

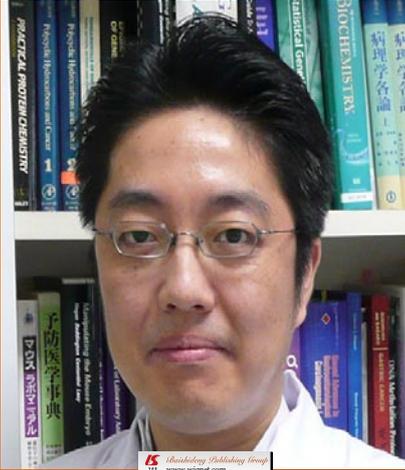
# World Journal of Gastrointestinal Pathophysiology

2013 Bound Volume 4 Issue 1-4: 1-125

ISSN 2150-5330 (online)

World Journal of  
*Gastrointestinal Pathophysiology*

World J Gastrointest Pathophysiol 2013 February 15; 4(1): 1-27



*Baishideng Publishing Group*  
www.wjnet.com

ISSN 2150-5330 (online)

World Journal of  
*Gastrointestinal Pathophysiology*

World J Gastrointest Pathophysiol 2013 May 15; 4(2): 28-42



*Baishideng Publishing Group*  
www.wjnet.com

ISSN 2150-5330 (online)

World Journal of  
*Gastrointestinal Pathophysiology*

World J Gastrointest Pathophysiol 2013 August 15; 4(3): 43-64



*Baishideng Publishing Group*  
www.wjnet.com

ISSN 2150-5330 (online)

World Journal of  
*Gastrointestinal Pathophysiology*

World J Gastrointest Pathophysiol 2013 November 15; 4(4): 65-125  
Volume End



*Baishideng Publishing Group*  
www.wjnet.com



*Baishideng Publishing Group*  
www.wjnet.com



*Baishideng Publishing Group*  
www.wjnet.com

## Editorial Board

2011-2015

The *World Journal of Gastrointestinal Pathophysiology* Editorial Board consists of 526 members, representing a team of worldwide experts in gastrointestinal pathophysiology. They are from 45 countries, including Argentina (2), Australia (14), Austria (3), Belgium (9), Brazil (10), Brunei Darussalam (1), Canada (20), China (30), Croatia (1), Czech Republic (2), Denmark (4), Egypt (1), Estonia (1), Finland (1), France (8), Germany (22), Greece (7), Hungary (5), India (11), Indonesia (1), Iran (2), Ireland (2), Israel (8), Italy (43), Japan (47), Lebanon (3), Malaysia (1), Mexico (2), Netherlands (8), Norway (1), Poland (4), Portugal (1), Romania (1), Russia (1), Singapore (4), South Korea (13), Spain (23), Sweden (11), Switzerland (4), Thailand (2), Turkey (6), Ukraine (1), United Kingdom (10), United States (174), and Venezuela (1).

### EDITOR-IN-CHIEF

Thomas Y Ma, *Albuquerque*

### STRATEGY ASSOCIATE EDITOR-IN-CHIEF

Hirota Akiho, *Fukuoka*  
Jean-Francois Beaulieu, *Sherbrooke*  
Michael W Bradbury, *Erie*  
Sharon DeMorrow, *Temple*

### GUEST EDITORIAL BOARD MEMBERS

Jia-Ming Chang, *Taipei*  
Wai-Keung Chow, *Taichung*  
Chien-Wei Hsu, *Kaohsiung*  
Ming-Tsan Lin, *Taipei*  
Bor-Shyang Sheu, *Tainan*  
Jin-Town Wang, *Taipei*

### MEMBERS OF THE EDITORIAL BOARD



#### Argentina

Bernabé Matías Quesada, *Buenos Aires*  
Marcelo G Roma, *Rosario*



#### Australia

Chris Richard Abbiss, *Joondalup*  
Guy D Eslick, *Penrith*  
Montri Gururatsakul, *Adelaide*  
Chandana Herath, *Melbourne*  
Michael Horowitz, *Adelaide*  
Mustafa Khasraw, *Geelong*  
Shu-Chuen Li, *Callaghan*  
Antonina Mikocka-Walus, *Adelaide*  
Nam Quoc Nguyen, *Adelaide*

Kulmira Nurgali, *St Albans*  
Nicholas John Spencer, *Flagstaff Hill*  
Nick Spencer, *Adelaide*  
Deborah Verran, *Camperdown*  
Shu-Feng Zhou, *Melbourne*



#### Austria

Cord Langner, *Graz*  
Dietmar Ofner-Velano, *Salzburg*  
Michael Trauner, *Graz*



#### Belgium

Kathleen Blondeau, *Leuven*  
Robaey Geert, *Genk*  
Ilse Maria Hoffman, *Leuven*  
Michael H J Maes, *Wilrijk*  
Theodoor Abram Niewold, *Heverlee*  
Xavier Sagaert, *Leuven*  
Jean-Marie Vanderwinden, *Brussels*  
Kristin Verbeke, *Leuven*  
Mathieu Vinken, *Roeselare*



#### Brazil

Uilian Andreis, *Botucatu*  
Everson L A Artifon, *Vila Mariana*  
João Batista Calixto, *Trindade*  
Niels O Saraiva Câmara, *Vila Clementino*  
Julio Chebli, *Juiz de Fora*  
Fernando Fornari, *Passo Fundo*  
Clélia Akiko Hiruma-Lima, *Botucatu*  
Marcel C C Machado, *Sao Paulo*  
Juarez Quaresma, *Belem*  
Wagner Vilegas, *Araraquara*



#### Brunei Darussalam

Vui Heng Chong, *Bandar Seri Begawan*



#### Canada

Fernando Alvarez, *Montréal*  
Francois Boudreau, *Sherbrooke*  
George A Bubenik, *Guelph*  
Wang-Xue Chen, *Ottawa*  
Jan D Huizinga, *Puslinch*  
Kusum K Kharbanda, *Omaha*  
Wolfgang Kunze, *Hamilto*  
Jian-Jun Li, *Ottawa*  
Roderick John Macleod, *Kingston*  
Michele Molinari, *Halifax*  
Nathalie Rivard, *Sherbrooke*  
Kirill Rosen, *Halifax*  
Manuela Santos, *Montreal*  
Caroline Saucier, *Quebec*  
Jean Sévigny, *Quebec*  
Eldon A Shaffer, *Calgary*  
Manuel A Silva, *Hamilton*  
Alan B R Thomson, *Edmonton*  
Pierre H Vachon, *Sherbrooke*



#### China

Kai-Xing Ai, *Shanghai*  
Zhao-Xiang Bian, *Hong Kong*  
Min-Hu Chen, *Guangzhou*  
CH Cho, *Hong Kong*  
Zhong-Hong Gao, *Wuhan*  
Jun-Ming Guo, *Ningbo*  
Jing-Yan Han, *Beijing*

Jian-Dong Huang, *Hong Kong*  
 Jia-Fu Ji, *Beijing*  
 Shi Liu, *Wuhan*  
 Zhan-Ju Liu, *Shanghai*  
 Xiao-Hong Wang, *Beijing*  
 Zhen-Ning Wang, *Shenyang*  
 Wei Wei, *Hefei*  
 Dong-Ping Xie, *Shanghai*  
 Wen-Xie Xu, *Shanghai*  
 Hua Yang, *Chongqing*  
 Xiao Yang, *Beijing*  
 Wei-Zhen Zhang, *Beijing*  
 Hua-Chuan Zheng, *Shenyang*  
 Da-Ling Zhu, *Harbin*  
 Jin-Xia Zhu, *Beijing*  
 Min-Sheng Zhu, *Nanjing*  
 Yong-Liang Zhu, *Hangzhou*



#### Croatia

Alen Protic, *Rijeka*



#### Czech Republic

Pavel Hladik, *Semily*  
 Martin Vokurka, *Prague*



#### Denmark

Lars Arendt-Nielsen, *Aalborg*  
 Frank Vinholt Schiodt, *Copenhagen*  
 Jonas Worsoe, *Aarhus*  
 Jing-Bo Zhao, *Aalborg*



#### Egypt

Mahmoud Aboelneen Khattab, *Minia*



#### Estonia

Enn Seppet, *Tartu*



#### Finland

Pauli Antero Puolakkainen, *Turku*



#### France

Bruno Bonaz, *Grenoble*  
 Pierre Marie Dechelotte, *Rouen*  
 Jean-Paul Lallès, *Saint-Gilles*  
 Charles-Henri Malbert, *Saint-Gilles*  
 Thierry Piche, *Nice*  
 Pascale Plaisancié, *Lyon*  
 Michelina Plateroti, *Lyon*  
 Veronique Vitton, *Marseille*



#### Germany

Hans Gunter Beger, *Ulm*  
 Carsten Bergmann, *Ingelheim*  
 Elke Cario, *Essen*

Arno J Dormann, *Koln*  
 Nikolaus Gassler, *Aachen*  
 Werner Hartwig, *Heidelberg*  
 Marion Hewicker-Trautwein, *Hannover*  
 Jens Hoepfner, *Freiburg*  
 Tobias Keck, *Freiburg*  
 Jorg Kleeff, *Munich*  
 Peter Malfertheiner, *Magdeburg*  
 Oliver Mann, *Hamburg*  
 Christoph Michalski, *Munich*  
 Andreas Klaus Nussler, *Munich*  
 Christian Pehl, *Vilsbiburg*  
 Peter Schemmer, *Heidelberg*  
 Marc Stemmler, *Freiburg*  
 Frank Tacke, *Aachen*  
 Sya Nomna Ukena, *Hannover*  
 Brigitte Vollmar, *Rostock*  
 Thomas Michael Wex, *Magdeburg*  
 Margot Zoller, *Heidelberg*



#### Greece

Stelios F Assimakopoulos, *Patras*  
 George N Dalekos, *Larissa*  
 Alkiviadis Efthymiou, *thessaloniki*  
 Maria Gazouli, *Athens*  
 Ioannis E Koutroubakis, *Heraklion*  
 Gerassimos J Mantzaris, *Athens*  
 George Papatheodoridis, *Athens*



#### Hungary

Mária Bagyánszki, *Szeged*  
 Mihály Boros, *Szeged*  
 Laszlo Czako, *Szeged*  
 Pal Miheller, *Budapest*  
 Zoltan Rakonczay, *Szeged*



#### India

Anil Kumar Agarwal, *Delhi*  
 Uday Bandyopadhyay, *Kolkata*  
 Sriparna Basu, *Varanasi*  
 Chandra Kanti Chakraborti, *Rourkela*  
 Nilesh M Dagia, *Mumbai*  
 Rajeev Garg, *Punjab*  
 Chandra P Sharma, *Thiruvananthapuram*  
 Shailesh V Shrikhande, *Mumbai*  
 Virendra Singh, *Chandigarh*  
 Nicholas James Skill, *Indianapolis*  
 Prabhakar R Veerareddy, *Andhra Pradesh*



#### Indonesia

Laurentius A Lesmana, *Jakarta*



#### Iran

Gholamreza Roshandel, *Gorgan*  
 Shahram Shahabi, *Urmia*



#### Ireland

Billy Bourke, *Dublin*

Stephen Keely, *Dublin*



#### Israel

Yosefa Avraham, *Jerusalem*  
 Yaron Bar-Dayana, *Holon*  
 Shomron Ben-Horin, *Hashomer*  
 Boris Kirshtein, *Beer Sheva*  
 Stephen Malnick, *Rehovot*  
 Yaakov Maor, *Tel-Hashomer*  
 Rifaat Safadi, *Jerusalem*  
 Nachum Vaisman, *Tel Aviv*



#### Italy

Rosaria Acquaviva, *Catania*  
 Dario Acuna-Castroviejo, *Armilla*  
 Alessandro Antonelli, *Pisa*  
 Giacosa Attilio, *Genova*  
 Salvatore Auricchio, *Naples*  
 Guido Basilisco, *Milano*  
 Antonio Basoli, *Rome*  
 Claudio Bassi, *Verona*  
 Massimo Bellini, *Pisa*  
 Luigi Bonavina, *Milano*  
 Alfio Brogna, *Catania*  
 Giuseppe Calamita, *Bari*  
 Raffaele Capasso, *Naples*  
 Ignazio Castagliuolo, *Padova*  
 Enrico Stefano Corazziari, *Rome*  
 Francesco Cresi, *Torino*  
 Rosario Cuomo, *Napoli*  
 Salvatore Cuzzocrea, *Gazzi*  
 Mario M D'Elios, *Florence*  
 Cinzia Domeneghini, *Milan*  
 Luca Elli, *Milano*  
 Cresi Francesco, *Torino*  
 Walter Fries, *Messina*  
 Eugenio Gaudio, *Rome*  
 Marco Gobetti, *Bari*  
 Fabio Grizzi, *Milan*  
 Enzo Grossi, *Milanese*  
 Enzo Ierardi, *Foggia*  
 Pietro Invernizzi, *Milan*  
 Angelo A Izzo, *Naples*  
 Anna Kohn, *Rome*  
 Giovanni Latella, *L'Aquila*  
 Massimo Marignani, *Rome*  
 F Marotta, *Milano*  
 Sergio Morini, *Rome*  
 Raffaele Pezzilli, *Bologna*  
 Cristiano Rumio, *Milan*  
 Giovanni Sarnelli, *Naples*  
 Edoardo Vincenzo Savarino, *Genoa*  
 Pierpaolo Sileri, *Rome*  
 Annamaria Staiano, *Naples*  
 Giacomo Carlo Sturniolo, *Padova*  
 Claudio Tiribelli, *Triest*



#### Japan

Akihiro Asakawa, *Kagoshima*  
 Hisashi Aso, *Sendai*  
 Yasu-Taka Azuma, *Osaka*  
 Shotaro Enomoto, *Wakayama*  
 Mikihiro Fujiya, *Hokkaido*  
 Takahisa Furuta, *Hamamatsu*

Akira Hokama, *Okinawa*  
 Ryota Hokari, *Saitama*  
 Yuichi Hori, *Kobe*  
 Hideki Iijima, *Osaka*  
 Masahiro Iizuka, *Akita*  
 Motohiro Imano, *Osaka*  
 Hajime Isomoto, *Nagasaki*  
 Tatehiro Kagawa, *Isehara*  
 Takumi Kawaguchi, *Kurume*  
 Haruki Kitazawa, *Sendai*  
 Xiao-Kang Li, *Tokyo*  
 Noriaki Manabe, *Okayama*  
 Atsushi Masamune, *Sendai*  
 Hiroyuki Matsubayashi, *Shizuoka*  
 Kazuyuki Matsushita, *Chuo-ku*  
 Reiko Miyazawa, *Gunma*  
 Kazunari Murakami, *Oita*  
 Hikaru Nagahara, *Tokyo*  
 Yuji Naito, *Kyoto*  
 Atsushi Nakajima, *Atsushi Nakajima*  
 Shoji Natsugoe, *Kagoshima*  
 Tsutomu Nishida, *Osaka*  
 Koji Nomoto, *Tokyo*  
 Naoaki Sakata, *Miyagi*  
 Shouji Shimoyama, *Tokyo*  
 Goshi Shiota, *Yonago*  
 Ikuo Shoji, *Hyogo*  
 Hidekazu Suzuki, *Tokyo*  
 Hitoshi Takagi, *Gunma*  
 Toru Takahashi, *Okayama*  
 Yoshihisa Takahashi, *Tokyo*  
 Kan Uchiyama, *Chiba*  
 Takato Ueno, *Kurume*  
 Yoshiyuki Ueno, *Sendai*  
 Hisayuki Uneyama, *Kwasaki*  
 Mitsunori Yamakawa, *Yamagata*  
 Takayuki Yamamoto, *Mie*  
 Yutaka Yata, *Gunma*  
 Naohisa Yoshida, *Kyoto*  
 Hitoshi Yoshiji, *Nara*



#### Lebanon

Costantine Fouad Daher, *Byblos*  
 Assaad M Soweid, *Beirut*  
 Julnar Usta, *Beirut*



#### Malaysia

Andrew Chua, *Perak*



#### Mexico

José María de la Roca-Chiapas, *Leon*  
 María Raquel Huerta Franco, *Guanajuato*



#### Netherland

Wouter J de Jonge, *Amsterdam*  
 Aldo Grefhorst, *Groningen*  
 Ruben Hummelen, *Rotterdam*  
 Daniel Keszthelyi, *Maastricht*  
 Cornelis F M Sier, *Leiden*  
 Pieter J Tanis, *Amsterdam*  
 Luc JW van der Laan, *Rotterdam*  
 Sander van der Marel, *Leiden*



#### Norway

Anne Marie Bakke, *Oslo*



#### Poland

Stanislaw Hac, *Gdańsk*  
 Stanisław Jan Konturek, *Kraków*  
 Agata Mulak, *Wrocław*  
 Napoleon Waszkiewicz, *Choroszcz*



#### Portugal

Ricardo Marcos, *Porto*



#### Romania

Mihai Ciocirlan, *Bucharest*



#### Russia

Ludmila Filaretova, *Petersburg*



#### Singapore

Madhav Bhatia, *Singapore*  
 Brian K P Goh, *Singapore*  
 Khek Yu Ho, *Singapore*  
 Cliff K S Ong, *Singapore*



#### South Korea

Jae Hee Cheon, *Seoul*  
 Myung Haing Cho, *Seoul*  
 Jae Bock Chung, *Seoul*  
 Ki-Baik Hahm, *Incheon*  
 Ho Jae Han, *Gwangju*  
 Chang Duk Jun, *Gwangju*  
 Hong Joo Kim, *Seoul*  
 Jin Kyung Kim, *Gyeongsan-Si*  
 Sang Geon Kim, *Seoul*  
 Won Jae Lee, *Seoul*  
 Kwan Kyu Park, *Daegu*  
 Seung Ha Park, *Busan*  
 Sung Joo Park, *Jeonbuk*



#### Spain

Raquel Abalo, *Alcorcón*  
 Juan G Abraldes, *Barcelona*  
 Agustin Albillos, *Madrid*  
 Maria-Angeles Aller, *Madrid*  
 Fernando Azpiroz, *Barcelona*  
 Ramon Bataller, *Barcelona*  
 Marco Bustamante, *Valencia*  
 Andres Cardenas, *Barcelona*  
 Dario Acuna Castroviejo, *Armillá*  
 Joan Claria, *Barcelona*  
 Pere Clave, *Barcelona*  
 Manuel Giner, *Madrid*

Angel I Lanas, *Zaragoza*  
 Maite Martin, *Barcelona*  
 Maria Teresa Martin, *Barcelona*  
 Vicente Martinez, *Barcelona*  
 Jose M Matés, *Malaga*  
 Julio M Mayol, *Madrid*  
 Marçal Pastor-Anglada, *Barcelona*  
 María Eugenia Sáez, *Seville*  
 Yolanda Sanz, *Burjassot*  
 Carlos Taxonera, *Madrid*  
 Maria D Yago, *Granada*



#### Sweden

Marco Del Chiaro, *Stockholm*  
 Frida Fak, *Gothenburg*  
 Gunnar FA Flemstrom, *Uppsala*  
 Evangelos Kalaitzakis, *Gothenburg*  
 Kristina Lamas, *Umea*  
 Bob Roger Olsson, *Göteborg*  
 Sara Maria Regné, *Malmö*  
 Peter thelin Schmidt, *Stockholm*  
 Xiao-Feng Sun, *Linköping*  
 Henrik Thorlacius, *Malmö*  
 Curt Tysk, *Orebro*



#### Switzerland

Jyrki J Eloranta, *Zurich*  
 Andreas Geier, *Zurich*  
 Remy Meier, *Liestal*  
 Catherine Pastor, *Geneva*



#### Thailand

Thawatchai Akaraviputh, *Bangkok*  
 Weekitt Kittisupamongkol, *Bangkok*



#### Turkey

Mehmet Bektas, *Ankara*  
 Mukaddes Esrefoglu, *Malatya*  
 Ahmet Guven, *Ankara*  
 Muammer Karadeniz, *Manisa*  
 Elvan Ozbek, *Erzuru*  
 Ilhami Yuksel, *Ankara*



#### Ukraine

Oksana S Zayavhivska, *Lviv*



#### United Kingdom

Geoffrey Burnstock, *London*  
 Janice E Drew, *Aberdeen*  
 Girish Gupte, *Birmingham*  
 David C Hay, *Edinburgh*  
 Nusrat Husain, *Cheshire*  
 Michael Leslie Lucas, *Glasgow*  
 Jamie Murphy, *London*  
 Vadim Sumbayev, *Kent*  
 Wing-Kin Syn, *Birmingham*  
 Andrea Varro, *Liverpool*



## United States

Sami Rene Achem, *Jacksonville*  
 Tauseef Ali, *Oklahoma*  
 David H Alpers, *St Louis*  
 Gianfranco D Alpini, *Temple*  
 Shrikant Anant, *Oklahoma*  
 M Sawkat Anwer, *North Grafton*  
 Andrew Aronsohn, *Chicago*  
 Toms Augustin, *Sayre*  
 Gyorgy Baffy, *Boston*  
 Michael T Bailey, *Columbus*  
 Kim Elaine Barrett, *San Diego*  
 Marc D Basson, *Lansing*  
 Robert L Bell, *New Haven*  
 David H Berger, *Houston*  
 Urs A Boelsterli, *Storrs*  
 Richard G Boles, *Los Angeles*  
 Edward L Bradley III, *Sarasota*  
 Qiang Cai, *Atlanta*  
 Wei-Biao Cao, *Providence*  
 Justin M M Cates, *Nashville*  
 Subhash C Chauhan, *Sioux Falls*  
 Jian-De Chen, *Galveston*  
 Tao-Sheng Chen, *Memphis*  
 John Chiang, *Rootstown*  
 Mashkoor A Choudhry, *Maywood*  
 Parimal Chowdhury, *Little Rock*  
 Eric Cohen, *Boston*  
 Robert Cormier, *Duluth*  
 Srinivasan Dasarathy, *Cleveland*  
 Edwin A Deitch, *Newark*  
 Dan A Dixon, *Columbia*  
 James P Dolan, *Portland*  
 H Henry Dong, *Pittsburgh*  
 Hui Dong, *La Jolla*  
 Ashkan Farhadi, *Irvine*  
 Bin Feng, *Pittsburgh*  
 Jenifer Fenton, *East Lansing*  
 Alessandro Fichera, *Chicago*  
 Mitchell P Fink, *Pittsburgh*  
 P Marco Fisichella, *Maywood*  
 Leo R Fitzpatrick, *Hummelstown*  
 Robert Armour Forse, *Omaha*  
 Glenn Tsuyoshi Furuta, *Aurora*  
 Juan F Gallegos-Orozco, *Scottsdale*  
 Pandu R Gangula, *Nashville*  
 Timothy Gardner, *Lebanon*  
 Shannon Stroud Glaser, *Temple*  
 Francisco Gondim, *St. Louis*  
 John R Grider, *Richmond*  
 Yan-Fang Guan, *Cincinnati*  
 Gregory M Holmes, *Baton Rouge*  
 Ai-Xuan Le Holterman, *Chicago*  
 Richard Hu, *Los Angeles*  
 Hartmut Jaeschke, *Kansas*  
 Robert Thomas Jensen, *Los Angeles*  
 Sreenivasa S Jonnalagadda, *Louis*  
 Michel Kahaleh, *Charlottesville*

Andreas Martin Kaiser, *Los Angeles*  
 Randeep Singh Kashyap, *Rochester*  
 Laurie Keefer, *Chicago*  
 Richard Kellermayer, *Houston*  
 Chris Kevil, *Shreveport*  
 Sandeep Khurana, *Baltimore*  
 Pawel R Kiela, *Tucson*  
 Tammy Lyn Kindel, *Cincinnati*  
 Gordana Kosutic, *Durham*  
 David Kravetz, *San Diego*  
 Ashok Kumar, *Detroit*  
 John H Kwon, *Chicago*  
 Muriel Larauche, *Los Angeles*  
 I Michael Leitman, *New York*  
 Felix W Leung, *North Hills*  
 Suthat Liangpunsakul, *Indianapolis*  
 Feng-Xin Lu, *Boston*  
 Pauline Kay Lund, *Chapel Hill*  
 George Luo, *Lexington*  
 Guang-Xiang Luo, *Lexington*  
 Jay Luther, *Ann Arbor*  
 Ram I Mahato, *Memphis*  
 Akhil Maheshwari, *Birmingham*  
 Kenneth Maiese, *Newark*  
 Adhip P N Majumdar, *Detroit*  
 Jose E Manautou, *Storrs*  
 Craig J McClain, *Louisville*  
 Dermot McGovern, *Los Angeles*  
 B Greenwood-van Meerveld, *Oklahoma*  
 Douglas Scott Merrel, *Bethesda*  
 Murielle Mimeault, *Omaha*  
 Emiko Mizoguchi, *Boston*  
 Huan-Biao Mo, *Denton*  
 Adam Moeser, *Raleigh*  
 Ramzi M Mohammad, *Detroit*  
 Satdarshan Singh Monga, *Pittsburgh*  
 Roger Klein Moreira, *New York*  
 Sandeep Mukherjee, *Omaha*  
 Karnam S Murthy, *Richmond*  
 Michael J Nowicki, *Jackson*  
 Shuji Ogino, *Boston*  
 Mary Francis Otterson, *Wisconsin*  
 Chung Owyang, *Ann Arbor*  
 Helieh S Oz, *Lexington*  
 Marco G Patti, *Chicago*  
 Timothy Michael Pawlik, *Baltimore*  
 Sara Peleg, *Houston*  
 Nicholas C Popescu, *Bethesda*  
 Li-Ya Qiao, *Richmond*  
 Chao Qin, *Oklahoma*  
 Parvaneh Rafiee, *Milwaukee*  
 Sigrid A Rajasekaran, *Wilmington*  
 Vazhaikurichi M Rajendran, *Morgantown*  
 Jean Pierre Raufman, *Baltimore*  
 Ramesh M Ray, *Memphis*  
 Arie Regev, *Indianapolis*  
 Douglas K Rex, *Carmel*  
 Yehuda Ringel, *Chapel Hill*  
 Richard A Rippe, *Rockville*  
 Chantal A Rivera, *Bossier*

Andrea Romani, *Cleveland*  
 Praveen K Roy, *Albuquerque*  
 Paul A Rufo, *Boston*  
 David B Sachar, *New York*  
 Bimaljit Singh Sandhu, *Richmond*  
 Sanjaya Kumar Satapathy, *New Hyde Park*  
 Anthony Senagore, *Los Angeles*  
 Muhammad Y Sheikh, *Fresno*  
 Bo Shen, *Cleveland*  
 Le Shen, *Chicago*  
 Frank A Simmen, *Little Rock*  
 Steven Mitchell Singer, *Washington*  
 Shailinder Jit Singh, *Washington*  
 Adam Jan Smolka, *Charleston*  
 Ned Snyder, *Houston*  
 Zhen-Yuan Song, *Chicago*  
 Gagan K Sood, *Houston*  
 Rhonda F Souza, *Dallas*  
 Stuart Jon Spechler, *Dallas*  
 Subbaramiah Sridha, *Augusta*  
 Catia Sternini, *Los Angeles*  
 Veedamali S Subramanian, *Long Beach*  
 Jun Sun, *Rochester*  
 Yvette Taché, *Los Angeles*  
 Xiao-Di Tan, *Chicago*  
 Paul Daniel Terry, *Atlanta*  
 Jennifer Tirnauer, *Farmington*  
 Andrea Todisco, *Ann Arbor*  
 George C Tsokos, *Boston*  
 Vic Velanovich, *Detroit*  
 Raj Vuppalanchi, *Indianapolis*  
 Estela Wajcberg, *Cranford*  
 Arnold Wald, *Madison*  
 Li-Xin Wang, *Los Angeles*  
 Horst Christian Weber, *Boston*  
 Steven D Wexner, *Weston*  
 Jackie D Wood, *Columbus*  
 Guo-Yao Wu, *College Station*  
 Christian Wunder, *Bethesda*  
 Zuo-Liang Xiao, *Cleveland*  
 Guang-Yin Xu, *Galveston*  
 Guo-Rong Xu, *East Orange*  
 Guang-Yu Yang, *Chicago*  
 Jay A Yelon, *Valhalla*  
 Yamaoka Yoshio, *Houston*  
 Shao-Yong Yu, *Hershey*  
 Yana Zavros, *Cincinnati*  
 Joerg Zehetner, *Los Angeles*  
 Jian X Zhang, *Charlotte*  
 Zhi Zhong, *Charleston*  
 Hui-Ping Zhou, *Richmond*  
 Zhan-Xiang Zhou, *Kannapolis*  
 Qing Zhu, *Bethesda*  
 Yao-Hui Zhu, *Stanford*



## Venezuela

Fabian Michelangeli, *Caracas*

# World Journal of *Gastrointestinal Pathophysiology*

*World J Gastrointest Pathophysiol* 2013 February 15; 4(1): 1-27



**Contents**

Quarterly Volume 4 Number 1 February 15, 2013

- |                             |    |   |
|-----------------------------|----|---|
| <b>REVIEW</b>               | 1  | Therapeutic potential of targeting the renin angiotensin system in portal hypertension<br><i>Herath CB, Grace JA, Angus PW</i>  |
| <hr/>                       |    |   |
| <b>BRIEF ARTICLE</b>        | 12 | Assessment of platelet activation and phagocytic activity in gastric cancer patients<br><i>Matowicka-Karna J, Kamocki Z, Kemono H</i>   |
|                             | 18 | Cytomegalovirus positive ulcerative colitis: A single center experience and literature review<br><i>Kopylov U, Sasson G, Geyshe B, Oikawa MT, Barshack I, Eliakim R, Ben-Horin S</i>  |
| <hr/>                       |    |   |
| <b>LETTER TO THE EDITOR</b> | 24 | Serum cytokine profile in patients with hepatitis B e antigen-negative chronic active hepatitis B and inactive hepatitis B virus carriers<br><i>Dimitropoulou D, Karakantza M, Theodorou GL, Leonidou L, Assimakopoulos SF, Mouzaki A, Gogos CA</i> |

## Contents

*World Journal of Gastrointestinal Pathophysiology*  
Volume 4 Number 1 February 15, 2013

**APPENDIX** I-V Instructions to authors

**ABOUT COVER** Editorial Board Member of *World Journal of Gastrointestinal Pathophysiology*, Shotaro Enomoto, MD, PhD, the Second Department of Internal Medicine, Wakayama Medical University, 811-1, Kimiidera, Wakayama 641-0012, Japan

**AIM AND SCOPE** *World Journal of Gastrointestinal Pathophysiology* (*World J Gastrointest Pathophysiol, WJGP*, online ISSN 2150-5330, DOI: 10.4291), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

*WJGP* is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. *WJGP* specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.

We encourage authors to submit their manuscripts to *WJGP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

**INDEXING/ABSTRACTING** *World Journal of Gastrointestinal Pathophysiology* is now indexed in Digital Object Identifier.

**FLYLEAF** I-IV Editorial Board

### EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Shuai Ma*  
Responsible Electronic Editor: *Ya-Jing Lu*  
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Huan-Huan Zhai*

**NAME OF JOURNAL**  
*World Journal of Gastrointestinal Pathophysiology*

**ISSN**  
ISSN 2150-5330 (online)

**LAUNCH DATE**  
April 15, 2010

**Frequency**  
Quarterly

**EDITOR-IN-CHIEF**  
Thomas Y Ma, MD, PhD, Professor, Chief, Division of Gastroenterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States

**EDITORIAL OFFICE**  
Jin-Lei Wang, Director  
Xiu-Xia Song, Vice Director  
*World Journal of Gastrointestinal Pathophysiology*

Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China  
Telephone: +86-10-85381892  
Fax: +86-10-85381893  
E-mail: [wjgp@wjgnet.com](mailto:wjgp@wjgnet.com)  
<http://www.wjgnet.com>

**PUBLISHER**  
Baishideng Publishing Group Co., Limited  
Flat C, 23/F, Lucky Plaza,  
315-321 Lockhart Road, Wan Chai,  
Hong Kong, China  
Telephone: +852-6555-7188  
Fax: +852-3177-9906  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
<http://www.wjgnet.com>

**PUBLICATION DATE**  
February 15, 2013

### COPYRIGHT

© 2013 Baishideng. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

### SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

### INSTRUCTIONS TO AUTHORS

Full instructions are available online at [http://www.wjgnet.com/2150-5330/g\\_info\\_20100316080008.htm](http://www.wjgnet.com/2150-5330/g_info_20100316080008.htm)

**ONLINE SUBMISSION**  
<http://www.wjgnet.com/esp/>

## Therapeutic potential of targeting the renin angiotensin system in portal hypertension

Chandana B Herath, Josephine A Grace, Peter W Angus

Chandana B Herath, Josephine A Grace, Department of Medicine, University of Melbourne, Austin Health, Heidelberg, Victoria 3084, Australia

Josephine A Grace, Peter W Angus, Department of Gastroenterology, Austin Health, Heidelberg, Victoria 3084, Australia

Author contributions: Herath CB wrote and reviewed the manuscript; Grace JA contributed to the work and reviewed the manuscript; Angus PW reviewed and assisted with editing the manuscript.

Supported by Grant from the National Health and Medical Research Council of Australia

Correspondence to: Chandana B Herath, PhD, Department of Medicine, University of Melbourne, Austin Health, 145 Studley Road, Heidelberg, Victoria 3084, Australia. [cherath@unimelb.edu.au](mailto:cherath@unimelb.edu.au)

Telephone: +61-3-94962549 Fax: +61-3-94575485

Received: September 24, 2012 Revised: November 15, 2012

Accepted: January 5, 2013

Published online: February 15, 2013

### Abstract

Portal hypertension is responsible for the bulk of the morbidity and mortality in patients with cirrhosis. Drug therapy to reduce portal pressure involves targeting two vascular beds. The first approach is to reduce intra hepatic vascular tone induced by the activity of powerful vasoconstrictors such as angiotensin II, endothelin-1 and the sympathetic system and mediated *via* contraction of perisinusoidal myofibroblasts and perversular smooth muscle cells. The second approach is to reduce mesenteric and portal blood flow. Non-selective  $\beta$ -blockers are widely used and have been shown to prolong patient survival and reduce oesophageal variceal bleeding in advanced cirrhosis. However many patients are unable to tolerate these drugs and they are ineffective in a significant proportion of patients. Unfortunately there are no other drug therapies that have proven efficacy in the treatment of portal hypertension and prevention of variceal bleeding. This review briefly outlines current therapeutic approaches to the

management of portal hypertension, and the evidence supporting the role of the renin angiotensin system (RAS) and the use of RAS blockers in this condition. It will also outline recent advances in RAS research that could lead to the development of new treatments focusing in particular on the recently discovered "alternate axis" of the RAS.

© 2013 Baishideng. All rights reserved.

**Key words:** Angiotensin-(1-7); Portal hypertension; Intrahepatic resistance; Mesenteric vasodilatation; Variceal bleeding; Non-selective  $\beta$ -blockers; Renin angiotensin system; Mas receptor; Angiotensin receptor; Cirrhosis

Herath CB, Grace JA, Angus PW. Therapeutic potential of targeting the renin angiotensin system in portal hypertension. *World J Gastrointest Pathophysiol* 2013; 4(1): 1-11 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i1/1.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i1.1>

### INTRODUCTION

Hepatic fibrosis and its end-stage sequelae of cirrhosis and liver cancer are major causes of morbidity and mortality throughout the world and their prevalence is rising, largely due to the increasing impact of chronic viral hepatitis and non alcoholic steatohepatitis. Much of the morbidity and mortality that occurs in cirrhosis is due to the development of portal hypertension. However, despite major advances in our understanding of the pathogenesis of portal hypertension, current treatment options are limited.

It is clear that the renin angiotensin system (RAS) contributes to organ dysfunction and chronic tissue injury in a range of conditions including diabetes, cardiovascular and renal disease, primarily through the vasoactive and profibrotic effects of its key effector peptide, angiotensin II<sup>[1]</sup>. More recently, the RAS has also been

implicated in the pathogenesis of both hepatic fibrosis and portal hypertension<sup>[2,3]</sup>. This is supported by studies which have shown that RAS blockers are able to reduce fibrosis in experimental models of chronic liver injury and that they can lower portal pressure in both animal models and in man, primarily by inhibiting angiotensin II mediated intrahepatic vasoconstriction<sup>[4-6]</sup>. This review will briefly outline current therapeutic approaches to the management of portal hypertension, and focus on the evidence supporting the role of the RAS and the use of RAS blockers in this condition, and recent advances in RAS research that could lead to the development of new treatments.

## CURRENT TREATMENT OF PORTAL HYPERTENSION

The initiating mechanism for the development of portal hypertension is thought to be the development of increased intrahepatic resistance to portal inflow. This is mostly caused by increased deposition of extracellular matrix and disruption of the normal hepatic vascular architecture<sup>[2]</sup>. However, a significant proportion of portal resistance is attributable to intrahepatic vasoconstriction which is caused by the contraction of activated perisinusoidal hepatic stellate cells and of vascular smooth muscle cells in portal venules<sup>[7]</sup>. It is this variable component of intrahepatic resistance, mediated by powerful intrahepatic vasoconstrictors such as angiotensin II and endothelin<sup>[8-11]</sup>, which is potentially amenable to pharmacological therapies.

The second and equally important contributor in the development of portal hypertension is splanchnic vasodilatation which increases portal blood flow<sup>[12]</sup>. The mechanisms responsible for splanchnic vasodilatation in portal hypertension are incompletely understood, however, there is considerable evidence from both animal and human studies to suggest that nitric oxide (NO) generated by endothelial NO synthase (NOS) plays a central role<sup>[13-16]</sup>. One of the key consequences of portal hypertension is the development of portosystemic collaterals, of which the most important clinically are oesophageal varices. These vessels divert much of the increased mesenteric inflow away from the liver. However, even when portal blood flow is entirely diverted through collaterals, portal hypertension persists because of concomitant increases in portal venous inflow caused by increasing splanchnic vasodilatation. It should be noted that although the formation of these collaterals has been assumed to be the result of dilatation of pre-existing vascular channels, recent studies have implicated a process of neoangiogenesis which has been shown to contribute to both the formation of portosystemic collaterals and increased splanchnic blood flow<sup>[17]</sup>.

Bleeding from oesophageal varices is responsible for much of the mortality and morbidity that occurs in patients with portal hypertension. Varices are present in about 50% of patients at diagnosis, and this increases to

about 90% on long-term follow up. The risk of variceal rupture is 10%-30% per year depending on their size and appearance and the severity of liver disease, and the risk of mortality from a single episode is around 15%-20%<sup>[18]</sup>. Thus prevention or control of variceal bleeding has been the primary aim of the drug treatments that have been used in an attempt to lower portal pressure.

In theory portal pressure should fall in response to drugs that reduce portal inflow or those that lower intra hepatic resistance to portal inflow. The drugs most widely used in the prevention of variceal bleeding are non-selective  $\beta$ -blockers (NSBB). They reduce portal pressure by reducing cardiac output *via*  $\beta$ -1 receptors and causing splanchnic vasoconstriction by blocking  $\beta$ -2 receptors, resulting in unopposed  $\alpha$ -1 activity. Randomized clinical trials showed NSBB reduce portal pressure and the risk of bleeding from oesophageal varices<sup>[18-26]</sup>. However around 15% of cirrhotic patients are intolerant of NSBB treatment, and up to 60% fail to achieve the treatment response required to prevent variceal bleeding defined as a fall in hepatic venous pressure gradient (HVPG) to less than 12 mmHg or a decrease of greater than 20% from baseline<sup>[27]</sup>. Although portal pressure is directly correlated with the presence of varices, lowering pressure with NSBB does not prevent the development of varices in patients with cirrhosis<sup>[28]</sup>.

Another approach is to reduce intrahepatic resistance with drugs that increase the delivery of NO to the intrahepatic circulation (*e.g.*, nitrates), or drugs that block  $\alpha$ -adrenergic activity (*e.g.*, prazosin, clonidine). Although modest reduction in HVPG can be achieved with these drugs, their use as monotherapy is not recommended as they not only act on the intrahepatic circulation but also exert a vasodilatory effect on the systemic circulation, leading to arterial hypotension<sup>[29]</sup>. A recent small placebo-controlled randomized study showed that simvastatin, a drug that originally developed for hypercholesterolemia and shown to act through the posttranslational modification of endothelial NOS (eNOS), significantly reduced HVPG in cirrhotic patients without altering the blood flow. This suggested that simvastatin improved HVPG by reducing intrahepatic vascular resistance<sup>[30]</sup>.

A number of vasoconstrictor drugs which increase splanchnic vascular tone have been shown to be effective in controlling acute variceal bleeding. The vasopressin analogue, terlipressin, acts on vascular V1 receptors in both the mesenteric and systemic arterial beds to mediate vasoconstriction, and as a result, the drug lowers mesenteric inflow and portal pressure. Terlipressin is generally well-tolerated, but there remains a small incidence of ischaemic events which respond to cessation of the drug<sup>[31]</sup>. This drug reduces the relative risk of mortality from acute variceal bleeding by approximately one third<sup>[32]</sup>. Moreover, terlipressin (plus albumin) is the only treatment shown to prolong short-term survival in type 1 hepatorenal syndrome (HRS)<sup>[33]</sup>. Somatostatin and its analogues octreotide and vapreotide are splanchnic

vasoconstrictors which act by inhibiting glucagon secretion and by a local mesenteric vasoconstrictive effect<sup>[34]</sup>. Although these drugs have a role in the treatment of acute variceal bleeding in combination with endoscopic therapy<sup>[35]</sup>, they do not reduce mortality in this setting compared to endoscopic therapy alone<sup>[35,36]</sup> and are ineffective in HRS<sup>[37]</sup>.

In summary, NSBBs are widely accepted as the main pharmacotherapy currently available for prevention of variceal bleeding. However, a significant proportion of patient fail to achieve an optimal response or do not tolerate treatment<sup>[27]</sup> and no other drugs have an established role in the long-term treatment of portal hypertension. Thus there remains a major need to develop more safe and effective treatment options for the treatment of portal hypertension.

## NEW CONCEPTS IN RAS PHYSIOLOGY

In recent years it has been shown that the RAS is a much more complex enzymatic pathway than previously thought. It has been long recognized that the RAS plays a central role in cardiovascular and fluid homeostasis *via* the formation of the potent vasoconstrictor angiotensin II<sup>[38]</sup>. However it is now clear that in addition to its vasoactive roles the “classical” axis of the RAS, comprising angiotensin II, angiotensin converting enzyme (ACE) and the angiotensin II type 1 receptor (AT1R), plays a role in the wound healing response to chronic tissue injury and contributes to inflammation, cell proliferation and fibrogenesis<sup>[39,42]</sup>. In addition an “alternate” axis of the RAS has been characterized comprising ACE2, a structural homologue of ACE, its peptide product angiotensin-(1-7) and the Mas receptor, which has effects that counterbalance those mediated by the classical axis (Figure 1).

Early studies showed that angiotensin-(1-7) can be generated from angiotensin I by the actions of endopeptidases such as prolyl oligopeptidase<sup>[43]</sup> and thimet oligopeptidase<sup>[44]</sup> in tissue, and in the circulation by neutral endopeptidase<sup>[45]</sup>. Whilst the various endopeptidases have been shown to produce angiotensin-(1-7) depending upon their tissue localization and access to substrates, emerging evidence suggests that ACE2 which has a distinct enzyme activity<sup>[46]</sup>, plays a key role in angiotensin-(1-7) production in several tissues. ACE2 is able to generate angiotensin-(1-7) from angiotensin I indirectly through an intermediary peptide angiotensin-(1-9); however, in comparison, ACE2 has an approximately 400-fold higher substrate preference for angiotensin II<sup>[47]</sup> which suggests that ACE2 is important not only for production of angiotensin-(1-7) but also in degrading angiotensin II. Recently, Westwood and Chappell described another pathway in which angiotensin-(1-7) is generated directly from angiotensin-(1-12) or *via* angiotensin I generation<sup>[48]</sup>.

Angiotensin-(1-7), an effector peptide of the alternate axis of the RAS, is a vasodilator in several vascular

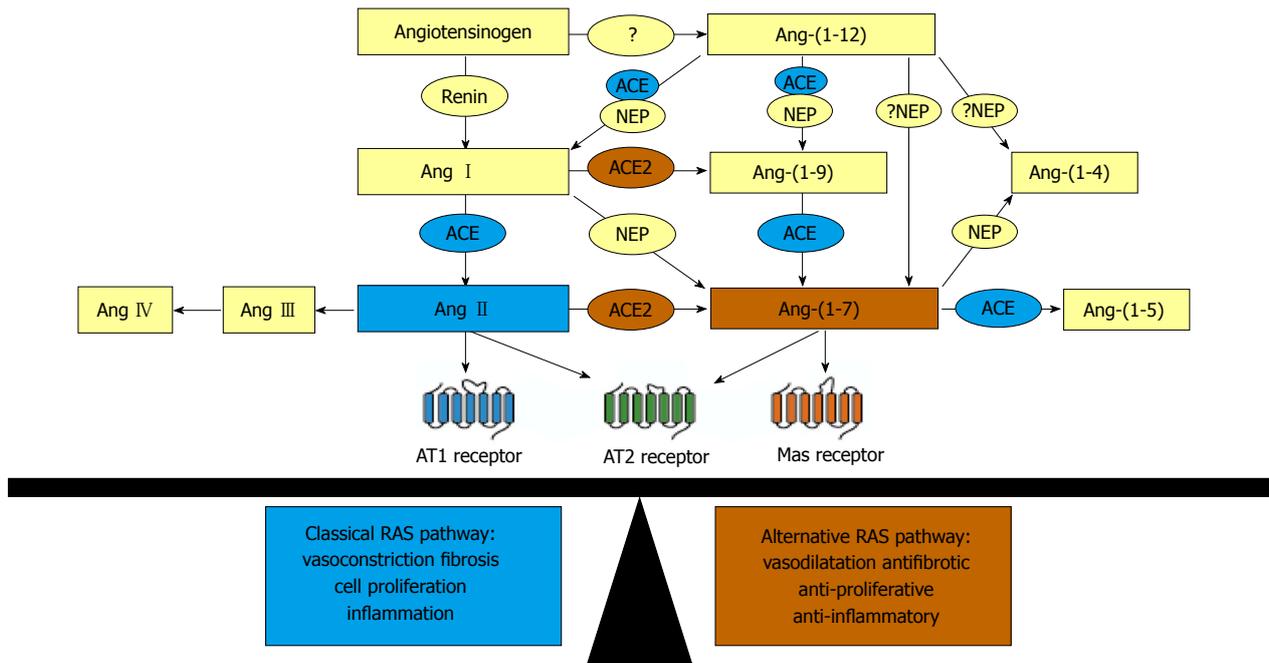
beds and has been shown to act mainly *via* its receptor Mas<sup>[49,54]</sup>. However, the existence of a receptor population that is insensitive to blockade with Mas receptor blocker A779 has also been reported<sup>[54,55]</sup>. It appears that angiotensin-(1-7), upon binding to its receptor, activates diverse pathways of intracellular signalling, leading to vasodilatation. For example, vasodilatory prostacyclin and/or NO appear to be involved in the response to angiotensin-(1-7) in the regional vascular beds<sup>[50,53,54,56-58]</sup>. It therefore appears that angiotensin-(1-7)-stimulated intracellular signaling leading to vasodilatation is depending upon the vascular bed under study and under differing pathophysiological condition.

Most components of the RAS are expressed in the liver, which is the primary source of angiotensinogen synthesis. Recent findings from our laboratory and others suggest that this intrahepatic RAS plays an important role in liver fibrosis since it is markedly upregulated in liver injury<sup>[11,59-61]</sup>, and blockade of the RAS improves experimental hepatic fibrosis<sup>[5,62-64]</sup>. The alternative axis of the RAS is also expressed in the liver and upregulated in liver disease leading to the generation of angiotensin-(1-7)<sup>[11,61,63]</sup>. The major pathway responsible for the generation of angiotensin-(1-7) in the cirrhotic liver is degradation of angiotensin II by ACE2 (Figure 2)<sup>[60]</sup> confirming previous *in vitro* findings that ACE2 has the highest substrate preference towards angiotensin II<sup>[47]</sup>. However there is limited data regarding the possible role of the alternate RAS in liver fibrosis and in modulating intrahepatic blood flow.

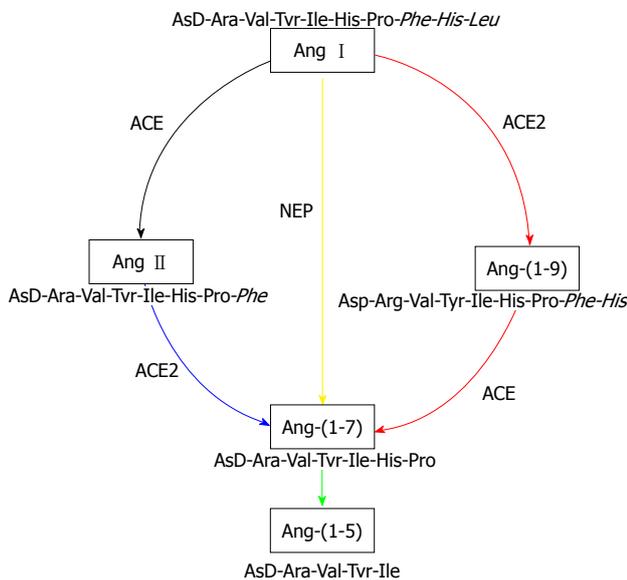
## RAS AND INTRAHEPATIC RESISTANCE

As outlined above, in patients with cirrhosis, the development of portal hypertension results from both an increase in the intrahepatic resistance to portal flow and an associated vasodilatation of the mesenteric vascular bed which leads to an increase in mesenteric blood flow. Splanchnic and systemic vasodilatation leads to secondary activation of vasoconstrictor pathways such as the sympathetic nervous system and the RAS in an attempt to maintain systemic vascular filling and blood pressure<sup>[67]</sup>. However these changes fail to correct the underlying circulatory hemodynamics<sup>[68]</sup>. There is now increasing evidence that in addition to its well recognized role in the homeostatic response to vasodilatation in cirrhosis the RAS may also play a primary role in the pathogenesis and maintenance of portal hypertension.

Hepatic structural changes such as tissue remodeling and scarring play a central role in increasing hepatic resistance to portal flow in the cirrhotic liver. However, when activated, hepatic stellate cells adopt a contractile myofibroblast phenotype, express the AT1R and have been shown *in vitro* to contract in response to angiotensin II and other vasoconstrictors such as endothelin-1<sup>[10,69]</sup>. The vasoconstriction response to angiotensin II is markedly increased in the perfused cirrhotic liver compared to normal livers, presumably mediated *via*



**Figure 1 Overview of the renin-angiotensin system.** The effects of the renin-angiotensin system (RAS) are determined by the balance between the angiotensin II (Ang II)-mediated “classical” axis, depicted in blue, which is vasoconstrictive and the angiotensin-(1-7) [Ang-(1-7)]-mediated “alternate” axis, depicted in orange, which is vasodilatory. Both Ang II and Ang-(1-7) can stimulate the Ang II type 2 (AT2) receptor, depicted in green; the effects of which are often analogous to those mediated by the Ang-(1-7) receptor Mas. Recent evidence indicates that a new member, Ang-(1-12), which is cleaved from angiotensinogen, also contributes either indirectly via Ang I or directly to the pool of Ang-(1-7). NEP: Neutral endopeptidase; ACE: Angiotensin converting enzyme; ACE2: Angiotensin converting enzyme 2; AT1 receptor: Angiotensin II type 1 receptor.

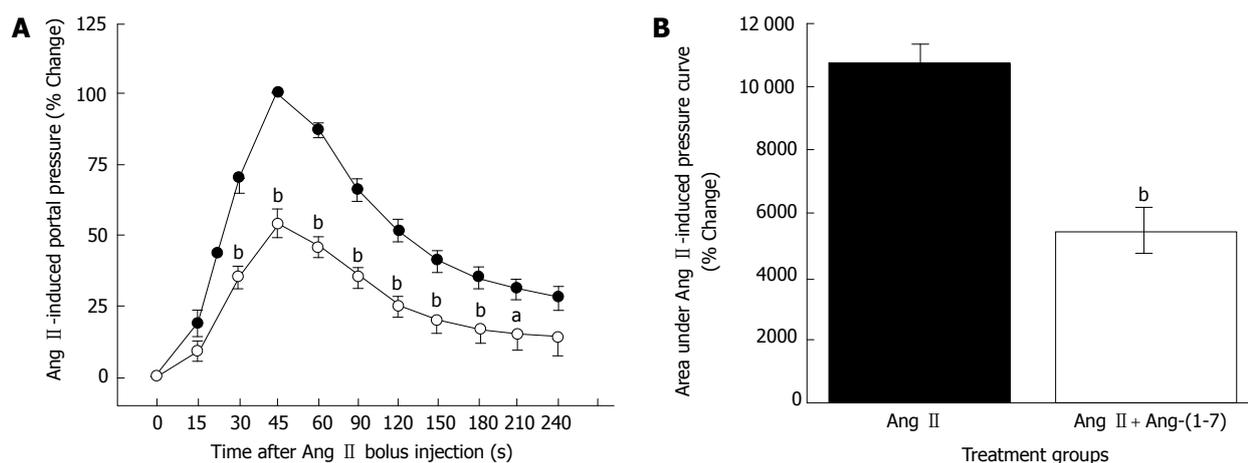


**Figure 2 Intrahepatic enzymatic pathways of renin-angiotensin system peptide production.** Schematic representation of the pathways responsible for the generation of vasodilator peptide angiotensin-(1-7) [Ang-(1-7)] in rat liver. The colour of the arrows represents the relative contribution of each pathway for *ex vivo* formation of Ang-(1-7) from the substrates angiotensin I (Ang I) and angiotensin II (Ang II) in cirrhotic rat liver. The most efficient pathway is represented by blue line, followed by green line. The black line is the least efficient pathway whilst red represents the efficiency less than green but higher than black. The yellow colour represents an efficient pathway to generate Ang-(1-7) directly from Ang I by the action of neutral endopeptidase (NEP) but it appears that this pathway is masked by angiotensin converting enzyme 2 in cirrhotic rat liver. Ang-(1-9): Angiotensin-(1-9); Ang-(1-5): Angiotensin-(1-5); ACE: Angiotensin converting enzyme; ACE2: Angiotensin converting enzyme 2; NEP: Neutral endopeptidase. Figure was adapted from our previous publication.

upregulated AT1R and AT1R expressing perivascular myofibroblasts<sup>[66]</sup>. Furthermore intrahepatic angiotensin II generation is increased in the cirrhotic liver<sup>[66]</sup>. These findings provide a rationale for the use of RAS blockers in the management of portal hypertension.

There is considerable evidence that another important contributor to elevated vascular tone in the cirrhotic liver is endothelial dysfunction of the hepatic microcirculation which diminishes the response to vasodilators<sup>[70]</sup>. It has been proposed that the reduced activity of hepatic vascular eNOS with concomitant reduction in NO synthesis impairs intrahepatic vasodilatation and thus, shifts the balance towards vasoconstriction<sup>[71]</sup>. This reduction in eNOS activity is linked to an increase in the expression of caveolin, a protein which is highly expressed in endothelial cells of the hepatic vasculature with predominant expression found in venous and sinusoidal endothelial cells in cirrhotic livers<sup>[72,73]</sup>. Interestingly, the calcium binding protein calmodulin competitively binds eNOS and reduces caveolin binding, thus increasing eNOS activity<sup>[72]</sup>.

Recent findings from our laboratory demonstrated that in *in-situ* perfused cirrhotic rat liver elicited a marked endothelium-dependent vasodilatory effect of exogenous angiotensin-(1-7) on the vasoconstrictive response evoked by angiotensin II (Figure 3)<sup>[55]</sup>. This finding suggests that as in other vascular beds<sup>[50,51,53,54]</sup>, in the cirrhotic liver angiotensin-(1-7) may cause a vasodilatory response that antagonizes the increase in portal pressure mediated by angiotensin II and other local vasoconstrictors. Although eNOS activity was not measured in this



**Figure 3 Angiotensin-(1-7)-induced portal pressure reduction.** Portal pressure changes in response to angiotensin II (Ang II) in cirrhotic rat liver. Portal pressure responses were measured using a vertically positioned graduated fluid-filled column open to atmospheric pressure. Ang II bolus (60 pmole) was injected into the portal vein of *in-situ* perfused cirrhotic rat liver preparations in the presence of an angiotensin converting enzyme (ACE) inhibitor lisinopril (0.7  $\mu$ mol/L). A: Percentage increases in pressure in response to Ang II bolus in the absence (closed circles) or presence (open circles) of angiotensin-(1-7) [Ang-(1-7), 0.7  $\mu$ mol/L]. The highest pressure that was recorded 45 s after the Ang II bolus injection was designated as 100% response and all other responses were calculated relative to this maximal response. B: The total area under Ang II response curve (AUC) in the absence (filled bar) or presence (open bar) of Ang-(1-7). Portal pressure change at each time point in panel A and AUC in B represents the mean  $\pm$  SEM from 21 cirrhotic rat liver preparations. Pre-incubation with Ang-(1-7) significantly reduced the portal pressure response evoked by Ang II bolus injection. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$  vs those with Ang II alone.

study, the eNOS inhibitor nitro-L-arginine methyl ester, L-NAME, completely abolished eNOS phosphorylation at Ser1177 and the response to angiotensin-(1-7)<sup>[55]</sup>. Whilst bradykinin *via* its B2 receptor mediates vasodilatation in response to angiotensin-(1-7) in porcine and canine coronary arteries<sup>[56,58,74]</sup>, it increases intrahepatic resistance and portal pressure<sup>[75]</sup>, possibly through acting on B2 receptors on stellate cells. However, angiotensin-(1-7)-induced vasodilatation in the cirrhotic liver was not affected by bradykinin B2 receptor blockade<sup>[55]</sup>. Possible mechanisms for these effects of angiotensin-(1-7) in the cirrhotic rat liver include increased phosphorylation of eNOS Ser1177<sup>[55]</sup> with simultaneous dephosphorylation at Thr495 and/or effects on calmodulin binding<sup>[76]</sup>. These findings suggest that it may be possible to reduce intrahepatic resistance and portal pressure by targeting the alternate axis of the RAS in the liver.

## RAS AND SPLANCHNIC VASODILATION

In contrast to intrahepatic hypervascular tone, the systemic and splanchnic circulation in cirrhosis is characterized by vasodilatation and hyporesponsiveness to vasoconstrictors including angiotensin II<sup>[8,67,77]</sup>. Interestingly, recent studies have shown that systemic levels of the vasodilatory peptide angiotensin-(1-7) increase as liver fibrosis progresses, whereas angiotensin II levels do not generally rise until cirrhosis is established with concurrent portal hypertension<sup>[65,78]</sup>. Furthermore, regional levels of the hormone are different from systemic levels such that in cirrhotic patients at transplant, the angiotensin-(1-7)/angiotensin II ratio is elevated in the splanchnic compared to the peripheral circulation, and negatively cor-

relates with systemic vascular resistance<sup>[78]</sup>. The recent findings suggest that angiotensin-(1-7) might contribute to vasodilatation in cirrhosis. Recent work from our laboratory provides support for this hypothesis<sup>[79]</sup>. We have shown that ACE2 is upregulated in cirrhotic mesenteric vessels and although angiotensin-(1-7) has no effect in the normal mesenteric circulation, it significantly reduces mesenteric vascular contractility in cirrhotic mesenteric beds *via* activation of the Mas receptor and the release of NO<sup>[79]</sup>.

Studies using isolated vessel preparations from cirrhotic animal models or portal hypertensive rats have led to the concept that vasodilatation is also linked to an intrinsic vascular hyporesponsiveness to endogenous vasoconstrictors such as angiotensin II,  $\alpha$ -adrenergic agonists and endothelin-1<sup>[80-82]</sup>. This concept is supported by the findings that peripheral vessels are hyporeactive to angiotensin II,  $\alpha$ -adrenergic agonists and endothelin-1 from cirrhotic animals and cirrhotic patients<sup>[77,83-90]</sup>, despite the fact that expression of AT1R and  $\alpha$ -adrenergic receptor subtypes 1a and 1b in the peripheral vessels is either normal or upregulated in both cirrhotic animals and patients<sup>[79,87,91]</sup>. However, previous studies in patients with cirrhosis or in peripheral resistance vessels obtained from such patients reported variable results in this regard<sup>[84,90,92]</sup>, probably attributable to the differences between conditions in different studies. Indeed, small resistance omental vessels from cirrhotic patients had a larger vasoconstriction response to  $\alpha$ -adrenergic agonists norepinephrine and methoxamine than similar vessels from healthy controls<sup>[92]</sup>. The same vessels vasodilated in response to substance P but this was inhibited by blocking NO or prostacyclin synthesis, suggesting that intrinsic hyporeactivity that is present in the peripheral circulation in cirrhosis is related to increased levels of NO

and prostanoids<sup>[93-95]</sup>. Hyporeactivity to angiotensin II infusion is also improved after inhibiting systemic NO production using an NO-clamp in cirrhotic patients<sup>[85]</sup>. This is in keeping with a wealth of literature suggesting that vasodilatory molecules including angiotensin-(1-7) are produced in excess in cirrhosis and that the final *in vivo* pressor effect is governed by a balance between the pressor and depressor arms of the circulation<sup>[96]</sup>.

There are also data, from both *in vitro* and *in vivo* studies, suggesting that vascular hyporeactivity to a range of endogenous pressors is attributed to changes that are downstream of the G-protein coupled receptors<sup>[86-88]</sup>. Evidence supporting the existence of vascular endothelium/NO independent pathways comes from studies in which endothelium denudation and pharmacological blockade of NOS in isolated vessels from cirrhotic animals and patients failed to improve vascular hyporeactivity to a range of vasoconstrictors<sup>[83,84,86,87,97-99]</sup>. One of the important NO-independent pathways is an impaired signaling by RhoA and Rho kinase, leading to a decreased phosphorylation of Ca<sup>2+</sup>-sensitizing proteins and increased myosin light chain phosphatase activity<sup>[87]</sup>. Moreover, increased expression of receptor desensitizing proteins, G protein-coupled receptor kinase 2 and  $\beta$ -arrestin-2, have also been implicated in this hyporeactivity to angiotensin II in vessels isolated from cirrhotic patients and rats<sup>[87]</sup>. It was also shown that mesenteric arteries from portal hypertensive rats had a reduced level of membrane associated RhoA, probably reflecting a diminished activity of RhoA/Rho kinase pathway which in turn results in increased activity of myosin light chain phosphatase and vasodilatation<sup>[68,100]</sup>.

## THERAPIES TARGETING THE RAS IN PORTAL HYPERTENSION

### **Therapeutic potential of the classical RAS**

The evidence from studies in experimental cirrhosis showing the angiotensin II contributes to the variable component of intrahepatic resistance in portal hypertension have provided a rationale for a number of trials examining the effects of ACE inhibitors and angiotensin receptors blockers (ARBs)<sup>[5,23,87]</sup> on portal pressure. Unfortunately many of these studies are small or non-randomized and there is very little long-term data. However Tandon *et al*<sup>[23]</sup> in a recent meta-analysis of individual patient data from three and nine studies that used ACE inhibitors and ARBs, respectively, showed that patients with Child Pugh A cirrhosis receiving ARBs/ACE inhibitors had a similar reduction in HVPG (17%) compared to patients with Pugh A cirrhosis that received NSBB (21%). There was no improvement of HVPG in patients with Child Pugh B/C cirrhosis receiving ARBs. Furthermore, several studies reported that RAS blockade can result in significant hypotension and renal impairment in patients with decompensated (Child Pugh B/C) cirrhosis<sup>[23]</sup> in whom there is activation of the systemic RAS.

Thus, although ARBs/ACE inhibitors do lower por-

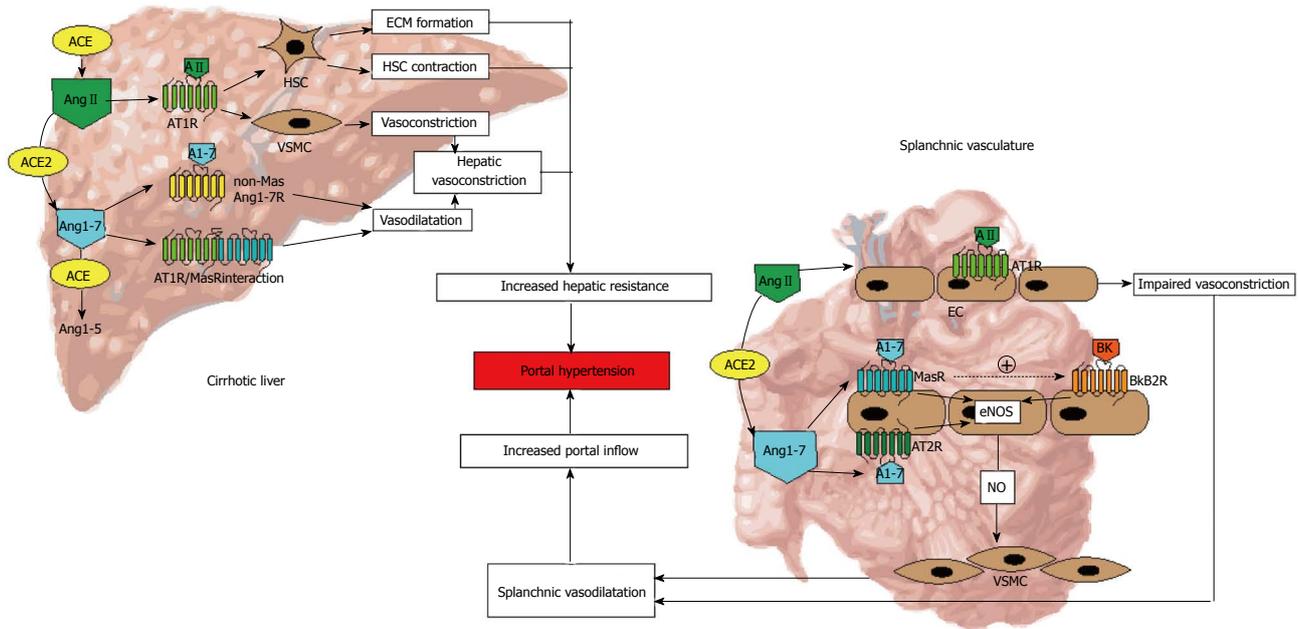
tal pressure in early cirrhotic patients where the activation of RAS may be a predominant pathway responsible for increased intrahepatic tone, they have less effect in late stages of cirrhosis. This probably reflects the fact that the hypotension induced by RAS blockers increases activation of other vasoconstrictive pathways such as the sympathetic nervous system that in turn increase intrahepatic vascular tone<sup>[28,101-103]</sup>. Further studies are needed to clarify whether this class of drugs could be useful in the prevention of variceal bleeding in patients with compensated cirrhosis as an alternative to or possibly even in combination with  $\beta$ -blockers.

### **Alternate RAS-a novel potential target for the treatment of portal hypertension**

Recent animal studies focusing on the alternate RAS have led to the suggestion that new generation antihypertensives developed to target this axis could serve as effective therapeutic agents to treat arterial, pulmonary and portal hypertension<sup>[1,65,104,105]</sup>. Recent work outlined in this review demonstrates the presence of all of the key components of the alternate RAS in the liver and mesenteric vasculature of both healthy and cirrhotic animals as well as in the liver of healthy and cirrhotic patients<sup>[11,61,66,79]</sup>. Furthermore circulating angiotensin-(1-7) levels are increased<sup>[11,65,78]</sup> and the system is upregulated in the liver and mesenteric circulation in cirrhosis suggesting that it plays an important role in the pathophysiology of hepatic fibrosis and portal hypertension (Figure 4). This evidence linking elevated angiotensin-(1-7) levels to mesenteric and systemic vasodilatation in cirrhosis suggests that blocking the alternate axis could reduce mesenteric flow and thus lower portal pressure. In line with this, Mas receptor blockade would provide an intervention option in portal hypertension<sup>[79]</sup> as this treatment regime does not appear to compromise the vasodilatory response of angiotensin-(1-7) within the hepatic vasculature in experimental cirrhosis<sup>[55]</sup>. Further studies are clearly needed examining the effects of Mas blockade and angiotensin-(1-7) on hepatic and mesenteric haemodynamics in experimental cirrhosis *in vivo*.

## CONCLUSION

Recent developments in our understanding of the complexities of the RAS and its role in the pathogenesis of chronic liver disease and portal hypertension have opened up new therapeutic possibilities. It is clear that the classical axis of the RAS and its key effector peptide angiotensin II play a central role in hepatic fibrogenesis and in regulating intrahepatic vascular tone in cirrhosis and that despite the mixed results achieved in previous trials, consideration should be given to further prospective studies examining the effects of RAS blockers in patients with compensated cirrhosis. There is also fascinating new evidence showing that there is increased regional production of angiotensin-(1-7) in the mesenteric vascular bed in cirrhosis, and that this vasodilatory



**Figure 4** Overview of the renin-angiotensin system-mediated pathophysiological changes in portal hypertension. In cirrhosis, the effects of classical axis of the renin-angiotensin system (RAS), mediated by its potent vasoconstrictor peptide angiotensin II (Ang II), are predominant within the hepatic vasculature, resulting in increased hepatic resistance to portal inflow. In this, apart from fixed barrier due to increased deposition of extracellular matrix (ECM) proteins, vascular tone is exacerbated by Ang II action on myfibroblastic cells (activated hepatic stellate cells-HSCs) and vascular smooth muscle cells (VSMCs). In contrast, the effects of the alternate axis of the RAS, mediated by its vasodilator peptide angiotensin-(1-7) (Ang1-7), are predominant within the splanchnic vasculature, resulting in increased nitric oxide (NO) production by vascular endothelial cells (ECs). This consequently exacerbates portal hypertension as a result of increased inflow to the portal circulation. Ang1-5: Angiotensin-(1-5); ACE: Angiotensin converting enzyme; ACE2: Angiotensin converting enzyme 2; eNOS: Endothelial nitric oxide synthase; Bk: Bradykinin; BkB2R: Bradykinin B2 receptor; AT1R: Angiotensin II type 1 receptor; AT2R: Angiotensin II type 2 receptor; MasR: Mas receptor.

peptide of the alternate axis of the RAS, contributes to mesenteric vasodilatation and the hyperdynamic circulation in cirrhosis. These novel data suggest that ACE2-angiotensin-(1-7)-Mas receptor axis is a potential target for the management of portal hypertension.

## REFERENCES

- Ferrario CM, Trask AJ, Jessup JA. Advances in biochemical and functional roles of angiotensin-converting enzyme 2 and angiotensin-(1-7) in regulation of cardiovascular function. *Am J Physiol Heart Circ Physiol* 2005; **289**: H2281-H2290 [PMID: 16055515 DOI: 10.1152/ajpheart.00618.2005]
- Bataller R, Brenner DA. Liver fibrosis. *J Clin Invest* 2005; **115**: 209-218 [PMID: 15690074 DOI: 10.1172/JCI200524282]
- Töx U, Steffen HM. Impact of inhibitors of the Renin-Angiotensin-aldosterone system on liver fibrosis and portal hypertension. *Curr Med Chem* 2006; **13**: 3649-3661 [PMID: 17168728 DOI: 10.2174/092986706779026138]
- Dickstein K, Kjekshus J. Effects of losartan and captopril on mortality and morbidity in high-risk patients after acute myocardial infarction: the OPTIMAAL randomised trial. *Lancet* 2002; **360**: 752-760 [PMID: 12241832 DOI: 10.1016/S0140-6736(02)09895-1]
- Paizis G, Gilbert RE, Cooper ME, Murthi P, Schembri JM, Wu LL, Rumble JR, Kelly DJ, Tikellis C, Cox A, Smallwood RA, Angus PW. Effect of angiotensin II type 1 receptor blockade on experimental hepatic fibrogenesis. *J Hepatol* 2001; **35**: 376-385 [PMID: 11592599 DOI: 10.1016/S0168-8278(01)00146-5]
- Parving HH, Persson F, Lewis JB, Lewis EJ, Hollenberg NK. Aliskiren combined with losartan in type 2 diabetes and nephropathy. *N Engl J Med* 2008; **358**: 2433-2446 [PMID: 18525041 DOI: 10.1056/NEJMoa0708379]
- Pinzani M, Gentilini P. Biology of hepatic stellate cells and their possible relevance in the pathogenesis of portal hypertension in cirrhosis. *Semin Liver Dis* 1999; **19**: 397-410 [PMID: 10643625 DOI: 10.1055/s-2007-1007128]
- Angus PW. Role of endothelin in systemic and portal resistance in cirrhosis. *Gut* 2006; **55**: 1230-1232 [PMID: 16905694 DOI: 10.1136/gut.2005.088633]
- Bataller R, Gäbele E, Schoonhoven R, Morris T, Lehnert M, Yang L, Brenner DA, Rippe RA. Prolonged infusion of angiotensin II into normal rats induces stellate cell activation and proinflammatory events in liver. *Am J Physiol Gastrointest Liver Physiol* 2003; **285**: G642-G651 [PMID: 12773299]
- Bataller R, Gines P, Nicolas JM, Gorbis MN, Garcia-Ramallo E, Gasull X, Bosch J, Arroyo V, Rodes J. Angiotensin II induces contraction and proliferation of human hepatic stellate cells. *Gastroenterology* 2000; **118**: 1149-1156 [PMID: 10833490 DOI: 10.1016/S0016-5085(00)70368-4]
- Herath CB, Warner FJ, Lubel JS, Dean RG, Jia Z, Lew RA, Smith AI, Burrell LM, Angus PW. Upregulation of hepatic angiotensin-converting enzyme 2 (ACE2) and angiotensin-(1-7) levels in experimental biliary fibrosis. *J Hepatol* 2007; **47**: 387-395 [PMID: 17532087 DOI: 10.1016/j.jhep.2007.03.008]
- Wiest R, Groszmann RJ. Nitric oxide and portal hypertension: its role in the regulation of intrahepatic and splanchnic vascular resistance. *Semin Liver Dis* 1999; **19**: 411-426 [PMID: 10643626 DOI: 10.1055/s-2007-1007129]
- Albornoz L, Motta A, Alvarez D, Estevez A, Bandi JC, McCormack L, Matera J, Bonofiglio C, Ciardullo M, De Santibanes E, Gimeno M, Gadan A. Nitric oxide synthase activity in the splanchnic vasculature of patients with cirrhosis: relationship with hemodynamic disturbances. *J Hepatol* 2001; **35**: 452-456 [PMID: 11682028 DOI: 10.1016/S0168-8278(01)00168-4]
- Sarela AI, Mihaimeed FM, Batten JJ, Davidson BR, Mathie

- RT. Hepatic and splanchnic nitric oxide activity in patients with cirrhosis. *Gut* 1999; **44**: 749-753 [PMID: 10205218 DOI: 10.1136/gut.44.5.749]
- 15 **Sieber CC**, Lopez-Talavera JC, Groszmann RJ. Role of nitric oxide in the in vitro splanchnic vascular hyporeactivity in ascitic cirrhotic rats. *Gastroenterology* 1993; **104**: 1750-1754 [PMID: 8388839]
  - 16 **Wiest R**, Shah V, Sessa WC, Groszmann RJ. NO overproduction by eNOS precedes hyperdynamic splanchnic circulation in portal hypertensive rats. *Am J Physiol* 1999; **276**: G1043-G1051 [PMID: 10198349]
  - 17 **Fernandez M**, Vizzutti F, Garcia-Pagan JC, Rodes J, Bosch J. Anti-VEGF receptor-2 monoclonal antibody prevents portal-systemic collateral vessel formation in portal hypertensive mice. *Gastroenterology* 2004; **126**: 886-894 [PMID: 14988842 DOI: 10.1053/j.gastro.2003.12.012]
  - 18 **D'Amico G**, Pagliaro L, Bosch J. The treatment of portal hypertension: a meta-analytic review. *Hepatology* 1995; **22**: 332-354 [PMID: 7601427 DOI: 10.1002/hep.1840220145]
  - 19 **García-Pagán JC**, Villanueva C, Albillos A, Bañares R, Morillas R, Abraldes JG, Bosch J. Nadolol plus isosorbide mononitrate alone or associated with band ligation in the prevention of recurrent bleeding: a multicentre randomised controlled trial. *Gut* 2009; **58**: 1144-1150 [PMID: 19218249]
  - 20 **Groszmann RJ**, Bosch J, Grace ND, Conn HO, Garcia-Tsao G, Navasa M, Alberts J, Rodes J, Fischer R, Bermann M. Hemodynamic events in a prospective randomized trial of propranolol versus placebo in the prevention of a first variceal hemorrhage. *Gastroenterology* 1990; **99**: 1401-1407 [PMID: 2210246]
  - 21 **Pagliaro L**, D'Amico G, Sørensen TI, Lebrec D, Burroughs AK, Morabito A, Tiné F, Politi F, Traina M. Prevention of first bleeding in cirrhosis. A meta-analysis of randomized trials of nonsurgical treatment. *Ann Intern Med* 1992; **117**: 59-70 [PMID: 1350716]
  - 22 **Patch D**, Sabin CA, Goulis J, Gerunda G, Greenslade L, Merkel C, Burroughs AK. A randomized, controlled trial of medical therapy versus endoscopic ligation for the prevention of variceal rebleeding in patients with cirrhosis. *Gastroenterology* 2002; **123**: 1013-1019 [PMID: 12360462 DOI: 10.1053/gast.2002.35955]
  - 23 **Tandon P**, Abraldes JG, Berzigotti A, Garcia-Pagan JC, Bosch J. Renin-angiotensin-aldosterone inhibitors in the reduction of portal pressure: a systematic review and meta-analysis. *J Hepatol* 2010; **53**: 273-282 [PMID: 20570385 DOI: 10.1016/j.jhep.2010.03.013]
  - 24 **Villanueva C**, Aracil C, Colomo A, Lopez-Balaguer JM, Piqueras M, Gonzalez B, Torras X, Guarner C, Balanzo J. Clinical trial: a randomized controlled study on prevention of variceal rebleeding comparing nadolol + ligation vs. hepatic venous pressure gradient-guided pharmacological therapy. *Aliment Pharmacol Ther* 2009; **29**: 397-408 [PMID: 19006538 DOI: 10.1111/j.1365-2036.2008.03880.x]
  - 25 **Villanueva C**, Balanzó J, Novella MT, Soriano G, Sáinz S, Torras X, Cussó X, Guarner C, Vilardell F. Nadolol plus isosorbide mononitrate compared with sclerotherapy for the prevention of variceal rebleeding. *N Engl J Med* 1996; **334**: 1624-1629 [PMID: 8628357 DOI: 10.1056/NEJM199606203342502]
  - 26 **Villanueva C**, Miñana J, Ortiz J, Gallego A, Soriano G, Torras X, Sáinz S, Boadas J, Cussó X, Guarner C, Balanzó J. Endoscopic ligation compared with combined treatment with nadolol and isosorbide mononitrate to prevent recurrent variceal bleeding. *N Engl J Med* 2001; **345**: 647-655 [PMID: 11547718 DOI: 10.1056/NEJMoa003223]
  - 27 **Feu F**, Garcia-Pagan JC, Bosch J, Luca A, Teres J, Escorsell A, Rodes J. Relation between portal pressure response to pharmacotherapy and risk of recurrent variceal haemorrhage in patients with cirrhosis. *Lancet* 1995; **346**: 1056-1059 [PMID: 7564785 DOI: 10.1016/S0140-6736(95)91740-3]
  - 28 **Groszmann RJ**, Garcia-Tsao G, Bosch J, Grace ND, Burroughs AK, Planas R, Escorsell A, Garcia-Pagan JC, Patch D, Matloff DS, Gao H, Makuch R. Beta-blockers to prevent gastroesophageal varices in patients with cirrhosis. *N Engl J Med* 2005; **353**: 2254-2261 [PMID: 16306522 DOI: 10.1056/NEJMoa044456]
  - 29 **García-Tsao G**. Current management of the complications of cirrhosis and portal hypertension: variceal hemorrhage, ascites, and spontaneous bacterial peritonitis. *Gastroenterology* 2001; **120**: 726-748 [PMID: 11179247 DOI: 10.1053/gast.2001.22580]
  - 30 **Abraldes JG**, Albillos A, Bañares R, Turnes J, González R, García-Pagán JC, Bosch J. Simvastatin lowers portal pressure in patients with cirrhosis and portal hypertension: a randomized controlled trial. *Gastroenterology* 2009; **136**: 1651-1658 [PMID: 19208350 DOI: 10.1053/j.gastro.2009.01.043]
  - 31 **Testro AG**, Wongseelashote S, Angus PW, Gow PJ. Long-term outcome of patients treated with terlipressin for types 1 and 2 hepatorenal syndrome. *J Gastroenterol Hepatol* 2008; **23**: 1535-1540 [PMID: 17784863 DOI: 10.1111/j.1440-1746.2007.05176.x]
  - 32 **Ioannou G**, Doust J, Rockey DC. Terlipressin for acute esophageal variceal hemorrhage. *Cochrane Database Syst Rev* 2003; (1): CD002147 [PMID: 12535432]
  - 33 **Gluud LL**, Christensen K, Christensen E, Krag A. Systematic review of randomized trials on vasoconstrictor drugs for hepatorenal syndrome. *Hepatology* 2010; **51**: 576-584 [PMID: 19885875 DOI: 10.1002/hep.23286]
  - 34 **Wiest R**, Tsai MH, Groszmann RJ. Octreotide potentiates PKC-dependent vasoconstrictors in portal-hypertensive and control rats. *Gastroenterology* 2001; **120**: 975-983 [PMID: 11231951 DOI: 10.1053/gast.2001.22529]
  - 35 **Bañares R**, Albillos A, Rincón D, Alonso S, González M, Ruiz-del-Arbol L, Salcedo M, Molinero LM. Endoscopic treatment versus endoscopic plus pharmacologic treatment for acute variceal bleeding: a meta-analysis. *Hepatology* 2002; **35**: 609-615 [PMID: 11870374 DOI: 10.1053/jhep.2002.31354]
  - 36 **D'Amico G**, Politi F, D'Antonio A, Giannuoli G, Pasta L, Vizziani G, Traina M, Morabito A, Pagliaro L. Second study shows that octreotide may prevent early rebleeding in cirrhosis. *BMJ* 1998; **316**: 1320 [PMID: 9554914 DOI: 10.1136/bmj.316.7140.1320]
  - 37 **Pomier-Layrargues G**, Paquin SC, Hassoun Z, Lafortune M, Tran A. Octreotide in hepatorenal syndrome: a randomized, double-blind, placebo-controlled, crossover study. *Hepatology* 2003; **38**: 238-243 [PMID: 12830007 DOI: 10.1053/jhep.2003.50276]
  - 38 **Lubel JS**, Herath CB, Burrell LM, Angus PW. Liver disease and the renin-angiotensin system: recent discoveries and clinical implications. *J Gastroenterol Hepatol* 2008; **23**: 1327-1338 [PMID: 18557800 DOI: 10.1111/j.1440-1746.2008.05461.x]
  - 39 **Kagami S**, Border WA, Miller DE, Noble NA. Angiotensin II stimulates extracellular matrix protein synthesis through induction of transforming growth factor-beta expression in rat glomerular mesangial cells. *J Clin Invest* 1994; **93**: 2431-2437 [PMID: 8200978 DOI: 10.1172/JCI117251]
  - 40 **Marrero MB**, Schieffer B, Paxton WG, Heerdt L, Berk BC, Delafontaine P, Bernstein KE. Direct stimulation of Jak/STAT pathway by the angiotensin II AT1 receptor. *Nature* 1995; **375**: 247-250 [PMID: 7746328 DOI: 10.1038/375247a0]
  - 41 **Mezzano SA**, Ruiz-Ortega M, Egido J. Angiotensin II and renal fibrosis. *Hypertension* 2001; **38**: 635-638 [PMID: 11566946 DOI: 10.1161/hy09t1.094234]
  - 42 **Suzuki Y**, Ruiz-Ortega M, Lorenzo O, Ruperez M, Esteban V, Egido J. Inflammation and angiotensin II. *Int J Biochem Cell Biol* 2003; **35**: 881-900 [PMID: 12676174 DOI: 10.1016/S1357-2725(02)00271-6]
  - 43 **Santos RA**, Brosnihan KB, Jacobsen DW, DiCorleto PE, Ferrario CM. Production of angiotensin-(1-7) by human vascular endothelium. *Hypertension* 1992; **19**: II56-II61 [PMID: 1310484 DOI: 10.1161/01.HYP.19.2\_Suppl.II56]

- 44 **Chappell MC**, Gomez MN, Pirro NT, Ferrario CM. Release of angiotensin-(1-7) from the rat hindlimb: influence of angiotensin-converting enzyme inhibition. *Hypertension* 2000; **35**: 348-352 [PMID: 10642323 DOI: 10.1161/01.HYP.35.1.348]
- 45 **Yamamoto K**, Chappell MC, Brosnihan KB, Ferrario CM. In vivo metabolism of angiotensin I by neutral endopeptidase (EC 3.4.24.11) in spontaneously hypertensive rats. *Hypertension* 1992; **19**: 692-696 [PMID: 1317352 DOI: 10.1161/01.HYP.19.6.692]
- 46 **Warner FJ**, Smith AI, Hooper NM, Turner AJ. Angiotensin-converting enzyme-2: a molecular and cellular perspective. *Cell Mol Life Sci* 2004; **61**: 2704-2713 [PMID: 15549171 DOI: 10.1007/s00018-004-4240-7]
- 47 **Vickers C**, Hales P, Kaushik V, Dick L, Gavin J, Tang J, Godbout K, Parsons T, Baronas E, Hsieh F, Acton S, Patane M, Nichols A, Tummino P. Hydrolysis of biological peptides by human angiotensin-converting enzyme-related carboxypeptidase. *J Biol Chem* 2002; **277**: 14838-14843 [PMID: 11815627 DOI: 10.1074/jbc.M200581200]
- 48 **Westwood BM**, Chappell MC. Divergent pathways for the angiotensin-(1-12) metabolism in the rat circulation and kidney. *Peptides* 2012; **35**: 190-195 [PMID: 22490446 DOI: 10.1016/j.peptides.2012.03.025]
- 49 **Benter IF**, Yousif MH, Anim JT, Cojocel C, Diz DI. Angiotensin-(1-7) prevents development of severe hypertension and end-organ damage in spontaneously hypertensive rats treated with L-NAME. *Am J Physiol Heart Circ Physiol* 2006; **290**: H684-H691 [PMID: 16403946 DOI: 10.1152/ajp-heart.00632.2005]
- 50 **Castro CH**, Santos RA, Ferreira AJ, Bader M, Alenina N, Almeida AP. Evidence for a functional interaction of the angiotensin-(1-7) receptor Mas with AT1 and AT2 receptors in the mouse heart. *Hypertension* 2005; **46**: 937-942 [PMID: 16157793 DOI: 10.1161/01.HYP.0000175813.04375.8a]
- 51 **Lemos VS**, Silva DM, Walther T, Alenina N, Bader M, Santos RA. The endothelium-dependent vasodilator effect of the nonpeptide Ang(1-7) mimic AVE 0991 is abolished in the aorta of mas-knockout mice. *J Cardiovasc Pharmacol* 2005; **46**: 274-279 [PMID: 16116331 DOI: 10.1097/01.fjc.0000175237.41573.63]
- 52 **Neves LA**, Williams AF, Averill DB, Ferrario CM, Walkup MP, Brosnihan KB. Pregnancy enhances the angiotensin (Ang)-(1-7) vasodilator response in mesenteric arteries and increases the renal concentration and urinary excretion of Ang-(1-7). *Endocrinology* 2003; **144**: 3338-3343 [PMID: 12865311 DOI: 10.1210/en.2003-0009]
- 53 **Santos RA**, Simoes e Silva AC, Maric C, Silva DM, Machado RP, de Buhr I, Heringer-Walther S, Pinheiro SV, Lopes MT, Bader M, Mendes EP, Lemos VS, Campagnole-Santos MJ, Schultheiss HP, Speth R, Walther T. Angiotensin-(1-7) is an endogenous ligand for the G protein-coupled receptor Mas. *Proc Natl Acad Sci USA* 2003; **100**: 8258-8263 [PMID: 12829792 DOI: 10.1073/pnas.1432869100]
- 54 **Silva DM**, Vianna HR, Cortes SF, Campagnole-Santos MJ, Santos RA, Lemos VS. Evidence for a new angiotensin-(1-7) receptor subtype in the aorta of Sprague-Dawley rats. *Peptides* 2007; **28**: 702-707 [PMID: 17129638 DOI: 10.1016/j.peptides.2006.10.007]
- 55 **Herath CB**, Mak K, Burrell LM, Angus PW. Angiotensin-(1-7) reduces the perfusion pressure response to angiotensin II and methoxamine via an endothelial nitric oxide-mediated pathway in cirrhotic rat liver. *Am J Physiol Gastrointest Liver Physiol* 2013; **304**: G99-G108 [PMID: 23086915 DOI: 10.1152/ajpgi.00163.2012]
- 56 **Brosnihan KB**, Li P, Ferrario CM. Angiotensin-(1-7) dilates canine coronary arteries through kinins and nitric oxide. *Hypertension* 1996; **27**: 523-528 [PMID: 8613197 DOI: 10.1161/01.HYP.27.3.523]
- 57 **Ferrario CM**, Chappell MC, Tallant EA, Brosnihan KB, Diz DI. Counterregulatory actions of angiotensin-(1-7). *Hypertension* 1997; **30**: 535-541 [PMID: 9322978 DOI: 10.1161/01.HYP.30.3.535]
- 58 **Pörsti I**, Bara AT, Busse R, Hecker M. Release of nitric oxide by angiotensin-(1-7) from porcine coronary endothelium: implications for a novel angiotensin receptor. *Br J Pharmacol* 1994; **111**: 652-654 [PMID: 8019744 DOI: 10.1111/j.1476-5381.1994.tb14787.x]
- 59 **Bataller R**, Gäbele E, Parsons CJ, Morris T, Yang L, Schoonhoven R, Brenner DA, Rippe RA. Systemic infusion of angiotensin II exacerbates liver fibrosis in bile duct-ligated rats. *Hepatology* 2005; **41**: 1046-1055 [PMID: 15841463 DOI: 10.1002/hep.20665]
- 60 **Paizis G**, Cooper ME, Schembri JM, Tikellis C, Burrell LM, Angus PW. Up-regulation of components of the renin-angiotensin system in the bile duct-ligated rat liver. *Gastroenterology* 2002; **123**: 1667-1676 [PMID: 12404241 DOI: 10.1053/gast.2002.36561]
- 61 **Paizis G**, Tikellis C, Cooper ME, Schembri JM, Lew RA, Smith AI, Shaw T, Warner FJ, Zuilli A, Burrell LM, Angus PW. Chronic liver injury in rats and humans upregulates the novel enzyme angiotensin converting enzyme 2. *Gut* 2005; **54**: 1790-1796 [PMID: 16166274 DOI: 10.1136/gut.2004.062398]
- 62 **Jonsson JR**, Clouston AD, Ando Y, Kelemen LI, Horn MJ, Adamson MD, Purdie DM, Powell EE. Angiotensin-converting enzyme inhibition attenuates the progression of rat hepatic fibrosis. *Gastroenterology* 2001; **121**: 148-155 [PMID: 11438504 DOI: 10.1053/gast.2001.25480]
- 63 **Yoshiji H**, Kuriyama S, Fukui H. Angiotensin-I-converting enzyme inhibitors may be an alternative anti-angiogenic strategy in the treatment of liver fibrosis and hepatocellular carcinoma. Possible role of vascular endothelial growth factor. *Tumour Biol* 2002; **23**: 348-356 [PMID: 12677092 DOI: 10.1159/000069792]
- 64 **Yoshiji H**, Kuriyama S, Noguchi R, Ikenaka Y, Yoshiji J, Yanase K, Namisaki T, Kitade M, Yamazaki M, Asada K, Akahane T, Tsujimoto T, Uemura M, Fukui H. Amelioration of liver fibrogenesis by dual inhibition of PDGF and TGF-beta with a combination of imatinib mesylate and ACE inhibitor in rats. *Int J Mol Med* 2006; **17**: 899-904 [PMID: 16596278]
- 65 **Lubel JS**, Herath CB, Tchongue J, Grace J, Jia Z, Spencer K, Casley D, Crowley P, Sievert W, Burrell LM, Angus PW. Angiotensin-(1-7), an alternative metabolite of the renin-angiotensin system, is up-regulated in human liver disease and has antifibrotic activity in the bile-duct-ligated rat. *Clin Sci (Lond)* 2009; **117**: 375-386 [PMID: 19371232 DOI: 10.1042/CS20080647]
- 66 **Herath CB**, Lubel JS, Jia Z, Velkoska E, Casley D, Brown L, Tikellis C, Burrell LM, Angus PW. Portal pressure responses and angiotensin peptide production in rat liver are determined by relative activity of ACE and ACE2. *Am J Physiol Gastrointest Liver Physiol* 2009; **297**: G98-G106 [PMID: 19389807 DOI: 10.1152/ajpgi.00045.2009]
- 67 **Macgillchrist AJ**, Howes LG, Hawksby C, Reid JL. Plasma noradrenaline in cirrhosis: a study of kinetics and temporal relationship to ascites formation. *Eur J Clin Invest* 1991; **21**: 238-243 [PMID: 1905638 DOI: 10.1111/j.1365-2362.1991.tb01816.x]
- 68 **Hennenberg M**, Trebicka J, Sauerbruch T, Heller J. Mechanisms of extrahepatic vasodilation in portal hypertension. *Gut* 2008; **57**: 1300-1314 [PMID: 18445644 DOI: 10.1136/gut.2007.144584]
- 69 **Pinzani M**, Milani S, De Franco R, Grappone C, Caligiuri A, Gentilini A, Tosti-Guerra C, Maggi M, Failli P, Ruocco C, Gentilini P. Endothelin 1 is overexpressed in human cirrhotic liver and exerts multiple effects on activated hepatic stellate cells. *Gastroenterology* 1996; **110**: 534-548 [PMID: 8566602 DOI: 10.1053/gast.1996.v110.pm8566602]
- 70 **Hernández-Guerra M**, García-Pagán JC, Bosch J. Increased hepatic resistance: a new target in the pharmacologic therapy of portal hypertension. *J Clin Gastroenterol* 2005; **39**:

- S131-S137 [PMID: 15758648]
- 71 **Rockey DC.** Vasoactive agents in intrahepatic portal hypertension and fibrogenesis: implications for therapy. *Gastroenterology* 2000; **118**: 1261-1265 [PMID: 10833501 DOI: 10.1016/S0016-5085(00)70379-9]
  - 72 **Shah V,** Cao S, Hendrickson H, Yao J, Katusic ZS. Regulation of hepatic eNOS by caveolin and calmodulin after bile duct ligation in rats. *Am J Physiol Gastrointest Liver Physiol* 2001; **280**: G1209-G1216 [PMID: 11352814]
  - 73 **Shah V,** Toruner M, Haddad F, Cadelina G, Papapetrooulos A, Choo K, Sessa WC, Groszmann RJ. Impaired endothelial nitric oxide synthase activity associated with enhanced caveolin binding in experimental cirrhosis in the rat. *Gastroenterology* 1999; **117**: 1222-1228 [PMID: 10535886 DOI: 10.1016/S0016-5085(99)70408-7]
  - 74 **Gorelik G,** Carhini LA, Scicli AG. Angiotensin 1-7 induces bradykinin-mediated relaxation in porcine coronary artery. *J Pharmacol Exp Ther* 1998; **286**: 403-410 [PMID: 9655885]
  - 75 **Gioli-Pereira L,** Nascimento EA, Santos EL, Bracht A, Juliano MA, Pesquero JB, Borges DR, Kouyoumdjian M. Fate of bradykinin on the rat liver when administered by the venous or arterial route. *J Gastroenterol Hepatol* 2005; **20**: 463-473 [PMID: 15740493 DOI: 10.1111/j.1440-1746.2005.03580.x]
  - 76 **Fleming I,** Fisslthaler B, Dimmeler S, Kemp BE, Busse R. Phosphorylation of Thr(495) regulates Ca(2+)/calmodulin-dependent endothelial nitric oxide synthase activity. *Circ Res* 2001; **88**: E68-E75 [PMID: 11397791 DOI: 10.1161/hh1101.092677]
  - 77 **Ferlitsch A,** Pleiner J, Mittermayer F, Schaller G, Homoncik M, Peck-Radosavljevic M, Wolzt M. Vasoconstrictor hyporeactivity can be reversed by antioxidants in patients with advanced alcoholic cirrhosis of the liver and ascites. *Crit Care Med* 2005; **33**: 2028-2033 [PMID: 16148476]
  - 78 **Vilas-Boas WW,** Ribeiro-Oliveira A, Pereira RM, Ribeiro Rda C, Almeida J, Nadu AP, Simões e Silva AC, dos Santos RA. Relationship between angiotensin-(1-7) and angiotensin II correlates with hemodynamic changes in human liver cirrhosis. *World J Gastroenterol* 2009; **15**: 2512-2519 [PMID: 19469002 DOI: 10.3748/wjg.15.2512]
  - 79 **Grace JA,** Jia Z, Herath CB, Burrell LM, Angus PW. Angiotensin-(1-7) is a mesenteric vasodilator in experimental cirrhosis. *Hepatology* 2010; **52**: 919A-1020A
  - 80 **Miñano C,** Garcia-Tsao G. Clinical pharmacology of portal hypertension. *Gastroenterol Clin North Am* 2010; **39**: 681-695 [PMID: 20951924 DOI: 10.1016/j.gtc.2010.08.015]
  - 81 **Schrier RW,** Arroyo V, Bernardi M, Epstein M, Henriksen JH, Rodés J. Peripheral arterial vasodilation hypothesis: a proposal for the initiation of renal sodium and water retention in cirrhosis. *Hepatology* 1988; **8**: 1151-1157 [PMID: 2971015 DOI: 10.1002/hep.1840080532]
  - 82 **Sieber CC,** Groszmann RJ. In vitro hyporeactivity to methoxamine in portal hypertensive rats: reversal by nitric oxide blockade. *Am J Physiol* 1992; **262**: G996-G1001 [PMID: 1616049]
  - 83 **Hartleb M,** Moreau R, Cailmail S, Gaudin C, Lebrec D. Vascular hyporesponsiveness to endothelin 1 in rats with cirrhosis. *Gastroenterology* 1994; **107**: 1085-1093 [PMID: 7523215]
  - 84 **Heller J,** Schepke M, Gehnen N, Molderings GJ, Muller A, Erhard J, Spengler U, Sauerbruch T. Altered adrenergic responsiveness of endothelium-denuded hepatic arteries and portal veins in patients with cirrhosis. *Gastroenterology* 1999; **116**: 387-393 [PMID: 9922320 DOI: 10.1016/S0016-5085(99)70136-8]
  - 85 **Helmy A,** Newby DE, Jalan R, Johnston NR, Hayes PC, Webb DJ. Nitric oxide mediates the reduced vasoconstrictor response to angiotensin II in patients with preascitic cirrhosis. *J Hepatol* 2003; **38**: 44-50 [PMID: 12480559 DOI: 10.1016/S0168-8278(02)00319-7]
  - 86 **Hennenberg M,** Biecker E, Trebicka J, Jochem K, Zhou Q, Schmidt M, Jakobs KH, Sauerbruch T, Heller J. Defective RhoA/Rho-kinase signaling contributes to vascular hypocontractility and vasodilation in cirrhotic rats. *Gastroenterology* 2006; **130**: 838-854 [PMID: 16530523 DOI: 10.1053/j.gastro.2005.11.029]
  - 87 **Hennenberg M,** Trebicka J, Biecker E, Schepke M, Sauerbruch T, Heller J. Vascular dysfunction in human and rat cirrhosis: role of receptor-desensitizing and calcium-sensitizing proteins. *Hepatology* 2007; **45**: 495-506 [PMID: 17256744 DOI: 10.1002/hep.21502]
  - 88 **Hennenberg M,** Trebicka J, Kohistani AZ, Heller J, Sauerbruch T. Vascular hyporesponsiveness to angiotensin II in rats with CCl(4)-induced liver cirrhosis. *Eur J Clin Invest* 2009; **39**: 906-913 [PMID: 19522833 DOI: 10.1111/j.1365-2362.2009.02181.x]
  - 89 **Kiel JW,** Pitts V, Benoit JN, Granger DN, Shepherd AP. Reduced vascular sensitivity to norepinephrine in portal-hypertensive rats. *Am J Physiol* 1985; **248**: G192-G195 [PMID: 3970200]
  - 90 **Newby DE,** Jalan R, Masumori S, Hayes PC, Boon NA, Webb DJ. Peripheral vascular tone in patients with cirrhosis: role of the renin-angiotensin and sympathetic nervous systems. *Cardiovasc Res* 1998; **38**: 221-228 [PMID: 9683925 DOI: 10.1016/S0008-6363(98)00008-X]
  - 91 **Neef M,** Biecker E, Heller J, Schepke M, Nischalke HD, Wolff M, Spengler U, Reichen J, Sauerbruch T. Portal hypertension is associated with increased mRNA levels of vasopressor G-protein-coupled receptors in human hepatic arteries. *Eur J Clin Invest* 2003; **33**: 249-255 [PMID: 12641544 DOI: 10.1046/j.1365-2362.2003.01131.x]
  - 92 **Vaughan RB,** Angus PW, Angus JA. Enhanced vascular responses to noradrenaline in isolated omental arteries from patients with advanced cirrhosis: evidence against intrinsic vascular hyporeactivity in cirrhosis. *Gut* 2004; **53**: 1544-1545 [PMID: 15361510]
  - 93 **Langer DA,** Shah VH. Nitric oxide and portal hypertension: interface of vasoreactivity and angiogenesis. *J Hepatol* 2006; **44**: 209-216 [PMID: 16297493 DOI: 10.1016/j.jhep.2005.10.004]
  - 94 **Shah V.** Cellular and molecular basis of portal hypertension. *Clin Liver Dis* 2001; **5**: 629-644 [PMID: 11565134 DOI: 10.1016/S1089-3261(05)70185-9]
  - 95 **Vaughan RB,** Angus JA, Angus PW. Vasoconstrictor responses are normal but prostanoid-mediated vasodilatation is enhanced in human cirrhotic mesenteric arteries. *J Gastroenterol Hepatol* 2005; **20**: 1158-1164 [PMID: 16048562 DOI: 10.1111/j.1440-1746.2005.03946.x]
  - 96 **Wiest R,** Groszmann RJ. The paradox of nitric oxide in cirrhosis and portal hypertension: too much, not enough. *Hepatology* 2002; **35**: 478-491 [PMID: 11826425 DOI: 10.1053/jhep.2002.31432]
  - 97 **Colle IO,** De Vriese AS, Van Vlierberghe HR, Lameire NH, De Vos MM. Vascular hyporesponsiveness in the mesenteric artery of anaesthetized rats with cirrhosis and portal hypertension: an in-vivo study. *Eur J Gastroenterol Hepatol* 2004; **16**: 139-145 [PMID: 15075986 DOI: 10.1097/00042737-200402000-00004]
  - 98 **Mathie RT,** Ralevic V, Burnstock G. Portal vascular responsiveness to sympathetic stimulation and nitric oxide in cirrhotic rats. *J Hepatol* 1996; **25**: 90-97 [PMID: 8836907 DOI: 10.1016/S0168-8278(96)80333-3]
  - 99 **Schepke M,** Heller J, Paschke S, Thomas J, Wolff M, Neef M, Malago M, Molderings GJ, Spengler U, Sauerbruch T. Contractile hyporesponsiveness of hepatic arteries in humans with cirrhosis: evidence for a receptor-specific mechanism. *Hepatology* 2001; **34**: 884-888 [PMID: 11679958 DOI: 10.1053/jhep.2001.28794]
  - 100 **Zhang HY,** Shirasawa Y, Chen X, Yu H, Benoit JN. Impaired agonist-dependent myosin phosphorylation and decreased RhoA in rat portal hypertensive mesenteric vasculature. *Am J Physiol Gastrointest Liver Physiol* 2005; **288**: G603-G608 [PMID: 15513955 DOI: 10.1152/ajpgi.00116.2004]
  - 101 **Alam I,** Bass NM, Bacchetti P, Gee L, Rockey DC. He-

- patic tissue endothelin-1 levels in chronic liver disease correlate with disease severity and ascites. *Am J Gastroenterol* 2000; **95**: 199-203 [PMID: 10638583 DOI: 10.1111/j.1572-0241.2000.01684.x]
- 102 **Gracia-Sancho J**, Laviña B, Rodríguez-Vilarrupla A, García-Calderó H, Bosch J, García-Pagán JC. Enhanced vasoconstrictor prostanoid production by sinusoidal endothelial cells increases portal perfusion pressure in cirrhotic rat livers. *J Hepatol* 2007; **47**: 220-227 [PMID: 17459512 DOI: 10.1016/j.jhep.2007.03.014]
- 103 **Guturu P**, Shah V. New insights into the pathobiology of portal hypertension. *Hepatol Res* 2009; **39**: 1016-1019 [PMID: 19796039 DOI: 10.1111/j.1872-034X.2009.00553.x]
- 104 **Ferreira AJ**, Raizada MK. Are we poised to target ACE2 for the next generation of antihypertensives? *J Mol Med (Berl)* 2008; **86**: 685-690 [PMID: 18449520 DOI: 10.1007/s00109-008-0339-x]
- 105 **Shenoy V**, Qi Y, Katovich MJ, Raizada MK. ACE2, a promising therapeutic target for pulmonary hypertension. *Curr Opin Pharmacol* 2011; **11**: 150-155 [PMID: 21215698 DOI: 10.1016/j.coph.2010.12.002]

**P- Reviewers** Gurvits GE, Zhao D, Saavedra J  
**S- Editor** Song XX **L- Editor** A **E- Editor** Lu YJ



## Assessment of platelet activation and phagocytic activity in gastric cancer patients

Joanna Matowicka-Karna, Zbigniew Kamocki, Halina Kemonia

Joanna Matowicka-Karna, Halina Kemonia, Department of Clinical Laboratory Diagnostics, Medical University of Bialystok, 15-269 Bialystok, Poland  
Zbigniew Kamocki, the Second Department of General and Gastroenterological Surgery, Medical University of Bialystok, 15-269 Bialystok, Poland

Author contributions: Matowicka-Karna J designed and performed the experiments, analysed data and wrote the manuscript; Kamocki Z selected patients and analyzed data; Kemonia H has approved the version to be published.

Correspondence to: Joanna Matowicka-Karna, PhD, Department of Clinical Laboratory Diagnostics, Medical University of Bialystok, Waszyngtona 15A, 15-269 Bialystok, Poland. [matowic@umb.edu.pl](mailto:matowic@umb.edu.pl)

Telephone: +48-85-7468584 Fax: +48-85-7468584

Received: July 24, 2012 Revised: October 1, 2012

Accepted: December 6, 2012

Published online: February 15, 2013

platelet count and stimulation of thrombocytopoiesis, the phagocytic functions of blood platelets were markedly impaired. Tumor development seems to impair metabolic processes.

**CONCLUSION:** A decreasing phagocytic activity can promote both inflammatory processes and cancer growth.

© 2013 Baishideng. All rights reserved.

**Key words:** Gastric cancer; Interleukin-6; Blood platelet; Phagocytic activity; Soluble platelet selectin

Matowicka-Karna J, Kamocki Z, Kemonia H. Assessment of platelet activation and phagocytic activity in gastric cancer patients. *World J Gastrointest Pathophysiol* 2013; 4(1): 12-17 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i1/12.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i1.12>

### Abstract

**AIM:** To assess the activation of platelets and their phagocytic activity in the course of gastric cancer.

**METHODS:** Forty-three gastric cancer patients were recruited to the study. The patients were divided into 3 groups depending on tumor stage. Group E included 6 patients with early gastric cancer; group A 18 patients with locally advanced cancer; and group M-19 with metastatic cancer. The investigations were performed twice, prior to surgery and 12-14 d afterwards.

**RESULTS:** The platelet count and the level of soluble platelet selectin (sP-selectin) were found to increase with the disease progression. The level of sP-selectin was lowest in early cancer and was observed to increase after surgery in all the study patients. Irrespective of tumor stage, a statistically significant decrease was noted in the percentage of phagocytizing platelets and in the phagocytic index in gastric cancer patients as compared to healthy subjects. Despite increased

### INTRODUCTION

Gastric cancer is the second most common cause of deaths due to cancer worldwide<sup>[1]</sup>. Approximately 900 000 new cases of gastric cancer are diagnosed every year. The risk factors include first of all *Helicobacter pylori* (*H. pylori*) infection, which leads to chronic inflammation of the gastric mucosa and intestinal metaplasia. Irrespective of histological type, gastric cancer is characterized by high malignancy and high incidence of lymph node involvement and distant metastases. Survival prognosis depends on tumor staging and the presence of metastases. Gastric cancer is usually located in the pyloric antrum and in the pylorus, but in 25% of cases in the body and bottom of the stomach. As the clinical symptoms of gastric cancer are frequently nonspecific, it is diagnosed only at the advanced stage<sup>[2]</sup>. Therefore, there is a continuous search for alternative uses of diagnostic markers that would allow early detection of gastric cancer<sup>[3]</sup>.

The proinflammatory cytokines, interleukin (IL)-6 and

IL-23, which are released during *H. pylori* infection and which strongly inhibit secretion of gastric juice, are involved in the pathogenesis and development of tumors. They facilitate tumor growth by inhibiting apoptosis of tumor cells and by inducing angiogenesis within the tumor. Platelets play an active part in cancerous diseases through secreted proinflammatory factors, chemokines and growth factors. Tumor cells exhibit prothrombotic effect by inducing platelet aggregation and by platelet activation. Upon platelet activation, platelet surface P-selectin binds to ligand CD24 present on tumor cells, causing their adherence to endothelial cells. P-selectin plays a key role both in inflammatory processes and in the pathogenesis of thrombosis in cancer patients<sup>[4]</sup>.

Blood platelets also exhibit the ability to phagocytize viruses, bacteria or immune complexes. They become activated and change in shape and surface properties: this is accompanied by the formation of phagosomes containing proteolytic enzymes<sup>[5-7]</sup>.

The study objective was to find out whether and how gastric cancer-related inflammation affects platelet activation and whether it changes their phagocytic functions. IL-6 was used as an inflammatory marker in gastric cancer patients.

## MATERIALS AND METHODS

The study involved 43 gastric cancer patients treated at the Second Department of General and Gastroenterological Surgery, Medical University of Bialystok. The group consisted of 32 men (M) and 11 women (F), aged 31-76 years (mean age 55.4 years). The patients were divided into 3 groups by cancer (adenocarcinoma) stage<sup>[8]</sup>. Group I (E) included 6 patients (4 M, 2 F) with early stage, group II (A) had 18 patients (13 M, 5 F) with locally advanced stage and group III (M) had 19 patients (15 M, 4 F) with cancer metastases. According to the UICC classification, in group E there were 5 stage I patients (12%) and 1 stage II patient (2%), in group A - 7 stage IIIA patients (16%) and 11 stage IIIB patients (26%), and in group M - 19 stage IV patients (44%) (Table 1). All the patients were qualified for surgery. In 34 cases the surgery involved total gastrectomy and regional lymphadenectomy, omentectomy and splenectomy. In 8 of these patients, treatment was extended to pancreatic tail resection. Four patients with less advanced cancer underwent Billroth II gastrectomy with regional lymphadenectomy. Due to the stage of the cancer, in 5 patients treatment was limited to gastrointestinal anastomosis. Examinations were carried out twice, before surgery (E1, A1, M1) and 12-14 d after surgery (E2, A2, M2).

The control group (C) consisted of 40 healthy subjects (aged 20-45 years), including 22 M and 18 F. Statistical analysis of all the parameters, including age and sex, was performed within the study groups and in the control group. Results were analyzed using Statistical 8.0 program.

All the patients gave their consent to the study in ac-

cordance with the Guidelines for Good Clinical Practice. The study was approved by the Bioethics Committee of the Medical University of Bialystok (RI-002/154/2009). Venous blood collected for clot was used as the material for analysis.

### Assessment of platelet phagocytic activity

About 1 mL of ACD (a mixture of citric acid and glucose) was added to 5 mL of venous blood collected for heparin at a concentration of 50 IU/mL blood. ACD prevents platelet aggregation by inhibiting thromboxane synthesis by platelets and reduces plasma pH, to 6.5, which decreases the physiological ability of platelets to agglutinate. Blood platelets were isolated by double centrifugation, taking advantage of the differences in the specific weight of the respective morphotic elements of blood. Whole blood was centrifuged at 250 *g* for 12 min, plasma was withdrawn to another test-tube and then centrifugation was repeated at 1500 *g* for 20 min. Some plasma (2 mL) was left over the platelet sediment and the sedimented platelets were suspended in it by gentle stirring with a plastic pipette. This platelet-rich plasma (PRP) was supplemented with 0.1 mL ACD to prevent platelet aggregation. In the PRP obtained in this way, erythrocytes and leukocytes did not exceed 1/1000 platelets<sup>[9]</sup>.

### Bacteria used for the phagocytic activity of blood platelets

*Staphylococcus aureus* ATCC 6538P strain was used for the study. The bacteria were cultured on an agar slant for 20 min, then rinsed off with 10 mL of phosphate buffer (PBS) and centrifuged at 1500 *g* for 20 min. The centrifuged bacterial sediment was rinsed with PBS three times and centrifuged again. The number of bacteria in the suspension was determined spectrophotometrically, by measuring optical density of the suspension. The suspension containing  $120 \times 10^6$  -  $140 \times 10^6$  bacteria, used to examine the phagocytic and bactericidal activity, corresponds to an optical density of T-63 at a wavelength of 540 nm. The bacteria were grown on the agar medium and counted. The suspension was diluted with PBS to obtain the platelet/bacterium ratio of 1:1.

Platelet phagocytic activity was determined by measuring the percentage phagocytizing cells and the phagocytic index; whereas bactericidal activity was determined by calculating the difference between the number of bacteria phagocytized by platelets and the number of phagocytized bacteria which survived within platelets.

### Determination of the percentage of phagocytizing platelets

Bacterial suspension and PRP were incubated at 37 °C for 6 min, and then mixed at 75 *g*. After incubation, smears were made and then stained with the Pappenheim method, for 1 h using Giemsa reagent. The stained preparations were evaluated in a light microscope at  $\times 1400$  magnification. The percentage of phagocytizing platelets was described as the percentage of phagocytizing platelets per 1000 other cells in the preparation.

**Table 1** Stage of gastric cancer *n* (%)

Group tested	
Tumor stage (TNM classification)	I + II
	6 (14)
	III
	18 (42)
	IV
Depth of tumor invasion	19 (44)
	T1
	2 (6)
	T2
	4 (13)
Lymph node metastases	T3
	15 (47)
	T4
	11 (34)
	N0
Distant metastases	7 (17)
	N1
	4 (9)
	N2
	13 (31)
	N3
	18 (43)
	M0
	9 (47)
	M1
	10 (53)

### Determination of the phagocytic index

The phagocytic index was determined using 100 phagocytizing platelets. The index was calculated as the mean number of phagocytized bacteria per single platelet according to the formula: phagocytic index = number of phagocytized bacteria/number of phagocytizing platelets. Platelet count (PLT) was determined using a hematology analyzer (ADVIA 2120, Siemens). The level of soluble platelet selectin (sP-selectin) was determined by the enzyme-linked immunosorbent assay (ELISA) method using Quantikine human kit (R and D Systems, United States). The level of IL-6 was determined by the ELISA method using Quantikine human kit (R and D Systems, United States).

The results were subjected to statistical analysis using the program Statistica 8.0. The differences were considered statistically significant when the value of the test function was at the level of significance set at  $P < 0.05$ . The Kolmogorov consistency test was used for features consistent with normal distribution, the *t*-Student test for comparisons between the groups and the Mann-Whitney test for traits inconsistent with this distribution.

## RESULTS

PLT in patients with early gastric cancer (E1 and E2) and with locally advanced cancer (A1 and A2) before and after surgery differed statistically significantly ( $P < 0.05$ ). PLT in these groups increased statistically significantly after surgery (Table 2).

The level of sP-selectin was lowest in group E1 and

only this value was found to differ statistically significantly compared to the control group ( $P < 0.05$ ). In the other groups, the increase was not statistically significant in comparison with the control group. A statistically significant difference in the level of sP-selectin was observed between the groups A1 and A2 (Table 2).

The level of IL-6 was found to be markedly higher in the groups prior to surgery (E1, A1 and M1) as compared to the control, the differences being statistically significant (Table 2).

Statistically significant differences were noted in the percentage of phagocytizing platelets between groups E and A, before and after surgery. In the study groups E1, A1 and M1 the percentage values of phagocytizing platelets were statistically significantly lower than those obtained in the control group ( $P < 0.001$ ) (Table 3).

The phagocytic index was statistically significantly lower in the groups before surgery (E1, A1 and M1) as compared to the control (Table 3).

## DISCUSSION

Gastric cancer is accompanied by inflammation and at the same time by impairment of immune mechanisms. *H. pylori* is responsible for gastric mucosa inflammation, resulting in the stimulation of neutrophils and macrophages, and in an increased production of reactive oxygen species. Chronic inflammation and reduced secretion of hydrochloric acid lead to stomach ulceration or, by causing intestinal metaplasia of the stomach, turn into dysplasia, which is considered a precancerous condition<sup>[10]</sup>. Platelets take an active part in the initiation and development of the inflammatory process by adhering to the cells of the vascular walls and by the release of chemokines, cytokines, proteases and procoagulants<sup>[11]</sup>. Cancer cells undergo proliferation under the effect of epidermal growth factor, platelet-derived growth factor (PDGF) and insulin-like growth factor-1 released from blood platelets. Vascular endothelial growth factor (VEGF) and angiopoietin exert a pro-angiogenic effect<sup>[12-14]</sup>. Fibrinogen, fibronectin, vitronectin and vWF enhance platelet-endothelium adhesion by the formation of connections between GP II b-III a and integrin  $\alpha v \beta 3$  or intercellular adhesion molecule-1. Adhesion molecules belonging to the P-selectin group have a special function during aggregation of platelets and cancer cells. P-selectin translocated from  $\alpha$ -granules onto the platelet surface during platelet activation facilitates their interaction with endothelial cells, monocytes, neutrophils and lymphocytes. Cancerous diseases are usually accompanied by overproduction of platelets. Thrombocytosis, which can be the cause of increased risk of metastases, has been observed in the cancer of the stomach, colon, lungs, kidneys, prostate and reproductive organs<sup>[15,16]</sup>.

IL-6 is a mediator of platelet production and a direct stimulator of megakaryocytes<sup>[15]</sup>. Ikeguchi *et al.*<sup>[17]</sup> showed the usefulness of IL-6 determination in the diagnosis of gastric cancer. They found a statistically significant cor-

**Table 2** Mean values of platelet counts, levels of soluble platelet-selectin and interleukin-6 in cancer patients-early carcinoma-E advanced carcinoma with lymph node involvement-A, metastatic cancer-M and in control group (mean  $\pm$  SD)

	PLT	sP-selectin	IL-6
Study group E1	247.33 $\pm$ 42.92	67.93 $\pm$ 74.49	20.68 $\pm$ 6.31
	E1 vs C	E1 vs C	E1 vs C
	0.8 < P < 0.9	P < 0.05	P < 0.01
Study group E2	662.83 $\pm$ 335.23	153.37 $\pm$ 174.59	10.90 $\pm$ 8.43
	E1 vs E2	E1 vs E2	E1 vs E2
	P < 0.05	0.5 < P < 0.6	0.1 < P < 0.2
	E2 vs C	E2 vs C	E2 vs C
	P < 0.05	0.6 < P < 0.7	0.05 < P < 0.1
Study group A1	344.28 $\pm$ 266.40	193.23 $\pm$ 143.70	11.51 $\pm$ 8.44
	A1 vs C	A1 vs C	A1 vs C
	0.05 < P < 0.1	0.05 < P < 0.1	P < 0.001
Study group A2	584.25 $\pm$ 237.54	278.54 $\pm$ 158.64	20.42 $\pm$ 12.88
	A1 vs A2	A1 vs A2	A1 vs A2
	P < 0.05	P < 0.001	0.2 < P < 0.3
	A2 vs C	A2 vs C	A2 vs C
	P < 0.01	0.2 < P < 0.3	P < 0.05
Study group M1	335.84 $\pm$ 154.08	173.71 $\pm$ 116.79	8.44 $\pm$ 5.63
	M1 vs C	M1 vs C	M1 vs C
	P < 0.01	0.1 < P < 0.2	P < 0.001
Study group M2	451.55 $\pm$ 210.74	178.96 $\pm$ 84.56	15.26 $\pm$ 12.73
	M1 vs M2	M1 vs M2	M1 vs M2
	0.2 < P < 0.3	0.1 < P < 0.2	0.1 < P < 0.2
	M2 vs C	M2 vs C	M2 vs C
	P < 0.01	0.5 < P < 0.6	P < 0.05
Control group C	247.78 $\pm$ 44.74	130.38 $\pm$ 48.35	2.45 $\pm$ 1.44

PLT: Platelet count; sP-selectin: Soluble platelet selectin; IL: Interleukin.

**Table 3** Assessment of phagocytic activity of blood platelets in gastric cancer patients-E, -A, -M and in control group (mean  $\pm$  SD)

	Percentage of phagocytizing platelets	Phagocytic index
Study group early carcinoma E1	1.08 $\pm$ 0.08	1.02 $\pm$ 0.04
	E1 vs C	E1 vs C
	P < 0.001	P < 0.001
Study group early carcinoma E2	1.27 $\pm$ 0.10	1.07 $\pm$ 0.05
	E1 vs E2	E1 vs E2
	P < 0.05	0.05 < P < 0.1
	E2 vs C	E2 vs C
	P < 0.01	P < 0.01
Study group advanced carcinoma with lymph node involvement A1	1.10 $\pm$ 0.11	1.02 $\pm$ 0.11
	A1 vs C	A1 vs C
	P < 0.001	P < 0.001
Study group advanced carcinoma with lymph node involvement A2	1.23 $\pm$ 0.08	1.03 $\pm$ 0.06
	A1 vs A2	A1 vs A2
	P < 0.05	0.7 < P < 0.8
	A2 vs C	A2 vs C
	P < 0.001	P < 0.001
Study group metastatic cancer M1	1.13 $\pm$ 0.10	1.0 $\pm$ 0.07
	M1 vs C	M1 vs C
	P < 0.001	P < 0.001
Study group metastatic cancer M2	1.20 $\pm$ 0.12	1.05 $\pm$ 0.08
	M1 vs M2	M1 vs M2
	0.2 < P < 0.3	0.7 < P < 0.8
	M2 vs C	M2 vs C
	P < 0.001	P < 0.001
Control group C	2.26 $\pm$ 0.57	1.83 $\pm$ 0.37

relation between IL-6 and tumor stage. An elevated level

of IL-6 indicated disease progression and higher malignancy, and was thus an unfavorable prognostic factor. The level of IL-6 was found to correlate with the disease stage and increased in gastric cancer relapse<sup>[18]</sup>. Ashizawa *et al*<sup>[19]</sup> described the importance of IL-6 measurement in gastric cancer patients with local lymph node involvement and distant metastases. Due to its pyrogenic action, IL-6 is responsible for cachexia, fever, body mass reduction and other devastating symptoms of tumor progression. Determination of IL-6 can be a prognostic factor of survival in gastric cancer patients<sup>[19-21]</sup>. In our study, the level of IL-6 was found to increase in all gastric cancer patients. Its values were the highest in patients with early cancer (over an 8-fold increase), whereas the lowest in those with metastases (over a 3-fold increase), as compared to the control group. This seems to confirm that inflammation can be the primary stage of the neoplastic process. When the inflammatory process begins to extinguish, neoformation develops intensively. PLT was found to increase along with disease progression.

Platelets are a rich source of pro- and anti-angiogenic factors that regulate the process of tumor growth. The angiogenesis-stimulating factors include tumor necrosis factor- $\alpha$ , PDGF, granulocyte-macrophage colony-stimulating factor, IL-6 and metalloproteinases. After angiogenesis initiation, platelets adhere to vascular endothelium and aggregate. It has been shown that VEGF released from platelets stimulates the development of megakaryocytes and platelets, activates endothelial cells to release vWF, and thus facilitates platelet adhesion to

the vascular wall<sup>[22,23]</sup>.

According to Park *et al*<sup>[24]</sup>, the statistically significantly lower values of MPC in gastric cancer patients are caused by degranulation and release of granular contents following platelet activation. Osada *et al*<sup>[25]</sup> found no statistically significant differences in the platelet count between gastric cancer patients and control subjects. The number of CD62P antigens on the platelet surface after TRAP activation in gastric cancer patients increased from 6 to 12 times as compared to a 3-fold increase in healthy subjects. The large number of glycoproteins on the platelet surface observed in gastric cancer after platelet activation *in vitro* may indicate prothrombotic tendencies. Platelets exhibited far greater activity, with their number remaining unchanged.

During platelet activation, the membranes of platelet  $\alpha$ -granules combine with the platelet cell membrane, whereas P-selectin is expressed on the surface as CD62P receptor. Part of the extracellular domain is rinsed off to the blood and occurs as sP-selectin, which has a long half-life and is a good plasma marker to assess platelet activation. An increase in the level of sP-selectin is accompanied by a decrease in P-selectin expression on the platelet surface. Soluble P-selectin is a marker of platelet hyperactivity, endothelial dysfunction and inflammation, and may serve as a biomarker associated with venous thrombosis in the course of cancer. At the same time it plays a major role linking inflammation with thrombosis<sup>[26]</sup>.

Ikeda *et al*<sup>[27]</sup> reported a slight, statistically insignificant difference in the level of sP-selectin determined in gastric cancer patients before and after surgery. However, in our study the level of sP-selectin was statistically significantly lower only in patients with early cancer (group E) as compared to the control, and it was accompanied by the lowest platelet count. In this group, the level of IL-6 was the highest, which suggested acute inflammation. The decrease in the platelet count may be due to their consumption at the site of the inflammatory and then neoplastic process. The lowest level of sP-selectin seems to confirm earlier activation of platelets and disintegration of sP-selectin. In the other groups of patients, an increase in platelet count was accompanied by a rise in sP-selectin, although the differences were not statistically significant. The increased level of sP-selectin reflects intravascular platelet activation.

P-selectin takes part in the process of platelet aggregation and in interactions with cancer cells. Platelets bind to cancer cells and form aggregates around them, thus protecting them against the host immune system and in consequence allowing survival and formation of metastases. Factors released from tumor cells, *e.g.*, cancer procoagulants, thrombin, adenosine diphosphate, tissue factor show the capacity of direct platelet activation. Therefore, a correlation can be found between the level of sP-selectin, platelet count and their activation. Adhesion molecules, such as integrins and glycoproteins are involved in the formation of the platelet-tumor cell aggregates and metastases. The increased expression of

P-selectin on the platelet surface is strongly related to the occurrence of metastases and therefore the blockade of P-selectin can be used to inhibit the formation of cancer metastases<sup>[13]</sup>.

Blood platelets exhibit the capacity of phagocytizing and digesting bacteria independently. The process of phagocytosis lasts only 6 min, *i.e.*, it is 5-times faster than in granulocytes. It also seems to be more efficient as the platelet count is 40-times higher and platelets remain in the circulation after digesting bacteria<sup>[6]</sup>. Platelet activation involves release of defensins, as well as Platelet factor 4 and CXC chemokine ligand 4 belonging to the family of chemokines, which show bactericidal properties. These actions are aided by H<sub>2</sub>O<sub>2</sub> and reactive oxygen species that have a toxic effect on bacteria<sup>[28]</sup>. However, White<sup>[29]</sup> questions the ability of platelets to phagocytize bacteria, claiming that platelets can absorb but not kill them, which is due to the fact that platelet lysosomes do not contain myeloperoxidase. He believes that the mechanism of bacterial absorption is in their sequestration by the open canalicular system, through which P-selectins are transported to the platelet surface.

We noted a statistically significant decrease in the percentage of phagocytizing platelets and the phagocytic index in gastric cancer patients irrespective of staging, as compared to healthy subjects. Despite increased platelet count and thrombocytopoiesis stimulation, the phagocytic functions of blood platelets are markedly impaired. It seems that cancer development reduces the effectiveness of the ongoing metabolic processes. The decreasing phagocytic activity of platelets can promote both inflammatory processes and neoplastic metastases.

A decreased phagocytic activity of blood platelets indicates directly that the mechanisms of nonspecific immunity are impaired. On the other hand, an increased level of sP-selectin suggests platelet stimulation and their continuous activation in the course of gastric cancer. A growing tumor promotes platelet stimulation and activation. The lowest platelet count and the lowest level of sP-selectin were observed in early cancer, which seems to confirm the involvement of platelets in the formation of inflammatory foci and their intravascular activation.

## COMMENTS

### Background

*Helicobacter pylori* (*H. pylori*) is responsible for gastric mucosa inflammation, resulting in the stimulation of neutrophils and macrophages, and in an increased production of reactive oxygen species. The proinflammatory cytokines, interleukin (IL)-6 and IL-23, which are released during *H. pylori* infection and which strongly inhibit secretion of gastric juice, are involved in the pathogenesis and development of tumors.

### Research frontiers

Platelets play an active part in cancerous diseases through the secreted proinflammatory factors, chemokines and growth factors. Tumor cells exhibit prothrombotic effect by inducing platelet aggregation and by platelet activation.

### Innovations and breakthroughs

The authors noted a statistically significant decrease in the percentage of phagocytizing platelets and the phagocytic index in gastric cancer patients irrespective of staging, as compared to healthy subjects. Despite increased platelet

count and thrombocytopoiesis stimulation, the phagocytic functions of blood platelets are markedly impaired.

### Applications

The lowest platelet count and the lowest level of soluble platelet selectin were observed in early cancer, which seems to confirm the involvement of platelets in the formation of inflammatory foci and their intravascular activation.

### Peer review

In this study, the level of IL-6 was found to increase in all gastric cancer patients. Its values were the highest in patients with early cancer, whereas the lowest in those with metastases, as compared to the control group. This seems to confirm that inflammation can be the primary stage of the neoplastic process.

## REFERENCES

- Brenner H**, Rothenbacher D, Arndt V. Epidemiology of stomach cancer. *Methods Mol Biol* 2009; **472**: 467-477 [PMID: 19107449 DOI: 10.1007/978-1-60327-492-0\_23]
- Hartgrink HH**, Jansen EP, van Grieken NC, van de Velde CJ. Gastric cancer. *Lancet* 2009; **374**: 477-490 [PMID: 19625077 DOI: 10.1016/S0140-6736(09)60617-6]
- Bornschein J**, Kandulski A, Selgrad M, Malfertheiner P. From gastric inflammation to gastric cancer. *Dig Dis* 2010; **28**: 609-614 [PMID: 21088411 DOI: 10.1159/000320061]
- Läubli H**, Borsig L. Selectins promote tumor metastasis. *Semin Cancer Biol* 2010; **20**: 169-177 [PMID: 20452433 DOI: 10.1016/j.semcancer.2010.04.005]
- George JN**. Platelets. *Lancet* 2000; **355**: 1531-1539 [PMID: 10801186 DOI: 10.1016/S0140-6736(00)02175-9]
- Kemona H**, Andrzejewska A, Prokopowicz J, Nowak H, Mantur M. Phagocytic activity of human blood platelets examined by electron microscopy. *Folia Haematol Int Mag Klin Morphol Blutforsch* 1986; **113**: 696-702 [PMID: 2435636]
- Kerrigan SW**, Cox D. Platelet-bacterial interactions. *Cell Mol Life Sci* 2010; **67**: 513-523 [PMID: 20091082 DOI: 10.1007/s00018-009-0207-z]
- Osch J**. The new TNM classification in gastroenterology (1997). *Endoscopy* 1998; **30**: 643-649 [PMID: 9826145]
- Mantur M**, Wołosowicz N, Prokopowicz J, Kemona H. System for testing the phagocytic capacity of human blood platelets. *Folia Haematol Int Mag Klin Morphol Blutforsch* 1986; **113**: 685-689 [PMID: 2435634]
- Ernst P**. Review article: the role of inflammation in the pathogenesis of gastric cancer. *Aliment Pharmacol Ther* 1999; **13** Suppl 1: 13-18 [PMID: 10209682]
- Blair P**, Flaumenhaft R. Platelet alpha-granules: basic biology and clinical correlates. *Blood Rev* 2009; **23**: 177-189 [PMID: 19450911 DOI: 10.1016/j.blre.2009.04.001]
- Borsig L**, Wong R, Hynes RO, Varki NM, Varki A. Synergistic effects of L- and P-selectin in facilitating tumor metastasis can involve non-mucin ligands and implicate leukocytes as enhancers of metastasis. *Proc Natl Acad Sci USA* 2002; **99**: 2193-2198 [PMID: 11854515 DOI: 10.1073/pnas.261704098]
- Borsig L**. The role of platelet activation in tumor metastasis. *Expert Rev Anticancer Ther* 2008; **8**: 1247-1255 [PMID: 18699763 DOI: 10.1586/14737140.8.8.1247]
- Kitadai Y**. Angiogenesis and lymphangiogenesis of gastric cancer. *J Oncol* 2010; **2010**: 468725 [PMID: 20369064 DOI: 10.1155/2010/468725]
- Heras P**, Hatzopoulos A, Kritikos N, Kritikos K. Platelet count and tumor progression in gastric cancer patients. *Scand J Gastroenterol* 2010; **45**: 1005-1006 [PMID: 20377467 DOI: 10.3109/0036551003797221,]
- Nash GF**, Turner LF, Scully MF, Kakkar AK. Platelets and cancer. *Lancet Oncol* 2002; **3**: 425-430 [PMID: 12142172 DOI: 10.1016/S1470-2045(02)00789-1]
- Ikeguchi M**, Hatada T, Yamamoto M, Miyake T, Matsunaga T, Fukumoto Y, Yamada Y, Fukuda K, Saito H, Tatebe S. Serum interleukin-6 and -10 levels in patients with gastric cancer. *Gastric Cancer* 2009; **12**: 95-100 [PMID: 19562463 DOI: 10.1007/s10120-009-0509-8]
- Wu CW**, Wang SR, Chao MF, Wu TC, Lui WY, P'eng FK, Chi CW. Serum interleukin-6 levels reflect disease status of gastric cancer. *Am J Gastroenterol* 1996; **91**: 1417-1422 [PMID: 8678006]
- Ashizawa T**, Okada R, Suzuki Y, Takagi M, Yamazaki T, Sumi T, Aoki T, Ohnuma S, Aoki T. Clinical significance of interleukin-6 (IL-6) in the spread of gastric cancer: role of IL-6 as a prognostic factor. *Gastric Cancer* 2005; **8**: 124-131 [PMID: 15864720 DOI: 10.1007/s10120-005-0315-x]
- Kim DK**, Oh SY, Kwon HC, Lee S, Kwon KA, Kim BG, Kim SG, Kim SH, Jang JS, Kim MC, Kim KH, Han JY, Kim HJ. Clinical significances of preoperative serum interleukin-6 and C-reactive protein level in operable gastric cancer. *BMC Cancer* 2009; **9**: 155 [PMID: 19457231]
- Wang CS**, Sun CF. C-reactive protein and malignancy: clinico-pathological association and therapeutic implication. *Chang Gung Med J* 2009; **32**: 471-482 [PMID: 19840504]
- Pinedo HM**, Verheul HM, D'Amato RJ, Folkman J. Involvement of platelets in tumour angiogenesis? *Lancet* 1998; **352**: 1775-1777 [PMID: 9848370 DOI: 10.1016/S0140-6736(98)05095-8]
- Yasui W**, Oue N, Aung PP, Matsumura S, Shutoh M, Nakayama H. Molecular-pathological prognostic factors of gastric cancer: a review. *Gastric Cancer* 2005; **8**: 86-94 [PMID: 15864715 DOI: 10.1007/s10120-005-0320-0]
- Park Y**, Schoene N, Harris W. Mean platelet volume as an indicator of platelet activation: methodological issues. *Platelets* 2002; **13**: 301-306 [PMID: 12189016 DOI: 10.1080/095371002220148332]
- Osada J**, Rusak M, Kamocki Z, Dabrowska MI, Kedra B. Platelet activation in patients with advanced gastric cancer. *Neoplasma* 2010; **57**: 145-150 [PMID: 20099978 DOI: 10.4149/neo\_2010\_02\_145]
- Paduch R**. Metastasis-the role of tumour cells aggregation with platelets. *Onkol Pol* 2005; **8**: 229-238
- Ikedai M**, Furukawa H, Imamura H, Shimizu J, Ishida H, Masutani S, Tatsuta M, Kawasaki T, Satomi T. Surgery for gastric cancer increases plasma levels of vascular endothelial growth factor and von Willebrand factor. *Gastric Cancer* 2002; **5**: 137-141 [PMID: 12378339 DOI: 10.1007/s101200200024]
- Clemetson KJ**. Platelets and pathogens. *Cell Mol Life Sci* 2010; **67**: 495-498 [PMID: 19936618 DOI: 10.1007/s00018-009-0204-2]
- White JG**. Why human platelets fail to kill bacteria. *Platelets* 2006; **17**: 191-200 [PMID: 16702047 DOI: 10.1080/09537100500441234]

P- Reviewers Guo JM, Enomoto S S- Editor Zhai HH  
L- Editor Hughes D E- Editor Yan JL



## Cytomegalovirus positive ulcerative colitis: A single center experience and literature review

Uri Kopylov, Gila Sasson, Bella Geysheis, Michal Tepperberg Oikawa, Iris Barshack, Rami Eliakim, Shomron Ben-Horin

Uri Kopylov, Gila Sasson, Rami Eliakim, Shomron Ben-Horin, Department of Gastroenterology, Sheba Medical Center, Ramat Gan 52621, Israel

Bella Geysheis, Internal Medicine D, Sheba Medical Center, Ramat Gan 52621, Israel

Michal Tepperberg Oikawa, Central Virology Laboratory, Public Health Laboratories, Israel Ministry of Health, Ramat Gan 52621, Israel

Iris Barshack, Department of Pathology, Sheba Medical Center, Ramat Gan 52621, Israel

Author contributions: Kopylov U and Ben-Horin S contributed to collection and analysis of data, drafting of the manuscript; Sasson G and Geysheis B contributed to collection of data; Oikawa MT contributed to data collection and analysis; Barshack I and Eliakim R contributed to critical review of the data and the manuscript for important scientific input; all authors reviewed the draft and final version of the manuscript.

Supported by Lecturer Fees from Abbott and Shering-Plough  
Correspondence to: Uri Kopylov, MD, Department of Gastroenterology, Sheba Medical Center, Tel Hashomer, Ramat Gan 52621, Israel. [ukopylov@gmail.com](mailto:ukopylov@gmail.com)

Telephone: +972-3-5302660 Fax: +972-3-5302660

Received: November 8, 2012 Revised: January 28, 2013

Accepted: February 5, 2013

Published online: February 15, 2013

### Abstract

**AIM:** To compare the clinical outcome of cytomegalovirus (CMV)-positive ulcerative colitis (UC) patients with and without antiviral therapy.

**METHODS:** This was a retrospective case-controlled study. The database of UC patients in our institution was scanned for documented presence of CMV on colonic biopsies. Demographics, clinical data, endoscopy findings and pathology reports were extracted from the patients' charts and electronic records. When available, the data from colonoscopies preceding and following the diagnosis of colonic CMV infection were also ex-

tracted. The primary outcomes of the study were colectomy/death during hospitalization and the secondary outcomes were colectomy/death through the course of the follow-up.

**RESULTS:** Thirteen patients were included in the study, 7 (53.5%) of them were treated with gancyclovir and 6 (46.5%) were not. Patients treated with antivirals presented with a more severe disease and 57% of them were treated with cyclosporine or infliximab before initiation of gancyclovir, while none of the patients without antivirals required rescue therapy. One patient died and another patient underwent urgent colectomy during hospitalization, both of them from the gancyclovir-treatment group. For the entire follow-up time ( $13 \pm 13$  mo), a total of 3 colectomies and one death occurred, all among the antiviral-treated patients (for colectomy: 3/7 vs 0/6 patients,  $P = 0.19$ ; for combined adverse outcome: 4/7 vs 0/6 patients,  $P = 0.07$ ). In 9/13 patients, immunohistochemistry for CMV was performed on biopsies obtained during a subsequent colonoscopy and was positive in one patient only.

**CONCLUSION:** Gancyclovir-treated patients had a more severe disease and outcome, probably unrelated to antiviral therapy. Immunohistochemistry-CMV-positive patients with mild disease may recover without antiviral therapy.

© 2013 Baishideng. All rights reserved.

**Key words:** Ulcerative colitis; Cytomegalovirus; Gancyclovir; Cyclosporine; Immunohistochemistry

Kopylov U, Sasson G, Geysheis B, Oikawa MT, Barshack I, Eliakim R, Ben-Horin S. Cytomegalovirus positive ulcerative colitis: A single center experience and literature review. *World J Gastrointest Pathophysiol* 2013; 4(1): 18-23 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i1/18.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i1.18>

## INTRODUCTION

Cytomegalovirus (CMV) infection is very common in the healthy adult population, with reported rates of CMV-IgG positivity of up to 100%, depending on the age and geographical location<sup>[1]</sup>. In immuno-compromised patients (post-solid organ transplantation, chemotherapy-treated, human immunodeficiency virus, recipients of immunosuppressive drugs, for instance), CMV infection or reactivation may lead to a systemic disease or end-organ involvement manifesting as severe pneumonitis, hepatitis or colitis<sup>[2]</sup>.

In patients with acute severe ulcerative colitis, CMV has been reported to be present in the colonic tissue of 21%-34% of patients and of 33%-36% of steroid refractory cases, respectively<sup>[3]</sup>.

The clinical significance of detecting CMV in UC patients remains debatable<sup>[4,5]</sup>. It has been suggested that CMV infection may be a marker of a more severe disease that is more likely to be refractory to corticosteroid and immunosuppressive therapy<sup>[6,7]</sup>. Conversely, some authors suggest that CMV may only be an "innocent bystander", reflecting a remote infection of the involved mucosa and lacking a significant impact on patient outcome<sup>[8,9]</sup>.

Several laboratory techniques are employed for the detection of CMV colitis, including hematoxylin and eosin (HE) staining for evidence of a cytopathic damage in epithelial cells, immunohistochemistry (IHC) and polymerase chain reaction (PCR). As most of the patients who are tested for CMV in the colonic mucosa are afflicted with a severe and treatment-resistant disease, the vast majority of them are treated with antiviral medications (gancyclovir, foscarnet) upon the diagnosis of CMV. However, the clinical benefit of this strategy is still not clear due to a paucity of reports of the outcome of CMV-positive patients who were not treated with antivirals.

In this manuscript, we describe the clinical course and outcome of 6 patients with ulcerative colitis who tested positive for CMV in the colonic mucosa but did not receive antiviral therapy, in comparison to 7 patients who were treated with anti-viral medications.

## MATERIALS AND METHODS

### Patient selection

This was a retrospective case-cohort study. The study cohort included all patients who were in Sheba Medical Center between 2007 and 2011 for an exacerbation of ulcerative colitis and who tested positive for CMV by IHC in colonic biopsies. Patients with Crohn's disease or indeterminate colitis were excluded from the study.

### Data collection

Clinical, endoscopic and laboratory data were retrieved from the patients' charts and electronic records. When available, endoscopic and pathological data from previous (before the index episode) and subsequent colonoscopies were also extracted.

### Histological examination

All samples were examined by an experienced gastrointestinal pathologist for evidence of a cytopathic damage (inclusion bodies) on HE stains.

**CMV immunostaining:** IHC staining for CMV was performed on all samples. Briefly, formalin fixed tissues from patients were dehydrated, embedded in paraffin and sectioned at 4  $\mu$ m. A positive control was added on the right side of the slides. The CMV immunostaining was calibrated on a Benchmark XT staining module (Ventana Medical Systems). The slides were warmed to 60 °C for 1 h and after that processed to a fully automated protocol. Briefly, after sections were dewaxed and rehydrated, a Protease 2 (Ventana Medical Systems) pretreatment during 8 min for antigen retrieval was selected. The CMV antibody (Clones CCH2 + DDG9, M0854, Dako) was diluted 1:25 and incubated for 24 min at 37 °C. Detection was performed with iView DAB Detection kit (Ventana Medical Systems). Counterstaining was performed for 4 min in hematoxylin (Ventana Medical Systems). After the automated staining was completed, the slides were dehydrated in 70% ethanol, 95% ethanol and 100% ethanol for 1 min in each ethanol concentration. Before cover-slipping, the sections were cleared in xylene for 1 min and mounted with Entellan.

### Statistical analysis

The demographic and clinical parameters of CMV-positive patients treated with gancyclovir were compared to those of patients who did not receive antiviral therapy.

Continuous variables were analyzed by student *t*-test for continuous variables and Fisher exact test for categorical variables.  $P < 0.05$  was considered significant. The analysis was performed with Medcalc statistical software version 11 (Mariakerke, Belgium).

## RESULTS

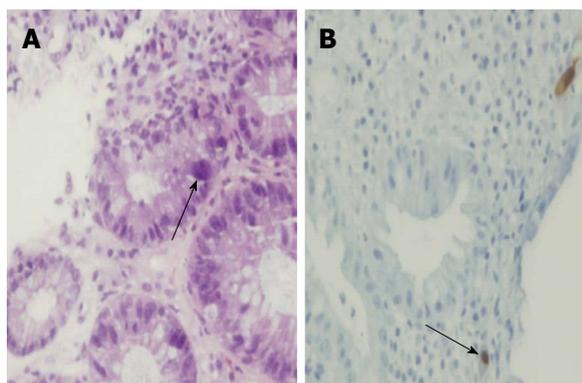
### Patient characteristics

Thirteen patients were included in the study, 7 of whom received antivirals. The clinical and demographic characteristics of the patients are shown in Table 1. Patients in the antiviral-treated group had a longer duration of disease ( $14.2 \pm 9.3$  years *vs*  $3.5 \pm 1.8$  years,  $P = 0.008$ ). Ten patients required hospitalization for severe exacerbation of ulcerative colitis, whereas three were treated as outpatients.

Nine out of the 10 (90%) hospitalized patients were treated with systemic corticosteroids on admission. Four patients received second-line treatment (3 patients-cyclosporine, 1 patient-infliximab).

### Diagnosis of CMV

Immunochemistry for CMV was positive in all patients (Figure 1A). Cytopathic changes consistent with CMV infection in the form of inclusion bodies (Figure 1B) were detectable in only 2/13 (15%) of the patients (both



**Figure 1** Cytomegalovirus demonstrated on a colonic biopsy in a patient with ulcerative colitis (arrows). A: Hematoxylin eosin staining; B: Immunohistochemistry.

patients received antivirals).

Additional tests for detection of CMV infection were performed in only a few of the patients. CMV IgM was assessed in 2 patients, one of them was seropositive. Five patients were tested for CMV DNA in the peripheral blood by quantitative PCR, of whom only one was found to be positive (1900 copies/mL). This patient had a severe and resistant disease and underwent total colectomy during the hospitalization after failure of intravenous corticosteroid, cyclosporine and antiviral therapy.

### Antiviral treatment

Seven patients received antivirals. One patient received oral valgancyclovir as an outpatient for 1 mo. Six patients were treated with intravenous gancyclovir for  $10.3 \pm 7.8$  d. One of the patients died and another underwent colectomy. Three out of 4 remaining patients were discharged with oral valgancyclovir. Cyclosporine was discontinued in 2/3 patients upon detection of CMV in the colonic mucosa.

### Patients' outcomes

In the antiviral-treated group, one patient died from uncontrolled gastroduodenal bleeding and one patient underwent colectomy during the initial hospitalization for severe treatment-resistant colitis. Two additional patients (both received gancyclovir) underwent colectomy during the course of follow-up ( $13 \pm 13$  mo) (3/7 vs 0/6 patients,  $P = 0.19$ ). Thus, 4/7 patients in the anti-viral treated group experienced an adverse outcome (colectomy/death) compared to 0/6 adverse outcomes in those who did not receive antivirals ( $P = 0.07$ ).

### Evolution of CMV status

In 2 patients, negative CMV status (both IHC and HE) was documented on a colonoscopy preceding the index examination. CMV status was assessed by a subsequent colonoscopy in 9 patients (after  $4.7 \pm 5.5$  mo). Immunostaining for CMV was positive in only one of these patients.

## DISCUSSION

In this case series, we have compared the outcome of CMV-positive UC patients who were treated with antivirals with the outcome of patients who received conventional anti-inflammatory therapy. In general, the latter group presented with a milder disease, as reflected by the fact that only half of them (3/6) were hospitalized and none required salvage cyclosporine or infliximab treatment. The long-term outcome of these patients during extended follow-up was also favorable.

Although evidence of CMV infection in the inflamed colonic mucosa of inflammatory bowel disease (IBD) patients is quite common, reportedly higher in steroid-resistant patients<sup>[10,11]</sup>, the actual clinical significance of this finding remains unclear. Cytomegalovirus is trophic for inflamed and replicating tissue and commonly affects patients on systemic immunosuppression<sup>[3,4]</sup>. Evidence of viral shedding and replication is often found in IBD patients, almost exclusively in the inflamed mucosa<sup>[1]</sup>. However, it remains unsettled whether the presence of virus in the tissue is a trigger or a byproduct of the inflammation. Moreover, studies of the outcome of CMV colitis in IBD rarely include a detailed description of the non-treated cohort, thereby hampering our knowledge on the true impact of this viral infection. In addition, the definitions of CMV infection vary significantly and depend on the diagnostic technique employed.

Earlier reports have included steroid-resistant patients with evidence of CMV-induced cytopathic damage on HE staining ("inclusion bodies")<sup>[10,12,13]</sup>. These patients had a severe disease and high rates of colectomy (up to 67%)<sup>[12]</sup>. A detection of inclusion bodies on HE staining is clinically relevant<sup>[3]</sup> and implies an ongoing destruction of colonocytes by the virus. Unfortunately, this technique has low sensitivity (10%-87%)<sup>[1]</sup>, primarily due to a sampling error, and potentially misses a significant number of patients. Immunohistochemistry with a monoclonal antibody targeting early CMV antigen may improve the diagnostic sensitivity to the range of 78%-93%<sup>[1,6]</sup>. Current European Crohn's and Colitis Organization guidelines recommend the combination of HE staining and IHC for detection of CMV infection in patients with UC flare-up<sup>[14]</sup>. In addition to these techniques, CMV DNA can be detected in various substrates, such as serum or full blood, by PCR. Although very sensitive, this method is subject to a significant heterogeneity stemming from employment of non-standardized commercial kits and lack of a standardized cutoff value for active infection. Moreover, there is a poor correlation between viral replication in the blood and the presence or pathogenic viral activity in the gut in IBD patients<sup>[1]</sup>. The same is true for detection of viral antigens in the blood (such as pp65) which is also poorly associated with viral-mediated gut injury.

In addition to its detection in the blood, CMV DNA can be also demonstrated in the colonic tissue by PCR. This highly sensitive technique often leads to the detection of the virus in the absence of histological evidence

**Table 1** Clinical and demographic characteristics of the included patients (mean  $\pm$  SD)

Patient characteristics	Treated (n = 7)	Untreated (n = 6)	P value
Age (yr)	50.0 $\pm$ 14.6	45.0 $\pm$ 13.6	0.540
Gender (male/female)	4/3	3/3	0.400
Extent of disease			
Pancolitis	6	5	0.540
Left-sided	1	1	0.540
Age on diagnosis of UC, yr	35.7 $\pm$ 13.3	41.5 $\pm$ 13.3	0.530
Duration of disease, yr	14.2 $\pm$ 9.3	3.5 $\pm$ 1.8	0.008
Hospitalized patients	6	4	0.560
Prehospitalization treatment			
SC	4	2	0.560
Thiopurines	3	2	1.000
Infliximab	1 <sup>1</sup>	0	1.000
5-asa	5	4	1.000
SC + thiopurines	2	1	1.000
Treatment during hospitalization			
SC	6	3	0.400
Infliximab	1	0	1.000
Cyclosporine	3	0	0.200
Timing of colonoscopy (d)	3.8 $\pm$ 2.4	2.7 $\pm$ 3.4	0.600
Positive cytopathic changes on HE	2	0	0.460
Hospitalization outcome			
Death	1	0	1.000
Colectomy	1	0	1.000
Outcome by the of the follow-up			
Colectomy	3	0	0.190
Death	1	0	1.000

<sup>1</sup>Combined with systemic corticosteroids and thiopurine. Treated: Patients who received antiviral therapy; Untreated: Patients who did not receive antiviral therapy; Timing of colonoscopy: Number of days from hospital admission; SC: Systemic corticosteroids; HE: Hematoxylin eosin; IHC: Immunohistochemistry; UC: Ulcerative colitis.

of tissue damage, thereby possibly representing a remote infection or a low-key viral replication of unclear significance<sup>[1]</sup>.

Recently, several reports utilizing quantitative real-time PCR for detection of CMV infection in patients with UC have been published. Yoshino *et al.*<sup>[15]</sup> tested the colonic biopsies of thirty patients with severe immunosuppressive-resistant ulcerative colitis. CMV DNA (defined as > 10 copies/ $\mu$ g by quantitative real-time PCR) was demonstrated in 56.7% of the patients and was limited to the inflamed mucosa. Seventy percent of these patients were treated with gancyclovir and 83.3% of them achieved remission. In contrast, 93.3% of the CMV-DNA negative patients achieved remission with immunosuppressive therapy. In the study by Roblin *et al.*<sup>[6]</sup>, CMV DNA load above 250 copies/mg in tissue was predictive of resistance to three successive anti-inflammatory regimens.

The impact of detectable CMV on the clinical course of a UC flare-up is still debated. Some authors suggest that CMV reactivation is associated with a worse clinical outcome and with a treatment-refractory disease<sup>[6,7,11,17,18]</sup>, but other studies have not supported this association<sup>[8,15,19]</sup>. The impact of antiviral therapy on the outcome of CMV-positive patients with UC is also debatable<sup>[4,5]</sup>. Based on several series, the cumulative rate of short-term response

of CMV-positive patients to gancyclovir therapy is 72%<sup>[1,5,11,15]</sup>. The outcome of patients who did not receive gancyclovir has not been extensively reported. Kim *et al.*<sup>[20]</sup> described the outcome of 31 CMV-positive patients with ulcerative colitis. Only steroid-resistant patients were treated with gancyclovir and 11/14 responded to the treatment. All the steroid responsive CMV-positive patients had a favorable outcome regardless of anti-viral treatment. In the study by Roblin *et al.*<sup>[6]</sup>, 37.5% (6/16) of patients with CMV DNA load > 250 copies/mg still responded to successive lines of immunosuppressive therapy. This could indicate that CMV presence in the tissue may not in itself preclude a possible response to intensified immunosuppression without antivirals.

In this context, our study provides several important observations. Primarily, the outcome of patients who were not treated with antivirals was favorable (no deaths or colectomies). However, the patients in the antiviral-treated group seem to have presented with a more severe disease, as reflected by their greater need for rescue cyclosporine/infliximab therapy and by their greater need of hospitalization. Antiviral therapy was usually withheld if clinical improvement was noted by the time CMV results were received from the laboratory. Therefore, there was probably a bias towards administration of antiviral therapy to the patients with a more severe disease who failed to improve on standard therapy during their hospitalization. It appears that the decision to start antiviral therapy was guided by the severity of the disease rather than merely the histological findings. The underlying severity of the disease is probably responsible for the inferior clinical outcome of the patients who received antiviral therapy, indicating once more that the sheer presence of CMV in tissue probably does not in itself dictate the clinical outcome.

Interestingly, 9/13 patients were tested for the presence of CMV by IHC on a subsequent colonoscopy and only one patient (who was not treated with antivirals) was positive. This is consistent with the findings reported by Matsuoka *et al.*<sup>[9]</sup>, who demonstrated frequent cycles of reactivation (defined by positive CMV-antigenemia or plasma PCR) and spontaneous clearance of CMV in immunosuppressed patients<sup>[21-23]</sup>. Therefore, in at least a subgroup of patients with exacerbation of UC, the presence of CMV may be an epiphenomenon of the inflammatory process rather than a causative agent.

Our study has several important limitations, primarily stemming from its retrospective design and small sample size. Long-term follow-up was available for only a minority of the patients. The treatment and control groups were significantly different regarding severity of the disease. Indeed, the decision to withhold antiviral therapy was generally adopted for patients who improved clinically by the time the histological results were available, thereby underlining once more the dissimilarity between the two groups. Finally, we did not routinely perform assays for CMV IgM, CMV DNA or pp65 in the serum. None of these methods reliably reflect the presence of

the virus in the colonic tissue, although they are useful in diagnosis of disseminated CMV infection.

Despite these limitations, our findings imply that patients with clear histological evidence of CMV presence in the colonic tissue may not universally require antiviral therapy and may respond to conventional anti-inflammatory therapy. The presence of CMV does not necessarily bear a significant impact on the course of the flare-up in all patients. In particular, less severe patients may probably be treated safely with conventional anti-inflammatory therapy alone, as long as they are responsive. Clearly, larger prospective placebo-controlled trials are called for in order to resolve the etiological role of CMV in severe UC and to elucidate the benefit of anti-viral treatment for these cases. Such studies, preferably utilizing a quantitative rather than qualitative CMV-detection technique, may help to establish a threshold differentiating active infection from low key reactivation and may therefore prove useful in guiding the management of suspected CMV involvement in UC patients.

## COMMENTS

### Background

Cytomegalovirus (CMV) is often present in the mucosa of patients with ulcerative colitis. However, the clinical impact of this finding on the course of the disease and the impact of antiviral therapy on the clinical outcome are still subject to significant debate.

### Research frontiers

In the retrospective cohort, the patients who were treated with antivirals were hospitalized with a more severe disease and had more adverse outcomes compared to the patients who did not receive antivirals. This higher rate of adverse outcome probably stems from the severity of the disease rather than the treatment itself.

### Innovations and breakthroughs

These findings suggest that CMV-positive ulcerative colitis (UC) patients with relatively mild disease may not require specific antiviral treatment as long as they respond to the conventional anti-inflammatory treatment.

### Applications

This is a small retrospective study. Large prospective controlled studies are required in order to identify the optimal treatment strategy in CMV-positive UC patients.

### Terminology

CMV belongs to the family of herpesviridae. It is a very frequent pathogen in healthy subjects as well as immunocompromised patients. CMV can be frequently demonstrated in the mucosa of patients with ulcerative colitis by a conventional staining with hematoxylin and eosin or by immunostaining (immunohistochemistry) with specific antibodies that are able to identify the traces of the viral genome in the tissue.

### Peer review

The results are well presented and discussed in a very balanced and comprehensive way. Despite the small number of patients included in this study, the work deserves publication as it may have important implications for the treatment of ulcerative colitis patients.

## REFERENCES

- 1 **Lawlor G**, Moss AC. Cytomegalovirus in inflammatory bowel disease: pathogen or innocent bystander? *Inflamm Bowel Dis* 2010; **16**: 1620-1627 [PMID: 20232408 DOI: 10.1002/ibd.21275]
- 2 **Manuel O**, Perrotet N, Pascual M. Valganciclovir to prevent or treat cytomegalovirus disease in organ transplantation. *Expert Rev Anti Infect Ther* 2011; **9**: 955-965 [PMID: 22029513 DOI: 10.1586/eri.11.116]
- 3 **Kandiel A**, Lashner B. Cytomegalovirus colitis complicating inflammatory bowel disease. *Am J Gastroenterol* 2006; **101**: 2857-2865 [PMID: 17026558 DOI: 10.1111/j.1572-0241.2006.00869.x]
- 4 **Hommel DW**, Sterringa G, van Deventer SJ, Tytgat GN, Weel J. The pathogenicity of cytomegalovirus in inflammatory bowel disease: a systematic review and evidence-based recommendations for future research. *Inflamm Bowel Dis* 2004; **10**: 245-250 [PMID: 15290919 DOI: 10.1097/00054725-200405000-00011]
- 5 **Domènech E**, Vega R, Ojanguren I, Hernández A, Garcia-Planella E, Bernal I, Rosinach M, Boix J, Cabré E, Gassull MA. Cytomegalovirus infection in ulcerative colitis: a prospective, comparative study on prevalence and diagnostic strategy. *Inflamm Bowel Dis* 2008; **14**: 1373-1379 [PMID: 18452205 DOI: 10.1002/ibd.20498]
- 6 **Kambham N**, Vij R, Cartwright CA, Longacre T. Cytomegalovirus infection in steroid-refractory ulcerative colitis: a case-control study. *Am J Surg Pathol* 2004; **28**: 365-373 [PMID: 15104299 DOI: 10.1097/0000478-200403000-00009]
- 7 **Minami M**, Ohta M, Ohkura T, Ando T, Ohmiya N, Niwa Y, Goto H. Cytomegalovirus infection in severe ulcerative colitis patients undergoing continuous intravenous cyclosporine treatment in Japan. *World J Gastroenterol* 2007; **13**: 754-760 [PMID: 17278199]
- 8 **Lévêque N**, Brixi-Benmansour H, Reig T, Renois F, Tal-mud D, Brodard V, Coste JF, De Champs C, Andréoletti L, Diebold MD. Low frequency of cytomegalovirus infection during exacerbations of inflammatory bowel diseases. *J Med Virol* 2010; **82**: 1694-1700 [PMID: 20827767 DOI: 10.1002/jmv.21877]
- 9 **Matsuoka K**, Iwao Y, Mori T, Sakuraba A, Yajima T, Hisamatsu T, Okamoto S, Morohoshi Y, Izumiya M, Ichikawa H, Sato T, Inoue N, Ogata H, Hibi T. Cytomegalovirus is frequently reactivated and disappears without antiviral agents in ulcerative colitis patients. *Am J Gastroenterol* 2007; **102**: 331-337 [PMID: 17156136 DOI: 10.1111/j.1572-0241.2006.00989.x]
- 10 **Berk T**, Gordon SJ, Choi HY, Cooper HS. Cytomegalovirus infection of the colon: a possible role in exacerbations of inflammatory bowel disease. *Am J Gastroenterol* 1985; **80**: 355-360 [PMID: 2859801]
- 11 **Cottone M**, Pietrosi G, Martorana G, Casà A, Pecoraro G, Oliva L, Orlando A, Rosselli M, Rizzo A, Pagliaro L. Prevalence of cytomegalovirus infection in severe refractory ulcerative and Crohn's colitis. *Am J Gastroenterol* 2001; **96**: 773-775 [PMID: 11280549 DOI: 10.1111/j.1572-0241.2001.03620.x]
- 12 **Begos DG**, Rappaport R, Jain D. Cytomegalovirus infection masquerading as an ulcerative colitis flare-up: case report and review of the literature. *Yale J Biol Med* 1996; **69**: 323-328 [PMID: 9273986]
- 13 **Loftus EV**, Alexander GL, Carpenter HA. Cytomegalovirus as an exacerbating factor in ulcerative colitis. *J Clin Gastroenterol* 1994; **19**: 306-309 [PMID: 7876511 DOI: 10.1097/00004836-199412000-00010]
- 14 **Rahier JF**, Ben-Horin S, Chowers Y, Conlon C, De Munter P, D'Haens G, Domènech E, Eliakim R, Eser A, Frater J, Gassull M, Giladi M, Kaser A, Lémann M, Moreels T, Moschen A, Pollok R, Reinisch W, Schunther M, Stange EF, Tilg H, Van Assche G, Vigez N, Vucelic B, Walsh A, Weiss G, Yazdandanpanah Y, Zabana Y, Travis SP, Colombel JF. European evidence-based Consensus on the prevention, diagnosis and management of opportunistic infections in inflammatory bowel disease. *J Crohns Colitis* 2009; **3**: 47-91 [PMID: 21172250 DOI: 10.1016/j.crohns.2009.02.010]
- 15 **Yoshino T**, Nakase H, Ueno S, Uza N, Inoue S, Mikami S, Matsuura M, Ohmori K, Sakurai T, Nagayama S, Hasegawa S, Sakai Y, Chiba T. Usefulness of quantitative real-time PCR assay for early detection of cytomegalovirus infection in patients with ulcerative colitis refractory to immunosup-

- pressive therapies. *Inflamm Bowel Dis* 2007; **13**: 1516-1521 [PMID: 17828781 DOI: 10.1002/ibd.20253]
- 16 **Roblin X**, Pillet S, Oussalah A, Berthelot P, Del Tedesco E, Phelip JM, Chambonnière ML, Garraud O, Peyrin-Biroulet L, Pozzetto B. Cytomegalovirus load in inflamed intestinal tissue is predictive of resistance to immunosuppressive therapy in ulcerative colitis. *Am J Gastroenterol* 2011; **106**: 2001-2008 [PMID: 21788989 DOI: 10.1038/ajg.2011.202]
  - 17 **Kishore J**, Ghoshal U, Ghoshal UC, Krishnani N, Kumar S, Singh M, Ayyagari A. Infection with cytomegalovirus in patients with inflammatory bowel disease: prevalence, clinical significance and outcome. *J Med Microbiol* 2004; **53**: 1155-1160 [PMID: 15496396 DOI: 10.1099/jmm.0.45629-0]
  - 18 **Papadakis KA**, Tung JK, Binder SW, Kam LY, Abreu MT, Targan SR, Vasilias EA. Outcome of cytomegalovirus infections in patients with inflammatory bowel disease. *Am J Gastroenterol* 2001; **96**: 2137-2142 [PMID: 11467645 DOI: 10.1111/j.1572-0241.2001.03949.x]
  - 19 **Dimitroulia E**, Spanakis N, Konstantinidou AE, Legakis NJ, Tsakris A. Frequent detection of cytomegalovirus in the intestine of patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2006; **12**: 879-884 [PMID: 16954807 DOI: 10.1097/01.mib.0000231576.11678.57]
  - 20 **Kim YS**, Kim YH, Kim JS, Cheon JH, Ye BD, Jung SA, Park YS, Choi CH, Jang BI, Han DS, Yang SK, Kim WH. Cytomegalovirus infection in patients with new onset ulcerative colitis: a prospective study. *HepatoGastroenterology* 2012; **59**: 1098-1101 [PMID: 22580660 DOI: 10.5754/hge10217]
  - 21 **Lavagna A**, Bergallo M, Daperno M, Sostegni R, Costa C, Leto R, Crocellà L, Molinaro G, Rocca R, Cavallo R, Pera A. Infliximab and the risk of latent viruses reactivation in active Crohn's disease. *Inflamm Bowel Dis* 2007; **13**: 896-902 [PMID: 17345605 DOI: 10.1002/ibd.20131]
  - 22 **Scheinberg P**, Fischer SH, Li L, Nunez O, Wu CO, Sloand EM, Cohen JI, Young NS, John Barrett A. Distinct EBV and CMV reactivation patterns following antibody-based immunosuppressive regimens in patients with severe aplastic anemia. *Blood* 2007; **109**: 3219-3224 [PMID: 17148582 DOI: 10.1182/blood-2006-09-045625]
  - 23 **Torre-Cisneros J**, Del Castillo M, Castón JJ, Castro MC, Pérez V, Collantes E. Infliximab does not activate replication of lymphotropic herpesviruses in patients with refractory rheumatoid arthritis. *Rheumatology (Oxford)* 2005; **44**: 1132-1135 [PMID: 15927999 DOI: 10.1093/rheumatology/keh696]

**P- Reviewer** Nevels M **S- Editor** Song XX  
**L- Editor** Roemmele A **E- Editor** Lu YJ



## Serum cytokine profile in patients with hepatitis B e antigen-negative chronic active hepatitis B and inactive hepatitis B virus carriers

Dimitra Dimitropoulou, Marina Karakantza, Georgios L Theodorou, Lydia Leonidou, Stelios F Assimakopoulos, Athanasia Mouzaki, Charalambos A Gogos

Dimitra Dimitropoulou, Lydia Leonidou, Stelios F Assimakopoulos, Charalambos A Gogos, Department of Internal Medicine, University Hospital of Patras, 26504 Patras, Greece  
Marina Karakantza, Georgios L Theodorou, Athanasia Mouzaki, Division of Hematology, Department of Internal Medicine, University Hospital of Patras, 26504 Patras, Greece  
Author contributions: Gogos CA, Karakantza M and Mouzaki A designed the study; Dimitropoulou D, Theodorou GL and Leonidou L acquired the data; Assimakopoulos SF performed the statistical analyses; Dimitropoulou D and Gogos CA interpreted the results; Dimitropoulou D wrote the paper; Gogos CA and Assimakopoulos SF critically revised the manuscript for intellectual content.

Correspondence to: Stelios F Assimakopoulos, MD, PhD, Department of Internal Medicine, University Hospital of Patras, 26504 Patras, Greece. [sassim@upatras.gr](mailto:sassim@upatras.gr)

Telephone: +30-2610-999583 Fax: +30-2610-993982

Received: July 20, 2012 Revised: December 5, 2012

Accepted: January 29, 2013

Published online: February 15, 2013

### Abstract

An insufficient cellular immune response seems to be critical for the immunopathogenesis of chronic hepatitis B virus infection. We have previously demonstrated no differences of T-lymphocyte subsets in blood between inactive hepatitis B s antigen (HBsAg) carriers and patients with HBeAg-negative chronic active hepatitis B. This study investigated the peripheral blood cytokine profile in patients with HBeAg-negative chronic active hepatitis B infection (Group A,  $n = 21$ ) and inactive HBsAg carriers (Group B,  $n = 13$ ). Serum cytokines [interferon (IFN)- $\gamma$ , tumor necrosis factor- $\alpha$ , interleukin (IL)-1b, IL-4, IL-12, IL-10, IL-2, IL-5, IL-8] were analyzed by using flow cytometry. Patients with chronic active disease presented with significantly decreased levels of IFN- $\gamma$  and IL-10 compared to inac-

tive carriers ( $P = 0.048$  and  $P = 0.008$ , respectively). In HBeAg-negative chronic active hepatitis B patients, a significant negative correlation of IFN- $\gamma$  levels with serum hepatitis B viral load was noted ( $P = 0.021$ ). In conclusion, patients with HBeAg-negative chronic active hepatitis B and HBsAg inactive carriers display a different cytokine profile. Decreased Th1 response observed in patients with chronic active hepatitis B could be implicated in the persistence of virus replication and ongoing progression of liver disease.

© 2013 Baishideng. All rights reserved.

**Key words:** Cytokines; Hepatitis B; Flow cytometry; Immunoreactive fibronectin- $\gamma$

Dimitropoulou D, Karakantza M, Theodorou GL, Leonidou L, Assimakopoulos SF, Mouzaki A, Gogos CA. Serum cytokine profile in patients with hepatitis B e antigen-negative chronic active hepatitis B and inactive hepatitis B virus carriers. *World J Gastrointest Pathophysiol* 2013; 4(1): 24-27 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i1/24.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i1.24>

### TO THE EDITOR

Chronic hepatitis B (CHB) is a highly heterogeneous disease regarding the levels of virus replication, liver disease activity and humoral responses. Hepatitis B virus (HBV) is not directly cytopathic for the infected cells and the host immune response, mainly the T-cell-mediated, plays a pivotal role in the immunopathogenesis of hepatitis B<sup>[1,2]</sup>. To date, there are only limited studies examining the immune responses in hepatitis B e antigen (HBeAg)-negative CHB, which is the main type of CHB in Greece and other Mediterranean countries. In a recent study,

**Table 1** Characteristics of hepatitis B e antigen-negative chronic active hepatitis B patients and asymptomatic hepatitis B virus carriers

Parameters	Group A (n = 21)	Group B (n = 13)	P value
Males (%)	12/21 (57)	7/13 (54)	NS
Age (yr)	44 (19-60)	39.5 (32-47)	NS
HBV DNA (c/mL)	1.25×10 <sup>6</sup> (0.04×10 <sup>6</sup> -3×10 <sup>6</sup> )	< 1000	N/A
ALT (IU/L)	90 (73-108)	32 (22-39)	< 0.001
AST (IU/L)	85 (69-102)	28 (17-39)	< 0.001
Histopathology		N/A	
HAI score			
Category A	2 (1-4)		
Category B	0 (0-1)		
Category C	2 (0-3)		
Category D	3 (1-4)		
Total score	7 (4-13)		
Stage	2		

Data expressed as median (min-max), upper limit of normal for both aminotransferases: 40 IU/L. Group A: Hepatitis B e antigen-negative chronic active hepatitis B patients; Group B: Asymptomatic hepatitis B virus carriers. NS: Non-significant; NA: Not applicable; HBV: Hepatitis B virus; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; HAI: Hepatitis activity index.

investigating T-lymphocyte subsets in peripheral blood and liver tissue of patients with HBeAg-negative CHB, we demonstrated evidence of an insufficient cellular immune response that might be critical for the ineffective virus clearance and liver damage in CHB<sup>[3]</sup>. However, no differences in T-lymphocyte subsets in blood were detected between inactive HBsAg carriers and patients with HBeAg-negative chronic active hepatitis B. Therefore, the question of how most HBsAg patients are able to maintain a low replication level and mild liver inflammation (inactive carriers), while a number of them develop chronic active hepatitis with an enhanced HBV replication level and severe liver damage, remains unanswered. The present study was focused on this specific question, investigating the produced cytokine profiles in patients with HBeAg (-) chronic active HBV infection and chronic inactive HBsAg carriers.

The study enrolled twenty-one patients with positive serum HBsAg for at least 6 mo, positive serum HBV DNA with high viral load (> 20 000 copies/mL), measured at least twice in a period of 12 mo, and aminotransferase levels higher than twice the upper normal limits, who were not currently treated nor had ever been treated with any antiviral agent (HBeAg-negative chronic hepatitis B - Group A) and thirteen patients with positive serum HBsAg for at least 6 mo, undetectable HBV DNA and normal serum aminotransferase levels (inactive carriers - Group B). All HBV infected patients were HBeAg negative, anti-Hbe positive and anti-HDV negative. Exclusion criteria were the presence of decompensated cirrhosis, HBV/HBV co-infection, alcohol abuse, human immunodeficiency virus or human T-cell lymphoma virus infection, any immunosuppressive treatment and other liver diseases, such as drug hepatotoxicity,  $\alpha$ -1 antitrypsin deficiency, Wilson's disease, hemochromatosis, autoimmune hepatitis and liver cancer. Liver biopsies were obtained only from the patients with

positive serum HBV DNA and elevated liver enzymes and were indicative of chronic active hepatitis B infection. In each biopsy, several histological features were assessed and finally, the hepatitis activity index was applied and the architectural grade recorded<sup>[4]</sup>.

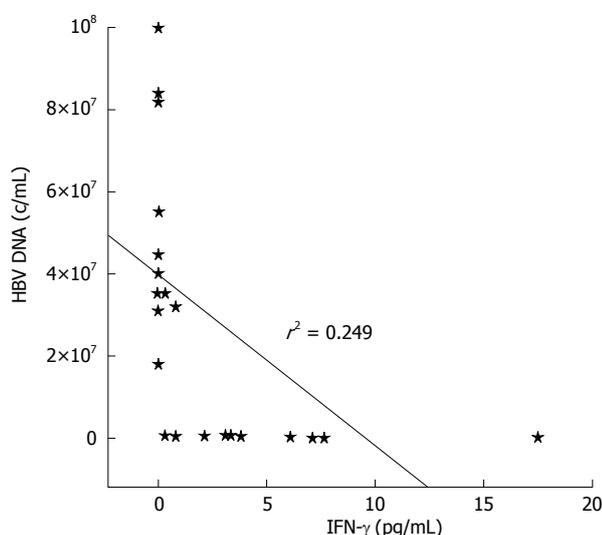
Blood tests were obtained before the initiation of any kind of treatment and analyzed for serum aminotransferases with an automatic Olympus AU 640 system (Olympus, Rungis, France), whilst serum HBV DNA load was assessed with the real time fluorescent quantitative polymerase chain reaction method (real time PCR), with a lower limit of detection of about 1000 viral genome copies/mL. Serum cytokine levels [interferon (IFN)- $\gamma$ , tumor necrosis factor- $\alpha$ , interleukin (IL)-1b, IL-4, IL-12, IL-10, IL-2, IL-5, IL-8] were evaluated by using a commercially available Flow Cytomix Human Basic Kit Assay (Bender MedSystems, Vienna, Austria), following the manufacturer's instructions. Quantitation measurements were performed by flow cytometer instrument FC 500 and accompanying CXP Software (Beckman Coulter, Calif, United States). Flow Cytomix Pro 2.3 Software was used to perform calculations (Bender MedSystems). Standard curves for each cytokine were generated with manufacturer-supplied reference analyte (pg/mL concentrations). Statistical analyses were performed using the Mann-Whitney *U* test since data was not normally distributed (Shapiro-Wilk Test). Correlation between paired variables in patients with HBeAg (-) chronic active hepatitis B was estimated by a non-parametric Spearman correlation test. In all cases, a *P*-value of less than 0.05 was considered as significant.

Patients' characteristics, serum aminotransferases and HBV DNA levels are shown in Table 1. Inactive HBV carriers (Group B) had significantly increased production of IFN- $\gamma$  and IL-10 cytokines compared with HBeAg-negative chronic active hepatitis B patients (Group A) (*P* = 0.048 and *P* = 0.008, respectively, Table 2). In HBeAg-negative

**Table 2** Cytokine levels in hepatitis B e antigen-negative chronic active hepatitis B patients and asymptomatic hepatitis B virus carriers

Cytokines (pg/mL)	Group A (n = 21)	Group B (n = 13)	P value
IFN- $\gamma$	0.3 (0-17.5)	18.3 (0-5137)	0.048
TNF- $\alpha$	101.3 (0-462.2)	202.6 (0-1044)	NS
IL-1b	98.5 (0-25196)	61.0 (0-11149)	NS
IL-4	285.4 (0-10885)	4.9 (0-4313)	NS
IL-12	154.6 (0-39042)	172.6 (0-10618)	NS
IL-10	0 (0-41.4)	18.6 (0-14393)	0.008
IL-8	314.8 (51.1-32592)	383.0 (67.8-14075)	NS
IL-5	78.4 (0-515.5)	198.0 (0-853.7)	NS
IL-2	158.9 (28.2-126842)	146.4 (29.5-163730)	NS

Data expressed as median (min-max). Group A: Hepatitis B e antigen-negative chronic active hepatitis B patients; Group B: Asymptomatic HBV carriers. NS: Non-significant; IFN- $\gamma$ : Interferon- $\gamma$ ; TNF- $\alpha$ : Tumor necrosis factor- $\alpha$ ; IL: Interleukin.



**Figure 1** Correlation between serum-interferon- $\gamma$  levels and viral load in patients with hepatitis B e antigen (-) chronic active hepatitis B. IFN- $\gamma$ : Interferon- $\gamma$ ; HBV: Hepatitis B virus.

chronic active hepatitis B patients, a significant negative correlation between serum HBV viral load and IFN- $\gamma$  production was noted (Figure 1), whilst no correlation existed between IFN- $\gamma$  and alanine aminotransferase levels.

This study demonstrates that patients with HBeAg negative chronic hepatitis B display a different cytokine profile depending on the degree of viremia and liver inflammation. A potential limitation of the present study is the relatively small number of patients included. According to our results, HBeAg inactive carriers displayed a strong production of IFN- $\gamma$  (Th1 type immune response) and IL-10 (Th2 type immune response) in peripheral blood compared to patients with HBeAg-negative chronic active hepatitis B. Both Th1 and Th2 T-cells mediate humoral and cellular immunity able to neutralize HBV by antibodies and inhibit HBV replication through cytokines<sup>[5]</sup>. Therefore, we can speculate that HBeAg inactive carriers suppress HBV replication through their capability to produce the Th1 type antiviral cytokine

IFN- $\gamma$ . In support of this theory, we demonstrated a negative correlation between IFN- $\gamma$  and the levels of viremia in chronic active hepatitis B patients; however, the other side of the coin might be that the continuing presence of viral load in serum could induce an impairment of IFN- $\gamma$ <sup>[6-8]</sup>. Normal aminotransferases levels in HBeAg inactive carriers might indicate that IFN- $\gamma$  promotes viral clearance through non-cytolytic mechanism(s)<sup>[9-11]</sup>. Alternatively, it could be explained by a counterbalancing effect of the observed increased production of IL-10 (Th2 type cytokine) on the excessive Th1 action, although IL-10 exerts a regulatory effect on Th2 type response as well<sup>[9-11]</sup>.

In conclusion, this study demonstrates that T-cell immunity is functionally impaired in chronic active hepatitis B patients. In addition, an inverse correlation was shown between the increase of one of the major determinants of Th1 response (IFN- $\gamma$  cytokine) and the decline of HBV load in blood samples of patients with chronic active hepatitis B. These findings suggest that impaired immunity could be associated with the persistence of HBV load and the elevation of serum aminotransferases in patients with active disease. On this basis, we are tempted to speculate that not only drugs with antiviral potency but also immunomodulating agents that can restore T cell function might be effective for a successful treatment of chronic HBV infection.

## REFERENCES

- 1 **Ferrari C**, Penna A, Bertoletti A, Valli A, Antoni AD, Giuberti T, Cavalli A, Petit MA, Fiaccadori F. Cellular immune response to hepatitis B virus-encoded antigens in acute and chronic hepatitis B virus infection. *J Immunol* 1990; **145**: 3442-3449 [PMID: 2230128]
- 2 **Chisari FV**, Ferrari C. Hepatitis B virus immunopathogenesis. *Annu Rev Immunol* 1995; **13**: 29-60 [PMID: 7612225 DOI: 10.1146/annurev.iy.13.040195.000333]
- 3 **Dimitropoulou D**, Karakantza M, Tsamandas AC, Mouzaki A, Theodorou G, Gogos CA. T-lymphocyte subsets in peripheral blood and liver tissue of patients with chronic hepatitis B and C. *In Vivo* 2011; **25**: 833-840 [PMID: 21753143]

- 4 **Ishak K**, Baptista A, Bianchi L, Callea F, De Groote J, Gudat F, Denk H, Desmet V, Korb G, MacSween RN. Histological grading and staging of chronic hepatitis. *J Hepatol* 1995; **22**: 696-699 [PMID: 7560864]
- 5 **Guidotti LG**. The role of cytotoxic T cells and cytokines in the control of hepatitis B virus infection. *Vaccine* 2002; **20** Suppl 4: A80-A82 [PMID: 12477433 DOI: 10.1016/S0264-410X(02)00392-4]
- 6 **Bertoletti A**, Gehring AJ. The immune response during hepatitis B virus infection. *J Gen Virol* 2006; **87**: 1439-1449 [PMID: 16690908 DOI: 10.1099/vir.0.81920-0]
- 7 **Baumert TF**, Thimme R, von Weizsäcker F. Pathogenesis of hepatitis B virus infection. *World J Gastroenterol* 2007; **13**: 82-90 [PMID: 17206757]
- 8 **Lai CL**, Yuen MF. The natural history of chronic hepatitis B. *J Viral Hepat* 2007; **14** Suppl 1: 6-10 [PMID: 17958636 DOI: 10.1111/j.1365-2893.2007.00909.x]
- 9 **Bertoletti A**, Maini M, Williams R. Role of hepatitis B virus specific cytotoxic T cells in liver damage and viral control. *Antiviral Res* 2003; **60**: 61-66 [PMID: 14638399 DOI: 10.1016/j.antiviral.2003.08.012]
- 10 **Guidotti LG**, Ando K, Hobbs MV, Ishikawa T, Runkel L, Schreiber RD, Chisari FV. Cytotoxic T lymphocytes inhibit hepatitis B virus gene expression by a noncytolytic mechanism in transgenic mice. *Proc Natl Acad Sci USA* 1994; **91**: 3764-3768 [PMID: 8170985 DOI: 10.1073/pnas.91.9.3764]
- 11 **Guidotti LG**, Rochford R, Chung J, Shapiro M, Purcell R, Chisari FV. Viral clearance without destruction of infected cells during acute HBV infection. *Science* 1999; **284**: 825-829 [PMID: 10221919 DOI: 10.1126/science.284.5415.825]

P- Reviewer Imazeki F S- Editor Zhai HH

L- Editor Roemmele A E- Editor Yan JL



**GENERAL INFORMATION**

*World Journal of Gastrointestinal Pathophysiology* (*World J Gastrointest Pathophysiol*, *WJGP*, online ISSN 2150-5330, DOI: 10.4291) is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

**Aim and scope**

*WJGP* is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. *WJGP* specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.

We encourage authors to submit their manuscripts to *WJGP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

*WJGP* is edited and published by Baishideng Publishing Group (BPG). BPG has a strong professional editorial team composed of science editors, language editors and electronic editors. BPG currently publishes 42 OA clinical medical journals, including 41 in English, has a total of 15 471 editorial board members or peer reviewers, and is a world first-class publisher.

**Columns**

The columns in the issues of *WJGP* will include: (1) Editorial: The editorial board members are invited to make comments on an important topic in their field in terms of its current research status and future directions to lead the development of this discipline; (2) Frontier: The editorial board members are invited to select a highly cited cutting-edge original paper of his/her own to summarize major findings, the problems that have been resolved and remain to be resolved, and future research directions to help readers understand his/her important academic point of view and future research directions in the field; (3) Diagnostic Advances: The editorial board members are invited to write high-quality diagnostic advances in their field to improve the diagnostic skills of readers. The topic covers general clinical diagnosis, differential diagnosis, pathological diagnosis, laboratory diagnosis, imaging diagnosis, endoscopic diagnosis, biotechnological diagnosis, functional diagnosis, and physical diagnosis; (4) Therapeutics Advances: The editorial board members are invited to write high-quality therapeutic advances in their field to help improve the therapeutic skills of readers. The topic covers medication therapy, psychotherapy, physical therapy, replacement therapy, interventional therapy, minimally invasive therapy, endoscopic therapy, transplantation therapy, and surgical therapy; (5) Field of Vision: The editorial board members are invited to write commentaries on classic articles, hot topic articles, or latest articles to keep readers at the forefront of research and increase their levels of clinical research. Classic articles refer to papers that are included in Web of Knowledge and have received a large number of citations (ranking in the top 1%) after being published for more than years, reflecting the quality and impact of papers. Hot topic articles refer to papers that are included in

Web of Knowledge and have received a large number of citations after being published for no more than 2 years, reflecting cutting-edge trends in scientific research. Latest articles refer to the latest published high-quality papers that are included in PubMed, reflecting the latest research trends. These commentary articles should focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions. Basic information about the article to be commented (including authors, article title, journal name, year, volume, and inclusive page numbers); (6) Minireviews: The editorial board members are invited to write short reviews on recent advances and trends in research of molecular biology, genomics, and related cutting-edge technologies to provide readers with the latest knowledge and help improve their diagnostic and therapeutic skills; (7) Review: To make a systematic review to focus on the status quo of research, the most important research topics, the problems that have now been resolved and remain to be resolved, and future research directions; (8) Topic Highlight: The editorial board members are invited to write a series of articles (7-10 articles) to comment and discuss a hot topic to help improve the diagnostic and therapeutic skills of readers; (9) Medical Ethics: The editorial board members are invited to write articles about medical ethics to increase readers' knowledge of medical ethics. The topic covers international ethics guidelines, animal studies, clinical trials, organ transplantation, etc.; (10) Clinical Case Conference or Clinicopathological Conference: The editorial board members are invited to contribute high-quality clinical case conference; (11) Original Articles: To report innovative and original findings in gastrointestinal pathophysiology; (12) Brief Articles: To briefly report the novel and innovative findings in gastrointestinal pathophysiology; (13) Meta-Analysis: To evaluate the clinical effectiveness in gastrointestinal pathophysiology by using data from two or more randomised control trials; (14) Case Report: To report a rare or typical case; (15) Letters to the Editor: To discuss and make reply to the contributions published in *WJGP*, or to introduce and comment on a controversial issue of general interest; (16) Book Reviews: To introduce and comment on quality monographs of gastrointestinal pathophysiology; and (17) Autobiography: The editorial board members are invited to write their autobiography to provide readers with stories of success or failure in their scientific research career. The topic covers their basic personal information and information about when they started doing research work, where and how they did research work, what they have achieved, and their lessons from success or failure.

**Name of journal**

*World Journal of Gastrointestinal Pathophysiology*

**ISSN**

ISSN 2150-5330 (online)

**Launch date**

April 26, 2010

**Frequency**

Quarterly

**Editor-in-Chief**

Thomas Y Ma, MD, PhD, Professor, Chief, Division of Gastro-

## Instructions to authors

enterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States

### Editorial office

Jin-Lei Wang, Director

Xiu-Xia Song, Vice Director

*World Journal of Gastrointestinal Pathophysiology*

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-59080039

Fax: +86-10-85381893

E-mail: [wjgp@wjgnet.com](mailto:wjgp@wjgnet.com)

<http://www.wjgnet.com>

### Publisher

Baishideng Publishing Group Co., Limited

Flat C, 23/F, Lucky Plaza,

315-321 Lockhart Road, Wan Chai,

Hong Kong, China

Telephone: +852-6555-7188

Fax: +852-3177-9906

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

<http://www.wjgnet.com>

### Production center

Beijing Baishideng BioMed Scientific Co., Limited

Room 903, Building D, Ocean International Center,

No. 62 Dongsihuan Zhonglu, Chaoyang District,

Beijing 100025, China

Telephone: +86-10-85381892

Fax: +86-10-85381893

### Representative office

USA Office

8226 Regency Drive,

Pleasanton, CA 94588-3144, United States

### Instructions to authors

Full instructions are available online at [http://www.wjgnet.com/2150-5330/g\\_info\\_20100316161927.htm](http://www.wjgnet.com/2150-5330/g_info_20100316161927.htm).

### Indexed and Abstracted in

PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.

---

## SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

### Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Redit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

### Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any poten-

tial bias, *WJGP* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: [http://www.icmje.org/ethical\\_4conflicts.html](http://www.icmje.org/ethical_4conflicts.html).

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

### Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

### Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

---

## SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we en-

courage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

### Online submissions

Manuscripts should be submitted through the Online Submission System at: <http://www.wjgnet.com/esps/>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS ([http://www.wjgnet.com/2150-5330/g\\_info\\_20100316161927.htm](http://www.wjgnet.com/2150-5330/g_info_20100316161927.htm)) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to [wjgp@wjgnet.com](mailto:wjgp@wjgnet.com), or by telephone: +86-10-85381892. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

## MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

### Title page

**Title:** Title should be less than 12 words.

**Running title:** A short running title of less than 6 words should be provided.

**Authorship:** Authorship credit should be in accordance with the standard proposed by ICMJE, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

**Institution:** Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

**Author contributions:** The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

**Supportive foundations:** The complete name and number of supportive foundations should be provided, e.g. Supported by National Natural Science Foundation of China, No. 30224801

**Correspondence to:** Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor

of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. [montgomery.bissell@ucsf.edu](mailto:montgomery.bissell@ucsf.edu)

**Telephone and fax:** Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g. Telephone: +86-10-85381892 Fax: +86-10-85381893

**Peer reviewers:** All articles received are subject to peer review. Normally, three experts are invited for each article. Decision on acceptance is made only when at least two experts recommend publication of an article. All peer-reviewers are acknowledged on Express Submission and Peer-review System website.

### Abstract

There are unstructured abstracts (no less than 200 words) and structured abstracts. The specific requirements for structured abstracts are as follows:

An informative, structured abstract should accompany each manuscript. Abstracts of original contributions should be structured into the following sections: AIM (no more than 20 words; Only the purpose of the study should be included. Please write the Aim in the form of "To investigate/study/..."), METHODS (no less than 140 words for Original Articles; and no less than 80 words for Brief Articles), RESULTS (no less than 150 words for Original Articles and no less than 120 words for Brief Articles; You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g.  $6.92 \pm 3.86$  vs  $3.61 \pm 1.67$ ,  $P < 0.001$ ), and CONCLUSION (no more than 26 words).

### Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

### Core tip

Please write a summary of less than 100 words to outline the most innovative and important arguments and core contents in your paper to attract readers.

### Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRODUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both.

### Illustrations

Figures should be numbered as 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ... *etc.* It is our principle to publish high resolution-figures for the E-versions.

### Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

### Notes in tables and illustrations

Data that are not statistically significant should not be noted. <sup>a</sup> $P < 0.05$ ,

## Instructions to authors

<sup>b</sup> $P < 0.01$  should be noted ( $P > 0.05$  should not be noted). If there are other series of  $P$  values, <sup>c</sup> $P < 0.05$  and <sup>d</sup> $P < 0.01$  are used. A third series of  $P$  values can be expressed as <sup>e</sup> $P < 0.05$  and <sup>f</sup> $P < 0.01$ . Other notes in tables or under illustrations should be expressed as <sup>1</sup>F, <sup>2</sup>F, <sup>3</sup>F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, etc., in a certain sequence.

### Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

## REFERENCES

### Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability<sup>[1,2]</sup>". If references are cited directly in the text, they should be put together within the text, for example, "From references<sup>[19,22-24]</sup>, we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

### PMID and DOI

Please provide PubMed citation numbers to the reference list, e.g. PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

### Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.539f].

### Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

### Format

#### Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaobua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA*

2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS A Careaction* 2002; 1-6 [PMID: 12154804]

### Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

### Statistical data

Write as mean  $\pm$  SD or mean  $\pm$  SE.

### Statistical expression

Express  $t$  test as  $t$  (in italics),  $F$  test as  $F$  (in italics), chi square test as

$\chi^2$  (in Greek), related coefficient as *r* (in italics), degree of freedom as  $\nu$  (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

### Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h, blood glucose concentration, *c* (glucose)  $6.4 \pm 2.1$  mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5  $\mu\text{g/L}$ ; CO<sub>2</sub> volume fraction, 50 mL/L CO<sub>2</sub>, not 5% CO<sub>2</sub>; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23243641.

The format for how to accurately write common units and quantum numbers can be found at: [http://www.wjgnet.com/2150-5330/g\\_info\\_20100312200347.htm](http://www.wjgnet.com/2150-5330/g_info_20100312200347.htm).

### Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

### Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, *etc.*

Restriction enzymes: *EcoRI*, *HindI*, *BamHI*, *Kho I*, *Kpn I*, *etc.*

Biology: *H. pylori*, *E. coli*, *etc.*

### Examples for paper writing

All types of articles' writing style and requirement will be found in the link: <http://www.wjgnet.com/esps/NavigationInfo.aspx?id=15>

## RESUBMISSION OF THE REVISED MANUSCRIPTS

Authors must revise their manuscript carefully according to the revision policies of Baishideng Publishing Group Co., Limited. The revised version, along with the signed copyright transfer agreement, responses to the reviewers, and English language Grade B certificate (for non-native speakers of English), should be submitted to the

online system via the link contained in the e-mail sent by the editor. If you have any questions about the revision, please send e-mail to [esps@wjgnet.com](mailto:esps@wjgnet.com).

### Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A.

### Copyright assignment form

Please download a Copyright assignment form from [http://www.wjgnet.com/2150-5330/g\\_info\\_20100312200118.htm](http://www.wjgnet.com/2150-5330/g_info_20100312200118.htm).

### Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: [http://www.wjgnet.com/2150-5330/g\\_info\\_20100312195923.htm](http://www.wjgnet.com/2150-5330/g_info_20100312195923.htm).

### Proof of financial support

For papers supported by a foundation, authors should provide a copy of the approval document and serial number of the foundation.

### Links to documents related to the manuscript

WJGP will be initiating a platform to promote dynamic interactions between the editors, peer reviewers, readers and authors. After a manuscript is published online, links to the PDF version of the submitted manuscript, the peer-reviewers' report and the revised manuscript will be put on-line. Readers can make comments on the peer reviewer's report, authors' responses to peer reviewers, and the revised manuscript. We hope that authors will benefit from this feedback and be able to revise the manuscript accordingly in a timely manner.

### Publication fee

WJGP is an international, peer-reviewed, OA online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium and format, provided the original work is properly cited. The use is non-commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. Publication fee: 600 USD per article. All invited articles are published free of charge.

# World Journal of *Gastrointestinal Pathophysiology*

*World J Gastrointest Pathophysiol* 2013 May 15; 4(2): 28-42





**BRIEF ARTICLE**

- 28 Duodenal epithelial transport in functional dyspepsia: Role of serotonin  
*Witte AB, D'Amato M, Poulsen SS, Laurent A, Knuhtsen S, Bindslev N, Hansen MB, Schmidt PT*
- 37 Impact of body mass index for patients undergoing pancreaticoduodenectomy  
*Del Chiaro M, Rangelova E, Ansoerge C, Blomberg J, Segersvård R*

## Contents

*World Journal of Gastrointestinal Pathophysiology*  
Volume 4 Number 2 May 15, 2013

**APPENDIX** I-V Instructions to authors

**ABOUT COVER** **Editorial Board Member of *World Journal of Gastrointestinal Pathophysiology***, Marco Del Chiaro, MD, PhD, Division of Surgery, Department of Clinical Science, Intervention and Technology (CLINTEC), Karolinska Institutet at Karolinska University Hospital, Huddinge K53, 14186 Stockholm, Sweden

### AIM AND SCOPE

*World Journal of Gastrointestinal Pathophysiology* (*World J Gastrointest Pathophysiol*, *WJGP*, online ISSN 2150-5330, DOI: 10.4291), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

*WJGP* is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. *WJGP* specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.

We encourage authors to submit their manuscripts to *WJGP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

### INDEXING/ ABSTRACTING

PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.

**FLYLEAF** I-IV Editorial Board

### EDITORS FOR THIS ISSUE

**Responsible Assistant Editor:** *Xin-Xin Che*  
**Responsible Electronic Editor:** *Jin-Li Yan*  
**Proofing Editor-in-Chief:** *Lian-Sheng Ma*

**Responsible Science Editor:** *Huan-Huan Zhai*

**NAME OF JOURNAL**  
*World Journal of Gastrointestinal Pathophysiology*

**ISSN**  
ISSN 2150-5330 (online)

**LAUNCH DATE**  
April 15, 2010

**Frequency**  
Quarterly

**EDITOR-IN-CHIEF**  
**Thomas Y Ma, MD, PhD, Professor, Chief**, Division of Gastroenterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States

**EDITORIAL OFFICE**  
Jin-Li Wang, Director  
Xiu-Xia Song, Vice Director  
*World Journal of Gastrointestinal Pathophysiology*

Room 903, Building D, Ocean International Center,  
No. 62 Dongsihuan Zhonglu, Chaoyang District,  
Beijing 100025, China  
Telephone: +86-10-85381892  
Fax: +86-10-85381893  
E-mail: [wjgp@wjgnet.com](mailto:wjgp@wjgnet.com)  
<http://www.wjgnet.com>

**PUBLISHER**  
Baishideng Publishing Group Co., Limited  
Flat C, 23/F, Lucky Plaza,  
315-321 Lockhart Road, Wan Chai,  
Hong Kong, China  
Telephone: +852-6555-7188  
Fax: +852-3177-9906  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
<http://www.wjgnet.com>

**PUBLICATION DATE**  
May 15, 2013

### COPYRIGHT

© 2013 Baishideng. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

### SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

### INSTRUCTIONS TO AUTHORS

Full instructions are available online at [http://www.wjgnet.com/2150-5330/g\\_info\\_20100316080008.htm](http://www.wjgnet.com/2150-5330/g_info_20100316080008.htm)

### ONLINE SUBMISSION

<http://www.wjgnet.com/esps/>

## Duodenal epithelial transport in functional dyspepsia: Role of serotonin

Anne-Barbara Witte, Mauro D'Amato, Steen Seier Poulsen, Agneta Laurent, Svend Knuhtsen, Niels Bindsløv, Mark Berner Hansen, Peter Thelin Schmidt

Anne-Barbara Witte, Peter Thelin Schmidt, Department of Medicine Solna, Unit of Gastroenterology and Hepatology, Karolinska University Hospital Solna, Karolinska Institute, SE-171 76 Stockholm, Sweden

Mauro D'Amato, Agneta Laurent, Department of Biosciences and Nutrition, Karolinska Institute Campus Huddinge, 141 86 Stockholm, Sweden

Steen Seier Poulsen, Niels Bindsløv, Department of Biomedical Sciences, Panum Institute, University of Copenhagen, 2200 Copenhagen N, Denmark

Svend Knuhtsen, Mark Berner Hansen, Department of Surgery K, Bispebjerg Hospital, University of Copenhagen, 2400 Copenhagen NV, Denmark

Mark Berner Hansen, AstraZeneca Research and Development, 431 83 Mölndal, Sweden

**Author contributions:** The study was designed by Witte AB, Poulsen SS, Bindsløv N, Hansen MB and Schmidt PT; Witte AB performed the research with the exception of PCR experiments and the histological staining process and analysed as well as summarized the data; Knuhtsen S performed the endoscopic examinations and assisted in characterizing patients and controls; Bindsløv N and Hansen MB provided technical guidance and facilities for biopsy sampling and Ussing experiments; D'Amato M and Laurent A performed the expression studies; Poulsen SS contributed with histological evaluation and guidance in the field; Schmidt PT participated in the data analysis and held overall responsibility for ethical and economic aspects; all authors participated in the revision of the manuscript and approved the final version.

**Supported by** Grants from the Snedkermester Sophus Jacobsen and Wife Astrid Jacobsen's Foundation, Else and Mogens Wedell-Wedellsborg Foundation, the Ihre Foundation, the Nanna Svartz Foundation as well as the Swedish Research Council; A Graduate Studentship Award from the Karolinska Faculty Foundation to Witte AB

**Correspondence to:** Dr. Anne-Barbara Witte, Department of Medicine Solna, Unit of Gastroenterology and Hepatology, Karolinska University Hospital Solna, Karolinska Institute, Solna, SE-171 76 Stockholm, Sweden. [anne-barbara.witte@ki.se](mailto:anne-barbara.witte@ki.se)  
Telephone: +46-81-2000990 Fax: +46-85-1775768

Received: December 5, 2012 Revised: February 16, 2013

Accepted: April 13, 2013

Published online: May 15, 2013

### Abstract

**AIM:** To investigate functional duodenal abnormalities in functional dyspepsia (FD) and the role of serotonin 5-hydroxytryptamine (5-HT) in mucosal ion transport and signalling.

**METHODS:** Duodenal mucosal biopsies were obtained from 15 patients with FD and 18 healthy controls. Immunohistochemistry was used to study the number of 5-HT-containing cells and real-time polymerase chain reaction for expression of 5-HT receptors 1A, 1B, 2A, 2B, 3A, 3B, 3C, 3D, 3E, 4 and 7, as well as expression of the serotonin re-uptake transporter (SERT) gene *SLC6A4* and tryptophan hydroxylase 1 (TPH1). Biopsies were mounted in Ussing chambers for evaluation of basal and 5-HT-stimulated short-circuit current (SCC).

**RESULTS:** Conductance was lower in FD [ $42.4 \pm 4.7$  mS/cm<sup>2</sup> ( $n = 15$ ) vs  $62.5 \pm 4.5$  mS/cm<sup>2</sup> ( $n = 18$ ),  $P = 0.005$ ]. 5-HT induced a dose dependent rise in SCC in both FD ( $n = 8$ ) and controls ( $n = 9$ ), the rise was lower in FD ( $P < 0.001$ ). Mean number of 5-HT stained cells per high power field was the same [ $34.4 \pm 8.4$  in FD ( $n = 15$ ) and  $30.4 \pm 3.7$  in controls ( $n = 18$ ),  $P = 0.647$ ]. The following genes were highly expressed: 5-HT receptor *HTR3E*, *HTR4*, *HTR7*, SERT gene (*SLC6A4*) and *TPH1*. Differences in expression levels were observed for *HTR3E* (higher expression in FD,  $P = 0.008$ ), *HTR7* (lower expression in FD,  $P = 0.027$ ), *SLC6A4* (higher expression in FD,  $P = 0.033$ ) and *TPH1* (lower expression in FD,  $P = 0.031$ ).

**CONCLUSION:** Duodenal ion transport in response to exogenous 5-HT is abnormal in FD patients and associated with high expression of the *HTR3E* receptor and the serotonin transporter.

© 2013 Baishideng. All rights reserved.

**Key words:** Dyspepsia; Ion transport; Gene expression; Duodenum; Epithelium; Serotonin

**Core tip:** The majority of patients with chronic symptoms from the gastro-duodenal region do not present signs of organic disease during routine examination, which commonly leads to the diagnosis of functional dyspepsia (FD). Our study strongly indicates the involvement of 5-hydroxytryptamine related duodenal mucosal mechanisms in FD pathogenesis. Future studies should further investigate alterations in up- and downstream effects related to HTR3E and HTR7 receptors.

Witte AB, D'Amato M, Poulsen SS, Laurent A, Knuhtsen S, Bindslev N, Hansen MB, Schmidt PT. Duodenal epithelial transport in functional dyspepsia: Role of serotonin. *World J Gastrointest Pathophysiol* 2013; 4(2): 28-36 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i2/28.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i2.28>

## INTRODUCTION

The majority of patients with chronic symptoms from the gastro-duodenal region do not present signs of organic disease during routine examination, which commonly leads to the diagnosis of functional dyspepsia (FD). FD is known for its high prevalence, association with decreased quality of life<sup>[1]</sup> and difficult management, which pose a considerable economic burden<sup>[2-4]</sup>. Highly debated pathogenetic mechanisms include alterations in stomach function, *e.g.*, hypersensitivity and emptying problems. Little is known about duodenal epithelial function in patients with FD and a broader understanding of the mucosal defence barrier, and insight in associated signalling systems is needed<sup>[5,6]</sup>.

It is widely recognized that therapies that modulate serotonin 5-hydroxytryptamine (5-HT) activity are partly effective in patients with functional intestinal disorders. There are indications of increased transcription of tryptophan hydroxylase 1 (TPH1) and lowered serotonin reuptake transporter (SERT) expression throughout the gut in isolated FD<sup>[7]</sup>. In addition, SERT polymorphisms appear to play a pathogenetic role in patients with postprandial distress syndrome<sup>[8]</sup>.

5-HT in humans is mainly found in the duodenum, with the highest concentration in the lamina mucosa<sup>[9,10]</sup>, and recent studies have revealed high expression levels of SERT, the 5-HT receptor HTR3 and HTR4 in the duodenum compared to other gut regions<sup>[11]</sup>. 5-HT is released by duodenal mucosal cells in response to luminal acidification<sup>[12]</sup> and together with other transmitters is involved in pathways mediating the response to luminal acid<sup>[13-15]</sup>. Furthermore, 5-HT has been shown to be a potent stimulator of electrogenic secretion in human duodenal mucosal biopsies<sup>[16]</sup>.

We hypothesized that disturbances in the duodenal

secretory reflex arches involving 5-HT play a pathogenic role in FD. Measurement of ion transport in human epithelia *in vivo* requires invasive methods; however a modified Ussing chamber for human endoscopic biopsies has been developed in our laboratory<sup>[17]</sup>.

In the present study, 5-HT induced ion transport is measured in FD patients and healthy controls. Furthermore, duodenal mucosal biopsies are evaluated for expression of 5-HT receptors real-time polymerase chain reaction (RT-PCR) and the density of 5-HT-containing cells (immunohistochemistry).

## MATERIALS AND METHODS

### Study population and protocol

The study was conducted in accordance with the Helsinki V declaration and approved by the Scientific Ethical Committee of Copenhagen. All participants received both written and oral information prior to the study and provided written informed consent.

Consecutive patients with dyspeptic symptoms referred for gastroscopy at Bispebjerg Hospital (Copenhagen, Denmark) who fulfilled the ROME III criteria<sup>[18]</sup> were invited to participate in the study. Healthy controls between 18-70 years were recruited. Two questionnaires, the gastrointestinal symptom rating scale and a non-validated Danish translation of a dyspepsia questionnaire developed by Tack *et al.*<sup>[19,20]</sup>, were used in order to confirm the diagnosis of FD, exclude gastroesophageal reflux disease as well as abdominal symptoms in the healthy controls. The gastrointestinal symptom rating scale is a validated instrument comprising 15 items for the assessment of gastrointestinal symptoms in irritable bowel syndrome (IBS) and peptic ulcer disease<sup>[21]</sup>.

Seventeen patients with FD and 20 healthy controls were included in the study. Median age was 39 years (range 24-54 years) in the FD group (7 men, 10 women) and 24 years (range 21-50 years) in the healthy group (5 men, 16 women). With the exception of two subjects who used oral contraception, healthy controls were medication free. Four of the FD patients reported use of proton pump inhibitors on a non-regular basis. None had used proton pump inhibitor treatment one week prior to endoscopy, nor *acetylsalicylic acid* or non-steroid anti-inflammatory drugs. Over-consumption of alcohol was not present in any subject. One of the FD patients and three of the healthy subjects reported being smokers.

During gastroscopy, biopsies were obtained from the duodenum at the border between the duodenal bulb and the descending duodenum using standard biopsy forceps (Radial Jaw 4, outside diameter 2.4 mm, Boston Scientific, Denmark). In two FD patients and one healthy control a major part of the biopsies could not be obtained because the procedure was too distressing, while esophageal pathology was found in another healthy control. This meant that only 15 FD patients and 18 healthy

controls were included, each with 8-10 biopsies available (out of 10 planned). Three of the biopsies were snap-frozen on dry ice for gene expression studies, one was stored in 4% buffered paraformaldehyde solution for subsequent immunohistochemical evaluation and up to four biopsies were placed in ice-cold Ringer solution for immediate mounting in Ussing chambers. Finally, one biopsy from the gastric antrum and one from the gastric corpus were stored in 4% buffered paraformaldehyde solution for subsequent histological analysis for *Helicobacter Pylori* detection.

### Mounting of biopsies and electrical measurements

Duodenal biopsies were transported to the laboratory in ice-cold bicarbonate-Ringer solution and 2-4 successfully mounted within 30 min in modified Ussing air suction chambers. Use of 10 times magnification through a stereomicroscope (Nikon, Tokyo) ensured correct mucosa-serosa orientation and appropriate fixation. Biopsies were fixed by constant air suction<sup>[17]</sup>. The exposed tissue area varied from 3.4 to 5 mm<sup>2</sup>, depending on the used insert, which was chosen to match the tissue size. The height of the (air) suction sleeve was 50 µm. Both sides of the tissue were bathed in bicarbonate-Ringer solution containing (in mmol/L) 140 Na<sup>+</sup>, 4 K<sup>+</sup>, 121 Cl<sup>-</sup>, 1 Ca<sup>2+</sup>, 0.5 Mg<sup>2+</sup>, 0.5 SO<sub>4</sub><sup>2-</sup> and 25 HCO<sub>3</sub><sup>-</sup>. In addition, 11 mmol/L *D*-glucose was applied to the serosal side and 11 mmol/L *D*-sorbitol to the mucosal side. Temperature was maintained at 37 °C with the help of water jackets and oxygenation was ensured by constant input of gas-lift-circulating 95% O<sub>2</sub> + 5% CO<sub>2</sub>. Short-circuit current (SCC) and slope conductance were continuously recorded by an automated voltage-clamp device (MFI 1-425) and measured as µA/cm<sup>2</sup> and mS/cm<sup>2</sup> respectively. Solution-resistance correction was performed immediately before mounting. Baseline values were recorded after an equilibration period of 15-30 min when stable values were reached. Further experiments were performed with different stimulating agents, but here we present results from the application of 5-HT, which was performed in nine FD patients and ten healthy controls as follows:

5-HT was applied in a dose-increasing manner ranging from 3 to 243 µmol/L (final concentration in the bathing solution) with 5-min intervals between applications. 5-HT was added to the serosal side as previously described<sup>[16]</sup>. For evaluation of tissue viability, transport capacity and correct tissue orientation, 4 mmol/L glucose and 0.1 mmol/L phloridzin, both purchased from Sigma, Denmark, were sequentially added to the mucosal side. A glucose response of > 5 µA/cm<sup>2</sup> or response to phloridzin < 5 µA/cm<sup>2</sup> was considered equivalent to full tissue viability. Some of the biopsies did not fulfil these criteria and were excluded from further analysis, resulting in a final number of eight FD patients and nine controls from whom 5-HT-stimulated values were obtained.

### Histopathology and immunohistochemistry

In addition to one un-mounted biopsy from each subject, all the mounted biopsies were gently collected upon conclusion of the Ussing chamber experiments and fixed in 4% buffered paraformaldehyde for a minimum of 24 h. They were then dehydrated, embedded in paraffin and cut into 10 µm sections, which were stained with hematoxylin/periodic acid Schiff for histological staining and examination. Protocols were blinded to the examiner. Assessment included possible pathology and the extent of epithelial damage, which was graded by a severity score ranging from 0 to 3; with 0 indicating no mucosal damage and 3 severe mucosal damages as previously described<sup>[16]</sup>. A Leitz Ortoplan microscope (Wetzlar, Germany) fitted with a cooled camera (Evolution MP, Media Cybernetics, Wokingham, Berkshire, United Kingdom) was used for examination and photography. For IHC analysis of anti-5-HT staining cells, sections of approximately 5 microns were deparaffinized and incubated for 5 min in 2% bovine serum albumin followed by 18 h at 4 °C with the primary antibody, monoclonal mouse anti-serotonin (M0758, DakoPatts) diluted 1:200. The immunoreactions were visualized by means of biotinylated rabbit anti-mouse immunoglobulins (E354, Dako) diluted 1:200, as the second layer, followed by streptavidin-peroxidase complex, VECTASTAIN PK-4000, diluted 1:100, for 30 min as the third layer. The sections were finally stained by means of 3,3'-diaminobenzidine for 30 min and counterstained with hematoxylin.

To determine the number, 5-HT-immunoreactive cells were counted in an objective manner from representative photomicrographs of the immunostained biopsies by means of Image-Pro Plus 6.0. Both the number of stained cells and the stained area per high power field were assessed. A field magnified 10 times was suitable to cover the biopsy area with only minor tissue deficiency, which correlates with a tissue area of 1.03 mm × 0.79 mm.

Gastric biopsies were immediately fixed in 4% buffered paraformaldehyde and after sectioning and staining with Giemsa (Merck, Darmstadt, Germany) examined for HP presence in the light microscope. Participants were considered HP positive if bacteria were found in either the antrum or the corpus fundus biopsies. These protocols were also blinded to the examiner.

### mRNA expression analysis

Samples for PCR were stored at -80 °C until tissue collection was completed for one-series analysis. Total RNA was extracted from biopsies with commercially available kits (Qiagen, Hilden, Germany) and cDNA synthesized from 0.5-1.0 µg of RNA with SuperScript™ III reverse transcriptase (Invitrogen, Carlsbad, CA, United States) in accordance with the manufacturer's instructions. In total, mRNA expression analysis was carried out on 10 FD patients and 16 controls.

For each gene examined, mRNA expression levels were measured by quantitative RT-PCR in ABI Prism 7500 Sequence Detection Systems (Applied Biosystems) with specific TaqMan Gene Expression Assays (Applied Biosystems), in accordance with the manufacturer's instructions. Gene expression of HTR1A (Assay ID Hs00265014), 1B (Hs00265286), 2A (Hs01033524), 2B (Hs00168362), 3A (Hs00168375), 3B (Hs00175775), 3C (Hs00365674), 3D (Hs00699391), 3E (Hs00704511), 4 (Hs00410577) and 7 (Hs00989028), the SERT gene (*SLC6A4*, Hs00984355) and TPH1 (Hs00188220) were studied. RT-PCR reactions were performed in triplicate on each sample and, after normalization to internal endogenous controls (glyceraldehyde 3-phosphate dehydrogenase, Hs99999905), mRNA expression levels for each gene and in each sample were determined by the comparative Ct method of relative quantification. Samples were only included in the analysis when gene expression was detected before cycle 35 in the PCR reaction (cycle threshold < 35) for a given gene, which resulted in slightly different *N*-values for each gene. Results were expressed in arbitrary units relative to a randomly chosen reference sample.

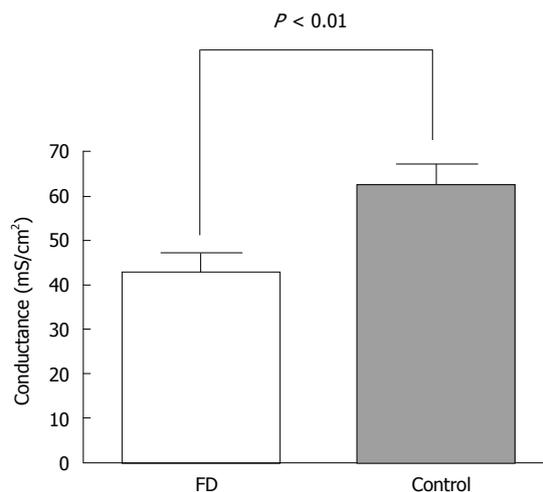
### Statistical analysis

The effect of the drugs applied in the Ussing chambers was defined as SCC before versus after application. Mean results from the 2-4 biopsies from each subject were used for further analysis and are presented as mean  $\pm$  SE. The unpaired *t*-test was used to compare the effects of glucose and the difference in basal SCC and slope conductance. The relationship between the concentration of 5-HT and induced SCC was investigated using a linear model, with treatment and log-transformed concentration as fixed effects. The number of 5-HT immune-reactive cells was compared between subject groups using the unpaired *t*-test. The Mann-Whitney *U*-test was employed to identify statistically significant differences between patients and controls in the RT-PCR gene expression analysis (mRNA). Commercially available software (PRISMA version 5.0 and SAS version 8.2) was used for all statistical analyses and two-tailed  $P < 0.05$  was considered significant.

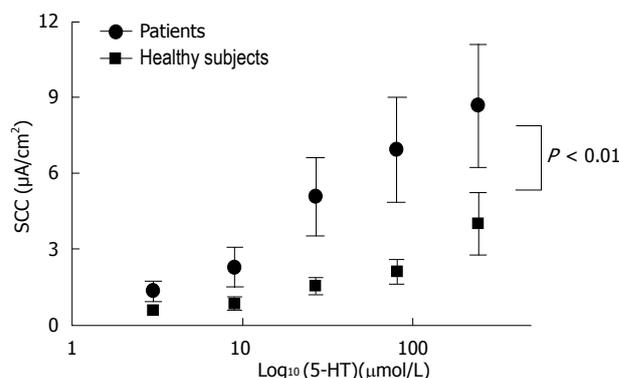
## RESULTS

### Electrophysiological measurements

Mean basal SCC was  $19.8 \pm 3.0 \mu\text{A}/\text{cm}^2$  for FD patients ( $n = 15$ ) and  $21.4 \pm 3.7 \mu\text{A}/\text{cm}^2$  for controls ( $n = 18$ ) with no significant difference between groups ( $P = 0.749$ ). As shown in Figure 1, comparison of basal conductance revealed significantly lower values for FD patients compared to healthy controls ( $42.4 \pm 4.7 \text{ mS}/\text{cm}^2$  and  $62.4 \pm 4.5 \text{ mS}/\text{cm}^2$  respectively,  $P = 0.005$ ). Glucose control values after 5-HT stimulation yielded a mean magnitude of  $12.5 \pm 2.0 \mu\text{A}/\text{cm}^2$  for the FD group and  $12.1$



**Figure 1 Basal slope conductance of duodenal mucosa as measured in a modified air-suction Ussing chamber.** Conductance, in millisiemens per square cm ( $\text{mS}/\text{cm}^2$ ), is significantly higher in healthy controls ( $n = 18$ ) compared to patients with functional dyspepsia (FD) ( $n = 15$ ),  $P = 0.005$  vs FD, mean  $\pm$  SE.

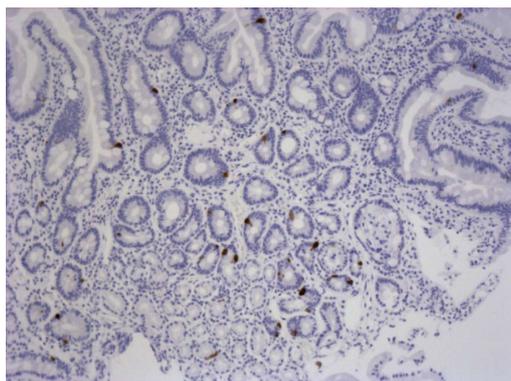


**Figure 2 Dose-response of 5-hydroxytryptamine-induced short circuit current.** Addition of 5-hydroxytryptamine (5-HT) in cumulative concentrations by steps of a factor three from 3 to 243  $\mu\text{mol}/\text{L}$  on the serosal side of duodenal biopsies mounted in an Ussing chamber resulted in an increased short circuit ( $\mu\text{A}/\text{cm}^2$ ), in both functional dyspepsia patients ( $n = 8$ ) and healthy controls ( $n = 9$ ), with significantly lower values in the dyspeptic group ( $P < 0.001$  for the overall difference) (mean  $\pm$  SE). SCC: Short circuit current.

$\pm 2.5 \mu\text{A}/\text{cm}^2$  for controls ( $P = 0.906$ ). 5-HT induced a dose dependent SCC rise in both healthy controls and FD patients (Figure 2). The 5-HT-induced rise in SCC was significantly lower in the latter ( $P < 0.001$ ).

### Histology

Histology revealed some variation with regard to biopsy depth; however, the surface epithelium and entire lamina propria were intact in all samples before and after mounting. Several biopsies also included the lamina muscularis mucosa and in some cases the submucosal layer contained part of Brunner's glands. Epithelial damage before mounting ranged from 0-3 in both groups with a mean score of 0.7 in dyspeptic and 1.1 in healthy subjects ( $P = 0.179$ ). 5-HT stained cells were found in the



**Figure 3** Paraformaldehyde-fixed duodenal tissue from a patient with functional dyspepsia. The tissue was stained immunohistochemically using a polyclonal antibody against 5-hydroxytryptamine (5-HT). There are many 5-HT immunoreactive cells in the surface epithelium, both in the crypts and the villi. The biopsy has an intact surface epithelium without signs of mechanical damage (magnification  $\times 220$ ).

epithelium and Brunner’s glands, but only the number of cells in the epithelium was included in the evaluation since not all biopsies contained sub-mucosal tissue (Figure 3). In un-mounted biopsies, the mean number of cells per high power field was  $34.4 \pm 8.4$  in the FD group and  $30.4 \pm 3.70$  in controls ( $P = 0.647$ ).

Helicobacter staining was positive in one FD patient and none of the healthy subjects.

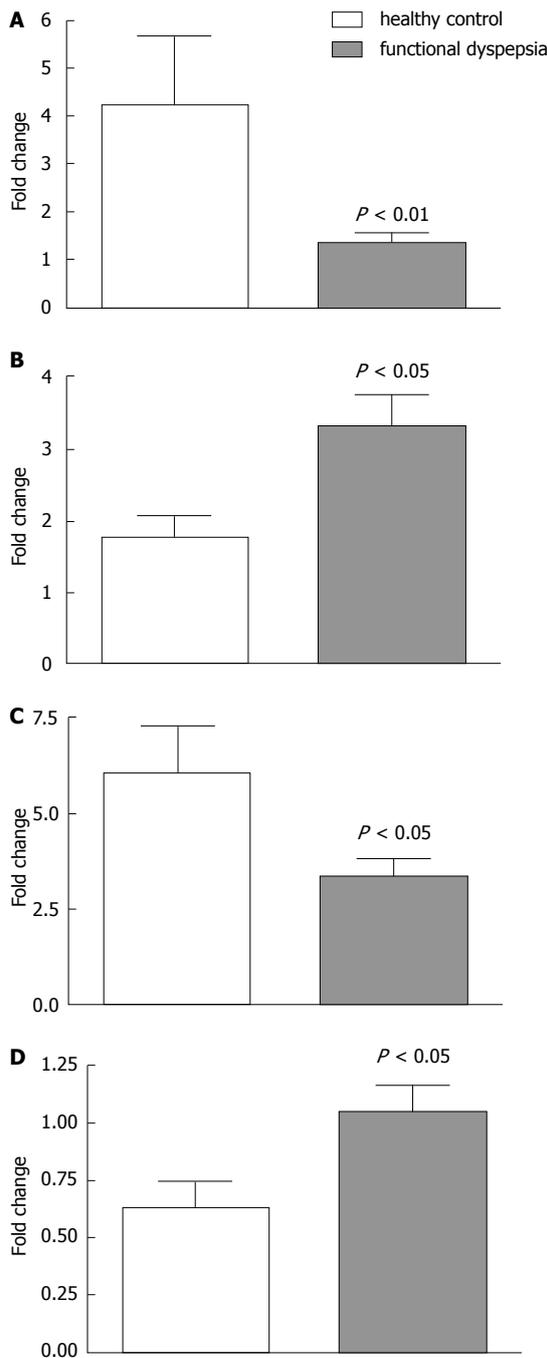
**Expression levels**

Expression levels were undetectable for three genes, *HTR1A*, *HTR3B* and *HTR3D*, while there was only very low expression of *HTR1B*, *HTR2A*, *HTR2B*, *HTR3A* and *HTR3C*. The following genes were strongly expressed: *HTR3E*, *HTR4*, *HTR7*, SERT gene (*SLC6A4*) and *TPH1*. Genes that exhibited statistically significant differences in expression level between FD patients ( $n = 8-10$ ) and healthy subjects ( $n = 13-16$ ) include *HTR3E* (higher expression in dyspepsia,  $P = 0.008$ ), *HTR7* (lower expression in dyspepsia,  $P = 0.027$ ), *SLC6A4* (higher expression in dyspepsia,  $P = 0.033$ ) and *TPH1* (lower expression in dyspepsia,  $P = 0.031$ ) (Figure 4).

**DISCUSSION**

5-HT increased the short circuit current in both healthy controls and FD patients in a dose-dependent manner, although the values were lower in FD patients (Figure 2). Slope conductance was significantly lower in FD patients.

Increased SCC is a direct result of secretion of negative ions to the duodenal lumen or flow of positive ions in the opposite direction and in the duodenum, bicarbonate secretion and hydrogen ion absorption are considered to be the most relevant fluxes. Disturbances in the response to duodenal acid have previously been described as related to dyspeptic symptoms<sup>[22,23]</sup>. Lower



**Figure 4** Gene expression (mRNA) levels differed significantly in duodenal mucosa from functional dyspepsia patients compared to healthy controls. A: *HTR3E*; B: *HTR7*; C: *SLC4A6*; D: *TPH1*. Empty bars represent patients ( $n = 10$  for *HTR3E*, *SLC4A6* and *TPH1*, and  $n = 8$  for *HTR7*) and filled bars represent controls ( $n = 16$  for *SLC4A6* and *TPH1*,  $n = 15$  for *HTR3E* and  $n = 13$  for *HTR7*). All levels were measured by quantitative real-time polymerase chain reaction and expressed as mean fold change  $\pm$  SE relative to the values obtained for a control sample arbitrarily chosen as a reference.  $P < 0.05$ ,  $P < 0.01$  vs control group.

SCC values in our FD group might result from dysfunction in bicarbonate transport and subsequent impaired secretion.

Generally, the SCC values obtained in this study were distinctly lower than previously described by Engelmann *et al*<sup>[16]</sup>, which could be explained by the fact that

the studies were performed in different settings and electrophysiological measurements are easily affected by environmental factors. One should also bear in mind that the study by Engelmann *et al.*<sup>[16]</sup> included patients with different dyspeptic conditions, which were not characterized by ROME III, and had no control group. In order to validate our results from the electrophysiological measurements, raw data were re-analysed by a second investigator blinded to the protocol and the analysis confirmed the results.

Reims *et al.*<sup>[24]</sup> reported the applicability of Ussing chamber-measured epithelial electrical resistance for quantifying permeability changes in duodenal biopsies. In our case, biopsies from patients with FD had significantly lower conductance (thus higher resistance) compared to healthy controls, which may indicate decreased permeability. This result is rather surprising, because duodenal mucosal disease is generally associated with mucosal damage and impaired barrier function, thus one would expect higher permeability in patients with FD compared to healthy controls. Furthermore, increased epithelial permeability has been demonstrated in IBS sub-groups<sup>[25-27]</sup>. Routine histology did not reveal morphological alterations in our biopsies and there was no evidence of increased edge leakage in the healthy controls. One could argue that the resistance measured in our experiments reflects disturbance in electrolyte transport rather than nutrient or large molecule passage. Thus, it would be more reasonable to assume lower conductance of bicarbonate or chloride ions because of channel dysfunction or, for example, lowered tight junction capacity. With the present measurements of SCC and slope conductance we are unable to discriminate between an altered ionic passage through the cell compartment and/or the paracellular pathway. Alteration in potassium channels both apically and basolaterally in epithelial cells may account for the observed changes in conductance as well as in other channels involved in the bicarbonate secretory process. A molecular reconfiguration in the paracellular pathway as an explanation for the observed difference in conductance is also a possibility.

Expression of HTR3E was considerably increased in FD patients, as was that of SLC6A4. However, HTR7 and TPH1 were significantly less expressed in patients with FD (Figure 4).

*SLC6A4* is the approved gene symbol for SERT. The transporter gene was highly expressed in the duodenal samples and there was a significant difference between experimental groups with higher expression in patients with FD. A relationship between *SERT* gene alterations and FD has been suggested in a number of reviews<sup>[8,28]</sup>. There are also several studies that have pointed out significant associations between *SERT* polymorphism and IBS<sup>[29-33]</sup>. To our knowledge, the only result that confirms the relevance of *SERT* polymorphisms in relation to dyspeptic symptoms comes from studies of postprandial distress syndrome<sup>[8]</sup>, while at least two other inves-

tigations failed to detect significant associations<sup>[34-37]</sup>. Gene expression levels have previously been described in dyspeptic female patients<sup>[7]</sup>, where decreased SERT expression was found in the patient group. Our results suggest the opposite, although the small sample size reduces reliability. As this issue remains unclear, further studies to detect a significant relation between alterations in SERT expression and FD are necessary. Both HTR3E and HTR4 have been reported to be prevalent in duodenal mucosa<sup>[11]</sup>, which is supported by our study. Furthermore, we found elevated expression of the HTR3E sub-type in patients with FD. *HTR3E* polymorphism possibly related to increased HTR3E production has been described in IBS<sup>[38]</sup> and our results might thus account for involvement of this receptor in FD pathology. In IBS, HTR3 antagonists are already in clinical use, and the cation-selective ligand-gated ion channels have been found to be involved in motility and bicarbonate secretion. Interestingly, expressions of other HTR3 sub-types, *i.e.*, A and B, were low in our samples. A strong genetic correlation between HTR3C and HTR3E in human duodenal tissue has recently been proposed<sup>[35]</sup> and patch clamp studies have revealed that HTR3C, HTR3D and HTR3E are non-functional when expressed alone<sup>[39]</sup>. *HTR3A* gene polymorphism has recently been associated with severe dyspeptic symptoms<sup>[40]</sup>. As HTR3A expression was generally low in our biopsies, differences between FD patients and controls might not have become apparent.

HTR7 is one of the 5-HT receptors about which very little is known, mainly due to the lack of specific ligands. A role in upper intestinal smooth muscle relaxation<sup>[41,42]</sup>, activation of afferent neurons<sup>[43]</sup> as well as high expression in the stomach and ileum<sup>[44]</sup> has been revealed previously, and we found high expression in duodenal samples. Our results also indicate significant differences in expression level between patients with FD and controls. HTR7 has been proposed as a novel target for treatment of FD and related disorders<sup>[45]</sup> and further elucidation of its pathophysiological role would definitely be valuable.

*TPH1* gene abnormalities have recently been suggested in IBS<sup>[46]</sup> and lower mRNA expression levels have been observed in such patients<sup>[30]</sup>. We found lower expression values in our FD group, indicating that decreased 5-HT availability might be part of the disease pathology. In contrast, Foxx-Orenstein *et al.*<sup>[7]</sup> presented results demonstrating higher expression in duodenal mucosa of female patients with FD. A recent study of TPH1 expression in gastric mucosal biopsies from children with FD found no difference between these patients and controls<sup>[47]</sup>.

As we did not observe any significant differences in the content of 5-HT-positive cells between FD patients and controls, further studies including a larger number of subjects may be needed to conclusively address this issue. 5-HT content in duodenal mucosa has been studied immunohistochemically in patients with post-

infectious FD and found to be significantly reduced in patients 6 mo or more after the infection, but not in recovered controls<sup>[48]</sup>.

The low number of subjects in the patient and control groups and differences in terms of age and smoking status (which can affect the mucous layer and bicarbonate secretion) are a limitation of this study.

In conclusion, our study strongly indicates the involvement of 5-HT related duodenal mucosal mechanisms in FD pathogenesis. Future studies should further investigate alterations in up- and downstream effects related to HTR3E and HTR7 receptors.

## ACKNOWLEDGMENTS

We thank Heidi Paulsen, Lise Strange and Anne-Marie Møller for technical assistance as well as Sofia Zetterstrand for help with the statistical analysis.

## COMMENTS

### Background

The majority of patients with chronic symptoms from the gastro-duodenal region do not present signs of organic disease during routine examination, which commonly leads to the diagnosis of functional dyspepsia (FD).

### Research frontiers

It is widely recognized that therapies that modulate serotonin 5-hydroxytryptamine (5-HT) activity are partly effective in patients with functional intestinal disorders.

### Innovations and breakthroughs

Duodenal ion transport in response to exogenous 5-HT is abnormal in FD patients and associated with high expression of the HTR3E receptor and the serotonin transporter. Their study strongly indicates the involvement of 5-HT related duodenal mucosal mechanisms in FD pathogenesis. Future studies should further investigate alterations in up- and downstream effects related to HTR3E and HTR7 receptors.

### Peer review

The authors present a study about an involvement of 5-HT in FD. Due to the prevalence of FD, it is important to reveal the mechanisms involved as a basis for the development of new therapies.

## REFERENCES

- 1 **Aro P**, Talley NJ, Agréus L, Johansson SE, Bolling-Sternevald E, Storskrubb T, Ronkainen J. Functional dyspepsia impairs quality of life in the adult population. *Aliment Pharmacol Ther* 2011; **33**: 1215-1224 [PMID: 21443537 DOI: 10.1111/j.1365-2036.2011.04640.x]
- 2 **Alander T**, Svärdsudd K, Agréus L. Functional gastrointestinal disorder is associated with increased non-gastrointestinal healthcare consumption in the general population. *Int J Clin Pract* 2008; **62**: 234-240 [PMID: 18021207 DOI: 10.1111/j.1742-1241.2007.01549.x]
- 3 **Ford AC**, Moayyedi P. Managing dyspepsia. *Curr Gastroenterol Rep* 2009; **11**: 288-294 [PMID: 19615304 DOI: 10.1007/s11894-009-0042-2]
- 4 **Talley NJ**. Functional gastrointestinal disorders as a public health problem. *Neurogastroenterol Motil* 2008; **20** Suppl 1: 121-129 [PMID: 18402649 DOI: 10.1111/j.1365-2982.2008.01097.x]
- 5 **van Boxel OS**, ter Linde JJ, Siersema PD, Smout AJ. Role of chemical stimulation of the duodenum in dyspeptic symptom generation. *Am J Gastroenterol* 2010; **105**: 803-811; quiz 802, 812 [PMID: 20234343 DOI: 10.1038/ajg.2010.100]

- 6 **Lee KJ**, Tack J. Duodenal implications in the pathophysiology of functional dyspepsia. *J Neurogastroenterol Motil* 2010; **16**: 251-257 [PMID: 20680163 DOI: 10.5056/jnm.2010.16.3.251]
- 7 **Foxx-Orenstein A**, Camilleri M, Gershon MD, Linden DR, Mawe GM, Lewis JT, Jensen KL, Talley NJ, Szurszewski JH, Zinsmeister A. Alterations in intestinal serotonin expression in dyspepsia and irritable bowel syndrome. *Gastroenterology* 2007; **132**: A72
- 8 **Oshima T**, Toyoshima F, Nakajima S, Fukui H, Watari J, Miwa H. Genetic factors for functional dyspepsia. *J Gastroenterol Hepatol* 2011; **26** Suppl 3: 83-87 [PMID: 21443717 DOI: 10.1111/j.1440-1746.2011.06639.x]
- 9 **Meyer T**, Brinck U. Differential distribution of serotonin and tryptophan hydroxylase in the human gastrointestinal tract. *Digestion* 1999; **60**: 63-68 [PMID: 9892800 DOI: 10.1159/00007590]
- 10 **Sjölund K**, Sandén G, Håkanson R, Sundler F. Endocrine cells in human intestine: an immunocytochemical study. *Gastroenterology* 1983; **85**: 1120-1130 [PMID: 6194039]
- 11 **van Lelyveld N**, Ter Linde J, Schipper ME, Samsom M. Regional differences in expression of TPH-1, SERT, 5-HT(3) and 5-HT(4) receptors in the human stomach and duodenum. *Neurogastroenterol Motil* 2007; **19**: 342-348 [PMID: 17509016 DOI: 10.1111/j.1365-2982.2006.00891.x]
- 12 **Kellum JM**, Jaffe BM. Release of immunoreactive serotonin following acid perfusion of the duodenum. *Ann Surg* 1976; **184**: 633-636 [PMID: 10857 DOI: 10.1097/0000658-197611000-00018]
- 13 **Allen A**, Flemström G. Gastroduodenal mucus bicarbonate barrier: protection against acid and pepsin. *Am J Physiol Cell Physiol* 2005; **288**: C1-C19 [PMID: 15591243]
- 14 **Nylander O**, Pihl L. Luminal hypotonicity increases duodenal mucosal permeability by a mechanism involving 5-hydroxytryptamine. *Acta Physiol (Oxf)* 2006; **186**: 45-58 [PMID: 16497179 DOI: 10.1111/j.1748-1716.2005.01507.x]
- 15 **Smith AJ**, Chappell AE, Buret AG, Barrett KE, Dong H. 5-Hydroxytryptamine contributes significantly to a reflex pathway by which the duodenal mucosa protects itself from gastric acid injury. *FASEB J* 2006; **20**: 2486-2495 [PMID: 17142798 DOI: 10.1096/fj.06-6391com]
- 16 **Engelmann BE**, Bindslev N, Poulsen SS, Larsen R, Hansen MB. Functional characterization of serotonin receptor subtypes in human duodenal secretion. *Basic Clin Pharmacol Toxicol* 2006; **98**: 142-149 [PMID: 16445586 DOI: 10.1111/j.1742-7843.2006.pto\_262.x]
- 17 **Larsen R**, Mertz-Nielsen A, Hansen MB, Poulsen SS, Bindslev N. Novel modified Ussing chamber for the study of absorption and secretion in human endoscopic biopsies. *Acta Physiol Scand* 2001; **173**: 213-222 [PMID: 11683679 DOI: 10.1046/j.1365-201X.2001.00865.x]
- 18 **Tack J**, Talley NJ, Camilleri M, Holtmann G, Hu P, Malagelada JR, Stanghellini V. Functional gastroduodenal disorders. *Gastroenterology* 2006; **130**: 1466-1479 [PMID: 16678560 DOI: 10.1053/j.gastro.2005.11.059]
- 19 **Tack J**, Piessevaux H, Coulie B, Caenepeel P, Janssens J. Role of impaired gastric accommodation to a meal in functional dyspepsia. *Gastroenterology* 1998; **115**: 1346-1352 [PMID: 9834261 DOI: 10.1016/S0016-5085(98)70012-5]
- 20 **Tack J**, Caenepeel P, Fischler B, Piessevaux H, Janssens J. Symptoms associated with hypersensitivity to gastric distention in functional dyspepsia. *Gastroenterology* 2001; **121**: 526-535 [PMID: 11522735 DOI: 10.1053/gast.2001.27180]
- 21 **Svedlund J**, Sjödin I, Dotevall G. GSRs--a clinical rating scale for gastrointestinal symptoms in patients with irritable bowel syndrome and peptic ulcer disease. *Dig Dis Sci* 1988; **33**: 129-134 [PMID: 3123181 DOI: 10.1007/BF01535722]
- 22 **Samsom M**, Verhagen MA, vanBerge Henegouwen GP,

- Smout AJ. Abnormal clearance of exogenous acid and increased acid sensitivity of the proximal duodenum in dyspeptic patients. *Gastroenterology* 1999; **116**: 515-520 [PMID: 10029608 DOI: 10.1016/S0016-5085(99)70171-X]
- 23 **Lee KJ**, Demarchi B, Demedts I, Sifrim D, Raeymaekers P, Tack J. A pilot study on duodenal acid exposure and its relationship to symptoms in functional dyspepsia with prominent nausea. *Am J Gastroenterol* 2004; **99**: 1765-1773 [PMID: 15330916 DOI: 10.1111/j.1572-0241.2004.30822.x]
- 24 **Reims A**, Strandvik B, Sjövall H. Epithelial electrical resistance as a measure of permeability changes in pediatric duodenal biopsies. *J Pediatr Gastroenterol Nutr* 2006; **43**: 619-623 [PMID: 17130738 DOI: 10.1097/01.mpg.0000232573.33526.f5]
- 25 **Dunlop SP**, Hebden J, Campbell E, Naesdal J, Olbe L, Perkins AC, Spiller RC. Abnormal intestinal permeability in subgroups of diarrhea-predominant irritable bowel syndromes. *Am J Gastroenterol* 2006; **101**: 1288-1294 [PMID: 16771951 DOI: 10.1111/j.1572-0241.2006.00672.x]
- 26 **Marshall JK**, Thabane M, Garg AX, Clark W, Meddings J, Collins SM. Intestinal permeability in patients with irritable bowel syndrome after a waterborne outbreak of acute gastroenteritis in Walkerton, Ontario. *Aliment Pharmacol Ther* 2004; **20**: 1317-1322 [PMID: 15606393 DOI: 10.1111/j.1365-2036.2004.02284.x]
- 27 **Piche T**, Barbara G, Aubert P, Bruley des Varannes S, Dainese R, Nano JL, Cremon C, Stanghellini V, De Giorgio R, Galmiche JP, Neunlist M. Impaired intestinal barrier integrity in the colon of patients with irritable bowel syndrome: involvement of soluble mediators. *Gut* 2009; **58**: 196-201 [PMID: 18824556 DOI: 10.1136/gut.2007.140806]
- 28 **Camilleri M**. Pharmacogenomics and serotonergic agents: research observations and potential clinical practice implications. *Neurogastroenterol Motil* 2007; **19** Suppl 2: 40-45 [PMID: 17620087 DOI: 10.1111/j.1365-2982.2007.00961.x]
- 29 **Pata C**, Erdal ME, Derici E, Yazar A, Kanik A, Ulu O. Serotonin transporter gene polymorphism in irritable bowel syndrome. *Am J Gastroenterol* 2002; **97**: 1780-1784 [PMID: 12135035]
- 30 **Coates MD**, Mahoney CR, Linden DR, Sampson JE, Chen J, Blaszyk H, Crowley MD, Sharkey KA, Gershon MD, Mawe GM, Moses PL. Molecular defects in mucosal serotonin content and decreased serotonin reuptake transporter in ulcerative colitis and irritable bowel syndrome. *Gastroenterology* 2004; **126**: 1657-1664 [PMID: 15188158 DOI: 10.1053/j.gastro.2004.03.013]
- 31 **Yeo A**, Boyd P, Lumsden S, Saunders T, Handley A, Stubbins M, Knaggs A, Asquith S, Taylor I, Bahari B, Crocker N, Rallan R, Varsani S, Montgomery D, Alpers DH, Dukes GE, Purvis I, Hicks GA. Association between a functional polymorphism in the serotonin transporter gene and diarrhoea predominant irritable bowel syndrome in women. *Gut* 2004; **53**: 1452-1458 [PMID: 15361494 DOI: 10.1136/gut.2003.035451]
- 32 **Park JM**, Choi MG, Park JA, Oh JH, Cho YK, Lee IS, Kim SW, Choi KY, Chung IS. Serotonin transporter gene polymorphism and irritable bowel syndrome. *Neurogastroenterol Motil* 2006; **18**: 995-1000 [PMID: 17040410 DOI: 10.1111/j.1365-2982.2006.00829.x]
- 33 **Sikander A**, Rana SV, Sinha SK, Prasad KK, Arora SK, Sharma SK, Singh K. Serotonin transporter promoter variant: Analysis in Indian IBS patients and control population. *J Clin Gastroenterol* 2009; **43**: 957-961 [PMID: 19687750 DOI: 10.1097/MCG.0b013e3181b37e8c]
- 34 **Camilleri CE**, Carlson PJ, Camilleri M, Castillo EJ, Locke GR, Geno DM, Stephens DA, Zinsmeister AR, Urrutia R. A study of candidate genotypes associated with dyspepsia in a U.S. community. *Am J Gastroenterol* 2006; **101**: 581-592 [PMID: 16464220 DOI: 10.1111/j.1572-0241.2006.00481.x]
- 35 **van Lelyveld N**, Linde JT, Schipper M, Samsom M. Candidate genotypes associated with functional dyspepsia. *Neurogastroenterol Motil* 2008; **20**: 767-773 [PMID: 18331431 DOI: 10.1111/j.1365-2982.2008.01102.x]
- 36 **Toyoshima F**, Oshima T, Nakajima S, Sakurai J, Tanaka J, Tomita T, Hori K, Matsumoto T, Miwa H. Serotonin transporter gene polymorphism may be associated with functional dyspepsia in a Japanese population. *BMC Med Genet* 2011; **12**: 88 [PMID: 21714874 DOI: 10.1186/1471-2350-12-88]
- 37 **Park CS**, Uhm JH. Polymorphisms of the Serotonin Transporter Gene and G-Protein  $\beta 3$  Subunit Gene in Korean Children with Irritable Bowel Syndrome and Functional Dyspepsia. *Gut Liver* 2012; **6**: 223-228 [PMID: 22570752 DOI: 10.5009/gnl.2012.6.2.223]
- 38 **Kapeller J**, Houghton LA, Mönnikes H, Walstab J, Möller D, Bönisch H, Burwinkel B, Autschbach F, Funke B, Lasitschka F, Gassler N, Fischer C, Whorwell PJ, Atkinson W, Fell C, Büchner KJ, Schmidtman M, van der Voort I, Wisser AS, Berg T, Rappold G, Niesler B. First evidence for an association of a functional variant in the microRNA-510 target site of the serotonin receptor-type 3E gene with diarrhea predominant irritable bowel syndrome. *Hum Mol Genet* 2008; **17**: 2967-2977 [PMID: 18614545 DOI: 10.1093/hmg/ddn195]
- 39 **Holbrook JD**, Gill CH, Zebda N, Spencer JP, Leyland R, Rance KH, Trinh H, Balmer G, Kelly FM, Yusaf SP, Courtenay N, Luck J, Rhodes A, Modha S, Moore SE, Sanger GJ, Gunthorpe MJ. Characterisation of 5-HT<sub>3C</sub>, 5-HT<sub>3D</sub> and 5-HT<sub>3E</sub> receptor subunits: evolution, distribution and function. *J Neurochem* 2009; **108**: 384-396 [PMID: 19012743 DOI: 10.1111/j.1471-4159.2008.05775.x]
- 40 **Mujakovic S**, ter Linde JJ, de Wit NJ, van Marrewijk CJ, Franssen GA, Onland-Moret NC, Laheij RJ, Muris JW, Grobbee DE, Samsom M, Jansen JB, Knottnerus A, Numans ME. Serotonin receptor 3A polymorphism c.-42C > G; T is associated with severe dyspepsia. *BMC Med Genet* 2011; **12**: 140 [PMID: 22014438 DOI: 10.1186/1471-2350-12-140]
- 41 **Janssen P**, Prins NH, Moreaux B, Meulemans AL, Lefebvre RA. Characterization of 5-HT<sub>7</sub>-receptor-mediated gastric relaxation in conscious dogs. *Am J Physiol Gastrointest Liver Physiol* 2005; **289**: G108-G115 [PMID: 15746214 DOI: 10.1152/ajpgi.00012.2005]
- 42 **Tonini M**, Vicini R, Cervio E, De Ponti F, De Giorgio R, Barbara G, Stanghellini V, Dellabianca A, Sternini C. 5-HT<sub>7</sub> receptors modulate peristalsis and accommodation in the guinea pig ileum. *Gastroenterology* 2005; **129**: 1557-1566 [PMID: 16285955 DOI: 10.1053/j.gastro.2005.08.005]
- 43 **Meuser T**, Pietruck C, Gabriel A, Xie GX, Lim KJ, Pierce Palmer P. 5-HT<sub>7</sub> receptors are involved in mediating 5-HT-induced activation of rat primary afferent neurons. *Life Sci* 2002; **71**: 2279-2289 [PMID: 12215375 DOI: 10.1016/S0024-3205(02)02011-8]
- 44 **Bard JA**, Zgombick J, Adham N, Vaysse P, Branchek TA, Weinshank RL. Cloning of a novel human serotonin receptor (5-HT<sub>7</sub>) positively linked to adenylate cyclase. *J Biol Chem* 1993; **268**: 23422-23426 [PMID: 8226867]
- 45 **Tonini M**. 5-Hydroxytryptamine effects in the gut: the 3, 4, and 7 receptors. *Neurogastroenterol Motil* 2005; **17**: 637-642 [PMID: 16185301 DOI: 10.1111/j.1365-2982.2005.00716.x]
- 46 **Jun S**, Kohen R, Cain KC, Jarrett ME, Heitkemper MM. Associations of tryptophan hydroxylase gene polymorphisms with irritable bowel syndrome. *Neurogastroenterol Motil* 2011; **23**: 233-239, e116 [PMID: 21073637 DOI: 10.1111/j.1365-2982.2010.01623.x]
- 47 **Faure C**, Patey N, Gauthier C, Brooks EM, Mawe GM. Serotonin signaling is altered in irritable bowel syndrome with

diarrhea but not in functional dyspepsia in pediatric age patients. *Gastroenterology* 2010; **139**: 249-258 [PMID: 20303355  
DOI: 10.1053/j.gastro.2010.03.032]

48 **Dizdar V**, Spiller R, Singh G, Hanevik K, Gilja OH, El-Salhy

M, Hausken T. Relative importance of abnormalities of CCK and 5-HT (serotonin) in Giardia-induced post-infectious irritable bowel syndrome and functional dyspepsia. *Aliment Pharmacol Ther* 2010; **31**: 883-891 [PMID: 20132151]

**P- Reviewer** Pehl C **S- Editor** Gou SX **L- Editor** A  
**E- Editor** Yan JL



## Impact of body mass index for patients undergoing pancreaticoduodenectomy

Marco Del Chiaro, Elena Rangelova, Christoph Ansoerge, John Blomberg, Ralf Segersvärd

Marco Del Chiaro, Elena Rangelova, Christoph Ansoerge, John Blomberg, Ralf Segersvärd, Division of Surgery, Department of Clinical Science, Intervention and Technology, Karolinska Institute at Karolinska University Hospital, 14186 Stockholm, Sweden

Author contributions: Del Chiaro M designed the study, analyzed the results and operates the patients; Rangelova E, Ansoerge C and Blomberg J operated the patients; Segersvärd R contributed to writing the paper and operating the patients.

Supported by Division of Surgery, Department of Clinical Science, Intervention and Technology, Karolinska Institute at Karolinska University Hospital, Stockholm, Sweden

Correspondence to: Marco Del Chiaro, MD, PhD, Division of Surgery, Department of Clinical Science, Intervention and Technology, Karolinska Institute at Karolinska University Hospital, Huddinge K53, 14186 Stockholm, Sweden. marco.del-chiaro@karolinska.se

Telephone: +46-8-58582340 Fax: +46-8-58582340

Received: January 10, 2013 Revised: April 9, 2013

Accepted: April 13, 2013

Published online: May 15, 2013

### Abstract

**AIM:** To evaluate the impact of body mass index (BMI) on short and long term results after pancreaticoduodenectomies (PD).

**METHODS:** A consecutive series of PDs performed at the Karolinska University Hospital from 2004 till 2010 were retrieved from our prospective database. The patients were divided by BMI into overweight/obese (O; BMI  $\geq$  25 kg/m<sup>2</sup>) and controls (C; BMI < 25 kg/m<sup>2</sup>). Demographics, peri-operative data, morbidity, mortality, pancreatic fistula (PF) rate, length of stay (LOS), hospital costs, histology, and survival were analyzed. An additional sub analysis of survival was performed in patients with a diagnosis of pancreatic ductal adenocarcinoma (PDAC) and divided in underweight, normal-weight, overweight and obese.

**RESULTS:** A total of 367 PDs were included (O = 141/ C = 226). No differences were found between O and C regarding demographics, peri-operative data, costs, morbidity or mortality. O was associated with higher intra-operative blood loss (1392  $\pm$  115 mL vs 1121  $\pm$  83 mL;  $P = 0.01$ ), rate of PF (20% vs 9.5%;  $P = 0.006$ ) and marginally longer LOS (18  $\pm$  0.9 d vs 15  $\pm$  1.1 d;  $P = 0.05$ ). An increasing risk for PF was observed with increasing BMI. The 1, 3 and 5 years survival rate was similar in O and C in PDAC (68.7%, 26.4% and 8.8% vs 66.1%, 30.9% and 17.9% respectively;  $P = 0.9$ ). When the survival was analyzed using 4 different categories of BMI (underweight, normal, overweight and obese), a trend was seen toward a difference in survival, with a worse prognosis for the underweight and obese patients compared to normal weight and overweight patients.

**CONCLUSION:** Overweight increases the risk for intra-operative bleeding and PF, but do not otherwise alter short or long term outcome after PD for pancreatic cancer.

© 2013 Baishideng. All rights reserved.

**Key words:** Pancreas surgery; Pancreatectomy; Body mass index; Pancreatico- duodenectomy; Oncology; Pancreas cancer

**Core tip:** In the last decades, the number of overweight individuals has increased dramatically in Western countries. No data are available in the literature that show clearly whether this comorbidity has an impact on short-term or long-term outcomes in these patients or on procedure-related costs. Some studies have shown that pancreatectomies in overweight patients are associated with an increased risk of post-operative complications. The data are even more confusing regarding long-term and oncologic outcomes. In our study, based on a large series of consecutive

pancreaticoduodenectomies (PD) performed in a high volume center for pancreatic surgery, we showed that body mass index (BMI) is a risk factor for intra-operative bleeding and post-operative pancreatic fistula, but does not increase the overall morbidity and have no impact on survival of patients with pancreatic ductal adenocarcinoma. Based on these results, BMI should not be considered, per-se, an exclusion criteria for candidates for PD.

Del Chiaro M, Rangelova E, Ansoorge C, Blomberg J, Segersvärd R. Impact of body mass index for patients undergoing pancreaticoduodenectomy. *World J Gastrointest Pathophysiol* 2013; 4(2): 37-42 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i2/37.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i2.37>

## INTRODUCTION

In the last decades, the number of overweight individuals has increased dramatically in Western countries. It is estimated that today there are about 1.6 billion individuals in the world who are overweight and 400 million who are obese<sup>[1]</sup>. At the same time, obesity is a well-recognized risk factor for several chronic degenerative diseases such as cardiovascular diseases<sup>[2,3]</sup>, diabetes<sup>[4]</sup> and a number of types of cancer<sup>[5]</sup>, including pancreatic cancer<sup>[6]</sup>. In fact, obese individuals have a twofold to threefold increase in the risk of death from all causes compared to the general population<sup>[7]</sup>. It is estimated that treatment of obesity-related diseases consumes about 10% of healthcare expenditures in the United States<sup>[8]</sup>.

These data clearly suggest that the number of overweight or obese patients who are candidates for pancreaticoduodenectomy will increase in the future, but no data are available in the literature that show clearly whether this comorbidity has an impact on short-term or long-term outcomes in these patients or on procedure-related costs.

Some studies have shown that pancreatectomies in overweight patients are associated with an increased risk of post-operative complications<sup>[9]</sup> including a longer hospital stay<sup>[9-11]</sup>, greater blood loss<sup>[12]</sup>, more wound infections<sup>[13-15]</sup>, and an increased rate of pancreatic fistula (PF)<sup>[12]</sup>. However, some other studies have not shown an overall increase of post-operative morbidity in overweight patients<sup>[16,17]</sup>. Analysis is even more difficult for long-term and oncologic outcomes. In these areas, few data are available, and the results are not consistent. In a recent paper, Fleming showed shorter long-term survival and an increase in the number of positive node specimens in patients with a body mass index (BMI) > 35 kg/m<sup>2</sup> who underwent pancreaticoduodenectomy for pancreatic ductal adenocarcinoma (PDAC)<sup>[18]</sup>. In contrast, a large single-institution study by Tsai *et al*<sup>[12]</sup> found a longer survival time in overweight patients who underwent

pancreaticoduodenectomy for PDAC. Due to the worldwide increase in numbers of overweight patients who are candidates for pancreaticoduodenectomy and the lack of data concerning short-term and long-term outcomes for that procedure in these patients, we decided to analyze our own results and investigate the correlation between BMI and survival, post-operative complications, and the cost of pancreaticoduodenectomy in overweight patients.

## MATERIALS AND METHODS

Patients who had undergone pancreatoduodenectomy at Karolinska University Hospital 2004–2010 were retrieved from the hospital database containing prospectively collected pre-, intra- and post-operative data. The cohort was divided by BMI into overweight/obese (BMI ≥ 25 kg/m<sup>2</sup>) and controls (BMI < 25 kg/m<sup>2</sup>). Demographics, peri-operative data, morbidity, in-hospital mortality, PF rate, length of stay (LOS), histology, and survival were analyzed. Financial files for the patients in the study were obtained from the Economic Department at the Karolinska University Hospital and analyzed. The cost analysis included all the in-hospital diagnostic and therapeutic procedures, such as X rays, blood samples, other tests (*e.g.*, electrocardiogram and spirometry), operations, and drugs, as well as pre- and post-operative stays, and intensive care unit (ICU) or sub-ICU stays. All patients underwent a conventional Whipple resection with vascular resections of large retropancreatic vessels whenever necessary and with radical lymphadenectomy, as described in the Castelfranco Veneto Classification<sup>[19]</sup>. In all patients, the reconstruction was done by an end-to-side, duct to mucosa, pancreaticojejunostomy, an end-to-side conventional hepatico-jejunostomy and an antecolic gastro-entero anastomosis. The study was approved by the local ethics committee. GraphPad Prism software (R) was used to compare costs, pathological results, intra-operative and post-operative outcomes, and long-term survival between the two groups. The Student's *t* test was used for comparison of the means of continuous variables. Fisher's exact test was used to compare categorical variables. Long-term survival was analyzed using a non-parametric method (Kaplan Meier). Differences in survival were estimated using the Log-rank test.

### Statistical analysis

Statistical methods should be described when they are used to verify the results. Choose suitable techniques for the statistical treatments; for example, *t*-test (group or paired comparisons),  $\chi^2$  test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance (ANOVA), analysis of covariance, *etc.*

## RESULTS

Three hundred sixty-seven consecutive patients were identified of which 141 (38.4%) were overweight/obese. As shown in Table 1, the overweight/obese group

**Table 1** Pre-operative characteristics of patients

Patient characteristics	Overweight (n = 141)	Controls (n = 226)	P value
BMI (kg/m <sup>2</sup> )	28.6	22.1	< 0.0001
Male/Female	75/66	127/99	0.5
Mean age (yr)	63.7	65	0.3
ASA risk			
I	10.40%	5.10%	0.06
II	58.30%	53.40%	0.40
III	29.60%	36.40%	0.20
IV	1.70%	5.10%	0.30
Vascular resection	15.60%	22.10%	0.10

ASA: American Society of Anesthesiology; BMI: Body mass index.

had significantly higher mean BMI than the controls (28.6 kg/m<sup>2</sup> *vs* 22.1 kg/m<sup>2</sup>;  $P < 0.0001$ ). No significant differences were found in sex distribution, mean age, pre-operative risk assessment according to the American Society of Anesthesiology classification system, or the number of procedures associated with resection of peri-pancreatic vessels.

### Operative and pathology details

The overweight/obese group had higher intra-operative blood loss (1392 ± 115 mL *vs* 1121 ± 83 mL;  $P = 0.01$ ) and a marginally longer LOS (18.0 ± 0.9 d *vs* 15.0 ± 1.1 d;  $P = 0.05$ ) compared to the control group. No differences were found between the groups in operation time (446 ± 8 min *vs* 431 ± 7 min;  $P = 0.2$ ). Both groups showed a similar panorama of final histological diagnoses even if more patients were treated for chronic pancreatitis in the controls group and for other tumor types in the overweight group (Table 2); about half of the patients were treated for PDAC.

### Peri-operative mortality and morbidity

Overall, 13 of the 367 patients died post-operatively (3.5% in-hospital mortality). No difference in mortality rate was found between the two groups: 5 patients died in the overweight group (3.4%) and 8 in the control group (3.5%). No differences in overall morbidity (47% *vs* 54%;  $P = 0.2$ ) or severe post-operative complications (grade ≥ 3b according to the Clavien classification)<sup>[20]</sup> (15.6% *vs* 17.2%;  $P = 0.7$ ) were found between the overweight and control groups. In contrast the overweight patients developed more post-operative medical specific complications (24.1% *vs* 15%;  $P = 0.03$ ). When specific surgical complications were compared, only PF occurred more frequently in the overweight group compared to the control group (20% *vs* 9.5%;  $P = 0.006$ ). Otherwise, no significant differences were found in the incidence of delayed gastric emptying, bile leakage, wound infection, or intra-abdominal or gastro-intestinal bleeding (Table 2). When the patients were divided into 4 different sub-groups according to BMI, the risk for developing post-operative PF was directly correlated with increasing BMI. Pancreatic fistula developed in none of

**Table 2** Histological diagnosis of resected specimens and main surgical specific post-operative complications

	Overweight (n = 141)	Controls (n = 226)	P value
Histology			
Ductal adenocarcinoma of the pancreas	42.5%	49.5%	0.2
Periampullary cancers	20.6%	17.4%	0.5
Cholangiocarcinoma	4.9%	3.9%	0.8
Cystic tumors	12.8%	10.6%	0.6
Neuroendocrine tumors	5.7%	5.8%	1.0
Chronic pancreatitis	3.6%	9.7%	0.03
Other tumor types	9.9%	3.1%	0.009
Surgical complications			
Pancreatic fistula	20.0%	9.5%	0.006
Delayed gastric emptying	14.2%	11.5%	0.5
Bile leakage	2.8%	1.7%	0.5
Abdominal bleeding	4.2%	4.4%	1.0
Gastro-intestinal bleeding	2.1%	2.6%	1.0
Wound infection	4.9%	2.6%	0.3

the underweight patients (BMI < 18.5 kg/m<sup>2</sup>), 10% of normal weight patients (BMI ≥ 18.5 kg/m<sup>2</sup> and ≤ 24.9 kg/m<sup>2</sup>), in 16% of overweight patients (BMI ≥ 25 kg/m<sup>2</sup> and ≤ 29.9 kg/m<sup>2</sup>), and 32% of obese (BMI ≥ 30 kg/m<sup>2</sup>).

### Cost analysis

The mean overall cost for the overweight group was 30700 ± 1938 euros and 33140 ± 2692 euros in the control group, which was not statistically different ( $P = 0.4$ ) (Figure 1).

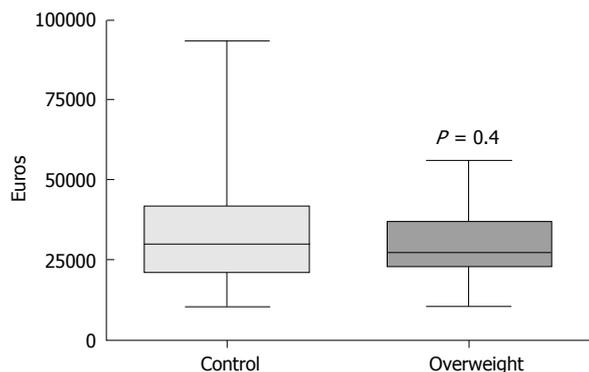
### Survival analysis

Overall, 60 patients (42.5%) in the overweight group and 112 patients (49.5%) in the control group were resected because of PDAC. One-year, 3-year, and 5-year actuarial survival rates (Figure 2A) were not different between the overweight group and the control group respectively (68.7%, 26.4%, and 8.8% *vs* 66.1%, 30.9%, and 17.9%;  $P = 0.9$ ). When the survival of PDAC patients was analyzed using 4 different categories of BMI (underweight, normal, overweight and obese), a trend was seen toward a difference in survival, with a worse prognosis at 1, 3 and 5 years for the underweight and obese patients (57.1%, 0%, 0% and 77.8%, 51.8%, 0%, respectively), compared to normal weight and overweight patients (67%, 32%, 18.4% and 66.5%, 20.6%, 10.3%, respectively) (Figure 2B). However, the survival comparison of the four groups was not significantly different ( $P = 0.8$ ).

## DISCUSSION

### General considerations

Overweight and obesity is becoming one of the most important health problems in Western countries. It is estimated that over \$140 million are spent each year on obesity-related diseases in the United States<sup>[17,21,22]</sup>. Few data are available concerning the impact of overweight on post-operative outcomes after pancreaticoduodenec-



**Figure 1** Comparison of in-hospital costs in the overweight and control groups.

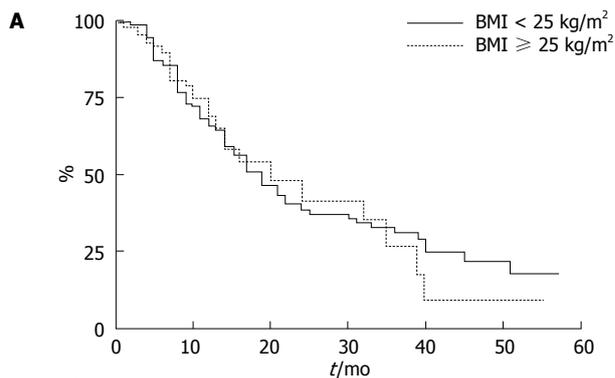
tomy and on the costs associated with the procedure. In addition, the limitations of the published studies make it difficult to analyze the data that are available. The available data were either collected retrospectively, or different criteria were used for dividing patients into sub-groups in the different investigations, or the study populations were not comparable.

**Impact of BMI in the short-term outcome of pancreaticoduodenectomies**

Some data suggest that pancreaticoduodenectomies (PD) is a more complicated operation in overweight patients. Our study is consistent with others<sup>[10,12]</sup> that have found comparable operative times in the control and overweight patients but higher intra-operative bleeding in the overweight group. Some studies have reported that overall morbidity after PD was not higher in overweight patients<sup>[10]</sup>. Similarly, we did not find any differences in overall rates of post-operative complications or in the severity of complications, even though overweight patients seem to be more prone to develop post-operative medical complications. We found an increased risk of PF in the overweight group, which is consistent with many other papers<sup>[11,12,23]</sup>, and found that the risk increased progressively in higher BMI categories. This result may be due to different pancreatic textures in the different BMI groups with an increasing percentage of fat infiltration in overweight patients that affected the quality of the pancreatico-jejunostomy anastomosis<sup>[11,24]</sup>. In our series, we did not find an increased risk of other BMI-related post-operative complications, such as wound infections, although such an increase has been reported in some other studies<sup>[9,15]</sup>. Previous papers have not looked at mortality rates, costs, or the LOS in overweight patients after PD. We did not find any differences in those areas, which is not surprising since there were no differences in the overall complication rate.

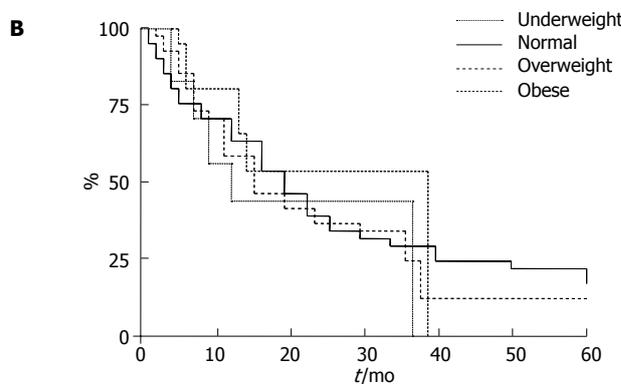
**Impact of BMI in the long-term outcome of PD**

Our study didn't show any statistically significant differences in survival in overweight patients compared to the control group. However, Fleming *et al*<sup>[18]</sup> reported decreased



# patients  
At risk (kg/m<sup>2</sup>)

BMI < 25	109	76	45	26	13	8	3
BMI ≥ 25	58	41	19	9	2	2	1



# patients  
At risk

Underweight	7	76	45	26	13	8	3
Normal weight	100	41	19	9	2	2	1
Overweight	48	76	45	26	13	8	3
Obese	9	41	19	9	2	2	1

**Figure 2** Actuarial survival curves. A: In overweight and control patients who underwent pancreaticoduodenectomy for pancreatic ductal adenocarcinoma (PDAC); B: For underweight, normal-weight, overweight, and obese patients who underwent pancreaticoduodenectomy for PDAC. BMI: Body mass index.

survival after PD in patients with a BMI >35 kg/m<sup>2</sup>. In contrast, a more recent study by Tsai *et al*<sup>[12]</sup> found that a higher BMI can be considered a positive long-term prognostic factor for patients undergoing PD for PDAC. In our analysis of patients treated for PDAC, we did not find any difference in survival between the overweight and control subgroups. In our sub-analysis using 4 different BMI categories, the underweight and obese patients had a worse prognosis than the normal weight individuals, although there were no differences between the normal weight and overweight patients. It is possible that the division of patients in two categories did not show a difference because the survival time of controls was negatively affected by the group of underweight patients and the overweight patients had a negative impact on the survival analysis of obese patients. Unfortunately, in our series, the extreme groups of BMI (underweight and obese) were too small to give a definitive answer to

this question. It would be interesting to study the impact of BMI on the long-term outcome of PD in a larger number of PDAC patients.

In conclusion, our data shows that PD can be done safely in overweight patients, even though with somewhat higher intra-operative blood loss, postoperative medical complications and PF rate. We also demonstrate that overweight/obesity do not impact survival rates negatively after PD for pancreatic cancer. However, the limited number of patients in the extreme groups of BMI limited the possibility to do a proper sub analysis useful to show the impact of underweight and obese patients in the short and long term outcome<sup>[25]</sup>. A sub analysis of fat distribution can also offer another area of research to predict the short and long term outcome of these patients<sup>[26]</sup>.

## COMMENTS

### Background

In the last decades, the number of overweight individuals has increased dramatically in Western countries. No data are available in the literature that show clearly whether this comorbidity has an impact on short-term or long-term outcomes in these patients or on procedure-related costs.

### Research frontiers

Some studies have shown that pancreatectomies in overweight patients are associated with an increased risk of post-operative complications. The data are even more confusing regarding long-term and oncologic outcomes.

### Innovations and breakthroughs

In their study, based on a large series of consecutive pancreaticoduodenectomies (PD) performed in a high volume center for pancreatic surgery, authors showed that body mass index (BMI) is a risk factor for intra-operative bleeding and post-operative pancreatic fistula, but does not increase the overall morbidity and have no impact on survival of patients with pancreatic ductal adenocarcinoma. Based on these results, BMI should not be considered, per-se, an exclusion criteria for candidates for PD.

### Peer review

In the present study, the authors investigated the impact of BMI on short and long term results after PD. This paper clearly demonstrated the relationship between BMI and outcome after PD for pancreatic cancer. It is worth publishing.

## REFERENCES

- Centers for Disease Control and Prevention (CDC). State-specific prevalence of obesity among adults--United States, 2005. *MMWR Morb Mortal Wkly Rep* 2006; **55**: 985-988
- Hubert HB, Feinleib M, McNamara PM, Castelli WP. Obesity as an independent risk factor for cardiovascular disease: a 26-year follow-up of participants in the Framingham Heart Study. *Circulation* 1983; **67**: 968-977 [PMID: 6219830 DOI: 10.1161/01.CIR.67.5.968]
- Manson JE, Colditz GA, Stampfer MJ, Willett WC, Rosner B, Monson RR, Speizer FE, Hennekens CH. A prospective study of obesity and risk of coronary heart disease in women. *N Engl J Med* 1990; **322**: 882-889 [PMID: 2314422 DOI: 10.1056/NEJM199003293221303]
- Hu FB, Manson JE, Stampfer MJ, Colditz G, Liu S, Solomon CG, Willett WC. Diet, lifestyle, and the risk of type 2 diabetes mellitus in women. *N Engl J Med* 2001; **345**: 790-797 [PMID: 11556298 DOI: 10.1056/NEJMoa010492]
- Bergström A, Pisani P, Tenet V, Wolk A, Adami HO. Overweight as an avoidable cause of cancer in Europe. *Int J Cancer* 2001; **91**: 421-430 [PMID: 11169969 DOI: 10.1002/1097-0215(200002)9999:9999<::AID-IJC1053>3.0.CO;2-T]
- Larsson SC, Orsini N, Wolk A. Body mass index and pancreatic cancer risk: A meta-analysis of prospective studies. *Int J Cancer* 2007; **120**: 1993-1998 [PMID: 17266034 DOI: 10.1002/ijc.22535]
- Adams KF, Schatzkin A, Harris TB, Kipnis V, Mouw T, Ballard-Barbash R, Hollenbeck A, Leitzmann MF. Overweight, obesity, and mortality in a large prospective cohort of persons 50 to 71 years old. *N Engl J Med* 2006; **355**: 763-778 [PMID: 16926275 DOI: 10.1056/NEJMoa055643]
- Finkelstein EA, Fiebelkorn IC, Wang G. National medical spending attributable to overweight and obesity: how much, and who's paying? *Health Aff (Millwood)* 2003; Suppl Web Exclusives: W3-219-W3-226 [PMID: 14527256 DOI: 10.1377/hlthaff.w3.219]
- Bennis M, Woodall C, Scoggins C, McMasters K, Martin R. The impact of obesity on outcomes following pancreatectomy for malignancy. *Ann Surg Oncol* 2009; **16**: 2565-2569 [PMID: 19557479 DOI: 10.1245/s10434-009-0573-7]
- Williams TK, Rosato EL, Kennedy EP, Chojnacki KA, Andrej J, Hyslop T, Doria C, Sauter PK, Bloom J, Yeo CJ, Berger AC. Impact of obesity on perioperative morbidity and mortality after pancreaticoduodenectomy. *J Am Coll Surg* 2009; **208**: 210-217 [PMID: 19228532 DOI: 10.1016/j.jamcollsurg.2008.10.019]
- Noun R, Riachy E, Ghorra C, Yazbeck T, Tohme C, Abboud B, Naderi S, Chalhoub V, Ayoub E, Yazbeck P. The impact of obesity on surgical outcome after pancreaticoduodenectomy. *JOP* 2008; **9**: 468-476 [PMID: 18648138]
- Tsai S, Choti MA, Assumpcao L, Cameron JL, Gleisner AL, Herman JM, Eckhauser F, Edil BH, Schulick RD, Wolfgang CL, Pawlik TM. Impact of obesity on perioperative outcomes and survival following pancreaticoduodenectomy for pancreatic cancer: a large single-institution study. *J Gastrointest Surg* 2010; **14**: 1143-1150 [PMID: 20431978 DOI: 10.1007/s11605-010-1201-3]
- Dindo D, Muller MK, Weber M, Clavien PA. Obesity in general elective surgery. *Lancet* 2003; **361**: 2032-2035 [PMID: 12814714]
- Su Z, Koga R, Saiura A, Natori T, Yamaguchi T, Yamamoto J. Factors influencing infectious complications after pancreaticoduodenectomy. *J Hepatobiliary Pancreat Sci* 2010; **17**: 174-179 [PMID: 19517055 DOI: 10.1007/s00534-009-0128-0]
- House MG, Fong Y, Arnaoutakis DJ, Sharma R, Winston CB, Protic M, Gonen M, Olson SH, Kurtz RC, Brennan MF, Allen PJ. Preoperative predictors for complications after pancreaticoduodenectomy: impact of BMI and body fat distribution. *J Gastrointest Surg* 2008; **12**: 270-278 [PMID: 18060467]
- Gaujoux S, Cortes A, Couvelard A, Noullet S, Clavel L, Rebours V, Lévy P, Sauvanet A, Ruszniewski P, Belghiti J. Fatty pancreas and increased body mass index are risk factors of pancreatic fistula after pancreaticoduodenectomy. *Surgery* 2010; **148**: 15-23 [PMID: 20138325 DOI: 10.1016/j.surg.2009.12.005]
- Balentine CJ, Enriquez J, Cruz G, Hodges S, Bansal V, Jo E, Ahern C, Sangsiry S, Petersen N, Silberfein E, Brunicaudi FC, Berger DH, Fisher W. Obesity does not increase complications following pancreatic surgery. *J Surg Res* 2011; **170**: 220-225 [PMID: 21514600 DOI: 10.1016/j.jss.2011.03.048]
- Fleming JB, Gonzalez RJ, Petzel MQ, Lin E, Morris JS, Gomez H, Lee JE, Crane CH, Pisters PW, Evans DB. Influence of obesity on cancer-related outcomes after pancreatectomy to treat pancreatic adenocarcinoma. *Arch Surg* 2009; **144**: 216-221 [PMID: 19289659 DOI: 10.1001/archsurg.2008.580]
- Jones L, Russell C, Mosca F, Boggi U, Sutton R, Slavin J, Hartley M, Neoptolemos JP. Standard Kausch-Whipple pancreatoduodenectomy. *Dig Surg* 1999; **16**: 297-304 [PMID: 10449974 DOI: 10.1159/000018739]
- Dindo D, Demartines N, Clavien PA. Classification of surgical complications: a new proposal with evaluation in a cohort of 6336 patients and results of a survey. *Ann*

- Surg* 2004; **240**: 205-213 [PMID: 15273542 DOI: 10.1097/01.sla.0000133083.54934.ae]
- 21 **Finkelstein EA**, Trogon JG, Cohen JW, Dietz W. Annual medical spending attributable to obesity: payer-and service-specific estimates. *Health Aff (Millwood)* 2009; **28**: w822-w831 [PMID: 19635784 DOI: 10.1377/hlthaff.28.5.w822]
- 22 **Ogden CL**, Carroll MD, Curtin LR, McDowell MA, Tabak CJ, Flegal KM. Prevalence of overweight and obesity in the United States, 1999-2004. *JAMA* 2006; **295**: 1549-1555 [PMID: 16595758 DOI: 10.1001/jama.295.13.1549]
- 23 **Ramsey AM**, Martin RC. Body mass index and outcomes from pancreatic resection: a review and meta-analysis. *J Gastrointest Surg* 2011; **15**: 1633-1642 [PMID: 21484490 DOI: 10.1007/s11605-011-1502-1]
- 24 **Mathur A**, Marine M, Lu D, Swartz-Basile DA, Saxena R, Zyromski NJ, Pitt HA. Nonalcoholic fatty pancreas disease. *HPB (Oxford)* 2007; **9**: 312-318 [PMID: 18345311 DOI: 10.1080/13651820701504157]
- 25 **Vaid S**, Bell T, Grim R, Ahuja V. Predicting risk of death in general surgery patients on the basis of preoperative variables using american college of surgeons national surgical quality improvement program data. *Perm J* 2012; **16**: 10-17 [PMID: 23251111 DOI: 10.7812/TPP/12-019]
- 26 **Gaujoux S**, Torres J, Olson S, Winston C, Gonen M, Brennan MF, Klimstra DS, D'Angelica M, DeMatteo R, Fong Y, House M, Jarnagin W, Kurtz RC, Allen PJ. Impact of obesity and body fat distribution on survival after pancreaticoduodenectomy for pancreatic adenocarcinoma. *Ann Surg Oncol* 2012; **19**: 2908-2916 [PMID: 22411205 DOI: 10.1245/s10434-012-2301-y]

**P- Reviewers** Hokama A, Miyazawa R **S- Editor** Gou SX  
**L- Editor** A **E- Editor** Yan JL



# World Journal of *Gastrointestinal Pathophysiology*

*World J Gastrointest Pathophysiol* 2013 August 15; 4(3): 43-64





**Contents**

Quarterly Volume 4 Number 3 August 15, 2013

- |                              |    |   |
|------------------------------|----|---|
| <b>FRONTIER</b>              | 43 | Primary clarithromycin resistance to <i>Helicobacter pylori</i> : Is this the main reason for triple therapy failure?<br><i>Giorgio F, Principi M, De Francesco V, Zullo A, Losurdo G, Di Leo A, Ierardi E</i>      |
| <b>MINIREVIEWS</b>           | 47 | Probiotics for the treatment of <i>Clostridium difficile</i> associated disease<br><i>Fitzpatrick LR</i>  |
| <b>BRIEF ARTICLE</b>         | 53 | Altered molecular pattern of mucosal healing in Crohn's disease fibrotic stenosis<br><i>Ierardi E, Giorgio F, Piscitelli D, Principi M, Cantatore S, Fiore MG, Rossi R, Barone M, Di Leo A, Panella C</i>           |
| <b>CASE REPORT</b>           | 59 | Severe hemorrhagic colitis in a patient with chronic myeloid leukemia in the blastic phase after dasatinib use<br><i>Kmira Z, Nesrine BS, Houneida Z, Wafa BF, Aida S, Yosra BY, Monia Z, Sriha B, Abderrahim K</i> |
| <b>LETTERS TO THE EDITOR</b> | 63 | Why is damage limited to the mucosa in ulcerative colitis but transmural in Crohn's disease?<br><i>Qin X</i>  |

**Contents***World Journal of Gastrointestinal Pathophysiology*  
Volume 4 Number 3 August 15, 2013**APPENDIX** I-V Instructions to authors**ABOUT COVER** Editorial Board Member of *World Journal of Gastrointestinal Pathophysiology*, Leo R Fitzpatrick, PhD, Associate Professor, Department of Pharmacology, Penn State College of Medicine, 1214 Research Boulevard, Hummelstown, PA 17036, United States**AIM AND SCOPE***World Journal of Gastrointestinal Pathophysiology* (*World J Gastrointest Pathophysiol*, *WJGP*, online ISSN 2150-5330, DOI: 10.4291), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.*WJGP* is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. *WJGP* specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.We encourage authors to submit their manuscripts to *WJGP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.**INDEXING/ ABSTRACTING***World Journal of Gastrointestinal Pathophysiology* is now indexed in PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.**FLYLEAF** I-IV Editorial Board**EDITORS FOR THIS ISSUE**Responsible Assistant Editor: *Xin-Xin Che*  
Responsible Electronic Editor: *Jun-Yao Li*  
Proofing Editor-in-Chief: *Lian-Sheng Ma*Responsible Science Editor: *Ling-Ling Wen***NAME OF JOURNAL**  
*World Journal of Gastrointestinal Pathophysiology***ISSN**  
ISSN 2150-5330 (online)**LAUNCH DATE**  
April 15, 2010**Frequency**  
Quarterly**EDITOR-IN-CHIEF**  
**Thomas Y Ma, MD, PhD, Professor, Chief**, Division of Gastroenterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States**EDITORIAL OFFICE**  
Jin-Lei Wang, Director  
Xiu-Xia Song, Vice Director*World Journal of Gastrointestinal Pathophysiology*  
Room 903, Building D, Ocean International Center,  
No. 62 Dongsihuan Zhonglu, Chaoyang District,  
Beijing 100025, China  
Telephone: +86-10-85381892  
Fax: +86-10-85381893  
E-mail: [wjgp@wjgnet.com](mailto:wjgp@wjgnet.com)  
<http://www.wjgnet.com>**PUBLISHER**  
Baishideng Publishing Group Co., Limited  
Flat C, 23/F, Lucky Plaza,  
315-321 Lockhart Road, Wan Chai,  
Hong Kong, China  
Telephone: +852-6555-7188  
Fax: +852-3177-9906  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
<http://www.wjgnet.com>**PUBLICATION DATE**  
August 15, 2013**COPYRIGHT**

© 2013 Baishideng. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

**INSTRUCTIONS TO AUTHORS**Full instructions are available online at [http://www.wjgnet.com/2150-5330/g\\_info\\_20100316080008.htm](http://www.wjgnet.com/2150-5330/g_info_20100316080008.htm)**ONLINE SUBMISSION**<http://www.wjgnet.com/esp/>

## Primary clarithromycin resistance to *Helicobacter pylori*: Is this the main reason for triple therapy failure?

Floriana Giorgio, Mariabeatrice Principi, Vincenzo De Francesco, Angelo Zullo, Giuseppe Losurdo, Alfredo Di Leo, Enzo Ierardi

Floriana Giorgio, Vincenzo De Francesco, Enzo Ierardi, Division of Gastroenterology, Department of Medical and Surgical Sciences, University of Foggia, 71100 Foggia, Italy  
Mariabeatrice Principi, Giuseppe Losurdo, Alfredo Di Leo, Division of Gastroenterology, Department of Emergency and Organ Transplantation, University of Bari, 70124 Bari, Italy  
Angelo Zullo, Department of Gastroenterology and Digestive Endoscopy, "Nuovo Regina Margherita" Hospital, 00153 Rome, Italy

Author contributions: De Francesco V, Zullo A, Di Leo A and Ierardi E designed the study, revised the manuscript and approved the final version; Giorgio F and Principi M and Losurdo G collected the data.

Correspondence to: Enzo Ierardi, Professor, Division of Gastroenterology, Department of Medical and Surgical Sciences, University of Foggia, AOU Ospedali Riuniti, Viale Pinto, 71100 Foggia, Italy. [enzo.ierardi@fastwenet.it](mailto:enzo.ierardi@fastwenet.it)

Telephone: +39-8-81736204 Fax: +39-8-81733848

Received: May 16, 2013 Revised: June 20, 2013

Accepted: July 4, 2013

Published online: August 15, 2013

### Abstract

Conventional triple therapies for *Helicobacter pylori* (*H. pylori*) eradication have recently shown a disappointing reduction in effectiveness in many countries. The main reason for failure was found to be bacterial resistance to one of the most commonly used antibiotics, clarithromycin. An additional problem for conventional triple therapy is the high rate of resistance to metronidazole found in Europe, America and Asia. In Italy, in the last 15 years a 2-fold increase in resistance has occurred. A recent study of the whole of Italy included about 20 patients from each region at the first endoscopic diagnosis of *H. pylori* infection. The most surprising result was the patchy distribution of resistance, which was almost absent in two regions (one northern and one southern), although the highest prevalence was found in some regions of the South. In the paediatric

population we found a 25% prevalence of resistance in a sample of *H. pylori* positive children observed between 2002 and 2007, mirroring data obtained in southern European countries. Clarithromycin resistance assessment is currently based on phenotypic detection performed after culture the agar dilution method or E-test, and genotypic methods based on polymerase chain reaction (PCR). In a recent comparative study we found a 71.2% agreement between the two methods. Culture-free techniques are highly accurate in finding even minimal traces of genotypically resistant strains. Moreover, PCR-based tools are accurate in detecting a heteroresistant status, defined as the co-existence of some strains that are susceptible and some resistant to the same antibiotic in an individual patient. Three point mutations, namely *A2143G*, *A2142G* and *A2142C*, are responsible for 90% of cases of primary clarithromycin resistance in *H. pylori* strains isolated in Western countries, although we previously demonstrated that the presence of the *A2143G* mutation, but not *A2142G* or *A2142C*, significantly lowered the *H. pylori* eradication rate. Treatment failure has considerable cost/benefit implications because of "waste" of National Health System and patient resources, in terms of drugs, further diagnostic tests and medical examination expenses. Therefore, in future it would be very useful to be able to test for clarithromycin resistance before starting conventional triple therapy. Hopefully, fast, effective non-invasive tests may soon be devised to determine this condition.

© 2013 Baishideng. All rights reserved.

**Key words:** *Helicobacter pylori*; Clarithromycin; Genotypic resistance; Phenotypic resistance; Therapy failure cost; Resistance epidemiology

**Core tip:** Clarithromycin resistance is the main reason for the failure of conventional therapies for *Helicobacter pylori*. We evaluate the scope of the problem,

as reported in literature and especially on the basis of personal data in adult and paediatric populations. Another issue is the detection of resistance using phenotypic and genotypic methods; comparison is made of the limits and advantages of these approaches. Cost/benefit analysis of unsuccessful eradication therapy is performed. Based on these considerations, the best solution in future seems likely to be the detection of resistant strains before starting treatment.

Giorgio F, Principi M, De Francesco V, Zullo A, Losurdo G, Di Leo A, Ierardi E. Primary clarithromycin resistance to *Helicobacter pylori*: Is this the main reason for triple therapy failure? *World J Gastrointest Pathophysiol* 2013; 4(3): 43-46 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i3/43.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i3.43>

## BACKGROUND

*Helicobacter pylori* (*H. pylori*) eradication may have clinical implications with a very high impact: healing of peptic ulcer disease, resolution of low-grade lymphoma in about 80% of cases and the prevention of gastric adenocarcinoma<sup>[1,2]</sup>. However, some problems have arisen in clinical practice in recent years, namely the decreased eradication rate of conventional therapies and the onset and diffusion of antibiotic resistances worldwide<sup>[3]</sup>. Triple therapy, comprising treatment with two antibiotics, amoxicillin and clarithromycin, and a proton pump inhibitor for a week, was recommended as the treatment of choice at several consensus conferences. However, this treatment has recently shown a disappointing reduction in effectiveness in many countries and the main reason for failure has been found to be bacterial resistance to one of the most commonly used antibiotics, clarithromycin<sup>[4]</sup>.

## PREVALENCE OF RESISTANCE TO CLARITHROMYCIN

An assessment of primary clarithromycin resistance made about 10 years ago demonstrated differences between children and adults in Europe. In adults, a marked difference has been observed between Northern countries, where the overall prevalence was about 5%, and Southern countries, where the resistance rate was up to 20%. This difference was not observed in children. Indeed, in all European countries a virtually equal, high prevalence has been reported, ranging from 12.5% to 23.5%. An additional problem for conventional triple therapy is the high rate of resistance to metronidazole<sup>[5,6]</sup> found in Europe (from 23.5% in Spain to 40.3% in the United Kingdom), America (from 21.6% in the United States to 76.3% in Mexico) and Asia (from 9%-12% in Japan to 41.9% in Korea).

In Italy, data obtained 10 years ago showed a strong difference between the resistance rates in Northern (2%)

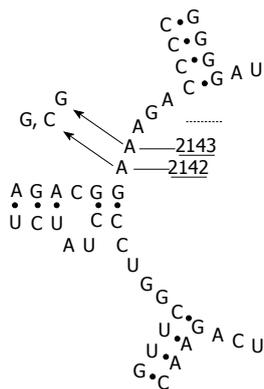
and Central (20%) areas<sup>[7,8]</sup>. However, in the same period a study by our group found a median value of 28.7% in two areas in Southern and Central Italy<sup>[9]</sup>. In 2007, in a retrospective study performed over a 15-year period in the same geographical area, we found a 10.2% prevalence of primary clarithromycin resistance in 147 patients in the 1990 period, whereas by 2004-2005 the resistance found in 178 patients had increased significantly, to 21.3% or 2-fold higher<sup>[10]</sup>. The phenomenon was found to be more evident in females. A possible explanation for this finding may be a wider use of the antibiotic in the course of infections with a greater prevalence in the female sex, such as urinary tract infections<sup>[11,12]</sup>.

A more recent study (2011) was focused on the whole of Italy, including about 20 patients from each region at the first endoscopic diagnosis of *H. pylori* infection. The most surprising result was the patchy distribution of resistance, which was almost absent in two regions (one Northern and one Southern) although the highest prevalence was found in some regions of the South (20%-25%). Due to this distribution variability, the overall resistance rate was lower than expected (about 10%). Resistances were higher in patients with non-ulcer dyspepsia than in those with peptic ulcer, in agreement with findings in previous investigations<sup>[13]</sup>.

In the paediatric population we found a 25% prevalence of resistance in a sample of 168 *H. pylori* positive children observed between 2002 and 2007, mirroring other data from European countries, where high values reflect the environmental conditions of southern areas<sup>[14]</sup>.

## METHODS FOR CLARITHROMYCIN RESISTANCE DETECTION

Clarithromycin resistance assessment is currently based on phenotypic detection performed after culture and the agar dilution method or E-test. However, in the past decade, different polymerase chain reaction (PCR) based approaches have been developed as alternative tools<sup>[15]</sup>. These techniques allow assessment of mutations in the peptidyltransferase region encoded in domain V of the *H. pylori* 23S ribosomal RNA region that confers clarithromycin resistance<sup>[16,17]</sup>. Undeniably, both culture and PCR-based methods have both advantages and limitations<sup>[18]</sup>. Bacterial culture allows an overall evaluation of *H. pylori* clarithromycin resistance, regardless of the intrinsic mechanism involved (point mutations, RNA methylations, efflux pumps, *etc.*). Nevertheless, *H. pylori* is a troublesome bacterium and culture may be difficult even in expert hands. Indeed, sensitivity values of culture as low as 55%-73% have been reported in some trials<sup>[19]</sup>. By contrast, PCR-based culture-free techniques are highly accurate in finding even minimal traces of genotypically resistant strains. Moreover, PCR-based tools are accurate in detecting a heteroresistant status, defined as the co-existence of some strains that are susceptible and some resistant to the same antibiotic in an individual patient. These techniques may be used even on paraffin-embed-



**Figure 1** Secondary structure model of the peptidyltransferase region in domain V of the *Helicobacter pylori* 23S rRNA. The site of the A2142C, A2142G and A2143G mutations is underlined.

ded tissue<sup>[20]</sup>. Nevertheless, these approaches are unable to detect clarithromycin resistance when it is based on uncommon genetic mechanisms (deletions, RNA methylations, *etc.*). Although several mutations have been detected, it has been found that three point mutations, namely A2143G, A2142G and A2142C (Figure 1), are responsible for 90% of cases of primary clarithromycin resistance in *H. pylori* strains isolated in Western countries<sup>[21,22]</sup>. Some studies showed that these point mutations are associated with different MIC values for clarithromycin resistance assessed by culture *in vitro*<sup>[23]</sup>, suggesting that they might have a different impact on the therapeutic outcome. Indeed, we previously found that the presence of the A2143G mutation, but not A2142G or A2142C, significantly lowered the *H. pylori* eradication rate<sup>[24,25]</sup>.

In a recent comparative study we found that the prevalence of clarithromycin phenotypic resistance was significantly lower than that of genotypic resistance (18.4% *vs* 37.6%,  $P < 0.001$ ). An agreement between the two methods was present in 71.2% of cases. A significant difference in the eradication rate was seen between clarithromycin-susceptible and resistant strains, when assessed with either the *E* test (92.4% *vs* 55.5%,  $P < 0.001$ ) or a PCR-based method (94.5% *vs* 70.9%,  $P < 0.001$ ). Notably, the eradication rate showed the lowest value (30.7%) when the phenotypic bacterial resistance was genetically linked to the A2143G point mutation. The conclusion of this study was that there is a disagreement between the two methods; clarithromycin resistance markedly reduces *H. pylori* eradication only when it is linked to a specific point mutation<sup>[26]</sup>.

On these grounds, it has been suggested that MIC values for clarithromycin resistance should be dropped to 0.5 in order to improve the efficacy of phenotypic detection<sup>[27]</sup>.

## COST/BENEFIT IMPLICATIONS IN ITALY

Conventional 7-d triple therapy failure leads to a “waste” of about 27 € in each case of resistance to clarithromycin. The sum is doubled when the therapy duration is prolonged to 10-14 d, as suggested by some reports<sup>[28]</sup>. If

sequential therapy is used, the cost of treatment is about 40 € but the therapeutic gain is unsatisfactory.

In addition to the net cost of drugs, the cost of support treatment used in cases of side effects (probiotics, prokinetics, anti-acids), accounts for a further 5-25 € on average. In cases of therapeutic failure the patient will undergo a new medical examination (average cost: 36 €) and new surveys for *H. pylori* status by non-invasive (urea breath test: 70 €, stool antigen: 30-40 €) and invasive methods (esophagogastroduodenoscopy: 220 + 36 € of individual contribution; biopsy: 40 €; histological examination: 95 + 36 € of individual contribution)<sup>[29]</sup>.

In this scenario it is clear that inappropriate use of an anti *H. pylori* regimen incurs heavy costs not only in terms of the initial, ineffective treatment but also of the later additional visits and investigations.

## FUTURE PERSPECTIVES

In future it will be more effective to test for clarithromycin resistance before starting conventional triple therapy. The limit of this approach is the high cost of molecular analysis, as well as the difficulty of performing phenotypic investigations, even for experts. Other issues hindering a wider spread of these assessments in clinical practice are the long time between testing and obtaining the results on which to plan therapy, and the need to perform an invasive examination (esophagogastro-duodenoscopy). In conclusion, it would be very useful to be able to test for clarithromycin resistance before starting conventional triple therapy. Hopefully, fast, effective non-invasive tests may soon be devised to determine this condition.

## REFERENCES

- 1 **Graham DY**, Lew GM, Klein PD, Evans DG, Evans DJ, Saeed ZA, Malaty HM. Effect of treatment of Helicobacter pylori infection on the long-term recurrence of gastric or duodenal ulcer. A randomized, controlled study. *Ann Intern Med* 1992; **116**: 705-708 [PMID: 1558340 DOI: 10.7326/0003-4819-116-9-705]
- 2 **Malfertheiner P**, Mégraud F, O’Morain C, Hungin AP, Jones R, Axon A, Graham DY, Tytgat G. Current concepts in the management of Helicobacter pylori infection--the Maastricht 2-2000 Consensus Report. *Aliment Pharmacol Ther* 2002; **16**: 167-180 [PMID: 11860399 DOI: 10.1046/j.1365-2036.2002.01169.x]
- 3 **De Francesco V**, Giorgio F, Hassan C, Manes G, Vannella L, Panella C, Ierardi E, Zullo A. Worldwide H. pylori antibiotic resistance: a systematic review. *J Gastrointest Liver Dis* 2010; **19**: 409-414 [PMID: 21188333]
- 4 **Mégraud F**. H pylori antibiotic resistance: prevalence, importance, and advances in testing. *Gut* 2004; **53**: 1374-1384 [PMID: 15306603 DOI: 10.1136/gut.2003.022111]
- 5 **Perez Aldana L**, Kato M, Nakagawa S, Kawarasaki M, Nagasaki T, Mizushima T, Oda H, Kodaira J, Shimizu Y, Komatsu Y, Zheng R, Takeda H, Sugiyama T, Asaka M. The relationship between consumption of antimicrobial agents and the prevalence of primary Helicobacter pylori resistance. *Helicobacter* 2002; **7**: 306-309 [PMID: 12390210 DOI: 10.1046/j.1523-5378.2002.00096.x]
- 6 **Kato M**, Yamaoka Y, Kim JJ, Reddy R, Asaka M, Kashima K, Osato MS, El-Zaatari FA, Graham DY, Kwon DH. Regional

- differences in metronidazole resistance and increasing clarithromycin resistance among *Helicobacter pylori* isolates from Japan. *Antimicrob Agents Chemother* 2000; **44**: 2214-2216 [PMID: 10898707]
- 7 **Toracchio S**, Marzio L. Primary and secondary antibiotic resistance of *Helicobacter pylori* strains isolated in central Italy during the years 1998-2002. *Dig Liver Dis* 2003; **35**: 541-545 [PMID: 14567457 DOI: 10.1016/S1590-8658(03)00265-2]
  - 8 **Pilotto A**, Rassu M, Leandro G, Franceschi M, Di Mario F. Prevalence of *Helicobacter pylori* resistance to antibiotics in Northeast Italy: a multicentre study. GISU. Interdisciplinary Group for the Study of Ulcer. *Dig Liver Dis* 2000; **32**: 763-768 [PMID: 11215555 DOI: 10.1016/S1590-8658(00)80352-7]
  - 9 **de Francesco V**, Margiotta M, Zullo A, Hassan C, Valle ND, Burattini O, Cea U, Stoppino G, Amoruso A, Stella F, Morini S, Panella C, Ierardi E. Primary clarithromycin resistance in Italy assessed on *Helicobacter pylori* DNA sequences by TaqMan real-time polymerase chain reaction. *Aliment Pharmacol Ther* 2006; **23**: 429-435 [PMID: 16423002 DOI: 10.1111/j.1365-2036.2006.02769.x]
  - 10 **De Francesco V**, Margiotta M, Zullo A, Hassan C, Giorgio F, Burattini O, Stoppino G, Cea U, Pace A, Zotti M, Morini S, Panella C, Ierardi E. Prevalence of primary clarithromycin resistance in *Helicobacter pylori* strains over a 15 year period in Italy. *J Antimicrob Chemother* 2007; **59**: 783-785 [PMID: 17329269 DOI: 10.1093/jac/dkm005]
  - 11 **August SL**, De Rosa MJ. Evaluation of the prevalence of urinary tract infection in rural Panamanian women. *PLoS One* 2012; **7**: e47752 [PMID: 23094080]
  - 12 **Boyanova L**, Ilieva J, Gergova G, Davidkov L, Spassova Z, Kamburov V, Katsarov N, Mitov I. Numerous risk factors for *Helicobacter pylori* antibiotic resistance revealed by extended anamnesis: a Bulgarian study. *J Med Microbiol* 2012; **61**: 85-93 [PMID: 21873378 DOI: 10.1099/jmm.0.035568-0]
  - 13 **De Francesco V**, Giorgio F, Ierardi E, Zotti M, Neri M, Milano A, Varasano V, Luzzza F, Suraci E, Marmo R, Marone A, Manta R, Mirante VG, de Mattheis M, Pedroni A, Manes G, Pallotta S, Usai P, Liggi M, Gatto G, Peri V, Sacco R, Bresci G, Monica F, Hassan C, Zullo A. Primary clarithromycin resistance in *Helicobacter pylori*: the Multicentric Italian Clarithromycin Resistance Observational (MICRO) study. *J Gastrointest Liver Dis* 2011; **20**: 235-239 [PMID: 21961089]
  - 14 **Francavilla R**, Lionetti E, Castellaneta S, Margiotta M, Piscitelli D, Polimeno L, Cavallo L, Ierardi E. Clarithromycin-Resistant Genotypes and Eradication of *Helicobacter Pylori*. *J Pediatr* 2010; **157**: 228-232 [DOI: 10.1016/j.jpeds.2010.02.007]
  - 15 **Francesco VD**, Zullo A, Hassan C, Giorgio F, Rosania R, Ierardi E. Mechanisms of *Helicobacter pylori* antibiotic resistance: An updated appraisal. *World J Gastrointest Pathophysiol* 2011; **2**: 35-41 [PMID: 21860834 DOI: 10.4291/wjgp.v2.i3.35]
  - 16 **Wada T**, Maeda S, Tamaru A, Imai S, Hase A, Kobayashi K. Dual-probe assay for rapid detection of drug-resistant *Mycobacterium tuberculosis* by real-time PCR. *J Clin Microbiol* 2004; **42**: 5277-5285 [PMID: 15528726 DOI: 10.1128/JCM.42.1.5277-5285.2004]
  - 17 **Baglan PH**, Bozdayi G, Ozkan M, Ahmed K, Bozdayi AM, Ozden A. Clarithromycin resistance prevalence and *IcaA* gene status in *Helicobacter Pylori* clinical isolates in Turkish patients with duodenal ulcer and functional dyspepsia. *J Microbiol* 2006; **44**: 409-416 [PMID: 16953176]
  - 18 **Monno R**, Giorgio F, Carmine P, Soleo L, Cinquepalmi V, Ierardi E. *Helicobacter pylori* clarithromycin resistance detected by Etest and TaqMan real-time polymerase chain reaction: a comparative study. *APMIS* 2012; **120**: 712-717 [PMID: 22882260 DOI: 10.1111/j.1600-0463.2012.02896.x]
  - 19 **Zullo A**, Hassan C, Lorenzetti R, Winn S, Morini S. A clinical practice viewpoint: to culture or not to culture *Helicobacter pylori*? *Dig Liver Dis* 2003; **35**: 357-361 [PMID: 12846409 DOI: 10.1016/S1590-8658(03)00081-1]
  - 20 **Paireder S**, Werner B, Bailer J, Werther W, Schmid E, Patzak B, Cichna-Markl M. Comparison of protocols for DNA extraction from long-term preserved formalin fixed tissues. *Anal Biochem* 2013; **439**: 152-160 [PMID: 23603300 DOI: 10.1016/j.ab.2013.04.006]
  - 21 **Matsumura M**, Hikiba Y, Ogura K, Togo G, Tsukuda I, Ushikawa K, Shiratori Y, Omata M. Rapid detection of mutations in the 23S rRNA gene of *Helicobacter pylori* that confers resistance to clarithromycin treatment to the bacterium. *J Clin Microbiol* 2001; **39**: 691-695 [PMID: 11158129]
  - 22 **Oleastro M**, Ménard A, Santos A, Lamouliatte H, Monteiro L, Barthélémy P, Mégraud F. Real-time PCR assay for rapid and accurate detection of point mutations conferring resistance to clarithromycin in *Helicobacter pylori*. *J Clin Microbiol* 2003; **41**: 397-402 [PMID: 12517879 DOI: 10.1128/JCM.41.1.397-402.2003]
  - 23 **De Francesco V**, Zullo A, Ierardi E, Vaira D. Minimal inhibitory concentration (MIC) values and different point mutations in the 23S rRNA gene for clarithromycin resistance in *Helicobacter pylori*. *Dig Liver Dis* 2009; **41**: 610-611 [PMID: 19200790 DOI: 10.1016/j.dld.2009.01.001]
  - 24 **De Francesco V**, Margiotta M, Zullo A, Hassan C, Troiani L, Burattini O, Stella F, Di Leo A, Russo F, Marangi S, Monno R, Stoppino V, Morini S, Panella C, Ierardi E. Clarithromycin-resistant genotypes and eradication of *Helicobacter pylori*. *Ann Intern Med* 2006; **144**: 94-100 [PMID: 16418408 DOI: 10.7326/0003-4819-144-2-200601170-00006]
  - 25 **De Francesco V**, Zullo A, Ierardi E, Giorgio F, Perna F, Hassan C, Panella C, Vaira D. The A2143G point mutation of clarithromycin resistance affects *Helicobacter pylori* eradication. *J Clin Gastroenterol* 2009; **43**: 386 [PMID: 19077730 DOI: 10.1097/MCG.0b013e318178d1e7]
  - 26 **De Francesco V**, Zullo A, Ierardi E, Giorgio F, Perna F, Hassan C, Morini S, Panella C, Vaira D. Phenotypic and genotypic *Helicobacter pylori* clarithromycin resistance and therapeutic outcome: benefits and limits. *J Antimicrob Chemother* 2010; **65**: 327-332 [PMID: 20008044 DOI: 10.1093/jac/dkp445]
  - 27 European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 2.0, valid from 2012-01-01: 67. Available from: URL: [http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Disk\\_test\\_documents/EUCAST\\_breakpoints\\_v\\_2.0\\_120101.pdf](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Disk_test_documents/EUCAST_breakpoints_v_2.0_120101.pdf)
  - 28 **De Francesco V**, Della Valle N, Stoppino V, Amoruso A, Muscatiello N, Panella C, Ierardi E. Effectiveness and pharmaceutical cost of sequential treatment for *Helicobacter pylori* in patients with non-ulcer dyspepsia. *Aliment Pharmacol Ther* 2004; **19**: 993-998 [PMID: 15113366 DOI: 10.1111/j.1365-2036.2004.01877.x]
  - 29 Italian pharmaceutical reference book 2013. Health Care Network. Euromedia s. r. l. Available from: URL: <http://www.prontuariofarmaceutico.it/>

**P- Reviewers** Figura N, Schwarz SM **S- Editor** Gou SX  
**L- Editor** Hughes D **E- Editor** Li JY



## Probiotics for the treatment of *Clostridium difficile* associated disease

Leo R Fitzpatrick

Leo R Fitzpatrick, Department of Pharmacology, Penn State College of Medicine Hummelstown, PA 17036, United States

Author contributions: Fitzpatrick LR wrote the paper.

Correspondence to: Leo R Fitzpatrick, PhD, Associate Professor, Department of Pharmacology, Penn State College of Medicine, 1214 Research Boulevard, Hummelstown, PA 17036, United States. [lfitzpatrick@psu.edu](mailto:lfitzpatrick@psu.edu)

Telephone: +1-717-310393 Fax: +1-717-5315013

Received: April 15, 2013 Revised: May 15, 2013

Accepted: June 1, 2013

Published online: August 15, 2013

### Abstract

The purpose of this review paper is to update the current and potential future role of probiotics for *Clostridium difficile*-associated disease (CDAD). Included in this review, is an update on the testing of newer probiotics (*e.g.*, *Bacillus coagulans* GBI-30, 6086) in animal models of CDAD. There is a focus on the modulation of signal transduction pathways (*i.e.*, transcription factors like cAMP response element-binding, activator protein 1, and nuclear factor kappa B), as well as the inhibition of certain kinases (*e.g.*, p38 mitogen activated protein kinases) by probiotics. Inhibition of signal transduction by probiotics, such as *Saccharomyces boulardii*, result in multiple effects on intestinal fluid secretion, neutrophil influx into the colon, inflammation, and colonocyte apoptosis that may positively impact CDAD. Recent clinical approaches with probiotics, for the prevention of primary and recurrent CDAD, are also summarized in this review paper. Future directions for the treatment of CDAD by probiotics are also mentioned in this review. In particular, the use of multi-strain probiotic formulations such as Ecologic<sup>®</sup> AAD and VSL #3<sup>®</sup> may represent a rationale pharmacological approach, particularly as adjunctive therapies for CDAD. Understanding the mechanistic basis of CDAD, and how probiotics interfere at certain steps in the pathogenic process, may also present the opportunity to design other multi-strain

probiotics that could have a future impact on CDAD.

© 2013 Baishideng. All rights reserved.

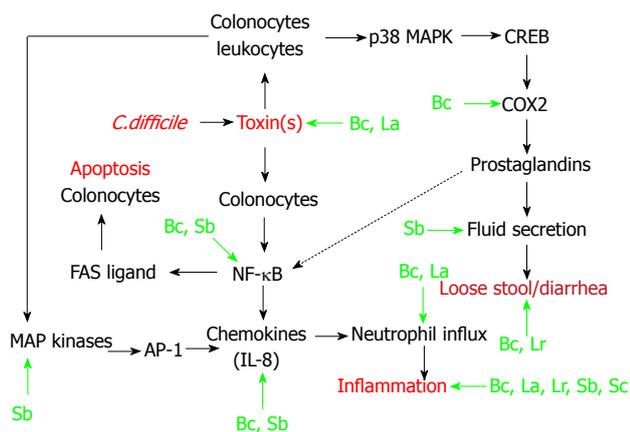
**Key words:** *Clostridium difficile*; Colitis; Probiotics; Mechanisms of action; Immune modulation; Transcription factors; *Saccharomyces boulardii*; VSL#3

**Core tip:** Certain probiotics can inhibit signal transduction pathways (*i.e.*, transcription factors like cAMP response element-binding, activator protein 1, and nuclear factor kappa B), as well as attenuate the activation of certain kinases (*e.g.*, p38 mitogen activated protein kinases). Inhibition of these intracellular signaling pathways by probiotics results in effects on intestinal fluid secretion, neutrophil influx into the colon, inflammation and colonocyte apoptosis that may positively impact *Clostridium difficile*-associated disease (CDAD). Understanding the mechanistic basis of CDAD, and how probiotics interfere at certain steps in the pathogenic process, may allow the development of novel probiotics that could have a future pharmacological impact on CDAD.

Fitzpatrick LR. Probiotics for the treatment of *Clostridium difficile* associated disease. *World J Gastrointest Pathophysiol* 2013; 4(3): 47-52 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i3/47.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i3.47>

### INTRODUCTION

*Clostridium difficile* (*C. difficile*) infection can cause nosocomial-related diarrhea and other distinct disease characteristics, which can affect the structural integrity of the intestine<sup>[1,2]</sup>. The spectrum of *C. difficile*-associated disease (CDAD) ranges from mild antibiotic associated diarrhea to severe pseudomembranous colitis that can lead



**Figure 1 Immunomodulation by probiotics for *Clostridium difficile*-associated disease.** *Clostridium difficile* (*C. difficile*) associated toxins (red font) engage colonic epithelial cells (colonocytes) leading to nuclear factor-kappa B (NF-κB) activation, interleukin (IL)-8 production, neutrophil influx and inflammation. These toxins also bind to receptors on colonocytes and leukocytes leading to p38 mitogen activated protein kinases (p38 MAPK) and cyclic-AMP response binding protein (CREB) activation. CREB, through cyclooxygenase 2 (COX2), is critical for the production of prostaglandin E2. In turn, this prostaglandin plays an important role in the fluid secretion/diarrhea associated with CDAD. *C. difficile* associated toxins also lead to the activation of other MAP kinases (ERK 1/2) and activator protein-1 (AP-1), which also plays a role in IL-8 production. There is also cross talk (dotted line) between the various pathways. For example, prostaglandin E2 can stimulate Fas ligand expression and apoptosis in colonic epithelial cells. The green arrows in this figure represent specific points of intervention by certain probiotics, resulting in immunomodulation by these agents. The abbreviations indicate the specific probiotics, which can modulate these signal transduction pathways. These probiotics (green font) include: *Saccharomyces boulardii* (Sb); *Bacillus coagulans* GBI-30, 6086 (Bc); *Lactobacillus acidophilus* (La); *Lactobacillus rhamnosus* (Lr); and *Saccharomyces cerevisiae*, strain 905 (Sc).

to mortality<sup>[1,2]</sup>. CDAD is caused by the actions of two exotoxins (toxin A and toxin B), which are produced by various pathogenic strains of *C. difficile*<sup>[2,3]</sup>.

CDAD is often treated successfully with standard antibiotics such as vancomycin and metronidazole<sup>[4-6]</sup>. However, recurrence occurs in many patients<sup>[4-6]</sup>. Some clinical studies have focused on combined treatment with vancomycin and probiotics such as *Saccharomyces boulardii* for the treatment of recurrence<sup>[7-10]</sup>. Therefore, initial treatment regimens with probiotics, or their use for prevention of recurrent disease, may be attractive as part of the overall therapeutic strategy for CDAD<sup>[11-13]</sup>.

Probiotics are live microorganisms that when ingested can confer health benefits<sup>[14]</sup>. Typically, probiotics include various strains of *Lactobacillus* and/or *Bifidobacteria* species. They exist as either single entities, or as combination products (e.g., VSL#3)<sup>[15,16]</sup>. Other known probiotics include certain non-pathogenic *Escherichia coli* strains like Nissle 1917 and M-17<sup>[14,17]</sup>.

Overall, the pertinent mechanisms explaining the potential role of probiotics as anti-colitis therapies have been reviewed in detail elsewhere<sup>[15,18-20]</sup>. The purpose of this review paper is to provide an update on the current and potential future role of probiotics for CDAD. Included in this review will be an update on the recent

testing of some probiotics in animal models of CDAD, as well as how certain probiotics can modulate signal transduction pathways.

## MECHANISMS OF ACTION FOR PROBIOTICS

### Focus on modulation of signal transduction (immunomodulation)

In an excellent review paper, Hell *et al*<sup>[19]</sup> cited potential mechanisms by which probiotics could prevent or reverse CDAD. These mechanisms included: (1) competitive exclusion; (2) bacterial metabolic activity; (3) preservation of gut-barrier function; (4) influence on water and ion channels; (5) influence on the innate nervous system; (6) modulation of signal transduction; (7) stimulation of the innate immune system; and (8) induction of adaptive immunity<sup>[19]</sup>. Specific details on these mechanisms are provided elsewhere in the relevant literature<sup>[18,19]</sup>.

In this review, I will focus on the modulation of signal transduction pathways (*i.e.*, immunomodulation) by probiotics, as related to CDAD<sup>[19,21,22]</sup>. As shown in Figure 1, endogenous colonic epithelial cells (colonocytes) seem to play an integral role in CDAD<sup>[23-25]</sup>. However, cells of the innate immune system (macrophages, neutrophils) also play a role in the etiology of CDAD<sup>[26,27]</sup>. In these cellular populations within the intestine, *C. difficile* associated toxins (particularly toxin A) result in the activation of three transcription factors (Figure 1). Nuclear factor-kappa B (NF-κB) is involved in chemokine production, and also plays a role in colonocyte apoptosis<sup>[23,28]</sup>. Activator protein-1 (AP-1) also plays a role in interleukin (IL)-8 production, in response to stimulation of colonocytes with toxin A<sup>[24]</sup>. Cyclic-AMP response binding protein (CREB) is critical for the production of Prostaglandin E2<sup>[23]</sup>. This prostaglandin plays an important role in the fluid secretion/diarrhea associated with CDAD (Figure 1). As shown in the figure, there is also cross talk between the various pathways. For example, prostaglandin E2 can stimulate Fas ligand expression and apoptosis in colonic epithelial cells<sup>[28,29]</sup>.

Specific points of intervention, resulting in immunomodulation by certain probiotics, are shown in Figure 1<sup>[21,30-39]</sup>. The non-pathogenic yeast probiotic, *Saccharomyces boulardii* has the most well described immunomodulatory actions<sup>[21]</sup>. *Saccharomyces boulardii* can inhibit toxin-A receptor binding to target cells, by release of a protease that digests both the exotoxin and its receptor binding sites<sup>[21,31,32]</sup>. Indirectly, this prevents the downstream activation of relevant MAP kinases, as well as transcription factor activation by toxin A (Figure 1). The same group of investigators showed that *Saccharomyces boulardii* supernatants could inhibit (*in vitro* or *in vivo*) toxin A-induced MAP kinase (ERK 1/2) activation, IL-8 production, fluid secretion, and intestinal inflammation<sup>[21,30]</sup>. *Saccharomyces boulardii* also reportedly inhibits activation of the key transcription factor NF-κB<sup>[21]</sup>.

*Bacillus coagulans* GBI-30, 6086 (Bc) is a novel probi-

otic, which can attenuate chemokine release both *in vitro* and *in vivo*<sup>[34,35]</sup>. Correspondingly, this probiotic reduced neutrophil influx and colonic inflammation associated with CDAD in mice<sup>[34,35]</sup>. Of note, *Bacillus coagulans* GBI-30 reduced the expression (by immunohistochemistry) of COX-2 in the colons of mice with CDAD (Figure 1)<sup>[34,35]</sup>.

*Lactobacillus acidophilus* (*L. acidophilus*) substantially improved cyclosporine-induced *C. difficile* infection in mice<sup>[36,39]</sup>. Various parameters of infectious colitis were attenuate by probiotic treatment, including myeloperoxidase and histopathology, as well as titers of toxins A and B derived from the cecal contents of mice (Figure 1)<sup>[36,39]</sup>. Another lactobacillus species, *Lactobacillus rhamnosus* (*L. rhamnosus*) improved *C. difficile*-induced inflammation and damage to the ileum of hamsters, with less evidence of diarrhea<sup>[37]</sup>.

Martins *et al*<sup>[38]</sup> developed a screening paradigm for yeast probiotic strains, based upon protection against enteric pathogens including *C. difficile*. These investigators found that *Saccharomyces cerevisiae*, strain 905 protected the cecum of gnotobiotic mice from *C. difficile*-induced pathological changes in the cecum (Figure 1)<sup>[38]</sup>.

## PROBIOTICS AND PRECLINICAL MODELS OF CDAD

Table 1 shows a list of some probiotics that were tested in pre-clinical models of *C. difficile*-induced colitis. Early studies, which were conducted approximately 30 years ago, showed that *Saccharomyces boulardii* could prevent Clindamycin (and by association *C. difficile*)-induced mortality in hamsters, with improvement in the histological appearance of the intestine in these animals<sup>[40,41]</sup>. In the same time period, Corthier *et al*<sup>[42]</sup> found that *Saccharomyces boulardii* could limit mortality in gnotobiotic mice that were infected with *C. difficile*. Of note, this probiotic also modulated fecal cytotoxin production (Figure 1)<sup>[42]</sup>.

More recent studies, showed that *Saccharomyces cerevisiae* strain 905 and two lactobacillus strains (*L. rhamnosus* and *acidophilus*) were effective against CDAD in rodents (Figure 1 and Table 1)<sup>[36-39]</sup>. My laboratory found that the novel probiotic strain *Bacillus coagulans* GBI-30, 6086 could improve both the initial phase of colitis in mice following *C. difficile* infection, as well the recurrence of CDAD following vancomycin withdrawal<sup>[34,35]</sup>. This probiotic most profoundly affected the stool consistency in these mice (Figure 1)<sup>[34,35]</sup>.

## CLINICAL USE OF PROBIOTICS FOR CDAD

Since 2011, several comprehensive reviews have been published regarding the use of probiotics for CDAD. Specific details from these reviews can be found in the relevant literature<sup>[19,43-46]</sup>. Floch *et al*<sup>[43]</sup> gave probiotics a B/C recommendation for both the prevention of CDAD,

**Table 1** Effects of probiotics in animal models of *Clostridium difficile*-induced colitis

Probiotic	Species	Efficacy	Reference
<i>Saccharomyces boulardii</i>	Hamster	Yes	[40,41]
<i>Saccharomyces boulardii</i>	Mice	Yes	[42]
<i>Saccharomyces cerevisiae</i> 905	Mice	Yes	[38]
<i>Lactobacillus rhamnosus</i>	Hamster	Yes	[37]
<i>Lactobacillus acidophilus</i>	Mice	Yes	[36,39]
<i>Bacillus coagulans</i> GBI-30, 6086	Mice	Yes	[34,35]

and also the prevention of recurrent CDAD. Their somewhat arbitrary rating system suggested some positive clinical studies, but also the presence of some negative studies (B rating), or inadequate clinical experience (C rating). In their evaluations, the investigators focused mainly on studies involving *Saccharomyces boulardii* and *Lactobacillus GG*<sup>[43]</sup>. In another review, Hickson<sup>[44]</sup> suggested that the evidence supporting the use of probiotics for CDAD is overall equivocal. Musgrave *et al*<sup>[45]</sup> reported that probiotics could be considered for the prevention of *C. difficile* infection, or as an adjunctive therapy in otherwise healthy (non-immunocompromised) patients. Davidson *et al*<sup>[46]</sup> suggested the possible co-administration of probiotics for prevention of CDAD in patients at increased risk for developing disease. However, they did not recommend adjunctive probiotics for the routine treatment of CDAD<sup>[46]</sup>. The most recent Cochrane review (from 2008) on probiotics for CDAD in adults concluded that insufficient evidence existed to recommend probiotics as an adjunct to antibiotic therapy for *C. difficile* colitis<sup>[47]</sup>. Moreover, reportedly there was no evidence to support the use of probiotics alone for *C. difficile* colitis<sup>[47]</sup>.

## FUTURE USE OF PROBIOTICS FOR CDAD

In a review article published in 2009, Imhoff *et al*<sup>[13]</sup> asked this question: Is there a future for probiotics in preventing *clostridium difficile*-associated disease and treatment of recurrent episodes? This statement remains a pertinent question in 2013. Recently, there has been a renewed interest in fecal microbiota transplantation therapy for recurrent CDAD, and new studies suggest efficacy for this indication<sup>[48-52]</sup>. Therefore, what about the future use of probiotics in CDAD beyond 2013? Because CDAD is a condition associated with disrupted endogenous gut flora, it is logical to employ treatment strategies that can reconstitute/restore the physiological intestinal flora. In a broad sense, both probiotics and fecal microbiota transplantation therapy attempt to accomplish this restoration of physiological bacterial species, but by different administration methods<sup>[13,53,54]</sup>. Certainly, fecal transplantation has yielded some interesting efficacy results<sup>[48-54]</sup>. However, it typically requires an invasive procedure (*e.g.*, colonoscopy), as well as an overall technique that is still aesthetically displeasing to some patients<sup>[48-54]</sup>. In contrast, probiotics can be easily ingested, but are often not optimally formulated to survive transit through the GI tract

for colonization in the colon<sup>[13,19]</sup>. Moreover, probiotics have demonstrated questionable efficacy for CDAD<sup>[43-47]</sup>. A recent publication further compares the pros and cons of probiotics versus fecal transplantation for intestinal diseases<sup>[55]</sup>.

With respect to the future of probiotics for CDAD, Hell *et al*<sup>[19]</sup> have provided some good insights, as well as interesting initial clinical data. They postulated that a multi-strain probiotic, resembling a healthy human microbiota, would be most effective for treating CDAD<sup>[19]</sup>. Therefore, these investigators developed a probiotic mixture (Ecologic<sup>®</sup> AAD) comprised of several *Bifidobacterium* and *Lactobacillus* strains, as well as *Enterococcus faecium*<sup>[19]</sup>. In a small series of 10 patients (five with recurrent disease) excellent results were obtained in all evaluable patients with *C. difficile* infection, following combined treatment with the multi-strain probiotic plus vancomycin<sup>[19]</sup>. This type of therapeutic paradigm seems to represent a logical future scientific approach for probiotic treatment in CDAD. Another probiotic preparation that could be tested for CDAD is VSL#3. This probiotic mixture contains 4 *Lactobacillus* strains, three strains of bifidobacteria and *Streptococcus salivarius*<sup>[56,57]</sup>. VSL#3 has been tested previously in both IBD and pouchitis patient populations, with some evidence of efficacy<sup>[56,57]</sup>. Moreover, the pertinent mechanism(s) of action for VSL#3 suggest that it would represent a rational pharmacological approach for CDAD<sup>[14,58]</sup>. Finally, it may be possible to utilize known mechanism of action diagrams, like in Figure 1, to create novel probiotic mixtures that could potentially be effective for CDAD.

While perhaps focusing on multi-strain probiotics, newer single strain probiotics of potential interest for CDAD could include *Bifidobacterium animalis* AHCT<sup>[59-61]</sup>. This probiotic can inhibit NF- $\kappa$ B activation, reduce *C. difficile* levels in the canine colon, and resolve idiopathic diarrhea in dogs. The pharmacological profile of *Bifidobacterium animalis* AHCT suggests that it could be an interesting candidate for further testing related to CDAD. Another single strain probiotic of interest is *Clostridium butyricum* MIYARI 588, which is being used for the prevention of CDAD in Japan<sup>[62]</sup>.

## REFERENCES

- 1 Johnson S, Gerding DN. Clostridium difficile-associated diarrhea. *Clin Infect Dis* 1998; **26**: 1027-1034; quiz 1035-1036 [PMID: 9597221 DOI: 10.1086/520276]
- 2 Kelly CP, Pothoulakis C, LaMont JT. Clostridium difficile colitis. *N Engl J Med* 1994; **330**: 257-262 [PMID: 8043060 DOI: 10.1056/NEJM199401273300406]
- 3 Pothoulakis C, Lamont JT. Microbes and microbial toxins: paradigms for microbial-mucosal interactions II. The integrated response of the intestine to Clostridium difficile toxins. *Am J Physiol Gastrointest Liver Physiol* 2001; **280**: G178-G183 [PMID: 11208538]
- 4 Bartlett JG. Treatment of antibiotic-associated pseudomembranous colitis. *Rev Infect Dis* 1984; **6** Suppl 1: S235-S241 [PMID: 6718937 DOI: 10.1093/clinids/6.Supplement\_1.S235]
- 5 Leffler DA, Lamont JT. Treatment of Clostridium difficile-associated disease. *Gastroenterology* 2009; **136**: 1899-1912 [PMID: 19457418 DOI: 10.1053/j.gastro.2008.12.070]
- 6 Martinez FJ, Leffler DA, Kelly CP. Clostridium difficile outbreaks: prevention and treatment strategies. *Risk Manag Healthc Policy* 2012; **5**: 55-64 [PMID: 22826646 DOI: 10.2147/RMHP.S13053]
- 7 Surawicz CM, McFarland LV, Elmer G, Chinn J. Treatment of recurrent Clostridium difficile colitis with vancomycin and Saccharomyces boulardii. *Am J Gastroenterol* 1989; **84**: 1285-1287 [PMID: 2679049]
- 8 McFarland LV, Surawicz CM, Greenberg RN, Fekety R, Elmer GW, Moyer KA, Melcher SA, Bowen KE, Cox JL, Noorani Z. A randomized placebo-controlled trial of Saccharomyces boulardii in combination with standard antibiotics for Clostridium difficile disease. *JAMA* 1994; **271**: 1913-1918 [PMID: 8201735 DOI: 10.1001/jama.1994.03510480037031]
- 9 Surawicz CM, McFarland LV, Greenberg RN, Rubin M, Fekety R, Mulligan ME, Garcia RJ, Brandmarker S, Bowen K, Borjal D, Elmer GW. The search for a better treatment for recurrent Clostridium difficile disease: use of high-dose vancomycin combined with Saccharomyces boulardii. *Clin Infect Dis* 2000; **31**: 1012-1017 [PMID: 11049785 DOI: 10.1086/318130]
- 10 Kimmey MB, Elmer GW, Surawicz CM, McFarland LV. Prevention of further recurrences of Clostridium difficile colitis with Saccharomyces boulardii. *Dig Dis Sci* 1990; **35**: 897-901 [PMID: 2364845 DOI: 10.1007/BF01536805]
- 11 Tung JM, Dolovich LR, Lee CH. Prevention of Clostridium difficile infection with Saccharomyces boulardii: a systematic review. *Can J Gastroenterol* 2009; **23**: 817-821 [PMID: 20011734]
- 12 Johnston BC, Ma SS, Goldenberg JZ, Thorlund K, Vandvik PO, Loeb M, Guyatt GH. Probiotics for the prevention of Clostridium difficile-associated diarrhea: a systematic review and meta-analysis. *Ann Intern Med* 2012; **157**: 878-888 [PMID: 23362517 DOI: 10.7326/0003-4819-157-12-201212180-00563]
- 13 Imhoff A, Karpa K. Is there a future for probiotics in preventing Clostridium difficile-associated disease and treatment of recurrent episodes? *Nutr Clin Pract* 2009; **24**: 15-32 [PMID: 19244145 DOI: 10.1177/0884533608329232]
- 14 Petrof EO, Kojima K, Ropeleski MJ, Musch MW, Tao Y, De Simone C, Chang EB. Probiotics inhibit nuclear factor-kappaB and induce heat shock proteins in colonic epithelial cells through proteasome inhibition. *Gastroenterology* 2004; **127**: 1474-1487 [PMID: 15521016 DOI: 10.1053/j.gastro.2004.09.001]
- 15 Fedorak RN, Madsen KL. Probiotics and prebiotics in gastrointestinal disorders. *Curr Opin Gastroenterol* 2004; **20**: 146-155 [PMID: 15703637 DOI: 10.1097/00054725-200405000-00018]
- 16 Shanahan F. Physiological basis for novel drug therapies used to treat the inflammatory bowel diseases I. Pathophysiological basis and prospects for probiotic therapy in inflammatory bowel disease. *Am J Physiol Gastrointest Liver Physiol* 2005; **288**: G417-G421 [PMID: 15701620]
- 17 Fitzpatrick LR, Small J, Hoerr RA, Bostwick EF, Maines L, Koltun WA. In vitro and in vivo effects of the probiotic Escherichia coli strain M-17: immunomodulation and attenuation of murine colitis. *Br J Nutr* 2008; **100**: 530-541 [PMID: 18279557 DOI: 10.1017/S0007114508930373]
- 18 Parkes GC, Sanderson JD, Whelan K. The mechanisms and efficacy of probiotics in the prevention of Clostridium difficile-associated diarrhoea. *Lancet Infect Dis* 2009; **9**: 237-244 [PMID: 19324296 DOI: 10.1016/S1473-3099(09)70059-3]
- 19 Hell M, Bernhofer C, Stalzer P, Kern JM, Claassen E. Probiotics in Clostridium difficile infection: reviewing the need for a multistrain probiotic. *Benef Microbes* 2013; **4**: 39-51 [PMID: 23434948 DOI: 10.3920/BM2012.0049]
- 20 Ng SC, Hart AL, Kamm MA, Stagg AJ, Knight SC. Mechanisms of action of probiotics: recent advances. *Inflamm Bowel Dis* 2009; **15**: 300-310 [PMID: 18626975 DOI: 10.1002/ibd.20602]
- 21 Pothoulakis C. Review article: anti-inflammatory mecha-

- nisms of action of *Saccharomyces boulardii*. *Aliment Pharmacol Ther* 2009; **30**: 826-833 [PMID: 19706150 DOI: 10.1111/j.1365-2036.2009.04102.x]
- 22 **Galdeano CM**, de Moreno de LeBlanc A, Vinderola G, Bonet ME, Perdigon G. Proposed model: mechanisms of immunomodulation induced by probiotic bacteria. *Clin Vaccine Immunol* 2007; **14**: 485-492 [PMID: 17360855 DOI: 10.1128/CVI.00406-06]
  - 23 **Kim H**, Rhee SH, Kokkotou E, Na X, Savidge T, Moyer MP, Pothoulakis C, LaMont JT. *Clostridium difficile* toxin A regulates inducible cyclooxygenase-2 and prostaglandin E2 synthesis in colonocytes via reactive oxygen species and activation of p38 MAPK. *J Biol Chem* 2005; **280**: 21237-21245 [PMID: 15767259 DOI: 10.1074/jbc.M413842200]
  - 24 **Lee JY**, Park HR, Oh YK, Kim YJ, Youn J, Han JS, Kim JM. Effects of transcription factor activator protein-1 on interleukin-8 expression and enteritis in response to *Clostridium difficile* toxin A. *J Mol Med (Berl)* 2007; **85**: 1393-1404 [PMID: 17639289]
  - 25 **Lica M**, Schulz F, Schelle I, May M, Just I, Genth H. Difference in the biological effects of *Clostridium difficile* toxin B in proliferating and non-proliferating cells. *Naunyn-Schmiedeberg's Arch Pharmacol* 2011; **383**: 275-283 [PMID: 21212934 DOI: 10.1007/s00210-010-0595-5]
  - 26 **Chen X**, Katchar K, Goldsmith JD, Nanthakumar N, Cheknis A, Gerding DN, Kelly CP. A mouse model of *Clostridium difficile*-associated disease. *Gastroenterology* 2008; **135**: 1984-1992 [PMID: 18848941 DOI: 10.1053/j.gastro.2008.09.002]
  - 27 **Alcantara C**, Stenson WF, Steiner TS, Guerrant RL. Role of inducible cyclooxygenase and prostaglandins in *Clostridium difficile* toxin A-induced secretion and inflammation in an animal model. *J Infect Dis* 2001; **184**: 648-652 [PMID: 11474431 DOI: 10.1086/322799]
  - 28 **Kim H**, Rhee SH, Pothoulakis C, Lamont JT. Inflammation and apoptosis in *Clostridium difficile* enteritis is mediated by PGE2 up-regulation of Fas ligand. *Gastroenterology* 2007; **133**: 875-886 [PMID: 17854595 DOI: 10.1053/j.gastro.2007.06.063]
  - 29 **O'Callaghan G**, Kelly J, Shanahan F, Houston A. Prostaglandin E2 stimulates Fas ligand expression via the EP1 receptor in colon cancer cells. *Br J Cancer* 2008; **99**: 502-512 [PMID: 18648368 DOI: 10.1038/sj.bjc.6604490]
  - 30 **Chen X**, Kokkotou EG, Mustafa N, Bhaskar KR, Sougioultzis S, O'Brien M, Pothoulakis C, Kelly CP. *Saccharomyces boulardii* inhibits ERK1/2 mitogen-activated protein kinase activation both in vitro and in vivo and protects against *Clostridium difficile* toxin A-induced enteritis. *J Biol Chem* 2006; **281**: 24449-24454 [PMID: 16816386]
  - 31 **Pothoulakis C**, Kelly CP, Joshi MA, Gao N, O'Keane CJ, Castagliuolo I, Lamont JT. *Saccharomyces boulardii* inhibits *Clostridium difficile* toxin A binding and enterotoxicity in rat ileum. *Gastroenterology* 1993; **104**: 1108-1115 [PMID: 8462799]
  - 32 **Castagliuolo I**, LaMont JT, Nikulasson ST, Pothoulakis C. *Saccharomyces boulardii* protease inhibits *Clostridium difficile* toxin A effects in the rat ileum. *Infect Immun* 1996; **64**: 5225-5232 [PMID: 8945570]
  - 33 **Jensen GS**, Benson KF, Carter SG, Endres JR. GanedenBC30 cell wall and metabolites: anti-inflammatory and immune modulating effects in vitro. *BMC Immunol* 2010; **11**: 15 [PMID: 20331905 DOI: 10.1186/1471-2172-11-15]
  - 34 **Fitzpatrick LR**, Small JS, Greene WH, Karpa KD, Keller D. *Bacillus Coagulans* GBI-30 (BC30) improves indices of *Clostridium difficile*-induced colitis in mice. *Gut Pathog* 2011; **3**: 16 [PMID: 22014083 DOI: 10.1186/1757-4749-3-16]
  - 35 **Fitzpatrick LR**, Small JS, Greene WH, Karpa KD, Farmer S, Keller D. *Bacillus coagulans* GBI-30, 6086 limits the recurrence of *Clostridium difficile*-induced colitis following vancomycin withdrawal in mice. *Gut Pathog* 2012; **4**: 13 [PMID: 23088680 DOI: 10.1186/1757-4749-4-13]
  - 36 **Kaur S**, Vaishnavi C, Ray P, Kochhar R, Prasad KK. Effect of biotherapeutics on cyclosporin-induced *Clostridium difficile* infection in mice. *J Gastroenterol Hepatol* 2010; **25**: 832-838 [PMID: 20074161 DOI: 10.1111/j.1440-1746.2009.06135.x]
  - 37 **Naaber P**, Mikelsaar RH, Salminen S, Mikelsaar M. Bacterial translocation, intestinal microflora and morphological changes of intestinal mucosa in experimental models of *Clostridium difficile* infection. *J Med Microbiol* 1998; **47**: 591-598 [PMID: 9839563 DOI: 10.1099/00222615-47-7-591]
  - 38 **Martins FS**, Nardi RM, Arantes RM, Rosa CA, Neves MJ, Nicoli JR. Screening of yeasts as probiotic based on capacities to colonize the gastrointestinal tract and to protect against enteropathogen challenge in mice. *J Gen Appl Microbiol* 2005; **51**: 83-92 [PMID: 15942869 DOI: 10.2323/jgam.51.83]
  - 39 **Kaur S**, Vaishnavi C, Prasad KK, Ray P, Kochhar R. Effect of *Lactobacillus acidophilus* & amp; epidermal growth factor on experimentally induced *Clostridium difficile* infection. *Indian J Med Res* 2011; **133**: 434-441 [PMID: 21537099]
  - 40 **Massot J**, Sanchez O, Couchy R, Astoin J, Parodi AL. Bacterio-pharmacological activity of *Saccharomyces boulardii* in clindamycin-induced colitis in the hamster. *Arzneimittelforschung* 1984; **34**: 794-797 [PMID: 6541916]
  - 41 **Toothaker RD**, Elmer GW. Prevention of clindamycin-induced mortality in hamsters by *Saccharomyces boulardii*. *Antimicrob Agents Chemother* 1984; **26**: 552-556 [PMID: 6517545 DOI: 10.1128/AAC.26.4.552]
  - 42 **Corthier G**, Dubos F, Ducluzeau R. Prevention of *Clostridium difficile* induced mortality in gnotobiotic mice by *Saccharomyces boulardii*. *Can J Microbiol* 1986; **32**: 894-896 [PMID: 3815159 DOI: 10.1139/m86-164]
  - 43 **Floch MH**, Walker WA, Madsen K, Sanders ME, Macfarlane GT, Flint HJ, Dieleman LA, Ringel Y, Guandalini S, Kelly CP, Brandt LJ. Recommendations for probiotic use-2011 update. *J Clin Gastroenterol* 2011; **45** Suppl: S168-S171 [PMID: 21992958 DOI: 10.1097/MCG.0b013e318230928b]
  - 44 **Hickson M**. Probiotics in the prevention of antibiotic-associated diarrhoea and *Clostridium difficile* infection. *Therap Adv Gastroenterol* 2011; **4**: 185-197 [PMID: 21694803 DOI: 10.1177/1756283X11399115]
  - 45 **Musgrave CR**, Bookstaver PB, Sutton SS, Miller AD. Use of alternative or adjuvant pharmacologic treatment strategies in the prevention and treatment of *Clostridium difficile* infection. *Int J Infect Dis* 2011; **15**: e438-e448 [PMID: 21596604 DOI: 10.1016/j.ijid.2011.03.017]
  - 46 **Davidson LE**, Hibber PL. *Clostridium difficile* and probiotics. Up to date 2013, published online, Wolters Kluwer, Alphen aan den Rijn, the Netherlands. Available from: URL: <http://www.uptodate.com/contents/clostridium-difficile-and-probiotics>
  - 47 **Pillai A**, Nelson R. Probiotics for treatment of *Clostridium difficile*-associated colitis in adults. *Cochrane Database Syst Rev* 2008; **23**: CD004611 [PMID: 18254055]
  - 48 **Kelly CP**. Fecal microbiota transplantation--an old therapy comes of age. *N Engl J Med* 2013; **368**: 474-475 [PMID: 23323865 DOI: 10.1056/NEJMe1214816]
  - 49 **Suwantarat N**, Bobak DA. Fecal Bacteriotherapy for Recurrent *Clostridium difficile* Infection: What's Old Is New Again? *Curr Infect Dis Rep* 2013; **15**: 101-103 [PMID: 23549617]
  - 50 **Zucca M**, Scutera S, Savoia D. Novel avenues for *Clostridium difficile* infection drug discovery. *Expert Opin Drug Discov* 2013; **8**: 459-477 [PMID: 23427910 DOI: 10.1517/17460441.2013.770466]
  - 51 **Agito MD**, Atreja A, Rizk MK. Fecal microbiota transplantation for recurrent *C difficile* infection: ready for prime time? *Cleve Clin J Med* 2013; **80**: 101-108 [PMID: 23376915 DOI: 10.3949/ccjm.80a.12110]
  - 52 **Brandt LJ**, Aroniadis OC, Mellow M, Kanatzar A, Kelly C, Park T, Stollman N, Rohlke F, Surawicz C. Long-term follow-up of colonoscopic fecal microbiota transplant for recurrent *Clostridium difficile* infection. *Am J Gastroenterol* 2012; **107**:

- 1079-1087 [PMID: 22450732 DOI: 10.1038/ajg.2012.60]
- 53 **Kelly CR**, de Leon L, Jasutkar N. Fecal microbiota transplantation for relapsing *Clostridium difficile* infection in 26 patients: methodology and results. *J Clin Gastroenterol* 2012; **46**: 145-149 [PMID: 22157239 DOI: 10.1097/MCG.0b013e318234570b]
- 54 **Kleger A**, Schnell J, Essig A, Wagner M, Bommer M, Seufferlein T, Härter G. Fecal transplant in refractory *Clostridium difficile* colitis. *Dtsch Arztebl Int* 2013; **110**: 108-115 [PMID: 23468820]
- 55 **Kruis W**. Specific probiotics or 'fecal transplantation'. *Dig Dis* 2012; **30** Suppl 3: 81-84 [PMID: 23295696 DOI: 10.1159/000342611]
- 56 **Chapman TM**, Plosker GL, Figgitt DP. VSL#3 probiotic mixture: a review of its use in chronic inflammatory bowel diseases. *Drugs* 2006; **66**: 1371-1387 [PMID: 16903771 DOI: 10.2165/00003495-200666100-00006]
- 57 **Chapman TM**, Plosker GL, Figgitt DP. Spotlight on VSL#3 probiotic mixture in chronic inflammatory bowel diseases. *BioDrugs* 2007; **21**: 61-63 [PMID: 17263590 DOI: 10.2165/00063030-200721010-00007]
- 58 **Petrof EO**. Probiotics and Gastrointestinal Disease: Clinical Evidence and Basic Science. *Antiinflamm Antiallergy Agents Med Chem* 2009; **8**: 260-269 [PMID: 20890386]
- 59 **O'Mahony D**, Murphy S, Boileau T, Park J, O'Brien F, Groeger D, Konieczna P, Ziegler M, Scully P, Shanahan F, Kiely B, O'Mahony L. *Bifidobacterium animalis* AHC7 protects against pathogen-induced NF- $\kappa$ B activation in vivo. *BMC Immunol* 2010; **11**: 63 [PMID: 21176205 DOI: 10.1186/1471-2172-11-63]
- 60 **Kelley RL**, Park JS, O'Mahony L, Minikhiem D, Fix A. Safety and tolerance of dietary supplementation with a canine-derived probiotic (*Bifidobacterium animalis* strain AHC7) fed to growing dogs. *Vet Ther* 2010; **11**: E1-14 [PMID: 20960415]
- 61 **O'Mahony D**, Murphy KB, MacSharry J, Boileau T, Sunvold G, Reinhart G, Kiely B, Shanahan F, O'Mahony L. Portrait of a canine probiotic *Bifidobacterium*--from gut to gut. *Vet Microbiol* 2009; **139**: 106-112 [PMID: 19523775 DOI: 10.1016/j.vetmic.2009.05.002]
- 62 **Woo TD**, Oka K, Takahashi M, Hojo F, Osaki T, Hanawa T, Kurata S, Yonezawa H, Kamiya S. Inhibition of the cytotoxic effect of *Clostridium difficile* in vitro by *Clostridium butyricum* MIYAIRI 588 strain. *J Med Microbiol* 2011; **60**: 1617-1625 [PMID: 21700738 DOI: 10.1099/jmm.0.033423-0]

**P- Reviewers** Hokama A, Schwarz SM **S- Editor** Gou SX  
**L- Editor** A **E- Editor** Li JY



## Altered molecular pattern of mucosal healing in Crohn's disease fibrotic stenosis

Enzo Ierardi, Floriana Giorgio, Domenico Piscitelli, Mariabeatrice Principi, Santina Cantatore, Maria Grazia Fiore, Roberta Rossi, Michele Barone, Alfredo Di Leo, Carmine Panella

Enzo Ierardi, Floriana Giorgio, Santina Cantatore, Michele Barone, Carmine Panella, Gastroenterology, Department of Medical and Surgical Sciences, University of Foggia, 71100 Foggia, Italy

Domenico Piscitelli, Mariabeatrice Principi, Maria Grazia Fiore, Roberta Rossi, Alfredo Di Leo, Gastroenterology, Department of Emergency and Organ Transplantation, University of Bari, 70124 Bari, Italy

**Author contributions:** Ierardi E, Barone M, Di Leo A and Panella C designed the study, revised the manuscript and approved the final version; Giorgio F, Piscitelli D, Principi M, Fiore MG and Rossi R collected the data; Cantatore S performed immunohistochemistry.

**Correspondence to:** Enzo Ierardi, Professor, Gastroenterology, Department of Medical and Surgical Sciences, University of Foggia, AOU Ospedali Riuniti, Viale Pinto, 71100 Foggia, Italy. [enzo.ierardi@fastwebnet.it](mailto:enzo.ierardi@fastwebnet.it)  
Telephone: +39-8-81736204 Fax: +39-8-81733848  
Received: April 6, 2013 Revised: June 17, 2013  
Accepted: July 18, 2013  
Published online: August 15, 2013

### Abstract

**AIM:** To investigate tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), syndecan 1 and basic fibroblast growth factor (bFGF) balance in Crohn's disease (CD) strictures.

**METHODS:** Our study was performed on 24 surgical specimens of CD fibrotic stenosis. Ten histological normal surgical samples were retrieved for both the large and small bowel from patients with benign conditions and healthy tissue represented control collection. Sex and age in controls did not differ from CD group. Three endoscopic biopsy specimens taken after informed consent in subjects with normal colon were also used as negative controls. TNF- $\alpha$ , syndecan 1 and bFGF were detected by both reverse transcriptase reverse transcriptase polymerase chain reaction after mRNA extraction (results expressed as fold-change) and immunohistochemistry.

**RESULTS:** TNF- $\alpha$  did not show any significant difference between CD and control specimens ( $1.54 \pm 1.19$ ;  $P > 0.05$ ). Very high levels of bFGF were observed in CD ( $11.76 \pm 4.65$ ;  $P < 0.001$ ) unlike syndecan 1 which showed a moderate increase ( $5.53 \pm 2.18$ ;  $P < 0.005$ ). analysis of variance (ANOVA) plus Student-Neumann-Keuls showed: bFGF > syndecan 1 > TNF- $\alpha$  = control. Immunoreactivity for bFGF was observed in epithelial, stromal, endothelial cells and even in the muscular layer, whilst in normal tissue it was almost unexpressed. Syndecan 1 and TNF- $\alpha$  staining was confined to mucosal epithelial and stromal cells, while in controls syndecan 1 was found in its normal site, *i.e.*, basolateral area of the crypts and TNF- $\alpha$  very poorly expressed.

**CONCLUSION:** Fibrotic stenosis of CD may be the final result of an irreversible transformation of different cells into fibrogenic phenotype no longer inhibited by post-transcriptional regulation.

© 2013 Baishideng. All rights reserved.

**Key words:** Crohn's disease; Fibrotic stenosis; Tumor necrosis factor- $\alpha$ ; Syndecan 1; Basic fibroblast growth factor

**Core tip:** The present manuscript reports a study of molecular pattern in the course of stenotic complication of Crohn's disease. We have studied the interaction among the main cytokine [tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )], an adhesion molecule (syndecan 1) and a growth factor basic fibroblast growth factor which are strictly involved in damage repair and mucosal healing, as showed in a previous study. In this study we demonstrated that a deep dysregulation of interaction of these three factors may support stenotic fibrosis in Crohn's disease and suggest that this condition needs to be investigated before a biological treatment since TNF- $\alpha$  lack downregulation could further stimulate fibrogenesis.

Ierardi E, Giorgio F, Piscitelli D, Principi M, Cantatore S, Fiore MG, Rossi R, Barone M, Di Leo A, Panella C. Altered molecular pattern of mucosal healing in Crohn's disease fibrotic stenosis. *World J Gastrointest Pathophysiol* 2013; 4(3): 53-58 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i3/53.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i3.53>

## INTRODUCTION

Considerable downregulation of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) may occur after infliximab therapy in inflammatory bowel diseases (IBD) in those patients who achieve disorder remission (responders: about the 70%)<sup>[1]</sup>. Such downregulation may be associated with a dramatic regression of endoscopic lesions with a feature suitable of "mucosal healing"<sup>[2]</sup>.

TNF- $\alpha$  downregulation presumably interacts with the adhesion molecules (syndecan 1)<sup>[3]</sup> and growth factors [basic fibroblast growth factor (bFGF)]<sup>[4]</sup> in the process of mucosal repair. Indeed, syndecan 1 is located in the basolateral region of the columnar epithelium and plays a relevant role in the course of IBD damage reversal<sup>[5-7]</sup>. This role seems to be due to its ability to change bFGF morphology and modulate the structure of its receptors, allowing its binding to repair dedicated epithelial and stromal cells located at the margins of ulcerative lesions<sup>[8-10]</sup>. The relevance of syndecan 1 also may be due to its aptitude to inhibit bFGF proteolysis, which restricts tissue repair, thus maintaining mucosal damage<sup>[9]</sup>. In normal tissues bFGF is present in basement membranes and in the subendothelial extracellular matrix of blood vessels. It stays membrane-bound as long as there is no signal peptide<sup>[11]</sup>. In intestinal inflammation both molecules are found in either stromal cells and matrix<sup>[12]</sup> with an increased expression.

In a previous study, we have analyzed the mucosal levels of TNF- $\alpha$  as well as bFGF/syndecan 1 link in a selected group of ulcerative colitis (UC) responder patients designed for an observational 6-mo "post-hoc" investigation<sup>[13]</sup>. Our results showed that the cytokine decrease induced by infliximab treatment is accompanied by a simultaneous decrease in both adhesion molecule and growth factor in the presence of mucosal healing. Additionally, we have found that syndecan 1 overexpression in stromal cells and apical epithelium is reversed after infliximab successful therapy on mucosal lesions.

Therefore, infliximab therapy downregulation of bFGF/syndecan 1 link could represent a possible molecular pathway of mucosal healing in ulcerative colitis despite our results do not clarify completely the reliability of the hypothesis, since the parallel pattern of TNF- $\alpha$ , syndecan 1 and bFGF could only be a simultaneous consequence of the control of inflammation.

Similarly, Verspaget *et al*<sup>[14]</sup> reported that healing of fistulizing/perianal Crohn's disease (CD) seems to be reflected by a decrease in high serum bFGF as well as in immunohistochemical mucosal labeling index. Neverthe-

**Table 1 Characteristics of study population with Crohn's disease undergoing surgery for fibrotic stenosis**

	Crohn's disease (n = 24)
Age (yr)	36 ± 17
Gender (male)	15 (62)
CDAI	197 ± 41
Steroids	7 (29)
AZA/6-MP	7 (29)
Infliximab	5 (20)
ADA	6 (25)
Extraintestinal manifestations	5 (20)
Intestinal fistula	3 (8)
Previous surgical interventions	0 (0)
Disease duration (mo)	59 ± 62

CDAI: Crohn's disease activity index; AZA: Azathioprine; 6-MP: 6-mercaptopurine; ADA: American Diabetes Association.

less, bFGF levels do not correlate with the therapeutic response in active disease, when indicated by Crohn's disease activity index (CDAI) score. Authors explained this result with the inclusion of several subjective parameters in the overall CDAI score, which may mask the real inflammatory bowel picture.

We performed the present study in order to verify the simultaneous configuration of TNF- $\alpha$ , bFGF and syndecan 1 in patients with a severe complication of CD, *i.e.*, intestinal resection for severe fibrotic stenosis. Aim of the work was to investigate whether a dysregulation of their balance observed in mucosal healing could indicate a molecular pattern for stenotic complication.

## MATERIALS AND METHODS

### Patient enrolment

Our study was performed on surgical specimens from 24 patients undergoing intestinal resection for fibrotic stenosis. Baseline demographic and clinical characters are reported in Table 1. Ten histological normal surgical samples were respectively retrieved for both the large and small bowel from patients operated for large adenomas (3 patients), acute diverticular perforation (2 patients), intestinal volvulus (2 patients), internal hernia passing into the bursa omentalis for mesenteric malformation (1 patient), acute mesenteric ischemia (1 patient) and post-surgical small bowel obstruction (1 patient). Strikingly, healthy tissue surrounding diseased one was taken and represented the control collection, which did not differ for sex and age distribution from the CD group. Additionally, 3 endoscopic biopsy specimens taken after informed consent in subjects undergoing colonoscopy for intestinal cancer familiarity were used as negative controls in each assay.

### Molecular analysis

As reported above<sup>[15]</sup>, reverse transcriptase polymerase chain reaction has the ability to reflect the altered pattern of the expression of genes dedicated to the synthesis of a specific molecule and quantify its transcription levels. Therefore, in this study, the technique allowed the

amount of mRNA codifying for the synthesis of TNF- $\alpha$ , syndecan 1 and bFGF to be estimated. The amount was expressed by a numerical value (*i.e.*, the fold change compared with controls)<sup>[16]</sup>. The relative expression of the studied gene levels was calculated using the 2<sup>-CT</sup> method. RNA was extracted from at least five sections of 10  $\mu$ m of paraffin blocks, using the RNeasy paraffin-embedded Formalin-Fixed and Paraffin-Embedded (FFPE) Kit (Qiagen, GmbH, Germany). The choice of this kit was suggested, since it has been specifically designed for the purification of total RNA from (FFPE) tissue sections. Briefly, 10  $\mu$ m thick sections were cut (at least 5) using a specific microtome (Leica Microsystems, Wetzlar, Germany). Five hundred microlitres of xylene were added to the sections to yield a solution that was then vortexed for 10 s and then incubated for 10 min at room temperature (25 °C). The next step was the addition of 500  $\mu$ L of absolute ethanol, the novel solution was again vortexed vigorously for 10 s and centrifuged for 2 min at 11000 *g*. The supernatant was carefully removed by pipetting without disturbing the pellet. Then, any residual xylene/ethanol was carefully removed using a fine pipette tip.

The lid was kept open until air induced the drying of the pellet (about 5 min at room temperature: 25 °C). After deparaffinisation, the method proceeded according to the manufacturer's instructions for RNA extraction. In detail, final mRNA concentrations were estimated by ultraviolet absorbance at 260/280 nm. Aliquots of total mRNA (1 mg) were reverse-transcribed using random hexamers and TaqMan Reverse Transcription Reagents (Applied Biosystems, Foster City, CA) with 3.125 U/mL of MultiScribe Reverse Transcriptase in a final volume of 50  $\mu$ L. A series of six serial dilutions (from 20 to 0.1 ng/mL) of colon tissue DNA (cDNA) was used as a template. Two-step reverse transcription polymerase chain reaction (PCR) was performed using first-strand with a final concentration of 13 TaqMan gene expression assay, that is, the analysed molecules plus reference gene (glyceraldehyde-3-phosphate-dehydrogenase) (Applied Biosystems). The final reaction volume was 25  $\mu$ L, and this was analysed in triplicate (all experiments were repeated twice). A non-template control (Rnase-free water) was included on every plate. Our method was validated further, moreover, by enclosing in each assay fresh samples from three normal patients and frozen at -90 °C until the analysis, as above reported. These samples were treated with the same technique as for paraffin-embedded samples with the exclusion of paraffin removal and rehydration procedures. Specific thermal cycler conditions were employed using a realtime PCR System (Applied Biosystems). A standard curve plus validation experiment were performed for each primer/probe set.

### Immunohistochemistry

Syndecan 1 immunohistochemical staining was additionally performed for each sample from both patients and controls in order to establish and semiquantitatively evaluate its location. Its stromal expression was reflected

by the percentage of positive cells, evaluated in three randomised fields for a total of at least 1000 cells, while its epithelial expression of basolateral and apical amount was reflected by a subjective score already used by our group for other studies<sup>[17]</sup>. Syndecan-1 expression was assessed on paraffin-embedded sections using a monoclonal mouse antibody (Santa Cruz Biotechnology, Santa Cruz, CA) diluted 1:80 in buffer phosphate.

Immunohistochemistry for bFGF was performed as follows. The primary antibody was a polyclonal antibody directed against bFGF (code No. sc-79-G; Santa Cruz Biotechnology Inc., Santa Cruz, CA). This rabbit affinity-purified antibody to bFGF was raised against the amino-terminal domain of human bFGF. Secondary biotinylated antibodies (code No. E 0432), *i.e.*, goat antirabbit immunoglobulin G (IgG), were purchased from Dako, Denmark. The colour reaction was developed by 3-amino-9-ethylcarbazole in acetate buffer containing H<sub>2</sub>O<sub>2</sub>. Specificity of the immunohistochemical staining was assessed by neutralization incubation of the primary antibodies with a 5- to 10-fold (by weight) excess of blocking peptide (code no. sc-79-P) also obtained from Santa Cruz Biotechnology, Inc., and incubation of preimmune serum or buffer instead of the primary antibody, all showing negative staining.

### Statistical analysis

Statistical analysis was performed using analysis of variance (ANOVA) plus the Student-Newman-Keuls test in order to compare numerical values indicating TNF- $\alpha$ , syndecan 1 and bFGF expression either in patients with complicated IBD and controls. Statistical analyses were performed using specific software (Statsoft 6.0 program for Windows 98).

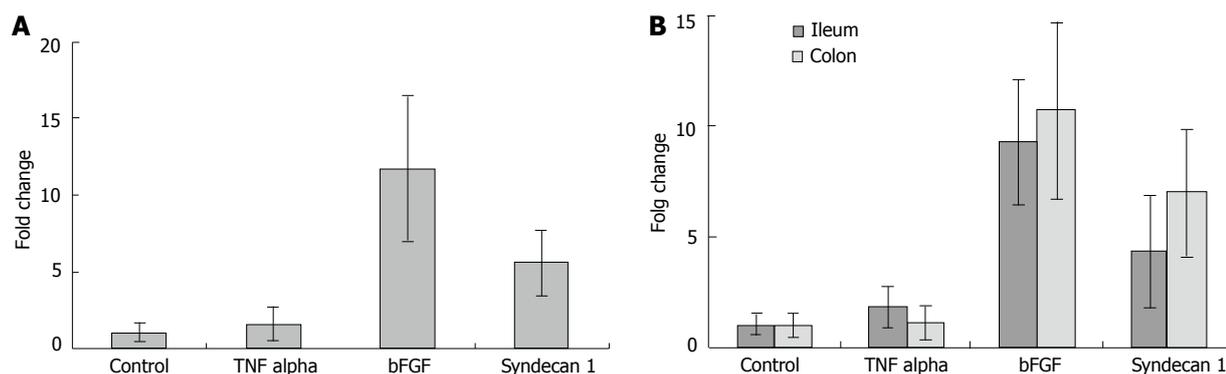
## RESULTS

### Molecular analysis

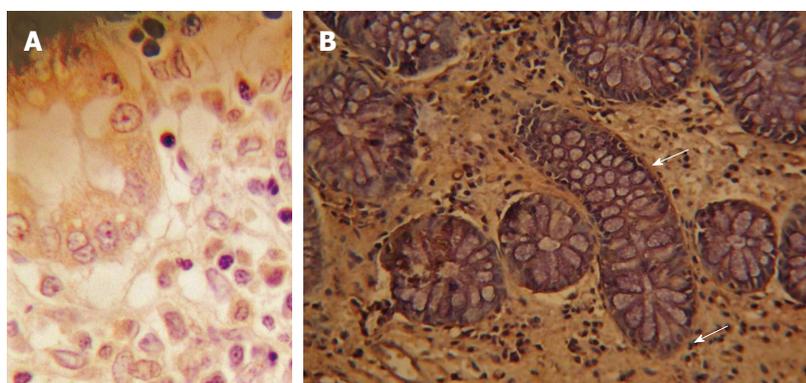
Figure 1A reports the levels of the three investigated molecules in specimens with complicated CD and controls. The levels of TNF- $\alpha$  did not show a significant difference between the two groups (Crohn's value:  $1.54 \pm 1.19$ ;  $P > 0.05$ ). However, very high levels of bFGF were observed in CD ( $11.76 \pm 4.65$ ;  $P < 0.001$ ), while syndecan 1 showed a moderate increase, as characteristic of inflamed tissue ( $5.53 \pm 2.18$ ;  $P < 0.005$ ). ANOVA plus Student-Neumann-Keuls showed: bFGF > syndecan 1 > TNF- $\alpha$  = control. The pattern remained unchanged even when the samples were divided into large and small bowel specimens as demonstrated in Figure 1B.

### Immunohistochemistry

As reported in Figure 2A, immunohistochemistry showed a syndecan 1 pattern reflecting that of inflammation even if confined at mucosal layer (*i.e.*, diffusion to stromal cells and to the apical surface of epithelium). In detail, the labeling index in stromal cells was  $42.57 \pm 9.21$ . In normal tissue no stromal cell was positive for the molecule as



**Figure 1 Levels of the three investigated molecules.** A: Specimens with complicated Crohn's disease and controls. The levels of tumor necrosis factor (TNF)- $\alpha$  do not show a significant difference from that observed in normal tissue ( $1.54 \pm 1.19$ ;  $P > 0.05$ ). High levels of basic fibroblast growth factor (bFGF) are observed in Crohn's disease ( $11.76 \pm 4.65$ ;  $P < 0.001$ ), while syndecan 1 shows a moderate increase ( $5.53 \pm 2.18$ ;  $P < 0.005$ ). Analysis of variance plus Student-Neumann-Keuls showed: bFGF > syndecan 1; B: The same parameters on the bases of their site (small and large bowel). No significant difference is observed in relation to the site of resection.



**Figure 2 Immunohistochemical staining of syndecan 1 in the lamina propria in controls (A) and Crohn's disease (B) stenotic complication.** In normal tissue staining is confined to basolateral area of the crypts (arrow). In Crohn's disease, positive stromal cells and apical epithelium are observed stained brown (diaminobenzidine). Negative cells are counterstained blue (haematoxylin) ( $\times 400$ ).

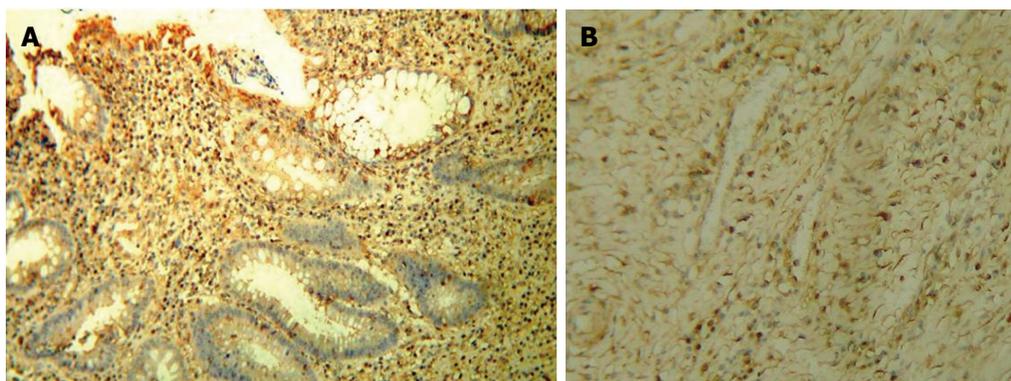
demonstrated in Figure 2B. Additionally, in inflamed mucosa epithelium was diffusely stained, whilst the staining was almost absent in the basolateral crypt area. However, in controls a mild staining confined to basal area of the crypts represented the only expression of this molecule. Immunoreactivity for bFGF was observed in epithelial, stromal and endothelial cells. In areas with signs of severe inflammation, the bFGF reaction in the extracellular matrix was relatively intense (Figure 3A). A positive staining for bFGF was finally observed even in the muscular layer (Figure 3B). In controls, it was almost unexpressed.

## DISCUSSION

In this study we investigated the pattern of TNF- $\alpha$ , syndecan 1 and bFGF in patients with CD complicated by fibrotic stenosis undergoing surgical resection. The basis of our investigation was represented by our previous finding showing that infliximab therapy downregulation of bFGF/syndecan 1 link could represent a possible molecular pathway of mucosal healing in ulcerative colitis<sup>[13]</sup>. Indeed, the link between the adhesion molecule and the growth factor is essential for mucosal repair since syndecan 1 has the ability to change bFGF morphology and modulate the structure of its receptors, allowing its binding to repair dedicated epithelial and stromal cells. Moreover, syndecan 1 has a peculiar aptitude to inhibit bFGF

proteolysis, which restricts tissue repair, thus maintaining mucosal inflammatory damage<sup>[5-7,10]</sup>. We found that the decrease of TNF- $\alpha$  induced by infliximab treatment is accompanied by a decrease in both adhesion molecule and growth factor in the presence of mucosal healing. Nevertheless, our results did not completely clarify whether infliximab therapy downregulation of bFGF/syndecan 1 link could represent a molecular pathway of mucosal healing, since the parallel pattern of TNF- $\alpha$ , syndecan 1 and bFGF could only be a simultaneous consequence of the control of inflammation. Furthermore, we experienced the "timing" of TNF- $\alpha$  decrease and bFGF/syndecan 1 reversal to normal levels and sites in cultured biopsy samples taken from patients with IBD and incubated in a medium containing an amount of infliximab similar to that reached in the serum of treated patients<sup>[18,19]</sup>. After 24 h we assayed TNF- $\alpha$ , syndecan 1 and bFGF in tissue homogenates. The final finding was that TNF- $\alpha$  decreased, while syndecan 1 and bFGF levels were still high when evaluated by both molecular method and immunohistochemistry<sup>[20]</sup>.

In the present study we surprisingly observed that TNF- $\alpha$  mucosal levels were not significantly increased in patients with fibrotic stenosis. A possible explanation of this finding may be that an overgrowth of fibrotic tissue may become evident as a successive step after active inflammation, characterized by TNF- $\alpha$  raise. Therefore, at



**Figure 3** Immunohistochemical staining of basic fibroblast growth factor in the lamina propria (A) and the muscular layer (B). A: Positive cells are stained red (aminoethylcarbazole) and negative ones blue (haematoxylin). A massive staining of stromal cells is observed, but also epithelium is involved by molecule expression ( $\times 100$ ); B: Positive cells are observed in the muscle and endothelium ( $\times 100$ ).

the stage of fibrotic stenosis requiring surgery, inflammatory mucosal changes may be an irrelevant phenomenon in most patients. Nevertheless, we cannot exclude that TNF- $\alpha$  decrease could have been also affected by immunosuppressive therapies assumed by most patients of our series, as shown in Table 1.

Syndecan 1 levels were significantly increased with a pattern similar to what observed in the active phase of inflammatory bowel diseases, *i.e.*, almost absent at the basolateral crypt level and strongly evident in stromal cells and apical surface of epithelium<sup>[5]</sup>. A possible explanation for this pattern is that the molecule location, despite limited to mucosal layer, shows a configuration which reflects the attempt of bFGF modulation for tissue damage reversal. However, this function cannot be completely and effectively articulated for the overexpression of bFGF and especially for its location along the whole intestinal wall, *i.e.*, outside the district in which syndecan 1 could operate.

Indeed, the main result of this study was the bFGF overexpression enclosing all intestinal wall layers. The process is known to involve various cells (epithelial, fibroblasts, myofibroblasts, monocytes, macrophages and neutrophils and endothelial cells)<sup>[21]</sup>. Additionally, its extension reached the smooth muscle layer. The explanation of this aspect may be argued from different likely reasons. First, it is presumable that the low levels of TNF- $\alpha$  may provoke a failure in cytokine induced bFGF proteolysis. A further enlightenment may be that the presence of syndecan 1 is limited to the mucosal layer with a consequent only partial regulation of bFGF binding to specific receptors dedicated to tissue repair. A final explanation could be an irreversible transformation of different type of cells into the fibrogenic phenotype, thus provoking the prevalence of fibrotic on inflammatory stenotic lesions<sup>[22]</sup>. During this process the exceeding of extracellular matrix cannot be inhibited by the regulatory mechanisms of the phenomenon according to what hypothesized by Pucilowska *et al.*<sup>[23,24]</sup>.

A speculative clinical consideration of our results may be the need of an accurate evaluation in the case of stenosis in the course of CD using all diagnostic available

tools (histology, ultrasonography with Doppler evaluation of resistance index, biochemical indices of inflammation) in order to distinguish inflammatory from fibrotic stenosis. This could allow addressing anti-TNF- $\alpha$  therapy only to the first case avoiding that the cytokine decrease may be a factor supporting the fibrotic complication<sup>[25]</sup>.

## COMMENTS

### Background

Downregulation of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) interacts with syndecan 1 and basic fibroblast growth factor (bFGF) in inducing mucosal healing in inflammatory bowel disease. In detail, after TNF- $\alpha$  fall, syndecan 1 and bFGF decrease and disappear from stromal area returning to the original location, *i.e.*, basolateral area of the crypts and basement membranes and in subendothelial extracellular matrix of blood vessels, respectively.

### Research frontiers

The pattern of TNF- $\alpha$ , syndecan 1 and bFGF remains unknown when a complication occurs in inflammatory bowel disease. In detail, the pathway of fibrotic stenosis of Crohn's disease (CD) requiring a surgical operation may be an interesting research hotspot for the possibility of investigating a possible alteration of the molecular configuration occurring in mucosal healing.

### Innovations and breakthroughs

The main result was the bFGF overexpression enclosing all intestinal wall layers until smooth muscle. The explanation of this aspect may be argued from different likely reasons. First, it is presumable that the low levels of TNF- $\alpha$  may provoke a failure in cytokine induced bFGF proteolysis. A further enlightenment may be that the presence of syndecan 1 is limited to the mucosal layer with a consequent only partial regulation of bFGF binding to specific receptors dedicated to tissue repair. A final explanation could be an irreversible transformation of different type of cells into the fibrogenic phenotype, thus provoking the prevalence of fibrotic on inflammatory stenotic lesions. During this process the exceeding of extracellular matrix cannot be inhibited by the regulatory mechanisms

### Applications

A speculative clinical consideration of our results may be the need of an accurate evaluation in the case of stenosis in the course of CD using all diagnostic available tools in order to distinguish inflammatory from fibrotic stenosis. This could allow addressing anti-TNF- $\alpha$  therapy only to the first case avoiding that the cytokine decrease may be a factor supporting the fibrotic complication.

### Terminology

CD is the main chronic inflammatory bowel disease involving the whole intestinal wall; fibrotic stenosis is a stricture requiring a surgical resection in order to avoid or resolve an intestinal occlusion; TNF- $\alpha$  is the main cytokine involved in inflammatory bowel disease related damage; syndecan 1 is an adhesion molecule implicated in tissue repair; bFGF is a strong fibrogenesis stimulator aimed

to restore damaged tissues.

### Peer review

This is a retrospective study in which authors analyze the simultaneous configuration of TNF- $\alpha$ , bFGF and syndecan 1 in patients with a severe complication of CD, *i.e.*, intestinal resection for severe fibrotic stenosis. A deep dysregulation of their balance was observed thus indicating a molecular pattern for stenotic complication.

## REFERENCES

- Hassan C, Ierardi E, Burattini O, De Francesco V, Zullo A, Stoppino G, Panella C, Morini S. Tumour necrosis factor alpha down-regulation parallels inflammatory regression in ulcerative colitis patients treated with infliximab. *Dig Liver Dis* 2007; **39**: 811-817 [PMID: 17652038]
- Chevaux JB, Vavricka SR, Rogler G, Lakatos PL, Schoepfer A, Peyrin-Biroulet L. Mucosal healing with anti-TNF antibodies. *Digestion* 2012; **86** Suppl 1: 16-22 [PMID: 23051722 DOI: 10.1159/000341957]
- Mali M, Jaakkola P, Arvilommi AM, Jalkanen M. Sequence of human syndecan indicates a novel gene family of integral membrane proteoglycans. *J Biol Chem* 1990; **265**: 6884-6889 [PMID: 2324102]
- Kim HS. Assignment1 of the human basic fibroblast growth factor gene FGF2 to chromosome 4 band q26 by radiation hybrid mapping. *Cytogenet Cell Genet* 1998; **83**: 73 [PMID: 9925931 DOI: 10.1159/000015129]
- Day R, Forbes A. Heparin, cell adhesion, and pathogenesis of inflammatory bowel disease. *Lancet* 1999; **354**: 62-65 [PMID: 10406379]
- Day R, Ilyas M, Daszak P, Talbot I, Forbes A. Expression of syndecan-1 in inflammatory bowel disease and a possible mechanism of heparin therapy. *Dig Dis Sci* 1999; **44**: 2508-2515 [PMID: 10630505]
- Saunders S, Jalkanen M, O'Farrell S, Bernfield M. Molecular cloning of syndecan, an integral membrane proteoglycan. *J Cell Biol* 1989; **108**: 1547-1556 [PMID: 2494194]
- Beck PL, Podolsky DK. Growth factors in inflammatory bowel disease. *Inflamm Bowel Dis* 1999; **5**: 44-60 [PMID: 10028449]
- Yoshida S, Ono M, Shono T, Izumi H, Ishibashi T, Suzuki H, Kuwano M. Involvement of interleukin-8, vascular endothelial growth factor, and basic fibroblast growth factor in tumor necrosis factor alpha-dependent angiogenesis. *Mol Cell Biol* 1997; **17**: 4015-4023 [PMID: 9199336]
- Rahmoune H, Chen HL, Gallagher JT, Rudland PS, Fernig DG. Interaction of heparan sulfate from mammary cells with acidic fibroblast growth factor (FGF) and basic FGF. Regulation of the activity of basic FGF by high and low affinity binding sites in heparan sulfate. *J Biol Chem* 1998; **273**: 7303-7310 [PMID: 9516424]
- Ornitz DM, Itoh N. Fibroblast growth factors. *Genome Biol* 2001; **2**: REVIEWS3005 [PMID: 11276432 DOI: 10.1186/gb-2001-2-3-reviews3005]
- Florkiewicz RZ, Shibata F, Barankiewicz T, Baird A, Gonzalez AM, Florkiewicz E, Shah N. Basic fibroblast growth factor gene expression. *Ann N Y Acad Sci* 1991; **638**: 109-126 [PMID: 1785797 DOI: 10.1111/j.1749-6632.1991.tb49022.x]
- Ierardi E, Giorgio F, Zotti M, Rosania R, Principi M, Marangi S, Della Valle N, De Francesco V, Di Leo A, Ingrosso M, Panella C. Infliximab therapy downregulation of basic fibroblast growth factor/syndecan 1 link: a possible molecular pathway of mucosal healing in ulcerative colitis. *J Clin Pathol* 2011; **64**: 968-972 [PMID: 21945924 DOI: 10.1136/jcp.2010.086892]
- Verspaget HW, van der Zon AM, Gao Q, van Hogezaand RA. Infliximab for Crohn's disease. *Aliment Pharmacol Ther* 2001; **15**: 2041-2042 [PMID: 11736736]
- Ierardi E, Giorgio F, Rosania R, Zotti M, Principe S, Della Valle N, De Francesco V, Panella C. Mucosal assessment of tumor necrosis factor alpha levels on paraffined samples: a comparison between immunohistochemistry and real time polymerase chain reaction. *Scand J Gastroenterol* 2010; **45**: 1007-1008 [PMID: 20446801 DOI: 10.3109/00365521.2010.483739]
- Bustin SA. Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. *J Mol Endocrinol* 2000; **25**: 169-193 [PMID: 11013345]
- Principi M, Day R, Marangi S, Burattini O, De Francesco V, Ingrosso M, Pisani A, Panella C, Forbes A, Di Leo A, Francavilla A, Ierardi E. Differential immunohistochemical expression of syndecan-1 and tumor necrosis factor alpha in colonic mucosa of patients with Crohn's disease. *Immunopharmacol Immunotoxicol* 2006; **28**: 185-195 [PMID: 16873088]
- Moorghen M, Chapman M, Appleton DR. An organ-culture method for human colorectal mucosa using serum-free medium. *J Pathol* 1996; **180**: 102-105 [PMID: 8943824]
- Dame MK, Bhagavathula N, Mankey C, DaSilva M, Paruchuri T, Aslam MN, Varani J. Human colon tissue in organ culture: preservation of normal and neoplastic characteristics. *In Vitro Cell Dev Biol Anim* 2010; **46**: 114-122 [PMID: 19915935]
- Della Valle N, Giorgio F, Cantatore S, Zotti M, Panella C, Ierardi E. Tumor Necrosis Factor alpha, syndecan 1 and basic fibroblast growth factor levels and site in cultured biopsy specimens of patients with inflammatory bowel diseases after incubation with infliximab. *Dig Liver Dis* 2013; **45**: S107
- Rieder F, Brenmoehl J, Leeb S, Schölmerich J, Rogler G. Wound healing and fibrosis in intestinal disease. *Gut* 2007; **56**: 130-139 [PMID: 17172588 DOI: 10.1136/gut.2006.090456]
- Sappino AP, Schürch W, Gabbiani G. Differentiation repertoire of fibroblastic cells: expression of cytoskeletal proteins as marker of phenotypic modulations. *Lab Invest* 1990; **63**: 144-161 [PMID: 2116562]
- Pucilowska JB, McNaughton KK, Mohapatra NK, Hoyt EC, Zimmermann EM, Sartor RB, Lund PK. IGF-I and procollagen alpha1(I) are coexpressed in a subset of mesenchymal cells in active Crohn's disease. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G1307-G1322 [PMID: 11093955]
- Pucilowska JB, Williams KL, Lund PK. Fibrogenesis. IV. Fibrosis and inflammatory bowel disease: cellular mediators and animal models. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G653-G659 [PMID: 11005750]
- Sorrentino D, Avellini C, Beltrami CA, Pasqual E, Zearo E. Selective effect of infliximab on the inflammatory component of a colonic stricture in Crohn's disease. *Int J Colorectal Dis* 2006; **21**: 276-281 [PMID: 15951989]

P- Reviewer Guo H S- Editor Zhai HH L- Editor A  
E- Editor Li JY



## Severe hemorrhagic colitis in a patient with chronic myeloid leukemia in the blastic phase after dasatinib use

Zahra Kmira, Ben Sayed Nesrine, Zaghouani Houneida, Ben Fredj Wafa, Slama Aida, Ben Youssef Yosra, Zaier Monia, Badreddine Sriha, Khelif Abderrahim

Zahra Kmira, Ben Sayed Nesrine, Ben Fredj Wafa, Ben Youssef Yosra, Zaier Monia, Khelif Abderrahim, Department of Clinical Hematology, University Hospital Farhat Hached, Sousse 4000, Tunisia

Zaghouani Houneida, Department of Radiology, University Hospital Farhat Hached, Sousse 4000, Tunisia

Slama Aida, Department of Gastroenterology, University Hospital Sahloul, Sousse 4000, Tunisia

Badreddine Sriha, Department of Anatomopathology, University Hospital Farhat Hached, Sousse 4000, Tunisia

Author contributions: All authors contributed to this work.

Correspondence to: Dr. Zahra Kmira, Department of Clinical Hematology, University Hospital Farhat Hached, Avenue Ibn Eljazzar, Sousse 4000, Tunisia. [kmira\\_zahra@yahoo.fr](mailto:kmira_zahra@yahoo.fr)

Telephone: +216-21-135521

Received: June 23, 2013 Revised: July 17, 2013

Accepted: August 4, 2013

Published online: August 15, 2013

### Abstract

Dasatinib is a second-line tyrosine kinase inhibitor used in patients with imatinib resistant or intolerant chronic myeloid leukemia (CML) and Philadelphia chromosome-positive acute leukemia. Gastrointestinal bleeding may occur in up to 7% of patients using dasatinib, although, severe dasatinib-related acute colitis had rarely been reported. Here, we present the case of a 36-year-old female who progressed to acute myeloid leukemia after fourteen months of receiving imatinib for CML in the chronic phase and was treated with a dasatinib-containing chemotherapy regimen. On day 34 of treatment, the patient developed moderate abdominal pain and bloody diarrhea with mucous. Analyses of stool specimens were negative for parasites, *Clostridium difficile*, and other pathogenic bacteria. The cytomegalovirus pp65 antigen was negative in her blood leukocytes. A colonoscopy revealed acute colitis, and a mucosal biopsy showed non-specific colitis. The patient was treated with broad-spectrum antibiotics, bowel rest and hydration, and dasatinib

treatment was stopped. Her bloody diarrhea improved within 72 h. After confirming cytological remission, the patient received initial course of consolidation, and dasatinib treatment was reinstated. However, hemorrhagic colitis recurred. After discontinuing dasatinib, hemorrhagic colitis drastically improved and did not recur following the administration of nilotinib. The characteristics of our patient suggest that dasatinib treatment can lead to hemorrhagic colitis, which typically resolves after discontinuation of the drug.

© 2013 Baishideng. All rights reserved.

**Key words:** Philadelphia chromosome; Chronic myeloid leukemia; Dasatinib; Colitis

**Core tip:** Dasatinib is a second-line tyrosine kinase inhibitor used in imatinib resistant or intolerant chronic myeloid leukemia (CML) and Philadelphia chromosome-positive acute leukemia patients. Dasatinib, which binds to the active and inactive conformation of the BCR-ABL oncoprotein, demonstrates greater potency than imatinib for wild-type and mutant BCR-ABL cases, with the exception of the T315I mutation. The most frequent adverse effects include myelosuppression, diarrhea, nausea and peripheral edema. Severe dasatinib-related acute colitis without thrombocytopenia, coagulation abnormalities or colonic ulcers has rarely been reported. Here, we report the case of an adult patient with Philadelphia chromosome positive CML in the blastic phase who developed acute colitis after dasatinib use.

Kmira Z, Nesrine BS, Houneida Z, Wafa BF, Aida S, Yosra BY, Monia Z, Sriha B, Abderrahim K. Severe hemorrhagic colitis in a patient with chronic myeloid leukemia in the blastic phase after dasatinib use. *World J Gastrointest Pathophysiol* 2013; 4(3): 59-62 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i3/59.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i3.59>

## INTRODUCTION

Dasatinib, an oral inhibitor of ABL and SRC family tyrosine kinases, is an effective drug for patients with Philadelphia chromosome positive (Ph+) leukemia, especially for those who develop resistance or who are intolerant to imatinib<sup>[1]</sup>. Mild to moderate thrombocytopenia and neutropenia occurred in approximately 50% of patients, but these conditions are generally well tolerated. Other side effects include diarrhea, headache, weakness, pleural effusion, nausea and peripheral edema. In addition, gastrointestinal (GI) bleeding may occur in up to 7% of patients using dasatinib<sup>[2]</sup>, although severe dasatinib-related hemorrhagic colitis without thrombocytopenia, coagulation abnormalities or colonic ulcer has been rarely reported. Here, we report the case of an adult patient with Ph+ chronic myeloid leukemia (CML) in the blastic phase who suffered from acute colitis after dasatinib use.

## CASE REPORT

A 36-year-old female, who has been treated with fourteen months imatinib for CML in the chronic phase, progressed to acute myeloid leukemia. The patient was given a course of systemic chemotherapy according to the protocol for AML, consisting of rubidomycin (45 mg/m<sup>2</sup> daily for 3 d), cytosine arabinoside (200 mg/m<sup>2</sup> continuous infusion for seven days) and dasatinib (140 mg once a day). After the end of chemotherapy, dasatinib was continued as maintenance therapy. On day 34 of treatment, the patient developed moderate abdominal pain and bloody diarrhea with mucous (4-6 bowel movements a day). Physical examination revealed the absence of fever and mild abdominal tenderness upon palpation. The laboratory results were as follows: hemoglobin 100 g/L, white blood cells 4 × 10<sup>9</sup>/L with an absolute neutrophil count of 1.5 × 10<sup>9</sup>/L, platelets 185 × 10<sup>9</sup>/L, prothrombin time 15 s, active partial thromboplastin time 33 s and an international normalized ratio of 1.3. The analyses of stool specimens were negative for parasites, *Clostridium difficile*, and other pathogenic bacteria. The cytomegalovirus pp65 antigen was negative in her blood leukocytes. An abdominal ultrasound showed the presence of uniform circumferential thickening of the transverse colon and splenic flexure with pericolic fat infiltration, indicating potential colitis. An abdominal computed tomography scan revealed bowel wall thickening up to 1 cm, involving the entire colon with infiltration of the mesenteric fat and a pelvic peritoneal effusion consistent with pancolitis. A total colonoscopy revealed no active bleeding, but there were multiple millimetric, nodular, hyperemic lesions on the mucosa involving the entire colon (Figure 1). A mucosal biopsy showed nonspecific colitis with a well-preserved crypt structure and lymphocytic infiltration in the lamina propria (Figure 2). Infiltrative lymphocytes expressed a high proportion of CD3 and sparse of CD20. No viral inclusion or apoptotic bodies were observed. The patient was treated with broad-spectrum



Figure 1 Colonoscopy showed multiple millimetric nodular hyperemic lesions on the mucosa.

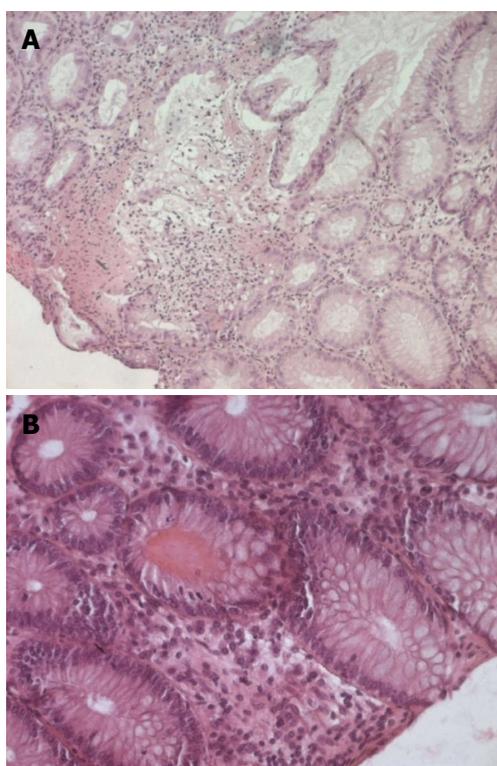


Figure 2 Mucosal biopsy of colon. A: A well-preserved crypt structure with lymphocytes infiltration in the lamina propria (original magnification × 100); B: Viral inclusions and apoptotic bodies were absent (original magnification × 200).

antibiotics, bowel rest and hydration, and dasatinib treatment was stopped. Improvement in the bloody diarrhea was evident after 72 h, and a control colonoscopy was performed ten days later and showed that the colonic mucosa was quite normal. After confirming the achievement of cytological remission (4% of medullary blasts), the patient received the first course of consolidation treatment (cytosine arabinoside + etoposide + rubidomycin), and dasatinib was reinstated. On day 6 of treatment, the patient again developed severe diarrhea with a large amount of intestinal hemorrhage, and a repeat colonoscopy was consistent with acute colitis (Figure 3). Again, dasatinib treatment was stopped, and the hemorrhagic

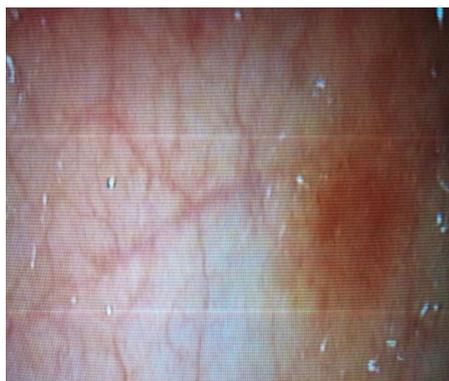


Figure 3 Macroscopic findings include erythema, edema and friability.

colitis drastically improved. A repeat colonoscopy was normal after discontinuing dasatinib treatment. Nilotinib (800 mg/d) was started as an alternative to dasatinib for CML treatment, and the hemorrhagic colitis did not recur. After confirming cytogenetic and molecular remission, the patient received a bone marrow transplantation from an HLA-identical, intrafamilial donor. She is still alive and in complete remission.

## DISCUSSION

Diarrhea and nausea are generally observed in approximately about 30% and 20% of patients respectively, during dasatinib therapy<sup>[3]</sup>. GI bleeding may occur during dasatinib treatment, but is generally mild and easily handled. Data from 84 patients treated with dasatinib show that grade 3 or 4 GI bleeding occurred in approximately 6% of cases, although, detailed information on the mechanisms of action was not reported<sup>[4]</sup>. Herein, we reported the case of a patient with Ph+ CML in the blastic phase who had severe hemorrhagic colitis during dasatinib therapy. Acute colitis is a pathological phenomenon characterized by the infiltration of inflammatory cells into the lamina propria. The pathogenesis of acute colitis is complex and many immune mediators and cells play roles in this poorly understood condition<sup>[5]</sup>. Moreover, the colitis induced by dasatinib may not be attributed simply to the tyrosine kinase inhibition because our patient did not experience similar GI symptoms during imatinib therapy. Although the precise mechanism is unknown, one possible explanation is that dasatinib may have a stronger inhibitory effect on tyrosine kinases and/or have some kinase-specific inhibitory effects on SRC-family kinases, BCR-ABL, c-KIT, EPHA2, and the PDGF receptor<sup>[6,7]</sup>. The SRC-family kinases are also expressed in normal B cells and are likely inhibited by dasatinib. Fei *et al.*<sup>[8]</sup> showed that dasatinib inhibited the proliferation and function of T regulatory cells by decreasing the expression of box P3 transcription factor, 5-glucocorticoid induced tumor necrosis factor receptor, cytotoxic T-cell-associated protein 4 and inducing apoptosis in the G<sub>0</sub>/G<sub>1</sub> phase of the cell cycle in T regulatory cells.

Further-more, some studies have shown that dasatinib suppresses the function of natural killer cells and T cells by inhibiting SRC-family kinases<sup>[9,10]</sup>. Therefore, dasatinib may cause acute colitis by decreasing immune tolerance to intestinal microflora through reducing the number of immunoregulatory cells and inhibiting signal transduction pathways.

In conclusion, hemorrhagic colitis may develop during dasatinib treatment although this condition improves upon discontinuation of the drug. Additional studies with greater numbers of patients are necessary to elucidate the relationship between hemorrhagic colitis and dasatinib therapy.

## REFERENCES

- 1 **Steinberg M.** Dasatinib: a tyrosine kinase inhibitor for the treatment of chronic myelogenous leukemia and Philadelphia chromosome-positive acute lymphoblastic leukemia. *Clin Ther* 2007; **29**: 2289-2308 [PMID: 18158072 DOI: 10.1016/j.clinthera.2007.11.005]
- 2 **Guilhot F, Apperley J, Kim DW, Bullorsky EO, Baccarani M, Roboz GJ, Amadori S, de Souza CA, Lipton JH, Hochhaus A, Heim D, Larson RA, Branford S, Muller MC, Agarwal P, Gollerkeri A, Talpaz M.** Dasatinib induces significant hematologic and cytogenetic responses in patients with imatinib-resistant or -intolerant chronic myeloid leukemia in accelerated phase. *Blood* 2007; **109**: 4143-4150 [PMID: 17264298 DOI: 10.1182/blood-2006-09-046839]
- 3 **Kantarjian H, Pasquini R, Hamerschlak N, Rousselot P, Holowiecki J, Jootar S, Robak T, Khoroshko N, Masszi T, Skotnicki A, Hellmann A, Zaritsky A, Golenkov A, Radich J, Hughes T, Countouriotis A, Shah N.** Dasatinib or high-dose imatinib for chronic-phase chronic myeloid leukemia after failure of first-line imatinib: a randomized phase 2 trial. *Blood* 2007; **109**: 5143-5150 [PMID: 17317857 DOI: 10.1182/blood-2006-11-056028]
- 4 **Talpaz M, Shah NP, Kantarjian H, Donato N, Nicoll J, Paquette R, Cortes J, O'Brien S, Nicaise C, Bleickardt E, Blackwood-Chirchir MA, Iyer V, Chen TT, Huang F, Decillis AP, Sawyers CL.** Dasatinib in imatinib-resistant Philadelphia chromosome-positive leukemias. *N Engl J Med* 2006; **354**: 2531-2541 [PMID: 16775234]
- 5 **Bouma G, Strober W.** The immunological and genetic basis of inflammatory bowel disease. *Nat Rev Immunol* 2003; **3**: 521-533 [PMID: 12876555]
- 6 **Piccaluga PP, Paolini S, Martinelli G.** Tyrosine kinase inhibitors for the treatment of Philadelphia chromosome-positive adult acute lymphoblastic leukemia. *Cancer* 2007; **110**: 1178-1186 [PMID: 17701954 DOI: 10.1002/cncr.22881]
- 7 **Schittenhelm MM, Shiraga S, Schroeder A, Corbin AS, Griffith D, Lee FY, Bokemeyer C, Deininger MW, Druker BJ, Heinrich MC.** Dasatinib (BMS-354825), a dual SRC/ABL kinase inhibitor, inhibits the kinase activity of wild-type, juxtamembrane, and activation loop mutant KIT isoforms associated with human malignancies. *Cancer Res* 2006; **66**: 473-481 [PMID: 16397263 DOI: 10.1158/0008-5472.CAN-05-2050]
- 8 **Fei F, Yu Y, Schmitt A, Rojewski MT, Chen B, Götz M, Döhner H, Bunjes D, Schmitt M.** Dasatinib inhibits the proliferation and function of CD4+CD25+ regulatory T cells. *Br J Haematol* 2009; **144**: 195-205 [PMID: 19016717 DOI: 10.1111/j.1365-2141.2008.07433.X]
- 9 **Blake SJ, Bruce Lyons A, Fraser CK, Hayball JD, Hughes TP.** Dasatinib suppresses in vitro natural killer cell cytotoxicity. *Blood* 2008; **111**: 4415-4416 [PMID: 18398058 DOI:

Kmira Z *et al.* Colitis due to dasatinib use

10.1182/blood-2008-02-138701]

- 10 **Schade AE**, Schieven GL, Townsend R, Jankowska AM, Susulic V, Zhang R, Szpurka H, Maciejewski JP. Dasatinib,

a small-molecule protein tyrosine kinase inhibitor, inhibits T-cell activation and proliferation. *Blood* 2008; **111**: 1366-1377 [PMID: 17962511 DOI: 10.1182/blood-2007-04-084814]

**P- Reviewer** Quaresma J **S- Editor** Song XX **L- Editor** A  
**E- Editor** Li JY



## Why is damage limited to the mucosa in ulcerative colitis but transmural in Crohn's disease?

Xiaofa Qin

Xiaofa Qin, Department of Surgery, University of Medicine and Dentistry of New Jersey, New Jersey Medical School, Newark, NJ 07103, United States

Author contributions: Qin X solely contributed to this paper. Correspondence to: Xiaofa Qin, MD, PhD, Department of Surgery, University of Medicine and Dentistry of New Jersey, New Jersey Medical School, 185 South Orange Avenue, Newark, NJ 07103, United States. [qinxi@umdnj.edu](mailto:qinxi@umdnj.edu)

Telephone: +1-973-9722896 Fax: +1-973-9726803

Received: May 12, 2013 Revised: June 8, 2013

Accepted: June 19, 2013

Published online: August 15, 2013

© 2013 Baishideng. All rights reserved.

**Key words:** Inflammatory bowel disease; Ulcerative colitis; Crohn's disease; Pathogenesis; Neutrophils; Macrophages

**Core tip:** In my opinion, ulcerative colitis is more like bacterial pneumonia with the involvement of mainly neutrophils, capacious exudates into the cavity but limited damage of the tissue, while Crohn's disease is more like pneumoconiosis, such as silicosis or berylliosis, or tuberculosis of the lung with the involvement of mainly macrophages and manifested as granulomatous inflammation in the interstices, destruction of the tissue, extensive proliferation of fibroblasts and formation of fibrosis.

### Abstract

It has been a big puzzle as why the inflammation of ulcerative colitis (UC) is limited to the mucosa, while in Crohn's disease (CD) the inflammation is transmural and can be seen in all layers of the gut. Here, I give a tentative explanation extended from the unified hypothesis I proposed on the etiology of inflammatory bowel disease. This hypothesis suggested that both UC and CD are caused by weakening of the gut barrier due to damage of the protective mucus layer and the underlying tissue by the poorly inactivated digestive proteases resulting from a reduction of gut bacteria by dietary chemicals like saccharin and sucralose. However, the large amounts of bacteria in the colon make the recruitment of neutrophils and formation of crypt abscess the main manifestation of UC, while the infiltration of antigens and dietary particles in the small and large intestine mainly cause the recruitment of macrophages and formation of granulomas as the main manifestations in CD. The fast reacting and short life span of neutrophils make the fight and damage limited to the surface of the mucosa. In contrast, the long life span and constant movement of macrophages may bring the harmful agents deep into the tissue. Therefore, the pathogenesis of UC may be more like bacterial pneumonia, while CD may be more like pneumoconiosis or tuberculosis of the lung.

Qin X. Why is damage limited to the mucosa in ulcerative colitis but transmural in Crohn's disease? *World J Gastrointest Pathophysiol* 2013; 4(3): 63-64 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i3/63.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i3.63>

### TO THE EDITOR

As we know, the inflammation of ulcerative colitis (UC) is limited to the mucosa, while in Crohn's disease (CD) the inflammation is transmural and can be seen in all layers of the gut<sup>[1,2]</sup>. These key features reflect the root mechanism of pathogenesis of the disease but a coherent explanation remains lacking. Here, I give a tentative explanation extended from a unified hypothesis I proposed on the etiology of inflammatory bowel disease (IBD), including the cause and mechanism of IBD, as well as the relationship between UC and CD<sup>[3]</sup>. This hypothesis suggested that both UC and CD are caused by weakening of the gut barrier due to damage of the

protective mucus layer and the underlying tissue by the poorly inactivated digestive proteases resulting from a reduction of gut bacteria by dietary chemicals like saccharin and sucralose. However, the large amounts of bacteria in the colon make the recruitment of neutrophils and formation of crypt abscess the main manifestation of UC, while the infiltration of antigens and dietary particles in the small and large intestine mainly causes the recruitment of macrophages and formation of granulomas as the main manifestations in CD. With a life span of just a couple of days, neutrophils are the fast reaction army of the body and will be dispatched to the very front and fight vigorously there until they die. However, macrophages are the cleaner and order keeper inside the body, with a primary duty of clearing up debris like the dead cells. They have a life span of months and can fulfill their job by eating the debris, digesting them, moving around and picking up again for quite a while and distance. However, things like dietary particles and multiple species of bacteria, such as *Mycobacterium*, *Salmonella*, *Listeria*, *Shigella* and adherent-invasive *Escherichia coli*, are beyond the ability of macrophages to digest. Hence, the macrophages may have carried and moved them quite far inside before they die. Then, the particles or bacteria will be released and picked up by other macrophages and moved further inside. The persistent existence of these foreign bodies results in the recruitment of more macrophages and other immune cells, eventually leading to the initiation of the quarantine process and formation of granulomas<sup>[4,5]</sup>. In my opinion, UC is more like bacterial pneumonia (but long lasting due to the continuous exist-

tence of large amounts of bacteria in the colon) with the involvement of mainly neutrophils, capacious exudates into the cavity (crypts and lumen in the gut and alveoli in the lung) but limited damage of the tissue<sup>[6]</sup>, while CD is more like pneumoconiosis, such as silicosis or berylliosis<sup>[7]</sup>, or tuberculosis of the lung with the involvement of mainly macrophages and manifested as granulomatous inflammation in the interstices, destruction of the tissue, extensive proliferation of fibroblasts and formation of fibrosis.

## REFERENCES

- 1 **Colombel JF**, Watson AJ, Neurath MF. The 10 remaining mysteries of inflammatory bowel disease. *Gut* 2008; **57**: 429-433 [PMID: 18079284 DOI: 10.1136/gut.2007.122192]
- 2 **Xavier RJ**, Podolsky DK. Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 2007; **448**: 427-434 [PMID: 17653185 DOI: 10.1038/nature06005]
- 3 **Qin X**. Etiology of inflammatory bowel disease: a unified hypothesis. *World J Gastroenterol* 2012; **18**: 1708-1722 [PMID: 22553395 DOI: 10.3748/wjg.v18.i15.1708]
- 4 **Turk JL**, Narayanan RB. The monocyte-macrophage system in granulomatous inflammation. *Haematol Blood Transfus* 1981; **27**: 101-107 [PMID: 7327427]
- 5 **Epstein WL**, Fukuyama K. Mechanisms of granulomatous inflammation. *Immunol Ser* 1989; **46**: 687-721 [PMID: 2488875]
- 6 **Montón C**, Torres A. Lung inflammatory response in pneumonia. *Monaldi Arch Chest Dis* 1998; **53**: 56-63 [PMID: 9632909]
- 7 **Chong S**, Lee KS, Chung MJ, Han J, Kwon OJ, Kim TS. Pneumoconiosis: comparison of imaging and pathologic findings. *Radiographics* 2006; **26**: 59-77 [PMID: 16418244 DOI: 10.1148/rg.261055070]

P- Reviewer Duman U S- Editor Zhai HH  
L- Editor Roemmele A E- Editor Li JY



# World Journal of *Gastrointestinal Pathophysiology*

*World J Gastrointest Pathophysiol* 2013 November 15; 4(4): 65-125

Volume End





**Contents**

Quarterly Volume 4 Number 4 November 15, 2013

- |                    |     |   |
|--------------------|-----|---|
| <b>EDITORIAL</b>   | 65  | Genetic contribution to motility disorders of the upper gastrointestinal tract<br><i>Sarnelli G, D'Alessandro A, Pesce M, Palumbo I, Cuomo R</i>                              |
| <b>REVIEWS</b>     | 74  | Pancreatic cancer diagnosis by free and exosomal miRNA<br><i>Zöller M</i>   |
| <b>MINIREVIEWS</b> | 91  | Prospects and challenges for intestinal microbiome therapy in pediatric gastrointestinal disorders<br><i>Kellermayer R</i>  |
|                    | 94  | Intestinal barrier: Molecular pathways and modifiers<br><i>Jeon MK, Klaus C, Kaemmerer E, Gassler N</i>   |
|                    | 100 | Fibrogenesis and fibrosis in inflammatory bowel diseases: Good and bad side of same coin?<br><i>Principi M, Giorgio F, Losurdo G, Neve V, Contaldo A, Di Leo A, Ierardi E</i> |
|                    | 108 | Effects of occupational stress on the gastrointestinal tract<br><i>Huerta-Franco MR, Vargas-Luna M, Tienda P, Delgadillo-Holtfort I, Balleza-Ordaz M, Flores-Hernandez C</i>  |
|                    | 119 | Usefulness of percutaneous endoscopic gastrostomy for supportive therapy of advanced aerodigestive cancer<br><i>Ogino H, Akiho H</i>  |

**Contents***World Journal of Gastrointestinal Pathophysiology*  
Volume 4 Number 4 November 15, 2013**APPENDIX** I-V Instructions to authors**ABOUT COVER** Editorial Board Member of *World Journal of Gastrointestinal Pathophysiology*, Maria-Raquel Huerta-Franco, PhD, Department of Applied Science and Labor Research, DCS Campus Leon, University of Guanajuato, Aquiles Serdan No. 924, Colonia Obregon, Leon, CP 37150, Guanajuato, Mexico**AIM AND SCOPE***World Journal of Gastrointestinal Pathophysiology* (*World J Gastrointest Pathophysiol*, *WJGP*, online ISSN 2150-5330, DOI: 10.4291), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.*WJGP* is to report rapidly the most recent results in basic and clinical research on gastrointestinal pathophysiology, including all aspects of normal or abnormal function of the gastrointestinal tract, hepatobiliary system, and pancreas. *WJGP* specifically covers growth and development, digestion, secretion, absorption, metabolism and motility relative to the gastrointestinal organs, as well as immune and inflammatory processes, and neural, endocrine and circulatory control mechanisms that affect these organs. This journal will also report new methods and techniques in gastrointestinal pathophysiological research.We encourage authors to submit their manuscripts to *WJGP*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.**INDEXING/ ABSTRACTING***World Journal of Gastrointestinal Pathophysiology* is now indexed in PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.**FLYLEAF** I-IV Editorial Board**EDITORS FOR THIS ISSUE**Responsible Assistant Editor: *Xin-Xin Che*  
Responsible Electronic Editor: *Ya-Jing Lu*  
Proofing Editor-in-Chief: *Lian-Sheng Ma*Responsible Science Editor: *Huan-Huan Zhai***NAME OF JOURNAL**  
*World Journal of Gastrointestinal Pathophysiology***ISSN**  
ISSN 2150-5330 (online)**LAUNCH DATE**  
April 15, 2010**Frequency**  
Quarterly**EDITOR-IN-CHIEF**  
**Thomas Y Ma, MD, PhD, Professor, Chief**, Division of Gastroenterology and Hepatology, University of New Mexico, MSC10 5550, 1 UNM, Albuquerque, NM 87131, United States**EDITORIAL OFFICE**  
Jin-Lei Wang, Director  
Xiu-Xia Song, Vice Director*World Journal of Gastrointestinal Pathophysiology*  
Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China  
Telephone: +86-10-85381892  
Fax: +86-10-85381893  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
<http://www.wjgnet.com>**PUBLISHER**  
Baishideng Publishing Group Co., Limited  
Flat C, 23/F, Lucky Plaza, 315-321 Lockhart Road, Wan Chai, Hong Kong, China  
Telephone: +852-6555-7188  
Fax: +852-3177-9906  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
<http://www.wjgnet.com>**PUBLICATION DATE**  
November 15, 2013**COPYRIGHT**

© 2013 Baishideng Publishing Group Co., Limited. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

**INSTRUCTIONS TO AUTHORS**Full instructions are available online at [http://www.wjgnet.com/2150-5330/g\\_info\\_20100316080008.htm](http://www.wjgnet.com/2150-5330/g_info_20100316080008.htm)**ONLINE SUBMISSION**<http://www.wjgnet.com/esp/>

## Genetic contribution to motility disorders of the upper gastrointestinal tract

Giovanni Sarnelli, Alessandra D'Alessandro, Marcella Pesce, Ilaria Palumbo, Rosario Cuomo

Giovanni Sarnelli, Alessandra D'Alessandro, Marcella Pesce, Ilaria Palumbo, Rosario Cuomo, Gastroenterology Unit, Department of Clinical Medicine and Surgery, University of Naples "Federico II", 80131 Naples, Italy

Author contributions: Sarnelli G contributed to the conception, made revisions and helped in writing; D'Alessandro A and Pesce M wrote the paper and contributed to revision of the literature; Palumbo I provided supportive contributions; Cuomo R contributed to revision of the literature.

Correspondence to: Giovanni Sarnelli, MD, PhD, Gastroenterology Unit, Department of Clinical Medicine and Surgery, University of Naples "Federico II", Via Sergio Pansini, 5, 80131 Naples, Italy. [sarnelli@unina.it](mailto:sarnelli@unina.it)

Telephone: +39-81-7463488 Fax: +39-81-7462753

Received: June 27, 2013 Revised: August 9, 2013

Accepted: October 17, 2013

Published online: November 15, 2013

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Achalasia; Functional dyspepsia; Genetic predisposition; Hypertrophic pyloric stenosis; Motility disorder

**Core tip:** Achalasia, functional dyspepsia and hypertrophic pyloric stenosis represent the main motility disorders of upper gastrointestinal tract. All these diseases have a less known pathophysiology and a presumable genetic predisposition in common. This review outlines the current knowledge on genes involved in the onset of these pathologies in order to promote further association studies which can help to explain this complex picture and find new therapeutic targets.

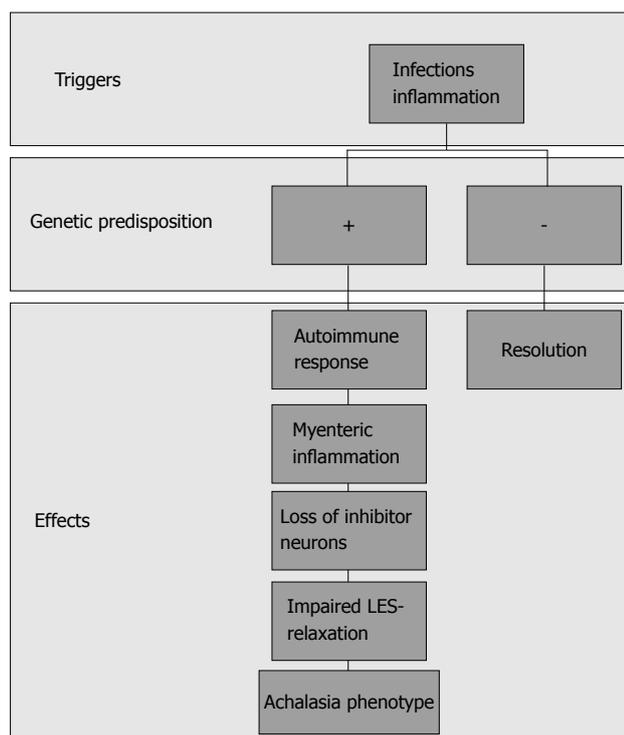
### Abstract

Motility disorders of the upper gastrointestinal tract encompass a wide range of different diseases. Esophageal achalasia and functional dyspepsia are representative disorders of impaired motility of the esophagus and stomach, respectively. In spite of their variable prevalence, what both diseases have in common is poor knowledge of their etiology and pathophysiology. There is some evidence showing that there is a genetic predisposition towards these diseases, especially for achalasia. Many authors have investigated the possible genes involved, stressing the autoimmune or the neurological hypothesis, but there is very little data available. Similarly, studies supporting a post-infective etiology, based on an altered immune response in susceptible individuals, need to be validated. Further association studies can help to explain this complex picture and find new therapeutic targets. The aim of this review is to summarize current knowledge of genetics in motility disorders of the upper gastrointestinal tract, addressing how genetics contributes to the development of achalasia and functional dyspepsia respectively.

Sarnelli G, D'Alessandro A, Pesce M, Palumbo I, Cuomo R. Genetic contribution to motility disorders of the upper gastrointestinal tract. *World J Gastrointest Pathophysiol* 2013; 4(4): 65-73 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/65.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.65>

### INTRODUCTION

Digestive motility is a highly coordinated process which enables mixing, absorption and propulsion of the ingesta through the gastrointestinal tract up to expulsion of residues. This function depends on a finely balanced integration between smooth muscle contractility and the related pacemaker activity evoked by the interstitial cells of Cajal (ICCs) that are finely regulated by intrinsic and extrinsic innervations [*i.e.*, enteric nervous system (ENS) and sympathetic and parasympathetic nerves, respectively]<sup>[1,2]</sup>. A disturbed digestive motility can occur as a result of a variety of abnormalities affecting each of these elements (alone or in combination), with consequent altered physiology of gut peristalsis and symptom generation.



**Figure 1** Supposed pathophysiology of achalasia: triggering external factors are able to induce a loss of inhibitor neurons in individuals genetically predisposed, causing an impaired lower esophageal sphincter relaxation.

The symptom complex is site specific and depends on the gastrointestinal tract involved. Thus, patients with disturbed esophageal motility may complain of dysphagia, regurgitation or chest pain, whereas patients with gastric dysmotility may report symptoms of nausea, vomiting, postprandial fullness, early satiety or epigastric pain. Esophageal achalasia and functional dyspepsia are the most representative motility disorders of the upper gastrointestinal (GI) tract, with the first rare and well characterized and the latter highly prevalent and less defined.

Like the majority of functional gastrointestinal disorders (FGIDs), they are characterized by the persistence of symptoms in the absence of reliable biological markers. As for other complex diseases, the pathophysiology of impaired upper GI tract motility diseases is supposed to be multifactorial, with triggering external factors that are able to generate the gastrointestinal dysfunction in individuals who are more or less genetically predisposed (Figure 1). The idea that an individual genotype may contribute to the development of FGIDs is suggested by clinical observations and prospective studies indicating that there is clustering of FGIDs within families<sup>[3,4]</sup> and an increased concordance in monozygotic twins<sup>[5-7]</sup>. The most commonly used approach in functional dyspepsia (FD) to date has been to search for correlations of a candidate gene polymorphism with the symptom based phenotype. More recently, a few studies have aimed to identify correlations between gene polymorphism and biological intermediate phenotypes, such as impaired motility.

In this review, we summarize the current knowledge

on the genetic contribution to upper GI motor disorders. We evaluate the most recent literature on the genetic epidemiology of representative motility disorders of the upper GI tract (UGT). We describe the putative genetic contributions that have been addressed and the potential association with single mechanisms, such as receptors, transporters and translation, or transduction mechanisms involved in the disturbed motility of the esophagus, stomach and pylorus, respectively impaired in esophageal achalasia, functional dyspepsia and hypertrophic pyloric stenosis. Each of the diseases can be considered paradigmatic because of the following: (1) hypertrophic pyloric stenosis (HPS) is characteristic of infants and there is significant evidence about the role of genetic factors; (2) isolated idiopathic achalasia can be considered the prototype of defective esophageal motility in adults and the role of genetic factors is emerging as a major challenge to explain the disease; and (3) functional dyspepsia cannot be paradigmatically considered a primary gastric motor disorder, but is characterized by a significant association between impaired gastric motility and symptoms to which genetic factors contribute.

## IMPAIRED MOTILITY OF THE ESOPHAGUS

### Achalasia

Idiopathic achalasia is the best recognized primary motor disorder of the esophagus. It is characterized by incomplete relaxation of the lower esophageal sphincter (LOS) and absence of peristalsis that causes bolus impaction and generation of symptoms like dysphagia, regurgitation and chest pain<sup>[8]</sup>. Although the pathogenesis of achalasia is still largely unknown, it is now clear that a major issue is the loss of neurons in the esophageal myenteric plexus. We know that neurons gradually disappear in the lower part of the esophagus of achalasia patients. In most cases, this process is the result of a localized infiltration of immune cells, determining an inflammatory-based neurodegeneration<sup>[9,10]</sup>. Interestingly, this process is prevalent for inhibitory neurons containing nitric oxide (NO) and vasoactive intestinal polypeptides (VIP) and accounts for the loss of inhibitory inputs, with consequent abnormal esophageal function<sup>[11-13]</sup>. Notwithstanding this histological background, we do not know the precise pathogenic mechanism of achalasia; consequently, hereditary, degenerative, autoimmune and infectious factors have all been claimed to be possible causes of the disease.

A certain degree of genetic influence in achalasia is suggested by the familial occurrence and twin concordance. However, a systematic analysis of the literature revealed that twin concordance was significant but still inconclusive<sup>[6]</sup>. On the other hand, the only family study of achalasia conducted to date is biased by the small sample size and because the diagnosis of achalasia was based on a self-reported questionnaire<sup>[14]</sup>. In addition, achalasia may present as part of a genetic syndrome or in association with isolated abnormalities or diseases<sup>[15]</sup>.

**Table 1 Overview of genetic association studies in achalasia**

Protein (gene)	Polymorphism	Finding	Ref.
PTPN22	C1858T	Risk factor	[28]
IL-10	GCC	Protective	[29]
IL-23R	G381A	Protective	[30]
iNOS	iNOS22*A/A	No association	[35]
eNOS	eNOS*4*4°	No association	[35]
iNOS	(CCTTT) n > 12	Risk factor	[36]
cKit	Rs6554199	Risk factor	[37]
VIPR type 1	Rs437876 and rs896	Risk factor	[38]

iNOS: Inducible nitric oxide synthase; IL: Interleukin; PTPN22: Polymorphism C1858T of phosphatase N22.

An achalasia phenotype is indeed present in well characterized genetic syndromes, like Down<sup>[16]</sup> and Allgrove syndromes<sup>[17]</sup>, the familial visceral neuropathy<sup>[18]</sup> and the achalasia microcephaly syndrome<sup>[19]</sup>. In many of the above mentioned syndromes, achalasia does occur in the majority of the patients, but it generally presents in infancy.

Although these findings indicate that genetic factors are involved in the development of an achalasia phenotype, they do not provide insights into the pathogenesis of the sporadic form of achalasia, which affects the vast majority of patients and presents with adult-onset achalasia. As for other complex diseases, it is likely that the etiology of this form of achalasia is multifactorial, *i.e.*, a combination of the cumulative effect of variants in various risk genes and environmental factors leads to the disorder. The idea of an infectious origin of achalasia was first suggested by the evidence of viruses or signs of viral infection in the esophageal tissues of achalasia patients<sup>[20-22]</sup>, but the possibility that the presence of viruses could be sufficient *per se* to explain the disease was ruled out by other observations<sup>[23]</sup>. More recently, the concept that achalasia could be the result of an immune mediated inflammatory disorder induced by a virus has been strongly repurposed<sup>[24]</sup>. Facco *et al*<sup>[24]</sup> demonstrated that HSV-1 or HSV-like antigens were responsible for a significant activation of CD3<sup>+</sup>T cells infiltrating the LES in achalasia patients, likely resulting in an immune-mediated destruction of the myenteric neurons of the LOS. The reasons whereby this process occurs only in esophageal tissues of achalasia patients are unknown, but it is reasonable to assume that some genetic influence may affect the disease phenotype, making some individuals more susceptible to the disease. In addition and most interestingly, several evidences strongly support the idea that genes encoding for proteins involved in the immune response are likely candidates in achalasia.

A significant association between HLA DR or DQ, especially DQA1 \*0103 and DQB1 \*0603, and achalasia has been indeed described<sup>[25-27]</sup>. The increased risk for the development of achalasia in individuals with specific polymorphisms of genes involved in the immune response was also supported by the finding that the polymorphism C1858T of phosphatase N22 (PTPN22

gene, chromosome 1), which is a down-regulator of T-cell activation, is significantly associated with achalasia in Spanish women<sup>[28]</sup>. The same researchers have also demonstrated that the GCC haplotype of *IL-10* gene promoter is a protective factor for achalasia. This specific polymorphism enhances the release of IL-10, an anti-inflammatory cytokine, resulting in a downregulation of immune response<sup>[29]</sup>. In a similar manner, a single nucleotide polymorphism (SNP) of the IL 23 receptor (G381A), which regulates T cell differentiation, appears to be protective against achalasia. De León *et al*<sup>[30]</sup> reported that the coding variant 381Gln of *IL-23R* was significantly more common in patients with achalasia compared with healthy controls.

This evidence sustains the role of both genetic predisposition and immune alteration in the pathogenesis of idiopathic achalasia. All data are summarized in Table 1.

### Contribution of genes with a dual effect on motility and immunity

A disturbed inhibitory neurotransmission is a trademark of achalasia<sup>[11]</sup>. In keeping with this, several studies have addressed the role of NO and VIP that are involved in both defense against infections and inhibitory neurotransmission and may represent ideal candidates to explain the spread of inflammation and inhibitory nerve degeneration<sup>[12,13,19,31-33]</sup>.

Nitric oxide is produced by three different forms of NO synthases: neuronal (nNOS), inducible (iNOS) and endothelial (eNOS). As NO production is genetically regulated, Mearin *et al*<sup>[34]</sup> firstly investigated whether SNP in the different *NOS* genes were respectively involved in the susceptibility to suffer from achalasia. Although a trend toward a higher prevalence of genotypes iNOS22?A/A and eNOS\*4a4a in patients than in controls was observed, the authors failed to find a significant association between *NOS* gene polymorphisms and susceptibility to achalasia. Although the simplest conclusion was that NO is not involved in the pathogenesis of achalasia, this study is biased by the small sample size and by the fact that the SNPs analyzed do not play a major role in gene expression. The lack of any association between the same SNP in the iNOS was also confirmed by a Spanish group in a larger population of achalasia patients, further suggesting that these specific polymorphisms play a minor role in the functional expression of the iNOS gene<sup>[35]</sup>. More recently, our group showed a significant association between the pentanucleotide (CCTTT) polymorphism in the *iNOS* gene promoter and achalasia. Since *in vitro* data showed that the iNOS promoter activity increases in parallel with the repeat number of (CCTTT)n, we concluded that individuals carrying longer forms have an increased risk of achalasia by higher nitric oxide production.

Moreover, growing evidence suggests that esophageal interstitial cells of Cajal, a group of specialized cells that constitutively expresses c-kit and contribute to nitrergic neurotransmission, may be involved in the pathogenesis

of achalasia. Alahdab *et al.*<sup>[36]</sup> have indeed shown that the rs6554199, but not rs2237025, c-kit polymorphism is significantly prevalent in achalasia patients in a Turkish population. Importantly, alterations in ICC have already been reported in other congenital diseases with abnormal peristalsis, emphasizing the key role of these cells in the regulation of GI motor function.

The implication of VIP in the pathogenesis of achalasia was also recently reported<sup>[37]</sup>. Two different SNPs, rs437876 and rs896, in the VIP-receptor type 1 gene were found to be significantly associated with the late onset achalasia. Interestingly, the authors suggest that this was probably related to a genetically based abnormal VIP-R1 signaling that may protect individuals carrying the specific genotype by delaying the immune-mediated neurodegeneration. Although these data need to be replicated, they are extremely interesting because they may suggest that early and late onset forms of idiopathic achalasia are genetically distinct disorders<sup>[38]</sup> (Table 1).

## IMPAIRED MOTILITY OF THE STOMACH

### FD

FD is defined by the presence of persistent symptoms in the upper part of the abdomen in the absence of organic or metabolic pathology<sup>[39]</sup>. On the basis of the Rome III criteria, patients who suffer from functional dyspepsia in the absence of any organic disease are categorized as having postprandial distress syndrome or epigastric pain syndrome<sup>[40]</sup>.

In spite of this, the pathogenesis of FD is still largely unknown and, although delayed gastric emptying, impaired gastric accommodation and visceral hypersensitivity have been all claimed as major underlying mechanisms, it is supposed to be a multifactorial disease<sup>[39]</sup>.

The classic assumption in studies addressing the association between genetic factors and a single or a cluster of diseases postulates that a specific functional gene polymorphism that results in altered protein function may play a role in disease pathophysiology<sup>[41]</sup>. This paradigm that implies a clear correlation between impaired physiological function and symptom generation, however, cannot be applied to FD whose pathophysiology is complex and not necessarily associated with specific symptoms. Since functional dyspepsia is one of the most prevalent FGIDs, a certain genetic influence is suggested by both symptom familial clustering and twin studies reported for FD and IBS<sup>[3]</sup>. So far, most of the studies conducted were designed to search for correlations of a candidate gene polymorphism with the symptom-based phenotype of FD. The candidate genes that have been studied for possible associations with functional dyspepsia are summarized in Table 1.

In a subset of dyspepsia patients, *Helicobacter pylori* (*H. pylori*) infection, even in the absence of ulceration or erosion in gastroduodenal mucosa, has been proposed to play a role in generation of symptoms. A Japanese

group found an association between TLR-2 -196 to -174 allele with a lower risk for developing FD in *Helicobacter pylori* positive subjects. Since the same group had previously demonstrated that the TLR-2 -196 to -174 increases the severity of *H. pylori*-induced inflammation, it is reasonable to hypothesize that the TLR-2 genotype influences the onset of dyspeptic symptoms modulating the degree of inflammatory response<sup>[42]</sup>.

Serotonin (5-HT) is a key signaling molecule affecting upper GI motor and sensory functions<sup>[43]</sup> and thus genes of the serotonergic system are critical candidates in assessing the role of genetic determinants in FD. The action of 5-HT is terminated by the 5-HT transporter (serotonin transporter, SERT) mediated uptake, thereby determining 5-HT availability at the receptor level. A single nucleotide insertion/deletion in the SERT gene, by creating a long (L) and short (S) allelic variant, results in reduced SERT expression and 5-HT uptake<sup>[44]</sup>. Although the SS genotype of the SERT promoter polymorphism has been reported to be associated with diarrhea-predominant irritable bowel syndrome (IBS), two different studies from the United States<sup>[45]</sup> and Europe<sup>[46]</sup> failed to find any significant association between such polymorphisms and FD. Similarly, association studies between SNPs of the 5-HT<sub>3</sub>, 5-HT<sub>1A</sub> and 5-HT<sub>2A</sub> receptors in FD patients failed to yield significant association<sup>[45,46]</sup>. Although both studies were conducted on small populations of patients, the results suggest that the serotonergic pathway plays a minor role in the pathophysiology or, at least in part, in the generation of dyspeptic symptoms.

Another genetic association reported in functional dyspepsia is the link to GNB $\beta$ <sup>[47]</sup>. G-proteins mediate the response to the release of 5-HT and several other neurotransmitters modulating gastroduodenal sensory and motor function.

A common polymorphism of GNB $\beta$  gene has been described and is associated with different genotypes that are predictive of an enhanced G-protein activation. A significant association between different GNB $\beta$  polymorphisms and both uninvestigated and investigated dyspepsia has been reported in different studies from the United States<sup>[45]</sup>, Europe<sup>[46,47]</sup> and Asia<sup>[48,49]</sup>. Although the sample sizes remain relatively small in all of these studies, which may account for some of the variability in the associations detected, it is of note that the different classifications of dyspepsia also contribute to the diverse distribution of polymorphisms. In fact, while homozygous GNB $\beta$  825T allele was found to influence the susceptibility to EPS-like dyspepsia in a Japanese population of dyspeptics<sup>[49]</sup>, in a report from the United States, the same polymorphism was associated with postprandial dyspeptic symptoms and lower fasting gastric volumes<sup>[45]</sup>. Other studies failed to identify single genetic factors as a predominant factor in the pathogenesis of functional dyspepsia and symptom generation. As sympathetic adrenergic dysfunctions may affect both gastric sensitivity, Camilleri *et al.*<sup>[45]</sup> analyzed the role of presynaptic inhibi-

**Table 2 Overview of genetic association studies in functional dyspepsia**

Protein (gene)	Polymorphism	Finding	Ref.
TLR-2	(192-174)del	Protective for <i>Helicobacter pylori</i> -infected patients	[43]
SERT	SS variant	No association	[45]
5-HT1A	-Pro16Leu	No association	[46,47]
5-HT2A	-1438 G/A	No association	[46,47]
HTR3A	C178T	No association	[46,47]
GNB3	C825T	Risk factor	[48-50]
GNB3	CT and TT carriers	Risk factor for PDS	[48-50]
GNB3	CC and TT carriers	Risk factor	[48-50]
GNB3	C825T	Risk factor for EPS	[50]
GNB3	TT carriers	Risk factor for EPS	[50]
Presynaptic inhibitory $\alpha$ 2A and $\alpha$ 2C adrenoceptor	-1291 C>G ( $\alpha$ 2A) and -del 1322-325 ( $\alpha$ 2A)	No association	[46]
Fatty acid amide hydrolase	C385A	No association	[51]
TRPV1	G315C	Risk factor	[52]
Na (V) 1.8	SCN10A 3218CC	Protective	[53]

PDS: Post discharge surveillance; EPS: Encapsulating peritoneal sclerosis; TLR: Toll-like receptor; TRPV: Transient receptor potential vanilloid.

tory  $\alpha$ 2A and  $\alpha$ 2B adrenoceptors polymorphism, but they failed to find any significant association with dyspepsia symptoms. The same group also failed to find any significant association between the genetic variation in the endocannabinoid metabolism (*i.e.*, a SNP in the human fatty acid amide hydrolase gene-C385A-) and both impaired fundus accommodation and delayed gastric emptying in a small subgroup of dyspepsia patients<sup>[50]</sup>. Conversely, in a recent report, the involvement of the transient receptor potential vanilloid 1 (TRPV1) was investigated in a small population of Japanese dyspeptic patients. The authors found that in a population of 109 dyspeptics, individuals carrying the G315C polymorphism, known to affect the *TRPV1* gene and to alter its protein level, were at lower risk for both epigastric pain and postprandial distress syndrome. In addition, the authors showed that dyspeptics with that specific polymorphism had a baseline and cold water induced symptom severity<sup>[47]</sup>. Although this finding needs to be reproduced in different ethnic populations and validated on a large sample, it is of note that TRPV1 pathways may be ideal candidates as they are involved in nociception and in acid sensitivity, with the latter being claimed to have a role in the generation of dyspepsia symptoms. On the same line, the same group have found a significant association between a polymorphism of the tetrodotoxin-resistant (TTX-r) sodium channel Na (V) 1.8, a channel expressed by C-fibers and involved in nociception, and functional dyspepsia. Indeed, the SCN10A 3218 CC variant, which determines a lower activity of this Na (V)-channel, was significantly associated with a decreased risk for the de-

velopment of FD<sup>[52]</sup> (all data are summarized in Table 2).

### Post-infectious dyspepsia

By summarizing what we have described above, we can say that although it is unlikely that a single genetic factor causes FD, it is more likely that a genetic factor (or factors) modulates the risk of developing the abnormalities after exposure to one or more specific environmental factors<sup>[53]</sup>. This paradigm is well supported by the hypothesis of a post-infectious origin of dyspepsia. Indeed, it is now evident that in a subgroup of patients with functional dyspepsia, acute GI infections may precipitate symptoms. Data from the literature indicate that in presumed post-infectious dyspepsia patients there is an increased prevalence of impaired gastric accommodation to the meal that is likely dependent on inflammatory-induced impaired nitergic innervation of the gastric wall<sup>[54]</sup>. However, only one study systematically addressed the genetic contribution to post-infectious dyspepsia and the data presented are quite controversial since a significant association between macrophage migration inhibitory factor-173C and IL17F-7488T polymorphisms was only observed in the subgroup of patients with symptoms suggestive of EPS-like dyspepsia<sup>[55]</sup>. Indirect evidence for a role of inflammatory products gene in the genesis of FD is also suggested by a Japanese study in which a significant association between a COX1 polymorphism and EPS-like symptoms was observed in a subgroup of dyspeptic female patients<sup>[56]</sup>. However, a very recent study investigating the role of a polymorphism in the receptor for neuropeptide S receptor gene (NPSR1) that is involved in inflammation, anxiety and nociception failed to reveal any significant association with FD<sup>[57]</sup> (Table 3).

### Hypertrophic pyloric stenosis

Isolated HPS is a common condition characterized by the hypertrophy of the muscle surrounding the pylorus, with impaired sphincter relaxation and consequent gastric outlet obstruction which causes severe non-bilious vomiting. The etiology of this disease is still largely unknown; however, epidemiological studies indicate that genetic factors play an important role in the pathophysiology of this entity<sup>[58]</sup>. Krogh *et al.*<sup>[59]</sup> have shown a 200-fold increased risk of HPS in monozygotic twins and a 20-fold increased risk among siblings of affected children. In particular, different studies have demonstrated a higher prevalence of HPS in offspring of an affected mother compared with offspring of an affected father, suggesting a major role of maternal factors; however, this evidence was not confirmed in further studies<sup>[60]</sup>. In addition, HPS is associated with several well-defined genetic syndromes, sustaining the hypothesis that genetic factors are largely involved in the pathogenesis of this disease<sup>[58]</sup>.

A reduced expression of NOS1 was demonstrated at the mRNA level in pyloric tissue of patients with HPS, suggesting a pathogenic role of nNOS in this disease.

**Table 3** Overview of genetic association studies in post-infectious dyspepsia

Protein (gene)	Polymorphism	Finding	Ref.
<i>MIF</i>	-173C	Risk factor for EPS	[56]
<i>IL17</i>	-7488T	Risk factor for EPS	[56]
<i>COX-1</i>	T1676C	Risk factor for EPS	[57]
<i>NPS-R</i>	rs2609234, rs6972158, and rs1379928	No association	[58]

IL: Interleukin; COX-1: Cyclooxygenase-1; EPS: Encapsulating peritoneal sclerosis; MIF: Macrophage migration inhibitory factor.

For this reason, several researchers have analyzed the gene of *NOS1*; however, only a Chinese group found a linkage between HPS and NOS1a on chromosome 12q<sup>[61]</sup>, but this result was not confirmed by another study<sup>[62]</sup>. Moreover, the analysis of the complete coding region of *NOS1* in patients and controls revealed no significant difference, confirming that the impaired nNOS function in the pylorus of these patients is not related to a direct mutation of the gene encoding for *NOS1*<sup>[63]</sup>.

Several studies have demonstrated a linkage between HPS and different loci in affected patients of the same family, but larger studies on additional families were usually unsuccessful. A Chinese group described a linkage of a SNPs of chromosome 16 (16p12-p13) in 10 affected members of the same family, but this association was not observed in 10 additional families<sup>[64]</sup>. Moreover, the major candidate genes of this region encoding for *MYH11* and *GRIN2A*, proteins involved in smooth muscle relaxation, have not shown mutations<sup>[64]</sup>. In members of the same family, Everett *et al.*<sup>[65]</sup> found an association with *SLC7A5* (16q24), a gene which influences NO activity; however, they failed to confirm these data on additional 14 families. In a genome-wide linkage study, the same group demonstrated an association between HPS and some loci on chromosome 11q14-q22 and Xq23, both encoding for protein involved in the functioning of ion channels (*TRPC5* and *TRPC6*) in 81 families with 206 affected members<sup>[66,67]</sup>; however, a subsequent Chinese study failed to replicate the association with *TRPC6*<sup>[68]</sup>.

Finally, a Danish group in a recent genome-wide association study (GWAS) on 1001 individuals affected by HPS and 2401 healthy controls demonstrated an association between HPS and three SNPs of *MBNL1* and *NKX 2-5*, genes involved in the splicing and transcription processes<sup>[69]</sup>, but it was the first GWAS and these results could explain only a very small proportion of HPS cases (all reviewed associations are summarized in Table 4).

In conclusion, hypertrophic pyloric stenosis is a multifactorial disease and both genetic and environmental factors could contribute to the pathophysiology of this condition. Especially for the syndromic form of HPS, different mutations in different genes involved in different functions have been associated with the onset of this disease, but we are still far from a single unifying pathogenetic factor. Conversely, in the sporadic form of

**Table 4** Overview of genetic association studies in hypertrophic pyloric stenosis

Protein (gene)	Chromosome	Finding	Ref.
<i>NOS1</i>	12q	Association	[62,63]
<i>MYH11-GRIN2A</i>	16p12-p13	Association	[65]
<i>SLC7A5</i>	16q24	Association	[66]
<i>TRPC5 and TRPC6</i>	11q14-q22 and Xq23	Association	[67-69]

the disease, the contribution of the genetic background is even more scanty and complex and no clear triggering factors have been described yet, further sustaining the great heterogeneity of this disease.

## CONCLUSION

Several association studies have established a genetic component in the genesis of motility disorders of the upper gastrointestinal tract, like esophageal achalasia, functional dyspepsia and hypertrophic pyloric stenosis. Although candidate gene studies have identified a few gene polymorphisms that may be correlated with these syndromes, small sample size, lack of reproducibility in large data sets, and the unreliability of the clinical phenotype represent a major limit to identify a unifying factor in the pathophysiology of these syndromes in any of the reported polymorphisms.

Whether the genetic contribution plays a crucial role in the generation of upper GI tract symptoms therefore deserves further studies. More specifically, the recruitment of large case-control samples appears to be mandatory in order to provide a powerful tool for the identification of risk genes, especially for diseases like achalasia and functional dyspepsia, in which a multifactorial inheritance is assumed. In this direction, genome-wide association studies will allow for the unbiased and systematic identification of risk genes and may represent the greatest challenge for all future studies of upper GI tract motility disorders.

## REFERENCES

- 1 **Goyal RK**, Hirano I. The enteric nervous system. *N Engl J Med* 1996; **334**: 1106-1115 [PMID: 8598871 DOI: 10.1056/NEJM199604253341707]
- 2 **Wood JD**. Neural and humoral regulation of gastrointestinal motility. In: Schuster MM, Crowell MD, Koch KL, editors. *Gastrointestinal motility in health and disease*. London: BC Decker, 2002: 19-42
- 3 **Locke GR**, Zinsmeister AR, Talley NJ, Fett SL, Melton LJ. Familial association in adults with functional gastrointestinal disorders. *Mayo Clin Proc* 2000; **75**: 907-912 [PMID: 10994826 DOI: 10.4065/75.9.907]
- 4 **Tryhus MR**, Davis M, Griffith JK, Ablin DS, Gogel HK. Familial achalasia in two siblings: significance of possible hereditary role. *J Pediatr Surg* 1989; **24**: 292-295 [PMID: 2709295 DOI: 10.1016/S0022-3468(89)80016-8]
- 5 **Morris-Yates A**, Talley NJ, Boyce PM, Nandurkar S, Andrews G. Evidence of a genetic contribution to functional bowel disorder. *Am J Gastroenterol* 1998; **93**: 1311-1317 [PMID: 9707057 DOI: 10.1111/j.1572-0241.1998.440.j.x]

- 6 **Stein DT**, Knauer CM. Achalasia in monozygotic twins. *Dig Dis Sci* 1982; **27**: 636-640 [PMID: 7200858 DOI: 10.1007/BF01297220]
- 7 **Lenbo T**, Zaman MS, Chavez NF, Krueger R, Jones MP, Talley NJ. Concordance of IBS among monozygotic and dizygotic twins. *Gastroenterol* 2001; **120** (suppl 1): A-66
- 8 **Richter JE**. Oesophageal motility disorders. *Lancet* 2001; **358**: 823-828 [PMID: 11564508 DOI: 10.1016/S0140-6736(01)05973-6]
- 9 **Goldblum JR**, Rice TW, Richter JE. Histopathologic features in esophagomyotomy specimens from patients with achalasia. *Gastroenterology* 1996; **111**: 648-654 [PMID: 8780569 DOI: 10.1053/gast.1996.v111.pm8780569]
- 10 **Clark SB**, Rice TW, Tubbs RR, Richter JE, Goldblum JR. The nature of the myenteric infiltrate in achalasia: an immunohistochemical analysis. *Am J Surg Pathol* 2000; **24**: 1153-1158 [PMID: 10935657 DOI: 10.1097/00000478-200008000-00014]
- 11 **Mearin F**, Mourelle M, Guarner F, Salas A, Riveros-Moreno V, Moncada S, Malagelada JR. Patients with achalasia lack nitric oxide synthase in the gastro-oesophageal junction. *Eur J Clin Invest* 1993; **23**: 724-728 [PMID: 7508398 DOI: 10.1111/j.1365-2362.1993.tb01292.x]
- 12 **Konturek JW**, Thor P, Lukaszuk A, Gabryelewicz A, Konturek SJ, Domschke W. Endogenous nitric oxide in the control of esophageal motility in humans. *J Physiol Pharmacol* 1997; **48**: 201-209 [PMID: 9223025]
- 13 **Goyal RK**, Rattan S, Said SI. VIP as a possible neurotransmitter of non-cholinergic non-adrenergic inhibitory neurones. *Nature* 1980; **288**: 378-380 [PMID: 6107863 DOI: 10.1038/288378a0]
- 14 **Mayberry JF**, Atkinson M. A study of swallowing difficulties in first degree relatives of patients with achalasia. *Thorax* 1985; **40**: 391-393 [PMID: 4023994 DOI: 10.1136/thx.40.5.391]
- 15 **Gockel HR**, Schumacher J, Gockel I, Lang H, Haaf T, Nöthen MM. Achalasia: will genetic studies provide insights? *Hum Genet* 2010; **128**: 353-364 [PMID: 20700745 DOI: 10.1007/s00439-010-0874-8]
- 16 **Okawada M**, Okazaki T, Yamataka A, Lane GJ, Miyano T. Down's syndrome and esophageal achalasia: a rare but important clinical entity. *Pediatr Surg Int* 2005; **21**: 997-1000 [PMID: 16261371 DOI: 10.1007/s00383-005-1528-0]
- 17 **Allgrove J**, Clayden GS, Grant DB, Macaulay JC. Familial glucocorticoid deficiency with achalasia of the cardia and deficient tear production. *Lancet* 1978; **1**: 1284-1286 [PMID: 78049 DOI: 10.1016/S0140-6736(78)91268-0]
- 18 **Barnett JL**, McDonnell WM, Appelman HD, Dobbins WO. Familial visceral neuropathy with neuronal intranuclear inclusions: diagnosis by rectal biopsy. *Gastroenterology* 1992; **102**: 684-691 [PMID: 1310083]
- 19 **Khalifa MM**. Familial achalasia, microcephaly, and mental retardation. Case report and review of literature. *Clin Pediatr (Phila)* 1988; **27**: 509-512 [PMID: 3048841 DOI: 10.1177/00092288802701009]
- 20 **de la Concha EG**, Fernandez-Arquero M, Conejero L, Lazaro F, Mendoza JL, Sevilla MC, Diaz-Rubio M, Ruiz de Leon A. Presence of a protective allele for achalasia on the central region of the major histocompatibility complex. *Tissue Antigens* 2000; **56**: 149-153 [PMID: 11019915 DOI: 10.1034/j.1399-0039.2000.560206.x]
- 21 **Jones DB**, Mayberry JF, Rhodes J, Munro J. Preliminary report of an association between measles virus and achalasia. *J Clin Pathol* 1983; **36**: 655-657 [PMID: 6853731 DOI: 10.1136/jcp.36.6.655]
- 22 **Robertson CS**, Martin BA, Atkinson M. Varicella-zoster virus DNA in the oesophageal myenteric plexus in achalasia. *Gut* 1993; **34**: 299-302 [PMID: 8386130 DOI: 10.1136/gut.34.3.299]
- 23 **Birgisson S**, Galinski MS, Goldblum JR, Rice TW, Richter JE. Achalasia is not associated with measles or known herpes and human papilloma viruses. *Dig Dis Sci* 1997; **42**: 300-306 [PMID: 9052510]
- 24 **Facco M**, Brun P, Baesso I, Costantini M, Rizzetto C, Berto A, Baldan N, Palù G, Semenzato G, Castagliuolo I, Zaninotto G. T cells in the myenteric plexus of achalasia patients show a skewed TCR repertoire and react to HSV-1 antigens. *Am J Gastroenterol* 2008; **103**: 1598-1609 [PMID: 18557707 DOI: 10.1111/j.1572-0241.2008.01956.x]
- 25 **De la Concha EG**, Fernandez-Arquero M, Mendoza JL, Conejero L, Figueredo MA, Perez de la Serna J, Diaz-Rubio M, Ruiz de Leon A. Contribution of HLA class II genes to susceptibility in achalasia. *Tissue Antigens* 1998; **52**: 381-384 [PMID: 9820602 DOI: 10.1111/j.1399-0039.1998.tb03059.x]
- 26 **Verne GN**, Hahn AB, Pineau BC, Hoffman BJ, Wojciechowski BW, Wu WC. Association of HLA-DR and -DQ alleles with idiopathic achalasia. *Gastroenterology* 1999; **117**: 26-31 [PMID: 10381906 DOI: 10.1016/S0016-5085(99)70546-9]
- 27 **Wong RK**, Maydonovitch CL, Metz SJ, Baker JR. Significant DQw1 association in achalasia. *Dig Dis Sci* 1989; **34**: 349-352 [PMID: 2920639 DOI: 10.1007/BF01536254]
- 28 **Santiago JL**, Martínez A, Benito MS, Ruiz de León A, Mendoza JL, Fernández-Arquero M, Figueredo MA, de la Concha EG, Urcelay E. Gender-specific association of the PTPN22 C1858T polymorphism with achalasia. *Hum Immunol* 2007; **68**: 867-870 [PMID: 17961776 DOI: 10.1016/j.humimm.2007.07.005]
- 29 **Núñez C**, García-González MA, Santiago JL, Benito MS, Mearin F, de la Concha EG, de la Serna JP, de León AR, Urcelay E, Vigo AG. Association of IL10 promoter polymorphisms with idiopathic achalasia. *Hum Immunol* 2011; **72**: 749-752 [PMID: 21641950 DOI: 10.1016/j.humimm.2011.05.017]
- 30 **de León AR**, de la Serna JP, Santiago JL, Sevilla C, Fernández-Arquero M, de la Concha EG, Núñez C, Urcelay E, Vigo AG. Association between idiopathic achalasia and IL23R gene. *Neurogastroenterol Motil* 2010; **22**: 734-738, e218 [PMID: 20367798 DOI: 10.1111/j.1365-2982.2010.01497.x]
- 31 **Tøttrup A**, Svane D, Forman A. Nitric oxide mediating NANC inhibition in opossum lower esophageal sphincter. *Am J Physiol* 1991; **260**: G385-G389 [PMID: 2003605]
- 32 **Croen KD**. Evidence for antiviral effect of nitric oxide. Inhibition of herpes simplex virus type 1 replication. *J Clin Invest* 1993; **91**: 2446-2452 [PMID: 8390481 DOI: 10.1172/JCI116479]
- 33 **Delgado M**, Pozo D, Ganea D. The significance of vasoactive intestinal peptide in immunomodulation. *Pharmacol Rev* 2004; **56**: 249-290 [PMID: 15169929 DOI: 10.1124/pr.56.2.7]
- 34 **Mearin F**, García-González MA, Strunk M, Zárata N, Malagelada JR, Lanás A. Association between achalasia and nitric oxide synthase gene polymorphisms. *Am J Gastroenterol* 2006; **101**: 1979-1984 [PMID: 16848803 DOI: 10.1111/j.1572-0241.2006.00762.x]
- 35 **Vigo AG**, Martínez A, de la Concha EG, Urcelay E, Ruiz de León A. Suggested association of NOS2A polymorphism in idiopathic achalasia: no evidence in a large case-control study. *Am J Gastroenterol* 2009; **104**: 1326-1327 [PMID: 19337240 DOI: 10.1038/ajg.2009.72]
- 36 **Alahdab YO**, Eren F, Giral A, Gunduz F, Kedrah AE, Atug O, Yilmaz Y, Kalayci O, Kalayci C. Preliminary evidence of an association between the functional c-kit rs6554199 polymorphism and achalasia in a Turkish population. *Neurogastroenterol Motil* 2012; **24**: 27-30 [PMID: 21951831 DOI: 10.1111/j.1365-2982.2011.01793.x]
- 37 **Paladini F**, Cocco E, Cascino I, Belfiore F, Badiali D, Piretta L, Alghisi F, Anzini F, Fiorillo MT, Corazzari E, Sorrentino R. Age-dependent association of idiopathic achalasia with vasoactive intestinal peptide receptor 1 gene. *Neurogastroenterol Motil* 2009; **21**: 597-602 [PMID: 19309439 DOI: 10.1111/j.1365-2982.2009.01284.x]
- 38 **Sarnelli G**. Impact of genetic polymorphisms on the pathogenesis of achalasia: an age-dependent paradigm? *Neurogastroenterol Motil* 2009; **21**: 575-578 [PMID: 19646069 DOI: 10.1111/j.1365-2982.2009.01319.x]

- 39 **Tack J**, Bisschops R, Sarnelli G. Pathophysiology and treatment of functional dyspepsia. *Gastroenterology* 2004; **127**: 1239-1255 [PMID: 15481001 DOI: 10.1053/j.gastro.2004.05.030]
- 40 **Tack J**, Talley NJ, Camilleri M, Holtmann G, Hu P, Malagelada JR, Stanghellini V. Functional gastroduodenal disorders. *Gastroenterology* 2006; **130**: 1466-1479 [PMID: 16678560 DOI: 10.1053/j.gastro.2005.11.059]
- 41 **Holtmann G**, Liebrechts T, Siffert W. Molecular basis of functional gastrointestinal disorders. *Best Pract Res Clin Gastroenterol* 2004; **18**: 633-640 [PMID: 15324704 DOI: 10.1016/j.bpg.2004.04.006]
- 42 **Tahara T**, Shibata T, Wang F, Yamashita H, Hirata I, Arisawa T. Genetic Polymorphisms of Molecules Associated with Innate Immune Responses, TRL2 and MBL2 Genes in Japanese Subjects with Functional Dyspepsia. *J Clin Biochem Nutr* 2010; **47**: 217-223 [PMID: 21103030 DOI: 10.3164/jcbn.10-40]
- 43 **Tack J**, Sarnelli G. Serotonergic modulation of visceral sensation: upper gastrointestinal tract. *Gut* 2002; **51** Suppl 1: i77-i80 [PMID: 12077073 DOI: 10.1136/gut.51.suppl\_1.i77]
- 44 **Camilleri M**, Atanasova E, Carlson PJ, Ahmad U, Kim HJ, Viramontes BE, McKinzie S, Urrutia R. Serotonin-transporter polymorphism pharmacogenetics in diarrhea-predominant irritable bowel syndrome. *Gastroenterology* 2002; **123**: 425-432 [PMID: 12145795 DOI: 10.1053/gast.2002.34780]
- 45 **Camilleri CE**, Carlson PJ, Camilleri M, Castillo EJ, Locke GR, Geno DM, Stephens DA, Zinsmeister AR, Urrutia R. A study of candidate genotypes associated with dyspepsia in a U.S. community. *Am J Gastroenterol* 2006; **101**: 581-592 [PMID: 16464220 DOI: 10.1111/j.1572-0241.2006.00481.x]
- 46 **van Lelyveld N**, Linde JT, Schipper M, Samsom M. Candidate genotypes associated with functional dyspepsia. *Neurogastroenterol Motil* 2008; **20**: 767-773 [PMID: 18331431]
- 47 **Holtmann G**, Siffert W, Haag S, Mueller N, Langkafel M, Senf W, Zotz R, Talley NJ. G-protein beta 3 subunit 825 CC genotype is associated with unexplained (functional) dyspepsia. *Gastroenterology* 2004; **126**: 971-979 [PMID: 15057736 DOI: 10.1053/j.gastro.2004.01.006]
- 48 **Tahara T**, Arisawa T, Shibata T, Wang F, Nakamura M, Sakata M, Hirata I, Nakano H. Homozygous 825T allele of the GNB3 protein influences the susceptibility of Japanese to dyspepsia. *Dig Dis Sci* 2008; **53**: 642-646 [PMID: 17717746 DOI: 10.1007/s10620-007-9923-0]
- 49 **Oshima T**, Nakajima S, Yokoyama T, Toyoshima F, Sakurai J, Tanaka J, Tomita T, Kim Y, Hori K, Matsumoto T, Miwa H. The G-protein beta3 subunit 825 TT genotype is associated with epigastric pain syndrome-like dyspepsia. *BMC Med Genet* 2010; **11**: 13 [PMID: 20102604 DOI: 10.1186/1471-2350-11-13]
- 50 **Camilleri M**, Carlson P, McKinzie S, Grudell A, Busciglio I, Burton D, Baxter K, Ryks M, Zinsmeister AR. Genetic variation in endocannabinoid metabolism, gastrointestinal motility, and sensation. *Am J Physiol Gastrointest Liver Physiol* 2008; **294**: G13-G19 [PMID: 17962356 DOI: 10.1152/ajpgi.00371.2007]
- 51 **Tahara T**, Shibata T, Nakamura M, Yamashita H, Yoshioka D, Hirata I, Arisawa T. Homozygous TRPV1 315C influences the susceptibility to functional dyspepsia. *J Clin Gastroenterol* 2010; **44**: e1-e7 [PMID: 19770677]
- 52 **Arisawa T**, Tahara T, Shiroeda H, Minato T, Matsue Y, Saito T, Fukuyama T, Otsuka T, Fukumura A, Nakamura M, Shibata T. Genetic polymorphisms of SCN10A are associated with functional dyspepsia in Japanese subjects. *J Gastroenterol* 2013; **48**: 73-80 [PMID: 22618805 DOI: 10.1007/s00535-012-0602-3]
- 53 **Holtmann G**, Talley NJ. Hypothesis driven research and molecular mechanisms in functional dyspepsia: the beginning of a beautiful friendship in research and practice? *Am J Gastroenterol* 2006; **101**: 593-595 [PMID: 16542295 DOI: 10.1111/j.1572-0241.2006.00480.x]
- 54 **Tack J**, Demedts I, Dehondt G, Caenepeel P, Fischler B, Zandecki M, Janssens J. Clinical and pathophysiological characteristics of acute-onset functional dyspepsia. *Gastroenterology* 2002; **122**: 1738-1747 [PMID: 12055579 DOI: 10.1053/gast.2002.33663]
- 55 **Arisawa T**, Tahara T, Shibata T, Nagasaka M, Nakamura M, Kamiya Y, Fujita H, Yoshioka D, Arima Y, Okubo M, Hirata I, Nakano H. Genetic polymorphisms of molecules associated with inflammation and immune response in Japanese subjects with functional dyspepsia. *Int J Mol Med* 2007; **20**: 717-723 [PMID: 17912466]
- 56 **Arisawa T**, Tahara T, Shibata T, Nagasaka M, Nakamura M, Kamiya Y, Fujita H, Yoshioka D, Arima Y, Okubo M, Hirata I, Nakano H. Genetic polymorphisms of cyclooxygenase-1 (COX-1) are associated with functional dyspepsia in Japanese women. *J Womens Health (Larchmt)* 2008; **17**: 1039-1043 [PMID: 18582172 DOI: 10.1089/jwh.2007.0720]
- 57 **Camilleri M**, Carlson P, Zinsmeister AR, McKinzie S, Busciglio I, Burton D, Zucchelli M, D'Amato M. Neuropeptide S receptor induces neuropeptide expression and associates with intermediate phenotypes of functional gastrointestinal disorders. *Gastroenterology* 2010; **138**: 98-107. e4 [PMID: 19732772 DOI: 10.1053/j.gastro.2009.08.051]
- 58 **Peeters B**, Benninga MA, Hennekam RC. Infantile hypertrophic pyloric stenosis--genetics and syndromes. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 646-660 [PMID: 22777173]
- 59 **Krogh C**, Fischer TK, Skotte L, Biggar RJ, Øyen N, Skytthe A, Goertz S, Christensen K, Wohlfahrt J, Melbye M. Familial aggregation and heritability of pyloric stenosis. *JAMA* 2010; **303**: 2393-2399 [PMID: 20551410 DOI: 10.1001/jama.2010.784]
- 60 **Mitchell LE**, Risch N. The genetics of infantile hypertrophic pyloric stenosis. A reanalysis. *Am J Dis Child* 1993; **147**: 1203-1211 [PMID: 8237916]
- 61 **Chung E**, Curtis D, Chen G, Marsden PA, Twells R, Xu W, Gardiner M. Genetic evidence for the neuronal nitric oxide synthase gene (NOS1) as a susceptibility locus for infantile pyloric stenosis. *Am J Hum Genet* 1996; **58**: 363-370 [PMID: 8571963]
- 62 **Söderhäll C**, Nordenskjöld A. Neuronal nitric oxide synthase, nNOS, is not linked to infantile hypertrophic pyloric stenosis in three families. *Clin Genet* 1998; **53**: 421-422 [PMID: 9660065 DOI: 10.1111/j.1399-0004.1998.tb02758.x]
- 63 **Serra A**, Schuchardt K, Genuneit J, Leriche C, Fitz G. Genomic variants in the coding region of neuronal nitric oxide synthase (NOS1) in infantile hypertrophic pyloric stenosis. *J Pediatr Surg* 2011; **46**: 1903-1908 [PMID: 22008325 DOI: 10.1016/j.jpedsurg.2011.05.021]
- 64 **Capon F**, Reece A, Ravindrarajah R, Chung E. Linkage of monogenic infantile hypertrophic pyloric stenosis to chromosome 16p12-p13 and evidence for genetic heterogeneity. *Am J Hum Genet* 2006; **79**: 378-382 [PMID: 16826529 DOI: 10.1086/505952]
- 65 **Everett KV**, Capon F, Georgoula C, Chioza BA, Reece A, Jaswon M, Pierro A, Puri P, Gardiner RM, Chung EM. Linkage of monogenic infantile hypertrophic pyloric stenosis to chromosome 16q24. *Eur J Hum Genet* 2008; **16**: 1151-1154 [PMID: 18478043 DOI: 10.1038/ejhg.2008.86]
- 66 **Everett KV**, Chioza BA, Georgoula C, Reece A, Capon F, Parker KA, Cord-Udy C, McKeigue P, Mitton S, Pierro A, Puri P, Mitchison HM, Chung EM, Gardiner RM. Genome-wide high-density SNP-based linkage analysis of infantile hypertrophic pyloric stenosis identifies loci on chromosomes 11q14-q22 and Xq23. *Am J Hum Genet* 2008; **82**: 756-762 [PMID: 18308288 DOI: 10.1016/j.ajhg.2007.12.023]
- 67 **Everett KV**, Chioza BA, Georgoula C, Reece A, Gardiner RM, Chung EM. Infantile hypertrophic pyloric stenosis: evaluation of three positional candidate genes, TRPC1, TRPC5 and TRPC6, by association analysis and re-sequencing. *Hum Genet* 2009; **126**: 819-831 [PMID: 19701773 DOI: 10.1007/s00439-009-0735-5]

- 68 **Ju JJ**, Gao H, Li H, Lu Y, Wang LL, Yuan ZW. No association between the SNPs (rs56134796; rs3824934; rs41302375) in the TRPC6 gene promoter and infantile hypertrophic pyloric stenosis in Chinese people. *Pediatr Surg Int* 2011; **27**: 1267-1270 [PMID: 21822655 DOI: 10.1007/s00383-011-2961-x]
- 69 **Feenstra B**, Geller F, Krogh C, Hollegaard MV, Gørtz S, Boyd HA, Murray JC, Hougaard DM, Melbye M. Common variants near MBNL1 and NKX2-5 are associated with infantile hypertrophic pyloric stenosis. *Nat Genet* 2012; **44**: 334-337 [PMID: 22306654 DOI: 10.1038/ng.1067]

**P- Reviewers:** Akiba Y, Lindberg G **S- Editor:** Wen LL  
**L- Editor:** A **E- Editor:** Lu YJ



## Pancreatic cancer diagnosis by free and exosomal miRNA

Margot Zöller

Margot Zöller, Department of Tumor Cell Biology, University Hospital of Surgery, D 69120 Heidelberg, Germany  
Author contributions: Zöller M solely contributed to this paper.  
Supported by The Wilhelm Sander Stiftung  
Correspondence to: Dr. Margot Zöller, Department of Tumor Cell Biology, University Hospital of Surgery, Im Neuenheimer Feld 365, D 69120 Heidelberg, Germany. [margot.zoeller@uni-heidelberg.de](mailto:margot.zoeller@uni-heidelberg.de)  
Telephone: +49-6221-565146 Fax: +49-6221-565199  
Received: May 25, 2013 Revised: August 1, 2013  
Accepted: September 18, 2013  
Published online: November 15, 2013

### Abstract

Patients with pancreatic adenocarcinoma (PaCa) have a dismal prognosis. This is in part due to late diagnosis prohibiting surgical intervention, which provides the only curative option as PaCa are mostly chemo- and radiation resistance. Hope is raised on a reliable non-invasive/minimally invasive diagnosis that is still missing. Recently two diagnostic options are discussed, serum MicroRNA (miRNA) and serum exosomes. Serum miRNA can be free or vesicle-, particularly, exosomes-enclosed. This review will provide an overview on the current state of the diagnostic trials on free serum miRNA and proceed with an introduction of exosomes that use as a diagnostic tool in serum and other body fluids has not received sufficient attention, although serum exosome miRNA in combination with protein marker expression likely will increase the diagnostic and prognostic power. By their crosstalk with host cells, which includes binding-initiated signal transduction, as well as reprogramming target cells *via* the transfer of proteins, mRNA and miRNA exosomes are suggested to become a most powerful therapeutics. I will discuss which hurdles have still to be taken as well as the different modalities, which can be envisaged to make therapeutic use of exosomes. PaCa are known to most intensely crosstalk with the host as apparent by desmoplasia and frequent paraneoplastic syndromes. Thus, there is hope that the therapeutic application of

exosomes brings about a major breakthrough.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Pancreatic cancer; Exosomes; MicroRNA; Diagnosis; Serum

**Core tip:** Patients with pancreatic adenocarcinoma have a dismal prognosis due to late diagnosis prohibiting surgical intervention, which is further burdened by chemo- and radiation resistance. Hope is raised on a minimally invasive diagnosis that is still missing. Recently two diagnostic options are discussed, serum microRNA (miRNA) and serum exosomes. Serum miRNA can be free or vesicle-, particularly, exosomes-enclosed. This review presents an overview on the current state on miRNA as a cancer diagnostics and discusses arguments in favor of tumor exosomes as a diagnostic tool that additionally could provide a powerful therapeutic option in the near future.

Zöller M. Pancreatic cancer diagnosis by free and exosomal miRNA. *World J Gastrointest Pathophysiol* 2013; 4(4): 74-90  
Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/74.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.74>

### INTRODUCTION

Pancreatic adenocarcinoma (PaCa) ranks fourth in mortality among cancer-related deaths. With an overall 5-year survival rate of below 1% and a mean survival time of 4-6 mo it is the deadliest cancer<sup>[1,2]</sup>. There has been considerable progress in the treatment of patients with early stage PaCa. But late initial diagnosis that prohibits resection, chemotherapy and radiation resistance and the early metastatic spread of PaCa account for the non-satisfactory progress in therapy<sup>[3,4]</sup>. Thus, research has focused on defining a reliable non-invasive or minimally invasive diagnosis. So far, serum markers allowing for a non-invasive diagnosis and follow up studies are rare.

CA19-9 is still the most reliable diagnostic serum marker, but should be used in conjunction with other diagnostic tools. Additional markers are carcino-embryonic antigen (CEA), CA125 and CA242, their specificity and particularly sensitivity being below that of CA19-9<sup>[5-7]</sup>. However, recently, two non-invasive diagnostic tools have come into focus. First, serum microRNA (miRNA) was repeatedly described to allow for differential diagnosis of cancer, where PaCa patients' serum miRNA might allow differentiating between benign and malignant tumors as well as inflammation<sup>[8,9]</sup>. Second, tumor-derived exosomes are readily detected in body fluids. Their protein, mRNA and miRNA profiles might well serve as diagnostic tools<sup>[10]</sup>. In addition, exosomes are hotly debated as potent therapeutics<sup>[11-13]</sup>.

## TUMOR DIAGNOSIS AND miRNA

### Recovery of non-coding RNA in body fluids

A new class of small noncoding RNA known as miRNA endogenously regulates gene expression at the posttranscriptional level<sup>[14]</sup>. miRNA range in size from 19 to 25 nucleotides. They regulate translation and degradation of mRNA through base pairing to complementary sites mostly in the untranslated region<sup>[15]</sup>. miRNAs constitute only 1%-3% of the human genome, but control about 30% of the coding genes<sup>[16]</sup>, most miRNAs controlling multiple mRNAs<sup>[17]</sup>. miRNA biogenesis is a multistep process, where a long primary transcript (pri-miR) is processed into a 70-100 nt hairpin precursor pre-miR. The pre-miR is translocated to the cytoplasm, where it is cleaved by the ribonuclease Dicer into a mature miR duplex, which is incorporated into the RNA-induced silencing complex (RISC) resulting in degradation of the duplex and binding to target mRNA by complementary base pairing at the 3'-untranslated region<sup>[14]</sup>. Seed sequence complementarity of about 7 base pairs enables miRNA to bind the target mRNA, which results in inhibition of translation or a reduction in mRNA stability<sup>[18]</sup>. miRNA in the serum may derive from necrosis, apoptosis<sup>[19]</sup> or be actively released in microvesicles<sup>[20]</sup>. Free extracellular miRNA is associated with argonaute proteins (Ago) The Ago2-miRNA complex accounts for the stability of the free miRNA<sup>[21,22]</sup>.

In advance of discussing serum miRNA as a potential diagnostic tool, it should be stated that data normalization is an important factor and that due to any fluctuation, epigenetic factors or others, like age, gender, diurnal changes and many more, cohort sizes should be large. Also due to these variabilities, it is very unlikely that a set of reference housekeeping miRNA with universal applicability can be identified<sup>[23,24]</sup>. Furthermore, it has to be kept in mind that most miRNAs regulate more than one mRNA. Thus, in turn, a given miRNA may be deregulated in multiple diseases, including different types of cancer<sup>[25,26]</sup>.

### miRNA and cancer

The increased knowledge on miRNA greatly fostered

progress in oncology, where miRNA could be linked to prognosis, disease progression, local recurrence and metastasis<sup>[24,27-29]</sup>. As summarized in a recent review<sup>[30]</sup> miRNA plays an important role in epithelial-mesenchymal transition (EMT), maintenance of cancer stem cells as well as tumor invasion and migration. EMT is regulated by the miR-200 family, miR-141, miR-429 and miR-205. The expression level of miR-200 negatively correlates with zinc finger E-box-binding homeobox (ZEB)1 and 2, which inhibit E-cadherin expression<sup>[31]</sup>. In PaCa, downregulation of miR-30 correlates with EMT, targets being vimentin and snail-1<sup>[32]</sup>. Examples for the involvement of miRNAs in cancer stem cell (CSC) control, including pancreatic cancer, are the tumor suppressor miR-34 that regulates Notch and Bcl2<sup>[33,34]</sup> and miR-21 that correlates with chemoresistance<sup>[35]</sup>. Instead, miR-9, regulating E-cadherin expression, is suggested to be of major importance for metastasis-associated mobility and invasiveness<sup>[36,37]</sup>. miR-34a overexpression can inhibit metastasis by regulating CD44<sup>[38]</sup> and miR-340 suppresses invasion and metastasis by regulating c-Met and *via* c-Met MMP2 and MMP9<sup>[39,40]</sup>.

For PaCa Jamieson *et al*<sup>[41]</sup> performed microarray analysis on resected PaCa tissue on a cohort of 48 and 24 patients. They describe associations with lymph node involvement, tumor grading and overall survival, where high expression of miR-21 and low expression of miR-34a significantly correlated with poor survival. Additional studies on PaCa tissue, non-transformed pancreatic ductal cells, CP samples and on PaCa culture lines by array or RT-PCR<sup>[42-46]</sup> have been summarized by Li *et al*<sup>[47]</sup>, which also provides an overview on their function as tumor suppressors (miR-15a, miR-34a, miR-96, miR-375) or oncogenes (miR-27a, miR-132, miR-155, miR-194, miR-200b, miR-220c, miR-429, miR-212, miR-214, miR-301a, miR-421, miR-483-3p) and potential molecular targets, which include besides others WNT3A, p53, K-Ras, Akt, 14-3-3zeta and Smad4<sup>[43,48-56]</sup>.

Taken together, there is increasing evidence that miRNA plays a central role in carcinogenesis and tumor progression, where the recovery of miRNA in body fluids may, additionally, provide a minimally invasive diagnostic tool. This has created hope particularly for most deadly PaCa, late diagnosis considerably contributing to the poor prognosis.

### Serum miRNA as a diagnostic tool in pancreatic cancer

The stability of free miRNA in serum and other body fluids has fostered the hope for a minimally invasive diagnostic tool that may also be of prognostic value<sup>[57-59]</sup>, which meanwhile has been experimentally supported for different types of cancer<sup>[60-62]</sup> including PaCa, where it will be particularly important as late diagnosis prohibits a curative intervention.

In an earlier study 4 miRNAs, miR-21, miR-210, miR-155 and miR-196a have been found to differentiate PaCa patients' serum from that of healthy controls, where miR-155 is a biomarker of early PaCa and miR-196a cor-

relates with progression<sup>[63]</sup>. Evaluating a combination of CA19-9 with plasma miRNA in PaCa revealed 4 miRNA, miR-155, miR-181a, miR-181b and miR-196a, to differ significantly from healthy donors' miRNA, where only miR-16 and miR-196a allowed for discrimination from chronic pancreatitis (CP). Including CA19-9 increased sensitivity and specificity of the analysis, 85.2% of PaCa samples being positive even at stage 1<sup>[64]</sup>. An elegant recent study on serum miRNA in PaCa based on sequencing of pooled samples, a selection phase based on quantitative reverse transcriptase PCR (qRT-PCR) followed by a testing phase revealed upregulation of miR-20a, miR-21, miR-24, miR-25, miR-99a, miR-185 and miR-191 in the serum of PaCa patients as compared to healthy controls. The authors also confirmed that these 7 miRNA allowed for differentiation towards CP, where expression in CP did not differ significantly from that of healthy donors<sup>[9]</sup>. Additional studies mostly confirmed abundance of miR-21, miR-155, miR-196a, miR-210 and miR-16 in PaCa patients' sera<sup>[65-69]</sup>. A statistical meta-analysis, which includes 9 studies, from which 5 were performed with tissue and 4 with serum or plasma<sup>[9,63,64]</sup> suggests a potential role for miRNA assays in screening for and confirming PaCa diagnosis<sup>[70]</sup>. However, the authors also point out that none of these miRNA is selective for PaCa. An additional concern should be mentioned. A differential analysis of free versus vesicular, particularly, exosomal miRNA in the serum of PaCa patients appears to be missing. An exosomal miRNA analysis may well be advantageous as exosomal miRNA derives from living cells, whereas free miRNA may mostly derive from dead cells and thus could significantly change particularly during therapy or in late stage PaCa<sup>[19,71,72]</sup>. Serum exosome screening could have an additional advantage. Membrane integrated PaCa markers will be recovered on exosomes, thus allowing for a concomitant screening of miRNA and proteins.

## EXOSOMES AS A DIAGNOSTIC TOOL

Exosomes are small 40-100 nm vesicles, which derive from the fusion of the intraluminal vesicles of multivesicular bodies (MVB) with the plasma membrane<sup>[10,73]</sup>. Their homogeneous size is one of the major criteria to differentiate exosomes from apoptotic blebs, microparticles and microvesicles, which vary in size<sup>[74]</sup>. Exosomes are composed of a lipid bilayer; they contain selected proteins, mRNA and miRNA<sup>[75]</sup>. Exosomes are secreted by many cells and abundantly by tumor cells<sup>[76]</sup> and are found in all body fluids<sup>[77]</sup>. Due to their presence in all body fluids and the expression of selected markers, exosomes are suggested to be optimal candidates for non-invasive diagnosis<sup>[78,79]</sup>. Exosomal proteins, mRNA and miRNA being functionally active<sup>[80,81]</sup> and transferred into target cells<sup>[13,81-85]</sup>, exosomes are the most important intercellular communicators<sup>[75]</sup> and are suggested to become a very powerful therapeutic tool<sup>[12,86,87]</sup>. To reach the goals of exosomes as diagnostics and therapeutics great efforts are

taken to elaborate the prerequisites, such as exosome assembly and exosomal message transfer.

### Exosome assembly and secretion

It is well known that the relative abundance of proteins, mRNA and miRNAs differs between exosomes and donor cells, which implies active sorting into MVB. Indeed, the sorting of proteins into exosomes is a highly regulated process, where monoubiquitinylation as well as the endosomal sorting complex required for transport (ESCRT) play a role, some components of ESCRT, like Tsg101 and Alix being recovered in exosomes. The ESCRT machinery consists of 3 complexes, ESCRT I, II and III, where Tsg1 in the ESCRT complex I binds ubiquitinated protein and recruits ESCRT II. ESCRT III becomes recruited *via* ESCRT II or Alix. ESCRT III recruits a deubiquitinating enzyme that removes the ubiquitin tag from the cargo proteins prior to sorting into MVB<sup>[88,89]</sup>. However, not all proteins require the ESCRT complex for incorporation into exosomes. Alternatively, proteins in detergent resistant membrane complexes can become incorporated into MVB like MHC II molecules in dendritic cells<sup>[90]</sup>. Lipid affinity also can account for MVB incorporation<sup>[91]</sup>. Tetraspanins and other proteins with high affinity for cholesterol and sphingolipids are partitioned into membrane domains which according to their physical properties are prone for internalization<sup>[92-95]</sup>. Proteins also may become recruited by associated proteins such as integrins associated with tetraspanins or the transferrin receptor (TfR), which associates with heat shock proteins (HSP)<sup>[94]</sup>. In particular for tetraspanin-associated molecules it has been described that protein complexes rather than singular molecules are recruited into exosomes. This complex binding severely influences exosome targeting and the crosstalk with target structures<sup>[96-98]</sup>. Besides members of the tetraspanin family (CD9, CD63, CD81, CD82, CD151, Tspan8), where tetraspanins are constitutive components of exosomes<sup>[91,99]</sup> and are frequently used to differentiate exosomes from other extracellular vesicles<sup>[75,91]</sup>, additional molecules most abundantly recovered in exosomes are HSP<sup>[100,101]</sup>, proteases<sup>[102,103]</sup>, MHC molecules, cytoskeletal proteins and signal transduction molecules<sup>[104]</sup>, where engulfment of cytosolic proteins involves proteins located close to the outer membrane of MVB by autophagy<sup>[105]</sup>.

Interest in exosomes has steeply increased, when it was reported that exosomes contain mRNA and miRNA that will be transferred into target cells<sup>[106]</sup>. Exosomal mRNA and miRNA also differs from that in the donor cell. mRNA recruitment can be guided by a zip code in the 3'-UTR<sup>[107]</sup>. Exosomal mRNA is less abundant than exosomal miRNA. Exosomal mRNA are mostly involved in cell cycle progression, angiogenesis, migration, or histone modification<sup>[98,108,109]</sup>. Exosomes also contain selected miRNA. miRNA recruitment is facilitated by coupling of RISCs (RNA-induced silencing complexes) to components of the sorting complex<sup>[110,111]</sup>, the release

of miRNA being controlled through ceramide-dependent machinery associated with exosome secretion<sup>[112]</sup>. Exosomes contain > 120 miRNA from a selected number of genes. Network based analysis of exosomal miRNA points towards an involvement in stem cell differentiation (let-7), organogenesis (miR-1), hematopoiesis (miR-181) tumorigenesis (miR-17, miR-18, miR-19a, miR-20, miR-19b-1, miR-93-1)<sup>[113,114]</sup> and metastasis<sup>[105]</sup>.

As exosomes are found in all body fluids<sup>[77]</sup>, the selective enrichment of “marker” proteins as well as of miRNA makes exosomes a very attractive means for non-invasive diagnosis<sup>[104,113]</sup>.

### Tumor diagnosis by serum exosomes

Exosomes are separated by sequential centrifugation steps followed or preceded by 0.2 µm filtering. For pre-evaluation exosomes should be further purified by sucrose density gradient centrifugation<sup>[114,115]</sup>. This, however may not be possible for large sample number evaluation and also may not be feasible with the amount of available serum. According to our experience and in line with literature reports, 1 mL of serum will be sufficient for screening of a limited number of proteins and miRNA. Particularly for miRNA screening, recently a thorough comparative evaluation of mRNA preparation has been published<sup>[116]</sup>, which should be taken into account as in dependence of the exosome source minor differences may lead to a pronounced loss of miRNA. Besides these “home made” exosomes, several commercially available exosome purification kits are available that were described to reveal comparable results. In addition, there are special diagnostic kits on the market, which will be helpful, if a clearly defined question is to be answered, e.g., searching for one marker or a few selected miRNA. As far as one is interested to find out the protein marker or miRNA profile of exosomes of a tumor entity that has not yet already been analyzed, it may be preferable to start open minded without any preselection. In concern of the readout system, I strongly recommend for miRNA the protocol of Liu *et al*<sup>[9]</sup> described above for free serum miRNA, starting with a microarray of pooled serum exosomes from patients and control donors. According to our unpublished experience the ten most abundant miRNA are with high likelihood recovered in exosome pools of different patients. As the serum contains much more exosomes that are not tumor-derived, taking into account that only platelets account for roughly 50% of serum exosomes<sup>[117]</sup>, the comparison to healthy donors’ exosomes provides already a good means to select out non-tumor exosomal miRNA. As an additional control, I would recommend exosomes from culture supernatant of tumor lines from the same cancer type.

It also should be remembered that exosome collect a limited number of mRNA and miRNA that does not correlate to the mRNA or miRNA profile of the cell, which we confirmed for a rat pancreatic cancer line and exosomes derived thereof<sup>[109]</sup>. Our unpublished study on

human PaCa serum exosomes confirms this inasmuch as the miRNA profile of serum exosomes and of culture supernatant exosomes show abundance of the same miRNA. In addition, the absence of a miRNA that is recovered in serum exosomes from healthy donors and PaCa patients provides a strong hint towards this miRNA being not derived from tumor exosomes. Having selected for miRNA abundant in pools of PaCa patients serum exosomes, one can proceed with verification by qRT-PCR.

In concern of serum exosome marker profiles one should also start with pooled healthy donors serum exosomes and select for markers that are undetectable on healthy donors’ exosomes. Antibodies against constitutive exosome markers may serve as controls. After this screening one can either proceed with enzyme-linked immunosorbent assay (ELISA)<sup>[118]</sup> or flow cytometry, where latex beads can be coated with antibody in advance or latex beads are loaded with exosomes and marker expression is evaluated by incubation with antibodies after blocking free binding sites of the latex beads<sup>[114,119,120]</sup>. Both procedures have advantages and disadvantages and it depends on the individual question, which to prefer. For diagnostic purposes several kits are commercially available.

So far, at least to my knowledge studies being concerned about serum diagnosis or diagnosis in other body fluids, like the urine, by miRNA have rarely taken into account the particular profile of exosomes. To give a few examples. In glioblastoma serum exosomes miR-21 was 40-fold increased<sup>[108]</sup>. In serum exosomes from ovarian cancer patients, 8 miRNA were significantly increased<sup>[121]</sup> and in prostate cancer urine exosomal miR-107 and miR-574-3p are upregulated<sup>[122]</sup>.

In concern of the comparably rare trials on serum or other body fluids exosomes as diagnostic tool, I want to stress again that only exosomal miRNA is delivered by live cells<sup>[19,71,72]</sup>. Thus, this miRNA can be expected to be particularly selected for functional relevance. In addition, CSC/migrating tumor cells are suggested to be enriched in the serum<sup>[123,124]</sup> and could well contribute to the serum exosome pool and to its diagnostic validity, cancer progression relying on the small population of CSC, which account for drug resistance, metastasis and late recurrence<sup>[125-127]</sup>. Finally, exosomes being delivered by live tumor cells, the amount of exosomal miRNA may change with the size of the tumor, but the miRNA profile most likely will be stable.

Serum exosomes as a diagnostic tool have an additional advantage, as besides tumor miRNA, membrane bound tumor markers can be searched for. Thus, in ovarian cancer, CD24<sup>+</sup> and EpCAM<sup>+</sup> exosomes were recovered in ascites of tumor patients and in serum CD24<sup>+</sup> exosomes were detected, the absence of EpCAM<sup>+</sup> exosomes in serum being due to cleavage by exosomal ADAM10<sup>[128,129]</sup>. Also in ovarian cancer claudin4 was upregulated in 32 of 63 patients’ serum exosomes, but only in 1 of 50 control serum exosomes<sup>[130]</sup>. In plasma exo-

somes of prostate cancer patients' survivin is upregulated compared to controls and benign prostate hyperplasia<sup>[131]</sup>. In urine exosomes of prostate cancer also PCA3 and TMPRSS2: ERG, deriving from a chromosomal rearrangement were detected, verifying body fluid exosomes as diagnostic marker<sup>[132]</sup>, though in another study on prostate cancer urinary exosomes PSA and PSMA were detected, but exosomes in urine showed great variability<sup>[133]</sup>. Also in plasma exosomes from melanoma patients caveolin-1 and CD63 were consistently elevated<sup>[134]</sup> and tumor exosomes could be efficiently isolated with anti-HER2/neu from ascites of cancer patients<sup>[135]</sup>. Last, not least, the tumor-specific epidermal growth factor receptor VIII (EGFRVIII) was detected in 7 out of 25 glioblastoma patients serum exosomes<sup>[108]</sup> and our ongoing study on pancreatic cancer serum exosomes confirms recovery of exosomes carrying PaCa stem cell markers<sup>[124]</sup>.

Taken together, comparably few studies on cancer patients serum/plasma or urinary exosomes confirmed the suggestion that exosomes in body fluids can serve as a diagnostic tool. Unfortunately, at least according to my stage of knowledge, PaCa serum exosomes have not yet been evaluated, where I strongly recommend to take into account that exosomes offer the possibility to evaluate both miRNA and protein markers. Our ongoing studies strongly suggest that combining the analysis of these two parameters most likely will bring about a considerable improvement in early PaCa diagnosis.

## EXOSOMES AS A THERAPEUTIC TOOL

Exosomes are hotly debated as the most potent gen therapeutic option of the future<sup>[12]</sup>. In advance of discussing this option, I should briefly introduce what is known so far about the interaction between exosomes and target cells. I will first discuss exosome binding and uptake and proceed giving a brief overview on exosome binding and uptake-induced target modulation.

### Exosome binding and uptake

In advance of considering options for the therapeutic use of exosomes, it is a *conditio sine qua non* to be aware, which cells in the body are potentially targeted by exosomes. Though it is well appreciated that exosomes only interact with selected targets<sup>[97,98,136]</sup>, the mode of selection requires further clarification. Several options, which are mutually not exclusive are discussed, receptor-ligand interactions, attachment, fusion with the target cell membrane, or internalization<sup>[136-138]</sup>.

Due to inward budding of endosomes into MVB, the outer membrane of exosomes is characterized by phosphatidylserine (PS), which can trigger exosome uptake by binding to scavenger receptors, integrins, complement receptors and PS receptors (TIM), particularly TIM-4<sup>[139,140]</sup>. In line with this, macrophages (M $\phi$ ) very rapidly bind exosomes, binding being efficiently blocked by anti-CD11b<sup>[141]</sup>. However, *in vivo* studies did not provide evidence that exosome uptake is dictated by scavenger

receptors. Furthermore, the selectivity of exosome uptake argues for PS facilitating binding, but not for being involved in exosome uptake<sup>[97,141,142]</sup>.

Instead, already in 2004 evidence was presented that exosome uptake by dendritic cells (DC), Kupffer cells and some macrophages (M $\phi$ ) involves, besides PS, milk fat globulin-E8, CD11a, CD54, CD9 and CD81 on exosomes and requires  $\alpha v \beta 3$ , CD11a and CD54 as ligands on DC<sup>[143]</sup> suggesting exosome binding and uptake to involve receptor-ligand interactions that may vary depending on the protein pattern on exosomes and target cells<sup>[144]</sup>. Notably, this early study also pointed towards a later on confirmed contribution of tetraspanins<sup>[97,145,146]</sup>. We additionally unraveled that target cell ligands are also located in internalization prone protein clusters, which include annexins, chaperons, molecules involved in vesicular transport, tetraspanins and tetraspanin-associated molecules<sup>[97]</sup>. Thus, internalization by donor cells and the exosome uptake by target cells use similar fusion/fission machineries, maintenance of internalization complexes and re-use of these complexes for exosome uptake apparently being a common theme<sup>[146-148]</sup>. Furthermore, antibody blocking of CD91, a common receptor for several HSP interferes with exosome activity<sup>[149]</sup>. Of note, exosomes also bind with high avidity several matrix proteins<sup>[102]</sup>, where matrix protein binding is selective and requires defined tetraspanin-adhesion molecule complexes<sup>[103]</sup>. Less is known about the discussed mechanism allowing for fusion of exosomes with their target cell. However, it has been shown that exosome fusion is facilitated or requires an acid pH<sup>[150]</sup>.

Thus, exosomes display target cell selectivity, which at least partly builds on the engagement of protein complexes in internalization prone membrane domains.

### Target modulation by exosomes

First to note, exosomal proteins, mRNA and miRNA are function competent<sup>[112,145]</sup>. Accordingly, there are several modes, whereby exosomes can modulate their targets. Binding-induced target modulation mostly relies on activation of exosome ligands and protein cleavage by exosomal proteases. Exosome uptake-initiated changes can be brought about by transferred proteins, mRNA and miRNA. These distinct activities of exosomes are far from being comprehensively understood, but all have exemplarily been confirmed. I will mention some examples, as I feel it is important to be aware of this ongoing research to understand the potential power of an exosome based therapy.

### Exosome-binding induced target modulation

Exosomes are rich in proteases<sup>[102]</sup>, which modulate the exosomes protein profile as well the ECM and target cells.

A tumor creates its own matrix, but also influences the host matrix to generate surroundings promoting tumor cell migration and survival. The phenomenon is poorly understood and the impact of tumor exosomes is largely unexplored. First to note, exosome proteases

modulate the exosome protein profile, described for L1 and CD44 shedding by ADAM10 and for EpCAM, CD46, TNFR1 by unknown metalloproteinases<sup>[151-153]</sup>. Exosomal proteases also modulate the ECM, where exosomal tetraspanins due to their association with proteases and integrins become important<sup>[152,154-156]</sup>. The collagenolytic and laminin-degrading activity of exosomes facilitates angiogenesis and metastasis<sup>[142,157-162]</sup>, degradation of aggrecan increases invasiveness<sup>[163,164]</sup> and exosomal MMP2, MMP9, MMP14 and cathepsinB correlate with invasiveness<sup>[160,165]</sup>. Focalizing exosomal matrix degrading enzymes allows for paving the path of metastasizing CSC towards the premetastatic niche, which we confirmed for a rat metastasizing pancreatic adenocarcinoma<sup>[103,166]</sup>. As the ECM also is a storage of bioactive compounds<sup>[167]</sup>, modulation of the ECM by exosomal proteases<sup>[168]</sup> can account for cytokine/chemokine and protease liberation and generation of cleavage products that promote motility, angiogenesis and stroma cell activation<sup>[102]</sup>. Thus, the modulation of the ECM by exosomal proteases creates a path for migrating cells, favors a tumor growth promoting microenvironment, angiogenesis and premetastatic niche establishment.

**Exosome-initiated signal transduction:** Exosome-initiated signal transduction can be promoted by exosome binding and exosome uptake, which in most instances is experimentally difficult to decipher. Nonetheless, the impact of tumor exosome binding-initiated signal transduction on tumor immunity, angiogenesis, tumor growth/metastasis has been convincingly demonstrated.

DC-exosomes are one of the best explored examples for exosome binding-initiated signal transduction. DC-exosomes can replace DC in immune response induction and exosome-based therapy was first explored using DC-exosomes as a cancer vaccine. DC also take up exosomes secreted by other cells, including tumor cells, which they internalize and process for presentation. Thus, DC use exosomes as a source of antigen and produce exosomes that suffice for T cell activation, both features expanding the operational range of DC<sup>[143,169-171]</sup>.

Tumor exosomes also affect the immune system<sup>[172]</sup>. Tumor exosomes inhibit CD4<sup>+</sup> T cell proliferation, which is accompanied by up-regulation and stronger suppressive activity of regulatory T cells (Treg) due to exosome-associated transforming growth factor beta 1 (TGF-β1)<sup>[168]</sup>. NK activity also becomes impaired *via* tumor exosome inhibiting activation of Stat5, Jak3, cyclinD3 expression and perforin release<sup>[173]</sup> or due to blocking NK cells *via* NKG2D binding<sup>[174]</sup>. Induction of myeloid-derived suppressor cell (MDSC) is promoted by exosomal TGFβ and PGE2<sup>[175]</sup>. *Via* stimulating TGFβ1 secretion by Mφ, tumor exosomes suppress anti-tumor immune responses allowing for tumor growth and metastasis formation in allogeneic mice<sup>[176]</sup> and by high ICAM1 expression, tumor exosomes block the interaction between T cells and endothelial cells, thereby decreasing T cell recruitment<sup>[177]</sup>. On the other hand, high level HSP expression on tumor

exosomes-HSP functioning as an endogenous danger signal-promotes NK activation and tumor cell lysis<sup>[178,179]</sup> and supports T cell activation and effector functions<sup>[180]</sup> as well as induction of costimulatory molecule expression in DC<sup>[181,182]</sup>. Tumor exosomal chemokines attract and activate DC and T cells, such that intratumoral injection efficiently inhibits tumor growth<sup>[183]</sup>. Tumor exosomes also can be an efficient antigen source, which induce a potent Th, CTL and B cell response, even where lysates of the same tumor are non-immunogenic<sup>[141,184]</sup>.

Taken together, there is an intense crosstalk between tumor exosomes and the immune system that may be due predominantly to exosome binding-initiated signal transduction. Depending on the individual tumor's exosome composition, immune responses are suppressed, but also can be strengthened and in combination with DC tumor exosomes could well contribute to cancer immunotherapy.

Angiogenesis induction being one of the hallmarks of cancer, intense efforts have been taken to elaborate the contribution of tumor exosomes. Tumor exosomes containing tumor necrosis factor alpha (TNF-α), IL1β, TGFβ and TNFR1 recruit endothelial cell (EC) progenitors, promote angiogenesis<sup>[107]</sup> and stimulate EC by paracrine signaling<sup>[185]</sup>. Delta-like4 bearing tumor exosomes confer a tip cell phenotype to EC with filopodia formation, enhanced vessel density and branching<sup>[186]</sup>, which involves activation of PPARα and NFκB activation<sup>[187]</sup>. In a feedback, prostate cancer exosomes lead to activation of fibroblasts, which then shed exosomes that increase tumor cell migration *via* CX3C-CX3CR1<sup>[188]</sup>.

Another elegant examples of tumor exosome-mediated signal transduction describes overexpression of CD9 or CD82 promoting formation and secretion of exosomes that contain β-catenin, thereby reducing its cellular content and impairing Wnt signaling, which proceeds *via* tetraspanin-associated E-cadherin<sup>[189]</sup>. Besides indicating that the cargo of exosomes differs depending on ESCRT- or tetraspanin-initiated internalization, this study demonstrates that by depletion of inhibitors or stimulators tumor exosomes can oppositely affect signal transduction<sup>[190]</sup>. Also, tumor exosome-promoted tumor growth may vary for individual tumors. Thus, a deficit in Rab27a leading to reduced exosome production affected growth of a tumor line that required recruitment of neutrophils, but not of another neutrophil-independent line<sup>[191]</sup>.

Briefly, binding of tumor exosomes to hematopoietic cells, EC and stroma cells can severely affect the target cell, which may become activated or suppressed. Additionally, the export of proteins into tumor exosomes affects the tumor cell itself. It also has to be kept in mind that tumor exosome-initiated signaling varies with the origin and composition of tumor exosomes. Last and importantly, the strength of tumor exosome initiated signaling relies on their accessibility throughout the body.

**Exosome uptake promoted target cell modulation:** Early reports on the information transfer *via* exosomes

showed that embryonic stem cell exosomes transfer messages into hematopoietic progenitor cells that promoted survival and expression of early pluripotency markers<sup>[20]</sup>. Adult tissue exosomes, too, had the capacity to alter the phenotype of their target such that upon coculture bone marrow cells (BMC) express markers found on the exosome donor cell<sup>[192]</sup>, where uptake of exosome proteins, mRNA and miRNA are contributing. These findings also account for tumor exosomes, which transfer receptor and oncoproteins or miRNA<sup>[20,193]</sup>.

One of the first evidences to support tumor exosome-uptake plays a critical role in autocrine stimulation of tumor growth revealed that the intercellular transfer of the oncogenic receptor EGFRVIII *via* tumor exosomes to glioma cells, lacking this receptor, causes transformation of indolent glioma cells<sup>[194]</sup> and reprograms growth factor pathways in EC<sup>[126]</sup>. Other oncogenes, like Ras, Myc, SV40T also induce signaling and gene expression<sup>[195-197]</sup>, where *e.g.*, exosomal amphiregulin, an EGFR ligand, increased tumor invasiveness 5-fold compared to the recombinant protein<sup>[198]</sup>.

Tumor exosome uptake-induced changes in recipient non-tumor cells can be transient, but also suffice to drive tumor growth as described for tissue transglutaminase and fibronectin<sup>[199]</sup> or high level c-Met uptake by BMC, which leads to their re-education to support premetastatic niche formation for melanoma cells, where in melanoma patients, too, circulating BM-derived cells express Met<sup>[200]</sup>. Tumor exosomes also transport apoptosis inhibitory proteins<sup>[201]</sup> and present TGF $\beta$ . This drives differentiation of fibroblasts towards myofibroblasts, which support tumor growth<sup>[202]</sup>. Adipose-tissue derived mesenchymal stem cells (MSC) also can be driven into myofibroblasts by tumor exosomes<sup>[203]</sup>. Lung cancer tumor exosome uptake stimulates IL8, VEGF, LIF, oncostatin and MMP secretion, which promotes tumor growth<sup>[204]</sup>. Instead, uptake of tumor suppressor genes from non-transformed cells can mitigate cancer cell aggressiveness<sup>[12,205]</sup>.

An involvement of exosomes in metastasis was first described for platelet-derived exosomes, which transferred the  $\alpha$  IIb integrin chain to lung cancer cells, stimulated the MAPK pathway and increased expression of MT1-MMP, cyclin D2 and angiogenic factors and enhanced adhesion to fibrinogen and human umbilical vein EC<sup>[206]</sup>. We explored that exosomes from a PaCa together with a soluble tumor matrix facilitated recruitment of hematopoietic progenitors from the BM as well as activation of stroma cells and leukocytes in premetastatic lymph nodes such that a non-metastatic tumor line settled and formed metastases<sup>[207]</sup>. The recruitment of tumor cells also becomes facilitated by exosomal HSP90, a complex of exosomal HSP90 with MMP2 and tissue plasminogen activator promoting together with exosomal annexin II plasmin activation tumor cell motility<sup>[208]</sup>. As already mentioned, the transfer of c-Met contributes to premetastatic niche formation mostly *via* bone marrow cell modulation<sup>[200]</sup>. Thus, tumor exosomes

enhance migration and homing of tumor cells in sentinel lymph nodes due to stroma and hematopoietic cell as well as matrix modulation<sup>[77,108,200,207]</sup>. Finally, uptake of exosomes from non-transformed cells in the tumor surrounding can affect tumor cells such that fibroblast-exosomes promote breast cancer motility *via* Wnt planar polarity signaling<sup>[209]</sup>.

Tumor exosome uptake also accounts for EC modulation. Colorectal cancer exosomes, enriched in cell cycle-related mRNA, promote EC proliferation<sup>[210]</sup>. Glioblastoma-exosome-induced angiogenesis relies on the transfer of exosomal proteins and mRNA<sup>[108]</sup>. Uptake of EGFR-positive tumor exosomes by EC elicit EGFR-dependent responses including activation of the MAPK and Akt pathway and VEGFR2 expression<sup>[211]</sup>. Transfer of exosomal Notch-ligand-delta-like-4 increases angiogenesis<sup>[183]</sup> and tumor exosomes expressing a complex of Tspan8 with CD49d preferentially are taken up by EC and EC progenitors, which initiates progenitor maturation and EC activation including VEGFR transcription<sup>[60]</sup>. Chronic myeloid leukemia (CML)-exosomes induce angiogenic activity in EC, where a Src inhibitor affects exosome production as well as vascular differentiation<sup>[212]</sup>.

As mentioned tumor exosome uptake-induced target cell modulation frequently represent the net result of protein transfer-initiated signal transduction, transferred mRNA translation and mRNA silencing by miRNA. Though a separation between these activities appears somewhat artificial, a few reports describing preferential activities of mRNA and miRNA should be mentioned.

By the transfer of miR-150 in AML-exosomes to hematopoietic progenitors CXCR4 expression becomes reduced and HSC migration is impaired<sup>[213]</sup>. CD105<sup>+</sup> renal cell CSC exosomes carry proangiogenic mRNA and miRNA, which trigger the angiogenic switch<sup>[158]</sup>. mRNA and miRNA of exosomes from a metastasizing PaCa are recovered in lymph node stroma and lung fibroblasts, and transferred miRNA significantly affects mRNA translation, which was exemplified for abundant exosomal miR-494 and miR-542-3p, which target cadherin17. Concomitantly, MMP transcription, accompanying cadherin17 downregulation, was up-regulated in lymph node stroma cells transfected with miR-494 or miR-542-3p or co-cultured with tumor exosomes. Thus, tumor exosome miRNA uptake affected premetastatic organ stroma cells towards supporting tumor cell hosting<sup>[109]</sup>. Exosomes from virus transfected cells transfer viral miRNA<sup>[214,215]</sup>. Leukemia cell exosomes contain miR-92a that is transferred into EC, downregulates CD49e and increases migration and tube formation<sup>[216]</sup>. In lung cancer exosomes miR-21 and miR-29a act as a ligand of mouse TLR7 or human TLR8, functioning as agonist and leading to NF $\kappa$ B activation and IL6 and TNF $\alpha$  secretion, which promotes metastasis<sup>[217]</sup>. Hepatocellular carcinoma exosomes abundantly contain miR-584, miR-517c, one of the potential targets, TGF $\beta$  activated kinase 1, activates JNK and MAPK pathway and NF $\kappa$ B, where transfer of exosomal miRNA in coculture promoted anchorage-

independent growth and apoptosis resistance<sup>[218]</sup>.

Stroma cells also release exosomes, whose miRNA can influence tumor cells. BM stroma cell exosomes inhibit the growth of multiple myeloma, but those derived from patients with multiple myeloma force multiple myeloma progression, the latter exosomes showing a lower content of tumor suppressor miR-15a, but high levels of oncogenic proteins, cytokines and adhesion molecules<sup>[219]</sup>. Tumor-associated M $\phi$  secret exosomes with high miR-223, that binds Mef2c, causing nuclear accumulation of  $\beta$ -catenin<sup>[220]</sup>. Monocyte exosomal miR-150, when transferred to EC, promotes migration<sup>[221]</sup>.

Taken together, transferred exosomal miRNA can re-program target cells, the linkage between exosomal miRNA and the targeted mRNA remaining to be elaborated in detail in many instances. In concern of the described impact of transferred proteins and mRNA, the question on long-lasting *in vivo* efficacy awaits clarification. Exosomes being a most powerful means of intercellular communication that function across long distance, it is utmost important to answer these open questions. Nonetheless, therapeutic exploitation of exosomes appears promising.

## EXOSOMES AS THERAPEUTICS

Exosomes are discussed as most potent gene delivery system, as they are easy to manipulate and efficiently transfer proteins and genes. This could offer a means to interfere with tumor exosome promoted angiogenesis and metastasis, two major targets in cancer therapy<sup>[191,222]</sup>. In addition, exosomes are discussed as cancer vaccine<sup>[172]</sup>. Nonetheless, in advance of discussing the possibilities to interfere with tumor growth and progression *via* exosomes, I want to stress three points. First, uptake by selective target cells needs to be most thoroughly controlled. Second, the pathway whereby exosomes affect a selected target cells has to be well defined. Besides the still open question, whether transferred proteins, mRNA and miRNA or a combination account for observed effects, the multiple targets of individual miRNA could create problems such that side effects at the present state of knowledge can not be excluded. Third, it should be mentioned that the indispensability of exosome transfer in human cancer remains questionable. In A431 PS blocking inhibits uptake of exosomes by EC, but the antiangiogenic effect was only transient<sup>[194]</sup>. Also a blockade of cellular vesiculation (TSAP6, acidic sphingomyelinase) did not prevent tumorigenesis<sup>[223,224]</sup>. Furthermore, blocking of Rab27a involved in exosome biogenesis exerts distinct effects on primary versus metastatic tumor growth and also differs between tumors<sup>[200,225]</sup>. These findings should not be taken to discourage attempts to translate experimental studies on the power of exosomes into therapeutic settings, but should foster the point that clinical translation in many instances essentially awaits progress in elaborating the mode of exosome activities. These clauses account particularly for active interference

with tumor exosomes. Instead, DC exosomes are already used as a vaccine<sup>[226,227]</sup>.

### Exosomes to substitute or support dendritic cells

Exosome research became highly stimulated, when it was noted that antigen presenting cells release exosomes derived from MVB of the MHC class II compartment, which can stimulate T cells *in vitro* and *in vivo*<sup>[228]</sup>. Several studies report that DC-exosomes were well tolerated, induced an antigen-specific response and or NK recovery and that the disease-free survival time was mostly prolonged. For the therapeutic translation it is also beneficial that exosomes can be stored at -80 °C and that recovery is high. Limitation were mostly restricted to the requirement of large amounts of DC-exosomes<sup>[229-231]</sup>.

Though tumor exosomes can be immunosuppressive, this does not affect their use for loading DC. Several groups report that exosomes delivered from DC after coculture with tumor exosomes might be superior to exosomes derived from peptide-pulsed DC. DC pulsed with exosomes of an AML line provoked a strong anti-leukemia response<sup>[232]</sup>. In line with this, directing tumor-associated, non-mutated antigens like CEA and HER2 to exosomes by coupling to lactadherin increased their immunogenicity<sup>[233]</sup>. Targeting prostate-specific antigen or prostatic acid phosphatase *via* lactadherin to exosomes also induced a superior immune response<sup>[234]</sup>. Furthermore, anticancer drug force the release of HSP-bearing exosomes, which efficiently activate NK cells<sup>[235]</sup>. Taking this into account, tumor exosomes should be particularly helpful as antigen source, when immunogenic entities of a tumor are unknown.

### Competing with tumor exosomes

Even taking into account that an individual tumor may not essentially depend on exosomes for survival and progression, tumor exosomes doubtless support the tumor by modulating the host. Thus, competing with tumor exosomes might be a means to retard metastasis formation.

Blocking of exosome uptake could be performed at the exosome or the target cell level<sup>[229]</sup>, where PS blocking of tumor exosomes only transiently inhibited angiogenesis<sup>[194]</sup>. Instead, in a rat PaCa, where exosomes expressing the tetraspanin Tspan8 induced a lethal systemic consumption coagulopathy, blocking exosomes by a Tspan8-specific antibody completely prevented undue angiogenesis, although primary tumor growth was not impaired<sup>[236,237]</sup>. Based on this finding and our ongoing studies that exosomes bind *via* tetraspanin-complexes to ligands also located in internalization prone membrane domains<sup>[59]</sup>, we speculate that a scrutinized analysis of an individual tumors' exosome-binding complex should provide the information for hampering undue tumor exosome-initiated angiogenesis and premetastatic niche formation, where exosomes from non-transformed cells modulated to express the tumor exosome-binding complex will be most promising<sup>[59]</sup>. As an alternative

approach, tumor exosomes can be removed by affinity plasmapheresis known as Aethlon ADAPT™<sup>[238]</sup>. Blocking of tumor exosomes also can affect drug and radiation resistance due to enhanced release of export transporter MRP2, ATP7A and ATP7B or Annexin A3<sup>[239,240]</sup>.

### Tailored exosomes for drug delivery

Greatest hope in exosome therapy is based on the discovery of horizontal transfer of mRNA and miRNA<sup>[106,241]</sup>, which can be translated or mediate RNA silencing<sup>[20,73,242]</sup>.

As exosomes are natural products, are small and flexible, which allows them to cross biological membranes and to protect their cargo from degradation by a lipid bilayer<sup>[138]</sup>, they are discussed as ideal and possibly the most potent gene delivery system<sup>[73,86,138,241,243]</sup>. Notably, exosome electroporation efficiently transfers siRNA into exosomes<sup>[112]</sup>. Furthermore, special devices can be developed, e.g., to cross the blood-brain barrier, which was explored for the delivery of BACE1 siRNA, where mast cell exosomes were equipped with a brain penetrating peptide fused to the vesicular membrane protein Lamp2<sup>[244,245]</sup>. Also, curcumin or Stat3 inhibitor delivery confirmed exosomes to be well suited for drug delivery<sup>[246,247]</sup>, where chemotherapeutic drug efficacy was increased by lowering the pH of exosomes<sup>[150,248]</sup>. Adenoviral vectors associated with exosomes displayed higher transduction efficacy than purified AAV vectors<sup>[249]</sup>. As exosomes from non-tumor cells contain tumor-suppressive miRNA, it was suggested to use exosomes loaded with those miRNA, which was exemplified for miR-143 as a therapeutic strategy in cancer<sup>[213]</sup>. In a mouse hepatoma, systemic administration of miR-26a, inducing cell cycle arrest, exerted a dramatic protective effect without toxicity<sup>[250]</sup>. Additional approaches like miRNA inhibitors (miRNA sponges), antagomirs, locked-nucleic-acid-modified oligonucleotides are reviewed in<sup>[23]</sup>.

At the present state of knowledge miRNA based therapies have to be considered as double-edged sword as most miRNA have a multitude of targets. However, as soon as the above mentioned hurdles are solved, rapid progress in clinical translation can be expected<sup>[251,252]</sup>.

### CONCLUSION

The recovery of tumor-associated miRNA and of tumor exosomes in serum and other body fluids has created hope for non/minimally invasive diagnostics, where our own, unpublished data indicate that an exosome-based screening may be advantageous as it offers the possibility to search concomitantly for tumor-related protein markers as well as tumor-associated miRNA. Taking into account that the poor prognosis of PaCa patients despite considerable progress in surgical treatment is mostly due to late diagnosis, a reliable serum-based diagnosis at early stages could already significantly contribute improving the rate of curative treatment.

Beyond diagnosis, the discovery of exosomes as intercellular communicators throughout the body fostered

reconsideration of many aspects of tumor biology and is hoped to bring a major breakthrough in therapy. The power of exosomes is due to their ubiquitous presence, their particular protein profile and their equipment with mRNA and miRNA as well as their most efficient transfer in target cells. Together with the ease of transfecting exosomes, there should be hardly any limits in the use of exosomes as therapeutics. The therapeutic use of exosomes from non-transformed cells to compete, to induce an immune response or to silence immunosuppression should not become a danger for the patient's organism. Instead, therapeutic approaches based on tailored tumor exosomes still awaits answers to the targeting receptors and their ligands, which most likely will offer modalities to further restrict the panel of potential targets of natural tumor exosomes and a precise knowledge on miRNA targets and consequences on release from repression. Answering these questions will take time, but is not an insurmountable hurdle.

PaCa are burdened by desmoplasia and early metastatic spread. Both features essentially depend on the crosstalk with the host, which has been convincingly demonstrated to be to a considerably degree mediated by tumor exosomes. Thus, it is my personal opinion that PaCa treatment/diagnosis will particularly profit from unraveling the option of exosome-based therapy.

### REFERENCES

- 1 **Jemal A**, Siegel R, Xu J, Ward E. Cancer statistics, 2010. *CA Cancer J Clin* 2010; **60**: 277-300 [PMID: 20610543 DOI: 10.3322/caac.20073]
- 2 **Loos M**, Kleeff J, Friess H, Büchler MW. Surgical treatment of pancreatic cancer. *Ann N Y Acad Sci* 2008; **1138**: 169-180 [PMID: 18837898 DOI: 10.1196/annals.1414.024]
- 3 **Wang Z**, Li Y, Ahmad A, Banerjee S, Azmi AS, Kong D, Sarkar FH. Pancreatic cancer: understanding and overcoming chemoresistance. *Nat Rev Gastroenterol Hepatol* 2011; **8**: 27-33 [PMID: 21102532 DOI: 10.1038/nrgastro.2010.188]
- 4 **Paulson AS**, Tran Cao HS, Tempero MA, Lowy AM. Therapeutic advances in pancreatic cancer. *Gastroenterology* 2013; **144**: 1316-1326 [PMID: 23622141 DOI: 10.1053/j.gastro.2013.01.078]
- 5 **Guo J**, Wang W, Liao P, Lou W, Ji Y, Zhang C, Wu J, Zhang S. Identification of serum biomarkers for pancreatic adenocarcinoma by proteomic analysis. *Cancer Sci* 2009; **100**: 2292-2301 [PMID: 19775290 DOI: 10.1111/j.1349-7006.2009.01324]
- 6 **Giovinazzo F**, Turri G, Zanini S, Butturini G, Scarpa A, Bassi C. Clinical implications of biological markers in Pancreatic Ductal Adenocarcinoma. *Surg Oncol* 2012; **21**: e171-e182 [PMID: 22981281 DOI: 10.1016/j.suronc.2012.07.004]
- 7 **Kaur S**, Baine MJ, Jain M, Sasson AR, Batra SK. Early diagnosis of pancreatic cancer: challenges and new developments. *Biomark Med* 2012; **6**: 597-612 [PMID: 23075238 DOI: 10.2217/bmm.12.69]
- 8 **Bhat K**, Wang F, Ma Q, Li Q, Mallik S, Hsieh TC, Wu E. Advances in biomarker research for pancreatic cancer. *Curr Pharm Des* 2012; **18**: 2439-2451 [PMID: 22372502]
- 9 **Liu R**, Chen X, Du Y, Yao W, Shen L, Wang C, Hu Z, Zhuang R, Ning G, Zhang C, Yuan Y, Li Z, Zen K, Ba Y, Zhang CY. Serum microRNA expression profile as a biomarker in the diagnosis and prognosis of pancreatic cancer. *Clin Chem* 2012; **58**: 610-618 [PMID: 22194634 DOI: 10.1373/clinchem.2011.172767]
- 10 **György B**, Szabó TG, Pásztói M, Pál Z, Misják P, Aradi B, László V, Pállinger E, Pap E, Kittel A, Nagy G, Falus A,

- Buzás EI. Membrane vesicles, current state-of-the-art: emerging role of extracellular vesicles. *Cell Mol Life Sci* 2011; **68**: 2667-2688 [PMID: 21560073 DOI: 10.1007/s00018-011-0689-3]
- 11 O'Loughlin AJ, Woffindale CA, Wood MJ. Exosomes and the emerging field of exosome-based gene therapy. *Curr Gene Ther* 2012; **12**: 262-274 [PMID: 22856601]
- 12 Lee Y, El Andaloussi S, Wood MJ. Exosomes and microvesicles: extracellular vesicles for genetic information transfer and gene therapy. *Hum Mol Genet* 2012; **21**: R125-R134 [PMID: 22872698]
- 13 Corrado C, Raimondo S, Chiesi A, Ciccia F, De Leo G, Alesandro R. Exosomes as intercellular signaling organelles involved in health and disease: basic science and clinical applications. *Int J Mol Sci* 2013; **14**: 5338-5366 [PMID: 23466882 DOI: 10.3390/ijms14035338]
- 14 Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; **116**: 281-297 [PMID: 14744438]
- 15 Calin GA, Croce CM. MicroRNA signatures in human cancers. *Nat Rev Cancer* 2006; **6**: 857-866 [PMID: 17060945]
- 16 Filipowicz W, Bhattacharyya SN, Sonenberg N. Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? *Nat Rev Genet* 2008; **9**: 102-114 [PMID: 18197166 DOI: 10.1038/nrg2290]
- 17 Lim LP, Lau NC, Garrett-Engele P, Grimson A, Schelter JM, Castle J, Bartel DP, Linsley PS, Johnson JM. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* 2005; **433**: 769-773 [PMID: 15685193]
- 18 Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; **136**: 215-233 [PMID: 19167326 DOI: 10.1016/j.cell.2009.01.002]
- 19 Tzimogiorgis G, Michailidou EZ, Kritis A, Markopoulos AK, Kouidou S. Recovering circulating extracellular or cell-free RNA from bodily fluids. *Cancer Epidemiol* 2011; **35**: 580-589 [PMID: 21514265 DOI: 10.1016/j.canep.2011.02.016]
- 20 Ratajczak J, Miekus K, Kucia M, Zhang J, Reca R, Dvorak P, Ratajczak MZ. Embryonic stem cell-derived microvesicles reprogram hematopoietic progenitors: evidence for horizontal transfer of mRNA and protein delivery. *Leukemia* 2006; **20**: 847-856 [PMID: 16453000]
- 21 Turchinovich A, Weiz L, Langheinz A, Burwinkel B. Characterization of extracellular circulating microRNA. *Nucleic Acids Res* 2011; **39**: 7223-7233 [PMID: 21609964 DOI: 10.1093/nar/gkr254]
- 22 Johnston M, Hutvagner G. Posttranslational modification of Argonautes and their role in small RNA-mediated gene regulation. *Silence* 2011; **2**: 5 [PMID: 21943311 DOI: 10.1186/1758-907X-2-5]
- 23 Ajit SK. Circulating microRNAs as biomarkers, therapeutic targets, and signaling molecules. *Sensors (Basel)* 2012; **12**: 3359-3369 [PMID: 22737013 DOI: 10.3390/s120303359]
- 24 Orlova IA, Alexander GM, Qureshi RA, Sacan A, Graziano A, Barrett JE, Schwartzman RJ, Ajit SK. MicroRNA modulation in complex regional pain syndrome. *J Transl Med* 2011; **9**: 195 [PMID: 22074333 DOI: 10.1186/1479-5876-9-195]
- 25 White NM, Fatoohi E, Metias M, Jung K, Stephan C, Yousef GM. Metastamirs: a stepping stone towards improved cancer management. *Nat Rev Clin Oncol* 2011; **8**: 75-84 [PMID: 21045789 DOI: 10.1038/nrclinonc.2010.173]
- 26 Reid G, Kirschner MB, van Zandwijk N. Circulating microRNAs: Association with disease and potential use as biomarkers. *Crit Rev Oncol Hematol* 2011; **80**: 193-208 [PMID: 21145252 DOI: 10.1016/j.critrevonc.2010.11.004]
- 27 Allen KE, Weiss GJ. Resistance may not be futile: microRNA biomarkers for chemoresistance and potential therapeutics. *Mol Cancer Ther* 2010; **9**: 3126-3136 [PMID: 20940321 DOI: 10.1158/1535-7163.MCT-10-0397]
- 28 Heneghan HM, Miller N, Kerin MJ. Circulating microRNAs: promising breast cancer Biomarkers. *Breast Cancer Res* 2011; **13**: 402; author reply 403 [PMID: 21345257 DOI: 10.1186/bcr2798]
- 29 Cortez MA, Bueso-Ramos C, Ferdin J, Lopez-Berestein G, Sood AK, Calin GA. MicroRNAs in body fluids--the mix of hormones and biomarkers. *Nat Rev Clin Oncol* 2011; **8**: 467-477 [PMID: 21647195 DOI: 10.1038/nrclinonc.2011.76]
- 30 Zhao L, Chen X, Cao Y. New role of microRNA: carcinogenesis and clinical application in cancer. *Acta Biochim Biophys Sin (Shanghai)* 2011; **43**: 831-839 [PMID: 21908856 DOI: 10.1093/abbs/gmr080]
- 31 Park SM, Gaur AB, Lengyel E, Peter ME. The miR-200 family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors ZEB1 and ZEB2. *Genes Dev* 2008; **22**: 894-907 [PMID: 18381893 DOI: 10.1101/gad.1640608]
- 32 Joglekar MV, Patil D, Joglekar VM, Rao GV, Reddy DN, Mitnala S, Shouche Y, Hardikar AA. The miR-30 family microRNAs confer epithelial phenotype to human pancreatic cells. *Islets* 2009; **1**: 137-147 [PMID: 21099261 DOI: 10.4161/isl.1.2.9578]
- 33 Wang Z, Zhang Y, Li Y, Banerjee S, Liao J, Sarkar FH. Down-regulation of Notch-1 contributes to cell growth inhibition and apoptosis in pancreatic cancer cells. *Mol Cancer Ther* 2006; **5**: 483-493 [PMID: 16546962]
- 34 Ji Q, Hao X, Zhang M, Tang W, Yang M, Li L, Xiang D, Desano JT, Bommer GT, Fan D, Fearon ER, Lawrence TS, Xu L. MicroRNA miR-34 inhibits human pancreatic cancer tumorigenic cells. *PLoS One* 2009; **4**: e6816 [PMID: 19714243 DOI: 10.1371/journal.pone.0006816]
- 35 Misawa A, Katayama R, Koike S, Tomida A, Watanabe T, Fujita N. AP-1-Dependent miR-21 expression contributes to chemoresistance in cancer stem cell-like SP cells. *Oncol Res* 2010; **19**: 23-33 [PMID: 21141738]
- 36 Khew-Goodall Y, Goodall GJ. Stromal miR-320 keeps an oncogenic secretome in check. *Nat Cell Biol* 2012; **14**: 124-125 [PMID: 22298040 DOI: 10.1038/ncb2431]
- 37 Uchida N. MicroRNA-9 controls a migratory mechanism in human neural progenitor cells. *Cell Stem Cell* 2010; **6**: 294-296 [PMID: 20362531 DOI: 10.1016/j.stem.2010.03.010]
- 38 Liu C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, Li H, Patrawala L, Yan H, Jeter C, Honorio S, Wiggins JF, Bader AG, Fagin R, Brown D, Tang DG. The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Nat Med* 2011; **17**: 211-215 [PMID: 21240262 DOI: 10.1038/nm.2284]
- 39 Wu ZS, Wu Q, Wang CQ, Wang XN, Huang J, Zhao JJ, Mao SS, Zhang GH, Xu XC, Zhang N. miR-340 inhibition of breast cancer cell migration and invasion through targeting of oncoprotein c-Met. *Cancer* 2011; **117**: 2842-2852 [PMID: 21692045 DOI: 10.1002/cncr.25860]
- 40 Le XF, Merchant O, Bast RC, Calin GA. The Roles of MicroRNAs in the Cancer Invasion-Metastasis Cascade. *Cancer Microenviron* 2010; **3**: 137-147 [PMID: 21209780 DOI: 10.1007/s12307-010-0037-4]
- 41 Jamieson NB, Morran DC, Morton JP, Ali A, Dickson EJ, Carter CR, Sansom OJ, Evans TR, McKay CJ, Oien KA. MicroRNA molecular profiles associated with diagnosis, clinicopathologic criteria, and overall survival in patients with resectable pancreatic ductal adenocarcinoma. *Clin Cancer Res* 2012; **18**: 534-545 [PMID: 22114136 DOI: 10.1158/1078-0432.CCR-11-0679]
- 42 Lee EJ, Gusev Y, Jiang J, Nuovo GJ, Lerner MR, Frankel WL, Morgan DL, Postier RG, Brackett DJ, Schmittgen TD. Expression profiling identifies microRNA signature in pancreatic cancer. *Int J Cancer* 2007; **120**: 1046-1054 [PMID: 17149698]
- 43 Kent OA, Mullendore M, Wentzel EA, López-Romero P, Tan AC, Alvarez H, West K, Ochs MF, Hidalgo M, Arking DE, Maitra A, Mendell JT. A resource for analysis of microRNA expression and function in pancreatic ductal adenocarcinoma cells. *Cancer Biol Ther* 2009; **8**: 2013-2024

- [PMID: 20037478]
- 44 **Zhang Y**, Li M, Wang H, Fisher WE, Lin PH, Yao Q, Chen C. Profiling of 95 microRNAs in pancreatic cancer cell lines and surgical specimens by real-time PCR analysis. *World J Surg* 2009; **33**: 698-709 [PMID: 19030927 DOI: 10.1007/s00268-008-9833-0]
  - 45 **Bloomston M**, Frankel WL, Petrocca F, Volinia S, Alder H, Hagan JP, Liu CG, Bhatt D, Taccioli C, Croce CM. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. *JAMA* 2007; **297**: 1901-1908 [PMID: 17473300]
  - 46 **Szafrańska AE**, Davison TS, John J, Cannon T, Sipos B, Maghnouj A, Labourier E, Hahn SA. MicroRNA expression alterations are linked to tumorigenesis and non-neoplastic processes in pancreatic ductal adenocarcinoma. *Oncogene* 2007; **26**: 4442-4452 [PMID: 17237814]
  - 47 **Li W**, Lebrun DG, Li M. The expression and functions of microRNAs in pancreatic adenocarcinoma and hepatocellular carcinoma. *Chin J Cancer* 2011; **30**: 540-550 [PMID: 21801602 DOI: 10.5732/cjc.011.10197]
  - 48 **Gironella M**, Seux M, Xie MJ, Cano C, Tomasini R, Gommeaux J, Garcia S, Nowak J, Yeung ML, Jeang KT, Chaix A, Fazli L, Motoo Y, Wang Q, Rocchi P, Russo A, Gleave M, Dagorn JC, Iovanna JL, Carrier A, Pébusque MJ, Dusetti NJ. Tumor protein 53-induced nuclear protein 1 expression is repressed by miR-155, and its restoration inhibits pancreatic tumor development. *Proc Natl Acad Sci USA* 2007; **104**: 16170-16175 [PMID: 17911264]
  - 49 **Mees ST**, Mardin WA, Wendel C, Baeumer N, Willscher E, Senninger N, Schleicher C, Colombo-Benkmann M, Haier J. EP300—a miRNA-regulated metastasis suppressor gene in ductal adenocarcinomas of the pancreas. *Int J Cancer* 2010; **126**: 114-124 [PMID: 19569050 DOI: 10.1002/ijc.24695]
  - 50 **Hao J**, Zhang S, Zhou Y, Liu C, Hu X, Shao C. MicroRNA 421 suppresses DPC4/Smad4 in pancreatic cancer. *Biochem Biophys Res Commun* 2011; **406**: 552-557 [PMID: 21352803 DOI: 10.1016/j.bbrc.2011.02.086]
  - 51 **Hao J**, Zhang S, Zhou Y, Hu X, Shao C. MicroRNA 483-3p suppresses the expression of DPC4/Smad4 in pancreatic cancer. *FEBS Lett* 2011; **585**: 207-213 [PMID: 21112326 DOI: 10.1016/j.febslet.2010.11.039]
  - 52 **Park JK**, Henry JC, Jiang J, Esau C, Gusev Y, Lerner MR, Postier RG, Brackett DJ, Schmittgen TD. miR-132 and miR-212 are increased in pancreatic cancer and target the retinoblastoma tumor suppressor. *Biochem Biophys Res Commun* 2011; **406**: 518-523 [PMID: 21329664 DOI: 10.1016/j.bbrc.2011.02.065]
  - 53 **Ma Y**, Yu S, Zhao W, Lu Z, Chen J. miR-27a regulates the growth, colony formation and migration of pancreatic cancer cells by targeting Sprouty2. *Cancer Lett* 2010; **298**: 150-158 [PMID: 20638779 DOI: 10.1016/j.canlet.2010.06.012]
  - 54 **Lu Z**, Li Y, Takwi A, Li B, Zhang J, Conklin DJ, Young KH, Martin R, Li Y. miR-301a as an NF- $\kappa$ B activator in pancreatic cancer cells. *EMBO J* 2011; **30**: 57-67 [PMID: 21113131 DOI: 10.1038/emboj.2010.296]
  - 55 **Zhang XJ**, Ye H, Zeng CW, He B, Zhang H, Chen YQ. Dysregulation of miR-15a and miR-214 in human pancreatic cancer. *J Hematol Oncol* 2010; **3**: 46 [PMID: 21106054 DOI: 10.1186/1756-8722-3-46]
  - 56 **Yu S**, Lu Z, Liu C, Meng Y, Ma Y, Zhao W, Liu J, Yu J, Chen J. miRNA-96 suppresses KRAS and functions as a tumor suppressor gene in pancreatic cancer. *Cancer Res* 2010; **70**: 6015-6025 [PMID: 20610624 DOI: 10.1158/0008-5472.CAN-09-4531]
  - 57 **Cortez MA**, Calin GA. MicroRNA identification in plasma and serum: a new tool to diagnose and monitor diseases. *Expert Opin Biol Ther* 2009; **9**: 703-711 [PMID: 19426115 DOI: 10.1517/14712590902932889]
  - 58 **Metias SM**, Lianidou E, Yousef GM. MicroRNAs in clinical oncology: at the crossroads between promises and problems. *J Clin Pathol* 2009; **62**: 771-776 [PMID: 19734473 DOI: 10.1136/jcp.2009.064717]
  - 59 **Ferracin M**, Veronese A, Negrini M. Micromarkers: miRNAs in cancer diagnosis and prognosis. *Expert Rev Mol Diagn* 2010; **10**: 297-308 [PMID: 20370587 DOI: 10.1586/erm.10.11]
  - 60 **Allegra A**, Alonci A, Campo S, Penna G, Petrunaro A, Gerace D, Musolino C. Circulating microRNAs: new biomarkers in diagnosis, prognosis and treatment of cancer (review). *Int J Oncol* 2012; **41**: 1897-1912 [PMID: 23026890 DOI: 10.3892/ijo.2012.1647]
  - 61 **Zen K**, Zhang CY. Circulating microRNAs: a novel class of biomarkers to diagnose and monitor human cancers. *Med Res Rev* 2012; **32**: 326-348 [PMID: 22383180 DOI: 10.1002/med.20215]
  - 62 **Qu H**, Xu W, Huang Y, Yang S. Circulating miRNAs: promising biomarkers of human cancer. *Asian Pac J Cancer Prev* 2011; **12**: 1117-1125 [PMID: 21875254]
  - 63 **Wang J**, Chen J, Chang P, LeBlanc A, Li D, Abbruzzese JL, Frazier ML, Killary AM, Sen S. MicroRNAs in plasma of pancreatic ductal adenocarcinoma patients as novel blood-based biomarkers of disease. *Cancer Prev Res (Phila)* 2009; **2**: 807-813 [PMID: 19723895 DOI: 10.1158/1940-6207.CAPR-09-0094]
  - 64 **Liu J**, Gao J, Du Y, Li Z, Ren Y, Gu J, Wang X, Gong Y, Wang W, Kong X. Combination of plasma microRNAs with serum CA19-9 for early detection of pancreatic cancer. *Int J Cancer* 2012; **131**: 683-691 [PMID: 21913185 DOI: 10.1002/ijc.26422]
  - 65 **Bauer AS**, Keller A, Costello E, Greenhalf W, Bier M, Borries A, Beier M, Neoptolemos J, Büchler M, Werner J, Giese N, Hoheisel JD. Diagnosis of pancreatic ductal adenocarcinoma and chronic pancreatitis by measurement of microRNA abundance in blood and tissue. *PLoS One* 2012; **7**: e34151 [PMID: 22511932 DOI: 10.1371/journal.pone.0034151]
  - 66 **Tavano F**, di Mola FF, Piepoli A, Panza A, Copetti M, Burbacì FP, Latiano T, Pellegrini F, Maiello E, Andriulli A, di Sebastiano P. Changes in miR-143 and miR-21 expression and clinicopathological correlations in pancreatic cancers. *Pancreas* 2012; **41**: 1280-1284 [PMID: 22836856 DOI: 10.1097/MPA.0b013e31824c11f4]
  - 67 **LaConti JJ**, Shivapurkar N, Preet A, Deslattes Mays A, Peran I, Kim SE, Marshall JL, Riegel AT, Wellstein A. Tissue and serum microRNAs in the Kras(G12D) transgenic animal model and in patients with pancreatic cancer. *PLoS One* 2011; **6**: e20687 [PMID: 21738581 DOI: 10.1371/journal.pone.0020687]
  - 68 **Morimura R**, Komatsu S, Ichikawa D, Takeshita H, Tsujiura M, Nagata H, Konishi H, Shiozaki A, Ikoma H, Okamoto K, Ochiai T, Taniguchi H, Otsuji E. Novel diagnostic value of circulating miR-18a in plasma of patients with pancreatic cancer. *Br J Cancer* 2011; **105**: 1733-1740 [PMID: 22045190 DOI: 10.1038/bjc.2011.453]
  - 69 **Kawaguchi T**, Komatsu S, Ichikawa D, Morimura R, Tsujiura M, Konishi H, Takeshita H, Nagata H, Arita T, Hirajima S, Shiozaki A, Ikoma H, Okamoto K, Ochiai T, Taniguchi H, Otsuji E. Clinical impact of circulating miR-221 in plasma of patients with pancreatic cancer. *Br J Cancer* 2013; **108**: 361-369 [PMID: 23329235 DOI: 10.1038/bjc.2012.546]
  - 70 **Wan C**, Shen Y, Yang T, Wang T, Chen L, Wen F. Diagnostic value of microRNA for pancreatic cancer: a meta-analysis. *Arch Med Sci* 2012; **8**: 749-755 [PMID: 23185182 DOI: 10.5114/aoms.2012.31609]
  - 71 **Mo MH**, Chen L, Fu Y, Wang W, Fu SW. Cell-free Circulating miRNA Biomarkers in Cancer. *J Cancer* 2012; **3**: 432-448 [PMID: 23074383 DOI: 10.7150/jca.4919]
  - 72 **Gallo A**, Tandon M, Alevizos I, Illei GG. The majority of microRNAs detectable in serum and saliva is concentrated in exosomes. *PLoS One* 2012; **7**: e30679 [PMID: 22427800 DOI: 10.1371/journal.pone.0030679]
  - 73 **Simons M**, Raposo G. Exosomes--vesicular carriers for

- intercellular communication. *Curr Opin Cell Biol* 2009; **21**: 575-581 [PMID: 19442504 DOI: 10.1016/j.ceb.2009.03.007]
- 74 **Vlassov AV**, Magdaleno S, Setterquist R, Conrad R. Exosomes: current knowledge of their composition, biological functions, and diagnostic and therapeutic potentials. *Biochim Biophys Acta* 2012; **1820**: 940-948 [PMID: 22503788 DOI: 10.1016/j.bbagen.2012.03.017]
- 75 **Mathivanan S**, Ji H, Simpson RJ. Exosomes: extracellular organelles important in intercellular communication. *J Proteomics* 2010; **73**: 1907-1920 [PMID: 20601276 DOI: 10.1016/j.jprot.2010.06.006]
- 76 **Kharaziha P**, Ceder S, Li Q, Panaretakis T. Tumor cell-derived exosomes: a message in a bottle. *Biochim Biophys Acta* 2012; **1826**: 103-111 [PMID: 22503823 DOI: 10.1016/j.bbcan.2012.03.006]
- 77 **Lee TH**, D'Asti E, Magnus N, Al-Nedawi K, Meehan B, Rak J. Microvesicles as mediators of intercellular communication in cancer--the emerging science of cellular 'debris'. *Semin Immunopathol* 2011; **33**: 455-467 [PMID: 21318413 DOI: 10.1007/s00281-011-0250-3]
- 78 **Simpson RJ**, Jensen SS, Lim JW. Proteomic profiling of exosomes: current perspectives. *Proteomics* 2008; **8**: 4083-4099 [PMID: 18780348 DOI: 10.1002/pmic.200800109]
- 79 **Simpson RJ**, Lim JW, Moritz RL, Mathivanan S. Exosomes: proteomic insights and diagnostic potential. *Expert Rev Proteomics* 2009; **6**: 267-283 [PMID: 19489699 DOI: 10.1586/epr.09.17]
- 80 **Rak J**. Microparticles in cancer. *Semin Thromb Hemost* 2010; **36**: 888-906 [PMID: 21049390 DOI: 10.1055/s-0030-1267043]
- 81 **Record M**, Subra C, Silvente-Poirot S, Poirot M. Exosomes as intercellular signalosomes and pharmacological effectors. *Biochem Pharmacol* 2011; **81**: 1171-1182 [PMID: 21371441 DOI: 10.1016/j.bcp.2011.02.011]
- 82 **Ramachandran S**, Palanisamy V. Horizontal transfer of RNAs: exosomes as mediators of intercellular communication. *Wiley Interdiscip Rev RNA* 2012; **3**: 286-293 [PMID: 22012863 DOI: 10.1002/wrna.115]
- 83 **Martins VR**, Dias MS, Hainaut P. Tumor-cell-derived microvesicles as carriers of molecular information in cancer. *Curr Opin Oncol* 2013; **25**: 66-75 [PMID: 23165142 DOI: 10.1097/CCO.0b013e32835b7c81]
- 84 **Gusachenko ON**, Zenkova MA, Vlassov VV. Nucleic acids in exosomes: disease markers and intercellular communication molecules. *Biochemistry (Mosc)* 2013; **78**: 1-7 [PMID: 23379554 DOI: 10.1134/S000629791301001X]
- 85 **Boon RA**, Vickers KC. Intercellular transport of microRNAs. *Arterioscler Thromb Vasc Biol* 2013; **33**: 186-192 [PMID: 23325475 DOI: 10.1161/ATVBAHA.112.300139]
- 86 **Lässer C**. Exosomal RNA as biomarkers and the therapeutic potential of exosome vectors. *Expert Opin Biol Ther* 2012; **12** Suppl 1: S189-S197 [PMID: 22506888 DOI: 10.1517/14712598.2012.680018]
- 87 **Raposo G**, Stoorvogel W. Extracellular vesicles: exosomes, microvesicles, and friends. *J Cell Biol* 2013; **200**: 373-383 [PMID: 23420871 DOI: 10.1083/jcb.201211138]
- 88 **Katzmann DJ**, Odorizzi G, Emr SD. Receptor downregulation and multivesicular-body sorting. *Nat Rev Mol Cell Biol* 2002; **3**: 893-905 [PMID: 12461556]
- 89 **Février B**, Raposo G. Exosomes: endosomal-derived vesicles shipping extracellular messages. *Curr Opin Cell Biol* 2004; **16**: 415-421 [PMID: 15261674]
- 90 **Buschow SI**, Nolte-t Hoen EN, van Niel G, Pols MS, ten Broeke T, Lauwen M, Ossendorp F, Melief CJ, Raposo G, Wubbolts R, Wauben MH, Stoorvogel W. MHC II in dendritic cells is targeted to lysosomes or T cell-induced exosomes via distinct multivesicular body pathways. *Traffic* 2009; **10**: 1528-1542 [PMID: 19682328 DOI: 10.1111/j.1600-0854.2009.00963.x]
- 91 **Zöllner M**. Tetraspanins: push and pull in suppressing and promoting metastasis. *Nat Rev Cancer* 2009; **9**: 40-55 [PMID: 19078974 DOI: 10.1038/nrc2543]
- 92 **Blanc L**, Vidal M. Reticulocyte membrane remodeling: contribution of the exosome pathway. *Curr Opin Hematol* 2010; **17**: 177-183 [PMID: 20173636 DOI: 10.1097/MOH.0b013e328337b4e3]
- 93 **Hurley JH**, Emr SD. The ESCRT complexes: structure and mechanism of a membrane-trafficking network. *Annu Rev Biophys Biomol Struct* 2006; **35**: 277-298 [PMID: 16689637]
- 94 **Trajkovic K**, Hsu C, Chiantia S, Rajendran L, Wenzel D, Wieland F, Schwille P, Brügger B, Simons M. Ceramide triggers budding of exosome vesicles into multivesicular endosomes. *Science* 2008; **319**: 1244-1247 [PMID: 18309083 DOI: 10.1126/science.1153124]
- 95 **Fang Y**, Wu N, Gan X, Yan W, Morrell JC, Gould SJ. Higher-order oligomerization targets plasma membrane proteins and HIV gag to exosomes. *PLoS Biol* 2007; **5**: e158 [PMID: 17550307]
- 96 **Rana S**, Claas C, Kretz CC, Nazarenko I, Zoeller M. Activation-induced internalization differs for the tetraspanins CD9 and Tspan8: Impact on tumor cell motility. *Int J Biochem Cell Biol* 2011; **43**: 106-119 [PMID: 20937409 DOI: 10.1016/j.biocel.2010.10.002]
- 97 **Rana S**, Yue S, Stadel D, Zöllner M. Toward tailored exosomes: the exosomal tetraspanin web contributes to target cell selection. *Int J Biochem Cell Biol* 2012; **44**: 1574-1584 [PMID: 22728313 DOI: 10.1016/j.biocel.2012.06.018]
- 98 **Nazarenko I**, Rana S, Baumann A, McAlear J, Hellwig A, Trendelenburg M, Lochnit G, Preissner KT, Zöllner M. Cell surface tetraspanin Tspan8 contributes to molecular pathways of exosome-induced endothelial cell activation. *Cancer Res* 2010; **70**: 1668-1678 [PMID: 20124479 DOI: 10.1158/0008-5472.CAN-09-2470]
- 99 **Pols MS**, Klumperman J. Trafficking and function of the tetraspanin CD63. *Exp Cell Res* 2009; **315**: 1584-1592 [PMID: 18930046 DOI: 10.1016/j.yexcr.2008.09.020]
- 100 **Cho JA**, Lee YS, Kim SH, Ko JK, Kim CW. MHC independent anti-tumor immune responses induced by Hsp70-enriched exosomes generate tumor regression in murine models. *Cancer Lett* 2009; **275**: 256-265 [PMID: 19036499 DOI: 10.1016/j.canlet.2008.10.021]
- 101 **Xie Y**, Bai O, Zhang H, Yuan J, Zong S, Chibbar R, Slatery K, Qureshi M, Wei Y, Deng Y, Xiang J. Membrane-bound HSP70-engineered myeloma cell-derived exosomes stimulate more efficient CD8(+) CTL- and NK-mediated antitumor immunity than exosomes released from heat-shocked tumour cells expressing cytoplasmic HSP70. *J Cell Mol Med* 2010; **14**: 2655-2666 [PMID: 19627400 DOI: 10.1111/j.1582-4934.2009.00851.x]
- 102 **Shimoda M**, Khokha R. Proteolytic factors in exosomes. *Proteomics* 2013; **13**: 1624-1636 [PMID: 23526769 DOI: 10.1002/pmic.201200458]
- 103 **Mu W**, Rana S, Zöllner M. Host matrix modulation by tumor exosomes promotes motility and invasiveness. *Neoplasia* 2013; **15**: 875-877 [PMID: 23908589]
- 104 **Henderson MC**, Azorsa DO. High-throughput RNAi screening for the identification of novel targets. *Methods Mol Biol* 2013; **986**: 89-95 [PMID: 23436407 DOI: 10.3389/fonc.2012.00038]
- 105 **Sahu R**, Kaushik S, Clement CC, Cannizzo ES, Scharf B, Follenzi A, Potolicchio I, Nieves E, Cuervo AM, Santambrogio L. Microautophagy of cytosolic proteins by late endosomes. *Dev Cell* 2011; **20**: 131-139 [PMID: 21238931 DOI: 10.1016/j.devcel.2010.12.003]
- 106 **Valadi H**, Ekström K, Bossios A, Sjöstrand M, Lee JJ, Lötvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol* 2007; **9**: 654-659 [PMID: 17486113]
- 107 **Pant S**, Hilton H, Burczynski ME. The multifaceted exosome: biogenesis, role in normal and aberrant cellular function, and frontiers for pharmacological and biomarker opportunities. *Biochem Pharmacol* 2012; **83**: 1484-1494 [PMID: 22230477 DOI: 10.1016/j.bcp.2011.12.037]
- 108 **Skog J**, Würdinger T, van Rijn S, Meijer DH, Gainche L, Sena-Esteves M, Curry WT, Carter BS, Krichevsky AM,

- Breakefield XO. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell Biol* 2008; **10**: 1470-1476 [PMID: 19011622 DOI: 10.1038/ncb1800]
- 109 **Rana S**, Malinowska K, Zöller M. Exosomal tumor microRNA modulates premetastatic organ cells. *Neoplasia* 2013; **15**: 281-295 [PMID: 23479506]
- 110 **Gibbins DJ**, Ciaudo C, Erhardt M, Voinnet O. Multivesicular bodies associate with components of miRNA effector complexes and modulate miRNA activity. *Nat Cell Biol* 2009; **11**: 1143-1149 [PMID: 19684575 DOI: 10.1038/ncb1929]
- 111 **Lee YS**, Pressman S, Andress AP, Kim K, White JL, Cassidy JJ, Li X, Lubell K, Lim do H, Cho IS, Nakahara K, Preall JB, Bellare P, Sontheimer EJ, Carthew RW. Silencing by small RNAs is linked to endosomal trafficking. *Nat Cell Biol* 2009; **11**: 1150-1156 [PMID: 19684574 DOI: 10.1038/ncb1930]
- 112 **Kosaka N**, Iguchi H, Yoshioka Y, Takeshita F, Matsuki Y, Ochiya T. Secretory mechanisms and intercellular transfer of microRNAs in living cells. *J Biol Chem* 2010; **285**: 17442-17452 [PMID: 20353945 DOI: 10.1074/jbc.M110.107821]
- 113 **Wittmann J**, Jäck HM. Serum microRNAs as powerful cancer biomarkers. *Biochim Biophys Acta* 2010; **1806**: 200-207 [PMID: 20637263 DOI: 10.1016/j.bbcan.2010.07.002]
- 114 **Tauro BJ**, Greening DW, Mathias RA, Ji H, Mathivanan S, Scott AM, Simpson RJ. Comparison of ultracentrifugation, density gradient separation, and immunoaffinity capture methods for isolating human colon cancer cell line LIM1863-derived exosomes. *Methods* 2012; **56**: 293-304 [PMID: 22285593 DOI: 10.1016/j.ymeth.2012.01.002]
- 115 **Cantin R**, Diou J, Bélanger D, Tremblay AM, Gilbert C. Discrimination between exosomes and HIV-1: purification of both vesicles from cell-free supernatants. *J Immunol Methods* 2008; **338**: 21-30 [PMID: 18675270 DOI: 10.1016/j.jim.2008.07.007]
- 116 **Lässer C**, Eldh M, Lötvall J. Isolation and characterization of RNA-containing exosomes. *J Vis Exp* 2012; **9**: e3037 [PMID: 22257828 DOI: 10.3791/3037]
- 117 **Burger D**, Schock S, Thompson CS, Montezano AC, Hakim AM, Touyz RM. Microparticles: biomarkers and beyond. *Clin Sci (Lond)* 2013; **124**: 423-441 [PMID: 23249271 DOI: 10.1042/CS20120309]
- 118 **Chen CL**, Lai YF, Tang P, Chien KY, Yu JS, Tsai CH, Chen HW, Wu CC, Chung T, Hsu CW, Chen CD, Chang YS, Chang PL, Chen YT. Comparative and targeted proteomic analyses of urinary microparticles from bladder cancer and hernia patients. *J Proteome Res* 2012; **11**: 5611-5629 [PMID: 23082778 DOI: 10.1021/pr3008732]
- 119 **Orozco AF**, Lewis DE. Flow cytometric analysis of circulating microparticles in plasma. *Cytometry A* 2010; **77**: 502-514 [PMID: 20235276 DOI: 10.1002/cyto.a.20886]
- 120 **Kim G**, Yoo CE, Kim M, Kang HJ, Park D, Lee M, Huh N. Noble polymeric surface conjugated with zwitterionic moieties and antibodies for the isolation of exosomes from human serum. *Bioconjug Chem* 2012; **23**: 2114-2120 [PMID: 23025585 DOI: 10.1021/bc300339b]
- 121 **Taylor DD**, Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol Oncol* 2008; **110**: 13-21 [PMID: 18589210 DOI: 10.1016/j.ygyno.2008.04.033]
- 122 **Bryant RJ**, Pawlowski T, Catto JW, Marsden G, Vessella RL, Rhee B, Kuslich C, Visakorpi T, Hamdy FC. Changes in circulating microRNA levels associated with prostate cancer. *Br J Cancer* 2012; **106**: 768-774 [PMID: 22240788 DOI: 10.1038/bjc.2011.595]
- 123 **Dick JE**. Stem cell concepts renew cancer research. *Blood* 2008; **112**: 4793-4807 [PMID: 19064739 DOI: 10.1182/blood-2008-08-077941]
- 124 **Wang H**, Rana S, Giese N, Büchler MW, Zöller M. Tspan8, CD44v6 and  $\alpha 6\beta 4$  are biomarkers of migrating pancreatic cancer-initiating cells. *Int J Cancer* 2013; **133**: 416-426 [PMID: 23338841 DOI: 10.1002/ijc.28044]
- 125 **Ischenko I**, Seeliger H, Kleespies A, Angele MK, Eichhorn ME, Jauch KW, Bruns CJ. Pancreatic cancer stem cells: new understanding of tumorigenesis, clinical implications. *Langenbecks Arch Surg* 2010; **395**: 1-10 [PMID: 19421768 DOI: 10.1007/s00423-009-0502-z]
- 126 **Quante M**, Wang TC. Stem cells in gastroenterology and hepatology. *Nat Rev Gastroenterol Hepatol* 2009; **6**: 724-737 [PMID: 19884893 DOI: 10.1038/nrgastro.2009.195]
- 127 **Lonardo E**, Hermann PC, Heeschen C. Pancreatic cancer stem cells - update and future perspectives. *Mol Oncol* 2010; **4**: 431-442 [PMID: 20580623 DOI: 10.1016/j.molonc.2010.06.002]
- 128 **Rupp AK**, Rupp C, Keller S, Brase JC, Ehehalt R, Fogel M, Moldenhauer G, Marmé F, Sültmann H, Altevogt P. Loss of EpCAM expression in breast cancer derived serum exosomes: role of proteolytic cleavage. *Gynecol Oncol* 2011; **122**: 437-446 [PMID: 21601258 DOI: 10.1016/j.ygyno.2011.04.035]
- 129 **Keller S**, König AK, Marmé F, Runz S, Wolterink S, Koensgen D, Mustea A, Sehoul J, Altevogt P. Systemic presence and tumor-growth promoting effect of ovarian carcinoma released exosomes. *Cancer Lett* 2009; **278**: 73-81 [PMID: 19188015 DOI: 10.1016/j.canlet.2008.12.028]
- 130 **Li J**, Sherman-Baust CA, Tsai-Turton M, Bristow RE, Roden RB, Morin PJ. Claudin-containing exosomes in the peripheral circulation of women with ovarian cancer. *BMC Cancer* 2009; **9**: 244 [PMID: 19619303 DOI: 10.1186/1471-2407-9-244]
- 131 **Khan S**, Jutzy JM, Valenzuela MM, Turay D, Aspe JR, Ashok A, Mirshahidi S, Mercola D, Lilly MB, Wall NR. Plasma-derived exosomal survivin, a plausible biomarker for early detection of prostate cancer. *PLoS One* 2012; **7**: e46737 [PMID: 23091600 DOI: 10.1371/journal.pone.0046737]
- 132 **Nilsson J**, Skog J, Nordstrand A, Baranov V, Mincheva-Nilsson L, Breakefield XO, Widmark A. Prostate cancer-derived urine exosomes: a novel approach to biomarkers for prostate cancer. *Br J Cancer* 2009; **100**: 1603-1607 [PMID: 19401683 DOI: 10.1038/sj.bjc.6605058]
- 133 **Mitchell PJ**, Welton J, Staffurth J, Court J, Mason MD, Tabi Z, Clayton A. Can urinary exosomes act as treatment response markers in prostate cancer? *J Transl Med* 2009; **7**: 4 [PMID: 19138409 DOI: 10.1186/1479-5876-7-4]
- 134 **Logozzi M**, De Milito A, Lugini L, Borghi M, Calabrò L, Spada M, Perdicchio M, Marino ML, Federici C, Iessi E, Brambilla D, Venturi G, Lozupone F, Santinami M, Huber V, Maio M, Rivoltini L, Fais S. High levels of exosomes expressing CD63 and caveolin-1 in plasma of melanoma patients. *PLoS One* 2009; **4**: e5219 [PMID: 19381331 DOI: 10.1371/journal.pone.0005219]
- 135 **Koga K**, Matsumoto K, Akiyoshi T, Kubo M, Yamanaka N, Tasaki A, Nakashima H, Nakamura M, Kuroki S, Tanaka M, Katano M. Purification, characterization and biological significance of tumor-derived exosomes. *Anticancer Res* 2005; **25**: 3703-3707 [PMID: 16302729]
- 136 **Bauer M**, Pelkmans L. A new paradigm for membrane-organizing and -shaping scaffolds. *FEBS Lett* 2006; **580**: 5559-5564 [PMID: 16996501]
- 137 **Nguyen J**, Szoka FC. Nucleic acid delivery: the missing pieces of the puzzle? *Acc Chem Res* 2012; **45**: 1153-1162 [PMID: 22428908 DOI: 10.1021/ar3000162]
- 138 **Vickers KC**, Remaley AT. Lipid-based carriers of microRNAs and intercellular communication. *Curr Opin Lipidol* 2012; **23**: 91-97 [PMID: 22418571 DOI: 10.1097/MOL.0b013e328350a425]
- 139 **Zakharova L**, Svetlova M, Fomina AF. T cell exosomes induce cholesterol accumulation in human monocytes via phosphatidylserine receptor. *J Cell Physiol* 2007; **212**: 174-181 [PMID: 17299798]
- 140 **Feng D**, Zhao WL, Ye YY, Bai XC, Liu RQ, Chang LF, Zhou Q, Sui SF. Cellular internalization of exosomes occurs through phagocytosis. *Traffic* 2010; **11**: 675-687 [PMID: 20136776 DOI: 10.1111/j.1600-0854.2010.01041.x]
- 141 **Zech D**, Rana S, Büchler MW, Zöller M. Tumor-exosomes and

- leukocyte activation: an ambivalent crosstalk. *Cell Commun Signal* 2012; **10**: 37 [PMID: 23190502 DOI: 10.1186/1478-811X-10-37]
- 142 **Runz S**, Keller S, Rupp C, Stoeck A, Issa Y, Koensgen D, Mustea A, Sehouli J, Kristiansen G, Altevogt P. Malignant ascites-derived exosomes of ovarian carcinoma patients contain CD24 and EpCAM. *Gynecol Oncol* 2007; **107**: 563-571 [PMID: 17900673]
- 143 **Morelli AE**, Larregina AT, Shufesky WJ, Sullivan ML, Stolz DB, Papworth GD, Zahorchak AF, Logar AJ, Wang Z, Watkins SC, Falo LD, Thomson AW. Endocytosis, intracellular sorting, and processing of exosomes by dendritic cells. *Blood* 2004; **104**: 3257-3266 [PMID: 15284116]
- 144 **Atay S**, Gercel-Taylor C, Taylor DD. Human trophoblast-derived exosomal fibronectin induces pro-inflammatory IL-1 $\beta$  production by macrophages. *Am J Reprod Immunol* 2011; **66**: 259-269 [PMID: 21410811 DOI: 10.1111/j.1600-0897.2011.00995.x]
- 145 **Mathivanan S**, Lim JW, Tauro BJ, Ji H, Moritz RL, Simpson RJ. Proteomics analysis of A33 immunoaffinity-purified exosomes released from the human colon tumor cell line LIM1215 reveals a tissue-specific protein signature. *Mol Cell Proteomics* 2010; **9**: 197-208 [PMID: 19837982 DOI: 10.1074/mcp.M900152-MCP200]
- 146 **Perez-Hernandez D**, Gutiérrez-Vázquez C, Jorge I, López-Martín S, Ursa A, Sánchez-Madrid F, Vázquez J, Yáñez-Mó M. The intracellular interactome of tetraspanin-enriched microdomains reveals their function as sorting machineries toward exosomes. *J Biol Chem* 2013; **288**: 11649-11661 [PMID: 23463506 DOI: 10.1074/jbc.M112.445304]
- 147 **Lakkaraju A**, Rodriguez-Boulan E. Itinerant exosomes: emerging roles in cell and tissue polarity. *Trends Cell Biol* 2008; **18**: 199-209 [PMID: 18396047 DOI: 10.1016/j.tcb.2008.03.002]
- 148 **Tian T**, Wang Y, Wang H, Zhu Z, Xiao Z. Visualizing of the cellular uptake and intracellular trafficking of exosomes by live-cell microscopy. *J Cell Biochem* 2010; **111**: 488-496 [PMID: 20533300 DOI: 10.1002/jcb.22733]
- 149 **Skokos D**, Botros HG, Demeure C, Morin J, Peronet R, Birkenmeier G, Boudaly S, Mécheri S. Mast cell-derived exosomes induce phenotypic and functional maturation of dendritic cells and elicit specific immune responses in vivo. *J Immunol* 2003; **170**: 3037-3045 [PMID: 12626558]
- 150 **Parolini I**, Federici C, Raggi C, Lugini L, Palleschi S, De Miliato A, Coscia C, Iessi E, Logozzi M, Molinari A, Colone M, Tatti M, Sargiacomo M, Fais S. Microenvironmental pH is a key factor for exosome traffic in tumor cells. *J Biol Chem* 2009; **284**: 34211-34222 [PMID: 19801663 DOI: 10.1074/jbc.M109.041152]
- 151 **Xu D**, Sharma C, Hemler ME. Tetraspanin12 regulates ADAM10-dependent cleavage of amyloid precursor protein. *FASEB J* 2009; **23**: 3674-3681 [PMID: 19587294 DOI: 10.1096/fj.09-133462]
- 152 **Arduise C**, Abache T, Li L, Billard M, Chabanon A, Ludwig A, Mauduit P, Boucheix C, Rubinstein E, Le Naour F. Tetraspanins regulate ADAM10-mediated cleavage of TNF- $\alpha$  and epidermal growth factor. *J Immunol* 2008; **181**: 7002-7013 [PMID: 18981120]
- 153 **Gutiérrez-López MD**, Gilsanz A, Yáñez-Mó M, Ovalle S, Lafuente EM, Domínguez C, Monk PN, González-Alvaro I, Sánchez-Madrid F, Cabañas C. The sheddase activity of ADAM17/TACE is regulated by the tetraspanin CD9. *Cell Mol Life Sci* 2011; **68**: 3275-3292 [PMID: 21365281 DOI: 10.1007/s00018-011-0639-0]
- 154 **Potolicchio I**, Carven GJ, Xu X, Stipp C, Riese RJ, Stern LJ, Santambrogio L. Proteomic analysis of microglia-derived exosomes: metabolic role of the aminopeptidase CD13 in neuropeptide catabolism. *J Immunol* 2005; **175**: 2237-2243 [PMID: 16081791]
- 155 **Le Naour F**, André M, Boucheix C, Rubinstein E. Membrane microdomains and proteomics: lessons from tetraspanin microdomains and comparison with lipid rafts. *Proteomics* 2006; **6**: 6447-6454 [PMID: 17109380]
- 156 **Yáñez-Mó M**, Barreiro O, Gonzalo P, Batista A, Megías D, Genís L, Sachs N, Sala-Valdés M, Alonso MA, Montoya MC, Sonnenberg A, Arroyo AG, Sánchez-Madrid F. MT1-MMP collagenolytic activity is regulated through association with tetraspanin CD151 in primary endothelial cells. *Blood* 2008; **112**: 3217-3226 [PMID: 18663148 DOI: 10.1182/blood-2008-02-139394]
- 157 **Hendrix A**, Westbroek W, Bracke M, De Wever O. An ex(o) citing machinery for invasive tumor growth. *Cancer Res* 2010; **70**: 9533-9537 [PMID: 21098711 DOI: 10.1158/0008-5472.CAN-10-3248]
- 158 **Grange C**, Tapparo M, Collino F, Vitillo L, Damasco C, Deregibus MC, Tetta C, Bussolati B, Camussi G. Microvesicles released from human renal cancer stem cells stimulate angiogenesis and formation of lung premetastatic niche. *Cancer Res* 2011; **71**: 5346-5356 [PMID: 21670082 DOI: 10.1158/0008-5472.CAN-11-0241]
- 159 **Lafleur MA**, Xu D, Hemler ME. Tetraspanin proteins regulate membrane type-1 matrix metalloproteinase-dependent pericellular proteolysis. *Mol Biol Cell* 2009; **20**: 2030-2040 [PMID: 19211836 DOI: 10.1091/mbc.E08-11-1149]
- 160 **Hakulinen J**, Sankkila L, Sugiyama N, Lehti K, Keski-Oja J. Secretion of active membrane type 1 matrix metalloproteinase (MMP-14) into extracellular space in microvesicular exosomes. *J Cell Biochem* 2008; **105**: 1211-1218 [PMID: 18802920 DOI: 10.1002/jcb.21923]
- 161 **Nieuwland R**, van der Post JA, Lok CA, Kenter G, Sturk A. Microparticles and exosomes in gynecologic neoplasias. *Semin Thromb Hemost* 2010; **36**: 925-929 [PMID: 21049392 DOI: 10.1055/s-0030-1267046]
- 162 **Park JE**, Tan HS, Datta A, Lai RC, Zhang H, Meng W, Lim SK, Sze SK. Hypoxic tumor cell modulates its microenvironment to enhance angiogenic and metastatic potential by secretion of proteins and exosomes. *Mol Cell Proteomics* 2010; **9**: 1085-1099 [PMID: 20124223 DOI: 10.1074/mcp.M900381-MCP200]
- 163 **Nakada M**, Miyamori H, Kita D, Takahashi T, Yamashita J, Sato H, Miura R, Yamaguchi Y, Okada Y. Human glioblastomas overexpress ADAMTS-5 that degrades brevican. *Acta Neuropathol* 2005; **110**: 239-246 [PMID: 16133547]
- 164 **Lo Cicero A**, Majkowska I, Nagase H, Di Liegro I, Troeberg L. Microvesicles shed by oligodendrogloma cells and rheumatoid synovial fibroblasts contain aggrecanase activity. *Matrix Biol* 2012; **31**: 229-233 [PMID: 22406378 DOI: 10.1016/j.matbio.2012.02.005]
- 165 **Ginestra A**, La Placa MD, Saladino F, Cassarà D, Nagase H, Vittorelli ML. The amount and proteolytic content of vesicles shed by human cancer cell lines correlates with their in vitro invasiveness. *Anticancer Res* 1998; **18**: 3433-3437 [PMID: 9858920]
- 166 **Ngora H**, Galli UM, Miyazaki K, Zöller M. Membrane-bound and exosomal metastasis-associated C4.4A promotes migration by associating with the  $\alpha(6)\beta(4)$  integrin and MT1-MMP. *Neoplasia* 2012; **14**: 95-107 [PMID: 22431918]
- 167 **Sangaletti S**, Colombo MP. Matricellular proteins at the crossroad of inflammation and cancer. *Cancer Lett* 2008; **267**: 245-253 [PMID: 18471960 DOI: 10.1016/j.canlet.2008.03.027]
- 168 **Clayton A**, Mitchell JP, Court J, Mason MD, Tabi Z. Human tumor-derived exosomes selectively impair lymphocyte responses to interleukin-2. *Cancer Res* 2007; **67**: 7458-7466 [PMID: 17671216]
- 169 **Delcayre A**, Shu H, Le Pecq JB. Dendritic cell-derived exosomes in cancer immunotherapy: exploiting nature's antigen delivery pathway. *Expert Rev Anticancer Ther* 2005; **5**: 537-547 [PMID: 16001959]
- 170 **Artavanis-Tsakonas K**, Kasperkovitz PV, Papa E, Cardenas ML, Khan NS, Van der Veen AG, Ploegh HL, Vyas JM. The tetraspanin CD82 is specifically recruited to fungal and bacterial phagosomes prior to acidification. *Infect Immun* 2011; **79**: 1098-1106 [PMID: 21149584 DOI: 10.1128/IAI.01135-10]
- 171 **Tumne A**, Prasad VS, Chen Y, Stolz DB, Saha K, Ratner DM,

- Ding M, Watkins SC, Gupta P. Noncytotoxic suppression of human immunodeficiency virus type 1 transcription by exosomes secreted from CD8+ T cells. *J Virol* 2009; **83**: 4354-4364 [PMID: 19193788 DOI: 10.1128/JVI.02629-08]
- 172 **Taylor DD**, Gercel-Taylor C. Exosomes/microvesicles: mediators of cancer-associated immunosuppressive microenvironments. *Semin Immunopathol* 2011; **33**: 441-454 [PMID: 21688197 DOI: 10.1007/s00281-010-0234-8]
- 173 **Zhang HG**, Kim H, Liu C, Yu S, Wang J, Grizzle WE, Kimberly RP, Barnes S. Curcumin reverses breast tumor exosomes mediated immune suppression of NK cell tumor cytotoxicity. *Biochim Biophys Acta* 2007; **1773**: 1116-1123 [PMID: 17555831]
- 174 **Ashiru O**, Boutet P, Fernández-Messina L, Agüera-González S, Skepper JN, Valés-Gómez M, Reyburn HT. Natural killer cell cytotoxicity is suppressed by exposure to the human NKG2D ligand MICA\*008 that is shed by tumor cells in exosomes. *Cancer Res* 2010; **70**: 481-489 [PMID: 20068167 DOI: 10.1158/0008-5472.CAN-09-1688]
- 175 **Yuan XK**, Zhao XK, Xia YC, Zhu X, Xiao P. Increased circulating immunosuppressive CD14(+)-HLA-DR(-)/low cells correlate with clinical cancer stage and pathological grade in patients with bladder carcinoma. *J Int Med Res* 2011; **39**: 1381-1391 [PMID: 21986138]
- 176 **Lima LG**, Chammas R, Monteiro RQ, Moreira ME, Barcinski MA. Tumor-derived microvesicles modulate the establishment of metastatic melanoma in a phosphatidylserine-dependent manner. *Cancer Lett* 2009; **283**: 168-175 [PMID: 19401262 DOI: 10.1016/j.canlet.2009.03.041]
- 177 **Lee HM**, Choi EJ, Kim JH, Kim TD, Kim YK, Kang C, Gho YS. A membranous form of ICAM-1 on exosomes efficiently blocks leukocyte adhesion to activated endothelial cells. *Biochem Biophys Res Commun* 2010; **397**: 251-256 [PMID: 20529672 DOI: 10.1016/j.bbrc.2010.05.094]
- 178 **Khalil AA**, Kabapy NF, Deraz SF, Smith C. Heat shock proteins in oncology: diagnostic biomarkers or therapeutic targets? *Biochim Biophys Acta* 2011; **1816**: 89-104 [PMID: 21605630 DOI: 10.1016/j.bbcan.2011.05.001]
- 179 **Elsner L**, Muppala V, Gehrman M, Lozano J, Malzahn D, Bickeböller H, Brunner E, Zientkowska M, Herrmann T, Walter L, Alves F, Multhoff G, Dressel R. The heat shock protein HSP70 promotes mouse NK cell activity against tumors that express inducible NKG2D ligands. *J Immunol* 2007; **179**: 5523-5533 [PMID: 17911639]
- 180 **Dai S**, Wan T, Wang B, Zhou X, Xiu F, Chen T, Wu Y, Cao X. More efficient induction of HLA-A\*0201-restricted and carcinoembryonic antigen (CEA)-specific CTL response by immunization with exosomes prepared from heat-stressed CEA-positive tumor cells. *Clin Cancer Res* 2005; **11**: 7554-7563 [PMID: 16243831]
- 181 **Hurwitz MD**, Kaur P, Nagaraja GM, Bausero MA, Manola J, Asea A. Radiation therapy induces circulating serum Hsp72 in patients with prostate cancer. *Radiother Oncol* 2010; **95**: 350-358 [PMID: 20430459 DOI: 10.1016/j.radonc.2010.03.024]
- 182 **Xiu F**, Cai Z, Yang Y, Wang X, Wang J, Cao X. Surface anchorage of superantigen SEA promotes induction of specific antitumor immune response by tumor-derived exosomes. *J Mol Med (Berl)* 2007; **85**: 511-521 [PMID: 17219095]
- 183 **Chen T**, Guo J, Yang M, Zhu X, Cao X. Chemokine-containing exosomes are released from heat-stressed tumor cells via lipid raft-dependent pathway and act as efficient tumor vaccine. *J Immunol* 2011; **186**: 2219-2228 [PMID: 21242526 DOI: 10.4049/jimmunol.1002991]
- 184 **Zeelenberg IS**, van Maren WW, Boissonnas A, Van Hout-Kuijper MA, Den Brok MH, Wagenaars JA, van der Schaaf A, Jansen EJ, Amigorena S, Théry C, Figdor CG, Adema GJ. Antigen localization controls T cell-mediated tumor immunity. *J Immunol* 2011; **187**: 1281-1288 [PMID: 21705625 DOI: 10.4049/jimmunol.1003905]
- 185 **Hood JL**, San RS, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. *Cancer Res* 2011; **71**: 3792-3801 [PMID: 21478294 DOI: 10.1158/0008-5472.CAN-10-4455]
- 186 **Sheldon H**, Heikamp E, Turley H, Dragovic R, Thomas P, Oon CE, Leek R, Edelmann M, Kessler B, Sainson RC, Sargent I, Li JL, Harris AL. New mechanism for Notch signaling to endothelium at a distance by Delta-like 4 incorporation into exosomes. *Blood* 2010; **116**: 2385-2394 [PMID: 20558614 DOI: 10.1182/blood-2009-08-239228]
- 187 **Benameur T**, Tual-Chalot S, Andriantsitohaina R, Martínez MC. PPAR $\alpha$  is essential for microparticle-induced differentiation of mouse bone marrow-derived endothelial progenitor cells and angiogenesis. *PLoS One* 2010; **5**: e12392 [PMID: 20811625 DOI: 10.1371/journal.pone.0012392]
- 188 **Castellana D**, Zobairi F, Martinez MC, Panaro MA, Mitolo V, Freyssinet JM, Kunzelmann C. Membrane microvesicles as actors in the establishment of a favorable prostatic tumoral niche: a role for activated fibroblasts and CX3CL1-CX3CR1 axis. *Cancer Res* 2009; **69**: 785-793 [PMID: 19155311 DOI: 10.1158/0008-5472.CAN-08-1946]
- 189 **Chairoungdua A**, Smith DL, Pochard P, Hull M, Caplan MJ. Exosome release of  $\beta$ -catenin: a novel mechanism that antagonizes Wnt signaling. *J Cell Biol* 2010; **190**: 1079-1091 [PMID: 20837771 DOI: 10.1083/jcb.201002049]
- 190 **Hupalowska A**, Miaczynska M. The new faces of endocytosis in signaling. *Traffic* 2012; **13**: 9-18 [PMID: 21752167 DOI: 10.1111/j.1600-0854.2011.01249.x]
- 191 **Bobrie A**, Krumeich S, Reyat F, Recchi C, Moita LF, Seabra MC, Ostrowski M, Théry C. Rab27a supports exosome-dependent and -independent mechanisms that modify the tumor microenvironment and can promote tumor progression. *Cancer Res* 2012; **72**: 4920-4930 [PMID: 22865453 DOI: 10.1158/0008-5472.CAN-12-0925]
- 192 **Aliotta JM**, Lee D, Puente N, Faradyan S, Sears EH, Amaral A, Goldberg L, Dooner MS, Pereira M, Quesenberry PJ. Progenitor/stem cell fate determination: interactive dynamics of cell cycle and microvesicles. *Stem Cells Dev* 2012; **21**: 1627-1638 [PMID: 22214238 DOI: 10.1089/scd.2011.0550]
- 193 **Pan Q**, Ramakrishnaiah V, Henry S, Fouraschen S, de Ruyter PE, Kwekkeboom J, Tilanus HW, Janssen HL, van der Laan LJ. Hepatic cell-to-cell transmission of small silencing RNA can extend the therapeutic reach of RNA interference (RNAi). *Gut* 2012; **61**: 1330-1339 [PMID: 22198713 DOI: 10.1136/gutjnl-2011-300449]
- 194 **Al-Nedawi K**, Meehan B, Micallef J, Lhotak V, May L, Guha A, Rak J. Intercellular transfer of the oncogenic receptor EGFRvIII by microvesicles derived from tumour cells. *Nat Cell Biol* 2008; **10**: 619-624 [PMID: 18425114 DOI: 10.1038/ncb1725]
- 195 **Balaj L**, Lessard R, Dai L, Cho YJ, Pomeroy SL, Breakefield XO, Skog J. Tumour microvesicles contain retrotransposon elements and amplified oncogene sequences. *Nat Commun* 2011; **2**: 180 [PMID: 21285958 DOI: 10.1038/ncomms1180]
- 196 **Demory Beckler M**, Higginbotham JN, Franklin JL, Ham AJ, Halvey PJ, Imasuen IE, Whitwell C, Li M, Liebler DC, Coffey RJ. Proteomic analysis of exosomes from mutant KRAS colon cancer cells identifies intercellular transfer of mutant KRAS. *Mol Cell Proteomics* 2013; **12**: 343-355 [PMID: 23161513 DOI: 10.1074/mcp.M112.022806]
- 197 **Verweij FJ**, Middeldorp JM, Pegtel DM. Intracellular signaling controlled by the endosomal-exosomal pathway. *Commun Integr Biol* 2012; **5**: 88-93 [PMID: 22482020]
- 198 **Higginbotham JN**, Demory Beckler M, Gephart JD, Franklin JL, Bogatcheva G, Kremers GJ, Piston DW, Ayers GD, McConnell RE, Tyska MJ, Coffey RJ. Amphiregulin exosomes increase cancer cell invasion. *Curr Biol* 2011; **21**: 779-786 [PMID: 21514161 DOI: 10.1016/j.cub.2011.03.043]
- 199 **Antonyak MA**, Li B, Boroughs LK, Johnson JL, Druso JE, Bryant KL, Holowka DA, Cerione RA. Cancer cell-derived microvesicles induce transformation by transferring tissue transglutaminase and fibronectin to recipient cells. *Proc Natl*

- Acad Sci USA* 2011; **108**: 4852-4857 [PMID: 21368175 DOI: 10.1073/pnas.1017667108]
- 200 **Peinado H**, Alečković M, Lavotshkin S, Matei I, Costa-Silva B, Moreno-Bueno G, Hergueta-Redondo M, Williams C, García-Santos G, Ghajar C, Nitadori-Hoshino A, Hoffman C, Badal K, Garcia BA, Callahan MK, Yuan J, Martins VR, Skog J, Kaplan RN, Brady MS, Wolchok JD, Chapman PB, Kang Y, Bromberg J, Lyden D. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. *Nat Med* 2012; **18**: 883-891 [PMID: 22635005 DOI: 10.1038/nm.2753]
- 201 **Khan S**, Jutzy JM, Aspe JR, McGregor DW, Neidigh JW, Wall NR. Survivin is released from cancer cells via exosomes. *Apoptosis* 2011; **16**: 1-12 [PMID: 20717727 DOI: 10.1007/s10495-010-0534-4]
- 202 **Webber J**, Steadman R, Mason MD, Tabi Z, Clayton A. Cancer exosomes trigger fibroblast to myofibroblast differentiation. *Cancer Res* 2010; **70**: 9621-9630 [PMID: 21098712 DOI: 10.1158/0008-5472.CAN-10-1722]
- 203 **Cho JA**, Park H, Lim EH, Lee KW. Exosomes from breast cancer cells can convert adipose tissue-derived mesenchymal stem cells into myofibroblast-like cells. *Int J Oncol* 2012; **40**: 130-138 [PMID: 21904773 DOI: 10.3892/ijo.2011.1193]
- 204 **Wysoczynski M**, Ratajczak MZ. Lung cancer secreted microvesicles: underappreciated modulators of microenvironment in expanding tumors. *Int J Cancer* 2009; **125**: 1595-1603 [PMID: 19462451 DOI: 10.1002/ijc.24479]
- 205 **Putz U**, Howitt J, Doan A, Goh CP, Low LH, Silke J, Tan SS. The tumor suppressor PTEN is exported in exosomes and has phosphatase activity in recipient cells. *Sci Signal* 2012; **5**: ra70 [PMID: 23012657]
- 206 **Janowska-Wieczorek A**, Wysoczynski M, Kijowski J, Marquez-Curtis L, Machalinski B, Ratajczak J, Ratajczak MZ. Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *Int J Cancer* 2005; **113**: 752-760 [PMID: 15499615]
- 207 **Jung T**, Castellana D, Klingbeil P, Cuesta Hernández I, Vitacolonna M, Orlicky DJ, Roffler SR, Brodt P, Zöllner M. CD44v6 dependence of premetastatic niche preparation by exosomes. *Neoplasia* 2009; **11**: 1093-1105 [PMID: 19794968]
- 208 **McCreedy J**, Sims JD, Chan D, Jay DG. Secretion of extracellular hsp90 $\alpha$  via exosomes increases cancer cell motility: a role for plasminogen activation. *BMC Cancer* 2010; **10**: 294 [PMID: 20553606 DOI: 10.1186/1471-2407-10-294]
- 209 **Luga V**, Zhang L, Vilorio-Petit AM, Ogunjimi AA, Inanlou MR, Chiu E, Buchanan M, Hosein AN, Basik M, Wrana JL. Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in breast cancer cell migration. *Cell* 2012; **151**: 1542-1556 [PMID: 23260141 DOI: 10.1016/j.cell.2012.11.024]
- 210 **Hong BS**, Cho JH, Kim H, Choi EJ, Rho S, Kim J, Kim JH, Choi DS, Kim YK, Hwang D, Gho YS. Colorectal cancer cell-derived microvesicles are enriched in cell cycle-related mRNAs that promote proliferation of endothelial cells. *BMC Genomics* 2009; **10**: 556 [PMID: 19930720 DOI: 10.1186/1471-2164-10-556]
- 211 **Al-Nedawi K**, Meehan B, Rak J. Microvesicles: messengers and mediators of tumor progression. *Cell Cycle* 2009; **8**: 2014-2018 [PMID: 19535896]
- 212 **Mineo M**, Garfield SH, Taverna S, Flugy A, De Leo G, Alesandro R, Kohn EC. Exosomes released by K562 chronic myeloid leukemia cells promote angiogenesis in a Src-dependent fashion. *Angiogenesis* 2012; **15**: 33-45 [PMID: 22203239 DOI: 10.1007/s10456-011-9241-1]
- 213 **Huan J**, Hornick NI, Shurtleff MJ, Skinner AM, Goloviznina NA, Roberts CT, Kurre P. RNA trafficking by acute myelogenous leukemia exosomes. *Cancer Res* 2013; **73**: 918-929 [PMID: 23149911 DOI: 10.1158/0008-5472.CAN-12-2184]
- 214 **Meckes DG**, Shair KH, Marquitz AR, Kung CP, Edwards RH, Raab-Traub N. Human tumor virus utilizes exosomes for intercellular communication. *Proc Natl Acad Sci USA* 2010; **107**: 20370-20375 [PMID: 21059916 DOI: 10.1073/pnas.1014194107]
- 215 **Gourzoues C**, Gelin A, Bombik I, Klibi J, Vérylaud B, Guigay J, Lang P, Téمام S, Schneider V, Amiel C, Bacconnais S, Jimenez AS, Busson P. Extra-cellular release and blood diffusion of BART viral micro-RNAs produced by EBV-infected nasopharyngeal carcinoma cells. *Virology* 2010; **7**: 271 [PMID: 20950422 DOI: 10.1186/1743-422X-7-271]
- 216 **Roccaro AM**, Sacco A, Maiso P, Azab AK, Tai YT, Reagan M, Azab F, Flores LM, Campigotto F, Weller E, Anderson KC, Scadden DT, Ghobrial IM. BM mesenchymal stromal cell-derived exosomes facilitate multiple myeloma progression. *J Clin Invest* 2013; **123**: 1542-1555 [PMID: 23454749]
- 217 **Yang M**, Chen J, Su F, Yu B, Su F, Lin L, Liu Y, Huang JD, Song E. Microvesicles secreted by macrophages shuttle invasion-potentiating microRNAs into breast cancer cells. *Mol Cancer* 2011; **10**: 117 [PMID: 21939504 DOI: 10.1186/1476-4598-10-117]
- 218 **Zhang Y**, Liu D, Chen X, Li J, Li L, Bian Z, Sun F, Lu J, Yin Y, Cai X, Sun Q, Wang K, Ba Y, Wang Q, Wang D, Yang J, Liu P, Xu T, Yan Q, Zhang J, Zen K, Zhang CY. Secreted monocytic miR-150 enhances targeted endothelial cell migration. *Mol Cell* 2010; **39**: 133-144 [PMID: 20603081 DOI: 10.1016/j.molcel.2010.06.010]
- 219 **Umezue T**, Ohyashiki K, Kuroda M, Ohyashiki JH. Leukemia cell to endothelial cell communication via exosomal miRNAs. *Oncogene* 2013; **32**: 2747-2755 [PMID: 22797057 DOI: 10.1038/onc.2012.295]
- 220 **Fabbri M**, Paone A, Calore F, Galli R, Gaudio E, Santhanam R, Lovat F, Fadda P, Mao C, Nuovo GJ, Zanoni N, Crawford M, Ozer GH, Wernicke D, Alder H, Caligiuri MA, Nana-Sinkam P, Perrotti D, Croce CM. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc Natl Acad Sci USA* 2012; **109**: E2110-E2116 [PMID: 22753494 DOI: 10.1073/pnas.1209414109]
- 221 **Kogure T**, Lin WL, Yan IK, Braconi C, Patel T. Intercellular nanovesicle-mediated microRNA transfer: a mechanism of environmental modulation of hepatocellular cancer cell growth. *Hepatology* 2011; **54**: 1237-1248 [PMID: 21721029 DOI: 10.1002/hep.24504]
- 222 **Pap E**, Pällinger E, Pásztói M, Falus A. Highlights of a new type of intercellular communication: microvesicle-based information transfer. *Inflamm Res* 2009; **58**: 1-8 [PMID: 19132498 DOI: 10.1007/s00011-008-8210-7]
- 223 **Lespagnol A**, Duflaut D, Beekman C, Blanc L, Fiucci G, Marine JC, Vidal M, Amson R, Telerman A. Exosome secretion, including the DNA damage-induced p53-dependent secretory pathway, is severely compromised in TSAP6/Steap3-null mice. *Cell Death Differ* 2008; **15**: 1723-1733 [PMID: 18617898 DOI: 10.1038/cdd.2008]
- 224 **Bianco F**, Perrotta C, Novellino L, Francolini M, Riganti L, Menna E, Saglietti L, Schuchman EH, Furlan R, Clementi E, Matteoli M, Verderio C. Acid sphingomyelinase activity triggers microparticle release from glial cells. *EMBO J* 2009; **28**: 1043-1054 [PMID: 19300439 DOI: 10.1038/emboj.2009.45]
- 225 **Tarabozetti G**, D'Ascenzo S, Borsotti P, Giavazzi R, Pavan A, Dolo V. Shedding of the matrix metalloproteinases MMP-2, MMP-9, and MT1-MMP as membrane vesicle-associated components by endothelial cells. *Am J Pathol* 2002; **160**: 673-680 [PMID: 11839588]
- 226 **Hao S**, Moyana T, Xiang J. Review: cancer immunotherapy by exosome-based vaccines. *Cancer Biother Radiopharm* 2007; **22**: 692-703 [PMID: 17979572]
- 227 **Tan A**, De La Peña H, Seifalian AM. The application of exosomes as a nanoscale cancer vaccine. *Int J Nanomedicine* 2010; **5**: 889-900 [PMID: 21116329 DOI: 10.2147/IJN.S13402]
- 228 **Bobrie A**, Théry C. Unraveling the physiological functions of exosome secretion by tumors. *Oncimmunology* 2013; **2**: e22565 [PMID: 23483742]
- 229 **Morse MA**, Garst J, Osada T, Khan S, Hobeika A, Clay TM, Valente N, Shreenivas R, Sutton MA, Delcayre A, Hsu DH,

- Le Pecq JB, Lyerly HK. A phase I study of dexosome immunotherapy in patients with advanced non-small cell lung cancer. *J Transl Med* 2005; **3**: 9 [PMID: 15723705]
- 230 **Dai S**, Wei D, Wu Z, Zhou X, Wei X, Huang H, Li G. Phase I clinical trial of autologous ascites-derived exosomes combined with GM-CSF for colorectal cancer. *Mol Ther* 2008; **16**: 782-790 [PMID: 18362931 DOI: 10.1038/mt.2008.1]
- 231 **Viaud S**, Théry C, Ploix S, Tursz T, Lapierre V, Lantz O, Zitvogel L, Chaput N. Dendritic cell-derived exosomes for cancer immunotherapy: what's next? *Cancer Res* 2010; **70**: 1281-1285 [PMID: 20145139 DOI: 10.1158/0008-5472.CAN-09-3276]
- 232 **Shen C**, Hao SG, Zhao CX, Zhu J, Wang C. Antileukaemia immunity: effect of exosomes against NB4 acute promyelocytic leukaemia cells. *J Int Med Res* 2011; **39**: 740-747 [PMID: 21819704]
- 233 **Hartman ZC**, Wei J, Glass OK, Guo H, Lei G, Yang XY, Osada T, Hobeika A, Delcayre A, Le Pecq JB, Morse MA, Clay TM, Lyerly HK. Increasing vaccine potency through exosome antigen targeting. *Vaccine* 2011; **29**: 9361-9367 [PMID: 22001882 DOI: 10.1016/j.vaccine.2011.09.133]
- 234 **Rountree RB**, Mandl SJ, Nachtwey JM, Dalpozzo K, Do L, Lombardo JR, Schoonmaker PL, Brinkmann K, Dirmeier U, Laus R, Delcayre A. Exosome targeting of tumor antigens expressed by cancer vaccines can improve antigen immunogenicity and therapeutic efficacy. *Cancer Res* 2011; **71**: 5235-5244 [PMID: 21670078 DOI: 10.1158/0008-5472.CAN-10-4076]
- 235 **Lv LH**, Wan YL, Lin Y, Zhang W, Yang M, Li GL, Lin HM, Shang CZ, Chen YJ, Min J. Anticancer drugs cause release of exosomes with heat shock proteins from human hepatocellular carcinoma cells that elicit effective natural killer cell antitumor responses in vitro. *J Biol Chem* 2012; **287**: 15874-15885 [PMID: 22396543 DOI: 10.1074/jbc.M112.340588]
- 236 **Claas C**, Seiter S, Claas A, Savelyeva L, Schwab M, Zöller M. Association between the rat homologue of CO-029, a metastasis-associated tetraspanin molecule and consumption coagulopathy. *J Cell Biol* 1998; **141**: 267-280 [PMID: 9531564]
- 237 **Gesierich S**, Berezovskiy I, Ryschich E, Zöller M. Systemic induction of the angiogenesis switch by the tetraspanin D6.1A/CO-029. *Cancer Res* 2006; **66**: 7083-7094 [PMID: 16849554]
- 238 **Marleau AM**, Chen CS, Joyce JA, Tullis RH. Exosome removal as a therapeutic adjuvant in cancer. *J Transl Med* 2012; **10**: 134 [PMID: 22738135 DOI: 10.1186/1479-5876-10-134]
- 239 **Safaei R**, Larson BJ, Cheng TC, Gibson MA, Otani S, Naerdemann W, Howell SB. Abnormal lysosomal trafficking and enhanced exosomal export of cisplatin in drug-resistant human ovarian carcinoma cells. *Mol Cancer Ther* 2005; **4**: 1595-1604 [PMID: 16227410]
- 240 **Yin J**, Yan X, Yao X, Zhang Y, Shan Y, Mao N, Yang Y, Pan L. Secretion of annexin A3 from ovarian cancer cells and its association with platinum resistance in ovarian cancer patients. *J Cell Mol Med* 2012; **16**: 337-348 [PMID: 21435174 DOI: 10.1111/j.1582-4934.2011.01316.x]
- 241 **Chen X**, Liang H, Zhang J, Zen K, Zhang CY. microRNAs are ligands of Toll-like receptors. *RNA* 2013; **19**: 737-739 [PMID: 23554231 DOI: 10.1261/rna.036319.112]
- 242 **Lotvall J**, Valadi H. Cell to cell signalling via exosomes through esRNA. *Cell Adh Migr* 2007; **1**: 156-158 [PMID: 19262134]
- 243 **Tan A**, Rajadas J, Seifalian AM. Exosomes as nano-therapeutic delivery platforms for gene therapy. *Adv Drug Deliv Rev* 2013; **65**: 357-367 [PMID: 22820532 DOI: 10.1016/j.addr.2012.06.014]
- 244 **Seow Y**, Wood MJ. Biological gene delivery vehicles: beyond viral vectors. *Mol Ther* 2009; **17**: 767-777 [PMID: 19277019 DOI: 10.1038/mt.2009.41]
- 245 **Alvarez-Erviti L**, Seow Y, Yin H, Betts C, Lakhali S, Wood MJ. Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nat Biotechnol* 2011; **29**: 341-345 [PMID: 21423189 DOI: 10.1038/nbt.1807]
- 246 **Ohno S**, Takanashi M, Sudo K, Ueda S, Ishikawa A, Matsuyama N, Fujita K, Mizutani T, Ohgi T, Ochiya T, Gotoh N, Kuroda M. Systemically injected exosomes targeted to EGFR deliver antitumor microRNA to breast cancer cells. *Mol Ther* 2013; **21**: 185-191 [PMID: 23032975 DOI: 10.1038/mt.2012.180]
- 247 **Sun D**, Zhuang X, Xiang X, Liu Y, Zhang S, Liu C, Barnes S, Grizzle W, Miller D, Zhang HG. A novel nanoparticle drug delivery system: the anti-inflammatory activity of curcumin is enhanced when encapsulated in exosomes. *Mol Ther* 2010; **18**: 1606-1614 [PMID: 20571541 DOI: 10.1038/mt.2010.105]
- 248 **Zhuang X**, Xiang X, Grizzle W, Sun D, Zhang S, Axtell RC, Ju S, Mu J, Zhang L, Steinman L, Miller D, Zhang HG. Treatment of brain inflammatory diseases by delivering exosome encapsulated anti-inflammatory drugs from the nasal region to the brain. *Mol Ther* 2011; **19**: 1769-1779 [PMID: 21915101 DOI: 10.1038/mt.2011.164]
- 249 **Chalmin F**, Ladoire S, Mignot G, Vincent J, Bruchard M, Remy-Martin JP, Boireau W, Rouleau A, Simon B, Lanneau D, De Thonel A, Multhoff G, Hamman A, Martin F, Chauffert B, Solary E, Zitvogel L, Garrido C, Ryffel B, Borg C, Apetoh L, Rébé C, Ghiringhelli F. Membrane-associated Hsp72 from tumor-derived exosomes mediates STAT3-dependent immunosuppressive function of mouse and human myeloid-derived suppressor cells. *J Clin Invest* 2010; **120**: 457-471 [PMID: 20093776 DOI: 10.1172/JCI40483]
- 250 **Maguire CA**, Balaj L, Sivaraman S, Crommentuijn MH, Ericsson M, Mincheva-Nilsson L, Baranov V, Gianni D, Tannous BA, Sena-Estevés M, Breakefield XO, Skog J. Microvesicle-associated AAV vector as a novel gene delivery system. *Mol Ther* 2012; **20**: 960-971 [PMID: 22314290 DOI: 10.1038/mt.2011.303]
- 251 **Kota J**, Chivukula RR, O'Donnell KA, Wentzel EA, Montgomery CL, Hwang HW, Chang TC, Vivekanandan P, Torbenson M, Clark KR, Mendell JR, Mendell JT. Therapeutic microRNA delivery suppresses tumorigenesis in a murine liver cancer model. *Cell* 2009; **137**: 1005-1017 [PMID: 19524505 DOI: 10.1016/j.cell.2009.04.021]
- 252 **Kosaka N**, Takeshita F, Yoshioka Y, Hagiwara K, Katsuda T, Ono M, Ochiya T. Exosomal tumor-suppressive microRNAs as novel cancer therapy: "exocure" is another choice for cancer treatment. *Adv Drug Deliv Rev* 2013; **65**: 376-382 [PMID: 22841506 DOI: 10.1016/j.addr.2012.07.011]

P- Reviewer: Guo CY S- Editor: Wen LL  
L- Editor: A E- Editor: Liu XM



## Prospects and challenges for intestinal microbiome therapy in pediatric gastrointestinal disorders

Richard Kellermayer

Richard Kellermayer, Department of Pediatrics, Section of Pediatric Gastroenterology, Hepatology and Nutrition, Baylor College of Medicine, Texas Children's Hospital, Houston, TX 77030-2399, United States

Author contributions: Richard Kellermayer wrote the manuscript.

Correspondence to: Dr. Richard Kellermayer, MD, PhD, Department of Pediatrics, Section of Pediatric Gastroenterology, Hepatology and Nutrition, Baylor College of Medicine, Texas Children's Hospital, 6621 Fannin St., CC1010.00, Houston, TX 77030-2399, United States. [kellerma@bcm.edu](mailto:kellerma@bcm.edu)

Telephone: +1-713-7980319 Fax: +1-832-8253633

Received: April 30, 2013 Revised: July 23, 2013

Accepted: August 8, 2013

Published online: November 15, 2013

### Abstract

Fecal microbiome (microbiota) transplantation is an emerging treatment not only for refractory/recurrent *Clostridium difficile* infections and chronic gastrointestinal diseases, but also for metabolic syndrome, and even possibly for neurological disorders. This non-conventional therapy has been perhaps more appropriately designated as fecal bacteriotherapy (FB) as well. The employment of FB is spreading into pediatric gastroenterology. This focused review highlights the pediatric applications of FB and discusses hypotheses for its mechanism of action. We propose that intestinal microbiome therapy may be a more appropriate term for FB, which integrates its potential future applications.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Microbiome; Fecal transplant; Children; Inflammatory bowel disease; Ulcerative colitis; *Clostridium difficile*

**Core tip:** This review provides a focused overview of fecal bacteriotherapy and discusses possible mechanisms

of action for this unconventional treatment. It also highlights the challenges, which this therapy faces.

Kellermayer R. Prospects and challenges for intestinal microbiome therapy in pediatric gastrointestinal disorders. *World J Gastrointest Pathophysiol* 2013; 4(4): 91-93 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/91.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.91>

### INTRODUCTION

The alarming increase in recurrent *Clostridium difficile* (*C. difficile*) infections (CDI) and associated deaths<sup>[1]</sup> geared the attention of gastroenterologists around the world towards fecal microbiome (or microbiota) transplantation (FMT)<sup>[2]</sup>. This non-conventional therapeutic approach has also been designated as fecal bacteriotherapy (FB)<sup>[3]</sup>.

Human fecal preparations have been used for centuries in traditional Chinese medicine to treat various disorders<sup>[4]</sup>. However, it was not until 1958 when fecal preparations from healthy donors were employed by bold surgeons as enemas to treat critically ill patients with pseudomembranous colitis (PC)<sup>[5]</sup>. In spite of the surgeons' dramatic success, fecal bacteriotherapy has received less attention up to the 1980's perhaps secondary to the recognition that *C. difficile* is the pathogen for PC and that it can be effectively treated with antibiotics. Indeed, the short term efficacy of current antimicrobials is around 90% against CDI<sup>[6]</sup>. However, the infection may recur in 13%-24% of cases within 4 wk<sup>[6]</sup>. In such instances FB has been utilized with a cure approaching 90% irrespective of the mode of delivery (*i.e.*, upper gastrointestinal, colonoscopic, or large volume retention enema)<sup>[7]</sup>. The first randomized control trial comparing FB with vancomycin therapy for recurrent CDI showed the overwhelming superiority of the fecal preparation<sup>[8]</sup>. In spite of this finding we do not understand clearly

how FB works. It appears that live bacteria are required for FB to be efficacious based on mouse model studies<sup>[9]</sup>. Many researchers argue that it is true engraftment of the donor microbiota that occurs in the recipients through FB, hence is designation as “transplantation”<sup>[10]</sup>. Only limited high-throughput metagenomic studies have addressed this question, especially over a prolonged time course after the treatments. Work with an artificial fecal bacterial preparation of 33 species found that there was a steady decline in the transplanted strains within the stool of the 2 recipients studied<sup>[11]</sup>. More specifically, only about 25%-30% of the species received remained in the recipient community by 6 mo after the “transplant”. This result shows that some donor bacteria truly populate the recipient microbiome at least for several months. However, I propose that FB works by shock therapy or “enslayment” of the recipient microbiota, rather than just engrafting absent bacterial species into the recipient population. More specifically, a brief shock from a healthy donor bacterial community may restructure the recipient microbiota, which acts as a dynamic organ. The short-term to long-term engraftment of a few bacterial species from the donor stool into the restructuring recipient microbiota may aid/participate in this process. I use the example of crystallization induced by a bit of crystal placed into an over-saturated solution, such as in the case of sodium acetate (<http://www.youtube.com/watch?v=HnSg2cl09PI>) to demonstrate my hypothesis. In the case of FB, the stool donor is the bit of crystal and the dysbiotic recipient microbiota is the over-saturated solution. Upon the induction from the healthy donor stool, the recipient microbiota reverts back (“crystallizes”) to a healthier state of the microbial community supporting its ability to overcome CDI. The arguments for the shock therapy are: (1) a single FB enema works as effectively as any other mode of delivery. Enema volumes are about 5%-10% of the colonic volume. Those reach only the hepatic flexure at best, and are evacuated within a few hours after delivery. It is physiologically rather difficult to imagine that such a preparation could truly transplant the whole intestine of the recipient with microbes; and (2) a simple cultured mixture of 10 bacterial species worked as effectively in treating CDI as a retention enema preparation<sup>[12]</sup>. It is unlikely that the treated recipients harbored only the 10 “transplanted” species following the resolution of CDI.

Based on the above, FB appears to be a more appropriate designation of this treatment modality than fecal transplantation. Even more, “intestinal microbiome therapy” (IMT) may be the most proper term for FB, since the future will likely bring the development of restricted microbial communities for the treatment of human diseases. In fact, FB has been used with benefit in inflammatory bowel diseases (IBD)<sup>[13]</sup>, chronic constipation and irritable bowel syndrome<sup>[14]</sup>, metabolic syndrome<sup>[15]</sup>, and even in isolated cases of neurological diseases<sup>[2]</sup>. As for IBD, it was an academic physician, Justin Bennet, who innovatively treated his own ulcerative colitis (UC) with serial large volume retention enemas

of stool preparation from a healthy donor<sup>[16]</sup>. Thereafter, Thomas Borody and colleagues treated 6 UC patients with 5 consecutive daily enemas resulting in over 1 year remission off all medications in all<sup>[17]</sup>. One of these patients has been in remission for over 11 years implicating the potential curative nature of FB for UC. However, in a recent review Borody *et al.*<sup>[18]</sup> states that more than 5 enemas are needed for most patients, but does not define how many. This statement leads to valid concerns raised about FB in the medical community<sup>[19]</sup>. The absence of consensus in regards to volume, route, donor screening, safety measures, and the potential lack of medical supervision has been discussed. Consequently, a fecal microbiota FMT workgroup has formed, and established guidelines for donor screening and recipient selection primarily for CDI<sup>[20]</sup>. To further standardization, “universal” frozen stool preparations to treat CDI were employed with success<sup>[21]</sup>. The establishment and adherence to stringent guidelines and methods should aid the safety and future utilization of this unconventional treatment option for various human diseases.

FB has been less investigated in children, most likely secondary to safety concerns. In the meantime, parents of children suffering from UC, for example, are eager for this treatment option to become available<sup>[22]</sup>. There are only two case reports in children supporting the safety and efficacy of FB for pediatric recurrent CDI<sup>[23,24]</sup>. Additionally, a very recent publication demonstrated that serial retention enemas from healthy donors can be of benefit for pediatric and young-adult patients with mild to moderately active UC<sup>[25]</sup>. The study participants experienced only mild to moderate, self-resolving side effects from FB. These publications clearly indicate the potential utility of FB in pediatric gastrointestinal disorders as well.

At present, FB appears to hold great prospects, but also significant challenges for the treatment of human diseases. For chronic disorders, such as IBD, the end point of therapy may be difficult to define. Long-term potential side effects such as modified metabolism, changes in mood and affect, altered susceptibility to malignancies, *etc.* have not been examined. The most optimal route of delivery may vary between diseases, and between differing phenotypes of a single disease. The need for donor-recipient matching is also of question. The importance of age, gender, race, and microbiome composition (among others) are also unknown in this respect. The potential significance of fungi and viruses during IMT has not even been addressed to date. Perhaps the greatest challenge for the future will be to define restricted microbial communities for specific diseases. Only dedicated academic scientists will be able to meet these challenges and optimize metagenomic medicine for current and future generations to come.

## REFERENCES

- 1 Peery AF, Dellon ES, Lund J, Crockett SD, McGowan CE, Bulsiewicz WJ, Gangarosa LM, Thiny MT, Stizenberg K,

- Morgan DR, Ringel Y, Kim HP, Dibonaventura MD, Carroll CF, Allen JK, Cook SF, Sandler RS, Kappelman MD, Shaheen NJ. Burden of gastrointestinal disease in the United States: 2012 update. *Gastroenterology* 2012; **143**: 1179-1187. e1-e3 [PMID: 22885331 DOI: 10.1053/j.gastro.2012.08.002]
- 2 **Brandt LJ**. American Journal of Gastroenterology Lecture: Intestinal microbiota and the role of fecal microbiota transplant (FMT) in treatment of *C. difficile* infection. *Am J Gastroenterol* 2013; **108**: 177-185 [PMID: 23318479 DOI: 10.1038/ajg.2012.450]
  - 3 **Suwantarat N**, Bobak DA. Fecal Bacteriotherapy for Recurrent *Clostridium difficile* Infection: What's Old Is New Again? *Curr Infect Dis Rep* 2013; **15**: 101-103 [PMID: 23549617 DOI: 10.1007/s11908-013-0314-8]
  - 4 **Zhang F**, Luo W, Shi Y, Fan Z, Ji G. Should we standardize the 1,700-year-old fecal microbiota transplantation? *Am J Gastroenterol* 2012; **107**: 1755; author reply 1755-1756 [PMID: 23160295 DOI: 10.1038/ajg.2012.251]
  - 5 **EISEMAN B**, SILEN W, BASCOM GS, KAUVAR AJ. Fecal enema as an adjunct in the treatment of pseudomembranous enterocolitis. *Surgery* 1958; **44**: 854-859 [PMID: 13592638]
  - 6 **Louie TJ**, Miller MA, Mullane KM, Weiss K, Lentnek A, Golan Y, Gorbach S, Sears P, Shue YK. Fidaxomicin versus vancomycin for *Clostridium difficile* infection. *N Engl J Med* 2011; **364**: 422-431 [PMID: 21288078 DOI: 10.1056/NEJMoa0910812]
  - 7 **Sofi AA**, Silverman AL, Khuder S, Garborg K, Westerink JM, Nawras A. Relationship of symptom duration and fecal bacteriotherapy in *Clostridium difficile* infection-pooled data analysis and a systematic review. *Scand J Gastroenterol* 2013; **48**: 266-273 [PMID: 23163886 DOI: 10.3109/00365521.2012.743585]
  - 8 **van Nood E**, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, Visser CE, Kuijper EJ, Bartelsman JF, Tijssen JG, Speelman P, Dijkgraaf MG, Keller JJ. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med* 2013; **368**: 407-415 [PMID: 23323867 DOI: 10.1056/NEJMoa1205037]
  - 9 **Lawley TD**, Clare S, Walker AW, Stares MD, Connor TR, Raisen C, Goulding D, Rad R, Schreiber F, Brandt C, Deakin LJ, Pickard DJ, Duncan SH, Flint HJ, Clark TG, Parkhill J, Dougan G. Targeted restoration of the intestinal microbiota with a simple, defined bacteriotherapy resolves relapsing *Clostridium difficile* disease in mice. *PLoS Pathog* 2012; **8**: e1002995 [PMID: 23133377 DOI: 10.1371/journal.ppat.1002995]
  - 10 **Hamilton MJ**, Weingarden AR, Unno T, Khoruts A, Sadowsky MJ. High-throughput DNA sequence analysis reveals stable engraftment of gut microbiota following transplantation of previously frozen fecal bacteria. *Gut Microbes* 2013; **4**: 125-135 [PMID: 23333862 DOI: 10.4161/gmic.23571]
  - 11 **Petrof EO**, Gloor GB, Vanner SJ, Weese SJ, Carter D, Daigneault MC, Brown EM, Schroeter K, Allen-Vercoe E. Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: 'RePOOPulating' the gut. *Microbiome* 2013; **1**: 3 [DOI: 10.1186/2049-2618-1-3]
  - 12 **Tvede M**, Rask-Madsen J. Bacteriotherapy for chronic relapsing *Clostridium difficile* diarrhoea in six patients. *Lancet* 1989; **1**: 1156-1160 [PMID: 2566734]
  - 13 **Anderson JL**, Edney RJ, Whelan K. Systematic review: faecal microbiota transplantation in the management of inflammatory bowel disease. *Aliment Pharmacol Ther* 2012; **36**: 503-516 [PMID: 22827693 DOI: 10.1111/j.1365-2036.2012.05220]
  - 14 **Andrews PJ**, Borody TJ. "Putting back the bugs": bacterial treatment relieves chronic constipation and symptoms of irritable bowel syndrome. *Med J Aust* 1993; **159**: 633-634 [PMID: 8155121]
  - 15 **Vrieze A**, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JF, Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druesne A, Van Hylckama Vlieg JE, Bloks VW, Groen AK, Heilig HG, Zoetendal EG, Stros ES, de Vos WM, Hoekstra JB, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* 2012; **143**: 913-916. e7 [PMID: 22728514 DOI: 10.1053/j.gastro.2012.06.031]
  - 16 **Bennet JD**, Brinkman M. Treatment of ulcerative colitis by implantation of normal colonic flora. *Lancet* 1989; **1**: 164 [PMID: 2563083]
  - 17 **Borody TJ**, Warren EF, Leis S, Surace R, Ashman O. Treatment of ulcerative colitis using fecal bacteriotherapy. *J Clin Gastroenterol* 2003; **37**: 42-47 [PMID: 12811208]
  - 18 **Borody TJ**, Campbell J. Fecal microbiota transplantation: current status and future directions. *Expert Rev Gastroenterol Hepatol* 2011; **5**: 653-655 [PMID: 22017691 DOI: 10.1586/egh.11.71]
  - 19 **El-Matary W**, Simpson R, Ricketts-Burns N. Fecal microbiota transplantation: are we opening a can of worms? *Gastroenterology* 2012; **143**: e19; author reply e19-e20 [PMID: 22732575 DOI: 10.1053/j.gastro.2012.04.055]
  - 20 **Bakken JS**, Borody T, Brandt LJ, Brill JV, Demarco DC, Franzos MA, Kelly C, Khoruts A, Louie T, Martinelli LP, Moore TA, Russell G, Surawicz C. Treating *Clostridium difficile* infection with fecal microbiota transplantation. *Clin Gastroenterol Hepatol* 2011; **9**: 1044-1049 [PMID: 21871249 DOI: 10.1016/j.cgh.2011.08.014]
  - 21 **Hamilton MJ**, Weingarden AR, Sadowsky MJ, Khoruts A. Standardized frozen preparation for transplantation of fecal microbiota for recurrent *Clostridium difficile* infection. *Am J Gastroenterol* 2012; **107**: 761-767 [PMID: 22290405 DOI: 10.1038/ajg.2011.482]
  - 22 **Kahn SA**, Gorawara-Bhat R, Rubin DT. Fecal bacteriotherapy for ulcerative colitis: patients are ready, are we? *Inflamm Bowel Dis* 2012; **18**: 676-684 [PMID: 21618362 DOI: 10.1002/ibd.21775]
  - 23 **Russell G**, Kaplan J, Ferraro M, Michelow IC. Fecal bacteriotherapy for relapsing *Clostridium difficile* infection in a child: a proposed treatment protocol. *Pediatrics* 2010; **126**: e239-e242 [PMID: 20547640 DOI: 10.1542/peds.2009-3363]
  - 24 **Kahn SA**, Young S, Rubin DT. Colonoscopic fecal microbiota transplant for recurrent *Clostridium difficile* infection in a child. *Am J Gastroenterol* 2012; **107**: 1930-1931 [PMID: 23211865 DOI: 10.1038/ajg.2012.351]
  - 25 **Kunde S**, Pham A, Bonczyk S, Crumb T, Duba M, Conrad H, Cloney D, Kugathasan S. Safety, tolerability, and clinical response after fecal transplantation in children and young adults with ulcerative colitis. *J Pediatr Gastroenterol Nutr* 2013; **56**: 597-601 [PMID: 23542823 DOI: 10.1097/MPG.0b013e318292fa0d]

P- Reviewer: Zhang FM S- Editor: Qi Y

L- Editor: A E- Editor: Liu XM



## Intestinal barrier: Molecular pathways and modifiers

Min Kyung Jeon, Christina Klaus, Elke Kaemmerer, Nikolaus Gassler

Min Kyung Jeon, Christina Klaus, Elke Kaemmerer, Nikolaus Gassler, Institute of Pathology, RWTH Aachen University, 52074 Aachen, Germany

Author contributions: Jeon MK and Gassler N designed and wrote the article; Klaus C and Kaemmerer E critical reviewed the literature and made technical notes.

Correspondence to: Nikolaus Gassler, MA, Professor, Institute of Pathology, RWTH Aachen University, Pauwelsstraße 30, 52074 Aachen, Germany. [ngassler@ukaachen.de](mailto:ngassler@ukaachen.de)

Telephone: +49-241-8088897 Fax: +49-241-8082439

Received: June 24, 2013 Revised: August 28, 2013

Accepted: September 3, 2013

Published online: November 15, 2013

### Abstract

The gastrointestinal tract is frequently challenged by pathogens/antigens contained in food and water and the intestinal epithelium must be capable of rapid regeneration in the event of tissue damage. Disruption of the intestinal barrier leads to a number of immune-mediated diseases, including inflammatory bowel disease, food allergy, and celiac disease. The intestinal mucosa is composed of different types of epithelial cells in specific barrier functions. Epithelial cells control surface-associated bacterial populations without disrupting the intestinal microflora that is crucial for host health. They are also capable of modulating mucosal immune system, and are thus essential in maintaining homeostasis in the gut. Thus, the regulation of intestinal epithelial homeostasis is crucial for the maintenance of the structure of the mucosa and the defensive barrier functions. Recent studies have demonstrated that multiple molecular pathways are involved in the regulation of intestinal epithelial cell polarity. These include the Wnt, Notch, Hippo, transforming growth factor- $\beta$  (TGF- $\beta$ )/bone morphogenetic protein (BMP) and Hedgehog pathways, most of which were identified in lower organisms where they play important roles during embryogenesis. These pathways are also used in adult organisms to regulate multiple self-renewing organs. Understanding the interactions between these

molecular mechanisms and intestinal barrier function will therefore provide important insight into the pathogenesis of intestinal-based immune-mediated diseases.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

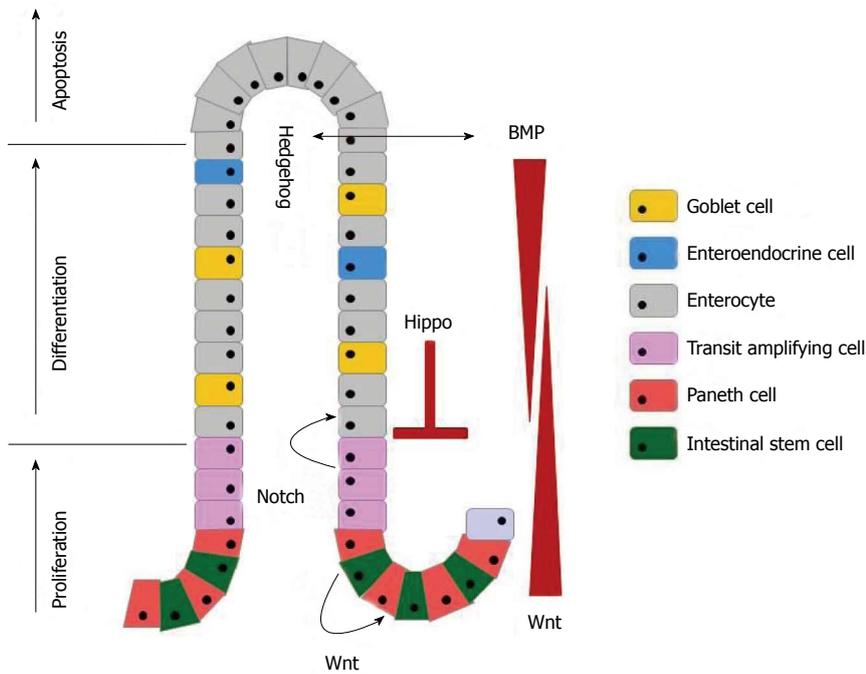
**Key words:** Intestinal epithelium; Mucosal barrier; Homeostasis; Molecular pathways; Immune-mediated disease

**Core tip:** The pathogenesis of gastrointestinal diseases is associated with important molecular pathways such as Wnt, Notch, Hippo, transforming growth factor- $\beta$ /bone morphogenetic protein or Hedgehog in controlling cell-fate determination. Here, we discuss how they contribute to homeostasis of intestinal epithelium.

Jeon MK, Klaus C, Kaemmerer E, Gassler N. Intestinal barrier: Molecular pathways and modifiers. *World J Gastrointest Pathophysiol* 2013; 4(4): 94-99 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/94.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.94>

### INTRODUCTION

The intestinal epithelium is a single-cell layer that serves as a protective barrier against the external environment. It supports nutrients and water transport, while maintaining a defense against intraluminal toxins, bacteria, and antigens. The epithelial barrier is governed by the expression of adherens junctions (AJs) and tight junctions (TJs), including cadherins, claudins, occludin, and junctional adhesion molecules (JAM) proteins, which seal adjacent cells together<sup>[1]</sup>. Expression of AJ and TJ proteins is regulated by phosphorylation and it can either promote or destabilize TJ formation<sup>[2]</sup>. The AJ and TJ complexes also play a crucial role in the regulation of cellular polarization, proliferation, and differentiation<sup>[3]</sup>.



**Figure 1** The molecular regulatory pathways in intestinal epithelial homeostasis. Wnt signaling promotes the proliferation of TA/stem cells and drives Paneth cell maturation. Notch signaling cooperates with Wnt to drive proliferation of intestinal stem cells and regulates maintenance of undifferentiated TA/stem cells. Hippo and bone morphogenetic protein (BMP) signaling inhibits proliferation and induces maturation of all secretory cell types. Hedgehog signaling activates the expression of BMP in the mesenchyme.

The intestinal epithelium is populated by distinct types of cells derived from stem cells, such as the absorptive cells (enterocytes) and the secretory cells (mucus-secreting goblet cells, hormone-secreting enteroendocrine cells, Tuft cells, and antimicrobial peptides-secreting Paneth cells). All but the Paneth cells differentiate into mature forms from the crypts to replace cells extruded from the tips of the villi<sup>[4,5]</sup>. This continuous replenishment of intestinal epithelium takes generally 4-7 d and it is important for the maintenance of epithelial integrity. Several conserved signaling cascades, *i.e.*, the Wnt, transforming growth factor- $\beta$  (TGF- $\beta$ )/bone morphogenetic protein (BMP), Notch, Hippo, and Hedgehog pathway are associated with the maintenance of the morphological and functional features of diverse epithelial cell types (Figure 1).

Disruption of this delicate balance causes a variety of intestinal-related inflammatory and autoimmune syndromes<sup>[6-8]</sup>. For example, healthy first-degree relatives of patients with inflammatory bowel disease (IBD) and celiac disease have increased intestinal permeability and epithelial apoptosis<sup>[9,10]</sup>. Intestinal barrier dysfunction also contributes to the severity of food allergen-induced clinical symptoms<sup>[11]</sup>. In this review, we summarize current studies for major molecular signaling pathways in intestinal homeostasis.

## SIGNALING PATHWAYS REGULATING INTESTINAL HOMEOSTASIS

### WNT PATHWAY

Wnt signaling plays multiple roles during intestinal ho-

meostasis. It contains the canonical and non-canonical pathway. The central player in the canonical Wnt signaling cascade is  $\beta$ -catenin, a cytoplasmic protein the stability of which is regulated by the APC tumor suppressor. Complexes of frizzled seven transmembrane molecules and low-density lipoprotein receptor related proteins (LRP-5/-6) serve as receptors for secreted Wnts<sup>[12]</sup>. When Wnt receptor complexes are not engaged, casein kinase-1 and GSK3- $\beta$ , both residing in the APC complex, sequentially phosphorylate  $\beta$ -catenin at a series of highly conserved Ser/Thr residues near its N-terminus and are thereby tagged for ubiquitination and proteasomal degradation. In contrast, Wnt stimulation blocks the intrinsic kinase activity of the APC complex, leading to  $\beta$ -catenin accumulation. Unphosphorylated  $\beta$ -catenin translocates into the nucleus where it engages transcription factors of the TCF/LEF family, thereby translating the Wnt signal into the transcription of a TCF target gene<sup>[13]</sup>.

The canonical Wnt pathway is essential for epithelial proliferation and crypt maintenance<sup>[14,15]</sup>. Mutations in the *APC* gene, a negative regulator of Wnt signaling, also results in hyperproliferation of the epithelium followed by the development of adenomas<sup>[16]</sup>. Moreover,  $\beta$ -catenin plays a role as an AJ component, thereby linking it to the cytoskeleton of epithelial cells. Wnt signaling is necessary for positioning and maturation of Paneth cells in the crypts, and for separating proliferating and differentiated cells. These processes are controlled by the Wnt-dependent expression of specific ephrin receptors in the intestine<sup>[17]</sup>. Non-canonical ( $\beta$ -catenin

independent) Wnt signaling is termed the planar cell polarity pathway, which is activated by the GTPases Rho and Rac. These induce cytoskeletal rearrangements and help to form new crypts<sup>[18]</sup>.

## NOTCH PATHWAY

Notch signaling is active in intestinal crypt compartments and assists the Wnt pathway in promoting stem cell proliferation and negatively regulates differentiation into the secretory lineages<sup>[19]</sup>. Interaction of the Notch receptor (NOTCH 1-4) and ligand (Delta-like 1, 3, and 4; Jagged 1 and 2) between two adjacent cells results in proteolytic cleavages of the receptor by the  $\gamma$ -secretase enzyme complex, leading to the translocation of the Notch intracellular domain (NICD) into the nucleus. There, NICD binds to the transcription factor CSL (CBF1 in human, RBPjk in mice, Su (H) in *Drosophila*, Lag-1 in *C. elegans*) and activates target genes such as *HES-1* (Hairy and enhancer of split 1), *HES-3*, and *HES-5*, which are important for the differentiation of absorptive cells. The transcription factor MATH-1 is a downstream target of HES-1 repression in the intestine and its activity leads to generation of the secretory cell lineages<sup>[20,21]</sup>. These results suggest that the absorptive versus secretory epithelial cell type fate decision is established through the HES/MATH1 axis. In addition, Gfi-1 and neurogenin-3 (Ngn-3), other transcription factors, compete for selection of enteroendocrine versus goblet or Paneth cell fates<sup>[22]</sup>.

The dysregulation of Notch activity is related to the pathogenesis of IBDs, such as ulcerative colitis (UC) and Crohn's disease (CD). A histological study in UC revealed that the depletion of goblet cells with loss of ATOH1 expression and CD is caused by dysregulation of secretory cell differentiation<sup>[23-25]</sup>. Additional evidence supporting such an important role for Notch in the intestine is derived from studies of  $\gamma$ -secretase inhibitors (GSIs). Treatment of mice with GSIs induced colitis due to inhibition of Notch signaling<sup>[26]</sup>. Notch activity may thus contribute to the regenerating epithelium and enhance the barrier function of the intestinal epithelium.

## HIPPO PATHWAY

The Hippo pathway plays a crucial role in controlling organ size by inhibiting cell proliferation and apoptosis in response to cell-cell contact. This tumor suppressor pathway regulates intestinal regeneration and tumorigenesis<sup>[27,28]</sup>. When the Hippo pathway is active, the downstream effector of this pathway Yes-associated protein (YAP) is phosphorylated at S127 by the LAT1/2 kinases. Phosphorylated YAP remains in the cytoplasm and inhibits its proliferative and anti-apoptotic function in the nucleus, which is mediated by its binding to TEAD1-4 transcription factors. Cytoplasmic YAP has the Wnt antagonizing effects, thereby contributing to the prevention of proliferation and intestinal stem cell expansion<sup>[29]</sup>.

In contrast, the deletion of Hippo pathway component Mst1/2 in mouse intestinal epithelial cells results in an expansion of undifferentiated stem cells and an absence of all secretory lineages<sup>[30]</sup>.

In the small intestinal and the colonic epithelium of the normal mouse, YAP protein is found in the crypts and under normal conditions YAP makes no contribution to intestinal epithelial proliferation. However, it is required for tissue regeneration caused by injury. YAP protein is overexpressed in the crypts of the recovery phase from the DSS-treated mice<sup>[31]</sup>. Treatment of GSIs suppressed the intestinal dysplasia caused by YAP and this result suggests that YAP stimulates Notch signaling. YAP is commonly overexpressed in colorectal cancers<sup>[32]</sup>. Therefore, the activation of YAP for regulating intestinal stem cells regeneration indicates that a deficiency of Hippo signaling may contribute to tumorigenesis in the intestine.

## HEDGEHOG PATHWAY

The Hedgehog signaling is initiated through the binding of Hedgehog ligands to the patched homolog 1 (Ptch1) receptor. In the absence of ligands, Ptch1 inhibits the activity of smoothed (Smo). Hedgehog binding inactivates Ptch1 and in contrast, Smo inhibition is released and these mechanisms lead to the translocation of members of the GLI family (GLI-1, GLI-2, and GLI-3 in mammals) of Zn-finger transcription factors from the cytoplasm to the nucleus. Upon nuclear translocation, they activate the transcription of target genes, such as Ptch1, Gli-1, Bmp4, Hhip1<sup>[33]</sup>. Three Hedgehog homologs, such as sonic hedgehog (Shh), indian hedgehog (Ihh), and desert hedgehog (Dhh) are known to be highly conserved in mammals and of these, only Shh and Ihh are expressed in epithelium during development, but are redistributed after villus formation to be localized to the intervillus region<sup>[34]</sup>. Thus, Hedgehog signaling is required for the formation of villi. Shh has been localized to the region of the crypts and Shh plays a role in the regulation of epithelial proliferation<sup>[35]</sup>. Furthermore, Hedgehog ligands have been reported to be anti-inflammatory epithelial modulators in the intestine<sup>[36]</sup>. Hedgehog signaling pathway inhibitors induced hypoplasia of Paneth cells, thus this signaling pathway may influence intestinal epithelium repair partly through the regulation of Paneth cells<sup>[37]</sup>. Hedgehog signaling decreases during the injury phase and increases during the repair phase<sup>[38]</sup>. It confirms an important role of Hedgehog signaling in the repair of intestinal epithelium after injury.

## TGF- $\beta$ /BMP PATHWAY

TGF- $\beta$  signaling regulates embryonic development, wound healing, proliferation, and cell differentiation<sup>[39,40]</sup>. The TGF- $\beta$  family consists of cytokines including TGF- $\beta$  isoform, BMPs, and activins. Signaling is induced by ligand binding to type II serine/threonine kinase receptors, which results in the phosphorylation of

the type I receptor. Then, signaling of these activated receptors is transduced through three classes of SMAD proteins: receptor-regulated SMADs (R-SMADs: SMAD-1, -2, -3, -5, and -8), common SMAD (SMAD-4), and inhibitory SMADs (I-SMADs: SMAD-6 and -7). R-SMADs become phosphorylated by activated type I receptors and subsequent translocate to the nucleus. The SMAD complex interacts with transcription factors, thereby inducing target gene expression. TGF- $\beta$  signaling components are expressed in the differentiated compartment of the intestine<sup>[41]</sup>. Inactivation of TGF- $\beta$  signaling components has also been identified at the adenoma-to-carcinoma transition<sup>[42]</sup>.

BMP signaling mediates the action of hedgehog, blocking the formation of ectopic crypts, and the expression of BMP antagonist noggin in the crypts prevents the activity of BMP, thereby enabling proliferation to continue<sup>[43]</sup>. TGF- $\beta$  ligands signal through SMAD-2 and -3, whereas BMP signaling is mediated through SMAD-1, -5 and -8. Phosphorylated SMAD-1, -5, and -8 are observed in the villus epithelium and this modification prevents the villus epithelium from adopting a crypt-like proliferative character<sup>[44]</sup>. BMP signaling antagonizes the Wnt pathway within the differentiated compartment, thereby positioning transient amplifying cells in the crypts<sup>[45]</sup>. In addition, humans with germline mutations in SMAD-4 or BMP receptor type 1A (BMPRI1A) are associated with up to 50% of JPS (Juvenile polyposis syndrome) cases<sup>[46,47]</sup>.

## CONCLUSION

Intestinal epithelial barrier dysfunction has been implicated as a critical role in the predisposition to a number of gastrointestinal diseases such as IBD, food allergy, and celiac disease. It could lead to defects in multiple aspects of microbial, epithelial, and immune interactions, and thus injury to the epithelial lining thought to be an important factor in the pathogenesis of intestinal-based immune-mediated diseases<sup>[48-50]</sup>.

The homeostasis of the intestinal epithelium is maintained by a complex interplay of multiple regulatory mechanisms. Together, the Wnt, Hedgehog and TGF- $\beta$ /BMP pathways maintain the crypt-villus architecture. The Wnt, Notch, and Hippo signaling pathways combine to control the cell fate choices from stem cells. Thus and understanding of critical signaling pathways for maintaining intestinal homeostasis is essential.

## ACKNOWLEDGMENTS

We would like to thank P Akens for her help in typing and proofreading the manuscript.

## REFERENCES

1 **Laukoetter MG**, Bruewer M, Nusrat A. Regulation of the intestinal epithelial barrier by the apical junctional complex.

*Curr Opin Gastroenterol* 2006; **22**: 85-89 [PMID: 16462161 DOI: 10.1097/01.mog.0000203864.48255.4f]

- 2 **Atkinson KJ**, Rao RK. Role of protein tyrosine phosphorylation in acetaldehyde-induced disruption of epithelial tight junctions. *Am J Physiol Gastrointest Liver Physiol* 2001; **280**: G1280-G1288 [PMID: 11352822]
- 3 **Hartsock A**, Nelson WJ. Adherens and tight junctions: structure, function and connections to the actin cytoskeleton. *Biochim Biophys Acta* 2008; **1778**: 660-669 [PMID: 17854762 DOI: 10.1016/j.bbame.2007.07.012]
- 4 **Porter EM**, Bevins CL, Ghosh D, Ganz T. The multifaceted Paneth cell. *Cell Mol Life Sci* 2002; **59**: 156-170 [PMID: 11846026]
- 5 **Sancho E**, Batlle E, Clevers H. Signaling pathways in intestinal development and cancer. *Annu Rev Cell Dev Biol* 2004; **20**: 695-723 [PMID: 15473857 DOI: 10.1146/annurev.cellbio.20.010403.092805]
- 6 **Brandt EB**, Strait RT, Hershko D, Wang Q, Muntel EE, Scribner TA, Zimmermann N, Finkelman FD, Rothenberg ME. Mast cells are required for experimental oral allergen-induced diarrhea. *J Clin Invest* 2003; **112**: 1666-1677 [PMID: 14660743 DOI: 10.1172/JCI19785]
- 7 **Schulzke JD**, Bentzel CJ, Schulzke I, Riecken EO, Fromm M. Epithelial tight junction structure in the jejunum of children with acute and treated celiac sprue. *Pediatr Res* 1998; **43**: 435-441 [PMID: 9544995 DOI: 10.1203/00006450-199804000-00001]
- 8 **D'Incà R**, Di Leo V, Corrao G, Martines D, D'Odorico A, Mestriner C, Venturi C, Longo G, Sturniolo GC. Intestinal permeability test as a predictor of clinical course in Crohn's disease. *Am J Gastroenterol* 1999; **94**: 2956-2960 [PMID: 10520851 DOI: 10.1111/j.1572-0241.1999.01444.x]
- 9 **Peeters M**, Geypens B, Claus D, Nevens H, Ghooys Y, Verbeke G, Baert F, Vermeire S, Vlietinck R, Rutgeerts P. Clustering of increased small intestinal permeability in families with Crohn's disease. *Gastroenterology* 1997; **113**: 802-807 [PMID: 9287971]
- 10 **Gitter AH**, Bendfeldt K, Schulzke JD, Fromm M. Leaks in the epithelial barrier caused by spontaneous and TNF- $\alpha$ -induced single-cell apoptosis. *FASEB J* 2000; **14**: 1749-1753 [PMID: 10973924 DOI: 10.1096/fj.99-0898com]
- 11 **Ventura MT**, Polimeno L, Amoroso AC, Gatti F, Annoscia E, Marinaro M, Di Leo E, Matino MG, Buquicchio R, Bonini S, Turisi A, Francavilla A. Intestinal permeability in patients with adverse reactions to food. *Dig Liver Dis* 2006; **38**: 732-736 [PMID: 16880015 DOI: 10.1016/j.dld.2006.06.012]
- 12 **Giles RH**, van Es JH, Clevers H. Caught up in a Wnt storm: Wnt signaling in cancer. *Biochim Biophys Acta* 2003; **1653**: 1-24 [PMID: 12781368]
- 13 **Aberle H**, Bauer A, Stappert J, Kispert A, Kemler R.  $\beta$ -catenin is a target for the ubiquitin-proteasome pathway. *EMBO J* 1997; **16**: 3797-3804 [PMID: 9233789 DOI: 10.1093/emboj/16.13.3797]
- 14 **Korinek V**, Barker N, Moerer P, van Donselaar E, Huls G, Peters PJ, Clevers H. Depletion of epithelial stem-cell compartments in the small intestine of mice lacking Tcf-4. *Nat Genet* 1998; **19**: 379-383 [PMID: 9697701 DOI: 10.1038/1270]
- 15 **Pinto D**, Gregorieff A, Begthel H, Clevers H. Canonical Wnt signals are essential for homeostasis of the intestinal epithelium. *Genes Dev* 2003; **17**: 1709-1713 [PMID: 12865297 DOI: doi: ]
- 16 **Oshima M**, Oshima H, Kitagawa K, Kobayashi M, Itakura C, Taketo M. Loss of Apc heterozygosity and abnormal tissue building in nascent intestinal polyps in mice carrying a truncated Apc gene. *Proc Natl Acad Sci USA* 1995; **92**: 4482-4486 [PMID: 7753829]
- 17 **Stappenbeck TS**, Mills JC, Gordon JI. Molecular features of adult mouse small intestinal epithelial progenitors. *Proc Natl Acad Sci USA* 2003; **100**: 1004-1009 [PMID: 12552106 DOI: 10.1073/pnas.242735899]

- 18 **Nusse R.** Wnt signaling. *Cold Spring Harb Perspect Biol* 2012; **4**: 1749-1753 [PMID: 22550232 DOI: 10.1101/cshperspect.a011163]
- 19 **VanDussen KL,** Carulli AJ, Keeley TM, Patel SR, Puthoff BJ, Magness ST, Tran IT, Maillard I, Siebel C, Kolterud Å, Grosse AS, Gumucio DL, Ernst SA, Tsai YH, Dempsey PJ, Samuelson LC. Notch signaling modulates proliferation and differentiation of intestinal crypt base columnar stem cells. *Development* 2012; **139**: 488-497 [PMID: 22190634 DOI: 10.1242/dev.070763]
- 20 **De Strooper B,** Annaert W, Cupers P, Saftig P, Craessaerts K, Mumm JS, Schroeter EH, Schrijvers V, Wolfe MS, Ray WJ, Goate A, Kopan R. A presenilin-1-dependent gamma-secretase-like protease mediates release of Notch intracellular domain. *Nature* 1999; **398**: 518-522 [PMID: 10206645 DOI: 10.1038/19083]
- 21 **Ueo T,** Imayoshi I, Kobayashi T, Ohtsuka T, Seno H, Nakase H, Chiba T, Kageyama R. The role of Hes genes in intestinal development, homeostasis and tumor formation. *Development* 2012; **139**: 1071-1082 [PMID: 22318232 DOI: 10.1242/dev.069070]
- 22 **Shroyer NF,** Wallis D, Venken KJ, Bellen HJ, Zoghbi HY. Gfi1 functions downstream of Math1 to control intestinal secretory cell subtype allocation and differentiation. *Genes Dev* 2005; **19**: 2412-2417 [PMID: 16230531 DOI: 10.1101/gad.1353905]
- 23 **Zheng X,** Tsuchiya K, Okamoto R, Iwasaki M, Kano Y, Sakamoto N, Nakamura T, Watanabe M. Suppression of hath1 gene expression directly regulated by hes1 via notch signaling is associated with goblet cell depletion in ulcerative colitis. *Inflamm Bowel Dis* 2011; **17**: 2251-2260 [PMID: 21987298 DOI: 10.1002/ibd.21611]
- 24 **Gersemann M,** Becker S, Kübler I, Koslowski M, Wang G, Herrlinger KR, Griger J, Fritz P, Fellermann K, Schwab M, Wehkamp J, Stange EF. Differences in goblet cell differentiation between Crohn's disease and ulcerative colitis. *Differentiation* 2009; **77**: 84-94 [PMID: 19281767 DOI: 10.1016/j.diff.2008.09.008]
- 25 **Wehkamp J,** Salzman NH, Porter E, Nuding S, Weichen-thal M, Petras RE, Shen B, Schaeffeler E, Schwab M, Linz-meier R, Feathers RW, Chu H, Lima H, Fellermann K, Ganz T, Stange EF, Bevins CL. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci USA* 2005; **102**: 18129-18134 [PMID: 16330776 DOI: 10.1073/pnas.0505256102]
- 26 **Okamoto R,** Tsuchiya K, Nemoto Y, Akiyama J, Nakamura T, Kanai T, Watanabe M. Requirement of Notch activation during regeneration of the intestinal epithelia. *Am J Physiol Gastrointest Liver Physiol* 2009; **296**: G23-G35 [PMID: 19023031 DOI: 10.1152/ajpgi.90225.2008]
- 27 **Karpowicz P,** Perez J, Perrimon N. The Hippo tumor suppressor pathway regulates intestinal stem cell regeneration. *Development* 2010; **137**: 4135-4145 [PMID: 21098564 DOI: 10.1242/dev.060483]
- 28 **Cai J,** Zhang N, Zheng Y, de Wilde RF, Maitra A, Pan D. The Hippo signaling pathway restricts the oncogenic potential of an intestinal regeneration program. *Genes Dev* 2010; **24**: 2383-2388 [PMID: 21041407 DOI: 10.1101/gad.1978810]
- 29 **Chen L,** Qin F, Deng X, Avruch J, Zhou D. Hippo pathway in intestinal homeostasis and tumorigenesis. *Protein Cell* 2012; **3**: 305-310 [PMID: 22492181 DOI: 10.1007/s13238-012-2913-9]
- 30 **Zhou D,** Zhang Y, Wu H, Barry E, Yin Y, Lawrence E, Dawson D, Willis JE, Markowitz SD, Camargo FD, Avruch J. Mst1 and Mst2 protein kinases restrain intestinal stem cell proliferation and colonic tumorigenesis by inhibition of Yes-associated protein (Yap) overabundance. *Proc Natl Acad Sci USA* 2011; **108**: E1312-E1320 [PMID: 22042863 DOI: 10.1073/pnas.1110428108]
- 31 **Ren F,** Wang B, Yue T, Yun EY, Ip YT, Jiang J. Hippo signaling regulates Drosophila intestine stem cell proliferation through multiple pathways. *Proc Natl Acad Sci USA* 2010; **107**: 21064-21069 [PMID: 21078993 DOI: 10.1073/pnas.1012759107]
- 32 **Steinhardt AA,** Gayyed MF, Klein AP, Dong J, Maitra A, Pan D, Montgomery EA, Anders RA. Expression of Yes-associated protein in common solid tumors. *Hum Pathol* 2008; **39**: 1582-1589 [PMID: 18703216 DOI: 10.1016/j.humpath.2008.04.012]
- 33 **Ingham PW,** McMahon AP. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 2001; **15**: 3059-3087 [PMID: 11731473 DOI: 10.1101/gad.938601]
- 34 **Watt FM.** Unexpected Hedgehog-Wnt interactions in epithelial differentiation. *Trends Mol Med* 2004; **10**: 577-580 [PMID: 15567325 DOI: 10.1016/j.molmed.2004.10.008]
- 35 **Barker N,** van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegbarth A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* 2007; **449**: 1003-1007 [PMID: 17934449 DOI: 10.1038/nature06196]
- 36 **Zacharias WJ,** Li X, Madison BB, Kretovich K, Kao JY, Merchant JL, Gumucio DL. Hedgehog is an anti-inflammatory epithelial signal for the intestinal lamina propria. *Gastroenterology* 2010; **138**: 2368-2377, 2377. e1-e4 [PMID: 20206176 DOI: 10.1053/j.gastro.2010.02.057]
- 37 **van den Brink GR.** Hedgehog signaling in development and homeostasis of the gastrointestinal tract. *Physiol Rev* 2007; **87**: 1343-1375 [PMID: 17928586 DOI: 10.1152/physrev.00054.2006]
- 38 **Liang R,** Morris P, Cho SS, Abud HE, Jin X, Cheng W. Hedgehog signaling displays a biphasic expression pattern during intestinal injury and repair. *J Pediatr Surg* 2012; **47**: 2251-2263 [PMID: 23217885 DOI: 10.1016/j.jpedsurg.2012.09.016]
- 39 **Blobe GC,** Schiemann WP, Lodish HF. Role of transforming growth factor beta in human disease. *N Engl J Med* 2000; **342**: 1350-1358 [PMID: 10793168 DOI: 10.1056/NEJM200005043421807]
- 40 **Shi Y,** Massagué J. Mechanisms of TGF- $\beta$  signaling from cell membrane to the nucleus. *Cell* 2003; **113**: 685-700 [PMID: 12809600]
- 41 **Winesett MP,** Ramsey GW, Barnard JA. Type II TGF(beta) receptor expression in intestinal cell lines and in the intestinal tract. *Carcinogenesis* 1996; **17**: 989-995 [PMID: 8640948]
- 42 **Fearon ER,** Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; **61**: 759-767 [PMID: 2188735]
- 43 **Haramis AP,** Begthel H, van den Born M, van Es J, Jonkheer S, Offerhaus GJ, Clevers H. De novo crypt formation and juvenile polyposis on BMP inhibition in mouse intestine. *Science* 2004; **303**: 1684-1686 [PMID: 15017003 DOI: 10.1126/science.1093587]
- 44 **He XC,** Zhang J, Tong WG, Tawfik O, Ross J, Scoville DH, Tian Q, Zeng X, He X, Wiedemann LM, Mishina Y, Li L. BMP signaling inhibits intestinal stem cell self-renewal through suppression of Wnt-beta-catenin signaling. *Nat Genet* 2004; **36**: 1117-1121 [PMID: 15378062 DOI: 10.1038/ng1430]
- 45 **Radtke F,** Clevers H, Riccio O. From gut homeostasis to cancer. *Curr Mol Med* 2006; **6**: 275-289 [PMID: 16712475]
- 46 **Zhou XP,** Woodford-Richens K, Lehtonen R, Kurose K, Aldred M, Hampel H, Launonen V, Virta S, Pilarski R, Salovaara R, Bodmer WF, Conrad BA, Dunlop M, Hodgson SV, Iwama T, Järvinen H, Kellokumpu I, Kim JC, Leggett B, Markie D, Mecklin JP, Neale K, Phillips R, Pirus J, Rozen P, Houlston RS, Aaltonen LA, Tomlinson IP, Eng C. Germline mutations in BMPRI1/ALK3 cause a subset of cases of juvenile polyposis syndrome and of Cowden and Bannayan-Riley-Ruvalcaba syndromes. *Am J Hum Genet* 2001; **69**: 704-711 [PMID: 11536076 DOI: 10.1086/323703]
- 47 **Howe JR,** Bair JL, Sayed MG, Anderson ME, Mitros FA, Petersen GM, Velculescu VE, Traverso G, Vogelstein B. Germline mutations of the gene encoding bone morphogenetic protein receptor 1A in juvenile polyposis. *Nat Genet* 2001;

- 28: 184-187 [PMID: 11381269 DOI: 10.1038/88919]
- 48 **Wu Y**, Cain-Hom C, Choy L, Hagenbeek TJ, de Leon GP, Chen Y, Finkle D, Venook R, Wu X, Ridgway J, Schahin-Reed D, Dow GJ, Shelton A, Stawicki S, Watts RJ, Zhang J, Choy R, Howard P, Kadyk L, Yan M, Zha J, Callahan CA, Hymowitz SG, Siebel CW. Therapeutic antibody targeting of individual Notch receptors. *Nature* 2010; **464**: 1052-1057 [PMID: 20393564 DOI: 10.1038/nature08878]
- 49 **Terzić J**, Grivennikov S, Karin E, Karin M. Inflammation and colon cancer. *Gastroenterology* 2010; **138**: 2101-2114.e5 [PMID: 20420949 DOI: 10.1053/j.gastro.2010.01.058]
- 50 **Koch S**, Nava P, Addis C, Kim W, Denning TL, Li L, Parkos CA, Nusrat A. The Wnt antagonist Dkk1 regulates intestinal epithelial homeostasis and wound repair. *Gastroenterology* 2011; **141**: 259-268, 268.e1-8 [PMID: 21440550 DOI: 10.1053/j.gastro.2011.03.043]

**P- Reviewers:** Jeung EB, Morini S

**S- Editor:** Song XX **L- Editor:** A **E- Editor:** Liu XM



## Fibrogenesis and fibrosis in inflammatory bowel diseases: Good and bad side of same coin?

Mariabeatrice Principi, Floriana Giorgio, Giuseppe Losurdo, Viviana Neve, Antonella Contaldo, Alfredo Di Leo, Enzo Ierardi

Mariabeatrice Principi, Floriana Giorgio, Giuseppe Losurdo, Viviana Neve, Antonella Contaldo, Alfredo Di Leo, Enzo Ierardi, Gastroenterology, Department of Emergency and Organ Transplantation, University of Bari, 70124 Bari, Italy  
Author contributions: Ierardi E, Principi M, Giorgio F and Di Leo A designed the study, revised the manuscript and approved the final version; Losurdo G, Neve V and Contaldo A collected the data.

Correspondence to: Enzo Ierardi, Professor, Gastroenterology, Department of Emergency and Organ Transplantation, University of Bari, Piazza Giulio Cesare 11, 70124 Bari, Italy. [enzo.ierardi@fastwebnet.it](mailto:enzo.ierardi@fastwebnet.it)

Telephone: +39-80-5502577 Fax: +39-80-5593251

Received: June 28, 2013 Revised: September 9, 2013

Accepted: October 16, 2013

Published online: November 15, 2013

### Abstract

Fibrogenesis in inflammatory bowel diseases is a complex phenomenon aimed at mucosal repair. However, it may provoke intestinal fibrosis with the development of strictures which require surgery. Therefore, fibrogenesis may be considered as a "two-faced" process when related to chronic intestinal inflammation. Many types of cells may be converted into the fibrogenic phenotype at different levels of the intestinal wall. A complex interaction of cytokines, adhesion molecules and growth factors is involved in the process. We report an overview of recent advances in molecular mechanisms of stricturing Crohn's disease (CD) including the potential role of transforming growth factor beta, protein kinase C and Ras, Raf and ERK proteins. Fibrotic growth factors such as vascular endothelial growth factor and platelet-derived growth factor, as well as the Endothelial-to-Mesenchymal Transition induced by transforming growth factor- $\beta$ , are considered. Finally, our experience, focused on tu-

mor necrosis factor  $\alpha$  (the main cytokine of inflammatory bowel diseases) and the link between syndecan 1 (a heparan sulphate adhesion molecule) and basic fibroblast growth factor (a strong stimulator of collagen synthesis) is described. We hypothesize a possible molecular pattern for mucosal healing as well as how its deregulation could be involved in fibrotic complications of CD. A final clinical point is the importance of performing an accurate evaluation of the presence of fibrotic strictures before starting anti-tumor necrosis  $\alpha$  treatment, which could worsen the lesions.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Fibrogenesis; Fibrosis; Tumor necrosis factor  $\alpha$ ; Syndecan 1; Basic fibroblast growth factor; Cellular fibrogenic phenotype; Inflammatory bowel diseases

**Core tip:** The present minireview reports an outline of the mechanisms of fibrogenesis in inflammatory bowel diseases. Potential fibrogenetic cells and their characterization are detailed. Recent advances in possible molecular mechanisms are highlighted. Our experience, suggesting the hypothesis of a possible molecular mechanism of mucosal healing, is described. The modalities whereby a deregulation of this molecular pattern may lead to fibrotic strictures in Crohn's disease are also illustrated. Finally, possible clinical implications are outlined.

Principi M, Giorgio F, Losurdo G, Neve V, Contaldo A, Di Leo A, Ierardi E. Fibrogenesis and fibrosis in inflammatory bowel diseases: Good and bad side of same coin? *World J Gastrointest Pathophysiol* 2013; 4(4): 100-107 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/100.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.100>

## INTRODUCTION

Crohn's disease (CD) and ulcerative colitis (UC) are immunologically mediated disorders of the gastrointestinal tract in which inflammation and damage are the main findings. Fibrosis may be a complication of both processes, occurring more frequently in CD and often requiring surgical treatment due to the development of stenosis and hence bowel occlusion. However, fibrogenesis is a phenomenon that is intimately involved in mucosal repair<sup>[1]</sup> and, therefore, fibrotic complications of the disorders are paradoxically closely linked to a physiological phenomenon aimed at restoring damaged mucosa.

On these bases, a better understanding of the modalities of the evolution of fibrogenesis into fibrosis is essential, and the issues of how fibrosis differs from normal tissue repair, as well as the identification of cellular mediators for testing possible therapies, are intriguing emerging research concepts.

## CELLULAR BASIS OF FIBROSIS

Fibrosis in CD can be viewed as an extreme healing response to injury. This model predicts that injury causes an initial activation of normal intestinal mesenchymal cells, with a shift to a "fibrogenic" phenotype<sup>[2,3]</sup>. These cells are characterized by an enhanced ability to trigger extracellular matrix (ECM) synthesis. Following acute injury, however, the normal intestinal architecture is restored because post-transcriptional and post-translational mechanisms prevent the accumulation of ECM, while fibrogenic cells are eliminated. By contrast, in fibrosis the mechanisms serving to degrade ECM are not operative at appropriate levels and fibrogenic cells are not only maintained but increase in number. The mechanisms regulating these effects are unknown but may include factors associated with CD, such as cytokines or transmural inflammation.

The normal intestine has a large, heterogeneous population of mesenchymal cells, some of which synthesize significant amounts of collagen. These cells could be considered to have a fibrogenic phenotype and are mainly constituted by fibroblasts and smooth muscle cells or myofibroblasts, as shown by their immunostaining properties with antibodies to vimentin (V) and  $\alpha$ -smooth muscle actin ( $\alpha$ -SMA)<sup>[2,3]</sup>.

Fibroblasts are V+/A-, while smooth muscle cells are V-/A+ and predominate in the normal muscularis mucosa and muscularis propria. Subepithelial myofibroblasts (SEMF) with a V+/A+ phenotype are found adjacent to epithelial cells. However, some of these, that share common features with V+/A+ myofibroblasts, do not express  $\alpha$  but  $\gamma$ -SMA<sup>[2,3]</sup>.

Interstitial cells of Cajal (ICC) are a myofibroblast-related subtype specific to the intestine. They are located between enteric smooth muscle layers and serve to regulate gut motility. The c-kit receptor, which binds the pro-

tooncogene stem cell or steel factor, is a marker of cells of Cajal. Recent studies suggest that in normal human intestine these elements also express vimentin, but not  $\alpha$ -SMA. It has been suggested that ICC could transform into a collagen-expressing fibroblast or myofibroblast phenotype. Another possibility is that ICC are destroyed during fibrosis and replaced by cells with a fibroblast phenotype<sup>[2,3]</sup>.

Inflammatory cells that infiltrate the gut in UC and CD include macrophages, lymphocytes, and plasma cells. These may have important interactions with mesenchymal cells and thereby impact fibrosis<sup>[4,5]</sup>.

In normal intestine, SEMF and fibroblasts found in submucosa, serosa, and intermuscular connective tissue are the primary sites of expression of collagen mRNA and protein. In UC, collagen mRNA expression is up-regulated in SEMF, suggesting that chronic inflammation further increases the activity of fibrogenic cells.

Recent studies in fibrotic intestine from CD patients indicate that V+/A- or V+/A+ fibroblasts and myofibroblasts are the major sites of increased collagen mRNA expression and collagen deposition<sup>[6]</sup>. An overgrowth of the muscularis mucosa and muscularis propria occurs in CD but not UC, and this contributes to the development of stenosis, strictures, and obstruction<sup>[7]</sup>. Muscularis overgrowth also occurs in some animal models of chronic intestinal inflammation and these data support the concept that in muscularis overgrowth in CD, but not in UC, a change in enteric smooth muscle cells towards a fibroblast or myofibroblast phenotype<sup>[8]</sup> is implicated.

## MOLECULAR MECHANISMS OF FIBROGENESIS

Cytokines are a heterogeneous class of secretory proteins produced by several types of cells. For some of them, they act as growth factors, for others as regulators of cellular division and finally, cytokines may paradoxically trigger mechanisms which mediate cell death. All these effects occur *via* regulation of the immune system and inflammation, so cytokines are currently subdivided into pro and anti-inflammatory types. The main cytokine involved in the pathogenesis of both UC and CD is tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )<sup>[9,10]</sup>. The sites of TNF- $\alpha$  production are the mononuclear phagocytes, antigen activated T-lymphocytes, activated mast cells and natural killer cells. Conventional stimulators of TNF- $\alpha$  production are the lipopolysaccharides of the Gram negative bacteria cellular wall, since they are the main mediators of the host response to these organisms. However, TNF- $\alpha$  may be seen as two-faced, since it is able to trigger a closed circle in which, starting from tissue damage, it generates an inflammatory response that exacerbates the damage itself. Moreover, increasing doses of TNF- $\alpha$  may have a lethal effect. Nevertheless, TNF- $\alpha$  plays a key role in the maintenance of tuberculous granuloma, allowing Koch's bacilli to "be walled alive" and thus pre-

venting their spread in the body of an infected person (miliary tuberculosis)<sup>[11]</sup>.

Cell adhesion molecules (CAMs) are proteins located on the cell surface involved in binding with other cells or with the ECM. Essentially, cell adhesion molecules help cells to join together and to their surroundings. These proteins are typically transmembrane receptors and are composed of three domains: an intracellular domain that interacts with the cytoskeleton, a transmembrane domain, and an extracellular domain that interacts either with other CAMs of the same kind (homophilic binding) or with other CAMs or the extracellular matrix (heterophilic binding)<sup>[12]</sup>. A subtype of adhesion molecules containing heparan sulphate (syndecan family) is chemically a proteo-glycan and plays a significant role in tissue repair<sup>[13]</sup>. At intestinal level, syndecan 1 is located in the basolateral region of the columnar epithelium<sup>[14]</sup> and is a relevant factor in the reversal of inflammatory bowel disease (IBD) damage<sup>[15,16]</sup>. Indeed, in inflamed mucosa, these molecules are mainly located in the cells of the stroma and apical epithelium, where the repair of ulcerative lesions will presumably occur.

Basic fibroblast growth factor (bFGF) is a member of the fibroblast growth factor family<sup>[17]</sup>, comprising at least 22 factors with pleiotropic functions<sup>[18]</sup>. This peptide is able to repair ulcerative lesions because of its capacity to bind epithelial and stromal cells. In normal tissue, basic fibroblast growth factor is present in basement membranes and in the subendothelial extracellular matrix of blood vessels. It stays membrane-bound as long as there is no signal peptide, when it acts as a potent angiogenic factor in patho-physiological processes that include wound healing and tissue repair<sup>[19,22]</sup>. The bFGF has been shown to promote proliferation of endothelial cells, to increase the number of fibroblasts and myofibroblasts and to activate these fibroblasts. The induction of collagen secretion from CD and UC fibroblasts by bFGF may be one of the mechanisms inherent to the stromal processes of the disease, including transmural fibrosis and stricture formation, as well as tissue repair and healing.

Klagsbrun *et al*<sup>[23]</sup> suggested that heparan sulphate proteoglycans (and, therefore, syndecan 1) change the bFGF morphology and modulate the structure of its receptors, allowing it to bind to the cells dedicated to the repair process, such as those located at the margins of an ulcerative lesion<sup>[24,25]</sup>. bFGF, when not activated by syndecan 1, is destroyed by luminal and circulating proteases, which may be activated by TNF- $\alpha$ , thus impeding the tissue restoration process.

### **Molecular mechanisms: recent advances**

A relevant actor in the pathogenesis of fibrotic complications of CD is the cytokine transforming growth factor beta (TGF- $\beta$ )<sup>[26]</sup>. TGF- $\beta$  is secreted by many cell types, including macrophages, and has a controlling role in cellular proliferation, differentiation and apoptosis, immunoregulation, supervision of the inflammatory

response, as well as fibrosis and other functions including tissue healing. It is known that TGF- $\beta$  promotes collagen expression by intestinal fibroblasts and smooth muscle cells<sup>[27,28]</sup>. This process is mediated by an intracellular signaling pathway in which a cascade constituted by protein kinase C and Ras, Raf and ERK proteins plays a key role<sup>[26,29,30]</sup>. Moreover, it has been hypothesized that TGF- $\beta$  promotes the overexpression of adhesion molecules (*e.g.*, intracellular adhesion molecule-1)<sup>[31]</sup> by fibroblasts and other pro-fibrotic growth factors such as vascular endothelial growth factor<sup>[32]</sup> and platelet-derived growth factor<sup>[33]</sup>. Finally, a recently revealed mechanism of fibrogenesis in CD, is the Endothelial-to-Mesenchymal Transition induced by TGF- $\beta$ <sup>[34]</sup>. This cytokine is able to induce a protein expression pattern in endothelium that leads to a de-differentiation of these cells and to a transformation to a fibrogenetic phenotype, similar to fibroblasts.

In conclusion, the overexpression of TGF- $\beta$  and its receptors in both the intestinal wall, and in fibroblast cultures taken from sites of intestinal stricture in patients with CD, suggests a potential regulatory role for this cytokine in intestinal fibrogenesis<sup>[35]</sup>.

Adipokines are cell-to-cell signaling proteins produced by adipose tissue. The best known adipokines are leptin, adiponectin and resistin. They play a key role in regulating energy intake and expenditure, including appetite and hunger, metabolism, and behavior: indeed, their role has been widely studied in diseases like metabolic syndrome and type 2 diabetes. Recently, however, further functions of these molecules have been discovered, as mediators of systemic inflammation. It has been shown that obesity per se, and in particular visceral adiposity, is associated with systemic microinflammation, and disturbed circulating adipokines levels<sup>[36,37]</sup>. This is why numerous studies have been focused on the role of adipokines in the pathogenesis of IBD<sup>[38]</sup>.

The outer intestinal wall in patients affected by IBD is enveloped by fat deposits called “wrapping” or “creeping” fat that seem to play an important role in the pathogenesis of IBD. An overexpression of leptin mRNA in mesenteric visceral adipose tissue (mWAT) has also been found in IBD subjects<sup>[39]</sup>, and is correlated with local macrophages infiltration, which drives a high expression of interleukin (IL)-10, IL-6 and TNF- $\alpha$ <sup>[40]</sup>. On the other hand, adiponectin seems to have a protective function against inflammation in IBD<sup>[41,42]</sup>. Rodrigues *et al*<sup>[43]</sup> evaluated serum adiponectin and leptin by enzyme-linked immunosorbent assay in patients with active CD (ACD group), CD in remission (RCD group) and in six healthy controls, and found that serum adiponectin was lower in the ACD group as compared to controls, whereas there were no differences between the ACD and RCD groups.

The Janus kinase/signal transducer and activator of transcription (JAK/STAT) pathway is involved in the control of the expression of DNA sequences that affect basic cell functions, like cell growth, differentiation and death<sup>[44]</sup>. This pathway is triggered by the binding

of a ligand (usually a pro-inflammatory cytokine such as IL-6<sup>[45]</sup>, IL-12, IL-1, TNF- $\alpha$  or interferon gamma<sup>[46]</sup>) to a tyrosine-kinase membrane receptor. Wu *et al*<sup>[47]</sup> evaluated whether intestinal myofibroblasts could produce nitric oxide (NO) in response to the IBD-associated cytokines IL-1b, TNF- $\alpha$ , and interferon gamma in a JAK-STAT dependant pathway, using intestinal myofibroblasts isolated from mice and cultured. The result was an increasing expression of inducible nitric oxide synthase (iNOS) mRNA (evaluated by real time polymerase chain reaction, RT-PCR), but not endothelial NOS or neuronal NOS. This mechanism was shown to be enhanced by a protein cascade constituted by JAK-STAT, Akt and NF- $\kappa$ B. The importance of NO in the pathogenesis of IBD has long been known and is widely discussed in literature<sup>[48,49]</sup>. Furthermore, genetic studies investigating polymorphisms in the *JAK* gene<sup>[50]</sup> revealed that same genetic variants (the G allele of rs744166 and the C allele of rs3816769) increase not only the risk of onset of CD, but even the risk of strictures requiring surgery, because of the interaction with the *CARD15* gene<sup>[51]</sup>. Considering the JAK2 rs10758669 polymorphism, the homozygous C/C or heterozygous A/C genotypes had a higher risk of CD as compared with the homozygous A/A (OR = 1.76, 95%CI: 1.26-2.45 and OR= 2.36, 95%CI: 1.44-3.86, respectively). On these bases, future therapy with JAK inhibitors for their anti-inflammatory effects appears promising<sup>[52]</sup>.

A novel field of genetics which is attracting the attention of researchers is epigenetics, *i.e.*, the study of all the heritable modifications that vary gene expression while not altering the DNA sequence. Micro RNAs (miRNA)<sup>[53]</sup> are just one example. miRNAs are small non-coding RNA oligonucleotides that can regulate the expression of a large number of genes and have been implicated in different human diseases like cancer<sup>[54]</sup> and inflammatory diseases<sup>[55,56]</sup> including IBD<sup>[57]</sup>. A very recent study<sup>[58]</sup> performed on NCM460 human colonocytes incubated with interleukin-6 and on colon biopsies from pediatric and adult patients with UC revealed that a deregulation (low levels) of miRNA-124 can cause a hyper-phosphorylation of STAT3 (and consequently, hyper-activation), *via* a mechanism induced by IL-6, very likely resulting in a pathogenic system leading to IBD. miRNA-124 is only a second lead on the crowded stage of miRNAs: miRNA-192, which is normally expressed in colonic epithelial cells, is significantly reduced in tissues of patients with active UC<sup>[59]</sup>; miR-150 is up-regulated in mice with dextran sulfate sodium-induced colitis and in colon tissues from patients with UC<sup>[60]</sup>; an overexpression of miRNA-21, which promotes inflammation, has been reported in several studies of patients with active UC and CD colitis<sup>[61]</sup>.

The process of DNA methylation is another form of epigenetic regulation of gene expression. It consists in the binding of a methyl group to cytosines that are part of cytosine-guanine dinucleotides (CpG), and has a gene silencing function. The main genes whose methylation

is involved in the pathogenesis of IBD are the CDH1<sup>[62]</sup>, BCL3, STAT3, OSM, STAT5<sup>[63]</sup> proteins involved in the IL-12 and IL-23 pathways.

Finally, histone modifications are an epigenetic process that may modify genomic expression. Histones are alkaline proteins that package and order the DNA into structural units called nucleosomes and chromatin<sup>[64]</sup>. They have N-terminal amino acid tails that protrude and can be modified by acetylation, methylation, ubiquitination, and phosphorylation. Acetylation, however, is the most closely studied phenomenon, because it improves gene transcription and recruitment of transcriptional factors. In the course of IBD, the main genes targeted by histone acetylation are *p53*, *STAT3*, and *HIF1 $\alpha$* <sup>[65]</sup>. Furthermore, an innate immune response to microbiota has been proposed to link histone modifications with inflammation: butyrate, an endogenous metabolite formed during fermentation of dietary fibers by the intestinal microbiota, is a histone deacetylase inhibitor. Butyrate increases the expression of NOD2 by increasing histone acetylation in its promoter region<sup>[66]</sup>.

The importance of all these epigenetic mechanisms lies in possible future therapeutic applications: inhibitors of deacetylation, demethylating agents and miRNA produced by genetic engineering could be potential targets in IBD<sup>[67]</sup>.

### **Molecular mechanisms in mucosal healing and strictures: our experience**

Our previous investigations<sup>[68]</sup> demonstrated that a decrease of TNF- $\alpha$  induced by anti-TNF- $\alpha$  (infliximab) treatment is accompanied by a decrease in both syndecan 1 and bFGF when mucosal healing occurs. A possible explanation is that infliximab therapy may downregulate, *via* a marked reduction of TNF- $\alpha$  mucosal levels, the bFGF/syndecan 1 link. This molecular profile could represent a pathway of mucosal healing. However, the parallel trend of TNF- $\alpha$ , syndecan 1 and bFGF could be just a simultaneous consequence of the control of inflammation. To dispel this doubt, we analyzed the “timing” of the TNF- $\alpha$  decrease and bFGF/syndecan 1 reversal to normal levels and sites in cultured biopsy samples taken from patients with both CD and UC and incubated in a medium containing comparable amounts of infliximab similar to those reached in the serum of treated patients. After 24 h we assayed TNF- $\alpha$ , syndecan 1 and bFGF in tissue homogenates. TNF- $\alpha$  was found to be decreased, while syndecan 1 and bFGF levels were still high when evaluated by both a molecular method (reverse transcriptase real time polymerase chain reaction) and immunohistochemistry<sup>[69]</sup>. This last finding supports our primary hypothesis that a mucosal TNF- $\alpha$  reset, induced by biological drugs, is followed by a mucosal restoration in which syndecan 1 modulates the strong reparative bFGF aptitudes. Finally, in healed mucosa, cytokines, adhesion molecules and growth factors resume their normal pattern.

A report by Bousvaros *et al*<sup>[70]</sup> showed that in chil-

dren with CD there was a strong correlation between the bFGF level and disease activity. The relationship of bFGF with disease activity persisted even after adjusting for other covariates (including age, sex, hematocrit, albumin, and sedimentation rate) in a multivariate linear regression model. There was also a statistically significant, although less strong, correlation between the bFGF level and disease activity in UC. Although bFGF is not a specific marker for CD, its serum levels reflect disease activity. Therefore, bFGF release may be important in mediating the angiogenesis and wound healing seen in CD. This report, as well as our experience both *in vivo* and *in vitro*<sup>[68,69]</sup>, suggest a similar molecular mechanism for mucosal healing both in CD and UC.

In a further study, we investigated the pattern of TNF- $\alpha$ , syndecan 1 and bFGF in patients with CD complicated by fibrotic stenosis undergoing surgical resection<sup>[71]</sup>. We observed that TNF- $\alpha$  mucosal levels were not significantly increased. A possible explanation for this finding may be that an overgrowth of fibrotic tissue may be a successive stage after inflammation, where the increase in TNF- $\alpha$  is the peculiar aspect. Therefore, at the stage of fibrotic stenosis requiring surgery, inflammatory mucosal changes may be an irrelevant phenomenon in most patients. Syndecan 1 levels were increased, showing a pattern similar to the one observed in damaged tissues. It may be presumed that the molecule location, albeit limited to the mucosal layer, reflects an attempt at bFGF modulation. However, this function cannot be effectively carried out due to the bFGF overexpression and location all along the intestinal wall, *i.e.*, outside the district where syndecan 1 could operate<sup>[14]</sup>. Indeed, bFGF overexpression affects all intestinal wall layers, being expressed in epithelium, stroma, muscularis propria and endothelium. It is possible that: (1) the low levels of TNF- $\alpha$  may provoke a failure in cytokine induced bFGF proteolysis; (2) the presence of syndecan 1 is limited to the mucosal layer with a consequent very partial regulation of bFGF binding to specific receptors dedicated to tissue repair; and (3) an irreversible transformation of different type of cells to the fibrogenetic phenotype occurs, thus provoking the prevalence of fibrotic over inflammatory stenotic lesions<sup>[72]</sup>.

In strictures, it is possible that the excess extracellular matrix cannot be inhibited by the regulatory mechanisms of the phenomenon, in accordance with the hypothesis proposed by Pucilowska *et al.*<sup>[72]</sup>. On these bases, we may conclude that the different molecular patterns in repair dedicated fibrogenesis and stricturizing fibrosis in CD could be the consequence of different mucosal levels of TNF- $\alpha$ . These are very high in the active disorder, but undergo a progressive depletion in the long term, and this event may trigger a polymorphic regulation of the syndecan 1/bFGF system.

## FINAL REMARKS

Fibrogenesis in inflammatory bowel diseases is a phenomenon aimed at tissue repair. Many cellular types are

involved in this process and cytokines, adhesion molecules and growth factors interact to achieve mucosal repair. However, a deregulation of the healing molecular pathway can progress towards fibrosis and stenotic complications often requiring surgical therapy<sup>[73,74]</sup>. Therefore, fibrogenesis and fibrosis may represent the good and bad sides of the same coin, the former allowing lesion healing but the latter leading to severe complications. The main tool for discriminating between them is, in our experience, the presence/absence of inflammation (and, consequently, the level of TNF- $\alpha$  expression).

A final clinical consideration is the importance of making an accurate evaluation<sup>[75]</sup> in cases of stenosis in the course of CD using all available diagnostic tools (histology, ultrasonography<sup>[76]</sup> with Doppler evaluation of the resistance index, magnetic resonance<sup>[77]</sup>, computed tomography<sup>[78]</sup>, biochemical indices of inflammation<sup>[79,80]</sup>) in order to distinguish inflammatory from fibrotic stenosis. This could orient anti-TNF- $\alpha$  therapy<sup>[81]</sup>, that should be limited to the first case, avoiding the risk that the cytokine decrease could support fibrotic complications rather than mucosal healing.

## REFERENCES

- 1 **Specia S**, Giusti I, Rieder F, Latella G. Cellular and molecular mechanisms of intestinal fibrosis. *World J Gastroenterol* 2012; **18**: 3635-3661 [PMID: 22851857 DOI: 10.3748/wjg.v18.i28.3635]
- 2 **Powell DW**, Mifflin RC, Valentich JD, Crowe SE, Saada JI, West AB. Myofibroblasts. II. Intestinal subepithelial myofibroblasts. *Am J Physiol* 1999; **277**: C183-C201 [PMID: 10444394]
- 3 **Pucilowska JB**, McNaughton KK, Mohapatra NK, Hoyt EC, Zimmermann EM, Sartor RB, Lund PK. IGF- I and procollagen  $\alpha 1(I)$  are coexpressed in a subset of mesenchymal cells in active Crohn's disease. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G1307-G1322 [PMID: 11093955]
- 4 **Fiocchi C**. Inflammatory bowel disease: etiology and pathogenesis. *Gastroenterology* 1998; **115**: 182-205 [PMID: 9649475]
- 5 **Matthes H**, Herbst H, Schuppan D, Stallmach A, Milani S, Stein H, Riecken EO. Cellular localization of procollagen gene transcripts in inflammatory bowel diseases. *Gastroenterology* 1992; **102**: 431-442 [PMID: 1732114]
- 6 **Dvorak AM**, Osage JE, Monahan RA, Dickersin GR. Crohn's disease: transmission electron microscopic studies. III. Target tissues. Proliferation of and injury to smooth muscle and the autonomic nervous system. *Hum Pathol* 1980; **11**: 620-634 [PMID: 6161074]
- 7 **Becker JM**. Surgical therapy for ulcerative colitis and Crohn's disease. *Gastroenterol Clin North Am* 1999; **28**: 371-390, viii-ix [PMID: 10372273]
- 8 **Williams KL**, Fuller CR, Dieleman LA, DaCosta CM, Haldeman KM, Sartor RB, Lund PK. Enhanced survival and mucosal repair after dextran sodium sulfate-induced colitis in transgenic mice that overexpress growth hormone. *Gastroenterology* 2001; **120**: 925-937 [PMID: 11231946]
- 9 **Locksley RM**, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* 2001; **104**: 487-501 [PMID: 11239407 DOI: 10.1016/S0092-8674(01)00237-9]
- 10 **Ierardi E**, Giorgio F, Rosania R, Zotti M, Prencipe S, Della Valle N, De Francesco V, Panella C. Mucosal assessment of tumor necrosis factor  $\alpha$  levels on paraffined samples: a comparison between immunohistochemistry and real time polymerase chain reaction. *Scand J Gastroenterol* 2010; **45**:

- 1007-1008 [PMID: 20446801]
- 11 **Xie X**, Li F, Chen JW, Wang J. Risk of tuberculosis infection in anti-TNF- $\alpha$  biological therapy: From bench to bedside. *J Microbiol Immunol Infect* 2013 May 30; [Epub ahead of print] [PMID: 23727394 DOI: 10.1016/j.jmii.2013.03.005]
  - 12 **Vainer B**. Role of cell adhesion molecules in inflammatory bowel diseases. *Scand J Gastroenterol* 1997; **32**: 401-410 [PMID: 9175198]
  - 13 **Saunders S**, Jalkanen M, O'Farrell S, Bernfield M. Molecular cloning of syndecan, an integral membrane proteoglycan. *J Cell Biol* 1989; **108**: 1547-1556 [PMID: 2494194]
  - 14 **Principi M**, Day R, Marangi S, Burattini O, De Francesco V, Ingrosso M, Pisani A, Panella C, Forbes A, Di Leo A, Francavilla A, Ierardi E. Differential immunohistochemical expression of syndecan-1 and tumor necrosis factor  $\alpha$  in colonic mucosa of patients with Crohn's disease. *Immunopharmacol Immunotoxicol* 2006; **28**: 185-195 [PMID: 16873088]
  - 15 **Day R**, Forbes A. Heparin, cell adhesion, and pathogenesis of inflammatory bowel disease. *Lancet* 1999; **354**: 62-65 [PMID: 10406379]
  - 16 **Day R**, Ilyas M, Daszak P, Talbot I, Forbes A. Expression of syndecan-1 in inflammatory bowel disease and a possible mechanism of heparin therapy. *Dig Dis Sci* 1999; **44**: 2508-2515 [PMID: 10630505]
  - 17 **Kim HS**. Assignment1 of the human basic fibroblast growth factor gene FGF2 to chromosome 4 band q26 by radiation hybrid mapping. *Cytogenet Cell Genet* 1998; **83**: 73 [PMID: 9925931 DOI: 10.1159/000015129]
  - 18 **Ornitz DM**, Itoh N. Fibroblast growth factors. *Genome Biol* 2001; **2**: REVIEWS3005 [PMID: 11276432]
  - 19 **Fernig DG**, Gallagher JT. Fibroblast growth factors and their receptors: an information network controlling tissue growth, morphogenesis and repair. *Prog Growth Factor Res* 1994; **5**: 353-377 [PMID: 7780086]
  - 20 **Okada-Ban M**, Thiery JP, Jouanneau J. Fibroblast growth factor-2. *Int J Biochem Cell Biol* 2000; **32**: 263-267 [PMID: 10716624]
  - 21 **Szebenyi G**, Fallon JF. Fibroblast growth factors as multifunctional signaling factors. *Int Rev Cytol* 1999; **185**: 45-106 [PMID: 9750265]
  - 22 **Galzie Z**, Kinsella AR, Smith JA. Fibroblast growth factors and their receptors. *Biochem Cell Biol* 1997; **75**: 669-685 [PMID: 9599656]
  - 23 **Klagsbrun M**, Baird A. A dual receptor system is required for basic fibroblast growth factor activity. *Cell* 1991; **67**: 229-231 [PMID: 1655276]
  - 24 **Beck PL**, Podolsky DK. Growth factors in inflammatory bowel disease. *Inflamm Bowel Dis* 1999; **5**: 44-60 [PMID: 10028449]
  - 25 **Yoshida S**, Ono M, Shono T, Izumi H, Ishibashi T, Suzuki H, Kuwano M. Involvement of interleukin-8, vascular endothelial growth factor, and basic fibroblast growth factor in tumor necrosis factor  $\alpha$ -dependent angiogenesis. *Mol Cell Biol* 1997; **17**: 4015-4023 [PMID: 9199336]
  - 26 **Mulsow JJ**, Watson RW, Fitzpatrick JM, O'Connell PR. Transforming growth factor-beta promotes pro-fibrotic behavior by serosal fibroblasts via PKC and ERK1/2 mitogen activated protein kinase cell signaling. *Ann Surg* 2005; **242**: 880-887, discussion 887-889 [PMID: 16327498]
  - 27 **Stallmach A**, Schuppan D, Riese HH, Matthes H, Riecken EO. Increased collagen type III synthesis by fibroblasts isolated from strictures of patients with Crohn's disease. *Gastroenterology* 1992; **102**: 1920-1929 [PMID: 1587410]
  - 28 **Graham MF**, Bryson GR, Diegelmann RF. Transforming growth factor beta 1 selectively augments collagen synthesis by human intestinal smooth muscle cells. *Gastroenterology* 1990; **99**: 447-453 [PMID: 2365193]
  - 29 **Chen Y**, Blom IE, Sa S, Goldschmeding R, Abraham DJ, Leask A. CTGF expression in mesangial cells: involvement of SMADs, MAP kinase, and PKC. *Kidney Int* 2002; **62**: 1149-1159 [PMID: 12234285]
  - 30 **Leask A**, Holmes A, Black CM, Abraham DJ. Connective tissue growth factor gene regulation. Requirements for its induction by transforming growth factor-beta 2 in fibroblasts. *J Biol Chem* 2003; **278**: 13008-13015 [PMID: 12571253]
  - 31 **Brannigan AE**, Watson RW, Beddy D, Hurley H, Fitzpatrick JM, O'Connell PR. Increased adhesion molecule expression in serosal fibroblasts isolated from patients with inflammatory bowel disease is secondary to inflammation. *Ann Surg* 2002; **235**: 507-511 [PMID: 11923606]
  - 32 **Beddy D**, Watson RW, Fitzpatrick JM, O'Connell PR. Increased vascular endothelial growth factor production in fibroblasts isolated from strictures in patients with Crohn's disease. *Br J Surg* 2004; **91**: 72-77 [PMID: 14716797]
  - 33 **Kumagai S**, Ohtani H, Nagai T, Funa K, Hiwatashi NO, Shimosegawa H. Platelet-derived growth factor and its receptors are expressed in areas of both active inflammation and active fibrosis in inflammatory bowel disease. *Tohoku J Exp Med* 2001; **195**: 21-33 [PMID: 11780721]
  - 34 **Rieder F**, Kessler SP, West GA, Bhilocha S, de la Motte C, Sadler TM, Gopalan B, Stylianou E, Fiocchi C. Inflammation-induced endothelial-to-mesenchymal transition: a novel mechanism of intestinal fibrosis. *Am J Pathol* 2011; **179**: 2660-2673 [PMID: 21945322 DOI: 10.1016/j.ajpath.2011.07.042.]
  - 35 **Burke JP**, Mulsow JJ, O'Keane C, Docherty NG, Watson RW, O'Connell PR. Fibrogenesis in Crohn's disease. *Am J Gastroenterol* 2007; **102**: 439-448 [PMID: 17156147]
  - 36 **Olszanecka-Glinianowicz M**, Chudek J, Kocelak P, Szromek A, Zahorska-Markiewicz B. Body fat changes and activity of tumor necrosis factor  $\alpha$  system--a 5-year follow-up study. *Metabolism* 2011; **60**: 531-536 [PMID: 20580040 DOI: 10.1016/j.metabol.2010.04.023]
  - 37 **Olszanecka-Glinianowicz M**, Zahorska-Markiewicz B, Janowska J, Zurakowski A. Serum concentrations of nitric oxide, tumor necrosis factor (TNF)- $\alpha$  and TNF soluble receptors in women with overweight and obesity. *Metabolism* 2004; **53**: 1268-1273 [PMID: 15375781]
  - 38 **Olszanecka-Glinianowicz M**, Handzlik-Orlik G, Orlik B, Chudek J. Adipokines in the pathogenesis of idiopathic inflammatory bowel disease. *Endokrynol Pol* 2013; **64**: 226-231 [PMID: 23873428]
  - 39 **Barbier M**, Vidal H, Desreumaux P, Dubuquoy L, Bourreille A, Colombel JF, Cherbut C, Galmiche JP. Overexpression of leptin mRNA in mesenteric adipose tissue in inflammatory bowel diseases. *Gastroenterol Clin Biol* 2003; **27**: 987-991 [PMID: 14732844]
  - 40 **Kredel LI**, Batra A, Stroh T, Kühl AA, Zeitz M, Erben U, Siegmund B. Adipokines from local fat cells shape the macrophage compartment of the creeping fat in Crohn's disease. *Gut* 2013; **62**: 852-862 [PMID: 22543156 DOI: 10.1136/gutjnl-2011-301424]
  - 41 **Karmiris K**, Koutroubakis IE, Xidakis C, Polychronaki M, Voudouri T, Kouroumalis EA. Circulating levels of leptin, adiponectin, resistin, and ghrelin in inflammatory bowel disease. *Inflamm Bowel Dis* 2006; **12**: 100-105 [PMID: 16432373]
  - 42 **Fantuzzi G**. Adiponectin and inflammation: consensus and controversy. *J Allergy Clin Immunol* 2008; **121**: 326-330 [PMID: 18061654]
  - 43 **Rodrigues VS**, Milanski M, Fagundes JJ, Torsoni AS, Ayrisono ML, Nunez CE, Dias CB, Meirelles LR, Dalal S, Coy CS, Velloso LA, Leal RF. Serum levels and mesenteric fat tissue expression of adiponectin and leptin in patients with Crohn's disease. *Clin Exp Immunol* 2012; **170**: 358-364 [PMID: 23121676 DOI: 10.1111/j.1365-2249.2012.04660.x]
  - 44 **Aaronson DS**, Horvath CM. A road map for those who don't know JAK-STAT. *Science* 2002; **296**: 1653-1655 [PMID: 12040185]
  - 45 **Ito H**. IL-6 and Crohn's disease. *Curr Drug Targets Inflamm Allergy* 2003; **2**: 125-130 [PMID: 14561164]

- 46 **O'Shea JJ**, Gadina M, Schreiber RD. Cytokine signaling in 2002: new surprises in the Jak/Stat pathway. *Cell* 2002; **109** Suppl: S121-S131 [PMID: 11983158]
- 47 **Wu J**, Chitapanarux T, Chen Y, Soon RK, Yee HF. Intestinal myofibroblasts produce nitric oxide in response to combinatorial cytokine stimulation. *J Cell Physiol* 2013; **228**: 572-580 [PMID: 22833357 DOI: 10.1002/jcp.24164]
- 48 **Lundberg JO**, Hellström PM, Lundberg JM, Alving K. Greatly increased luminal nitric oxide in ulcerative colitis. *Lancet* 1994; **344**: 1673-1674 [PMID: 7996962]
- 49 **Boughton-Smith NK**, Evans SM, Hawkey CJ, Cole AT, Balsitis M, Whittle BJ, Moncada S. Nitric oxide synthase activity in ulcerative colitis and Crohn's disease. *Lancet* 1993; **342**: 338-340 [PMID: 7687730]
- 50 **Ferguson LR**, Han DY, Fraser AG, Huebner C, Lam WJ, Morgan AR, Duan H, Karunasinghe N. Genetic factors in chronic inflammation: single nucleotide polymorphisms in the STAT-JAK pathway, susceptibility to DNA damage and Crohn's disease in a New Zealand population. *Mutat Res* 2010; **690**: 108-115 [PMID: 20109474 DOI: 10.1016/j.mrfmmm.2010.01.017]
- 51 **Barrett JC**, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, Brant SR, Silverberg MS, Taylor KD, Barmada MM, Bitton A, Dassopoulos T, Datta LW, Green T, Griffiths AM, Kistner EO, Murtha MT, Regueiro MD, Rotter JJ, Schumm LP, Steinhardt AH, Targan SR, Xavier RJ, Libioulle C, Sandor C, Lathrop M, Belaiche J, Dewit O, Gut I, Heath S, Laukens D, Mni M, Rutgeerts P, Van Gossum A, Zelenika D, Franchimont D, Hugot JP, de Vos M, Vermeire S, Louis E, Cardon LR, Anderson CA, Drummond H, Nimmo E, Ahmad T, Prescott NJ, Onnie CM, Fisher SA, Marchini J, Ghori J, Bumpstead S, Gwilliam R, Tremelling M, Deloukas P, Mansfield J, Jewell D, Satsangi J, Mathew CG, Parkes M, Georges M, Daly MJ. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 2008; **40**: 955-962 [PMID: 18587394 DOI: 10.1038/ng.175]
- 52 **Coskun M**, Salem M, Pedersen J, Nielsen OH. Involvement of JAK/STAT signaling in the pathogenesis of inflammatory bowel disease. *Pharmacol Res* 2013; **76**: 1-8 [PMID: 23827161 DOI: 10.1016/j.phrs.2013.06.007]
- 53 **Bartel DP**. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; **136**: 215-233 [PMID: 19167326 DOI: 10.1016/j.cell.2009.01.021]
- 54 **Hatziapostolou M**, Polyarchou C, Aggelidou E, Drakaki A, Poultsides GA, Jaeger SA, Ogata H, Karin M, Struhl K, Hadzopoulou-Cladaras M, Iliopoulos D. An HNF4 $\alpha$ -miRNA inflammatory feedback circuit regulates hepatocellular oncogenesis. *Cell* 2011; **147**: 1233-1247 [PMID: 22153071 DOI: 10.1016/j.cell.2011.10.043]
- 55 **Stagakis E**, Bertias G, Verginis P, Nakou M, Hatziapostolou M, Kritikos H, Iliopoulos D, Boumpas DT. Identification of novel microRNA signatures linked to human lupus disease activity and pathogenesis: miR-21 regulates aberrant T cell responses through regulation of PDCD4 expression. *Ann Rheum Dis* 2011; **70**: 1496-1506 [PMID: 21602271 DOI: 10.1136/ard.2010.139857]
- 56 **Du C**, Liu C, Kang J, Zhao G, Ye Z, Huang S, Li Z, Wu Z, Pei G. MicroRNA miR-326 regulates TH-17 differentiation and is associated with the pathogenesis of multiple sclerosis. *Nat Immunol* 2009; **10**: 1252-1259 [PMID: 19838199 DOI: 10.1038/ni.1798]
- 57 **Dalal SR**, Kwon JH. The Role of MicroRNA in Inflammatory Bowel Disease. *Gastroenterol Hepatol (N Y)* 2010; **6**: 714-722 [PMID: 21437020]
- 58 **Koukos G**, Polyarchou C, Kaplan JL, Morley-Fletcher A, Gras-Mirallas B, Kokkotou E, Baril-Dore M, Pothoulakis C, Winter HS, Iliopoulos D. MicroRNA-124 Regulates STAT3 Expression and Is Down-regulated in Colon Tissues of Pediatric Patients With Ulcerative Colitis. *Gastroenterology* 2013; **145**: 842-852. e2 [PMID: 23856509 DOI: 10.1053/j.gastro.2013.07.001]
- 59 **Wu F**, Zikusoka M, Trindade A, Dassopoulos T, Harris ML, Bayless TM, Brant SR, Chakravarti S, Kwon JH. MicroRNAs are differentially expressed in ulcerative colitis and alter expression of macrophage inflammatory peptide-2  $\alpha$ . *Gastroenterology* 2008; **135**: 1624-1635. e24 [PMID: 18835392 DOI: 10.1053/j.gastro.2008.07.068]
- 60 **Bian Z**, Li L, Cui J, Zhang H, Liu Y, Zhang CY, Zen K. Role of miR-150-targeting c-Myb in colonic epithelial disruption during dextran sulphate sodium-induced murine experimental colitis and human ulcerative colitis. *J Pathol* 2011; **225**: 544-553 [PMID: 21590770 DOI: 10.1002/path.2907]
- 61 **Fasseu M**, Tréton X, Guichard C, Pedruzzi E, Cazals-Hatem D, Richard C, Aparicio T, Daniel F, Soulé JC, Moreau R, Bouhnik Y, Laburthe M, Groyer A, Ogier-Denis E. Identification of restricted subsets of mature microRNA abnormally expressed in inactive colonic mucosa of patients with inflammatory bowel disease. *PLoS One* 2010; **5**: pii: e13160 [PMID: 20957151 DOI: 10.1371/journal.pone.0013160]
- 62 **Saito S**, Kato J, Hiraoka S, Horii J, Suzuki H, Higashi R, Kaji E, Kondo Y, Yamamoto K. DNA methylation of colon mucosa in ulcerative colitis patients: correlation with inflammatory status. *Inflamm Bowel Dis* 2011; **17**: 1955-1965 [PMID: 21830274 DOI: 10.1002/ibd.21573]
- 63 **Lin Z**, Hegarty JP, Yu W, Cappel JA, Chen X, Faber PW, Wang Y, Poritz LS, Fan JB, Koltun WA. Identification of disease-associated DNA methylation in B cells from Crohn's disease and ulcerative colitis patients. *Dig Dis Sci* 2012; **57**: 3145-3153 [PMID: 22821069 DOI: 10.1007/s10620-012-2288-z]
- 64 **Kouzarides T**. Chromatin modifications and their function. *Cell* 2007; **128**: 693-705 [PMID: 17320507]
- 65 **Glauben R**, Siegmund B. Inhibition of histone deacetylases in inflammatory bowel diseases. *Mol Med* 2011; **17**: 426-433 [PMID: 21365125]
- 66 **Leung CH**, Lam W, Ma DL, Gullen EA, Cheng YC. Butyrate mediates nucleotide-binding and oligomerisation domain (NOD) 2-dependent mucosal immune responses against peptidoglycan. *Eur J Immunol* 2009; **39**: 3529-3537 [PMID: 19830732 DOI: 10.1002/eji.200939454]
- 67 **Ventham NT**, Kennedy NA, Nimmo ER, Satsangi J. Beyond gene discovery in inflammatory bowel disease: the emerging role of epigenetics. *Gastroenterology* 2013; **145**: 293-308 [PMID: 23751777 DOI: 10.1053/j.gastro.2013.05.050]
- 68 **Ierardi E**, Giorgio F, Zotti M, Rosania R, Principi M, Marangi S, Della Valle N, De Francesco V, Di Leo A, Ingrosso M, Panella C. Infliximab therapy downregulation of basic fibroblast growth factor/syndecan 1 link: a possible molecular pathway of mucosal healing in ulcerative colitis. *J Clin Pathol* 2011; **64**: 968-972 [PMID: 21945924 DOI: 10.1136/jcp.2010.086892]
- 69 **Della Valle N**, Giorgio F, Cantatore S, Zotti M, Ierardi E, Panella C. Tumor Necrosis Factor  $\alpha$ , syndecan 1 and basic fibroblast growth factor levels and site in cultured biopsy specimens of patients with inflammatory bowel diseases after incubation with infliximab (Abstract). *Dig Liver Dis* 2013; **45**: S107 [DOI: 10.1016/S1590-8658(13)60296-0]
- 70 **Bousvaros A**, Zurakowski D, Fishman SJ, Keough K, Law T, Sun C, Leichtner AM. Serum basic fibroblast growth factor in pediatric Crohn's disease. Implications for wound healing. *Dig Dis Sci* 1997; **42**: 378-386 [PMID: 9052523]
- 71 **Ierardi E**, Giorgio F, Piscitelli D, Principi M, Cantatore S, Fiore MG, Rossi R, Barone M, Di Leo A, Panella C. Altered molecular pattern of mucosal healing in Crohn's disease fibrotic stenosis. *World J Gastrointest Pathophysiol* 2013; **4**: 53-58 [PMID: 23946888 DOI: 10.4291/wjgp.v4.i3.53]
- 72 **Pucilowska JB**, Williams KL, Lund PK. Fibrogenesis. IV. Fibrosis and inflammatory bowel disease: cellular mediators and animal models. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G653-G659 [PMID: 11005750]
- 73 **Schoepfer AM**, Safroneeva E, Vavricka SR, Peyrin-Biroulet

- L, Mottet C. Treatment of fibrostenotic and fistulizing Crohn's disease. *Digestion* 2012; **86** Suppl 1: 23-27 [PMID: 23051723 DOI: 10.1159/000341961]
- 74 **Ono Y**, Hirai F, Matsui T, Beppu T, Yano Y, Takatsu N, Takaki Y, Nagahama T, Hisabe T, Yao K, Higashi D, Futami K. Value of concomitant endoscopic balloon dilation for intestinal stricture during long-term infliximab therapy in patients with Crohn's disease. *Dig Endosc* 2012; **24**: 432-438 [PMID: 23078435 DOI: 10.1111/j.1443-1661.2012.01315.x]
- 75 **Panes J**, Bouhnik Y, Reinisch W, Stoker J, Taylor SA, Baumgart DC, Danese S, Halligan S, Marincek B, Matos C, Peyrin-Biroulet L, Rimola J, Rogler G, van Assche G, Ardizzone S, Ba-Ssalamah A, Bali MA, Bellini D, Biancone L, Castiglione F, Ehehalt R, Grassi R, Kucharzik T, Maccioni F, Maconi G, Magro F, Martín-Comín J, Morana G, Pendsé D, Sebastian S, Signore A, Tolan D, Tielbeek JA, Weishaupt D, Wiarda B, Laghi A. Imaging techniques for assessment of inflammatory bowel disease: Joint ECCO and ESGAR evidence-based consensus guidelines. *J Crohns Colitis* 2013; **7**: 556-785 [PMID: 23583097 DOI: 10.1016/j.crohns.2013.02.020]
- 76 **Novak KL**, Wilson SR. The role of ultrasound in the evaluation of inflammatory bowel disease. *Semin Roentgenol* 2013; **48**: 224-233 [PMID: 23796373 DOI: 10.1053/j.ro.2013.03.003.]
- 77 **Lawrance IC**, Welman CJ, Shipman P, Murray K. Correlation of MRI-determined small bowel Crohn's disease categories with medical response and surgical pathology. *World J Gastroenterol* 2009; **15**: 3367-3375 [PMID: 19610137]
- 78 **Zhang LH**, Zhang SZ, Hu HJ, Gao M, Zhang M, Cao Q, Zhang QW. Multi-detector CT enterography with iso-osmotic mannitol as oral contrast for detecting small bowel disease. *World J Gastroenterol* 2005; **11**: 2324-2329 [PMID: 15818746]
- 79 **Hauser G**, Tkalcic M, Pletikosic S, Grabar N, Stimac D. Erythrocyte sedimentation rate - possible role in determining the existence of the low grade inflammation in Irritable Bowel Syndrome patients. *Med Hypotheses* 2012; **78**: 818-820 [PMID: 22513237 DOI: 10.1016/j.mehy.2012.03.020]
- 80 **Ricanek P**, Brackmann S, Perminow G, Lyckander LG, Sponheim J, Holme O, Høie O, Rydning A, Vatn MH. Evaluation of disease activity in IBD at the time of diagnosis by the use of clinical, biochemical, and fecal markers. *Scand J Gastroenterol* 2011; **46**: 1081-1091 [PMID: 21619483 DOI: 10.3109/00365521.2011.584897]
- 81 **Sorrentino D**, Avellini C, Beltrami CA, Pasqual E, Zearo E. Selective effect of infliximab on the inflammatory component of a colonic stricture in Crohn's disease. *Int J Colorectal Dis* 2006; **21**: 276-281 [PMID: 15951989]

**P- Reviewers:** Feo F, Grizzi F, Sipos F, Tarantino G  
**S- Editor:** Zhai HH **L- Editor:** A **E- Editor:** Liu XM



## Effects of occupational stress on the gastrointestinal tract

María-Raquel Huerta-Franco, Miguel Vargas-Luna, Paola Tienda, Isabel Delgadillo-Holtfort, Marco Balleza-Ordaz, Corina Flores-Hernandez

María-Raquel Huerta-Franco, Paola Tienda, Corina Flores-Hernandez, Department of Applied Science and Labor Research, DCS Campus Leon, University of Guanajuato, Leon, CP 37320, Mexico

Miguel Vargas-Luna, Isabel Delgadillo-Holtfort, Marco Balleza-Ordaz, Department of Physical Engineering, DCI, Campus Leon, University of Guanajuato, Leon, CP 37150, Mexico

Author contributions: Huerta-Franco MR and Vargas-Luna M designed and conducted the study and wrote the manuscript; Tienda P, Delgadillo-Holtfort I, Flores-Hernandez C and Balleza-Ordaz M contributed to the data collection, reviewed the literature and provided analytic input.

Supported by Dirección de Apoyo a la Investigación y al Posgrado (DAIP); University of Guanajuato (2012-2013); and Programa Integral de Fortalecimiento Institucional (PIFI-SEP) 2012

Correspondence to: María-Raquel Huerta-Franco, PhD, Department of Applied Science and Labor Research, DCS Campus Leon, University of Guanajuato, Aquiles Serdan No. 924, Colonia Obregon, Leon, CP 37150, Guanajuato, Mexico. [huertafranco@hotmail.com](mailto:huertafranco@hotmail.com)

Telephone: +52-477-2569688 Fax: +52-477-7885100

Received: July 2, 2013 Revised: September 12, 2013

Accepted: October 16, 2013

Published online: November 15, 2013

### Abstract

The aim of this review is to provide a general overview of the relationship between occupational stress and gastrointestinal alterations. The International Labour Organization suggests occupational health includes psychological aspects to achieve mental well-being. However, the definition of health risks for an occupation includes biological, chemical, physical and ergonomic factors but does not address psychological stress or other affective disorders. Nevertheless, multiple investigations have studied occupational stress and its physiological consequences, focusing on specific risk groups and occupations considered stressful. Among the physiological effects of stress, gastrointestinal tract (GIT) alterations are highly prevalent. The relationship

between occupational stress and GIT diseases is evident in everyday clinical practice; however, the usual strategy is to attack the effects but not the root of the problem. That is, in clinics, occupational stress is recognized as a source of GIT problems, but employers do not ascribe it enough importance as a risk factor, in general, and for gastrointestinal health, in particular. The identification, stratification, measurement and evaluation of stress and its associated corrective strategies, particularly for occupational stress, are important topics to address in the near future to establish the basis for considering stress as an important risk factor in occupational health.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Stress; Occupation; Gastric alterations; Gastrointestinal tract diseases; Health risks

**Core tip:** In workers, the combination of personality patterns (anxiety/depression), stress and negative emotions contribute to gastrointestinal tract (GIT) alterations. In particular, jobs that produce privation, fatigue, chronic mental anxiety and a long past history of tension, frustration, resentment, psychological disturbance or emotional conflict have been shown to produce gastric ulcers. Irritable bowel syndrome and functional dyspepsia also have significant co-morbidity with mood alterations. Workers with unipolar depression have been shown to be more prone to present irritable bowel syndrome-like symptoms. Moreover, three systems are known to participate in the GIT alterations of workers: sympathetic autonomic nervous system, the hypothalamic-pituitary-adrenal axis and genetic factors.

Huerta-Franco MR, Vargas-Luna M, Tienda P, Delgadillo-Holtfort I, Balleza-Ordaz M, Flores-Hernandez C. Effects of occupational stress on the gastrointestinal tract. *World J Gastrointest Pathophysiol* 2013; 4(4): 108-118 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/108.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.108>

## INTRODUCTION

Stress is a term that is often used by the global population. This term was first described as a “syndrome produced by diverse noxious agents” in the 1930s by Selye<sup>[1]</sup> and was later called General Adaptation Syndrome. Stress refers to the consequences of the failure of a living organism (*i.e.*, human or animal) to respond appropriately to emotional or physical threats, whether actual or imagined<sup>[2]</sup>. Stress can be defined as any threat to an organism’s homeostasis<sup>[2,3]</sup>. The function of the stress response is to maintain homeostasis and may involve both physiological and behavioral adaptations<sup>[2]</sup>. Currently, stress is a condition that affects people daily. Environmental factors, such as work pressures, financial conditions, family situations and social issues, contribute to stress. Factors related to job stress include the need for counseling, lack of leisure time, daily shift work, dissatisfaction with the workplace, work absenteeism due to health problems and insufficient work incentives. All of these situations produce psychological stress that may affect different physiological functions in the gastrointestinal tract (GIT)<sup>[3]</sup>, including gastric secretion, gut motility, mucosal permeability, mucosal barrier function, visceral sensitivity and mucosal blood flow<sup>[4]</sup>. There have been several studies on the effects of psychological stress on the GIT that debate if these effects constitute a physiological response of the body or if they can be considered a pathology<sup>[2-5]</sup>. In relation to occupations, it is difficult to define a psychological stress classification for determining stress levels, exposure duration, exposure limits to stressors, the sensitivity of the worker, *etc.*; therefore, as evidenced in most of the literature, the delimitation of the problem to be addressed is almost forced. In this review article, we consider general occupational stress as assessed using multiple approaches. Additionally, we focus on occupational stress associated with specific GIT problems in any worker group, but particularly in those groups working stressful jobs.

## EFFECT OF PSYCHOLOGICAL STRESS AND EMOTIONS ON THE GASTROINTESTINAL TRACT

The relationship between psychological stress and disease has already been recognized by the ancient Greeks, who hypothesized that moods affect the body. Hippocrates described how psychosomatic disorders produce abnormal physical reactions due to stressful emotions, and Galen supported the idea that emotions and pain are diseases of the soul. However, in 1637, Descartes changed the paradigm by proposing the separation of the thinking mind from the material body<sup>[5]</sup>. Currently, there are an increasing number of reports regarding cognitive and psychological declines related to stress, including occupational stress, in subjects without psychiatric pre-morbidities or major life trauma<sup>[6]</sup>.

The relationship between emotions and gastric mo-

tility has been documented since the nineteenth century and the beginning of the twentieth century by Charles Cabanis and William Beaumont and thereafter by Ivan Pavlov, Walter Canon and Stewart Wolf, who were the pioneers in determining the gastric response after an emotional stimulus in animal models<sup>[7,8]</sup>. Based on these antecedents, researchers and clinicians were curious about the relationship between stress and gastric motility. For example, Muth *et al.*<sup>[9]</sup> reported the case of a fistulated patient who displayed increased gastric motility when he was angry but decreased gastric motility when he was fearful. Nevertheless, in general, the role of occupational stress in gastric motility has not been closely examined. The main limitation, as far as we perceive, occurs because most of the techniques used for GIT evaluations are invasive<sup>[10]</sup>.

The combination of personality patterns and emotional stress also has an important contribution to GIT alterations. In a review by Alp *et al.*<sup>[11]</sup>, studies were mentioned that suggested privation, fatigue and mental anxiety frequently coincided with the presence of gastric ulceration<sup>[12]</sup> and that a psychological disturbance or emotional conflict might be transformed into an organic disease, *e.g.*, a peptic ulcer<sup>[13,14]</sup>. The same paper also mentioned that a significant correlation exists between the onset of peptic ulcer symptoms and domestic upset, financial stress or an extensive past history of tension<sup>[15]</sup>. Furthermore, anxiety, frustration, resentment and fatigue were suggested to be important aggravating factors in the symptomology of peptic ulceration<sup>[16]</sup>. Many of the emotional states previously described by these researchers are related to psychological stress.

In relation to other GIT alterations, such as irritable bowel syndrome (IBS) and functional dyspepsia, a significant co-morbidity reportedly exists between mood alterations (*e.g.*, anxiety and depression) and functional gastrointestinal syndromes<sup>[17]</sup>. However, the exact pathophysiological link between emotions and the gut is not yet well established (see below). A model of an emotional motor system (EMS) that reacts to interoceptive and exteroceptive stress was proposed by Karling *et al.*<sup>[18]</sup>. These investigators found that recurrent unipolar depression patients who were experiencing remission did not have a greater number of IBS-like symptoms than the controls, indicating that GIT dysfunctions may resolve when depression is treated to remission. Apparently, there is a relationship between mood alterations (*e.g.*, anxiety and depression) and IBS-like symptoms in patients with unipolar depression, in patients with IBS and in a sample of the normal population. In addition, the investigators suggested that, during the regulation of the emotional-motor system, there is a participation of the following three systems (the interrelationships of which are presented schematically in Figure 1): (1) the sympathetic autonomic nervous system (ANS), which explains symptoms that occur when patients change their body position; (2) the hypothalamic-pituitary-adrenal (HPA) axis, which explains the symptoms of diarrhea and early satiety by the stimulation of

Corticotrophin-releasing hormone (CRH) receptors and (3) the *val158met* COMT polymorphism (single nucleotide polymorphism in the *COMT* gene that encodes Catechol-O-methyl-transferase), which is associated with IBS-like symptoms. IBS patients tend to have a lower frequency of the heterozygous val/met genotype, and this genotype may be protective against IBS/IBS-like symptoms. Moreover, a higher frequency of the val/val genotype is associated with diarrhea symptoms<sup>[19]</sup>.

## JOB, OCCUPATION AND PSYCHOLOGICAL STRESS

A job is defined in the OECD Glossary of statistical terms<sup>[20]</sup> as a set of tasks and duties executed, or meant to be executed, by one person. Based on this description, an occupation is defined as a set of jobs whose main tasks and duties are characterized by a high degree of similarity. An occupational classification is a tool for organizing all jobs in an establishment, an industry or a country into a clearly defined set of groups according to the tasks and duties undertaken in the job. The occupation classification is generally not based on health risk factors. However, the concept of occupational health has been well-defined since 1950, when the International Labour Organization (ILO) and the World Health Organization (WHO) created a common definition through the ILO/WHO Committee on Occupational Health. The definition reads as follows: “Occupational health should aim at the promotion and maintenance of the highest degree of physical, mental and social well-being of workers in all occupations...the placing and maintenance of the worker in an occupational environment adapted to his (her) physiological and psychological capabilities...”<sup>[21]</sup>. In this definition, the mental well-being and the psychological capabilities are mentioned; moreover, the main focus of occupational health describes the promotion of a “positive social climate”. The term health, in relation to work, indicates not merely the absence of disease or infirmity but also includes the physical and mental elements affecting health, which are directly related to safety and hygiene at work<sup>[22]</sup>.

In 2001, the ILO published the ILO-OSH 2001 document titled “Guidelines on Occupational Safety and Health Management Systems” to assist organizations introducing OSH management systems<sup>[23]</sup>. Usually, occupational health hazards are considered as physical, chemical, biological and/or ergonomic factors; almost everywhere, psychological factors, such as stress<sup>[24]</sup>, are not even mentioned. We consider that one of the main reasons for minimizing psycho-social factors is because the principles of occupational hygiene are the “recognition and/or identification” of occupational health hazards, the “measurement” of the level or concentration of such factors, the “evaluation” of the likelihood and severity of harm and the “control strategies” available to reduce or eliminate risks. Usually, recognition and identification implies the obvious correlation between

cause and effect, with the effect being associated with physical illness or injury. In our case, psychological stress has been recognized and correlated with physical effects in many studies<sup>[25]</sup>. Furthermore, the control strategies (although a subjective topic) appear to be well known by professionals and therefore are well-established<sup>[26]</sup>. The measurement of levels or concentrations and the evaluation of the likelihood and severity of the stress factors are the most complicated factors to be studied quantitatively, and we consider these factors to be poorly established aspects of the problem.

Despite the discussion above, stress, particularly occupational stress, has become one of the most serious health issues in the modern world<sup>[27]</sup>. The concept of occupational stress can be observed as a natural extension of the classical concept of stress introduced by Selye<sup>[1]</sup> to a specific form of human activity, namely work. Steers<sup>[28]</sup> indicated that occupational stress has become an important topic study of organizational behavior for several reasons: (1) stress has harmful psychological and physiological effects on employees; (2) stress is a major cause of employee turnover and absenteeism; (3) stress experienced by one employee can affect the safety of other employees and (4) by controlling dysfunctional stress, individuals and organizations can be managed more effectively. More recently, Beheshtifar and Modaber<sup>[29]</sup> described five types of sources of occupational stress: (1) causes intrinsic to the job, including factors such as poor physical working conditions, work overload or time pressures; (2) the role in the organization, including role ambiguity and role conflict; (3) career development, including lack of job security and under/over promotion; (4) relationships at work, including poor relationships with the boss or colleagues, an extreme component of which is mobbing in the workplace; and (5) the organizational structure and climate, including the experience of having little involvement in decision-making and office politics.

The large diversity of stress factors, the complexity of labour activities and the large number of worker health-status levels make it very difficult to complete an exhaustive review of the relationships among GIT alterations, stress and occupational activity. Nevertheless, many delimited investigations have been performed around these topics, mainly for specific activities, delimited work places and/or specific groups of people. For example, in a controlled study of lorry drivers, de Croon *et al.*<sup>[30]</sup> investigated the result of specific job demands on job stress (fatigue and job dissatisfaction), thereby identifying the risk factors associated with the psychosocial work environment to begin building an effective stress-reducing strategy. Another study of telemarketers directly addressed occupational stress by reporting the prevalence of stressors affecting job performance<sup>[31]</sup>. In a non-specific occupational study of migrants in Spain, Ronda *et al.*<sup>[32]</sup> reported the occupational health-risk differences between local and foreign-born workers. These investigators listed what they called psychosocial factors, of which most were identified with stressful work condi-

tions. Based on the self-reported exposure, this study revealed a larger difference in females in non-service jobs; although no specific attention was paid to psychosocial factors, the prevalence of exposure to occupational risk factors appeared to be, on average, higher for migrants. The same research group in Spain searched for risk factors during pregnancy using self-reports, which are the predominant method to report psychosocial risks. The results of these investigations revealed that the prevalence of the psychosocial risks was, on average, higher than any other chemical, physical or biological factors<sup>[33]</sup>.

Several researchers studied the psychological stress in a specific pathology, for example, in diabetes. Golmohammadi *et al.*<sup>[34]</sup> investigated the occupational stress in diabetic workers from Iran and found that the type of occupation was not an important factor in psychological stress, although a difference was evident in the patients compared with the control group. The researchers concluded that occupational stress may be a risk factor in the development of diabetes. Specific studies addressing jobs identified as stressful have been conducted. Examples of this type of work are the study of stress-factor risks in nurses from England<sup>[35]</sup> or in psychiatric nurses (*i.e.*, a job with more exposure to psychological risks) in Japan<sup>[36]</sup>. Other stressful jobs are those related to the army and security services. Martins *et al.*<sup>[37]</sup> studied the military hierarchy in peace times in the Brazilian Army, finding correlations with common mental disorders. Berg *et al.*<sup>[38]</sup> studied security service workers, focusing on personality, anxiety and depression. Another type of occupation considered stressful is the mental health profession<sup>[39,40]</sup>. These professionals, similar to other workers exposed to long-term occupational stress, often experience the stage known as burnout. According to Selye's definition, if stress is associated with adaptation, then occupational stress should be identified conceptually with a temporary adaptation to work that is associated with psychological and physical symptoms. The long-term process of adaptation to certain jobs yields chronic physical and psychological symptoms. The final stage in a breakdown during this adaptation is known as burnout and is caused by prolonged occupational stress<sup>[39,40]</sup>. In 1982, Belcastro<sup>[41]</sup> stated that several somatic complaints have been suggested to be associated with burnout, including gastrointestinal disturbances, nausea and loss of weight, which are some of the most common symptoms. The author suggested that specific illnesses appear to be associated with burnout, including colitis, gastrointestinal problems and drug and alcohol addiction, among others. Developed countries face different challenges than do non-developed countries, specifically, economic<sup>[42]</sup> and cultural differences, which have been considered in stress research<sup>[43]</sup>. Nonetheless, it is difficult to describe the occupational psychological-stress classifications, levels, exposure durations, exposure limits, sensitivity, *etc.* Therefore, in most of the literature, the delimitation of the problem to be addressed is almost forced. In this review article, we focus on occupational stress in gen-

eral, assessed in any manner. Additionally, we focus on specific GIT problems in any worker group, but mainly in those groups dedicated to some jobs well-identified as stressful.

## TYPE OF JOB, STRESS AND GIT ALTERATIONS

Currently, gastric ulcers are identified as an extremely common chronic disease in working-age adults. The first description of an association between stress and peptic ulcer disease was in men with supervisory jobs; these individuals had a higher ulcer prevalence than executives or artisans. Cobb and Rose<sup>[44]</sup> found that air traffic controllers, particularly those individuals with higher stress levels in their workplace, were almost twice as likely to have ulcers than the civilian copilots. Hui *et al.*<sup>[45]</sup> noted that the numbers of positive and negative life events were similar in both subjects with dyspepsia and control subjects, but the former had a higher negative perception of major life events and daily stresses. Police officers' work-stress reactions have been classified as physiological, emotional and behavioral reactions<sup>[46]</sup>. Physiological reactions have been termed as having a higher-than-normal probability of death from certain illnesses; after cardiovascular problems, stomach problems are the most frequent. Changing work shifts has also been associated with changes in the digestive system, circadian rhythm and other bodily reactions. Angolla<sup>[46]</sup> studied 229 police officers (163 males and 66 females) who answered a questionnaire consisting of five parts: demographics, external and internal work environments, coping mechanisms and symptoms. This investigator found that police work is highly stressful, and the highest rated symptoms were as follows: feeling a lack of energy, loss of personal enjoyment, increased appetite, feeling depressed, trouble concentrating, feeling restless, nervousness and indigestion. Satija *et al.*<sup>[47]</sup> evaluated 150 professional workers (100 males and 50 females) who self-completed the Emotional Intelligence and Occupational Stress Scale. The authors' findings demonstrated a negative correlation between emotional intelligence and occupational stress; those professionals with a high score in overall emotional intelligence suffered less stress. Shigemi *et al.*<sup>[48]</sup> evaluated 585 employees (296 males and 289 females), all of whom were working at a middle-sized company in Japan. A self-administered questionnaire about smoking habits and perceived job stress was administered, and the patients were followed for two years. In addition, previous or current gastric or duodenal ulcers were evaluated. The researchers found 32 incidences of peptic ulcers over the two years, and the risk ratio (RR) was 2.13 (95%CI: 1.09-4.16) between job stress and peptic ulcers. Susheela *et al.*<sup>[49]</sup> evaluated 462 smelter workers, 60 supervisors working in the smelter unit and 62 non-smelter workers (control group). The participants' state of health and gastrointestinal complaints were recorded and included the following symptoms: nausea/loss of ap-

petite, gas formation, pain in the stomach, constipation, diarrhea (intermittent) and headaches. The researchers found that the total number of complaints reported by the study groups was significantly higher than in the control group. The prevalence of gastrointestinal complaints in the smelter workers was significantly higher ( $P < 0.001$ ) than in the non-smelter workers (control group). In 2006, Nakadaira *et al.*<sup>[50]</sup> investigated the effects of tanshin funin (*i.e.*, working far from one's hometown and therefore far from one's family) on the health of married male workers. A prospective study using the pair-matched method was performed in 129 married male tanshin funin workers who were 40-50 years of age. Matched workers living with their families also participated. These researchers demonstrated that fewer tanshin funin workers ate breakfast every day. Moreover, these workers more frequently suffered from stress due to daily chores and from stress-related health problems, namely, headache and gastric/duodenal ulcers (21% and 2.4%, respectively). The levels of gamma-glutamyl-transpeptidase in workers reluctant to work under tanshin funin conditions and in workers who spent less than two years in tanshin funin conditions increased significantly, although the corresponding levels in the matched regular workers did not exhibit significant changes. The investigators concluded that abrupt changes in lifestyle and elevated mental stress were thus important effects of tanshin funin.

Although jobs and occupations are considered a risk factor for stress and morbidity for gastric and duodenal ulcers<sup>[9,11,48]</sup>, there are studies that report discrepancies. For example, Westerling *et al.*<sup>[51]</sup> evaluated the socioeconomic differences in avoidable mortality in a Swedish population from 1986 to 1990. Using the population of 21- to 64-year-old individuals, the researchers performed analyses for different socioeconomic groups [blue-collar workers (BCWs), white-collar workers (WCWs) and the self-employed] and for individuals outside the labor market. The researchers demonstrated that the largest differences were found in ulcers of the stomach and duodenum, in addition to other symptoms. For these causes of death, the risk of dying was between 3.1 and 7.5 times greater in the non-working population than in the workforce. The differences in avoidable mortality between BCWs and WCWs and the self-employed were much smaller. However, the death rate for ulcers of the stomach and duodenum in BCWs was 2.8 times higher than for other work categories. The GIT and mortality problems reported by Westerling *et al.*<sup>[51]</sup> used standardized mortality ratios for the occupied population. The ratios for malignant neoplasms of the large intestine, except for the rectum, were 95 in BCWs and 104 in WCWs and for malignant neoplasms in the rectum and rectum-sigmoid junction were 103 and 100 for BCWs and WCWs, respectively. However, for gastric and duodenal ulcers, the investigators reported ratios of 163 and 59 for these BCWs and WCWs, respectively. Moreover, the causes for mortality reported for abdominal hernia, cholelithiasis

and cholecystitis were 127 and 86 for BCWs and WCWs, respectively. However, the researchers suggested that, as in most other studies, the follow-up period was short and that the exposure data from earlier censuses would be advantageous.

In Table 1, we summarize the international literature regarding the GIT disorders most frequently reported by workers experiencing job-related psychological stress and other alterations in their affective states. In a cross-sectional study with a population of 2237 subjects from San Marino, Italy, Gasbarrini *et al.*<sup>[52]</sup> demonstrated that the prevalence of *Helicobacter pylori* (*H. pylori*) was 51%; this prevalence increased with age from 23% (20-29 years) to 68% (> 70 years) and was higher among manual workers. In San Marino, there was a higher incidence of clinically relevant gastroduodenal diseases, such as peptic ulcer and gastric cancer (25 of 10000 and 8 of 10000 in 1990, respectively). With respect to Italy and other European countries, Gasbarrini *et al.*<sup>[52]</sup> demonstrated that *H. pylori* infections tended to be more frequent among BCWs ( $P < 0.001$ ), especially those doing manual work (miners 78%, road sweepers 65%, plumbers/painters 61%, housekeepers 60% and cooks 58%) compared with WCWs (physicians 15%, clerks 21%, secretaries 37%, nurses 38%, general managers and lawyers 38%, teachers 39% and shop workers 50%). A high prevalence of infection was also noted among social workers (74%). The researchers concluded that social workers with a high educational standard had a higher rate of seropositivity to *H. pylori* (74%) than did subjects with a similar socioeconomic status but a different type of job, thereby emphasizing the relevance of direct person-to-person spread of the infection. This result was also recently demonstrated in nurses and in cohabiting children. These findings suggest that poor hygienic standards and a low socioeconomic status (which frequently reflects the former) are important factors for acquiring *H. pylori* during the first years of life, thereby confirming previous findings regarding the differences between developed and developing countries and the importance of overcrowding and close person-to-person contact during childhood.

In 2009, Lin *et al.*<sup>[53]</sup> investigated 289 call-center workers (mean age of 33.6 years) to investigate how these workers perceived their job stress and health status and the relationships among inbound (incoming calls) versus outbound (outgoing calls) workers in a Taiwanese bank. Data were obtained on individual factors, health complaints, perceived job stress levels and major job stressors (using the 22-item Job Content Questionnaire, C-JCQ). For inbound services, operators handled approximately 120 to 150 calls during each 8 h per day. Outbound operators were primarily responsible for sales and handled approximately 120 calls daily. The subjects completed the self-administered questionnaires during their leisure time (taking between 15 and 20 min). The results demonstrated that 33.5% of outbound service-call center workers and 27.1% of inbound service-call

**Table 1 Summary of the international literature on the gastrointestinal tract disorders most frequently reported by workers experiencing job-related psychological stress and other alterations in their affective states**

GIT alteration	Ref.	Study/procedure	Conclusions
Generalized GIT disturbances/IBS	Konturek <i>et al</i> <sup>[4]</sup>	Impact of stress on the GIT. The study addresses the role of stress in the pathophysiology of the most common GIT diseases.	The exposure to stress is the major risk factor in the pathogenesis of various GIT diseases.
	Bhatia <i>et al</i> <sup>[5]</sup>	Association between stress and various GIT pathologies.	The mind directly influences the gut. The enteric nervous system is connected bidirectionally to the brain by the parasympathetic and sympathetic pathways, forming the brain-gut axis.
	Karling <i>et al</i> <sup>[18]</sup>	Pre- and post-dexamethasone morning serum cortisol levels were analyzed in 124 subjects with symptoms of IBS.	There is a relationship between mood alterations (anxiety/depression) and IBS-like symptoms in patients with unipolar depression, in patients with IBS and in a sample of the normal population.
	Karling <i>et al</i> <sup>[19]</sup>	In total, 867 subjects representative of the general population and 70 patients with IBS were genotyped for the val158met polymorphism. The IBS patients completed the Hospital Anxiety and Depression Scale questionnaire.	There is an association between the val/val genotype of the val158met COMT gene and IBS and with the specific IBS-related bowel pattern in IBS patients.
	Belcastro <i>et al</i> <sup>[41]</sup>	Estimation of the relationship between teachers' somatic complaints and illnesses and their self-reported job-related stresses. Stress group: teachers	Several somatic complaints have been suggested to be associated with burnout.
	Hui <i>et al</i> <sup>[45]</sup>	Perception of life events and the role of daily "hassles" (stressful events) in 33 dyspeptic patients vs 33 controls of comparable sex, age and social class.	Patients with non-ulcer dyspepsia have a higher negative perception of major life events than controls. Psychological factors may play a role in the pathogenesis of non-ulcer dyspepsia.
	Angolla <i>et al</i> <sup>[46]</sup>	Empirical study of police-work stress, symptoms and coping strategies among police service workers in Botswana measured by a questionnaire. Sample size <i>n</i> = 229 (163 males and 66 females). Stress group: the Botswana Police Service.	Police duties are highly stressful. The highest rated symptoms were as follows: feeling a lack of energy, loss of personal enjoyment, increased appetite, feeling depressed, trouble concentrating, feeling restless, nervousness and indigestion.
Ulcers	Alp <i>et al</i> <sup>[11]</sup>	Comparative study using a neuroticism-scale questionnaire administered to 181 patients with previously diagnosed gastric ulcers and 181 controls without any previous history of gastric ulcers.	People with a past history of chronic gastric ulcers have an increased incidence of domestic and financial stress compared with age- and sex-matched individuals with no previous history of gastric ulcers.
	Cobb <i>et al</i> <sup>[44]</sup>	Review of aeromedical certification examinations of 4325 traffic controllers and 8435 second-class aviators. Stress group: air traffic controllers.	Air traffic controllers were almost twice as likely to have stomach ulcers as civilian copilots.
	Shigemi <i>et al</i> <sup>[48]</sup>	Two-year study to examine the role of perceived job stress on the relationship between smoking and peptic ulcers.	These results suggest that specific and perceived job stress is an effect modifier in the relationship between the history of peptic ulcers and smoking.
	Nakadaira <i>et al</i> <sup>[50]</sup>	Effects of working far from family on the health of 129 married male workers (40-50 yr of age) compared with the control group.	The tanshin funin workers had higher rates of missing breakfast, stress due to daily chores and stress-related health problems ( <i>e.g.</i> , headache, gastric/duodenal ulcers and common colds/bronchitis).
	Westerling <i>et al</i> <sup>[51]</sup>	1985 study of the Swedish population (21-64 yr of age). Analyses of standardized mortality ratios (avoidable mortality) of blue-collar workers, white-collar workers, self-employed workers, and individuals outside the labor market. Stress group: Unemployed individuals	The death rates for the non-workers were higher than for the workers. The largest differences were found for stomach and duodenal ulcers.
	Lin <i>et al</i> <sup>[53]</sup>	289 call center workers in Taiwan, 19 to 54 yr of age. Health complaints, perceived level of job stress and major job stressors were considered. Stress group: call center workers.	Workers who perceived higher job stress had significantly increased risks of multiple health problems, including hoarse or sore throat, irritable stomach and peptic ulcers.
	Gastric motility alterations	Mawdsley <i>et al</i> <sup>[2]</sup>	Review of recent advances in the understanding of the pathogenic role of psychological stress in IBD, with an emphasis on the necessity of investigating the therapeutic potential of stress reduction.
Huerta-Franco <i>et al</i> <sup>[3]</sup>		Bioimpedance technique. In this study, 57 healthy women (40-60 yr of age) were analyzed.	Assessment of the changes in gastric motility induced by acute psychological stress.
Mai <i>et al</i> <sup>[7]</sup>		Description or tracking of 238 experiments conducted over more than 10 yr on a young man (Beaumont) with digestive disorders.	Emotions can cause bile reflux into the stomach and may delay gastric emptying.
Wolf <i>et al</i> <sup>[8]</sup>		Description of the work of the French physiologist Cabanis.	Inhibitory and excitatory effects of gastric secretory and motor function were described.

Muth <i>et al.</i> <sup>[9]</sup>	Electrogastrograms were recorded, and the inter-beat intervals were obtained from electrocardiographic recordings from 20 subjects during baseline and in response to a shock avoidance task (shock stimulus) and forehead cooling (dive stimulus).	Acute stress can evoke arousal and dysrhythmic gastric myoelectrical activity. These acute changes, which occur in healthy individuals, may provide insight into functional gastrointestinal disorders.
-----------------------------------	---	---

GIT: Gastrointestinal tract; IBD: Inflammatory bowel disease; IBS: Irritable bowel syndrome; COMT: catechol-O-methyltransferase.

center workers were classified as suffering from high stress, which was considerably higher than figures from a wider survey of the working population in Taiwan (7.6%). The researchers demonstrated a relationship between the perceived job stress and health complaints, indicating that workers who perceived a higher job stress had a significantly increased risk of multiple health problems (OR ranging from 2.13 to 8.24), including an irritable stomach and peptic ulcers [42% and 57% for inbound and outbound operators, respectively ( $P < 0.05$ )]. For example, the OR of irritable stomach and peptic ulcers when the stress was moderate (*i.e.*, sometimes feeling extremely stressed at work) was 3.03 (95%CI: 1.40-6.55) and when the stress was high (*i.e.*, often or always feeling extremely stressed at work) was 8.24 (95%CI: 3.56-19.09). The researchers concluded that there is an association between the perceived job stress and health complaints, as workers who perceived a high level of job stress had significantly increased risks of irritable stomach and peptic ulcers.

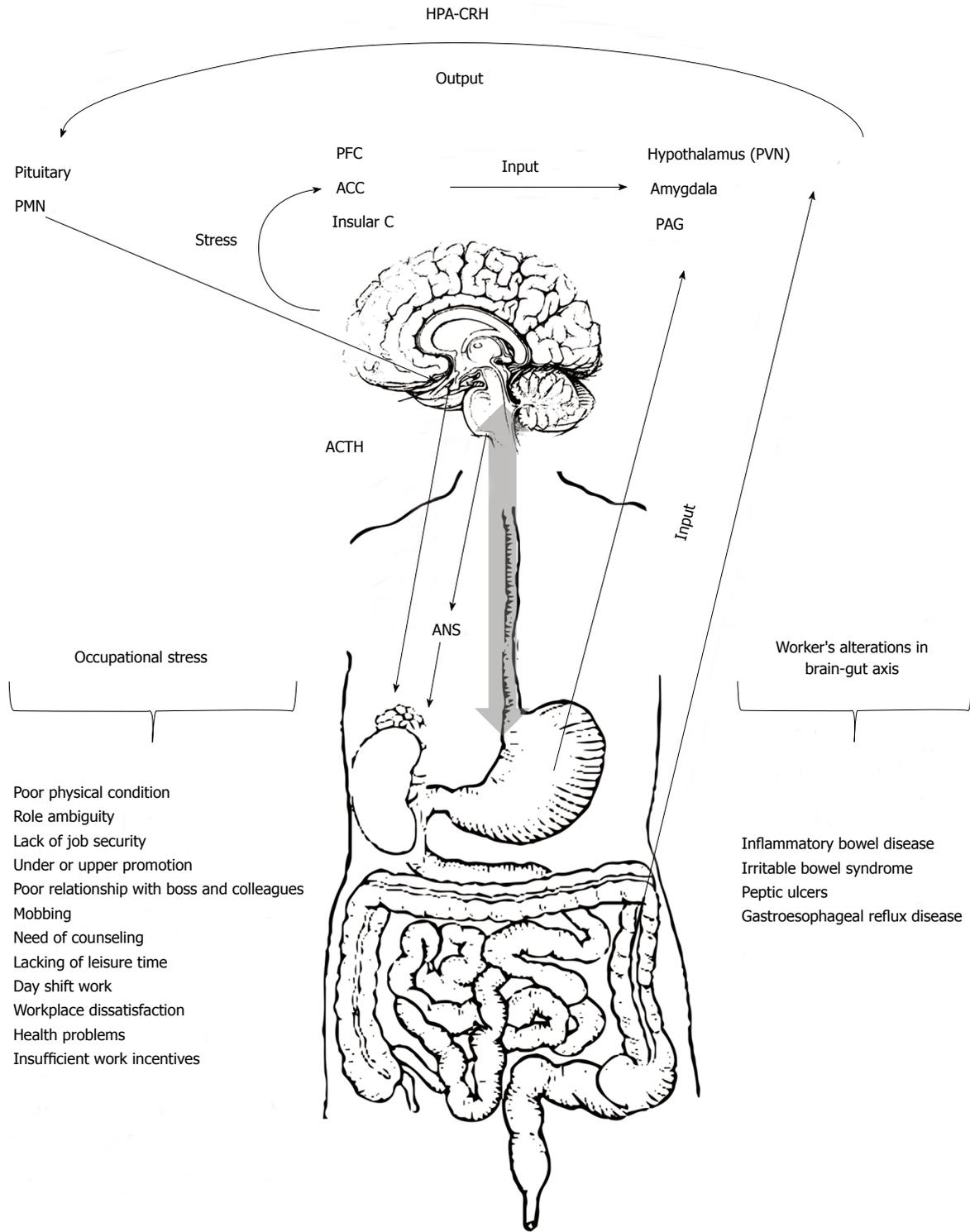
In 2011, Nabavizadeh<sup>[54]</sup> demonstrated that physical and psychological stress increase gastric acid and pepsin secretions possibly by raising the gastric tissue nitric oxide level. In return, the increased gastric acid and pepsin secretions cause necrotic and inflammatory changes in the gastric and duodenal tissue.

## STRESS IN THE PATHOPHYSIOLOGY OF GASTROINTESTINAL ALTERATIONS

Currently, many adults die from diseases caused by the relationship between stress, moods and vital organs; among these diseases, GIT has become a major clinical problem. Stress is an acute threat to the homeostasis of an organism, either physically or psychologically. A number of studies have shown that stress can delay gastric emptying, impair gastro-duodenal motility<sup>[3]</sup>, modify gastric secretions<sup>[55]</sup> and pancreatic output and alter intestinal transit and colonic motility. Owing to its considerable effects on physiological and pathophysiological processes of gastrointestinal (GI) motility, stress is thought to play an important role in the development, maintenance and exacerbation of symptoms related to functional GI disorders. To analyze the effects of occupational stress on GIT function, it is important to have an understanding of the physiology of GIT motility and emptying. Similar to almost every other system, the physiological processes that occur in the GIT are wide-ranging; the major function of the GIT includes swallowing, motility, emptying (of every section), assimilation and elimination. Motility

enables swallowing, transit, emptying and elimination, and all these functions are essential for proper assimilation<sup>[56]</sup>. Beginning with swallowing and ending with elimination, motility is required for GIT function<sup>[57,58]</sup>. Two variables related to GIT motility are particularly important: (1) peristalsis, which is a function of the frequency and magnitude of the gastric contractions that are generated by the pacemaker area and (2) gastric emptying, which is a measure of the average time the stomach takes to empty half of its luminal content. The ANS regulates GIT motility, controlling peristaltic activity through the myenteric system<sup>[59,60]</sup> (Figure 1). In fact, alterations in GIT motility are frequently viewed as signs of neuropathy of the myenteric plexus or other pathologies of neuropathic origin. Abnormal gastric emptying is considered a clinical marker for a gastric or intestinal motility disorder<sup>[61,62]</sup>. Quigley and other researchers have found a relationship between stress and delayed gastric emptying or other motor disturbances<sup>[63-65]</sup>. This factor is understood by analyzing how the human body reacts defensively when threatened by the environment and when attempting to achieve both physical and psychological balance. However, activation of these adaptive or allostatic systems can become maladaptive because of frequent, chronic or excessive stress and can cause a predisposition to disease<sup>[5]</sup>. This explanation leads to the concept of brain-gut interaction described by Mawdsley and Rampton<sup>[2]</sup>. These authors mentioned that, to maintain homeostasis, a living organism must constantly adapt to environmental alterations at a molecular, cellular, physiological and behavioral level. As presented in Figure 1, these investigators hypothesized that exposure to psychological stress causes alterations of the brain-gut interactions (brain-gut axis), ultimately leading to the development of a broad array of GIT disorders, including inflammatory bowel disease, irritable bowel syndrome, other functional GIT diseases, food antigen-related adverse responses, peptic ulcers and gastro-esophageal reflux disease (GERD)<sup>[16,19]</sup>. For instance, IBS is presumed to be a disorder of the brain-gut link associated with an exaggerated response to stress<sup>[66]</sup>.

More recently, relative importance has been ascribed to the hypothesis that emotional and environmental states in females play an important role in the genesis of IBS<sup>[2]</sup>. This hypothesis has been proved by demonstrating that, worldwide, women present the highest prevalence of physical and psychological symptoms compared with males<sup>[67]</sup>. Men may be more apt to experience stress due to unfamiliar house chores than women, and women are more likely to experience occupational stress than



**Figure 1 Representation of the hypothetical mechanism by which occupational stress produces gastrointestinal tract alterations in workers.** Stress during job development (see occupational stress) generates a response of the network integrated by the hypothalamus (paraventricular nucleus), amygdala and periaqueductal grey. These brain regions receive input from visceral and somatic afferents and from the medial prefrontal cortex (PFC) and the anterior cingulate (ACC) and insular cortices (Insular C). In turn, output from this integrated network to the pituitary and ponto-medullary nuclei (PMN) mediates the neuroendocrine and autonomic responses in the body. The final output of this central stress circuitry is called the emotional motor system and includes the autonomic neurotransmitters norepinephrine and epinephrine and neuroendocrine (hypothalamus-pituitary-adrenal axis, HPA) and pain modulatory systems. PVN: Paraventricular nucleus; PAG: Periaqueductal grey; GIT: Gastrointestinal tract; ACTH: Adrenocorticotropic hormone; CRH: Corticotrophin-releasing hormone; ANS: Autonomic nervous system.

men<sup>[50]</sup>. The emotions and stress experienced by workers (Figure 1) play an important role in exaggerated gut responses. Stress affects the relationship between the brain

and gut, leading these systems to act defensively to real or imaginary threats.

In Figure 1, we present the pathophysiological mech-

anism by which stress has been proposed to produce GIT alterations in workers. The workers' responses to stress are generated by a network comprising integrated brain structures, particularly sub-regions of the hypothalamus (paraventricular nucleus), amygdala and periaqueductal grey. These brain regions receive input from visceral and somatic afferents and from cortical regions, including the medial prefrontal cortex and sub-regions of the anterior cingulate and insular cortices. In turn, output from this integrated network to the pituitary and ponto-medullary nuclei mediates the neuroendocrine and autonomic responses in the body<sup>[4,18,19]</sup>. The final output of this central stress circuitry is called the emotional-motorsystem and includes the autonomic neurotransmitters norepinephrine and epinephrine, the neuroendocrine HPA axis and the pain modulatory systems. This circuit is under feedback control by serotonergic neurons from the raphe nuclei and noradrenergic neurons from the locus coeruleus<sup>[5]</sup>.

The neuroendocrine response to stress is mediated by corticotropin-releasing hormone (CRH). In the brain-gut-axis, CRH is considered a major mediator of the stress response. Particularly, the stress-related activation of CRH receptors has been reported to produce alterations in GIT function. Physical and psychological stress delays gastric emptying, accelerates colonic transit and evokes colonic motility in rats. Accelerated colonic motor function can be produced by the central or peripheral administration of CRH and is blocked by treatment with a variety of CRH antagonists. In a clinical trial, Sagami *et al*<sup>[68]</sup> administered a non-selective CRH antagonist (10 ug/kg of  $\alpha$ hCRH) to 10 IBS patients and 10 healthy controls. The researchers demonstrated that the peripheral administration of  $\alpha$ hCRH improved GIT motility, visceral perception and negative moods in response to gut stimulation, without affecting the HPA axis in IBS patients. This response was significantly suppressed in IBS patients but not in controls after the administration of  $\alpha$ hCRH<sup>[68]</sup>. IBS is considered a disorder of the brain-gut-link. Psychological stress induces colonic segmental contractions, which are exaggerated in IBS patients. Similarly, the peripheral administration of CRH affects colonic motility, induces abdominal symptoms and stimulates adrenocorticotropic hormone (ACTH) secretion, all of which are also exaggerated in IBS patients<sup>[69]</sup>. Two CRH receptor subtypes, R1 and R2, have been suggested to mediate increased colonic motor activity and slowed gastric emptying, respectively, in response to stress<sup>[5]</sup>.

The genesis of gastric ulcers by stress was demonstrated in the study of Saxena *et al*<sup>[70]</sup>, who investigated the gastro-protective effect of citalopram (an antidepressant drug) both as a single dose pre-treatment and 14-d repeated pre-treatment for animals exposed to cold restraint stress (CRS). The results revealed that the plasma corticosterone level significantly increased in the stress group compared with the control group. Furthermore, mucosal ulceration, epithelial cell loss and a ruptured gastric mucosal layer at the ulcer site were observed in

the gastric mucosa of rats exposed to CRS. Repeated citalopram pretreatment decreased the CRS-induced enhancement in the corticosterone level. The researchers also demonstrated that citalopram at doses of 5, 10 and 20 mg/kg significantly attenuated the CRS-induced gastric mucosal lesions.

In summary, gastric ulcers are identified as an extremely common chronic disease in working-age adults. In workers, the combination of personality patterns (*e.g.*, anxiety and depression), stress and negative emotions significantly contribute to GIT alterations. Particular jobs that produce privation, fatigue or chronic mental anxiety and a long past history of tension, frustration, resentment, psychological disturbance or emotional conflict lead to gastric ulcers (*e.g.*, in traffic controllers, police officers, smelter workers, tanshin funin workers, health professionals and manual workers). Irritable bowel syndrome and functional dyspepsia also exhibit significant co-morbidities between mood alterations in workers (*i.e.*, anxiety and depression). Workers with unipolar depression were shown to be more prone to present IBS-like symptoms. Moreover, three systems are known to participate in the mechanism of GIT alterations in workers: (1) the sympathetic ANS, (2) the HPA axis and (3) genetic factors.

Subjective evaluations of stress (mainly self-reported) are extremely common in the clinic and in research. However, much work must first be done to quantitatively identify the psychological stress (*i.e.*, the occupational stress level in this case), considering the particularity of each worker (*i.e.*, the general health, the social and physical adaptation capacity and the physical and psychological vulnerability). The unique demands of each occupation require unique profiles of each worker. It is essential to train workers not only in specific skills but also in human and social aspects that include stress control strategies. Although the word "stress" has been included in our everyday language (even in research), the term continues to be a vague concept, even with the clear definition coined in 1936 by Selye.

## REFERENCES

- 1 **Selye H.** A syndrome produced by diverse noxious agents. 1936. *J Neuropsychiatry Clin Neurosci* 1998; **10**: 230-231 [PMID: 9722327]
- 2 **Mawdsley JE, Rampton DS.** Psychological stress in IBD: new insights into pathogenic and therapeutic implications. *Gut* 2005; **54**: 1481-1491 [PMID: 16162953 DOI: 10.1136/gut.2005.064261]
- 3 **Huerta-Franco MR, Vargas-Luna M, Montes-Frausto JB, Morales-Mata I, Ramirez-Padilla L.** Effect of psychological stress on gastric motility assessed by electrical bioimpedance. *World J Gastroenterol* 2012; **18**: 5027-5033 [PMID: 23049210 DOI: 10.3748/wjg.v18.i36.5027]
- 4 **Konturek PC, Brzozowski T, Konturek SJ.** Stress and the gut: pathophysiology, clinical consequences, diagnostic approach and treatment options. *J Physiol Pharmacol* 2011; **62**: 591-599 [PMID: 22314561]
- 5 **Bhatia V, Tandon RK.** Stress and the gastrointestinal tract. *J Gastroenterol Hepatol* 2005; **20**: 332-339 [PMID: 15740474 DOI: 10.1111/j.1440-1746.2004.03508.x]

- 6 **Blix E**, Perski A, Berglund H, Savic I. Long-term occupational stress is associated with regional reductions in brain tissue volumes. *PLoS One* 2013; **8**: e64065 [PMID: 23776438 DOI: 10.1371/journal.pone.0064065]
- 7 **Mai FM**. Beaumont's contribution to gastric psychophysiology: a reappraisal. *Can J Psychiatry* 1988; **33**: 650-653 [PMID: 3058293]
- 8 **Wolf S**. The stomach's link to the brain. *Fed Proc* 1985; **44**: 2889-2893 [PMID: 2865172]
- 9 **Muth ER**, Koch KL, Stern RM, Thayer JF. Effect of autonomic nervous system manipulations on gastric myoelectrical activity and emotional responses in healthy human subjects. *Psychosom Med* 1999; **61**: 297-303 [PMID: 10367609]
- 10 **Huerta-Franco MR**, Vargas-Luna M, Montes-Frausto JB, Flores-Hernández C, Morales-Mata I. Electrical bioimpedance and other techniques for gastric emptying and motility evaluation. *World J Gastrointest Pathophysiol* 2012; **3**: 10-18 [PMID: 22368782 DOI: 10.4291/wjgp.v3.i1.10]
- 11 **Alp MH**, Court JH, Grant AK. Personality pattern and emotional stress in the genesis of gastric ulcer. *Gut* 1970; **11**: 773-777 [PMID: 5473609 DOI: 10.1136/gut.11.9.773]
- 12 **Brinton W**. Lectures on the Diseases of the Stomach. 2nd ed. Philadelphia: Lea & Blanchard, 1865
- 13 **Alexander F**. Psychosomatic medicine: its principles and applications. New York: Norton, 1950
- 14 **Højer-Pedersen W**. On the significance of psychic factors in the development of peptic ulcer; a comparative personality investigation in male duodenal ulcer-patients and controls. *Acta Psychiatr Neurol Scand Suppl* 1958; **119**: 1-232 [PMID:13532963]
- 15 **Davies DT**, Wilson ATM. Observations of the life history of chronic peptic ulcer. *The Lancet* 1937; **230**: 1353-1360 [DOI: 10.1016/S0140-6736(00)88967-9]
- 16 **Jones FA**. Clinical and social problems of peptic ulcer. *Br Med J* 1957; **1**: 719-723; contd [PMID: 13404283 DOI: 10.1136/bmj.1.5021.719]
- 17 **O'Malley D**, Quigley EM, Dinan TG, Cryan JF. Do interactions between stress and immune responses lead to symptom exacerbations in irritable bowel syndrome? *Brain Behav Immun* 2011; **25**: 1333-1341 [PMID: 21536124 DOI: 10.1016/j.bbi.2011.04.009]
- 18 **Karling P**, Norrback KF, Adolfsson R, Danielsson A. Gastrointestinal symptoms are associated with hypothalamic-pituitary-adrenal axis suppression in healthy individuals. *Scand J Gastroenterol* 2007; **42**: 1294-1301 [PMID: 17852841 DOI: 10.1080/00365520701395945]
- 19 **Karling P**, Danielsson Å, Wikgren M, Söderström I, Del-Favero J, Adolfsson R, Norrback KF. The relationship between the val158met catechol-O-methyltransferase (COMT) polymorphism and irritable bowel syndrome. *PLoS One* 2011; **6**: e18035 [PMID: 21437260 DOI: 10.1371/journal.pone.0018035]
- 20 **International Labour Office**. Structure, group definitions and correspondence tables. In: International standard classification of occupations: ISCO-08. Geneva: International Labour Office, 2012: 1-420
- 21 **Lee PCB**. Going beyond career plateau: Using professional plateau to account for work outcomes. *JMD* 2003; **22**: 538-551 [DOI: 10.1108/02621710310478503]
- 22 **International Labour Organization**. Occupational safety and health convention: C155. 1981. Cited 2013-02-15. Available from: URL: [http://www.ilo.org/dyn/normlex/en/f?p=1000:12100:0::NO::P12100\\_INSTRUMENT\\_ID:312300#A1](http://www.ilo.org/dyn/normlex/en/f?p=1000:12100:0::NO::P12100_INSTRUMENT_ID:312300#A1)
- 23 **International Labour Office**. Guidelines on occupational safety and health management systems: ILO-OSH 2001. Geneva: International Labour Office, 2001
- 24 **Huang D**, Zhang J, Liu M. Application of a health risk classification method to assessing occupational hazard in China. In: Proceedings of the 3rd International Conference on Bioinformatics and Biomedical Engineering; 2009 Jun 11-13; Beijing: ICBBE, 2009: 1-5
- 25 **Rostamkhani F**, Zardooz H, Zahediasl S, Farrokhi B. Comparison of the effects of acute and chronic psychological stress on metabolic features in rats. *J Zhejiang Univ Sci B* 2012; **13**: 904-912 [PMID: 23125083 DOI: 10.1631/jzus.B1100383]
- 26 **Pereira MA**, Barbosa MA. Teaching strategies for coping with stress--the perceptions of medical students. *BMC Med Educ* 2013; **13**: 50 [PMID: 23565944 DOI: 10.1186/1472-6920-13-50]
- 27 **Lee PCB**. Going beyond career plateau: using professional plateau to account for work outcomes. *J Manag Dev* 2003; **22**: 538-551 [DOI: 10.1108/02621710310478503]
- 28 **Steers RM**. Introduction to organizational behavior. Glenview: Scott Foresman Publishing, 1981
- 29 **Beheshtifar M**, Modaber H. The investigation of relation between occupational stress and career plateau. *Interdisciplinary J Contemp Res Bus* 2013; **4**: 650-660
- 30 **de Croon EM**, Blonk RW, de Zwart BC, Frings-Dresen MH, Broersen JP. Job stress, fatigue, and job dissatisfaction in Dutch lorry drivers: towards an occupation specific model of job demands and control. *Occup Environ Med* 2002; **59**: 356-361 [PMID: 12040108 DOI: 10.1136/oem.59.6.356]
- 31 **Santos AC**, Vianna MI. Prevalence of stress reaction among telemarketers and psychological aspects related to occupation. *Epidemiol Comm Health* 2011; **65**: A416 [DOI: 10.1136/jech.2011.142976o.29]
- 32 **Ronda E**, Agudelo-Suárez AA, García AM, López-Jacob MJ, Ruiz-Frutos C, Benavides FG. Differences in exposure to occupational health risks in Spanish and foreign-born workers in Spain (ITSAL Project). *J Immigr Minor Health* 2013; **15**: 164-171 [PMID: 22739799 DOI: 10.1007/s10903-012-9664-9]
- 33 **García AM**, González-Galarzo MC, Ronda E, Ballester F, Estarlich M, Guxens M, Lertxundia A, Martínez-Argüelles B, Santa Marina L, Tardón A, Vrijheid M. Prevalence of exposure to occupational risks during pregnancy in Spain. *Int J Public Health* 2012; **57**: 817-826 [PMID: 22760548 DOI: 10.1007/s00038-012-0384-7]
- 34 **Golmohammadi R**, Abdulrahman B. Relationship between occupational stress and non-insulin-dependent diabetes in different occupation in Hamadan (West of Iran). *J Med Sci* 2006; **6**: 241-244 [DOI: 10.3923/jms.2006.241.244]
- 35 **Mark G**, Smith AP. Occupational stress, job characteristics, coping, and the mental health of nurses. *Br J Health Psychol* 2012; **17**: 505-521 [PMID: 22107162 DOI: 10.1111/j.2044-8287.2011.02051.x]
- 36 **Leka S**, Hassard J, Yanagida A. Investigating the impact of psychosocial risks and occupational stress on psychiatric hospital nurses' mental well-being in Japan. *J Psychiatr Ment Health Nurs* 2012; **19**: 123-131 [PMID: 22070548 DOI: 10.1111/j.1365-2850.2011.01764.x]
- 37 **Martins LC**, Lopes CS. Military hierarchy, job stress and mental health in peacetime. *Occup Med (Lond)* 2012; **62**: 182-187 [PMID: 22402895 DOI: 10.1093/occmed/kqs006]
- 38 **Berg AM**, Hem E, Lau B, Ekeberg Ø. An exploration of job stress and health in the Norwegian police service: a cross sectional study. *J Occup Med Toxicol* 2006; **1**: 26 [PMID: 17156489 DOI: 10.1186/1745-6673-1-26]
- 39 **Lasalvia A**, Tansella M. Occupational stress and job burn-out in mental health. *Epidemiol Psychiatr Sci* 2011; **20**: 279-285 [PMID: 22201203 DOI: 10.1017/S2045796011000576]
- 40 **Rössler W**. Stress, burnout, and job dissatisfaction in mental health workers. *Eur Arch Psychiatry Clin Neurosci* 2012; **262** Suppl 2: S65-S69 [PMID: 22926058 DOI: 10.1007/s00406-012-0353-4]
- 41 **Belcastro PA**. Burnout and its relationship to teachers' somatic complaints and illnesses. *Psychol Rep* 1982; **50**: 1045-1046 [PMID: 7111569 DOI: 10.2466/pr0.1982.50.3c.1045]
- 42 **Haq Z**, Iqbal Z, Rahman A. Job stress among community health workers: a multi-method study from Pakistan. *Int J Ment Health Syst* 2008; **2**: 15 [PMID: 18954470 DOI: 10.1186/1752-4458-2-15]
- 43 **Taylor SE**, Welch WT, Kim HS, Sherman DK. Cultural differences in the impact of social support on psychological

- and biological stress responses. *Psychol Sci* 2007; **18**: 831-837 [PMID: 17760781 DOI: 10.1111/j.1467-9280.2007.01987.x]
- 44 **Cobb S**, Rose RM. Hypertension, peptic ulcer, and diabetes in air traffic controllers. *JAMA* 1973; **224**: 489-492 [PMID: 4739607 DOI: 10.1001/jama.1973.03220170019004]
- 45 **Hui WM**, Shiu LP, Lam SK. The perception of life events and daily stress in nonulcer dyspepsia. *Am J Gastroenterol* 1991; **86**: 292-296 [PMID: 1998310]
- 46 **Angolla EJ**. Occupational Stress among police officers: the case of Botswana Police Service. *J Bus Manag* 2009; **3**: 25-35 [DOI: 10.3923/rjbm.2009.25.35]
- 47 **Satija S**, Khan W. Emotional intelligence as predictor of occupational Stress among working Professionals. *Prin. LN Welingkar Institute of Management Development & Research* 2013; **15**: 79-97
- 48 **Shigemi J**, Mino Y, Tsuda T. The role of perceived job stress in the relationship between smoking and the development of peptic ulcers. *J Epidemiol* 1999; **9**: 320-326 [PMID: 10616265 DOI: 10.2188/jea.9.320]
- 49 **Susheela AK**, Mondal NK, Singh A. Exposure to fluoride in smelter workers in a primary aluminum industry in India. *Int J Occup Environ Med* 2013; **4**: 61-72 [PMID: 23567531]
- 50 **Nakadaira H**, Yamamoto M, Matsubara T. Mental and physical effects of Tanshin funin, posting without family, on married male workers in Japan. *J Occup Health* 2006; **48**: 113-123 [PMID: 16612040 DOI: 10.1539/joh.48.113]
- 51 **Westerling R**, Gullberg A, Rosén M. Socioeconomic differences in 'avoidable' mortality in Sweden 1986-1990. *Int J Epidemiol* 1996; **25**: 560-567 [PMID: 8671557 DOI: 10.1093/ije/25.3.560]
- 52 **Gasbarrini G**, Pretolani S, Bonvicini F, Gatto MR, Tonelli E, Mégraud F, Mayo K, Ghironzi G, Giulianelli G, Grassi M. A population based study of Helicobacter pylori infection in a European country: the San Marino Study. Relations with gastrointestinal diseases. *Gut* 1995; **36**: 838-844 [PMID: 7615270 DOI: 10.1136/gut.36.6.838]
- 53 **Lin YH**, Chen CY, Hong WH, Lin YC. Perceived job stress and health complaints at a bank call center: comparison between inbound and outbound services. *Ind Health* 2010; **48**: 349-356 [PMID: 20562511 DOI: 10.2486/indhealth.48.349]
- 54 **Nabavizadeh F**, Vahedian M, Sahraei H, Adeli S, Salimi E. Physical and psychological stress have similar effects on gastric acid and pepsin secretions in rat. *J Stress Physiology Biochem* 2011; **7**: 164-174
- 55 **Huerta-Franco R**, Vargas-Luna M, Hernandez E, Capacione K, Cordova T. Use of short-term bio-impedance for gastric motility assessment. *Med Eng Phys* 2009; **31**: 770-774 [PMID: 19303803 DOI: 10.1016/j.medengphys.2009.02.008]
- 56 **Wenger MA**, Engel BT, Clemens TL, Cullen TD. Stomach motility in man as recorded by the magnetometer method. *Gastroenterology* 1961; **41**: 479-485 [PMID: 14006124]
- 57 **Doglietto F**, Prevedello DM, Jane JA, Han J, Laws ER. Brief history of endoscopic transsphenoidal surgery--from Philipp Bozzini to the First World Congress of Endoscopic Skull Base Surgery. *Neurosurg Focus* 2005; **19**: E3 [PMID: 16398480 DOI: 10.3171/foc.2005.19.6.4]
- 58 **Janssen P**, Vanden Berghe P, Verschuere S, Lehmann A, Depoortere I, Tack J. Review article: the role of gastric motility in the control of food intake. *Aliment Pharmacol Ther* 2011; **33**: 880-894 [PMID: 21342212 DOI: 10.1111/j.1365-2036.2011.04609.x]
- 59 **Sobreira LF**, Zucoloto S, Garcia SB, Troncon LE. Effects of myenteric denervation on gastric epithelial cells and gastric emptying. *Dig Dis Sci* 2002; **47**: 2493-2499 [PMID: 12452385 DOI: 10.1023/A:1020508009213]
- 60 **Quintana E**, Hernández C, Alvarez-Barrientos A, Esplugues JV, Barrachina MD. Synthesis of nitric oxide in postganglionic myenteric neurons during endotoxemia: implications for gastric motor function in rats. *FASEB J* 2004; **18**: 531-533 [PMID: 14715697]
- 61 **Quigley EMM**. Gastric motor and sensory function and motor disorders of the stomach. In: Feldman M, Friedman LS, Sleisenger MH, editors. *Gastrointestinal and liver disease*. 7th ed. Philadelphia: W.B. Saunders, 2002: 691-713
- 62 **Giouvanoudi A**, Amaee WB, Sutton JA, Horton P, Morton R, Hall W, Morgan L, Freedman MR, Spyrou NM. Physiological interpretation of electrical impedance epigastrography measurements. *Physiol Meas* 2003; **24**: 45-55 [PMID: 12636186 DOI: 10.1088/0967-3334/24/1/304]
- 63 **Quigley EM**. Review article: gastric emptying in functional gastrointestinal disorders. *Aliment Pharmacol Ther* 2004; **20** Suppl 7: 56-60 [PMID: 15521856 DOI: 10.1111/j.1365-2036.2004.02186.x]
- 64 **Stanghellini V**, Malagelada JR, Zinsmeister AR, Go VL, Kao PC. Stress-induced gastroduodenal motor disturbances in humans: possible humoral mechanisms. *Gastroenterology* 1983; **85**: 83-91 [PMID: 6303893]
- 65 **Stanghellini V**, Tosetti C, Paternico A, Barbara G, Morselli-Labate AM, Monetti N, Marengo M, Corinaldesi R. Risk indicators of delayed gastric emptying of solids in patients with functional dyspepsia. *Gastroenterology* 1996; **110**: 1036-1042 [PMID: 8612991]
- 66 **Delvaux M**. Role of visceral sensitivity in the pathophysiology of irritable bowel syndrome. *Gut* 2002; **51** Suppl 1: i67-i71 [PMID: 12077070 DOI: 10.1136/gut.51.suppl.1.i67]
- 67 **Huerta R**, Brizuela-Gamiño OL. Interaction of pubertal status, mood and self-esteem in adolescent girls. *J Reprod Med* 2002; **47**: 217-225 [PMID: 11933687]
- 68 **Sagami Y**, Shimada Y, Tayama J, Nomura T, Satake M, Endo Y, Shoji T, Karahashi K, Hongo M, Fukudo S. Effect of a corticotropin releasing hormone receptor antagonist on colonic sensory and motor function in patients with irritable bowel syndrome. *Gut* 2004; **53**: 958-964 [PMID: 15194643 DOI: 10.1136/gut.2003.018911]
- 69 **Sagami Y**, Hongo M. [The gastrointestinal motor function in irritable bowel syndrome (IBS)]. *Nihon Rinsho* 2006; **64**: 1441-1445 [PMID: 16898609]
- 70 **Saxena B**, Singh S. Investigations on gastroprotective effect of citalopram, an antidepressant drug against stress and pyloric ligation induced ulcers. *Pharmacol Rep* 2011; **63**: 1413-1426 [PMID: 22358089]

P- Reviewer: Acuna-Castroviejo D

S- Editor: Zhai HH L- Editor: A E- Editor: Liu XM



## Usefulness of percutaneous endoscopic gastrostomy for supportive therapy of advanced aerodigestive cancer

Haruei Ogino, Hirotada Akiho

Haruei Ogino, Hirotada Akiho, Department of Gastroenterology, Kitakyushu Municipal Medical Center, Kitakyushu 802-0077, Japan  
Author contributions: Both authors contributed extensively to this manuscript; Ogino H provided a significant editorial and literature contribution; Akiho H provided literature-related comments and review.

Correspondence to: Hirotada Akiho, MD, PhD, Department of Gastroenterology, Kitakyushu Municipal Medical Center, 2-1-1 Bashaku, Kokura-kitaku, Kitakyushu 802-0077, Japan. [akiho@med.kyushu-u.ac.jp](mailto:akiho@med.kyushu-u.ac.jp)

Telephone: +81-93-5411831 Fax: +81-93-5338693

Received: June 10, 2013 Revised: September 6, 2013

Accepted: October 16, 2013

Published online: November 15, 2013

### Abstract

Aerodigestive cancer, like esophageal cancer or head and neck cancer, is well known to have a poor prognosis. It is often diagnosed in the late stages, with dysphagia being the major symptom. Insufficient nutrition and lack of stimulation of the intestinal mucosa may worsen immune compromise due to toxic side effects. A poor nutritional status is a significant prognostic factor for increased mortality. Therefore, it is most important to optimize enteral nutrition in patients with aerodigestive cancer before and during treatment, as well as during palliative treatment. Percutaneous endoscopic gastrostomy (PEG) may be useful for nutritional support. However, PEG tube placement is limited by digestive tract stenosis and is an invasive endoscopic procedure with a risk of complications. There are three PEG techniques. The pull/push and introducer methods have been established as standard techniques for PEG tube placement. The modified introducer method, namely the direct method, allows for direct placement of a larger button-bumper-type catheter device. PEG tube placement using the introducer method or the direct method may be a much safer alternative than the pull/push method. PEG may be recommended in patients with aerodigestive cancer because of the im-

proved complication rate.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Aerodigestive cancer; Percutaneous endoscopic gastrostomy; Direct method; Introducer method; Pull/push method; Complications

**Core tip:** Aerodigestive cancer is well known to have a poor prognosis and is often diagnosed in the late stages with dysphagia. Insufficient nutrition and lack of stimulation of the intestinal mucosa may worsen immune compromise. Therefore, it is most important to optimize enteral nutrition before and during treatment, as well as during palliative treatment. Percutaneous endoscopic gastrostomy (PEG) may be useful for nutritional support. PEG tube placement using the introducer method or the direct method may be a much safer alternative than the pull/push method. PEG may be recommended in patients with aerodigestive cancer because of the improved complication rate.

Ogino H, Akiho H. Usefulness of percutaneous endoscopic gastrostomy for supportive therapy of advanced aerodigestive cancer. *World J Gastrointest Pathophysiol* 2013; 4(4): 119-125 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/119.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.119>

### INTRODUCTION

Tumors of the esophagus and gastroesophageal junction or head and neck are some of the most malignant cancers with high mortality rates because many patients are diagnosed in the advanced stages<sup>[1]</sup>. Dysphagia, or difficulty swallowing, is one of the most distressing and debilitating symptoms. Dysphagia leads to nutritional compromise and deterioration of quality of life<sup>[2,3]</sup>. When the esophageal lumen becomes stenotic to less

than 14 mm in diameter, dysphagia generally develops. It first becomes difficult to swallow solid food. Next, it becomes difficult to swallow semisolid food. Finally, fluids and even saliva are difficult to swallow<sup>[4]</sup>. Patients develop anorexia and significant weight loss secondary to the tumor effects and may present with varying degrees of malnutrition. A poor nutritional status is a significant prognostic factor for increased mortality<sup>[5]</sup>.

Selection of therapy for aerodigestive cancer is dependent upon the tumor stage, location and histological type, and the physician's experience and preference. Therapeutic options include surgical resection of the primary tumor, chemotherapy and radiotherapy. Therapies are sometimes combined, such as chemotherapy plus surgery or chemotherapy and radiotherapy plus surgery. Many of these patients find that their initial dysphagia worsens during this treatment because of side effects such as esophagitis and oral mucositis. Moreover, insufficient nutrition and lack of stimulation of the intestinal mucosa may worsen immune compromise due to toxic side effects<sup>[6]</sup>. During these periods, it is most important to optimize enteral nutrition. Early enteral nutrition reduces the incidence of life-threatening surgical complications in patients who undergo esophagectomy or esophagogastrectomy for esophageal carcinoma<sup>[7-11]</sup>. Nutrition is administered through a transnasal feeding tube for short-term feeding when oral intake is not possible. When chemotherapy and/or radiotherapy are intended to be curative, they frequently compromise oral intake for a long period of time. Nasogastric tubes are easy to place but they are poorly tolerated for prolonged periods of feeding. Percutaneous endoscopic gastrostomy (PEG) may be one of the best options for nutritional support.

A majority of patients are destined to receive palliation only, which is associated with a severely impaired health-related quality of life. These patients require palliative treatment, including brachytherapy, chemotherapy and endoscopic palliation techniques, such as esophageal dilatation, intraluminal stents and laser therapy, to relieve progressive dysphagia<sup>[12,13]</sup>. The two most commonly used strategies for improving swallowing are stent insertion and radiation, including intraluminal brachytherapy. They allow for an almost normal oral intake. Unfortunately, some patients develop restenosis symptoms after palliative therapy and some develop severe treatment-related side effects such as mucositis from radiation therapy. Stent insertion is difficult in some patients with proximal esophageal cancers or head and neck cancers. For these patients, PEG or nasal tubes may be the best options for nutritional support.

## PEG PROCEDURE

There are three PEG tube insertion methods. The pull/push and introducer methods have been established as standard techniques for PEG tube placement. In the pull/push method, the feeding tube is introduced through the mouth. In the introducer method, balloon-

type catheter feeding tubes can be inserted directly into the stomach through the abdominal wall. The third method is the modified introducer method (*i.e.*, direct method). The direct method allows for direct placement of a larger button-bumper-type catheter device. Use of the direct method is spreading in Japan, but it is not yet used worldwide<sup>[14]</sup>. Each method has advantages and disadvantages.

### **Pull/push method**

The pull/push PEG technique is based on the standard Ponsky technique in which a guidewire is inserted through the abdominal wall under endoscopic guidance, grasped by a snare through a port on the endoscope, and subsequently advanced in a retrograde manner through the patient's mouth. The remaining end exits the patient through the anterior abdominal wall. A 20-French Ross Flexiflo Inverta-PEG tube (Abbott Laboratories, Columbus, OH) is then secured to the transoral end of the patient's mouth and abdominal wall by pulling the extra-abdominal end of the wire to advance the gastrostomy tube<sup>[15]</sup>.

### **Introducer method**

The introducer PEG technique is based on the Russell introducer method of PEG placement. After the endoscope is inserted and the PEG site is marked, four T-fasteners are placed before gastrostomy tube insertion to secure the stomach to the anterior abdominal wall. This prevents gastric wall displacement while inserting the gastrostomy tube. Using the Seldinger technique, a short guidewire is then passed transabdominally under endoscope visualization. Serial dilators are passed over the guidewire to create a stoma tract; the endoscope remains in place for visualization and verification of gastrostomy tube placement. An 18-French Ross Flexiflo gastrostomy tube (Abbott Laboratories) is then inserted or pushed over the guidewire, directly through the anterior abdominal wall<sup>[16]</sup>.

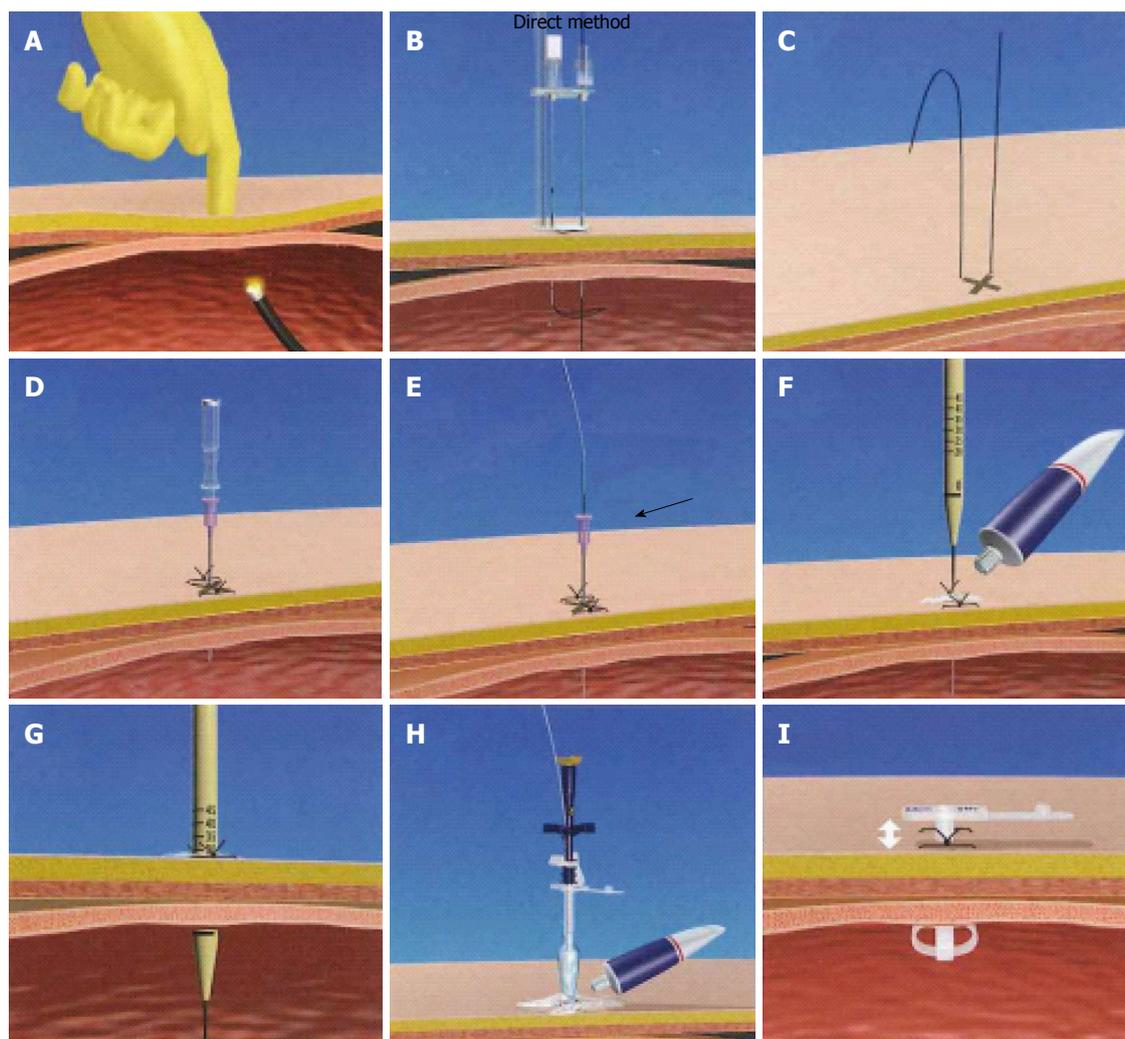
### **Direct method**

The direct method is a modified version of the introducer method (Direct Ideal PEG kit; Olympus Corp., Tokyo, Japan). After the stomach is secured to the anterior abdominal wall, the skin incision is dilated by passing a dilator percutaneously into the stomach over the guidewire as the same as introducer method. After the dilator is removed, a 24-French PEG tube is inserted using an obturator<sup>[14]</sup> (Figure 1).

## OUTCOMES OF PEG

### **PEG in patients with aerodigestive cancer**

PEG tube feeding is the preferred method with which to provide long-term tube feeding and its use is currently widespread. Many studies have examined the usefulness of PEG for aerodigestive cancer. A PEG tube was inserted in patients with oral intake difficulties for the



**Figure 1 Direct method.** A: The transilluminated area on the abdominal wall was pushed with a finger; B, C: The stomach was punctured using a double-lumen gastroscopy device; D: A needle with an outer plastic sheath (18-French) was introduced into the stomach under endoscopic control; E: The needle was removed and the guidewire was replaced; F, G: The skin incision was dilated by passing a dilator percutaneously into the stomach over the guidewire under endoscopic visualization; H: After the dilator was removed, a 24-French percutaneous endoscopic gastrostomy tube using an obturator was inserted over the guidewire; I: The tube was fixed to the abdominal wall.

purpose of nutrition support in all stages and locations, including patients who had undergone chemotherapy and chemoradiation therapy with curative intent<sup>[17-22]</sup>. Chemotherapy or chemoradiation therapy is frequently associated with mucositis, dysphagia, loss of taste and anorexia. Chemotherapy, chemoradiation therapy and hyperfractionated radiation therapy are usually associated with even more severe treatment-related side effects and greater impairment of swallowing function. These treatments are long-term. Therefore, during these periods, PEG tube insertion may be one of the best options for nutritional support if the complication and mortality rates of PEG are low. Nasogastric tubes are easy to place but they are poorly tolerated for prolonged periods of feeding because they are associated with frequent ulceration, esophageal reflux and general discomfort. PEG tubes are better tolerated but they must be used selectively in patients who can be predicted to have a long-term need for nutritional support<sup>[23]</sup>.

There are more reports of patients with head and

neck cancer than patients with esophageal cancer. One of the reasons is that in the operation planned in esophageal cancer patients, PEG may limit the reconstruction of the stomach after esophagectomy because of the adhesion of the stomach and the abdominal wall, or the possibility of the injury for the right gastroepiploic artery which is needed in the reconstruction of the stomach<sup>[17]</sup>. Another reason for this is that stent insertion and brachytherapy are the first-choice palliative treatments in patients with middle and low esophageal cancers in many institutions. In terms of nutritional support, the most important factor is maintenance of oral food intake, which should stabilize or even improve quality of life. Dysphagia improves more rapidly after stent placement<sup>[12,15]</sup> and long-term relief of dysphagia is better after brachytherapy<sup>[24,25]</sup>. Therefore, stent placement may be reserved for esophageal cancer patients with severe dysphagia in combination with a short life expectancy who need more rapid relief of dysphagia and for patients with persistent or recurrent tumor growth after

**Table 1 Comparison of the advantages and disadvantages of the pull, introducer and direct percutaneous endoscopic gastrostomy placement methods**

	Advantages	Disadvantages
Pull method	Bumper type device inside stomach prevents misplacement of catheter Large-bore catheters can be used immediately after placement	Catheter may be contaminated during passage through mouth/esophagus → Increased risk of wound infection and tumor implantation Endoscope must be inserted twice to confirm correct placement
Introducer method	Adherence to aseptic technique guarantees low risk of wound infection Endoscope must be inserted only once	Risk of bleeding and incorrect puncture with large trocar Only small-lumen catheters can be used immediately after placement Catheter size must be increased step by step
Direct PEG Kit	Adherence to aseptic technique guarantees low risk of wound infection Endoscope must be inserted only once Small puncture needle and blunt dilator → small wound One-step insertion of bumper type device Large-bore catheters can be used immediately after placement	High probability of catheter misplacement (if using balloon type) Risk of bleeding

PEG: Percutaneous endoscopic gastrostomy.

brachytherapy<sup>[12,13]</sup>. When these modalities are technically not possible, nutritional support with a nasogastric feeding tube or PEG tube should be considered to maintain adequate calorie intake. Grilo *et al.*<sup>[22]</sup> suggest that PEG should be considered as a nutritional support method in patients with upper esophageal cancer that is unsuitable for esophageal stenting. For patients who suffer from restenosis symptoms after palliative therapy or who have proximal esophageal cancers or head and neck cancer, PEG may be one of the best options for nutritional support.

Thus, depending on the treatment, disease and the degree of stenosis, the following situations are indications for PEG. First, in aerodigestive cancer patients undergoing chemotherapy or chemoradiation therapy who are suffering from dysphagia, PEG is the first choice. Stenosis, even if not severe, and if lesions are located in the upper esophagus or head and neck, is an indication for PEG because difficult long-term oral intake is expected due to mucositis and esophagitis during the treatment. Next, in the operation planned for head and neck cancer patients, PEG is indicated because the stomach is not used for reconstruction. Lastly, in palliative treatment, patients with lesions of the upper esophagus or head and neck with the difficulty of a stent are indications for PEG. In addition, PEG will be indicated in patients in whom stenosis is severe even after palliative radiation therapy or a stent (Figure 2).

However, studies on this topic have weaknesses typical to retrospective studies. Nugent *et al.*<sup>[26]</sup> and Locher *et al.*<sup>[27]</sup> reported that there is insufficient evidence to determine the optimal method of enteral feeding for patients with head and neck cancer receiving radiotherapy and/or chemoradiotherapy. Larger studies of enteral feeding in patients with esophageal cancer are needed.

## COMPLICATIONS

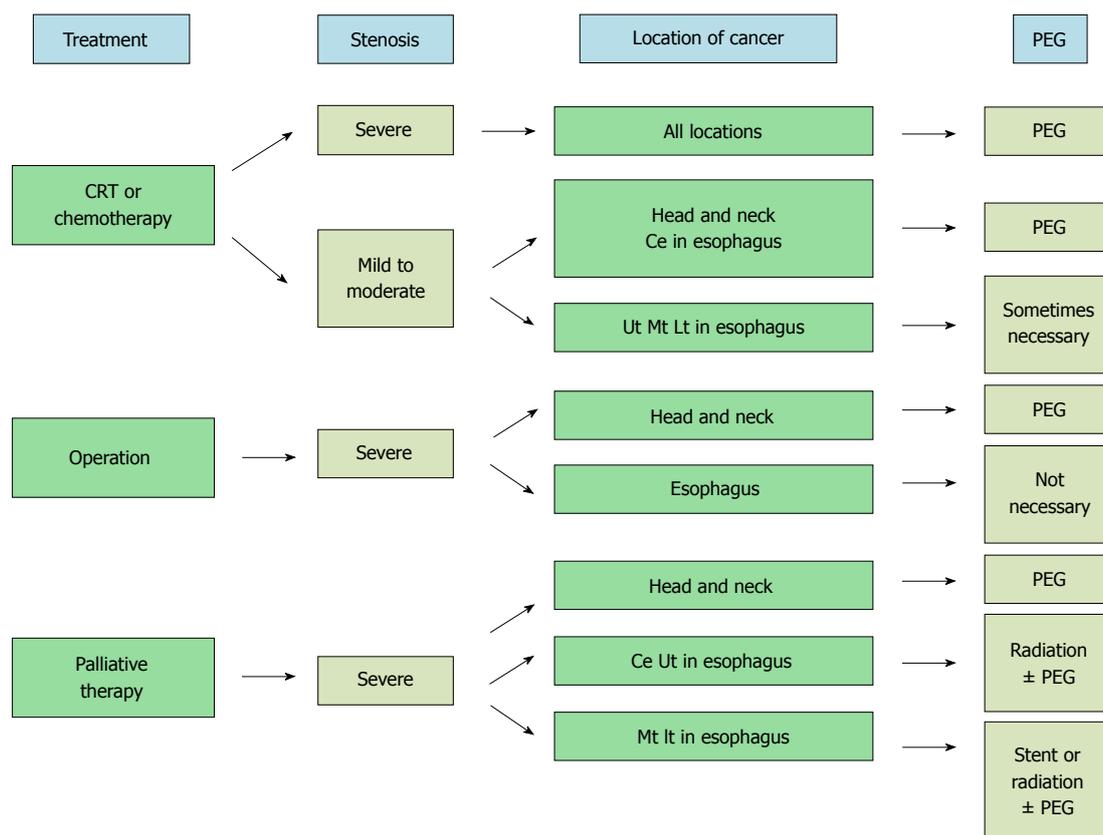
PEG tube placement is an invasive endoscopic procedure with a risk of complications. Minor complications

resulting from PEG tube placement include cellulitis, ileus, peristomal leakage, extrusion, tube obstruction and gastric wall hematoma formation. Major complications include peritonitis, hemorrhage, airway aspiration, peristomal wound infection, buried bumper syndrome, tumor implantation and gastrocolic fistula<sup>[28,29]</sup> (Table 1).

The major complications of the standard pull/push method, which requires an esophageal lumen sufficient to pass a standard endoscope<sup>[30]</sup>, include peristomal wound infections, presumably resulting from contamination of the gastrostomy catheter as it passes through the oral cavity<sup>[14,31]</sup>, and tumor implantation at the PEG site<sup>[28,32]</sup> which are specific for pull/push method in the aerodigestive cancer patients. In the literature on patients with cancer, the overall complication and mortality rates of the pull/push method in patients with head and neck cancer are 10.9%-42.0% and 0%-5%, respectively<sup>[15,17,18,20-22,33-36]</sup>.

An overall complication rate of 0%-11% and mortality rate of 0% have been reported with the introducer method<sup>[15,16,37,38]</sup> compared with the pull/push method in patients with aerodigestive cancer. In the pull/push method, one reason for the high complication rate may be that it is necessary to dilate the lumen before treatment when the stenosis caused by the tumor is severe. In many aerodigestive cancer patients, PEG tube placement by pull/push method can be limited by digestive tract stenosis. PEG tube placement using an introducer is the safest alternative in this group of patients but use of the available devices is difficult to implement.

In the past, the introducer technique was technically more demanding and associated with a lower success rate. This problem was solved by the use of T-fasteners to secure the anterior stomach to the abdominal wall<sup>[39,40]</sup>. Therefore, recent data on the introducer method using T-fasteners show low complication rates of less than 11% and no mortality<sup>[38,41-45]</sup>. However, Dyck's study shows that severe short-term complications may occur in patients with esophageal or head and neck tumors after place-



**Figure 2** Algorithm of percutaneous endoscopic gastrostomy for aerodigestive cancer. CRT: Chemoradiation therapy; Ce: Cervical esophagus; Ut: Upper thoracic esophagus; Mt: Middle thoracic esophagus; Lt: Lower thoracic esophagus; PEG: Percutaneous endoscopic gastrostomy.

ment of the introducer PEG tube with T-fasteners, leading to urgent surgical intervention and even death in a substantial number of patients<sup>[20]</sup>. Why the complication and mortality rates were high in Dyck's study is unclear. Selection bias may be one reason. Van Dyck *et al.*<sup>[20]</sup> reported that better follow-up of PEG tube daily care might be necessary. In almost all studies, the complication and mortality rates were low. Larger studies on the introducer method in patients with esophageal cancer are needed.

One disadvantage of the introducer method is that only small diameter balloon-type catheters are available and the requirement for frequent catheter changes when long-term tube feeding is needed<sup>[42,43]</sup>. The modification of the PEG device using the introducer technique is improved in this respect. It allows for the use of a larger-caliber tube with low complication rates and no procedure-related mortality. The direct method reduces the incidence of catheter changes compared with the 20-French catheter in the standard pull/push method. It is also feasible, safe and efficient in outpatients with obstructive head and neck cancer. However, procedure-related severe bleeding associated with the direct method has been reported<sup>[46]</sup>.

## TIMING OF PEG TUBE PLACEMENT

Cady<sup>[47]</sup> reported that patients who require therapeutic

PEG tube placement in response to significant weight loss during treatment suffer greater morbidity than patients who receive PEG tubes prophylactically. Patients who have a PEG tube at treatment initiation experience less overall weight loss and fewer hospitalizations and toxicity-related treatment interruptions. However, Locher *et al.*<sup>[27]</sup> reported that systematic evidence assessing both the benefits and harm associated with prophylactic PEG tube placement in patients undergoing treatment for head and neck cancer is weak and the benefits and potential for harm have not been established.

## CONCLUSION

An optimal supportive treatment for aerodigestive carcinoma is not yet available. PEG has many advantages for aerodigestive cancer, although there is insufficient evidence to determine the optimal method of enteral feeding. Enteral nutrition by the introducer method or the direct method must be studied with an emphasis on the long-term effectiveness and safety of supportive therapy of the aerodigestive cancer.

## REFERENCES

- 1 **Pisani P**, Parkin DM, Bray F, Ferlay J. Estimates of the worldwide mortality from 25 cancers in 1990. *Int J Cancer* 1999; 83: 18-29 [PMID: 10449602]
- 2 **Javle M**, Ailawadhi S, Yang GY, Nwogu CE, Schiff MD,

- Nava HR. Palliation of malignant dysphagia in esophageal cancer: a literature-based review. *J Support Oncol* 2006; **4**: 365-373, 379 [PMID: 17004508]
- 3 **Conigliaro R**, Battaglia G, Repici A, De Pretis G, Ghezzi L, Bittinger M, Messmann H, Demarquay JF, Togni M, Bianchi S, Filiberti R, Conio M. Polyflex stents for malignant oesophageal and oesophagogastric stricture: a prospective, multicentric study. *Eur J Gastroenterol Hepatol* 2007; **19**: 195-203 [PMID: 17301645]
  - 4 **Dakkak M**, Hoare RC, Maslin SC, Bennett JR. Oesophagitis is as important as oesophageal stricture diameter in determining dysphagia. *Gut* 1993; **34**: 152-155 [PMID: 8432464]
  - 5 **Miyata H**, Yano M, Yasuda T, Hamano R, Yamasaki M, Hou E, Motoori M, Shiraishi O, Tanaka K, Mori M, Doki Y. Randomized study of clinical effect of enteral nutrition support during neoadjuvant chemotherapy on chemotherapy-related toxicity in patients with esophageal cancer. *Clin Nutr* 2012; **31**: 330-336 [PMID: 22169459]
  - 6 **Motoori M**, Yano M, Yasuda T, Miyata H, Peng YF, Yamasaki M, Shiraishi O, Tanaka K, Ishikawa O, Shiozaki H, Doki Y. Relationship between immunological parameters and the severity of neutropenia and effect of enteral nutrition on immune status during neoadjuvant chemotherapy on patients with advanced esophageal cancer. *Oncology* 2012; **83**: 91-100 [PMID: 22777298]
  - 7 **Gabor S**, Renner H, Matzi V, Ratzenhofer B, Lindenmann J, Sankin O, Pinter H, Maier A, Smolle J, Smolle-Jüttner FM. Early enteral feeding compared with parenteral nutrition after oesophageal or oesophagogastric resection and reconstruction. *Br J Nutr* 2005; **93**: 509-513 [PMID: 15946413]
  - 8 **Fujita T**, Daiko H, Nishimura M. Early enteral nutrition reduces the rate of life-threatening complications after thoracic esophagectomy in patients with esophageal cancer. *Eur Surg Res* 2012; **48**: 79-84 [PMID: 22377820]
  - 9 **Bozzetti F**, Braga M, Gianotti L, Gavazzi C, Mariani L. Postoperative enteral versus parenteral nutrition in malnourished patients with gastrointestinal cancer: a randomised multicentre trial. *Lancet* 2001; **358**: 1487-1492 [PMID: 11705560]
  - 10 **Wu GH**, Liu ZH, Wu ZH, Wu ZG. Perioperative artificial nutrition in malnourished gastrointestinal cancer patients. *World J Gastroenterol* 2006; **12**: 2441-2444 [PMID: 16688841]
  - 11 **Braga M**, Gianotti L, Gentilini O, Parisi V, Salis C, Di Carlo V. Early postoperative enteral nutrition improves gut oxygenation and reduces costs compared with total parenteral nutrition. *Crit Care Med* 2001; **29**: 242-248 [PMID: 11246300]
  - 12 **Siersema PD**. New developments in palliative therapy. *Best Pract Res Clin Gastroenterol* 2006; **20**: 959-978 [PMID: 16997172]
  - 13 **Homs MY**, Kuipers EJ, Siersema PD. Palliative therapy. *J Surg Oncol* 2005; **92**: 246-256 [PMID: 16299791]
  - 14 **Horiuchi A**, Nakayama Y, Tanaka N, Fujii H, Kajiyama M. Prospective randomized trial comparing the direct method using a 24 Fr bumper-button-type device with the pull method for percutaneous endoscopic gastrostomy. *Endoscopy* 2008; **40**: 722-726 [PMID: 18773341]
  - 15 **Tucker AT**, Gourin CG, Ghegan MD, Porubsky ES, Martindale RG, Terris DJ. 'Push' versus 'pull' percutaneous endoscopic gastrostomy tube placement in patients with advanced head and neck cancer. *Laryngoscope* 2003; **113**: 1898-1902 [PMID: 14603043]
  - 16 **Foster JM**, Filocamo P, Nava H, Schiff M, Hicks W, Rigual N, Smith J, Loree T, Gibbs JF. The introducer technique is the optimal method for placing percutaneous endoscopic gastrostomy tubes in head and neck cancer patients. *Surg Endosc* 2007; **21**: 897-901 [PMID: 17180272]
  - 17 **Stockeld D**, Fagerberg J, Granström L, Backman L. Percutaneous endoscopic gastrostomy for nutrition in patients with oesophageal cancer. *Eur J Surg* 2001; **167**: 839-844 [PMID: 11848238]
  - 18 **Rabie AS**. Percutaneous endoscopic gastrostomy (PEG) in cancer patients; technique, indications and complications. *Gulf J Oncology* 2010; **(7)**: 37-41 [PMID: 20164007]
  - 19 **Yagishita A**, Kakushima N, Tanaka M, Takizawa K, Yamaguchi Y, Matsubayashi H, Ono H. Percutaneous endoscopic gastrostomy using the direct method for aerodigestive cancer patients. *Eur J Gastroenterol Hepatol* 2012; **24**: 77-81 [PMID: 22081009]
  - 20 **Van Dyck E**, Macken EJ, Roth B, Pelckmans PA, Moreels TG. Safety of pull-type and introducer percutaneous endoscopic gastrostomy tubes in oncology patients: a retrospective analysis. *BMC Gastroenterol* 2011; **11**: 23 [PMID: 21410958]
  - 21 **Zuercher BF**, Grosjean P, Monnier P. Percutaneous endoscopic gastrostomy in head and neck cancer patients: indications, techniques, complications and results. *Eur Arch Otorhinolaryngol* 2011; **268**: 623-629 [PMID: 21046412]
  - 22 **Grilo A**, Santos CA, Fonseca J. Percutaneous endoscopic gastrostomy for nutritional palliation of upper esophageal cancer unsuitable for esophageal stenting. *Arq Gastroenterol* 2012; **49**: 227-231 [PMID: 23011248]
  - 23 **Corry J**, Poon W, McPhee N, Milner AD, Cruickshank D, Porceddu SV, Rischin D, Peters LJ. Randomized study of percutaneous endoscopic gastrostomy versus nasogastric tubes for enteral feeding in head and neck cancer patients treated with (chemo) radiation. *J Med Imaging Radiat Oncol* 2008; **52**: 503-510 [PMID: 19032398]
  - 24 **Sur R**, Donde B, Falkson C, Ahmed SN, Levin V, Nag S, Wong R, Jones G. Randomized prospective study comparing high-dose-rate intraluminal brachytherapy (HDRILBT) alone with HDRILBT and external beam radiotherapy in the palliation of advanced esophageal cancer. *Brachytherapy* 2004; **3**: 191-195 [PMID: 15607150]
  - 25 **Homs MY**, Eijkenboom WM, Coen VL, Haringsma J, van Blankenstein M, Kuipers EJ, Siersema PD. High dose rate brachytherapy for the palliation of malignant dysphagia. *Radiother Oncol* 2003; **66**: 327-332 [PMID: 12742273]
  - 26 **Nugent B**, Lewis S, O'Sullivan JM. Enteral feeding methods for nutritional management in patients with head and neck cancers being treated with radiotherapy and/or chemotherapy. *Cochrane Database Syst Rev* 2013; **1**: CD007904 [PMID: 23440820 DOI: 10.1002/14651858]
  - 27 **Locher JL**, Bonner JA, Carroll WR, Caudell JJ, Keith JN, Kilgore ML, Ritchie CS, Roth DL, Tajeu GS, Allison JJ. Prophylactic percutaneous endoscopic gastrostomy tube placement in treatment of head and neck cancer: a comprehensive review and call for evidence-based medicine. *JPEN J Parenter Enteral Nutr* 2011; **35**: 365-374 [PMID: 21527598 DOI: 10.1177/0148607110377097]
  - 28 **Ellrichmann M**, Sergeev P, Bethge J, Arlt A, Topalidis T, Ambrosch P, Wiltfang J, Fritscher-Ravens A. Prospective evaluation of malignant cell seeding after percutaneous endoscopic gastrostomy in patients with oropharyngeal/esophageal cancers. *Endoscopy* 2013; **45**: 526-531 [PMID: 23780843]
  - 29 **Lin HS**, Ibrahim HZ, Kheng JW, Fee WE, Terris DJ. Percutaneous endoscopic gastrostomy: strategies for prevention and management of complications. *Laryngoscope* 2001; **111**: 1847-1852 [PMID: 11801956]
  - 30 **Ferguson DR**, Harig JM, Kozarek RA, Kelsey PB, Picha GJ. Placement of a feeding button ("one-step button") as the initial procedure. *Am J Gastroenterol* 1993; **88**: 501-504 [PMID: 8470628]
  - 31 **Maetani I**, Tada T, Ukita T, Inoue H, Sakai Y, Yoshikawa M. PEG with introducer or pull method: a prospective randomized comparison. *Gastrointest Endosc* 2003; **57**: 837-841 [PMID: 12776029]
  - 32 **Brown MC**. Cancer metastasis at percutaneous endoscopic gastrostomy stomata is related to the hematogenous or lymphatic spread of circulating tumor cells. *Am J Gastroenterol* 2000; **95**: 3288-3291 [PMID: 11095357]

- 33 **Baredes S**, Behin D, Deitch E. Percutaneous endoscopic gastrostomy tube feeding in patients with head and neck cancer. *Ear Nose Throat J* 2004; **83**: 417-419 [PMID: 15266879]
- 34 **Hujala K**, Sipilä J, Pulkkinen J, Grenman R. Early percutaneous endoscopic gastrostomy nutrition in head and neck cancer patients. *Acta Otolaryngol* 2004; **124**: 847-850 [PMID: 15370571]
- 35 **Ehrsson YT**, Langius-Eklöf A, Bark T, Laurell G. Percutaneous endoscopic gastrostomy (PEG) - a long-term follow-up study in head and neck cancer patients. *Clin Otolaryngol Allied Sci* 2004; **29**: 740-746 [PMID: 15533171]
- 36 **Chandu A**, Smith AC, Douglas M. Percutaneous endoscopic gastrostomy in patients undergoing resection for oral tumors: a retrospective review of complications and outcomes. *J Oral Maxillofac Surg* 2003; **61**: 1279-1284 [PMID: 14613083]
- 37 **Saunders JR**, Brown MS, Hirata RM, Jaques DA. Percutaneous endoscopic gastrostomy in patients with head and neck malignancies. *Am J Surg* 1991; **162**: 381-383 [PMID: 1951893]
- 38 **Giordano-Nappi JH**, Maluf-Filho F, Ishioka S, Hondo FY, Matuguma SE, Simas de Lima M, Lera dos Santos M, Retes FA, Sakai P. A new large-caliber trocar for percutaneous endoscopic gastrostomy by the introducer technique in head and neck cancer patients. *Endoscopy* 2011; **43**: 752-758 [PMID: 21656456 DOI: 10.1055/s-0030-1256495]
- 39 **Brown AS**, Mueller PR, Ferrucci JT. Controlled percutaneous gastrostomy: nylon T-fastener for fixation of the anterior gastric wall. *Radiology* 1986; **158**: 543-545 [PMID: 2934763]
- 40 **Robertson FM**, Crombleholme TM, Latchaw LA, Jacir NN. Modification of the "push" technique for percutaneous endoscopic gastrostomy in infants and children. *J Am Coll Surg* 1996; **182**: 215-218 [PMID: 8603240]
- 41 **Wejda BU**, Deppe H, Huchzermeyer H, Dormann AJ. PEG placement in patients with ascites: a new approach. *Gastrointest Endosc* 2005; **61**: 178-180 [PMID: 15672085]
- 42 **Dormann AJ**, Glosemeyer R, Leistner U, Deppe H, Roggel R, Wigglinghaus B, Huchzermeyer H. Modified percutaneous endoscopic gastrostomy (PEG) with gastropexy--early experience with a new introducer technique. *Z Gastroenterol* 2000; **38**: 933-938 [PMID: 11194881]
- 43 **Dormann AJ**, Wejda B, Kahl S, Huchzermeyer H, Ebert MP, Malfertheiner P. Long-term results with a new introducer method with gastropexy for percutaneous endoscopic gastrostomy. *Am J Gastroenterol* 2006; **101**: 1229-1234 [PMID: 16771943]
- 44 **Shastri YM**, Hoepffner N, Tessmer A, Ackermann H, Schroeder O, Stein J. New introducer PEG gastropexy does not require prophylactic antibiotics: multicenter prospective randomized double-blind placebo-controlled study. *Gastrointest Endosc* 2008; **67**: 620-628 [PMID: 18374024]
- 45 **Shigoka H**, Maetani I, Tominaga K, Gon K, Saitou M, Takenaka Y. Comparison of modified introducer method with pull method for percutaneous endoscopic gastrostomy: prospective randomized study. *Dig Endosc* 2012; **24**: 426-431 [PMID: 23078434 DOI: 10.1111/j.1443-1661]
- 46 **Koide T**, Inamori M, Kusakabe A, Uchiyama T, Watanabe S, Iida H, Endo H, Hosono K, Sakamoto Y, Fujita K, Takahashi H, Yoneda M, Tokoro C, Yasuzaki H, Goto A, Abe Y, Kobayashi N, Kubota K, Saito S, Nahajima A. Early complications following percutaneous endoscopic gastrostomy: results of use of a new direct technique. *Hepatogastroenterology* 2010; **57**: 1639-1644 [PMID: 21443135]
- 47 **Cady J**. Nutritional support during radiotherapy for head and neck cancer: the role of prophylactic feeding tube placement. *Clin J Oncol Nurs* 2007; **11**: 875-880 [PMID: 18063546]

**P- Reviewers:** Bustamante-Balen M, Levine EA, Nishida T, Shimoyama S

**S- Editor:** Zhai HH

**L- Editor:** Roemmele A **E- Editor:** Liu XM





百世登

**Baishideng**®

Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza,

315-321 Lockhart Road, Wan Chai, Hong Kong, China

Telephone: +852-6555-7188

Fax: +852-3177-9906

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

<http://www.wjgnet.com>

