOI: 10.1111/j.1365-2982.2007.01056.x]
- 18 **Vaarala O**, Atkinson MA, Neu J. The “perfect storm” for type 1 diabetes: the complex interplay between intestinal microbiota, gut permeability, and mucosal immunity. *Diabetes* 2008; **57**: 2555-2562 [PMID: 18820210 DOI: 10.2337/db08-0331]
 - 19 **D’Addio F**, La Rosa S, Maestroni A, Jung P, Orsenigo E, Ben Nasr M, Tezza S, Bassi R, Finzi G, Marando A, Vergani A, Frego R, Albarello L, Andolfo A, Manuguerra R, Viale E, Staudacher C, Corradi D, Battle E, Breault D, Secchi A, Folli F, Fiorina P. Circulating IGF-I and IGFBP3 Levels Control Human Colonic Stem Cell Function and Are Disrupted in Diabetic Enteropathy. *Cell Stem Cell* 2015; **17**: 486-498 [PMID: 26431183 DOI: 10.1016/j.stem.2015.07.010]
 - 20 **Brownlee M**. Glycation products and the pathogenesis of diabetic complications. *Diabetes Care* 1992; **15**: 1835-1843 [PMID: 1464241]
 - 21 **Giacco F**, Brownlee M. Oxidative stress and diabetic complications. *Circ Res* 2010; **107**: 1058-1070 [PMID: 21030723 DOI: 10.1161/CIRCRESAHA.110.223545]
 - 22 **Forbes JM**, Cooper ME. Mechanisms of diabetic complications. *Physiol Rev* 2013; **93**: 137-188 [PMID: 23303908 DOI: 10.1152/physrev.00045.2011]
 - 23 **Bytzer P**, Talley NJ, Hammer J, Young LJ, Jones MP, Horowitz M. GI symptoms in diabetes mellitus are associated with both poor glycemic control and diabetic complications. *Am J Gastroenterol* 2002; **97**: 604-611 [PMID: 11922554 DOI: 10.1111/j.1572-0241.2002.05537.x]
 - 24 **Mohamed Q**, Gillies MC, Wong TY. Management of diabetic retinopathy: a systematic review. *JAMA* 2007; **298**: 902-916 [PMID: 17712074 DOI: 10.1001/jama.298.8.902]
 - 25 **Tanaka K**, Hara S, Hattori M, Sakai K, Onishi Y, Yoshida Y, Kawazu S, Kushiya A. Role of elevated serum uric acid levels at the onset of overt nephropathy in the risk for renal function decline in patients with type 2 diabetes. *J Diabetes Investig* 2015; **6**: 98-104 [PMID: 25621139 DOI: 10.1111/jdi.12243]
 - 26 **Furuta K**, Ishihara S, Sato S, Miyake T, Ishimura N, Koshino K, Tobita H, Moriyama I, Amano Y, Adachi K, Ohta A, Kinoshita Y. [Development and verification of the Izumo Scale, new questionnaire for quality of life assessment of patients with gastrointestinal symptoms]. *Nihon Shokakibyo Gakkai Zasshi* 2009; **106**: 1478-1487 [PMID: 19834295]
 - 27 **Kakuta E**, Yamashita N, Katsube T, Kushiya Y, Suetsugu H, Furuta K, Kinoshita Y. Abdominal symptom-related QOL in individuals visiting an outpatient clinic and those attending an annual health check. *Intern Med* 2011; **50**: 1517-1522 [PMID: 21804275]
 - 28 **Kinoshita Y**, Chiba T. Characteristics of Japanese patients with chronic gastritis and comparison with functional dyspepsia defined by ROME III criteria: based on the large-scale survey, FUTURE study. *Intern Med* 2011; **50**: 2269-2276 [PMID: 22001450]
 - 29 **Yoshioka T**, Okimoto N, Okamoto K, Sakai A. A comparative study of the effects of daily minodronate and weekly alendronate on upper gastrointestinal symptoms, bone resorption, and back pain in postmenopausal osteoporosis patients. *J Bone Miner Metab* 2013; **31**: 153-160 [PMID: 23076293 DOI: 10.1007/s00774-012-0393-x]
 - 30 **Okimoto E**, Ishimura N, Morito Y, Mikami H, Shimura S, Uno G, Tamagawa Y, Aimi M, Oshima N, Kawashima K, Kazumori H, Sato S, Ishihara S, Kinoshita Y. Prevalence of gastroesophageal reflux disease in children, adults, and elderly in the same community. *J Gastroenterol Hepatol* 2015; **30**: 1140-1146 [PMID: 25611309 DOI: 10.1111/jgh.12899]
 - 31 **Kinoshita Y**, Chiba T. Therapeutic effects of famotidine on chronic symptomatic gastritis: subgroup analysis from FUTURE study. *J Gastroenterol* 2012; **47**: 377-386 [PMID: 22183857 DOI: 10.1007/s00535-011-0503-x]
 - 32 **Lovell RM**, Ford AC. Global prevalence of and risk factors for irritable bowel syndrome: a meta-analysis. *Clin Gastroenterol Hepatol* 2012; **10**: 712-721.e4 [PMID: 22426087 DOI: 10.1016/j.cgh.2012.02.029]
 - 33 **Galmiche JP**, Clouse RE, Bálint A, Cook IJ, Kahrilas PJ, Paterson WG, Smout AJ. Functional esophageal disorders. *Gastroenterology* 2006; **130**: 1459-1465 [PMID: 16678559 DOI: 10.1053/j.gastro.2005.08.060]
 - 34 **Tack J**, Talley NJ, Camilleri M, Holtmann G, Hu P, Malagelada JR, Stanghellini V. Functional gastroduodenal disorders. *Gastroenterology* 2006; **130**: 1466-1479 [PMID: 16678560 DOI: 10.1053/j.gastro.2005.11.059]
 - 35 **Longstreth GF**, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorders. *Gastroenterology* 2006; **130**: 1480-1491 [PMID: 16678561 DOI: 10.1053/j.gastro.2005.11.061]
 - 36 **Mjörnheim AC**, Finizia C, Blohmé G, Attvall S, Lundell L, Ruth M. Gastrointestinal symptoms in type 1 diabetic patients, as compared to a general population. A questionnaire-based study. *Digestion* 2003; **68**: 102-108 [PMID: 14593236]
 - 37 **Bytzer P**, Talley NJ, Leemon M, Young LJ, Jones MP, Horowitz M. Prevalence of gastrointestinal symptoms associated with diabetes mellitus: a population-based survey of 15,000 adults. *Arch Intern Med* 2001; **161**: 1989-1996 [PMID: 11525701]
 - 38 **Kushiya A**, Yoshida Y, Kikuchi T, Suzawa N, Yamamoto M, Tanaka K, Okayasu M, Tahara T, Takao T, Onishi Y, Kawazu S. Twenty-year trend of increasing obesity in young patients with poorly controlled type 2 diabetes at first diagnosis in urban Japan. *J Diabetes Investig* 2013; **4**: 540-545 [PMID: 24843707 DOI: 10.1111/jdi.12090]
 - 39 **Clouse RE**, Lustman PJ. Gastrointestinal symptoms in diabetic patients: lack of association with neuropathy. *Am J Gastroenterol* 1989; **84**: 868-872 [PMID: 2756978]
 - 40 **Williams-Herman D**, Engel SS, Round E, Johnson J, Golm GT, Guo H, Musser BJ, Davies MJ, Kaufman KD, Goldstein BJ. Safety and tolerability of sitagliptin in clinical studies: a pooled analysis of data from 10,246 patients with type 2 diabetes. *BMC Endocr Disord* 2010; **10**: 7 [PMID: 20412573 DOI: 10.1186/1472-6823-10-7]
 - 41 **Otsuki H**, Kosaka T, Nakamura K, Shimomura F, Kuwahara Y, Tsukamoto T. Safety and efficacy of teneligliptin: a novel DPP-4 inhibitor for hemodialysis patients with type 2 diabetes. *Int Urol Nephrol* 2014; **46**: 427-432 [PMID: 24014134 DOI: 10.1007/s11255-013-0552-6]
 - 42 **Kobayashi M**, Yamazaki K, Hirao K, Oishi M, Kanatsuka A, Yamauchi M, Takagi H, Kawai K. The status of diabetes control and antidiabetic drug therapy in Japan--a cross-sectional survey of 17,000 patients with diabetes mellitus (JDDM 1). *Diabetes Res Clin Pract* 2006; **73**: 198-204 [PMID: 16621117 DOI: 10.1016/j.diabetes.2006.01.013]
 - 43 **Kosachunhanun N**, Benjasuratwong Y, Mongkolsomlit S, Rawdaree P, Plengvidhya N, Leelawatana R, Bunnag P, Pratiapanawatr T, Krittiyawong S, Suwanwalaikorn S, Deerochanawong C, Chetthakul T, Ngarmukos C, Komoltri C. Thailand diabetes registry project: glycemic control in Thai type 2 diabetes and its relation to hypoglycemic agent usage. *J Med Assoc Thai* 2006; **89** Suppl 1: S66-S71 [PMID: 17715836]
 - 44 **Scheiman JM**. Prevention of damage induced by aspirin in the GI tract. *Best Pract Res Clin Gastroenterol* 2012; **26**: 153-162 [PMID: 22542153 DOI: 10.1016/j.bpg.2012.01.005]
 - 45 **Nadatani Y**, Watanabe T, Tanigawa T, Sogawa M, Yamagami H, Shiba M, Watanabe K, Tominaga K, Fujiwara Y, Yoshiyama M, Arakawa T. Incidence and risk factors of gastrointestinal bleeding in patients on low-dose aspirin therapy after percutaneous coronary intervention in Japan. *Scand J Gastroenterol* 2013; **48**: 320-325 [PMID: 23298342 DOI: 10.3109/00365521.2012.758771]
 - 46 **Dobretsov M**, Romanovsky D, Stimers JR. Early diabetic neuropathy: triggers and mechanisms. *World J Gastroenterol* 2007; **13**: 175-191 [PMID: 17226897 DOI: 10.3748/wjg.v13.i2.175]
 - 47 **Ekberg K**, Brismar T, Johansson BL, Jonsson B, Lindström P, Wahren J. Amelioration of sensory nerve dysfunction by C-Peptide in patients with type 1 diabetes. *Diabetes* 2003; **52**: 536-541 [PMID: 12540632]
 - 48 **Talley NJ**, Holtmann G, Agréus L, Jones M. Gastrointestinal

- symptoms and subjects cluster into distinct upper and lower groupings in the community: a four nations study. *Am J Gastroenterol* 2000; **95**: 1439-1447 [PMID: 10894576 DOI: 10.1111/j.1572-0241.2000.02075.x]
- 49 **Talley NJ**, Dennis EH, Schettler-Duncan VA, Lacy BE, Olden KW, Crowell MD. Overlapping upper and lower gastrointestinal symptoms in irritable bowel syndrome patients with constipation or diarrhea. *Am J Gastroenterol* 2003; **98**: 2454-2459 [PMID: 14638348 DOI: 10.1111/j.1572-0241.2003.07699.x]
- 50 **de Bortoli N**, Martinucci I, Bellini M, Savarino E, Savarino V, Blandizzi C, Marchi S. Overlap of functional heartburn and gastroesophageal reflux disease with irritable bowel syndrome. *World J Gastroenterol* 2013; **19**: 5787-5797 [PMID: 24124323 DOI: 10.3748/wjg.v19.i35.5787]
- 51 **Yantiss RK**, Odze RD. Optimal approach to obtaining mucosal biopsies for assessment of inflammatory disorders of the gastrointestinal tract. *Am J Gastroenterol* 2009; **104**: 774-783 [PMID: 19209164 DOI: 10.1038/ajg.2008.108]

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Observational Study

Serum angiotensin-converting enzyme level for evaluating significant fibrosis in chronic hepatitis B

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Abstract

AIM

To evaluate the diagnostic performance of angiotensin-converting enzyme (ACE) on significant liver fibrosis in patients with chronic hepatitis B (CHB).

METHODS

In total, 100 patients with CHB who underwent liver biopsy in our hospital were enrolled, and 70 patients except for 30 patients with hypertension, fatty liver or habitual alcoholic consumption were analyzed. We compared histological liver fibrosis and serum ACE levels and evaluated the predictive potential to diagnose significant liver fibrosis by comparison with several biochemical marker-based indexes such as the aspartate aminotransferase (AST)-to-platelet ratio index (APRI), the fibrosis index based on four factors (FIB-4), the Mac-2 binding protein glycosylation isomer (M2BPGi) level and the number of platelets (Plt).

RESULTS

Serum ACE levels showed moderately positive correlation with liver fibrotic stages ($R^2 = 0.181$). Patients with significant, advanced fibrosis and cirrhosis (F2-4) had significantly higher serum ACE levels than those with

early-stage fibrosis and cirrhosis (F0-1). For significant fibrosis (\geq F2), the 12.8 U/L cut-off value of ACE showed 91.7% sensitivity and 75.0% specificity. The receiver-operating characteristic (ROC) curves analysis revealed that the area under the curve (AUC) value of ACE was 0.871, which was higher than that of APRI, FIB-4, M2BPGi and Plt.

CONCLUSION

The serum ACE level could be a novel noninvasive, easy, accurate, and inexpensive marker of significant fibrosis stage in patients with CHB.

Key words: Angiotensin-converting enzyme; Hepatitis B virus; Liver fibrosis; Noninvasive fibrosis marker; Aspartate aminotransferase-to-platelet ratio index; Fibrosis index based on four factors; Mac-2 binding protein glycosylation isomer

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Core tip: Liver fibrosis is one of key factors to determine therapeutic intervention for patients with chronic hepatitis B (CHB). However, the noninvasive prediction of CHB-related liver fibrosis is difficult. Angiotensin-converting enzyme (ACE) is reportedly involved in liver fibrogenesis. In this paper, we demonstrate that serum ACE levels are elevated in patients with CHB and show the predictive potential to diagnose significant fibrosis (\geq F2), which is the therapeutically adapted stage, with higher accuracy as compared with other fibrotic markers including APRI, FIB-4, M2BPGi and Plt. The serum ACE level could be a novel noninvasive marker of significant fibrosis stage in CHB.

Noguchi R, Kaji K, Namisaki T, Moriya K, Kitade M, Takeda K, Kawaratani H, Okura Y, Aihara Y, Furukawa M, Mitoro A, Yoshiji H. Serum angiotensin-converting enzyme level for evaluating significant fibrosis in chronic hepatitis B. *World J Gastroenterol* 2017; 23(36): 6705-6714 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6705.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6705>

INTRODUCTION

Hepatitis B virus (HBV) annually affects 350-400 million people and causes 1 million deaths worldwide^[1,2]. Chronic HBV infection results in a risk of progressive liver fibrosis, leading to cirrhosis with decreased liver reserve and hepatocellular carcinoma (HCC)^[3]. The annual incidence of HCC is reported to be 10%-17% in HBV-induced liver cirrhosis, and the fibrotic status stepwisely increases the risk of HCC, as in case of chronic infection of hepatitis C virus (HCV)^[4]. Therefore, an early assessment of liver fibrosis is required for not only the prevention of disease progression but also the judgment of therapeutic intervention in patients

with HBV infection. Liver biopsy is currently the gold standard for estimating fibrosis progression, although frequent application of this procedure is limited because of sampling error, invasiveness, and some other complications^[5]. These reasons have prompted many investigators to explore noninvasive predictors for liver fibrosis. Of several noninvasive methods, two tests, namely the aspartate aminotransferase (AST)-to-platelet index (APRI) and the fibrosis index based on four factors (FIB-4), are utilized for relatively accurate detection of liver fibrosis especially induced by HCV infection^[6-8]. At present, the accuracy of these models has been externally validated in patients with chronic viral hepatitis, nonalcoholic fatty liver disease (NAFLD), primary biliary cholangitis (PBC), and primary sclerosing cholangitis (PSC). With regard to chronic hepatitis B (CHB), a recent meta-analysis suggested that APRI and FIB-4 could identify CHB-related fibrosis with moderate sensitivity and accuracy^[9]. However, more methods are required to circumstantially classify liver fibrosis.

The renin-angiotensin-aldosterone system (RAAS) is a key mediator in the regulation of arterial blood pressure and body fluid homeostasis^[10]. RAAS also reportedly plays an important role in the hemodynamics of several organs^[11,12]. RAAS is frequently activated in patients with chronic liver diseases such as cirrhosis^[13,14]. Angiotensin-I-converting enzyme (ACE), a central component of RAAS, converts the inactive decapeptide angiotensin I (AT-I) into the octapeptide angiotensin II (AT-II), which shows many physiological activities, including vascular hormonal secretion and tissue growth^[15]. AT-II has been considered to be a potential mediator of portal hypertension because its plasma level is markedly increased in patients with cirrhosis and its administration induces the elevation of portal pressure^[13,16-18]. AT-II is pathologically recognized to induce the contractility and proliferation of hepatic stellate cells (HSCs), which play a pivotal role in the progression of liver fibrosis^[19-21]. The aim of the present study was to evaluate the behavior of circulating ACE in patients with CHB and assess the relationship between CHB-related liver fibrosis and serum levels of ACE.

MATERIALS AND METHODS

Patients

In total, 100 patients who were diagnosed with serologically and histologically confirmed CHB at Nara Medical University between 2013 and 2015 were enrolled. Chronic HBV infection was diagnosed in patients according to the following criteria: (1) detectable hepatitis B surface antigen (HBsAg) for \geq 6 mo; and (2) serum HBV-DNA \geq 1.3 log IU/mL. The status of other HBV markers such as HB envelope antigen (HBeAg), anti-HBe, and anti-HB core (HBc) IgG was not considered as a criterion for the current assessment. All the patients who fulfilled these criteria underwent

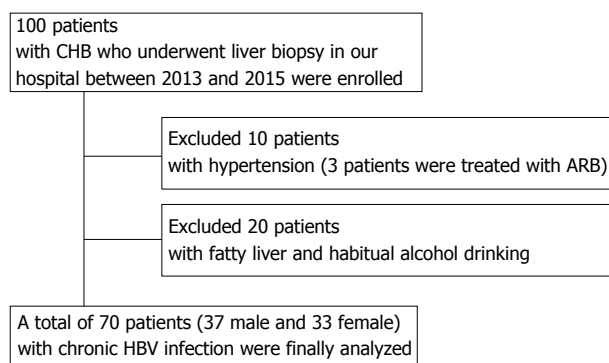


Figure 1 The selection of the study population. 70 patients except for 10 patients with hypertension and 20 patients with fatty liver or habitual alcoholic consumption were finally analyzed.

routine liver biopsies before therapeutic intervention. All pathological specimens were evaluated by at least two experienced pathologists. The degree of hepatic fibrosis was assessed and graded according to the METAVIR score for chronic hepatitis graded 0-4: F0, no fibrosis; F1, portal fibrosis without septa; F2, portal fibrosis with few septa; F3, numerous septa without cirrhosis; and F4, cirrhosis^[22,23]. Hepatic steatosis is histologically defined with more than 5% of accumulated lipid droplets in the hepatic parenchyma, and habitual alcohol consumption is defined with a daily ethanol intake of > 20 g (female) or > 30 g (male). Patients were excluded if they had other concomitant chronic liver diseases, including chronic hepatitis C (CHC), autoimmune hepatitis (AIH), PBC, PSC, hemochromatosis, or Wilson's disease as well as cancer, severe cardiopulmonary or renal diseases, diabetes mellitus (DM), sarcoidosis, dysthyroidism, or a previous history of liver transplantation. 10 patients treated with antihypertensive agents were also excluded regardless of the type of drug because high blood pressure was considered to affect serum ACE levels (Figure 1). The study was conducted in accordance with the standards of the Helsinki Declaration, and written informed consents were provided by all the study subjects. The protocols used were approved by the Ethics Committee of Nara Medical University (Nara, Japan; Approval number 1077) and other facilities.

Laboratory analysis and measurement of angiotensin-converting enzyme levels

Serum samples from all patients were collected when liver biopsy was performed and were stored and used for the present study. The following laboratory parameters were routinely measured: complete blood count, AST, alanine aminotransferase (ALT), and albumin (Alb). To assess liver fibrosis, we evaluated four serum fibrotic markers, namely, hyaluronic acid, type 4 collagen 7S, type 3 procollagen-N-peptide (P-III-P), and Mac-2-binding protein glycosylation isomer (M2BPGi), as well as the number of platelets (Plt). APRI and FIB-4 were also used as noninvasive tests for the assessment of liver fibrosis. In these tests, evaluation

was performed using the following formula: $APRI = [(AST \text{ of the sample/reference AST}) \times 100] / \text{platelets}$; $FIB-4 = (\text{age} \times AST) / [(\text{platelets}) \times (ALT)^{1/2}]$. Serum ACE levels were measured using Kasahara's colorimetry-based methods^[24]. A serum ACE level between 8.3 and 21.4 IU/L was considered to be within the normal range according to the manufacturer's instruction.

Statistical analysis

Continuous variables of patients with discordance and those without discordance were compared using independent *t*-tests or Mann-Whitney *U* tests, as appropriate. The χ^2 or Fisher's exact tests were used for categorical variables. Area under the receiver operating characteristic (AUROC) curves and other statistical analyses were performed using R software as described previously^[25].

RESULTS

Serum ACE levels in patients with fatty liver and alcohol abuse

ACE activity is susceptible to the presence of fatty liver and/or habitual alcoholic consumption (FL/AL), as reported previously^[26]. Thus, we initially evaluated the serum ACE activity in patients histologically diagnosed with hepatic steatosis to exclude patients with FL/AL from the current subjects. Histological analysis demonstrated that 20 patients showed hepatic steatosis. Of these patients, 8 patients habitually drank alcohol. Similar to several reports, serum ACE levels were significantly higher in patients with CHB with FL/AL than in those without FL/AL (Figure 2A), although other fibrotic markers, including APRI, FIB-4, and M2BPGi, were not affected by the coexistence of FL/AL (Figure 2B-2D). Interestingly, a fibrosis stage-matched comparison showed that this difference in serum ACE levels was prevalently observed in patients with early-stage (F0 and F1) liver fibrosis (Figure 2E). Consequently, 70 patients with CHB and without hypertension and/or FL/AL were included as subjects in the present study.

Characteristic features of patients

The demographic and clinicopathological characteristics of the patients in the final analysis are presented in Table 1. In total, 70 patients with CHB without FL/AL (37 males and 33 females) with a median age of 48.6 ± 14.1 years were included in the present study. The fibrosis stages were F0, F1, F2, F3 and F4 in nine (12.8%), 25 (35.7%), 17 (24.3%), 13 (18.6%), and six (8.6%) patients, respectively. All the patients in F4 were classified as Child-Pugh A. There were no differences in serum Alb levels and HBV-DNA and HBsAg values among each stage of fibrosis.

Elevated serum ACE level above F2 stage in liver fibrosis

In patients with CHB, the mean serum ACE level was

Table 1 The clinicopathological characteristics of the patients with hepatitis B

Variables	Patients with HB (n = 70)
Sex (males/females)	37/33
Age	48.6 ± 14.1
Fibrosis stage (F0/F1/F2/F3/F4)	9/25/17/13/6
Platelet (× 10 ⁴ μL) (F0/F1/F2/F3/F4)	17.9 ± 5.0 (23.2 ± 4.7/18.4 ± 3.6/18.7 ± 3.8/15.1 ± 5.4/18.9 ± 3.3)
Alb (g/dL)	4.1 ± 0.4
AST (IU/L)	35.8 ± 26.7
ALT (IU/L)	42.6 ± 39.4
HBV DNA (Log IU/mL)	4.04 ± 2.24
HBsAg (IU/mL)	22697.1 ± 52927.9
Hyaluronic acid (ng/mL)	62.4 ± 92.1
Type 4 collagen 7S (ng/mL)	4.1 ± 2.0
Serum ACE (U/L) (F0/F1/F2/F3/F4)	14.1 ± 5.1 (10.8 ± 2.3/11.1 ± 4.7/17.9 ± 4.4/6.4 ± 3.9/15.0 ± 2.9)
APRI (F0/F1/F2/F3/F4)	1.0 ± 1.3 (0.3 ± 0.1/0.7 ± 0.5/0.9 ± 0.6/2.3 ± 2.1/1.0 ± 0.9)
FIB4 (F0/F1/F2/F3/F4)	2.0 ± 1.9 (1.3 ± 0.6/1.6 ± 1.0/1.3 ± 0.8/2.8 ± 1.7/1.7 ± 0.7)
M2BPGi (COI) (F0/F1/F2/F3/F4)	1.2 ± 1.1 (0.9 ± 0.9/0.9 ± 0.6/1.1 ± 1.0/2.2 ± 1.8/1.3 ± 0.6)

HBV: Hepatitis B virus; HBsAg: Hepatitis B surface antigen; Alb: Albumin; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; ACE: Angiotensin-I-converting enzyme; APRI: Aspartate aminotransferase-to-platelet index; FIB4: Fibrosis index based on four factors; M2BPGi: Mac-2-binding protein glycosylation isomer.

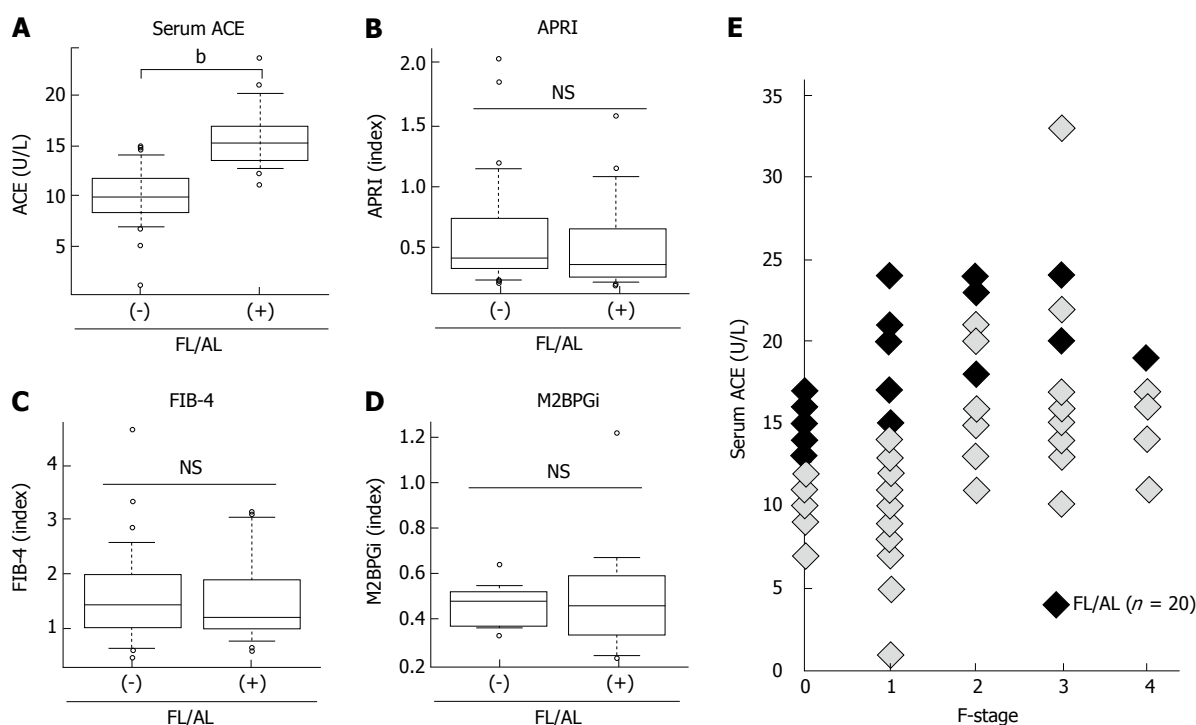


Figure 2 Serum levels of angiotensin-converting enzyme and fibrotic markers in patients with fatty liver and/or habitual alcoholic drinking. A: Serum angiotensin-converting enzyme (ACE) level; B: Aspartate aminotransferase to platelet index (APRI); C: Fibrosis index based on the four factors (FIB-4); D: Serum Mac-2 binding protein glycosylation isomer (M2BPGi) level in chronic hepatitis B patients with and without fatty liver and/or habitual alcoholic drinking (FL/AL). Serum ACE levels were significantly higher in the patients with FL/AL than those without FL/AL; E: Fibrosis stage-matched comparison showed that this difference in serum ACE levels between with and without FL/AL was prevalently observed in early stages (F0 and F1) of liver fibrosis. Data are means ± SD, ^b*P* < 0.01.

14.1 ± 5.1 U/L and serum ACE levels at F0, F1, F2, F3, and F4 were 10.8 ± 2.3 U/L, 11.1 ± 4.7 U/L, 17.9 ± 4.4 U/L, 16.4 ± 3.9 U/L, and 15.0 ± 2.9 U/L, respectively. Pearson's correlation coefficient showed moderately positive correlation between serum ACE levels and fibrotic stages ($R^2 = 0.181$), and Mann-Whitney *U* tests demonstrated that patients with significant, advanced and cirrhotic stages (F2-4) showed markedly higher serum ACE levels than those with early-stage

fibrosis and cirrhosis (F0-1) (Figure 3A). To validate the diagnostic performance of the serum ACE level in predicting significant liver fibrosis, we next performed receiver-operating characteristic (ROC) curve analysis. For significant fibrosis ($\geq F2$), ROC curve analysis revealed that the optimal ACE level cut-off point was 12.8 U/L (sensitivity, 91.7%; specificity, 75.0%; PPV, 80.5%; NPV, 89.7%; accuracy, 84.3%), and the area under the curve (AUC) value for significant fibrosis (\geq

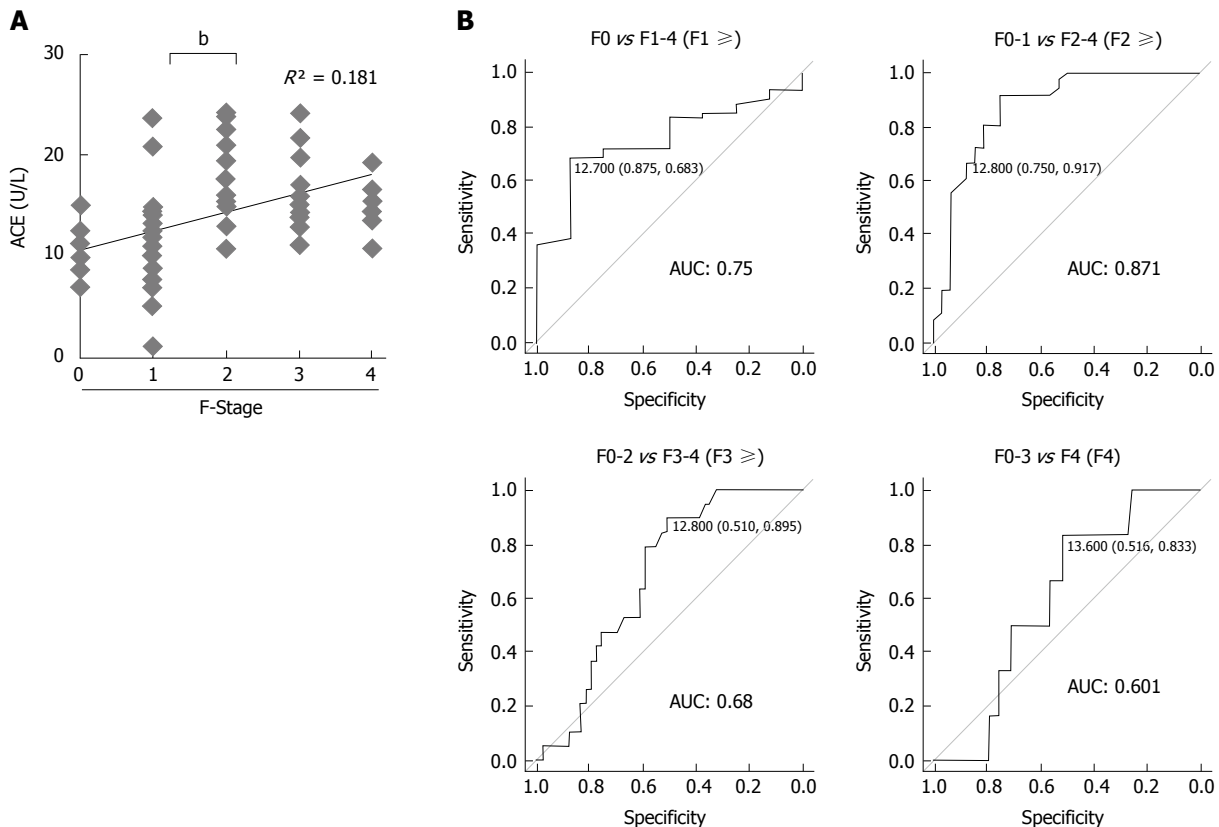


Figure 3 Serum angiotensin-converting enzyme levels and liver fibrosis development in patients with chronic hepatitis B. A: Correlation between serum angiotensin-converting enzyme (ACE) level and liver fibrosis stage (F-Stage) ($R = 0.42$, $R^2 = 0.181$); B: Receiver operating characteristic curve analysis and area under curve (AUC) value for diagnostic performance of serum ACE level for predicting each stage of liver fibrosis. The optimal ACE level cut-off point for significant fibrosis was 12.8 U/L. Data are means \pm SD, $^bP < 0.01$.

F2) was 0.871 (Figure 3B). Meanwhile, the AUC values to predict mild ($\geq F1$), advanced ($\geq F3$) and cirrhotic (F4) stage of fibrosis were 0.75 (sensitivity, 68.3%; specificity, 87.5%), 0.68 (sensitivity, 89.5%; specificity, 51.0%) and 0.601 (sensitivity, 83.3%; specificity, 51.6%), respectively which were remarkably lower than significant fibrosis (Figure 3B). These findings suggest that serum ACE level manifests its efficient performance in diagnosing significant fibrosis ($\geq F2$).

Diagnostic performance in other markers to predict significant stage liver fibrosis

Next, to evaluate predictive potential for significant fibrosis in other markers, we performed similar analysis for well-known fibrotic parameters, including APRI, FIB-4, M2BPGi and Plt. Unlike the ACE level, there was a significant difference below and above the F3 stage in these parameters, APRI (F2, 0.86 ± 0.6 vs F3, 2.27 ± 2.0 ; $P < 0.05$), FIB-4 (F2, 1.34 ± 0.6 vs F3, 2.76 ± 1.7 ; $P < 0.05$), M2BPGi (F2, 1.1 ± 1.0 vs F3, 2.2 ± 1.8 ; $P < 0.05$) and Plt (F2, 18.7 ± 3.8 vs F3, $15.1 \pm 5.4 \times 10^4/\mu\text{L}$; $P = 0.0616$) (Figure 4A-D). Similarly, the established fibrosis markers such as hyaluronic acid, type 4 collagen 7S, and P-III-P indicated an advanced fibrosis stage ($\geq F3$) (Figure 4E-G). Furthermore, we assessed the diagnostic performance of APRI, FIB-4, M2BPGi and Plt to predict significant fibrosis ($\geq F2$)

by ROC curve analysis. For significant fibrosis ($\geq F2$), the APRI cut-off point was 0.57 (sensitivity, 83.3%; specificity, 69.7%; PPV, 76.0%; NPV, 71.1%; accuracy, 73%), FIB-4 cut-off point was 2.23 (sensitivity, 40.0%; specificity, 84.8%; PPV, 70.6%; NPV, 39.1%; accuracy, 47.6%), M2BPGi cut-off point was 0.81 (sensitivity, 50.0%; specificity, 48.9%; PPV, 69.7%; NPV, 29.4%; accuracy, 49.3%), and Plt cut-off point was $17.8 \times 10^4/\mu\text{L}$ (sensitivity, 68.8%; specificity, 71.9%; PPV, 33.3%; NPV, 72.4%; accuracy, 50.8%) (Supplementary Figure 1). The AUC value of serum ACE (0.871) was higher than those of APRI (0.83, $P = 0.224$), FIB-4 (0.641, $P = 0.0012$), M2BPGi (0.717, $P = 0.0239$), and Plt (0.70, $P = 0.016$) (Figure 5A-D), indicating that compared with other noninvasive markers, the serum ACE level is distinctively capable of the enclosure of significant liver fibrosis ($\geq F2$) in patients with CHB.

APRI is a beneficial marker in predicting advanced liver fibrosis with the highest accuracy

As the next step to closely diagnose significant fibrosis in patients with CHB, patients with $> F2$ fibrosis need to be further classified into significant (F2) and advanced (F3) fibrosis. Therefore, we compared the AUC values to predict advanced fibrosis stages ($\geq F3$) among APRI, FIB-4, M2BPGi, and Plt to evaluate diagnostic

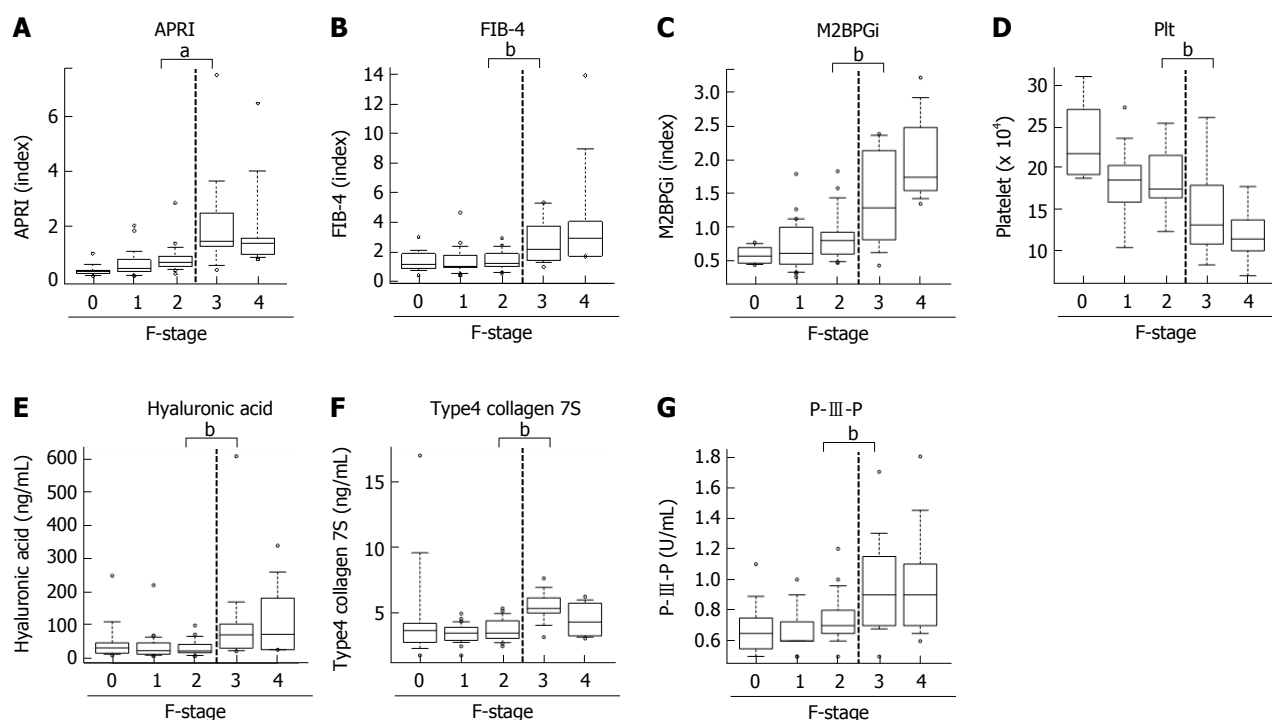


Figure 4 Serum levels of other markers and liver fibrosis in patients with chronic hepatitis B. A: Aspartate aminotransferase to platelet index (APRI); B: Fibrosis index based on the four factors (FIB-4); C: Serum levels of Mac-2 binding protein glycosylation isomer (M2BPGi); D: The number of platelets (Plt); E: Serum hyaluronic acid level; F: Type4 collagen 7S; and G: P-III-P levels in patients with each liver fibrotic stage (F-Stage). Data are means \pm SD, ^a $P < 0.05$, ^b $P < 0.01$.

performance of these parameters for advanced fibrosis. The ROC curve analysis demonstrated that the optimum cut-off points for advanced fibrosis (\geq F3) are APRI; 1.27 (sensitivity, 75%; specificity, 91.5%), FIB-4; 2.51 (sensitivity, 56.2%; specificity, 93.6%), M2BPGi; 0.82 (sensitivity, 87.5%; specificity, 64.0%), and Plt; $15.0 \times 10^4/\mu\text{L}$ (sensitivity, 70.6%; specificity, 87.2%) (Supplementary Figure 1). The AUC value is the highest in APRI (0.879) as compared with FIB-4 (0.803), M2BPGi (0.791), and Plt (0.813).

DISCUSSION

Identifying the degree of liver fibrosis is a clinical requisite for the treatment of chronic liver diseases regardless of the etiology because it plays a key role in predicting therapy responses and long-term outcomes of patients. In the case of CHB, nucleos(t)ide analogs (NAs) inhibit HBV-DNA replication and reduce the serum HBV level to achieve therapeutic improvement. Meanwhile, NAs do not play any role in the complete elimination of HBV and do not provide HBsAg clearance or persistent HBeAg seroconversion. Moreover, NAs frequently cause drug resistance and relapse after the termination of therapy. These pharmacological properties of NAs require long-term administration for patients to avoid the risk of liver decompensation due to cirrhosis and HCC progression, suggesting that the ideal time to start the therapy should be carefully decided considering the fibrotic stage as well as HBV-DNA and ALT levels which are extensively recognized

as guideline for treatment. At present, APRI and FIB-4 indexes are widely known as noninvasive predictors to evaluate liver fibrosis^[7,8]. Xiao *et al.*^[9] systematically reviewed the performance of two indexes in HBV-associated liver fibrosis. The total AUC values of APRI/FIB-4 for the diagnosis of significant fibrosis, advanced fibrosis, and cirrhosis were 0.7407, 0.7844 and 0.7347/0.8165, and 0.7268 and 0.8448, respectively. These data indicate that APRI and FIB-4 can identify CHB-related fibrosis with moderate sensitivity and accuracy. However, additional markers are required for diagnosis with higher accuracy and sensitivity. Recent evidences demonstrated that several serum markers are possibly beneficial for prediction of earlier fibrosis. Oztas *et al.*^[27] reported that soluble ST2, a receptor for the Th2 cytokine IL-33, could be used for differentiating significant fibrosis from mild fibrosis in CHB patients, and Deng *et al.*^[27] showed that serum complement 5a concentration significantly decreased in severe HBV fibrosis stages and earlier cirrhosis^[28].

Our results show that serum ACE levels were markedly higher in patients with CHB who were histologically diagnosed with significant, advanced fibrosis and cirrhosis than in those with early fibrosis, while other predictors distinguished advanced fibrosis and cirrhosis (\geq F3) but not significant fibrosis (F2). These results indicate that the serum ACE level has a distinctive potential to enclose significant fibrosis (\geq F2), which is the therapeutically adapted stage for NAs^[23]. As confirmed by AUC, the diagnostic value of serum ACE for detecting significant fibrosis above F2 was 87.1%,

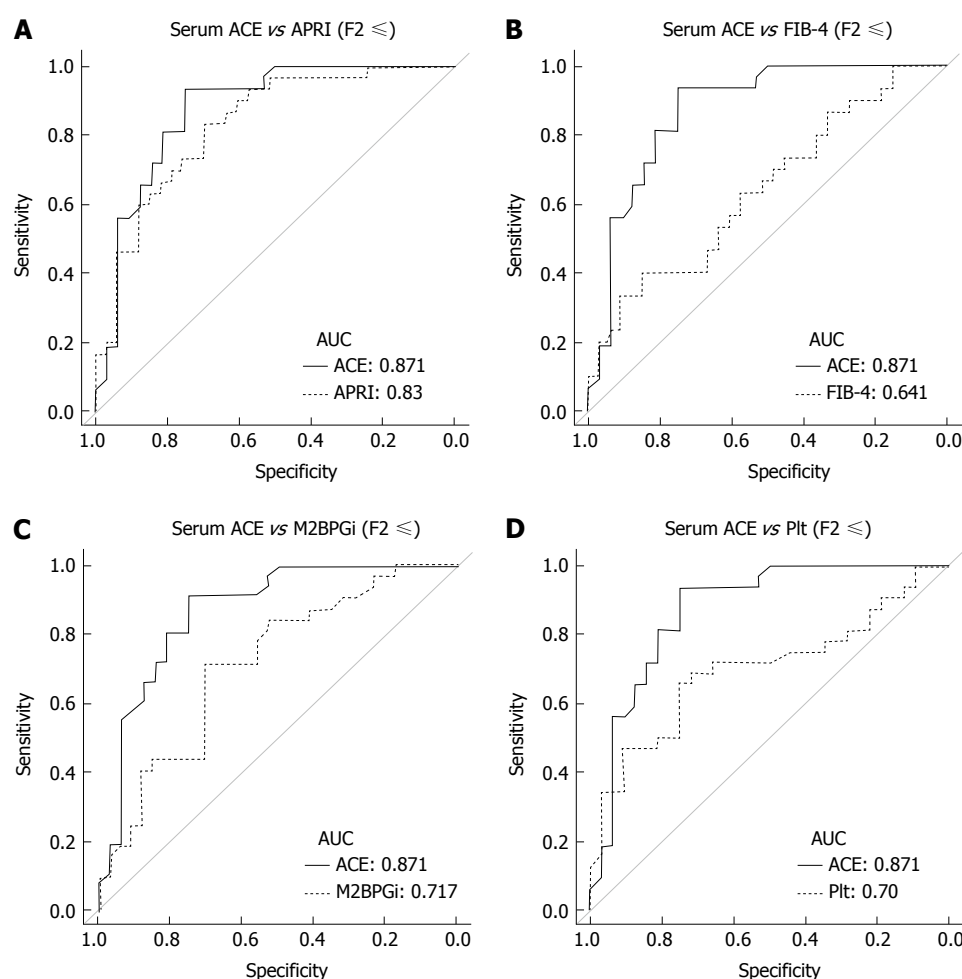


Figure 5 Receiver operating characteristic curve analysis for diagnostic performance for predicting significant liver fibrosis. Compared to other markers, the area under curve (AUC) value in serum angiotensin-converting enzyme (ACE) level, 0.871, was higher than that in A: Aspartate aminotransferase to platelet index (APRI), 0.83 ($P = 0.224$); B: Fibrosis index based on the four factors (FIB-4), 0.641 ($P = 0.0012$); C: Serum levels of Mac-2 binding protein glycosylation isomer (M2BPGi), 0.717 ($P = 0.0239$); D: The number of platelets (Plt), 0.70 ($P = 0.016$).

which was higher than other that of fibrotic markers, including APRI, FIB-4, M2BPGi and Plt.

As described in previous reports, AT-II, which is produced by ACE converted from AT-I, plays an important role in liver fibrosis development^[21,29,30]. AT-II induces HSC proliferation, upregulates transforming growth factor- β and collagen-I gene expression, and promotes extracellular matrix synthesis^[19,20]. Additionally, previous animal study demonstrated that ACE gene was up-regulated in the bile duct ligation-induced fibrotic liver^[31]. Recent evidence also suggested that upregulation of hepatic ACE is accelerated particularly during fibrogenesis and is dampened in the developed fibrosis state^[32]. Correspondingly, our results showed that serum ACE level was increased with moderately positive correlation with liver fibrosis development, and reached a threshold in the significant fibrosis (F2) (Figure 3A). These findings suggest that the suppression of RAAS is an inevitable strategy for preventing liver fibrosis progression. Our recent animal studies have revealed that ACE inhibitor (ACE-I) and AT receptor blocker (ARB) show significant antifibrotic effects on experimental liver fibrosis along with the

suppression of activated HSC^[33-36]. A clinical study also demonstrated that the group of patients with HCV treated using angiotensin-blocking agents exhibited lesser fibrosis than those without hypertension^[37]. Another report showed that RAAS blocker-treated hypertensive patients with NAFLD prevalently had a mild degree of liver fibrosis^[38]. Liver fibrogenesis progresses through multiple processes that are dependent on the etiology. Therefore, the clinical findings in patients with HCV and NAFLD may provide a basis for the assessment of the antifibrotic properties of RAAS blockers in HBV. A previous report actually showed that circulating ACE levels were elevated in patients with chronic HBV and more prominent elevation was observed in patients with advanced fibrosis^[39].

Other reports have validated whether the serum ACE level has the clinical potential to predict liver fibrosis induced by other chronic liver diseases. Efe *et al*^[40] suggested that ACE sustains hepatic fibrogenesis in AIH. They demonstrated that serum ACE levels increased for each fibrosis score in 73 patients with AIH, and AUC values of serum ACE for the diagnosis of significant fibrosis ($\geq F2$), advanced fibrosis

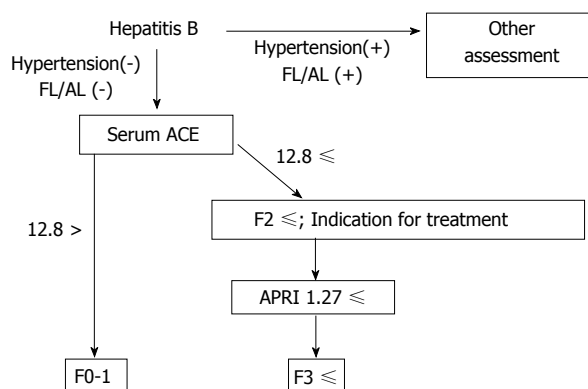


Figure 6 Schematic algorithm for noninvasive diagnosis of chronic hepatitis B. ACE: Angiotensin-converting enzyme; FL: Fatty liver; AL: Habitual alcoholic drinking; APRI: Aspartate aminotransferase to platelet index.

($\geq F3$), and cirrhosis (F4) were 0.89, 0.91, and 0.95, respectively. The diagnostic performance for HCV-related fibrosis is controversial. Raslan *et al.*^[41] showed that ACE polymorphism was associated with the progression of hepatic fibrosis in chronic HCV infection. In contrast, Forrest *et al.*^[42] reported that no association was identified between four RAAS polymorphisms and fibrosis in chronic HCV infection. Our other assessment also suggested that serum ACE levels in patients with CHC did not correlate with the fibrosis stage (data not shown). This discrepancy may be explained by the interaction between HCV infection and steatosis. HCV infection induces metabolic abnormalities such as insulin resistance, leading to hepatic steatosis^[43,44]. Interestingly, our data show that serum ACE levels were elevated in patients with hepatic steatosis (Figure 2). The evidence indicates that hepatic steatosis induced by HCV infection may destabilize serum ACE levels.

In our results, we also found that APRI, FIB-4, M2BPGi and Plt might be good predictive markers for advanced fibrosis in patients with CHB, and APRI showed a potential with the highest accuracy in these parameters. Based on these data, we can propose a strategy to noninvasively evaluate CHB-related liver fibrosis in combination with serum ACE and APRI (Figure 6). This algorithm may be utilized to enclose significant fibrosis, particularly in patients with CHB without fatty liver and/or habitual alcoholic consumption.

Our data have some limitations such as the lack of clinicopathological and prognostic data and a small sample size. Moreover, our study was not performed prospectively. Sequential measurements of serum ACE are required in longitudinal studies to assess the change in the serum ACE level associated with the degree of fibrosis. In addition, the optimum cut-off value for significant fibrosis of 12.8 U/L deviates from the value of 52.5 U/L in the previous study because a different method is used to measure the serum ACE level^[39,40]. Therefore, comparison with data from other reports to validate the integration is difficult.

In conclusion, we indicated that the serum ACE

level is a beneficial noninvasive marker to evaluate significant fibrosis and to further determine whether therapeutic intervention with NAs is necessary for patients with CHB. Higher accuracy will be expected by combination with other markers, including APRI, FIB4, M2BPGi and Plt. Moreover, our results suggest that blockade of RAAS is an effective new therapeutic strategy against CHB-related fibrosis.

COMMENTS

Background

Chronic hepatitis B virus (HBV) infection results in a risk of progressive liver fibrosis, leading to cirrhosis with decreased liver reserve and hepatocellular carcinoma. Therefore, an early assessment of liver fibrosis is required for not only the prevention of disease progression but also the judgment of therapeutic intervention. Currently, several noninvasive markers are explored for estimating early stage of HBV-related fibrosis alternatively to liver biopsy. However, it is required to identify both easier and more beneficial methods.

Research frontiers

Angiotensin-I-converting enzyme (ACE), a central component of renin-angiotensin-aldosterone system, plays a key role in the progression of liver fibrosis, however, its diagnostic performance to evaluate HBV-related earlier fibrosis.

Innovations and breakthroughs

The present study indicates that the patients with significant, advanced fibrosis and cirrhosis (F2-4) have significantly higher serum ACE levels than those with early-stage fibrosis and cirrhosis (F0-1), and serum ACE shows higher accuracy than other markers including APRI, FIB-4, M2BPGi and Plt in diagnostic performance to differentiate significant fibrosis (F2) from mild fibrosis (F1).

Applications

The serum ACE level is a beneficial noninvasive marker to evaluate significant fibrosis and to further determine whether therapeutic intervention is necessary for patients with chronic HBV.

Terminology

ACE converts the inactive decapeptide angiotensin I (AT-I) into the octapeptide angiotensin II (AT-II), which shows many physiological activities, including vascular hormonal secretion and tissue growth. AT-II has been pathologically recognized to induce the contractility and proliferation of hepatic stellate cells, which play a pivotal role in the progression of liver fibrosis.

Peer-review

The authors report an interesting approach about the diagnostic value evaluation of serum ACE for the detection of earlier HBV-related fibrosis. The sensitivity of the test discriminating F0-F1 from F2-F4 proved to be higher than that of other tests. Overall, the study is well planned and results are presented clearly.

REFERENCES

- Schweitzer A, Horn J, Mikolajczyk RT, Krause G, Ott JJ. Estimations of worldwide prevalence of chronic hepatitis B virus infection: a systematic review of data published between 1965 and 2013. *Lancet* 2015; **386**: 1546-1555 [PMID: 26231459 DOI: 10.1016/S0140-6736(15)61412-X]
- Stanaway JD, Flaxman AD, Naghavi M, Fitzmaurice C, Vos T, Abubakar I, Abu-Raddad LJ, Assadi R, Bhala N, Cowie B, Forouzanfar MH, Groeger J, Hanafiah KM, Jacobsen KH, James SL, MacLachlan J, Malekzadeh R, Martin NK, Mokdad AA, Mokdad AH, Murray CJL, Plass D, Rana S, Rein DB, Richardus JH, Sanabria J, Saylan M, Shahraz S, So S, Vlassov VV, Weiderpass E, Wiersma ST, Younis M, Yu C, El Sayed Zaki

- M, Cooke GS. The global burden of viral hepatitis from 1990 to 2013: findings from the Global Burden of Disease Study 2013. *Lancet* 2016; **388**: 1081-1088 [PMID: 27394647 DOI: 10.1016/S0140-6736(16)30579-7]
- 3 **de Martel C**, Maucourt-Boulch D, Plummer M, Franceschi S. World-wide relative contribution of hepatitis B and C viruses in hepatocellular carcinoma. *Hepatology* 2015; **62**: 1190-1200 [PMID: 26146815 DOI: 10.1002/hep.27969]
- 4 **Burns GS**, Thompson AJ. Viral hepatitis B: clinical and epidemiological characteristics. *Cold Spring Harb Perspect Med* 2014; **4**: a024935 [PMID: 25359547 DOI: 10.1101/cshperspect.a024935]
- 5 **Bravo AA**, Sheth SG, Chopra S. Liver biopsy. *N Engl J Med* 2001; **344**: 495-500 [PMID: 11172192 DOI: 10.1056/NEJM200102153440706]
- 6 **Lin ZH**, Xin YN, Dong QJ, Wang Q, Jiang XJ, Zhan SH, Sun Y, Xuan SY. Performance of the aspartate aminotransferase-to-platelet ratio index for the staging of hepatitis C-related fibrosis: an updated meta-analysis. *Hepatology* 2011; **53**: 726-736 [PMID: 21319189 DOI: 10.1002/hep.24105]
- 7 **Vallet-Pichard A**, Mallet V, Nalpas B, Verkarre V, Nalpas A, Dhalluin-Venier V, Fontaine H, Pol S. FIB-4: an inexpensive and accurate marker of fibrosis in HCV infection. comparison with liver biopsy and fibrotest. *Hepatology* 2007; **46**: 32-36 [PMID: 17567829 DOI: 10.1002/hep.21669]
- 8 **Shaheen AA**, Myers RP. Diagnostic accuracy of the aspartate aminotransferase-to-platelet ratio index for the prediction of hepatitis C-related fibrosis: a systematic review. *Hepatology* 2007; **46**: 912-921 [PMID: 17705266 DOI: 10.1002/hep.21835]
- 9 **Xiao G**, Yang J, Yan L. Comparison of diagnostic accuracy of aspartate aminotransferase to platelet ratio index and fibrosis-4 index for detecting liver fibrosis in adult patients with chronic hepatitis B virus infection: a systemic review and meta-analysis. *Hepatology* 2015; **61**: 292-302 [PMID: 25132233 DOI: 10.1002/hep.27382]
- 10 **Cooper ME**. The role of the renin-angiotensin-aldosterone system in diabetes and its vascular complications. *Am J Hypertens* 2004; **17**: 16S-20S; quiz A2-4 [PMID: 15539106 DOI: 10.1016/j.amjhyper.2004.08.004]
- 11 **Atlas SA**. The renin-angiotensin aldosterone system: pathophysiological role and pharmacologic inhibition. *J Manag Care Pharm* 2007; **13**: 9-20 [PMID: 17970613 DOI: 10.18553/jmcp.2007.13.s8-b.9]
- 12 **Wolf G**. Novel aspects of the renin-angiotensin-aldosterone-system. *Front Biosci* 2008; **13**: 4993-5005 [PMID: 18508564]
- 13 **Helmy A**, Jalan R, Newby DE, Hayes PC, Webb DJ. Role of angiotensin II in regulation of basal and sympathetically stimulated vascular tone in early and advanced cirrhosis. *Gastroenterology* 2000; **118**: 565-572 [PMID: 10702208]
- 14 **Munshi MK**, Uddin MN, Glaser SS. The role of the renin-angiotensin system in liver fibrosis. *Exp Biol Med* (Maywood) 2011; **236**: 557-566 [PMID: 21508249 DOI: 10.1258/ebm.2011.010375]
- 15 **Stergiou GS**, Skeva II. Renin-angiotensin system blockade at the level of the angiotensin converting enzyme or the angiotensin type-1 receptor: similarities and differences. *Curr Top Med Chem* 2004; **4**: 473-481 [PMID: 14965313]
- 16 **Lugo-Baruqui A**, Muñoz-Valle JF, Arévalo-Gallegos S, Armendáriz-Borunda J. Role of angiotensin II in liver fibrosis-induced portal hypertension and therapeutic implications. *Hepatol Res* 2010; **40**: 95-104 [PMID: 19737316 DOI: 10.1111/j.1872-034X.2009.00581.x]
- 17 **Beyazit Y**, Ibis M, Purnak T, Turhan T, Kekilli M, Kurt M, Sayilir A, Onal IK, Turhan N, Tas A, Köklü S, Haznedaroglu IC. Elevated levels of circulating angiotensin converting enzyme in patients with hepatoportal sclerosis. *Dig Dis Sci* 2011; **56**: 2160-2165 [PMID: 21290180 DOI: 10.1007/s10620-011-1580-7]
- 18 **Klein S**, Rick J, Lehmann J, Schierwagen R, Schierwagen IG, Verbeke L, Hittatiya K, Uschner FE, Manekeller S, Strassburg CP, Wagner KU, Sayeski PP, Wolf D, Laleman W, Sauerbruch T, Trebicka J. Janus-kinase-2 relates directly to portal hypertension and to complications in rodent and human cirrhosis. *Gut* 2017; **66**: 145-155 [PMID: 26385087 DOI: 10.1136/gutjnl-2015-309600]
- 19 **Batalier R**, Ginès P, Nicolás JM, Görbig MN, Garcia-Ramallo E, Gasull X, Bosch J, Arroyo V, Rodés J. Angiotensin II induces contraction and proliferation of human hepatic stellate cells. *Gastroenterology* 2000; **118**: 1149-1156 [PMID: 10833490]
- 20 **Yoshiji H**, Kuriyama S, Yoshii J, Ikenaka Y, Noguchi R, Nakatani T, Tsujinoue H, Fukui H. Angiotensin-II type 1 receptor interaction is a major regulator for liver fibrosis development in rats. *Hepatology* 2001; **34**: 745-750 [PMID: 11584371 DOI: 10.1053/jhep.2001.28231]
- 21 **Yoshiji H**, Kuriyama S, Noguchi R, Ikenaka Y, Kitade M, Kaji K, Yoshii J, Yanase K, Yamazaki M, Asada K, Tsujimoto T, Akahane T, Uemura M, Fukui H. Angiotensin-II and vascular endothelial growth factor interaction plays an important role in rat liver fibrosis development. *Hepatol Res* 2006; **36**: 124-129 [PMID: 16919500 DOI: 10.1016/j.hepres.2006.07.003]
- 22 Intraobserver and interobserver variations in liver biopsy interpretation in patients with chronic hepatitis C. The French METAVIR Cooperative Study Group. *Hepatology* 1994; **20**: 15-20 [PMID: 8020885]
- 23 **Drafting Committee for Hepatitis Management Guidelines and the Japan Society of Hepatology**. JSH Guidelines for the Management of Hepatitis B Virus Infection. *Hepatol Res* 2014; **44** Suppl S1: 1-58 [PMID: 24397839 DOI: 10.1111/hepr.12269]
- 24 **Kasahara Y**, Ashihara Y. Colorimetry of angiotensin-I converting enzyme activity in serum. *Clin Chem* 1981; **27**: 1922-1925 [PMID: 6271420]
- 25 **Kanda Y**. Investigation of the freely available easy-to-use software 'EZ' for medical statistics. *Bone Marrow Transplant* 2013; **48**: 452-458 [PMID: 23208313 DOI: 10.1038/bmt.2012.244]
- 26 **Saiki A**, Ohira M, Endo K, Koide N, Oyama T, Murano T, Watanabe H, Miyashita Y, Shirai K. Circulating angiotensin II is associated with body fat accumulation and insulin resistance in obese subjects with type 2 diabetes mellitus. *Metabolism* 2009; **58**: 708-713 [PMID: 19375596 DOI: 10.1016/j.metabol.2009.01.013]
- 27 **Oztas E**, Kuzu UB, Zengin NI, Kalkan IH, Onder FO, Yildiz H, Celik HT, Akdogan M, Kilic MY, Koksas AS, Odemis B, Suna N, Kayacetin E. Can Serum ST2 Levels Be Used as a Marker of Fibrosis in Chronic Hepatitis B Infection? *Medicine* (Baltimore) 2015; **94**: e1889 [PMID: 26632683 DOI: 10.1097/MD1889]
- 28 **Deng Y**, Zhao H, Zhou J, Yan L, Wang G; China HepB-Related Fibrosis Assessment Research Group. Complement 5a is an indicator of significant fibrosis and earlier cirrhosis in patients chronically infected with hepatitis B virus. *Infection* 2017; **45**: 75-81 [PMID: 27605044 DOI: 10.1007/s15010-016-0942-7]
- 29 **Yoshiji H**. Anti-fibrotic therapy: Are matrix metalloproteinases friends or foes? *Hepatol Res* 2009; **39**: 748-750 [PMID: 19709329 DOI: 10.1111/j.1872-034X.2009.00573.x]
- 30 **Kaji K**, Yoshiji H, Ikenaka Y, Noguchi R, Aihara Y, Shirai Y, Douhara A, Fukui H. Possible involvement of angiogenesis in chronic liver diseases: interaction among renin-angiotensin-aldosterone system, insulin resistance and oxidative stress. *Curr Med Chem* 2012; **19**: 1889-1898 [PMID: 22376037]
- 31 **Paizis G**, Cooper ME, Schembri JM, Tikellis C, Burrell LM, Angus PW. Up-regulation of components of the renin-angiotensin system in the bile duct-ligated rat liver. *Gastroenterology* 2002; **123**: 1667-1676 [PMID: 12404241]
- 32 **Warner FJ**, Lubel JS, McCaughan GW, Angus PW. Liver fibrosis: a balance of ACEs? *Clin Sci* (Lond) 2007; **113**: 109-118 [PMID: 17600527 DOI: 10.1042/CS20070026]
- 33 **Yoshiji H**, Kuriyama S, Fukui H. Blockade of renin-angiotensin system in antifibrotic therapy. *J Gastroenterol Hepatol* 2007; **22** Suppl 1: S93-S95 [PMID: 17567477 DOI: 10.1111/j.1440-1746.2006.04663.x]
- 34 **Kaji K**, Yoshiji H, Kitade M, Ikenaka Y, Noguchi R, Shirai Y, Aihara Y, Namisaki T, Yoshii J, Yanase K, Tsujimoto T, Kawaratani H, Fukui H. Combination treatment of angiotensin II type I receptor blocker and new oral iron chelator attenuates progression of nonalcoholic steatohepatitis in rats. *Am J Physiol Gastrointest Liver Physiol* 2011; **300**: G1094-G1104 [PMID: 21372165 DOI: 10.1152/ajpgi.00365.2010]

- 35 **Noguchi R**, Yoshiji H, Ikenaka Y, Kaji K, Aihara Y, Shirai Y, Namisaki T, Kitade M, Douhara A, Moriya K, Fukui H. Dual blockade of angiotensin-II and aldosterone suppresses the progression of a non-diabetic rat model of steatohepatitis. *Hepatol Res* 2013; **43**: 765-774 [PMID: 23163573 DOI: 10.1111/hepr.12008]
- 36 **Okura Y**, Namisaki T, Moriya K, Kitade M, Takeda K, Kaji K, Noguchi R, Nishimura N, Seki K, Kawaratani H, Takaya H, Sato S, Sawada Y, Shimozato N, Furukawa M, Nakanishi K, Saikawa S, Kubo T, Asada K, Yoshiji H. Combined treatment with dipeptidyl peptidase-4 inhibitor (sitagliptin) and angiotensin-II type 1 receptor blocker (losartan) suppresses progression in a non-diabetic rat model of steatohepatitis. *Hepatol Res* 2016; Epub ahead of print [PMID: 28029729 DOI: 10.1111/hepr.12860]
- 37 **Colmenero J**, Bataller R, Sancho-Bru P, Domínguez M, Moreno M, Forns X, Bruguera M, Arroyo V, Brenner DA, Ginès P. Effects of losartan on hepatic expression of nonphagocytic NADPH oxidase and fibrogenic genes in patients with chronic hepatitis C. *Am J Physiol Gastrointest Liver Physiol* 2009; **297**: G726-G734 [PMID: 19628656 DOI: 10.1152/ajpgi.00162.2009]
- 38 **Goh GB**, Pagadala MR, Dasarathy J, Unalp-Arida A, Sargent R, Hawkins C, Sourianarayanan A, Khiyami A, Yerian L, Pai R, McCullough AJ, Dasarathy S. Renin-angiotensin system and fibrosis in non-alcoholic fatty liver disease. *Liver Int* 2015; **35**: 979-985 [PMID: 24905085 DOI: 10.1111/liv.12611]
- 39 **Purnak T**, Beyazit Y, Oztas E, Yesil Y, Efe C, Torun S, Celik T, Tenlik I, Kurt M, Ozaslan E. Serum angiotensin-converting enzyme level as a marker of fibrosis in patients with chronic hepatitis B. *J Renin Angiotensin Aldosterone Syst* 2012; **13**: 244-249 [PMID: 22277254 DOI: 10.1177/1470320311434241]
- 40 **Efe C**, Cengiz M, Kahramanoğlu-Aksoy E, Yılmaz B, Özşeker B, Beyazit Y, Tanoğlu A, Purnak T, Kav T, Turhan T, Ozenirler S, Ozaslan E, Wahlin S. Angiotensin-converting enzyme for noninvasive assessment of liver fibrosis in autoimmune hepatitis. *Eur J Gastroenterol Hepatol* 2015; **27**: 649-654 [PMID: 25860719 DOI: 10.1097/MEG.0000000000000355]
- 41 **Raslan HM**, Amr KS, Elhosary YA, Ezzat WM, Abdullah NA, El-Batae HE. Possible role of angiotensin-converting enzyme polymorphism on progression of hepatic fibrosis in chronic hepatitis C virus infection. *Trans R Soc Trop Med Hyg* 2011; **105**: 396-400 [PMID: 21546048 DOI: 10.1016/j.trstmh.2011.03.005]
- 42 **Forrest EH**, Thorburn D, Spence E, Oien KA, Inglis G, Smith CA, McCrudden EA, Fox R, Mills PR. Polymorphisms of the renin-angiotensin system and the severity of fibrosis in chronic hepatitis C virus infection. *J Viral Hepat* 2005; **12**: 519-524 [PMID: 16108768 DOI: 10.1111/j.1365-2893.2005.00630.x]
- 43 **Shintani Y**, Fujie H, Miyoshi H, Tsutsumi T, Tsukamoto K, Kimura S, Moriya K, Koike K. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. *Gastroenterology* 2004; **126**: 840-848 [PMID: 14988838]
- 44 **Fartoux L**, Poujol-Robert A, Guéchet J, Wendum D, Poupon R, Serfaty L. Insulin resistance is a cause of steatosis and fibrosis progression in chronic hepatitis C. *Gut* 2005; **54**: 1003-1008 [PMID: 15951550 DOI: 10.1136/gut.2004.050302]

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Observational Study

Wilson's disease in Lebanon and regional countries: Homozygosity and hepatic phenotype predominance

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Abstract

AIM

To determine the phenotypes and predominant disease-causing mutations in Lebanese patients with Wilson's disease, as compared to regional non-European data.

METHODS

The clinical profile of 36 patients diagnosed in Lebanon was studied and their mutations were determined by molecular testing. All patients underwent full physical exam, including ophthalmologic slit-lamp examination ultrasound imaging of the liver, as well as measurement of serum ceruloplasmin and 24-h urinary-Cu levels. In addition, genetic screening using PCR followed by sequencing to determine disease-causing mutations and polymorphisms in the *ATP7B* gene was carried on extracted DNA from patients and immediate family members. Our phenotypic-genotypic findings were then compared to reported mutations in Wilson's disease patients from regional Arab and non-European countries.

RESULTS

Patients belonged to extended consanguineous families. The majority were homozygous for the disease-causing mutation, with no predominant mutation identified.

The most common mutation, detected in 4 out of 13 families, involved the ATP hinge region and was present in patients from Lebanon, Egypt, Iran and Turkey. Otherwise, mutations in Lebanese patients and those of the region were scattered over 17 exons of *ATP7B*. While the homozygous exon 12 mutation Trp939Cys was only detected in patients from Lebanon but none from the regional countries, the worldwide common mutation H1069Q was not present in the Lebanese and was rare in the region. Pure hepatic phenotype was predominant in patients from both Lebanon and the region (25%-65%). Furthermore, the majority of patients, including those who were asymptomatic, had evidence of some hepatic dysfunction. Pure neurologic phenotype was rare.

CONCLUSION

Findings do not support presence of a founder effect. Clinical and genetic screening is recommended for family members with index patients and unexplained hepatic dysfunction.

Key words: Wilson Disease; Cu-metabolism; Phenotype; Genotype; ATP7B; Hepatic manifestations

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Core tip: We report on the genotype-phenotype of 36 Lebanese patients with Wilson's disease from 13 different families. The majority were homozygous for disease-causing mutations. The most common mutation worldwide, His1069Trp, was absent in our patients. The ATP hinge region may comprise a hot spot for mutations, as it was detected in 4 families. Hepatic phenotypes were predominant in both symptomatic and asymptomatic patients. Neurologic phenotypes were rare. Compared to findings reported in regional Arab and non-European countries, our results do not support a founder effect. Mutations are scattered over 17 exons, with no common or frequent mutation characterizing the region.

Barada K, El Haddad A, Katerji M, Jomaa M, Usta J. Wilson's disease in Lebanon and regional countries: Homozygosity and hepatic phenotype predominance. *World J Gastroenterol* 2017; 23(36): 6715-6725 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6715.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6715>

INTRODUCTION

Wilson's disease (WD) is an autosomal recessive disorder of copper (Cu) metabolism, resulting from defects in the *ATP7B* gene protein. It is characterized by failure of Cu incorporation into ceruloplasmin (Cp) and decreased biliary Cu excretion. As a consequence, Cu accumulates in various organs, primarily

liver and brain. The clinical presentations of WD are characterized by substantial diversity. Patients can present at any age in variable combinations of liver impairment, neurologic dysfunction and/or osseomuscular symptom. Hepatic manifestations include asymptomatic transaminitis, acute or chronic hepatitis, fulminant hepatic failure and/or cirrhosis, while neurologic symptoms vary from mild tremors, articulating problems, dysarthria, Parkinson-like features, seizures and cognitive dysfunction. Some patients have mixed hepato-neurologic presentation^[1]. Ophthalmologic involvement with Kaiser-Fleischer (KF) rings is common.

Traditionally, the diagnosis of WD is based on low serum-Cp (< 20 mg/dL), high 24-h urinary Cu and high hepatic Cu content (250 µg/g dry tissue)^[2,3]. Recent guidelines for the diagnosis of WD were published by the European Association for the Study of the Liver (EASL)^[4]. Nonetheless, the diagnosis of WD may be difficult based on clinical and laboratory criteria, and in some patients it is delayed, leading to detrimental consequences^[5]. This is why molecular testing and genotypic analysis may be warranted for confirming and/or supporting a diagnosis of WD, particularly in asymptomatic patients^[3].

More than 500 mutations have been identified in WD with a very high allelic heterogeneity. Most patients are compound heterozygous, rendering it difficult to ascribe a phenotype to a specific genotype^[6]. Furthermore, a large number of mutations are rare, making it impractical to screen populations for all WD-causing mutations^[7]. Some mutations, however, are relatively frequent and population-specific, like the p.His1069Gln on exon 14 in Northern and Eastern European patients^[8], the p.Arg778Leu and the p.Arg778Gly mutations on exon 8 among Chinese and Taiwanese patients respectively^[9], and the deletion in the 5' regulatory region in Sardinian patients^[10]. These findings facilitate molecular diagnosis based on patients' ethnic background. In the Arab World, consanguinity and marriage among individuals belonging to the same ethnic background is very common, thereby increasing the prevalence of genetic disorders, including WD^[11]. However, it is not known whether there is a predominant WD mutation in the Arab world, and if so what its phenotypic associations are.

In a cohort of Egyptian patients, genotypic and phenotypic profiles were described, but no prevalent mutation was identified^[12]. Moreover, previous reports from Lebanon on a limited number of families suggested an association of liver presentation with homozygous missense mutations: Gly691Arg and non-His1069Trp in exons 7 and 14 of the *ATP7B* gene respectively^[13,14]. Whether a specific WD mutation prevails in Lebanon is not known.

In this study, we described the spectrum and frequency of mutations and phenotypes in 36 Lebanese WD patients. We also conducted a comprehensive

literature search for regional studies on WD in Arab and non-European countries in the Middle East. In order to determine whether there is a frequent mutation characterizing the region, a comparative study was undertaken to identify common mutations in the region, and to compare them to our data. We also determined if common mutations in the region were associated with similar clinical phenotypes.

MATERIALS AND METHODS

A total of 36 patients (P₁-P₃₆) from 13 unrelated Lebanese families (U, Or, S, Ah, T, B, H, Ha, Is, Z, Ri, Sc and Gh) were enrolled in the study. Most patients were diagnosed at the American University of Beirut (AUB) Medical Center, a major tertiary referral center in Lebanon. All participating subjects were asked to sign a written consent form (Protocol No. BioCh.JU.01) that was approved by both the Institutional Review Board and Research Committee at AUB.

Clinical testing

Patients' data for evaluation included: history, date of birth, age of onset of symptoms, age at diagnosis, and findings from full physical exam, ophthalmologic slit-lamp examination, and biochemical tests, including liver function tests, serum-Cp and 24-h urinary-Cu levels. Abdominal ultrasound imaging of the liver was performed on all patients and, when necessary, brain magnetic resonance imaging was done. Phenotypic classifications were designated following Ferenci's classification as hepatic, neurologic, mixed or asymptomatic^[3]. Diagnosis was further established by computing total WD score developed at the 8th international meeting^[15]. Family members (siblings, parents) of all WD-confirmed index patients were also subjected to physical, biochemical and genotypic testing.

Genotypic screening

DNA screening for disease-causing mutations and single nucleotide polymorphisms was performed on all recruited subjects and their immediate family members. Extraction of DNA from blood samples followed by amplification (using PCR) of the 21 *ATP7B* exons was carried out as described^[6,14]. Amplified PCR products were purified, sequenced and compared to published normal sequences in the following databases: Blat at University of California Santa-Cruz, Genome Bio-informatics (<http://www.Genome.ucsc.edu/cgi-bin/hgBlat>) or Blast at National Center for Biotechnology Information (<http://www.ncbi.nih.gov/blast>).

WD in regional countries

After identifying the disease-causing mutations in our patients, we compared them to reported mutations in WD patients from regional Arab and Non-European countries. A comprehensive literature search of

PubMed and Medline, as well as of the University of Alberta database (<http://www.wilsonsdisease.med.ualberta.ca/database.asp>), was conducted for articles published from the regional Arab and non-European countries. Index terms used were Wilson Disease, genotype, phenotype, and each of the following countries: Lebanon, Syria, Jordan, Egypt, Iraq, Saudi Arabia, Kuwait, Bahrain, Qatar, UAE, Yemen, Tunisia, Morocco, Libya, Mauritania, Turkey, Iran and Oman. We included studies in which both the genotype and phenotype were identified. In some studies, it was not clearly indicated whether patients presenting to one medical center with a certain mutation belonged to the same family or to different ethnic groups^[16,17]. This made it difficult to estimate the most frequent genotype. We, therefore, opted to identify common mutations between Lebanon and the region, and to determine the frequent regional mutations as indicated by the authors of the various reports.

RESULTS

Clinical presentation

In this study, 36 Lebanese WD patients, including 15 females and 21 males, were recruited from different regions in Lebanon. Patients belonged to 13 unrelated families, referred to as: U (P₁-P₉); Or (P₁₀); S (P₁₁-P₁₉); Ah (P₂₀); T (P₂₁-P₂₃); B (P₂₄-P₂₅); H (P₂₆-P₂₈); Ha (P₂₉-P₃₀); Is (P₃₁); Z (P₃₂-P₃₃); Ri (P₃₄); Sc (P₃₅); and Gh (P₃₆) families. Consanguinity was present in the parents of 27 of the patients (75%), who belonged to the U, S, B, H, Ha, and Z families (Table 1). WD scores computed following EASL guidelines ranged between 4 and 12 (Table 2), confirming the diagnosis.

The clinical profiles of affected subjects are summarized in Table 2. Age at diagnosis ranged between 1 and 39 years. All patients had low Cp level (< 0.2 g/L), except for P₄ and P₁₄. Out of 31 patients, 18 (58%) had KF rings (5/36 were NAV). Out of the 36 WD patients, 24 were symptomatic (67%; 16 males and 8 females) and presented clinically at an average age of 14.5 years. Data on P₆-P₉ were not available. Twelve patients were asymptomatic (33%), diagnosed by genetic screening of family members of index patients. Their average age was 7.6 years.

Pure hepatic phenotype was the most common in our symptomatic patients [9/32: P₁, P₁₆-P₁₇, P₁₈, P₂₀, P₂₅, P₂₇-P₂₈ and P₃₂]. Neurologic presentation was noted in 12.5% of patients (4/32: P₁₂, P₁₄, P₃₄ and P₃₆). Mixed presentation was observed in 25% of patients (7/32: P₂, P₁₀, P₂₄, P₂₆, P₂₉, P₃₁ and P₃₅), two of whom had suicidal attempts/disposition (P₁₀ and P₂₆). Notably, liver cirrhosis was present in 12 symptomatic (38%) patients (P₁, P₂, P₁₀, P₁₆-P₁₈, P₂₀, P₂₄, P₂₆-P₂₈ and P₃₂), including 4 patients with mixed presentation.

Of the asymptomatic subjects who were diagnosed by screening, 10/12 patients had evidence of liver disease, ranging from transaminitis (P₃-P₅, P₁₁, P₁₃,

Table 1 Spectrum of mutations in the *ATP7B* gene of Lebanese patients with Wilson's disease

Family	ID	Sex	Birth date	AD	Exon	Mutation(s)	Region of protein
U	P ₁	M	1966	7	7	Gly691Arg	TM2
	P ₂	M	1985	9	7	Gly691Arg	TM2
	P ₃	F	1986	13 ¹	7	Gly691Arg	TM2
	P ₄	F	1990	9 ¹	7	Gly691Arg	TM2
	P ₅	M	1996	3 ¹	7	Gly691Arg	TM2
	P ₆	M	NAV	3 ²	7	Gly691Arg	TM2
	P ₇	M	NAV	7 ²	7	Gly691Arg	TM2
	P ₈	M	NAV	12 ²	7	Gly691Arg	TM2
	P ₉	F	NAV	NAV	7	Gly691Arg	TM2
Or	P ₁₀	F	1986	21	7/10	Gly691Arg/Val845Ser	TM2/Td
S	P ₁₁	F	1993	5 ¹	8	2299insC/2299insC	TM4
	P ₁₂	M	1973	12	8	2299insC/2299insC	TM4
	P ₁₃	F	1997	10 ¹	8	2299insC/2299insC	TM4
	P ₁₄	M	1980	16	8	2299insC/2299insC	TM4
	P ₁₅	M	2007	1 ¹	8	2299insC/2299insC	TM4
	P ₁₆	M	1981	16	8/13	2299insC/p.Ala1003Thr	TM4/Ch-TM6
	P ₁₇	F	1983	14	8/13	2299insC/p.Ala1003Thr	TM4/Ch-TM6
	P ₁₈	F	1993	12	8/13	2299insC/p.Ala1003Thr	TM4/Ch-TM6
	P ₁₉	F	1989	15 ¹	8/13	2299insC/p.Ala1003Thr	TM4/Ch-TM6
Ah	P ₂₀	M	1992	15	8	2299insC/2299insC	TM4
T	P ₂₁	F	1998	7	12	Trp939Cys	Td
	P ₂₂	M	2001	8 ¹	12	Trp939Cys	Td
	P ₂₃	M	2006	3 ¹	12	Trp939Cys	Td
B	P ₂₄	M	1992	13	12	Trp939Cys	Td
	P ₂₅	M	2002	5 ³	12	Trp939Cys	Td
H	P ₂₆	F	1985	18	18	Asn1270Ser	ATP hinge
	P ₂₇	F	1987	18 ¹	18	Asn1270Ser	ATP hinge
	P ₂₈	F	1991	8	18	Asn1270Ser	ATP hinge
Ha	P ₂₉	F	1998	14	18	Asn1270Ser	ATP hinge
	P ₃₀	F	2002	11	18	Asn1270Ser	ATP hinge
Is	P ₃₁	M	1995	13	18	Asn1270Ser	ATP hinge
Z	P ₃₂	M	1990	15	18	Pro1273Leu	ATP hinge
	P ₃₃	M	2000	6 ¹	18	Pro1273Leu	ATP hinge
Ri	P ₃₄	M	2009	3	19	Arg1319stop	TM7
Sc	P ₃₅	M	1979	22	15/19	Thr1092Met/Arg1319stop	ATP loop/TM7
Gh	P ₃₆	M	1970	39	-	None identified	-

¹Screening; ²Died at age; ³Deceased. AD: Age at diagnosis; NAV: Not available.

P₂₁-P₂₃, P₃₀ and P₃₃) and hepatomegaly detected by abdominal ultrasound (P₁₁, P₁₃ and P₂₁) to full blown cirrhosis (P₂₂-P₂₃). Overall, 27/32 patients (84%) on whom we had clinical information presented with some form of hepatic dysfunction.

Patients with the neurologic phenotype presented at an average age of 22.3 years, while those with hepatic and mixed phenotypes presented at 12.2 and 14 years, respectively. KF rings were present in 17 symptomatic patients (5 symptomatic were NAV) and absent in two (P₂₅, P₃₄). They were not identified in the asymptomatic patients, except for patient P₁₉ who had KF rings with no evidence of hepatic or neurologic dysfunctions.

Mutation analysis

Sequencing of the *ATP7B* gene revealed (Table 1) 9 different disease-causing mutations in 70 chromosomes (35 patients), which were distributed as: 7 missense (exons: 7, 12, 10, 13, 15 and 18), 1 non-sense (exon 19), and 1 frame-shift (exon 8). Out of 70 chromosomes, missense/frameshift and/non-sense

mutation(s) were detected in 51:16:3 chromosomes at 72.8%:22.8%:4.3% frequency respectively. No mutation was identified in P₃₆, who had been diagnosed based on KF rings, and clinical and biochemical testing.

Out of 35 patients, 29 were homozygous (82.8%) for a disease-causing mutation and 6 were compound heterozygous (17.1%). Parents of our index patients were carriers for the disease-causing mutations. Mutations were most frequent in the exon 18 motif encoding the conserved ATP hinge region of WD gene product. Four out of the 13 unrelated families (H, Ha, Is and Z) had, in this motif, missense mutations in the homozygous state; these were Asn1270Ser in 6 patients (P₂₆-P₃₁) and Pro1273Leu in 2 patients (P₃₂-P₃₃), accounting for 17% and 5.7% of chromosomes respectively. Other identified mutations (Table 1) included missense mutations in exon 7 (Gly691Arg; 10 patients: U and Or) and exon 12 (Trp939Cys; 5 patients: T and B), frameshift in exon 8 (2299insC; 10 patients: S and Ah), and nonsense mutation in exon 19 (Arg1319stop; 2 patients: Ri and Sc), accounting for a chromosome frequency of 27%, 14%, 23% and

Table 2 Phenotypic and genotypic profiles of Lebanese patients with Wilson's disease

ID	Mutation(s)	GI manifestation(s)	Neurological manifestations	KF rings	Cp	Urinary Cu	Score
P ₁	Gly691Arg	Liver cirrhosis	Absent	Present	NAV	718.8	8
P ₂	Gly691Arg	Liver cirrhosis	Change in school performance	Present	0.11	1998	10
P ₃	Gly691Arg	Asymptomatic ¹	Absent	Absent	0.03	148.5	8
P ₄	Gly691Arg	Asymptomatic ¹	Absent	Absent	0.22	304	6
P ₅	Gly691Arg	Asymptomatic ¹	Absent	Absent	0.02	65.9	7
P ₆	Gly691Arg	NAV	NAV	NAV	NAV	NAV	4
P ₇	Gly691Arg	NAV	NAV	NAV	NAV	NAV	4
P ₈	Gly691Arg	NAV	NAV	NAV	NAV	NAV	4
P ₉	Gly691Arg	NAV	NAV	NAV	NAV	NAV	4
P ₁₀	Gly691Arg/ Val845Ser	Liver cirrhosis	Suicidal attempts	Present	0.08	2184	12
P ₁₁	2299insC	Asymptomatic	Absent	Absent	0.04	99	7
P ₁₂	2299insC	Absent	Slurred speech, ataxia, tremors	Present	0.072	512	12
P ₁₃	2299insC	Asymptomatic	Absent	Absent	0.03	152.8	8
P ₁₄	2299insC	Absent	Choreoathetosis, tremors, rigidity	Present	0.423	2300	10
P ₁₅	2299insC	Asymptomatic	Absent	Absent	0.019	10	6
P ₁₆	2299insC/ p.Ala1003Thr	Liver cirrhosis	Absent	Present	0.096	775	10
P ₁₇	2299insC/ p.Ala1003Thr	Liver cirrhosis	Absent	Present	0.096	590	10
P ₁₈	2299insC/ p.Ala1003Thr	Liver cirrhosis	Absent	Present	0.17	645	9
P ₁₉	2299insC/ p.Ala1003Thr	Absent	Absent	Present	0.12	487	9
P ₂₀	2299insC	Liver cirrhosis	Absent	NAV	0.023	651	8
P ₂₁	Trp939Cys	Asymptomatic	Absent	Absent	0.02	77.6	7
P ₂₂	Trp939Cys	Asymptomatic	Absent	Absent	0.02	20	6
P ₂₃	Trp939Cys	Asymptomatic	Absent	Absent	0.02	41.5	6
P ₂₄	Trp939Cys	Liver cirrhosis, ascites	Jaw drooping, hypersalivation, slurred speech, narrow based gait, intention tremors	Present	0.021	744	12
P ₂₅	Trp939Cys	Liver cirrhosis, Hepatic encephalopathy, Hepatomegaly, Mild to moderate ascites	Absent	Absent	0.04	NAV	6
P ₂₆	Asn1270Ser	Liver cirrhosis	Psychiatric symptoms and suicidal attempts	Present	0.03	27.6	10
P ₂₇	Asn1270Ser	Liver cirrhosis	Absent	Present	0.03	65.1	9
P ₂₈	Asn1270Ser	Ascites, liver cirrhosis	Absent	Present	0.04	55	9
P ₂₉	Asn1270Ser	Transaminitis	Neurodevelopmental	Present	0.078	171	11
P ₃₀	Asn1270Ser	Asymptomatic	Absent	Absent	0.03	116	8
P ₃₁	Asn1270Ser	Chronic liver parenchymal disease	Dysarthria and left-sided dystonia	Present	0.029	402.3	12
P ₃₂	Pro1273Leu	Ascites, Liver cirrhosis, Hepatic encephalopathy	Absent	Present	0.17	1041.1	9
P ₃₃	Pro1273Leu	Asymptomatic	Absent	Absent	0.19	89.7	6
P ₃₄	Arg1319stop	Asymptomatic	Delay in speech	Absent	0.02	92	8
P ₃₅	Thr1092Met/ Arg1319stop	Chronic liver disease and early portal hypertension	Clenching of mandible, left side dystonia, sialorrhea, dysarthria, head tremors	Present	0.025	199	12
P ₃₆	None identified	Absent	Drooling, dysarthria, difficulty concentrating, dysphagia	Present	0.085	NAV	6

¹Developed later. Normal range: Serum ceruloplasmin: 0.2 to 0.6 g/L; Urine copper: 15 to 50 µg/24 h. Score = Ferrenci Score of diagnosis. 2 or less: Very unlikely; 3: Possible, more tests needed; 4 or more: Established^[4]. Cp: Ceruloplasmin (g/L); KF: Keiser-Fleischer; NAV: Not available; Urinary Cu: 24-h urine copper (µg/24 h).

4.3% respectively. Compound heterozygous mutations were identified in exons 10 (Or: P₁₀), 13 (S: P₁₆-P₁₉) and 15 (P₃₅).

Eight polymorphisms were detected in exons 2, 3, 10, 12, 13 and 16 (Table 3) in patients and normal chromosomes obtained from related and unrelated individuals. Three polymorphisms (Lys832Arg, Arg952Lys and Val1140Ala) were present in the homo-

zygous state in 94% (34/36) of patients and in the heterozygous state in 5% (P₁₁ and P₁₉), in addition to others in exons 2, 3 and 13 (Table 3).

WD patients: Lebanon vs regional countries

A search of the literature for population studies on the spectrum of mutations in WD patients in the region, including Arab and non-European countries,

Table 3 Identified polymorphisms in the *ATP7B* gene of Lebanese patients with Wilson's disease

Polymorphism	Asp96Gly	Ser406Ala	Val456Leu	Lys832Arg	Arg952Lys	Ala1003 Ala	Val1140Ala	Ser1166Ser
Exon	2	2	3	10	12	13	16	16
Base change	GAC → GGC	TCT → GCT	GTG → CTG	AAG → AGG	AGA → AAA	GCG → GCA	GTC → GCC	AGC → AGT
Domain	Cu1-4	Cu4 binding	Cu4/Cu5	Td	Tm5	ATP binding/Tm6	ATP loop	ATP loop
Family								
U				HM	HM		HM	
Or		HM	HM	HM	HM		HM	
S								
P1, P2, P31, P41, P59				HM	HM		HM	
P7, P8				HT	HT		HT	
P3, P4					HM		HM	
AH			HM	HM	HM		HM	
TF				HM	HM	HM	HM	HM
B				HM	HM	HM	HM	
H		HM	HM	HM	HM		HM	
Ha								HM
Is			HM	HM	HM		HM	
Z		HM	HM	HM	HM		HM	
Ri	HM			HM			HM	
Sc			HT	HM	HM		HM	
Gh		HM	HM	HM	HM		HM	
Ah			HM	HM	HM		HM	

was conducted. A total of 77 articles on WD patients were initially identified, but only those reporting the genotypes and/or the phenotypes were considered. Consanguinity, homozygosity and frequency of mutation were also noted when indicated.

Seventeen articles were included and distributed as follows: Saudi Arabia^[18-21], Egypt^[12,22-24], Turkey^[25,26], Iran^[27,28], Oman^[29] and Lebanon^[6,13,14,30]. Two reports on WD from Iraq were not included, as they had no genotypic information. There were no reports on WD from Jordan, Libya, Tunisia, Morocco or Syria.

Homozygosity was highly prevalent in Lebanese WD patients (83%), and ranged between 68%-85.7% and 50%-53% in Egyptian and Saudi Arabian patients respectively. This finding is attributed to high consanguinity (Table 4) that is common in our societies, or the high prevalence of the same mutation in carriers. Frequency of asymptomatic cases was relatively similar in Lebanon, Egypt and Saudi Arabia. Similar to Lebanese patients, many of asymptomatic patients had evidence of hepatic dysfunction on laboratory and/or imaging studies. Hepatic phenotype was more common than neurologic phenotype in patients from Lebanon, Egypt, Turkey, Iran and Saudi Arabia. Taking into account patients who are asymptomatic and those with mixed phenotype, the vast majority of patients in those countries have some form of hepatic dysfunction. A minority of patients had pure neurologic phenotype. Also, the frequency of patients having KF rings was high and was similar in the 5 countries (Table 4). In a report on a single family from Oman, 78% of patients were asymptomatic and 21% had neurologic phenotype. No patients had a hepatic phenotype in that study.

In conducting our analysis of genotypes, we con-

sidered a mutation to be frequent if it was present in multiple unrelated families. We compared genotypic changes in the *ATP7B* gene of Lebanese patients with those from regional Arab and non-European patients. In our patients, the conserved ATP hinge region (exon 18) was the most frequently mutated region identified in 4 unrelated families (Table 1).

Table 3 shows that Lebanese patients share in common with: (1) Egypt, Iran and Turkey, the Val845Ser and Asp1270Ser mutations in exons 10 and 18 respectively; (2) Egypt, the Pro1273Leu mutation in exon 18; (3) Egypt and Turkey, the Arg1319X mutation in exon 19; and (4) Turkey, the Ala1003Thr mutation in exon 13 and the exon 7 mutation (Gly691Arg) reported in one Turkish patient^[26]. More interestingly, the mutation in exon 12 (Trp939Cys) was only detected in Lebanese patients and in none of the searched/ listed countries. Whereas the worldwide exon 14 mutation (His1069Gln) was detected in some patients from Egypt, Iran and Turkey, it was not identified in Lebanese or in Saudi Arabian patients.

DISCUSSION

The diagnosis of WD based on clinical grounds alone is often difficult. Thus, it may be necessary to resort to genetic testing. In this study, involving more than 500 patients from Lebanon and the region, we found a great deal of genetic heterogeneity with no common or population specific mutation. This reflects the extensive ethnic diversity of people in this part of the world and argues against the presence of a founder gene, even in highly consanguineous populations. It also implies that patients suspected to have WD without a family history, *i.e.*, without a known mutation

Table 4 Lebanese vs regional Arab and non-European Wilson's disease patients: Genotype-phenotype

	Lebanon	Egypt	Iran	Turkey	Saudi Arabia	Oman
Number of patients	36	198	88	46	152 ¹	14
Number of families	13	135	-	46	53	1
% Homozygosity	83%	68.4% - 85.7%	NAV	NAV	50%-53%	NAV
% Consanguinity	75%	39.5% - 78.9%	NAV	NAV	36.6%-88.8%	NAV
% Hepatic manifestation	28%	45.5% - 84.2%	65.20%	43.50%	25%-54.9%	0%
% Neurologic manifestation	12.50%	4.2%-15.8%	4.30%	34.80%	0%-25%	21.40%
% Mixed manifestation	21.80%	0%-20.9%	21.70%	21.70%	19.6%-55.6%	0%
% Asymptomatic	37%	0%-35.1%	-	0%	30.35%	78.60%
% KF rings	58%	26.3%-69.2%	65.20%	67.40%	50.7%-59%	NAV
Mutation						
E2		Glu396stop		Gly457stop		
E3			No common mutations identified			
E4-6						
E7	Gly691Arg			Gly691Arg		
E8	2299insC	c. 2304-5insC Cys703Tyr	Trp779Gly	Gly710Ser Pro767 Arg	Ser744Pro	
E9			No common or frequent mutations identified			
E10	Val845Ser	Val845Ser	Val845Ser	Val845Ser		
E11			No common or frequent mutations identified			
E12	Trp939Cys					
E13	Ala1003Thr					
E14			3061-1G>A sp	Ala1003Thr		Deletion of E13
E15	Thr1092Met	Thr1076Ile				
E16-17		His1069Gln	His1069Gln	His1069Gln		
E18	Asn1270Ser Pro1273Leu	His1126fs Asn1270Ser Pro1273Leu IVS18-2A>G	Ile1102Thr No common or frequent mutations identified Asn1270Ser	No common or frequent mutations identified Asn1270Ser		
E19	Arg1319stop	Arg1319stop		Arg1319stop		
E20					Gly1341Ser	
E21	Barada <i>et al</i> ^[13,30]	Abdelghaffar <i>et al</i> ^[12,22]	Dastsooz <i>et al</i> ^[27]	Simsek Papur <i>et al</i> ^[25]	Gln1399Arg	
Ref.	Usta <i>et al</i> ^[6,14]	El-Karakasy <i>et al</i> ^[23] El-Mougy <i>et al</i> ^[24]	Zali <i>et al</i> ^[28]	Loudianos <i>et al</i> ^[26]	Al Jumah <i>et al</i> ^[18] Al Fadda <i>et al</i> ^[19] Majumdar <i>et al</i> ^[20,21]	Al-Tobi <i>et al</i> ^[29]

¹152 patients described in articles from Saudi Arabia, of which 5 patients were from Yemen and Syria.

in their family, may need to be screened for mutations in all exons of the *ATP7B* gene. In view of clustering of WD patients within families, their members should be screened for mutations identified in index patients. This is important as it could prevent the silent progression of WD, which may occur as early as 1 year of age, and facilitate management.

Based on the recently published EASL criteria for diagnosis, all our symptomatic and asymptomatic patients had a composite score > 4 (range: 6-12), confirming

the diagnosis beyond doubt. In many of our patients, confirmation of the diagnosis required mutation analysis. Traditionally WD was diagnosed on the basis of low Cp level, KF ring presence and increased 24-h urine Cu level in the context of hepatic and/or neurologic manifestations^[31]. In our experience, many patients with WD do not satisfy all these criteria. For example, patients P₄ and P₁₄ had normal Cp, 13 did not have KF rings and 4 had normal 24-h urinary Cu. This highlights the difficulties and challenges of making a diagnosis of WD based on clinical grounds alone, particularly in asymptomatic patients.

Worldwide, the majority of WD patients are compound heterozygous^[32]. In contrast, in our community, the high rate of consanguinity increases the chance of homozygosity, which is present in 83% of our patients. Only 17% of our patients were compound heterozygous. Missense mutations were the most predominant in Lebanese patients, as worldwide^[33]. These occurred in 8 exons of *ATP7B*. One possible hot spot of the WD gene in our patients is that of the conserved ATP hinge region in exon 18. Two mutations in the homozygous state, Pro1273Leu and Asn1270Ser, were the most frequent, being identified in 8 patients from 4 unrelated families. None of the possible hot spot mutations in Lebanon were shared with those of Asia, Latin America or Europe.

One of our WD patients had no identifiable mutations in the coding region of the *ATP7B* gene. Mutations may be present in the promoter or the transcription factor regions which control protein translation and function. In such cases, detailed clinical testing and family history may be of help in diagnosis, such as P₃₆, in whom the diagnosis was based on clinical assessment showing low Cp and presence of KF rings in the context of neurologic manifestation. Finally, all our patients had multiple genetic polymorphisms that may influence the final folded conformation, affinity and/or the function of the Wilson protein and possibly the phenotype of WD patients^[34].

Remarkable differences in phenotypes and age at diagnosis were noted among patients and even among family members carrying the same genotype. The age of onset of the disease varied between 1-22 years, with one (P₃₆) diagnosed at 39 years. In the B family, patient P₂₅ was diagnosed at 5 and passed away before the onset of his brother's symptoms at the age of 13 (P₂₄). Variation in age at diagnosis was also observed in asymptomatic cases. During a checkup at the age of 7, the female index patient in family T (P₂₁) was found to have transaminitis and hepatomegaly. She was confirmed to have WD and was homozygous for a mutation in exon 12. Genetic screening of her 2 brothers, P₂₂ (8 years) and P₂₃ (3 years), confirmed WD. Though they were asymptomatic, it was surprising to find that both already had evidence of liver cirrhosis on liver imaging. This raises the question as to whether sex plays a role in the clinical manifestations of disease^[15]. Verification of this, however, requires a cohort study with a larger number of patients. Such

phenotypic diversity has been reported even among monozygotic twins^[35], suggesting a role for epigenetic and/or environmental factors in the expression of WD^[36-38].

Diversity in clinical presentation introduces yet another obstacle in the diagnosis of WD, regardless of whether the patient is symptomatic or asymptomatic. A patient, at age of diagnosis, may have mild to severe hepatic and/or neurologic symptoms with or without KF rings. This emphasizes, again, the limitations of pure clinical evaluation and argues for genetic testing of all family members of an affected sibling. In our patients, 28% had pure hepatic manifestations ranging from transaminitis and hepatomegaly to clinically unapparent or overt cirrhosis and portal hypertension. On the other hand, only 9% of our patients had pure neurologic symptoms ranging from weak school performance, slurred speech, tremors, drooling, dysarthria, dysphagia and ataxia to suicidal attempts in some (P₁₀ and P₂₆).

Our asymptomatic patients (36%) were found to have liver involvement (transaminitis, fatty liver and cirrhosis) with no KF rings, except for P₁₉. Changes such as fatty liver were detected at the age of 1 year in P₁₅, diagnosed by genetic screening. Therefore, early diagnosis is important in families with index patient(s), to mitigate against progression of the disease. This is in line with the EASL recommendations to perform genetic testing for WD, in individuals with liver disease or neurologic movement disorders of unclear etiology. Whether genetic testing for WD in patients with unexplained hepatic dysfunction will turn out to be cost effective or not in this part of the world is unclear.

Few studies from the Arab world on WD from Lebanon, Egypt, Saudi Arabia and Oman have been published. Similar to Lebanon, the majority of patients from Egypt and Saudi Arabia have consistently shown a high prevalence of consanguinity and homozygosity, with a great deal of genetic heterogeneity, and no mutation characteristic of the region identified. The predominant phenotype of WD in the region was also hepatic, suggesting the benefits of screening for WD in patients with unexplained hepatic dysfunction.

Lebanese and Egyptian patients share missense mutations in exons 8, 10, 18 and 19 (Table 4). However, mutations Gly691Arg and Trp939Cys were identified in Lebanese patients but not in Saudi Arabian or Egyptian ones. There were also common mutations with Turkish WD patients, including exon 7 (Gly691Arg), exon 10 (Val845Ser), exon 13 (Ala1003Thr), exon 18 (Asp1270Ser) and exon 19 (Arg1319stop). Only exon 10 (Val845Ser), and exon 18 (Asp1270Ser) were shared with Iranian patients. Interestingly, the exon 12 mutation of Trp939Cys was detected in Lebanon but not in any regional country. We reported this mutation in the homozygous state in 5 Lebanese patients, while worldwide it was only detected in 1 Hungarian patient in the heterozygous state^[39]. This extensive genotypic diversity argues for testing patients suspected to have WD for mutations in all exons of *ATP7B*. The shared

mutations with the region may be attributed to common ancestors (Turkey and Egypt) who ruled Lebanon in the past. The origin of the Trp939Cys mutation, however, remains undetermined.

To our surprise, the His1069Gln mutation which is common in diverse populations in North America, Europe and several Mediterranean countries^[40] was not present in Lebanese patients, but was reported in a minority of patients from Egypt, Iran and Turkey. We did not identify a predominant mutation in Lebanon or the region. Whether mutations in the ATP hinge region in exon 18 may turn out to be a hot spot in this part of the world requires further studies on larger numbers of WD patients.

One major strength of our study is that it involves more than 500 patients from Lebanon and the region. It includes a comprehensive clinical and genetic assessment of WD patients in Lebanon, as well as studies from the region clearly stating the genotype and phenotype. Our patients belonged to extended consanguineous families having similar environmental exposures and dietary habits, which helped in reducing the effects of compounding factors on the genotype and phenotype of patients. In addition, our study has some limitations including the absence of true population studies and the lack of long-term follow-up to determine reliably the true phenotype of patients. It is possible that many WD patients in Lebanon and the region remain undiagnosed or unreported, hence missing new mutations and other genotype-phenotype associations.

In conclusion, WD in Lebanon and the region is characterized by extensive genotypic and phenotypic diversity, and by high rates of consanguinity and homozygosity. No predominant mutation has been identified in the region, while the predominant phenotype seems to be hepatic. Clinical and/or genetic testing of all family members for WD, as well as those with unexplained hepatic dysfunction may increase the detection rate of the disease. This could facilitate early institution of therapy and reduce the mortality and morbidity of this condition.

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COMMENTS

Background

Wilson's disease (WD) is an autosomal recessive disorder of copper metabolism, characterized by extensive phenotypic diversity. Most of the patients are compound heterozygotes, having different mutation on each of the *ATP7B* alleles. Attempts to establish genotype-phenotype correlations was hampered by the large number of mutations in the *ATP7B* gene and difficulty

in ascribing a phenotype to one allele. This, however, may be overcome by examining WD in homozygous patients. In Lebanon, consanguinity is quite prevalent, increasing the probability of homozygosity and possibility of establishing a phenotype-genotype correlation. They hereby report the spectrum of mutations and phenotypes of 36 Lebanese patients diagnosed with WD. In addition, we examine if a frequent mutation characterizing the region exists by comparing our findings with those reported from regional studies on WD in Arab and non-European countries.

Research frontiers

This manuscript examines whether genotype-phenotype correlation exists in Lebanese patients diagnosed with WD. It also determines if a frequent mutation characteristic of the Lebanese patients and/or the region occurs.

Innovations and breakthroughs

This is the first comparative study that attempts to identify a frequent mutation characterizing WD patients from Lebanon and regional Arab and non-European countries. Although this region is characterized by high rates of consanguinity and homozygosity, no frequent mutation has been identified in the region but predominance of the hepatic phenotype was noted.

Applications

This study improves our understanding of WD pathogenesis and the genetic determinants of patients' phenotype. It emphasizes the importance of genetic screening for WD in family members with index patients, as well in patients with unexplained hepatic dysfunction. This would surely facilitate diagnosis and early management prior to onset of symptoms, thereby preventing the progressive clinical deterioration of the patient.

Terminology

WD is a rare disease of copper homeostasis that results from a defect in the *ATP7B* gene encoding a copper transporter. Ceruloplasmin is the major copper carrying protein in blood with ferroxidase activity. Kaiser-Fleischer rings refer to copper deposition circumscribing the iris of the eye, diagnostic of WD.

Peer-review

It is a very interesting manuscript.

REFERENCES

- 1 **Schilsky ML.** Wilson disease: Clinical manifestations, diagnosis, and natural history. Available from: URL: <https://www.uptodate.com/contents/wilson-disease-clinical-manifestations-diagnosis-and-natural-history>
- 2 **Sternlieb I.** The outlook for the diagnosis of Wilson's disease. *J Hepatol* 1993; **17**: 263-264 [PMID: 8315256]
- 3 **Ferenci P, Caca K, Loudianos G, Mieli-Vergani G, Tanner S, Sternlieb I, Schilsky M, Cox D, Berr F.** Diagnosis and phenotypic classification of Wilson disease. *Liver Int* 2003; **23**: 139-142 [PMID: 12955875]
- 4 **European Association for Study of Liver.** EASL Clinical Practice Guidelines: Wilson's disease. *J Hepatol* 2012; **56**: 671-685 [PMID: 22340672 DOI: 10.1016/j.jhep.2011.11.007]
- 5 **Hahn SH.** Population screening for Wilson's disease. *Ann N Y Acad Sci* 2014; **1315**: 64-69 [PMID: 24731025 DOI: 10.1111/nyas.12423]
- 6 **Usta J, Wehbeh A, Rida K, El-Rifai O, Estiphan TA, Majarian T, Barada K.** Phenotype-genotype correlation in Wilson disease in a large Lebanese family: association of c.2299insC with hepatic and of p. Ala1003Thr with neurologic phenotype. *PLoS One* 2014; **9**: e109727 [PMID: 25390358 DOI: 10.1371/journal.pone.0109727]
- 7 **Gupta A.** Low-density oligonucleotide microarrays - A major step in Wilson's disease diagnosis. *Indian J Med Res* 2015; **141**: 145-147 [PMID: 25900946]
- 8 **Thomas GR, Forbes JR, Roberts EA, Walshe JM, Cox DW.** The Wilson disease gene: spectrum of mutations and their consequences. *Nat Genet* 1995; **9**: 210-217 [PMID: 7626145 DOI: 10.1038/ng0995210]

- 10.1038/ng0295-210]
- 9 **Chuang LM**, Wu HP, Jang MH, Wang TR, Sue WC, Lin BJ, Cox DW, Tai TY. High frequency of two mutations in codon 778 in exon 8 of the ATP7B gene in Taiwanese families with Wilson disease. *J Med Genet* 1996; **33**: 521-523 [PMID: 8782057]
 - 10 **Gialluisi A**, Incollu S, Pippucci T, Lepori MB, Zappu A, Loudianos G, Romeo G. The homozygosity index (HI) approach reveals high allele frequency for Wilson disease in the Sardinian population. *Eur J Hum Genet* 2013; **21**: 1308-1311 [PMID: 23486543 DOI: 10.1038/ejhg.2013.43]
 - 11 **Tadmouri GO**, Nair P, Obeid T, Al Ali MT, Al Khaja N, Hamamy HA. Consanguinity and reproductive health among Arabs. *Reprod Health* 2009; **6**: 17 [PMID: 19811666 DOI: 10.1186/1742-4755-6-17]
 - 12 **Abdelghaffar TY**, Elsayed SM, Elsobky E, Bochow B, Büttner J, Schmidt H. Mutational analysis of ATP7B gene in Egyptian children with Wilson disease: 12 novel mutations. *J Hum Genet* 2008; **53**: 681-687 [PMID: 18483695 DOI: 10.1007/s10038-008-0298-7]
 - 13 **Barada K**, Nemer G, ElHajj II, Touma J, Cortas N, Boustany RM, Usta J. Early and severe liver disease associated with homozygosity for an exon 7 mutation, G691R, in Wilson's disease. *Clin Genet* 2007; **72**: 264-267 [PMID: 17718866 DOI: 10.1111/j.1399-0004.2007.00853.x]
 - 14 **Usta J**, Abu Daya H, Halawi H, Al-Shareef I, El-Rifai O, Malli AH, Sharara AI, Habib RH, Barada K. Homozygosity for Non-H1069Q Missense Mutations in ATP7B Gene and Early Severe Liver Disease: Report of Two Families and a Meta-analysis. *JIMD Rep* 2012; **4**: 129-137 [PMID: 23430908 DOI: 10.1007/8904_2011_91]
 - 15 **Ferenci P**. Whom and how to screen for Wilson disease. *Expert Rev Gastroenterol Hepatol* 2014; **8**: 513-520 [PMID: 24650289 DOI: 10.1586/17474124.2014.899898]
 - 16 **Kalinsky H**, Funes A, Zeldin A, Pel-Or Y, Korostishevsky M, Gershoni-Baruch R, Farrer LA, Bonne-Tamir B. Novel ATP7B mutations causing Wilson disease in several Israeli ethnic groups. *Hum Mutat* 1998; **11**: 145-151 [PMID: 9482578 DOI: 10.1002/(SICI)1098-1004(1998)11:2<145::AID-HUMU7>3.0.CO;2-I]
 - 17 **Hussein H**, Jabbar A. Mutation analysis in Iraqi patients with Wilson's disease: Identification of four novel mutations. *Wasit J Sci Med* 2014; **7**: 149-158
 - 18 **Al Jumah M**, Majumdar R, Al Rajeh S, Awada A, Al Zaben A, Al Traif I, Al Jumah AR, Rehana Z. A clinical and genetic study of 56 Saudi Wilson disease patients: identification of Saudi-specific mutations. *Eur J Neurol* 2004; **11**: 121-124 [PMID: 14748773]
 - 19 **Al Fadda M**, Al Quaiz M, Al Ashgar H, Al Kahtani K, Helmy A, Al Benmoussa A, Abdulla M, Peedikayil M. Wilson disease in 71 patients followed for over two decades in a tertiary center in Saudi Arabia: a retrospective review. *Ann Saudi Med* 2012; **32**: 623-629 [PMID: 23396027 DOI: 10.5144/0256-4947.2012.623]
 - 20 **Majumdar R**, Al Jumah M, Al Rajeh S, Fraser M, Al Zaben A, Awada A, Al Traif I, Paterson M. A novel deletion mutation within the carboxyl terminus of the copper-transporting ATPase gene causes Wilson disease. *J Neurol Sci* 2000; **179**: 140-143 [PMID: 11054498]
 - 21 **Majumdar R**, Al Jumah M, Fraser M. 4193delC, a common mutation causing Wilson's disease in Saudi Arabia: rapid molecular screening of patients and carriers. *Mol Pathol* 2003; **56**: 302-304 [PMID: 14514926]
 - 22 **Abdel Ghaffar TY**, Elsayed SM, Elnaghy S, Shaded A, Elsobky ES, Schmidt H. Phenotypic and genetic characterization of a cohort of pediatric Wilson disease patients. *BMC Pediatr* 2011; **11**: 56 [PMID: 21682854 DOI: 10.1186/1471-2431-11-56]
 - 23 **El-Karakasy H**, Fahmy M, El-Raziky MS, El-Hawary M, El-Sayed R, El-Koofy N, El-Mougy F, El-Hennawy A, El-Shabrawi M. A clinical study of Wilson's disease: The experience of a single Egyptian Paediatric Hepatology Unit. *Arab J Gastroenterol* 2011; **12**: 125-130 [PMID: 22055589 DOI: 10.1016/j.ajg.2011.07.007]
 - 24 **El-Mougy FA**, Sharaf SA, Elsharkawy MM, Mandour IA, El-Essawy RA, Eldin AM, Helmy HM, Soliman DH, Selim LH, Sharafeldin HM, Mogahed EA, El-Karakasy HM. Gene mutations in Wilson disease in Egyptian children: report on two novel mutations. *Arab J Gastroenterol* 2014; **15**: 114-118 [PMID: 25465132 DOI: 10.1016/j.ajg.2014.10.005]
 - 25 **Simsek Papur O**, Akman SA, Cakmur R, Terzioğlu O. Mutation analysis of ATP7B gene in Turkish Wilson disease patients: identification of five novel mutations. *Eur J Med Genet* 2013; **56**: 175-179 [PMID: 23333878 DOI: 10.1016/j.ejmg.2013.01.003]
 - 26 **Loudianos G**, Dessi V, Lovicu M, Angius A, Nurchi A, Sturniolo GC, Marcellini M, Zancan L, Bragetti P, Akar N, Yagci R, Vegnente A, Cao A, Pirastu M. Further delineation of the molecular pathology of Wilson disease in the Mediterranean population. *Hum Mutat* 1998; **12**: 89-94 [PMID: 9671269 DOI: 10.1002/(SICI)1098-1004(1998)12:2<89::AID-HUMU3>3.0.CO;2-G]
 - 27 **Dastsooz H**, Imanieh MH, Dehghani SM, Haghighat M, Moini M, Fardaei M. Multiplex ARMS PCR to Detect 8 Common Mutations of ATP7B Gene in Patients With Wilson Disease. *Hepat Mon* 2013; **13**: e8375 [PMID: 24003324 DOI: 10.5812/hepatmon.8375]
 - 28 **Zali N**, Mohebbi SR, Esteghamat S, Chiani M, Haghighi MM, Hosseini-Asl SM, Derakhshan F, Mohammad-Alizadeh AH, Malek-Hosseini SA, Zali MR. Prevalence of ATP7B Gene Mutations in Iranian Patients With Wilson Disease. *Hepat Mon* 2011; **11**: 890-894 [PMID: 22308153 DOI: 10.5812/kowsar.1735143X.762]
 - 29 **Al-Tobi M**, Kashoob M, Joshi S, Bayoumi R. A Novel Splice-site Allelic Variant is Responsible for Wilson Disease in an Omani Family. *Sultan Qaboos Univ Med J* 2011; **11**: 357-362 [PMID: 22087377]
 - 30 **Barada K**, El-Atrache M, El-Hajj II, Rida K, El-Hajjar J, Mahfoud Z, Usta J. Homozygous mutations in the conserved ATP hinge region of the Wilson disease gene: association with liver disease. *J Clin Gastroenterol* 2010; **44**: 432-439 [PMID: 20485189 DOI: 10.1097/MCG.0b013e3181ce5138]
 - 31 **Das SK**, Ray K. Wilson's disease: an update. *Nat Clin Pract Neurol* 2006; **2**: 482-493 [PMID: 16932613 DOI: 10.1038/ncpneuro0291]
 - 32 **Ferenci P**. Wilson's Disease. *Clin Gastroenterol Hepatol* 2005; **3**: 726-733 [PMID: 16233999]
 - 33 **Lv T**, Li X, Zhang W, Zhao X, Ou X, Huang J. Recent advance in the molecular genetics of Wilson disease and hereditary hemochromatosis. *Eur J Med Genet* 2016; **59**: 532-539 [PMID: 27592149 DOI: 10.1016/j.ejmg.2016.08.011]
 - 34 **Lu CX**, Qing Lin, Huang WQ, Tzeng CM. New mutations and polymorphisms of the ATP7B gene in sporadic Wilson disease. *Eur J Med Genet* 2014; **57**: 498-502 [PMID: 24878384 DOI: 10.1016/j.ejmg.2014.04.016]
 - 35 **Członkowska A**, Gromadzka G, Chabik G. Monozygotic female twins discordant for phenotype of Wilson's disease. *Mov Disord* 2009; **24**: 1066-1069 [PMID: 19306278 DOI: 10.1002/mds.22474]
 - 36 **Fraga MF**, Ballestar E, Paz MF, Ropero S, Setien F, Ballestar ML, Heine-Suñer D, Cigudosa JC, Urioste M, Benitez J, Boix-Chornet M, Sanchez-Aguilera A, Ling C, Carlsson E, Poulsen P, Vaag A, Stephan Z, Spector TD, Wu YZ, Plass C, Esteller M. Epigenetic differences arise during the lifetime of monozygotic twins. *Proc Natl Acad Sci USA* 2005; **102**: 10604-10609 [PMID: 16009939 DOI: 10.1073/pnas.0500398102]
 - 37 **Leggio L**, Gasbarrini G, Addolorato G. Wilson's disease. *Lancet* 2007; **369**: 902 [PMID: 17368142 DOI: 10.1016/S0140-6736(07)60437-1]
 - 38 **Takeshita Y**, Shimizu N, Yamaguchi Y, Nakazono H, Saitou M, Fujikawa Y, Aoki T. Two families with Wilson disease in which siblings showed different phenotypes. *J Hum Genet* 2002; **47**: 543-547 [PMID: 12376745 DOI: 10.1007/s100380200082]
 - 39 **Folhoffer A**, Ferenci P, Csak T, Horvath A, Hegedus D, Firneisz G, Osztovits J, Kosa JP, Willheim-Polli C, Szonyi L, Abonyi M, Lakatos PL, Szalay F. Novel mutations of the ATP7B gene among 109 Hungarian patients with Wilson's disease. *Eur J Gastroenterol*

Hepatol 2007; **19**: 105-111 [PMID: 17272994 DOI: 10.1097/01.meg.0000223904.70492.0b]

- 40 **Shah AB**, Chernov I, Zhang HT, Ross BM, Das K, Lutsenko S, Parano E, Pavone L, Evgrafov O, Ivanova-Smolenskaya IA, Annerén G, Westermarck K, Urrutia FH, Penchaszadeh GK,

Sternlieb I, Scheinberg IH, Gilliam TC, Petrukhin K. Identification and analysis of mutations in the Wilson disease gene (ATP7B): population frequencies, genotype-phenotype correlation, and functional analyses. *Am J Hum Genet* 1997; **61**: 317-328 [PMID: 9311736 DOI: 10.1086/514864]

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Observational Study

Short-term outcomes of overlapped delta-shaped anastomosis, an innovative intracorporeal anastomosis technique, in totally laparoscopic colectomy for colon cancer

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Abstract

AIM

To introduce an innovative intracorporeal anastomosis technique named overlapped delta-shaped anastomosis (ODA) for colon cancer cases undergoing totally laparoscopic colectomy (TLC) and to assess its feasibility and safety.

METHODS

From January 2016 to March 2017, a total of 20 consecutive patients with colon cancer accepted TLC and the ODA technique at our medical center. Patient demographics, operative outcomes, perioperative complications, and pathological results were collected and analyzed.

RESULTS

We successfully completed TLC and the ODA procedure in all 20 cases, including 6 (30%) males and 14 (70%) females. In total, 11 (55%), 2 (10%), and 7 (35%) cases accepted right hemicolectomy, transverse hemicolectomy, and left hemicolectomy, respectively. None of the surgeries were converted to an open operation. Mean operative time was 178.5 min, and mean estimated blood loss was 58.5 mL. Mean time to first flatus was 2.5 d, and mean postoperative hospitalization duration was 6.8 d. No severe complications occurred, such as anastomotic leakage, snastomotic stenosis, anastomotic bleeding, and wound infection, except for one case who suffered from an abdominal infection and another case who suffered from gastric paralysis syndrome. Tumor recurrence was not observed in any patient during the follow-up period.

CONCLUSION

The ODA technique for colon cancer cases undergoing TLC appears to be safe and feasible, although our current results need to be verified in further studies.

Key words: Overlapped delta-shaped anastomosis; Safety; Totally laparoscopic colectomy; Intracorporeal anastomosis; Colon cancer

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Core tip: Intracorporeal anastomosis technique is one of the biggest difficulties encountered by surgeons during the totally laparoscopic colectomy procedure. In this paper, we introduce an innovative intracorporeal anastomosis technique named overlapped delta-shaped anastomosis and assess its feasibility and safety.

Zhou HT, Wang P, Liang JW, Su H, Zhou ZX. Short-term outcomes of overlapped delta-shaped anastomosis, an innovative intracorporeal anastomosis technique, in totally laparoscopic colectomy for colon cancer. *World J Gastroenterol* 2017; 23(36): 6726-6732 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6726.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6726>

INTRODUCTION

With the improvement of living standards and the extension of life expectancy, the incidence of colon cancer is increasingly rising and will continue to rise in China^[1,2]. Surgery is the mainstay of treatment for colon cancer. Over the past three decades, minimally invasive surgery for colon cancer has drawn more and more attention^[3]. Nowadays, laparoscopic assisted colectomy (LAC) is widely used for cases with colon cancer and the advantages of this procedure have

been widely verified^[4-6].

With the advances in surgical devices and the improvements in surgical performance, totally laparoscopic colectomy (TLC) has gradually been adopted by experienced surgeons^[7-8]. In theory, the TLC procedure conforms more to the concept of minimally invasive surgery and the principle of the tumor-free technique, while the intracorporeal anastomosis (IA) technique still represents one of the biggest difficulties for surgeons during TLC.

As a new technique, delta-shaped anastomosis was first presented by a Japanese scholar named Kanaya in 2002 after he completed the first case of totally laparoscopic gastroenterostomy (TLG)^[9]. This procedure has been widely adopted for TLG and has proven to be both safe and feasible^[10-12]. However, delta-shaped anastomosis for TLC is rarely reported. Since January 2016, an innovative technique named "overlapped delta-shaped anastomosis (ODA)" has been applied to colon cancer cases undergoing TLC at the National Cancer Center/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College. Herein, we introduce this surgical innovation and assess its feasibility and safety.

MATERIALS AND METHODS

Methods

Prior to surgery, the advantages and disadvantages of TLC and ODA were explained to all patients in detail, and then preoperative informed content was acquired. All patients underwent the same preoperative examinations including routine blood tests, chest X-ray, electrocardiogram examination, and computerized tomography (CT) of the abdominal and pelvic cavity to exclude cases with surgical contraindications and distant organ metastasis. Patients with intestinal obstruction were excluded from our study. The American Joint Committee on Cancer (AJCC) staging system (the seventh edition) was used for tumor staging. Postoperative pain was evaluated by the patients on a subjective visual pain scale ranging from 0 to 10, with 0 representing no pain and 10 representing the worst pain imaginable. The ethics committee at our institution approved this study, and this retrospective study conformed to the ethical standards of the World Medical Association Declaration of Helsinki.

Patients

From January 2016 to March 2017, a total of 20 consecutive patients with colon cancer underwent TLC and the ODA procedure. All of these cases accepted colonoscopy examination, and colon cancer was diagnosed by pathology. Preoperative mechanical bowel preparation was performed using polyethylene glycol electrolyte powder. Preventative antibiotics were administered by intravenous drip 30 min preoperatively and continued for 24 h after the operation.



Figure 1 Five-port technique.

Surgical procedure (taking right hemicolectomy as an example)

Step 1: Positioning the patient and placing trocars:

The patient was placed in the modified lithotomy position. A five-port technique was used: a 12 mm sub-umbilical port as the observation port, a 12 mm port located in the left upper quadrant as the primary operating port, and three 5 mm ports located in the left lower quadrant, the right lower quadrant and the right upper quadrant, respectively, as the secondary operating ports (Figure 1). Abdominal pressure was maintained at approximately 15 mmHg, and then the patient was placed in the Trendelenburg position and left tilt applied in order to expose the mesenteric root, ileocolic vessels, and superior mesenteric vessels.

Step 2: Clearing lymph nodes and tailoring the mesentery:

Along the surface of the superior mesenteric vessels, the mesocolon was dissected in a bottom-up fashion, and then the ileocolic vessels, ascending colon vessels, and the right branch of the transverse colon vessels were exposed and carefully severed (Figure 2A). During this process, attention should be paid to the gastroduodenal trunk, pancreatic head, duodenum, and right gastroepiploic vessels, such that these structures are protected. Next, adhesions between the right abdominal wall and the ascending colon were dissected in a similar bottom-up fashion. Finally, the mesentery of the terminal ileum approximately 15 cm away from the ileocecal region, and the right half of the transverse colon, was tailored, respectively.

Step 3: Transecting and anastomosing the bowel:

The terminal ileum and the right half of the transverse colon were transected using two endoscopic linear cutter staplers (Johnson and Johnson, PSE60A and ECR60B) (Figure 2B). The broken ends were sterilized using alcohol gauze swabs. Then, the proximal ileum and the distal transverse colon were fixed using a piece of absorbable suture; this was performed in an overlapped fashion in order to

facilitate anastomosis (Figure 2C). Two small openings located at the ileum and the transverse colon were created using an ultrasound scalpel. After imbedding the lumens with another endoscopic linear cutter stapler, intestinal walls with no mesentery were got through (Figure 2D). Finally, the common opening was closed using an endoscopic linear cutter stapler (Figure 2E). The specimen was removed from the abdominal cavity using a transverse incision above the symphysis pubis (Figure 2F).

The ways the transverse and left colectomy was performed were similar to the method above mentioned in the right hemicolectomy, including the number of trocars, type of staplers, and same anisoperistaltic anastomosis.

Follow-up

The first day after surgery represented the beginning of the follow-up period. Patients were asked to visit doctors every three months after leaving hospital until two years after surgery, biannually for the next three years and then annually, and the deadline of follow-up period was March 31, 2017.

Statistical analysis

Statistical Package for the Social Sciences (SPSS) software (21.0 version for Windows; SPSS Inc. Chicago, IL, United States) was used for data analyses. Quantitative data following a normal distribution are provided as mean and its range. Qualitative data are provided as number and its percentage. The statistical methods of this study were reviewed by Wang M from our institution.

RESULTS

We successfully completed TLC and the ODA procedure in all 20 cases, including 6 (30%) males and 14 (70%) females. None of the surgeries were converted to an open operation. Mean patient age was 52.6 years (range: 38-67 years), and mean body mass index (BMI) was 22.9 kg/m² (range: 20.2-25.5 kg/m²). Our study included 10 (50%) cases with ascending colon cancer, 4 (20%) with transverse colon cancer, and 6 (30%) with descending colon cancer. In total, 11 (55%), 2 (10%), and 7 (35%) cases accepted right hemicolectomy, transverse hemicolectomy, and left hemicolectomy, respectively. Four (20%) out of the 20 cases accepted preoperative chemotherapy (Table 1).

The mean operative time was 178.5 min (range: 155-225 min), and mean estimated blood loss was 58.5 mL (range: 30-100 mL). For all 20 cases, the mean time to first flatus was 2.5 d (range: 1-3 d), and mean first time to oral intake was 3.0 d (range: 2-4 d). Mean postoperative hospitalization period was 6.8 d (range 5-8 d). Patients only reported slight pain (scoring 2.8 on average) on the first day after the operation, and there was almost no pain (scoring 0.7 on average)

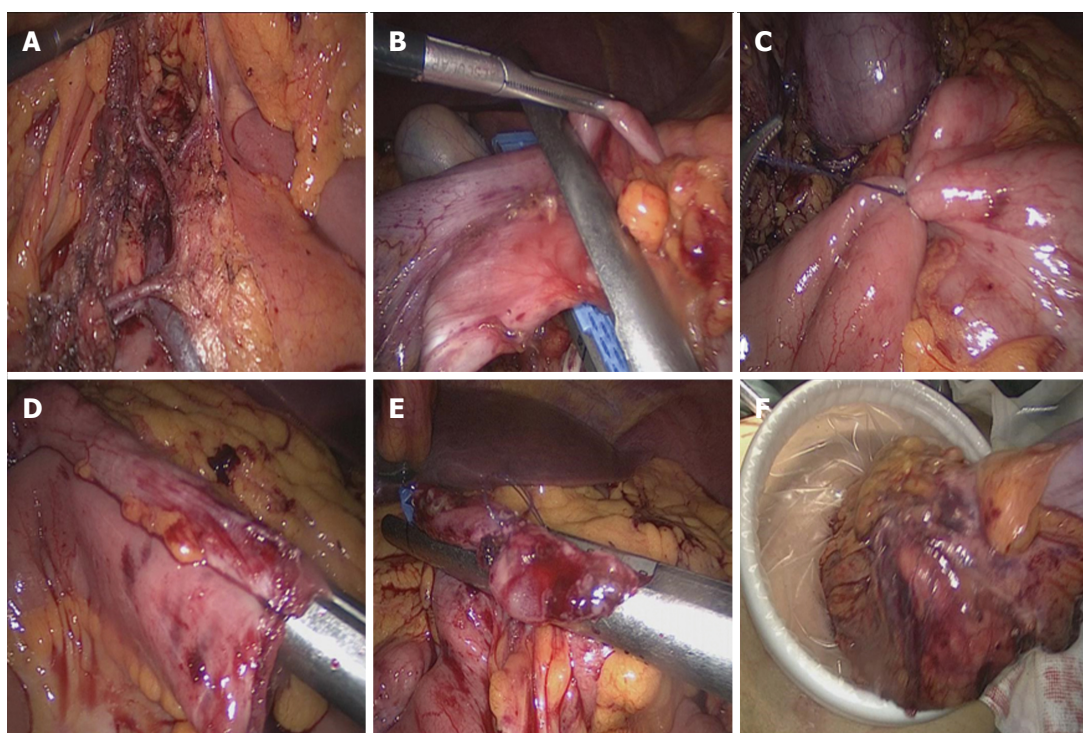


Figure 2 Surgical procedure. A: The ileocolic vessels, ascending colon vessels, and the right branch of the transverse colon vessels were exposed; B: The right half of the transverse colon was transected using endoscopic linear cutter staplers; C: The proximal ileum and the distal transverse colon were fixed in an overlapped fashion using a piece of absorbable suture to facilitate anastomosis; D: After imbedding the lumens with an endoscopic linear cutter stapler, intestinal walls with no mesentery were got through; E: The common opening was then closed using an endoscopic linear cutter stapler; F: Finally, the specimen was removed from the abdominal cavity using a transverse incision above the symphysis pubis.

Table 1 Patient demographics *n* (%)

Parameter	
Gender	
Male	6 (30)
Female	14 (70)
Age, yr, mean (range)	52.6 (38-67)
BMI, kg/m ² , mean (range)	22.9 (20.2-25.5)
ASA score	
1	10 (50)
2	7 (35)
3	3 (15)
Tumor site	
Ascending colon	10 (50)
Transverse colon	4 (20)
Descending colon	6 (30)
Surgical procedure	
Right hemicolectomy	11 (55)
Transverse hemicolectomy	2 (10)
Left hemicolectomy	7 (35)
Preoperative chemotherapy	4 (20)

BMI: Body mass index; ASA: American Society of Anesthesiologists.

by the third day after the operation. No severe complications were encountered, such as anastomotic leakage, anastomotic stenosis, anastomotic bleeding, deep vein thrombosis, or intestinal obstruction, except one case who suffered from abdominal infection due to chylous fistula, and another case who suffered from gastric paralysis syndrome. However, both of these

patients made a healthy recovery with conservative treatments. There were no deaths during the peri-operative period (Table 2).

Pathological results are shown in Table 3. Mean tumor size was 4.2 cm (range: 2.8-6.3 cm) and mean proximal and distal resection margins were 19.5 cm (range: 13.8-23.5 cm) and 17.8 cm (range: 12.2-21.6 cm), respectively. The mean number of lymph nodes harvested was 32.4 (range: 23-45). Among these cases, there were five (25%) cases of stage I, eight (40%) cases of stage II, and seven (35%) cases of stage III disease. A typical specimen and the transverse incision above the symphysis pubis are shown in Figure 3. The mean follow-up time was 8.5 mo (range: 1-15 mo). No patient was lost to follow-up and no cases experienced recurrence during the follow-up period.

DISCUSSION

Surgery plays an important role in the treatment of colon cancer and the goal of every surgeon is to create as little trauma as necessary and encourage a quick recovery. With recent advances in surgical devices and improvements in surgical performance, surgeons have achieved a significant breakthrough in the treatment of colon cancer by changing from open surgery to laparoscopic surgery^[3]. Minimally invasive surgery and quick recovery following laparoscopic surgery have



Figure 3 Typical specimen (A) and a transverse incision above the symphysis pubis (B).

Table 2 Operative outcomes and perioperative complications	
Parameter	
Operative outcomes	
Operative time, min, mean (range)	178.5 (155-225)
Estimated blood loss, mL, mean (range)	58.5 (30-100)
Time to first flatus, d, mean (range)	2.5 (1-3)
Time to first oral intake, d, mean (range)	3.0 (2-4)
Postoperative hospitalization, d, mean (range)	6.8 (5-8)
Length of transverse incision, cm, mean (range)	4.8 (4-6)
Postoperative pain score	
The first day, mean (range)	2.8 (2-4)
The second day, mean (range)	1.5 (1-3)
The third day, mean (range)	0.7 (0-1)
Perioperative complications (%)	
Anastomotic leakage	0 (0)
Anastomotic stenosis	0 (0)
Anastomotic bleeding	0 (0)
Abdominal infection	1 (5)
Deep-vein thrombosis	0 (0)
Wound infection	0 (0)
Intestinal obstruction	0 (0)
Gastric paralysis syndrome	1 (5)
Reoperation (%)	0 (0)

been well verified and documented. As a consequence, laparoscopic surgery has become incredibly popular over the last three decades and several new laparoscopic techniques, including robotic-assisted colectomy, single incision laparoscopic surgery, natural orifice specimen extraction surgery, and natural orifice transluminal endoscopic surgery, have also been reported^[3,13-17]. Although, totally laparoscopic surgery has been carried out by experienced surgeons^[7-8], IA technique still represents a significant difficulty during TLC. Therefore, an easy and feasible anastomosis technique is very important.

In 2002, the Japanese scholar Kanaya first reported the delta-shaped anastomosis of TLG^[9]. In 2011, he summarized 100 cases of delta-shaped anastomosis performed by eight surgeons, and the results showed that, on average, only 13 min were needed to complete anastomosis, and that all eight surgeons experienced a short learning curve. In addition, this earlier study reported that patients had an early time of first oral intake^[18]. Subsequently, more and more studies con-

Table 3 Pathological results	
Parameter	
Tumor size, cm, mean (range)	4.2 (2.8-6.3)
Proximal resection margin, cm, mean (range)	19.5 (13.8-23.5)
Distal resection margin, cm, mean (range)	17.8 (12.2-21.6)
No. of lymph nodes harvested, mean (range)	32.4 (23-45)
pTNM stage (%)	
I	5 (25)
II	8 (40)
III	7 (35)

pTNM: Pathological tumor node metastasis.

firmed the advantages of this procedure^[10-12].

In recent years, several studies have described IA for patients with colon cancer. Wang *et al.*^[17] reported that 11 patients with sigmoid cancer undergoing totally laparoscopic sigmoid colectomy with delta-shaped anastomosis and transvaginal specimen extraction, and these procedures were successful without serious complications. Jian-Cheng *et al.*^[19] reported 56 cases with colon cancer, who had accepted total laparoscopic right hemicolectomy (LRH) with 3-step stapled intracorporeal isoperistaltic ileocolic anastomosis (TLG group). Compared to cases in the extracorporeal anastomosis group (LG group), cases in the TLG group had shorter IA time (9.9-15.5 min vs 13.5-18.2 min, $P < 0.001$), lower mean intraoperative blood loss (83.2 mL vs 93.3 mL, $P < 0.001$), faster recovery of bowel function ($P < 0.001$), and lower postoperative pain score ($P < 0.001$). van Oostendorp *et al.*^[20] conducted a meta-analysis including 12 non-randomized comparative studies and the results showed that IA in the LRH group was associated with reduced short-term morbidity and decreased length of hospital stay suggesting faster recovery. In addition, Wu *et al.*^[21] also reported that the IA for LRH could improve cosmetic effect and result in better postoperative recovery outcomes without increasing intraoperative and postoperative complications. However, the difficulties of IA procedure behind the satisfactory results were all reported by above-mentioned studies.

In the present study, we performed TLC in 20

colon cancer cases using a new IA method named the ODA procedure; all surgeries were carried out smoothly and none of the cases were converted to open operation. Patients had slight pain after surgery and showed early time of first flatus and oral intake. The results also showed that the ODA procedure did not increase postoperative complications and that radical resection was also guaranteed. In particular, the transverse incision above the symphysis pubis was only 4.8 cm in length, much shorter and more covert than that required for LAC. Collectively, these observations provide promising results with regard to its feasibility and safety. It thus appears that TLC and the ODA procedure are more in line with the concept of minimally invasive surgery and enhance recovery following surgery.

This was a retrospective study and only 20 patients were included; these factors may, therefore, represent potential limitations. However, all operations were successfully accomplished and no severe complications occurred. Prospective randomized controlled trials with larger sample sizes and longer follow-up periods are now needed to confirm our results.

The ODA technique for cases with TLC appears to be safe and feasible for suitable patients when performed by experienced surgeons. This promising result now allows us to cater for colon cancer patients who require medical cosmetology.

COMMENTS

Background

With the advances in surgical devices and the improvements in surgical performance, totally laparoscopic colectomy (TLC) has gradually been adopted by experienced surgeons in large medical centers. However, the intracorporeal anastomosis (IA) technique is one of the biggest difficulties encountered by surgeons during the TLC procedure, and the currently adopted delta-shaped anastomosis technique is difficult to operate.

Research frontiers

Now, the mainstream of colectomy for colon cancer is still laparoscopic assisted colectomy. TLC is only adopted by experienced surgeons in large medical centers for the difficulties of IA technique.

Innovations and breakthroughs

In this study, a modified delta-shaped anastomosis technique named overlapped delta-shaped anastomosis (ODA) has been applied to colon cancer cases undergoing TLC in this medical institution. The results show that the ODA technique is a feasible and safe procedure.

Applications

In the present study, we have reported, for the first time, the ODA technique adopted during the TLC procedure. The perfect result has proven its feasibility and safety. Therefore, the ODA procedure can be applied to colon cancer patients who have no contraindications for laparoscopic surgery.

Terminology

The ODA procedure is currently used for the anastomosis between the proximal and distal intestinal canals for colon cancer cases who accepted TLC.

Peer-review

IA technique is one of the biggest difficulties during the TLC procedure. The

authors report an innovative IA technique named the "ODA technique". The short-term outcomes are exciting and it deserves further promotion.

REFERENCES

- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. *CA Cancer J Clin* 2016; **66**: 115-132 [PMID: 26808342 DOI: 10.3322/caac.21338]
- Zeng WG, Liu MJ, Zhou ZX, Hou HR, Liang JW, Wang Z, Zhang XM, Hu JJ. Outcome of Laparoscopic Versus Open Resection for Transverse Colon Cancer. *J Gastrointest Surg* 2015; **19**: 1869-1874 [PMID: 26197767 DOI: 10.1007/s11605-015-2891-3]
- Zeng WG, Zhou ZX. Mini-invasive surgery for colorectal cancer. *Chin J Cancer* 2014; **33**: 277-284 [PMID: 24589210 DOI: 10.5732/cjc.013.10182]
- Briggs A, Goldberg J. Tips, Tricks, and Technique for Laparoscopic Colectomy. *Clin Colon Rectal Surg* 2017; **30**: 130-135 [PMID: 28381944 DOI: 10.1055/s-0036-1597313]
- Wang Y, Zhang C, Feng YF, Fu Z, Sun YM. Comparison of short-term outcomes between laparoscopic-assisted and open complete mesocolic excision (CME) for the treatment of transverse colon cancer. *Chin Clin Oncol* 2017; **6**: 6 [PMID: 28285536 DOI: 10.21037/cco.2017.01.01]
- Agarwal S, Gincher M, Birnbaum E, Fleshman JW, Mutch M. Comparison of long-term follow up of laparoscopic versus open colectomy for transverse colon cancer. *Proc (Bayl Univ Med Cent)* 2015; **28**: 296-299 [PMID: 26130871]
- Swaid F, Sroka G, Madi H, Shteinberg D, Somri M, Matter I. Totally laparoscopic versus laparoscopic-assisted left colectomy for cancer: a retrospective review. *Surg Endosc* 2016; **30**: 2481-2488 [PMID: 26335075 DOI: 10.1007/s00464-015-4502-5]
- Lascarides C, Buscaglia JM, Denoya PI, Nagula S, Bucobo JC, Bergamaschi R. Laparoscopic right colectomy vs laparoscopic-assisted colonoscopic polypectomy for endoscopically unresectable polyps: a randomized controlled trial. *Colorectal Dis* 2016; **18**: 1050-1056 [PMID: 27038277 DOI: 10.1111/codi.13346]
- Kanaya S, Gomi T, Momoi H, Tamaki N, Isobe H, Katayama T, Wada Y, Ohtoshi M. Delta-shaped anastomosis in totally laparoscopic Billroth I gastrectomy: new technique of intraabdominal gastroduodenostomy. *J Am Coll Surg* 2002; **195**: 284-287 [PMID: 12168979 DOI: 10.1016/S1072-7515(02)01239-5]
- Gao B, Huang Q, Dong J. [Clinical research of delta-shaped anastomosis technology in laparoscopic distal gastrectomy and digestive tract reconstruction]. *Zhonghua Weichang Waikē Zazhi* 2017; **20**: 73-78 [PMID: 28105624 DOI: 10.3760/cma.j.issn.1671-0274.2017.01.015]
- Lin M, Zheng CH, Huang CM, Li P, Xie JW, Wang JB, Lin JX, Lu J, Chen QY, Cao LL, Tu RH. Totally laparoscopic versus laparoscopy-assisted Billroth-I anastomosis for gastric cancer: a case-control and case-matched study. *Surg Endosc* 2016; **30**: 5245-5254 [PMID: 27008576 DOI: 10.1007/s00464-016-4872-3]
- Luo R, Ge Y, Wu X, Zhang J. [Long-term survival of total laparoscopic radical distal gastrectomy with delta-shaped anastomosis]. *Zhonghua Weichang Waikē Zazhi* 2016; **19**: 549-552 [PMID: 27215524 DOI: 10.3760/cma.j.issn.1671-0274]
- Cai JC, Hong XY. Laparoscopic-Assisted Natural Orifice Specimen Extraction Radical Descending Colectomy Using a Cai Tube. *World J Surg* 2016; **40**: 2803-2807 [PMID: 27338816 DOI: 10.1007/s00268-016-3597-8]
- Karagul S, Kayaalp C, Sumer F, Ertugrul I, Kirmizi S, Tardu A, Yagci MA. Success rate of natural orifice specimen extraction after laparoscopic colorectal resections. *Tech Coloproctol* 2017; **21**: 295-300 [PMID: 28447167 DOI: 10.1007/s10151-017-1611-2]
- Kayaalp C, Yagci MA, Soyer V. Laparoscopic and natural orifice transluminal restorative proctocolectomy: no abdominal incision for specimen extraction or ileostomy. *Wideochir Inne Tech Maloinwazyjne* 2016; **11**: 115-120 [PMID: 27458493 DOI: 10.5114/wiitm.2016.59578]
- Ngu J, Wong AS. Transanal natural orifice specimen extraction in colorectal surgery: bacteriological and oncological concerns. *ANZ J Surg* 2016; **86**: 299-302 [PMID: 26603221 DOI: 10.1111/

- ans.13383]
- 17 **Wang Z**, Zhang XM, Zhou HT, Liang JW, Zhou ZX. New technique of intracorporeal anastomosis and transvaginal specimen extraction for laparoscopic sigmoid colectomy. *Asian Pac J Cancer Prev* 2014; **15**: 6733-6736 [PMID: 25169517 DOI: 10.7314/apjcp.2014.15.16.6733]
 - 18 **Kanaya S**, Kawamura Y, Kawada H, Iwasaki H, Gomi T, Satoh S, Uyama I. The delta-shaped anastomosis in laparoscopic distal gastrectomy: analysis of the initial 100 consecutive procedures of intracorporeal gastroduodenostomy. *Gastric Cancer* 2011; **14**: 365-371 [PMID: 21573920 DOI: 10.1007/s10120-011-0054-0]
 - 19 **Jian-Cheng T**, Shu-Sheng W, Bo Z, Jian F, Liang Z. Total laparoscopic right hemicolectomy with 3-step stapled intracorporeal isoperistaltic ileocolic anastomosis for colon cancer: An evaluation of short-term outcomes. *Medicine (Baltimore)* 2016; **95**: e5538 [PMID: 27902621 DOI: 10.1097/MD.0000000000005538]
 - 20 **van Oostendorp S**, Elfrink A, Borstlap W, Schoonmade L, Sietses C, Meijerink J, Tuynman J. Intracorporeal versus extracorporeal anastomosis in right hemicolectomy: a systematic review and meta-analysis. *Surg Endosc* 2017; **31**: 64-77 [PMID: 27287905 DOI: 10.1007/s00464-016-4982-y]
 - 21 **Wu Q**, Jin C, Hu T, Wei M, Wang Z. Intracorporeal Versus Extracorporeal Anastomosis in Laparoscopic Right Colectomy: A Systematic Review and Meta-Analysis. *J Laparoendosc Adv Surg Tech A* 2017; **27**: 348-357 [PMID: 27768552 DOI: 10.1089/lap.2016.0485]

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Prospective Study

Effect of local wound infiltration with ropivacaine on postoperative pain relief and stress response reduction after open hepatectomy

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Abstract

AIM

To prospectively evaluate the effect of local wound infiltration with ropivacaine on postoperative pain relief and stress response reduction after open hepatectomy.

METHODS

A total of 56 patients undergoing open hepatectomy

were randomly divided into two groups: a ropivacaine group (wound infiltration with ropivacaine solution) and a control group (infiltration with isotonic saline solution). A visual analog scale (VAS) at rest and on movement was used to measure postoperative pain for the first 48 h after surgery. Mean arterial pressure (MAP), heart rate (HR), time to bowel recovery, length of hospitalization after surgery, cumulative sufentanil consumption, and incidence of nausea and vomiting were compared between the two groups. Surgical stress hormones (epinephrine, norepinephrine, and cortisol) were detected using enzyme-linked immunosorbent assay, and the results were compared.

RESULTS

VAS scores both at rest and on movement at 24 h and 48 h were similar between the two groups. Significantly lower VAS scores were detected at 0, 6, and 12 h in the ropivacaine group compared with the control group ($P < 0.05$ for all). MAP was significantly lower at 6, 12, and 24 h ($P < 0.05$ for all); HR was significantly lower at 0, 6, 12, and 24 h ($P < 0.05$ for all); time to bowel recovery and length of hospitalization after surgery ($P < 0.05$ for both) were significantly shortened; and cumulative sufentanil consumption was significantly lower at 6, 12, 24, and 36 h ($P < 0.05$ for all) in the ropivacaine group than in the control group, although the incidence of nausea and vomiting showed no significant difference between the two groups. The levels of epinephrine, norepinephrine, and cortisol were significantly lower in the ropivacaine group than in the control group at 24 and 48 h ($P < 0.01$ for all).

CONCLUSION

Local wound infiltration with ropivacaine after open hepatectomy can improve postoperative pain relief, reduce surgical stress response, and accelerate postoperative recovery.

Key words: Local wound infiltration; Ropivacaine; Open hepatectomy; Postoperative pain; Surgical stress

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Core tip: This study prospectively evaluated the effect of local wound infiltration with ropivacaine on postoperative pain relief and stress response reduction after open hepatectomy. Wound infiltration with ropivacaine could provide more effective analgesia both at rest and on movement in the first 48 h after surgery, with lower mean arterial pressure, heart rate and sufentanil consumption, accelerated postoperative recovery, and reduced stress response. These results suggest that local wound infiltration with ropivacaine is a simple, convenient and effective analgesic method that can provide postoperative analgesia and short-term benefits after open hepatectomy.

Wang B, Wang XY, Jin B. Effect of local wound infiltration with ropivacaine on postoperative pain relief and stress response reduction after open hepatectomy. *World J Gastroenterol* 2017; 23(36): 6733-6740 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6733.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6733>

INTRODUCTION

Postoperative pain is an important issue for surgeons, anesthetists, patients, and other related personnel. The intensive pain caused by upper abdominal laparotomy may influence postoperative recovery, prolong hospitalization, and cause stress response and complications, including respiratory and cardiovascular depression and gastrointestinal and neuroendocrine dysfunction^[1,2]. Currently, intravenous analgesia or epidural analgesia (EA) with a patient-controlled analgesia pump is the most common analgesic approach for controlling postoperative pain after laparotomy^[3]. Although favored after major laparotomy, patient-controlled intravenous analgesia (PCIA) can delay postoperative recovery because of nausea and vomiting, excessive sedation, and dizziness; moreover, this analgesic mode involves the risk of addiction with large opioid dosages over long periods^[4,5]. As to EA, which provides better analgesia than PCIA, it is restricted by contraindications, epidural puncture failure, and side effects^[6,7]. Therefore, finding other analgesic strategies with fewer potentially serious adverse effects will be beneficial for patients suffering from postoperative pain.

Postoperative analgesia is a crucial section of perioperative management, and local anesthetic methods are more effective than systemic analgesia regardless of the operation type^[8]. Currently, wound infiltration with local anesthetics, which is a simple, effective, and inexpensive method, is performed in various surgical procedures and provides satisfactory analgesia without major side effects^[9]. Ropivacaine and bupivacaine, as long-acting local anesthetics, are commonly used for local anesthesia and pain management in China^[10]. Ropivacaine has the same analgesic effects as bupivacaine and levobupivacaine, but it is associated with a low incidence of motor block^[11]. Thus, ropivacaine appears to be an important component for local anesthesia and postoperative analgesia. Meanwhile, surgical pain frequently increases the systemic stress response during the perioperative period, which can induce the excessive release of catecholamines (epinephrine and norepinephrine) and cortisol. Optimistically, local anesthesia provides considerable advantages over general anesthesia by suppressing catecholamines and cortisol levels^[12].

In this study, we aimed to assess the effect of local wound infiltration with ropivacaine on postoperative pain control, mean arterial pressure (MAP), heart rate

(HR), cumulative sufentanil consumption, incidence of nausea and vomiting, time to bowel recovery, and length of hospitalization after open hepatectomy. The changes of three stress hormones, namely, epinephrine, norepinephrine, and cortisol, were evaluated in patients undergoing wound infiltration with and without ropivacaine.

MATERIALS AND METHODS

A total of 56 patients undergoing open hepatectomy, which was performed by the same experienced surgical team at the Department of Hepatobiliary Surgery of Qilu Hospital of Shandong University from January 2016 to March 2017, participated in this study. The study was approved by the Medical Ethics Committee of Qilu Hospital of Shandong University (No. 2017052), and written informed consent was obtained from all patients. The inclusion criteria included adult patients (aged 18–75 years) who would undergo open hepatectomy and were classified as grades I–III according to the American Society of Anesthesiologists (ASA) Physical Status Classification System. Patients with a history of known allergy to local anesthetics, chronic preoperative opioid consumption, or any other analgesic treatment for chronic pain before surgery, psychiatric or neurological diseases, or acquired or genetic hemostatic abnormality were excluded from the study.

On the day of surgery, the patients were randomly divided into two groups with a table of random numbers. Surgeons and patients were kept blinded to the assigned treatment groups throughout the study. Wound infiltration was performed with a 7.5 mg/mL ropivacaine solution in the ropivacaine group and with an isotonic saline solution in the control group. Solutions were prepared and provided by the anesthetist, and surgeons were blinded to patient allocation. When closing the abdomen at the end of the surgical procedure, 20 mL of the prearranged solution was used to infiltrate the subcutaneous tissues, deep muscular fascia, and parietal peritoneum. Moreover, one or two drainage tubes were routinely placed near the cutting surface of the liver and then pulled out and fixed on the abdominal skin. In the presence of tube incision or pulling of the tube during movement or when turning over, the surrounding tissues of the tube were also infiltrated with the solution. Infiltration was performed under direct vision by the surgeon. All patients were given unrestricted access to sufentanil through a 100 mL disposable patient-controlled analgesic (PCA) device containing 1 µg/mL sufentanil that was delivered at a rate of 2 µg/h and a bolus of 0.5 µg with a 15 min lockout time. When the skin was closed, the PCA pump was connected to the venous catheter and routinely removed 36 h after the operation.

The intensity of postoperative pain at rest was

measured on a visual analogue scale (VAS) graded from 0 (no pain) to 10 (very severe pain) for the first 48 h after surgery. Movement pain was scored using VAS when coughing or turning over. Pain scores were recorded both by nurses and surgeons blinded to patient allocation. Pain measurements were performed at 0, 6, 12, 24, and 48 h after the surgery. Other variables were recorded, including time to bowel recovery, length of hospitalization after surgery, hemodynamic data represented by MAP and HR, cumulative sufentanil consumption, and postoperative nausea and vomiting (PONV). The three surgical stress hormones, namely, epinephrine, norepinephrine, and cortisol, were detected using commercial enzyme-linked immunosorbent assay kits (Cusabio Biotech Co., Ltd., Wuhan, China). Time to bowel recovery was defined as the time to first anal exhaust. PONV was recorded with a three-point rating scale: 1, no nausea and vomiting; 2, nausea without vomiting; 3, nausea with vomiting.

Statistical analyses were performed with SPSS 19.0 (SPSS Inc., Chicago, IL, United States). All data were checked for normal distribution and the results are expressed as mean ± SD for continuous variables. The *t* test, χ^2 test, Fisher's exact test, or analysis of variance was carried out where appropriate. *P* < 0.05 was considered statistically significant.

RESULTS

All patients successfully received the surgical procedure, including the wound infiltration with a prearranged solution. However, three patients (two in the ropivacaine group and one in the control group) were dropped from the study for postoperative bleeding and bile leakage; finally, 26 patients were enrolled in the ropivacaine group and 27 enrolled in the control group. The demographic characteristics of the patients assigned to the two groups were comparable in terms of age, gender, weight, ASA grade, incision length, and postoperative pathology, except for operation type, which showed a statistical difference but had no clinical significance (Table 1).

The VAS scores both at rest and on movement were similar between the two groups at 24 h and 48 h after open hepatectomy (Figure 1A and B). Significant differences in VAS scores at rest were detected at 0 h (*P* = 0.0106), 6 h (*P* = 0.0032), and 12 h (*P* = 0.0002). Moreover, significant differences in VAS scores on movement were observed at 0 h (*P* = 0.0208), 6 h (*P* = 0.0043), and 12 h (*P* = 0.0089). The details are shown in Table 2.

Hemodynamic data are presented in Figure 1C and D and Table 2. MAP was significantly lower in the ropivacaine group than in the control group at 6 h (*P* = 0.0241), 12 h (*P* = 0.0001), and 24 h (*P* = 0.002). In the ropivacaine group, HR was significantly lower at 0 h (*P* = 0.0103), 6 h (*P* = 0.0087), 12 h (*P* < 0.0001), and

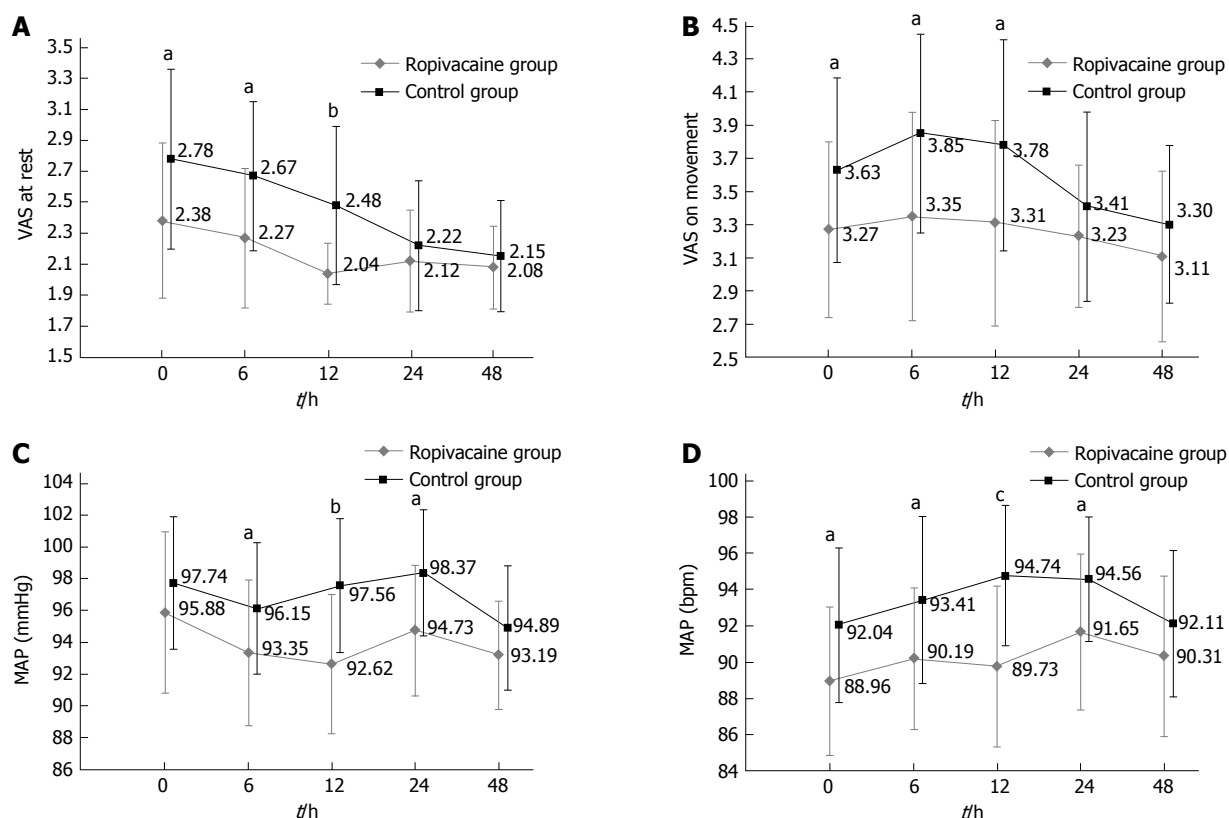


Figure 1 Visual analogue scale scores at rest and on movement, mean arterial pressure, and heart rate during the first 48 h after surgery. A: VAS scores at rest; B: VAS scores on movement; C: MAP; D: HR. ^a $P < 0.05$; ^b $P < 0.001$; ^c $P < 0.0001$. VAS: Visual analogue scale; MAP: Mean arterial pressure; HR: Heart rate.

Table 1 Demographic characteristics of the patients studied (mean \pm SD)

Characteristic	Ropivacaine group	Control group	t/χ^2	P value
Age (yr)	48.38 \pm 11.74	49.59 \pm 12.42	-0.36	0.7176
Gender				
Male/female	18/8	18/9	0.04	0.8415
Weight (kg)	63.04 \pm 9.21	66.04 \pm 9.86	-1.14	0.2583
ASA grade				
I/II/III	4/17/5	7/15/5	0.92	0.6298
Incision length (cm)	24.65 \pm 1.83	24.22 \pm 2.76	0.67	0.5075
Operation type				
Left hepatectomy	8	5		0.0086
Right hepatectomy	13	8		
Mesohepatectomy	1	0		
Caudate lobectomy	2	1		
Irregular hepatectomy	2	13		
Postoperative pathology				
Hepatocellular carcinoma	22	18		0.3292
Intra-and extrahepatic cholangiolithiasis	3	7		
Hepatic focal nodular hyperplasia	1	1		
Hepatocellular adenoma	0	1		

ASA: American Society of Anesthesiologists.

24 h ($P = 0.0089$).

No statistically significant difference was observed in baseline levels (0 h) of epinephrine, norepinephrine, or cortisol between the two groups. The levels of epinephrine at 24 and 48 h were significantly lower in the ropivacaine group than in the control group ($P = 0.0064$, $P = 0.0078$). Similarly, the values of

norepinephrine and cortisol at 24 and 48 h were significantly reduced in the ropivacaine group ($P < 0.0001$ for all), as shown in Figure 2A-C and Table 3.

Cumulative sufentanil consumption at 36 h after surgery is presented in Figure 2D and Table 2. The consumption was significantly lower in the ropivacaine group than in the control group at 6 h ($P = 0.022$),

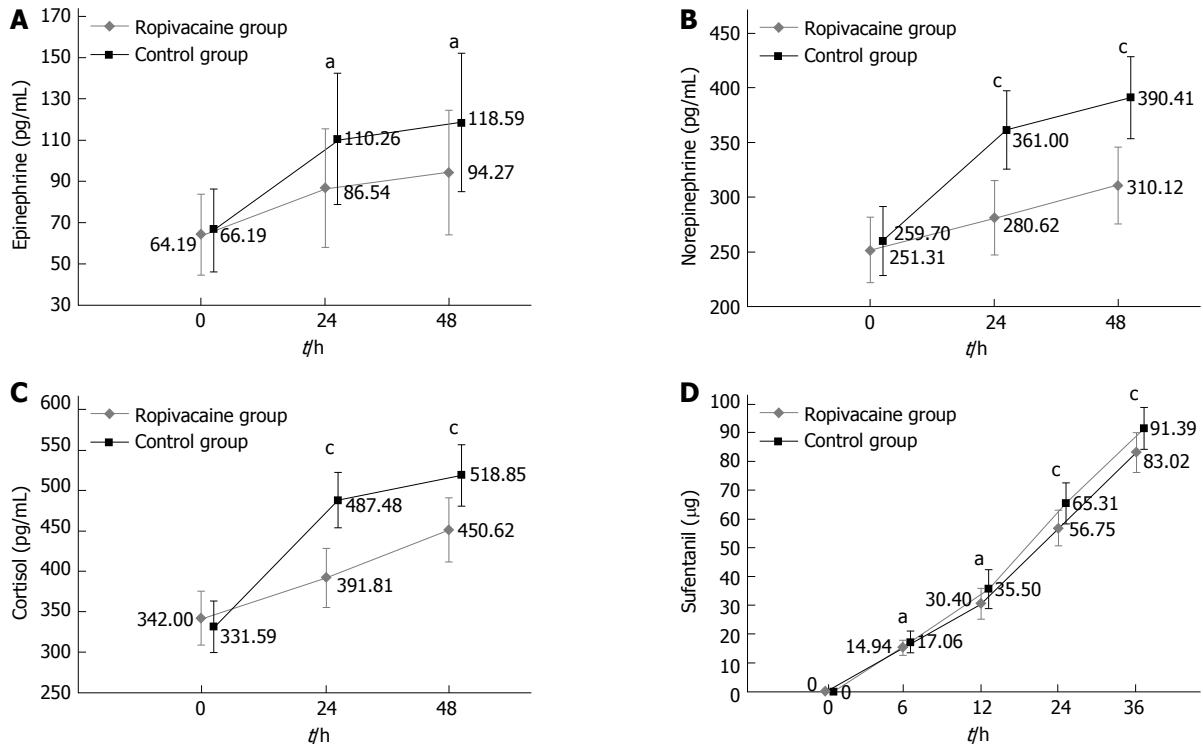


Figure 2 Plasma levels of epinephrine, norepinephrine and cortisol and cumulative sufentanil consumption during the first 48 h after surgery. A: Epinephrine; B: Norepinephrine; C: Cortisol; and D: Cumulative sufentanil consumption. ^a $P < 0.05$; ^c $P < 0.0001$.

Table 2 Visual analog scale scores at rest and on movement, mean arterial pressure, heart rate, and cumulative sufentanil consumption

Characteristic	0 h	6 h	12 h	24 h	48 h ¹
VAS at rest					
Ropivacaine group	2.38 ± 0.50	2.27 ± 0.45	2.04 ± 0.20	2.12 ± 0.33	2.08 ± 0.27
Control group	2.78 ± 0.58	2.67 ± 0.48	2.48 ± 0.51	2.22 ± 0.42	2.15 ± 0.36
<i>t</i>	-2.65	-3.17	-4.21	-1.03	-0.81
<i>P</i> value	0.0106	0.0032	0.0002	0.3096	0.4230
VAS on movement					
Ropivacaine group	3.27 ± 0.53	3.35 ± 0.63	3.31 ± 0.62	3.23 ± 0.43	3.11 ± 0.52
Control group	3.63 ± 0.56	3.85 ± 0.60	3.78 ± 0.64	3.41 ± 0.57	3.30 ± 0.47
<i>t</i>	-2.39	-2.99	-2.72	-1.27	-1.34
<i>P</i> value	0.0208	0.0043	0.0089	0.2110	0.1857
MAP (mmHg)					
Ropivacaine group	95.88 ± 5.08	93.35 ± 4.63	92.62 ± 4.43	94.73 ± 4.16	93.19 ± 3.41
Control group	97.74 ± 4.17	96.15 ± 4.14	97.56 ± 4.23	98.37 ± 3.99	94.89 ± 3.90
<i>t</i>	-1.46	-2.32	-4.16	-3.25	-1.69
<i>P</i> value	0.1515	0.0241	0.0001	0.0020	0.0981
HR (bpm)					
Ropivacaine group	88.96 ± 4.12	90.19 ± 3.92	89.73 ± 4.45	91.65 ± 4.30	90.31 ± 4.45
Control group	92.04 ± 4.27	93.41 ± 4.62	94.74 ± 3.88	94.56 ± 3.43	92.11 ± 4.05
<i>t</i>	-2.66	-2.73	-4.37	-2.72	-1.54
<i>P</i> value	0.0103	0.0087	< 0.0001	0.0089	0.1289
Cumulative sufentanil consumption (μg)					
Ropivacaine group	0	14.94 ± 2.56	30.40 ± 5.39	56.75 ± 6.20	83.02 ± 7.05
Control group	0	17.06 ± 3.81	35.50 ± 6.91	65.31 ± 7.09	91.39 ± 7.34
<i>t</i>		-2.36	-2.99	-4.67	-4.23
<i>P</i> value		0.0220	0.0043	< 0.0001	< 0.0001

¹As to cumulative sufentanil consumption, the time point was set at 36 h. VAS: Visual analog scale; MAP: Mean arterial pressure; HR: Heart rate.

12 h ($P = 0.0043$), 24 h ($P < 0.0001$), and 36 h ($P < 0.0001$). Even so, the incidence of nausea and vomiting, the side effects of sufentanil, between the

two groups had no significant difference (Table 4). Moreover, in the ropivacaine group, time to bowel recovery ($P = 0.0133$) and hospitalization after surgery

Table 3 Plasma concentrations of epinephrine, norepinephrine and cortisol (pg/mL)

Stress hormone	0 h	24 h	48 h
Epinephrine			
Ropivacaine group	64.19 ± 19.62	86.54 ± 28.64	94.27 ± 30.10
Control group	66.19 ± 20.30	110.26 ± 31.88	118.59 ± 33.65
<i>t</i>	-0.36	-2.85	-2.77
<i>P</i> value	0.7180	0.0064	0.0078
Norepinephrine			
Ropivacaine group	251.31 ± 30.19	280.62 ± 34.22	310.12 ± 35.15
Control group	259.70 ± 31.72	361.00 ± 36.06	390.41 ± 37.73
<i>t</i>	-0.99	-8.32	-8.01
<i>P</i> value	0.3286	< 0.0001	< 0.0001
Cortisol			
Ropivacaine group	342.00 ± 33.72	391.81 ± 36.53	450.62 ± 39.39
Control group	331.59 ± 31.92	487.48 ± 34.36	518.85 ± 38.21
<i>t</i>	1.15	-9.82	-6.40
<i>P</i> value	0.2537	< 0.0001	< 0.0001

Table 4 Time to bowel recovery, postoperative nausea and vomiting, and hospitalization length in the two groups

Characteristic	Ropivacaine group	Control group	<i>t/χ²</i>	<i>P</i> value
Time to bowel recovery (d)	3.15 ± 1.01	3.93 ± 1.17	-2.56	0.0133
PONA				
No PONA	7	3		0.2729
Nausea without vomiting	16	18		
Nausea with vomiting	3	6		
Hospitalization (d)	8.65 ± 2.43	10.52 ± 3.49	-2.25	0.0289

PONA: Postoperative nausea and vomiting.

(P = 0.0289) were significantly shortened (Table 4).

DISCUSSION

Laparoscopic hepatectomy is commonly adopted in clinical settings because of its many advantages, including little trauma, low pain, fast recovery, and short hospitalization; however, open hepatectomy remains irreplaceable, especially in the presence of lesions close to or invading the root of the hepatic veins or the inferior vena cava, history of previous hepatectomy or any previous surgery potentially causing severe adhesion around the liver, and concomitant cardiopulmonary disease^[13]. A right subcostal incision or reversed L-shaped incision (> 20 cm) is often made for open hepatectomy, and either of these two incision types is the most important source of postoperative pain. Thus, finding an effective way to reduce postoperative pain is urgent and necessary.

Local anesthetic wound infiltration is a useful and important component of a multimodality approach to postoperative pain control, and it can be applied in many types of surgery, including lumbar spine surgery,

breast surgery, and inguinal hernia repair^[9,14,15]. Local anesthetics used in the wound can block parietal afferents, reduce the sensitization of spinal dorsal horn neurons, and provide analgesia by inhibiting the transmission of noxious impulses from the incision^[16]. Moreover, local anesthetics can suppress local inflammatory responses to incision injury that could sensitize nociceptive receptors and contribute to hyperalgesia^[17]. Ropivacaine, a pure levorotatory stereoisomer and long-acting amide local anesthetic agent, has been widely used for local anesthesia and postoperative analgesia, and its reduced lipophilicity is associated with decreased incidence of central nervous system toxicity and cardiotoxicity^[18]. Postoperative pain comes from superficial structures and deep muscular-peritoneal components; therefore, ropivacaine infiltrated not only the subcutaneous tissues but also the parietal peritoneum and deep muscular fasciae in our study. Our results showed that in the first 12 h after surgery, the local anesthetic ropivacaine significantly relieved the pain intensity at rest and on movement, demonstrating the potential of local wound infiltration with ropivacaine as a reliable analgesic strategy after open hepatectomy.

Surgical stress could cause a spectrum of changes in the body, involving the neuroendocrine, metabolic, immunological, and hematological systems^[12]. The body's surgical stress response is mainly determined by the surgical wound severity, including the length of the incision in the abdominal wall from the skin to the parietal peritoneum^[19]. The incision of open hepatectomy often exceeds 20 cm, and the surgical stress is thought to be high. Therefore, using local anesthetics to block surgical stress is feasible. Surgical stress response to injury causes a series of hormone changes; moreover, catecholamines (epinephrine and norepinephrine) and cortisol, as the main and most reliable peripheral hormones, correlate well with the extent of surgical stress. In this study, surgical stress was significantly reduced in the first 48 h after surgery as revealed by the levels of epinephrine, norepinephrine, and cortisol. Changes in MAP and HR were recorded, and the results demonstrated that the indexes of the ropivacaine group were obviously decreased. These results indicate that local wound infiltration with ropivacaine could also reduce surgical stress responses.

Opioids are commonly used for postoperative analgesia *via* venous access. However, they are associated with a potential risk of addiction, especially in large doses over long periods. Moreover, opioids possess potentially serious side effects, such as nausea, vomiting, constipation, respiratory depression, excessive sedation, and liver function impairment; hence, sparing opioids may reduce the incidence of the above side effects^[20]. Reducing the dosage and duration of opioid usage is regarded suitable for avoiding

potentially serious adverse effects. Wound infiltration, as part of an opioid-sparing, multimodal analgesic regime, should therefore be recommended. Our current study showed that cumulative sufentanil consumption was significantly reduced in the ropivacaine group. Moreover, the time to bowel recovery was shorter in the ropivacaine group than in the control group. This may be caused by a combination of several reasons. First, sufentanil inhibits gut motility and propulsive activity by combining the μ -2 and κ receptors in the digestive tract^[21]. Second, a previous animal study demonstrated that catecholamines reduce gut motility^[22] and that the level of catecholamines in the ropivacaine group was reduced. Finally, ropivacaine could accelerate postoperative intestinal motility by reducing the inflammatory response.

The current study may have some limitations. The sample size was relatively small, and thus, more patients are needed in future studies to confirm our results. Compared with a previous study using catheters as a continuous wound infiltration method to deliver ropivacaine into the wound^[23], we used single-shot ropivacaine infiltration into the superficial and deep muscular-peritoneal layers to achieve an analgesic effect. In our study, the drainage tube was routinely placed beside the liver resection surface and fixed outside. Movement and turning over could drag the tube and cause intensive pain, and thus, infiltration around the tube was highly effective. Moreover, the catheter under the wound could bring potential risks, such as infection and delayed wound healing, and the delivery rates and volumes of local anesthetics remain unidentified. Thus, single-shot infiltration with ropivacaine is a simple, convenient, and effective analgesic method that can bring short-term benefits for patients who underwent open hepatectomy. A previous study suggested that local anesthesia and stress response reduction could decrease cancer formation and that local anesthesia and analgesia may improve overall patient survival after oncologic surgery^[24]. Thus, future research about local anesthesia, tumor recurrence, and patient survival after open hepatectomy is required.

In conclusion, local wound infiltration with ropivacaine after open hepatectomy can decrease acute postoperative pain and surgical stress response. This simple, convenient, and effective analgesic method provides postoperative analgesia and short-term benefits after open hepatectomy.

COMMENTS

Background

The postoperative pain caused by laparotomy delays patients' recovery and incurs stress response. Although commonly used to control pain, intravenous analgesia and epidural analgesia still have their contraindications and side effects. Local wound infiltration is a simple and effective method that can provide satisfactory analgesia without major side effects. The current study was designed to evaluate the effect of local wound infiltration with ropivacaine on

postoperative pain and stress response after open hepatectomy.

Research frontiers

Postoperative analgesia is an indispensable component of fast track surgery for surgical patients, especially those who undergo laparotomy. Local anesthetics can effectively provide analgesia by inhibiting the transmission of noxious impulses from the wound and suppress local inflammatory responses to wound injury.

Innovations and breakthroughs

Wound infiltration with ropivacaine could provide effective analgesia in the first 48 h after open hepatectomy, with lower mean arterial pressure, heart rate and sufentanil consumption, accelerated postoperative recovery, and reduced stress response. These results suggest that this method is a simple, convenient and effective analgesic method that can provide postoperative analgesia and short-term benefits after open hepatectomy.

Applications

This study provides additional evidence supporting that local wound infiltration with ropivacaine after open hepatectomy can improve postoperative pain relief, reduce surgical stress response, and accelerate postoperative recovery.

Terminology

The intensity of postoperative pain at rest was measured on a visual analogue scale graded from 0 (no pain) to 10 (very severe pain) after surgery. Postoperative nausea and vomiting was recorded with a three-point rating scale: 1, no nausea and vomiting; 2, nausea without vomiting; 3, nausea with vomiting.

Peer-review

The study was well written and its findings are informative. Local wound infiltration with ropivacaine has good effects for pain relief and stress response reduction after open hepatectomy.

REFERENCES

- 1 **Wightman JA.** A prospective survey of the incidence of postoperative pulmonary complications. *Br J Surg* 1968; **55**: 85-91 [PMID: 5635926]
- 2 **Latimer RG, Dickman M, Day WC, Gunn ML, Schmidt CD.** Ventilatory patterns and pulmonary complications after upper abdominal surgery determined by preoperative and postoperative computerized spirometry and blood gas analysis. *Am J Surg* 1971; **122**: 622-632 [PMID: 4939329]
- 3 **Zhu Z, Wang C, Xu C, Cai Q.** Influence of patient-controlled epidural analgesia versus patient-controlled intravenous analgesia on postoperative pain control and recovery after gastrectomy for gastric cancer: a prospective randomized trial. *Gastric Cancer* 2013; **16**: 193-200 [PMID: 22806415 DOI: 10.1007/s10120-012-0168-z]
- 4 **Hankin CS, Schein J, Clark JA, Panchal S.** Adverse events involving intravenous patient-controlled analgesia. *Am J Health Syst Pharm* 2007; **64**: 1492-1499 [PMID: 17617499 DOI: 10.2146/ajhp060220]
- 5 **Choi JB, Shim YH, Lee YW, Lee JS, Choi JR, Chang CH.** Incidence and risk factors of postoperative nausea and vomiting in patients with fentanyl-based intravenous patient-controlled analgesia and single antiemetic prophylaxis. *Yonsei Med J* 2014; **55**: 1430-1435 [PMID: 25048507 DOI: 10.3349/ymj.2014.55.5.1430]
- 6 **Mohta M, Ophrii LE, Agarwal D, Bhatt S, Sethi AK, Chilkoti G.** Vocal cord palsy: an unusual complication of paravertebral block. *Anaesth Intensive Care* 2011; **39**: 969-971 [PMID: 21970149]
- 7 **Lucas SD, Higdon T, Boezaart AP.** Unintended epidural placement of a thoracic paravertebral catheter in a patient with severe chest trauma. *Pain Med* 2011; **12**: 1284-1289 [PMID: 21714843 DOI: 10.1111/j.1526-4637.2011.01180.x]

- 8 **Wu CL**, Cohen SR, Richman JM, Rowlingson AJ, Courpas GE, Cheung K, Lin EE, Liu SS. Efficacy of postoperative patient-controlled and continuous infusion epidural analgesia versus intravenous patient-controlled analgesia with opioids: a meta-analysis. *Anesthesiology* 2005; **103**: 1079-1088; quiz 1109-1110 [PMID: 16249683]
- 9 **Scott NB**. Wound infiltration for surgery. *Anaesthesia* 2010; **65** Suppl 1: 67-75 [PMID: 20377548 DOI: 10.1111/j.1365-2044.2010.06241.x]
- 10 **Tam KW**, Chen SY, Huang TW, Lin CC, Su CM, Li CL, Ho YS, Wang WY, Wu CH. Effect of wound infiltration with ropivacaine or bupivacaine analgesia in breast cancer surgery: A meta-analysis of randomized controlled trials. *Int J Surg* 2015; **22**: 79-85 [PMID: 26277531 DOI: 10.1016/j.ijsu.2015.07.715]
- 11 **Li M**, Wan L, Mei W, Tian Y. Update on the clinical utility and practical use of ropivacaine in Chinese patients. *Drug Des Devel Ther* 2014; **8**: 1269-1276 [PMID: 25246768 DOI: 10.2147/DDDT.S57258]
- 12 **Iwasaki M**, Edmondson M, Sakamoto A, Ma D. Anesthesia, surgical stress, and “long-term” outcomes. *Acta Anaesthesiol Taiwan* 2015; **53**: 99-104 [PMID: 26235899 DOI: 10.1016/j.aat.2015.07.002]
- 13 **Kawaguchi Y**, Otsuka Y, Kaneko H, Nagai M, Nomura Y, Yamamoto M, Otani M, Ohashi Y, Sugawara K, Koike D, Ishida T, Kokudo N, Tanaka N. Comparisons of financial and short-term outcomes between laparoscopic and open hepatectomy: benefits for patients and hospitals. *Surg Today* 2016; **46**: 535-542 [PMID: 26021453 DOI: 10.1007/s00595-015-1189-0]
- 14 **Byager N**, Hansen MS, Mathiesen O, Dahl JB. The analgesic effect of wound infiltration with local anaesthetics after breast surgery: a qualitative systematic review. *Acta Anaesthesiol Scand* 2014; **58**: 402-410 [PMID: 24617619 DOI: 10.1111/aas.12287]
- 15 **Kjærgaard M**, Moiniche S, Olsen KS. Wound infiltration with local anesthetics for post-operative pain relief in lumbar spine surgery: a systematic review. *Acta Anaesthesiol Scand* 2012; **56**: 282-290 [PMID: 22260370 DOI: 10.1111/j.1399-6576.2011.02629.x]
- 16 **Brennan TJ**, Zahn PK, Pogatzki-Zahn EM. Mechanisms of incisional pain. *Anesthesiol Clin North America* 2005; **23**: 1-20 [PMID: 15763408 DOI: 10.1016/j.atc.2004.11.009]
- 17 **Kawamata M**, Takahashi T, Kozuka Y, Nawa Y, Nishikawa K, Narimatsu E, Watanabe H, Namiki A. Experimental incision-induced pain in human skin: effects of systemic lidocaine on flare formation and hyperalgesia. *Pain* 2002; **100**: 77-89 [PMID: 12435461]
- 18 **Kuthiala G**, Chaudhary G. Ropivacaine: A review of its pharmacology and clinical use. *Indian J Anaesth* 2011; **55**: 104-110 [PMID: 21712863 DOI: 10.4103/0019-5049.79875]
- 19 **Krikri A**, Alexopoulos V, Zoumakis E, Katsaronis P, Balafas E, Kouraklis G, Karayannacos PE, Chrousos GP, Skalkas G. Laparoscopic vs. open abdominal surgery in male pigs: marked differences in cortisol and catecholamine response depending on the size of surgical incision. *Hormones (Athens)* 2013; **12**: 283-291 [PMID: 23933697]
- 20 **White PF**. The changing role of non-opioid analgesic techniques in the management of postoperative pain. *Anesth Analg* 2005; **101**: S5-S22 [PMID: 16334489]
- 21 **Ducrotté P**, Caussé C. The Bowel Function Index: a new validated scale for assessing opioid-induced constipation. *Curr Med Res Opin* 2012; **28**: 457-466 [PMID: 22236136 DOI: 10.1185/03007995.2012.657301]
- 22 **Fruhwald S**, Herk E, Petnehazy T, Scheidl S, Holzer P, Hammer F, Metzler H. Sufentanil potentiates the inhibitory effect of epinephrine on intestinal motility. *Intensive Care Med* 2002; **28**: 74-80 [PMID: 11819004 DOI: 10.1007/s00134-001-1167-4]
- 23 **Xin Y**, Hong Y, Yong LZ. Efficacy of postoperative continuous wound infiltration with local anesthesia after open hepatectomy. *Clin J Pain* 2014; **30**: 571-576 [PMID: 24281275 DOI: 10.1097/AJP.0000000000000032]
- 24 **Cata JP**, Hernandez M, Lewis VO, Kurz A. Can regional anesthesia and analgesia prolong cancer survival after orthopaedic oncologic surgery? *Clin Orthop Relat Res* 2014; **472**: 1434-1441 [PMID: 24081665 DOI: 10.1007/s11999-013-3306-y]

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Pedicated omental patch as a bridging procedure for iatrogenic bile duct injury

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Abstract

Iatrogenic bile duct injuries during cholecystectomy can present as fulminant intra-abdominal sepsis which precludes immediate repair or biliary reconstruction. We report the case of a 29-year-old female patient who sustained a bile duct injury after an open cholecystectomy in a neighboring country. She presented to our institution 22 d after initial surgery with septic shock and multiple intra-abdominal collections. Endoscopic retrograde cholangiography revealed a large common hepatic duct defect corresponding to a Strasberg type D bile duct injury. Definitive reconstruction such as a hepaticojejunostomy cannot be performed due to the presence of dense adhesions with infected and friable tissues. She underwent a combination of endoscopic biliary stenting and pedicled omental patch repair of the bile duct to control bile leak and sepsis as a bridging procedure to definite hepaticojejunostomy three months later.

Key words: Cholecystectomy; Endoscopic retrograde cholangiopancreatography; Case reports; Bile ducts; Abdominal abscess

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Core tip: Iatrogenic bile duct injury is a challenging condition to treat. This case report describes a novel and innovative surgical technique, whereby a pedicled omental patch repair was performed as a bridging procedure to definitive repair such as a hepaticojejunostomy, in a patient who presented in a delayed fashion with severe intra-abdominal sepsis.

Ng JJ, Kow AWC. Pedicled omental patch as a bridging procedure for iatrogenic bile duct injury. *World J Gastroenterol* 2017; 23(36): 6741-6746 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v23/i36/6741.htm> DOI: <http://dx.doi.org/10.3748/wjg.v23.i36.6741>

INTRODUCTION

Iatrogenic bile duct injuries (BDIs) are uncommon but can lead to significant morbidity and mortality in patients who had undergone cholecystectomy. Although the incidence of BDIs in LC was reported to be almost tenfold that of OC during the 1990s, LC has now become the gold standard in the treatment of gallstone disease, with a low and declining incidence of BDIs^[1,2]. Several studies have reported the rate of BDIs during LC to be between 0.4% to 0.7%, but a more recent retrospective review of more than 10000 patients who had undergone LC showed that the incidence of BDIs was 0.2%, which was comparable to that of OC^[3-5]. Approximately one-third of BDIs is diagnosed intra-operatively and are amenable to immediate repair or reconstruction. However, the majority of BDIs are recognized only in the post-operative period with varying clinical presentations such as abdominal pain, jaundice, bile peritonitis or shock^[6-8]. BDIs can be classified using the Strasberg classification, with most BDIs manifesting as biliary leaks (Strasberg type A) that can be managed using endoscopic techniques^[6]. The other large group of BDIs are common bile duct or common hepatic duct transections (Strasberg type E1 to E3) which usually warrants surgical repair^[9]. A tension-free Roux-en-Y hepaticojejunostomy is the preferred surgical procedure for reconstitution of bilio-enteric continuity in major common bile duct or common hepatic duct transections, but rarely, in some patients, the presence of severe intra-abdominal sepsis precludes immediate biliary reconstruction^[8]. We report a novel method of using a pedicled omental patch, combined with endoscopic biliary stenting, as a bridging procedure to control bile leak, in a patient with severe intra-abdominal sepsis from a large common hepatic duct transection after OC, before attempting definitive hepaticojejunostomy.

CASE REPORT

A 29-year-old female underwent an open cholecystectomy for acute calculous cholecystitis in a hospital from a neighbouring country. Exact intra-operative findings and details of her post-operative recovery were not made available to us. She developed obstructive jaundice on post-operative day three with bilious effluent from her abdominal drain on post-operative day seven. She was initially treated expectantly but due to persistent high bilious output from her abdominal drain, she underwent an exploratory laparotomy on post-operative day 16. The surgeons noted a collapsed common bile duct, but were not able to locate the exact site of bile leak. She was subsequently transferred to our institution on post-operative day 22 for further management for further management of her bile leak.

Upon presentation to our institution, the patient

was in septic shock. There was active leakage of bilious fluid from her a right subcostal incision. Her abdomen was distended with generalized tenderness. She was resuscitated and started on broad spectrum antibiotics. Laboratory investigations revealed elevated infective markers with a white cell count of $19 \times 10^9/L$ and a C-reactive protein level of 48 mg/L. Serum bilirubin was normal. Computed tomography scan of the abdomen revealed a 3 cm biloma at the gallbladder fossa (Figure 1A) with multiple rim-enhancing abdominal collections around the upper abdomen (Figure 1B) - the largest being a 9.3 cm \times 8.5 cm perisplenic collection (Figure 1C). Endoscopic retrograde cholangiography performed on post-operative day 23 revealed a large common hepatic duct defect just below the bifurcation, corresponding to a Strasberg type D BDI with contrast seen extravasating into the sub-hepatic recess (Figure 2A). After much difficulty, guide-wires were placed across the bile duct defect and plastic biliary stents were deployed into the left and right hepatic ducts respectively (Figure 2B).

In view of persistent sepsis with the presence of multiple intra-abdominal collections, a decision to perform damage control surgery was taken. At the exploratory laparotomy performed on post-operative day 24, a large 2 cm defect at the anterolateral aspect of the common hepatic duct with biliary stents visible from within was found (Figure 3A). There was a large perisplenic abscess with presence of turbid bile and debris in the sub-hepatic recess which was drained and washed out thoroughly. Neither primary repair of the biliary defect nor biliary reconstruction with a hepaticojejunostomy were suitable options due to dense adhesions around the hepatic hilum with infected and friable surrounding tissues. The biliary drains were unable to completely exclude the biliary system entirely due to the size of the biliary defect. Hence, a well-vascularized and healthy pedicled omental patch was harvested and tagged down circumferentially to the biliary defect using absorbable sutures (Figure 3B). An illustration of the pedicled omental patch is as demonstrated (Figure 4). Drains were placed and the abdomen was closed primarily. The patient improved after surgery with resolution of pyrexia and sepsis. Abdominal drains were removed within eight days of surgery. Laboratory infective markers normalized and serum bilirubin levels remained normal. Intra-abdominal cultures grew *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* and the patient was sent home to complete four weeks of intravenous antibiotics.

A repeat computed tomography scan of the abdomen three months later revealed complete resolution of the intra-abdominal collections. Although the biliary stents were in-situ, there was mild intrahepatic biliary dilatation suggesting the possibility of a common hepatic duct stricture. The patient subsequently underwent creation of a hepaticojejunostomy one week later. Intra-operatively, there were dense adhesions around the porta hepatis with evidence of early stricture

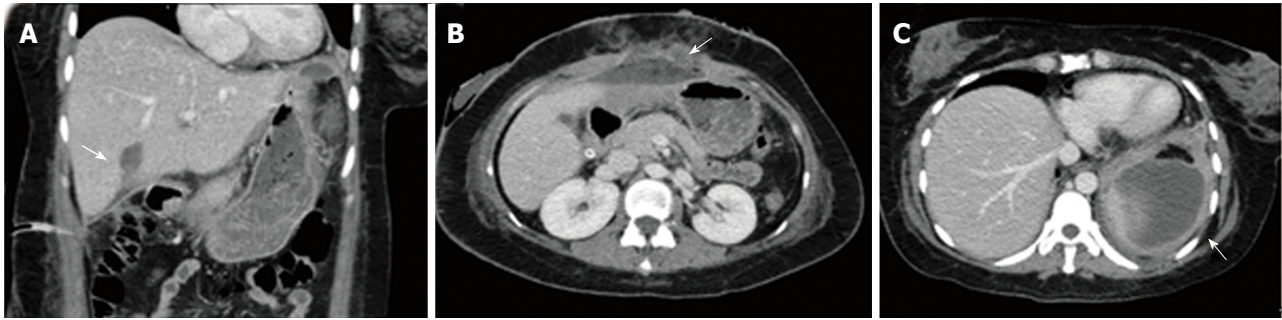


Figure 1 Computed tomography scan. A: Initial computed tomography scan of the abdomen revealing a 3 cm biloma at the gallbladder fossa with bile tracking into the sub-hepatic recess; B: Multiple rim-enhancing smaller intra-abdominal collections were also present in the upper abdomen; C: The largest intra-abdominal collection was a 9.3 cm × 8.5 cm perisplenic collection.

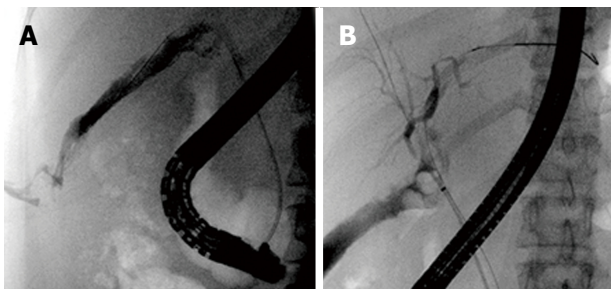


Figure 2 Endoscopic retrograde cholangiography. A: Endoscopic retrograde cholangiography revealed a large Strasberg Type D common hepatic duct defect with contrast seen immediately in the sub-hepatic recess; B: Plastic biliary stents inserted across the biliary defect into the left and right hepatic ducts.

formation at the previous site of BDI. The patient had an uneventful recovery after surgery and was discharged on post-operative day five.

DISCUSSION

BDIs rarely present in such a delayed fashion with fulminant intra-abdominal sepsis. The initial management of BDIs especially for patients that present late in the post-operative period is directed at delineating the type and extent of BDI, controlling sepsis and ongoing bile leak^[8]. We describe a technique that can be utilized as a bridge to definitive bilioenteric reconstruction such as a hepaticojejunostomy when prevailing conditions such as severe intra-abdominal sepsis renders immediate or early reconstruction unfavourable and not ideal. Our technique fulfils the above-mentioned principles of BDI management - first by using endoscopic retrograde cholangiogram to characterize the BDI with placement of biliary stents endoscopically to help control the bile leak. The stents will negate the function of the sphincter of Oddi as well as direct bile into the duodenum. Subsequent laparotomy will allow source control of sepsis *via* drainage of intra-abdominal abscesses whilst the placement of a pedicled omental patch will ensure proper sealing of the biliary defect and

exclusion of the biliary system from the peritoneal cavity. This restores continuity of bile flow, allows for resolution of intra-abdominal sepsis, inflammation and fibrosis, after which definitive bilioenteric reconstruction can be performed at an elective setting as the presence of peritonitis has been reported to confer a poorer outcome in patients undergoing biliary reconstruction^[10].

The use of omentum as an autologous graft to seal, patch or reinforce tissues during surgery is widely practiced, and its ability to support and adhere to local tissue is due to its abundant blood supply, angiogenic activity, innate immune function and high concentration of tissue factor^[11,12]. Moreover, animal studies have shown that the use of a pedicled omental flap in tissue reconstruction confers an anti-inflammatory effect, which can lead to an improved healing response^[13].

Omentum has been commonly used as a patch to close perforations in the gastro-intestinal tract such as a perforated duodenal ulcer. There have been several reports describing the use of omentum in biliary reconstruction. Meissner described his technique of "T-tube stented omentoplasty" used to successfully treat a common hepatic duct stricture. After incising the strictured segment of bile duct resulting in a 4 cm non-circumferential biliary defect, a large bore T-tube was placed within and a pedicled omental flap was then sutured down over the horizontal limb of the T-tube to bridge any remnant biliary defect as well as to envelop the vertical limb. The T-tube was removed after 6 mo with no sequelae and repeat ERC 19 years later showed normal biliary anatomy^[14]. Ebata *et al*^[15] also described a similar technique of "hilar cholangioplasty" where he used a pedicled omental flap with a 16F T-tube to seal a 2 cm × 3 cm ductal defect due to a bilio-biliary fistula. T-tube cholangioscopy 1 mo after surgery revealed a yellowish polypoid mass at the previous ductal defect and a biopsy taken revealed normal biliary epithelium lining the surface of the grafted omentum. The patient was followed up for 3 years without evidence of cholangitis or abnormal liver function. Another similar technique was described

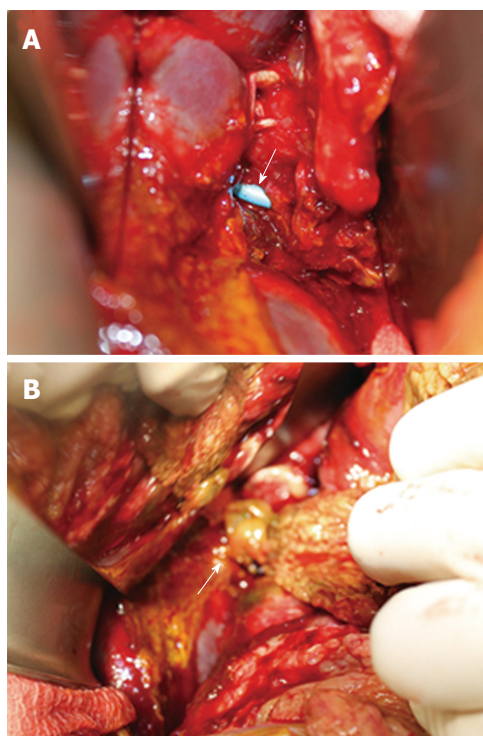


Figure 3 Damage control surgery. A: At the time of exploratory laparotomy, endoscopically placed biliary stents were visible within a large 2 cm anterolateral common hepatic duct defect; B: A pedicled omental patch was harvested and secured to the biliary defect using absorbable sutures.

by Chang but he used the falciform ligament instead of the omentum to patch a large bile duct defect due clip necrosis after laparoscopic cholecystectomy^[16]. Some authors have also used pedicled omental flaps to reinforce high risk biliary anastomoses in liver transplantation with good results^[17]. These examples, although limited to individual case reports reaffirms the ideal properties of omentum that make it a suitable autologous graft to aid in biliary reconstruction.

We postulate that our technique could also have been used as a stand-alone procedure without the need of a subsequent hepaticojejunostomy. The biliary stents could then be retrieved endoscopically after 10 to 12 wk. The above-mentioned case reports have shown good long-term patency of the reconstructed bile duct but established data is lacking. A hepaticojejunostomy would still confer the best long-term outcomes, especially in our patient who is young^[18]. Using our technique as a stand-alone procedure remains experimental and may be suitable in patients who cannot tolerate longer periods of general anaesthesia needed in a hepaticojejunostomy. Intensive follow-up would be necessary for such patients due to the risk of biliary strictures. We performed a definitive hepaticojejunostomy for our patient as she was due to return to a rural area with no access to regular follow-up for the development of biliary strictures.

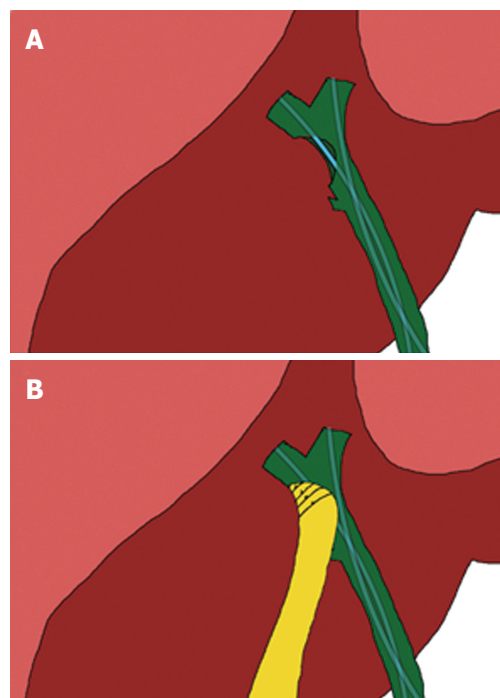


Figure 4 An illustration of the pedicled omental patch. A: An illustration showing the location of the common hepatic duct defect and endoscopic biliary stents placed across; B: The harvested pedicled omental patch was placed over the biliary defect and secured using absorbable sutures that run through the anterior and posterior margins of the biliary defect.

Alternative management strategies that could be deployed in our patient would be the placement of percutaneous trans-hepatic biliary drainage tubes or endoscopic biliary stents, combined with either percutaneous or surgically inserted intra-abdominal drains. This might be feasible if the ductal defect was small. However, if the ductal defect is large, the bile leak not be controlled leading to persistent biliary peritonitis and the development of intra-abdominal collections. Moreover, the drains would have to be placed on active suction for a significant period of time to be able to successfully divert and control the bile leak. More often than not, these drains would have to be kept for weeks to months until the patient undergoes definitive bilioenteric reconstruction. In a patient like ours who present with uncontrolled bile leak, intra-abdominal collections and sepsis, the utilization of our technique would allow for control of bile leak without any other additional percutaneous procedures or prolonged drain placement. Another management strategy that could be considered would be use of a T-tube in combination with an omental pedicled flap similar to techniques described above. We were, however, with the assistance of an experienced operator, able to cannulate the left and right hepatic ducts individually and place biliary stents across the biliary defect obviating the need of a T-tube.

Lastly, and most importantly, our case highlights the

need for early referral to an institution with experienced hepatobiliary surgeons, endoscopists, or interventional radiologists once a BDI is diagnosed. It is irrefutable evidence that best outcomes are obtained when first repair is performed in experienced tertiary centres^[18,19].

This case highlights the need for early referral of BDIs to specialist tertiary centres for further management once detected. Immediate repair or biliary reconstruction is contraindicated in patients with BDIs who present in a delayed fashion with severe intra-abdominal sepsis. We describe an alternative technique which combines endoscopic biliary stenting with a pedicled omental patch as a bridging procedure to definitive hepaticojejunostomy in this group of patients.

COMMENTS

Case characteristics

A 29-year-old female presented with septic shock and generalized abdominal pain following an open cholecystectomy performed three weeks ago in another hospital.

Clinical diagnosis

Physical examination revealed generalized abdominal tenderness and leakage of bile stained fluid from a right subcostal incision.

Differential diagnosis

Bile duct injury, duodenal injury, small bowel injury.

Laboratory diagnosis

Initial laboratory investigations revealed an elevated total white cell count and C-reactive protein levels.

Imaging diagnosis

Computed tomography scan of the abdomen revealed multiple large intra-abdominal collections. Endoscopic retrograde cholangiography revealed a large common hepatic duct defect (Strasberg type D bile duct injury).

Treatment

An exploratory laparotomy with a pedicled omental patch repair of the common hepatic duct was performed initially, followed by a definitive hepaticojejunostomy three months later.

Related reports

This is the first report describing the use of a pedicled omental patch combined with biliary stenting for treatment of a large bile duct injury. Two other case reports have described the use of a pedicled omental patch in conjunction with a T tube for similar bile duct injuries.

Experiences and lessons

In patients who present with severe intra-abdominal sepsis after bile duct injury, up-front creation of a definitive hepaticojejunostomy may not be possible. Instead, a pedicled omental patch repair of the biliary defect may be performed as a bridging procedure to a hepaticojejunostomy, or even as a stand-alone procedure.

Peer-review

The authors presented a novel and alternative technique by using a pedicled omental patch repair as a bridge to a definitive procedure in bile duct injuries.

REFERENCES

- Dolan JP**, Diggs BS, Sheppard BC, Hunter JG. Ten-year trend in the national volume of bile duct injuries requiring operative repair. *Surg Endosc* 2005; **19**: 967-973 [PMID: 15920680 DOI: 10.1007/s00464-004-8942-6]
- McPartland KJ**, Pomposelli JJ. Iatrogenic biliary injuries: classification, identification, and management. *Surg Clin North Am* 2008; **88**: 1329-1343; ix [PMID: 18992598 DOI: 10.1016/j.suc.2008.07.006]
- Adamsen S**, Hansen OH, Funch-Jensen P, Schulze S, Stage JG, Wara P. Bile duct injury during laparoscopic cholecystectomy: a prospective nationwide series. *J Am Coll Surg* 1997; **184**: 571-578 [PMID: 9179112]
- Flum DR**, Cheadle A, Prela C, Dellinger EP, Chan L. Bile duct injury during cholecystectomy and survival in medicare beneficiaries. *JAMA* 2003; **290**: 2168-2173 [PMID: 14570952 DOI: 10.1001/jama.290.16.2168]
- Pekolj J**, Alvarez FA, Palavecino M, Sánchez Clariá R, Mazza O, de Santibañes E. Intraoperative management and repair of bile duct injuries sustained during 10,123 laparoscopic cholecystectomies in a high-volume referral center. *J Am Coll Surg* 2013; **216**: 894-901 [PMID: 23518251 DOI: 10.1016/j.jamcollsurg.2013.01.051]
- Strasberg SM**, Hertl M, Soper NJ. An analysis of the problem of biliary injury during laparoscopic cholecystectomy. *J Am Coll Surg* 1995; **180**: 101-125 [PMID: 8000648]
- Sicklick JK**, Camp MS, Lillemoe KD, Melton GB, Yeo CJ, Campbell KA, Talamini MA, Pitt HA, Coleman J, Sauter PA, Cameron JL. Surgical management of bile duct injuries sustained during laparoscopic cholecystectomy: perioperative results in 200 patients. *Ann Surg* 2005; **241**: 786-792; discussion 793-795 [PMID: 15849514]
- Lau WY**, Lai EC, Lau SH. Management of bile duct injury after laparoscopic cholecystectomy: a review. *ANZ J Surg* 2010; **80**: 75-81 [PMID: 20575884 DOI: 10.1111/j.1445-2197.2009.05205.x]
- Pitt HA**, Sherman S, Johnson MS, Hollenbeck AN, Lee J, Daum MR, Lillemoe KD, Lehman GA. Improved outcomes of bile duct injuries in the 21st century. *Ann Surg* 2013; **258**: 490-499 [PMID: 24022441 DOI: 10.1097/SLA.0b013e3182a1b25b]
- Goykhman Y**, Kory I, Small R, Kessler A, Klausner JM, Nakache R, Ben-Haim M. Long-term outcome and risk factors of failure after bile duct injury repair. *J Gastrointest Surg* 2008; **12**: 1412-1417 [PMID: 18493825 DOI: 10.1007/s11605-008-0538-3]
- Goldsmith HS**. The omentum: Research and clinical applications. Springer-Verlag, New York, 1990
- Logmans A**, Schoenmakers CH, Haensel SM, Koolhoven I, Trimbois JB, van Lent M, van Ingen HE. High tissue factor concentration in the omentum, a possible cause of its hemostatic properties. *Eur J Clin Invest* 1996; **26**: 82-83 [PMID: 8682161]
- Uchibori T**, Takanari K, Hashizume R, Amoroso NJ, Kamei Y, Wagner WR. Use of a pedicled omental flap to reduce inflammation and vascularize an abdominal wall patch. *J Surg Res* 2017; **212**: 77-85 [PMID: 28550925 DOI: 10.1016/j.jss.2016.11.052]
- Meissner K**. Successful repair of Bismuth type 2 bile duct stricture by T-tube stented omentoplasty: 19 years follow-up. *HPB (Oxford)* 2001; **3**: 241-244 [PMID: 18333023 DOI: 10.1080/136518201752422299]
- Ebata T**, Takagi K, Nagino M. Hilar cholangioplasty using omentum for ductal defect in biliobiliary fistula. *J Hepatobiliary Pancreat Sci* 2011; **18**: 458-462 [PMID: 20886358 DOI: 10.1007/s00534-010-0332-y]
- Chang EG**. Repair of common bile duct injury with the round and falciform ligament after clip necrosis: case report. *JSLs* 2000; **4**: 163-165 [PMID: 10917125]
- Ye QF**, Niu Y, She XG, Ming YZ, Cheng K, Ma Y, Ren ZH. Pedicled greater omentum flap for preventing bile leak in liver transplantation patients with poor biliary tract conditions.

Hepatobiliary Pancreat Dis Int 2007; **6**: 470-473 [PMID: 17897907]

- 18 **Stewart L**, Way LW. Bile duct injuries during laparoscopic cholecystectomy. Factors that influence the results of treatment. *Arch Surg* 1995; **130**: 1123-1128; discussion 1129 [PMID:

7575127]

- 19 **Lillemoe KD**, Melton GB, Cameron JL, Pitt HA, Campbell KA, Talamini MA, Sauter PA, Coleman J, Yeo CJ. Postoperative bile duct strictures: management and outcome in the 1990s. *Ann Surg* 2000; **232**: 430-441 [PMID: 10973393]

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