

Editorial Board

2011-2015

The *World Journal of Pharmacology* Editorial Board consists of 476 members, representing a team of worldwide experts in pharmacology. They are from 44 countries, including Argentina (1), Australia (12), Austria (3), Belarus (1), Belgium (3), Brazil (5), Bulgaria (1), Canada (13), Chile (2), China (45), Czech Republic (2), Denmark (2), Egypt (2), Finland (3), France (13), Germany (7), Greece (17), Hungary (6), Iceland (1), India (10), Iran (4), Ireland (1), Israel (13), Italy (40), Japan (31), Malaysia (1), Mexico (1), Netherlands (11), New Zealand (2), Poland (3), Portugal (2), Russia (1), Saint Kitts and Nevis (1), Saudi Arabia (1), Serbia (1), Singapore (7), South Korea (10), Spain (22), Sweden (4), Switzerland (2), Thailand (2), Turkey (6), United Kingdom (21), and United States (140).

EDITOR-IN-CHIEF

Geoffrey Burnstock, *London*

GUEST EDITORIAL BOARD MEMBERS

Chia-Hsiang Chen, *Zhunan*
Jong-Yuh Cherng, *Chia-yi*
Jia-You Fang, *Taoyuan*
Ming-Fa Hsieh, *Chung Li*
Dong-Ming Huang, *Miaoli County*
Tsong-Long Hwang, *Taoyuan*
Jiiang-Huei Jeng, *Taipei*
Mei-Chuan Ko, *Taipei*
Po-Lin Kuo, *Kaohsiung*
Hsien-Yuan Lane, *Taichung*
Chen-Lung Steve Lin, *Kaohsiung*
Min-Hsiung Pan, *Kaohsiung*
Joen-Rong Sheu, *Taipei*
Chih-Hsin Tang, *Taichung*
Chin-Hsiao Tseng, *Taipei*
Chih-Shung Wong, *Taipei*
Sheng-Nan Wu, *Tainan*
Wen-Bin Wu, *Taipei*
Chuen-Mao Yang, *Taoyuan*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Alicia Beatriz Motta, *Buenos Aires*



Australia

Jonathon C Arnold, *Sydney*
Alexander Bobik, *Melbourne*

Stephen John Clarke, *Artarmon*
Brian Dean, *Melbourne*
Xiao-Jun Du, *Melbourne*
Cherrie A Galletly, *Adelaide*
Andrew John Lawrence, *Parkville Vic*
Johnson Mak, *Victoria*
Des Raymond Richardson, *Sydney*
Shaun L Sandow, *Sydney*
Karly Calliopi Sourris, *Victoria*
Fanfan Zhou, *Sydney*



Austria

Andreas Bernkop-Schnurch, *Innsbruck*
Martin Hohenegger, *Vienna*
Siegfried Kasper, *Vienna*



Belarus

Peter Gregor Rytik, *Minsk*



Belgium

Van Dam D Charlotte Josephine, *Wilrijk*
Mark Van de Castele, *Brussels*
Mathieu Vinken, *Brussels*



Brazil

Mohammad Abdollahi, *Minas Gerais*
Frederic Frezard, *Minas Gerais*
Maria de N Correia Soeiro, *Rio de Janeiro*
Waldiceu Aparecido Verri Jr, *Londrina*
Angelina Zanesco, *Sao Paulo*



Bulgaria

Stanislav Gueorguiev Yanev, *Sofia*



Canada

Sylvain G Bourgoin, *Quebec*
Subrata Chakrabarti, *Ontario*
Thomas K H Chang, *Vancouver*
Janos G Filep, *Montreal*
Pierre A Guertin, *Quebec*
Bernard Le Foll, *Toronto*
Suhayla Mukaddam-Daher, *Quebec*
Claude Rouillard, *Quebec*
Jean Sevigny, *Quebec*
Ashok K Srivastava, *Quebec*
Margarey Danielle Weiss, *Vancouver*
Jonathan P Wong, *Medicine Hat*
Xi Yang, *Manitoba*



Chile

Javier Palacios, *Antofagasta*
Armando Rojas, *Talca*



China

George G Chen, *Hong Kong*
Chi-Hin Cho, *Hong Kong*
Li-Wu Fu, *Guangzhou*
Qin He, *Chengdu*
Qing-Yu He, *Guangzhou*
Yu Huang, *Hong Kong*
Xi-Qun Jiang, *Nanjing*

Tai-Yi Jin, *Shanghai*
 Yiu Wa Kwan, *Hong Kong*
 Ke Lan, *Chengdu*
 Pak-Heng George Leung, *Hong Kong*
 Jian-Jun Li, *Beijing*
 Peng Liang, *Shenyang*
 Zhi-Xiu Lin, *Hong Kong*
 Xiao-Dong Liu, *Nanjing*
 Xin-Yong Liu, *Jinan*
 Yong-Yong Shi, *Shanghai*
 Jing-Fang Wang, *Shanghai*
 Yong-Qing Wang, *Nanjing*
 William Ka Kei Wu, *Hong Kong*
 Ruian (Ray) Xu, *Xiamen*
 Xiaoqiang Yao, *Hong Kong*
 Wei-Hai Ying, *Shanghai*
 Shu-Biao Zhang, *Dalian*
 Yu Zhang, *Beijing*
 Cheng-Gang Zou, *Kunming*



Czech Republic

Vladimir Krystof, *Olomouc*
 Kamil Kuca, *Hradec Kralove*



Denmark

Morten Grunnet, *Copenhagen*
 Yasser Ahmed Mahmmoud, *Aarhus*



Egypt

Nagwa M Nour El Din, *Alexandria*
 Manar Mahfouz Salem, *Tanta*



Finland

Seppo Kahkonen, *Helsinki*
 Hannu Ilmari Kankaanranta, *Seinajoki*
 Helder Almeida Santos, *Helsinki*



France

Christian Bronner, *Strasbourg*
 Rene Bruno, *Marseille*
 Marie-Chantal Canivenc-Lavier, *Dijon*
 Bertrand Cariou, *Nantes*
 Emmanuelle Corruble, *Le Kremlin Bicêtre*
 Boue-Grabot Eric, *Bordeaux*
 Siest Gerard, *Nancy*
 Laurent Karila, *Villejuif*
 Frederic Lagarce, *Angers*
 Tanguy Nicolas Maurice, *Montpellier*
 Fernando Rodrigues-Lima, *Paris*
 Jean-Marc Sabatier, *Marseille*
 Steeve Herve Thany, *Angers*



Germany

Axel Becker, *Magdeburg*
 Thomas Efferth, *Mainz*
 Walter E Haefeli, *Heidelberg*
 Florian Lang, *Tubingen*
 Huijie Li, *Mainz*

Frank Thevenod, *Witten*
 Michael Wink, *Heidelberg*



Greece

Panagiotis G Anagnostis, *Thessaloniki*
 Ekaterini Chatzaki, *Alexandroupolis*
 Vassilis J Demopoulos, *Thessaloniki*
 Moses Elisaf, *Ioannina*
 Panagiotis Ferentinos, *Athens*
 Dimitrios Galaris, *Ioannina*
 George Kolios, *Alexandroupolis*
 Tzortzis Nomikos, *Athens*
 Constantinos M Paleos, *Aghia Paraskevi*
 George Panagis, *Rethymno*
 Andreas Papapetropoulos, *Patras*
 Kosmas I Paraskevas, *Athens*
 George P Patrinos, *Patras*
 Evangelos Rizos, *Ioannina*
 Despina Sanoudou, *Athens*
 Kostas Syrigos, *Athens*
 Ioannis S Vizirianakis, *Thessaloniki*



Hungary

Albert Császár, *Budapest*
 Peter Hamar, *Budapest*
 Peter Krajcsi, *Budapest*
 Gabor Maksay, *Budapest*
 Attila Janos Miseta, *Cserkut*
 Joseph Molnar, *Szeged*



Iceland

Hekla Sigmundsdottir, *Reykjavik*



India

VN Balaji, *Bangalore*
 Chiranjib Chakraborty, *Vellore*
 Naibedya Chattopadhyay, *Lucknow*
 SJS Flora, *Gwalior*
 Srinivas Gopala, *Thiruvananthapuram*
 Seetharamappa Jaldappagari, *Dharwad*
 Basavaraj K Nanjwade, *Karnataka*
 Kishore Madhukar Paknikar, *Pune*
 Vikas Anand Saharan, *Sri Ganganagar*
 Abdus Samad, *delhi*



Iran

Mohammad Abdollahi, *Tehran*
 Ahmad Reza Dehpour, *Tehran*
 Mehrdad Hamidi, *Zanjan*
 Arash Mowla, *Bushehr*



Ireland

Marek Witold Radomski, *Dublin*



Israel

Galila Agam, *Beer-Sheva*

Robert Henry Belmaker, *Beersheva*
 Shomron Ben-Horin, *Tel-Hashomer*
 Arik Dahan, *Beer-Sheva*
 Hagit Eldar-Finkelman, *Rehovot*
 Eliezer Flescher, *Tel Aviv*
 Moshe Gavish, *Haifa*
 Jacob George, *Rehovot*
 Israel Hanukoglu, *Ariel*
 Joseph Kost, *Beer-Sheva*
 Irena Manov, *Haifa*
 Mordechai Muszkat, *Jerusalem*
 Michal Schwartz, *Rehovot*



Italy

Giuseppe Barbaro, *Rome*
 Francesca Borrelli, *Naples*
 Franco Borsini, *Pomezia*
 Silvio Caccia, *Milan*
 Giuseppe Maurizio Campo, *Messina*
 Raffaele Capasso, *Naples*
 Mauro Antonio Maria Carai, *Cagliari*
 Dario Cattaneo, *Milan*
 Davide Cervia, *Viterbo*
 Giuseppe Cirino, *Napoli*
 Emilio Clementi, *Milano*
 Massimo Collino, *Torino*
 Vincenzo Cuomo, *Rome*
 Francesca Fallarino, *Perugia*
 Tullio Florio, *Genova*
 Vittorio Gentile, *Napoli*
 Guido Grassi, *Milan*
 Mario Grassi, *Trieste*
 Annalisa Guaragna, *Napoli*
 Milena Gusella, *Trecenta*
 Francesco Impagnatiello, *Milan*
 Angelo A Izzo, *Naples*
 Luca La Colla, *Parma*
 Giovanni Landoni, *Milan*
 Aurelio Leone, *Castelnuovo Magra*
 Mauro Magnani, *Urbino*
 Mario Marchi, *Genova*
 Silvia Marinelli, *Rome*
 Robert Nistico, *Rome*
 Francesco Parmeggiani, *Ferrara*
 Sabina Passamonti, *Trieste*
 Emilio Perucca, *Pavia*
 Carlo Riccardi, *Perugia*
 Graziano Riccioni, *Manfredonia*
 Sergio Rutella, *Rome*
 Gianni Sava, *Trieste*
 Pier Andrea Serra, *Sassari*
 Luca Steardo, *Rome*
 Claudiu T Supuran, *Florence*
 Gianluca Tettamanti, *Varese*



Japan

Katsuya Dezaki, *Tochigi*
 Jun Fang, *Kumamoto*
 Takahisa Furuta, *Hamamatsu*
 Mitsuko Furuya, *Yokohama*
 Osamu Handa, *Kyoto*
 Hideaki Hara, *Gifu*
 Kenji Hashimoto, *Chiba*
 Zhi-Qing Hu, *Tokyo*
 Toru Kobayashi, *Niigata*
 Hiroshi Kunugi, *Tokyo*
 Makoto Makishima, *Tokyo*

Takayuki Masaki, *Oita*
 Shin-ichiro Miura, *Fukuoka*,
 Noboru-Motohashi, *Tokyo*
 Yuji Naito, *Kyoto*
 Toshio Nakaki, *Tokyo*
 Satomi Onoue, *Shizuoka*
 Honoo Satake, *Osaka*
 Masaharu Seno, *Okayama*
 Yasuyuki Shimada, *Yuri-Honjo*
 Mitsushige Sugimoto, *Hamamatsu*
 Masafumi Takahashi, *Tochigi*
 Shinji Takai, *Takatsuki*
 Yoh Takuwa, *Kanazawa*
 Shingo Tsuji, *Osaka*
 Hirokazu Tsukahara, *Okayama*
 Motoko Unoki, *Fukuoka*
 Shizuo Yamada, *Shizuoka*
 Norio Yasui-Furukori, *Hirosaki*
 Yukio Yoneda, *Kanazawa*
 Kiyotsugu Yoshida, *Bunkyo-ku*



Malaysia

Johnson Stanslas, *Serdang*



Mexico

Esus Adolfo Garcia-Sainz, *Col. Nápoles*



Netherlands

Arjan Blokland, *Maastricht*
 Eliyahu Dremencov, *Groningen*
 Elisa Giovannetti, *Amsterdam*
 Hidde J Haisma, *Groningen*
 Godefridus J Peters, *Amsterdam*
 Frank A Redegeld, *Utrecht*
 Harald H H W Schmidt, *Maastricht*
 Martina Schmidt, *Groningen*
 Frederik M van der Veen, *Rotterdam*
 Charles J Vecht, *The Hague*
 Joris Cornelis Verster, *Utrecht*



New Zealand

Hesham Al-Sallami, *Dunedin*
 Lin Yang, *Dunedin*



Poland

Thomas Michal Brzozowski, *Cracow*
 Wladyslawa Anna Daniel, *Krakow*
 Andrzej Pilc, *Krakow*



Portugal

Bruno Filipe C Cardoso Sarmiento, *Porto*
 Cristina Maria Sena, *Coimbra*



Russia

Roman Gerbertovich Efremov, *Moscow*



Saint Kitts and Nevis

Ignacio Lizarraga, *Baseterre*



Saudi Arabia

Mohamed Haidara, *Abha*



Serbia

Milan Jokanovic, *Belgrade*



Singapore

Jinsong Bian, *Singapore*
 Gavin S Dawe, *Singapore*
 Chang Ming Li, *Singapore*
 Haishu Lin, *Singapore*
 Rajkumar Ramamoorthy, *Singapore*
 Gautam Sethi, *Singapore*
 WS Fred Wong, *Singapore*



South Korea

Ki Churl Chang, *Jinju*
 Joohun Ha, *Seoul*
 Sang June Hahn, *Seoul*
 Byeongmoon Jeong, *Seoul*
 Myung Gull Lee, *Bucheon*
 Won Suk Lee, *Yongsan*
 Seung-Yeol Nah, *Seoul*
 Kyoungsoo Park, *Daegu*
 Young-Hyun Yoo, *Pusan*
 Soh Yunjo, *Jeonju*



Spain

José Luis Arias-Mediano, *Granada*
 Pedro Emilio Bermejo, *Madrid*
 Fermín Sánchez de Medina, *Granada*
 Guillermo Elizondo, *Mexico*
 Leandro Fernández-Pérez, *Las Palmas*
 Cristina Fillat, *Barcelona*
 J Adolfo Garcia-Sainz, *Mexico*
 Angel Luis Montejo Gonzalez, *Salamanca*
 Tomas Herraiz, *Madrid*
 Miguel JA Lainez, *Valencia*
 Jose Martinez Lanao, *Salamanca*
 Angel Lanas, *Zaragoza*
 Vicente Martinez, *Barcelona*
 Faustino Mollinedo, *Salamanca*
 Virginia Motilva, *Sevilla*
 Gorka Orive, *Vitoria-Gasteiz*
 Ricardo Enrique Perez-Tomas, *Barcelona*
 S Rodriguez-Couto, *Donostia-San Sebastian*
 Maria Eugenia Saez, *Seville*
 Juan Sastre, *Valencia*
 Juan L Tamargo, *Madrid*
 Salvador Ventura Zamora, *Barcelona*



Sweden

Aleksander A Mathe, *Stockholm*

Sharma Hari Shanker, *Uppsala*
 Marie-Louise G Wadenberg, *Kalmar*
 Cang-Bao Xu, *Lund*



Switzerland

Stefan J Borgwardt, *Basel*
 Felicien Karege, *Geneva*



Thailand

Rumi Ghosh, *Rayong*
 Kanokwan Jarukamjorn, *Khon Kaen*



Turkey

Cengiz Abdollahi Akkaya, *Bursa*
 Sule Apikoglu-Rabus, *Istanbul*
 Fatih Canan, *Bolu*
 Saygin S Eker, *Bursa*
 Nese Tuncel, *Eskisehir*
 Mehmet Yaman, *Elazig*



United Kingdom

Charalambos Antoniadis, *Oxford*
 Sabine Bahn, *Cambridge*
 Christopher John Bushe, *New Malden*
 David J Chambers, *London*
 Michael J Curtis, *London*
 Rossen M Donev, *Swansea*
 Marco Falasca, *London*
 David James Grieve, *Belfast*
 Alan Jeffrey Hargreaves, *Nottingham*
 Mahmoud M Iravani, *London*
 Nigel Irwin, *Coleraine*
 Lin-Hua Jiang, *Leeds*
 Veena Kumari, *London*
 Kim Lawson, *Sheffield*
 Debbi MacMillan, *Glasgow*
 Elek-Molnar, *Bristol*
 Stuart Anthony Rushworth, *Norwich*
 Sunita Suri, *Nottingham*
 Jinsheng Xu, *Bristol*
 Alexander Victor Zholos, *Belfast*



United States

Nihal Ahmad, *Madison*
 James David Adams Jr, *Los Angeles*
 Gustav Akk, *St. Louis*
 Karim A Alkadhi, *Houston*
 Charles Antzelevitch, *Utica*
 Hugo Ruben Arias, *Glendale*
 Dominick L Auci, *Escondido*
 Ross J Baldessarini, *Belmont*
 Oleg A Barski, *Louisville*
 Bjorn Bauer, *Duluth*
 Chengpeng Bi, *Kansas*
 Marco Bortolato, *Los Angeles*
 Josh Burk, *Williamsburg*
 William K Chan, *Stockton*
 James J Chen, *Jefferson*
 Zhe-Sheng Chen, *New York*
 Beek Yoke Chin, *Boston*
 Ting-Chao Chou, *New York*

Olivier Civelli, *Irvine*
 Brian S Cummings, *Athens*
 John A Dani, *Houston*
 Igor Elman, *Belmont*
 Keith M Erikson, *Greensboro*
 Eric R Fedyk, *Cambridge*
 Pingfu Feng, *Cleveland*
 William Douglas Figg, *Bethesda*
 Mitchell Phillip Fink, *Los Angeles*
 Masayuki Fukata, *Miami*
 Bolin Geng, *Waltham*
 Arup K Ghose, *West Chester*
 Alasdair M Gilfillan, *Bethesda*
 Neeraj Gupta, *Cambridge*
 James P Hardwick, *Rootstown*
 David W Hein, *Louisville*
 Huixiao Hong, *Jefferson*
 Andrew G Horti, *Baltimore*
 Eric Huang, *San Diego*
 Peng Huang, *Houston*
 Ying Huang, *Syracuse*
 Sally A Huston, *Athens*
 Basalingappa L Hungund, *Orangeburg*
 Kenneth A Jacobson, *Bethesda*
 Sabzali Javadov, *San Juan*
 Douglas Lee Jennings, *Detroit*
 Robert Thomas Jensen, *New York*
 Guang-Liang Jiang, *Irvine*
 Zhi-Gen Jiang, *Portland*
 Harish C Joshi, *Atlanta*
 Thomas Harold Kelly, *Lexington*
 Raouf A Khalil, *Boston*
 Arifulla Khan, *Seattle*
 Mattheos Koffas, *Buffalo*
 Zbigniew K Krowicki, *New Orleans*
 Macus Tien Kuo, *Houston*
 Young Jik Kwon, *Irvine*
 Lorenzo Leggio, *Tehran*
 Jinhe Li, *Abbott Park*
 Liwu Li, *Blacksburg*
 Ching-Shwun Lin, *San Francisco*
 Yong Lin, *Albuquerque*
 Dong min Liu, *Blacksburg*
 Jie Liu, *Kansas City*
 Ming-Cheh Liu, *Toledo*
 Xiu Liu, *Jackson*
 Edythe D London, *Los Angeles*
 Jian Lu, *Baltimore*
 Rudolf Lucas, *Augusta*
 Qing Ma, *Buffalo*
 Iddo Magen, *Los Angeles*
 Gerald A Maguire, *Orange*
 Kenneth Maiese, *Newark*
 Stuart Maudsley, *Baltimore*
 Christopher Robert McCurdy, *Mississippi*
 Michael Robert McDevitt, *New York*
 Pamela A McKinley, *Detroit*
 Beverley-G-Van Meerveld, *Oklahoma City*
 Kapil-Mehta, *Houston*
 Murielle Mimeault, *Nebraska*
 Ashim Kumar Mitra, *Kansas City*
 Agostino Molteni, *Kansas City*
 Nader H Moniri, *Atlanta*
 Valentina Echeverria Moran, *Bay Pines*
 Sandeep Mukherjee, *Omaha*
 Masanori Onda, *Bethesda*
 Murat OZ, *Baltimore*
 Pal Pacher, *Bethesda*
 Hui-Lin Pan, *Houston*
 Weihong Pan, *Baton Rouge*
 Giulio Maria Pasinetti, *New York*
 Kennerly Sexton Patrick, *Charleston*
 George Perry, *San Antonio*
 James Porter, *Grand Forks*
 Lucas Pozzo-Miller, *Birmingham*
 Mei Qiang, *San Antonio*
 Baskaran Rajasekaran, *Pittsburgh*
 Jeff Reagan, *Woodside*
 Victoria Risbrough, *San Diego*
 Michael A Rogawski, *Sacramento*
 Steven Alan Rosenzweig, *Charleston*
 Uwe Rudolph, *Belmont*
 Arnold E Ruoho, *Madison*
 Wolfgang Sadee, *Columbus*
 Ahmad R Safa, *Indianapolis*
 Stephen H Safe, *Houston*
 Shakil Ahmed Saghir, *Midland*
 Sanjeev Shangary, *Ann Arbor*
 Mahesh Chandra Sharma, *Washington*
 Anantha Shekhar, *Indianapolis*
 Riyi Shi, *West Lafayette*
 Amruthesh C Shivachar, *Houston*
 Blair Karina Simone, *Bethesda*
 Brij Bhan Singh, *Grand Forks*
 Xue-Long Sun, *Cleveland*
 Manjunath N Swamy, *El Paso*
 Yvette France Tache, *Los Angeles*
 Kevin Scott Thorneloe, *King of Prussia*
 Robin L Thurmond, *San Diego*
 Guochuan Emil Tsai, *Torrance*
 Tove Tuntland, *San Diego*
 N D Vaziri, *Orange*
 Libor Velisek, *New York*
 Christoph F Adam Vogel, *Sacramento*
 Christian Waeber, *Charlestown*
 Yu-Jui Yvonne Wan, *Kansas City*
 Qin Wang, *Birmingham*
 R Clinton Webb, *Augusta*
 Thomas Wisniewski, *New York*
 Wing Tak Jack Wong, *Stanford*
 Jie Wu, *Phoenix*
 Zheng-Xiong Xi, *Baltimore*
 Da-Liao Xiao, *Loma Linda*
 Lixia Yao, *King of Prussia*
 Hao Yin, *Cambridge*
 Xiaozhong Yu, *Seattle*
 Chang-Guo Zhan, *Lexington*
 Hanting Zhang, *Morgantown*
 Qunwei Zhang, *Louisville*
 Shuxing Zhang, *Houston*
 Bao-Ting Zhu, *Kansas City*
 Chang Zhi Zhu, *Abbott Park*

World Journal of *Pharmacology*

World J Pharmacol 2014 March 9; 3(1): 1-17





FRONTIER

- 1 Implantable (Bio)sensors as new tools for wireless monitoring of brain neurochemistry in real time

Farina D, Alvau MD, Puggioni G, Calia G, Bazzu G, Migheli R, Sechi O, Rocchitta G, Desole MS, Serra PA

Contents

World Journal of Pharmacology
Volume 3 Number 1 March 9, 2014

APPENDIX I-V Instructions to authors

ABOUT COVER Editorial Board Number of *World Journal of Pharmacology*, Pier Andrea Serra, Professor of Pharmacology, Department of Neuroscience, Medical School, University of Sassari, V.le S. Pietro 43/b, 07100 Sassari, Italy

AIM AND SCOPE *World Journal of Pharmacology* (*World J Pharmacol*, *WJP*, online ISSN 2220-3192, DOI: 10.5497) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJP covers topics concerning neuropsychiatric pharmacology, cerebrovascular pharmacology, geriatric pharmacology, anti-inflammatory and immunological pharmacology, antitumor pharmacology, anti-infective pharmacology, metabolic pharmacology, gastrointestinal and hepatic pharmacology, respiratory pharmacology, blood pharmacology, urinary and reproductive pharmacology, pharmacokinetics and pharmacodynamics, clinical pharmacology, and drug toxicology.

We encourage authors to submit their manuscripts to *WJP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING *World Journal of Pharmacology* is now indexed in Digital Object Identifier.

FLYLEAF I-IV Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Su-Qing Liu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xiu-Xia Song*

NAME OF JOURNAL
World Journal of Pharmacology

ISSN
ISSN 2220-3192 (online)

LAUNCH DATE
February 9, 2012

FREQUENCY
Quarterly

EDITOR-IN-CHIEF
Geoffrey Burnstock, PhD, DSc, FAA, FRCS (Hon), FRCP (Hon), FmedSci, FRS, Professor, Autonomic Neuroscience Centre, University College Medical School, Royal Free Campus, Rowland Hill Street, London NW3 2PF, United Kingdom

EDITORIAL OFFICE
Jin-Lei Wang, Director
Xiu-Xia Song, Vice Director

World Journal of Pharmacology
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLISHER
Baishideng Publishing Group Inc
8226 Regency Drive,
Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLICATION DATE
March 9, 2014

COPYRIGHT

© 2014 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

Full instructions are available online at http://www.wjgnet.com/2220-3192/g_info_20100722180909.htm

ONLINE SUBMISSION

<http://www.wjgnet.com/esp/>

Implantable (Bio)sensors as new tools for wireless monitoring of brain neurochemistry in real time

Donatella Farina, Maria D Alvau, Giulia Puggioni, Giammario Calia, Gianfranco Bazzu, Rossana Migheli, Ottavio Sechi, Gaia Rocchitta, Maria S Desole, Pier Andrea Serra

Donatella Farina, Maria D Alvau, Giulia Puggioni, Giammario Calia, Gianfranco Bazzu, Rossana Migheli, Ottavio Sechi, Gaia Rocchitta, Maria S Desole, Pier Andrea Serra, Department of Clinical and Experimental Medicine, Medical School, University of Sassari, 07100 Sassari, Italy

Author contributions: Farina D and Calia G contributed to acetylcholine biosensor and references reorganization; Alvau MD and Puggioni G contributed to glucose and lactate biosensors; Bazzu G and Migheli R contributed to dopamine, norepinephrine, and serotonin microsensors; Rocchitta G contributed to glutamate biosensor, ascorbic acid microsensor, ethanol biosensor, and biotelemetry; Sechi O contributed to oxygen and nitric oxide microsensors and ethanol biosensor; Desole MS and Serra PA contributed to title, abstract, introduction, conclusion, references reorganization, and manuscript overview; Farina D and Alvau MD equally contributed to this study.

Supported by The Regione autonoma della Sardegna (fund P. O. R. SARDEGNA F. S. E. 2007-2013-Obiettivo competitività regionale e occupazione, Asse IV Capitale umano, Linea di Attività I. 3. 1)

Correspondence to: Pier Andrea Serra, MD, PhD, Department of Clinical and Experimental Medicine, Medical School, University of Sassari, V.le S. Pietro 43/b, 07100 Sassari, Italy. paserra@uniss.it

Telephone: +39-079-228558 Fax: +39-079-228525

Received: January 15, 2014 Revised: March 3, 2014

Accepted: March 6, 2014

Published online: March 9, 2014

Abstract

Implantable electrochemical microsensors are characterized by high sensitivity, while amperometric biosensors are very selective in virtue of the biological detecting element. Each sensor, specific for every neurochemical species, is a miniaturized high-technology device resulting from the combination of several factors: electrode material, shielding polymers, applied electrochemical technique, and in the case of biosensors, biological sensing material, stabilizers, and entrapping chemical nets. In this paper, we summarize

the available technology for the *in vivo* electrochemical monitoring of neurotransmitters (dopamine, norepinephrine, serotonin, acetylcholine, and glutamate), bioenergetic substrates (glucose, lactate, and oxygen), neuromodulators (ascorbic acid and nitric oxide), and exogenous molecules such as ethanol. We also describe the most represented biotelemetric technologies in order to wirelessly transmit the signals of the above-listed neurochemicals. Implantable (Bio)sensors, integrated into miniaturized telemetry systems, represent a new generation of analytical tools that could be used for studying the brain's physiology and pathophysiology and the effects of different drugs (or toxic chemicals such as ethanol) on neurochemical systems.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Electrochemical microsensors; Amperometric biosensors; Neurotransmitters; Bioenergetic substrates; Wireless biotelemetric technologies

Core tip: Electrochemical microsensors and amperometric biosensors arouse enormous scientific interest because of their low-cost technology and because they guarantee real-time monitoring of changes of the most important brain compounds. In conjunction with miniaturized telemetric devices, the electrochemical sensors, allow the neurochemical monitoring of extracellular space of discrete brain regions in awake, untethered animals for days or weeks. This new scientific approach opens new frontiers for studying the physiological and physiopathological pathways in wild-type animals and in genetic models of the most widespread neurodegenerative diseases.

Farina D, Alvau MD, Puggioni G, Calia G, Bazzu G, Migheli R, Sechi O, Rocchitta G, Desole MS, Serra PA. Implantable (Bio)sensors as new tools for wireless monitoring of brain neurochemistry in real time. *World J Pharmacol* 2014; 3(1): 1-17

INTRODUCTION

The identification, observation, and quantification of extracellular biomolecules in the central nervous system (CNS) is a field of growing interest for studying the brain in physiological conditions and for identifying neurochemical changes during neurological diseases. The study of neurochemistry in real time is very important in preclinical (and recently also in clinical) research and for developing new therapeutic strategies for many neuropsychiatric diseases, such as schizophrenia, depression, epilepsy, multiple sclerosis, and neurodegenerative diseases (*i.e.*, Parkinson's and Alzheimer's diseases), and also for neural conditions that deeply influence individual and social behavior such as addiction.

For decades, the extracellular neurochemistry of the CNS has been studied using *in vivo* microdialysis. Microdialysis is a minimally invasive technique suitable for measuring low-molecular-weight compounds in the extracellular compartment of several organs, tissues, or specific brain regions^[1]. The microdialysis idea originated in the 1970s with the aim of implanting a hollow dialysis fiber (microdialysis probe) into a tissue for simulating the role of a blood capillary and recovering molecules from the extracellular compartment to highlight their regional changes in concentration^[2,3]. When implanted in the brain, the microdialysis probe is perfused with an appropriate Ringer solution (that mimics the composition of the extracellular space fluid) so that neurochemicals are able to diffuse down their concentration gradients out of the probe. The recovered microdialysis samples are analyzed using different analytical methods. The poor temporal resolution and the need to have an available expensive analytical laboratory (for analyzing microdialysis samples) represent the major limitations of this technique.

In recent decades, implantable electrochemical sensors and biosensors have been emerging because of their versatility, their multiple applications, and most of all, their high spatial and temporal resolution^[4-6]. In particular, implantable amperometric sensors have been proven to be very sensitive so as to allow the detection of very low concentrations of the studied analytes^[5]. The basic idea of implantable electrochemical sensors is to "concentrate" an entire analytical laboratory "on the tip of a pin" without the need of an expensive analytical apparatus or of a dedicated laboratory.

In the past years, despite their high sensitivity, the main limitation for the use of electrochemical sensors was related to their poor selectivity. Recently, the development of new sensing materials and new shielding polymers and, mainly, the introduction of biological elements such as molecular recognition sites have allowed

overcoming this limitation in a large part.

Today, each sensor, specific for every neurochemical species, is a miniaturized high-technology device resulting from the combination of several factors: electrode material, shielding polymers, applied electrochemical technique, and in the case of biosensors, biological sensing material, stabilizers, and entrapping chemical nets.

The dimensions of implantable electrochemical sensors vary from a few micrometers (5-10) up to 125 μm (always lower than those of a microdialysis probe, around 220 μm), and their sensing surface can be increased without increasing their invasiveness using new nanomaterials (*i.e.*, carbon nanotubes); this process is often indicated as "nanostructuration" or simply "nano-on-micro". But one of the most exciting perspectives, for future development and applications, is to combine implantable sensors with miniaturized electronic devices in order to transmit neurochemical signals at a distance so that awake animals are allowed to be totally free to move^[4-6].

In this study, we highlight the state-of-art of electrochemical microsensors and biosensors, already used in preclinical research for recording neurochemical changes, suitable to be integrated in biotelemetry systems for the wireless monitoring of brain neurochemistry.

IMPLANTABLE (BIO)SENSORS

We have chosen to describe the available technology for the *in vivo* electrochemical monitoring of neurotransmitters (dopamine, norepinephrine, serotonin, acetylcholine, and glutamate), bioenergetic substrates (glucose, lactate, and oxygen), neuromodulators (ascorbic acid and nitric oxide), and exogenous molecules such as ethanol. In the next section, we also describe the most represented biotelemetric technologies to combine with the sensors in order to wirelessly transmit the signals of the above-listed neurochemicals.

Dopamine, Norepinephrine, and Serotonin

Brain neurotransmitters such as the tyrosine derivatives dopamine, norepinephrine and the neuroactive tryptophan derivative serotonin have been implicated in the neurochemistry and physiology of mental diseases and neurological disorders.

Catecholamine biosynthesis is a common pathway from tyrosine^[7], where the hydroxylation of tyrosine to L-3,4-dihydroxyphenylalanine by tyrosine hydroxylase is the rate-limiting step. Dopamine, a catechol-like neurotransmitter derived by L-3,4-dihydroxyphenylalanine decarboxylation, is actively involved in reward pathways^[8,9] and in cognitive functions^[10]. Its metabolism mainly occurs by reaction with monoamine oxidase and catechol-O-methyltransferase with the formation of dihydroxyphenylacetic acid, homovanillic acid, and 3-methoxytyramine. Neuronal death of catecholaminergic cells in the substantia nigra, with a consequent significant reduction of dopamine levels^[11] as well as dihydroxyphenylacetic acid, homovanillic acid^[12] and

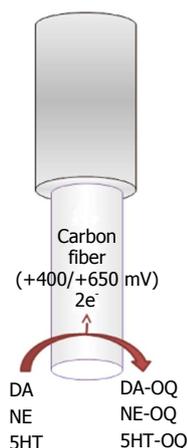
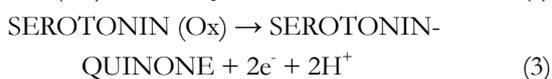


Figure 1 Schematic representation of the carbon-based microsensor used for detecting dopamine, norepinephrine, and 5-hydroxytryptamine in the central nervous system of awake, freely moving animals. DA: Dopamine; NE: Norepinephrine; 5HT: 5-Hydroxytryptamine, serotonin; DA-OQ: DA-derived orthoquinones; NE-OQ: NE-derived orthoquinones; 5HT-OQ: 5HT-derived orthoquinones.

3-methoxytyramine^[13] in the striatum is a hallmark in Parkinson's disease^[1]. On the other hand, an increase in dopaminergic levels is involved in the etiopathogenesis of schizophrenia^[14,15].

Formed by β -hydroxylation of dopamine, norepinephrine plays multiple roles as a hormone and a neurotransmitter. Norepinephrine is involved in directly increasing heart rate, suppressing neuroinflammation^[16], and triggering the glycogenolysis and the release of glucose from energy stores^[17], and along with serotonin, it is implicated in depression and anxiety disorders^[18]. Moreover, the serotonergic system is also implicated in several neuroregulatory processes such as stress, aggression, pain, sleep, appetite, reproduction, circadian rhythm, and cardiovascular and respiratory functions^[19].

All of these compounds are electrochemically active, show a similar 2-electron oxidation reaction with similar peak potentials at physiological pH, and can be directly detected by electrochemical oxidation of the molecule^[20].



The electroactive neurotransmitters can be directly detected *in vitro* and *in vivo* using different electrochemical techniques (Figure 1) such as constant potential amperometry (CPA)^[21], chronoamperometry^[22,23], differential pulse voltammetry (DPV)^[24], and fast-scan cyclic voltammetry (FSCV)^[8,25-27]. Different microelectrodes for voltammetric recordings in the CNS are available, such as carbon paste microelectrodes, where carbon powder is mixed with silicon oil^[10]; epoxy carbon microelectrodes, where epoxy resin is mixed with carbon paste; and carbon fiber, gold, and platinum (Pt) microelectrodes^[20].

Along carbon-fiber microelectrodes, FSCV is the

most common technique used for dopamine, norepinephrine and serotonin *in vivo* monitoring.

Carbon-fiber microelectrodes (Figure 1) are made by inserting a carbon fiber (outer diameter ranging between 5 and 30 μm , most commonly about 7 μm) into a glass capillary, which is pulled with a pipette puller and sealed by epoxy resin with 25 to 100 μm of the fiber protruding from the glass. The final geometry of the electrode, cylindrical^[28] or disk shaped^[29], is obtained by cutting or polishing the protruding carbon fiber^[30]. Because of their dimension, carbon-fiber microelectrodes minimize distortion caused by ohmic drop, and then, coupled with a minimal tissue damages when implanted into the brain, they are suitable for high-temporal-resolution measurements^[28]. In addition, a 7 μm carbon fiber does not stimulate glial reaction^[25], in agreement with the evidence that probes that are less than 12 μm in diameter are not encapsulated as demonstrated by previous studies^[31]. FSCV is a technique with high resolution and selectivity, where the potential applied to the microsensor is cycled between the reduction and the oxidation peaks of the analyte of interest^[20]. For dopamine and norepinephrine recordings, a scan rate in a triangle fashion at 400 V/s is applied. The potential of the carbon-fiber microelectrode is ramped linearly from -400 mV *vs* Ag/AgCl to +1.3 V and back and held at -400 mV between scans^[32]. To obtain the 5HT recording, an N-waveform scan rate is used, in which the applied potential is scanned first from 0 mV to +1200 mV then to -600 mV and back to 0 *vs* Ag/AgCl^[27]. Typically, the waveform is applied for 10 ms, and voltammetric scans are repeated at 100 ms intervals. During the anodic sweep, the catecholamine (dopamine and/or norepinephrine) and serotonin present at the electrode surface are oxidized into corresponding orthoquinone and then reduced back at the original form during the cathodic sweep. The number of molecules that undergo electrolysis is directly proportional to the measured current^[21]. The peak positions during oxidation and the reduction sweep as well as the peak shape can be used to distinguish different analytes^[33].

Using fast-scan cyclic voltammetry, dopamine, norepinephrine, and serotonin have been shown a similar oxidation peak at approximately +650 mV *vs* Ag/AgCl^[33-35] and a single reduction peak around -200 mV for dopamine and norepinephrine or Wdouble reduction peaks around 0 and -500 mV *vs* Ag/AgCl for serotonin^[27].

Because they are virtually identical, voltammograms alone cannot be used to distinguish dopamine and norepinephrine^[36], but histology and pharmacology, such as the use of dopamine drugs (raclopride, GBR 12909), can aid in this distinction even in simultaneous measurements with FSCV^[37]. Ascorbic acid is the main electroactive interference molecule in the extracellular fluid (ECF) of the brain for electrochemical measurements. Ascorbic acid is 10^4 - 10^6 times higher than the concentrations of catecholamines in the ECF of the brain, and its concentration is approximately 0.5 mmol/L^[37,38]. The carbon-fiber microsensor selectivity for catecholamines can be enhanced

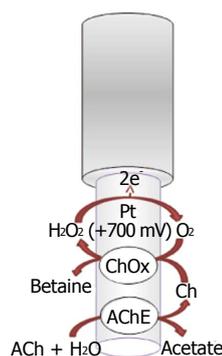


Figure 2 Schematic representation of the platinum-based biosensor used for detecting acetylcholine in the brain of awake, freely moving animals. ACh: Acetylcholine; Ch: Choline; ChOx: Choline oxidase; AChE: Acetylcholinesterase.

by applying on fibers a negatively charged resin (Nafion) able to concentrate cations such as dopamine on the active surface of the sensor and, at the same time, to repel anions such as ascorbic acid and dihydroxyphenylacetic acid^[22,39].

Although carbon-fiber microelectrodes are the most used sensors for dopamine and norepinephrine for *in vivo* recording, new strategies are developed to monitor catecholamines real time in the brain.

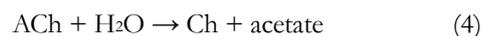
As recently suggested by Njagi *et al.*^[40], an amperometric biosensor can be fabricated depositing an enzyme, such as tyrosinase, onto the surface of a carbon-fiber electrode. The enzyme immobilized in a biocompatible matrix and with a final diameter of about 100 μm provides an alternative to FSCV for *in vivo* monitoring of dopamine^[40].

Acetylcholine

The neurotransmitter acetylcholine and its metabolite choline play a critical role in various functions of the CNS^[41]. The concentration of acetylcholine in the ECF of the brain is 0.1-6 nmol/L^[42]; the abnormalities in their concentrations are related to several neural diseases^[43]. In particular, it is involved in learning and memory formation^[44], in the development and maintenance of addiction^[45], and in neurodegenerative disorders such as Alzheimer's disease^[46] and Parkinson's disease^[47,48]; dysregulation of cholinergic transmission is correlated to cognitive alterations such as those manifested in Alzheimer's disease^[49]. Furthermore, organophosphorus (OP) and carbamate pesticides and neurotoxic compounds are capable to inhibit the acetylcholinesterase enzyme (AChE), which is responsible of the hydrolysis of acetylcholine^[50].

Therefore, the *in vivo* determination of acetylcholine and choline is important because a rapid and an effective method for simultaneous determination of levels of acetylcholine and choline is needed for the characterization of cholinergic transmission in normal and pathological physiology^[51,52]. The most common methods developed for the simultaneous determination of acetylcholine and choline require a conversion into more easily detectable compounds^[52].

A lot of strategies have been used to obtain selective detection for acetylcholine and choline with biosensors. Among all acetylcholinesterase-based biosensors, amperometric acetylcholinesterase/choline oxidase (ChOx) biosensor is especially performing because of its potential high sensitivity, reproducibility, and excellent selectivity for *in vivo* simultaneous determination of neurotransmitters; these devices are usable for *in situ* determination of choline and acetylcholine and have been implanted in rat brain^[51]. The working mechanism of acetylcholinesterase (Figure 2) is based on the following biochemical reaction^[53]:



While the choline, in the presence of oxygen, is oxidized by choline oxidase, forming hydrogen peroxide (H_2O_2), which can be easily oxidized onto electrode surface:



The oxidation current of hydrogen peroxide can be used for the evaluation of acetylcholine, choline, and acetylcholinesterase activity. Acetylcholine signal is attenuated by acetylcholinesterase inhibitors such as neostigmine or physostigmine^[54,55]. The enzymes acetylcholinesterase and choline oxidase are immobilized on the solid electrode surface such as platinum-iridium (Pt/Ir)^[51,56] (Figure 2) or carbon fibers^[57]. In order to prevent signal of interferents, different shielding strategies are currently used different. For example, ascorbate oxidase (AAO) is used to minimize interference from ascorbic acid, which is present in relatively high concentrations in the brain ECF^[58]; polymeric films are also used onto the sensor surface that limit the access of potential interferences due to electrostatic repulsion (*e.g.*, Nafion) and nonconducting polymers [*e.g.*, poly-(phenylenediamines) (PPD)] that restrict the permeability of small organic molecules (*e.g.*, major interferences ascorbate and urate) while retaining a high permeability to small species such as hydrogen peroxide^[59]. The acetylcholinesterase/choline oxidase layer is trapped onto the surface electrode by the cross-linking of amino groups of the enzymes with glutaraldehyde^[51]. Moreover, the enzyme layer also includes bovine serum albumin (BSA) that provides stabilization of the enzyme activity in the immobilized state^[51].

Hence, the amperometric sensors for acetylcholine and choline are successfully applied and provide a useful tool to analyze basic mechanisms of cholinergic physiology in normal and pathological conditions and those involved in the activity of pharmacological cholinergic drugs.

Glutamate and ascorbic acid

Even if glutamate is a nonessential amino acid, it has been shown to be the most abundant in the brain. As fully described, glutamate represents the most important excitatory neurotransmitter. In plasma, glutamate concentrations reach 50-100 $\mu\text{mol/L}$ while in the whole brain, they are 10-12 mmol/L, but we must take into account that glutamate reaches only 0.5-2.0 $\mu\text{mol/L}$ in ECFs^[60].

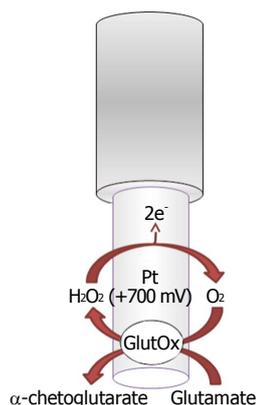


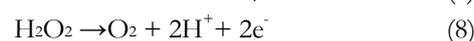
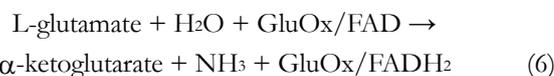
Figure 3 Scheme of glutamate biosensor. The transducer is made of a platinum (Pt) wire that immobilizes the glutamate oxidase (GluOx) enzyme that selectively transforms glutamate in alpha-ketoglutarate, producing H_2O_2 that is then oxidized on the Pt surface.

Glutamate is well known to be involved in most phases of normal brain functions such as memory and learning, cognition, cell migration, differentiation, and death; but at the same time, it is known to play important roles as a highly toxic endogenous excitotoxin^[61]. Recently, some authors have highlighted its involvement not only in the development of the CNS, particularly related to neuronal survival, growth, and differentiation, but also in the development of several circuits^[62]. In this regard, for example, it has been widely shown that low glutamate levels during neurogenesis may have a key role in the development of schizophrenia^[63], and high glutamate levels can also interfere with astroglial proliferation and neuronal differentiation^[61]. Glutamate has been of particular importance because of its possible involvement in neurodegenerative diseases such as amyotrophic lateral sclerosis, multiple sclerosis, Parkinson's disease, and others. In fact, the chronic overexcitation of neurons, stimulated by glutamate, is a newer concept that has linked glutamate excitotoxicity to neurodegeneration in amyotrophic lateral sclerosis, Huntington's disease, Parkinson's disease, and Alzheimer's dementia^[64].

The importance of glutamate has generated a strong interest in the development of several tools for the detection of this amino acid. Different methods have been developed to determine glutamate, including optical methods, patch clamp, and microdialysis^[65], but also including fluorometric, chromatographic, or spectrophotometric techniques, which, however, have some intrinsic limitations, such as being time-consuming, requiring pretreatment of the sample, being labor intensive, and requiring skilled handling. Nowadays, electrochemical methods are considered as one of the most promising approaches because of easiness, high spatial resolution, high sensitivity, and specificity^[66]. From the neurochemical point of view, a wide range of amperometric biosensor designs, based mainly on glutamate oxidase enzyme loading [GluOx; molecular weight, 140 kDa; solution Michaelis constant (KM), 0.21 mmol/L in neutral buffer; pI, 6.2], have been developed^[67-75].

The aim of monitoring brain glutamate using amperometric biosensors, however, is very challenging, mainly because the baseline ECF concentration of glutamate is estimated to be $\leq 5 \mu\text{mol/L}$ ^[76-103].

Glutamate oxidase-based biosensors (Figure 3) exploit the capability of the oxidase to selectively convert L-glutamate as follows:



The byproduct hydrogen peroxide is then oxidized, on the transducer surface, by applying a positive potential generating a current flow directly proportional to the glutamate concentrations.

Pt generally is the electrode material of choice for electrooxidation of hydrogen peroxide^[77,78]. Various strategies are as well realized in order to shield the biosensor from electroactive interfering substances that usually occur in ECF: first of all, ascorbic acid, through the electrochemical deposition of polymers^[68-74]; the use of anionic substances such as Nafion^[68,70,79], or the coimmobilization of the ascorbate oxidase enzyme^[75].

The amperometric biosensors have been proven to be interesting devices for *in vivo* measurement of glutamate concentrations and also for their response time, which has been estimated to be about a few seconds^[73,74], making these biosensors suitable for the study of the rapid changes in the concentrations of glutamate both in physiological conditions or during pharmacological treatments.

Ascorbic acid is a water-soluble vitamin. It is widely known for its role as an antioxidant, but it is as much recognized as a cofactor in several enzymatic reactions, including those concerning the synthesis of catecholamines, carnitine, or cholesterol^[80].

Because humans are lacking the enzyme L-gulonolactone oxidase, they cannot synthesize ascorbate, so they, therefore, have efficient machineries for both absorption and recycling of this vitamin^[81]. Among them is the transporter sodium-dependent vitamin C transporter-1 (SVCT1) involved in the body homeostasis of ascorbic acid, and the transporter SVCT2 that is necessary for the defense of active cells against oxidative stress^[82]. Even the ubiquitous GLUT-type glutamate transporters play a key role in the homeostasis of this vitamin inasmuch as they are involved in the uptake of dehydroascorbate, the oxidized form of ascorbate, in order to be recycled to ascorbate^[83].

In the CNS, ascorbic acid is an essential micronutrient, and although the entire brain concentrations are between 1 and 2 mmol/L, the neuronal concentrations have been evaluated to be as high as 10 mmol/L, whereas concentrations in glial cells are about 1 mmol/L^[84,85]. At the same time, the ascorbate concentrations present in brain ECF have been estimated to comprise between 200 and 400 $\mu\text{mol/L}$ ^[81].

Those findings suggest not only that ascorbate has a

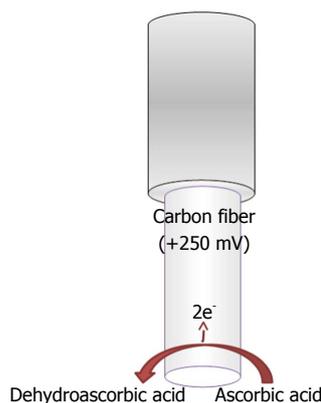
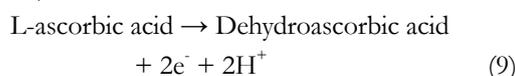


Figure 4 Scheme of AA sensor used in constant potential amperometry. In this representation, the transducer is made of a carbon fiber. The AA is oxidized by applying mild potentials (+250 mV or less) needed for oxidizing the AA to dehydroascorbic acid.

significant role in normal neuronal physiology but also that, given the structural characteristics as an electron donor and free-radical scavenger, it has assumed its role as a neuroprotective molecule and as an important component of the neuronal antioxidant pool^[81].

Neurons and glia are able to interact with each other in order to conserve CNS ascorbate, using the mechanism of heteroexchange in which ascorbate release is related principally to glutamate uptake^[86,87].

Ascorbic acid is easily oxidized in the following manner (Figure 4)



by applying a mild anodic potential^[4] at the transducer surface (Figure 4), when a constant potential is applied, and generating a current flow directly proportional to the ascorbate concentrations.

For ascorbate *in vivo* monitoring, the transducer is typically made of composite materials of carbon such as carbon paste^[87,88] or fibers^[89] and multiwalled carbon nanotube (MWNT)-modified carbon fibers^[90].

The transducer surface is sometimes modified for excluding electroactive interfering species such as positive catecholamines, so the electrode modification is carried out by the deposition of overoxidized poly (1,2-phenylenediamine)^[89].

Cyclic voltammetry (CV)^[89,90], square-wave voltammetry^[89], and differential pulse voltammetry^[91] have been used for *in vivo* measurements of ascorbic acid in the brain of animal models. The latter methods have been proven to be the most sensitive for sensing and biosensing because they change the potential pulsing from one potential to another in a relatively short range of time, different to what happens for the CV where the potential is constantly modified in a linear way^[92].

Constant potential voltammetric techniques have also been used for *in vivo* monitoring of ascorbic acid in the brain by applying mild positive potentials such as +120 mV *vs* Ag/AgCl, when this is the implanted reference

electrode (RE)^[4], or +250 mV when the implanted RE is Ag⁺^[93].

All the applied techniques have confirmed what was found with other methods that the ascorbate concentrations present in neuronal extracellular spaces are close to 500 μmol/L, emphasizing the reliability and specificity of the reading of the ascorbic acid sensors.

Glucose and lactate

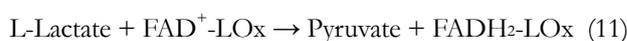
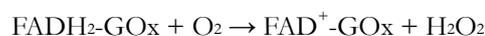
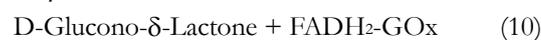
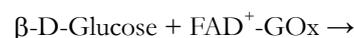
Glucose, a main nutrient in the brain^[94], is the most important factor for its energetic metabolism^[95-98] and is actively involved in ATP synthesis; it is an important modulator of memory in multiple tasks and improves memory in patients with Alzheimer's disease and Down's syndrome^[99,100].

Lactate is another important molecule involved in brain energetic metabolism as energetic substrate for neurons^[96] or product of glycolysis under anaerobic condition^[94,97].

For a long time, lactate production in the brain was viewed as a lack of oxygen, as the lack of an aerobic oxidation process, or as a mismatch between glycolytic and oxidative rates, but it has recently been identified as an alternative food to glucose^[97,100,101].

Contemporary studies in the amount of glucose and lactate in the brain are significant both in physiological conditions and in the presence of disease^[102-104].

The recognition and quantification levels of glucose and lactate are possible by using innovative devices such as biosensors constituted by an electric transducer and a biological component such as enzymes; for example, glucose oxidase (GOx), L-lactate oxidase (LOx), or L-lactate dehydrogenase (LDH) is commonly used in the design, respectively, of glucose and lactate amperometric biosensors and their exploiting simple enzymatic reactions and relatively easy sensor design configuration^[105]. In particular, amperometric methods have been widely used in glucose and lactate sensing. The biochemical reactions, in presence of oxygen, occurring at glucose and lactate biosensors are as follows^[5,106,107]:



In the electrochemical biosensor (Figures 5 and 6), the hydrogen peroxide byproduct from oxidase enzymes is directly proportional to the quantity of substrate glucose or lactate transformed by the enzymes as shown below in equation (8)^[4].

Many studies of neuronal applying biosensors in experimental models *in vivo* are present in literature^[108]. These studies show different types of biosensor designs, made with several transducer materials. Biosensors are mainly composed of noble metals, such as gold and/or

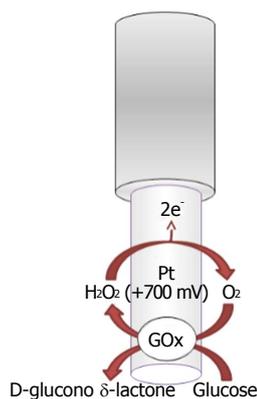


Figure 5 Schematic representation of the platinum-based biosensor used for detecting extracellular glucose in the central nervous system of freely moving animals. The immobilized glucose oxidase (GOx) selectively transforms glucose in D-gluconolactone in the presence of molecular O₂ and generates H₂O₂ that is promptly oxidized on platinum surface.

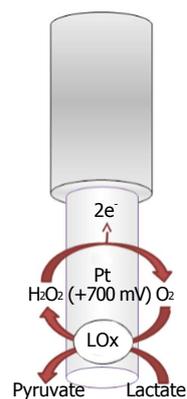


Figure 6 Schematic representation of the platinum-based biosensors for detecting extracellular lactate in the central nervous system of awake animals. In the presence of O₂, the immobilized enzyme [lactate oxidase (LOx)] selectively converts the substrate (lactate) in the corresponding product (pyruvate) and generates H₂O₂ that is oxidized on the Pt surface.

Pt, although recently, other systems use conductive carbon based materials.

A new approach for the simultaneous detection of brain glucose and lactate in real time is reached by the use of a biometric device fixed on the head of the animal^[109-111].

In a previous study^[6], O-phenylenediamine (OPD) monomers were electrodeposited onto a Pt/Ir cylinder electrode (diameter, 125 μm) surface. The next step was to immobilize GOx, stabilized with polyethylenimine (PEI), by immersing the transducer in the BSA solution and after in the glutaraldehyde solution (GTA). The lactate biosensor was initially made in the same way by changing the oxidase enzyme, but substituting the BSA/GTA with a final layer of polyurethane (PU)^[6] for increasing the linear region. CPA was used, fixing the applied potential for hydrogen peroxide oxidation at +700 mV *vs* Ag/AgCl RE.

There are numerous problems with this approach because it is necessary to apply a high potential to detect hydrogen peroxide (+700 mV)^[112,113] and the concentration of oxygen can change in the region in which the biosensor is implanted and the resulting current is not directly correlated with the extracellular concentrations of lactate^[113-115].

Furthermore, the presence of interfering electroactive species in the tissues and the reactions of biopolymerization are needed to be considered^[116,117]. In the nineties, to solve these problems, Karyakin proposed to modify the transduction element using carbon compounds coated with a thin film of Prussian blue (PB), Fe₄ [Fe(CN)₆]₃^[113,114,118-121].

After the introduction of PB in the field of biosensors were formulated different materials as supports and methodologies of deposition to improve its electrocatalytic properties and stability^[122]. In recent years, some research groups have worked on glucose and lactate microbiosensors based on PB electrodes made of carbon fiber (CFE) modified to detect enzyme-generated hydrogen

peroxide low applied potential (0 mV).

Afterward, the enzyme stabilizer PEI was added to improve the performance of the enzyme^[122], and GOx and LOx were subsequently immobilized. In order to avoid signal of interferents, OPD was electrodeposited^[122]. For the first time, a glucose and lactate microbiosensor, based on PB-modified CFE, is able to detect physiological changes in molecular levels at a low applied potential in the CNS^[123].

Moreover, the ultrasmall biosensor size is apposite for *in vivo* neuroscience studies. In contrast, the first generation of microbiosensor transducers based on noble metals have high dimensions (diameter, approximately 100 μm) even if they have been used successfully over the last few decades for the monitoring of neurochemical species^[116]. Consequently, the use of carbon-fiber microbiosensors (diameter, approximately 10 μm), modified with PB, seems to be more suitable for use in these studies because it reduces brain damage during insertion^[124] and provides an even higher temporal resolution, allowing the real-time correlation with animal behavior^[125].

Oxygen and nitric oxide

Oxygen and endogenous nitric oxide are gaseous molecules playing a pivotal role in mediating important biological processes yet are involved in very distinct aspects of organism physiology. Oxygen is indispensable for animal life; an adequate tissue oxygen content, delivered by hemoglobin through the bloodstream, is fundamental to supply cellular metabolic demands, as oxygen is involved in energy production as well as in aerobic cellular metabolism^[126].

In contrast, an insufficient oxygen concentration in tissues leads to hypoxia, a severe altered condition in which low oxygen availability prevents aerobic metabolism and oxidative phosphorylation in the cell, yielding to impoverishment of high-energy compounds such as ATP and, lastly, inducing cellular dysfunction and death^[127,128].

Though oxygen is a crucial substrate for cellular

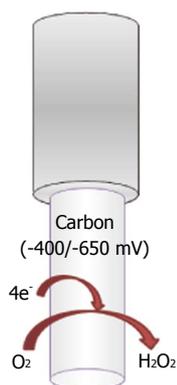


Figure 7 Schematic representation of the carbon-based sensor used for detecting the molecular O₂ dissolved in the extracellular space of the brain of freely moving animals. The O₂ is reduced on the carbon surface at low potentials and converted to water in a one- or two-step reaction (see text).

functions, it also provokes damage because of the toxicity of oxygen-derived reactive species (ROS), such as hydrogen peroxide, singlet oxygen, hydroxyl radicals, and superoxide anion^[129]. ROS free radicals attack lipids, proteins, DNA, and RNA and expose cells to oxidative stress, which has been demonstrated to be involved in the pathogenesis of several neurodegenerative diseases^[129,130].

Endogenous nitric oxide is a gaseous signaling molecule released in low concentration (tens of nanomoles to low micromoles), characterized by possessing a lifetime of a few seconds^[131], as nitric oxide is a highly reactive free-radical species. Nitric oxide production mainly involves the enzymes NO-synthases, which catalyze nitric oxide formation as a byproduct of the reduction of the amino acid L-arginine into L-citrulline^[132,133]. Nitric oxide acts as a transitory paracrine and autocrine signaling molecule, by activating the soluble guanylyl cyclase, increasing cellular cyclic guanosine monophosphate (c)^[134].

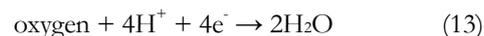
Since its discovery in 1987^[135-137], when first nitric oxide was recognized as being involved in the physiological actions of endothelium-derived relaxing factor, mediating vasodilatation, the knowledge of the important role that nitric oxide plays in physiopathology and pharmacology exponentially increased. In fact, further studies revealed how nitric oxide actions are implicated in the cardiovascular system, in the immune response^[138], as well as in the nervous systems, mediating neurotransmission^[131,139]. Furthermore, nitric oxide is a mediator of both antitumor and antimicrobial activities^[140].

Otherwise, the disruption of nitric oxide production seems to be involved in diseases such as atherosclerosis^[141], hypertension, cerebral and coronary vasospasm, and ischemia-reperfusion injury. In fact, nitric oxide is attacked by ROS, specifically by superoxide anion, forming peroxynitrite, which generates further reactive nitrogen species (RNS) such as nitrogen dioxide and dinitrogen trioxide. Like ROS, RNS damage lipids, proteins, and other macromolecules, thus also contributing to the onset of diabetes and neurodegenerative diseases^[141-143].

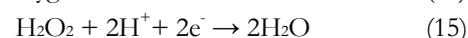
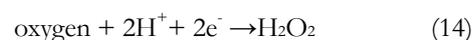
The detection of oxygen and nitric oxide tension in the brain has been studied *in vivo*, providing critical infor-

mation about the physiopathology and pharmacological implications of these molecules.

A wide variety of O₂-sensitive microsensors have been developed. Electrochemical devices exploiting amperometric techniques of detection, such as CPA, differential-pulse amperometry (DPA), CV, and fast-scan voltammetry (FCV), allow the reliable direct reduction of oxygen. Carbon paste and noble metal transducers are the most commonly diffused. Reactions involved in the electrochemical reduction of oxygen at the electrode's surface can occur *via* two mechanisms: a single-step reaction yields to detectable intermediates (Figure 7):



In the second mechanism, two-step O₂ reduction forms H₂O₂ as measurable intermediate:



Changes after physiological stimulations or pharmacological treatments were recorded in the extracellular space of the striatum, by using optic microfibers, assessing that oxygen concentration is about 50 μmol/L^[143].

Electrochemical oxygen microelectrodes using CPA at a noble metal transducer bare, such as gold or Pt, allowed the long-term monitoring of oxygen subcutaneous and venous dynamics^[144,145].

Nevertheless, several groups preferred to use carbon-paste electrodes (CPEs) because of their longer *in vivo* stability, less surface fouling^[146], and quite easy manufacture^[147] (Figure 7). Venton *et al.*^[148] used the FCV technique in a study in which dissolved oxygen was measured in the rat caudate-putamen, by using 5 μm Nafion-coated carbon fibers with a subsecond time resolution. FCV was used also in a study that targeted oxygen levels in the striatum of primates during reward delivery. In this case, the diameter of the carbon fibers ranged from 12 to 33 μm^[149].

Lowry *et al.*^[101,150,151] largely used carbon paste-based miniaturized electrodes in an experimental session in which the effects of anesthesia were studied *in vivo*, as well as the effects of hypoxia and hyperoxia on brain energy metabolism in the striatum^[147-149]. Changes in oxygen at CPEs were usually monitored by using the DPA technique^[151,152]. Two equally sized cathodic pulses were applied: the first from a resting potential at -150 to -350 mV, corresponding to the foot of the reduction wave for oxygen, and the second, which corresponds to the peak of the reduction wave, from -350 to -550 mV.

In addition, oxygen microsensors were used by Finnerty *et al.*^[153] in real-time monitoring of oxygen levels in an animal model of schizophrenia, coupled with the use of a glucose biosensor and an nitric oxide microsensor. Oxygen reduction at CPEs has been widely detected also *via* CPA^[152]. For example, by applying a constant cathodic potential of -650 mV *vs* a saturated calomel RE, oxygen reduction was recorded in real time in the hippocampus of freely moving rats^[115].

Furthermore, CPEs of 200 μm in diameter were implanted in the dorsal and the ventral hippocampus of rats

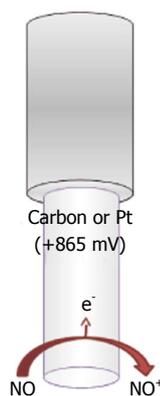


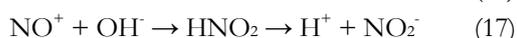
Figure 8 Schematic representation of the more widely used sensor for detecting NO in the brain of freely moving animals. The NO is directly oxidized on a carbon (or platinum) surface to NO⁺. This sensor is particularly sensitive to electroactive interferences in virtue of the very high oxidation potentials.

to investigate spatial processing and anxiety. Even in this case, the applied potential was -650 mV *vs* a silver wire REF^[154]. The CPA technique was also used by Bazzu *et al.*^[110] to monitor striatal oxygen levels in a telemetric *in vivo* study. Working electrodes, consisting of miniaturized conical-shaped epoxy-carbon electrodes (180 μm), allowed oxygen detection by fixing the reduction potential at -400 mV *vs* Ag/AgCl REF.

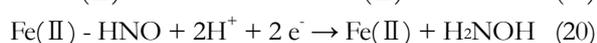
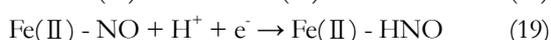
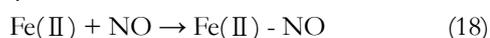
Recently, oxygen amperometry was applied to a behavioral study of reward processing in the rat nucleus accumbens. CPEs (200 μm in diameter) were used by applying a constant potential of -650 mV *vs* a silver wire REF to reduce oxygen. Data showed similar results to those obtained in human fMRI studies, confirming how oxygen amperometry is a powerful technique for the measurement of brain function^[155].

In the attempts of monitoring the concentration of the unstable nitric oxide molecule *in vivo* and to test nitric oxide donor drugs, several microsensors have been developed since the 1990s^[156]. The majority exploits electrochemical amperometric techniques to directly detect nitric oxide. Commonly, an oxidant potential is applied (higher than +850 mV *vs* Ag/AgCl), in view of the fact that nitric oxide and oxygen reduction potential are very close, so oxygen interferes with nitric oxide measurement (at nitric oxide-reducing potentials) (Figure 8).

Basically, a double reaction occurs at the transducer's face, usually carbon fiber or noble metals^[157-161], involving the formation of NO⁺, which is further converted into nitrite (Figure 8):



Otherwise, metalloporphyrin-modified sensors^[162-164] are also largely used:



Because of the enormous interest kindled by the wide

range of actions of nitric oxide, several *in vivo* experiments were conducted to monitor nitric oxide release on different tissues^[165-168]. Friedemann *et al.*^[169] developed an electrochemical electrode using carbon fiber as a transducer, coated with Nafion and further electropolymerized with OPD. Nitric oxide was quantified amperometrically using differential pulse voltammetry^[169].

Wu *et al.*^[170,171] research group conducted several experiments in which physiological nitric oxide actions on a cat's brain were investigated. Nitric oxide concentration was measured in real-time using voltammetry techniques, implanting Nafion-/porphyrin-/OPD-coated carbon-fiber electrodes. A highly sensitive and selective NO electrode was used to measure the nitric oxide concentration in a rat hippocampus^[172]. In addition, an electrochemical nitric oxide microbiosensor based on cytochrome C, immobilized onto a functionalized conducting polymer layer, was implanted in the striatum. Nafion was used for its shielding properties toward interference electroactive molecules present in the brain, chiefly ascorbic acid^[173]. Brown *et al.*^[174] and Finnerty *et al.*^[175] obtained a simple and useful design by modifying a Pt sensor with multicoated Nafion layers. This electrochemical sensor was successfully implanted in the striatum of freely moving rats, allowing the real-time nitric oxide at Nafion-coated Pt. Santos *et al.*^[176] recently developed an electrochemical biomimetic sensor based on nanocomposite hemin-based microelectrode, measuring exogenous NO in the rat hippocampus *in vivo* using CV.

Ethanol

In the last decades, ethanol has become the most widespread psychotropic toxic substance in Western countries because it is widely legally accepted and also because it is available at a low cost. Acute, subacute and chronic exposure to ethanol may have important effects on the CNS, therefore it becomes significant to monitor ethanol kinetic and its effects on the brain using the most appropriate techniques^[177]. The main effects of ethanol consumption cause significant effects on the CNS, principally enhancing the action of the neurotransmitter GABA and generating disinhibition, ataxia, and sedation^[178]. Subchronic exposure to ethanol enhances the dopamine neurotransmission in the mesolimbic system^[179,180] and increases dopamine levels in the nucleus accumbens^[181], playing an important role as a "rewarding" molecule^[182-184].

Recently, implantable electrochemical biosensors have been developed for monitoring the real-time changes of ethanol concentrations in the brain ECFs of freely moving animals (Figure 9). As previously described for other implantable biosensors, the ethanol biosensor exploits the presence of an enzyme, the alcohol oxidase, to selectively quantify ethanol using the production of a directly oxidizable byproduct (hydrogen peroxide), electrochemically detectable on the surface of a Pt transducer^[185,186]. The main characteristic of this biosensor is its capability of monitoring ethanol changes second by second and over

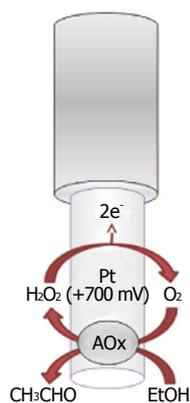


Figure 9 Schematic representation of the biosensor for the detection of exogenous ethanol in the brain of freely moving animals. EtOH: Ethanol; CH₃CHO: Acetaldehyde; AOx: Alcohol oxidase.

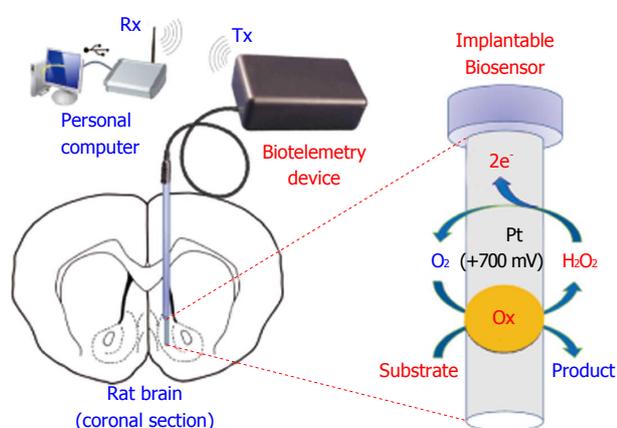


Figure 10 Schematic representation of the biotelemetry system, connected to a constant potential amperometry-based amperometric biosensor, for the real-time monitoring of brain neurochemistry in freely moving animals. Ox: Oxidase enzyme.

a period of two weeks. This neurochemical tool has been proven to be successful, especially when associated with a miniaturized telemetric system (see next paragraph). According to the results of previous studies^[177,185,186], the ethanol biosensor has been demonstrated to be a reliable device for the short-time monitoring of exogenous ethanol in the CNS, and it could be used for studying ethanol pharmacokinetics during addiction and the real-time effect of drugs on ethanol levels in the CNS.

BIOTELEMETRY

Biotelemetry has been defined as the recording of physiological parameters by uni- or bidirectional electromagnetic signals^[6,187], or more simply, it represents a variety of techniques intended for real-time monitoring of physiological parameters. Innovative biotelemetry systems (Figure 10) have been developed for studying brain neurochemistry^[188], in particular for monitoring CNS dopamine in freely moving animals^[189-191] and, more recently, in humans^[192]. The wireless detection of dopamine requires complex waveform generation and high-resolution synchronization; indeed, as previously shown, FSCV allows

the redox detection of dopamine up to ten times per second^[189-191]. Also chronoamperometry and differential pulse voltammetry techniques have been demonstrated to work in conjunction with telemetric devices^[158,193-196]; the resulting systems are very complex, not easily miniaturizable, and difficult to use in small rodents. On the contrary, non-pulsed techniques, such as CPA, free the microcontroller unit (MCU) from high-density calculations, allowing an increase in the number of implantable sensors and facilitating the miniaturization of the electronics^[109,197]. The battery-powered biotelemetric device (Figure 10), composed of an amperometric module, an MCU, and a transmitter, polarizes the sensors and sends sensor data to a receiving unit connected to a PC. The system electronics exhibits low power consumption, high stability, and good linear response^[3]. A CPA-based biotelemetry device may be easily interfaced with amperometric microsensors and biosensors^[6,109,197] and leave enough MCU computing power available for other tasks such as motion detection using inertial physical sensors. Indeed, in a previous study, we described this new approach with the simultaneous detection of brain glucose, lactate, and movements in real time using a biotelemetric device fixed to the head of a freely moving rat^[6].

COMPARISON BETWEEN VOLTAMMETRY AND MICRODIALYSIS

Although voltammetric techniques have been widely used in last decades, microdialysis still remains the “gold standard” for *in vivo* neurochemical study of the brain extracellular compartment. The advantages in using this technique include the possibility of measuring several neurochemicals at the same time with high sensitivity and very high selectivity, providing a more complete picture of the ECF. Its invasiveness, associated with low temporal resolution, and the necessity of using connecting tubes to carry out the experiments do not make it particularly suitable for monitoring fast neurochemical changes and do not allow the application of wireless techniques. As an alternative, electrochemical sensors are increasingly-used tools to study the neurochemical modifications in the ECF. The main characteristics of these devices are represented by very low invasiveness (carbon fibers in particular), when compared with microdialysis probes, and, most of all, their capability of monitoring variations of analytes in seconds or fractions. Furthermore, some electrochemical sensors have been demonstrated to be effective for weeks or months when implanted in the brain and, as described in this review, they are the optimal candidates for wireless detection. The Table 1 summarizes the principal characteristics of the main techniques indicated in this review.

CONCLUSION

Implantable (Bio)sensors, integrated into miniaturized telemetric systems, represent a new generation of analytical tools for studying brain neurochemistry of awake, freely

Table 1 Principal characteristics of the main techniques indicated in this review and used for *in vivo* monitoring of brain neurochemistry

Characteristics of the technique	Technique				
	Electrochemical techniques (voltammetry)				Microdialysis
	CPA	CA	DPV	FSCV	
Brain invasiveness	+	+	+	+	++
Selectivity	+	+	++	++	+++
Sensitivity	++	++	+	+	+++
Concentration range	nmol/L-mmol/L	nmol/L-mmol/L	nmol/L-mmol/L	nmol/L-mmol/L	fmol/L-mmol/L
Temporal resolution	++++	+++	++	+++	+
Spatial resolution	++	+++	++	+++	+
Monitoring period	d/wk	d/wk	d/wk	d/wk	h/d
Untethered detection	++	+	+	+	-

CPA: Constant potential amperometry; CA: Chronoamperometry; DPV: Differential pulse voltammetry; FSCV: Fast-scan cyclic voltammetry.

moving animals in real time. This approach, based on simple and inexpensive components, could be used as a rapid and reliable model for studying the physiology, the pathophysiology, and the effects of different drugs (or toxic compounds such as ethanol) on brain neurochemical systems.

REFERENCES

- 1 **Delgado JM**, DeFeudis FV, Roth RH, Ryugo DK, Mitruka BM. Dialytrode for long term intracerebral perfusion in awake monkeys. *Arch Int Pharmacodyn Ther* 1972; **198**: 9-21 [PMID: 4626478]
- 2 **Ungerstedt U**. In: Robinson TE and Justice JB (eds.). Microdialysis in the Neurosciences. Netherlands: Elsevier Science BV, 1991: 3-18
- 3 **Calia G**, Rocchitta G, Migheli R, Puggioni G, Spissu Y, Bazzu G, Mazzarello V, Lowry JP, O'Neill RD, Desole MS, Serra PA. Biotelemetric monitoring of brain neurochemistry in conscious rats using microsensors and biosensors. *Sensors* (Basel) 2009; **9**: 2511-2523 [PMID: 22574029 DOI: 10.3390/s90402511]
- 4 **Bazzu G**, Biosa A, Farina D, Spissu Y, Dedola S, Calia G, Puggioni G, Rocchitta G, Migheli R, Desole MS, Serra PA. Dual asymmetric-flow microdialysis for *in vivo* monitoring of brain neurochemicals. *Talanta* 2011; **85**: 1933-1940 [PMID: 21872041 DOI: 10.1016/j.talanta.2011.07.018]
- 5 **Serra PA**, Puggioni G, Bazzu G, Calia G, Migheli R, Rocchitta G. Design and construction of a distributed sensor NET for biotelemetric monitoring of brain energetic metabolism using microsensors and biosensors. In: Serra PA, editor. Croatia: Intech, 2010: 241-260 [DOI: 10.5772/7213]
- 6 **Rocchitta G**, Secchi O, Alvau MD, Farina D, Bazzu G, Calia G, Migheli R, Desole MS, O'Neill RD, Serra PA. Simultaneous telemetric monitoring of brain glucose and lactate and motion in freely moving rats. *Anal Chem* 2013; **85**: 10282-10288 [PMID: 24102201 DOI: 10.1021/ac402071w]
- 7 **Cooper JR**, Bloom FE, Roth RH, editors. The Biochemical Basis of Neuropharmacology. Eighth Edition, 2003
- 8 **Flagel SB**, Clark JJ, Robinson TE, Mayo L, Czuj A, Willuhn I, Akers CA, Clinton SM, Phillips PE, Akil H. A selective role for dopamine in stimulus-reward learning. *Nature* 2011; **469**: 53-57 [PMID: 21150898 DOI: 10.1038/nature09588]
- 9 **Wightman RM**, Robinson DL. Transient changes in mesolimbic dopamine and their association with 'reward'. *J Neurochem* 2002; **82**: 721-735 [PMID: 12358778 DOI: 10.1046/j.1471-4159.2002.01005.x]
- 10 **O'Neill RD**. Long-term monitoring of brain dopamine metabolism *in vivo* with carbon paste electrodes. *Sensors* 2005; **5**: 317-342 [DOI: 10.3390/s5060317]
- 11 **Bazzu G**, Rocchitta G, Migheli R, Alvau MD, Zinellu M, Puggioni G, Calia G, Mercanti G, Giusti P, Desole MS, Serra PA. Effects of the neurotoxin MPTP and pargyline protection on extracellular energy metabolites and dopamine levels in the striatum of freely moving rats. *Brain Res* 2013; **1538**: 159-171 [PMID: 24080403 DOI: 10.1016/j.brainres.2013.09.037]
- 12 **Serra PA**, Sciola L, Delogu MR, Spano A, Monaco G, Miele E, Rocchitta G, Miele M, Migheli R, Desole MS. The neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine induces apoptosis in mouse nigrostriatal glia. Relevance to nigral neuronal death and striatal neurochemical changes. *J Biol Chem* 2002; **277**: 34451-34461 [PMID: 12084711 DOI: 10.1074/jbc.M202099200]
- 13 **Serra PA**, Pluchino S, Marchetti B, Desole MS, Miele E. The MPTP mouse model: cues on DA release and neural stem cell restorative role. *Parkinsonism Relat Disord* 2008; **14** Suppl 2: S189-S193 [PMID: 18579428 DOI: 10.1016/j.parkreldis.2008.04.029]
- 14 **Howes OD**, Murray RM. Schizophrenia: an integrated socio-developmental-cognitive model. *Lancet* 2014; **383**: 1677-1687 [PMID: 24315522 DOI: 10.1016/S0140-6736(13)62036-X]
- 15 **Howes OD**, Kambeitz J, Kim E, Stahl D, Slifstein M, Abi-Dargham A, Kapur S. The nature of dopamine dysfunction in schizophrenia and what this means for treatment. *Arch Gen Psychiatry* 2012; **69**: 776-786 [PMID: 22474070 DOI: 10.1001/archgenpsychiatry.2012.169]
- 16 **Heneka MT**, Nadrigny F, Regen T, Martinez-Hernandez A, Dumitrescu-Ozimek L, Terwel D, Jardanhazi-Kurutz D, Walter J, Kirchhoff F, Hanisch UK, Kummer MP. Locus ceruleus controls Alzheimer's disease pathology by modulating microglial functions through norepinephrine. *Proc Natl Acad Sci USA* 2010; **107**: 6058-6063 [PMID: 20231476 DOI: 10.1073/pnas.0909586107]
- 17 **Fillenz M**. *In vivo* neurochemical monitoring and the study of behaviour. *Neurosci Biobehav Rev* 2005; **29**: 949-962 [PMID: 15963566 DOI: 10.1016/j.neubiorev.2005.02.003]
- 18 **Ressler KJ**, Nemeroff CB. Role of serotonergic and noradrenergic systems in the pathophysiology of depression and anxiety disorders. *Depress Anxiety* 2000; **12** Suppl 1: 2-19 [PMID: 11098410]
- 19 **Kermorgant M**, Lancien F, Mimassi N, Le Mével JC. Central ventilatory and cardiovascular actions of serotonin in trout. *Respir Physiol Neurobiol* 2014; **192**: 55-65 [PMID: 24325919 DOI: 10.1016/j.resp.2013.12.001]
- 20 **Robinson DL**, Hermans A, Seipel AT, Wightman RM. Monitoring rapid chemical communication in the brain. *Chem Rev* 2008; **108**: 2554-2584 [PMID: 18576692 DOI: 10.1021/cr068081q]
- 21 **Migheli R**, Puggioni G, Dedola S, Rocchitta G, Calia G, Bazzu G, Esposito G, Lowry JP, O'Neill RD, Desole MS, Miele E, Serra PA. Novel integrated microdialysis-amperometric sys-

- tem for in vitro detection of dopamine secreted from PC12 cells: design, construction, and validation. *Anal Biochem* 2008; **380**: 323-330 [PMID: 18577368 DOI: 10.1016/j.ab.2008.05.050]
- 22 **Nevalainen N**, Af Bjerken S, Lundblad M, Gerhardt GA, Strömberg I. Dopamine release from serotonergic nerve fibers is reduced in L-DOPA-induced dyskinesia. *J Neurochem* 2011; **118**: 12-23 [PMID: 21534956 DOI: 10.1111/j.1471-4159.2011.07292.x]
 - 23 **Gratton A**, Hoffer BJ, Gerhardt GA. In vivo electrochemical studies of monoamine release in the medial prefrontal cortex of the rat. *Neuroscience* 1989; **29**: 57-64 [PMID: 2710348 DOI: 10.1016/0306-4522(89)90332-1]
 - 24 **Ozel RE**, Wallace KN, Andreescu S. Chitosan coated carbon fiber microelectrode for selective in vivo detection of neurotransmitters in live zebrafish embryos. *Anal Chim Acta* 2011; **695**: 89-95 [PMID: 21601035 DOI: 10.1016/j.aca.2011.03.057]
 - 25 **Clark JJ**, Sandberg SG, Wanat MJ, Gan JO, Horne EA, Hart AS, Akers CA, Parker JG, Willuhn I, Martinez V, Evans SB, Stella N, Phillips PE. Chronic microsensors for longitudinal, subsecond dopamine detection in behaving animals. *Nat Methods* 2010; **7**: 126-129 [PMID: 20037591 DOI: 10.1038/nmeth.1412]
 - 26 **Willuhn I**, Burgeno LM, Everitt BJ, Phillips PE. Hierarchical recruitment of phasic dopamine signaling in the striatum during the progression of cocaine use. *Proc Natl Acad Sci USA* 2012; **109**: 20703-20708 [PMID: 23184975 DOI: 10.1073/pnas.1213460109]
 - 27 **John CE**, Jones SR. Fast scan cyclic voltammetry of dopamine and serotonin in mouse brain slices. In: Michael AC, Borland LM (eds.), *Electrochemical Methods for Neuroscience*. Boca Raton (FL): CRC Press, 2006: 49-62 [PMID: 21204393 DOI: 10.1201/9781420005868.ch4]
 - 28 **Peters JL**, Miner LH, Michael AC, Sesack SR. Ultrastructure at carbon fiber microelectrode implantation sites after acute voltammetric measurements in the striatum of anesthetized rats. *J Neurosci Methods* 2004; **137**: 9-23 [PMID: 15196823 DOI: 10.1016/j.jneumeth.2004.02.006]
 - 29 **Dressman SF**, Peters JL, Michael AC. Carbon fiber microelectrodes with multiple sensing elements for in vivo voltammetry. *J Neurosci Methods* 2002; **119**: 75-81 [PMID: 12234638 DOI: 10.1016/S0165-0270(02)00180-2]
 - 30 **Kawagoe KT**, Zimmerman JB, Wightman RM. Principles of voltammetry and microelectrode surface states. *J Neurosci Methods* 1993; **48**: 225-240 [PMID: 8412305 DOI: 10.1016/0165-0270(93)90094-8]
 - 31 **Seymour JP**, Kipke DR. Neural probe design for reduced tissue encapsulation in CNS. *Biomaterials* 2007; **28**: 3594-3607 [PMID: 17517431 DOI: 10.1016/j.biomaterials.2007.03.024]
 - 32 **Park J**, Takmakov P, Wightman RM. In vivo comparison of norepinephrine and dopamine release in rat brain by simultaneous measurements with fast-scan cyclic voltammetry. *J Neurochem* 2011; **119**: 932-944 [PMID: 21933188 DOI: 10.1111/j.1471-4159.2011.07494.x]
 - 33 **Heien ML**, Johnson MA, Wightman RM. Resolving neurotransmitters detected by fast-scan cyclic voltammetry. *Anal Chem* 2004; **76**: 5697-5704 [PMID: 15456288 DOI: 10.1021/ac0491509]
 - 34 **Park J**, Kile BM, Wightman RM. In vivo voltammetric monitoring of norepinephrine release in the rat ventral bed nucleus of the stria terminalis and anteroventral thalamic nucleus. *Eur J Neurosci* 2009; **30**: 2121-2133 [PMID: 20128849 DOI: 10.1111/j.1460-9568.2009.07005.x]
 - 35 **Hashemi P**, Dankoski EC, Wood KM, Ambrose RE, Wightman RM. In vivo electrochemical evidence for simultaneous 5-HT and histamine release in the rat substantia nigra pars reticulata following medial forebrain bundle stimulation. *J Neurochem* 2011; **118**: 749-759 [PMID: 21682723]
 - 36 **Herr NR**, Park J, McElligott ZA, Belle AM, Carelli RM, Wightman RM. In vivo voltammetry monitoring of electrically evoked extracellular norepinephrine in subregions of the bed nucleus of the stria terminalis. *J Neurophysiol* 2012; **107**: 1731-1737 [PMID: 22190618 DOI: 10.1152/jn.00620.2011]
 - 37 **Mefford IN**, Oke AF, Adams RN. Regional distribution of ascorbate in human brain. *Brain Res* 1981; **212**: 223-226 [PMID: 7225858 DOI: 10.1016/0006-8993(81)90056-1]
 - 38 **Nagy G**, Rice ME, Adams RN. A new type of enzyme electrode: the ascorbic acid eliminator electrode. *Life Sci* 1982; **31**: 2611-2616 [PMID: 6130453 DOI: 10.1016/0024-3205(82)90736-6]
 - 39 **Willuhn I**, Wanat MJ, Clark JJ, Phillips PE. Dopamine signaling in the nucleus accumbens of animals self-administering drugs of abuse. *Curr Top Behav Neurosci* 2010; **3**: 29-71 [PMID: 21161749 DOI: 10.1007/7854_2009_27]
 - 40 **Njagi J**, Chernov MM, Leiter JC, Andreescu S. Amperometric detection of dopamine in vivo with an enzyme based carbon fiber microbiosensor. *Anal Chem* 2010; **82**: 989-996 [PMID: 20055419 DOI: 10.1021/ac9022605]
 - 41 **Sarter M**, Bruno JP, Givens B. Attentional functions of cortical cholinergic inputs: what does it mean for learning and memory? *Neurobiol Learn Mem* 2003; **80**: 245-256 [PMID: 14521867 DOI: 10.1016/S1074-7427(03)00070-4]
 - 42 **Uutela P**, Reinilä R, Piepponen P, Ketola RA, Kostianen R. Analysis of acetylcholine and choline in microdialysis samples by liquid chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom* 2005; **19**: 2950-2956 [PMID: 16180202 DOI: 10.1002/rcm.2160]
 - 43 **Watanabe T**, Yamagata N, Takasaki K, Sano K, Hayakawa K, Katsurabayashi S, Egashira N, Mishima K, Iwasaki K, Fujiwara M. Decreased acetylcholine release is correlated to memory impairment in the Tg2576 transgenic mouse model of Alzheimer's disease. *Brain Res* 2009; **1249**: 222-228 [PMID: 18996097 DOI: 10.1016/j.brainres.2008.10.029]
 - 44 **Anagnostaras SG**, Murphy GG, Hamilton SE, Mitchell SL, Rahnema NP, Nathanson NM, Silva AJ. Selective cognitive dysfunction in acetylcholine M1 muscarinic receptor mutant mice. *Nat Neurosci* 2003; **6**: 51-58 [PMID: 12483218 DOI: 10.1038/nn992]
 - 45 **Dani JA**, Ji D, Zhou FM. Synaptic plasticity and nicotine addiction. *Neuron* 2001; **31**: 349-352 [PMID: 11516393 DOI: 10.1016/S0896-6273(01)00379-8]
 - 46 **Klucken J**, McLean PJ, Gomez-Tortosa E, Ingelsson M, Hyman BT. Neuritic alterations and neural system dysfunction in Alzheimer's disease and dementia with Lewy bodies. *Neurochem Res* 2003; **28**: 1683-1691 [PMID: 14584822 DOI: 10.1023/A:1026061021946]
 - 47 **Calabresi P**, Picconi B, Parnetti L, Di Filippo M. A convergent model for cognitive dysfunctions in Parkinson's disease: the critical dopamine-acetylcholine synaptic balance. *Lancet Neurol* 2006; **5**: 974-983 [PMID: 17052664 DOI: 10.1016/S1474-4422(06)70600-7]
 - 48 **Zhu W**, Wang D, Zheng J, An Y, Wang Q, Zhang W, Jin L, Gao H, Lin L. Effect of (R)-salsolinol and N-methyl-(R)-salsolinol on the balance impairment between dopamine and acetylcholine in rat brain: involvement in pathogenesis of Parkinson disease. *Clin Chem* 2008; **54**: 705-712 [PMID: 18238832 DOI: 10.1373/clinchem.2007.097725]
 - 49 **Kuo MF**, Grosch J, Fregni F, Paulus W, Nitsche MA. Focusing effect of acetylcholine on neuroplasticity in the human motor cortex. *J Neurosci* 2007; **27**: 14442-14447 [PMID: 18160652 DOI: 10.1523/JNEUROSCI.4104-07.2007]
 - 50 **Hildebrandt A**, Brago's R, Lacorte S, Marty JL. Performance of a portable biosensor for the analysis of organophosphorus and carbamate insecticides in water and food. *Sens Actuators B Chem* 2008; **133**: 195-201 [DOI: 10.1016/j.snb.2008.02.017]
 - 51 **Mitchell KM**. Acetylcholine and choline amperometric enzyme sensors characterized in vitro and in vivo. *Anal Chem* 2004; **76**: 1098-1106 [PMID: 14961744 DOI: 10.1021/ac034757v]
 - 52 **Khan A**, Ab Ghani S. Multienzyme microbiosensor based on electropolymerized o-phenylenediamine for simultaneous in vitro determination of acetylcholine and choline. *Biosens*

- Bioelectron* 2012; **31**: 433-438 [PMID: 22154168 DOI: 10.1016/j.bios.2011.11.007]
- 53 **Chen Q**, Kobayashi Y, Takeshita H, Hoshi T, Anzai J. Avadin biotin system-based enzyme multilayer membranes for biosensor application: Optimization of loading of choline esterase and choline oxidase in the bienzyme membrane for acetylcholine biosensors. *Electroanalysis* 1998; **10**: 94-97
- 54 **Sarter M**, Parikh V. Choline transporters, cholinergic transmission and cognition. *Nat Rev Neurosci* 2005; **6**: 48-56 [PMID: 15611726 DOI: 10.1038/nrn1588]
- 55 **Zamfir LG**, Rotariu L, Bala C. Acetylcholinesterase biosensor for carbamate drugs based on tetrathiafulvalene-tetracyanoquinodimethane/ionic liquid conductive gels. *Biosens Bioelectron* 2013; **46**: 61-67 [PMID: 23500478 DOI: 10.1016/j.bios.2013.02.018]
- 56 **Wu BY**, Hou SH, Yin F, Zhao ZX, Wang YY, Wang XS, Chen Q. Amperometric glucose biosensor based on multilayer films via layer-by-layer self-assembly of multi-wall carbon nanotubes, gold nanoparticles and glucose oxidase on the Pt electrode. *Biosens Bioelectron* 2007; **22**: 2854-2860 [PMID: 17212983 DOI: 10.1016/j.bios.2006.11.028]
- 57 **Garguilo MG**, Michael AC. Amperometric microsensors for monitoring choline in the extracellular fluid of brain. *J Neurosci Methods* 1996; **70**: 73-82 [PMID: 8982984 DOI: 10.1016/S0165-0270(96)00105-7]
- 58 **Cammack J**, Ghasemzadeh B, Adams RN. Electrochemical monitoring of brain ascorbic acid changes associated with hypoxia, spreading depression, and seizure activity. *Neurochem Res* 1992; **17**: 23-27 [PMID: 1347161 DOI: 10.1007/BF00966861]
- 59 **Curulli A**, Dragulescu S, Cremisini C, Palleschi G. Bienzyme amperometric probes for choline and choline esters assembled with nonconducting electrosynthesized polymers. *Electroanalysis* 2001; **13**: 236-242
- 60 **Hawkins RA**. The blood-brain barrier and glutamate. *Am J Clin Nutr* 2009; **90**: 867S-874S [PMID: 19571220 DOI: 10.3945/ajcn.2009.27462BB]
- 61 **Sundaram RS**, Gowtham L, Nayak BS. The role of excitatory neurotransmitter glutamate in brain physiology and pathology. *Asian J Pharm Clin Res* 2012; **5**: 1-7
- 62 **Suzuki K**, Martin PM. Neurotoxicants and developing brain. In: Harry GJ, editor. *Developmental Neurotoxicology*. Boca Raton: CRC Press, 1994: 9-32
- 63 **Hirsch SR**, Das I, Garey LJ, de Bellerocche J. A pivotal role for glutamate in the pathogenesis of schizophrenia, and its cognitive dysfunction. *Pharmacol Biochem Behav* 1997; **56**: 797-802 [PMID: 9130307 DOI: 10.1016/S0091-3057(96)00428-5]
- 64 **Lau A**, Tymianski M. Glutamate receptors, neurotoxicity and neurodegeneration. *Pflugers Arch* 2010; **460**: 525-542 [PMID: 20229265 DOI: 10.1007/s00424-010-0809-1]
- 65 **McLamore ES**, Mohanty S, Shi J, Claussen J, Jedlicka SS, Rickus JL, Porterfield DM. A self-referencing glutamate biosensor for measuring real time neuronal glutamate flux. *J Neurosci Methods* 2010; **189**: 14-22 [PMID: 20298719 DOI: 10.1016/j.jneumeth.2010.03.001]
- 66 **Batra B**, Pundir CS. An amperometric glutamate biosensor based on immobilization of glutamate oxidase onto carboxylated multiwalled carbon nanotubes/gold nanoparticles/chitosan composite film modified Au electrode. *Biosens Bioelectron* 2013; **47**: 496-501 [PMID: 23628843 DOI: 10.1016/j.bios.2013.03.063]
- 67 **Tolosa VM**, Wassum KM, Maidment NT, Monbouquette HG. Electrochemically deposited iridium oxide reference electrode integrated with an electroenzymatic glutamate sensor on a multi-electrode array microprobe. *Biosens Bioelectron* 2013; **42**: 256-260 [PMID: 23208095 DOI: 10.1016/j.bios.2012.10.061]
- 68 **Wassum KM**, Tolosa VM, Tseng TC, Balleine BW, Monbouquette HG, Maidment NT. Transient extracellular glutamate events in the basolateral amygdala track reward-seeking actions. *J Neurosci* 2012; **32**: 2734-2746 [PMID: 22357857 DOI: 10.1523/JNEUROSCI.5780-11.2012]
- 69 **Frey O**, Holtzman T, McNamara RM, Theobald DE, van der Wal PD, de Rooij NF, Dalley JW, Koudelka-Hep M. Enzyme-based choline and L-glutamate biosensor electrodes on silicon microprobe arrays. *Biosens Bioelectron* 2010; **26**: 477-484 [PMID: 20705443 DOI: 10.1016/j.bios.2010.07.073]
- 70 **Wahono N**, Qin S, Oomen P, Cremers TI, de Vries MG, Westerink BH. Evaluation of permselective membranes for optimization of intracerebral amperometric glutamate biosensors. *Biosens Bioelectron* 2012; **33**: 260-266 [PMID: 22326702 DOI: 10.1016/j.bios.2012.01.019]
- 71 **Rothwell SA**, Kinsella ME, Zain ZM, Serra PA, Rocchitta G, Lowry JP, O'Neill RD. Contributions by a novel edge effect to the permselectivity of an electrosynthesized polymer for microbiosensor applications. *Anal Chem* 2009; **81**: 3911-3918 [PMID: 19371060 DOI: 10.1021/ac900162c]
- 72 **Tian F**, Gourine AV, Huckstepp RT, Dale N. A microelectrode biosensor for real time monitoring of L-glutamate release. *Anal Chim Acta* 2009; **645**: 86-91 [PMID: 19481635 DOI: 10.1016/j.aca.2009.04.048]
- 73 **McMahon CP**, Rocchitta G, Kirwan SM, Killoran SJ, Serra PA, Lowry JP, O'Neill RD. Oxygen tolerance of an implantable polymer/enzyme composite glutamate biosensor displaying polycation-enhanced substrate sensitivity. *Biosens Bioelectron* 2007; **22**: 1466-1473 [PMID: 16887344 DOI: 10.1016/j.bios.2006.06.027]
- 74 **McMahon CP**, Rocchitta G, Serra PA, Kirwan SM, Lowry JP, O'Neill RD. Control of the oxygen dependence of an implantable polymer/enzyme composite biosensor for glutamate. *Anal Chem* 2006; **78**: 2352-2359 [PMID: 16579619 DOI: 10.1021/ac0518194]
- 75 **Rahman MA**, Kwon NH, Won MS, Choe ES, Shim YB. Functionalized conducting polymer as an enzyme-immobilizing substrate: an amperometric glutamate microbiosensor for in vivo measurements. *Anal Chem* 2005; **77**: 4854-4860 [PMID: 16053298 DOI: 10.1021/ac050558v]
- 76 **Miele M**, Boutelle MG, Fillenz M. The source of physiologically stimulated glutamate efflux from the striatum of conscious rats. *J Physiol* 1996; **497** (Pt 3): 745-751 [PMID: 9003559]
- 77 **Hamdi N**, Wang J, Monbouquette HG. Polymer films as permselective coatings for H₂O₂-sensing electrodes. *J Electroanal Chem* 2005; **581**: 258-264 [DOI: 10.1016/j.jelechem.2005.04.028]
- 78 **O'Neill RD**, Chang SC, Lowry JP, McNeil CJ. Comparisons of platinum, gold, palladium and glassy carbon as electrode materials in the design of biosensors for glutamate. *Biosens Bioelectron* 2004; **19**: 1521-1528 [PMID: 15093225 DOI: 10.1016/j.bios.2003.12.004]
- 79 **Yao T**, Okano G. Simultaneous determination of L-glutamate, acetylcholine and dopamine in rat brain by a flow-injection biosensor system with microdialysis sampling. *Anal Sci* 2008; **24**: 1469-1473 [PMID: 18997377 DOI: 10.2116/analsci.24.1469]
- 80 **Harrison FE**, May JM. Vitamin C function in the brain: vital role of the ascorbate transporter SVCT2. *Free Radic Biol Med* 2009; **46**: 719-730 [PMID: 19162177 DOI: 10.1016/j.freeradbiomed.2008.12.018]
- 81 **May JM**. Vitamin C transport and its role in the central nervous system. *Subcell Biochem* 2012; **56**: 85-103 [PMID: 22116696 DOI: 10.1007/978-94-007-2199-9_6]
- 82 **Savini I**, Rossi A, Pierro C, Avigliano L, Catani MV. SVCT1 and SVCT2: key proteins for vitamin C uptake. *Amino Acids* 2008; **34**: 347-355 [PMID: 17541511 DOI: 10.1007/s00726-007-0555-7]
- 83 **Vera JC**, Rivas CI, Fischbarg J, Golde DW. Mammalian facilitative hexose transporters mediate the transport of dehydroascorbic acid. *Nature* 1993; **364**: 79-82 [PMID: 8316303 DOI: 10.1038/364079a0]
- 84 **Rice ME**, Russo-Menna I. Differential compartmentalization of brain ascorbate and glutathione between neurons and glia. *Neuroscience* 1998; **82**: 1213-1223 [PMID: 9466441]

- 85 **Rice ME.** Ascorbate regulation and its neuroprotective role in the brain. *Trends Neurosci* 2000; **23**: 209-216 [PMID: 10782126 DOI: 10.1016/S0166-2236(99)01543-X]
- 86 **O'Neill RD.** The measurement of brain ascorbate in vivo and its link with excitatory amino acid neurotransmission. In: Boulton A, Baker G, Adams RN (eds.). Humana Press Inc, 221-268
- 87 **Miele M, Boutelle MG, Fillenz M.** The physiologically induced release of ascorbate in rat brain is dependent on impulse traffic, calcium influx and glutamate uptake. *Neuroscience* 1994; **62**: 87-91 [PMID: 7816214 DOI: 10.1016/0306-4522(94)90316-6]
- 88 **O'Neill RD, Fillenz M, Sundstrom L, Rawlins JN.** Voltammetrically monitored brain ascorbate as an index of excitatory amino acid release in the unrestrained rat. *Neurosci Lett* 1984; **52**: 227-233 [PMID: 6521967]
- 89 **Hocevar SB, Zivin M, Milutinovic A, Hawlina M, Hutton E, Ogorevc B.** Simultaneous in vivo measurement of dopamine, serotonin and ascorbate in the striatum of experimental rats using voltammetric microprobe. *Front Biosci* 2006; **11**: 2782-2789 [PMID: 16720351]
- 90 **Gonon F, Buda M, Cespuglio R, Jouvét M, Pujol JF.** Voltammetry in the striatum of chronic freely moving rats: detection of catechols and ascorbic acid. *Brain Res* 1981; **223**: 69-80 [PMID: 7284811]
- 91 **Zhang M, Liu K, Xiang L, Lin Y, Su L, Mao L.** Carbon nanotube-modified carbon fiber microelectrodes for in vivo voltammetric measurement of ascorbic acid in rat brain. *Anal Chem* 2007; **79**: 6559-6565 [PMID: 17676820 DOI: 10.1021/ac0705871]
- 92 **Chen A, Shah B.** Electrochemical sensing and biosensing based on square wave voltammetry. *Anal. Methods* 2013; **5**: 2158-2173 [DOI: 10.1039/c3ay40155c]
- 93 **Miele M, Fillenz M.** In vivo determination of extracellular brain ascorbate. *J Neurosci Methods* 1996; **70**: 15-19 [PMID: 8982976 DOI: 10.1016/S0165-0270(96)00094-5]
- 94 **Yao T, Yano T, Nanjyo Y, Nishino H.** Simultaneous determination of glucose and L-lactate in rat brain by an electrochemical in vivo flow-injection system with an on-line microdialysis sampling. *Anal Sci* 2003; **19**: 61-65 [PMID: 12558025 DOI: 10.2116/analsci.19.61]
- 95 **Lowry JP, Miele M, O'Neill RD, Boutelle MG, Fillenz M.** An amperometric glucose-oxidase/poly(o-phenylenediamine) biosensor for monitoring brain extracellular glucose: in vivo characterisation in the striatum of freely-moving rats. *J Neurosci Methods* 1998; **79**: 65-74 [PMID: 9531461 DOI: 10.1016/S0165-0270(97)00171-4]
- 96 **Magistretti PJ, Pellerin L.** Cellular mechanisms of brain energy metabolism and their relevance to functional brain imaging. *Philos Trans R Soc Lond B Biol Sci* 1999; **354**: 1155-1163 [PMID: 10466143 DOI: 10.1098/rstb.1999.0471]
- 97 **Fillenz M.** The role of lactate in brain metabolism. *Neurochem Int* 2005; **47**: 413-417 [PMID: 16039756 DOI: 10.1016/j.neuint.2005.05.011]
- 98 **Chen C, Xie Q, Yang D, Xiao H, Fu Y, Tan Y, Yao S.** Recent advances in electrochemical glucose biosensors: a review. *RSC Adv* 2013; **3**: 4473-4491 [DOI: 10.1039/c2ra22351a]
- 99 **Manning CA, Honn VJ, Stone WS, Jane JS, Gold PE.** Glucose effects on cognition in adults with Down's syndrome. *Neuropsychology* 1998; **12**: 479-484 [PMID: 9674002 DOI: 10.1037/0894-4105.12.3.479]
- 100 **Manning CA, Ragozzino ME, Gold PE.** Glucose enhancement of memory in patients with probable senile dementia of the Alzheimer's type. *Neurobiol Aging* 1993; **14**: 523-528 [PMID: 8295654 DOI: 10.1016/0197-4580(93)90034-9]
- 101 **Lowry JP, Fillenz M.** Real-time monitoring of brain energy metabolism in vivo using microelectrochemical sensors: the effects of anesthesia. *Bioelectrochemistry* 2001; **54**: 39-47 [PMID: 11506973 DOI: 10.1016/S1567-5394(01)00109-8]
- 102 **Ahmad F, Yusof AP, Bainbridge M, Ab Ghani S.** The application of glucose biosensor in studying the effects of insulin and anti-hypertensive drugs towards glucose level in brain striatum. *Biosens Bioelectron* 2008; **23**: 1862-1868 [PMID: 18440218 DOI: 10.1016/j.bios.2008.03.006]
- 103 **Baker DA, Gough DA.** A continuous, implantable lactate sensor. *Anal Chem* 1995; **67**: 1536-1540 [DOI: 10.1021/ac00105a010]
- 104 **Parra A, Casero E, Vázquez L, Pariente F, E. Lorenzo E.** Design and characterization of a lactate biosensor based on immobilized lactate oxidase onto gold surfaces. *Anal Chim Acta* 2006; **555**: 308-315 [DOI: 10.1016/j.aca.2005.09.025]
- 105 **Rassaei L, Olthuis W, Tsujimura S, Sudhölter EJ, van den Berg A.** Lactate biosensors: current status and outlook. *Anal Bioanal Chem* 2014; **406**: 123-137 [PMID: 24037614 DOI: 10.1007/s00216-013-7307-1]
- 106 **Dixon BM, Lowry JP, O'Neill RD.** Characterization in vitro and in vivo of the oxygen dependence of an enzyme/polymer biosensor for monitoring brain glucose. *J Neurosci Methods* 2002; **119**: 135-142 [PMID: 12323417 DOI: 10.1016/S0165-0270(02)00170-X]
- 107 **Nesakumar N, Sethuraman S, Krishnan UM, Rayappan JB.** Fabrication of lactate biosensor based on lactate dehydrogenase immobilized on cerium oxide nanoparticles. *J Colloid Interface Sci* 2013; **410**: 158-164 [PMID: 24034216 DOI: 10.1016/j.jcis.2013.08.009]
- 108 **Jaffari SA, Turner AP.** Recent advances in amperometric glucose biosensors for in vivo monitoring. *Physiol Meas* 1995; **16**: 1-15 [PMID: 7749351 DOI: 10.1088/0967-3334/16/1/001]
- 109 **Serra PA, Rocchitta G, Bazzu G, Manca A, Puggioni GM, Lowry JP, O'Neill RD.** Design and construction of a low cost single-supply embedded telemetry system for amperometric biosensor applications. *Sens Actuat B* 2007; **122**: 118-126 [DOI: 10.1016/j.snb.2006.05.013]
- 110 **Bazzu G, Puggioni GG, Dedola S, Calia G, Rocchitta G, Migheli R, Desole MS, Lowry JP, O'Neill RD, Serra PA.** Real-time monitoring of brain tissue oxygen using a miniaturized biotelemetric device implanted in freely moving rats. *Anal Chem* 2009; **81**: 2235-2241 [PMID: 19222224 DOI: 10.1021/ac802390f]
- 111 **Van Gompel JJ, Chang SY, Goerss SJ, Kim IY, Kimble C, Bennet KE, Lee KH.** Development of intraoperative electrochemical detection: wireless instantaneous neurochemical concentration sensor for deep brain stimulation feedback. *Neurosurg Focus* 2010; **29**: E6 [PMID: 20672923 DOI: 10.3171/2010.5.FOCUS10110]
- 112 **Liu J, Wang J.** A novel improved design for the first-generation glucose biosensor. *Food Technol Biotechnol* 2001; **39**: 55-58
- 113 **Stoytcheva M, Zlatev R, Velkova Z, Valdez B, Ovalle M.** Analytical characteristics of electrochemical biosensors. *Portugaliae Electrochim Acta* 2009; **27**: 353-362 [DOI: 10.4152/pea.200903353]
- 114 **McMahon CP, Killoran SJ, O'Neill RD.** Design variations of a polymer-enzyme composite biosensor for glucose: enhanced analyte sensitivity without increased oxygen dependence. *J Electroanal Chem* 2005; **580**: 193-202 [DOI: 10.1016/j.jelechem.2005.03.026]
- 115 **Kealy J, Bennett R, Lowry JP.** Simultaneous recording of hippocampal oxygen and glucose in real time using constant potential amperometry in the freely-moving rat. *J Neurosci Methods* 2013; **215**: 110-120 [PMID: 23499196 DOI: 10.1016/j.jneumeth.2013.02.016]
- 116 **O'Neill RD, Lowry JP, Rocchitta G, McMahon CP, Serra PA.** Designing sensitive and selective polymer/enzyme composite biosensors for brain monitoring in vivo. *Trends Anal Chem* 2008; **27**: 78-88 [DOI: 10.1016/j.trac.2007.11.008]
- 117 **Palmisano F, Zamboni PG.** Ascorbic acid interferences in hydrogen peroxide detecting biosensors based on electrochemically immobilized enzymes. *Anal Chem* 1993; **65**: 2690-2692 [DOI: 10.1021/ac00067a024]
- 118 **Karyakin AA, Gitelmacher OV, Karyakina EE.** A high sensitive glucose amperometric biosensor based on Prussian Blue modified electrodes. *Anal Lett* 1994; **27**: 2861-2869 [DOI:

- 10.1080/00032719408000297]
- 119 **Karyakin AA**, Karyakina EE, Gorton L. Prussian-Blue-based amperometric biosensors in flow-injection analysis. *Talanta* 1996; **43**: 1597-1606 [PMID: 18966641 DOI: 10.1016/0039-9140(96)01909-1]
 - 120 **Huang J**, Song Z, Li J, Yang Y, Shi H, Wu B, Anzai J, Osa T, Chen Q. A highly-sensitive L-lactate biosensor based on sol-gel film combined with multi-walled carbon nanotubes (MWCNTs) modified electrode. *Material Science and Engineering: C* 2007; **27**: 29-34 [DOI: 10.1016/j.msec.2006.01.001]
 - 121 **Salazar P**, O'Neill RD, Martín M, Rochea R, González-Mora JL. Amperometric glucose microbiosensor based on a Prussian Blue modified carbon fiber electrode for physiological applications. *Sensors and Actuators B* 2011; **152**: 137-143 [DOI: 10.1016/j.snb.2010.11.056]
 - 122 **Salazar P**, Martín M, O'Neill RD, Roche R, González-Mora JL. Biosensors based on Prussian blue modified carbon fibers electrodes for monitoring lactate in the extracellular space of brain tissue. *Int J Electrochem Sci* 2012; **7**: 5910-5926
 - 123 **Salazar P**, Martín M, Roche R, González-Mora JL, O'Neill RD. Microbiosensors for glucose based on Prussian Blue modified carbon fiber electrodes for in vivo monitoring in the central nervous system. *Biosens Bioelectron* 2010; **26**: 748-753 [PMID: 20656470 DOI: 10.1016/j.bios.2010.06.045]
 - 124 **Plaxco KW**, Soh HT. Switch-based biosensors: a new approach towards real-time, in vivo molecular detection. *Trends Biotechnol* 2011; **29**: 1-5 [PMID: 21106266 DOI: 10.1016/j.tibtech.2010.10.005]
 - 125 **Wilson GS**, Gifford R. Biosensors for real-time in vivo measurements. *Biosens Bioelectron* 2005; **20**: 2388-2403 [PMID: 15854814 DOI: 10.1016/j.bios.2004.12.003]
 - 126 **Schultz JE**. Brain Energy Metabolism. Von B. K. Siesjö, John Wiley and Sons, Chichester, New York, Brisbane Toronto, 1978, 607 S., DM 78,65. *Pharmazie in unserer Zeit* 1978; **7**: 192 [DOI: 10.1002/pauz.19780070610]
 - 127 **Bardt TF**, Unterberg AW, Härtl R, Kiening KL, Schneider GH, Lanksch WR. Monitoring of brain tissue PO₂ in traumatic brain injury: effect of cerebral hypoxia on outcome. *Acta Neurochir Suppl* 1998; **71**: 153-156 [PMID: 9779171]
 - 128 **Beck T**, Kriegelstein J. Cerebral circulation, metabolism, and blood-brain barrier of rats in hypocapnic hypoxia. *Am J Physiol* 1987; **252** (3 Pt 2): H504-H512
 - 129 **Carraway MS**, Piantadosi CA. Oxygen toxicity. *Respir Care Clin N Am* 1999; **5**: 265-295 [PMID: 10333451]
 - 130 **Zuo L**, Motherwell MS. The impact of reactive oxygen species and genetic mitochondrial mutations in Parkinson's disease. *Gene* 2013; **532**: 18-23 [PMID: 23954870 DOI: 10.1016/j.gene.2013.07.085]
 - 131 **Moncada S**, Palmer RM, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 1991; **43**: 109-142 [PMID: 1852778]
 - 132 **Palmer RM**. The L-arginine: nitric oxide pathway. *Curr Opin Nephrol Hypertens* 1993; **2**: 122-128 [PMID: 7522910]
 - 133 **Moncada S**, Palmer RM, Higgs EA. Biosynthesis of nitric oxide from L-arginine. A pathway for the regulation of cell function and communication. *Biochem Pharmacol* 1989; **38**: 1709-1715 [PMID: 2567594 DOI: 10.1016/0006-2952(89)90403-6]
 - 134 **Ignarro LJ**. Nitric oxide. A novel signal transduction mechanism for transcellular communication. *Hypertension* 1990; **16**: 477-483 [PMID: 1977698]
 - 135 **Ignarro LJ**, Buga GM, Wood KS, Byrns RE, Chaudhuri G. Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc Natl Acad Sci USA* 1987; **84**: 9265-9269 [PMID: 2827174 DOI: 10.1073/pnas.84.24.9265]
 - 136 **Ignarro LJ**, Byrns RE, Buga GM, Wood KS. Endothelium-derived relaxing factor from pulmonary artery and vein possesses pharmacologic and chemical properties identical to those of nitric oxide radical. *Circ Res* 1987; **61**: 866-879 [PMID: 2890446]
 - 137 **Moncada S**, Radomski MW, Palmer RM. Endothelium-derived relaxing factor. Identification as nitric oxide and role in the control of vascular tone and platelet function. *Biochem Pharmacol* 1988; **37**: 2495-2501 [PMID: 3291879]
 - 138 **Holán V**, Krulová M, Zajícová A, Pindjácová J. Nitric oxide as a regulatory and effector molecule in the immune system. *Mol Immunol* 2002; **38**: 989-995 [PMID: 12009578 DOI: 10.1016/S0161-5890(02)00027-5]
 - 139 **Solomonson LP**. Nitric oxide. New discoveries, biomedical implications. *J Fla Med Assoc* 1991; **78**: 107-109 [PMID: 2026996]
 - 140 **Lowenstein CJ**, Hill SL, Lafond-Walker A, Wu J, Allen G, Landavere M, Rose NR, Herskowitz A. Nitric oxide inhibits viral replication in murine myocarditis. *J Clin Invest* 1996; **97**: 1837-1843 [PMID: 8621766 DOI: 10.1172/JCI118613]
 - 141 **Ignarro LJ**, Napoli C. Novel features of nitric oxide, endothelial nitric oxide synthase, and atherosclerosis. *Curr Diab Rep* 2005; **5**: 17-23 [PMID: 15663912]
 - 142 **Schmidt HH**, Warner TD, Ishii K, Sheng H, Murad F. Insulin secretion from pancreatic B cells caused by L-arginine-derived nitrogen oxides. *Science* 1992; **255**: 721-723 [PMID: 1371193 DOI: 10.1126/science.1371193]
 - 143 **Vincent SR**. Nitric oxide and arginine-evoked insulin secretion. *Science* 1992; **258**: 1376-1378 [PMID: 1455235]
 - 144 **Ward WK**, Wood MD, Slobodzian EP. Continuous amperometric monitoring of subcutaneous oxygen in rabbit by telemetry. *J Med Eng Technol* 2002; **26**: 158-167 [PMID: 12396331]
 - 145 **Holmström N**, Nilsson P, Carlsten J, Bowald S. Long-term in vivo experience of an electrochemical sensor using the potential step technique for measurement of mixed venous oxygen pressure. *Biosens Bioelectron* 1998; **13**: 1287-1295 [PMID: 9883563 DOI: 10.1016/S0956-5663(98)00091-8]
 - 146 **Bolger FC**, Lowry JP. Brain tissue oxygen: In vivo monitoring with carbon paste electrodes. *Sensors* 2005; **5**: 473-487 [DOI: 10.3390/s5110473]
 - 147 **O'Neill RD**, Grünewald RA, Fillenz M, Albery WJ. Linear sweep voltammetry with carbon paste electrodes in the rat striatum. *Neuroscience* 1982; **7**: 1945-1954 [PMID: 6127652]
 - 148 **Venton BJ**, Michael DJ, Wightman RM. Correlation of local changes in extracellular oxygen and pH that accompany dopaminergic terminal activity in the rat caudate-putamen. *J Neurochem* 2003; **84**: 373-381 [PMID: 12558999 DOI: 10.1046/j.1471-4159.2003.01527.x]
 - 149 **Ariansen JL**, Heien ML, Hermans A, Phillips PE, Hernadi I, Bermudez MA, Schultz W, Wightman RM. Monitoring extracellular pH, oxygen, and dopamine during reward delivery in the striatum of primates. *Front Behav Neurosci* 2012; **6**: 36 [PMID: 22783176 DOI: 10.3389/fnbeh.2012.00036]
 - 150 **Lowry JP**, Demestre M, Fillenz M. Relation between cerebral blood flow and extracellular glucose in rat striatum during mild hypoxia and hyperoxia. *Dev Neurosci* 1998; **20**: 52-58 [PMID: 9600390]
 - 151 **Lowry JP**, Boutelle MG, O'Neill RD, Fillenz M. Characterization of carbon paste electrodes in vitro for simultaneous amperometric measurement of changes in oxygen and ascorbic acid concentrations in vivo. *Analyst* 1996; **121**: 761-766 [PMID: 8763205]
 - 152 **Bolger FB**, McHugh SB, Bennett R, Li J, Ishiwari K, Francois J, Conley MW, Gilmour G, Bannerman DM, Fillenz M, Tricklebank M, Lowry JP. Characterisation of carbon paste electrodes for real-time amperometric monitoring of brain tissue oxygen. *J Neurosci Methods* 2011; **195**: 135-142 [PMID: 21115045 DOI: 10.1016/j.jneumeth.2010.11.013]
 - 153 **Finnerty NJ**, Bolger FB, Pålsson E, Lowry JP. An investigation of hypofrontality in an animal model of schizophrenia using real-time microelectrochemical sensors for glucose, oxygen, and nitric oxide. *ACS Chem Neurosci* 2013; **4**: 825-831 [PMID: 23578219 DOI: 10.1021/cn4000567]
 - 154 **McHugh SB**, Fillenz M, Lowry JP, Rawlins JN, Bannerman

- DM. Brain tissue oxygen amperometry in behaving rats demonstrates functional dissociation of dorsal and ventral hippocampus during spatial processing and anxiety. *Eur J Neurosci* 2011; **33**: 322-337 [PMID: 21105915 DOI: 10.1111/j.1460-9568.2010.07497.x]
- 155 **Francois J**, Conway MW, Lowry JP, Tricklebank MD, Gilmour G. Changes in reward-related signals in the rat nucleus accumbens measured by in vivo oxygen amperometry are consistent with fMRI BOLD responses in man. *Neuroimage* 2012; **60**: 2169-2181 [PMID: 22361256 DOI: 10.1016/j.neuroimage.2012.02.024]
- 156 **Shibuki K**. An electrochemical microprobe for detecting nitric oxide release in brain tissue. *Neurosci Res* 1990; **9**: 69-76 [PMID: 2175870 DOI: 10.1016/0168-0102(90)90048-J]
- 157 **Kashevskii AV**, Lei J, Safronov AY, Ikeda O. Electrocatalytic properties of meso-tetraphenylporphyrin cobalt for nitric oxide oxidation in methanolic solution and in Nafion® film. *J Electroanal Chem* 2002; **531**: 71-79 [DOI: 10.1016/S0022-0728(02)01048-3]
- 158 **Kashevskii AV**, Safronov A.Y, Ikeda O. Behaviors of H2TPP and CoTPPCL in Nafion® film and the catalytic activity for nitric oxide oxidation. *J Electroanal Chem* 2001; **510**: 86-95 [DOI: 10.1016/S0022-0728(01)00550-2]
- 159 **Mao L**, Shi G, Tian Y, Liu H, Jin L, Yamamoto K, Tao S, Jin J. A novel thin-layer amperometric detector based on chemically modified ring-disc electrode and its application for simultaneous measurements of nitric oxide and nitrite in rat brain combined with in vivo microdialysis. *Talanta* 1998; **46**: 1547-1556 [PMID: 18967286 DOI: 10.1016/S0039-9140(98)00027-7]
- 160 **Bedioui F**, Trevin S, Devynck J, Lantoine F, Brunet A, Devynck MA. Elaboration and use of nickel planar macrocyclic complex-based sensors for the direct electrochemical measurement of nitric oxide in biological media. *Biosens Bioelectron* 1997; **12**: 205-212 [PMID: 9115688 DOI: 10.1016/S0956-5663(97)85338-9]
- 161 **Trevin S**, Bedioui F, Devynck J. Electrochemical and spectrophotometric study of the behavior of electropolymerized nickel porphyrin films in the determination of nitric oxide in solution. *Talanta* 1996; **43**: 303-311 [PMID: 18966491 DOI: 10.1016/0039-9140(95)01752-6]
- 162 **Ricciardolo FL**, Vergnani L, Wiegand S, Ricci F, Manzoli N, Fischer A, Amadesi S, Fellin R, Geppetti P. Detection of nitric oxide release induced by bradykinin in guinea pig trachea and main bronchi using a porphyrinic microsensor. *Am J Respir Cell Mol Biol* 2000; **22**: 97-104 [PMID: 10615071]
- 163 **Zhang X**, Li H, Jin H, Ebin Z, Brodsky S, Goligorsky MS. Effects of homocysteine on endothelial nitric oxide production. *Am J Physiol Renal Physiol* 2000; **279**: F671-F678 [PMID: 10997917]
- 164 **Joshi MS**, Lancaster JR, Liu X, Ferguson TB. In situ measurement of nitric oxide production in cardiac isografts and rejecting allografts by an electrochemical method. *Nitric Oxide* 2001; **5**: 561-565 [PMID: 11730363 DOI: 10.1006/niox.2001.0369]
- 165 **Griveau S**, Dumézy C, Séguin J, Chabot GG, Scherman D, Bedioui F. In vivo electrochemical detection of nitric oxide in tumor-bearing mice. *Anal Chem* 2007; **79**: 1030-1033 [PMID: 17263331 DOI: 10.1021/ac061634c]
- 166 **Isik S**, Castillo J, Blöchl A, Csöregi E, Schuhmann W. Simultaneous detection of L-glutamate and nitric oxide from adherently growing cells at known distance using disk shaped dual electrodes. *Bioelectrochemistry* 2007; **70**: 173-179 [PMID: 16733097 DOI: 10.1016/j.bioelect.2006.03.037]
- 167 **Zhang X**. Real time and in vivo monitoring of nitric oxide by electrochemical sensors—from dream to reality. *Front Biosci* 2004; **9**: 3434-3446 [PMID: 15353368]
- 168 **Dalbasti T**, Kilinc E. Microelectrode for in vivo real-time detection of NO. *Methods Enzymol* 2005; **396**: 584-592 [PMID: 16291265 DOI: 10.1016/S0076-6879(05)96050-3]
- 169 **Friedemann MN**, Robinson SW, Gerhardt GA. o-Phenylene-diamine-modified carbon fiber electrodes for the detection of nitric oxide. *Anal Chem* 1996; **68**: 2621-2628 [PMID: 8694261 DOI: 10.1021/ac960093w]
- 170 **Wu WC**, Wang Y, Su CK, Chai CY. The nNOS/cGMP signal transducing system is involved in the cardiovascular responses induced by activation of NMDA receptors in the rostral ventrolateral medulla of cats. *Neurosci Lett* 2001; **310**: 121-124 [PMID: 11585582 DOI: 10.1016/S0304-3940(01)02100-0]
- 171 **Wu WC**, Wang Y, Kao LS, Tang FI, Chai CY. Nitric oxide reduces blood pressure in the nucleus tractus solitarius: a real time electrochemical study. *Brain Res Bull* 2002; **57**: 171-177 [PMID: 11849823 DOI: 10.1016/S0361-9230(01)00737-7]
- 172 **Zheng X**, Ning G, Yang Y. Study on the technology of nitric oxide (NO) detection in vitro and in vivo. *Clin Hemorheol Microcirc* 2006; **34**: 347-352 [PMID: 16543656]
- 173 **Alvin Koh WC**, Rahman MA, Choe ES, Lee DK, Shim YB. A cytochrome c modified-conducting polymer microelectrode for monitoring in vivo changes in nitric oxide. *Biosens Bioelectron* 2008; **23**: 1374-1381 [PMID: 18242975 DOI: 10.1016/j.bios.2007.12.008]
- 174 **Brown FO**, Finnerty NJ, Lowry JP. Nitric oxide monitoring in brain extracellular fluid: characterisation of Nafion-modified Pt electrodes in vitro and in vivo. *Analyst* 2009; **134**: 2012-2020 [PMID: 19768208]
- 175 **Finnerty NJ**, O'Riordan SL, Brown FO, Serra P, O'Neill RD, Lowry JP. In vivo characterisation of a Nafion®-modified Pt electrode for real-time nitric oxide monitoring in brain extracellular fluid. *Analytical Methods* 2012; **4**: 550-557
- 176 **Santos RM**, Rodrigues MS, Laranjinha J, Barbosa RM. Biomimetic sensor based on hemin/carbon nanotubes/chitosan modified microelectrode for nitric oxide measurement in the brain. *Biosens Bioelectron* 2013; **44**: 152-159 [PMID: 23419387 DOI: 10.1016/j.bios.2013.01.015]
- 177 **Rocchitta G**, Serra PA. Direct monitoring of ethanol in the brain. *OA Alcohol* 2013; **1**: 15
- 178 **Hardman JG**, Limbird LE. Goodman and Gilman's the Pharmacological Basis of Therapeutics, 11th ed. New York: McGraw-Hill, 2005
- 179 **Brodie MS**, Shefner SA, Dunwiddie TV. Ethanol increases the firing rate of dopamine neurons of the rat ventral tegmental area in vitro. *Brain Res* 1990; **508**: 65-69 [PMID: 2337793]
- 180 **Gessa GL**, Muntoni F, Collu M, Vargiu L, Mereu G. Low doses of ethanol activate dopaminergic neurons in the ventral tegmental area. *Brain Res* 1985; **348**: 201-203 [PMID: 2998561 DOI: 10.1016/0006-8993(85)90381-6]
- 181 **Theile JW**, Morikawa H, Gonzales RA, Morrisett RA. GABAergic transmission modulates ethanol excitation of ventral tegmental area dopamine neurons. *Neuroscience* 2011; **172**: 94-103 [PMID: 20974231 DOI: 10.1016/j.neuroscience.2010.10.046]
- 182 **Karahanian E**, Quintanilla ME, Tampier L, Rivera-Meza M, Bustamante D, Gonzalez-Lira V, Morales P, Herrera-Marschitz M, Israel Y. Ethanol as a prodrug: brain metabolism of ethanol mediates its reinforcing effects. *Alcohol Clin Exp Res* 2011; **35**: 606-612 [PMID: 21332529 DOI: 10.1111/j.1530-0277.2011.01439.x]
- 183 **Jamal M**, Ameno K, Kumihashi M, Ameno S, Kubota T, Wang W, Ijiri I. Microdialysis for the determination of acetaldehyde and ethanol concentrations in the striatum of freely moving rats. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003; **798**: 155-158 [PMID: 14630370]
- 184 **Munson PL**, Mueller RA, Breese GR. Principles of Pharmacology: Basic Concepts and Clinical Application, 1st ed. New York: Chapman and Hall Inc., 1995
- 185 **Secchi O**, Zinellu M, Spissu Y, Pirisinu M, Bazzu G, Migheli R, Desole MS, O'Neill RD, Serra PA, Rocchitta G. Further in vitro characterization of an implantable biosensor for ethanol monitoring in the brain. *Sensors (Basel)* 2013; **13**: 9522-9535 [PMID: 23881145 DOI: 10.3390/s130709522]

- 186 **Rocchitta G**, Secchi O, Alvau MD, Migheli R, Calia G, Bazzu G, Farina D, Desole MS, O'Neill RD, Serra PA. Development and characterization of an implantable biosensor for telemetric monitoring of ethanol in the brain of freely moving rats. *Anal Chem* 2012; **84**: 7072-7079 [PMID: 22823474 DOI: 10.1021/ac301253h]
- 187 **FCC (Federal Communication Commission) (U.S.A.)**. Amendment of Parts 2 and 95 of the Commission's Rules to Create a Wireless Medical Telemetry Service. Washington, DC, 2000
- 188 **Crespi F**, Dalessandro D, Annovazzi-Lodi V, Heidbreder C, Norgia M. In vivo voltammetry: from wire to wireless measurements. *J Neurosci Methods* 2004; **140**: 153-161 [PMID: 15589345 DOI: 10.1016/j.jneumeth.2004.06.018]
- 189 **Garris PA**, Ensmann R, Poehlman J, Alexander A, Langley PE, Sandberg SG, Greco PG, Wightman RM, Rebec GV. Wireless transmission of fast-scan cyclic voltammetry at a carbon-fiber microelectrode: proof of principle. *J Neurosci Methods* 2004; **140**: 103-115 [PMID: 15589340 DOI: 10.1016/j.jneumeth.2004.04.043]
- 190 **Garris PA**, Greco PG, Sandberg SG, Howes G, Pongmaytegul S, Heidenreich BA, Casto JM, Ensmann R, Poehlman J, Alexander A, Rebec GV. In: Michael AC, Borland LM. *Electrochemical Methods for Neuroscience- In vivo voltammetry with telemetry*. Boca Raton (FL): CRC Press, 2007: 233-260
- 191 **Johnson DA**, Wilson GS. In: Michael A.C, Borland LM. *Electrochemical Methods for Neuroscience- Telemetry for biosensor systems*. Boca Raton (FL): CRC Press, 2007: 451-464
- 192 **Kasasbeh A**, Lee K, Bieber A, Bennet K, Chang SY. Wireless neurochemical monitoring in humans. *Stereotact Funct Neurosurg* 2013; **91**: 141-147 [PMID: 23445903 DOI: 10.1159/000345111]
- 193 **Imeri L**, De Simoni MG, Giglio R, Clavenna A, Mancina M. Changes in the serotonergic system during the sleep-wake cycle: simultaneous polygraphic and voltammetric recordings in hypothalamus using a telemetry system. *Neuroscience* 1994; **58**: 353-358 [PMID: 7512239 DOI: 10.1016/0306-4522(94)90042-6]
- 194 **Imeri L**, Gemma C, De Simoni MG, Opp MR, Mancina M. Hypothalamic serotonergic activity correlates better with brain temperature than with sleep-wake cycle and muscle tone in rats. *Neuroscience* 1999; **89**: 1241-1246 [PMID: 10362311 DOI: 10.1016/S0306-4522(98)00395-9]
- 195 **De Simoni MG**, De Luigi A, Imeri L, Algeri S. Miniaturized optoelectronic system for telemetry of in vivo voltammetric signals. *J Neurosci Methods* 1990; **33**: 233-240 [PMID: 2232871 DOI: 10.1016/0165-0270(90)90027-D]
- 196 **Annovazzi-Lodi V**, Donati S. An optoelectronic interconnection for bidirectional transmission of biological signals. *IEEE Trans Biomed Eng* 1988; **35**: 595-606 [PMID: 3169810 DOI: 10.1109/10.4592]
- 197 **Rocchitta G**, Migheli R, Dedola S, Calia G, Desole MS, Miele E, Lowry JP, O'Neill RD, Serra PA. Development of a distributed, fully automated, bidirectional telemetry system for amperometric microsensor and biosensor applications. *Sensor Actuat B Chem* 2007; **126**: 700-709 [DOI: 10.1016/j.snb.2007.04.019]

P- Reviewers: Chawla M, Fang Y, Ju HX, Panagis G, Trohman R

S- Editor: Song XX **L- Editor:** A **E- Editor:** Liu SQ



GENERAL INFORMATION

World Journal of Pharmacology (*World J Pharmacol*, *WJP*, online ISSN 2220-3192, DOI: 10.5497) is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

Aim and scope

WJP covers topics concerning neuropsychiatric pharmacology, cerebrovascular pharmacology, geriatric pharmacology, anti-inflammatory and immunological pharmacology, antitumor pharmacology, anti-infective pharmacology, metabolic pharmacology, gastrointestinal and hepatic pharmacology, respiratory pharmacology, blood pharmacology, urinary and reproductive pharmacology, pharmacokinetics and pharmacodynamics, clinical pharmacology, and drug toxicology. The current columns of *WJP* include editorial, frontier, therapeutics advances, field of vision, mini-reviews, review, topic highlight, medical ethics, original articles, case report, clinical case conference (Clinicopathological conference), and autobiography.

We encourage authors to submit their manuscripts to *WJP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

WJP is edited and published by Baishideng Publishing Group (BPG). BPG has a strong professional editorial team composed of science editors, language editors and electronic editors. BPG currently publishes 43 OA clinical medical journals, including 42 in English, has a total of 15471 editorial board members or peer reviewers, and is a world first-class publisher.

Columns

The columns in the issues of *WJP* will include: (1) Editorial: The editorial board members are invited to make comments on an important topic in their field in terms of its current research status and future directions to lead the development of this discipline; (2) Frontier: The editorial board members are invited to select a highly cited cutting-edge original paper of his/her own to summarize major findings, the problems that have been resolved and remain to be resolved, and future research directions to help readers understand his/her important academic point of view and future research directions in the field; (3) Diagnostic Advances: The editorial board members are invited to write high-quality diagnostic advances in their field to improve the diagnostic skills of readers. The topic covers general clinical diagnosis, differential diagnosis, pathological diagnosis, laboratory diagnosis, imaging diagnosis, endoscopic diagnosis, biotechnological diagnosis, functional diagnosis, and physical diagnosis; (4) Therapeutics Advances: The editorial board members are invited to write high-quality therapeutic advances in their field to help improve the therapeutic skills of readers. The topic covers medication therapy, psychotherapy, physical therapy, replacement therapy, interventional therapy, minimally invasive therapy, endoscopic therapy, transplantation therapy, and surgical therapy; (5) Field of Vision: The editorial board members are invited to write commentaries on classic articles, hot topic articles, or latest articles to keep readers at the forefront of research and increase their levels of clinical research. Classic articles refer to papers that are included in Web of Knowledge and have received a large number of citations (ranking in the top 1%) after being published for more than years, reflecting the quality and impact of papers. Hot topic articles refer to papers that are included in Web of Knowledge and have received a large number of citations after being published for no more

than 2 years, reflecting cutting-edge trends in scientific research. Latest articles refer to the latest published high-quality papers that are included in PubMed, reflecting the latest research trends. These commentary articles should focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions. Basic information about the article to be commented (including authors, article title, journal name, year, volume, and inclusive page numbers); (6) Minireviews: The editorial board members are invited to write short reviews on recent advances and trends in research of molecular biology, genomics, and related cutting-edge technologies to provide readers with the latest knowledge and help improve their diagnostic and therapeutic skills; (7) Review: To make a systematic review to focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions; (8) Topic Highlight: The editorial board members are invited to write a series of articles (7-10 articles) to comment and discuss a hot topic to help improve the diagnostic and therapeutic skills of readers; (9) Medical Ethics: The editorial board members are invited to write articles about medical ethics to increase readers' knowledge of medical ethics. The topic covers international ethics guidelines, animal studies, clinical trials, organ transplantation, *etc.*; (10) Clinical Case Conference or Clinicopathological Conference: The editorial board members are invited to contribute high-quality clinical case conference; (11) Original Articles: To report innovative and original findings in pharmacology; (12) Research Report: To briefly report the novel and innovative findings in pharmacology; (13) Meta-Analysis: Covers the systematic review, mixed treatment comparison, meta-regression, and overview of reviews, in order to summarize a given quantitative effect, *e.g.*, the clinical effectiveness and safety of clinical treatments by combining data from two or more randomized controlled trials, thereby providing more precise and externally valid estimates than those which would stem from each individual dataset if analyzed separately from the others; (14) Case Report: To report a rare or typical case; (15) Letters to the Editor: To discuss and make reply to the contributions published in *WJP*, or to introduce and comment on a controversial issue of general interest; (16) Book Reviews: To introduce and comment on quality monographs of pharmacology; and (17) Autobiography: The editorial board members are invited to write their autobiography to provide readers with stories of success or failure in their scientific research career. The topic covers their basic personal information and information about when they started doing research work, where and how they did research work, what they have achieved, and their lessons from success or failure.

Name of journal

World Journal of Pharmacology

ISSN

ISSN 2220-3192 (online)

Launch date

February 9, 2012

Frequency

Quarterly

Editor-in-Chief

Geoffrey Burnstock, PhD, DSc, FAA, FRCS (Hon), FRCP

Instructions to authors

(Hon), FmedSci, FRS, Professor, Autonomic Neuroscience Centre, University College Medical School, Royal Free Campus, Rowland Hill Street, London NW3 2PF, United Kingdom

Editorial office

Jin-Lei Wang, Director
Xiu-Xia Song, Vice Director
World Journal of Pharmacology
Room 903, Building D, Ocean International Center,
No. 62 Dongsihuan Zhonglu, Chaoyang District,
Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: editorialoffice@wjnet.com
Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>
<http://www.wjnet.com>

Publisher

Baishideng Publishing Group Inc
8226 Regency Drive,
Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjnet.com
Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>
<http://www.wjnet.com>

Instructions to authors

Full instructions are available online at http://www.wjnet.com/2220-3192/g_info_20100722180909.htm.

Indexed and Abstracted in

Digital Object Identifier.

SPECIAL STATEMENT

All articles published in journals owned by the BPG represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Redit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word “significantly” should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJP* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read “Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest” from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serv-

ing as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of BPG, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at <http://www.wjgnet.com/esps/>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/2220-3192/g_info_20100722180909.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjpharmaco@wjgnet.com, or by telephone: +86-10-85381892. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, *e.g.*, Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, *e.g.*, Telephone: +86-10-85381892 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision on

acceptance is made only when at least two experts recommend publication of an article. All peer-reviewers are acknowledged on Express Submission and Peer-review System website.

Abstract

There are unstructured abstracts (no less than 200 words) and structured abstracts. The specific requirements for structured abstracts are as follows:

An informative, structured abstract should accompany each manuscript. Abstracts of original contributions should be structured into the following sections: AIM (no more than 20 words; Only the purpose of the study should be included. Please write the Aim in the form of "To investigate/study/..."), METHODS (no less than 140 words for Original Articles; and no less than 80 words for Brief Articles), RESULTS (no less than 150 words for Original Articles and no less than 120 words for Brief Articles; You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, *e.g.*, 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$), and CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Core tip

Please write a summary of less than 100 words to outline the most innovative and important arguments and core contents in your paper to attract readers.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both.

Illustrations

Figures should be numbered as 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ...*etc.* It is our principle to publish high resolution-figures for the E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a*P* < 0.05, ^b*P* < 0.01 should be noted (*P* > 0.05 should not be noted). If there are other series of *P* values, ^c*P* < 0.05 and ^d*P* < 0.01 are used. A third series of *P* values can be expressed as ^e*P* < 0.05 and ^f*P* < 0.01. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, *etc.*, in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^[1,2]". If references are cited directly in the text, they should be put together within the text, for example, "From references^[19,22-24], we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g., PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaohua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMCID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS-A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as ν (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, m (B) = 78 kg; blood pres-

sure, p (B) = 16.2/12.3 kPa; incubation time, t (incubation) = 96 h, blood glucose concentration, c (glucose) 6.4 ± 2.1 mmol/L; blood CEA mass concentration, p (CEA) = 8.6 24.5 $\mu\text{g/L}$; CO_2 volume fraction, 50 mL/L CO_2 , not 5% CO_2 ; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantums can be found at: http://www.wjgnet.com/2220-3192/g_info_20100725073806.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: t time or temperature, c concentration, A area, l length, m mass, V volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kbo I*, *Kpn I*, *etc.*

Biology: *H. pylori*, *E coli*, *etc.*

Examples for paper writing

All types of articles' writing style and requirement will be found in the link: <http://www.wjgnet.com/esps/NavigationInfo.aspx?pid=15>

RESUBMISSION OF THE REVISED MANUSCRIPTS

Authors must revise their manuscript carefully according to the revision policies of BPG. The revised version, along with the signed copyright transfer agreement, responses to the reviewers, and English language Grade A certificate (for non-native speakers of English), should be submitted to the online system *via* the link contained in the e-mail sent by the editor. If you have any questions about the revision, please send e-mail to esps@wjgnet.com.

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2220-3192/g_info_20100725073726.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2220-3192/g_info_20100725073445.htm.

Proof of financial support

For papers supported by a foundation, authors should provide a copy of the approval document and serial number of the foundation.

STATEMENT ABOUT ANONYMOUS PUBLICATION OF THE PEER REVIEWERS' COMMENTS

In order to increase the quality of peer review, push authors to carefully revise their manuscripts based on the peer reviewers' comments, and promote academic interactions among peer reviewers, authors and readers, we decide to anonymously publish the reviewers' comments and author's responses at the same time the manuscript is published online.

PUBLICATION FEE

WJJP is an international, peer-reviewed, OA online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium and format, provided the original work is properly cited. The use is non-commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 698 USD per article. All invited articles are published free of charge.

World Journal of *Pharmacology*

World J Pharmacol 2014 June 9; 3(2): 18-32





REVIEW

- 18 Arsenic exposure decreases rhythmic contractions of vascular tone through sodium transporters and K⁺ channels
Palacios J, Nwokocha CR, Cifuentes F

MINIREVIEWS

- 24 Pharmacological management of neuropathic pain in patients with vestibular schwannomas: Experience of the Atlantic Lateral Skull Base Clinic
Hebb ALO, Sawynok J, Bance M, Walling S, Chisholm K, Morris DP

APPENDIX I-V Instructions to authors

ABOUT COVER Editorial Board Number of *World Journal of Pharmacology*, Hugo Ruben Arias, PhD, Professor of Pharmacology and Biochemistry, Department of Medical Education, College of Medicine, California Northstate University, 9700 W. Taron Dr., Elk Grove, CA 95757, United States

AIM AND SCOPE *World Journal of Pharmacology* (*World J Pharmacol*, *WJP*, online ISSN 2220-3192, DOI: 10.5497) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJP covers topics concerning neuropsychiatric pharmacology, cerebrovascular pharmacology, geriatric pharmacology, anti-inflammatory and immunological pharmacology, antitumor pharmacology, anti-infective pharmacology, metabolic pharmacology, gastrointestinal and hepatic pharmacology, respiratory pharmacology, blood pharmacology, urinary and reproductive pharmacology, pharmacokinetics and pharmacodynamics, clinical pharmacology, and drug toxicology.

We encourage authors to submit their manuscripts to *WJP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING *World Journal of Pharmacology* is now indexed in Digital Object Identifier.

FLYLEAF I-IV Editorial Board

EDITORS FOR THIS ISSUE Responsible Assistant Editor: *Xiang Li* Responsible Science Editor: *Fang-Fang Ji*
 Responsible Electronic Editor: *Su-Qing Liu* Proofing Editorial Office Director: *Xiu-Xia Song*
 Proofing Editor-in-Chief: *Lian-Sheng Ma*

NAME OF JOURNAL
World Journal of Pharmacology

ISSN
 ISSN 2220-3192 (online)

LAUNCH DATE
 February 9, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Geoffrey Burnstock, PhD, DSc, FAA, FRCS (Hon), FRCP (Hon), FmedSci, FRS, Professor, Autonomic Neuroscience Centre, University College Medical School, Royal Free Campus, Rowland Hill Street, London NW3 2PF, United Kingdom

EDITORIAL OFFICE
 Jin-Lei Wang, Director
 Xiu-Xia Song, Vice Director

World Journal of Pharmacology
 Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
 Telephone: +86-10-85381891
 Fax: +86-10-85381893
 E-mail: editorialoffice@wjnet.com
 Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>
<http://www.wjnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 8226 Regency Drive,
 Pleasanton, CA 94588, USA
 Telephone: +1-925-223-8242
 Fax: +1-925-223-8243
 E-mail: bpgoffice@wjnet.com
 Help Desk: <http://www.wjnet.com/esps/helpdesk.aspx>
<http://www.wjnet.com>

PUBLICATION DATE
 June 9, 2014

COPYRIGHT
 © 2014 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjnet.com/2220-3192/g_info_20100722180909.htm

ONLINE SUBMISSION
<http://www.wjnet.com/esps/>

Arsenic exposure decreases rhythmic contractions of vascular tone through sodium transporters and K⁺ channels

Javier Palacios, Chukwuemeka R Nwokocha, Fredi Cifuentes

Javier Palacios, Departamento de Ciencias Químicas y Farmacéuticas, Universidad Arturo Prat, Iquique 1110939, Chile

Chukwuemeka R Nwokocha, Department of Basic Medical Science, Physiology Section, University of The West Indies, Mona Kingston JMAKN, Jamaica

Fredi Cifuentes, Experimental Physiology Laboratory, Instituto Antofagasta, Universidad de Antofagasta, Antofagasta 02800, Chile

Author contributions: Palacios J wrote the paper; Nwokocha CR and Cifuentes F revised the paper.

Correspondence to: Javier Palacios, PhD, Departamento de Ciencias Químicas y Farmacéuticas, Universidad Arturo Prat, Av. Arturo Prat Chacón, 2120, Iquique 1110939, Chile. palacios.unap@gmail.com

Telephone: +56-57-2526910 Fax: +56-57-2526210

Received: April 5, 2014 Revised: June 4, 2014

Accepted: June 5, 2014

Published online: June 9, 2014

blood flow. Since vascular rhythmic contractions of blood vessels are involved in modulating the vascular resistance, the blood flow, and the systemic pressure, we suggest a model explaining the participation of the sodium pump and NKCC1 co-transporter in low dose arsenic exposure effects on vasomotion and vascular dysfunction.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Arsenic; Vasomotion; Na⁺/K⁺-ATPase; Na⁺-K⁺-2Cl⁻; K⁺ channels; Nitric oxide; Prostaglandin; Vascular

Core tip: Vascular tone is regulated in part by cytosolic calcium oscillations. Arsenic can induce an increase in vascular tone and resistance. We suggest a model explaining the participation of the sodium pump and Na⁺-K⁺-2Cl⁻ co-transporter in low dose arsenic exposure effects on vasomotion and vascular dysfunction.

Abstract

Arsenic-contaminated drinking water is a public health problem in countries such as Taiwan, Bangladesh, United States, Mexico, Argentina, and Chile. The chronic ingestion of arsenic-contaminated drinking water increases the risk for ischemic heart disease, cerebrovascular disease, and prevalence of hypertension. Although toxic arsenic effects are controversial, there is evidence that a high concentration of arsenic may induce hypertension through increase in vascular tone and resistance. Vascular tone is regulated by the rhythmic contractions of the blood vessels, generated by calcium oscillations in the cytosol of vascular smooth muscle cells. To regulate the cytosolic calcium oscillations, the membrane oscillator model involves the participation of Ca²⁺ channels, calcium-activated K⁺ channels, Na⁺/Ca²⁺ exchange, plasma membrane Ca²⁺-ATPase, and the Na⁺/K⁺-ATPase. However, little is known about the role of K⁺ uptake by sodium transporters [Na⁺/K⁺-ATPase or Na⁺-K⁺-2Cl⁻ (NKCC1)] on the rhythmic contractions. Vascular rhythmic contractions, or vasomotion are a local mechanism to regulate vascular resistance and

Palacios J, Nwokocha CR, Cifuentes F. Arsenic exposure decreases rhythmic contractions of vascular tone through sodium transporters and K⁺ channels. *World J Pharmacol* 2014; 3(2): 18-23 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i2/18.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i2.18>

INTRODUCTION

Arsenic toxicity is a global environmental health problem. The toxicity of this metalloid has been observed in various countries, including Taiwan^[1], Bangladesh^[2], Mexico^[3], United States^[4], Hungary^[5], Argentina^[6], and Chile^[7]. Volcanic emission is one of the natural sources of arsenic, and individuals are majorly exposed through contaminated drinking water^[8]. Smelting companies are also an important source of individual and population exposure to these kinds of heavy metals contamination. Contamination has been reported in Russia^[9], United States^[10], Mexico^[11], Peru^[12], and Chile^[13]. There are few

studies showing that Chinese workers in copper smelter, steel or iron have high levels of total arsenic in urine (50 g/g creatinine). These studies include those reported for Fushun city^[14], Yunnan province^[15], and Fuxin city^[16].

CHRONIC ARSENIC EXPOSURE AND VASCULAR DISEASES

There are epidemiologic studies that showed an association between chronic arsenic exposure and vascular diseases^[17,18]. In fact, the ingestion of the arsenic-contaminated drinking water produced an increased risk for ischemic heart disease, cerebrovascular disease, and peripheral vascular resistance^[19]. Other studies report positive associations between chronic arsenic exposure in drinking water, and the prevalence of hypertension^[20-24].

Currently, arsenic effects on systemic blood pressure are controversial^[25,26]. However, there is ample evidence that arsenic exposure mainly increases the vascular peripheral resistance^[19,27], which defines the difficulty to blood flow through the blood vessels, particularly the small arteries.

Vascular rhythmic contractions, or vasomotion, are local mechanisms that regulate the vascular resistance and blood flow^[28-30]. For instance, an increase in the amplitude of the rhythmic contractions cause an increased blood flow because the vascular resistance is reduced^[31]. Since vascular rhythmic contractions of blood vessels are involved in modulating the vascular resistance, the blood flow, and the systemic pressure^[28,29], the effects of chronic low dose exposures to arsenic on vascular rhythmic contractions becomes of great interest.

VASCULAR RHYTHMIC CONTRACTIONS

Vascular rhythmic contractions may be considered as a compensatory mechanism to preserve the perfusion of tissues^[31], especially in patients with hypertension^[32,33] or ischemia^[34]. The mechanisms of the vascular rhythmic contractions may account for 3 states of contraction in blood vessels with different levels of calcium. These include small, medium, and tonic contraction, but only the medium concentrations produce rhythmic contractions^[35]. The changes of vascular tone are generated by calcium oscillations in the cytosol of vascular smooth muscle cells^[36]. To regulate the cytosolic calcium oscillations, the membrane oscillator model considers that activity of Ca^{2+} channels, calcium-activated K^+ channels, $\text{Na}^+/\text{Ca}^{2+}$ exchange, plasma membrane Ca^{2+} -ATPase, and the Na^+/K^+ -ATPase, voltage-dependent calcium channel, and transient receptor potential channel are essential for maintaining calcium oscillations^[37].

ROLE OF Na^+/K^+ -ATPASE AND $\text{Na}^+-\text{K}^+-2\text{Cl}^-$ COTRANSPORTER ON RHYTHMIC CONTRACTIONS

Little is known about the role of K^+ uptake through

Na^+/K^+ -ATPase and $\text{Na}^+-\text{K}^+-2\text{Cl}^-$ (NKCC1) on the rhythmic contractions. Na^+/K^+ -ATPase and NKCC1 cotransporter are responsible for the major K^+ uptake in vascular smooth muscle cells^[38-40]. Recent reports demonstrates that rhythmic contractions were associated with tonic and phasic responses, the tonic dependent on $[\text{Ca}^{2+}]_i$ and the phasic on potassium efflux (through K^+ channels) and potassium uptake^[41,42].

Na^+/K^+ -ATPase is responsible for the electrochemical gradient of sodium and potassium ions, it also plays a vital role in the regulations of ionic homeostasis in tissues and cells. In vascular smooth muscle cells, Na^+/K^+ -ATPase plays a major role in the regulation of vascular tone^[43,44], an increase in Na^+/K^+ -ATPase activity leads to hyperpolarization and relaxation of smooth muscle^[45], while its inhibition blunts rhythmic contractions in vascular smooth muscle cells^[46].

It was postulated that the inhibition of K_{ATP} channels reduces extracellular K^+ and Na^+/K^+ -ATPase activity, increases intracellular calcium concentration *via* $\text{Na}^+/\text{Ca}^{2+}$ exchanger, uncouples vascular smooth muscle cells *via* gap junctions, and eliminates vascular rhythmic contractions^[47,48]. Also, the inhibition of inward-rectifier K^+ channels (Kir) decrease Na^+/K^+ -ATPase activity in vascular smooth muscle cells^[49]. It is important to remember that the Na^+/K^+ -ATPase participates in relaxation of vascular smooth muscle cells through K^+ channels. For instance, Na^+/K^+ -ATPase is involved in K^+ -induced vasodilatation of hamster cremasteric arterioles^[50], and vasodilation in the human forearm^[51]. When K^+ (1 to 15 mmol/L) accumulates in the extracellular space, Na^+/K^+ -ATPase activity increases efflux of potassium through Kir. This leads to hyperpolarization and vasodilatation of the vascular smooth muscle cells^[49,52]. In contrast, the opening of calcium-activated K^+ channels inhibits the Na^+/K^+ -ATPase function^[53,54], and vascular rhythmic contractions^[28].

NKCC1 is an obligatory symport system with an apparent stoichiometry of 1:1:2 sodium, potassium and chloride ratios respectively. Although the co-transporter is bidirectional in resting vascular smooth muscle cells, the sum of the electrochemical gradients for the three transported ion species determines net influx^[55].

Evidence for the role of NKCC1 co-transporter on vascular rhythmic contractions is scanty, but it is worthy of note that the inward current of Cl^- decreases rhythmic contractions by increasing vasoconstriction^[47]. NKCC1 is responsible in part to keep intracellular Cl^- concentration above the electrochemical equilibrium^[56] as such helping to maintain the electrochemical gradient and cellular reactivity. Phenylephrine-induced stimulation of NKCC1 increases intracellular Cl^- concentration, depolarize vascular smooth muscle cells^[57], open L-type calcium channels^[58] and produce vasoconstriction. In the vascular oscillator model^[59], the release of intracellular Ca^{2+} from the reticulum stimulates the inward current of Cl^- *via* the calcium-activated Cl^- channel^[60] and cyclic guanosine monophosphate (cGMP)-activated Ca^{2+} -dependent Cl^- channels^[61]. This leads to membrane depolarization, opening

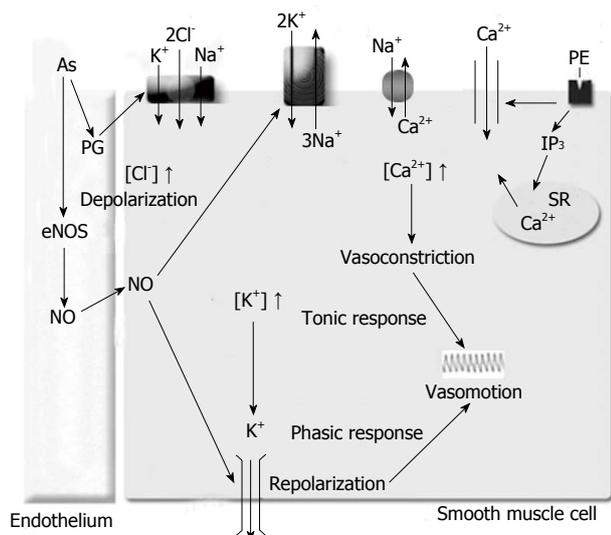


Figure 1 Putative model of arsenic effect on vasomotion phenomenon in blood vessels. The figure shows the stimulation of the Na^+/K^+ -ATPase by endothelial nitric oxide (NO) and stimulation of the Na^+/K^+ - 2Cl^- cotransporter by endothelial prostaglandins (PG). Arsenic would reduce NO bioavailability or would increase PG level, both of them would produce an increase in vasoconstriction or a decrease in the repolarization of the cell membrane, respectively, and then would reduce vasomotion. PE: Phenylephrine; As: Arsenic; eNOS: Endothelial nitric oxide synthase; SR: Sarcoplasmic reticulum.

L-type calcium channels and reduction in the oscillations of vascular tone. Therefore these findings suggest that the cotransporter NKCC1 would be responsible, in part, for vasoconstriction by chloride.

EFFECT OF ARSENIC ON VASCULAR RHYTHMIC CONTRACTIONS

Vascular rhythmic contractions are dependent in part on endothelial nitric oxide (NO)^[46], but there are few studies showing that the arsenic reduces vasomotion (vascular rhythmic contractions) by decreasing the NO bioavailability^[62].

It is well established that heavy metals such as arsenic induce increases in vascular resistance by inducing vascular endothelial dysfunction (VED)^[62,63]. VED consists of a reduction in endothelium-dependent vasorelaxation caused by a decrease in the release of endothelial NO^[64]. Arsenic-induced VED is caused in part by oxidative stress.

Oxidative stress from pollutants like arsenic causes an increase in the reactive oxygen species, this leads to a modification of amino acids of proteins, mainly sulfur-containing amino acids methionine and cysteine^[65]. Arsenic causes oxidative stress through peroxynitrite generation in aortic endothelial cells, producing loss of biological activity in enzymes and proteins^[66,67]. In this context we had shown that chronic arsenic exposure in drinking water reduced acetylcholine-induced relaxation in female rat aorta^[68], impairment of the endothelial nitric oxide synthase activity and decreasing of endothelial NO production^[69,70].

NO is reported to activates Na^+/K^+ -ATPase func-

tion^[71], we observed that acetylcholine and sodium nitroprusside (SNP) induces activation of Na^+/K^+ -ATPase activity, and SNP effect is abolished by inhibition of PKG (KT-5823)^[72]. Cogolludo *et al*^[73] (2001) showed that SNP activates Na^+/K^+ -ATPase in mesenteric piglet's arteries while Tamaoki *et al*^[74] (1997) found that cGMP activates Na^+/K^+ -ATPase in pulmonary artery smooth muscle cells.

Since arsenic decreases the NO bioavailability^[62], and the NO increases Na^+/K^+ -ATPase function^[71] which enhances the vascular rhythmic contractions, we may suggest that arsenic decreases the vascular rhythmic contractions by Na^+/K^+ -ATPase function (Figure 1). Similar conclusions would be expected with the Kir channel, as Chen *et al*^[75] (2010) demonstrated that arsenic trioxide produces down-regulation of Kir channel in cardiomyocytes of rats, and the Kir channel function increases Na^+/K^+ -ATPase activity^[49].

Although the endothelial NO does not affect NKCC1 co-transporter function^[76], the endothelial prostaglandins increase NKCC1 activity thereby enhancing the contractile response to agonist in rat aorta^[77-80]. Moreover, the endothelial prostaglandins increase agonist-induced rhythmic contractions in rat aorta^[81], rat mesenteric artery^[82], and arterioles of the cheek pouch of male hamsters^[42]. Furthermore, arsenic increases the cyclooxygenase-2 (COX-2) protein in aortic endothelial cells^[67], COX-2 in HUVEC^[83], and enhances COX-1 and COX-2 activities in hind paw muscle of male rats^[84]. Therefore, as a result of the prostaglandins effect on the vascular contractility through NKCC1 described above, arsenic might increase the vascular rhythmic contractions by NKCC1 co-transporter function.

The major toxic species of arsenic used in several studies are arsenite (trivalent inorganic arsenic, *i.e.*, arsenic trioxide) or arsenate (pentavalent inorganic arsenic). Although the concentration of arsenate in drinking water is higher than those of arsenite, toxic effects of arsenate have not been properly documented. Arsenate is mainly metabolized by organisms as monomethylarsonic acid and dimethylarsinic acid, which significantly are not toxic^[85]. However, this theory of the methylation of inorganic arsenic as a detoxification process has been revised^[86] as other trivalent methylated species with higher toxicity have been reported^[87]. Possibly, the biological effect of arsenate is mainly by reduction to arsenite^[88].

REFERENCES

- 1 **Guo HR**, Chiang HS, Hu H, Lipsitz SR, Monson RR. Arsenic in drinking water and incidence of urinary cancers. *Epidemiology* 1997; **8**: 545-550 [PMID: 9270957 DOI: 10.1097/00001648-199709000-00012]
- 2 **Smith AH**, Lingas EO, Rahman M. Contamination of drinking-water by arsenic in Bangladesh: a public health emergency. *Bull World Health Organ* 2000; **78**: 1093-1103 [PMID: 11019458]
- 3 **Del Razo LM**, Arellano MA, Cebrián ME. The oxidation states of arsenic in well-water from a chronic arsenicism area

- of northern Mexico. *Environ Pollut* 1990; **64**: 143-153 [PMID: 15092299 DOI: 10.1016/0269-7491(90)90111-O]
- 4 **Lewis DR**, Southwick JW, Ouellet-Hellstrom R, Rench J, Calderon RL. Drinking water arsenic in Utah: A cohort mortality study. *Environ Health Perspect* 1999; **107**: 359-365 [PMID: 10210691 DOI: 10.1289/ehp.99107359]
 - 5 **Börzsönyi M**, Bereczky A, Rudnai P, Csanady M, Horvath A. Epidemiological studies on human subjects exposed to arsenic in drinking water in southeast Hungary. *Arch Toxicol* 1992; **66**: 77-78 [PMID: 1580796 DOI: 10.1007/BF02307274]
 - 6 **Hopenhayn-Rich C**, Biggs ML, Fuchs A, Bergoglio R, Tello EE, Nicolli H, Smith AH. Bladder cancer mortality associated with arsenic in drinking water in Argentina. *Epidemiology* 1996; **7**: 117-124 [PMID: 8834549 DOI: 10.1097/00001648-199603000-00003]
 - 7 **Borgoño JM**, Vicent P, Venturino H, Infante A. Arsenic in the drinking water of the city of Antofagasta: epidemiological and clinical study before and after the installation of a treatment plant. *Environ Health Perspect* 1977; **19**: 103-105 [PMID: 908283 DOI: 10.2307/3428458]
 - 8 **Nordstrom DK**. Public health. Worldwide occurrences of arsenic in ground water. *Science* 2002; **296**: 2143-2145 [PMID: 12077387 DOI: 10.1126/science.1072375]
 - 9 **Bustueva KA**, Revich BA, Bezpalko LE. Cadmium in the environment of three Russian cities and in human hair and urine. *Arch Environ Health* 1994; **49**: 284-288 [PMID: 8031186 DOI: 10.1080/00039896.1994.9937481]
 - 10 **Hwang YH**, Bornschein RL, Grote J, Menrath W, Roda S. Environmental arsenic exposure of children around a former copper smelter site. *Environ Res* 1997; **72**: 72-81 [PMID: 9012374 DOI: 10.1006/enrs.1996.3691]
 - 11 **Diaz-Barriga F**, Santos MA, Mejía JJ, Batres L, Yáñez L, Carrizales L, Vera E, del Razo LM, Cebrián ME. Arsenic and cadmium exposure in children living near a smelter complex in San Luis Potosí, Mexico. *Environ Res* 1993; **62**: 242-250 [PMID: 8344231 DOI: 10.1006/enrs.1993.1109]
 - 12 **Ramírez AV**. [Environmental pollution by cadmium in a metallurgy plant]. *Bol Oficina Sanit Panam* 1986; **101**: 514-521 [PMID: 2947598]
 - 13 **Rivara MI**, Cebrián M, Corey G, Hernández M, Romieu I. Cancer risk in an arsenic-contaminated area of Chile. *Toxicol Ind Health* 1997; **13**: 321-338 [PMID: 9200798 DOI: 10.1177/074823379701300217]
 - 14 **Xi S**, Zheng Q, Zhang Q, Sun G. Metabolic profile and assessment of occupational arsenic exposure in copper- and steel-smelting workers in China. *Int Arch Occup Environ Health* 2011; **84**: 347-353 [PMID: 21132326 DOI: 10.1007/s00420-010-0574-7]
 - 15 **Wen J**, Wen W, Li L, Liu H. Methylation capacity of arsenic and skin lesions in smelter plant workers. *Environ Toxicol Pharmacol* 2012; **34**: 624-630 [PMID: 22885843 DOI: 10.1016/j.etap.2012.07.003]
 - 16 **Xu Y**, Wang Y, Zheng Q, Li B, Li X, Jin Y, Lv X, Qu G, Sun G. Clinical manifestations and arsenic methylation after a rare subacute arsenic poisoning accident. *Toxicol Sci* 2008; **103**: 278-284 [PMID: 18308700 DOI: 10.1093/toxsci/kfn041]
 - 17 **Chiou HY**, Huang WI, Su CL, Chang SF, Hsu YH, Chen CJ. Dose-response relationship between prevalence of cerebrovascular disease and ingested inorganic arsenic. *Stroke* 1997; **28**: 1717-1723 [PMID: 9303014 DOI: 10.1161/01.STR.28.9.1717]
 - 18 **Wang CH**, Hsiao CK, Chen CL, Hsu LI, Chiou HY, Chen SY, Hsueh YM, Wu MM, Chen CJ. A review of the epidemiologic literature on the role of environmental arsenic exposure and cardiovascular diseases. *Toxicol Appl Pharmacol* 2007; **222**: 315-326 [PMID: 17433393 DOI: 10.1016/j.taap.2006.12.022]
 - 19 **Chen WY**, Yen TS. Experimental studies on the drinking water of blackfoot endemic area. 2. studies on the effects of drinking water of blackfoot endemic area on peripheral vascular perfusion of the hind limbs of frogs. *Tsa Chih Gastroxiong Yi Xue Yuan Tong Xue Hui* 1964; **63**: 150-158 [PMID: 14199915]
 - 20 **Borgoño JM**, Greiber R. [Epidemiologic study of arsenic poisoning in the city of Antofagasta]. *Rev Med Chil* 1971; **99**: 702-707 [PMID: 5157219]
 - 21 **Rosenberg HG**. Systemic arterial disease and chronic arsenicism in infants. *Arch Pathol* 1974; **97**: 360-365 [PMID: 4825098]
 - 22 **Zaldívar R**. Arsenic contamination of drinking water and foodstuffs causing endemic chronic poisoning. *Beitr Pathol* 1974; **151**: 384-400 [PMID: 4838015 DOI: 10.1016/S0005-8165(74)80047-8]
 - 23 **Zierold KM**, Knobloch L, Anderson H. Prevalence of chronic diseases in adults exposed to arsenic-contaminated drinking water. *Am J Public Health* 2004; **94**: 1936-1937 [PMID: 15514231 DOI: 10.2105/AJPH.94.11.1936]
 - 24 **Chen Y**, Factor-Litvak P, Howe GR, Graziano JH, Brandt-Rauf P, Parvez F, van Geen A, Ahsan H. Arsenic exposure from drinking water, dietary intakes of B vitamins and folate, and risk of high blood pressure in Bangladesh: a population-based, cross-sectional study. *Am J Epidemiol* 2007; **165**: 541-552 [PMID: 17164464 DOI: 10.1093/aje/kwk037]
 - 25 **Abir T**, Rahman B, D'Este C, Farooq A, Milton AH. The Association between Chronic Arsenic Exposure and Hypertension: A Meta-Analysis. *J Toxicol* 2012; **2012**: 198793 [PMID: 22523484]
 - 26 **Islam MR**, Khan I, Attia J, Hassan SM, McEvoy M, D'Este C, Azim S, Akhter A, Akter S, Shahidullah SM, Milton AH. Association between hypertension and chronic arsenic exposure in drinking water: a cross-sectional study in Bangladesh. *Int J Environ Res Public Health* 2012; **9**: 4522-4536 [PMID: 23222207 DOI: 10.3390/ijerph9124522]
 - 27 **Lee MY**, Lee YH, Lim KM, Chung SM, Bae ON, Kim H, Lee CR, Park JD, Chung JH. Inorganic arsenite potentiates vasoconstriction through calcium sensitization in vascular smooth muscle. *Environ Health Perspect* 2005; **113**: 1330-1335 [PMID: 16203242 DOI: 10.1289/ehp.8000]
 - 28 **Nilsson H**, Aalkjaer C. Vasomotion: mechanisms and physiological importance. *Mol Interv* 2003; **3**: 79-89, 51 [PMID: 14993429]
 - 29 **Funk W**, Endrich B, Messmer K, Intaglietta M. Spontaneous arteriolar vasomotion as a determinant of peripheral vascular resistance. *Int J Microcirc Clin Exp* 1983; **2**: 11-25 [PMID: 6678836]
 - 30 **Gratton RJ**, Gandley RE, McCarthy JF, Michaluk WK, Slinker BK, McLaughlin MK. Contribution of vasomotion to vascular resistance: a comparison of arteries from virgin and pregnant rats. *J Appl Physiol* (1985) 1998; **85**: 2255-2260 [PMID: 9843550]
 - 31 **Pradhan RK**, Chakravarthy VS. Informational dynamics of vasomotion in microvascular networks: a review. *Acta Physiol (Oxf)* 2011; **201**: 193-218 [PMID: 20887358 DOI: 10.1111/j.1748-1716.2010.02198.x]
 - 32 **Hollenberg NK**, Sandor T. Vasomotion of renal blood flow in essential hypertension. Oscillations in xenon transit. *Hypertension* 1984; **6**: 579-585 [PMID: 6746087 DOI: 10.1161/01.HYP.6.4.579]
 - 33 **Frielingdorf J**, Kaufmann P, Seiler C, Vassalli G, Suter T, Hess OM. Abnormal coronary vasomotion in hypertension: role of coronary artery disease. *J Am Coll Cardiol* 1996; **28**: 935-941 [PMID: 8837571 DOI: 10.1016/S0735-1097(96)00260-4]
 - 34 **Intaglietta M**. Arteriolar vasomotion: implications for tissue ischemia. *Blood Vessels* 1991; **28** Suppl 1: 1-7 [PMID: 1932763]
 - 35 **Koenigsberger M**, Sauser R, Bény JL, Meister JJ. Role of the endothelium on arterial vasomotion. *Biophys J* 2005; **88**: 3845-3854 [PMID: 15792979 DOI: 10.1529/biophysj.104.054965]
 - 36 **Peng H**, Matchkov V, Ivarsen A, Aalkjaer C, Nilsson H. Hypothesis for the initiation of vasomotion. *Circ Res* 2001; **88**: 810-815 [PMID: 11325873 DOI: 10.1161/hh0801.089603]
 - 37 **Parthimos D**, Edwards DH, Griffith TM. Minimal model of arterial chaos generated by coupled intracellular and membrane Ca²⁺ oscillators. *Am J Physiol* 1999; **277**: H1119-H1144

- [PMID: 10484436]
- 38 **Garrahan PJ**, Glynn IM. The sensitivity of the sodium pump to external sodium. *J Physiol* 1967; **192**: 175-188 [PMID: 6051802]
- 39 **Sachs JR**. Ouabain-insensitive sodium movements in the human red blood cell. *J Gen Physiol* 1971; **57**: 259-282 [PMID: 5544793 DOI: 10.1085/jgp.57.3.259]
- 40 **Russell JM**. Sodium-potassium-chloride cotransport. *Physiol Rev* 2000; **80**: 211-276 [PMID: 10617769]
- 41 **Palacios J**, Vega JL, Paredes A, Cifuentes F. Effect of phenylephrine and endothelium on vasomotion in rat aorta involves potassium uptake. *J Physiol Sci* 2013; **63**: 103-111 [PMID: 23180009 DOI: 10.1007/s12576-012-0240-9]
- 42 **de Souza Md**, Bouskela E. Arteriolar diameter and spontaneous vasomotion: importance of potassium channels and nitric oxide. *Microvasc Res* 2013; **90**: 121-127 [PMID: 23948594 DOI: 10.1016/j.mvr.2013.08.001]
- 43 **Blaustein MP**. Sodium ions, calcium ions, blood pressure regulation, and hypertension: a reassessment and a hypothesis. *Am J Physiol* 1977; **232**: C165-C173 [PMID: 324293]
- 44 **Clausen T**, Nielsen OB. The Na⁺,K⁽⁺⁾-pump and muscle contractility. *Acta Physiol Scand* 1994; **152**: 365-373 [PMID: 7701937 DOI: 10.1111/j.1748-1716.1994.tb09818.x]
- 45 **Rapoport RM**, Schwartz K, Murad F. Effects of Na⁺,K⁺-pump inhibitors and membrane depolarizing agents on acetylcholine-induced endothelium-dependent relaxation and cyclic GMP accumulation in rat aorta. *Eur J Pharmacol* 1985; **110**: 203-209 [PMID: 2985409 DOI: 10.1016/0014-2999(85)90212-2]
- 46 **Gustafsson H**, Nilsson H. Rhythmic contractions in isolated small arteries of rat: role of K⁺ channels and the Na⁺,K⁽⁺⁾-pump. *Acta Physiol Scand* 1994; **150**: 161-170 [PMID: 8191895 DOI: 10.1111/j.1748-1716.1994.tb09673.x]
- 47 **Matchkov VV**, Gustafsson H, Rahman A, Briggs Boedtkjer DM, Gorintin S, Hansen AK, Bouzinova EV, Praetorius HA, Aalkjaer C, Nilsson H. Interaction between Na⁺/K⁺-pump and Na⁺/Ca²⁺-exchanger modulates intercellular communication. *Circ Res* 2007; **100**: 1026-1035 [PMID: 17347477 DOI: 10.1161/01.RES.0000262659.09293.56]
- 48 **Glavind-Kristensen M**, Matchkov V, Hansen VB, Forman A, Nilsson H, Aalkjaer C. KATP-channel-induced vasodilation is modulated by the Na₂S₂O₈-pump activity in rabbit coronary small arteries. *Br J Pharmacol* 2004; **143**: 872-880 [PMID: 15504751 DOI: 10.1038/sj.bjp.0706016]
- 49 **Haddy FJ**, Vanhoutte PM, Feletou M. Role of potassium in regulating blood flow and blood pressure. *Am J Physiol Regul Integr Comp Physiol* 2006; **290**: R546-R552 [PMID: 16467502 DOI: 10.1152/ajpregu.00491.2005]
- 50 **Burns WR**, Cohen KD, Jackson WF. K⁺-induced dilation of hamster cremasteric arterioles involves both the Na⁺/K⁺-ATPase and inward-rectifier K⁺ channels. *Microcirculation* 2004; **11**: 279-293 [PMID: 15280082 DOI: 10.1080/10739680490425985]
- 51 **Dawes M**, Sieniawska C, Delves T, Dwivedi R, Chowienczyk PJ, Ritter JM. Barium reduces resting blood flow and inhibits potassium-induced vasodilation in the human forearm. *Circulation* 2002; **105**: 1323-1328 [PMID: 11901043 DOI: 10.1161/hc1102.105651]
- 52 **Ulusoy HB**, Kaya MG. Potassium induced dilation in bovine coronary artery involves both inward rectifier potassium channels and Na⁺ /K⁺ ATPase. *Acta Physiol Hung* 2009; **96**: 427-436 [PMID: 19942549 DOI: 10.1556/APhysiol.96.2009.4.3]
- 53 **Dora KA**, Ings NT, Garland CJ. K(Ca) channel blockers reveal hyperpolarization and relaxation to K⁺ in rat isolated mesenteric artery. *Am J Physiol Heart Circ Physiol* 2002; **283**: H606-H614 [PMID: 12124208]
- 54 **Weston AH**, Richards GR, Burnham MP, Félétou M, Vanhoutte PM, Edwards G. K⁺-induced hyperpolarization in rat mesenteric artery: identification, localization and role of Na⁺/K⁺-ATPases. *Br J Pharmacol* 2002; **136**: 918-926 [PMID: 12110616 DOI: 10.1038/sj.bjp.0704787]
- 55 **O'Donnell ME**, Owen NE. Regulation of ion pumps and carriers in vascular smooth muscle. *Physiol Rev* 1994; **74**: 683-721 [PMID: 8036250]
- 56 **Chipperfield AR**, Harper AA. Chloride in smooth muscle. *Prog Biophys Mol Biol* 2000; **74**: 175-221 [PMID: 11226512 DOI: 10.1016/S0079-6107(00)00024-9]
- 57 **Davis JP**, Chipperfield AR, Harper AA. Accumulation of intracellular chloride by (Na-K-Cl) co-transport in rat arterial smooth muscle is enhanced in deoxycorticosterone acetate (DOCA)/salt hypertension. *J Mol Cell Cardiol* 1993; **25**: 233-237 [PMID: 8510166 DOI: 10.1006/jmcc.1993.1029]
- 58 **Anfinogenova YJ**, Baskakov MB, Kovalev IV, Kilin AA, Dulin NO, Orlov SN. Cell-volume-dependent vascular smooth muscle contraction: role of Na⁺, K⁺, 2Cl⁻ cotransport, intracellular Cl⁻ and L-type Ca²⁺ channels. *Pflugers Arch* 2004; **449**: 42-55 [PMID: 15293051 DOI: 10.1007/s00424-004-1316-z]
- 59 **Berridge MJ**. Smooth muscle cell calcium activation mechanisms. *J Physiol* 2008; **586**: 5047-5061 [PMID: 18787034 DOI: 10.1113/jphysiol.2008.160440]
- 60 **Haddock RE**, Hill CE. Differential activation of ion channels by inositol 1,4,5-trisphosphate (IP₃) and ryanodine-sensitive calcium stores in rat basilar artery vasomotion. *J Physiol* 2002; **545**: 615-627 [PMID: 12456838 DOI: 10.1113/jphysiol.2002.027904]
- 61 **Piper AS**, Large WA. Single cGMP-activated Ca⁽⁺⁾-dependent Cl⁽⁻⁾ channels in rat mesenteric artery smooth muscle cells. *J Physiol* 2004; **555**: 397-408 [PMID: 14724180 DOI: 10.1113/jphysiol.2003.057646]
- 62 **Lee MY**, Jung BI, Chung SM, Bae ON, Lee JY, Park JD, Yang JS, Lee H, Chung JH. Arsenic-induced dysfunction in relaxation of blood vessels. *Environ Health Perspect* 2003; **111**: 513-517 [PMID: 12676608 DOI: 10.1289/ehp.5916]
- 63 **Jindal S**, Singh M, Balakumar P. Effect of bis (maltolato) oxovanadium (BMOV) in uric acid and sodium arsenite-induced vascular endothelial dysfunction in rats. *Int J Cardiol* 2008; **128**: 383-391 [PMID: 17658639 DOI: 10.1016/j.ijcard.2007.05.031]
- 64 **Tsou TC**, Tsai FY, Hsieh YW, Li LA, Yeh SC, Chang LW. Arsenite induces endothelial cytotoxicity by down-regulation of vascular endothelial nitric oxide synthase. *Toxicol Appl Pharmacol* 2005; **208**: 277-284 [PMID: 16239170 DOI: 10.1016/j.taap.2005.03.001]
- 65 **Beckman JS**, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *Am J Physiol* 1996; **271**: C1424-C1437 [PMID: 8944624]
- 66 **Del Razo LM**, Quintanilla-Vega B, Brambila-Colombres E, Calderón-Aranda ES, Manno M, Alboreas A. Stress proteins induced by arsenic. *Toxicol Appl Pharmacol* 2001; **177**: 132-148 [PMID: 11740912 DOI: 10.1006/taap.2001.9291]
- 67 **Bunderson M**, Coffin JD, Beall HD. Arsenic induces peroxynitrite generation and cyclooxygenase-2 protein expression in aortic endothelial cells: possible role in atherosclerosis. *Toxicol Appl Pharmacol* 2002; **184**: 11-18 [PMID: 12392964 DOI: 10.1006/taap.2002.9492]
- 68 **Cifuentes F**, Bravo J, Norambuena M, Stegen S, Ayavire A, Palacios J. Chronic exposure to arsenic in tap water reduces acetylcholine-induced relaxation in the aorta and increases oxidative stress in female rats. *Int J Toxicol* 2009; **28**: 534-541 [PMID: 19966145 DOI: 10.1177/1091581809345924]
- 69 **Pi J**, Horiguchi S, Sun Y, Nikaido M, Shimojo N, Hayashi T, Yamauchi H, Itoh K, Yamamoto M, Sun G, Waalkes MP, Kumagai Y. A potential mechanism for the impairment of nitric oxide formation caused by prolonged oral exposure to arsenate in rabbits. *Free Radic Biol Med* 2003; **35**: 102-113 [PMID: 12826260 DOI: 10.1016/S0891-5849(03)00269-7]
- 70 **Kumagai Y**, Pi J. Molecular basis for arsenic-induced alteration in nitric oxide production and oxidative stress: implication of endothelial dysfunction. *Toxicol Appl Pharmacol* 2004; **198**: 450-457 [PMID: 15276426 DOI: 10.1016/j.taap.2003.10.031]
- 71 **Pavlovic D**, Hall AR, Kennington EJ, Aughton K, Bogu-

- slavskiy A, Fuller W, Despa S, Bers DM, Shattock MJ. Nitric oxide regulates cardiac intracellular Na⁺ and Ca²⁺ by modulating Na/K ATPase via PKC ϵ and phospholemman-dependent mechanism. *J Mol Cell Cardiol* 2013; **61**: 164-171 [PMID: 23612119 DOI: 10.1016/j.yjmcc.2013.04.013]
- 72 **Palacios J**, Marusic ET, Lopez NC, Gonzalez M, Michea L. Estradiol-induced expression of N(+)-K(+)-ATPase catalytic isoforms in rat arteries: gender differences in activity mediated by nitric oxide donors. *Am J Physiol Heart Circ Physiol* 2004; **286**: H1793-H1800 [PMID: 14704224 DOI: 10.1152/ajpheart.00990.2003]
- 73 **Cogolludo AL**, Pérez-Vizcaíno F, Zaragoza-Arnáez F, Ibarra M, López-López G, López-Miranda V, Tamargo J. Mechanisms involved in SNP-induced relaxation and [Ca²⁺]_i reduction in piglet pulmonary and systemic arteries. *Br J Pharmacol* 2001; **132**: 959-967 [PMID: 11181438 DOI: 10.1038/sj.bjp.0703894]
- 74 **Tamaoki J**, Tagaya E, Nishimura K, Isono K, Nagai A. Role of Na(+)-K+ ATPase in cyclic GMP-mediated relaxation of canine pulmonary artery smooth muscle cells. *Br J Pharmacol* 1997; **122**: 112-116 [PMID: 9298536 DOI: 10.1038/sj.bjp.0701351]
- 75 **Chen X**, Shan H, Zhao J, Hong Y, Bai Y, Sun I, Pan Z, Zhang Y, Yang B, Du Z. L-type calcium current (I_{Ca,L}) and inward rectifier potassium current (I_{K1}) are involved in QT prolongation induced by arsenic trioxide in rat. *Cell Physiol Biochem* 2010; **26**: 967-974 [PMID: 21220927 DOI: 10.1159/000324005]
- 76 **Koltsova SV**, Kotelevtsev SV, Tremblay J, Hamet P, Orlov SN. Excitation-contraction coupling in resistance mesenteric arteries: evidence for NKCC1-mediated pathway. *Biochem Biophys Res Commun* 2009; **379**: 1080-1083 [PMID: 19150334 DOI: 10.1016/j.bbrc.2009.01.018]
- 77 **Palacios J**, Espinoza F, Munita C, Cifuentes F, Michea L. Na⁺-K⁺-2Cl⁻ cotransporter is implicated in gender differences in the response of the rat aorta to phenylephrine. *Br J Pharmacol* 2006; **148**: 964-972 [PMID: 16799647 DOI: 10.1038/sj.bjp.0706818]
- 78 **Oppermann M**, Hansen PB, Castrop H, Schnermann J. Vasodilatation of afferent arterioles and paradoxical increase of renal vascular resistance by furosemide in mice. *Am J Physiol Renal Physiol* 2007; **293**: F279-F287 [PMID: 17494095 DOI: 10.1152/ajprenal.00073.2007]
- 79 **Mtabaji JP**, Manku MS, Horrobin DF. Vascular actions of furosemide and bumetanide on the rat superior mesenteric vascular bed: interactions with prolactin and prostaglandins. *Can J Physiol Pharmacol* 1976; **54**: 357-366 [PMID: 953864 DOI: 10.1139/y76-050]
- 80 **Pickkers P**, Dormans TP, Russel FG, Hughes AD, Thien T, Schaper N, Smits P. Direct vascular effects of furosemide in humans. *Circulation* 1997; **96**: 1847-1852 [PMID: 9323071 DOI: 10.1161/01.CIR.96.6.1847]
- 81 **Mauban JR**, Wier WG. Essential role of EDHF in the initiation and maintenance of adrenergic vasomotion in rat mesenteric arteries. *Am J Physiol Heart Circ Physiol* 2004; **287**: H608-H616 [PMID: 15059779 DOI: 10.1152/ajpheart.01084.2003]
- 82 **Okazaki K**, Seki S, Kanaya N, Hattori J, Tohse N, Namiki A. Role of endothelium-derived hyperpolarizing factor in phenylephrine-induced oscillatory vasomotion in rat small mesenteric artery. *Anesthesiology* 2003; **98**: 1164-1171 [PMID: 12717138 DOI: 10.1097/00000542-200305000-00019]
- 83 **Tsai SH**, Liang YC, Chen L, Ho FM, Hsieh MS, Lin JK. Arsenite stimulates cyclooxygenase-2 expression through activating I κ B kinase and nuclear factor κ B in primary and ECV304 endothelial cells. *J Cell Biochem* 2002; **84**: 750-758 [PMID: 11835400 DOI: 10.1002/jcb.10096]
- 84 **Ahmad W**, Prawez S, Chandrasekara HH, Tandan SK, Sankar P, Sarkar SN. Subacute arsenic exposure through drinking water reduces the pharmacodynamic effects of ketoprofen in male rats. *Environ Toxicol Pharmacol* 2012; **33**: 267-276 [PMID: 22236721 DOI: 10.1016/j.etap.2011.12.013]
- 85 **Vahter M**, Concha G. Role of metabolism in arsenic toxicity. *Pharmacol Toxicol* 2001; **89**: 1-5 [PMID: 11484904 DOI: 10.1034/j.1600-0773.2001.d01-128.x]
- 86 **Kitchin KT**. Recent advances in arsenic carcinogenesis: modes of action, animal model systems, and methylated arsenic metabolites. *Toxicol Appl Pharmacol* 2001; **172**: 249-261 [PMID: 11312654 DOI: 10.1006/taap.2001.9157]
- 87 **Dopp E**, von Recklinghausen U, Diaz-Bone R, Hirner AV, Rettenmeier AW. Cellular uptake, subcellular distribution and toxicity of arsenic compounds in methylating and non-methylating cells. *Environ Res* 2010; **110**: 435-442 [PMID: 19758587 DOI: 10.1016/j.envres.2009.08.012]
- 88 **Huang RN**, Lee TC. Cellular uptake of trivalent arsenite and pentavalent arsenate in KB cells cultured in phosphate-free medium. *Toxicol Appl Pharmacol* 1996; **136**: 243-249 [PMID: 8619232 DOI: 10.1006/taap.1996.0031]

P- Reviewer: Chen F, Sandow SL **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Liu SQ



Pharmacological management of neuropathic pain in patients with vestibular schwannomas: Experience of the Atlantic Lateral Skull Base Clinic

Andrea LO Hebb, Jana Sawynok, Manohar Bance, Simon Walling, Ken Chisholm, David P Morris

Andrea LO Hebb, Atlantic Lateral Skull Base Clinic, Division of Neurosurgery, Dalhousie University, QEII Halifax Infirmary Site, Halifax NS B3H 3A7, Canada

Andrea LO Hebb, Capital District Health Authority, QEII Halifax Infirmary Site, 3rd Floor, Division of Neurosurgery, Halifax NS B3H 3A7, Canada

Jana Sawynok, Department of Pharmacology, Dalhousie University, Nova Scotia B3H 4R2, Canada

Manohar Bance, David P Morris, Division Otolaryngology, Dalhousie University, QEII Health Sciences Centre, Halifax NS B3H 2Y9, Canada

Simon Walling, Division of Neurosurgery, Dalhousie University, Halifax Infirmary Site, QEII Health Sciences Centre, Halifax NS B3H 3A7, Canada

Ken Chisholm, Department of Anesthesia and Pain Management, Dalhousie University, Halifax NS B3H 2Y9, Canada

Author contributions: All authors had contributed substantially to conception and design, or acquisition of data, or analysis and interpretation of data; drafted the article or revised it critically for important intellectual content and gave final approval of the version to be published.

Correspondence to: Dr. Andrea LO Hebb, MSc, PhD, RN, Capital District Health Authority, QEII Halifax Infirmary Site, 3rd Floor, Division of Neurosurgery, 1796 Summer Street, Halifax NS B3H 3A7, Canada. andrea.hebb@cdha.nshealth.ca
Telephone: +1-902-473-4824 Fax: +1-902-425-4176

Received: April 29, 2014 Revised: May 31, 2014

Accepted: June 5, 2014

Published online: June 9, 2014

Abstract

Neuropathic pain is chronic pain generated by disorders of the peripheral and central nervous system, including skull base tumours. A skull base tumour can be any type of tumour that forms in the skull base, and this includes vestibular schwannomas which arise from the sheath of the inner ear vestibulocochlear nerve (eighth cranial nerve). Growth of the tumour, surgical resection, and/or stereotactic radiotherapy may result in

compression and/or irritation of the fifth cranial nerve (trigeminal nerve) resulting in facial pain and/or numbness. Non-trigeminal afferent input may contribute to the wide constellation of symptoms seen in orofacial pain patients. The purpose of this report was to develop a decision tool to guide the recognition and treatment of neuropathic pain in this specialized population. Recommendations for treatment are based on evidence presented in Canadian and international neuropathic treatment guidelines. Algorithms are included for assessment and treatment of adult patients with agents that are recognized to have analgesic efficacy within the broad context of neuropathic pain.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Acoustic neuroma; Stereotactic radiotherapy; Tricyclic antidepressants; Serotonin-norepinephrine reuptake inhibitors; Calcium channel modulators; Tramadol; Opioids

Core tip: The complexity of managing trigeminal neuralgia and neuropathic pain conditions among patients with skull base tumors requires a simple albeit comprehensive treatment algorithm that can be employed effectively by general practitioners, surgeons and other primary care prescribers in acute care or ambulatory clinical settings. We describe a simple treatment algorithm formulated on recommended best practice and based on clinical experience. It is intended to guide treatment, facilitate management and evaluation of outcome data (self-reported pain, quality of life measures) to elucidate the use of standardized approaches to pain management in patients with skull base etiology.

Hebb ALO, Sawynok J, Bance M, Walling S, Chisholm K, Morris DP. Pharmacological management of neuropathic pain in patients with vestibular schwannomas: Experience of the Atlantic

Lateral Skull Base Clinic. *World J Pharmacol* 2014; 3(2): 24-32
 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i2/24.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i2.24>

INTRODUCTION

This report considers pharmacological approaches to be used within the skull base tumour health community. Its intent is to offer assistance in treating patients who present with neuropathic pain (NP) in the context of orofacial, head and neck pain, including trigeminal neuralgia (TN), which is the most common craniofacial pain syndrome that is neuropathic in origin. The impetus for assembling the information stems from the experiences of practitioners within an interdisciplinary clinic who consistently rely on formularies and consultation with colleagues for advice in treating neuralgia and NP. The document offers suggested algorithms for assessment and treatment of patients with agents that are recognized to have analgesic efficacy within the broader context of NP. It makes the distinction between generalized NP^[1] and NP due to skull base tumours affecting the head, neck, face and glossopharyngeal region^[2,3], but also recognizes commonalities in mechanisms underlying various forms of NP^[1].

The Atlantic Lateral Skull Base Clinic

Vestibular schwannomas (VS) are slow growing, benign neoplasms that may be life threatening due to compression of central structures. Extra-axial tumours arising from the Schwann cell sheath of the vestibular or cochlear nerve (8th cranial nerve) are referred to as acoustic neuromas or, more appropriately, VS. Clinical diagnosis of a VS suggests an incidence of 0.7-1 per 100000 people, although incidental (*i.e.*, non-clinically relevant) discovery of VS suggests an incidence as high as 2 per 10000 people^[4]. In some cases, VS are associated with NP and TN, often secondary to tumour compression or stereotactic radiation treatment affecting cranial nerves within the cerebellopontine angle (CPA).

The Atlantic Lateral Skull Base Clinic provides coordinated care through Neurootology (Division of Otolaryngology), Neurosurgery and the Stereotactic Radiotherapy Group to patients with unilateral or bilateral VS, other CPA tumours, as well as lesions of the petrous apex and jugular foramen. This program is unique in Canada, offering a single centre, multidisciplinary approach for lateral skull base lesions. The Atlantic Lateral Skull Base Clinic serves a population of more than 2 million people in a catchment area that includes Newfoundland, Prince Edward Island, New Brunswick and Nova Scotia^[5]. Treatment options for skull base tumours include monitoring clinical progression, surgery, stereotactic radiation therapy (SRT), balance and hearing rehabilitation. An interdisciplinary clinic provides an ideal environment in which to identify and intervene in the treatment and management of NP affecting the head and neck region. The focus

of this work is on clinical experiences within our clinic. With low incidence, it will be very unlikely there will ever be randomized clinical trials that provide direct guidance for pain management in patients with VS. In that event, one must rely on extrapolation, and what we report is implementation of that approach and what it looks like in clinic.

Disease background

NP is a chronic pain state that is initiated by peripheral and central nervous system injury caused by trauma, inflammation, infection or metabolic disease, and includes conditions such as distal polyneuropathy due to diabetes [diabetic neuropathy (DN)] and post-herpetic neuralgia (PHN)^[1]. The Canadian Pain Society (CPS) estimates (based on 8.2% chronic NP prevalence in the general population) that 1 million Canadians live with NP^[6-8]. Neuropathic pain interferes with activities of daily living and work performance, impairs mood, decreases quality of living and generates three-fold increases in health care costs relative to matched controls^[9]. NP conditions involve spontaneous (paroxysmal or ongoing) and stimulus-evoked (*e.g.*, mechanical and thermal) symptoms; continuous or intermittent spontaneous pain is frequently described as burning, stabbing, shooting or shock-like; stimulus-evoked pain includes allodynia (pain in response to non-painful stimulation, extreme sensitivity to touch) and hyperalgesia (enhanced response to painful stimuli); NP can also involve tingling and numbness^[1,6,9].

TN involves irritation or compression of the 5th cranial nerve (trigeminal nerve), which evokes paroxysmal episodic stabbing pain of the facial area. Classically, pain is described as a sharp, shooting, electric shock-like, unilateral pain with acute onset and termination in distribution of the trigeminal nerve; this usually involves the V2 (maxillary) and V3 (mandibular) divisions but is rare in the V1 (ocular) division^[10]. TN has an incidence of 4-28 per 100000 person years^[11,12]. It can arise due to vascular alterations, non-vascular lesions, or tumours and other skull base abnormalities which exert pressure on the trigeminal nerve located in the CPA. Trigeminal NP is more continuous, and is characterized as burning, aching, throbbing^[10]. VS, or acoustic neuromas, are the most frequent CPA tumour to cause TN-like symptoms^[13]. Non-trigeminal nociceptive input, concurrent with induced masticatory responses (*i.e.*, hypoglossal, spinal accessory, facial, glossopharyngeal and vagal motor centers), may contribute to the wide constellation of symptoms seen in orofacial pain patients^[14]. The distinction between TN and trigeminal NP is important, as there are different treatment recommendations for each^[1,10].

Current algorithms

Treatment guidelines and decision rules improve patient outcomes. Recent literature providing strategies for the treatment of NP include the consensus statement and guidelines from the CPS^[6], as well as the Neuropathic Pain Special Interest Group of the International Associa-

tion for the Study of Pain^[15,16]. It needs to be emphasized that most recommendations have been derived from studies on PHN and DN. The European Federation of Neurological Societies (EFNS) Task Force included an approach to the management of NP pain associated with damage to the trigeminal nerve^[17].

There remains a paucity of information on how to assess, diagnose and treat pain in patients with VS or other skull base tumours, pain that appears to be of neuropathic origin and typically resides within the head, face, neck and glossopharyngeal area. In order to provide guidance relating to this specialized patient population, we constructed an algorithm for NP and craniofacial pain, including TN, based on Canadian contemporary standards of care and existing NP treatment algorithms. Recommendations consider pharmaceutical agents with evidence of efficacy in neuropathic pain, patient tolerability of the dose range expected to be needed, and actual therapeutic efficacy observed within our clinical practice. To date, there has not been a published tool that provides a clearly-defined algorithm for the assessment and treatment of NP in patients with skull base tumours. It is intended that this report provides guidance to primary care practitioners treating NP in patients with skull base tumours, using specific drugs or combinations of drugs, to improve outcomes in clinical practice with respect to patient self-reports of pain and quality of life.

CHARACTERIZATION OF PAIN ASSOCIATED WITH SKULL BASE TUMOURS

Skull base tumours involve the proliferation of abnormal cells in the part of the brain that meets the base of the skull. Symptoms of skull base tumours may include twitching, paralysis or facial pain. Craniofacial NP disorders include neuropathies, neuromas and neuralgias. Although significant interpractitioner and institutional variability exists, facial neuropathy and trigeminal nerve disturbances are relatively uncommon in comparison with unilateral hearing loss, tinnitus and vertigo or even idiopathic headache^[18,19]. Even if uncommon, when a patient presents with what appears to be NP, the origin of the pain and appropriate treatment actions may be more difficult to determine than the existence of pain. At the very least, the challenge is to identify where the pain is coming from and to distinguish it from idiopathic headaches. Headaches are present in 50%-60% of patients with unilateral VS at the time of diagnosis^[4,20]. Clinically, headaches that are unresponsive to over-the-counter analgesics may be a subtle cue that the pain originates from compression of the cranial nerves by the tumour^[21]. It has been reported that 3% to 45% of patients with CPA VS experience facial paresis, facial neuropathy and trigeminal nerve disturbances (hypoesthesia, paresthesia, and neuralgia) due to compression by the tumour on the ipsilateral^[13,22], and less commonly, on contralateral^[23] cra-

nial nerves; up to 93% are at risk secondary to irradiation treatment^[24-27].

The incidence of pain following SRT treatment is common. It has been estimated that 93% of lesions treated with SRT leave the patient at risk of radiation-induced TN^[26]. Other centers report that trigeminal symptoms occur in $\geq 3\%$ of patients whose tumours approach the level of the trigeminal nerve^[25]. It is also important to distinguish NP from pain associated with hydrocephalus which may occur following tumour irradiation^[19]. As such, patients presenting with pain should be referred for a magnetic resonance imaging (MRI) or computerized tomography scan. It is evident, given the diversity of pain mechanisms and individual patient responses that no single drug works for all NP states. In this respect, successful management of pain syndromes first necessitates accurate assessment.

PHARMACOLOGICAL MANAGEMENT OF NEUROPATHIC PAIN

Assessment of pain and associated symptoms is necessary for diagnosis and management of NP. The self-report version of the Leeds Assessment of Neuropathic Symptoms and Signs scale and the French Neuropathic Pain Group clinician-administered questionnaire called DN4 may be used, based on their high sensitivity and selectivity^[1,28,29]. The use of these tools is meant to complement but not replace clinical judgment.

Current treatment guidelines provide an evidence-based approach to the treatment of NP. Treatment guidelines have been developed based on data collected from randomized controlled clinical trials of anticonvulsants (carbamazepine, oxcarbazepine), tricyclic antidepressants (TCAs) (amitriptyline, nortriptyline), serotonin-norepinephrine reuptake inhibitors (SNRIs) (duloxetine, venlafaxine), calcium channel ligands (gabapentin, pregabalin), local anesthetics (5% lidocaine patch), opioids (morphine, methadone) and opioid-like hybrid drugs (tramadol)^[6,16,17]. It should be mentioned that if the 5% lidocaine patch is unavailable, a lidocaine gel formulation or compounded cream may be substituted, although limited efficacy in non-post-herpetic pain has been reported^[30]. A general overview of our suggested algorithm for the management of NP in our clinic is presented in Figures 1 and 2.

The algorithms presented below are based on published clinical guidelines to simplify the management and evaluation of NP in patients with lateral skull base tumours. Suggested first, second and third line agents [determined by efficacy, indicated by number-needed-to-treat (NNT), and patient tolerability as indicated by number-needed-to-harm (NNH), or side-effects] are listed in Figure 3. NNT and NNH vary according to the etiology of the pain and reference consulted^[31,32]. Algorithms for individual pharmacological agents include initial starting doses, titration doses, temporal intervals and maximal dosing schedules^[33]. In addition, TCAs, SNRIs, gabapentinoids, opioid analgesics and tramadol must all be used

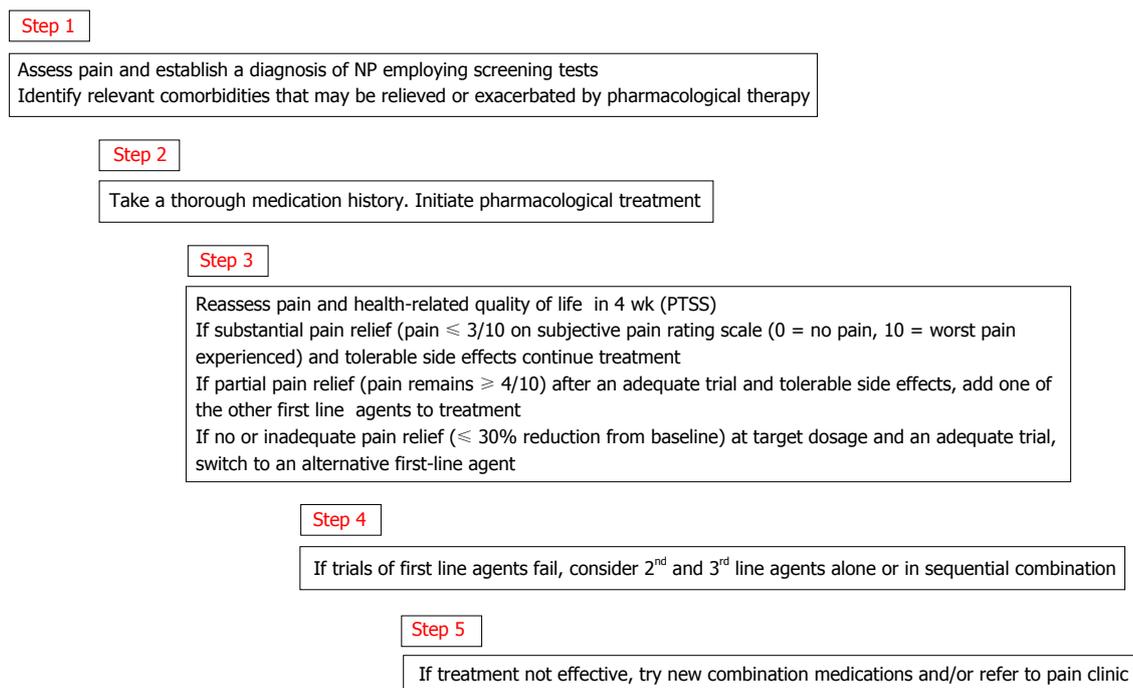


Figure 1 Assessment and management of neuropathic pain. NP: Neuropathic pain; PTSS: Pain treatment satisfaction scale.

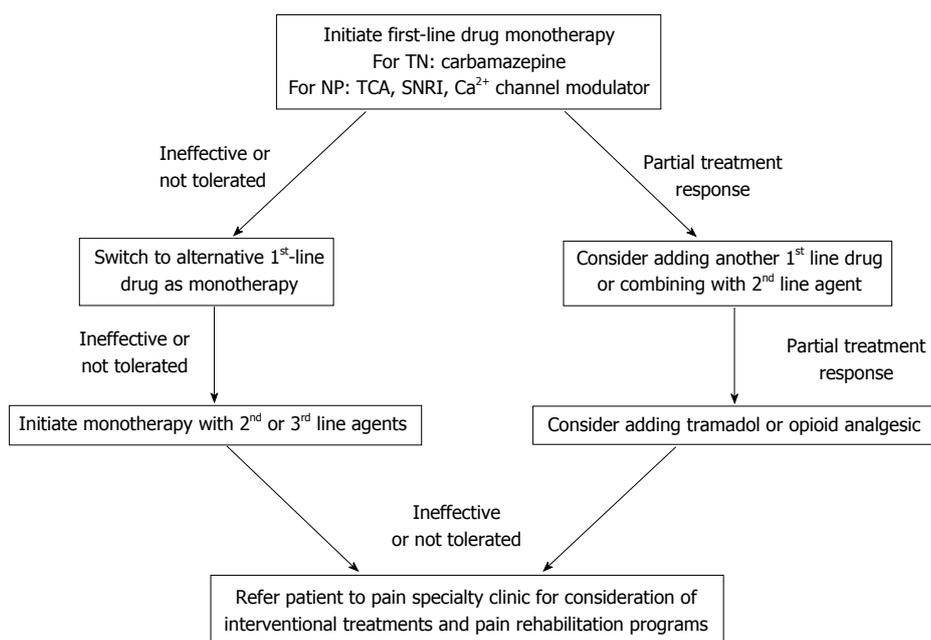


Figure 2 Management of neuropathic pain or trigeminal neuralgia in the Atlantic Lateral Skull Base Clinic. NP: Neuropathic pain; TN: Trigeminal neuralgia; SNRI: Serotonin-norepinephrine reuptake inhibitor.

with caution in elderly patients because of the risk of falls and cognitive impairment^[15,16]. Recommendations are also made regarding the duration of adequate trials at maximum tolerated dosages to evaluate the impact on self-reported pain.

Initial and subsequent agents

For management of classical TN (characterized by paroxysmal, unilateral pain), carbamazepine is the first choice, but otherwise it is not used; this agent has a NNT around

1.9 with virtually complete pain relief^[6,10,34-36] (Table 1). Oxcarbazepine can, and should, be substituted for carbamazepine if there is an unacceptable side effect profile^[10,34-36] (Table 1).

For management of NP, there are several TCAs available, but amitriptyline and nortriptyline are commonly used, and exhibit NNT values of 1.3-3.6^[32,37-39]; NNH values are 3-6 for minor, and 14-28 for major, harm. The analgesic properties of TCAs are independent of their antidepressant effects, and several mechanisms, in

Table 1 Summary of drugs used for management of trigeminal neuralgia and neuropathic pain

Drug	Action	Dosing	Common side effects ¹
CBZ	Blocks Na ⁺ and Ca ²⁺ channels	300-1000 mg; 100 mg BID initially, increase by 200 mg weekly Adequate trial 8-12 wk, 2 wk at maximal dose	Drowsiness, ataxia, headaches, nausea, vomiting, constipation, blurred vision, rash Drug interactions Taper doses when discontinuing
OXC	Keto derivative of CBZ, same actions	Equivalent efficacy to CBZ; 300-2400 mg; 300 mg BID initially, increase by 600 mg weekly Adequate trial 8-12 wk, 2 wk at maximal dose	Improved tolerability compared to CBZ Vertigo, fatigue, dizziness, nausea, hyponatraemia in high doses No major drug interactions Taper doses when discontinuing
TCAs or tricyclic antidepressants: nortriptyline, amitriptyline	Block NA and 5-HT reuptake, block Na ⁺ channels, interact with several neurotransmitter systems	Nortriptyline 10 mg (elderly) or 25 mg (adult) at bedtime; increase dose by 10 or 25 mg every 3-7 d; up to 75-100 mg daily Adequate trial 6-8 wk, 2 wk at maximal dose Amitriptyline doses similar	Nortriptyline is better tolerated than amitriptyline Dry mouth, constipation, blurred vision, sedation, orthostatic hypotension Taper doses when discontinuing
SNRIs or serotonin-noradrenaline reuptake inhibitors: Duloxetine, Venlafaxine	Similar actions to TCAs, but fewer interactions with receptor systems	Duloxetine: 60 mg/d, increase to 120 mg after 1 wk Venlafaxine: 75 mg/d, increase to 225 mg over 3 wk Adequate trial 4-6 wk, 2 wk at maximum dose	Headache, nausea, dry mouth, sleepiness, fatigue, constipation, dizziness, decreased appetite, and increased sweating. Taper doses when discontinuing Drowsiness, dizziness, weakness, feeling nervous, tinnitus, increased sweating, blurred vision, dry mouth, changes in appetite or weight, facial flushing, mild nausea, constipation, sexual side-effects Taper doses over 7-10 d when discontinuing
Gabapentin	Ca ²⁺ channel modulator	100-300 mg TID, increase dose every 1-7 d; maximum dose 3600 mg daily Adequate trial 3-8 wk titration, 2-8 wk at maximal dose	Dizziness, sedation, weight gain, weakness, tiredness, nausea, diarrhea, constipation, blurred vision, headache, breast swelling, dry mouth, fatigue, myalgia, loss of balance or coordination Taper doses when discontinuing
Pregabalin	Ca ²⁺ channel modulator	50 mg OD or 25 mg BID, double dose each week; maximum daily dose 600 mg Adequate trial is 4 wk at maximal dose	Dizziness, drowsiness, loss of balance or coordination, problems with memory or concentration, anxiety, depersonalization, hypertonia, hypesthesia, decreased libido, nystagmus, paresthesia, twitching, breast swelling, tremors, dry mouth, constipation Taper doses when discontinuing (minimum of one week)
Tramadol	Inhibits NA and 5-HT reuptake, binds opioid receptors	50 mg BID, increase by 50-100 mg daily in divided doses over 3-7 d as tolerated; 400 mg is maximum dose (300 mg in elderly) Adequate trial is 4 wk at maximum dose	Dizziness, spinning sensation, constipation, upset stomach, headache, drowsiness, feeling nervous or anxious
Morphine	Interacts with mu opioid receptors in spinal cord and brain, regulates synaptic activity in pain pathways	10-15 mg q4h or prn (equianalgesic doses for other opioids); after 1-2 wk, convert to long-acting opioid (e.g., CR hydromorphone) Titration allows for dose escalation Adequate trial is 4-6 wk	Sedation, pruritus, constipation, diarrhea, weight loss, nausea, vomiting, stomach pain, loss of appetite, flushing (warmth, redness, tingling), headache, dizziness, spinning sensation, memory problems, sleep problems (insomnia), strange dreams Taper doses when discontinuing

¹These are not a complete list of side effects and others may occur. CBZ: Carbamazepine; 5-HT: Serotonin; NA: Noradrenaline; OXC: Oxcarbazepine; q4h: Every four hours; BID: Twice a day; TCA: Tricyclic antidepressant; SNRI: Serotonin-norepinephrine reuptake inhibitors; prn: Pro re nata; TID: Three times a day; OD: Once daily; CR: Continuous release.

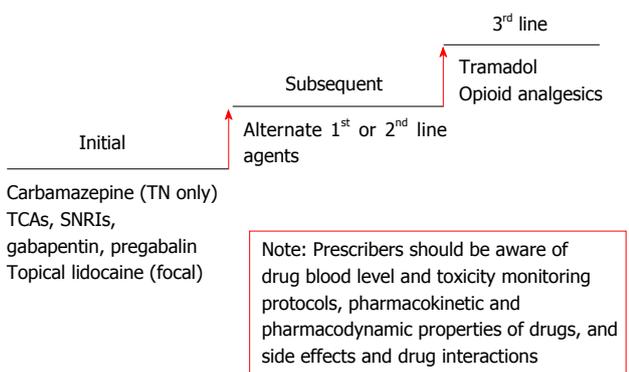


Figure 3 Treatment algorithm for trigeminal neuralgia or neuropathic craniofacial pain. TN: Trigeminal neuralgia; SNRIs: Serotonin-norepinephrine reuptake inhibitors; TCA: Tricyclic antidepressant.

addition to blockage of serotonin and norepinephrine reuptake, are involved in their actions^[40]. It should be emphasized that caution must be employed with the use of TCAs in older patients because of anticholinergic adverse effects, sedation, risk of falls, and risk of cardiac toxicity. Nortriptyline, a metabolite of amitriptyline with similar pharmacological effectiveness reflected in similar NNT values^[6,37,39], has a lower incidence of adverse effects compared to amitriptyline^[32,41]. The lowest effective dose of TCA should be used in NP patients, avoiding patients with ischemic heart disease or increased risk of cardiac death^[15] (Table 1). Where available, lidocaine medicated plaster (NNT = 4.4, NNH = 29), a topical formulation, can be considered first-line treatment if the pain is focal and there are tolerability issues for oral for-

mulations^[16,37].

The EFNS and Neuropathic Pain Special Interest Group (NeuPSIG) recommend the two SNRIs, duloxetine and venlafaxine, as first-line options, while Canadian guidelines consider these second-line options for treatment of NP. Duloxetine (NNT = 5-6; NNH = 7-9 minor, 13-15 major) has primarily been examined in DN, while venlafaxine (NNT = 3-5, NNH = 7-9 minor, 16 major) has been examined in a broader range of NP conditions^[31,37-39] (Table 1). Venlafaxine may be employed in combination with gabapentin to increase its efficacy^[16,30,42,43].

The Consensus Statement and Guidelines from the Canadian Pain Society list gabapentin and pregabalin as first-line agents in the treatment of NP^[6]. Most of the literature and guidelines for NP are based on PHN and DN and may not be applicable to all NP conditions^[9,15]. Gabapentin (NNT = 4.1-6.4, NNH = 3.7) and pregabalin (NNT = 3.3-6.0, NNH = 3.7-8.8) (Table 1)^[32,37,38] can be considered as first- or second-line agents for management of orofacial NP associated with skull base tumours. The EFNS Task Force has identified the usefulness of combination therapy, including TCA-gabapentin for the management of NP and TN^[17] (Figure 2).

Gabapentin and pregabalin decrease the release of several neurotransmitters involved in pain through binding to the $\alpha 2\text{-}\delta$ subunit of voltage-gated calcium channels, synaptic γ -aminobutyric acid modulation and synaptogenesis^[44]. Side effects of gabapentin include dry mouth, dizziness, gastrointestinal disturbances and cognitive impairment. There is some evidence that the efficacy of gabapentin is increased in painful DN, when combined with venlafaxine or morphine^[31,45]. Combination of gabapentin with nortriptyline was superior to either drug alone, and combination of gabapentin with opioids has shown increased efficacy for the treatment of TN and NP^[17] (Figure 2).

Third-line agents

Tramadol, a hybrid drug with SNRI and μ -opioid agonist properties, has a NNT = 3.4-4.9, and a NNH = 7.7; it should not be combined with TCAs due to increased risk of serotonin syndrome^[32,37] (Table 1). Opioids are generally safe if titrated slowly^[46]. While there is some evidence to support opioids in NP^[15,32,37,39], opioids as primary therapy are not always effective, and combination therapy may be needed^[31,42,44]. The analgesic efficacy of a combination of morphine and gabapentin was increased compared to each individual agent in patients with PHN and painful DN; however, while the maximal tolerated doses were lower in combination therapy, there was report of increased adverse effects^[16].

There are many liabilities and controversies surrounding the use of opioids in those with chronic non-cancer pain^[28,46]. For treating NP with opioids we follow recommendations of the 2010 National Opioid Use Guideline Group^[46]; referral to a chronic pain service is indicated when combination therapies involve primary and adjunct treatment beyond the common agents listed.

Other agents

Although beyond the scope of this document and better reserved for pain specialty clinics, interventional procedures, compounded drugs (such as carbamazepine, gabapentin, antidepressants, lidocaine) delivered as topical formulations for the orofacial region^[47], invasive techniques (may include intravenous lidocaine)^[6] or intradermal botulinum toxin^[16] may be considered. Several recent case series reports and a controlled study support the efficacy and safety of botulinum toxin for TN^[48-50]. Future treatment guidelines will position these options within current schemes.

CASE EXAMPLES

Two case examples of the presentation and treatment of neuropathic pain resulting from VS are outlined below.

Case 1

Presentation and diagnosis: RG, a 72-year-old retired fireman, presented to the Atlantic Lateral Skull Base Clinic in August 2010 with progressive right-sided hearing loss and tinnitus over 1-2 years with a new presentation of numbness and tingling on the right side of his face including the tip of his tongue. MRI revealed a right-sided cerebellar pontine angle tumour (25 mm \times 27 mm) in the axial plane with mass effect on the brainstem consistent with a vestibular schwannoma/acoustic neuroma. RG opted for stereotactic radiation therapy (SRT) over surgery or a more conservative "wait and scan" approach.

Symptoms prior to treatment: In December 2010, prior to SRT therapy, lidocaine hydrochloride swish and spit was ineffective in controlling symptoms. In January 2011, he reported exacerbation of symptoms on the right side of his face, dysesthesia rather than numbness, constant burning involving the tip of the tongue and the bottom of the tongue on the right hand side; symptoms were exacerbated by eating. He had lost 30 pounds due to decreased food intake. Pregabalin 75 mg twice a day and dexamethasone 4 mg every morning with breakfast was prescribed. Over the next month, pregabalin was increased to 150 mg twice a day as he noted some early improvement. The pregabalin dose was again increased to 300 mg and RG gained 3 pounds.

Treatment and follow up: SRT began in February 2011. RG noticed an immediate improvement in his tinnitus. In March 2011, gabapentin replaced pregabalin and was increased to 1800 mg/d while dexamethasone weaning began. RG experienced hypotension in response to the increase in gabapentin and the dose was decreased to 300 mg three times a day. SRT was completed in April 2011. At this time, RG reported increased facial pain, so gabapentin was increased to 1200 and then 1500 mg/d. RG was not reporting any relief from symptoms, and numbness and right-sided trigeminal neuralgia persisted. In October 2011, he was weaned off gabapentin, and amitriptyline was prescribed for facial pain 10 mg every night, and increased to 20 mg after 1 wk. In November

2011, he reported side effects which included sleepiness and nausea. His family doctor changed the prescription to nortriptyline 10 mg for 5 d, then 20 mg/d for 5 d, and then 30 mg/d, with a plan to increase this again in the next 5 d to 50 mg/d, his current dose in February 2012. He has improved facial pain and reports improvements in eating and tasting food. He still has some paresthesia and dysesthesia in the right side of his face in the trigeminal distribution in all 3 branches. His vestibular schwannoma has shrunk by 0.5 cm.

Case 2

Presentation and diagnosis: DB aged 71, presented to the Atlantic Lateral Skull Base Clinic in March 2008 reporting a recent history of loud noises prompting headaches. She subsequently had an audiogram which showed slight decrease in useful hearing on the right side compared to the left. An MRI (November 2007) revealed a small right-sided acoustic neuroma (18 mm × 9 mm in the axial plane). General ears, nose and throat exam was normal, other than the slight decrease in useful hearing; no tinnitus or vertigo was reported. The Unterberger stepping test revealed a moderate pulling to the right; cerebellar and oculomotor function was normal. Her facial nerve was completely normal with no facial weakness, facial twitching or facial numbness. She remained stable for a year.

Symptoms prior to treatment: In July 2009, her hearing deteriorated, to 60% of normal, and she was experiencing disequilibrium. MRI revealed slight growth in the neuroma (20 mm in longest dimension, 13 mm perpendicular to the petrous apex). A subsequent MRI in November 2009 showed further growth (20 mm × 15 mm).

Treatment and follow up: DB was referred for SRT, and this was completed June 2010. MRI in January 2011 revealed central cystic changes in tumour composition consistent with SRT. DB reported no new clinical symptoms other than further decrease in hearing on the right side.

In March 2011, new neurological symptoms emerged. Initially she described the right side of her tongue as heavy. She began to have some right-sided painful secondary paresthesia on her face in cheek area, upper and lower lips and tongue [maxillary (V2) and mandibular (V3) divisions of the trigeminal nerve] that she describes as a persistent burning sensation that is made worse by eating. She also described discomfort in her right eye and right-sided facial twitching. MRI in March 2011 showed swelling of the tumour in the right CPA consistent with SRT-induced changes. She was started on gabapentin 300 mg once daily for one week, increasing the dose to twice daily the second week, and thrice daily for the third week if tolerated.

In November of 2011 she returned. Gabapentin 300 mg once daily reduced her symptoms; however, it was associated with intolerable constipation and myalgia in the upper arms. She was weaned off gabapentin and her constipation and myalgia dissipated. The right-sided acoustic neuroma had further decreased in size, approximately 18 mm × 12 mm (May 2013) compared to 22 mm × 16 mm

(April 2012), yet she continued to experience persistent chronic orofacial pain. While it is possible that DB would be refractory to other pharmacologic interventions, she has declined further interventions, despite the pain and effect on her quality of life.

CONCLUSION

The present report describes evidence for application of pain management strategies in patients with VS. A decision tool and treatment algorithm is presented to facilitate evaluation and management of patients with NP resulting from skull base disorders. A pharmacological algorithm, with primary and adjuvant treatment, summarizes pharmaceutical choice and management of NP and TN in patients with VS and can be made available in the clinic for quick review by the treating physicians. This instrument is intended to guide treatment of neuropathic/trigeminal pain in patients in acute care or ambulatory clinical settings. Essential to the comprehensive management of patients is evaluation of the effects of the intervention on quality-of-life and patient satisfaction with pain management. Patient satisfaction with pain management, using the Pain Treatment Satisfaction Scale^[51], as well as the American Pain Society Satisfaction Survey, was influenced by effectiveness of medication on pain severity, independent of initial pain intensity, and by communication^[52]. Comparison of outcome data (self-reported pain and quality of life) will elucidate the use of standardized approaches to managing pain among patients with specific skull base etiology. The purpose of the present treatment algorithm was to develop a common scheme that may be utilized by beginning practitioners for treating this relatively uncommon, but clinically challenging, condition.

REFERENCES

- 1 **Baron R**, Binder A, Wasner G. Neuropathic pain: diagnosis, pathophysiological mechanisms, and treatment. *Lancet Neurol* 2010; **9**: 807-819 [PMID: 20650402 DOI: 10.1016/S1474-4422(10)70143-5]
- 2 **Elias WJ**, Burchiel KJ. Trigeminal neuralgia and other neuropathic pain syndromes of the head and face. *Curr Pain Headache Rep* 2002; **6**: 115-124 [PMID: 11872182 DOI: 10.1007/s11916-002-0007-8]
- 3 **Zakrzewska JM**. Diagnosis and differential diagnosis of trigeminal neuralgia. *Clin J Pain* 2002; **18**: 14-21 [PMID: 11803298 DOI: 10.1097/00002508-200201000-00003]
- 4 **Kutz JW**. Acoustic neuroma. 2011. Available from: URL: <http://emedicine.medscape.com/article/882876-overview>
- 5 **Statistics Canada**. Canada's population estimates. 2011. Available from: URL: <http://www.statcan.gc.ca/daily-quotidien/100929/t100929b3-eng.htm>
- 6 **Moulin DE**, Clark AJ, Gilron I, Ware MA, Watson CP, Sessle BJ, Coderre T, Morley-Forster PK, Stinson J, Boulanger A, Peng P, Finley GA, Taenzer P, Squire P, Dion D, Chokan A, Gilani A, Gordon A, Henry J, Jovey R, Lynch M, Mailis-Gagnon A, Panju A, Rollman GB, Velly A. Pharmacological management of chronic neuropathic pain-consensus statement and guidelines from the Canadian Pain Society. *Pain Res Manag* 2007; **12**: 13-21 [PMID: 17372630]
- 7 **Torrance N**, Smith BH, Bennett MI, Lee AJ. The epidemiol-

- ogy of chronic pain of predominantly neuropathic origin. Results from a general population survey. *J Pain* 2006; **7**: 281-289 [PMID: 16618472 DOI: 10.1016/j.jpain.2005.11.008]
- 8 **Smith BH**, Torrance N. Epidemiology of neuropathic pain and its impact on quality of life. *Curr Pain Headache Rep* 2012; **16**: 191-198 [PMID: 22395856 DOI: 10.1007/s11916-012-0256-0]
 - 9 **Gilron I**, Watson CP, Cahill CM, Moulin DE. Neuropathic pain: a practical guide for the clinician. *CMAJ* 2006; **175**: 265-275 [PMID: 16880448 DOI: 10.1503/cmaj.060146]
 - 10 **Zakrzewska JM**. Medical management of trigeminal neuropathic pains. *Expert Opin Pharmacother* 2010; **11**: 1239-1254 [PMID: 20426709 DOI: 10.1517/14656561003767449]
 - 11 **Koopman JS**, Deilman JP, Huygen FJ, de Mos M, Martin CG, Sturkenboom MC. Incidence of facial pain in the general population. *Pain* 2009; **147**: 122-127 [PMID: 19783099 DOI: 10.1016/j.pain.2009.08.023]
 - 12 **van Hecke O**, Austin SK, Khan RA, Smith BH, Torrance N. Neuropathic pain in the general population: a systematic review of epidemiological studies. *Pain* 2014; **155**: 654-662 [PMID: 24291734 DOI: 10.1016/j.pain.2013.11.013]
 - 13 **Matsuka Y**, Fort ET, Merrill RL. Trigeminal neuralgia due to an acoustic neuroma in the cerebellopontine angle. *J Orofac Pain* 2000; **14**: 147-151 [PMID: 11203749]
 - 14 **Türp JC**, Kowalski CJ, O'Leary N, Stohler CS. Pain maps from facial pain patients indicate a broad pain geography. *J Dent Res* 1998; **77**: 1465-1472 [PMID: 9649175 DOI: 10.1177/00220345980770061101]
 - 15 **Dworkin RH**, O'Connor AB, Backonja M, Farrar JT, Finnerup NB, Jensen TS, Kalso EA, Loeser JD, Miaskowski C, Nurmikko TJ, Portenoy RK, Rice AS, Stacey BR, Treede RD, Turk DC, Wallace MS. Pharmacologic management of neuropathic pain: evidence-based recommendations. *Pain* 2007; **132**: 237-251 [PMID: 17920770 DOI: 10.1016/j.pain.2007.08.033]
 - 16 **Dworkin RH**, O'Connor AB, Audette J, Baron R, Gourlay GK, Haanpää ML, Kent JL, Krane EJ, LeBel AA, Levy RM, Mackey SC, Mayer DC, Miaskowski C, Raja SN, Rice AS, Schmadder KE, Stacey B, Stanos S, Treede RD, Turk DC, Walco GA, Wells CD. Recommendations for the pharmacological management of neuropathic pain: an overview and literature update. *Mayo Clin Proc* 2010; **85**: S3-S14 [PMID: 20194146 DOI: 10.4065/mcp.2009.0649]
 - 17 **Attal N**, Cruccu G, Baron R, Haanpää M, Hansson P, Jensen TS, Nurmikko T. EFNS guidelines on the pharmacological treatment of neuropathic pain: 2010 revision. *Eur J Neurol* 2010; **17**: 1113-e88 [PMID: 20402746 DOI: 10.1111/j.1468-1331.2010.02999.x]
 - 18 **Morrison GA**, Sterkers JM. Unusual presentations of acoustic tumours. *Clin Otolaryngol Allied Sci* 1996; **21**: 80-83 [PMID: 8674229 DOI: 10.1111/j.1365-2273.1996.tb01030.x]
 - 19 **Shin YJ**, Lapeyre-Mestre M, Gafsi I, Cognard C, Deguine O, Tremoulet M, Frayssé B. Neurotological complications after radiosurgery versus conservative management in acoustic neuromas: a systematic review-based study. *Acta Otolaryngol* 2003; **123**: 59-64 [PMID: 12625575 DOI: 10.1081/003655402100028084]
 - 20 **Walter J**, Roland PS, Isaacson B. Acoustic Neuroma. Medscape 2011. Available from: URL: <http://emedicine.medscape.com/article/882876-overview#showall>
 - 21 **Hankinson T**, Haque R, Kellner CP, Bruce JN, Sinson GP, Reiter GT. Skull Base Tumours. Medscape 2011. Available from: URL: <http://emedicine.medscape.com/article/250237-overview>
 - 22 **Matthies C**, Samii M. Management of 1000 vestibular schwannomas (acoustic neuromas): clinical presentation. *Neurosurgery* 1997; **40**: 1-10 [DOI: 10.1097/00006123-199701000-0-00001]
 - 23 **Chamadoira C**, Cerejo A, Duarte F, Vaz R. [Trigeminal neuralgia caused by contra lateral cerebellopontine angle tumor. A case report]. *Neurocirugia (Astur)* 2010; **21**: 50-52 [PMID: 20186375]
 - 24 **Chung WY**, Liu KD, Shiao CY, Wu HM, Wang LW, Guo WY, Ho DM, Pan DH. Gamma knife surgery for vestibular schwannoma: 10-year experience of 195 cases. *J Neurosurg* 2005; **102** Suppl: 87-96 [PMID: 15662787 DOI: 10.3171/jns.2005.102.s_supplement.0087]
 - 25 **Lunsford LD**, Niranjan A, Flickinger JC, Maitz A, Kondziolka D. Radiosurgery of vestibular schwannomas: summary of experience in 829 cases. *J Neurosurg* 2005; **102** Suppl: 195-199 [PMID: 15662809 DOI: 10.3171/jns.2005.102.s_supplement.0195]
 - 26 **Combs SE**, Thilmann C, Debus J, Schulz-Ertner D. Long-term outcome of stereotactic radiosurgery (SRS) in patients with acoustic neuromas. *Int J Radiat Oncol Biol Phys* 2006; **64**: 1341-1347 [PMID: 16464537 DOI: 10.1016/j.ijrobp.2005.10.024]
 - 27 **Sughrue ME**, Yang I, Han SJ, Aranda D, Kane AJ, Amoils M, Smith ZA, Parsa AT. Non-audiofacial morbidity after Gamma Knife surgery for vestibular schwannoma. *Neurosurg Focus* 2009; **27**: E4 [PMID: 19951057 DOI: 10.3171/2009.9.FOCUS09198]
 - 28 **Bennett MI**, Smith BH, Torrance N, Potter J. The S-LANSS score for identifying pain of predominantly neuropathic origin: validation for use in clinical and postal research. *J Pain* 2005; **6**: 149-158 [PMID: 15772908 DOI: 10.1016/j.jpain.2004.11.007]
 - 29 **Bouhassira D**, Attal N, Alchaar H, Boureau F, Brochet B, Bruxelle J, Cunin G, Fermanian J, Ginies P, Grun-Overdyking A, Jafari-Schluep H, Lanteri-Minet M, Laurent B, Mick G, Serrie A, Valade D, Vicaut E. Comparison of pain syndromes associated with nervous or somatic lesions and development of a new neuropathic pain diagnostic questionnaire (DN4). *Pain* 2005; **114**: 29-36 [PMID: 15733628 DOI: 10.1016/j.pain.2004.12.010]
 - 30 **O'Connor AB**, Dworkin RH. Treatment of neuropathic pain: an overview of recent guidelines. *Am J Med* 2009; **122**: S22-S32 [PMID: 19801049]
 - 31 **Finnerup NB**, Otto M, McQuay HJ, Jensen TS, Sindrup SH. Algorithm for neuropathic pain treatment: an evidence based proposal. *Pain* 2005; **118**: 289-305 [PMID: 16213659 DOI: 10.1016/j.pain.2005.08.013]
 - 32 **Jefferies K**. Treatment of neuropathic pain. *Semin Neurol* 2010; **30**: 425-432 [PMID: 20941675 DOI: 10.1055/s-0030-1267286]
 - 33 **E-CPS**. Compendium of pharmaceuticals and specialties. 2012 (online). Available from: URL: <http://www.pharmacists.ca/inline.cfm/function/store/PublicationDetail.cfm?pPub=5>
 - 34 **Cruccu G**, Gronseth G, Alksne J, Argoff C, Brainin M, Burchiel K, Nurmikko T, Zakrzewska JM. AAN-EFNS guidelines on trigeminal neuralgia management. *Eur J Neurol* 2008; **15**: 1013-1028 [PMID: 18721143 DOI: 10.1111/j.1468-1331.2008.02185.x]
 - 35 **Cruccu G**, Truini A. Refractory trigeminal neuralgia. Non-surgical treatment options. *CNS Drugs* 2013; **27**: 91-96 [PMID: 23225488 DOI: 10.1007/s40263-012-0023-0]
 - 36 **Jorns TP**, Zakrzewska JM. Evidence-based approach to the medical management of trigeminal neuralgia. *Br J Neurosurg* 2007; **21**: 253-261 [PMID: 17612914 DOI: 10.1080/02688690701219175]
 - 37 **Lindsay TJ**, Rodgers BC, Savath V, Hettlinger K. Treating diabetic peripheral neuropathic pain. *Am Fam Physician* 2010; **82**: 151-158 [PMID: 20642268]
 - 38 **Moore RA**, Derry S, Aldington D, Cole P, Wiffen PJ. Amitriptyline for neuropathic pain and fibromyalgia in adults. *Cochrane Database Syst Rev* 2012; **12**: CD008242 [DOI: 10.1002/14651858]
 - 39 **Watson CP**, Gilron I, Sawynok J, Lynch ME. Nontricyclic antidepressant analgesics and pain: are serotonin norepinephrine reuptake inhibitors (SNRIs) any better? *Pain* 2011; **152**: 2206-2210 [PMID: 21723037 DOI: 10.1016/j.pain.2011.05.032]
 - 40 **Verdu B**, Decosterd I, Buclin T, Stiefel F, Berner A. Antidepressants for the treatment of chronic pain. *Drugs* 2008; **68**: 2611-2632 [PMID: 19093703 DOI: 10.2165/0003495-200868180-00007]
 - 41 **Watson CP**, Vernich L, Chipman M, Reed K. Nortriptyline

- versus amitriptyline in postherpetic neuralgia: a randomized trial. *Neurology* 1998; **51**: 1166-1171 [PMID: 9781549 DOI: 10.1212/WNL.51.4.1166]
- 42 **Backonja MM**, Irving G, Argoff C. Rational multidrug therapy in the treatment of neuropathic pain. *Curr Pain Headache Rep* 2006; **10**: 34-38 [PMID: 16499828 DOI: 10.1007/s11916-006-0007-1]
- 43 **Vorobeychik Y**, Gordin V, Mao J, Chen L. Combination therapy for neuropathic pain: a review of current evidence. *CNS Drugs* 2011; **25**: 1023-1034 [PMID: 22133325 DOI: 10.2165/11596280-000000000-00000]
- 44 **Taylor CP**. Mechanisms of analgesia by gabapentin and pregabalin--calcium channel alpha2-delta [Cavalpha2-delta] ligands. *Pain* 2009; **142**: 13-16 [PMID: 19128880 DOI: 10.1016/j.pain.2008.11.019]
- 45 **Gilron I**, Bailey JM, Tu D, Holden RR, Weaver DF, Houlden RL. Morphine, gabapentin, or their combination for neuropathic pain. *N Engl J Med* 2005; **352**: 1324-1334 [PMID: 15800228 DOI: 10.1056/NEJMoa042580]
- 46 **NOUGG (National Opioid Use Guideline Group)**. Canadian Guideline for Safe and Effective Use of Opioids for Chronic Non-Cancer Pain. 2010. Available from: URL: <http://nationalpaincentre.mcmaster.ca/opioid/>
- 47 **Heir G**, Karolchek S, Kalladka M, Vishwanath A, Gomes J, Khatri R, Nasri C, Eliav E, Ananthan S. Use of topical medication in orofacial neuropathic pain: a retrospective study. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2008; **105**: 466-469 [PMID: 18329583 DOI: 10.1016/j.tripleo.2007.09.030]
- 48 **Zúñiga C**, Díaz S, Piedimonte F, Micheli F. Beneficial effects of botulinum toxin type A in trigeminal neuralgia. *Arq Neuropsiquiatr* 2008; **66**: 500-503 [PMID: 18813708 DOI: 10.1590/S0004-282X2008000400012]
- 49 **Bohluli B**, Motamedi MH, Bagheri SC, Bayat M, Lassemi E, Navi F, Moharamnejad N. Use of botulinum toxin A for drug-refractory trigeminal neuralgia: preliminary report. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2011; **111**: 47-50 [PMID: 20674409 DOI: 10.1016/j.tripleo.2010.04.043]
- 50 **Wu CJ**, Lian YJ, Zheng YK, Zhang HF, Chen Y, Xie NC, Wang LJ. Botulinum toxin type A for the treatment of trigeminal neuralgia: results from a randomized, double-blind, placebo-controlled trial. *Cephalalgia* 2012; **32**: 443-450 [PMID: 22492424 DOI: 10.1177/0333102412441721]
- 51 **Evans CJ**, Trudeau E, Mertzanis P, Marquis P, Peña BM, Wong J, Mayne T. Development and validation of the Pain Treatment Satisfaction Scale (PTSS): a patient satisfaction questionnaire for use in patients with chronic or acute pain. *Pain* 2004; **112**: 254-266 [PMID: 15561380 DOI: 10.1016/j.pain.2004.09.005]
- 52 **Carlson J**, Youngblood R, Dalton JA, Blau W, Lindley C. Is patient satisfaction a legitimate outcome of pain management? *J Pain Symptom Manage* 2003; **25**: 264-275 [PMID: 12614961 DOI: 10.1016/S0885-3924(02)00677-2]

P- Reviewer: Gottschalk A, Noll-Hussong M, Tao F

S- Editor: Ji FF **L- Editor:** A **E- Editor:** Liu SQ



World Journal of *Pharmacology*

World J Pharmacol 2014 September 9; 3(3): 33-38





MINIREVIEWS

- 33 Overview on metabolomics in traditional Chinese medicine
Qiu S, Zhang AH, Sun H, Yan GL, Wang XJ

Contents*World Journal of Pharmacology*
Volume 3 Number 3 September 9, 2014**APPENDIX** I-V Instructions to authors**ABOUT COVER** Editorial Board Number of *World Journal of Pharmacology*, Xue-Long Sun, Associate Professor, Department of Chemistry, Cleveland State University, 2121 Euclid Avenue, SI 313 Cleveland, OH 44115, United States**AIM AND SCOPE** *World Journal of Pharmacology* (*World J Pharmacol*, *WJP*, online ISSN 2220-3192, DOI: 10.5497) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.
WJP covers topics concerning neuropsychiatric pharmacology, cerebrovascular pharmacology, geriatric pharmacology, anti-inflammatory and immunological pharmacology, antitumor pharmacology, anti-infective pharmacology, metabolic pharmacology, gastrointestinal and hepatic pharmacology, respiratory pharmacology, blood pharmacology, urinary and reproductive pharmacology, pharmacokinetics and pharmacodynamics, clinical pharmacology, and drug toxicology.
We encourage authors to submit their manuscripts to *WJP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.**INDEXING/ABSTRACTING** *World Journal of Pharmacology* is now indexed in Digital Object Identifier.**FLYLEAF** I-IV Editorial Board**EDITORS FOR THIS ISSUE**Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xiu-Xia Song*NAME OF JOURNAL
*World Journal of Pharmacology*ISSN
ISSN 2220-3192 (online)LAUNCH DATE
February 9, 2012FREQUENCY
QuarterlyEDITOR-IN-CHIEF
Geoffrey Burnstock, PhD, DSc, FAA, FRCS (Hon), FRCP (Hon), FmedSci, FRS, Professor, Autonomic Neuroscience Centre, University College Medical School, Royal Free Campus, Rowland Hill Street, London NW3 2PF, United KingdomEDITORIAL OFFICE
Jin-Lei Wang, Director
Xiu-Xia Song, Vice Director*World Journal of Pharmacology*
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
Telephone: +86-10-85381891
Fax: +86-10-85381893
E-mail: editorialoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>PUBLISHER
Baishideng Publishing Group Inc
8226 Regency Drive,
Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>PUBLICATION DATE
September 9, 2014**COPYRIGHT**

© 2014 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORSFull instructions are available online at http://www.wjgnet.com/2220-3192/g_info_20100722180909.htm**ONLINE SUBMISSION**<http://www.wjgnet.com/esp/>

Overview on metabolomics in traditional Chinese medicine

Shi Qiu, Ai-Hua Zhang, Hui Sun, Guang-Li Yan, Xi-Jun Wang

Shi Qiu, Ai-Hua Zhang, Hui Sun, Guang-Li Yan, Xi-Jun Wang, National TCM Key Laboratory of Serum Pharmacochimistry, Laboratory of Metabolomics and Chinmedomics, Department of Pharmaceutical Analysis, Heilongjiang University of Chinese Medicine, Harbin 150040, Heilongjiang Province, China
Author contributions: Qiu S and Sun H conceived of the study and wrote the manuscript; Zhang AH and Wang XJ helped revise the manuscript; Yan GL contributed to manuscript generation.

Supported by National Natural Science Foundation of China, Nos. 81173500, 81373930, 81302905, 81102556, and 81202639; National Key Technology Research and Development Program of the Ministry of Science and Technology of China, Nos. 2011BAI03B03, 2011BAI03B06, and 2011BAI03B08; National Key Subject of Drug Innovation, No. 2009ZX09502-005; and Foundation of Heilongjiang University of Chinese Medicine, No. 201209

Correspondence to: Xi-Jun Wang, Professor, National TCM Key Laboratory of Serum Pharmacochimistry, Laboratory of Metabolomics and Chinmedomics, Department of Pharmaceutical Analysis, Heilongjiang University of Chinese Medicine, Heping Road 24, Harbin 150040, Heilongjiang Province, China. xijunwangls@126.com

Telephone: +86-451-82110818 Fax: +86-451-82110818

Received: March 26, 2014 Revised: July 2, 2014

Accepted: July 27, 2014

Published online: September 9, 2014

Abstract

Metabolomics has been widely used in the modern research of traditional Chinese medicine (TCM). At the same time, the world is increasingly concerned about TCM, and many studies have been conducted to investigate different aspects of TCM. Among these studies, metabolomic approach has been implemented to facilitate TCM development. The current methods for TCM research are diverse, including nuclear magnetic resonance, gas chromatography-mass spectrometry, and liquid chromatography-mass spectrometry. Using these techniques, some advantageous results have been obtained in the studies of TCM, such as diagnosis and treatment, quality control, and mechanisms of action. It is believed that the further development of metabolomic analytical techniques is beneficial to the modern-

ization of TCM. This review summarizes potential applications of metabolomics in the area of TCM. Guidelines for good practice for the application of metabolomics in TCM research are also proposed, and the special role of metabolomics in TCM is highlighted.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Traditional Chinese medicine; Metabolomics; Metabolite; Biomarker; Liquid chromatography-mass spectrometry

Core tip: Traditional Chinese medicine (TCM) has been used for thousands of years to treat or prevent diseases. Actual value of TCM has not been fully recognized worldwide due to the lack of scientific approaches. Metabolomics has become a hot topic in TCM research. Metabolomics is the best method to fit the holistic concept of TCM, and it can not only interpret the essence of syndrome but also elucidate the scientific connotation of prescription. This combination of TCM with metabolomics in modern health care systems may lead to a revolution in TCM therapy.

Qiu S, Zhang AH, Sun H, Yan GL, Wang XJ. Overview on metabolomics in traditional Chinese medicine. *World J Pharmacol* 2014; 3(3): 33-38 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i3/33.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i3.33>

INTRODUCTION

Traditional Chinese medicine (TCM) has been important in health protection and disease control in East Asia for thousands of years. Now, it is getting more and more popular in the whole world for improving health conditions of human beings and preventing or healing diseases. Moreover it shows some advantages in early intervention, individuation medicine and combination therapies. TCM has a distinctive feature, *i.e.*, its systems

theory, which includes the holistic view and the dialectical view. It takes the human body as a whole from the key concepts of “qi, blood, yin-yang, viscera (Zang-Fu), and meridian and channel”^[1]. TCM is also a natural combination of philosophy and ancient science disciplines. The overall concept and differential treatment are the most basic features. Chinese herbal medicine treats a disease by regulating and mobilizing the whole body rather than just regulating a single factor. However, the development of TCM is restricted in the world. The lack of scientific and technologic approaches makes TCM face serious challenges and suffer from inadequate modern research.

Metabolomics is a new subject concerned with the comprehensive characterization of the small molecule metabolites in biological systems. It can distinguish between diseased and non-diseased status information through the assessment of global metabolic profiles in approximative biofluids and biomarker discovery^[2]. Studying the metabolome can highlight changes in networks and pathways and provide advice to physiological and pathological states^[3]. New dimensions are adding to the field of metabolomics by developments in new technology, flux analysis and biochemical modeling^[4-6]. The holistic analyses in the context of metabolomics will give the current essay and the foreseeable developments some relatively clear conclusions^[7].

TCM recognizes the human body by system discrimination and in a cybernetic way. TCM can be characterized as holistic, emphasizing the integrity of the human body. It also pays close attention to the relationship between human and their social and natural environment^[8]. Metabolomics is a promising way for research of TCM and opens a new way for using metabolomic platform to resolve TCM issues^[9]. The metabolomic platforms provide a way to extend the understanding of the mechanisms of action of TCM formulae and the analysis of Chinese herbs, TCM syndromes, mineral medicine, and acupuncture^[10]. They offer a useful tool to identify biomarkers and provide a new method for studying the efficacies and mechanisms of TCM in treating diseases^[11]. Conclusively, metabolomics has become a key to resolving special TCM issues. Here, we give an overview of the applications of metabolomic approaches in research of TCM in recent years.

METABOLOMICS IN DIAGNOSIS AND TREATMENT IN TCM

The superiority and soul of TCM are diagnosis and treatment, and syndrome is the basic concept of the theory. The accuracy of syndrome differentiation determines the effectiveness of TCM treatment. The metabolomic technologies have been used in objectively differentiating syndromes and exploring their biological mechanisms by studying the functional activities of the human body from a system-wide perspective. It will impact our understanding of the theory behind the evidence-based Chinese medicine^[12]. Lu *et al.*^[13] performed the overall biologi-

cal characterization of the urine of psoriasis patients with Blood Stasis Syndrome. Simultaneously, they investigated the therapeutic metabolomic mechanism of the Optimized Yinxieling formula. The findings enhanced the understanding of the metabolic influence in Blood Stasis Syndrome in psoriasis patients and the mechanism of action of optimized Yinxieling^[13]. In addition, that study demonstrated that metabolomics was a powerful tool in diagnosis and treatment of primary dysmenorrhea by providing information on changes in metabolites and endocrinal, neural and immune pathways. Xiang-Fu-Si-Wu Formula intervention can affect some significant perturbations in sphingolipid metabolism, glycerophospholipid metabolism and steroid hormone biosynthesis to make the metabolic discrepancy return to the normal level^[14].

The personalized diagnosis in TCM can help to distinguish different types of diabetics. Metabolomics provides biomarkers for disease subtypes as a potential platform. It has been proved that combining metabolomics with TCM diagnosis can reveal metabolic characteristics for pre-diabetic subtypes^[15]. Xu *et al.*^[16] used the orthogonal signal correction-partial least squares method to confirm the existence of metabolite differences among different TCM syndromes. Additionally, a new method has been developed to distinguish the difference between healthy controls and patients with TCM deficiency syndromes by uncorrelated linear discriminant analysis. It provides important information assisting TCM clinical diagnosis^[16]. Metabolomics has the potential to become a diagnostic tool for diseases and provide a new way to understand pathophysiologic mechanisms. Metabolic pathways including alanine and aspartate were found to be disturbed in jaundice syndrome patients. Using this method, 44 marker metabolites have been identified to distinguish patients with jaundice syndrome from matched healthy controls^[17].

METABOLOMICS IN ACUPUNCTUROLOGY

Acupuncture, as an alternative and complementary therapy, has been used for disease treatment and prevention in TCM^[18]. But, the underlying mechanism of acupuncture is unclear, which precludes its widespread use. Metabolomics is similar to acupuncturology in terms of dynamic changes and comprehensiveness^[19]. The high-throughput metabolomics can identify potential factors for acupuncture effects and provide valuable information towards understanding therapeutic mechanisms. Wang *et al.*^[17] assessed the acupuncture treatment at the “Zusanli” acupoint *via* marker metabolites, based on the perturbed signatures and pathways after acupuncture^[20,21].

Many studies show the potential of an NMR-based metabolomic approach in the research of biological effects of acupuncture. It was used to investigate the metabolic change of plasma before and after electro-acupuncture in senescence-prone mice, providing a method to assess the effects of acupuncture and to understand the

underlying mechanism in neurodegenerative diseases^[22]. Wu *et al.*^[23] have shown that acupuncture demonstrates its therapeutic effects in the relief of functional dyspepsia symptoms. After treatment, the levels of leucine/isoleucine, lactate and glucose in patients significantly changed and lipid levels slightly changed towards those of the healthy controls^[23]. Acupuncture could make the metabolite network recover. An UPLC-MS-based metabolomic method has been developed to investigate the biological effect of acupuncture in acute gouty arthritis and to understand the underlying mechanism^[24].

APPLICATION OF METABOLOMICS TO MATERIAL FOUNDATION

Metabolic profiling is benefit to screening active components in medicinal plants. Li *et al.*^[25] have used metabolomics to find metabolites with antitussive and expectorant activities. It has been shown that chlorogenic acid, 3,5-dicaffeoylquinic acid, and rutin may be closely associated with the antitussive and expectorant activities^[25]. Wang *et al.*^[26] established a UPLC/MS method for analyzing the chemical constituents after oral administration of Yinchenhao Tang (YCHT), which was used for treatment of jaundice syndrome. Forty-five compounds *in vitro* and 21 compounds *in vivo* were detected^[26,27]. The three components of YCHT are *Artemisia annua* L., *Gardenia jasminoids* Ellis, and *Rheum Palmatum* L., whose major active ingredients are 6,7-dimethylsculetin (D), geniposide (G), and rhein (R), respectively. The D/G/R combination had a more robust synergistic effect than any one or two of the three individual compounds by acting upon multiple target proteins^[28].

ACTION MECHANISM RESEARCH

Action mechanisms of most of Chinese medicines are difficult to determine. In an attempt to address the benefits of Chinese medicine using current biomedical approaches, we regard the metabolomics technology as a powerful tool. Metabolomic techniques are promising for identifying biomarkers, clarifying mechanisms of disease, and highlighting insights into drug discovery. Zhao *et al.*^[29] identified 19 metabolites as potential biomarkers of chronic kidney disease, and 10 biomarkers returned to the control levels in *Poria cocos*-treated groups. Furthermore, they found that topical treatment with *Poria cocos* intervenes some primary metabolic pathways^[29]. Scoparone has a potential effect against carbon tetrachloride-induced liver injury through regulating multiple perturbed pathways to the normal state^[30]. The dried root of Kansui (*Euphorbia kansui* L.), an effective TCM, has been researched by NMR analysis. It provides new clues to the toxicity of Kansui from a systematic and holistic view^[31].

Modified Sinisan can have an effect on liver injury through partially regulating the perturbed pathways, such as phenylalanine metabolism, tyrosine and tryptophan biosynthesis, tryptophan metabolism, retinol metabolism,

and tyrosine metabolism^[32]. Gou *et al.*^[33] investigated the effect of Xia Yu Xue Decoction on liver fibrosis by a urinary metabolomic method, based on gas chromatography coupled with GC/MS. It was suggested that the mechanism of action of Xia Yu Xue Decoction may affect ten potential biomarkers associated with microflora metabolism^[33]. Chen *et al.*^[34] have studied the therapeutic mechanism of a traditional Chinese medicine Jiu Wei Qiang Huo decoction effects against H1N1-induced pneumonia by a metabolomic approach. The findings provided a systematic view and a basis for understanding of prevention and treatment^[34].

POTENTIAL OF METABOLOMICS FOR STUDING CHINESE MEDICAL FORMULAE

Chinmedomics, defined as “elucidating the therapeutic and synergistic properties and metabolism of traditional Chinese medical formulae (chinmediformulae) and related metabolic analysis by modern techniques”, has recently showed potential in evaluating TCM^[35]. It supplies a way to translate chinmediformulae into practices. TCM therapy will be revolutionized by the way, which combines chinmedomics with chinmediformulae in modern health care systems^[36]. A developed and validated UPLC-MS/MS method has been used to test the plasma pharmacokinetics, tissue distribution and excretion of schisan-drin (the main component of shengmaisan) in rats after oral administration of shengmaisan. This method can be used to investigate the *in vivo* behaviors of the TCM components in formulae^[37]. The pathogenic mechanism of yinhuang syndrome was investigated by a metabolomics method, which had identified 19 biomarkers for the progression of the yinhuang syndrome^[38].

METABONOMICS IS A ROAD TO QUALITY CONTROL OF TCM

The quality control of Chinese medicine, referring to the TCM preparations comprising more than one herb, is challenging due to their extreme chemical complexity. A chemical fingerprint technique for quality control has been established for identifying herbs from different origins. Fingerprinting analysis could provide a platform to identify herbs from different origins by GC-MS, which is beneficial to quality control. In this way, Longae rhizome samples have been identified as the characteristic components for distinguishing these samples of various geographical origins, which is good for quality control^[39]. A practical quality control method for *A. Radix* has been set up by recognizing GC-based metabolic markers. It identified sorbitol and a glucose/4-aminobutyric acid combination as bio-markers for discriminating species and cultivation area^[40].

Twelve active components in a methanol extract of Weichang'an pill were simultaneously determined using the HPLC-DAD-ESI-MS/MS technique^[41]. Wang *et al.*^[42]

have separated and determined 18 major active ingredients of Banxia Xiexin decoction in order to achieve quality control by UPLC-MS/MS. A fingerprint profile of Niu Huang Shangqing pill has been established and 190 compounds was characterized by HPLC/qTOF-MS. It is a significant method to implement and provides a potential approach to achieve the holistic quality control of complex TCM preparations^[43].

METABONOMICS PROVIDES INSIGHTS INTO THE GLOBAL ISSUES OF TCM TOXICOLOGY

Metabolomics has showed potential to improve the discovery of biomarkers for detection of toxicity. Dong *et al*^[44] utilized global metabolomics to find 17 metabolites, which were regarded as phenotypic biomarkers for toxicity of Chuan Wu. Additionally, the mechanisms of Fuzi's toxicity and potential tissue-specific biomarkers for the toxicity have been explored by metabolomics. Significant changes of 14 lipid metabolites were considered the potential biomarkers for toxicity of Fuzi^[45].

In TCM, a principle called “Jun-Chen-Zuo-Shi” may be used to formulate a herbal formula that can mitigate the toxicity of the main ingredient. NMR-based metabolomics approach has been used to research the toxicity of realgar after being counterbalanced by other TCMs in a TCM prescription named Niu Huang Jiedu Tablet. The counterbalanced realgar in Niu Huang Jiedu Tablet was more secure and much less toxic^[46]. A TCM Paozhi approach can increase potency and reduce toxicity. An RPLC-Q-TOF/MS method based on metabolomic analysis has been explored to help improve the understanding of the transformation mechanisms underlying Paozhi. Twenty-two key biomarkers responding to detoxifying actions of Paozhi were identified^[47]. Sun *et al*^[48] also proved that metabolomic method greatly contributes to the investigation of processed Fuzi and provides useful information for the potential activity and toxicity of processed Fuzi.

CONCLUSION

Metabolomics is a modern technology in the post-genome era and has been used widely in modern Chinese medicine^[49,50]. Metabolomics reflects the function of organisms from terminal symptoms of metabolic network and can help understand metabolic changes of a complete system caused by interventions in the holistic context. Its character is consistent with the whole thinking of TCM, and it may be beneficial to provide an opportunity to scientifically express the meaning of evidence-based Chinese medicine. It shows potential in both TCM research and drug discovery. Metabolomic applications in TCM field related to drug development from natural sources and drug discovery aim at raising the potential of metabolomics in reducing the gap between

TCM and modern drug discovery, and highlight the key role of biomarkers for drug discovery and development of traditional oriental medicine. It is expected that current metabolomic technologies can impel the development of TCM, especially in the understanding of the concept of Chinmedomics. Currently, systems biology is in accordance with the holistic concept and practices of TCM and will help to understand the mechanisms of TCM. As one part of “Omic”, metabolomics, playing an import role in systems biology, has a non-selective approach and can thus lead to the identification of all the metabolites. Metabolomics should be devoted to establishing and improving its own databases, linking other genomics together to solve the problems of TCM, in order to enhance its self-worth in the field of TCM research. Overall, incorporation of metabolomics technologies into TCM can make it possible to study the mechanism of TCM.

REFERENCES

- 1 **Xutian S**, Zhang J, Louise W. New exploration and understanding of traditional Chinese medicine. *Am J Chin Med* 2009; **37**: 411-426 [PMID: 19606504 DOI: 10.1142/S0192415X09006941]
- 2 **Nicholson JK**, Lindon JC. Systems biology: Metabonomics. *Nature* 2008; **455**: 1054-1056 [PMID: 18948945 DOI: 10.1038/4551054a]
- 3 **Sreekumar A**, Poisson LM, Rajendiran TM, Khan AP, Cao Q, Yu J, Laxman B, Mehra R, Lonigro RJ, Li Y, Nyati MK, Ahsan A, Kalyana-Sundaram S, Han B, Cao X, Byun J, Omenn GS, Ghosh D, Pennathur S, Alexander DC, Berger A, Shuster JR, Wei JT, Varambally S, Beecher C, Chinnaiyan AM. Metabolomic profiles delineate potential role for sarcosine in prostate cancer progression. *Nature* 2009; **457**: 910-914 [PMID: 19212411 DOI: 10.1038/nature07762]
- 4 **Kim JA**, Choi HJ, Kwon YK, Ryu do H, Kwon TH, Hwang GS. 1H NMR-based metabolite profiling of plasma in a rat model of chronic kidney disease. *PLoS One* 2014; **9**: e85445 [PMID: 24465563 DOI: 10.1371/journal.pone.0085445]
- 5 **Liu X**, Ser Z, Locasale JW. Development and quantitative evaluation of a high-resolution metabolomics technology. *Anal Chem* 2014; **86**: 2175-2184 [PMID: 24410464 DOI: 10.1021/ac403845u]
- 6 **Di Girolamo F**, Lante I, Muraca M, Putignani L. The Role of Mass Spectrometry in the “Omics” Era. *Curr Org Chem* 2013; **17**: 2891-2905 [PMID: 24376367]
- 7 **Jia J**, Yu Y, Deng JH, Robinson N, Bovey M, Cui YH, Liu HR, Ding W, Wu HG, Wang XM. A review of Omics research in acupuncture: the relevance and future prospects for understanding the nature of meridians and acupoints. *J Ethnopharmacol* 2012; **140**: 594-603 [PMID: 22322253 DOI: 10.1016/j.jep.2012.01.034]
- 8 **Lu AP**, Jia HW, Xiao C, Lu QP. Theory of traditional Chinese medicine and therapeutic method of diseases. *World J Gastroenterol* 2004; **10**: 1854-1856 [PMID: 15222022]
- 9 **Yang B**, Zhang A, Sun H, Dong W, Yan G, Li T, Wang X. Metabolomic study of insomnia and intervention effects of Suanzaoren decoction using ultra-performance liquid-chromatography/electrospray-ionization synapt high-definition mass spectrometry. *J Pharm Biomed Anal* 2012; **58**: 113-124 [PMID: 22019702 DOI: 10.1016/j.jpba.2011.09.033]
- 10 **Zhang A**, Sun H, Wang Z, Sun W, Wang P, Wang X. Metabolomics: towards understanding traditional Chinese medicine. *Planta Med* 2010; **76**: 2026-2035 [PMID: 21058239 DOI: 10.1055/s-0030-1250542]

- 11 **Tan G**, Liao W, Dong X, Yang G, Zhu Z, Li W, Chai Y, Lou Z. Metabonomic profiles delineate the effect of traditional Chinese medicine sini decoction on myocardial infarction in rats. *PLoS One* 2012; **7**: e34157 [PMID: 22493681 DOI: 10.1371/journal.pone.0034157]
- 12 **Zhang A**, Sun H, Qiu S, Wang X. Advancing drug discovery and development from active constituents of yinchenhao tang, a famous traditional chinese medicine formula. *Evid Based Complement Alternat Med* 2013; **2013**: 257909 [PMID: 24191164 DOI: 10.1155/2013/257909]
- 13 **Lu C**, Deng J, Li L, Wang D, Li G. Application of metabolomics on diagnosis and treatment of patients with psoriasis in traditional Chinese medicine. *Biochim Biophys Acta* 2014; **1844**: 280-288 [PMID: 23747921 DOI: 10.1016/j.bbapap.2013.05.019]
- 14 **Liu P**, Duan J, Wang P, Qian D, Guo J, Shang E, Su S, Tang Y, Tang Z. Biomarkers of primary dysmenorrhea and herbal formula intervention: an exploratory metabolomics study of blood plasma and urine. *Mol Biosyst* 2013; **9**: 77-87 [PMID: 23111557 DOI: 10.1039/c2mb25238d]
- 15 **Wei H**, Pasman W, Rubingh C, Wopereis S, Tienstra M, Schroen J, Wang M, Verheij E, van der Greef J. Urine metabolomics combined with the personalized diagnosis guided by Chinese medicine reveals subtypes of pre-diabetes. *Mol Biosyst* 2012; **8**: 1482-1491 [PMID: 22414982 DOI: 10.1039/c2mb05445k]
- 16 **Xu W**, Zhang L, Huang Y, Yang Q, Xiao H, Zhang D. [Plasma fatty acid metabolic profiles for traditional Chinese medicine syndrome differentiation in diabetic patients using uncorrelated linear discriminant analysis]. *Se Pu* 2012; **30**: 864-869 [PMID: 23285965]
- 17 **Wang X**, Zhang A, Han Y, Wang P, Sun H, Song G, Dong T, Yuan Y, Yuan X, Zhang M, Xie N, Zhang H, Dong H, Dong W. Urine metabolomics analysis for biomarker discovery and detection of jaundice syndrome in patients with liver disease. *Mol Cell Proteomics* 2012; **11**: 370-380 [PMID: 22505723 DOI: 10.1074/mcp.M111.016006]
- 18 **Zhao L**, Chen J, Liu CZ, Li Y, Cai DJ, Tang Y, Yang J, Liang FR. A review of acupoint specificity research in china: status quo and prospects. *Evid Based Complement Alternat Med* 2012; **2012**: 543943 [PMID: 23243454 DOI: 10.1155/2012/543943]
- 19 **Gao J**, Liu XG, Yan XZ, Yu SG, Wu QF, Du HB, Liang FR. [Primary analysis on the methodology and strategies for studying mechanisms of acu-moxibustion by using metabolomics]. *Zhen Ci Yan Jiu* 2011; **36**: 296-301 [PMID: 21942185]
- 20 **Zhang Y**, Zhang A, Yan G, Cheng W, Sun H, Meng X, Liu L, Xie N, Wang X. High-throughput metabolomic approach revealed the acupuncture exerting intervention effects by perturbed signatures and pathways. *Mol Biosyst* 2014; **10**: 65-73 [PMID: 24150485 DOI: 10.1039/c3mb70352e]
- 21 **Yan G**, Zhang A, Sun H, Cheng W, Meng X, Liu L, Zhang Y, Xie N, Wang X. Dissection of Biological Property of Chinese Acupuncture Point Zusanli Based on Long-Term Treatment via Modulating Multiple Metabolic Pathways. *Evid Based Complement Alternat Med* 2013; **2013**: 429703 [PMID: 24073005 DOI: 10.1155/2013/429703]
- 22 **Qiao-feng W**, Ling-ling G, Shu-guang Y, Qi Z, Sheng-feng L, Fang Z, Hai-yan Y, Yong T, Xian-zhong Y. A(1)H NMR-based metabonomic study on the SAMP8 and SAMR1 mice and the effect of electro-acupuncture. *Exp Gerontol* 2011; **46**: 787-793 [PMID: 21741463 DOI: 10.1016/j.exger.2011.06.002]
- 23 **Wu Q**, Zhang Q, Sun B, Yan X, Tang Y, Qiao X, Chen Q, Yu S, Liang F. 1H NMR-based metabonomic study on the metabolic changes in the plasma of patients with functional dyspepsia and the effect of acupuncture. *J Pharm Biomed Anal* 2010; **51**: 698-704 [PMID: 19854601 DOI: 10.1016/j.jpba.2009.09.042]
- 24 **Wen SL**, Liu YJ, Yin HL, Zhang L, Xiao J, Zhu HY, Xue JT, Ye LM. Effect of acupuncture on rats with acute gouty arthritis inflammation: a metabonomic method for profiling of both urine and plasma metabolic perturbation. *Am J Chin Med* 2011; **39**: 287-300 [PMID: 21476206]
- 25 **Li ZY**, Zhi HJ, Zhang FS, Sun HF, Zhang LZ, Jia JP, Xing J, Qin XM. Metabolomic profiling of the antitussive and expectorant plant *Tussilago farfara* L. by nuclear magnetic resonance spectroscopy and multivariate data analysis. *J Pharm Biomed Anal* 2013; **75**: 158-164 [PMID: 23261808 DOI: 10.1016/j.jpba.2012.11.023]
- 26 **Wang X**, Sun W, Sun H, Lv H, Wu Z, Wang P, Liu L, Cao H. Analysis of the constituents in the rat plasma after oral administration of Yin Chen Hao Tang by UPLC/Q-TOF-MS/MS. *J Pharm Biomed Anal* 2008; **46**: 477-490 [PMID: 18164893 DOI: 10.1016/j.jpba.2007.11.014]
- 27 **Lv H**, Sun H, Wang X, Sun W, Jiao G, Zhou D, Zhao L, Cao H, Zhang G. Simultaneous determination by UPLC-ESI-MS of scoparone, capillaridin, rhein, and emodin in rat urine after oral administration of Yin Chen Hao Tang preparation. *J Sep Sci* 2008; **31**: 659-666 [PMID: 18264991 DOI: 10.1002/jssc.200700596]
- 28 **Wang X**, Zhang A, Wang P, Sun H, Wu G, Sun W, Lv H, Jiao G, Xu H, Yuan Y, Liu L, Zou D, Wu Z, Han Y, Yan G, Dong W, Wu F, Dong T, Yu Y, Zhang S, Wu X, Tong X, Meng X. Metabolomics coupled with proteomics advancing drug discovery toward more agile development of targeted combination therapies. *Mol Cell Proteomics* 2013; **12**: 1226-1238 [PMID: 23362329 DOI: 10.1074/mcp.M112.021683]
- 29 **Zhao YY**, Lei P, Chen DQ, Feng YL, Bai X. Renal metabolic profiling of early renal injury and renoprotective effects of *Poria cocos* epidermis using UPLC Q-TOF/HSMS/MSE. *J Pharm Biomed Anal* 2013; **81-82**: 202-209 [PMID: 23670099 DOI: 10.1016/j.jpba.2013.03.028]
- 30 **Zhang A**, Sun H, Dou S, Sun W, Wu X, Wang P, Wang X. Metabolomics study on the hepatoprotective effect of scoparone using ultra-performance liquid chromatography/electrospray ionization quadrupole time-of-flight mass spectrometry. *Analyst* 2013; **138**: 353-361 [PMID: 23152956 DOI: 10.1039/c2an36382h]
- 31 **Tang B**, Ding J, Wu F, Chen L, Yang Y, Song F. 1H NMR-based metabonomics study of the urinary biochemical changes in Kansui treated rat. *J Ethnopharmacol* 2012; **141**: 134-142 [PMID: 22406398 DOI: 10.1016/j.jep.2012.02.011]
- 32 **Liu CG**, Wang XL, Du XW, Jiang DY, Geng NZ, Zhang SX, Zhou YY, Kuang HX. Metabolomic profiling for identification of potential biomarkers in the protective effects of modified Sinsan against liver injury in dimethylnitrosamine treated rats. *Biol Pharm Bull* 2013; **36**: 1700-1707 [PMID: 24189414]
- 33 **Gou X**, Tao Q, Feng Q, Peng J, Sun S, Cao H, Zheng N, Zhang Y, Hu Y, Liu P. Urinary metabolomics characterization of liver fibrosis induced by CCl₄ in rats and intervention effects of Xia Yu Xue Decoction. *J Pharm Biomed Anal* 2013; **74**: 62-65 [PMID: 23245234 DOI: 10.1016/j.jpba.2012.09.021]
- 34 **Chen L**, Fan J, Li Y, Shi X, Ju D, Yan Q, Yan X, Han L, Zhu H. Modified Jiu Wei Qiang Huo decoction improves dysfunctional metabolomics in influenza A pneumonia-infected mice. *Biomed Chromatogr* 2014; **28**: 468-474 [PMID: 24132661 DOI: 10.1002/bmc.3055]
- 35 **Wang X**, Zhang B. [Elucidation of compatibility principle and scientific value of Chinese medical formulae based on pharmacometabolomics]. *Zhongguo Zhong Yao Za Zhi* 2010; **35**: 1346-1348 [PMID: 20707212]
- 36 **Wang X**, Zhang A, Sun H. Future perspectives of Chinese medical formulae: chinmedomics as an effector. *OMICS* 2012; **16**: 414-421 [PMID: 22734809 DOI: 10.1089/omi.2011.0138]
- 37 **Lu SW**, Zhang AH, Sun H, Yan GL, Han Y, Wu XH, Wang XJ. Ultra-performance liquid-chromatography with tandem mass spectrometry for rapid analysis of pharmacokinetics, biodistribution and excretion of schisandrin after oral administration of Shengmaisan. *Biomed Chromatogr* 2013; **27**: 1657-1663 [PMID: 23852935 DOI: 10.1002/bmc.2976]

- 38 **Tong X**, Sun H, Yan GL, Dong W, Sun WJ, Yu Y, Wang P, Han Y, Wang XJ. Evaluation study on urine metabolomics in yinhuang rat model induced by triplet factors of rhubarb, ethanol, and α -naphthylisothiolyanate. *Chin J Integr Med* 2011; **17**: 369-375 [PMID: 21611901 DOI: 10.1007/s11655-011-0728-9]
- 39 **Hu Y**, Kong W, Yang X, Xie L, Wen J, Yang M. GC-MS combined with chemometric techniques for the quality control and original discrimination of *Curcumae longae* rhizome: analysis of essential oils. *J Sep Sci* 2014; **37**: 404-411 [PMID: 24311554 DOI: 10.1002/jssc.201301102]
- 40 **Kobayashi S**, Putri SP, Yamamoto Y, Donghyo K, Bamba T, Fukusaki E. Gas chromatography-mass spectrometry based metabolic profiling for the identification of discrimination markers of *Angelicae Radix* and its application to gas chromatography-flame ionization detector system. *J Biosci Bioeng* 2012; **114**: 232-236 [PMID: 22633242 DOI: 10.1016/j.jbiosc.2012.03.022]
- 41 **Zhang J**, Gao W, Liu Z, Zhang Z. Identification and Simultaneous Determination of Twelve Active Components in the Methanol Extract of Traditional Medicine Weichang'an Pill by HPLC-DAD-ESI-MS/MS. *Iran J Pharm Res* 2013; **12**: 15-24 [PMID: 24250567]
- 42 **Wang Y**, Xu R, Xiao J, Zhang J, Wang X, An R, Ma Y. Quantitative analysis of flavonoids, alkaloids and saponins of Banxia Xiexin decoction using ultra-high performance liquid chromatography coupled with electrospray ionization tandem mass spectrometry. *J Pharm Biomed Anal* 2014; **88**: 525-535 [PMID: 24189040 DOI: 10.1016/j.jpba.2013.10.002]
- 43 **Wang DD**, Liang J, Yang WZ, Hou JJ, Yang M, Da J, Wang Y, Jiang BH, Liu X, Wu WY, Guo DA. HPLC/qTOF-MS-oriented characteristic components data set and chemometric analysis for the holistic quality control of complex TCM preparations: Niu Huang Shangqing pill as an example. *J Pharm Biomed Anal* 2014; **89**: 130-141 [PMID: 24284229 DOI: 10.1016/j.jpba.2013.10.042]
- 44 **Dong H**, Zhang A, Sun H, Wang H, Lu X, Wang M, Ni B, Wang X. Inguenuity pathways analysis of urine metabolomics phenotypes toxicity of Chuanwu in Wistar rats by UPLC-Q-TOF-HDMS coupled with pattern recognition methods. *Mol Biosyst* 2012; **8**: 1206-1221 [PMID: 22282765 DOI: 10.1039/c1mb05366c]
- 45 **Cai Y**, Gao Y, Tan G, Wu S, Dong X, Lou Z, Zhu Z, Chai Y. Myocardial lipidomics profiling delineate the toxicity of traditional Chinese medicine *Aconiti Lateralis radix praeparata*. *J Ethnopharmacol* 2013; **147**: 349-356 [PMID: 23541933 DOI: 10.1016/j.jep.2013.03.017]
- 46 **Xu W**, Wang H, Chen G, Li W, Xiang R, Pei Y. (1)H NMR-based metabolomics study on the toxicity alleviation effect of other traditional Chinese medicines in Niu Huang Jiedu tablet to realgar (*As₂S₂*). *J Ethnopharmacol* 2013; **148**: 88-98 [PMID: 23583735 DOI: 10.1016/j.jep.2013.03.073]
- 47 **Li Y**, Wang Y, Su L, Li L, Zhang Y. Exploring potential chemical markers by metabolomics method for studying the processing mechanism of traditional Chinese medicine using RPLC-Q-TOF/MS: a case study of *Radix Aconiti*. *Chem Cent J* 2013; **7**: 36 [PMID: 23432780 DOI: 10.1186/1752-153X-7-36]
- 48 **Sun H**, Ni B, Zhang A, Wang M, Dong H, Wang X. Metabolomics study on Fuzi and its processed products using ultra-performance liquid-chromatography/electrospray-ionization synapt high-definition mass spectrometry coupled with pattern recognition analysis. *Analyst* 2012; **137**: 170-185 [PMID: 22030742 DOI: 10.1039/c1an15833c]
- 49 **Dong Y**, Ding Y, Liu PZ, Song HY, Zhao YP, Li M, Shi JR. Investigation of the Material Basis Underlying the Correlation between Presbycusis and Kidney Deficiency in Traditional Chinese Medicine via GC/MS Metabolomics. *Evid Based Complement Alternat Med* 2013; **2013**: 762092 [PMID: 24371466 DOI: 10.1155/2013/762092]
- 50 **Ding X**, Hu J, Wen C, Ding Z, Yao L, Fan Y. Rapid resolution liquid chromatography coupled with quadrupole time-of-flight mass spectrometry-based metabolomics approach to study the effects of jieduquyuzi Yin prescription on systemic lupus erythematosus. *PLoS One* 2014; **9**: e88223 [PMID: 24505438 DOI: 10.1371/journal.pone.0088223]

P- Reviewer: Tang WF, Wang XP **S- Editor:** Ji FF
L- Editor: Wang TQ **E- Editor:** Lu YJ



World Journal of *Pharmacology*

World J Pharmacol 2014 December 9; 3(4): 39-216

Volume End



REVIEW

- 39 Dried-leaf *Artemisia annua*: A practical malaria therapeutic for developing countries?
Weathers PJ, Towler M, Hassanali A, Lutgen P, Engeu PO
- 56 Asthma in pregnancy
Blackburn HK, Allington DR, Procacci KA, Rivey MP
- 72 Targeted approaches for HER2 breast cancer therapy: News from nanomedicine?
Mazzucchelli S, Truffi M, Fiandra L, Sorrentino L, Corsi F
- 86 Telomerase activity: An attractive target for cancer therapeutics
Picariello L, Grappone C, Polvani S, Galli A
- 97 Patents on antivirulence therapies
López M, Barbosa B, Gato E, Bou G, Tomás M
- 110 Harnessing pharmacological knowledge for personalized medicine and pharmacotyping: Challenges and lessons learned
Vizirianakis IS
- 120 Phosphoprotein phosphatase 1-interacting proteins as therapeutic targets in prostate cancer
Felgueiras J, Fardilha M
- 140 Potential ability of xanthophylls to prevent obesity-associated cancer
Terasaki M, Mutoh M, Fujii G, Takahashi M, Ishigamori R, Masuda S

MINIREVIEWS

- 153 Pharmacological role of efflux transporters: Clinical implications for medication use during breastfeeding
Ahmadzai H, Tee LBG, Crowe A
- 162 Pharmacophore approaches in protein kinase inhibitors design
Starosyla SA, Volynets GP, Bdzhola VG, Golub AG, Yarmoluk SM

174 Role of antipsychotics for treating behavioral and psychological symptoms of dementia

Yap KZ, Chan SY

186 Use of eltrombopag in thrombocytopenia of liver disease

Sharma V

**THERAPEUTICS
ADVANCES**

193 Perspective of antiviral therapeutics for hepatitis C after liver transplantation

Ho CM, Hu RH, Lee PH

**EVIDENCE-BASED
MEDICINE**

199 Vitamin D and bone fracture healing

Ray M

SYSTEMATIC REVIEWS

209 Association of serum bilirubin and non-alcoholic fatty liver disease: A feasible therapeutic avenue?

Anwar MS, Dillon JF, Miller MH

APPENDIX I-V Instructions to authors

ABOUT COVER Editorial Board Number of *World Journal of Pharmacology*, Won Suk Lee, MD, PhD Professor, Department of Pharmacology, Pusan National University School of Medicine, Yangsan, Gyeongsangnam-do 626-870, South Korea

AIM AND SCOPE *World Journal of Pharmacology* (*World J Pharmacol*, *WJP*, online ISSN 2220-3192, DOI: 10.5497) is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJP covers topics concerning neuropsychiatric pharmacology, cerebrovascular pharmacology, geriatric pharmacology, anti-inflammatory and immunological pharmacology, antitumor pharmacology, anti-infective pharmacology, metabolic pharmacology, gastrointestinal and hepatic pharmacology, respiratory pharmacology, blood pharmacology, urinary and reproductive pharmacology, pharmacokinetics and pharmacodynamics, clinical pharmacology, and drug toxicology.

We encourage authors to submit their manuscripts to *WJP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING *World Journal of Pharmacology* is now indexed in Digital Object Identifier.

FLYLEAF I-IV Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
 Responsible Electronic Editor: *Ya-Jing Lu*
 Proofing Editor-in-Chief: *Lian-Sheng Ma*
 Responsible Science Editor: *Yue-Li Tian*
 Proofing Editorial Office Director: *Jin-Lei Wang*

NAME OF JOURNAL
World Journal of Pharmacology

ISSN
 ISSN 2220-3192 (online)

LAUNCH DATE
 February 9, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Geoffrey Burnstock, PhD, DSc, FAA, FRCS (Hon), FRCP (Hon), FmedSci, FRS, Professor, Autonomic Neuroscience Centre, University College Medical School, Royal Free Campus, Rowland Hill Street, London NW3 2PF, United Kingdom

EDITORIAL OFFICE
 Jin-Lei Wang, Director
 Xiu-Xia Song, Vice Director

World Journal of Pharmacology
 Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China
 Telephone: +86-10-85381891
 Fax: +86-10-85381893
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 8226 Regency Drive, Pleasanton, CA 94588, USA
 Telephone: +1-925-223-8242
 Fax: +1-925-223-8243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esp/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLICATION DATE
 December 9, 2014

COPYRIGHT
 © 2014 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjgnet.com/2220-3192/g_info_20100722180909.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/esp/>

Dried-leaf *Artemisia annua*: A practical malaria therapeutic for developing countries?

Pamela J Weathers, Melissa Towler, Ahmed Hassanali, Pierre Lutgen, Patrick Ogwang Engeu

Pamela J Weathers, Melissa Towler, Department of Biology and Biotechnology, Worcester Polytechnic Institute, Worcester, MA 01609, United States

Ahmed Hassanali, School of Pure and Applied Sciences, Kenyatta University, Nairobi 20100, Kenya

Pierre Lutgen, IFBV-BELHERB, PO Box 98, L-6908 Niederanven, Luxembourg

Patrick Ogwang Engeu, Natural Chemotherapeutics Research Institute, Ministry of Health, PO Box 4864 Kampala, Uganda

Author contributions: Weathers PJ, Towler M, Hassanali A, Lutgen P and Engeu PO all participated in writing the article; Hassanali A, Lutgen P and Engeu PO provided clinical data; Weathers PJ and Towler M conducted analyses of lab and field samples.

Supported by Worcester Polytechnic Institute and University of Massachusetts Center for Clinical and Translational Science partially; partially by Award Number NIH-R15AT008277-01 from the National Center for Complementary and Alternative Medicine

Correspondence to: Pamela J Weathers, Professor, Department of Biology and Biotechnology, Worcester Polytechnic Institute, 100 Institute Rd., Worcester, MA 01609, United States. weathers@wpi.edu

Telephone: +1-508-8315196 Fax: +1-508-8315936

Received: June 25, 2014 Revised: September 9, 2014

Accepted: October 1, 2014

Published online: December 9, 2014

(mainly mono and sesqui), flavonoids, and polyphenolic acids. In addition, polysaccharide constituents of *A. annua* may enhance bioavailability of artemisinin. Rodent pharmacokinetics showed longer $T_{1/2}$ and T_{max} and greater C_{max} and AUC in *Plasmodium chabaudi*-infected mice treated with *A. annua* dried leaves than in healthy mice. Pharmacokinetics of deoxyartemisinin, a liver metabolite of artemisinin, was more inhibited in infected than in healthy mice. In healthy mice, artemisinin serum levels were > 40-fold greater in dried leaf fed mice than those fed with pure artemisinin. Human trial data showed that when delivered as dried leaves, 40-fold less artemisinin was required to obtain a therapeutic response compared to pure artemisinin. ACTs are still unaffordable for many malaria patients, and cost estimates for *A. annua* dried leaf tablet production are orders of magnitude less than for ACT, despite improvements in the production capacity. Considering that for > 2000 years this plant was used in traditional Chinese medicine for treatment of fever with no apparent appearance of artemisinin drug resistance, the evidence argues for inclusion of affordable *A. annua* dried leaf tablets into the arsenal of drugs to combat malaria and other artemisinin-susceptible diseases.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Malaria; Infectious disease; *Artemisia annua*; Artemisinin; Combination therapy; Artemisinin combination therapy

Core tip: Artemisinin, extracted from the plant *Artemisia annua* (*A. annua*) L., and artemisinin derivatives are the current best antimalarial therapeutics and are delivered as artemisinin combination therapy (ACT). Availability and cost are problematic for the developing world where malaria is endemic. Oral consumption of *A. annua* dried leaves is more effective than the pure drug. A tea infusion of the leaves has prophylactic effects. Cost of producing and delivering the tea and *A. annua* dried leaf tablets is much more affordable than ACT.

Abstract

Artemisinin from the plant *Artemisia annua* (*A. annua*) L., and used as artemisinin combination therapy (ACT), is the current best therapeutic for treating malaria, a disease that hits children and adults especially in developing countries. Traditionally, *A. annua* was used by the Chinese as a tea to treat "fever". More recently, investigators have shown that tea infusions and oral consumption of the dried leaves of the plant have prophylactic and therapeutic efficacy. The presence of a complex matrix of chemicals within the leaves seems to enhance both the bioavailability and efficacy of artemisinin. Although about 1000-fold less potent than artemisinin in their antiplasmodial activity, these plant chemicals are mainly small molecules that include other artemisinic compounds, terpenes

Weathers PJ, Towler M, Hassanali A, Lutgen P, Engeu PO. Dried-leaf *Artemisia annua*: A practical malaria therapeutic for developing countries? *World J Pharmacol* 2014; 3(4): 39-55 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/39.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.39>

INTRODUCTION

Nearly three billion people are affected by malaria with almost a million deaths annually, especially in Africa and amongst children^[1]. Currently extracted from *Artemisia annua* (*A. annua*) L., artemisinin (Figure 1) is delivered in concert with another antimalarial drug [artemisinin combination therapy (ACT)] as the preferred treatment to slow emergence of drug resistance. Despite these efforts, artemisinin resistance is appearing^[2] and persistent and/or asymptomatic malaria may also be playing a role in disease transmission^[3-5]. Moreover, for developing countries ACT is costly and the supply is inadequate^[6-9].

Artemisinin is a sesquiterpene lactone that is produced and stored in the glandular trichomes that are mainly on the leaves and floral buds of *A. annua*, a GRAS medicinal herb^[10-12]. The plant also produces > 40 flavonoids^[13], many polyphenols, and a variety of other terpenes including mono-, sesqui-, di-, and triterpenes^[14]. As discussed later, many of these have weak antimalarial activity, and, based on transcriptome analyses, many also seem to be produced and/or stored in the glandular trichomes that also contain artemisinin^[15].

We and others proposed direct consumption of *A. annua* either as a tea infusion^[16-19] or by oral consumption of the leaves^[20-24]. In contrast to the oral consumption of pure artemisinin, we showed that the presence of plant material significantly enhanced appearance of artemisinin in the serum of healthy and *Plasmodium chabaudi*-infected mice^[22]. Because of the plethora of mild antimalarial compounds naturally present in the dried leaves of the plant, we have termed this orally consumed dried leaf therapeutic plant-based artemisinin combination therapy, or pACT. These whole plant approaches are similar to the more than 2000 year traditional use of the plant by the Chinese^[25].

To produce a therapeutically effective drug using a complex material like a medicinal plant requires that a number of key factors be met: the medicinal herbal product must be therapeutically effective; levels of key chemical components in the herb must be verifiably consistent; production must also be cost effective. Here we summarize and update our recent review^[26] on the effects of *A. annua* on malaria and further discuss the bioavailability and therapeutic efficacy of pACT and how such an herbal drug could inexpensively be produced with a consistent dose.

PROPHYLACTIC USE OF *A. ANNUA*

Tea infusion, its chemistry, and in vitro studies

Until recently, there have been, to our knowledge, few

well-controlled studies examining extraction, recovery, and stability of artemisinin and other compounds in *A. annua* tea infusion. A systematic study of preparations of *A. annua* therapeutic tea infusion was performed by van der Kooy *et al*^[27] and showed that nearly 93% of available artemisinin was extracted from dried *A. annua* leaves, but only under certain conditions. Best preparation method was: 9 g DW leaves/L, for 5 min at 100 °C. Subsequent storage of the tea infusion at room temperature showed that artemisinin concentration was stable for > 24 h, important for malaria-endemic locations where there is no refrigeration. Artemisinin water solubility is approximately 50 mg/L^[27], so the amount of artemisinin recovered from hot water tea infusions is reasonable. Other studies using the same extraction protocol also measured extraction and stability of artemisinin and some key flavonoids in the tea. Artemisinin was found to be stable at room temperature for up to 48 h^[28]; however, some flavonoids were poorly extracted and not stable at room temperature^[29].

Carbonara *et al*^[28] detected an assortment of phenolics, including 0.06 mg/g DW cirsilineol, in an *A. annua* tea infusion prepared at about a 4-10 fold higher proportion (approximately 38 g DW/L) than that proposed as optimal (9 g DW/L) by van der Kooy *et al*^[27]. Most of the measured phenolics in the tea remained constant at room temperature for 48 h post-infusion. More recently, Suberu *et al*^[19] identified milligram amounts of phenolic acids, flavonoids, and sesquiterpenes in a liter of *A. annua* tea, all of which demonstrated IC₅₀ values in the micromolar or less range (Table 1). Indeed, the IC₅₀ of the tea infusion itself was 7.6 and 2.9 nmol/L for the chloroquine (CQ)-sensitive HB3 and CQ-insensitive Dd2 strains of *P. falciparum*, respectively, and better than artemisinin alone suggesting synergism of constituents in the tea mixture. Clearly if a tea infusion is to be a therapeutic option, it must be consistently and reliably prepared and ingested. As suggested by van der Kooy *et al*^[27], ideally a liter of tea infusion would be prepared daily and consumed in equal aliquots of about 250 mL over 24 h for several days.

Tea infusion clinical trials

Ogwang *et al*^[30,31] tested *Artemisia* tea as a prophylaxis against malaria in 132 adult farm workers, aged 18-60 years, for 12 mo in a randomized clinical trial in Uganda. Tea infusion was consumed once a week at 2.5 g dried leaves per adult infusion dose with 55-100 mg artemisinin/L. Malaria was tracked for 9 mo while adverse clinical effects were tracked for 12 mo. Among those who used *Artemisia* tea there were 80% fewer fever-related hospital visits. Indeed, some patients reported using *A. annua* tea for > 7 years with no incidence of malaria and no serious adverse events. Although this study suggested that once weekly consumption of *A. annua* tea infusion may offer prophylactic protection, there were no children or elderly in the study, so additional clinical trials need to be conducted with different populations and age groups. Authors argued that since a single weekly dose was effec-

Table 1 Antimalarial compounds in *Artemisia annua* vs falciparum malaria

Compound	Compound IC ₅₀ (μmol/L)	Compound + artemisinin IC ₅₀ (nmol/L)	Ref.
Terpenes			
Artemisinin	0.033	Not applicable	Liu <i>et al</i> ^[52]
	0.022, 0.023 ¹		
Artemisinic acid	77.8, 61.6 ¹	No numerical value provided; response depended on concentration of compound tested with artemisinin	Suberu <i>et al</i> ^[19]
Arteannuin B	3.2, 4.8 ¹		
Dihydroartemisinic acid	21.1, 17.7 ¹		
Nerolidol	9 ⁴	Interaction with artemisinin	van Zyl <i>et al</i> ^[55]
α-pinene	1 ⁴	not yet tested	
1,8-cineole (eucalyptol)	70 ⁴		
Limonene	533 ⁴		
Phenolic acids			
Chlorogenic acid	69.4, 61.4 ¹	No numerical value provided; response depended on concentration of compound tested with artemisinin	Suberu <i>et al</i> ^[19]
Rosmarinic acid	65.1, 65.0 ¹		
Flavonoids			
Artemetin	26	26	Liu <i>et al</i> ^[52]
Casticin	24	26	
Cirsilineol	23	22.5	
Chrysopenol-D	32	15	
Chrysopenetin	36	16	
Eupatorin	65	30	
Isovitexin	72.5, 48.1 ¹	Interaction with artemisinin	Suberu <i>et al</i> ^[19]
Luteolin	11, 12 ²	not yet tested	Lehane <i>et al</i> ^[54]
Kaempferol	33, 25 ²		
Myricetin	40, 76 ²		
Quercetin	15, 14 ² ,		Ganesh <i>et al</i> ^[58]
	14.7, 4.11, 2.94 ³		
Rutin	7.1, 3.5, 10.38 ³		

¹Against CQ-sensitive HB3 and CQ-resistant Dd2 strains, respectively; ²Against CQ-sensitive 3D7 and CQ-resistant 7G8 strains, respectively; ³Against fresh Bangladeshi isolates, CQ-sensitive 3D7, and CQ-resistant K1 strains, respectively; ⁴Against CQ-resistant FCR-3. CQ: Chloroquine.

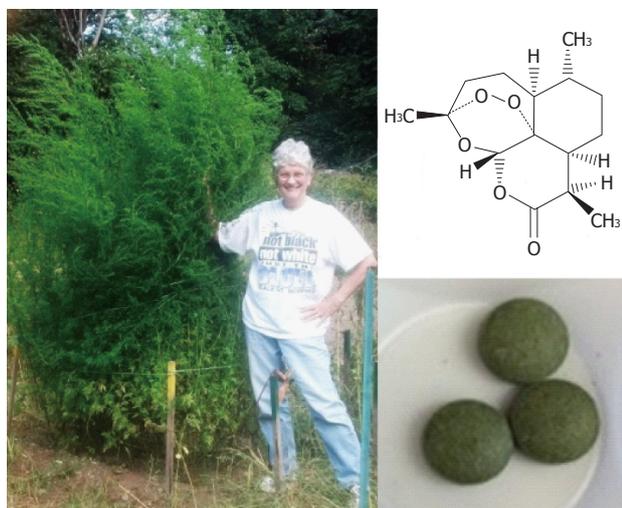


Figure 1 *Artemisia annua* (single clone of *Artemisia annua* cultivar at approximately 2 m height at floral bud formation), artemisinin and plant-based artemisinin combination therapy tablets.

tive, compounds other than artemisinin may have played the prophylactic role since artemisinin itself has short plasma half-life.

THERAPEUTIC USE OF *A. ANNUA*

Tea infusion

Reports on the efficacy of *A. annua* (cv. Artemis) tea on

human malaria patients by Mueller *et al*^[17,32] and Blanke *et al*^[33] yielded at times conflicting results. Their tea infusions contained 47-94 mg artemisinin/L, but recrudescence was much lower in the quinine-treated control group, so parasite reappearance in the tea-treated patients was ascribed to recrudescence and not re-infection^[17]. In the Blanke *et al*^[33] trial that included a placebo tea, recrudescence was consistently lower in the tea patients than in those treated with 500 mg pure artemisinin. More recently, however, De Donno *et al*^[34] showed that 5 g dried leaves in one liter of *A. annua* tea infusion was effective against both CQ-resistant (W2) and CQ-sensitive (D10) strains of *P. falciparum* with IC₅₀ values of 5.60 nmol/L and 7.08 nmol/L, respectively, results also consistent with those of Suberu *et al*^[19] as already highlighted. These latter *in vitro* studies suggested that tea should be efficacious, so why the discrepancy with the earlier human trials? Preparation methodology is crucial for preserving as much biochemical integrity of the plant as possible^[27]. The more recent *in vitro* studies likely used more consistently prepared tea infusions than the earlier human trials, so variations in chemical composition of the infusions and in the plant source material could explain the different responses.

The argument that tea is a monotherapy is unsubstantiated considering the now well-established chemical complexity and related antiparasmodial activity of tea infusions of *A. annua* and its components. Although data from therapeutic tea trials in animals and in humans cor-

Table 2 Kenyan human trial data^[20] for orally delivered dried leaf *Artemisia annua* (plant-based artemisinin combination therapy)

pACT (dried leaf <i>A. annua</i> tablets, ea 500 mg, 3.7 mg artemisinin/tablet)					
Artemisinin dose (mg)		No. of patients	Leaf DW (g)		% Recrudescence
Day 1	Days 2-6		Day 1	Days 2-6	
7.4 × 2	3.7 × 2	12	2	1	25
11.1 × 2	7.4 × 2	12	3	2	9.1
14.8 × 2	11.1 × 2	12	4	3	16.7
18.5 × 2	14.8 × 2	12	5	4	9.1
Compare to orally delivered pure artemisinin ^[39]					
Day 1	Day 2-7				
500 × 2	500	227	NA		24

A. annua: *Artemisia annua*; pACT: Plant-based artemisinin combination therapy; NA: Not available.

relate well, unfortunately, they do not support use of *A. annua* tea for treating malaria because animal and human data are comparably negative, the artemisinin dose is not easily controlled, and other potentially synergistic components in the tea are not readily controlled or extracted. Nevertheless, use of the tea could play a role in malaria prophylaxis to reduce incidence of malaria in different communities, or in temporary relief from malaria, mainly in prevention of coma or “to buy time” to enable an infected person from a rural area to travel to a hospital or clinic stocked with ACT.

Dried leaf *A. annua* - pACT

Recently, Elfawal *et al.*^[23] measured parasitemia in mice infected with *P. chabaudi* that were fed two different doses (0.6 or 3.0 mg artemisinin; 24 and 120 mg/kg) of either pure artemisinin in mouse chow or as pACT. Artemisinin delivered *via* pACT was at least five times more effective, and with a longer lasting response, than pure artemisinin in reducing parasitemia. Excluding artemisinin there are > 600 phytochemicals that have been identified in *Artemisia annua*^[35], but there is currently a lack of information on the chemistry, effect of the preparation method (harvesting, drying, storage, *etc.*), and overall bioavailability of these chemicals^[36].

Clinical trials using dried leaf *A. annua* are scarce in the scientific literature and few, other than those in Democratic Republic of Congo by Mueller *et al.*^[17,32], are published. Despite the fact that WHO does not encourage either whole plant or tea infusion clinical trials^[37], some African universities have been conducting their own trials, many of which have not been published nor results assessed by polymerase chain reaction (PCR) as later done for clinical trials with ACTs (personal comm from C. Kasongo to P. Lutgen). Many of these trials used *A. annua* infusions, and compared to controls or even other antimalarial drugs, *e.g.*, artesunate-amodiaquine, showed significantly greater sensitivity of the infusion with fewer late therapeutic failures. For example, in Democratic Republic of Congo, 54 malaria-infected volunteers were treated for 10 d with capsules containing powdered leaves

of *A. annua*. Each patient was given 15 g dried leaves containing 15 mg of artemisinin (artemisinin content in leaves = 0.1%^[38]). After 2 d all were free of fever and 51 (or 94%) were parasite free after 10 d.

In a study aimed at preventing severe post-operative malaria at Bangui, Central Africa, powdered leaves of *A. annua* were administered in capsules to 25 patients, 22 of them children aged 1-16 years^[24]. Treatment duration ranged from 3-4 d with a dose of 0.4-0.5 g/d of *A. annua* dried leaves (0.1% artemisinin leaf content) delivering 0.4-0.5 mg/d artemisinin. In spite of the very low administered daily dose of artemisinin, average parasitemia dropped by 62% in the patients with an added benefit of a strong antinociceptive response, especially beneficial to post-operative patients.

The most clinically definitive study to date of pACT efficacy was conducted at the International Centre of Insect Physiology and Ecology (ICIPE) Mbita Field campus, Suba District, in Western Kenya. This was a collaborative project between ICIPE and Kenya Medical Research Institute^[20] (Table 2^[39]) and was an open-label, non-randomized clinical trial mainly targeted to assess efficacy, safety, and tolerance of increasing doses of pACT delivered as tablets. The tablets were made by a Tanzania-based NGO, Natural Uwemba System for Health, from a hybrid of *A. annua* grown in the Tanzania highlands (2000-2200 m altitude). Leaves were harvested just before flowering, dried for approximately 3 wk under shade, then crushed, powdered, homogenized, and pressed into 500 mg tablets under ambient temperature. Tablets were robust with no excipient required. Using HPLC with diode array detector, analysis of hexane extracts of randomly selected batches of 100 tablets showed artemisinin content of the tablets was consistent at 0.74% ± 0.06% (*i.e.*, approximately 3.7 mg per tablet).

The four cohorts of the trial each had 12 consenting patients aged 15-56 years (average 23.42) with *P. falciparum* malaria. Based on Giemsa-stained blood smears counted against 200 wbc, parasitemia was 0.02%-4% and hemoglobin levels > 8 mg/dL. Each cohort received one of four increasing numbers of *A. annua* tablets, ranging from 2-5 tablets twice on day 1, followed by 1-4 tablets twice daily for the next 5 d (Table 2). A week following the treatments, three patients scattered throughout different cohorts showed re-appearance of parasites in blood smears; however, all doses were effective in clinical and parasitological regression of malaria, with 9%-20% recrudescence at day 28 and no measurable toxicity.

Compared to the usual large pure artemisinin doses of 1000 mg on day 1 followed by 500 mg on each of days 2-7 that were administered to 227 malaria patients^[39], patients treated with pACT had generally better therapeutic outcomes (Table 2). The measured pACT cure rate also was comparable to or exceeded other results using pure artemisinin^[40,41], and similar levels of artemisinin (artesunate, artemether, *etc.*)^[42]. Furthermore, the positive therapeutic response using pACT appeared somewhat independent of dose beyond the second level of dose test-

ed (Table 2^[20]). Although oral doses used in the ICIPE^[20] trials were far less than any tea studies, levels of recrudescence were much lower than tea and often better than in studies using pure artemisinin^[39] (Table 2). Indeed, about 100 total mg of total artemisinin delivered *via* pACT for a full malaria treatment yielded a better recrudescence rate than the 4000 mg of pure artemisinin used by Gao *et al.*^[39] (Table 2). This 40-fold difference correlates well with the early pharmacokinetic studies by Weathers *et al.*^[21] that showed 45-fold enhanced bioavailability of the drug when delivered as pACT.

These results suggest that the natural phytochemical blend in pACT is important especially when orally administered as tablets. The results are also consistent with a study in China on mice infected with *P. berghei*, which compared the effects of pure artemisinin with crude *A. annua* extracts^[43], and the studies by Elfawal *et al.*^[23] and Weathers *et al.*^[22]. In all three studies the administered products had comparable levels of artemisinin, but crude preparations and pACT were at least 3.5 times more effective in reducing parasitemia than pure artemisinin, suggesting a synergistic role for non-artemisinin constituents in the extracts and orally consumed dried leaves.

COMPARATIVE PHARMACOKINETICS AND BIOAVAILABILITY

Orally delivered artemisinin

When given orally or rectally, dihydroartemisinin showed higher bioavailability in humans than artemisinin in an early pharmacokinetic study by Zhao *et al.*^[44]. The C_{max} , T_{max} , and $T_{1/2}$ for orally delivered dihydroartemisinin were 0.13-0.71 mg/L, 1.33 h, approximately 1.6 h, respectively; for pure artemisinin they were 0.09 mg/L, 1.5 h, and 2.27 h, respectively. Alin *et al.*^[45] compared orally delivered artemisinin and artemisinin-mefloquine combination therapy for treatment of *P. falciparum* malaria. Infected and uninfected patients had similar pharmacokinetic parameters. After a single dose, bioavailability of artemisinin was not altered. Interestingly, pharmacokinetics were similar when comparing treatment failures with successes, suggesting that studies that only measure artemisinin pharmacokinetics were inadequate for predicting therapeutic success^[45]. Ilet *et al.*^[46] also reviewed artemisinin pharmacokinetics in patients with falciparum malaria and reported a dose of 9.1 mg/kg, which was comparable to that of Alin *et al.*^[45]. C_{max} and T_{max} values did not differ much from those reported by Alin *et al.*^[45].

In the Ilet *et al.*^[46] review of pharmacokinetic parameters of artemisinin and its derivatives, oral pure artemisinin doses ranged from about 6-11 mg kg/L in healthy subjects and C_{max} was 0.15-0.39 mg/L. Dose seemed to have no major effect. An earlier study by Ashton *et al.*^[47] compared increasing artemisinin doses of 250, 500, and 1000 mg per person and both C_{max} and $T_{1/2}$ showed dose-dependent increases of 0.21, 0.45, and 0.79 mg/L, and 1.38, 2.0, and 2.8 h, respectively, but T_{max} remained relatively constant at 2.3-2.8 h.

Diet is an important consideration for any orally delivered drug, and when Dien *et al.*^[48] compared artemisinin oral doses given with and without food, C_{max} values were similar between subjects who fasted and those who did not. Food consumption along with artemisinin did not seem to affect artemisinin absorption. In contrast, a later rodent study by Weathers *et al.*^[21] observed that when artemisinin was consumed as part of a complex plant material, pACT, approximately 45-fold more drug entered the serum of mice than orally administered pure drug. Similarly, when pure artemisinin was fed to mice, it was not detectable in the serum after 60 min. However, artemisinin was detected in the serum when consumed in conjunction with mouse chow, which consists of a variety of plant materials including soy, oats, wheat, alfalfa, beet pulp, corn, *etc.*^[22].

In a study by Ashton *et al.*^[49], artemisinin at 9.1 mg/kg was given daily for 7 d, and measurements taken on days 1, 4, 7, and 21. On day 1 plasma C_{max} and $T_{1/2}$ were similar and comparable to data from other studies using a similar dose. On day 4 and 7, however, C_{max} decreased, while $T_{1/2}$ increased, indicating that although artemisinin was delivered daily for 7 d, it was either not readily absorbed or it degraded after the first dose. After the third dose, C_{max} fell from 0.31 to 0.11 mg/L, and $T_{1/2}$ increased from 3.0 to 4.8 h. These results suggested that either artemisinin was metabolized or accumulated elsewhere in the body.

In the liver, cytochrome P450 (CYP450) enzymes metabolize artemisinin to deoxyartemisinin, deoxydihydroartemisinin, 9,10-dihydrodeoxyartemisinin, and a metabolite named "crystal 7"^[50]. Extended artemisinin dosing may not be beneficial as shown by Svensson *et al.*^[50] using human liver microsomes where activity of CYP450s, CYP2B6 in particular, correlated with decreasing artemisinin serum levels. In intermittent dosing studied by Ashton *et al.*^[49], the P450 levels were allowed to decline for 14 d before delivery of another dose, and C_{max} rose from 0.11 to 0.20 mg/L, and $T_{1/2}$ decreased from 4.8 to 2.7 h. Generally, maximum concentration of artemisinin in the body increased with increasing doses with $T_{1/2}$ ranging from about 1.4-4.8 h for reported trials using oral pure artemisinin. Thus, increased and extended artemisinin treatment may reduce recrudescence.

Tea infusion delivered artemisinin

Other than R ath *et al.*^[16], there are few reports on the pharmacokinetics of tea infusion artemisinin delivered in humans. In the R ath *et al.*^[16] study, artemisinin C_{max} was 0.24 mg/L at 0.6 h post consumption. Tea infusion containing 94.5 mg artemisinin had a C_{max} equivalent to a dose of 250 mg pure artemisinin, but at a significantly shorter T_{max} , 0.6 h *vs* 2.8 h^[47]. Compared to pure artemisinin, the shorter half-life of artemisinin in the tea infusion may account for the observed higher recrudescence. Although tea-delivered artemisinin seemed more bioavailable, its shorter $T_{1/2}$ of 0.9 h compared with about 2 h for pure artemisinin, suggested that more than two doses per day may be more beneficial; indeed, four doses a day were

recommended.

The unacceptably high recrudescence rates in clinical tea infusion trials were attributed to low plasma concentrations, almost 40% lower than that for traditional doses (500 mg per person of 60 kg or 8.3 mg artemisinin/kg) of pure artemisinin. Although not specified, tea trial doses have been estimated at about 1.5 mg/kg, close to the 1.1 mg/kg dose of pure artemisinin used by Zhao *et al.*^[44], which is far below the 8.3 mg/kg that is traditionally accepted as pharmacologically effective. Nevertheless, the C_{max} of 0.24 mg/L artemisinin for the tea dose is nearly twice that for pure artemisinin ($C_{max} = 0.13$ mg/L) as measured by Zhao *et al.*^[44]. *A. annua* tea also showed potent antiparasmodial activity against 40 field isolates of *P. falciparum* collected in Pikine, Senegal (mean IC_{50} 0.095 μ g/mL^[51]).

Dried leaf (pACT) delivered artemisinin

There are as yet no pharmacokinetic studies of pACT in humans. In a small PK study of healthy mice fed artemisinin there was about 45-fold more artemisinin delivered *via* pACT than when delivered as the pure drug^[21]. More recently, pharmacokinetics of artemisinin and one of its liver metabolites, deoxyartemisinin, were compared over 120 min in healthy and *P. chabaudi*-infected mice treated with dried *A. annua* leaves at a 100 mg/kg body weight dose of artemisinin^[22]. In pACT-treated healthy mice, the first order elimination rate constant for artemisinin was estimated to be 0.80/h, corresponding to a $T_{1/2}$ of 51.6 min. C_{max} and T_{max} were 4.33 mg/L and 60 min, respectively. The AUC was 299.5 μ gmin/mL. The first order absorption rate constant was estimated at 1.39/h. In contrast, the AUC for pACT-treated infected mice was greater at 435.6 μ g min/mL. Serum levels of artemisinin in the infected mice continued to increase over the 120 min of the study period. As a result, the elimination half-life, $T_{1/2}$ could not be determined, so C_{max} and T_{max} could only be estimated at ≥ 6.64 mg/L and ≥ 120 min, respectively. Nevertheless, both C_{max} and T_{max} of artemisinin were greater in infected than in healthy mice.

Generally, artemisinin concentrations decreased with a concomitant rise in deoxyartemisinin levels only in healthy subjects^[22]. In contrast, artemisinin levels in infected mice continued to rise over the study period whilst deoxyartemisinin levels fell and then leveled, so infection seemed to retard the capacity of the mice to process artemisinin into deoxyartemisinin over the two-hour period. Many compounds in *A. annua* inhibit *P. falciparum*^[52-53] and CYP3A^[56]. At the high (100 mg/kg) dose used in the study, nearly equal amounts of artemisinin and deoxyartemisinin were measured in the serum, indicating that an excessive dose of artemisinin was used.

The presence of plant material affected artemisinin pharmacokinetics. At 60 min no artemisinin was detected in serum of mice fed pure artemisinin at 100 mg/kg body weight. When plant material was present, however, as mouse chow or *A. annua* pACT, artemisinin level in the serum rose to 2.44 and 4.32 μ g/mL, respectively,

demonstrating that the presence of plant material, even mouse chow, had a major positive impact on the appearance of artemisinin in the blood^[22]. To our knowledge, these are the only data available on pharmacokinetics for orally delivered *A. annua* in animals or humans.

NON-ARTEMISININ THERAPEUTIC COMPOUNDS IN *A. ANNUA*

Flavonoids

A. annua is rich in essential oils, coumarins, polyphenols, polysaccharides, saponins, terpenes, and flavonoids. The levels of flavonoids and other compounds in *A. annua* change with developmental growth stage, with some being highest during full bloom^[57]. There are > 40 flavonoids^[13], and at least 11, including artemetin, casticin, chrysoplenetin, chrysoplenol-D, cirsilineol, eupatorin, kaempferol, luteolin, myricetin, quercetin, and rutin, are reported to have weak therapeutic efficacy against falciparum malaria (Table 1^[52-54,58]). Some of these flavonoids were shown to improve the IC_{50} of artemisinin against *P. falciparum* *in vitro* by as much as 50%, suggesting synergy (Table 1^[52]). Elford *et al.*^[53] also showed that while casticin [5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-3,6,7-trimethoxychromen-4-one] showed synergism with artemisinin, it did not synergize with chloroquine, suggesting a different interactive mechanism. Combining casticin with artemisinin inhibited parasite-mediated transport systems that control influx of myo-inositol and L-glutamine in malaria-infected erythrocytes. These apparent synergistic actions between flavonoids and artemisinin suggest that flavonoids are likely to be important for efficacious use of *A. annua* consumed either as whole dried leaves or as tea.

Many flavonoids have antiplasmodial effects and inhibit *P. falciparum* growth in liver cells *in vitro* as reported for dietary flavonoids^[54]. To our knowledge, there are no reports on pharmacokinetics of *A. annua* delivered flavonoids. Some flavonoids are reported to have long plasma half-lives; *e.g.*, quercetin, found in *A. annua* and most fruits, has a plasma half-life of 27 h^[59]. Quercetin [2-(3,4-dihydroxyphenyl)-3,5,7-trihydroxy-4H-chromen-4-one], also found in garlic, inhibits parasite growth with differential activity against different strains of *Plasmodium* (Table 1^[54,58]). Rutin, which is a rutinose [α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranose] glycoside of quercetin, showed similar results, suggesting that the sugar moiety did not significantly affect antimalarial activity (Table 1^[58]). Flavonoids are known to persist in the body for > 5 d; this may explain the once a week dose inducing a prophylactic effect from *A. annua* tea infusion that was reported by Ogwang *et al.*^[30,31]. Many dietary flavonoids inhibit *Plasmodium* growth *in vitro*, but amounts in the diets are reportedly insufficient to offer protection against malaria^[54]. Plants such as *A. annua* with high concentrations of flavonoids (*e.g.*, up to 0.6%) may, however, work in concert with artemisinin to prevent malaria when consumed regularly.

The flavone luteolin [2-(3,4-dihydroxyphenyl)-5,7-

dihydroxy-4-chromenone] comprises up to 0.0023% DW in *Artemisia*^[14] and has been used for a variety of ailments including cough, diarrhea, dysentery, diabetes, cancer, and malaria. Although luteolin has an IC₅₀ value around 11 μmol/L^[54] and is one of the more active antiplasmodial flavonoids found in *A. annua*, one cannot compare its role between studies as indicated by Ganesh *et al.*^[58] (see Table 1). The antimalarial response of different flavonoids seems to be affected by the strain of *Plasmodium* being tested. Luteolin also prevents completion of a full intra-erythrocytic cycle by inhibiting progression of parasite growth beyond the young trophozoite stage. The mechanism of this antiplasmodial activity seems to be related to the inhibition of parasite fatty acid biosynthesis. These lipids are required by the parasite to detoxify heme into hemozoin^[60]. Independent of the human host, apicomplexan parasites use a fatty acid biosynthetic pathway. Enzymes in the pathway, like the NADPH-dependent *b*-ketoacyl-ACP reductase (FabG), are potential antimalarial targets. Among 30 flavonoids studied, luteolin and quercetin had the lowest IC₅₀ values for the inhibition of these enzymes and also showed *in vitro* activity in the sub-micromolar range against multiple strains of *P. falciparum*^[60].

Isovitexin {5,7-dihydroxy-2-(4-hydroxyphenyl)-6-[(2S,3R,4R,5S,6R)-3,4,5-trihydroxy-6-(hydroxymethyl)oxan-2-yl]chromen-4-one} is another flavone, the 6-*C*-glucoside of apigenin, that was found in *A. annua* tea infusion at > 100 mg/L with micromolar antiplasmodial activity (Table 1^[19,28]). Isovitexin inhibits lipid peroxidation and xanthine oxidase activity and protects cells from ROS damage with an overall LD₅₀ > 400 μmol/L^[61].

Terpenes

Limonene (1-methyl-4-(1-methylethenyl)-cyclohexene) is part of the “cineole cassette” that includes 1,8-cineole (eucalyptol), limonene, myrcene, α-pinene, β-pinene, sabinene, and α-terpineol^[62]; many of these affect particular stages of *Plasmodium* species. For example, limonene is often present at 7 mg/kg in *A. annua*^[14] and inhibits isoprenoid biosynthesis in *Plasmodium*^[63] and development at the ring and trophozoite stages^[64]. Eucalyptol affects the trophozoite stage^[65]. Limonene also arrests protein isoprenylation in *P. falciparum*, halting parasite development within 48 h of treatment^[64]. The IC₅₀ against *in vitro Plasmodium* in these trials was 2.27 mmol/L, more than twice the IC₅₀ of 533 μmol/L measured by van Zyl *et al.*^[55]. Limonene and its metabolites remain in the plasma for at least 48 h^[66], so the pharmacokinetics is favorable, which is important for elimination of gametocytes and malaria transmission.

The volatile monoterpene α-pinene (4,6,6-trimethylbicyclo[3.1.1]hept-3-ene) is present in the plant at levels up to 0.05% of dry weight^[14]; it has an IC₅₀ of 1.2 μmol/L, in the range of quinine at 0.29 μmol/L^[55]. Eucalyptol (1,8-cineole) may comprise up to 30% [0.24%-0.42% (V/DW)] of the essential oil in *A. annua*^[67] and is a strong inhibitor of the pro-inflammatory cytokines tumor necrosis factor (TNF)-α, interleukin (IL)-6 and IL-8^[68]. Both

chloroquine-resistant and chloroquine-sensitive *Plasmodium* strains are affected at the early trophozoite stage^[65].

Eucalyptol (1,3,3-trimethyl-2-oxabicyclo[2,2,2]octane) is also volatile and rapidly enters the blood when delivered either as an inhalant or orally^[69,70]. At an IC₅₀ of 0.02 mg/mL and low toxicity (LD₅₀ of approximately 25 mg/mL), either oral or inhalation delivery is reasonable^[65,71]. Indeed eucalyptol concentrations can reach 15 μg/mL in 60 min^[69] suggesting its possible use as an antimalarial inhalant.

Artemisia ketone (3,3,6-trimethyl-1,5-heptadien-4-one), a major constituent of some cultivars of *A. annua*, has barely been studied. Other ketones like curcumin^[72] have been implicated as inhibitors of β-hematin synthesis, so artemisia ketone may play a similar role and affect hemozoin formation. Although hemoglobin is required for *Plasmodium* survival and multiplication in merozoites inside the red blood cell, it leaves toxic debris like heme. The parasite subsequently oxidizes Fe²⁺ in heme to Fe³⁺ forming hematin, a nontoxic insoluble polymeric crystal called β-hematin (also known as hemozoin), which also inhibits cell-mediated immunity against the parasite. Water extracts of *A. annua* inhibit hemozoin synthesis^[73].

Essential oils often contain a large amount of monoterpenes that may enhance the antimalarial effect of artesunate and even reverse the observed resistance of *P. berghei* against artesunate^[74]. Monoterpenes tend to be higher in the pre-flowering phase of *A. annua*^[75], but are drastically reduced by high drying temperatures or drying in the sun^[13,76] and, of particular concern, during compression of dried leaves into tablets^[77]. Although monoterpenes have some antimalarial potential, most are rather volatile and thus they may be therapeutically less important than the nonvolatile flavonoids, phenolic acids, and higher molecular weight sesquiterpenes.

Unlike α-pinene and eucalyptol, camphor (1,7,7-trimethylbicyclo[2.2.1]heptan-2-one) has no reported antimalarial activity, but it may comprise as much as 43.5% of the essential oil of *A. annua*^[78]. Considering camphor is less volatile than either eucalyptol or α-pinene (melting points of 204 °C, 176 °C, and 155 °C, and flash points of 54 °C, 49 °C, and 33 °C, respectively), it may instead play a role in enhanced transport of hydrophobic molecules like artemisinin from pACT across the intestinal wall into the bloodstream^[21,22]. Camphor may also affect thymocyte viability and aid in developing malaria immunity through production of T-cells^[79]. At 50 μg/mL, camphor increased viability of cultured thymocytes^[80].

The sesquiterpene nerolidol (3,7,11-trimethyl-1,6,10-dodecatrien-3-ol) has an IC₅₀ of 0.99 μmol/L and arrests development of the intraerythrocytic stages of the parasite (Table 1^[55]). Indians of the Amazon basin in Brazil treated malaria using the vapors of the leaves of *Viola surinamensis*; nerolidol was identified as the active constituent leading to 100% growth inhibition at the schizont stage^[81]. Nerolidol levels vary with the cultivar tested, with one of the highest values found in plants from Ethiopia^[82]. There is a greater concentration of this sesquiterpene in stems

than leaves of *A. annua*^[83].

Other sesquiterpenes found in the artemisinin biosynthetic pathway were only recently shown to have antiplasmodial activity at $\mu\text{mol/L}$ levels, similar to that of other compounds found in the plant (Table 1^[19]). These artemisinic compounds were extracted into *A. annua* tea infusions and showed varying interactions with artemisinin depending on their relative concentrations and the target parasite strain. For example, arteannuin B showed an additive interaction with artemisinin against the CQ-sensitive *Plasmodium* HB3 strain, while against the CQ-insensitive Dd2 strain the interaction was synergistic.

Phenolic acids

Rosmarinic ((2''R'')-2-[[[(2''E'')-3-(3,4-dihydroxyphenyl)-1-oxo-2-propenyl]]oxy]-3-(3,4-dihydroxyphenyl) propanoic acid) and chlorogenic ((1S,3R,4R,5R)-3-[(2Z)-3-(3,4-dihydroxyphenyl)prop-2-enyl]oxy}-1,4,5-trihydroxycyclohexanecarboxylic acid) acids are strong antioxidants found in a wide variety of *A. annua* cultivars^[56]. In Caco-2 studies, these acids significantly inhibited activity of CYP3A4, one of the hepatic P450s responsible for metabolism of artemisinin to deoxyartemisinin, an inactive form of the drug^[50]. These and other phenolic acids are present in *A. annua* tea infusion^[19]. Both phenolic acids have an IC_{50} of about $65 \mu\text{mol/L}$ (Table 1) and also significantly reduced secretion of cytokines IL-6 and IL-8, and thus enhanced antimalarial activity while reducing inflammation^[56].

Other compounds often found in *A. annua* and that may affect pACT efficacy

Although polysaccharides in other medicinal plants have been more extensively studied, they seem to have been rather overlooked in *A. annua*, probably because most *Artemisia* extracts are obtained using organic solvents and polysaccharides are only soluble in water. Polysaccharides extracted from *Artemisia wayomogi* showed hydroxyl radical scavenging activity three times stronger than glutathione or caffeic acid, and ROS inhibition was twice as strong as ascorbic acid^[84]. In *A. wayomogi*, more polysaccharides were found in stems than in leaves and their solubility was also higher from stem than from leaf tissue^[84].

The combination of polysaccharides with lipophilic molecules like artemisinin may lead to a higher bioavailability of the antimalarial constituents when delivered via *A. annua*, which may explain the lower effective therapeutic dose against malaria observed for pACT than for pure artemisinin^[20,23,26]. Indeed, Han^[85] showed that ginseng polysaccharides had preventive and curative antimalarial activities and synergized with artesunate in malaria-infected mice. Sulfated polysaccharides inhibited the *in vitro* invasion of merozoites into erythrocytes and interfered with merozoite surface protein^[86-88]. Heparin and other sulfated polysaccharides have been shown to inhibit blood-stage growth of plasmodium^[89,90]. Some sulfated polysaccharides inhibited the formation of rosettes between infected red blood cells (iRBC) and unin-

fectured RBCs, as well as adhesion of iRBCs to placental chondroitin sulfate A, which is linked to severe disease outcome in pregnancy-associated malaria^[91].

Saponins, common in many plants, have an important role in human and animal nutrition and are reportedly present in *A. annua*, but only as measured in alcoholic extracts using the nonquantitative foaming test^[92,93] (Weathers, unpublished). These soap-like amphiphilic (lypo- and hydro-philic) bioactive compounds are mainly produced by plants. Recently, there has been interest in the clinical use of saponins as chemotherapeutic agents^[94], and as adjuvants for vaccines^[95]. At very low doses saponins are efficient, have hemolytic properties, produce 40-50 Å pores in erythrocyte membranes, and modulate the sodium pump and ATPase^[96]. Saponins also have a hypoglycemic effect mainly by inhibiting intestinal permeability and absorption of glucose and may therefore inhibit the growth of *P. falciparum*, which needs glucose to grow^[97]. Better identification, quantification, and investigation into the role of saponins in pACT efficacy are warranted.

The coumarin, scopoletin (7-hydroxy-6-methoxychromen-2-one), also known for its antinociceptive properties^[98,99], is commonly found in most *Artemisia* species at, for example, about 0.2% (w/w) in a Luxembourg cultivar. Known for its anti-oxidant, hepatoprotective, and anti-inflammatory activities, scopoletin scavenging capacity for hydroxyl radical, DPPH, superoxide anion, hydrogen peroxide, and Fe^{2+} chelating activity is almost at the level of α -tocopherol (Vitamin E)^[100].

Although not antiplasmodial, scopoletin inhibits TNF- α , IL-6, and IL-8 at millimolar concentrations, and is thus likely one of the major anti-inflammatory and antipyretic constituents of *A. annua*^[101]. Coumarins can activate lymphocytes, thereby stimulating immunological functions^[102]. Indeed, scopoletin induced cell proliferation in normal lymphocytes with an immunomodulatory effect^[101]. In uninfected erythrocytes internal Na concentration is much lower than external concentration, but the K concentration is higher; in infected blood cells this situation is drastically reversed^[103]. Scopoletin significantly stimulated erythrocyte membrane ATPases at $0.1 \mu\text{mol/L}$, in particular Na-K-ATPase vs Ca-ATPase or Mg-ATPase^[104], so scopoletin may affect malaria infection. A significant hormetic effect was also noticed; stimulation was higher at scopoletin concentrations of $10 \mu\text{g/mL}$ than at 1 or at $100 \mu\text{g/mL}$. In addition scopoletin also inhibited ADP-platelet aggregation at a range of 0.1 to $5 \mu\text{mol/L}$ and improved blood rheology^[105].

Scopoletin may also affect the interaction between malaria and uric acid. Cyclical fevers and high levels of inflammation characterize malaria and this likely aids parasite clearance. Excessive and persistent inflammation, on the other hand, can lead to severe malaria^[106]. In the cytoplasm of their parasitophorous vacuole, *Plasmodium*-infected erythrocytes contain uric acid precipitates that are released upon erythrocyte rupture. Uric acid precipitates are mediators for inflammatory cytokines IL-6, IL-8, and are considered a danger signal for innate immu-

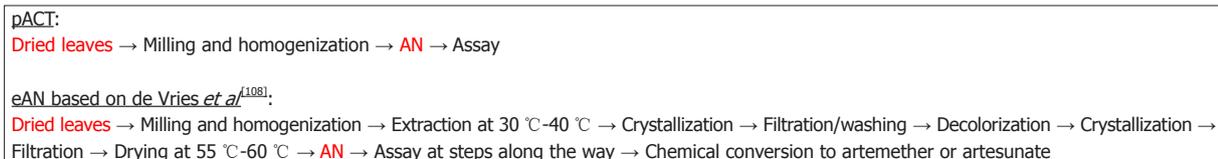


Figure 2 Comparison between plant-based artemisinin combination therapy production and extracted artemisinin from dry harvested leaves to product ready either for packaging (plant-based artemisinin combination therapy) or conversion to artemether or artesunate (extracted artemisinin). AN: Artemisinin; eAN: Extracted artemisinin; pACT: Plant-based artemisinin combination therapy.

nity. Uric acid is also the causative agent in gout. These precipitates could offer a novel molecular target for anti-inflammatory therapies in malaria. Scopoletin inhibits the activity of xanthine oxidase in hyperuricemic mice after peritoneal administration, and this hypouricemic effect is fast and dose-dependent^[107].

Toxicology

Although many of the compounds in *A. annua* have not been tested for their toxicity in, a survey of available MSDS data showed that the LD₅₀ levels for orally administered compounds in rodents ranged from about 160 mg/kg for quercetin to > 8000 mg/kg for nerolidol. The artemisinin LD₅₀ measured *via* oral dose in a mouse was 4228 mg/kg. Therefore, at the estimated amounts of dried leaves of pACT that may be orally consumed by a malaria patient, most of the compounds reported thus far in *A. annua* are at concentrations that are orders of magnitude below their LD₅₀ toxicity values.

Toxicology of the dried leaf tablets used in the Kenyan human trial measured the following components: serum levels of urea, serum proteins, creatinine, γ -glutamyl transferase, serum glutamic pyruvic transaminase, serum glutamic oxaloacetic transaminase, or alkaline phosphatase levels, hemoglobin, and pre- and post-electrocardiograms^[20]. Compared to levels prior to treatment with pACT, there was no significant change post-treatment.

PRODUCTION CONSIDERATIONS

Production comparisons with traditional extraction

Because production costs are usually closely held secrets, there are few cost estimates that are publicly available to compare pACT production with extracted artemisinin. However, costs can be estimated from a study by de Vries *et al.*^[108] where they reported a 1 kg recovery of artemisinin from *A. annua* containing 0.6% artemisinin. Downstream processing costs and product losses increase with increasing number of unit operations (unit ops), a fact often not generally appreciated^[109]. Indeed for biotechnology processes, recovery can be anywhere from 9%-51%^[110]. As an example, if each step of a 4 step process is 95% efficient, then the overall process has a final efficiency of about 81%, while a single step process at 95% efficiency has a 95% overall recovery. The described process steps for extracted artemisinin (eAN) *vs* pACT-AN are shown in Figure 2. From the point of harvested dried leaves to material ready for packaging or conversion to the delivered drug (*e.g.*, artesunate or artemether), pACT has one

unit op and eAN has eight^[108]. Extraction solvents and other chemicals are clearly no longer part of the cost. Because there is one *vs* eight unit ops for eAN and at least two of the eAN unit ops involve significant amounts of heat, pACT energy cost is significantly reduced by at least 90%. Costs for labor, interest, depreciation, and maintenance are all also affected by the number of unit ops^[109], so we estimated that with seven fewer unit op steps those costs would reduce by approximately 88%. Although better extraction processes may be in play^[111], using the de Vries *et al.*^[108] analysis our estimate of cost reduction for producing pACT is about 30% less than the cost of producing eAN. Data provided by de Vries *et al.*^[108] was based on 0.6% artemisinin content, so if a higher producing cultivar was harvested, costs would drop proportionately. Moreover, cost drops again because with pACT there is no need to convert artemisinin to artesunate or artemether; those conversions were necessary because they have higher bioavailability than pure artemisinin, which is not an issue with pACT^[121,22].

The de Vries *et al.*^[108] process cost estimation focuses on a production yield of 1 kg of artemisinin from 500 kg dried leaves, so per Gao *et al.*^[39] that amount of pure artemisinin would treat only 250 patients. Based on the data shown in Table 3 from Kenyan or WPI *A. annua* at 0.7 and 1.4% artemisinin, 15 and 7.5 g DW leaves, respectively, are required for a total adult pACT treatment; so from 500 kg leaves, 33300 and 66600 patients could be treated, respectively. This represents more than a 130-fold increase in patients treated compared to pure artemisinin with proportionate reduction in price.

A. annua dry leaf production varies around the globe. "In East Africa yields average 2.5 T/ha (range = 0.75-4.2)..."^[112]. Based on our field trials^[113], the reported average *A. annua* leaf production in E. Africa^[112], and the doses used in the Kenyan human trial^[20], one can estimate the amount of dry leaf production, and depending on the amount of artemisinin in the biomass, estimate possible number of adult patients that could be treated with pACT (Table 3).

Current ACT drugs vs pACT

Using the dosing information obtained from the Kenyan human malaria trial^[20], each adult needs about 100 mg artemisinin total over 6 d for a malaria treatment, so for *A. annua* leaves with 0.7% artemisinin, 15 g of dried leaves would be needed for a 6 d treatment course. At 2 ton of dried leaves harvested per hectare, 127260 adult patients could be treated for malaria (Table 3). For leaves

Table 3 Estimated numbers of adult patients treatable from plant-based artemisinin combination therapy¹

For <i>A. annua</i> cultivar containing	Number of patients treated at various dry leaf tonnage			
	2 T/ha ²	3 T/ha	4 T/ha ³	5 T/ha
0.7% artemisinin/g DW (Kenyan cultivar)	127260	190890	254520	318150
1.4% artemisinin/g DW (WPI cultivar)	254520	381780	509040	636300

¹Assumptions: each adult needs 100 mg artemisinin (AN) over 6 d for a cure; at 0.7% and 1.4% AN that is approximately 15 and 7.5 g DW leaves, respectively, for a single adult total malaria treatment; ²Below the average of 2.5 T/ha reported for all of East Africa; ³Equal to the maximum obtained growing *A. annua* SAM in the Stow, MA, United States field trials. *A. annua*: *Artemisia annua*.

Table 4 Estimated number of patient treatments by current artemisinin combination therapy *vs* plant-based artemisinin combination therapy

Combination therapy drug	Adult treatments per ton of artemisinin
AL ¹	1.76 million
AS/AQ ¹	2.5 million
pACT leaves with 0.7% artemisinin ²	8.6 million

¹http://www.rollbackmalaria.org/partnership/wg/wgprocurementsupply/docs/psmwg_ppACT-API.pdf, p.2 [cited May 27, 2014]; ²Assumes a 6 day treatment with pACT, with each patient receiving 15 g dried leaves per full malaria treatment for leaves with 0.7% artemisinin. To obtain an amount of artemisinin equal to 1 T of the extracted drug, one would have to harvest 142.8 tons of dried *A. annua* leaves containing 0.7% artemisinin. AL: Artemether/lumefantrine; AS/AQ: Artesunate/amodiaquine; pACT: Plant-based artemisinin combination therapy.

containing 1.4% artemisinin, only 7.5 g of dried leaves are required, so from a hectare of land producing 2 tons of leaves twice as many patients could be treated (Table 3). Clearly choosing cultivars that have higher levels of artemisinin in their leafy biomass will dramatically increase the number of patients that can be treated from 1 ha.

According to Roll Back Malaria, from one ton of purified artemisinin current ACT therapy can provide 1.76 million adult malaria treatments using artemether/lumefantrine, and 2.5 million adult treatments using artesunate/amodiaquine^[114] (Table 4). Using the same one ton artemisinin equivalent, but delivering the drug *via* pACT with 0.7% artemisinin content, one would have harvested about 142.8 tons of dried *A. annua* leaves. Assuming 15 g dried leaves per patient from the dosing data in the Kenyan human malaria trial (Table 2^[20]), 8.64 million adult patients could be treated, about a four-fold increase over either of the current ACT drugs. The actual cost of pACT, therefore, mainly depends on the cost of the dried leaves and their artemisinin content.

As yet unpublished data from the Rich and Weathers labs demonstrated that pACT prevents emergence of artemisinin drug resistance; the plant itself seems to function as its own ACT (pACT). This would obviate the need for inclusion of a co-drug as used in currently administered ACTs. The co-drug costs at least as much as the artemisinin portion of the drug^[6]. Consequently, elimination of the added co-drug could result in at least an additional 50% reduction in cost, so that the final pACT cost reduction is conservatively estimated to be far below that of a current course of ACT therapy.

Considering that *A. annua* is nontoxic and safe to consume orally, dose may not have to be adjusted for children. On the other hand, the leaves taste bitter, so masking the taste, perhaps with sugar, should help with pediatric treatment. Our recent simulated digestion study showed that adding table sugar (sucrose) to pACT did not significantly alter the amount of artemisinin released after digestion, with the added benefit of doubling the amount of flavonoids released^[115].

Comparison with emerging artemisinin sources or other newer antimalarial drugs

There are at least three other emerging antimalarial therapeutic technologies: synthetic artemisinin^[116], semi-synthetic artemisinin (SSA) production from genetically engineered microbes^[117], and a single dose drug, OZ439^[118]. In early 2013, Sanofi/PATH Drug Development Programme, announced they would have the capacity to produce up to 60 MT of SSA in 2014 at about \$400/kg, depending on quantity; Sanofi now has WHO prequalification for its SSA^[119]. Although not much cheaper than the current price of about \$550/kg^[120], supply would be more or less unlimited. Despite what might seem as an advantage to large amounts of SSA production, there are also some serious disadvantages, and comparison of some advantages and disadvantages for each of these new synthetic antimalarial drugs and pACT is noted in Table 5.

QUALITY ASSURANCE

CONSIDERATIONS

Agricultural quality

The traditional and least costly method for cultivating *A. annua* uses seeds and in developing countries farmers prefer to save seeds from one growing season to the next. However, seed generated plants of *A. annua* will vary widely from generation to generation even with high quality starting stock (see review by Ferreira *et al*^[10]). Stem cuttings of *A. annua* readily root in about two weeks, so clonal propagation *via* rooted cutting is recommended to eliminate this variability. Although this method of propagation is not cost effective for large plantations, it would work for a few hectares or for controlled environment agriculture. Given the large numbers of patients that could be treated from growing just a few hectares of *A. annua* (Table 3), clonal propagation by rooted stem

Table 5 Comparison of emerging antimalarial therapeutic technologies with plant-based artemisinin combination therapy

Technology	Advantages	Disadvantages
Synthetic AN ^[116]	Fully synthetic method giving AN = compound Lowers AN cost compared to extraction	Requires co-drug to obviate emergence of AN drug resistance Not yet in production Needs sophisticated process Likely all under Western control
Semi-synthetic AN ^[117]	Semi-synthetic method giving authentic AN Lowers AN cost compared to extraction	Challenging patient compliance due to multiday dosing Requires co-drug to obviate emergence of AN drug resistance Production began <i>via</i> Sanofi Needs sophisticated process Likely all under Western control
OZ439 ^[118]	Single dose cure insures patient compliance In successful Phase 2 trials Mechanism of action not the same as AN Probably low cost due to full synthesis	Challenging patient compliance due to multiday dosing Requires co-drug to obviate emergence of AN drug resistance Not yet in production Needs sophisticated process Likely all under Western control
pACT ^[20-24]	Has its own in planta co-drug to obviate emergence of AN drug resistance Very low cost Very consistent product Can be used to treat other diseases Can be locally owned, produced, managed, and distributed	Not yet in production Likely to meet push back from pharmaceutical industry Challenging patient compliance due to multiday dosing

AN: Artemisinin.

cuttings is recommended. Since pACT therapy involves the direct consumption of the dried leaves of the plant, harvested leaf material must be kept clean, which is easiest to do in controlled environment agriculture and following Good Agricultural Procedures^[121], particularly as applied to fresh produce^[122]. However, controlled agriculture would probably result in loss of agricultural jobs, a concern to be assessed locally. Alternatively, great care must be taken during field harvest and post-harvest storage, so as not to affect the quality of the product. WHO has established good agricultural practices specifically for *A. annua* for purposes of artemisinin extraction^[123], for general medicinal plants^[124], and to minimize contamination of herbal medicines^[125].

Chemical consistency and quantification

To deliver a reliable dose of therapeutics to a patient, the dried leaves of harvested *A. annua* must have a reliable and consistent composition. Clonal propagation provides the required consistency. Recently we showed that of 10 crops harvested from vegetative and early flowering plants grown over three years under diverse conditions in the lab, field, and home garden, the artemisinin content of a single clone of *A. annua* (SAM) was 1.38% ± 0.26% (w/w)^[77]. Thus, despite variations in culture and environmental conditions, a consistent level of the main therapeutic constituent can be achieved. Moreover, the content of harvested leaves is certainly not a guarantee of finished product, *e.g.*, compressed leaf tablets. Analyses by Weathers *et al.*^[77] showed that although artemisinin content was very stable after tablet compression, other constituents varied significantly. For example, although flavonoids increased with tablet compression, the more volatile monoterpenes decreased substantially. Thus, it is critical to monitor the composition profile of both in-

coming harvested material as well as the final product.

Complex and expensive analytical procedures have been used to analyze the many products found in *A. annua*, but they are not necessary to measure and assure product quality. Artemisinin is easily extracted and then can be quantified using a variety of thin layer chromatography (TLC) methods and visualized with *p*-anisaldehyde stain^[126,127]. Other key constituents like the flavonoids are also readily separated using TLC and visualized under either UV ± AlCl₃ reagent^[128]. Total flavonoids also can be quantified using inexpensive visible spectroscopy *via* the AlCl₃ method with quercetin used as an inexpensive standard. To our knowledge no inexpensive, reliable spectrophotometric assay is available to measure artemisinin in complex plant extracts.

SOCIOECONOMIC BENEFITS

Other diseases

Artemisinin and its derivatives are also effective against a number of viruses^[129], a variety of human cancer cell lines^[130-133], and several neglected tropical diseases including schistosomiasis^[134], leishmaniasis^[135,136], trypanosomiasis^[137], and some livestock diseases^[133,138].

Although they rank below malaria in terms of public health importance, schistosomiasis, leishmania, and trypanosomiasis result in estimated annual infections of about 240 million, 1.3 million (0.3 visceral and 1.0 cutaneous), and 30000, respectively^[139]. These diseases along with many others respond to treatment with artemisinins. Although the IC₅₀ is about 1000-fold greater than for *Plasmodium* sp., the greater apparent bioavailability of artemisinin *via* oral pACT^[20-22] would likely reduce the amount of drug required for treatment. At present, pACT has not been tested *in vivo* for diseases other than

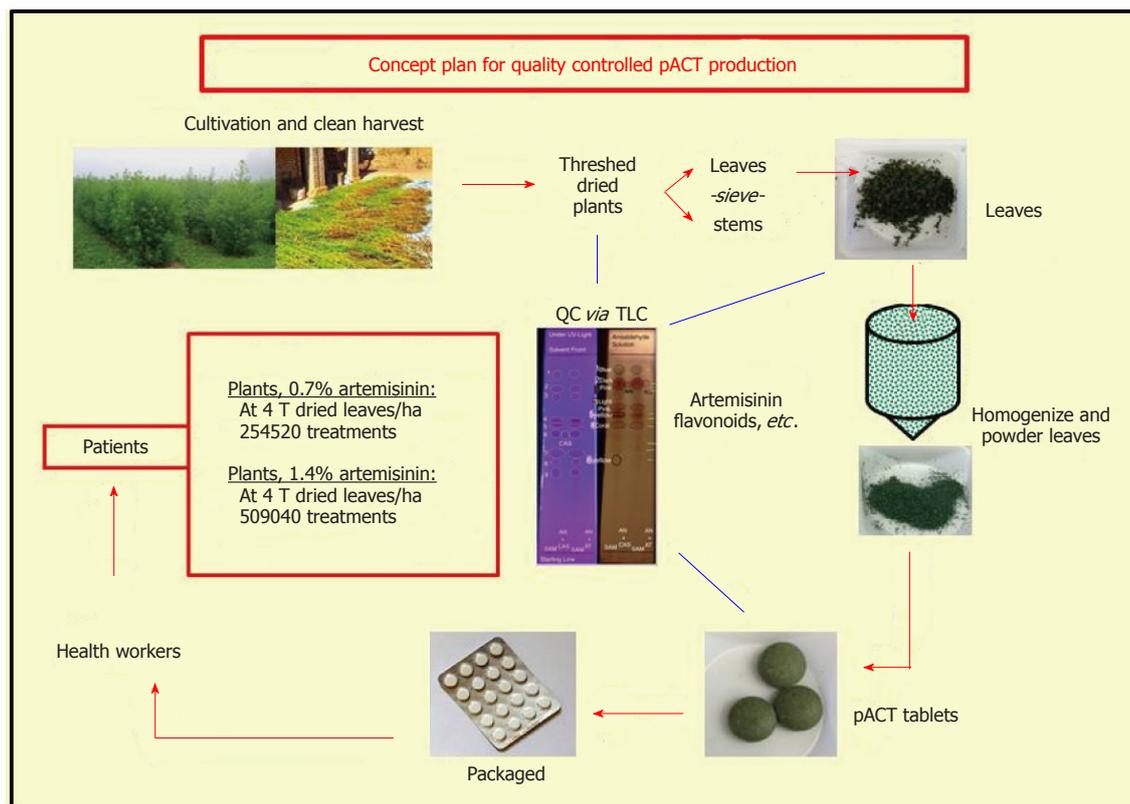


Figure 3 Overall scheme for plant-based artemisinin combination therapy production. pACT: Plant-based artemisinin combination therapy; TLC: Thin layer chromatography.

malaria.

Malaria treatment is further complicated for Human immunodeficiency virus/acquired immune deficiency syndrome (HIV/AIDS) patients. Malaria and HIV co-infection represents a major health burden in Africa mainly because it is now “well established that HIV infection results in a higher incidence and more severe manifestations of malaria”^[140]. With a weakened immune system, AIDS patients are more susceptible to malaria and also respond slower to malaria therapy^[140-142]. Furthermore, in a meta-analysis by Tusting *et al.*^[143], socioeconomic development strongly correlated with better malaria therapeutic outcomes. Recently, *A. annua* has demonstrated anti HIV activity^[126,144] and thus oral consumption of the dried leaves of this herb will not only treat malaria, but should also enhance the well-being of HIV/AIDS patients.

Agriculture, jobs and self-determination

A. annua is grown in more than 75 countries^[145]. In 2011 about 163 MT of artemisinin were extracted from plantations and small stakeholder farms mainly located in China, Vietnam, and Eastern Africa including Madagascar; value was about \$550/kg^[120]. With the advent of the production of semi synthetic artemisinin by Sanofi, 60 MT were projected for 2014 with an anticipated price of about \$400/kg^[119]. As this new source of artemisinin becomes available, the Netherlands Royal Tropical Institute projected that the market for natural *Artemisia* will signifi-

cantly destabilize, undermining the security of farmers. The Tropical Institute was further concerned that “pharmaceutical companies will accumulate control and power over the production process; *Artemisia* producers will lose a source of income; and local production, extraction and (possibly) manufacturing of ACT in regions where malaria is prevalent will shift to the main production sites of Western pharmaceutical companies”, disrupting the fragile economics of these already impoverished countries^[120]. The average small stakeholder crop area is about 0.2 ha in China and Africa^[120], so while implementation of pACT may not require as much agricultural land as for extracted artemisinin, it could still help provide small stakeholders with a source of income. We have estimated that localized micro manufacturing plants could be constructed for < \$50000 USD, and produce quality-controlled pACT tablets with readily verifiable contents. Our overall approach, schematically illustrated in Figure 3, leads to local control of malaria and possibly other artemisinin susceptible diseases while also improving the socioeconomic status of the populations.

CONCLUSION

Evidence is mounting for the therapeutic efficacy of the use of dried leaves of *A. annua*, pACT, to treat malaria and possibly other diseases. The complex mixture of antiparasitic compounds in the plant seems to account for its therapeutic activity with animal and human trials

supporting this claim. It is also clear that the cost of using pACT is a fraction of that for any other current or emerging antimalarial therapeutic. Likewise, the recent evidence of persistent and/or asymptomatic malaria suggests that a more prophylactic approach to malaria using pACT or even *A. annua* tea may be warranted. Considering that for > 2000 years this plant was used in traditional Chinese medicine for treatment of fever with no apparent appearance of artemisinin drug resistance, taken together the cumulative evidence argues for inclusion of pACT into the arsenal of drugs to combat malaria, and very likely, other diseases.

REFERENCES

- 1 WHO. World Malaria Report 2013. Available from: URL: http://www.who.int/malaria/publications/world_malaria_report_2013/en/
- 2 Phyto AP, Nkhoma S, Stepniewska K, Ashley EA, Nair S, McGready R, Iler Moo C, Al-Saai S, Dondorp AM, Lwin KM, Singhasivanon P, Day NP, White NJ, Anderson TJ, Nosten F. Emergence of artemisinin-resistant malaria on the western border of Thailand: a longitudinal study. *Lancet* 2012; **379**: 1960-1966 [PMID: 22484134 DOI: 10.1016/S0140-6736(12)60484-X]
- 3 Betson M, Sousa-Figueiredo JC, Atuhaire A, Arinaitwe M, Adriko M, Mwesigwa G, Nabonge J, Kabatereine NB, Sutherland CJ, Stothard JR. Detection of persistent Plasmodium spp. infections in Ugandan children after artemether-lumefantrine treatment. *Parasitology* 2014; **16**: 1-11 [PMID: 24837880 DOI: 10.1017/S003118201400033X]
- 4 Beshir KB, Sutherland CJ, Sawa P, Drakeley CJ, Okell L, Mweresa CK, Omar SA, Shekalaghe SA, Kaur H, Ndaro A, Chilongola J, Schallig HD, Sauerwein RW, Hallett RL, Bousema T. Residual Plasmodium falciparum parasitemia in Kenyan children after artemisinin-combination therapy is associated with increased transmission to mosquitoes and parasite recurrence. *J Infect Dis* 2013; **208**: 2017-2024 [PMID: 23945376 DOI: 10.1093/infdis/jit431]
- 5 Lindblade KA, Steinhardt L, Samuels A, Kachur SP, Slutsker L. The silent threat: asymptomatic parasitemia and malaria transmission. *Expert Rev Anti Infect Ther* 2013; **11**: 623-639 [PMID: 23750733 DOI: 10.1586/eri.13.45]
- 6 Yeung S, Van Damme W, Socheat D, White NJ, Mills A. Cost of increasing access to artemisinin combination therapy: the Cambodian experience. *Malar J* 2008; **7**: 84 [PMID: 18492245 DOI: 10.1186/1475-2875-7-84]
- 7 O'Connell KA, Gatakaa H, Poyer S, Njogu J, Evance I, Munroe E, Solomon T, Goodman C, Hanson K, Zinsou C, Akulayi L, Raharinjatovo J, Arogundade E, Buyungo P, Mpasela F, Adjibabi CB, Agbango JA, Ramarosandratana BF, Coker B, Rubahika D, Hamainza B, Chapman S, Shewchuk T, Chavasse D. Got ACTs? Availability, price, market share and provider knowledge of anti-malarial medicines in public and private sector outlets in six malaria-endemic countries. *Malar J* 2011; **10**: 326 [PMID: 22039838 DOI: 10.1186/1475-2875-10-326]
- 8 Davis B, Ladner J, Sams K, Tekinturhan E, de Korte D, Saba J. Artemisinin-based combination therapy availability and use in the private sector of five AMFm phase 1 countries. *Malar J* 2013; **12**: 135 [PMID: 23607504 DOI: 10.1186/1475-2875-12-135]
- 9 Mikkelsen-Lopez I, Shango W, Barrington J, Ziegler R, Smith T, deSavigny D. The challenge to avoid anti-malarial medicine stock-outs in an era of funding partners: the case of Tanzania. *Malar J* 2014; **13**: 181 [PMID: 24885420 DOI: 10.1186/1475-2875-13-181]
- 10 Ferreira JFS, Laughlin JC, Delabays N, de Magalhães PM. Cultivation and genetics of *Artemisia annua* L. for increased production of the antimalarial artemisinin. *Plant Gen Res* 2005; **3**: 206-229 [DOI: 10.1079/PGR200585]
- 11 Duke JA. Handbook of phytochemical constituents of GRAS herbs and other economic plants. Boca Raton, FL: CRC Press LLC; 2001: 70
- 12 Duke MV, Paul RN, Elshohly HN, Sturtz G, Duke SO. Localization of artemisinin and artemisitene in foliar tissues of glanded and glandless biotypes of *Artemisia annua* L. *Int J Plant Sci* 1994; **155**: 365-372
- 13 Ferreira JF, Luthria DL, Sasaki T, Heyerick A. Flavonoids from *Artemisia annua* L. as antioxidants and their potential synergism with artemisinin against malaria and cancer. *Molecules* 2010; **15**: 3135-3170 [PMID: 20657468 DOI: 10.3390/molecules15053135]
- 14 Bhakuni RS, Jain DC, Sharma RP, Kumar S. Secondary metabolites of *Artemisia annua* and their biological activity. *Curr Sci* 2001; **80**: 35-48
- 15 Wang W, Wang Y, Zhang Q, Qi Y, Guo D. Global characterization of *Artemisia annua* glandular trichome transcriptome using 454 pyrosequencing. *BMC Genomics* 2009; **10**: 465 [PMID: 19818120 DOI: 10.1186/1471-2164-10-465]
- 16 Rāth K, Taxis K, Walz G, Gleiter CH, Li SM, Heide L. Pharmacokinetic study of artemisinin after oral intake of a traditional preparation of *Artemisia annua* L. (annual wormwood). *Am J Trop Med Hyg* 2004; **70**: 128-132 [PMID: 14993622]
- 17 Mueller MS, Runyambo N, Wagner I, Borrmann S, Dietz K, Heide L. Randomized controlled trial of a traditional preparation of *Artemisia annua* L. (Annual Wormwood) in the treatment of malaria. *Trans R Soc Trop Med Hyg* 2004; **98**: 318-321 [PMID: 15109558]
- 18 Silva LF, Magalhães PM, Costa MR, Alecrim Md, Chaves FC, Hidalgo Ade F, Pohlit AM, Vieira PP. *In vitro* susceptibility of Plasmodium falciparum Welch field isolates to infusions prepared from *Artemisia annua* L. cultivated in the Brazilian Amazon. *Mem Inst Oswaldo Cruz* 2012; **107**: 859-866 [PMID: 23147140]
- 19 Suberu JO, Gorka AP, Jacobs L, Roepe PD, Sullivan N, Barker GC, Lapkin AA. Anti-plasmodial polyvalent interactions in *Artemisia annua* L. aqueous extract—possible synergistic and resistance mechanisms. *PLoS One* 2013; **8**: e80790 [PMID: 24244716 DOI: 10.1371/journal.pone.0080790]
- 20 ICIPE. Whole-leaf *Artemisia annua*-based antimalarial drug: report on proof-of-concepts studies. 2005. [cited 2014 May 25]. Available from: URL: <http://www.google.com/url?sa=t&rct=j&q=&esrc=s&frm=1&source=web&cd=2&ved=0CDgQFjAB&url=http://www.iwerliwen.org/index.php/component/edocman/?task=document.download&id=96&Itemid=181&ei=J2miUbnFN080QGoi4GACw&usq=AFQjCNHoLJmPt4n0AkKyBIXPSy15W7rc6w&sig2=ppM08X1tZgJQLLiaoJZx1w&bv=bv.47008514,d.dmQ>
- 21 Weathers PJ, Arsenault PR, Covello P, McMickle A, Reed D, Teoh KH. Artemisinin production in *Artemisia annua*: studies in planta and results of a novel delivery method for treating malaria and other neglected diseases. *Phytochem Rev* 2011; **10**: 173-183 [PMID: 21643453 DOI: 10.1007/s11101-010-9166-0]
- 22 Weathers PJ, Elfawal MA, Towler MJ, Acquah-Mensah GK, Rich SM. Pharmacokinetics of artemisinin delivered by oral consumption of *Artemisia annua* dried leaves in healthy vs. Plasmodium chabaudi-infected mice. *J Ethnopharmacol* 2014; **153**: 732-736 [PMID: 24661969 DOI: 10.1016/j.jep.2014.03.037]
- 23 Elfawal MA, Towler MJ, Reich NG, Golenbock D, Weathers PJ, Rich SM. Dried whole plant *Artemisia annua* as an antimalarial therapy. *PLoS One* 2012; **7**: e52746 [PMID: 23289055 DOI: 10.1371/journal.pone.0052746]
- 24 Onimus M, Carteron S, Lutgen P. The surprising efficiency of *Artemisia annua* powder capsules. *Medicin Aromat Plants* 2013; **2**: 3 [DOI: 10.4172/2167-0412.1000125]

- 25 **Hsu E.** The history of qing hao in the Chinese materia medica. *Trans R Soc Trop Med Hyg* 2006; **100**: 505-508 [PMID: 16566952]
- 26 **Weathers PJ**, Reed K, Hassanali A, Lutgen P, Engeu PO. Chapter 4: Whole plant approaches to therapeutic use of *Artemisia annua* L. (Asteraceae). In: Aftab T, Ferreira JFS, Khan MMA, Naeem M, editors. *Artemisia annua - Pharmacology and Biotechnology*. Heidelberg, GDR: Springer, 2014: 51-74
- 27 **van der Kooy F**, Verpoorte R. The content of artemisinin in the *Artemisia annua* tea infusion. *Planta Med* 2011; **77**: 1754-1756 [PMID: 21544776 DOI: 10.1055/s-0030-1271065]
- 28 **Carbonara T**, Pascale R, Argentieri MP, Papadia P, Fanizzi FP, Villanova L, Avato P. Phytochemical analysis of a herbal tea from *Artemisia annua* L. *J Pharm Biomed Anal* 2012; **62**: 79-86 [PMID: 22305080 DOI: 10.1016/j.jpba.2012.01.015]
- 29 **Weathers PJ**, Towler MJ. The flavonoids casticin and artemetin are poorly extracted and are unstable in an *Artemisia annua* tea infusion. *Planta Med* 2012; **78**: 1024-1026 [PMID: 22673829 DOI: 10.1055/s-0032-1314949]
- 30 **Ogwang PE**, Ogwal-Okeng J, Kasasa S, Ejobi F, Kabasa D, Obua C. Use of *Artemisia annua* L. infusion for malaria prevention: mode of action and benefits in a Ugandan community. *British J Pharm Res* 2011; **1**: 124-132 [DOI: 10.9734/BJPR/2011/392]
- 31 **Ogwang PE**, Ogwal JO, Kasasa S, Olila D, Ejobi F, Kabasa D, Obua C. *Artemisia annua* L. infusion consumed once a week reduces risk of multiple episodes of malaria: a randomized trial in a Ugandan community. *Trop J Pharm Res* 2012; **13**: 445-453 [DOI: 10.4314/tjpr.v11i3.14]
- 32 **Mueller MS**, Karhagomba IB, Hirt HM, Wemakor E. The potential of *Artemisia annua* L. as a locally produced remedy for malaria in the tropics: agricultural, chemical and clinical aspects. *J Ethnopharmacol* 2000; **73**: 487-493 [PMID: 11091003]
- 33 **Blanke CH**, Naisabha GB, Balema MB, Mbaruku GM, Heide L, Müller MS. Herba *Artemisiae annuae* tea preparation compared to sulfadoxine-pyrimethamine in the treatment of uncomplicated falciparum malaria in adults: a randomized double-blind clinical trial. *Trop Doct* 2008; **38**: 113-116 [PMID: 18453510 DOI: 10.1258/td.2007.060184]
- 34 **De Donno A**, Grassi T, Idolo A, Guido M, Papadia P, Caccioppola A, Villanova L, Merendino A, Bagordo F, Fanizzi FP. First-time comparison of the *in vitro* antimalarial activity of *Artemisia annua* herbal tea and artemisinin. *Trans R Soc Trop Med Hyg* 2012; **106**: 696-700 [PMID: 22986092 DOI: 10.1016/j.trstmh.2012.07.008]
- 35 **Brown GD.** The biosynthesis of artemisinin (Qinghaosu) and the phytochemistry of *Artemisia annua* L. (Qinghao). *Molecules* 2010; **15**: 7603-7698 [PMID: 21030913 DOI: 10.3390/molecules15117603]
- 36 **van der Kooy F**, Sullivan SE. The complexity of medicinal plants: the traditional *Artemisia annua* formulation, current status and future perspectives. *J Ethnopharmacol* 2013; **150**: 1-13 [PMID: 23973523 DOI: 10.1016/j.jep.2013.08.021]
- 37 **WHO Position Statement.** Effectiveness of Non-Pharmaceutical Forms of *Artemisia annua* L. against malaria. 2012. Available from: URL: http://www.who.int/malaria/position_statement_herbal_remedy_artemisia_annua_l.pdf
- 38 **Tiruneh G**, Kebede Y, Yizgaw T. Use of the plant *Artemisia annua* as a natural anti-malarial herb in Arbaminch town. *Ethiop J Health Biomed Sci* 2010; **2**: 76-82
- 39 **Giao PT**, Binh TQ, Kager PA, Long HP, Van Thang N, Van Nam N, de Vries PJ. Artemisinin for treatment of uncomplicated falciparum malaria: is there a place for monotherapy? *Am J Trop Med Hyg* 2001; **65**: 690-695 [PMID: 11791958]
- 40 **Hien TT.** An overview of the clinical use of artemisinin and its derivatives in the treatment of falciparum malaria in Viet Nam. *Trans R Soc Trop Med Hyg* 1994; **88** Suppl 1: S7-S8 [PMID: 8053033]
- 41 **McIntosh HM**, Olliaro P. Artemisinin derivatives for treating severe malaria. *Cochrane Database Syst Rev* 2000; **2**: CD000527 [PMID: 10796551 DOI: 10.1002/14651858.CD000527]
- 42 **de Vries PJ**, Dien TK. Clinical pharmacology and therapeutic potential of artemisinin and its derivatives in the treatment of malaria. *Drugs* 1996; **52**: 818-836 [PMID: 8957153]
- 43 **Wan YD**, Zang QZ, Wang JS. [Studies on the antimalarial action of gelatin capsule of *Artemisia annua*]. *Zhongguo Jishengchongxue Yu Jishengchongbing Zazhi* 1992; **10**: 290-294 [PMID: 1303339]
- 44 **Zhao KC**, Song ZY. [Pharmacokinetics of dihydroqinghaosu in human volunteers and comparison with qinghaosu]. *Ya-oxue Xuebao* 1993; **28**: 342-346 [PMID: 8237378]
- 45 **Alin MH**, Ashton M, Kihamia CM, Mtey GJ, Björkman A. Clinical efficacy and pharmacokinetics of artemisinin monotherapy and in combination with mefloquine in patients with falciparum malaria. *Br J Clin Pharmacol* 1996; **41**: 587-592 [PMID: 8799526]
- 46 **Ilet KF**, Batty KT. Artemisinin and its derivatives. In: Yu VL, Edwards G, McKinnon PS, Peloquin CA, Morse G, editors. *Antimicrobial Therapy and Vaccines*. Pittsburgh USA: ESun Technologies, 2005: 981-1002
- 47 **Ashton M**, Gordi T, Trinh NH, Nguyen VH, Nguyen DS, Nguyen TN, Dinh XH, Johansson M, Le DC. Artemisinin pharmacokinetics in healthy adults after 250, 500 and 1000 mg single oral doses. *Biopharm Drug Dispos* 1998; **19**: 245-250 [PMID: 9604124]
- 48 **Dien TK**, de Vries PJ, Khanh NX, Koopmans R, Binh LN, Duc DD, Kager PA, van Boxtel CJ. Effect of food intake on pharmacokinetics of oral artemisinin in healthy Vietnamese subjects. *Antimicrob Agents Chemother* 1997; **41**: 1069-1072 [PMID: 9145871]
- 49 **Ashton M**, Hai TN, Sy ND, Huong DX, Van Huong N, Niêu NT, Công LD. Artemisinin pharmacokinetics is time-dependent during repeated oral administration in healthy male adults. *Drug Metab Dispos* 1998; **26**: 25-27 [PMID: 9443848]
- 50 **Svensson US**, Ashton M. Identification of the human cytochrome P450 enzymes involved in the *in vitro* metabolism of artemisinin. *Br J Clin Pharmacol* 1999; **48**: 528-535 [PMID: 10583023]
- 51 **Gueye PEO**, Diallo M, deme AB, Badiane A, Dior DM, Ahouidi A, Abdoul AN, Dieng T, Lutgen P, Mbopup S, Sarr O. Tea *Artemisia annua* inhibits Plasmodium falciparum isolates collected in Pikine, Senegal. *Af J Biochem Res* 2013; **7**: 107-113 [DOI: 10.5897/AJBR12.022]
- 52 **Liu KC**, Yang SL, Roberts MF, Elford BC, Phillipson JD. Antimalarial activity of *Artemisia annua* flavonoids from whole plants and cell cultures. *Plant Cell Rep* 1992; **11**: 637-640 [PMID: 24213368 DOI: 10.1007/BF00236389]
- 53 **Elford BC**, Roberts MF, Phillipson JD, Wilson RJ. Potentiation of the antimalarial activity of qinghaosu by methoxylated flavones. *Trans R Soc Trop Med Hyg* 1987; **81**: 434-436 [PMID: 3318019]
- 54 **Lehane AM**, Saliba KJ. Common dietary flavonoids inhibit the growth of the intraerythrocytic malaria parasite. *BMC Res Notes* 2008; **1**: 26 [PMID: 18710482 DOI: 10.1186/1756-0500-1-26]
- 55 **van Zyl RL**, Seatlholo ST, van Vuuren SF, Viljoen AM. The biological activities of 20 nature identical essential oil constituents. *J Essent Oil Res* 2006; **18**: Special Edition 129-133
- 56 **Melillo de Magalhães P**, Dupont I, Hendrickx A, Joly A, Raas T, Dessy S, Sergent T, Schneider YJ. Anti-inflammatory effect and modulation of cytochrome P450 activities by *Artemisia annua* tea infusions in human intestinal Caco-2 cells. *Food Chem* 2012; **134**: 864-871 [PMID: 23107701 DOI: 10.1016/j.foodchem.2012.02.195]
- 57 **Baraldi R**, Isacchi B, Predieri S, Marconi G, Vincieri FF, Bilia AR. Distribution of artemisinin and bioactive flavonoids from *Artemisia annua* L. during plant growth. *Biochem Syst Ecol* 2008; **36**: 340-348 [DOI: 10.1016/j.bse.2007.11.002]
- 58 **Ganesh D**, Fuehrer HP, Starzengrüber P, Swoboda P, Khan

- WA, Reismann JA, Mueller MS, Chiba P, Noedl H. Antiplasmodial activity of flavonol quercetin and its analogues in *Plasmodium falciparum*: evidence from clinical isolates in Bangladesh and standardized parasite clones. *Parasitol Res* 2012; **110**: 2289-2295 [PMID: 22215188 DOI: 10.1007/s00436-011-2763-z]
- 59 **Manach C**, Donovan JL. Pharmacokinetics and metabolism of dietary flavonoids in humans. *Free Radic Res* 2004; **38**: 771-785 [PMID: 15493450]
- 60 **Tasdemir D**, Lack G, Brun R, Rüedi P, Scapozza L, Perozzo R. Inhibition of *Plasmodium falciparum* fatty acid biosynthesis: evaluation of FabG, FabZ, and FabI as drug targets for flavonoids. *J Med Chem* 2006; **49**: 3345-3353 [PMID: 16722653]
- 61 **Lin CM**, Chen CT, Lee HH, Lin JK. Prevention of cellular ROS damage by isovitexin and related flavonoids. *Planta Med* 2002; **68**: 365-367 [PMID: 11988866]
- 62 **Raguso RA**, Schlumpberger BO, Kaczorowski RL, Holtsford TP. Phylogenetic fragrance patterns in *Nicotiana* sections *Alatae* and *Suaevolentes*. *Phytochemistry* 2006; **67**: 1931-1942 [PMID: 16843507]
- 63 **Rodrigues Goulart H**, Kimura EA, Peres VJ, Couto AS, Aquino Duarte FA, Katzin AM. Terpenes arrest parasite development and inhibit biosynthesis of isoprenoids in *Plasmodium falciparum*. *Antimicrob Agents Chemother* 2004; **48**: 2502-2509 [PMID: 15215101]
- 64 **Moura IC**, Wunderlich G, Uhrig ML, Couto AS, Peres VJ, Katzin AM, Kimura EA. Limonene arrests parasite development and inhibits isoprenylation of proteins in *Plasmodium falciparum*. *Antimicrob Agents Chemother* 2001; **45**: 2553-2558 [PMID: 11502528]
- 65 **Su V**, King D, Woodrow I, McFadden G, Gleadow R. *Plasmodium falciparum* growth is arrested by monoterpenes from eucalyptus oil. *Flavour Frag J* 2008; **23**: 315-318 [DOI: 10.1002/ffj.1880]
- 66 **Miller JA**, Hakim IA, Chew W, Thompson P, Thomson CA, Chow HH. Adipose tissue accumulation of d-limonene with the consumption of a lemonade preparation rich in d-limonene content. *Nutr Cancer* 2010; **62**: 783-788 [PMID: 20661827 DOI: 10.1080/01635581003693066]
- 67 **Charles DJ**, JE Simon, Wood KV, Heinstein P. Germplasm variation in artemisinin content of *Artemisia annua* using an alternative method of artemisinin analysis from crude plant extracts. *J Nat Prod* 1990; **53**: 157-160 [DOI: 10.1021/np50067a021]
- 68 **Juergens UR**, Engelen T, Racké K, Stöber M, Gillissen A, Vetter H. Inhibitory activity of 1,8-cineol (eucalyptol) on cytokine production in cultured human lymphocytes and monocytes. *Pulm Pharmacol Ther* 2004; **17**: 281-287 [PMID: 15477123]
- 69 **Kovar KA**, Gropper B, Friess D, Ammon HP. Blood levels of 1,8-cineole and locomotor activity of mice after inhalation and oral administration of rosemary oil. *Planta Med* 1987; **53**: 315-318 [PMID: 3671550]
- 70 **Stimpfl T**, Nasel B, Binder R, Vycudilik W, Buchbauer G. Concentration of 1,8-cineol in human blood during prolonged inhalation. *Chem Senses* 1995; **20**: 349-350 [PMID: 7552045]
- 71 **Kengne RDC**. Characterisation physico-chimique de *Artemisia annua* (asteraceae), plante medicinale cultivee au Cameroun. MS thesis in Organic Chemistry. Univ de Dschang Republic of Cameroun; 2010
- 72 **Akhtar F**, Rizvi MM, Kar SK. Oral delivery of curcumin bound to chitosan nanoparticles cured *Plasmodium yoelii* infected mice. *Biotechnol Adv* 2012; **30**: 310-320 [PMID: 21619927 DOI: 10.1016/j.biotechadv.2011.05.009]
- 73 **Akkawi M**, Jaber S, Abu-Remeleh Q, Ogwang PE, Lutgen P. Investigations of *Artemisia annua* and *Artemisia sieberi* water extracts inhibitory effects on β -hematin formation. *Medicin Arom Plants* 2014; **3**: 150 [DOI: 10.4172/2167-0412.100150]
- 74 **Liu AR**, Yu ZY, Lu LL, Sui ZY. [The synergistic action of guanghuoxiang volatile oil and sodium artesunate against *Plasmodium berghei* and reversal of SA-resistant *Plasmodium berghei*]. *Zhongguo Jishengchongxue Yu Jishengchongbing Zazhi* 2000; **18**: 76-78 [PMID: 12567719]
- 75 **Yang ZN**, Zhu SQ, Yu ZW. Comparison of terpene components from flowers of *Artemisia annua*. *Bangladesh J Pharmacol* 2012; **7**: 114-119 [DOI: 10.3329/bjp.v7i2.10815]
- 76 **Khangholil S**, Rezaeinodehi A. Effect of drying temperature on essential oil content and composition of sweet wormwood (*Artemisia annua*) growing wild in Iran. *Pak J Biol Sci* 2008; **11**: 934-937 [PMID: 18814660]
- 77 **Weathers PJ**, Towler MJ. Changes in key constituents of clonally propagated *Artemisia annua* L. during preparation of compressed leaf tablets for possible therapeutic use. *Ind Crop Prod* 2014; In press [DOI: 10.1016/j.indcrop.2014.08.033]
- 78 **Juteau F**, Masotti V, Bessière JM, Dherbomez M, Viano J. Antibacterial and antioxidant activities of *Artemisia annua* essential oil. *Fitoterapia* 2002; **73**: 532-535 [PMID: 12385883]
- 79 **Roberts DW**, Rank RG, Weidanz WP, Finerty JF. Prevention of recrudescence malaria in nude mice by thymic grafting or by treatment with hyperimmune serum. *Infect Immun* 1977; **16**: 821-826 [PMID: 330396]
- 80 **Cherneva E**, Pavlovic V, Smelcerovic A, Yancheva D. The effect of camphor and borneol on rat thymocyte viability and oxidative stress. *Molecules* 2012; **17**: 10258-10266 [PMID: 22926306 DOI: 10.3390/molecules170910258]
- 81 **Lopes NP**, Kato MJ, Andrade EH, Maia JG, Yoshida M, Planchart AR, Katzin AM. Antimalarial use of volatile oil from leaves of *Virola surinamensis* (Rol.) Warb. by Waiãpi Amazon Indians. *J Ethnopharmacol* 1999; **67**: 313-319 [PMID: 10617066]
- 82 **Muzemil A**. Determination of artemisinin and essential oil contents of *Artemisia annua* L. grown in Ethiopia and in vivo antimalarial activity of its crude extracts against *Plasmodium berghei* in mice. Ethiopia: MS Thesis in Medicinal Chemistry, Addis Ababa University, 2008
- 83 **Li Y**, Hu HB, Zheng XD, Zhu JH, Liu LP. Composition and antimicrobial activity of essential oil from the aerial part of *Artemisia annua*. *J Medicin Plants Res* 2011; **5**: 3629-3633
- 84 **Ahn BY**, Jung MY. Antioxidant and protective activity of polysaccharide extract from *Artemisia iwamomogi* Kitamura stems on UVB-damaged mouse epidermis. *J Appl Biol Chem* 2011; **54**: 184-189 [DOI: 10.3839/jabc.2011.031]
- 85 **Han H**. Antimalarial Activity of Ginseng Polysaccharides and Bulgaria Inquinans Polysaccharides. MS Thesis in Biochem & Molec Biol. Changchun PRC: Northeast Normal Univ., 2008
- 86 **Andrews KT**, Klatt N, Adams Y, Mischnick P, Schwartz-Albiez R. Inhibition of chondroitin-4-sulfate-specific adhesion of *Plasmodium falciparum*-infected erythrocytes by sulfated polysaccharides. *Infect Immun* 2005; **73**: 4288-4294 [PMID: 15972521]
- 87 **Xiao L**, Yang C, Patterson PS, Udhayakumar V, Lal AA. Sulfated polyanions inhibit invasion of erythrocytes by plasmodial merozoites and cytoadherence of endothelial cells to parasitized erythrocytes. *Infect Immun* 1996; **64**: 1373-1378 [PMID: 8606103]
- 88 **Clark DL**, Su S, Davidson EA. Saccharide anions as inhibitors of the malaria parasite. *Glycoconj J* 1997; **14**: 473-479 [PMID: 9249145]
- 89 **Munir M**, Tjandra H, Rampengan TH, Mustadjab I, Wulur FH. Heparin in the treatment of cerebral malaria. *Paediatr Indones* 1980; **20**: 47-50 [PMID: 6988763]
- 90 **Rampengan TH**. Cerebral malaria in children. Comparative study between heparin, dexamethasone and placebo. *Paediatr Indones* 1991; **31**: 59-66 [PMID: 1852471]
- 91 **Adams Y**, Freeman C, Schwartz-Albiez R, Ferro V, Parish CR, Andrews KT. Inhibition of *Plasmodium falciparum*

- growth in vitro and adhesion to chondroitin-4-sulfate by the heparan sulfate mimetic PI-88 and other sulfated oligosaccharides. *Antimicrob Agents Chemother* 2006; **50**: 2850-2852 [PMID: 16870784]
- 92 **Ashok PK**, Upadhyaya K. Preliminary phytochemical screening and physico-chemical parameters of *Artemisia absinthium* and *Artemisia annua*. *J Pharmacog Phytochem* 2013; **1**: 229-235
- 93 **Massiha A**, Khoshkholgh-Pahlaviani MM, Issazadeh K, Bidarigh S, Zarrabi S. Antibacterial activity of essential oils and plant extracts of *Artemisia* (*Artemisia annua* L.) in vitro. *Zahedan J Res Med Sci* 2013; **15**: 14-18
- 94 **Podolak I**, Galanty A, Sobolewska D. Saponins as cytotoxic agents: a review. *Phytochem Rev* 2010; **9**: 425-474 [PMID: 20835386]
- 95 **Sun HX**, Xie Y, Ye YP. Advances in saponin-based adjuvants. *Vaccine* 2009; **27**: 1787-1796 [PMID: 19208455 DOI: 10.1016/j.vaccine.2009.01.091]
- 96 **Haruna M**, Tanaka M, Sugimoto T, Kojima R, Suzuki Y, Konoshima T, Kozuka M, Ito K. Alteration of Na permeability in human erythrocytes as studied by ²³Na-NMR and inhibition of the Na₂K-ATPase activities with saponins: Interactions of Gleditsia saponins with human erythrocyte membranes. *Bioorg Med Chem Lett* 1995; **5**: 827-830 [DOI: 10.1016/0960-894X(95)00121-9]
- 97 **Francis G**, Kerem Z, Makkar HP, Becker K. The biological action of saponins in animal systems: a review. *Br J Nutr* 2002; **88**: 587-605 [PMID: 12493081]
- 98 **Meotti FC**, Ardenghi JV, Pretto JB, Souza MM, d' Avila Moura J, Junior AC, Soldi C, Pizzolatti MG, Santos AR. Antinociceptive properties of coumarins, steroid and dihydrosteryl-2-pyrone from *Polygala sabulosa* (Polygalaceae) in mice. *J Pharm Pharmacol* 2006; **58**: 107-112 [PMID: 16393470]
- 99 **Chang TN**, Deng JS, Chang YC, Lee CY, Jung-Chun L, Lee MM, Peng WH, Huang SS, Huang GJ. Ameliorative Effects of Scopoletin from *Crossostephium chinensis* against Inflammation Pain and Its Mechanisms in Mice. *Evid Based Complement Alternat Med* 2012; **2012**: 595603 [PMID: 22991572 DOI: 10.1155/2012/595603]
- 100 **Malik A**, Kushnoor A, Saini V, Singhal IS, Kumar S, Yadav YC. In vitro antioxidant properties of scopoletin. *J Chem Pharm Res* 2011; **3**: 659-665
- 101 **Moon PD**, Lee BH, Jeong HJ, An HJ, Park SJ, Kim HR, Ko SG, Um JY, Hong SH, Kim HM. Use of scopoletin to inhibit the production of inflammatory cytokines through inhibition of the I κ B/NF- κ B signal cascade in the human mast cell line HMC-1. *Eur J Pharmacol* 2007; **555**: 218-225 [PMID: 17113069]
- 102 **Cherng JM**, Chiang W, Chiang LC. Immunomodulatory activities of common vegetables and spices of Umbelliferae and its related coumarins and flavonoids. *Food Chem* 2008; **106**: 944-950 [DOI: 10.1016/j.foodchem.2007.07.005]
- 103 **Surono IS**, Nishigaki T, Endaryanto A, Waspodo P. Indonesian biodiversities, from microbes to herbal plants as potential functional foods. *J Fac Agric Shinshu U* 2008; **44**: 23-27
- 104 **Ezeokonkwo CA**, Obidoa O. Effect of scopoletin on erythrocyte membrane ion motive ATPases. *Nigerian J Nat Prod Med* 2001; **5**: 37-40 [DOI: 10.4314/njnp.v5i1.11721]
- 105 **Dunn MJ**. Alterations of red blood cell sodium transport during malarial infection. *J Clin Invest* 1969; **48**: 674-684 [PMID: 4975361]
- 106 **Clark IA**, Alleva LM, Mills AC, Cowden WB. Pathogenesis of malaria and clinically similar conditions. *Clin Microbiol Rev* 2004; **17**: 509-539, table of contents [PMID: 15258091]
- 107 **Ding Z**, Dai Y, Wang Z. Hypouricemic action of scopoletin arising from xanthine oxidase inhibition and uricosuric activity. *Planta Med* 2005; **71**: 183-185 [PMID: 15729630]
- 108 **de Vries PJ**, de Vries PJ, Nguyen GC, de Goeje P, de Goeje P. Production and Application of Artemisinin in Vietnam. Institute of Materia Medica, Viet Nam and the University of Amsterdam: Gioi Publishers, 1999
- 109 **Atkinson B**, Mavituna F. *Biochemical Engineering and Biotechnology Handbook*. 2nd ed. USA: Stockton Press, 1991: 1059-1110
- 110 **Lim JAC**, Patkar A, McDonagh G, Sinclair A, Lucy P. Modeling bioprocess cost: the economic benefits of expression technology based on *Pseudomonas fluorescens*. *Bioprocess Intnl* 2010; **8**: 62-70
- 111 **Lapkin AA**, Plucinski PK, Cutler M. Comparative assessment of technologies for extraction of artemisinin. *J Nat Prod* 2006; **69**: 1653-1664 [PMID: 17125242]
- 112 **Griffee P**, Diemer P. *Artemisia annua*; the plant, production, processing and medicinal applications. Food and Agriculture Organization of the United Nations. Available from: URL: <http://ecoport.org/ep?SearchType=earticleView&earticleId=727&page=2#section5675>
- 113 **Sipler D**, Weathers PJ. *Artemisia annua* as a high value crop and weed control. Available from: URL: <http://small-farm.org/SARE/FNE12-766.html>
- 114 RBM Active pharmaceutical ingredient requirements for the manufacture of ACTs. Available from: URL: http://www.rollbackmalaria.org/partnership/wg/wg_procurementsupply/docs/psmwg_ppACT-API.pdf
- 115 **Weathers PJ**, Jordan NJ, Lasin P, Towler MJ. Simulated digestion of dried leaves of *Artemisia annua* consumed as a treatment (pACT) for malaria. *J Ethnopharmacol* 2014; **151**: 858-863 [PMID: 24316176 DOI: 10.1016/j.jep.2013.11.043]
- 116 **Zhu C**, Cook SP. A concise synthesis of (+)-artemisinin. *J Am Chem Soc* 2012; **134**: 13577-13579 [PMID: 22866604 DOI: 10.1021/ja3061479]
- 117 **Paddon CJ**, Westfall PJ, Pitera DJ, Benjamin K, Fisher K, McPhee D, Leavell MD, Tai A, Main A, Eng D, Polichuk DR, Teoh KH, Reed DW, Treynor T, Lenihan J, Fleck M, Bajad S, Dang G, Dengrove D, Diola D, Dorin G, Ellens KW, Fickes S, Galazzo J, Gaucher SP, Geistlinger T, Henry R, Hepp M, Horning T, Iqbal T, Jiang H, Kizer L, Lieu B, Melis D, Moss N, Regentin R, Secret S, Tsuruta H, Vazquez R, Westblade LF, Xu L, Yu M, Zhang Y, Zhao L, Lievense J, Covello PS, Keasling JD, Reiling KK, Renninger NS, Newman JD. High-level semi-synthetic production of the potent antimalarial artemisinin. *Nature* 2013; **496**: 528-532 [PMID: 23575629 DOI: 10.1038/nature12051]
- 118 **Charman SA**, Arbe-Barnes S, Bathurst IC, Brun R, Campbell M, Charman WN, Chiu FC, Chollet J, Craft JC, Creek DJ, Dong Y, Matile H, Maurer M, Morizzi J, Nguyen T, Papastogiannidis P, Scheurer C, Shackelford DM, Sriraghavan K, Stingelin L, Tang Y, Urwyler H, Wang X, White KL, Wittlin S, Zhou L, Vennerstrom JL. Synthetic ozonide drug candidate OZ439 offers new hope for a single-dose cure of uncomplicated malaria. *Proc Natl Acad Sci USA* 2011; **108**: 4400-4405 [PMID: 21300861 DOI: 10.1073/pnas.1015762108]
- 119 **A2s2**. [cited 2014 May 25]. Available from: URL: <http://www.a2s2.org>
- 120 **ETC Group**. Synthetic Biology: Livelihoods and Biodiversity. Artemisinin. Cited: 2014-05-25. Available from: URL: http://www.etcgroup.org/files/CBD_Artemisinin_case_study_TA.pdf
- 121 **Kentucky Cooperative Extension Service**. GAP Good Agricultural Practices (GAP). Cited: 2014-05-25. Available from: URL: <http://www.uky.edu/Ag/CCD/introsheets/gap.pdf>
- 122 **Pewtrusts**. GAP Comparison of gaps governing the growing and harvesting of fresh produce. Cited: 2014-05-27. Available from: URL: http://www.pewhealth.org/uploadedFiles/PHG/Content_Level_Pages/Reports/PSP-RPT-GAP-Governing-Fresh-Produce.pdf
- 123 **World Health Organization**. WHO monograph on good agricultural and collection practices (GACP) for *Artemisia annua* L. Available from: URL: <http://www.who.int/malaria/publications/atoz/9241594438/en/>

- 124 **World Health Organization.** WHO guidelines on good agricultural and collection practices (GACP) for medicinal plants. Available from: URL: <http://whqlibdoc.who.int/publications/2003/9241546271.pdf>
- 125 **World Health Organization.** WHO guidelines for assessing quality of herbal medicines with reference to contaminants and residues. Available from: URL: <http://apps.who.int/medicinedocs/documents/s14878e/s14878e.pdf>
- 126 **Marchand E,** Atemnkeng MA, Vanermen S, Plaizier-Vercammen J. Development and validation of a simple thin layer chromatographic method for the analysis of artemisinin in *Artemisia annua* L. plant extracts. *Biomed Chromatogr* 2008; **22**: 454-459 [PMID: 18059047]
- 127 **Koobkokkrud T,** Chochai A, Kerdmanee C, De-Eknamkul W. TLC-densitometric analysis of artemisinin for the rapid screening of high-producing plantlets of *Artemisia annua* L. *Phytochem Anal* 2007; **18**: 229-234 [PMID: 17500366]
- 128 **Arvouet-Grand A,** Vennat B, Pourrat A, Legret P. [Standardization of propolis extract and identification of principal constituents]. *J Pharm Belg* 1994; **49**: 462-468 [PMID: 7884635]
- 129 **Efferth T,** Romero MR, Wolf DG, Stamminger T, Marin JJ, Marschall M. The antiviral activities of artemisinin and artesunate. *Clin Infect Dis* 2008; **47**: 804-811 [PMID: 18699744 DOI: 10.1086/591195]
- 130 **Efferth T.** Artemisinin: a versatile weapon from traditional Chinese medicine. In: Ramawat KG. Herbal drugs: ethnomedicine to modern medicine. Heidelberg: Springer Verlag, 2009: 179-194
- 131 **Efferth T,** Herrmann F, Tahrani A, Wink M. Cytotoxic activity of secondary metabolites derived from *Artemisia annua* L. towards cancer cells in comparison to its designated active constituent artemisinin. *Phytomedicine* 2011; **18**: 959-969 [PMID: 21831619 DOI: 10.1016/j.phymed.2011.06.008]
- 132 **Firestone GL,** Sundar SN. Anticancer activities of artemisinin and its bioactive derivatives. *Expert Rev Mol Med* 2009; **11**: e32 [PMID: 19883518 DOI: 10.1017/S1462399409001239]
- 133 **Brisibe EA,** Umoren UE, Brisibe F, Magalhães PM, Ferreira JFS, Luthria D, Wu X, Prior RL. Nutritional characterization and antioxidant capacity of different tissues of *Artemisia annua* L. *Food Chem* 2009; **115**: 1240-1246 [DOI: 10.1016/j.foodchem.2009.01.033]
- 134 **Utzinger J,** Xiao S, Keiser J, Chen M, Zheng J, Tanner M. Current progress in the development and use of artemether for chemoprophylaxis of major human schistosome parasites. *Curr Med Chem* 2001; **8**: 1841-1860 [PMID: 11772354 DOI: 10.2174/0929867013371581]
- 135 **Avery MA,** Muraleedharan KM, Desai PV, Bandyopadhyaya AK, Furtado MM, Tekwani BL. Structure-activity relationships of the antimalarial agent artemisinin. 8. design, synthesis, and CoMFA studies toward the development of artemisinin-based drugs against leishmaniasis and malaria. *J Med Chem* 2003; **46**: 4244-4258 [PMID: 13678403]
- 136 **Sen R,** Bandyopadhyay S, Dutta A, Mandal G, Ganguly S, Saha P, Chatterjee M. Artemisinin triggers induction of cell-cycle arrest and apoptosis in *Leishmania donovani* promastigotes. *J Med Microbiol* 2007; **56**: 1213-1218 [PMID: 17761485 DOI: 10.1099/jmm.0.47364-0]
- 137 **Mishina YV,** Krishna S, Haynes RK, Meade JC. Artemisinins inhibit *Trypanosoma cruzi* and *Trypanosoma brucei* rhodesiense in vitro growth. *Antimicrob Agents Chemother* 2007; **51**: 1852-1854 [PMID: 17339374 DOI: 10.1128/AAC.01544-06]
- 138 **Ferreira JF,** Peadar P, Keiser J. In vitro trematocidal effects of crude alcoholic extracts of *Artemisia annua*, *A. absinthium*, *Asimina triloba*, and *Fumaria officinalis*: trematocidal plant alcoholic extracts. *Parasitol Res* 2011; **109**: 1585-1592 [PMID: 21562762 DOI: 10.1007/s00436-011-2418-0]
- 139 **World Health Organization.** The 17 neglected tropical diseases. Available from: URL: http://www.who.int/neglected_diseases/diseases/en/
- 140 **Marconi VC.** Commentary: Malaria and HIV transmission: old meets new in a deadly partnership or an opportunity for healthcare synergism? *Int J Epidemiol* 2011; **40**: 940-944 [PMID: 21393253 DOI: 10.1093/ije/dyr038]
- 141 **Kamya MR,** Gasasira AF, Yeka A, Bakyaite N, Nsohya SL, Francis D, Rosenthal PJ, Dorsey G, Havlir D. Effect of HIV-1 infection on antimalarial treatment outcomes in Uganda: a population-based study. *J Infect Dis* 2006; **193**: 9-15 [PMID: 16323126]
- 142 **Ezeamama AE,** Spiegelman D, Hertzmark E, Bosch RJ, Manji KP, Duggan C, Kupka R, Lo MW, Okuma JO, Kisenge R, Aboud S, Fawzi WW. HIV infection and the incidence of malaria among HIV-exposed children from Tanzania. *J Infect Dis* 2012; **205**: 1486-1494 [PMID: 22457274 DOI: 10.1093/infdis/jis234]
- 143 **Tusting LS,** Willey B, Lucas H, Thompson J, Kafy HT, Smith R, Lindsay SW. Socioeconomic development as an intervention against malaria: a systematic review and meta-analysis. *Lancet* 2013; **382**: 963-972 [PMID: 23790353 DOI: 10.1016/S0140-6736(13)60851-X]
- 144 **Lubbe A,** Seibert I, Klimkait T, van der Kooy F. Ethnopharmacology in overdrive: the remarkable anti-HIV activity of *Artemisia annua*. *J Ethnopharmacol* 2012; **141**: 854-859 [PMID: 22465592 DOI: 10.1016/j.jep.2012.03.024]
- 145 **Willcox ML,** Burton S, Oyweka R, Namyalo R, Challand S, Lindsey K. Evaluation and pharmacovigilance of projects promoting cultivation and local use of *Artemisia annua* for malaria. *Malar J* 2011; **10**: 84 [PMID: 21481234 DOI: 10.1186/1475-2875-10-84]

P- Reviewer: Masocha W S- Editor: Ji FF L- Editor: A
E- Editor: Lu YJ



Asthma in pregnancy

Hayley K Blackburn, Douglas R Allington, Kendra A Procacci, Michael P Rivey

Hayley K Blackburn, Douglas R Allington, Kendra A Procacci, Michael P Rivey, Department of Pharmacy Practice, University of Montana Skaggs School of Pharmacy, Missoula, MT 59812, United States

Hayley K Blackburn, Douglas R Allington, Michael P Rivey, Department of Pharmacy, Community Medical Center, Missoula, MT 59804, United States

Kendra A Procacci, Department of Pharmacy, Grant Creek Family Practice, Missoula, MT 59804, United States

Author contributions: Blackburn HK, Allington DR, Procacci KA and Rivey MP were all involved in the conception and writing of the manuscript.

Correspondence to: Hayley K Blackburn, PhD, Department of Pharmacy Practice, University of Montana Skaggs School of Pharmacy, 32 Campus Dr, Missoula, MT 59812, United States. hblackburn@communitymed.org

Telephone: +1-406-2434624 Fax: +1-706-6536645

Received: July 1, 2014 Revised: August 1, 2014

Accepted: September 16, 2014

Published online: December 9, 2014

Abstract

Asthma affects approximately 8% of women during pregnancy. Pregnancy results in a variable course for asthma control, likely contributed to by physiological changes affecting the respiratory, immune, and hormonal systems. While asthma during pregnancy has been associated with an increased risk of maternal and fetal complications including malformations, available data also suggest that active asthma management and monitoring can decrease the risk of adverse outcomes. The diagnosis, disease classification, and goals for asthma management in the pregnant woman are the same as for nonpregnant patients. However, evidence shows that pregnant asthmatics are more likely to be undertreated, resulting in asthma exacerbations occurring in approximately one third and hospitalization in one tenth of patients. Pharmacotherapeutic management of asthma exacerbations in pregnant patients follows standard treatment guidelines. In contrast, the principles of asthma maintenance therapy are slightly modified in the pregnant patient. Patients and practitioners may

avoid use of asthma medications due to concern for a risk of fetal complications and malformations. A variable amount of information is available regarding the risk of a given asthma medication to cause adverse fetal outcomes, and it is preferable to use an inhaled product. Nevertheless, based on available data, the majority of asthma medications are regarded as safe for use during pregnancy. And, any increased risk to either the mother or fetus from medication use appears to be small compared to that associated with poor asthma control.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Asthma; Pregnancy; Fetal outcomes; Maternal outcomes; Management of asthma; Pharmacotherapy

Core tip: This comprehensive review of the impact of asthma during pregnancy provides information regarding proposed pathophysiological alterations and fetal and maternal outcomes associated with asthma during pregnancy. In addition, we outline the treatment of acute exacerbations and the maintenance management of asthma throughout pregnancy, including specific information on the various classes of medication used to treat asthma.

Blackburn HK, Allington DR, Procacci KA, Rivey MP. Asthma in pregnancy. *World J Pharmacol* 2014; 3(4): 56-71 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/56.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.56>

INTRODUCTION

Asthma is a common condition affecting approximately 8% of pregnant women^[1]. Epidemiological evidence demonstrates that the course of asthma during pregnancy is variable and unpredictable, with approximately one-third of women experiencing an improvement, one-third experiencing a worsening, and one-third having no

changes in asthma symptoms^[2,3]. It is also apparent that poor control of maternal asthma leads to increased risk of adverse maternal and fetal outcomes. Because of these risks and the unpredictable course of asthma symptoms, it is especially important to provide appropriate monitoring and management of the asthmatic patient throughout pregnancy. This review will discuss the physiologic changes associated with asthma during pregnancy, asthma control and its effect on pregnancy outcomes, and the management of asthma in pregnancy.

PHYSIOLOGICAL CHANGES AND ASTHMA IN PREGNANCY

The relationship between the changes in body physiology as pregnancy progresses and the physiological processes driving asthma symptoms is not well understood, but it is evident that the relationship is bidirectional and complex^[3-5]. It is thought that changes including alterations in pulmonary physiology, maternal immune function and hormonal balance contribute to the unpredictable course of asthma during pregnancy.

Pulmonary changes

As pregnancy progresses, the uterus expands and causes elevation of the diaphragm by 4-5 centimeters, resulting in a decrease in lung functional residual capacity (FRC) of 10%-25%. However, the decrease in FRC does not typically result in significant changes to forced vital capacity, peak expiratory flow rate, or forced expiratory volume in 1 second (FEV1). Minute ventilation (VE) may be elevated as much as 50% by the third trimester of pregnancy as a result of progesterone-driven increases in tidal volume and respiratory rate^[6]. Concomitantly, oxygen consumption can increase up to 35%^[7]. Respiratory alkalosis occurs as a result of the increase in VE but is compensated for by increased renal excretion of bicarbonate. Typical arterial blood gas values in pregnancy are altered only slightly from the nonpregnant state, with a normal pH of 7.40-7.45 and pCO₂ of 28-32 mmHg^[7-9].

Due to the pulmonary changes during pregnancy, dyspnea is common and manifests as shortness of breath with rest or mild exertion. The pulmonary changes are often magnified in an asthmatic patient, and may contribute to the perception of changing symptoms during pregnancy^[6].

Immunologic changes

Physiological immunosuppression is characteristic of pregnancy and results in fetomaternal tolerance required for completion of a normal gestation^[10]. Changes in immune characteristics during pregnancy include a shift in the helper T cell (Th1)/Th2 ratio toward a Th2-predominant immune state and an increase in regulatory T cells (Tregs) that work to suppress activation of effector T cells and natural killer cells. This immune deviation is thought to prevent Th1-induced fetal rejection

as paternally originated antigens are expressed during development^[10]. A number of immune-mediated disease states can be affected by this Th2-predominant shift during pregnancy. For example, rheumatoid arthritis is a Th1-mediated disease that goes into remission during pregnancy in the majority of patients^[11]. Asthma, on the other hand, has traditionally been categorized as a Th2-predominant disease state, with allergic Th2-type inflammation leading to airway hyperresponsiveness in patients. Evidence suggests that the pregnancy-associated Th2 immunological shift leads to worsening of the Th2-driven manifestations of asthma^[10,12,13].

Immune changes in pregnant asthmatic women have not been well elucidated but recent studies have helped to better characterize the interplay of immunologic processes associated with pregnancy and asthma. Results of several studies provide evidence that exaggerated Th2 responses in pregnant women with uncontrolled asthma contribute to worsening of maternal symptoms, as well as low birth weight in neonates^[14,15]. In contrast, no differences in the Th1/Th2 ratio were observed between healthy pregnant women and pregnant women with well-controlled asthma, suggesting that pregnancy and asthma do not have additive effects in terms of Th2 prevalence if asthma is well-controlled with medication therapy^[15,16].

The relative number of peripheral Treg cells has been found to be lower in asthmatic compared to healthy pregnant women. It is thought that a decrease in Treg cells results in decreased suppression of the effects of pro-inflammatory Th17 cells and may contribute both to worsening of symptoms as well as increased likelihood of poor fetal outcomes in asthmatic patients^[16]. Increased numbers of pro-inflammatory Th17 cells, have been observed in pregnant asthmatic women, and are hypothesized to contribute to impaired intrauterine growth (Figure 1)^[3]. Continued research is needed to further characterize the complex and bidirectional relationships between T-cell subpopulations and the immunologic processes of asthma, in order to understand their role and importance in asthma during pregnancy.

Another factor associated with immunological changes of pregnancy relates to women's changing susceptibility to respiratory pathogens. The pregnancy-associated decrease in cell-mediated immunity is also known to make pregnant women more susceptible to viral respiratory infections, a common precipitating factor in asthma exacerbations during pregnancy^[17,18].

Hormonal changes

As a pregnancy progresses, concentrations of circulating maternal hormones increase to varying degrees. As such, inter-individual variations in hormonal changes could contribute to the unpredictable course seen in maternal asthma. Pregnancy is associated with an increase in serum free cortisol, a hormone with endogenous anti-inflammatory activity which can improve asthma^[19]. Evidence also suggests that levels of sex steroids including estrogen and progesterone can affect asthma symptoms. Changes

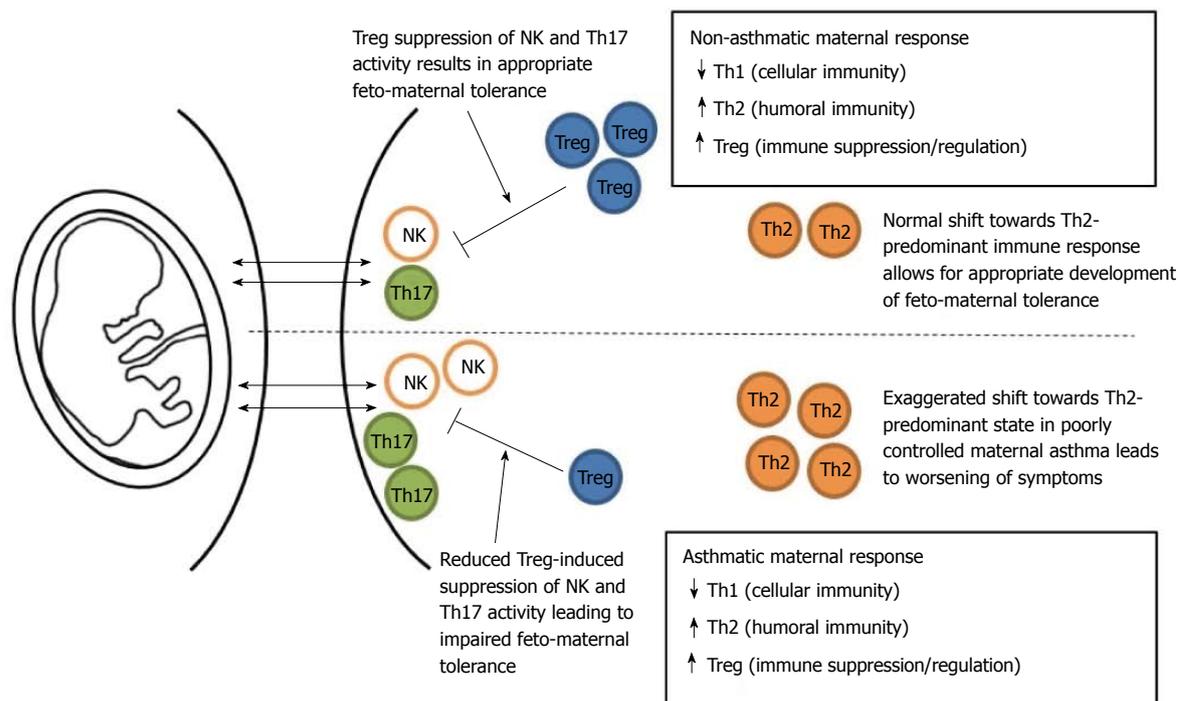


Figure 1 Immunology of Asthma in pregnancy. NK: Natural killer; Treg: Regulatory T cells.

in asthma symptoms are known to occur throughout the menstrual cycle, with up to 40% of females experiencing premenstrual asthma worsening during the follicular phase when progesterone and estrogen levels are normally low^[20-23].

Estrogen levels have been shown to be correlated with the quantity of peripheral Tregs, suggesting an interrelationship between the hormonal and immunological effects of pregnancy. Arruvito *et al*^[24] observed a strong correlation between increasing estradiol levels in fertile women and the percentage of Tregs found in the total number of CD4⁺ T lymphocytes. An elevation of estrogen during the third trimester has been associated with increased bronchial mucus production and airway edema, thereby increasing symptoms of asthma^[6,25]. Increased progesterone levels during pregnancy can result in a multitude of effects as a potent smooth muscle relaxant. While smooth muscle relaxation in the lungs would likely improve asthma symptoms, relaxation of the smooth muscle controlling the esophageal sphincter could result in increased gastroesophageal reflux, a condition known to exacerbate the symptoms of asthma^[26].

Additionally, serum progesterone levels have been shown to exert effects on the regulation of β 2-adrenoreceptors in female asthmatics, which could in turn effect changes in asthma symptoms and medication response. In a study of seven asthmatic females receiving exogenous progesterone during the follicular phase of their menstrual cycle, a significant decrease in lymphocyte β 2-adrenoreceptors was determined when compared to baseline, with a trend towards decreased responsiveness to exogenously administered isoproterenol. Results of this study suggest that downregulation of β 2-adrenoreceptors in response to increases in serum

progesterone may result in loss of responsiveness to both endogenous catecholamines and exogenously administered beta-agonists^[27].

Available evidence also suggests there may be inherent differences in the course of asthma during pregnancy based on fetal sex, with a worsening of maternal asthma more likely in the presence of a female fetus^[28-31]. In a study of pregnant women with asthma, there was a significantly increased dose requirement of inhaled corticosteroid (ICS) and a significant rise in circulating monocytes that progressed throughout the pregnancy in women with a female compared to those carrying a male fetus^[29]. It has been proposed that factors such as differences in hormone or protein production between male and female fetuses account for the variability in maternal asthma severity and treatment response, although currently there is no specific evidence to support this hypothesis.

ASTHMA CONTROL AND PREGNANCY OUTCOMES

Maternal outcomes

Compared to pregnant women without asthma, asthma during pregnancy is consistently associated with higher rates of preeclampsia, pregnancy-induced hypertension, transient hypertension of pregnancy, gestational diabetes, placenta previa or placental abruption, premature labor or delivery, cesarean section and postpartum hemorrhage^[32-37]. Results from a recent retrospective cohort study conducted in the United States from 2002-2008 of obstetric complications among women with asthma ($n = 17044$) compared to women without asthma ($n =$

206468) found asthma was associated with increased rates of preeclampsia [adjusted odds ratio (aOR) = 1.14; 95%CI: 1.06-1.22], superimposed hypertension (aOR 1.34; 95%CI: 1.15-1.56), placenta previa (aOR = 1.30; 95%CI: 1.08-1.56), placental abruption (aOR = 1.22; 95%CI: 1.09-1.36), preterm premature rupture of membranes (aOR = 1.18; 95%CI: 1.07-1.30), gestational diabetes (aOR = 1.11; 95%CI: 1.03-1.19), maternal hemorrhage (aOR = 1.09; 95%CI: 1.03-1.16), breech presentation (aOR = 1.13; 95%CI: 1.05-1.22), prelabor cesarean delivery (aOR = 1.16; 95%CI: 1.09-1.23) and maternal intensive care unit admission (aOR = 1.34; 95%CI: 1.04-1.72)^[38]. Blais *et al.*^[39] reported women with asthma have a higher rate of spontaneous abortion (OR = 1.41; 95%CI: 1.33-1.49) but a lower rate of induced abortion (OR = 0.92; 0.88-0.97); neither finding was influenced by baseline asthma severity. Actively managing asthma during pregnancy has resulted in reducing risks of certain maternal outcomes to non-significant levels such as preterm delivery and low birth weight but without effect on pre-eclampsia^[40].

Fetal outcomes

Fetal complications associated with maternal asthma include premature delivery (< 37 wk), small for gestational age (SGA, defined as < 10% percentile for matched age), low birth weight (LBW, defined as < 2500 gm), intrauterine growth restriction, mortality, and congenital malformations. Asthma during pregnancy has been associated with increased rates of neonatal sepsis and hospitalization in some studies but results are inconsistent^[33,41,42]. Clifton *et al.*^[30] completed a meta-analysis of literature describing perinatal outcomes in women with asthma. The meta-analysis included data from 11 prospective and 15 retrospective cohort studies with total subject populations exceeding 1500000. Subgroup analysis was conducted to compare outcomes of prospective *vs* retrospective studies, as well as outcomes of studies with active asthma management as compared to those without active asthma management. Maternal asthma without active management was associated with increased rates of preterm delivery [risk ratio (RR) = 1.41; 95%CI: 1.22-1.61], pre-eclampsia (RR = 1.54; 95%CI: 1.32-1.81), LBW (RR = 1.46; 95%CI: 1.22-1.75), and SGA (RR = 1.22; 95%CI: 1.14-1.31). Studies in the meta-analysis that included active asthma management reduced the relative risk of preterm labor and delivery to non-significant levels^[40].

Maternal asthma is associated with increased risk of fetal malformations but the magnitude of the risk appears to be only slightly greater than risk associated in non-asthmatic pregnant women. Tata *et al.*^[43] performed a case-controlled study using 5124 fetal cases of major congenital malformations compared with over 30000 matched controls. Results indicated the malformation risk due to maternal asthma was marginally greater (aOR = 1.10; 95%CI: 1.01 to 1.20) than that in controls and that the risk could be modified by asthma therapy in the year before and during pregnancy^[44]. Additionally, a meta-

analysis and systematic review of maternal asthma and its influence on fetal outcomes was recently conducted by Murphy and colleagues. When compared to control groups consisting of pregnant women without asthma, maternal asthma was associated with a significantly increased RR for neonatal sepsis (RR = 2.27; 95%CI: 1.12-4.58), hospitalization (RR = 1.50; 95%CI: 1.03-2.20) and perinatal mortality (RR = 1.25; 95%CI: 1.05-1.50). Investigators also found an overall increased risk of congenital malformations (RR = 1.11; 95%CI: 1.02-1.21) but the increased risk was only significant in the subcategory of retrospective cohort studies that did not include active asthma management^[44].

MANAGEMENT OF ASTHMA IN PREGNANCY

The goals of therapy and principles of management of asthma in pregnant women are similar to those for non-pregnant women^[9]. However, pregnant women are more likely to be undertreated by physicians for a given level of asthma severity^[45,46]. Evidence has also shown that pregnant asthma patients often avoid medications they believe are potentially harmful to the fetus. Rocklin *et al.*^[36] tracked the prescription claims data in a cohort of 112171 pregnant women who were enrolled in the Tennessee Medicaid program. Subjects significantly decreased ($P \leq 0.0005$) their use of asthma medications during the 5th-13th weeks of pregnancy. Utilization rates during the first trimester declined by 23% for ICS prescriptions, 13% for short-acting B2-agonists (SABA) prescriptions and 54% for rescue corticosteroid prescriptions^[47].

It is known that critical fetal organ development occurs predominately during the first trimester, and available data suggest that first trimester asthma exacerbations in the mother lead to higher rates of malformation in the offspring. Infants born to mothers with a first trimester asthma exacerbation, compared to those without, were more likely to suffer at least one malformation affecting 9.2% of infants (aOR = 1.48 with 95%CI: 1.04-2.09) and a major malformation affecting 6.0% of infants (aOR = 1.32 with 95%CI: 0.86-2.04)^[48]. Such evidence highlights the need for careful management of maintenance therapy in pregnant asthmatics, and suggests the value of a multidisciplinary approach to assessment and treatment of asthma to promote appropriate therapy and medication adherence.

Diagnosis and classification of asthma

The diagnosis of asthma in a pregnant patient is the same as in the nonpregnant population, ideally with confirmation by spirometry showing at least partially reversible obstruction of the airways. If a pregnant patient presents with symptoms of new-onset asthma without spirometric confirmation of diagnosis, they should be treated with appropriate asthma therapy only after other diagnoses are excluded. Differential diagnoses associated with new-

onset dyspnea during pregnancy include physiologic dyspnea of pregnancy, pulmonary embolism, amniotic fluid embolism, pneumonia or bronchitis, GERD, and/or vocal cord dysfunction^[49,50]. It should be noted that testing of bronchial hyperresponsiveness with methacholine challenge is contraindicated during pregnancy due to a lack of safety data^[49].

Evidence clearly shows that the course of asthma is unpredictable as the pregnancy progresses^[17,51]. While baseline asthma severity, the frequency of asthma exacerbations, and the level of asthma control in prior pregnancies have been used as predictors for asthma outcomes during pregnancy, monitoring and assessment of patients with all levels of asthma severity is important. In a study of 1739 women, patients' asthma severity was classified at the onset of pregnancy based on standard criteria including FEV1, symptoms and rescue inhaler use. It was found that 13%, 16%, and 52% of women classified as having mild, moderate, and severe asthma, respectively, suffered at least one asthma exacerbation during pregnancy, suggesting a correlation between baseline asthma severity and the risk of an exacerbation during pregnancy. However, results of the study also demonstrated that 30% of patients initially classified with mild asthma progressed to moderate or severe disease throughout the course of pregnancy, while 23% of patients categorized as having baseline moderate or severe asthma improved to the mild disease category^[51]. Results of this study and other studies (Table 1) illustrate the unpredictable nature of asthma in pregnancy and emphasize the ongoing need for monitoring of asthma symptoms regardless of initial asthma severity. Generally, a patient's asthma course and severity will revert to their pre-pregnancy status approximately 90 d postpartum^[52].

Acute asthma exacerbation management

Most women with asthma complete their pregnancy without incident. But data also indicate 20%-36% of patients will experience an asthma exacerbation, 9%-11% will require hospitalization and some will need ICU management and rarely (< 1%), intubation^[9,38,51]. Exacerbations during pregnancy most commonly occur during the 25th-36th week of pregnancy with fewer episodes occurring during labor and the peripartum period^[53-55]. In addition, evidence suggests that pregnant black women with asthma are more likely to experience and require medical care for exacerbations^[9].

In the Emergency Department (ED), the pregnant asthmatic patient should receive a thorough physical examination, spirometry or peak flow meter assessment and arterial blood gas evaluation. Assessment of maternal oxygen saturation *via* pulse oximetry should be conducted to ensure oxygen saturation is maintained at or above 95%. Spirometry or peak flow meter results can be compared to the patient's baseline measurements or their predicted personal best. Arterial blood gas results usually demonstrate a compensated respiratory alkalosis common to pregnancy. As such, an otherwise normal value

for arterial pCO₂ (pCO₂ of 40 mmHg) in some cases may signal relative hypercapnia and could be an indicator of respiratory fatigue in the pregnant patient^[5].

The health status of the fetus must also be ascertained. Specific recommendations for fetal assessment are determined by the stage of pregnancy but a biophysical profile that combines ultrasound and a non-stress test is routine. These tests are used to measure amniotic fluid volume and fetal heart rate, muscle tone, breathing episodes and gross movements^[9,56].

Recommended initial medical treatment for an acute asthma exacerbation in a pregnant woman presenting to the ED follows standard treatment guidelines. In addition to oxygen supplementation, inhaled albuterol every 20 min up to three doses in the first hour is recommended. If the exacerbation is severe, 500 µg of inhaled ipratropium bromide can supplement albuterol administrations. Oral or intravenous corticosteroids are recommended for individuals with inadequate response to bronchodilator therapy, for individuals who have required multiple short courses of steroids throughout their pregnancy or for those receiving systemic corticosteroids at time of presentation to the ED^[5,9]. If the pregnant patient responds favorably to the bronchodilators and/or corticosteroids, generally within 4 h of presentation to the ED, she may be discharged. On discharge from the ED, a short 5-10 d course of oral prednisone given at 40-80 mg as a single or divided daily doses is recommended to prevent asthma relapses^[5,9].

Alternatively, hospitalization is recommended if a maternal oxygenation saturation of 95% or greater cannot be maintained on room air after appropriate medication administration, if FEV1 or PEF measurements are persistently less than 70% despite therapy, or if fetal distress is evident. Subcutaneous or intravenous terbutaline can be utilized on a case-by-case basis if inhaled SABA have been maximized whereas systemic epinephrine should be avoided^[9]. Life threatening asthma episodes are characterized by significant maternal hypoxemia (PaO₂ < 60 mmHg), hypercapnia (PaCO₂ > 40), respiratory acidosis, maternal respiratory fatigue and/or fetal distress. Intubation and mechanical ventilation can be required in these life-threatening circumstances and on rare occasion, delivery of the newborn by cesarean section is indicated^[9,56].

Maintenance therapy

Due to the increased risk of adverse pregnancy outcomes associated with poor asthma control, optimal management of maternal asthma through optimization of maintenance therapy becomes especially important. Although evidence exists to support the safety of most major classes of medications used for asthma management during pregnancy, patients and providers often remain apprehensive about the use of any drug therapy. Unfortunately, the consequence of decreasing or discontinuing asthma medications during pregnancy is an increase in the likelihood of poor asthma control and its associated risks to both mother and fetus. Recent research has high-

Table 1 Pregnancy associated asthma morbidity by severity classification

	Mild asthma <i>n</i> (%)		Moderate asthma <i>n</i> (%)		Severe asthma <i>n</i> (%)	
	Schatz <i>et al.</i> ^[51]	Murphy <i>et al.</i> ^[55]	Schatz <i>et al.</i> ^[51]	Murphy <i>et al.</i> ^[55]	Schatz <i>et al.</i> ^[51]	Murphy <i>et al.</i> ^[55]
	(<i>n</i> = 873)	(<i>n</i> = 63)	(<i>n</i> = 814)	(<i>n</i> = 34)	(<i>n</i> = 52)	(<i>n</i> = 49)
Asthma Exacerbation	110 (12.6)	5 (8)	209 (25.7) ^b	16 (47)	27 (51.9) ^d	32 (65)
Unscheduled physician or ED presentation	99 (11.3)	4 (6.3)	157 (19.3) ^b	14 (41)	19 (36.5) ^b	20 (41)
Oral corticosteroid use	19 (2.2)	0 (0)	71 (8.7) ^b	4 (11.8)	20 (38.5) ^b	19 (38.8)
Hospitalization	20 (2.3)	2 (3.2)	55 (6.8) ^b	1 (2.9)	14 (26.9) ^b	9 (18.4)

^b*P* < 0.0001, ^d*P* < 0.001 *vs* preceding severity group (moderate to mild; severe to moderate). ED: Emergency Department.

lighted the importance of a multidisciplinary approach to patient education and management of maternal asthma by involving physicians, pharmacists, midwives and others associated with perinatal care, in order to ensure appropriate treatment and promote patient adherence^[57,58].

Regular visits to evaluate asthma control are recommended throughout pregnancy in all patients regardless of disease severity. If there is any indication that maternal asthma symptoms are worsening, more frequent monitoring would be indicated. Disease evaluations should include objective assessment of lung function with the use of spirometry or peak flow meter, as well as assessment of symptoms using a validated questionnaire such as the Asthma Control Test or Asthma Control Questionnaire (ACQ)^[59,60]. More recent research from a double-blind, parallel-group, controlled trial focused on the use of the fraction of exhaled nitric oxide (F_{ENO}) as a marker of airway inflammation in asthma during pregnancy. Results showed a 50% reduction in asthma exacerbations using a treatment algorithm guided by F_{ENO} compared to that guided by symptom assessment^[60]. These early results are encouraging but the use of F_{ENO} is not yet widely available, and its use has yet to be included in guidelines as a standard of care.

As in nonpregnant patients, the pharmacological treatment of asthma should be implemented in a step-wise fashion using current guidelines, with “step-up” therapy indicated if the patient is not adequately controlled with current therapy. Prior to medication regimen changes, issues such as poor medication adherence, improper inhaler technique, and other conditions associated with worsening dyspnea including pneumonia, pulmonary embolism or amniotic embolism, should be assessed^[5,50].

There are, however, several exceptions to current asthma treatment guidelines in pregnant women. First, when a step-up in controller medication is indicated, an ICS should be initially trialed in preference to a combination ICS/long-acting bronchodilator (LABA) product due to the safety concerns associated with LABAs that will be discussed below (Table 2). Second, current asthma guidelines recommend step-down therapy to be considered in non-pregnant patients who are well controlled on a regimen for a minimum of three months^[50]. In contrast, maintenance therapy should not be altered or escalated during pregnancy in asthmatics who are well controlled since fetal risks associated with the loss of disease con-

trol outweigh the benefits associated with a reduction in maintenance therapy^[59,61].

Education

Patient education is an important component of appropriate management in any patient with asthma. Several studies have highlighted the importance of asthma education during pregnancy, using strategies to provide information to patients about the disease and its treatment, as well as improving medication adherence^[55,57]. One recent study by Lim and colleagues showed that 70% of surveyed women were unaware of the risks associated with poor asthma control, while 32% discontinued or changed medications during pregnancy without discussing the changes with a healthcare professional^[57]. Asthma education can directly address these issues and promote improved outcomes.

Key education topics for patients should include general information about asthma, potential complications and their relationship to pregnancy, proper use of inhaler devices, appropriate self-monitoring, adherence to medications, and optimal control of environmental factors. A written asthma action plan should be established to assist patients with self-monitoring and treatment in response to asthma control based on symptoms and/or peak flow monitoring. The plan should be developed in coordination with a healthcare provider and communicated to all those involved in the treatment of the patient. Patients should receive follow-up education and assessment of medication adherence and inhaler technique at every visit. The use of regular education and monitoring through a multidisciplinary team approach has been shown to significantly decrease ACQ scores when compared to groups receiving usual asthma care without education^[50,57-59].

Nonpharmacologic measures and immunizations

Nonpharmacologic approaches can improve asthma symptoms while decreasing the use of “as needed” medication, thereby minimizing any associated maternal or fetal risk. The identification and avoidance or removal of indoor and outdoor environmental asthma “triggers” may greatly reduce the risk of asthma exacerbation. Common triggers including mold, dust, animal dander, cockroaches, pollens, and perfumes are often impossible to avoid completely but minimization of exposure is a treatment goal. Furthermore, smoking cessation and/or avoidance

Table 2 Stepwise approach to asthma therapy in pregnant and nonpregnant patients^[49,60,62]

	Step	Preferred therapy in nonpregnant patients	Preferred therapy in pregnant patients	Alternative therapy in pregnant patients
Intermittent asthma	1	SABA, as needed ¹	SABA, as needed ¹	N/A
Persistent asthma	2	Low-dose ICS	Low-dose ICS	LTRA
	3	Low-dose ICS + LABA, or medium-dose ICS	Medium-dose ICS	LTRA
	4	Medium-dose ICS + LABA	Low-dose ICS + LABA	Medium-dose ICS, or high-dose ICS, or low-dose ICS + LABA + LTRA
	5	High-dose ICS + LABA	Medium-dose ICS + LABA, or high-dose ICS + LABA	LTRA + theophylline
	6	High-dose ICS + LABA + oral corticosteroid	High-dose ICS + LABA + oral corticosteroids	Omalizumab

¹SABA should be included as quick-acting rescue medication to be used as needed in all patients. SABA: Short-acting beta-agonist; LABA: Long-acting beta-agonist; ICS: Inhaled corticosteroid.

of secondhand smoke always should be incorporated to treatment plans of pregnant asthmatic patients^[50,62].

Immunization against influenza is strongly recommended in both pregnant and postpartum patients with asthma, as influenza is more likely to cause severe illness in these populations, leading to serious disease exacerbations that pose risk to maternal and fetal wellbeing. Pregnant women should receive an inactivated influenza vaccine by injection while postpartum women who are breastfeeding may receive either the live or attenuated vaccine given *via* the intranasal route or by injection. Recommendations regarding administration of pneumococcal vaccine in asthmatic patients vary between countries, and some controversy regarding the effectiveness of pneumococcal vaccination in asthma exists. Although no evidence of maternal or fetal harm has been demonstrated following administration of the pneumococcal vaccine (PPSV23) during pregnancy, providers should make every effort to vaccinate women with asthma prior to pregnancy. Pneumococcal vaccine is not, however, contraindicated in breastfeeding^[63-65].

Management of comorbid conditions

While the use of allergen immunotherapy is known to be effective for improving asthma symptoms in patients with allergies, anaphylaxis is the greatest risk accompanying allergen injections in the asthmatic patient and has the potential to result in maternal and/or fetal death. The risk is especially high earlier in the course of immunotherapy when allergen doses are being increased. Consideration of the benefits and risks of allergen immunotherapy generally favors continuation of the treatment if a patient has reached a maintenance or near-maintenance dose without adverse reactions prior to a pregnancy. However, initiation of allergen immunotherapy during pregnancy is not recommended^[50].

As in nonpregnant patients, rhinitis and gastroesophageal reflux may lead to exacerbation of asthma symptoms during pregnancy. Management of these comorbid conditions should be considered an integral part of patient care because pregnancy can result in physiological alterations that lead to worsening of the conditions. Details of

the management of these conditions during pregnancy are beyond the scope of this review; however, the interested reader is referred to references for further information^[66-71].

ASTHMA MEDICATIONS USED DURING PREGNANCY

Data from three studies describing the association of congenital malformations with maternal asthma medication use have been recently published^[44,72,73]. The National Birth Defects Prevention Study by Källén *et al.*^[73] included 2853 infants with one or more specific malformations compared to a control group of 6726 unaffected infants. Mothers of cases and controls were contacted by telephone and asked to describe their medication use beginning one month prior to and through their third month of pregnancy. Other potential risk factors such as tobacco and alcohol use, co-morbid chronic diseases and exposures at home and work were also solicited. Congenital malformations included esophageal atresia, small intestinal atresia, anorectal atresia, limb deficiencies, diaphragmatic hernia, omphalocele, or neural tube defects. Significant associations were determined for omphalocele with both bronchodilators (SABA and/or LABA use) and anti-inflammatory medication use (aOR = 4.13; 95%CI: 1.43-11.95), isolated anorectal atresia with anti-inflammatory use (aOR = 2.12; 95%CI: 1.09-4.12) and isolated esophageal atresia with bronchodilator use (aOR = 2.39; 95%CI: 1.23-4.66). No other positive associations with other birth defects were determined^[72].

Cydulka *et al.*^[45] conducted a systematic review and meta-analysis of the literature concerning the association of maternal asthma disease management with the risk of congenital malformations. Data from 12 cohort studies (four prospective and eight retrospective studies) of women with asthma stratified according to disease severity, exacerbation history, corticosteroid use, or bronchodilator use were included in the analysis. In accordance with other studies, maternal asthma was associated with a significantly increased risk of malformations for the en-

tire group (RR = 1.11; 95%CI: 1.02-1.21) but no increase was observed in the subset of patients in the prospective studies with active asthma medication management. While the presence of asthma was associated with an overall increased risk of congenital malformations, significant associations were not found for any specific factors related to asthma including maternal asthma exacerbation history, bronchodilator use, or ICS use^[44].

Mendola *et al.*^[38] used data from the Swedish Medical Birth Register for the period 1996-2011 to investigate the risk of congenital malformations in infants born to women who had received medications for asthma during early pregnancy. Maternal drug use information was obtained from midwife interview records of patients during the first perinatal care appointment that typically occurred during the 10th-12th wk of pregnancy. The data spanned a 15-year timeframe with over 1.5 million births, including those of 44772 (2.9%) patients who received asthma medications from at least one of the following medication classes: inhaled adrenergics (SABA and/or LABA), ICS, anticholinergics, anti-allergics, xanthines, and leukotriene receptor antagonists. Women receiving antiasthmatic medications were compared to women who did not receive a drug from the listed classes, with adjustments made for year of delivery, maternal age, parity, smoking, and BMI. Those receiving antiasthmatic drugs were further stratified into specific medication classes.

Results indicated the OR for bearing an infant with a major congenital malformation was 1.09 (95%CI: 1.03-1.15) for women receiving any antiasthmatic medication *vs* those with no exposure to asthma medications. Median cleft palate (but not cleft lip/palate) with an OR of 1.45 (95%CI: 1.06-1.98), cardiovascular defects with an OR of 1.13 (95%CI: 1.04-1.23), and pyloric stenosis with an OR of 1.42 (95%CI: 1.06-1.91) were determined to be significantly increased malformations in infants born to mothers who took asthma medications. Risk estimates for the associations of the number of different asthma medications taken by the mother with a major malformation were significant for use of medication from a single group 1.11 (95%CI: 1.04-1.19) and use of medications from three or more groups 1.18 (95%CI: 1.01-1.38). In regard to specific medication classes, significantly increased odds or risk ratio OR/RR were found for the use of SABA (OR/RR = 1.10; 95%CI: 1.04-1.10) and ICS (OR/RR = 1.08; 95%CI: 1.01-1.16). However, there was no examination of asthma severity and its potential links to fetal outcomes in this study. Furthermore, as with all of these recently published studies, increased congenital risks could not be linked to specific, individual medications within a given medication group^[73].

***β*2-agonists**

SABAs are effective bronchodilators for quick-acting relief of asthma symptoms and are generally considered safe for use during pregnancy and breastfeeding. While SABA use was associated with a small increased risk of congenital malformation in some of the large studies

described above, most studies evaluating maternal SABA use during pregnancy have not shown significant increases in adverse maternal or fetal outcomes associated with drug use^[6,74,75]. In other studies that did show significant increases in adverse events potentially correlated with SABA use during pregnancy, reference groups of healthy non-asthmatic women or mixed asthmatic plus non-asthmatic women were used, making it impossible to discern if observed adverse outcomes were attributable to medication use or the disease^[73,74,76-80]. Results of a single population-based case-control study of 511 pregnant women with asthma demonstrated an increased risk of congenital malformations with fenoterol use, but no association with other SABA use^[79]. Due to the preponderance of evidence supporting the safety of SABA use in pregnancy, the drugs should be used according to guidelines for the quick-relief of asthma symptoms.

There is limited data regarding the safety of LABAs during pregnancy. In a population-based retrospective cohort study of β -2 agonist use in pregnancy, Eltonsy and colleagues observed a nonsignificant trend for an increased risk of major congenital malformations in infants of women who used LABAs during the first trimester. In the same study, SABA use was not associated with any increased risk of malformations^[74]. On further analysis of those using a LABA in the first trimester (*n* = 165), investigators found that while there was no significant increase in all major malformations (defined as malformations that were life-threatening, caused major cosmetic defects, or resulted in at least one hospitalization within the first year of life), there were significant increased risks for the subtype of major cardiac malformations (aOR = 2.38, 95%CI: 1.11-5.10), genital organ malformations (aOR = 6.84, 95%CI: 2.58-18.10) and major "other and unspecified congenital malformations" (aOR = 3.97, 95%CI: 1.29-12.20)^[75]. The authors of the study offered explanations for the observed trend of adverse outcomes associated with LABA use beyond that of a true causal relationship. First, while the investigators attempted to correct for asthma severity in the analysis of the data, it was possible that there was residual confounding of results by asthma disease severity. Second, specific interactions between concurrent LABA and steroid use have been identified including effects on protein kinase A (PKA) and ligand-independent activation of glucocorticoid receptors^[81]. Because LABAs were used concomitantly with ICS for asthma in this study, authors suggest that observed increases in fetal malformations might be due to an effect of LABA use on steroid function, leading to potentiation of steroid-associated adverse effects^[75].

Other studies examining LABA use have not found significant association between LABA use and major fetal malformations, and evidence supporting differences in the safety profiles of individual LABAs is lacking^[75,77,80-83]. A recent study by Cossette *et al.*^[84] failed to find any statistically significant differences in low birth weight or preterm birth for infants of mothers who had used sal-

meterol *vs* formoterol during pregnancy.

Due to a lack of robust evidence for the safety of LABA use in pregnancy, the medications should only be used if asthma control cannot be achieved using medium-dose steroids in addition to SABAs. As previously noted, this recommendation is a deviation from asthma guidelines for nonpregnant patients (Table 2). Maternal plasma concentrations of inhaled LABAs have been shown to be undetectable or minimal, however, and the use of the agents is not considered a contraindication to breastfeeding^[85].

Inhaled and systemic corticosteroids

Due to potent and predictable anti-inflammatory effects, ICS form the foundation of maintenance therapy in patients with persistent asthma. As a drug class, ICS have generally been shown to decrease the risk of asthma exacerbations among pregnant women, with no increased rate in adverse maternal or fetal outcomes^[86-88]. Systemic absorption of an ICS is typically very low, with data demonstrating very low to undetectable plasma concentrations of triamcinolone, fluticasone, ciclesonide and beclomethasone after inhalation^[89]. Inhaled budesonide has approximately 39% bioavailability, but results of studies of inhaled budesonide in lactation demonstrated a negligible amount transferred to the breastfeeding infant^[90]. Additionally, similar incidences of adverse pregnancy outcomes were observed in a randomized, controlled trial comparing the use of inhaled budesonide *vs* placebo in pregnant women with asthma^[90]. Information from other studies of ICS use in asthmatic patients during pregnancy provide similar evidence indicating no significant increased risk for neonatal adverse events including oral clefts, cardiac defects, spina bifida and other congenital malformations beyond those expected in the general population^[88,91-93].

Concerns regarding the safety of corticosteroids in pregnancy have been specifically addressed in a number of studies. Data from most studies support the safety of ICS use for asthma during pregnancy^[55,76,88,93-96]. However, in a retrospective cohort study of 817 asthmatic women during pregnancy, Lim *et al*^[97] found statistically significant increases in the rates of pregnancy-induced hypertension (OR = 1.7, 95%CI: 1.0-2.9) and neonatal hyperbilirubinemia (OR = 1.9, 95%CI: 1.1-3.4) associated with the use of ICS or oral corticosteroid as compared to those using no medication. It is important to note, however, that outcomes associated with oral and inhaled corticosteroids were combined in this analysis, rather than independently assessed. Additionally, the authors cite the inability to distinguish well-controlled from uncontrolled asthma as an important limitation to this study^[96].

Comparative studies of various ICS and commonly used dosages are somewhat lacking. All ICS except budesonide are classified as pregnancy category C by the United States Federal Drug Administration (FDA). Budesonide was moved to category B based on evidence of its safety from Dombrowski *et al*^[93]; however, no spe-

cific data exists to suggest that other ICS are less safe for use during pregnancy. The United States National Heart Lung and Blood Institute guidelines of 2008 state that budesonide is the preferred ICS during pregnancy, but states that other ICS may be continued^[58,59]. More recent global guidelines do not distinguish a preferred ICS for treatment of asthma during pregnancy, consistent with evidence from studies that has not shown significant differences in adverse maternal or fetal outcomes between patients using an ICS with beclomethasone, budesonide or fluticasone during pregnancy^[50,77,88].

Few studies contain information regarding dosage ranges for the ICS used, making it difficult to determine if any risk of fetal adverse effects is dose-related^[98]. Investigators in several studies did detect trends towards increased rates of SGA infants and increased congenital malformations with increasing doses of ICS, but the differences did not reach statistical significance and authors could not rule out confounding of results due to asthma severity^[98,99]. In summary, there is no compelling evidence to substantiate a correlation between the use of an ICS during pregnancy and an increased risk of adverse infant outcomes; currently, these agents should be used when necessary to maintain asthma control.

Systemic corticosteroids should be reserved for use in acute exacerbations for all asthma patients or in those patients unable to achieve disease control using other agents. In contrast to ICS, oral corticosteroids have been associated with a higher incidence of maternal adverse effects, including preeclampsia and gestational diabetes^[94,96,100,101]. A meta-analysis and systematic literature review conducted by Murphy *et al*^[44] could not rule out the possibility of increased malformation risk associated with maternal oral corticosteroid use during a critical period for fetal lip and palate closure. The suggestion is based on data from case control studies that have indicated cleft lip and or cleft palate may not only be associated with maternal asthma but also with exposure to first-trimester oral corticosteroids^[101,102]. As with other asthma therapies, the benefits associated with gaining control of severe uncontrolled asthma symptoms often outweigh the risks of adverse events associated with systemic steroid use. Nevertheless, systemic corticosteroids should be used judiciously in pregnancy and patients should be closely monitored for adverse effects.

Leukotriene receptor antagonists

Leukotrienes are potent mediators in the signaling pathways of allergic inflammation and thus play a central role in the pathophysiology of asthma. Leukotriene antagonists (LTRAs) function to reduce inflammation through this pathway and can reduce asthma exacerbations and improve lung function in persistent asthma. Few studies have been conducted to analyze the effects of this medication class exclusively and large, well-designed studies of LTRA use in pregnancy are lacking.

Limited data from available studies have shown conflicting results regarding maternal and fetal adverse out-

comes associated with the use of LTRAs^[103-108]. A study of 180 pregnant asthmatics examined the effects of montelukast exposure compared to two separate groups of pregnant women: 180 disease-matched controls using inhalers but with no exposure to LTRAs and 180 age-matched healthy controls with no known teratogen exposure. Investigators found that in asthmatic women who used montelukast during the first trimester of pregnancy, there were significantly increased rates of infant LBW, preterm delivery and fetal distress when compared to healthy non-asthmatic maternal controls. Although only 47.4% of pregnant women taking montelukast in the first trimester continued the medication throughout pregnancy, a subgroup analysis of these patients demonstrated no significant difference in rates of fetal distress or preterm delivery when compared to asthmatic controls. This finding suggested a protective effect of montelukast likely due to improved asthma control throughout pregnancy. Moreover, investigators found no significant differences in adverse effects in pregnant women with asthma exposed to LTRAs compared to disease-matched asthmatic maternal controls^[108].

In contrast, a previously published study generated results that indicated a nonsignificant increase in the rate of malformations in 96 asthma patients who received a LTRA throughout pregnancy as compared to 122 pregnant asthmatics who received SABA monotherapy. Malformations were observed in 5.95% of LTRA compared to 3.9% of SABA users, respectively ($P = 0.524$). Notably, the study had important limitations in addition to a relatively small sample size. The women taking LTRAs also were exposed during pregnancy to other asthma medications including SABAs, ICS, and oral corticosteroids. Also, LTRA use was associated with an increased patient baseline asthma severity which was not adjusted for in this study, and could account for the increased rate of malformations^[104]. Further studies are needed in order to determine the safety of LTRA agents during pregnancy and/or lactation, but available data suggest that if necessary, montelukast would be the preferred LTRA due to a greater amount of evidence supporting its safety and a safer lactation profile.

Theophylline

Theophylline is a drug with mild bronchial anti-inflammatory effects^[105]. While it is not a preferred agent in the treatment of asthma due to prevalent adverse effects, drug-drug interactions and the need for monitoring of serum concentrations, theophylline may be beneficial in selected patients. In a prospective study, 153 women with asthma including 85 receiving theophylline were followed throughout the course of their pregnancy. Results of the study demonstrated a significantly reduced risk of preeclampsia in patients treated with compared to those not receiving theophylline. Investigators suggested that theophylline's ability to increase cAMP levels and thereby reduce vascular reactivity and platelet aggregation may result in the decreased incidence of preeclampsia^[106].

A subsequent study compared theophylline with inhaled beclomethasone therapy in 398 pregnant females with mild or moderate persistent asthma. No significant differences in adverse obstetric outcomes including preeclampsia, preterm delivery and oligohydramnios were detected between patient groups. Additionally, there was no significant difference in the number of asthma exacerbations between the two groups, although there was a significant increase in the proportion of women with a FEV1 less than 80% of predicted among the theophylline group^[107]. Available evidence suggests that use of theophylline in pregnancy is likely safe; the drug is currently classified as a category C medication by the United States FDA^[85,108].

Mast-cell stabilizers

Mast-cell stabilizers prevent mast-cell release of histamine and other inflammatory mediators during allergic response. Although they are not commonly compared to other asthma medications, they are considered effective second-line agents for asthma control. Very few studies have evaluated the use of mast-cell stabilizers for asthma during pregnancy and major limitations of available studies include small patient sample size, concurrent use of other medications, and comparison of treatment groups to healthy, non-asthmatic controls. Nevertheless, cromolyns are considered safe for use during pregnancy due to limited systemic bioavailability, and could be an appropriate adjunctive therapy in some patients^[86,109].

Omalizumab

Omalizumab is a recombinant monoclonal anti-IgE therapy that works by binding and neutralizing the effects of IgE in basophils and mast cells, thereby preventing downstream allergic inflammation. The biologic therapy is reserved for patients with moderate to severe persistent asthma who are unable to be controlled by medium- to high-dose ICS plus LABA therapy. As a relatively new therapy, evidence for safety of omalizumab use in pregnancy is very limited. Currently, the Xolair® Pregnancy Registry (EXPECT) is collecting data for an ongoing observational study designed to monitor outcomes in women exposed to omalizumab during the time period starting eight weeks prior to conception and continuing throughout the pregnancy. Of the 128 known outcomes from preliminary data in this registry, there were 119 live births, with 117 singletons and two pairs of twins for a total of 121 infants. Of these infants, 16% were premature (gestational age less than 37 wk) and 7% had a birth weight less than 2.5 kg. The rate of major birth defects was 4%, with observed defects including patent foramen ovale, cutaneous mastocytosis, hemangioma, hypospadias, and bilateral renal pelvis dilation^[110]. It is important to note that this agent is typically reserved for patients with moderate to severe asthma, and thus, it is difficult for effects related to omalizumab use to be differentiated from effects due to disease severity. Currently, the United States FDA has classified this agent as pregnancy cat-

Table 3 Asthma medications in pregnancy and lactation

Medication	United States FDA pregnancy category ¹	Australian Drug Evaluation Committee pregnancy category ²	German pregnancy risk category ³	Lactation ^[85]
Inhaled corticosteroids				
Beclomethasone	C	B3	Group 3	Unknown
Budesonide	B	A	Group 3	Unknown
Ciclesonide	C	B3	--	Unknown
Fluticasone	C	B3	Group 5	Unknown
Mometasone	C	B3	Group 5	Unknown
Short-acting β -agonists				
Albuterol	C	A	--	Likely safe
Levalbuterol	C	A	--	Unknown
Terbutaline	C	A	--	Likely safe
Long-acting β -agonists				
Formoterol	C	B3	Group 4	Unknown
Salmeterol	C	B3	Group 5	Unknown
Leukotriene inhibitors				
Montelukast	B	B1	Group 5	Likely safe
Zafirlukast	B	B1	--	Possibly unsafe
Zileuton	C	--	--	Likely safe
Mast-cell stabilizers				
Nedocromil	B	B1	Group 4	Unknown
Cromolyn	B	A	Group 1	Unknown
Systemic corticosteroids				
Dexamethasone	C	A	--	Likely safe
Methylprednisolone	C	A	Group 3	Likely safe
Prednisone	C	A	Group 3	Likely safe
Theophylline	C	A	--	Likely safe
Omalizumab	B	B1	Group 4	Unknown

¹United States Federal Drug Association pregnancy categories^[111]: Category A: Adequate and well-controlled studies have failed to demonstrate a risk to the fetus in the first trimester of pregnancy (and there is no evidence of risk in later trimesters); Category B: Animal reproduction studies have failed to demonstrate a risk to the fetus and there are no adequate and well-controlled studies in pregnant women; Category C: Animal reproduction studies have shown an adverse effect on the fetus and there are no adequate and well-controlled studies in humans, but potential benefits may warrant use of the drug in pregnant women despite potential risks; Category D: There is positive evidence of human fetal risk based on adverse reaction data from investigational or marketing experience or studies in humans, but potential benefits may warrant use of the drug in pregnant women despite potential risks; Category X: Studies in animals or humans have demonstrated fetal abnormalities and/or there is positive evidence of human fetal risk based on adverse reaction data from investigational or marketing experience, and the risks involved in use of the drug in pregnant women clearly outweigh potential benefits; ²Australian Drug Evaluation Committee pregnancy categories^[111]: Category A: Drugs which have been taken by a large number of pregnant women and women of childbearing age without any proven increase in the frequency of malformations or other direct or indirect harmful effects on the fetus having been observed; Category B1: Drugs that have been taken by only a limited number of pregnant women and women of childbearing age, without an increase in the frequency of malformation or other direct or indirect harmful effects on the human fetus having been observed. Studies in animals have not shown evidence of an increased occurrence of fetal damage; Category B2: Drugs that have been taken by only a limited number of pregnant women and women of childbearing age, without an increase in the frequency of malformation or other direct or indirect harmful effects on the human fetus having been observed. Studies in animals are inadequate or may be lacking, but available data show no evidence of an increased occurrence of fetal damage; Category B3: Drugs that have been taken by only a limited number of pregnant women and women of childbearing age, without an increase in the frequency of malformation or other direct or indirect harmful effects on the human fetus having been observed. Studies in animals have shown evidence of an increased occurrence of fetal damage, the significance of which is considered uncertain in humans; Category C: Drugs that, owing to their pharmacological effects, have caused or may be suspected of causing harmful effects on the human fetus or neonate without causing malformations. These effects may be reversible; Category D: Drugs that have caused or are suspected to have caused or may be expected to cause an increased incidence of human fetal malformations or irreversible damage. These drugs may also have adverse pharmacological effects; Category X: Drugs that have such a high risk of causing permanent damage to the fetus that they should not be used in pregnancy or when there is a possibility of pregnancy; ³German pregnancy risk categories^[113]: Group 1: Extensive human tests and animal studies have not shown the drug to be embryotoxic/teratogenic; Group 2: Extensive human tests of the drug have not shown the drug to be embryotoxic/teratogenic; Group 3: Extensive human tests of the drug have not shown the drug to be embryotoxic/teratogenic. However, the drug appears to be embryotoxic/teratogenic in animals; Group 4: No adequate and well-controlled studies of the drug's effects on humans are available. Animal studies have shown no embryotoxic/teratogenic effects; Group 5: No adequate and well-controlled studies of the drug's effects on humans are available; Group 6: No adequate and well-controlled studies of the drug's effects on humans are available. Animal studies have shown embryotoxic/teratogenic effects"; Group 7: There is a risk that the drug is embryotoxic/teratogenic in humans, at least in the first trimester; Group 8: There is a risk that the drug is toxic to fetuses throughout the second and third trimesters; Group 9: There is a risk that the drug causes prenatal complications or abnormalities; Group 10: There is a risk that the drug causes hormone specific action on the human fetus; Group 11: There is a known risk that the drug is a mutagen/carcinogen.

egory B based on evidence from animal studies (Table 2). Due to the small amount of human safety data, appropriate risk-benefit analysis should be undertaken before use in pregnancy.

DISCUSSION

Despite large amounts of data related to the influence of asthma and its treatment on maternal and fetal outcomes,

there are a number of limitations to these studies. Many early studies evaluating the influence of asthma control on maternal and fetal outcomes failed to assess, or poorly defined baseline asthma severity as well as the frequency, timing and severity of asthma exacerbations during pregnancy. However, data are available from other studies that have corrected for race, smoking status, age of mother, mean gestational age at enrollment, baseline asthma severity classification, severity of asthma exacerbations and other important covariates^[33,35,42,111]. Based on existing information, it is clear that poor maternal asthma control has serious implications for both maternal and fetal health.

While congenital malformations are believed to be caused by a variety of factors associated with maternal asthma during pregnancy, establishing an association between maternal asthma effects and neonatal congenital malformations has also been challenging. Small sample sizes, varying study designs (case-control *vs* cohort), the timing of maternal study enrollment, a lack of correction for multiple testing and other confounders are routinely cited as key limitations^[43,44,73]. Additionally, separating the impact of maternal asthma from effects caused by asthma medications on resultant fetal malformations is a daunting task. Given the low overall rate of congenital malformations in the general population (3%), a power analysis indicates that nearly 12000 women with asthma would be needed to detect a relatively small 15% increase for a major congenital malformation, given an alpha level of significance of 0.05 and a beta of 0.80^[73]. Generally, data from large studies support a small increased risk of malformations from asthma medication use, although this risk is difficult to delineate from confounding factors including asthma severity, asthma control during pregnancy, fetal hypoxia at birth or simply chance^[44,73,73]. Further studies that control for these confounding factors are required in order to truly separate the effects of disease *vs* the effects of medication use in pregnancy.

Although patients may express concerns regarding possible fetal adverse effects related to medication use, the majority of medications used for asthma maintenance therapy are regarded as safe (Table 3). In 2008, the United States FDA proposed the elimination of current pregnancy categories A, B, C, D, and X in favor of more detailed information for drug safety in pregnancy in lactation. The new format of pregnancy and lactation labeling aims to provide improved information for risk analysis and patient counseling on package inserts for all drugs. With its final version currently undergoing review and clearance, these changes are expected to improve the data available for the sometimes difficult clinical decision-making regarding the use of prescription drugs during pregnancy and lactation^[112,113].

Most evidence indicates that improved maternal and fetal outcomes are correlated with improved asthma control with medications during pregnancy, suggesting the greatest adverse fetal risks are associated with poor

asthma control^[48]. Outcomes can be further improved through appropriate disease monitoring and management, patient education, and optimization of nonpharmacologic interventions to improve asthma control^[50,57-59].

CONCLUSION

As a common condition in pregnancy, asthma can have a significant impact on both maternal and fetal health. Frequent monitoring and optimization of both pharmacological and nonpharmacological modalities are crucial to maintaining asthma control throughout pregnancy. Asthma management should also focus on education to promote patient understanding of the risks associated with uncontrolled asthma, avoidance of asthma triggers, proper inhaler technique and appropriate adherence to asthma therapy. Although some patients and providers will be concerned about the use of asthma medications during pregnancy, evidence shows the greatest risk of adverse maternal and perinatal outcomes is associated with uncontrolled asthma, and that the benefits of maintaining asthma control outweigh the risks associated with medication use.

REFERENCES

- 1 **Kwon HL**, Triche EW, Belanger K, Bracken MB. The epidemiology of asthma during pregnancy: prevalence, diagnosis, and symptoms. *Immunol Allergy Clin North Am* 2006; **26**: 29-62 [PMID: 16443142 DOI: 10.1016/j.iac.2005.11.002]
- 2 **Barron WM**, Leff AR. Asthma in pregnancy. *Am Rev Respir Dis* 1993; **147**: 510-511 [PMID: 8442579]
- 3 **Tamási L**, Horváth I, Bohács A, Müller V, Losonczy G, Schatz M. Asthma in pregnancy--immunological changes and clinical management. *Respir Med* 2011; **105**: 159-164 [PMID: 21145223 DOI: 10.1016/j.rmed.2010.11.006]
- 4 **Kircher S**, Schatz M, Long L. Variables affecting asthma course during pregnancy. *Ann Allergy Asthma Immunol* 2002; **89**: 463-466 [PMID: 12452203 DOI: 10.1016/S1081-1206(10)62082-0]
- 5 **Maselli DJ**, Adams SG, Peters JI, Levine SM. Management of asthma during pregnancy. *Ther Adv Respir Dis* 2013; **7**: 87-100 [PMID: 23129568 DOI: 10.1177/1753465812464287]
- 6 **Guy ES**, Kirumaki A, Hanania NA. Acute asthma in pregnancy. *Crit Care Clin* 2004; **20**: 731-745, x [PMID: 15388199 DOI: 10.1016/j.ccc.2004.05.013]
- 7 **Prowse CM**, Gaensler EA. Respiratory and acid-base changes during pregnancy. *Anesthesiology* 1965; **26**: 381-392 [PMID: 14313450]
- 8 **Soothill PW**, Nicolaidis KH, Rodeck CH, Gamsu H. Blood gases and acid-base status of the human second-trimester fetus. *Obstet Gynecol* 1986; **68**: 173-176 [PMID: 3090491]
- 9 **Hanania NA**, Belfort MA. Acute asthma in pregnancy. *Crit Care Med* 2005; **33**: S319-S324 [PMID: 16215354 DOI: 10.1097/01.CCM.0000182789.14710.A1]
- 10 **Saito S**, Shiozaki A, Sasaki Y, Nakashima A, Shima T, Ito M. Regulatory T cells and regulatory natural killer (NK) cells play important roles in fetomaternal tolerance. *Semin Immunopathol* 2007; **29**: 115-122 [PMID: 17621697 DOI: 10.1007/s00281-007-0067-2]
- 11 **Ostensen M**, Villiger PM. Immunology of pregnancy-pregnancy as a remission inducing agent in rheumatoid arthritis. *Transpl Immunol* 2002; **9**: 155-160 [PMID: 12180824 DOI: 10.1016/S0966-3274(02)00017-5]
- 12 **Robinson DS**, Hamid Q, Ying S, Tscicopoulos A, Barkans J,

- Bentley AM, Corrigan C, Durham SR, Kay AB. Predominant TH2-like bronchoalveolar T-lymphocyte population in atopic asthma. *N Engl J Med* 1992; **326**: 298-304 [PMID: 1530827 DOI: 10.1056/NEJM199201303260504]
- 13 **Mazzarella G**, Bianco A, Catena E, De Palma R, Abbate GF. Th1/Th2 lymphocyte polarization in asthma. *Allergy* 2000; **55** Suppl 61: 6-9 [PMID: 10919498 DOI: 10.1034/j.1398-9995.2000.00511.x]
 - 14 **Tamási L**, Bohács A, Pállinger E, Falus A, Rigó J, Müller V, Komlósi Z, Magyar P, Losonczy G. Increased interferon-gamma- and interleukin-4-synthesizing subsets of circulating T lymphocytes in pregnant asthmatics. *Clin Exp Allergy* 2005; **35**: 1197-1203 [PMID: 16164448 DOI: 10.1111/j.1365-2222.2005.02322.x]
 - 15 **Toldi G**, Molvarec A, Stenczer B, Müller V, Eszes N, Bohács A, Bikov A, Rigó J, Vásárhelyi B, Losonczy G, Tamási L. Peripheral T(h)1/T(h)2/T(h)17/regulatory T-cell balance in asthmatic pregnancy. *Int Immunol* 2011; **23**: 669-677 [PMID: 21937455 DOI: 10.1111/j.1600-0897.2010.00878.x]
 - 16 **Bohács A**, Cseh A, Stenczer B, Müller V, Gálffy G, Molvarec A, Rigó J, Losonczy G, Vásárhelyi B, Tamási L. Effector and regulatory lymphocytes in asthmatic pregnant women. *Am J Reprod Immunol* 2010; **64**: 393-401 [PMID: 20528830]
 - 17 **Palmer GW**, Claman HN. Pregnancy and immunology: selected aspects. *Ann Allergy Asthma Immunol* 2002; **89**: 350-359; quiz 350-359; 428 [PMID: 12392378 DOI: 10.1016/S1081-1206(10)62034-0]
 - 18 **Denney JM**, Nelson EL, Wadhwa PD, Waters TP, Mathew L, Chung EK, Goldenberg RL, Culhane JF. Longitudinal modulation of immune system cytokine profile during pregnancy. *Cytokine* 2011; **53**: 170-177 [PMID: 21123081 DOI: 10.1016/j.cyto.2010.11.005]
 - 19 **Nelson-Piercy C**. Asthma in pregnancy. *Thorax* 2001; **56**: 325-328 [PMID: 11254828 DOI: 10.1136/thorax.56.4.325]
 - 20 **Tan KS**, McFarlane LC, Lipworth BJ. Modulation of airway reactivity and peak flow variability in asthmatics receiving the oral contraceptive pill. *Am J Respir Crit Care Med* 1997; **155**: 1273-1277 [PMID: 9105066 DOI: 10.1164/ajrccm.155.4.9105066]
 - 21 **Tan KS**. Premenstrual asthma: epidemiology, pathogenesis and treatment. *Drugs* 2001; **61**: 2079-2086 [PMID: 11735634]
 - 22 **Chandler MH**, Schuldheisz S, Phillips BA, Muse KN. Premenstrual asthma: the effect of estrogen on symptoms, pulmonary function, and beta 2-receptors. *Pharmacotherapy* 1997; **17**: 224-234 [PMID: 9085312 DOI: 10.1002/j.1875-9114.1997.tb03703.x]
 - 23 **Vrieze A**, Postma DS, Kerstjens HA. Perimenstrual asthma: a syndrome without known cause or cure. *J Allergy Clin Immunol* 2003; **112**: 271-282 [PMID: 12897732 DOI: 10.1067/mai.2003.1676]
 - 24 **Arruvito L**, Sanz M, Banham AH, Fainboim L. Expansion of CD4+CD25+and FOXP3+ regulatory T cells during the follicular phase of the menstrual cycle: implications for human reproduction. *J Immunol* 2007; **178**: 2572-2578 [PMID: 17277167 DOI: 10.4049/jimmunol.178.4.2572]
 - 25 **Brancazio LR**, Laifer SA, Schwartz T. Peak expiratory flow rate in normal pregnancy. *Obstet Gynecol* 1997; **89**: 383-386 [PMID: 9052590]
 - 26 **Juniper EF**. Effect of asthma on quality of life. *Can Respir J* 1998; **5** Suppl A: 77A-84A [PMID: 9753523]
 - 27 **Tan KS**, McFarlane LC, Lipworth BJ. Paradoxical down-regulation and desensitization of beta2-adrenoceptors by exogenous progesterone in female asthmatics. *Chest* 1997; **111**: 847-851 [PMID: 9106558 DOI: 10.1378/chest.111.4.847]
 - 28 **Beecroft N**, Cochrane GM, Milburn HJ. Effect of sex of fetus on asthma during pregnancy: blind prospective study. *BMJ* 1998; **317**: 856-857 [PMID: 9748178 DOI: 10.1136/bmj.317.7162.856]
 - 29 **Murphy VE**, Gibson PG, Giles WB, Zakar T, Smith R, Bisits AM, Kessell CG, Clifton VL. Maternal asthma is associated with reduced female fetal growth. *Am J Respir Crit Care Med* 2003; **168**: 1317-1323 [PMID: 14500261 DOI: 10.1164/rccm.200303-374OC]
 - 30 **Clifton VL**, Murphy VE. Maternal asthma as a model for examining fetal sex-specific effects on maternal physiology and placental mechanisms that regulate human fetal growth. *Placenta* 2004; **25** Suppl A: S45-S52 [PMID: 15033307 DOI: 10.1016/j.placenta.2004.01.004]
 - 31 **Bakhireva LN**, Schatz M, Jones KL, Tucker CM, Slymen DJ, Klonoff-Cohen HS, Gresham L, Johnson D, Chambers CD. Fetal sex and maternal asthma control in pregnancy. *J Asthma* 2008; **45**: 403-407 [PMID: 18569234 DOI: 10.1080/02770900801971826]
 - 32 **Macmullen NJ**, Shen JJ, Tymkow C. Adverse maternal outcomes in women with asthma versus women without asthma. *Appl Nurs Res* 2010; **23**: e9-e13 [PMID: 20122503 DOI: 10.1016/j.apnr.2009.03.004]
 - 33 **Liu S**, Wen SW, Demissie K, Marcoux S, Kramer MS. Maternal asthma and pregnancy outcomes: a retrospective cohort study. *Am J Obstet Gynecol* 2001; **184**: 90-96 [PMID: 11174486 DOI: 10.1067/mob.2001.108073]
 - 34 **Demissie K**, Breckenridge MB, Rhoads GG. Infant and maternal outcomes in the pregnancies of asthmatic women. *Am J Respir Crit Care Med* 1998; **158**: 1091-1095 [PMID: 9769265 DOI: 10.1164/ajrccm.158.4.9802053]
 - 35 **Enriquez R**, Griffin MR, Carroll KN, Wu P, Cooper WO, Gebretsadik T, Dupont WD, Mitchel EF, Hartert TV. Effect of maternal asthma and asthma control on pregnancy and perinatal outcomes. *J Allergy Clin Immunol* 2007; **120**: 625-630 [PMID: 17658591 DOI: 10.1016/j.jaci.2007.05.044]
 - 36 **Rocklin RE**. Asthma, asthma medications and their effects on maternal/fetal outcomes during pregnancy. *Reprod Toxicol* 2011; **32**: 189-197 [PMID: 21684328 DOI: 10.1016/j.reprotox.2011.05.023]
 - 37 **Källén B**, Rydhstroem H, Aberg A. Asthma during pregnancy—a population based study. *Eur J Epidemiol* 2000; **16**: 167-171 [PMID: 10845267]
 - 38 **Mendola P**, Laughon SK, Männistö TI, Leishear K, Reddy UM, Chen Z, Zhang J. Obstetric complications among US women with asthma. *Am J Obstet Gynecol* 2013; **208**: 127.e1-127.e8 [PMID: 23159695 DOI: 10.1016/j.ajog.2012.11.007]
 - 39 **Blais L**, Kettani FZ, Forget A. Relationship between maternal asthma, its severity and control and abortion. *Hum Reprod* 2013; **28**: 908-915 [PMID: 23427230 DOI: 10.1093/humrep/det024]
 - 40 **Murphy VE**, Namazy JA, Powell H, Schatz M, Chambers C, Attia J, Gibson PG. A meta-analysis of adverse perinatal outcomes in women with asthma. *BJOG* 2011; **118**: 1314-1323 [PMID: 21749633 DOI: 10.1111/j.1471-0528.2011.03055.x]
 - 41 **Bracken MB**, Triche EW, Belanger K, Saftlas A, Beckett WS, Leaderer BP. Asthma symptoms, severity, and drug therapy: a prospective study of effects on 2205 pregnancies. *Obstet Gynecol* 2003; **102**: 739-752 [PMID: 14551004]
 - 42 **Dombrowski MP**, Schatz M, Wise R, Momirova V, Landon M, Mabie W, Newman RB, McNellis D, Hauth JC, Lindheimer M, Caritis SN, Leveno KJ, Meis P, Miodovnik M, Wapner RJ, Paul RH, Varner MW, O'Sullivan MJ, Thurnau GR, Conway DL. Asthma during pregnancy. *Obstet Gynecol* 2004; **103**: 5-12 [PMID: 14704237]
 - 43 **Tata LJ**, Lewis SA, McKeever TM, Smith CJ, Doyle P, Smeeth L, Gibson JE, Hubbard RB. Effect of maternal asthma, exacerbations and asthma medication use on congenital malformations in offspring: a UK population-based study. *Thorax* 2008; **63**: 981-987 [PMID: 18678701 DOI: 10.1136/thx.2008.098244]
 - 44 **Murphy VE**, Wang G, Namazy JA, Powell H, Gibson PG, Chambers C, Schatz M. The risk of congenital malformations, perinatal mortality and neonatal hospitalisation among pregnant women with asthma: a systematic review and meta-analysis. *BJOG* 2013; **120**: 812-822 [PMID: 23530780 DOI: 10.1111/1471-0528.12224]

- 45 **Cydulka RK**, Emerman CL, Schreiber D, Molander KH, Woodruff PG, Camargo CA. Acute asthma among pregnant women presenting to the emergency department. *Am J Respir Crit Care Med* 1999; **160**: 887-892 [PMID: 10471614 DOI: 10.1164/ajrccm.160.3.9812138]
- 46 **McCallister JW**, Benninger CG, Frey HA, Phillips GS, Mastronarde JG. Pregnancy related treatment disparities of acute asthma exacerbations in the emergency department. *Respir Med* 2011; **105**: 1434-1440 [PMID: 21700439 DOI: 10.1016/j.rmed.2011.05.015]
- 47 **Enriquez R**, Wu P, Griffin MR, Gebretsadik T, Shintani A, Mitchel E, Carroll KN, Hartert TV. Cessation of asthma medication in early pregnancy. *Am J Obstet Gynecol* 2006; **195**: 149-153 [PMID: 16631099 DOI: 10.1016/j.ajog.2006.01.065]
- 48 **Blais L**, Forget A. Asthma exacerbations during the first trimester of pregnancy and the risk of congenital malformations among asthmatic women. *J Allergy Clin Immunol* 2008; **121**: 1379-1384, 1384.e1 [PMID: 18410961 DOI: 10.1016/j.jaci.2008.02.038]
- 49 **Vatti RR**, Teuber SS. Asthma and pregnancy. *Clin Rev Allergy Immunol* 2012; **43**: 45-56 [PMID: 21858482]
- 50 Global Strategy for Asthma Management and Prevention. Global Initiative for Asthma (GINA), 2013. (Date last updated, 2013). Available from: URL: <http://www.ginasthma.org>
- 51 **Schatz M**, Dombrowski MP, Wise R, Thom EA, Landon M, Mabie W, Newman RB, Hauth JC, Lindheimer M, Caritis SN, Leveno KJ, Meis P, Miodovnik M, Wapner RJ, Paul RH, Varner MW, O'sullivan MJ, Thurnau GR, Conway D, McNellis D. Asthma morbidity during pregnancy can be predicted by severity classification. *J Allergy Clin Immunol* 2003; **112**: 283-288 [PMID: 12897733 DOI: 10.1067/mai.2003.1516]
- 52 **Elsayegh D**, Shapiro JM. Management of the obstetric patient with status asthmaticus. *J Intensive Care Med* 2008; **23**: 396-402 [PMID: 18794165 DOI: 10.1177/0885066608324295]
- 53 **Hodder R**, Loughheed MD, FitzGerald JM, Rowe BH, Kaplan AG, McIvor RA. Management of acute asthma in adults in the emergency department: assisted ventilation. *CMAJ* 2010; **182**: 265-272 [PMID: 19901044 DOI: 10.1503/cmaj.080073]
- 54 **Chan AL**, Juarez MM, Gidwani N, Albertson TE. Management of Critical Asthma Syndrome During Pregnancy. *Clin Rev Allergy Immunol* 2013 Nov 21; Epub ahead of print [PMID: 24258096 DOI: 10.1007/s12016-013-8397-4]
- 55 **Murphy VE**, Gibson PG, Talbot PI, Kessell CG, Clifton VL. Asthma self-management skills and the use of asthma education during pregnancy. *Eur Respir J* 2005; **26**: 435-441 [PMID: 16135724 DOI: 10.1183/09031936.05.00135604]
- 56 **Schatz M**, Dombrowski MP. Clinical practice. Asthma in pregnancy. *N Engl J Med* 2009; **360**: 1862-1869 [PMID: 19403904 DOI: 10.1056/NEJMcp0809942]
- 57 **Lim AS**, Stewart K, Abramson MJ, Walker SP, Smith CL, George J. Multidisciplinary Approach to Management of Maternal Asthma (MAMMA): a randomized controlled trial. *Chest* 2014; **145**: 1046-1054 [PMID: 24522786 DOI: 10.1378/chest.13-2276]
- 58 **National Heart Lung and Blood Institute**. Expert Panel Report 3: Guidelines for the Diagnosis and Management of Asthma (EPR-3), 2007. (Date last updated, 2008). Available from: URL: <http://www.nhlbi.nih.gov/guidelines/asthma/asthdln.htm>
- 59 NAEPP expert panel report. Managing asthma during pregnancy: recommendations for pharmacologic treatment-2004 update. *J Allergy Clin Immunol* 2005; **115**: 34-46 [PMID: 15637545 DOI: 10.1016/j.jaci.2004.10.023]
- 60 **Powell H**, Murphy VE, Taylor DR, Hensley MJ, McCaffery K, Giles W, Clifton VL, Gibson PG. Management of asthma in pregnancy guided by measurement of fraction of exhaled nitric oxide: a double-blind, randomised controlled trial. *Lancet* 2011; **378**: 983-990 [PMID: 21907861 DOI: 10.1016/S0140-6736(11)60971-9]
- 61 **Dombrowski MP**, Schatz M. ACOG practice bulletin: clinical management guidelines for obstetrician-gynecologists number 90, February 2008: asthma in pregnancy. *Obstet Gynecol* 2008; **111**: 457-464 [PMID: 18238988 DOI: 10.1097/AOG.0b013e3181665ff4]
- 62 **Oberg M**, Jaakkola MS, Woodward A, Peruga A, Prüss-Ustün A. Worldwide burden of disease from exposure to second-hand smoke: a retrospective analysis of data from 192 countries. *Lancet* 2011; **377**: 139-146 [PMID: 21112082 DOI: 10.1016/S0140-6736(10)61388-8.]
- 63 **Sheikh A**, Alves B, Dhami S. Pneumococcal vaccine for asthma. *Cochrane Database Syst Rev* 2002; **(1)**: CD002165 [PMID: 11869626 DOI: 10.1002/14651858.CD002165]
- 64 Updated recommendations for prevention of invasive pneumococcal disease among adults using the 23-valent pneumococcal polysaccharide vaccine (PPSV23). *MMWR Morb Mortal Wkly Rep* 2010; **59**: 1102-1106 [PMID: 20814406]
- 65 **Pebody RG**, Leino T, Nohynek H, Hellenbrand W, Salmaso S, Ruutu P. Pneumococcal vaccination policy in Europe. *Euro Surveill* 2005; **10**: 174-178 [PMID: 16280609]
- 66 **Osur SL**. The management of asthma and rhinitis during pregnancy. *J Womens Health (Larchmt)* 2005; **14**: 263-276 [PMID: 15857273 DOI: 10.1089/jwh.2005.14.263]
- 67 **Dombrowski MP**. Asthma and pregnancy. *Obstet Gynecol* 2006; **108**: 667-681 [PMID: 16946229]
- 68 **Demoly P**, Piette V, Daures JP. Treatment of allergic rhinitis during pregnancy. *Drugs* 2003; **63**: 1813-1820 [PMID: 12921487]
- 69 **Namazy JA**, Schatz M. Asthma and rhinitis during pregnancy. *Mt Sinai J Med* 2011; **78**: 661-670 [PMID: 21913197 DOI: 10.1002/msj.20284]
- 70 **Pasternak B**, Hviid A. Use of proton-pump inhibitors in early pregnancy and the risk of birth defects. *N Engl J Med* 2010; **363**: 2114-2123 [PMID: 21105793 DOI: 10.1056/NEJMoa1002689]
- 71 **Majithia R**, Johnson DA. Are proton pump inhibitors safe during pregnancy and lactation? Evidence to date. *Drugs* 2012; **72**: 171-179 [PMID: 22239714 DOI: 10.2165/11597290]
- 72 **Lin S**, Munsie JP, Herdt-Losavio ML, Druschel CM, Campbell K, Browne ML, Romitti PA, Olney RS, Bell EM. Maternal asthma medication use and the risk of selected birth defects. *Pediatrics* 2012; **129**: e317-e324 [PMID: 22250027 DOI: 10.1542/peds.2010-2660]
- 73 **Källén B**. Maternal asthma and use of antiasthmatic drugs in early pregnancy and congenital malformations in the offspring. *J Pulm Respir Med* 2014; **4**: 166 [DOI: 10.4172/2161-105X.1000166]
- 74 **Eltonsy S**, Forget A, Blais L. Beta2-agonists use during pregnancy and the risk of congenital malformations. *Birth Defects Res A Clin Mol Teratol* 2011; **91**: 937-947 [PMID: 21948561 DOI: 10.1002/bdra.22850]
- 75 **Eltonsy S**, Kettani FZ, Blais L. Beta2-agonists use during pregnancy and perinatal outcomes: a systematic review. *Respir Med* 2014; **108**: 9-33 [PMID: 24360293 DOI: 10.1016/j.rmed.2013.07.009]
- 76 **Bakhireva LN**, Jones KL, Schatz M, Johnson D, Chambers CD. Asthma medication use in pregnancy and fetal growth. *J Allergy Clin Immunol* 2005; **116**: 503-509 [PMID: 16159616 DOI: 10.1016/j.jaci.2005.05.027]
- 77 **Lin S**, Munsie JP, Herdt-Losavio ML, Bell E, Druschel C, Romitti PA, Olney R. Maternal asthma medication use and the risk of gastroschisis. *Am J Epidemiol* 2008; **168**: 73-79 [PMID: 18436535 DOI: 10.1093/aje/kwn098]
- 78 **Lin S**, Herdt-Losavio M, Gensburg L, Marshall E, Druschel C. Maternal asthma, asthma medication use, and the risk of congenital heart defects. *Birth Defects Res A Clin Mol Teratol* 2009; **85**: 161-168 [PMID: 19067406 DOI: 10.1002/bdra.20523]
- 79 **Tamási L**, Somoskövi A, Müller V, Bártfai Z, Acs N, Puhó E,

- Czeizel AE. A population-based case-control study on the effect of bronchial asthma during pregnancy for congenital abnormalities of the offspring. *J Asthma* 2006; **43**: 81-86 [PMID: 16448971]
- 80 **Munsie JW**, Lin S, Browne ML, Campbell KA, Caton AR, Bell EM, Rasmussen SA, Romitti PA, Druschel CM. Maternal bronchodilator use and the risk of orofacial clefts. *Hum Reprod* 2011; **26**: 3147-3154 [PMID: 21926056 DOI: 10.1093/humrep/der315]
- 81 **Kips JC**, Pauwels RA. Long-acting inhaled beta(2)-agonist therapy in asthma. *Am J Respir Crit Care Med* 2001; **164**: 923-932 [PMID: 11587972 DOI: 10.1164/ajrccm.164.6.2010107]
- 82 **Källén B**. The safety of asthma medications during pregnancy. *Expert Opin Drug Saf* 2007; **6**: 15-26 [PMID: 17181448 DOI: 10.1517/14740338.6.1.15]
- 83 **Källén B**, Otterblad Olausson P. Use of anti-asthmatic drugs during pregnancy. 3. Congenital malformations in the infants. *Eur J Clin Pharmacol* 2007; **63**: 383-388 [PMID: 17279357 DOI: 10.1007/s00228-006-0259-z]
- 84 **Cossette B**, Beauchesne MF, Forget A, Lemièrre C, Larivée P, Rey E, Blais L. Relative perinatal safety of salmeterol vs formoterol and fluticasone vs budesonide use during pregnancy. *Ann Allergy Asthma Immunol* 2014; **112**: 459-464 [PMID: 24656659 DOI: 10.1016/j.anai.2014.02.010]
- 85 Lexi-Comp, Inc. (Lexi-DrugsTM). Lexi-Comp, Inc.; February 9, 2014
- 86 **Wendel PJ**, Ramin SM, Barnett-Hamm C, Rowe TF, Cunningham FG. Asthma treatment in pregnancy: a randomized controlled study. *Am J Obstet Gynecol* 1996; **175**: 150-154 [PMID: 8694041 DOI: 10.1016/S0002-9378(96)70265-X]
- 87 **Stenius-Aarniala BS**, Hedman J, Teramo KA. Acute asthma during pregnancy. *Thorax* 1996; **51**: 411-414 [PMID: 8733495]
- 88 **Namazy J**, Schatz M, Long L, Lipkowitz M, Lillie MA, Voss M, Deitz RJ, Petitti D. Use of inhaled steroids by pregnant asthmatic women does not reduce intrauterine growth. *J Allergy Clin Immunol* 2004; **113**: 427-432 [PMID: 15007341 DOI: 10.1016/j.jaci.2003.11.046]
- 89 **Fält A**, Bengtsson T, Kennedy BM, Gyllenberg A, Lindberg B, Thorsson L, Stråndgarden K. Exposure of infants to budesonide through breast milk of asthmatic mothers. *J Allergy Clin Immunol* 2007; **120**: 798-802 [PMID: 17825891 DOI: 10.1016/j.jaci.2007.07.023]
- 90 **Silverman M**, Sheffer A, Diaz PV, Lindmark B, Radner F, Broddene M, de Verdier MG, Pedersen S, Pauwels RA. Outcome of pregnancy in a randomized controlled study of patients with asthma exposed to budesonide. *Ann Allergy Asthma Immunol* 2005; **95**: 566-570 [PMID: 16400897 DOI: 10.1016/S1081-1206(10)61020-4]
- 91 **Gluck PA**, Gluck JC. A review of pregnancy outcomes after exposure to orally inhaled or intranasal budesonide. *Curr Med Res Opin* 2005; **21**: 1075-1084 [PMID: 16004676 DOI: 10.1185/030079905X50570]
- 92 **Källén B**, Rydhstroem H, Aberg A. Congenital malformations after the use of inhaled budesonide in early pregnancy. *Obstet Gynecol* 1999; **93**: 392-395 [PMID: 10074986]
- 93 **Dombrowski MP**, Brown CL, Berry SM. Preliminary experience with triamcinolone acetonide during pregnancy. *J Matern Fetal Med* 1996; **5**: 310-313 [PMID: 8972405]
- 94 **Schatz M**, Zeiger RS, Harden K, Hoffman CC, Chilingar L, Petitti D. The safety of asthma and allergy medications during pregnancy. *J Allergy Clin Immunol* 1997; **100**: 301-306 [PMID: 9314340 DOI: 10.1016/S0091-6749(97)70241-0]
- 95 **Norjavaara E**, de Verdier MG. Normal pregnancy outcomes in a population-based study including 2,968 pregnant women exposed to budesonide. *J Allergy Clin Immunol* 2003; **111**: 736-742 [PMID: 12704351 DOI: 10.1067/mai.2003.1340]
- 96 **Alexander S**, Dodds L, Armson BA. Perinatal outcomes in women with asthma during pregnancy. *Obstet Gynecol* 1998; **92**: 435-440 [PMID: 9721785]
- 97 **Lim A**, Stewart K, König K, George J. Systematic review of the safety of regular preventive asthma medications during pregnancy. *Ann Pharmacother* 2011; **45**: 931-945 [PMID: 21712513 DOI: 10.1345/aph.1P764]
- 98 **Blais L**, Beauchesne MF, Lemièrre C, Elftouh N. High doses of inhaled corticosteroids during the first trimester of pregnancy and congenital malformations. *J Allergy Clin Immunol* 2009; **124**: 1229-1234.e4 [PMID: 19910032 DOI: 10.1016/j.jaci.2009.09.025]
- 99 **Stenius-Aarniala B**, Piirilä P, Teramo K. Asthma and pregnancy: a prospective study of 198 pregnancies. *Thorax* 1988; **43**: 12-18 [PMID: 2895502]
- 100 **Perlow JH**, Montgomery D, Morgan MA, Towers CV, Porto M. Severity of asthma and perinatal outcome. *Am J Obstet Gynecol* 1992; **167**: 963-967 [PMID: 1415433]
- 101 **Park-Wyllie L**, Mazzotta P, Pastuszak A, Moretti ME, Beique L, Hunnisset L, Friesen MH, Jacobson S, Kasapinovic S, Chang D, Diav-Citrin O, Chitayat D, Nulman I, Einarson TR, Koren G. Birth defects after maternal exposure to corticosteroids: prospective cohort study and meta-analysis of epidemiological studies. *Teratology* 2000; **62**: 385-392 [PMID: 11091360]
- 102 **Pradat P**, Robert-Gnansia E, Di Tanna GL, Rosano A, Lisi A, Mastroiacovo P. First trimester exposure to corticosteroids and oral clefts. *Birth Defects Res A Clin Mol Teratol* 2003; **67**: 968-970 [PMID: 14745915 DOI: 10.1002/bdra.10134]
- 103 **Sarkar M**, Koren G, Kalra S, Ying A, Smorlesi C, De Santis M, Diav-Citrin O, Avgil M, Lavigne SV, Berkovich M, Einarson A. Montelukast use during pregnancy: a multicentre, prospective, comparative study of infant outcomes. *Eur J Clin Pharmacol* 2009; **65**: 1259-1264 [PMID: 19707749 DOI: 10.1007/s00228-009-0713-9]
- 104 **Bakhireva LN**, Jones KL, Schatz M, Klonoff-Cohen HS, Johnson D, Slymen DJ, Chambers CD. Safety of leukotriene receptor antagonists in pregnancy. *J Allergy Clin Immunol* 2007; **119**: 618-625 [PMID: 17336611 DOI: 10.1016/j.jaci.2006.12.618]
- 105 **Weinberger M**, Hendeles L. Theophylline in asthma. *N Engl J Med* 1996; **334**: 1380-1388 [PMID: 8614425]
- 106 **Dombrowski MP**, Bottoms SF, Boike GM, Wald J. Incidence of preeclampsia among asthmatic patients lower with theophylline. *Am J Obstet Gynecol* 1986; **155**: 265-267 [PMID: 3740136]
- 107 **Dombrowski MP**, Schatz M, Wise R, Thom EA, Landon M, Mabie W, Newman RB, McNellis D, Hauth JC, Lindheimer M, Caritis SN, Leveno KJ, Meis P, Miodovnik M, Wapner RJ, Varner MW, O'Sullivan MJ, Conway DL. Randomized trial of inhaled beclomethasone dipropionate versus theophylline for moderate asthma during pregnancy. *Am J Obstet Gynecol* 2004; **190**: 737-744 [PMID: 15042007 DOI: 10.1016/j.ajog.2003.09.071]
- 108 **U.S. Food and Drug Administration**. Drug development and approval process: pregnancy and lactation labeling. Available from: URL: <http://www.fda.gov/Drugs/DevelopmentApprovalProcess/DevelopmentResources/Labeling/ucm093307.htm>
- 109 **Lim AS**, Stewart K, Abramson MJ, George J. Management of asthma in pregnant women by general practitioners: a cross sectional survey. *BMC Fam Pract* 2011; **12**: 121 [PMID: 22047491 DOI: 10.1186/1471-2296-12-121]
- 110 **Namazy JA**, Murphy VE, Powell H, Gibson PG, Chambers C, Schatz M. Effects of asthma severity, exacerbations and oral corticosteroids on perinatal outcomes. *Eur Respir J* 2013; **41**: 1082-1090 [PMID: 22903964 DOI: 10.1183/09031936.00195111]
- 111 **Tan KS**, Thomson NC. Asthma in pregnancy. *Am J Med* 2000; **109**: 727-733 [PMID: 11137489 DOI: 10.1016/S0002-9343(00)00615-X]
- 112 **U.S. Food and Drug Administration**. Content and Format of Labeling for Human Prescription Drug and Biological Products; Requirements for Pregnancy and Lactation Labeling, 73 Fed. Reg. 30831-68 (May 29, 2008) Available from: URL: <https://www.federalregister.gov/regulations/0910->

AF11/content-and-format-of-labeling-for-human-prescription-drugs-and-biologics-requirements-for-pregnancy
113 **Ramoz LL**, Patel-Shori NM. Recent changes in pregnancy

and lactation labeling: retirement of risk categories. *Pharmacotherapy* 2014; **34**: 389-395 [PMID: 24390829 DOI: 10.1002/phar.1385]

P- Reviewer: Dong L, Lee SH **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ



Targeted approaches for HER2 breast cancer therapy: News from nanomedicine?

Serena Mazzucchelli, Marta Truffi, Luisa Fiandra, Luca Sorrentino, Fabio Corsi

Serena Mazzucchelli, Luisa Fiandra, "L. Sacco" University Hospital, 20157 Milan, Italy

Marta Truffi, Fabio Corsi, Department of Biomedical and Clinical Sciences "L. Sacco", University of Milan, 20157 Milan, Italy
Luca Sorrentino, Fabio Corsi, Surgery Division, "Luigi Sacco" Hospital, 20157 Milan, Italy

Author contributions: All authors contributed to this paper.

Correspondence to: Fabio Corsi, MD, Professor, Department of Biomedical and Clinical Sciences "L. Sacco", University of Milan, via G. B. Grassi, 74, 20157 Milan, Italy. fabio.corsi@unimi.it

Telephone: +39-2-39043449 Fax: +39-2-50319846

Received: July 28, 2014 Revised: August 29, 2014

Accepted: September 23, 2014

Published online: December 9, 2014

Abstract

About 30% of human breast cancers are human epidermal growth factor receptor 2 (HER2)⁺. This particular biological portrait is characterized by the overexpression of HER2 receptor with the subsequent deregulation of downstream pathways, which control cellular survival and proliferation. The most effective treatment for HER2⁺ cancer is represented by therapy with HER2-targeted agents. Anti-HER2 therapy dramatically improves clinical outcomes, although it shows some limitations in achieving a proper treatment. These drawbacks of HER2-targeted therapy may be overcome with the development of HER2-targeted drug delivery nanodevices. These nanoparticles possess an internal three-dimensional compartmentalization, which allows to combine the specific target recognition with their capability to act as a drug reservoir for the selective delivery of chemotherapeutics to tumor sites. Moreover, nanoparticles useful in photothermal ablation or in photodynamic therapy have been functionalized in order to match specificity in tumor cell recognition and suitable chemical properties. Here, we summarize the state of the art concerning the HER2⁺ breast cancer and anti-HER2 therapy, in particular deepening the contribution of the nanomedicine. Description of preclinical studies performed with HER2-targeted na-

noparticles for HER2⁺ breast cancer therapy will be preceded by an overview on HER2-targeting molecules and nano-conjugation strategies. Further investigation will be necessary to introduce these nano-drugs in clinical practice; however promising results encourage an upcoming translation of this research for the next future.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Human epidermal growth factor receptor 2; Human epidermal growth factor receptor 2⁺-breast cancer; Nanomedicine; Nanoparticle; Targeted-therapy

Core tip: About 30% of human breast cancers are characterized by the overexpression of human epidermal growth factor receptor 2 (HER2) receptor, which determines the deregulation of cell survival and proliferation pathways. The HER2-targeted therapy is the most effective treatment, despite some related limitations, which could be bypassed with the development of nanoparticles for HER2-targeted drug delivery, photothermal ablation or photodynamic therapy. Here, we describe HER2⁺ breast cancer features and anti-HER2 therapy, and focus on the contribution of nanomedicine in this context, by reporting HER2-targeted nanoparticles under preclinical investigations. Promising results suggest upcoming clinical application of these nanocompounds in the next future.

Mazzucchelli S, Truffi M, Fiandra L, Sorrentino L, Corsi F. Targeted approaches for HER2 breast cancer therapy: News from nanomedicine? *World J Pharmacol* 2014; 3(4): 72-85 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/72.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.72>

INTRODUCTION

The erythroblastic leukemia viral oncogene homolog (ErbB) family of receptors and associated pathways main-

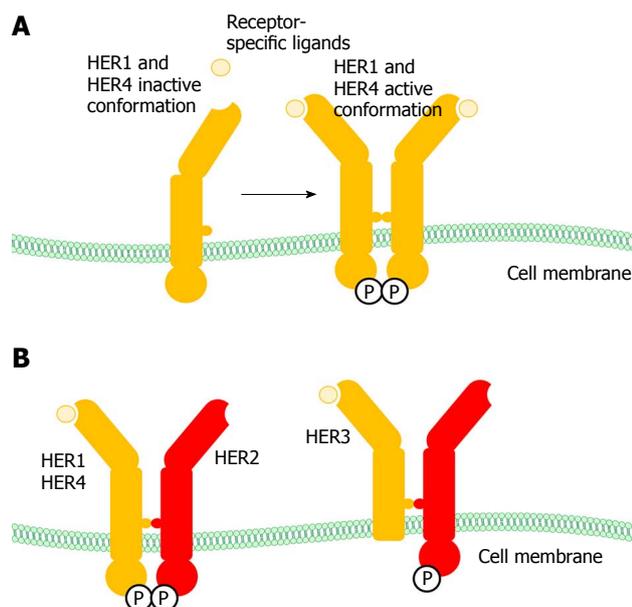


Figure 1 Characteristic features of ErbB receptors family. A: Schematic representation of HER1 and HER4 conformational change upon ligand interaction; B: Schematic representation of HER2 heterodimers. HER: Human epidermal growth factor receptor 2; ErbB: erythroblastic leukemia viral oncogene homolog.

ly regulate cell survival and proliferation. They include many actors, which cross-talk with each other and which may have overlapping functions. The typical redundancy of robust physiological processes, including the ErbB pathways, is employed by normal cells to guarantee their survival. However, it may also be highly dangerous during the early stages of tumor development, since it contributes to increase the proliferative potential of cancer cells^[1]. Actually, the main goal in clinical practice is to selectively kill the tumor cell before it can acquire the capability of metastasizing and to reduce the onset of severe side effects related to chemotherapies. Based on this rationale, several nanotechnological devices have been developed to target delivery of chemotherapeutic agents toward cancer cells in order to minimize their toxic effects on healthy tissues while increasing the antitumor efficacy^[2]. Although the number of nanoparticle strategies developed for drug delivery is increasing rapidly, they can be classified into two major groups: (1) particles containing organic molecules; and (2) particles that use inorganic elements, usually metals, as a core.

In this review, we focus on the ErbB receptor human epidermal growth factor receptor 2 (HER2) with the aim to summarize the state of the art of HER2⁺ breast cancer and related targeted therapy. In particular, we wish to explore the contribution of nanotechnology in the development of HER2-targeted nanoparticles for therapeutic purpose.

HER2 AND THE ErbB FAMILY OF PROTEINS

HER2 is a cell membrane-bound tyrosine kinase receptor that is overexpressed in 20%-30% of breast cancer

in humans. It belongs to the ErbB family of proteins, consisting of four different membrane receptors: epidermal growth factor receptor 1 (EGFR, HER1, ErbB1), 2 (HER2, ErbB2), 3 (HER3, ErbB3) and 4 (HER4, ErbB4)^[3]. Each receptor includes an extracellular domain recognized and bound by the ligand, an α -helical transmembrane portion and an intracellular tyrosine-kinase domain^[4]. Within the ErbB family there are also 13 polypeptide ligands that share the conserved epidermal growth factor (EGF) domain. The EGF family of polypeptides specifically binds the ErbB receptor and generally include three classes of proteins. The first one contains several EGFR ligands such as EGF, transforming growth factor (TGF)- α , amphiregulin and Epigen. The second group is constituted of β -cellulin, heparin binding EGF and epiregulin, which display dual specificity toward EGFR and HER4. The last group contains neuregulins (NRGs), which are divided into two subclasses depending on recognition of HER3 and HER4 (NRG-1 and NRG-2) or HER4 only (NRG-3 and NRG-4)^[5]. Generally, ErbB receptors take on an inactive conformation. The binding of the physiological ligand determines and stabilizes a conformational change that makes the dimerization domain within the extracellular portion accessible to other receptors of the family (Figure 1A). The receptor dimerization is essential for ErbB function and for activating the downstream cascade of signal transduction. Dimerization can take place between two different ErbB receptors (heterodimerization) or between two identical ErbB molecules (homodimerization). The receptor dimerization causes transactivation of the tyrosine-kinase domain by phosphorylation, so that each receptor activates its partner^[4]. In the ErbB family, only HER3 and HER2 are non-autonomous: the first one does not have intrinsic kinase activity since it is unable to bind adenosine triphosphate (ATP), whereas HER2 is an orphan receptor, since it lacks a physiological ligand^[3,4].

PHYSIOLOGICAL MECHANISM OF ACTION: FROM THE RECEPTOR TO THE PATHWAYS

Since it is an orphan receptor, HER2 is always in a constitutively active conformation, which exposes the dimerization domain to other receptors of the ErbB family. Therefore, HER2 cannot homodimerize and needs to be activated by heterodimerization with ligand-activated HER1, HER3 or HER4 (Figure 1B)^[3]. After dimerization, the cross-phosphorylation of dimer partner creates docking sites for the engagement of downstream signaling actors. Depending on the type of ligand and the type of ErbB receptor recruited by HER2, different adaptor proteins are engaged and different pathways are activated^[4]. Two key pathways are activated by HER2: the mitogen-activated protein kinase (MAPK) pathway and the phosphoinositide 3-kinase (PI3K)/Akt pathway, which promote proliferation and cell survival, respectively. The activation of the MAPK pathway is due to

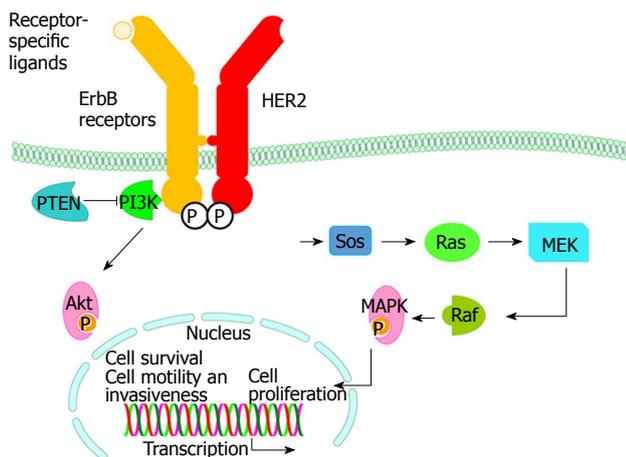


Figure 2 Schematic representation of HER2 physiological pathways. Ras: Rat sarcoma protein; Sos: Son of sevenless; PI3K: Phosphoinositide 3-kinase; MAPK: Mitogen-activated protein kinase; HER: Human epidermal growth factor receptor; ErbB: Erythroblastic leukemia viral oncogene homolog.

the recruitment and activation of the rat sarcoma protein (Ras) by the transducer molecule son of sevenless, which is in turn activated by growth-factor-receptor-bound-2 previously activated by Src-Homology-2 containing (Shc). Shc is activated upon interaction with the phosphorylated tyrosine residues within the HER2 intracellular domain. Activation of Ras kinase triggers the activation of the MAPK signaling cascade, which includes the phosphorylation of Raf, MEK and MAPK. Upon phosphorylation by MEK, MAPK translocates into the nucleus, where it regulates the transcription of genes involved in angiogenesis, proliferation and cell cycle control (Figure 2).

Differently, the PI3K pathway is activated through interaction of the phosphorylated serine or threonine residues of the receptor with the PI3K or with one of its transducer proteins, such the ubiquitin ligase Cbl. The activation of PI3K leads to the conversion of Phosphatidyl inositol 2 (PI) into PI3, with subsequent activation by phosphorylation of the Akt kinase. Once phosphorylated, Akt interacts with several transcription factors involved in cell cycle control, suppression of apoptosis and cell survival, such as mTOR, p27, nuclear factor- κ B, glycogen synthase kinase-3 β , and modulates their activation/inhibition^[4,6]. The formation of PI3 is antagonized by the phosphatase PTEN, which acts reverting PI3 in PI2. Interestingly, HER2 is also able to translocate into the nucleus, where it interacts with the cyclooxygenase-2 promoter and directly activates the transcription of specific HER2-dependent genes (Figure 2)^[7].

MOLECULAR FEATURES AND PATHOGENESIS OF HER2⁺ BREAST CANCERS

HER2⁺ breast cancer is characterized by HER2 overexpression due to Her2 gene amplification or aneuploidy in more than 90% of cases^[8]. In addition to gene amplifica-

tion and aneuploidy, HER2 overexpression may derive from transcriptional deregulation involving *cis*-acting enhancer elements near Her2 promoter or overexpression of transcription factors that bind this region^[9]. As a result of HER2 overexpression, many intracellular signaling proteins and physiological pathways are activated^[1]. Moreover, the negative regulatory loops usually active in normal cells are impaired, further contributing to pathology onset^[10]. Frequently, in HER2⁺ breast cancer the deregulation of the PI3K/Akt pathway takes place. Indeed, the PI3K activity is maintained high by the preferred interaction of HER2 with HER3. HER3 has impaired kinase activity and is unable to form homodimers but it contains six docking sites for the PI3K interaction that makes it the major PI3K activating receptor of the ErbB family. HER3 is the preferred partner of HER2 and the HER3/HER2 dimer functions as an oncogenic unit^[5]. The activation of PI3K leads to the phosphorylation and subsequent activation of Akt, which determines, among others, many important downstream effects in the oncogenic process, such as the downregulation of cyclin D1 and p27, which increase tumor cell proliferation and survival^[11]. Another typical outcome of HER2 overexpression is the hyperactivation of the MAPK pathway that results in the transcription of genes that drive cell proliferation and migration, thus conferring to tumor cells poor differentiation, invasiveness and metastatic behavior^[11,11].

Generally, HER2 overexpression is also combined with increased angiogenesis, since HER2 is able to modulate the balance between pro- and anti-angiogenic factors. In particular, high HER2 expression has been related to high levels of the pro-angiogenic molecules VEGF, IL-8 and angiopoietin-2^[11].

It has to be noted that HER2 extracellular portion is subjected to metalloproteinase cleavage, which generates a kinase-active p95 fragment. At present, it is unknown whether this activated fragment undergoes nuclear translocation and regulates HER2-dependent genes expression^[12]. Moreover, decreased levels of phosphatase expression (*e.g.*, PTEN), increased expression of ErbB receptor partners and/or their ligands^[12,13], cross-talk with other tyrosine-kinases (*e.g.*, IGF-IR) are alternative mechanisms leading to HER2 hyperactivation even in absence of HER2 overexpression^[1,3,8,14].

CLINICAL FEATURES OF HER2⁺ BREAST CANCER

As widely stated in literature, breast cancer is a heterogeneous disease and includes various subsets with distinct biological portraits. HER2⁺ breast cancer is characterized by a poor clinical outcome when anti-HER2 therapy is not administered. Notoriously, HER2 overexpression is related to lower hormonal receptor (HR) positivity, higher index of mitosis, and frequent p53 mutations. Clinical implications of these features include shorter metastasis-free and overall survival^[13]. A retrospective

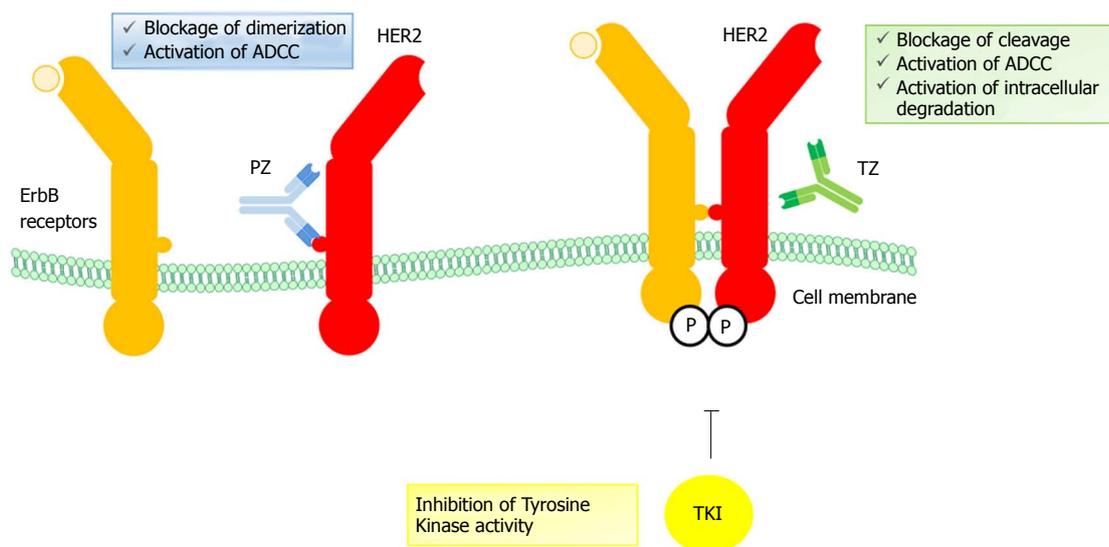


Figure 3 Schematic representation of mechanism of action of HER2-targeted drugs. PZ recognizes an epitope within the HER2 dimerization domain, thus preventing interaction with other activated ErbB receptors. Moreover PZ recruits natural killer cells, which mediate ADCC. TKI act on HER2 tyrosine kinase activity, by blocking intracellular signaling. TZ binds the juxtamembrane portion of HER2, thus preventing receptor cleavage and stimulating ADCC response and receptor degradation after endocytosis of the HER2-TZ complex. PZ: Pertuzumab; TZ: Trastuzumab; TKI: Tyrosine kinase inhibitors; ADCC: Antibody dependent cellular cytotoxicity; HER: Human epidermal growth factor receptor; ErbB: erythroblastic leukemia viral oncogene homolog.

study on 892 breast cancer patients showed a significantly higher frequency of distant metastases for HER2⁺ tumors, together with a lower 9-year disease free survival and a lower 7-year overall survival compared to the other subgroups^[16]. However, the clinical behavior of HER2⁺ cancers may also depend on HR status^[17]. A recent prospective cohort study was conducted on 3394 HER2⁺ breast tumors: among these, HR⁻ cancers more likely presented higher T stage (T3 to T4 in 17% *vs* 10%, $P < 0.001$), nodal involvement (52% *vs* 45%, $P < 0.001$), and higher histologic grade (81% *vs* 60%, $P < 0.001$)^[18]. Interestingly, HER2⁺/HR⁻ cancers were associated with more frequent brain recurrences (OR = 1.75, $P = 0.033$), and less frequent bone metastases as a first distant recurrence (OR = 0.53, $P = 0.005$), thus indicating a more aggressive disease. Therefore, HER2⁺ cancers may be divided into two distinct clinical entities based on HR status, although further studies are needed.

MANAGEMENT OF HER2⁺ BREAST CANCER

Among the different therapeutic strategies employed for HER2⁺ breast cancer, Trastuzumab (TZ) is the most widely used agent. TZ is a humanized monoclonal antibody developed starting from the murine antibody 4D5 and constituted of two antigen-binding sites that recognize the juxtamembrane portion of HER2 receptor. It functions by blocking the downstream signaling activity of HER2, thus causing cell cycle arrest and reduced angiogenesis^[4]. TZ inhibits the PI3K survival pathway by increasing PTEN membrane localization and activity, with resulting inhibition of proliferation^[14]. The inhibition of PI3K signaling may also result from HER2

internalization and degradation upon TZ interaction. However, it is still under debate whether HER2 may effectively be downregulated by TZ or not^[19,20]. Moreover, there is evidence that TZ-HER2 interaction activates the immunological response mediated by the antibody-dependent cellular cytotoxicity (ADCC), through recruitment of natural killer (NK) cells. NK cells express on their surface the Fcγ receptor IIIa, which recognizes and binds the Fc domain of TZ^[1,4,12]. In addition, the interaction of TZ with HER2 prevents the proteolytic cleavage of HER2 extracellular domain, its serum-release and the production of the truncated p95 kinase active fragment by masking the cleavage site to metalloproteinase (Figure 3)^[12,13]. At present, it is still unknown if TZ can act directly on HER2 intracellular partners^[14,21]; however TZ likely inhibits signaling downstream HER2-HER1 heterodimers^[4]. Finally, TZ treatment blocks the cell cycle in the G₁ phase, leading to reduced proliferation. This event is coupled to reduced expression of proteins involved in the sequestration of the cyclin-dependent kinase inhibitor p27KIP1, including cyclin D1. This results in increased expression of p27KIP1 protein, which causes cell cycle arrest in S phase^[1,12].

The significant improvement in overall survival and disease free survival achieved with TZ in HER2⁺ breast cancer may be considered a paradigm of the importance of targeted therapy in clinical practice, since TZ-based chemotherapy regimens have changed the clinical course of the disease. A phase II randomized clinical trial on HER2⁺ metastatic breast cancers showed that the addition of TZ resulted in a significantly improved overall response rate (61% *vs* 34%, $P = 0.0002$), and overall survival (31.2 mo *vs* 22.7 mo, $P = 0.0325$)^[22]. In a recent Cochrane systematic review on a total of 1497 patients in which TZ was administered in combination with chemotherapy,

the efficacy of TZ was confirmed, with an improved overall survival (HR = 0.82, $P = 0.004$), progression-free survival (HR = 0.61, $P < 0.00001$), and overall response rate (RR = 1.58, $P < 0.00001$), although an increased risk in congestive heart failure was evident^[23]. Because of the great efficacy as a first-line and adjuvant treatment, TZ has been successfully introduced also in the neoadjuvant setting. Interestingly, the first phase III randomized trial on neoadjuvant TZ was prematurely stopped due to the evident superiority of TZ-based chemotherapy^[24]. Other more extensive trials, such as GeparQuattro and NOAH, demonstrated similar results, with a substantial improvement of pathologic complete response rates and 5-year event-free survival in the TZ arm^[25,26].

Besides TZ, other antibodies against HER2 are currently under investigation. In particular, Pertuzumab (PZ) is a humanized monoclonal antibody that recognizes an epitope within the HER2 dimerization domain. It is able to inhibit heregulin-induced activation of HER2 phosphorylation and cell growth. Differently from TZ, PZ blocks the heterodimerization of HER2 with HER3, which is extremely relevant in tumorigenesis. However, PZ is not able to prevent the formation of EGFR-HER2 dimers, thus limiting its therapeutic efficacy^[27,28]. As observed for TZ, PZ efficacy is also mediated by the recruitment of the ADCC system (Figure 3). Because of its capability to inhibit HER2 dimerization with HER3, PZ has been approved by FDA for clinical use in association with TZ, thus helping to overcome resistance to anti-HER2 treatment. This approval has been obtained on the basis of a phase III Clinical Evaluation of PZ and TZ (CLEOPATRA) trial, in which placebo plus TZ plus docetaxel was compared to PZ plus TZ plus docetaxel for first-line treatment of 808 HER2⁺ metastatic breast cancer^[4]. Median progression-free survival was significantly higher in PZ group (18.5 mo *vs* 12.4 mo), and preliminary analysis showed also a favorable trend about overall survival. Currently other studies are investigating the role of PZ in HER2⁺ breast cancer in progression under TZ treatment, confirming a synergistic role between the two antibodies^[29].

In the last ten years, an antibody-drug conjugate, named T-DM1 (Genentech), has been developed, and it is constituted by a TZ molecule conjugated with the anti-microtubule agent DM1. TZ-DM1 recognizes HER2, is internalized and release DM-1 into the cytoplasm of HER2⁺ cells^[30]. In February 2013, T-DM1 was approved by FDA for treatment of metastatic HER2⁺ breast cancer previously treated with TZ and taxanes. The efficacy of T-DM1 in this setting has been assessed in comparison with Lapatinib on 991 patients, with an overall survival of 30.9 mo *vs* 25.1 mo ($P < 0.001$): in particular, T-DM1 was associated with a higher objective response rate (43.6% *vs* 30.8%, $P < 0.001$) and a lower toxicity profile^[31].

Another class of biological drugs for HER2-targeted therapy is represented by tyrosine-kinase inhibitors (TKI). They are small molecules that bind the ATP binding site of ErbB receptors, and prevent the activation of both

PI3K and MAPK signaling pathways, thus increasing apoptosis and reducing proliferation (Figure 3)^[4]. Among them, the most clinically advanced is Lapatinib, a dual inhibitor of HER2 and EGFR^[32]. Lapatinib has the advantage to act also on p95 activated fragment of HER2, which strongly correlates with poor prognosis^[33,34]. Lapatinib has gained a great interest for breast cancer treatment mainly for two reasons: its orally available formulation and its efficacy in the treatment of TZ-resistant metastatic HER2-positive breast cancer, with a reduction in risk of death by 26%^[35]. Moreover, Lapatinib has been recently studied in the neoadjuvant setting in association with TZ (NeoALTO Trial), with a pathologic complete response of 51.3% *vs* 29.5% with TZ alone^[36]. Other TKIs, such as HKI-272, ARRY-334543 and BIBW-2992, are under clinical investigation for breast cancer^[4].

Finally, some inhibitors of the Heat Shock Protein 90 (Hsp90) have been developed for breast cancer therapy. Indeed, Hsp90 has a role in controlling the stability of nascent and mature forms of HER2. Inhibition of its activity results in HER2 ubiquitination and subsequent proteasomal degradation, thus blocking HER2 downstream signaling pathway^[3]. A phase II trial has been conducted on 31 patients with HER2⁺ breast cancer in progression after TZ treatment, subsequently treated with the Hsp90 inhibitor tanespimycin: the objective response rate was 22% with a progression-free survival of 6 mo, therefore demonstrating the efficacy of the drug against this subset of breast cancer^[37]. However, tanespimycin has been suspended for further clinical studies, and other novel Hsp90 inhibitors are currently studied.

ONSET OF RESISTANCE

Frequently, HER2⁺ cancers develop resistance to HER2-targeted therapies^[38]. In particular, the development of resistance toward the widely used TZ has been extensively examined. Generally, resistance to TZ occurs because of three different mechanisms: (1) epitope masking; (2) upregulation of HER2 signaling; and (3) alterations of the immune response^[39]. As regards to epitope masking, two candidates have been identified: Mucin 4 (MUC4) and the CD44/hyaluronan polymer complex. MUC4 is an O-glycosylated membrane-associated protein, which is upregulated in TZ-resistant JIMT-1 cells. Binding of TZ to HER2 was reduced in JIMT-1, while it was restored after knockdown of MUC4^[40]. A similar result was observed with the CD44/hyaluronan polymer complex, where knockdown of CD44 or chemical inhibition of hyaluronan synthesis restored TZ-HER2 recognition in JIMT-1 cells. In both cases, the TZ-resistance is probably due to the steric hindrance of the complex that prevents TZ binding and internalization, without altering HER2 signaling^[41]. Upregulation of HER2-signaling is another mechanism found to bypass TZ hurdle. It results from the overexpression of some ErbB family members and the subsequent increase in heterodimer formation. Indeed, in presence of an excess of ErbB ligands the

resulting heterodimers drive cells towards proliferation and inhibition of apoptosis, thus interfering with TZ action^[42]. However, the HER2/HER1 complex may also undergo antibody-induced internalization, ubiquitination, and proteolysis, that disable its transforming activity^[1]. Moreover, up to 30% of HER2⁺ breast cancers express p95, an amino-terminal truncated form of HER2. Since p95 is a constitutively active kinase lacking the TZ binding site, it is able to confer TZ resistance^[43]. In this situation, treatment with PZ or with one of TKIs may replace responsiveness to anti-HER2 therapy^[44]. Another mechanism to bypass the TZ-mediated blockade of HER2 signaling is the activation of downstream effectors by alternative routes, *e.g.*, *via* the insulin-like growth factor 1 receptor (IGF-1R) or c-Met, often overexpressed in TZ-resistant cells, and able to hyperactivate the PI3K/Akt pathway. Treatments with inhibitors of IGF-1R or c-Met may restore TZ sensitivity^[18,45]. Decreased expression of the Akt inhibitor PTEN is another crucial factor in TZ resistance. TZ upregulates the microRNA miRNA-21, a physiological inhibitor of PTEN phosphatase^[46]. The reduction of PTEN expression maintains Akt active, and diminishes TZ efficacy^[14]. Moreover, the hyperactivity of the PI3K pathway causes epigenetic changes, which result in the inhibition of FoxO, the transcription of antiapoptotic genes^[47] and the downregulation of p27KIP1^[44]. Finally, the alteration of the immune response may cause TZ resistance in tumor cells. It is well known that TZ treatment induces ATCC, which triggers tumor cell death^[48]. It exists a FcγRIIIa polymorphism, which makes it less effective at inducing ATCC. This mechanism of resistance is common to both TZ and PZ treatments^[49].

Besides these three main mechanisms, other minor ones are involved in TZ resistance and have to be taken into account. These concern the discovery of HER2 mutants with modulated receptor activities, and subsequent more aggressive tumor phenotype^[16], and the increased activation of Notch receptors upon TZ or Lapatinib treatment, which contributes to the development of resistance^[50,51].

To overcome the previously described mechanisms of resistance to TZ a new agent against HER2⁺ cancers, called Neratinib, is being investigated for clinical use. Neratinib is a pan-HER irreversible TKI, also available for oral administration, and ErbB2 mutations were found to be sensitive to Neratinib in some preclinical studies^[52]. In a phase II trial on patients with or without previous treatment with TZ, Neratinib was administered daily at 240 mg dosage. The 16-wk progression-free survival rates was 59% for patients with prior TZ and 78% for the other group of patients and the most common adverse event was diarrhea^[53]. Interestingly, Neratinib was recently administered in combination with weekly paclitaxel and TZ in a phase I trial on metastatic HER2⁺ positive cancers previously treated with TZ, Lapatinib or T-DM1, with an objective response in 38% of patients and a median time to progression of 3.7 mo, therefore suggesting that dual anti-HER blockade with Neratinib and TZ may be more

effective than single-agent inhibition^[54]. Therefore Neratinib represents a promising tool for HER2⁺ TZ-resistant breast tumors, and a phase III trial comparing Neratinib plus Capecitabine and Lapatinib plus Capecitabine in metastatic HER2⁺ breast cancer is ongoing^[55].

THERAPEUTIC IMPROVEMENT FROM NANOTECHNOLOGY

Over the last thirty years, we all have witnessed the great development of nanotechnologies. In particular, nanomedicine has shown promising scenarios for clinical practice with the development of more effective, less toxic and smart therapeutics^[56]. The novel field of nanoncology was created and several nanodevices for tumors treatment have been developed in order to overcome limitations of conventional therapies. Indeed, chemotherapy lacks of selectivity toward tumor cells and therefore it is highly toxic toward healthy tissues. It has limited accessibility to the tumor tissues, and requires high doses to be efficient^[2]. Moreover, conventional chemotherapies are usually unable to cross biological barriers, thus bearing limited efficacy at several metastatic sites^[57]. Nanoparticles possess physical and chemical properties suitable for molecular and cellular interactions, partially due to their high surface-to-volume ratio. Moreover, their capability to form internal 3D nanostructures gives them the appropriate flexibility to be exploited as drug delivery devices able to overcome biological barriers, and to transport hydrophobic and poorly water-soluble drugs. Nanoparticles can be designed to have a large therapeutic payload and to be applied in combinatorial therapy since they can accommodate multiple drugs. Moreover, nanoparticles protect embedded drugs, thus allowing in certain cases to overcome drug resistance, which is crucial for effectiveness of cancer treatment. Finally, nanoparticle surface can be engineered with antibodies, peptides or other biologically active molecules in order to achieve a selective targeting of tumor malignancies^[58].

In the next paragraphs, we will overview the ligands and the conjugation employed for the development of HER2-targeted nanoparticles, and we will report some nanotechnological approaches for the targeted-therapy of HER2⁺ breast cancer.

HER2-TARGETED LIGANDS FOR NANOPARTICLES BIOENGINEERING

An active targeting strategy relies on the coupling of a targeting moiety to the surface of nanoparticles, thus providing specific binding to cancer biomarkers overexpressed at the target site. Such a targeting mechanism increases specific recognition of tumor cells and internalization of the nanocomplex through receptor-mediated endocytosis^[59-61]. The influence of a targeting molecule on the pharmacokinetics, biodistribution and tumor accumulation of nanoparticles depends on several

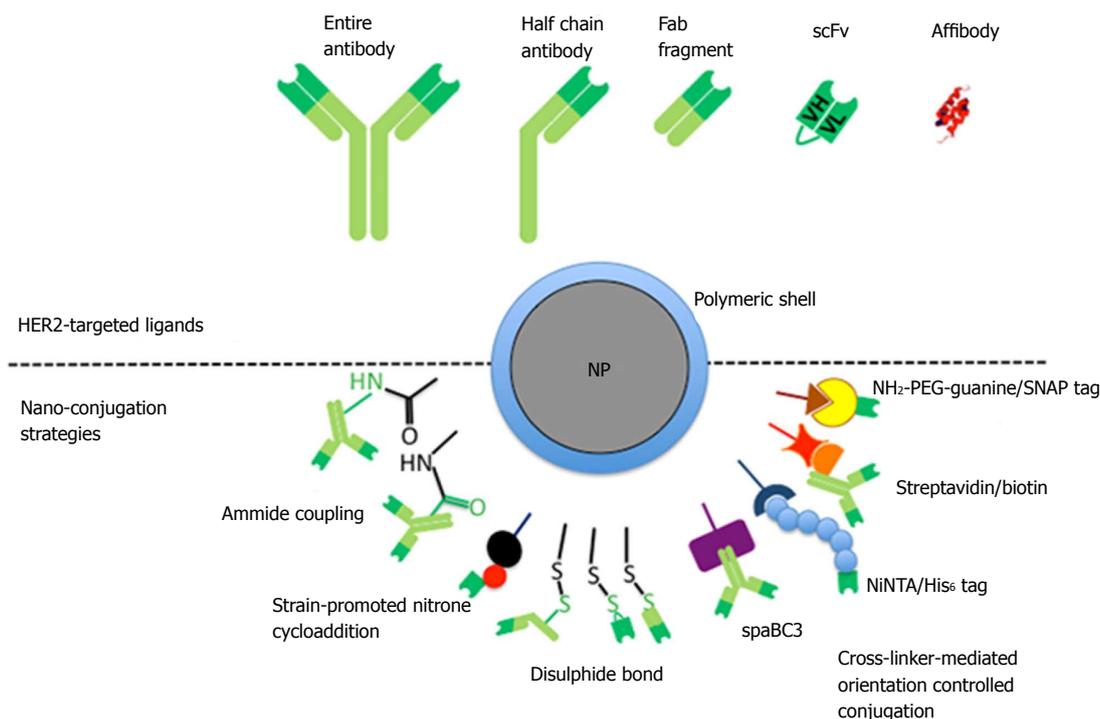


Figure 4 Schematic representation of HER2-targeting ligands and conjugation strategies employed for NPs functionalization. HER: Human epidermal growth factor receptor; scFv: Single-chain fragment variable antibodies.

factors, including the nature of the ligand, its density on the surface of the nanoparticle and its activity^[62]. Targeting moieties exploited for nanoparticle functionalization include peptides, proteins, oligonucleotides, aptamers, carbohydrates, lipids, and other biologically active molecules. Among them monoclonal antibodies and antibody-derived ligands, are widely used (Figure 4). Indeed, antibody-based therapy has received wide attention because of its stability to selectively target tumor cells through receptor-specific interactions^[63-65]. The selective tumor targeting capability of antibodies can be exploited in nanotechnology by covalently coupling antibodies directed against HER2 to the surface of colloidal nanoparticles, thus achieving higher cellular uptake and improved antitumor efficacy of nanoformulated drugs. In the context of HER2-targeted nano-therapy the most studied example of nanoparticle-conjugated ligand is TZ, a humanized monoclonal antibody already used as single agent after chemotherapy or in combination with chemotherapy in HER2-overexpressing metastatic breast cancer treatment. TZ has been used as a targeting moiety to be conjugated onto the surface of different nanoparticles, including quantum dots, magnetic and gold nanoparticles, in order to achieve selective recognition of HER2⁺ tumor cells for both imaging and therapeutic applications, with promising results obtained in preclinical studies on HER2⁺ breast cancer-bearing animal models. Different kinds of TZ-functionalized nanoparticles have been extensively reported^[1,66]. However, conjugating entire antibodies onto nanoparticles may lead to increased immunogenicity of the resulting nano-compound and reduced circulation time and tissue penetration^[64]. Recombinant

antibodies with small size have been developed in order to overcome such problems. Nano-conjugation of the half-chain of the monoclonal antibody TZ dramatically improves the intracellular trafficking and the long-term stability of the nano-compound in both *in vitro* and *in vivo* settings^[67]. Anti-HER2 Fab fragment of the monoclonal antibody TZ has also been shown to enhance tumor cell uptake resulted from HER2-mediated internalization of HER2-targeted liposomes^[68,69]. Innovative and intriguing ligands, which have provided promising results, are single-chain fragment variable antibodies (scFv), variable V_H and V_L regions of antibodies connected through a synthetic loop^[63]. Anti-HER2 scFv immobilized onto the surface of magnetic nanoparticles has proved to be highly effective in selectively targeting HER2⁺-breast cancer cells, and has shown faster cellular interaction and incorporation of nanoparticles when compared to entire TZ ligand^[67]. A number of nanoparticles have also been functionalized with HER2 affibody molecules, small proteins mimicking the active portion of the Fab region of TZ^[70,71]. Nanoparticle-affibody conjugates have shown highly specific targeting and efficiency toward HER2⁺-breast cancer, thus representing another promising class of targeting ligands with simple, robust, and precise structure and high affinity.

Besides antibodies and antibody-derived ligands, other active molecules directed toward HER2 have been proposed as interesting ligands for nano-formulation. In particular, Lapatinib is a dual inhibitor of the tyrosine kinase receptors EGFR and HER2 used to treat advanced breast cancers, and its poor water solubility has been overcome by conjugation with lipoprotein-like nanopar-

ticles (LTNPs). Such nano-compounds could be taken up by breast tumor cells by endosomes through clathrin-dependent pinocytosis and macropinocytosis, with subsequent escape from endosomes to the cytoplasm. Within tumor cells, LTNPs induce a significant cell arrest at G₀/G₁ phase compared with equal concentrations of classical lapatinib. They also could passively accumulate into the tumor *in vivo* via the enhanced permeability and retention effect where they induce elevated anti-tumor activity^[72,73].

NANO-CONJUGATION OF HER2-TARGETED LIGANDS

When designing nano-devices for targeted treatment, a crucial issue concerns the optimization of functionalization strategies to achieve an efficient and specific targeting. The structural features of a nano-compound may affect its biological functions; hence many efforts have been focused on development of new strategies for nanoparticle surface bioengineering (Figure 4). In particular, fine control of positioning, spatial orientation and conservation of the activity of targeting biomolecules have revealed essential for the generation of nano-compounds with well-defined and reproducible properties^[74,75]. Reliable conjugation strategies include physical adsorption and formation of covalent chemical connections, often through coupling with appropriate crosslinkers^[75,76]. Physical adsorption is usually related to protein ligands destabilization. Moreover, ligand orientation, number of immobilized molecules and bond stability are completely out of control. Instead the covalent coupling between the ligand and the nanoparticle gives some advantages in terms of stability of the ligand conjugation and versatility of the conjugation strategy. Indeed, chemical properties of nanoparticles have sometimes to be modulated with different functionalities depending on the functional groups found on the targeting ligands. Frequently superficial amino and carboxylic groups on the surface of nanoparticles are employed for amide coupling, thus obtaining covalent binding between the ligand and the biocompatible polymers coating the nanoparticles surface. Cysteine residues have also been found as preferred conjugation sites on proteins in general, and further exploited for HER2-targeted ligands bioconjugation. Such cysteines, either naturally present in the polypeptide sequence or introduced at specific positions by site-directed mutagenesis in case of recombinant ligands, can be activated with reducing agents and used to form disulfide bonds with properly modified nanoparticles surface^[77,78]. Traditionally, poly ethylene glycol (PEG) or poly ethylene oxide molecules are used to coat nanoparticles surface in order to reduce eventual aspecific interactions of the nanoparticle with the cells and function as spacer. Anti-HER2 antibodies have been conjugated to PEGylated nanoparticles, by covalent attachment to superficial amino and carboxylic groups^[79-81]. Polyvinyl-pyrrolidone (PVP) and poly-D,L-lactic-co-glycolic acid (PLGA) are other clinically safe polymers used

to coat nanoparticles, which can interact with a variety of agents^[82]. Vivek *et al.*^[83] have developed TZ-conjugated PVP-PLGA nanoparticles for targeted delivery of drugs to HER2-overexpressing breast cancer cells.

Optimization of nanoparticle functionalization has led to the development of smart conjugation techniques, which allow fine-tuning of the orientation of the targeting biomolecules, in order to maintain and/or further exploit the targeting capability and the therapeutic efficacy of HER2-directed ligands^[75,84]. In several cases both TZ and the nanoparticle surface have been modified with heterobifunctional linkers, such as N-succinimidyl-3-(2-pyridyldithio)propionate, succinimidyl-4-(N-maleimidomethyl)cyclohexane-1-carboxylate and succinimidyl iodoacetate, commercially available and widely used crosslinkers for bioconjugation. They allow to determine the exact number of reactive amines on the nanoparticle surface, thus widely contributing in controlling ligand density onto the surface of the resulting nano-compound^[77,84-86]. A classical approach, although not applicable in medicine, is based on the strength of streptavidin-biotin complex. Basically biotinylated TZ reacts with streptavidin-modified nanoparticles, thus generating HER2-targeted nano-compounds^[87]. Another smart conjugation strategy consists in taking advantage of spaBC3, a monodomain variant of protein A, a natural peptide linker endowed with high affinity for IgGs. It has been used to bind iron oxide and gold nanoparticles for tight TZ immobilization through the Fc fragment, thus achieving an optimal presentation of the target-directed Fab fragments and keeping full binding capacity of the bound antibody^[88]. In several cases the use of a protein biolinker is suited for a controlled site-specific conjugation of HER2-targeted ligands and it also contribute in stabilizing nanoparticle while producing.

Recombinant ligands offer an extremely desirable versatility in terms of nanoparticle conjugation, since their polypeptide sequence can be easily genetically engineered leading to generation of useful functionalities for nanoparticles conjugation. Anti HER2 scFv were modified inserting a His-tag in N-terminal position leading to conjugation of NiNTA functionalized nanoparticles. Otherwise, the mutation of a serine residue with a cysteine within the V_H-V_L linker region or the insertion of a N-terminal serine have been probed to nanoparticle conjugation through disulphide bridges formation nanoparticles or nitrene cycloaddition^[78]. These different chemical immobilization strategies of anti-HER2 scFv have been developed and tested, thus leading to multiple scFv specific and uniform orientations on the surface of nanoparticles and demonstrated subsequent effect on the targeting efficiency of the nano-compound. Another recently explored bioconjugation approach exploits the genetic fusion between the scFv module and a small enzyme (*i.e.*, SNAP tag), which works as “capture” unit. Nanoparticles have been functionalized with a suicide inhibitor of the enzyme, allowing covalent, irreversible and specific immobilization of the scFv on the nanoparticle

Table 1 Nanoparticles for breast cancer therapy

Nanoparticle	Delivered active molecule	Activity on HER2 + breast cancer cells or tumors
TZ-polymeric NPs ^[90]	DOX	nuclear drug delivery and apoptotic effect (<i>in vitro</i> study)
TZ-PLGA-PEG NPs ^[91]	DTX	HER2 specific targeting, cellular internalization and cytotoxic activity (<i>in vitro</i> study)
anti-HER2 affibody-PLA-PEG-Mal NPs ^[70]	Ptxl	HER2 specific targeting, cellular internalization and cytotoxic activity (<i>in vitro</i> study)
rhuMAbHER2 (Fab')-PLGA NPs ^[92]	PE38KDEL	HER2 specific targeting, cellular internalization and cytotoxic activity (<i>in vitro</i> study); anti-tumor activity (<i>in vivo</i> study)
scFv-PEG-PLA NPs ^[96]	<i>siPlk1</i>	cellular internalization, Plk1 silencing, apoptotic effect (<i>in vitro</i> study); <i>siPlk1</i> accumulation in tumors and anti-tumor activity (<i>in vivo</i> study)
Herceptin-PLA-TPGS+TPGS-COOH NPs ^[97]	DTX/IOs	cellular internalization and cytotoxic activity (<i>in vitro</i> study)
anti HER2-PEG-gold NPs ^[98]	phthalocyanine	HER2 specific cytotoxic activity (<i>in vitro</i> study)

TZ: Trastuzumab; DOX: Doxorubicin; DTX: Docetaxel; PLGA: Poly-D,L-lactic-co-glycolic acid; PEG: Poly ethylene glycol; IOs: Iron oxides; HER2: Human epidermal growth factor receptor 2.

surface. In this case the immobilized molecules are fully active since the bioconjugation reaction takes place in mild conditions without affecting scFv stability^[89].

HER2-TARGETED NANOPARTICLES FOR BREAST CANCER THERAPY

In breast cancer therapy, many studies have been devoted to the development of HER2-targeted nanodevices as delivery system for chemotherapics (anthracyclines and taxanes) or other molecules exerting an anti-tumor effect (Table 1). Most of them are mainly focused on the development and characterization of bioengineered NPs for HER2⁺ breast cancer cells targeting, and have demonstrated their cytotoxic effect *in vitro*. In 2009, Shi and collaborators developed an amphiphilic copolymeric NP, where surface furan groups were used to bind, by a simple diels-alder coupling chemistry, both an anti-HER2 antibody and the chemotherapeutic doxorubicin (DOX)^[90]. The DOX-conjugated immuno-NPs were able to efficiently deliver DOX into the cytoplasm, and then into the nucleus of HER2⁺ breast cancer cells, where DOX exerts its function. This intracellular DOX accumulation was significantly higher than that measured when DOX was delivered by non-functionalized NPs. These results demonstrated, for the first time, the great prospective of a surface-conjugation strategy for the development of nanoformulated DOX, which proved to be more efficient than the conventional encapsulation for the nuclear delivery of this drug. The enhanced HER2-mediated intracellular uptake of DOX also resulted in increased apoptosis of HER2⁺ breast cancer cells, when compared to non-functionalized DOX-NPs.

In 2011, Koopaei *et al.*^[91] developed a copolymeric immuno-nanocarrier for the active delivery of docetaxel (DTX) to human breast cancer cells. DTX was encapsulated in PLGA-PEG nanoparticles functionalized with TZ. A fast and over time sustained release of DTX from NPs was first observed *in vitro*, together with a specific interaction of DTX-PLGA-TZ with HER2⁺ breast cancer cells. Cytotoxicity of HER2-targeted DTX-PLGA was

compared with that of free DTX and non-specifically targeted nanoformulates. The greatest cytotoxic effect was obtained with the immune NPs as results of their specific interaction with HER2 receptors on cancer cell surface.

Another taxane commonly used in clinical practice, the paclitaxel (Ptxl), has been nanoformulated to be actively delivered to breast cancer cells. An interesting study of Alexis and colleagues^[70] addressed the numerous drawbacks of the antibody-based approach for an efficient drug delivery to tumors, mainly related to the large hydrodynamic size of the ligand. Here, it was sponsored the use of an anti-HER2 affibody, which shows several advantages in comparison to the entire monoclonal antibody: (1) smaller size (15 kDa *vs* 150 kDa); (2) considerable distance between the functional end group and the conjugation site; and (3) high *in vitro* and *in vivo* stability. Copolymeric NPs conjugated to the anti-HER2 affibody (NPs-Affb) efficiently bound HER2⁺ cancer cells and were internalized. The cytotoxic effect of Ptxl encapsulated into targeted NP-Affb was then evaluated in comparison to that of nude NPs, non-targeted NPs (Ptxl), NPs-Affb and free Ptxl. A significant decrease of cells viability was observed both with free Ptxl and non-targeted NPs (Ptxl) after 2 h, but a further significant decrease in cell viability was obtained with NP-Affb (Ptxl).

Despite the conspicuous literature about the *in vitro* therapeutic potential of nanostructured chemotherapics, only few researchers have assessed the efficacy of biofunctionalized nanodevices *in vivo*. In 2009, Gao and collaborators decided to encapsulate the anti-cancer *Pseudomonas* exotoxin A (PE)-based immunotoxin into PLGA nanoparticles^[92]. In particular, PE38KDEL, a 38 kDa mutant form of PE, was loaded into PLGA nanoparticles targeting HER2 (PE-NP-HER), where the anti-HER2 portion was represented by a Fab' fragment of a humanized anti-HER2 monoclonal antibody (rhuMAbHER2). Once assessed that the integrity and the potent activity of PE38KDEL were maintained after encapsulation in PLGA particles, *in vitro* interaction of PE-NP-HER with HER2⁺ breast cancer was compared to that obtained with

HER2-negative cells and the cytotoxic effect on the two cell types was also evaluated. PE-NP-HER were exclusively internalized by HER2⁺ cells and a strong cytotoxicity occurred specifically in these cells. *In vivo* toxicity studies were performed upon intravenous injection of PE-NP-HER, PE-NP, PE-HER and PE38KDEL in mice. A 3-fold lower LD₅₀ (mg/Kg) and no influence on hepatic functionality were observed for PLGA-loaded PE, compared to non-encapsulated toxins. A dose-dependent inhibition of tumor growth was observed in mice injected both with PE-HER and PE-NP-HER, even though a 2-fold higher dose of the PE-HER was necessary to obtain the same effects of the nanoformulated immunotoxin.

A recent nanotechnological approach proposes the employment of NPs for the delivery of small interfering RNA (siRNA)^[93-95]. In the current year, it has been developed a nanocarrier for the delivery of siRNA targeting the gene encoding polo-like kinase 1 (Plk1)^[96]. The siPlk1 was encapsulated in a PEG-PLA shell functionalized with the anti-HER2 scFv (ScFv^{HER2}-NP^{siPlk1}), to exert an active targeting of HER2⁺ breast cancer. ScFv^{HER2}-NP^{siPlk1} were efficiently internalized by cancer cells and promoted Plk1 silencing, inducing tumor cell apoptosis. Nano-complex-mediated accumulation of siPlk1 in HER2⁺ breast tumors was also observed *in vivo*, in parallel with a dose-dependent anti-tumor efficacy: ScFv^{HER2}-NP^{siPlk1} significantly increased the inhibition of tumor growth, when compared to non-targeted NP^{siPlk1}, and allowed to reduce the active dose of injected siRNA.

Nanotechnology has found a great application also in thermal therapy where gold or magnetic NPs have proved to be very useful in triggering ablation of cancer cells. In 2012, Mi and colleagues identified a multimodal strategy for breast cancer treatment, where the chemotherapy DTX was formulated with a PLA-tocopheryl-PEG-succinate and carboxyl group-terminated TPGS (TPGS-COOH) copolymer, containing iron oxides (IOs) for hyperthermia therapy^[97]. TPGS-COOH molecules were conjugated with TZ for HER2 targeting. The *in vitro* therapeutic efficiency of these multimodal NPs was tested on HER2⁺ breast cancer cells. A stronger cytotoxic activity was observed on cells incubated with TZ-IO-NPs under the exposure to an alternating current field, or with TZ-DTX-NPs, in comparison to the corresponding non-targeted NPs.

In photodynamic therapy of cancer, irradiation with visible and/or near-infrared light induces the activation of photosensitizer drugs, able to generate reactive oxygen species and trigger apoptotic or necrotic response of target cells, thus leading to cell death. Stuchinskaya and collaborators developed a PEG-gold NP conjugated to the phthalocyanine and functionalized with an anti-HER2 antibody on PEG chains. Upon red laser irradiation a strong cytotoxic effect of these NPs was observed on HER2⁺ cells and not on HER2-negative cells^[98].

CONCLUSION

About 30% of breast cancers are associated with HER2 receptor overexpression, which strongly correlates with

a poor prognosis. Indeed, HER2 regulates several highly redundant pathways involved in cellular survival and proliferation, which are deregulated in HER2⁺ cancer. At present, conventional therapy with biological drugs, such as Trastuzumab, Pertuzumab or Lapatinib, has provided satisfactory results, although still shows some limitations in achieving a proper treatment. In this context, the development of HER2-targeted nanoparticles exploited as drug delivery systems may overcome these drawbacks. Specific HER2 ligands have been conjugated on the surface of nanoparticles, thus providing a specific recognition of HER2⁺ cancer cells. Their specific target recognition is combined with the nanoparticles capability to act as a drug reservoir for a selective delivery to tumor sites. In addition, therapeutic efficiency can be reached also by combining targeting molecules with nanoparticle useful for photothermal ablation. In this review we have extensively analysed HER2⁺ breast cancer features and related targeted therapy, particularly underlining the precious contribution that nanomedicine may provide. Moreover, we have described various molecules used to target HER2 and related nano-conjugation strategies, and provided a detailed overview of preclinical studies performed with HER2-targeted nanoparticles developed for cancer therapy. Further investigations and synergic collaborations between nanotechnologists and physicians will hopefully allow to achieve the introduction of these nano-drugs in clinical.

REFERENCES

- 1 **Colombo M**, Corsi F, Foschi D, Mazzantini E, Mazzucchelli S, Morasso C, Occhipinti E, Polito L, Proserpi D, Ronchi S, Verderio P. HER2 targeting as a two-sided strategy for breast cancer diagnosis and treatment: Outlook and recent implications in nanomedical approaches. *Pharmacol Res* 2010; **62**: 150-165 [PMID: 20117211 DOI: 10.1016/j.phrs.2010.01.013]
- 2 **Shapira A**, Livney YD, Broxterman HJ, Assaraf YG. Nanomedicine for targeted cancer therapy: towards the overcoming of drug resistance. *Drug Resist Updat* 2011; **14**: 150-163 [PMID: 21330184 DOI: 10.1016/j.drug.2011.01.003]
- 3 **Citri A**, Yarden Y. EGF-ERBB signalling: towards the systems level. *Nat Rev Mol Cell Biol* 2006; **7**: 505-516 [PMID: 16829981 DOI: 10.1038/nrm1962]
- 4 **Baselga J**, Swain SM. Novel anticancer targets: revisiting ERBB2 and discovering ERBB3. *Nat Rev Cancer* 2009; **9**: 463-475 [PMID: 19536107 DOI: 10.1038/nrc2656]
- 5 **Hynes NE**, MacDonald G. ErbB receptors and signaling pathways in cancer. *Curr Opin Cell Biol* 2009; **21**: 177-184 [PMID: 19208461 DOI: 10.1016/j.ceb.2008.12.010]
- 6 **Kau TR**, Way JC, Silver PA. Nuclear transport and cancer: from mechanism to intervention. *Nat Rev Cancer* 2004; **4**: 106-117 [PMID: 14732865 DOI: 10.1038/nrc1274]
- 7 **Chen J**, Saeki F, Wiley BJ, Cang H, Cobb MJ, Li ZY, Au L, Zhang H, Kimmey MB, Li X, Xia Y. Gold nanocages: bioconjugation and their potential use as optical imaging contrast agents. *Nano Lett* 2005; **5**: 473-477 [PMID: 15755097 DOI: 10.1021/nl047950t]
- 8 **Jin Q**, Esteva FJ. Cross-talk between the ErbB/HER family and the type I insulin-like growth factor receptor signaling pathway in breast cancer. *J Mammary Gland Biol Neoplasia* 2008; **13**: 485-498 [PMID: 19034632 DOI: 10.1007/s10911-008-9107-3]
- 9 **Freudenberg JA**, Wang Q, Katsumata M, Drebin J, Nagato-

- mo I, Greene MI. The role of HER2 in early breast cancer metastasis and the origins of resistance to HER2-targeted therapies. *Exp Mol Pathol* 2009; **87**: 1-11 [PMID: 19450579 DOI: 10.1016/j.yexmp.2009.05.001]
- 10 **Amit I**, Wides R, Yarden Y. Evolvable signaling networks of receptor tyrosine kinases: relevance of robustness to malignancy and to cancer therapy. *Mol Syst Biol* 2007; **3**: 151 [PMID: 18059446 DOI: 10.1038/msb4100195]
 - 11 **Dean-Colomb W**, Esteva FJ. Her2-positive breast cancer: herceptin and beyond. *Eur J Cancer* 2008; **44**: 2806-2812 [PMID: 19022660 DOI: 10.1016/j.ejca.2008.09.013]
 - 12 **Suzuki E**, Toi M. Improving the efficacy of trastuzumab in breast cancer. *Cancer Sci* 2007; **98**: 767-771 [PMID: 17428260 DOI: 10.1111/j.1349-7006.2007.00455.x]
 - 13 **Molina MA**, Codony-Servat J, Albanell J, Rojo F, Arribas J, Baselga J. Trastuzumab (herceptin), a humanized anti-Her2 receptor monoclonal antibody, inhibits basal and activated Her2 ectodomain cleavage in breast cancer cells. *Cancer Res* 2001; **61**: 4744-4749 [PMID: 11406546]
 - 14 **Nagata Y**, Lan KH, Zhou X, Tan M, Esteva FJ, Sahin AA, Klos KS, Li P, Monia BP, Nguyen NT, Hortobagyi GN, Hung MC, Yu D. PTEN activation contributes to tumor inhibition by trastuzumab, and loss of PTEN predicts trastuzumab resistance in patients. *Cancer Cell* 2004; **6**: 117-127 [PMID: 15324695 DOI: 10.1016/j.ccr.2004.06.022]
 - 15 **Ferrero-Poüs M**, Hacène K, Bouchet C, Le Doussal V, Tubiana-Hulin M, Spyratos F. Relationship between c-erbB-2 and other tumor characteristics in breast cancer prognosis. *Clin Cancer Res* 2000; **6**: 4745-4754 [PMID: 11156229]
 - 16 **Wang SE**, Narasanna A, Perez-Torres M, Xiang B, Wu FY, Yang S, Carpenter G, Gazdar AF, Muthuswamy SK, Arteaga CL. HER2 kinase domain mutation results in constitutive phosphorylation and activation of HER2 and EGFR and resistance to EGFR tyrosine kinase inhibitors. *Cancer Cell* 2006; **10**: 25-38 [PMID: 16843263 DOI: 10.1016/j.ccr.2006.05.023]
 - 17 **Lee HJ**, Park IA, Park SY, Seo AN, Lim B, Chai Y, Song IH, Kim NE, Kim JY, Yu JH, Ahn JH, Gong G. Two histopathologically different diseases: hormone receptor-positive and hormone receptor-negative tumors in HER2-positive breast cancer. *Breast Cancer Res Treat* 2014; **145**: 615-623 [PMID: 24820412 DOI: 10.1007/s10549-014-2983-x]
 - 18 **Vaz-Luis I**, Ottesen RA, Hughes ME, Marcom PK, Moy B, Rugo HS, Theriault RL, Wilson J, Niland JC, Weeks JC, Lin NU. Impact of hormone receptor status on patterns of recurrence and clinical outcomes among patients with human epidermal growth factor-2-positive breast cancer in the National Comprehensive Cancer Network: a prospective cohort study. *Breast Cancer Res* 2012; **14**: R129 [PMID: 23025714 DOI: 10.1186/bcr3324]
 - 19 **Valabrega G**, Montemurro F, Sarotto I, Petrelli A, Rubini P, Tacchetti C, Aglietta M, Comoglio PM, Giordano S. TGF α expression impairs Trastuzumab-induced HER2 down-regulation. *Oncogene* 2005; **24**: 3002-3010 [PMID: 15735715 DOI: 10.1038/sj.onc.1208478]
 - 20 **Austin CD**, De Mazière AM, Pisacane PI, van Dijk SM, Eigenbrot C, Sliwkowski MX, Klumperman J, Scheller RH. Endocytosis and sorting of ErbB2 and the site of action of cancer therapeutics trastuzumab and geldanamycin. *Mol Biol Cell* 2004; **15**: 5268-5282 [PMID: 15385631 DOI: 10.1091/mbc.E04-07-0591]
 - 21 **Delord JP**, Allal C, Canal M, Mery E, Rochaix P, Hennebelle I, Pradines A, Chatelut E, Bugat R, Guichard S, Canal P. Selective inhibition of HER2 inhibits AKT signal transduction and prolongs disease-free survival in a micrometastasis model of ovarian carcinoma. *Ann Oncol* 2005; **16**: 1889-1897 [PMID: 16219625 DOI: 10.1093/annonc/mdi405]
 - 22 **Marty M**, Cognetti F, Maraninchi D, Snyder R, Mauriac L, Tubiana-Hulin M, Chan S, Grimes D, Antón A, Lluch A, Kennedy J, O'Byrne K, Conte P, Green M, Ward C, Mayne K, Extra JM. Randomized phase II trial of the efficacy and safety of trastuzumab combined with docetaxel in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer administered as first-line treatment: the M77001 study group. *J Clin Oncol* 2005; **23**: 4265-4274 [PMID: 15911866 DOI: 10.1200/JCO.2005.04.173]
 - 23 **Balduzzi S**, Mantarro S, Guarneri V, Tagliabue L, Pistotti V, Moja L, D'Amico R. Trastuzumab-containing regimens for metastatic breast cancer. *Cochrane Database Syst Rev* 2014; **6**: CD006242 [PMID: 24919460 DOI: 10.1002/14651858.CD006242.pub2]
 - 24 **Buzdar AU**, Ibrahim NK, Francis D, Booser DJ, Thomas ES, Theriault RL, Puztai L, Green MC, Arun BK, Giordano SH, Cristofanilli M, Frye DK, Smith TL, Hunt KK, Singletary SE, Sahin AA, Ewer MS, Buchholz TA, Berry D, Hortobagyi GN. Significantly higher pathologic complete remission rate after neoadjuvant therapy with trastuzumab, paclitaxel, and epirubicin chemotherapy: results of a randomized trial in human epidermal growth factor receptor 2-positive operable breast cancer. *J Clin Oncol* 2005; **23**: 3676-3685 [PMID: 15738535 DOI: 10.1200/JCO.2005.07.032]
 - 25 **Untch M**, Rezai M, Loibl S, Fasching PA, Huober J, Tesch H, Bauerfeind I, Hilfrich J, Eidtmann H, Gerber B, Hanusch C, Kühn T, du Bois A, Blohmer JU, Thomssen C, Dan Costa S, Jackisch C, Kaufmann M, Mehta K, von Minckwitz G. Neoadjuvant treatment with trastuzumab in HER2-positive breast cancer: results from the GeparQuattro study. *J Clin Oncol* 2010; **28**: 2024-2031 [PMID: 20308670 DOI: 10.1200/JCO.2009.23.8451]
 - 26 **Gianni L**, Eiermann W, Semiglazov V, Lluch A, Tjulandin S, Zambetti M, Moliterni A, Vazquez F, Byakhov MJ, Lichinitser M, Climent MA, Ciruelos E, Ojeda B, Mansutti M, Bozhok A, Magazzù D, Heinzmann D, Steinseifer J, Valagussa P, Baselga J. Neoadjuvant and adjuvant trastuzumab in patients with HER2-positive locally advanced breast cancer (NOAH): follow-up of a randomised controlled superiority trial with a parallel HER2-negative cohort. *Lancet Oncol* 2014; **15**: 640-647 [PMID: 24657003 DOI: 10.1016/S1470-2045(14)70080-4]
 - 27 **Cai CQ**, Peng Y, Buckley MT, Wei J, Chen F, Liebes L, Gerald WL, Pincus MR, Osman I, Lee P. Epidermal growth factor receptor activation in prostate cancer by three novel missense mutations. *Oncogene* 2008; **27**: 3201-3210 [PMID: 18193092 DOI: 10.1038/sj.onc.1210983]
 - 28 **Gaborit N**, Larbouret C, Vallaghe J, Peyrusson F, Bascoul-Mollevi C, Crapez E, Azria D, Chardès T, Poul MA, Mathis G, Bazin H, Pèlerin A. Time-resolved fluorescence resonance energy transfer (TR-FRET) to analyze the disruption of EGFR/HER2 dimers: a new method to evaluate the efficiency of targeted therapy using monoclonal antibodies. *J Biol Chem* 2011; **286**: 11337-11345 [PMID: 21282108 DOI: 10.1020/JCO.2011.37.4207]
 - 29 **Cortés J**, Fumoleau P, Bianchi GV, Petrella TM, Gelmon K, Pivot X, Verma S, Albanell J, Conte P, Lluch A, Salvagni S, Servent V, Gianni L, Scaltriti M, Ross GA, Dixon J, Szado T, Baselga J. Pertuzumab monotherapy after trastuzumab-based treatment and subsequent reintroduction of trastuzumab: activity and tolerability in patients with advanced human epidermal growth factor receptor 2-positive breast cancer. *J Clin Oncol* 2012; **30**: 1594-1600 [PMID: 22393084]
 - 30 **Lewis Phillips GD**, Li G, Dugger DL, Crocker LM, Parsons KL, Mai E, Blättler WA, Lambert JM, Chari RV, Lutz RJ, Wong WL, Jacobson FS, Koeppen H, Schwall RH, Kenkare-Mitra SR, Spencer SD, Sliwkowski MX. Targeting HER2-positive breast cancer with trastuzumab-DM1, an antibody-cytotoxic drug conjugate. *Cancer Res* 2008; **68**: 9280-9290 [PMID: 19010901 DOI: 10.1158/0008-5472.CAN-08-1776]
 - 31 **Verma S**, Miles D, Gianni L, Krop IE, Welslau M, Baselga J, Pegram M, Oh DY, Diéras V, Guardino E, Fang L, Lu MW, Olsen S, Blackwell K. Trastuzumab emtansine for HER2-positive advanced breast cancer. *N Engl J Med* 2012; **367**:

- 1783-1791 [PMID: 23020162 DOI: 10.1056/NEJMoa1209124]
- 32 **Spector NL**, Xia W, Burris H, Hurwitz H, Dees EC, Dowlati A, O'Neil B, Overmoyer B, Marcom PK, Blackwell KL, Smith DA, Koch KM, Stead A, Mangum S, Ellis MJ, Liu L, Man AK, Bremer TM, Harris J, Bacus S. Study of the biologic effects of lapatinib, a reversible inhibitor of ErbB1 and ErbB2 tyrosine kinases, on tumor growth and survival pathways in patients with advanced malignancies. *J Clin Oncol* 2005; **23**: 2502-2512 [PMID: 15684311 DOI: 10.1200/JCO.2005.12.157]
- 33 **Pedersen K**, Angelini PD, Laos S, Bach-Faig A, Cunningham MP, Ferrer-Ramón C, Luque-García A, García-Castillo J, Parra-Palau JL, Scaltriti M, Ramón y Cajal S, Baselga J, Arribas J. A naturally occurring HER2 carboxy-terminal fragment promotes mammary tumor growth and metastasis. *Mol Cell Biol* 2009; **29**: 3319-3331 [PMID: 19364815 DOI: 10.1128/MCB.01803-08]
- 34 **Scaltriti M**, Rojo F, Ocaña A, Anido J, Guzman M, Cortes J, Di Cosimo S, Matias-Guiu X, Ramon y Cajal S, Arribas J, Baselga J. Expression of p95HER2, a truncated form of the HER2 receptor, and response to anti-HER2 therapies in breast cancer. *J Natl Cancer Inst* 2007; **99**: 628-638 [PMID: 17440164 DOI: 10.1093/jnci/djk134]
- 35 **Blackwell KL**, Burstein HJ, Storniolo AM, Rugo H, Sledge G, Koehler M, Ellis C, Casey M, Vukelja S, Bischoff J, Baselga J, O'Shaughnessy J. Randomized study of Lapatinib alone or in combination with trastuzumab in women with ErbB2-positive, trastuzumab-refractory metastatic breast cancer. *J Clin Oncol* 2010; **28**: 1124-1130 [PMID: 20124187 DOI: 10.1200/JCO.2008.21.4437]
- 36 **Baselga J**, Bradbury I, Eidtmann H, Di Cosimo S, de Azambuja E, Aura C, Gómez H, Dinh P, Fauria K, Van Dooren V, Aktan G, Goldhirsch A, Chang TW, Horváth Z, Coccia-Portugal M, Domont J, Tseng LM, Kunz G, Sohn JH, Semiglazov V, Lerzo G, Palacova M, Probachai V, Pusztai L, Untch M, Gelber RD, Piccart-Gebhart M. Lapatinib with trastuzumab for HER2-positive early breast cancer (Neo-ALTTO): a randomised, open-label, multicentre, phase 3 trial. *Lancet* 2012; **379**: 633-640 [PMID: 22257673 DOI: 10.1016/S0140-6736(11)61847-3]
- 37 **Modi S**, Stopeck A, Linden H, Solit D, Chandarlapaty S, Rosen N, D'Andrea G, Dickler M, Moynahan ME, Sugarman S, Ma W, Patil S, Norton L, Hannah AL, Hudis C. HSP90 inhibition is effective in breast cancer: a phase II trial of tanespimycin (17-AAG) plus trastuzumab in patients with HER2-positive metastatic breast cancer progressing on trastuzumab. *Clin Cancer Res* 2011; **17**: 5132-5139 [PMID: 21558407 DOI: 10.1158/1078-0432.CCR-11-0072]
- 38 **Zhang H**, Berezov A, Wang Q, Zhang G, Drebin J, Murali R, Greene MI. ErbB receptors: from oncogenes to targeted cancer therapies. *J Clin Invest* 2007; **117**: 2051-2058 [PMID: 17671639 DOI: 10.1172/JCI32278]
- 39 **Martin HL**, Smith L, Tomlinson DC. Multidrug-resistant breast cancer: current perspectives. *Breast Cancer* (Dove Med Press) 2014; **6**: 1-13 [PMID: 24648765 DOI: 10.2147/BCTT.S37638]
- 40 **Nagy P**, Friedländer E, Tanner M, Kapanen AI, Carraway KL, Isola J, Jovin TM. Decreased accessibility and lack of activation of ErbB2 in JIMT-1, a herceptin-resistant, MUC4-expressing breast cancer cell line. *Cancer Res* 2005; **65**: 473-482 [PMID: 15695389]
- 41 **Pályi-Krek Z**, Barok M, Isola J, Tammi M, Szöllosi J, Nagy P. Hyaluronan-induced masking of ErbB2 and CD44-enhanced trastuzumab internalisation in trastuzumab resistant breast cancer. *Eur J Cancer* 2007; **43**: 2423-2433 [PMID: 17911008 DOI: 10.1016/j.ejca.2007.08.018]
- 42 **Diermeier S**, Horváth G, Knuechel-Clarke R, Hofstaedter F, Szöllosi J, Brockhoff G. Epidermal growth factor receptor co-expression modulates susceptibility to Herceptin in HER2/neu overexpressing breast cancer cells via specific erbB-receptor interaction and activation. *Exp Cell Res* 2005; **304**: 604-619 [PMID: 15748904 DOI: 10.1016/j.yexcr.2004.12.008]
- 43 **Gajria D**, Chandarlapaty S. HER2-amplified breast cancer: mechanisms of trastuzumab resistance and novel targeted therapies. *Expert Rev Anticancer Ther* 2011; **11**: 263-275 [PMID: 21342044 DOI: 10.1586/era.10.226]
- 44 **Nahta R**, Yuan LX, Zhang B, Kobayashi R, Esteva FJ. Insulin-like growth factor-I receptor/human epidermal growth factor receptor 2 heterodimerization contributes to trastuzumab resistance of breast cancer cells. *Cancer Res* 2005; **65**: 11118-11128 [PMID: 16322262 DOI: 10.1158/0008-5472.CAN-04-3841]
- 45 **Shattuck DL**, Miller JK, Carraway KL, Sweeney C. Met receptor contributes to trastuzumab resistance of Her2-overexpressing breast cancer cells. *Cancer Res* 2008; **68**: 1471-1477 [PMID: 18316611 DOI: 10.1158/0008-5472.CAN-07-5962]
- 46 **Gong C**, Yao Y, Wang Y, Liu B, Wu W, Chen J, Su F, Yao H, Song E. Up-regulation of miR-21 mediates resistance to trastuzumab therapy for breast cancer. *J Biol Chem* 2011; **286**: 19127-19137 [PMID: 21471222 DOI: 10.1074/jbc.M110.216887]
- 47 **Chakrabarty A**, Bhola NE, Sutton C, Ghosh R, Kuba MG, Dave B, Chang JC, Arteaga CL. Trastuzumab-resistant cells rely on a HER2-PI3K-FoxO-survivin axis and are sensitive to PI3K inhibitors. *Cancer Res* 2013; **73**: 1190-1200 [PMID: 23204226 DOI: 10.1158/0008-5472.CAN-12-2440]
- 48 **Clynes RA**, Towers TL, Presta LG, Ravetch JV. Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets. *Nat Med* 2000; **6**: 443-446 [PMID: 10742152 DOI: 10.1038/74704]
- 49 **Musulino A**, Naldi N, Bortesi B, Pezzuolo D, Capelletti M, Missale G, Laccabue D, Zerbini A, Camisa R, Bisagni G, Neri TM, Ardizzoni A. Immunoglobulin G fragment C receptor polymorphisms and clinical efficacy of trastuzumab-based therapy in patients with HER-2/neu-positive metastatic breast cancer. *J Clin Oncol* 2008; **26**: 1789-1796 [PMID: 18347005 DOI: 10.1200/JCO.2007.14.8957]
- 50 **Osipo C**, Patel P, Rizzo P, Clementz AG, Hao L, Golde TE, Miele L. ErbB-2 inhibition activates Notch-1 and sensitizes breast cancer cells to a gamma-secretase inhibitor. *Oncogene* 2008; **27**: 5019-5032 [PMID: 18469855 DOI: 10.1038/onc.2008.149]
- 51 **Politi K**, Feirt N, Kitajewski J. Notch in mammary gland development and breast cancer. *Semin Cancer Biol* 2004; **14**: 341-347 [PMID: 15288259 DOI: 10.1016/j.semcancer.2004.04.013]
- 52 **Greulich H**, Kaplan B, Mertins P, Chen TH, Tanaka KE, Yun CH, Zhang X, Lee SH, Cho J, Ambrogio L, Liao R, Imielinski M, Banerji S, Berger AH, Lawrence MS, Zhang J, Pho NH, Walker SR, Winckler W, Getz G, Frank D, Hahn WC, Eck MJ, Mani DR, Jaffe JD, Carr SA, Wong KK, Meyererson M. Functional analysis of receptor tyrosine kinase mutations in lung cancer identifies oncogenic extracellular domain mutations of ERBB2. *Proc Natl Acad Sci USA* 2012; **109**: 14476-14481 [PMID: 22908275 DOI: 10.1073/pnas.1203201109]
- 53 **Burstein HJ**, Sun Y, Dirix LY, Jiang Z, Paridaens R, Tan AR, Awada A, Ranade A, Jiao S, Schwartz G, Abbas R, Powell C, Turnbull K, Vermette J, Zacharchuk C, Badwe R. Neratinib, an irreversible ErbB receptor tyrosine kinase inhibitor, in patients with advanced ErbB2-positive breast cancer. *J Clin Oncol* 2010; **28**: 1301-1307 [PMID: 20142587 DOI: 10.1200/JCO.2009.25.8707]
- 54 **Jankowitz RC**, Abraham J, Tan AR, Limentani SA, Tierno MB, Adamson LM, Buyse M, Wolmark N, Jacobs SA. Safety and efficacy of neratinib in combination with weekly paclitaxel and trastuzumab in women with metastatic HER2-positive breast cancer: an NSABP Foundation Research Program phase I study. *Cancer Chemother Pharmacol* 2013; **72**: 1205-1212 [PMID: 24077916]
- 55 **Mohamed A**, Krajewski K, Cakar B, Ma CX. Targeted therapy for breast cancer. *Am J Pathol* 2013; **183**: 1096-1112 [PMID: 23988612 DOI: 10.1016/j.ajpath.2013.07.005]
- 56 **Schroeder A**, Heller DA, Winslow MM, Dahlman JE, Pratt

- GW, Langer R, Jacks T, Anderson DG. Treating metastatic cancer with nanotechnology. *Nat Rev Cancer* 2012; **12**: 39-50 [PMID: 22193407 DOI: 10.1038/nrc3180]
- 57 **Esteve FJ**, Valero V, Pusztai L, Boehnke-Michaud L, Buzdar AU, Hortobagyi GN. Chemotherapy of metastatic breast cancer: what to expect in 2001 and beyond. *Oncologist* 2001; **6**: 133-146 [PMID: 11306725]
- 58 **Desai N**. Challenges in development of nanoparticle-based therapeutics. *AAPS J* 2012; **14**: 282-295 [PMID: 22407288 DOI: 10.1208/s12248-012-9339-4]
- 59 **Lammers T**, Kiessling F, Hennink WE, Storm G. Nanotheranostics and image-guided drug delivery: current concepts and future directions. *Mol Pharm* 2010; **7**: 1899-1912 [PMID: 20822168 DOI: 10.1021/mp100228v]
- 60 **Torchilin VP**. Passive and active drug targeting: drug delivery to tumors as an example. *Handb Exp Pharmacol* 2010; **(197)**: 3-53 [PMID: 20217525 DOI: 10.1007/978-3-642-00477-3_1]
- 61 **Yu MK**, Park J, Jon S. Targeting strategies for multifunctional nanoparticles in cancer imaging and therapy. *Theranostics* 2012; **2**: 3-44 [PMID: 22272217 DOI: 10.7150/thno.3463]
- 62 **Allen TM**. Ligand-targeted therapeutics in anticancer therapy. *Nat Rev Cancer* 2002; **2**: 750-763 [PMID: 12360278 DOI: 10.1038/nrc903]
- 63 **Holliger P**, Hudson PJ. Engineered antibody fragments and the rise of single domains. *Nat Biotechnol* 2005; **23**: 1126-1136 [PMID: 16151406 DOI: 10.1038/nbt1142]
- 64 **Sanz L**, Cuesta AM, Compte M, Alvarez-Vallina L. Antibody engineering: facing new challenges in cancer therapy. *Acta Pharmacol Sin* 2005; **26**: 641-648 [PMID: 15916728 DOI: 10.1111/j.1745-7254.2005.00135.x]
- 65 **Slamon DJ**, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, Fleming T, Eiermann W, Wolter J, Pegram M, Baselga J, Norton L. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N Engl J Med* 2001; **344**: 783-792 [PMID: 11248153 DOI: 10.1056/NEJM200103153441101]
- 66 **Corsi F**, Fiandra L, De Palma C, Colombo M, Mazzucchelli S, Verderio P, Allevi R, Tosoni A, Nebuloni M, Clementi E, Prosperi D. HER2 expression in breast cancer cells is down-regulated upon active targeting by antibody-engineered multifunctional nanoparticles in mice. *ACS Nano* 2011; **5**: 6383-6393 [PMID: 21790185 DOI: 10.1021/nn201570n]
- 67 **Fiandra L**, Mazzucchelli S, De Palma C, Colombo M, Allevi R, Sommaruga S, Clementi E, Bellini M, Prosperi D, Corsi F. Assessing the in vivo targeting efficiency of multifunctional nanoconstructs bearing antibody-derived ligands. *ACS Nano* 2013; **7**: 6092-6102 [PMID: 23758591 DOI: 10.1021/nn4018922]
- 68 **Hoang B**, Ekdawi SN, Reilly RM, Allen C. Active targeting of block copolymer micelles with trastuzumab Fab fragments and nuclear localization signal leads to increased tumor uptake and nuclear localization in HER2-overexpressing xenografts. *Mol Pharm* 2013; **10**: 4229-4241 [PMID: 24066900 DOI: 10.1021/mp400315p]
- 69 **Kirpotin DB**, Drummond DC, Shao Y, Shalaby MR, Hong K, Nielsen UB, Marks JD, Benz CC, Park JW. Antibody targeting of long-circulating lipidic nanoparticles does not increase tumor localization but does increase internalization in animal models. *Cancer Res* 2006; **66**: 6732-6740 [PMID: 16818648 DOI: 10.1158/0008-5472.CAN-05-4199]
- 70 **Alexis F**, Basto P, Levy-Nissenbaum E, Radovic-Moreno AF, Zhang L, Pridgen E, Wang AZ, Marein SL, Westerhof K, Molnar LK, Farokhzad OC. HER-2-targeted nanoparticle-affibody bioconjugates for cancer therapy. *ChemMedChem* 2008; **3**: 1839-1843 [PMID: 19012296 DOI: 10.1002/cmdc.200800122]
- 71 **Gao J**, Chen K, Miao Z, Ren G, Chen X, Gambhir SS, Cheng Z. Affibody-based nanoprobe for HER2-expressing cell and tumor imaging. *Biomaterials* 2011; **32**: 2141-2148 [PMID: 21147502 DOI: 10.1016/j.biomaterials.2010.11.053]
- 72 **Gao H**, Cao S, Chen C, Cao S, Yang Z, Pang Z, Xi Z, Pan S, Zhang Q, Jiang X. Incorporation of lapatinib into lipoprotein-like nanoparticles with enhanced water solubility and anti-tumor effect in breast cancer. *Nanomedicine (Lond)* 2013; **8**: 1429-1442 [PMID: 23451915 DOI: 10.2217/nmm.12.180]
- 73 **Zhang L**, Zhang S, Ruan SB, Zhang QY, He Q, Gao HL. Lapatinib-incorporated lipoprotein-like nanoparticles: preparation and a proposed breast cancer-targeting mechanism. *Acta Pharmacol Sin* 2014; **35**: 846-852 [PMID: 24902791 DOI: 10.1038/aps.2014.26]
- 74 **Algar WR**, Prasuhn DE, Stewart MH, Jennings TL, Blanco-Canosa JB, Dawson PE, Medintz IL. The controlled display of biomolecules on nanoparticles: a challenge suited to bioorthogonal chemistry. *Bioconjug Chem* 2011; **22**: 825-858 [PMID: 21585205 DOI: 10.1021/bc200065z]
- 75 **Avvakumova S**, Colombo M, Tortora P, Prosperi D. Biotechnological approaches toward nanoparticle biofunctionalization. *Trends Biotechnol* 2014; **32**: 11-20 [PMID: 24182737 DOI: 10.1016/j.tibtech.2013.09.006]
- 76 **Anand G**, Sharma S, Dutta AK, Kumar SK, Belfort G. Conformational transitions of adsorbed proteins on surfaces of varying polarity. *Langmuir* 2010; **26**: 10803-10811 [PMID: 20433160 DOI: 10.1021/la100613z]
- 77 **Yoon TJ**, Yu KN, Kim E, Kim JS, Kim BG, Yun SH, Sohn BH, Cho MH, Lee JK, Park SB. Specific targeting, cell sorting, and bioimaging with smart magnetic silica core-shell nanomaterials. *Small* 2006; **2**: 209-215 [PMID: 17193022 DOI: 10.1002/sml.200500360]
- 78 **Mazzucchelli S**, Sommaruga S, O'Donnel M, Galeffi P, Tortora P, Prosperi D, Colombo M. Dependence of nanoparticle-cell recognition efficiency on the surface orientation of scFv targeting ligands. *Biomaterial Sciences* 2013; **1**: 728-735 [DOI: 10.1039/c3bm60068h]
- 79 **Han H**, Davis ME. Single-antibody, targeted nanoparticle delivery of camptothecin. *Mol Pharm* 2013; **10**: 2558-2567 [PMID: 23676007 DOI: 10.1021/mp300702x]
- 80 **Kouchakzadeh H**, Shojaosadati SA, Tahmasebi F, Shokri F. Optimization of an anti-HER2 monoclonal antibody targeted delivery system using PEGylated human serum albumin nanoparticles. *Int J Pharm* 2013; **447**: 62-69 [PMID: 23454849 DOI: 10.1016/j.ijpharm.2013.02.043]
- 81 **Zhao J**, Feng SS. Effects of PEG tethering chain length of vitamin E TPGS with a Herceptin-functionalized nanoparticle formulation for targeted delivery of anticancer drugs. *Biomaterials* 2014; **35**: 3340-3347 [PMID: 24461325 DOI: 10.1016/j.biomaterials.2014.01.003]
- 82 **Duncan R**. The dawning era of polymer therapeutics. *Nat Rev Drug Discov* 2003; **2**: 347-360 [PMID: 12750738 DOI: 10.1038/nrd1088]
- 83 **Vivek R**, Thangam R, NipunBabu V, Rejeeth C, Sivasubramanian S, Gunasekaran P, Muthuchelian K, Kannan S. Multifunctional HER2-antibody conjugated polymeric nanocarrier-based drug delivery system for multi-drug-resistant breast cancer therapy. *ACS Appl Mater Interfaces* 2014; **6**: 6469-6480 [PMID: 24780315 DOI: 10.1021/am406012g]
- 84 **Occhipinti E**, Verderio P, Natalello A, Galbiati E, Colombo M, Mazzucchelli S, Salvadè A, Tortora P, Doglia SM, Prosperi D. Investigating the structural biofunctionality of antibodies conjugated to magnetic nanoparticles. *Nanoscale* 2011; **3**: 387-390 [PMID: 20877896 DOI: 10.1039/c0nr00436g]
- 85 **Chen H**, Wang L, Yu Q, Qian W, Tiwari D, Yi H, Wang AY, Huang J, Yang L, Mao H. Anti-HER2 antibody and ScFvEGFR-conjugated antifouling magnetic iron oxide nanoparticles for targeting and magnetic resonance imaging of breast cancer. *Int J Nanomedicine* 2013; **8**: 3781-3794 [PMID: 24124366 DOI: 10.2147/IJN.S49069]
- 86 **Shukla R**, Thomas TP, Peters JL, Desai AM, Kukowska-Latallo J, Patri AK, Kotlyar A, Baker JR. HER2 specific tumor targeting with dendrimer conjugated anti-HER2 mAb. *Bioconjug Chem* 2006; **17**: 1109-1115 [PMID: 16984117 DOI:

- 10.1021/bc050348p]
- 87 **Artemov D**, Mori N, Okollie B, Bhujwala ZM. MR molecular imaging of the Her-2/neu receptor in breast cancer cells using targeted iron oxide nanoparticles. *Magn Reson Med* 2003; **49**: 403-408 [PMID: 12594741 DOI: 10.1002/mrm.10406]
- 88 **Colombo M**, Mazzucchelli S, Collico V, Avvakumova S, Pandolfi L, Corsi F, Porta F, Prosperi D. Protein-assisted one-pot synthesis and biofunctionalization of spherical gold nanoparticles for selective targeting of cancer cells. *Angew Chem Int Ed Engl* 2012; **51**: 9272-9275 [PMID: 22833476 DOI: 10.1002/anie.201204699]
- 89 **Colombo M**, Mazzucchelli S, Montenegro JM, Galbiati E, Corsi F, Parak WJ, Prosperi D. Protein oriented ligation on nanoparticles exploiting O6-alkylguanine-DNA transferase (SNAP) genetically encoded fusion. *Small* 2012; **8**: 1492-1497 [PMID: 22431243 DOI: 10.1002/smll.201102284]
- 90 **Shi M**, Ho K, Keating A, Shoichet MS. Doxorubicin-conjugated immuno-nanoparticles for intracellular anticancer drug delivery. *Adv Funct Mater* 2009; **19**: 1-8 [DOI: 10.1002/adfm.200801271]
- 91 **Koopaei MN**, Dinarvand R, Amini M, Rabbani H, Emami S, Ostad SN, Atyabi F. Docetaxel immunonanocarriers as targeted delivery systems for HER 2-positive tumor cells: preparation, characterization, and cytotoxicity studies. *Int J Nanomedicine* 2011; **6**: 1903-1912 [PMID: 21931485 DOI: 10.2147/IJN.S23211]
- 92 **Gao J**, Kou G, Wang H, Chen H, Li B, Lu Y, Zhang D, Wang S, Hou S, Qian W, Dai J, Zhao J, Zhong Y, Guo Y. PE38KDEL-loaded anti-HER2 nanoparticles inhibit breast tumor progression with reduced toxicity and immunogenicity. *Breast Cancer Res Treat* 2009; **115**: 29-41 [PMID: 18481173 DOI: 10.1007/s10549-008-0043-0]
- 93 **Piao L**, Li H, Teng L, Yung BC, Sugimoto Y, Brueggemeier RW, Lee RJ. Human serum albumin-coated lipid nanoparticles for delivery of siRNA to breast cancer. *Nanomedicine* 2013; **9**: 122-129 [PMID: 22542825 DOI: 10.1016/j.nano.2012.03.008]
- 94 **Meng H**, Mai WX, Zhang H, Xue M, Xia T, Lin S, Wang X, Zhao Y, Ji Z, Zink JJ, Nel AE. Codelivery of an optimal drug/siRNA combination using mesoporous silica nanoparticles to overcome drug resistance in breast cancer in vitro and in vivo. *ACS Nano* 2013; **7**: 994-1005 [PMID: 23289892 DOI: 10.1021/nn3044066]
- 95 **Deng ZJ**, Morton SW, Ben-Akiva E, Dreaden EC, Shopsowitz KE, Hammond PT. Layer-by-layer nanoparticles for systemic codelivery of an anticancer drug and siRNA for potential triple-negative breast cancer treatment. *ACS Nano* 2013; **7**: 9571-9584 [PMID: 24144228 DOI: 10.1021/nn4047925]
- 96 **Dou S**, Yang X-Z, Xiong M-H, Sun C-Y, Yao Y-D, Zhu Y-H, Wang J. ScFv-Decorated PEG-PLA-Based Nanoparticles for Enhanced siRNA Delivery to Her2(+) Breast Cancer. *Adv Healthc Mater* 2014; **3**: 1792-803 [PMID: 24947820 DOI: 10.1002/adhm.201400037]
- 97 **Mi Y**, Liu X, Zhao J, Ding J, Feng SS. Multimodality treatment of cancer with herceptin conjugated, thermomagnetic iron oxides and docetaxel loaded nanoparticles of biodegradable polymers. *Biomaterials* 2012; **33**: 7519-7529 [PMID: 22809649 DOI: 10.1016/j.biomaterials.2012.06.100]
- 98 **Stuchinskaya T**, Moreno M, Cook MJ, Edwards DR, Russell DA. Targeted photodynamic therapy of breast cancer cells using antibody-phthalocyanine-gold nanoparticle conjugates. *Photochem Photobiol Sci* 2011; **10**: 822-831 [PMID: 21455532 DOI: 10.1039/c1pp05014a]

P- Reviewers: Langdon S, Zhang H **S- Editor:** Ji FF

L- Editor: A **E- Editor:** Lu YJ



Telomerase activity: An attractive target for cancer therapeutics

Lucia Picariello, Cecilia Grappone, Simone Polvani, Andrea Galli

Lucia Picariello, Cecilia Grappone, Simone Polvani, Andrea Galli, Gastroenterology Unit, Department of Experimental and Clinical Biomedical Sciences, University of Florence, 50139 Florence, Italy

Author contributions: Picariello L, Grappone C, Polvani S and Galli A contributed to this paper.

Correspondence to: Andrea Galli, MD, PhD, Professor, Gastroenterology Unit, Department of Experimental and Clinical Biomedical Sciences, University of Florence, Viale Pieraccini n°6, 50139 Florence, Italy. andrea.galli@unifi.it

Telephone: +39-5-54271419 Fax: +39-5-54271297

Received: July 28, 2014 Revised: October 1, 2014

Accepted: October 28, 2014

Published online: December 9, 2014

Abstract

Telomeres are non-coding tandem repeats of 1000-2000 TTAGGG nucleotide DNA sequences on the 3' termini of human chromosomes where they serve as protective "caps" from degradation and loss of genes. The "cap" at the end of chromosome required to protect its integrity is a 150-200 nucleotide-long single stranded G-rich 3' overhang that forms two higher order structures, a T-loop with Sheltering complex, or a G-quadruplex complex. Telomerase is a human ribonucleoprotein reverse transcriptase that continually added single stranded TTAGGG DNA sequences onto the single strand 3' of telomere in the 5' to 3' direction. Telomerase activity is detected in male germ line cells, proliferative cells of renewal tissues, some adult pluripotent stem cells, embryonic cells, but in most somatic cells is not detected. Re-expression or up-regulation of telomerase in tumours cells is considered as a critical step in cell tumorigenesis and telomerase is widely considered as a tumour marker and a target for anticancer drugs. Different approaches have been used in anticancer therapeutics targeting telomerase. Telomerase inhibitors can block directly Human Telomerase Reverse Transcriptase (hTERT) or Human Telomerase RNA telomerase subunits activity, or G-quadruplex and Sheltering com-

plex components, shortening telomeres and inhibiting cell proliferation. Telomerase can become an immune target and GV1001, Vx-001, I540 are the most widespread vaccines used with encouraging results. Another method is to use hTERT promoter to drive suicide gene expression or to control a lytic virus replication. Recently telomerase activity was used to activate pro-drugs such as Acycloguanosyl 5'-thymidyltriphosphate, a synthetic ACV-derived molecule when it is activated by telomerase it does not require any virus or host active immune response to induce suicide gene therapy. Advantage of all these therapies is that target only neoplastic cells without any effects in normal cells, avoiding toxicity and adverse effects of the current chemotherapy. However, as not all the approaches are equally efficient, further studies will be necessary.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Human telomerase reverse transcriptase; Immunotherapy; Suicide gene therapy; Acycloguanosyl 5'-thymidyltriphosphate; Telomerase inhibition

Core tip: One of the hallmark of cancer is the replicative immortality of tumor cells guaranteed by telomerase activity that counteracts progressive telomere shortening during cellular replication: this makes telomerase a tumor marker and a target for anticancer drugs. In this review we summarize and update the most recent innovative studies and results on the different strategies that consider telomerase as a target for cancer therapy. In particular, we try to point out the advantages and the potentialities of some innovative approaches, compared to other, equally promising, but that need further investigations.

Picariello L, Grappone C, Polvani S, Galli A. Telomerase activity: An attractive target for cancer therapeutics. *World J Pharmacol* 2014; 3(4): 86-96 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/86.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.86>

TELOMERES, TELOMERASE AND CANCER

Telomeres are non-coding tandem repeats of 1000-2000 TTAGGG nucleotide DNA sequences on the 3' termini of human chromosomes^[1-3] where they serve as protective "caps" from degradation and loss of genes. In this way cells can discriminate between double strand breaks and natural chromosome ends^[4,5]. In human somatic cells, telomeres become critically short after successive cell divisions (number of divisions depending on the length of their telomeres), cells stop division and replicative senescence occurs^[6]. As a consequence, telomeres can reach a critical length that is no longer suitable to assemble into T-loop: this triggers a localized DNA damage response and p53-mediated cell cycle arrest^[7-9]. However, cells that have inactivated the p53-pathway cell cycle checkpoint, are able to continue dividing, bypassing senescence, losing telomeric sequence with each division^[9,10] and reach a "crisis" stage^[11,12]. In this way telomeres become so short that cannot protect chromosome ends, so that they fuse together to produce a dicentric chromosome, inducing an increase aneuploidy and genomic instability that finally will lead to p53-independent apoptosis^[13,14]. Bypassing crisis rarely occurs in human cells (1 in 10⁻⁶ in epithelial cells and 1 in 10⁻⁷ in human fibroblasts) and this leads to cell immortality and cancer cell progression, characterized by capability to continue to proliferate without limits.

The "cap" at the end of chromosome required to protect its integrity is a 150-200 nucleotide-long single stranded G-rich 3' overhang that forms two higher order structures, a T-loop with Sheltering complex, or a G-quadruplex complex. Sheltering complex is represented by six proteins (TRF1 and TRF2, POT1, TPP1, TIN2, RAP1) responsible for maintaining the T-loop structure. G-quadruplex is stabilized with BRACO19, RHS4 and telomestatin proteins. Sheltering complex with T-loop, G-quadruplex and its stabilizers can lock the telomeric 3' overhang and block telomerase from accessing telomeres^[15] (Figure 1).

Telomerase is a human ribonucleoprotein reverse transcriptase that continually adds single stranded TTAGGG DNA sequences onto the single strand 3' of telomere in the 5' to 3' direction and translocates to the new terminus^[16,17]. This cycle goes on as far as telomerase dissociates from telomere^[18,19]. Telomerase is composed of two main subunits: the catalytic protein Human TELOmerase Reverse Transcriptase (hTERT) and the ribonucleoprotein template Human TELOmerase RNA (hTER)^[15-17]. In particular hTER consists of 451 nucleotides of which only nucleotides 46 through 56 (5'-CUAACCCUAAC-3') represent a template for new telomeric added DNA sequences (Figure 2).

Many proteins associated to the core components hTERT and hTER are required and are necessary for stability regulation, recruitment and activity of the holoenzyme^[20]. hTER is expressed in all human cells, as well as normal and tumour cells, so telomerase activity is limited

by of hTERT expression, whereas is present^[21,22].

Telomerase activity is detected in proliferative cells of renewal tissues, in some adult pluripotent stem cells, male germ line cells, embryonic cells, but not in most somatic cells^[23]. However, telomerase activity is found in almost all human cancer cell lines and in about 85%-90% of primary tumours^[24]. In fact, one of the hallmark of cancer is the replicative immortality and so the ability to endlessly growth is synonymous of telomerase reverse transcriptase reactivation. Up-regulation or re-expression of telomerase in tumour cells is considered as a critical step in cell tumorigenesis and telomerase is widely considered as a tumour marker and a target for anticancer drugs. Progressive telomere shortening during cellular replication is counteracted by telomerase activity^[1,25].

One of the advantages of anticancer therapies targeting telomerase is that the telomeres of highly proliferating cancer cells are shorter (5 kb) compared to that in normal somatic cells and stem cells (10-20 kb) that have not yet reached critical lengths as a result of aging^[26,27]. The difference in telomerase activity and telomere lengths in normal and cancer cells leads to a more selective therapeutics cytotoxicity on cancer cells and a minimal impact on normal cells with a limitation of collateral effects that can be evaluated^[28].

TELOMERASE INHIBITION AS A THERAPY

Telomerase inhibitors can be employed as a selective anticancer therapy, disrupting telomerase-positive cancer cells replicative capacity^[29].

To target telomerase in cancer treatment we can find two types of approaches: the first one is blocking directly telomerase hTERT or hTER subunits activity, with consequent shortening of telomeres leading to the arrest of cell replication. The second approach is to block telomerase by an indirect method, targeting G-quadruplex stabilizers or Sheltering complex components with the consequence of preventing telomerase interaction with telomeres or binding of proteins associated with telomerase; this leads to telomere uncapping and cell apoptosis^[30].

Antisense oligonucleotides-targeting hTER

One of the most recent strategy for a direct telomerase enzymatic inhibition, is the use of antisense oligonucleotides inhibitors. These molecules are complementary to the 11-base template region of telomerase (hTER) and can be used to block the translation of sense RNA. In order to hybridize the hTER-template the antisense oligonucleotides must get to the hTER region without being degraded by nucleases. For this reason the challenge for this kind of drugs is both access and stability. To better get its target, antisense oligonucleotides have been modified and significantly improved in the past years.

Currently GRN163L (Imetelstat[®]) is one of the first generation most promising telomerase inhibitor targeting hTER used in cancer treatment; it is a lipid modified

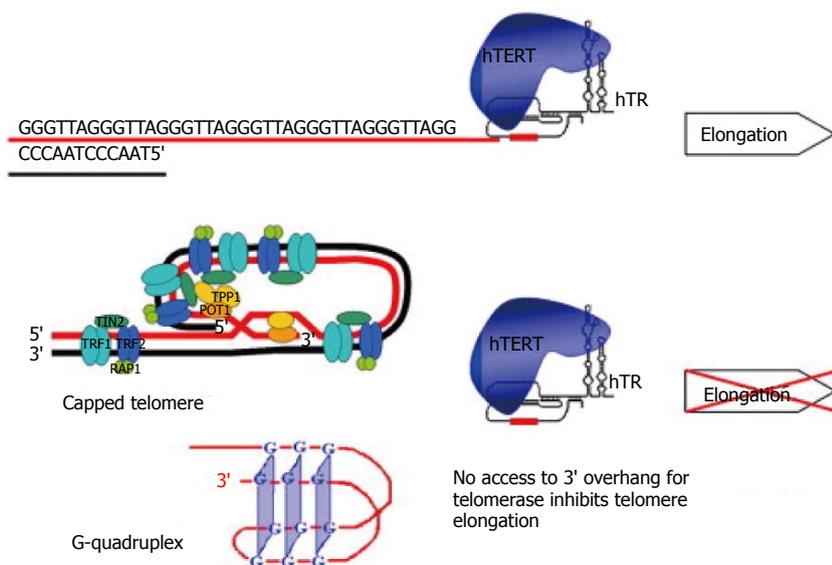


Figure 1 Impact of open or capped telomere structure on the telomerase activity. (Reproduced with permission from Philippi C, Loretz B, Schaefer UF, Lehr CM. *J Control Release* 2010; 146: 228-240. Copyright Clearance Center, Inc.). hTERT: Human Telomerase Reverse Transcriptase.

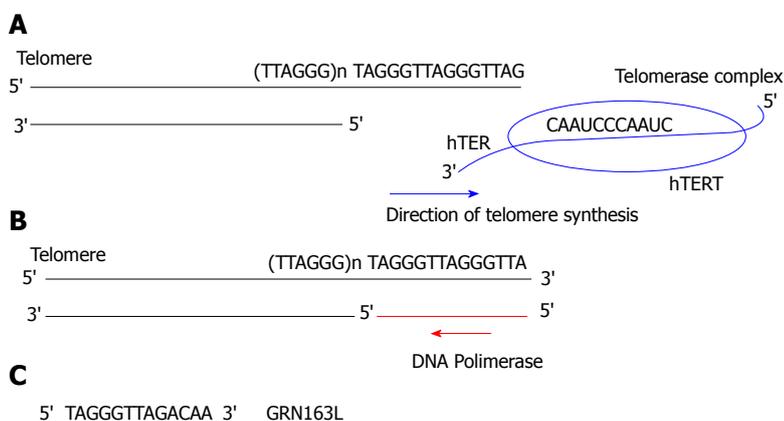


Figure 2 Telomeres as a telomerase complex. (TTAGG)_n sequences form a 3'-overhang on the 3' end of chromosome. Telomerase is composed by hTERT and hTER subunits; hTER is the RNA template for DNA synthesis to add new telomeric TTAGG sequences on 3'-overhang; B: DNA polymerase completes the lagging strand; C: GRN163L sequence complementary. hTERT: Human Telomerase Reverse Transcriptase; hTER: Human Telomerase RNA.

version of GRN163, a 13-mer oligonucleotide N3'-P5'-thio-phosphoramidate, that required a lipid carrier molecule and a lipid-base transfection agent to adequately enter tissue and cellular membranes^[31,32]. On the contrary, GRN163L with a covalently bound lipophylic palmitoyl (C16) group linked to its 5'-thio-phosphate^[33] is lipid soluble, and shows an higher drug availability and bio-distribution, without any lipid carrier supply^[32]. GRN163L in part overlaps the hTER template region by binding with high affinity and specificity at its active site, acting as competitive telomerase inhibitor and causing a total enzyme inhibition^[32,33] (Figure 2).

The GRN163L inhibitory effect on telomerase activity has been evaluated in different cancer cell lines^[34] and its effects were evident as well as “*in vitro*” and “*in vivo*” models; in fact, long term treatment with GRN163L reduced cell viability in cancer cells derived from bladder^[33] glioblastoma^[35], multiple myeloma^[36], Barrett’s adenocarcinoma^[37], as well as breast^[38,39], lung^[40], liver cancer^[41] and prostate^[42].

Recently, the effects of GRN163L have been tested on a panel of ten pancreatic cancer cell lines, and the results indicated that the inhibitory effect of the drug was maintained also after its removal^[43]: in fact, only three weeks after the GRN163L removal, a telomerase recovery was ob-

served, but the enzyme was less processive. This suggests that to maintain continuous telomerase inhibition and to reduce side effects risk after a pharmacological treatment of a patient with GRN163L, a maintenance dose given once every other week might be sufficient. However, the reversible effects of Imetelstat have been also previously demonstrated on rat mesenchymal stem cells^[44].

A combined treatment where homologous recombination and telomerase inhibition are associated, causes a significant increase in telomeres attrition, relative to each treatment alone, leading to senescence and apoptosis in Barrett’s adenocarcinoma^[45].

Tamakawa *et al*^[46] showed that the DNA damage induced in S/G₂ phase of the cell cycle, by genotoxic stimulus was potentiated by the telomerase inhibition induced by GRN163L in breast and colorectal cancer cells^[46].

In previous studies, synergies between GRN163L and various anticancer treatments such as microtubule inhibition, inhibition of oncogenic signals and ionizing radiation, were considered to be dependent on longer-term changes associated with chromatin status^[47] and telomere length^[48].

Telomere shortening induced by telomerase inhibitors would affect the self-renewal properties of cancer stem cells (CSCs), normally not responding to standard chemotherapy, but capable of inducing initiation and currency

in different hematologic and solid tumours^[49,50].

Many studies showed that CSCs can represent the Imetelstat target in different cancers^[35,42,51], and that a telomere shortening-independent as well as dependent Imetelstat mechanism of action on CSCs subpopulation, can be suggested^[52,53]. The effect of Imetelstat was evaluated on both the bulk cancer cells and putative CSCs of breast and pancreatic cancer cell lines. The *in vitro* treatment inhibited telomerase activity, cell growth, self renewal in bulk cancer cells and putative CSCs, with a consequent reduced cancer engraftment in nude mice^[52]; in particular an increased sensitivity of CSCs to Imetelstat did not correlate with differences between telomerase activity expression levels or telomere length of CSCs and bulk tumour cells suggesting a telomere shortening-independent mechanism of action for the Imetelstat effects on CSCs subpopulation.

All these studies support the hypothesis that conventional therapies often fail to target CSCs while the use of telomerase inhibitor could have the potential role for more durable clinical response in many tumors, reducing relapse recurrence.

Imetelstat is currently in phase II clinical development for breast cancer, non-small cell lung carcinoma, multiple myeloma, and other tumor types^[30].

Inhibitors targeting hTERT: BIBR1532

BIBR1532 [2-(E-3-naphthalen-2-yl-but-2-enylamino)-benzoic acid] is actually a promising hTERT inhibitor among the few TERT inhibitors developed. BIBR1532 is a small synthetic non-nucleic compound that linking hTERT in its active site, inhibits telomerase in a non-competitive manner: BIBR1532 does not cause chain termination events but rather leads to an overall reduction in the number of added TTAGGG repeats^[54]; in particular the drug could act translocating the enzyme-DNA-substrate complex, or favouring the DNA substrate disjunction from the enzyme during the copy of the template^[55].

In the last few years, different studies showed that BIBR1532 treatment induced telomerase activity reduction with consequent cell growth arrest in different human cancer cell lines^[54,56-60], without affecting normal stem cells^[61]. In addition telomeres targeting might represent a valid strategy for the re-sensitization of chemoresistant chondrosarcomas^[56], and a rapid induction of a high level telomere dysfunction appears to be a crucial parameter for the development of future telomerase-based therapeutic^[62]. However, although some human squamous cell carcinoma cell lines are resistant to telomerase inhibition^[63] some works suggest that a valid strategy for the treatment of both drug-resistant and drug-sensitive cancers may be pharmacological telomerase inhibition in combination therapy^[64-66].

IMMUNOTHERAPY FOR TELOMERASE EXPRESSING CANCER

As previously described, nearly all cancer cells over-ex-

press functional active telomerase, and hTERT-specific epitopes are expressed on tumour cells, but not on normal cells. In this way, telomerase become an immune target, and can be eradicated by the stimulation of the immune system with specific vaccines. Telomerase-target immunotherapy sensitizes immune cells against tumor cells expressing hTERT peptides as surface antigens^[67]. The consequent expansion of telomerase-specific CD8+ cytotoxic T lymphocytes is directed to target and kill telomerase positive cancer cells^[68,69].

Recently, multiple peptides are known to induce hTERT-specific immune responses^[68] and several vaccine strategies are being developed and used: among these GV1001, Vx-001, I540, are the most widespread therapeutic approaches. As almost all human tumor-associated antigens are self-proteins, their specific T cells are often tolerated: this is the major problem of cancer immunotherapy. For this reason, overcoming tumor-specific self-tolerance is a principal goal in cancer immunotherapy.

Self-tolerance is commonly directed against “dominant” (high affinity for HLA) but not against “cryptic” (low affinity for HLA) peptides^[70,71], so the simplest way to circumvent tolerance is to use these cryptic peptides^[72] as for example Vx-001 (9-mer cryptic TERT 572 peptide) that was developed as tumour-associated antigen of hTERT to induce cytotoxic T lymphocyte responses^[73,74].

Immunological response associated with extended survival were evident in patients with advanced non-small-cell lung cancer treated with Vx-001 vaccine (TERT572Y peptide)^[74]; in patients with various types of chemo-resistant advanced solid tumours (stages III and IV) the vaccination with Vx-001 stimulates TERT572-specific reactive T cells in a great number of patients independently of the disease stage or clinical status before vaccination and a late immune response correlated with longer survival was induced^[73,75].

State of the art of clinical trials using anti-telomerase cancer immunotherapy is encouraging. In fact, vaccines are tested in breast, lung, melanoma, prostate, and pancreatic cancer^[76-82] and these trials have widely induced a specific immune response against hTERT positive cancer cells. Encouraging results have been also obtained in patients with advanced melanoma, where immunity to hTERT has been safely generated^[83]. The combination of cancer vaccination with chemotherapy showed that temozolomide and GV1001 induced immune and clinical response in 78% of stage IV melanoma patients, that developed long-term T-cell memory and survived more than those rapidly losing their responses^[84]. Vaccination with GV1001 was well tolerated and immunized the great part of non-small cell lung cancer patients establishing durable T-cell memory^[85]. However, GV1001 vaccination was not effective in cutaneous T cell lymphoma patients, raising concerns about also its safety^[86]. The survival data indicated that patients with non-resectable pancreatic cancer treated with GV1001 showed that immune response correlated with an extended survival, suggesting that the vaccine could be the new goal for pancreatic cancer patients treatment and encouraging further clinical

studies^[82]. On the contrary, in patients with advanced and metastatic pancreatic cancer the use of GV1001 telomerase vaccination in combination with chemotherapy, induced a weak and transient immune response and did not improve overall survival^[80,81]. Likewise, a low dose cyclophosphamide treatment in combination with GV1001 vaccination in patients with advanced hepatocellular carcinoma did not show antitumor efficacy^[87]. Further studies and new strategies are needed to analyze and to enhance the immune response effect of telomerase vaccination during chemotherapy, in patients with both pancreatic and hepatocellular cancers.

Vaccination with autologous dendritic cells transfected with hTERT mRNA (GRNVAC1) represents another anticancer approach that induced immunological response in human. Immunotherapy targeting the hTERT subunit of telomerase has been demonstrated to induce an important immune responses in cancer patients after vaccination with single hTERT peptides, while vaccination with dendritic cells transfected with hTERT mRNA has a key role in inducing efficient immune responses to multiple hTERT epitopes. In this way this kind of therapy can be an attractive approach to more efficient immunotherapy^[88-90].

TELOMERASE-EXPRESSING CELLS AS TARGET OF ONCOLYTIC VIRUSES

Recently has been shown that the use of hTERT promoter to drive the expression of a suicide gene and/or control the replication of a lytic virus, can be a successful approach to target cancer cells.

To drive the expression of a suicide gene, the expression of a pro-apoptotic protein, like TRAIL (tumour necrosis factor-related apoptosis-inducing ligand) or pro-drug-activating enzyme^[91-96] is controlled by the hTERT promoter, generally active in cancer cells expressing telomerase. These cells are injected with viruses carrying the suicide gene and then killed by a toxin derived from the administration of a pro-drug activated by the pro-drug-activating enzyme.

A second clinical approach, is to use the hTERT promoter to control the replication of a lytic virus. Oncolytic effects on tumors can be mediated by oncolytic viruses, tumor selective viruses genetically modified and engineered to replicate in and kill only cancer cells. For this purpose, the *E1* gene expresses viral proteins E1A and E1B necessary for adenovirus replication, but the modified virus can replicate only in cells which express telomerase if gene itself is redesigned to be controlled by the hTERT promoter^[97-100]. One such virus is telomelysin (OBP-301) that in pre-clinical studies targets selectively only telomerase-expressing cells.

The modified viruses induce cytolysis in several kinds of human cancer cell lines in which can replicate; when human lung, prostate or liver cancer cells were used in xenotransplantation models, intratumoral injection of the virus reduced tumor growth and improved mice sur-

vival^[97-100].

The potential role of oncolytic virotherapy has recently been demonstrated to be a promising strategy in the management of human gastrointestinal cancer^[101]. Studies about OBP-301 have been shown that it mediates the effective *in vivo* purging of metastatic tumor cells from regional lymph nodes and moreover it co-operates to optimize treatment of human gastrointestinal malignancies^[102]. Moreover, telomerase-specific oncolytic viruses is a potential treatment of human squamous cell carcinoma of head and neck^[103], while in pancreatic cancer the combination therapy with gemcitabine has been tried, exhibiting enhanced cytotoxic effects both "*in vitro*" and "*in vivo*"^[104]. In addition, preclinical study showed that OBP-301 can be used for treatment of human hepatocellular carcinoma and that its tumor-killing activity persists after multiple injections^[105].

Data regarding combination therapy with OBP-301 and chemotherapeutic agents are preliminary but encouraging^[106]. In particular Boozari *et al.*^[107] showed that the combination of intratumoural virotherapy with an antitumoural vaccine, could represent a promising immunotherapeutic strategy against hepatocellular carcinoma and metastasis.

TELOMERASE CANONICAL ACTIVITY AS A THERAPY

Recent studies revealed that telomerase canonical activity can be exploited for therapeutic purpose.

The evidence that telomerase is expressed in almost all tumor cells, preventing telomeres shortening by continually adding single stranded TTAGGG DNA sequences, prompted us to develop a thymidine analogue pro-drug, acycloguanosyl 5'-thymidyltriphosphate (ACV-TP-T) (Figure 3). This molecule is a synthetic ACV modification that is metabolized by telomerase, and this reaction releases the active form of acyclovir able to reduce pancreatic and hepatocellular carcinoma cells growth as well as "*in vitro*" and "*in vivo*"^[108,109].

ACV is a nucleoside analogue acting as a DNA chain terminator that could be used in the suicide gene therapy^[110]. ACV or the ACV analogue ganciclovir^[110,111] when used as antiviral agent needs a first phosphorylation to ACV monophosphate by herpes virus thymidine kinase (TK) carried by wild-type herpes virus or, in the suicide gene therapy, by engineered adenovirus (Figure 3), then cellular kinases perform the two remaining phosphorylation to obtain the ACV triphosphate. This active metabolite is incorporated into DNA during its replication causing DNA chain termination.

On the contrary, ACV-TP-T, may be metabolized by telomerase that incorporates thymidine in replicating telomeres and releases ACV diphosphate. This process skips the viral TK phosphorylation, allowing the cellular kinases to go on with further phosphorylation to obtain the active drug^[108,109]. The results showed that after activation of ACV-TP-T by telomerase, cell proliferation is significantly

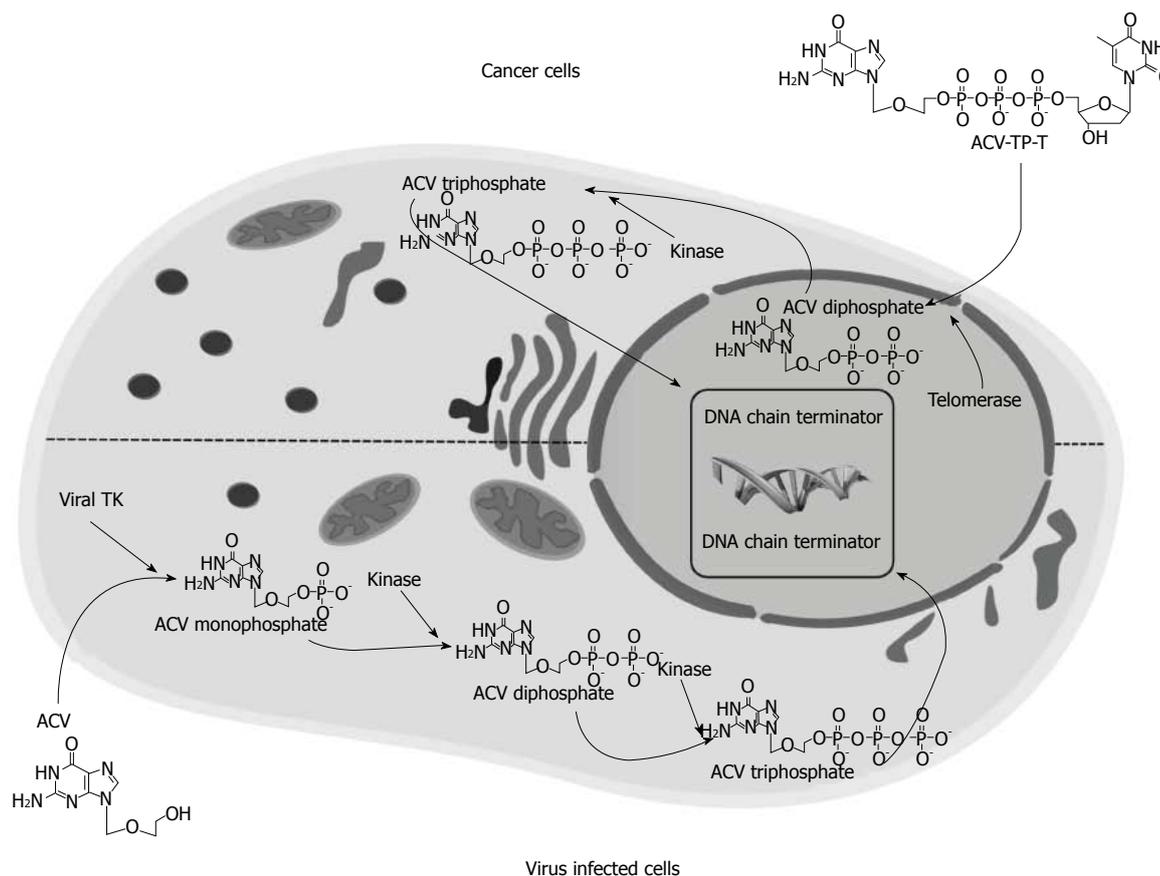


Figure 3 Structure and schematic mode of action of Acycloguanosyl 5'-thymidyltriphosphate in comparison with ACV. For activation, ACV requires to be phosphorylated to ACV monophosphate by viral TK carried either by wildtype herpes virus or, in the suicide gene therapy, engineered adenovirus. ACV monophosphate is then further phosphorylated by cellular kinases to the triphosphated active form. Conversely, ACV-TP-T is substrate of telomerase that incorporates the thymidine in the replicating telomeres and directly release ACV diphosphate skipping the viral TK phosphorylation step. (Reproduced with permission from Ref [108]. Copyright 2011 AGA Institute). ACV-TP-T: Acycloguanosyl 5'-thymidyltriphosphate; TK: Thymidine kinase.

reduced and apoptosis is increased in different human pancreatic adenocarcinoma cell lines. High and low telomerase activity is related with low and high IC₅₀ of the drug, respectively. On the other hand, the cytosine-containing pro-drug ACV-TP-dC, which is not a telomerase substrate, is not able to reduce pancreatic cancer cell proliferation. Moreover, ACV-TP-T administration increases apoptosis, reduces growth, proliferation and vascularization of pancreatic xenograft tumors in mice^[108].

Analogue results were obtained in human and murine hepatocellular carcinoma cell lines and in transgenic and orthotopic murine models of hepatic cancers^[109]. Furthermore, in orthotopic syngenic mice, ACV-TP-T has been used alone or in combination with the approved standard of care, Sorafenib, a multikinase inhibitor. Combination therapy showed a synergistic effect between Sorafenib and ACV-TP-T.

Advantages of this strategy are evident. Despite recent improvements in suicide gene therapy, the application of adenovirus-mediated therapy is limited by many factors: the low and transient expression levels of the transgene^[110,112,113], the induction of immune response in the host^[108], and a late carcinogenesis^[112]. In addition ethical concerns regarding the use of virus in patients^[112,113] could be a limitation.

The use of telomerase promoter^[114] and the introduction of conditionally replication-competent adenovirus^[115] only partially overcome the above mentioned disadvantages. Moreover, the immunotherapy based on vaccination for telomerase^[84] relies on the induction of an active immune response that often is deregulated in the oncology treated patients^[116].

In this contest, the use of ACV-TP-T represents a new therapeutic strategy that exploits the enzymatic activity of telomerase. This approach is efficient only in neoplastic cells without any effects in normal cells, it avoids the toxicity and the adverse effects of the current chemotherapy, and finally, it does not require the use of any viruses or an active immune response of the host.

As a paradox in this contest telomerase switches from being a target of anticancer therapy, to an integral part of the therapy. Preliminary evidences suggest the possible use of ACV-TP-T molecule for the treatment of other tumors characterized by high telomerase expression and activity such as ovarian and adrenocortical cancers.

NON CANONICAL EFFECTS OF TELOMERASE

Telomerase activation may have both telomere-dependent

and telomere-independent implications for cancer progression: in particular, telomerase reverse transcriptase may exert some biological functions independently of its telomere maintenance enzymatic activity.

Different studies support a role of telomerase in some telomere-independent activities in cancer progression; nevertheless, apart from its role in telomere maintenance, the molecular mechanism by which telomerase promotes cancer is still not fully understood. Zhou *et al.*^[117] showed that hTER regulated vascular endothelial growth factor (VEGF) expression at the transcriptional level, independently of telomerase activity^[117]; previous studies reported that VEGF induced hTERT expression and activity in normal^[118] and cancer cells^[119]. All these results suggested a positive feedback regulation that could contribute to a mutual and collaborative function of VEGF and telomerase in cancer progression.

Wu *et al.*^[120] in a recent review focused on various signaling pathways and genes involved in the feedback regulation of TERT. The expression of numerous genes involved in different cellular processes, as well as cell cycle and cellular signaling, could be regulated by TERT, indicating that telomerase is both an effector and a regulator in carcinoma. However, the mechanisms underlying the interaction between TERT and its target genes are still not completely understood.

Ghosh *et al.*^[121] suggested a functional interplay between TERT and nuclear factor (NF- κ B) signaling, further reinforced by the observation that telomerase over expression resulted in enhanced expression of NF- κ B target genes, whereas telomerase null mice were refractory to NF- κ B activation; in addition, it seems that also hTER could regulate the expression of some NF- κ B target genes. The function of hTER in gene expression regulation is not clear, in fact, hTERT can form complexes with or without hTER^[122].

hTERT could be involved also in a negative feedback loop system with pRb/E2F pathway in cancer, as well as in a positive feedback loop with Wnt/ β -catenin signaling, or in multiple interactions with phosphoinositide 3 kinase/Akt pathway^[120]. In addition, Liu *et al.*^[123] demonstrated a potential role of hTERT in epithelial mesenchymal transition.

Although the mechanisms underlying the interaction between TERT and its target genes are still not completely understood, all the above observations, strengthen the idea that telomerase non-telomeric functions could be used as a new therapeutic target for cancer.

CONCLUSION

Although recent and ongoing results support an important role for telomerase targeting therapeutics in cancer treatment, additional preclinical and clinical trials are necessary to improve some of these strategies.

In fact, if difficulties with dendritic cells derivation will be easily overcome^[124], vaccination with dendritic cells transfected with hTERT mRNA could potentially

become an attractive approach to a more potent immunotherapy. In addition, further studies are necessary to enhance the effects of telomerase vaccination in combination with intratumoral virotherapy and with standard chemotherapeutic agents.

On the contrary, beside more promising approach offered by GRN163L that seems to target also CSC, BIBR1532 could be preferred therapy if used also in combination with standard chemotherapy for the treatment of drug-resistant cancers.

Finally, ACV-TP-T use is very promising and deserves further studies. In fact, preclinical evidences showed that this new pro-drug may be considered for treatment of hepatocellular and pancreatic carcinoma, as well as of other tumors characterized by high telomerase expression and activity.

REFERENCES

- 1 **Artandi SE**, DePinho RA. Telomeres and telomerase in cancer. *Carcinogenesis* 2010; **31**: 9-18 [PMID: 19887512 DOI: 10.1093/carcin/bgp268]
- 2 **Meyerson M**. Role of telomerase in normal and cancer cells. *J Clin Oncol* 2000; **18**: 2626-2634 [PMID: 10893296]
- 3 **Batista LF**, Artandi SE. Telomere uncapping, chromosomes, and carcinomas. *Cancer Cell* 2009; **15**: 455-457 [PMID: 19477422 DOI: 10.1016/j.ccr.2009.05.006]
- 4 **Moyzis RK**, Buckingham JM, Cram LS, Dani M, Deaven LL, Jones MD, Meyne J, Ratliff RL, Wu JR. A highly conserved repetitive DNA sequence, (TTAGGG)_n, present at the telomeres of human chromosomes. *Proc Natl Acad Sci USA* 1988; **85**: 6622-6626 [PMID: 3413114]
- 5 **Blackburn EH**. Telomere states and cell fates. *Nature* 2000; **408**: 53-56 [PMID: 11081503]
- 6 **Harley CB**, Futcher AB, Greider CW. Telomeres shorten during ageing of human fibroblasts. *Nature* 1990; **345**: 458-460 [PMID: 2342578 DOI: 10.1038/345458a0]
- 7 **Takai H**, Smogorzewska A, de Lange T. DNA damage foci at dysfunctional telomeres. *Curr Biol* 2003; **13**: 1549-1556 [PMID: 12956959 DOI: 10.1016/S0960-9822(03)00542-6]
- 8 **d'Adda di Fagnana F**, Reaper PM, Clay-Farrace L, Fiegler H, Carr P, Von Zglinicki T, Saretzki G, Carter NP, Jackson SP. A DNA damage checkpoint response in telomere-initiated senescence. *Nature* 2003; **426**: 194-198 [PMID: 14608368]
- 9 **Karlseder J**, Broccoli D, Dai Y, Hardy S, de Lange T. p53- and ATM-dependent apoptosis induced by telomeres lacking TRF2. *Science* 1999; **283**: 1321-1325 [PMID: 10037601 DOI: 10.1126/science.283.5406.1321]
- 10 **Shay JW**, Pereira-Smith OM, Wright WE. A role for both RB and p53 in the regulation of human cellular senescence. *Exp Cell Res* 1991; **196**: 33-39 [PMID: 1652450 DOI: 10.1016/0014-4827(91)90453-2]
- 11 **Wright WE**, Pereira-Smith OM, Shay JW. Reversible cellular senescence: implications for immortalization of normal human diploid fibroblasts. *Mol Cell Biol* 1989; **9**: 3088-3092 [PMID: 2779554]
- 12 **Wright WE**, Shay JW. The two-stage mechanism controlling cellular senescence and immortalization. *Exp Gerontol* 1992; **27**: 383-389 [PMID: 1333985 DOI: 10.1016/0531-5565(92)90069-C]
- 13 **Macera-Bloch L**, Houghton J, Lenahan M, Jha KK, Ozer HL. Termination of lifespan of SV40-transformed human fibroblasts in crisis is due to apoptosis. *J Cell Physiol* 2002; **190**: 332-344 [PMID: 11857449 DOI: 10.1002/jcp.10062]
- 14 **Zhang X**, Mar V, Zhou W, Harrington L, Robinson MO. Telomere shortening and apoptosis in telomerase-inhibited human tumor cells. *Genes Dev* 1999; **13**: 2388-2399 [PMID: 10500096 DOI: 10.1101/gad.13.18.2388]

- 15 **Ruden M**, Puri N. Novel anticancer therapeutics targeting telomerase. *Cancer Treat Rev* 2013; **39**: 444-456 [PMID: 22841437 DOI: 10.1016/j.ctrv.2012.06.007]
- 16 **Blackburn EH**. Telomerase and Cancer: Kirk A. Landon--AACR prize for basic cancer research lecture. *Mol Cancer Res* 2005; **3**: 477-482 [PMID: 16179494 DOI: 10.1158/1541-7786.MCR-05-0147]
- 17 **Tian X**, Chen B, Liu X. Telomere and telomerase as targets for cancer therapy. *Appl Biochem Biotechnol* 2010; **160**: 1460-1472 [PMID: 19412578 DOI: 10.1007/s12010-009-8633-9]
- 18 **Cong YS**, Wright WE, Shay JW. Human telomerase and its regulation. *Microbiol Mol Biol Rev* 2002; **66**: 407-425, table of contents [PMID: 12208997 DOI: 10.1128/MMBR.66.3.407-425.2002]
- 19 **Kelleher C**, Teixeira MT, Förstemann K, Lingner J. Telomerase: biochemical considerations for enzyme and substrate. *Trends Biochem Sci* 2002; **27**: 572-579 [PMID: 12417133 DOI: 10.1016/S0968-0004(02)02206-5]
- 20 **Cohen SB**, Graham ME, Lovrecz GO, Bache N, Robinson PJ, Reddel RR. Protein composition of catalytically active human telomerase from immortal cells. *Science* 2007; **315**: 1850-1853 [PMID: 17395830]
- 21 **Ducrest AL**, Szutorisz H, Lingner J, Nabholz M. Regulation of the human telomerase reverse transcriptase gene. *Oncogene* 2002; **21**: 541-552 [PMID: 11850779 DOI: 10.1038/sj.onc.1205081]
- 22 **Takakura M**, Kyo S, Kanaya T, Tanaka M, Inoue M. Expression of human telomerase subunits and correlation with telomerase activity in cervical cancer. *Cancer Res* 1998; **58**: 1558-1561 [PMID: 9537264]
- 23 **Wright WE**, Piatyszek MA, Rainey WE, Byrd W, Shay JW. Telomerase activity in human germline and embryonic tissues and cells. *Dev Genet* 1996; **18**: 173-179 [PMID: 8934879]
- 24 **Kim NW**, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PL, Coviello GM, Wright WE, Weinrich SL, Shay JW. Specific association of human telomerase activity with immortal cells and cancer. *Science* 1994; **266**: 2011-2015 [PMID: 7605428 DOI: 10.1126/science.7605428]
- 25 **Collins K**, Mitchell JR. Telomerase in the human organism. *Oncogene* 2002; **21**: 564-579 [PMID: 11850781 DOI: 10.1038/sj.onc.1205083]
- 26 **Kelland LR**. Overcoming the immortality of tumour cells by telomere and telomerase based cancer therapeutics--current status and future prospects. *Eur J Cancer* 2005; **41**: 971-979 [PMID: 15862745]
- 27 **Phatak P**, Burger AM. Telomerase and its potential for therapeutic intervention. *Br J Pharmacol* 2007; **152**: 1003-1011 [PMID: 17603541]
- 28 **Corey DR**. Telomerase: an unusual target for cytotoxic agents. *Chem res Toxicol* 2000; **13**: 957-960 [PMID: 11080041]
- 29 **Hanahan D**, Weinberg RA. The hallmarks of cancer. *Cell* 2000; **100**: 57-70 [PMID: 10647931 DOI: 10.1016/S0092-8674(00)81683-9]
- 30 **Harley CB**. Telomerase and cancer therapeutics. *Nat Rev Cancer* 2008; **8**: 167-179 [PMID: 18256617 DOI: 10.1038/nrc2275]
- 31 **Rankin AM**, Faller DV, Spanjaard RA. Telomerase inhibitors and 'T-oligo' as cancer therapeutics: contrasting molecular mechanisms of cytotoxicity. *Anticancer Drugs* 2008; **19**: 329-338 [PMID: 18454043 DOI: 10.1097/CAD.0b013e3282f5d4c2]
- 32 **Röth A**, Harley CB, Baerlocher GM. Imetelstat (GRN163L)-telomerase-based cancer therapy. *Recent Results Cancer Res* 2010; **184**: 221-234 [PMID: 20072842]
- 33 **Dikmen ZG**, Wright WE, Shay JW, Gryaznov SM. Telomerase targeted oligonucleotide thio-phosphoramidates in T24-luc bladder cancer cells. *J Cell Biochem* 2008; **104**: 444-452 [PMID: 18044713]
- 34 **Herbert BS**, Gellert GC, Hochreiter A, Pongracz K, Wright WE, Zielinska D, Chin AC, Harley CB, Shay JW, Gryaznov SM. Lipid modification of GRN163, an N3'-& gt; P5' thio-phosphoramidate oligonucleotide, enhances the potency of telomerase inhibition. *Oncogene* 2005; **24**: 5262-5268 [PMID: 15940257 DOI: 10.1038/sj.onc.1208760]
- 35 **Marian CO**, Cho SK, McEllin BM, Maher EA, Hatanpaa KJ, Madden CJ, Mickey BE, Wright WE, Shay JW, Bachoo RM. The telomerase antagonist, imetelstat, efficiently targets glioblastoma tumor-initiating cells leading to decreased proliferation and tumor growth. *Clin Cancer Res* 2010; **16**: 154-163 [PMID: 20048334 DOI: 10.1158/1078-0432.CCR-09-2850]
- 36 **Shammas MA**, Koley H, Bertheau RC, Neri P, Fulciniti M, Tassone P, Blotta S, Protopopov A, Mitsiades C, Batchu RB, Anderson KC, Chin A, Gryaznov S, Munshi NC. Telomerase inhibitor GRN163L inhibits myeloma cell growth in vitro and in vivo. *Leukemia* 2008; **22**: 1410-1418 [PMID: 18449204 DOI: 10.1038/leu.2008.81]
- 37 **Shammas MA**, Qazi A, Batchu RB, Bertheau RC, Wong JY, Rao MY, Prasad M, Chanda D, Ponnazhagan S, Anderson KC, Steffes CP, Munshi NC, De Vivo I, Beer DG, Gryaznov S, Weaver DW, Goyal RK. Telomere maintenance in laser capture microdissection-purified Barrett's adenocarcinoma cells and effect of telomerase inhibition in vivo. *Clin Cancer Res* 2008; **14**: 4971-4980 [PMID: 18676772 DOI: 10.1158/1078-0432.CCR-08-0473]
- 38 **Gellert GC**, Dikmen ZG, Wright WE, Gryaznov S, Shay JW. Effects of a novel telomerase inhibitor, GRN163L, in human breast cancer. *Breast Cancer Res Treat* 2006; **96**: 73-81 [PMID: 16319992 DOI: 10.1007/s10549-005-9043-5]
- 39 **Hochreiter AE**, Xiao H, Goldblatt EM, Gryaznov SM, Miller KD, Badve S, Sledge GW, Herbert BS. Telomerase template antagonist GRN163L disrupts telomere maintenance, tumor growth, and metastasis of breast cancer. *Clin Cancer Res* 2006; **12**: 3184-3192 [PMID: 16707619 DOI: 10.1158/1078-0432.CCR-05-2760]
- 40 **Dikmen ZG**, Gellert GC, Jackson S, Gryaznov S, Tressler R, Dogan P, Wright WE, Shay JW. In vivo inhibition of lung cancer by GRN163L: a novel human telomerase inhibitor. *Cancer Res* 2005; **65**: 7866-7873 [PMID: 16140956]
- 41 **Djojoseburo MW**, Chin AC, Go N, Schaezlein S, Manns MP, Gryaznov S, Harley CB, Rudolph KL. Telomerase antagonists GRN163 and GRN163L inhibit tumor growth and increase chemosensitivity of human hepatoma. *Hepatology* 2005; **42**: 1127-1136 [PMID: 16114043 DOI: 10.1002/hep.20822]
- 42 **Marian CO**, Wright WE, Shay JW. The effects of telomerase inhibition on prostate tumor-initiating cells. *Int J Cancer* 2010; **127**: 321-331 [PMID: 19908230 DOI: 10.1002/ijc.25043]
- 43 **Burchett KM**, Yan Y, Ouellette MM. Telomerase inhibitor Imetelstat (GRN163L) limits the lifespan of human pancreatic cancer cells. *PLoS One* 2014; **9**: e85155 [PMID: 24409321 DOI: 10.1371/journal.pone.0085155]
- 44 **Tokcaer-Keskin Z**, Dikmen ZG, Ayaloglu-Butun F, Gultekin S, Gryaznov SM, Akcali KC. The effect of telomerase template antagonist GRN163L on bone-marrow-derived rat mesenchymal stem cells is reversible and associated with altered expression of cyclin d1, cdk4 and cdk6. *Stem Cell Rev* 2010; **6**: 224-233 [PMID: 20180048]
- 45 **Lu R**, Pal J, Buon L, Nanjappa P, Shi J, Fulciniti M, Tai YT, Guo L, Yu M, Gryaznov S, Munshi NC, Shammas MA. Targeting homologous recombination and telomerase in Barrett's adenocarcinoma: impact on telomere maintenance, genomic instability and tumor growth. *Oncogene* 2014; **33**: 1495-1505 [PMID: 23604115 DOI: 10.1038/onc.2013.103]
- 46 **Tamakawa RA**, Fleisig HB, Wong JM. Telomerase inhibition potentiates the effects of genotoxic agents in breast and colorectal cancer cells in a cell cycle-specific manner. *Cancer Res* 2010; **70**: 8684-8694 [PMID: 20837664 DOI: 10.1158/0008-5472.CAN-10-2227]
- 47 **Goldblatt EM**, Gentry ER, Fox MJ, Gryaznov SM, Shen C, Herbert BS. The telomerase template antagonist GRN163L alters MDA-MB-231 breast cancer cell morphology, inhibits growth, and augments the effects of paclitaxel.

- Mol Cancer Ther* 2009; **8**: 2027-2035 [PMID: 19509275 DOI: 10.1158/1535-7163.MCT-08-1188]
- 48 **Gomez-Millan J**, Goldblatt EM, Gryaznov SM, Mendonca MS, Herbert BS. Specific telomere dysfunction induced by GRN163L increases radiation sensitivity in breast cancer cells. *Int J Radiat Oncol Biol Phys* 2007; **67**: 897-905 [PMID: 17175117 DOI: 10.1016/j.ijrobp.2006.09.038]
- 49 **Allan AL**, Vantyghem SA, Tuck AB, Chambers AF. Tumor dormancy and cancer stem cells: implications for the biology and treatment of breast cancer metastasis. *Breast Dis* 2006; **26**: 87-98 [PMID: 17473368]
- 50 **Hermann PC**, Huber SL, Herrler T, Aicher A, Ellwart JW, Guba M, Bruns CJ, Heeschen C. Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 2007; **1**: 313-323 [PMID: 18371365 DOI: 10.1016/j.stem.2007.06.002]
- 51 **Castelo-Branco P**, Zhang C, Lipman T, Fujitani M, Hansford L, Clarke I, Harley CB, Tressler R, Malkin D, Walker E, Kaplan DR, Dirks P, Tabori U. Neural tumor-initiating cells have distinct telomere maintenance and can be safely targeted for telomerase inhibition. *Clin Cancer Res* 2011; **17**: 111-121 [PMID: 21208905 DOI: 10.1158/1078-0432.CCR-10-2075]
- 52 **Joseph I**, Tressler R, Bassett E, Harley C, Buseman CM, Pattamatta P, Wright WE, Shay JW, Go NF. The telomerase inhibitor imetelstat depletes cancer stem cells in breast and pancreatic cancer cell lines. *Cancer Res* 2010; **70**: 9494-9504 [PMID: 21062983 DOI: 10.1158/0008-5472.CAN-10-0233]
- 53 **Brennan SK**, Wang Q, Tressler R, Harley C, Go N, Bassett E, Huff CA, Jones RJ, Matsui W. Telomerase inhibition targets clonogenic multiple myeloma cells through telomere length-dependent and independent mechanisms. *PLoS One* 2010; **5**: [PMID: 20824134 DOI: 10.1371/journal.pone.0012487]
- 54 **El-Daly H**, Kull M, Zimmermann S, Pantic M, Waller CF, Martens UM. Selective cytotoxicity and telomere damage in leukemia cells using the telomerase inhibitor BIBR1532. *Blood* 2005; **105**: 1742-1749 [PMID: 15507522 DOI: 10.1182/blood-2003-12-4322]
- 55 **Pascolo E**, Wenz C, Lingner J, Huel N, Pripke H, Kauffmann I, Garin-Chesa P, Rettig WJ, Damm K, Schnapp A. Mechanism of human telomerase inhibition by BIBR1532, a synthetic, non-nucleosidic drug candidate. *J Biol Chem* 2002; **277**: 15566-15572 [PMID: 11854300]
- 56 **Parsch D**, Brassat U, Brümmendorf TH, Fellenberg J. Consequences of telomerase inhibition by BIBR1532 on proliferation and chemosensitivity of chondrosarcoma cell lines. *Cancer Invest* 2008; **26**: 590-596 [PMID: 18584350 DOI: 10.1080/07357900802072905]
- 57 **Röth A**, Dürig J, Himmelreich H, Bug S, Siebert R, Dührsen U, Lansdorf PM, Baerlocher GM. Short telomeres and high telomerase activity in T-cell prolymphocytic leukemia. *Leukemia* 2007; **21**: 2456-2462 [PMID: 17898784]
- 58 **Brassat U**, Balabanov S, Bali D, Dierlamm J, Braig M, Hartmann U, Sirma H, Günes C, Wege H, Fehse B, Gontarewicz A, Dikomey E, Borgmann K, Brümmendorf TH. Functional p53 is required for effective execution of telomerase inhibition in BCR-ABL-positive CML cells. *Exp Hematol* 2011; **39**: 66-76. e1-e2 [PMID: 20940029 DOI: 10.1016/j.exphem.2010.10.001]
- 59 **Bashash D**, Ghaffari SH, Zaker F, Hezave K, Kazerani M, Ghavamzadeh A, Alimoghaddam K, Mosavi SA, Gharebaghian A, Vossough P. Direct short-term cytotoxic effects of BIBR 1532 on acute promyelocytic leukemia cells through induction of p21 coupled with downregulation of c-Myc and hTERT transcription. *Cancer Invest* 2012; **30**: 57-64 [PMID: 22236190 DOI: 10.3109/07357907.2011.629378]
- 60 **Nakashima M**, Nandakumar J, Sullivan KD, Espinosa JM, Cech TR. Inhibition of telomerase recruitment and cancer cell death. *J Biol Chem* 2013; **288**: 33171-33180 [PMID: 24097987 DOI: 10.1074/jbc.M113.518175]
- 61 **El Daly H**, Martens UM. Telomerase inhibition and telomere targeting in hematopoietic cancer cell lines with small non-nucleosidic synthetic compounds (BIBR1532). *Methods Mol Biol* 2007; **405**: 47-60 [PMID: 18369817 DOI: 10.1007/978-1-60327-070-0_6]
- 62 **Pantic M**, Zimmermann S, Waller CF, Martens UM. The level of telomere dysfunction determines the efficacy of telomerase-based therapeutics in a lung cancer cell line. *Int J Oncol* 2005; **26**: 1227-1232 [PMID: 15809713 DOI: 10.3892/ijo.26.5.1227]
- 63 **Bojovic B**, Crowe DL. Resistance to telomerase inhibition by human squamous cell carcinoma cell lines. *Int J Oncol* 2011; **38**: 1175-1181 [PMID: 21305252]
- 64 **Ward RJ**, Autexier C. Pharmacological telomerase inhibition can sensitize drug-resistant and drug-sensitive cells to chemotherapeutic treatment. *Mol Pharmacol* 2005; **68**: 779-786 [PMID: 15939802 DOI: 10.1124/mol.105.011494]
- 65 **Park YP**, Kim KD, Kang SH, Yoon do Y, Park JW, Kim JW, Lee HG. Human telomerase reverse transcriptase (hTERT): a target molecule for the treatment of cisplatin-resistant tumors. *Korean J Lab Med* 2008; **28**: 430-437 [PMID: 19127107 DOI: 10.3343/kjlm.2008.28.6.430]
- 66 **Meng E**, Taylor B, Ray A, Shevde LA, Rocconi RP. Targeted inhibition of telomerase activity combined with chemotherapy demonstrates synergy in eliminating ovarian cancer spheroid-forming cells. *Gynecol Oncol* 2012; **124**: 598-605 [PMID: 22115853 DOI: 10.1016/j.ygyno.2011.11.018]
- 67 **Vonderheide RH**, Hahn WC, Schultze JL, Nadler LM. The telomerase catalytic subunit is a widely expressed tumor-associated antigen recognized by cytotoxic T lymphocytes. *Immunity* 1999; **10**: 673-679 [PMID: 10403642 DOI: 10.1016/S1074-7613(00)80066-7]
- 68 **Liu JP**, Chen W, Schwarer AP, Li H. Telomerase in cancer immunotherapy. *Biochim Biophys Acta* 2010; **1805**: 35-42 [PMID: 19751801 DOI: 10.1016/j.bbcan.2009.09.001]
- 69 **Vonderheide RH**. Prospects and challenges of building a cancer vaccine targeting telomerase. *Biochimie* 2008; **90**: 173-180 [PMID: 17716803 DOI: 10.1016/j.biochi.2007.07.005]
- 70 **Robbins PF**, Kawakami Y. Human tumor antigens recognized by T cells. *Curr Opin Immunol* 1996; **8**: 628-636 [PMID: 8902387 DOI: 10.1016/S0952-7915(96)80078-1]
- 71 **Van Pel A**, van der Bruggen P, Coulie PG, Brichard VG, Lethé B, van den Eynde B, Uyttenhove C, Renaud JC, Boon T. Genes coding for tumor antigens recognized by cytolytic T lymphocytes. *Immunol Rev* 1995; **145**: 229-250 [PMID: 7590828]
- 72 **Tourdot S**, Scardino A, Saloustrou E, Gross DA, Pascolo S, Cordopatis P, Lemonnier FA, Kosmatopoulos K. A general strategy to enhance immunogenicity of low-affinity HLA-A2. 1-associated peptides: implication in the identification of cryptic tumor epitopes. *Eur J Immunol* 2000; **30**: 3411-3421 [PMID: 11093159]
- 73 **Vetsika EK**, Konsolakis G, Aggouraki D, Kotsakis A, Papadimitraki E, Christou S, Menez-Jamet J, Kosmatopoulos K, Georgoulas V, Mavroudis D. Immunological responses in cancer patients after vaccination with the therapeutic telomerase-specific vaccine Vx-001. *Cancer Immunol Immunother* 2012; **61**: 157-168 [PMID: 21858533 DOI: 10.1007/s00262-011-1093-4]
- 74 **Bolonaki I**, Kotsakis A, Papadimitraki E, Aggouraki D, Konsolakis G, Vagia A, Christophylakis C, Nikoloudi I, Magganis E, Galanis A, Cordopatis P, Kosmatopoulos K, Georgoulas V, Mavroudis D. Vaccination of patients with advanced non-small-cell lung cancer with an optimized cryptic human telomerase reverse transcriptase peptide. *J Clin Oncol* 2007; **25**: 2727-2734 [PMID: 17602077 DOI: 10.1200/JCO.2006.10.3465]
- 75 **Kotsakis A**, Vetsika EK, Christou S, Hatzidaki D, Vardakis N, Aggouraki D, Konsolakis G, Georgoulas V, Christophylakis Ch, Cordopatis P, Kosmatopoulos K, Mavroudis D. Clinical outcome of patients with various advanced cancer types vaccinated with an optimized cryptic human telom-

- erase reverse transcriptase (TERT) peptide: results of an expanded phase II study. *Ann Oncol* 2012; **23**: 442-449 [PMID: 21873272 DOI: 10.1093/annonc/mdr396]
- 76 **Vonderheide RH**, Domchek SM, Schultze JL, George DJ, Hoar KM, Chen DY, Stephans KF, Masutomi K, Loda M, Xia Z, Anderson KS, Hahn WC, Nadler LM. Vaccination of cancer patients against telomerase induces functional antitumor CD8+ T lymphocytes. *Clin Cancer Res* 2004; **10**: 828-839 [PMID: 14871958]
- 77 **Brunsvig PF**, Aamdal S, Gjertsen MK, Kvalheim G, Markowski-Grimsrud CJ, Sve I, Dyrhaug M, Trachsel S, Møller M, Eriksen JA, Gaudernack G. Telomerase peptide vaccination: a phase I/II study in patients with non-small cell lung cancer. *Cancer Immunol Immunother* 2006; **55**: 1553-1564 [PMID: 16491401 DOI: 10.1007/s00262-006-0145-7]
- 78 **Inderberg-Suso EM**, Trachsel S, Lislerud K, Rasmussen AM, Gaudernack G. Widespread CD4+ T-cell reactivity to novel hTERT epitopes following vaccination of cancer patients with a single hTERT peptide GV1001. *Oncoimmunology* 2012; **1**: 670-686 [PMID: 22934259 DOI: 10.4161/onci.20426]
- 79 **Chen Z**, Koenenman KS, Corey DR. Consequences of telomerase inhibition and combination treatments for the proliferation of cancer cells. *Cancer Res* 2003; **63**: 5917-5925 [PMID: 14522918]
- 80 **Middleton G**, Silcocks P, Cox T, Valle J, Wadsley J, Propper D, Coxon F, Ross P, Madhusudan S, Roques T, Cunningham D, Falk S, Wadd N, Harrison M, Corrie P, Iveson T, Robinson A, McAdam K, Eatock M, Evans J, Archer C, Hickish T, Garcia-Alonso A, Nicolson M, Steward W, Anthony A, Greenhalf W, Shaw V, Costello E, Naisbitt D, Rawcliffe C, Nanson G, Neoptolemos J. Gemcitabine and capecitabine with or without telomerase peptide vaccine GV1001 in patients with locally advanced or metastatic pancreatic cancer (TeloVac): an open-label, randomised, phase 3 trial. *Lancet Oncol* 2014; **15**: 829-840 [PMID: 24954781 DOI: 10.1016/S1470-2045(14)70236-0]
- 81 **Staff C**, Mozaffari F, Frödin JE, Mellstedt H, Liljefors M. Telomerase (GV1001) vaccination together with gemcitabine in advanced pancreatic cancer patients. *Int J Oncol* 2014; **45**: 1293-1303 [PMID: 24919654 DOI: 10.3892/ijo.2014.2496]
- 82 **Bernhardt SL**, Gjertsen MK, Trachsel S, Møller M, Eriksen JA, Meo M, Buanes T, Gaudernack G. Telomerase peptide vaccination of patients with non-resectable pancreatic cancer: A dose escalating phase I/II study. *Br J Cancer* 2006; **95**: 1474-1482 [PMID: 17060934]
- 83 **Hunger RE**, Kernland Lang K, Markowski CJ, Trachsel S, Møller M, Eriksen JA, Rasmussen AM, Braathen LR, Gaudernack G. Vaccination of patients with cutaneous melanoma with telomerase-specific peptides. *Cancer Immunol Immunother* 2011; **60**: 1553-1564 [PMID: 21681371 DOI: 10.1007/s00262-011-1061-z]
- 84 **Kyte JA**. Cancer vaccination with telomerase peptide GV1001. *Expert Opin Investig Drugs* 2009; **18**: 687-694 [PMID: 19388882 DOI: 10.1517/13543780902897631]
- 85 **Brunsvig PF**, Kyte JA, Kersten C, Sundström S, Møller M, Nyakas M, Hansen GL, Gaudernack G, Aamdal S. Telomerase peptide vaccination in NSCLC: a phase II trial in stage III patients vaccinated after chemoradiotherapy and an 8-year update on a phase I/II trial. *Clin Cancer Res* 2011; **17**: 6847-6857 [PMID: 21918169 DOI: 10.1158/1078-0432.CCR-11-1385]
- 86 **Schlapbach C**, Yerly D, Daubner B, Yawalkar N, Hunger RE. Telomerase-specific GV1001 peptide vaccination fails to induce objective tumor response in patients with cutaneous T cell lymphoma. *J Dermatol Sci* 2011; **62**: 75-83 [PMID: 21377838 DOI: 10.1016/j.jdermsci.2011.02.001]
- 87 **Greten TF**, Forner A, Korangy F, N'Kontchou G, Barget N, Ayuso C, Ormandy LA, Manns MP, Beaugrand M, Bruix J. A phase II open label trial evaluating safety and efficacy of a telomerase peptide vaccination in patients with advanced hepatocellular carcinoma. *BMC Cancer* 2010; **10**: 209 [PMID: 20478057 DOI: 10.1016/S0168-8278(10)60217-6]
- 88 **Su Z**, Vieweg J, Weizer AZ, Dahm P, Yancey D, Turaga V, Higgins J, Boczkowski D, Gilboa E, Dannull J. Enhanced induction of telomerase-specific CD4(+) T cells using dendritic cells transfected with RNA encoding a chimeric gene product. *Cancer Res* 2002; **62**: 5041-5048 [PMID: 12208759]
- 89 **Su Z**, Dannull J, Yang BK, Dahm P, Coleman D, Yancey D, Sichi S, Niedzwiecki D, Boczkowski D, Gilboa E, Vieweg J. Telomerase mRNA-transfected dendritic cells stimulate antigen-specific CD8+ and CD4+ T cell responses in patients with metastatic prostate cancer. *J Immunol* 2005; **174**: 3798-3807 [PMID: 15749921 DOI: 10.4049/jimmunol.174.6.3798]
- 90 **Suso EM**, Dueland S, Rasmussen AM, Vetrusu T, Aamdal S, Kvalheim G, Gaudernack G. hTERT mRNA dendritic cell vaccination: complete response in a pancreatic cancer patient associated with response against several hTERT epitopes. *Cancer Immunol Immunother* 2011; **60**: 809-818 [PMID: 21365467 DOI: 10.1007/s00262-011-0991-9]
- 91 **Katz MH**, Spivack DE, Takimoto S, Fang B, Burton DW, Moossa AR, Hoffman RM, Bouvet M. Gene therapy of pancreatic cancer with green fluorescent protein and tumor necrosis factor-related apoptosis-inducing ligand fusion gene expression driven by a human telomerase reverse transcriptase promoter. *Ann Surg Oncol* 2003; **10**: 762-772 [PMID: 12900367 DOI: 10.1245/ASO.2003.01.021]
- 92 **Liu J**, Zou WG, Lang MF, Luo J, Sun LY, Wang XN, Qian QJ, Liu XY. Cancer-specific killing by the CD suicide gene using the human telomerase reverse transcriptase promoter. *Int J Oncol* 2002; **21**: 661-666 [PMID: 12168115 DOI: 10.3892/ijo.21.3.661]
- 93 **Majumdar AS**, Hughes DE, Lichtsteiner SP, Wang Z, Lebkowski JS, Vasserot AP. The telomerase reverse transcriptase promoter drives efficacious tumor suicide gene therapy while preventing hepatotoxicity encountered with constitutive promoters. *Gene Ther* 2001; **8**: 568-578 [PMID: 11319624 DOI: 10.1038/sj.gt.3301421]
- 94 **Plumb JA**, Bilsland A, Kakani R, Zhao J, Glasspool RM, Knox RJ, Evans TR, Keith WN. Telomerase-specific suicide gene therapy vectors expressing bacterial nitroreductase sensitize human cancer cells to the pro-drug CB1954. *Oncogene* 2001; **20**: 7797-7803 [PMID: 11753658 DOI: 10.1038/sj.onc.1204954]
- 95 **Schepelmann S**, Ogilvie LM, Hedley D, Friedlos F, Martin J, Scanlon I, Chen P, Marais R, Springer CJ. Suicide gene therapy of human colon carcinoma xenografts using an armed oncolytic adenovirus expressing carboxypeptidase G2. *Cancer Res* 2007; **67**: 4949-4955 [PMID: 17510425 DOI: 10.1158/0008-5472.CAN-07-0297]
- 96 **Zhou JH**, Tang B, Liu XL, He DW, Yang DT. hTERT-targeted E. coli purine nucleoside phosphorylase gene/6-methylpurine deoxyribose therapy for pancreatic cancer. *Chin Med J* 2007; **120**: 1348-1352
- 97 **Irving J**, Wang Z, Powell S, O'Sullivan C, Mok M, Murphy B, Cardoza L, Lebkowski JS, Majumdar AS. Conditionally replicative adenovirus driven by the human telomerase promoter provides broad-spectrum antitumor activity without liver toxicity. *Cancer Gene Ther* 2004; **11**: 174-185 [PMID: 14726958 DOI: 10.1038/sj.cgt.7700666]
- 98 **Kawashima T**, Kagawa S, Kobayashi N, Shirakiya Y, Umeoka T, Teraishi F, Taki M, Kyo S, Tanaka N, Fujiwara T. Telomerase-specific replication-selective virotherapy for human cancer. *Clin Cancer Res* 2004; **10**: 285-292 [PMID: 14734481 DOI: 10.1158/1078-0432.CCR-1075-3]
- 99 **Lanson NA**, Friedlander PL, Schwarzenberger P, Kolls JK, Wang G. Replication of an adenoviral vector controlled by the human telomerase reverse transcriptase promoter causes tumor-selective tumor lysis. *Cancer Res* 2003; **63**: 7936-7941 [PMID: 14633724]
- 100 **Wirth T**, Zender L, Schulte B, Mundt B, Plentz R, Rudolph KL, Manns M, Kubicka S, Kühnel F. A telomerase-dependent

- conditionally replicating adenovirus for selective treatment of cancer. *Cancer Res* 2003; **63**: 3181-3188 [PMID: 12810646]
- 101 **Fujiwara T.** A novel molecular therapy using bioengineered adenovirus for human gastrointestinal cancer. *Acta Med Okayama* 2011; **65**: 151-162
- 102 **Fujiwara T, Shirakawa Y, Kagawa S.** Telomerase-specific oncolytic virotherapy for human gastrointestinal cancer. *Expert Rev Anticancer Ther* 2011; **11**: 525-532 [PMID: 21504319 DOI: 10.1586/era.10.200]
- 103 **Fujiwara T.** Telomerase-specific virotherapy for human squamous cell carcinoma. *Expert Opin Biol Ther* 2009; **9**: 321-329 [PMID: 19216621 DOI: 10.1517/14712590802715731]
- 104 **Onimaru M, Ohuchida K, Nagai E, Mizumoto K, Egami T, Cui L, Sato N, Uchino J, Takayama K, Hashizume M, Tanaka M.** Combination with low-dose gemcitabine and hTERT-promoter-dependent conditionally replicative adenovirus enhances cytotoxicity through their crosstalk mechanisms in pancreatic cancer. *Cancer Lett* 2010; **294**: 178-186 [PMID: 20163915 DOI: 10.1016/j.canlet.2010.01.034]
- 105 **Lin WH, Yeh SH, Yang WJ, Yeh KH, Fujiwara T, Nii A, Chang SS, Chen PJ.** Telomerase-specific oncolytic adenoviral therapy for orthotopic hepatocellular carcinoma in HBx transgenic mice. *Int J Cancer* 2013; **132**: 1451-1462 [PMID: 22886913 DOI: 10.1002/ijc.27770]
- 106 **Fujiwara T, Kagawa S, Tazawa H.** Synergistic interaction of telomerase-specific oncolytic virotherapy and chemotherapeutic agents for human cancer. *Curr Pharm Biotechnol* 2012; **13**: 1809-1816 [PMID: 21740362 DOI: 10.2174/138920112800958887]
- 107 **Boozari B, Mundt B, Woller N, Strüver N, Gürlevik E, Schache P, Kloos A, Knocke S, Manns MP, Wirth TC, Kubicka S, Kühnel F.** Antitumoural immunity by virus-mediated immunogenic apoptosis inhibits metastatic growth of hepatocellular carcinoma. *Gut* 2010; **59**: 1416-1426 [PMID: 20675696 DOI: 10.1136/gut.2009.196519]
- 108 **Polvani S, Calamante M, Foresta V, Ceni E, Mordini A, Quattrone A, D'Amico M, Luchinat C, Bertini I, Galli A.** Acycloguanosyl 5'-thymidyltriphosphate, a thymidine analogue prodrug activated by telomerase, reduces pancreatic tumor growth in mice. *Gastroenterology* 2011; **140**: 709-720.e9 [PMID: 21044629 DOI: 10.1053/j.gastro.2010.10.050]
- 109 **Tarocchi M, Polvani S, Peired AJ, Marroncini G, Calamante M, Ceni E, Rhodes D, Mello T, Pieraccini G, Quattrone A, Luchinat C, Galli A.** Telomerase activated thymidine analogue pro-drug is a new molecule targeting hepatocellular carcinoma. *J Hepatol* 2014; **61**: 1064-1072 [PMID: 24862448 DOI: 10.1016/j.jhep.2014.05.027]
- 110 **Ghosh SS, Gopinath P, Ramesh A.** Adenoviral vectors: a promising tool for gene therapy. *Appl Biochem Biotechnol* 2006; **133**: 9-29 [PMID: 16622281 DOI: 10.1385/ABAB:133:1:9]
- 111 **Kaplan JM.** Adenovirus-based cancer gene therapy. *Curr Gene Ther* 2005; **5**: 595-605 [PMID: 16457649 DOI: 10.2174/156652305774964677]
- 112 **Painter RG, Lanson NA, Jin Z, Park F, Wang G.** Conditional expression of a suicide gene by the telomere reverse transcriptase promoter for potential post-therapeutic deletion of tumorigenesis. *Cancer Sci* 2005; **96**: 607-613 [PMID: 16128746 DOI: 10.1111/j.1349-7006.2005.00085.x]
- 113 **Bonini C, Bondanza A, Perna SK, Kaneko S, Traversari C, Cicceri F, Bordignon C.** The suicide gene therapy challenge: how to improve a successful gene therapy approach. *Mol Ther* 2007; **15**: 1248-1252 [PMID: 17505474 DOI: 10.1038/sj.mt.6300190]
- 114 **Wirth T, Kühnel F, Kubicka S.** Telomerase-dependent gene therapy. *Curr Mol Med* 2005; **5**: 243-251 [PMID: 15974879 DOI: 10.2174/1566524053586536]
- 115 **Kirby TO, Rivera A, Rein D, Wang M, Ulasov I, Breidenbach M, Kataram M, Contreras JL, Krumdieck C, Yamamoto M, Rots MG, Haisma HJ, Alvarez RD, Mahareshti PJ, Curiel DT.** A novel ex vivo model system for evaluation of conditionally replicative adenoviruses therapeutic efficacy and toxicity. *Clin Cancer Res* 2004; **10**: 8697-8703 [PMID: 15623655 DOI: 10.1158/1078-0432.CCR-04-1166]
- 116 **Steer HJ, Lake RA, Nowak AK, Robinson BW.** Harnessing the immune response to treat cancer. *Oncogene* 2010; **29**: 6301-6313 [PMID: 20856204]
- 117 **Zhou L, Zheng D, Wang M, Cong YS.** Telomerase reverse transcriptase activates the expression of vascular endothelial growth factor independent of telomerase activity. *Biochem Biophys Res Commun* 2009; **386**: 739-743 [PMID: 19559675 DOI: 10.1016/j.bbrc.2009.06.116]
- 118 **Zaccagnini G, Gaetano C, Della Pietra L, Nanni S, Grasselli A, Mangoni A, Benvenuto R, Fabrizi M, Truffa S, Germani A, Moretti F, Pontecorvi A, Sacchi A, Bacchetti S, Capogrossi MC, Farsetti A.** Telomerase mediates vascular endothelial growth factor-dependent responsiveness in a rat model of hind limb ischemia. *J Biol Chem* 2005; **280**: 14790-14798 [PMID: 15687494 DOI: 10.1074/jbc.M414644200]
- 119 **Bermudez Y, Yang H, Saunders BO, Cheng JQ, Nicosia SV, Kruk PA.** VEGF- and LPA-induced telomerase in human ovarian cancer cells is Sp1-dependent. *Gynecol Oncol* 2007; **106**: 526-537 [PMID: 17559911 DOI: 10.1016/j.ygyno.2007.05.005]
- 120 **Wu XQ, Huang C, He X, Tian YY, Zhou DX, He Y, Liu XH, Li J.** Feedback regulation of telomerase reverse transcriptase: new insight into the evolving field of telomerase in cancer. *Cell Signal* 2013; **25**: 2462-2468 [PMID: 23993966 DOI: 10.1016/j.cellsig.2013.08009]
- 121 **Ghosh A, Saginc G, Leow SC, Khattar E, Shin EM, Yan TD, Wong M, Zhang Z, Li G, Sung WK, Zhou J, Chng WJ, Li S, Liu E, Tergaonkar V.** Telomerase directly regulates NF- κ B-dependent transcription. *Nat Cell Biol* 2012; **14**: 1270-1281 [PMID: 23159929 DOI: 10.1038/ncb2621]
- 122 **Zhou J, Ding D, Wang M, Cong YS.** Telomerase reverse transcriptase in the regulation of gene expression. *BMB Rep* 2014; **47**: 8-14 [PMID: 24388106 DOI: 10.5483/BMBRep.2014.47.1.284]
- 123 **Brower V.** Telomerase-based therapies emerging slowly. *J Natl Cancer Inst* 2010; **102**: 520-521 [PMID: 20388877 DOI: 10.1093/jnci/djq145]
- 124 **Liu Z, Li Q, Li K, Chen L, Li W, Hou M, Liu T, Yang J, Lindvall C, Björkholm M, Jia J, Xu D.** Telomerase reverse transcriptase promotes epithelial-mesenchymal transition and stem cell-like traits in cancer cells. *Oncogene* 2013; **32**: 4203-4213 [PMID: 23045275]

P- Reviewer: Majumdar APN, Paraskevas KI **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Lu YJ



Patents on antivirulence therapies

María López, Beatriz Barbosa, Eva Gato, Germán Bou, María Tomás

María López, Eva Gato, Germán Bou, María Tomás, Servicio de Microbiología, Complejo Hospitalario Universitario A Coruña-INIBIC, 15006 La Coruña, Spain

Beatriz Barbosa, Departamento de Microbiologia, Imunologia e Parasitologia - FCM/UERJ, 20551-030 Rio de Janeiro, Brazil

Author contributions: All authors contributed to this work.

Supported by Instituto de Salud Carlos III FEDER, Spanish Network for the Research in Infectious Diseases, No. REIPI RD12/0015; by the Spanish Ministry of Health and FEDER funding, No. FIS PI10/00056-PI13/02390 (to Tomás M) and PI12/00552 (to Bou G); and by the Miguel Servet Programme (C.H.U.A. Coruña and ISCIII) (to Tomás M)

Correspondence to: María Tomás, MD, PhD, Servicio de Microbiología, Complejo Hospitalario Universitario A Coruña-INIBIC, As Xubias, 84, 15006 La Coruña,

Spain. ma.del.mar.tomas.carmona@sergas.es

Telephone: +34-98-1176399 Fax: +34-98-1178273

Received: June 28, 2014 Revised: October 30, 2014

Accepted: November 7, 2014

Published online: December 9, 2014

Abstract

Antivirulence therapy inhibits bacterial virulence factors, thus preventing the development of infection without affecting bacterial growth. The development of new antibiotics is complicated by the increasing incidence of antibiotic resistance in pathogenic bacteria. Antivirulence therapy is a promising alternative to traditional antibiotic therapy for the treatment of infectious disease, either alone or in combination with antibiotic treatment. In this review, we consider patents concerning inhibition of several bacterial virulence factors: adhesion/colonization, secretion systems, cellular signalling systems and antimicrobial resistance mechanisms. Finally, we emphasize the importance of analyzing new targets and/or molecules in this field and of considering possible resistance mechanisms.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Patents; Quorum sensing; Adhesion; Bacterial secretory systems; Resistance

Core tip: Antimicrobial resistance in nosocomial pathogens has increased dramatically in recent years. The development of new molecules, therapies and/or new combinations for the eradication of these pathogens is therefore imperative. A new line of research in this area is called "Antivirulence Therapy".

López M, Barbosa B, Gato E, Bou G, Tomás M. Patents on antivirulence therapies. *World J Pharmacol* 2014; 3(4): 97-109 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/97.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.97>

INTRODUCTION

Microbial virulence is the ability of a microbe to cause disease. Antivirulence therapies are constituted on inhibition of bacterial virulence and do not influence bacterial growth. Bacteria appreciate their environment and, once in the host they respond by starting a plan determined for the activation of virulence factors. Hence, antivirulence strategies have the ability to interfere in the recognition of the host signals that alarm the bacteria localized in the place of infection and/or that activate specific virulence factors implicated to development of the infection. If the development of virulence factors is prevented, the bacteria will be less able to colonize. Moreover, this tactic will not directly kill bacteria, so initially the evolutionary pressure for the development of resistant strains would be lower than with conventional antibiotics^[1]. Inhibition of the following systems enables interruption of the process of bacterial infection: toxin production, adhesion and colonization, bacterial secretory systems, cell-to-cell signalling pathways, and antibiotic resistance mechanisms, such as efflux pumps (multidrug resistance) (Figure 1)^[2]. In this review, we provide details of patents concerning the inhibition of each of these mechanisms, except for toxin production, which is specific to certain pathogens such as *Bacillus anthracis* (which causes anthrax) and *Clostridium spp.* (which causes gangrene)^[1].

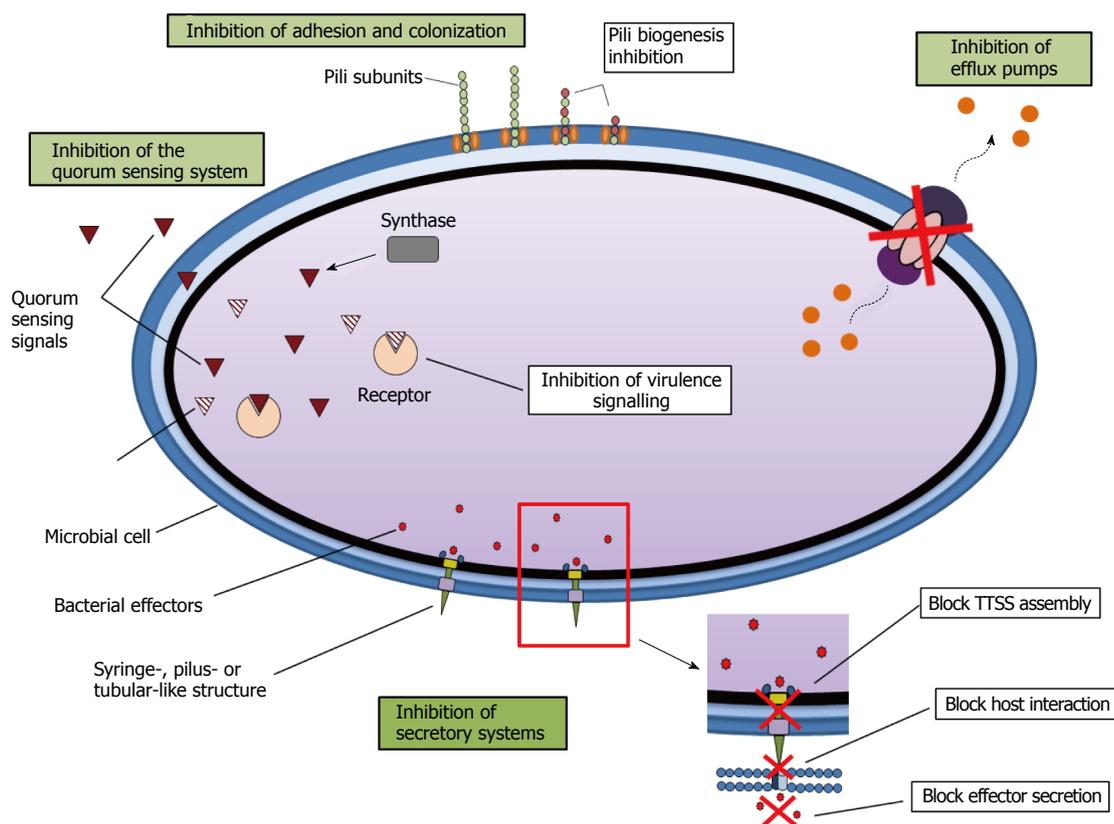


Figure 1 Anti-virulence strategies to combat bacteria-mediated disease (quorum quenchers). TTSS: Type III secretion system.

ADHESION AND COLONIZATION MECHANISMS

Microorganisms adhere to host cells in order to colonize the host and begin infection. The majority of the bacteria own a determined host interval and will only infect hosts that express specific receptors for bacterial adhesion traits on their cell surface. Besides, once inside the host, bacteria will only infect the cells (tissue) that have the adequate receptor. Attachment of bacteria to a host cell is a complicated process managed by adhesin on the bacteria and the receptor on the cell. However, adherence is frequently only the first step in the infection course, that besides implicates internalization, deeper tissue penetration and likely systemic spread. Bacteria have different kinds of elements for adhere to the host surface, including- but not only-pili, fimbriae and in some cases flagella^[3]. Adhesion can be inhibited by the following strategies: (1) prevention of adhesion complex assembly [as in the pili of uropathogenic *Escherichia coli* (*E. coli*)], for which the compounds bicyclic 2-pyridones (pilicides)^[4] and Virstatin^[5] have been developed; and (2) prevention of elongation and formation of a functional pilus.

A search carried out in patent databases^[6,7] revealed a total of 26 patent applications related to strategies that interfere with adhesion and colonization mechanisms (Table 1). These include the use of probiotics such as *Lactobacillus reuteri*, *Bifidobacterium infantis*, *Bifidobacterium lactis*, *Lactobacillus acidophilus* and *Lactobacillus casei*, for in-

hibition of *Candida* colonization, and also *Enterococcus faecium* LJS-01, which displays a strong capacity to adhere to intestinal epithelial cells and also good antimicrobial activity. The following proteins have also been described: decorin binding protein, which prevents colonization by *Borrelia*; collagen binding protein, isolated from *Staphylococcus aureus*; C3 binding polypeptide, isolated from *Streptococcus agalactiae*; novel fluorinated linker compounds; and Zn-releasing calcium phosphate. Finally, the following targets have been identified for vaccine development: capsular polysaccharide, EtpA flagellin and pyruvate-ferredoxin oxidoreductase adhesin protein.

BACTERIAL SECRETORY SYSTEMS

Many bacteria have a specialized excretory system that resembles a syringe through which bacterial toxins (effector proteins) are injected into the host cell. These systems work by imitating host proteins, thus altering the signalling pathways and enabling development of the disease^[8]. Three different secretion systems are implicated in the translocation of bacterial effectors into host cells, III, IV and VI^[9].

The type III secretion system (TTSS) comprises some proteins that form a spire-like construction through which the bacteria inject the effector proteins from the bacterial cytoplasm to the cytoplasm of eukaryotic host cells. These secreted effector proteins often modify signal transduction in the host cells to improve microbial survival, invasion or attachment^[10,11]. The type IV secretory

Table 1 Patents concerning the inhibition of bacterial adhesion and colonization

Patent title	Description	Application date	Inventors	Publication number
Capsular polysaccharide adhesion antigen preparation, purification and use	General method for preparing pure capsular exopolysaccharide adhesins strains of adhesin coagulase-negative staphylococci to produce vaccines	1994	Pier Gerald B	US5980910 (A)
Method for inhibiting microbial adhesion on surfaces	A method for inhibiting microbial adhesion on surfaces in contact with an aqueous system is disclosed and involves adding a treatment comprising an alkyl sulfosuccinate surfactant to the system	1995	Wright J Barry; Michalopoulos Daniel L	US5512186
Method and apparatus for preventing adhesion and colonization of bacteria in medical devices	Activation of compounding photochemicals for preventing and eliminating adherence and colonization of bacteria	1996	Prescott Marvin A	WO9806340 (A1)
Decorin binding protein compositions and methods of use	DNA segments encoding these proteins and anti-(decorin binding protein) antibodies for use in the prevention of <i>Borrelia</i> colonization in an animal	1996	Guo Betty P; Hoeoek Magnus; Hanson Mark	WO9727301 (A1)
Collagen binding protein compositions and methods of use	Disclosed are the <i>cna</i> gene and <i>cna</i> -derived nucleic acid segments from <i>Staphylococcus aur.</i> Also disclosed are Col Binding Protein (CBP) for use in the treatment of pathological infections, and in particular, for use in the prevention of bacterial adhesion to Col	1997	Hoeek Magnus; Patti Joseph M; House-Pompeo Karen; Sthanam Narayana; Symersky Jindrich	US6288214 (B1)
Surfactants for reducing bacterial adhesion onto surfaces	Inhibiting microbial colonization (ethylene oxide units) of a surface in contact with an aqueous system	1997	Donlan Rodney M; Elliot David L; Kapp Nancy J; Wiatr Christopher L; Rey Paula	US6039965 (A)
Composition of treatment of Candidiasis	Inhibition of adhesion of <i>Candida</i> colonization by using probiotics (<i>Lactobacillus reuteri</i> , <i>Bifidobacterium infantis</i> , <i>Bifidobacterium lactis</i> , <i>Lactobacillus acidophilus</i> not viable and non-viable <i>Lactobacillus casei</i>)	1998	Dohnlek Margaret H; Wagner Robert Doug; Balish Edward; Hilty Milo D	WO9917788 (A1)
Antimicrobial adhesion surface	The invention provides an implantable medical device with a hydrophilic coating to limit <i>in vivo</i> colonization of bacteria and fungi	1999	Zhong Samuel P	US6468649 (B1)
Anti-bacterial compounds directed against pilus biogenesis, adhesion and activity; co-crystals of pilus subunits and methods of use thereof	The invention relates to novel compounds that mimic a chaperone G1 beta-strand or an amino terminal motif of a pilus subunit	2000	Hultgren Scott J; Sauer Frederic G; Waksman Gabriel; Fuetterer Klaus; Choudhury Devapriya; Knight Stefan D; Barnhart Michelle	US7041465 (B1)
C3 binding polypeptide of <i>Streptococcus agalactiae</i> group b-Streptococcus	The invention involves the identification of a human complement C3 binding polypeptide and the nucleic acid that encodes the polypeptide from <i>Streptococcus agalactiae</i>	2000	Smith Beverly L; Ferrieri Patricia	US6582950 (B1)
Compounds directed against pilus biogenesis and activity in pathogenic bacteria, methods and compositions for synthesis thereof	Novel fluorinated linker compounds and methods of synthesis are provided. Methods for using the fluorinated linker compounds in methods of solid-phase synthesis of the N-substituted amino acid compounds are also disclosed (inhibiting or preventing the formation of a pilus chaperone-subunit complex)	2000	Kihlberg Jan; Larsson Andreas; Svensson Anette; Fex Tomas; Hultgren Scott J; Pinkner Jerry	WO2001020995
DbpA compositions and methods of use	The DBP gene and decorin protein compositions of <i>Borrelia burgdorferi</i> are disclosed The DBP protein and antigenic epitopes derived from them are contemplated for use in preventing bacterial adhesion to decorin	2000	Guo Betty P; Hook Magnus	US6312907
Composition and method for controlling microbial adhesion and biofilm formation of surfaces	The invention describes how coating of surfaces with an extract, particularly a fish extract, can significantly reduce microbial adhesion, attachment, colonization and biofilm formation on surfaces	2003	Gram Lone Kirsten; Vogel Birtefonnesbech; Bagge-Ravn Dorthe	WO03092382 (A1)
Packaged antimicrobial medical device and method of preparing same	An antimicrobial suture assembly (halogenated hydroxyl ethers, acyloxydiphenyl ethers, and combinations thereof) to substantially inhibit bacterial colonization	2004	Scalzo Howard; Fischer Jerome A; Rothenburger Stephen	US2004220614 (A1)
Sealing material	A sealing material is presented (fluoropolymer layer, a reinforcing layer and an adhesive) to hinder growth and colonization of bacteria	2004	Patel Malay; Napolitano Michael; Hanrahan James R; Chu Chaokang	US2005250398 (A1)

Zn-releasing calcium phosphate (Zn-CaP) compounds for antimicrobial coating on orthodontic appliances and dental implants	Compositions of Zn-releasing calcium phosphate (Zn-CaP) compounds for use as anti-bacterial coatings for orthodontic brackets and dental implants	2006	Legeros Racquel Z; Legeros John P; Park Jae Hyun; Mijares Dindo	WO2007022211 (A2)
Composition for the administration of biologically active principles in gynaecological and rectal conditions and uses thereof	The invention relates to a composition for the administration of biologically active substances in gynaecological and rectal conditions, as well as the uses of said composition	2007	Strozzi Gianpaolo; Mogna Luca	US2010092440 (A1)
Enhanced treatments to kill or debilitate pathogenic microorganisms of a mammalian body	The novel treatments involve the use of anti-adhesive polysaccharide molecules to abolish or reduce the adhesion of <i>Helicobacter pylori</i>	2008	Nifantiev Nikolay; Wieland Gerhard D	US2011245198 (A1)
Non-leaching surface-active film compositions for microbial adhesion prevention	Coating (surfactant) useful to prevent bacterial colonization on a variety of surface including surfaces of medical devices	2008	Gruening Rainer; Qu Xin; Merritt Karen; Chen Paul N; Falevich Vitaly	MX2008009326 (A)
Prevention and treatment of Gram-negative, flagellated bacterial infections	EtpA which binds to the conserved region of the flagellin protein located at the tip of the flagella in Gram-negative bacteria (development vaccine)	2008	Fleckenstein James M	US2011206694 (A1)
Method for coating medical device ¹	Method for coating a medical device to prevent bacterial adhesion, colonization and device-associated infection (isocyanate-terminated polymer)	2010	Stopek Joshua	JP2011019902 (A)
Novel <i>Enterococcus faecium</i> LJS-01 and its use as a probiotic ¹	<i>Enterococcus faecium</i> LJS-01 shows good antimicrobial activity and strong capacity to adhere to intestinal epithelial cells	2010	Lin Chuen-FU; Wu Cheng-Nan; Lu Cheng-Hsiung; Hsu Wei-Li; Chiou Ming-Tang	TW201143631 (A)
Method for detecting colonization characteristic of lactobacillus in gastrointestinal tract on basis of green fluorescent protein ¹	The invention relates to a method for detecting the colonization characteristic of lactobacillus in the gastrointestinal tract on the basis of green fluorescent protein	2011	Yanping Wang; Jingrui Wang; Jinju Wang	CN102604877 (A)
Prevention of bacterial adhesion ¹	Prevention of adhesion of microorganisms on hard surfaces by the semi-permanent modification thereof during the cleaning process. A cleaning agent that contains surface-active polymers is used to prevent the bacterial colonization of hard surfaces	2011	Veith Birgit; Weide Mirko; Corbellini Francesca; Giesen Brigitte; Stumpe Stefan; Breves Roland; Barreleiro Paula; Karten Stefan; Bockmuehl Dirkl; Meier Frank	WO2012010700 (A1)
Pyruvate-ferredoxin oxidoreductase (PFO) adhesive protein as a target for inhibiting the adherence of <i>Trichomonas vaginalis</i> and as a diagnosis and vaccinal target for trichomoniasis ¹	Novel function of PFO upon participating in the cytoadherence of the <i>Trichomonas vaginalis</i> parasite to the hosting cell. The present invention enables development of vaccines for preventing the adhesion (and therefore the colonization) of parasites to the vaginal mucosa	2011	Verastegui Rossana Arroyo	MX2011011361 (A)
Vacuum assisted percutaneous appliance ¹	This device is stabilized by fibroblast in-growth and inhibits bacterial colonization	2012	Kantrowitz Allen B; Mortin Chris; Wadsworth JR Daniel C	US2013006186 (A1)

¹Those published from 2010 onwards are highlighted.

system is utilized to transfer bacterial DNA or bacterial effector proteins to eukaryotic cells. This system also forms a duct between the bacterial and eukaryotic cell cytoplasm. It is a pilus-like structure rather than a spire construction^[9]. Type VI secretion systems form tubular construction; however, exactly how these systems assemble and give effector proteins into the eukaryotic host cells stays in great measure unknown^[12].

Although a lot of various types of TTSS are known, there are a restricted number of manners of inhibiting them: (1) prevention of assembly of the TTSS; (2) inhibition of interplay with the eukaryotic host cells; and (3) inhibition of secretion of the effector proteins.

Three components that are capable of inhibiting bacterial secretion systems have been reported^[13]: (1) inhibitors of the type III secretion systems such as acylated hydrazones

of salicylaldehydes in *Chlamydia* and *Shigella* infections; (2) 2-amino-5-arylidene thiazolidinone in *Salmonella*, *Pseudomonas* and *Yersinia* infections; and (3) dirylacrylonitrile, which inhibits sortase A and has shown *in vitro* activity against *S. aureus*.

A search of the patent database (up to April 2014) revealed 22 patents involving inhibition of the proteins related to secretion systems (Table 2). All of these are based on methods that describe how to identify inhibitors and target proteins of bacterial secretion systems. Two proteins groups are associated with these secretion systems: Inc and HpaB group proteins.

CELL-TO-CELL SIGNALLING: QUORUM SENSING

Cell-cell communication, or quorum sensing (QS), is a

Table 2 Patents concerning the inhibition of bacterial secretion systems

Patent title	Description	Application date	Inventors	Publication number
Method for screening for inhibitors and activators of type III secretion machinery in <i>Gram-negative</i> bacteria	This invention relates to mutant strains of Gram-negative bacteria that constitutively secrete proteins <i>via</i> the type III secretion machinery and to methods of identifying molecules that are able to activate or inhibit secretion in wild-type strains of Gram-negative bacteria	2001	Demers Brigitte; Sansonetti Philippe; Parsot Claude	US6696249 (B1)
Method of detecting substance inhibiting type III secretion mechanism of bacterium and the function of secretory protein thereof	A method whereby a substance specifically inhibiting the type III secretion mechanism and the function of a type III secretory protein secreted therefrom can be detected in large amounts within a short period of time without depending on any animal infection experiments	2001	Omura Satoshi; Abe Akio	KR1020020086208
Secreted <i>Chlamydia</i> polypeptides and method for identifying such polypeptides by their secretion by a type III secretion pathway of a <i>Gram-negative</i> bacteria	The present invention uses a heterologous secretion system, namely a type III system, to investigate whether some <i>Chlamydia</i> proteins, especially Inc proteins and other proteins exhibiting a similar hydropathy profile, might be secreted and demonstrates that these hybrid proteins are secreted by the type III secretion system of <i>Shigella flexneri</i>	2003	Subtil Agathe; Parsot Claude; Dautry-Varsat Alice	US2004131624 (A1)
Bacterial system for protein transport in eukaryotic cells	Development of a system for the targeted transport of proteins into eukaryotic cells by using a type III secretion system and bacteria strains that are mutated in hpaB or homogenous genes. The inventive bacterial system is used to transport bacterial proteins into eukaryotic cells, in order to influence or modify cellular processes such as gene expression, growth, development and defence/resistance mechanisms	2005	Bonas Ulla; Buettner Daniela	WO2005085417 (A2)
Methods of identifying modulators of bacterial type III protein secretion system	Provides methods for identifying inhibitors or activators of bacterial type III protein secretion system by using a recombinant beta-lactamase that can be secreted by a type III protein secretion system. The assay could be easily adapted to a high throughput mode to allow daily screening of several tens of thousands compounds	2005	Goldschmidt Raul; Loeloff Michael	WO2005113791 (A2)
Pharmaceutical composition for the treatment of bacterial infections and sepsis	The invention involves a pharmaceutical composition comprising at least one glycogen synthase kinase 3 β ; inhibitor, at least one Rho-kinase inhibitor, and an optional adequate pharmaceutical carrier for producing a drug for the preventive or therapeutic treatment of bacterial infectious diseases by synergistically increasing synthesis and secretion of type II. A secretory phospholipase A2 into the bloodstream so as to boost the body's inherent resistance to infections	2005	Menschikowski Mario; Hagelgans Albert; Siegert Gabriele	WO2005120475
Pyridone compounds as inhibitors of bacterial type III protein secretion systems	Provides compounds that inhibit type III protein secretion useful for the treatment and prevention of bacterial infections, particularly those caused by <i>Gram-negative</i> bacteria, and methods for their use	2005	Li Xiaobing	US2005256137 (A1)
Methods for stimulating an immune response using bacterial antigen delivery system	Provides methods for stimulating and/or increasing an immune response against tumor antigens through the use of the type III secretion system of bacteria. The invention also relates to the preparation of antigen presenting cells from peripheral blood mononuclear cells by using bacteria with a type III secretion system	2006	Old Lloyd J; Ritter Gerd; Nishikawa Hiroyoshi; Gnjjatic Sacha; Galan Jorge E	US2009324651 (A1)
Screening system for inhibitors and activators of type III secretion machinery in <i>Gram-negative</i> bacteria	Provides a screening system (comprising inhibitors and activators of type III secretion machinery) that directly transfers pathogenic proteins of <i>Gram-negative</i> bacteria into a host cell to identify substances capable of activating or inhibiting the secretion of type III protein secretion system	2006	Hwang In Gyu; Moon Jae Sun; Kim Sung Uk	KR20080051240 (A)
Application of bovine lactoferrin for preparing a medicinal agent for inhibition of bacteria growth	The invention refers to a new application of bovine lactoferrin for preparing a medicinal agent for inhibiting bacteria growth. The bovine lactoferrin inhibits the growth of bacterial pathogens expressing the type III secretory system	2007	Makmakhon Robert Dzh; Kliari Tomas; Ochoa Tereza	RU2007140789 (A)
Bacterial secretion system and uses	-	2007	Gey Van Pittius Nicolaas Claudius; Warren Robin Mark; Van Helden Paul David	ZA200706520 (A)
Biopolymer and protein production using type III secretion systems of <i>Gram-negative</i> bacteria	Provides proteins, polynucleotide, expression cassette, vector and bacterium compositions for obtaining proteins of interest by expression of same in Gram-negative bacteria with a type III secretion system. Also provides uses for the proteins obtained in the manufacture of isolated proteins and pharmaceutical compositions	2007	Voigt Christopher Ashby; Widmaier Daniel Matthew	WO2008019183 (A2)

Use of the <i>Pseudomonas syringae</i> effector protein HopU1 related to its ability to ADP-ribosylate eukaryotic RNA binding proteins	The invention provides novel methods for modulation of the innate immune response of a plant to infection caused by <i>Pseudomonas syringae</i> , which injects effector proteins into host cells via a type III protein secretion system. Also provides methods for enhancing or suppressing the innate immune response of the plant	2007	Alfano James R; Fu Zheng Qing; Elthon Thomas E	WO2008042026 (A2)
Method and means for preventing and inhibiting type III secretion in infections caused by Gram-negative bacteria	Discloses a means of decreasing bacterial virulence in a mammal or in a plant by inhibition of the type III secretion system at concentrations that do not prevent or substantially reduce bacterial growth. Also disclosed are a therapeutic method and a pharmaceutical composition	2008	Elofsson Mikael	US2010099674 (A1)
Carboplatin compound inhibiting secretion system of phytopathogenic Gram-negative bacteria and biocontrol agent of plant diseases with this compound	Provides an agent for preventing plant diseases, containing carboplatin compounds, to selectively suppress secretion system related to plant pathogenicity	2009	-	KR20110048335 (A)
Inhibition of quorum sensing-mediated processes in bacteria	Provides methods for identifying molecules that can be used to positively and negatively manipulate quorum-sensing-mediated communication to control bacterial behavior. Methods of inhibiting quorum sensing-mediated activity in Gram-negative bacteria are provided wherein the activity is pathogenicity, bioluminescence, siderophore production, type III secretion, or metalloprotease production	2009	Bassler Bonnie; Swem Lee	US2011123586 (A1)
Type III secretion inhibitors, analogs and uses thereof	The invention relates to compounds and compositions useful for inhibiting type III secretion systems in pathogenic bacteria, such as <i>Yersinia pestis</i> , and uses of such inhibitors in the treatment and prevention of disease	2009	Goguen Jon; Pan Ning; Lee Kyungae	US2011034463 (A1)
5-substituted-2-imino-thiazolidinone compounds and their use as inhibitors of bacterial infection ¹	Provides a method for inhibiting Gram-negative bacterial pathogenesis, a method of screening for compounds that inhibit type III secretion in <i>Gram-negative</i> bacteria, and compounds that inhibit type III secretion in <i>Gram-negative</i> bacteria	2010	Felise Heather B; Miller Samuel I; Kline Toni	US2011039849 (A1)
Methods for Identifying Inhibitors of the type III Secretion System ¹	Provides a method for determining whether a test compound can inhibit the function of the type III secretion system. The method identifies drug candidates that are highly specific anti-bacterial agents for treating diseases caused by Gram-negative bacteria with a T3SS	2010	Marlovits Thomas C; Radics Julia; Schmied Wolfgang	US2013130283 (A1)
Attenuated <i>Salmonella</i> inducible secretory expression oral vaccine presentation system and application thereof ¹	The invention comprises an attenuated salmonella inducible secretory expression oral vaccine presentation system containing an antigen expression carrier. The system is controlled by a promoter induced by induction of a bacteria excretion signal, and it uses the attenuated salmonella as the host of the antigen expression carrier	2011	Zichun Hua; Guo Chen	CN102335421 (A)
Bacterial mediated delivery of nuclear protein into pluripotent and differentiated cells ¹	A modified <i>Pseudomonas aeruginosa</i> type III secretion system has been developed that efficiently delivers selected proteins into a host cell	2011	Jin Shouguang; Bichsel Candace	WO2012012605 (A2)
Inhibitors of bacterial type III secretion system ¹	Discloses organic compounds showing the ability to inhibit effector toxin secretion or translocation mediated by bacterial type III secretion systems. These inhibitor compounds are useful for combating infections by <i>Gram-negative</i> bacteria with such type III secretion systems	2012	Moir Donald T.; Aiello Daniel; Peet Norton P; Williams John D; Torhan Matthew	US2014142134 (A1)

¹Those published from 2010 onwards are highlighted.

widespread phenomenon in bacteria that is used to coordinate gene expression between local populations^[13]. Bacterial populations can use QS communication to coordinate the execution of important biological functions, many of which are involved in pathogen virulence, *e.g.*, biofilm formation, extracellular polysaccharide production, host colonization, motility, bioluminescence, transfer of plasmids by conjugation, and biosynthesis of antibiotics and siderophores.

All QS systems utilize small, secreted signalling molecules known as autoinducers (AIs): (1) AI-1 molecules are N-acyl-homoserine lactones (AHLs); (2) AI-2 molecules are heterocyclic furanosyl-borates; (3) AI-3 signals are catecholamines and finally; and (4) AI-4 signals are cyclic peptides. Some other QS signals go beyond these classes,

e.g., *Pseudomonas* quinolone signal and diffusible signal factor. New molecules will undoubtedly be discovered as the study of QS expands to species of bacteria yet to be investigated.

Targeting bacterial virulence (quorum quenchers, Figure 1) is an alternative focusing to antimicrobial therapy that offers a hopeful opportunity to inhibit pathogenesis and its consequences without producing immediately the death the target bacterium. Bacterial virulence factors have been shown to be potential targets for drug design and therapeutic intervention for Gram-negative pathogens^[1]. Numerous quorum sensing inhibitors have been reported in the literature^[1,2].

In 2012, Romero *et al.*^[14] published an article about patents concerning quorum quenching (QQ) (*i.e.*, the

mechanisms that cause the inactivation of QS communication systems)^[15].

A search of patent databases (up to August 2011) revealed a total of 45 applications related to strategies for interfering with QS systems as a method of fighting microbial infections. Following the bias in the literature, the vast majority of the patented technologies based on the inhibition of QS mechanisms target AHL signals, whereas only 5 out of 45 patent cases are based on the inhibition of AI-2 signals and only 4 are based on the inhibition of peptide-based QS signals from Gram-positive bacteria. In this review, a search of more recent reports (up to April 2014) was conducted, revealing 32 patents concerning the inhibition of QS systems (Table 3). QQ occurs in *Lysobacter enzymogenes*, *Shewanella piezotolerans*, *Bacillus pumilus*, *Tenacibaculum discolor* (cect 7426) and novel alpha-proteobacteria. The molecules involved in QQ distinguish inhibitors of the AI-2 signals (triazol derivatives, furan compounds and phosphorylated, branched dihydroxy-pentane-dione) and inhibitors of AHLs [(oxododecanoyl)-L-homoserine lactone and bicyclic furanones with low toxicity]. Furanones, which are naturally occurring compounds, appear to be the most widely studied QQ compounds. These compounds are toxic to *Artemia* and rotifers, which will limit their use in humans^[15]. However, the use of C-30, a synthetic furanone, at non toxic concentrations, significantly reduced the pathogenicity of *Vibrio anguillarum* in rainbow trout^[16]. Other patented compounds involved in QQ include honaucin A, 2-methylthiopyrrolidines, lovastatin and hydroxytyrosol. Finally, one enzyme (OLB-26) is known to be involved in QQ.

MECHANISMS OF RESISTANCE: MULTIDRUG RESISTANCE PUMPS

Multidrug resistance (MDR) efflux pumps have multiple functions in natural microbial ecosystems. In clinical environments, MDR pumps are implicated in the following: (1) resistance to antimicrobial compounds localized on mucosal surfaces (colonization factor)^[17,18]; (2) efflux virulence factors^[19]; (3) QS-regulated expression of virulence traits^[20]; and (4) antibiotic resistance, which is a code element in patients under treatment^[21]. All of these roles are important for the survival, colonization and pathogenic outcome of virulent bacteria in clinical environments. In nonclinical environments, MDR pumps may be associated to resistance to heavy metals^[22] and organic solvents^[23] (plant colonization factor). Bacterial efflux pumps are classified into five families according to their composition, number of transmembrane spanning regions, energy sources and substrates: the resistance-nodulation-division (RND) family, the major facilitator superfamily, the adenosine triphosphate-binding cassette superfamily, the small multidrug resistance family, and the multidrug and toxic compound extrusion family^[17,18].

Several MDR pump inhibitors were published^[24]. We consider two examples of RND inhibitors pumps: (1)

1-(1-naphthylmethyl)-piperazine and phenyl-arginine- β -naphthylamide. These act as inhibitors of RND efflux pumps and virulence traits in *Vibrio cholerae*, such as the cholera toxin and the toxin-coregulated pilus^[25], and were suggested as a suitable tool for the treatment of cholera infections; and (2) Of 12 trifluoromethyl ketone compounds tested, 6 proved to be effective inhibitors of the quorum-sensing response by *Chromobacterium violaceum* 026, as well as inhibitors of the RND efflux pumps of CV026 and *E. coli*. This result is of clinical applicability and may be used for the prevention of QS responses of infecting bacteria^[26].

We found 22 patents related to the inhibition of the MDR efflux in patent databases up to April 2014 (Table 4). Most of these are screening methods for microbial efflux pump inhibitors. Moreover, pump inhibitors are described as potentiators of the action of antiseptics, disinfectants and antimicrobial agents such as tigecycline. Finally, polyamine molecules are inhibitors of bacterial efflux pumps that could be used in combination with other drugs such as antibiotics, as well as pharmaceutical compositions thereof.

FUTURE PROSPECTS: RESISTANCE TO ANTIVIRULENCE COMPOUNDS

Although antivirulence therapies are novel in the field of treatment of infectious diseases, several studies involving clinical strains have demonstrated the development of mechanisms of resistance, especially to Quorum Quenching compounds^[27]. *Vibrio cholerae* strains that are resistant to virstatin have also been described; the mechanisms whereby these strains colonize their hosts are independent of the elaboration of the toxin co-regulated pilus^[28].

The first evidence that cells develop resistance to QQ compounds has been reported by Maeda *et al.*^[29] (published ahead of print in 2011). These authors worked with a concentration of brominated furanone C-30 [the gold standard for QQ compounds, and which is a synthetic brominated furanone 4-bromo-5-(bromomethylene)-2(5H)-furanone] that did not influence growth in rich medium (*i.e.*, it did not inhibit growth) and utilized both transposon mutagenesis and spontaneous mutants to detect resistant bacteria.

The mechanism of this resistance was overexpression of the MexAB-OprM multidrug resistance operon due to mutations in the gene repressors *mexR* and *nalC*, resulting in efflux of the compound C-30. This quorum quenching compound showed a reduced capacity to decrease some QS-controlled virulence traits and phenotypes in the *mexR* mutant, and the pathogenicity of the *mexR* mutant against the nematode *Caenorhabditis elegans* was not decrease by the inclusion of C-30. Importantly, these authors also worked with cells from cystic fibrosis patients (Liverpool epidemic strain 12142) with *mexR* and *nalC* mutations^[30] to demonstrate that, even in the absence of the QS inhibitor, cells develop resistance to quorum quenching compounds in the pathogenic state when

Table 3 Patents concerning the inhibition of quorum sensing systems

Patent title	Description	Application date	Inventors	Publication number
Anti-inflammatory and quorum sensing inhibition compounds and methods of making and using them	This invention generally relates to novel compositions based on a structure designated as "Honaucin A", including Honaucin A variants and analogs, and pharmaceutical compositions, liposomes and nanoparticles comprising them, and methods of making and using them	2011	Gerwick William H; Gerwick Lena; Choi Huykjae; Villa Francisco A; Smith Jennifer; Rowley David C	WO2011153502 (A2)
Composition for oral use	A method for suppressing dental caries by regulating biofilm formation by the bacteria that cause dental caries instead of controlling these bacteria	2011	Tsugane Takanori; Saeki Yoji	EP2620160
Conjugates of acyl homoserine lactone and catalase a from <i>Pseudomonas aeruginosa</i>	The present invention relates to the acyl homoserine lactone N-3-(oxododecanoyl)-L-homoserine lactone or butyryl L-homoserine lactone and <i>Pseudomonas aeruginosa</i> KatA protein, or an antigenic portion conjugate thereof, used to treat <i>Pseudomonas aeruginosa</i> infections by limiting biofilm formation and inhibiting a range of quorum-sensing dependent virulence factors	2011	Kyd Jennelle M; Cooley Margaret	WO/2012/083382
Enzyme bag containing quorum quenching enzyme immobilized silica for inhibiting biofilm formation and membrane bioreactor system for water treatment system using the bag	This invention relates to an enzyme bag containing silica-immobilized enzyme for inhibiting biofilm formation. A membrane bioreactor system for water treatment using the bag is provided for stable implementation of filtering operations over a long period of time by improving the performance of operational processes	2011	Lee Chung Hak; Yang Cheon Seok; Lee Jung Kee; Han Jong Yun; Lee Chung Hak; Yang Cheon Seok; Lee Jung Kee; Han Jong Yun	KR20120134724 (A)
Fluidizable carrier with biofilm formation-inhibiting microorganisms immobilized therein and membrane water treatment apparatus using the same	This invention relates to a biofilm formation-inhibiting microorganism immobilized fluidizable carrier in which a biofilm formation-inhibiting microorganism is fixed therein and a membrane water treatment apparatus including the same are provided to increase a membrane cleaning period	2011	Lee Chung Hak; Kim Sang Ryoung; Lee Jung Kee	KR20130034935 (A)
Methods of disrupting quorum sensing to affect microbial population cell density	The invention relates to the modulation of quorum sensing mechanisms in a microorganism for the purpose of exploiting the fermentation capabilities of the microorganism	2011	Marrs Barry; Swalla Brian M	US2011124522 (A1)
Quorum-sensing signal molecular preparation and application thereof in tobacco waste treatment	The invention relates to the field of environmental biotechnology, in particular to preparation of a quorum sensing signal molecule and application in processing tobacco waste	2011	Meizhen Wang; Hongzhen He; Huajun Feng; Xin Zheng; Dongsheng Shen; Zhenmei LV; Hang Min	CN102392051 (A)
Synthetic analogs of bacterial quorum sensors	The invention relates to synthetic analogs of bacterial quorum sensing molecules, and methods of use of these	2011	Iyer Rash; Ganguly Kumkum; Silks Louis A	US2012071430 (A1)
System and method for reversing the antibiotic tolerance of bacterial persister cells	The present invention relates to antibiotics and, more particularly, to a system and method for decreasing the tolerance of bacterial persister cells to antibiotics	2011	Ren Dacheng; Pan Jiachuan	EP2603576
Triazole compounds as well as preparation method and application thereof	The present invention relates to triazole derivatives, the preparation method and as the autoinducer-2 (AI-2) quorum sensing inhibitors, belonging to anti-AI-2 quorum sensing type drug technology	2011	Minyong Li; Lvpei Du; Peng Zhu	CN102219753 (A)
Use of a novel alpha-proteobacteria for quorum quenching	The invention (concerning the fields of biology, molecular biology, and aquaculture) specifically relates to a new alpha-proteobacteria capable of degrading/V-acyl-homoserine lactones (AHLs) for control of bacterial infectious diseases and prevention of biofilm formation	2011	Otero Casal Ana Maria; Romero Bernardez Manuel	WO2011154585 (A1)
2-methylthiopyrrolidines and their use for modulating bacterial quorum sensing	Formula (I) compounds are disclosed and their use in inhibiting quorum sensing in bacteria is reported	2012	Malladi Venkata L; Schnepfer Lisa; Sobczak Adam J; Mathee Kalai; Wnuk Stanislaw F	WO/2012/174511
Compositions for regulating or modulating quorum sensing in bacteria, methods of using the compounds, and methods of regulating or modulating quorum sensing in bacteria	The report encompasses compounds and compositions that are useful as specific AI-2 antagonists for the control of bacterial quorum sensing and methods for inhibiting or attenuating microbial virulence, biofilm formation and drug resistance	2012	Wang Binghe; Ni Nanting; Wang Junfeng; Lu Chung-Dar; Chou Han-Ting; Li Minyong; Zheng Shilong; Cheng Yunfeng; Peng Hanjing	EP2529793 (A2)
Construction and application of unmarked <i>Lysobacter enzymogenes</i> engineering strain capable of preventing plant bacteriosis	The invention (within the field of microbial genetic engineering), specifically relates to a plant bacterial disease that can prevent dissolving enzyme production strains of <i>Bacillus</i> unmarked engineering construction and application	2012	Liu Fengquan; Qian Guoliang	CN102943061 (A)

Furan compound and preparation method and application of furan compound	The invention relates to furan derivatives, the method of preparation and as the AI-2 quorum sensing inhibitors, are anti-AI-2 type of quorum sensing	2012	Minyong Li; Peng Zhu	CN102603683 (A)
Method for increasing output of microbial lovastatin based on quorum sensing mechanism	The invention relates to the pharmaceutical raw material fermentation industry, in particular to a method of increasing microbial production of lovastatin	2012	Li Haoming	CN102925509 (A)
Method for quickly identifying food-borne pathogen bacterial biofilm formation inhibitor	The invention relates to the field of food microbiology control technology, in particular to the rapid identification of an inhibitor of foodborne bacterial biofilm formation	2012	Wenyang Zhang; Hongmei Zhang; Zhihua Tao; Wenyuan Zhou	CN102706821 (A)
Phosphorylated and branched dihydroxy-pentane-one analogs as quorum sensing inhibitors in bacteria	The invention provides compositions and methods for modulating quorum sensing in microbes and can be used in prophylactic methods or therapy for bacterial infections and for reduction of biofilms. The compounds are AI-2 analogs and as such have structures similar to 4,5-dihydroxy-2,3-pentanedione that can act as agonists/antagonists of quorum sensing	2012	Sintim Herman; Bentley William E; Roy Yarnika; Smith Jacqueline	US2012294900 (A1)
Preparation method of imprinted polymer of bacterial quorum sensing signal molecule AI-1	The present invention relates to bacterial quorum sensing signal molecules AI-1 imprinted polymer preparation	2012	Xin Li; Ling Wang	CN102604010 (A)
Probiotics for biological control against <i>Vibrio</i> sp.	The invention relates to probiotics for biological control against <i>Vibrio</i> sp., and in particular, to a newly isolated bacillus strain that degrades quorum-sensing signal molecules of the pathogenic bacteria <i>Vibrio</i> sp., and inhibits biofilm formation	2012	Yang Si Yong; Woo Seo Hyung; Kang In Hye; Im Hyun Jung	WO2012105805
Quorum sensing inhibitor against a pathogenic microorganism, and an antibacterial composition using the same	A quorum sensing inhibitor and an antibacterial composition using the same are provided to suppress quorum sensing between bacteria, and to prevent or treat infection or diseases	2012	Undescribed	KR101243696
<i>Shewanella piezotolerans</i> 34# and application thereof to algae inhibition	The invention relates to the field of biotechnology, in particular to a marine bacterium <i>Shewanella</i> and the inhibition of algal growth	2012	Zhou Jin; Yin Peng	CN103173383 (A)
Simple method for testing disease resistance of pathogenic bacteria quorum-quenching gene prokaryotic expression product	The invention relates to a prokaryotic expression product of disease resistance testing methods, in particular to test pathogen populations prokaryotic expression product quenching effect of a simple disease, and belongs to the field of gene function identification techniques	2012	Ouyang Lejun; Huang Zhenchi; Zeng Fuhua; Li Limei; Li Heng	CN102972220 (A)
Use of quorum sensing inhibitors and biofilm dispersing agents for controlling biofilm-associated implantable medical device related infections	The invention generally relates to implantable medical devices and, more specifically, to the use of quorum sensing inhibitors and/or biofilm dispersing agents to control biofilm-associated infections related to the use of implantable medical devices	2012	Samade Richard; Dinesh Prashant; Nabutovsky Yelena; Bornzin Gene A; Poore John W; Karicherla Annapurna; Dalal Nirav	US2014005605 (A1)
<i>Bacillus pumillus</i> microbial preparation with quorum sensing system inhibiting effect	The invention relates to <i>Bacillus pumillus</i> with a quorum sensing system inhibiting effect. The invention has the advantages that <i>Chromobacterium violaceum</i> is used for screening out a bacterial strain F3-1. It can be used to produce a microbial preparation capable of preventing and treating aquatic bacterial diseases	2013	Song Zengfu; Fan Bin; Chen Biao	CN103525723 (A)
Bicyclic furanones with low toxicity for microbial control	The invention relates to a class of bicyclic brominated furanone structures with reduced toxicity and high activity for inhibiting biofilm formation and quorum sensing by microbes	2013	Luk Yan-Yeung; Yang Sijie	US2013197077 (A1)
Method for detecting quorum sensing quenching bacterial strain	The invention relates to a method for detecting a quorum sensing quenching bacterial strain. The method comprises the addition of a bacterial strain to be detected in a PIPES (1,4-piperazinediethanesulfonic acid) buffer solution of pH 6	2013	Zhang Xiaohua; Tang Kaihao; Shi Xiaochong; Zhang Yunhui	CN103215342 (A)
Quorum-quenching enzyme OLB-26, and coding gene and application thereof	The invention relates to a quorum sensing quenching enzyme OLB-26 and its coding gene and application	2013	Zhou Zhigang; Zhang Meichao; Yang Yalin; Xu Li; He Suxu; Li Qing; Yu Qiang	CN103275949 (A)
Targeted enzymatic degradation of quorum-sensing peptides	The present invention generally relates to the fields of microbiology and wound care. More particularly, it concerns methods and compositions for inhibiting biofilms in wounds and on medical devices	2013	Alarcon Rodolfo; McNulty Amy K	US20130253382
Use of ellagitannins as inhibitors of bacterial quorum sensing	Materials and methods for the inhibition of bacterial QS are described. Methods of treating bacterial infections by administration of one or more ellagitannins in amounts effective for inhibiting bacterial QS are also provided	2013	Athee Kalai; Adonizio Allison L; Ausubel Frederick; Clardy Jon; Bennett Bradley; Downum Kelsey	US2013317094 (A1)
Use of hydroxytyrosol and derivatives thereof as quorum quenchers	The quorum quenching activity of formula (I) or (II) compounds, such as hydroxytyrosol (HT), hydroxytyrosol acetate (HTA), 3,4-dihydroxyphenylacetic acid (DOPAC) and derivatives thereof are described	2013	Au Ón Calles David; Allende Prieto Ana; Fábregas Casal Jaime; Gómez-Acebo Gullón Eduardo	WO2014060581 (A1)
Use of the cect 7426 strain for generating quorum quenching of the autoinducer-2 signal (ai-2)	The invention relates to the use of a bacterial strain of the species <i>Tenacibaculum discolor</i> in the control of infectious diseases and for inhibiting biofilm formation caused by bacteria, through the inhibition of quorum sensing signals type AI-2. The invention applies to the field of molecular biology	2013	Otero Casal, Ana María; Romero Bernárdez Manuel	WO2014057151 (A1)

Table 4 Patents concerning the inhibition of multidrug resistance systems

Patent title	Description	Application date	Inventors	Publication number
Method for screening for non-tetracycline efflux pump inhibitors	Provides screening methods for inhibitors of microbial efflux pumps and pharmaceutical compositions containing such efflux pump inhibitors as well as methods for treating microbial infections by use of these compositions	1995	Trias; Joaquim Chamberland; Suzanne Hecker; Scott J Lee; Ving J	US5989832 A
Efflux pump inhibitors	Provides screening methods for inhibitors of microbial efflux pumps and pharmaceutical compositions containing such efflux pump inhibitors. Also provides methods for treating microbial infections using those compositions	1996	Trias Joaquim; Hecker Scott J; Chamberland Suzanne; Lee Ving J	CA2217865
Methods and compositions for reducing bacterial tolerance of disinfectants and organic solvents	Methods and compositions useful for manipulating bacterial resistance to non-antibiotic antibacterial compositions, disinfectants and organic solvents, and for rendering bacterial cells susceptible to non-antibiotic antibacterial compositions	1997	Levy Stuart B	WO9917607 (A2)
Methods and compositions for reducing bacterial tolerance to antibacterials, disinfectants and organic solvents	Methods and compositions useful for manipulating bacterial resistance to non-antibiotic antibacterial compositions, disinfectants and organic solvents, and methods for rendering bacterial cells susceptible to non-antibiotic antibacterial compositions	1997	Levy Stuart B	US6068972 (A)
Inhibitors of cellular efflux pumps of microbes	Describes compounds that are inhibitors of efflux pump in bacteria. Also describes methods of preparing and using such compounds and the pharmaceutical compositions that include such compounds	2001	De Souza Noel John; Patel Mahesh Vithalbhaj; Gupte Shrikant V; Upadhyay Dilip J; Shukla Milind Chintaman; Chaturvedi Nishith C; Bhawsar Satish B; Nair Sheela Chandresekharan; Jafri Mohammed A; Khorakiwala Habil Fakhruddin	US2002177559 (A1)
Drug discovery and increased potency of antiseptics and disinfectants based on high extracellular pH, the disablement of cellular efflux pumps, and the unexpected synergism therebetween	Methods for increasing the therapeutic potency of amphipathic compounds, <i>e.g.</i> , antiseptics and disinfectants, and the exploitation of these discoveries in the screening of small molecules, and libraries thereof, for biological activity in prokaryotes and eukaryotes	2002	Lewis Kim; Hsieh Peichung	US2003118541 (A1)
Methods to study and mechanisms of biofilm-related antibiotic resistance	Discloses various genes that encode proteins shown to play a role in microbial resistance of an organism in a biofilm and homologs thereof. Describes methods of identifying a compound that modulates microbial resistance of an organism in a biofilm and also genes encoding proteins that play a role in biofilm resistance	2002	O'Toole George A; Mah Thien-Fah	US2003166030 (A1)
Minicell-based gene therapy	Provides compositions and methods for the production of achromosomal and anucleate cells useful for applications such as diagnostic and therapeutic uses, as well as research tools and agents for drug discovery	2002	Sabbadini Roger A; Berkley Neil; Surber Mark W	US2003199088 (A1)
Potentiators of antibacterial activity	The invention relates to compounds that potentiate the activity of antibacterials, compositions useful in treating bacterial infection, and related methods. Also relates to a method of inhibiting bacterial efflux of an antibiotic, thereby increasing its efficacy	2004	Grossman Trudy H	US20040887719 20040709
Substituted polyamines as inhibitors of bacterial efflux pumps	Methods of treating bacterial infections, including those caused by multidrug resistant bacteria, by using polyamine efflux pump inhibiting compounds, optionally in combination with other drugs such as antibiotics. The pharmaceutical compositions of the polyamines are also reported	2004	Nelson Mark L; Alekshun Michael N	US2004204378 (A1)
Essential and important genes of <i>Pseudomonas aeruginosa</i> and the use thereof to design or identify antibacterial agents	Database of candidate essential genes in <i>Pseudomonas aeruginosa</i> , and other important genes that, when mutated, produce a growth attenuated phenotype. The invention includes methods for confirming the need for or importance of candidate genes, methods for using those genes to screen for new antibacterial drugs, the antibacterial agents identified using the disclosed methods, and also methods of using the same for treating and preventing <i>Pseudomonas</i> infection	2005	Bruce Kim F; Warrener Paul; Hou Kevin	US2007196829 (A1)
Method for increasing the susceptibility of peptide deformylase inhibitors by using efflux pump inhibitors	Provides methods and compositions for increasing the susceptibility of PDF inhibitors against Gram-negative organisms by using efflux pump inhibitors	2005	Dean Charles Richard; Ryder Neil Stewart	CA2569681

Products and methods for <i>in vivo</i> secretion of monatin	Provides products and methods for the <i>in vivo</i> production of monatin sweetener. The products include microorganisms that are genetically modified to secrete or to improve secretion of monatin, to produce monatin, and to both secrete/ improve secretion and produce monatin	2006	Laplaza Jose; Anderson James C; Desouza Mervyn L; Hicks Paula M; Kollmann Sherry R	WO2006113897
Rhamnose-inducible expression systems and methods	Describes rhamnose-inducible expression constructs which may include at least one rhamnose-inducible regulatory element expressing a regulatory protein and one promoter that is inducible by the regulatory protein. An open reading frame expressing a protein of interest may be placed under control of the promoter. Also describes optimized Shine-Dalgarno sequences for use with the promoter	2006	Surber Mark W	US2007122881 (A1)
Enhancement of tigecycline potency using efflux pump inhibitors	Discloses Efflux Pump Inhibitor (EPI) compounds that can be co-administered with antimicrobial agents for the treatment of infections caused by drug resistant pathogens and methods of treatment and pharmaceutical compositions for co-administering tigecycline with an EPI	2007	Glinka Tomasz; Bostian Keith; Lomovskaya Olga; Surber Mark; Sun Dongxu	US20070832626 20070801
Method or agent for inhibiting the function of efflux pump of <i>Pseudomonas aeruginosa</i>	Discloses a method comprised of modifying any amino acid residue selected from 100 th to 109 th and 311 th to 320 th amino acid residues in an amino acid sequence for mature OprM protein, as well as an agent for inhibiting the function of an efflux pump of <i>Pseudomonas aeruginosa</i> with good efficiency. Further disclosed is a method for screening the agent	2007	Yoshihara Eisaku; Inoko Hidatoshi	CA2641988
Method or agent for inhibiting the function of efflux pump of <i>Pseudomonas aeruginosa</i>	Provides a method for inhibiting the function of the drug efflux pump of <i>Pseudomonas aeruginosa</i> , comprising modification of any amino acid residue selected from 100 th to 109 th or 311 th to 320 th amino acid residues in the amino acid sequence of mature OprM protein. Also reports an agent with such inhibitory effect, as well as a screening method	2007	Yoshihara Eisaku; Inoko Hidatoshi	WO2007/091395
Microbial production of aromatic acids	Method for the microbial production of aromatic acids from a fermentable carbon substrate using a host cell capable of producing said aromatic acid, and comprising an efflux pump for said aromatic acid. A preferred host cell comprises a member of the proton-dependent resistance/nodulation/cell division (RND) family of efflux pumps	2007	Wery Jan	US2007259409 (A1)
Near-infrared electromagnetic modification of cellular steady-state membrane potentials	Discloses systems and methods for applying near-infrared optical energies and dosimetries to alter the bioenergetic steady-state trans-membrane and mitochondrial potentials (DeltaPsi-steady) of all irradiated cells through an optical depolarization effect. This membrane depolarization provides the ability to potentiate antimicrobial, antifungal and/or antineoplastic drugs against only targeted undesirable cells	2007	Bornstein Eric	US2008139992
<i>In vivo</i> gene sensors	Describes methods and compositions for the detection of target genes that permit the selective expression of an effector gene in those cells expressing the target gene, thus selectively targeting these cells for treatment or elimination. The methods and compositions described may also permit the selective expression of an agent such as a therapeutic gene product, in a targeted population of cells	2009	Collins James J; Lu Timothy Kuan-Ta	WO2009/137136
Methods of reducing microbial resistance to drugs	Provides methods of treating infection, screening for inhibitors of AcrAB-like efflux pumps, and enhancing antimicrobial activity of drugs. Pharmaceutical composition comprising an inhibitor of an AcrAB-like efflux pump and an antimicrobial agent are also provided	2009	Oethinger Margaret; Levy Stuart B	US2009298873
Minicells displaying antibodies or derivatives thereof and comprising biologically active compounds ¹	Minicells are used to deliver biologically active compounds, including polypeptides, nucleic acids, small molecules, drug molecules, and chemotherapeutic agents. In some cases, the minicell displays ligands or binding moieties that target the minicell to a desired host cell	2011	Sabbadini Roger A; Berkley Neil; Surber Mark W; Klepper Robert	US20070725196 20070316

¹Those published from 2010 onwards are highlighted.

coexisted with the pressures of antibiotic treatment; so, antimicrobial treatment can induce to resistance to QQ compounds.

Intensified efforts are needed to establish whether resistance may develop to other QQ compounds, as is the case of lactonase or acylase enzymes in connection with AHL autoinducers.

CONCLUSION

Antimicrobial resistance in nosocomial pathogens is in-

creasing worldwide. Mortality rates of patients infected with drug-resistant pathogens have increased by approximately 50% in recent years. Hospitals have become breeding grounds for extremely resistant pathogens, exacerbating the risk associated with hospitalization. These pathogens are extremely resistant to last line antimicrobials. If the current trend continues, the beginning of a “post-antibiotic era” is predicted. Development of novel antibiotics has almost totally ceased, at least against Gram-negative bacteria, and the prospects for a reversal of this trend are bleak. It is therefore imperative to de-

velop new molecules, therapies and/or new combinations of these for the eradication of resistant pathogens. In this review, we discuss some examples of patented molecules that act by inhibiting different bacterial virulence mechanisms (adhesion/colonization and quorum sensing mechanisms, and secretory and efflux pump systems) and which open the way to studying potential new treatments for infections caused by multiresistant bacteria.

Novel targets and molecules must be discovered for antivirulence therapies, taking into account the possible development of resistance mechanisms. Further study of combinations of these compounds with other antimicrobials for the treatment of infectious diseases is also important.

REFERENCES

- Rasko DA**, Sperandio V. Anti-virulence strategies to combat bacteria-mediated disease. *Nat Rev Drug Discov* 2010; **9**: 117-128 [PMID: 20081869 DOI: 10.1038/nrd3013]
- Beceiro A**, Tomás M, Bou G. Antimicrobial resistance and virulence: a successful or deleterious association in the bacterial world? *Clin Microbiol Rev* 2013; **26**: 185-230 [PMID: 23554414]
- Proft T**, Baker EN. Pili in Gram-negative and Gram-positive bacteria - structure, assembly and their role in disease. *Cell Mol Life Sci* 2009; **66**: 613-635 [PMID: 18953686 DOI: 10.1007/s00018-008-8477-4]
- Svensson A**, Larsson A, Emtenäš H, Hedenström M, Fex T, Hultgren SJ, Pinkner JS, Almqvist F, Kihlberg J. Design and evaluation of plicicides: potential novel antibacterial agents directed against uropathogenic *Escherichia coli*. *Chembiochem* 2001; **2**: 915-918 [PMID: 11948880]
- Shakhnovich EA**, Hung DT, Pierson E, Lee K, Mekalanos JJ. Virstatin inhibits dimerization of the transcriptional activator ToxT. *Proc Natl Acad Sci USA* 2007; **104**: 2372-2377 [PMID: 17283330]
- European Patent Office**. Espacenet: Patent search. [accessed 2014 April]. Available from: URL: <http://worldwide.espacenet.com>
- United States Patent and Trademark Office**. Patent Full-Text and Image Database. [accessed 2014 April]. Available from: URL: <http://patft.uspto.gov/netahtml/PTO/search-adv.htm>
- Galán JE**, Cossart P. Host-pathogen interactions: a diversity of themes, a variety of molecular machines. *Curr Opin Microbiol* 2005; **8**: 1-3 [PMID: 15694849]
- Filloux A**, Hachani A, Bleves S. The bacterial type VI secretion machine: yet another player for protein transport across membranes. *Microbiology* 2008; **154**: 1570-1583 [PMID: 18524912 DOI: 10.1099/mic.0.2008/016840-0]
- Keyser P**, Elofsson M, Rosell S, Wolf-Watz H. Virulence blockers as alternatives to antibiotics: type III secretion inhibitors against Gram-negative bacteria. *J Intern Med* 2008; **264**: 17-29 [PMID: 18393958 DOI: 10.1111/j.1365-2796.2008.01941.x]
- Stavrinides J**, McCann HC, Guttman DS. Host-pathogen interplay and the evolution of bacterial effectors. *Cell Microbiol* 2008; **10**: 285-292 [PMID: 18034865]
- Filloux A**. The type VI secretion system: a tubular story. *EMBO J* 2009; **28**: 309-310 [PMID: 19225443 DOI: 10.1038/emboj.2008.301]
- Fuqua WC**, Winans SC, Greenberg EP. Quorum sensing in bacteria: the LuxR-LuxI family of cell density-responsive transcriptional regulators. *J Bacteriol* 1994; **176**: 269-275 [PMID: 8288518]
- Romero M**, Acuña L, Otero A. Patents on quorum quenching: interfering with bacterial communication as a strategy to fight infections. *Recent Pat Biotechnol* 2012; **6**: 2-12 [PMID: 22420877]
- Dong YH**, Wang LY, Zhang LH. Quorum-quenching microbial infections: mechanisms and implications. *Philos Trans R Soc Lond B Biol Sci* 2007; **362**: 1201-1211 [PMID: 17360274]
- Rasch M**, Buch C, Austin B, Slierendrecht WJ, Ekmann KS, Larsen JL, Johansen C, Riedel K, Eberl L, Givskov M, Gram L. An inhibitor of bacterial quorum sensing reduces mortalities caused by Vibriosis in rainbow trout (*Oncorhynchus mykiss*, Walbaum). *Syst Appl Microbiol* 2004; **27**: 350-359 [PMID: 15214641]
- Jerse AE**, Sharma ND, Simms AN, Crow ET, Snyder LA, Shafer WM. A gonococcal efflux pump system enhances bacterial survival in a female mouse model of genital tract infection. *Infect Immun* 2003; **71**: 5576-5582 [PMID: 14500476]
- Martínez JL**, Sánchez MB, Martínez-Solano L, Hernández A, Garmendia L, Fajardo A, Alvarez-Ortega C. Functional role of bacterial multidrug efflux pumps in microbial natural ecosystems. *FEMS Microbiol Rev* 2009; **33**: 430-449 [PMID: 19207745 DOI: 10.1111/j.1574-6976.2008.00157.x]
- Hirakata Y**, Srikumar R, Poole K, Gotoh N, Suematsu T, Kohno S, Kamihira S, Hancock RE, Speert DP. Multidrug efflux systems play an important role in the invasiveness of *Pseudomonas aeruginosa*. *J Exp Med* 2002; **196**: 109-118 [PMID: 12093875]
- Pearson JP**, Feldman M, Iglewski BH, Prince A. *Pseudomonas aeruginosa* cell-to-cell signaling is required for virulence in a model of acute pulmonary infection. *Infect Immun* 2000; **68**: 4331-4334 [PMID: 10858254]
- Martínez JL**, Baquero F. Interactions among strategies associated with bacterial infection: pathogenicity, epidemicity, and antibiotic resistance. *Clin Microbiol Rev* 2002; **15**: 647-679 [PMID: 12364374]
- Silver S**, Phung LT. Bacterial heavy metal resistance: new surprises. *Annu Rev Microbiol* 1996; **50**: 753-789 [PMID: 8905098]
- Ramos JL**, Duque E, Gallegos MT, Godoy P, Ramos-Gonzalez MI, Rojas A, Teran W, Segura A. Mechanisms of solvent tolerance in gram-negative bacteria. *Annu Rev Microbiol* 2002; **56**: 743-768 [PMID: 12142492]
- Kourtesi C**, Ball AR, Huang YY, Jachak SM, Vera DM, Khondkar P, Gibbons S, Hamblin MR, Tegos GP. Microbial efflux systems and inhibitors: approaches to drug discovery and the challenge of clinical implementation. *Open Microbiol J* 2013; **7**: 34-52 [PMID: 23569468 DOI: 10.2174/1874285801307010034]
- Bina XR**, Philippart JA, Bina JE. Effect of the efflux inhibitors 1-(1-naphthylmethyl)-piperazine and phenyl-arginine-beta-naphthylamide on antimicrobial susceptibility and virulence factor production in *Vibrio cholerae*. *J Antimicrob Chemother* 2009; **63**: 103-108 [PMID: 19010827 DOI: 10.1093/jac/dkn466]
- Varga ZG**, Armada A, Cerca P, Amaral L, Mior Ahmad Subki MA, Savka MA, Szegedi E, Kawase M, Motohashi N, Molnár J. Inhibition of quorum sensing and efflux pump system by trifluoromethyl ketone proton pump inhibitors. *In Vivo* 2012; **26**: 277-285 [PMID: 22351670]
- García-Contreras R**, Maeda T, Wood TK. Resistance to quorum-quenching compounds. *Appl Environ Microbiol* 2013; **79**: 6840-6846 [PMID: 24014536 DOI: 10.1128/AEM.02378-13]
- Shakhnovich EA**, Sturtevant D, Mekalanos JJ. Molecular mechanisms of virstatin resistance by non-O1/non-O139 strains of *Vibrio cholerae*. *Mol Microbiol* 2007; **66**: 1331-1341 [PMID: 17986190]
- Maeda T**, García-Contreras R, Pu M, Sheng L, Garcia LR, Tomás M, Wood TK. Quorum quenching quandary: resistance to antivirulence compounds. *ISME J* 2012; **6**: 493-501 [PMID: 21918575 DOI: 10.1038/ismej.2011.122]
- Tomás M**, Doumith M, Warner M, Turton JF, Beceiro A, Bou G, Livermore DM, Woodford N. Efflux pumps, OprD porin,

AmpC beta-lactamase, and multiresistance in *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. *Antimicrob*

Agents Chemother 2010; **54**: 2219-2224 [PMID: 20194693 DOI: 10.1128/AAC.00816-09]

P- Reviewer: Galgoczy L, Hays J, Zhang ZM **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Lu YJ



Harnessing pharmacological knowledge for personalized medicine and pharmacotyping: Challenges and lessons learned

Ioannis S Vizirianakis

Ioannis S Vizirianakis, Laboratory of Pharmacology, Department of Pharmaceutical Sciences, School of Health Sciences, Aristotle University of Thessaloniki, GR-54124 Thessaloniki, Greece

Author contributions: Vizirianakis IS contributed to this manuscript from its concept to writing and submission

Correspondence to: Ioannis S Vizirianakis, PhD, Laboratory of Pharmacology, Department of Pharmaceutical Sciences, School of Health Sciences, Aristotle University of Thessaloniki, GR-54124 Thessaloniki, Greece. ivizir@pharm.auth.gr

Telephone: +30-2310-997658 Fax: +30-2310-997645

Received: June 12, 2014 Revised: October 17, 2014

Accepted: October 28, 2014

Published online: December 9, 2014

Abstract

The contribution of the genetic make-up to an individual's capacity has long been recognized in modern pharmacology as a crucial factor leading to therapy inefficiency and toxicity, negatively impacting the economic burden of healthcare and restricting the monitoring of diseases. In practical terms, and in order for drug prescription to be improved toward meeting the personalized medicine concept in drug delivery, the maximum clinical outcome for most, if not all, patients must be achieved, *i.e.*, pharmacotyping. Such a direction although promising and of high expectation from the society, it is however hardly to be afforded for healthcare worldwide. To overcome any existed hurdles, this means that practical clinical utility of personalized medicine decisions have to be documented and validated in the clinical setting. The latter implies for drug delivery the efficient implementation of previously gained *in vivo* pharmacology experience with pharmacogenomics knowledge. As an approach to work faster and in a more productive way, the elaboration of advanced physiologically based pharmacokinetics models is discussed. And in better clarifying this topic, the example of tamoxifen is thoroughly presented. Overall, pharmacotyping represents a major

challenge in modern therapeutics for which pharmacologists need to work in successfully fulfilling this task.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Pharmacology; Pharmacogenomics; Personalized medicine; Pharmacokinetics; Pharmacodynamics; Pharmacotyping; Translational medicine; Drug delivery; Education; Curricula

Core tip: Drug prescription in order to be improved, the drug delivery process needs to confront the challenges of genomics knowledge translation to ensure the maximum clinical outcome for most, if not all, patients, *i.e.*, achieving pharmacotyping. The practical clinical utility of personalized medicine decisions needs to be documented and validated in the clinical setting. Physiologically based pharmacokinetic models represent an approach by which the faster and more efficient implementation of pharmacogenomics knowledge in evidence-based medicine could be achieved. Pharmacotyping represents a major challenge in modern therapeutics for which pharmacologists need to work both in academia and research in successfully fulfilling this task.

Vizirianakis IS. Harnessing pharmacological knowledge for personalized medicine and pharmacotyping: Challenges and lessons learned. *World J Pharmacol* 2014; 3(4): 110-119 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/110.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.110>

INTRODUCTION

Unanimously nowadays, nanotechnology and nanomedicine in parallel with pharmacology and pharmacogenomics (PGx) contribute knowledge and methodologies permitting individualized treatment decisions to enter

in everyday clinical practice. The personalized medicine concept along with the interdisciplinary efforts needed to achieve the desired practical clinical utility for personalized medicine decisions worldwide is extensively and thoroughly described elsewhere^[1]. The latter also implies that PGx by bridging pharmacology with genetics/genomics provides additional advantage for translational medicine to positively impact drug development and delivery outcomes. This means that the molecular etiology of drug response variability, by clinically assessing the genetic factors that contribute to pharmacokinetics (PK) and pharmacodynamics (PD), is now considered an integral part of modern pharmacology and therapeutics. As a consequence, the drug administration has been changed allowing for pharmacotyping (PTx) to emerge in the prescription process *e.g.*, the individual patient (personalized) specific medicine selection and administration scheme, as proposed earlier^[1-4]. From a historical point of view, pharmacogenetics as a term has been introduced by Friedrich Vogel (1959), whereas the first example of pharmacogenetics described ever is flavism disorder by Pythagoras (580-500 BC; ancient Greek mathematician). By following-up chronologically until nowadays such pharmacogenetics-related scientific breakthroughs for pharmacology, it is obvious that these focused efforts have been successful by efficiently translating multidisciplinary-based experimental data that enabled pharmacological improvements both in research and the clinical setting (for such a detailed chronological description of pharmacogenetics/PGx breakthroughs see elsewhere^[1]). However from the experience gained thus far, it needs considerable effort and, more importantly, to invent focused as well as interdisciplinary-oriented “smart and sophisticated” experimental approaches to move all the way through establishing personalized medicine decisions of broader practical clinical utility. The molecular etiology of illness pathophysiology and the elucidation of genetic factors contributing to pharmacological profiles of drugs in the body are hardly experimentally approachable, especially in being thoroughly understood for all marketed therapeutics. Moreover, the interplay of genes with therapeutics implies that their interaction is also modulating drug delivery outcomes, since the mutational status of genes (gene polymorphism) and drug-regulated gene expression profiles contribute to drug response variability (Figure 1). The latter, it leads to patient phenotypic (pharmacological) response modulation, or alternatively into pharmacological response heterogeneity. Complementary, in trying to minimize the emergence of drug response variation amongst population and in order to achieve improved profiles of administered drugs worldwide, a new interdisciplinary infrastructure needs to be created and integrated in clinical practice^[4]. The latter, will also help in adjusting the regulatory environment to improve drug development productivity by minimizing the emergence of adverse drug reactions (ADRs), avoiding drug interactions and thus finally improving the clinical outcome.

Moreover, by considering the issue of education in

pharmacy and medicine, the better training in pharmacology will be achieved through the development of new curricula aiming to advance skills of medical and pharmacy students in implementing *in vivo* pharmacology experience with PGx information. But how this task could be attainable and productive? Already, academics in relevant disciplines confront with obstacles in trying to integrate knowledge from PGx and personalized medicine concepts into teaching curricula and enrich the skills of students toward better handling modern therapeutics issues of practical clinical utility. The previously well-established background bridge between pharmacology and other disciplines (*e.g.*, physiology/pathophysiology, chemistry) created in order future practitioners to understand drug behavior and actions in the body is being expanded by incorporating translational information extracted (*e.g.*, biochemical, biological, molecular) including that from bioinformatics and also material sciences and nanotechnology. Unanimously, the molecular approaches applied to predict and/or assess the behavior of therapeutics in the living organisms enrich the knowledge of students and also strengthen their capacity in drug prescription for better dosage-scheme selection of administered medicines in the clinical setting. And for sure, the better education by covering the concept of PGx as well as personalized medicine will be the maximum positive impact for both academics involved and healthcare practitioners would happen; the latter, however, further necessitates the proper adjustments in academia to restructure and organize relevant innovative medical and pharmacy curricula worldwide^[4-6].

DEVELOPMENT OF ADVANCED PK/PD MODELS TO IMPLEMENT MOLECULAR PHARMACOLOGY FOR ENRICHING TRANSLATIONAL MEDICINE CAPACITY IN DRUG DELIVERY

Nowadays, it has been evidenced that mechanism-based PK/PD modeling has been a necessity in modern pharmacology toward speeding up early achievements in drug discovery and ensuring improved efficacy and safety profiles of candidate molecules before their final clinical development. The latter, implies that improved prediction capabilities of crucial drug-related parameters can be documented by extrapolating *in vitro* experimentation data into *in vivo* clinical variables across species^[7,8]. Importantly, however, PK/PD modeling in order to contribute greatest benefits at the preclinical and the clinical era, it also needs to be embraced across regulatory bodies and pharmaceutical industrial sector, as well as the educational process in academia^[1,9]. The recent advancement of genomic medicine and systems pharmacology, however, forms the baselines for multidisciplinary translational approaches by crossing the borders between molecular pharmacology with pathophysiology,

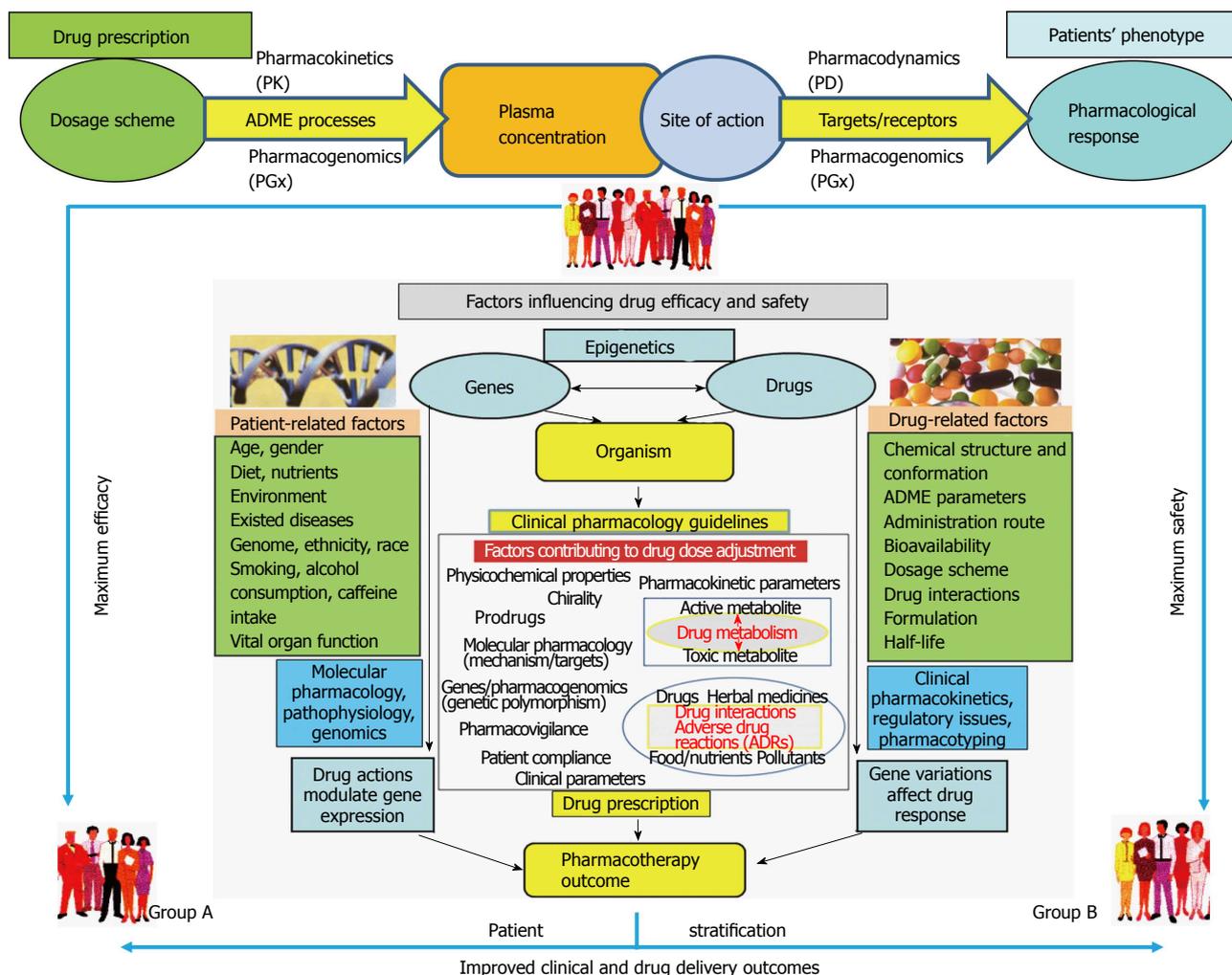


Figure 1 A roadmap of pharmacological response stages to efficiently address the pharmacotyping concepts in drug prescription. The processes and the factors related to pharmacological effects along with the scientific environment contributing to drug delivery outcomes in terms of efficacy, safety are depicted above. The need for enriching pharmacological knowledge to advance personalized medicine decisions in the clinical setting through drug dosage scheme adjustment (*i.e.*, PTx) is exemplified. The *in vivo* pharmacology experience gained thus far and it already appears in the drug regulatory environment is stressfully demanded to be empowered by pharmacogenomics knowledge in terms of PD/PK drug parameters assessment methodologies, the clinical pharmacology guidelines development and the prescription process. Complementary to this, the development of information-based workflow platforms in clinical practice incorporating algorithms to assess the efficient translation of clinical, biological, genomic and chemical information is also eagerly expected. Such a direction of major pharmacological importance permits the maximum efficacy and safety outcomes to be reached in a timely and worldwide basis for everyday healthcare (see text for more details). PTx: Pharmacotyping; PGx: Pharmacogenomics; PK: Pharmacokinetics; PD: Pharmacodynamics.

clinical sciences and genomics. In particular, systems pharmacology aims to understand the effects of drugs including ADRs in terms of pharmacological targets and within the molecular networks context that evidently has been allowing the integration of the systems biology-level in understanding the behavior of drugs in the body^[10]. This direction could be proven helpful especially for complex and multifactorial illnesses where the more thorough elucidation of their molecular pathophysiology is needed; complementary, such task in turn it represents a crucial prerequisite parameter upon attempting to improve pharmacotherapy outcomes in these diseases. By enabling network analyses of interactions mediated both pathophysiological and PD/PK drug responses through the different organization levels, (from the molecular level through organ and tissue into finally the entire organism), in an integrated approach, the faster and more

cost-affordable manner to empower the practical clinical utility of personalized medicine decisions will be clearly achieved^[1,11-14].

It is evident, that the improved translational medicine capacity means the successful adjustment of clinical pharmacology guidelines toward personalized medicine concepts. Complementary to this point, the issue on how the already gained *in vivo* pharmacology experience can be adjusted by being enriched with relevant systems pharmacology approaches and methodologies clearly emerges in a way that the implementation in real time with the PGx knowledge could happen^[15-17]. Alternatively, what it has been already established through the previously gained experience of *in vivo* pharmacology approaches, is the fact that drug pharmacological responses are evidenced by two dynamic processes being interrelated, that of PD and PK. PD describes what medicines do to the body (*i.e.*, drug-

receptor interactions), whereas PK is associated with what an organism does to therapeutics (*i.e.*, absorption, distribution, metabolism, excretion; ADME processes). As a consequence, a question then rises; on how in the new drug delivery era, maximum benefits could be ensured for all patients in terms of drug efficacy and safety? The latter task can be fulfilled only if the molecular mechanisms underlying PD/PK drug effects could decipher issues addressing either the emergence of idiosyncratic (genetic/genomic) toxic effects and ADRs in a given individual, or the involvement of environmental and epigenetic factors^[1,18,19] (Figure 1). The application of predictive bioinformatic approaches and computational methodologies in evaluating PK and PD profiles of drugs represents an established approach, especially the last few decades, throughout the drug development as well as delivery processes^[20]. *In silico* methods use and application of technologies to enhance the predictive capacity to ultimately improve productivity and drug delivery safety and efficacy profiles is now considered a major advancement^[20-23]. In parallel, specific information-based workflow computerized healthcare systems are being developed to contribute in the exploitation of knowledge coming from interdisciplinary resources with an affordable for the end-user manner^[24]. The latter, also implies the proper application of the translated knowledge into information standing types being capable to be simultaneously used in everyday healthcare approaches upon illness prognosis, diagnosis and administration of therapeutics (Figure 1).

Undoubtedly, the development of suitable translational medicine-enriched clinical pharmacology guidelines for providing instructions upon drug prescription (*e.g.*, dosage scheme adjustment, disease prognosis/diagnosis profile improvement) will efficiently facilitate the successful implementation of PGx concepts into everyday clinical practice. The latter, also refers to the information systems applied in routine patient care. Moreover importantly as it has been proposed recently, by working within this direction the maximum benefits from both nanomedicine and personalized medicine efforts is expected to happen empowering clinical outcome through the advent of personalized nanomedicine concepts at both the research and the clinical setting^[25]. Complementary to this, personalized medicine is paving the way toward broader practical clinical utility of translational advancements, thus contributing toward PTx in drug prescription as well as medicine and pharmacy in general. The latter, refers to the development of clinically-applied algorithms in drug prescription regarding the genetic variables being able to affect PK and PD behavior of marketed therapeutics. The use of information-based systems into everyday healthcare has clearly shown the need of developing such unified information systems with adherence to various healthcare environments worldwide (Figure 1). To this end, the more successful development of quantitative PGx models for translation medicine is being achieved then the best benefits in PTx-based drug delivery from genetically-guided drug dose adjustment is expected^[1,26,27].

In fulfilling this task of practical clinical utility, the close collaboration of clinicians with pharmacologists will pace PTx in drug prescription in a faster and more efficient manner.

HARNESSING PHARMACOLOGICAL AND TOXICOLOGICAL GENOMICS KNOWLEDGE FOR ADVANCING THE SKILLS OF FUTURE HEALTHCARE PROFESSIONALS TO IMPLEMENT PTX CONCEPTS IN DRUG PRESCRIPTION

The use of PD/PK tools implemented with biostatistics approaches in courses related to pharmacology, model-based drug development as well as predictive modeling and simulation upon pharmacological assessment empower the teaching process and ensure greatest benefits for researchers, educators, as well as students. Alternatively, in order to achieve this task for strengthening students' knowledge and skills in clinical and molecular pharmacology, PGx expertise as well as personalized medicine decision-making means of being capable to simultaneously: (1) assess and predict clinically relevant drug interactions, thus minimizing ADRs emergence risk; and (2) advance the profiles of drugs in terms of efficacy and safety for individual patients by inter-correlating clinical data, drug properties and genetic/genomic characteristics^[1-6]. Moving forward in this manner for education, future healthcare practitioners will be instructed on how to more efficiently and in real time apply personalized medicine approaches in clinical practice, a fact that impose health and societal benefits in general. In addition, the successful implementation of systems pharmacology and pathophysiology approaches with *in vivo* pharmacology experience better ensures both productivity and clinical outcome for innovative molecularly-targeted therapeutics, "smart/genius" drug delivery systems, translational medicine efforts, as well as nanomedicine applications (Figure 1).

Although PGx advancements contribute clinically relevant genomics knowledge, it is also obvious however, that modern pharmacology is gaining major benefits in experimenting with new sophisticated technological methodologies in drug development and delivery era. By projecting such changes that are expected to happen for therapeutics in the near future, it is important mainly for pharmacologists in academia to be actively engaged in providing their students with strong background and skills of *in vivo* pharmacology enriched with PGx knowledge. Since the latter represents a dynamic knowledge module and an ever changing scientific environment, it means that personalized medicine concepts must be taught to allow therapeutic decisions in real time and for all pharmacological drug classes. In such case, young healthcare practitioners will be trained in getting capable for individualized prescription of drug dosage schemes,

thus minimizing the risk for toxicity, the emergence of interactions and ADRs in clinical practice. It is thus crucial for pharmacologists to prioritize the steps and the process needed to be considered in pharmacology curricula, as well as to set-up a pharmacology-focused roadmap for achieving broader utility of personalized medicine decisions.

Nowadays, laboratory medicine techniques have received major impact from genomics methodologies and experimentation. The availability of methodologies and tools allowing the simultaneous assessment of various source data of clinical relevance (*e.g.*, drug-related, genomics-focused, clinical measurements) clearly contribute toward individualized therapeutic decisions in routine healthcare. Moving forward and in order to improve pharmacology-related productivity and clinical outcome issues, this means that the creation of platforms where the pharmacological assessment and the clinical exploitation of PK/PD-related molecular targets is happening throughout the drug discovery and development process; for example, the improvement of PK/PD behavior of the designed molecularly-targeted therapeutics in the body will be better served and secured^[28]. Moreover, the beneficial implementation of functional mapping framework in pharmacology by smoothly addressing PGx data integration into PK/PD processes will be greatly benefited, as proposed^[29]. In that case, the already applied PK/PD-related mathematical models could be effectively coupled with PGx data referring to specific pharmacology-focused molecular networks and signaling pathways. The interdisciplinary nature of the framework and infrastructure needed represents, however, tedious and long-standing processes that obviously also rely on elucidation of illness etiology and drug behavior profiles. Besides, the establishment of selective molecular biomarkers with broader clinical validity and utility for most, if not all, pathological disorders and marketed therapeutics have to be clearly addressed. Moreover upon formulating personalized medicine decisions, the developed genomics-related methodology profiling (*e.g.*, genome-wide linkage analysis including proteomics-, genotyping-, gene array-, transcriptomics- and/or metabolomics-related data) must exhibit broader advantages in laboratory medicine applications for all patients worldwide^[28-33]. The latter, means that the capability of molecular diagnostics to help addressing routine therapeutic decisions in real time is the most desirable goal nowadays for PTx and personalized medicine concepts.

ESTABLISHMENT OF NETWORK AND SYSTEMS PHARMACOLOGY METHODOLOGIES IN ENRICHING *IN VIVO* PHARMACOLOGY EXPERIENCE TOWARD FORMULATING PTX-BASED CLINICAL PHARMACOLOGY GUIDELINES

Unanimously, the availability of predictive tools to effec-

tively address issues related to safety and efficacy profiles of therapeutics within the body represent a major task toward improving productivity and clinical outcomes of drug candidate molecules. Especially by considering the whole drug discovery and development process, that need is even more stressful at preclinical-clinical phase of development; such capacity in predicting safety and efficacy therapeutic outcomes very early is considered crucial toward establishing focused personalized medicine decisions of broader clinical utility. To do so, the implementation of clinical pharmacology guidelines has to be achieved through the knowledge coming from the use of cost-affordable PGx molecular diagnostics^[3,34]. For example, the organization and development of evidence-based PGx guidelines in clinical practice represents an obstacle hindering translational medicine efforts in drug discovery and development from bench to bedside^[35-37]. To this end and although various issues (*e.g.*, reimbursement, social, cost-benefit and ethical) should be simultaneously addressed, specific efforts for formulating PGx guidelines for dose recommendation schemes in specific pharmacological drug classes has been initiated and proposed^[38-43].

The development and application of physiologically based pharmacokinetic (PBPK) modeling for key PK-related processes have been greatly appreciated in drug delivery and development era. The PBPK models capacity also permits to assess, predict and evaluate in a quantitative basis the potential clinical effect of drug interactions along with any impact related to disease status, genetic make-up, environmental factors and/or drug formulation properties^[44,45]. Such an effort is presented in more detail elsewhere^[34]. As far as the PGx data exploitation is concerned, it is crucial for example to understand that only in the circumstances where the genetic variation represents the rate-limiting PK/PD step it would be possible to directly inter-correlate such molecular knowledge with the predictability profile in drug plasma concentration; and then beneficial for the practical utility of personalized medicine to proceed toward adjusting dosage scheme for individual patients based on their genetic variation (Figure 1). Such a direction upon drug prescription, in order not to be restricted in clinical pharmacology guidelines have to also effectively integrate and address issues related to the PGx concepts, the drug interactions knowledge, as well as the emergence of ADRs^[46]. For having PTx success, this relies on the ability to use drug interactions knowledge being efficiently inter-correlated with PGx knowledge for genes mediated the PK/PD behavior of therapeutics. Moving ahead, such direction necessitates a structure where suitably constructed PGx models assembling large cohort studies will be established to better serve: (1) the interdisciplinary data assessment; (2) the dissemination and broader clinical utility of personal genetic information upon illness risk prevention; and also (3) the use of PTx-based concepts upon medicines prescription^[1,34]. Moreover, the importance of having in these scientific attempts expert pharmacologists to participate is crucial,

since pharmacologists would be capable to verify: (1) the enrichment of *in vivo* pharmacology experience; (2) the efficient translation of PGx information to implement therapeutics decisions; and last, but not least, (3) the adjustment of drug dosage schemes, thus making personalized medicine decisions to benefit routine clinical practice, or alternatively, to achieve PTx for individual patient populations, if not all patients.

PBPK MODELING APPROACHES TO IMPLEMENT *IN VIVO* PHARMACOLOGY EXPERIENCE WITH PGX KNOWLEDGE FOR ENSURING PTX IN CANCER THERAPY: THE CASE OF TAMOXIFEN

PGx of anticancer drugs is now considered an integral part of cancer therapy^[47]. Indeed, a number of predictive PGx biomarkers to assess the safety and efficacy clinical profiles of individual marketed anticancer drugs has been validated by drug regulatory agencies (*e.g.*, the FDA and EMA) and are shown in Table 1. As mentioned above, however, the development of PBPK models implemented with systems pharmacology approaches, (assessing predictive PGx biomarkers), represents a platform where in real time the assessment of both patient-related and drug-related factors can be intercorrelated to achieve maximum efficacy and safety outcome for individual populations, *i.e.*, PTx (Figure 1). Alternatively, the latter means the elaboration of a multidisciplinary environment in order both the assessment of drug interactions and PGx data to be effectively incorporated to guide drug prescription. To better clarify this issue by analyzing the complexity existed and the hurdles needed to be overcome, the example of tamoxifen and serotonin reuptake inhibitors (SSRIs) will be further considered.

Accumulated evidence over the previous years have clearly postulated the contribution of genetic polymorphic variants of CYP2C19 and CYP2D6 (drug metabolizing enzymes) to the pharmacological response of psychotropic drugs in clinical practice^[48-50]. To this end, a specific guideline for psychiatrists providing practical recommendations upon the prescription of psychotropic drugs based both on clinical drug-related as well as CYP2D6 and CYP2C19 PGx data for individual patient populations has been proposed. Although, the broader clinical applicability of such instructions is still elusive, however, the improvement of PK and PD profile toward achieving personalized medicine decisions for psychotropic drug coincides with the capacity to simultaneously assess *CYP* genes variants in routine healthcare^[51-54].

The fact that metabolism of psychotropic drugs including antidepressants represents a rate-limiting step in their pharmacological profile means that specific CYP polymorphic variant forms (*e.g.*, CYP2D6) contribute either to toxicity and/or drug inefficiency in specific individual populations. Moreover, since some antidepressants

are also CYP2D6 inhibitors, clinically-relevant drug interactions are expected upon their co-administration with other drugs whose pharmacological activity is based on CYP2D6 function like tamoxifen^[55]. The clinical efficacy of tamoxifen varies widely among breast cancer women depending on their CYP2D6 genotype. Tamoxifen is a pro-drug which means that its active metabolite 4-hydroxytamoxifen and endoxifen being produced through the function of CYP2D6 mediates the pharmacological anti-estrogen action in the body. At the same time, co-administration of SSRIs antidepressants had previously been a common routine clinical practice and prescribed to treat hot flashes in women who take tamoxifen^[56-61]. But at what extent, however, would be clinically validated the predictive capacity in dosage scheme to ensure tamoxifen efficacy (active metabolites plasma concentration) and safety (toxicity, *i.e.*, hot flashes) based on CYP2D6 function affected by genotypes and SSRIs inhibitory behavior? Moving ahead, it has been proven that either women exhibiting polymorphic null-activity for CYP2D6 (PMs; CYP2D6 poor-metabolizers), or patients under tamoxifen chemotherapy co-prescribed with potent CYP2D6 inhibitors (*e.g.*, antidepressant drugs fluoxetine and paroxetine) show a greater risk of breast cancer recurrence and mortality due to decreased levels of active tamoxifen metabolites formed in their organism. That knowledge now proposes that personalized dosage schemes of tamoxifen administration to individual or population of patients are achieved through: (1) for CYP2D6 PM women by avoiding the co-prescription of tamoxifen with SSRIs or other medicines acting as potent CYP2D6 inhibitors; (2) Alternatively, similar improvement can be achieved through the proper dose adjustment of tamoxifen, or alternatively by switching into another hormonal therapy drug class (*e.g.*, aromatase inhibitors); and (3) For breast cancer patients exhibiting normal CYP2D6 metabolism (phenotype of CYP2D6 extensive metabolizers) by selecting the co-administration of an antidepressant that exhibits no inhibitor activity for CYP2D6 (*e.g.*, venlafaxine which represents a weak CYP2D6 inhibitor). But even in this case, the clinical effectiveness and the cost-effectiveness of CYP2D6 genotyping for the management of women with breast cancer treated with tamoxifen still needs to be validated^[62]. However, having this knowledge in mind one can further consider the possibility of developing advanced PBPK models in order to: (1) more thoroughly exploit clinical, pharmacological and PGx data of drugs; (2) develop proper algorithms to implement drug prescription; and (3) to facilitate new drug development productivity through predicting PD/PK behavior and reduce attrition rates in potential drug candidate molecules. Recently, the successful PBPK model development for tamoxifen delivery and for the evaluation of PK of patients with cancer clearly shows the dynamics of such scientific approaches^[63,64]. And more importantly this dynamics of PBPK modeling is further strengthened from research efforts reshaping the field of PD/PK modeling by enhancing the capacity to efficiently predict drug

Table 1 Genes used as predictive pharmacogenomics biomarkers to assess the safety and efficacy clinical profiles of individual marketed anticancer drugs^{1,2}

Drug	Gene	Safety/efficacy profile
Anastrozole	<i>ER</i>	Lower efficacy; no response in cancer patients with tumor <i>ER</i> -negative expression
Capecitabine	<i>DPYD</i>	Lower safety; ADRs; Orodigestive neutropenia
Cetuximab	<i>EGFR</i>	Lower efficacy; no response in cancer patients with tumor <i>EGFR</i> -negative expression
	<i>K-RAS</i>	Lower efficacy; no response in cancer patients with tumor specific <i>K-RAS</i> mutations
Cisplatin	<i>TPMT</i>	Lower safety; ADRs; cytotoxicity associated with hearing loss in children
Crizotinib	<i>ALK</i>	Efficacy; indication only in patients bearing <i>ALK</i> gene rearrangement positive tumors (<i>EML4-ALK</i> translocation)
Dabrafenib	<i>BRAF</i>	Efficacy; indicated only in melanoma patients with <i>BRAF</i> V600E mutation
	<i>G6PD</i>	Safety; ADRs; toxicity in <i>G6PD</i> deficient patients
Dasatinib	<i>Ph+</i>	Efficacy; indicated only for <i>Ph</i> ⁺ tumors
Erlotinib	<i>EGFR</i>	Lower efficacy; no response in cancer patients with tumor <i>EGFR</i> -negative expression
Everolimus	<i>Her2/Neu</i>	Efficacy; indicated in <i>HER2</i> protein overexpression negative in breast cancer women
	<i>ER</i>	Efficacy; indicated for breast cancer women bearing <i>ER</i> positive tumors (<i>ESR1</i> ⁺)
Exemestane	<i>ER</i>	Lower efficacy; no response in cancer patients with tumor <i>ER</i> -negative expression
Imatinib	<i>Ph+</i>	Efficacy; indicated only for <i>Ph</i> ⁺ tumors
	<i>PDGFR</i>	Efficacy; indicated in myelodysplastic- myeloproliferative syndromes with <i>PGFR</i> gene rearrangements
	<i>FIP1L1-PDGFR</i>	Efficacy; assessment of <i>FIP1L1-PDGFR</i> translocation -fusion kinase in tumors
	<i>c-kit</i>	Lower efficacy; no response in cancer patients with absence of tumor activating <i>c-Kit</i> mutations
Irinotecan	<i>UGT1A1</i>	Lower safety; ADRs; diarrhea, increased risk for severe neutropenia in high doses of irinotecan
Lapatinib	<i>Her2/Neu</i>	Efficacy; indicated for over-expressing <i>Her2/Neu</i> advanced or metastatic breast cancer
Letrozole	<i>ER</i>	Lower efficacy; no response in cancer patients with tumor <i>ER</i> -negative expression
Nilotinib	<i>Ph+</i>	Efficacy; indicated only for <i>Ph</i> ⁺ tumors
	<i>UGT1A1</i>	Safety; increased risk of hyperbilirubinemia in patients with <i>UGT1A1</i> *28 genotype
6-Mercaptopurine	<i>TPMT</i>	Lower safety; ADRs; neutropenia
Panitumumab	<i>EGFR</i>	Lower efficacy; no response in cancer patients with tumor <i>EGFR</i> -negative expression
	<i>K-RAS</i>	Lower efficacy; no response in cancer patients with tumor specific <i>K-RAS</i> mutations
Pertuzumab	<i>HER2/Neu</i>	Efficacy; indicated only for <i>HER2/Neu</i> ⁺ breast cancer
Tamoxifen	<i>ER</i>	Lower efficacy; no response in cancer patients with tumor <i>ER</i> -negative expression
	<i>CYP2D6</i>	Lower efficacy; loss of therapeutic benefit for PMs and/or upon co-administration with <i>CYP2D6</i> inhibitors; lower plasma levels of active metabolite endoxifen achieved
	<i>FV</i>	Safety; ADRs; risk for venous thromboembolism in breast cancer women also bearing factor V Leiden (FLV) mutations
	<i>F2</i>	Safety; ADRs; risk for venous thromboembolism in breast cancer women also bearing factor II (prothrombin) mutations
Thioguanine	<i>TPMT</i>	Lower safety; ADRs; Neutropenia
Trastuzumab	<i>HER2/Neu</i>	Lower efficacy; no response in cancer patients with tumor <i>HER2/Neu</i> -negative expression
Vemurafenib	<i>BRAF</i>	Efficacy; indicated only in melanoma patients whose tumors has a mutation at amino acid 600 of the B-raf protein (V600E and/or V600K <i>BRAF</i> mutations)

¹See also the table of PGx biomarkers in drug labeling at the FDA that can be accessed at: <http://www.fda.gov/drugs/scienceresearch/researchareas/pharmacogenetics/ucm083378.htm> (Accessed on June 27, 2014); ²Additional data can be seen in "The Pharmacogenomics Knowledge Base (PharmGKB) at: <https://www.pharmgkb.org/> (Accessed on June 29, 2013). ADRs: Adverse drug reactions; G6PD: Glucose-6-phosphate dehydrogenase deficiency; PM: Poor metabolizers; ER: Estrogen receptor; TMTP: Thiopurine methyltransferase gene; DPYD: Dihydropyrimidine dehydrogenase gene; Ph: Philadelphia chromosome; FV: Factor V; F2: Factor II (prothrombin).

effects in the body^[65-68]. This is an example for PTx on how the pharmacological knowledge of drug interactions covering both clinical and biochemical knowledge can be efficiently inter-correlated with PGx information of genes mediating the PK/PD behavior of therapeutics to improve delivered medicines clinical outcomes. The need for well-educated physicians and pharmacists, the proper clinical pharmacology/PGx guidelines development and adjustment, as well as healthcare infrastructure organization equipped with suitable clinically-validated technological methodologies is now, more than ever, stressful and demanding.

The analysis presented above for tamoxifen and SSRIs imply that the broader clinical utility of personalized medicine as well as PTx will be also strengthened by developing pharmacology-focused functional mapping

frameworks for most, if not all, specific pharmacological drug classes. In such a case, additional benefits for translational medicine will be gained; alternatively, this refers to the successful implementation of PD/PK data and the new drug development environment with PGx knowledge^[34]. Such modeling approaches clearly strengthen healthcare efforts toward the establishment of PTx as a new drug prescription "philosophy" in drug delivery. The latter, means that the practical utility of genomics information is conceptually exploited to ensure maximum safety and efficacy profiles. This way toward achieving PTx represents a very complex task that is clearly documented in the case of tamoxifen prescription where the *CYP2D6* pharmacogenomics assessment by healthcare decision makers well documented the steps still needed to be addressed^[69,70]. In the meantime, however, the

proper education of healthcare professionals has to be adjusted to fulfill expectations for the PTx roadmap in personalized medicine. Importantly, to highlight education needs and also to facilitate the teaching process in the revised curricula of various professionals engaged in this topic, a recently edited book volume has been organized and released as a first attempt to fill the gap in terms of the multidisciplinary perspective for personalized medicine^[1].

REFERENCES

- Vizirianakis IS, editor. Personalized medicine: Advances in nanotechnology, drug delivery and therapy. Singapore: Pan Stanford Publishing, 2014 [DOI: 10.1201/b15465]
- Vizirianakis IS. Challenges in current drug delivery from the potential application of pharmacogenomics and personalized medicine in clinical practice. *Curr Drug Deliv* 2004; **1**: 73-80 [PMID: 16305372]
- Vizirianakis IS. Clinical translation of genotyping and haplotyping data: implementation of in vivo pharmacology experience leading drug prescription to pharmacotyping. *Clin Pharmacokinet* 2007; **46**: 807-824 [PMID: 17854232]
- Vizirianakis IS. Nanomedicine and personalized medicine toward the application of pharmacotyping in clinical practice to improve drug-delivery outcomes. *Nanomedicine* 2011; **7**: 11-17 [PMID: 21094279 DOI: 10.1016/j.nano.2010.11.002]
- Vizirianakis IS. Pharmaceutical education in the wake of genomic technologies for drug development and personalized medicine. *Eur J Pharm Sci* 2002; **15**: 243-250 [PMID: 11923056]
- Vizirianakis IS. From defining bioinformatics and pharmacogenomics to developing information-based medicine and pharmacotyping in healthcare. In: Gad SC, editor. Handbook of Pharmaceutical Biotechnology. New York: John Wiley & Sons, Inc., 2007: 201-228
- Mager DE, Jusko WJ. Development of translational pharmacokinetic-pharmacodynamic models. *Clin Pharmacol Ther* 2008; **83**: 909-912 [PMID: 18388873 DOI: 10.1038/clpt.2008.52]
- Ploeger BA, van der Graaf PH, Danhof M. Incorporating receptor theory in mechanism-based pharmacokinetic-pharmacodynamic (PK-PD) modeling. *Drug Metab Pharmacokinet* 2009; **24**: 3-15 [PMID: 19252332]
- Rajman I. PK/PD modelling and simulations: utility in drug development. *Drug Discov Today* 2008; **13**: 341-346 [PMID: 18405847 DOI: 10.1016/j.drudis.2008.01.003]
- Wist AD, Berger SI, Iyengar R. Systems pharmacology and genome medicine: a future perspective. *Genome Med* 2009; **1**: 11 [PMID: 19348698 DOI: 10.1186/gm11]
- Zhao S, Iyengar R. Systems pharmacology: network analysis to identify multiscale mechanisms of drug action. *Annu Rev Pharmacol Toxicol* 2012; **52**: 505-521 [PMID: 22235860 DOI: 10.1146/annurev-pharmtox-010611-134520]
- Berger SI, Iyengar R. Network analyses in systems pharmacology. *Bioinformatics* 2009; **25**: 2466-2472 [PMID: 19648136 DOI: 10.1093/bioinformatics/btp465]
- Carpenter AE, Sabatini DM. Systematic genome-wide screens of gene function. *Nat Rev Genet* 2004; **5**: 11-22 [PMID: 14708012]
- Hardiman G. Microarray platforms--comparisons and contrasts. *Pharmacogenomics* 2004; **5**: 487-502 [PMID: 15212585]
- Ekins S. Predicting undesirable drug interactions with promiscuous proteins in silico. *Drug Discov Today* 2004; **9**: 276-285 [PMID: 15003246]
- Gardner SP. Ontologies and semantic data integration. *Drug Discov Today* 2005; **10**: 1001-1007 [PMID: 16023059]
- Kurland L, Lind L, Melhus H. Using genotyping to predict responses to anti-hypertensive treatment. *Trends Pharmacol Sci* 2005; **26**: 443-447 [PMID: 16055200]
- Weng L, Zhang L, Peng Y, Huang RS. Pharmacogenetics and pharmacogenomics: a bridge to individualized cancer therapy. *Pharmacogenomics* 2013; **14**: 315-324 [PMID: 23394393 DOI: 10.2217/pgs.12.213]
- O'Kane DJ, Weinshilboum RM, Moyer TP. Pharmacogenomics and reducing the frequency of adverse drug events. *Pharmacogenomics* 2003; **4**: 1-4 [PMID: 12517278]
- In Vivo Pharmacology Training Group. The fall and rise of in vivo pharmacology. *Trends Pharmacol Sci* 2002; **23**: 13-18 [PMID: 11804646]
- Walker MJ, Soh ML. Challenges facing pharmacology--the in vivo situation. *Trends Pharmacol Sci* 2006; **27**: 125-126 [PMID: 16480779]
- British Pharmacological Society and the Physiological Society. Tackling the need to teach integrative pharmacology and physiology: problems and ways forward. *Trends Pharmacol Sci* 2006; **27**: 130-133 [PMID: 16500714]
- Farahani P, Levine M. Pharmacovigilance in a genomic era. *Pharmacogenomics J* 2006; **6**: 158-161 [PMID: 16415916]
- Pirmohamed M. Pharmacogenetics: past, present and future. *Drug Discov Today* 2011; **16**: 852-861 [PMID: 21884816 DOI: 10.1016/j.drudis.2011.08.006]
- Vizirianakis IS, Fatouros DG. Personalized nanomedicine: paving the way to the practical clinical utility of genomics and nanotechnology advancements. *Adv Drug Deliv Rev* 2012; **64**: 1359-1362 [PMID: 22983333 DOI: 10.1016/j.addr.2012.09.034]
- Guo Y, Shafer S, Weller P, Usuka J, Peltz G. Pharmacogenomics and drug development. *Pharmacogenomics* 2005; **6**: 857-864 [PMID: 16296948]
- Cascorbi I, Tyndale R. Progress in pharmacogenomics: bridging the gap from research to practice. *Clin Pharmacol Ther* 2014; **95**: 231-235 [PMID: 24548984 DOI: 10.1038/clpt.2013.235]
- Vizirianakis IS, Chatzopoulou M, Bonovolias ID, Nicolaou I, Demopoulos VJ, Tsiptsoglou AS. Toward the development of innovative bifunctional agents to induce differentiation and to promote apoptosis in leukemia: clinical candidates and perspectives. *J Med Chem* 2010; **53**: 6779-6810 [PMID: 20925433 DOI: 10.1021/jm100189a]
- Ahn K, Luo J, Berg A, Keefe D, Wu R. Functional mapping of drug response with pharmacodynamic-pharmacokinetic principles. *Trends Pharmacol Sci* 2010; **31**: 306-311 [PMID: 20488563 DOI: 10.1016/j.tips.2010.04.004]
- Plenge RM, Scolnick EM, Altshuler D. Validating therapeutic targets through human genetics. *Nat Rev Drug Discov* 2013; **12**: 581-594 [PMID: 23868113 DOI: 10.1038/nrd4051]
- La Thangue NB, Kerr DJ. Predictive biomarkers: a paradigm shift towards personalized cancer medicine. *Nat Rev Clin Oncol* 2011; **8**: 587-596 [PMID: 21862978 DOI: 10.1038/nrclinonc.2011.121]
- Dequeker E, Ramsden S, Grody WW, Stenzel TT, Barton DE. Quality control in molecular genetic testing. *Nat Rev Genet* 2001; **2**: 717-723 [PMID: 11533720]
- Nicolaidis NC, O'Shannessy DJ, Albone E, Grasso L. Co-development of diagnostic vectors to support targeted therapies and theranostics: essential tools in personalized cancer therapy. *Front Oncol* 2014; **4**: 141 [PMID: 24982846 DOI: 10.3389/fonc.2014.00141]
- Vizirianakis IS. Advancement of pharmacogenomics toward pharmacotyping in drug prescription: Concepts, challenges, and perspectives for personalized medicine. In: Vizirianakis IS. (Ed). Personalized medicine: Advances in nanotechnology, drug delivery and therapy. Singapore: Pan Stanford Publishing, 2014: 893-952 [DOI: 10.1201/b15465-22]
- Relling MV, Klein TE. CPIC: Clinical Pharmacogenetics Implementation Consortium of the Pharmacogenomics Research Network. *Clin Pharmacol Ther* 2011; **89**: 464-467 [PMID: 21270786 DOI: 10.1038/clpt.2010.279]
- Swen JJ, Nijenhuis M, de Boer A, Grandia L, Maitland-van der Zee AH, Mulder H, Rongen GA, van Schaik RH, Schale-

- kamp T, Touw DJ, van der Weide J, Wilffert B, Deneer VH, Guchelaar HJ. Pharmacogenetics: from bench to byte--an update of guidelines. *Clin Pharmacol Ther* 2011; **89**: 662-673 [PMID: 21412232 DOI: 10.1038/clpt.2011.34]
- 37 **Amstutz U**, Carleton BC. Pharmacogenetic testing: time for clinical practice guidelines. *Clin Pharmacol Ther* 2011; **89**: 924-927 [PMID: 21508939 DOI: 10.1038/clpt.2011.18]
- 38 **Relling MV**, Gardner EE, Sandborn WJ, Schmiegelow K, Pui CH, Yee SW, Stein CM, Carrillo M, Evans WE, Hicks JK, Schwab M, Klein TE. Clinical pharmacogenetics implementation consortium guidelines for thiopurine methyltransferase genotype and thiopurine dosing: 2013 update. *Clin Pharmacol Ther* 2013; **93**: 324-325 [PMID: 23422873 DOI: 10.1038/clpt.2013.4]
- 39 **Johnson JA**, Gong L, Whirl-Carrillo M, Gage BF, Scott SA, Stein CM, Anderson JL, Kimmel SE, Lee MT, Pirmohamed M, Wadelius M, Klein TE, Altman RB. Clinical Pharmacogenetics Implementation Consortium Guidelines for CYP2C9 and VKORC1 genotypes and warfarin dosing. *Clin Pharmacol Ther* 2011; **90**: 625-629 [PMID: 21900891 DOI: 10.1038/clpt.2011.185]
- 40 **Scott SA**, Sangkuhl K, Gardner EE, Stein CM, Hulot JS, Johnson JA, Roden DM, Klein TE, Shuldiner AR. Clinical Pharmacogenetics Implementation Consortium guidelines for cytochrome P450-2C19 (CYP2C19) genotype and clopidogrel therapy. *Clin Pharmacol Ther* 2011; **90**: 328-332 [PMID: 21716271 DOI: 10.1038/clpt.2011.132]
- 41 **Vizirianakis IS**. Improving pharmacotherapy outcomes by pharmacogenomics: from expectation to reality? *Pharmacogenomics* 2005; **6**: 701-711 [PMID: 16207147]
- 42 **Goldstein DB**. Pharmacogenetics in the laboratory and the clinic. *N Engl J Med* 2003; **348**: 553-556 [PMID: 12571264]
- 43 **Voora D**, Ginsburg GS. A hub for bench-to bedside pharmacogenomic-based research. *Pharmacogenomics* 2011; **12**: 1095-1098 [PMID: 21843063 DOI: 10.2217/pgs.11.62]
- 44 **Rowland M**, Peck C, Tucker G. Physiologically-based pharmacokinetics in drug development and regulatory science. *Annu Rev Pharmacol Toxicol* 2011; **51**: 45-73 [PMID: 20854171 DOI: 10.1146/annurev-pharmtox-010510-100540]
- 45 **Wu R**, Tong C, Wang Z, Mauger D, Tantisira K, Szeffler SJ, Chinchilli VM, Israel E. A conceptual framework for pharmacodynamic genome-wide association studies in pharmacogenomics. *Drug Discov Today* 2011; **16**: 884-890 [PMID: 21920452 DOI: 10.1016/j.drudis.2011.09.001]
- 46 **Gharani N**, Keller MA, Stack CB, Hodges LM, Schmidlen TJ, Lynch DE, Gordon ES, Christman MF. The Coriell personalized medicine collaborative pharmacogenomics appraisal, evidence scoring and interpretation system. *Genome Med* 2013; **5**: 93 [PMID: 24134832 DOI: 10.1186/gm499]
- 47 **Wheeler HE**, Maitland ML, Dolan ME, Cox NJ, Ratain MJ. Cancer pharmacogenomics: strategies and challenges. *Nat Rev Genet* 2013; **14**: 23-34 [PMID: 23183705 DOI: 10.1038/nrg3352]
- 48 **Zandi PP**, Judy JT. The promise and reality of pharmacogenetics in psychiatry. *Clin Lab Med* 2010; **30**: 931-974 [PMID: 20832660 DOI: 10.1016/j.cll.2010.07.004]
- 49 **Kirchheiner J**, Nickchen K, Bauer M, Wong ML, Licinio J, Roots I, Brockmüller J. Pharmacogenetics of antidepressants and antipsychotics: the contribution of allelic variations to the phenotype of drug response. *Mol Psychiatry* 2004; **9**: 442-473 [PMID: 15037866]
- 50 **Zhou SF**. Polymorphism of human cytochrome P450 2D6 and its clinical significance: Part I. *Clin Pharmacokinet* 2009; **48**: 689-723 [PMID: 19817501]
- 51 **de Leon J**, Armstrong SC, Cozza KL. Clinical guidelines for psychiatrists for the use of pharmacogenetic testing for CYP450 2D6 and CYP450 2C19. *Psychosomatics* 2006; **47**: 75-85 [PMID: 16384813]
- 52 **Horstmann S**, Binder EB. Pharmacogenomics of antidepressant drugs. *Pharmacol Ther* 2009; **124**: 57-73 [PMID: 19563827 DOI: 10.1016/j.pharmthera.2009.06.007]
- 53 **Reynolds GP**, McGowan OO, Dalton CF. Pharmacogenomics in psychiatry: the relevance of receptor and transporter polymorphisms. *Br J Clin Pharmacol* 2014; **77**: 654-672 [PMID: 24354796 DOI: 10.1111/bcp.12312]
- 54 **Perlis RH**. Pharmacogenomic testing and personalized treatment of depression. *Clin Chem* 2014; **60**: 53-59 [PMID: 24281779 DOI: 10.1373/clinchem.2013.204446]
- 55 **Jin Y**, Desta Z, Stearns V, Ward B, Ho H, Lee KH, Skaar T, Storniolo AM, Li L, Araba A, Blanchard R, Nguyen A, Ullmer L, Hayden J, Lemler S, Weinschilboum RM, Rae JM, Hayes DF, Flockhart DA. CYP2D6 genotype, antidepressant use, and tamoxifen metabolism during adjuvant breast cancer treatment. *J Natl Cancer Inst* 2005; **97**: 30-39 [PMID: 15632378]
- 56 **Borges S**, Desta Z, Li L, Skaar TC, Ward BA, Nguyen A, Jin Y, Storniolo AM, Nikoloff DM, Wu L, Hillman G, Hayes DF, Stearns V, Flockhart DA. Quantitative effect of CYP2D6 genotype and inhibitors on tamoxifen metabolism: implication for optimization of breast cancer treatment. *Clin Pharmacol Ther* 2006; **80**: 61-74 [PMID: 16815318]
- 57 **Henry NL**, Stearns V, Flockhart DA, Hayes DF, Riba M. Drug interactions and pharmacogenomics in the treatment of breast cancer and depression. *Am J Psychiatry* 2008; **165**: 1251-1255 [PMID: 18829880 DOI: 10.1176/appi.ajp.2008.08040482]
- 58 **Hoskins JM**, Carey LA, McLeod HL. CYP2D6 and tamoxifen: DNA matters in breast cancer. *Nat Rev Cancer* 2009; **9**: 576-586 [PMID: 19629072 DOI: 10.1038/nrc2683]
- 59 **Kelly CM**, Juurlink DN, Gomes T, Duong-Hua M, Pritchard KL, Austin PC, Paszat LF. Selective serotonin reuptake inhibitors and breast cancer mortality in women receiving tamoxifen: a population based cohort study. *BMJ* 2010; **340**: c693 [PMID: 20142325 DOI: 10.1136/bmj.c693]
- 60 **Sideras K**, Ingle JN, Ames MM, Loprinzi CL, Mrazek DP, Black JL, Weinschilboum RM, Hawse JR, Spelsberg TC, Goetz MP. Coprescription of tamoxifen and medications that inhibit CYP2D6. *J Clin Oncol* 2010; **28**: 2768-2776 [PMID: 20439629 DOI: 10.1200/JCO.2009.23.8931]
- 61 **Lash TL**, Rosenberg CL. Evidence and practice regarding the role for CYP2D6 inhibition in decisions about tamoxifen therapy. *J Clin Oncol* 2010; **28**: 1273-1275 [PMID: 20124162 DOI: 10.1200/JCO.2009.26.7906]
- 62 **Fleeman N**, Martin Saborido C, Payne K, Boland A, Dickson R, Dunder Y, Fernández Santander A, Howell S, Newman W, Oyee J, Walley T. The clinical effectiveness and cost-effectiveness of genotyping for CYP2D6 for the management of women with breast cancer treated with tamoxifen: a systematic review. *Health Technol Assess* 2011; **15**: 1-102 [PMID: 21906462 DOI: 10.3310/hta15330]
- 63 **Dickschen K**, Willmann S, Thelen K, Lippert J, Hempel G, Eissing T. Physiologically Based Pharmacokinetic Modeling of Tamoxifen and its Metabolites in Women of Different CYP2D6 Phenotypes Provides New Insight into the Tamoxifen Mass Balance. *Front Pharmacol* 2012; **3**: 92 [PMID: 22661948 DOI: 10.3389/fphar.2012.00092]
- 64 **Cheeti S**, Budha NR, Rajan S, Dresser MJ, Jin JY. A physiologically based pharmacokinetic (PBPK) approach to evaluate pharmacokinetics in patients with cancer. *Biopharm Drug Dispos* 2013; **34**: 141-154 [PMID: 23225350 DOI: 10.1002/bdd.1830]
- 65 **Chen Y**, Jin JY, Mukadam S, Malhi V, Kenny JR. Application of IVIVE and PBPK modeling in prospective prediction of clinical pharmacokinetics: strategy and approach during the drug discovery phase with four case studies. *Biopharm Drug Dispos* 2012; **33**: 85-98 [PMID: 22228214 DOI: 10.1002/bdd.1769]
- 66 **Sayama H**, Komura H, Kogayu M, Iwaki M. Development of a hybrid physiologically based pharmacokinetic model with drug-specific scaling factors in rat to improve prediction of human pharmacokinetics. *J Pharm Sci* 2013; **102**: 4193-4204 [PMID: 24018828 DOI: 10.1002/jps.23726]

- 67 **Jones HM**, Mayawala K, Poulin P. Dose selection based on physiologically based pharmacokinetic (PBPK) approaches. *AAPS J* 2013; **15**: 377-387 [PMID: 23269526 DOI: 10.1208/s12248-012-9446-2]
- 68 **Shaffer CL**, Scialis RJ, Rong H, Obach RS. Using Simcyp to project human oral pharmacokinetic variability in early drug research to mitigate mechanism-based adverse events. *Biopharm Drug Dispos* 2012; **33**: 72-84 [PMID: 22213407 DOI: 10.1002/bdd.1768]
- 69 **de Souza JA**, Olopade OI. CYP2D6 genotyping and tamoxifen: an unfinished story in the quest for personalized medicine. *Semin Oncol* 2011; **38**: 263-273 [PMID: 21421116 DOI: 10.1053/j.seminoncol.2011.01.002]
- 70 **Blue Cross Blue Shield Association; Kaiser Foundation Health Plan; Southern California Permanente Medical Group**. CYP2D6 pharmacogenomics of tamoxifen treatment. *Technol Eval Cent Assess Program Exec Summ* 2014; **28**: 1-4 [PMID: 24730084]

P- Reviewer: Cepeda C, Lee TM **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ



Phosphoprotein phosphatase 1-interacting proteins as therapeutic targets in prostate cancer

Juliana Felgueiras, Margarida Fardilha

Juliana Felgueiras, Margarida Fardilha, Laboratory of Signal Transduction, Centre for Cell Biology, Biology Department and Health Sciences Department, University of Aveiro, Campus de Santiago, 3810-193 Aveiro, Portugal

Author contributions: Felgueiras J and Fardilha M contributed to this paper.

Supported by Fundação para a Ciência e Tecnologia (FCT) (PTDC/QUI-BIQ/118492/2010) and Fundo Europeu de Desenvolvimento Regional (FEDER) (FCOMP-01-0124-FEDER-020895), Portugal

Correspondence to: Margarida Fardilha, PhD, Laboratory of Signal Transduction, Centre for Cell Biology, Biology Department and Health Sciences Department, University of Aveiro, Campus de Santiago, 3810-193 Aveiro, Portugal. mfardilha@ua.pt

Telephone: +351-918-143947 Fax: +351-234-377220

Received: June 29, 2014 Revised: September 1, 2014

Accepted: September 23, 2014

Published online: December 9, 2014

Abstract

Prostate cancer is a major public health concern worldwide, being one of the most prevalent cancers in men. Great improvements have been made both in terms of early diagnosis and therapeutics. However, there is still an urgent need for reliable biomarkers that could overcome the lack of cancer-specificity of prostate-specific antigen, as well as alternative therapeutic targets for advanced metastatic cases. Reversible phosphorylation of proteins is a post-translational modification critical to the regulation of numerous cellular processes. Phosphoprotein phosphatase 1 (PPP1) is a major serine/threonine phosphatase, whose specificity is determined by its interacting proteins. These interactors can be PPP1 substrates, regulators, or even both. Deregulation of this protein-protein interaction network alters cell dynamics and underlies the development of several cancer hallmarks. Therefore, the identification of PPP1 interactome in specific cellular context is of crucial importance. The knowledge on PPP1 complexes in prostate cancer remains scarce, with

only 4 holoenzymes characterized in human prostate cancer models. However, an increasing number of PPP1 interactors have been identified as expressed in human prostate tissue, including the tumor suppressors TP53 and RB1. Efforts should be made in order to identify the role of such proteins in prostate carcinogenesis, since only 26 have yet well-recognized roles. Here, we revise literature and human protein databases to provide an in-depth knowledge on the biological significance of PPP1 complexes in human prostate carcinogenesis and their potential use as therapeutic targets for the development of new therapies for prostate cancer.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Prostate cancer; Reversible phosphorylation; Phosphoprotein phosphatase 1; Phosphoprotein phosphatase 1-interacting proteins; Protein complexes; Therapeutic targets

Core tip: Protein kinases and phosphatases are challenging and valuable therapeutic targets for cancer. Here, we revise the relevance of phosphoprotein phosphatase 1 and its interactors for prostate carcinogenesis. Although only 4 complexes are characterized in human prostate cancer models, 81 additional interactors are expressed in human prostate tissue and, at least, 29 of which are involved in prostate carcinogenesis. This complex network has promising roles in the development of new therapies for prostate cancer. Therefore, efforts should be made in order to characterize their biological significance in prostate carcinogenesis.

Felgueiras J, Fardilha M. Phosphoprotein phosphatase 1-interacting proteins as therapeutic targets in prostate cancer. *World J Pharmacol* 2014; 3(4): 120-139 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/120.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.120>

INTRODUCTION

Prostate cancer (PCa) is the second most commonly diagnosed non-cutaneous cancer and a leading cause of cancer-related death in men worldwide^[1]. In spite of the recent advances in early diagnosis and therapeutic management of the disease, prognoses are still poor once the disease progresses to castration-resistant and metastasizes, mainly to bone^[2,3]. The urgent need for a panel of reliable biomarkers that could overcome the lack of cancer-specificity of prostate-specific antigen (PSA), as well as alternative therapeutic targets is challenging the scientific community^[4].

Reversible phosphorylation of proteins regulates more than 70% of all eukaryotic cellular processes^[5]. Phosphorylation at serine (Ser) and threonine (Thr) residues is accomplished by protein Ser/Thr kinases (PSTKs) and reversed by protein Ser/Thr phosphatases (PSTPs)^[6]. Deregulation of the counterbalanced action between PSTKs and PSTPs is frequently associated with system-wide disruption of signal transduction and malignant transformation of cells^[7,8]. For this reason, emergent studies have been focused on large-scale examination of PCa phosphoproteome^[9-12]. Androgen receptor (AR), for instance, is a phosphoprotein vital to the development and progression of PCa that presents, at least, 15 Ser and Thr phosphorylated residues. Phosphorylation of such residues modulates the transcriptional activity, subcellular localization, and stability of the AR^[13].

The mammalian genome encodes considerably more PSTKs than PSTPs-nearly 10 to 1^[14]. Hence, the success of the protein reversible phosphorylation system depends on the ability of PSTPs to form stable complexes with other proteins, giving rise to a huge number of distinct holoenzymes. This is particularly true for phosphoprotein phosphatase 1 (PPP1), a major PSTP of eukaryotic cells that controls a myriad of processes: glycogen metabolism, muscle contraction, RNA splicing, apoptosis, protein synthesis, cell cycle, among others^[15-18]. PPP1 exhibits an effective catalytic machinery, but lacks substrate specificity. Therefore, a number of regulatory subunits, also known as PPP1-interacting proteins (PIPs), have been associated with the spatiotemporal regulation of PPP1 activity^[19]. Given the key roles of PIPs, efforts have been made to characterize PPP1 interactomes in human tissues and to identify disease relevant PIPs^[20-24].

In contrast to PSTKs, whose therapeutic benefits have been largely explored, PSTPs had been considered not “drug-targetable” for years and, thus, remain understudied^[25,26]. In the case of PPP1, this vision is changing due to the increasing number of PPP1 holoenzymes that have been described, and which seem to be attractive targets for the development of new therapies^[27]. In fact, pharmaceutical companies are being encouraged to pursue approaches that aim the inhibition or activation of PPP1 holoenzymes.

Here, we revise literature and human protein databases to provide an in-depth knowledge on the relevance of PIPs, expressed in human prostate tissue, for prostate

carcinogenesis. Moreover, we address the biological significance of their interaction with PPP1 and consider their potential use as therapeutic targets for the management of human PCa.

RESEARCH

A comprehensive literature search of studies involving human samples or human cell lines was performed to identify articles on PPP1 and its interactors in human PCa. The Pubmed database was searched until May 2014 using the Medical Subject Heading (MeSH) whenever possible-for terms not included in MeSH (*e.g.*, “PP1-interacting protein” or “PP1 interactor”) a basic Pubmed search was employed instead. MeSH terms included: (“protein phosphatase 1” or “(PIP abbreviation) protein, human”) and (“prostate” or “prostatic neoplasms”). Reference lists of included studies and review articles were manually searched. The search was restricted to English-language literature.

For the sake of completeness, databases were reviewed: TissueNet and HIPPIE were used to identify additional PIPs expressed in human prostate tissue; BioGPS and The Human Protein Atlas were used to assess mRNA and protein expression levels, respectively; Gene Ontology Consortium was used to identify the biological processes in which proteins are involved; and, ScanProsite was used to identify PPP1 binding motifs for each PIP.

OVERVIEW OF PHOSPHOPROTEIN PHOSPHATASE 1 STRUCTURE

PPP1 is one of the most conserved proteins in eukaryotic species^[28,29]. In mammals, three genes encode the catalytic isoforms of the enzyme-PPP1CA, PPP1CB, and PPP1CC-which are ubiquitously expressed. Additionally, *PPP1CC* gene can generate two splice variants-PPP1CC1 and PPP1CC2-with the latter one being testis-enriched. This catalytic core is analogous, both in terms of structure and mechanism of action, to all members of the phosphoprotein phosphatase superfamily^[30]. The major divergences among PPP1 isoforms are found at NH₂- and COOH-terminal sequences^[13]. Interestingly, the N-terminal was shown to influence the properties of the active site and, consequently, the function of the enzyme and its sensitivity to inhibitors^[31,32].

PPP1 catalytic isoforms are not found freely in cells. PPP1 catalytic subunit (PPP1C) interacts with diverse regulatory subunits, known as PIPs, thus enabling the formation of distinct PPP1 multimeric holoenzymes. The nature of the relationship between PPP1 and PIPs greatly varies: (1) PIPs can be substrates for PPP1, with their functions being directly controlled through dephosphorylation by PPP1; (2) PIPs can determine the substrate specificity of PPP1 by either targeting PPP1C to specific subcellular compartments or enhancing/suppressing PPP1C activity towards different substrates; and

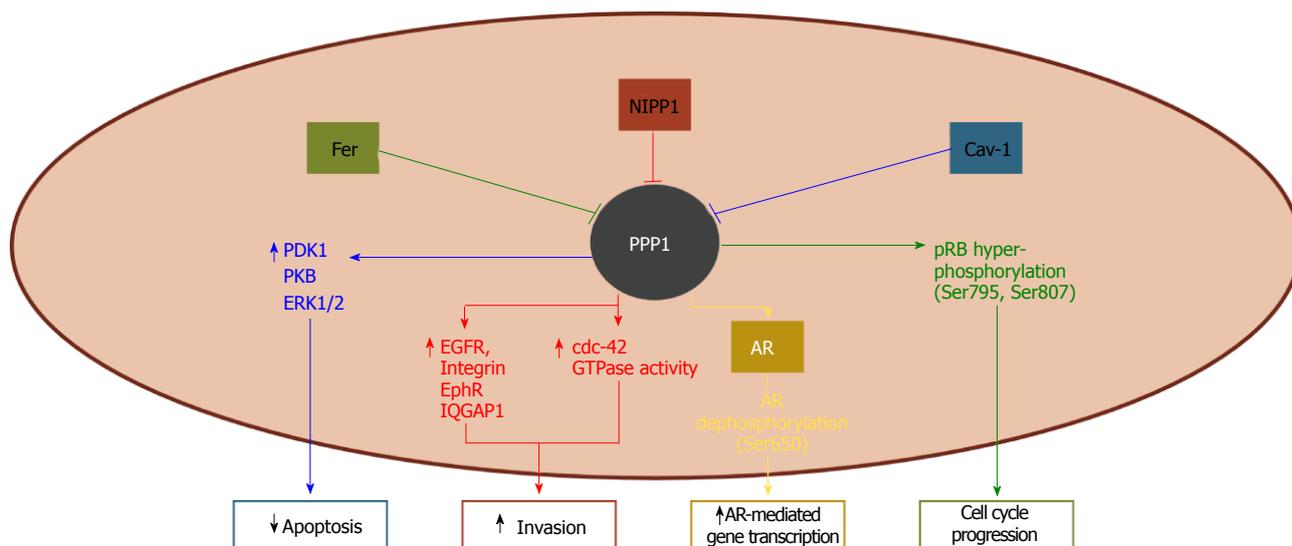


Figure 1 Phosphoprotein phosphatase-1 complexes described in human prostate cancer. Phosphoprotein phosphatase-1 (PPP1) dephosphorylates pRb at Ser795 and Ser807 and contributes to its hypophosphorylated and activated state. The activity of Fer tyrosine kinase leads to the inhibition of PPP1 and consequent hyperphosphorylation of pRb, which culminate in poor G-S transition control. The inhibition of PPP1 by the nuclear inhibitor of protein phosphatase 1 (NIPP1) increases the expression of epidermal growth factor receptor (EGFR), integrin, epherin receptor (EphR), and Ras GTPase-activating-like protein IQGAP1, as well as enhances the activity of cdc-42 GTPase. This, in turn, promotes invasiveness of tumor cells. Caveolin-1 (Cav-1) inhibits PPP1 and potentiates the activity of phosphoinositide-dependent kinase-1 (PDK1), protein kinase B (PKB), and extracellular signal-regulated kinase 1/2 (ERK1/2), increasing cell survival. PPP1 specifically dephosphorylates androgen receptor (AR) at Ser650, thus inhibiting AR nuclear export and enhancing AR-mediated gene transcription.

(3) some PIPs are simultaneously substrates and regulators of PPP1^[15,19]. More than 200 holoenzymes have already been identified and characterized, but thousand more remain unknown^[15,33].

PPP1 is not able to recognize a consensus sequence near the phosphorylated residue of its phosphotarget. Instead, PPP1C binding is mostly mediated by a short sequence (usually four to eight residues long), remote from the active site, commonly referred to as docking motif^[19]. A number of novel PPP1 binding sites have been mapped in PIPs, such as SILK and MyPhoNE, but the RVxF motif (x = any amino acid except proline) is still the most frequent, described for more than 70% of PIPs^[34,35]. It was also reported that some PIPs display isoform-specificity, suggesting that they possess isoform-specific docking motifs with putative location at the N- or C-terminal^[19].

PHOSPHOPROTEIN PHOSPHATASE 1 COMPLEXES CHARACTERIZED IN HUMAN PROSTATE CANCER

Imbalances in the protein phosphorylation system strongly contribute to carcinogenesis. In addition to the constitutive activation of oncogenic protein kinases, there is also evidence that the gain and loss of phosphorylation sites in relevant signaling proteins occur in human cancers^[36,37].

PPP1 has been shown to take part of various complexes that control cancer hallmarks^[24,38]. However, whether the role of PPP1 is pro- or anti-cancer largely depends on its interacting partners and cellular context. For instance, the interaction between PPP1CA and tensin1 impairs the

migration and invasion of cancer cells, and the interaction between PPP1CC and hScrib downregulates extracellular signal-regulated kinase (ERK) signaling, thus suppressing oncogene-induced transformation of primary rodent cells^[39,40]. On the other hand, the interplay between PPP1 and transforming growth factor- β (TGF β) drives malignant transformation of premalignant oral lesion cells^[41].

The knowledge on the involvement of PPP1 complexes in human PCa remains scarce. Few complexes have actually been characterized and even those are not fully understood; nonetheless, such complexes seem to have central roles in prostate carcinogenesis (Figure 1).

Androgen receptor/Phosphoprotein phosphatase 1

AR plays a key role in the development of PCa and, accordingly, androgen deprivation therapy is the standard hormonal treatment for the disease^[42,43]. AR is regulated by phosphorylation at multiple Ser residues and PPP1CA was shown to specifically reverse phosphorylation at Ser650^[44]. Since phosphoSer650 mediates AR nuclear export, PPP1CA dephosphorylation increases the stability and the transcriptional activity of the AR (Figure 1)^[44,45]. Besides being a substrate for PPP1, AR may also regulate the phosphatase activity by targeting it to chromatin, where PPP1 can modulate transcription and splicing events^[44].

Nuclear inhibitor of protein phosphatase 1 / Phosphoprotein phosphatase 1

Nuclear inhibitor of protein phosphatase 1 (NIPP1) is a ubiquitously expressed scaffold protein that was firstly identified as a PPP1 inhibitor^[46]. The interaction between NIPP1 and PPP1 was shown to orientate cell migration

by regulating the expression of integrin and growth factor receptors, and the activity of Cdc42 GTPase (Figure 1). Genetic disruption of this complex decreases the directional migration of PC-3 cells and impairs their migratory potential^[47].

Fer tyrosine kinase /Phosphoprotein phosphatase 1

Fer tyrosine kinase is highly expressed in human malignant prostate tissues compared to normal or benign tissues, which suggests its involvement in PCa progression^[48]. Fer interacts with signal transducer and activator of transcription 3 (STAT3) and phosphorylates AR at Tyr223, thus contributing to interleukin-6 (IL-6)-mediated AR activation and cell growth^[49,50]. Downregulation of Fer results in the activation of PPP1CA and consequent hypo-phosphorylation and activation of retinoblastoma protein (RB1), which, in turn, leads to cells arrest at the G₀/G₁ phase (Figure 1)^[51]. Accordingly, downregulation of Fer impairs the proliferation of PCa cells and their ability to form colonies in soft agar^[48].

Caveolin-1 /Phosphoprotein phosphatase 1

Caveolin-1 (Cav-1) is overexpressed in human PCa and correlates positively with Gleason score, thus being suggested as a potential prognostic marker^[52-55]. It has also been proposed as biomarker to monitor the response to treatments with dasatinib and sunitinib^[56]. Cav-1-mediated cell survival depends on its interaction with and inhibition of PPP1 (and also PPP2), leading to the increased activity of phosphoinositide-dependent kinase-1, v-akt murine thymoma viral oncogene homolog (AKT), and ERK1/2 (Figure 1)^[57].

ADDITIONAL PHOSPHOPROTEIN PHOSPHATASE 1-INTERACTING PROTEINS EXPRESSED IN HUMAN PROSTATE TISSUE

In spite of the limited number of PPP1 complexes experimentally characterized in human PCa models, several additional PIPs have already been identified as expressed in human prostate tissue. The human prostate proteome includes a total of 81 hitherto experimentally detected PIPs (Table 1)^[58,59], but many more may remain unknown. None of the interactors is prostate-specific; however, 28 are highly expressed in prostate tissue, namely BAD, BCL2, CCND1, CUEDC2, GABARAPL2, HCFC1, HDAC1, HDAC10, HEYL, IKBKB, LMTK2, MAP1LC3B, MYC, NOM1, PPP1R3D, PPP1R7, PPP1R11, PPP1R13B, PPP1R37, RB1, RRP1B, RYR2, SH2D4A, STAM, STAU1, SYTL2, TRIM28, and ZFYVE9 (Table 1)^[60].

Of the interactions identified, 67 were described for a specific PPP1 isoform, while 6 seem to be common to all isoforms (Table 1)^[58,59]. In the vast majority, binding to PPP1 is assured *via* RVxF motif, although less described SILK, MyPhoNE, RARA, and other motifs (*e.g.*, apop-

totic signature motifs and inhibitor-2 degenerate motif) are also present in some PIPs (Table 1).

INTERACTORS OF PHOSPHOPROTEIN PHOSPHATASE 1 IN PROSTATE CARCINOGENESIS: VALUABLE TOOLS FOR CANCER MANAGEMENT

PIPs expressed in human prostate tissue are key mediators of several signaling pathways and cellular processes, such as apoptosis, transcription, cell cycle, development/differentiation, and immunology/inflammation.

In the context of human prostate carcinogenesis, only 31 of these proteins have well-recognized functions. In this section, we revise the contribution of these PIPs to prostate carcinogenesis, focusing on studies involving human tissue samples or cell lines.

Apoptotic protease-activating factor 1

Apoptotic protease-activating factor 1 (APAF1) is responsible for the cleavage of procaspase-9 and mitochondria-mediated activation of caspases-9, being a major effector of apoptosis^[61].

An alternative splicing product of APAF1, known as APAF1-ALT, was found in LNCaP cells. APAF1-ALT exhibits a defective pro-apoptotic function and its expression was shown to be increased under infective conditions. Therefore, this spliced form may be particularly involved in inflammation and carcinogenesis, since it compromises the apoptotic pathway^[62].

Resveratrol, sulforaphane, and vitamin D3 exert their tumor suppressive functions through changes in the gene that encodes for APAF1, at least in part^[63-65]. APAF1 apoptosome is also involved in malignant cells-selective induction of apoptosis by apoptin^[66].

Ataxia telangiectasia mutated kinase

Ataxia telangiectasia mutated (ATM) is a ubiquitously expressed Ser/Thr kinase with a wide spectrum of downstream targets involved in cell-cycle control, DNA repair after radiation-induced damage, and apoptosis^[67].

The expression levels of ATM are similar or higher in PCa samples compared to normal prostate tissue; however, its activation is higher in precursor stages of prostate tumorigenesis, like PIN^[68,69].

Variants of the *ATM* gene have been associated with the risk of PCa development, and might be useful predictive markers of adverse responses to radiotherapy^[70-72]. ATM maintains telomeres' length and mediates tumor surveillance^[68,69,73].

Downregulation of ATM increases LNCaP, DU-145, and PC-3 cells' sensitivity to radiation-induced apoptosis^[74-77]. The molecular events that arose from ATM inhibition include increased mitotic index, augmented expression of E2F transcription factor and proliferating cell nuclear antigen, and inhibition of G₂ arrest in response

Table 1 Interactors of phosphoprotein phosphatase-1 expressed in human prostate tissue

PIP	Uniprot ID	Biological processes	PPP1 specificity	PPP1 binding motif
AKAP11	Q9UKA4	Intracellular signal transduction	PPP1CB, PPP1CC	RVxF, other motifs
APAF1	O14727	Apoptosis	PPP1CA	RVxF, other motifs
ATM	Q13315	Cell cycle; response to DNA damage; protein phosphorylation	PPP1CA	RVxF, SILK
AXIN1	O15169	Intracellular signal transduction; apoptosis; regulation of protein phosphorylation; transcription	PPP1CA	-
BAD ¹	Q92934	Apoptosis	PPP1CA	other motifs
BCL2 ¹	P10415	Apoptosis; response to DNA damage; transmembrane transport	PPP1CA, PPP1CB	RVxF, other motifs
BCL2L2	Q92843	Apoptosis	PPP1CA	RVxF, other motifs
BRCA1	P38398	Cell cycle; DNA repair; lipid metabolism	PPP1CA, PPP1CB, PPP1CC	RVxF, other motifs
CCND1 ¹	P24385	Cell cycle; response to DNA damage; transcription	PPP1CB	-
CCND3	P30281	Cell cycle; intracellular signal transduction; protein phosphorylation	PPP1CB	-
CDC5L	Q99459	Cell cycle; transcription; mRNA splicing	PPP1CA	RVxF
CDC34	P49427	Cell cycle; protein ubiquitination; intracellular signal transduction	PPP1CB	other motifs
CDK2	P24941	Cell cycle; DNA repair; meiosis; mitosis; intracellular signal transduction; cell proliferation	PPP1CA	-
CDK4	P11802	Cell cycle; protein phosphorylation; cell proliferation	PPP1CA	-
CSRN2	Q9H175	Apoptosis; transcription; protein phosphorylation	PPP1CA	RVxF, SILK
CUEDC2 ¹	Q9H467	Intracellular signal transduction	PPP1CA	-
CUL1	Q13616	Host-virus interaction; intracellular signal transduction	PPP1CA	-
EED	O75530	Transcription	PPP1CA	RVxF
EIF2AK2	P19525	Transcription; immunity; host-virus interaction	PPP1CA	other motifs
GABARAP	O95166	Apoptosis; autophagy; transport	PPP1CC	other motifs
GABARAPL2 ¹	P60520	Autophagy; transport	PPP1CC	-
GRB2	P62993	Host-virus interaction	PPP1CB	RVxF
GSK3B	P49841	Carbohydrate metabolism; differentiation; intracellular signal transduction	PPP1CA	-
HCFC1 ¹	P51610	Cell cycle; host-virus interaction	PPP1CA	RVxF, RARA
HDAC1 ¹	Q13547	Transcription; host-virus interaction; biological rhythms	PPP1CC	RVxF
HDAC6	Q9UBN7	Transcription; autophagy	PPP1CC	RVxF
HDAC8	Q9BY41	Transcription	PPP1CC	-
HDAC10 ¹	Q96958	Transcription	PPP1CC	-
HEYL ¹	Q9NQ87	Transcription; intracellular signal transduction	PPP1CA	RVxF
HSPA8	P11142	Host-virus interaction; mRNA processing; transcription	PPP1CA	other motifs
IKBK ¹	O14920	Intracellular signal transduction	PPP1CA	other motifs
IKBK ²	Q9Y6K9	Transcription; host-virus interaction	PPP1CB, PPP1CC	RARA
LMTK2 ¹	Q8IWU2	Protein phosphorylation; intracellular transport; receptor recycling	PPP1CA	RVxF, other motifs
MAP1LC3A	Q9H492	Autophagy; intracellular signal transduction	PPP1CC	other motifs
MAP1LC3B ¹	Q9GZQ8	Autophagy; intracellular signal transduction	PPP1CC	-
MAP3K3	Q99759	Intracellular signal transduction; protein phosphorylation	PPP1CA, PPP1CC	RVxF
MAX	P61244	Transcription	PPP1CA, PPP1CB	-
MDM4	O15151	Cell cycle; cell proliferation; apoptosis; response to DNA damage and hypoxia; protein stabilization; protein complex assembly	PPP1CA, PPP1CB, PPP1CC	-
MPHOSPH10	O00566	Ribosome biogenesis; RNA processing	PPP1CA	RVxF
MYC ¹	P01106	Transcription	PPP1CA	RVxF
NCL	P19338	Transcription; angiogenesis	PPP1CB	other motifs
NCOR1	O75376	Transcription	PPP1CA, PPP1CB, PPP1CC	RVxF, other motifs
NOC2L	Q9Y3T9	Apoptosis; transcription	PPP1CA	RVxF
NOM1 ¹	Q5C9Z4	Targets PPP1CA to the nucleolus	PPP1CA	RVxF, SILK
PAK6	Q9NQ55	Transcription; protein phosphorylation; cytoskeleton organization; apoptosis	-	Other motifs
PLCL2	Q9UPR0	Intracellular signal transduction; lipid metabolic process	PPP1CA	RVxF
PPP1R2	P41236	Regulation of phosphoprotein phosphatase activity; regulation of signal transduction; carbohydrate and glycogen metabolism	PPP1CB, PPP1CC	SILK, other motifs
PPP1R3B	Q86X16	Carbohydrate and glycogen metabolism	PPP1CA	RVxF
PPP1R3D ¹	O95685	Regulation of protein dephosphorylation; carbohydrate and glycogen metabolism	PPP1CC	RVxF
PPP1R7 ¹	Q15435	Regulation of protein dephosphorylation; regulation of catalytic activity	PPP1CB	other motifs
PPP1R10	Q96QC0	Regulation of catalytic activity; transcription; protein import into nucleus	PPP1CA	RVxF
PPP1R11 ¹	O60927	Regulation of catalytic activity	PPP1CB	RVxF
PPP1R12A	O14974	Regulation of catalytic activity; intracellular transport; cell cycle; regulation of cell adhesion; protein dephosphorylation; intracellular signal transduction	PPP1CB	RVxF, MyPhoNE
PPP1R13B ¹	Q96KQ4	Cell cycle; apoptosis	PPP1CA	RVxF
PPP1R14B	Q96C90	Regulation of phosphorylation; regulation of catalytic activity	PPP1CC	RVxF
PPP1R15A	O75807	Apoptosis; regulation of translation; stress response	PPP1CA, PPP1CB, PPP1CC	RVxF, RARA
PPP1R15B	Q5SWA1	Regulation of translation; stress response; dephosphorylation	PPP1CA	RVxF, other motifs
PPP1R26	Q5T8A7	Regulation of phosphatase activity	PPP1CA	RVxF

PPP1R37 ¹	O75864	Regulation of phosphatase activity	PPP1CA	RVxF
PTEN	P60484	Lipid metabolism; apoptosis; neurogenesis	PPP1CA	RVxF
PTK2	Q05397	Angiogenesis	PPP1CB	SILK
RB1 ¹	P06400	Transcription; cell cycle; host-virus interaction	PPP1CA	RVxF, SILK, other motifs
RIPK3	Q9Y572	Necrosis	PPP1CB, PPP1CC	RVxF
RPAP2	Q8IXW5	Transcription	PPP1CA	RVxF
RPAP3	Q9H6T3	Alternative splicing; polymorphism	PPP1CA	other motifs
RRP1B ¹	Q14684	Regulation of phosphatase activity; RNA processing	PPP1CA	RVxF, other motifs
RUVBL2	Q9Y230	DNA repair; growth regulation; transcription	PPP1CA	-
RYR21	Q92736	Intracellular transport	PPP1CA, PPP1CB, PPP1CC	RVxF, other motifs
SF3A2	Q15428	mRNA processing and splicing	PPP1CA	-
SH2D4A ¹	Q9H788	Regulation of phosphatase activity	PPP1CB	RVxF, MyPhoNE
SKP1	P63208	Intracellular signal transduction	PPP1CA	RVxF
SMARCB1	Q12824	Transcription; cell cycle; host-virus interaction; neurogenesis	PPP1CA, PPP1CB, PPP1CC	RVxF
SPRED1	Q7Z699	Regulation of protein phosphorylation; regulation of protein deacetylation; response to DNA damage; development	PPP1CA	RVxF
STAM ¹	Q92783	Protein transport	PPP1CA	RVxF
STAU1 ¹	O95793	Intracellular mRNA localization	PPP1CA	RVxF, other motifs
SYTL2 ¹	Q9HCH5	Intracellular transport; exocytosis; regulation of phosphatase activity	PPP1CA	RVxF, SILK, other motifs
TMEM33	P57088	-	PPP1CB	-
TP53	P04637	Apoptosis; cell cycle; host-virus interaction; necrosis; transcription	PPP1CA	-
TP53BP2	Q13625	Intracellular signal transduction; apoptosis; cell cycle; embryo development; heart development; response to ionizing radiation	PPP1CA, PPP1CC	RVxF, other motifs
TRIM28 ¹	Q13263	Transcription; DNA repair; protein ubiquitination; protein sumoylation; protein oligomerization; gene expression; epithelial to mesenchymal transition; innate immune response; regulation of viral release from host cell	PPP1CA, PPP1CB, PPP1CC	RVxF
TUSC3	Q13454	Intracellular transport	PPP1CA	RVxF, other motifs
USF1	P22415	Transcription	PPP1CC	RVxF, other motifs
ZFYVE9 ¹	O95405	Intracellular signal transduction	PPP1CA, PPP1CB, PPP1CC	RVxF, other motifs
ZFYVE16	Q7Z3T8	Intracellular signal transduction; regulation of endocytosis; protein targeting	PPP1CA	RVxF, other motifs

¹Highly expressed in human prostate (criteria selection: prostate mRNA expression higher than the mean mRNA expression taking into account all tissues analyzed). Other motifs include apoptotic signature motifs and inhibitor-2 degenerate motif. PPP1: Phosphoprotein phosphatase-1; ATM: Ataxia telangiectasia mutated; GSK3B: Glycogen synthase kinase-3 β ; SMARCB1: SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily B member 1; BRCA1: Breast cancer type 1 susceptibility protein.

to DNA damage^[78]. Therefore, *ATM* gene therapy and the use of *ATM* inhibitors have been explored as adjuvants to radiation therapy in PCa.

Axis inhibition protein 1

Axis inhibition protein 1 (AXIN1) is a tumor suppressor that integrates the β -catenin destruction complex, along with adenomatous polyposis coli protein and glycogen synthase kinase-3 β (GSK3B).

Wnt/ β -catenin signaling pathway has been extensively explored due to its impact on development, proliferation, and tumorigenesis^[79]. Mutations in the signaling mediators of such system are reported in several types of cancer. For instance, 7 variations in the DNA sequence of axin-1 were found in specimens with abnormal β -catenin immunohistochemistry and 4 different polymorphisms were observed in LNCaP, DU145, PC-3, 22Rv1, and P69SV40T cell lines, as well as in the sublines M12, M2182, M2205^[80].

Bcl-2 family members: BCL2, BCL2L2 and BAD

Members of the Bcl-2 family of proteins are pivotal regulators of apoptosis. Bcl-2 (BCL2) and Bcl-2-like pro-

tein 2 (BCL2L2) are anti-apoptotic proteins, while Bcl-2 antagonist of cell death (BAD) has proapoptotic functions^[81].

The expression of BCL2 is not observed in normal prostate epithelial cells, but is found in PIN and increases in advanced PCa (further details in Catz *et al.*^[82]). Higher BCL2 expression is also found in patients that underwent radiotherapy before surgery than those who received surgical treatment as first choice^[83]. BCL2 upregulation is required for the acquisition of castration-resistance, in part by suppressing TGF β and dihydrotestosterone-mediated induction of caspase-1 expression and activation^[84,85].

In conformity with BCL2, the expression of BAD is found elevated in highly proliferative states, in spite of not being helpful in the discrimination between benign and malignant prostate tissues^[86]. The overexpression of proapoptotic proteins in highly proliferative states seems paradoxical since cancer cells normally take advantage of the molecular machinery to evade apoptosis. However, BAD overexpression was shown to stimulate PCa cells proliferation and enhance tumor growth^[87]. On the other hand, overexpression of BAD in LNCaP cells, which are resistant to tumor necrosis factor-related apoptosis-

inducing ligand (TRAIL)-induced apoptosis, renders the cells sensitive to TRAIL effects^[88].

Several studies have reported the great value of targeting apoptotic molecules in order to increase the sensitivity to apoptosis-inducing agents. Polygene therapy and other combinatorial approaches are receiving increased attention due to their effectiveness^[89]. Since BCL2 is associated with increased resistance to androgen deprivation in LNCaP cells, a number of approaches aim to decrease its expression and phosphorylation state^[90,91]. The targeting of BCL2 has been explored not only in already settled castration-resistant cases, but also to delay the progression to this advanced state^[92,93]. In the case of BCL2L2, it was shown to be a target of miR-205-modulated chemosensitivity^[94]. Pharmacological interventions targeting BAD intent to increase its expression and decrease its phosphorylation state^[95-97].

Breast cancer type 1 susceptibility protein

Breast cancer type 1 susceptibility protein (BRCA1) has long been described as a tumor suppressor that regulates gene transcription and DNA damage repair^[98]. In spite of the relevance of BRCA1 mutations in other types of cancers, evidences of their association with PCa development have been inconsistent and at times contradictory. While some studies point to their irrelevance in PCa development, others state that carriers of BRCA1 mutations have more aggressive phenotype and are more prone to develop distant metastasis^[99-102].

The expression profile of BRCA1 during PCa progression is also very heterogeneous, although it tends to be higher in PCa compared to normal prostate epithelium^[103,104]. Some works actually suggest that its expression correlates with increased tumor proliferative index and development of lethal cancer, being, therefore, considered a potential prognostic marker^[105].

The mode of action of BRCA1 in PCa is complex and seeks clarification^[102]. BRCA1 mediates apoptosis, cell-cycle arrest, and the response to doxorubicin treatment in PC-3 cells by targeting a wide variety of genes (*e.g.*, *CCND1*, *BLM*, *BRCA2*, *DDB2*, *FEN1*, *H3F3B*, *CCNB2*, *MAD2L1*, and *GADD153*)^[106]. It was also shown to negatively regulate the transcription of insulin-like growth factor I receptor in an AR-dependent manner^[104].

The use of anticancer drugs that inhibit poly ADP-ribose polymerase (PARP), such as niraparib and olaparib, has demonstrated efficacy in PCa patients with BRCA1 mutations^[107,108].

Cyclins D1 and D3

Cyclins are key mediators of the cell cycle. Cyclin D1 (CCND1) is scarcely found in non-neoplastic tissues, but its levels are increased in the majority of localized tumors, where distinct subcellular localizations are observed according to tumor grading^[109]. Likewise, CCND3 displays higher expression in PCa than in BPH and its expression correlates positively with PSA serum levels^[110].

In addition to the roles of cyclins in the regulation

cell cycle, CCND1/3 interact with AR. CCND1 suppresses the activity of the AR either directly, with the main mediator being the repressor domain of CCND1, or indirectly *via* histone deacetylases^[111]. In this fashion, CCND1 differentially regulates the expression of several androgen-sensitive genes-it represses some genes, such as *KLK3/PSA*, while induces the transcription of others, as *CDC6* and *MCM2*. Further effects of CCND1 include alteration of transcription factor-chromatin interactions, restraining of TGF β , Snail, Twist, and Goosecoid signaling pathways, enhancement of *Wnt* and *ES* gene expression, and enlargement of a prostate stem cell population^[112,113]. The association between CCND3 and the AR represses ligand-dependent activation through cyclin-dependent kinase 4 (CDK4)-independent mechanisms and appears androgen-dependent proliferation^[114].

CCND1 has been proposed as a prognostic marker for poor clinical outcome in PCa biochemical-free recurrence. A number of strategies targets CCND1, including miR-153 and perhaps miR-449a, piperine, and L-mimosine, with the effect of the latter being only observed in PC-3 cells^[115-118].

Cyclin-dependent kinases 2 and 4

CDK family of proteins regulates cell cycle progression and is involved in AR-mediated cell proliferation. CDK2 mRNA levels decrease after castration, increase after testosterone propionate treatment, and are expressed at high levels in recurrent human xenograft CWR22 tumors^[119]. The expression of CDK2 and CDK4 is up-regulated within hours of androgen treatment; nevertheless, castration-resistant PC-3 cells, which do not respond to androgen stimulation, show constitutively high basal expression of both kinases^[120].

The activity of CDK2 kinase is stimulated by androgen^[119,120]. Increased CDK2 activity correlates with PCa cells insensitivity to TGF- β 1, while even modest depletion of CDK2 in LNCaP cells results in strong growth repression^[121,122].

CDK4 protein expression was not found elevated in localized prostate tumors, but its overexpression overcome 3,9-dihydroxy-2-prenylcoumestan-induced G₀/G₁ arrest in castration-resistant cells^[123].

Decreased expression and inhibition of the activity of CDK2 and CDK4 are observed upon treatment with anti-proliferative agents, such as resveratrol, BZL 101, and inositol hexaphosphate^[124-126]. As phosphorylation of CDK2 on Thr160 is essential for the kinase activity, the manipulation of this phospho-residue has also been analyzed^[127].

GSK3B

The multifunctional GSK3B exhibits potent tumor suppressor qualities and is upregulated in many types of tumor, including PCa. The expression pattern of GSK3B differs between normal prostate and PCa cells - nuclear GSK3B is higher in normal prostate, whereas cytoplasmic GSK3B is higher in PCa. Increased GSK3B cytoplasmic

levels might determine PCa development and progression due to their correlation with high Ki-67 labeling index, low apoptotic index by TUNEL, high levels of AR and phosphorylated AKT, extracapsular extension, high Gleason score, lymph node metastasis, and biochemical recurrence-free survival^[128].

GSK3B mediates both estrogen and AR signaling (for details see Mulholland *et al.*^[129]). GSK3B phosphorylates AR and represses AR-mediated transcription and growth^[130,131]. GSK3B/AR complex, which locates within the cytoplasm and nucleus, contributes to AR stability, nuclear translocation, and consequent modulation of PCa cells' response to androgen^[132]. The signaling pathway AKT/GSK3B is also involved in nuclear factor α (NF α)-induced epithelial-mesenchymal transition (EMT) in PC-3 cells by contributing to Snail stability^[133]. On the other hand, suppression of GSK3B sensitizes PCa cells to TRAIL-induced apoptosis, which might suggest its involvement during resistance acquisition^[134]. The suppression of GSK3B expression or phosphorylation state has also demonstrated positive results in inhibiting proliferation of PCa cells^[135,136].

AKT/GSK3B signaling pathway has been targeted by a number of antiproliferative and apoptosis-induced agents, namely thiazolidinediones and isoflavone, as well as agents that impair cell migration and invasion, such as fenretinide^[137-139].

Histone deacetylases-1, -6 and -8

Histone deacetylases (HDACs) are a large family of enzymes that regulates the nucleosomal histone acetylation. Members of HDACs' family are divided into four classes (classes I-IV), according to their homology with yeast proteins, with class II being further subdivided into class II a and II b. HDAC1 and -8 belong to the class I histone deacetylases, whereas HDAC6 belongs to the class II b. All members of class I were found to be deregulated in many types of cancers (for review see^[140]).

HDAC1 is expressed in normal prostate tissues, where it locates exclusively in the nucleus, cancer precursor lesions, and PCa, and its expression was shown to be lower in stromal cells^[141,142]. Conversely, HDAC8 is primarily found in the cytoplasm of stromal cells^[142].

HDAC1 expression levels correlate with tumor dedifferentiation, high Gleason score, high pT stage, and high biochemical recurrence rates^[141,143]. HDAC1 is a major repressor of AR and E-cadherin, thereby regulating AR-transcriptional activity, cell proliferation and motility, and invasion^[144-146].

HDAC6 deacetylates and activates HSP90 chaperone protein, which, in turn, binds to AR^[147]. Indeed, HDAC6 regulates AR hypersensitivity to androgens, nuclear localization, and attenuation of its degradation^[147,148]. HDAC6 might establish important interactions with other proteins since its decrease is also observed in PC-3 cells, which are castration-resistant^[149,150].

The use of HDAC inhibitors in the prevention and treatment of cancer has become an area of intense re-

search. The repression of HDAC1 expression by miR-449a induces growth arrest in PCa and its inhibition by maspin prevents pathologic gene silencing, increasing tumor cell's sensitivity to drug-induced apoptosis^[151,152]. The deacetylase activity of HDAC6 decreases after sulforaphane treatment and it might be responsible for the selective effects of this agent in both hormone-sensitive and castration-resistant cells, while normal cells remain intact^[149,150].

Heyl

Heyl is a member of the hairy/enhancer-of-split-related with YRPW-like motif family of transcriptional repressors. Of the three members of the referred protein family, Heyl is the more potent AR corepressor and reduces the growth of LNCaP cells. The repression of AR activity by Heyl occurs through HDAC1/2-independent mechanisms. Heyl was shown to be excluded from the nucleus in malignant cells but not in benign tissue, thus nuclear exclusion of the protein might be involved in tumor progression^[153].

Inhibitor of nuclear factor- κ B kinase subunit beta and gamma

Inhibitor of nuclear factor- κ B (NF- κ B) kinase subunit β and γ (IKBKB and IKBKG, respectively) are involved in the activation of NF- κ B^[154], a transcription factor that regulates cell growth, apoptosis, inflammation, angiogenesis, and metastasis (for review see^[155]).

Studies on human prostate cell lines have not revealed significant differences between primary prostate cells, hormone-sensitive, and castration-resistant PCa cells^[156]. However, IKBKB expression is higher in PCa tissue than in benign non-atrophic and atrophic glands^[157].

The effects of sulforaphane and phenethyl isothiocyanate are mediated, at least in part, through the inhibition of IKBKB phosphorylation^[158]. The loss of IKBKB and IKBKG are also involved in proteasome inhibitors-induced apoptosis^[159].

Lemur tyrosine kinase 2

Lemur tyrosine kinase 2 (LMTK2) is a Ser/Thr transmembrane protein kinase mainly involved in endosomal membrane trafficking^[160]. In LNCaP cells, this function is achieved, at least in part, by the interaction with myosin VI and consequent recruitment of this protein to the surface of endosomes^[161]. Interestingly, the gene that encodes for LMTK2 is one of the novel common alleles associated with PCa^[162]. LMTK2 is underexpressed in PCa tissue compared to non-malignant BPH tissue due to alterations in intron 9; however, the mechanism by which this alteration leads to the increased risk of PCa is not properly understood^[163]. LMTK2 functions depend on its interaction with other proteins, which includes CDK/P35 complex and PPP1, besides the already mentioned myosin VI^[164,165].

MYC family of proteins: MYC and MAX

MYC deregulation is a well-established mechanism in car-

cinogenesis^[166]. The role of MYC in PCa, nevertheless, is not fully understood. MYC overexpression is frequently observed in PCa, which can be partially explained by locus amplification, mainly in advanced cancers^[167]. MYC stabilizes the length of telomeres and is required for EMT^[168,169]. MYC and MAX interact with and regulate the AR^[170].

MYC amplification status and its overexpression have been suggested as a valuable prognostic tool^[171,172]. The existence of a panel of markers that encompass MYC, PTEN, and Ki67 shown benefits in predicting progression-free survival in men receiving adjuvant docetaxel after prostatectomy^[173]. Cells that exhibit resistance to treatment with docetaxel have constitutive activation of MYC signaling^[174].

Nucleolin

Nucleolin (NCL) is an abundant nucleolar phosphoprotein involved in various stages of ribosome synthesis. The expression and phosphorylation of NCL are extremely sensitive to androgens-with both decreasing following androgen deprivation. Thus, the control of NCL expression and phosphorylation by androgens may be an important nucleolar control mechanisms involved in the growth of prostate cells^[175]. NCL can also be found in the cell surface, where it may function as a hepatocyte growth factor receptor. Cell surface NCL was shown to be upregulated during PCa progression^[176].

Nuclear corepressor 1

Nuclear corepressor 1 (NCOR1), an AR co-repressor, is overexpressed in PCa cell lines compared to normal prostate cells. NCOR1 expression is confined to the S phase of cell cycle; therefore, during this time NCOR1 represses the expression of AR target genes^[177]. In PCa cells, the activity of NCOR1 is positively regulated by protein kinase A (PKA)^[178].

The increased expression and activity of NCOR1 impair peroxisome proliferator activated receptor α/γ -mediated expression of key target genes, such as *CDKN1A* and *TGFBRAP1*, thus contributing to the loss of ligands anti-proliferative responsiveness in PCa cells^[179]. NCOR1 might also be important during the process of castration-resistant acquisition^[180].

Serine/threonine-protein kinase PAK 6

The expression of PAK6 is increased in primary and metastatic PCa, and correlates with cells' sensitivity to androgens^[181,182]. PAK6 co-localizes with AR in the cytoplasm of normal prostate epithelium and translocates into the nucleus in malignant phenotypes, where it represses both AR- and ER-mediated gene transcription^[181,183,184]. It was also shown that PAK6 phosphorylates the AR at Ser578, promoting the association of AR-E3 ligase murine double minute-2 (Mdm2) and guiding AR degradation^[185].

The knockdown of PAK6 impairs PCa growth and improves chemosensitivity of docetaxel and sensitivity to radiation^[186,187].

Phosphatase and tensin homolog

Phosphatase and tensin homolog (PTEN) is a dual speci-

ficity phosphatase and a recognized tumor suppressor. Inactivation of *PTEN* has been associated with many different types of cancer, including PCa, and assumes preponderant roles (further details on^[188,189]). A number of molecules have been recently shown to contribute to PTEN downregulation, including lamin A/C and a subset of microRNAs (*e.g.*, miR-19b, miR-23b, miR-26a, miR-92a, and miR-153)^[117,190,191]. Loss of PTEN determines PCa progression through several downstream effectors and signaling pathways, including PI3K/AKT, BIM1, CXCL12/CXCR4, and PDGF D/ β -PDGFR^[192-195]. The loss of PTEN is also associated with increased risk of capsular penetration^[196]. Interestingly, it was recently shown that PTEN is incorporated in the cargo of exosomes prevent from cancer cells, but not in those derived from non-malignant cells. Exosomes are able to transfer PTEN to other cells, which in turn recover the tumor-suppressor activity^[197].

Recent evidences support the usefulness of PTEN in PCa management. Blood exosomes of PCa patients contain PTEN, contrarily to the exosomes isolated from normal subjects, which may indicate exosomal PTEN as a putative diagnostic tool^[197]. The loss of cytoplasmic PTEN was shown to accurately distinct intraductal carcinoma from prostatic intraepithelial neoplasia, since the latter does not manifest PTEN loss at all^[198]. PTEN status might also be useful in the prognostic evaluation of men with localized PCa^[199,200].

Moreover, a phase II clinical trial reported that the activity of PTEN determines the improvement of progression-free survival and is potentially required for the efficacy of cetuximab in metastatic castration-resistant PCa^[201]. PTEN expression is enhanced by resveratrol-mediated AR inhibition^[202].

Protein tyrosine kinase 2

Protein tyrosine kinase 2 (PTK2) regulates adhesion and motility of cells. Its upregulation and activation was observed in localized and castration-resistant PCa, in spite of being more evident in the latter case^[203,204]. The complexes that PTK2 forms with paxillin and p50csk are mainly observed in metastatic PCa and contribute to the metastatic behavior^[204]. PTK2 is also involved in the migration and invasion mediated by IL-8 and CXCL13-CXCR5^[205,206].

Treatments with FTY720 and the combinatorial therapy with curcumin and methylseleninic acid compromises PTK2 activity, and PTK2 inhibition was shown to delay the progression of PCa^[205,207]. PTK2 is also a target of genistein-mediated morphologic changes^[208].

Tripartite motif-containing protein 28

Tripartite motif-containing protein 28 (TRIM28) is a substrate of ATM kinase involved in the maintenance of chromatin in condensed states^[209]. The expression of TRIM28 is observed in prostate cancer lines, despite being lower in castration-resistant cell lines^[210]. TRIM28 was recently identified as an activator of the AR and is also

involved in the response of prostate cells to DNA damage^[210,211].

Tumor suppressor pathways: RB1 and TP53

Retinoblastoma-associated protein (RB1) and cellular tumor antigen p53 (TP53) are major tumor suppressors whose functions in PCa have been broadly explored. Loss of RB1 and TP53 is strictly associated with AR misregulation and progression to castration-resistant disease. Both their mechanism and their potential roles in managing PCa are extensively revised in Aparicio *et al.*^[212], Dean *et al.*^[213] and Lee *et al.*^[214].

Ryanodine receptor 2

Ryanodine receptor 2 (RZR2) is expressed in PWR-1E non-tumor cells, as well as in LNCaP and DU145 PCa cells, with the latter registering the lowest expression^[215,216]. RZR2 mobilizes Ca²⁺ from intracellular stores, which is essential to the regulation of apoptosis^[215].

SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily B member 1

SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily B member 1 (SMARCB1) is a core subunit of the SWI/SNF family of nucleosome-remodeling complexes^[217]. In aggressive PCa, the expression of SWI/SNF target genes is impaired by the binding to SchLAP1, which was shown to be aberrantly upregulated^[218].

BIOLOGICAL SIGNIFICANCE OF INTERACTIONS: PPP1 REGULATORS OR SUBSTRATES?

The relationship between PPP1 and the majority of the PIPs here referred, as well as possible alterations in the dynamics of such complexes during prostate carcinogenesis, require further elucidation.

Some of the PIPs are already characterized as PPP1 regulators, substrates, or both (Figure 2). For a significant number of PIPs identified in protein-protein interaction screenings, nevertheless, the functional significance of their interaction with PPP1 remains poorly understood. Therefore, efforts should be made in order to understand PPP1 interaction with CCND1, CCND3, CDC5L, CDC34, CDK4, EED, EIF2AK2, GABARAP, GABARAPL2, GRB2, HDAC8, HDAC10, HSPA8, HEYL, MAX, NCL, IKBKB, IKBKG, MAP1LC3A, MAP1LC3B, MAP3K3, MPHOSPH10, MYC, NCOR1, NOC2L, PAK6, PLCL2, PPP1R3B, PPP1R12A, PPP1R13B, PTEN, RIPK3, RPAP2, RPAP3, RRP1B, RUVBL2, SF3A2, SH2D4A, SKP1, SPRED1, STAM, STAU1, SYTL2, and USF1.

In other cases, the biological significance of the complex is partially known, although it is not established whether the PIP is the regulator or the substrate (or even both). For instance, HDAC6 directly binds to PPP1 and the complex controls microtubule dynamics by maintaining

α -tubulin in a deacetylated state, but the exact mechanism remains poorly understood^[219].

Phosphoprotein phosphatase 1 regulators

AKAP11 acts as a targeting subunit of PPP1 and it can also inhibit the phosphatase activity^[220,221]. BCL2L2 and CUEDC2 target PPP1 to protein complexes: BCL2L2 recruits PPP1 to BAD, forming a complex that is involved in the control of apoptosis^[222], and, CUEDC2 targets PPP1 to IKK, thereby promoting the dephosphorylation and inactivation of the kinase^[223]. NOM1 acts as a PPP1 nucleolar targeting subunit, PPP1R10 targets PPP1 to the nucleus, and PPP1R15A targets PPP1 to the endoplasmic reticulum^[224-227]. PPP1R10/PPP1 holoenzyme is known to regulate chromosome decondensation and apoptosis in response to cellular stresses^[226,228].

PPP1C positive regulators include ATM, GSK3B, and SMARCB1. In response to ionizing radiation, PPP1 is dephosphorylated and activated by ATM^[229]. ATM-mediated activation of PPP1 could occur, at least, *via* two mechanisms: (1) phosphorylation of I-2 and consequent dissociation of the complex I-2/PPP1; or, (2) dephosphorylation of PPP1C at Thr320 to amplify its activity^[230]. As a result, PPP1 dephosphorylates HDAC1, leading to the dissociation of the HDAC1-PPP1-Rb complex^[231,232]. In similar way to ATM, GSK3B activates PPP1 *via* phosphorylation of I-2 and consequent disruption of the I-2/PPP1 complex^[233]. SMARCB1 forms a tricomplex with PPP1R15A and PPP1, and weakly stimulates PPP1 activity^[234].

AKAP11, BRCA1, CDK2, LMTK2, HCFC1, PPP1R7, PPP1R11, and TP53BP2 inhibit the activity of PPP1^[229,235-239].

Phosphoprotein phosphatase 1 substrates

PPP1C is a key regulator of the two major tumor suppressors: it inhibits TP53 and activates RB1 (further details on^[24]). The apoptotic process is strictly controlled by reversible phosphorylation^[240]. The phosphorylation of APAF1 by the 90-kDa ribosomal S6 kinase (RSK) compromises the formation of the apoptosome, impairs cells' sensitivity to cytochrome c, and inhibits apoptosis. PPP1CA was shown to reverse the RSK-mediated phosphorylation of APAF1, thus enhancing its pro-apoptotic activities^[241]. BAD is also dephosphorylated by PPP1CA in a dependent way of the anti-apoptotic members BCL2, BCL2L2, and BCL-XL^[222,242,243]. While BAD overexpression provides proliferative advantage to tumor cells, BAD dephosphorylation increases their sensitivity to apoptosis^[87].

PPP1 exerts a positive control on Wnt signaling through dephosphorylation of AXIN1. As a result, β -catenin destruction complex dissociates, the free phospho- β -catenin accumulates in the cytoplasm, and the transcriptional activity of β -catenin is promoted^[244].

In addition of being regulators, BRCA1 and GSK3B are also substrates for PPP1C. PPP1C dephosphorylates BRCA1 and enhances its DNA repair function^[235,245,246]. GSK3B is also dephosphorylated and disinhibited by PPP1-mediated dephosphorylation^[247].

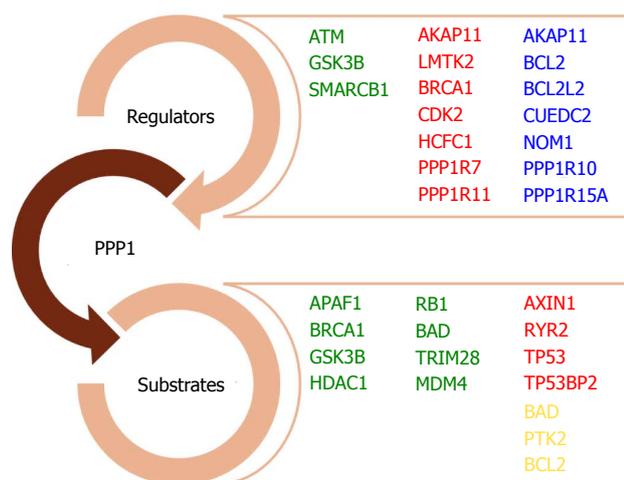


Figure 2 Phosphoprotein phosphatase 1 interacting proteins can be regulators, substrate or both. Green represents positive regulation of phosphoprotein phosphatase 1 (PPP1) (in case of regulators) or PPP1-mediated upregulation of substrates. Red represents negative regulation of PPP1 (in case of regulators) or PPP1-mediated downregulation of substrates. Blue characterizes PPP1-interacting proteins (PIPs) exhibiting PPP1 targeting ability. Yellow corresponds to proteins that are identified as substrates, but whose interaction with PPP1 are not fully understood. ATM: Ataxia telangiectasia mutated; GSK3B: Glycogen synthase kinase-3 β ; SMARCB1: SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily B member 1; BRCA1: Breast cancer type 1 susceptibility protein.

HDAC1 activity is promoted through dephosphorylation of Ser133 by PPP1, which leads to dissociation of HDAC1-PP1-Rb complex, and consequent increase of HDAC1 activity^[232,248]. Similarly, HDAC6 and -8, which can be phosphorylated, might also be PPP1 substrates, although this fact has not been confirmed yet.

RYRs are regulated through reversible phosphorylation, and PPP1 reverts PKA-mediated phosphorylation and activation of RYR2^[249-251]. PPP1 dephosphorylates TRIM28 (Ser824), enhancing its sumoylation state, and MDM4 (Ser367), enhancing its stability and leading to the consequent inhibition of TP53 activity^[252]. PPP1 also reverts the phosphorylation of PTK2 and BCL2^[253-255].

CHALLENGE OF TARGETING PPP1 ACTIVITY

Since PPP1 is a Ser/Thr phosphatase with major roles in several pathological processes, the manipulation of its activity is a valuable therapeutic tool that had been misjudged for years. PPP1 activity could be manipulated through direct or indirect inhibition of the catalytic site (for review see^[27]).

The dissociation of PPP1 complexes through the targeting of PIPs is challenging and might overcome the problems that arise from direct inhibition of PPP1C. However, this area remains understudied and only two complexes are currently being targeted: PPP1C/HDAC and PPP1C/PPP1R15A. Trichostatin A disrupts PPP1C/HDAC and is used in the treatment of glioblastoma and PCa cells. LBH589, an inhibitor of HDAC, was also shown to be

able to dissociate this complex^[256]. As a consequence, AKT is dephosphorylated and its activity decreases. PPP1C/PPP1R15A complex is disrupted upon salubrinal treatment, thereby dephosphorylating eIF2 α ^[257,258].

The increasing number of PPP1 docking motifs identified offers excellent opportunities for targeting specific complexes. In fact, the docking motif found in Bad has inspired the designing of a peptide that interferes with PPP1/BAD complex and is able to induce cell death^[259]. Also, the PPP1 docking motif R/Kx(0,1)V/IxFxxR/KxR/K, a new PPP1C-dependent apoptotic signature, might be a useful tool for drug design^[260].

The disruption or enhancement of several other complexes might contribute to the enhancement of PCa management, enabling more efficient therapies for advanced castration-resistant PCa. Therefore, the identification of PPP1 complexes in human prostate and their characterization in prostate carcinogenesis is imperative for the search of new therapeutic targets.

REFERENCES

- 1 **Jemal A**, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; **61**: 69-90 [PMID: 21296855 DOI: 10.3322/caac.20107]
- 2 **Bubendorf L**, Schöpfer A, Wagner U, Sauter G, Moch H, Willi N, Gasser TC, Mihatsch MJ. Metastatic patterns of prostate cancer: an autopsy study of 1,589 patients. *Hum Pathol* 2000; **31**: 578-583 [PMID: 10836297]
- 3 **Kapoor A**. What's new in prostate cancer research? Highlights of GU-ASCO 2014. *Can Urol Assoc J* 2014; **8**: S8-S12 [PMID: 24860635 DOI: 10.5489/auaj.2013]
- 4 **Felgueiras J**, Silva JV, Fardilha M. Prostate cancer: the need for biomarkers and new therapeutic targets. *J Zhejiang Univ Sci B* 2014; **15**: 16-42 [PMID: 24390742 DOI: 10.1631/jzus. B1300106]
- 5 **Olsen JV**, Vermeulen M, Santamaria A, Kumar C, Miller ML, Jensen LJ, Gnad F, Cox J, Jensen TS, Nigg EA, Brunak S, Mann M. Quantitative phosphoproteomics reveals widespread full phosphorylation site occupancy during mitosis. *Sci Signal* 2010; **3**: ra3 [PMID: 20068231 DOI: 10.1126/scisignal.2000475]
- 6 **Brautigan DL**. Protein Ser/Thr phosphatases--the ugly ducklings of cell signalling. *FEBS J* 2013; **280**: 324-345 [PMID: 22519956 DOI: 10.1111/j.1742-4658.2012.08609.x]
- 7 **Stebbing J**, Lit LC, Zhang H, Darrington RS, Melaiu O, Rudraraju B, Giamas G. The regulatory roles of phosphatases in cancer. *Oncogene* 2014; **33**: 939-953 [PMID: 23503460 DOI: 10.1038/onc.2013.80]
- 8 **Lim YP**. Mining the tumor phosphoproteome for cancer markers. *Clin Cancer Res* 2005; **11**: 3163-3169 [PMID: 15867208 DOI: 10.1158/1078-0432.CCR-04-2243]
- 9 **Chen L**, Giorgianni F, Beranova-Giorgianni S. Characterization of the phosphoproteome in LNCaP prostate cancer cells by in-gel isoelectric focusing and tandem mass spectrometry. *J Proteome Res* 2010; **9**: 174-178 [PMID: 20044836 DOI: 10.1021/pr900338q]
- 10 **Lescarbeau R**, Kaplan DL. Correlating phosphoproteomic signaling with castration resistant prostate cancer survival through regression analysis. *Mol Biosyst* 2014; **10**: 605-612 [PMID: 24413303 DOI: 10.1039/c3mb70403c]
- 11 **Lescarbeau RM**, Kaplan DL. Quantitative analysis of castration resistant prostate cancer progression through phosphoproteome signaling. *BMC Cancer* 2014; **14**: 325 [PMID: 24885093 DOI: 10.1186/1471-2407-14-325]
- 12 **Giorgianni F**, Zhao Y, Desiderio DM, Beranova-Giorgianni

- S. Toward a global characterization of the phosphoproteome in prostate cancer cells: identification of phosphoproteins in the LNCaP cell line. *Electrophoresis* 2007; **28**: 2027-2034 [PMID: 17487921 DOI: 10.1002/elps.200600782]
- 13 **van der Steen T**, Tindall DJ, Huang H. Posttranslational modification of the androgen receptor in prostate cancer. *Int J Mol Sci* 2013; **14**: 14833-14859 [PMID: 23863692 DOI: 10.3390/ijms140714833]
- 14 **Moorhead GB**, Trinkle-Mulcahy L, Ulke-Lemée A. Emerging roles of nuclear protein phosphatases. *Nat Rev Mol Cell Biol* 2007; **8**: 234-244 [PMID: 17318227 DOI: 10.1038/nrm2126]
- 15 **Ceulemans H**, Bollen M. Functional diversity of protein phosphatase-1, a cellular economizer and reset button. *Physiol Rev* 2004; **84**: 1-39 [PMID: 14715909 DOI: 10.1152/physrev.00013.2003]
- 16 **Bollen M**. Combinatorial control of protein phosphatase-1. *Trends Biochem Sci* 2001; **26**: 426-431 [PMID: 11440854]
- 17 **Cohen PT**. Protein phosphatase 1--targeted in many directions. *J Cell Sci* 2002; **115**: 241-256 [PMID: 11839776]
- 18 **Flores-Delgado G**, Liu CW, Sposto R, Berndt N. A limited screen for protein interactions reveals new roles for protein phosphatase 1 in cell cycle control and apoptosis. *J Proteome Res* 2007; **6**: 1165-1175 [PMID: 17274640 DOI: 10.1021/pr060504h]
- 19 **Bollen M**, Peti W, Ragusa MJ, Beullens M. The extended PP1 toolkit: designed to create specificity. *Trends Biochem Sci* 2010; **35**: 450-458 [PMID: 20399103 DOI: 10.1016/j.tibs.2010.03.002]
- 20 **Fardilha M**, Esteves SL, Korrodi-Gregório L, Vintém AP, Domingues SC, Rebelo S, Morrice N, Cohen PT, da Cruz e Silva OA, da Cruz e Silva EF. Identification of the human testis protein phosphatase 1 interactome. *Biochem Pharmacol* 2011; **82**: 1403-1415 [PMID: 21382349 DOI: 10.1016/j.bcp.2011.02.018]
- 21 **Esteves SL**, Domingues SC, da Cruz e Silva OA, Fardilha M, da Cruz e Silva EF. Protein phosphatase 1 α interacting proteins in the human brain. *OMICS* 2012; **16**: 3-17 [PMID: 22321011 DOI: 10.1089/omi.2011.0041]
- 22 **Fardilha M**, Esteves SL, Korrodi-Gregório L, Pelech S, da Cruz E Silva OA, da Cruz E Silva E. Protein phosphatase 1 complexes modulate sperm motility and present novel targets for male infertility. *Mol Hum Reprod* 2011; **17**: 466-477 [PMID: 21257602 DOI: 10.1093/molehr/gar004]
- 23 **Esteves SL**, Korrodi-Gregório L, Cotrim CZ, van Kleeff PJ, Domingues SC, da Cruz e Silva OA, Fardilha M, da Cruz e Silva EF. Protein phosphatase 1 γ isoforms linked interactions in the brain. *J Mol Neurosci* 2013; **50**: 179-197 [PMID: 23080069 DOI: 10.1007/s12031-012-9902-6]
- 24 **Figueiredo J**, da Cruz E Silva OA, Fardilha M. Protein phosphatase 1 and its complexes in carcinogenesis. *Curr Cancer Drug Targets* 2014; **14**: 2-29 [PMID: 24200083]
- 25 **Zhang J**, Yang PL, Gray NS. Targeting cancer with small molecule kinase inhibitors. *Nat Rev Cancer* 2009; **9**: 28-39 [PMID: 19104514 DOI: 10.1038/nrc2559]
- 26 **McConnell JL**, Wadzinski BE. Targeting protein serine/threonine phosphatases for drug development. *Mol Pharmacol* 2009; **75**: 1249-1261 [PMID: 19299564 DOI: 10.1124/mol.108.053140]
- 27 **Chatterjee J**, Köhn M. Targeting the untargetable: recent advances in the selective chemical modulation of protein phosphatase-1 activity. *Curr Opin Chem Biol* 2013; **17**: 361-368 [PMID: 23647984 DOI: 10.1016/j.cbpa.2013.04.008]
- 28 **Sangrador A**, Andrés I, Eguiraun A, Lorenzo ML, Ortiz JM. Growth arrest of *Schizosaccharomyces pombe* following overexpression of mouse type 1 protein phosphatases. *Mol Gen Genet* 1998; **259**: 449-456 [PMID: 9790575]
- 29 **Ceulemans H**, Stalmans W, Bollen M. Regulator-driven functional diversification of protein phosphatase-1 in eukaryotic evolution. *Bioessays* 2002; **24**: 371-381 [PMID: 11948623 DOI: 10.1002/bies.10069]
- 30 **Shi Y**. Serine/threonine phosphatases: mechanism through structure. *Cell* 2009; **139**: 468-484 [PMID: 19879837 DOI: 10.1016/j.cell.2009.10.006]
- 31 **Xie X**, Huang W, Xue C, Wei Q. The nonconserved N-terminus of protein phosphatases 1 influences its active site. *BMB Rep* 2008; **41**: 881-885 [PMID: 19123980]
- 32 **Xie XJ**, Huang W, Xue CZ, Wei Q. The N-terminal domain influences the structure and property of protein phosphatase 1. *Mol Cell Biochem* 2009; **327**: 241-246 [PMID: 19242655 DOI: 10.1007/s11010-009-0062-0]
- 33 **Egloff MP**, Johnson DF, Moorhead G, Cohen PT, Cohen P, Barford D. Structural basis for the recognition of regulatory subunits by the catalytic subunit of protein phosphatase 1. *EMBO J* 1997; **16**: 1876-1887 [PMID: 9155014 DOI: 10.1093/emboj/16.8.1876]
- 34 **Hendrickx A**, Beullens M, Ceulemans H, Den Abt T, Van Eynde A, Nicolaescu E, Lesage B, Bollen M. Docking motif-guided mapping of the interactome of protein phosphatase-1. *Chem Biol* 2009; **16**: 365-371 [PMID: 19389623 DOI: 10.1016/j.chembiol.2009.02.012]
- 35 **Meiselbach H**, Sticht H, Enz R. Structural analysis of the protein phosphatase 1 docking motif: molecular description of binding specificities identifies interacting proteins. *Chem Biol* 2006; **13**: 49-59 [PMID: 16426971 DOI: 10.1016/j.chembiol.2005.10.009]
- 36 **Radivojac P**, Baenziger PH, Kann MG, Mort ME, Hahn MW, Mooney SD. Gain and loss of phosphorylation sites in human cancer. *Bioinformatics* 2008; **24**: i241-i247 [PMID: 18689832 DOI: 10.1093/bioinformatics/btn267]
- 37 **Bellacosa A**, Kumar CC, Di Cristofano A, Testa JR. Activation of AKT kinases in cancer: implications for therapeutic targeting. *Adv Cancer Res* 2005; **94**: 29-86 [PMID: 16095999 DOI: 10.1016/S0065-230X(05)94002-5]
- 38 **Korrodi-Gregório L**, Silva JV, Santos-Sousa L, Freitas MJ, Felgueiras J, Fardilha M. TGF- β cascade regulation by PPP1 and its interactors -impact on prostate cancer development and therapy. *J Cell Mol Med* 2014; **18**: 555-567 [PMID: 24629090 DOI: 10.1111/jcmm.12266]
- 39 **Hall EH**, Daugherty AE, Choi CK, Horwitz AF, Brautigan DL. Tensin1 requires protein phosphatase-1 α in addition to RhoGAP DLC-1 to control cell polarization, migration, and invasion. *J Biol Chem* 2009; **284**: 34713-34722 [PMID: 19826001 DOI: 10.1074/jbc.M109.059592]
- 40 **Nagasaka K**, Seiki T, Yamashita A, Massimi P, Subbaiah VK, Thomas M, Kranjec C, Kawana K, Nakagawa S, Yano T, Taketani Y, Fujii T, Kozuma S, Banks L. A novel interaction between hScrib and PP1 γ downregulates ERK signaling and suppresses oncogene-induced cell transformation. *PLoS One* 2013; **8**: e53752 [PMID: 23359326 DOI: 10.1371/journal.pone.0053752]
- 41 **Walsh JE**, Young MR. TGF-beta regulation of focal adhesion proteins and motility of premalignant oral lesions via protein phosphatase 1. *Anticancer Res* 2011; **31**: 3159-3164 [PMID: 21965722]
- 42 **Grossmann M**, Cheung AS, Zajac JD. Androgens and prostate cancer; pathogenesis and deprivation therapy. *Best Pract Res Clin Endocrinol Metab* 2013; **27**: 603-616 [PMID: 24054933 DOI: 10.1016/j.beem.2013.05.001]
- 43 **Wen S**, Niu Y, Lee SO, Chang C. Androgen receptor (AR) positive vs negative roles in prostate cancer cell deaths including apoptosis, anoikis, entosis, necrosis and autophagic cell death. *Cancer Treat Rev* 2014; **40**: 31-40 [PMID: 23993415 DOI: 10.1016/j.ctrv.2013.07.008]
- 44 **Chen S**, Kesler CT, Paschal BM, Balk SP. Androgen receptor phosphorylation and activity are regulated by an association with protein phosphatase 1. *J Biol Chem* 2009; **284**: 25576-25584 [PMID: 19622840 DOI: 10.1074/jbc.M109.043133]
- 45 **Gioeli D**, Black BE, Gordon V, Spencer A, Kesler CT, Eblen ST, Paschal BM, Weber MJ. Stress kinase signaling regulates androgen receptor phosphorylation, transcription, and localization. *Mol Endocrinol* 2006; **20**: 503-515 [PMID: 16282370]

- DOI: 10.1210/me.2005-0351]
- 46 **Beullens M**, Van Eynde A, Stalmans W, Bollen M. The isolation of novel inhibitory polypeptides of protein phosphatase 1 from bovine thymus nuclei. *J Biol Chem* 1992; **267**: 16538-16544 [PMID: 1322907]
 - 47 **Martin-Granados C**, Prescott AR, Van Dessel N, Van Eynde A, Arocena M, Klaska IP, Görnemann J, Beullens M, Bollen M, Forrester JV, McCaig CD. A role for PP1/NIPPI in steering migration of human cancer cells. *PLoS One* 2012; **7**: e40769 [PMID: 22815811 DOI: 10.1371/journal.pone.0040769]
 - 48 **Allard P**, Zoubeidi A, Nguyen LT, Tessier S, Tanguay S, Chevrette M, Aprikian A, Chevalier S. Links between Fer tyrosine kinase expression levels and prostate cell proliferation. *Mol Cell Endocrinol* 2000; **159**: 63-77 [PMID: 10687853 DOI: 10.1016/S0303-7207(99)00205-1]
 - 49 **Zoubeidi A**, Rocha J, Zouanat FZ, Hamel L, Scarlata E, Aprikian AG, Chevalier S. The Fer tyrosine kinase cooperates with interleukin-6 to activate signal transducer and activator of transcription 3 and promote human prostate cancer cell growth. *Mol Cancer Res* 2009; **7**: 142-155 [PMID: 19147545 DOI: 10.1158/1541-7786.MCR-08-0117]
 - 50 **Rocha J**, Zouanat FZ, Zoubeidi A, Hamel L, Benidir T, Scarlata E, Brimo F, Aprikian A, Chevalier S. The Fer tyrosine kinase acts as a downstream interleukin-6 effector of androgen receptor activation in prostate cancer. *Mol Cell Endocrinol* 2013; **381**: 140-149 [PMID: 23906537 DOI: 10.1016/j.mce.2013.07.017]
 - 51 **Pasder O**, Shpungin S, Salem Y, Makovsky A, Vilchick S, Michaeli S, Malovani H, Nir U. Downregulation of Fer induces PP1 activation and cell-cycle arrest in malignant cells. *Oncogene* 2006; **25**: 4194-4206 [PMID: 16732323 DOI: 10.1038/sj.onc.1209695]
 - 52 **Yang G**, Truong LD, Timme TL, Ren C, Wheeler TM, Park SH, Nasu Y, Bangma CH, Kattan MW, Scardino PT, Thompson TC. Elevated expression of caveolin is associated with prostate and breast cancer. *Clin Cancer Res* 1998; **4**: 1873-1880 [PMID: 9717814]
 - 53 **Yang G**, Truong LD, Wheeler TM, Thompson TC. Caveolin-1 expression in clinically confined human prostate cancer: a novel prognostic marker. *Cancer Res* 1999; **59**: 5719-5723 [PMID: 10582690]
 - 54 **Karam JA**, Lotan Y, Roehrborn CG, Ashfaq R, Karakiewicz PI, Shariat SF. Caveolin-1 overexpression is associated with aggressive prostate cancer recurrence. *Prostate* 2007; **67**: 614-622 [PMID: 17299799 DOI: 10.1002/Pros.20557]
 - 55 **Thompson TC**, Tahir SA, Li L, Watanabe M, Naruishi K, Yang G, Kadmon D, Logothetis CJ, Troncso P, Ren C, Goltsov A, Park S. The role of caveolin-1 in prostate cancer: clinical implications. *Prostate Cancer Prostatic Dis* 2010; **13**: 6-11 [PMID: 19581923 DOI: 10.1038/pcan.2009.29]
 - 56 **Tahir SA**, Kurosaka S, Tanimoto R, Goltsov AA, Park S, Thompson TC. Serum caveolin-1, a biomarker of drug response and therapeutic target in prostate cancer models. *Cancer Biol Ther* 2013; **14**: 117-126 [PMID: 23114714 DOI: 10.4161/cbt.22633]
 - 57 **Li L**, Ren CH, Tahir SA, Ren C, Thompson TC. Caveolin-1 maintains activated Akt in prostate cancer cells through scaffolding domain binding site interactions with and inhibition of serine/threonine protein phosphatases PP1 and PP2A. *Mol Cell Biol* 2003; **23**: 9389-9404 [PMID: 14645548 DOI: 10.1128/Mcb.23.24.9389-9404.2003]
 - 58 **Barshir R**, Basha O, Eluk A, Smoly IY, Lan A, Yeger-Lotem E. The TissueNet database of human tissue protein-protein interactions. *Nucleic Acids Res* 2013; **41**: D841-D844 [PMID: 23193266 DOI: 10.1093/nar/gks1198]
 - 59 **Schaefer MH**, Fontaine JF, Vinayagam A, Porras P, Wanker EE, Andrade-Navarro MA. HIPPIE: Integrating protein interaction networks with experiment based quality scores. *PLoS One* 2012; **7**: e31826 [PMID: 22348130 DOI: 10.1371/journal.pone.0031826]
 - 60 **Wu C**, Orozco C, Boyer J, Leglise M, Goodale J, Batalov S, Hodge CL, Haase J, Janes J, Huss JW, Su AI. BioGPS: an extensible and customizable portal for querying and organizing gene annotation resources. *Genome Biol* 2009; **10**: R130 [PMID: 19919682 DOI: 10.1186/gb-2009-10-11-r130]
 - 61 **Zou H**, Li Y, Liu X, Wang X. An APAF-1-cytochrome c multimeric complex is a functional apoptosome that activates procaspase-9. *J Biol Chem* 1999; **274**: 11549-11556 [PMID: 10206961 DOI: 10.1074/jbc.274.17.11549]
 - 62 **Ogawa T**, Shiga K, Hashimoto S, Kobayashi T, Horii A, Furukawa T. APAF-1-ALT, a novel alternative splicing form of APAF-1, potentially causes impeded ability of undergoing DNA damage-induced apoptosis in the LNCaP human prostate cancer cell line. *Biochem Biophys Res Commun* 2003; **306**: 537-543 [PMID: 12804598]
 - 63 **Choi S**, Lew KL, Xiao H, Herman-Antosiewicz A, Xiao D, Brown CK, Singh SV. D,L-Sulforaphane-induced cell death in human prostate cancer cells is regulated by inhibitor of apoptosis family proteins and Apaf-1. *Carcinogenesis* 2007; **28**: 151-162 [PMID: 16920735 DOI: 10.1093/carcin/bgl144]
 - 64 **Narayanan BA**, Narayanan NK, Re GG, Nixon DW. Differential expression of genes induced by resveratrol in LNCaP cells: P53-mediated molecular targets. *Int J Cancer* 2003; **104**: 204-212 [PMID: 12569576 DOI: 10.1002/ijc.10932]
 - 65 **Guzey M**, Luo J, Getzenberg RH. Vitamin D3 modulated gene expression patterns in human primary normal and cancer prostate cells. *J Cell Biochem* 2004; **93**: 271-285 [PMID: 15368355 DOI: 10.1002/jcb.20182]
 - 66 **Burek M**, Maddika S, Burek CJ, Daniel PT, Schulze-Osthoff K, Los M. Apoptin-induced cell death is modulated by Bcl-2 family members and is Apaf-1 dependent. *Oncogene* 2006; **25**: 2213-2222 [PMID: 16288204 DOI: 10.1038/sj.onc.1209258]
 - 67 **Rotman G**, Shiloh Y. ATM: a mediator of multiple responses to genotoxic stress. *Oncogene* 1999; **18**: 6135-6144 [PMID: 10557105 DOI: 10.1038/sj.onc.1203124]
 - 68 **Fan C**, Quan R, Feng X, Gillis A, He L, Matsumoto ED, Salama S, Cutz JC, Kapoor A, Tang D. ATM activation is accompanied with earlier stages of prostate tumorigenesis. *Biochim Biophys Acta* 2006; **1763**: 1090-1097 [PMID: 16997395 DOI: 10.1016/j.bbamer.2006.08.026]
 - 69 **Angèle S**, Falconer A, Foster CS, Taniere P, Eeles RA, Hall J. ATM protein overexpression in prostate tumors: possible role in telomere maintenance. *Am J Clin Pathol* 2004; **121**: 231-236 [PMID: 14983937 DOI: 10.1309/JTKG-GGKU-RFX3-XMGT]
 - 70 **Angèle S**, Falconer A, Edwards SM, Dörk T, Bremer M, Moullan N, Chapot B, Muir K, Houlston R, Norman AR, Bullock S, Hope Q, Meitz J, Dearnaley D, Dowe A, Southgate C, Ardern-Jones A, Easton DF, Eeles RA, Hall J. ATM polymorphisms as risk factors for prostate cancer development. *Br J Cancer* 2004; **91**: 783-787 [PMID: 15280931 DOI: 10.1038/sj.bjc.6602007]
 - 71 **Meyer A**, Wilhelm B, Dörk T, Bremer M, Baumann R, Karstens JH, Machtens S. ATM missense variant P1054R predisposes to prostate cancer. *Radiother Oncol* 2007; **83**: 283-288 [PMID: 17502119 DOI: 10.1016/j.radonc.2007.04.029]
 - 72 **Cesaretti JA**, Stock RG, Lehrers S, Atencio DA, Bernstein JL, Stone NN, Wallenstein S, Green S, Loeb K, Kollmeier M, Smith M, Rosenstein BS. ATM sequence variants are predictive of adverse radiotherapy response among patients treated for prostate cancer. *Int J Radiat Oncol Biol Phys* 2005; **61**: 196-202 [PMID: 15629612 DOI: 10.1016/j.ijrobp.2004.09.031]
 - 73 **Farooqi AA**, Fayyaz S, Rashid S. Upon the tightrope in prostate cancer: two acrobats on the same tightrope to cross the finishline. *Mol Cell Biochem* 2012; **364**: 53-57 [PMID: 22200977 DOI: 10.1007/s11010-011-1204-8]
 - 74 **Fan Z**, Chakravarty P, Alfieri A, Pandita TK, Vikram B, Guha C. Adenovirus-mediated antisense ATM gene transfer sensitizes prostate cancer cells to radiation. *Cancer Gene Ther* 2000; **7**: 1307-1314 [PMID: 11059687 DOI: 10.1038/

- sj.cgt.7700242]
- 75 **Collis SJ**, Swartz MJ, Nelson WG, DeWeese TL. Enhanced radiation and chemotherapy-mediated cell killing of human cancer cells by small inhibitory RNA silencing of DNA repair factors. *Cancer Res* 2003; **63**: 1550-1554 [PMID: 12670903]
 - 76 **Truman JP**, Gueven N, Lavin M, Leibel S, Kolesnick R, Fuks Z, Haimovitz-Friedman A. Down-regulation of ATM protein sensitizes human prostate cancer cells to radiation-induced apoptosis. *J Biol Chem* 2005; **280**: 23262-23272 [PMID: 15837784 DOI: 10.1074/jbc.M503701200]
 - 77 **Shaheen FS**, Znojek P, Fisher A, Webster M, Plummer R, Gaughan L, Smith GC, Leung HY, Curtin NJ, Robson CN. Targeting the DNA double strand break repair machinery in prostate cancer. *PLoS One* 2011; **6**: e20311 [PMID: 21629734 DOI: 10.1371/journal.pone.0020311]
 - 78 **Mukhopadhyay UK**, Senderowicz AM, Ferbeyre G. RNA silencing of checkpoint regulators sensitizes p53-defective prostate cancer cells to chemotherapy while sparing normal cells. *Cancer Res* 2005; **65**: 2872-2881 [PMID: 15805289 DOI: 10.1158/0008-5472.CAN-04-2502]
 - 79 **Luo W**, Lin SC. Axin: a master scaffold for multiple signaling pathways. *Neurosignals* 2004; **13**: 99-113 [PMID: 15067197 DOI: 10.1159/000076563]
 - 80 **Yardy GW**, Bicknell DC, Wilding JL, Bartlett S, Liu Y, Winney B, Turner GD, Brewster SF, Bodmer WF. Mutations in the AXIN1 gene in advanced prostate cancer. *Eur Urol* 2009; **56**: 486-494 [PMID: 18514389 DOI: 10.1016/j.eururo.2008.05.029]
 - 81 **Ola MS**, Nawaz M, Ahsan H. Role of Bcl-2 family proteins and caspases in the regulation of apoptosis. *Mol Cell Biochem* 2011; **351**: 41-58 [PMID: 21210296 DOI: 10.1007/s11010-010-0709-x]
 - 82 **Catz SD**, Johnson JL. BCL-2 in prostate cancer: a minireview. *Apoptosis* 2003; **8**: 29-37 [PMID: 12510149]
 - 83 **Rosser CJ**, Reyes AO, Vakar-Lopez F, Levy LB, Kuban DA, Hoover DC, Lee AK, Pisters LL. Bcl-2 is significantly overexpressed in localized radio-recurrent prostate carcinoma, compared with localized radio-naïve prostate carcinoma. *Int J Radiat Oncol Biol Phys* 2003; **56**: 1-6 [PMID: 12694817]
 - 84 **Lin Y**, Fukuchi J, Hiipakka RA, Kokontis JM, Xiang J. Up-regulation of Bcl-2 is required for the progression of prostate cancer cells from an androgen-dependent to an androgen-independent growth stage. *Cell Res* 2007; **17**: 531-536 [PMID: 17404601 DOI: 10.1038/cr.2007.12]
 - 85 **Bruckheimer EM**, Kyprianou N. Bcl-2 antagonizes the combined apoptotic effect of transforming growth factor-beta and dihydrotestosterone in prostate cancer cells. *Prostate* 2002; **53**: 133-142 [PMID: 12242728 DOI: 10.1002/pros.10143]
 - 86 **Royuela M**, Arenas MI, Bethencourt FR, Sánchez-Chapado M, Fraile B, Paniagua R. Immunoections of p21, Rb, mcl-1 and bad gene products in normal, hyperplastic and carcinomatous human prostates. *Eur Cytokine Netw* 2001; **12**: 654-663 [PMID: 11781193]
 - 87 **Smith AJ**, Karpova Y, D'Agostino R, Willingham M, Kulik G. Expression of the Bcl-2 protein BAD promotes prostate cancer growth. *PLoS One* 2009; **4**: e6224 [PMID: 19593445 DOI: 10.1371/journal.pone.0006224]
 - 88 **Taghiyev AF**, Guseva NV, Harada H, Knudson CM, Rokhlin OW, Cohen MB. Overexpression of BAD potentiates sensitivity to tumor necrosis factor-related apoptosis-inducing ligand treatment in the prostatic carcinoma cell line LNCaP. *Mol Cancer Res* 2003; **1**: 500-507 [PMID: 12754297]
 - 89 **Zhang Y**, Yu J, Unni E, Shao TC, Nan B, Snabbon T, Kasper S, Andriani F, Denner L, Marcelli M. Monogene and polygene therapy for the treatment of experimental prostate cancers by use of apoptotic genes bax and bad driven by the prostate-specific promoter ARR(2)PB. *Hum Gene Ther* 2002; **13**: 2051-2064 [PMID: 12490000 DOI: 10.1089/10430340260395901]
 - 90 **Kajiwara T**, Takeuchi T, Ueki T, Moriyama N, Ueki K, Kakizoe T, Kawabe K. Effect of Bcl-2 overexpression in human prostate cancer cells in vitro and in vivo. *Int J Urol* 1999; **6**: 520-525 [PMID: 10533903]
 - 91 **Tolcher AW**. Preliminary phase I results of G3139 (bcl-2 antisense oligonucleotide) therapy in combination with docetaxel in hormone-refractory prostate cancer. *Semin Oncol* 2001; **28**: 67-70 [PMID: 11685732]
 - 92 **Gleave ME**, Miayake H, Goldie J, Nelson C, Tolcher A. Targeting bcl-2 gene to delay androgen-independent progression and enhance chemosensitivity in prostate cancer using antisense bcl-2 oligodeoxynucleotides. *Urology* 1999; **54**: 36-46 [PMID: 10606283]
 - 93 **Chi KN**. Targeting Bcl-2 with oblimersen for patients with hormone refractory prostate cancer. *World J Urol* 2005; **23**: 33-37 [PMID: 15723221 DOI: 10.1007/s00345-004-0477-x]
 - 94 **Bhatnagar N**, Li X, Padi SK, Zhang Q, Tang MS, Guo B. Downregulation of miR-205 and miR-31 confers resistance to chemotherapy-induced apoptosis in prostate cancer cells. *Cell Death Dis* 2010; **1**: e105 [PMID: 21368878 DOI: 10.1038/cddis.2010.85]
 - 95 **Gapter L**, Wang Z, Glinski J, Ng KY. Induction of apoptosis in prostate cancer cells by pachymic acid from *Poria cocos*. *Biochem Biophys Res Commun* 2005; **332**: 1153-1161 [PMID: 15913545 DOI: 10.1016/j.bbrc.2005.05.044]
 - 96 **Sambantham S**, Radha M, Paramasivam A, Anandan B, Malathi R, Chandra SR, Jayaraman G. Molecular mechanism underlying hesperetin-induced apoptosis by in silico analysis and in prostate cancer PC-3 cells. *Asian Pac J Cancer Prev* 2013; **14**: 4347-4352 [PMID: 23992001]
 - 97 **Cho YS**, Cho-Chung YS. Antisense protein kinase A RI-alpha acts synergistically with hydroxycamptothecin to inhibit growth and induce apoptosis in human cancer cells: molecular basis for combinatorial therapy. *Clin Cancer Res* 2003; **9**: 1171-1178 [PMID: 12631623]
 - 98 **Wang Q**, Zhang H, Fishel R, Greene MI. BRCA1 and cell signaling. *Oncogene* 2000; **19**: 6152-6158 [PMID: 11156529 DOI: 10.1038/sj.onc.1203974]
 - 99 **Fachal L**, Gómez-Caamaño A, Celeiro-Muñoz C, Peleteiro P, Blanco A, Carballo A, Forteza J, Carracedo A, Vega A. BRCA1 mutations do not increase prostate cancer risk: results from a meta-analysis including new data. *Prostate* 2011; **71**: 1768-1779 [PMID: 21520156 DOI: 10.1002/pros.21394]
 - 100 **Castro E**, Goh C, Olmos D, Saunders R, Leongamornlert D, Tymrakiewicz M, Mahmud N, Dadaev T, Govindasami K, Guy M, Sawyer E, Wilkinson R, Ardern-Jones A, Ellis S, Frost D, Peock S, Evans DG, Tischkowitz M, Cole T, Davidson R, Eccles D, Brewer C, Douglas F, Porteous ME, Donaldson A, Dorkins H, Izatt L, Cook J, Hodgson S, Kennedy MJ, Side LE, Eason J, Murray A, Antoniou AC, Easton DF, Kote-Jarai Z, Eeles R. Germline BRCA mutations are associated with higher risk of nodal involvement, distant metastasis, and poor survival outcomes in prostate cancer. *J Clin Oncol* 2013; **31**: 1748-1757 [PMID: 23569316 DOI: 10.1200/JCO.2012.43.1882]
 - 101 **Douglas JA**, Levin AM, Zuhlke KA, Ray AM, Johnson GR, Lange EM, Wood DP, Cooney KA. Common variation in the BRCA1 gene and prostate cancer risk. *Cancer Epidemiol Biomarkers Prev* 2007; **16**: 1510-1516 [PMID: 17585057 DOI: 10.1158/1055-9965.EPI-07-0137]
 - 102 **Sundararajan S**, Ahmed A, Goodman OB. The relevance of BRCA genetics to prostate cancer pathogenesis and treatment. *Clin Adv Hematol Oncol* 2011; **9**: 748-755 [PMID: 22252577]
 - 103 **Rabiau N**, Déchelotte P, Adjakly M, Kemeny JL, Guy L, Boiteux JP, Kwiatkowski F, Bignon YJ, Bernard-Gallon D. BRCA1, BRCA2, AR and IGF-I expression in prostate cancer: correlation between RT-qPCR and immunohistochemical detection. *Oncol Rep* 2011; **26**: 695-702 [PMID: 21667031 DOI: 10.3892/or.2011.1339]
 - 104 **Schayek H**, Haugk K, Sun S, True LD, Plymate SR, Werner H. Tumor suppressor BRCA1 is expressed in prostate cancer and controls insulin-like growth factor I receptor (IGF-IR) gene transcription in an androgen receptor-dependent man-

- ner. *Clin Cancer Res* 2009; **15**: 1558-1565 [PMID: 19223505 DOI: 10.1158/1078-0432.ccr-08-1440]
- 105 **Fiorentino M**, Judson G, Penney K, Flavin R, Stark J, Fiore C, Fall K, Martin N, Ma J, Sinnott J, Giovannucci E, Stampfer M, Sesso HD, Kantoff PW, Finn S, Loda M, Mucci L. Immunohistochemical expression of BRCA1 and lethal prostate cancer. *Cancer Res* 2010; **70**: 3136-3139 [PMID: 20388772 DOI: 10.1158/0008-5472.CAN-09-4100]
- 106 **De Luca P**, Vazquez ES, Moiola CP, Zalazar F, Cotignola J, Gueron G, Gardner K, De Siervi A. BRCA1 loss induces GADD153-mediated doxorubicin resistance in prostate cancer. *Mol Cancer Res* 2011; **9**: 1078-1090 [PMID: 21700680 DOI: 10.1158/1541-7786.MCR-11-0155]
- 107 **Sandhu SK**, Schelman WR, Wilding G, Moreno V, Baird RD, Miranda S, Hylands L, Riisnaes R, Forster M, Omlin A, Kreischer N, Thway K, Gevensleben H, Sun L, Loughney J, Chatterjee M, Toniatti C, Carpenter CL, Iannone R, Kaye SB, de Bono JS, Wenham RM. The poly(ADP-ribose) polymerase inhibitor niraparib (MK4827) in BRCA mutation carriers and patients with sporadic cancer: a phase 1 dose-escalation trial. *Lancet Oncol* 2013; **14**: 882-892 [PMID: 23810788 DOI: 10.1016/S1470-2045(13)70240-7]
- 108 Olaparib shows promise in multiple tumor types. *Cancer Discov* 2013; **3**: OF5 [PMID: 23847380 DOI: 10.1158/2159-8290.CD-NB2013-082]
- 109 **Comstock CE**, Revelo MP, Buncher CR, Knudsen KE. Impact of differential cyclin D1 expression and localisation in prostate cancer. *Br J Cancer* 2007; **96**: 970-979 [PMID: 17375037 DOI: 10.1038/sj.bjc.6603615]
- 110 **Nikoleishvili D**, Pertia A, Trsintsadze O, Gogokhia N, Managadze L, Chkhotua A. Expression of p27((Kip1)), cyclin D3 and Ki67 in BPH, prostate cancer and hormone-treated prostate cancer cells. *Int Urol Nephrol* 2008; **40**: 953-959 [PMID: 18317945 DOI: 10.1007/s11255-008-9350-y]
- 111 **Schiewer MJ**, Morey LM, Burd CJ, Liu Y, Merry DE, Ho SM, Knudsen KE. Cyclin D1 repressor domain mediates proliferation and survival in prostate cancer. *Oncogene* 2009; **28**: 1016-1027 [PMID: 19079343 DOI: 10.1038/onc.2008.446]
- 112 **Ju X**, Casimiro MC, Gormley M, Meng H, Jiao X, Katiyar S, Crosariol M, Chen K, Wang M, Quong AA, Lisanti MP, Ertel A, Pestell RG. Identification of a cyclin D1 network in prostate cancer that antagonizes epithelial-mesenchymal restraint. *Cancer Res* 2014; **74**: 508-519 [PMID: 24282282 DOI: 10.1158/0008-5472.CAN-13-1313]
- 113 **Comstock CE**, Augello MA, Schiewer MJ, Karch J, Burd CJ, Ertel A, Knudsen ES, Jessen WJ, Aronow BJ, Knudsen KE. Cyclin D1 is a selective modifier of androgen-dependent signaling and androgen receptor function. *J Biol Chem* 2011; **286**: 8117-8127 [PMID: 21212260 DOI: 10.1074/jbc.M110.170720]
- 114 **Olshavsky NA**, Groh EM, Comstock CE, Morey LM, Wang Y, Revelo MP, Burd C, Meller J, Knudsen KE. Cyclin D3 action in androgen receptor regulation and prostate cancer. *Oncogene* 2008; **27**: 3111-3121 [PMID: 18084330 DOI: 10.1038/sj.onc.1210981]
- 115 **Chung LC**, Tsui KH, Feng TH, Lee SL, Chang PL, Juang HH. L-Mimosine blocks cell proliferation via upregulation of B-cell translocation gene 2 and N-myc downstream regulated gene 1 in prostate carcinoma cells. *Am J Physiol Cell Physiol* 2012; **302**: C676-C685 [PMID: 22116304 DOI: 10.1152/ajpcell.00180.2011]
- 116 **Ouyang DY**, Zeng LH, Pan H, Xu LH, Wang Y, Liu KP, He XH. Piperine inhibits the proliferation of human prostate cancer cells via induction of cell cycle arrest and autophagy. *Food Chem Toxicol* 2013; **60**: 424-430 [PMID: 23939040 DOI: 10.1016/j.fct.2013.08.007]
- 117 **Wu Z**, He B, He J, Mao X. Upregulation of miR-153 promotes cell proliferation via downregulation of the PTEN tumor suppressor gene in human prostate cancer. *Prostate* 2013; **73**: 596-604 [PMID: 23060044 DOI: 10.1002/pros.22600]
- 118 **Noonan EJ**, Place RF, Basak S, Pookot D, Li LC. miR-449a causes Rb-dependent cell cycle arrest and senescence in prostate cancer cells. *Oncotarget* 2010; **1**: 349-358 [PMID: 20948989]
- 119 **Gregory CW**, Johnson RT, Presnell SC, Mohler JL, French FS. Androgen receptor regulation of G1 cyclin and cyclin-dependent kinase function in the CWR22 human prostate cancer xenograft. *J Androl* 2001; **22**: 537-548 [PMID: 11451350]
- 120 **Lu S**, Tsai SY, Tsai MJ. Regulation of androgen-dependent prostatic cancer cell growth: androgen regulation of CDK2, CDK4, and CKI p16 genes. *Cancer Res* 1997; **57**: 4511-4516 [PMID: 9377562]
- 121 **Flores O**, Wang Z, Knudsen KE, Burnstein KL. Nuclear targeting of cyclin-dependent kinase 2 reveals essential roles of cyclin-dependent kinase 2 localization and cyclin E in vitamin D-mediated growth inhibition. *Endocrinology* 2010; **151**: 896-908 [PMID: 20147522 DOI: 10.1210/en.2009-1116]
- 122 **Cipriano SC**, Chen YQ. Insensitivity to growth inhibition by TGF-beta1 correlates with a lack of inhibition of the CDK2 activity in prostate carcinoma cells. *Oncogene* 1998; **17**: 1549-1556 [PMID: 9794232 DOI: 10.1038/sj.onc.1202069]
- 123 **Gulappa T**, Reddy RS, Suman S, Nyakeriga AM, Damodaran C. Molecular interplay between cdk4 and p21 dictates G0/G1 cell cycle arrest in prostate cancer cells. *Cancer Lett* 2013; **337**: 177-183 [PMID: 23684928 DOI: 10.1016/j.canlet.2013.05.014]
- 124 **Fang Y**, DeMarco VG, Nicholl MB. Resveratrol enhances radiation sensitivity in prostate cancer by inhibiting cell proliferation and promoting cell senescence and apoptosis. *Cancer Sci* 2012; **103**: 1090-1098 [PMID: 22417066 DOI: 10.1111/j.1349-7006.2012.02272.x]
- 125 **Marconett CN**, Morgenstern TJ, San Roman AK, Sundar SN, Singhal AK, Firestone GL. BZL101, a phytochemical extract from the *Scutellaria barbata* plant, disrupts proliferation of human breast and prostate cancer cells through distinct mechanisms dependent on the cancer cell phenotype. *Cancer Biol Ther* 2010; **10**: 397-405 [PMID: 20574166]
- 126 **Agarwal C**, Dhanalakshmi S, Singh RP, Agarwal R. Inositol hexaphosphate inhibits growth and induces G1 arrest and apoptotic death of androgen-dependent human prostate carcinoma LNCaP cells. *Neoplasia* 2004; **6**: 646-659 [PMID: 15548374 DOI: 10.1593/neo.04232]
- 127 **Ukomadu C**, Dutta A. Inhibition of cdk2 activating phosphorylation by mevastatin. *J Biol Chem* 2003; **278**: 4840-4846 [PMID: 12475985 DOI: 10.1074/jbc.M208658200]
- 128 **Li R**, Erdamar S, Dai H, Sayeeduddin M, Frolov A, Wheeler TM, Ayala GE. Cytoplasmic accumulation of glycogen synthase kinase-3beta is associated with aggressive clinicopathological features in human prostate cancer. *Anticancer Res* 2009; **29**: 2077-2081 [PMID: 19528467]
- 129 **Mulholland DJ**, Dedhar S, Wu H, Nelson CC. PTEN and GSK3beta: key regulators of progression to androgen-independent prostate cancer. *Oncogene* 2006; **25**: 329-337 [PMID: 16421604 DOI: 10.1038/sj.onc.1209020]
- 130 **Wang L**, Lin HK, Hu YC, Xie S, Yang L, Chang C. Suppression of androgen receptor-mediated transactivation and cell growth by the glycogen synthase kinase 3 beta in prostate cells. *J Biol Chem* 2004; **279**: 32444-32452 [PMID: 15178691 DOI: 10.1074/jbc.M313963200]
- 131 **Salas TR**, Kim J, Vakar-Lopez F, Sabichi AL, Troncoso P, Jenster G, Kikuchi A, Chen SY, Shemshedini L, Suraokar M, Logothetis CJ, DiGiovanni J, Lippman SM, Menter DG. Glycogen synthase kinase-3 beta is involved in the phosphorylation and suppression of androgen receptor activity. *J Biol Chem* 2004; **279**: 19191-19200 [PMID: 14985354 DOI: 10.1074/jbc.M309560200]
- 132 **Schütz SV**, Cronauer MV, Rinnab L. Inhibition of glycogen synthase kinase-3beta promotes nuclear export of the androgen receptor through a CRM1-dependent mechanism in prostate cancer cell lines. *J Cell Biochem* 2010; **109**: 1192-1200 [PMID: 20127713 DOI: 10.1002/jcb.22500]

- 133 **Wang H**, Fang R, Wang XF, Zhang F, Chen DY, Zhou B, Wang HS, Cai SH, Du J. Stabilization of Snail through AKT/GSK-3 β signaling pathway is required for TNF- α -induced epithelial-mesenchymal transition in prostate cancer PC3 cells. *Eur J Pharmacol* 2013; **714**: 48-55 [PMID: 23769744 DOI: 10.1016/j.ejphar.2013.05.046]
- 134 **Liao X**, Zhang L, Thrasher JB, Du J, Li B. Glycogen synthase kinase-3 β suppression eliminates tumor necrosis factor-related apoptosis-inducing ligand resistance in prostate cancer. *Mol Cancer Ther* 2003; **2**: 1215-1222 [PMID: 14617795]
- 135 **Lu Y**, Nie D, Witt WT, Chen Q, Shen M, Xie H, Lai L, Dai Y, Zhang J. Expression of the fat-1 gene diminishes prostate cancer growth in vivo through enhancing apoptosis and inhibiting GSK-3 β phosphorylation. *Mol Cancer Ther* 2008; **7**: 3203-3211 [PMID: 18852124 DOI: 10.1158/1535-7163.MCT-08-0494]
- 136 **Ban JO**, Oh JH, Son SM, Won D, Song HS, Han SB, Moon DC, Kang KW, Song MJ, Hong JT. Troglitazone, a PPAR agonist, inhibits human prostate cancer cell growth through inactivation of NF κ B via suppression of GSK-3 β expression. *Cancer Biol Ther* 2011; **12**: 288-296 [PMID: 21613824]
- 137 **Li Y**, Wang Z, Kong D, Li R, Sarkar SH, Sarkar FH. Regulation of Akt/FOXO3a/GSK-3 β /AR signaling network by isoflavone in prostate cancer cells. *J Biol Chem* 2008; **283**: 27707-27716 [PMID: 18687691 DOI: 10.1074/jbc.M802759200]
- 138 **Zhu H**, Han B, Pan X, Qi H, Xu L. Thiazolidinediones induce tumour-cell apoptosis through the Akt-GSK3 β pathway. *J Clin Pharm Ther* 2012; **37**: 65-70 [PMID: 21410737 DOI: 10.1111/j.1365-2710.2011.01251.x]
- 139 **Benelli R**, Monteghirfo S, Venè R, Tosetti F, Ferrari N. The chemopreventive retinoid 4HPR impairs prostate cancer cell migration and invasion by interfering with FAK/AKT/GSK3 β pathway and beta-catenin stability. *Mol Cancer* 2010; **9**: 142 [PMID: 20537156 DOI: 10.1186/1476-4598-9-142]
- 140 **Barneda-Zahonero B**, Parra M. Histone deacetylases and cancer. *Mol Oncol* 2012; **6**: 579-589 [PMID: 22963873 DOI: 10.1016/j.molonc.2012.07.003]
- 141 **Weichert W**, Röske A, Gekeler V, Beckers T, Stephan C, Jung K, Fritzsche FR, Niesporek S, Denkert C, Dietel M, Kristiansen G. Histone deacetylases 1, 2 and 3 are highly expressed in prostate cancer and HDAC2 expression is associated with shorter PSA relapse time after radical prostatectomy. *Br J Cancer* 2008; **98**: 604-610 [PMID: 18212746 DOI: 10.1038/sj.bjc.6604199]
- 142 **Waltregny D**, North B, Van Mellaert F, de Leval J, Verdin E, Castronovo V. Screening of histone deacetylases (HDAC) expression in human prostate cancer reveals distinct class I HDAC profiles between epithelial and stromal cells. *Eur J Histochem* 2004; **48**: 273-290 [PMID: 15590418]
- 143 **Song Y**, Shiota M, Tamiya S, Kuroiwa K, Naito S, Tsuneyoshi M. The significance of strong histone deacetylase 1 expression in the progression of prostate cancer. *Histopathology* 2011; **58**: 773-780 [PMID: 21438903 DOI: 10.1111/j.1365-2559.2011.03797.x]
- 144 **Kim NH**, Kim SN, Kim YK. Involvement of HDAC1 in E-cadherin expression in prostate cancer cells; its implication for cell motility and invasion. *Biochem Biophys Res Commun* 2011; **404**: 915-921 [PMID: 21184735 DOI: 10.1016/j.bbrc.2010.12.081]
- 145 **Gaughan L**, Logan IR, Cook S, Neal DE, Robson CN. Tip60 and histone deacetylase 1 regulate androgen receptor activity through changes to the acetylation status of the receptor. *J Biol Chem* 2002; **277**: 25904-25913 [PMID: 11994312 DOI: 10.1074/jbc.M203423200]
- 146 **Halkidou K**, Gaughan L, Cook S, Leung HY, Neal DE, Robson CN. Upregulation and nuclear recruitment of HDAC1 in hormone refractory prostate cancer. *Prostate* 2004; **59**: 177-189 [PMID: 15042618 DOI: 10.1002/pros.20022]
- 147 **Kovacs JJ**, Murphy PJ, Gaillard S, Zhao X, Wu JT, Nicchitta CV, Yoshida M, Toft DO, Pratt WB, Yao TP. HDAC6 regulates Hsp90 acetylation and chaperone-dependent activation of glucocorticoid receptor. *Mol Cell* 2005; **18**: 601-607 [PMID: 15916966 DOI: 10.1016/j.molcel.2005.04.021]
- 148 **Ai J**, Wang Y, Dar JA, Liu J, Liu L, Nelson JB, Wang Z. HDAC6 regulates androgen receptor hypersensitivity and nuclear localization via modulating Hsp90 acetylation in castration-resistant prostate cancer. *Mol Endocrinol* 2009; **23**: 1963-1972 [PMID: 19855091 DOI: 10.1210/me.2009-0188]
- 149 **Gibbs A**, Schwartzman J, Deng V, Alumkal J. Sulforaphane destabilizes the androgen receptor in prostate cancer cells by inactivating histone deacetylase 6. *Proc Natl Acad Sci USA* 2009; **106**: 16663-16668 [PMID: 19805354 DOI: 10.1073/pnas.0908908106]
- 150 **Clarke JD**, Hsu A, Yu Z, Dashwood RH, Ho E. Differential effects of sulforaphane on histone deacetylases, cell cycle arrest and apoptosis in normal prostate cells versus hyperplastic and cancerous prostate cells. *Mol Nutr Food Res* 2011; **55**: 999-1009 [PMID: 21374800 DOI: 10.1002/mnfr.201000547]
- 151 **Noonan EJ**, Place RF, Pookot D, Basak S, Whitson JM, Hirata H, Giardina C, Dahiya R. miR-449a targets HDAC-1 and induces growth arrest in prostate cancer. *Oncogene* 2009; **28**: 1714-1724 [PMID: 19252524 DOI: 10.1038/onc.2009.19]
- 152 **Li X**, Kaplun A, Lonardo F, Heath E, Sarkar FH, Irish J, Sakr W, Sheng S. HDAC1 inhibition by maspin abrogates epigenetic silencing of glutathione S-transferase pi in prostate carcinoma cells. *Mol Cancer Res* 2011; **9**: 733-745 [PMID: 21622623 DOI: 10.1158/1541-7786.MCR-10-0505]
- 153 **Lavery DN**, Villarronga MA, Walker MM, Patel A, Belandia B, Bevan CL. Repression of androgen receptor activity by HEYL, a third member of the Hairy/Enhancer-of-split-related family of Notch effectors. *J Biol Chem* 2011; **286**: 17796-17808 [PMID: 21454491 DOI: 10.1074/jbc.M110.198655]
- 154 **Mercurio F**, Zhu H, Murray BW, Shevchenko A, Bennett BL, Li J, Young DB, Barbosa M, Mann M, Manning A, Rao A. IKK-1 and IKK-2: cytokine-activated I κ B kinases essential for NF- κ B activation. *Science* 1997; **278**: 860-866 [PMID: 9346484]
- 155 **Baud V**, Karin M. Is NF- κ B a good target for cancer therapy? Hopes and pitfalls. *Nat Rev Drug Discov* 2009; **8**: 33-40 [PMID: 19116625 DOI: 10.1038/nrd2781]
- 156 **Gasparian AV**, Yao YJ, Kowalczyk D, Lyakh LA, Karseladze A, Slaga TJ, Budunova IV. The role of IKK in constitutive activation of NF- κ B transcription factor in prostate carcinoma cells. *J Cell Sci* 2002; **115**: 141-151 [PMID: 11801732]
- 157 **Häggarth L**, Hägglöf C, Jaraj SJ, Wester K, Pontén F, Ostman A, Egevad L. Diagnostic biomarkers of prostate cancer. *Scand J Urol Nephrol* 2011; **45**: 60-67 [PMID: 21034352 DOI: 10.3109/00365599.2010.526141]
- 158 **Xu C**, Shen G, Chen C, Gélinas C, Kong AN. Suppression of NF- κ B and NF- κ B-regulated gene expression by sulforaphane and PEITC through I κ B α , IKK pathway in human prostate cancer PC-3 cells. *Oncogene* 2005; **24**: 4486-4495 [PMID: 15856023 DOI: 10.1038/sj.onc.1208656]
- 159 **Shirley RB**, Kaddour-Djebbar I, Patel DM, Lakshmiathan V, Lewis RW, Kumar MV. Combination of proteasomal inhibitors lactacystin and MG132 induced synergistic apoptosis in prostate cancer cells. *Neoplasia* 2005; **7**: 1104-1111 [PMID: 16354593]
- 160 **Inoue T**, Kon T, Ohkura R, Yamakawa H, Ohara O, Yokota J, Sutoh K. BREK/LMTK2 is a myosin VI-binding protein involved in endosomal membrane trafficking. *Genes Cells* 2008; **13**: 483-495 [PMID: 18429820 DOI: 10.1111/j.1365-2443.2008.01184.x]
- 161 **Puri C**, Chibalina MV, Arden SD, Kruppa AJ, Kendrick-Jones J, Buss F. Overexpression of myosin VI in prostate cancer cells enhances PSA and VEGF secretion, but has no effect on endocytosis. *Oncogene* 2010; **29**: 188-200 [PMID: 19855435 DOI: 10.1038/onc.2009.328]
- 162 **Guy M**, Kote-Jarai Z, Giles GG, Al Olama AA, Jugurnauth SK, Mulholland S, Leongamornlert DA, Edwards SM,

- Morrison J, Field HI, Southey MC, Severi G, Donovan JL, Hamdy FC, Dearnaley DP, Muir KR, Smith C, Bagnato M, Arderm-Jones AT, Hall AL, O'Brien LT, Gehr-Swain BN, Wilkinson RA, Cox A, Lewis S, Brown PM, Jhavar SG, Tymrakiewicz M, Lophatananon A, Bryant SL, Horwich A, Huddart RA, Khoo VS, Parker CC, Woodhouse CJ, Thompson A, Christmas T, Ogden C, Fisher C, Jameson C, Cooper CS, English DR, Hopper JL, Neal DE, Easton DF, Eeles RA. Identification of new genetic risk factors for prostate cancer. *Asian J Androl* 2009; **11**: 49-55 [PMID: 19050691 DOI: 10.1038/aja.2008.18]
- 163 **Harries LW**, Perry JR, McCullagh P, Crundwell M. Alterations in LMTK2, MSMB and HNF1B gene expression are associated with the development of prostate cancer. *BMC Cancer* 2010; **10**: 315 [PMID: 20569440 DOI: 10.1186/1471-2407-10-315]
- 164 **Kesavapany S**, Lau KF, Ackerley S, Banner SJ, Shemilt SJ, Cooper JD, Leigh PN, Shaw CE, McLoughlin DM, Miller CC. Identification of a novel, membrane-associated neuronal kinase, cyclin-dependent kinase 5/p35-regulated kinase. *J Neurosci* 2003; **23**: 4975-4983 [PMID: 12832520]
- 165 **Eto M**, Elliott E, Prickett TD, Brautigan DL. Inhibitor-2 regulates protein phosphatase-1 complexed with NimA-related kinase to induce centrosome separation. *J Biol Chem* 2002; **277**: 44013-44020 [PMID: 12221103 DOI: 10.1074/jbc.M208035200]
- 166 **Tansey WP**. Mammalian MYC Proteins and Cancer. *New J Sci* 2014; **2014**: 27 [DOI: 10.1155/2014/757534]
- 167 **Gurel B**, Iwata T, Koh CM, Jenkins RB, Lan F, Van Dang C, Hicks JL, Morgan J, Cornish TC, Sutcliffe S, Isaacs WB, Luo J, De Marzo AM. Nuclear MYC protein overexpression is an early alteration in human prostate carcinogenesis. *Mod Pathol* 2008; **21**: 1156-1167 [PMID: 18567993 DOI: 10.1038/modpathol.2008.111]
- 168 **Gil J**, Kerai P, Lleonart M, Bernard D, Cigudosa JC, Peters G, Carnero A, Beach D. Immortalization of primary human prostate epithelial cells by c-Myc. *Cancer Res* 2005; **65**: 2179-2185 [PMID: 15781629 DOI: 10.1158/0008-5472.CAN-03-4030]
- 169 **Amatangelo MD**, Goodyear S, Varma D, Stearns ME. c-Myc expression and MEK1-induced Erk2 nuclear localization are required for TGF-beta induced epithelial-mesenchymal transition and invasion in prostate cancer. *Carcinogenesis* 2012; **33**: 1965-1975 [PMID: 22791812 DOI: 10.1093/carcin/bgs227]
- 170 **Grad JM**, Dai JL, Wu S, Burnstein KL. Multiple androgen response elements and a Myc consensus site in the androgen receptor (AR) coding region are involved in androgen-mediated up-regulation of AR messenger RNA. *Mol Endocrinol* 1999; **13**: 1896-1911 [PMID: 10551783 DOI: 10.1210/mend.13.11.0369]
- 171 **Hawksworth D**, Ravindranath L, Chen Y, Furusato B, Sesterhenn IA, McLeod DG, Srivastava S, Petrovics G. Overexpression of C-MYC oncogene in prostate cancer predicts biochemical recurrence. *Prostate Cancer Prostatic Dis* 2010; **13**: 311-315 [PMID: 20820186 DOI: 10.1038/pcan.2010.31]
- 172 **Fromont G**, Godet J, Peyret A, Irani J, Celhay O, Rozet F, Cathelineau X, Cussenot O. 8q24 amplification is associated with Myc expression and prostate cancer progression and is an independent predictor of recurrence after radical prostatectomy. *Hum Pathol* 2013; **44**: 1617-1623 [PMID: 23574779 DOI: 10.1016/j.humpath.2013.01.012]
- 173 **Antonarakis ES**, Keizman D, Zhang Z, Gurel B, Lotan TL, Hicks JL, Fedor HL, Carducci MA, De Marzo AM, Eisenberger MA. An immunohistochemical signature comprising PTEN, MYC, and Ki67 predicts progression in prostate cancer patients receiving adjuvant docetaxel after prostatectomy. *Cancer* 2012; **118**: 6063-6071 [PMID: 22674438 DOI: 10.1002/cncr.27689]
- 174 **Hatano K**, Yamaguchi S, Nimura K, Murakami K, Nagahara A, Fujita K, Uemura M, Nakai Y, Tsuchiya M, Nakayama M, Nonomura N, Kaneda Y. Residual prostate cancer cells after docetaxel therapy increase the tumorigenic potential via constitutive signaling of CXCR4, ERK1/2 and c-Myc. *Mol Cancer Res* 2013; **11**: 1088-1100 [PMID: 23788635 DOI: 10.1158/1541-7786.MCR-13-0029-T]
- 175 **Tawfic S**, Goueli SA, Olson MO, Ahmed K. Androgenic regulation of phosphorylation and stability of nucleolar protein nucleolin in rat ventral prostate. *Prostate* 1994; **24**: 101-106 [PMID: 8309845]
- 176 **Tate A**, Isotani S, Bradley MJ, Sikes RA, Davis R, Chung LW, Edlund M. Met-Independent Hepatocyte Growth Factor-mediated regulation of cell adhesion in human prostate cancer cells. *BMC Cancer* 2006; **6**: 197 [PMID: 16869958 DOI: 10.1186/1471-2407-6-197]
- 177 **Altintas DM**, Vlaeminck V, Angelov D, Dimitrov S, Samarut J. Cell cycle regulated expression of NCoR might control cyclic expression of androgen responsive genes in an immortalized prostate cell line. *Mol Cell Endocrinol* 2011; **332**: 149-162 [PMID: 20974212 DOI: 10.1016/j.mce.2010.10.007]
- 178 **Choi HK**, Yoo JY, Jeong MH, Park SY, Shin DM, Jang SW, Yoon HG, Choi KC. Protein kinase A phosphorylates NCoR to enhance its nuclear translocation and repressive function in human prostate cancer cells. *J Cell Physiol* 2013; **228**: 1159-1165 [PMID: 23129261 DOI: 10.1002/jcp.24269]
- 179 **Battaglia S**, Maguire O, Thorne JL, Hornung LB, Doig CL, Liu S, Sucheston LE, Bianchi A, Khanim FL, Gommersall LM, Coulter HS, Rakha S, Giddings I, O'Neill LP, Cooper CS, McCabe CJ, Bunce CM, Campbell MJ. Elevated NCOR1 disrupts PPARalpha/gamma signaling in prostate cancer and forms a targetable epigenetic lesion. *Carcinogenesis* 2010; **31**: 1650-1660 [PMID: 20466759 DOI: 10.1093/carcin/bgq086]
- 180 **Wang Y**, Li JQ, Shao C, Shi CH, Liu F, Yang ZY, Qiu JX, Li YM, Fu Q, Zhang W, Xue W, Lei YH, Gao JY, Wang JY, Gao XP, Yuan JL, Bao TY, Zhang YT. Androgen receptor coregulators NOCR1, TIF2, and ARA70 may account for the hydroxyflutamide insensitivity of prostate cancer cells. *Ir J Med Sci* 2011; **180**: 865-872 [PMID: 21748440 DOI: 10.1007/s11845-011-0714-4]
- 181 **Schranz N**, da Silva Correia J, Fowler B, Ge Q, Sun Z, Bokoch GM. Mechanism of p21-activated kinase 6-mediated inhibition of androgen receptor signaling. *J Biol Chem* 2004; **279**: 1922-1931 [PMID: 14573606 DOI: 10.1074/jbc.M311145200]
- 182 **Kaur R**, Yuan X, Lu ML, Balk SP. Increased PAK6 expression in prostate cancer and identification of PAK6 associated proteins. *Prostate* 2008; **68**: 1510-1516 [PMID: 18642328 DOI: 10.1002/pros.20787]
- 183 **Yang F**, Li X, Sharma M, Zarnegar M, Lim B, Sun Z. Androgen receptor specifically interacts with a novel p21-activated kinase, PAK6. *J Biol Chem* 2001; **276**: 15345-15353 [PMID: 11278661 DOI: 10.1074/jbc.M010311200]
- 184 **Lee SR**, Ramos SM, Ko A, Masiello D, Swanson KD, Lu ML, Balk SP. AR and ER interaction with a p21-activated kinase (PAK6). *Mol Endocrinol* 2002; **16**: 85-99 [PMID: 11773441 DOI: 10.1210/mend.16.1.0753]
- 185 **Liu T**, Li Y, Gu H, Zhu G, Li J, Cao L, Li F. p21-Activated kinase 6 (PAK6) inhibits prostate cancer growth via phosphorylation of androgen receptor and tumorigenic E3 ligase murine double minute-2 (Mdm2). *J Biol Chem* 2013; **288**: 3359-3369 [PMID: 23132866 DOI: 10.1074/jbc.M112.384289]
- 186 **Wen X**, Li X, Liao B, Liu Y, Wu J, Yuan X, Ouyang B, Sun Q, Gao X. Knockdown of p21-activated kinase 6 inhibits prostate cancer growth and enhances chemosensitivity to docetaxel. *Urology* 2009; **73**: 1407-1411 [PMID: 19362342 DOI: 10.1016/j.urology.2008.09.041]
- 187 **Zhang M**, Siedow M, Saia G, Chakravarti A. Inhibition of p21-activated kinase 6 (PAK6) increases radiosensitivity of prostate cancer cells. *Prostate* 2010; **70**: 807-816 [PMID: 20054820 DOI: 10.1002/pros.21114]

- 188 **Uzoh CC**, Perks CM, Bahl A, Holly JM, Sugiono M, Persad RA. PTEN-mediated pathways and their association with treatment-resistant prostate cancer. *BJU Int* 2009; **104**: 556-561 [PMID: 19220271 DOI: 10.1111/j.1464-410X.2009.08411.x]
- 189 **Shen MM**, Abate-Shen C. Pten inactivation and the emergence of androgen-independent prostate cancer. *Cancer Res* 2007; **67**: 6535-6538 [PMID: 17638861 DOI: 10.1158/0008-5472.CAN-07-1271]
- 190 **Tian L**, Fang YX, Xue JL, Chen JZ. Four microRNAs promote prostate cell proliferation with regulation of PTEN and its downstream signals in vitro. *PLoS One* 2013; **8**: e75885 [PMID: 24098737 DOI: 10.1371/journal.pone.0075885]
- 191 **Kong L**, Schäfer G, Bu H, Zhang Y, Zhang Y, Klocker H. Lamin A/C protein is overexpressed in tissue-invasive prostate cancer and promotes prostate cancer cell growth, migration and invasion through the PI3K/AKT/PTEN pathway. *Carcinogenesis* 2012; **33**: 751-759 [PMID: 22301279 DOI: 10.1093/carcin/bgs022]
- 192 **Conley-LaComb MK**, Saliganan A, Kandagatla P, Chen YQ, Cher ML, Chinni SR. PTEN loss mediated Akt activation promotes prostate tumor growth and metastasis via CXCL12/CXCR4 signaling. *Mol Cancer* 2013; **12**: 85 [PMID: 23902739 DOI: 10.1186/1476-4598-12-85]
- 193 **Conley-LaComb MK**, Huang W, Wang S, Shi D, Jung YS, Najy A, Fridman R, Bonfil RD, Cher ML, Chen YQ, Kim HR. PTEN regulates PDGF ligand switch for β -PDGFR signaling in prostate cancer. *Am J Pathol* 2012; **180**: 1017-1027 [PMID: 22209699 DOI: 10.1016/j.ajpath.2011.11.021]
- 194 **Fan C**, He L, Kapoor A, Rybak AP, De Melo J, Cutz JC, Tang D. PTEN inhibits BMI1 function independently of its phosphatase activity. *Mol Cancer* 2009; **8**: 98 [PMID: 19903340 DOI: 10.1186/1476-4598-8-98]
- 195 **Chetram MA**, Odero-Marrah V, Hinton CV. Loss of PTEN permits CXCR4-mediated tumorigenesis through ERK1/2 in prostate cancer cells. *Mol Cancer Res* 2011; **9**: 90-102 [PMID: 21076047 DOI: 10.1158/1541-7786.MCR-10-0235]
- 196 **Nagle RB**, Algotar AM, Cortez CC, Smith K, Jones C, Sathyanarayana UG, Yun S, Riley J, Nagy D, Dittamore R, Dalkin B, Brosh L, Pestano G. ERG overexpression and PTEN status predict capsular penetration in prostate carcinoma. *Prostate* 2013; **73**: 1233-1240 [PMID: 23653096 DOI: 10.1002/pros.22675]
- 197 **Gabriel K**, Ingram A, Austin R, Kapoor A, Tang D, Majeed F, Qureshi T, Al-Nedawi K. Regulation of the tumor suppressor PTEN through exosomes: a diagnostic potential for prostate cancer. *PLoS One* 2013; **8**: e70047 [PMID: 23936141 DOI: 10.1371/journal.pone.0070047]
- 198 **Lotan TL**, Gumuskaya B, Rahimi H, Hicks JL, Iwata T, Robinson BD, Epstein JI, De Marzo AM. Cytoplasmic PTEN protein loss distinguishes intraductal carcinoma of the prostate from high-grade prostatic intraepithelial neoplasia. *Mod Pathol* 2013; **26**: 587-603 [PMID: 23222491 DOI: 10.1038/modpathol.2012.201]
- 199 **Cuzick J**, Yang ZH, Fisher G, Tikishvili E, Stone S, Lanchbury JS, Camacho N, Merson S, Brewer D, Cooper CS, Clark J, Berney DM, Møller H, Scardino P, Sangale Z. Prognostic value of PTEN loss in men with conservatively managed localised prostate cancer. *Br J Cancer* 2013; **108**: 2582-2589 [PMID: 23695019 DOI: 10.1038/bjc.2013.248]
- 200 **Chaux A**, Peskoe SB, Gonzalez-Roibon N, Schultz L, Albaine R, Hicks J, De Marzo AM, Platz EA, Netto GJ. Loss of PTEN expression is associated with increased risk of recurrence after prostatectomy for clinically localized prostate cancer. *Mod Pathol* 2012; **25**: 1543-1549 [PMID: 22684219 DOI: 10.1038/modpathol.2012.104]
- 201 **Cathomas R**, Rothermundt C, Klingbiel D, Bubendorf L, Jaggi R, Betticher DC, Brauchli P, Cotting D, Droege C, Winterhalder R, Siciliano D, Berthold DR, Pless M, Schiess R, von Moos R, Gillessen S. Efficacy of cetuximab in metastatic castration-resistant prostate cancer might depend on EGFR and PTEN expression: results from a phase II trial (SAKK 08/07). *Clin Cancer Res* 2012; **18**: 6049-6057 [PMID: 22977195 DOI: 10.1158/1078-0432.CCR-12-2219]
- 202 **Wang Y**, Romigh T, He X, Orloff MS, Silverman RH, Heston WD, Eng C. Resveratrol regulates the PTEN/AKT pathway through androgen receptor-dependent and -independent mechanisms in prostate cancer cell lines. *Hum Mol Genet* 2010; **19**: 4319-4329 [PMID: 20729295 DOI: 10.1093/hmg/ddq354]
- 203 **Menon R**, Deng M, Rüenauer K, Queisser A, Peifer M, Oferrmann A, Boehm D, Vogel W, Scheble V, Fend F, Kristiansen G, Wernert N, Oberbeckmann N, Biskup S, Rubin MA, Shaikhibrahim Z, Perner S. Somatic copy number alterations by whole-exome sequencing implicates YWHAZ and PTK2 in castration-resistant prostate cancer. *J Pathol* 2013; **231**: 505-516 [PMID: 24114522 DOI: 10.1002/path.4274]
- 204 **Tremblay L**, Hauck W, Aprikian AG, Begin LR, Chapdelaine A, Chevalier S. Focal adhesion kinase (pp125FAK) expression, activation and association with paxillin and p50CSK in human metastatic prostate carcinoma. *Int J Cancer* 1996; **68**: 164-171 [PMID: 8900422]
- 205 **Lee LF**, Louie MC, Desai SJ, Yang J, Chen HW, Evans CP, Kung HJ. Interleukin-8 confers androgen-independent growth and migration of LNCaP: differential effects of tyrosine kinases Src and FAK. *Oncogene* 2004; **23**: 2197-2205 [PMID: 14767470 DOI: 10.1038/sj.onc.1207344]
- 206 **El Haibi CP**, Sharma PK, Singh R, Johnson PR, Suttles J, Singh S, Lillard JW. PI3Kp110-, Src-, FAK-dependent and DOCK2-independent migration and invasion of CXCL13-stimulated prostate cancer cells. *Mol Cancer* 2010; **9**: 85 [PMID: 20412587 DOI: 10.1186/1476-4598-9-85]
- 207 **Guo X**, Yin S, Dong Y, Fan L, Ye M, Lu J, Hu H. Enhanced apoptotic effects by the combination of curcumin and methylseleninic acid: potential role of Mcl-1 and FAK. *Mol Carcinog* 2013; **52**: 879-889 [PMID: 22711297 DOI: 10.1002/mc.21933]
- 208 **Bergan R**, Kyle E, Nguyen P, Trepel J, Ingui C, Neckers L. Genistein-stimulated adherence of prostate cancer cells is associated with the binding of focal adhesion kinase to beta-1-integrin. *Clin Exp Metastasis* 1996; **14**: 389-398 [PMID: 8878413]
- 209 **Ziv Y**, Bielopolski D, Galanty Y, Lukas C, Taya Y, Schultz DC, Lukas J, Bekker-Jensen S, Bartek J, Shiloh Y. Chromatin relaxation in response to DNA double-strand breaks is modulated by a novel ATM- and KAP-1 dependent pathway. *Nat Cell Biol* 2006; **8**: 870-876 [PMID: 16862143 DOI: 10.1038/ncb1446]
- 210 **Van Tilborgh N**, Spans L, Helsen C, Clinckemalie L, Dubois V, Lerut E, Boonen S, Vanderschueren D, Claessens F. The transcription intermediary factor 1 β coactivates the androgen receptor. *J Endocrinol Invest* 2013; **36**: 699-706 [PMID: 23563173 DOI: 10.3275/8927]
- 211 **Zhang Z**, Yang Z, Jäämaa S, Liu H, Pellakuru LG, Iwata T, af Hällström TM, De Marzo AM, Laiho M. Differential epithelium DNA damage response to ATM and DNA-PK pathway inhibition in human prostate tissue culture. *Cell Cycle* 2011; **10**: 3545-3553 [PMID: 22030624 DOI: 10.4161/cc.10.20.17841]
- 212 **Aparicio A**, Den RB, Knudsen KE. Time to stratify? The retinoblastoma protein in castrate-resistant prostate cancer. *Nat Rev Urol* 2011; **8**: 562-568 [PMID: 21811228 DOI: 10.1038/nrurol.2011.107]
- 213 **Dean JL**, Knudsen KE. The role of tumor suppressor dysregulation in prostate cancer progression. *Curr Drug Targets* 2013; **14**: 460-471 [PMID: 23410128]
- 214 **Lee JT**, Lehmann BD, Terrian DM, Chappell WH, Stivala F, Libra M, Martelli AM, Steelman LS, McCubrey JA. Targeting prostate cancer based on signal transduction and cell cycle pathways. *Cell Cycle* 2008; **7**: 1745-1762 [PMID: 18594202]
- 215 **Mariot P**, Prevarskaya N, Roudbaraki MM, Le Bourhis X, Van Coppenolle F, Vanoverberghe K, Skryma R. Evidence of functional ryanodine receptor involved in apoptosis of prostate cancer (LNCaP) cells. *Prostate* 2000; **43**: 205-214

- [PMID: 10797495]
- 216 **Kobylewski SE**, Henderson KA, Eckhart CD. Identification of ryanodine receptor isoforms in prostate DU-145, LNCaP, and PWR-1E cells. *Biochem Biophys Res Commun* 2012; **425**: 431-435 [PMID: 22846571 DOI: 10.1016/j.bbrc.2012.07.119]
- 217 **Tang L**, Nogales E, Ciferri C. Structure and function of SWI/SNF chromatin remodeling complexes and mechanistic implications for transcription. *Prog Biophys Mol Biol* 2010; **102**: 122-128 [PMID: 20493208 DOI: 10.1016/j.pbiomolbio.2010.05.001]
- 218 **Prensner JR**, Iyer MK, Sahu A, Asangani IA, Cao Q, Patel L, Vergara IA, Davicioni E, Erho N, Ghadessi M, Jenkins RB, Triche TJ, Malik R, Bedenis R, McGregor N, Ma T, Chen W, Han S, Jing X, Cao X, Wang X, Chandler B, Yan W, Siddiqui J, Kunju LP, Dhanasekaran SM, Pienta KJ, Feng FY, Chinnaiyan AM. The long noncoding RNA SChLAP1 promotes aggressive prostate cancer and antagonizes the SWI/SNF complex. *Nat Genet* 2013; **45**: 1392-1398 [PMID: 24076601 DOI: 10.1038/Ng.2771]
- 219 **Brush MH**, Guardiola A, Connor JH, Yao TP, Shenolikar S. Deacetylase inhibitors disrupt cellular complexes containing protein phosphatases and deacetylases. *J Biol Chem* 2004; **279**: 7685-7691 [PMID: 14670976 DOI: 10.1074/jbc.M310997200]
- 220 **Schillace RV**, Scott JD. Association of the type 1 protein phosphatase PP1 with the A-kinase anchoring protein AKAP220. *Curr Biol* 1999; **9**: 321-324 [PMID: 10209101]
- 221 **Schillace RV**, Voltz JW, Sim AT, Shenolikar S, Scott JD. Multiple interactions within the AKAP220 signaling complex contribute to protein phosphatase 1 regulation. *J Biol Chem* 2001; **276**: 12128-12134 [PMID: 11152471 DOI: 10.1074/jbc.M010398200]
- 222 **Ayllón V**, Cayla X, García A, Fleischer A, Rebollo A. The anti-apoptotic molecules Bcl-xL and Bcl-w target protein phosphatase 1alpha to Bad. *Eur J Immunol* 2002; **32**: 1847-1855 [PMID: 12115603 DOI: 10.1002/1521-4141(200207)32:7<1847::AID-IMMU1847>3.0.CO;2-7]
- 223 **Li HY**, Liu H, Wang CH, Zhang JY, Man JH, Gao YF, Zhang PJ, Li WH, Zhao J, Pan X, Zhou T, Gong WL, Li AL, Zhang XM. Deactivation of the kinase IKK by CUEDC2 through recruitment of the phosphatase PP1. *Nat Immunol* 2008; **9**: 533-541 [PMID: 18362886 DOI: 10.1038/ni.1600]
- 224 **Gunawardena SR**, Ruis BL, Meyer JA, Kapoor M, Conklin KF. NOM1 targets protein phosphatase 1 to the nucleolus. *J Biol Chem* 2008; **283**: 398-404 [PMID: 17965019 DOI: 10.1074/jbc.M706708200]
- 225 **Kim YM**, Watanabe T, Allen PB, Kim YM, Lee SJ, Greengard P, Nairn AC, Kwon YG. PNUTS, a protein phosphatase 1 (PP1) nuclear targeting subunit. Characterization of its PP1- and RNA-binding domains and regulation by phosphorylation. *J Biol Chem* 2003; **278**: 13819-13828 [PMID: 12574161 DOI: 10.1074/jbc.M209621200]
- 226 **Landsverk HB**, Kirkhus M, Bollen M, Küntziger T, Collas P. PNUTS enhances in vitro chromosome decondensation in a PP1-dependent manner. *Biochem J* 2005; **390**: 709-717 [PMID: 15907195 DOI: 10.1042/BJ20050678]
- 227 **Brush MH**, Weiser DC, Shenolikar S. Growth arrest and DNA damage-inducible protein GADD34 targets protein phosphatase 1 alpha to the endoplasmic reticulum and promotes dephosphorylation of the alpha subunit of eukaryotic translation initiation factor 2. *Mol Cell Biol* 2003; **23**: 1292-1303 [PMID: 12556489]
- 228 **Lee SJ**, Lim CJ, Min JK, Lee JK, Kim YM, Lee JY, Won MH, Kwon YG. Protein phosphatase 1 nuclear targeting subunit is a hypoxia inducible gene: its role in post-translational modification of p53 and MDM2. *Cell Death Differ* 2007; **14**: 1106-1116 [PMID: 17318220 DOI: 10.1038/sj.cdd.4402111]
- 229 **Guo CY**, Brautigan DL, Larner JM. Ionizing radiation activates nuclear protein phosphatase-1 by ATM-dependent dephosphorylation. *J Biol Chem* 2002; **277**: 41756-41761 [PMID: 12202491 DOI: 10.1074/jbc.M207519200]
- 230 **Tang X**, Hui ZG, Cui XL, Garg R, Kastan MB, Xu B. A novel ATM-dependent pathway regulates protein phosphatase 1 in response to DNA damage. *Mol Cell Biol* 2008; **28**: 2559-2566 [PMID: 18250156 DOI: 10.1128/MCB.01711-07]
- 231 **Shimada M**, Haruta M, Niida H, Sawamoto K, Nakanishi M. Protein phosphatase 1 γ is responsible for dephosphorylation of histone H3 at Thr 11 after DNA damage. *EMBO Rep* 2010; **11**: 883-889 [PMID: 20948546 DOI: 10.1038/embor.2010.152]
- 232 **Guo C**, Mi J, Brautigan DL, Larner JM. ATM regulates ionizing radiation-induced disruption of HDAC1: PP1: Rb complexes. *Cell Signal* 2007; **19**: 504-510 [PMID: 17008050 DOI: 10.1016/j.cellsig.2006.08.001]
- 233 **Sakashita G**, Shima H, Komatsu M, Urano T, Kikuchi A, Kikuchi K. Regulation of type 1 protein phosphatase/inhibitor-2 complex by glycogen synthase kinase-3 β in intact cells. *J Biochem* 2003; **133**: 165-171 [PMID: 12761178]
- 234 **Wu DY**, Tkachuck DC, Roberson RS, Schubach WH. The human SNF5/INI1 protein facilitates the function of the growth arrest and DNA damage-inducible protein (GADD34) and modulates GADD34-bound protein phosphatase-1 activity. *J Biol Chem* 2002; **277**: 27706-27715 [PMID: 12016208 DOI: 10.1074/jbc.M200955200]
- 235 **Liu Y**, Virshup DM, White RL, Hsu LC. Regulation of BRCA1 phosphorylation by interaction with protein phosphatase 1 α . *Cancer Res* 2002; **62**: 6357-6361 [PMID: 12438214]
- 236 **Manser C**, Guillot F, Vagnoni A, Davies J, Lau KF, McLoughlin DM, De Vos KJ, Miller CC. Lemur tyrosine kinase-2 signalling regulates kinesin-1 light chain-2 phosphorylation and binding of Smad2 cargo. *Oncogene* 2012; **31**: 2773-2782 [PMID: 2196745 DOI: 10.1038/onc.2011.437]
- 237 **Ajuh PM**, Browne GJ, Hawkes NA, Cohen PT, Roberts SG, Lamond AI. Association of a protein phosphatase 1 activity with the human factor C1 (HCF) complex. *Nucleic Acids Res* 2000; **28**: 678-686 [PMID: 10637318]
- 238 **Lesage B**, Beullens M, Pedelini L, Garcia-Gimeno MA, Waelkens E, Sanz P, Bollen M. A complex of catalytically inactive protein phosphatase-1 sandwiched between Sds22 and inhibitor-3. *Biochemistry* 2007; **46**: 8909-8919 [PMID: 17630778 DOI: 10.1021/bi7003119]
- 239 **Helps NR**, Barker HM, Elledge SJ, Cohen PT. Protein phosphatase 1 interacts with p53BP2, a protein which binds to the tumour suppressor p53. *FEBS Lett* 1995; **377**: 295-300 [PMID: 8549741 DOI: 10.1016/0014-5793(95)01347-4]
- 240 **García A**, Cayla X, Guergnon J, Dessauge F, Hospital V, Rebollo MP, Fleischer A, Rebollo A. Serine/threonine protein phosphatases PP1 and PP2A are key players in apoptosis. *Biochimie* 2003; **85**: 721-726 [PMID: 14585537]
- 241 **Parrish AB**. Regulation of Apoptosis Following Mitochondrial Cytochrome c Release. Department of Pharmacology and Molecular Cancer Biology. USA: Duke University, 2010
- 242 **Ayllón V**, Martínez-A C, García A, Cayla X, Rebollo A. Protein phosphatase 1alpha is a Ras-activated Bad phosphatase that regulates interleukin-2 deprivation-induced apoptosis. *EMBO J* 2000; **19**: 2237-2246 [PMID: 10811615 DOI: 10.1093/emboj/19.10.2237]
- 243 **Ayllón V**, Cayla X, García A, Roncal F, Fernández R, Albar JP, Martínez C, Rebollo A. Bcl-2 targets protein phosphatase 1 alpha to Bad. *J Immunol* 2001; **166**: 7345-7352 [PMID: 11390485]
- 244 **Luo W**, Peterson A, Garcia BA, Coombs G, Kofahl B, Heinrich R, Shabanowitz J, Hunt DF, Yost HJ, Virshup DM. Protein phosphatase 1 regulates assembly and function of the beta-catenin degradation complex. *EMBO J* 2007; **26**: 1511-1521 [PMID: 17318175 DOI: 10.1038/sj.emboj.7601607]
- 245 **Yu YM**, Pace SM, Allen SR, Deng CX, Hsu LC. A PP1-binding motif present in BRCA1 plays a role in its DNA repair function. *Int J Biol Sci* 2008; **4**: 352-361 [PMID: 18953404]
- 246 **Hsu LC**. Identification and functional characterization of a PP1-binding site in BRCA1. *Biochem Biophys Res Commun* 2007; **360**: 507-512 [PMID: 17603999 DOI: 10.1016/

- j.bbrc.2007.06.090]
- 247 **Szatmari E**, Habas A, Yang P, Zheng JJ, Hagg T, Hetman M. A positive feedback loop between glycogen synthase kinase 3 β and protein phosphatase 1 after stimulation of NR2B NMDA receptors in forebrain neurons. *J Biol Chem* 2005; **280**: 37526-37535 [PMID: 16155008 DOI: 10.1074/jbc.M502699200]
- 248 **Canettieri G**, Morantte I, Guzmán E, Asahara H, Herzig S, Anderson SD, Yates JR, Montminy M. Attenuation of a phosphorylation-dependent activator by an HDAC-PP1 complex. *Nat Struct Biol* 2003; **10**: 175-181 [PMID: 12567184 DOI: 10.1038/nsb895]
- 249 **Zhao S**, Brandt NR, Caswell AH, Lee EY. Binding of the catalytic subunit of protein phosphatase-1 to the ryanodine-sensitive calcium release channel protein. *Biochemistry* 1998; **37**: 18102-18109 [PMID: 9922179]
- 250 **Sonnleitner A**, Fleischer S, Schindler H. Gating of the skeletal calcium release channel by ATP is inhibited by protein phosphatase 1 but not by Mg²⁺. *Cell Calcium* 1997; **21**: 283-290 [PMID: 9160164 DOI: 10.1016/S0143-4160(97)90116-0]
- 251 **Xiao B**, Tian X, Xie W, Jones PP, Cai S, Wang X, Jiang D, Kong H, Zhang L, Chen K, Walsh MP, Cheng H, Chen SR. Functional consequence of protein kinase A-dependent phosphorylation of the cardiac ryanodine receptor: sensitization of store overload-induced Ca²⁺ release. *J Biol Chem* 2007; **282**: 30256-30264 [PMID: 17693412 DOI: 10.1074/jbc.M703510200]
- 252 **Lu Z**, Wan G, Guo H, Zhang X, Lu X. Protein phosphatase 1 inhibits p53 signaling by dephosphorylating and stabilizing Mdmx. *Cell Signal* 2013; **25**: 796-804 [PMID: 23277204 DOI: 10.1016/j.cellsig.2012.12.014]
- 253 **Fresu M**, Bianchi M, Parsons JT, Villa-Moruzzi E. Cell-cycle-dependent association of protein phosphatase 1 and focal adhesion kinase. *Biochem J* 2001; **358**: 407-414 [PMID: 11513739]
- 254 **Brichese L**, Valette A. PP1 phosphatase is involved in Bcl-2 dephosphorylation after prolonged mitotic arrest induced by paclitaxel. *Biochem Biophys Res Commun* 2002; **294**: 504-508 [PMID: 12051739 DOI: 10.1016/S0006-291X(02)00505-3]
- 255 **Li X**, Lin HH, Chen H, Xu X, Shih HM, Ann DK. SUMOylation of the transcriptional co-repressor KAP1 is regulated by the serine and threonine phosphatase PP1. *Sci Signal* 2010; **3**: ra32 [PMID: 20424263 DOI: 10.1126/scisignal.2000781]
- 256 **Chuang MJ**, Wu ST, Tang SH, Lai XM, Lai HC, Hsu KH, Sun KH, Sun GH, Chang SY, Yu DS, Hsiao PW, Huang SM, Cha TL. The HDAC inhibitor LBH589 induces ERK-dependent prometaphase arrest in prostate cancer via HDAC6 inactivation and down-regulation. *PLoS One* 2013; **8**: e73401 [PMID: 24023871 DOI: 10.1371/journal.pone.0073401]
- 257 **Boyce M**, Bryant KF, Jousse C, Long K, Harding HP, Scheuner D, Kaufman RJ, Ma D, Coen DM, Ron D, Yuan J. A selective inhibitor of eIF2 α dephosphorylation protects cells from ER stress. *Science* 2005; **307**: 935-939 [PMID: 15705855 DOI: 10.1126/science.1101902]
- 258 **Chen CS**, Weng SC, Tseng PH, Lin HP, Chen CS. Histone acetylation-independent effect of histone deacetylase inhibitors on Akt through the reshuffling of protein phosphatase 1 complexes. *J Biol Chem* 2005; **280**: 38879-38887 [PMID: 16186112 DOI: 10.1074/jbc.M505733200]
- 259 **Guergnon J**, Dessauge F, Dominguez V, Viallet J, Bonnefoy S, Yuste VJ, Mercereau-Puijalon O, Cayla X, Rebollo A, Susin SA, Bost PE, Garcia A. Use of penetrating peptides interacting with PP1/PP2A proteins as a general approach for a drug phosphatase technology. *Mol Pharmacol* 2006; **69**: 1115-1124 [PMID: 16387795 DOI: 10.1124/mol.105.019364]
- 260 **Godet AN**, Guergnon J, Maire V, Croset A, Garcia A. The combinatorial PP1-binding consensus Motif (R/K)_x(0,1)V/IxRx(R/K)_x(R/K) is a new apoptotic signature. *PLoS One* 2010; **5**: e9981 [PMID: 20376316 DOI: 10.1371/journal.pone.0009981]

P- Reviewer: Choi CY, Moens U **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ



Potential ability of xanthophylls to prevent obesity-associated cancer

Masaru Terasaki, Michihiro Mutoh, Gen Fujii, Mami Takahashi, Rikako Ishigamori, Sonoko Masuda

Masaru Terasaki, Sonoko Masuda, Department of Health and Environmental Sciences, School of Pharmaceutical Sciences, Health Sciences University of Hokkaido, Ishikari-Tobetsu, Hokkaido 061-0293, Japan

Michihiro Mutoh, Gen Fujii, Rikako Ishigamori, Division of Cancer Prevention Research, National Cancer Center Research Institute, Chuo-ku, Tokyo 104-0045, Japan

Mami Takahashi, Central Animal Division, National Cancer Center Research Institute, Chuo-ku, Tokyo 104-0045, Japan

Author contributions: Terasaki M, Mutoh M, Fujii G, Takahashi M, Ishigamori R and Masuda S contributed to this paper.

Supported by National Cancer Center Research and Development Fund No. 25-A-15; and by The Research Grant of the Princess Takamatsu Cancer Research Fund

Correspondence to: Michihiro Mutoh, MD, PHD, Division of Cancer Prevention Research, National Cancer Center Research Institute, 5-1-1 Tsukiji, Chuo-ku, Tokyo 104-0045, Japan. mimutoh@ncc.go.jp

Telephone: +81-3-35422511 Fax: +81-3-35439305

Received: June 27, 2014 Revised: October 2, 2014

Accepted: October 31, 2014

Published online: December 9, 2014

Abstract

Obesity-associated cancers, including colon cancer and breast cancer, are increasing in Asian countries with Westernized lifestyles as exemplified by reduced physical activity and increased fat/sugar consumption. An excessive accumulation of visceral adipose tissue causes insulin resistance, dyslipidemia and adipocytokine imbalance, and these factors are suggested to be involved in cancer promotion. To prevent obesity-associated cancers, researcher attention is increasing on the so-called "functional foods". In addition, new approaches to cancer control are in high demand, and using "functional foods" as supplemental or adjuvant agents in chemotherapy is thought to be a promising approach. One of these functional ingredients is xanthophylls, which are natural fat-soluble pigments found in fruits, vegetables, algae and other plants. Xanthophylls belong to the carotenoid class and have struc-

tures containing oxygen. Some studies have revealed that xanthophylls improve the inflammation status, serum triglyceride levels, blood pressure levels and liver function test values. Furthermore, recent studies show that xanthophylls possess high anti-cancer, anti-diabetic, anti-obesity and anti-oxidant properties. In this review, we highlight the recent findings for five xanthophylls, namely astaxanthin, β -cryptoxanthin, fucoxanthin, neoxanthin and zeaxanthin/lutein, and their relevance to cancer prevention.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Cancer prevention; Xanthophylls

Core tip: Xanthophylls belong to the class of carotenoids, and are natural fat-soluble pigments found in fruits, vegetables, algae and so on. It has been shown that the versatile functions of xanthophylls have great potential for the prevention of metabolic syndrome and cancers. Xanthophylls have proved safety, and several xanthophylls provide other health benefits, including improvement of inflammation, dyslipidemia, hypertension and liver function. These findings indicate that xanthophylls could be useful to prevent obesity-associated cancer.

Terasaki M, Mutoh M, Fujii G, Takahashi M, Ishigamori R, Masuda S. Potential ability of xanthophylls to prevent obesity-associated cancer. *World J Pharmacol* 2014; 3(4): 140-152 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/140.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.140>

INTRODUCTION

Obesity has recently attracted much interest as a risk factor for several cancers, such as breast cancer and colorectal cancer^[1,2]. Both metabolic syndrome that is characterized by obesity, hyperlipidemia, type 2 diabetes and hypertension

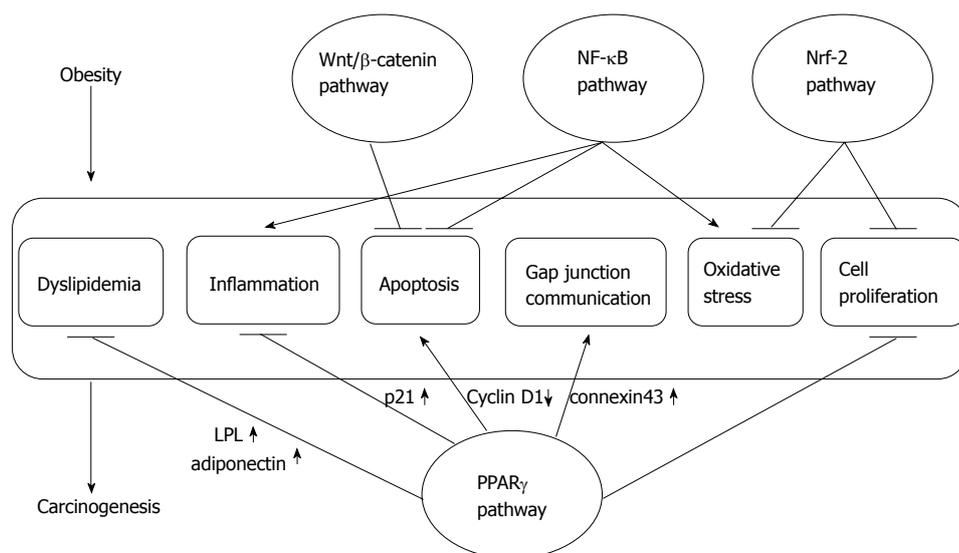


Figure 1 Possible mechanisms for obesity-associated cancer prevention. LPL: Lipoprotein lipase; NF-κB: Nuclear factor kappa B; Nrf2: Nuclear factor-erythroid 2 related factor 2; PPAR: Peroxisome proliferator-activated receptor.

Table 1 Obesity-associated cancers

Type of cancer
Breast (postmenopausal)
Colorectum
Endometrium
Esophagus
Gallbladder
Kidney
Pancreas
Thyroid

and obesity-associated cancers (Table 1) are extremely common in Western countries, and they are currently increasing in Eastern countries, including Japan. The factors linking obesity and cancer are becoming apparent, and they are insulin resistance, dyslipidemia and a subsequent adipocytokine imbalance (Figure 1)^[1,2]. Carotenoid intake is reported to be inversely associated with obesity and with the risk of many cancers^[3-6].

Carotenoids are fat-soluble pigments found in fruits, vegetables, algae and other plants. Humans cannot synthesize carotenoids, and we should therefore consume them as part of our diet. Carotenoids belong to the tetraterpenoid category, and they can be divided into xanthophylls and carotenes according to whether the structure contains oxygen or not. Carotenoids with structures containing oxygen are xanthophylls. As the name indicates, the color of xanthophylls is usually yellow, and they are usually lipophilic because of the long unsaturated aliphatic chain in their structure.

Because conventional chemotherapy has failed to reduce the mortality rates of common cancers, including obesity-associated cancers, new approaches to controlling the development of cancer are in great demand^[7]. One approach is the use of functional foods/plant-derived agents as supplemental or adjuvant agents in chemo-

therapy^[8,9]. Another approach is chemoprevention for the control of cancer development^[8,9]. In both methods, using xanthophylls seems to be an attractive approach. As shown in this review, xanthophylls provided health benefits, such as improvements in inflammation, dyslipidemia, hypertension and liver function. Moreover, the biological significance of xanthophylls as important candidates for the chemoprevention of cancer is becoming clearer, and the safety of xanthophylls has been affirmed, as described in this review. Another candidate called β -carotene is the most abundant dietary carotenoid, and long-term supplementation with this compound has been shown to be ineffective for cancer chemoprevention in several recent large-scale intervention trials^[10-12].

In this review, we focus on recent findings for five xanthophylls as follows: astaxanthin, β -cryptoxanthin, fucoxanthin, neoxanthin and zeaxanthin/lutein, and their relevance to cancer prevention (Figure 2).

ASTAXANTHIN

Distribution and nature of astaxanthin

Astaxanthin (AX) is a natural fat-soluble red pigment and belongs to the xanthophyll subclass of carotenoids. Dietary sources of AX are eggs of salmon and trout, skin of red sea bream, crabs, shrimps and lobsters. AX is synthesized in microalgae (*Chlorella zofingiensis*, *Chlorococcum* and *Haematococcus pluvialis*). Krills (*Euphausia superba*) feed on the microalgae and in turn are fed upon by fishes. The microalga, *H. pluvialis*, is the main source of natural AX and is able to accumulate up to 4% AX on dry weight basis^[13-15]. AX extracted from *H. pluvialis* is used as a food dye in many countries. AX exists in stereoisomers and geometric isomers. *H. pluvialis* biosynthesizes the (3*S*, 3'*S*)-isomer, meanwhile *P. rhodozyma* biosynthesizes the (3*R*, 3'*R*)-isomer. AX has two hydroxyl groups and is able to react with fatty acids and proteins. AX is found as free, mono- and di-ester forms in organisms^[13].

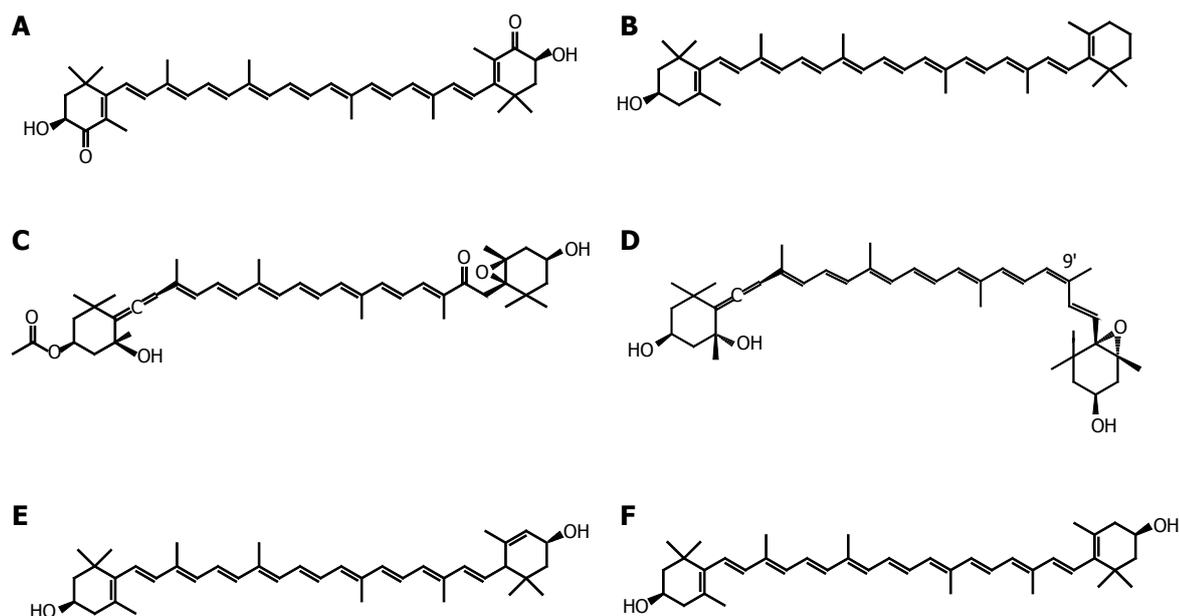


Figure 2 Structure of xanthophylls. A: Astaxanthin; B: β -cryptoxanthin; C: Fucoxanthin; D: 9'-cis-neoxanthin; E: Lutein; F: Zeaxanthin.

AX can take to transverse cell membrane orientation, and shows strong antioxidative activity^[13,15]. After oral administration of AX, AX changes to all-*E*, 9*Z*-, 13*Z*-geometrical isomers and 3*R*,3'*R*-, 3*R*,3'*S* meso-, 3*S*,3'*S*-optical isomers, all of which can be detected in human blood^[16].

Safety profile

Many experimental and clinical studies have demonstrated the safety of AX^[13,17]. In a subchronic toxicity study in rats, feeding AX-rich microalgae biomass corresponding to doses of 465 and 557 mg AX/kg per day for 90 d in male and female rats, respectively, revealed no adverse events^[18]. A randomized, double-blind, placebo-controlled study has demonstrated that it is safe to administer 6 mg/d AX in healthy adults for 8 wk^[19], and a significant decrease of triglycerides and increase of adiponectin and high density lipoprotein cholesterol in participants with mild dyslipidemia by administration of AX at doses of 12 mg/d and 18 mg/d for 12 wk^[20].

Preclinical studies and anti-cancer mechanisms

Oxidative stress and inflammation are closely related to carcinogenesis (Figure 1), and many antioxidants, including carotenoids have been demonstrated to decrease cancer development in experimental animal models^[14]. There are papers on preventive effects of AX on urinary bladder^[21], oral^[22,23], and colorectal^[24,25] carcinogenesis. In mouse urinary bladder cancer induced by *N*-butyl-*N*-(4-hydroxybutyl)nitrosamine (OH-BBN), AX administration at a dose of 50 ppm in water for 20 wk after OH-BBN exposure for 20 wk resulted in a decrease in the incidences of precancerous lesions and bladder cancer^[21]. In rat oral carcinogenesis induced by 4-nitroquinoline 1-oxide (4-NQO), the incidence of oral precancerous le-

sions in rats treated with 20 ppm 4-NQO and 100 ppm AX was smaller compared to those of the non-treatment group, and oral neoplasms did not observed in rats fed AX among the 4-NQO exposure^[22]. In these studies, AX decreased cell growth activity in the non-cancerous epithelial tissues of 4-NQO-exposed animals^[21,22]. AX has also been demonstrated to show preventive effects in 7,12-dimethylbenz[*a*]anthracene (DMBA)-induced hamster buccal pouch carcinogenesis *via* nuclear factor-erythroid 2 related factor 2 (Nrf2) activation^[23]. Moreover, AX has been shown to inhibit nuclear factor kappa B (NF- κ B) and Wnt/ β -catenin signaling pathways^[26]. Related to colorectal carcinogenesis, AX at 500 ppm in diet significantly decreased the development of aberrant crypt foci (ACF) and the incidence and multiplicity of colorectal tumors induced by azoxymethane (AOM)^[24]. AX at 200 ppm in diet also suppressed mucosal ulcers induced by dextran sulfate sodium (DSS), and development of dysplastic ACF and colonic adenocarcinoma induced by both treatment of DSS and AOM^[25]. In addition, AX reduced the number and size of aflatoxin B1-induced liver preneoplastic foci in rats^[27]. Growth of WAZ-2T cells, mammary tumor cells, inoculated into the mice mammary fat pad was also inhibited by AX at 100 ppm or 400 ppm in diet^[28]. Lipid peroxidation activity in tumors was reduced in tumors treated with 400 ppm^[28]. AX markedly attenuated the promotion of hepatic metastasis of P815 mastocytoma cells in a syngenic graft model under restraint stress^[29]. In *in vitro* cell culture systems, AX suppressed invasion of rat ascites hepatoma AH109A cells^[30]. AX inhibited cell proliferation and decreased cell viability of leukemia K562 cells *via* induction of apoptosis along with up-regulation of peroxisome proliferator-activated receptor (PPAR) γ and p21, and down-regulation

of cyclin D1^[31]. Induction of connexin 43, gap junction protein, through activation of PPAR γ is suggested to be one of the anti-tumor mechanisms of AX^[32,33]. Up-regulation of the Nrf2 pathway is also involved in antioxidant activity of AX^[23,31,34], and may improve mitochondrial function^[35]. However, the role of Nrf2 activation in anti-tumorigenesis is controversial. The oncogenic *K-ras* gene induces Nrf2 expression, and detoxification of reactive oxygen species promotes tumor growth^[36]. Deficiency of Nrf2 has been reported to increase induction of tumors in urethane-induced mouse lung carcinogenesis, but reduce the number of malignant tumors harboring activated mutation in the *K-ras* gene, indicating that Nrf2 prevents initiation but accelerates progression under the activation of the K-ras signaling pathway^[37]. Indeed, there is a report that effects of AX differ at the stages of initiation and the stage of promotion in mammary tumors. AX fed before tumor initiation delayed mammary tumor growth and modulated immune response, but AX supplementation after tumor initiation resulted in more rapid tumor growth^[38]. Thus, use of antioxidants for cancer prevention is considered to be useful at the time before tumor initiation, but more caution is required in using them after the stage accompanied with activated K-ras signaling.

Clinical studies

A randomized, double-blind, placebo-controlled study has demonstrated that AX reduces oxidation of fatty acids^[39], decreases oxidative stress markers^[40] and inflammation^[41], and improves dyslipidemia^[20] and age-related dysfunction of eyes^[42,43] and brain^[44]. However, human cancer prevention studies using AX have not yet been reported.

β -CRYPTOXANTHIN

Distribution and nature of β -cryptoxanthin

β -cryptoxanthin (β -CRX) is one of the naturally occurring carotenoid pigments, and is also classified as a xanthophyll. Its unique character is that it is found in specific fruits and vegetables such as mango, papaya, loquat, Japanese persimmon, peach, sweet red peppers and citrus fruits of the mandarin family^[45,46]. Satsuma mandarin, *Citrus unshiu*, is one of the most β -CRX rich fruits in Japan. The content of β -CRX in *C. unshiu* reaches several mg/100g wet weight. The level of β -CRX in Valencia orange is very low and grapefruit has been found to be devoid of it.

In the human body, β -CRX is easily converted to vitamin A (retinol) and is therefore considered as a pro-vitamin A. It is also known that β -CRX might be easily absorbed^[47], and is accumulated in various organs^[48]. Moreover, it can be stored for several months in the human body^[49]. Serum β -CRX concentration could be around 96 $\mu\text{g}/\text{dL}$ ^[50]. It is also reported that β -CRX concentration in Japanese mother's milk and serum are nearly parallel with their intake of the Satsuma mandarin, and

are higher than other countries^[51,52].

Epidemiologic studies

Many epidemiological studies showed the intake of β -CRX was significantly associated with reduced risks of type 2 diabetes [relative risk (RR) = 0.58]^[53] and rheumatoid arthritis (RR = 0.59)^[54]. β -CRX supplementation significantly decreased cigarette smoke-induced lung squamous metaplasia and inflammation^[55]. Regarding cancer risk, several observational epidemiologic studies suggest that β -CRX could potentially prevent cancer development. The demonstrated cancer risks for lung, esophageal and bladder were 0.76 (RR), 0.16 [odds ratio (OR)] and 0.74 (RR), respectively, comparing the highest to lowest quintile of intake^[56-58]. A greater intake of β -CRX was also inversely associated with developing undetermined cervical atypical squamous cells (OR = 0.4)^[59]. Interestingly, the serum level of β -CRX is lower in the patients of liver cancer than that in healthy counterparts^[60]. These results suggest that a high serum β -CRX concentration or intake of β -CRX is beneficial to human health.

Safety profile

The scientific panel on additives and products or substances used in animal feed (FEEDAP) panel members considered β -CRX to appear not to be mutagenic and show no clastogenic activity^[61]. In subchronic studies, The FEEDAP panel could not find any adverse effects^[61]. Also an acceptable daily intake has not been determined^[61]. Previously, we have reported the chemoprevention effect of β -CRX against chemically-induced bladder carcinogenesis in ICR mice^[62]. Mice were fed with 1, 5 and 25 ppm of β -CRX for 24 wk, and no clinical signs of toxicity and poor condition, low survival or histopathological changes were found^[62]. Many epidemiological studies^[53-60,63-68] indicated that administration of β -CRX is safe for human health.

Preclinical studies and anti-cancer mechanisms

Various functions of β -CRX have been reported recently. β -CRX is an antioxidant phytochemical and may help prevent oxidative damage^[69]. Thus, it is believed that β -CRX has health benefits for people with risk of chronic diseases.

Numerous possible mechanisms for the anti-carcinogenic potential of β -CRX have been proposed. These include the antioxidant function that is associated with the enhancement of DNA repair^[55,69], suppression of efficacy of key proinflammatory cytokine expression, such as tumor necrosis factor- α ^[55] and an apoptotic induction effect^[70]. Also, β -CRX is known to stimulate the expression of the RB gene (a tumor-suppressor gene) and *p73* gene (a *p53*-related gene)^[71] and reduce the expression of NF- κ B and activator protein-1 (AP-1), that induces numerous genes including inflammation, cell proliferation, and apoptosis^[55]. These mechanisms indicate that β -CRX may be a promising chemopreventive agent against cancer. Indeed, β -CRX exerts an anti-tumor promoter action *in vitro*^[72] and

inhibits chemically induced carcinogenesis *in vivo*^[62,71,73,74]. Previously, we investigated the effects of β -CRX extracted from *C. unshiu* oranges on OH-BBN-induced urinary bladder carcinogenesis in male ICR mice^[62]. OH-BBN-exposed mice were fed with 1, 5 and 25 ppm of β -CRX for 24 wk starting 1 wk after the cessation of OH-BBN exposure. Feeding with β -CRX decreased the incidence and multiplicity of precancerous and cancerous urinary bladder lesions. Especially, 25 ppm β -CRX markedly reduced the occurrence of bladder cancer. Meanwhile, β -CRX is also reported to reduce mouse skin^[71], mouse lung^[74] and rat colon^[71] carcinogenesis. In our report, β -CRX lowered ratios of cyclin D1-positive cell in various urinary bladder lesions, meaning that reduction in the incidence of precancerous and cancerous urinary bladder lesions is due to reduced cell cycle progression^[62].

Clinical studies

The efficacy of β -CRX supplementation on obesity have been investigated^[75]. Seventeen postmenopausal obese women were provided 200 mL of a beverage containing β -CRX (1.56 mg/serving and 4.7 mg/d) for 3 wk^[75]. As a result, the levels of serum β -CRX were significantly elevated from 0.28 (initial period) to 1.15 mg/mL, and high molecular weight-adiponectin was also elevated from 9.8 to 11.1 mg/mL^[75]. At the end of the study, the levels of serum triglyceride ($P = 0.057$) and total plasminogen activator inhibitor-1 (PAI-1) ($P = 0.052$) tended to decrease. Nishino *et al.*^[60] reported an intervention study where β -CRX-rich mandarin orange juice (3 mg β -CRX in 80 mL) was provided for 12 wk to obese men or obese men with elevated serum γ -glutamyl transpeptidase (γ GTP) levels^[60]. After drinking β -CRX for 12 wk, body weight ($P < 0.001$), BMI ($P < 0.001$) and β -GTP levels ($P < 0.005$) were decreased.

An intervention trial regarding prevention of liver cancer has also been reported^[60]. Viral hepatitis with cirrhosis patients were randomly assigned into two groups in the trial. The treatment group was administered mandarin orange juice enriched with β -CRX and with the carotenoids mixture (lycopene, β -carotene and α -carotene). Patients in the control group were administered a carotenoids mixture alone. At year 2.5, cumulative incidence of liver cancer/hepatocellular carcinoma development in the mandarin orange juice group was lower than that of the carotenoids mixture alone group ($P = 0.05$). The combinational use of natural carotenoids containing β -CRX might be valuable for the prevention of liver cancer in hepatitis virus infected patients with cirrhosis.

FUCOXANTHIN

Distribution and nature of fucoxanthin

Brown seaweeds include *Undaria pinnatifida* (wakame), *Hizikia fusiforme* (hijiki), *Laminaria japonica* (ma-kombu) and *Sargassum fulvellum*. The Japanese have been estimated to intake wakame at 1 g/d^[76]. Brown seaweeds are known to contain many bioactive components, *i.e.*,

fucoxanthin (FX), fucoidan, vitamins, minerals, dietary fibers, proteins, ω -3 polyunsaturated fatty acids (PUFAs), polysaccharides, other carotenoids and various functional polyphenols. Fucoidan is a sulfated polysaccharide that is one of the major bioactive components in seaweed^[77,78], but we would like to focus on FX in this review. FX is a xanthophyll belonging to non-provitamin A carotenoids, constructed with an unusual allenic bond, an epoxide group, and a conjugated carbonyl group in a polyene chain^[79]. Some reports demonstrated that the FX content of *U. pinnatifida* is approximately 1.0-3.0 mg/g dry weight through one life cycle^[80,81]. It has been proven that mice convert FX into keto-carotenoids by oxidation of the secondary hydroxyl groups ($\text{FX} + \text{H}_2\text{O} \rightarrow \text{FuOH}$; $\text{FuOH} + \text{NAD}^+ \rightarrow \text{amarouciaxanthin A} + \text{NADH}$)^[79]. On the other hand, oral administration of kombu extract containing FX in humans revealed that the FuOH and the *cis*-isomer of FuOH could be found in the serum, detected by HPLC^[82].

Safety profile

FX has been proved to be safe with no side effects by single (1000 or 2000 mg/kg BW) and repeated (500 or 1000 mg/kg BW for 30 d) oral dose toxicity studies in male and female mice^[83]. In the repeated doses study, histological examination of the gonadal tissues, kidneys, liver and spleen revealed no abnormal changes^[83]. In rats, 13-wk oral subchronic toxicity studies suggested that more than 2000 mg/kg BW of microalgal FX oil induce the 50% lethal^[84].

Preclinical studies and anti-cancer mechanisms

Many studies suggested FX possesses anti-cancer potential, especially shown in colon cancer cell lines (Caco-2, DLD-1 and HT-29), liver cell lines (HepG2), prostate cancer cell lines (DU 145, LNCaP and PC-3) and urinary bladder^[85-88]. The main biomolecules involved in anti-cancer mechanism is assumed to be the biomolecules related to apoptosis and cell cycle^[89,90] and those may associate with antioxidant activity through their free radical scavenging action^[91]. Moreover, inhibition of PI3K/Akt and NF- κ B signals were reported in human cervical and breast cancer cells, respectively^[92,93].

Its metabolite fucoxanthinol (FuOH) also has inhibitory effects on cancer cell growth^[94,95], and 1,2-dimethylhydrazine-induced formation of colonic ACF in mice and AOM/DSS-induced colon carcinogenesis^[25,96]. To find new cancer prevention approaches, we investigated the combination effect of FuOH and 1 α ,25-dihydroxyvitamin D₃ (1 α ,25(OH)₂D₃), and found inhibition of cell viability and induction of apoptosis in DLD-1 and HT-29 cells^[97]. Down-regulation of PPAR γ and NF- κ B p52 were suggested to be involved in the inhibition of cell viability due to the combination of FuOH and 1 α ,25(OH)₂D₃. It has been shown that activation of PPAR γ suppresses intestinal polyp development in *Apc*-mutant mice and AOM-induced colonic ACF development in obese KK-*A*⁰ mice^[98,99].

Clinical studies

FX has been reported to provide health benefits in humans, such as improvement of obesity, reduction of inflammation, healthy triglyceride levels, and improvements in blood pressure levels^[100,101].

After daily intake of *U. pinnatifida*, FuOH is detectable in human plasma^[82]. Although metabolites of FX could be measured as a marker of exposure, effects of FX or FuOH in human carcinogenesis have not been reported to date. From the aspect of obesity-associated cancer, we here introduce one study that has been conducted to assess the effects of FX supplementation on weight loss. FX supplementation on obese patients with non-alcoholic fatty liver disease results in the improvement of liver inflammatory markers, such as alanine aminotransferase, aspartate aminotransferase, C-reactive protein, γ -glutamyltransferase (γ GT, GGT)^[101]. Of note, it has been demonstrated that increased of GGT plasma levels are associated with an increased risk of pancreatic cancer^[102,103], nevertheless GGT has no causative role itself.

It is also interesting to mention that intake of 5 g/d *U. pinnatifida* stimulated a significant 50% reduction in urinary urokinase-like plasminogen activator receptor (uPAR) proteins in postmenopausal women. uPAR, is the membrane receptor for uPA, responsible for extracellular membrane proteins degradation and PAI-1, responsible for the inhibition of plasminogen activation^[104]. Generally, uPAR is known to be higher in postmenopausal women as well as in breast cancer patients^[105]. Moreover, it has been reported that uPA and/or PAI-1 is positively correlated with poor prognosis in patients with breast cancer, *i.e.*, correlation with cancer metastatic potential^[106,107]. Thus, uPA, PAI-1 and uPAR might be used as prognostic markers for breast cancer^[108], and FX may reduce such a tumor marker.

NEOXANTHIN

Distribution and nature of Neoxanthin

Neoxanthin (NX), a non-provitamin A carotenoid, has an unusual allenic bond and a 5,6-monoepoxide as well as FX. NX is widely present in terrestrial and marine biota and the occurrence of two geometric *cis/trans* isomers is known to be species dependent^[109-111]. The 9'-*cis* form of NX (9'-*cis* NX) is mainly localized and used in the photosynthetic organs of spinach leaves and marine algae such as *Euglenophyta*. It is also used as a precursor of abscisic acid, a plant hormone^[112,113]. Whereas the all-*trans* form of NX is predominant in the petals of globeflower and yellow rose, this xanthophyll is not involved in the photosynthetic system^[111,114]. We mainly obtain the 9'-*cis* NX from leafy green vegetables. Fresh spinach contains 9'-*cis* NX around 5 mg/100 g in fresh leaf^[110]. It has been estimated that 9'-*cis* NX exists at 0.95 μ mol/L in digested fluid (9 L/d), when we ingest 100 g/d spinach.

The 5,6-monoepoxide moiety in 9'-*cis* NX is easily isomerized to 5,8-epoxide under the acidic conditions of the stomach and generates almost equal amounts of

(8'-R/S)-neochrome^[115,116]. After a 1-wk spinach intervention (3 mg 9'-*cis* NX/day), highly hydrophilic xanthophylls of 9'-*cis* NX and (8'-R and 8'-S)-neochromes appeared at a very low level in human plasma (about 1 nmol/L)^[117]. It is known that the uptake of various carotenoids by human colon cancer cells (Caco-2 cells) positively correlates with the lipophilicity of the carotenoids^[118]. The highly hydrophilic xanthophylls such as NX, FX and violaxanthin could be detected slightly in human plasma, when we intake purified forms and food matrices^[79,101,117,119-121]. Because of the poor intestinal absorption of NX, a considerable amount of ingested 9'-*cis* NX and (8'-R and 8'-S)-neochrome would be delivered to the colon, and even if absorbed in the small intestine, they would be metabolized easily.

Preclinical studies and anti-cancer mechanisms

It has been reported that both 9'-*cis* NX and all-*trans* NX possess strong potential of cell growth inhibition and apoptosis induction in human prostate cancer cells^[87,94,115,122], human colon cancer^[122-124], mouse melanoma^[122] and mouse embryonic mesenchymal cells^[125]. In addition, several researchers have reported that 9'-*cis* NX, all-*trans* NX and (8'-R/S)-neochrome have cancer preventive effects^[126], and also anti-tumor promoter functions^[70]. Moreover, induction of cell cycle arrest^[115], anti-oxidant properties^[127] and anti-obesity properties^[128] have been reported. Recently, we additionally demonstrated that 9'-*cis* NX rapidly accumulated in the mitochondria, caused mitochondria $\Delta\Psi$ loss and thereafter the release of cytochrome *c* and production of apoptosis-inducing factor in human colon cancer cells^[123]. It is regrettable that there is little information about the anti-cancer mechanisms of dietary NX in mammals, except for that described above.

Safety profile and clinical studies

No safety profile and clinical studies have been reported on 9'-*cis* NX, any -*trans* NX and (8'-R/S)-neochromes. However, epidemiological data show that higher intake of fruits and vegetables, rich in highly hydrophilic epoxyxanthophylls such as NX, is associated with a lower risk of colorectal cancer^[129,130]. Further studies are required to elucidate the clinical beneficial properties of NX.

ZEAXANTHIN/LUTEIN

Distribution and nature of zeaxanthin / lutein

Zeaxanthin (ZX) and lutein also belong to the xanthophyll family. Their unique character is that they are the only carotenoid among more than 600 species of carotenoid existing in eye tissue, especially in the retina^[131]. Lutein can be photochemically transformed to meso-ZX. They are stereoisomer of each other, differ by the location of a double bond. Lutein is abundant in egg yolk, and in dark-green leafy vegetables, such as broccoli, brussels sprouts, kale and spinach^[132]. In the human body, lutein is distributed at the skin, breasts, cervix uteri, and also found in serum in high amounts. Serum lutein and

ZX levels are reported to be around 180 and 20 ng/mL, respectively^[133]. They are assumed to play a critical role in ocular health because they act as strong anti-oxidants and filtered out high-energy blue light^[134]. Of note, no correlation between plasma concentrations of lutein/zeaxanthin and BMI or insulin resistance has been reported^[135].

Epidemiologic studies

In many papers, target organs for lutein are reported to be the eyeballs, the skin and the heart. Regarding ocular conditions, age-related macular degeneration, cataracts, and retina pigmentosa have been reported to have some correlation with lutein. Lutein also possesses a preventive function of cardiovascular diseases/stroke^[131,134,136,137].

Regarding lung cancer, some epidemiologic studies state lutein has an important cancer preventive function^[4,14]. A ten-year study of 120000 United States people revealed that lung cancer incidence was significantly reduced in those who ingested a high amount of total carotenoids, including lutein and ZX^[138]. Similar relationships were found in Fijians, when compared to the other South Pacific islands' people. Fijians intake 25 mg lutein daily on an average (200 g dark greens), whereas other 20 South Pacific countries intake less lutein in diets^[139]. Thus, there was a clear inverse association with lutein intake and lung cancer incidence.

Regarding colorectal cancer, inverse associations with dietary lutein intake have been reported^[124], and serum ZX concentration by Okuyama *et al.*^[140]. However, no association has been detected between the levels of plasma lutein and the risk of gastric cancer^[141].

Regarding skin cancer risk, the specific effects of lutein are not fully known. The only reported data is that a combination of carotenoids may protect erythema development in human skin^[142], and that may be correlated with the presence of skin cancer or precancerous lesions^[124].

Regarding breast cancer, there is some possibility for protective effects of lutein^[6,14,143]. Intake of lutein-rich foods significantly lowered the risk of premenopausal breast cancer. The Nurse's Health Study demonstrated a weak inverse association, but significant, between lutein and ZX intake and the breast cancer risk among premenopausal women^[6]. Of note, the protective effect of lutein and ZX was strongest in patients have a family history of breast cancer. Also there is a report that increasing serum levels of lutein and ZX were associated with a reduced breast cancer risk, but the trend was only marginally significant in a case-control study^[143]. There is a report comparing biopsy samples from breast cancer tissue and benign mammary tissue. In this report, increasing lutein and ZX concentrations tends to decrease the risk of breast cancer^[144]. Meanwhile, Other studies have shown that there are no differences of lutein and ZX concentrations in mammary adipose tissue between benign breast tumors and breast cancer^[145]. New York University Women's Health Study, a nested case-control prospective study, demonstrated an inverse relationship

between plasma levels of lutein, but not ZX, and risk of breast cancer^[146].

Regarding other cancers, significant inverse relations were observed for lutein and ZX in oral cavity and pharyngeal cancer^[147].

Safety profile

No toxicities or adverse reactions for intake of lutein/ZX have been reported at doses up to 40 mg/d for 2 mo^[131,148]. High doses of β -carotene supplements (> 30 mg/d) are well known to be associated with carotenodermia^[149], and the same could happen when we consume high doses of lutein and ZX. Also it has been demonstrated that lutein has no mutagenic effect in the Ames test^[150].

Preclinical study and anticancer mechanism

Lutein/ZX is thought to have a superior anti-oxidant ability to scavenge free radicals than other carotenoids. An *in vitro* study showed that lutein could quench peroxy radicals and play a guarding role against oxidative injury^[151,152]. In this experiment, a synergistic antioxidant effect was obtained with a combination of lutein and lycopene^[153]. Carotenoids also show a superb function for immune response^[154].

Lutein could also function as an anti-carcinogenic reagent, such as a modulator of cell growth and apoptosis signaling. Lutein induces cell cycle arrest in human prostate and esophageal cancer cell^[155,156]. Lutein induces apoptosis in transformed cancer cells but do not induce apoptosis in normal human mammary cells through modulating the ratio of Bcl-xL/Bax protein expression^[157]. Meanwhile, ZX, structural isomer of lutein, induced cell cycle arrest in human breast cancer cells^[158]. Lutein stimulates some genes involved in T-cell transformations activated by antigens, cytokines and mitogens^[159]. Lutein interacts with carcinogens such as 1-nitropyrene and aflatoxin B1, and lowered its carcinogenic activity^[150,160]. In a recent report, female BALB/c mice were fed a diet containing lutein for 14 d, and then inoculated with 0 to 2.5×10^3 mammary tumor cells. The results demonstrated that 0.002% and 0.02% lutein lowered both mammary tumor incidence and tumor growth^[161].

FUTURE ASPECTS

The versatile functions of xanthophylls have shown great potential for the prevention of metabolic syndrome and cancers, both *in vitro* and *in vivo*. Xanthophylls have been verified as safe with no side events, and several xanthophylls provide other health benefits, including improvements in inflammation, dyslipidemia, hypertension and liver function, as shown in this review. The accumulated evidence indicates the functionality of xanthophylls as anti-obesity and anti-insulin-resistance functional foods, implying that xanthophylls could be useful in preventing obesity-associated cancer.

The chemical synthesis of each xanthophyll is not impossible, but it may be very expensive. However, the

promising results obtained from *in vivo* studies encourage researchers to undertake more clinical studies in humans. We have some information about xanthophylls trials, and we should further promote human clinical studies to obtain information about the adequate dosage of xanthophylls needed to prevent cancers.

REFERENCES

- Fujii G, Yamamoto M, Takahashi M, Mutoh M. Role of adipocytokines in colorectal carcinogenesis. *Curr Res in Cancer* 2011; **5**: 39-48
- Ishino K, Mutoh M, Totsuka Y, Nakagama H. Metabolic syndrome: A novel high-risk state for colorectal cancer. *Cancer Lett* 2013; **334**: 56-61 [PMID: 23085010 DOI: 10.1016/j.canlet.2012.10.012]
- Eastwood MA. Interaction of dietary antioxidants in vivo: how fruit and vegetables prevent disease? *QJM* 1999; **92**: 527-530 [PMID: 10627873 DOI: 10.1093/qjmed/92.9.527]
- Holick CN, Michaud DS, Stolzenberg-Solomon R, Mayne ST, Pietinen P, Taylor PR, Virtamo J, Albanes D. Dietary carotenoids, serum beta-carotene, and retinol and risk of lung cancer in the alpha-tocopherol, beta-carotene cohort study. *Am J Epidemiol* 2002; **156**: 536-547 [PMID: 12226001 DOI: 10.1093/aje/kwf072]
- Rock CL. Carotenoid update. *J Am Diet Assoc* 2003; **103**: 423-425 [PMID: 12668998 DOI: 10.1016/S0002-8223(03)00164-0]
- Zhang S, Hunter DJ, Forman MR, Rosner BA, Speizer FE, Colditz GA, Manson JE, Hankinson SE, Willett WC. Dietary carotenoids and vitamins A, C, and E and risk of breast cancer. *J Natl Cancer Inst* 1999; **91**: 547-556 [PMID: 10088626 DOI: 10.1093/jnci/91.6.547]
- Sporn MB, Suh N. Chemoprevention: an essential approach to controlling cancer. *Nat Rev Cancer* 2002; **2**: 537-543 [PMID: 12094240 DOI: 10.1038/nrc844]
- Ferguson LR, Schlothauer RC. The potential role of nutritional genomics tools in validating high health foods for cancer control: broccoli as example. *Mol Nutr Food Res* 2012; **56**: 126-146 [PMID: 22147677 DOI: 10.1002/mnfr.201100507]
- Temraz S, Mukherji D, Shamseddine A. Potential targets for colorectal cancer prevention. *Int J Mol Sci* 2013; **14**: 17279-17303 [PMID: 23975167 DOI: 10.3390/ijms140917279]
- Heinonen OP, Albanes D. The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers. The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group. *N Engl J Med* 1994; **330**: 1029-1035 [PMID: 8127329 DOI: 10.1056/NEJM199404143301501]
- Hennekens CH, Buring JE, Manson JE, Stampfer M, Rosner B, Cook NR, Belanger C, LaMotte F, Gaziano JM, Ridker PM, Willett W, Peto R. Lack of effect of long-term supplementation with beta carotene on the incidence of malignant neoplasms and cardiovascular disease. *N Engl J Med* 1996; **334**: 1145-1149 [PMID: 8602179 DOI: 10.1056/NEJM199605023341801]
- Omenn GS, Goodman GE, Thornquist MD, Balmes J, Cullen MR, Glass A, Keogh JP, Meyskens FL, Valanis B, Williams JH, Barnhart S, Hammar S. Effects of a combination of beta carotene and vitamin A on lung cancer and cardiovascular disease. *N Engl J Med* 1996; **334**: 1150-1155 [PMID: 8602180 DOI: 10.1056/NEJM199605023341802]
- Ambati RR, Phang SM, Ravi S, Aswathanarayana RG. Astaxanthin: sources, extraction, stability, biological activities and its commercial applications--a review. *Mar Drugs* 2014; **12**: 128-152 [PMID: 24402174 DOI: 10.3390/md12010128]
- Tanaka T, Shnimizu M, Moriwaki H. Cancer chemoprevention by carotenoids. *Molecules* 2012; **17**: 3202-3242 [PMID: 22418926 DOI: 10.3390/molecules17033202]
- Kidd P. Astaxanthin, cell membrane nutrient with diverse clinical benefits and anti-aging potential. *Altern Med Rev* 2011; **16**: 355-364 [PMID: 22214255]
- Coral-Hinojosa GN, Ytrestøl T, Ruyter B, Bjerkeng B. Plasma appearance of unesterified astaxanthin geometrical E/Z and optical R/S isomers in men given single doses of a mixture of optical 3 and 3'R/S isomers of astaxanthin fatty acyl diesters. *Comp Biochem Physiol C Toxicol Pharmacol* 2004; **139**: 99-110 [PMID: 15556071 DOI: 10.1016/j.cca.2004.09.011]
- Fassett RG, Coombes JS. Astaxanthin in cardiovascular health and disease. *Molecules* 2012; **17**: 2030-2048 [PMID: 22349894 DOI: 10.3390/molecules17022030]
- Stewart JS, Lignell A, Pettersson A, Elfving E, Soni MG. Safety assessment of astaxanthin-rich microalgae biomass: Acute and subchronic toxicity studies in rats. *Food Chem Toxicol* 2008; **46**: 3030-3036 [PMID: 18588938 DOI: 10.1016/j.fct.2008.05.038]
- Spiller GA, Dewell A. Safety of an astaxanthin-rich Haematococcus pluvialis algal extract: a randomized clinical trial. *J Med Food* 2003; **6**: 51-56 [PMID: 12804020 DOI: 10.1089/109662003765184741]
- Yoshida H, Yanai H, Ito K, Tomono Y, Koikeda T, Tsukahara H, Tada N. Administration of natural astaxanthin increases serum HDL-cholesterol and adiponectin in subjects with mild hyperlipidemia. *Atherosclerosis* 2010; **209**: 520-523 [PMID: 19892350 DOI: 10.1016/j.atherosclerosis.2009.10.012]
- Tanaka T, Morishita Y, Suzui M, Kojima T, Okumura A, Mori H. Chemoprevention of mouse urinary bladder carcinogenesis by the naturally occurring carotenoid astaxanthin. *Carcinogenesis* 1994; **15**: 15-19 [PMID: 8293542 DOI: 10.1093/carcin/15.1.15]
- Tanaka T, Makita H, Ohnishi M, Mori H, Satoh K, Hara A. Chemoprevention of rat oral carcinogenesis by naturally occurring xanthophylls, astaxanthin and canthaxanthin. *Cancer Res* 1995; **55**: 4059-4064 [PMID: 7664280]
- Kavitha K, Thiagarajan P, Rathna Nandhini J, Mishra R, Nagini S. Chemopreventive effects of diverse dietary phytochemicals against DMBA-induced hamster buccal pouch carcinogenesis via the induction of Nrf2-mediated cytoprotective antioxidant, detoxification, and DNA repair enzymes. *Biochimie* 2013; **95**: 1629-1639 [PMID: 23707664 DOI: 10.1016/j.biochi.2013.05.004]
- Tanaka T, Kawamori T, Ohnishi M, Makita H, Mori H, Satoh K, Hara A. Suppression of azoxymethane-induced rat colon carcinogenesis by dietary administration of naturally occurring xanthophylls astaxanthin and canthaxanthin during the postinitiation phase. *Carcinogenesis* 1995; **16**: 2957-2963 [PMID: 8603470 DOI: 10.1093/carcin/16.12.2957]
- Yasui Y, Hosokawa M, Mikami N, Miyashita K, Tanaka T. Dietary astaxanthin inhibits colitis and colitis-associated colon carcinogenesis in mice via modulation of the inflammatory cytokines. *Chem Biol Interact* 2011; **193**: 79-87 [PMID: 21621527 DOI: 10.1016/j.cbi.2011.05.006]
- Kavitha K, Kowshik J, Kishore TK, Baba AB, Nagini S. Astaxanthin inhibits NF- κ B and Wnt/ β -catenin signaling pathways via inactivation of Erk/MAPK and PI3K/Akt to induce intrinsic apoptosis in a hamster model of oral cancer. *Biochim Biophys Acta* 2013; **1830**: 4433-4444 [PMID: 23726989 DOI: 10.1016/j.bbagen.2013.05.032]
- Gradelet S, Le Bon AM, Bergès R, Suschetet M, Astorg P. Dietary carotenoids inhibit aflatoxin B1-induced liver preneoplastic foci and DNA damage in the rat: role of the modulation of aflatoxin B1 metabolism. *Carcinogenesis* 1998; **19**: 403-411 [PMID: 9525273 DOI: 10.1093/carcin/19.3.403]
- Chew BP, Park JS, Wong MW, Wong TS. A comparison of the anticancer activities of dietary beta-carotene, canthaxanthin and astaxanthin in mice in vivo. *Anticancer Res* 1999; **19**: 1849-1853 [PMID: 10470126]
- Kurihara H, Koda H, Asami S, Kiso Y, Tanaka T. Contribution of the antioxidative property of astaxanthin to its protective effect on the promotion of cancer metastasis in mice treated with restraint stress. *Life Sci* 2002; **70**: 2509-2520

- [PMID: 12173414]
- 30 **Kozuki Y**, Miura Y, Yagasaki K. Inhibitory effects of carotenoids on the invasion of rat ascites hepatoma cells in culture. *Cancer Lett* 2000; **151**: 111-115 [PMID: 10766430 DOI: 10.1016/S0304-3835(99)00418-8]
 - 31 **Zhang X**, Zhao WE, Hu L, Zhao L, Huang J. Carotenoids inhibit proliferation and regulate expression of peroxisome proliferators-activated receptor gamma (PPAR γ) in K562 cancer cells. *Arch Biochem Biophys* 2011; **512**: 96-106 [PMID: 21620794 DOI: 10.1016/j.abb.2011.05.004]
 - 32 **Hix LM**, Lockwood SF, Bertram JS. Bioactive carotenoids: potent antioxidants and regulators of gene expression. *Redox Rep* 2004; **9**: 181-191 [PMID: 15479561]
 - 33 **Vine AL**, Bertram JS. Upregulation of connexin 43 by retinoids but not by non-provitamin A carotenoids requires RARs. *Nutr Cancer* 2005; **52**: 105-113 [PMID: 16091010 DOI: 10.1207/s15327914nc5201_13]
 - 34 **Saw CL**, Yang AY, Guo Y, Kong AN. Astaxanthin and omega-3 fatty acids individually and in combination protect against oxidative stress via the Nrf2-ARE pathway. *Food Chem Toxicol* 2013; **62**: 869-875 [PMID: 24157545 DOI: 10.1016/j.fct.2013.10.023]
 - 35 **Wolf AM**, Asoh S, Hiranuma H, Ohsawa I, Iio K, Satou A, Ishikura M, Ohta S. Astaxanthin protects mitochondrial redox state and functional integrity against oxidative stress. *J Nutr Biochem* 2010; **21**: 381-389 [PMID: 19423317 DOI: 10.1016/j.jnutbio.2009.01.011]
 - 36 **DeNicola GM**, Karreth FA, Humpton TJ, Gopinathan A, Wei C, Frese K, Mangal D, Yu KH, Yeo CJ, Calhoun ES, Scrimieri F, Winter JM, Hruban RH, Iacobuzio-Donahue C, Kern SE, Blair IA, Tuveson DA. Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. *Nature* 2011; **475**: 106-109 [PMID: 21734707 DOI: 10.1038/nature10189]
 - 37 **Satoh H**, Moriguchi T, Takai J, Ebina M, Yamamoto M. Nrf2 prevents initiation but accelerates progression through the Kras signaling pathway during lung carcinogenesis. *Cancer Res* 2013; **73**: 4158-4168 [PMID: 23610445 DOI: 10.1158/0008-5472.CAN-12-4499]
 - 38 **Nakao R**, Nelson OL, Park JS, Mathison BD, Thompson PA, Chew BP. Effect of dietary astaxanthin at different stages of mammary tumor initiation in BALB/c mice. *Anticancer Res* 2010; **30**: 2171-2175 [PMID: 20651366]
 - 39 **Karppi J**, Rissanen TH, Nyssönen K, Kaikkonen J, Olsson AG, Voutilainen S, Salonen JT. Effects of astaxanthin supplementation on lipid peroxidation. *Int J Vitam Nutr Res* 2007; **77**: 3-11 [PMID: 17685090]
 - 40 **Choi HD**, Kim JH, Chang MJ, Kyu-Youn Y, Shin WG. Effects of astaxanthin on oxidative stress in overweight and obese adults. *Phytother Res* 2011; **25**: 1813-1818 [PMID: 21480416 DOI: 10.1002/ptr.3494]
 - 41 **Park JS**, Chyun JH, Kim YK, Line LL, Chew BP. Astaxanthin decreased oxidative stress and inflammation and enhanced immune response in humans. *Nutr Metab (Lond)* 2010; **7**: 18 [PMID: 20205737 DOI: 10.1186/1743-7075-7-18]
 - 42 **Parisi V**, Tedeschi M, Gallinaro G, Varano M, Saviano S, Piermarocchi S. Carotenoids and antioxidants in age-related maculopathy Italian study: multifocal electroretinogram modifications after 1 year. *Ophthalmology* 2008; **115**: 324-333. e2 [PMID: 17716735 DOI: 10.1016/j.ophtha.2007.05.029]
 - 43 **Piermarocchi S**, Saviano S, Parisi V, Tedeschi M, Panozzo G, Scarpa G, Boschi G, Lo Giudice G. Carotenoids in Age-related Maculopathy Italian Study (CARMIS): two-year results of a randomized study. *Eur J Ophthalmol* 2012; **22**: 216-225 [PMID: 22009916 DOI: 10.5301/ejo.5000069]
 - 44 **Katagiri M**, Satoh A, Tsuji S, Shirasawa T. Effects of astaxanthin-rich Haematococcus pluvialis extract on cognitive function: a randomised, double-blind, placebo-controlled study. *J Clin Biochem Nutr* 2012; **51**: 102-107 [PMID: 22962526 DOI: 10.3164/jcfn.11-00017]
 - 45 **Yano M**, Kato M, Ikoma Y, Kawasaki A, Fukazawa Y, Sugiura M, Matsumoto H, Oohara Y, Nagao A, Ogawa K. Quantitation of carotenoids in raw and processed fruits in Japan. *Food Sci Technol Res* 2005; **11**: 13-18 [DOI: 10.3136/fstr.11.13]
 - 46 **Holden JM**, Eldridge AL, Beecher GR, Buzzard IM, Bhagwat S, Davis CS, Douglass LW, Gebhardt S, Haytowitz D, Schakel S. Carotenoid content of US foods: An update of the database. *J Food Comp Anal* 1999; **12**: 169-196 [DOI: 10.1006/jfca.1999.0827]
 - 47 **Wahlqvist ML**, Wattanapenpaiboon N, Macrae FA, Lambert JR, MacLennan R, Hsu-Hage BH. Changes in serum carotenoids in subjects with colorectal adenomas after 24 mo of beta-carotene supplementation. Australian Polyp Prevention Project Investigators. *Am J Clin Nutr* 1994; **60**: 936-943 [PMID: 7985637]
 - 48 **Sugiura M**, Ogawa K, Yano M. Absorption, storage and distribution of β -cryptoxanthin in rat after chronic administration of Satsuma mandarin (*Citrus unshiu* MARC.) juice. *Biol Pharm Bull* 2013; **36**: 147-151 [PMID: 23302648 DOI: 10.1248/bpb.b12-00836]
 - 49 **Sugiura M**, Kato M, Matsumoto H, Nagao A, Yano M. Serum concentration of beta-cryptoxanthin in Japan reflects the frequency of Satsuma mandarin (*Citrus unshiu* Marc.) consumption. *J Health Sci* 2002; **48**: 350-353 [DOI: 10.1248/jhs.48.350]
 - 50 **Sugiura M**, Matsumoto H, Kato M, Ikoma Y, Yano M, Nagao A. Seasonal changes in the relationship between serum concentration of beta-cryptoxanthin and serum lipid levels. *J Nutr Sci Vitaminol (Tokyo)* 2004; **50**: 410-415 [PMID: 15895516]
 - 51 **Sugiura M**, Matsumoto H, Kato M, Ikoma Y, Yano M, Nagao A. Multiple linear regression analysis of the seasonal changes in the serum concentration of beta-cryptoxanthin. *J Nutr Sci Vitaminol (Tokyo)* 2004; **50**: 196-202 [PMID: 15386932]
 - 52 **Canfield LM**, Clandinin MT, Davies DP, Fernandez MC, Jackson J, Hawkes J, Goldman WJ, Pramuk K, Reyes H, Sablan B, Sonobe T, Bo X. Multinational study of major breast milk carotenoids of healthy mothers. *Eur J Nutr* 2003; **42**: 133-141 [PMID: 12811470]
 - 53 **Montonen J**, Knekt P, Järvinen R, Reunanen A. Dietary antioxidant intake and risk of type 2 diabetes. *Diabetes Care* 2004; **27**: 362-366 [PMID: 14747214]
 - 54 **Cerhan JR**, Saag KG, Merlino LA, Mikuls TR, Criswell LA. Antioxidant micronutrients and risk of rheumatoid arthritis in a cohort of older women. *Am J Epidemiol* 2003; **157**: 345-354 [PMID: 12578805 DOI: 10.1093/aje/kwf205]
 - 55 **Liu C**, Bronson RT, Russell RM, Wang XD. β -Cryptoxanthin supplementation prevents cigarette smoke-induced lung inflammation, oxidative damage, and squamous metaplasia in ferrets. *Cancer Prev Res (Phila)* 2011; **4**: 1255-1266 [PMID: 21421799 DOI: 10.1158/1940-6207.CAPR-10-0384]
 - 56 **Männistö S**, Smith-Warner SA, Spiegelman D, Albanes D, Anderson K, van den Brandt PA, Cerhan JR, Colditz G, Feskanih D, Freudenheim JL, Giovannucci E, Goldbohm RA, Graham S, Miller AB, Rohan TE, Virtamo J, Willett WC, Hunter DJ. Dietary carotenoids and risk of lung cancer in a pooled analysis of seven cohort studies. *Cancer Epidemiol Biomarkers Prev* 2004; **13**: 40-48 [PMID: 14744731 DOI: 10.1158/1055-9965.EPI-038-3]
 - 57 **De Stefani E**, Brennan P, Boffetta P, Ronco AL, Mendilaharsu M, Deneo-Pellegrini H. Vegetables, fruits, related dietary antioxidants, and risk of squamous cell carcinoma of the esophagus: a case-control study in Uruguay. *Nutr Cancer* 2000; **38**: 23-29 [PMID: 11341040 DOI: 10.1207/S15327914NC381_4]
 - 58 **Zeegers MP**, Goldbohm RA, van den Brandt PA. Are retinol, vitamin C, vitamin E, folate and carotenoids intake associated with bladder cancer risk? Results from the Netherlands Cohort Study. *Br J Cancer* 2001; **85**: 977-983 [PMID: 11592769 DOI: 10.1054/bjoc.2001.1968]

- 59 **Goodman MT**, McDuffie K, Hernandez B, Hankin JH, Wilkens LR, Franke AA, Kolonel LN, Kuypers J, Kiviat N, Bertram CC, Kessel B, Sunoo C, Nakamura J, Killeen J. The Association of Plasma Micronutrients with the Risk of Cervical Atypical Squamous Cells of Undetermined Significance (ASCUS). *Asian Pac J Cancer Prev* 2000; **1**: 337-345 [PMID: 12716311]
- 60 **Nishino H**, Murakoshi M, Satomi Y. Health promotion by antioxidants. *Functional Foods in Health and Disease* 2011; **1**: 574-581. Available from: URL: <http://www.functionalfood-science.net/files/48097641.pdf>
- 61 Opinion of the scientific panel on additives and products or substances used in animal feed on the request from the commission on the safety of use of colouring agents in animal nutrition. *The EFSA Journal* 2006; **386**: 1-40. Available from: URL: <http://www.efsa.europa.eu/en/efsajournal/doc/320.pdf>
- 62 **Miyazawa K**, Miyamoto S, Suzuki R, Yasui Y, Ikeda R, Kohno H, Yano M, Tanaka T, Hata K, Suzuki K. Dietary beta-cryptoxanthin inhibits N-butyl-N-(4-hydroxybutyl)nitrosamine-induced urinary bladder carcinogenesis in male ICR mice. *Oncol Rep* 2007; **17**: 297-304 [PMID: 17203164 DOI: 10.3892/or.17.2.297]
- 63 **Sugiura M**, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. Serum carotenoid concentrations are inversely associated with serum aminotransferases in hyperglycemic subjects. *Diabetes Res Clin Pract* 2006; **71**: 82-91 [PMID: 16005096 DOI: 10.1016/j.atherosclerosis.2005.04.006]
- 64 **Sugiura M**, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. High serum carotenoids are inversely associated with serum gamma-glutamyltransferase in alcohol drinkers within normal liver function. *J Epidemiol* 2005; **15**: 180-186 [PMID: 16195638]
- 65 **Nakamura M**, Sugiura M, Aoki N. High beta-carotene and beta-cryptoxanthin are associated with low pulse wave velocity. *Atherosclerosis* 2006; **184**: 363-369 [PMID: 15936762]
- 66 **Sugiura M**, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. The homeostasis model assessment-insulin resistance index is inversely associated with serum carotenoids in non-diabetic subjects. *J Epidemiol* 2006; **16**: 71-78 [PMID: 16537987 DOI: 10.2188/jea.16.71]
- 67 **Sugiura M**, Nakamura M, Ogawa K, Ikoma Y, Matsumoto H, Ando F, Shimokata H, Yano M. Associations of serum carotenoid concentrations with the metabolic syndrome: interaction with smoking. *Br J Nutr* 2008; **100**: 1297-1306 [PMID: 18445303 DOI: 10.1017/S0007114508978302]
- 68 **Sugiura M**, Nakamura M, Ogawa K, Ikoma Y, Ando F, Shimokata H, Yano M. Dietary patterns of antioxidant vitamin and carotenoid intake associated with bone mineral density: findings from post-menopausal Japanese female subjects. *Osteoporos Int* 2011; **22**: 143-152 [PMID: 20480147 DOI: 10.1007/s00198-010-1239-9]
- 69 **Lorenzo Y**, Azqueta A, Luna L, Bonilla F, Domínguez G, Collins AR. The carotenoid beta-cryptoxanthin stimulates the repair of DNA oxidation damage in addition to acting as an antioxidant in human cells. *Carcinogenesis* 2009; **30**: 308-314 [PMID: 19056931 DOI: 10.1093/carcin/bgn270]
- 70 **Uchiyama S**, Yamaguchi M. Beta-cryptoxanthin stimulates apoptotic cell death and suppresses cell function in osteoclastic cells: change in their related gene expression. *J Cell Biochem* 2006; **98**: 1185-1195 [PMID: 16514646 DOI: 10.1002/jcb.20824]
- 71 **Nishino H**, Tokuda H, Murakoshi M, Satomi Y, Masuda M, Onozuka M, Yamaguchi S, Takayasu J, Tsuruta J, Okuda M, Khachik F, Narisawa T, Takasuka N, Yano M. Cancer prevention by natural carotenoids. *Biofactors* 2000; **13**: 89-94 [PMID: 11237205]
- 72 **Tsushima M**, Maoka T, Katsuyama M, Kozuka M, Matsuno T, Tokuda H, Nishino H, Iwashima A. Inhibitory effect of natural carotenoids on Epstein-Barr virus activation activity of a tumor promoter in Raji cells. A screening study for anti-tumor promoters. *Biol Pharm Bull* 1995; **18**: 227-233 [PMID: 7742789]
- 73 **Narisawa T**, Fukaura Y, Oshima S, Inakuma T, Yano M, Nishino H. Chemoprevention by the oxygenated carotenoid beta-cryptoxanthin of N-methylnitrosourea-induced colon carcinogenesis in F344 rats. *Jpn J Cancer Res* 1999; **90**: 1061-1065 [PMID: 10595732]
- 74 **Iskandar AR**, Liu C, Smith DE, Hu KQ, Choi SW, Ausman LM, Wang XD. β -cryptoxanthin restores nicotine-reduced lung SIRT1 to normal levels and inhibits nicotine-promoted lung tumorigenesis and emphysema in A/J mice. *Cancer Prev Res (Phila)* 2013; **6**: 309-320 [PMID: 23275008 DOI: 10.1158/1940-6207.CAPR-12-0368]
- 75 **Iwamoto M**, Imai K, Ohta H, Shirouchi B, Sato M. Supplementation of highly concentrated β -cryptoxanthin in a satsuma mandarin beverage improves adipocytokine profiles in obese Japanese women. *Lipids Health Dis* 2012; **11**: 52 [PMID: 22584034 DOI: 10.1186/1476-511X-11-52]
- 76 **Ministry of Health**. Labour and Welfare Japan (2004) The National Nutrition Survey in Japan [in Japanese]. Tokyo, Japan: Dai-ichi shuppan, 2002
- 77 **Cumashi A**, Ushakova NA, Preobrazhenskaya ME, D'Incecco A, Piccoli A, Totani L, Tinari N, Morozevich GE, Berman AE, Bilan MI, Usov AI, Ustyuzhanina NE, Grachev AA, Sanderson CJ, Kelly M, Rabinovich GA, Iacobelli S, Nifantiev NE. A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. *Glycobiology* 2007; **17**: 541-552 [PMID: 17296677 DOI: 10.1093/glycob/cwm014]
- 78 **Shibata H**, Iimuro M, Uchiya N, Kawamori T, Nagaoka M, Ueyama S, Hashimoto S, Yokokura T, Sugimura T, Wakabayashi K. Preventive effects of Cladosiphon fucoidan against *Helicobacter pylori* infection in Mongolian gerbils. *Helicobacter* 2003; **8**: 59-65 [PMID: 12603617 DOI: 10.1046/j.1523-5378.2003.00124.x]
- 79 **Yonekura L**, Kobayashi M, Terasaki M, Nagao A. Keto-carotenoids are the major metabolites of dietary lutein and fucoxanthin in mouse tissues. *J Nutr* 2010; **140**: 1824-1831 [PMID: 20739451 DOI: 10.3945/jn.110.126466]
- 80 **Campbell SJ**, Bité JS, Burrige TR. Seasonal patterns in the photosynthetic capacity, tissue pigment and nutrient content of different developmental stages of *Undaria pinnatifida* (Phaeophyta: Laminariales) in port phillip bay, south-eastern Australia. *Bot Mar* 1999; **42**: 231-242 [DOI: 10.1515/BOT.1999.027]
- 81 **Terasaki M**, Narayan B, Kamogawa H, Nomura M, Stephen NM, Kawagoe C, Hosokawa M, Miyashita K. Carotenoid profile of edible Japanese seaweeds: An improved HPLC method for separation of major carotenoids. *J Aquatic Food Prod Tech* 2012; **21**: 468-479 [DOI: 10.1080/10498850.2011.610025]
- 82 **Hashimoto T**, Ozaki Y, Mizuno M, Yoshida M, Nishitani Y, Azuma T, Komoto A, Maoka T, Tanino Y, Kanazawa K. Pharmacokinetics of fucoxanthinol in human plasma after the oral administration of kombu extract. *Br J Nutr* 2012; **107**: 1566-1569 [PMID: 21920061 DOI: 10.1017/S0007114511004879]
- 83 **Beppu F**, Niwano Y, Tsukui T, Hosokawa M, Miyashita K. Single and repeated oral dose toxicity study of fucoxanthin (FX), a marine carotenoid, in mice. *J Toxicol Sci* 2009; **34**: 501-510 [PMID: 19797858 DOI: 10.2131/jts.34.501]
- 84 **Iio K**, Okada Y, Ishikura M. [Single and 13-week oral toxicity study of fucoxanthin oil from microalgae in rats]. *Shokuhin Eiseigaku Zasshi* 2011; **52**: 183-189 [PMID: 21720124 DOI: <http://dx.odi.org/10.3358/shokueishi.52.183>]
- 85 **Hosokawa M**, Kudo M, Maeda H, Kohno H, Tanaka T, Miyashita K. Fucoxanthin induces apoptosis and enhances the antiproliferative effect of the PPAR γ ligand, troglitazone, on colon cancer cells. *Biochim Biophys Acta* 2004;

- 1675: 113-119 [PMID: 15535974]
- 86 **Das SK**, Hashimoto T, Kanazawa K. Growth inhibition of human hepatic carcinoma HepG2 cells by fucoxanthin is associated with down-regulation of cyclin D. *Biochim Biophys Acta* 2008; **1780**: 743-749 [PMID: 18230364 DOI: 10.1016/j.bbagen.2008.01.003]
- 87 **Kotake-Nara E**, Kushiro M, Zhang H, Sugawara T, Miyashita K, Nagao A. Carotenoids affect proliferation of human prostate cancer cells. *J Nutr* 2001; **131**: 3303-3306 [PMID: 11739884]
- 88 **Zhang Z**, Zhang P, Hamada M, Takahashi S, Xing G, Liu J, Sugiura N. Potential chemoprevention effect of dietary fucoxanthin on urinary bladder cancer EJ-1 cell line. *Oncol Rep* 2008; **20**: 1099-1103 [PMID: 18949407 DOI: 10.3892/or_00000115]
- 89 **Miyashita K**, Nishikawa S, Beppu F, Tsukui T, Abe M, Hosokawa M. The allenic carotenoid fucoxanthin, a novel marine nutraceutical from brown seaweeds. *J Sci Food Agric* 2011; **91**: 1166-1174 [PMID: 21433011 DOI: 10.1002/jsfa.4353]
- 90 **Liu CL**, Huang YS, Hosokawa M, Miyashita K, Hu ML. Inhibition of proliferation of a hepatoma cell line by fucoxanthin in relation to cell cycle arrest and enhanced gap junctional intercellular communication. *Chem Biol Interact* 2009; **182**: 165-172 [PMID: 19737546 DOI: 10.1016/j.cbi.2009.08.017]
- 91 **Liu CL**, Chiu YT, Hu ML. Fucoxanthin enhances HO-1 and NQO1 expression in murine hepatic BNL CL.2 cells through activation of the Nrf2/ARE system partially by its pro-oxidant activity. *J Agric Food Chem* 2011; **59**: 11344-11351 [PMID: 21919437 DOI: 10.1021/jf2029785]
- 92 **Ye G**, Lu Q, Zhao W, Du D, Jin L, Liu Y. Fucoxanthin induces apoptosis in human cervical cancer cell line HeLa via PI3K/Akt pathway. *Tumour Biol* 2014; **35**: 11261-11267 [PMID: 25113250 DOI: 10.1007/s13277-014-2337-7]
- 93 **Rwigemera A**, Mamelona J, Martin LJ. Inhibitory effects of fucoxanthinol on the viability of human breast cancer cell lines MCF-7 and MDA-MB-231 are correlated with modulation of the NF-kappaB pathway. *Cell Biol Toxicol* 2014; **30**: 157-167 [PMID: 24760606 DOI: 10.1007/s10565-014-9277-2]
- 94 **Kotake-Nara E**, Asai A, Nagao A. Neoxanthin and fucoxanthin induce apoptosis in PC-3 human prostate cancer cells. *Cancer Lett* 2005; **220**: 75-84 [PMID: 15737690 DOI: 10.1016/j.canlet.2004.07.048]
- 95 **Tafuku S**, Ishikawa C, Yasumoto T, Mori N. Anti-neoplastic effects of fucoxanthin and its deacetylated product, fucoxanthinol, on Burkitt's and Hodgkin's lymphoma cells. *Oncol Rep* 2012; **28**: 1512-1518 [PMID: 22859062 DOI: 10.3892/or.2012.1947]
- 96 **Kim JM**, Araki S, Kim DJ, Park CB, Takasuka N, Baba-Toriyama H, Ota T, Nir Z, Khachik F, Shimidzu N, Tanaka Y, Osawa T, Uraji T, Murakoshi M, Nishino H, Tsuda H. Chemopreventive effects of carotenoids and curcumins on mouse colon carcinogenesis after 1,2-dimethylhydrazine initiation. *Carcinogenesis* 1998; **19**: 81-85 [PMID: 9472697 DOI: 10.1093/carcin/19.1.81]
- 97 **Terasaki M**, Nagao A, Maeda H, Miyashita K, Masuda S. Combined antiproliferative effect of dietary PPAR γ suppressing lipids fucoxanthinol and 1 α ,25-dihydroxyvitamin D3 in human colon cancer cells. (Proceeding of The Japanese Society for Carotenoid Research) *Carotenoid Science* 2012; **17**: 40-43
- 98 **Mutoh M**, Niho N, Wakabayashi K. Concomitant suppression of hyperlipidemia and intestinal polyp formation by increasing lipoprotein lipase activity in Apc-deficient mice. *Biol Chem* 2006; **387**: 381-385 [PMID: 16606335 DOI: 10.1515/BC.2006.051]
- 99 **Ueno T**, Teraoka N, Takasu S, Nakano K, Takahashi M, Yamamoto M, Fujii G, Komiya M, Yanaka A, Wakabayashi K, Mutoh M. Suppressive effect of pioglitazone, a PPAR gamma ligand, on azoxymethane-induced colon aberrant crypt foci in KK-Ay mice. *Asian Pac J Cancer Prev* 2012; **13**: 4067-4073 [PMID: 23098518]
- 100 **Maeda H**, Hosokawa M, Sashima T, Funayama K, Miyashita K. Effect of medium-chain triacylglycerols on anti-obesity effect of fucoxanthin. *J Oleo Sci* 2007; **56**: 615-621 [PMID: 17992001 DOI: 10.5650/jos.56.615]
- 101 **Abidov M**, Ramazanov Z, Seifulla R, Grachev S. The effects of Xanthigen in the weight management of obese premenopausal women with non-alcoholic fatty liver disease and normal liver fat. *Diabetes Obes Metab* 2010; **12**: 72-81 [PMID: 19840063 DOI: 10.1111/j.1463-1326.2009.01132.x]
- 102 **Penn R**, Worthington DJ. Is serum gamma-glutamyltransferase a misleading test? *Br Med J (Clin Res Ed)* 1983; **286**: 531-535 [PMID: 6130816]
- 103 **Hori M**, Takahashi M, Hiraoka N, Yamaji T, Mutoh M, Ishigamori R, Furuta K, Okusaka T, Shimada K, Kosuge T, Kanai Y, Nakagama H. Association of pancreatic fatty infiltration with pancreatic ductal adenocarcinoma. *Clin Transl Gastroenterol* 2014; **5**: e53 [PMID: 24622469 DOI: 10.1038/ctg.2014.5]
- 104 **Eden G**, Archinti M, Furlan F, Murphy R, Degryse B. The urokinase receptor interactome. *Curr Pharm Des* 2011; **17**: 1874-1889 [PMID: 21711237 DOI: 10.2174/138161211796718215]
- 105 **Teas J**, Vena S, Cone DL, Irhimeh M. The consumption of seaweed as a protective factor in the etiology of breast cancer: proof of principle. *J Appl Phycol* 2013; **25**: 771-779 [PMID: 23678231 DOI: 10.1007/s10811-012-9931-0]
- 106 **Jänicke F**, Prechtel A, Thomssen C, Harbeck N, Meisner C, Untch M, Sweep CG, Selbmann HK, Graeff H, Schmitt M. Randomized adjuvant chemotherapy trial in high-risk, lymph node-negative breast cancer patients identified by urokinase-type plasminogen activator and plasminogen activator inhibitor type 1. *J Natl Cancer Inst* 2001; **93**: 913-920 [PMID: 11416112 DOI: 10.1093/jnci/93.12.913]
- 107 **Look MP**, van Putten WL, Duffy MJ, Harbeck N, Christensen IJ, Thomssen C, Kates R, Spyrtos F, Fernö M, Eppenberger-Castori S, Sweep CG, Ulm K, Peyrat JP, Martin PM, Magdelenat H, Brünner N, Duggan C, Lisboa BW, Bendahl PO, Quillien V, Daver A, Ricolleau G, Meijer-van Gelder ME, Manders P, Fiets WE, Blankenstein MA, Brøt P, Romain S, Daxenbichler G, Windbichler G, Cufer T, Borstnar S, Kueng W, Beex LV, Klijn JG, O'Higgins N, Eppenberger U, Jänicke F, Schmitt M, Foekens JA. Pooled analysis of prognostic impact of urokinase-type plasminogen activator and its inhibitor PAI-1 in 8377 breast cancer patients. *J Natl Cancer Inst* 2002; **94**: 116-128 [PMID: 11792750 DOI: 10.1093/jnci/94.2.116]
- 108 **Harris L**, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, Somerfield MR, Hayes DF, Bast RC. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. *J Clin Oncol* 2007; **25**: 5287-5312 [PMID: 17954709]
- 109 **Goodwin TW**. The biochemistry of the carotenoids, Vol. I. Plants. 2nd ed. New York, NY: Chapman and Hall, 1980
- 110 **Khachik F**, Beecher GR, Whittaker NF. Separation, identification, and quantification of the major carotenoid and chlorophyll constituents in extracts of several green vegetables by liquid chromatography. *J Agric Food Chem* 1986; **34**: 603-616 [DOI: 10.1021/jf00070a006]
- 111 **Takaichi S**, Mimuro M. Distribution and geometric isomerism of neoxanthin in oxygenic phototrophs: 9'-cis, a sole molecular form. *Plant Cell Physiol* 1998; **39**: 968-977 [DOI: 10.1093/oxfordjournals.pcp.a029461]
- 112 **Ruban AV**, Lee PJ, Wentworth M, Young AJ, Horton P. Determination of the stoichiometry and strength of binding of xanthophylls to the photosystem II light harvesting complexes. *J Biol Chem* 1999; **274**: 10458-10465 [PMID: 10187836 DOI: 10.1074/jbc.274.15.10458]
- 113 **Seo M**, Koshiha T. Complex regulation of ABA biosynthesis in plants. *Trends Plant Sci* 2002; **7**: 41-48 [PMID: 11804826]
- 114 **Marki-Fischer E**, Eugster CH. Neoflor and 6-epineoflor from flowers of *Trollius europaeus*; highfield 1H-NMR spectra of (all-E)-neoxanthin and (9'Z)-neoxanthin. *Helv Chim Acta* 1990; **73**: 1637-1643 [DOI: 10.1002/hlca.19900730608]

- 115 **Asai A**, Terasaki M, Nagao A. An epoxide-furanoid rearrangement of spinach neoxanthin occurs in the gastrointestinal tract of mice and in vitro: formation and cytostatic activity of neochrome stereoisomers. *J Nutr* 2004; **134**: 2237-2243 [PMID: 15333710]
- 116 **Eugster CH** (1995) Chemical derivatization: microscale tests for the presence of common functional groups in carotenoids. In: Carotenoids Vol. 1A: Isolation and Analysis. Britton G, Liaaen-Jensen S, Pfander H, editors. Birkhäuser Verlag, Basel, Switzerland, 1995: 71-80
- 117 **Asai A**, Yonekura L, Nagao A. Low bioavailability of dietary epoxyxanthophylls in humans. *Br J Nutr* 2008; **100**: 273-277 [PMID: 18186952 DOI: 10.1017/S0007114507895468]
- 118 **Sugawara T**, Kushiro M, Zhang H, Nara E, Ono H, Nagao A. Lysophosphatidylcholine enhances carotenoid uptake from mixed micelles by Caco-2 human intestinal cells. *J Nutr* 2001; **131**: 2921-2927 [PMID: 11694619]
- 119 **Barua AB**, Olson JA. Xanthophyll epoxides, unlike beta-carotene monoepoxides, are not detectably absorbed by humans. *J Nutr* 2001; **131**: 3212-3215 [PMID: 11739868]
- 120 **Hashimoto T**, Ozaki Y, Taminato M, Das SK, Mizuno M, Yoshimura K, Maoka T, Kanazawa K. The distribution and accumulation of fucoxanthin and its metabolites after oral administration in mice. *Br J Nutr* 2009; **102**: 242-248 [PMID: 19173766 DOI: 10.1017/S0007114508199007]
- 121 **Pérez-Gálvez A**, Martin HD, Sies H, Stahl W. Incorporation of carotenoids from paprika oleoresin into human chylomicrons. *Br J Nutr* 2003; **89**: 787-793 [PMID: 12828795]
- 122 **Kotake-Nara E**, Sugawara T, Nagao A. Antiproliferative effect of neoxanthin and fucoxanthin on culture cells. *Fish Sci* 2005; **71**: 459-461. Available from: URL: <http://link.springer.com/article/10.1111/j.1444-2906.2005.00986.x#page-1>
- 123 **Terasaki M**, Asai A, Zhang H, Nagao A. A highly polar xanthophyll of 9'-cis-neoxanthin induces apoptosis in HCT116 human colon cancer cells through mitochondrial dysfunction. *Mol Cell Biochem* 2007; **300**: 227-237 [PMID: 17186379 DOI: 10.1007/s11010-006-9387-0]
- 124 **Ugocsai K**, Varga A, Molnár P, Antus S, Molnár J. Effects of selected flavonoids and carotenoids on drug accumulation and apoptosis induction in multidrug-resistant colon cancer cells expressing MDR1/LRP. *In Vivo* 2005; **19**: 433-438 [PMID: 15796208]
- 125 **Chang JM**, Lin JK. Isolation of neoxanthin from spinach and its prevention on lipid peroxidation. *J Chin Med* 1993; **4**: 235-245. Available from: URL: http://tao.wordpedia.com/show_pdf.ashx?sess=m3o4bi2vfuk2zou5qjretbmz&file_name=JO00000295_4-3_235-245&file_type=r
- 126 **Chang JM**, Chen WC, Hong D, Lin JK. The inhibition of DMBA-induced carcinogenesis by neoxanthin in hamster buccal pouch. *Nutr Cancer* 1995; **24**: 325-333 [PMID: 8610051 DOI: 10.1080/01635589509514421]
- 127 **Murakami A**, Nakashima M, Koshihara T, Maoka T, Nishino H, Yano M, Sumida T, Kim OK, Koshimizu K, Ohgashi H. Modifying effects of carotenoids on superoxide and nitric oxide generation from stimulated leukocytes. *Cancer Lett* 2000; **149**: 115-123 [PMID: 10737715]
- 128 **Okada T**, Nakai M, Maeda H, Hosokawa M, Sashima T, Miyashita K. Suppressive effect of neoxanthin on the differentiation of 3T3-L1 adipose cells. *J Oleo Sci* 2008; **57**: 345-351 [PMID: 18469497]
- 129 **Mayne ST**. Beta-carotene, carotenoids, and disease prevention in humans. *FASEB J* 1996; **10**: 690-701 [PMID: 8635686]
- 130 **Slattery ML**, Benson J, Curtin K, Ma KN, Schaeffer D, Potter JD. Carotenoids and colon cancer. *Am J Clin Nutr* 2000; **71**: 575-582 [PMID: 10648274]
- 131 Lutein and zeaxanthin. Monograph. *Altern Med Rev* 2005; **10**: 128-135 [PMID: 15989382]
- 132 **Sommerburg O**, Keunen JE, Bird AC, van Kuijk FJ. Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes. *Br J Ophthalmol* 1998; **82**: 907-910 [PMID: 9828775 DOI: 10.1136/bjo.82.8.907]
- 133 **Kelly ER**, Plat J, Haenen GR, Kijlstra A, Berendschot TT. The effect of modified eggs and an egg-yolk based beverage on serum lutein and zeaxanthin concentrations and macular pigment optical density: results from a randomized trial. *PLoS One* 2014; **9**: e92659 [PMID: 24675775 DOI: 10.1371/journal.pone.0092659]
- 134 **Landrum JT**, Bone RA. Lutein, zeaxanthin, and the macular pigment. *Arch Biochem Biophys* 2001; **385**: 28-40 [PMID: 11361022 DOI: 10.1006/abbi.2000.2171]
- 135 **Ben Amara N**, Tourniaire F, Maraninchi M, Attla N, Amiot-Carlin MJ, Raccach D, Valéro R, Landrier JF, Darmon P. Independent positive association of plasma β -carotene concentrations with adiponectin among non-diabetic obese subjects. *Eur J Nutr* 2014 [PMID: 24906472]
- 136 **Kritchevsky SB**. beta-Carotene, carotenoids and the prevention of coronary heart disease. *J Nutr* 1999; **129**: 5-8 [PMID: 9915867]
- 137 **Ascherio A**, Rimm EB, Hernán MA, Giovannucci E, Kawachi I, Stampfer MJ, Willett WC. Relation of consumption of vitamin E, vitamin C, and carotenoids to risk for stroke among men in the United States. *Ann Intern Med* 1999; **130**: 963-970 [PMID: 10383366 DOI: 10.7326/0003-4819-130-12-199906150-00003]
- 138 **Michaud DS**, Feskanich D, Rimm EB, Colditz GA, Speizer FE, Willett WC, Giovannucci E. Intake of specific carotenoids and risk of lung cancer in 2 prospective U.S. cohorts. *Am J Clin Nutr* 2000; **72**: 990-997 [PMID: 11010942]
- 139 **Le Marchand L**, Hankin JH, Bach F, Kolonel LN, Wilkens LR, Staceywicz-Sapuntzakis M, Bowen PE, Beecher GR, Laudon F, Baque P. An ecological study of diet and lung cancer in the South Pacific. *Int J Cancer* 1995; **63**: 18-23 [PMID: 7558446]
- 140 **Okuyama Y**, Ozasa K, Oki K, Nishino H, Fujimoto S, Watanabe Y. Inverse associations between serum concentrations of zeaxanthin and other carotenoids and colorectal neoplasia in Japanese. *Int J Clin Oncol* 2014; **19**: 87-97 [PMID: 23380957 DOI: 10.1007/s10147-013-0520-2]
- 141 **Tsubono Y**, Tsugane S, Gey KF. Plasma antioxidant vitamins and carotenoids in five Japanese populations with varied mortality from gastric cancer. *Nutr Cancer* 1999; **34**: 56-61 [PMID: 10453442]
- 142 **Stahl W**, Heinrich U, Jungmann H, Sies H, Tronnier H. Carotenoids and carotenoids plus vitamin E protect against ultraviolet light-induced erythema in humans. *Am J Clin Nutr* 2000; **71**: 795-798 [PMID: 10702175]
- 143 **Dorgan JF**, Sowell A, Swanson CA, Potischman N, Miller R, Schussler N, Stephenson HE. Relationships of serum carotenoids, retinol, alpha-tocopherol, and selenium with breast cancer risk: results from a prospective study in Columbia, Missouri (United States) *Cancer Causes Control* 1998; **9**: 89-97 [PMID: 9486468]
- 144 **Zhang S**, Tang G, Russell RM, Mayzel KA, Stampfer MJ, Willett WC, Hunter DJ. Measurement of retinoids and carotenoids in breast adipose tissue and a comparison of concentrations in breast cancer cases and control subjects. *Am J Clin Nutr* 1997; **66**: 626-632 [PMID: 9280184]
- 145 **Yeum KJ**, Ahn SH, Rupp de Paiva SA, Lee-Kim YC, Krinsky NI, Russell RM. Correlation between carotenoid concentrations in serum and normal breast adipose tissue of women with benign breast tumor or breast cancer. *J Nutr* 1998; **128**: 1920-1926 [PMID: 9808643]
- 146 **Toniolo P**, Van Kappel AL, Akhmedkhanov A, Ferrari P, Kato I, Shore RE, Riboli E. Serum carotenoids and breast cancer. *Am J Epidemiol* 2001; **153**: 1142-1147 [PMID: 11415946 DOI: 10.1093/aje/153.12.1142]
- 147 **Bravi F**, Bosetti C, Filomeno M, Levi F, Garavello W, Galimberti S, Negri E, La Vecchia C. Foods, nutrients and the risk of oral and pharyngeal cancer. *Br J Cancer* 2013; **109**: 2904-2910 [PMID: 24149181 DOI: 10.1038/bjc.2013.667]

- 148 **Dagnelie G**, Zorge IS, McDonald TM. Lutein improves visual function in some patients with retinal degeneration: a pilot study via the Internet. *Optometry* 2000; **71**: 147-164 [PMID: 10970259]
- 149 **Granado F**, Olmedilla B, Gil-Martínez E, Blanco I. Lutein ester in serum after lutein supplementation in human subjects. *Br J Nutr* 1998; **80**: 445-449 [PMID: 9924266 DOI: 10.1017/S0007114598001512]
- 150 **González de Mejía E**, Ramos-Gómez M, Loarca-Piña G. Antimutagenic activity of natural xanthophylls against aflatoxin B1 in *Salmonella typhimurium*. *Environ Mol Mutagen* 1997; **30**: 346-353 [PMID: 9366914 DOI: 10.1002/(SICI)1098-2280(1997)30:3<346::AID-EM14>3.0.CO;2-D]
- 151 **Kawashima T**. A marine carotenoid, fucoxanthin, induces regulatory T cells and inhibits Th17 cell differentiation in vitro. *Biosci Biotechnol Biochem* 2011; **75**: 2066-2069 [PMID: 21979096]
- 152 **Iannone A**, Rota C, Bergamini S, Tomasi A, Canfield LM. Antioxidant activity of carotenoids: An electron-spin resonance study on β -carotene and lutein interaction with free radicals generated in a chemical system. *J Biochem Mol Toxicol* 1998; **1**: 299-304 [PMID: 9664236 DOI: 10.1002/(SICI)1099-0461(1998)12:5<299::AID-JBT6>3.0.CO;2-G]
- 153 **Sujak A**, Gabrielska J, Grudzinski W, Borc R, Mazurek P, Gruszecki WI. Lutein and zeaxanthin as protectors of lipid membranes against oxidative damage: The structural aspects. *Arch Biochem Biophys* 1999; **371**: 301-307 [PMID: 10545218 DOI: 10.1006/abbi.1999.1437]
- 154 **Stahl W**, Junghans A, de Boer B, Driomina ES, Briviba K, Sies H. Carotenoid mixtures protect multilamellar liposomes against oxidative damage: Synergistic effects of lycopene and lutein. *FEBS Lett* 1998; **427**: 305-308 [PMID: 9607334 DOI: 10.1016/S0014-5793(98)00434-7]
- 155 **Rafi MM**, Kanakasabai S, Gokam SV, Krueger EG, Bright JJ. Dietary Lutein Modulates Growth and Survival Genes in Prostate Cancer Cells. *J Med Food* 2014 Aug 27; Epub ahead of print [PMID: 25162762 DOI: 10.1089/jmf.2014.0003]
- 156 **Pei YX**, Heng ZC, Duan GC, Wang MC. [The mechanisms and effects of lutein on inducing the cell differentiation of human esophagus cancer EC9706]. *Sichuan Daxue Xuebao Yixueban* 2007; **38**: 629-632 [PMID: 17718427]
- 157 **Sumantran VN**, Zhang R, Lee DS, Wicha MS. Differential regulation of apoptosis in normal versus transformed mammary epithelium by lutein and retinoic acid. *Cancer Epidemiol Biomarkers Prev* 2000; **9**: 257-263 [PMID: 10750663]
- 158 **Li Z**, Wang Y, Mo B. [The effects of carotenoids on the proliferation of human breast cancer cell and gene expression of bcl-2]. *Zhonghua Yufangyixue Zazhi* 2002; **36**: 254-257 [PMID: 12411207]
- 159 **Park JS**, Chew BP, Wong TS, Zhang JX, Magnuson NS. Dietary lutein but not astaxanthin or beta-carotene increases pim-1 gene expression in murine lymphocytes. *Nutr Cancer* 1999; **33**: 206-212 [PMID: 10368818 DOI: 10.1207/S15327914NC330214]
- 160 **Gonzalez de Mejia E**, Loarca-Pina G, Ramos-Gomez M. Antimutagenicity of xanthophylls present in Aztec Marigold (*Tagetes erecta*) against 1-nitropyrene. *Mutat Res* 1997; **389**: 219-226 [PMID: 9093387 DOI: 10.1016/S1383-5718(96)00151-9]
- 161 **Park JS**, Chew BP, Wong TS. Dietary lutein from marigold extract inhibits mammary tumor development in BALB/c mice. *J Nutr* 1998; **128**: 1650-1656 [PMID: 9772131]

P- Reviewer: Gagliardi G, Muscarella P, Schweiger U
S- Editor: Ji FF L- Editor: A E- Editor: Lu YJ



Pharmacological role of efflux transporters: Clinical implications for medication use during breastfeeding

Hilai Ahmadzai, Lisa BG Tee, Andrew Crowe

Hilai Ahmadzai, Lisa BG Tee, Andrew Crowe, School of Pharmacy and Curtin Health Innovation Research Institute, Curtin University, Bentley, WA 6102, Australia

Author contributions: All authors contributed to this paper equally.

Correspondence to: Dr. Andrew Crowe, School of Pharmacy and Curtin Health Innovation Research Institute, Curtin University, Kent Street, Bentley, WA 6102, Australia. a.p.crowe@curtin.edu.au

Telephone: +61-8-92663423 Fax: +61-8-92662769

Received: June 28, 2014 Revised: September 3, 2014

Accepted: September 23, 2014

Published online: December 9, 2014

Abstract

The World Health Organisation recommends exclusive breastfeeding for the first six months of an infant's life and in combination with solid food thereafter. This recommendation was introduced based on research showing numerous health benefits of breastfeeding for both the mother and the infant. However, there is always concern regarding the transfer of medications from mother to their breastfed baby *via* milk. Pharmacokinetic properties of a drug are usually used to predict its transferability into breast milk. Although most drugs are compatible with breastfeeding, cases of toxic drug exposure have been reported. This is thought to be due to active transport mechanisms whereby efflux transporter proteins expressed in the epithelial cells of the mammary gland actively secrete drugs into milk. An example of such efflux transporters including the breast cancer resistance protein which is strongly induced during lactation and this could result in contamination of milk with the substrates of this transporter which may place the suckling infant at risk of toxicity. Furthermore, there is little known about the substrate specificity of most efflux transporters as we have highlighted in this review. There also exists some degree of contradiction between *in vivo* and *in vitro* studies which makes it difficult to conclusively predict outcomes and drug-drug

interactions.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Active efflux transporters; Lactation; Breast-feeding; Mammary gland; Breast cancer resistance protein; P-glycoprotein; Breast milk; ABC transporters

Core tip: The aim of this review was to analyse the available literature on psychoactive drugs specifically selective serotonin reuptake inhibitors, antipsychotics and antiepileptic drugs that are commonly prescribed during lactation and pregnancy. This review investigated whether these drugs are substrates and/or inhibitors of efflux transporters especially of P-glycoprotein and breast cancer resistance protein and whether this has any effect on adverse outcomes in the breastfed infant of mothers who use these pharmacotherapeutic agents. Current evidence on acute adverse effects in breastfed infants due to the aforementioned drug groups either as sole treatment or their use in combination with other drugs was also explored.

Ahmadzai H, Tee LBG, Crowe A. Pharmacological role of efflux transporters: Clinical implications for medication use during breastfeeding. *World J Pharmacol* 2014; 3(4): 153-161 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/153.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.153>

INTRODUCTION

It is widely accepted that breastfeeding is the best way of ensuring a good start in an infant's life as it not only has a favourable nutrient content but also provides passive immunity and various growth hormones to the breastfed infant^[1-4]. However, there is always concern regarding the transfer of medications taken by the mother to the infant *via* breast milk. To understand the safety of medicines in a

nursing mother, it is important to elucidate the mechanism of such drug transfer.

The major determinant of the transferability of drugs from mother to baby *via* breast milk is usually calculated using parameters such as the physicochemical properties of the drug and the composition of the milk^[4]. However, research has shown that active transport *via* efflux transporters may have a significant role in the transfer of drugs from the maternal plasma to breast fed infant. It has been shown that breast cancer resistance protein (BCRP) (also known as ABCG2), belonging to the ATP-binding cassette (ABC) superfamily of transporters is strongly induced during lactation^[5]. There are also many other transporters belonging to the ABC family such as P-glycoprotein (P-gp) (also known as ABCB1 or MDR1) which like BCRP have some toxicological significance in lactation. Other transporters belonging to the solute carrier family such as the organic cation and anion transporters, peptide and nucleoside transporters also found in the mammary gland play an important role in the active transport of many nutrients, endogenous substances and xenobiotics^[6].

The protective role of P-gp in the blood brain barrier is well established. It inhibits a wide variety of substrates from entering the central nervous system^[7]. The main focus of research has so far been on the role of these transporters in the gastrointestinal tract and the blood brain barrier (BBB) affecting bioavailability of drugs, pharmacoresistance leading to ineffective drug treatment. Modulation of these efflux transporters can have an impact on drug absorption, disposition and consequently therapeutic outcome. However, we know that P-gp is also widely expressed in many other human tissues including the liver, kidneys, testes, placenta and the mammary epithelial cells^[8]. It is also well known that these ABC transporters play a crucial role in the protective mechanism during embryogenesis in the placenta which is continued during lactation providing foetal protection against naturally occurring toxins^[9]. Our area of interest is the role of these transporters in the lactating human mammary epithelial cells (HMEC) where they could potentially influence transfer of drugs from mother to their breastfed baby *via* breast milk. However, this area remains poorly researched and there is lack of controlled studies that can provide conclusive evidence. On the basis of other organ systems, co-administration of medications that are substrates or inhibitors of these transporters in a nursing mother could have significant drug-drug interactions which may lead to adverse effects in their breastfed infant. Several *in vitro* and animal studies have been conducted which address this area of concern. However, the applicability of these *in vitro* findings may not always be conclusive and are often contradictory due to differences in experimental design and use of species other than humans^[10-12]. Most animal species such as rats and mice used in the laboratory have two genes (*Mdr1a* and *Mdr1b*) that code for P-gp, which further complicates issues regarding induction, expression and drug-drug interactions^[11].

TRANSFER OF DRUGS FROM MATERNAL PLASMA TO THE BREASTFED INFANT

The transfer of drugs from maternal plasma to breast milk occurs *via* passive and active mechanisms^[9,13]. The critical determinants of passive transfer include drug protein binding, drug ionisation and fat partitioning^[4,13]. These factors can be used to predict milk to plasma (M:P) ratio where passive diffusion is thought to predominate. Other pharmacokinetic parameters such as half-life of the drug, protein binding, water and lipid solubility, route of drug administration, bioavailability, dissociation constant, volume of distribution, molecular size and ionisation potential can further help to determine the transfer of drugs from mother's plasma into the breast milk^[14]. Drugs with the shortest plasma half-life, highest protein binding and lowest lipid solubility usually have the lowest ductal milk transport. The dose of a drug that an infant receives during breastfeeding depends on the amount excreted into the breast milk, the daily volume of the milk ingested and the average plasma concentration of the mother. Thus, M:P ratio has large inter-subject variability^[15]. Although the transfer of most drugs into breast milk can be explained by passive diffusion theories, a review of the literature shows that there are several drugs where the actual measured M:P ratio is significantly greater than predicted^[16-19]. Nitrofurantoin, acyclovir and cimetidine are some drugs which exhibited a significantly higher observed M:P ratio than predicted^[5,7,16,18,20]. In one study nitrofurantoin had an observed M:P ratio of 6 as opposed to the predicted 0.28^[17]. It has also been shown that several members of the ABC drug efflux transporters significantly affect the pharmacokinetic disposition of drugs such as the quinolones, thereby increasing their secretion into breast milk^[17,19].

ROLE OF EFFLUX TRANSPORTERS IN THE TRANSFER OF DRUGS FROM MATERNAL PLASMA TO THE BREASTFED INFANT

The extent of the involvement of these ABC transporters in the transfer of many nutrients including essential vitamins and drugs into the breast milk has been recently considered^[21]. There are several efflux transporters in human mammary epithelial cells that line the alveoli within the mammary gland^[8,22,23]. This leads us to believe that there may be a more substantial role of these transporter proteins in the transfer of many compounds from maternal plasma to the breast milk than currently perceived. Alcorn *et al*^[8] have shown that there is some of variability between the level of RNA expression of various transporters in the HMEC from lactating *vs* non lactating breast tissue, indicating a graded expression change during induction of the lactation process that could lead to significant changes in substrate transport

during lactation. Using immunocytochemical analysis and functional studies in primary human mammary epithelial cells culture, our group have demonstrated the presence of MDR1 (ABCB1), MDR3 (ABCB4) and MRP1 (ABCC1) in these cells^[21].

Gilchrist and colleagues showed that there is a stage dependent change in the expression of transporters in rat mammary gland and isolated mammary epithelial organoids^[24]. Using quantitative reverse transcription polymerase chain reaction, they demonstrated that the various solute carrier and ABC transporters showed a changing pattern in the different stages of lactation. Ling *et al*^[25] studied the M:P ratio of cefepime, an actively transported drug at four and ten days post-partum in rats and found a significant reduction in the amount of cefepime excreted at these two time points. This leads us to believe that as lactation progresses from stages of mammogenesis to lactogenesis to galactopoiesis, changes in the expression of efflux transporters along with changing hormones may influence the transfer of endogenous and exogenous substances from mother to baby *via* breast milk. However, currently there are no studies in humans to confirm this. Our laboratory is presently investigating whether the expression of efflux transporters in humans follow a stage dependent pattern as seen in animal studies.

The expression of MRP1 (ABCC1) and MDR1 (ABCB1) are significantly lower in the lactating HMEC as compared to non-lactating HMEC^[8] whereas that of BCRP is significantly higher^[5]. It is important to note that there is a substantial overlap in the substrate specificities of these transporters^[6]. These findings highlight the importance of possible drug-drug interactions between various transporter substrates and/or inhibitors when co-administered at different stages of lactation. In addition, it is important to take into account the localization of these transporters, such that their presence in the apical surface (MDR1 and BCRP) may pump drugs into milk and further place the suckling infant at risk of xenobiotic exposure^[20]. Alternatively, if these transporters are located in the basolateral membrane of the cell (MRP1), then the substrate will be pumped out of the milk and into the mother's blood, thereby reducing infant exposure.

The active efflux transporters usually help in preventing accumulation of drugs into the tissues as they work against a concentration gradient and push drugs from the tissues back into the blood^[7,8,24]. There have also been reports of transporter proteins being involved in the transfer of essential nutrients and vitamins to the breast fed infant which are mostly located in the basolateral side^[7,22,26]. However, the extent to which they affect drug transfer in the mammary gland is not fully known^[6]. Similar to P-gp, BCRP has a protective role at the blood side of many organ systems by facilitating the extrusion of toxins, xenobiotics and drugs out of the capillaries prior to interstitial accumulation with an important role being in the blood-placental barrier, where they protect the foetus from endogenous and exogenous toxins^[27]. A current vexing question is how relevant is the role of efflux transporters at other blood-tissue barriers such as in HMECs of a lactating female who is breastfeeding her infant compared

to the plethora of studies examining gastrointestinal or BBB transport. Can P-gp or BCRP modulation by an inhibitor drug that the mother consumes cause less drug to be transferred to the breastfed infant *via* milk? Also if P-gp is located on the apical membrane of the HMEC, the evidence that RNA expression of P-gp is lower in the lactating HMEC^[8] could have a relatively protective effect on the breastfed infant, by virtue of less P-gp to excrete drug into the milk (Figure 1).

The expression of BCRP in pregnant mice was found to be strongly induced during late pregnancy and lactation, which is the opposite to that of P-gp^[5]. Lindner and colleagues reported that the expression of BCRP in the mammary glands of several species of animals including sheep, goats and cows were significantly increased (up to 10 fold) during pregnancy and lactation^[28]. Both P-gp and BCRP are located in the apical membrane of alveolar epithelial cells of the mammary gland and actively transport their substrates into breast milk as confirmed by animal studies^[5,20]. BCRP has a significant role in accumulation of drugs and xenotoxins in breast milk which could be either beneficial or detrimental to the breastfed infant's health depending on the drug administered^[29]. A BCRP substrate that is toxic can accumulate in milk and result in adverse effects in the infant whereas the accumulation of a drug such as aciclovir could be beneficial in reducing transmission of milk borne viruses from mother to baby. There is some evidence that the role of P-gp in the lactating HMEC is relatively insignificant in the transfer of medications which could possibly be due to its down regulation in lactation. Animal studies have shown that the transfer of nelfinavir, a known P-gp substrate is not affected by P-gp in the mammary gland^[28]. However, many other studies have shown that the role of BCRP is much more significant, given that BCRP is strongly induced during pregnancy and lactation^[5,16,18,30].

As there is a significant overlap between P-gp, BCRP, other efflux transporters and CYP3A4 substrates^[6], it is crucial that each drug or the combination of drugs is considered with respect to these transporters and metabolism pathways in addition to the usual pharmacokinetic parameters to ensure minimum inadvertent exposure to a breastfed infant.

SPECIFIC DRUGS

Antidepressants - selective serotonin reuptake inhibitors

Post-natal depression is considered to be a significant problem in women of child bearing age with approximately 14% of all women affected by this condition at some stage^[31]. Psychotherapy is considered quite useful in the management of post natal depression but due to a lack of adequate service provision in the community, it is often necessary to treat women with pharmacotherapeutic agents^[31]. Selective serotonin reuptake inhibitors (SSRI) are considered the mainstay of postnatal depression due to their perceived low transferability into breast milk and safety profile unlike tricyclic antidepressants that may have a higher breast milk ex-

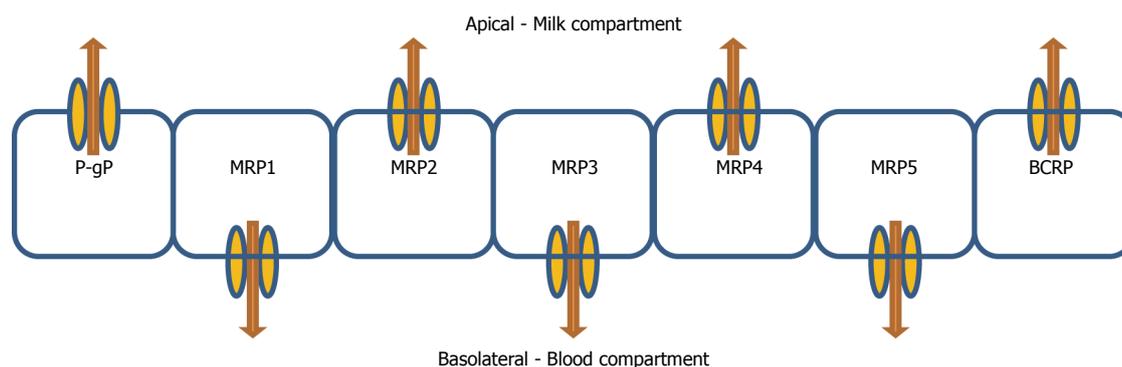


Figure 1 Schematic representation of expected localisation of drug efflux transporters in the human mammary epithelial cells. P-gp: P-glycoprotein; BCRP: Breast cancer resistance protein; MRP: Multidrug resistance protein.

posure leading to reduced usage^[31]. The recommended SSRIs for post-natal depression are sertraline and paroxetine as they have been widely studied and are not associated with many adverse effects^[32]. Fluoxetine and citalopram have measurable plasma levels in some infants^[33,34]. However, these levels were usually low. Some women may choose to continue using one of the other SSRIs that they have been stabilised on before and during pregnancy. Nonetheless, most SSRIs are considered to be fairly safe in breastfeeding.

Paroxetine and fluoxetine are thought to be P-gp substrates^[35]. Citalopram and its enantiomer, escitalopram have also been found to be substrates of P-gp in animal studies as demonstrated by Weiss *et al*^[36]. The implications and relevance for this evidence in lactation remains elusive. However, *in vivo* it has been shown that the peak plasma concentration and area under the curve of paroxetine, is significantly increased by itraconazole (a P-gp and CYP3A4 inhibitor)^[37]. Again, this could mean that a P-gp substrate which normally has low transferability into breast milk may potentially transfer in significantly higher amounts if given with a P-gp inhibitor.

Furthermore, it has been found that sertraline, a P-gp substrate and inhibitor, can modulate P-gp both *in vivo* and *in vitro* at the BBB and blood testes barrier sites^[38]. This finding suggests that concurrent administration of other P-gp substrates with sertraline potentially increase CNS penetration of that substrate^[39]. Again, the significance and implication of this finding in lactation needs to be investigated further as another study found modulation of P-gp by sertraline was site specific with different tissues reacting to sertraline in different ways^[40]. It is important to investigate whether this finding has any potential for causing interactions in the lactating HMEC leading to adverse effects in the breastfed infant as this tissue was not directly investigated. A different study by Bhuiyan *et al*^[41] found that a single dose of sertraline does not affect the pharmacokinetic profile of fexofenadine, another P-gp substrate but paroxetine and fluvoxamine do. There was no data with regards to BCRP substrate specificity in the selective serotonin reuptake inhibitors.

Of other new antidepressants, duloxetine, which is a serotonin noradrenaline reuptake inhibitor (SNRI), not used often in pregnancy and breastfeeding due to lack of

safety data in this population, was found to cause no immediate adverse effects in an exclusively breastfed 32 d old infant^[42]. The measured M:P ratio and the relative infant dose for this drug were also found to be very low^[43]. Venlafaxine, another SNRI was found to induce BCRP in brain tissue and is thought to be a P-gp substrate whereas desvenlafaxine, the active metabolite of venlafaxine was found to have no effect on BCRP induction or P-gp modulation^[44,45].

Another case study on sertraline reported signs of serotonergic overstimulation in a preterm baby whose mother had therapeutic levels of both the drug and its metabolite, desmethylsertraline. The symptoms disappeared on discontinuation of breastfeeding^[46]. This adverse reaction was attributed to immaturity in the development of the infant's clearance mechanisms and lack of development of the BBB. Interestingly, the plasma sertraline and desmethylsertraline levels of the infant were significantly below the threshold levels considered to cause symptoms. There was no record of other medications that may have been used by the mother acutely or long term during the postpartum period. Hence, it raises the question of whether another drug(s) administered acutely, able to modulate P-gp activity, could have resulted in the adverse effects experienced by the breastfed baby. The lack of a full medication record makes it difficult to draw conclusions regarding the drug-drug interactions.

Current guidelines place paroxetine and sertraline amongst the recommended antidepressant drugs for use in lactation. It is imperative that infants of mothers taking these drugs are regularly monitored for adverse effects especially when the mothers are also treated acutely or chronically with another pharmacologic agent that could modulate active transporters as these drugs have the potential to do^[35,47].

Antipsychotics

About one third of pregnant women with psychotic illness use antipsychotics at least once during pregnancy or whilst breastfeeding^[48]. About 10% of women of child bearing age have a postpartum psychiatric disorder, with a significant number warranting the use of an antipsychotic medication^[49]. Although the second generation (atypical)

antipsychotics are considered the best treatment option for schizophrenia, female patients who are pregnant or breastfeeding are often excluded from this treatment option due to safety concerns.

Several atypical antipsychotics are substrates for P-gp in therapeutic concentrations. These include amisulpride, aripiprazole, olanzapine, risperidone, quetiapine and paliperidone, with quetiapine and risperidone having high affinity for P-gp^[50]. Friedman *et al*^[50] found olanzapine and quetiapine to be P-gp substrates whereas another *in vitro* study by Müller *et al*^[47] contradicted this finding and identified that quetiapine, haloperidol, olanzapine and clozapine are not P-gp substrates. As the first study assessed P-gp activity by measuring ATPase activity whereas the second study was an inhibition study carried out using Caco-2 cell monolayers, these different methods of P-gp substrate identification may be responsible for their different conclusions. Our studies using Caco-2 monolayers support the latter work, demonstrating the lack of P-gp mediated efflux associated with quetiapine and olanzapine. Hence, it is important to exercise caution when interpreting results from different cell lines and membrane systems^[55]. *In vivo* studies using knockout and wild type mice may be more reliable and may provide a clearer picture of functional consequence^[51]. However, it is worth noting that these rodents may have more than one gene coding for P-gp, hence may differ from human P-gp structurally.

Several antipsychotics including clozapine, quetiapine, paliperidone and chlorpromazine also exert some inhibitory effects on BCRP^[52]. Risperidone has major inhibitory effects on BCRP, making it a potential contributor of adverse effects with co-administered with BCRP modulators such as commonly prescribed pantoprazole and omeprazole^[52,53]. Aripiprazole, an atypical antipsychotic was found to have an inhibitory effect on BCRP^[54]. Most antipsychotics are thought to act as inhibitors of P-gp/BCRP and therefore can influence plasma and brain concentrations of other substrates^[55].

A prospective controlled observational study of olanzapine use in 30 pregnant women who were taking olanzapine during pregnancy and whilst breastfeeding found that no adverse effects were imputable to the use of olanzapine by the mother. However, the rate of breastfeeding was significantly lower in the treated group. Also three out of thirty babies (10%) experienced withdrawal symptoms after birth and interestingly all three mums were on multiple medications including zuclopenthixol, lithium and paroxetine^[56]. A case report of a lactating patient who was taking olanzapine after a psychotic episode reported low infant plasma levels of olanzapine and no adverse effects in the breastfed baby^[57]. Another case study reported no adverse effects in the infant of a woman who was initiated on olanzapine during her third trimester and continued breastfeeding six months post-partum^[49]. There is no mention of concurrent therapy if any. A case report of risperidone by Lutz *et al*^[58] showed that risperidone, a P-gp substrate, and its metabolite

9-hydroxyrisperidone were moderately transferred into breast milk. A dose (concentration) of less than 10% of that received by the mother (on a mg/kg basis) has been suggested and is widely accepted as a “safe” dose (or concentration) in the infant^[45]. Although, the amount transferred (4.3% of maternal dose) was below the notional 10% threshold, the mother was encouraged not to breastfeed due to concerns for the safety of her infant. Again, no information was provided on whether the mother was on any other concurrent medications. Certainly, we would prefer clinical studies to state that there were no other medications in the study rather than just omitting this crucial information. Often when the focus is on one particular drug and snapshot studies such as M:P ratio are being investigated, other medications do not make it into the clinical notes. Given our presumption that in psychiatry multiple medications are often used concurrently, this makes subsequent contextual analysis very difficult. Furthermore, a series of case reports by Illet's group^[59] confirmed the findings from the first report that risperidone on its own is not transferred into breast milk in levels high enough to be considered a clinical issue for the safety of the breastfed infant.

Ziprasidone, an atypical antipsychotic was found to be excreted in very low concentrations in breast milk from a treated patient while no adverse effects were observed in the infant of a different patient with psychotic depression treated with citalopram and ziprasidone^[60,61]. It is not known whether ziprasidone is a P-gp substrate or BCRP modulator at this time^[55]. Further research and more long term studies are warranted to ensure the safety of the newer antipsychotics on infant growth and development.

Amisulpride, another atypical antipsychotic has been used in one woman who was breastfeeding her 13 mo old infant. An unusually high M:P ratio for this drug compared to predicted values based on pharmacokinetic parameters was found^[62]. This high M:P ratio was attributed to amisulpride being a P-gp substrate^[47,63]. Amisulpride used in conjunction with desvenlafaxine in a partially breastfed infant yielded no higher than expected relative infant dose. Given that Amisulpride is a P-gp substrate, the combination with desvenlafaxine did not appear to alter its pharmacokinetics reaffirming the *in vitro* evidence that desvenlafaxine does not modulate efflux transporters^[44,45]. Olanzapine is considered a weak substrate whereas data for quetiapine are contradictory with one study identifying it as a substrate and the other not a substrate^[47,63]. Another *in vitro* study found that olanzapine and risperidone may inhibit P-gp activity. Most other drugs in this therapeutic group though, such as clozapine, haloperidol chlorpromazine and quetiapine did not inhibit P-gp^[64]. Much of the studies discussed above were cell based, and projecting this data into clinical studies has been sorely lacking. Nonetheless, it is appropriate to exercise caution when using these agents especially in combination with another agent which may modulate P-gp especially in a lactating mother who is breastfeeding or exclusively breastfeeding, given that some studies sup-

port the notion that blocking P-gp (or other efflux transporters) can elevate milk concentration of these drugs.

Antiepileptic drugs

Many women with epilepsy require treatment with anti-epileptic drugs (AED) during pregnancy and post-partum when they may be breastfeeding their baby. Usually women with epilepsy do not have a choice to discontinue treatment while pregnant or breastfeeding. All antiepileptic drugs are transferred across the placenta and to a lesser extent into breast milk in varying amounts. The implications of AED exposure *via* breast milk is still not fully understood. A large prospective study of epileptic mothers on AED prenatally showed adverse development in their children regardless of breastfeeding status^[65,66]. Some other large studies also showed no damaging effects on neurodevelopment of breastfed children of women who were prescribed AED during breastfeeding^[67]. Nevertheless, most antiepileptic drugs are known to be teratogenic and increase the risk of foetal malformations^[68].

The role of efflux transporters such as the MRP group and P-gp in the transport of AED showed variable results^[69]. Luna-Tortós *et al.*^[10] initially found that several AED were substrates of the human P-gp. Lamotrigine and phenobarbital were also found to be MRP substrates. However, in a subsequent study they indicated that AED were not substrates of the human P-gp and that the different interpretations from the two studies were attributed to the different experimental designs used^[11]. Several other studies also suggested that AED were not human P-glycoprotein substrates^[12,68-70]. Studies performed in animals yielded conflicting results showing that AED were indeed substrates of P-gp^[71-74]. The variability in these results may be attributed to the differences between human and rat P-gp. The cell lines and experimental designs used for conducting these studies can also have a significant impact on the results^[71]. Another study in mice found that levetiracetam, topiramate and phenytoin demonstrate biphasic modulation of P-gp in BBB whereby at therapeutic doses they act as inducers of efflux. Sodium valproate and lamotrigine were found not to interact with P-gp^[11,73,75]. Carbamazepine, itself was not a P-gp substrate but its metabolites were^[76,77]. Dickens and co-workers also identified that lamotrigine and carbamazepine were not affected by P-gp^[78,79]. Several case reports show that lamotrigine was transferred into breast milk in moderate amounts and in one case, where the mother was on 850 mg/d, it led to a severe apnoeic reaction in the exclusively breast fed infant^[80]. It is not specified whether the mother was on any other medications at the time that could have contributed to this adverse reaction. Another study has shown that although lamotrigine's M:P ratio is highly variable, it is transferred into breast milk in moderate amounts^[67].

In vitro studies carried out on MDCKII cells by Nakanishi *et al.*^[75] found that phenobarbital, clobazam, zonisamide, gabapentin and levetiracetam were BCRP substrates. Contrary to their findings, another group

found that phenobarbital as well as other AED including phenytoin, ethosuximide, primidone, sodium valproate, carbamazepine, clonazepam, and lamotrigine did not interact with BCRP^[79]. The differences in the experimental design and human and animal transporters may partly explain the variations in these results, with the animal studies appearing to favour anti-epileptic P-gp affinity while the human P-gp studies do not support this^[68,77,79].

It is interesting that genetic polymorphism of P-gp is not associated with drug response in epileptic patients^[80,81]. Again this concurs with the evidence that AED may not be transported by efflux transporters such as P-gp and BCRP^[81]. Weiss *et al.*^[36] indicate that P-gp may not be of great significance in the transport of AED. Further studies are required to elucidate if there are any other active transport mechanisms which may have significant clinical implications in breastfeeding mothers.

DISCUSSION

It is well known that exposure to medications in utero is significantly higher than exposure *via* breast milk. However, exposure *via* breast milk is voluntary as opposed to in utero where they are inadvertently exposed to medications through placental transfer. Infant exposure to medications *via* breast milk, especially in the first six months when the infant is likely to be exclusively breastfed can have severe adverse effects possibly due to the underdeveloped metabolic and excretory mechanisms in the infant during this time. Hence, even small exposure to medication *via* breast milk may result in accumulation of drug at a level that may cause side effects in the infant, which calls into question the notional 10% of mother's plasma concentration in the milk as the threshold for concern. It is also important not to discontinue breastfeeding unnecessarily as there are many benefits associated with breastfeeding. Drug pharmacokinetic parameters usually give a good indication of the transferability of the drug from mother to baby *via* breast milk. However, there have been incidents when the actual M:P ratio of certain drugs have been much higher than predicted which is thought to be due to the involvement of active transport mechanisms. Yet, the significance of such active transport mechanisms and efflux transporters in the human mammary epithelial cells is still not fully understood. *In vivo* and *in vitro* studies have confirmed their presence but the clinical relevance of their role remains elusive. There is also contradiction and conflict with regards to the current findings as to whether particular drugs are substrates of these transporters and the degree of commonality between transporter substrates. Data can vary depending on the experimental design and the cell lines used. There is also significant inter species variability which can affect interpretation of results. Nonetheless, it is important to exercise caution when prescribing CNS acting drugs, such as psychotropics to breastfeeding mothers as even small unnecessary drug exposure may have disturbing side effects in the very young.

REFERENCES

- 1 **Lawrence RA**, Lawrence RM. Biochemistry of human milk. In: Breastfeeding: a guide for the medical profession. Missouri, USA: Mosby, 2011: 98-152
- 2 **Duncan B**, Ey J, Holberg CJ, Wright AL, Martinez FD, Taussig LM. Exclusive breast-feeding for at least 4 months protects against otitis media. *Pediatrics* 1993; **91**: 867-872 [PMID: 8474804]
- 3 **Lawrence RA**, Lawrence RM. Host-resistance factors and immunologic significance of human milk. In: Breastfeeding: a guide for the medical profession. Missouri, USA: Mosby, 2011: 153-195
- 4 **Fleishaker JC**, Desai N, McNamara PJ. Factors affecting the milk-to-plasma drug concentration ratio in lactating women: physical interactions with protein and fat. *J Pharm Sci* 1987; **76**: 189-193 [PMID: 3585733]
- 5 **Jonker JW**, Merino G, Musters S, van Herwaarden AE, Bolscher E, Wagenaar E, Mesman E, Dale TC, Schinkel AH. The breast cancer resistance protein BCRP (ABCG2) concentrates drugs and carcinogenic xenotoxins into milk. *Nat Med* 2005; **11**: 127-129 [PMID: 15685169 DOI: 10.1038/nm1186]
- 6 **Ito S**, Alcorn J. Xenobiotic transporter expression and function in the human mammary gland. *Adv Drug Deliv Rev* 2003; **55**: 653-665 [PMID: 12706548]
- 7 **Schinkel AH**, Jonker JW. Mammalian drug efflux transporters of the ATP binding cassette (ABC) family: an overview. *Adv Drug Deliv Rev* 2003; **55**: 3-29 [PMID: 12535572]
- 8 **Alcorn J**, Lu X, Moscow JA, McNamara PJ. Transporter gene expression in lactating and nonlactating human mammary epithelial cells using real-time reverse transcription-polymerase chain reaction. *J Pharmacol Exp Ther* 2002; **303**: 487-496 [PMID: 12388627 DOI: 10.1124/jpet.102.038315]
- 9 **Vähäkangas K**, Myllynen P. Drug transporters in the human blood-placental barrier. *Br J Pharmacol* 2009; **158**: 665-678 [PMID: 19788499 DOI: 10.1111/j.1476-5381]
- 10 **Luna-Tortós C**, Fedrowitz M, Löscher W. Several major antiepileptic drugs are substrates for human P-glycoprotein. *Neuropharmacology* 2008; **55**: 1364-1375 [PMID: 18824002 DOI: 10.1016/j.neuropharm.2008.08.032]
- 11 **Luna-Tortós C**, Fedrowitz M, Löscher W. Evaluation of transport of common antiepileptic drugs by human multidrug resistance-associated proteins (MRP1, 2 and 5) that are overexpressed in pharmacoresistant epilepsy. *Neuropharmacology* 2010; **58**: 1019-1032 [PMID: 20080116 DOI: 10.1016/j.neuropharm.2010.01.007]
- 12 **Rivers F**, O'Brien TJ, Callaghan R. Exploring the possible interaction between anti-epilepsy drugs and multidrug efflux pumps; in vitro observations. *Eur J Pharmacol* 2008; **598**: 1-8 [PMID: 18835265 DOI: 10.1016/j.ejphar.2008.09.014]
- 13 **Begg EJ**, Atkinson HC. Modelling of the passage of drugs into milk. *Pharmacol Ther* 1993; **59**: 301-310 [PMID: 8309993]
- 14 **Ito S**, Lee A. Drug excretion into breast milk--overview. *Adv Drug Deliv Rev* 2003; **55**: 617-627 [PMID: 12706545]
- 15 **Schaefer PP**, Miller R. Drugs during pregnancy and lactation- Treatment options and risk assessment. 2nd ed. London GB: Academic Press, 2007
- 16 **Buhimschi CS**, Weiner CP. Medications in pregnancy and lactation: part 1. Teratology. *Obstet Gynecol* 2009; **113**: 166-188 [PMID: 19104374 DOI: 10.1097/AOG.0b013e31818d6788]
- 17 **Dostal LA**, Weaver RP, Schwetz BA. Excretion of high concentrations of cimetidine and ranitidine into rat milk and their effects on milk composition and mammary gland nucleic acid content. *Toxicol Appl Pharmacol* 1990; **102**: 430-442 [PMID: 1690458]
- 18 **Gerk PM**, Kuhn RJ, Desai NS, McNamara PJ. Active transport of nitrofurantoin into human milk. *Pharmacotherapy* 2001; **21**: 669-675 [PMID: 11401180]
- 19 **Alvarez AI**, Pérez M, Prieto JG, Molina AJ, Real R, Merino G. Fluoroquinolone efflux mediated by ABC transporters. *J Pharm Sci* 2008; **97**: 3483-3493 [PMID: 18200507 DOI: 10.1002/jps.21233]
- 20 **Merino G**, Jonker JW, Wagenaar E, van Herwaarden AE, Schinkel AH. The breast cancer resistance protein (BCRP/ABCG2) affects pharmacokinetics, hepatobiliary excretion, and milk secretion of the antibiotic nitrofurantoin. *Mol Pharmacol* 2005; **67**: 1758-1764 [PMID: 15709111 DOI: 10.1124/mol.104.010439]
- 21 **Tee LBG**, Williams D, Crowe A, Illett K. Drug transporters in human mammary epithelial cells. Denmark: 16th World Congress of Basic and Clinical Pharmacology, 2010
- 22 **van Herwaarden AE**, Schinkel AH. The function of breast cancer resistance protein in epithelial barriers, stem cells and milk secretion of drugs and xenotoxins. *Trends Pharmacol Sci* 2006; **27**: 10-16 [PMID: 16337280 DOI: 10.1016/j.tips.2005.11.007]
- 23 **van Herwaarden AE**, Wagenaar E, Merino G, Jonker JW, Rosing H, Beijnen JH, Schinkel AH. Multidrug transporter ABCG2/breast cancer resistance protein secretes riboflavin (vitamin B2) into milk. *Mol Cell Biol* 2007; **27**: 1247-1253 [PMID: 17145775 DOI: 10.1128/mcb.01621-06]
- 24 **Gilchrist SE**, Alcorn J. Lactation stage-dependent expression of transporters in rat whole mammary gland and primary mammary epithelial organoids. *Fundam Clin Pharmacol* 2010; **24**: 205-214 [PMID: 19702690 DOI: 10.1111/j.1472-8206.2009.00760.x]
- 25 **Ling B**, Alcorn J. Lactation stage influences drug milk-to-serum values and neonatal exposure risk. *Int J Toxicol* 2010; **29**: 411-417 [PMID: 20457592 DOI: 10.1177/1091581810367949]
- 26 **Vlaming ML**, Lagas JS, Schinkel AH. Physiological and pharmacological roles of ABCG2 (BCRP): recent findings in Abcg2 knockout mice. *Adv Drug Deliv Rev* 2009; **61**: 14-25 [PMID: 19118589 DOI: 10.1016/j.addr.2008.08.007]
- 27 **Mo W**, Zhang JT. Human ABCG2: structure, function, and its role in multidrug resistance. *Int J Biochem Mol Biol* 2012; **3**: 1-27 [PMID: 22509477]
- 28 **Lindner S**, Halwachs S, Wassermann L, Honscha W. Expression and subcellular localization of efflux transporter ABCG2/BCRP in important tissue barriers of lactating dairy cows, sheep and goats. *J Vet Pharmacol Ther* 2013; **36**: 562-570 [PMID: 23473424 DOI: 10.1111/jvp.12045]
- 29 **Edwards JE**, Alcorn J, Savolainen J, Anderson BD, McNamara PJ. Role of P-glycoprotein in distribution of nelfinavir across the blood-mammary tissue barrier and blood-brain barrier. *Antimicrob Agents Chemother* 2005; **49**: 1626-1628 [PMID: 15793156 DOI: 10.1128/aac.49.4.1626-1628.2005]
- 30 **Perez M**, Blazquez AG, Real R, Mendoza G, Prieto JG, Merino G, Alvarez AI. In vitro and in vivo interaction of moxidecetin with BCRP/ABCG2. *Chem Biol Interact* 2009; **180**: 106-112 [PMID: 19428349 DOI: 10.1016/j.cbi.2009.02.009]
- 31 **Stuart S**, Koleva H. Psychological treatments for perinatal depression. *Best Pract Res Clin Obstet Gynaecol* 2014; **28**: 61-70 [PMID: 24269903 DOI: 10.1016/j.bpobgyn.2013.09.004]
- 32 **Gentile S**. Tricyclic antidepressants in pregnancy and puerperium. *Expert Opin Drug Saf* 2014; **13**: 207-225 [PMID: 24383525 DOI: 10.1517/14740338.2014.869582]
- 33 **Guille C**, Newman R, Fryml LD, Lifton CK, Epperson CN. Management of postpartum depression. *J Midwifery Womens Health* 2013; **58**: 643-653 [PMID: 24131708]
- 34 **Chad L**, Pupco A, Bozzo P, Koren G. Update on antidepressant use during breastfeeding. *Can Fam Physician* 2013; **59**: 633-634 [PMID: 23766044]
- 35 **Berle JO**, Spigset O. Antidepressant Use During Breastfeeding. *Curr Womens Health Rev* 2011; **7**: 28-34 [PMID: 22299006 DOI: 10.2174/157340411794474784]
- 36 **Weiss J**, Dormann SM, Martin-Facklam M, Kerpen CJ, Ketabi-Kiyavash N, Haefeli WE. Inhibition of P-glycoprotein by newer antidepressants. *J Pharmacol Exp Ther* 2003; **305**: 197-204 [PMID: 12649369 DOI: 10.1124/jpet.102.046532]
- 37 **Karlsson L**, Carlsson B, Hiemke C, Ahlner J, Bengtsson F,

- Schmitt U, Kugelberg FC. Altered brain concentrations of citalopram and escitalopram in P-glycoprotein deficient mice after acute and chronic treatment. *Eur Neuropsychopharmacol* 2013; **23**: 1636-1644 [PMID: 23428338 DOI: 10.1016/j.euroneuro.2013.01.003]
- 38 Yasui-Furukori N, Saito M, Niioka T, Inoue Y, Sato Y, Kaneko S. Effect of itraconazole on pharmacokinetics of paroxetine: the role of gut transporters. *Ther Drug Monit* 2007; **29**: 45-48 [PMID: 17304149 DOI: 10.1097/FTD.0b013e31802bb20d]
- 39 Kapoor A, Iqbal M, Petropoulos S, Ho HL, Gibb W, Matthews SG. Effects of sertraline and fluoxetine on p-glycoprotein at barrier sites: in vivo and in vitro approaches. *PLoS One* 2013; **8**: e56525 [PMID: 23468867 DOI: 10.1371/journal.pone.0056525]
- 40 Wang JS, Zhu HJ, Gibson BB, Markowitz JS, Donovan JL, DeVane CL. Sertraline and its metabolite desmethylsertraline, but not bupropion or its three major metabolites, have high affinity for P-glycoprotein. *Biol Pharm Bull* 2008; **31**: 231-234 [PMID: 18239278]
- 41 Bhuiyan M, Petropoulos S, Gibb W, Matthews SG. Sertraline alters multidrug resistance phosphoglycoprotein activity in the mouse placenta and fetal blood-brain barrier. *Reprod Sci* 2012; **19**: 407-415 [PMID: 22510699 DOI: 10.1177/1933719111424438]
- 42 Saruwatari J, Yasui-Furukori N, Niioka T, Akamine Y, Takashima A, Kaneko S, Uno T. Different effects of the selective serotonin reuptake inhibitors fluvoxamine, paroxetine, and sertraline on the pharmacokinetics of fexofenadine in healthy volunteers. *J Clin Psychopharmacol* 2012; **32**: 195-199 [PMID: 22367658 DOI: 10.1097/JCP.0b013e318248ddb9]
- 43 Briggs GG, Ambrose PJ, Ilett KF, Hackett LP, Nageotte MP, Padilla G. Use of duloxetine in pregnancy and lactation. *Ann Pharmacother* 2009; **43**: 1898-1902 [PMID: 19809008 DOI: 10.1345/aph.1M317]
- 44 Konieczna A, Erdösová B, Lichnovská R, Jandl M, Cížková K, Ehrmann J. Differential expression of ABC transporters (MDR1, MRP1, BCRP) in developing human embryos. *J Mol Histol* 2011; **42**: 567-574 [PMID: 22012127 DOI: 10.1007/s10735-011-9363-1]
- 45 Bachmeier C, Levin GM, Beaulieu-Abdelahad D, Reed J, Mullan M. Effect of venlafaxine and desvenlafaxine on drug efflux protein expression and biodistribution in vivo. *J Pharm Sci* 2013; **102**: 3838-3843 [PMID: 23897419 DOI: 10.1002/jps.23680]
- 46 Bachmeier CJ, Beaulieu-Abdelahad D, Ganey NJ, Mullan MJ, Levin GM. Induction of drug efflux protein expression by venlafaxine but not desvenlafaxine. *Biopharm Drug Dispos* 2011; **32**: 233-244 [PMID: 21446053 DOI: 10.1002/bdd.753]
- 47 Müller MJ, Preuß C, Paul T, Streit F, Brandhorst G, Seeliger S. Serotonergic overstimulation in a preterm infant after sertraline intake via breastmilk. *Breastfeed Med* 2013; **8**: 327-329 [PMID: 23249132 DOI: 10.1089/bfm.2012.0084]
- 48 El Ela AA, Härtter S, Schmitt U, Hiemke C, Spahn-Langguth H, Langguth P. Identification of P-glycoprotein substrates and inhibitors among psychoactive compounds--implications for pharmacokinetics of selected substrates. *J Pharm Pharmacol* 2004; **56**: 967-975 [PMID: 15285840 DOI: 10.1211/0022357043969]
- 49 Iqbal MM, Aneja A, Rahman A, Megna J, Freemont W, Shiplo M, Nihilani N, Lee K. The potential risks of commonly prescribed antipsychotics: during pregnancy and lactation. *Psychiatry (Edgmont)* 2005; **2**: 36-44 [PMID: 21152171]
- 50 Friedman SH, Rosenthal MB. Treatment of perinatal delusional disorder: a case report. *Int J Psychiatry Med* 2003; **33**: 391-394 [PMID: 15152788]
- 51 Boulton DW, DeVane CL, Liston HL, Markowitz JS. In vitro P-glycoprotein affinity for atypical and conventional antipsychotics. *Life Sci* 2002; **71**: 163-169 [PMID: 12031686]
- 52 Linnet K, Ejsing TB. A review on the impact of P-glycoprotein on the penetration of drugs into the brain. Focus on psychotropic drugs. *Eur Neuropsychopharmacol* 2008; **18**: 157-169 [PMID: 17683917 DOI: 10.1016/j.euroneuro.2007.06.003]
- 53 Wang JS, Zhu HJ, Markowitz JS, Donovan JL, Yuan HJ, Devane CL. Antipsychotic drugs inhibit the function of breast cancer resistance protein. *Basic Clin Pharmacol Toxicol* 2008; **103**: 336-341 [PMID: 18834354 DOI: 10.1111/j.1742-7843.2008.00298.x]
- 54 Breedveld P, Zelcer N, Plum D, Sönmezer O, Tibben MM, Beijnen JH, Schinkel AH, van Tellingen O, Borst P, Schellens JH. Mechanism of the pharmacokinetic interaction between methotrexate and benzimidazoles: potential role for breast cancer resistance protein in clinical drug-drug interactions. *Cancer Res* 2004; **64**: 5804-5811 [PMID: 15313923 DOI: 10.1158/0008-5472.can-03-4062]
- 55 Nagasaka Y, Oda K, Iwatsubo T, Kawamura A, Usui T. Effects of aripiprazole and its active metabolite dehydroaripiprazole on the activities of drug efflux transporters expressed both in the intestine and at the blood-brain barrier. *Biopharm Drug Dispos* 2012; **33**: 304-315 [PMID: 22847220 DOI: 10.1002/bdd.1801]
- 56 Moons T, de Roo M, Claes S, Dom G. Relationship between P-glycoprotein and second-generation antipsychotics. *Pharmacogenomics* 2011; **12**: 1193-1211 [PMID: 21843066 DOI: 10.2217/pgs.11.55]
- 57 Gilad O, Merlob P, Stahl B, Klinger G. Outcome of infants exposed to olanzapine during breastfeeding. *Breastfeed Med* 2011; **6**: 55-58 [PMID: 21034242 DOI: 10.1089/bfm.2010.0027]
- 58 Lutz UC, Wiater G, Orlikowsky T, Gaertner HJ, Bartels M. Olanzapine treatment during breast feeding: a case report. *Ther Drug Monit* 2008; **30**: 399-401 [PMID: 18520614 DOI: 10.1097/FTD.0b013e31816850e2]
- 59 Hill RC, McIvor RJ, Wojnar-Horton RE, Hackett LP, Ilett KF. Risperidone distribution and excretion into human milk: case report and estimated infant exposure during breast-feeding. *J Clin Psychopharmacol* 2000; **20**: 285-286 [PMID: 10770482]
- 60 Ilett KF, Hackett LP, Kristensen JH, Vaddadi KS, Gardiner SJ, Begg EJ. Transfer of risperidone and 9-hydroxyrisperidone into human milk. *Ann Pharmacother* 2004; **38**: 273-276 [PMID: 14742766 DOI: 10.1345/aph.1D326]
- 61 Schlotterbeck P, Saur R, Hiemke C, Gründer G, Vehren T, Kircher T, Leube D. Low concentration of ziprasidone in human milk: a case report. *Int J Neuropsychopharmacol* 2009; **12**: 437-438 [PMID: 19203410 DOI: 10.1017/s1461145709009936]
- 62 Werremeyer A. Ziprasidone and citalopram use in pregnancy and lactation in a woman with psychotic depression. *Am J Psychiatry* 2009; **166**: 1298 [PMID: 19884241 DOI: 10.1176/appi.ajp.2009.09060765]
- 63 Teoh S, Ilett KF, Hackett LP, Kohan R. Estimation of rac-amisulpride transfer into milk and of infant dose via milk during its use in a lactating woman with bipolar disorder and schizophrenia. *Breastfeed Med* 2011; **6**: 85-88 [PMID: 20925494 DOI: 10.1089/bfm.2010.0016]
- 64 Schmitt U, Kirschbaum KM, Poller B, Kusch-Poddar M, Drewe J, Hiemke C, Gutmann H. In vitro P-glycoprotein efflux inhibition by atypical antipsychotics is in vivo nicely reflected by pharmacodynamic but less by pharmacokinetic changes. *Pharmacol Biochem Behav* 2012; **102**: 312-320 [PMID: 22525746 DOI: 10.1016/j.pbb.2012.04.002]
- 65 Wang JS, Zhu HJ, Markowitz JS, Donovan JL, DeVane CL. Evaluation of antipsychotic drugs as inhibitors of multidrug resistance transporter P-glycoprotein. *Psychopharmacology (Berl)* 2006; **187**: 415-423 [PMID: 16810505 DOI: 10.1007/s00213-006-0437-9]
- 66 Meador KJ. Breastfeeding and antiepileptic drugs. *JAMA* 2014; **311**: 1797-1798 [PMID: 24794373 DOI: 10.1212/WNL.0b013e3181ffe4a9]
- 67 Veiby G, Engelsen BA, Gilhus NE. Early child development and exposure to antiepileptic drugs prenatally and through breastfeeding: a prospective cohort study on children of women with epilepsy. *JAMA Neurol* 2013; **70**: 1367-1374 [PMID: 24061295 DOI: 10.1001/jamaneurol.2013.4290]
- 68 Mintzer S. To test our guess that breast is best: anticon-

- vulsants and breastfeeding. *Epilepsy Curr* 2011; **11**: 116-117 [PMID: 21852881 DOI: 10.5698/1535-7511-11.4.116]
- 69 **Pennell PB**, Gidal BE, Sabers A, Gordon J, Perucca E. Pharmacology of antiepileptic drugs during pregnancy and lactation. *Epilepsy Behav* 2007; **11**: 263-269 [PMID: 17996633 DOI: 10.1016/j.yebeh.2007.08.018]
- 70 **Crowe A**, Teoh YK. Limited P-glycoprotein mediated efflux for anti-epileptic drugs. *J Drug Target* 2006; **14**: 291-300 [PMID: 16882549 DOI: 10.1080/10611860600720814]
- 71 **Schinkel AH**, Wagenaar E, Mol CA, van Deemter L. P-glycoprotein in the blood-brain barrier of mice influences the brain penetration and pharmacological activity of many drugs. *J Clin Invest* 1996; **97**: 2517-2524 [PMID: 8647944 DOI: 10.1172/jci118699]
- 72 **Mahar Doan KM**, Humphreys JE, Webster LO, Wring SA, Shampine LJ, Serabjit-Singh CJ, Adkison KK, Polli JW. Passive permeability and P-glycoprotein-mediated efflux differentiate central nervous system (CNS) and non-CNS marketed drugs. *J Pharmacol Exp Ther* 2002; **303**: 1029-1037 [PMID: 12438524 DOI: 10.1124/jpet.102.039255]
- 73 **Löscher W**, Potschka H. Drug resistance in brain diseases and the role of drug efflux transporters. *Nat Rev Neurosci* 2005; **6**: 591-602 [PMID: 16025095]
- 74 **Feng B**, Mills JB, Davidson RE, Mireles RJ, Janiszewski JS, Troutman MD, de Morais SM. In vitro P-glycoprotein assays to predict the in vivo interactions of P-glycoprotein with drugs in the central nervous system. *Drug Metab Dispos* 2008; **36**: 268-275 [PMID: 17962372 DOI: 10.1124/dmd.107.017434]
- 75 **Nakanishi H**, Yonezawa A, Matsubara K, Yano I. Impact of P-glycoprotein and breast cancer resistance protein on the brain distribution of antiepileptic drugs in knockout mouse models. *Eur J Pharmacol* 2013; **710**: 20-28 [PMID: 23588114 DOI: 10.1016/j.ejphar.2013.03.049]
- 76 **Moerman L**, Wyffels L, Slaets D, Raedt R, Boon P, De Vos F. Antiepileptic drugs modulate P-glycoproteins in the brain: a mice study with (11)C-desmethyloperamide. *Epilepsy Res* 2011; **94**: 18-25 [PMID: 21277169 DOI: 10.1016/j.eplepsyres.2010.12.013]
- 77 **Zhang C**, Zuo Z, Kwan P, Baum L. In vitro transport profile of carbamazepine, oxcarbazepine, eslicarbazepine acetate, and their active metabolites by human P-glycoprotein. *Epilepsia* 2011; **52**: 1894-1904 [PMID: 21692796 DOI: 10.1111/j.1528-1167.2011.03140.x]
- 78 **Baltes S**, Gastens AM, Fedrowitz M, Potschka H, Kaever V, Löscher W. Differences in the transport of the antiepileptic drugs phenytoin, levetiracetam and carbamazepine by human and mouse P-glycoprotein. *Neuropharmacology* 2007; **52**: 333-346 [PMID: 17045309 DOI: 10.1016/j.neuropharm.2006.07.038]
- 79 **Dickens D**, Yusof SR, Abbott NJ, Weksler B, Romero IA, Couraud PO, Alfirevic A, Pirmohamed M, Owen A. A multi-system approach assessing the interaction of anticonvulsants with P-gp. *PLoS One* 2013; **8**: e64854 [PMID: 23741405 DOI: 10.1371/journal.pone.0064854]
- 80 **Nordmo E**, Aronsen L, Wasland K, Småbrekke L, Vorren S. Severe apnea in an infant exposed to lamotrigine in breast milk. *Ann Pharmacother* 2009; **43**: 1893-1897 [PMID: 19826099 DOI: 10.1345/aph.1M254]
- 81 **Cervený L**, Pavěk P, Maláková J, Staud F, Fendrich Z. Lack of interactions between breast cancer resistance protein (bcrp/abcg2) and selected antiepileptic agents. *Epilepsia* 2006; **47**: 461-468 [PMID: 16529607 DOI: 10.1111/j.1528-1167.2006.00453.x]

P- Reviewer: Catania VA, Pan W **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ



Pharmacophore approaches in protein kinase inhibitors design

Sergiy A Starosyla, Galyna P Volynets, Volodymyr G Bdzholo, Andriy G Golub, Sergiy M Yarmoluk

Sergiy A Starosyla, Galyna P Volynets, Volodymyr G Bdzholo, Sergiy M Yarmoluk, Department of Medicinal Chemistry, Institute of Molecular Biology and Genetics, NAS of Ukraine, 03680 Kyiv, Ukraine

Andriy G Golub, OTAVA Ltd., 400 Applewood Crescent, Unit 100, Vaughan, Ontario L4K0C3, Canada

Author contributions: Starosyla SA and Volynets GP contributed equally to this work, generated the figures and wrote the manuscript; Bdzholo VG and Golub AG contributed to the writing of the manuscript; Yarmoluk SM designed the aim of the editorial and wrote the manuscript.

Correspondence to: Dr. Sergiy M Yarmoluk, Sci., Professor, Department of Medicinal Chemistry, Institute of Molecular Biology and Genetics, NAS of Ukraine, 150 Zabolotnogo St., 03680 Kyiv, Ukraine. sergiy@yarmoluk.org.ua

Telephone: +38-44-5222458 Fax: +38-44-5222458

Received: June 27, 2014 Revised: October 15, 2014

Accepted: October 28, 2014

Published online: December 9, 2014

Abstract

Protein kinases constitute a superfamily of therapeutic targets for a number of human and animal diseases that include more than 500 members accordingly to sequencing data of the human genome. The well characterized nature of protein kinases makes them excellent targets for drug development. Pharmacophore approaches have become one of the major tools in the area of drug discovery. Application of pharmacophore modeling approaches allows reducing of expensive overall cost associated with drug development project. Pharmacophore models are important functional groups of atoms in the proper spatial position for interaction with target protein. Various ligand-based and structure-based methods have been developed for pharmacophore model generation. Despite the successes in pharmacophore models generation these approaches have not reached their full capacity in application for drug discovery. In the following review, we summarize the published data on pharmacophore models for inhibitors

of tyrosine protein kinases (EGFR, HER2, VEGFR, JAK2, JAK3, Syk, ZAP-70, Tie2) and inhibitors of serine/threonine kinases (Clk, Dyrk, Chk1, IKK2, CDK1, CDK2, PLK, JNK3, GSK3, mTOR, p38 MAPK, PKB). Here, we have described the achievements of pharmacophore modeling for protein kinase inhibitors, which provide key points for further application of generated pharmacophore hypotheses in virtual screening, de novo design and lead optimization.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Protein kinase; Inhibitor; Pharmacophore model; Receptor-based method; Ligand-based method

Core tip: In the following review, we summarize the published data on pharmacophore models for inhibitors of tyrosine protein kinases (EGFR, HER2, VEGFR, JAK2, JAK3, Syk, ZAP-70, Tie2) and inhibitors of serine/threonine kinases (Clk, Dyrk, Chk1, IKK2, CDK1, CDK2, PLK, JNK3, GSK3, mTOR, p38 MAPK, PKB). Here, we have described the achievements of pharmacophore modeling for protein kinase inhibitors, which provide key points for further application of generated pharmacophore hypotheses in virtual screening, de novo design and lead optimization.

Starosyla SA, Volynets GP, Bdzholo VG, Golub AG, Yarmoluk SM. Pharmacophore approaches in protein kinase inhibitors design. *World J Pharmacol* 2014; 3(4): 162-173 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/162.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.162>

INTRODUCTION

Protein kinases are a group of enzymes which covalently modify proteins by adding phosphate groups from adenosine triphosphate (ATP) to serine, threonine or tyro-

sine residues and therefore, transduce a variety of signals in eukaryotic cells^[1]. Kinases play a vital role in diverse cellular processes, functions, deregulations and now represent the second most important class of drug targets for pharmaceutical industry, after G-protein-coupled receptors^[2]. Over the past decade about 20 drugs targeting kinases have been approved for clinical application, and much more are currently undergoing clinical studies^[3].

Pharmacophore modeling is an important tool in drug development. A pharmacophore is the ensemble of steric and electronic features that is necessary to ensure the optimal supramolecular interactions with a specific biological target structure and to trigger (or to block) its biological response^[4]. There are two approaches for pharmacophore construction-receptor-based methods that allow building pharmacophore models based on the interactions of ligands with receptors, and ligand-based methods allowing generation of pharmacophore models based on the training sets of active compounds.

The pharmacophore models are applicable for screening large compound libraries *in silico* for the search of new small molecule inhibitors because they allow select compounds exhibiting binding features complementary oriented to an active binding pocket^[5].

In this review we discuss the published data on pharmacophore models for inhibitors of several tyrosine protein kinases and serine/threonine protein kinases (Table 1).

PHARMACOPHORE MODELING FOR PROTEIN TYROSINE KINASE INHIBITORS

Protein tyrosine kinases (PTKs) are a family of enzymes that can transfer phosphate group from ATP to tyrosine amino acid residues of target proteins in cell. This covalent post-translational modification is a crucial event for regulation of various biological processes including growth, metabolism, differentiation and apoptosis. Recent advances have demonstrated that tyrosine kinases play significant role in development of different diseases suggesting PTKs as attractive targets in the search for therapeutic agents.

Tyrosine kinases are classified as receptor tyrosine kinases such as FGFR, epidermal growth factor receptor (EGFR), Vascular endothelial growth factor receptor (VEGFR), TEK tyrosine kinase, endothelial (Tie2) and non-receptor tyrosine kinases such as Spleen tyrosine kinase (Syk), Zeta-chain-associated protein kinase 70 (ZAP-70), ABL, SRC, FAK, Janus kinase (JAK)^[6]. Each of receptor tyrosine kinases contains extracellular ligand-binding domain, transmembrane hydrophobic helix, and intracellular tyrosine protein kinase domain^[7]. Non-receptor tyrosine kinases are cytosolic proteins, possessing considerable structural variability. The non-receptor tyrosine kinases have a kinase domain and often include several additional protein-protein interacting domains like SH2, SH3 and the PH domain^[6].

EGFR inhibitors pharmacophore models

The EGFR family comprises four cell surface receptors:

HER1 (EGFR/erbB1), HER2 (erbB2), HER3 (erbB3) and HER4 (erbB4)^[7]. Binding of specific ligands to three of these receptors causes their dimerization and activation. HER2 is called an “orphan receptor” because it does not interact with any ligand, but it dimerizes with other ligand-bound members of EGFR family^[8].

HER1 (EGFR/erbB1): HER1 overexpression and over-activity are often associated with a wide range of cancers, including prostate, gastric, breast, colorectal, pancreatic, ovarian, lung cancers, head and neck squamous cell carcinoma and glioma^[9]. Aberrant EGFR signaling has been implicated in psoriasis, eczema and atherosclerosis^[10,11]. Therefore, the EGFR inhibitors can be used for the amelioration of these diseases.

Furet *et al*^[12] reported the first data concerning pharmacophore model for ATP-competitive inhibitors of EGFR. Accordingly to this Novartis pharmacophore hypothesis, the ATP-binding pocket in protein tyrosine kinases can be divided into five regions. In these five regions, three regions, namely, adenine region, sugar pocket, and hydrophobic region I, are primarily important to the binding affinity. Two other regions, hydrophobic region II and phosphate binding region are not of primary significance with respect to binding affinity, though they can be useful to enhance the inhibitor selectivity.

Also, pseudoreceptor model for EGFR was developed using a method FLARM. This model indicates the possible interactions between the receptor and ligand including two hydrogen bonds, one hydrophobic interaction and one sulfur-aromatic interaction, which are in accord with those in the Novartis pharmacophore model. Pharmacophore can be obtained according to the Novartis pharmacophore model and the pseudoreceptor model given by the FLARM method. 3D searching can then be done with the compound databases to find the lead compound of EGFR inhibitors^[13].

HER2 (erbB2): HER2 has been demonstrated to play an essential role in the development and progression of about 25%-30% of human primary breast and ovarian cancers^[14]. It was shown that the application of herceptin (a monoclonal antibody toward the HER2 receptor ectodomain) in combination with chemotherapy, leads to considerable regression of HER2-overexpressing metastatic breast tumors^[15]. Therefore, the inhibition of HER2 has been considered a promising way of controlling malignant tumors.

Two groups of small molecules have been found to possess inhibitory activity toward HER2. A ligand-based approach was used for building HER2 pharmacophore model^[16]. In the course of that work pharmacophore model generation was performed with Catalyst applying the Poling algorithm. From the calculated results, the best hypothesis bore good correlation with four features such as hydrogen bond donor, hydrogen bond acceptor, aliphatic and aromatic hydrophobic points. It seems that the formations of hydrogen bonds and the hydrophobic

Table 1 Protein kinases discussed in the review

Tyrosine protein kinases	
Epidermal growth factor receptor (EGFR; Erb-1; HER1 in humans)	Receptor
Human epidermal growth factor receptor 2 (HER2; erbB2; protooncogene Neu)	Receptor
Vascular endothelial growth factor receptor 2	Receptor
Janus kinase 2	Non-receptor
Janus kinase 3	Non-receptor
Spleen tyrosine kinase	Non-receptor
Zeta-chain-associated protein kinase 70	Non-receptor
TEK tyrosine kinase, endothelial	Receptor
Serine/threonine protein kinases	
Dual-specificity tyrosine-phosphorylation regulated kinase 1A	Non-receptor
Cdc2-like kinase	Non-receptor
Checkpoint kinase 1	Non-receptor
Human inhibitor nuclear-factor κB kinase 2	Non-receptor
Cyclin-dependent kinase 1	Non-receptor
Cyclin-dependent kinase 2	Non-receptor
Polo-like kinase	Non-receptor
c-Jun N-terminal kinase 3	Non-receptor
Glycogen synthase kinase 3	Non-receptor
Mammalian target of rapamycin	Non-receptor
p38 mitogen-activated protein kinase	Non-receptor
Protein kinase B	Non-receptor

interactions are crucial for ligand binding.

Pharmacophore modeling for VEGFR inhibitors

VEGFR signaling regulates vascular development, angiogenesis, lymphangiogenesis^[17] and has been involved in a wide range of human pathologies including cancer, atherosclerosis, and inflammatory diseases^[18]. Therefore, VEGFR has been emerged as an attractive therapeutic target. The pharmacophore models for VEGFR inhibitors were reported in^[18,19].

The ligand-based pharmacophore models were generated with Catalyst using the Poling algorithm and the “best conformational analysis” method. The best obtained hypothesis comprised four pharmacophore features: hydrogen bond donor, hydrogen bond acceptor, hydrophobic, and ring aromatic. The three-dimensional structure of ligand extracted from the crystal structure of 1YWN has been taken for shape query generation. The combined shape and hypothesis model was further used as a search query to screen Maybridge database. The query has been effectively performed to find one novel promising inhibitor of VEGFR kinase which possesses activity in cell lines^[18].

Two structure-based pharmacophore models of VEGFR-2 kinase inhibitors were built using the SBF software. The first pharmacophoric hypothesis was based on the crystal structure of 1Y6A, with the selection hydrogen bonding interaction for Glu915, Cys917, and Asn921. The second pharmacophoric model was based on the crystal structure of 1YWN; the backbone amide-NH of Cys917 and Asp1044 were used as hydrogen bond donors; and the backbone carbonyl oxygen of Glu883 was the hydrogen bond acceptor. The results suggest the importance of the five features for pharmacophores: the presence of two hydrogen bond donors, one hydrogen bond acceptor and two hydrophobic groups. The screening accuracy was assessed using a series of known inhibitors^[19].

JAK 2 and JAK 3: JAK2 and JAK3 are non-receptor protein tyrosine kinases involved in B-cell- and T-cell-mediated diseases^[20]. The inhibition of these kinases can be a potential strategy for the treatment of lymphoid-derived disorders.

Pharmacophore models for JAK2 and JAK3 were generated with PHASE, a high-performance program module of Schrödinger for ligand-based drug design. PHASE provides six pharmacophoric features: hydrogen bond donor (D), hydrogen bond acceptor (A), positively charged (P), negatively charged (N), hydrophobic (H), and aromatic ring (R) features. Two ligand-based pharmacophore hypotheses were constructed for the dataset of inhibitor molecules of JAK2 and JAK3 to dig out the essential structural features required for inhibition of both enzymes. These models can be helpful for screening of novel molecules having inhibitory activity toward both enzymes. The best hypothesis of JAK2 was ADRR, indicating that JAK2 inhibitors have one hydrogen bond acceptor (A), one hydrogen bond donor (D) and two ring aromatic (R) features. The best model of JAK3 was AD-DRR. Pearson correlation coefficient calculated for test set molecules demonstrated excellent predictive power of these hypotheses^[20].

Syk and ZAP-70: Syk and ZAP-70 are cytoplasmic non-receptor tyrosine kinases which play critical roles in the intracellular signal transduction of hematopoietic cells^[21]. Syk is a key mediator of immunoreceptor signalling in B-lymphocytes, mast cells, macrophages and neutrophils^[22-24], and ZAP-70 in T-lymphocytes, basophils and natural killer cells^[24,25]. Syk was shown to be an attractive drug target for therapy of type I hypersensitivity reactions such as allergic rhinitis, asthma, urticaria, anaphylaxis and autoimmune diseases such as multiple sclerosis, rheumatoid arthritis and systemic lupus erythematosus^[23,26].

For a number of Syk and ZAP-70 small molecule inhibitors two reliable pharmacophore models were built with PHASE. The generated pharmacophore hypotheses combined with docking calculations were taken for further multi-step systematic virtual screening and finally 27 dual inhibitors of Syk and ZAP-70 were obtained as hits^[24].

Also, 3D pharmacophore model of Syk inhibitors was developed by other authors applying HipHop and HypoRefine modules within Catalyst program package. Based on this model six compounds with good inhibitory potency against Syk were found^[21].

Tie2: Tie2 is a receptor tyrosine protein kinase expressed almost exclusively in endothelial cells which plays an important role in blood vessel formation. This receptor negatively regulates the inflammatory response in endothelial cells, suppressing VEGF- and TNF α -induced expression of leukocyte adhesion molecules and pro-coagulant tissue factor^[27]. Tie2 signaling also regulates pathologic angiogenesis, which includes tumor, psoriasis, choroidal neovascularization and rheumatoid arthritis angiogenesis^[28]. The implication of Tie2 in pathologic angiogenesis makes this cellular receptor an attractive therapeutic target.

All the pharmacophore modeling calculations of type I and type II kinase inhibitors of Tie2 were performed with HipHop and HypoRefine modules within Catalyst program package. In connection with the lack of highly active type I protein kinase inhibitors and restricted their structural diversity, only qualitative HipHop pharmacophore models were generated for this type inhibitors of Tie2. The best hypothesis comprised five pharmacophore features, namely, hydrogen bond donor, hydrogen bond acceptor, general hydrophobic, hydrophobic aromatic and ring aromatic. For type II kinase inhibitors of Tie2, at the first step, a HipHop model was built with the aim to identify the common pharmacophore features which can be essential for potent inhibitors. Then, based on the information obtained from the HipHop hypothesis for type II kinase inhibitors, the quantitative pharmacophore models were created with the aid of HypoRefine module. The best HypoRefine hypothesis included two hydrogen bond donors, one hydrophobic aromatic, two general hydrophobic features, and two excluded volumes. The validation of this HypoRefine model with the test set method demonstrated good correlation between the experimental and estimated IC₅₀ values, suggesting a good predictive power^[29].

PHARMACOPHORE MODELING FOR SERINE/THREONINE PROTEIN KINASE INHIBITORS

Serine/threonine protein kinases phosphorylate hydroxyl groups of serine or threonine residues of target proteins. Eukaryotic serine/threonine kinases can be classified into six groups: AGC, CaMK (for calcium-calmodulin dependent), CMGC (for CDK, MAP kinase, GSK and CDK-

like), STE (homologs of STE11 and STE20), CK1 (for casein kinase-1), and TKL group (tyrosine kinase like). Accordingly to analysis of available structural data for members of each of the large groups it was revealed that the protein kinases possess similar architecture^[30].

Dual-specificity tyrosine-phosphorylation regulated kinases and Cdc2-like kinases

Dual-specificity tyrosine-phosphorylation regulated kinases (Dyrk) proteins are defined as dual-specificity protein kinases because they can phosphorylate serine, threonine and tyrosine residues. Dyrk1A has increased expression in Down Syndrown individuals and is implicated in the development of other pathologies, such as neurodegeneration, cardiac hypertrophy and bone homeostasis^[31,32]. Hence, inhibition of Dyrk1A may have possible application as a therapeutical strategy for treatment of these diseases. Cdc2-like kinases (Clk) is implicated in the regulation of alternative splicing of mRNA isoforms, indicating that small molecule compounds able to modulate Clk activity may represent an important mechanism for the control of mRNA splicing^[33].

Pharmacophore models of Dyrk1A and Clk4 inhibitors were built based on the structure of the five most active compounds. Both hypotheses are represented with AAARR, indicating they comprise three hydrogen bond acceptors and two hydrophobic groups. The models associated with Dyrk1A and Clk4 have pharmacophore features located at the similar positions, considering both active sets have common structural cores. For both models, two hydrogen bond acceptors and one hydrophobic group are mapped to the quinazoline ring, which is shared among all studied compounds. The other two features, or one hydrogen bond acceptor and one hydrophobic group, are mapped to the R3 substituent 1,3-benzodioxol, which is common for tested inhibitors^[34].

Checkpoint kinase 1

Checkpoint kinase 1 (Chk1) is a serine/threonine protein kinase which plays an integral role in the regulation of cell cycle progression, normal cell division and is critical component for DNA damage response. The inhibition of Chk1 kinase has been shown to result in interrupting of the G₂/M checkpoint, which would permit premature mitotic entry in the presence of DNA damage, leading to cell death. This suggests a potential therapeutic use of Chk1 inhibitors in cancer therapy^[35].

All the pharmacophore modeling calculations for Chk1 were performed with Catalyst software package. The common pharmacophore features essential for promising Chk1 inhibitors were found with HipHop module. The best model involves four types of features, namely, hydrogen bond donor, hydrogen bond acceptor, ring aromatic and hydrophobic feature, indicating that the four types of features are important for potent Chk1 inhibitors^[36].

Human inhibitor nuclear-factor κ B kinase 2

The human inhibitor nuclear-factor κ B kinase 2 (hIKK-2) is a serine/threonine protein kinase which belongs to the

IKK complex and implicated in the activation of nuclear-factor κ B transcription factor under inflammatory conditions. The inhibitors of hIKK-2 could have strong therapeutic potential for treatment of chronic inflammatory diseases.

The structure-based pharmacophore model for hIKK-2 was built by using LigandScout software based on the protein-ligand complexes which were obtained by the docking process of ATP-competitive inhibitors into active site of hIKK-2.

The ligand poses which satisfied the common pharmacophore features of protein kinase inhibitors necessary for interaction with ATP-binding site [ability to form hydrogen bonds with the amino acid residues in the hinge region (segment 96-99 in hIKK-2 sequence) and hydrophobic interactions with the hydrophobic cavity in the active site of hIKK-2 (for example, Val29, Lys44, Ile65 and Val152)] were taken as knowledge-based coherent. As a result of this analysis, 43 poses of the 21 hIKK-2 inhibitors were considered as knowledge-based coherent, and their corresponding sites (functional groups which form intermolecular interactions with the kinase domain of hIKK-2) were selected to generate structure-based pharmacophore model. This hypothesis comprised two hydrogen bond donors, one hydrogen bond acceptor and one hydrophobic group common to most of the 43 poses^[37].

Cyclin-dependent kinase 1

Cyclin-dependent kinase 1 (CDK1) is a serine/threonine protein kinase which plays a key role in promoting mitosis^[38]. It was shown, that CDK1 inhibitors effectively blocked cell cycle progression in human tumor cell lines, indicating their potential clinical application as anticancer drugs^[39].

A number of reliable binding hypotheses for CDK1 inhibitors were constructed with HypoGen module within Catalyst software package. HypoGen identifies a three-dimensional array of a maximum of five chemical features shared among the active ligands from training set providing relative alignment for each input molecule compatible with binding to target protein active site. The considered pharmacophore features can be hydrogen bond donors, hydrogen bond acceptors, aromatic planes, aliphatic, hydrophobic, positive and negative ionizable groups. The conformational flexibility of compounds from training set is modeled by generating multiple conformers covering a specified energy range for each input molecule. Successful pharmacophore models are complemented with exclusion spheres. Optimal sterically refined models obtained for CDK1 inhibitors were selected as search queries to screen the NCI, drugs and agrochemicals libraries. As a result, ten compounds demonstrated low micromolar activity toward CDK1, suggesting that generated pharmacophore hypothesis can be useful for search of potential anti-CDK1 agents^[40].

Cyclin-dependent kinase 2

Cyclin-dependent kinase 2 (CDK2) is important protein

kinase for initiation of DNA synthesis in higher eukaryotes and is required for promoting the cell division cycle and for successful progression through S and G₂ phases^[41]. The importance of CDK2 for cell cycle progression has led to an active search of small molecule compounds inhibiting this enzyme as potential anticancer drugs.

Several ligand-based pharmacophore models for CDK2 small molecule inhibitors were generated independently with Catalyst by Hecker *et al.*^[42], Toba *et al.*^[43] and Vadivelan *et al.*^[44]. The multicomplex-based comprehensive pharmacophore map was built with LigandScout software by Zou *et al.*^[45]. It should be noted that during pharmacophore model construction Catalyst software takes into consideration only ligand information whereas LigandScout adds pharmacophore feature to the model when important interaction pattern between inhibitor and receptor is identified.

The authors of multicomplex-based comprehensive pharmacophore map compared their hypothesis with other reported CDK2 inhibitors models. It was revealed that each pharmacophore feature in the ligand-based models was mapped to the corresponding feature in comprehensive pharmacophore map suggesting that the last one includes more information over all three other models. Detailedly, during alignment of the final Hecker model and multicomplex-based comprehensive map it was shown that hydrogen bond acceptor feature in Hecker model was mapped to the feature of comprehensive map reflecting the interaction of small-molecule inhibitor with hinge region (A1); the hydrogen bond donor feature in Hecker model was matched to the feature representing the interaction with Gln131 (D5); the hydrophobic features were mapped to the features located at the solvent-accessible region (H1) and in the ribose-phosphate binding site (H3). The pharmacophore model generated by Toba *et al.*^[43] comprised two hydrogen bond donor features and three hydrophobic features. Each of these features was matched to the corresponding features D1, D5, H1, H2 and H3 of comprehensive pharmacophore map. The pharmacophore hypothesis constructed by Vadivelan *et al.*^[44] included two hydrogen bond acceptors, one hydrogen bond donor, and one hydrophobic feature. In comparison with comprehensive pharmacophore map, one of the hydrogen bond acceptor features was mapped to the feature that is located near Asp86 (A3), other one was matched to the feature representing the interaction with Lys33 (A4). The hydrogen bond donor feature and hydrophobic feature of Vadivelan model correspond to D1 and H3 features of multicomplex-based comprehensive pharmacophore map, respectively^[45].

Polo-like kinases

Polo-like kinases (PLKs) belong to a family of serine/threonine protein kinases and exist in four isoforms, namely, PLK1, PLK2, PLK3, and PLK4. The only one of these isoforms, PLK1, is shown to be implicated in the regulation of chromosome segregation, centrosome maturation, bipolar spindle formation and execution of

cytokinesis^[46]. The activity of PLK1 is increased in many tumor types, including lung, breast, colon, pancreatic, prostate and ovarian indicating its capability as a drug target^[47].

The chemical feature-based pharmacophore models of PLK1 inhibitors were constructed by using HipHop and HypoGen modules within Catalyst program package. The best qualitative HipHop pharmacophore hypothesis contains seven features, namely, hydrophobic aromatic feature, two hydrophobic aliphatic moieties, three hydrogen bond acceptors and one hydrogen bond donor. The best quantitative HypoGen pharmacophore model, possessing the lowest rmsd value and the highest correlation coefficient, includes four features, namely, general hydrophobic, hydrophobic aliphatic, hydrogen bond acceptor and hydrogen bond donor. The results of validation for HypoGen pharmacophore model, which were obtained with the aid of the test set method, have demonstrated a really good correlation between the experimental and estimated IC₅₀ values suggesting a good predictive ability^[48].

c-Jun N-terminal kinase 3

The c-Jun N-terminal kinase 3 (JNK3) is a member of the mitogen-activated protein kinase (MAPK) family, which activates signaling pathways under environmental stress conditions^[49]. JNK3 is expressed selectively in brain, heart, and testis^[50]. It was shown, that JNK3 phosphorylates β -amyloid precursor protein, a conserved and ubiquitously expressed transmembrane glycoprotein involved in the development of Alzheimer's disease^[51]. Therefore, JNK3 appears to be an attractive therapeutic target for this neurodegenerative disease.

Pharmacophore models for JNK3 small-molecule inhibitors were generated with Catalyst software package. X-ray crystal structure of JNK3 (PDB ID: 2R9S) was taken for structure-based pharmacophore modeling and molecular docking simulations. A structure-based pharmacophore hypothesis was built using interaction generation module implemented in Discovery Studio. The most important interaction patterns were transformed into pharmacophore features, such as hydrophobes, hydrogen bond donors and hydrogen bond acceptors along with their direction vectors. The final refined structure-based pharmacophore model included hydrogen bond donor features with Lys68, Gly71, Ser72, Gln155, Met149 and hydrogen bond acceptors with Lys68, Gly71, Met149, Gln155 and three hydrophobic features.

The features obtained in the ligand-based pharmacophore model are well compared to the features of structure-based pharmacophore model. But, structure-based pharmacophore model had three additional features, not present in ligand-based pharmacophore hypothesis, which would be helpful for development of novel JNK3 inhibitors^[49].

Glycogen synthase kinase 3

Glycogen synthase kinase 3 (GSK-3) is a serine/threonine protein kinase highly expressed in the nervous

system, which regulates glycogen metabolism by insulin, and is involved in many different biological processes such as tumorigenesis, cell survival, and developmental patterning^[52]. GSK-3 has recently emerged as a promising therapeutic target for the search of small molecule inhibitors which can be potential novel drug candidates for treatment of several human pathologies, including cancer, Alzheimer's disease, stroke, bipolar disorders, type II diabetes and chronic inflammatory processes^[53].

Pharmacophore models for GSK-3 inhibitors were constructed with the HypoGen module within the Catalyst software package based on list of 152 GSK-3 inhibitors. HypoGen allows automatic pharmacophore generation based on a library of at least 16 molecules with inhibitory activity toward proposed molecular target ranging over 4 orders of magnitude^[5].

3D pharmacophore mapping methodology based on distance comparison technique was designed for the three GSK-3 inhibitors using DISCOtech™ module implemented in SYBYL 8.0. DISCOtech™ is a well established module in constructing pharmacophoric map. Taking into consideration a set of molecules which are characterized by the ability to interact with the same protein receptor, DISCOtech™ identifies features that could be components in a pharmacophore hypothesis. DISCOtech™ operates in distance space and can perform clique detection to build pharmacophore models on up to 300 conformers per molecule. Therefore, DISCOtech™ can be efficiently applied with at least 3-5 compounds to design reliable pharmacophore hypotheses^[54].

Mammalian target of rapamycin

Mammalian target of rapamycin (mTOR) is a ubiquitous serine/threonine protein kinase that regulates several important physiological functions like protein synthesis, metabolism, cell growth, proliferation, and autophagy. mTOR is also critical for a number of brain-specific mechanisms, such as synaptic plasticity, learning, and cortical development^[55]. Recent studies have implicated mTOR to several human pathologies including cancer, diabetes, obesity, cardiovascular diseases and neurological disorders^[56]. The pharmaceutical attractiveness of small molecule mTOR inhibitors coupled with the deficiency of crystallographic structural data for mTOR kinase domain, were starting point for development of ligand-based QSAR and pharmacophore models^[57].

The Hip-Hop pharmacophore model was created with the Common Feature Pharmacophore Generation module implemented in Accelrys Discovery Studio 2.1. This pharmacophore hypothesis provides a geometrical representation of the features necessary for ligands to interact favorably with a receptor site and demonstrate biological activity. Hip-Hop identifies configurations or three-dimensional spatial arrangements of chemical features that are shared among all molecules in the set. Under the Common Feature Pharmacophore Generation protocol, were used four features such as hydrogen bond donor, hydrogen bond acceptor, ring aromatic, and hy-

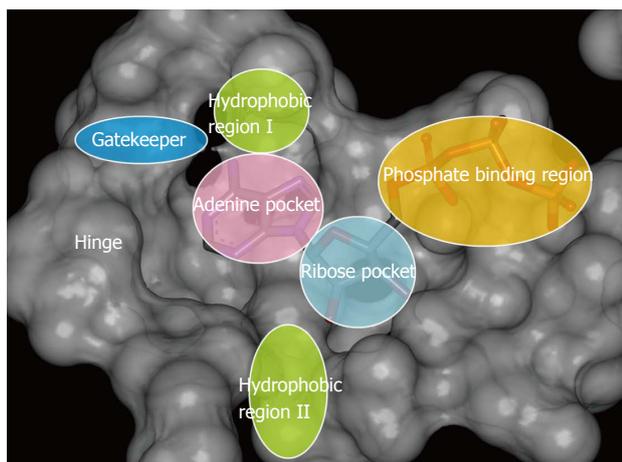


Figure 1 Pharmacophore model for type I protein kinase inhibitors.

drophobic to build the pharmacophore model.

The best pharmacophore hypothesis generated from 27 ATP-competitive inhibitors of mTOR comprised two hydrogen bond acceptors, one hydrophobic feature and one aromatic ring feature^[58].

p38 mitogen-activated protein kinase

The p38 mitogen-activated protein (MAP) kinase (p38MAPK) is a serine/threonine protein kinase which plays a very important role in the pathophysiology of several inflammatory human diseases, such as, asthma, osteoarthritis and rheumatoid arthritis, a chronic obstructive pulmonary disease. Therefore, the inhibition of p38MAPK can be an effective strategy to prevent the development of these diseases^[59].

Catalyst HypoGen pharmacophore approach was applied to obtain models for a collection of p38MAPK inhibitors^[59,60]. Eight out of ten best hypotheses comprise the identical four features: one hydrogen bond acceptor, two hydrophobic aromatic and one hydrophobic feature, which indicates the stability of the models^[59]. The obtained hypotheses are readily interpretable and can be applied for the rational discovery of new p38MAPK inhibitors^[60].

Protein kinase B (PKB; Akt)

The protein kinase B (PKB; Akt) family of serine/threonine kinases consists of three members: Akt1/PKB α , Akt2/PKB β , and Akt3/PKB γ ^[61]. Akt is a central component in cell signaling pathways regulated by growth factors, cytokines, and other cellular stimuli. The activation of Akt leads to cell cycle progression (inhibiting apoptosis)^[62]. Ligand-based pharmacophore model of Akt inhibitors was built using DISCOtech and GASP (genetic algorithm similarity program) module^[63].

A crystal structure of Akt2 complexed with a known inhibitor (PDB ID: 3E8D) was taken for construction of structure-based pharmacophore hypothesis. The Interaction Generation protocol within DS program was used to create pharmacophoric features corresponding to all important interaction points at the ATP-binding pocket of

Akt2. The obtained pharmacophore model consisted of seven pharmacophoric features, namely, hydrogen bond donor (HD), two hydrogen bond acceptors (HA1-2), and four hydrophobic groups (HY1-4), besides, eighteen exclusion volume spheres were also taken into account. HD is at the neighborhood of the carboxyl group of Asp293. HA1 is positioned to interact with the amino group of Ala232. HA2 is located near amino group of Phe294 and Asp293. Groups in accordance with these pharmacophoric features potentially able form hydrogen bonds with adjacent amino acid residues. HY1 is located in a hydrophobic pocket formed by Ala178, Met282 and Phe439. HY2 is situated in another hydrophobic pocket composed by Gly159, Gly161, Gly164 and Val166. HY3 is close to Lys181 and Met229 and HY3 is near to Phe294. There are short distances between HY4 and hydrophobic amino acids Phe163 and Lys181. Groups in accordance with these hydrophobic features may be involved in hydrophobic interactions with enzyme. Therefore, small-molecule compounds matching with some of these features may be potential inhibitors of Akt2^[64].

DISCUSSION

The methods for pharmacophore model generation are divided in two categories: receptor-based and ligand-based. Receptor-based approaches can be used when the structure of molecular target is determined. In other case, ligand-based approaches can be applied for pharmacophore hypothesis generation.

During analysis of the pharmacophore approaches in protein kinase inhibitors design, it was revealed that despite of large amount of the structural data for protein kinases, the ligand-based approaches are more widely used for protein kinase pharmacophore model generation than the receptor-based. Ligand-based methods for pharmacophore elucidation include ALADDIN, DISCO, GERM, COMPASS, GASP, Catalyst HipHop, SCAMPI, Catalyst HypoGen, Phase, CLEW GAMMA, PARM, DANTE, *etc.*

3D-QSAR methods Catalyst HypoGen and Phase which use the known activity values of the small molecule inhibitors in the training set to build the hypothesis, are the most applied for protein kinase inhibitors pharmacophore models generation. These models include features common only for highly active compounds and also can contain excluded volumes (obtained based on the structure of inactive compounds), which couldn't be occupied by inhibitors. The methods Catalyst HipHop, PharmaGist, DISCO can be used only for qualitative pharmacophore models generation which don't take into consideration the information concerning activity of compounds. Qualitative pharmacophore model can be taken as a basis for further 3D-QSAR hypothesis generation.

The receptor-based methods of pharmacophore model elucidation are more rarely used for protein kinase pharmacophore design. These approaches can be useful

- 2 **Cohen P.** Protein kinases--the major drug targets of the twenty-first century? *Nat Rev Drug Discov* 2002; **1**: 309-315 [PMID: 12120282 DOI: 10.1038/nrd773]
- 3 **Cohen P, Alessi DR.** Kinase drug discovery--what's next in the field? *ACS Chem Biol* 2013; **8**: 96-104 [PMID: 23276252 DOI: 10.1021/cb300610s]
- 4 **Wermuth CG, Ganellin CR, Lindberg P, Mitscher LA.** Glossary of terms used in medicinal chemistry. *Annu Rep Med Chem* 1998; **33**: 385-395 [DOI: 10.1016/S0065-7743(08)611101-X]
- 5 **Taha MO, Bustanji Y, Al-Ghussein MA, Mohammad M, Zalloum H, Al-Masri IM, Atallah N.** Pharmacophore modeling, quantitative structure-activity relationship analysis, and in silico screening reveal potent glycogen synthase kinase-3beta inhibitory activities for cimetidine, hydroxychloroquine, and gemifloxacin. *J Med Chem* 2008; **51**: 2062-2077 [PMID: 18324764 DOI: 10.1021/jm7009765]
- 6 **Paul MK, Mukhopadhyay AK.** Tyrosine kinase - Role and significance in Cancer. *Int J Med Sci* 2004; **1**: 101-115 [PMID: 15912202 DOI: 10.7150/ijms.1.101]
- 7 **Dowsett M, Cooke T, Ellis I, Gullick WJ, Gusterson B, Mallon E, Walker R.** Assessment of HER2 status in breast cancer: why, when and how? *Eur J Cancer* 2000; **36**: 170-176 [PMID: 10741274 DOI: 10.1016/S0959-8049(99)00264-6]
- 8 **Brennan PJ, Kumagai T, Berezov A, Murali R, Greene MI.** HER2/Neu: mechanisms of dimerization/oligomerization. *Oncogene* 2002; **21**: 328 [PMID: 11840330 DOI: 10.1038/sj/onc1205119]
- 9 **Giaccone G.** HER1/EGFR-targeted agents: predicting the future for patients with unpredictable outcomes to therapy. *Ann Oncol* 2005; **16**: 538-548 [PMID: 15746148 DOI: 10.1093/annonc/mdi129]
- 10 **Jost M, Kari C, Rodeck U.** The EGF receptor - an essential regulator of multiple epidermal functions. *Eur J Dermatol* 2000; **10**: 505-510 [PMID: 11056418]
- 11 **Dreux AC, Lamb DJ, Modjtahedi H, Ferns GA.** The epidermal growth factor receptors and their family of ligands: their putative role in atherogenesis. *Atherosclerosis* 2006; **186**: 38-53 [PMID: 16076471 DOI: 10.1016/j.atherosclerosis.2005.06.038]
- 12 **Furet P, Caravatti G, Lydon N, Priestle JP, Sowadski JM, Trinks U, Traxler P.** Modelling study of protein kinase inhibitors: binding mode of staurosporine and origin of the selectivity of CGP 52411. *J Comput Aided Mol Des* 1995; **9**: 465-472 [PMID: 8789188]
- 13 **Peng T, Pei J, Zhou J.** 3D-QSAR and receptor modeling of tyrosine kinase inhibitors with flexible atom receptor model (FLARM). *J Chem Inf Comput Sci* 2003; **43**: 298-303 [PMID: 12546565 DOI: 10.1021/ci0256034]
- 14 **Slamon DJ, Godolphin W, Jones LA, Holt JA, Wong SG, Keith DE, Levin WJ, Stuart SG, Udove J, Ullrich A.** Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 1989; **244**: 707-712 [PMID: 2470152 DOI: 10.1126/science.2470152]
- 15 **Arteaga CL.** Trastuzumab, an appropriate first-line single-agent therapy for HER2-overexpressing metastatic breast cancer. *Breast Cancer Res* 2003; **5**: 96-100 [PMID: 12631388 DOI: 10.1186/bcr574]
- 16 **Zhu LL, Hou TJ, Chen LR, Xu XJ.** 3D QSAR analyses of novel tyrosine kinase inhibitors based on pharmacophore alignment. *J Chem Inf Comput Sci* 2001; **41**: 1032-1040 [PMID: 11500121 DOI: 10.1021/ci010002i]
- 17 **Olsson AK, Dimberg A, Kreuger J, Claesson-Welsh L.** VEGF receptor signalling - in control of vascular function. *Nat Rev Mol Cell Biol* 2006; **7**: 359-371 [PMID: 16633338 DOI: 10.1038/nrm1911]
- 18 **Yu H, Wang Z, Zhang L, Zhang J, Huang Q.** The discovery of novel vascular endothelial growth factor receptor tyrosine kinases inhibitors: pharmacophore modeling, virtual screening and docking studies. *Chem Biol Drug Des* 2007; **69**: 204-211 [PMID: 17441906 DOI: 10.1111/j.1747-0285.2007.00488.x]
- 19 **Lee K, Jeong KW, Lee Y, Song JY, Kim MS, Lee GS, Kim Y.** Pharmacophore modeling and virtual screening studies for new VEGFR-2 kinase inhibitors. *Eur J Med Chem* 2010; **45**: 5420-5427 [PMID: 20869793 DOI: 10.1016/j.ejmech.2010.09.002]
- 20 **Jasuja H, Chadha N, Kaur M, Silakari O.** Dual inhibitors of Janus kinase 2 and 3 (JAK2/3): designing by pharmacophore- and docking-based virtual screening approach. *Mol Divers* 2014; **18**: 253-267 [PMID: 24415188 DOI: 10.1007/s11030-013-9497-z]
- 21 **Xie HZ, Li LL, Ren JX, Zou J, Yang L, Wei YQ, Yang SY.** Pharmacophore modeling study based on known spleen tyrosine kinase inhibitors together with virtual screening for identifying novel inhibitors. *Bioorg Med Chem Lett* 2009; **19**: 1944-1949 [PMID: 19254842 DOI: 10.1016/j.bmcl.2009.02.049]
- 22 **Riccaboni M, Bianchi I, Petrillo P.** Spleen tyrosine kinases: biology, therapeutic targets and drugs. *Drug Discov Today* 2010; **15**: 517-530 [PMID: 20553955 DOI: 10.1016/j.drudis.2010.05.001]
- 23 **Wong BR, Grossbard EB, Payan DG, Masuda ES.** Targeting Syk as a treatment for allergic and autoimmune disorders. *Expert Opin Investig Drugs* 2004; **13**: 743-762 [PMID: 15212616 DOI: 10.1517/13543784.13.7.743]
- 24 **Kaur M, Kumari A, Bahia MS, Silakari O.** Designing of new multi-targeted inhibitors of spleen tyrosine kinase (Syk) and zeta-associated protein of 70kDa (ZAP-70) using hierarchical virtual screening protocol. *J Mol Graph Model* 2013; **39**: 165-175 [PMID: 23280414 DOI: 10.1016/j.jmgm.2012.11.011]
- 25 **Wang H, Kadlecck TA, Au-Yeung BB, Goodfellow HE, Hsu LY, Freedman TS, Weiss A.** ZAP-70: an essential kinase in T-cell signaling. *Cold Spring Harb Perspect Biol* 2010; **2**: a002279 [PMID: 20452964 DOI: 10.1101/cshperspect.a002279]
- 26 **Mazuc E, Villoutreix BO, Malbec O, Roumier T, Fleury S, Leonetti JP, Dombrowicz D, Daëron M, Martineau P, Dariavach P.** A novel druglike spleen tyrosine kinase binder prevents anaphylactic shock when administered orally. *J Allergy Clin Immunol* 2008; **122**: 188-194, 194.e1-3 [PMID: 18539317 DOI: 10.1016/j.jaci.2008.04.026]
- 27 **Hughes DP, Marron MB, Brindle NP.** The antiinflammatory endothelial tyrosine kinase Tie2 interacts with a novel nuclear factor-kappaB inhibitor ABIN-2. *Circ Res* 2003; **92**: 630-636 [PMID: 12609966 DOI: 10.1161/01.RES.0000063422.38690.DC]
- 28 **Martin V, Liu D, Fueyo J, Gomez-Manzano C.** Tie2: a journey from normal angiogenesis to cancer and beyond. *Histol Histopathol* 2008; **23**: 773-780 [PMID: 18366015]
- 29 **Xie QQ, Xie HZ, Ren JX, Li LL, Yang SY.** Pharmacophore modeling studies of type I and type II kinase inhibitors of Tie2. *J Mol Graph Model* 2009; **27**: 751-758 [PMID: 19138543 DOI: 10.1016/j.jmgm.2008.11.008]
- 30 **Goldsmith EJ, Akella R, Min X, Zhou T, Humphreys JM.** Substrate and docking interactions in Ser/Thr protein kinases. *Chem Rev* 2007; **107**: 5065-5081 [DOI: 10.1021/cr068221w]
- 31 **Aranda S, Laguna A, de la Luna S.** DYRK family of protein kinases: evolutionary relationships, biochemical properties, and functional roles. *FASEB J* 2011; **25**: 449-462 [PMID: 21048044 DOI: 10.1096/fj.10-165837]
- 32 **Park J, Song WJ, Chung KC.** Function and regulation of Dyrk1A: towards understanding Down syndrome. *Cell Mol Life Sci* 2009; **66**: 3235-3240 [PMID: 19685005 DOI: 10.1007/s00018-009-0123-2]
- 33 **Mott BT, Tanega C, Shen M, Maloney DJ, Shinn P, Leister W, Marugan JJ, Inglese J, Austin CP, Misteli T, Auld DS, Thomas CJ.** Evaluation of substituted 6-arylquinazolin-4-amines as potent and selective inhibitors of cdc2-like kinases (Clk). *Bioorg Med Chem Lett* 2009; **19**: 6700-6705 [PMID: 19837585 DOI: 10.1016/j.bmcl.2009.09.121]
- 34 **Pan Y, Wang Y, Bryant SH.** Pharmacophore and 3D-QSAR characterization of 6-arylquinazolin-4-amines as Cdc2-like

- kinase 4 (Clk4) and dual specificity tyrosine-phosphorylation-regulated kinase 1A (Dyrk1A) inhibitors. *J Chem Inf Model* 2013; **53**: 938-947 [PMID: 23496085 DOI: 10.1021/ci300635c]
- 35 **McNeely S**, Beckmann R, Bence Lin AK. CHEK again: revisiting the development of CHK1 inhibitors for cancer therapy. *Pharmacol Ther* 2014; **142**: 1-10 [PMID: 24140082 DOI: 10.1016/j.pharmthera.2013.10.005]
- 36 **Chen JJ**, Liu TL, Yang LJ, Li LL, Wei YQ, Yang SY. Pharmacophore modeling and virtual screening studies of checkpoint kinase 1 inhibitors. *Chem Pharm Bull (Tokyo)* 2009; **57**: 704-709 [PMID: 19571415 DOI: 10.1248/cpb.57.704]
- 37 **Sala E**, Guasch L, Iwazskiewicz J, Mulero M, Salvadó MJ, Pinent M, Zoete V, Grosdidier A, Garcia-Vallvé S, Michielin O, Pujadas G. Identification of human IKK-2 inhibitors of natural origin (part I): modeling of the IKK-2 kinase domain, virtual screening and activity assays. *PLoS One* 2011; **6**: e16903 [PMID: 21390216 DOI: 10.1371/journal.pone.0016903]
- 38 **Chow JP**, Poon RY, Ma HT. Inhibitory phosphorylation of cyclin-dependent kinase 1 as a compensatory mechanism for mitosis exit. *Mol Cell Biol* 2011; **31**: 1478-1491 [PMID: 21262764 DOI: 10.1128/MCB.00891-10]
- 39 **Chen S**, Chen L, Le NT, Zhao C, Sidduri A, Lou JP, Michoud C, Portland L, Jackson N, Liu JJ, Konzelmann F, Chi F, Tovar C, Xiang Q, Chen Y, Wen Y, Vassilev LT. Synthesis and activity of quinolinyl-methylene-thiazolinones as potent and selective cyclin-dependent kinase 1 inhibitors. *Bioorg Med Chem Lett* 2007; **17**: 2134-2138 [PMID: 17303421 DOI: 10.1016/j.bmcl.2007.01.081]
- 40 **Al-Sha'er MA**, Taha MO. Discovery of novel CDK1 inhibitors by combining pharmacophore modeling, QSAR analysis and in silico screening followed by in vitro bioassay. *Eur J Med Chem* 2010; **45**: 4316-4330 [PMID: 20638755 DOI: 10.1016/j.ejmech.2010.06.034]
- 41 **Hu B**, Mitra J, van den Heuvel S, Enders GH. S and G2 phase roles for Cdk2 revealed by inducible expression of a dominant-negative mutant in human cells. *Mol Cell Biol* 2001; **21**: 2755-2766 [PMID: 11283255 DOI: 10.1128/MCB.21.8.27-55-27.66.2001]
- 42 **Hecker EA**, Duraiswami C, Andrea TA, Diller DJ. Use of catalyst pharmacophore models for screening of large combinatorial libraries. *J Chem Inf Comput Sci* 2002; **42**: 1204-1211 [PMID: 12377010 DOI: 10.1021/ci020368a]
- 43 **Toba S**, Srinivasan J, Maynard AJ, Sutter J. Using pharmacophore models to gain insight into structural binding and virtual screening: an application study with CDK2 and human DHFR. *J Chem Inf Model* 2006; **46**: 728-735 [PMID: 16563003 DOI: 10.1021/ci050410c]
- 44 **Vadivelan S**, Sinha BN, Irudayam SJ, Jagarlapudi SA. Virtual screening studies to design potent CDK2-cyclin A inhibitors. *J Chem Inf Model* 2007; **47**: 1526-1535 [PMID: 17523616 DOI: 10.1021/ci7000742]
- 45 **Zou J**, Xie HZ, Yang SY, Chen JJ, Ren JX, Wei YQ. Towards more accurate pharmacophore modeling: Multicomplex-based comprehensive pharmacophore map and most-frequent-feature pharmacophore model of CDK2. *J Mol Graph Model* 2008; **27**: 430-438 [PMID: 18786843 DOI: 10.1016/j.jmkgm.2008.07.004]
- 46 **Stewart HJ**, Kishikova L, Powell FL, Wheatley SP, Chevassut TJ. The polo-like kinase inhibitor BI 2536 exhibits potent activity against malignant plasma cells and represents a novel therapy in multiple myeloma. *Exp Hematol* 2011; **39**: 330-338 [PMID: 21184800 DOI: 10.1016/j.exphem.2010.12.006]
- 47 **Chopra P**, Sethi G, Dastidar SG, Ray A. Polo-like kinase inhibitors: an emerging opportunity for cancer therapeutics. *Expert Opin Investig Drugs* 2010; **19**: 27-43 [PMID: 20001553 DOI: 0.1517/13543780903483191]
- 48 **Wang HY**, Cao ZX, Li LL, Jiang PD, Zhao YL, Luo SD, Yang L, Wei YQ, Yang SY. Pharmacophore modeling and virtual screening for designing potential PLK1 inhibitors. *Bioorg Med Chem Lett* 2008; **18**: 4972-4977 [PMID: 18762425 DOI: 10.1016/j.bmcl.2008.08.033]
- 49 **Kumar BV**, Kotla R, Buddiga R, Roy J, Singh SS, Gundla R, Ravikumar M, Sarma JA. Ligand-based and structure-based approaches in identifying ideal pharmacophore against c-Jun N-terminal kinase-3. *J Mol Model* 2011; **17**: 151-163 [PMID: 20393763 DOI: 10.1007/s00894-010-0701-0]
- 50 **Kyriakis JM**, Avruch J. Mammalian MAPK signal transduction pathways activated by stress and inflammation: a 10-year update. *Physiol Rev* 2012; **92**: 689-737 [PMID: 22535895 DOI: 10.1152/physrev.00028.2011]
- 51 **Kimberly WT**, Zheng JB, Town T, Flavell RA, Selkoe DJ. Physiological regulation of the beta-amyloid precursor protein signaling domain by c-Jun N-terminal kinase JNK3 during neuronal differentiation. *J Neurosci* 2005; **25**: 5533-5543 [PMID: 15944381 DOI: 10.1523/JNEUROSCI.4883-04.2005]
- 52 **Martinez A**, Castro A, Dorronsoro I, Alonso M. Glycogen synthase kinase 3 (GSK-3) inhibitors as new promising drugs for diabetes, neurodegeneration, cancer, and inflammation. *Med Res Rev* 2002; **22**: 373-384 [PMID: 12111750 DOI: 10.1002/med.10011]
- 53 **Dorronsoro I**, Castro A, Martinez A. Inhibitors of glycogen synthase kinase-3: future therapy for unmet medical needs? *Expert Opin Ther Patents* 2002; **12**: 1527-1536 [DOI: 10.1517/13543776.12.10.1527]
- 54 **Khanfar MA**, Asal BA, Mudit M, Kaddoumi A, El Sayed KA. The marine natural-derived inhibitors of glycogen synthase kinase-3beta phenylmethylene hydantoins: In vitro and in vivo activities and pharmacophore modeling. *Bioorg Med Chem* 2009; **17**: 6032-6039 [PMID: 19616957 DOI: 10.1016/j.bmc.2009.06.054]
- 55 **Wong M**. Mammalian target of rapamycin (mTOR) pathways in neurological diseases. *Biomed J* 2013; **36**: 40-50 [PMID: 23644232 DOI: 10.4103/2319-4170.110365]
- 56 **Tsang CK**, Qi H, Liu LF, Zheng XF. Targeting mammalian target of rapamycin (mTOR) for health and diseases. *Drug Discov Today* 2007; **12**: 112-124 [PMID: 17275731 DOI: 10.1016/j.drudis.2006.12.008]
- 57 **Khanfar MA**, AbuKhader MM, Alqtaishat S, Taha MO. Pharmacophore modeling, homology modeling, and in silico screening reveal mammalian target of rapamycin inhibitory activities for sotalol, glyburide, metipranolol, sulfamethizole, glipizide, and pioglitazone. *J Mol Graph Model* 2013; **42**: 39-49 [PMID: 23545333 DOI: 10.1016/j.jmkgm.2013.02.009]
- 58 **Tanneeru K**, Guruprasad L. Ligand-based 3-D pharmacophore generation and molecular docking of mTOR kinase inhibitors. *J Mol Model* 2012; **18**: 1611-1624 [PMID: 21805127 DOI: 10.1007/s00894-011-1184-3]
- 59 **Sarma R**, Sinha S, Ravikumar M, Kishore Kumar M, Mahmood SK. Pharmacophore modeling of diverse classes of p38 MAP kinase inhibitors. *Eur J Med Chem* 2008; **43**: 2870-2876 [PMID: 18406015 DOI: 10.1016/j.ejmech.2008.02.014]
- 60 **Xiao Z**, Varma S, Xiao YD, Tropsha A. Modeling of p38 mitogen-activated protein kinase inhibitors using the Catalyst HypoGen and k-nearest neighbor QSAR methods. *J Mol Graph Model* 2004; **23**: 129-138 [PMID: 15363455 DOI: 10.1016/j.jmkgm.2004.05.001]
- 61 **Dillon RL**, Muller WJ. Distinct biological roles for the akt family in mammary tumor progression. *Cancer Res* 2010; **70**: 4260-4264 [PMID: 20424120 DOI: 10.1158/0008-5472]
- 62 **Manning BD**, Cantley LC. AKT/PKB signaling: navigating downstream. *Cell* 2007; **129**: 1261-1274 [PMID: 17604717 DOI: 10.1016/j.cell.2007.06.009]
- 63 **Vyas VK**, Ghatge M, Goel A. Pharmacophore modeling, virtual screening, docking and in silico ADMET analysis of protein kinase B (PKB β) inhibitors. *J Mol Graph Model* 2013; **42**: 17-25 [PMID: 23507201 DOI: 10.1016/j.jmkgm.2013.01.010]
- 64 **Fei J**, Zhou L, Liu T, Tang XY. Pharmacophore modeling, virtual screening, and molecular docking studies for discovery of novel Akt2 inhibitors. *Int J Med Sci* 2013; **10**: 265-275

[PMID: 23372433 DOI: 10.7150/ijms.5344]

65 **Traxler P**, Furet P. Strategies toward the design of novel and se-

lective protein tyrosine kinase inhibitors. *Pharmacol Ther* 1999; **82**:
195-206 [PMID: 10454197 DOI: 10.1016/s0163-7258(98)00044-8]

P- Reviewer: Choi CY, Huang Y, Moens U **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Lu YJ



Role of antipsychotics for treating behavioral and psychological symptoms of dementia

Kai Zhen Yap, Sui Yung Chan

Kai Zhen Yap, Sui Yung Chan, Department of Pharmacy, National University of Singapore, Singapore 117543, Singapore
Author contributions: Yap KZ and Chan SY contributed 70% and 30% respectively to this work, from the conception, review of evidence, draft and amendment of this review manuscript.
Correspondence to: Kai Zhen Yap, PhD, Instructor, Department of Pharmacy, National University of Singapore, 18 Science Drive 4, Singapore 117543, Singapore. phaykz@nus.edu.sg
Telephone: +65-66013479 Fax: +65-67791554
Received: July 29, 2014 Revised: October 2, 2014
Accepted: October 23, 2014
Published online: December 9, 2014

Abstract

Over the past three decades, concerns about the high prevalence of antipsychotic use in the nursing homes (NHs) for the management of behavioral and psychological symptoms of dementia continue to be emphasized and intervened by many. However, despite the numerous side effects and the recent blackbox warning by the United States Food and Drug Administration about the increased risks for stroke and sudden death associated with the use of antipsychotics in dementia, the prevalence of antipsychotic use in NHs remains high. While the use of antipsychotics appeared to have modest efficacy in reducing symptoms of aggression and psychosis in dementia, there is insufficient evidence to routinely recommend the use of alternative psychopharmacological treatments for these symptoms. Hence, clinicians have to balance the safety warnings against the need to treat these symptoms in order to prevent harm to the resident that may result from his/her dangerous behaviors. Although the use of antipsychotics may be warranted in some cases, organizational, resource and training support should be provided to encourage and equip NH staff to participate in interventions so as to minimize inappropriate use of these medicines in NHs. This review will discuss the place in therapy, the trend and appropriateness of antipsychotic use in NHs, as well as the effectiveness

of current and future strategies for reducing antipsychotic use in the NHs.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Antipsychotic agents; Psychotropic drugs; Prescribing appropriateness; Dementia; Behavioral and psychological symptoms of dementia; Nursing homes

Core tip: While antipsychotics may be used to manage symptoms of severe aggression and psychosis when the safety of the resident is threatened, there should be routine reviews of the appropriateness of antipsychotic use as well as training and support of the care staff in providing psychosocial intervention to treat the symptoms so as to reduce antipsychotic use in nursing homes. Reported studies evaluating interventions to improve antipsychotic use appropriateness in nursing homes are limited by the small sample sizes and absence of control groups. Future research should address these methodological issues while exploring safer therapeutic alternatives to manage these symptoms.

Yap KZ, Chan SY. Role of antipsychotics for treating behavioral and psychological symptoms of dementia. *World J Pharmacol* 2014; 3(4): 174-185 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/174.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.174>

OVERVIEW OF BEHAVIORAL AND PSYCHOLOGICAL SYMPTOMS OF DEMENTIA

With the rapidly aging population worldwide, the number of persons with dementia is projected to double every 20 years, from 35.6 million in year 2010, to 65.7 million, and 115.4 million by years 2030 and 2050 respectively^[1]. Dementia is marked by features of progressively worsening

memory impairment and cognitive disturbances^[2]. As the illness advances, the resulting decline in functional capacity naturally exerts its toll on the patient's family and/or the society, demanding significant expenditure in time, energy and resources in caregiving for extended periods. In year 2010, the cost of informal care (unpaid care provided by family and others) and direct cost of social care (provided by care professionals in the community and in residential long-term care institutions) for dementia each contributed to about 42% of the estimated USD 604 billion total cost^[3].

In addition to delaying cognitive and functional decline, dementia-related research has become increasingly focused on defining, measuring and managing behavioral and psychological symptoms of dementia (BPSD)^[4]. BPSD is a term that encompasses a heterogeneous range of non-cognitive symptoms, such as disturbed perception, thought content, mood, and behavior^[4]; and are broadly classified as "behavioral" or "psychological"^[5]. These symptoms may be present in up to 97% of persons with dementia over a five-year period^[6], and were reported to be a significant source of patient distress, caregiver stress^[7,8], increased costs of care and nursing home (NH) admissions^[9]. Hence, it was not surprising that higher point prevalence of BPSD were reported in the NHs compared to that in the community care setting^[10].

EFFICACY AND SAFETY OF ANTIPSYCHOTICS FOR TREATING BPSD

Over many decades, antipsychotics have been prescribed for managing BPSD, particularly symptoms of severe agitation, aggression and psychosis. Although the precise mechanism of antipsychotics' influence on agitation is not known, their antagonistic effect on postsynaptic dopamine receptors was postulated to play a role in the amelioration of psychotic symptoms in dementia^[11]. In a meta-analysis, haloperidol appeared to be clinically effective in reducing symptoms of aggression in dementia compared to controls^[12]. The lack of a significant difference in the overall drop-out rates between the haloperidol treatment and control groups despite more drop-outs from the haloperidol group due to the presence of side effects further suggests the possible effectiveness of the drug^[12]. However, the authors concluded that the routine use of haloperidol for the treatment of agitation in dementia should not be recommended, due to insufficient evidence of benefits from this treatment^[12].

Among the atypical antipsychotics, risperidone and olanzapine were deemed by Sink and his team to have the best evidence for efficacy in the treatment of aggression and psychosis^[13]. Specifically, a meta-analysis reported that risperidone (1 to 2 mg/d) was effective in the treatment of aggression and psychosis related to dementia, while the use of olanzapine (5 to 10 mg/d) showed significant benefits in reducing aggression when compared to placebo^[14]. These findings were consistent with that

reported in another meta-analysis^[15] and the CATIE-AD trial^[16,17]. In another randomized, placebo-controlled study, the use of aripiprazole resulted in a reduction of psychotic symptoms^[18]. Although quetiapine was observed by Tariot and his co-investigators to improve symptoms of agitation^[19], this finding was not replicated in other studies^[16,17,20].

While typical antipsychotics are primarily D₂ receptor antagonists and inhibit dopaminergic neurotransmission in a dose-dependent manner, atypical antipsychotics vary in their binding affinities to other receptors^[21]. As a result, each atypical antipsychotic has a different side effect profile. Unlike haloperidol, atypical antipsychotics generally have lesser propensities to cause neuroleptic-induced movement disorders or extrapyramidal symptoms (EPS)^[22] due to a lower D₂ receptor blocking effect or a partial D₂ agonistic effect^[23]. However, EPS may become more apparent with risperidone at doses higher than 2 mg/d due to its dose-dependent dopamine receptor blocking effect^[14]. Other side effects and adverse events of risperidone also included somnolence, peripheral edema, cerebrovascular adverse events, urinary incontinence, urinary tract infection and falls^[14]. For olanzapine and quetiapine, somnolence side effect was more prominent compared to other antipsychotics due to their higher affinity to block H₁ receptors^[24]. In addition, significant weight gains, increased waist circumferences and decreased high-density lipoprotein cholesterol levels were also reported as characteristic side effects of olanzapine and quetiapine^[14,25]. A decline in cognitive function could be a side effect of antipsychotic use, but this was found to be more significant among individuals who were treated with olanzapine and risperidone^[26], possibly due to the more pronounced anticholinergic effects of these antipsychotics^[21].

Besides the above-mentioned side effects, several studies and reviews have also highlighted the safety concerns regarding the use of antipsychotics in dementia. In a systematic review by Trifiró *et al.*^[27], an increase in risk for mortality was reported to be associated with the use of both typical and atypical antipsychotics in a dose-dependent fashion, where the highest risk was estimated to be shortly after exposure^[27]. Although the related causes were postulated to include cerebrovascular event, pneumonia, peripheral vascular effects and/or metabolic effects, the differential risks of the individual antipsychotics and predisposing patient factors have yet to be established^[27].

Another safety concern was the increased risk for cerebrovascular events (CVEs) associated with antipsychotics, particularly olanzapine and risperidone, which were linked to a threefold increase in risk^[14,28]. Yet, based on currently reported studies, a fair conclusion on the difference in risks between individual antipsychotic agents cannot be drawn. The plausible mechanisms of antipsychotic-related CVEs were deemed to be linked to side effects of antipsychotics including EPS^[29], orthostatic hypotension, hyperprolactinemia^[30], thromboem-

bolic events and excessive sedation^[29,31]. However, while the elevated risk was thought to be temporal with the potential to decrease over time^[32], the contribution of predisposing patient factors to the development of CVEs (such as presence of vascular dementia) independent of antipsychotic use was not ascertained^[27].

Antipsychotic use was also associated with an increased risk for pneumonia. Specifically, threefold and 1.6-fold increases in risk were observed when atypical and typical antipsychotics were used respectively^[33]. These could possibly be due to aspiration pneumonia linked to underlying mechanisms of antipsychotic-induced side effects including dysphagia, sedation and EPS^[34]. Due to the poor prognosis of pneumonia in older persons with dementia, this adverse event may in turn contribute to the increased risk of their mortality associated with the use of these medicines.

ROLE OF ALTERNATIVE PSYCHOPHARMACOLOGICAL AGENTS

Benzodiazepines

Compared to antipsychotics, there was no or little evidence for recommending the use of other psychopharmacologicals such as benzodiazepines and anticonvulsant mood stabilizers to treat symptoms of severe agitation, aggression and psychosis. Specifically, the use of benzodiazepines among older persons for the treatment of agitation, especially in the presence of dementia, should be avoided as these individuals are more sensitive to side effects including sedation, ataxia and withdrawal symptoms, which may potentiate confusion, falls and fractures leading to adverse clinical outcomes^[35]. To the authors' knowledge, there are also no published systematic review, meta-analysis or randomized controlled study to provide any evidence to support the treatment of severe agitation, aggression and psychosis in dementia with benzodiazepines.

Anticonvulsant mood stabilizers

With regards to anticonvulsant mood stabilizers, valproate was ineffective in reducing BPSD or agitation symptoms according to two recent reviews^[13,36] and a meta-analysis^[37]. Although carbamazepine appeared to have some effect in reducing symptoms of aggression^[38,39], these reports were countered by negative findings of another study^[40]. Furthermore, carbamazepine has clinically significant drug-drug interactions with medicines commonly used by older persons such as verapamil^[41,42] and warfarin^[43]. Carbamazepine also carries black box warnings for potentially fatal severe adverse drug reactions, specifically hematologic toxicity and serious dermatologic reactions especially for individuals with HLA-B*1502 allele^[44,45]. As such, the clinical use of carbamazepine, especially in the NHs, would be highly inconvenient due to the need for pre-treatment genotype screening and regular hematological monitoring to minimize the occurrence of these serious adverse drug reactions. Therefore,

anticonvulsant mood stabilizers should not be used to treat aggression and psychosis related to dementia.

Antidepressants

While serotonin has been postulated to be involved in the underlying pathophysiological mechanisms for psychosis and aggression^[46,47], the evidence for the clinical use of antidepressants is primarily for the treatment of depression in dementia^[48]. Although Lyketsos and his colleagues observed a beneficial reduction in non-mood behavioral symptom scores of the Neuropsychiatric Inventory (NPI-NM)^[49,50] among individuals with Alzheimer's disease who had responded fully to the treatment of depression with sertraline, the difference in the reported NPI-NM scores between the treatment and control groups of the study was not statistically significant^[51]. In a recent meta-analysis^[52], the use of serotonin reuptake inhibitors (SSRIs) sertraline and citalopram were associated with a larger mean change in the Cohen Mansfield Agitation Inventory^[53] score (compared to placebo: -0.89, 95%CI: -1.22 to -0.57) and appeared to be better tolerated than typical and atypical antipsychotics. However, these findings were limited by the small sample sizes^[52]. Furthermore, an evaluation of the effectiveness of citalopram for the treatment of BPSD noted improvements that were limited to symptoms of agitation and lability. The results were also potentially biased with a high dropout rate of more than 50% due to possible side effects and lack of efficacy^[54]. Hence, more large-scale studies would be required to ascertain the safety and efficacy of SSRIs in the treatment of aggression and psychotic symptoms in dementia.

Acetylcholinesterase inhibitors and memantine

Besides improving cognitive symptoms, the effects of acetylcholinesterase inhibitors (donepezil, rivastigmine and galantamine) and an N-methyl D-aspartate antagonist (memantine) in reducing BPSD were described in many case reports^[55-57], clinical studies^[58-64], randomized controlled trials^[65-74] and systematic reviews^[63,75-77]. A review on the randomized controlled trials concluded that the findings for donepezil and memantine appeared to be conflicting^[13]. In addition, there is no landmark head-to-head study to offer a fair comparison for the differences in efficacies of these pharmacological agents. Meta-analyses of these drugs were also limited by different methodologies and measures of BPSD used in the clinical trials of each drug^[78]. Furthermore, there were also reports of paradoxical worsening of both behavioral symptoms related to the use of donepezil in frontotemporal dementia^[79] and parkinsonism associated with the use of donepezil in dementia with Lewy bodies^[57]. Significant side effects of rivastigmine were also observed, which included nausea, vomiting, tremor and dizziness^[66]. In all, it appears however, that rivastigmine^[64,66,77] and memantine^[71] may be safer alternatives for the management of aggression and psychotic symptoms, particularly for individuals with Parkinson's disease dementia and dementia with Lewy bodies, as they are likely to be susceptible to

the severe adverse effects of antipsychotics such as worsening of Parkinsonian symptoms and life-threatening severe neuroleptic malignant syndrome^[80,81].

GUIDELINES AND TRENDS OF ANTIPSYCHOTIC USE IN DEMENTIA

Although there is limited evidence supporting the efficacy of non-pharmacological interventions for reducing aggression and psychosis related to dementia, these are recommended as the first-line strategy over the use of antipsychotics in all practice guidelines^[82-84]. The obvious reasons are the numerous side effects^[12,25,26,85] and higher risks for stroke and death associated with antipsychotics, which out-weigh their modest efficacies^[86-89], and their limited benefits with long-term use^[90]. Despite the introduction of the black box warning for antipsychotics by the United States Food and Drug Administration in 2005 against its use in view of the increased risks for stroke and death, the reported prevalence of antipsychotic use in most NHs in the United States remained unchanged^[91]. A recent report by the Centers for Medicare and Medicaid Services estimated that about 40% of NH residents with dementia were prescribed with antipsychotics in 2010^[92]. Interestingly, this corresponded with the prevalence of delusion (54%) and hallucination (39%) found among these residents with dementia^[93]. In a cross-national comparison, while about a quarter of NH residents in the United States were prescribed with antipsychotics, this prevalence varied between 11%-40% in Hong Kong, Canada, Switzerland and Finland and other countries^[94].

Within NHs, comparisons between the older persons with dementia residing in special dementia units *vs* traditional care wards found that antipsychotics were used more often in the former, as these residents were more likely to exhibit behavioral problems^[95,96]. However, no statistical difference in the prevalence of antipsychotic use between these two cohorts of elderly NH residents with dementia was reported in another study, where the researchers attributed it to the effect of increased number of activities and psychosocial interventions which reduced the need for antipsychotics^[97].

Nevertheless, the use of antipsychotics in the NHs will continue due to the lack of alternative evidence-based pharmacological treatments for dementia-related severe agitation, aggression and psychosis for the residents with risk of physical harm as a result of uncontrolled behavior^[98,99]. In addition, although many guidelines advocate prescribing antipsychotics for a minimal duration with attempts to taper off and discontinue at least once every 6 mo, a recent study suggested that the use of antipsychotics for up to 9 mo in individuals with severe baseline symptoms may confer benefits of having a reduction in symptom relapses compared to those who were taken off antipsychotics after 4 mo^[100]. Similarly, another review concluded that though antipsychotics can be withdrawn within 6 mo without detrimental effects on behavior for most individuals, the use of antipsychotics

could be extended for those with more severe symptoms at baseline to prevent relapses^[90]. Yet, concerns regarding the inappropriate use of antipsychotics in NHs were raised, which included the lack of proper documentation (especially pertaining to indications for use)^[92,101,102], prescribing of inappropriately high doses^[103] and inadequate monitoring^[104] for managing adverse effects and evaluating the need for continued use.

APPROPRIATENESS OF ANTIPSYCHOTIC PRESCRIBING

While there appears to be a lack of specific clinical guidance for antipsychotic prescribing in dementia^[105], the literature is replete with criteria for defining what are considered as “inappropriate”. Firstly, clinicians have to balance the safety warnings associated with antipsychotic use in dementia against the need to alleviate the caregiving stress of providing the basic needs of the aggressive patient and to protect the resident from his/her own dangerous behaviors^[99,106,107]. A failure to address these needs is considered “inappropriate”^[99,108]. On the other hand, the patient’s and/or family’s wishes to refrain from antipsychotic use have to be considered and respected^[108].

Secondly, antipsychotic prescribing decisions without documented reasons are considered as inappropriate. As suggested in the algorithm by Osborne *et al.*^[102], proper documentation of prescribing rationale when initiating antipsychotics would include the specific description of the target behavior and/or symptoms and its impact on the patient that the prescribed medicine was intended to treat and resolve. During subsequent medical follow-ups and reviews, documentation of the patients’ responses to the prescribed treatment in terms of the changes in the details and impact of the target indications would be required to make informed decision for attempting a dose reduction or continuing with the antipsychotic treatment at the minimum effective dose, according to the guiding principle of “start low and go slow”^[109]. Since the recommended antipsychotic doses for managing aggression and psychosis in dementia is generally much lower than that for treatment of psychiatric conditions, the prescribing of high doses and/or quick upward titration of doses may inappropriately expose the older person with dementia to unnecessary side effects such as drug-induced movement disorders, gait disturbances, and somnolence, potentially contributing to falls and adverse events such as fractures^[12,85,110].

Lastly, prescribing of antipsychotics for use in older persons with dementia would be inappropriate without proper monitoring for clinical responses and side effects. As prescribing decisions made during the short consultation time often depend on feedback from the caregivers, detailed and specific accounts of the changes in behavior, symptoms and complaints of patients may prompt timely interventions such as titrating the antipsychotic dose downwards for abating side effects. Specifically in the NH setting, the lack of proper monitoring and feedback

processes may be attributed to the low staff-to-resident ratio^[111], nurse-resident miscommunication^[112], inadequate training and the lack of a structured monitoring framework for antipsychotic use^[113], resulting in a lack of proper documentation of the rationale for antipsychotic use for each resident throughout his stay at the NH^[104].

INTERVENTIONS TO REDUCE INAPPROPRIATE PRESCRIBING OF ANTIPSYCHOTICS IN NURSING HOMES

In order to reduce inappropriate prescribing of antipsychotics in the NHs, many interventions have been detailed. The first widespread changes in antipsychotic use trends were reported across most NHs in the US during the early 1990s. This was in response to the implementation of the OBRA'87 legislation which restricted the unjustified use of antipsychotics as a chemical restraint in the NHs for the management of difficult behaviors such as wandering, restlessness, anxiety and uncooperativeness^[114]. In tandem with this legislation was the mandatory conduct of routine drug regimen reviews by pharmacists^[115]. Although these brought about remarkable reductions in antipsychotic use, evidence of its positive impact on other clinical outcomes such as reduction in adverse events among NH residents was elusive. In contrast, a retrospective cross-sectional study noted that the NH residents in the United States were more likely to sustain falls, despite lower prevalence of psychotropic use, compared to those in Denmark, Iceland, Italy, Japan and Sweden^[116]. Furthermore, adequate level of staffing could be more crucial towards the successful reduction of antipsychotic use and improved outcomes in the NHs^[117,118].

A literature search using a combination of terms “antipsychotics”, “neuroleptics”, “prescribing”, “nursing homes” and “intervention” was conducted to identify original studies that reported interventions targeting to reduce inappropriate antipsychotic use in NHs. A total of 12 interventions involving strategies such as audit-feedback processes, education and training for prescribers and/or nurses, medication review, multi-disciplinary case conferencing, early screening and intervention, structured monitoring program, as well as patient-centered psychosocial interventions are discussed in this review (Table 1).

Interventions involving audits and feedback

Among the interventions using the audits and feedback approach, Westbury's and Castle's reports showed statistically significant reduction in the use of antipsychotics^[119,120]. However, Westbury's group postulated that the positive outcome was likely to be attributed to the impact of the academic detailing with physicians, nursing staff training and follow-up medication review component^[119], but the outcome was not sustainable after 18 mo and the antipsychotic use prevalence returned to baseline levels^[121]. This suggests for the management of BPSD

to be a long-term process, requiring constant reviews to ensure the appropriateness of antipsychotic prescribing. Similarly, another study showed that the audit-feedback process resulted in a reduction in antipsychotic use when it was carried out in combination with providing education and practical tools for nurses on how to document behaviors as well as the use of non-pharmacological interventions by nurses to manage agitation and challenging behaviors^[122]. However, the intervention employed by Castle^[120] did not include the component of education. Instead, it focused on communicating “important legislative efforts to reduce the prevalence of antipsychotics” as well as each NH's “performance” compared to other NHs across the board, in a manner which facilitated the use of these information by the NHs to change their care processes^[120]. However, as inadequate information was provided on the selection or randomization of the NHs included in Castle's study, there could be a potential bias of more NHs that are motivated in change in the intervention group.

Interventions involving education of healthcare professionals

Unlike educational interventions previously reported in the early 1990s^[123,124], two studies published more recently in 2005 and 2010 did not report significant reduction of antipsychotic use in the intervention NHs^[125,126]. Reasons for these could be related to the small sample size of residents using antipsychotics at baseline as well as the use of non comparable control NHs with regards to the BPSD severity and use of antipsychotics of the NH residents. However, Monette *et al.*^[127] reported a large number of antipsychotic discontinuation and dose reduction following an inter-disciplinary educational intervention, which was coupled with active monthly clinical follow-up by pharmacists to remind physicians to review antipsychotic prescriptions, monthly charting of residents' BPSD severity by nurses, as well as regular inter-disciplinary team meetings. A re-evaluation of this complex intervention involving education and inter-disciplinary efforts five years' later continued to demonstrate a reduction in the prevalence of antipsychotic use from 30.5% in year 2004 to 17.2% in year 2009^[128].

Interventions involving medication review

In the United States, pharmacist-conducted medication reviews yielded a positive impact on improving the appropriateness of antipsychotic use in nursing homes^[116]. This effect was also observed in Northern Ireland where a significant reduction in antipsychotic use was observed among NH residents receiving a structured pharmacist-led medication review program compared to those receiving usual care with no pharmacist intervention^[129]. Despite the involvement of resident interviews and multidisciplinary meetings with nurses and physicians, this intervention was estimated to be more cost-effective than usual care^[130]. Positive reduction in antipsychotic use was also reported in a medication review intervention led by

Table 1 Selected original intervention studies aimed to improve antipsychotic use in the nursing home

Intervention type	Study design	Study duration	Healthcare disciplines providing intervention	Changes in antipsychotic use
Audit-feedback				
Castle ^[120]	CT	1 yr	NA	4.8% reduction in intervention group; SS
Westbury <i>et al</i> ^[119]	CT	26 wk	P, Ph, N	1.7% reduction in intervention group; SS
Watson-Wolfe <i>et al</i> ^[122]	SSBAS	2 mo	N	4.9% reduction
Education				
Hagen <i>et al</i> ^[126]	CT	1 yr	Ph	Increases in antipsychotic use; no SS in intervention group but SS in control group
Testad <i>et al</i> ^[125]	CRCT	1 yr	P	Increases in antipsychotic use; no SS
Monette <i>et al</i> ^[127]	SSBAS	7 mo	P, Psy, Ph, N	49% discontinued antipsychotics, 13.6% had dose reduction
Medication review				
Patterson <i>et al</i> ^[129]	CRCT	1 yr	P, Ph, N	9.4% reduction in intervention group; odds ratio of antipsychotic use for intervention group <i>vs</i> control = 0.26 (95%CI: 0.14–0.49); SS
Chakraborty <i>et al</i> ^[131]	MSBAS	2 yr	Psy, N	13.4% reduction
Case conferencing				
Dahl <i>et al</i> ^[133]	SSBAS	1 yr	P, Ph, N	1.3% reduction
Structured monitoring				
Yap <i>et al</i> ^[104]	SSBAS	24 wk	P, Ph, N	4 times increase in antipsychotic prescribing decisions due to side-effects reported; SS
Psychosocial intervention				
Fossey <i>et al</i> ^[134]	CRCT	10 mo	Psychologist, occupational therapist, N	19.1% reduction in intervention group; lower prevalence in intervention group (19.1% <i>vs</i> 42.1%); SS
Bird <i>et al</i> ^[135]	CT	9 mo	P, Psy, N, Psychologist	15.7% reduction in intervention group; SS

CRCT: Cluster-randomized controlled trial; CT: Controlled trial (non-randomized); MSBAS: Multi-site, before-and-after study; N: Nurse; NA: Not assessed; P: Physician; Ph: Pharmacist; Psy: Psychiatrist; SSBAS: Single site, before-and-after study; SS: Statistically significant.

psychiatrists and nurses^[131]. However, the study did not include a control group or cost-effective analysis. The frequency and duration of visits by the psychiatrist-nurse team was also not known. Interestingly, the NHs in this study had an overall higher prevalence of antipsychotic use compared to non-nursing residential homes. This could be attributed to NH residents having more severe BPSD, which supports the continuous need to identify safer and more effective approaches to manage BPSD in NHs.

Interventions involving multi-disciplinary case conferencing

A regular multidisciplinary team intervention study reported a significant decrease in the prevalence (-19%) of antipsychotic use^[132]. However, at the end of the study, the prevalence of use remained at 19% after the intervention study period, while only 5% of the study population had psychotic disorders. Additional tools for facilitating the assessment, documentation of symptoms and reporting during multi-disciplinary meetings were described in other published studies^[104,133]. In order to circumvent the challenge of coordinating the schedules of visiting physicians, psychiatrists and pharmacists for face-to-face case conferencing at the NHs, the intervention reported by Yap's group^[104] emphasized on the process of monitoring, documentation and feedback of changes in residents' behavior, clinical responses and side effects to the prescribed antipsychotics by the nursing staff, including nursing aides and healthcare attendants. These

caregivers were motivated in providing the intervention as the monitoring-feedback processes were readily incorporated into their usual duties and did not require them to perform additional interviews or physical assessments on the residents. This resulted in a significant increase in prescribing decisions, specifically dose reduction and switching of agents to one with less propensity for drug-induced movement disorders, in response to side effects of antipsychotics reported by the nursing staff. Hence, this intervention would be useful in settings with low staff-to-resident ratios and where potential language and/or cultural barriers between the care staff and residents are present.

Interventions involving psychosocial intervention

The use of psychosocial care was found to be an effective alternative to the use of antipsychotics in managing BPSD according to two studies. Specifically, the use of person-centred care approach for managing BPSD demonstrated significant reduction in antipsychotic use in NHs^[134,135]. However, it may be challenging for NHs with staffing caps to implement full psychosocial care in managing BPSD as it is labor-intensive. The provision of continuous support from prescribers and NH administrators as well as training of staff^[134] and adequate staff-to-resident ratio is needed^[135].

Overall, the majority of the published interventions with positive outcomes of reducing inappropriate antipsychotic use involved education for clinicians and care staff. While nurses were involved in all the interventions

with positive changes in antipsychotic use, they were not part of those interventions which reported no significant reduction in antipsychotic use. This observation suggests that interventions should involve healthcare providers from more than one discipline, especially the nursing staff, as their input as direct caregivers would be a significant influence on antipsychotic prescribing in the NHs^[136]. Furthermore, it was noted that many interventions focused on reducing the use of antipsychotics, which is synonymous with preventing the “overuse” and “mis-use” of antipsychotics, while only the intervention reported by Yap’s group^[104] expressly addressed the potential “underuse” of antipsychotics due to under- or mis-identification of symptoms such as psychosis, which could respond to short-term antipsychotic treatment^[83].

Most of the intervention studies cited in this review employed a variety of intervention types and methodological designs, and some are limited by small sample sizes and the lack of a suitable control. As only one study evaluated the cost-effectiveness of the pharmacist-led medication review intervention^[130], the comparisons of effectiveness in the other studies were descriptive at best. However, a study making direct comparison of the 4 interventions, namely medication review, recreational therapy, exercise and patient-centered care is ongoing^[137]. Its results would provide deeper insights on the effectiveness of these interventions. Although some of the intervention studies^[125,127,134] included in this review measured the change in BPSD using various instruments, positive results for this outcome measure can not be entirely attributed to the appropriateness of antipsychotic use as BPSD, specifically agitation, is intermittent in nature^[138]. Hence, future studies should address the highlighted methodological concerns and measure the long-term effects of reducing antipsychotic use on BPSD and adverse outcomes among NH residents^[139].

CONCLUSION

It appears that despite the modest efficacy and concerns for adverse outcomes of antipsychotic use in the management of BPSD, the use of these medications in the NHs is inevitable. However, future research should continue to explore the use of safer alternatives for the treatment of these symptoms. Although current guidelines recommend the use of psychosocial care over antipsychotics for the management of BPSD, organizational, resource and training support are essential to encourage and equip the NH staff to participate and provide these interventions. At present, there is no alternative solution to antipsychotic treatment and no gold standard in clinical practice to reduce inappropriate antipsychotic use. While the use of antipsychotics to manage BPSD symptoms may be warranted in cases when the safety of the NH resident and others around him/her is threatened, multidisciplinary interventions such as routine medication reviews to promote the appropriate use of antipsychotics may contribute as a long-term sustainable solution.

REFERENCES

- 1 **Prince M**, Bryce R, Albanese E, Wimo A, Ribeiro W, Ferri CP. The global prevalence of dementia: a systematic review and metaanalysis. *Alzheimers Dement* 2013; **9**: 63-75.e2 [PMID: 23305823 DOI: 10.1016/j.jalz.2012.11.007]
- 2 **Work group on Alzheimer's disease and other dementias**. Treatment of patients with Alzheimer's disease and other dementias. 2nd ed. [accessed 2014 September 29]. Available from: URL: <http://psychiatryonline.org/content.aspx?bookid=28&ionid=1679489-152238>
- 3 **World Alzheimer Report**. The global economic impact of dementia. 2010. [Accessed 2014 September 23]. Available from: URL: <http://www.alz.co.uk/research/files/WorldAlzheimerReport2010.pdf>
- 4 **Finkel SI**, Costa e Silva J, Cohen G, Miller S, Sartorius N. Behavioral and psychological signs and symptoms of dementia: a consensus statement on current knowledge and implications for research and treatment. *Int Psychogeriatr* 1996; **8** Suppl 3: 497-500 [PMID: 9154615 DOI: 10.1017/S1041610297003943]
- 5 **Finkel S**. Introduction to behavioural and psychological symptoms of dementia (BPSD). *Int J Geriatr Psychiatry* 2000; **15** Suppl 1: S2-S4 [PMID: 10767742]
- 6 **Steinberg M**, Shao H, Zandi P, Lyketsos CG, Welsh-Bohmer KA, Norton MC, Breitner JC, Steffens DC, Tschanz JT. Point and 5-year period prevalence of neuropsychiatric symptoms in dementia: the Cache County Study. *Int J Geriatr Psychiatry* 2008; **23**: 170-177 [PMID: 17607801 DOI: 10.1002/gps.1858]
- 7 **Heok KE**, Li TS. Stress of caregivers of dementia patients in the Singapore Chinese family. *Int J Geriatr Psychiatry* 1997; **12**: 466-469 [PMID: 9178051 DOI: 10.1002/(SICI)1099-1166(199704)12:4<466::AID-GPS517>3.3.CO;2-L]
- 8 **Tan LL**, Wong HB, Allen H. The impact of neuropsychiatric symptoms of dementia on distress in family and professional caregivers in Singapore. *Int Psychogeriatr* 2005; **17**: 253-263 [PMID: 16050434 DOI: 10.1017/S1041610205001523]
- 9 **O'Donnell BF**, Drachman DA, Barnes HJ, Peterson KE, Swearer JM, Lew RA. Incontinence and troublesome behaviors predict institutionalization in dementia. *J Geriatr Psychiatry Neurol* 1992; **5**: 45-52 [PMID: 1571074 DOI: 10.1177/002383099200500108]
- 10 **Margallo-Lana M**, Swann A, O'Brien J, Fairbairn A, Reichelt K, Potkins D, Mynt P, Ballard C. Prevalence and pharmacological management of behavioural and psychological symptoms amongst dementia sufferers living in care environments. *Int J Geriatr Psychiatry* 2001; **16**: 39-44 [PMID: 11180484 DOI: 10.1002/1099-1166(200101)16:1<39::AID-GPS269>3.0.CO;2-F]
- 11 **White KE**, Cummings JL. Schizophrenia and Alzheimer's disease: clinical and pathophysiologic analogies. *Compr Psychiatry* 1996; **37**: 188-195 [PMID: 8732586 DOI: 10.1016/S0010-440X(96)90035-8]
- 12 **Lonergan E**, Luxenberg J, Colford J. Haloperidol for agitation in dementia. *Cochrane Database Syst Rev* 2001; **2**: CD002852 [PMID: 11687166 DOI: 10.1002/14651858.CD002852]
- 13 **Sink KM**, Holden KF, Yaffe K. Pharmacological treatment of neuropsychiatric symptoms of dementia: a review of the evidence. *JAMA* 2005; **293**: 596-608 [PMID: 15687315 DOI: 10.1001/jama.293.5.596]
- 14 **Ballard C**, Waite J. The effectiveness of atypical antipsychotics for the treatment of aggression and psychosis in Alzheimer's disease. *Cochrane Database Syst Rev* 2006; **1**: CD003476 [PMID: 16437455 DOI: 10.1002/14651858.CD003476.pub2]
- 15 **Maher AR**, Maglione M, Bagley S, Suttrop M, Hu JH, Ewing B, Wang Z, Timmer M, Sultzer D, Shekelle PG. Efficacy and comparative effectiveness of atypical antipsychotic medications for off-label uses in adults: a systematic review and meta-analysis. *JAMA* 2011; **306**: 1359-1369 [PMID: 21954480 DOI: 10.1001/jama.2011.1360]

- 16 **Schneider LS**, Tariot PN, Dagerman KS, Davis SM, Hsiao JK, Ismail MS, Lebowitz BD, Lyketsos CG, Ryan JM, Stroup TS, Sultzer DL, Weintraub D, Lieberman JA. Effectiveness of atypical antipsychotic drugs in patients with Alzheimer's disease. *N Engl J Med* 2006; **355**: 1525-1538 [PMID: 17035647 DOI: 10.1056/NEJMoa061240]
- 17 **Sultzer DL**, Davis SM, Tariot PN, Dagerman KS, Lebowitz BD, Lyketsos CG, Rosenheck RA, Hsiao JK, Lieberman JA, Schneider LS. Clinical symptom responses to atypical antipsychotic medications in Alzheimer's disease: phase 1 outcomes from the CATIE-AD effectiveness trial. *Am J Psychiatry* 2008; **165**: 844-854 [PMID: 18519523 DOI: 10.1176/appi.ajp.2008.07111779]
- 18 **De Deyn P**, Jeste DV, Swanink R, Kostic D, Breder C, Carson WH, Iwamoto T. Aripiprazole for the treatment of psychosis in patients with Alzheimer's disease: a randomized, placebo-controlled study. *J Clin Psychopharmacol* 2005; **25**: 463-467 [PMID: 16160622 DOI: 10.1097/01.jcp.0000178415.22309.8f]
- 19 **Tariot PN**, Schneider L, Katz IR, Mintzer JE, Street J, Copenhaver M, Williams-Hughes C. Quetiapine treatment of psychosis associated with dementia: a double-blind, randomized, placebo-controlled clinical trial. *Am J Geriatr Psychiatry* 2006; **14**: 767-776 [PMID: 16905684 DOI: 10.1097/01.JGP.0000196628.12010.35]
- 20 **Ballard C**, Margallo-Lana M, Juszcak E, Douglas S, Swann A, Thomas A, O'Brien J, Everatt A, Sadler S, Maddison C, Lee L, Bannister C, Elvish R, Jacoby R. Quetiapine and rivastigmine and cognitive decline in Alzheimer's disease: randomised double blind placebo controlled trial. *BMJ* 2005; **330**: 874 [PMID: 15722369 DOI: 10.1136/bmj.38369.459988.8f]
- 21 **Gareri P**, Segura-García C, Manfredi VG, Bruni A, Ciambrone P, Cerminara G, De Sarro G, De Fazio P. Use of atypical antipsychotics in the elderly: a clinical review. *Clin Interv Aging* 2014; **9**: 1363-1373 [PMID: 25170260 DOI: 10.2147/CIA.S63942]
- 22 **Wirshing WC**. Movement disorders associated with neuroleptic treatment. *J Clin Psychiatry* 2001; **62** Suppl 21: 15-18 [PMID: 11584982]
- 23 **Divac N**, Prostran M, Jakovcevski I, Cerovac N. Second-generation antipsychotics and extrapyramidal adverse effects. *Biomed Res Int* 2014; **2014**: 656370 [PMID: 24995318 DOI: 10.1155/2014/656370]
- 24 **Street JS**, Clark WS, Gannon KS, Cummings JL, Bymaster FP, Tamura RN, Mitan SJ, Kadam DL, Sanger TM, Feldman PD, Tollefson GD, Breier A. Olanzapine treatment of psychotic and behavioral symptoms in patients with Alzheimer disease in nursing care facilities: a double-blind, randomized, placebo-controlled trial. The HGEU Study Group. *Arch Gen Psychiatry* 2000; **57**: 968-976 [PMID: 11015815 DOI: 10.1001/archpsyc.57.10.968]
- 25 **Zheng L**, Mack WJ, Dagerman KS, Hsiao JK, Lebowitz BD, Lyketsos CG, Stroup TS, Sultzer DL, Tariot PN, Vigen C, Schneider LS. Metabolic changes associated with second-generation antipsychotic use in Alzheimer's disease patients: the CATIE-AD study. *Am J Psychiatry* 2009; **166**: 583-590 [PMID: 19369318]
- 26 **Vigen CL**, Mack WJ, Keefe RS, Sano M, Sultzer DL, Stroup TS, Dagerman KS, Hsiao JK, Lebowitz BD, Lyketsos CG, Tariot PN, Zheng L, Schneider LS. Cognitive effects of atypical antipsychotic medications in patients with Alzheimer's disease: outcomes from CATIE-AD. *Am J Psychiatry* 2011; **168**: 831-839 [PMID: 21572163 DOI: 10.1176/appi.ajp.2011.08121844]
- 27 **Trifirò G**, Sultana J, Spina E. Are the safety profiles of antipsychotic drugs used in dementia the same? An updated review of observational studies. *Drug Saf* 2014; **37**: 501-520 [PMID: 24859163 DOI: 10.1007/s40264-014-0170-y]
- 28 **Wooltorton E**. Olanzapine (Zyprexa): increased incidence of cerebrovascular events in dementia trials. *CMAJ* 2004; **170**: 1395 [PMID: 15111472 DOI: 10.1503/cmaj.1040539]
- 29 **Herrmann N**, Lanctôt KL. Do atypical antipsychotics cause stroke? *CNS Drugs* 2005; **19**: 91-103 [PMID: 15697324 DOI: 10.2165/00023210-200519020-00001]
- 30 **Wallaschofski H**, Lohmann T, Hild E, Kobsar A, Siegemund A, Spilcke-Liss E, Hentschel B, Stumpf C, Daniel WG, Garlich CD, Eigenthaler M. Enhanced platelet activation by prolactin in patients with ischemic stroke. *Thromb Haemost* 2006; **96**: 38-44 [PMID: 16807649 DOI: 10.1160/TH05-09-0634]
- 31 **Smith DA**, Beier MT. Association between risperidone treatment and cerebrovascular adverse events: examining the evidence and postulating hypotheses for an underlying mechanism. *J Am Med Dir Assoc* 2004; **5**: 129-132 [PMID: 15008183 DOI: 10.1016/S1525-8610(04)70069-9]
- 32 **Kleijer BC**, van Marum RJ, Egberts AC, Jansen PA, Knol W, Heerdink ER. Risk of cerebrovascular events in elderly users of antipsychotics. *J Psychopharmacol* 2009; **23**: 909-914 [PMID: 18635700 DOI: 10.1177/0269881108093583]
- 33 **Knol W**, van Marum RJ, Jansen PA, Souverein PC, Schobben AF, Egberts AC. Antipsychotic drug use and risk of pneumonia in elderly people. *J Am Geriatr Soc* 2008; **56**: 661-666 [PMID: 18266664 DOI: 10.1111/j.1532-5415.2007.01625.x]
- 34 **Trifirò G**. Antipsychotic drug use and community-acquired pneumonia. *Curr Infect Dis Rep* 2011; **13**: 262-268 [PMID: 21394430 DOI: 10.1007/s11908-011-0175-y]
- 35 American Geriatrics Society updated Beers Criteria for potentially inappropriate medication use in older adults. *J Am Geriatr Soc* 2012; **60**: 616-631 [PMID: 22376048 DOI: 10.1111/j.1532-5415.2012.03923.x]
- 36 **Konovalev S**, Muralee S, Tampi RR. Anticonvulsants for the treatment of behavioral and psychological symptoms of dementia: a literature review. *Int Psychogeriatr* 2008; **20**: 293-308 [PMID: 18047764 DOI: 10.1017/S1041610207006540]
- 37 **Lonegan E**, Luxenberg J. Valproate preparations for agitation in dementia. *Cochrane Database Syst Rev* 2009; **3**: CD003945 [PMID: 19588348 DOI: 10.1002/14651858.CD003945.pub3]
- 38 **Cooney C**, Mortimer A, Smith A, Newton K, Wrigley M. Carbamazepine use in aggressive behaviour associated with senile dementia. *Int J Geriatr Psychiatry* 1996; **11**: 901-905 [DOI: 10.1002/(SICI)1099-1166(199610)11:10<901::AID-GPS409>3.0.CO;2-7]
- 39 **Tariot PN**, Erb R, Podgorski CA, Cox C, Patel S, Jakimovich L, Irvine C. Efficacy and tolerability of carbamazepine for agitation and aggression in dementia. *Am J Psychiatry* 1998; **155**: 54-61 [PMID: 9433339]
- 40 **Olin JT**, Fox LS, Pawluczyk S, Taggart NA, Schneider LS. A pilot randomized trial of carbamazepine for behavioral symptoms in treatment-resistant outpatients with Alzheimer disease. *Am J Geriatr Psychiatry* 2001; **9**: 400-405 [PMID: 11739066 DOI: 10.1176/appi.ajgp.9.4.400]
- 41 **Beattie B**, Biller J, Mehlhaus B, Murray M. Verapamil-induced carbamazepine neurotoxicity. A report of two cases. *Eur Neurol* 1988; **28**: 104-105 [PMID: 3371379 DOI: 10.1159/000116239]
- 42 **Bahls FH**, Ozuna J, Ritchie DE. Interactions between calcium channel blockers and the anticonvulsants carbamazepine and phenytoin. *Neurology* 1991; **41**: 740-742 [PMID: 2027492 DOI: 10.1212/WNL.41.5.740]
- 43 **Massey EW**. Effect of carbamazepine on Coumadin metabolism. *Ann Neurol* 1983; **13**: 691-692 [PMID: 6881938 DOI: 10.1002/ana.410130629]
- 44 **Hung SI**, Chung WH, Liu ZS, Chen CH, Hsieh MS, Hui RC, Chu CY, Chen YT. Common risk allele in aromatic antiepileptic-drug induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Han Chinese. *Pharmacogenomics* 2010; **11**: 349-356 [PMID: 20235791 DOI: 10.2217/pgs.09.162]
- 45 Information for healthcare professionals: Dangerous or even fatal skin reactions - Carbamazepine (marketed as Carbatrol, Equetro, Tegretol, and generics). 2013. [accessed 2014 September 19]. Available from: URL: <http://www.fda.gov/Drugs/DrugSafety/PostmarketDrugSafetyInformationforPatientsandProviders/ucm124718.htm>
- 46 **Mintzer JE**. Underlying mechanisms of psychosis and aggression in patients with Alzheimer's disease. *J Clin Psychia-*

- try 2001; **62** Suppl 21: 23-25 [PMID: 11584984]
- 47 **Lancôt KL**, Herrmann N, Mazzotta P. Role of serotonin in the behavioral and psychological symptoms of dementia. *J Neuropsychiatry Clin Neurosci* 2001; **13**: 5-21 [PMID: 11207325 DOI: 10.1176/appi.neuropsych.13.1.5]
- 48 **Bains J**, Birks J, Dening T. Antidepressants for treating depression in dementia. *Cochrane Database Syst Rev* 2002; **4**: CD003944 [PMID: 12519625 DOI: 10.1002/14651858.CD003944]
- 49 **Cummings JL**. The Neuropsychiatric Inventory: assessing psychopathology in dementia patients. *Neurology* 1997; **48**: S10-S16 [PMID: 9153155 DOI: 10.1212/WNL.48.5_Suppl_6.10S]
- 50 **Frisoni GB**, Rozzini L, Gozzetti A, Binetti G, Zanetti O, Bianchetti A, Trabucchi M, Cummings JL. Behavioral syndromes in Alzheimer's disease: description and correlates. *Dement Geriatr Cogn Disord* 1999; **10**: 130-138 [PMID: 10026387 DOI: 10.1159/000017113]
- 51 **Lyketsos CG**, DelCampo L, Steinberg M, Miles Q, Steele CD, Munro C, Baker AS, Sheppard JM, Frangakis C, Brandt J, Rabins PV. Treating depression in Alzheimer disease: efficacy and safety of sertraline therapy, and the benefits of depression reduction: the DIADS. *Arch Gen Psychiatry* 2003; **60**: 737-746 [PMID: 12860778 DOI: 10.1001/archpsyc.60.7.737]
- 52 **Seitz DP**, Adunuri N, Gill SS, Gruneir A, Herrmann N, Rochon P. Antidepressants for agitation and psychosis in dementia. *Cochrane Database Syst Rev* 2011; **2**: CD008191 [PMID: 21328305 DOI: 10.1002/14651858.CD008191.pub2]
- 53 **Cohen-Mansfield J**, Marx MS, Rosenthal AS. A description of agitation in a nursing home. *J Gerontol* 1989; **44**: M77-M84 [PMID: 2715584 DOI: 10.1093/geronj/44.3.M77]
- 54 **Pollock BG**, Mulsant BH, Rosen J, Sweet RA, Mazumdar S, Bharucha A, Marin R, Jacob NJ, Huber KA, Kastango KB, Chew ML. Comparison of citalopram, perphenazine, and placebo for the acute treatment of psychosis and behavioral disturbances in hospitalized, demented patients. *Am J Psychiatry* 2002; **159**: 460-465 [PMID: 11870012 DOI: 10.1176/appi.ajp.159.3.460]
- 55 **Lancôt KL**, Herrmann N. Donepezil for behavioural disorders associated with Lewy bodies: a case series. *Int J Geriatr Psychiatry* 2000; **15**: 338-345 [PMID: 10767734 DOI: 10.1002/(SICI)1099-1166(200004)15:4<338::AID-GPS119>3.0.CO;2-U]
- 56 **Fergusson E**, Howard R. Donepezil for the treatment of psychosis in dementia with Lewy bodies. *Int J Geriatr Psychiatry* 2000; **15**: 280-281 [PMID: 10713588 DOI: 10.1002/(SICI)1099-1166(200003)15:3<280::AID-GPS108>3.3.CO;2-E]
- 57 **Shea C**, MacKnight C, Rockwood K. Donepezil for treatment of dementia with Lewy bodies: a case series of nine patients. *Int Psychogeriatr* 1998; **10**: 229-238 [PMID: 9785144 DOI: 10.1017/S1041610298005341]
- 58 **Weiner MF**, Martin-Cook K, Foster BM, Saine K, Fontaine CS, Svetlik DA. Effects of donepezil on emotional/behavioral symptoms in Alzheimer's disease patients. *J Clin Psychiatry* 2000; **61**: 487-492 [PMID: 10937606 DOI: 10.4088/JCP.v61n0705]
- 59 **Matthews HP**, Korbey J, Wilkinson DG, Rowden J. Donepezil in Alzheimer's disease: eighteen month results from Southampton Memory Clinic. *Int J Geriatr Psychiatry* 2000; **15**: 713-720 [PMID: 10960883 DOI: 10.1002/1099-1166(200008)15:8<713::AID-GPS187>3.0.CO;2-I]
- 60 **Cummings JL**, McRae T, Zhang R. Effects of donepezil on neuropsychiatric symptoms in patients with dementia and severe behavioral disorders. *Am J Geriatr Psychiatry* 2006; **14**: 605-612 [PMID: 16816014 DOI: 10.1097/01.JGP.0000221293.91312.d3]
- 61 **Rösler M**, Retz W, Retz-Junginger P, Dennler HJ. Effects of two-year treatment with the cholinesterase inhibitor rivastigmine on behavioural symptoms in Alzheimer's disease. *Behav Neurol* 1998; **11**: 211-216 [PMID: 11568422 DOI: 10.1155/1999/168023]
- 62 **Aupperle PM**, Koumaras B, Chen M, Rabinowicz A, Mirski D. Long-term effects of rivastigmine treatment on neuropsychiatric and behavioral disturbances in nursing home residents with moderate to severe Alzheimer's disease: results of a 52-week open-label study. *Curr Med Res Opin* 2004; **20**: 1605-1612 [PMID: 15462693 DOI: 10.1185/030079904125004204]
- 63 **Finkel SI**. Effects of rivastigmine on behavioral and psychological symptoms of dementia in Alzheimer's disease. *Clin Ther* 2004; **26**: 980-990 [PMID: 15336465 DOI: 10.1016/S0149-2918(04)90172-5]
- 64 **McKeith I**, Del Ser T, Spano P, Emre M, Wesnes K, Anand R, Cicin-Sain A, Ferrara R, Spiegel R. Efficacy of rivastigmine in dementia with Lewy bodies: a randomised, double-blind, placebo-controlled international study. *Lancet* 2000; **356**: 2031-2036 [PMID: 11145488]
- 65 **Erkinjuntti T**, Kurz A, Gauthier S, Bullock R, Lilienfeld S, Damaraju CV. Efficacy of galantamine in probable vascular dementia and Alzheimer's disease combined with cerebrovascular disease: a randomised trial. *Lancet* 2002; **359**: 1283-1290 [PMID: 11965273 DOI: 10.1016/S0140-6736(02)08267-3]
- 66 **Emre M**, Aarsland D, Albanese A, Byrne EJ, Deuschl G, De Deyn PP, Durif F, Kulisevsky J, van Laar T, Lees A, Poewe W, Robillard A, Rosa MM, Wolters E, Quarg P, Tekin S, Lane R. Rivastigmine for dementia associated with Parkinson's disease. *N Engl J Med* 2004; **351**: 2509-2518 [PMID: 15590953 DOI: 10.1056/NEJMoa041470]
- 67 **Holmes C**, Wilkinson D, Dean C, Vethanayagam S, Olivieri S, Langley A, Pandita-Gunawardena ND, Hogg F, Clare C, Dams J. The efficacy of donepezil in the treatment of neuropsychiatric symptoms in Alzheimer disease. *Neurology* 2004; **63**: 214-219 [PMID: 15277611 DOI: 10.1212/01.WNL.0000129990.32253.7B]
- 68 **Courtney C**, Farrell D, Gray R, Hills R, Lynch L, Sellwood E, Edwards S, Hardyman W, Raftery J, Crome P, Lendon C, Shaw H, Bentham P. Long-term donepezil treatment in 565 patients with Alzheimer's disease (AD2000): randomised double-blind trial. *Lancet* 2004; **363**: 2105-2115 [PMID: 15220031 DOI: 10.1016/S0140-6736(04)16499-4]
- 69 **Tariot PN**, Cummings JL, Katz IR, Mintzer J, Perdomo CA, Schwam EM, Whalen E. A randomized, double-blind, placebo-controlled study of the efficacy and safety of donepezil in patients with Alzheimer's disease in the nursing home setting. *J Am Geriatr Soc* 2001; **49**: 1590-1599 [PMID: 11843990 DOI: 10.1111/j.1532-5415.2001.49266.x]
- 70 **Feldman H**, Gauthier S, Hecker J, Vellas B, Subbiah P, Whalen E. A 24-week, randomized, double-blind study of donepezil in moderate to severe Alzheimer's disease. *Neurology* 2001; **57**: 613-620 [PMID: 11524468 DOI: 10.1212/WNL.57.4.613]
- 71 **Emre M**, Tsolaki M, Bonuccelli U, Destée A, Tolosa E, Kutzelnigg A, Ceballos-Baumann A, Zdravkovic S, Bladström A, Jones R. Memantine for patients with Parkinson's disease dementia or dementia with Lewy bodies: a randomised, double-blind, placebo-controlled trial. *Lancet Neurol* 2010; **9**: 969-977 [PMID: 20729148 DOI: 10.1016/S1474-4422(10)70194-0]
- 72 **Reisberg B**, Doody R, Stöffler A, Schmitt F, Ferris S, Möbius HJ. Memantine in moderate-to-severe Alzheimer's disease. *N Engl J Med* 2003; **348**: 1333-1341 [PMID: 12672860 DOI: 10.1056/NEJMoa013128]
- 73 **Tariot PN**, Farlow MR, Grossberg GT, Graham SM, McDonald S, Gergel I. Memantine treatment in patients with moderate to severe Alzheimer disease already receiving donepezil: a randomized controlled trial. *JAMA* 2004; **291**: 317-324 [PMID: 14734594 DOI: 10.1001/jama.291.3.317]
- 74 **Tariot PN**, Solomon PR, Morris JC, Kershaw P, Lilienfeld S, Ding C. A 5-month, randomized, placebo-controlled trial of galantamine in AD. The Galantamine USA-10 Study Group. *Neurology* 2000; **54**: 2269-2276 [PMID: 10881251 DOI: 10.1212/WNL.54.12.2269]
- 75 **Loy C**, Schneider L. Galantamine for Alzheimer's disease and mild cognitive impairment. *Cochrane Database Syst Rev* 2006; **1**: CD001747 [PMID: 16437436 DOI: 10.1002/14651858.CD001747.pub3]
- 76 **Trinh N-H**, Hoblyn J, Mohanty S, Yaffe K. Efficacy of cho-

- linesterase inhibitors in the treatment of neuropsychiatric symptoms and functional impairment in Alzheimer disease. *JAMA* 2003; **289**: 210-216 [DOI: 10.1001/jama.289.2.210]
- 77 **Wild R**, Pettit T, Burns A. Cholinesterase inhibitors for dementia with Lewy bodies. *Cochrane Database Syst Rev* 2003; **3**: CD003672 [PMID: 12917981 DOI: 10.1002/14651858.CD003672]
- 78 **Kindermann SS**, Dolder CR, Bailey A, Katz IR, Jeste DV. Pharmacological treatment of psychosis and agitation in elderly patients with dementia: four decades of experience. *Drugs Aging* 2002; **19**: 257-276 [PMID: 12038878 DOI: 10.2165/00002512-200219040-00002]
- 79 **Mendez MF**, Shapira JS, McMurtray A, Licht E. Preliminary findings: behavioral worsening on donepezil in patients with frontotemporal dementia. *Am J Geriatr Psychiatry* 2007; **15**: 84-87 [PMID: 17194818 DOI: 10.1097/01.JGP.0000231744.69631.33]
- 80 **Aarsland D**, Perry R, Larsen JP, McKeith IG, O'Brien JT, Perry EK, Burn D, Ballard CG. Neuroleptic sensitivity in Parkinson's disease and parkinsonian dementias. *J Clin Psychiatry* 2005; **66**: 633-637 [PMID: 15889951 DOI: 10.4088/JCP.v66n0514]
- 81 **Ballard C**, Grace J, McKeith I, Holmes C. Neuroleptic sensitivity in dementia with Lewy bodies and Alzheimer's disease. *Lancet* 1998; **351**: 1032-1033 [PMID: 9546516 DOI: 10.1016/S0140-6736(05)78999-6]
- 82 MOH Clinical Practice Guidelines. Dementia. 2013. [accessed 2014 September 10]. Available from: URL: http://www.moh.gov.sg/content/moh_web/healthprofessionalsportal/doctors/guidelines/cpg_medical/2013/cpgmed_dementia_revised.html
- 83 **American Psychiatric Association**. Practice guideline for the treatment of patients with Alzheimer's disease and other dementias. 2007. [accessed 2014 September 29]. Available from: URL: <http://psychiatryonline.org/guidelines.aspx>
- 84 **Azermai M**, Petrovic M, Elseviers MM, Bourgeois J, Van Bortel LM, Vander Stichele RH. Systematic appraisal of dementia guidelines for the management of behavioural and psychological symptoms. *Ageing Res Rev* 2012; **11**: 78-86 [PMID: 21856452 DOI: 10.1016/j.arr.2011.07.002]
- 85 **Jalbert JJ**, Eaton CB, Miller SC, Lapane KL. Antipsychotic use and the risk of hip fracture among older adults afflicted with dementia. *J Am Med Dir Assoc* 2010; **11**: 120-127 [PMID: 20142067 DOI: 10.1016/j.jamda.2009.10.001]
- 86 **Schneider LS**, Dagerman KS, Insel P. Risk of death with atypical antipsychotic drug treatment for dementia: meta-analysis of randomized placebo-controlled trials. *JAMA* 2005; **294**: 1934-1943 [PMID: 16234500 DOI: 10.1001/jama.294.15.1934]
- 87 **Ballard C**, Hanney ML, Theodoulou M, Douglas S, McShane R, Kossakowski K, Gill R, Juszcak E, Yu LM, Jacoby R. The dementia antipsychotic withdrawal trial (DART-AD): long-term follow-up of a randomised placebo-controlled trial. *Lancet Neurol* 2009; **8**: 151-157 [PMID: 19138567 DOI: 10.1016/S1474-4422(08)70295-3]
- 88 **Gill SS**, Bronskill SE, Normand SL, Anderson GM, Sykora K, Lam K, Bell CM, Lee PE, Fischer HD, Herrmann N, Gurwitz JH, Rochon PA. Antipsychotic drug use and mortality in older adults with dementia. *Ann Intern Med* 2007; **146**: 775-786 [PMID: 17548409 DOI: 10.7326/0003-4819-146-11-200706050-00006]
- 89 **Kales HC**, Valenstein M, Kim HM, McCarthy JF, Ganoczy D, Cunningham F, Blow FC. Mortality risk in patients with dementia treated with antipsychotics versus other psychiatric medications. *Am J Psychiatry* 2007; **164**: 1568-1576; quiz 1623 [PMID: 17898349 DOI: 10.1176/appi.ajp.2007.06101710]
- 90 **Declercq T**, Petrovic M, Azermai M, Vander Stichele R, De Sutter AL, van Driel ML, Christiaens T. Withdrawal versus continuation of chronic antipsychotic drugs for behavioural and psychological symptoms in older people with dementia. *Cochrane Database Syst Rev* 2013; **3**: CD007726 [PMID: 23543555 DOI: 10.1002/14651858.CD007726.pub2]
- 91 **Lester P**, Kohen I, Stefanacci RG, Feuerman M. Antipsychotic drug use since the FDA black box warning: survey of nursing home policies. *J Am Med Dir Assoc* 2011; **12**: 573-577 [PMID: 21450177 DOI: 10.1016/j.jamda.2010.04.005]
- 92 **Mitka M**. CMS seeks to reduce antipsychotic use in nursing home residents with dementia. *JAMA* 2012; **308**: 119, 121 [PMID: 22782393 DOI: 10.1001/jama.2012.7422]
- 93 **Zuidema S**, Koopmans R, Verhey F. Prevalence and predictors of neuropsychiatric symptoms in cognitively impaired nursing home patients. *J Geriatr Psychiatry Neurol* 2007; **20**: 41-49 [PMID: 17341770 DOI: 10.1177/0891988706292762]
- 94 **Feng Z**, Hirdes JP, Smith TF, Finne-Soveri H, Chi I, Du Pasquier JN, Gilgen R, Ikegami N, Mor V. Use of physical restraints and antipsychotic medications in nursing homes: a cross-national study. *Int J Geriatr Psychiatry* 2009; **24**: 1110-1118 [PMID: 19280680 DOI: 10.1002/gps.2232]
- 95 **Gruneir A**, Lapane KL, Miller SC, Mor V. Is dementia special care really special? A new look at an old question. *J Am Geriatr Soc* 2008; **56**: 199-205 [PMID: 18179483 DOI: 10.1111/j.1532-5415.2007.01559.x]
- 96 **Phillips CD**, Spry KM, Sloane PD, Hawes C. Use of physical restraints and psychotropic medications in Alzheimer special care units in nursing homes. *Am J Public Health* 2000; **90**: 92-96 [PMID: 10630143 DOI: 10.2105/AJPH.90.1.92]
- 97 **Weyerer S**, Schäufele M, Hendlmeier I. Evaluation of special and traditional dementia care in nursing homes: results from a cross-sectional study in Germany. *Int J Geriatr Psychiatry* 2010; **25**: 1159-1167 [PMID: 20054837 DOI: 10.1002/gps.2455]
- 98 **Serby MJ**, Roane DM, Lantz MS, Cohen AJ, Turok A, Perlis TE. Current attitudes regarding treatment of agitation and psychosis in dementia. *Am J Geriatr Psychiatry* 2009; **17**: 174 [PMID: 19155750 DOI: 10.1097/JGP.0b013e31818cd38f]
- 99 **Schultz SK**. Atypical antipsychotic medications in Alzheimer's disease: effectiveness versus expectations. *Am J Psychiatry* 2008; **165**: 787-789 [PMID: 18593779 DOI: 10.1176/appi.ajp.2008.08040517]
- 100 **Devanand DP**, Mintzer J, Schultz SK, Andrews HF, Sultzer DL, de la Pena D, Gupta S, Colon S, Schimming C, Pelton GH, Levin B. Relapse risk after discontinuation of risperidone in Alzheimer's disease. *N Engl J Med* 2012; **367**: 1497-1507 [PMID: 23075176 DOI: 10.1056/NEJMoa1114058]
- 101 **Mamun K**, Goh-Tan CY, Ng LL. Prescribing psychoactive medications in nursing homes: current practice in Singapore. *Singapore Med J* 2003; **44**: 625-629 [PMID: 14770256]
- 102 **Oborne CA**, Hooper R, Li KC, Swift CG, Jackson SH. An indicator of appropriate neuroleptic prescribing in nursing homes. *Age Ageing* 2002; **31**: 435-439 [PMID: 12446288 DOI: 10.1093/ageing/31.6.435]
- 103 **Briesacher BA**, Limcangco MR, Simoni-Wastila L, Doshi JA, Levens SR, Shea DG, Stuart B. The quality of antipsychotic drug prescribing in nursing homes. *Arch Intern Med* 2005; **165**: 1280-1285 [PMID: 15956008 DOI: 10.1001/archinte.165.11.1280]
- 104 **Yap KZ**, Kua EH, Chan SY, Lee JY-C. Improving the appropriateness of antipsychotic prescribing for behavioral and psychological symptoms of dementia (BPSD): A pilot study of the Psychotropic Use Monitoring (PUM) Program. *O J Psych* 2014; **4**: 153-162 [DOI: 10.4236/ojpsych.2014.42020]
- 105 **Bowman CE**. Education, guidance, and equality are needed to address problem of antipsychotic prescribing in nursing homes. *BMJ* 2012; **344**: e2421 [PMID: 22474266 DOI: 10.1136/bmj.e2421]
- 106 **Barber N**. What constitutes good prescribing? *BMJ* 1995; **310**: 923-925 [PMID: 7719188 DOI: 10.1136/bmj.310.6984.923]
- 107 **Sylliaas H**, Selbaek G, Bergland A. Do behavioral disturbances predict falls among nursing home residents? *Ageing Clin Exp Res* 2012; **24**: 251-256 [PMID: 23114551]
- 108 **Barber N**, Bradley C, Barry C, Stevenson F, Britten N, Jenkins L. Measuring the appropriateness of prescribing in pri-

- mary care: are current measures complete? *J Clin Pharm Ther* 2005; **30**: 533-539 [PMID: 16336285]
- 109 **British Columbia Ministry of Health.** Best Practice Guideline for Accommodating and Managing Behavioural and Psychological Symptoms of Dementia in Residential Care. 2012. [accessed 2014 September 29]. Available from: URL: <http://www.health.gov.bc.ca/library/publications/year/2012/bpsd-guideline.pdf>
- 110 **Leipzig RM,** Cumming RG, Tinetti ME. Drugs and falls in older people: a systematic review and meta-analysis: I. Psychotropic drugs. *J Am Geriatr Soc* 1999; **47**: 30-39 [PMID: 9920227]
- 111 **Crystal S,** Olfson M, Huang C, Pincus H, Gerhard T. Broadened use of atypical antipsychotics: safety, effectiveness, and policy challenges. *Health Aff (Millwood)* 2009; **28**: w770-w781 [PMID: 19622537 DOI: 10.1377/hlthaff.28.5.w770]
- 112 **Khatutsky G,** Wiener JM, Anderson WL. Immigrant and non-immigrant certified nursing assistants in nursing homes: how do they differ? *J Aging Soc Policy* 2010; **22**: 267-287 [PMID: 20589554 DOI: 10.1080/08959420.2010.485526]
- 113 **Barber ND,** Allred DP, Raynor DK, Dickinson R, Garfield S, Jesson B, Lim R, Savage I, Standage C, Buckle P, Carpenter J, Franklin B, Woloshynowych M, Zermansky AG. Care homes' use of medicines study: prevalence, causes and potential harm of medication errors in care homes for older people. *Qual Saf Health Care* 2009; **18**: 341-346 [PMID: 19812095 DOI: 10.1136/qshc.2009.034231]
- 114 **Turnham H.** Federal Nursing Home Reform Act from the Omnibus Budget Reconciliation Act of 1987 or simply OBRA '87. Summary. [accessed 2014 September 29]. Available from: URL: <http://www.ltombudsman.org/NORC-library>
- 115 Code of Federal Regulation. Title 42 - Public Health. Chapter IV - Centers for Medicare & Medicaid Services, Department of Health and Human Services (Continued) Volume 3 (Part 483). Requirements for States and long term care facilities. Available from: URL: <http://www.gpo.gov/fdsys/search/pagedetails.action?sessionid=By16Pv7NTHxsD2yTpy00LWfHQhGwNfhZp9vQYvWKKPXnFkVnyf1830174162!744597377?collectionCode=CFR&searchPath=Title42/ChapterIV&granuleId=&packageId=CFR-2002-title42-vol1&oldPath=Title42/ChapterIV&fromPageDetails=true&collapse=false&yord=357>
- 116 **Hughes CM,** Lapane KL, Mor V, Ikegami N, Jónsson PV, Ljunggren G, Sgadari A. The impact of legislation on psychotropic drug use in nursing homes: a cross-national perspective. *J Am Geriatr Soc* 2000; **48**: 931-937 [PMID: 10968297]
- 117 **Shorr RI,** Fought RL, Ray WA. Changes in antipsychotic drug use in nursing homes during implementation of the OBRA-87 regulations. *JAMA* 1994; **271**: 358-362 [PMID: 8283585]
- 118 **Ray WA,** Blazer DG, Schaffner W, Federspiel CF. Reducing antipsychotic drug prescribing for nursing home patients: a controlled trial of the effect of an educational visit. *Am J Public Health* 1987; **77**: 1448-1450 [PMID: 2889382 DOI: 10.2105/AJPH.77.11.1448]
- 119 **Westbury J,** Jackson S, Gee P, Peterson G. An effective approach to decrease antipsychotic and benzodiazepine use in nursing homes: the RedUse project. *Int Psychogeriatr* 2010; **22**: 26-36 [PMID: 19814843 DOI: 10.1017/S1041610209991128]
- 120 **Castle NG.** Providing outcomes information to nursing homes: can it improve quality of care? *Gerontologist* 2003; **43**: 483-492 [PMID: 12937327 DOI: 10.1093/geront/43.4.483]
- 121 **Westbury J,** Tichelaar L, Peterson G, Gee P, Jackson S. A 12-month follow-up study of "RedUse": a trial aimed at reducing antipsychotic and benzodiazepine use in nursing homes. *Int Psychogeriatr* 2011; **23**: 1260-1269 [PMID: 21429285 DOI: 10.1017/S1041610211000421]
- 122 **Watson-Wolfe K,** Galik E, Klinedinst J, Brandt N. Application of the Antipsychotic Use in Dementia Assessment audit tool to facilitate appropriate antipsychotic use in long term care residents with dementia. *Geriatr Nurs* 2014; **35**: 71-76 [PMID: 24139205 DOI: 10.1016/j.gerinurse.2013.09.002]
- 123 **Avorn J,** Soumerai SB, Everitt DE, Ross-Degnan D, Beers MH, Sherman D, Salem-Schatz SR, Fields D. A randomized trial of a program to reduce the use of psychoactive drugs in nursing homes. *N Engl J Med* 1992; **327**: 168-173 [PMID: 1608408 DOI: 10.1056/NEJM199207163270306]
- 124 **Ray WA,** Taylor JA, Meador KG, Lichtenstein MJ, Griffin MR, Fought R, Adams ML, Blazer DG. Reducing antipsychotic drug use in nursing homes. A controlled trial of provider education. *Arch Intern Med* 1993; **153**: 713-721 [PMID: 8447709 DOI: 10.1001/archinte.153.6.713]
- 125 **Testad I,** Ballard C, Brønnick K, Aarsland D. The effect of staff training on agitation and use of restraint in nursing home residents with dementia: a single-blind, randomized controlled trial. *J Clin Psychiatry* 2010; **71**: 80-86 [PMID: 20129008 DOI: 10.4088/JCP.09m054860i]
- 126 **Hagen BF,** Armstrong-Esther C, Quail P, Williams RJ, Norton P, Le Navenec CL, Ikuta R, Osis M, Congdon V, Zieb R. Neuroleptic and benzodiazepine use in long-term care in urban and rural Alberta: characteristics and results of an education intervention to ensure appropriate use. *Int Psychogeriatr* 2005; **17**: 631-652 [PMID: 16246262 DOI: 10.1017/S1041610205002188]
- 127 **Monette J,** Champoux N, Monette M, Fournier L, Wolfson C, du Fort GG, Sourial N, Le Cruguel JP, Gore B. Effect of an interdisciplinary educational program on antipsychotic prescribing among nursing home residents with dementia. *Int J Geriatr Psychiatry* 2008; **23**: 574-579 [PMID: 17968860 DOI: 10.1002/gps.1934]
- 128 **Vida S,** Monette J, Wilchesky M, Monette M, Friedman R, Nguyen A, Dastoor D, Cristache G, Sourial N, Tremblay L, Gore B. A long-term care center interdisciplinary education program for antipsychotic use in dementia: program update five years later. *Int Psychogeriatr* 2012; **24**: 599-605 [PMID: 22126992 DOI: 10.1017/S1041610211002225]
- 129 **Patterson SM,** Hughes CM, Crealey G, Cardwell C, Lapane KL. An evaluation of an adapted U.S. model of pharmaceutical care to improve psychoactive prescribing for nursing home residents in northern Ireland (fleetwood northern Ireland study). *J Am Geriatr Soc* 2010; **58**: 44-53 [PMID: 20002510 DOI: 10.1111/j.1532-5415.2009.02617.x]
- 130 **Patterson SM,** Hughes CM, Cardwell C, Lapane KL, Murray AM, Crealey GE. A cluster randomized controlled trial of an adapted U.S. model of pharmaceutical care for nursing home residents in Northern Ireland (Fleetwood Northern Ireland study): a cost-effectiveness analysis. *J Am Geriatr Soc* 2011; **59**: 586-593 [PMID: 21453379 DOI: 10.1111/j.1532-5415.2011.03354.x]
- 131 **Chakraborty A,** Linton CR. Antipsychotic prescribing in dementia patients in care homes: proactive in-reach service improved quality of care. *Int J Geriatr Psychiatry* 2012; **27**: 1097-1098 [PMID: 22945348 DOI: 10.1002/gps.2827]
- 132 **Schmidt I,** Claesson CB, Westerholm B, Nilsson LG, Svarstad BL. The impact of regular multidisciplinary team interventions on psychotropic prescribing in Swedish nursing homes. *J Am Geriatr Soc* 1998; **46**: 77-82 [PMID: 9434669]
- 133 **Dahl LJ,** Wright R, Xiao A, Keeven A, Carr DB. Quality improvement in long term care: the psychotropic assessment tool (PAT). *J Am Med Dir Assoc* 2008; **9**: 676-683 [PMID: 18992701 DOI: 10.1016/j.jamda.2008.07.002]
- 134 **Fossey J,** Ballard C, Juszcak E, James I, Alder N, Jacoby R, Howard R. Effect of enhanced psychosocial care on antipsychotic use in nursing home residents with severe dementia: cluster randomised trial. *BMJ* 2006; **332**: 756-761 [PMID: 16543297 DOI: 10.1136/bmj.38782.575868.7C]
- 135 **Bird M,** Jones RH, Kortan A, Smithers H. A controlled trial of a predominantly psychosocial approach to BPSD: treating causality. *Int Psychogeriatr* 2007; **19**: 874-891 [PMID: 17234041 DOI: 10.1017/S1041610206004790]
- 136 **Sørensen L,** Foldspang A, Gulmann NC, Munk-Jørgensen P. Determinants for the use of psychotropics among nurs-

- ing home residents. *Int J Geriatr Psychiatry* 2001; **16**: 147-154 [PMID: 11241719 DOI: 10.1002/1099-1166(200102)16:2<147::AID-GPS286>3.0.CO;2-4]
- 137 **Whitaker R**, Ballard C, Stafford J, Orrell M, Moniz-Cook E, Woods RT, Murray J, Knapp M, Carlton BW, Fossey J. Feasibility study of an optimised person-centred intervention to improve mental health and reduce antipsychotics amongst people with dementia in care homes: study protocol for a randomised controlled trial. *Trials* 2013; **14**: 13 [PMID: 23305152 DOI: 10.1186/1745-6215-14-13]
- 138 **Aalten P**, de Vugt ME, Jaspers N, Jolles J, Verhey FR. The course of neuropsychiatric symptoms in dementia. Part I: findings from the two-year longitudinal Maasbed study. *Int J Geriatr Psychiatry* 2005; **20**: 523-530 [PMID: 15920712 DOI: 10.1002/gps.1316]
- 139 **Nishtala PS**, McLachlan AJ, Bell JS, Chen TF. Psychotropic prescribing in long-term care facilities: impact of medication reviews and educational interventions. *Am J Geriatr Psychiatry* 2008; **16**: 621-632 [PMID: 18669940 DOI: 10.1097/JGP.0b013e31817c6abe]

P- Reviewer: Myers CS, Mauri MC **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Lu YJ



Use of eltrombopag in thrombocytopenia of liver disease

Vishal Sharma

Vishal Sharma, Department of Gastroenterology, Postgraduate Institute of Medical Education and Research, Chandigarh 160011, India

Author contributions: Sharma V solely contributed to this paper.

Correspondence to: Vishal Sharma, Assistant Professor, Department of Gastroenterology, Postgraduate Institute of Medical Education and Research, Sector 12, Chandigarh 160011, India. docvishalsharma@gmail.com

Telephone: +91-950-1013399

Received: May 16, 2014 Revised: July 4, 2014

Accepted: September 17, 2014

Published online: December 9, 2014

Abstract

Second generation thrombopoietin agonists including eltrombopag and romiplostim act on the thrombopoietin receptor to increase the megakaryocyte production. These agents were needed as use of first generation recombinant products was associated with formation of autoantibodies. Eltrombopag is an oral thrombopoietin agonist found effective in raising platelet counts in patients with immune thrombocytopenia. The drug has now been found to be useful in raising platelet counts in thrombocytopenia related to liver disease including cirrhosis and chronic viral hepatitis. Although the drug may help enable adequate interferon therapy in patients with HCV infection and help carry out invasive procedures in patients with cirrhosis, concerns have been raised of possible thrombotic complications including portal vein thrombosis. Randomized trials have shown that use of eltrombopag concomitant with pegylated interferon and ribavirin increased the chances of sustained virologic response while decreasing the dose reductions of interferon. The data on use of romiplostim in these clinical indications is also emerging. However, in the future, availability of interferon free regimens is likely to decrease the use of eltrombopag for enabling antiviral therapy. The review discusses the role of eltrombopag in management of liver disease related thrombocytopenia in wake of recent data as also the dosage, precautions and adverse effects associated with its use.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Cirrhosis; Thrombopoietin; Eltrombopag; Romiplostim; Hepatitis; Hepatitis C virus; Splenectomy; Thrombocytopenia

Core tip: Thrombocytopenia associated with liver disease is multifactorial. Eltrombopag, a thrombopoietin agonist, has been found useful in increasing platelet counts in these patients. It has been clinically used to increase platelet counts in cirrhotic patients prior to invasive procedures and in patients with chronic hepatitis C to enable administration of interferon based antiviral therapy. However, there are concerns regarding its safety and possible increased risk of portal vein thrombosis.

Sharma V. Use of eltrombopag in thrombocytopenia of liver disease. *World J Pharmacol* 2014; 3(4): 186-192 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/186.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.186>

INTRODUCTION

The recent advances in management of immune thrombocytopenia seem to have rubbed off on the management of thrombocytopenia in liver disease. The availability of thrombopoietin agonists in recent times has added to the armamentarium to manage liver disease related thrombocytopenia. The present review focuses on the evidence regarding clinical use of eltrombopag (marketed as Revolade and Promacta) in liver disease related thrombocytopenia.

THROMBOCYTOPENIA IN LIVER DISEASE

Thrombocytopenia is an important complication of chronic liver disease but may accompany non-cirrhotic liver disease. Although various authors have used different definitions, any level of platelets below $150000/\text{mm}^3$

Table 1 Mechanisms of thrombocytopenia in liver disease

In chronic liver disease
Decreased Thrombopoietin production
Splenic sequestration
Autoantibodies against platelets
Expansion of plasma volume
Bone marrow suppression (Alcohol)
In specific liver diseases
Viral or alcohol related marrow suppression
Autoimmune thrombocytopenia
Cryoglobulins
Drugs: Interferon mediated marrow suppression

would qualify as thrombocytopenia^[1]. The incidence of thrombocytopenia has been reported from 15% to 75%^[2,3]. Multiple mechanisms may contribute to the genesis of thrombocytopenia in association with liver disease^[2,4]. These may include sequestration in the enlarged spleen and reduced thrombopoietin production by the diseased liver (Table 1). The role of antiplatelet antibodies has also been alluded to in the genesis of liver disease related thrombocytopenia^[5]. Antibodies may also account for thrombocytopenia associated with viral hepatitis^[6,7]. The improvement in thrombocytopenia after liver transplantation has been ascribed to the normalization of thrombopoietin production^[8].

The incidence of thrombocytopenia in chronic hepatitis C is higher than the general population and variable incidence (0.16%–45%) has been reported from multiple reports^[1,7,9,10]. The reason is that different definitions have been used to define thrombocytopenia and different disease stage of liver disease of included patients. A report indicated that the likelihood of having a platelet count of less than 100000/mm³ was 12 in the cirrhotic population vis-à-vis the general population^[10]. Indeed thrombocytopenia is considered an indirect marker of severity of chronic liver disease and may predict the presence of cirrhotic complication especially esophageal varices^[11,12]. In fact a ratio of platelet to spleen size may help predict the presence of esophageal varices in patients with liver disease^[12]. Thrombocytopenia of liver disease is usually not life-threatening. The importance of liver disease related thrombocytopenia relates to the difficulties in management of such patients including in administration of antiviral therapy in hepatitis C virus (HCV), or the difficulty in doing invasive procedures in chronic liver disease^[2,13]. Most importantly low platelet counts are a contraindication to start pegylated interferon related therapy in patients with HCV infection. Interferon itself causes thrombocytopenia in around 30%–35% of patients^[14,15]. In spite of recent advances in management of HCV and number of interferon free regimens becoming available, interferon remains a therapy of first choice in many countries due to the lack of availability and high costs of the newer regimens^[16].

Traditionally many treatment options were available for management of liver disease related thrombocytopenia but these were either invasive or had significant risks associated with them. Platelet transfusion remained the

standard especially in emergent settings but with the caveat that there were attendant risks of transfusion transmitted infections, febrile reactions, lung injury and alloimmunisation^[17]. Other options included splenectomy or splenic artery embolization with an intent to reduce the effects of hypersplenism to raise thrombocyte counts^[2]. Danazol, in a dosage of 300–600 mg daily has also been found effective for management of thrombocytopenia in chronic hepatitis C thereby enabling administration of antiviral therapy with pegylated interferon and ribavirin^[18]. However the availability of thrombopoietin agonists has remarkably altered the management of liver disease related thrombocytopenia.

THROMBOPOIETIN AGONISTS

Formation of platelets is a complex process wherein pluripotent hematopoietic stem cells undergo maturation to form the megakaryocytes. The production of megakaryocytes is controlled and can increase ten folds in times of need^[19]. This increase involves regulation by many factors including interleukin (IL)-3, IL-6, IL-11 and most importantly thrombopoietin. Although its existence was postulated in 1958, thrombopoietin (TPO) was discovered in 1994 by five different laboratories independently^[20]. TPO is synthesized in the liver and mediates its actions through interaction with Human anti-thrombopoietin receptor resulting in downstream activation of various signaling pathways like janus kinase/signal transducer and activator of transcription, Shc/Ras/mitogen-activated protein kinase and phosphatidylinositol 3-kinase/Akt which result in activation and proliferation of erythroid, myeloid and megakaryocytic progenitors. Therefore, TPO has been also termed as a pan-hematopoietic cytokine^[21].

Discovery of TPO and its effects on megakaryocyte proliferation resulted in efforts to use TPO and its congeners in clinical situations. Two recombinant thrombopoietins entered clinical development: recombinant full length thrombopoietin (rhTPO) and pegylated recombinant human megakaryocyte growth and development factor (PEG-MGDF). The essential difference between the two is whilst rhTPO is a full length glycosylated form of TPO, PEG-MGDF is a truncated and non-glycosylated form of TPO^[22]. Both these agents were effective in elevating platelet counts and were used in various clinical indications. rhTPO resulted in elevation in platelet counts from day 4 to day 21 of administration with peak levels on day 12^[23]. Pharmacokinetics with PEG-MDGF were also similar. However further clinical development of these otherwise excellent drugs was halted because of development of neutralizing antibodies especially with PEG-MDGF^[24,25]. However the second generation of thrombopoietin agonists soon entered clinical realm and have since added to the armamentarium available for management of immune thrombocytopenia and other thrombocytopenic disorders. Romiplostim is a peptibody which was synthesized by combining a pair of 14 amino

acid TPO peptide into IgG type 1 heavy chain resulting in a drug with significant with potent action on megakaryocyte production^[26]. Romiplostim is used as once in a week subcutaneous injection^[20,27]. Since romiplostim has no molecular homology with the human TPO, no problem of neutralizing antibodies has been noted with its use^[26]. Eltrombopag is the other second generation molecule which is orally active and is a biaryl hydrazone^[27,28].

ELTROMBOPAG

Eltrombopag is a non-peptide TPO agonist which acts on the thrombopoietin receptor to increase the production of platelets. It was first approved in 2008 for treatment of immune thrombocytopenia. Since then its role has been recognized in management of thrombocytopenia of diverse etiologies^[19,27,29]. Being structurally different from the endogenous TPO, the interaction with TPO receptor is non-competitive and additive^[30]. In relapsing or refractory immune thrombocytopenia, eltrombopag was seen to exhibit a dose dependent increase in platelet counts over doses of 30, 50 and 75 mg/d^[31]. The dosages available are 12.5, 25, 50, 75 and 100 mg. In immune thrombocytopenia the lowest dosage which achieves a platelet count of 50000/ μ L is used. Use of higher doses in chronic liver disease may predispose to more chances of portal vein thrombosis. In such a situation an initial dosage of 25 mg/d for 2 wk has been recommended^[32]. Certain precautions need to be observed whilst prescribing eltrombopag. The drug must be taken empty stomach with a 1-2 h interval between the drug intake and the meals. Concomitant calcium supplements or other polyvalent cations must be avoided and the patient must not take the drug more than once in a 24 h period. In a pharmacokinetic study in healthy volunteers it was noted that intake of calcium or magnesium and aluminum containing antacids reduced the systemic availability of the drug^[33].

Eltrombopag is absorbed to the extent of around 50% after oral ingestion and peak plasma levels are achieved in 2-6 h^[27]. Twenty percent of the drug is excreted unchanged in faeces^[34]. With increasing hepatic impairment the area under curve increased suggesting that liver plays an important role in elimination of eltrombopag^[35]. Eltrombopag also has low-distribution performance and liver is the primary site for its distribution and elimination both^[34]. Apart from immune thrombocytopenia eltrombopag has also been used in myelodysplastic syndrome, chemotherapy related thrombocytopenia, and aplastic anemia^[19]. Eltrombopag is a fairly safe drug. The most common side effects noted include headache, malaise, fever, deranged liver function tests including transaminase elevations and indirect hyper-bilirubinemia^[36,37]. Other reported adverse events include cutaneous hyperpigmentation, erythroderma, pruritic exanthema and episodes of venous thrombosis at various sites^[37-39]. Although increased cataracts had also been reported but patients had received steroids for immune thrombocytopenia^[37]. Pos-

sible reasons for stopping eltrombopag treatment include severe adverse events like thrombosis, lack of response to maximal dose for 4 wk, or elevation in transaminases more than three folds of the baseline^[40].

ELTROMBOPAG IN HEPATITIS C THERAPY

Therapy for HCV has been an area of much contemporary interest and has seen many new drugs emerge which are likely to become available globally and will result in increased rates of sustained virologic response and reduced side-effects^[41]. Interferon free regimens are now a clinical reality for all HCV genotypes^[42]. The combinations of ledipasvir and sofosbuvir given for a 12 wk duration provide standard variable rate (SVR) of more than 90% in HCV genotype 1^[43,44]. Even for genotype 2 and 3 a combination of sofosbuvir and ribavirin for 12 and 24 wk respectively provides good SVR rates^[45,46]. However pegylated interferon and ribavirin combination remains the therapy of choice for many patients due to issues of affordability and availability of the newer agents.

As previously discussed HCV is known to cause thrombocytopenia. It may also increase the risk of developing immune thrombocytopenia^[47,48]. The matter is further complicated in the patients receiving pegylated interferon and ribavirin. Interferon is known to cause thrombocytopenia by causing bone marrow suppression^[49]. In a large report on interferon therapy in HCV patients, baseline thrombocytopenia was present in 44% of patients. In patients with severe thrombocytopenia (< 75000/ μ L), the need to stop interferon or reduce its dose was much higher. Severe bleeding events were uncommon but a platelet count of < 50000/ μ L predicted an increased risk of bleeding^[50].

ENABLE-1 and 2 trials provided data regarding use of eltrombopag to ensure initiation and completion of interferon and ribavirin therapy (Table 2). In ENABLE-1 trial patients with HCV infection and platelet count of < 75000/ μ L received progressively increasing doses of eltrombopag (25, 50, 75 and 100 mg) to achieve a platelet count of > 90000/ μ L. With this strategy initiation of interferon treatment was possible in 95% cases and only in 2% cases did the count not increase to the desired level. Also 88% patients benefited with a dose of 50 mg/d or less. The group was now randomized to either receive pegylated interferon-2a and ribavirin with placebo *vs* with eltrombopag. Although the rates of RVR were similar in the two groups the rates of EVR and SVR were increased in those receiving eltrombopag. Dose reductions of interferon were higher in the placebo arm. ENABLE-2 had a similar study design except for the use of pegylated interferon 2b instead of 2a used in ENABLE-1. The results were similar with 96% patients achieving the target platelet counts. Median time to achieve the target was 2 wk. Discontinuations were higher with the placebo arm but thromboembolic events including portal vein thrombosis

Table 2 Studies of Eltrombopag in patients with liver disease

Ref.	Population	Type	Results
McHutchison <i>et al</i> ^[55]	Compensated HCV cirrhosis with thrombocytopenia	Phase II RCT, placebo controlled	Dose dependent increase noted with eltrombopag
Kawaguchi <i>et al</i> ^[32]	Cirrhosis	Phase II Randomised Open label study	Risk of thrombotic phenomenon, recommends lower dose in Japanese
Afdhal <i>et al</i> ^[56] ELEVATE trial	Cirrhosis patients, peri-procedural use	Phase III, RCT, placebo controlled	Decreased platelet transfusion with eltrombopag with increased risk of portal vein thrombosis
Afdhal <i>et al</i> ^[51] ENABLE 1 and 2 trial	HCV related thrombocytopenia, to enable SVR	Phase III, RCT, placebo controlled	Decreased dose reduction in eltrombopag group, Higher SVR

HCV: Hepatitis C virus; SVR: Standard variable rate.

were higher in the eltrombopag arm as were the rates of hepatic decompensation^[51-53]. The occurrence of portal vein thrombosis may compromise the feasibility and outcomes of liver transplantation which may be needed in these patients. Interestingly neither the dosage of eltrombopag nor the platelet counts predicted the risk of thromboembolic events in the ENABLE trials^[49]. With advent of interferon free therapies, the use of eltrombopag or other thrombopoietin agonists for enabling interferon based therapies is likely to decrease in the near future. However, there is still time before the majority of world population especially in the low income countries has access to direct acting antivirals and till that time role of eltrombopag to support difficult to treat groups like those with cirrhosis will remain^[54].

ELTROMBOPAG IN CIRRHOSIS

Compensated cirrhosis with HCV is also an indication for treatment with interferon but the presence of thrombocytopenia complicates the management. In a trial evaluating multiple dose regimens of eltrombopag for management of thrombocytopenia in HCV related cirrhosis so as to initiate interferon and ribavirin therapy, 74 patients were assigned to receive placebo, 30, 50 or 75 mg of eltrombopag daily for 4 wk. Twelve weeks therapy with the antivirals was possible only in 6% patients receiving placebo whilst a progressively larger number of patients (36%, 53%, and 65%) were able to receive therapy with increasing doses of eltrombopag (30, 50, and 75 mg respectively)^[55]. In this trial three patients required withdrawal of eltrombopag for various reasons including new onset ascites, retinal exudates and neutropenia; effects not entirely related with use of eltrombopag. Further information on use of eltrombopag in cirrhosis came from the ELEVATE trial in which patients with cirrhosis and a platelet count of < 50000/ μL who were planned for an invasive procedure received either placebo or eltrombopag in a dosage of 75 mg daily for 2 wk before the planned procedure (Table 2). In a significantly higher number of patients receiving the drug (72%) vis-à-vis the placebo (19%), the transfusion of platelets could be avoided. However, there were no differences in significant bleeding episodes^[56]. This, however, came at an

increased risk of portal vein thrombosis in the treatment arm raising concerns about the safety. Interestingly the dosage used in this trial was a higher one at 75 mg and the risk increased with higher platelet count levels^[56,57]. There are reports suggesting a higher drug exposure in East Asian population and lower initial doses have been recommended^[58,59]. In a report from Japan on 38 patients with chronic liver disease even a dosage of 12.5 mg daily resulted in a mean platelet elevation of 24000/ μL suggesting that lower doses may be effective in this population. More side effects as also serious events like portal vein thrombosis were noted in the 37.5 mg group^[32]. Other case reports have also described similar events with the usage of eltrombopag or romiplostim in chronic liver disease^[60,61]. Romiplostim has also been effective for management of HCV and cirrhosis related thrombocytopenia^[62,63]. The use of romiplostim was reported to be effective in raising the platelet count in majority of patients (33 out of 35) with chronic hepatitis C related cirrhosis to a level of > 70000/ μL thereby enabling surgical procedures. No major bleeding or thrombotic episodes were reported in this Phase II study^[64,65].

CONCLUSION

Eltrombopag is effective in treatment of thrombocytopenia of liver disease and may help in certain clinical situations. The drug may be of use to initiate and complete interferon based anti-HCV therapy and may have a role prior to invasive procedures in patients with cirrhosis. However the use must be tempered by the possible risk of thrombotic complications including portal vein thrombosis. Importantly the minimum possible dose which can achieve the requisite platelet count should be used.

REFERENCES

- 1 **Mohamed SF.** Prevalence of thrombocytopenia in Egyptian patients with chronic hepatitis C virus. *J Egypt Soc Parasitol* 2013; **43**: 617-628 [PMID: 24640862]
- 2 **Poordad F.** Review article: thrombocytopenia in chronic liver disease. *Aliment Pharmacol Ther* 2007; **26** Suppl 1: 5-11 [PMID: 17958514 DOI: 10.1111/j.1365-2036.2007.03510.x]
- 3 **Dusheiko G.** Thrombopoietin agonists for the treatment

- of thrombocytopenia in liver disease and hepatitis C. *Clin Liver Dis* 2009; **13**: 487-501 [PMID: 19628164 DOI: 10.1016/j.cld.2009.05.012]
- 4 **Peck-Radosavljevic M.** Thrombocytopenia in liver disease. *Can J Gastroenterol* 2000; **14** Suppl D: 60D-66D [PMID: 11110614]
 - 5 **Kajihara M, Okazaki Y, Kato S, Ishii H, Kawakami Y, Ikeda Y, Kuwana M.** Evaluation of platelet kinetics in patients with liver cirrhosis: similarity to idiopathic thrombocytopenic purpura. *J Gastroenterol Hepatol* 2007; **22**: 112-118 [PMID: 17201890 DOI: 10.1111/j.1440-1746.2006.04359.x]
 - 6 **Dimitroulis D, Valsami S, Stamopoulos P, Kouraklis G.** Immunological HCV-associated thrombocytopenia: short review. *Clin Dev Immunol* 2012; **2012**: 378653 [PMID: 22829850 DOI: 10.1155/2012/378653]
 - 7 **Fouad YM.** Chronic hepatitis C-associated thrombocytopenia: aetiology and management. *Trop Gastroenterol* 2013; **34**: 58-67 [PMID: 24377151 DOI: 10.7869/tg.2012.99]
 - 8 **Martin TG, Somberg KA, Meng YG, Cohen RL, Heid CA, de Sauvage FJ, Shuman MA.** Thrombopoietin levels in patients with cirrhosis before and after orthotopic liver transplantation. *Ann Intern Med* 1997; **127**: 285-288 [PMID: 9265428 DOI: 10.7326/0003-4819-127-4-199708150-00005]
 - 9 **Louie KS, Micallef JM, Pimenta JM, Forssen UM.** Prevalence of thrombocytopenia among patients with chronic hepatitis C: a systematic review. *J Viral Hepat* 2011; **18**: 1-7 [PMID: 20796208 DOI: 10.1111/j.1365-2893.2010.01366.x]
 - 10 **Bashour FN, Teran JC, Mullen KD.** Prevalence of peripheral blood cytopenias (hypersplenism) in patients with nonalcoholic chronic liver disease. *Am J Gastroenterol* 2000; **95**: 2936-2939 [PMID: 11051371 DOI: 10.1111/j.1572-0241.2000.02325.x]
 - 11 **Wehmeyer MH, Krohm S, Kastein F, Lohse AW, Lüth S.** Prediction of spontaneous bacterial peritonitis in cirrhotic ascites by a simple scoring system. *Scand J Gastroenterol* 2014; **49**: 595-603 [PMID: 24673156 DOI: 10.3109/00365521.2013.848471]
 - 12 **Giannini EG, Botta F, Borro P, Dulbecco P, Testa E, Mansi C, Savarino V, Testa R.** Application of the platelet count/spleen diameter ratio to rule out the presence of oesophageal varices in patients with cirrhosis: a validation study based on follow-up. *Dig Liver Dis* 2005; **37**: 779-785 [PMID: 15996912 DOI: 10.1016/j.dld.2005.05.007]
 - 13 **Hayashi H, Beppu T, Shirabe K, Maehara Y, Baba H.** Management of thrombocytopenia due to liver cirrhosis: a review. *World J Gastroenterol* 2014; **20**: 2595-2605 [PMID: 24627595 DOI: 10.3748/wjg.v20.i10.2595]
 - 14 **Mac Nicholas R, Norris S.** Review article: optimizing SVR and management of the haematological side effects of peginterferon/ribavirin antiviral therapy for HCV - the role of epoetin, G-CSF and novel agents. *Aliment Pharmacol Ther* 2010; **31**: 929-937 [PMID: 20175767 DOI: 10.1111/j.1365-2036.2010.04269.x]
 - 15 **Zekry A, Freiman J.** Eltrombopag: Is this "24 karat gold platelet" treatment for thrombocytopenia in cirrhosis associated with hepatitis C? *Hepatology* 2008; **47**: 1418-1421 [PMID: 18366111 DOI: 10.1002/hep.22300]
 - 16 **Muir AJ.** The rapid evolution of treatment strategies for hepatitis C. *Am J Gastroenterol* 2014; **109**: 628-635; quiz 636 [PMID: 24732866 DOI: 10.1038/ajg.2014.66]
 - 17 **Blumberg N, Heal JM, Phillips GL.** Platelet transfusions: trigger, dose, benefits, and risks. *F1000 Med Rep* 2010; **2**: 5 [PMID: 20502614 DOI: 10.3410/M2-5]
 - 18 **Alvarez GC, Gómez-Galicia D, Rodríguez-Fragoso L, Marina VM, Dorantes LC, Sánchez-Alemán M, Méndez-Sánchez N, Esparza JR.** Danazol improves thrombocytopenia in HCV patients treated with peginterferon and ribavirin. *Ann Hepatol* 2011; **10**: 458-468 [PMID: 21911886]
 - 19 **Wörmann B.** Clinical indications for thrombopoietin and thrombopoietin-receptor agonists. *Transfus Med Hemother* 2013; **40**: 319-325 [PMID: 24273485 DOI: 10.1159/000355006]
 - 20 **Kuter DJ.** Milestones in understanding platelet production: a historical overview. *Br J Haematol* 2014; **165**: 248-258 [PMID: 24528208 DOI: 10.1111/bjh.12781]
 - 21 **Geddis AE, Linden HM, Kaushansky K.** Thrombopoietin: a pan-hematopoietic cytokine. *Cytokine Growth Factor Rev* 2002; **13**: 61-73 [PMID: 11750880 DOI: 10.1016/S1359-6101(01)00030-2]
 - 22 **Vadhan-Raj S.** Clinical findings with the first generation of thrombopoietic agents. *Semin Hematol* 2010; **47**: 249-257 [PMID: 20620436 DOI: 10.1053/j.seminhematol.2010.03.004]
 - 23 **Vadhan-Raj S, Murray LJ, Bueso-Ramos C, Patel S, Reddy SP, Hoots WK, Johnston T, Papadopolous NE, Hittelman WN, Johnston DA, Yang TA, Paton VE, Cohen RL, Hellmann SD, Benjamin RS, Broxmeyer HE.** Stimulation of megakaryocyte and platelet production by a single dose of recombinant human thrombopoietin in patients with cancer. *Ann Intern Med* 1997; **126**: 673-681 [PMID: 9139552 DOI: 10.7326/0003-4819-126-9-199705010-00001]
 - 24 **Li J, Yang C, Xia Y, Bertino A, Glaspy J, Roberts M, Kuter DJ.** Thrombocytopenia caused by the development of antibodies to thrombopoietin. *Blood* 2001; **98**: 3241-3248 [PMID: 11719360 DOI: 10.1182/blood.V98.12.3241]
 - 25 **Basser RL, O'Flaherty E, Green M, Edmonds M, Nichol J, Menchaca DM, Cohen B, Begley CG.** Development of pancytopenia with neutralizing antibodies to thrombopoietin after multicycle chemotherapy supported by megakaryocyte growth and development factor. *Blood* 2002; **99**: 2599-2602 [PMID: 11895799 DOI: 10.1182/blood.V99.7.2599]
 - 26 **Shimamoto G, Gegg C, Boone T, Quéva C.** Peptibodies: A flexible alternative format to antibodies. *MAbs* 2012; **4**: 586-591 [PMID: 22820181 DOI: 10.4161/mabs.21024]
 - 27 **Sharma V, Randhawa H, Sharma A, Aggarwal S.** Eltrombopag--an oral thrombopoietin agonist. *Eur Rev Med Pharmacol Sci* 2012; **16**: 743-746 [PMID: 22913204]
 - 28 **Nurden AT, Viallard JF, Nurden P.** New-generation drugs that stimulate platelet production in chronic immune thrombocytopenic purpura. *Lancet* 2009; **373**: 1562-1569 [PMID: 19324405 DOI: 10.1016/S0140-6736(09)60255-5]
 - 29 **Tillmann HL, McHutchison JG.** Use of thrombopoietic agents for the thrombocytopenia of liver disease. *Semin Hematol* 2010; **47**: 266-273 [PMID: 20620438 DOI: 10.1053/j.seminhematol.2010.04.003]
 - 30 **Erickson-Miller CL, Delorme E, Tian SS, Hopson CB, Landis AJ, Valoret EI, Sellers TS, Rosen J, Miller SG, Luengo JL, Duffy KJ, Jenkins JM.** Preclinical activity of eltrombopag (SB-497115), an oral, nonpeptide thrombopoietin receptor agonist. *Stem Cells* 2009; **27**: 424-430 [PMID: 19038790 DOI: 10.1634/stemcells.2008-0366]
 - 31 **Bussell JB, Cheng G, Saleh MN, Psaila B, Kovaleva L, Meddeb B, Kloczko J, Hassani H, Mayer B, Stone NL, Arning M, Provan D, Jenkins JM.** Eltrombopag for the treatment of chronic idiopathic thrombocytopenic purpura. *N Engl J Med* 2007; **357**: 2237-2247 [PMID: 18046028 DOI: 10.1056/NEJMoa073275]
 - 32 **Kawaguchi T, Komori A, Seike M, Fujiyama S, Watanabe H, Tanaka M, Sakisaka S, Nakamura M, Sasaki Y, Oketani M, Hattori T, Katsura K, Sata M.** Efficacy and safety of eltrombopag in Japanese patients with chronic liver disease and thrombocytopenia: a randomized, open-label, phase II study. *J Gastroenterol* 2012; **47**: 1342-1351 [PMID: 22674141 DOI: 10.1007/s00535-012-0600-5]
 - 33 **Williams DD, Peng B, Bailey CK, Wire MB, Deng Y, Park JW, Collins DA, Kapsi SG, Jenkins JM.** Effects of food and antacids on the pharmacokinetics of eltrombopag in healthy adult subjects: two single-dose, open-label, randomized-sequence, crossover studies. *Clin Ther* 2009; **31**: 764-776 [PMID: 19446149 DOI: 10.1016/j.clinthera.2009.04.010]
 - 34 **Takeuchi K, Sugiura T, Umeda S, Matsubara K, Horikawa M, Nakamichi N, Silver DL, Ishiwata N, Kato Y.** Pharmacokinetics and hepatic uptake of eltrombopag, a novel platelet-increasing agent. *Drug Metab Dispos* 2011; **39**: 1088-1096 [PMID: 21422191 DOI: 10.1124/dmd.110.037960]
 - 35 **Bauman JW, Vincent CT, Peng B, Wire MB, Williams DD,**

- Park JW. Effect of hepatic or renal impairment on eltrombopag pharmacokinetics. *J Clin Pharmacol* 2011; **51**: 739-750 [PMID: 20663991 DOI: 10.1177/0091270010372106]
- 36 **Yoshida M**, Kanashima H, Nakao T, Ogawa Y, Hino M, Nakane T, Ohta T, Kumura T, Manabe M, Yamamura R, Yamane T. Retrospective analysis of eltrombopag for the treatment of refractory primary immune thrombocytopenia in Japan. *Rinsho Ketsueki* 2013; **54**: 444-450 [PMID: 23727682]
- 37 **Saleh MN**, Bussel JB, Cheng G, Meyer O, Bailey CK, Arning M, Brainsky A. Safety and efficacy of eltrombopag for treatment of chronic immune thrombocytopenia: results of the long-term, open-label EXTEND study. *Blood* 2013; **121**: 537-545 [PMID: 23169778 DOI: 10.1182/blood-2012-04-425512]
- 38 **Braunstein I**, Wanat KA, Elenitsas R, Xu X, Frey N, Rosenbach M. Eltrombopag-associated hyperpigmentation. *JAMA Dermatol* 2013; **149**: 1112-1115 [PMID: 23884150 DOI: 10.1001/jamadermatol.2013.5107]
- 39 **Meyer SC**, Rovó A, Tsakiris DA, Scherer K, Tichelli A, Holbro A. Severe cutaneous toxicity related to Eltrombopag. *Br J Haematol* 2013; **160**: 412-414 [PMID: 23151239 DOI: 10.1111/bjh.12126]
- 40 **Danish FA**, Koul SS, Subhani FR, Rabbani AE, Yasmin S. Considerations in the management of hepatitis C virus-related thrombocytopenia with eltrombopag. *Saudi J Gastroenterol* 2010; **16**: 51-56 [PMID: 20065578 DOI: 10.4103/1319-3767.58772]
- 41 **Liang TJ**, Ghany MG. Current and future therapies for hepatitis C virus infection. *N Engl J Med* 2013; **368**: 1907-1917 [PMID: 23675659 DOI: 10.1056/NEJMra1213651]
- 42 **Liang TJ**, Ghany MG. Therapy of hepatitis C--back to the future. *N Engl J Med* 2014; **370**: 2043-2047 [PMID: 24795199 DOI: 10.1056/NEJMe1403619]
- 43 **Afdhal N**, Zeuzem S, Kwo P, Chojkier M, Gitlin N, Puoti M, Romero-Gomez M, Zarski JP, Agarwal K, Buggisch P, Foster GR, Bräu N, Buti M, Jacobson IM, Subramanian GM, Ding X, Mo H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Mangia A, Marcellin P. Ledipasvir and sofosbuvir for untreated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1889-1898 [PMID: 24725239 DOI: 10.1056/NEJMoa1402454]
- 44 **Afdhal N**, Reddy KR, Nelson DR, Lawitz E, Gordon SC, Schiff E, Nahass R, Ghalib R, Gitlin N, Herring R, Lalezari J, Younes ZH, Pockros PJ, Di Bisceglie AM, Arora S, Subramanian GM, Zhu Y, Dvory-Sobol H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Sulkowski M, Kwo P. Ledipasvir and sofosbuvir for previously treated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1483-1493 [PMID: 24725238 DOI: 10.1056/NEJMoa1316366]
- 45 **Zeuzem S**, Dusheiko GM, Salupere R, Mangia A, Flisiak R, Hyland RH, Illeperuma A, Svarovskaia E, Brainard DM, Symonds WT, Subramanian GM, McHutchison JG, Weiland O, Reesink HW, Ferenci P, Hézode C, Esteban R. Sofosbuvir and ribavirin in HCV genotypes 2 and 3. *N Engl J Med* 2014; **370**: 1993-2001 [PMID: 24795201 DOI: 10.1056/NEJMoa1316145]
- 46 **Jacobson IM**, Gordon SC, Kowdley KV, Yoshida EM, Rodriguez-Torres M, Sulkowski MS, Shiffman ML, Lawitz E, Everson G, Bennett M, Schiff E, Al-Assi MT, Subramanian GM, An D, Lin M, McNally J, Brainard D, Symonds WT, McHutchison JG, Patel K, Feld J, Pianko S, Nelson DR. Sofosbuvir for hepatitis C genotype 2 or 3 in patients without treatment options. *N Engl J Med* 2013; **368**: 1867-1877 [PMID: 23607593 DOI: 10.1056/NEJMoa1214854]
- 47 **Stasi R**, Chia LW, Kalkur P, Lowe R, Shannon MS. Pathobiology and treatment of hepatitis virus-related thrombocytopenia. *Mediterr J Hematol Infect Dis* 2009; **1**: e2009023 [PMID: 21415958 DOI: 10.4084/MJHID.2009.023]
- 48 **Chiao EY**, Engels EA, Kramer JR, Pietz K, Henderson L, Giordano TP, Landgren O. Risk of immune thrombocytopenic purpura and autoimmune hemolytic anemia among 120 908 US veterans with hepatitis C virus infection. *Arch Intern Med* 2009; **169**: 357-363 [PMID: 19237719 DOI: 10.1001/archinternmed.2008.576]
- 49 **Leber A**, Feld JJ. Does eltrombopag really ENABLE SVR? *Gastroenterology* 2014; **146**: 339-342 [PMID: 24361434 DOI: 10.1053/j.gastro.2013.12.021]
- 50 **Maan R**, van der Meer AJ, Hansen BE, Feld JJ, Wedemeyer H, Dufour JF, Zangneh HF, Lammert F, Manns MP, Zeuzem S, Janssen HL, de Knegt RJ, Veldt BJ. Effect of thrombocytopenia on treatment tolerability and outcome in patients with chronic HCV infection and advanced hepatic fibrosis. *J Hepatol* 2014; **61**: 482-491 [PMID: 24780302 DOI: 10.1016/j.jhep.2014.04.021]
- 51 **Afdhal NH**, Dusheiko GM, Giannini EG, Chen PJ, Han KH, Mohsin A, Rodriguez-Torres M, Rugina S, Bakulin I, Lawitz E, Shiffman ML, Tayyab GU, Poordad F, Kamel YM, Brainsky A, Geib J, Vasey SY, Patwardhan R, Campbell FM, Theodore D. Eltrombopag increases platelet numbers in thrombocytopenic patients with HCV infection and cirrhosis, allowing for effective antiviral therapy. *Gastroenterology* 2014; **146**: 442-452.e1 [PMID: 24126097 DOI: 10.1053/j.gastro.2013.10.012]
- 52 **Dusheiko G**, Afdhal N, Giannini EG, Chen PJ, Han KH, Rodriguez-Torres M, Rugina S, Lawitz E, Streinu-Cercel A, Shiffman ML, Poordad F, Mostafa Kamel Y, Brainsky A, Geib J, Vasey SY, Patwardhan R, Campbell F, Theodore D. 60 Results of enable 2, a phase 3, multicenter study of eltrombopag and peginterferon alfa-2b treatment in patients with hepatitis c and thrombocytopenia. *J of Hepatology* 2012; **56**: S27 [DOI: 10.1016/S0168-8278(12)60074-9]
- 53 **Giannini EG**, Afdhal NH. Eltrombopag in patients with chronic liver disease. *Expert Opin Pharmacother* 2013; **14**: 669-678 [PMID: 23452139 DOI: 10.1517/14656566.2013.775249]
- 54 **Jayasekera CR**, Barry M, Roberts LR, Nguyen MH. Treating hepatitis C in lower-income countries. *N Engl J Med* 2014; **370**: 1869-1871 [PMID: 24720680 DOI: 10.1056/NEJMp1400160]
- 55 **McHutchison JG**, Dusheiko G, Shiffman ML, Rodriguez-Torres M, Sigal S, Bourliere M, Berg T, Gordon SC, Campbell FM, Theodore D, Blackman N, Jenkins J, Afdhal NH. Eltrombopag for thrombocytopenia in patients with cirrhosis associated with hepatitis C. *N Engl J Med* 2007; **357**: 2227-2236 [PMID: 18046027 DOI: 10.1056/NEJMoa073255]
- 56 **Afdhal NH**, Giannini EG, Tayyab G, Mohsin A, Lee JW, Andriulli A, Jeffers L, McHutchison J, Chen PJ, Han KH, Campbell F, Hyde D, Brainsky A, Theodore D. Eltrombopag before procedures in patients with cirrhosis and thrombocytopenia. *N Engl J Med* 2012; **367**: 716-724 [PMID: 22913681 DOI: 10.1056/NEJMoa1110709]
- 57 **Tripodi A**, Primignani M. Nontransfusal approach to increased platelet count in patients with cirrhosis and thrombocytopenia. *Hepatology* 2013; **58**: 1177-1180 [PMID: 23703879 DOI: 10.1002/hep.26502]
- 58 **Gibiensky E**, Zhang J, Williams D, Wang Z, Ouellet D. Population pharmacokinetics of eltrombopag in healthy subjects and patients with chronic idiopathic thrombocytopenic purpura. *J Clin Pharmacol* 2011; **51**: 842-856 [PMID: 20663993 DOI: 10.1177/0091270010375427]
- 59 **Farrell C**, Hayes SC, Wire M, Zhang J. Population pharmacokinetic/pharmacodynamic modelling of eltrombopag in healthy volunteers and subjects with chronic liver disease. *Br J Clin Pharmacol* 2014; **77**: 532-544 [PMID: 24117976 DOI: 10.1111/bcp.12244]
- 60 **Kawano N**, Hasuike S, Iwakiri H, Nakamura K, Ozono Y, Kusumoto H, Nagata K, Kikuchi I, Yoshida S, Kuriyama T, Yamashita K, Muranaka T, Kawaguchi T, Sata M, Okamura T, Ueda A, Shimoda K. Portal vein thrombosis during eltrombopag treatment for immune thrombocytopenic purpura in a patient with liver cirrhosis due to hepatitis C viral infection. *J Clin Exp Hematop* 2013; **53**: 151-155 [PMID: 23995112 DOI: 10.3960/jslrt.53.151]
- 61 **Dultz G**, Kronenberger B, Azizi A, Mihm U, Vogl TJ, Sarrazin U, Sarrazin C, Zeuzem S, Hofmann WP. Portal vein

thrombosis as complication of romiplostim treatment in a cirrhotic patient with hepatitis C-associated immune thrombocytopenic purpura. *J Hepatol* 2011; **55**: 229-232 [PMID: 21310200 DOI: 10.1016/j.jhep.2011.01.020]

- 62 **Voican CS**, Naveau S, Perlemuter G. Successful antiviral therapy for hepatitis C virus-induced cirrhosis after an increase in the platelet count with romiplostim: two case reports. *Eur J Gastroenterol Hepatol* 2012; **24**: 1455-1458 [PMID: 22890208 DOI: 10.1097/MEG.0b013e328357d5f2]
- 63 **Buccoliero G**, Urbano T, Massa P, Resta F, Pisconti S. Romip-

lostim for severe thrombocytopenia in the treatment of chronic hepatitis C virus infection: a new option for clinicians? *New Microbiol* 2014; **37**: 97-101 [PMID: 24531177]

- 64 **Moussa MM**, Mowafy N. Preoperative use of romiplostim in thrombocytopenic patients with chronic hepatitis C and liver cirrhosis. *J Gastroenterol Hepatol* 2013; **28**: 335-341 [PMID: 22849409 DOI: 10.1111/j.1440-1746.2012.07246.x]
- 65 **Wai CT**. Correcting thrombocytopenia in patients with liver diseases: a difficult hurdle. *J Gastroenterol Hepatol* 2013; **28**: 207-208 [PMID: 23339384 DOI: 10.1111/jgh.12052]

P- Reviewer: Ali AEM, Chuang WL, Hekmatdoost A, Kurtoglu E
S- Editor: Song XX **L- Editor:** A **E- Editor:** Lu YJ



Perspective of antiviral therapeutics for hepatitis C after liver transplantation

Cheng-Maw Ho, Rey-Heng Hu, Po-Huang Lee

Cheng-Maw Ho, Rey-Heng Hu, Po-Huang Lee, Department of Surgery, National Taiwan University Hospital, Taipei 100, Taiwan

Author contributions: Ho CM conceived of the study and wrote the manuscript; Hu RH and Lee PH helped revise the manuscript. Correspondence to: Cheng-Maw Ho, MD, Department of Surgery, National Taiwan University Hospital, 7 Chun-Shan S Rd, Taipei 100, Taiwan. mingho@ntu.edu.tw

Telephone: +886-2-23123456 Fax: +886-2-23568810

Received: June 24, 2014 Revised: September 22, 2014

Accepted: October 1, 2014

Published online: December 9, 2014

Abstract

Hepatitis C virus (HCV) almost recurs after liver transplantation for HCV-related liver cirrhosis or hepatocellular carcinoma. Management of HCV recurrence after liver transplantation is challenging because the traditional interferon-based therapy is often patient-intolerable and inducing cytopenia, and dose reduction is needed. The response rate in liver recipients is inferior to those of chronic HCV infection. About 5 percent of liver recipients receiving interferon-based therapy would develop immune-mediated graft injury and may need retransplantation. Recent advances of anti-HCV therapy for chronic HCV infection has evolutionary changing the schema from interferon-based, to interferon-free, and even to ribavirin-free, all oral combinations for pan-genotypes. Management of HCV recurrence after liver transplantation is currently evolving too and promising results will soon come to the stage. This "fast-track" concise review focuses on the issues relevant to HCV recurrence after liver transplantation and provides up-to-date information of the trend of the management. A real-world case demonstration of management was presented here to illustrate the potential complications of anti-HCV therapy after liver transplantation.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Hepatitis C; Recurrence; Liver transplantation; Therapeutics; Fibrosis

Core tip: Management of hepatitis C virus (HCV) recurrence after liver transplantation used to be a bothering issue due mostly to the interferon-based therapy. Current available data from treatment of chronic HCV infection shows promising results of interferon-free, or even ribavirin-free, pan-genotypic, all oral medications will soon reform the treatment of HCV recurrence after liver transplantation.

Ho CM, Hu RH, Lee PH. Perspective of antiviral therapeutics for hepatitis C after liver transplantation. *World J Pharmacol* 2014; 3(4): 193-198 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/193.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.193>

INTRODUCTION

Hepatitis C-related liver cirrhosis or hepatocellular carcinoma is the main indication of liver transplantation worldwide^[1-4]. Almost all recipients experiences post-transplant recurrence of hepatitis C virus (HCV) and some degrees of long-term graft injury^[5,6]. Although hepatitis C is a potentially curable disease because it is caused by an RNA flavivirus with 6 major genotypes^[4], and, unlike hepatitis B virus which integrates its DNA into host DNA genome and causes viral clearance difficult, therapeutic outcomes were inferior in liver recipients compared to patients of chronic HCV infection^[7].

Ongoing evolutionary positive results in "general" (non-recipient) population shed promising lights on HCV liver recipients who are still struggling to suffer from the recurrence. The aim of this concise review is to illustrate the current and, more importantly, future pictures of the anti-HCV therapeutics after liver transplantation for non-hepatologists by summarizing a great varieties of reviews

Table 1 Category, mode of action and major side-effects of anti-hepatitis C virus therapy

Category	Specific target	Mechanism of action	Major side-effect
Interferon α	Host hepatocytes and immune cells	Enhance host immune response	Flu-like symptoms
Ribavirin (nucleoside inhibitor)	Nucleoside guanosine analogue	Stop viral RNA synthesis and viral mRNA capping	Cytopenia
DAA		Enhance interferon response	
Protease inhibitor (NS3 inhibitor or NS3/4A inhibitor)	NS 3 serine protease \pm NS4A cofactor	Inhibit cleavage of the HCV proteins from the polyprecursor	Transfusion-dependent anemia; drug interaction with calcineurin inhibitors
HCV polymerase inhibitor (NS 5B inhibitor)	NS 5B protein (RNA-dependent RNA polymerase)	Inhibit HCV replication	Minimal
NS 5A replication complex inhibitor	NS 5A protein (protein for viral RNA replication and inteferon-resistance)	Stop viral RNA replication	Minimal

DAA: Direct acting antiviral agent; NS: Non-structural; HCV: Hepatitis C virus.

and articles. Detailed or extensive dissections of single agents are beyond the scope of this fast-track review and will be suggested to references.

COURSES OF HEPATITIS C AFTER LIVER TRANSPLANTATION-CONSIDER MORE THAN JUST VIRAL LOAD IN LIVER RECIPIENTS

Early experiences showed that after liver transplantation for HCV-related cirrhosis, persistent HCV infection can cause severe graft damage, and such damage is more frequent in patients infected with HCV genotype 1b than with other genotypes^[8].

As more experiences accumulated around the world, the natural history of HCV is accelerated after liver transplantation with 20%-40% progressing to cirrhosis within 5 years^[9-12]. Evidence of markers and risk factors, including clinical, serum, histopathological, or donor-related, to early predict this group of patients is summarized well in Howell *et al*^[5] and Mariño *et al*^[13] review and is still being identified. Crespo *et al*^[6] proposed simplified algorithm, combining risk factors (old donors, female recipients, diabetes mellitus, cytomegalovirus infection and corticosteroid boluses) and non-invasive liver stiffness measurement, for management of HCV recurrence after liver transplantation and further consolidated the timing for antiviral intervention.

The primary goal in this scenario is prevention of graft loss from fibrosis progression^[12]. Viral load does not correspond to graft injury^[14]. Clinicians should note that progression of fibrosis is occasionally observed even in patients who responded to treatment; in these cases, progression may be related to other factors, such as smoldering rejection, nonalcoholic steatohepatitis, other graft-related issues, or older donor or patient age^[14]. About 5% of liver recipients receiving interferon-based regimens would develop rejection or graft fibrosis, in the absence of HCV^[14]. They could be related to interferon-induced innate alloimmune response or fluctuation of immunosuppressant levels *via* drug interactions^[15-18].

Once graft injury progresses irreversibly, either through fibrosis due to HCV recurrence or through therapeutic-induced, immune-mediated injury, retransplantation is often the only option of treatment^[19,20]. Patient and graft survival rates after retransplantation in these circumstances, however, are inferior to those after primary liver transplantation^[19] although it seems no different compared to those of retransplantation due to other reasons^[20].

TRENDS OF ANTI-HCV THERAPY RELEVANT TO LIVER TRANSPLANTATION- NEW INSIGHTS TOWARD THE NEAR FUTURE

HCV was first isolated and identified in 1989^[21]. Lai *et al*^[22] pioneer study found that interferon-based therapy increased the sustained virologic response and thereafter was considered as the standard of care for HCV treatment. Interferon-based therapy augments the innate immune response to cure the virus^[23]. Ribavirin synergistically increase the interferon effect through NK cell activation^[24]. As the 3D crystalized structure of HCV was identified, more and more targets disclosed and large upcoming amounts of new drugs designed to achieve better effect^[25,26]. Non-structural (NS)3/4A protease inhibitors were added to the interferon-based regimen and increased the sustained virological response (SVR) further^[25].

The effective therapy against chronic HCV infection has improved dramatically recently, with expected SVR rates of near 75% in all previously untreated patients and the current treatment guidelines was summarized by Gane *et al*^[12].

Recently, NS 5B polymerase inhibitor further raised the response rate substantially and open the era of interferon-free, all oral, regimens^[25]. NS 5B and NS 5A fixed, once daily combination further suggest the possibility of ribavirin-free regimens in the near future^[27,28]. Shorter treatment course and pan-genotypic response, in previous treatment failure or cirrhotic patients make anti-HCV treatment revolutionary^[27-29]. The evolutionary trend of

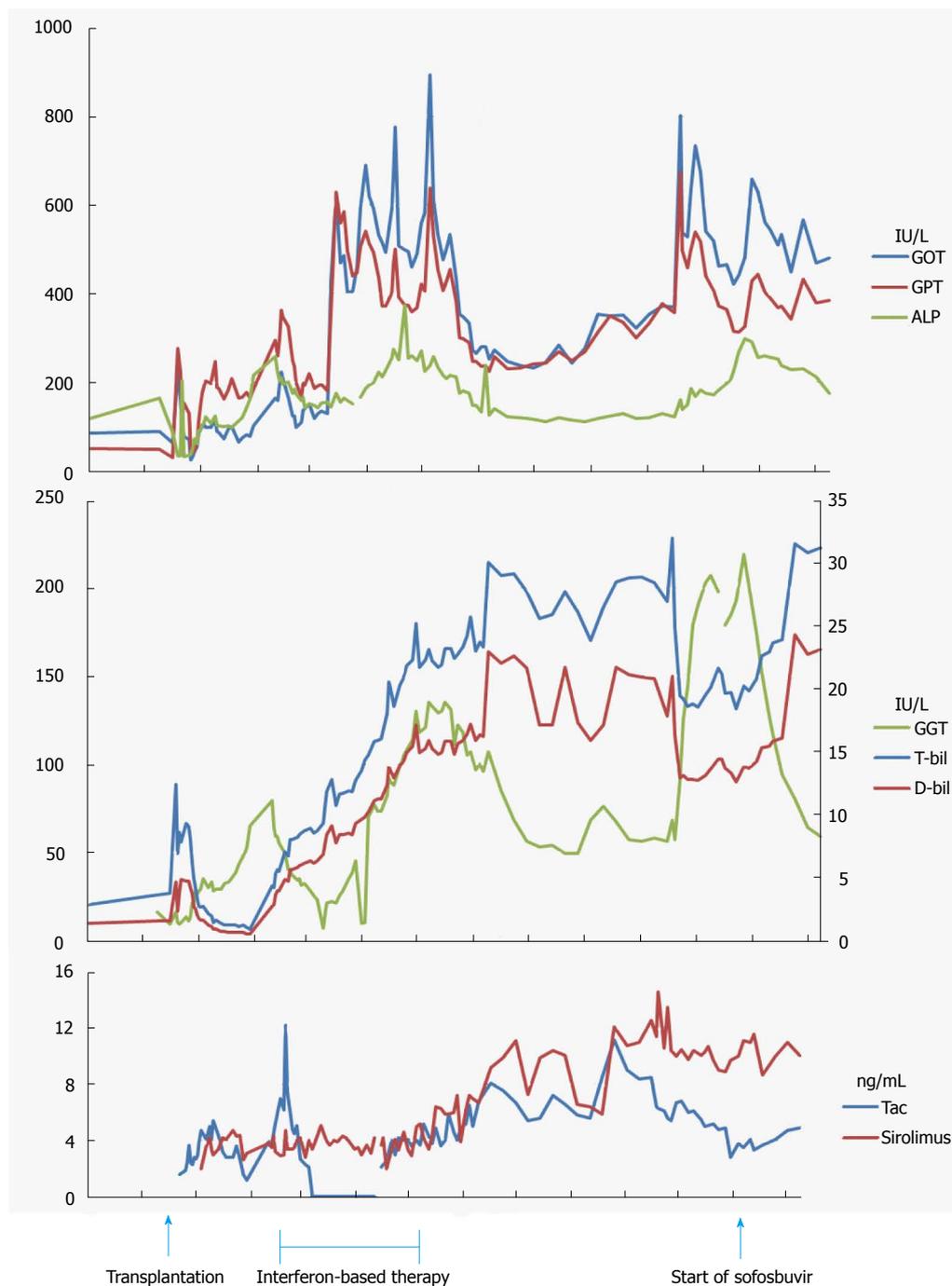


Figure 1 Post-transplant course of a liver recipient with hepatitis C recurrence.

anti-HCV therapy was nicely presented in Heim's work^[25].

Table 1 summarized the major anti-HCV therapeutic agents, mode of actions, and major side effects. High cost will be the major issue of anti-HCV therapy in the world instead^[50]. In the future, the indication for ribavirin would be limited only to non-nucleotide-based combinations or failure of other oral combination^[7,51].

ANTI-HCV THERAPY AFTER LIVER TRANSPLANTATION-THE REAL WORLD

Interferon-based regimens after liver transplantation

achieved 30% SVR in liver recipients, with higher rates achievable in patients with non-1 genotypes^[32,33]. A lot of HCV liver recipients, however, are rejected for the regimens from the start or early in the course of treatment because of the side effect intolerance. In fact, the most negative predictor for viral response is lack of tolerability from pegylated interferon/ribavirin-more than 80% of patients dose reduce and almost 30% cease therapy because of adverse effects^[12]. These results highlight the critical need for better tolerated and more efficacious HCV therapies for HCV transplant recipients^[12].

Interferon-based, NS 3A/4A added regimens after

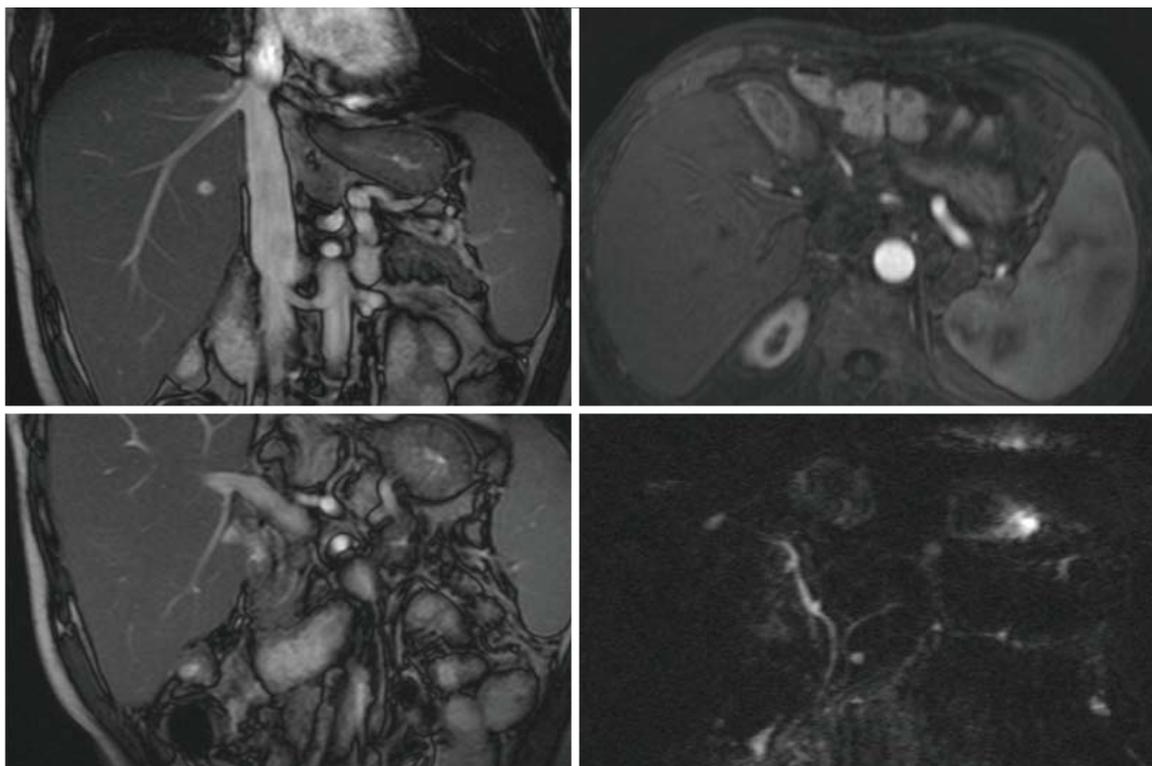


Figure 2 Magnetic resonance imaging screening for liver graft vascular or biliary complications.

liver transplantation could be further increased the SVR to 50%^[34]. Discontinuation rates were still observed high and over one third of patients need blood transfusion^[12]. In addition, antiviral therapy utilizing boceprevir in liver transplant recipients requires close monitoring of cyclosporine (5-fold) or tacrolimus (70-fold increase) due to the enzyme inhibition of the cytochrome P450 3A^[12].

A case report of the HCV liver recipient with good virological response using NS 5B inhibitor show promise of translating the success in “general” HCV population to HCV liver recipients^[35]. Sarkar *et al*^[31] reported preliminary, multi-center, promising results of sofosbuvir and ribavirin for post-transplant HCV recurrence (more than 80% had HCV genotype 1) in the Liver Meeting 2013. They found a rapidly decline of HCV after starting therapy, and over 70% of 40 recipients had SVR 4 wk after completing treatment^[36,37]. Only 2 (5%) had side effects that led to treatment discontinuation^[36,37]. These studies had actively formulating the future all-oral treatment regimens in HCV liver recipients.

REAL-WORLD CASE DEMONSTRATION

A 59 year-old man, received liver transplantation for HCV-related liver cirrhosis, was referred for prolonged and progressive jaundice since 2 mo following liver transplantation. Serial liver biopsies showed chronic hepatitis C and serum viral load was 5.8×10^6 IU/mL with the genotype 1B. Interferon-based therapy was initiated soon but lasted for 3 mo because of patient intolerance. The viral load was 1.3×10^4 IU/mL at this time. Jaundice,

however, was progressive. Laboratory data of liver profiles were shown in Figure 1. Image survey showed no evidence of vascular or biliary complications (Figure 2). Immunosuppressant regimens were tacrolimus-based initially, withdrawal transiently for the threat of drug-related cholestasis, and followed by re-initiation. The serum levels of immunosuppressant (tacrolimus and sirolimus) were shown in Figure 1. Interferon-based therapy with add-in sofosbuvir were restarted 9 mo after liver transplantation. Ribavirin and sofosbuvir were used 3 wk later without interferon because of the patient intolerance. Serum viral load dropped dramatically to undetectable 3 mo after the use of sofosbuvir, 11 mo after transplantation and remained thereafter. The latter serial liver biopsies (from 5 mo to 11 mo after liver transplantation), however, showed first acute rejection, and chronic rejection later. Steroid bolus therapy was not responsive. Liver re-transplantation was suggested.

CONCLUSION

In summary, with the rapid advances of anti-viral therapy in HCV hepatitis, the prognosis of HCV liver recipients is expected to improve greatly once the all oral, interferon-free, ribavirin-free regimens come to the stage of the standard of care with reasonable cost.

REFERENCES

- 1 Chinnadurai R, Velazquez V, Grakoui A. Hepatic transplant and HCV: a new playground for an old virus. *Am J*

- Transplant* 2012; **12**: 298-305 [PMID: 22044693 DOI: 10.1111/j.1600-6143.2011.03812.x]
- 2 **Williams R.** Global challenges in liver disease. *Hepatology* 2006; **44**: 521-526 [PMID: 16941687 DOI: 10.1002/hep.21347]
 - 3 **Lee MH, Yang HI, Lu SN, Jen CL, You SL, Wang LY, Wang CH, Chen WJ, Chen CJ.** Chronic hepatitis C virus infection increases mortality from hepatic and extrahepatic diseases: a community-based long-term prospective study. *J Infect Dis* 2012; **206**: 469-477 [PMID: 22811301 DOI: 10.1093/infdis/jis385]
 - 4 **Roche B, Samuel D.** Hepatitis C virus treatment pre- and post-liver transplantation. *Liver Int* 2012; **32** Suppl 1: 120-128 [PMID: 22212582 DOI: 10.1111/j.1478-3231.2011.02714.x]
 - 5 **Howell J, Angus P, Gow P.** Hepatitis C recurrence: the Achilles heel of liver transplantation. *Transpl Infect Dis* 2014; **16**: 1-16 [PMID: 24372756 DOI: 10.1111/tid.12173]
 - 6 **Crespo G, Mariño Z, Navasa M, Forns X.** Viral hepatitis in liver transplantation. *Gastroenterology* 2012; **142**: 1373-1383. e1 [PMID: 22537446 DOI: 10.1053/j.gastro.2012.02.011]
 - 7 **Liang TJ, Ghany MG.** Current and future therapies for hepatitis C virus infection. *N Engl J Med* 2013; **368**: 1907-1917 [PMID: 23675659 DOI: 10.1056/NEJMra1213651]
 - 8 **Gane EJ, Portmann BC, Naoumov NV, Smith HM, Underhill JA, Donaldson PT, Maertens G, Williams R.** Long-term outcome of hepatitis C infection after liver transplantation. *N Engl J Med* 1996; **334**: 815-820 [PMID: 8596547 DOI: 10.1056/NEJM199603283341302]
 - 9 **Garcia-Retortillo M, Forns X, Feliu A, Moitinho E, Costa J, Navasa M, Rimola A, Rodes J.** Hepatitis C virus kinetics during and immediately after liver transplantation. *Hepatology* 2002; **35**: 680-687 [PMID: 11870384 DOI: 10.1053/jhep.2002.31773]
 - 10 **Ballardini G, De Raffele E, Groff P, Bioulac-Sage P, Grassi A, Ghetti S, Susca M, Strazzabosco M, Bellusci R, Iemmolo RM, Grazi G, Zauli D, Cavallari A, Bianchi FB.** Timing of reinfection and mechanisms of hepatocellular damage in transplanted hepatitis C virus-reinfected liver. *Liver Transpl* 2002; **8**: 10-20 [PMID: 11799480 DOI: 10.1053/jlts.2002.30141]
 - 11 **Gane EJ, Naoumov NV, Qian KP, Mondelli MU, Maertens G, Portmann BC, Lau JY, Williams R.** A longitudinal analysis of hepatitis C virus replication following liver transplantation. *Gastroenterology* 1996; **110**: 167-177 [PMID: 8536853 DOI: 10.1053/gast.1996.v110.pm8536853]
 - 12 **Gane EJ, Agarwal K.** Directly acting antivirals (DAAs) for the treatment of chronic hepatitis C virus infection in liver transplant patients: "a flood of opportunity". *Am J Transplant* 2014; **14**: 994-1002 [PMID: 24730431 DOI: 10.1111/ajt.12714]
 - 13 **Mariño Z, Mensa L, Crespo G, Miquel R, Bruguera M, Pérez-Del-Pulgar S, Bosch J, Forns X, Navasa M.** Early periportal sinusoidal fibrosis is an accurate marker of accelerated HCV recurrence after liver transplantation. *J Hepatol* 2014; **61**: 270-277 [PMID: 24703854 DOI: 10.1016/j.jhep.2014.03.029]
 - 14 **Nair SP.** Management of hepatitis C virus infection in liver transplant recipients. *Gastroenterol Hepatol (N Y)* 2012; **8**: 56-59 [PMID: 22347835]
 - 15 **Shaked A.** The interrelation between recurrent hepatitis C, alloimmune response, and immunosuppression. *Liver Transpl* 2005; **11**: 1329-1331 [PMID: 16237699 DOI: 10.1002/lt.20588]
 - 16 **Nellore A, Fishman JA.** NK cells, innate immunity and hepatitis C infection after liver transplantation. *Clin Infect Dis* 2011; **52**: 369-377 [PMID: 21217184 DOI: 10.1093/cid/ciq156]
 - 17 **Levitsky J, Fiel MI, Norvell JP, Wang E, Watt KD, Curry MP, Tewani S, McCashland TM, Hoteit MA, Shaked A, Saab S, Chi AC, Tien A, Schiano TD.** Risk for immune-mediated graft dysfunction in liver transplant recipients with recurrent HCV infection treated with pegylated interferon. *Gastroenterology* 2012; **142**: 1132-1139. e1 [PMID: 22285805 DOI: 10.1053/j.gastro.2012.01.030]
 - 18 **Selzner N, Guindi M, Renner EL, Berenguer M.** Immune-mediated complications of the graft in interferon-treated hepatitis C positive liver transplant recipients. *J Hepatol* 2011; **55**: 207-217 [PMID: 21145865 DOI: 10.1016/j.jhep.2010.11.012]
 - 19 **Carrión JA, Navasa M, Forns X.** Retransplantation in patients with hepatitis C recurrence after liver transplantation. *J Hepatol* 2010; **53**: 962-970 [PMID: 20800307 DOI: 10.1016/j.jhep.2010.06.006]
 - 20 **Maggi U, Andorno E, Rossi G, De Carlis L, Cillo U, Bresadola F, Mazzaferro V, Risaliti A, Bertoli P, Consonni D, Barretta F, De Feo T, Scalapogna M.** Liver retransplantation in adults: the largest multicenter Italian study. *PLoS One* 2012; **7**: e46643 [PMID: 23071604 DOI: 10.1371/journal.pone.0046643]
 - 21 **Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M.** Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 1989; **244**: 359-362 [PMID: 2523562 DOI: 10.1126/science.2523562]
 - 22 **Lai MY, Kao JH, Yang PM, Wang JT, Chen PJ, Chan KW, Chu JS, Chen DS.** Long-term efficacy of ribavirin plus interferon alfa in the treatment of chronic hepatitis C. *Gastroenterology* 1996; **111**: 1307-1312 [PMID: 8898645 DOI: 10.1053/gast.1996.v111.pm8898645]
 - 23 **Feld JJ, Hoofnagle JH.** Mechanism of action of interferon and ribavirin in treatment of hepatitis C. *Nature* 2005; **436**: 967-972 [PMID: 16107837 DOI: 10.1038/nature04082]
 - 24 **Werner JM, Serti E, Chepa-Lotrea X, Stoltzfus J, Ahlenstiel G, Nouredin M, Feld JJ, Liang TJ, Rotman Y, Rehermann B.** Ribavirin improves the IFN- γ response of natural killer cells to IFN-based therapy of hepatitis C virus infection. *Hepatology* 2014; **60**: 1160-1169 [PMID: 24700342 DOI: 10.1002/hep.27092]
 - 25 **Heim MH.** 25 years of interferon-based treatment of chronic hepatitis C: an epoch coming to an end. *Nat Rev Immunol* 2013; **13**: 535-542 [PMID: 23743475 DOI: 10.1038/nri3463]
 - 26 **Madela K, McGuigan C.** Progress in the development of anti-hepatitis C virus nucleoside and nucleotide prodrugs. *Future Med Chem* 2012; **4**: 625-650 [PMID: 22458682 DOI: 10.4155/fmc.12.10]
 - 27 **Afdhal N, Zeuzem S, Kwo P, Chojkier M, Gitlin N, Puoti M, Romero-Gomez M, Zarski JP, Agarwal K, Buggisch P, Foster GR, Bräu N, Buti M, Jacobson IM, Subramanian GM, Ding X, Mo H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Mangia A, Marcellin P.** Ledipasvir and sofosbuvir for untreated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1889-1898 [PMID: 24725239 DOI: 10.1056/NEJMoa1402454]
 - 28 **Afdhal N, Reddy KR, Nelson DR, Lawitz E, Gordon SC, Schiff E, Nahass R, Ghalib R, Gitlin N, Herring R, Lalezari J, Younes ZH, Pockros PJ, Di Bisceglie AM, Arora S, Subramanian GM, Zhu Y, Dvory-Sobol H, Yang JC, Pang PS, Symonds WT, McHutchison JG, Muir AJ, Sulkowski M, Kwo P.** Ledipasvir and sofosbuvir for previously treated HCV genotype 1 infection. *N Engl J Med* 2014; **370**: 1483-1493 [PMID: 24725238 DOI: 10.1056/NEJMoa1316366]
 - 29 **Poordad F, Hezode C, Trinh R, Kowdley KV, Zeuzem S, Agarwal K, Shiffman ML, Wedemeyer H, Berg T, Yoshida EM, Forns X, Lovell SS, Da Silva-Tillmann B, Collins CA, Campbell AL, Podsadecki T, Bernstein B.** ABT-450/r-ombitasvir and dasabuvir with ribavirin for hepatitis C with cirrhosis. *N Engl J Med* 2014; **370**: 1973-1982 [PMID: 24725237 DOI: 10.1056/NEJMoa1402869]
 - 30 **Hoofnagle JH, Sherker AH.** Therapy for hepatitis C--the costs of success. *N Engl J Med* 2014; **370**: 1552-1553 [PMID: 24725236 DOI: 10.1056/NEJMe1401508]
 - 31 **Sarkar S, Lim JK.** Advances in interferon-free hepatitis C therapy: 2014 and beyond. *Hepatology* 2014; **59**: 1641-1644 [PMID: 24590916 DOI: 10.1002/hep.27055]
 - 32 **Berenguer M.** Systematic review of the treatment of established recurrent hepatitis C with pegylated interferon in combination with ribavirin. *J Hepatol* 2008; **49**: 274-287 [PMID: 18571272 DOI: 10.1016/j.jhep.2008.05.002]

- 33 **Xirouchakis E**, Triantos C, Manousou P, Sigalas A, Calvaruso V, Corbani A, Leandro G, Patch D, Burroughs A. Pegylated-interferon and ribavirin in liver transplant candidates and recipients with HCV cirrhosis: systematic review and meta-analysis of prospective controlled studies. *J Viral Hepat* 2008; **15**: 699-709 [PMID: 18673428 DOI: 10.1111/j.1365-2893.2008.01019.x]
- 34 **Joshi D**, Carey I, Agarwal K. Review article: the treatment of genotype 1 chronic hepatitis C virus infection in liver transplant candidates and recipients. *Aliment Pharmacol Ther* 2013; **37**: 659-671 [PMID: 23432320 DOI: 10.1111/apt.12260]
- 35 **Fontana RJ**, Hughes EA, Bifano M, Appelman H, Dimitrova D, Hindes R, Symonds WT. Sofosbuvir and daclatasvir combination therapy in a liver transplant recipient with severe recurrent cholestatic hepatitis C. *Am J Transplant* 2013; **13**: 1601-1605 [PMID: 23593993 DOI: 10.1111/ajt.12209]
- 36 Charlton MR, Gane EJ, Manns MP, Brown RS, Curry MP, Kwo P, Fontana R, Gilroy R, Teperman L, Muir A, McHutchinson JG, Symonds WT, Denning J, McNair L, Arterburn S, Terrault N, Samuel D, Forns X. Sofosbuvir and ribavirin for the treatment of established recurrent hepatitis C infection after liver transplantation: preliminary results of a prospective, multicenter study. The 64th Annual Meeting of the American Association for the Study of Liver Diseases; 2013 Nov 1-5; Washington, DC
- 37 Highleyman L. AASLD 2013: Sofosbuvir taken before or after liver transplant reduces HCV recurrence. [updated 2013 November 8]. 2013. Available from: URL: <http://www.hivandhepatitis.com/hcv-treatment/experimental-hcv-drugs/4400-aasld-2013-sofosbuvir-taken-before-or-after-liver-transplant-reduces-hepatitis-c-recurrence>

P- Reviewer: Jr JDA, Simkhovich BZ S- Editor: Tian YL
L- Editor: A E- Editor: Lu YJ



Vitamin D and bone fracture healing

Marks Ray

Marks Ray, Department of Health, Physical Education, Gerontological Studies and Services, School of Health and Behavioral Sciences, City University of New York, York College, New York, NY 11451, United States

Marks Ray, Department of Health and Behavior Studies, Teachers College, Columbia University, New York, NY 10027, United States

Author contributions: Ray M contributed to this paper.

Correspondence to: Dr. Marks Ray, Department of Health and Behavior Studies, Teachers College, Columbia University, Box 114, 525W, 120th Street, New York, NY 10027, United States. rm226@columbia.edu

Telephone: +1-212-6783445 Fax: +1-212-6788259

Received: July 28, 2014 Revised: September 22, 2014

Accepted: October 14, 2014

Published online: December 9, 2014

Abstract

AIM: To examine whether vitamin D is of potential relevance in the healing process of fractures.

METHODS: The present narrative review examined the bulk of the evidence based literature on the topic of vitamin D and bone healing in key electronic data bases from 1980 onwards using the terms vitamin D and bone healing, callus, fracture healing. All data were examined carefully and categorized according to type of study. A summary of the diverse terms and approaches employed in the research, as well as the rationale for hypothesizing vitamin D has a role in fracture healing was detailed.

RESULTS: The results show very few human studies have been conducted to examine if vitamin D is effective at promoting post fracture healing, and the different animal models that have been studied provide no consensus on this topic. The terms used in the related literature, as well as the methods used to arrive at conclusions on this clinical issue are highly diverse, there is no standardization of either of these important terms and methodologies, hence no conclusive statements or clinical guidelines can be forthcoming. There is a strong rationale for

continuing to examine if vitamin D supplements should be administered post-fracture, and ample evidence vitamin D is an essential hormone for functioning in general, as well as bone health and muscle as this relates to bone density.

CONCLUSION: Whether those with low vitamin D levels can benefit from supplements if their nutritional practices do not cover recommended daily amounts, remains in question.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Bone healing; Callus formation; Fractures; Fracture healing; Vitamin D

Core tip: This work describes the status of research on the role of vitamin D in bone healing, and offers suggestions for future research and current clinical practice.

Ray M. Vitamin D and bone fracture healing. *World J Pharmacol* 2014; 3(4): 199-208 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/199.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.199>

INTRODUCTION

Bone fractures are an important cause of morbidity and often, premature mortality among the older population. Among athletes and others, bone fractures due to trauma or excessive stress can seriously impair function and future activities and aspirations. In both older persons as well as younger persons minimizing the bone healing time, while maximizing bone strength of the fracture site during healing are important outcomes of the therapeutic process. Because inactivity as a result of a fracture is detrimental both to bone healing and health, and may exacerbate or foster vitamin D insufficiency or deficiency, it appears early or accelerated fracture healing would be highly desirable for returning fracture patients to function as soon as possible with minimal side effects.

The term vitamin D or cholecalciferol, which refers to a group of structurally related metabolites obtained either from dietary sources, supplementation, or sunlight and, bound by vitamin D binding protein is transported to the liver where hydroxylating enzymes initially catalyze it to form 25(OH)D (25-hydroxycholecalciferol). This product is then transported to the kidney where a second hydroxyl group is added to form 1,25-dihydroxycholecalciferol, the biologically active form of vitamin D^[1]. Vitamin D is critically important for the development, growth, and maintenance of a healthy skeleton. Calcitriol or 1,25(OH)₂D₃, the dominant D(3)-hormone and active form produces a wide array of biological responses by interacting with vitamin D nuclear receptors [VDR(nuc)] that regulate gene transcription in over 30 target organs and with a putative cell membrane receptor [VDR(mem1,25)] that mediates rapid biological responses^[2]. A second type of receptor is a cell surface vitamin D receptor^[1].

Not surprisingly, even though the nomenclature is highly varied in the related literature^[1], a substantive body of research implies low vitamin D levels can significantly increase fracture risk, as well as increase the risk of fragility fractures^[3]. By contrast, vitamin D supplements can reportedly reduce bone loss, especially at common fracture sites due to its effect on bone mineralization and maintenance^[4]. As well, physical activities alone, and especially those that improve muscular loading of bone may enhance bone health and reduce fracture risk, whilst inactivity or muscle weakness may increase the risk of falls and subsequent fractures, and here again vitamin D can play a positive role as suggested by research conducted by Beudart *et al*^[5] and Shuler *et al*^[6] and Tieland *et al*^[7].

As outlined by Schindeler *et al*^[8], fracture healing is a complex event involving a variety of differing processes. To better understand if fracture healing itself can be accelerated by the use of vitamin D supplements, either as a result of its impact on bone, or muscle or both, as suggested by Schunak^[2] and Smith *et al*^[3] this present review was designed to examine more closely, if vitamin D levels consistently predict the extent or rate of post-fracture bone healing, either directly through their osteogenic effects or indirectly through their effects on muscle function.

Since the literature remains equivocal about whether supplementation may be desirable for promoting bone healing in fracture cases, despite considerable prior discussions on this topic, it was felt a broad examination of the available literature would be helpful in this regard. The term fracture healing in this paper refers to the different stages during one of the four stages of fracture repair, but these are not strictly delineated as there is overlap in these stages, namely inflammation, soft callus formation, hard callus formation, and bone remodeling^[8]. The terminology adopted to describe vitamin D in this paper is that most commonly used in the related literature, rather than any generic term as there is considerable diversity in this respect and it is highly challenging to interpret or standardize successfully (Table 1).

That is, employing the terminology of the authors whose work is reviewed, this review sought to examine whether deficiencies or insufficiencies in serum levels of 25 hydroxyvitamin D, the metabolite recommended for determining vitamin D status in humans^[1], and 1,25-dihydroxyvitamin D, the hormone related to bone and muscle health, are specifically related to the fracture healing process.

At the same time it was hoped the review would provide recommendations for future research and practice in this area, given that the paper by Esche *et al*^[9] published in 2011 concluded there were too few human based studies to arrive at conclusive recommendations.

MATERIALS AND METHODS

Using the same search strategy as Esche *et al*^[9], the search term Vitamin D and Fracture Healing; produced 130 citations (of which 43 were relevant); Vitamin D and Bone Healing; produced 318 cited studies; Vitamin D and Callus Formation; produced 51 cited studies. Compared to Vitamin D alone: that had 59559 cited studies, it can be seen that although the topic is increasing in terms of citations, it is still understudied relative to other topics in the field. Accepted as valid sources of information were literature reviews, case studies, cross-sectional studies, prospective studies, and topics related to healing both direct and indirect that involved the topic of vitamin D and fracture healing or fracture non-union situations, and that appeared to address the topic of interest in this review.

RESULTS

Animal studies

Briggs *et al*^[10] mention that dihydroxylated vitamin D metabolites may play a key role on fracture healing as shown by enhanced serum levels of 24R, 25-dihydroxyvitamin D levels in the long bone post fracture period. This idea has been examined for almost three decades and was supported early on by a number of studies using various animal models, such as the chick^[11,12], mice^[13], rat^[14], and rabbit^[15].

Melhus *et al*^[16] who examined if osteoporosis and the healing of fractured osteoporotic bone were related, studied this issue in vitamin-D depleted ovariectomized rats known to induce weakening of the femoral neck. After initial ovariectomy, the rats were allocated to vitamin D deficient diets and sham operated rats received normal diets. At 12 wk, a fracture was induced in the tibia and fixed with a nail. Bone and callus formation were monitored with bone scans and vitamin D serum levels were measured. The results showed the experimental group had reduced bone mass, but no differences were found in the mechanical properties of the callus between the groups. The authors concluded that vitamin D is not crucial for fracture healing or for enhancing the mechanical properties of callus. This was a similar overall finding to that of Mao *et al*^[17] who examined the influence of

Table 1 Diverse vitamin D terminology and modes of assessment in the related literature and related source

Serum 25(OH)D, 24R,25(OH) ₂ D, 1,25(OH) ₂ D ^[10]
Vitamin D ₂ ^[25]
24R,25-dihydroxyvitamin D ₃ [24R,25(OH) ₂ D ₃], and 1 α,25-dihydroxyvitamin D ₃ [1 α, 25(OH) ₂ D ₃] hormonally active vitamin D metabolites ^[24]
Plasma 1,25-dihydroxyvitamin D ₃ 25(OH) ₂ D ₃ ^[28]
24R,25-dihydroxyvitamin D ₃ ^[29]
25OHD concentration ^[36]
Serum 25-hydroxyvitamin D ^[37]
Serum 25(OH)D ₃ ^[38]
Serum 25-hydroxyvitamin D (25-OH-D ₃ , 24,25 dihydroxyvitamin D ₃ [24,25(OH) ₂ D ₃], 1,25 dihydroxyvitamin D ₃ [1,25(OH) ₂ D ₃] ^[44]
Serum 25-hydroxyvitamin D, 1,25 dihydroxyvitamin D ₃ , 24,25 dihydroxyvitamin D ₃ metabolites ^[46]
25-hydroxyvitamin D [25(OH) ₂ D ₃], 1,25 dihydroxyvitamin D ₃ [1,25(OH) ₂ D ₃], and 24,25 dihydroxycholecalciferol; 24,25(OH) ₂ D ₃ -active metabolites of vitamin D ₃ ^[30]
1,25 dihydroxyvitamin D [1,25(OH) ₂ D]-biologically active metabolite of vitamin D; 24,25(OH) ₂ D ₃ -a metabolite of vitamin D ^[54]
1,25(OH)D ^[58]
Vitamin D 25(OH)D ^[63]

Vitamin D refers to an inactive compound ingested from the diet or produced after exposure of skin to sunlight. 25-hydroxyvitamin D [25-(OH)D] is an inactive metabolite produced in the liver that is hydroxylated in the kidney to form 1-α,25-dihydroxyvitamin D [1,25(OH)₂D] is the active form of vitamin D that binds to vitamin D receptor or VDR on target tissues^[11]. 24R,25-dihydroxyvitamin D₃ [24R,25(OH)₂D₃] is an essential vitamin D metabolite^[24,29].

both diabetes and vitamin D deficiency on bone repair in female mice. Although vitamin D deficiency aggravated the decrease in bone mineral density according to the diabetic state of the mice, it did not affect bone repair delayed by the diabetic state.

Hong *et al*^[18] examined the potential effects of vitamin D on bone regeneration in dogs. Their results indicated that when combined with calcium, vitamin D supplementation may have positive systemic effects that influence bone regeneration more speedily. Similarly, Fu *et al*^[19] found the effect of 1,25-dihydroxy vitamin D on fracture healing and bone remodeling in ovariectomized rat femora to favor fracture healing by improving the histological parameters of the bone, its mechanical strength, and tendency to increase transformation of woven bone into lamellar bone. Blahos *et al*^[14] who investigated the impact of 1,25-dihydroxycholecalciferol on local healing of artificially induced tibial fracture in the rat, found the contributory effect to increase the weight of the fractured tibias. This was explained by its stimulatory effect on callus formation. Omeroğlu *et al*^[15] found a single high-dose of vitamin D₃ did show positive effects in the healthy rabbit as far as fracture healing goes. This was supported by observations of increases in the sites mechanical strength after the administration of the high-dose vitamin D₃.

Likewise, Liu *et al*^[20] who examined the effect of vitamin D supplementation on the fixation of titanium implants in mice with chronic kidney disease—a problem that negatively affects bone regeneration and fracture healing, showed the bone-implant contact ratio and bone volume around the implant were significantly increased in the vitamin D supplementation group. It was concluded that these results implied vitamin D supplementation is an effective approach for improving titanium implants fixation in cases of chronic kidney disease. This is consistent with the finding by Gigante *et al*^[21] that vitamin D is able to stimulate osteoblast differentiation of fracture site derived mesenchymal stem cells, and that administra-

tion of 25-OH-vitamin D after a fracture can improve the fractured bone's mechanical strength^[22] and accelerate the initial mineralization process in the healing fracture region^[23]. It was also consistent with the observation by Kato *et al*^[24] that there is a biological role for 24R, and 25 (OH)₂D₃ forms of vitamin D in the fracture healing process. This group actually found the presence of its receptor/binding protein in a callus membrane fraction of a chick tibial fracture.

Contrary results however, were those of Sun *et al*^[25] who found vitamin D binding protein had no effect on enhancing healing in rat bone defects. Melhus *et al*^[16] too found vitamin D deficiency was not crucial for fracture healing or the mechanical properties of the callus, in rats with osteoporosis induced by ovariectomy. Lindgren *et al*^[26] produced evidence that 1,25(OH)₂D₃ actually impairs fracture healing/in the rabbit, as did Andreen and Larsson in the rat^[27]. Yet Jingushi *et al*^[28] found serum 1 α, 25 dihydroxy vitamin D₃ does accumulate into the fracture callous during rat femoral fracture healing. The authors suggested that plasma 1,25(OH)₂D₃ becomes localized in the callous, possibly regulating processes of fracture healing, a finding similar to that of Seo *et al*^[29] Dekel *et al*^[30] who examined fractures of the right tibia of chicks depleted of vitamin D, or given vitamin D₃ that were subsequently tested mechanically with respect to torsional stress, showed benefits of vitamin D. In this respect, they found repletion with 24,25(OH)₂D₃ and 1,25(OH)₂D₃ produced the most marked effects.

In sum, it is difficult to arrive at any consensus among the many approaches taken to examine the role of vitamin D on bone healing in the context of animal models. Results vary across models, as well as in the same models, and research approach, compounds, metabolites, and vitamin D derivatives are highly heterogeneous and unstandardized (Table 2).

Human studies

A good account of early clinical studies examining the

Table 2 Sample of studies using animal models to examine vitamin D influence on bone healing

Researchers	Model	Finding
Andreen <i>et al</i> ^[27]	Rat	Low doses 1,25(OH) ₂ D ₃ increased early callus mineralization
Blahos <i>et al</i> ^[14]	Rat tibia	1,25(OH) ₂ D ₃ may produce a general response
Brumbaugh <i>et al</i> ^[45]	Chick	Chicks without 1 α , 25 dihydroxy D ₃ supplementation showed prolonged fracture healing; 1 α , 25 dihydroxyvitamin D ₃ promotes bone repair in the absence vitamin D ₃ , 25 hydroxyvitamin D ₃ , and 24, 25 dihydroxyvitamin D ₃
Dekel <i>et al</i> ^[30]	Chick	24,25(OH) ₂ D ₃ , as well as 1,25(OH) ₂ D ₃ are essential for bone formation after fracture
Fu <i>et al</i> ^[19]	Rat	Vitamin D affected fracture healing positively for up to 12 wk compared to controls both biomechanically and histologically
Lindgren <i>et al</i> ^[49]	Adult rat	Rats given 1,25(OH) ₂ D ₃ had stronger fracture callus
Lindgren <i>et al</i> ^[26]	Rabbit	1,25(OH) ₂ D ₃ impairs fracture healing
Lidor <i>et al</i> ^[12]	Chick	Active metabolites of vitamin D ₃ are involved directly in fracture repair
Melhus <i>et al</i> ^[16]	Rat	Vitamin D deficiency does not impact fracture healing
Omeroglu <i>et al</i> ^[15]	Rabbit	A single high dose of vitamin D ₃ had Positive mechanical effects on fractured bone
Seo <i>et al</i> ^[29]	Chicken	24,25(OH) ₂ D ₃ levels increased during fracture repair
Steier <i>et al</i> ^[23]	Rat	Vitamin D ₂ accelerated initial mineralization in the fracture healing region

role of vitamin D in fracture healing has been provided by Gorter *et al*^[31] Among these studies, research by Doetsch *et al*^[32] tried to quantify the healing process of an osteoporotic fracture and to quantify the impact of vitamin D supplementation on the healing process among 30 women randomly assigned to a 800 IU vitamin D plus 1 g calcium or placebo in a double blinded prospective study. The researchers examined the mechanical properties of bone, as well as radiographs to evaluate healing. Bone mineral density was comparable among groups at baseline, and both increased over the 2 wk period. The authors found positive benefits of vitamin D₃ and calcium over the first 6 wk of the fracture for the active group.

Briggs *et al*^[10] conducted a prospective study to examine the extent of bioavailable levels of vitamin D metabolites among 28 patients after a cross-shaft fracture of the long bone. They measured serum concentrations of 3 vitamin D metabolites within 48 h of a fracture, and at 1 wk and 6 wk post fracture. They found no change in serum concentrations of 25(OH)D or 24R,25(OH)₂D at any time. Mean serum 1,25(OH)₂D declined 21% over the course of the study, but no changes in bioavailable concentrations of any vitamin D metabolite were seen over the course of the study.

In a case study reported by Parchi *et al*^[33] who examined the impact of vitamin D on the fracture healing process in a child, the authors found deficient vitamin D was a possible cause of the observed inadequate fracture healing process. More specifically, this research showed a significant effect on callus formation with the addition of vitamin D supplementation. Similarly, as reported by Pourfeizi *et al*^[34] who conducted a case control study of 30 patients with tibial non union compared with 32 patients with normal bone healing, a high percentage of vitamin D deficiency was observed in tibial unexplained nonunion compared to normal union. Accordingly, the authors suggested vitamin D deficiency was a possible explanation for nonunion of traumatic fractures. This finding, which generally supported the observation of Van

Denmark *et al*^[35] of a relationship between non union of a distal tibial stress fracture associated with vitamin D deficiency, was contrary to that reported by Boszczyk *et al*^[36] who compared vitamin D concentrations in patients with normal and impaired bone union. These authors found a vitamin D deficiency in 86% of examined patients. They found no difference though in 35 patients either with normal or with impaired bone healing. This was a retrospective case-control study, not a prospective randomized controlled study.

Ettehad *et al*^[37] who recently examined changes in the vitamin D levels in the serum during healing with respect to fractures of the tibial and femoral shafts found levels of vitamin D declined by the end of the third week after the fracture. They felt this was demonstrative of the fact vitamin D is important in the formation and mineralization of the callus, and consequently supplements of vitamin D administered during the healing process might be helpful in those patients with tibial or femoral shaft fractures. Again, Wöflfl *et al*^[38] who examined the time course of 25(OH)D during fracture healing in persons with fractures of healthy bone *vs* osteoporotic bone over an eight week period found no inter group differences, making it difficult to establish a definite role for vitamin D in fracture healing in this case controlled study.

A more positive finding in favor of supplementation with vitamin D was reported by Gomberg *et al*^[39]. This group described the outcome of efforts to heal subtrocchanteric stress fractures caused by excessive long term treatment with alendronate. They found treatment with large doses of oral vitamin D increased serum 25-hydroxyvitamin D₃ to normal levels in 2 mo, after which it remained in the normal range using a maintenance dosage. Although fractures appeared worse on magnetic resonance imaging at 2 mo, 6 mo later, in conjunction with teriparatide treatment and calcium, there was faint bridging of cortical bone, and complete fracture healing occurred over the next year. The combined treatment seemed beneficial to the patient. Inklebarger *et al*^[40] have also argued recently for the need to consider the presence

of low vitamin D levels when investigating the causes and possible interventions of femoral and tibial stress fractures in soldiers which may delay healing of these fractures, which is consistent with the finding that serum vitamin D levels are generally low in trauma cases in the United States^[41]. In accord with the favorable results of Kato *et al*^[42] in an *in vitro* experiment, and findings of low vitamin D levels among fallers^[43], the most common cause of hip fractures in older people. Alkalay *et al*^[44] found serum 1,25-dihydroxyvitamin D₃ [1,25(OH)₂D₃] was significantly reduced in the fracture patient, even though serum 25 hydroxyvitamin D₃(25-OH-D₃) and 24,25 dihydroxyvitamin D₃ [24, 25(OH)₂D₃] did not differ significantly between fracture patients and elective patients.

The data is confusing though because while Brumbaugh *et al*^[45] indicated 1 alpha, 25-hydroxyvitamin D₃ promotes bone repair, Haining *et al*^[46] found vitamin D metabolites had no influence in explaining fracture non-union, even though vitamin D supplementation after traction avulsion fracture was recommended by Inkelbarger *et al*^[47] and may have indirect beneficial bone effects^[48] and appeared to have favorable healing effects in adult rats^[49]. Tauber *et al*^[50] found blood levels of several active vitamin D metabolites were decreased in some fracture patients, but not others, and attributed the decrease to their consumption during fracture healing. Meller *et al*^[51] found a significant rise in plasma 24,25(OH)₂-D₃ on the day of the fracture compared to the level measured six weeks later, but no significant changes in plasma 25(OH)D₃ levels, in young patients with fractures, and suggested a physiological role for 24,25(OH)₂-D₃ in human fracture healing. In an animal model, Seo *et al*^[29] implied that 24,25(OH)D₃ seems to be involved in the early stage of fracture repair and there is some form of physiological communication between the fractured bone and kidney that results in an increase of the renal derived 24-hydroxylase and circulating concentration of this metabolite. However contrary to research by Hoikka *et al*^[52], Omeroglu *et al*^[53], and Lidor *et al*^[54], Osório *et al*^[55] found no changes in serum levels of 24R,25(OH)₂D, although levels of 1,25-dihydroxyvitamin D decreased after fracture over a 6 wk period.

In sum, as outlined in Table 1 and Table 3, the limited data in this area is highly variable and there is consequently little definitive data on whether vitamin D is helpful or not to the healing human fracture, although ample rationale for its post-fracture application exists (Table 4).

DISCUSSION

Fractures, especially those that occur among the elderly are considered to place an enormous burden on the individual, as well as on societies and their social and economic wellbeing. Considerable research shows that high rates of vitamin D insufficiency, referring to serum 25(OH)D concentrations less than 20 ng/mL^[1], currently prevail in a high proportion of cases who sustain traumatic fractures^[41], especially among the elderly. Consequently, improving vitamin D levels for these fracture patients has

been advocated^[41] to concentrations greater than 30 ng/mL^[1] is advocated. But is there sufficient evidence for this idea? Esche *et al*^[9] who conducted a short literature review that examined the question of whether vitamin D supplementation is beneficial for fracture healing found only two studies that were clinically oriented, and that most were studies using a wide variety of animal models. As they observed, both in the non human, as well as the human studies, there are negative, as well as positive results supporting vitamin D supplementation for enhancing fracture healing. As indicated by these authors, at a minimum, more research on larger samples, with more robust research designs, and a careful differentiation of baseline vitamin D status and agreed upon methods of determining vitamin D status is strongly recommended. In particular, more follow up studies, including a focus on events that take place at the four distinct phases of healing could be highly revealing, as opposed to those that simply measure short term fluctuations in vitamin D levels post fracture only, often with opposing results. Gorter *et al*^[31] who conducted an updated literature review of 75 *in vitro* and 30 *in vivo* studies found inconsistent results concerning the mechanism of action of vitamin D on fracture healing. They found only four studies that examined the effect of vitamin D deficiency on human fracture healing and that indicated no effect. No studies examined the specific benefits of supplementation alone and studies discussing the cellular effect of vitamin D in fracture healing were non-conclusive.

Because one fracture is often followed by another, and preliminary evidence strongly supports a role for 24,25(OH)D₂, a vitamin D metabolite, in mammalian fracture repair^[48], it would seem advantageous to strongly consider the use of nutritious sources, sunlight, and if not available, supplementary resources for those at greatest risk of second fractures, even if healing is not promoted. Given that a sizeable proportion of the population appears to suffer from vitamin D insufficiency^[48], and that optimal muscle function is contingent on appropriate vitamin D levels^[10-12], this alone might be helpful both in preventing future falls, and in enabling muscle forces around the fracture site to promote healing, while offering better protection of the bone while it is healing, even if the fracture site is not impacted directly. As well, the more generic benefits of vitamin D on physical wellbeing could serve to enhance activity levels that are key to building or maintaining bone mineral density, as well as preventing falls and future fractures, and fostering opportunities to be exposed to sunlight.

Cortier *et al*^[31] who specifically discussed the influence of vitamin D on bone mineralization and subsequent bone quality did not refer to the importance of vitamin D in fostering muscle function, as well as general wellbeing. Even though this group retrieved over 100 studies on this topic, the fact that they only found five *in vitro* studies performed on material from a fracture site, and only one *in vivo* study in the fracture patient, renders the role of vitamin D in this respect is very hard to discern.

Table 3 Sample of human studies designed to examine vitamin D influence on bone healing

Ref.	Type of Study	Finding
Briggs <i>et al</i> ^[10]	Examined vitamin D levels in 28 patents with diaphyseal long bone fractures at 48 h, 1 wk and 6 wk	Serum 1,25-dihydroxyvitamin D decreased from baseline, but serum 24R,25(OH) ₂ D levels did not change
Delgado-Martínez <i>et al</i> ^[22]	Investigated 25-OH-vitamin D effect in elderly with fractures	The addition of the vitamin D supplement improved strength of the fractured bone
Sun <i>et al</i> ^[25]	Examined effect of vitamin D ₃ on the differentiation of mesenchymal stem cells from a human fracture site	Vitamin D ₃ was able to modulate the the differentiation towards osteoblastic phenotype of the cells derived from fracture sites
Doetsch <i>et al</i> ^[32]	Quantified impact of vitamin D ₃ + calcium on healing of osteoporotic fracture	Bone mineral density at 6 wk was higher in actively treated group suggesting vitamin D ₃ had a positive effect 6 wk post fracture, but this was not maintained at 12 wk
Parchi <i>et al</i> ^[33]	Case report of child post-fracture	Hypovitaminosis D is a possible cause of inadequate fracture healing and refracture in children Vitamin D has a clear effect on callus formation
Boszczyk <i>et al</i> ^[36]	35 patients with inexplicable fracture healing impairments and controls were studied with regard to vitamin D	No impact of vitamin D deficiency noted
Ettehad <i>et al</i> ^[37]	Determined serum levels of vitamin D during fracture healing of 73 patients	Serum levels of vitamin D were reduced in curative period, suggesting vitamin D plays a role in the formation and mineralization of callus
Alkalay <i>et al</i> ^[44]	Assessed vitamin D metabolite levels in 28 patients after fracture, and 27 undergoing surgery	Serum 1,25-dihydroxyvitamin D ₃ was significantly reduced in the fracture cases
Tauber <i>et al</i> ^[50]	Determined active metabolites of vitamin D ₃ in 7 fracture patients	24,25(OH) ₂ D ₃ levels showed a relative decrease, and a decrease in 1,25(OH) ₂ D ₃ in 2 cases, suggesting these metabolites are consumed at fracture site during healing
Meller <i>et al</i> ^[51]	Levels of 25(OH)D ₃ + 24,25(OH) ₂ D ₃ were determined in 13 young patients with long bone fractures on admission and after 6-8 wk	Plasma 24,25(OH) ₂ -D ₃ levels rose over the 6 wk period, but no changes in 25(OH)D ₃ levels occurred
Hoikka <i>et al</i> ^[52]	Treated 37 osteoporotic fracture cases with 1 α ₁ -OHD ₃ - dosage 1 ug per day, plus 2.5 gm calcium	1 α ₁ -OHD ₃ impacts fracture healing although 5/19 cases developed hypercalcemia

Table 4 Rationale for hypothesizing vitamin D as beneficial in fracture healing

Plays an essential role in bone formation and maintenance ^[1,58]
Has positive benefits on muscle strength ^[5,58]
Is involved in calcium and bone metabolism ^[1,29,37,54,57,58,64]
Deficiency is associated with fractures ^[58]
Can modulate cell growth and neuromuscular function ^[57,65]
May influence the inflammation stage of bone healing positively, as well as the callus formation stage ^[31]
Can help regulate inflammation and bone marrow and intramuscular fat deposits ^[58]
Protects older people from osteoporosis ^[58]
Enhance fixation of implants ^[58]
Deficiency may be associated with refracture ^[33]
Deficiency is associated with non union ^[34,35,67,69]

Although very few studies were evident in the data bases reviewed, this group noted vitamin D deficiency does not seem to hinder fracture healing, while supplementation with calcium increases the extent of the fracture callus at the fracture site and promotes healing.

In other research, Briggs *et al*^[10] found decreased serum levels of 1,25-dihydroxyvitamin D in cases with diaphyseal long bone fractures but no changes in serum levels of vitamin D metabolites post fracture. However, Tauber *et al*^[50] found a relative decrease in 24,25(OH)₂D₃ levels as well as a partial decrease in 1,25(OH)₂D₃ in cases suffering from delayed non union and/or multiple fractures. Ettehad *et al*^[37] too found these metabolites were reduced during the curative period in cases with either tibial or femoral fractures. They related this finding to the possible role of vitamin D in the formation and mineralization of the callus. Suzuki *et al*^[43] found excessively low

levels of 25(OH)D to be independently and significantly associated with an increased risk of falling in the elderly. Since adequate or high levels of supplementary vitamin D are protective of bone, and many elderly with fractures are already vitamin D deficient at the time of a fall, the most common reason for fracturing a bone, it seems taking supplements as a precaution against future fractures, as well as attempting to enhance fracture healing is potentially of great importance as supported by findings of Hoikka *et al*^[52] who observed the addition of 1 alpha-OHD₃ to patients with osteoporotic hip fractures seemed to have a beneficial effect on fracture healing. Although this was also found to frequently cause hypercalcemia, and Boszczyk *et al*^[36] found no difference in vitamin D concentrations in normal and impaired bone union, and disturbances in vitamin D metabolism are unlikely to play a major role in maintenance of non-union fractures^[46],

vitamin D deficiency was present in 86% of examined patients. Inadequate levels of vitamin D were also found to prevail among patients undergoing orthopedic surgery who presented with bone healing complications^[47].

Given that hypovitaminosis D could affect bone formation adversely^[1], and that muscle strength capacity alone is found to benefit from vitamin D if taken orally^[55], and in combination with calcium may decrease the incidence of non-vertebral fractures in older persons with low vitamin D levels^[56] the sustained usage of these compounds may be more favorable than not for influencing fracture healing^[57-60], despite the negative findings of the RECORD trial^[61]. In addition, for those requiring internal fixation surgery post-fracture, the supplementation of vitamin D where this is found deficient may increase the bone-implant contact ratio and bone volume around the implant as reported by Liu *et al*^[20]. As well as fostering callus mineralization^[62], resistance of the implant is also expected to increase favorably with appropriate supplementation^[20].

However, the lack of definitive evidence precludes any conclusion or any set of useful guidelines concerning vitamin D supplementation post-fracture, where indicated, despite the magnitude of the societal burden incurred by the high prevalence of adults who experience delayed bone union, non-union, or future fractures due to suboptimal bone and muscle recovery post-fracture. Clearly, while *in vitro* models are helpful, a much greater effort in the clinical research arena appears warranted. In particular, more prospective long term follow-up studies of different vulnerable groups, and exposure to different levels and combinations of supplements appear desirable. For example, in the study by Omeroglu *et al*^[53] 116 guinea pigs who had received 50000 i.u./kg of vitamin D₃ intramuscularly benefited by this administration, suggesting this method of vitamin D delivery might be highly beneficial for accelerating the synthesis and organization of collagen fibers, the proliferation and differentiation of osteoprogenitor cells, and mineralization of the matrix. Alternately, Lidor *et al*^[11,54] found the implantation of D₃ compounds directly into the fractures accelerated healing and prevented non-union. Another mode of delivery, namely subcutaneous delivery after an experimental fracture improved fracture strength in a dose dependent manner^[22] as did vitamin D injections^[20]. Thus different modes of delivering vitamin D post fracture may produce positive, albeit differential impacts on the healing bone that might be worth investigating. Another area for research may extend to testing different vitamin D metabolites and the affinity of callus membrane receptor/binding proteins for these as observed by Kato *et al*^[42] in chick tibial fracture healing callus. Another form of study might be focused on assessing the viability of vitamin D receptors, and whether their functional status is linked to the outcomes of vitamin D analyses in the context of fracture healing, bearing in mind that vitamin D measures may not be useful for judging vitamin D in clinical studies. The consistent use of assays to examine plasma concentrations of 25-hydroxyvitamin D [25(OH)D] may

provide the best method for assessing the presence of any prevailing vitamin D deficiency^[63].

In sum, since the elderly in particular, who are highly prone to fractures, are at risk of vitamin D deficiency and insufficiency, as well as reduced exposure to sunlight, accelerated bone^[57] loss, skeletal fragility and reduced muscle power^[58], the application of post-fracture vitamin D supplements would appear beneficial^[64,65]. In particular, as outlined in Table 4, vitamin D in its different physiological forms is implicated in bone metabolism^[59], and muscle-bone interactions^[58] and potentially promotes fracture healing and mineralization^[62,66]. Consequently, identifying the optimal vitamin D level that is desirable in the post-fracture state, as well as the best mode of delivery appears highly warranted. Stressing the importance of compliance with recommendations regarding supplements, if indicated, acknowledging the importance of calcium supplementation when vitamin D levels are deficient, and applying doses of vitamin D known to have clinical efficacy is more likely than not to foster optimal post fracture bone remodeling processes and functional benefits especially for those at risk for osteoporotic fractures^[57], falls that lead to fractures^[58], fractures requiring fixation^[20] or atrophic fracture nonunion in the presence of vitamin D deficiency^[55]. As outlined by Maier *et al*^[60] about 20% of seniors receive vitamin D at the time of their fracture and after the event despite the documented 81% prevalence of vitamin D deficiency. In this regard, it appears reasonable to suggest efforts to improve vitamin D supplementation in seniors both before and after a fracture event are warranted, especially if it is confirmed that serum levels of 1,25-dihydroxyvitamin D are deficient^[61], non-union appears to prevail or is imminent^[67], or the diagnosis of a stress fracture is forthcoming^[68]. Based on findings of vitamin D deficiencies among patients with non-unions^[69], studies that show calcium and vitamin D₃ supplementation may enhance callus formation in the osteopenic or osteoporosis patient^[32], and animal models that show a combined effect of 1,25 (OH)₂D₃ on serum calcium and phosphate and bone matrix formation^[62], fracture healing rates as well as bone quality or both may be forthcoming^[70]. Alternately, it is possible, that by inadvertently delaying fracture healing, failure to provide adequate vitamin D supplementation in those suffering from vitamin D insufficiency may result in longer curative periods of inactivity and pain, thus potentially fostering further vitamin D insufficiency or depletion.

COMMENTS

Background

Fractures of the bone and their mechanisms of repair are topics that have been the subject of investigation for more than three decades. In both cases, both preventing and treating fractures and the role of vitamin D in both processes have received increasing attention in the literature due to the importance of minimizing post-fracture complications, especially among the older population.

Research frontiers

While vitamin D is of potential relevance in the healing process of fractures, it is unclear whether supplements should routinely follow fracture injuries, and if so

what is the evidence base for this.

Innovations and breakthroughs

The present narrative review examined the bulk of the literature present in key electronic data bases from 1980 onwards. The results show very few human studies have been conducted to examine if vitamin D is effective at promoting post fracture healing, and the different animal models that have been studied provide no consensus on this topic. While not new, this gap in the literature indicates much more attention is required in this realm than is currently evident.

Applications

Given that vitamin D is an essential hormone for functioning in general, those who have low levels of the hormone in general, can probably benefit from supplements in the post-fracture period if their nutritional practices do not cover recommended daily amounts, and they are at high risk for non union and/or subsequent fractures. Since those who experience non union or delayed union may inadvertently suffer from inadequate vitamin D exposure, and vitamin D insufficiency, or deficiency this approach appears worthwhile to contemplate.

Terminology

The term vitamin D in this paper refers to all forms of this hormone and/or its metabolites. The terms bone healing and fracture healing are used interchangeably.

Peer review

The review by Marks is well written.

REFERENCES

- Rodriguez WJ, Gromelski J. Vitamin d status and spine surgery outcomes. *ISRN Orthop* 2013; **2013**: 471695 [PMID: 24959360 DOI: 10.1155/2013/471695]
- Schunack W. [Vitamin D3--a prodrug of different D3-hormones]. *Med Klin (Munich)* 2006; **101** Suppl 1: 20-24 [PMID: 16826365 DOI: 10.1007/s00063-006-9004-8]
- Smith JT, Halim K, Palms DA, Okike K, Bluman EM, Chiodo CP. Prevalence of vitamin D deficiency in patients with foot and ankle injuries. *Foot Ankle Int* 2014; **35**: 8-13 [PMID: 24127268 DOI: 10.1177/1071100713509240]
- Hayes WC, Myers ER, Robinovitch SN, Van Den Kroonenberg A, Courtney AC, McMahon TA. Etiology and prevention of age-related hip fractures. *Bone* 1996; **18**: 77S-86S [PMID: 8717551 DOI: 10.1016/8756-3282(95)00383-5]
- Beudart C, Buckinx F, Rabenda V, Gillain S, Cavalier E, Slomian J, Petermans J, Reginster JY, Bruyère O. The effects of vitamin D on skeletal muscle strength, muscle mass and muscle power: a systematic review and meta-analysis of randomized controlled trials. *J Clin Endocrinol Metab* 2014; **99**: 4336-4345 [PMID: 25033068 DOI: 10.1210/jc.2014-1742]
- Shuler FD, Schlierf T, Wingate M. Preventing falls with vitamin D. *W V Med J* 2014; **110**: 10-12 [PMID: 24984399]
- Tieland M, Brouwer-Brolsma EM, Nienaber-Rousseau C, van Loon LJ, De Groot LC. Low vitamin D status is associated with reduced muscle mass and impaired physical performance in frail elderly people. *Eur J Clin Nutr* 2013; **67**: 1050-1055 [PMID: 23942175 DOI: 10.1038/ejcn.2013.144]
- Schindeler A, McDonald MM, Bokko P, Little DG. Bone remodeling during fracture repair: The cellular picture. *Semin Cell Dev Biol* 2008; **19**: 459-466 [PMID: 18692584 DOI: 10.1016/j.semcdb.2008.07.004]
- Eschle D, Aeschlimann AG. Is supplementation of vitamin d beneficial for fracture healing? A short review of the literature. *Geriatr Orthop Surg Rehabil* 2011; **2**: 90-93 [PMID: 23569676 DOI: 10.1177/2151458511408568]
- Briggs AD, Kuan V, Greiller CL, Maclaughlin BD, Ramachandran M, Harris T, Timms PM, Venton TR, Vieth R, Norman AW, Griffiths CJ, Martineau AR. Longitudinal study of vitamin D metabolites after long bone fracture. *J Bone Miner Res* 2013; **28**: 1301-1307 [PMID: 23281057 DOI: 10.1002/jbmr.1855]
- Lidor C, Dekel S, Hallel T, Edelstein S. Levels of active metabolites of vitamin D3 in the callus of fracture repair in chicks. *J Bone Joint Surg Br* 1987; **69**: 132-136 [PMID: 3029136]
- Lidor C, Dekel S, Edelstein S. The metabolism of vitamin D3 during fracture healing in chicks. *Endocrinology* 1987; **120**: 389-393 [PMID: 3023034 DOI: 10.1210/endo-120-1-389]
- St-Arnaud R. CYP24A1-deficient mice as a tool to uncover a biological activity for vitamin D metabolites hydroxylated at position 24. *J Steroid Biochem Mol Biol* 2010; **121**: 254-256 [PMID: 20144713 DOI: 10.1016/j.jsbmb.2010.02.002]
- Blahos J, Babický A, Porsová I, Kolár J. Effect of 1,25-dihydroxycholecalciferol on fracture healing and on general posttraumatic skeletal response in rats. *Endocrinol Exp* 1989; **23**: 287-294 [PMID: 2620660]
- Omeroğlu H, Ateş Y, Akkuş O, Korkusuz F, Biçimoğlu A, Akkaş N. Biomechanical analysis of the effects of single high-dose vitamin D3 on fracture healing in a healthy rabbit model. *Arch Orthop Trauma Surg* 1997; **116**: 271-274 [PMID: 9177802 DOI: 10.1007/BF00390051]
- Melhus G, Solberg LB, Dimmen S, Madsen JE, Nordsletten L, Reinholt FP. Experimental osteoporosis induced by ovariectomy and vitamin D deficiency does not markedly affect fracture healing in rats. *Acta Orthop* 2007; **78**: 393-403 [PMID: 17611855 DOI: 10.1080/17453670710013988]
- Mao L, Tamura Y, Kawao N, Okada K, Yano M, Okumoto K, Kaji H. Influence of diabetic state and vitamin D deficiency on bone repair in female mice. *Bone* 2014; **61**: 102-108 [PMID: 24378215 DOI: 10.1016/j.bone.2013.12.024]
- Hong HH, Chou TA, Yang JC, Chang CJ. The potential effects of cholecalciferol on bone regeneration in dogs. *Clin Oral Implants Res* 2012; **23**: 1187-1192 [PMID: 22092360 DOI: 10.1111/j.1600-0501.2011.02284.x]
- Fu L, Tang T, Miao Y, Hao Y, Dai K. Effect of 1,25-dihydroxy vitamin D3 on fracture healing and bone remodeling in ovariectomized rat femora. *Bone* 2009; **44**: 893-898 [PMID: 19442605 DOI: 10.1016/j.bone.2009.01.378]
- Liu W, Zhang S, Zhao D, Zou H, Sun N, Liang X, Dard M, Lanske B, Yuan Q. Vitamin D supplementation enhances the fixation of titanium implants in chronic kidney disease mice. *PLoS One* 2014; **9**: e95689 [PMID: 24752599 DOI: 10.1371/journal.pone.0095689]
- Gigante A, Torcianti M, Boldrini E, Manzotti S, Falcone G, Greco F, Mattioli-Belmonte M. Vitamin K and D association stimulates in vitro osteoblast differentiation of fracture site derived human mesenchymal stem cells. *J Biol Regul Homeost Agents* 2008; **22**: 35-44 [PMID: 18394316]
- Delgado-Martínez AD, Martínez ME, Carrascal MT, Rodríguez-Avial M, Munuera L. Effect of 25-OH-vitamin D on fracture healing in elderly rats. *J Orthop Res* 1998; **16**: 650-653 [PMID: 9877387 DOI: 10.1002/jor.1100160604]
- Steier A, Gedalia I, Schwarz A, Rodan A. Effect of vitamin D2 and fluoride on experimental bone fracture healing in rats. *J Dent Res* 1967; **46**: 675-680 [PMID: 5298503 DOI: 10.1177/00220345670460040801]
- Kato A, Seo EG, Einhorn TA, Bishop JE, Norman AW. Studies on 24R,25-dihydroxyvitamin D3: evidence for a nonnuclear membrane receptor in the chick tibial fracture-healing callus. *Bone* 1998; **23**: 141-146 [PMID: 9701473 DOI: 10.1016/S8756-3282(98)00085-4]
- Sun JS, Chen PY, Tsuang YH, Chen MH, Chen PQ. Vitamin-D binding protein does not enhance healing in rat bone defects: a pilot study. *Clin Orthop Relat Res* 2009; **467**: 3156-3164 [PMID: 19418105]
- Lindgren JU, DeLuca HF, Mazess RB. Effects of 1,25(OH)2D3 on bone tissue in the rabbit: studies on fracture healing, disuse osteoporosis, and prednisone osteoporosis. *Calcif Tissue Int* 1984; **36**: 591-595 [PMID: 6441632 DOI: 10.1007/BF02405372]
- Andreen O, Larsson SE. Effects of parathyroidectomy and vitamin D on fracture healing. Fracture biomechanics in rats after parathyroidectomy and treatment with 1,25-dihydroxycholecalciferol. *Acta Orthop Scand* 1983; **54**: 805-809

- [PMID: 6689464 DOI: 10.3109/17453678308992913]
- 28 **Jingushi S**, Iwaki A, Higuchi O, Azuma Y, Ohta T, Shida JL, Izumi T, Ikenoue T, Sugioka Y, Iwamoto Y. Serum 1 α ,25-dihydroxyvitamin D₃ accumulates into the fracture callus during rat femoral fracture healing. *Endocrinology* 1998; **139**: 1467-1473 [PMID: 9528922 DOI: 10.1210/endo.139.4.5883]
 - 29 **Seo EG**, Einhorn TA, Norman AW. 24R,25-dihydroxyvitamin D₃: an essential vitamin D₃ metabolite for both normal bone integrity and healing of tibial fracture in chicks. *Endocrinology* 1997; **138**: 3864-3872 [PMID: 9275076 DOI: 10.1210/endo.138.9.5398]
 - 30 **Dekel S**, Salama R, Edelstein S. The effect of vitamin D and its metabolites on fracture repair in chicks. *Clin Sci (Lond)* 1983; **65**: 429-436 [PMID: 6309464]
 - 31 **Gorter EA**, Hamdy NA, Appelman-Dijkstra NM, Schipper IB. The role of vitamin D in human fracture healing: a systematic review of the literature. *Bone* 2014; **64**: 288-297 [PMID: 24792958 DOI: 10.1016/j.bone.2014.04.026]
 - 32 **Doetsch AM**, Faber J, Lynnerup N, Wätjen I, Bliddal H, Danneskiold-Samsøe B. The effect of calcium and vitamin D₃ supplementation on the healing of the proximal humerus fracture: a randomized placebo-controlled study. *Calcif Tissue Int* 2004; **75**: 183-188 [PMID: 15386160 DOI: 10.1007/s00223-004-0167-0]
 - 33 **Parchi P**, Andreani L, Piolanti N, Niccolai F, Cervi V, Lisanti M. Effect of vitamin D in fracture healing in a child: case report. *Arch Osteoporos* 2014; **9**: 170 [PMID: 24452512 DOI: 10.1007/s11657-013-0170-z]
 - 34 **Pourfeizi HH**, Tabriz A, Elmi A, Aslani H. Prevalence of vitamin D deficiency and secondary hyperparathyroidism in nonunion of traumatic fractures. *Acta Med Iran* 2013; **51**: 705-710 [PMID: 24338144]
 - 35 **Van Demark RE**, Allard B, Van Demark RE. Nonunion of a distal tibial stress fracture associated with vitamin D deficiency: a case report. *S D Med* 2010; **63**: 87-91, 93 [PMID: 20301871]
 - 36 **Boszczyk AM**, Zakrzewski P, Pomianowski S. Vitamin D concentration in patients with normal and impaired bone union. *Pol Orthop Traumatol* 2013; **78**: 1-3 [PMID: 23306314]
 - 37 **Ettehad H**, Mirbolook A, Mohammadi F, Mousavi M, Ebrahimi H, Shirangi A. Changes in the serum level of vitamin d during healing of tibial and femoral shaft fractures. *Trauma Mon* 2014; **19**: e10946 [PMID: 24719823 DOI: 10.5812/traumamon.10946]
 - 38 **Wölfel C**, Englert S, Moghaddam AA, Zimmermann G, Schmidt-Gayk H, Höner B, Hogan A, Lehnhardt M, Grütznert PA, Kolios L. Time course of 25(OH)D₃ vitamin D₃ as well as PTH (parathyroid hormone) during fracture healing of patients with normal and low bone mineral density (BMD). *BMC Musculoskelet Disord* 2013; **14**: 6 [PMID: 23286544 DOI: 10.1186/1471-2474-14-16]
 - 39 **Gomberg SJ**, Wustrack RL, Napoli N, Arnaud CD, Black DM. Teriparatide, vitamin D, and calcium healed bilateral subtrochanteric stress fractures in a postmenopausal woman with a 13-year history of continuous alendronate therapy. *J Clin Endocrinol Metab* 2011; **96**: 1627-1632 [PMID: 21430030 DOI: 10.1210/jc.2010-2520]
 - 40 **Inklebarger J**, Griffin M, Taylor MJ, Dembry RB. Femoral and tibial stress fractures associated with vitamin D insufficiency. *J R Army Med Corps* 2014; **160**: 61-63 [PMID: 24109098 DOI: 10.1136/jramc-2013-000085]
 - 41 **Bee CR**, Sheerin DV, Wuest TK, Fitzpatrick DC. Serum vitamin D levels in orthopaedic trauma patients living in the northwestern United States. *J Orthop Trauma* 2013; **27**: e103-e106 [PMID: 22576645 DOI: 10.1097/BOT.0b013e31825cf8fb]
 - 42 **Kato A**, Bishop JE, Norman AW. Evidence for a 1 α ,25-dihydroxyvitamin D₃ receptor/binding protein in a membrane fraction isolated from a chick tibial fracture-healing callus. *Biochem Biophys Res Commun* 1998; **244**: 724-727 [PMID: 9535732 DOI: 10.1006/bbrc.1998.8318]
 - 43 **Suzuki T**. [Frontiers in vitamin D; basic research and clinical application. Vitamin D and falls]. *Clin Calcium* 2011; **21**: 71-79 [PMID: 22040823]
 - 44 **Alkalay D**, Shany S, Dekel S. Serum and bone vitamin D metabolites in elective patients and patients after fracture. *J Bone Joint Surg Br* 1989; **71**: 85-87 [PMID: 2783695]
 - 45 **Brumbaugh PF**, Speer DP, Pitt MJ. 1 α , 25-Dihydroxyvitamin D₃ a metabolite of vitamin D that promotes bone repair. *Am J Pathol* 1982; **106**: 171-179 [PMID: 6895976]
 - 46 **Haining SA**, Atkins RM, Guillard-Cumming DF, Sharrard WJ, Russell RG, Kanis JA. Vitamin D metabolites in patients with established non-union of fracture. *Bone Miner* 1986; **1**: 205-209 [PMID: 3509893]
 - 47 **Inklebarger J**, Taylor MJ, Griffin M, Clarke T. Fixation failure in an isolated tibial eminence ACL traction avulsion fracture in a paratrooper: is there an association with vitamin D deficiency? *J Surg Case Rep* 2014; **2014** [PMID: 24876463 DOI: 10.1093/jscr/rju029]
 - 48 **St-Arnaud R**, Naja RP. Vitamin D metabolism, cartilage and bone fracture repair. *Mol Cell Endocrinol* 2011; **347**: 48-54 [PMID: 21664253 DOI: 10.1016/j.mce.2011.05.018]
 - 49 **Lindgren JU**, Narechania RG, McBeath AA, Lange TA, DeLuca HF. Effects of 1,24 dihydroxyvitamin D₃ and calcitonin on fracture healing in adult rats. *Clin Orthop Relat Res* 1981; **160**: 304-308 [PMID: 6269785]
 - 50 **Tauber C**, Noff D, Noff M, Malkin C. Blood levels of active metabolites of vitamin D₃ in fracture repair in humans. A preliminary report. *Arch Orthop Trauma Surg* 1990; **109**: 265-267 [PMID: 2271359 DOI: 10.1007/BF00419941]
 - 51 **Meller Y**, Shainkin-Kestenbaum R, Shany S, Zuilli I, Yankowitz N, Giat J, Konforti A, Torok G. Parathyroid hormone, calcitonin, and vitamin D metabolites during normal fracture healing in humans. A preliminary report. *Clin Orthop Relat Res* 1984; **183**: 238-245 [PMID: 6697591]
 - 52 **Hoikka V**, Alhava EM, Aro A, Karjalainen P, Rehnberg V. Treatment of osteoporosis with 1-alpha-hydroxycholecalciferol and calcium. *Acta Med Scand* 1980; **207**: 221-224 [PMID: 6989171 DOI: 10.1111/j.0954-6820.1980.tb09709.x]
 - 53 **Omeroglu S**, Erdogan D, Omeroglu H. Effects of single high-dose vitamin D₃ on fracture healing. An ultrastructural study in healthy guinea pigs. *Arch Orthop Trauma Surg* 1997; **116**: 37-40 [PMID: 9006763]
 - 54 **Lidor C**, Dekel S, Meyer MS, Blaugrund E, Hallel T, Edelstein S. Biochemical and biomechanical properties of avian callus after local administration of dihydroxylated vitamin D metabolites. *J Bone Joint Surg Br* 1990; **72**: 137-140 [PMID: 2298772]
 - 55 **Osório J**. Bone: vitamin D metabolites and fracture healing. *Nat Rev Endocrinol* 2013; **9**: 130 [PMID: 23358363 DOI: 10.1038/nrendo.2013.9]
 - 56 **Lips P**, Gielen E, van Schoor NM. Vitamin D supplements with or without calcium to prevent fractures. *Bonekey Rep* 2014; **3**: 512 [PMID: 24818004 DOI: 10.1038/bonekey.2014.7]
 - 57 **Lips P**, Bouillon R, van Schoor NM, Vanderschueren D, Verschueren S, Kuchuk N, Milisen K, Boonen S. Reducing fracture risk with calcium and vitamin D. *Clin Endocrinol (Oxf)* 2010; **73**: 277-285 [PMID: 20796001 DOI: 10.1111/j.1365-2265.2009.03701.x]
 - 58 **Sanders KM**, Scott D, Ebeling PR. Vitamin D deficiency and its role in muscle-bone interactions in the elderly. *Curr Osteoporos Rep* 2014; **12**: 74-81 [PMID: 24488588 DOI: 10.1007/s11914-014-0193-4]
 - 59 **Sintov AC**, Yarmolinsky L, Dahan A, Ben-Shabat S. Pharmacological effects of vitamin D and its analogs: recent developments. *Drug Discov Today* 2014 Jun 16; Epub ahead of print [PMID: 24947685 DOI: 10.1016/j.drudis.2014.06.008]
 - 60 **Maier S**, Sidelnikov E, Dawson-Hughes B, Egli A, Theiler R, Platz A, Staehelin HB, Simmen HP, Meier C, Dick W, Grob D, von Eckardstein A, Bischoff-Ferrari HA. Before and after hip fracture, vitamin D deficiency may not be treated sufficiently. *Osteoporos Int* 2013; **24**: 2765-2773 [PMID: 23716038]

- 61 **Grant AM**, Avenell A, Campbell MK, McDonald AM, MacLennan GS, McPherson GC, Anderson FH, Cooper C, Francis RM, Donaldson C, Gillespie WJ, Robinson CM, Torgerson DJ, Wallace WA. Oral vitamin D3 and calcium for secondary prevention of low-trauma fractures in elderly people (Randomised Evaluation of Calcium Or vitamin D, RECORD): a randomised placebo-controlled trial. *Lancet* 2005; **365**: 1621-1628 [PMID: 15885294]
- 62 **Andreen O**, Larsson SE. Effects of 1,25-dihydroxycholecalciferol on fracture healing. Calcium, phosphate, and zinc in callus and serum. *Arch Orthop Trauma Surg* 1984; **103**: 257-262 [PMID: 6548905 DOI: 10.1007/BF00387331]
- 63 **Schmidt-Gayk H**, Bouillon R, Roth HJ. Measurement of vitamin D and its metabolites (calcidiol and calcitriol) and their clinical significance. *Scand J Clin Lab Invest Suppl* 1997; **227**: 35-45 [PMID: 9127467 DOI: 10.1080/00365519709168307]
- 64 **Ceglia L**, Harris SS. Vitamin D and its role in skeletal muscle. *Calcif Tissue Int* 2013; **92**: 151-162 [PMID: 22968766 DOI: 10.1007/s00223-012-9645-y]
- 65 Vitamin D. Fact Sheet for Professionals. 2014. Available from: URL: [http://ods.od.nih.gov/factsheets/VitaminD-HealthProfessional/associated with non union](http://ods.od.nih.gov/factsheets/VitaminD-HealthProfessional/associated%20with%20non%20union)
- 66 **Nampei A**, Hashimoto J. [Bone fracture and the healing mechanisms. Metabolic bone disease and skeletal healing]. *Clin Calcium* 2009; **19**: 648-652 [PMID: 19398831]
- 67 **Hobby B**, Lee MA. Managing atrophic nonunion in the geriatric population: incidence, distribution, and causes. *Orthop Clin North Am* 2013; **44**: 251-256 [PMID: 23544828 DOI: 10.1016/j.jocl.2013.01.011]
- 68 **McCabe MP**, Smyth MP, Richardson DR. Current concept review: vitamin D and stress fractures. *Foot Ankle Int* 2012; **33**: 526-533 [PMID: 22735329]
- 69 **Brinker MR**, O'Connor DP, Monla YT, Earthman TP. Metabolic and endocrine abnormalities in patients with non-unions. *J Orthop Trauma* 2007; **21**: 557-570 [PMID: 17805023 DOI: 10.1097/BOT.0b013e31814d4dc6]
- 70 **Carpintero P**, Caeiro JR, Carpintero R, Morales A, Silva S, Mesa M. Complications of hip fractures: A review. *World J Orthop* 2014; **5**: 402-411 [PMID: 25232517 DOI: 10.5312/wjo.v5.i4.402]

P- Reviewer: Makishima M, Peng JB S- Editor: Ji FF
L- Editor: A E- Editor: Lu YJ



Association of serum bilirubin and non-alcoholic fatty liver disease: A feasible therapeutic avenue?

Mohamed S Anwar, John F Dillon, Michael H Miller

Mohamed S Anwar, John F Dillon, Michael H Miller, Gut Group, Medical Research Institute, Ninewells Hospital and Medical School, University of Dundee, Dundee DD1 9SY, United Kingdom

Author contributions: Anwar MS, Dillon JF and Miller MH discussed and conceived the idea of this systematic review; Anwar MS and Miller MH performed the literature search and collated the data; all three authors wrote the manuscript.

Correspondence to: Dr. Mohamed S Anwar, Gut Group, Medical Research Institute, Ninewells Hospital and Medical School, University of Dundee, Nethergate, Dundee DD1 9SY, United Kingdom. mohamed-anwar@doctors.org.uk

Telephone: +44-1382-632334 Fax: +44-1382-632098

Received: May 21, 2014 Revised: October 15, 2014

Accepted: October 23, 2014

Published online: December 9, 2014

Abstract

AIM: To look at the current strength of evidence and the potential application of anti-oxidants in this setting.

METHODS: Two electronic databases (PubMed and Web of Knowledge) were searched to January 2013 to find studies addressing serum bilirubin levels in non-alcoholic fatty liver disease (NAFLD). The search used key word combinations in relation to NAFLD and serum bilirubin specific to human adults only. After screening selected studies were reviewed in depth by two independent reviewers. Data synthesis with further meta-analysis was planned but not possible due to the heterogeneity of the outcome measures in these studies.

RESULTS: Out of 416 studies screened only seven studies were considered suitable for inclusion. All seven studies consistently reported an inverse association of bilirubin with NAFLD despite the heterogeneous sample of studies. Only two studies were prospective. No negative studies were found.

CONCLUSION: Most studies suggest a correlation

between high bilirubin levels of any type are inversely correlated with NAFLD. But to date most of these studies have been poorly designed to allow meaningful conclusions, except one cohort study. There is a need for a large prospective cohort study in multiple populations to test this hypothesis fully before mechanistic associations can be established and therapeutic options of the apparent anti-oxidant effect of bilirubin be explored in NAFLD. Furthermore these studies should include analysis of UGT1A1 gene to expound upon underlying cause of unconjugated hyperbilirubinaemia.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

Key words: Review; Systematic; Bilirubin; Hyperbilirubinemia; Anti-oxidants; Adult; Non-alcoholic fatty liver disease

Core tip: This systematic review summarises and highlights the deficiencies in the current studies on the association of serum bilirubin with non-alcoholic fatty liver disease (NAFLD). It explores the potential underpinning of the mechanistic association of NAFLD with bilirubin. Potential novel therapeutic avenues of bilirubin are explored in NAFLD, a common condition with oxidative damage as a core pathogenetic factor. Although this area of study is still in its infancy, this review is a timely summary of current key studies in this subject area and provides an up to date thought perspective with focus on future direction and potential therapeutic application.

Anwar MS, Dillon JF, Miller MH. Association of serum bilirubin and non-alcoholic fatty liver disease: A feasible therapeutic avenue? *World J Pharmacol* 2014; 3(4): 209-216 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v3/i4/209.htm> DOI: <http://dx.doi.org/10.5497/wjp.v3.i4.209>

INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) has been

recognised as the most prevalent liver disease, with current estimations that it affects around 20%-30% of the general population in the western world^[1,2]. NAFLD is considered to be the hepatic manifestation of the metabolic syndrome as it is closely related to insulin resistance, obesity, hypercholesterolemia, type 2 diabetes, and coronary artery disease^[3-5]. NAFLD is composed of a histological spectrum of hepatic dysfunctions ranging from simple steatosis (SS) to non-alcoholic steatohepatitis (NASH), cirrhosis and hepatocellular carcinoma^[6]. The pathogenic processes underpinning NAFLD remain unclear, although oxidative stress, fat transportation and inflammation are implicated. Furthermore oxidative stress has been suggested as an aetiopathogenic mechanism in NAFLD^[7,8]. Additionally, mounting evidence suggests a link between serum ferritin, insulin resistance, and NAFLD^[9,10]. Excessive hepatic iron accumulation in NAFLD is likely one of the potential cofactors involved in the enhanced oxidative stress, which triggers liver cell necrosis and activation of hepatic stellate cells, both leading to fibrosis^[11]. The significance of the association serum ferritin in NAFLD may relate to heme catabolism and the anti-oxidant state of play, which will be discussed later. Studies involving administration of a free radical-generating azo compound to mice or rats induced fat accumulation in the liver by increasing triacylglycerol and decreasing phospholipids. Likewise fat accumulation in the liver was suppressed through the simultaneous administration of free radical-scavenging antioxidants such as Vitamin E, Therefore antioxidant agents have been proposed as an effective treatment^[12-15].

Bilirubin, the end product of heme catabolism, is known to be a potent physiological antioxidant cytoprotectant due to its inhibitory effect on the activity of NAD(P)H oxidase. In addition it scavenges peroxy radicals, hydroxyl radicals, and reactive nitrogen species preventing oxidation of intracellular lipids^[16-20]. Bilirubin has also been proposed as having an anti-inflammatory role and has major anti-fibrogenic properties *via* heme oxygenase-1 (HO-1)^[21-23]. Previous studies have shown that unconjugated hyperbilirubinemia is inversely associated with ischaemic heart disease, carotid stenosis, insulin resistance, diabetes, vascular complication of diabetes, peripheral vascular disease and even cancer^[24-29]. Furthermore there is strong clinical evidence for the beneficial cytoprotective effects of unconjugated bilirubin as observed in Gilbert's syndrome^[28-29].

Consequently, it can be hypothesised that elevated serum bilirubin levels reduce oxidative stress, decrease fibrosis and inflammation, and decrease the risk of NAFLD development and progression. If this hypothesis is confirmed then therapeutic options of inducing "iatrogenic" Gilbert's syndrome would be a key area of research. This systematic review evaluates the studies carried out to date to assess the reported association between bilirubin and NAFLD.

MATERIALS AND METHODS

Data sources

This systematic review included studies published in

electronic databases over the time period ranging from their inception to January 2013. We searched two main stream public-domain data bases, PubMed and Web of Knowledge. Three categories were devised (1) conditions (SS, NASH, NAFLD, FLD); (2) bilirubin (unconjugated hyperbilirubinemia, bilirubin, anti-oxidant, protective marker); and (3) subjects (human). Each possible combination was searched in the above two databases, also the bibliographies of relevant systematic reviews were manually searched.

Study selection

We included all types of studies, which investigated the relationship between bilirubin and NAFLD. Paediatric studies were excluded due to the potential alternate pathophysiology in this category. Outcome measure for each study was bilirubin, but it should be noted there are three methods of reporting bilirubin. Total bilirubin which consists of direct (conjugated) and indirect (unconjugated) bilirubin. Each study focused on one of these during statistical analysis.

Statistical analysis

Each selected study was assessed independently by two reviewers for methodology, outcome measures, results, limitations, risks of bias. Data synthesis with further meta-analysis was not possible due to the heterogeneity of outcome measures, study designs and statistical analysis in each study. Therefore instead we have provided a summary of this for each study.

RESULTS

Study selection, characteristics and analysis

We considered 416 potentially relevant articles, after screening the abstracts and titles, 407 studies were excluded, (Figure 1). Nine articles were fully evaluated, with a further two excluded. First of these articles was an editorial rather than an original study^[30] and the second article was not relevant, it discussed measures of oxidation stress in NASH^[31]. Of the included studies, three were cross sectional, two retrospective and two prospective. Two of these studies had a large sample size and were conducted in South Korea^[32,33]. The remaining five small sized studies used liver biopsy^[34-38] to diagnose NAFLD but notably this does not include every patient in the study. Also only two studies^[37,38] specifically states blinding of the pathologist to biochemical results and intention of study. All studies excluded patients using alcohol > 20 g/d, screened for viral hepatitis, alternative liver pathology and haemolysis, except the Tarantino^[38] study which did not assess patient for possible haemolysis. The characteristics of each study are shown in Table 1. Due to the heterogeneity of these studies, further pooled analysis could not be carried out and therefore a summary of key results is provided instead in Table 2. Notably no study reported an insignificant association of bilirubin with NAFLD.

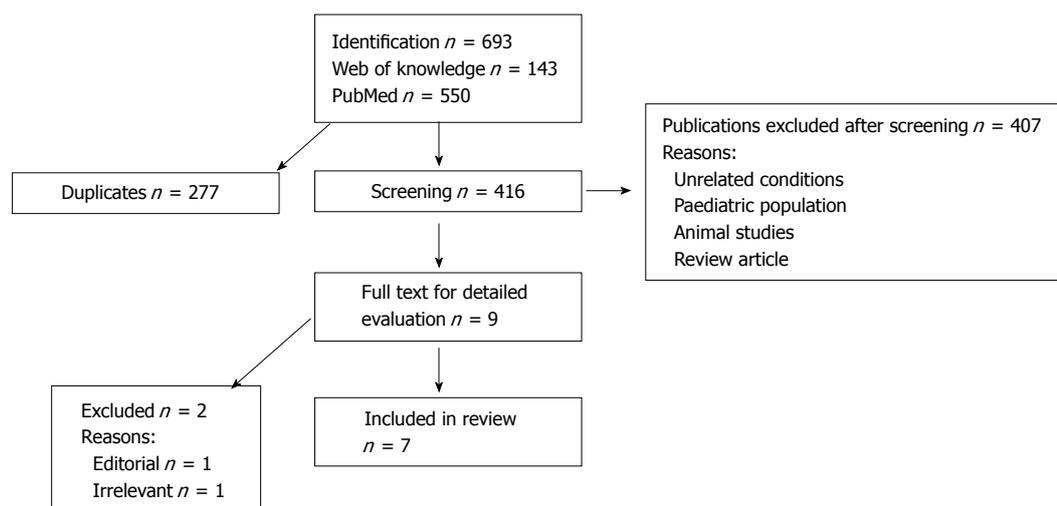


Figure 1 Flow chart describing the selection process for publication to be included in this review.

Of these studies, two were prospective^[32,33] and one^[32] of these was a large cohort study of young Korean men (5900) and showed all types of raised bilirubin were inversely associated with developing NAFLD but focused on conjugated hyperbilirubinaemia. In this study multivariate model analysis showed only conjugated hyperbilirubinaemia as independently association with risk of developing NAFLD. The study adjusted for confounding factors such as age, body mass index, current smoking, alcohol intake, exercise, diabetes mellitus, history of cardiovascular disease and history of malignancy, high density lipoprotein cholesterol, triglycerides, glucose, insulin, and uric acid.

The second^[33] large sample study was also carried out in South Korea and showed an inverse association between total bilirubin and NAFLD. But due to the cross-sectional design of this study a causal relationship cannot be confirmed. These large sample studies confirmed NAFLD on the basis of typical ultrasound findings instead of a liver biopsy. This limitation is essentially unavoidable in such a large sample size given the morbidity and mortality associated with this procedure.

The smaller studies which were based at tertiary centres did carry out liver biopsies albeit not for every patient. Duseja^[34] carried out the first study looking into association of hyperbilirubinaemia and NASH. This was a prospective study consisting of only 67 subjects and therefore did not show a statistically significant association between hyperbilirubinaemia and NASH. Given the small sample size of this study, it cannot be considered sufficient to have shown negative or positive correlation between hyperbilirubinemia and NASH. Furthermore the setting and criterion for patient selection is not adequately defined.

Hjelkrem *et al.*^[35] and Kumar *et al.*^[36] suggested that degree of fibrosis also appears to be related to bilirubin levels alongside NAFLD development and progression, although neither study blinded the pathologist to study intent. Interestingly although 508 patients had liver biopsy

in the Hjelkrem *et al.*^[35] study including only thirty five who had elevated unconjugated bilirubin, their statistical analysis did not suggest any association between NAFLD severity and unconjugated bilirubin but this is probably due to small sample of elevated unconjugated bilirubin patients in this group. Kumar *et al.*^[36] reported a negative correlation between serum unconjugated levels and histopathological NAS score, stages of fibrosis on the basis of biopsy results from 42 patients in total. Given the sample size and study design of these two studies, these results can only be regarded as speculative at this stage.

Chisholm *et al.*^[37] carried out a study in bariatric patients prior to surgery for evaluation of markers predictive of steatohepatitis, they devised a ROC curve for prediction curve of NASH which included 3 variables: total bilirubin, ALT and HOMA-IR (Homeostasis model of assessment-insulin resistance). Of 370 patients in this study 275 had a liver biopsy, with the pathologist blinded to the intent of the study but a cohort of patients were also given very low calorie diet to study the influence of this on histology findings. Unfortunately insufficient data is given to infer any association between degree of fibrosis and bilirubin level.

Tarantino *et al.*^[38] study analysed hepatocytes after exposing them to free fatty acids with expected mitochondrial damage for the presence of anti-oxidant substances such as cytochrome c, gamma-glutamyl transferase, triglycerides and unconjugated bilirubin in patients with NAFLD of differing severity ($n = 186$) and controls ($n = 27$). The study showed unconjugated bilirubin to be elevated in all spectra of NAFLD, except in healthy controls ($P = 0.008$). The study authors felt the elevated unconjugated bilirubin represented anti-oxidant effect in this setting and unconjugated bilirubin could be used to monitor response to disease modifying treatment. Interestingly the level of unconjugated bilirubin was the highest in NASH subgroup ($n = 44$) with the most severe histological findings, contradicting all previous studies. All previous studies had suggested unconjugated bilirubin was protective

Table 1 Summary of characteristics of studies on the association of bilirubin with non-alcoholic fatty liver disease

Ref.	Study design Total number of patients	Study population	Main outcome measure
Chang <i>et al</i> ^[32]	Cohort-longitudinal n = 5900	Korean men from a single large semiconductor company (South Korea)	Conjugated bilirubin
Kwak <i>et al</i> ^[33]	Cross-section n = 17348	Health check-general Population (South Korea)	Total bilirubin
Duseja <i>et al</i> ^[34]	Prospective n = 67	Hospital setting unclear, probably Tertiary (India)	Unconjugated bilirubin
Hjelkrem <i>et al</i> ^[35]	Retrospective n = 641	Tertiary hospital patients undergoing liver biopsy (United States)	Unconjugated bilirubin
Kumar <i>et al</i> ^[36]	Cross-sectional n = 204	Tertiary hospital, outpatient NAFLD clinic (India)	Unconjugated bilirubin
Chisholm <i>et al</i> ^[37]	Cross-sectional n = 370	Liver biopsy prior to bariatric surgery (United States)	Total bilirubin
Tarantino <i>et al</i> ^[38]	Cross-sectional n = 186	Tertiary hospital, outpatient likely liver clinic (Italy)	Unconjugated bilirubin

NAFLD: Non-alcoholic fatty liver disease.

Table 2 Results and analysis summary of the studies on the association of bilirubin with non-alcoholic fatty liver disease

Ref.	Study analysis	Results
Chang <i>et al</i> ^[32]	(+) Large sample (+) Prospective (-) NAFLD on US	Hazard ratio for NAFLD comparing the highest to the lower quartile of serum conjugated bilirubin = 0.61 (95%CI: 0.54-0.68), after adjusting metabolic parameters 0.86 ^a (95%CI : 0.76-0.98)
Kwak <i>et al</i> ^[33]	(+) Large sample (-) NAFLD on US	Total bilirubin inversely associated with NAFLD OR = 0.88, 95%CI: 0.80-0.97. An inverse, dose-dependent association between NAFLD and serum total bilirubin levels OR = 0.80 ^b , 95%CI: 0.71-0.90 in the fourth quartile <i>vs</i> lowest quartile
Duseja <i>et al</i> ^[34]	(-) Small sample (+) Prospective	Patient sample too small to make any meaningful statistical inference
Hjelkrem <i>et al</i> ^[35]	(+) NASH on biopsy (selected) (-) Retrospective design (-) Small sample (+) NAFLD on biopsy	Unconjugated hyperbilirubinaemia inversely associated with NASH OR = 16.1 ^b , 95%CI: 3.7-70.8
Kumar <i>et al</i> ^[36]	(-) Small sample (+/-) NAFLD on biopsy but not all patients	Unconjugated hyperbilirubinaemia (UCHB) and NAS score-negative correlation: r: -0.48 ^b . UCHB and stage of fibrosis: r: -0.28 ^d
Chisholm <i>et al</i> ^[37]	(-) Small sample (+) NAFLD on biopsy	Binary logistic regression showed independent association of total bilirubin with NASH ^c P = 0.016
Tarantino <i>et al</i> ^[38]	(-) Retrospective (-) Small sample (+) NASH on biopsy (selected)	Unconjugated bilirubin elevated in all spectra of NAFLD, except healthy control ^e P = 0.008 High UCHB in advanced NASH group

^aP < 0.039; ^bP < 0.001; ^dP = 0.007; ^cP = 0.016; ^eP = 0.008. (+) favourable design; (-) unfavourable design; OR: odds ration; US: ultrasound; r: relative coefficient; P: Probability; NAFLD: Non-alcoholic fatty liver disease; NASH: Non-alcoholic steatohepatitis.

for NASH and less commonly elevated in the advanced cohort of NASH. It is worth noting that the study did not demonstrate cytochrome c to be elevated as expected which contradicts multiple previous studies and notably did not check for underlying haemolysis in patient cohort either therefore a cautious approach should be taken with all results of this study.

DISCUSSION

Despite the heterogeneous nature of studies addressing the association of bilirubin with NAFLD including large variation in sample size and specific bilirubin type (s) measured there was consistent inverse association

of raised total bilirubin, conjugated and unconjugated hyperbilirubinemia with NAFLD, across populations in Asia and America. Although Tarantino *et al*^[38] data contradicted these findings, the poor study design and failure to demonstrate other expected findings make result interpretation from this study highly speculative. But given the sample size and study design variation of the remaining studies, these results can only be regarded as showing a tentative association with disease reduction but combined with a plausible biological mechanism, they raise the intriguing possibility of a causative relationship.

This review is limited due to the small number of studies on this topic and the heterogeneity of these studies. Given this pooled analysis could not be carried out.

Despite the shortcoming of the studies in this review, there is a consistent reported inverse association between high bilirubin and NAFLD, except in one study, which had very small sample size, and therefore no statistical inference can be attained from this study. Notably to date most studies have suggested undiagnosed Gilbert's syndrome as the cause of unconjugated hyperbilirubinemia although none of the studies have validated this with analysis of UGT1A1 genetic mutation. Although Gilbert's syndrome may account for some of these patients, another possibility is that unconjugated hyperbilirubinemia may be an initial acquired response to oxidative stress and thus represents the liver's intrinsic anti-oxidative capacity, but is not sustainable due to repeated insults. Notably Gilbert's syndrome is more commonly diagnosed in men and sex steroids may influence bilirubin metabolism with higher production in men^[39]. Two studies included in this review^[32,36] showed higher preponderance of men with unconjugated hyperbilirubinemia. Therefore future studies are required to verify the inverse correlation of bilirubin to NAFLD in large, prospective cohort study in multiple populations along with the analysis of *UGT1A1* gene to expound upon underlying cause of unconjugated hyperbilirubinemia in this cohort.

Although a direct effect of bilirubin maybe likely, other parts of the heme catabolism such as the effect on iron homeostasis may also be relevant. Heme catabolism represents a key function in mobilising macrophage iron derived from ingested erythrocytes. Importantly, the storage and processing of iron from erythrophagocytosis by macrophages within plaque appear to play a vital role in plaque progression^[40]. Accordingly, it has been demonstrated that erythrocytes induce plaque vulnerability in a dose-dependent manner in a rabbit model of intra-plaque haemorrhage^[41]. Furthermore, it has been found that the effect of HO-1 on iron homeostasis within macrophages may represent a new tool to prevent foam cell formation and atherosclerotic lesion progression. A protective effect of iron depletion that may have multiple beneficial consequences is decreased availability of redox-active iron *in vivo*. The amount of free iron available at sites of oxidative or inflammatory injury appears to be a function of the stored iron level. Indeed, a recent study found that the cytoprotective effect of HO-1 induction or forced expression (which usually leads to concomitant elevated serum bilirubin level) may derive from temporary elevated expression of ferritin, and consequent reduction of redox-active iron^[42]. Alongside this, there is accumulating evidence to suggest that excessive hepatic iron accumulation in NAFLD is one of the potential cofactors involved in the enhanced oxidative stress, which triggers liver cell necrosis and eventually fibrosis^[9-11]. There are potentially multiple mechanistic underpinnings for the beneficial association of bilirubin with NAFLD.

The development of antioxidant therapeutics is gaining prominence as the pathophysiological foundation of oxidative stress in cardiovascular disease, neoplasia and NAFLD is better understood^[43]. Significant numbers

of studies have focused on using vitamin based antioxidant therapy to assess prevention of cardiovascular or neoplastic disease. Despite replicable *in vitro* evidence to support antioxidant vitamin use, this has poor correlation with *in vivo* subjects showing conflicting results at best to date. Further ongoing large randomised controlled trials results are awaited^[44,45]. This disparity is likely due to any exogenous vitamin antioxidants' inability to meaningfully influence intracellular antioxidant levels^[46].

Bilirubin is recognised to have antioxidant activity so inducing "iatrogenic Gilbert's syndrome" is another strategy for advancing antioxidant therapeutics which involves using drugs that promote the unconjugated hyperbilirubinaemic state. This strategy may overcome the difficulties of achieving sufficient antioxidants at intracellular level as bilirubin's main antioxidant action appears to act not as a direct radical scavenger given its low concentration in tissue. But instead as a potent and specific inhibitor of the membrane bound NADPH oxidase, a key source oxidants in both phagocytic and non-phagocytic cells^[47]. Probenecid, a uricosuric agent is known to decrease hepatic glucuronidation activity, leading to hyperbilirubinaemia which has been observed only in multiple case reports but not formally evaluated in studies in dose-dependent manner^[47,48]. It is known to be well tolerated but decreasing glucuronidation activity will increase half life of commonly used medication such as paracetamol and lorazepam and thus may hinder the application of this drug and strategy^[49,50]. Rifampicin could be another possible candidate, it causes hyperbilirubinaemia by inhibiting the transporter which accelerates hepatocyte uptake of bilirubin^[51-53]. But rifampicin can cause significant side effects which include liver failure although in a small proportion of patients, this would entail closer monitoring. Sodium valproate has a similar mechanism of action to rifampicin but due to many side effects and poor tolerability would be unsuitable for this application.

Oral administration of bilirubin or its precursor biliverdin which is more soluble is another possible avenue but this strategy is limited by the lack of evidence showing increased bilirubin levels after oral administration other than in mice and rat models^[41,54]. Even if clear evidence was obtained the mass production of bilirubin is complex and costly thus would be a major obstacle to overcome^[55]. Phycobilins that are structural analogues of biliverdin are produced readily by plants, algae and cyanobacteria which may provide feasible alternatives to bilirubin^[56]. Both in animal models and *in vitro* studies phycobilins have been shown to have comparable effect to bilirubin^[57,58]. Further research in this area with human subjects is awaited.

To conclude once the protective association between unconjugated hyperbilirubinaemia and NAFLD is verified. The focus of preventing NAFLD progression and development should consider bilirubin induction therapy randomised double blind controlled trials to assess the clinical value of this treatment. Thereafter it remains to be shown how popular this treatment would be as patient

will be icteric. Although equally with weight reduction as the current mainstay of NAFLD prevention, this strategy has its own limitations and therefore optimal treatment options which are acceptable to patients continues to be a key challenge in this area.

COMMENTS

Background

Obesity is a growing epidemic in the western world and if it is not controlled and reversed it is associated with type 2 diabetes, hypercholesterolaemia and coronary artery disease. The hepatic manifestation of this multi-organ disease is non-alcoholic fatty liver disease (NAFLD). This hepatic dysfunction can eventually lead to cirrhosis and hepatocellular carcinoma. At present there is no effective treatment available and the key current management is prevention by weight reduction. High levels of bilirubin (hyperbilirubinaemia), which is naturally occurring anti-oxidant in the human body has been shown to prevent progression of NAFLD in some studies. This paper evaluates all studies in relation to this, to the look at the current strength of evidence and the potential application of anti-oxidants in this setting.

Research frontiers

Preventing NAFLD progression is a key area of research, inducing hyperbilirubinaemia represents a potential solution. Laboratory based research as well as on animal models has shown high levels of bilirubin to have protective and preventative effect on NAFLD and other diseases processes related to obesity. But this relationship needs to be substantiated in human subjects.

Innovations and breakthroughs

Although most studies evaluated in this review suggest there is likely a protective association between high bilirubin levels and NAFLD progression and prevention. The poor design of these studies is prohibitive to allow any meaningful conclusion. There is a need for good quality large prospective cohort study in multiple populations to test this hypothesis fully, which should also included analysis of UGT1A1 gene to expound upon underlying cause of unconjugated hyperbilirubinaemia.

Applications

This review allows readers to appreciate and evaluate current progress in this area of research. The case for protective association of hyperbilirubinaemia with NAFLD needs to be corroborated with good quality studies. They are multiple drug options available to induce hyperbilirubinaemia but these drugs need to be tested in randomised controlled trial setting.

Terminology

Bilirubin is a naturally occurring substance in the blood which comes from break down of red blood cells. It is an anti-oxidant (protects cells against harmful substances) and appears to also have an anti-inflammatory effect. High level of bilirubin gives the jaundiced appearance.

Peer review

It is a good review article, the authors provide updates of the fatty liver diseases with a Q and A format for easy reads.

REFERENCES

- 1 **Browning JD**, Szczepaniak LS, Dobbins R, Nuremberg P, Horton JD, Cohen JC, Grundy SM, Hobbs HH. Prevalence of hepatic steatosis in an urban population in the United States: impact of ethnicity. *Hepatology* 2004; **40**: 1387-1395 [PMID: 15565570 DOI: 10.1002/hep.20466]
- 2 **Neuschwander-Tetri BA**, Caldwell SH. Nonalcoholic steatohepatitis: summary of an AASLD Single Topic Conference. *Hepatology* 2003; **37**: 1202-1219 [PMID: 12717402 DOI: 10.1053/jhep.2003.50193]
- 3 **Choi SY**, Kim D, Kim HJ, Kang JH, Chung SJ, Park MJ, Kim YS, Kim CH, Choi SH, Kim W, Kim YJ, Yoon JH, Lee HS, Cho SH, Sung MW, Oh BH. The relation between non-alcoholic fatty liver disease and the risk of coronary heart disease in Koreans. *Am J Gastroenterol* 2009; **104**: 1953-1960 [PMID: 19491838 DOI: 10.1038/ajg.2009.238]
- 4 **Speliotis EK**, Massaro JM, Hoffmann U, Vasani RS, Meigs JB, Sahani DV, Hirschhorn JN, O'Donnell CJ, Fox CS. Fatty liver is associated with dyslipidemia and dysglycemia independent of visceral fat: the Framingham Heart Study. *Hepatology* 2010; **51**: 1979-1987 [PMID: 20336705 DOI: 10.1002/hep.23593]
- 5 **Chen SH**, He F, Zhou HL, Wu HR, Xia C, Li YM. Relationship between nonalcoholic fatty liver disease and metabolic syndrome. *J Dig Dis* 2011; **12**: 125-130 [PMID: 21401898 DOI: 10.1111/j.1751-2980.2011.00487.x]
- 6 **Salamone F**, Bugianesi E. Nonalcoholic fatty liver disease: the hepatic trigger of the metabolic syndrome. *J Hepatol* 2010; **53**: 1146-1147 [PMID: 20817302 DOI: 10.1016/j.jhep.2010.06.013]
- 7 **Oliveira CP**, da Costa Gayotto LC, Tatai C, Della Bina BI, Janiszewski M, Lima ES, Abdalla DS, Lopasso FP, Laurindo FR, Laudanna AA. Oxidative stress in the pathogenesis of nonalcoholic fatty liver disease, in rats fed with a choline-deficient diet. *J Cell Mol Med* 2002; **6**: 399-406 [PMID: 12417056 DOI: 10.1111/j.1582-4934.2002.tb00518.x]
- 8 **Sumida Y**, Nakashima T, Yoh T, Furutani M, Hirohama A, Kakisaka Y, Nakajima Y, Ishikawa H, Mitsuyoshi H, Okanoue T, Kashima K, Nakamura H, Yodoi J. Serum thioredoxin levels as a predictor of steatohepatitis in patients with non-alcoholic fatty liver disease. *J Hepatol* 2003; **38**: 32-38 [PMID: 12480557 DOI: 10.1016/S0168-8278(02)00331-8]
- 9 **Zelber-Sagi S**, Nitzan-Kaluski D, Halpern Z, Oren R. NAFLD and hyperinsulinemia are major determinants of serum ferritin levels. *J Hepatol* 2007; **46**: 700-707 [PMID: 17150278]
- 10 **Ryan MC**, Itsiopoulos C, Thodis T, Ward G, Trost N, Hofferberth S, O'Dea K, Desmond PV, Johnson NA, Wilson AM. The Mediterranean diet improves hepatic steatosis and insulin sensitivity in individuals with non-alcoholic fatty liver disease. *J Hepatol* 2013; **59**: 138-143 [PMID: 23485520 DOI: 10.1016/j.jhep.2013.02.012]
- 11 **Mascitelli L**, Goldstein MR. Might some of the beneficial effects of the Mediterranean diet on non-alcoholic fatty liver disease be mediated by reduced iron stores? *J Hepatol* 2013; **59**: 639 [PMID: 23707366 DOI: 10.1016/j.jhep.2013.03.041]
- 12 **Morita M**, Ishida N, Uchiyama K, Yamaguchi K, Itoh Y, Shichiri M, Yoshida Y, Hagihara Y, Naito Y, Yoshikawa T, Niki E. Fatty liver induced by free radicals and lipid peroxidation. *Free Radic Res* 2012; **46**: 758-765 [PMID: 22468959 DOI: 10.3109/10715762.2012.677840]
- 13 **Terao K**, Niki E. Damage to biological tissues induced by radical initiator 2,2'-azobis(2-amidinopropane) dihydrochloride and its inhibition by chain-breaking antioxidants. *J Free Radic Biol Med* 1986; **2**: 193-201 [PMID: 3571847]
- 14 **Shimasaki H**, Saypil WH, Ueta N. Free radical-induced liver injury. II. Effects of intraperitoneally administered 2,2'-azobis(2-amidinopropane) dihydrochloride on the fatty acid profiles of hepatic triacylglycerol and phospholipids. *Free Radic Res Commun* 1991; **14**: 247-252 [PMID: 1874455]
- 15 **Sanyal AJ**, Mofrad PS, Contos MJ, Sargeant C, Luketic VA, Sterling RK, Stravitz RT, Shiffman ML, Clore J, Mills AS. A pilot study of vitamin E versus vitamin E and pioglitazone for the treatment of nonalcoholic steatohepatitis. *Clin Gastroenterol Hepatol* 2004; **2**: 1107-1115 [PMID: 15625656 DOI: 10.1016/S1542-3565(04)00457-4]
- 16 **Baranano DE**, Rao M, Ferris CD, Snyder SH. Biliverdin reductase: a major physiologic cytoprotectant. *Proc Natl Acad Sci USA* 2002; **99**: 16093-16098 [PMID: 12456881 DOI: 10.1073/pnas.252626999]
- 17 **Stocker R**, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. *Science* 1987; **235**: 1043-1046 [PMID: 3029864 DOI: 10.1126/science.3029864]
- 18 **Stocker R**. Antioxidant activities of bile pigments. *Antioxid Redox Signal* 2004; **6**: 841-849 [PMID: 15345144 DOI: 10.1089/ars.2004.6.841]
- 19 **Inoguchi T**, Li P, Umeda F, Yu HY, Kakimoto M, Imamura

- M, Aoki T, Etoh T, Hashimoto T, Naruse M, Sano H, Utsumi H, Nawata H. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C--dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 2000; **49**: 1939-1945 [PMID: 11078463 DOI: 10.2337/diabetes.49.11.1939]
- 20 **Keshavan P**, Deem TL, Schwemberger SJ, Babcock GF, Cook-Mills JM, Zucker SD. Unconjugated bilirubin inhibits VCAM-1-mediated transendothelial leukocyte migration. *J Immunol* 2005; **174**: 3709-3718 [PMID: 15749910 DOI: 10.4049/jimmunol.174.6.3709]
- 21 **Willis D**, Moore AR, Frederick R, Willoughby DA. Heme oxygenase: a novel target for the modulation of the inflammatory response. *Nat Med* 1996; **2**: 87-90 [PMID: 8564848]
- 22 **Li L**, Grenard P, Nhieu JT, Julien B, Mallat A, Habib A, Lotersztajn S. Heme oxygenase-1 is an antifibrogenic protein in human hepatic myofibroblasts. *Gastroenterology* 2003; **125**: 460-469 [PMID: 12891549 DOI: 10.1016/S0016-5085(03)00906-5]
- 23 **Cheriyath P**, Gorrepati VS, Peters I, Nookala V, Murphy ME, Srouji N, Fischman D. High Total Bilirubin as a Protective Factor for Diabetes Mellitus: An Analysis of NHANES Data From 1999 - 2006. *J Clin Med Res* 2010; **2**: 201-206 [PMID: 21629541 DOI: 10.4021/jocmr425w]
- 24 **Lin JP**, O'Donnell CJ, Schwaiger JP, Cupples LA, Lingenhel A, Hunt SC, Yang S, Kronenberg F. Association between the UGT1A1*28 allele, bilirubin levels, and coronary heart disease in the Framingham Heart Study. *Circulation* 2006; **114**: 1476-1481 [PMID: 17000907 DOI: 10.1161/CIRCULATIONAHA.106.633206]
- 25 **Perlstein TS**, Pande RL, Creager MA, Weuve J, Beckman JA. Serum total bilirubin level, prevalent stroke, and stroke outcomes: NHANES 1999-2004. *Am J Med* 2008; **121**: 781-788. e1 [PMID: 18724968 DOI: 10.1016/j.amjmed.2008.03.045]
- 26 **Lin LY**, Kuo HK, Hwang JJ, Lai LP, Chiang FT, Tseng CD, Lin JL. Serum bilirubin is inversely associated with insulin resistance and metabolic syndrome among children and adolescents. *Atherosclerosis* 2009; **203**: 563-568 [PMID: 18775539 DOI: 10.1016/j.atherosclerosis.2008.07.021]
- 27 **Zucker SD**, Horn PS, Sherman KE. Serum bilirubin levels in the U.S. population: gender effect and inverse correlation with colorectal cancer. *Hepatology* 2004; **40**: 827-835 [PMID: 15382174]
- 28 **Vítek L**, Jirsa M, Brodanová M, Kalab M, Marecek Z, Danzig V, Novotný L, Kotal P. Gilbert syndrome and ischemic heart disease: a protective effect of elevated bilirubin levels. *Atherosclerosis* 2002; **160**: 449-456 [PMID: 11849670 DOI: 10.1016/S0021-9150(01)00601-3]
- 29 **Inoguchi T**, Sasaki S, Kobayashi K, Takayanagi R, Yamada T. Relationship between Gilbert syndrome and prevalence of vascular complications in patients with diabetes. *JAMA* 2007; **298**: 1398-1400 [PMID: 17895455 DOI: 10.1001/jama.298.12.1398-b]
- 30 **Jang BK**. Elevated serum bilirubin levels are inversely associated with nonalcoholic fatty liver disease. *Clin Mol Hepatol* 2012; **18**: 357-359 [PMID: 23323250 DOI: 10.3350/cmh.2012.18.4.357]
- 31 **Madan K**, Bhardwaj P, Thareja S, Gupta SD, Saraya A. Oxidant stress and antioxidant status among patients with non-alcoholic fatty liver disease (NAFLD). *J Clin Gastroenterol* 2006; **40**: 930-935 [PMID: 17063114 DOI: 10.1097/01mcg.000212608.59090.08]
- 32 **Chang Y**, Ryu S, Zhang Y, Son HJ, Kim JY, Cho J, Guallar E. A cohort study of serum bilirubin levels and incident non-alcoholic fatty liver disease in middle aged Korean workers. *PLoS One* 2012; **7**: e37241 [PMID: 22615952 DOI: 10.1371/journal.pone.0037241]
- 33 **Kwak MS**, Kim D, Chung GE, Kang SJ, Park MJ, Kim YJ, Yoon JH, Lee HS. Serum bilirubin levels are inversely associated with nonalcoholic fatty liver disease. *Clin Mol Hepatol* 2012; **18**: 383-390 [PMID: 23323254 DOI: 10.3350/cmh.2012.18.4.383]
- 34 **Duseja A**, Das A, Das R, Dhiman RK, Chawla Y, Bhansali A. Unconjugated hyperbilirubinemia in nonalcoholic steatohepatitis--is it Gilbert's syndrome? *Trop Gastroenterol* 2005; **26**: 123-125 [PMID: 16512459]
- 35 **Hjelkrem M**, Morales A, Williams CD, Harrison SA. Unconjugated hyperbilirubinemia is inversely associated with non-alcoholic steatohepatitis (NASH). *Aliment Pharmacol Ther* 2012; **35**: 1416-1423 [PMID: 22540836 DOI: 10.1111/j.1365-2036.2012.05114.x]
- 36 **Kumar R**, Rastogi A, Maras JS, Sarin SK. Unconjugated hyperbilirubinemia in patients with non-alcoholic fatty liver disease: a favorable endogenous response. *Clin Biochem* 2012; **45**: 272-274 [PMID: 22198578 DOI: 10.1016/j.clinbiochem.2011.11.017]
- 37 **Chisholm J**, Seki Y, Toouli J, Stahl J, Collins J, Kow L. Serologic predictors of nonalcoholic steatohepatitis in a population undergoing bariatric surgery. *Surg Obes Relat Dis* 2012; **8**: 416-422 [PMID: 21865094 DOI: 10.1016/j.soard.2011.06.010]
- 38 **Tarantino G**, Colao A, Capone D, Conca P, Tarantino M, Grimaldi E, Chianese D, Finelli C, Contaldo F, Scopacasa F, Savastano S. Circulating levels of cytochrome C, gamma-glutamyl transferase, triglycerides and unconjugated bilirubin in overweight/obese patients with non-alcoholic fatty liver disease. *J Biol Regul Homeost Agents* 2011; **25**: 47-56 [PMID: 21382273]
- 39 **Muraca M**, Fevery J. Influence of sex and sex steroids on bilirubin uridine diphosphate-glucuronosyltransferase activity of rat liver. *Gastroenterology* 1984; **87**: 308-313 [PMID: 6428963]
- 40 **Finn AV**, Nakano M, Polavarapu R, Karmali V, Saeed O, Zhao X, Yazdani S, Otsuka F, Davis T, Habib A, Narula J, Kolodgie FD, Virmani R. Hemoglobin directs macrophage differentiation and prevents foam cell formation in human atherosclerotic plaques. *J Am Coll Cardiol* 2012; **59**: 166-177 [PMID: 22154776 DOI: 10.1016/j.jacc.2011.10.852]
- 41 **Lin HL**, Xu XS, Lu HX, Zhang L, Li CJ, Tang MX, Sun HW, Liu Y, Zhang Y. Pathological mechanisms and dose dependency of erythrocyte-induced vulnerability of atherosclerotic plaques. *J Mol Cell Cardiol* 2007; **43**: 272-280 [PMID: 17628589 DOI: 10.1016/j.yjmcc.2007.05.023]
- 42 **Lanceta L**, Li C, Choi AM, Eaton JW. Haem oxygenase-1 overexpression alters intracellular iron distribution. *Biochem J* 2013; **449**: 189-194 [PMID: 22989377 DOI: 10.1042/BJ20120936]
- 43 **Schwertner HA**, Vitek L. Gilbert syndrome, UGT1A1*28 allele, and cardiovascular disease risk: possible protective effects and therapeutic applications of bilirubin. *Atherosclerosis* 2008; **198**: 1-11 [PMID: 18343383 DOI: 10.1016/j.atherosclerosis.2008.01.001]
- 44 **Christen WG**, Gaziano JM, Hennekens CH. Design of Physicians' Health Study II--a randomized trial of beta-carotene, vitamins E and C, and multivitamins, in prevention of cancer, cardiovascular disease, and eye disease, and review of results of completed trials. *Ann Epidemiol* 2000; **10**: 125-134 [PMID: 10691066]
- 45 **Hennekens CH**, Buring JE, Manson JE, Stampfer M, Rosner B, Cook NR, Belanger C, LaMotte F, Gaziano JM, Ridker PM, Willett W, Peto R. Lack of effect of long-term supplementation with beta carotene on the incidence of malignant neoplasms and cardiovascular disease. *N Engl J Med* 1996; **334**: 1145-1149 [PMID: 8602179 DOI: 10.1056/NEJM199605023341801]
- 46 **Juránek I**, Nikitovic D, Kouretas D, Hayes AW, Tsatsakis AM. Biological importance of reactive oxygen species in relation to difficulties of treating pathologies involving oxidative stress by exogenous antioxidants. *Food Chem Toxicol* 2013; **61**: 240-247 [PMID: 24025685 DOI: 10.1016/j.fct.2013.08.074]
- 47 **McCarty MF**. "Iatrogenic Gilbert syndrome"--a strategy for reducing vascular and cancer risk by increasing plasma unconjugated bilirubin. *Med Hypotheses* 2007; **69**: 974-994 [PMID: 17825497 DOI: 10.1016/j.mehy.2006.12.069]

- 48 **Bokoch GM**, Knaus UG. NADPH oxidases: not just for leukocytes anymore! *Trends Biochem Sci* 2003; **28**: 502-508 [PMID: 13678962 DOI: 10.1016/S0968-0004(03)00194-4]
- 49 **Abernethy DR**, Greenblatt DJ, Ameer B, Shader RI. Probenecid impairment of acetaminophen and lorazepam clearance: direct inhibition of ether glucuronide formation. *J Pharmacol Exp Ther* 1985; **234**: 345-349 [PMID: 4020675]
- 50 **Vree TB**, van den Biggelaar-Martea M, Verwey-van Wissen CP, van Ewijk-Beneken Kolmer EW. Probenecid inhibits the glucuronidation of indomethacin and O-desmethylin-domethacin in humans. A pilot experiment. *Pharm World Sci* 1994; **16**: 22-26 [PMID: 8156046]
- 51 **Turner KC**, Brouwer KL. In vitro mechanisms of probenecid-associated alterations in acetaminophen glucuronide hepatic disposition. *Drug Metab Dispos* 1997; **25**: 1017-1021 [PMID: 9311615]
- 52 **Murthy GD**, Byron D, Shoemaker D, Visweswaraiiah H, Pasquale D. The utility of rifampin in diagnosing Gilbert's syndrome. *Am J Gastroenterol* 2001; **96**: 1150-1154 [PMID: 11316162 DOI: 10.1111/j.1572-0241.2001.03693.x]
- 53 **Campbell SD**, de Morais SM, Xu JJ. Inhibition of human organic anion transporting polypeptide OATP 1B1 as a mechanism of drug-induced hyperbilirubinemia. *Chem Biol Interact* 2004; **150**: 179-187 [PMID: 15535988 DOI: 10.1016/j.cbi.2004.08.008]
- 54 **Vitek L**, Carey MC. Enterohepatic cycling of bilirubin as a cause of 'black' pigment gallstones in adult life. *Eur J Clin Invest* 2003; **33**: 799-810 [PMID: 12925040 DOI: 10.1046/j.1365-2362.2003.01214.x]
- 55 **Meyer UA**. [Heme biosynthesis]. *Schweiz Med Wochenschr* 1975; **105**: 1165-1168 [PMID: 1215898]
- 56 **Brown SB**, Houghton JD, Vernon DI. Biosynthesis of phyco-bilins. Formation of the chromophore of phytochrome, phycocyanin and phycoerythrin. *J Photochem Photobiol B* 1990; **5**: 3-23 [PMID: 2111391]
- 57 **Romay C**, González R. Phycocyanin is an antioxidant protector of human erythrocytes against lysis by peroxy radicals. *J Pharm Pharmacol* 2000; **52**: 367-368 [PMID: 10813544]
- 58 **Bhat VB**, Madyastha KM. C-phycocyanin: a potent peroxy radical scavenger in vivo and in vitro. *Biochem Biophys Res Commun* 2000; **275**: 20-25 [PMID: 10944434 DOI: 10.1006/bbrc.2000.3270]

P- Reviewer: Ahmed M, Mascitelli L, Oeda S, Shen WJ, Tamano M **S- Editor:** Song XX **L- Editor:** A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

