

World Journal of *Dermatology*

World J Dermatol 2017 February 2; 6(1): 1-16



Editorial Board

2017-2020

The *World Journal of Dermatology* Editorial Board consists of 139 members, representing a team of worldwide experts in dermatology. They are from 39 countries, including Argentina (1), Australia (1), Austria (1), Brazil (1), Bulgaria (1), Canada (4), China (10), Croatia (1), Denmark (1), Egypt (1), Finland (1), France (3), Germany (5), Greece (4), Hungary (2), India (3), Iran (3), Israel (1), Italy (16), Japan (5), Malaysia (1), Malta (1), Mexico (4), Netherlands (3), Nigeria (2), Norway (1), Oman (1), Poland (2), Portugal (1), Romania (1), Saudi Arabia (1), Singapore (2), South Korea (8), Spain (8), Swaziland (2), Thailand (2), Turkey (6), United Kingdom (9), United States (19).

EDITOR-IN-CHIEF

Santosh K Katiyar, *Birmingham*

GUEST EDITORIAL BOARD MEMBERS

Tsong-Min Chang, *Taipei*

Ching-Chi Chi, *Taipei*

Jia-You Fang, *Taipei*

Sindy Hu, *Taipei*

Stephen Chu-Sung Hu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

María D Hermida, *Buenos Aires*



Australia

Ronald Sluyter, *Wollongong*



Austria

Iris Zalaudek, *Graz*



Brazil

Cidia Vasconcellos, *São Paulo*



Bulgaria

Georgi Tchernev, *Sofia*



Canada

Eleftherios P Diamandis, *Toronto*

Tim Lee, *Vancouver*

Gang Li, *Vancouver*

Kursad Turksen, *Ottawa*



China

Henry Hin Lee Chan, *Hong Kong*

Min Li, *Nanjing*

Cheng Tan, *Nanjing*

Guo-You Zhang, *Wenzhou*

Min Zheng, *Hangzhou*



Croatia

Mariastefania Antica, *Zagreb*



Denmark

Lars Iversen, *Aarhus*



Egypt

Moetaz El-Domyati, *Cairo*



Finland

Kari J Syrjänen, *Turku*



France

Guinot J Christiane, *Neully sur Seine*

Roger Mouawad, *Paris*

Stephane Rocchi, *Nice*



Germany

Martin Leverkus, *Mannheim*

Roderick AF MacLeod, *Braunschweig*

Markus Meissner, *Frankfurt*

Enno Schmidt, *Luebeck*

Peter Schroeder, *Dusseldorf*



Greece

Ioannis D Bassukas, *Ioannina*

Maria Dalamaga, *Athens*

Andreas Katsambas, *Athens*

Eleni Sotiriou, *Thessaloniki*



Hungary

Arpad Farkas, *Szeged*

Janos Fodor, *Budapest*



India

Sujoy Khan, *Kolkata*

Harsh Mohan, *Chandigarh*

Davinder Parsad, *Chandigarh*

**Iran**

Alireza Firooz, *Tehran*
 Mohammad R Namazi, *Shiraz*
 Afshin Sadighha, *Ilam*

**Israel**

Ronni Wolf, *Rehovo*

**Italy**

Giuseppe Argenziano, *Naples*
 Laura Atzori, *Cagliari*
 Ettore D Capoluongo, *Rome*
 Dott V Di Lernia, *Reggio Emilia*
 Paolo Fabbri, *Florence*
 Gabriella Fabbrocini, *Naples*
 Silvano Gallus, *Milan*
 Torello Lotti, *Firenze*
 Clelia Miracco, *Cosenza*
 Agnese Molinari, *Rome*
 Pierfrancesco Morganti, *Rome*
 Luigi Naldi, *Bergamo*
 Luca Negosanti, *Bologna*
 Raffaele Palmirotta, *Rome*
 Mario Santinami, *Milano*
 Riccarda Serri, *Milano*

**Japan**

Masutaka Furue, *Fukuoka*
 Fukumi Furukawa, *Wakayama*
 Mohammad Ghazizadeh, *Kawasaki*
 Yohei Tanaka, *Matsumoto*
 Toshiyuki Yamamoto, *Tokyo*

**Malaysia**

Felix Boon-Bin Yap, *Kuala Lumpur*

**Malta**

Michael J Boffa, *Floriana*

**Mexico**

Roberto G Arenas, *Mexico*
 Sergio A Cuevas-Covarrubias, *Mexico*
 Leopoldo Flores-Romo, *Mexico*
 Maria Bertha Torres-alvarez, *San Luis Potosi*

**Netherlands**

Rosalie M Luiten, *Amsterdam*

Arnold P Oranje, *Rotterdam*
 Arnold C Spek, *Amsterdam*

**Nigeria**

Maurice E Asuquo, *Calabar*
 Joseph I Ikechebelu, *Nnewi*

**Norway**

Andrej M Grijbovski, *Oslo*

**Oman**

Mohamed Mabruk, *Muscat*

**Poland**

Andrzej Grzybowski, *Poznan*
 Lidia Rudnicka, *Warsaw*

**Portugal**

Bruno Sarmento, *Porto*

**Romania**

Liana Manolache, *Bucharest*

**Saudi Arabia**

Feroze Kaliyadan, *Hofuf*

**Singapore**

Wei-Sheng Chong, *Singapore*
 Hong Liang Tey, *Singapore*

**South Korea**

Dong-Seok Kim, *Seoul*
 Chang Hoon Lee, *Seoul*
 Jongsung Lee, *Seongnam City*
 Chil Hwan Oh, *Seoul*
 Byung Soon Park, *Seoul*
 Myung-Geun Shin, *Hwasun*
 Jong-Hyuk Sung, *Seoul*
 Young Kwan Sung, *Daegu*

**Spain**

Agustin Alomar, *Barcelona*
 Salvador Arias-Santiago, *Granada*
 Juan G Gavín, *Vigo*
 Marcos A Gonzalez-Lopez, *Santander*

Ramon Grimalt, *Barcelona*
 Husein Husein-ElAhmed, *Granada*
 Ander Izeta, *San Sebastian*
 Marcela Del Rio, *Madrid*

**Switzerland**

Gunther FL Hofbauer, *Zurich*
 Alexander A Navarini, *Zurich*

**Thailand**

Chirayu U Auewarakul, *Bangkok*
 Viroj Wiwanitkit, *Bangkok*

**Turkey**

Berna Aksoy, *Kocaeli*
 Fatma Aydin, *Samsun*
 Cem Dane, *Istanbul*
 Sibel Dogan, *Istanbul*
 Aylin T Ermertcan, *Manisa*
 Ozlem Su, *Istanbul*

**United Kingdom**

Theodoros Dimitroulas, *Dudley*
 Bernhard F Gibbs, *Chatham Maritime*
 Evmorfia Ladoyanni, *Stourbridge*
 Mark R Nelson, *London*
 Adrian V Pace, *Dudley*
 Anthony B Paul, *London*
 Sam Shuster, *Woodbridge*
 Olga Tura, *Edinburgh*
 Indre Verpetinske, *Stourbridge*

**United States**

Jeremy S Bordeaux, *Cleveland*
 Robert F Diegelmann, *Richmond*
 Q Ping Dou, *Detroit*
 Zeev Estrov, *Houston*
 Vincent Falanga, *Providence*
 Miranda A Farage, *Cincinnati*
 Markus H Frank, *Boston*
 W Scott Goebel, *Indianapolis*
 Dan-Ning Hu, *New York*
 Amor Khachemoune, *Brooklyn*
 Arash Kimyai-Asadi, *Houston*
 Michael S Kolodney, *Torrance*
 Feng Liu, *Chapel Hill*
 Senthamil R Selvan, *San Diego*
 Lei Shi, *Fort Worth*
 Animesh A Sinha, *East Lansing*
 Jeffrey M Weinberg, *New York*
 John A Zic, *Nashville*

W

J

D

World Journal of Dermatology

Contents

Quarterly Volume 6 Number 1 February 2, 2017

REVIEW

- 1 Evidence based review of negative pressure wound therapy

Panayi AC, Leavitt T, Orgill DP

ABOUT COVER

Editorial Board Member of *World Journal of Dermatology*, Salvador Arias-Santiago, MD, PhD, Assistant Professor, Granada School of Medicine, San Cecilio University Hospital, 1801 Granada, Spain

AIM AND SCOPE

World Journal of Dermatology (*World J Dermatol*, *WJD*, online ISSN 2218-6190, DOI: 10.5314), is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of dermatology. *WJD* covers fungal diseases, dermatitis and eczema, urticarial diseases, drug eruptions, pruritus, erythroderma desquamativum, connective tissue diseases, bullous skin diseases, vascular skin diseases, skin appendage diseases, pigmentary diseases, genetic diseases, nutritional and metabolic disorders, tumors, sexually transmitted diseases, AIDS, traditional medicine, integrated Chinese and Western medicine, evidence-based medicine, epidemiology and nursing. The journal also publishes original articles and reviews that report the results of applied and basic research in fields related to dermatology, such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Dermatology is now indexed in China National Knowledge Infrastructure (CNKI).

FLYLEAF

I-II Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xin-Xia Song*

NAME OF JOURNAL
World Journal of Dermatology

ISSN
 ISSN 2218-6190 (online)

LAUNCH DATE
 June 2, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Santosh K Katiyar, PhD, Professor, Department of Dermatology, University of Alabama at Birmingham, Birmingham, AL 35294, United States

EDITORIAL BOARD MEMBERS
 All editorial board members resources online at <http://www.wjgnet.com/2218-6190/editorialboard.htm>

EDITORIAL OFFICE
 Fang-Fang Ji, Director

World Journal of Dermatology
 Baishideng Publishing Group Inc
 8226 Regency Drive, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 8226 Regency Drive,
 Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>
<http://www.wjgnet.com>

PUBLICATION DATE
 February 2, 2017

COPYRIGHT

© 2017 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION

<http://www.wjgnet.com/esps/>

Evidence based review of negative pressure wound therapy

Adriana C Panayi, Tripp Leavitt, Dennis P Orgill

Adriana C Panayi, School of Clinical Medicine, University of Cambridge, Cambridge CB2 0SP, United Kingdom

Tripp Leavitt, Boston University School of Medicine, Boston, MA 02115, United States

Dennis P Orgill, Department of Surgery, Division of Plastic Surgery, Brigham and Women's Hospital, Harvard Medical School, Boston, MA 02115, United States

Author contributions: Panayi AC gathered and analyzed the data, drafted and edited the manuscript; Leavitt T assisted in revision of the manuscript and figures; Orgill DP supervised and assisted in the drafting and provided critical revision of the manuscript.

Conflict-of-interest statement: The authors have no conflict of interest related to the manuscript. Dr. Orgill is a consultant for KCI and receives research funding through grants to Brigham and Women's Hospital

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Dr. Dennis P Orgill, Department of Surgery, Division of Plastic Surgery, Brigham and Women's Hospital, Harvard Medical School, 75 Francis St., Boston, MA 02115, United States. dorgill@partners.org
Telephone: +1-617-5257837
Fax: +1-617-7302855

Received: August 26, 2016

Peer-review started: September 27, 2016

First decision: October 15, 2016

Revised: December 16, 2016

Accepted: January 11, 2017

Article in press: January 12, 2017

Published online: February 2, 2017

Abstract

Vacuum-assisted closure, sometimes referred to as microdeformational wound therapy or most commonly negative pressure wound therapy (NPWT), has significantly improved wound care over the past two decades. NPWT is known to affect wound healing through four primary mechanisms (macrodeformation, microdeformation, fluid removal, and alteration of the wound environment) and various secondary mechanisms (including neurogenesis, angiogenesis, modulation of inflammation, and alterations in bioburden) which are described in this review. In addition, the technique has many established uses, for example in wound healing of diabetic and pressure ulcers, as well as burn and blast wounds. This therapy also has many uses whose efficacy has yet to be confirmed, for example the use in digestive surgery. Modifications of the traditional NPWT have also been established and are described in detail. This therapy has various considerations and contraindications which are summarized in this review. Finally, future perspectives, such as the optimal cycling of the treatment and the most appropriate interface material, are touched upon in the final segment. Overall, despite the fact that questions remain to be answered about NPWT, this technology is a major breakthrough in wound healing with significant potential use both in the hospital but also in the community.

Key words: Negative pressure wound therapy; Chronic wounds; Microdeformational wound therapy; Vacuum assisted closure; Pressure ulcers

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Negative pressure wound therapy has been very beneficial in the wound care of many different kinds of wounds, from pressure ulcers to open fractures mostly

due to its mechanism of action which we explain in detail. We explain the original purpose of this technology and going into detail about the many different ways it is currently being used in a clinical setting. Our review also explains its advantages and disadvantages and how they could be overcome. The last part of this review discusses the future of this technology and how it will keep impacting the field of wound care.

Panayi AC, Leavitt T, Orgill DP. Evidence based review of negative pressure wound therapy. *World J Dermatol* 2017; 6(1): 1-16 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i1/1.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i1.1>

INTRODUCTION

Since its introduction 19 years ago by Argenta and Morykwas, negative pressure wound therapy (NPWT) has emerged as a common treatment for acute and chronic wounds, including diabetic wounds, pressure ulcers, and burns^[1]. In simple terms, NPWT refers to any device that tightly seals the wound creating a near airtight environment to which a vacuum can be applied resulting in a series of biological reactions that enhance wound healing.

The terms Vacuum Assisted Closure (VAC, KCI, San Antonio, TX) and microdeformational wound therapy (MDWT) are sometimes used interchangeably with NPWT. MDWT refers to devices (generally foam) that substantially deform the wound surface^[2]. VAC now commonly refers to a family of devices using a highly porous foam based on the first commercially available NPWT device. Much of the clinical and basic science literature is based on these early devices (Figure 1). "Negative pressure" is somewhat of a misnomer as technically all pressure values should be positive.

Research on the application of NPWT in treating chronic non-healing wounds has largely taken the form of case studies, single-center studies, non-randomized controlled trials, with few randomized controlled trials (RCTs). This paper will analyze the available literature in order to summarize the current understanding of NPWT in terms of its mechanism of action, its applications, complications, contraindications and its future.

NPWT

In NPWT the wound is first filled with a porous material such as foam or gauze, that facilitates pressure transmission within the wound. A drainage port is then attached above the porous material and the wound is sealed with an adhesive film dressing. The drainage port is connected to a controlled vacuum pump which maintains negative pressure, usually ranging from -50 to -150 mmHg^[2,3]. The pressure can be applied in a continuous, intermittent, or variable mode, with the continuous type being the most frequently used. In the variable mode, the suction level changes but is never turned off, whereas

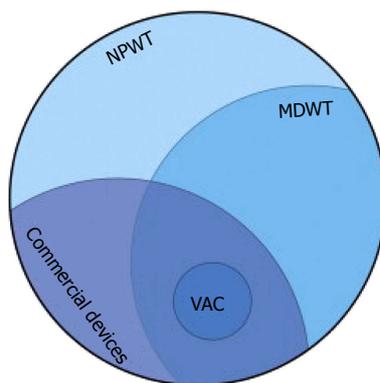


Figure 1 Visual representation of the definitions used in this article^[2]. Negative pressure wound therapy (NPWT) defines the entire field of wound therapy that applies differential suction to the wound. Microdeformational wound therapy (MDWT) refers specifically to the field as applied to the science of the foam wound interface that causes deformation. The Vacuum-assisted closure (VAC) therapy system is the most commonly used commercially available device. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

in the intermittent mode the pressure is switched on and off throughout the course of treatment.

The specific interface material that contacts the wound surface affects the biological response of the system. The most commonly used material is a reticulated open-pore polyurethane (PU) foam that forms a structure resembling a three-dimensional net. This lattice formation allows the vacuum to be evenly distributed throughout the foam and improves fluid drainage.

VAC

Three foam types are used in the VAC systems. Black polyurethane ether (VAC GranuFoam, KCI) is the most commonly used foam, and black polyurethane ester (VAC VeraFlow, KCI) is used in instillation systems. The white polyvinyl alcohol (VAC WhiteFoam, KCI) foam has very small pore sizes and is used to protect critical structures without inducing microdeformations, which will be discussed in the following section.

Gauze-based system

Usage of gauze in NPWT is based on the Chariker-Jeter method of application, which uses a moistened antimicrobial gauze (AMD; Covidien, Hampshire, United Kingdom) as a wound interface, along with 80 mmHg of negative pressure and a silicone drain^[4]. In one retrospective study with a mixed group of patients with challenging wounds, gauze used as a wound filler material was found to achieve reductions in wound size and volume comparable with published data from polyurethane foam-based systems^[5].

MECHANISMS OF ACTION

Primary mechanisms

NPWT is thought to promote wound healing *via* four

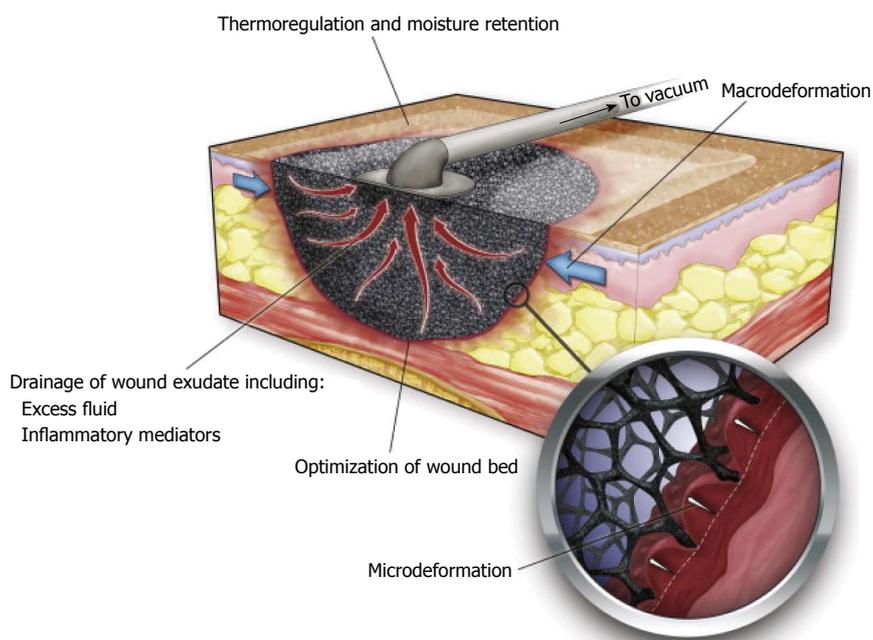


Figure 2 The 4 primary mechanisms of microdeformational wound therapy: (1) macrodeformation; (2) microdeformation (3) fluid removal; and (4) alteration of the wound environment^[2]. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

primary mechanisms: (1) macrodeformation; (2) microdeformation; (3) fluid removal; and (4) alteration of the wound environment (Figure 2).

Macrodeformation: Macrodeformation, or simply induced wound shrinkage, occurs when suction is applied to the foam causing pore collapse. This results in deformational forces being exerted on the wound edges, which draws them together. Macrodeformation can also induce compressive forces such as when these devices are used circumferentially on extremities^[6,7].

Studies in a porcine model showed that suction of 125 mmHg can decrease the volume of a PU foam by approximately 80% resulting in a substantial shrinkage of the wound^[8,9]. The extent of contraction is largely dependent on the deformability of the wound^[2].

The inherent tension in the dermis, which can cause wound margins to pull apart, and the attachment of the dermis to underlying structures vary in different parts of the body. Consequently different wounds contract to different degrees. For example, scalp skin is constrained by attachments to the underlying skull resulting in minimal deformation of the surrounding tissue when a foam based NPWT device is applied. In contrast, when a large open abdominal wound in an obese patient is treated with a similar device, the wound edges can be brought together in close approximation.

Microdeformation: Microdeformation describes the mechanical changes that occur on the microscopic scale when suction is applied to the porous material resulting in an undulated wound surface. For PU foam interfaces, treating wounds for several days results in a cobblestone appearance of the wound surface. Models that mimic the strain applied to a wound by the opposing forces of the suction and the sponge have been designed

to investigate these mechanical changes. Using finite element analysis (FEA), these models have shown that at 110 mmHg, MDWT results in a 5%-20% strain across the wound surface. This strain directly corresponds to the percentage change in length of the material exposed to the external forces^[10].

Mechanical forces, which include compression and tension from the foam, shear and hydrostatic forces from the extracellular fluid, and the effect of gravity, are transmitted throughout the tissue *via* the extracellular cell matrix (ECM). These forces vary greatly across the wound surface. For example, the tissue just underneath the foam struts is exposed to focal high compression, whereas the wound surface centrally in the pore is focally exposed to high tension^[10]. Microdeformation is the morphology that occurs due to the interplay between these forces.

Shear forces affect the cytoskeleton and activate a signaling cascade that upregulates granulation tissue formation and, hence, enhances wound healing^[11-13]. Furthermore, microdeformation is believed to stimulate vessel sprouting towards the wound^[2]. This is described in further detail in the secondary effects of NPWT. Microdeformation causes localized hypoxia that causes an increase in local vascularity. Factors known to affect the efficiency of microdeformation include the level of suction, the pore size and the consistency of the foam, the tissue being treated and the deformability of the surrounding tissues.

Fluid removal: Fluid accumulation in the extracellular space or edema, which often occurs in chronic wounds, inhibits healing by compressing local cells and tissues. For example, during wound healing in peripheral diabetic ulcers, cell proliferation occurs due to the intrinsic tension generated in the cells by the interaction of their

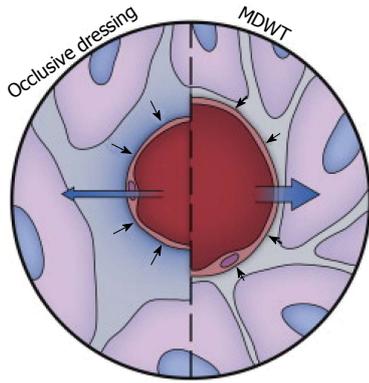


Figure 3 Removal of edema results in decreased hydrostatic compression of the capillaries and, hence, reduction of the required diffusion distance^[2]. The overall increase in tissue perfusion optimises wound healing. MDWT: Microdeformational wound therapy. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

cytoskeleton and the ECM^[14]. Fluid accumulation in the extracellular space elevates pressure in the interstitium and inhibits proliferation by decreasing the buildup of intrinsic tension. Since the fluids in the extracellular space communicate with the surface of the wound, vacuum application can remove fluid from the wound. Depending on the type of wound, significant amounts of fluid can be removed, as is the case with open abdominal and fasciotomy wounds. By removing fluid, the compression forces acting on the microvasculature allow increased blood flow and perfusion of the tissue (Figure 3)^[1]. The adhesive film dressing covering the wound is semipermeable and hence allows some air to enter the system preventing a fluid lock and enabling continuous fluid removal. Other devices have been designed to let a small amount of air into the system through a remote port.

NPWT is believed to affect the lymphatic system *via* two mechanisms. First, since edema is cleared *via* the lymphatic system, by removing fluid, NPWT concurrently reduces the burden on the lymphatic system. Second, NPWT promotes lymphatic drainage by inducing a gradual increase in the density of the lymphatics at the wound edges (Figure 4)^[15].

Alteration of the wound environment: When fluid is evacuated, electrolytes and proteins are removed that may stabilize osmotic and oncotic gradients at the wound surface^[14]. The foam and drape act as insulators maintaining a warm wound environment^[2]. The drape is semipermeable and helps maintain a sterile, moist environment by reducing wound contamination with microorganisms and minimizes water evaporation from the wound. In addition, NPWT can be more comfortable to patients by reducing the number of dressing changes. Special types of NPWT have been designed that serve to address specific issues in healing. For example, foam can be bound with antimicrobial silver or bioactive factors.

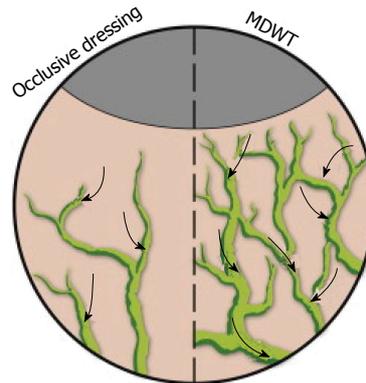


Figure 4 Lymphatic drainage is promoted through gradual increase in the density of the lymphatics at the wound edges^[2]. MDWT: Microdeformational wound therapy. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

Secondary effects

The four primary mechanisms of NPWT affect various wound healing processes including neurogenesis, hemostasis, angiogenesis, modulation of inflammation, cellular proliferation, differentiation, and migration, granulation formation, and alterations in bioburden.

Neurogenesis: MDWT has been linked to enhanced neural growth and neuropeptide expression through upregulation of neurotrophin nerve growth factor, substance P, and calcitonin gene-related peptide^[16]. Epinephrine and norepinephrine show a transient elevation, which is followed by a slower but more long-lasting elevation of substance P and neuropeptide Y (Figure 5). Neuropeptides are believed to be key homeostatic factors in the skin which play a role in the secondary effects of NPWT. The extent of neurogenesis has been directly linked to the level of microdeformation. In addition, intermittent suction results in greater neurogenesis than continuous MDWT^[16].

Hemostasis: NPWT is postulated to promote haemostasis *via* two mechanisms. First, the negative pressure is believed to constrict and occlude small blood vessels mechanically reducing hemorrhage. It should be noted that this constrictive effect persists even after negative pressure is discontinued. Second, compression due to negative pressure strongly apposes the dressings to the wound surface favoring clot formation by the gauze^[17]. In terms of the appropriate usage of the device, suction is applied when hemostasis is nearly complete, taking special care in patients with coagulopathies. In addition to the filler material, dressing, connecting tube and the vacuum pump, most systems also have a fluid collection canister which sounds an alarm when full, alerting clinicians of excessive blood loss^[18].

Angiogenesis and blood flow: MDWT treatment of chronic wounds results in increased microvessel density^[19]. Microdeformation causes temporary hypoperfusion to the

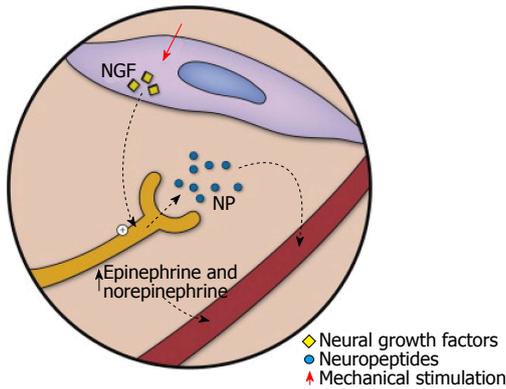


Figure 5 Microdeformational wound therapy promotes neurogenesis via upregulation of substance P, neurotrophin nerve growth factor, and calcitonin gene-related peptide^[2]. Plasma epinephrine and norepinephrine are also transiently elevated. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

wound edge resulting in localized hypoxia of the tissues, subsequent upregulation of hypoxia-inducible factor-1 α and in turn increased VEGF expression^[20]. Ultimately this leads to increased angiogenesis (Figure 6). Similar results to angiogenic response stimulation have been replicated in *in vitro* studies using intermittent MDWT^[21]. Furthermore, *in vivo* studies in patients have shown a difference between the initial and final stages of wound healing. Initially MDWT results in upregulation of angiogenin-2 (Ang-2) expression and downregulation of angiogenin-1 (Ang-1) expression, hence leading to decreased ratios of Ang-1/Ang-2. This favors destabilization and regression of microvessels leading to increased angiogenesis. In contrast, in the latter stages, Ang-1 is increased and the ratio of Ang-1/Ang-2 also increases. Phosphorylation of tyrosine kinase receptor-2 is activated, enhancing microvessel stabilization and promoting microvessel maturation^[22].

Using a deep tissue wound in a porcine model it was shown that a maximum fourfold increase in blood perfusion occurs when suction of 125 mmHg is applied to a PU foam. It was also shown that higher suction levels of 400 mmHg and above inhibit blood flow as the capillaries distort. In healthy human skin suction levels of up to 300 mmHg applied to a PU foam cause a fivefold increase of blood flow while suction on a Polyvinyl Alcohol (PVA) foam results in a threefold increase^[23].

Topical negative pressure has been shown to stimulate vessel proliferation and neo-angiogenesis. Topical negative pressure applied to chronic wounds of 16 patients (-125 mmHg) in preparation for reconstruction with free or pedicled flaps has been shown to considerably increase blood vessel density, reaching a maximum of approximately 200% in contrast to the vessel density prior to treatment^[24].

Modulation of inflammation: MDWT promotes active wound healing by simultaneously inducing inflammation while removing harmful components of inflammation such as infiltrating leucocytes, cytokines, and matrix metalloproteinases. Wounds treated with MDWT display

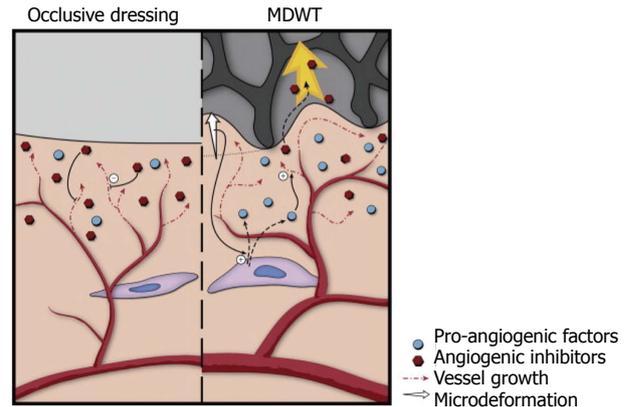


Figure 6 Angiogenesis is stimulated via several mechanisms, including microdeformation, upregulation of proangiogenic factors and removal of inhibitory factors^[2]. MDWT: Microdeformational wound therapy. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

increased cellularity of wound exudate with elevated erythrocytes and leukocytes, along with increased gene expression of leukocyte chemoattractants, such as CXCL5 and IL-8^[25].

Topical negative pressure with reticulated open cell foam (ROCF) has been shown to increase expression of various pro-healing anabolism-related genes. This includes increased expression of extracellular matrix genes resulting in increased production of the proteoglycans epiphygan and fibronectin. Other genes found to be upregulated include CD163 and macrophage scavenger receptor 1 which are involved in macrophage signalling^[26].

Cellular responses-proliferation, differentiation, and migration: Cells have long been known to undergo proliferation and division when exposed to mechanical stresses^[27]. Consequently, cells can be induced to undergo these cellular functions by exposure to dynamic physical inputs. MDWT is largely based on this principle; the tissue microdeformation stimulates cellular proliferation enhancing wound healing. This effect has been shown to upregulate angiogenesis and epithelialization in chronic wounds in a rabbit model^[28].

Studies using a diabetic mouse model found that application of short (6 h) intermittent MDWT results in increased expression of Ki-67, a marker for proliferation. The level of strain induced in the tissue by MDWT is the same level of strain required to induce cell proliferation *in vitro*^[13].

It is believed that isometric tension is heavily involved in promoting cellular proliferation^[29]. In the absence of isometric tension, growth factors and cell attachment to ECM proteins are essential but insufficient for cellular proliferation. Chronic wounds tend to lack the structural scaffolding needed for cell adherence and hence the development of isometric tension. Consequently cells undergo spherization and apoptosis. Suction applied

during MDWT is believed to generate the necessary forces within the tissues that enable isometric tension and hence cellular proliferation^[13].

In addition, gene ontology enrichment analysis of tissue treated with MDWT has shown increased epithelial cell migration in the tissue^[25]. Analysis of histologic penetration depth showed increased endothelial migration in tissues, whereas migration assays indicated enhanced dermal fibroblast migration^[30,31]. MDWT also enhances migration of resident skin mesenchymal cells and circulating progenitor cells into granulation tissue^[32].

Interestingly, although migration and proliferation of epithelial cells is increased by MDWT, differentiation is decreased. By downregulating keratin genes, such as KRT1 and KRT2, and major cornified envelope genes, such as annexin A9 and loricrin, MDWT inhibits keratinocyte differentiation^[25]. This decrease is believed to be due to the changes induced in the wound tissue matrix as mesenchymal stem cell lineages are highly specific to the mechanical characteristics of the ECM. For example, mesenchymal stem cell neurogenesis, myogenesis, and osteogenesis are guided along soft, stiffer and rigid matrices, respectively^[33]. Overall, MDWT is believed to promote wound healing by modulating cellular proliferation and migration, whilst inhibiting epidermal development and maturation^[25].

Granulation tissue formation: MDWT affects the proliferation stage of repair by inducing robust tissue granulation, cell proliferation, and blood vessel sprouting. Research on mast cell-deficient mice has shown that all three processes require early and continuous activation by mast cells^[34]. Collagen maturation is strictly dependent on mast cells in order to proliferate and remodel. Interestingly, MDWT application in mast cell-deficient mice, has no effect on collagen maturation, as expected, but does induce an increase in collagen production. In the control mice, both production and maturation were increased. Consequently, collagen production is not believed to be dependent on mast cells^[34].

MDWT relies heavily on mechanotransduction, whereby mechanical forces are transduced by cells into biological triggers for various processes, including gene expression^[35,36]. Mechanotransduction signalling in MDWT is a relatively new area of study. The current theory supports that molecules in the hypoxia pathway, such as nitric oxide, are involved^[18].

In vitro studies using tissue engineering bioreactors have held a dominant role in simulating the *in vivo* micromechanical environment and the foam-wound interface. For example, tissue analogues subjected to topical MDWT for 48 h in a 3-dimensional bioreactor model of the wound bed environment displayed that fibroblast cell bodies undergo morphological change, from elongated bipolar to thickened morphology. Another observation following this treatment was the presence of dense actin cortical structures^[37].

Research on a 3-dimensional fibrin matrix model found that MDWT increases cytochrome c oxidase levels,

energy charge, and the adenosine triphosphate (ATP)-adenosine diphosphate (ADP) ratio in fibroblasts. The increased energy was found to be utilized by healing biomechanisms. Two important factors required for collagen production during granulation formation, growth factor TGF- β and platelet-derived growth factors (PDGF) α and β , were also shown to increase with simultaneous application of subatmospheric pressure and a reticulated open-cell foam. In addition to upregulating collagen formation, PDGF α and β upregulate glycosaminoglycans and fibronectin synthesis in fibroblasts^[31]. Upregulation of fibroblast growth factor, TGF- β 1, Type I collagen α 1, and smooth muscle actin α 2 messenger RNA expression has also been observed in cells 48 h after been exposed to a suction, foam or perfusion bioreactor^[32].

The rate of granulation formation with NPWT therapy with a PU foam was measured in a porcine model by determining the decrease in wound volume over time. Increased rates of granulation formation were seen with continuous (63%) and intermittent (103%) application of suction. Continuous treatment is believed to be less effective than intermittent treatment because the cells in the wound become accommodated, and hence less responsive, to continuous physical forces^[38].

Intermittent suction application inactivates capillary autoregulation, hence increasing tissue perfusion, and enables the production of new cellular components by allowing time between cycles of cell division for the proliferating cells to rest. Continuous stimulation on the other hand is believed to switch off mitosis. Despite this, many clinicians prefer to use continuous treatment for the first 48 h, before switching to the intermittent mode, because it is better tolerated by patients^[39].

Alterations in bioburden: Changes in bioburden occur as a result of NPWT. However, studies related to this have produced mixed results. One study showed that NPWT results in decreased presence of non-fermentative gram-negative bacilli, but increased load of *Staphylococcus aureus*^[40]. Other studies found no significant difference in bacterial levels when foam dressings were used with and without suction. These experiments, however, used nonviable tissue and focused primarily on the effect of suction on bacterial load^[41]. It is believed that the decreased bacterial load occurs due to an interplay of multiple factors, not just due to the effect of suction^[42,43]. Experiments on foam material found high bacterial loads in sonicated foams, and very high polymicrobial bacterial loads in all foams studied^[44]. Porous PU foams on high suction (125 mmHg) were found to have a lower level of bacteria than PVA foams on lower suction. In addition, increased angiogenesis and blood perfusion may increase infection resistance by increasing inflow of oxygen in the wound tissue.

CLINICAL APPLICATIONS OF NPWT

NPWT has been used to treat wounds in numerous different anatomical locations, with different levels of

complexity and varying pathologies. The following section will review the evidence available on the application of NPWT in different wounds.

Open wounds

Basic applications of NPWT: At its most basic application, NPWT has been used in the management of open wounds, where the foam is directly applied to the wound bed. Common targets are poorly healing ulcers such as those caused by diabetes, venous or arterial pathologies and pressure necrosis.

More specifically, NPWT has been found to promote wound area reduction, wound bed granulation and clearance of microbial infection in diabetic foot ulcers^[45]. NPWT has been associated with a higher rate of limb salvage^[46]. Furthermore, treatment of diabetic, arterial and venous ulcers in high-risk patients using NPWT results in a higher rate of successful closure, with the greatest difference seen in venous ulcers^[47]. When NPWT is applied earlier on in the treatment these wounds display faster healing times^[47]. In nonoperative treatment of scleroderma ulcers, NPWT has been found to prevent digit amputation^[48]. In the treatment of pressure ulcers, NPWT has been shown to reduce the surface area, volume and depth of wounds, to enhance granulation and to decrease the likelihood of hospitalisation^[39,49,50]. Vowden K NPWT may also be effective in treating nonhealing deep-pressure ulcers covered by soft-necrotic tissue which require rapid formation of granulation tissue^[51]. It is important to be noted that NPWT needs to be used concurrently with disease specific treatment, for example medical treatment for vasculitis and pyoderma gangrenosum^[52].

In the treatment of surgical wounds, NPWT often acts as the pre-treatment before a skin flap or graft, or before secondary closure with NPWT. More specifically, in the excision of melanoma, NPWT enhances both functional (improves vascularity) and cosmetic (scar reduction) outcomes^[53], whereas in the postoperative treatment of lymphangioma in children it is believed to decrease the risk of recurrence and infection^[54].

Skin graft and dermal scaffold recipient site preparation: NPWT is often used to prepare a recipient site for skin grafts and dermal scaffolds. Large wounds, where granulation tissue spans the entire wound, can be rapidly closed with autologous skin grafting. One prospective RCT investigated the efficacy of NPWT prior to skin grafting in patients with acute traumatic wounds. NPWT improved total successful graft uptake, decreased regrafting, and required shorter lengths of hospital stay^[55].

Dermal scaffolds are often used in wounds where tendon or bone is exposed to induce vascularization of the wound bed in preparation for skin grafting^[56,57]. Concurrent treatment with NPWT and dermal scaffolds (Matriderm, Dr Suwelack Skin and Health Care AG, Billerbeck, Germany) enhances the contact between the wound surface and the scaffold and is believed to result in scars with higher elasticity and more natural skin pigmentation,

and a decreased occurrence of postoperative wound contamination one year post-operatively^[58].

Combination therapy: Various bioactive factors have been incorporated in NPWT to enhance efficacy.

Silver has been used in various wound dressings and has proven useful in burn care^[59]. In NPWT therapy, silver was added to the coating of the PU foam in order to decrease the bacterial load in the wound. In a goat model of complex infected orthopedic wounds, silver dressings placed beneath the negative pressure dressings resulted in a decrease in the bacterial load, most notably in the numbers of *S. aureus*^[60]. The MDWT foam can be modified to contain silver in order to act as an antimicrobial agent, as has been used in wound bed preparation for substantial split-thickness skin grafts (STSGs) to treat recalcitrant venous stasis ulcers^[61]. However, silver-infused dressings are not always indicated^[62].

A combined treatment of NPWT, platelet-rich plasma (PRP), STSGs and bilayered acellular matrix grafting was found to completely heal a large necrotizing fasciitis wound in a patient with diabetes^[63]. Furthermore, one study used a combination therapy of PRP and NPWT on patients with sternal osteomyelitis and sinus tract after thoracotomy. The treatment regimen was PRP gel on the day of surgery followed by continuous NPWT for 20 d. This combination therapy was found to shorten the sinus tract sealing time, wound healing time, and length of hospital stay. Secondary repair surgery was also avoided^[64].

Instillation therapy is the injection of fluid, such as normal saline, into the wound through a port on the NPWT connecting tube to enhance wound healing (Figure 7). This technique has been successfully applied in massive venous stasis wounds to reduce bacterial concentrations in the wound prior to STSG. A single-delivery-instillation system, whereby a series of intermittent cycles of MDWT was followed by a single injection of dilute sodium hypochlorate solution, appeared to create good wound bed preparation^[65].

In continuous-instillation MDWT a second port is connected to the continuous-drip system, which can allow continuous instillation of a fluid, for example insulin, to decrease the time required for wound healing^[66]. In full-thickness excisional wounds in a porcine model, NPWT with simultaneous irrigation with polyhexanide biguanide (PHMB) or saline showed improved wound healing with either irrigation solution when compared with NPWT alone^[67]. Other factors which can be instilled are dilute Betadine, doxycycline, phenytoin and lactoferrin. Further research is needed to investigate the efficacy of this concept^[68]. Instillation MDWT has been recommended in patients with multiple comorbidities with difficult wounds, although without high level of evidence to support these recommendations^[69].

Further examples of adjuvants include platelet gel, activated protein C, arginine-rich dietary supplements, and Manuka and *Leptospermum* honeys. Platelet gel, added to the wound bed following initial NPWT, has been used in

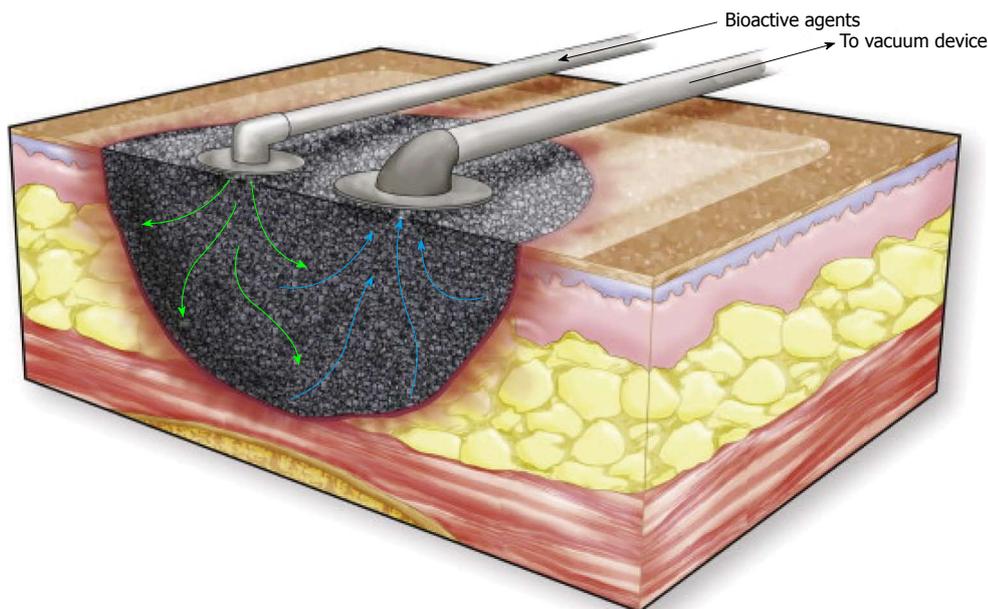


Figure 7 Instillation vacuum-assisted closure^[6]. Fluid, such as normal saline, is instilled into the wound through a port on the connecting tube to enhance wound healing. Reproduced from, Huang C, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; 51: 301-331. Copyright 2016 by Elsevier.

the treatment of a nonhealing ileocutaneous fistula leading to complete wound healing^[70]. In orthopedic wounds, activated protein C, an anticoagulant, was injected into the wound bed resulting in a decrease in the area and depth of the wound and an increase in granulation tissue formation^[71]. Studies on the use of arginine-rich dietary supplements, which are believed to enhance local circulation at the wound bed, have shown that treatment with the supplements results in complete healing of infection-induced wound dehiscence with only one month of treatment and no recurrence at the 6-mo follow-up^[72]. *Leptospermum* and Manuka honeys have been used in the treatment of a nonhealing postsurgical wound and an abdominal phlegmonous lesion, respectively^[73,74].

NPWT in burns: NPWT has been found to preserve perfusion in acute partial-thickness hand burns^[75]. It has also been used as a dressing over a dermal substitute in burn wounds, where it was believed to have no effect on graft adherence, but did improve long-term scar elasticity^[76]. When NPWT was used with porcine acellular dermal matrix (ADM) dressing as combination therapy in deep burn wounds it was found to decrease wound exudate and bacterial load and promote wound healing^[77]. NPWT has also been used in a patient with major third and fourth-degree high voltage electrical burns to enhance granulation tissue formation in preparation for skin grafting. Through the use of NPWT, major chronic soft tissue defects in the right leg were covered and amputation was avoided^[78].

Treatment of deep infected wounds: The efficacy of NPWT in deep wounds has been studied using soft tissue blast injuries in porcine models. In these models, NPWT

was found to decrease bacterial load, inhibit infection-induced tissue necrosis, and induce early initiation of granulation tissue formation^[79]. Research on humans has shown that NPWT is effective in controlling infection, specifically in thoracic and abdominal wounds.

NPWT in open fractures: NPWT has also been used in the treatment of open wounds with exposed bone or joints, where it is believed to keep the wound moist, warm and sterile by preventing external contamination. The rate of wound healing in open fractures has also been shown to be expedited with NPWT^[80]. Furthermore, the rate of deep infection in open tibial fractures is believed to be lower in NPWT treated wounds than in conventional treatment^[81]. Treatment with NPWT in an open left knee-joint wound, induced formation of granulated wound bed which fully covered the exposed bones and joint^[82]. NPWT is especially important in cases where free-flap transfer is contraindicated^[82,83]. NPWT in Gustilo grade III B open tibial fracture treatment helps reduce the size of the flap required but can also eliminate the need for a flap transfer all together. However, it should be noted that treatment with duration longer than 7 d was associated with higher likelihood of infection and amputation^[84]. NPWT in patients with grade III open fractures significantly reduced bacterial load at the wound site, as well as decreasing the risk of recurrent infection^[85].

Deep sternal wound infection: Use of NPWT as the first line of therapy in deep sternal wound infections (DSWI) has been shown to decrease rates of early reinfections, as well as reducing numbers of late chronic sternal infections and mortality^[86]. The length of hospital stay is also shortened by 1 wk^[87]. In methicillin-resistant DSWIs

following cardiovascular surgery, MDWT has been shown to decrease length of hospital stay, healing time, and infection recurrence^[88].

NPWT is believed to have the ability to stabilize the thoracic cage, improving hemodynamics and pulmonary status. NPWT in conjunction with a tissue flap can provide adequate control of infection preventing sepsis and hemodynamic instability^[89]. In the presence of poststernotomy osteomyelitis, adequate debridement and antibiotic therapy is still necessary with the use of NPWT^[90,91].

NPWT as an augmented surgical drain: NPWT enables improved drainage of the fluid which builds up in anatomical cavities or abscesses in deep wound infections. For example, application of NPWT in the treatment of a deep neck abscess reduced the need for open thoracotomy by preventing accumulation of purulent material^[92]. Concurrent open window thoracotomy and NPWT was found to eradicate local infection and hence control sepsis in postoperative or recurrent pleural empyema. In complex chest wall wounds NPWT inhibited empyema recurrence and enhanced lung expansion^[93]. A modified NPWT therapy has been used to treat deep cavitory defects, such as those caused by blast injuries or high-velocity projectiles. By connecting a superficial foam dressing to the surgical drain, deep cavitory defects can be converted into superficial ones. This modified therapy can apply suction deep in the wound cavity and result in a decrease in the dead space, reduced edema and lower risk of infection. In comparison to traditional VAC therapy, draining is enhanced, and the risk of deeper cavities closing off is minimized^[94]. VAC therapy has been successfully used in cases of Hidradenitis suppurativa where immediate primary closure was not possible secondary to the large size of the defect. An internal VAC was found to accelerate delayed closure and reduce the rate of recurrence in hidradenitis excisions^[95].

NPWT as biologic sampling device: NPWT is increasingly being used as a biologic sampling device, where mediastinal fluid is collected from the wound and cultured for microorganisms. In one study, NPWT was found to increase the rate of detection of microorganisms and was recommended as a replacement to traditional biologic sampling devices^[96]. On the other hand, microbiology of NPWT specimens in patients with prosthetic vascular graft infections was found to have limited diagnostic value, with anaerobe species being the most poorly identified in NPWT foam samples^[97]. Additional studies are, however, required before general conclusions on the efficacy of NPWT as a sampling device can be drawn.

Intra-abdominal NPWT: A further potential target for NPWT are deep intra-abdominal wound infections.

Specifically, in a case of acute necrotizing pancreatitis, placement of NPWT foam dressing in the opening created during lesser sac marsupialization in classical laparotomy

was found to accelerate wound closure, hence improving patient outcome^[98]. In addition, examination of wound secretions improved, abdominal compartment syndrome was prevented and care was overall simplified. NPWT has also been successfully used endoscopically in the treatment of rectal wall anastomotic disruptions^[99].

NPWT and gynecological laparotomy: NPWT has also been used as a prophylactic measure in laparotomy wounds in patients with gynecologic malignancies. The rate of wound complications was similar in patients who received traditional treatment and those receiving prophylactic NPWT dressing, despite those receiving NPWT having significantly higher BMIs^[100].

NPWT and soft-tissue sarcomas: NPWT has been shown to be safe and effective as an adjunct to wound closure in cases of wide tumor resection for soft-tissue sarcomas. Continuous suction, with pressures from -200 to -300 mmHg, was applied on the soft-tissue defects as preparation for wound closure. This treatment was found to decrease wound complications, such as post-operative infection and recurrence, while also reducing edema, draining exudate, and promoting granulation tissue formation^[101].

NPWT and congenital deformities: NPWT holds potential for the treatment of congenital deformities such as giant omphalocele^[102] or complex gastroschisis^[103]. However the efficacy for this has yet to be established and the treatment is not approved by the Food and Drug Administration (FDA). In pediatric patients, the granulation tissue response is often much more robust than in adults, often leading to more frequent dressing changes to avoid ingrowth into the interface material.

NPWT and digestive surgery: Most recently, there have been suggestions of using NPWT in digestive surgery, however this use is yet to be established. One previous pilot study investigated NPWT use following ileocecal resection in Crohn's disease and found that NPWT shortened length of hospital stay by 70%-80%^[104]. It is important to note that one major concern of NPWT use in digestive surgery is the development of enteric fistulas due to negative pressure^[105]. Prophylactic NPWT at the ostomy closure wound in patients with ulcerative colitis was found to be safe with no enterocutaneous fistula formation or postoperative bleeding. However, in this particular study, no effect on the duration of wound healing was observed and the prophylactic efficacy of NPWT could not be proven^[106]. Further studies are needed to prove the efficacy of NPWT in digestive surgery.

VARIATIONS OF THE TRADITIONAL NPWT SYSTEMS

Incisional NPWT

The literature supporting the use of NPWT over clean

incisions has mixed results. Clean, closed surgical incisions displayed decreased rates of seroma and hematoma formation in porcine models^[107]. Decreased rates of seroma, as well as haematoma formation have also been reported in post-bariatric patients receiving incisional NPWT^[108]. Topical NPWT applied to closed incisions also decreases the risk of infection^[109-112]. In total hip and knee arthroplasty, NPWT has been shown to be beneficial by decreasing excessive hospital stay and achieving a more predictable length of hospital stay. Wound complications, such as superficial wound infections and prolonged wound exudate, were also reduced^[113]. Furthermore, in total hip arthroplasty incisional NPWT was found to decrease the rate of postoperative seromas^[114], whereas in abdominal wall reconstruction it was found to reduce the incidence of incisional wound dehiscence^[114,115]. Incisional NPWT in the reconstructive surgery of poststernotomy mediastinitis was found to decrease the duration of required therapy, length of hospital stay, and failure of treatment^[116]. Peri-incisional lateral stress is reduced by approximately 50% following NPWT application and the directions of these stress vectors mimicked the distribution found in intact tissue^[117]. Evidence from this research has supported the development of systems such as Prevena™ Incision Management System (KCI, an Acelyty company, San Antonio, TX) which is specifically designed to be used in incisional wounds. Prevena™ has successfully been used in closed sternal incisions in cardiac patients where it has been shown to result in favorable outcomes within 30 d post-surgery^[118]. A recent literature review has recommended the use of incisional NPWT in all patients with high risk of developing surgical site occurrences and those undergoing a high-risk procedure or a procedure that would have morbid consequences if complications occurred^[119].

In contrast, VAC therapy in high risk patients with lower extremity and abdominal wound incisions had no significant effect on infection and dehiscence rates^[120]. It should be noted that one prospective analysis of 21 patients who received NPWT post primary knee arthroplasty found no benefit in wound healing with NPWT, with the only notable benefit being less wound leakage and better protection of the incisional site. This study was, however, limited by the small sample size and the results need to be validated by a larger prospective RCT^[121].

Skin graft immobilization

NPWT is used in STSGs in the place of a bolster, which is traditionally used to immobilize the graft by applying gentle pressure^[122,123]. NPWT stabilizes the graft, drains excess fluids, and promotes better contact for graft integration enhancing vascularisation^[122-125]. NPWT has been shown to decrease the risk of reoperation in cases of congested lower extremity pedicle and free flaps by decreasing venous insufficiency and tissue edema, promoting granulation and, hence, preventing further flap necrosis^[126]. Furthermore, NPWT decreases venous

congestion in random local flaps used in complex ankle wounds, hence decreasing the likelihood for ischemia and distal necrosis and enhancing their viability^[127]. NPWT is also used in the treatment of large back donor sites for head and neck free flaps where it is believed to decrease complications. NPWT has successfully been used in degloving injuries to immobilize skin grafts^[128,129], or as adjuvant treatment with a dermal regeneration template^[130].

CLINICAL CONSIDERATIONS

NPWT has been used in various wound types and its use in poorly healing wounds is FDA approved. The KCI VAC therapy system, which is widely used, lists acute, subacute, chronic, traumatic, dehisced wounds, ulcers, partial-thickness burns, grafts and flaps as indications for use in its manufacturer guidelines^[131].

Contraindications

Use of NPWT is contraindicated in untreated osteomyelitis, when necrotic tissue or malignancy is present in the wound, in nonenteric and unexplored fistulas, and when there is exposed vasculature, nerves, anastomotic sites, or organs^[107]. It should be noted that NPWT has been used successfully in contraindicated cases such as in cases of osteomyelitis^[91], exposed organs^[98], and exposed anastomotic sites^[132].

Numerous potential patient risk factors that require consideration have also been identified. NPWT is contraindicated in patients with high risk of bleeding or hemorrhage or those who are on chronic anticoagulation or antiplatelet treatment^[133]. In addition, the patient's body habitus needs to be considered.

Direct contact of exposed tendons, nerves, vasculature and organs with PU foam under vacuum forces can also result in complications. Hence, in cases where NPWT needs to be applied in close proximity to exposed structures, these structures can be covered with a nonadherent barrier layer such as petroleum gauze and nonadherent dressings^[134]. In addition, an isolation sterile bag has been described for intra-abdominal dressings in which the VAC foam is placed inside a sterile bag (3M SteriDrape Isolation Bag) whose surface has been perforated, allowing fluid drainage whilst simultaneously protecting the surrounding tissue^[97].

Possible complications

Complications that have been mentioned in the literature include infection and sepsis, foam retention in the wound, tissue adherence, bleeding, and pain (Figure 8). Although in the most serious cases bleeding and infection have led to death, these complications occur very rarely^[107]. Death associated with NPWT application at home or in long-term care facilities, is most commonly due to massive bleeding. Consequently, care needs to be exercised when selecting patients to apply NPWT at home, particularly in wounds of high risk for bleeding. In open abdominal



Figure 8 Infection and sepsis, foam retention in the wound, tissue adherence, bleeding, and pain^[2]. A: Treatment of L-1 partial paralysis and left ischial pressure ulcer with microdeformational wound therapy for 2 m; B: Discovery of retained foam; C: Wound healing following removal of foreign material and flap surgery. Courtesy of Brigham and Women's Hospital.

wounds the tension generated on the proximal bowel of the stoma during NPWT has been seen to cause stomal mucocutaneous dehiscence^[135].

The frequency of dressing changes can also affect treatment outcome. For example, in a case of NPWT treatment of a stage IV ischial pressure ulcer, spreading infections were masked, resulting in necrotizing fasciitis, as the dressing was changed at 5 d intervals^[136].

Special consideration should be taken when treating blast wounds with NPWT, as application has been linked to increased rates of sepsis^[137]. If applied before complete debridement is performed, it should be changed more frequently until the debridement is complete. Blast wounds have deep cavitory defects and the tail of the foam placed within the cavity has a higher chance of retention^[94]. Currently, most interface materials do not have any indicator that can be visualized on X-ray. Great care must therefore be taken when changing the interface material to ensure complete removal.

FUTURE PERSPECTIVES

Interface material

Various interfaces have been developed through the years. At the moment, reticulated open-pore foams have been the most carefully studied and are considered to be able to transmit suction over long distances and to induce tissue microdeformation. Furthermore, the pore size in the foam material is believed to be directly related to the level of granulation tissue formation, where larger

pores induce higher production of granulation tissue^[138]. Further research related to optimal interface materials for different clinical situations is necessary.

Optimal cycling

To date, research supports that short, intermittent NPWT induces a more robust tissue response in biological systems than continuous mechanical forces^[16]. However, in one study, continuous NPWT application and variable application (every 2 d for 4 h) were found to induce a similar granulation tissue response. Furthermore, very fast cycle times seem to decrease granulation tissue formation by causing damage to nascent granulation tissue^[8,139,140]. Intermittent therapy is also often not adhered to by patients because of patient discomfort. Additionally, optimal cycling varies with the type of wound. A chronic ulcer, for example, may be best treated with continuous suction throughout treatment, whereas an acute wound, may respond better to continuous suction for 48 h, followed by cycles of intermittent therapy^[135].

Optimal suction level

There has yet to be a definitive study that states the optimal NPWT suction level. A lower pressure is believed to be best for circumferential wounds and in cases where NPWT is used in conjunction with a free flap.

Adhesives

A limitation of the currently available NPWT devices is the inability to obtain a good seal at the edges of the device

which compromises the maintenance of suction. Further development of materials that enable better adhesion on curved and moist surfaces is necessary to allow use of the devices in difficult wounds.

CONCLUSION

NPWT may have proven successful as an adjunctive therapy in a wide variety of wounds. However, the currently available systems are still novel, and the number of high-level clinical studies investigating NPWT is lacking. More RCTs are needed to elucidate the details of NPWT efficacy, particularly in terms of its different indications and modalities. Overall, NPWT continues to hold great promise and with further research on the optimal parameters of its application this management option stands to continue to improve wound healing and patient care.

ACKNOWLEDGMENTS

We would like to acknowledge Mr. Christian Jones an MD candidate at the School of Clinical Medicine, University of Cambridge, Cambridge CB2 0SP United Kingdom for recording the Core Tip.

REFERENCES

- 1 **Argenta LC**, Morykwas MJ. Vacuum-assisted closure: a new method for wound control and treatment: clinical experience. *Ann Plast Surg* 1997; **38**: 563-576; discussion 577 [PMID: 9188971 DOI: 10.1097/0000637-199706000-00002]
- 2 **Huang C**, Leavitt T, Bayer LR, Orgill DP. Effect of negative pressure wound therapy on wound healing. *Curr Probl Surg* 2014; **51**: 301-331 [PMID: 24935079 DOI: 10.1067/j.cpsurg.2014.04.001]
- 3 **Malmsjö M**, Borgquist O. NPWT settings and dressing choices made easy. *Wounds International* 2010; **1**: 1-6
- 4 **Chariker ME**, Jeter KF, Tintle TE, Ottisford JE. Effective management of incisional and cutaneous fistulae with closed suction wound drainage. *Contemp Surg* 1989; **34**: 59-63
- 5 **Campbell PE**, Smith GS, Smith JM. Retrospective clinical evaluation of gauze-based negative pressure wound therapy. *Int Wound J* 2008; **5**: 280-286 [PMID: 18494633 DOI: 10.1111/j.1742-481X.2008.00485.x]
- 6 **Kairinos N**, Solomons M, Hudson DA. Negative-pressure wound therapy I: the paradox of negative-pressure wound therapy. *Plast Reconstr Surg* 2009; **123**: 589-598; discussion 599-600 [PMID: 19182617 DOI: 10.1097/PRS.0b013e3181956551]
- 7 **Kairinos N**, Solomons M, Hudson DA. The paradox of negative pressure wound therapy--in vitro studies. *J Plast Reconstr Aesthet Surg* 2010; **63**: 174-179 [PMID: 19036656 DOI: 10.1016/j.bjps.2008.08.037]
- 8 **Scherer SS**, Pietramaggiore G, Mathews JC, Prsa MJ, Huang S, Orgill DP. The mechanism of action of the vacuum-assisted closure device. *Plast Reconstr Surg* 2008; **122**: 786-797 [PMID: 18766042 DOI: 10.1097/PRS.0b013e31818237ac]
- 9 **Borgquist O**, Ingemansson R, Malmsjö M. The influence of low and high pressure levels during negative-pressure wound therapy on wound contraction and fluid evacuation. *Plast Reconstr Surg* 2011; **127**: 551-559 [PMID: 20966819 DOI: 10.1097/PRS.0b013e3181fed52a]
- 10 **Saxena V**, Hwang CW, Huang S, Eichbaum Q, Ingber D, Orgill DP. Vacuum-assisted closure: microdeformations of wounds and cell proliferation. *Plast Reconstr Surg* 2004; **114**: 1086-1096 [PMID: 15457017 DOI: 10.1097/01.prs.0000135338.78447.4e]
- 11 **Quinn TP**, Schlueter M, Soifer SJ, Gutierrez JA. Cyclic mechanical stretch induces VEGF and FGF-2 expression in pulmonary vascular smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol* 2002; **282**: L897-L903 [PMID: 11943652 DOI: 10.1152/ajplung.00044.2001]
- 12 **Rivlis I**, Milkiewicz M, Boyd P, Goldstein J, Brown MD, Egginton S, Hansen FM, Hudlicka O, Haas TL. Differential involvement of MMP-2 and VEGF during muscle stretch- versus shear stress-induced angiogenesis. *Am J Physiol Heart Circ Physiol* 2002; **283**: H1430-H1438 [PMID: 12234794 DOI: 10.1152/ajpheart.00082.2002]
- 13 **Urschel JD**, Scott PG, Williams HT. The effect of mechanical stress on soft and hard tissue repair; a review. *Br J Plast Surg* 1988; **41**: 182-186 [PMID: 3280056 DOI: 10.1016/0007-1226(88)90049-5]
- 14 **Orgill DP**, Manders EK, Sumpio BE, Lee RC, Attinger CE, Gurtner GC, Ehrlich HP. The mechanisms of action of vacuum assisted closure: more to learn. *Surgery* 2009; **146**: 40-51 [PMID: 19541009 DOI: 10.1016/j.surg.2009.02.002]
- 15 **Lancerotto L**, Bayer LR, Orgill DP. Mechanisms of action of microdeformational wound therapy. *Semin Cell Dev Biol* 2012; **23**: 987-992 [PMID: 23036531 DOI: 10.1016/j.semdb.2012.09.009]
- 16 **Younan G**, Ogawa R, Ramirez M, Helm D, Dastouri P, Orgill DP. Analysis of nerve and neuropeptide patterns in vacuum-assisted closure-treated diabetic murine wounds. *Plast Reconstr Surg* 2010; **126**: 87-96 [PMID: 20595860 DOI: 10.1097/PRS.0b013e3181da86d0]
- 17 **Kheirabadi BS**, Terrazas IB, Williams JF, Hanson MA, Dubick MA, Blackburne LH. Negative-pressure wound therapy: a hemostatic adjunct for control of coagulopathic hemorrhage in large soft tissue wounds. *J Trauma Acute Care Surg* 2012; **73**: 1188-1194 [PMID: 23117379 DOI: 10.1097/TA.0b013e31826f98ea]
- 18 **Saxena V**, Orgill D, Kohane I. A set of genes previously implicated in the hypoxia response might be an important modulator in the rat ear tissue response to mechanical stretch. *BMC Genomics* 2007; **8**: 430 [PMID: 18034909 DOI: 10.1186/1471-2164-8-430]
- 19 **Greene AK**, Puder M, Roy R, Arsenault D, Kwei S, Moses MA, Orgill DP. Microdeformational wound therapy: effects on angiogenesis and matrix metalloproteinases in chronic wounds of 3 debilitated patients. *Ann Plast Surg* 2006; **56**: 418-422 [PMID: 16557076 DOI: 10.1097/01.sap.0000202831.43294.02]
- 20 **Erba P**, Ogawa R, Ackermann M, Adini A, Miele LF, Dastouri P, Helm D, Mentzer SJ, D'Amato RJ, Murphy GF, Konerding MA, Orgill DP. Angiogenesis in wounds treated by microdeformational wound therapy. *Ann Surg* 2011; **253**: 402-409 [PMID: 21217515 DOI: 10.1097/SLA.0b013e31820563a8]
- 21 **Potter MJ**, Banwell P, Baldwin C, Clayton E, Irvine L, Linge C, Grobbelaar AO, Sanders R, Dye JF. In vitro optimisation of topical negative pressure regimens for angiogenesis into synthetic dermal replacements. *Burns* 2008; **34**: 164-174 [PMID: 18242874 DOI: 10.1016/j.burns.2007.06.020]
- 22 **Ma Z**, Shou K, Li Z, Jian C, Qi B, Yu A. Negative pressure wound therapy promotes vessel destabilization and maturation at various stages of wound healing and thus influences wound prognosis. *Exp Ther Med* 2016; **11**: 1307-1317 [PMID: 27073441 DOI: 10.3892/etm.2016.3083]
- 23 **Timmers MS**, Le Cessie S, Banwell P, Jukema GN. The effects of varying degrees of pressure delivered by negative-pressure wound therapy on skin perfusion. *Ann Plast Surg* 2005; **55**: 665-671 [PMID: 16327472 DOI: 10.1016/s1535-1513(08)70228-5]
- 24 **Malsiner CC**, Schmitz M, Horch RE, Keller AK, Leffler M. Vessel transformation in chronic wounds under topical negative pressure therapy: an immunohistochemical analysis. *Int Wound J* 2015; **12**: 501-509 [PMID: 24028468 DOI: 10.1111/iwj.12143]
- 25 **Nuutila K**, Siltanen A, Peura M, Harjula A, Nieminen T, Vuola J, Kankuri E, Aarnio P. Gene expression profiling of negative-pressure-treated skin graft donor site wounds. *Burns* 2013; **39**: 687-693 [PMID: 23141686 DOI: 10.1016/j.burns.2012.09.014]
- 26 **Leffler M**, Derrick KL, McNulty A, Malsiner C, Dragu A, Horch RE. Changes of anabolic processes at the cellular and molecular level in chronic wounds under topical negative pressure can be revealed by transcriptome analysis. *J Cell Mol Med* 2011; **15**: 1564-1571 [PMID: 20716124 DOI: 10.1111/j.1582-4934.2010.01147.x]
- 27 **Ingber DE**. The mechanochemical basis of cell and tissue regulation. *Mech Chem Biosyst* 2004; **1**: 53-68 [PMID: 16783946]
- 28 **Fabian TS**, Kaufman HJ, Lett ED, Thomas JB, Rawl DK, Lewis PL, Summitt JB, Merryman JI, Schaeffer TD, Sargent LA, Burns RP. The evaluation of subatmospheric pressure and hyperbaric oxygen in

- ischemic full-thickness wound healing. *Am Surg* 2000; **66**: 1136-1143 [PMID: 11149585]
- 29 **Huang S**, Ingber DE. The structural and mechanical complexity of cell-growth control. *Nat Cell Biol* 1999; **1**: E131-E138 [PMID: 10559956 DOI: 10.1038/13043]
- 30 **Baldwin C**, Potter M, Clayton E, Irvine L, Dye J. Topical negative pressure stimulates endothelial migration and proliferation: a suggested mechanism for improved integration of Integra. *Ann Plast Surg* 2009; **62**: 92-96 [PMID: 19131729 DOI: 10.1097/SAP.0b013e31817762fd]
- 31 **McNulty AK**, Schmidt M, Feeley T, Kieswetter K. Effects of negative pressure wound therapy on fibroblast viability, chemotactic signaling, and proliferation in a provisional wound (fibrin) matrix. *Wound Repair Regen* 2007; **15**: 838-846 [PMID: 18028132 DOI: 10.1111/j.1524-475X.2007.00287.x]
- 32 **Lu F**, Ogawa R, Nguyen DT, Chen B, Guo D, Helm DL, Zhan Q, Murphy GF, Orgill DP. Microdeformation of three-dimensional cultured fibroblasts induces gene expression and morphological changes. *Ann Plast Surg* 2011; **66**: 296-300 [PMID: 21233699 DOI: 10.1097/SAP.0b013e3181ea1e9b]
- 33 **Engler AJ**, Sen S, Sweeney HL, Discher DE. Matrix elasticity directs stem cell lineage specification. *Cell* 2006; **126**: 677-689 [PMID: 16923388 DOI: 10.1016/j.cell.2006.06.044]
- 34 **Younan GJ**, Heit YI, Dastouri P, Kekhiah H, Xing W, Gurish MF, Orgill DP. Mast cells are required in the proliferation and remodeling phases of microdeformational wound therapy. *Plast Reconstr Surg* 2011; **128**: 649e-658e [PMID: 22094766 DOI: 10.1097/PRS.0b013e318230e55d]
- 35 **Ingber DE**. Tensegrity-based mechanosensing from macro to micro. *Prog Biophys Mol Biol* 2008; **97**: 163-179 [PMID: 18406455 DOI: 10.1016/j.pbiomolbio.2008.02.005]
- 36 **Ingber DE**. Mechanobiology and diseases of mechanotransduction. *Ann Med* 2003; **35**: 564-577 [PMID: 14708967 DOI: 10.1080/07853890310016333]
- 37 **Wilkes RP**, McNulty AK, Feeley TD, Schmidt MA, Kieswetter K. Bioreactor for application of subatmospheric pressure to three-dimensional cell culture. *Tissue Eng* 2007; **13**: 3003-3010 [PMID: 17988192 DOI: 10.1089/ten.2007.0036]
- 38 **Morykwas MJ**, Faler BJ, Pearce DJ, Argenta LC. Effects of varying levels of subatmospheric pressure on the rate of granulation tissue formation in experimental wounds in swine. *Ann Plast Surg* 2001; **47**: 547-551 [PMID: 11716268 DOI: 10.1097/00000637-200111000-00013]
- 39 **Vowden K**. Conservative management of pressure ulcers. In: Banwell PE, Harding K, editors. Vacuum Assisted Closure TM Therapy: Science and Practice. London: MEP Ltd, 2006
- 40 **Mouës CM**, Vos MC, van den Bemd GJ, Stijnen T, Hovius SE. Bacterial load in relation to vacuum-assisted closure wound therapy: a prospective randomized trial. *Wound Repair Regen* 2004; **12**: 11-17 [PMID: 14974959 DOI: 10.1111/j.1067-1927.2004.12105.x]
- 41 **Assadian O**, Assadian A, Stadler M, Diab-Elschahawi M, Kramer A. Bacterial growth kinetic without the influence of the immune system using vacuum-assisted closure dressing with and without negative pressure in an in vitro wound model. *Int Wound J* 2010; **7**: 283-289 [PMID: 20550601 DOI: 10.1111/j.1742-481X.2010.00686.x]
- 42 **Morykwas MJ**, Argenta LC, Shelton-Brown EI, McGuirt W. Vacuum-assisted closure: a new method for wound control and treatment: animal studies and basic foundation. *Ann Plast Surg* 1997; **38**: 553-562 [PMID: 9188970 DOI: 10.1097/00000637-199706000-00001]
- 43 **Morykwas MJ**, Simpson J, Pungler K, Argenta A, Kremers L, Argenta J. Vacuum-assisted closure: state of basic research and physiologic foundation. *Plast Reconstr Surg* 2006; **117**: 121S-126S [PMID: 16799379 DOI: 10.1097/01.prs.0000225450.12593.12]
- 44 **Yusuf E**, Jordan X, Clauss M, Borens O, Mäder M, Trampuz A. High bacterial load in negative pressure wound therapy (NPWT) foams used in the treatment of chronic wounds. *Wound Repair Regen* 2013; **21**: 677-681 [PMID: 23927079 DOI: 10.1111/wrr.12088]
- 45 **Nather A**, Hong NY, Lin WK, Sakham JA. Effectiveness of bridge V.A.C. dressings in the treatment of diabetic foot ulcers. *Diabet Foot Ankle* 2011; **2**: 5893 [PMID: 22396825 DOI: 10.3402/dfa.v2i0.5893]
- 46 **Ulusal AE**, Sahin MS, Ulusal B, Cakmak G, Tuncay C. Negative pressure wound therapy in patients with diabetic foot. *Acta Orthop Traumatol Turc* 2011; **45**: 254-260 [PMID: 21908965 DOI: 10.3944/aott.2011.2283]
- 47 **Yao M**, Fabbri M, Hayashi H, Park N, Attala K, Gu G, French MA, Driver VR. A retrospective cohort study evaluating efficacy in high-risk patients with chronic lower extremity ulcers treated with negative pressure wound therapy. *Int Wound J* 2014; **11**: 483-488 [PMID: 23163962 DOI: 10.1111/j.1742-481X.2012.01113.x]
- 48 **Patel RM**, Nagle DJ. Nonoperative management of scleroderma of the hand with tadalafil and subatmospheric pressure wound therapy: case report. *J Hand Surg Am* 2012; **37**: 803-806 [PMID: 22305739 DOI: 10.1016/j.jhsa.2011.12.030]
- 49 **Joseph E**, Hamori CA, Bergman S, Roaf E, Swann NF, Anastasi GW. A prospective, randomized trial of vacuum-assisted closure versus standard therapy of chronic non healing wounds. *Wounds* 2000; **12**: 60-67
- 50 **Schwien T**, Gilbert J, Lang C. Pressure ulcer prevalence and the role of negative pressure wound therapy in home health quality outcomes. *Ostomy Wound Manage* 2005; **51**: 47-60 [PMID: 16230764]
- 51 **Nakayama M**. Applying negative pressure therapy to deep pressure ulcers covered by soft necrotic tissue. *Int Wound J* 2010; **7**: 160-166 [PMID: 20455957 DOI: 10.1111/j.1742-481X.2010.00667.x]
- 52 **Zutt M**, Haas E, Kruger U, Distler M, Neumann C. Successful use of vacuum-assisted closure therapy for leg ulcers caused by occluding vasculopathy and inflammatory vascular diseases—a case series. *Dermatology* 2007; **214**: 319-324 [PMID: 17460403 DOI: 10.1159/000100882]
- 53 **Oh BH**, Lee SH, Nam KA, Lee HB, Chung KY. Comparison of negative pressure wound therapy and secondary intention healing after excision of acral lentiginous melanoma on the foot. *Br J Dermatol* 2013; **168**: 333-338 [PMID: 23362968 DOI: 10.1111/bjd.12099]
- 54 **Katz MS**, Finck CM, Schwartz MZ, Moront ML, Prasad R, Timmapuri SJ, Arthur LG. Vacuum-assisted closure in the treatment of extensive lymphangiomas in children. *J Pediatr Surg* 2012; **47**: 367-370 [PMID: 22325392 DOI: 10.1016/j.jpedsurg.2011.11.030]
- 55 **Saaq M**, Hameed-Ud-Din MI, Chaudhery SM. Vacuum-assisted closure therapy as a pretreatment for split thickness skin grafts. *J Coll Physicians Surg Pak* 2010; **20**: 675-679 [PMID: 20943111]
- 56 **Heit YI**, Lancerotto L, Cortes R, Mesteri I, Ackermann M, Hollander R, Li Q, Douaiher J, Konerding MA, Orgill DP. Early kinetics of integration of collagen-glycosaminoglycan regenerative scaffolds in a diabetic mouse model. *Plast Reconstr Surg* 2013; **132**: 767e-776e [PMID: 24165628 DOI: 10.1097/PRS.0b013e3182a3c091]
- 57 **Kang GC**, Por YC, Tan BK. In vivo tissue engineering over wounds with exposed bone and tendon: Autologous dermal grafting and vacuum-assisted closure. *Ann Plast Surg* 2010; **65**: 70-73 [PMID: 20548234 DOI: 10.1097/SAP.0b013e3181a72f77]
- 58 **Bloemen MC**, van der Wal MB, Verhaegen PD, Nieuwenhuis MK, van Baar ME, van Zuijlen PP, Middelkoop E. Clinical effectiveness of dermal substitution in burns by topical negative pressure: a multicenter randomized controlled trial. *Wound Repair Regen* 2012; **20**: 797-805 [PMID: 23110478 DOI: 10.1111/j.1524-475X.2012.00845.x]
- 59 **Khundkar R**, Malic C, Burge T. Use of Acticoat dressings in burns: what is the evidence? *Burns* 2010; **36**: 751-758 [PMID: 20346592 DOI: 10.1016/j.burns.2009.04.008]
- 60 **Stinner DJ**, Waterman SM, Masini BD, Wenke JC. Silver dressings augment the ability of negative pressure wound therapy to reduce bacteria in a contaminated open fracture model. *J Trauma* 2011; **71**: S147-S150 [PMID: 21795872 DOI: 10.1097/TA.0b013e318221944a]
- 61 **Gerry R**, Kwei S, Bayer L, Breuing KH. Silver-impregnated vacuum-assisted closure in the treatment of recalcitrant venous stasis ulcers. *Ann Plast Surg* 2007; **59**: 58-62 [PMID: 17589262 DOI: 10.1097/01.sap.0000263420.70303.cc]
- 62 **Leaper D**. Appropriate use of silver dressings in wounds: international consensus document. *Int Wound J* 2012; **9**: 461-464 [PMID: 22994382 DOI: 10.1111/j.1742-481X.2012.01091.x]
- 63 **Deng W**, Boey J, Chen B, Byun S, Lew E, Liang Z, Armstrong DG. Platelet-rich plasma, bilayered acellular matrix grafting and negative pressure wound therapy in diabetic foot infection. *J Wound Care* 2016;

- 25: 393-397 [PMID: 27410393 DOI: 10.12968/jowc.2016.25.7.393]
- 64 **Hao D**, Feng G, Li T, Chu W, Chen Z, Li S, Zhang X, Zhao J, Zhao F. [Curative effects of platelet-rich plasma combined with negative-pressure wound therapy on sternal osteomyelitis and sinus tract after thoracotomy]. *Zhonghua Shaoshang Zazhi* 2016; **32**: 331-335 [PMID: 27321486]
- 65 **Raad W**, Lantis JC, Tyrie L, Gendics C, Todd G. Vacuum-assisted closure instill as a method of sterilizing massive venous stasis wounds prior to split thickness skin graft placement. *Int Wound J* 2010; **7**: 81-85 [PMID: 20529147 DOI: 10.1111/j.1742-481X.2010.00658.x]
- 66 **Scimeca CL**, Bharara M, Fisher TK, Kimbriel H, Mills JL, Armstrong DG. Novel use of insulin in continuous-instillation negative pressure wound therapy as "wound chemotherapy". *J Diabetes Sci Technol* 2010; **4**: 820-824 [PMID: 20663443 DOI: 10.1177/193229681000400408]
- 67 **Davis K**, Bills J, Barker J, Kim P, Lavery L. Simultaneous irrigation and negative pressure wound therapy enhances wound healing and reduces wound bioburden in a porcine model. *Wound Repair Regen* 2013; **21**: 869-875 [PMID: 24134060 DOI: 10.1111/wrr.12104]
- 68 **Kim PJ**, Attinger CE, Steinberg JS, Evans KK, Lehner B, Willy C, Lavery L, Wolvos T, Orgill D, Ennis W, Lantis J, Gabriel A, Schultz G. Negative-pressure wound therapy with instillation: international consensus guidelines. *Plast Reconstr Surg* 2013; **132**: 1569-1579 [PMID: 24005370 DOI: 10.1097/PRS.0b013e3182a80586]
- 69 **Kim PJ**, Attinger CE, Olawoye O, Crist BD, Gabriel A, Galiano RD, Gupta S, Lantis J, Lavery L, Lipsky BA, Teot L. Negative Pressure Wound Therapy With Instillation: Review of Evidence and Recommendations. *Wounds* 2015; **27**: S2-S19 [PMID: 26966814]
- 70 **Scala M**, Spagnolo F, Trapasso M, Strada P, Moresco L, Santi P. Association of vacuum-assisted closure and platelet gel for the definitive surgical repair of an enterocutaneous fistula: a case report. *In Vivo* 2012; **26**: 147-150 [PMID: 22210730]
- 71 **Wijewardena A**, Vandervord E, Lajevardi SS, Vandervord J, Jackson CJ. Combination of activated protein C and topical negative pressure rapidly regenerates granulation tissue over exposed bone to heal recalcitrant orthopedic wounds. *Int J Low Extrem Wounds* 2011; **10**: 146-151 [PMID: 21807809 DOI: 10.1177/1534734611417342]
- 72 **Masumoto K**, Nagata K, Oka Y, Kai H, Yamaguchi S, Wada M, Kusuda T, Hara T, Hirose S, Iwasaki A, Taguchi T. Successful treatment of an infected wound in infants by a combination of negative pressure wound therapy and arginine supplementation. *Nutrition* 2011; **27**: 1141-1145 [PMID: 21621390 DOI: 10.1016/j.nut.2011.01.006]
- 73 **Rudzka-Nowak A**, Łuczywek P, Gajos MJ, Piechota M. Application of manuka honey and GENADYNE A4 negative pressure wound therapy system in a 55-year-old woman with extensive phlegmonous and necrotic lesions in the abdominal integuments and lumbar region after traumatic rupture of the colon. *Med Sci Monit* 2010; **16**: CS138-CS142 [PMID: 20980964]
- 74 **Ganacias-Acuna EF**. Active Leptospermum honey and negative pressure wound therapy for nonhealing postsurgical wounds. *Ostomy Wound Manage* 2010; **56**: 10-12 [PMID: 20560236]
- 75 **Kamolz LP**, Andel H, Haslik W, Winter W, Meissl G, Frey M. Use of subatmospheric pressure therapy to prevent burn wound progression in human: first experiences. *Burns* 2004; **30**: 253-258 [PMID: 15082354 DOI: 10.1016/j.burns.2003.12.003]
- 76 **Honari S**, Gibran NS, Engrav LH. Three years' experience with 52 Integra (artificial skin) patients since FDA approval. *J Burn Care Rehabil* 2000; **21**: 190
- 77 **Liu W**, Li F, Chen X, Pan Q. [Clinical efficacy of negative-pressure wound therapy combined with porcine acellular dermal matrix for repairing deep burn wounds in limbs]. *Zhonghua Shaoshang Zazhi* 2016; **32**: 356-362 [PMID: 27321490]
- 78 **Tevanov I**, Enescu DM, Bălănescu R, Sterian G, Ulici A. Negative Pressure Wound Therapy (NPWT) to Treat Complex Defect of the Leg after Electrical Burn. *Chirurgia (Bucur)* 2016; **111**: 175-179 [PMID: 27172534]
- 79 **Li J**, Topaz M, Tan H, Li Y, Li W, Xun W, Yuan Y, Chen S, Li X. Treatment of infected soft tissue blast injury in swine by regulated negative pressure wound therapy. *Ann Surg* 2013; **257**: 335-344 [PMID: 23108116 DOI: 10.1097/SLA.0b013e318269d1ca]
- 80 **Arti H**, Khorami M, Ebrahimi-Nejad V. Comparison of negative pressure wound therapy (NPWT) & amp; conventional wound dressings in the open fracture wounds. *Pak J Med Sci* 2016; **32**: 65-69 [PMID: 27022347 DOI: 10.12669/pjms.321.8568]
- 81 **Blum ML**, Esser M, Richardson M, Paul E, Rosenfeldt FL. Negative pressure wound therapy reduces deep infection rate in open tibial fractures. *J Orthop Trauma* 2012; **26**: 499-505 [PMID: 22487900 DOI: 10.1097/BOT.0b013e31824133e3]
- 82 **Lee SY**, Niikura T, Miwa M, Sakai Y, Oe K, Fukazawa T, Kawakami Y, Kurosaka M. Negative pressure wound therapy for the treatment of infected wounds with exposed knee joint after patellar fracture. *Orthopedics* 2011; **34**: 211 [PMID: 21667911 DOI: 10.3928/01477447-7-20110427-27]
- 83 **Barnett TM**, Shilt JS. Use of vacuum-assisted closure and a dermal regeneration template as an alternative to flap reconstruction in pediatric grade IIIB open lower-extremity injuries. *Am J Orthop (Belle Mead NJ)* 2009; **38**: 301-305 [PMID: 19649348]
- 84 **Hou Z**, Irgit K, Strohecker KA, Matzko ME, Wingert NC, DeSantis JG, Smith WR. Delayed flap reconstruction with vacuum-assisted closure management of the open IIIB tibial fracture. *J Trauma* 2011; **71**: 1705-1708 [PMID: 22182878 DOI: 10.1097/TA.0b013e31822e2823]
- 85 **Krtička M**, Ira D, Nekuda V, Švancara J, Mašek M. [Effect of Negative Pressure Wound Therapy on Infectious Complications in Grade III Open Fractures]. *Acta Chir Orthop Traumatol Cech* 2016; **83**: 117-122 [PMID: 27167417]
- 86 **Steingrímsson S**, Gottfredsson M, Gudmundsdóttir I, Sjögren J, Guðbjartsson T. Negative-pressure wound therapy for deep sternal wound infections reduces the rate of surgical interventions for early re-infections. *Interact Cardiovasc Thorac Surg* 2012; **15**: 406-410 [PMID: 22691377 DOI: 10.1093/icvts/ivs254]
- 87 **Damiani G**, Pinnarelli L, Sommella L, Tocco MP, Marvulli M, Magrini P, Ricciardi W. Vacuum-assisted closure therapy for patients with infected sternal wounds: a meta-analysis of current evidence. *J Plast Reconstr Aesthet Surg* 2011; **64**: 1119-1123 [PMID: 21256819 DOI: 10.1016/j.bjps.2010.11.022]
- 88 **De Feo M**, Vicchio M, Nappi G, Cotrufo M. Role of vacuum in methicillin-resistant deep sternal wound infection. *Asian Cardiovasc Thorac Ann* 2010; **18**: 360-363 [PMID: 20719787 DOI: 10.1177/0218492310375854]
- 89 **Morisaki A**, Hosono M, Murakami T, Sakaguchi M, Suehiro Y, Nishimura S, Sakon Y, Yasumizu D, Kawase T, Shibata T. Effect of negative pressure wound therapy followed by tissue flaps for deep sternal wound infection after cardiovascular surgery: propensity score matching analysis. *Interact Cardiovasc Thorac Surg* 2016; **23**: 397-402 [PMID: 27199380 DOI: 10.1093/icvts/ivw141]
- 90 **Tocco MP**, Ballardini M, Masala M, Perozzi A. Post-sternotomy chronic osteomyelitis: is sternal resection always necessary? *Eur J Cardiothorac Surg* 2013; **43**: 715-721 [PMID: 22869252 DOI: 10.1093/ejcts/ezs449]
- 91 **Tocco MP**, Costantino A, Ballardini M, D'Andrea C, Masala M, Merico E, Mosillo L, Sordini P. Improved results of the vacuum assisted closure and Nitinol clips sternal closure after postoperative deep sternal wound infection. *Eur J Cardiothorac Surg* 2009; **35**: 833-838 [PMID: 19216084 DOI: 10.1016/j.ejcts.2008.12.036]
- 92 **Gallo O**, Deganello A, Meccariello G, Spina R, Peris A. Vacuum-assisted closure for managing neck abscesses involving the mediastinum. *Laryngoscope* 2012; **122**: 785-788 [PMID: 22252529 DOI: 10.1002/lary.22403]
- 93 **Sziklavari Z**, Grosser C, Neu R, Schemm R, Kortner A, Szöke T, Hofmann HS. Complex pleural empyema can be safely treated with vacuum-assisted closure. *J Cardiothorac Surg* 2011; **6**: 130 [PMID: 21978620 DOI: 10.1186/1749-8090-6-130]
- 94 **Rispoli DM**, Horne BR, Kryzak TJ, Richardson MW. Description of a technique for vacuum-assisted deep drains in the management of cavity defects and deep infections in devastating military and civilian trauma. *J Trauma* 2010; **68**: 1247-1252 [PMID: 20453774 DOI: 10.1097/TA.0b013e3181d3cc3c]
- 95 **Chen YE**, Gerstle T, Verma K, Treiser MD, Kimball AB, Orgill DP. Management of hidradenitis suppurativa wounds with an internal

- vacuum-assisted closure device. *Plast Reconstr Surg* 2014; **133**: 370e-377e [PMID: 24572882 DOI: 10.1097/PRS.0000000000000080]
- 96 **Cagnoni G**, Rimoldi SG, Pagani C, Savi C, Stefani F, Terzi R, Olivieri P, Tosi G, Parravicini C, Di Gregorio A, Antona C, Gismondo MR. Can Drainage Using a Negative-Pressure Wound Therapy Device Replace Traditional Sample Collection Methods? *Surg Infect (Larchmt)* 2016; **17**: 577-582 [PMID: 27348793 DOI: 10.1089/sur.2016.026]
- 97 **Scherrer AU**, Bloemberg G, Zbinden R, Zinkernagel AS, Fuchs C, Frauenfelder S, Rancic Z, Mayer D, Hasse B. Prosthetic Vascular Graft Infections: Bacterial Cultures from Negative-Pressure-Wound-Therapy Foams Do Not Improve Diagnostics. *J Clin Microbiol* 2016; **54**: 2190-2193 [PMID: 27252462 DOI: 10.1128/JCM.01102-16]
- 98 **Sermoneta D**, Di Mugno M, Spada PL, Lodoli C, Carvelli ME, Magalini SC, Cavicchioni C, Bocci MG, Martorelli F, Brizi MG, Gui D. Intra-abdominal vacuum-assisted closure (VAC) after necrosectomy for acute necrotising pancreatitis: preliminary experience. *Int Wound J* 2010; **7**: 525-530 [PMID: 20726923 DOI: 10.1111/j.1742-481X.2010.00727.x]
- 99 **Belmelan WA**. Vacuum assisted closure in coloproctology. *Tech Coloproctol* 2009; **13**: 261-263 [PMID: 19907919 DOI: 10.1007/s10151-009-0543-x]
- 100 **Lynam S**, Mark KS, Temkin SM. Primary Placement of Incisional Negative Pressure Wound Therapy at Time of Laparotomy for Gynecologic Malignancies. *Int J Gynecol Cancer* 2016; **26**: 1525-1529 [PMID: 27488215 DOI: 10.1097/IGC.0000000000000792]
- 101 **Chen YU**, Xu SF, Xu M, Yu XC. Use of negative pressure wound therapy as an adjunct to the treatment of extremity soft-tissue sarcoma with ulceration or impending ulceration. *Oncol Lett* 2016; **12**: 757-763 [PMID: 27347212 DOI: 10.3892/ol.2016.4654]
- 102 **Kilbride KE**, Cooney DR, Custer MD. Vacuum-assisted closure: a new method for treating patients with giant omphalocele. *J Pediatr Surg* 2006; **41**: 212-215 [PMID: 16410135 DOI: 10.1016/j.jpedsurg.2005.10.003]
- 103 **Choi WW**, McBride CA, Kimble RM. Negative pressure wound therapy in the management of neonates with complex gastroschisis. *Pediatr Surg Int* 2011; **27**: 907-911 [PMID: 21336610 DOI: 10.1007/s00383-011-2868-6]
- 104 **Pellino G**, Sciaudone G, Candilio G, Campitiello F, Selvaggi F, Canonico S. Effects of a new pocket device for negative pressure wound therapy on surgical wounds of patients affected with Crohn's disease: a pilot trial. *Surg Innov* 2014; **21**: 204-212 [PMID: 23883481 DOI: 10.1177/1553350613496906]
- 105 **Atema JJ**, Gans SL, Boermeester MA. Systematic review and meta-analysis of the open abdomen and temporary abdominal closure techniques in non-trauma patients. *World J Surg* 2015; **39**: 912-925 [PMID: 25446477 DOI: 10.1007/s00268-014-2883-6]
- 106 **Uchino M**, Hirose K, Bando T, Chohno T, Takesue Y, Ikeuchi H. Randomized Controlled Trial of Prophylactic Negative-Pressure Wound Therapy at Ostomy Closure for the Prevention of Delayed Wound Healing and Surgical Site Infection in Patients with Ulcerative Colitis. *Dig Surg* 2016; **33**: 449-454 [PMID: 27246708 DOI: 10.1159/000446550]
- 107 **Kilpadi DV**, Cunningham MR. Evaluation of closed incision management with negative pressure wound therapy (CIM): hematoma/seroma and involvement of the lymphatic system. *Wound Repair Regen* 2011; **19**: 588-596 [PMID: 22092797 DOI: 10.1111/j.1524-475X.2011.00714.x]
- 108 **Horch RE**. Incisional negative pressure wound therapy for high-risk wounds. *J Wound Care* 2015; **24**: 21-28 [PMID: 25853645 DOI: 10.12968/jowc.2015.24.Sup4b.21]
- 109 **Stannard JP**, Volgas DA, McGwin G, Stewart RL, Obremesky W, Moore T, Anglen JO. Incisional negative pressure wound therapy after high-risk lower extremity fractures. *J Orthop Trauma* 2012; **26**: 37-42 [PMID: 21804414 DOI: 10.1097/BOT.0b013e3182161e5]
- 110 **Zaidi A**, El-Masry S. Closed incision negative pressure therapy in high-risk general surgery patients following laparotomy: a retrospective study. *Colorectal Dis* 2016 Jul 15; Epub ahead of print [PMID: 27416813 DOI: 10.1111/codi.13458]
- 111 **Grauhan O**, Navasardyan A, Hofmann M, Müller P, Stein J, Hetzer R. Prevention of poststernotomy wound infections in obese patients by negative pressure wound therapy. *J Thorac Cardiovasc Surg* 2013; **145**: 1387-1392 [PMID: 23111014 DOI: 10.1016/j.jtcvs.2012.09.040]
- 112 **Semsarzadeh NN**, Tadisina KK, Maddox J, Chopra K, Singh DP. Closed Incision Negative-Pressure Therapy Is Associated with Decreased Surgical-Site Infections: A Meta-Analysis. *Plast Reconstr Surg* 2015; **136**: 592-602 [PMID: 26313829 DOI: 10.1097/PRS.0000000000001519]
- 113 **Karlakki SL**, Hamad AK, Whittall C, Graham NM, Banerjee RD, Kuiper JH. Incisional negative pressure wound therapy dressings (iNPWTd) in routine primary hip and knee arthroplasties: A randomised controlled trial. *Bone Joint Res* 2016; **5**: 328-337 [PMID: 27496913 DOI: 10.1302/2046-3758.58.BJR-2016-0022.R1]
- 114 **Pachowsky M**, Gusinde J, Klein A, Lehr S, Schulz-Drost S, Schlechtweg P, Pauser J, Gelse K, Brem MH. Negative pressure wound therapy to prevent seromas and treat surgical incisions after total hip arthroplasty. *Int Orthop* 2012; **36**: 719-722 [PMID: 21761149 DOI: 10.1007/s00264-011-1321-8]
- 115 **Condé-Green A**, Chung TL, Holton LH, Hui-Chou HG, Zhu Y, Wang H, Zahir H, Singh DP. Incisional negative-pressure wound therapy versus conventional dressings following abdominal wall reconstruction: a comparative study. *Ann Plast Surg* 2013; **71**: 394-397 [PMID: 22868327 DOI: 10.1097/SAP.0b013e31824c9073]
- 116 **Karlakki S**, Brem M, Giannini S, Khanduja V, Stannard J, Martin R. Incisional negative pressure wound therapy in reconstructive surgery of poststernotomy mediastinitis. *Bone Joint Res* 2013; **2**: 276, 284
- 117 **Wilkes RP**, Kilpad DV, Zhao Y, Kazala R, McNulty A. Closed incision management with negative pressure wound therapy (CIM): biomechanics. *Surg Innov* 2012; **19**: 67-75 [PMID: 21868417 DOI: 10.1177/1553350611414920]
- 118 **Reddy VS**. Use of Closed Incision Management with Negative Pressure Therapy for Complex Cardiac Patients. *Cureus* 2016; **8**: e506 [PMID: 27026831 DOI: 10.7759/cureus.506]
- 119 **Willy C**, Agarwal A, Andersen CA, Santis G, Gabriel A, Grauhan O, Guerra OM, Lipsky BA, Malas MB, Mathiesen LL, Singh DP, Reddy VS. Closed incision negative pressure therapy: international multidisciplinary consensus recommendations. *Int Wound J* 2016; Epub ahead of print [PMID: 27170231 DOI: 10.1111/iwj.12612]
- 120 **Masden D**, Goldstein J, Endara M, Xu K, Steinberg J, Attinger C. Negative pressure wound therapy for at-risk surgical closures in patients with multiple comorbidities: a prospective randomized controlled study. *Ann Surg* 2012; **255**: 1043-1047 [PMID: 22549748 DOI: 10.1097/SLA.0b013e3182501bae]
- 121 **Manoharan V**, Grant AL, Harris AC, Hazratwala K, Wilkinson MP, McEwen PJ. Closed Incision Negative Pressure Wound Therapy vs Conventional Dry Dressings After Primary Knee Arthroplasty: A Randomized Controlled Study. *J Arthroplasty* 2016; **31**: 2487-2494 [PMID: 27341973 DOI: 10.1016/j.arth.2016.04.016]
- 122 **Petkar KS**, Dhanraj P, Kingsly PM, Sreekar H, Lakshmanarao A, Lamba S, Shetty R, Zachariah JR. A prospective randomized controlled trial comparing negative pressure dressing and conventional dressing methods on split-thickness skin grafts in burned patients. *Burns* 2011; **37**: 925-929 [PMID: 21723044 DOI: 10.1016/j.burns.2011.05.013]
- 123 **Azzopardi EA**, Boyce DE, Dickson WA, Azzopardi E, Laing JH, Whitaker IS, Shokrollahi K. Application of topical negative pressure (vacuum-assisted closure) to split-thickness skin grafts: a structured evidence-based review. *Ann Plast Surg* 2013; **70**: 23-29 [PMID: 23249474 DOI: 10.1097/SAP.0b013e31826eab9e]
- 124 **Llanos S**, Danilla S, Barraza C, Armijo E, Piñeros JL, Quintas M, Searle S, Calderon W. Effectiveness of negative pressure closure in the integration of split thickness skin grafts: a randomized, double-masked, controlled trial. *Ann Surg* 2006; **244**: 700-705 [PMID: 17060762 DOI: 10.1097/01.sla.0000217745.56657.e5]
- 125 **Moisidis E**, Heath T, Boorer C, Ho K, Deva AK. A prospective, blinded, randomized, controlled clinical trial of topical negative pressure use in skin grafting. *Plast Reconstr Surg* 2004; **114**: 917-922 [PMID: 15468399 DOI: 10.1016/s1535-1513(08)70433-8]
- 126 **Vaienti L**, Gazzola R, Benanti E, Leone F, Marchesi A, Parodi PC, Riccio M. Failure by congestion of pedicled and free flaps for reconstruction of lower limbs after trauma: the role of negative-pressure wound therapy. *J Orthop Traumatol* 2013; **14**: 213-217

- [PMID: 23543100 DOI: 10.1007/s10195-013-0236-0]
- 127 **Goldstein JA**, Iorio ML, Brown B, Attinger CE. The use of negative pressure wound therapy for random local flaps at the ankle region. *J Foot Ankle Surg* 2010; **49**: 513-516 [PMID: 20801691 DOI: 10.1053/j.jfas.2010.07.001]
- 128 **Meara JG**, Guo L, Smith JD, Pribaz JJ, Breuing KH, Orgill DP. Vacuum-assisted closure in the treatment of degloving injuries. *Ann Plast Surg* 1999; **42**: 589-594 [PMID: 10382793 DOI: 10.1097/00000637-199906000-00002]
- 129 **Morris M**, Schreiber MA, Ham B. Novel management of closed degloving injuries. *J Trauma* 2009; **67**: E121-E123 [PMID: 19820564 DOI: 10.1097/TA.0b013e31803420be]
- 130 **Dini M**, Quercioli F, Mori A, Romano GF, Lee AQ, Agostini T. Vacuum-assisted closure, dermal regeneration template and degloved cryopreserved skin as useful tools in subtotal degloving of the lower limb. *Injury* 2012; **43**: 957-959 [PMID: 21492856 DOI: 10.1016/j.injury.2011.03.020]
- 131 V.A.C.s Therapy Forms and Brochures. [accessed 2016 Dec 8]. Available from: URL: <http://www.kci-medical.co.uk/UK-ENG/vacformsandbrochures>
- 132 **Weidenhagen R**, Gruetzner KU, Wiecken T, Spelsberg F, Jauch KW. Endoscopic vacuum-assisted closure of anastomotic leakage following anterior resection of the rectum: a new method. *Surg Endosc* 2008; **22**: 1818-1825 [PMID: 18095024 DOI: 10.1007/s00464-007-9706-x]
- 133 V.A.C. Therapy Indications and Contraindications. [accessed 2016 Dec 8]. Available from: URL: <http://www.kci-medical.sg/SG-ENG/indications>
- 134 **Mendez-Eastman S**. Guidelines for using negative pressure wound therapy. *Adv Skin Wound Care* 2001; **14**: 314-322; quiz 324-325 [PMID: 11794443]
- 135 **Steenvoorde P**, den Outer A, Neijenhuis P. Stomal mucocutaneous dehiscence as a complication of topical negative pressure used to treat an open abdomen: a case series. *Ostomy Wound Manage* 2009; **55**: 44-48 [PMID: 19564672]
- 136 **Citak M**, Backhaus M, Meindl R, Muhr G, Fehmer T. Rare complication after VAC-therapy in the treatment of deep sore ulcers in a paraplegic patient. *Arch Orthop Trauma Surg* 2010; **130**: 1511-1514 [PMID: 20306199 DOI: 10.1007/s00402-010-1091-6]
- 137 **Marsh DJ**, Abu-Sitta G, Patel H. The role of vacuum-assisted wound closure in blast injury. *Plast Reconstr Surg* 2007; **119**: 1978-1979 [PMID: 17440412 DOI: 10.1097/01.prs.0000259773.52889.68]
- 138 **Heit YI**, Dastouri P, Helm DL, Pietramaggiore G, Younan G, Erba P, Münster S, Orgill DP, Scherer SS. Foam pore size is a critical interface parameter of suction-based wound healing devices. *Plast Reconstr Surg* 2012; **129**: 589-597 [PMID: 22090246 DOI: 10.1097/PRS.0b013e3182402e89]
- 139 **Dastouri P**, Helm DL, Scherer SS, Pietramaggiore G, Younan G, Orgill DP. Waveform modulation of negative-pressure wound therapy in the murine model. *Plast Reconstr Surg* 2011; **127**: 1460-1466 [PMID: 21460654 DOI: 10.1097/PRS.0b013e31820a63cb]
- 140 **Malmsjö M**, Gustafsson L, Lindstedt S, Gesslein B, Ingemansson R. The effects of variable, intermittent, and continuous negative pressure wound therapy, using foam or gauze, on wound contraction, granulation tissue formation, and ingrowth into the wound filler. *Eplasty* 2012; **12**: e5 [PMID: 22292101]

P- Reviewer: Horch RE, Ichioka S, Lu SL **S- Editor:** Kong JX
L- Editor: A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



World Journal of *Dermatology*

World J Dermatol 2017 May 2; 6(2): 17-41



Editorial Board

2017-2020

The *World Journal of Dermatology* Editorial Board consists of 139 members, representing a team of worldwide experts in dermatology. They are from 39 countries, including Argentina (1), Australia (1), Austria (1), Brazil (1), Bulgaria (1), Canada (4), China (10), Croatia (1), Denmark (1), Egypt (1), Finland (1), France (3), Germany (5), Greece (4), Hungary (2), India (3), Iran (3), Israel (1), Italy (16), Japan (5), Malaysia (1), Malta (1), Mexico (4), Netherlands (3), Nigeria (2), Norway (1), Oman (1), Poland (2), Portugal (1), Romania (1), Saudi Arabia (1), Singapore (2), South Korea (8), Spain (8), Swaziland (2), Thailand (2), Turkey (6), United Kingdom (9), United States (19).

EDITOR-IN-CHIEF

Santosh K Katiyar, *Birmingham*

GUEST EDITORIAL BOARD MEMBERS

Tsong-Min Chang, *Tsichung*

Ching-Chi Chi, *Chiayi*

Jia-You Fang, *Taoyuan*

Sindy Hu, *Taipei*

Stephen Chu-Sung Hu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

María D Hermida, *Buenos Aires*



Australia

Ronald Sluyter, *Wollongong*



Austria

Iris Zalaudek, *Graz*



Brazil

Cidia Vasconcellos, *São Paulo*



Bulgaria

Georgi Tchernev, *Sofia*



Canada

Eleftherios P Diamandis, *Toronto*

Tim Lee, *Vancouver*

Gang Li, *Vancouver*

Kursad Turksen, *Ottawa*



China

Henry Hin Lee Chan, *Hong Kong*

Min Li, *Nanjing*

Cheng Tan, *Nanjing*

Guo-You Zhang, *Wenzhou*

Min Zheng, *Hangzhou*



Croatia

Mariastefania Antica, *Zagreb*



Denmark

Lars Iversen, *Aarhus*



Egypt

Moetaz El-Domyati, *Cairo*



Finland

Kari J Syrjänen, *Turku*



France

Guinot J Christiane, *Neully sur Seine*

Roger Mouawad, *Paris*

Stephane Rocchi, *Nice*



Germany

Martin Leverkus, *Mannheim*

Roderick AF MacLeod, *Braunschweig*

Markus Meissner, *Frankfurt*

Enno Schmidt, *Luebeck*

Peter Schroeder, *Dusseldorf*



Greece

Ioannis D Bassukas, *Ioannina*

Maria Dalamaga, *Athens*

Andreas Katsambas, *Athens*

Eleni Sotiriou, *Thessaloniki*



Hungary

Arpad Farkas, *Szeged*

Janos Fodor, *Budapest*



India

Sujoy Khan, *Kolkata*

Harsh Mohan, *Chandigarh*

Davinder Parsad, *Chandigarh*

**Iran**

Alireza Firooz, *Tehran*
 Mohammad R Namazi, *Shiraz*
 Afshin Sadighha, *Ilam*

**Israel**

Ronni Wolf, *Rehovo*

**Italy**

Giuseppe Argenziano, *Naples*
 Laura Atzori, *Cagliari*
 Ettore D Capoluongo, *Rome*
 Dott V Di Lernia, *Reggio Emilia*
 Paolo Fabbri, *Florence*
 Gabriella Fabbrocini, *Naples*
 Silvano Gallus, *Milan*
 Torello Lotti, *Firenze*
 Clelia Miracco, *Cosenza*
 Agnese Molinari, *Rome*
 Pierfrancesco Morganti, *Rome*
 Luigi Naldi, *Bergamo*
 Luca Negosanti, *Bologna*
 Raffaele Palmirotta, *Rome*
 Mario Santinami, *Milano*
 Riccarda Serri, *Milano*

**Japan**

Masutaka Furue, *Fukuoka*
 Fukumi Furukawa, *Wakayama*
 Mohammad Ghazizadeh, *Kawasaki*
 Yohei Tanaka, *Matsumoto*
 Toshiyuki Yamamoto, *Tokyo*

**Malaysia**

Felix Boon-Bin Yap, *Kuala Lumpur*

**Malta**

Michael J Boffa, *Floriana*

**Mexico**

Roberto G Arenas, *Mexico*
 Sergio A Cuevas-Covarrubias, *Mexico*
 Leopoldo Flores-Romo, *Mexico*
 Maria Bertha Torres-alvarez, *San Luis Potosi*

**Netherlands**

Rosalie M Luiten, *Amsterdam*

Arnold P Oranje, *Rotterdam*
 Arnold C Spek, *Amsterdam*

**Nigeria**

Maurice E Asuquo, *Calabar*
 Joseph I Ikechebelu, *Nnewi*

**Norway**

Andrej M Grijbovski, *Oslo*

**Oman**

Mohamed Mabruk, *Muscat*

**Poland**

Andrzej Grzybowski, *Poznan*
 Lidia Rudnicka, *Warsaw*

**Portugal**

Bruno Sarmento, *Porto*

**Romania**

Liana Manolache, *Bucharest*

**Saudi Arabia**

Feroze Kaliyadan, *Hofuf*

**Singapore**

Wei-Sheng Chong, *Singapore*
 Hong Liang Tey, *Singapore*

**South Korea**

Dong-Seok Kim, *Seoul*
 Chang Hoon Lee, *Seoul*
 Jongsung Lee, *Seongnam City*
 Chil Hwan Oh, *Seoul*
 Byung Soon Park, *Seoul*
 Myung-Geun Shin, *Hwasun*
 Jong-Hyuk Sung, *Seoul*
 Young Kwan Sung, *Daegu*

**Spain**

Agustin Alomar, *Barcelona*
 Salvador Arias-Santiago, *Granada*
 Juan G Gavín, *Vigo*
 Marcos A Gonzalez-Lopez, *Santander*

Ramon Grimalt, *Barcelona*
 Husein Husein-ElAhmed, *Granada*
 Ander Izeta, *San Sebastian*
 Marcela Del Rio, *Madrid*

**Switzerland**

Gunther FL Hofbauer, *Zurich*
 Alexander A Navarini, *Zurich*

**Thailand**

Chirayu U Auewarakul, *Bangkok*
 Viroj Wiwanitkit, *Bangkok*

**Turkey**

Berna Aksoy, *Kocaeli*
 Fatma Aydin, *Samsun*
 Cem Dane, *Istanbul*
 Sibel Dogan, *Istanbul*
 Aylin T Ermertcan, *Manisa*
 Ozlem Su, *Istanbul*

**United Kingdom**

Theodoros Dimitroulas, *Dudley*
 Bernhard F Gibbs, *Chatham Maritime*
 Evmorfia Ladoyanni, *Stourbridge*
 Mark R Nelson, *London*
 Adrian V Pace, *Dudley*
 Anthony B Paul, *London*
 Sam Shuster, *Woodbridge*
 Olga Tura, *Edinburgh*
 Indre Verpetinske, *Stourbridge*

**United States**

Jeremy S Bordeaux, *Cleveland*
 Robert F Diegelmann, *Richmond*
 Q Ping Dou, *Detroit*
 Zeev Estrov, *Houston*
 Vincent Falanga, *Providence*
 Miranda A Farage, *Cincinnati*
 Markus H Frank, *Boston*
 W Scott Goebel, *Indianapolis*
 Dan-Ning Hu, *New York*
 Amor Khachemoune, *Brooklyn*
 Arash Kimyai-Asadi, *Houston*
 Michael S Kolodney, *Torrance*
 Feng Liu, *Chapel Hill*
 Senthamil R Selvan, *San Diego*
 Lei Shi, *Fort Worth*
 Animesh A Sinha, *East Lansing*
 Jeffrey M Weinberg, *New York*
 John A Zic, *Nashville*



REVIEW

- 17 Review of the initial treatment and avoidance of scald injuries
Bourdon RT, Nelson-Cheeseman BB, Abraham JP

MINIREVIEWS

- 27 Cutaneous implications of essential oils
Vangipuram R, Mask-Bull L, Kim SJ

ORIGINAL ARTICLE

Retrospective Study

- 32 Use of a selective enzymatic debridement agent (Nexobrid®) for wound management: Learning curve
Palao R, Aguilera-Sález J, Serracanta J, Collado JM, Dos Santos BP, Barret JP

ABOUT COVER

Editorial Board Member of *World Journal of Dermatology*, Liana Manolache, MD, PhD, Department of Dermatology, Institution of Dali Medical, Bucharest 060816, Romania

AIM AND SCOPE

World Journal of Dermatology (*World J Dermatol*, *WJD*, online ISSN 2218-6190, DOI: 10.5314), is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of dermatology. *WJD* covers fungal diseases, dermatitis and eczema, urticarial diseases, drug eruptions, pruritus, erythroderma desquamativum, connective tissue diseases, bullous skin diseases, vascular skin diseases, skin appendage diseases, pigmentary diseases, genetic diseases, nutritional and metabolic disorders, tumors, sexually transmitted diseases, AIDS, traditional medicine, integrated Chinese and Western medicine, evidence-based medicine, epidemiology and nursing. The journal also publishes original articles and reviews that report the results of applied and basic research in fields related to dermatology, such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Dermatology is now indexed in China National Knowledge Infrastructure (CNKI).

FLYLEAF

I-II Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xin-Xia Song*

NAME OF JOURNAL
World Journal of Dermatology

ISSN
 ISSN 2218-6190 (online)

LAUNCH DATE
 June 2, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Santosh K Katiyar, PhD, Professor, Department of Dermatology, University of Alabama at Birmingham, Birmingham, AL 35294, United States

EDITORIAL BOARD MEMBERS
 All editorial board members resources online at <http://www.wjgnet.com/2218-6190/editorialboard.htm>

EDITORIAL OFFICE
 Fang-Fang Ji, Director

World Journal of Dermatology
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLICATION DATE
 May 2, 2017

COPYRIGHT

© 2017 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION

<http://www.f6publishing.com>

Review of the initial treatment and avoidance of scald injuries

Ryan T Bourdon, Brittany B Nelson-Cheeseman, John P Abraham

Ryan T Bourdon, Emergency Department, Regions Hospital, St. Paul, MN 55101, United States

Brittany B Nelson-Cheeseman, John P Abraham, School of Engineering, University of St. Thomas, St. Paul, MN 55105-1079, United States

Author contributions: Bourdon RT, Nelson-Cheeseman BB and Abraham JP contributed equally to this work; Abraham JP contributed to conceiving this paper and writing portions of this paper; Bourdon RT contributed to writing portions of this paper and reviewing the manuscript; Nelson-Cheeseman BB contributed to writing portions of this paper and reviewing the manuscript.

Conflict-of-interest statement: The authors declare no conflicts. John Abraham has served on burn injury court cases in the past.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Dr. John P Abraham, School of Engineering, University of St. Thomas, 2115 Summit Ave, St. Paul, MN 55105-1079, United States. jpabraham@stthomas.edu
Telephone: +1-651-9625766

Received: August 23, 2016

Peer-review started: August 25, 2016

First decision: September 27, 2016

Revised: January 10, 2017

Accepted: February 8, 2017

Article in press: February 10, 2017

Published online: May 2, 2017

Abstract

Scald injuries, which describe burns to living tissue from

hot liquids, are a very common injury that occur across geographical, social, economic, and national boundaries. Despite their ubiquitous nature, a complete understanding of the conditions which are required to cause scald burns is not yet available. In addition, clear guidance to medical practitioners is available through various guidelines however in actual situations, the extent of the burn is not fully known and this lack of knowledge complicates care. Here, a comprehensive review is made of the available knowledge of temperatures and scald durations which lead to skin-burn injuries. The range of volumes and liquid temperatures are typical of those found in heated consumer beverages. This review can help medical practitioners design initial treatment protocols and can be used by manufacturers of hot-liquid products to avoid the most severe burns. Next, within the context of this ability to quantify burn depths, a review of current burn treatment guidelines is given. Included in this review is a visual recognition of the extent of burns into the dermal layer as well as decision guidelines for selection of patients which would benefit from referral to a dedicated burn center. It is hoped that by bringing together both the quantified burn-depth information and current treatment guidelines, this review can be used as a resource for persons in the medical, manufacturing, beverage service, and other industries to reduce the human impact of scald injuries.

Key words: Scald injury; Skin burns; Biological heating; Hot beverages; Burn depth

© The Author(s) 2017. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: This paper presents a concise summary that relates hot-beverage spills to burn injury risk. Not only can this paper be used to predict the depth of burn injuries, but it can also show how service temperature and cooling time can be set to reduce the threat of injury. Results are presented in simple to use tables and graphs for ease to medical practitioners.

Bourdon RT, Nelson-Cheeseman BB, Abraham JP. Review of the initial treatment and avoidance of scald injuries. *World J Dermatol* 2017; 6(2): 17-26 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i2/17.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i2.17>

INTRODUCTION

Burn injuries are a common type of injury that can occur in many situations around the globe. Within the category of burn injuries, scald wounds caused by hot liquids are among the most common. Scald injuries can occur in kitchens, baths, industrial and manufacturing environments, restaurants, and other locations. The extent of scald injuries can vary from mild to death causing. Mild burns are almost always treated without medical attention, while extreme burn injuries often result in referral to dedicated burn centers. Because of the large variation in harm caused by burns, it is important to create environments which lessen burn injuries. It is also important for medical responders to quickly and correctly categorize a burn so that appropriate treatment is initiated.

Scald burns, which here refer to any thermal injury caused by heated liquid, can occur anywhere on exposed body; however, they are most common on the skin. Since the vast majority of scalds are skin burns, they will be the sole focus of this review.

The severity of a skin burn is quantified by two measurements. The first is the depth of the burn into the tissue. The second measure is an estimation of the size of the surface area involved. Both of these measures will now be discussed.

BURN CLASSIFICATION

Burn depth

Both within the medical community as well as with the general public, the degree classification is most often used to describe burn depth. Within this classification, the description first-degree burn describes burns that kill tissue only within the outermost layer of skin (the epidermis). As a consequence, the burns generally heal quickly and without medical attention. The injuries may be painful and result in hyperemia and flaking of the necrosed epidermis following the incident.

Second-degree burns pass through the epidermal layer and into the dermis. They lead to thermal necrosis of tissue within that layer and consequently damage the skin structures which are housed there, such as hair follicles, sweat glands, capillaries, among others.

Third-degree burns pass through the dermis and enter the underlying hypodermis tissue and possibly into muscle. Since the burn completely destroys the dermis, the functions of this layer are also halted (such as blood flow and sensation). Consequently, the skin is ischemic. While it may appear hyperemic, when the

skin is compressed, reperfusion of the tissue is very slow to occur if at all. The pain associated with third-degree burns is often less than that of second- or first-degree burns because the innervation to the tissue is compromised. For third-degree burns, slow healing occurs and medical attention such as surgical excision and grating is required. If large parts of the body are burned, shock, hypothermia, and infection can occur^[1-3].

Within the academic and bioheat transfer community, different descriptions are used. In this community, superficial burns are the equivalent to burns of first degree. Superficial-partial-thickness burns are those that pass through the epidermis and into the external part of the dermal layer - generally not beyond the dermal mid-plane. Next, deep-partial-thickness burns pass through the dermal midplane and into the lower half of this layer. Finally, full-thickness burns are essentially the same as third-degree burns - they extend through the dermal layer and into underlying tissue. As with third-degree burns, they often require skin grafting^[4-6]. The primary difference between the two modes of burn classification is that in the latter, a separate accounting is made for burns that either are confined to the outer half of the dermis and those that pass into the lower half of the dermal layer.

Regardless of the preferred nomenclature, it is a challenge to visually assess burn depth. Many times, the visual estimation is not accurate^[5,7-10]. A summary image is provided in Figure 1 to bring together the various burn extents which have already been discussed in the accompanying text.

Burn area

Total burn surface area (TBSA) is also an important measure of injury. TBSA is most commonly reported as a % of the body area that a burn covers. With this method, the body is subdivided into major regions (head, torso, legs, arms, etc.). Each region is allocated a numerical value which is a typical percentage of the region compared to the total body surface. An example of such a regional breakdown is provided in Figure 2. Since many of the body regions are multiples of 9%, this regional breakdown is often referred to as the Rule of Nines. The Rule of Nines is endorsed by both the American Burn Association and the European Practice Guidelines for Burn Care. While other burn area criteria have been studied, the widespread current usage of this metric motivated its inclusion here.

More information will be provided in a later section of this report related to decision making of medical professionals. First, however, quantitative information on the depth of the burn will be provided.

Prediction of burn depth

Burn depth studies have been performed since the late 1940s. Pioneering work^[11-14] quantified the complex temperature-time relationship required for burns on porcine and human skin. It was discovered that the physiologic

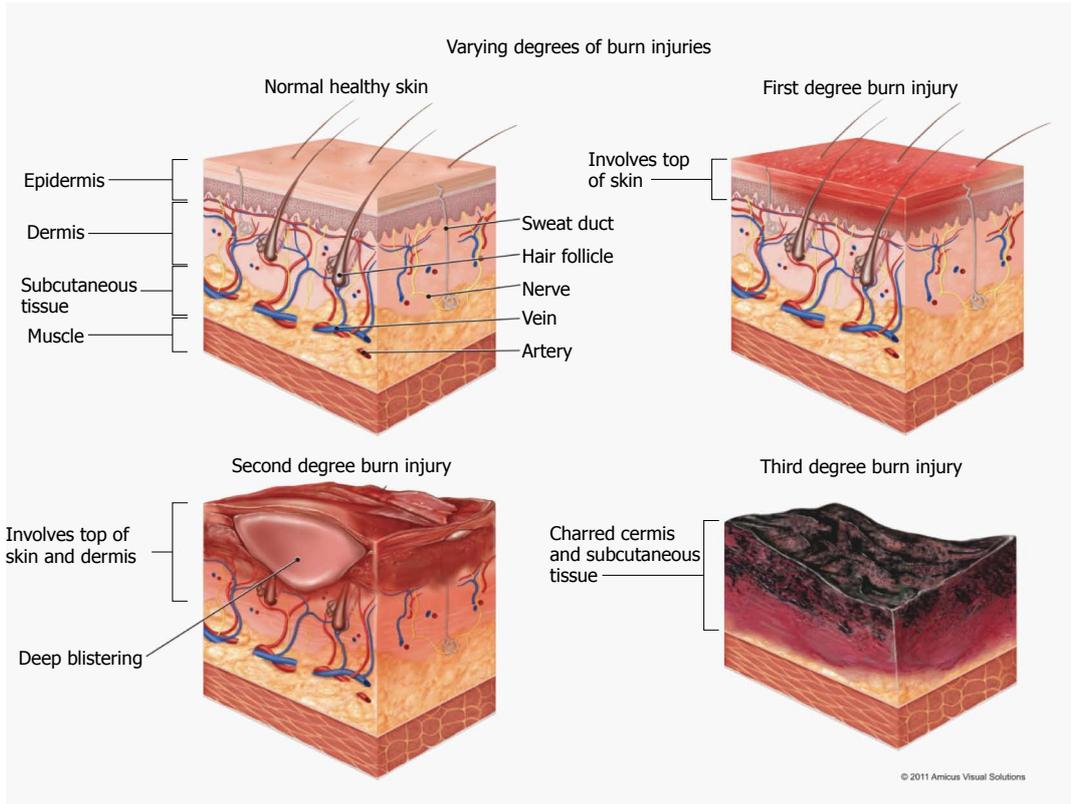


Figure 1 Illustration of burn depths and characteristics (courtesy of Amicus Medical Images).

| Rule of nines for adult body surface burns | |
|--|-----|
| Head and neck equal | 9% |
| Anterior trunk equals | 18% |
| Posterior trunk equals | 18% |
| Upper extremities (each 9%) | 18% |
| Lower extremities (each 18%) | 36% |
| Perineum | 1% |

| Rule of nines for small child body surface | |
|--|-----|
| Head and neck equal | 18% |
| Anterior trunk equals | 18% |
| Posterior trunk equals | 18% |
| Upper extremities (each 9%) | 18% |
| Lower extremities (each 13.5%) | 27% |
| Perineum | 1% |

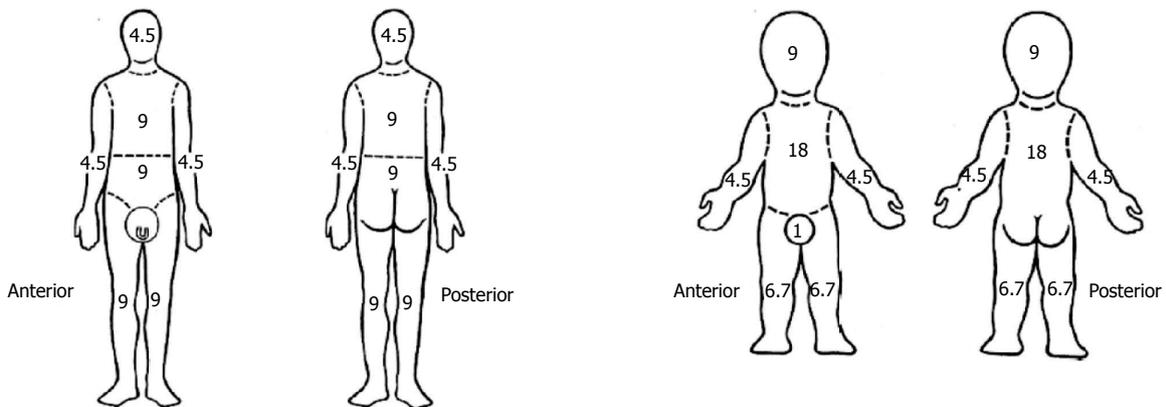


Figure 2 The Rule of Nines for adults and children (Courtesy of Brookside Associates).

response to elevated temperatures was complex and that for small increases in exposure temperature, large changes in burn injury rates were observed. The outcome of this

work was a quantification of the rate of cellular damage and the amount of viable cells still within the heated zone which is expressed mathematically as:

Table 1 Injury parameters for skin tissue

| Ref. | A (1/s) | Ea (J/kmol) |
|---------|------------------|-----------------|
| [14,23] | 3.10e98 | 6.28e08 |
| [25] | 2.90e37 | 2.44e08 |
| [26] | 9.09e37 | 2.49e08 |
| [27] | 4.33e64, T < 50 | 24.19e8, T < 50 |
| | 9.39e104, T > 50 | 6.70e8, T > 50 |
| [13,24] | 3.10e98 | 6.28e08 |
| [28] | 2.19e124, T < 50 | 7.78e8, T < 50 |
| | 1.82e51, T > 50 | 3.25e8, T > 50 |
| [29] | 1.43e72 | 4.57e08 |
| [29] | 2.86e69 | 4.61e08 |
| [29] | 4.32e54, T < 50 | 4.16e8, T < 50 |
| | 9.39e104, T < 60 | 6.65e8, T < 60 |
| [30] | 3.1e98, T < 55 | 6.27e8, T < 55 |
| | 5e45, T > 55 | 2.96e8, T > 55 |
| [31] | 2.19e124, T < 50 | 7.82e8, T < 50 |
| | 1.82e51, T > 50 | 3.27e8, T > 50 |
| [27] | 4.33e64, T < 50 | 4.18e8, T < 50 |
| | 9.39e104, T > 50 | 6.69e8, T > 50 |

$$\text{Damage index} = \Omega = \ln [c(0)/c(t)] = \int Ae^{-(Ea/RT)} dt \quad (1)$$

Here, *C* represents the concentration of viable cells either initially or at some time *t*. *A* is the frequency factor (1/s), *E_a* is the activation energy (J/mol), *R* is the universal gas constant (8.3143 J/mol-K), *T* is the temperature in Kelvin, *e* is the constant 2.718, and *t* is the time (s)^[15-31]. The fitted terms are obtained through experimental correlation, and they depend on the tissue type and body location. To illustrate the wide-ranging values of these parameters, Table 1 has been prepared.

The prediction of injury [through Equation (1)] is achieved by solving for the conduction of heat through perfused tissue using the Pennes model^[32] which is expressed in Equation (2).

$$(\rho C)_t (\partial T_t / \partial t) = k (\partial^2 T_t / \partial x^2) + S_{met} + (\rho C)_b \omega (T_b - T_t) \quad (2)$$

The calculation method outlined here has a long history of use and has been corroborated with experimental results. Results from those works have been adopted to predict skin burns using modern computational techniques^[33-57]. An outcome of these efforts is the ability to predict the distribution of an injury parameter throughout the various layers of tissue that have been heated. Values of are taken to correspond to completely necrosed tissue as they generally represent tissue which has too few viable cells to regenerate.

In the present study, focus is given on a subclass of scald burns - those caused by hot beverages. While the spill scenario can vary dramatically (volume of liquid spilled, temperature of liquid, spill pattern, presence or absence of clothing, speed of spill, thickness of skin, ability of victim to remove heat and apply cooling duration of hot liquid cooling prior to spill, among others), it is possible to generalize the results in a way that can provide meaningful information to treating physicians. To enable the generalization, the following ranges of parameters were used in the experiments.

Volume of liquid spilled: 8-16oz (237-473 mL).

Temperature of liquid spilled: 158 °F-203 °F (70 °C-95 °C).

Type of clothing: 1 layer of a cotton shirt.

Spills of cups that had caps as well as cups without caps.

Other parameters, such as the constitution of the cup, the presence or absence of an insulating sleeve, etc. had a very small impact on the results and so for simplicity are not included here^[46].

The experiments, which were carried out involving spills on living human tissue and on a tissue surrogate are described in^[46] and are not discussed in detail here. The experiments were carried out in typical ambient conditions of about 20 °C. The temperature during the cooling period for an 8 ounce (237 mL) beverages without a protective cover are shown below in Table 2. Corresponding information for covered beverages are provided in Table 3. These tables are followed immediately by information for larger volume beverages (16 ounces) both with and without protective caps (Tables 4 and 5).

The calculations were continued until the temperatures within the tissue reduced to levels which would no longer cause injury. Interested readers are invited to references^[46] and which discuss both the cooling effect^[56], the impact of rapid cooling after a burn, and the duration of time needed to bring tissue temperatures to safe levels.

From the experiments and calculations completed^[46], verified by a simplified model^[56] and by the comparison between models and physical observations which were contained therein, it is possible to create a visual reference which allows the estimation of burn risk caused by hot-liquid spills. The reference is provided in Figure 3.

The information presented in Figure 3 and the preceding tables can be brought together in a simple to use manner as seen in Tables 6-9. These tables list the initial beverage temperature, the volume, and the cooling time which should occur to bring the beverage to a low enough temperature so that a threshold mid-dermal burn may not occur. Table 6 corresponds to small (237 mL) cups cooling with a protective cap; Table 7 is the counterpart for the no-cap situation. Tables 8 and 9 present information for 473 mL cups with and without a protective cap, respectively.

These data show that beverage service temperatures, which are often in excess of 180 °F are at levels which have the potential to cause serious physical harm. Furthermore, some beverage service temperatures are above the preferred temperature for the consumers^[57-61]. It seems reasonable to promote the service of beverages at temperatures which are both preferred by consumers and safe so that serious mid-dermal burns are unlikely. The cooling results set forth here are corroborated by other mutually reinforcing studies^[62-64].

It should also be noted that while the above tables correspond to typical adults and children (whose skin thickness is approximately 70% that of an adult), great care must be given to their use^[55,65,66]. Also, as summarized in^[55], skin thickness varies by body location^[67-70]. For locations or persons whose skin is thinner than that used in the current study, lower temperatures or longer cooling durations are recommended. On the other hand, for persons whose skin

Table 2 Cooling of an 8 ounce (237 mL) heated beverage without a protective cap

| Cooling time (min) | Service temperature, °C (°F) | | | | | |
|--------------------|------------------------------|----------|----------|----------|----------|----------|
| | 70 (158) | 75 (167) | 80 (176) | 85 (185) | 90 (194) | 95 (203) |
| 0 | 70 | 75 | 80 | 85 | 90 | 95 |
| 5 | 58.1 | 61.9 | 61.9 | 69.5 | 73.4 | 77.2 |
| 10 | 50.4 | 53.4 | 53.4 | 59.5 | 62.5 | 65.5 |
| 15 | 45.1 | 47.6 | 47.6 | 52.6 | 55.2 | 57.7 |
| 20 | 42.4 | 44.6 | 44.6 | 49.1 | 51.4 | 53.6 |
| 25 | 42.2 | 44.4 | 44.4 | 48.9 | 51.1 | 53.3 |

Table 3 Cooling of an 8 ounce (237 mL) heated beverage with a protective cap

| Cooling time (min) | Service temperature, °C (°F) | | | | | |
|--------------------|------------------------------|----------|----------|----------|----------|----------|
| | 70 (158) | 75 (167) | 80 (176) | 85 (185) | 90 (194) | 95 (203) |
| 0 | 70 | 75 | 80 | 85 | 90 | 95 |
| 5 | 62.5 | 66.7 | 66.7 | 75.2 | 79.5 | 83.7 |
| 10 | 56.9 | 60.6 | 60.6 | 67.9 | 71.6 | 75.3 |
| 15 | 52.5 | 55.8 | 55.8 | 62.3 | 65.6 | 68.8 |
| 20 | 49.5 | 52.5 | 52.5 | 58.4 | 61.3 | 64.3 |
| 25 | 47.7 | 50.5 | 50.5 | 56.1 | 58.8 | 61.6 |

Table 4 Cooling of a 16 ounce (473 mL) heated beverage without a protective cap

| Cooling time (min) | Service temperature, °C (°F) | | | | | |
|--------------------|------------------------------|----------|----------|----------|----------|----------|
| | 70 (158) | 75 (167) | 80 (176) | 85 (185) | 90 (194) | 95 (203) |
| 0 | 70 | 75 | 80 | 85 | 90 | 95 |
| 5 | 61.5 | 65.7 | 69.8 | 74 | 78.1 | 82.3 |
| 10 | 55.9 | 59.5 | 63.1 | 66.7 | 70.3 | 73.9 |
| 15 | 51.9 | 55.1 | 58.3 | 61.5 | 64.7 | 67.9 |
| 20 | 49.6 | 52.5 | 55.5 | 58.5 | 61.4 | 64.4 |
| 25 | 48.9 | 51.8 | 54.7 | 57.5 | 60.4 | 63.3 |

Table 5 Cooling of a 16 ounce (473 mL) heated beverage with a protective cap

| Cooling time (min) | Service temperature, °C (°F) | | | | | |
|--------------------|------------------------------|----------|----------|----------|----------|----------|
| | 70 (158) | 75 (167) | 80 (176) | 85 (185) | 90 (194) | 95 (203) |
| 0 | 70 | 75 | 80 | 85 | 90 | 95 |
| 5 | 65 | 69.5 | 74 | 78.5 | 83 | 87.5 |
| 10 | 61.5 | 65.7 | 69.8 | 74 | 78.1 | 82.3 |
| 15 | 58.8 | 62.7 | 66.6 | 70.5 | 74.4 | 78.3 |
| 20 | 57 | 60.7 | 64.4 | 68.1 | 71.8 | 75.5 |
| 25 | 56 | 59.6 | 63.2 | 66.8 | 70.4 | 74 |

Table 6 Cooling times required for various service temperatures in order to cause threshold mid-dermal burns, 8-ounce (237 mL) cups with a protective cap

| Service temperature | | Cooling time for adult mid-dermal burns (min) | Cooling time for children mid-dermal burns (min) |
|---------------------|------|---|--|
| (°C) | (°F) | | |
| 95 | 203 | 8 | 12 |
| 90 | 194 | 5 | 9 |
| 85 | 185 | 2 | 6 |
| 80 | 176 | Not applicable | 3 |

is thicker than 2 mm, higher temperatures and/or shorter cooling durations can be used.

In addition to skin thickness, clothing type and the

ability of someone to remove heat quickly and apply cool or cold temperatures should be a consideration. For persons whose mobility is limited, such as children or elderly, a spill

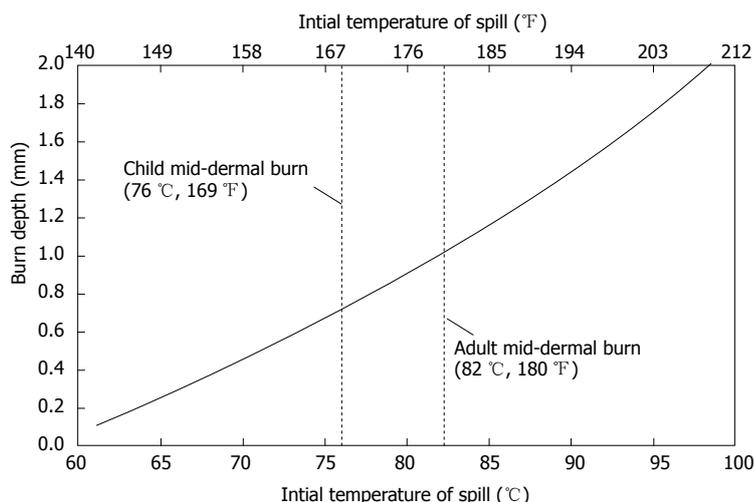


Figure 3 Burn depth and spill temperature relationship with annotations for mid dermal burns in adults and children.

Table 7 Cooling times required for various service temperatures in order to cause threshold mid-dermal burns, 8-ounce (237 mL) cups without a protective cap

| Service temperature | | Cooling time for adult mid-dermal burns (min) | Cooling time for children mid-dermal burns (min) |
|---------------------|------|---|--|
| (°C) | (°F) | | |
| 95 | 203 | 4 | 6 |
| 90 | 194 | 2 | 4 |
| 85 | 185 | 1 | 3 |
| 80 | 176 | Not applicable | 1 |

Table 8 Cooling times required for various service temperatures in order to cause threshold mid-dermal burns, 16-ounce (473 mL) cups with a protective cap

| Service temperature | | Cooling time for adult mid-dermal burns (min) | Cooling time for children mid-dermal burns (min) |
|---------------------|------|---|--|
| (°C) | (°F) | | |
| 95 | 203 | 11 | 18 |
| 90 | 194 | 7 | 12 |
| 85 | 185 | 2 | 8 |
| 80 | 176 | Not applicable | 3 |

Table 9 Cooling times required for various service temperatures in order to cause threshold mid-dermal burns, 16-ounce (473 mL) cups without a protective cap

| Service temperature | | Cooling time for adult mid-dermal burns (min) | Cooling time for children mid-dermal burns (min) |
|---------------------|------|---|--|
| (°C) | (°F) | | |
| 95 | 203 | 6 | 9 |
| 90 | 194 | 4 | 6 |
| 85 | 185 | 2 | 4 |
| 80 | 176 | Not applicable | 2 |

may remain in contact with skin for a longer duration than for someone who quickly removes the source of heat (the spilled liquid and any saturated clothing). Consequently, for these mobility-challenged persons, lower temperatures and/or longer cooling durations are recommended.

With respect to clothing, it has two competing effects. First, clothing can insulate the skin against the hottest temperatures of the liquid. Second, the clothing can hold the hot liquid against the skin, extending the scald and

delaying cooling. From a heat transfer perspective, a first responder should immediately remove the source of heat by removing saturated clothing if possible. In addition, the responder should apply cool/cold temperatures to quickly reduce the skin temperature. Room temperature liquids are often available and an excellent choice. It is recommended that if cool liquids such as room temperature water or other beverages are available nearby, they can be applied directly to the burn location and even through clothing. Burns occur

very quickly and even a few second delay in the application of cooling can make an impact on burn depth.

Not only does cooling reduce temperatures and thereby reduce burn depth^[55-56], but it also provides palliative relief^[71-84]. Many of these studies have investigated the speed at which cooling should be applied to maximize the benefit. Others have considered the optimal temperatures for the cooling. While the consensus is that temperatures in the range of 10 °C-20 °C (50 °F-70 °F) are effective, in our opinion, speed is of critical importance. A delay of a second or two can make the difference between a superficial-partial-thickness burn and a deep-partial-thickness burn. In some instances, ice is used to cause cooling. Ice should be used with care, because very cold temperatures can cause vasoconstriction which inhibits blood flow to the injured region and slows healing. Extended application of cold temperatures can even cause cryological injuries^[85-92].

BURN ASSESSMENT

Depth of the burn, TBSA involved, and the location of the burn all are necessary to develop a treatment plan. Burn depth occurs on a continuum and accurate assessment of burn depth is often difficult immediately after the injury^[3,6-10,13-21]. Burn depth assessment helps predict which injuries will require excision and grafting as deep-partial-thickness burns and full-thickness burns often require this treatment.

First-degree (superficial) burns are red, have a dry surface, and typically are associated with discomfort. These burns do not blister. The affected skin will blanch with pressure and quickly reperfuse. A typical sunburn is an example of a first-degree burn.

Superficial second-degree (superficial-partial-thickness) burns are painful and form blisters. The surface under the blister is typically red, hypersensitive and moist. This surface will blanch with pressure.

Deep second-degree (deep-partial-thickness) burns blister and may be less painful than superficial partial thickness burns. The tissue underlying the blister will be moist and may appear mottled, red, or even white. Reperfusion after blanching is slow or absent. Sensation of the damaged tissue maybe diminished when tested with pin prick.

Third-degree (full-thickness) burns extend into subcutaneous fat or connective tissue. These injuries appear white or tan and are insensate. They do not blanch as there is no perfusion to this area of the burn due to damage of the capillaries.

Fourth-degree (full-thickness) burns extend into deep structures such as muscle or bone. These injuries are easily identified.

The size of the burn is estimated based on the percentage of total body surface area involved. Only partial-thickness burns and deeper are used when estimating TBSA burned. This estimation informs intravenous fluid resuscitation needs and is used to determine if a patient

would benefit from a referral to a burn center. A common method for estimating TBSA burned is the Rule of Nines as described above. This method is endorsed by the American Burn Association, but several other methods exist. For smaller burns, the patient's hand (palm and fingers) can be used as a reference as this surface area is approximately 1% TBSA.

The final aspect of the initial burn assessment involves noting the area of the body involved. Partial thickness and deep burns involving the face, hands, feet, perineum, genitals, and major joints often require consultation with a burn center.

INITIAL MANAGEMENT AND TREATMENT CONSIDERATIONS FOR SCALD INJURIES

Prehospital care

Immediate removal of the burn source and cooling of the affected area is critical to prevent further tissue damage. Cooling is most easily accomplished by application of cool or even room temperature water. If immediate removal of any overlying clothing is possible this may be done before application of water. Otherwise, liquid can be poured directly on porous clothing to begin the cooling process. Cool water is a safe, effective, and soothing intervention. Therefore, continue with the application of cool water until the burning process is completely halted.

Initial emergency center care

Serious burns require initial treatment at an emergency center. A complete review of the initial care of the burned patient is beyond the scope of this paper. The following text will therefore focus on how the burn assessment informs subsequent treatment. After any life threatening injuries and medical complications have been addressed, the burning process has been halted, and any necessary analgesics have been administered a burn assessment can be performed. The clinician will need to assess all burns noting location and depth. Next, TBSA burned can be estimated using only partial thickness burns and deeper for this calculation.

Initial fluid resuscitation

Fluid loss through damaged skin and loss into the interstitial space can result in hypovolemic shock and thus inadequate tissue perfusion. This can lead to not only viable skin becoming nonviable, but also to end organ dysfunction. Resuscitation with intravenous fluid is the mainstay for addressing and preventing burn shock. As noted previously, first degree burns are not included in the TBSA estimation for fluid resuscitation calculations^[2,3]. Adult patients with less than 20% TBSA affected and pediatric patients will less than 10% TBSA affected can often be managed with oral hydration alone^[3]. Larger burns require intravenous fluid resuscitation with crystalloids and ringer's lactate solution is commonly

used. Multiple formulas exist for estimating intravenous fluid needs in the first 24 to 48 h after a burn. The Parkland Formula is one of the most popular. This formula uses the variables TBSA burned and patient weight in kilograms. The Parkland formula estimates the milliliters of intravenous fluid to be given in the first 24 h after the burn occurs. Half of the volume is administered over the first 8 h and the second half of the volume is administered over the following 16 h.

The Parkland Formula: Milliliters of IV fluid = 4 × TBSA burned as percent × weight in kilograms (3)

BURN CENTER REFERRAL CRITERIA

The American Burn Association has published guidelines to help clinicians determine which patients would benefit from referral to a burn center^[1].

Criteria recommended by the American Burn Association for burn center referrals are: (1) Partial thickness burns greater than 10% of body area; (2) burns of hands, feet, face, genitals, perineum, or major joints; (3) third degree burns; (4) electrical burns (including lightning); (5) chemical burns; (6) inhalation burns; (7) burns in patients with pre-existing conditions that could complicate management; (8) concomitant trauma which increases morbidity or mortality risk; (9) burns of minors treated in hospitals without qualified personnel or equipment; and (10) burn injury which requires social, emotional or rehabilitation intervention.

CONCLUDING REMARKS

Here, a two-fold presentation of information is given. First, the relationship of liquid temperature to burn depth is showcased with an easy-to-use graph. Included in the results is information regarding how the cooling time between hot-beverage service and a spill incident will reduce burn depth. With this information, it is possible to predict, within a reasonable degree of certainty, the depth of a burn injury. In addition, required beverage cooling times to avoid mid-dermal burns are listed.

In addition, the information presented here can be used by the beverage service industry to make safer hot-liquid beverages. Often times, service temperatures are above those preferred by consumers and these elevated temperatures pose an unnecessary risk of injury.

While the focus in this study was on mid-dermal burns, it should be recognized that lesser burns can also be injurious. Also, readers should recognize that mid-dermal burns can be caused at lower temperatures on areas where the skin is thinner, when victim mobility is challenged, or when the situation makes difficult the removal of the source of heat and the application of cooling.

With respect to medical care, there are a number of care phases that occur. First care starts prior to arrival at a medical center and involves the removal of heat and application of cooling. Next, there is a stage of initial medical care which may include fluid resuscitation.

For severe burn situations, referral to a burn center is made. Guidance for all these care phases is provided here with reference to published guidelines.

REFERENCES

- 1 Advanced Burn Life Support Course Provider Manual. USA: The American Burn Association, 2007
- 2 Singer AJ, Taira BR, Lee CC. Rosen's Emergency Medicine, 8th edition. Elsevier/Saunders: Thermal Burns, 2014: Chapter 63
- 3 Enoch S, Roshan A, Shah M. Emergency and early management of burns and scalds. *BMJ* 2009; **338**: b1037 [PMID: 19357185 DOI: 10.1136/bmj.b1037]
- 4 Merz KM, Pfau M, Blumenstock G, Tenenhaus M, Schaller HE, Rennekampff HO. Cutaneous microcirculatory assessment of the burn wound is associated with depth of injury and predicts healing time. *Burns* 2010; **36**: 477-482 [PMID: 19854578 DOI: 10.1016/j.burns.2009.06.195]
- 5 Johnson RM, Richard R. Partial-thickness burns: identification and management. *Adv Skin Wound Care* 2003; **16**: 178-187; quiz 188-189 [PMID: 12897674]
- 6 Monafu WW. Initial management of burns. *N Engl J Med* 1996; **335**: 1581-1586 [PMID: 8900093 DOI: 10.1056/NEJM199611213352108]
- 7 Singer AJ, Berruti L, Thode HC, McClain SA. Standardized burn model using a multiparametric histologic analysis of burn depth. *Acad Emerg Med* 2000; **7**: 1-6 [PMID: 10894235 DOI: 10.1111/j.1553-2712.2000.tb01881.x]
- 8 Durrant CAT, Simpson AR, Williams G. Thermal injury the first 24 h. *Current Anesthesia Crit Care* 2008; **19**: 256-263 [DOI: 10.1016/j.cacc.2008.09.014]
- 9 Heimbach DM, Afromowitz MA, Engrav LH, Marvin JA, Perry B. Burn depth estimation—man or machine. *J Trauma* 1984; **24**: 373-378 [PMID: 6371255 DOI: 10.1097/00005373-198405000-00001]
- 10 Palla RL. A heat transfer analysis of scald injury. USA: National Technology Information Service, U.S. Department of Commerce, 1981 [DOI: 10.6028/nbs.ir.81-2320]
- 11 Henriques FC, Moritz AR. Studies of Thermal Injury: I. The Conduction of Heat to and through Skin and the Temperatures Attained Therein. A Theoretical and an Experimental Investigation. *Am J Pathol* 1947; **23**: 530-549 [PMID: 19970945]
- 12 Moritz AR, Henriques FC. Studies of Thermal Injury: II. The Relative Importance of Time and Surface Temperature in the Causation of Cutaneous Burns. *Am J Pathol* 1947; **23**: 695-720 [PMID: 19970955]
- 13 Moritz AR. Studies of Thermal Injury: III. The Pathology and Pathogenesis of Cutaneous Burns. An Experimental Study. *Am J Pathol* 1947; **23**: 915-941 [PMID: 19970971]
- 14 Henriques FC. Studies of thermal injury; the predictability and the significance of thermally induced rate processes leading to irreversible epidermal injury. *Arch Pathol (Chic)* 1947; **43**: 489-502 [PMID: 20243514]
- 15 Pearce JA, Thomsen S. Kinetic models of laser-tissue fusion processes. *Biomed Sci Instrum* 1993; **29**: 355-360 [PMID: 8329613 DOI: 10.1117/12.137349]
- 16 McMurray T, Pearce JA. Thermal damage quantification utilizing tissue birefringence color image analysis. *Biomed Sci Instrum* 1993; **29**: 235-242 [PMID: 8329595 DOI: 10.1117/12.148628]
- 17 Sapareto SA, Dewey WC. Thermal dose determination in cancer therapy. *Int J Radiat Oncol Biol Phys* 1984; **10**: 787-800 [PMID: 6547421 DOI: 10.1016/0360-3016(84)90379-1]
- 18 Iwai M, Iwai Y, Suzumura S, Miyahara H, Imai S, Matsunaga T. Normal human salivary gland cells produce carcinoembryonic antigen-related antigen in collagen gels. *In Vitro Cell Dev Biol* 1991; **27A**: 759-762 [PMID: 1960143 DOI: 10.1115/1.3128671]
- 19 Pearce JA. Comparative analysis of mathematical models of cell death and thermal damage processes. *Int J Hyperthermia* 2013; **29**: 262-280 [PMID: 23738695 DOI: 10.3109/02656736.2013.786140]
- 20 Lepock JR. Cellular effects of hyperthermia: relevance to the minimum dose for thermal damage. *Int J Hyperthermia* 2003; **19**: 252-266 [PMID: 12745971 DOI: 10.1080/0265673031000065042]

- 21 **Arrhenius S.** Uber die reaktionsgeschwindigkeit bei der inversion von rohrzucker durch sauren (on the reaction rate in the inversion of cane sugar by acids). *Z Phys Chem* 1889; **4**: 226-248
- 22 **Arrhenius S.** Quantitative laws in biological chemistry. London: G. Bell & Sons, 1915: 184
- 23 **Tropea BI, Lee RC.** Thermal injury kinetics in electrical trauma. *J Biomech Eng* 1992; **114**: 241-250 [PMID: 1602768 DOI: 10.1115/1.2891378]
- 24 **Torvi DA, Dale JD.** A finite element model of skin subjected to a flash fire. *J Biomech Eng* 1994; **116**: 250-255 [PMID: 7799624 DOI: 10.1115/1.2895727]
- 25 **Taylor D.** Physical mechanisms of cellular injury in electrical trauma. USA: PhD Thesis, MIT, 1989
- 26 **Moussa NA, McGrath JJ, Cravalho EG, Asimacopoulos PJ.** Kinetics of Thermal Injury in Cells. *J Biomech Eng* 1977; **99**: 155-159 [DOI: 10.1115/1.3426283]
- 27 **Takata A.** Development of criterion for skin burns. *Aerosp Med* 1974; **45**: 634-637
- 28 **Weaver JA, Stoll AM.** Mathematical model of skin exposed to thermal radiation. *Aerosp Med* 1969; **40**: 24-30 [PMID: 5782654]
- 29 **Mehta AK, Wong F.** Measurement of flammability and burn potential of fabrics. Technical Report, COM-73-10950, NTIS, 1973
- 30 **Fugitt CE.** A rate process of thermal injury. Armed Forces Special Weapons Project, AFSWP-606, 1955
- 31 **Weaver JA, Stoll AM.** Mathematical model of skin exposed to thermal radiation. Us Nav Air Develop Center Aviat Med Accel Lab Rep Mr: National Air Defense Command Memo Report, 1967: 6708
- 32 **Pennes HH.** Analysis of tissue and arterial blood temperatures in resting human forearm. *J Applied Physiology* 1948; **1**: 93-133
- 33 **Dai W, Yu H, Nassar R.** A Fourth-Order compact finite-difference scheme for solving a 1-D Pennes' bioheat transfer equation in a triple-layered skin structure. *Num Heat Transfer B* 2004; **46**: 447-461 [DOI: 10.1080/104077990503014]
- 34 **Liu J, Zhu L, Xu LX.** Studies on the three-dimensional temperature transients in the canine prostate during transurethral microwave thermal therapy. *J Biomech Eng* 2000; **122**: 372-379 [PMID: 11036560 DOI: 10.1115/1.1288208]
- 35 **Diller KR.** Modeling thermal skin burns on a personal computer. *J Burn Care Rehabil* 1998; **19**: 420-429 [PMID: 9789178]
- 36 **Ng EY, Tan HM, Ooi EH.** Boundary element method with bioheat equation for skin burn injury. *Burns* 2009; **35**: 987-997 [PMID: 19427127 DOI: 10.1016/j.burns.2009.01.010]
- 37 **Diller KR, Hayes LJ, Blake GK.** Analysis of alternate models for simulating thermal burns. *J Burn Care Rehabil* 1991; **12**: 177-189 [PMID: 2050731 DOI: 10.1097/00004630-199103000-00020]
- 38 **Ng EY, Chua LT.** Prediction of skin burn injury. Part 1: Numerical modelling. *Proc Inst Mech Eng H* 2002; **216**: 157-170 [PMID: 12137283 DOI: 10.1243/0954411021536379]
- 39 **Ng EY, Chua LT.** Prediction of skin burn injury. Part 2: Parametric and sensitivity analysis. *Proc Inst Mech Eng H* 2002; **216**: 171-183 [PMID: 12137284 DOI: 10.1243/0954411021536388]
- 40 **Ng EY, Chua LT.** Comparison of one- and two-dimensional programmes for predicting the state of skin burns. *Burns* 2002; **28**: 27-34 [PMID: 11834326 DOI: 10.1016/S0305-4179(01)00066-3]
- 41 **Abraham JP, Sparrow EM, Ramadhyani S.** A mathematical model to predict tissue temperatures and necrosis during microwave thermal ablation of the prostate. New York: Numerical Heat Transfer 3: Numerical Implementation of Bioheat Models and Equations, Taylor and Francis, 2009
- 42 **Lovik RD, Abraham JP, Sparrow EM.** Potential tissue damage from transcutaneous recharge of neuromodulation implants. *Int J Heat Mass Trans* 2009; **52**: 3518-3524 [DOI: 10.1016/j.ijheatmasstransfer.2009.03.010]
- 43 **Smith DK, Lovik RD, Sparrow EM, Abraham JP.** Human tissue temperatures achieved during recharging of new-generation neuromodulation devices. *Int J Heat Mass Transfer* 2010; **53**: 3292-3299 [DOI: 10.1016/j.ijheatmasstransfer.2010.02.049]
- 44 **Abraham JP, Sparrow EM, Ramadhyani S.** Numerical simulation of a bph thermal therapy - a case study involving TUMT. *J Biomech Eng* 2007; **129**: 548-557 [DOI: 10.1115/1.2746377]
- 45 **Jiang SC, Ma N, Li HJ, Zhang XX.** Effects of thermal properties and geometrical dimensions on skin burn injuries. *Burns* 2002; **28**: 713-717 [PMID: 12464468 DOI: 10.1016/S0305-4179(02)00104-3]
- 46 **Abraham JP, Nelson-Cheeseman BB, Sparrow E, Wentz JE, Gorman JM, Wolf SE.** Comprehensive method to predict and quantify scald burns from beverage spills. *Int J Hyperthermia* 2016; **32**: 900-910 [PMID: 27405847 DOI: 10.1080/02656736.2016.1211752]
- 47 **Bourdon RT, Nelson-Cheeseman BB, Abraham JP.** Prediction, identification, and initial treatment guidelines for scald injuries. *Austin J Emerg Crit Care Med Special Issue on Burns* 2016; **3**: article no. 1043
- 48 **Diller KR, Hayes LJ.** A finite element model of burn injury in blood-perfused skin. *J Biomech Eng* 1983; **105**: 300-307 [PMID: 6632835 DOI: 10.1115/1.3138423]
- 49 **Viglianti BL, Dewhirst MW, Abraham JP, Gorman JM, Sparrow EM.** Rationalization of thermal injury quantification methods: application to skin burns. *Burns* 2014; **40**: 896-902 [PMID: 24418648 DOI: 10.1016/j.burns.2013.12.005]
- 50 **Diller KR.** Modeling of bioheat transfer processes at high and low temperatures. *Advances in Heat Transfer* 1992; **22**: 157-357 [DOI: 10.1016/S0065-2717(08)70345-9]
- 51 **Vallez LJ, Plourde BD, Abraham JP.** A new computational thermal model for the whole human body: applications to patient warming blankets. *Num Heat Transfer A* 2016; **69**: 227-241 [DOI: 10.1080/10407782.2015.1080573]
- 52 **Diller KR.** Analysis of burns caused by long-term exposure to a heating pad. *J Burn Care Rehabil* 1991; **12**: 214-217 [PMID: 1885636 DOI: 10.1097/00004630-199105000-00003]
- 53 **Abraham JP, Hennessey MP, Minkowycz WJ.** A simple algebraic model to predict burn depth and injury. *Int Comm Heat Mass Transfer* 2011; **38**: 1169-1171 [DOI: 10.1016/j.icheatmasstransfer.2011.07.004]
- 54 **Dai W, Wang H, Jordan PM, Mickens RE, Bejan A.** A Mathematical Model for Skin Burn Injury Induced by Radiation Heating. *Int J Heat Mass Trans* 2008; **51**: 5497-5510 [DOI: 10.1016/j.ijheatmasstransfer.2008.01.006]
- 55 **Johnson NN, Abraham JP, Helgeson ZI, Minkowycz WJ, Sparrow EM.** An archive of skin-layer thicknesses and properties and calculations of scald burns with comparisons to experimental observations. *J Thermal Sci Engin Appl* 2011; **3**: 011003 [DOI: 10.1115/1.4003610]
- 56 **Abraham JP, Plourde B, Vallez L, Stark J, Diller KR.** Estimating the time and temperature relationship for causation of deep-partial thickness skin burns. *Burns* 2015; **41**: 1741-1747 [PMID: 26188899 DOI: 10.1016/j.burns.2015.06.002]
- 57 **Brown F, Diller KR.** Calculating the optimum temperature for serving hot beverages. *Burns* 2008; **34**: 648-654 [PMID: 18226454 DOI: 10.1016/j.burns.2007.09.012]
- 58 **Pipatsattayanutong S, Lee HS, Lau S, O'Mahony M.** Hedonic R-Index measurement of temperature preferences for drinking black coffee. *J Sensor Studies* 2001; **16**: 517-536 [DOI: 10.1111/j.1745-459X.2001.tb00317.x]
- 59 **Borchgrevink CP, Susskind AM, Tarras JT.** Consumer preferred hot beverage temperatures. *Food Quality and Preference* 1999; **10**: 117-121 [DOI: 10.1016/S0950-3293(98)00053-6]
- 60 **Lee HS, O'Mahony M.** At what temperature do consumers like to drink coffee?: Mixing methods. *J Food Science* 2002; **67**: 2774-2777 [DOI: 10.1111/j.1365-2621.2002.tb08814.x]
- 61 **Jamnadas-Khoda B, See MS, Cubison CT, Dheansa BS.** How would you like your tea, vicar? *Burns* 2010; **36**: 356-359 [PMID: 19586723 DOI: 10.1016/j.burns.2009.04.024]
- 62 **Ramanathan C, Ekpenyong L, Stevenson JH.** Scald burns in children caused by hot drinks--the importance of the type of cup. *Burns* 1994; **20**: 111-114 [PMID: 8198713 DOI: 10.1016/S0305-4179(06)80005-7]
- 63 **Mercer NS.** With or without? A cooling study. *Burns Incl Therm Inj* 1988; **14**: 397-398 [PMID: 3228698 DOI: 10.1016/0305-4179(88)90010-1]
- 64 **Warner RM, Wilson Y, Chester DL.** Cooling properties of everyday liquids. *Burns* 2012; **38**: 1186-1191 [PMID: 22560336 DOI: 10.1016/j.burns.2012.03.008]
- 65 **Seidenari S, Giusti G, Bertoni L, Magnoni C, Pellacani G.** Thickness

- and echogenicity of the skin in children as assessed by 20-MHz ultrasound. *Dermatology* 2000; **201**: 218-222 [PMID: 11096192 DOI: 10.1159/000018491]
- 66 **Conti A**, Schiavi ME, Seidenari S. Capacitance, transepidermal water loss and causal level of sebum in healthy subjects in relation to site, sex and age. *Int J Cosmet Sci* 1995; **17**: 77-85 [PMID: 19250473 DOI: 10.1111/j.1467-2494.1995.tb00111.x]
- 67 **Laurent A**, Mistretta F, Bottiglioli D, Dahel K, Goujon C, Nicolas JF, Hennino A, Laurent PE. Echographic measurement of skin thickness in adults by high frequency ultrasound to assess the appropriate microneedle length for intradermal delivery of vaccines. *Vaccine* 2007; **25**: 6423-6430 [PMID: 17640778 DOI: 10.1016/j.vaccine.2007.05.046]
- 68 **Sandby-Møller J**, Poulsen T, Wulf HC. Epidermal thickness at different body sites: relationship to age, gender, pigmentation, blood content, skin type and smoking habits. *Acta Derm Venereol* 2003; **83**: 410-413 [PMID: 14690333 DOI: 10.1080/00015550310015419]
- 69 **Whitton JT**, Everall JD. The thickness of the epidermis. *Br J Dermatol* 1973; **89**: 467-476 [PMID: 4753709 DOI: 10.1111/j.1365-2133.1973.tb03007.x]
- 70 **Southwood WF**. The thickness of the skin. *Plast Reconstr Surg* (1946) 1955; **15**: 423-429 [PMID: 14384521 DOI: 10.1097/00006534-195505000-00006]
- 71 **Saffle JR**, Davis B, Williams P. Recent outcomes in the treatment of burn injury in the United States: a report from the American Burn Association Patient Registry. *J Burn Care Rehabil* 1995; **16**: 219-232; discussion 288-289 [PMID: 7673300 DOI: 10.1097/00004630-199505000-00002]
- 72 **Ahrenholz DH**, Clayton MC, Solem LD. Burns and wound management. *Otolaryngol Clin North Am* 1995; **28**: 1039-1055 [PMID: 8559571]
- 73 **Davies JW**. Prompt cooling of burned areas: a review of benefits and the effector mechanisms. *Burns Incl Therm Inj* 1982; **9**: 1-6 [PMID: 6184139]
- 74 **Wiedeman MP**, Brigham MP. The effects of cooling on the microvasculature after thermal injury. *Microvasc Res* 1971; **3**: 154-161 [PMID: 5110756 DOI: 10.1016/0026-2862(71)90019-7]
- 75 **Yuan J**, Wu C, Holland AJ, Harvey JG, Martin HC, La Hei ER, Arbuckle S, Godfrey TC. Assessment of cooling on an acute scald burn injury in a porcine model. *J Burn Care Res* 2007; **28**: 514-520 [PMID: 17438497 DOI: 10.1097/BCR.0B013E318053DB13]
- 76 **Rajan V**, Bartlett N, Harvey JG, Martin HC, La Hei ER, Arbuckle S, Godfrey C, Holland AJ. Delayed cooling of an acute scald contact burn injury in a porcine model: is it worthwhile? *J Burn Care Res* 2009; **30**: 729-734 [PMID: 19506512 DOI: 10.1097/BCR.0b013e3181ac059b]
- 77 **Jandera V**, Hudson DA, de Wet PM, Innes PM, Rode H. Cooling the burn wound: evaluation of different modalities. *Burns* 2000; **26**: 265-270 [PMID: 10741593 DOI: 10.1016/S0305-4179(99)00133-3]
- 78 **Ross DC**, Diller KR. Therapeutic effects of postburn cooling. *J Biomech Eng* 1978; **100**: 149-152 [DOI: 10.1115/1.3426205]
- 79 **Baxter CR**, Waeckerle JF. Emergency treatment of burn injury. *Ann Emerg Med* 1988; **17**: 1305-1315 [PMID: 3057947 DOI: 10.1016/S0196-0644(88)80356-1]
- 80 **Boykin JV**, Eriksson E, Sholley MM, Pittman RN. Cold-water treatment of scald injury and inhibition of histamine-mediated burn edema. *J Surg Res* 1981; **31**: 111-123 [PMID: 6115095 DOI: 10.1016/0022-4804(81)90038-X]
- 81 **Nguyen NL**, Gun RT, Sparnon AL, Ryan P. The importance of immediate cooling--a case series of childhood burns in Vietnam. *Burns* 2002; **28**: 173-176 [PMID: 11900942 DOI: 10.1016/S0305-4179(01)00094-8]
- 82 **Jakobsson OP**, Arturson G. The effect of prompt local cooling on oedema formation in scalded rat paws. *Burns Incl Therm Inj* 1985; **12**: 8-15 [PMID: 4063871 DOI: 10.1016/0305-4179(85)90177-9]
- 83 **Ernst E**, Fialka V. Ice freezes pain? A review of the clinical effectiveness of analgesic cold therapy. *J Pain Symptom Manage* 1994; **9**: 56-59 [PMID: 8169463 DOI: 10.1016/0885-3924(94)90150-3]
- 84 **Werner MU**, Lassen B, Pedersen JL, Kehlet H. Local cooling does not prevent hyperalgesia following burn injury in humans. *Pain* 2002; **98**: 297-303 [PMID: 12127031 DOI: 10.1016/S0304-3959(02)00030-1]
- 85 **Shulman AG**. Ice water as primary treatment of burns. Simple method of emergency treatment of burns to alleviate pain, reduce sequelae, and hasten healing. *JAMA* 1960; **173**: 1916-1919 [PMID: 14446305 DOI: 10.1001/jama.1960.03020350034007]
- 86 **Johnson NN**, McCaffrey KL, Rose KM, Abraham JP. Cryosurgical treatments for terine fibroids. ASME 2010 International Congress and Expo, Vancouver, CA, November 12-18, 2010
- 87 **Hoffmann NE**, Bischof JC. Cryosurgery of normal and tumor tissue in the dorsal skin flap chamber: Part II--injury response. *J Biomech Eng* 2001; **123**: 310-316 [PMID: 11563755 DOI: 10.1115/1.1385838]
- 88 **Abraham JP**, Plourde BD, Stark JR, Minkowycz WJ. Cryosurgical treatment of cancer: the importance of modeling. 4th World Congress on Cancer Science and Therapy, Chicago, October 20-22, 2014
- 89 **He X**, Bischof JC. Quantification of temperature and injury response in thermal therapy and cryosurgery. *Crit Rev Biomed Eng* 2003; **31**: 355-422 [PMID: 15139301 DOI: 10.1615/CritRevBiomedEng.v31.i56.10]
- 90 **Abraham JP**, Plourde BD, Stark JR. Cryosurgical treatment of cancer: the importance of modeling. *J Cancer Science Ther* 2014; **26**: 124
- 91 **Hayes L**, Diller KR, Chang HJ, Lee HS. Prediction of local cooling rates and cell survival during freezing of a cylindrical specimen. *Cryobiology* 1988; **25**: 67-872 [DOI: 10.1016/0011-2240(88)90022-3]
- 92 **Litvan GG**. Mechanism of cryoinjury in biological systems. *Cryobiology* 1972; **9**: 182-191 [PMID: 5045624 DOI: 10.1016/0011-2240(88)90022-3]

P- Reviewer: Chen XL, Suliman MT S- Editor: Kong JX
L- Editor: A E- Editor: Lu YJ



Cutaneous implications of essential oils

Ramya Vangipuram, Lisa Mask-Bull, Soo Jung Kim

Ramya Vangipuram, Center for Clinical Studies, Webster, TX 77598, United States

Lisa Mask-Bull, Soo Jung Kim, Department of Dermatology, Texas Tech University Health Sciences Center, Lubbock, TX 79430, United States

Author contributions: Vangipuram R, Mask-Bull L and Kim SJ contributed equally to this work; Vangipuram R wrote the paper; Mask-Bull L developed the idea and designed the outline; Kim SJ performed a critical revision of the manuscript.

Conflict-of-interest statement: All authors declare no conflict of interest.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Ramya Vangipuram, MD, Center for Clinical Studies, 451 N. Texas Ave, Webster, TX 77598, United States. rvangip@gmail.com
Telephone: +1-281-3332288-1109
Fax: +1-281-3334605

Received: October 20, 2016

Peer-review started: October 24, 2016

First decision: December 1, 2016

Revised: January 8, 2017

Accepted: February 10, 2017

Article in press: February 13, 2017

Published online: May 2, 2017

surge in the popularity of natural products, these oils have garnered increased attention. EOs are complex natural mixtures obtained plant materials, and have demonstrated potent biological effects *in vitro*. They have commercial value in the food, cosmetics, and fragrance industries, and also have also experienced a steady rise in personal and home use as part of aromatherapy. Currently, widespread acceptance and use of EOs is limited by a lack of large-scale clinical trials in humans. In addition, they are associated with notable side effects such as contact and allergic dermatitis, among a myriad of rare but serious systemic side effects. This review is intended to provide the clinician with key background information and biology of essentials oils, identify key trials demonstrating benefits, and describe adverse effects, with a focus on cutaneous presentations.

Key words: Essential oils; Photosensitization; Contact dermatitis; Aromatherapy

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Essential oils (EOs) have been used as home remedies for millennia. Currently, widespread acceptance and use of EOs is limited by a lack of large-scale clinical trials in humans. In addition, EOs are associated with notable side effects such as contact and allergic dermatitis, among a myriad of rare but serious systemic side effects. We review the current usage of EOs and identify pertinent cutaneous manifestations.

Vangipuram R, Mask-Bull L, Kim SJ. Cutaneous implications of essential oils. *World J Dermatol* 2017; 6(2): 27-31 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i2/27.htm>
DOI: <http://dx.doi.org/10.5314/wjd.v6.i2.27>

Abstract

Essential oils (EOs) as home remedies and for health benefits have been used for millennia, but with the recent

INTRODUCTION

Essential oils (EOs) are complex volatile substances extracted from plants, and used in food, cosmetic, and

Table 1 Commonly used essential oils (botanical origin)

| |
|--|
| Tea tree oil (<i>Melaleuca alternifolia</i>) |
| Jasmine absolute (<i>Jasminum officinale</i>) |
| Sweet bay (Laurel) oil (<i>Laurus nobilis</i>) |
| Cedarwood oil (<i>Juniperus virginiana</i>) |
| Patchouli oil (<i>Pogostemon cablin</i>) |
| Ylang-ylang oil (<i>Cananga odorata</i>) |
| Lemongrass oil (<i>Cymbopogon spp.</i>) |
| Clove oils (<i>Eugenia caryophyllus</i>) |
| Jasmine absolute (<i>Jasminum officinale</i>) |
| Sweet bay (Laurel) oil (<i>Laurus nobilis</i>) |
| Neroli oil (<i>Citrus aurantium</i> flower) |
| Peppermint oil (<i>Mentha piperita</i>) |
| Narcissus absolute (<i>Narcissus poeticus</i> Flower Extract) |
| Lemon oil (<i>Citrus medica limonum</i>) |
| Eucalyptus oil (<i>Eucalyptus globulus</i>) |
| Orange oil (<i>Citrus aurantium dulcis</i>) |

fragrance industries. They have gained the attention of the medical community for their biologically active effects and therapeutic potential for many illnesses. In addition, EOs are also experiencing a tremendous growth in aromatherapy and home use for their reported health benefits^[1]. EOs are well-known allergens and photosensitizers; however, there is a paucity of data on the dermal exposure of essential oil use in the United States. In addition, the production and use of EOs is not currently standardized or regulated and may pose an occupational hazard for those with close and repeated contact with EOs. With increasing popularity of essential oil consumption, clinicians can expect to come across more cases of cutaneous and systemic reactions to these complex substances. This review provides the most updated and relevant scientific information related essential oil use, primarily pertaining to cutaneous involvement.

BACKGROUND

EOs are secondary metabolites found in plants^[1]. They are derived from plant material, such as leaves, stems, flowers, bark, and roots^[1]. Common methods used to extract the components include steam distillation, or mechanical expression; oils produced with the aid of chemical solvents are not considered true EOs^[1]. The major chemical composition of EOs includes terpenes, esters, aldehydes, ketones, alcohols, phenols, and oxides^[2]. A given essential oil contains varying amounts of each of these compounds, which imparts a particular fragrance and determines its therapeutic characteristics^[2]. In contrast, a fragrance is chemically made to mimic the smell of a plant or flower.

EOs can be divided into two main distinct biosynthetic origins: The terpenes and terpenoids, and the aromatic and aliphatic components^[3]. There is great interest in the main biologically active component of EOs - terpenes and terpenoids. Terpenes are a large and diverse class of organic compounds that consist of five-carbon bases^[4]. Some terpenes, such as the diterpenes, are the building

blocks for biologically active compounds such as retinol, retinal, and taxol^[1]. Diterpenoids have antioxidant, antimicrobial, anticancer, anti-inflammatory, wound healing, antihypertensive, analgesic, and anxiolytic activities^[5-7].

APPLICATIONS OF EOS

Currently, of the approximately 3000 EOs that have been described, 300 are commercially important^[8,9]. The use of EOs is common in food flavoring, fragrance, and cosmetic industries. The United States Food and Drug administration has classified most EOs as "generally recognized as safe" at specified concentration limits^[3].

EOs comprise the key ingredient in aromatherapy, which is rapidly growing in popularity worldwide^[10-13]. Many spas, massage therapists, and practitioners of alternative medicine provide aromatherapy. The most commonly used EOs in aromatherapy include patchouli, cedarwood, lavender, tea tree oil, along with citrus-scented oils such as bergamot, lemon, and orange oils (Table 1). The oils are usually applied to the skin, but can also be given orally, by inhalation, or by diffusion through the air. Currently, aromatherapy products do not need approval by the FDA^[13].

Little is known about consumption habits and exposure to EOs, especially in the United States. The most comprehensive study of usage patterns was a 2014 study, which focused on the 12 most types of EOs among 1507 participants in France^[14]. Information about types of EOs used, skin areas exposed, frequencies and quantities were collected. Lavender (*Lavanda*) species are the most used EOs among both females and males, followed by *Eucalyptus* oil (Table 2)^[14]. The study notably pointed out the increased prevalence of female users for almost all types of Eos^[14]. In addition, females tend to apply EOs on their face and neck, while males applied the products on the chest^[14].

MEDICINAL USES

EOs are composed of many biologically active molecules, which may have promising therapeutic benefits in many diseases and ailments. EOs have been recognized for their antibacterial, antiviral, antifungal, and insecticidal properties, which led to their acceptance and wide-spread use in the food industry^[15-19]. Pre-clinical studies have shown that in addition to aforementioned properties, EOs also demonstrate potent anti-inflammatory, and antioxidant activity^[20-22]. Because of the great number and variety of constituents, EOs do not have specific cellular targets. They exert their cytotoxic effects through disruptions in the structure and functions of key intracellular lipids and proteins^[2]. In eukaryotic cells, EOs can change the fluidity of membranes, which become abnormally permeable resulting in leakage of radicals, cytochrome C, calcium ions and proteins^[2]. Permeabilization of outer and inner mitochondrial membranes leads to cell death through apoptosis and

Table 2 Usage patterns of essential oils by gender (percentage of use)

| | Females | Males |
|----------------|-------------------|------------------|
| Essential oils | Lavender (60%) | Lavender (50%) |
| | Eucalyptus (35%) | Eucalyptus (42%) |
| | Menthol (28%) | Ylang ylang (21) |
| | Ylang ylang (28%) | Tea Tree (19%) |
| | Tea Tree (24%) | Citrus (19%) |
| | Citrus (24%) | Menthol (18%) |
| | Vanilla (17%) | Vanilla (16%) |
| | Rosemary (16%) | Pine (15%) |
| | Ravintsara (16%) | Rosemary (14%) |
| | Pine (11%) | Neroli (9%) |

necrosis^[2]. Similar cytotoxic effects were observed *in vitro* in many gram positive and gram negative bacteria of relevance to the food industry including *S. aureus* and *E. coli*^[2].

Bottom of form

Of all the EOs, tea tree oil (TTO) is arguably the most recognized and investigated compound in dermatology. Numerous studies have demonstrated its tolerability and efficacy and against *P. acnes*^[22-24]. A 1990 single-blind randomized controlled trial (RCT) in 124 patients showed that 5% TTO gel has a comparable efficacy to that 5% benzoyl peroxide lotion^[22]. In 2007, a double-blind RCT was performed in 60 patients with mild to moderate facial acne vulgaris^[23]. A significant difference between TTO gel and placebo was observed based on decreases in total lesion counts and acne severity index scores^[23]. Most recently, the results of a 2016 phase II pilot study assessing tea tree oil for the treatment of mild to moderate acne further demonstrated its efficacy, and favorable side-effect profile^[24]. No serious adverse events were reported in this study and side effects were limited to self-resolving peeling, dryness and scaling^[24]. In addition, tea tree oil has shown promising results for other common dermatologic ailments such as seborrheic dermatitis^[25-27]. A 2002 single-blind parallel controlled trial of 126 patients with mild to moderate dandruff showed that the use of 5% TTO shampoo showed 41% improvement in dandruff, as measured by quadrant-area-severity score, compared with 11% in the placebo group ($P < 0.001$)^[27].

EOs have also been studied for the treatment of alopecia areata. A double-blind RCT involving 86 patients showed that a mixture of thyme, rosemary, lavender, and cedarwood EOs massaged into patients' scalps produced significant improvement when compared with the carrier oils alone (improvement in 54% and 21% of patients, respectively, $P = 0.08$)^[28]. The efficacy of the treatment was evaluated at initial assessment and 3 and 7 mo after treatment by dermatologists' visual scoring of photographs and a computerized analysis of traced areas of alopecia^[28]. However, the study had limited external validity, as the extent and severity of the alopecia areata in the subjects were not mentioned. At this time, there

are no further clinical trials using EOs for alopecia areata.

There is little doubt that EOs may have great relevance to the field of dermatology, and more studies should be performed given all of their *in vitro* findings. Further work on the antimicrobial, antiviral and antifungal effects of EOs may have immense potential in the treatment of dermatological diseases. Indeed, a 2012 study showed that a combination of TTO with iodine was superior to iodine alone in the treatment of molluscum contagiosum virus in 53 children^[29]. Moreover, EOs may have benefits in other cutaneous maladies, such as hyperpigmentation. The efficacy of α -bisabolol, a terpene derivative of the essential oil of *Matricaria chamomilla*, exerts an inhibitory effect on melanogenesis^[30]. In a 2010 study, α -bisabolol was evaluated in an 8-wk clinical trial of 28 Asian females, and led to a significant decrease in hyperpigmentation^[31].

ADVERSE EFFECTS

While safety testing on EOs has shown minimal adverse effects, the use of EOs still poses risks and allergic responses that clinicians should be aware of. Under normal conditions of established use, most oils appear to have a good safety profile^[12]. The majority of adverse events are mild, but serious toxic reactions from some EOs have been observed, including abortions or abnormalities in pregnancy, neurotoxicity manifesting as seizures or retardation of infant development, bronchial hyperreactivity, and hepatotoxicity^[12]. Accidental ingestion by young children has occasionally proved fatal^[32]. Repeated exposure to topical lavender and tea tree oils was associated with the development of prepubertal gynecomastia in a case-series of 3 subjects^[33]. This outcome was reversible upon discontinuation of the oils, and was attributed to the mild estrogenic and anti-androgenic activities of lavender and tea tree oils^[33].

Notably, the majority of adverse effects of EOs are cutaneous in nature. The field of dermatology has encountered an increase in the frequency of allergic reactions to EOs, likely secondary to the growing popularity of topical use of EOs^[12]. EOs are known sensitizers, and there is extensive evidence linking them to cases of contact allergy and allergic contact dermatitis^[34,35]. One case of airborne contact dermatitis secondary to sensitization after inhaled aromatherapy has also been described^[36]. As EOs age, they are often oxidized so their chemical composition changes, and may become more allergenic or prone to irritation^[13]. The most common allergens are ylang-ylang oils, lemongrass oil, jasmine absolute, sandalwood oil, and clove oil^[13]. However, in clinical practice, it may be difficult to identify specific EOs in many cases. For example, in aromatherapy, the practitioner commonly uses undefined mixtures of EOs without specifying the plant sources.

In addition, many EOs contain chemicals prone to causing sensitization, including limonene, linalool, citral, and cinnamyl alcohol (Table 3)^[14]. This is most commonly seen with citrus oils, such as bergamot, lemon, lime, and orange, which contain furocoumarins, in addition

Table 3 List of commonly used essential oils (botanical origin) and allergenic ingredients

| | |
|--|---|
| Ylang-ylang oil (<i>Cananga odorata</i>) | Linalool, Benzyl benzoate, Benzyl salicylate, Geraniol, Isoeugenol, Eugenol |
| Lemongrass oil (<i>Cymbopogon</i> spp.) | Citral, Geraniol, Limonene, Trans-isocitral, Eugenol, Linalool |
| Clove oils (<i>Eugenia caryophyllus</i>) | Eugenol, Isoeugenol |
| Jasmine absolute (<i>Jasminum officinale</i>) | Benzyl benzoate, Linalool, Eugenol, Benzyl salicylate, Isoeugenol |
| Sweet bay (Laurel) oil (<i>Laurus nobilis</i>) | Linalool, Limonene, Eugenol, Geraniol |
| Neroli oil (<i>Citrus aurantium</i> flower) | Linalool, Limonene, Geraniol, Citral |
| Peppermint oil (<i>Mentha piperita</i>) | Menthol, Limonene, Linalool |
| Narcissus absolute (<i>Narcissus poeticus</i> Flower Extract) | Benzyl benzoate, Cinnamyl alcohol, Isoeugenol |
| Lemon oil (<i>Citrus medica limonum</i>) | Limonene, Citral, Linalool, Geraniol |
| Eucalyptus oil (<i>Eucalyptus globulus</i>) | Limonene |
| Orange oil (<i>Citrus aurantium dulcis</i>) | Limonene, Linalool, Citral |
| Patchouli oil (<i>Pogostemon cablin</i>) | Cinnamyl alcohol |

to limonene, linalool, and citral. Linalool, a terpene derivative found in many EOs, is the most sensitizing components in many EOs^[36]. It is a fragrant chemical also found in lavender, ylang-ylang, and jasmine oils^[36]. Cinnamyl alcohol is found in patchouli oil^[15]. Factors influencing risk of photo-sensitization also include the amount of product applied and the area of exposure. This is important as the major study determining exposure patterns of topical essential oil use found that females tend to apply to areas such as the face and neck, thus placing themselves at greater risk of photosensitive reactions^[14].

NEED FOR FURTHER RESEARCH

Although it is well established that allergic contact dermatitis can result from essential oil use, the allergens in EOs are largely unknown. Moreover, patch testing currently does not provide accurate or particularly reliable information on EOs, as many EOs lack standardization in manufacturing and production^[37]. Finally, larger scale studies on exposure patterns are needed to reliably estimate the use of EOs. Many patients struggle with chronic cutaneous diseases and often wish to try to "natural" or "alternative" therapies, without being aware of the potential allergenic side effects.

CONCLUSION

The use of EOs, which are complex volatile substances with strong odors, has long been established in the fragrance and cosmetic industries. In addition, EOs have notable effects as antimicrobial agents, and are widely used in food industries. In recent times, EOs in the form of aromatherapy have experienced a resurgence in their popularity. They are notable for causing allergic and photosensitivity reactions, along with serious but rarely occurring side effects. More controlled clinical studies are needed to determine the benefits and risks of plant-derived products, especially EOs, in dermatology. This review describes historical and current results from scientific studies of essential oil components and highlights the areas in need of further research.

REFERENCES

- 1 **Islam MT**, da Mata AM, de Aguiar RP, Paz MF, de Alencar MV, Ferreira PM, de Carvalho Melo-Cavalcante AA. Therapeutic Potential of Essential Oils Focusing on Diterpenes. *Phytother Res* 2016; **30**: 1420-1444 [PMID: 27307034 DOI: 10.1002/ptr.5652]
- 2 **Bakkali F**, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils--a review. *Food Chem Toxicol* 2008; **46**: 446-475 [PMID: 17996351 DOI: 10.1016/j.fct.2007.09.106]
- 3 **Pichersky E**, Noel JP, Dudareva N. Biosynthesis of plant volatiles: nature's diversity and ingenuity. *Science* 2006; **311**: 808-811 [PMID: 16469917 DOI: 10.1126/science.1118510]
- 4 **Llana-Ruiz-Cabello M**, Pichardo S, Maisanaba S, Puerto M, Prieto AI, Gutiérrez-Praena D, Jos A, Cameán AM. In vitro toxicological evaluation of essential oils and their main compounds used in active food packaging: A review. *Food Chem Toxicol* 2015; **81**: 9-27 [PMID: 25865936 DOI: 10.1016/j.fct.2015.03.030]
- 5 **Devappa RK**, Makkar HPS, Becker K. 2011. Jatropa diterpenes: a review. *J Am Oil Chem Soc* 2011; **88**: 301-303 [DOI: 10.1007/s11746-010-1720-9]
- 6 **Cheenpracha S**, Yodsaoue O, Karalai C, Ponglimanont C, Subhadhirasakul S, Tewtrakul S, Kanjana-opas A. Potential anti-allergic ent-kaurene diterpenes from the bark of *Suregada multiflora*. *Phytochemistry* 2006; **67**: 2630-2634 [PMID: 17095024 DOI: 10.1016/j.phytochem.2006.09.031]
- 7 **Cho JH**, Lee JY, Sim SS, Whang WK, Kim CJ. Inhibitory effects of diterpene acids from root of *Aralia cordata* on IgE-mediated asthma in guinea pigs. *Pulm Pharmacol Ther* 2010; **23**: 190-199 [PMID: 20060054 DOI: 10.1016/j.pupt.2009.12.004]
- 8 **Gyawali R**, Ibrahim SA. Natural products as antimicrobial agents. *Food Contr* 2014; 12-29 [DOI: 10.1016/j.foodcont.2014.05.047]
- 9 **Hyltdgaard M**, Mygind T, Meyer RL. Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Front Microbiol* 2012; **3**: 12 [PMID: 22291693 DOI: 10.3389/fmicb.2012.00012]
- 10 **Rhind JP**. Essential Oils. A Handbook for Aromatherapy Practice. 2nd edition. London: Singing Dragon, 2012
- 11 **Lawless J**. The Encyclopedia of Essential Oils. 2nd edition. London: Harper Thorsons, 2014
- 12 **de Groot AC**, Schmidt E. Essential Oils, Part I: Introduction. *Dermatitis* 2016; **27**: 39-42 [PMID: 26983089 DOI: 10.1097/DER.000000000000175]
- 13 Aromatherapy and Essential Oils (PDQ®): Health Professional Version. PDQ Integrative, Alternative, and Complementary Therapies Editorial Board. Bethesda (MD): National Cancer Institute (US); 2002-2016 Apr 8 [PMID: 26389313]
- 14 **Dornic N**, Ficheux AS, Roudot AC, Saboureau D, Ezzedine K. Usage patterns of aromatherapy among the French general population: A descriptive study focusing on dermal exposure. *Regul Toxicol Pharmacol* 2016; **76**: 87-93 [PMID: 26826550 DOI: 10.1016/j.yrtph.2016.01.016]

- 15 **Sung SY**, Sin LT, Tee TT Soo-Tueen B, Rahmat AR, Rahman WA, Ann-Chen T, Vikhraman K. Antimicrobial agents for food packaging applications. *Trends Food Sci Technol* 2013; **33**: 110-123 [DOI: 10.1016/j.tifs.2013.08.001]
- 16 **Angioni A**, Barra A, Coroneo V, Dessi S, Cabras P. Chemical composition, seasonal variability, and antifungal activity of *Lavandula stoechas* L. ssp. *stoechas* essential oils from stem/leaves and flowers. *J Agric Food Chem* 2006; **54**: 4364-4370 [PMID: 16756368 DOI: 10.1021/jf0603329]
- 17 **Bajpai VK**, Baek KH, Kang SC. Control of Salmonella in foods by using essential oils: a review. *Food Res Int* 2012; **45**: 722-734 [DOI: 10.1016/j.foodres.2011.04.052]
- 18 **Abbaszadeh G**, Srivastava C, Walia S. Insecticidal and antifeedant activities of clerodane diterpenoids isolated from the Indian bhant tree, *Clerodendron infortunatum*, against the cotton bollworm, *Helicoverpa armigera*. *J Insect Sci* 2014; **14**: 29 [PMID: 25373176 DOI: 10.1093/jis/14.1.29]
- 19 **Tang W**, Wei X, Xu H, Zeng D, Long L. 13-Deoxyitol A, a new insecticidal isoryanodane diterpene from the seeds of *Itoa orientalis*. *Fitoterapia* 2009; **80**: 286-289 [PMID: 19318121 DOI: 10.1016/j.fitote.2009.03.006]
- 20 **Roby MHH**, Sarhana MA, Selima KAH, Khalela KI. Evaluation of antioxidant activity, total phenols and phenolic compounds in thyme (*Thymus vulgaris* L.), sage (*Salvia officinalis* L.), and marjoram (*Origanum majorana* L.) extracts. *Ind Crops Prod* 2013; **43**: 827-831 [DOI: 10.1016/j.indcrop.2012.08.029]
- 21 **Teixeira B**, Marques A, Ramos C, Batista I, Serrano C, Matos O, Neng NR, Nogueira J, Saraiva JA, Nunes ML. European pennyroyal (*Mentha pulegium*) from Portugal: chemical composition of essential oil and antioxidant and antimicrobial properties of extracts and essential oil. *Ind Crops Prod* 2012; **36**: 81-87 [DOI: 10.1016/j.indcrop.2011.08.011]
- 22 **Bassett IB**, Pannowitz DL, Barnetson RS. A comparative study of tea-tree oil versus benzoylperoxide in the treatment of acne. *Med J Aust* 1990; **153**: 455-458 [PMID: 2145499]
- 23 **Enshaieh S**, Jooya A, Siadat AH, Iraj F. The efficacy of 5% topical tea tree oil gel in mild to moderate acne vulgaris: a randomized, double-blind placebo-controlled study. *Indian J Dermatol Venereol Leprol* 2007; **73**: 22-25 [PMID: 17314442]
- 24 **Malhi HK**, Tu J, Riley TV, Kumarasinghe SP, Hammer KA. Tea tree oil gel for mild to moderate acne; a 12 week uncontrolled, open-label phase II pilot study. *Australas J Dermatol* 2016; Epub ahead of print [PMID: 27000386]
- 25 **Gupta AK**, Nicol K, Batra R. Role of antifungal agents in the treatment of seborrheic dermatitis. *Am J Clin Dermatol* 2004; **5**: 417-422 [PMID: 15663338]
- 26 **Waldroup W**, Scheinfeld N. Medicated shampoos for the treatment of seborrheic dermatitis. *J Drugs Dermatol* 2008; **7**: 699-703 [PMID: 18664167]
- 27 **Satchell AC**, Saurajen A, Bell C, Barnetson RS. Treatment of dandruff with 5% tea tree oil shampoo. *J Am Acad Dermatol* 2002; **47**: 852-855 [PMID: 12451368 DOI: 10.1067/mjd.2002.122734]
- 28 **Hay IC**, Jamieson M, Ormerod AD. Randomized trial of aromatherapy. Successful treatment for alopecia areata. *Arch Dermatol* 1998; **134**: 1349-1352 [PMID: 9828867]
- 29 **Markum E**, Baillie J. Combination of essential oil of *Melaleuca alternifolia* and iodine in the treatment of molluscum contagiosum in children. *J Drugs Dermatol* 2012; **11**: 349-354 [PMID: 22395586]
- 30 **Kim S**, Lee J, Jung E, Huh S, Park JO, Lee JW, Byun SY, Park D. Mechanisms of depigmentation by alpha-bisabolol. *J Dermatol Sci* 2008; **52**: 219-222 [PMID: 18692366 DOI: 10.1016/j.jdermsci.2008.06.005]
- 31 **Lee J**, Jun H, Jung E, Ha J, Park D. Whitening effect of alpha-bisabolol in Asian women subjects. *Int J Cosmet Sci* 2010; **32**: 299-303 [PMID: 20642768 DOI: 10.1111/j.1468-2494.2010.00560.x]
- 32 **Tisserand R**, Young R. Essential Oil Safety. 2nd ed. USA: Churchill Livingstone Elsevier, 2014
- 33 **Henley DV**, Lipson N, Korach KS, Bloch CA. Prepubertal gynecomastia linked to lavender and tea tree oils. *N Engl J Med* 2007; **356**: 479-485 [PMID: 17267908 DOI: 10.1056/NEJMoa064725]
- 34 **Bilsland D**, Strong A. Allergic contact dermatitis from the essential oil of French marigold (*Tagetes patula*) in an aromatherapist. *Contact Dermatitis* 1990; **23**: 55-56 [PMID: 2401143]
- 35 **Lapeere H**, Boone B, Verhaeghe E, Ongenae K, Lambert J. Contact dermatitis caused by lovage (*Levisticum officinalis*) essential oil. *Contact Dermatitis* 2013; **69**: 181-182 [PMID: 23948036 DOI: 10.1111/cod.12082]
- 36 **Schaller M**, Korting HC. Allergic airborne contact dermatitis from essential oils used in aromatherapy. *Clin Exp Dermatol* 1995; **20**: 143-145 [PMID: 8565250]
- 37 **Hagvall L**, Christensson JB. Patch Testing with Main Sensitizers Does Not Detect All Cases of Contact Allergy to Oxidized Lavender Oil. *Acta Derm Venereol* 2016; **96**: 679-683 [PMID: 26671837 DOI: 10.2340/00015555-2319]

P- Reviewer: Chen GS, Kaliyadan F, Manolache L, Vasconcellos C

S- Editor: Ji FF **L- Editor:** A **E- Editor:** Lu YJ



Retrospective Study

Use of a selective enzymatic debridement agent (Nexobrid®) for wound management: Learning curve

Ricardo Palao, Jorge Aguilera-Sáez, Jordi Serracanta, Jose Manuel Collado, Bruce Patrik Dos Santos, Juan Pedro Barret

Ricardo Palao, Jorge Aguilera-Sáez, Jordi Serracanta, Jose Manuel Collado, Bruce Patrik Dos Santos, Juan Pedro Barret, Plastic, Reconstructive and Burns Unit Service, Vall d'Hebron University Hospital, 08035 Barcelona, Spain

Author contributions: Palao R, Aguilera-Sáez J and Serracanta J designed and performed research; Palao R, Aguilera-Sáez J, Serracanta J, Collado JM, Dos Santos BP and Barret JP analysed de data; Palao R, Aguilera-Sáez J and Dos Santos BP wrote the paper.

Institutional review board statement: This study was reviewed and approved by the Ethics Committee of the Vall d'hebron University Hospital.

Informed consent statement: Patients were not required to give informed consent to the study because the analysis used anonymous clinical data that were obtained after each patient agreed to treatment by written consent.

Conflict-of-interest statement: The authors report no relevant conflicts of interest.

Data sharing statement: No additional data are available.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Ricardo Palao, MD, Staff Member, Plastic, Reconstructive and Burns Unit Service, Vall d'Hebron University Hospital, Passeig Vall d'Hebron 119-129, 08035 Barcelona, Spain. rpalao@vhebron.net
Telephone: +34-62-9330102

Received: August 23, 2016

Peer-review started: August 24, 2016

First decision: October 8, 2016

Revised: November 16, 2016

Accepted: January 20, 2017

Article in press: January 21, 2017

Published online: May 2, 2017

Abstract**AIM**

To evaluate the efficacy of Nexobrid® in the initial management of burns and lessons learned with the procedure.

METHODS

From January 27th 2015 until January 25th 2016, 25 patients aged between 18-94 years old with deep partial and full thickness burns were treated with Nexobrid® covering 1%-30% of their total body surface area (TBSA). The debridement was applied in the first 96 h post-injury following the protocol suggested for Nexobrid®. In patients with burns of more than 15% TBSA a second application of Nexobrid® was performed. After the removal of the product - 4 h post application and after a 2 h period of wet dressing - we used several products to cover the wound like Suprathel®, Biobrane®, Mepitel® with wet dressing, silver sulphadiazine 1% cream, and in some cases even autografts. We treated patients with inhalation injury as well. All the procedures were done under deep sedation, regional blocks in extremities or general anaesthesia in the intensive care unit room or in the operating theatre.

RESULTS

After these first 25 cases, we have observed that patients with partial thickness burns treated with Nexobrid®, experienced great benefits in the reduction of the need for autografting compared with the standard of care. This is

because after selective enzymatic debriding of the burn scar we can distinguish different wound beds, which can coexist in the same patient, and we also managed to associate each one to its ability to epithelize. In major burns, besides the improvement in wound healing, we observed an important improvement in their general state. This may be because SIRS significantly improved through a bloodless debridement of necrotic tissue, decreasing the requirements of vasoactive drugs and fluid resuscitation. Circumferential burns also benefited from enzymatic debridement, observing a decrease in the number of compartment syndromes and the need for escharotomies. At present, we have not observed a positive effect in the evolution and outcome of major burns with inhalation injury.

CONCLUSION

The introduction of Nexobrid® shows significant improvement in burn treatment. Cumulative experiences are necessary to adapt its application in all Burns Centres.

Key words: Burns; Eschar; Debridement; Enzymatic debridement; Wound bed; Nexobrid®; Epithelialization; Dermis preservation; Autograft

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Burns continue to be a common injury in western countries. It is difficult to assess the severity of burns, but knowing its mechanisms as well as its characteristics can be of help. An alternative to surgical debridement is the enzymatic or chemical debridement, although past reports that used it with patients show that its efficiency is limited. Nexobrid® is a new enzymatic debridement agent. We show our learning curve with it.

Palao R, Aguilera-Sáez J, Serracanta J, Collado JM, Dos Santos BP, Barret JP. Use of a selective enzymatic debridement agent (Nexobrid®) for wound management: Learning curve. *World J Dermatol* 2017; 6(2): 32-41 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i2/32.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i2.32>

INTRODUCTION

When a thermic, electric or chemical aggression acts in the skin for the necessary time and with enough intensity, then a local injury known as burn occurs. Nowadays, burns continue to be a common injury in western countries. Many of these burns will need urgent medical attention, hospital admission, and even surgery^[1]. It is difficult to assess the severity of burns, but knowing its mechanisms as well as its characteristics can be of help. Nonetheless, the majority of burns present with different degrees in all its extension through the skin. Consequently, it can be extremely complicated to differentiate between a superficial partial thickness burn and a deep partial thickness one^[2,3]. The diagnosis of burn depth is really

important in order to plan the treatment. While the epidermal and superficial partial thickness burns will heal spontaneously, the deep partial thickness as well as the full thickness burns will need surgery^[4]. For the latter, the tangential debridement followed by the coverage with skin autograft within the first five days after suffering the injury, is considered to be the current standard of care^[5-7]. An alternative to such surgical debridement is the enzymatic or chemical debridement, although past reports that used it with patients show that its efficiency is limited, it is a slow process and increases the infection risk due to maceration of necrotic tissue^[8]. Since 2004 many articles have been published on the use, efficiency and security that Nexobrid® (enzymatic debridement) provides. Nexobrid® consists of a mixture of proteolytic enzymes enriched in bromelain, which is extracted from the stem of the pineapple plant^[9-13]. The purpose of this manuscript is to share the experience acquired during the learning process with this enzymatic debridement used in the Burns Unit of Vall d'Hebron University Hospital at Barcelona.

MATERIALS AND METHODS

The Burns Unit of Vall d'Hebron University Hospital at Barcelona is the top reference center for Catalonia, Balearic Islands (Spain) and Andorra. Every year we receive approximately between 400 and 500 burns related entries. We also receive, but to a lesser extent, other entries due to exfoliative skin illnesses.

We considered initial entries of the learning curve the first 25 cases, which received the enzymatic debridement treatment. These were done between January of 2015 and January of 2016. We worked with the enzymatic debridement in the circumstances where there was at least an intermediate partial thickness burn with an extension not above 15%, according to the product indication, unless we organized the therapy in two separated days if the patient achieved a total enzymatic debridement of 30% of its total body surface area. Based on our experience we now have our own recommendations regarding its indications based on our learning and when not to use it. As it is a painful process pain management is indicated. We do this particularly at the extremities in which we can perform locoregional nerve block, or we can also do it either in the extremities or trunk if the patient has been intubated with sedoanalgesia for whatever reason.

The technique is always performed in a sterile environment, either the theater or the same room for major burns, removing devitalized tissue. Then, we will proceed to mix both components of the product, creating the Nexobrid® gel. After that, we will delimit the area to treat with a trail of Vaseline in order to avoid spillage of the product out of the treatment area. Once we have done this, we will proceed to use the cream - creating a uniform fine layer of 1.5-3 mm thick - over the area of the body that we want to debride. Afterwards, we will cover the extremity with a transparent plastic wrap so that the lotion will not move during the 4 h that the product

Table 1 Patients treated with Nexobrid® (January 2015-January 2016)

| Patient | Age | Sex | Etiology | Extension (%) | Depth | Time to escaectomy (d) | Treated area | Pre debridement dressing | Post debridement dressing | Surgery | Time to complete healing (d) |
|---------|-----|-----|--------------|---------------|-------------|------------------------|--------------------------------|------------------------------|---------------------------|---------|------------------------------|
| 1 | 24 | W | Scald | 2 | DPT | 4 | Foot | SSD | Mepitel one | Yes | 35 |
| 2 | 31 | M | Flame | 20 | SPT and DPT | 1 | Hands and lower limbs | SSD | Mepitel one | No | 21 |
| 3 | 33 | M | Flame | 60 | DPT and FT | 0 | Trunk and upper limbs and foot | SSD | Mepitel one | Yes | 15 |
| 4 | 46 | M | Chemical | 6 | DPT and FT | 4 | Trunk | SSD | Biobrane | Yes | 19 |
| 5 | 90 | M | Flame | 4 | DPT and FT | 2 | Forearm and foot | SSD | Suprathel | Yes | 33 |
| 6 | 58 | M | Flame | 53 | DPT and FT | 0 | Hands and lower limbs | SSD | Biobrane | Yes | Exitus |
| 7 | 18 | W | Scald | 2 | SPT and DPT | 1 | Lower limbs | WD | Suprathel | Yes | 30 |
| 8 | 64 | M | Scald | 10 | MPT | 3 | Lower limbs | SSD | Suprathel | No | 14 |
| 9 | 58 | M | Contact | 1 | SPT and DPT | 3 | Foot | WD | Suprathel | Yes | 20 |
| 10 | 44 | M | Deflagration | 15 | SPT and DPT | 1 | Upper limbs | WD | Suprathel | No | 27 |
| 11 | 69 | M | Deflagration | 8 | SPT and DPT | 4 | Upper limbs | WD | Suprathel | No | 21 |
| 12 | 51 | M | Scald | 2 | SPT and DPT | 2 | Upper limbs | Nitrofurazone | Biobrane | Yes | 51 |
| 13 | 21 | M | Flame | 10 | SPT and DPT | 1 | Upper limbs | WD | Aquacel Ag | No | 16 |
| 14 | 56 | M | Flash | 5 | MPT and DPT | 4 | Hands | WD | Suprathel | No | 19 |
| 15 | 40 | M | Deflagration | 6 | SPT and DPT | 2 | Upper limbs | WD | SSD | No | 22 |
| 16 | 94 | W | Flame | 10 | DPT and FT | 4 | Trunk and upper limbs | WD | SSD | Yes | 21 |
| 17 | 81 | M | Flame | 2 | SPT and FT | 2 | Lower limb | WD | WD | Yes | 31 |
| 19 | 72 | W | Deflagration | 14 | MPT | 1 | Lower limbs | WD | Suprathel | Yes | 27 |
| 20 | 38 | M | Deflagration | 7 | DPT | 1 | Hands | WD | Biobrane | Yes | 38 |
| 21 | 63 | M | Flame | 65 | MPT and FT | 0 | Lower limbs | SSD change to WD before 24 h | Mepitel and Nitrofurazone | Yes | 23 |
| 22 | 46 | W | Scald | 4 | MPT | 2 | Lower limb | WD | Suprathel | No | 37 |
| 23 | 62 | M | Flame | 18 | SPT and DPT | 1 | Upper and lower limbs | WD | Aquacel Ag | Yes | 36 |
| 24 | 56 | M | Flame | 28 | SPT and DPT | 2 | Lower limbs | WD | SSD | Yes | EXITUS |
| 25 | 38 | W | Abrasion | 1 | DPT | 2 | Hand | WD | Aquacel Ag | No | 17 |

SPT: Superficial partial thickness; MPT: Mid partial thickness; DPT: Deep partial thickness; SSD: Silver sulfadiazine; WD: Wet dressing; W: Women; M: Man.

needs in order to work. Always remember that we must try to maintain a uniform layer of gel when we cover the extremity with the plastic wrap. After the 4 h we remove the product as well as the plastic wrap, and the rest of necrotic tissue, vaseline, exudate and bleeding. We do this with an intense brushing or scratching with the scraper. Then, we apply a wet dressing with normal saline solution 0.9% that we will leave until the next day. Once we remove the wet dressing we must apply a new one that varies according to the appearance of the treated wound bed. In the Discussion we elaborate on what we use in each step.

RESULTS

Between January 2015 and January 2016 we treated 25 patients (Table 1), 19 of them were men, 6 women. Their mean age was 52.6 years old (ranging from 19 to 94 years old). The etiology of their burns was flame in 11 cases, deflagration in 5 cases, scald in 5 cases, electrical flash in 1 case, chemical burn in another one, abrasion in 1 case, and finally one case of contact burn. The mean burned body surface area was 14.4% (ranging from 1%

to 65% TBSA).

In all cases patients received enzymatic debridement treatment in their extremities, and 3 cases also required enzymatic debridement treatment of their anterior trunk. As mentioned previously, this was done within 4 d of injury. It is important to highlight that in most cases the treatment was successful (it didn't go well in 5 cases). The reason was 4 of the cases had received local silver sulphadiazine 1% cream during more than 24 h before being treated with the enzymatic debridement. This caused the consolidation of the characteristic pseudo-eschar that this cream forms. This explains why it was unsuccessful. In the fifth case, we believe that the treatment didn't work due to the use of an insufficient amount of the product.

Pre-Nexobrid® treatment, local dressing employed was silver sulphadiazine 1% cream in 9 cases. As mentioned, in 4 of those this was used for more than 24 h which lead to incomplete eschar debridement. In the rest of patients the enzymatic debridement was performed within the first 24 h (in 3 of them), or wound dressing was replaced with wet dressing also within the first 24 h (2 patients). In 15 cases a wet dressing of normal saline solution and

chlorhexidine was used. In one case, initial dressing was silicone sheets and nitrofurazone cream. The mean burned area treated with enzymatic debridement was 6.5% (ranging from 1% to 30%).

We essayed with a wide range of possibilities regarding local dressings once we have already performed the enzymatic debridement, and also after placing wet dressing: Suprathel® (9 cases), Biobrane® (4 cases), silver sulphadiazine (3 cases), Aquacel Ag® (3 cases), Mepitel one® (3 cases), Mepitel® and nitrofurazone cream (1 case), wet dressing (1 case).

Fifteen patients underwent surgery to cover the debrided areas with skin grafts. The other 10 patients showed spontaneous epithelialization. Those patients who needed surgery (apart from the 4 patients that died) healed within an average of 30.45 d, whereas those who didn't need surgery within 22.30 d.

Each patient has been followed-up until today, so they are still receiving the same post-burn care than the patients that have been treated with the standard treatment (tangential debridement plus skin graft coverage). The post-burn care means: Use of silicone patches, pressure therapy, massages, hydration, rehabilitation, and so on. As mentioned, 4 patients died, 3 of them because of a bad systemic evolution that caused a multiple organ failure, while the other death was due to a respiratory insufficiency in the context of smoke inhalation that caused burn in the air way.

DISCUSSION

When a new technique is first introduced we always need a period of time in order to learn its different new aspects. Once we verified the debriding capacity of the product, and with the knowledge of the publications on it, then we had some doubts about which where the most suitable circumstances in which we should use the product, how to assess the wound bed after the procedure or which local dressings to apply on it, that's what we want to discuss here from our experience.

Where to use Nexobrid®

In our opinion, this product should be used to treat circumferential burns that can lead to a compartment syndrome; intermediate and deep partial thickness burns as well as burns where there are different degrees, full thickness burns. We consider that children, elderly people and major burns victims, could specially benefit from this procedure for their specific circumstances. Now we will proceed to elaborate on the benefits of using the product in each of the circumstances mentioned above.

Compartment syndrome: When there is a suspicion that the patient is suffering a compartment syndrome in any of the extremities it is, in our opinion, one of the clearest cases in which we should use the product. It has been proved that the first 30 min after employing the product the compartment pressure descends below

30 mmHg. This descent will stand even after removing the product^[10]. Eliminating the secondary rigid eschar from the circumferential burn will allow the expansion of the muscles, which consequently will eliminate the possibility of suffering compartment syndrome^[12]. None of the 25 patients that we debrided with such technique required surgical escharotomy. It is true that the surgical escharotomy effects are immediate in relation to the compartment pressure fall, however we also need to highlight that an expert surgeon as well as basic surgical staff are needed. Sometimes the expert surgeon can't treat victims fast enough, which subsequently will cause the victims to suffer irreversible pain. Furthermore, surgical escharotomy is not exempted from possible complications such as haemorrhages, loss of fluids, subcutaneous tissue infection, and so on^[10].

Partial and full thickness burns: Nexobrid® can help us diagnose intermediate partial thickness burns. These burns are the ones that due to their appearance are difficult to classify upon initial presentation. This also occurs with superficial partial thickness burns and the deep partial thickness ones. The clinical assessment performed by an expert surgeon on the depth of a burn is only accurate in two-thirds of the examined cases^[14]. There are some equipment that help us to differentiate burn depth, such as Laser Doppler Imaging, which allows to clearly differentiate depth, because it informs about the status of the dermic vascularization with an positive predictive value around 90%^[14,15]. However, Nexobrid® has a considerable advantage against this equipment, as it shows the wound bed itself thus facilitating diagnosis, at the same time we are doing the escharectomy. Another advantage of the product is that it reduces the time elapsed between the burn injury and complete burn eschar removal^[13]. Later we will talk about how to assess the wound bed after enzymatic debridement. Nowadays there are no publications that exactly delineate the interpretation of the wound bed in intermediate partial thickness burns.

Children and elderly people: In children and elderly people we tend to be more conservative in the surgical aspects as they are more delicate patients than others. Both types of patients have less epithelial reserve, while only elderly people have commonly associated comorbidities^[16,17]. We believe that Nexobrid® is an ideal medical tool. First of all, it allows us to predict if the burn will need to be treated in a conservative manner or a surgical debridement is necessary, and it also let us know how to handle such burn (conservative or surgical treatment). What is more, the extension of the area treated will be smaller in comparison to standard therapy^[12,13]. If we limit the surgical areas, we will consequently limit the extension of the donor site, where we obtain the skin graft that will cover the debrided part of the body^[13]. Moreover, if we manage to avoid surgical debridement bleeding will also be avoided^[13]. However,



Figure 1 Patient treated with silver sulphadiazine cream for more than 24 h in the pre-Nexobrid® treatment. A: Patient where silver sulphadiazine cream was applied in the pre-Nexobrid® treatment for more than 24 h; B: After 4 h of exposure to Nexobrid®, the incomplete removal of eschar is shown.

we must say that no child has participated in this research in our Unit.

Major burn victims: In relation to major burn patients, we think that an early enzymatic debridement can contribute many benefits to them. Firstly, we consider that by reducing the time until the complete escharotomy, we are also reducing the amount of time in which the body is producing cytokines and inflammatory mediators as a systemic response to the burn^[18,19]. So, we believe that the syndrome caused by the systemic inflammatory response will be minor. Additionally, early removal of the necrotic tissue can lead to a reduced incidence of sepsis. There are no publications that support our arguments, but still we consider that it is a reasonable argument that needs to be studied in the future. Finally, it is also important to say that as these patients tend to show poor overall state, we believe that by limiting surgical aggression from tangential debridement this will be beneficial to them.

When not use Nexobrid®

On the other hand, we will not follow this principle in superficial partial thickness burns, as it doesn't result in any benefit. We also will not use it in electric burns because they have a complex injury mechanism and employing it would be useless. It also won't be used in small burns where the cost outweighs the benefits. We will not employ it in burns that had been treated during more than 24 h with silver sulphadiazine cream or dressings with silver as the pseudo-eschar formed over the burn limits the debriding action of the product, as is shown in Figure 1. We can support this information with the other experiences present in the bibliography^[9]. However, if the silver sulphadiazine has been working less than 24 h, then we will proceed to remove the pseudo-eschar through an intense scratching (with a surgical brush or scraper), and after that we will use Nexobrid®.

After 24 h the pseudo-eschar cannot be completely removed. This is the reason why we apply a wet dressing with normal saline 0.9% to the patients that we initially believe can benefit from enzymatic debridement

(Nexobrid®), while we wait.

Wound bed diagnosis

As previously stated, we believe Nexobrid® is a great tool for proper diagnosis of burn depth. Actually, the selective debridement allows us to perform a precise diagnosis of the total depth of the burn, through a direct view of the skin structure and vascular patterns^[12].

The current publications recommend grafting as soon as possible those wound beds that do not present valid dermic remainders; while those valid dermic remainders will be protected with an allograft or with a dressing that will consequently allow the epithelisation during 3 wk, the time in which the areas that remain denuded will be then grafted^[12]. This has been one of the most difficult aspects to understand, as until now we associated a white wound bed without much bleeding to a deep dermic burn. But there is nothing in relation to the pre-debridement inspection with the post Nexobrid® debridement one. However, when we were reviewing our clinical cases we managed to distinguish different dermic wound beds after performing enzymatic debridement with Nexobrid®, and we also managed to associate each one to its ability to epithelize. So, after using Nexobrid® we can face four different wound beds, which obviously can coexist in the same patient, because a burn can have different depths. Now we are going to analyse the different wound beds (Table 2).

Type I : Dermic wound bed with abundance of small diameter pin-point bleeders (uniform shades of red to pink). This bed corresponds to a superficial dermic burn (Figure 2).

Type II a: Dermic wound bed with a sparse pattern of larger diameter bleeders (irregular shades of pink to white) that it is not depressed in relation to the surrounding healthy skin, or the own bed surface skin burn. It corresponds to an intermediate/deep dermal burn, but the loss of tissue thickness cannot be macroscopically observed (Figures 2 and 3).

Type II b: Dermic wound bed with a sparse pattern of larger diameter bleeders (no shades, white colour) that it is depressed in relation to the surrounding healthy skin, or the own bed surface skin burn. It corresponds to a deep dermal burn where the loss of its tissue thickness can be macroscopically observed (Figures 3 and 4).

Type III: Fatty wound bed. It corresponds to a full thickness burn (Figure 4).

The importance of all this, in our experience, is that the types I and II a will heal spontaneously within a maximum period of 30 d; whereas the types II b and III won't heal spontaneously over a period of 30 d, so we will consider covering the burns with skin autografts. Despite 30 d for burns, at risk of hypertrophic scarring, is more time than the classically accepted for a spontaneous epithelialization (3 wk), it has been seen that in a long-

Table 2 Wound bed classification after debridement with Nexobrid®

| Wound bed | Appearance | Depression | Type of burn |
|-----------|--|------------|----------------|
| Type I | Abundance of small diameter pin-point bleeders (uniform shades of red to pink) | No | Superficial PT |
| Type II a | Sparse pattern of larger diameter bleeders (irregular shades of pink to white) | No | Mid/deep PT |
| Type II b | Sparse pattern of larger diameter bleeders (no shades, white colour) | Yes | Deep PT |
| Type III | Fatty | Yes | PT |

PT: Partial thickness.



Figure 2 Wound bed Type I and II a. A: Burn before Nexobrid® treatment; B: Burn after Nexobrid® treatment with two different wound bed. Type I, with abundance of small diameter pin-point bleeders (uniform shades of red to pink, superficial partial thickness burn). Type II a, with a sparse pattern of larger diameter bleeders (irregular shades of pink to white colour), not depressed either in relation to the surrounding healthy skin or the own bed surface skin bed (mid/deep partial thickness burn); C: Outcome 38 d post-burn.

term outcome the burn scars treated with Nexobrid® are not inferior than the ones who are subjected to the standard treatment^[13]. We think that two factors can have an influence on this. On the one hand, the spontaneous epithelialization in a inflammatory bed is different from the one produced in a clean dermal base, and secondly, the effectiveness of the postoperative care of these scars.

We believe that the difference between types II a and II b is the amount of epithelial reserve available for wound healing by reepithelialisation. When we talk about epithelial reserve, we are referring to the presence of cells capable of generating a new healthy epithelial tissue. In this case it would be the basal cells of the epidermis, which are damaged in both types (II a as well as II b). However, these basal cells of the epidermis are in continuity with the basal sebaceous gland cells and the cells of the outer hair shaft, as both share embryologic development. These cells have the ability to form new

keratinocytes that migrate into the wound^[9]. In addition, the cells of the protuberance, structure located in the hair follicle below the sebaceous glands, are pluripotential cells that can reconstitute all the epithelial lineages after suffering an insult, including the interfollicular epidermis, albeit they are not involved in the normal process of epidermal renewal. Furthermore, it has been proved that no less than 25% of the new cells that have repopulated a wound derive from this structure^[20].

It's obvious to think that in deep dermal burns healing by regeneration of a healthy epithelium will depend on the number of these structures that have survived the injury. According to our observations the beds type II a maintain a sufficient epithelial reserve for healing, while the beds type II b don't (if we are macroscopically able to see a depression it is logical to think that the remaining dermis corresponds to deeper layers, and therefore is poorer in these areas).



Figure 3 Wound bed II a and II b. A and B: Burn before Nexobrid® treatment; C and D: After Nexobrid® treatment with two different wound bed. Type II a and Type II b, sparse pattern of larger diameter bleeders (no shades, white colour) that is depressed in relation to the surrounding healthy skin, or the own bed surface skin burn (Deep partial thickness burn); E and F: Autografting after two weeks; G and H: Final outcome after 6 mo.

This explanation should be taken carefully in areas of thin skin, as well as in children and elderly people who already have a thin skin. In a minor skin thickness it can be assumed that an injury that doesn't produce this macroscopic dermal depression, if it is able to destroy

the epithelial reserve so that the injury cannot heal by regeneration. Therefore, we should also be cautious in areas without hair or with patients that have suffered a hair removal treatment that removes the hair follicle. These areas will have less epithelial reserve.



Figure 4 Different wound beds in the same burn. A: Burn before Nexobrid® treatment; B: After Nexobrid® treatment with three different wound bed. Type II a, Type II b and Type III, with a fatty wound bed (full thickness burn)

Factors that can influence wound healing

Once we have done the initial assessment of the wound bed after practicing the enzymatic debridement, we must be aware that there are some factors that can have an influence in relation to the evolution of the wound healing. Factors that may lead to the stasis zone or even cause irreversible damage. These factors are infection, edema or prolonged hypotension^[21]. The latter can occur to major burns patients for reasons such as distributive shock and the use of vasoactive drugs which may difficult the healing due to bad peripheral perfusion.

To sum up, once we have already performed the enzymatic debridement treatment and while we are waiting for the spontaneous healing or for the coverage with skin autografts, it is very important to remember that we must keep the wound bed wet in order to prevent it from drying out and therefore deepening, even more important than with a regular wound. As we previously stated, we will use different creams and dressings that we can easily find in our Unit: If we believe the wound bed to be type II b or III, then we will have to do surgery,

but first it is really important to previously apply to the patient some dressings to keep the wound bed hydrated. For example, dressings such as an hydrocolloid type that must be changed every 48 h until the moment that the general conditions of the patients and the availability of the operating rooms allow us to perform the surgery. If on the other hand we consider the wound bed is type I or II a, then the injury will heal spontaneously, so that we will only have to apply a dressing that does not interfere and promotes spontaneous epithelization. At this moment, our first choice to cover the wound bed is Suprathel® and nitrofurazone cream with Mepitel®. On Suprathel® we place a non-adherent meshed dressing and finally, dry gauzes to finish the dressing. We keep this for 10 d (despite the fact that at the fifth day we should have a look and check that there haven't been setbacks. Depending on the state of the affected area, after the 10th day we continue using the same treatment or change it. With nitrofurazone cream with Mepitel® we change the dressing every 48-72 h.

When we introduced the enzymatic debridement with

Nexobrid® in our armamentarium we could only use the nitrofurazone cream in one case, because the Producer stopped the production for some months. However, now it is available again, so we have introduced its use after Nexobrid® treatment of new patients (25 patients more since January 2016, 50 patients totally), combined with a silicone meshed non-adherent dressing (Mepitel®).

Our choice, to use one or the other, depends on either if the patient is going to be treated as in-patient or as out patient. In in-patients, we prefer to apply the nitrofurazone cream combined with Mepitel®, because it favours their comfort. We change the dressings every 2-3 d, which consequently allow their cleaning. On the other hand, in out patients we prefer to apply Suprathel® because it reduces the amount of dressing changes and controls.

It is important to highlight that we have used silver sulphadiazine 1% cream, but the pseudoeschar formation promotes delayed healing and the appearance of hypertrophic tissue granulation.

When we decide that autografting is necessary in type II B and III wound bed, we usually wait at least 3-4 d before doing it, because in our first patients some skin grafts failed when the surgery was performed in the first two days after the debridement. Our opinion is that some of the eschar and product still remain on the wound bed for several days after debridement. If we wait these days before surgery, we have removed them and then, the skin grafts intake is much better.

The introduction of the new enzymatic debridement agent Nexobrid® in our Burn Centre has shown significant improvement in wound healing in general and in the overall state of major burn patients. Cumulative experiences are necessary to adapt its utilisation to other Burn Centres.

COMMENTS

Background

When a thermic, electric or chemical aggression acts in the skin for the necessary time and with enough intensity, then a local injury known as burn occurs. Nowadays, burns continue to be a common injury in Western countries. Many of these burns will need urgent medical attention, hospital admission, and even surgery. The majority of burns present with different degrees in all its extension through the skin. Consequently, it can be extremely complicated to differentiate between a superficial partial thickness burn and a deep partial thickness one. The diagnosis of burn depth is really important in order to plan the treatment. While the epidermal and superficial partial thickness burns will heal spontaneously, the deep partial thickness as well as the full thickness burns will need surgery. For the latter, the tangential debridement followed by coverage with skin autograft is considered to be the current standard of care. An alternative to surgical debridement is enzymatic or chemical debridement, although past reports that used it on patients show that its efficiency is limited, it is a slow process and increases the infection risk due to maceration of necrotic tissue. The introduction of a new selective enzymatic debridement agent (Nexobrid®) in armamentarium for burn wound management has provided us some new concepts in the initial management and wound healing as well. In this study, the authors show learning curve of the procedure, lessons learned, advantages and disadvantages compared to the standard of care.

Research frontiers

Nexobrid® is a selective enzymatic debridement agent for burn wound

management. The results of this study contribute to a better understanding the procedure and to avoid mistakes in its use.

Innovations and breakthroughs

The efficacy of Nexobrid® for burn wound management has been previously well established. In this study, the authors show learning curve of the procedure and give some advises for a better understanding of the remaining wound bed after debridement, which can help in the diagnosis of burn depth and the plan of treatment.

Applications

This study suggests that Nexobrid® is useful for burn wound management, specially the mid-partial, deep-partial and full thickness burns which can benefit in the reduction of the need for autografting compared with the standard of care. In major burns, besides the improvement in wound healing, the authors observed an important improvement in their general state.

Peer-review

The manuscript is an interesting and well written study.

REFERENCES

- 1 **Hettiaratchy S**, Dziewulski P. ABC of burns. Introduction. *BMJ* 2004; **328**: 1366-1368 [PMID: 15178618 DOI: 10.1136/bmj.328.7452.1366]
- 2 **Hlava P**, Moserová J, Königová R. Validity of clinical assessment of the depth of a thermal injury. *Acta Chir Plast* 1983; **25**: 202-208 [PMID: 6199924]
- 3 **Palao R**. Quemados: Valoración y criterios de actuación. Barcelona: Marge Médica Books, 2011: 19-52
- 4 **Hettiaratchy S**, Papini R. Initial management of a major burn: II -assessment and resuscitation. *BMJ* 2004; **329**: 101-103 [PMID: 15242917 DOI: 101136/bmj.329.7457.101]
- 5 **Papini R**. Management of burn injuries of various depths. *BMJ* 2004; **329**: 158-160 [PMID: 15258073 DOI: 10.1136/bmj.329.7458.158]
- 6 **Gómez PA**, Palao R. Tratamiento de las quemaduras en el siglo XXI desde la cirugía. *Cir Plast Iberoamer* 2002; **28**: 69
- 7 **Heimbach DM**. Early burn excision and grafting. *Surg Clin North Am* 1987; **67**: 93-107 [PMID: 3544269 DOI: 10.1016/S0039-6109(16)44135-6]
- 8 **Klasen HJ**. A review on the nonoperative removal of necrotic tissue from burn wounds. *Burns* 2000; **26**: 207-222 [PMID: 10741585 DOI: 10.1016/S0305-4179(99)00117-5]
- 9 **Rosenberg L**, Lapid O, Bogdanov-Berezovsky A, Glesinger R, Krieger Y, Silberstein E, Sagi A, Judkins K, Singer AJ. Safety and efficacy of a proteolytic enzyme for enzymatic burn debridement: a preliminary report. *Burns* 2004; **30**: 843-850 [PMID: 15555800 DOI: 10.1016/j.burns.2004.04.010]
- 10 **Krieger Y**, Rosenberg L, Lapid O, Glesinger R, Bogdanov-Berezovsky A, Silberstein E, Sagi A, Judkins K. Escharotomy using an enzymatic debridement agent for treating experimental burn-induced compartment syndrome in an animal model. *J Trauma* 2005; **58**: 1259-1264 [PMID: 15995479 DOI: 10.1097/01.TA.0000169867.08607.F1]
- 11 **Rosenberg L**, Krieger Y, Silberstein E, Arnon O, Sinelnikov IA, Bogdanov-Berezovsky A, Singer AJ. Selectivity of a bromelain based enzymatic debridement agent: a porcine study. *Burns* 2012; **38**: 1035-1040 [PMID: 22385643 DOI: 10.1016/j.burns.2012.02.011]
- 12 **Krieger Y**, Bogdanov-Berezovsky A, Gurfinkel R, Silberstein E, Sagi A, Rosenberg L. Efficacy of enzymatic debridement of deeply burned hands. *Burns* 2012; **38**: 108-112 [PMID: 22103988 DOI: 10.1016/j.burns.2011.06.002]
- 13 **Rosenberg L**, Krieger Y, Bogdanov-Berezovski A, Silberstein E, Shoham Y, Singer AJ. A novel rapid and selective enzymatic debridement agent for burn wound management: a multi-center RCT. *Burns* 2014; **40**: 466-474 [PMID: 24074719 DOI: 10.1016/j.burns.2013.08.013]
- 14 **Monstrey S**, Hoeksema H, Verbelen J, Pirayesh A, Blondeel P. Assessment of burn depth and burn wound healing potential. *Burns* 2008; **34**: 761-769 [PMID: 18511202 DOI: 10.1016/j.burns.

- 2008.01.009]
- 15 **Khatib M**, Jabir S, Fitzgerald O'Connor E, Philp B. A systematic review of the evolution of laser Doppler techniques in burn depth assessment. *Plast Surg Int* 2014; **2014**: 621792 [PMID: 25180087 DOI: 10.1155/2014/621792]
 - 16 **Sheridan R**, Remensnyder J, Prelack K, Petras L, Lydon M. Treatment of the seriously burned infant. *J Burn Care Rehabil* 1998; **19**: 115-118 [PMID: 9556311 DOI: 10.1097/00004630-199803000-00005]
 - 17 **Keck M**, Lumenta DB, Andel H, Kamolz LP, Frey M. Burn treatment in the elderly. *Burns* 2009; **35**: 1071-1079 [PMID: 19520515 DOI: 10.1016/j.burns.2009.03.004]
 - 18 **Wilmore DW**, Long JM, Mason AD, Skreen RW, Pruitt BA. Catecholamines: mediator of the hypermetabolic response to thermal injury. *Ann Surg* 1974; **180**: 653-669 [PMID: 4412350 DOI: 10.1097/0000658-197410000-00031]
 - 19 **Yamada Y**, Endo S, Inada K. Plasma cytokine levels in patients with severe burn injury--with reference to the relationship between infection and prognosis. *Burns* 1996; **22**: 587-593 [PMID: 8982534 DOI: 10.1016/S0305-4179(96)00052-6]
 - 20 **Cotsarelis G**. Epithelial stem cells: a folliculocentric view. *J Invest Dermatol* 2006; **126**: 1459-1468 [PMID: 16778814 DOI: 10.1038/sj.jid.5700376]
 - 21 **Hettiaratchy S**, Dziewulski P. ABC of burns: pathophysiology and types of burns. *BMJ* 2004; **328**: 1427-1429 [PMID: 15191982 DOI: 10.1136/bmj.328.7453.1427]

P- Reviewer: Aksoy B, Negosanti L **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ





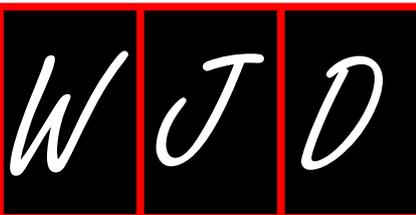
Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>



World Journal of *Dermatology*

World J Dermatol 2017 August 2; 6(3): 42-51





World Journal of Dermatology

A peer-reviewed, online, open-access journal of dermatology

Editorial Board

2017-2020

The *World Journal of Dermatology* Editorial Board consists of 139 members, representing a team of worldwide experts in dermatology. They are from 39 countries, including Argentina (1), Australia (1), Austria (1), Brazil (1), Bulgaria (1), Canada (4), China (10), Croatia (1), Denmark (1), Egypt (1), Finland (1), France (3), Germany (5), Greece (4), Hungary (2), India (3), Iran (3), Israel (1), Italy (16), Japan (5), Malaysia (1), Malta (1), Mexico (4), Netherlands (3), Nigeria (2), Norway (1), Oman (1), Poland (2), Portugal (1), Romania (1), Saudi Arabia (1), Singapore (2), South Korea (8), Spain (8), Swaziland (2), Thailand (2), Turkey (6), United Kingdom (9), United States (19).

EDITOR-IN-CHIEF

Santosh K Katiyar, *Birmingham*

GUEST EDITORIAL BOARD MEMBERS

Tsong-Min Chang, *Tsichung*

Ching-Chi Chi, *Chiayi*

Jia-You Fang, *Taoyuan*

Sindy Hu, *Taipei*

Stephen Chu-Sung Hu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

María D Hermida, *Buenos Aires*



Australia

Ronald Sluyter, *Wollongong*



Austria

Iris Zalaudek, *Graz*



Brazil

Cidia Vasconcellos, *São Paulo*



Bulgaria

Georgi Tchernev, *Sofia*



Canada

Eleftherios P Diamandis, *Toronto*

Tim Lee, *Vancouver*

Gang Li, *Vancouver*

Kursad Turksen, *Ottawa*



China

Henry Hin Lee Chan, *Hong Kong*

Min Li, *Nanjing*

Cheng Tan, *Nanjing*

Guo-You Zhang, *Wenzhou*

Min Zheng, *Hangzhou*



Croatia

Mariastefania Antica, *Zagreb*



Denmark

Lars Iversen, *Aarhus*



Egypt

Moetaz El-Domyati, *Cairo*



Finland

Kari J Syrjänen, *Turku*



France

Guinot J Christiane, *Neully sur Seine*

Roger Mouawad, *Paris*

Stephane Rocchi, *Nice*



Germany

Martin Leverkus, *Mannheim*

Roderick AF MacLeod, *Braunschweig*

Markus Meissner, *Frankfurt*

Enno Schmidt, *Luebeck*

Peter Schroeder, *Dusseldorf*



Greece

Ioannis D Bassukas, *Ioannina*

Maria Dalamaga, *Athens*

Andreas Katsambas, *Athens*

Eleni Sotiriou, *Thessaloniki*



Hungary

Arpad Farkas, *Szeged*

Janos Fodor, *Budapest*



India

Sujoy Khan, *Kolkata*

Harsh Mohan, *Chandigarh*

Davinder Parsad, *Chandigarh*

**Iran**

Alireza Firooz, *Tehran*
 Mohammad R Namazi, *Shiraz*
 Afshin Sadighha, *Ilam*

**Israel**

Ronni Wolf, *Rehovo*

**Italy**

Giuseppe Argenziano, *Naples*
 Laura Atzori, *Cagliari*
 Ettore D Capoluongo, *Rome*
 Dott V Di Lernia, *Reggio Emilia*
 Paolo Fabbri, *Florence*
 Gabriella Fabbrocini, *Naples*
 Silvano Gallus, *Milan*
 Torello Lotti, *Firenze*
 Clelia Miracco, *Cosenza*
 Agnese Molinari, *Rome*
 Pierfrancesco Morganti, *Rome*
 Luigi Naldi, *Bergamo*
 Luca Negosanti, *Bologna*
 Raffaele Palmirotta, *Rome*
 Mario Santinami, *Milano*
 Riccarda Serri, *Milano*

**Japan**

Masutaka Furue, *Fukuoka*
 Fukumi Furukawa, *Wakayama*
 Mohammad Ghazizadeh, *Kawasaki*
 Yohei Tanaka, *Matsumoto*
 Toshiyuki Yamamoto, *Tokyo*

**Malaysia**

Felix Boon-Bin Yap, *Kuala Lumpur*

**Malta**

Michael J Boffa, *Floriana*

**Mexico**

Roberto G Arenas, *Mexico*
 Sergio A Cuevas-Covarrubias, *Mexico*
 Leopoldo Flores-Romo, *Mexico*
 Maria Bertha Torres-alvarez, *San Luis Potosi*

**Netherlands**

Rosalie M Luiten, *Amsterdam*

Arnold P Oranje, *Rotterdam*
 Arnold C Spek, *Amsterdam*

**Nigeria**

Maurice E Asuquo, *Calabar*
 Joseph I Ikechebelu, *Nnewi*

**Norway**

Andrej M Grijbovski, *Oslo*

**Oman**

Mohamed Mabruk, *Muscat*

**Poland**

Andrzej Grzybowski, *Poznan*
 Lidia Rudnicka, *Warsaw*

**Portugal**

Bruno Sarmento, *Porto*

**Romania**

Liana Manolache, *Bucharest*

**Saudi Arabia**

Feroze Kaliyadan, *Hofuf*

**Singapore**

Wei-Sheng Chong, *Singapore*
 Hong Liang Tey, *Singapore*

**South Korea**

Dong-Seok Kim, *Seoul*
 Chang Hoon Lee, *Seoul*
 Jongsung Lee, *Seongnam City*
 Chil Hwan Oh, *Seoul*
 Byung Soon Park, *Seoul*
 Myung-Geun Shin, *Hwasun*
 Jong-Hyuk Sung, *Seoul*
 Young Kwan Sung, *Daegu*

**Spain**

Agustin Alomar, *Barcelona*
 Salvador Arias-Santiago, *Granada*
 Juan G Gavín, *Vigo*
 Marcos A Gonzalez-Lopez, *Santander*

Ramon Grimalt, *Barcelona*
 Husein Husein-ElAhmed, *Granada*
 Ander Izeta, *San Sebastian*
 Marcela Del Rio, *Madrid*

**Switzerland**

Gunther FL Hofbauer, *Zurich*
 Alexander A Navarini, *Zurich*

**Thailand**

Chirayu U Auewarakul, *Bangkok*
 Viroj Wiwanitkit, *Bangkok*

**Turkey**

Berna Aksoy, *Kocaeli*
 Fatma Aydin, *Samsun*
 Cem Dane, *Istanbul*
 Sibel Dogan, *Istanbul*
 Aylin T Ermertcan, *Manisa*
 Ozlem Su, *Istanbul*

**United Kingdom**

Theodoros Dimitroulas, *Dudley*
 Bernhard F Gibbs, *Chatham Maritime*
 Evmorfia Ladoyanni, *Stourbridge*
 Mark R Nelson, *London*
 Adrian V Pace, *Dudley*
 Anthony B Paul, *London*
 Sam Shuster, *Woodbridge*
 Olga Tura, *Edinburgh*
 Indre Verpetinske, *Stourbridge*

**United States**

Jeremy S Bordeaux, *Cleveland*
 Robert F Diegelmann, *Richmond*
 Q Ping Dou, *Detroit*
 Zeev Estrov, *Houston*
 Vincent Falanga, *Providence*
 Miranda A Farage, *Cincinnati*
 Markus H Frank, *Boston*
 W Scott Goebel, *Indianapolis*
 Dan-Ning Hu, *New York*
 Amor Khachemoune, *Brooklyn*
 Arash Kimyai-Asadi, *Houston*
 Michael S Kolodney, *Torrance*
 Feng Liu, *Chapel Hill*
 Senthamil R Selvan, *San Diego*
 Lei Shi, *Fort Worth*
 Animesh A Sinha, *East Lansing*
 Jeffrey M Weinberg, *New York*
 John A Zic, *Nashville*

MINIREVIEWS

- 42 Atopic eczema treatment now and in the future: Targeting the skin barrier and key immune mechanisms in human skin
Bell DC, Brown SJ

ABOUT COVER

Editorial Board Member of *World Journal of Dermatology*, Raffaele Palmirotta, MD, PhD, Professor, Oncogenomic Research Center, Department of Internal Medicine and Clinical Oncology, University of Bari "A. Moro", 00163 Rome, Italy

AIM AND SCOPE

World Journal of Dermatology (*World J Dermatol*, *WJD*, online ISSN 2218-6190, DOI: 10.5314), is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of dermatology. *WJD* covers fungal diseases, dermatitis and eczema, urticarial diseases, drug eruptions, pruritus, erythroderma desquamativum, connective tissue diseases, bullous skin diseases, vascular skin diseases, skin appendage diseases, pigmentary diseases, genetic diseases, nutritional and metabolic disorders, tumors, sexually transmitted diseases, AIDS, traditional medicine, integrated Chinese and Western medicine, evidence-based medicine, epidemiology and nursing. The journal also publishes original articles and reviews that report the results of applied and basic research in fields related to dermatology, such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Dermatology is now indexed in China National Knowledge Infrastructure (CNKI).

FLYLEAF

I-II Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Yuan Qi*

NAME OF JOURNAL
World Journal of Dermatology

ISSN
 ISSN 2218-6190 (online)

LAUNCH DATE
 June 2, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Santosh K Katiyar, PhD, Professor, Department of Dermatology, University of Alabama at Birmingham, Birmingham, AL 35294, United States

EDITORIAL BOARD MEMBERS
 All editorial board members resources online at <http://www.wjgnet.com/2218-6190/editorialboard.htm>

EDITORIAL OFFICE
 Fang-Fang Ji, Director

World Journal of Dermatology
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.f0publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.f0publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLICATION DATE
 August 2, 2017

COPYRIGHT
 © 2017 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION
<http://www.f0publishing.com>

Atopic eczema treatment now and in the future: Targeting the skin barrier and key immune mechanisms in human skin

David C Bell, Sara J Brown

David C Bell, Sara J Brown, Skin Research Group, School of Medicine, Ninewells Hospital and Medical School, University of Dundee, Dundee DD1 9SY, Scotland, United Kingdom

Author contributions: Bell DC performed the majority of the writing and produced the figures and table with advice from Brown SJ; Brown SJ edited and supplemented the text and provided advice in the development of the figures and table.

Conflict-of-interest statement: Bell DC has no conflict of interest to declare; Brown SJ receives funding from the Wellcome Trust (Senior Research Fellowship in Clinic Science reference 106865/Z/15/Z); she provides consultancy advice for CXR Biosciences.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Sara J Brown, Professor of Molecular and Genetic Dermatology, Wellcome Trust Senior Research Fellow in Clinical Science, Honorary Consultant Dermatologist, Skin Research Group, School of Medicine, Ninewells Hospital and Medical School, University of Dundee, James Arrott Drive, Dundee DD1 9SY, Scotland, United Kingdom. s.j.brown@dundee.ac.uk
Telephone: +44-1382-383210

Received: February 15, 2017

Peer-review started: February 15, 2017

First decision: March 7, 2017

Revised: March 14, 2017

Accepted: April 4, 2017

Article in press: April 5, 2017

Published online: August 2, 2017

Abstract

The skin facilitates a number of key roles but its functioning can be impaired by disease. Atopic eczema is a chronic inflammatory disease where the skin barrier has become leaky, and inflammation occurs. It affects up to 20% of children and 3% of adults worldwide, manifesting as red itchy patches of skin with varying severity. This review aims to investigate the leaky skin barrier and immune mechanisms from the perspective of potential novel treatments. The complexity of atopic eczema as a disease is what makes it difficult to treat. Genome-wide association studies have highlighted possible genetic variations associated with atopic eczema, however in some cases, individuals develop the disease without these genetic risk factors. Loss of function mutations in the filaggrin gene are one of these associations and this is plausible due to its key role in barrier function. The Th2 immune response is the link with regards to the immune mechanisms as atopic inflammation often occurs through increased levels of interleukin (IL)-4 and IL-13. Eczematous inflammation also creates susceptibility to colonisation and damage by bacteria such as *Staphylococcus aureus*. Potential novel treatments are becoming ever more specific, offering the hope of fewer side effects and better disease control. The best new treatments highlighted in this review target the immune response with human beta defensin 2, phosphodiesterase-4 inhibitors and monoclonal antibodies all showing promise.

Key words: Atopic eczema; Novel treatment; Filaggrin; Skin barrier; Immune dysfunction

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Atopic eczema (atopic dermatitis) is an itchy inflammatory skin disease with complex aetiology, including impairment in barrier function and concomitant inflammation. Increased understanding of the molecular

mechanisms in eczema pathogenesis has opened up opportunities for new therapeutic targets. This review summarizes current understanding and highlights some novel treatments in development.

Bell DC, Brown SJ. Atopic eczema treatment now and in the future: Targeting the skin barrier and key immune mechanisms in human skin. *World J Dermatol* 2017; 6(3): 42-51 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i3/42.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i3.42>

INTRODUCTION

Atopic eczema is a prevalent disease with substantial morbidity

Atopic eczema (also called atopic dermatitis) is a skin disease that has shown a rise in prevalence in Africa, Eastern Asia and Western Europe^[1]. It is a chronic inflammatory disease that manifests as red patches of itchy skin and in severe cases excoriated or infected lesions^[2]. Inflammation is believed to occur when the skin barrier becomes leaky and an immune response is stimulated; *vice versa*, the inflammatory response can itself impair the skin barrier function^[3].

There are two forms of eczema: Atopic, when an increased IgE response occurs, and non-atopic when this response is not observed. Eighty percent of cases are atopic^[4] and the feature of atopy (raised IgE) is what relates the disease to other allergic responses such as food allergies, allergic rhinitis and asthma which can all show an IgE response. The so-called "atopic march" describes the progressive acquisition of atopic diseases in a step-wise manner throughout childhood^[5]. Therefore children affected by one atopic disease will often show phenotypes of the others too^[5]. Atopic eczema is predominantly a childhood disease that affects up to 20% of children^[6] and this is one of the reasons why it is among the most common skin diseases worldwide^[2]. In the 2010 WHO article into the global burden skin diseases, atopic eczema was ranked first by causing the most number of days that people were not at full health^[7]. The effects of atopic eczema are not limited to skin. Children with more extreme atopic eczema suffer from reduced sleep and increased psychological problems^[8]. Atopic eczema not only has a large effect on the health of the sufferer but it also has a substantial economic effect. In the United States, the direct cost for the treatment of atopic eczema may be as high as \$3.8 billion per year^[9], studies in the United Kingdom are not recent enough to compare as one of the most recent was in 1996^[10,11]. This shows from an economic stance that research into a more cost effective treatment is of great importance.

Atopic eczema is a complex trait and current treatment options are sub-optimal

The development of specific treatment modalities in

atopic eczema is difficult due to the complexity of this disease. For example, there are a number of strong genetic risk factors associated with the disease and variations in these genes are often seen in atopic eczema; however in a subset of cases, a mutation is present but there is no disease^[12]. Another feature involved with atopic eczema is environmental allergens such as dust or animal hair which can precipitate the disease or elicit a flare up. Therefore the disease is now believed to be caused by a combination of both genetic and environmental factors^[13]; this complexity is why treatments are only partially effective and why there is currently no cure^[14].

Atopic eczema arises due to interactions between a leaky skin barrier and the immune response that occurs both in the skin and the systemic circulation; therefore current treatments aim to reduce the inflammation and repair the skin barrier at sites of inflamed or dry skin. Due to the complexity and range of severity of the disease, there are a number of different treatments^[15]. These have been summarised in Figure 1. The most commonly used treatment is the application of emollients; these act by increasing the lipid content in the stratum corneum (outermost layer of the skin) to repair the barrier, thereby improving hydration^[15]. However, in all but the mildest cases emollients are insufficiently effective so a combination therapy is used with another agent targeting the inflammatory response. Topical corticosteroids act through the corticosteroid-receptor complex to downregulate synthesis of the proteins involved in inflammation^[16]. Topical corticosteroids come in different forms from sprays to creams and ointments but more potent steroids must be used sparingly as they have been found to reduce dermal thickness^[17]. Topical calcineurin inhibitors also target the immune mechanisms and act by binding to intracellular protein macropilin-12, thereby decreasing the production of inflammatory cytokines interleukin (IL)-4 and IL-13; this treatment does not decrease dermal thickness^[18,19].

Other treatments include wet-wraps^[15,20], oral antihistamines^[13] and phototherapy^[21] where ultraviolet light is administered to the skin for its immunosuppressive effect. Atopic eczema sufferers frequently have to undergo two or more treatments (Figure 1), one for the disease itself and the other for co-existent bacterial or viral infection^[22].

Staphylococcus aureus (*S. aureus*) is a bacterium that may be carried in the nose and flexural skin of some individuals and on the apparently healthy skin of atopic individuals. In individuals with atopic eczema it can cause infections within actively inflamed lesions and lead to increased skin barrier damage^[22]. There is a more permissive environment for the growth of *S. aureus* because atopic skin shows a reduction in the expression of antimicrobial peptides^[23]. Topical corticosteroids have been found to reduce the colonisation by *S. aureus*^[24], but the most effective treatment involves combining these topical corticosteroids with an antimicrobial preparation^[25]. However, this combination therapy only shows

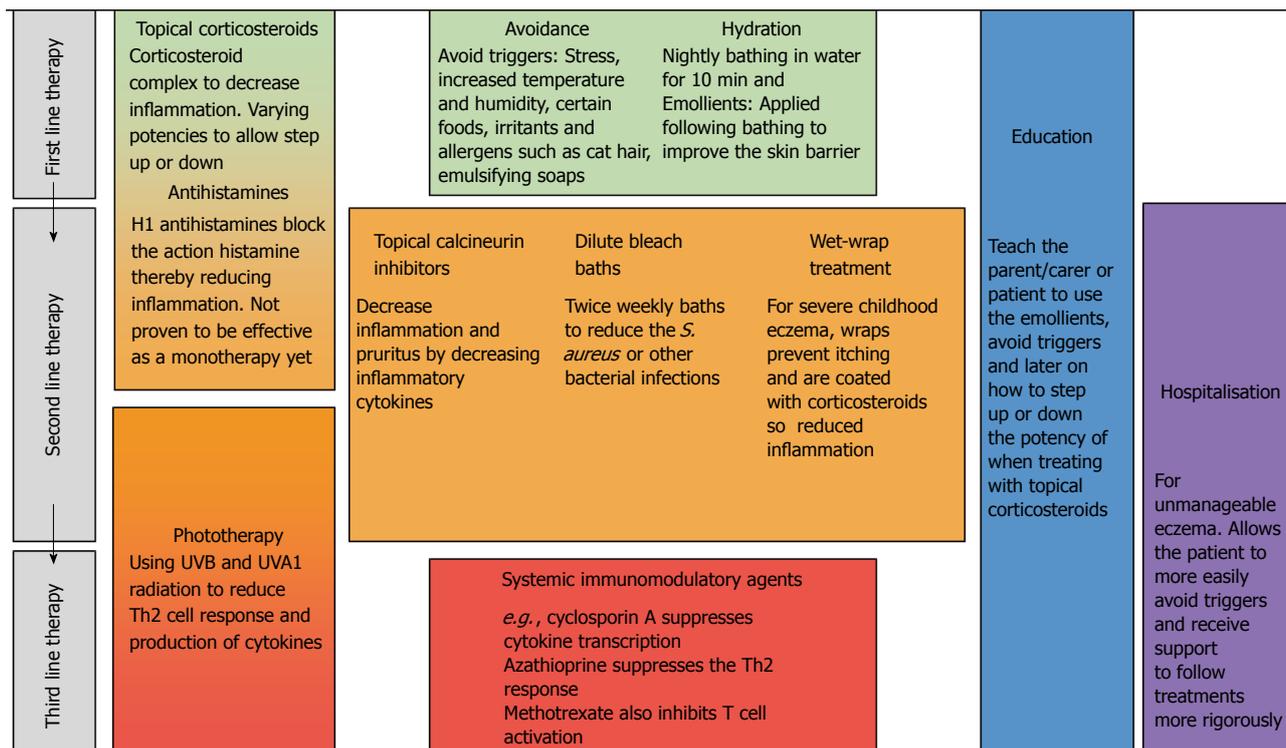


Figure 1 Current treatment options for atopic eczema. A summary of current treatment for atopic eczema, showing stepwise progression for more severe or treatment-resistant disease^[13,15-18,20,21].

efficacy in the short-term as over a long period there is no significant benefit in comparison to corticosteroids alone. Therefore a more effective treatment is required to prevent *S. aureus* re-infection.

The stepwise approach to treating atopic eczema, shown in Figure 1, highlights the varying severity in this disease. It also illustrates the range of treatments in use, because of the variation in individual response to each therapy. The currently available treatments target atopic eczema in a rather non-specific way; it is hoped that novel treatments, such as monoclonal antibodies, will be able to target the specific problem(s) in each individual's atopic eczema.

WHAT IS KNOWN ABOUT BARRIER FUNCTION AND IMMUNE MECHANISMS IN ATOPIC SKIN?

This review will focus on the skin barrier and the immune mechanisms of the skin and how irregular function of both lead to atopic eczema, as well as the novel and theoretical treatments designed for targeting each component of the disease.

The skin barrier plays an essential role in protecting the body against the entry of allergens and loss of water

The skin is the largest organ in the body^[26] and it plays a number of key roles in survival. A central function is to act as a physical barrier to prevent the entry of allergens and irritants while also vitally retaining water

within the body^[4]. The skin is composed of three main layers: epidermis, dermis and hypodermis^[27]. The outermost layer of the epidermis is called the stratum corneum (Figure 2); this contributes to the control of trans-epidermal water loss (TEWL)^[28], *i.e.*, water lost through evaporation. The stratum corneum includes 18-20 layers constructed from dead cells containing keratin called corneocytes; this is surrounded by a matrix of lipids mainly consisting of ceramides and cholesterol^[29]. The epidermis provides the physical skin barrier function through the matrix of lipids but also through corneodesmosomes and tight junctions with the stratum granulosum layer below^[29].

An essential component of the barrier function of the skin is filaggrin (filament-aggregating protein), an intracellular protein^[30] important for formation of the stratum corneum^[31]. Filaggrin is formed from the dephosphorylation and proteolysis of profilaggrin when the keratinocytes in the stratum granulosum are undergoing differentiation to the corneocytes of the stratum corneum^[32]. Filaggrin monomers bind to keratin molecules to strengthen the filament network and contribute to the changes in shape of keratinocytes as they mature into corneocytes. Filaggrin also plays a key role in control of TEWL. Filaggrin undergoes proteolysis to release hygroscopic amino acids at the surface of the stratum corneum, when the outer skin starts to become dehydrated. These amino acids contribute to natural moisturising factor (NMF) for the skin, maintaining hydration of the stratum corneum and controlling TEWL^[33]. Another key role played by filaggrin

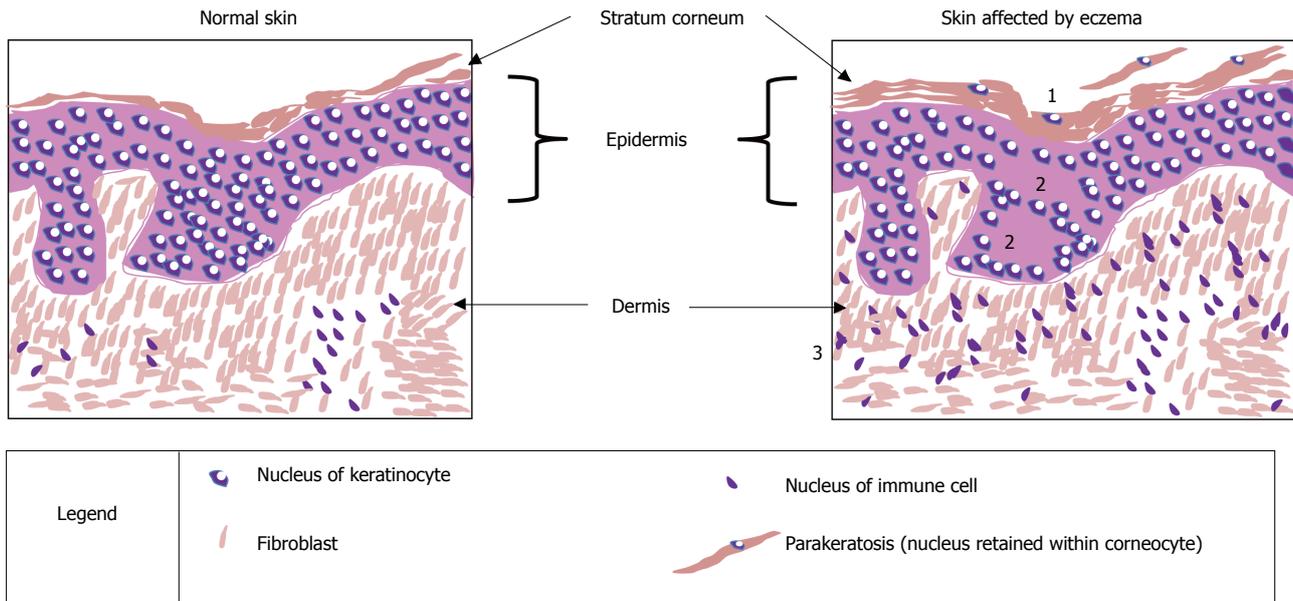


Figure 2 Histological features of atopic eczema. This figure represents sections of skin biopsies, stained with haematoxylin and eosin, to highlight the changes that can be observed under a light microscope, comparing skin affected by atopic eczema and a control sample. Three characteristic features of atopic eczema are illustrated: Increased thickness of the stratum corneum (hyperkeratosis and parakeratosis) is caused by a disruption to the cornification process; spongiosis occurs when there is a reduction or damage of the proteins involved in tight junctions, thereby leading to uncontrolled movement of fluids in the paracellular space; infiltration of the dermis by immune cells is a sign of the immune response as a primary feature of atopic eczema itself or in response to the entry of allergens through a leaky skin barrier. The effects of eczema: 1: Increased thickness of stratum corneum; 2: Spongiosis - oedema (water retained between cells); 3: Increased number of immune cells.

and its degradation products is in the control of skin pH. The acidic pH of skin acts as an antimicrobial defensive mechanism to limit bacterial colonisation^[34]. If the pH of the skin is increased, filaggrin proteolysis can contribute acidic amino acids to return it to the optimal slightly acidic pH^[34].

Tight junctions hold the keratinocytes together, control the flow of fluids paracellularly and are key as they act as another barrier; the stratum corneum acts as the first physical barrier to allergens and irritants while tight junctions form the second^[35]. This barrier is particularly important if the stratum corneum becomes diseased, since the tight junctions provide a second line of defence against allergens, to prevent their entry into the skin and the resultant immune response.

Atopic eczema has profound effects on the skin barrier of an affected individual

The difficulty in describing the causes of atopic eczema are that the mutations or genetic variants being proposed as the culprits of the skin barrier dysfunction only occur in a proportion of affected individuals. There have been a number of genetic studies aiming to highlight potential risk factors for atopic eczema; they have discovered links between certain mutations or genetic variants that are associated with increased risk for the disease^[36-40]. The majority of candidate gene association studies point to null mutations in the filaggrin gene, *FLG*, and genes involved with the type 2 T helper lymphocytes (Th2 cell) function^[41,42]. Loss of function mutations in *FLG* were first identified in 2006 and this remains the strongest genetic risk factor for

atopic eczema identified to date^[43].

The section above highlighted the importance of the protein filaggrin in a number of key roles involved with producing the skin barrier; therefore it is understandable that a loss of function mutation in *FLG* may increase the risk of atopic eczema. Other related diseases such as ichthyosis vulgaris, atopic asthma, allergic rhinitis and food allergies are also often associated with mutations in filaggrin^[44]. Filaggrin is key in cross-linking keratins to flatten the shape of cells, in the stratum corneum; consequently null mutations will lead to malformed corneocytes and allergens may be able to enter through this leaky skin barrier and incite an inflammatory response^[26]. In atopic eczema there is an increased rate of TEWL and again it is possible that filaggrin may play a role, as null mutations mean filaggrin cannot be degraded to form NMF, hence skin hydration is reduced^[26]. Another often vital part of the disease is the colonisation by bacteria and this is more likely to occur in the alkaline conditions of the skin when filaggrin is absent or reduced in amount^[45]. This is one of possibly several factors that allows binding of bacteria such as *S. aureus*, and it can contribute to the development of atopic eczema or worsen its severity^[46]. The main mechanism by which *S. aureus* damages the skin barrier is through secretion of SspA/V8 protease. This protease acts to degrade the proteins in the corneodesmosomes in the stratum corneum but also proteins in the tight junctions of the stratum granulosum, thereby compromising both elements of the skin barrier and allowing entry of allergens and irritants^[47,48].

In some cases of atopic eczema it has been shown that key proteins involved with tight junction function, claudin-1 and claudin-23, are reduced^[35]. It was believed that mutations in the filaggrin gene may also affect tight junction functionality, however mouse models demonstrated that filaggrin insufficiency did not have a direct effect^[49]. Other mouse models have demonstrated the importance of claudin-1 in maintaining normal levels of TEWL: When mice lack this protein, they die within a day^[50]. On closer inspection, it was observed that claudin-deficient mice died of dehydration as a result of increased TEWL and this was due to non-functioning tight junctions^[50]. Therefore, the reduction in claudin-1 seen in atopic eczema patients may contribute to their increased TEWL and dry skin. Tight junctions are vital for paracellular control of fluids, as well acting as a physical barrier, and what is often observed with atopic eczema is spongiosis, where oedema is occurring between the keratinocytes in the epidermis (Figure 2).

Another characteristic of atopic eczema which can be seen as scaliness (white flaky skin) when directly observing the skin, or by light microscopy of a histological section of diseased skin, is an increased thickness of the stratum corneum^[51]. Normally epidermal cells undergo transformation from keratinocytes of the stratum basale, in the lower epidermis, to corneocytes of the stratum corneum, in the upper epidermis, and begin to shed off the skin; however in atopic eczema this cornification process is disrupted^[52]. In atopic eczema the keratinocytes retain their nucleus and remain attached instead of shedding, contributing to the thickened stratum corneum. This feature can also be seen in Figure 2.

Immune dysfunction plays a key role in eczema pathogenesis

The balance of immune mechanisms in the skin is a closely regulated process, which involves a number of different immune and non-immune cells interacting to protect the body from pathogens^[53]. However, this balance is susceptible to a number of diseases. Above, we have mostly described how skin barrier dysfunction leads to an increased immune response thereby causing atopic eczema; nevertheless the disease may also be caused by immune dysfunction leading to skin barrier damage. It has been demonstrated that IL-4 and IL-13, the two cytokines that are greatly increased in atopic eczema, are able to significantly decrease the expression of filaggrin^[54]. When IL-4 and IL-13 were incubated with keratinocytes for 24 h they decrease the expression of filaggrin^[54]. Hence these two interleukins can cause damage to the barrier *via* their effects on keratinocyte differentiation^[54]. This study also highlighted that environmental allergens such as soap and detergents would cause the same damaging effects through increased levels of IL-4 and IL-13^[54]. A different study demonstrated that histamine may

also contribute to immune dysfunction causing a leaky skin barrier and atopic eczema^[55]. This study again observed keratinocyte differentiation as a measure of barrier damage but also investigated the important barrier proteins keratin 1 and keratin 10, loricrin and filaggrin^[55]. The study showed that expression of these proteins was reduced by as much as 80%-95% in the presence of histamine thereby affecting keratinocyte differentiation and the skin barrier^[55]. The final part of this study demonstrated that histamine also caused down-regulation of the claudins involved in tight junction formation and therefore this may also contribute to the leaky skin barrier in atopic eczema^[55].

The adaptive and innate immune responses have both been highlighted as possibly playing a role in atopic eczema^[56]. The candidate gene and genome-wide association studies mentioned earlier have illustrated that variation in genes involved with the adaptive response of the Th2 cells is associated with risk of atopic eczema. In a number of cases of atopic eczema there will be increased levels of the Th2 cell and its cytokines IL-4 and IL-13; these are important for instigating inflammation^[56]. IL-4 is key for production of eosinophils and importantly IgE, which then acts through Fc ϵ RI receptors to cause mast cells degranulation, stimulating inflammation^[57]. IL-13 has not been as extensively studied in skin, however its mechanism of inflammation appears to occur through interacting with the IL-4R α receptor^[58]. Another mechanism by which atopic eczema may occur is when someone begins to scratch, causing mechanical damage. The traumatised keratinocytes release thymic stromal lymphopoietin (TSLP), another cytokine. Studies have demonstrated that TSLP levels are increased in skin affected by atopic eczema compared to normal skin^[59]. TSLP acts on dendritic cells which activate Th2 cells producing IL-4 and IL-13 resulting in a cycle of increased inflammation and atopic eczema^[59,60].

The innate immune system may also play a role in causing an individual's atopic eczema. One of the first lines of response to pathogens is by antimicrobial peptides which are secreted and activated once toll-like receptors (TLR) identify pathogens^[61]. Defects in these TLRs have been implicated in potentially allowing the colonisation of bacteria and therefore instigating atopic eczema^[62]. A study using a mouse model found that mice with defective TLR4 or TIR-domain-containing adapter-inducing interferon- β (TRIF) had increased levels of TEWL, resulting in atopic eczema^[62]. The peptides themselves are found to be reduced in eczema-affected skin and therefore fail to prevent colonisation and damage by pathogens such as *S. aureus* or infection with herpes simplex resulting in eczema herpeticum^[63]. Cathelicidin and human beta defensin 2 (h β D-2) have been shown to be reduced in atopic eczema, lowering the threshold for this damage to occur^[64,65]. *S. aureus* releases alpha and delta toxins which activate the adaptive immune response resulting

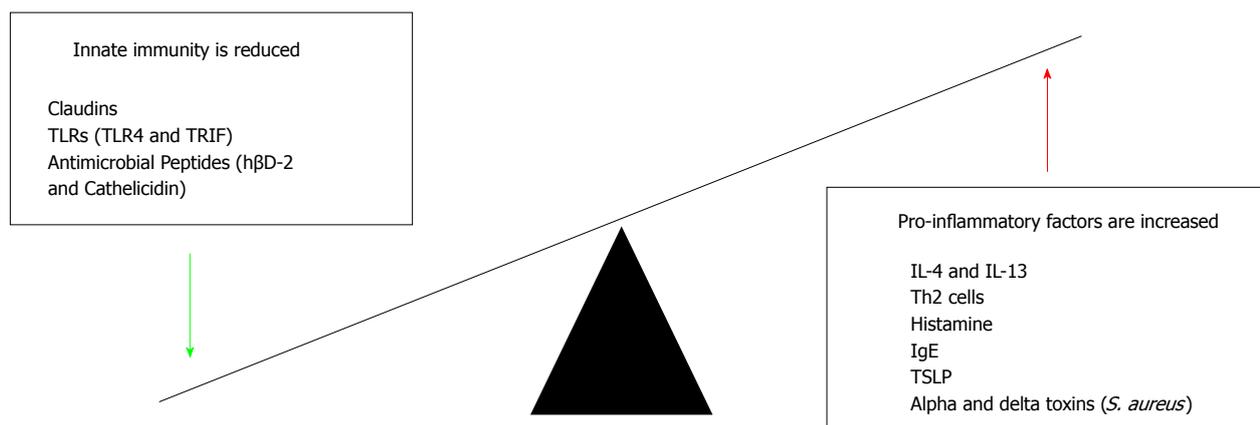


Figure 3 Immune dysfunction in atopic eczema. This figure summarizes the number of immune factors discussed in the review. Increased and decreased levels of different factors contribute to the pathogenesis atopic eczema. TLR: Toll-like receptors; hβD-2: Human beta defensin 2; *S. aureus*: *Staphylococcus aureus*; IL: Interleukin; TSLP: Thymic stromal lymphopoietin.

in increased Th2 cell activity^[53,66] and driving further inflammation (Figure 3).

HOW CAN THIS KNOWLEDGE BE APPLIED?

Novel treatments target the leaky skin barrier and immune mechanisms in eczema

This knowledge of the molecular mechanisms in skin barrier function and immune response is creating opportunities for novel treatment approaches for atopic eczema, summarised in Figure 4 and Table 1.

One potential therapeutic avenue may involve using vitamin D to decrease the severity of atopic eczema; studies have shown that it is important in both barrier repair and modulation of the immune mechanism^[67,68]. A study in mice demonstrated that those treated by phototherapy had greatly increased levels of filaggrin which also lead to decreased time for barrier repair to occur^[69]. This was believed to be due to the action of vitamin D on keratinocytes to increase levels of calcitriol, thereby normalizing the faulty keratinocyte differentiation seen in atopic eczema, to improve barrier function^[69,70]. Oral supplementation of vitamin D has not shown a therapeutic effect, so alternative methods of administration are required.

The knowledge of a central role for dry skin in atopic eczema has stimulated interest in the development of bespoke emollients as treatment for xerosis^[71]. In one study a standard emollient (the control) was compared to an emollient cream containing 5% urea, a skin ceramide N-stearoyl phytosphingosine (NP) and lactate^[71]. When skin that had previously been treated with the cream containing ceramide NP was changed to the control there was an increase in TEWL from 11.58 to 11.94 g/m² per hour; this suggested that skin barrier function was improved more by using investigated cream than the control^[71]. However this improvement between the creams may be due to the sodium lauryl sulfate in the control having an emulsifying effect

and increasing barrier damage^[71]. When hydration was also considered, the application of the ceramide NP cream showed greater hydration compared to the control, suggesting improvement of the stratum corneum barrier^[71]. There is the possibility of damage if the ceramide NP cream is used with atopic eczema as it increases pH slightly and further work is needed to define the optimal emollient treatment for atopic eczema.

Another potential treatment aimed at the leaky skin barrier involves using a synthetic elastic "second skin"^[72]. This skin is made of a cross-linked polymer (XPL) and has been used initially as anti-ageing solution where it can be applied to remove signs such as wrinkled skin; it has demonstrated dramatic results, especially around the eyes^[72]. It has been subsequently proposed as a potential treatment for atopic eczema as the XPL will act as an extra skin barrier, for up to 24 h, preventing entry of allergens or irritants to affected areas of skin^[72]. However this very interesting application remains speculative, as no research has yet been conducted.

It has recently been highlighted that increasing levels of the antimicrobial peptide hβD-2 can be used to reduce damage caused to the skin barrier by *S. aureus*^[48]. In atopic eczema there are reduced levels of hβD-2 so its IL-1β defensive mechanism cannot prevent damage by the SS_{PA}/V8 protease^[48]. It was demonstrated that purified recombinant or synthesised hβD-2 could decrease skin barrier damage by 15% and 10% respectively^[48]; this may be a useful avenue for future topical treatments.

Phosphodiesterase-4 (PDE-4) inhibitors can be taken orally to prevent cyclic-AMP degradation in cells involved in immune mechanisms of atopic eczema; however, this often leads to side effects such as nausea and diarrhoea^[73]. An ointment based PDE-4 inhibitor has therefore been developed called crisaborole; it is one of the few non-steroidal based ointments developed recently^[73]. It has just finished phase 3 clinical trials and has demonstrated improvements as high as 41%

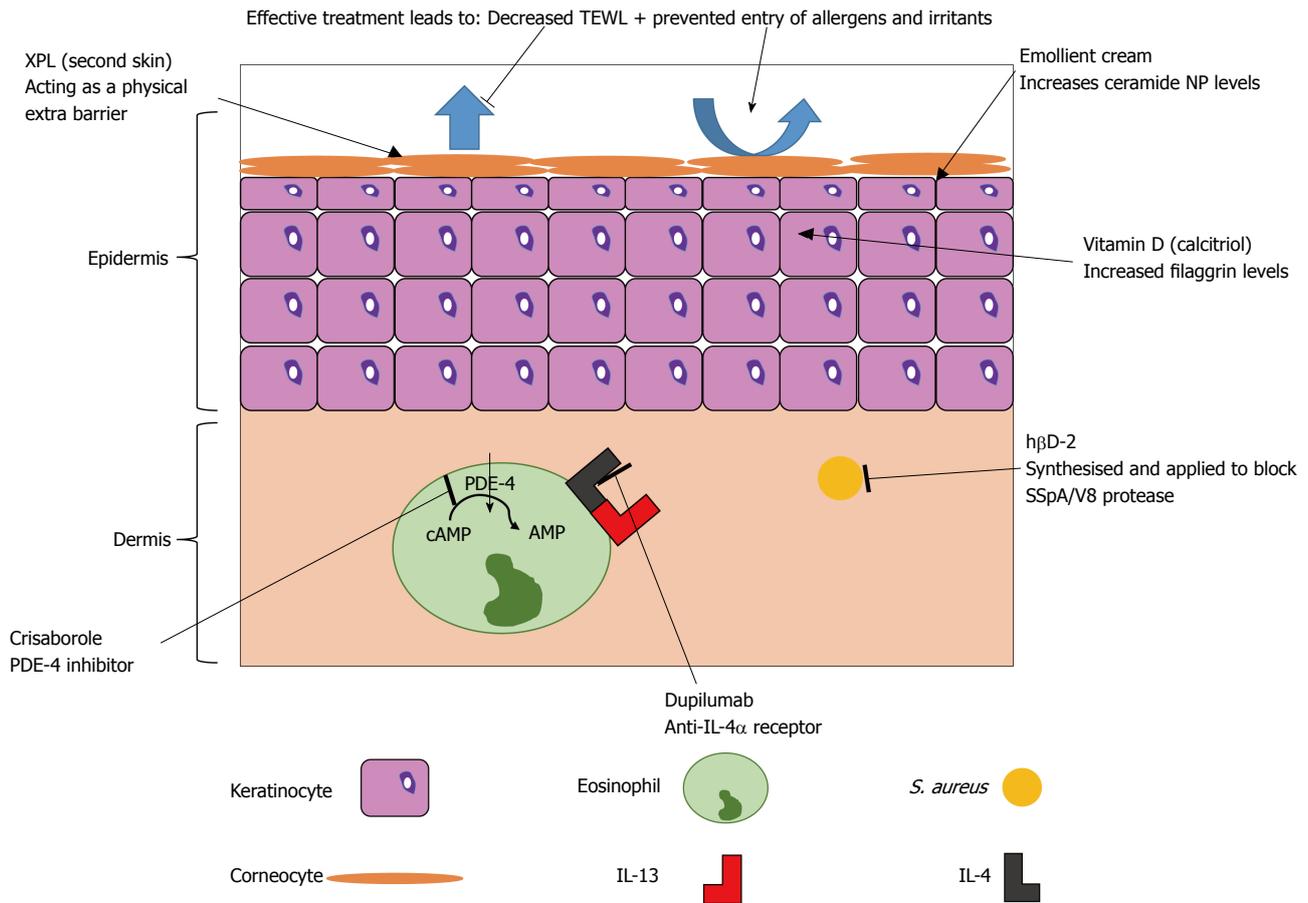


Figure 4 Novel treatment targets in atopic eczema. This figure highlights the keys roles of treatment: to decrease TEWL, prevent entry of irritants and allergens into the skin and to decrease the action of the immune response. TEWL: Trans-epidermal water loss; IL: Interleukin.

| Table 1 Novel therapeutic strategies targeting atopic eczema | |
|--|------------------------------|
| Repairing the damaged skin barrier | Reducing atopic inflammation |
| XPL (second skin) | Crisaborole |
| Emollients with increased ceramide NP levels | Dupilumab |
| Vitamin D (calcitriol) | hβD-2 |

Novel therapeutic strategies and mechanisms of action in atopic eczema. XPL: Cross-linked polymer; hβD-2: Human beta defensin 2.

compared to a placebo moisturiser, in terms of atopic eczema severity scores^[73]. This drug may soon be approved by the FDA for treatment of atopic eczema.

A novel treatment that presents the greatest opportunity to powerfully target atopic eczema inflammation involves using monoclonal antibodies which are currently being developed to treat several different atopic diseases^[74]. Currently one of the only commercially available monoclonal antibody treatments for atopic disease is omalizumab which is licenced for the treatment of asthma^[74]. It targets the IgE C ϵ 3 domain which leads to decreased severity in asthma and has the potential to be used in atopic eczema^[74].

However a more promising monoclonal antibody treatment is dupilumab, which is an anti-IL-4 α receptor,

so stops the action of IL-4, thereby preventing the inflammation by both IL-4 and IL-13^[74]. It is currently showing promising results in phase 2 trials (up to 85% of patients showing a 50% reduction in eczema severity score)^[75] and phase 3 trials (in which up to 38% of patients were clear or almost clear after 16 wk of treatment)^[76]. Drawbacks of this treatment are that it involves an injection which is more invasive than the other treatments and that the long-term safety is currently unknown^[74].

CONCLUSION

Atopic eczema is a complex and chronic inflammatory disease of the skin that affects a large proportion of people. The pathomechanisms include a leaky skin barrier and an immune response: Both are able to occur first thereby causing the other. The problems associated with atopic eczema extend far beyond skin disease, affecting a whole family and not just the individual or child affected, causing mental health problems as well as economic impact. Mutations in the gene encoding filaggrin (*FLG*) have been highlighted as an important part of the disease; filaggrin plays a number of roles in maintaining the skin barrier so this is plausible. However, not every case of atopic eczema

has these null mutations in *FLG* and the same can be seen with the immune mechanisms of the disease. The Th2 cell response often occurs in atopic eczema and is central to causing the atopic inflammation. Bacterial infection contributes to atopic eczema pathogenesis and this is potentiated *via* reduced antimicrobial peptides or mutations in filaggrin leading to a reduction in the acidic pH of skin.

The multitude of causes is what has brought about the variety of treatments for atopic eczema. Different treatments are effective or ineffective in different individuals. The ideal treatment for atopic eczema would be able to target and repair the leaky skin barrier but also normalise the increased immune response of atopic skin. In milder atopic eczema, the best treatments often involve educating patients and children to avoid their own triggers and more education may improve overall treatment. Novel treatments have become more specific in targeting molecular mechanisms in atopic eczema, which, it is hoped, will make them more effective and with fewer side effects. However a considerable amount of research is still required to develop the most effective treatment to target both key mechanisms - the skin barrier dysfunction and immune response - to fully control this complex disease.

ACKNOWLEDGMENTS

Sara J Brown holds a Wellcome Trust Senior Research Fellowship in Clinical Science (ref 106865/Z/15/Z).

REFERENCES

- 1 **Deckers IA**, McLean S, Linssen S, Mommers M, van Schayck CP, Sheikh A. Investigating international time trends in the incidence and prevalence of atopic eczema 1990-2010: a systematic review of epidemiological studies. *PLoS One* 2012; **7**: e39803 [PMID: 22808063 DOI: 10.1371/journal.pone.0039803]
- 2 **Zaniboni MC**, Samorano LP, Orfali RL, Aoki V. Skin barrier in atopic dermatitis: beyond filaggrin. *An Bras Dermatol* 2016; **91**: 472-478 [PMID: 27579743 DOI: 10.1590/abd1806-4841.20164412]
- 3 **Boguniewicz M**, Leung DY. Atopic dermatitis: a disease of altered skin barrier and immune dysregulation. *Immunol Rev* 2011; **242**: 233-246 [PMID: 21682749 DOI: 10.1111/j.1600-065X.2011.01027.x]
- 4 **Leung DY**. New insights into atopic dermatitis: role of skin barrier and immune dysregulation. *Allergol Int* 2013; **62**: 151-161 [PMID: 23712284 DOI: 10.2332/allergolint.13-RAI-0564]
- 5 **Cork MJ**, Danby SG, Vasilopoulos Y, Hadgraft J, Lane ME, Moustafa M, Guy RH, Macgowan AL, Tazi-Ahnini R, Ward SJ. Epidermal barrier dysfunction in atopic dermatitis. *J Invest Dermatol* 2009; **129**: 1892-1908 [PMID: 19494826 DOI: 10.1038/jid.2009.133]
- 6 **Asher MI**, Montefort S, Björkstén B, Lai CK, Strachan DP, Weiland SK, Williams H; ISAAC Phase Three Study Group. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet* 2006; **368**: 733-743 [PMID: 16935684 DOI: 10.1016/S0140-6736(06)69283-0]
- 7 **Hay RJ**, Johns NE, Williams HC, Bolliger IW, Dellavalle RP, Margolis DJ, Marks R, Naldi L, Weinstock MA, Wulf SK, Michaud C, J L Murray C, Naghavi M. The global burden of skin disease in 2010: an analysis of the prevalence and impact of skin conditions. *J Invest Dermatol* 2014; **134**: 1527-1534 [PMID: 24166134 DOI: 10.1038/jid.2013.446]
- 8 **Absolon CM**, Cottrell D, Eldridge SM, Glover MT. Psychological disturbance in atopic eczema: the extent of the problem in school-aged children. *Br J Dermatol* 1997; **137**: 241-245 [PMID: 9292073 DOI: 10.1046/j.1365-2133.1997.18121896.x]
- 9 **Arkwright PD**, Motala C, Subramanian H, Spergel J, Schneider LC, Wollenberg A; Atopic Dermatitis Working Group of the Allergic Skin Diseases Committee of the AAAAI. Management of difficult-to-treat atopic dermatitis. *J Allergy Clin Immunol Pract* 2013; **1**: 142-151 [PMID: 24565453 DOI: 10.1016/j.jaip.2012.09.002]
- 10 **Sach TH**, McManus E, Mcmonagle C, Levell N. Economic evidence for the prevention and treatment of atopic eczema: a protocol for a systematic review. *Syst Rev* 2016; **5**: 90 [PMID: 27230780 DOI: 10.1186/s13643-016-0262-0]
- 11 **Herd RM**, Tidman MJ, Prescott RJ, Hunter JA. The cost of atopic eczema. *Br J Dermatol* 1996; **135**: 20-23 [PMID: 8776353 DOI: 10.1111/j.1365-2133.1996.tb03601.x]
- 12 **Rodríguez E**, Baurecht H, Herberich E, Wagenpfeil S, Brown SJ, Cordell HJ, Irvine AD, Weidinger S. Meta-analysis of filaggrin polymorphisms in eczema and asthma: robust risk factors in atopic disease. *J Allergy Clin Immunol* 2009; **123**: 1361-70.e7 [PMID: 19501237]
- 13 **Apfelbacher CJ**, van Zuuren EJ, Fedorowicz Z, Jupiter A, Mattered U, Weisshaar E. Oral H1 antihistamines as monotherapy for eczema. *Cochrane Database Syst Rev* 2013; **(2)**: CD007770 [PMID: 23450580 DOI: 10.1002/14651858.CD012167]
- 14 **Nutten S**. Atopic dermatitis: global epidemiology and risk factors. *Ann Nutr Metab* 2015; **66** Suppl 1: 8-16 [PMID: 25925336 DOI: 10.1159/000370220]
- 15 **Nowicki R**, Trzeciak M, Wilkowska A, Sokołowska-Wojdyło M, Ługowska-Umer H, Barańska-Rybak W, Kaczmarek M, Kowalewski C, Kruszewski J, Maj J, Silny W, Śpiewak R, Petranjuk A. Atopic dermatitis: current treatment guidelines. Statement of the experts of the Dermatological Section, Polish Society of Allergology, and the Allergology Section, Polish Society of Dermatology. *Postepy Dermatol Alergol* 2015; **32**: 239-249 [PMID: 26366146 DOI: 10.5114/pdia.2015.53319]
- 16 **Lebwohl M**, Friedlander SF. New strategies for optimizing the treatment of inflammatory dermatoses with topical corticosteroids in an era of corticosteroid-sparing regimens. *J Am Acad Dermatol* 2005; **53**: S1-S2 [DOI: 10.1016/j.jaad.2005.04.025]
- 17 **Aschoff R**, Schmitt J, Knuschke P, Koch E, Bräutigam M, Meurer M. Evaluation of the atrophogenic potential of hydrocortisone 1% cream and pimecrolimus 1% cream in uninvolved forehead skin of patients with atopic dermatitis using optical coherence tomography. *Exp Dermatol* 2011; **20**: 832-836 [PMID: 21771098 DOI: 10.1111/j.1600-0625.2011.01335.x]
- 18 **Gutfreund K**, Bienias W, Szewczyk A, Kaszuba A. Topical calcineurin inhibitors in dermatology. Part I: Properties, method and effectiveness of drug use. *Postepy Dermatol Alergol* 2013; **30**: 165-169 [PMID: 24278069 DOI: 10.5114/pdia.2013.35619]
- 19 **Martins JC**, Martins C, Aoki V, Leonardi-Bee J, Gois AFT, Ishii HA, da Silva EMK. Topical tacrolimus for atopic dermatitis: a systematic review. *Brit J Dermatol* 2014; **170**: E43-E44
- 20 **Devillers AC**, Oranje AP. Wet-wrap treatment in children with atopic dermatitis: a practical guideline. *Pediatr Dermatol* 2012; **29**: 24-27 [PMID: 22256990 DOI: 10.1111/j.1525-1470.2011.01691.x]
- 21 **Darné S**, Leech SN, Taylor AE. Narrowband ultraviolet B phototherapy in children with moderate-to-severe eczema: a comparative cohort study. *Br J Dermatol* 2014; **170**: 150-156 [PMID: 23937117 DOI: 10.1111/bjd.12580]
- 22 **Ong PY**, Leung DY. Bacterial and Viral Infections in Atopic Dermatitis: a Comprehensive Review. *Clin Rev Allergy Immunol* 2016; **51**: 329-337 [PMID: 27377298 DOI: 10.1007/s12016-016-8548-5]
- 23 **Park HY**, Kim CR, Huh IS, Jung MY, Seo EY, Park JH, Lee DY, Yang JM. Staphylococcus aureus Colonization in Acute and Chronic Skin Lesions of Patients with Atopic Dermatitis. *Ann Dermatol* 2013; **25**: 410-416 [PMID: 24371386 DOI: 10.5021/ad.2013.25.4.410]
- 24 **Gong JQ**, Lin L, Lin T, Hao F, Zeng FQ, Bi ZG, Yi D, Zhao B. Skin colonization by Staphylococcus aureus in patients with eczema and atopic dermatitis and relevant combined topical therapy: a double-blind

- multicentre randomized controlled trial. *Br J Dermatol* 2006; **155**: 680-687 [PMID: 16965415 DOI: 10.1111/j.1365-2133.2006.07410.x]
- 25 **Friedman BC**, Goldman RD. Anti-staphylococcal treatment in dermatitis. *Can Fam Physician* 2011; **57**: 669-671 [PMID: 21673210]
- 26 **Matsui T**, Amagai M. Dissecting the formation, structure and barrier function of the stratum corneum. *Int Immunol* 2015; **27**: 269-280 [PMID: 25813515 DOI: 10.1093/intimm/dxv013]
- 27 **Chen AI**, Balter ML, Chen MI, Gross D, Alam SK, Maguire TJ, Yarmush ML. Multilayered tissue mimicking skin and vessel phantoms with tunable mechanical, optical, and acoustic properties. *Med Phys* 2016; **43**: 3117-3131 [PMID: 27277058 DOI: 10.1118/1.4951729]
- 28 **Engesland A**, Škalko-Basnet N, Flaten GE. In vitro models to estimate drug penetration through the compromised stratum corneum barrier. *Drug Dev Ind Pharm* 2016; **42**: 1742-1751 [PMID: 27019078 DOI: 10.3109/03639045.2016.1171334]
- 29 **Menon GK**, Cleary GW, Lane ME. The structure and function of the stratum corneum. *Int J Pharm* 2012; **435**: 3-9 [PMID: 22705878 DOI: 10.1016/j.ijpharm.2012.06.005]
- 30 **Gruber R**, Elias PM, Crumrine D, Lin TK, Brandner JM, Hachem JP, Presland RB, Fleckman P, Janecek AR, Sandilands A, McLean WH, Fritsch PO, Mildner M, Tschachler E, Schmuth M. Filaggrin genotype in ichthyosis vulgaris predicts abnormalities in epidermal structure and function. *Am J Pathol* 2011; **178**: 2252-2263 [PMID: 21514438 DOI: 10.1016/j.ajpath.2011.01.053]
- 31 **Kezic S**, Kemperman PM, Koster ES, de Jongh CM, Thio HB, Campbell LE, Irvine AD, McLean WH, Puppels GJ, Caspers PJ. Loss-of-function mutations in the filaggrin gene lead to reduced level of natural moisturizing factor in the stratum corneum. *J Invest Dermatol* 2008; **128**: 2117-2119 [PMID: 18305568 DOI: 10.1038/jid.2008.29]
- 32 **Cabanillas B**, Novak N. Atopic dermatitis and filaggrin. *Curr Opin Immunol* 2016; **42**: 1-8 [PMID: 27206013 DOI: 10.1016/j.coi.2016.05.002]
- 33 **Robinson M**, Visscher M, Laruffa A, Wickett R. Natural moisturizing factors (NMF) in the stratum corneum (SC). I. Effects of lipid extraction and soaking. *J Cosmet Sci* 2010; **61**: 13-22 [PMID: 20211113]
- 34 **Bandier J**, Johansen JD, Petersen LJ, Carlsen BC. Skin pH, atopic dermatitis, and filaggrin mutations. *Dermatitis* 2014; **25**: 127-129 [PMID: 24819286 DOI: 10.1097/DER.0000000000000045]
- 35 **De Benedetto A**, Rafaels NM, McGirt LY, Ivanov AI, Georas SN, Cheadle C, Berger AE, Zhang K, Vidyasagar S, Yoshida T, Boguniewicz M, Hata T, Schneider LC, Hanifin JM, Gallo RL, Novak N, Weidinger S, Beatty TH, Leung DY, Barnes KC, Beck LA. Tight junction defects in patients with atopic dermatitis. *J Allergy Clin Immunol* 2011; **127**: 773-786.e1-e7 [PMID: 21163515 DOI: 10.1016/j.jaci.2010.10.018]
- 36 **Barnes KC**. An update on the genetics of atopic dermatitis: scratching the surface in 2009. *J Allergy Clin Immunol* 2010; **125**: 16-29.e1-e11; quiz 30-31 [PMID: 20109730 DOI: 10.1016/j.jaci.2009.11.008]
- 37 **Esparza-Gordillo J**, Weidinger S, Fölster-Holst R, Bauerfeind A, Ruschendorf F, Patone G, Rohde K, Marenholz I, Schulz F, Kerscher T, Hubner N, Wahn U, Schreiber S, Franke A, Vogler R, Heath S, Baurecht H, Novak N, Rodriguez E, Illig T, Lee-Kirsch MA, Ciechanowicz A, Kurek M, Piskackova T, Macek M, Lee YA, Ruether A. A common variant on chromosome 11q13 is associated with atopic dermatitis. *Nat Genet* 2009; **41**: 596-601 [PMID: 19349984 DOI: 10.1038/ng.347]
- 38 **Sun LD**, Xiao FL, Li Y, Zhou WM, Tang HY, Tang XF, Zhang H, Schaarschmidt H, Zuo XB, Foelster-Holst R, He SM, Shi M, Liu Q, Lv YM, Chen XL, Zhu KJ, Guo YF, Hu DY, Li M, Li M, Zhang YH, Zhang X, Tang JP, Guo BR, Wang H, Liu Y, Zou XY, Zhou FS, Liu XY, Chen G, Ma L, Zhang SM, Jiang AP, Zheng XD, Gao XH, Li P, Tu CX, Yin XY, Han XP, Ren YQ, Song SP, Lu ZY, Zhang XL, Cui Y, Chang J, Gao M, Luo XY, Wang PG, Dai X, Su W, Li H, Shen CP, Liu SX, Feng XB, Yang CJ, Lin GS, Wang ZX, Huang JQ, Fan X, Wang Y, Bao YX, Yang S, Liu JJ, Franke A, Weidinger S, Yao ZR, Zhang XJ. Genome-wide association study identifies two new susceptibility loci for atopic dermatitis in the Chinese Han population. *Nat Genet* 2011; **43**: 690-694 [PMID: 21666691 DOI: 10.1038/ng.851]
- 39 **Paternoster L**, Standl M, Chen CM, Ramasamy A, Bønnelykke K, Duijts L, Ferreira MA, Alves AC, Thyssen JP, Albrecht E, Baurecht H, Feenstra B, Sleiman PM, Hysi P, Warrington NM, Curjurić I, Myhre R, Curtin JA, Groen-Blokhuis MM, Kerkhof M, Sääf A, Franke A, Ellinghaus D, Fölster-Holst R, Dermitzakis E, Montgomery SB, Prokisch H, Heim K, Hartikainen AL, Pouta A, Pekkanen J, Blakemore AI, Buxton JL, Kaakinen M, Duffy DL, Madden PA, Heath AC, Montgomery GW, Thompson PJ, Matheson MC, Le Souëf P; Australian Asthma Genetics Consortium (AAGC), St Pourcain B, Smith GD, Henderson J, Kemp JP, Timpson NJ, Deloukas P, Ring SM, Wichmann HE, Müller-Nurasyid M, Novak N, Klopp N, Rodríguez E, McArdle W, Linneberg A, Menné T, Nohr EA, Hofman A, Uitterlinden AG, van Duijn CM, Rivadeneira F, de Jongste JC, van der Valk RJ, Wjst M, Jogi R, Geller F, Boyd HA, Murray JC, Kim C, Mentch F, March M, Mangino M, Spector TD, Bataille V, Pennell CE, Holt PG, Sly P, Tiesler CM, Thierring E, Illig T, Imboden M, Nystad W, Simpson A, Hottenga JJ, Postma D, Koppelman GH, Smit HA, Söderhäll C, Chawes B, Kreiner-Møller E, Bisgaard H, Melén E, Boomsma DI, Custovic A, Jacobsson B, Probst-Hensch NM, Palmer LJ, Glass D, Hakonarson H, Melbye M, Jarvis DL, Jaddoe VW, Gieger C; Genetics of Overweight Young Adults (GOYA) Consortium, Strachan DP, Martin NG, Jarvelin MR, Heinrich J, Evans DM, Weidinger S; EARly Genetics & Lifecourse Epidemiology (EAGLE) Consortium. Meta-analysis of genome-wide association studies identifies three new risk loci for atopic dermatitis. *Nat Genet* 2011; **44**: 187-192 [PMID: 22197932 DOI: 10.1038/ng.1017]
- 40 **Cookson WO**, Ubhi B, Lawrence R, Abecasis GR, Walley AJ, Cox HE, Coleman R, Leaves NI, Trembath RC, Moffatt MF, Harper JJ. Genetic linkage of childhood atopic dermatitis to psoriasis susceptibility loci. *Nat Genet* 2001; **27**: 372-373 [PMID: 11279517 DOI: 10.1038/86867]
- 41 **Brown SJ**, Relton CL, Liao H, Zhao Y, Sandilands A, Wilson IJ, Burn J, Reynolds NJ, McLean WH, Cordell HJ. Filaggrin null mutations and childhood atopic eczema: a population-based case-control study. *J Allergy Clin Immunol* 2008; **121**: 940-46.e3 [PMID: 18313126 DOI: 10.1016/j.jaci.2008.01.013]
- 42 **Bin L**, Leung DY. Genetic and epigenetic studies of atopic dermatitis. *Allergy Asthma Clin Immunol* 2016; **12**: 52 [PMID: 27777593 DOI: 10.1186/s13223-016-0158-5]
- 43 **Smith FJ**, Irvine AD, Terron-Kwiatkowski A, Sandilands A, Campbell LE, Zhao Y, Liao H, Evans AT, Goudie DR, Lewis-Jones S, Arseculeratne G, Munro CS, Sergeant A, O'Regan G, Bale SJ, Compton JG, DiGiovanna JJ, Presland RB, Fleckman P, McLean WH. Loss-of-function mutations in the gene encoding filaggrin cause ichthyosis vulgaris. *Nat Genet* 2006; **38**: 337-342 [PMID: 16444271 DOI: 10.1038/ng1743]
- 44 **Irvine AD**, McLean WH, Leung DY. Filaggrin mutations associated with skin and allergic diseases. *N Engl J Med* 2011; **365**: 1315-1327 [PMID: 21991953 DOI: 10.1056/NEJMr1011040]
- 45 **Lambers H**, Piessens S, Bloem A, Pronk H, Finkel P. Natural skin surface pH is on average below 5, which is beneficial for its resident flora. *Int J Cosmet Sci* 2006; **28**: 359-370 [PMID: 18489300 DOI: 10.1111/j.1467-2494.2006.00344.x]
- 46 **Lin YT**, Wang CT, Chiang BL. Role of bacterial pathogens in atopic dermatitis. *Clin Rev Allergy Immunol* 2007; **33**: 167-177 [PMID: 18163223 DOI: 10.1007/s12016-007-0044-5]
- 47 **Tamber S**, Cheung AL. SarZ promotes the expression of virulence factors and represses biofilm formation by modulating SarA and agr in *Staphylococcus aureus*. *Infect Immun* 2009; **77**: 419-428 [PMID: 18955469 DOI: 10.1128/IAI.00859-08]
- 48 **Wang B**, McHugh BJ, Qureshi A, Campopiano DJ, Clarke DJ, Fitzgerald JR, Dorin JR, Weller R, Davidson DJ. IL-1 β -Induced Protection of Keratinocytes against *Staphylococcus aureus*-Secreted Proteases Is Mediated by Human β -Defensin 2. *J Invest Dermatol* 2017; **137**: 95-105 [PMID: 27702565 DOI: 10.1016/j.jid.2016.08.025]
- 49 **Yokouchi M**, Kubo A, Kawasaki H, Yoshida K, Ishii K, Furuse M, Amagai M. Epidermal tight junction barrier function is altered by skin inflammation, but not by filaggrin-deficient stratum corneum. *J Dermatol Sci* 2015; **77**: 28-36 [PMID: 25511077 DOI: 10.1016/j.jdermsci.2014.11.007]
- 50 **Furuse M**, Hata M, Furuse K, Yoshida Y, Haratake A, Sugitani Y, Noda T, Kubo A, Tsukita S. Claudin-based tight junctions are crucial

- for the mammalian epidermal barrier: a lesson from claudin-1-deficient mice. *J Cell Biol* 2002; **156**: 1099-1111 [PMID: 11889141 DOI: 10.1083/jcb.200110122]
- 51 **Alsaad KO**, Ghazarian D. My approach to superficial inflammatory dermatoses. *J Clin Pathol* 2005; **58**: 1233-1241 [PMID: 16311340 DOI: 10.1136/jcp.2005.027151]
- 52 **Yang M**, Chang JM. Successful treatment of refractory chronic hand eczema with calcipotriol/betamethasone ointment: A report of three cases. *Exp Ther Med* 2015; **10**: 1943-1946 [PMID: 26640577 DOI: 10.3892/etm.2015.2729]
- 53 **Pasparakis M**, Haase I, Nestle FO. Mechanisms regulating skin immunity and inflammation. *Nat Rev Immunol* 2014; **14**: 289-301 [PMID: 24722477 DOI: 10.1038/nri3646]
- 54 **Howell MD**, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A, Schneider L, Beck LA, Barnes KC, Leung DY. Cytokine modulation of atopic dermatitis filaggrin skin expression. *J Allergy Clin Immunol* 2009; **124**: R7-R12 [PMID: 19720210 DOI: 10.1016/j.jaci.2009.07.012]
- 55 **Gschwandtner M**, Mildner M, Mlitz V, Gruber F, Eckhart L, Werfel T, Gutzmer R, Elias PM, Tschachler E. Histamine suppresses epidermal keratinocyte differentiation and impairs skin barrier function in a human skin model. *Allergy* 2013; **68**: 37-47 [PMID: 23157658 DOI: 10.1111/all.12051]
- 56 **Brandt EB**, Sivaprasad U. Th2 Cytokines and Atopic Dermatitis. *J Clin Cell Immunol* 2011; **2**: 110 [PMID: 21994899 DOI: 10.4172/2155-9899.1000110]
- 57 **Stone KD**, Prussin C, Metcalfe DD. IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol* 2010; **125**: S73-S80 [PMID: 20176269 DOI: 10.1016/j.jaci.2009.11.017]
- 58 **Hershey GK**. IL-13 receptors and signaling pathways: an evolving web. *J Allergy Clin Immunol* 2003; **111**: 677-690; quiz 691 [PMID: 12704343 DOI: 10.1067/mai.2003.1333]
- 59 **Soumelis V**, Reche PA, Kanzler H, Yuan W, Edward G, Homey B, Gilliet M, Ho S, Antonenko S, Lauerma A, Smith K, Gorman D, Zurawski S, Abrams J, Menon S, McClanahan T, de Waal-Malefyt Rd R, Bazan F, Kastelein RA, Liu YJ. Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. *Nat Immunol* 2002; **3**: 673-680 [PMID: 12055625 DOI: 10.1038/ni805]
- 60 **Corrigan CJ**, Jayaratnam A, Wang Y, Liu Y, de Waal Malefyt R, Meng Q, Kay AB, Phipps S, Lee TH, Ying S. Early production of thymic stromal lymphopoietin precedes infiltration of dendritic cells expressing its receptor in allergen-induced late phase cutaneous responses in atopic subjects. *Allergy* 2009; **64**: 1014-1022 [PMID: 19187393 DOI: 10.1111/j.1398-9995.2009.01947.x]
- 61 **Schauber J**, Gallo RL. Antimicrobial peptides and the skin immune defense system. *J Allergy Clin Immunol* 2009; **124**: R13-R18 [PMID: 19720207 DOI: 10.1016/j.jaci.2009.07.014]
- 62 **Brandt EB**, Gibson AM, Bass S, Rydyznski C, Khurana Hershey GK. Exacerbation of allergen-induced eczema in TLR4- and TRIF-deficient mice. *J Immunol* 2013; **191**: 3519-3525 [PMID: 23997219 DOI: 10.4049/jimmunol.1300789]
- 63 **Howell MD**, Wollenberg A, Gallo RL, Flaig M, Streib JE, Wong C, Pavicic T, Boguniewicz M, Leung DY. Cathelicidin deficiency predisposes to eczema herpeticum. *J Allergy Clin Immunol* 2006; **117**: 836-841 [PMID: 16630942 DOI: 10.1016/j.jaci.2005.12.1345]
- 64 **Mallbris L**, Carlén L, Wei T, Heilborn J, Nilsson MF, Granath F, Ståhle M. Injury downregulates the expression of the human cathelicidin protein hCAP18/LL-37 in atopic dermatitis. *Exp Dermatol* 2010; **19**: 442-449 [PMID: 19645825 DOI: 10.1111/j.1600-0625.2009.00918.x]
- 65 **Ong PY**, Ohtake T, Brandt C, Strickland I, Boguniewicz M, Ganz T, Gallo RL, Leung DY. Endogenous antimicrobial peptides and skin infections in atopic dermatitis. *N Engl J Med* 2002; **347**: 1151-1160 [PMID: 12374875 DOI: 10.1056/NEJMoa021481]
- 66 **Breuer K**, Wittmann M, Kempe K, Kapp A, Mai U, Dittrich-Breiholz O, Kracht M, Mrabet-Dahbi S, Werfel T. Alpha-toxin is produced by skin colonizing *Staphylococcus aureus* and induces a T helper type 1 response in atopic dermatitis. *Clin Exp Allergy* 2005; **35**: 1088-1095 [PMID: 16120092 DOI: 10.1111/j.1365-2222.2005.02295.x]
- 67 **Russell M**. Assessing the relationship between vitamin D3 and stratum corneum hydration for the treatment of xerotic skin. *Nutrients* 2012; **4**: 1213-1218 [PMID: 23112909 DOI: 10.3390/nu4091213]
- 68 **Mutgi K**, Koo J. Update on the role of systemic vitamin D in atopic dermatitis. *Pediatr Dermatol* 2013; **30**: 303-307 [PMID: 22957498 DOI: 10.1111/j.1525-1470.2012.01850.x]
- 69 **Hong SP**, Kim MJ, Jung MY, Jeon H, Goo J, Ahn SK, Lee SH, Elias PM, Choi EH. Biopositive effects of low-dose UVB on epidermis: coordinate upregulation of antimicrobial peptides and permeability barrier reinforcement. *J Invest Dermatol* 2008; **128**: 2880-2887 [PMID: 18580964 DOI: 10.1038/jid.2008.169]
- 70 **Bikle DD**, Chang S, Crumrine D, Elalieh H, Man MQ, Dardenne O, Xie Z, Arnaud RS, Feingold K, Elias PM. Mice lacking 25OHD 1alpha-hydroxylase demonstrate decreased epidermal differentiation and barrier function. *J Steroid Biochem Mol Biol* 2004; **89-90**: 347-353 [PMID: 15225799 DOI: 10.1016/j.jsbmb.2004.03.113]
- 71 **Danby SG**, Brown K, Higgs-Bayliss T, Chittock J, Albenali L, Cork MJ. The Effect of an Emollient Containing Urea, Ceramide NP, and Lactate on Skin Barrier Structure and Function in Older People with Dry Skin. *Skin Pharmacol Physiol* 2016; **29**: 135-147 [PMID: 27251427 DOI: 10.1159/000445955]
- 72 **Yu B**, Kang SY, Akthakul A, Ramadurai N, Pilkenton M, Patel A, Nashat A, Anderson DG, Sakamoto FH, Gilchrist BA, Anderson RR, Langer R. An elastic second skin. *Nat Mater* 2016; **15**: 911-918 [PMID: 27159017 DOI: 10.1038/nmat4635]
- 73 **Jarnagin K**, Chanda S, Coronado D, Ciaravino V, Zane LT, Guttman-Yassky E, Leibold MG. Crisaborole Topical Ointment, 2%: A Nonsteroidal, Topical, Anti-Inflammatory Phosphodiesterase 4 Inhibitor in Clinical Development for the Treatment of Atopic Dermatitis. *J Drugs Dermatol* 2016; **15**: 390-396 [PMID: 27050693]
- 74 **Landolina N**, Levi-Schaffer F. Monoclonal antibodies: the new magic bullets for allergy: IUPHAR Review 17. *Br J Pharmacol* 2016; **173**: 793-803 [PMID: 26620589 DOI: 10.1111/bph.13396]
- 75 **Beck LA**, Thaçi D, Hamilton JD, Graham NM, Bieber T, Rocklin R, Ming JE, Ren H, Kao R, Simpson E, Ardeleanu M, Weinstein SP, Pirozzi G, Guttman-Yassky E, Suárez-Fariñas M, Hager MD, Stahl N, Yancopoulos GD, Radin AR. Dupilumab treatment in adults with moderate-to-severe atopic dermatitis. *N Engl J Med* 2014; **371**: 130-139 [PMID: 25006719 DOI: 10.1056/NEJMoa1314768]
- 76 **Simpson EL**, Bieber T, Guttman-Yassky E, Beck LA, Blauvelt A, Cork MJ, Silverberg JL, Deleuran M, Kataoka Y, Lacour JP, Kingo K, Worm M, Poulin Y, Wollenberg A, Soo Y, Graham NM, Pirozzi G, Akinlade B, Staudinger H, Mastey V, Eckert L, Gadkari A, Stahl N, Yancopoulos GD, Ardeleanu M; SOLO 1 and SOLO 2 Investigators. Two Phase 3 Trials of Dupilumab versus Placebo in Atopic Dermatitis. *N Engl J Med* 2016; **375**: 2335-2348 [PMID: 27690741 DOI: 10.1056/NEJMoa1610020]

P- Reviewer: Atzori L, Cuevas-Covarrubias SA, Husein-EIAhmed H, Kaliyadan F, Palmirota R **S- Editor:** Ji FF **L- Editor:** A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>



World Journal of *Dermatology*

World J Dermatol 2017 November 2; 6(4): 52-65



Editorial Board

2017-2020

The *World Journal of Dermatology* Editorial Board consists of 139 members, representing a team of worldwide experts in dermatology. They are from 39 countries, including Argentina (1), Australia (1), Austria (1), Brazil (1), Bulgaria (1), Canada (4), China (10), Croatia (1), Denmark (1), Egypt (1), Finland (1), France (3), Germany (5), Greece (4), Hungary (2), India (3), Iran (3), Israel (1), Italy (16), Japan (5), Malaysia (1), Malta (1), Mexico (4), Netherlands (3), Nigeria (2), Norway (1), Oman (1), Poland (2), Portugal (1), Romania (1), Saudi Arabia (1), Singapore (2), South Korea (8), Spain (8), Swaziland (2), Thailand (2), Turkey (6), United Kingdom (9), United States (19).

EDITOR-IN-CHIEF

Santosh K Katiyar, *Birmingham*

GUEST EDITORIAL BOARD MEMBERS

Tsong-Min Chang, *Tsichung*

Ching-Chi Chi, *Chiayi*

Jia-You Fang, *Taoyuan*

Sindy Hu, *Taipei*

Stephen Chu-Sung Hu, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Argentina

María D Hermida, *Buenos Aires*



Australia

Ronald Sluyter, *Wollongong*



Austria

Iris Zalaudek, *Graz*



Brazil

Cidia Vasconcellos, *São Paulo*



Bulgaria

Georgi Tchernev, *Sofia*



Canada

Eleftherios P Diamandis, *Toronto*

Tim Lee, *Vancouver*

Gang Li, *Vancouver*

Kursad Turksen, *Ottawa*



China

Henry Hin Lee Chan, *Hong Kong*

Min Li, *Nanjing*

Cheng Tan, *Nanjing*

Guo-You Zhang, *Wenzhou*

Min Zheng, *Hangzhou*



Croatia

Mariastefania Antica, *Zagreb*



Denmark

Lars Iversen, *Aarhus*



Egypt

Moetaz El-Domyati, *Cairo*



Finland

Kari J Syrjänen, *Turku*



France

Guinot J Christiane, *Neully sur Seine*

Roger Mouawad, *Paris*

Stephane Rocchi, *Nice*



Germany

Martin Leverkus, *Mannheim*

Roderick AF MacLeod, *Braunschweig*

Markus Meissner, *Frankfurt*

Enno Schmidt, *Luebeck*

Peter Schroeder, *Dusseldorf*



Greece

Ioannis D Bassukas, *Ioannina*

Maria Dalamaga, *Athens*

Andreas Katsambas, *Athens*

Eleni Sotiriou, *Thessaloniki*



Hungary

Arpad Farkas, *Szeged*

Janos Fodor, *Budapest*



India

Sujoy Khan, *Kolkata*

Harsh Mohan, *Chandigarh*

Davinder Parsad, *Chandigarh*

**Iran**

Alireza Firooz, *Tehran*
 Mohammad R Namazi, *Shiraz*
 Afshin Sadighha, *Ilam*

**Israel**

Ronni Wolf, *Rehovo*

**Italy**

Giuseppe Argenziano, *Naples*
 Laura Atzori, *Cagliari*
 Ettore D Capoluongo, *Rome*
 Dott V Di Lernia, *Reggio Emilia*
 Paolo Fabbri, *Florence*
 Gabriella Fabbrocini, *Naples*
 Silvano Gallus, *Milan*
 Torello Lotti, *Firenze*
 Clelia Miracco, *Cosenza*
 Agnese Molinari, *Rome*
 Pierfrancesco Morganti, *Rome*
 Luigi Naldi, *Bergamo*
 Luca Negosanti, *Bologna*
 Raffaele Palmirotta, *Rome*
 Mario Santinami, *Milano*
 Riccarda Serri, *Milano*

**Japan**

Masutaka Furue, *Fukuoka*
 Fukumi Furukawa, *Wakayama*
 Mohammad Ghazizadeh, *Kawasaki*
 Yohei Tanaka, *Matsumoto*
 Toshiyuki Yamamoto, *Tokyo*

**Malaysia**

Felix Boon-Bin Yap, *Kuala Lumpur*

**Malta**

Michael J Boffa, *Floriana*

**Mexico**

Roberto G Arenas, *Mexico*
 Sergio A Cuevas-Covarrubias, *Mexico*
 Leopoldo Flores-Romo, *Mexico*
 Maria Bertha Torres-alvarez, *San Luis Potosi*

**Netherlands**

Rosalie M Luiten, *Amsterdam*

Arnold P Oranje, *Rotterdam*
 Arnold C Spek, *Amsterdam*

**Nigeria**

Maurice E Asuquo, *Calabar*
 Joseph I Ikechebelu, *Nnewi*

**Norway**

Andrej M Grijbovski, *Oslo*

**Oman**

Mohamed Mabruk, *Muscat*

**Poland**

Andrzej Grzybowski, *Poznan*
 Lidia Rudnicka, *Warsaw*

**Portugal**

Bruno Sarmento, *Porto*

**Romania**

Liana Manolache, *Bucharest*

**Saudi Arabia**

Feroze Kaliyadan, *Hofuf*

**Singapore**

Wei-Sheng Chong, *Singapore*
 Hong Liang Tey, *Singapore*

**South Korea**

Dong-Seok Kim, *Seoul*
 Chang Hoon Lee, *Seoul*
 Jongsung Lee, *Seongnam City*
 Chil Hwan Oh, *Seoul*
 Byung Soon Park, *Seoul*
 Myung-Geun Shin, *Hwasun*
 Jong-Hyuk Sung, *Seoul*
 Young Kwan Sung, *Daegu*

**Spain**

Agustin Alomar, *Barcelona*
 Salvador Arias-Santiago, *Granada*
 Juan G Gavín, *Vigo*
 Marcos A Gonzalez-Lopez, *Santander*

Ramon Grimalt, *Barcelona*
 Husein Husein-ElAhmed, *Granada*
 Ander Izeta, *San Sebastian*
 Marcela Del Rio, *Madrid*

**Switzerland**

Gunther FL Hofbauer, *Zurich*
 Alexander A Navarini, *Zurich*

**Thailand**

Chirayu U Auewarakul, *Bangkok*
 Viroj Wiwanitkit, *Bangkok*

**Turkey**

Berna Aksoy, *Kocaeli*
 Fatma Aydin, *Samsun*
 Cem Dane, *Istanbul*
 Sibel Dogan, *Istanbul*
 Aylin T Ermertcan, *Manisa*
 Ozlem Su, *Istanbul*

**United Kingdom**

Theodoros Dimitroulas, *Dudley*
 Bernhard F Gibbs, *Chatham Maritime*
 Evmorfia Ladoyanni, *Stourbridge*
 Mark R Nelson, *London*
 Adrian V Pace, *Dudley*
 Anthony B Paul, *London*
 Sam Shuster, *Woodbridge*
 Olga Tura, *Edinburgh*
 Indre Verpetinske, *Stourbridge*

**United States**

Jeremy S Bordeaux, *Cleveland*
 Robert F Diegelmann, *Richmond*
 Q Ping Dou, *Detroit*
 Zeev Estrov, *Houston*
 Vincent Falanga, *Providence*
 Miranda A Farage, *Cincinnati*
 Markus H Frank, *Boston*
 W Scott Goebel, *Indianapolis*
 Dan-Ning Hu, *New York*
 Amor Khachemoune, *Brooklyn*
 Arash Kimyai-Asadi, *Houston*
 Michael S Kolodney, *Torrance*
 Feng Liu, *Chapel Hill*
 Senthamil R Selvan, *San Diego*
 Lei Shi, *Fort Worth*
 Animesh A Sinha, *East Lansing*
 Jeffrey M Weinberg, *New York*
 John A Zic, *Nashville*

MINIREVIEWS

- 52 Skin-gut axis: The relationship between intestinal bacteria and skin health
Vaughn AR, Notay M, Clark AK, Sivamani RK

CASE REPORT

- 59 Pleomorphic cutaneous xanthomas disclosing homozygous familial hypercholesterolemia
Mastrolorenzo A, D'Errico A, Pierotti P, Vannucchi M, Giannini S, Fossi F

ABOUT COVER

Editorial Board Member of *World Journal of Dermatology*, Jeremy S Bordeaux, MD, MPhil, Assistant Professor, Director, Department of Dermatology, University Hospitals, Cleveland, OH 44122, United States

AIM AND SCOPE

World Journal of Dermatology (*World J Dermatol*, *WJD*, online ISSN 2218-6190, DOI: 10.5314), is a peer-reviewed open access (OA) academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of dermatology. *WJD* covers fungal diseases, dermatitis and eczema, urticarial diseases, drug eruptions, pruritus, erythroderma desquamativum, connective tissue diseases, bullous skin diseases, vascular skin diseases, skin appendage diseases, pigmentary diseases, genetic diseases, nutritional and metabolic disorders, tumors, sexually transmitted diseases, AIDS, traditional medicine, integrated Chinese and Western medicine, evidence-based medicine, epidemiology and nursing. The journal also publishes original articles and reviews that report the results of applied and basic research in fields related to dermatology, such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

INDEXING/ABSTRACTING

World Journal of Dermatology is now indexed in China National Knowledge Infrastructure (CNKI).

FLYLEAF

I-II Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Fang-Fang Ji*
Proofing Editorial Office Director: *Xin-Xia Song*

NAME OF JOURNAL
World Journal of Dermatology

ISSN
 ISSN 2218-6190 (online)

LAUNCH DATE
 June 2, 2012

FREQUENCY
 Quarterly

EDITOR-IN-CHIEF
Santosh K Katiyar, PhD, Professor, Department of Dermatology, University of Alabama at Birmingham, Birmingham, AL 35294, United States

EDITORIAL BOARD MEMBERS
 All editorial board members resources online at <http://www.wjgnet.com/2218-6190/editorialboard.htm>

EDITORIAL OFFICE
 Fang-Fang Ji, Director

World Journal of Dermatology
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: editorialoffice@wjgnet.com
 Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLISHER
 Baishideng Publishing Group Inc
 7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
 Telephone: +1-925-2238242
 Fax: +1-925-2238243
 E-mail: bpgoffice@wjgnet.com
 Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

PUBLICATION DATE
 November 2, 2017

COPYRIGHT
 © 2017 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS
<http://www.wjgnet.com/bpg/gerinfo/204>

ONLINE SUBMISSION
<http://www.f6publishing.com>

Skin-gut axis: The relationship between intestinal bacteria and skin health

Alexandra R Vaughn, Manisha Notay, Ashley K Clark, Raja K Sivamani

Alexandra R Vaughn, Drexel University College of Medicine, Philadelphia, PA 19129, United States

Alexandra R Vaughn, Manisha Notay, Ashley K Clark, Raja K Sivamani, UC Davis Department of Dermatology, Sacramento, CA 95816, United States

Author contributions: All authors equally contributed to this paper with the design, literature review and analysis, drafting and critical revision and editing and final approval of the final version.

Conflict-of-interest statement: No potential conflict of interest. No financial support.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Raja K Sivamani, MD, UC Davis Department of Dermatology, 3301 C Street, Suite 1400, Sacramento, CA 95816, United States. rkshivamani@ucdavis.edu
Telephone: +1-916-7346550

Received: March 12, 2017

Peer-review started: March 15, 2017

First decision: May 12, 2017

Revised: September 7, 2017

Accepted: October 15, 2017

Article in press: October 16, 2017

Published online: November 2, 2017

Abstract

The gut microbiome is an emerging area of interest in

medicine. Imbalances in the gut microbiome have been linked to a number of disease states such as obesity and type 2 diabetes. The relationship between normally residing intestinal bacteria (the *gut microbiota*) and their potential role in the pathogenesis of skin diseases is an area of research for which we are only beginning to understand. Small studies have demonstrated underlying changes in the gut microbiome of patients with certain dermatological diseases. Interestingly, studies suggest that probiotics may have a role in the treatment of atopic dermatitis. However, the concept of the "skin-gut axis" is a newly emerging and important avenue of investigation, still lacking in pathobiological explanations. This review will introduce and describe the intestinal microbiome as it relates to skin health in a complex communication network between the immune system, endocrine system, metabolic system, and nervous system.

Key words: Gut microbiome; Skin; Bacteria; Probiotics; Dermatology

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The intestinal microbiome is a complex and dynamic bacterial community that plays an important role in human health. Alterations in microbiota composition have been related to different intestinal and extra-intestinal diseases such as psoriasis and rosacea. Studies have reported beneficial interactions between the human body and its microbiota and modulation through prebiotics and probiotics may prevent or resolve such diseases. Although the mechanisms for how the gut and skin communicate are not fully understood the association likely involves a complex connection between the nervous, immune, and endocrine systems as well as environmental factors.

Vaughn AR, Notay M, Clark AK, Sivamani RK. Skin-gut axis: The relationship between intestinal bacteria and skin health.

World J Dermatol 2017; 6(4): 52-58 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i4/52.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i4.52>

INTRODUCTION

The role of the gut microbiome as an important determinant of human health and disease has emerged as an exciting niche of research in many areas of medicine. An imbalance in the gut microbiome has been linked to obesity, type 2 diabetes, atopy and inflammatory bowel disease (IBD)^[1]. Furthermore, the relationship between normally residing intestinal bacteria (the *gut microbiota*) and their potential role in the pathogenesis of skin diseases is an area of research for which we are only now starting to gain an understanding. The small and large intestines provide residence for a vast community of bacteria and their metabolites and by-products, which we call the *gut microbiome*. Similarly, thousands of microbial organisms and their by-products inhabit the skin, referred to as the *skin microbiome*. In both the gut and the skin, a harmonial balance in these microflora is important in maintaining homeostasis^[2]. The skin and the gut have more similarities than one would suppose, and in fact, there is budding interest in learning how the skin and gut communicate and influence the health of one another^[3]. Both contain rich vascular supply, diverse microbial communities, and act as vital interfaces between the internal human body and the external environment. Additionally, the skin and gut both operate as neuro-immuno-endocrine organs, and participate in essential communication with the nervous system, immune system, and endocrine system. The "brain-gut axis" has been documented extensively in the literature, and was first described in 1930 when Stokes and Pillsbury attributed depression to altering the gut microbiome, leading to inflammatory skin diseases^[4]. However, the "skin-gut axis" is a newly emerging and important avenue of investigation, still sparse in pathobiological explanations. This review will introduce and describe the intestinal microbiome as it relates to skin health in a complex communication network between the immune system, endocrine system, metabolic system, and nervous system.

HUMAN INTESTINAL MICROBIOME

The "gut microbiome" refers to the diverse community of microbial organisms that normally inhabit the bowel and their metabolites/byproducts^[5]. There are more than 100 trillion bacteria present in the human gastrointestinal tract, consisting of over one thousand different species colonizing the intestines^[6,7]. A large proportion of the organisms found in the gut microbiome belong to two phyla: *Firmicutes* and *Bacteroidetes*^[5]. The density of the bacterial populations within the bowel differs by anatomical location. For instance, the density

is approximately 10^{2-3} colony forming units (CFU) per gram in the proximal ileum and jejunum, compared to the ascending colon which has approximately 10^{11-12} CFU per gram^[8]. There is significant variation in the gut microbiome communities among healthy individuals^[9]. The gut microbiome is relatively stable, however, studies have demonstrated that antibiotic therapy, international travel, and illness can all alter the normal gut microbiome. Aging can also lead to a shift in the predominant species within the gut microbiome. Research currently suggests that our long-term dietary patterns could have a large impact on the composition of our gut microbiome^[6].

The role of the gut microbiome is thought to include proper development and functioning of the immune system, protection against infections, digestion of polysaccharides, and synthesis of vitamins^[7]. The symbiotic relationship between resident gut bacterial flora and the host is vital to the normal immune system development and homeostasis of the host and regulation of epithelial growth and differentiation^[10].

PROBIOTICS/PREBIOTICS

Probiotic supplementation has become increasingly popular, with many commercially available products in capsule, powder, beverage, and food forms. According to the Food and Agricultural Organization of the United Nations and the World Health Organization, probiotics are considered to be "live microorganisms which when administered in adequate amounts confer a health benefit on the host"^[11]. The most frequently used bacteria are from the *Lactobacillus* and *Bifidobacterium* genera^[12]. There has been evidence to suggest that they are useful in the treatment of irritable bowel syndrome (IBS), diarrhea, and lactose intolerance^[13]. Probiotics may alleviate abnormal alterations of the gut microbiome, referred to as "dysbiosis". Dysbiosis of the gut microbiome has been linked to metabolic disorders, gastrointestinal infections, IBD, and irritable bowel syndrome (IBS)^[14]. Probiotics are thought to provide therapeutic benefits *via* multiple mechanisms. Firstly they are believed to prevent pathogenic bacteria from colonizing the gastrointestinal tract, which would otherwise subsequently lead to disease. Secondly, they are thought to improve the barrier function of the colonic mucosa. Thirdly, probiotics may help modulate the immune system, which may help shift away from pro-inflammatory immune reactivity^[12]. Fourth, they may synthesize and secrete metabolites that may have nutritional benefits and anti-inflammatory effects^[15]. Lastly, probiotics may even play a role in modulating central nervous system and enteric nervous system functions. In fact, in a randomized controlled trial patients with Alzheimer's disease who received probiotic supplementation for 12 wk had significant improvement in mental status score and had a significant decrease in serum c-reactive protein (Akbari, 2016 #991). Additionally, probiotic supplementation has demonstrated

improvement in multiple sclerosis symptoms and exacerbations (Dolan, 2016 #992).

Probiotics have not yet been widely studied in the treatment of dermatological diseases. Two meta-analyses failed to demonstrate any clinically significant changes in the severity of atopic dermatitis (AD) in children treated with probiotic supplementation^[16,17]. However, Lee *et al.*^[16] found a significant risk reduction (up to 61%) of pediatric AD in those who were treated with prenatal and/or postnatal probiotics. There are even fewer studies available regarding the treatment of adults with AD using probiotics. These small studies have demonstrated that there may be a clinical benefit in adults^[18-20]; however, larger trials are needed before any conclusions can be drawn. Probiotics are postulated to help in atopic dermatitis by improving the diversity of the intestinal flora, increase the barrier function of the skin and mucosa and by producing a mainly Th1 response^[13].

Prebiotics are non-digestible carbohydrates that help stimulate the growth of certain bacteria in the gut, which can lead to an improvement in the health of the host^[21]. A review by Osborn *et al.*^[22] of four clinical trials found that there was a statistically significant reduction in the incidence of infant eczema with prebiotic supplementation of galactooligosaccharides and fructooligosaccharides (RR 0.68). It has been demonstrated that milk glycoproteins are able to select for and stimulate the growth of *Bifidobacteria longum infantis* (*B. infantis*) in the gut microbiome^[23]. This is of clinical importance as *B. infantis* supplementation can reduce the risk of necrotizing enterocolitis in preterm infants. *B. infantis* colonization of the gastrointestinal tract is associated with improved immune response to vaccination and weight gain^[24].

However, further studies need to be conducted into the use of prebiotics and probiotics before recommendations regarding their use in the treatment or prevention of dermatological diseases can be made.

LINK BETWEEN SKIN DISEASE AND THE GUT

Gastrointestinal disorders can present with dermatological skin findings. IBD is linked to skin manifestations such as pyoderma gangrenosum, erythema nodosum, Sweet's Syndrome and oral lesions^[23]. Celiac disease is associated skin manifestations such as dermatitis herpetiformis, alopecia, vitiligo and oral mucosal lesions. Furthermore, psoriasis is more commonly found in patients with Crohn's disease than healthy people^[24].

There is emerging evidence linking certain dermatological disorders to gut dysbiosis. However, this is not a novel topic and in fact, in 1911 a gastroenterologist named Milton H. Mack wrote, "Acne and eczema are both traceable to this fountainhead of diseases... if in a case of urticarial we look to the intestinal track, why not in eczema and acne?"^[25]. Simultaneous gut and skin microbiome

dysbiosis has been observed in several inflammatory skin diseases, such as rosacea, psoriasis, and atopic dermatitis^[26].

Psoriasis

Interestingly, patients with psoriatic arthritis are at increased risk of developing IBD and have subclinical evidence of gut inflammation^[27]. A recent clinical study including 16 patients with psoriatic arthritis, 15 with psoriasis and 17 healthy controls analysed the gut microbiome across these three groups. The gut microbiome was less diverse in the psoriasis and psoriatic arthritis groups; with a decrease in the *Coprococcus* spp. Those with psoriatic arthritis experienced a reduction in important bacterial enterotypes such as *Akkermansia*, *Ruminococcus*, and *Pseudobutyrvibrio*. It is thought that these taxonomic changes cause to a reduction in the ability of the gut to regulate immune responses, which may lead to systemic or localized inflammation^[28].

In addition, a clinical trial has shown that treating psoriasis patients with probiotic *Bifidobacterium infantis* 35624 for eight weeks improved C-reactive protein (CRP), TNF-alpha and IL-6 levels. However, during this study no clinical assessments were performed after baseline. These results suggest that probiotic supplementation could modulate inflammation in this disorder^[29].

Rosacea

Rosacea has been linked to *Helicobacter pylori* (*H. pylori*) infection, however the efficacy of *H. pylori* eradication in rosacea therapy is unclear^[30]. Moreover, a study of 113 rosacea patients demonstrated that those with rosacea have a higher incidence of small intestinal bacterial overgrowth (SIBO) when compared to controls. Those with SIBO were treated with either rifaximin therapy for 10 d or placebo. Those who were treated with antibiotic therapy experienced an improvement in their symptoms for at least nine months^[31].

Atopic dermatitis

There is a well-documented association between gut microbiome dysbioses and low diversity within the gut microbiota with the development of allergic diseases (Melli, 2016 #993). Conversely, increased microbial diversity within the gut has been associated with reduced flares in inflammatory skin diseases, such as atopic dermatitis (Marrs, 2016 #994).

PROPOSED MECHANISMS REGARDING THE SKIN-GUT AXIS

At present, there is clinical evidence suggesting a close relationship between intestinal dysbiosis and dermatologic conditions. However, the mechanistic basis behind these observations has yet to be confirmed. The association between the gut and skin likely involves a complex and multifactorial interplay between the

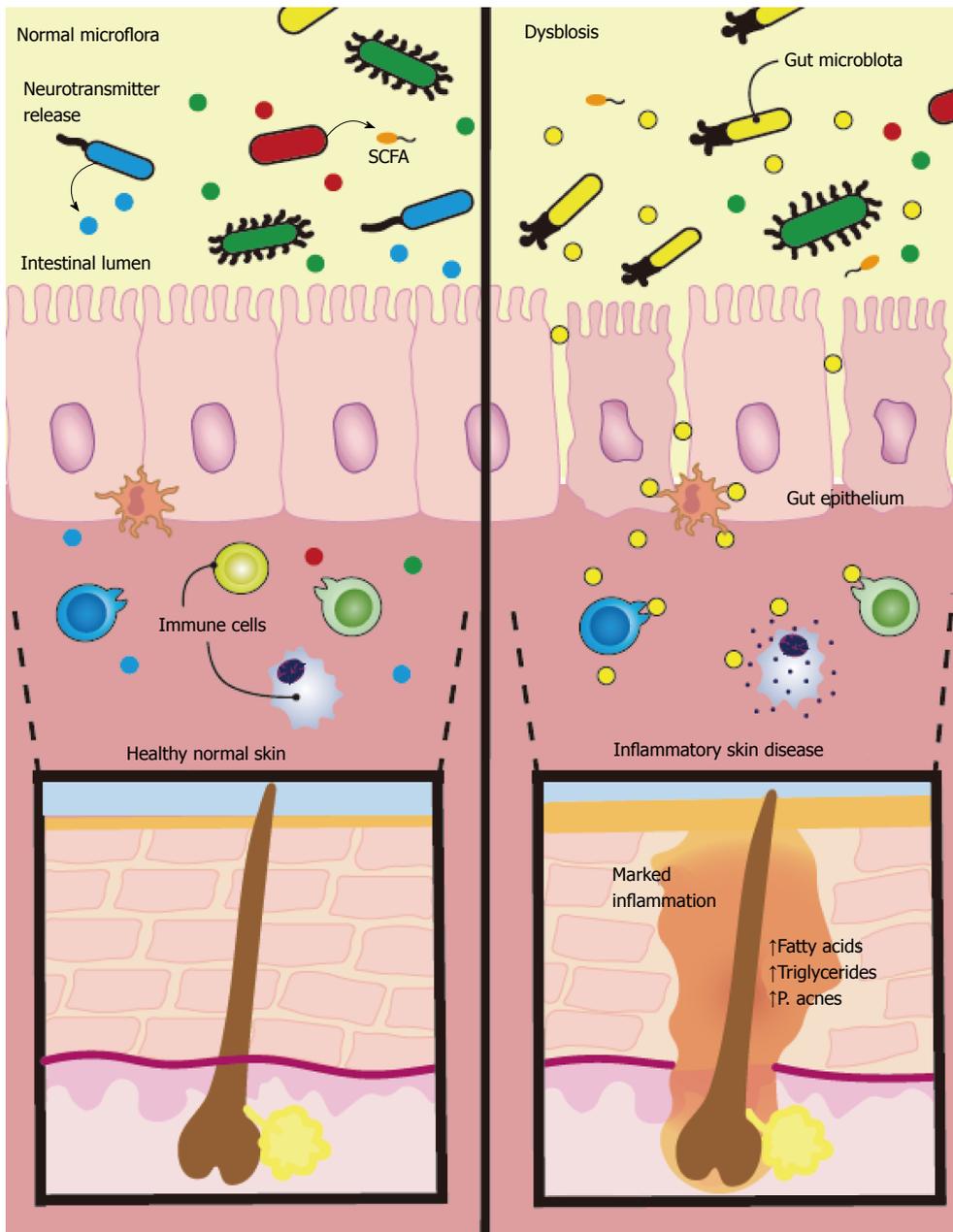


Figure 1 There is emerging evidence linking dermatological disorders to alterations in gut bacteria. Studies hypothesize intestinal flora produce neurotransmitters in response to stress that can modulate skin function. These neurotransmitters cross the intestinal epithelium enter the bloodstream and induce systemic effects. Along with neurotransmitters, the gut microflora also release short chain fatty acids (SCFAs), which can also enter systemic circulation and affect the skin. Additionally, diet may influence inflammation in the skin through nutrient signalling and release of long chain fatty acids, leading to excessive stimulation of sterol regulatory element-binding protein 1 and increased synthesis of fatty acids and triglycerides promoting *Propionibacterium acnes* overgrowth.

nervous, immune, and endocrine systems as well as environmental factors such as diet and medications (Figure 1).

Skin-gut axis and the neuroendocrine system

The “brain-gut-skin axis” has been eloquently documented by Arck *et al.*^[32] and Bowe and Logan^[4]. It is known that psychosocial stress is implicated in both exacerbation and the initiation of various skin conditions^[33]. It is plausible that the intestinal microflora produce neurotransmitters in response to stress and other external stimuli that could modulate skin function *via* neural pathways. For

instance, commensal organisms in the gut can produce norepinephrine, serotonin, and acetylcholine or may evoke the release of neuropeptides from nearby enteroendocrine cells^[34]. These neurotransmitters might cross the intestinal epithelium into the bloodstream and induce systemic effects^[35]. Along with neurotransmitters, the gut microflora also release short chain fatty acids (SCFAs), including propionic acid, butyric acid, acetic acid, and lactic acid derived from polysaccharide fermentation from food we eat^[36]. The majority of these SCFAs are produced in the large intestine, where the colon is highly efficient in the reabsorption of fatty acids, only allowing approximately

10% to remain in expelled feces^[37]. The true systemic levels of SCFA derived from the colon depend on individual dietary habits, rate of SCFA production by gut microbes, and the degree of absorption through the large intestine. It is not known whether these metabolites, along with many others produced by gut microbes, are able to reach clinically significant levels in the bloodstream in order to impact the skin^[38].

Immune system modulation

Health, including skin health and overall well being, require tightly integrated immune and hormone feedback systems that allow beneficial microbial to dominate in the gut and on the skin^[39]. The normal gut microbial residents continuously interact with the immune system to support host homeostasis. In general, immune system homeostasis requires a proper balance of pro-inflammatory and anti-inflammatory signals and molecules in response to internal and external environmental changes. If the microbiome composition changes for any given reason, the immune system reactivity could subsequently shift and eventually lead to inflammatory skin diseases^[40]. This idea was exemplified in a mouse study by Zanvit *et al* which demonstrated that mice treated with antibiotics neonatally had exacerbated imiquimod-induced psoriasis as an adult, while mice treated with the same antibiotics in adulthood had improved psoriasis (Zanvit, 2015 #990). This study demonstrates the importance of how neonatal gut dysbioses can affect skin inflammation, potentially triggering or exacerbating inflammatory skin diseases such as psoriasis later in adulthood. Interleukin-10 (IL-10) is generally considered to decrease pro-inflammatory molecules, such as IL-17^[41]. Animal models have shown that probiotic supplementation up regulates IL-10 and provides beneficial skin effects^[42]. In a recent article, Zákostelská *et al*^[43] hypothesize that certain beneficial families of intestinal bacteria, such as lactobacilli, are able to suppress the IL-23/Th17 axis, which is believed to play an important role in inflammation involved in psoriasis^[43]. This suppression may occur through certain gut commensal organisms' ability to down regulate IL-23 and transforming growth factor-beta (TGF- β) expression, and preventing Th17 cell-mediated release of proinflammatory IL-17^[44]. As a result of immune system dysfunction and deficiency in T regulatory cells, some autoimmune diseases can result in rampant inflammation and severe dermatitis, such as in IPEX syndrome (Halabi-Tawil, 2009 #996). The intestinal microbiome is responsible for regulating the expansion of T regulatory cells, Th1 and Th2 type cells to provide immune system homeostasis, and there has been recent research investigating how treating the gut microbiome could improve these types of skin conditions (He, 2017 #995). These are examples demonstrating the complex interplay between the immune system and gut commensal organisms. The true connection between skin health and gut bacteria induced immune system reactivity is poorly understood and still requires more extensive investigation.

Diet

Recent research continues to reveal the influence of the "western diet" in the obesity epidemic, and researchers have hypothesized that alterations in the gut microbiome due to high dietary fat intake could be partly to blame (Murphy, 2015 #997). In the literature, it is generally accepted that high fat diets lead to gut dysbioses, reflected by a decrease in *Bacteroidetes* species and an increase in *Firmicutes* species (Zhang, 2012 #998). Although the exact mechanisms are still under investigation, "western diet" induced gut dysbioses may be associated with cancer (Schulz, 2014 #999), atherosclerosis and heart disease (Gregory, 2015 #1000), insulin resistance (Carvalho, 2012 #1001), and even disorders of the central nervous system (Scheperjans, 2015 #1002). Until recently, conflicting opinions and inconclusive evidence have predominated regarding the link between diet and skin conditions. Although more mechanistic studies are warranted, there is growing evidence that diet plays an important role in the pathogenesis of skin diseases, with acne vulgaris being an example. For example, the western diet consisting of large amounts of saturated fats and high glycemic load has been strongly associated with acne^[45,46]. Researchers hypothesize this occurs from problems in nutrient signalling, ultimately leading to excessive stimulation of sterol regulatory element-binding protein 1 (SREBP-1) and increased synthesis of fatty acids (ex - free oleic acid) and triglycerides in sebum that promotes flourishing *Propionibacterium acnes* growth^[47]. The strong association between atopic dermatitis and food sensitivities similarly exemplifies the importance of food on the gut-skin relationship^[48]. The ability of diet to both positively and negatively influence skin function demonstrates the undeniable link between the skin and gut, however, the mechanisms surrounding this connection is likely multifactorial and at present based primarily on theory. Indeed, it is difficult to detangle the direct effects of food on the skin versus food's modulation of the intestinal microflora.

CONCLUSION

The intimate relationship between the gut and skin is undeniable. Possibly, both the intestinal bacteria themselves and their metabolic by-products influence skin physiology. The mechanisms are still under study but there are a few theories: (1) bacterial products and diet could alter the physiology of the gut epithelium, resulting in different secretory products that might circulate systemically and reach the skin; (2) neurotransmitters, hormones, and other bioactive chemicals such as SCFAs derived from the gut could all act on receptors within the skin and directly alter the skin or alter the skin's commensal bacteria; and (3) ingested compounds and chemicals may absorb and have a direct effect on the skin's appearance or function^[49].

Although not a new avenue of research, the relationship between the gut microbiome and skin health is emerging as an important and intriguing topic in dermatology and gastroenterology alike. It is especially important to understand how diet, medications, and psychosocial stress can influence or contribute to altered microbial communities in the gut, which may directly or indirectly affect skin health.

REFERENCES

- Bull MJ**, Plummer NT. Part 1: The Human Gut Microbiome in Health and Disease. *Integr Med (Encinitas)* 2014; **13**: 17-22 [PMID: 26770121]
- Barrett KE**, Ghishan FK, Mercant JL. Physiology of the Gastrointestinal Tract. 4th ed. New York: Elsevier
- Thrash B**, Patel M, Shah KR, Boland CR, Menter A. Cutaneous manifestations of gastrointestinal disease: part II. *J Am Acad Dermatol* 2013; **68**: 211.e1-e33; quiz 244-246 [PMID: 23317981 DOI: 10.1016/j.jaad.2012.10.036]
- Bowe WP**, Logan AC. Acne vulgaris, probiotics and the gut-brain-skin axis - back to the future? *Gut Pathog* 2011; **3**: 1 [PMID: 21281494 DOI: 10.1186/1757-4749-3-1]
- Ahmad OF**, Akbar A. Microbiome, antibiotics and irritable bowel syndrome. *Br Med Bull* 2016; **120**: 91-99 [PMID: 27737852 DOI: 10.1093/bmb/ldw038]
- Moschen AR**, Wieser V, Tilg H. Dietary Factors: Major Regulators of the Gut's Microbiota. *Gut Liver* 2012; **6**: 411-416 [PMID: 23170142 DOI: 10.5009/gnl.2012.6.4.411]
- D'Argenio V**, Salvatore F. The role of the gut microbiome in the healthy adult status. *Clin Chim Acta* 2015; **451**: 97-102 [PMID: 25584460 DOI: 10.1016/j.cca.2015.01.003]
- Jones RM**. The Influence of the Gut Microbiota on Host Physiology: In Pursuit of Mechanisms. *Yale J Biol Med* 2016; **89**: 285-297 [PMID: 27698613]
- Shreiner AB**, Kao JY, Young VB. The gut microbiome in health and in disease. *Curr Opin Gastroenterol* 2015; **31**: 69-75 [PMID: 25394236 DOI: 10.1097/MOG.0000000000000139]
- Neish AS**, Jones RM. Redox signaling mediates symbiosis between the gut microbiota and the intestine. *Gut Microbes* 2014; **5**: 250-253 [PMID: 24637602 DOI: 10.4161/gmic.27917]
- Hill C**, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, Morelli L, Canani RB, Flint HJ, Salminen S, Calder PC, Sanders ME. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol* 2014; **11**: 506-514 [PMID: 24912386 DOI: 10.1038/nrgastro.2014.66]
- Butel MJ**. Probiotics, gut microbiota and health. *Med Mal Infect* 2014; **44**: 1-8 [PMID: 24290962 DOI: 10.1016/j.medmal.2013.10.002]
- Baquerizo Nole KL**, Yim E, Keri JE. Probiotics and prebiotics in dermatology. *J Am Acad Dermatol* 2014; **71**: 814-821 [PMID: 24906613 DOI: 10.1016/j.jaad.2014.04.050]
- Hemarajata P**, Versalovic J. Effects of probiotics on gut microbiota: mechanisms of intestinal immunomodulation and neuromodulation. *Therap Adv Gastroenterol* 2013; **6**: 39-51 [PMID: 23320049 DOI: 10.1177/1756283X12459294]
- Stanton C**, Ross RP, Fitzgerald GF, Van Sinderen D. Fermented functional foods based on probiotics and their biogenic metabolites. *Curr Opin Biotechnol* 2005; **16**: 198-203 [PMID: 15831387 DOI: 10.1016/j.copbio.2005.02.008]
- Lee J**, Seto D, Bielory L. Meta-analysis of clinical trials of probiotics for prevention and treatment of pediatric atopic dermatitis. *J Allergy Clin Immunol* 2008; **121**: 116-121.e11 [PMID: 18206506 DOI: 10.1016/j.jaci.2007.10.043]
- Michail SK**, Stolfa A, Johnson T, Onady GM. Efficacy of probiotics in the treatment of pediatric atopic dermatitis: a meta-analysis of randomized controlled trials. *Ann Allergy Asthma Immunol* 2008; **101**: 508-516 [PMID: 19055205 DOI: 10.1016/S1081-1206(10)60290-6]
- Drago L**, De Vecchi E, Toscano M, Vassena C, Altomare G, Pigatto P. Treatment of atopic dermatitis eczema with a high concentration of Lactobacillus salivarius LS01 associated with an innovative gelling complex: a pilot study on adults. *J Clin Gastroenterol* 2014; **48** Suppl 1: S47-S51 [PMID: 25291127 DOI: 10.1097/MCG.0000000000000249]
- Inoue Y**, Kambara T, Murata N, Komori-Yamaguchi J, Matsukura S, Takahashi Y, Ikezawa Z, Aihara M. Effects of oral administration of Lactobacillus acidophilus L-92 on the symptoms and serum cytokines of atopic dermatitis in Japanese adults: a double-blind, randomized, clinical trial. *Int Arch Allergy Immunol* 2014; **165**: 247-254 [PMID: 25660281 DOI: 10.1159/000369806]
- Moroi M**, Uchi S, Nakamura K, Sato S, Shimizu N, Fujii M, Kumagai T, Saito M, Uchiyama K, Watanabe T, Yamaguchi H, Yamamoto T, Takeuchi S, Furue M. Beneficial effect of a diet containing heat-killed Lactobacillus paracasei K71 on adult type atopic dermatitis. *J Dermatol* 2011; **38**: 131-139 [PMID: 21269308 DOI: 10.1111/j.1346-8138.2010.00939.x]
- Holscher HD**. Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes* 2017; **8**: 172-184 [PMID: 28165863 DOI: 10.1080/19490976.2017.1290756]
- Osborn DA**, Sinn JK. Prebiotics in infants for prevention of allergy. *Cochrane Database Syst Rev* 2013; **(3)**: CD006474 [PMID: 23543544 DOI: 10.1002/14651858.CD006474.pub3]
- Tavarela Veloso F**. Review article: skin complications associated with inflammatory bowel disease. *Aliment Pharmacol Ther* 2004; **20** Suppl 4: 50-53 [PMID: 15352894 DOI: 10.1111/j.1365-2036.2004.02055.x]
- Saarialho-Kere U**. The gut-skin axis. *J Pediatr Gastroenterol Nutr* 2004; **39** Suppl 3: S734-S735 [PMID: 15167366]
- Mack M**. Intestinal toxemia. *Illinois Med J* 1911; **(20)**: 311-316
- Gallo RL**, Nakatsuji T. Microbial symbiosis with the innate immune defense system of the skin. *J Invest Dermatol* 2011; **131**: 1974-1980 [PMID: 21697881 DOI: 10.1038/jid.2011.182]
- Scher JU**, Littman DR, Abramson SB. Microbiome in Inflammatory Arthritis and Human Rheumatic Diseases. *Arthritis Rheumatol* 2016; **68**: 35-45 [PMID: 26331579 DOI: 10.1002/art.39259]
- Scher JU**, Ubeda C, Artacho A, Attur M, Isaac S, Reddy SM, Marmon S, Neimann A, Brusca S, Patel T, Manasson J, Pamer EG, Littman DR, Abramson SB. Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis Rheumatol* 2015; **67**: 128-139 [PMID: 25319745 DOI: 10.1002/art.38892]
- Groeger D**, O'Mahony L, Murphy EF, Bourke JF, Dinan TG, Kiely B, Shanahan F, Quigley EM. Bifidobacterium infantis 35624 modulates host inflammatory processes beyond the gut. *Gut Microbes* 2013; **4**: 325-339 [PMID: 23842110 DOI: 10.4161/gmic.25487]
- Holmes AD**. Potential role of microorganisms in the pathogenesis of rosacea. *J Am Acad Dermatol* 2013; **69**: 1025-1032 [PMID: 24011460 DOI: 10.1016/j.jaad.2013.08.006]
- Parodi A**, Paolino S, Greco A, Drago F, Mansi C, Rebora A, Parodi A, Savarino V. Small intestinal bacterial overgrowth in rosacea: clinical effectiveness of its eradication. *Clin Gastroenterol Hepatol* 2008; **6**: 759-764 [PMID: 18456568 DOI: 10.1016/j.cgh.2008.02.054]
- Arck P**, Handjiski B, Hagen E, Pincus M, Bruenahl C, Bienenstock J, Paus R. Is there a 'gut-brain-skin axis'? *Exp Dermatol* 2010; **19**: 401-405 [PMID: 20113345 DOI: 10.1111/j.1600-0625.2009.01060.x]
- Koo J**, Lebwohl A. Psycho dermatology: the mind and skin connection. *Am Fam Physician* 2001; **64**: 1873-1878 [PMID: 11764865]
- Lyte M**. Microbial endocrinology and the microbiota-gut-brain axis. *Adv Exp Med Biol* 2014; **817**: 3-24 [PMID: 24997027 DOI: 10.1007/978-1-4939-0897-4_1]
- Rea K**, Dinan TG, Cryan JF. The microbiome: A key regulator of stress and neuroinflammation. *Neurobiol Stress* 2016; **4**: 23-33 [PMID: 27981187 DOI: 10.1016/j.ynstr.2016.03.001]
- Cummings JH**, Macfarlane GT. Role of intestinal bacteria in nutrient metabolism. *JPEN J Parenter Enteral Nutr* 1997; **21**: 357-365 [PMID: 9406136]
- Wong JM**, de Souza R, Kendall CW, Emam A, Jenkins DJ. Colonic health: fermentation and short chain fatty acids. *J Clin Gastroenterol* 2006; **40**: 235-243 [PMID: 16633129]

- 38 **O'Neill CA**, Monteleone G, McLaughlin JT, Paus R. The gut-skin axis in health and disease: A paradigm with therapeutic implications. *Bioessays* 2016; **38**: 1167-1176 [PMID: 27554239 DOI: 10.1002/bies.201600008]
- 39 **Lee YK**, Mazmanian SK. Has the microbiota played a critical role in the evolution of the adaptive immune system? *Science* 2010; **330**: 1768-1773 [PMID: 21205662 DOI: 10.1126/science.1195568]
- 40 **Tlaskalová-Hogenová H**, Štěpánková R, Kozáková H, Hudcovic T, Vannucci L, Tučková L, Rossmann P, Hrnčič T, Kverka M, Zákostelská Z, Klimešová K, Přibyllová J, Bártová J, Sanchez D, Fundová P, Borovská D, Srůtková D, Zidek Z, Schwarzer M, Drastich P, Funda DP. The role of gut microbiota (commensal bacteria) and the mucosal barrier in the pathogenesis of inflammatory and autoimmune diseases and cancer: contribution of germ-free and gnotobiotic animal models of human diseases. *Cell Mol Immunol* 2011; **8**: 110-120 [PMID: 21278760 DOI: 10.1038/cmi.2010.67]
- 41 **Maynard CL**, Elson CO, Hattton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and immune system. *Nature* 2012; **489**: 231-241 [PMID: 22972296 DOI: 10.1038/nature11551]
- 42 **Levkovich T**, Poutahidis T, Smillie C, Varian BJ, Ibrahim YM, Lakritz JR, Alm EJ, Erdman SE. Probiotic bacteria induce a 'glow of health'. *PLoS One* 2013; **8**: e53867 [PMID: 23342023 DOI: 10.1371/journal.pone.0053867]
- 43 **Zákostelská Z**, Málková J, Klimešová K, Rossmann P, Hornová M, Novosádová I, Stehliková Z, Kostovčik M, Hudcovic T, Štěpánková R, Jůzlová K, Hercogová J, Tlaskalová-Hogenová H, Kverka M. Intestinal Microbiota Promotes Psoriasis-Like Skin Inflammation by Enhancing Th17 Response. *PLoS One* 2016; **11**: e0159539 [PMID: 27434104 DOI: 10.1371/journal.pone.0159539]
- 44 **Chen L**, Zou Y, Peng J, Lu F, Yin Y, Li F, Yang J. Lactobacillus acidophilus suppresses colitis-associated activation of the IL-23/Th17 axis. *J Immunol Res* 2015; **2015**: 909514 [PMID: 25973440 DOI: 10.1155/2015/909514]
- 45 **Cordain L**, Lindeberg S, Hurtado M, Hill K, Eaton SB, Brand-Miller J. Acne vulgaris: a disease of Western civilization. *Arch Dermatol* 2002; **138**: 1584-1590 [PMID: 12472346]
- 46 **Grossi E**, Cazzaniga S, Crotti S, Naldi L, Di Landro A, Ingordo V, Cusano F, Atzori L, Tripodi Cutri F, Musumeci ML, Pezzarossa E, Bettoli V, Caproni M, Bonci A; GISED Acne Study Group. The constellation of dietary factors in adolescent acne: a semantic connectivity map approach. *J Eur Acad Dermatol Venereol* 2016; **30**: 96-100 [PMID: 25438834 DOI: 10.1111/jdv.12878]
- 47 **Zouboulis CC**, Jourdan E, Picardo M. Acne is an inflammatory disease and alterations of sebum composition initiate acne lesions. *J Eur Acad Dermatol Venereol* 2014; **28**: 527-532 [PMID: 24134468 DOI: 10.1111/jdv.12298]
- 48 **Manam S**, Tsakok T, Till S, Flohr C. The association between atopic dermatitis and food allergy in adults. *Curr Opin Allergy Clin Immunol* 2014; **14**: 423-429 [PMID: 25153338 DOI: 10.1097/aci.0000000000000095]
- 49 **Whitehead RD**, Re D, Xiao D, Ozakinci G, Perrett DI. You are what you eat: within-subject increases in fruit and vegetable consumption confer beneficial skin-color changes. *PLoS One* 2012; **7**: e32988 [PMID: 22412966 DOI: 10.1371/journal.pone.0032988]

P- Reviewer: Antonakopoulos N, Rhoads JM **S- Editor:** Qi Y
L- Editor: A **E- Editor:** Lu YJ



Pleomorphic cutaneous xanthomas disclosing homozygous familial hypercholesterolemia

Antonio Mastrolorenzo, Antonietta D'Errico, Piera Pierotti, Margherita Vannucchi, Stefano Giannini, Fiammetta Fossi

Antonio Mastrolorenzo, Antonietta D'Errico, Department of Surgery and Translational Medicine, Section of Dermatology, University of Florence, Public Hospital Piero Palagi, Florence 50125, Italy

Piera Pierotti, Azienda Sanitaria di Firenze, Department of Infectious Diseases, Ospedale SM Annunziata, Florence 50012, Italy

Margherita Vannucchi, Histopathology and Molecular Diagnostics Institute, Careggi University Hospital, Florence 50134, Italy

Stefano Giannini, Diabetes and Metabolic Disease Agency, Careggi University Hospital, Florence 50134, Italy

Fiammetta Fossi, SODc of Transfusion Medicine and Cell Therapy, Careggi University Hospital, Florence 50134, Italy

Author contributions: Mastrolorenzo A and D'Errico A designed the report; Pierotti P performed the infectiology analyses; Mastrolorenzo A and D'Errico A collected the patient's clinical data; Giannini S and Fossi F performed the vascular and metabolic analyses, dyslipidemia management and critical revision; Vannucchi M performed the histopathological analyses; Mastrolorenzo A and D'Errico A analyzed the data and wrote the paper.

Institutional review board statement: The Case Report was reviewed and approved by the (Department of Surgery and Translational Medicine, Section of Dermatology, University of Florence) Institutional Review Board as required.

Informed consent statement: The patient involved in this case report has signed an informed consent allowing the use of pictures and information in an anonymous format.

Conflict-of-interest statement: The authors have no conflicts of interest and have not received any funding or financial consideration with respect to this article.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this

work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Correspondence to: Antonio Mastrolorenzo, MD, Department of Surgery and Translational Medicine, Section of Dermatology, University of Florence, Public Hospital Piero Palagi, Viale Michelangiolo 41, Florence 50125, Italy. amas@dada.it
Telephone: +39-055-6939655
Fax: +39-055-6939598

Received: July 12, 2017

Peer-review started: July 20, 2017

First decision: August 7, 2017

Revised: August 26, 2017

Accepted: October 15, 2017

Article in press: October 16, 2017

Published online: November 2, 2017

Abstract

Homozygous Familial Hypercholesterolemia is characterized by a presence of several types of cutaneous xanthomas with an abnormal lipid profile. Some of these could be pathognomonic. Although these could be initially interpreted as isolated and localized benign disorders and offered surgical treatment, it has become increasingly clear that they could be a part of a systemic pathology. Here we describe a case of this rare disorder in a 19 years old non-obese young man who presented multiple, intertriginous, tuberous and tendinous xanthomas and had an associated abnormal lipid profile with elevated low-density lipoprotein cholesterol levels. A detailed history with clinical assessment in the differential diagnosis and laboratory investigations led to a precise diagnosis.

Key words: Intertriginous xanthomas Homozygous Familial

Hypercholesterolemia; Familial hypercholesterolemia; Dyslipidemia; Xanthomas

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: This article describes a contemporary approach to the differential diagnosis of xanthomas, and the morphological classification from a review of the literature, specifically reflect the clinical findings evidenced in this case report.

Mastrolorenzo A, D'Errico A, Pierotti P, Vannucchi M, Giannini S, Fossi F. Pleomorphic cutaneous xanthomas disclosing homozygous familial hypercholesterolemia. *World J Dermatol* 2017; 6(4): 59-65 Available from: URL: <http://www.wjgnet.com/2218-6190/full/v6/i4/59.htm> DOI: <http://dx.doi.org/10.5314/wjd.v6.i4.59>

INTRODUCTION

The clinical picture of xanthomas is variable from yellow or orange dermal macules or papules, to soft or firm-hard subcutaneous plaques and tendinous nodules not attached to underlying structures, with normal-appearing overlying skin. In recent years, interest in xanthomas has been growing for several reasons, mainly because the pathogenetic mechanisms involved in the development seems to be similar to those in early stages of atherosclerotic plaques^[1]. Cholesterol accumulation in tissues produces common dermatological manifestations as several types of cutaneous xanthomas^[2]. The association of xanthomas with lipoprotein disorders was initially defined by Frederickson's classification^[3,4]. From that phenotypic classification the recent advances in molecular genetics led to the discovery of a broad group of disorders of the lipids metabolism disclosing the relationship between the development of xanthomas and hyperlipidemias^[1,5,6].

Xanthomas can be classified following clinical as well as patho-anatomical schemes, addressing special attention to the needs of dermatologists and internal medicine specialists respectively. These correlated issues gave rise to the following groups which are useful in clinical practice: Normolipidaemic xanthomas (NX), hyperlipidaemic xanthomas (HX), and necrobiotic xanthogranuloma (NXG)^[1,7,8]. Nevertheless, xanthomas may be seen either as a primary disorder (primary dyslipidemia, an inherited abnormality of lipoprotein metabolism) or secondary disorder (hyperlipidemia secondary to systemic disease or medication)^[1,4,9].

Cutaneous xanthomas may or may not be present with lipid metabolic disorders, usually depending on the severity of the lipid abnormality. Normolipidemic xanthomas mostly appear as diffuse flat skin lesions, while hyperlipidaemic types are polymorphous, often tuberous, and can affect either skin or tendons and joints. Recognition of these types of xanthomas may be

facilitated on the basis of clinical morphology, presence or absence of inflammation, anatomic distribution, and development pattern, defining the primary type of lesion and histologic level of involvement. From a dermatological point of view these can be categorized in two specific subsets and each one with distinctive clinical associated features: (1) papulonodular xanthomas: Eruptive and tubero-eruptive xanthoma, xanthoma tuberosum (the term "tuberous" refers to the nodular character of these xanthomas) and tendineum; and (2) plane xanthomas: Plane and intertriginous xanthoma, striated palmar xanthoma and xanthelasma palpebrarum^[1,4,9-11].

We report a case of a young man with multiple pleomorphic cutaneous xanthomas in association with a neglected Homozygous Familial Hypercholesterolemia (HoFH). This article thus presents a contemporary approach to the differential diagnosis of xanthomas, and the diagnostic criteria we propose was developed after a review of the literature, and reflect the clinical findings evidenced in our patient, seen at our dermatological facility.

CASE REPORT

A 19 years old non-obese young man presented as an outpatient to our hospital with multiple, bilateral and symmetrical slow growing yellowish lesions of various forms over the dorsum of the elbows, knees, buttocks, ears, feet and hands. Biopsy of three representative and different skin lesions revealed them to be xanthomas characterized by the presence in the dermis of cholesterol crystalline aggregates surrounded by fibrosis and foamy cells (Figure 1).

On dermatological examination each lesion was defined on morphological pattern. The following clinical forms have been recognized: (1) Xanthelasma: Involving the inner canthus of the left eye (Figure 2); (2) Intertriginous xanthomas and a confluence of plane-eruptive xanthomas (Figure 3): In finger web spaces (Figure 3A), toe web spaces (Figure 3C and D), and the flexural surfaces: the ankle crease (Figure 3D), the antecubital fossae (Figure 3E and F), the popliteal fossae (Figure 3G and H) and the creases of ears (Figure 3I and J); (3) Tendinous xanthomas (Figure 4): To form a single mass localized all over the Achilles tendon just above its insertion point to the calcaneal tuberosity; and (4) Tuberous xanthomas (Figure 5): Soft skin color or yellowish nodules and tumors, with a tendency to coalesce, localized on the knees (Figure 5A and B), malleolus (Figure 5C) and buttocks (Figure 5D).

The lesions appeared at about 2 years of age on both lateral malleolus, at 3 years of age over the buttocks; they were originally asymptomatic then progressively increased in size and extent. At present time the size of the lesions varied between 1 cm × 1 cm × 1 cm to 10 cm × 5 cm (over the Achilles tendon, Figure 4) and 10 cm × 10 cm (over the buttocks, Figure 5D). On detailed clinical history the patient had symptoms of discomfort and pain in the elbows for bilateral massive

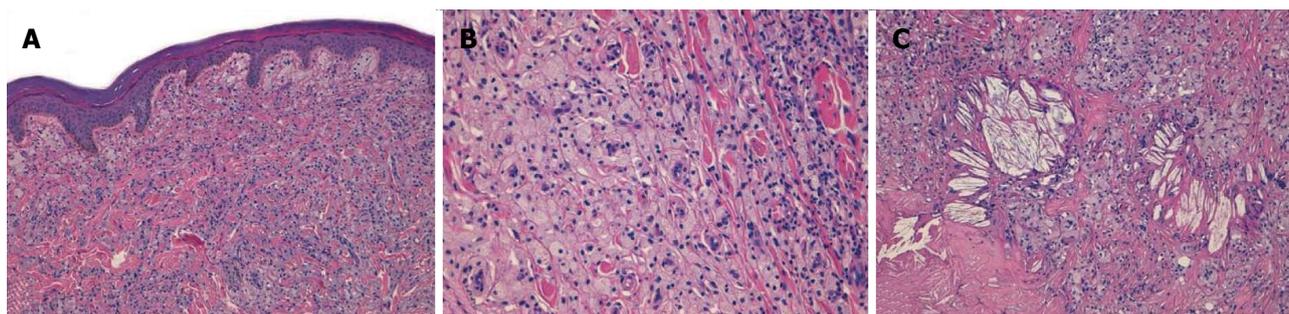


Figure 1 Histopathological examination. A: Foam cells infiltrate the superficial and deep dermis in cluster, separated by collagen fibers. Absence of any other significant inflammatory infiltrate (10 ×, EE); B: Xanthoma cells are filled with optically empty vacuoles, showing thin, well defined cytoplasmic membranes, and tend to be attached to each other. They can be multinucleate (20 ×, EE); C: Presence in the dermis of cholesterol crystalline aggregates surrounded by fibrosis and foamy cells (10 ×, EE).



Figure 2 Xanthelasma. Single yellow-orange papular lesion on the inner canthus of the eye.

tuberous xanthomas and at the age of 8, for significantly restricted joint mobility at these sites he had surgery in China in a rural hospital. Up to day the removed lesions did not recur (Figure 6). However, the discomfort and pain due to the large size of the masses of the buttocks and the limitation of his walking distance for the Achilles tendinous xanthomas progressively worsened resulting in significant disability.

Clinical examination did not reveal xanthomatous infiltration of cornea, oral, pharyngeal, and laryngeal mucosae. The patient's family history was remarkable in that both nonconsanguineous parents had a chronic hepatitis B and high total cholesterol (TC) and low-density lipoprotein cholesterol (LDL-C) levels. The 17 years old sister had a mild hypercholesterolemia, but no other family members have shown any other inherited disorders and such similar xanthomas. The patient's plasma TC level in the last six months ranged between 657 mg/dL and 990 mg/dL (reference value < 200), and LDL-C level was 557 mg/dL (reference value < 130). Triglycerides and high density lipoprotein cholesterol (HDL-C) levels were normal. Although not useful for the diagnosis or for clinical purposes, we also measured the plasma levels of other lipoproteins in order to better quantify the lipid profile of the patient. In particular plasma levels of apolipoprotein (Apo) A1 was

normal, while the level of ApoB 345 mg/dL (reference value 55-140) and atherogenic lipoprotein (aLp) 734 mg/dL (reference value < 300) were increased.

Blood pressure was 16/11 kPa. Renal function tests, hemogram, thyroid function tests, immunoglobulins, erythrocyte sedimentation rate were all normal. The patient was suffering of a chronic hepatitis B with no current liver damage. He received a first course of entecavir therapy 0.5 mg once a day for the last 10 mo because he was tested positive for the hepatitis B "s" antigen (HBsAg), fluctuating or minimally elevated liver enzymes [alanine transaminase (ALT) 118 IU/L (reference value < 50) and aspartate transaminase (AST) 62 IU/L (reference value < 50) and very high viral load (real time HBV DNA 158000000 IU/mL)]. The patient was referred to our STDs Centre in order to establish if the dermatologic disorder was HBV-related. New test results for liver enzymes, HBV DNA, and sonography of the liver were negative.

Abdomino-pelvic ultrasonography, chest X-ray, brain magnetic resonance imaging and upper and lower gastrointestinal endoscopy revealed no abnormality. No osseous pathology was noted on plain radiographs.

The patient was referred to the Metabolic Disease Centre of the University of Florence, Centre for dyslipidemia management. Echocardiography was normal. However, a transoesophageal echocardiogram revealed mild supravalvular aortic narrowing and a luminal irregularity of arch. The artery doppler ultrasound scan showed that the right carotid artery had the intima media 1.5 cm thick, and the right common femoral artery had formed atherosclerotic noncalcified plaques lesions and the intima media was 2.2 cm thick.

Based on the following findings including clinical picture, patient's clinical history, clinical conditions still present in his family, and pathological and serological analysis the patient was diagnosed with HoFH and multiple xanthomas.

At this time the patient is treated with a combined treatment regimen of atorvastatin (20 mg/d), ezetimibe (10 mg/d), a low dose aspirin (100 mg/d) and LDL-C apheresis therapy every two weeks while on the list for



Figure 3 Diffuse intertriginous xanthomas. Usually appear in a symmetric distribution as well-demarcated and slightly elevated noninflammatory plaques of ochre-yellow or yellow-brown discoloration. Typically found in intertriginous and flexural areas. A: In finger web spaces, and in this picture with metacarpophalangeal joint tendon xanthoma; B: At metacarpophalangeal palmar crease in linear band or single papules; C: Toe web spaces; D: In toe web spaces and ankle crease; E and F: At antecubital fossae, with the "eruptive" appearance of crops of yellow dermal soft, velvety papules; G and H: In popliteal fossae; I and J: At the creases of ears in a rare pattern of "plane xanthoma" as very thin flat patches, easily clinically missed, of yellow-orange macular discoloration.



Figure 4 Tendinous xanthomas. Bilateral Xanthomas of Achilles tendon. Each swelling was localized all over the tendon just above its insertion point to the calcaneal tuberosity. They appear as firm, mobile, painless slowly enlarging subcutaneous nodules which may join together to form a single mass or multilobated masses. They are covered by reddish-brown thickened skin.



Figure 5 Tuberos xanthomas. They are very common and clinically variable. They may appear as firm, painless, red-yellow, waxy-appearing nodules located in the dermis and subcutaneous tissue, from few millimeters to several centimeters in size. They often present with a cobblestone-like pattern developing around the pressure areas such as: A and B: The knees; C: Malleolus; D: Buttocks. Lesions can join together to form multilobated masses.

anti-PCSK9 monoclonal antibody therapy.

DISCUSSION

Familial hypercholesterolemia (FH), is a primary hyperlipoproteinemia characterized by an autosomal codominant genetic disorder due to mutations in the LDL receptor gene located on chromosome 19. There are two types of FH: A Homozygous FH (HoFH), in that the individuals with two mutant LDL receptor alleles are much more affected than those with one mutant allele, Heterozygous FH (HeFH)^[2,6,12]. HoFH is a rare form of inherited dyslipidemia often diagnosed early in childhood which in most cases is not detected. Originally, the



Figure 6 Bilateral massive tuberous xanthomas of the elbows did not recur after surgical excision.

prevalence of HoFH was estimated as 1 per million, with higher prevalence in countries with founder mutations, especially if consanguineous marriages were present. However, the HoFH prevalence is now estimated at one in 160000 to 300000^[2,13]. The heterozygous form is the most common with an incidence of 1 out of 500, in which the patient has usually diagnosed as adult^[12,13]. Despite published data, there is not agreement about how and when perform the screening in childhood but familial history of hypercholesterolemia in parents is crucial for detection and diagnosis of HoFH^[2,11,14-16]. FH is a disease characterized by a triad: Elevated LDL-C, tendon xanthomas, and premature coronary heart disease^[6]. HoFH should be suspected if both parents have HeFH where the probability of a child having HoFH is 1 in 4^[16]. In HoFH patients, markedly elevated LDL-C concentration may be present at birth, as well as cutaneous xanthomas but generally present by the age of four. Corneal arcus is common by age ten and tendon xanthomas develop inevitably while coronary artery disease (CAD) develops from childhood on with high risk for a fatal or non-fatal coronary event by age thirty^[5,13,15]. However, the presence of xanthomas increases the risk of CAD in patients with FH by as much as three fold. For a patient with cutaneous or tendon xanthomas, the probability of FH is very high; however, an absence of xanthoma does not rule out FH^[6,15]. Epidemiologic data on cutaneous xanthomas are limited. Xanthomas are rarely seen before age twenty although those associated with FH are an exception. They tend to occur in both males and females without any sex predilection, develop inevitably and an exaggerated phenotype may be observed in patients with HoFH as was in our case^[1,4,9,17,18]. The patient in the present study presented with multiple large xanthomas with a wide ranging distribution all over the body, and an onset at the age of two. The patient had an LDL-C level of 557 mg/dL, suggesting a high likelihood of HoFH. The patient was the offspring of two parents with HeFH, and appeared to have an inherited HoFH phenotype associated with an increased level of TC and serum LDL-C and more severe symptoms than the parents. The parents had mildly elevated levels of TC

(father 330 mg/mL; mother 300 mg/mL), which, when combined with the absence of xanthomas, suggests that the parents suffered from HeFH. Only a minority of patients with lipoprotein disorders have xanthomas thus the estimation of plasma lipid levels alone may not be enough to properly identify a specific lipid metabolic disorder, on the contrary the presence of xanthoma lesions represent a useful marker for these diseases^[4]. Therefore it seems logical that skin lesions have been described as the first symptom. Cutaneous xanthomas were first introduced in the medical field by Rayer^[19] in 1835, when he described "yellow lesions on the eyelids"^[19,20]. In 1851, Addison and Gull observed various forms of xanthomas naming those "vitiligoidea"^[20,21]. This term was soon replaced by xanthoma by W. Frank Smith in 1869 and descriptive terms were added, such as "planum", "multiplex" and "tuberosum"^[10,12,22]. The unique association of FH and tendon xanthomas was reported by Fagge in 1873^[6,23]. Xanthomas are seen in 40%-50% patients of FH and HeFH is the most common cause. Of the affected individuals 50% to 75% may complain of tendon xanthomas that rarely have been reported in the setting of normal plasma levels of cholesterol. The prevalence increases from 7% in the third decennium to 50% in the sixth decennium^[1,4,10,15]. They are not palpable in up to 20% of individuals. Thus, to identify these xanthomas sonography is the most appropriate technique and is superior to clinical assessment, and even if not present an abnormal texture and thickening of Achilles tendon were demonstrated in 68% of subjects with FH^[1,24,25]. Xanthelasma are seen in 23% of cases but they are the least specific of all xanthomas representing the vast majority of cases (> 95%) and because they are seen in many hyperlipidaemic and normolipidaemic states. About 65% of adult patients with xanthelasma may show normal plasma lipid levels. Tuberosus xanthomas are reported in 10%-15% of cases and intertriginous xanthomas occurring occasionally^[1,10]. The presence of tuberosus and intertriginous xanthomas in a child with a markedly elevated plasma cholesterol level is strongly suggestive of HoFH. Intertriginous xanthomas have not been seen in the HeFH, in which plasma LDL-C are less markedly increased. In contrast, tuberoeruptive xanthomas are associated with several forms of hyperlipoproteinemia and rarely occur in patients with FH^[1,4,9,10,12,16]. From detailed literature review and according to European Atherosclerosis Association Guidelines^[6,13,16,26] our patient has met clinical criteria for a definite diagnosis of HoFH, even in the absence of a mutation in genetic testing, and was based on the following data: (1) High serum TC and LDL-C levels with normal triglyceride levels; (2) Appearance of xanthomas in the first decade of life; (3) Documentation of mildly elevated levels of LDL-C and TC and absence of xanthomas in both parents and in one of the siblings; (4) The presence of signs of atherosclerosis; and (5) The presence of multiple large xanthomas with a wide ranging distribution and above all, the rare pathognomonic intertriginous xanthomas, which have

been described as a dermatological marker of this homozygous type.

In conclusion, this case highlights the importance of proper identification of nodular lesions and a differential diagnosis of specific subtypes of xanthomas by physicians and especially dermatologists. Xanthomas cannot be considered as simple cosmetic lesions as they are the earliest clinical indicators of lipidemic disorders. The publication of individual cases seems beneficial since this case study of HoFH wants to emphasise that this disorder remains critically under-diagnosed, and a delayed diagnosis could have potentially devastating consequences because these patients progress rapidly to atherosclerotic changes leading to aortic stenosis and CAD. Nonetheless a very important problem in these patients is that most of them do not feel ill enough until a severe CAD takes place.

ARTICLE HIGHLIGHTS

Case characteristics

Cutaneous xanthomas may or may not be present with lipid metabolic disorders, usually depending on the severity of the lipid abnormality.

Clinical diagnosis

Polymorphous cutaneous xanthomas in Homozygous Familial Hypercholesterolemia (HoFH).

Differential diagnosis

The presence of specific lesions represents a useful marker to properly identify a specific hyperlipidaemic disorder.

Laboratory diagnosis

High serum total cholesterol and low-density lipoprotein cholesterol (LDL-C) levels with normal triglyceride levels.

Pathological diagnosis

Presence in the dermis of cholesterol crystalline aggregates surrounded by fibrosis and foamy cells.

Experiences and lessons

HoFH is a rare form of inherited dyslipidemia now estimated with a prevalence of one in 160000 to 300000.

Treatment

Atorvastatin, ezetimibe, low dose aspirin and LDL-C apheresis.

Related report

The presence, the clinical and dermatological features of multiple large xanthomas with a wide ranging distribution and above all, the rare pathognomonic intertriginous xanthomas, have been described as a dermatological marker of the HoFH.

REFERENCES

- 1 **Zak A**, Zeman M, Slaby A, Vecka M. Xanthomas: clinical and pathophysiological relations. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2014; **158**: 181-188 [PMID: 24781043 DOI: 10.5507/bp.2014.016]
- 2 **Palacio CH**, Harring TR, Nguyen NT, Goss JA, O'Mahony CA. Homozygous familial hypercholesterolemia: case series and review of the literature. *Case Rep Transplant* 2011; **2011**: 154908 [PMID:

- 23213598 DOI: 10.1155/2011/154908]
- 3 **Fredrickson DS**, Levy RI, Lees RS. Fat transport in lipoproteins- an integrated approach to mechanisms and disorders. *N Engl J Med* 1967; **276**: 34-42 contd [PMID: 5333081 DOI: 10.1056/NEJM196701052760107]
 - 4 **Cruz PD Jr**, East C, Bergstresser PR. Dermal, subcutaneous, and tendon xanthomas: diagnostic markers for specific lipoprotein disorders. *J Am Acad Dermatol* 1988; **19**: 95-111 [PMID: 3042820]
 - 5 **Yuan G**, Wang J, Hegele RA. Heterozygous familial hypercholesterolemia: an underrecognized cause of early cardiovascular disease. *CMAJ* 2006; **174**: 1124-1129 [PMID: 16606962 DOI: 10.1503/cmaj.051313]
 - 6 **Mabuchi H**. Half a Century Tales of Familial Hypercholesterolemia (FH) in Japan. *J Atheroscler Thromb* 2017; **24**: 189-207 [PMID: 28179607 DOI: 10.5551/jat.RV16008]
 - 7 **Jeziorska M**, Hassan A, Mackness MI, Woolley DE, Tullo AB, Lucas GS, Durrington PN. Clinical, biochemical, and immunohistochemical features of necrobiotic xanthogranulomatosis. *J Clin Pathol* 2003; **56**: 64-68 [PMID: 12499438]
 - 8 **Szalat R**, Arnulf B, Karlin L, Rybojad M, Asli B, Malphettes M, Galicier L, Vignon-Pennamen MD, Harel S, Cordoliani F, Fuzibet JG, Oksenhendler E, Clauvel JP, Brouet JC, Fermand JP. Pathogenesis and treatment of xanthomatosis associated with monoclonal gammopathy. *Blood* 2011; **118**: 3777-3784 [PMID: 21757618 DOI: 10.1182/blood-2011-05-356907]
 - 9 **Vargas-Flores E**, Estrada-Alpizar L, Arenas-Osuna J. Tuberos Xanthomatosis as a Presentation of Familial Hypercholesterolemia. *JSM Clin Case Rep* 2016; **4**: 1114
 - 10 **Kumar N**, Anand S, Yadav C, Kataria H, Kumar P, Yadav S. Xanthomatosis and Orthopaedics: Review of literature. *Int Res J Basic Clin Stud* 2014; **2**: 78-81 [DOI: 10.14303/irjbc.2014.041]
 - 11 **Fernandes M**, Pereira P. A case of type IIa Homozygous Familial Hypercholesterolemia with cutaneous xanthomas. *IJMRHS* 2015; **1**: 1-3
 - 12 **Jayaram S**, Meera S, Kadi S, Sreenivasa N. An Interesting Case of Familial Homozygous Hypercholesterolemia-A Brief Review. *Indian J Clin Biochem* 2012; **27**: 309-313 [PMID: 26405394 DOI: 10.1007/s12291-011-0165-8]
 - 13 **Janus ED**. Homozygous familial hypercholesterolaemia - Early recognition and early treatment improve outcomes. *Atherosclerosis* 2017; **260**: 147-149 [PMID: 28341574 DOI: 10.1016/j.atherosclerosis.2017.02.008]
 - 14 **Bermudez EB**, Storey L, Mayo S, Simpson G. An Unusual Case of Multiple Tendinous Xanthomas Involving the Extremities and the Ears. *Case Rep Dermatol* 2015; **7**: 340-344 [PMID: 26955329 DOI: 10.1159/000441711]
 - 15 **Zhao C**, Kong M, Cao L, Zhang Q, Fang Y, Ruan W, Dou X, Gu X, Bi Q. Multiple large xanthomas: A case report. *Oncol Lett* 2016; **12**: 4327-4332 [PMID: 28101197 DOI: 10.3892/ol.2016.5282]
 - 16 **France M**. Homozygous familial hypercholesterolaemia: update on management. *Paediatr Int Child Health* 2016; **36**: 243-247 [PMID: 27967828 DOI: 10.1080/20469047.2016.1246640]
 - 17 **Huijgen R**, Stork AD, Defesche JC, Peter J, Alonso R, Cuevas A, Kastelein JJ, Duran M, Stroes ES. Extreme xanthomatosis in patients with both familial hypercholesterolemia and cerebrotendinous xanthomatosis. *Clin Genet* 2012; **81**: 24-28 [PMID: 21955034 DOI: 10.1111/j.1399-0004.2011.01793.x]
 - 18 **Wani IH**, Farooq M, Bhat MS, Lone A, Kamal Y, Latoo I. Lumpy bumpy Achilles with gait instability could be cerebrotendinous xanthomatosis: Two case reports. *FAOJ* 2014; **7**: 3 [DOI: 10.3827/faoj.2014.0701.0003]
 - 19 **Rayer P**. Plaques jaunes folliculeuses, développées sur la paupière supérieure. Atlas du Traité des maladies de la peau, 1835: Pl VIII, figure 16, and Pl XXII, figure 15
 - 20 **Fleischmajer R**, Schragger AH. The clinical significance of cutaneous xanthomas. *Postgrad Med J* 1970; **46**: 671-677 [PMID: 4924273]
 - 21 **Addison T**, Gull W. On a Certain Affection of the Skin: Vitiligoidea-Plana-Tuberosa, with Remarks and Plates. *Guy's Hosp Rep* 1851; **7**: 265-276
 - 22 **Smith FW**. On xanthoma or vitiligoidea. *Journal of Cut Med* 1869; **T.III**: 241
 - 23 **Fagge CH**. Xanthomatous disease of the skin. General xanthelasma or vitiligoidea. *Trans Pathol Soc Lond* 1873; **24**: 242-250
 - 24 **Junyent M**, Gilabert R, Zambón D, Núñez I, Vela M, Civeira F, Pocovi M, Ros E. The use of Achilles tendon sonography to distinguish familial hypercholesterolemia from other genetic dyslipidemias. *Arterioscler Thromb Vasc Biol* 2005; **25**: 2203-2208 [PMID: 16123315 DOI: 10.1161/01.ATV.0000183888.48105.d1]
 - 25 **Dagistan E**, Canan A, Kizildag B, Barut AY. Multiple tendon xanthomas in patient with heterozygous familial hypercholesterolaemia: sonographic and MRI findings. *BMJ Case Rep* 2013; **2013**: pii: bcr2013200755 [PMID: 24252837 DOI: 10.1136/bcr-2013-200755]
 - 26 **Cuchel M**, Bruckert E, Ginsberg HN, Raal FJ, Santos RD, Hegele RA, Kuivenhoven JA, Nordestgaard BG, Descamps OS, Steinhagen-Thiessen E, Tybjaerg-Hansen A, Watts GF, Averna M, Boileau C, Borén J, Catapano AL, Defesche JC, Hovingh GK, Humphries SE, Kovanen PT, Masana L, Pajukanta P, Parhofer KG, Ray KK, Stalenhoef AF, Stroes E, Taskinen MR, Wiegman A, Wiklund O, Chapman MJ; European Atherosclerosis Society Consensus Panel on Familial Hypercholesterolaemia. Homozygous familial hypercholesterolaemia: new insights and guidance for clinicians to improve detection and clinical management. A position paper from the Consensus Panel on Familial Hypercholesterolaemia of the European Atherosclerosis Society. *Eur Heart J* 2014; **35**: 2146-2157 [PMID: 25053660 DOI: 10.1093/eurheartj/ehu274]

P- Reviewer: Kaliyadan F, Palmirotta R, Vasconcellos C

S- Editor: Ji FF **L- Editor:** A **E- Editor:** Lu YJ





Published by **Baishideng Publishing Group Inc**
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA
Telephone: +1-925-223-8242
Fax: +1-925-223-8243
E-mail: bpgoffice@wjgnet.com
Help Desk: <http://www.f6publishing.com/helpdesk>
<http://www.wjgnet.com>

