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Case Control Study

Does low volume high-intensity interval training elicit superior benefits to continuous low to moderate-intensity training in cancer survivors?

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Abstract

AIM

To determine the impact of low volume high-intensity interval training (LVHIIT) and continuous low to moderate-

intensity exercise training (CLMIT) on cardiovascular disease (CVD) risk and health outcomes in cancer survivors.

METHODS

Sedentary cancer survivors ($n = 75$, aged 51 ± 12 year) within 24 months of diagnosis, were randomised into three groups for 12 wk of LVHIIT ($n = 25$), CLMIT ($n = 25$) or control group ($n = 25$). The exercise intervention involved 36 sessions (three sessions per week). The LVHIIT group performed 7 x 30 s intervals ($\geq 85\%$ predicted maximal heart rate) with a 60 s rest between intervals, and the CLMIT group performed continuous aerobic training for 20 min ($\leq 55\%$ predicted maximal heart rate) on a stationary bike. Outcome variables were measured at baseline and at 12 weeks and analysed using a 3 x 2 (group x time) repeated measures ANCOVA to evaluate main and interaction effects.

RESULTS

Significant improvements (time) were observed for seven of the 22 variables (ES 0.35-0.97, $P \leq 0.05$). There was an interaction effect ($P < 0.01$) after 12 wk in the LVHIIT group for six-minute walk test ($P < 0.01$; $d = 0.97$; 95%CI: 0.36, 1.56; large), sit to stand test ($P < 0.01$; $d = -0.83$; 95%CI: -1.40, -0.22; large) and waist circumference reduction ($P = 0.01$; $d = -0.48$; 95%CI: -1.10, 0.10; medium). An interaction effect ($P < 0.01$) was also observed for quality of life in both the LVHIIT ($d = 1.11$; 95%CI: 0.50, 1.72; large) and CLMIT ($d = 0.57$; 95%CI: -0.00, 1.20; moderate) compared with the control group ($d = -0.15$; 95%CI: -0.95, 0.65; trivial).

CONCLUSION

Low-volume high-intensity training shows promise as an effective exercise prescription within the cancer population, showing greater improvements in cardio-respiratory fitness, lower body strength and waist circumference compared with traditional CLMIT and control groups. Both LVHIIT and CLMIT improved quality of life. A proposed benefit of LVHIIT is the short duration (3 min) of exercise required, which may entice more cancer survivors to participate in exercise, improving health outcomes and lowering the risk of CVD.

Key words: High-intensity exercise; Health; Oncology; Exercise prescription

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Core tip: Low-volume high-intensity training is not commonly used in the rehabilitation of Cancer Survivors. In this study it shows promise as an effective exercise prescription, with greater improvements in cardio-respiratory fitness, lower body strength and waist circumference compared with traditional continuous low to moderate-intensity exercise training (CLMIT) and control groups. Low volume high-intensity interval

training (LVHIIT) and CLMIT improved quality of life. A proposed benefit of LVHIIT is the short duration (3 min) of exercise required, which may entice more cancer survivors to participate in exercise, improving health outcomes and lowering the risk of cardiovascular disease.

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INTRODUCTION

Worldwide there is an increase in cancer survival rates^[1]. This potentially raises the risk of cancer recurrence and other non-communicable diseases (NCDs) such as type II diabetes and cardiovascular disease (CVD)^[2]. These increased health risks may be due to the effects of cancer treatments and reductions in healthy lifestyle habits, such as physical activity (PA)^[3,4]. Physical activity decreases NCD risk in apparently healthy people^[5], though it is not conclusive if the same trends are evident in individuals with cancer. Exercise during and after cancer treatment has been shown to be safe, improve fitness levels, and quality of life (QoL)^[6,7]. Because of this, there is significant interest in the clinical use of exercise as an adjunctive therapy for improving cancer-related health outcomes.

Evidence is rapidly increasing regarding the benefits of exercise concurrent to treatment in the remediation of adverse clinical outcomes for cancer survivors^[8]. It is uncommon for cancer survivors to be advised by clinicians to participate in an exercise program, despite the existence of exercise guidelines^[9,10]. This may be due to the generic nature of the guidelines, limited practitioner expertise or the costs associated with some programs^[11,12]. The present exercise guidelines lack detail on the type, mode, duration and intensity of exercise necessary to achieve the best outcomes for cancer survivors. Additional research is required to fill the gaps in our current knowledge to further improve exercise recommendations, the evidence-based knowledge and exercise education for cancer survivors and their treating practitioners.

Low volume high-intensity interval training (LVHIIT) is the use of small doses of high-intensity exercise to elicit physiological responses such as improved VO₂ max and positive metabolic changes in skeletal muscle. The physiological changes could potentially be much higher or different to those currently being obtained with participation in other doses of activity, such as continuous, moderate-intensity exercise^[13]. LVHIIT has been shown to improve VO₂ peak and insulin sensitivity in apparently healthy individuals in as little as

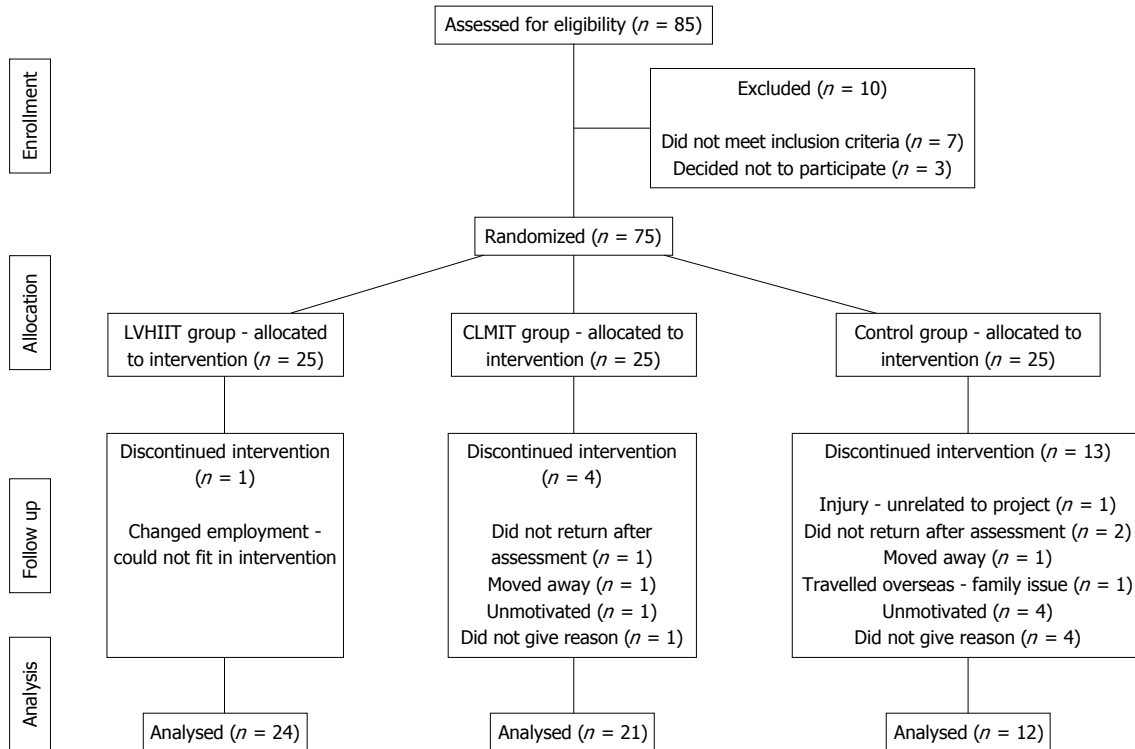


Figure 1 Consort diagram. LVHIIT: Low volume high-intensity interval training; CLMIT: Continuous low to moderate-intensity training.

four weeks^[14,15]. There is limited research examining the effects of LVHIIT in improving health outcomes for cancer survivors. Its application as a modality for use with other chronic disease populations is evolving^[14,16,17], potentially due to its ability to elicit positive physiological improvements in a short period. The current study aimed to investigate and compare the effects of LVHIIT and the more commonly prescribed continuous low to moderate-intensity training (CLMIT) on improving health outcomes and reducing cardiovascular disease (CVD) risk in cancer survivors.

MATERIALS AND METHODS

Participants

Eighty-five cancer survivors (83 female and 2 male, 51.48 ± 12.45 years) were recruited for the 12-wk study. Recruitment was conducted between September 2014 and June 2016 *via* email, pamphlet distribution, word of mouth and online social media. Referrals were obtained from the medical community and community organisations. The inclusion criteria included: (1) Participants within the first 24 mo of diagnosis; (2) in the post-treatment phase of the “physical activity across the cancer experience” (PEACE) organisational model, once the acute effects of medical treatments had dissipated^[18], and (3) sedentary, as described by the American College of Sports Medicine^[19]. Participants were excluded if they had: (1) Brain or metastatic bone cancers; (2) bone pain; (3) resting blood pressure $> 180/110$ mmHg; (4) were pregnant; (5) undergoing psychotherapy treatment; (6) had musculoskeletal

injuries or disabilities restricting their ability to participate in exercise.

Randomisation

Of the 85 participants, seven participants did not meet the inclusion criteria, and three decided not to participate due to the timing of exercise sessions. The 75 remaining participants were randomly assigned *via* an online randomisation tool into either LVHIIT ($n = 25$) or CLMIT ($n = 25$) or control ($n = 25$) group (Figure 1). A person independent of the research team used the research randomizer computer software^[20] to allocate participant codes into the three groups (LVHIIT, CLMIT, and control).

Of the 75 participants, 57 completed the study (76%). In the LVHIIT group ($n = 24$), one participant changed employment and could not complete the intervention. In the CLMIT group ($n = 21$), one participant did not return after baseline assessment, one moved interstate, one was not motivated to continue, and one failed to provide a reason. In the control group ($n = 12$) one participant sustained an injury (unrelated to the project), two did not return after baseline assessment, one moved interstate, one traveled overseas, four failed to respond to the final evaluation and four did not provide a reason (Figure 1).

Quality of life

The Functional Assessment of Cancer Therapy-General (FACT-G) questionnaire (version 4) was used to measure quality of life (QoL) and functional capacity^[21]. The FACT-G is a validated survey containing 27 items. The questions are in four categories: (1) physical well-

being; (2) social/family well-being; (3) emotional well-being; and (4) functional well-being. The questionnaire is regularly used to measure QoL in cancer survivors^[22] and was completed at baseline and then after the 12-wk intervention.

Anthropometrics

The Dual X-Ray Absorptiometry (DXA) scan (GE Healthcare, Sydney, NSW, Australia) was used to measure total body composition, including; lean mass, body weight and body fat percentage^[23]. The DXA scanner was calibrated each day, using a phantom spine. The manufacturers' guidelines were followed to carry out daily quality control checks. All scans were carried out by trained densitometrists. Participants were asked to fast overnight and wear no jewellery, while being scanned. Hip and waist circumferences were measured using a standard anthropometric tape measure^[24,25]. The same individual measured the circumferences at baseline and post-intervention using WHO STEPwise approach measurement protocols^[25,26].

Cardiovascular functioning

Pulse wave velocity (PWV) and pulse wave analysis (PWA) were measured using the SphygmoCor XCEL system (SphygmoCor; At-Cor Medical Pty Ltd., Sydney, Australia). Carotid-femoral PWV is the recognised gold standard measure of aortic stiffness, a strong independent predictor of cardiovascular risk^[27,28]. PWA, which included measures of resting heart rate (RHR), augmentation index (AIx), central systolic blood pressure (CSP), central diastolic blood pressure (CDP), central pulse pressure (CPP), augmentation pressure (AP), mean arterial pressure (MAP), systolic blood pressure (SBP) and diastolic blood pressure (DBP) were assessed at baseline and upon completion of the intervention. The pulse pressure (PP) waveform of the left carotid artery was measured with an applanation tonometer. Participants rested supine for 10 min before the measurements were obtained. Twenty continuous waveforms were essential for results to be considered valid and these were used to acquire the PP waveform of the aorta, and PWV was used as a marker of aortic stiffness^[29].

Biomarkers

Participants fasted overnight in preparation for blood sample analysis. Samples were analysed by an independent laboratory for high-sensitive C-reactive protein (CRP), insulin, glucose, and full blood count. The analysis was conducted offsite at Capital Pathology, Canberra, Australia.

Functional capacity

Lower body strength was measured using a repeated chair rise test (STS). The participants sat in a chair and were asked to stand and then sit as fast as possible five times, without the use of their arms^[30,31]. The six-minute walk test (6MWT) was used to delineate participants' cardiorespiratory fitness levels. Participants were asked

to walk as quickly as possible for six minutes and the distance traveled was recorded^[32]. The 6MWT is used extensively to assess cardiorespiratory fitness in cancer survivors^[33,34].

Intervention

An Accredited Exercise Physiologist (AEP) supervised all exercise sessions performed over the 12-wk intervention period (three sessions per week). Participants were asked to refrain from consuming food and caffeine and participating in any exercise for two hours before baseline and post-intervention assessments. Assessments were carried out within seven days before commencement and within seven days following completion of the program^[29]. The exercise sessions were carried out on a stationary bike. The LVHIIT group performed interval training ($\geq 85\%$ maximal heart rate), which consisted of a five-minute warm up, seven by 30 s intervals, with one-minute rest in between each interval, followed by a five-minute cool down (adapted from Gibala, 2012). A gradual increase in exercise was carried out by the LVHIIT group. Individuals started the first session with three intervals, with one interval added per week over the following four weeks and by the 5th week participants performed all seven intervals^[29]. The CLMIT group performed continuous aerobic training ($\leq 55\%$ predicted maximal heart rate) for 20 min also with a five-minute warm up and cool down^[29]. Age-predicted maximal heart rate was used to calculate relative intensity and the two exercise protocols were matched for appropriate energy expenditure using the calculation reported by Rognmo *et al.*^[36] (2004). For each participant, their heart rate (HR) and rate of perceived exertion (RPE) were logged every minute for the CLMIT sessions. The peak HR and RPE in each interval and the resting HR and RPE between each interval were recorded during the LVHIIT sessions. Blood pressure (BP) was monitored immediately before and after each exercise session.

Statistical analysis

Means and standard deviations were calculated for dependent variables. An ANOVA was performed to determine if pre-intervention differences existed between the three groups. A Bonferroni post-hoc multiple comparison test was conducted to determine where group differences existed. A 3 x 2 (group x time) repeated measures ANCOVA^[29] was used to evaluate main and intervention effects and was adjusted by baseline values. Effect sizes (ES) were calculated, and Cohen's *d* values were interpreted as follows: large ≥ 0.8 , medium ≥ 0.5 , small ≥ 0.2 and trivial < 0.2 ^[37]. The alpha level was set at $P < 0.05$, and SAS 9.3 (Cary, NC, United States) was used for all analyses.

RESULTS

Participant characteristics

Participant cancer diagnosis included 47 (82%) breast

Table 1 Baseline characteristics of participants

Characteristics	LVHIIT <i>n</i> = 24	CLMIT <i>n</i> = 21	Control <i>n</i> = 12
Sociodemographic			
Age, mean (SD), yr	48 (11.9)	52 (12.4)	57 (11.5)
Sex, <i>n</i> (%) male	F, 25 (0)	F, 21 (0)	F, 12 (0)
Comorbidity, <i>n</i> (%) yes	2 (0.8)	1 (0.4)	0 (0)
Diagnosis, <i>n</i> (%)			
Breast	21 (88)	16 (75)	10 (83)
Ovarian		1 (5)	1 (8.5)
Appendix		1 (5)	
Anal			1 (8.5)
Cervical	1 (4)		
Liver	1 (4)		
Esophageal	1 (4)		
Melanoma		1 (5)	
Leiomyosarcoma		1 (5)	
Unknown primary		1 (5)	
Stage of disease, <i>n</i> (%)			
Stage I - II	19 (79)	16 (76)	10 (83)
Stage II -IV	5 (21)	5 (24)	2 (17)
Type of treatment, <i>n</i> (%)			
Surgery	21 (88)	20 (95)	12 (100)
Radiation therapy	18 (75)	13 (62)	10 (83)
Hormone therapy	19 (79)	15 (71)	9 (75)
Types of chemotherapy, <i>n</i> (%)			
TAC	7 (29)	4 (19)	1 (8)
FEC	5 (21)	3 (14)	3 (25)
TAC/FEC combinations	2 (8)	3 (14)	2 (17)
Capecitabine and oxaliplatin		1 (5)	
Carboplatin and paclitaxel	1 (4)		1 (8)
Cisplatin	1 (4)		
CHOP	1 (4)		
ABVD	1 (4)	1 (5)	1 (8)
Other	2 (8)	3 (14)	
No chemotherapy	4 (17)	6 (29)	4 (33)

LVHIIT: Low volume high-intensity training; CLMIT: Continuous low to moderate-intensity training; TAC: Taxotere, adriamycin, cyclophosphamide; FEC: Fluorouracil, epirubicin, cyclophosphamide; CHOP: Cyclophosphamide, doxorubicin, vincristine, prednisone; ABVD: Doxorubicin, bleomycin, vinblastine, dacarbazine.

cancer, two (3%) ovarian cancer and one diagnosis of appendix, anal, cervical, liver, oesophageal, melanoma, leiomyosarcoma and unknown primary (15%).

The age of the participants was 51.48 ± 12.45 years, with a BMI of 26.43 ± 4.08 kg/m² (Table 1). Baseline values for all variables were similar across the three groups except for white blood cell count, which was higher in the LVHIIT group compared with both the CLMIT group and control groups ($P < 0.02$).

For the LVHIIT group, the average HR during the 36 sessions was 147 ± 11 beats per minute (bpm), while RPE was 6 ± 3 using the Borg 1-10 scale^[38,39]. The average HR and RPE at the end of each 60s recovery was 117 ± 8 bpm and RPE of 3 ± 2 . The average HR for the CLMIT group was 98 ± 6 bpm and RPE was 3 ± 2 during the sessions. Overall mean session compliance was 92% (72%-100%). Ten participants attended all 36 sessions, and one participant missed ten sessions over the intervention period (data were included). There were no adverse events reported in the study.

Fitness, functional and body composition measures

There was a significant interaction effect ($P < 0.01$)

for 6MWT. Distance walked was significantly longer for the LVHIIT group ($d = 0.97$; 95%CI: 0.36, 1.56; large) when compared with the CLMIT ($d = 0.17$; CI: -0.23, 0.99; trivial) and control ($d = -0.13$; 95%CI: -0.93, 0.67; trivial) groups (Figure 2A). There was a significant interaction effect ($P < 0.01$) for the STS test with a faster performance in the LVHIIT group ($d = -0.83$; 95%CI: -1.40, -0.22; large) compared with the CLMIT ($d = -0.59$; 95%CI: -1.20, 0.42; unclear) and control ($d = 0.36$; CI: -0.44, 1.17; unclear) groups (Figure 2B). There was a significant interaction ($P = 0.01$) effect for waist circumference; post results were significantly lower for LVHIIT ($d = -0.48$; 95%CI: -1.10, 0.10; medium) compared with CLMIT ($d = -0.05$; 95%CI: -0.66, 0.55; trivial) and control group ($d = 0.11$; 95%CI: -0.69, 0.91; trivial).

Quality of life

Overall: There was an interaction effect for overall QoL ($P < 0.01$). The QoL score was significantly improved in the LVHIIT group ($d = 1.11$; 95%CI: 0.50, 1.72; large) compared with the CLMIT group ($d = 0.57$; 95%CI: -0.00, 1.20; moderate) and the control group ($d =$

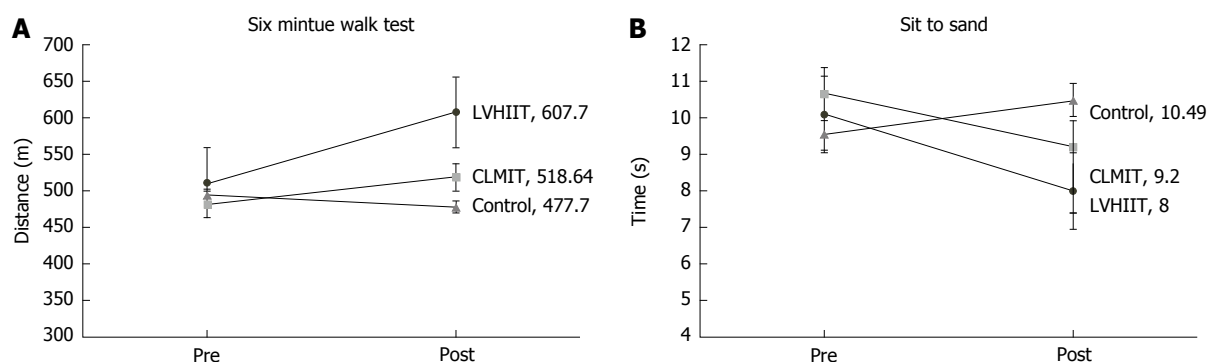


Figure 2 Changes in cardiorespiratory fitness and sit to stand for low volume high-intensity interval training, continuous low to moderate-intensity exercise training and control groups.

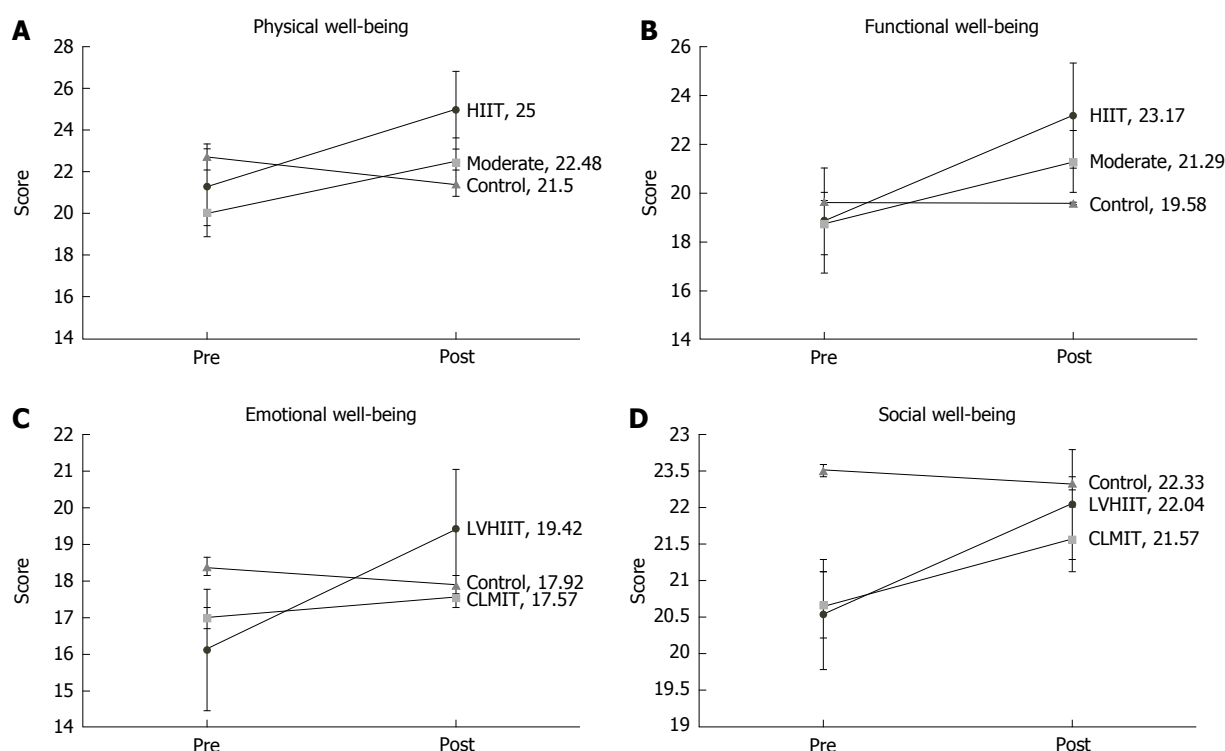


Figure 3 Changes in quality of life subscales for low volume high-intensity interval training, continuous low to moderate-intensity exercise training, and control groups.

-0.15; 95%CI: -0.95, 0.65; trivial) (Figure 3).

Well-being: An interaction effect was observed for physical well-being ($P = 0.02$). Improvements were seen in the LVHIIT ($d = 0.89$; 95%CI: 0.30, 1.48; large) and CLMIT ($d = 0.61$; 95%CI: -0.001, 1.23; medium) compared with the control group ($d = -0.30$; 95%CI: -1.09, 0.51; unclear) (Figure 3A). There was an interaction effect observed for functional well-being ($P = 0.02$). The LVHIIT group significantly improved ($d = 0.96$; 95%CI: 0.37, 1.56; large) compared to the CLMIT group ($d = 0.64$; 95%CI: 0.02, 1.26; medium) and the control group ($d = -0.02$; 95%CI: -0.82, 0.78; trivial) (Figure 3B). An interaction effect was also found for emotional well-being ($P < 0.01$). Improvements were observed for the LVHIIT group ($d = 1.04$, 95%CI:

0.43, 1.64; large) compared with the CLMIT group ($d = 0.15$; 95%CI: -0.45, 0.76; trivial) and the control group ($d = -0.11$; 95%CI: -0.92, 0.69; trivial) (Figure 3C). No interaction effect was observed for social well-being ($P = 0.057$) (Figure 3D). Changes were observed in the LVHIIT ($d = 0.35$; 95%CI: -0.22, 0.92; medium) compared to the CLMIT group ($d = 0.20$; 95%CI: -0.40, 0.80; unclear) and the control group ($d = -0.05$; 95%CI: -0.85, 0.75; trivial).

Cardiovascular function and biomarkers

There were no interaction effects for vessel stiffness parameters or biomarkers. However, time effects were found for markers of cardiovascular functioning. CSP ($P = 0.07$) in the LVHIIT group ($d = -0.51$; 95%CI: -1.07-0.10, moderate) which reduced by 4.94%. PP in

Table 2 Changes in risk factors from pre-to post intervention (Mean \pm SD)

Variable	LVHIIT group <i>n</i> = 24			CLMIT group <i>n</i> = 21			Control group <i>n</i> = 12		
	Pre	Post	ES (95%CI)	Pre	Post	ES (95%CI)	Pre	Post	ES (95%CI)
Weight (kg)	72.9 \pm 11.7	72.8 \pm 11.4	-0.01 (-0.57, 0.56)	69.4 \pm 14.3	70.4 \pm 14.9	0.06 (-0.54, 0.67)	65.5 \pm 13.1	65.9 \pm 13.9	0.03 (-0.77, 0.83)
Body fat (%)	42.2 \pm 7.9	41.4 \pm 7.8	-0.1 (-0.67, 0.46)	43.0 \pm 6.7	44.4 \pm 9.5	0.17 (-0.44, 0.78)	37.7 \pm 9.2	37.0 \pm 10.5	-0.07 (-0.87, 0.73)
Fat mass (kg)	30.6 \pm 9.1	29.2 \pm 8.7	-0.16 (-0.72, 0.41)	30.5 \pm 12.4	31.0 \pm 12.1	0.05 (-0.56, 0.65)	24.3 \pm 9.0	24.4 \pm 9.5	0.01 (-0.79, 0.81)
Lean mass (kg)	40.7 \pm 6.4	41.0 \pm 7.2	0.05 (-0.52, 0.61)	38.8 \pm 6.2	39.3 \pm 6.8	0.07 (-0.53, 0.68)	38.8 \pm 6.7	39.2 \pm 6.9	0.06 (-0.74, 0.86)
Waist (cm) ^{1,2}	91.1 \pm 11.7	85.7 \pm 10.5 ¹	-0.48 (-1.10, 0.10)	93.1 \pm 13.1	92.4 \pm 13.3	-0.05 (-0.66, 0.55)	89.1 \pm 9.5	90.3 \pm 11.6	0.11 (-0.69, 0.91)
Hip (cm) ¹	108.3 \pm 10.3	104.8 \pm 10.5 ¹	-0.37 (-0.91, 0.23)	107.0 \pm 11.3	107.0 \pm 14.8	-0.07 (-0.67, 0.54)	104.1 \pm 9.4	104.3 \pm 8.9	-0.02 (-0.82, 0.78)
Resting HR (bpm)	76.3 \pm 12.0	73.3 \pm 12.4	-0.24 (-0.81, 0.32)	73.9 \pm 11.9	72.3 \pm 9.8	-0.14 (-0.75, 0.46)	74.5 \pm 11.9	73.7 \pm 12.6	-0.07 (-0.87, 0.74)
SBP (mmHg)	127.3 \pm 18.3	121.8 \pm 9.7	-0.39 (-0.95, 0.20)	130.9 \pm 15.5	130.4 \pm 14.7	-0.58 (-0.64, 0.57)	123.9 \pm 15.4	128.1 \pm 12.7	0.3 (-0.51, 1.10)
DBP (mmHg)	79.7 \pm 8.3	77.8 \pm 6.9	-0.25 (-0.82, 0.32)	81.5 \pm 11.7	81.4 \pm 6.5	-0.01 (-0.62, 0.59)	78.2 \pm 5.3	79.9 \pm 6.1	0.30 (-0.51, 1.10)
MAP	97.2 \pm 10.8	93.9 \pm 7.9	-0.36 (-0.92, 0.22)	100.7 \pm 12.5	98.4 \pm 8.9	-0.21 (-0.82, 0.39)	96.6 \pm 6.2	95.9 \pm 8.6	-0.09 (-0.89, 0.71)
CSP (mmHg) ¹	118.6 \pm 17.7	111.3 \pm 10.7 ¹	-0.51 (-1.07, 0.10)	121.8 \pm 15.9	119.9 \pm 8.9	-0.14 (-0.75, 0.46)	116.7 \pm 12.1	115.3 \pm 13.5	-0.11 (-0.91, 0.69)
PP (mmHg)	35.7 \pm 12.8	30.8 \pm 6.8	-0.5 (-1.05, 0.10)	35.7 \pm 10.6	36.0 \pm 7.5	0.04 (-0.57, 0.64)	35.0 \pm 11.4	34.1 \pm 11.4	-0.08 (-0.88, 0.72)
AP (mmHg) ¹	8.1 \pm 8.1	5.9 \pm 4.4 ¹	-0.35 (-0.91, 0.23)	9.3 \pm 8.6	7.0 \pm 4.8	-0.34 (-0.94, 0.28)	10.3 \pm 6.6	8.0 \pm 9.2	-0.29 (-1.09, 0.52)
Aix (%)	19.9 \pm 14.7	18 \pm 11.6	-0.14 (-0.71, 0.42)	22.5 \pm 18.4	11.59 \pm 18.8	-0.24 (-1.20, 0.03)	26.6 \pm 12.5	10.0 \pm 18.3	-1.06 (-1.91, -0.20)
CDP (mmHg)	83.25 \pm 9.13	80.3 \pm 7.9	-0.34 (-0.92, 0.22)	85.9 \pm 10.6	82.8 \pm 7.1	-0.34 (-0.95, 0.27)	82.7 \pm 6.9	81.1 \pm 7.1	-0.22 (-1.03, 0.57)
PWV (m/s)	6.6 \pm 1.5	6.5 \pm 1.2	-0.12 (-0.64, 0.49)	6.8 \pm 1.5	6.9 \pm 1.0	0.11 (-0.53, 0.68)	6.8 \pm 1.5	6.2 \pm 1.4	-0.41 (-1.22, 0.40)
STS (s) ^{1,2}	10.1 \pm 2.8	8.1 \pm 2.1 ¹	-0.83 (-1.40, -0.22)	10.6 \pm 2.8	9.2 \pm 2.0	-0.59 (-1.20, 0.42)	9.6 \pm 2.3	10.49 \pm 2.7	0.36 (-0.44, 1.17)
6MWT (m) ^{1,2}	510.7 \pm 114.9	607.7 \pm 85.5 ¹	0.97 (0.36, 1.56)	483.1 \pm 72.3	518.6 \pm 94.5	0.17 (-0.23, 0.99)	494.2 \pm 128.7	477.7 \pm 127.1	-0.13 (-0.93, 0.67)
Glucose (mmol/L)	4.9 \pm 0.0	4.8 \pm 0.5	-0.28 (-0.85, 0.29)	5.1 \pm 0.6	4.9 \pm 0.7	-0.31 (-0.92, 0.30)	5.1 \pm 0.4	4.9 \pm 0.4	-0.5 (-1.31, 0.31)
CRP (mg/L)	2.9 \pm 3.5	2.7 \pm 3.2	-0.07 (-0.63, 0.50)	4.5 \pm 5.1	4.7 \pm 4.9	0.04 (-0.56, -0.64)	2.0 \pm 1.1	2.0 \pm 1.6	0 (-0.80, 0.80)
Insulin (mU/L)	11.4 \pm 6.8	9.0 \pm 4.9	-0.41 (-1.36, -0.19)	13.1 \pm 10.4	11.3 \pm 9.3	-0.18 (-0.79, -0.42)	9.9 \pm 5.6	10.7 \pm 6.3	0.13 (-0.67, 0.94)
WBC ($\times 10^9$ /L) ¹	6.9 \pm 2.5	5.7 \pm 2.5 ¹	-0.46 (-1.05, 0.10)	5.6 \pm 2.0	5.2 \pm 1.8	-0.21 (-0.82, -0.40)	5.4 \pm 0.7	4.9 \pm 0.9	-0.62 (-1.44, 0.20)

¹Time effect; ²Interaction effect. ES: Effect size; HR: Heart rate; SBP: Systolic blood pressure; DBP: Diastolic blood pressure; MAP: Mean arterial pressure; CSP: Central systolic pressure; PP: Central pulse pressure; AP: Arterial pressure; Aix: Augmentation index (stiffness); CDP: Central diastolic pressure; PWV: Pulse wave velocity; STS: Sit-to-stand test; 6MWT: 6-min walk test; CRP: C-reactive protein; WBC: White blood cell.

the LVHIIT group decreased by 7.96% ($d = 0.50$; 95%CI: -1.05 – 0.10, moderate). AP ($P = 0.02$) in the LVHIIT group ($d = -0.35$; 95%CI: -0.91 – 0.23, small) reduced by 27%. A 22% decrease was observed in the CLMIT group ($d = -0.34$; 95%CI: -0.94 – 0.28, small) (Table 2).

DISCUSSION

The aim of this study was to determine the impact of LVHIIT and CLMIT on CVD risk and health outcomes in cancer survivors. There were significant improvements in functional capacity, specifically cardiorespiratory fitness, lower-limb strength, and waist circumference in participants who completed the LVHIIT compared with the CLMIT and control groups. QoL improved in both the LVHIIT group and the CLMIT group when compared with the control group, however a greater effect was observed in the LVHIIT group.

Maintaining or improving functional capacity and lower limb strength in cancer survivors is essential as it enhances survivors' ability to move and carry out physical activities during and after treatment⁴⁰. It also provides the functional strength to start and adhere to an exercise program which increases levels of activity^{41,42}. Building lean mass through exercise builds a healthy metabolic profile, which is imperative as it assists in improving risk factors in those who have spent an increased volume of

time sedentary during and following, intensive cancer treatment^[43]. Participants in the LVHIIT group gained a larger effect in the sit to stand test compared with the other groups indicating improvements in lower limb strength. Possible mechanisms involved in this change could have been: (1) The increased power-output and the energy system/muscle fibre types recruited during the LVHIIT, similar to those recruited in the STS test; (2) an increased activation of the type IIa and type IIx fibres; and (3) an enhanced level of activation of the ATP/PC and glycolysis systems. An increased level of neuromuscular coordination is required at a higher-intensity of exercise, which also could have contributed to the improved strength and performance in the STS test seen in the LVHIIT group. Improvements were observed in the STS test in the LVHIIT group (20%), and in the CLMIT group (9.5%), highlighting greater increases in lower leg strength and functional capacity for those who completed the LVHIIT program. This finding was identified in our pilot study, however, due to small numbers, it was non-significant^[29].

Significant improvements in cardiorespiratory fitness levels were identified through the increased distance covered in the 6MWT, with a larger effect observed in the LVHIIT group. These findings were also detected in the authors pilot study^[29] and reported in other randomised controlled trials in cancer survivors^[7,29,44]. The greater improvements seen in the LVHIIT group suggests that more comprehensive cardiovascular adaptations may occur with the use of high-intensity exercise. Potential mechanisms involved in increases in cardiorespiratory fitness could be due to the increased level of mitochondrial enzymes recruited in high-intensity exercise^[45], which contributes to enhanced aerobic capacity of the skeletal muscles commonly seen after participating in high-intensity training. The positive impacts can contribute to changes in VO₂ difference, thereby increasing maximal aerobic capacity. Improvements in cardiorespiratory fitness using high-intensity interval training has been extensively reported in healthy people, but less in the cancer survivor population^[46]. LVHIIT could decrease the risk of developing additional chronic diseases such as CVD and diabetes^[47] and increase cancer survival rates^[48,49] faster than the commonly prescribed moderate-intensity exercise. It has been shown that cancer survivors have decreased levels of cardiorespiratory fitness levels in comparison with healthy people^[50].

A cancer diagnosis has a significant impact on behaviours which in turn can negatively affect QoL^[51]. QoL reductions can have a profound effect on the recovery of cancer survivors, reducing the probability of being able to move freely and maintain physical activity levels^[52]. A novel and clinically relevant finding of the present study was that QoL improvements were seen in both the LVHIIT and the CLMIT group, with a greater increase observed in the LVHIIT group. To date, very few studies have investigated how different exercise protocols or intensities impact QoL^[44]. While

current exercise recommendations for cancer survivors prescribe low to moderate intensity exercise, evidence is accumulating that there may be greater improvements in QoL when participating in more vigorous exercise^[11]. In most cases QoL changes are monitored from cancer diagnosis, to inform clinical decisions to improve patient outcomes. Linking exercise in with this conversation may assist clinicians in improving healthy behaviours to improve QoL^[53].

White blood cell count was significantly reduced from baseline to post-exercise intervention in the exercise groups, with the most pronounced reduction seen in the LVHIIT group, with a small effect found. Lung cancer participants who participated in a program using resistance band exercise over 12 wk in conjunction with chemotherapy treatment showed similar results^[54]. Normal WBC levels in cancer survivors are important because high levels of WBC's are associated with chronic inflammation, autonomic nervous system imbalances and may contribute to reductions in insulin sensitivity^[55]. Similarly, low WBC counts compromises an already compromised patient. Potentially both these factors would contribute to fatigue, obesity and increased risk for CVD^[56,57]. To date, little work has been carried out to determine the mechanism involved in the changes in WBC's in cancer survivors after different exercise programs^[57]. Insulin levels decreased in the LVHIIT group with a small effect and increased in the CLMIT group by 13.74% with an unclear effect. These results were not significant, yet they are potentially clinically relevant findings^[58]. Even a small decrease in insulin is essential in this population because of its reported role in the growth of cells, including the potential growth of tumour cells^[58-60].

In the current study, a moderate effect was found in the LVHIIT group for CSP and PP with unclear effects in the CLMIT and control groups. The importance of these changes relates to the negative impact of chemotherapy drugs on the cardiovascular system. Chemotherapy drugs have been found to have a long-lasting anti-angiogenic effect on the cardiovascular system^[61]. Specifically, chemotherapy has been shown to negatively impact vessel stiffness and CDP during the treatment period, with increases in BP remaining long after the treatment period^[61]. Anthracyclines, cyclophosphamide and tyrosine kinase inhibitors have all been shown to increase oxidative stress^[62,63]. Increases in oxidative stress can cause an overproduction of cytokines which damage the vessel wall causing endothelial dysfunction^[64-66]. Endothelial dysfunction can also be caused by radiotherapy; with the adverse effects potentially having a lifelong impact on cancer survivors^[65,67]. Little is known about the impact of exercise on endothelial dysfunction in cancer survivors. The current study suggests that exercise-induced cardiovascular improvements may be one mechanism whereby exercise diminishes the adverse effects that chemotherapy has on the cardiovascular system.

Limitations of this investigation were that diet; and

daily physical activity levels were not controlled for in the time periods between study sessions. Although 6MWT has been shown to be a valid and reliable measure of fitness in cancer survivors, a more robust cardiorespiratory fitness test such as VO₂ max testing could be used. There were no male participants in this study, and as such the data should be interpreted accordingly. Participants were prescribed different ongoing medications and treatment protocols, all of which could have impacted the results; this does however represent a real world group of cancer survivors.

In conclusion, this study shows promise for the use of LVHIIT in the cancer population. The encouraging results opens up the possibility of introducing LVHIIT in therapy programs, as a shorter and more efficacious exercise to increase the fitness levels in cancer survivors. The LVHIIT protocol improved fitness and functional capacity and decreased waist circumference compared with CLMIT. Both LVHIIT and CLMIT improved QoL. LVHIIT may be an effective alternative to traditional exercise prescription within this population. The benefit of LVHIIT is that for selected variables it produces more pronounced results compared with CLMIT and it is short in duration which could entice more cancer survivors to participate in exercise as time is a barrier^[68]. This study highlights that the most commonly prescribed CLMIT may not be enough to induce clinically relevant changes in cancer survivors. Additional research is required to fully understand the mechanisms involved in the changes identified in this study in relation to different doses of exercise. This research would be highly beneficial to assist clinicians in the optimisation of clinical exercise recommendations for cancer survivors.

ARTICLE HIGHLIGHTS

Research background

Research into the optimum exercise guidelines for cancer survivors are not conclusive. Little evidence exists for the use of low-volume high-intensity interval training (LVHIIT) within the cancer population, even though it shows promise in other chronic populations. LVHIIT has been used in populations such as stroke, diabetes, cardiovascular disease (CVD), cardiac rehabilitation showing more pronounced health benefits than the more commonly prescribed continuous low-moderate intensity training (CLMIT). Therefore, it should be further investigated for use with cancer survivors as it is a time efficient exercise modality, with greater health benefits.

Research motivation

Using LVHIIT in the cancer population shows promise as a more efficient exercise prescription. The encouraging results of this study has opens the possibility of introducing LVHIIT into rehabilitation programs. LVHIIT is a time efficient exercise modality, which could be used to increase the fitness levels in cancer survivors. The LVHIIT protocol in this study improved fitness and functional capacity and decreased waist circumference compared with CLMIT and the control group. Both LVHIIT and CLMIT improved QoL. LVHIIT may be an effective alternative to traditional exercise prescription within this population. The benefit of LVHIIT is that for selected variables it produces more pronounced results compared with CLMIT and it is short, which could entice more cancer survivors to participate in exercise as time is a barrier.

Research objectives

To determine the effectiveness of LVHIIT compared to CLMIT and a control

group and to determine if LVHIIT and CLMIT improved CVD risk and health outcomes in cancer survivors. The significance of these objectives is that this form of exercise can be used to achieve more pronounced improvements in health outcomes than the commonly prescribed CLMIT.

Research methods

The experiments and data analysis used in this study were a mix of validated methods used before within this population (6MWT, STS, DXA, hip/waist circumference) and unique protocols which have been used with other populations, but not commonly with cancer survivors (Sphygmocor). The use of the ANCOVA analysis and effect size analysis was chosen due to the robustness that this analysis provides for the data collected.

Research results

There were significant improvements in functional capacity, specifically cardiorespiratory fitness, lower-limb strength, and waist circumference in participants who completed the LVHIIT compared with the CLMIT and control groups. QoL improved in both the LVHIIT group and the CLMIT group when compared with the control group, however a greater effect was observed in the LVHIIT group. Additional research is required to fully understand the mechanisms involved in the changes identified in this study in relation to different doses of exercise. This research would be highly beneficial to assist clinicians in the optimisation of clinical exercise recommendations for cancer survivors.

Research conclusions

Present exercise guidelines for cancer survivors lack detail on the type, mode, duration and intensity of exercise necessary to achieve best outcomes. This current research was required to fill the gaps in current knowledge to further improve exercise recommendations. LVHIIT is the use of small doses of high-intensity exercise to elicit physiological responses such as improved VO₂ max and positive metabolic changes in skeletal muscle which seem greater than the commonly prescribed CLMIT. The LVHIIT physiological changes show potential for use in clinical practice in the rehabilitation of cancer survivors. At present there is limited research examining the effects of LVHIIT in improving health outcomes for cancer survivors. This study shows promise for the use of LVHIIT in the cancer population. The encouraging results opens up the possibility of introducing LVHIIT in therapy programs, as a shorter and more efficacious exercise to increase the fitness levels in cancer survivors. The benefit of LVHIIT is that for selected variables it produces more pronounced results compared with CLMIT and it is short in duration which could entice more cancer survivors to participate in exercise. This study highlights that the most commonly prescribed CLMIT may not be enough to induce clinically relevant changes in cancer survivors.

Research perspectives

The use of VO₂ max testing would be beneficial if it can be tolerated well by cancer survivors post treatment. Future research should analyse ways to measure the mechanisms involved in the changes seen in the study results and clarify how and why these changes occur.

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REFERENCES

1. **Allemani C**, Weir HK, Carreira H, Harewood R, Spika D, Wang X-S, Bannan F, Ahn JV, Johnson CJ, Bonaventure A. Global surveillance of cancer survival 1995–2009: analysis of individual data for 25 676 887 patients from 279 population-based registries

- in 67 countries (CONCORD-2). *The Lancet* 2015; **385**: 977-1010 [PMID: 25467588 DOI: 10.1016/S0140-6736(14)62038-9]
- 2 **Gallagher EJ**, LeRoith D. Obesity and Diabetes: The Increased Risk of Cancer and Cancer-Related Mortality. *Physiol Rev* 2015; **95**: 727-748 [PMID: 26084689 DOI: 10.1152/physrev.00030.2014]
- 3 **Toohy K**, Puma K, Cooke J, Semple S. Do Activity Patterns And Body Weight Change After A Cancer Diagnosis? A Retrospective Cohort Study. *IJHSR* 2016; **6**: 110-117
- 4 **Holmes MD**, Chen WY, Feskanich D, Kroenke CH, Colditz GA. Physical activity and survival after breast cancer diagnosis. *JAMA* 2005; **293**: 2479-2486 [PMID: 15914748 DOI: 10.1001/jama.293.20.2479]
- 5 **Mazzeo RS**, Cavanagh P, Evans WJ, Fiatarone M, Hagberg J, McAuley E, Startzell J. Exercise and physical activity for older adults. *Medicine and science in sports and exercise* 1998; **30**: 992-1008
- 6 **Mishra SI**. Are Exercise Programs Effective for Improving Health-Related Quality of Life Among Cancer Survivors? A Systematic Review and Meta-Analysis. *Oncol Nurs Forum* 2014; **41**: E326-E42 [PMID: 25355029 DOI: 10.1188/14.ONF.E326-E342]
- 7 **Jones LW**, Liang Y, Pituskin EN, Battaglini CL, Scott JM, Hornsby WE, Haykowsky M. Effect of exercise training on peak oxygen consumption in patients with cancer: a meta-analysis. *Oncologist* 2011; **16**: 112-120 [PMID: 21212429 DOI: 10.1634/theoncologist.2010-0197]
- 8 **Lakoski SG**, Willis BL, Barlow CE, Leonard D, Gao A, Radford NB, Farrell SW, Douglas PS, Berry JD, DeFina LF, Jones LW. Midlife Cardiorespiratory Fitness, Incident Cancer, and Survival After Cancer in Men: The Cooper Center Longitudinal Study. *JAMA Oncol* 2015; **1**: 231-237 [PMID: 26181028 DOI: 10.1001/jamaoncol.2015.0226]
- 9 **Schmitz KH**, Courneya KS, Matthews C, Demark-Wahnefried W, Galvão DA, Pinto BM, Irwin ML, Wolin KY, Segal RJ, Lucia A, Schneider CM, von Gruenigen VE, Schwartz AL; American College of Sports Medicine. American College of Sports Medicine roundtable on exercise guidelines for cancer survivors. *Med Sci Sports Exerc* 2010; **42**: 1409-1426 [PMID: 20559064 DOI: 10.1249/MSS.0b013e3181e0c112]
- 10 **Velthuis MJ**, Agasi-Idenburg SC, Aufdemkampe G, Wittink HM. The effect of physical exercise on cancer-related fatigue during cancer treatment: a meta-analysis of randomised controlled trials. *Clin Oncol (R Coll Radiol)* 2010; **22**: 208-221 [PMID: 20110159 DOI: 10.1016/j.clon.2009.12.005]
- 11 **Hayes SC**, Spence RR, Galvão DA, Newton RU. Australian Association for Exercise and Sport Science position stand: optimising cancer outcomes through exercise. *J Sci Med Sport* 2009; **12**: 428-434 [PMID: 19428291 DOI: 10.1016/j.jsams.2009.03.002]
- 12 **Donnelly JE**, Blair SN, Jakicic JM, Manore MM, Rankin JW, Smith BK; American College of Sports Medicine. American College of Sports Medicine Position Stand. Appropriate physical activity intervention strategies for weight loss and prevention of weight regain for adults. *Med Sci Sports Exerc* 2009; **41**: 459-471 [PMID: 19127177 DOI: 10.1249/MSS.0b013e3181949333]
- 13 **Gibala MJ**, Little JP, Macdonald MJ, Hawley JA. Physiological adaptations to low-volume, high-intensity interval training in health and disease. *J Physiol* 2012; **590**: 1077-1084 [PMID: 22289907 DOI: 10.1113/jphysiol.2011.224725]
- 14 **Weston KS**, Wisløff U, Coombes JS. High-intensity interval training in patients with lifestyle-induced cardiometabolic disease: a systematic review and meta-analysis. *Br J Sports Med* 2014; **48**: 1227-1234 [PMID: 24144531 DOI: 10.1136/bjsports-2013-092576]
- 15 **Boutcher SH**. High-intensity intermittent exercise and fat loss. *J Obes* 2011; **2011**: 868305 [PMID: 21113312 DOI: 10.1155/2011/868305]
- 16 **Aamot IL**, Forbord SH, Gustad K, Løckra V, Stensen A, Berg AT, Dalen H, Karlsen T, Støylen A. Home-based versus hospital-based high-intensity interval training in cardiac rehabilitation: a randomized study. *Eur J Prev Cardiol* 2014; **21**: 1070-1078 [PMID: 23613224 DOI: 10.1177/2047487313488299]
- 17 **Ulbrich AZ**, Angarten VG, Netto AS, Sties SW, Bündchen DC, de Mara LS, Cornelissen VA, de Carvalho T. Comparative effects of high intensity interval training versus moderate intensity continuous training on quality of life in patients with heart failure: study protocol for a randomized controlled trial. *Clinical Trials and Regulatory Science in Cardiology* 2016; **13**: 21-28 [DOI: 10.1016/j.ctrsc.2015.11.005]
- 18 **Courneya KS**, Friedenreich CM. Framework PEACE: an organizational model for examining physical exercise across the cancer experience. *Annals of Behavioral Medicine* 2001; **23**: 263-272 [DOI: 10.1207/S15324796ABM2304_5]
- 19 **American College of Sports M**. ACSM's guidelines for exercise testing and prescription: Lippincott Williams & Wilkins; 2013
- 20 **Urbanik G**, Plous S. Research Randomizer (Version 4.0)[Computer software]. Retrieved on June 22, 2013
- 21 **Esper P**, Mo F, Chodak G, Sinner M, Cella D, Pienta KJ. Measuring quality of life in men with prostate cancer using the functional assessment of cancer therapy-prostate instrument. *Urology* 1997; **50**: 920-928 [DOI: 10.1016/S0090-4295(97)00459-7]
- 22 **Overcash J**, Extermann M, Parr J, Perry J, Balducci L. Validity and reliability of the FACT-G scale for use in the older person with cancer. *Am J Clin Oncol* 2001; **24**: 591-596 [PMID: 11801761 DOI: 10.1097/00000421-200112000-00013]
- 23 **Bergman RN**, Stefanovski D, Buchanan TA, Sumner AE, Reynolds JC, Sebring NG, Xiang AH, Watanabe RM. A better index of body adiposity. *Obesity (Silver Spring)* 2011; **19**: 1083-1089 [PMID: 21372804 DOI: 10.1038/oby.2011.38]
- 24 **Dobbeltsteyn CJ**, Joffres MR, MacLean DR, Flowerdew G. A comparative evaluation of waist circumference, waist-to-hip ratio and body mass index as indicators of cardiovascular risk factors. The Canadian Heart Health Surveys. *Int J Obes Relat Metab Disord* 2001; **25**: 652-661 [PMID: 11360147 DOI: 10.1038/sj.ijo.0801582]
- 25 **Organization WH**. Waist circumference and waist-hip ratio: Report of a WHO expert consultation, Geneva, 8-11 December 2008. 2011
- 26 **Organization WH**. WHO STEPS surveillance manual: the WHO STEP wise approach to chronic disease risk factor surveillance. 2005
- 27 **McDonnell BJ**, Maki-Petaja KM, Munnery M, Yasmin, Wilkinson IB, Cockcroft JR, McEniery CM. Habitual exercise and blood pressure: age dependency and underlying mechanisms. *Am J Hypertens* 2013; **26**: 334-341 [PMID: 23382483 DOI: 10.1093/ajh/hps055]
- 28 **Sharma JE**, Lim R, Qasem AM, Coombes JS, Burgess MI, Franco J, Garrahy P, Wilkinson IB, Marwick TH. Validation of a generalized transfer function to noninvasively derive central blood pressure during exercise. *Hypertension* 2006; **47**: 1203-1208 [PMID: 16651459 DOI: 10.1161/01.HYP.0000223013.60612.72]
- 29 **Toohy K**, Puma KL, Arnold L, Cooke J, Yip D, Craft PS, Semple S. A pilot study examining the effects of low-volume high-intensity interval training and continuous low to moderate intensity training on quality of life, functional capacity and cardiovascular risk factors in cancer survivors. *Peer J* 2016; **4**: e2613 [PMID: 27781180 DOI: 10.7717/peerj.2613]
- 30 **Simmonds MJ**. Physical function in patients with cancer: psychometric characteristics and clinical usefulness of a physical performance test battery. *J pain and symptom management* 2002; **24**: 404-414 [DOI: 10.1016/S0885-3924(02)00502-X]
- 31 **Bohannon RW**. Test-retest reliability of the five-repetition sit-to-stand test: a systematic review of the literature involving adults. *J Strength Cond Res* 2011; **25**: 3205-3207 [PMID: 21904240 DOI: 10.1519/JSC.0b013e318234e59f]
- 32 **Cote CG**, Casanova C, Marín JM, Lopez MV, Pinto-Plata V, de Oca MM, Dordelly LJ, Nekach H, Celli BR. Validation and comparison of reference equations for the 6-min walk distance test. *Eur Respir J* 2008; **31**: 571-578 [PMID: 17989117 DOI: 10.1183/09031936.00104507]
- 33 **Enright PL**. The six-minute walk test. *Respir Care* 2003; **48**: 783-785 [PMID: 12890299]

- 34 **Schmidt K**, Vogt L, Thiel C, Jäger E, Banzer W. Validity of the six-minute walk test in cancer patients. *Int J Sports Med* 2013; **34**: 631-636 [PMID: 23444095 DOI: 10.1055/s-0032-1323746]
- 35 **Gillen JB**, Gibala MJ. Is high-intensity interval training a time-efficient exercise strategy to improve health and fitness? *Appl Physiol Nutr Metab* 2014; **39**: 409-412 [PMID: 24552392 DOI: 10.1139/apnm-2013-0187]
- 36 **Rognmo Ø**, Hetland E, Helgerud J, Hoff J, Slørdahl SA. High intensity aerobic interval exercise is superior to moderate intensity exercise for increasing aerobic capacity in patients with coronary artery disease. *Eur J Cardiovasc Prev Rehabil* 2004; **11**: 216-222 [PMID: 15179103 DOI: 10.1097/01.hjr.0000131677.96762.0c]
- 37 **Cohen J**. Statistical Power Analysis for the Behavioral Sciences (2nd ed) Hillsdale, NJ: Erlbaum; 1998
- 38 **Borg G**. Psychophysical scaling with applications in physical work and the perception of exertion. *Scand J Work Environ Health* 1990; **16** Suppl 1: 55-58 [PMID: 2345867 DOI: 10.5271/sjweh.1815]
- 39 **Borg G**, Linderholm H. Perceived exertion and pulse rate during graded exercise in various age groups. *J Internal Medicine* 1967; **181**: 194-206 [DOI: 10.1111/j.0954-6820.1967.tb12626.x]
- 40 **Galvão DA**, Newton RU. Review of exercise intervention studies in cancer patients. *J Clin Oncol* 2005; **23**: 899-909 [PMID: 15681536 DOI: 10.1200/JCO.2005.06.085]
- 41 **Mock V**, Pickett M, Ropka ME, Muscari Lin E, Stewart KJ, Rhodes VA, McDaniel R, Grimm PM, Krumm S, McCorkle R. Fatigue and quality of life outcomes of exercise during cancer treatment. *Cancer Pract* 2001; **9**: 119-127 [PMID: 11879296 DOI: 10.1046/j.1523-5394.2001.009003119.x]
- 42 **Irwin ML**, Crumley D, McTiernan A, Bernstein L, Baumgartner R, Gilliland FD, Kriska A, Ballard-Barbash R. Physical activity levels before and after a diagnosis of breast carcinoma: the Health, Eating, Activity, and Lifestyle (HEAL) study. *Cancer* 2003; **97**: 1746-1757 [PMID: 12655532 DOI: 10.1002/cncr.11227]
- 43 **Vardar-Yagli N**, Sener G, Saglam M, Calik-Kutukcu E, Arikani H, Inal-Ince D, Savci S, Altundag K, Kutluk T, Ozisik Y, Kaya EB. Associations among physical activity, comorbidity, functional capacity, peripheral muscle strength and depression in breast cancer survivors. *Asian Pac J Cancer Prev* 2015; **16**: 585-589 [PMID: 25684491 DOI: 10.7314/APJCP.2015.16.2.585]
- 44 **Kampshoff CS**, Chinapaw MJ, Brug J, Twisk JW, Schep G, Nijziel MR, van Mechelen W, Buffart LM. Randomized controlled trial of the effects of high intensity and low-to-moderate intensity exercise on physical fitness and fatigue in cancer survivors: results of the Resistance and Endurance exercise After ChemoTherapy (REACT) study. *BMC Med* 2015; **13**: 275 [PMID: 26515383 DOI: 10.1186/s12916-015-0513-2]
- 45 **Gibala MJ**, Little JP, van Essen M, Wilkin GP, Burgomaster KA, Safdar A, Raha S, Tarnopolsky MA. Short-term sprint interval versus traditional endurance training: similar initial adaptations in human skeletal muscle and exercise performance. *J Physiol* 2006; **575**: 901-911 [PMID: 16825308 DOI: 10.1113/jphysiol.2006.112094]
- 46 **Weston M**, Taylor KL, Batterham AM, Hopkins WG. Effects of low-volume high-intensity interval training (HIT) on fitness in adults: a meta-analysis of controlled and non-controlled trials. *Sports Med* 2014; **44**: 1005-1017 [PMID: 24743927 DOI: 10.1007/s40279-014-0180-z]
- 47 **Leon AS**, Connett J, Jacobs DR Jr, Rauramaa R. Leisure-time physical activity levels and risk of coronary heart disease and death. The Multiple Risk Factor Intervention Trial. *JAMA* 1987; **258**: 2388-2395 [PMID: 3669210 DOI: 10.1001/jama.1987.03400170074026]
- 48 **Laukkanen JA**, Lakka TA, Rauramaa R, Kuhanen R, Venäläinen JM, Salonen R, Salonen JT. Cardiovascular fitness as a predictor of mortality in men. *Arch Intern Med* 2001; **161**: 825-831 [PMID: 11268224 DOI: 10.1001/archinte.161.6.825]
- 49 **Schmid D**, Leitzmann M. Cardiorespiratory fitness as predictor of cancer mortality: a systematic review and meta-analysis. *Ann Oncol* 2015; **26**: 272-178 [PMID: 25009011 DOI: 10.1093/annonc/
- 50 **Schneider CM**, Repka CP, Brown JM, Lalonde TL, Dallow KT, Barlow CE, Hayward R. Demonstration of the need for cardiovascular and pulmonary normative data for cancer survivors. *Int J Sports Med* 2014; **35**: 1134-1137 [PMID: 24995960 DOI: 10.1055/s-0034-1375691]
- 51 **Zabora J**, BrintzenhofeSzoc K, Curbow B, Hooker C, Piantadosi S. The prevalence of psychological distress by cancer site. *Psychooncology* 2001; **10**: 19-28 [PMID: 11180574]
- 52 **Courneya KS**, Friedenreich CM. Physical exercise and quality of life following cancer diagnosis: a literature review. *Ann Behav Med* 1999; **21**: 171-179 [PMID: 10499138]
- 53 **Montazeri A**. Health-related quality of life in breast cancer patients: a bibliographic review of the literature from 1974 to 2007. *J Exp Clin Cancer Res* 2008; **27**: 32 [PMID: 18759983 DOI: 10.1186/1756-9966-27-32]
- 54 **Karvinen KH**, Esposito D, Raedeke TD, Vick J, Walker PR. Effect of an exercise training intervention with resistance bands on blood cell counts during chemotherapy for lung cancer: a pilot randomized controlled trial. *Springerplus* 2014; **3**: 15 [PMID: 24683529 DOI: 10.1186/2193-1801-3-15]
- 55 **Vozarova B**, Weyer C, Lindsay RS, Pratley RE, Bogardus C, Tataranni PA. High white blood cell count is associated with a worsening of insulin sensitivity and predicts the development of type 2 diabetes. *Diabetes* 2002; **51**: 455-461 [PMID: 11812755 DOI: 10.2337/diabetes.51.2.455]
- 56 **Saligan LN**, Olson K, Filler K, Larkin D, Cramp F, Yennurajalingam S, Escalante CP, del Giglio A, Kober KM, Kamath J, Palesh O, Mustian K; Multinational Association of Supportive Care in Cancer Fatigue Study Group-Biomarker Working Group. The biology of cancer-related fatigue: a review of the literature. *Support Care Cancer* 2015; **23**: 2461-2478 [PMID: 25975676 DOI: 10.1007/s00520-015-2763-0]
- 57 **LaVoy EC**, Fagundes CP, Dantzer R. Exercise, inflammation, and fatigue in cancer survivors. *Exerc Immunol Rev* 2016; **22**: 82-93 [PMID: 26853557]
- 58 **Wieser V**, Moschen AR, Tilg H. Inflammation, cytokines and insulin resistance: a clinical perspective. *Arch Immunol Ther Exp (Warsz)* 2013; **61**: 119-125 [PMID: 23307037 DOI: 10.1007/s00005-012-0210-1]
- 59 **Giovannucci E**. Insulin and colon cancer. *Cancer Causes & Control* 1995; **6**: 164-179 [DOI: 10.1007/BF00052777]
- 60 **Renahan AG**, Zwahlen M, Minder C, T O'Dwyer S, Shalet SM, Egger M. Insulin-like growth factor (IGF)-I, IGF binding protein-3, and cancer risk: systematic review and meta-regression analysis. *The Lancet* 2004; **363**: 1346-1353 [DOI: 10.1016/S0140-6736(04)16044-3]
- 61 **Moreo A**, Vallerio P, Ricotta R, Stucchi M, Pozzi M, Musca F, Meani P, Maloberti A, Facchetti R, Di Bella S, Giganti MO, Sartore-Bianchi A, Siena S, Mancia G, Giannattasio C. Effects of Cancer Therapy Targeting Vascular Endothelial Growth Factor Receptor on Central Blood Pressure and Cardiovascular System. *Am J Hypertens* 2016; **29**: 158-162 [PMID: 26031304]
- 62 **Gorriani C**, Harris IS, Mak TW. Modulation of oxidative stress as an anticancer strategy. *Nat Rev Drug Discov* 2013; **12**: 931-947 [PMID: 24287781 DOI: 10.1038/nrd4002]
- 63 **Reuter S**, Gupta SC, Chaturvedi MM, Aggarwal BB. Oxidative stress, inflammation, and cancer: how are they linked? *Free Radic Biol Med* 2010; **49**: 1603-1616 [PMID: 20840865 DOI: 10.1016/j.freeradbiomed.2010.09.006]
- 64 **Mozos I**, Borzak G, Caraba A, Mihaescu R. Arterial stiffness in hematologic malignancies. *Onco Targets Ther* 2017; **10**: 1381-1388 [PMID: 28424554 DOI: 10.2147/OTT.S126852]
- 65 **Vallerio P**, Sarno L, Stucchi M, Musca F, Casadei F, Maloberti A, Lestuzzi C, Mancia G, Moreo A, Palazzi M, Giannattasio C. Long-Term Effects of Radiotherapy on Arterial Stiffness in Breast Cancer Women. *Am J Cardiol* 2016; **118**: 771-776 [PMID: 27392510 DOI: 10.1016/j.amjcard.2016.06.001]
- 66 **Truskey GA**, Fernandez CE. Tissue-engineered blood vessels as

- 67 promising tools for testing drug toxicity. Taylor & Francis; 2015
- 67 **Lenihan DJ**, Cardinale DM. Late cardiac effects of cancer treatment. *J Clin Oncol* 2012; **30**: 3657-3664 [PMID: 23008297 DOI: 10.1200/JCO.2012.45.2938]
- 68 **Rogers LQ**, Courneya KS, Verhulst S, Markwell S, Lanzotti V, Shah P. Exercise barrier and task self-efficacy in breast cancer patients during treatment. *Support Care Cancer* 2006; **14**: 84-90 [PMID: 16007455 DOI: 10.1007/s00520-005-0851-2]

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Observational Study

Palliative surgery for Krukenberg tumors – 12-year experience and review of the literature

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Abstract

AIM

To determine the clinical characteristics of patients undergoing palliative surgery for Krukenberg tumors, including disease presentation, outcomes, and prognostic factors.

METHODS

This was a retrospective clinical study of all patients who underwent palliative surgery for Krukenberg tumors between January 2004 and December 2015. Patient information was obtained from inpatient and outpatient case notes as well as the hospital electronic records. Patients who underwent potentially curative resection, and patients with Krukenberg tumors who did not undergo surgery were also excluded from the study. Palliative surgery was defined as those performed for either alleviation of symptoms or for asymptomatic patients for whom surgical removal of the tumors were deemed necessary following a multidisciplinary consensus. Tumors were diagnosed pre-operatively by computed tomography

scans and all had histologic confirmation of the surgical specimens.

RESULTS

Over the study duration, 38 female patients underwent palliative surgery for Krukenberg tumors at our institution. Mean age was 54.2 ± 11.7 years. The colon was the most frequent primary source of metastases ($n = 21$) followed by the stomach ($n = 4$). Prophylactic palliative surgery was performed for eight (21.1%) asymptomatic patients. Median post-operative length of stay was 8 d (IQR 6-12 d). Five patients (13.2%) experienced post-operative complications, although high grade morbidity was only seen in one patient (2.6%). Median overall survival from surgery was 17 mo (95%CI: 12.1-21.9) at a median follow-up duration of 12 mo (IQR 8-17 mo). The median survival was shorter for patients who underwent emergency surgery, younger patients, those with a colorectal primary, larger tumors, or synchronous peritoneal or hepatic metastases.

CONCLUSION

Palliative surgery for Krukenberg tumors can be performed safely with acceptable complication rates. Bilateral oophorectomy should be performed to prevent the risk of symptomatic contralateral tumors.

Key words: Krukenberg tumor; Palliative surgery

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Core tip: Krukenberg tumors represent metastases to the ovary and convey a poor prognosis. By reporting our 12-year experience with palliative surgery for Krukenberg tumors and discussing the existing literature, we hope to shed light and establish best practices on the approach to management of the disease. Palliative surgery for patients with Krukenberg tumors can be performed safely in an experienced unit with acceptable complication rates. Where possible, bilateral oophorectomy should be performed to obviate the risk of the contralateral ovary developing symptomatic tumors. Appropriate selection by a multidisciplinary consensus is essential for asymptomatic patients who may benefit from prophylactic surgery.

Seow-En I, Hwang G, Tan GHC, Ho LML, Teo MCC. Palliative surgery for Krukenberg tumors – 12-year experience and review of the literature. *World J Clin Oncol* 2018; 9(1): 13-19 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i1/13.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i1.13>

INTRODUCTION

A Krukenberg tumor is a rare ovarian tumor which has metastasized from a primary site, accounting for 1%-2% of all tumors of the ovary^[1]. The stomach was previously described to be the most common primary site, followed

by the colon, appendix and breast^[1]. Recent literature reveals an increased incidence of tumors originating from the colon^[2]. Compared with primary ovarian tumors, the prognosis of patients with Krukenberg tumors is bleak, even with metastatic disease confined to the ovaries.

Various studies have been published over the past decade with regards to the prognostic factors and outcomes of surgery for Krukenberg tumors. Metastectomy, or surgical removal of one or both involved ovaries, has been found to improve overall survival^[3-5]. Cytoreductive surgery incorporating Krukenberg tumor removal has also been found to have a beneficial effect, with a 7% overall 5-year survival^[6]. However, extensive cytoreductive surgery may have significant associated morbidity and mortality. To our knowledge, no previous study focuses exclusively on the group of patients for whom surgery is considered palliative.

Here we discuss the existing literature and report our experience with palliative surgery for Krukenberg tumors, including clinical characteristics of patients, disease presentation, surgical outcomes, safety, and prognostic factors.

MATERIALS AND METHODS

Data was collected for all patients who underwent surgery for Krukenberg tumors between January 2004 and December 2015. Patient information was obtained from inpatient and outpatient case notes as well as the hospital electronic records. Protocols were approved by hospital Institutional Review Board and the data was analyzed retrospectively. Patients with ovarian metastases who underwent potentially curative resection were excluded from the study. Patients with Krukenberg tumors who did not undergo surgery were also excluded from the study.

Over this period, surgery for Krukenberg tumors was performed by experienced surgeons from 3 departments within the same institution; General Surgery, Surgical Oncology, as well as Obstetrics and Gynaecology. The diagnosis of metastatic disease was made pre-operatively at a multidisciplinary tumor board meeting in 37 cases and post-operatively in one patient who underwent emergency surgery. Most Krukenberg tumors were diagnosed pre-operatively by computed tomography (CT) scans and all had histologic confirmation based on the pathological evaluation of the surgical specimens.

Palliative surgery in our series was defined as those performed for either alleviation of symptoms caused by the Krukenberg tumor (e.g., abdominal pain, distension or obstruction), or for asymptomatic patients for whom surgical removal of the tumors were deemed necessary following a multidisciplinary consensus.

Statistical analyses were performed using SPSS version 20.0 (SPSS Inc., Chicago, IL, United States). Survival was defined as the time from the date of surgery for the Krukenberg tumor to the date of death. Patients in whom the event of death had not occurred

Table 1 Patient characteristics *n* (%)

Variable	Frequency
Age	
≥ 50	25 (65.8)
< 50	13 (34.2)
Race	
Chinese	33 (86.8)
Malay	3 (7.9)
Others	2 (5.3)
Presenting symptoms	
Pain	15 (39.5)
Distension	9 (23.7)
Intestinal obstruction	3 (7.9)
Obstructive symptoms	3 (7.9)
Asymptomatic	8 (21.1)
ASA score at KT surgery	
2	30 (78.9)
3	8 (21.1)

SD: Standard deviation; ASA: American Society of Anesthesiologists; KT: Krukenberg tumour.

at the time of this study were censored. Survival rates were calculated using the Kaplan-Meier method and the differences in median survival times between groups assessed using the log-rank test. Cox proportional hazards model was used to determine the hazard ratios of variables affecting survival. $P < 0.05$ was taken as significant.

RESULTS

Patient and disease characteristics

Between January 2004 and December 2015, 38 female patients underwent palliative surgery for Krukenberg tumors at our institution. Patient and disease characteristics are summarised in Tables 1 and 2 respectively. Mean patient age was 54.2 ± 11.7 years. Almost 40% of the patients with Krukenberg tumors presented with abdominal pain or had pain as their most significant symptom. Median tumor size was 11.8 cm [Interquartile range (IQR) 7.8–16.0 cm]. Apart from the ovaries, all patients had at least one other site of metastases, mostly commonly the peritoneum or liver. The colon was the most frequent primary source of metastases followed by the stomach. In one patient, the primary tumor was not identified despite extensive investigation. The Krukenberg tumor was diagnosed during the same setting as the primary in just under half of the patients. Amongst the patients who had ovarian metastases detected metachronously, the median duration of diagnosis of the Krukenberg tumor from the primary was 19.5 mo (IQR 9–24 mo).

Twenty-six patients (68.4%) had undergone chemotherapy prior to Krukenberg tumor surgery and of these, 19 continued post-operative palliative chemotherapy. A total of 27 patients (71.1%) received chemotherapy post-operatively, including eight patients who commenced chemotherapy for the first time after surgery. Four patients (10.5%) had never received chemotherapy.

Table 2 Disease characteristics *n* (%)

Variable	Frequency
Site of primary	
Stomach	4 (10.5)
Colon	21 (55.3)
Rectum	1 (2.6)
Pancreas	3 (7.9)
Breast	2 (5.3)
Peritoneum	2 (5.3)
Appendix	1 (2.6)
Endometrium	1 (2.6)
Bladder	1 (2.6)
Lung	1 (2.6)
Unknown	1 (2.6)
Previous surgery for primary	
Yes	17 (44.7)
No	21 (55.3)
Detection of KT and primary	
Synchronous	18 (47.4)
Metachronous	20 (52.6)
KT size	
≥ 12 cm	19 (50.0)
< 12 cm	19 (50.0)
Ovaries involved	
Unilateral	20 (52.6)
Bilateral	18 (47.4)
Peritoneal metastases	
Yes	21 (55.3)
No	17 (44.7)
Hepatic metastases	
Yes	16 (42.1)
No	22 (57.9)

KT: Krukenberg tumour; SD: Standard deviation.

Surgical characteristics

Surgical procedures performed were; a unilateral or bilateral salphingo-oophorectomy alone, or a total hysterectomy in addition to a bilateral salphingo-oophorectomy (Table 3). Two other patients underwent additional surgical procedures during the same sitting as the Krukenberg tumor surgery; one patient had a small bowel resection while the other had a right hemicolectomy performed. Prophylactic palliative surgery was performed for eight asymptomatic patients; two were performed for histological confirmation of metastases to guide subsequent treatment, two were for progression of the Krukenberg tumors while on chemotherapy despite adequate extra-ovarian disease control, and four were prophylactic for sizeable tumors which would have likely caused symptoms during the patient's lifetime. Five patients underwent emergent surgery; two for ruptured Krukenberg tumors, the third for a patient with fever and abdominal pain whose pre-operative CT scan was incorrectly reported as tubo-ovarian abscesses, the fourth for ovarian torsion, and in the last patient for acute intestinal obstruction secondary to compression of the bowel from the Krukenberg tumor.

Peri-operative morbidity

Median post-operative length of stay was 8 d (IQR 6–12 d). The majority of patients had an uneventful recovery period (86.8%). Five patients (13.2%) suffered complications; two Clavien-Dindo grade I, two grade

Table 3 Surgery characteristics *n* (%)

Variable	Frequency
Urgency	
Elective	33 (86.8)
Emergency	5 (13.2)
Pre-KT surgery chemo	
Yes	26 (68.4)
No	12 (31.6)
Surgery	
USO	10 (26.3)
BSO	12 (31.6)
THBSO	16 (42.1)
Post-op clavian-dindo score	
Grade 0	33 (86.8)
Grade I	2 (5.3)
Grade II	2 (5.3)
Grade IIIa	1 (2.6)

SD: Standard deviation; ASA: American Society of Anesthesiologists; KT: Krukenberg tumour; USO: Unilateral salphingo-oopherectomy; BSO: Bilateral salphingo-oopherectomy; THBSO: Total hysterectomy and bilateral salphingo-oopherectomy.

Table 4 Univariate analyses of variables affecting survival

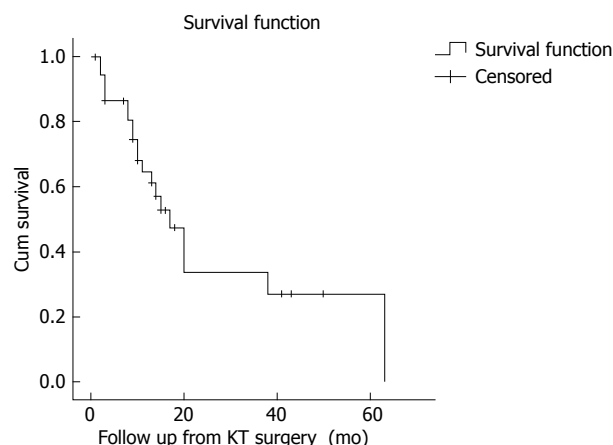
Variable	HR (95%CI)	P-value
Age < 50	2.169 (0.876-5.369)	0.094
ASA score 3	2.830 (0.873-9.171)	0.083
Colorectal primary	1.089 (0.424-2.795)	0.860
Previous surgery for primary	0.894 (0.359-2.226)	0.810
Synchronous lesion	1.240 (0.497-3.096)	0.645
Size ≥ 12 cm	1.106 (0.424-2.884)	0.837
Bilateral disease	1.473 (0.597-3.636)	0.400
Peritoneal metastases	1.337 (0.509-3.511)	0.555
Hepatic metastases	1.055 (0.419-2.656)	0.909
Emergency surgery	3.382 (1.051-10.89)	0.027
Pre-op chemotherapy	1.575 (0.521-4.758)	0.421
Serum CA 125 > 35 kU/L	0.750 (0.187-3.004)	0.684
Serum CEA ≥ 30 µg/L	0.785 (0.234-2.626) ¹	0.694 ^a

¹Among patients with a colorectal primary only. HR: Hazard ratio; CI: Confidence interval; ASA: American Society of Anesthesiologists; CEA: Carcinoembryonic antigen.

II, and one grade IIIa (Table 3). Grade I complications included a urinary tract infection requiring antibiotics and supraventricular tachycardia which aborted with medication. Both grade II complications were cases of post-operative ileus requiring total parenteral nutrition. In these two patients, peritoneal carcinomatosis was thought to be a likely contributory factor. The grade IIIa complication was a post-operative intraabdominal collection requiring radiological guided drainage. This patient had undergone emergency surgery for a large ruptured Krukenberg tumor.

Survival

The median follow-up duration was 12 mo (IQR 8-17 mo). Two patients were lost to follow-up and one patient returned to her country of origin for continued care shortly after discharge. Twenty patients (52.6%) died during the follow-up period. Median overall survival

**Figure 1** Kaplan-Meier survival analysis of 38 patients who underwent palliative surgery for Krukenberg tumours.

(OS) from Krukenberg tumor surgery was 17 mo (95%CI: 12.1-21.9) (Figure 1). Based on univariate analysis (Table 4), only emergency surgery was associated with a significantly worse survival outcome. The median survival for patients with a colorectal primary compared to all other origins was 15 mo (95%CI: 10.5-19.5) and 38 mo respectively (95%CI: 5.8-70.2), $P = 0.845$. The median OS was also shorter for patients younger than 50 years old (13 mo, 95%CI: 8.0-18.0 vs 20 mo, 95%CI: 16.4-23.6), patients with a pre-operative American Society of Anaesthesiologists (ASA) score of 3 vs 2 (8 mo, 95%CI: 4.5-12.1 vs 29 mo, 95%CI: 18.6-40.1), larger tumors ≥ 12 cm (14 mo, 95%CI: 9.4-18.6 vs 20 mo, 95%CI: 14.8-25.2), those with peritoneal metastases (17 mo, 95%CI: 11.3-22.7 vs 20 mo, 95%CI: 9.7-30.3) and those with hepatic metastases (14 mo, 95%CI: 8.4-19.6 vs 17 mo, 95%CI: 9.8-24.2).

Pre-operative serum tumor marker level did not correlate with OS. Serum CA 125 level was performed in 21 patients (55.3%) before Krukenberg tumor surgery, for which median CA 125 level was 42.4 kU/L (IQR 14.1-206.5). Pre-operative serum carcinoembryonic antigen (CEA) testing was done in 30 patients (78.9%), including 20 out of 21 patients who had a colorectal primary. For these 20 patients, the median CEA level was 30.3 µg/L (IQR 10.8-144.0). Using a cut-off value of 35 and 30 for CA 125 and CEA respectively (Table 4), it appeared that the pre-operative values of these tumor markers did not reflect patient prognosis. These values were chosen based on the normal reference range of CA 125 (0-35 kU/L) and the median CEA level amongst patients with a colorectal primary in our study.

DISCUSSION

The eponymous Krukenberg tumor was first described in 1896 by the German doctor Friedrich Krukenberg (1871-1946). Representing advanced metastatic disease, this entity clearly conveys a poor prognosis. Gastric cancer was previously reported to account for

the majority of primary tumors responsible^[7]. However, recent literature suggests an increased frequency of Krukenberg tumors with a colorectal origin^[2]. In our study, more than half of the patients had a colorectal primary compared to only 10% with a gastric primary. This is likely related to the increasing frequency of colorectal cancer locally as well as in other developed countries. In Singapore, the age-standardized incidence rate of colorectal cancer is one of the highest among ethnic Chinese populations in the world^[8]. With the longer survival generally seen in patients with metastatic colorectal cancer as compared with that of gastric cancer, the option of pursuing palliative surgery in these patients may be seemingly more attractive.

Many aspects of Krukenberg tumors still remain controversial, including the mechanism of metastases or even the pathologic tumor characteristics used for diagnosis. The current criteria used by the World Health Organisation, established in 1973 by Serov and Scully^[9], may not adequately reflect the complexity of the tumor considering the multiple different primary sites possible. Treatment generally consists of surgery, chemotherapy or radiotherapy but guidelines concerning treatment of choice and appropriate timing of intervention have yet to be established.

Over the past decade, a number of retrospective studies have attempted to determine the prognostic factors for patients with Krukenberg tumors. Current evidence suggests a survival benefit for patients who undergo metastasectomy, compared to those without surgery. A recent multivariate analysis of 128 patients with Krukenberg tumors (58 colorectal, 41 gastric origin) showed that synchronous tumors, pelvic invasion and ascites were independent factors predicting for a poorer overall survival^[10]. Other factors found to negatively influence overall prognosis include R1 or suboptimal resection^[3], metastatic disease beyond the ovaries^[11], tumors of gastric origin compared to colorectal origin^[2], and patients with poorer Karnofsky performance status scores^[4]. For gastric cancer, metastasectomy in addition to chemotherapy was also found to improve survival compared to palliative chemotherapy alone^[12]. For colorectal cancers, ovarian metastases have been shown to be less responsive to chemotherapy compared to extra-ovarian sites^[13]. Surgical resection was therefore recommended for these "metastatic sanctuaries" even in the palliative setting, as they would often progress and result in symptoms while on chemotherapy. In a separate study comparing 83 patients with ovarian metastases from a colorectal primary who underwent an oophorectomy vs 47 historical controls who did not undergo surgery, metastasectomy conferred a significantly longer OS at 20.8 mo vs 10.9 mo^[14]. In terms of overall survival, surgery for patients with a colorectal primary may confer a greater advantage over that for a gastric primary in view of less aggressive tumor biology.

In contrast to the usually straightforward oophorectomy, the operative circumstances for patients with Krukenberg tumors may be more challenging. These

patients often have undergone previous surgeries with resultant intraabdominal adhesions, or have synchronous pelvic peritoneal disease. Therefore, while the benefit of potentially curative metastasectomy is clear, clinicians may be hesitant to subject patients to palliative surgery, particularly if the patient is asymptomatic or in a debilitated state. In our series, five patients (13.2%) experienced post-operative complications, although high grade morbidity was only seen in one patient (2.6%). Both patients with ileus resumed enteral feeding within two weeks and were discharged well. The 30-d mortality rate was zero and median length of stay was about a week.

Close to half of all patients were found to have bilateral ovarian disease. Of these, the diagnosis of contralateral ovarian involvement was not apparent on pre-operative imaging and only established following surgery and histological examination of the resected specimen in a quarter of the cases ($n = 4$). Even if not synchronously affected there is a high chance of subsequent contralateral ovary involvement resulting in symptoms. We therefore recommend that bilateral ovarian resection be routine if surgery is to be performed for palliation.

Following our analyses, emergency surgery was the only factor that was found to significantly reduce survival prognosis. Although patients who were less than 50 years old, had a higher ASA score, with tumor size larger than 12 cm, or had hepatic or peritoneal metastases tended to have a shorter median survival, this was not found to be statistically significant. Surprisingly, tumors with a colorectal primary tended to fare worse than tumors of all other origins with a median overall survival of 15 mo vs 38 mo. Meaningful comparisons could not be performed for patients with a gastric primary in view of the small sample size ($n = 4$) with one patient defaulting follow-up and the other continuing further management overseas. While a single study found that a pre-operative CA 125 level of more than 75 kU/L significantly correlated to poorer survival^[15], in our series no association could be found with this value nor the reference range cut-off. Pre- and post-operative trending of CA 125 levels therefore remains of questionable utility. CEA levels do not appear to prognosticate survival in patients with a colorectal primary, but may serve as an adjunct to imaging for reflecting disease response to chemotherapy.

Over a fifth of the patients in our series were asymptomatic at the time of surgery, all of whom had uncomplicated post-surgery recovery periods. Appropriate patient selection for prophylactic surgery is essential, with patient fitness and prognosis along with tumor size, location and anticipated development of symptoms being key considerations. With emergency surgery conferring a significantly worse outcome, the role of prophylactic surgery should certainly be explored further. The possibility of Krukenberg tumors being chemo-resistant "metastatic sanctuaries" was also evident in two asymptomatic patients in our study who

had progression of the ovarian tumors despite good systemic control of disease elsewhere.

In conclusion, Palliative surgery for patients with Krukenberg tumors can be performed safely in an experienced unit with acceptable complication rates. The decision to proceed with metastasectomy is influenced by several factors including the presence of symptoms, synchronous disease, and tumor response to chemotherapy, and should be made as part of a multidisciplinary team consensus. Where possible, bilateral oophorectomy should be performed to obviate the significant risk of symptomatic contralateral ovarian involvement. Tumor markers need not be routinely trended peri-operatively. Proper selection is essential for asymptomatic patients who may benefit from prophylactic surgery. Further studies can be done to determine if symptom-free survival can be prolonged or quality of life improved with palliative surgery.

ARTICLE HIGHLIGHTS

Research background

A Krukenberg tumor is a rare ovarian tumor which has metastasized from a primary site, accounting for 1%-2% of all tumors of the ovary, and conveys a poor prognosis even with disease confirmed to the ovaries. A handful of studies have been published over the past decade with regards to the prognostic factors and outcomes of surgery for Krukenberg tumors, but no previous study focuses exclusively on the group of patients for whom surgery is considered palliative.

Research motivation

Many aspects of Krukenberg tumors still remain controversial, and guidelines concerning treatment of choice and appropriate timing of intervention have yet to be established. We report our experience with palliative resection of Krukenberg tumors over the past 12 years and discuss the existing literature, so as to shed light and establish best practices on the approach to management of the disease.

Research objectives

We aimed to determine the clinical characteristics of patients undergoing palliative surgery for Krukenberg tumors, including disease presentation, outcomes, and prognostic factors.

Research methods

This was a retrospective clinical study of all patients who underwent palliative surgery for Krukenberg tumors between January 2004 and December 2015. Patient information was obtained from inpatient and outpatient case notes as well as the hospital electronic records. Patients who underwent potentially curative resection, and patients with Krukenberg tumors who did not undergo surgery were also excluded from the study. Palliative surgery was defined as those performed for either alleviation of symptoms or for asymptomatic patients for whom surgical removal of the tumors were deemed necessary following a multidisciplinary consensus. Tumors were diagnosed pre-operatively by computed tomography scans and all had histologic confirmation of the surgical specimens.

Research results

Over the study duration, 38 female patients underwent palliative surgery for Krukenberg tumors at our institution. Mean age was 54.2 ± 11.7 years. The colon was the most frequent primary source of metastases ($n = 21$) followed by the stomach ($n = 4$). Prophylactic palliative surgery was performed for eight (21.1%) asymptomatic patients. Median post-operative length of stay was 8 d (IQR 6-12 d). Five patients (13.2%) experienced post-operative complications, although high grade morbidity was only seen in one patient (2.6%). Median

overall survival from surgery was 17 mo (95%CI: 12.1-21.9) at a median follow-up duration of 12 mo (IQR 8-17 mo). The median survival was shorter for patients who underwent emergency surgery, younger patients, those with a colorectal primary, larger tumors, or synchronous peritoneal or hepatic metastases.

Research conclusions

Palliative surgery for patients with Krukenberg tumors can be performed safely in an experienced unit with acceptable complication rates. The decision to proceed with metastasectomy is influenced by several factors including the presence of symptoms, synchronous disease, and tumor response to chemotherapy, and should be made as part of a multidisciplinary team consensus. Where possible, bilateral oophorectomy should be performed to obviate the significant risk of symptomatic contralateral ovarian involvement. Tumor markers need not be routinely trended peri-operatively. Proper selection is essential for asymptomatic patients who may benefit from prophylactic surgery.

Research perspectives

Further studies can be done to determine if symptom-free survival can be prolonged or quality of life improved with palliative surgery.

REFERENCES

- 1 Al-Agha OM, Nicastri AD. An in-depth look at Krukenberg tumor: an overview. *Arch Pathol Lab Med* 2006; **130**: 1725-1730 [PMID: 17076540]
- 2 Jeung YJ, Ok HJ, Kim WG, Kim SH, Lee TH. Krukenberg tumors of gastric origin versus colorectal origin. *Obstet Gynecol Sci* 2015; **58**: 32-39 [PMID: 25629016 DOI: 10.5468/ogs.2015.58.1.32]
- 3 Cheong JH, Hyung WJ, Chen J, Kim J, Choi SH, Noh SH. Surgical management and outcome of metachronous Krukenberg tumors from gastric cancer. *J Surg Oncol* 2004; **87**: 39-45 [PMID: 15221918 DOI: 10.1002/jso.20072]
- 4 Jiang R, Tang J, Cheng X, Zang RY. Surgical treatment for patients with different origins of Krukenberg tumors: outcomes and prognostic factors. *Eur J Surg Oncol* 2009; **35**: 92-97 [PMID: 18632244 DOI: 10.1016/j.ejso.2008.05.006]
- 5 Lu LC, Shao YY, Hsu CH, Hsu C, Cheng WF, Lin YL, Cheng AL, Yeh KH. Metastasectomy of Krukenberg tumors may be associated with survival benefits in patients with metastatic gastric cancer. *Anticancer Res* 2012; **32**: 3397-3401 [PMID: 22843921]
- 6 Kim WY, Kim TJ, Kim SE, Lee JW, Lee JH, Kim BG, Bae DS. The role of cytoreductive surgery for non-genital tract metastatic tumors to the ovaries. *Eur J Obstet Gynecol Reprod Biol* 2010; **149**: 97-101 [PMID: 20018420 DOI: 10.1016/j.ejogrb.2009.11.011]
- 7 Yada-Hashimoto N, Yamamoto T, Kamiura S, Seino H, Ohira H, Sawai K, Kimura T, Saji F. Metastatic ovarian tumors: a review of 64 cases. *Gynecol Oncol* 2003; **89**: 314-317 [PMID: 12713997 DOI: 10.1016/S0090-8258(03)00075-1]
- 8 Singapore Cancer Registry report. Cancer Incidence and Mortality 2003-2012 and Selected Trends 1973-2012 in Singapore. Ministry of Health Singapore 2015
- 9 Serov SF, Scully RE. International Histological Classification of Tumours. Vol. 9. Geneva: WHO; 1973. Histological typing of ovarian tumors 2015
- 10 Wu F, Zhao X, Mi B, Feng LU, Yuan NA, Lei F, Li M, Zhao X. Clinical characteristics and prognostic analysis of Krukenberg tumor. *Mol Clin Oncol* 2015; **3**: 1323-1328 [PMID: 26807242 DOI: 10.3892/mco.2015.634]
- 11 Kim HK, Heo DS, Bang YJ, Kim NK. Prognostic factors of Krukenberg's tumor. *Gynecol Oncol* 2001; **82**: 105-109 [PMID: 11426970 DOI: 10.1006/gyno.2001.6210]
- 12 Cho JH, Lim JY, Choi AR, Choi SM, Kim JW, Choi SH, Cho JY. Comparison of Surgery Plus Chemotherapy and Palliative Chemotherapy Alone for Advanced Gastric Cancer with Krukenberg Tumor. *Cancer Res Treat* 2015; **47**: 697-705 [PMID: 25648093 DOI: 10.4143/crt.2013.175]
- 13 Goéré D, Daveau C, Elias D, Boige V, Tomasic G, Bonnet S,

- Pocard M, Dromain C, Ducreux M, Lasser P, Malka D. The differential response to chemotherapy of ovarian metastases from colorectal carcinoma. *Eur J Surg Oncol* 2008; **34**: 1335-1339 [PMID: 18455357 DOI: 10.1016/j.ejso.2008.03.010]
- 14 **Lee SJ**, Lee J, Lim HY, Kang WK, Choi CH, Lee JW, Kim TJ, Kim BG, Bae DS, Cho YB, Kim HC, Yun SH, Lee WY, Chun HK, Park YS. Survival benefit from ovarian metastectomy in colorectal cancer patients with ovarian metastasis: a retrospective analysis. *Cancer Chemother Pharmacol* 2010; **66**: 229-235 [PMID: 19820936 DOI: 10.1007/s00280-009-1150-2]
- 15 **Kikkawa F**, Shibata K, Ino K, Nomura S, Kajiyama H, Suzuki T, Kawai M, Mizutani S. Preoperative findings in non-gynecologic carcinomas metastasizing to the ovaries. *Gynecol Obstet Invest* 2002; **54**: 221-227 [PMID: 12592066 DOI: 10.1159/000068388]

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Yttrium-90 microsphere selective internal radiation therapy for liver metastases following systemic chemotherapy and surgical resection for metastatic adrenocortical carcinoma

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Abstract

Adrenocortical carcinoma (ACC) is a rare malignancy with generally poor outcomes and limited treatment options. While surgical resection can be curative for early local disease, most patients present with advanced ACC owing to nonspecific symptoms. For those patients, treatment options include systemic chemotherapy and locoregional therapies including radiofrequency ablation and transarterial chemoembolization. We present the first reported case of utilizing yttrium-90 microsphere selective internal radiation therapy (SIRT) in combination with first line EDP-M (Etoposide, Doxorubicin, Cisplatin, Mitotane) chemotherapy and debulking surgical primary tumor resection for treatment of metastatic ACC. Stable complete radiologic response has been maintained after twelve months with resolution of clinical symptoms. These findings prompt the need for further consideration and studies to elucidate the role of SIRT in combination with systemic and surgical treatment for metastatic ACC.

Key words: Adrenocortical carcinoma; Hepatic metastases; Radioembolization; Yttrium-90

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Core tip: Adrenocortical carcinoma (ACC) is a rare malignancy with generally poor outcomes and limited treatment options. Approximately 70% of ACC patients have unresectable stage III or IV disease on initial presentation. Yttrium-90 microsphere selective internal radiation therapy was applied toward hepatic metastases for a patient with metastatic ACC in combination with first line chemotherapy and debulking surgical primary tumor resection. A stable complete radiologic response has been maintained for twelve months with resolution of clinical symptoms.

Makary MS, Krishner LS, Wuthrick EJ, Bloomston MP, Dowell JD. Yttrium-90 microsphere selective internal radiation therapy for liver metastases following systemic chemotherapy and surgical resection for metastatic adrenocortical carcinoma. *World J Clin Oncol* 2018; 9(1): 20-25 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i1/20.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i1.20>

INTRODUCTION

Adrenocortical carcinoma (ACC) is a rare malignancy that constitutes less than 5% of all adrenal incidentalomas and has an estimated annual incidence of 1-2 cases per million people^[1-3]. ACC carries a poor prognosis with an overall 5-year survival of 16%-44%^[4-6]. Surgical resection is the treatment of choice with those presenting with only local disease. However, most patients present with advanced disease and are not surgical candidates, and 80% of even those patients with local disease treated surgically develop local or distant recurrence following complete resection^[7]. Indeed, given the nonspecific ACC symptoms, approximately 70% of ACC patients have unresectable stage III or IV disease on initial presentation^[8]. In those cases, treatment options include mitotane, systemic chemotherapy, external beam radiotherapy, radiofrequency ablation (RFA), transarterial chemoembolization (TACE), or a combination of these therapies. In this case study, we present a complete radiologic and clinical response to yttrium-90 (Y-90) microsphere selective internal radiation therapy (SIRT) treatment of hepatic metastases in conjunction with surgery and chemotherapy for treatment of a patient with advanced disease. A critical review of the current therapies and treatment strategies is also presented.

CASE REPORT

A 60-year-old female patient with a past medical history significant only for poorly controlled hypertension despite

medical therapy presented with a large left adrenal mass incidentally discovered on MRA during workup for renal artery stenosis. Abdominal MR imaging revealed a heterogeneous, partially necrotic left adrenal mass measuring 5.6 cm × 8.9 cm as well as multiple bilobar enhancing liver metastases. Histologic (Figure 1A) and immunologic evaluation of a biopsied hepatic lesion showed metastatic carcinoma positive for inhibin (Figure 1B), melan-A (Figure 1C), and CKAE1/3 (Figure 1D) with an immunohistochemical staining profile consistent with metastatic ACC. Serology values revealed normal urine cortisol [14.1 mcg/24 h (normal range: 3-22)], low adrenocorticotrophic hormone (ACTH) [5.1 pg/mL (normal range: 9-50)], normal plasma metanephrines [0.20 nmol/L (normal range < 0.50)] and high aldosterone [38.8 ng/dL (normal range: 1-16)].

The Berruti chemotherapy regimen of Etoposide, Doxorubicin and Cisplatin plus Mitotane (EDP-M) was immediately initiated^[9] for a total of five 28-d cycles. Subsequent abdominal MRI and PET/CT evaluation demonstrated overall improvement in metastatic disease burden with interval decrease in size of the left adrenal mass. Given the response to chemotherapy and following a multidisciplinary tumor board discussion, decision was made to pursue an open adrenalectomy debulking surgery two months following chemotherapy completion to reduce the hormonal load for clinical symptomatic control. Intraoperatively, the entire adrenal mass was resected *en bloc* without difficulty. Adjacent paraaortic, omental, peripancreatic thickened pearly lymph nodes were resected with a clear periadrenal margin obtained. The adrenal vein and artery were identified with no tumoral involvement visualized. The patient had an uneventful post-operative hospital course and was discharged in 5 d in stable condition.

Additionally, to further control the bilobar liver metastases, decision was made to undergo two lobar Y-90 treatments using Y-90 TheraSphere microspheres (BTG, London, United Kingdom). The patient was evaluated for radioembolization with selective hepatic arteriography (Figure 2). Arterial administration of 4.1 mCi of 99mTc-MAA (Drax Image MAA kit; 99.7% purity) in the proper hepatic artery demonstrated a hepatopulmonary shunt of 0.8% on planar scintigraphy and SPECT, which is within acceptable range (< 10%). Additional pretreatment planning included evaluation with an abdominal CTA which revealed no variant vascular anatomy (Figure 2A). Based on imaging, estimated right hepatic lobe volume of 1339 ml and left hepatic lobe volume of 409 ml were used to calculate the radioactivity required to deliver a desired dose of 120 Gy to each lobe (Figure 2B). Patient underwent treatment of the right hepatic lobe (Figure 2C) followed by the left lobe within a month. Post-treatments, she tolerated the procedures well with low-grade fevers lasting for 24 h and intermittent right upper quadrant pain, both of which are common and expected following this treatment, which resolved within 2-3 wk.

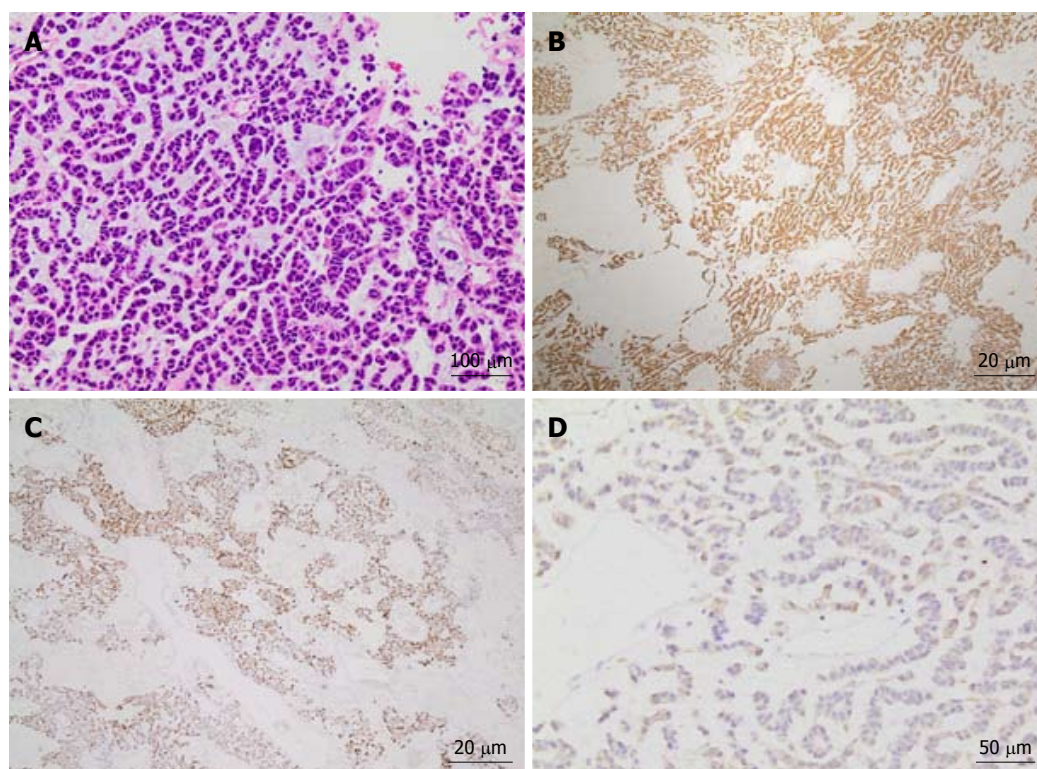


Figure 1 Tissue histopathology of a hepatic lesion confirms metastatic adrenocortical carcinoma. Hematoxylin and eosin (HE) staining (A) at 200 × magnification of tissue from the hepatic mass shows diffuse infiltrating cords of cells with hyperchromasia, eosinophilic cytoplasm, and a mild degree of nuclear pleomorphism in a background of myxoid stroma. Additional positive immunostaining with inhibin (B) and melan-A (C) at 40 × magnification as well as CKAE1/3 (D) at 100 × magnification confirms the diagnosis as adrenocortical carcinoma.

The patient tolerated the treatment well, experienced a drop in aldosterone levels (2.8 ng/dL), and has been normotensive and off of antihypertensive medication following intervention. Post Y-90 therapy imaging at 1 year revealed significantly smaller and less numerous hepatic lesions (Figure 3). Additionally, no mass was identified in the adrenal resection. Patient is currently in observation with periodic surveillance imaging and serology.

DISCUSSION

ACC is a rare malignancy but is highly lethal with limited treatment options. Given its rarity, ACC is difficult to study with most studies being retrospective case series with several key randomized trials^[9-11]. In a large landmark Phase II trial by Berruti *et al.*^[9] in 2005, the combination of etoposide, doxorubicin and cisplatin plus mitotane (EDP-M) in the treatment of advanced adrenocortical carcinoma demonstrated an overall response rate of 49%, a median time to progression of 9.1 mo, and an overall survival of 28.5 mo in a cohort of 72 patients^[9]. A previous study by Khan *et al.*^[10] in 2000 reported an overall response rate of 36% and a 2- and 5-year survival of 70% and 33%, respectively, with mitotane plus streptozotocin (M-S). In the First International Randomized trial in locally advanced and Metastatic Adrenocortical Carcinoma Treatment (FIRM-ACT) in 2012 of 304 patients, there was no significance

difference in the overall survival between EDP-M vs M-S (14.8 mo and 12.0 mo, respectively, $P = 0.07$), but the response rates (23.2% vs 9.2%, $P < 0.001$) and progression-free survival (5.0 mo vs 2.1 mo, $P < 0.001$) were significantly better with EDP-M^[11].

In addition to systemic chemotherapy, surgical resection and local treatment modalities such as RFA and TACE are key palliative treatment options. While adrenalectomy for local ACC disease can be curable and is first line therapy, surgical debulking is also often performed, as done here, in the setting of advanced metastatic disease for control of hormone excess^[12]. RFA has been also used in addition to surgery or as an alternative to surgery to reduce tumor burden. TACE has been utilized previously to treat liver metastases, although its efficacy has not been assessed specifically for ACC in clinical trials. ACC patients have been rendered disease free following hepatic RFA treatment in retrospective studies, but RFA has limited efficacy in patients with bilobar and multiple lesions as in our patient, highly vascularized metastases, or metastases in proximity to large vessels due to the heat-sink effect^[13,14]. In these patients, TACE, or the local intra-arterial delivery of chemotherapy to liver lesions with embolic agents, has been performed. Limited evaluation of TACE in the ACC literature demonstrates a partial response in 21% of patients and stabilization in 62%^[15].

In our case, we chose to pursue Y-90 SIRT for bilobar liver disease in a patient with metastatic ACC with the

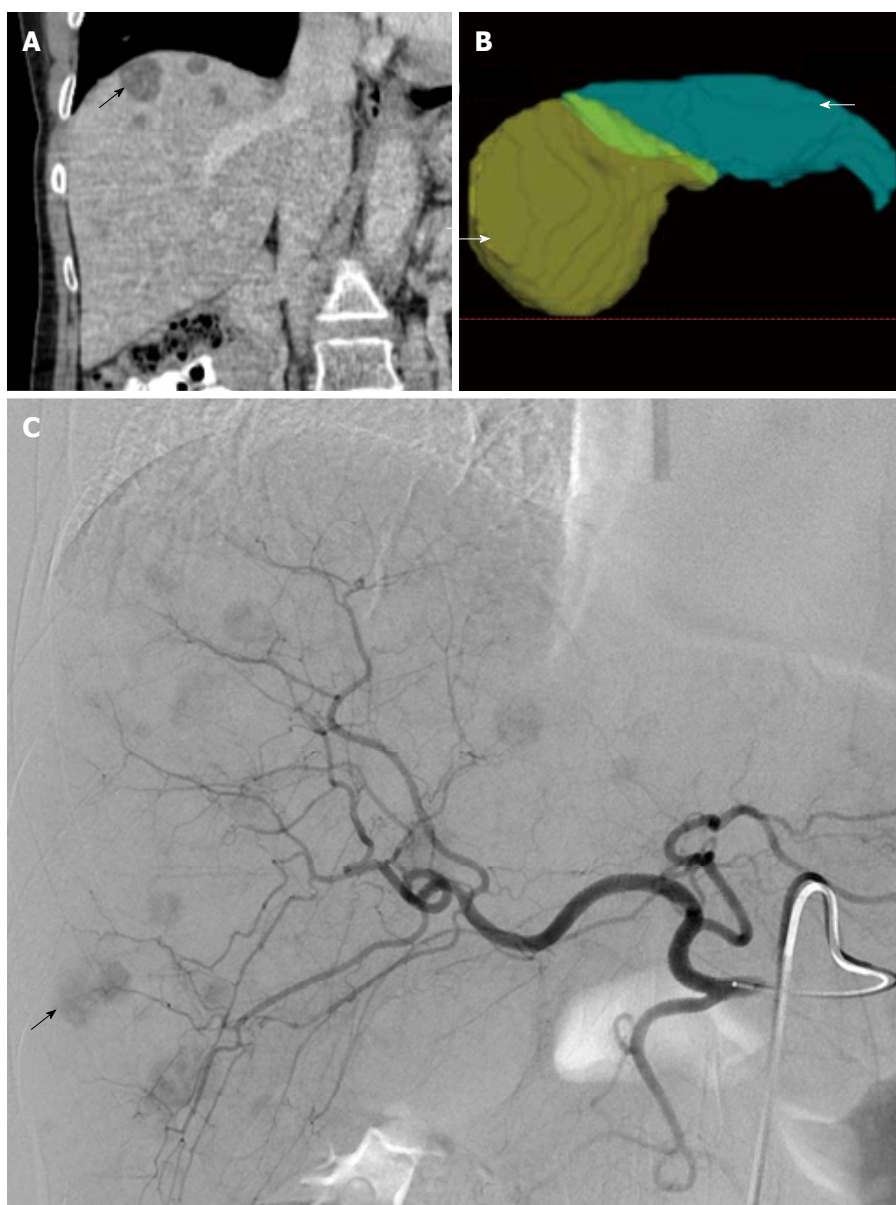


Figure 2 Yttrium-90 planning and radioembolization for treatment of liver metastases from adrenocortical carcinoma. A: Representative coronal CT abdomen image obtained prior to locoregional therapy shows low density lesions within the liver (arrow) consistent with metastatic disease; B: Three-dimensional reconstructions of the liver used for Yttrium-90 dose calculations revealed a proposed treatment volume of 409 mL for the left hepatic lobe (green colored, arrow) and 1339 mL for the right hepatic lobe (yellow colored, arrowhead) for a total hepatic volume of 1749 mL; C: Digitally subtracted contrast angiography reveals numerous hypervascular metastases within the right and left hepatic lobes (representative right hepatic lobe lesion demarked by arrow).

hopes of improving upon the outcomes previously seen with TACE. The benefit of Y-90 over TACE is that it can be delivered in an outpatient setting rather than the inpatient setting as reported previously for ACC, which offers convenience for the patients and is potentially more cost-effective^[16]. While Y-90 radioembolization has been primarily studied as a locoregional therapy for hepatic metastases from colorectal carcinoma or for multifocal hepatocellular carcinoma, we present its first successful utility when coupled with surgery and systemic chemotherapy to treat a patient with advanced ACC. Indeed, following a single session to each hepatic lobe, this patient has continued to demonstrate a positive radiologic and clinical response twelve months following treatment. Similar positive outcomes were observed by

Hendlisz *et al.*^[17] in a Phase III trial comparing fluorouracil infusion alone or combined with Y-90 radioembolization for metastatic colorectal cancer to the liver, which revealed a longer time to progression in the Y-90 arm (5.5 mo vs 2.1 mo, $P = 0.003$). Cosimelli *et al.*^[18] reported the results of Phase II trial showing that radioembolization alone produced meaningful response and disease stabilization in patients with advanced, unresectable and chemorefractory metastatic CRC^[18]. Our findings prompt further investigation through similar trials for patients with advanced ACC.

In conclusion, this case study presents the successful treatment of metastatic ACC by combining Y-90 locoregional therapy for bilobar liver disease with first line EDP-M chemotherapy and debulking surgery. This

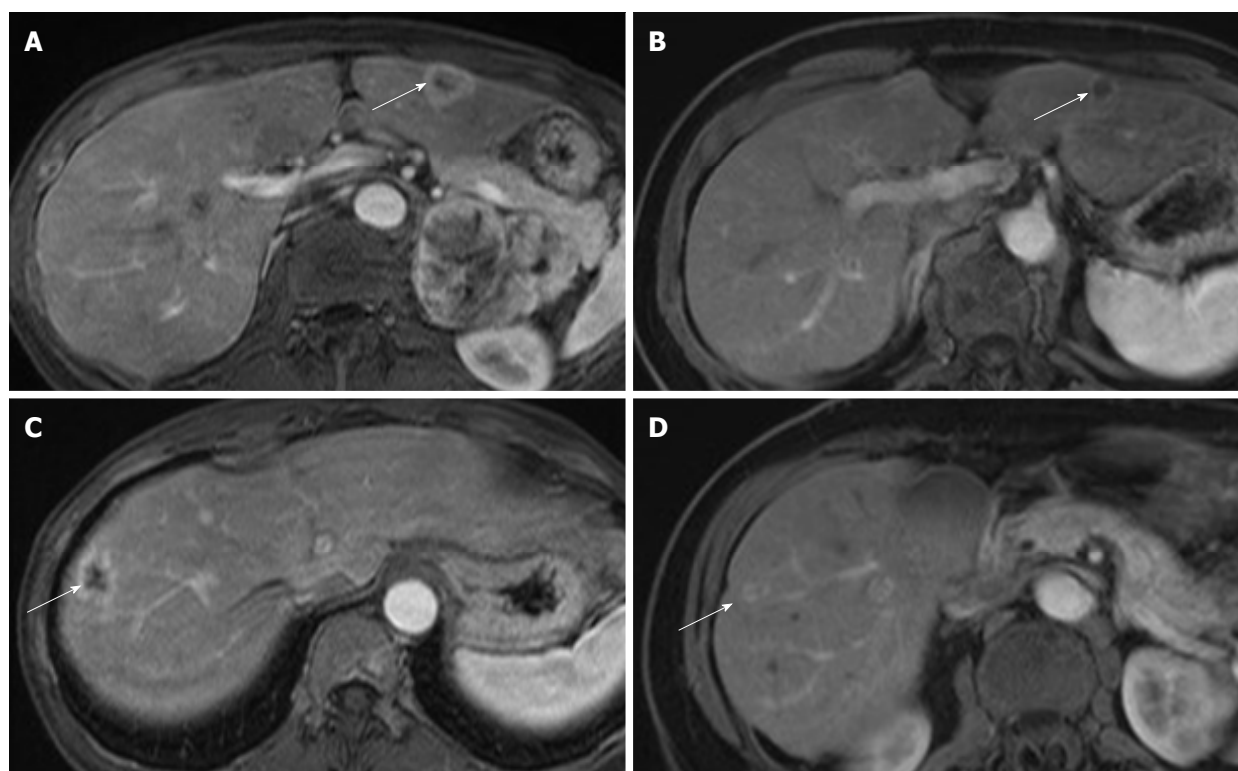


Figure 3 Hepatic metastases from adrenocortical carcinoma respond to Yttrium-90 radioembolization. Index hepatic lesions within the left hepatic lobe (A) and right hepatic lobe (C) demarked by arrows demonstrate significant reduction in size and perfusion one year following single dose treatment (B and D, respectively, arrows) by catheter-directed Yttrium-90 impregnated microspheres.

strategy allowed for disease management for at least 12 mo following a single outpatient Y-90 treatment for each hepatic lobe rather than the repeated inpatient treatments required previously through TACE with possibly superior results. Future prospective trials are needed to further elucidate the role of Y-90 radioembolization for treatment of advanced ACC.

ARTICLE HIGHLIGHTS

Case characteristics

A 60-year-old female patient with a past medical history significant only for poorly controlled hypertension despite medical therapy presented with a large left adrenal mass incidentally discovered on MRA during workup for renal artery stenosis.

Clinical diagnosis

The clinical examination of the patient revealed elevated blood pressure and abdominal pain.

Differential diagnosis

The differential diagnosis includes renal artery stenosis, adrenal pheochromocytoma, adrenocortical carcinoma, hyperaldosteronism, and hyperthyroidism.

Laboratory diagnosis

Serology values revealed normal urine cortisol [14.1 mcg/24 h (normal range: 3-22)], low adrenocorticotropic hormone (ACTH) [5.1 pg/mL (normal range: 9-50)], normal plasma metanephrines [0.20 nmol/L (normal range < 0.50)] and high aldosterone [38.8 ng/dL (normal range: 1-16)].

Imaging diagnosis

Abdominal MR imaging revealed a heterogeneous, partially necrotic left adrenal mass measuring 5.6 cm × 8.9 cm as well as multiple bilobar enhancing liver metastases.

Pathological diagnosis

Histologic and immunologic evaluation of a biopsied hepatic lesion showed metastatic carcinoma positive for inhibin, melan-A, and CKA1/3 with an immunohistochemical staining profile consistent with metastatic adrenocortical carcinoma.

Treatment

We present the first reported case of utilizing yttrium-90 microsphere selective internal radiation therapy (SIRT) in combination with first line EDP-M (Etoposide, Doxorubicin, Cisplatin, Mitotane) chemotherapy and debulking surgical primary tumor resection for treatment of metastatic adrenocortical carcinoma.

Related reports

While surgical resection can be curative for early local disease, most patients present with advanced ACC owing to nonspecific symptoms. For those patients, previous reports studied treatment options including systemic chemotherapy and locoregional therapies including radiofrequency ablation and transarterial chemoembolization.

Term explanation

Adrenocortical carcinoma (ACC) is a rare malignancy that constitutes less than 5% of all adrenal incidentalomas and has an estimated annual incidence of 1-2 cases per million people. EDP-M (etoposide, doxorubicin and cisplatin plus mitotane) is first line chemotherapy for advanced adrenocortical carcinoma demonstrating an overall response rate of 49%, a median time to progression of 9.1 mo, and an overall survival of 28.5 mo in prior studies.

Experiences and lessons

This case study presents the successful treatment of metastatic ACC by combining Y-90 locoregional therapy for bilobar liver disease with first line EDP-M chemotherapy and debulking surgery. This strategy allowed for disease management for at least 12 mo following a single outpatient Y-90 treatment for each hepatic lobe rather than the repeated inpatient treatments required previously through TACE with possibly superior results.

REFERENCES

- 1 **Zini L**, Porpiglia F, Fassnacht M. Contemporary management of adrenocortical carcinoma. *Eur Urol* 2011; **60**: 1055-1065 [PMID: 21831516 DOI: 10.1016/j.eururo.2011.07.062]
- 2 **Wandoloski M**, Bussey KJ, Demeure MJ. Adrenocortical cancer. *Surg Clin North Am* 2009; **89**: 1255-1267 [PMID: 19836496 DOI: 10.1016/j.suc.2009.06.019]
- 3 **Koschker AC**, Fassnacht M, Hahner S, Weismann D, Allolio B. Adrenocortical carcinoma -- improving patient care by establishing new structures. *Exp Clin Endocrinol Diabetes* 2006; **114**: 45-51 [PMID: 16570232 DOI: 10.1055/s-2006-923808]
- 4 **Lafemina J**, Brennan MF. Adrenocortical carcinoma: past, present, and future. *J Surg Oncol* 2012; **106**: 586-594 [PMID: 22473597 DOI: 10.1002/jso.23112]
- 5 **Bilimoria KY**, Shen WT, Elaraj D, Bentrem DJ, Winchester DJ, Kebebew E, Sturgeon C. Adrenocortical carcinoma in the United States: treatment utilization and prognostic factors. *Cancer* 2008; **113**: 3130-3136 [PMID: 18973179 DOI: 10.1002/cncr.23886]
- 6 **Berruti A**, Baudin E, Gelderblom H, Haak HR, Porpiglia F, Fassnacht M, Pentheroudakis G; ESMO Guidelines Working Group. Adrenal cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2012; **23** Suppl 7: vii131-vii138 [PMID: 22997446 DOI: 10.1093/annonc/mds231]
- 7 **Dackiw AP**, Lee JE, Gagel RF, Evans DB. Adrenal cortical carcinoma. *World J Surg* 2001; **25**: 914-926 [PMID: 11572033]
- 8 **Fay AP**, Elfiky A, Teló GH, McKay RR, Kaymakcalan M, Nguyen PL, Vaidya A, Ruan DT, Bellmunt J, Choueiri TK. Adrenocortical carcinoma: the management of metastatic disease. *Crit Rev Oncol Hematol* 2014; **92**: 123-132 [PMID: 24958272 DOI: 10.1016/j.critrevonc.2014.05.009]
- 9 **Berruti A**, Terzolo M, Sperone P, Pia A, Della Casa S, Gross DJ, Carnaghi C, Casali P, Porpiglia F, Mantero F, Reimondo G, Angeli A, Dogliotti L. Etoposide, doxorubicin and cisplatin plus mitotane in the treatment of advanced adrenocortical carcinoma: a large prospective phase II trial. *Endocr Relat Cancer* 2005; **12**: 657-666 [PMID: 16172198 DOI: 10.1677/erc.1.01025]
- 10 **Khan TS**, Imam H, Juhlin C, Skogseid B, Gröndal S, Tibblin S, Wilander E, Oberg K, Eriksson B. Streptozocin and o,p'DDD in the treatment of adrenocortical cancer patients: long-term survival in its adjuvant use. *Ann Oncol* 2000; **11**: 1281-1287 [PMID: 11106117]
- 11 **Fassnacht M**, Terzolo M, Allolio B, Baudin E, Haak H, Berruti A, Welin S, Schade-Brittinger C, Lacroix A, Jarzab B, Sorbye H, Torpy DJ, Stepan V, Schteingart DE, Arlt W, Kroiss M, Lebouilleux S, Sperone P, Sundin A, Hermesen I, Hahner S, Willenberg HS, Tabarin A, Quinkler M, de la Fouchardière C, Schlumberger M, Mantero F, Weismann D, Beuschlein F, Gelderblom H, Wilmink H, Sender M, Edgerly M, Kenn W, Fojo T, Müller HH, Skogseid B; FIRM-ACT Study Group. Combination chemotherapy in advanced adrenocortical carcinoma. *N Engl J Med* 2012; **366**: 2189-2197 [PMID: 22551107 DOI: 10.1056/NEJMoa1200966]
- 12 **Else T**, Kim AC, Sabolch A, Raymond VM, Kandathil A, Caoili EM, Jolly S, Miller BS, Giordano TJ, Hammer GD. Adrenocortical carcinoma. *Endocr Rev* 2014; **35**: 282-326 [PMID: 24423978 DOI: 10.1210/er.2013-1029]
- 13 **Ripley RT**, Kemp CD, Davis JL, Langan RC, Royal RE, Libutti SK, Steinberg SM, Wood BJ, Kammula US, Fojo T, Avital I. Liver resection and ablation for metastatic adrenocortical carcinoma. *Ann Surg Oncol* 2011; **18**: 1972-1979 [PMID: 21301973 DOI: 10.1245/s10434-011-1564-z]
- 14 **Bauditz J**, Quinkler M, Wermke W. Radiofrequency thermal ablation of hepatic metastases of adrenocortical cancer--a case report and review of the literature. *Exp Clin Endocrinol Diabetes* 2009; **117**: 316-319 [PMID: 19053031 DOI: 10.1055/s-0028-1087178]
- 15 **Gates VL**, Marshall KG, Salzig K, Williams M, Lewandowski RJ, Salem R. Outpatient single-session yttrium-90 glass microsphere radioembolization. *J Vasc Interv Radiol* 2014; **25**: 266-270 [PMID: 24332243 DOI: 10.1016/j.jvir.2013.11.005]
- 16 **de Baere T**, Teriitehau C, Deschamps F, Catherine L, Rao P, Hakime A, Auperin A, Goere D, Elias D, Hechelhammer L. Predictive factors for hypertrophy of the future remnant liver after selective portal vein embolization. *Ann Surg Oncol* 2010; **17**: 2081-2089 [PMID: 20237856 DOI: 10.1245/s10434-010-0979-2]
- 17 **Hendilisz A**, Van den Eynde M, Peeters M, Maleux G, Lambert B, Vannoote J, De Keukeleire K, Verslype C, Defreyne L, Van Cutsem E, Delatte P, Delaunoy T, Personeni N, Paesmans M, Van Laethem JL, Flamen P. Phase III trial comparing protracted intravenous fluorouracil infusion alone or with yttrium-90 resin microspheres radioembolization for liver-limited metastatic colorectal cancer refractory to standard chemotherapy. *J Clin Oncol* 2010; **28**: 3687-3694 [PMID: 20567019 DOI: 10.1200/JCO.2010.28.5643]
- 18 **Cosimelli M**, Golfieri R, Cagol PP, Carpanese L, Sciuto R, Maini CL, Mancini R, Sperduti I, Pizzi G, Diodoro MG, Perrone M, Giampalma E, Angelelli B, Fiore F, Lastoria S, Bacchetti S, Gasperini D, Geatti O, Izzo F; Italian Society of Locoregional Therapies in Oncology (SITIO). Multi-centre phase II clinical trial of yttrium-90 resin microspheres alone in unresectable, chemotherapy refractory colorectal liver metastases. *Br J Cancer* 2010; **103**: 324-331 [PMID: 20628388 DOI: 10.1038/sj.bjc.6605770]

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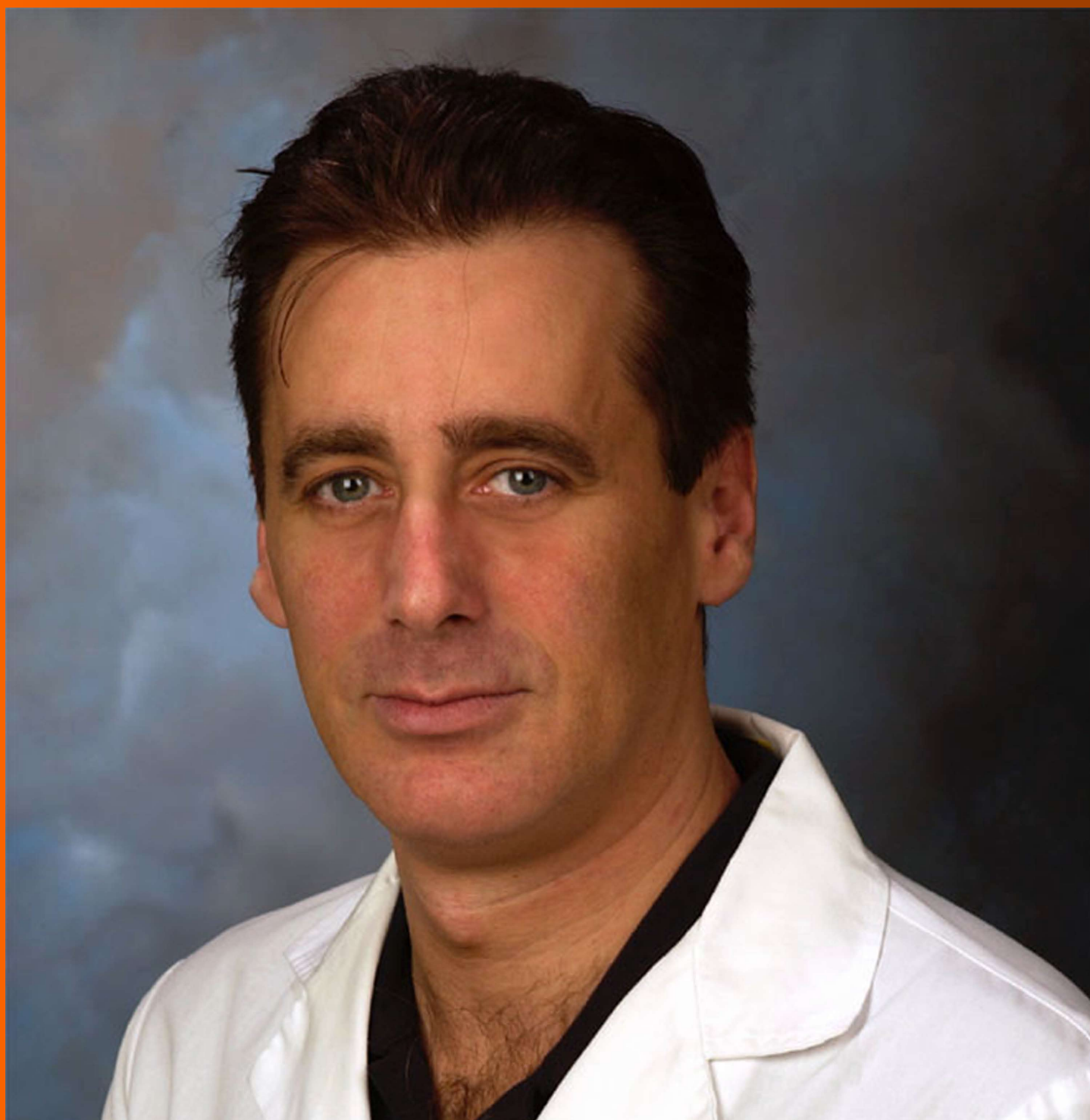


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Cell-free DNA integrity for the monitoring of breast cancer: Future perspectives?

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Abstract

Breast cancer (BC) is the most common cancer and the second cause of death in women worldwide. Therapeutic options are increasing, but the response to treatments is not always efficient and the risk of recurrence covers decades. In this perspective, the need to have a proper follow-up for the therapeutic responses and for anticipating recurrence it is urgent in the clinical setting. Liquid biopsy provides the basic principle for a non-invasive method for the routinely monitoring of BC. However, due to the heterogeneity of tumors during onset and progression, the search for tumor DNA mutations of targeted genes in plasma/serum is a limiting factor. A possible approach overtaking this problem comes from the measurement of cell-free DNA integrity, which is an independent factor from the mutational status and theoretically is representative of all tumors. This review summarizes the state-of-the-art of cell-free DNA integrity researches in BC, the controversies and the future perspective.

Key words: cfDNA integrity; Liquid biopsy; Breast cancer; ALU sequences; LINE-1 sequences

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Core tip: Despite the potentiality of cell-free DNA integrity as a useful tool for the monitoring of Breast Cancer (BC), evinced in some clinical studies, the scientific community has not reached agreeable conclusions to translate the results from the bench-to-the-bedside yet. The main controversy regards

the targets' choice and the size of circulating cell-free tumor DNA fragments. This work underlines the utility of cell-free DNA Integrity evaluation for BC follow-up and at the same time highlights the common concepts explaining the different results in line of future directions.

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INTRODUCTION

Breast cancer (BC) is still the most common cancer and the second cause of cancer-related death in women worldwide^[1]. A timely knowledge of its occurrence, responsiveness to therapies and recurrence is becoming of paramount importance for clinicians to adopt specific and more efficient approaches with regards to any single patient's health assistance. In clinical routine, the evaluation of serum markers as CEA or CA15-3 is still used for BC follow-up, but with a low specificity and sensibility^[2-5]. Up to now, one of the most promising frontiers in this field is the liquid biopsy. Recently, the meta-analysis on the clinical utility of circulating tumor cells (CTC) in early BC or in metastatic BC (MBC) provides a solid rationale for their use in oncological settings^[6-8]. However, their routinely use is still compromised by the relatively high cost of the technique.

Circulating cell-free DNA and qPCR measurement

From the blood circulation, it is possible to derive CTC, exosomes or cell-free nucleic acids (Figure 1). Cell-free DNA (cfDNA), consists of DNA fragments released after cell death processes from both tumor and normal cells. The circulating tumor DNA (ctDNA) can be differentiated from the rest of the cfDNA by looking at tumor-specific DNA changes, including mutations, gene amplifications, rearrangements and methylations^[9] proving it as a valid non-invasive biomarker to monitor tumor growth, spread, clonal evolution and response to therapies^[10]. This can be achieved either by a qualitative way (*i.e.*, type of mutations) or quantitative way (*i.e.*, copy number evaluation of mutated genes). However, the known mutations that can be used in liquid biopsy represent a limited percentage of patients. As an example, the most studied *PI3KCA* mutations all together have been found in about 30%-40% of BC patients^[11].

Here, both low-cost and easy-to-be-perform methods that are not bound to one or few specific genetic mutations to predict occurrence and monitor disease progression in BC patients will be described in line of what is currently known in literature.

Briefly, real-time polymerase chain reaction-or quantitative PCR (qPCR) is a powerful advancement of PCR technology that enables the measurement of the starting amount of nucleic acids in the reaction without the need for post-PCR gel analysis. This is achieved by the possibility to detect in a real-time manner the amplification process by fluorescence and to measure the amplification products of samples at exponential phases. Through this technology the expression of a target is measured by fluorescent probes or DNA-labelling dyes. Of note, the qPCR dyes do not discriminate between specific or non-specific amplicon products, thus there is a need for an accurate testing of the annealing conditions and buffer reagents to guarantee specificity of the reaction. The quantification of an unknown sample can be absolute by using an internal amplification standard curve obtained with known DNA quantities or it can be relative by comparison of the difference in cycle threshold values (Ct) of a unknown sample with respect to reference (mainly expressed as $\Delta\Delta\text{Ct}$ values)^[12,13]. Finally, to improve the accuracy of measurements, qPCR offers, together with the basic reagents, a passive fluorescein or ROX dyes to remove well-factors. The fluorescein acts as a passive reference dye, providing sufficient background fluorescence before the amplification reaction occurs, removing in this way the well factors-such as pipetting inaccuracies and fluorescence fluctuations-from the plate with the test samples.

Quantification of total circulating cell-free DNA

Some studies have focused on the quantification of total cfDNA levels using *GAPDH*, *Beta-globin*, *Beta2-Microglobulin*, *hTERT* or *LINE-1* as potential target genes, making the higher levels of cfDNA as a way to distinguish benign from malignant BC^[14-18]. Also SYBR Green's fluorescence to measure total serum cfDNA has been investigated^[19]. However, in our opinion, it is worth to consider how the total cfDNA levels are susceptible to increase also by the presence of other pathological conditions (*e.g.*, infection, inflammation, *etc.*), thus influencing the results.

Quantification of cell-free DNA integrity

The detection of ctDNA levels using cell-free DNA integrity (cfDI) measurement, as ratio between longer and shorter DNA fragments, is more specific than total serum cfDNA and has been explored in BC by qPCR by many authors using SYBRGreen fluorescent dye (Table 1). In principle, normal cells, undergoing apoptosis, release DNA fragments of about 200 bp as the result of enzymatic cleavage of nucleosome units; whereas, tumor cells undergo many different death processes, including necrosis and autophagy, and they can release DNA fragments of different sizes^[20,21]. Umetani *et al.*^[22], using ALU targets proposed cfDI for the first time as a valuable tool to identify primary BC, showing it could be suitable to define lymph node metastasis in a group of 83 patients compared to 51 healthy controls.

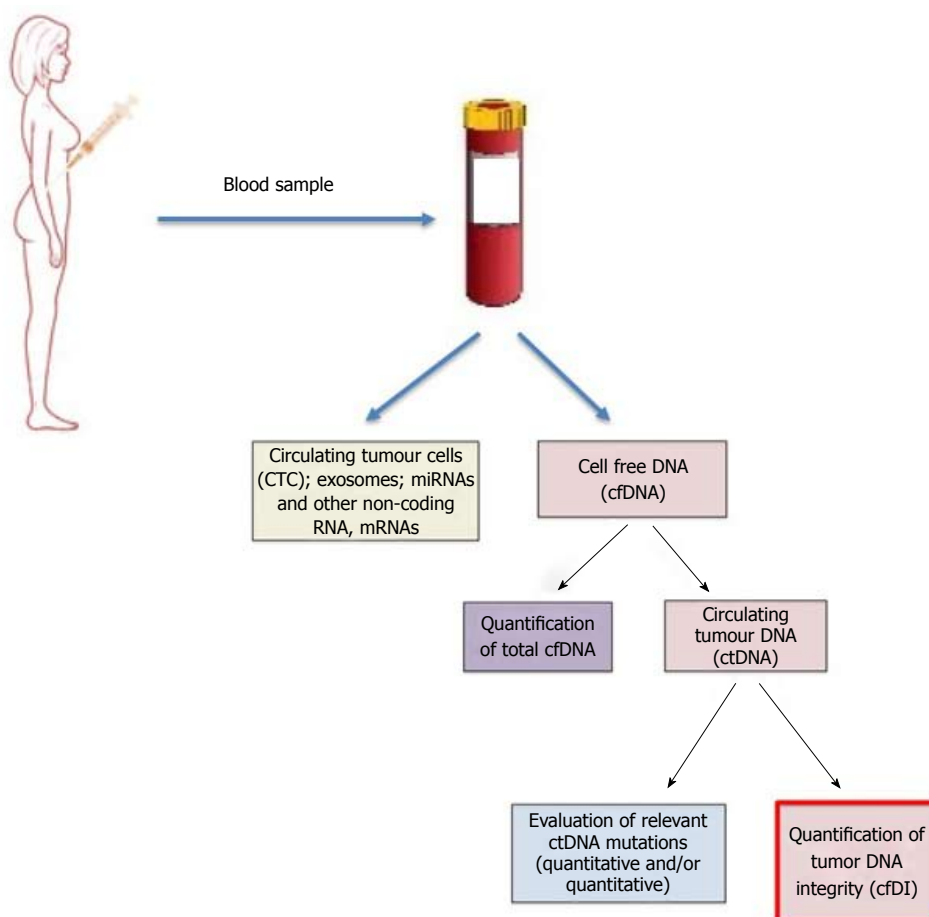


Figure 1 Diagram summarizing the possibility to monitor breast cancer from the blood circulating DNA.

They measured in serum shorter fragments of 115 bp that were considered as derived from apoptotic normal cells and larger ones of 247 bp as ctDNA, derived from necrosis/autophagy of cancer cells. The cfDI value calculated as the ratio quantity of longer over shorter fragments, ALU247/ALU115, was found to be higher in BC patients with high grade cancer compared to healthy controls. Accordingly to Umetani *et al.*^[22], Agostini *et al.*^[23] using the same ALU247 bp and ALU115bp targets demonstrated in plasma that cfDI value was twice higher in BC patients ($n = 39$) vs healthy controls ($n = 49$). Subsequently, Stötzer *et al.*^[24] proved in plasma that the ratio ALU247/115 were higher in patients with locally confined BC and MBC ($n = 47$) than benign BC ($n = 12$) ($P < 0.001$) but not vs healthy controls ($n = 28$). Moreover, this group evidenced that ALU concentrations alone were very interesting as markers for locally confined BC, while the use of cfDI was limited by the elevated levels found in some healthy controls. However, Iqbal *et al.*^[25] enrolling a larger number of women (148 patients vs 51 healthy controls) confirmed that the cfDI value, represented as ALU247/115 ratio, was significantly higher in serum of patients compared to healthy controls. Moreover, through a multivariate analysis, they showed a correlation between the cfDI value and

the tumor size to predict the overall survival (OS) at 5 years and disease-free survival (DFS) at 4 years. Madhavan *et al.*^[21] also considered cfDI as a useful biomarker for BC in the largest patients' cohort (82 BC and 201 MBC) by using different primer set for ALU sequences and introducing LINE-1 as another DNA repetitive element target. They quantified ALU 260 bp and LINE-1 266 bp amplicons vs ALU 111 bp and LINE-197 bp amplicons, respectively. They showed, differently than the other groups, cfDI value was lower in BC patients vs healthy control and positively correlated with a decrease in progression-free survival (PFS) ($P = 0.0025$ for ALU) and OS ($P < 0.0001$ for both ALU and LINE-1). Similarly, using the same ALU260/111 and LINE-1 266/197 ratios, Cheng *et al.*^[26] showed that cfDI was significantly lower in recurrent BC ($n = 37$) vs non-recurrent BC ($n = 175$) ($P < 0.001$ for both ALU and LINE-1 cfDI values) but they did not provide as an extra measure healthy controls. Interestingly, this latter research group showed that a higher risk of developing recurrence could be predicted by the reduction of cfDI value ($P = 0.020$ for ALU and $P = 0.019$ for LINE-1 cfDI values, respectively). Finally, it should be mentioned that Cheng *et al.*^[27] recently observed that higher cfDI values for both ALU and LINE-1 targets in MBC patients correlated

Table 1 cfDI evaluation for the monitoring of breast cancer

Targets, length of the amplicons and primers' sequences	Patients with primary BC	Results	Ref.
ALU, 115 bp FW: 5'-CCTGAGGTCAGGAGTTCGAG-3' RV: 5'-CCCGAGTAGCTGGGATTACA-3'	Healthy females ($n = 51$) and BC patients ($n = 83$) DNA from serum	The ratio ALU247/115 was higher in 51 patients with stage II ($P = 0.005$), stage III ($P < 0.0001$), stage IV (0.002) compared to healthy controls but not in 32 patients with stage 0 or I	Umetani <i>et al</i> ^[22] , 2006
ALU, 247 bp FW: 5'-GTGGCTCACGCCTGTAATC-3' RV: 5'-CAGGCTGGAGTGCAGTGG-3'	Healthy females ($n = 49$) and BC patients ($n = 39$) DNA from plasma	In the group of patients the ratio ALU247/115 was twice higher ($P < 0.0001$) than in the group of healthy controls	Agostini <i>et al</i> ^[23] , 2012
ALU, 115 bp FW: 5'-CCTGAGGTCAGGAGTTCGAG-3' RV: 5'-CCCGAGTAGCTGGGATTACA-3'	Healthy females ($n = 28$), benign breast disease patients ($n = 12$), locally confined BC patients ($n = 65$) and MBC patients ($n = 47$) DNA from plasma	The ratio ALU247/115 was higher in patients with locally confined BC and metastatic BC than in benign BC ($P < 0.001$), but not <i>vs</i> healthy controls	Stötzer <i>et al</i> ^[24] , 2014
ALU, 247 bp FW: 5'-GTGGCTCACGCCTGTAATC-3' RV: 5'-CAGGCTGGAGTGCAGTGG-3'	Healthy females ($n = 100$), primary BC patients ($n = 82$) and MBC patients ($n = 201$) DNA from plasma	Both the ratios ALU 260/111 and LINE-1 266/97 were lower in primary BC patients (ALU: $P = 0.046$; LINE-1 $P = 0.041$) In MBC patients the lower values of cfDI were related to both a decrease in PFS ($P = 0.0025$ for ALU) and OS ($P < 0.0001$ for both ALU and LINE-1 fragments)	Madhavan <i>et al</i> ^[21] , 2014
LINE-1, 97 bp FW: 5'-TGGACATATACACCATGGAA-3' RV: 5'TGAGAATGATGGTTTCCAATTTC-3'			
LINE-1, 266 bp FW: 5'-ACTTGAACCAACCCAAATG-3' RV: 5'-CACCACAGTCCCAGAGTG-3'			
ALU, 115 bp FW: 5'-CCTGAGGTCAGGAGTTCGAG-3' RV: 5'-CCCGAGTAGCTGGGATTACA-3'	Healthy females ($n = 51$) and BC patients ($n = 148$) DNA from serum	The ratio ALU 247/115 was significantly higher in patients compared to controls ($P < 0.001$)	Iqbal <i>et al</i> ^[25] , 2015
ALU, 247 bp FW: 5'-GTGGCTCACGCCTGTAATC-3' RV: 5'-CAGGCTGGAGTGCAGTGG-3'			
Beta-actin, 100 bp FW: 5'-GCACCACACCTTCTACAATGA-3' RV: 5'-GTATCTTCTCGCGGTGGC-3'	Healthy females ($n = 70$), benign lesions ($n = 95$) and BC patients ($n = 95$) DNA from plasma	cfDI value calculated as difference between 400 bp and 100 bp fragments Higher cfDI values were obtained in BC compared to benign lesions and healthy subjects ($P < 0.001$)	Kamel <i>et al</i> ^[20] , 2016
Beta-actin, 400 bp FW: 5-GCACCACACCTTCTACAATGA-3' (common primer) RV: 5'-TGTCACGCACGATTCCCC-3'			
HER2, 126 bp FW-5-CCAGGGTGTCTCCTCAGTTGT-3' RV-5- GGAGTTCCTGCAGAGGACAG-3'	Healthy females ($n = 10$), BC patients ($n = 79$) DNA from serum	The ratios BCAS1 266/129, MYC 264/128, PIK3CA 274/129 were significantly higher in patients compared to controls ($P = 0.002$, $P = 0.030$ and $P = 0.004$, respectively) No significant values for HER2 targets	Maltoni <i>et al</i> ^[28] , 2017
HER2, 295 bp FW-5'-CCAGGGTGTCTCCTCAGTTGT-3' RV-5'-TCAGTATGGCCTCACCTTC-3'			
MYC, 128 bp FW-5-GGCATTTAAATTCGGCTCA-3' RV-5-AAAAGCCAAATGCCAACTT-3'			
MYC, 264 bp FW-5'-TGGAGTAGGGACCGCATATC-3' RV-5'-ACCCAACACACGTCCTAAC-3'			
BCAS1, 129 bp FW-5-GGGTCAGAGCTTCTGTGAG-3' RV-5-TATCATGCCTTGGAGAACCA-3'			
BCAS1, 266 bp FW-5'-GGGTCAGAGCTTCTGTGAG-3' RV-5'-CGTGTCTGAAACAGAGCA-3'			
PIK3CA, 129 bp FW-5'CTCCACGACCATCATCATCAGGT-3' RV-5'-TGGTTATTAATGAGCCTCACGG-3'			
PIK3CA, 274 bp FW-5'-CTC CACGAC CAT CATCAGGT-3' RV-5'-CGAAGGTCACAAAGTCGTCT-3'			

ALU, 111 bp FW: 5'-CTGGCCAACATGGTGAAAC-3' RV: 5'-AGCGATTCTCCTGCCTCAG-3'	Non-recurrent BC patients (<i>n</i> = 175) <i>vs</i> recurrent-BC patients (<i>n</i> = 37) No healthy females reported DNA from plasma	Both the ratios ALU260/111 and LINE1-266/97 were significantly lower during follow-up in recurrent BC <i>vs</i> non recurrent BC (<i>P</i> < 0.001 for both ALU and LINE-1 cfDI). Moreover, BC patients with a lower cfDI had higher risk of developing recurrence compared to patients with higher cfDI (<i>P</i> = 0.020 for ALU cfDI and <i>P</i> = 0.019 for LINE-1 cfDI, respectively)	Cheng <i>et al</i> ^[26] , 2017
ALU, 260 bp FW: 5'-ACGCCTGTAATCCCAGCA-3' RV: 5'-CGGAGTCTCGTCTGTGCG-3'			
LINE-1, 97 bp FW: 5'-TGGCACATATACACCATGGAA-3' RV: 5'-TGAGAAATGATGGTTTCCAATTTC-3'			
LINE-1, 266 bp FW: 5'-ACTTGGAAACCAACCCAAATG-3' RV: 5'-CACCACAGTCCCCAGAGTG-3'			
ALU, 111 bp FW: 5'-CTGGCCAACATGGTGAAAC-3' RV: 5'-AGCGATTCTCCTGCCTCAG-3'	MBC patients (total <i>n</i> = 268) No healthy females DNA from plasma	Both the ratios ALU260/111 and LINE1-266/97 significantly increased in 268 MBC patients treated with one cycle of chemotherapy (MBCLB) compared to MBC at baseline (MBC1C) (<i>P</i> = 0.00017 for ALU -0.053 <i>vs</i> 0.063- and <i>P</i> = 0.0016 for LINE-1-0.45 <i>vs</i> 0.49)	Cheng <i>et al</i> ^[27] , 2018
ALU, 260 bp FW: 5'-ACGCCTGTAATCCCAGCA-3' RV: 5'-CGGAGTCTCGTCTGTGCG-3'		Moreover, in both MBCBL and MBC1C patients with a higher cfDI (for both ALU and LINE-1) correlated with a higher PFS and OS <i>vs</i> lower cfDI MBC patients	
LINE-1, 97 bp FW: 5'-TGGCACATATACACCATGGAA-3' RV: 5'-TGAGAAATGATGGTTTCCAATTTC-3'			
LINE-1, 266 bp FW: 5'-ACTTGGAAACCAACCCAAATG-3' RV: 5'-CACCACAGTCCCCAGAGTG-3'			

BC: Breast cancer; cfDNA: Cell-free DNA; cfDI: Cell-free DNA integrity; ctDNA: Circulating tumour DNA; DFS: Disease free survival; MBC: Metastatic breast cancer; PFS: Progression-free survival; OS: Overall survival; qPCR: Quantitative real-time PCR; ddPCR: Droplet digital PCR.

with longer PFS and OS. However, Kamel *et al*^[20] measuring the 400 bp and 100 bp amplicons of the *Beta-actin* from the DNA derived from plasma of 95 BC and 95 benign lesions *vs* 70 healthy controls estimated a cfDI- as difference between longer and shorter fragments- accordingly to Umetani *et al*^[22] and the other authors^[23-25], while yet differently from Madhavan *et al*^[21]. In fact cfDI was found significantly higher in BC samples compared to those of benign and healthy subjects (*P* < 0.001). Moreover, they related those higher values to TNM stage, suggesting a cut-off to identify the more aggressive BC^[20]. In agreement with Kamel *et al*^[20], Maltoni *et al*^[28] recently showed that tumour cells released longer DNA fragments than normal cells in the bloodstream. They quantified large fragments of 295 bp, 264 bp, 266 bp, 274 bp and short amplicons of 126, 128, 129, 129 bp from *HER2*, *MYC*, *BCAS1* and *PIK3CA*, respectively, from the serum of healthy females (*n* = 10), non-recurrent BC (*n* = 58) and recurrent BC (*n* = 21). They estimated cfDI as the ratio between longer and shorter amplicons of these genes and demonstrated that *BCAS1*, *MYC* and *PIK3CA* long/short amplicons were significantly higher in patients compared to healthy controls (*P* = 0.002, *P* = 0.030 and *P* = 0.004, respectively). On the other hand, there was no significant difference for long/short amplicons of *HER2*^[27].

DISCUSSION

The overall literature on cfDI is intriguing as it has an extraordinary potential for the monitoring of BC, but it remains to be clarified what is the expected value of cfDI: some authors claimed that ctDNA is made of longer amplicons than normal cfDNA, explaining

why the cfDI increased in BC^[20,22-25,27], whereas other research groups, using different primers, claimed the exact opposite^[21,26].

Most of the authors, in their measurement of cfDI through the ALU sequences, decided to use a standard DNA curve, as for Umetani *et al*^[22], to derive quantifications of their DNA^[21-25,27], and used the fluorescein or ROX passive reference dyes to improve the quality of their results^[23,25]. Additionally, the specificities of the amplification reactions for the different couple of primers described in the papers have been controlled by means of denaturation curves or gel electrophoresis. This implies that the different results by qPCR hardly can be attributable to the laboratory's methodology, although we cannot completely exclude some variability in sample collection in the studies here described. Of note, differently than the other groups, Stötzer *et al*^[24] have adopted a slightly different protocol for ALU amplifications by introducing UDP-DNA glycosidase.

Higher cfDI values in BC *vs* healthy controls were found in larger patients' cohorts derived from independent clinical settings and by using more different targets compared to studies claiming lower cfDI values in the tumor (Figure 2). Of note, higher cfDI in tumor than healthy controls were found in those studies that have analyzed mainly BCs, which did not reach the metastatic setting^[22,23,25], whereas lower cfDI than healthy controls were reported in a study using the largest MBC patients' cohort up-to-date^[21]. It is interesting to note that Umetani *et al*^[22] proposed an increased cfDI value to predict local micrometastasis and recently Cheng *et al*^[28] observed that cfDI value particularly decreased in BC patients with visceral metastasis. Thus we

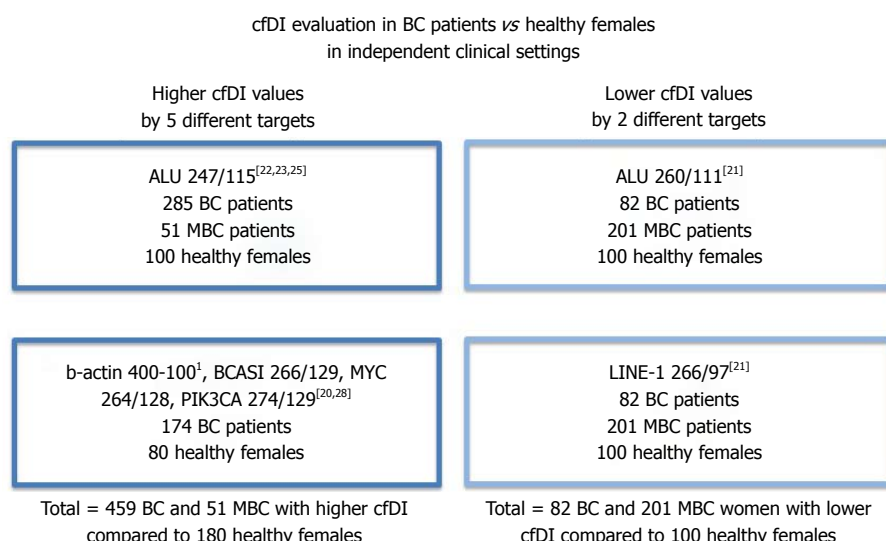


Figure 2 Summary of the literature data on cfDI determination in primary breast cancer vs healthy females. ¹Note that cfDI by β -actin was evaluated as difference between large and short amplicons and not as ratio longer to shorter amplicons. BC: Breast cancer; MBC: Metastatic breast cancer.

suggest that cfDI value can increase at initial stages of the BC and decrease in MBC. Surely, the most promising targets for the measurement of cfDI are represented by repetitive elements such as ALU and LINE-1 sequences, accounting for nearly 10% and 17% of the total genome, respectively. It is worth nothing that reproducible results were obtained when independent groups used the same ALU primer pairs, either those demonstrating higher cfDI^[22-25] and those demonstrating lower cfDI in BC^[21,26]. In our opinion, the methods of DNA extractions merely could have influenced the results. Interestingly, by looking with BLASTN genomic RefSeqGene Human at the target sites of ALU primers' pairs used by the research groups obtaining divergent results, we observed different target sites for ALU247/115 pairs compared to the ALU260/111 ones. We cannot exclude that this could contribute to the opposite cfDI values obtained by the different research groups comparing BC vs healthy controls. Moreover, we would like to point out that the qPCR methodology by SYBR Green is not very sensitive in quantifying very small DNA fragments in diluted solutions^[29], as it could be in liquid biopsy, and that the variability of amplification efficiency of a sample can be overtaken by many replicates and independent experiments, that are hard to performed with samples derived from liquid biopsy. In this respect, the determination of cfDI in liquid biopsy samples would benefit by more sensitive and accurate technologies such as digital droplet PCR (ddPCR).

In conclusion, monitoring primary and MBC through a non-invasive analysis such as that of circulating DNA remains one of the most interesting goals to achieve. Surely, the mutations in liquid biopsy are of paramount importance for targeted therapies and for monitoring response to treatment. However, the most interesting benefit-to-cost analysis for the follow-up of BC and its recurrence seems to be the evaluation

of circulating cfDI. Future investigations for cfDI by ddPCR are warranted for the (1) testing for the choice of best targets; (2) clarification of the clinical significance of larger and shorter DNA fragments origin in serum/plasma; and (3) a better understanding of the potential clinical impact of cfDI in anticipating recurrence and responsiveness to therapies for all patients, independently from the mutational signature of BC.

REFERENCES

- 1 **Torre LA**, Siegel RL, Ward EM, Jemal A. Global Cancer Incidence and Mortality Rates and Trends--An Update. *Cancer Epidemiol Biomarkers Prev* 2016; **25**: 16-27 [PMID: 26667886 DOI: 10.1158/1055-9965]
- 2 **Duffy MJ**, Evoy D, McDermott EW. CA 15-3: uses and limitation as a biomarker for breast cancer. *Clin Chim Acta* 2010; **411**: 1869-1874 [PMID: 20816948 DOI: 10.1016/j.cca.2010.08.039]
- 3 **Harris L**, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, Somerfield MR, Hayes DF, Bast RC Jr; American Society of Clinical Oncology. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. *J Clin Oncol* 2007; **25**: 5287-5312 [PMID: 17954709 DOI: 10.1200/JCO.2007.14.2364]
- 4 **Lauro S**, Trasatti L, Bordin F, Lanzetta G, Bria E, Gelibter A, Reale MG, Vecchione A. Comparison of CEA, MCA, CA 15-3 and CA 27-29 in follow-up and monitoring therapeutic response in breast cancer patients. *Anticancer Res* 1999; **19**: 3511-3515 [PMID: 10629644]
- 5 **Shao Y**, Sun X, He Y, Liu C, Liu H. Elevated Levels of Serum Tumor Markers CEA and CA15-3 Are Prognostic Parameters for Different Molecular Subtypes of Breast Cancer. *PLoS One* 2015; **10**: e0133830 [PMID: 26207909 DOI: 10.1371/journal.pone.0133830]
- 6 **Bidard FC**, Peeters DJ, Fehm T, Nolé F, Gisbert-Criado R, Mavroudis D, Grisanti S, Generali D, Garcia-Saenz JA, Stebbing J, Caldas C, Gazzaniga P, Manso L, Zamarchi R, de Lascoiti AF, De Mattos-Arruda L, Ignatiadis M, Lebofsky R, van Laere SJ, Meier-Stiegen F, Sandri MT, Vidal-Martinez J, Politaki E, Consoli F, Bottini A, Diaz-Rubio E, Krell J, Dawson SJ, Raimondi C, Rutten A, Janni W, Munzone E, Carañana V, Agelaki S, Almici C, Dirix L, Solomayer EF, Zorzino L, Johannes H, Reis-Filho JS, Pantel

- K, Pierga JY, Michiels S. Clinical validity of circulating tumour cells in patients with metastatic breast cancer: a pooled analysis of individual patient data. *Lancet Oncol* 2014; **15**: 406-414 [PMID: 24636208 DOI: 10.1016/S1470-2045(14)70069-5]
- 7 **Bidard FC**, Michiels S, Mueller V, Riethdorf S, Esserman L, Lucci A, Naume B, Horiguchi J, Gisbert-Criado R, Sleijfer S, Toi M, Garcia-Saenz J, Hartkopf A, Generali D, Rothe F, Smerage J, Muinelo L, Stebbing J, Viens P, Magbanua M, Hall C, Engebraten O, Takata D, Vidal-Martinez J, Onstenk W, Fujisawa N, Diaz-Rubio E, Taran FA, Cappelletti M, Ignatiadis M, Name N, Proudhon C, Wolf D, Bowman Bauldry J, Borgen E, Nagaoka R, Carañana V, Kraan J, Maestro M, Brucker S, Weber K, Rey F, Amara D, Gopalkrishna Karhade M, Ruud Mathiesen R, Tokiniwa H, Llombart-Cussac A, D'Hollander K, Cottu P, Park J, Loibl S, Pierga J Y, Pantel K. Abstract S3-01: IMENEO: International MEta-analysis of circulating tumor cell detection in early breast cancer patients treated by NEOadjuvant chemotherapy. *Cancer Res* 2017; **77**: S3-S1 [DOI: 10.1158/1538-7445.SABCS16-S3-01]
- 8 **Bonora M**, Wiecekowsk MR, Chinopoulos C, Kepp O, Kroemer G, Galluzzi L, Pinton P. Molecular mechanisms of cell death: central implication of ATP synthase in mitochondrial permeability transition. *Oncogene* 2015; **34**: 1608 [PMID: 25790189 DOI: 10.1038/bjc.2012.137]
- 9 **Schwarzenbach H**, Hoon DS, Pantel K. Cell-free nucleic acids as biomarkers in cancer patients. *Nat Rev Cancer* 2011; **11**: 426-437 [PMID: 21562580 DOI: 10.1038/nrc3066]
- 10 **De Mattos-Arruda L**, Caldas C. Cell-free circulating tumour DNA as a liquid biopsy in breast cancer. *Mol Oncol* 2016; **10**: 464-474 [PMID: 26776681 DOI: 10.1016/j.molonc.2015.12.001]
- 11 **Sobhani N**, Roviello G, Corona SP, Scaltriti M, Ianza A, Bortul M, Zanconati F, Generali D. The prognostic value of PI3K mutational status in breast cancer: a meta-analysis. *J Cell Biochem* 2018; Epub ahead of print [PMID: 29345357 DOI: 10.1002/jcb.26687]
- 12 **Livak KJ**, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001; **25**: 402-408 [PMID: 11846609 DOI: 10.1006/meth.2001.1262]
- 13 **Gal S**, Fidler C, Lo YM, Taylor M, Han C, Moore J, Harris AL, Wainscoat JS. Quantitation of circulating DNA in the serum of breast cancer patients by real-time PCR. *Br J Cancer* 2004; **90**: 1211-1215 [PMID: 15026803 DOI: 10.1038/sj.bjc.6601609]
- 14 **El Tarhouny S**, Seefeld M, Fan AX, Hahn S, Holzgreve W, Zhong XY. Comparison of serum VEGF and its soluble receptor sVEGFR1 with serum cell-free DNA in patients with breast tumor. *Cytokine* 2008; **44**: 65-69 [PMID: 18691902 DOI: 10.1016/j.cyto.2008.06.008]
- 15 **Bechmann T**, Andersen RF, Pallisgaard N, Madsen JS, Maae E, Jakobsen EH, Bak Jyelling AM, Steffensen KD, Jakobsen A. Plasma HER2 amplification in cell-free DNA during neoadjuvant chemotherapy in breast cancer. *J Cancer Res Clin Oncol* 2013; **139**: 995-1003 [PMID: 23479212 DOI: 10.1007/s00432-013-1413-5]
- 16 **Kohler C**, Radpour R, Barekati Z, Asadollahi R, Bitzer J, Wight E, Bürki N, Diesch C, Holzgreve W, Zhong XY. Levels of plasma circulating cell free nuclear and mitochondrial DNA as potential biomarkers for breast tumors. *Mol Cancer* 2009; **8**: 105 [PMID: 19922604 DOI: 10.1186/1476-4598-8-105]
- 17 **Huang ZH**, Li LH, Hua D. Quantitative analysis of plasma circulating DNA at diagnosis and during follow-up of breast cancer patients. *Cancer Lett* 2006; **243**: 64-70 [PMID: 16412565 DOI: 10.1016/j.canlet.2005.11.027]
- 18 **Sunami E**, Vu AT, Nguyen SL, Giuliano AE, Hoon DS. Quantification of LINE1 in circulating DNA as a molecular biomarker of breast cancer. *Ann N Y Acad Sci* 2008; **1137**: 171-174 [PMID: 18837943 DOI: 10.1196/annals.1448.011]
- 19 **Catarino R**, Ferreira MM, Rodrigues H, Coelho A, Nogal A, Sousa A, Medeiros R. Quantification of free circulating tumor DNA as a diagnostic marker for breast cancer. *DNA Cell Biol* 2008; **27**: 415-421 [PMID: 18694299 DOI: 10.1089/dna.2008.0744]
- 20 **Kamel AM**, Teama S, Fawzy A, El Deftar M. Plasma DNA integrity index as a potential molecular diagnostic marker for breast cancer. *Tumour Biol* 2016; **37**: 7565-7572 [PMID: 26684805 DOI: 10.1007/s13277-015-4624-3]
- 21 **Madhavan D**, Wallwiener M, Bents K, Zucknick M, Nees J, Schott S, Cuk K, Riethdorf S, Trumpp A, Pantel K, Sohn C, Schneeweiss A, Surowy H, Burwinkel B. Plasma DNA integrity as a biomarker for primary and metastatic breast cancer and potential marker for early diagnosis. *Breast Cancer Res Treat* 2014; **146**: 163-174 [PMID: 24838941 DOI: 10.1007/s10549-014-2946-2]
- 22 **Umetani N**, Giuliano AE, Hiramatsu SH, Amersi F, Nakagawa T, Martino S, Hoon DS. Prediction of breast tumor progression by integrity of free circulating DNA in serum. *J Clin Oncol* 2006; **24**: 4270-4276 [PMID: 16963729 DOI: 10.1200/JCO.2006.05.9493]
- 23 **Agostini M**, Enzo MV, Bedin C, Belardinelli V, Goldin E, Del Bianco P, Maschietto E, D'Angelo E, Izzi L, Saccani A, Zavagno G, Nitti D. Circulating cell-free DNA: a promising marker of regional lymphnode metastasis in breast cancer patients. *Cancer Biomark* 2012; **11**: 89-98 [PMID: 23011155 DOI: 10.3233/CBM-2012-0263]
- 24 **Stötzer OJ**, Lehner J, Fersching-Gierlich D, Nagel D, Holdenrieder S. Diagnostic relevance of plasma DNA and DNA integrity for breast cancer. *Tumour Biol* 2014; **35**: 1183-1191 [PMID: 24018822 DOI: 10.1007/s13277-013-1158-4]
- 25 **Iqbal S**, Vishnubhatla S, Raina V, Sharma S, Gogia A, Deo SS, Mathur S, Shukla NK. Circulating cell-free DNA and its integrity as a prognostic marker for breast cancer. *Springerplus* 2015; **4**: 265 [PMID: 26090312 DOI: 10.1186/s40064-015-1071-y]
- 26 **Cheng J**, Cuk K, Heil J, Golatta M, Schott S, Sohn C, Schneeweiss A, Burwinkel B, Surowy H. Cell-free circulating DNA integrity is an independent predictor of impending breast cancer recurrence. *Oncotarget* 2017; **8**: 54537-54547 [PMID: 28903362 DOI: 10.18632/oncotarget.17384]
- 27 **Cheng J**, Holland-Letz T, Wallwiener M, Surowy H, Cuk K, Schott S, Trumpp A, Pantel K, Sohn C, Schneeweiss A, Burwinkel B. Circulating free DNA integrity and concentration as independent prognostic markers in metastatic breast cancer. *Breast Cancer Res Treat* 2018; **169**: 69-82 [PMID: 29340881 DOI: 10.1007/s10549-018-4666-5]
- 28 **Maltoni R**, Casadio V, Ravaioli S, Foca F, Tumedei MM, Salvi S, Martignano F, Calistri D, Rocca A, Schirone A, Amadori D, Bravaccini S. Cell-free DNA detected by "liquid biopsy" as a potential prognostic biomarker in early breast cancer. *Oncotarget* 2017; **8**: 16642-16649 [PMID: 28186965 DOI: 10.18632/oncotarget.15120]
- 29 **Sedlackova T**, Repiska G, Celec P, Szemes T, Minarik G. Fragmentation of DNA affects the accuracy of the DNA quantitation by the commonly used methods. *Biol Proced Online* 2013; **15**: 5 [PMID: 23406353 DOI: 10.1186/1480-9222-15-5]

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Observational Study

Clinicopathological predictors of long-term benefit in breast cancer treated with neoadjuvant chemotherapy

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Institutional review board statement: This study was reviewed and approved by the Instituto Nacional de Enfermedades

Neoplasicas Institutional Review Board. Personal and filiation data including identity of every patient was protected with an added code in the Excel table. This is a retrospective case series that did not have any activity or contact with the patients.

Informed consent statement: Patients were not required to give informed consent to the study because the analysis used anonymous clinical data that were obtained after each patient agreed to treatment by written consent.

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Abstract

AIM

To investigate the survival impact of clinicopathological factors, including pathological complete response (pCR) and tumor-infiltrating lymphocytes (sTIL) levels according to subtypes, in breast cancer (BC) patients who received neo-adjuvant chemotherapy (NAC).

METHODS

We evaluated 435 BC patients who presented and received NAC at the Instituto Nacional de Enfermedades Neoplasias from 2003 to 2014. sTIL was analyzed as the proportion of tumor stroma occupied by lymphocytes, and was prospectively evaluated on hematoxylin and eosin-stained sections of the preNAC core biopsy. pCR was considered in the absence of infiltrating cancer cells in primary tumor and axillary lymph nodes. Analysis of statistical association between clinical pathological features, sTIL, pCR and survival were carried out using SPSSv19.

RESULTS

Median age was 49 years (range 24-84 years) and the most frequent clinical stage was III B (58.3%). Luminal A, Luminal B, HER2-enriched and (triple-negative) TN phenotype was found in 24.6%, 37.9%, 17.7% and 19.8%, respectively. pCR was observed in 11% and median percentage of sTIL was 40% (2%-95%) in the whole population. pCR was associated to Ct1-2 ($P = 0.045$) and to high sTIL ($P = 0.029$) in the whole population. There was a slight trend towards significance for sTIL ($P = 0.054$) in Luminal A. sTIL was associated with grade III ($P < 0.001$), no-Luminal A subtype ($P < 0.001$), RE-negative ($P < 0.001$), PgR-negative ($P < 0.001$), HER2-positive ($P = 0.002$) and pCR ($P = 0.029$) in the whole population. Longer disease-free survival was associated with grade I - II ($P = 0.006$), cN0 ($P < 0.001$), clinical stage II ($P = 0.004$), ER-positive ($P < 0.001$), PgR-positive ($P < 0.001$), luminal A ($P < 0.001$) and pCR ($P = 0.002$). Longer disease-free survival was associated with grade I - II in Luminal A ($P < 0.001$), N0-1 in Luminal A ($P = 0.045$) and TNBC ($P = 0.01$), clinical stage II in Luminal A ($P = 0.003$) and TNBC ($P = 0.038$), and pCR in TNBC ($P < 0.001$). Longer overall survival was associated with grade I - II ($P < 0.001$), ER-positive ($P < 0.001$), PgR-positive ($P < 0.001$), Luminal A ($P < 0.001$), cN0 ($P = 0.002$) and pCR ($P = 0.002$) in the whole population. Overall survival was associated with clinical stage II ($P = 0.017$) in Luminal A, older age ($P = 0.042$) in Luminal B, and pCR in TNBC ($P = 0.005$).

CONCLUSION

Predictive and prognostic values of clinicopathological features, like pCR and sTIL, differ depending on the evaluated molecular subtype.

Key words: Breast cancer; Subtype; Tumor-infiltrating lymphocytes; Neoadjuvant therapy; Pathological complete response; Survival

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Core tip: The authors evaluated a series of 435 breast cancer (BC) patients who received neoadjuvant chemotherapy. They evaluated the association between stromal tumor-infiltrating lymphocytes levels and pCR in preneoadjuvant chemotherapy samples according to molecular subtypes. The results confirm differences in the predictive and prognostic role of stromal tumor-infiltrating lymphocytes and pathological complete response depending on the tumor subtype. Additionally, the authors evaluate the value of traditional prognostic features in every BC subset. The results increase the understanding of biomarkers in the heterogeneous scenario of BC.

Galvez M, Castaneda CA, Sanchez J, Castillo M, Rebaza LP, Calderon G, De La Cruz M, Cotrina JM, Abugattas J, Dunstan J, Guerra H, Mejia O, Gomez HL. Clinicopathological predictors of long-term benefit in breast cancer treated with neoadjuvant chemotherapy. *World J Clin Oncol* 2018; 9(2): 33-41 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i2/33.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i2.33>

INTRODUCTION

Breast cancer (BC) is the second most common cancer in the world and the most frequent cancer among women, with an estimated 1.67 million new cancer cases diagnosed in 2012 (25% of all cancers), and is the fifth cause of death from cancer overall (522000 deaths)^[1]. Neoadjuvant chemotherapy (NAC) is the standard therapy for locally advanced BC and could improve both surgical options and long-term outcome^[2]. Response to NAC is considered an *in vivo* test of tumor sensitivity to NAC, and the achievement of a pathological complete response (pCR) is associated with longer disease-free survival (DFS) and greater overall survival (OS)^[3-7]. Tumor-infiltrating lymphocytes (TILs) serve to evaluate the host immune system response against a tumor and also constitutes a valuable predictive biomarker of NAC response and survival^[8-11].

BC is a heterogeneous disease, and intrinsically different subtypes of BC have been identified in the past years based on gene expression profiles and on the combined immunohistochemical status of hormone and HER2 receptors. Responsiveness to preoperative therapies and outcome after surgery can be predicted by BC subtypes^[12-14].

In this study, we investigated the survival impact of different clinicopathological factors, including pCR and TIL levels, according to the subtypes in BC patients who received NAC. The predictive role of different clinicopathological features for having high density TIL and obtaining pCR according to subtypes was also

determined.

MATERIALS AND METHODS

We found 435 patients diagnosed with BC at clinical stage II B to III C at the Medical Department of the Instituto Nacional de Enfermedades Neoplásicas from 2003 to 2014. Eligibility criteria for this retrospective study were a histological diagnosis based on a core needle biopsy, having received NAC regimen and having undergone surgery after NAC. Patient characteristics such as age, clinical stage, histological subtype and grade, presence of estrogen receptors (ERs), progesterone receptors (PgRs) and HER2, and molecular subtype was obtained from the pathology report of preNAC core biopsy. pCR was defined as absence of invasive cancer in the breast and axillary nodes, irrespective of carcinoma *in situ* (ypT0/is ypN0), as previously described^[4,15]. Phenotype classification was prospectively concluded through the evaluation of ER, PgR, HER2 and Ki67 as well as histological grade (in cases without Ki67 information): Luminal A (ER \geq 10%, PgR \geq 20%, HER2-negative and Ki67 < 15% or HG- I - II), Luminal B (ER \geq 10% and any PgR < 20%, HER2-positive, Ki67 < 15% or HG-III), HER2-enriched (ER < 10%, PgR < 10% and HER2-positive) and triple-negative (TN) (ER < 10%, PgR < 10% and HER2-negative). Stromal (s)TIL was prospectively evaluated in preNAC core biopsy and was defined as percentage of stromal area covered by lymphocytes^[16].

Follow-up and recurrence information (date and location) were obtained from patient files. Time-from-last-chemotherapy-to-surgery was considered as the number of months from the date of the last NAC administration to surgery of the primary tumor. OS was calculated from surgery date of the primary breast tumor to death or last follow-up date, and DFS was calculated from surgery date of the primary breast tumor to recurrence or last follow-up date.

Statistical analysis

Categorical comparisons and association analysis between clinical pathological features and pCR were carried out using the chi-square statistic or Fisher's exact test. Survival analysis, regarding OS and DFS, was performed using the Kaplan-Meier method, and differences between curves were estimated by log-rank test. In all cases, the level of alpha was set at 0.05 *a priori*. Statistical analysis was performed using SPSS v19 (IBM Corp., Armonk, NY, United States).

RESULTS

Clinicopathological description

There were 435 patients included in this study, with median age at diagnosis of 49 years (range: 24-84 years), median tumor size of 6.5 cm (range: 1.0-24.0 cm), T3 in 27.8% and T4 in 63.9%. Inflammatory

disease was found in 29.2%. The most frequent clinical stages were III B (60.5%) and III A (18.6%). Ductal histology was found in 93.3%, high grade in 52.2%, ER+ status in 62.8%, PgR+ status in 51% and HER2+++ in 32.4%. Luminal A, Luminal B, HER2-enriched and TN phenotype was found in 24.6%, 37.9%, 17.7% and 19.8%, respectively. The most frequent NACs were doxorubicin-cyclophosphamide for 4 cycles followed by 12 weekly paclitaxel (67.18%), doxorubicin-cyclophosphamide for 4 cycles followed by every 3 wk paclitaxel in 4 cycles (18.85%) and doxorubicin-cyclophosphamide for 4 cycles alone (7.32%). The median time from the last chemotherapy to surgery was 63 d (maximum: 982 d). pCR was observed in 48 (11%) patients. Median percentage of sTILs was 40% (2%-95%) in the entire population and 70% (60%-95%) in patients with pCR. Recurrence was found in 35.7%. Median DFS was 7.54 and median OS was 5.16 years (95%CI: 4.16-6.15 years) (Table 1).

Clinicopathological factors associated to pCR according to BC subtypes

Association analysis found that pCR was associated with T1-2 ($P = 0.045$) and to high sTIL level ($P = 0.029$) in the entire population (Table 1). Higher sTIL level had a slight trend towards association with pCR ($P = 0.054$) in Luminal A, and smaller tumor size had a trend towards association with pCR ($P = 0.098$) in Luminal A. Clinical involvement of axillary lymph nodes was not associated to variation of pCR (Table 2). An additional analysis by level of axillary involvement found that N2-3 had lower rates of pCR than N0-1 only in TNBC ($P = 0.018$).

Clinicopathological factors associated with sTIL according to BC subtypes

Association analysis found that sTIL level was associated with grade III ($P < 0.001$), no-Luminal A subtype ($P < 0.001$), ER-negative ($P < 0.001$), PgR-negative ($P < 0.001$), HER2-positive ($P = 0.002$) and pCR ($P = 0.029$) in the entire population (Table 1). Within each BC subtype, sTIL level remained associated with grade III in Luminal B ($P = 0.011$) and TN ($P = 0.006$) subtypes, as well as cN+ in Luminal B ($P = 0.02$) (Table 3).

Prognostic clinicopathological factors according to BC subtypes

Survival analysis found longer DFS was associated with grade I - II ($P = 0.006$), cN0 ($P < 0.001$), clinical stage II ($P = 0.004$), ER-positive ($P < 0.001$), PgR-positive ($P < 0.001$), Luminal A ($P < 0.001$) and pCR ($P = 0.002$). Longer DFS was associated with grade I - II in Luminal A ($P = 0.033$), N0-1 in Luminal A ($P = 0.045$) and TNBC ($P = 0.01$), clinical stage II in Luminal A ($P = 0.003$) and TNBC ($P = 0.038$), and pCR in TNBC ($P = 0.001$) (Table 1).

Table 1 Clinical-pathological features *n* (%)

	Cases 435	sTIL ≥ 50% 181	<i>P</i> value	pCR 48	<i>P</i> value	Overall Survival at 5 yr (OS = 50.1%)	<i>P</i> value	Progression free survival at 5 yr (DFS = 57.8%)	<i>P</i> value
Age (yr), median (range)	49 (24-84)	49 (24-84)	0.923	47 (28-80)	0.472		0.512		0.833
< 50	231 (53.1)	96 (35.2)		28 (12.1)		48.8%		59.7%	
≥ 50	204 (46.9)	85 (36.7)		20 (9.8)		51.7%		55.9%	
Histological subtypes			0.928		0.234		0.512		0.497
Ductal	406 (93.3)	169 (43.6)		43 (10.6)		49.0%		57.5%	
Lobular	21 (4.8)	7 (3.6.8)		2 (9.5)		61.0%		55.2%	
Others	8 (1.8)	5 (6.2.5)		3 (37.5)		-		-	
Histological grade			< 0.001		0.170		0.001		0.006
G1-G2	200 (46.0)	59 (32.6)		17 (8.5)		57.1%		64.6%	
G3	227 (52.2)	119 (65.7)		29 (12.8)		42.8%		52.2%	
NR	8 (1.8)	3 (1.7)		2 (2.5)		83.3%		45.7%	
ER			< 0.001		0.098		< 0.001		0.000
No	162 (37.2)	89 (57.8)		23 (14.2)		36.1%		47.1%	
Yes	273 (62.8)	92 (35.2)		25 (9.2)		58.2%		64.3%	
PgR			0.003		0.246		< 0.001		0.000
No	213 (49)	104 (51.0)		27 (12.7)		41.0%		50.0%	
Yes	222 (51)	77 (36.5)		21 (9.5)		58.4%		64.8%	
HER2			0.002		0.135		0.334		0.135
No	294 (67.6)	106 (38.3)		28 (9.5)		53.7%		60.4%	
Yes	141 (32.4)	75 (54.3)		20 (14.2)		40.8%		52.3%	
Molecular subtypes			< 0.001		0.233		< 0.001		< 0.001
Luminal A	107 (24.6)	30 (29.7)		13 (12)		72.0%		76.1%	
Luminal B	165 (37.9)	61 (38.4)		12 (7)		50.6%		57.7%	
HER2-enriched	77 (17.7)	50 (66.7)		10 (13)		41.5%		54.9%	
Triple-Negative	86 (19.8)	40 (50.0)		13 (15)		32.5%		40.3%	
Tumor size (cm)			0.183		0.019		0.490		0.250
Median (range)	6.5 (1-24)	6.5 (1-16)		6.0 (2-15)					
cT									
cT1-cT2	36 (8.3)	19 (54.3)		8 (22.2)		55.0%		69.2%	
cT3-cT4	399 (91.7)	162 (42.6)		40 (10)		49.6%		56.8%	
cN			0.084		0.743		0.007		0.001
cN0	83 (19.1)	28 (35.0)		10 (12)		65.8%		77.0%	
cN1-cN2-cN3	352 (80.9)	153 (45.7)		38 (10.8)		47.2%		54.2%	
Clinical stage			0.192		0.088		0.155		0.004
II	72 (16.6)	26 (36.6)		12 (16.7)		62.1%		74.3%	
III	363 (83.4)	155 (45.1)		36 (9.9)		48.1%		55.4%	
sTIL %					0.002		0.598		0.747
Median (range)	40 (2-95)	70 (60-95)		65 (5-95)					
< 50%	266 (61.1)	0 (0)		20 (7.5)		49.6%		55.7%	
≥ 50%	149 (34.3)	181 (100)		26 (17.4)		53.9%		63.1%	
Missing data	20 (4.6)	20 (0)		2 (10)		-		-	
TLCS (d)			0.411		0.633		0.317		0.156
Median (range)	63 (5-982)	58 (8-982)		65 (8-281)					
Shorter than median	207 (47.6)	91 (45.5)		22 (10.6)		48.5%		55.0%	
Longer than median	211 (48.5)	82 (41.4)		26 (12.3)		56.7%		61.2%	
Missing data	17 (3.9)	8 (47.1)		0 (0)		17.6%		46.3%	
pCR			0.029				0.002		0.002
No	387 (89)	154 (41.7)		0 (0)		47.4%		55.1%	
Yes	48 (11)	27 (58.7)		48 (100)		85.1%		84.9%	
Relapse			0.895		< 0.001		< 0.001		
No	284 (65.3)	118 (43.4)		42 (14.8)		81.6%		-	
Yes	151 (34.7)	63 (44.1)		6 (4)		8.58%		-	

TIL: Tumor-infiltrating lymphocytes; pCR: Pathological complete response; OS: Overall survival; DFS: Disease free survival; PgR: Progesterone; TLCS: Time-From-Last-Chemotherapy-To-Surgery.

Longer OS was associated with grade I - II ($P < 0.001$), ER-positive ($P < 0.001$), PgR-positive ($P < 0.001$), Luminal A ($P < 0.001$), cN0 ($P = 0.007$) and pCR ($P = 0.002$) in the entire population. It was also associated with older age in Luminal B ($P = 0.042$), to clinical stage II in Luminal A ($P = 0.017$), and to cN0 ($P = 0.045$) and pCR in TNBC ($P = 0.005$) (Figure 1). Differences in TILs did not affect survival in the entire

nor molecular subtype populations (Table 1 and Figure 2).

DISCUSSION

The biological heterogeneity of BC has been extensively described, and differences between intrinsic subtypes have been confirmed in the recent decade. We explored differences in the survival impact

Table 2 Association between response and Clinical-pathological features regarding molecular subtype *n* (%)

	Lum A			Lum B			HER2			TN		
	Total 107	pCR 13	<i>P</i> value	Total 165	pCR 12	<i>P</i> value	Total 77	pCR 10	<i>P</i> value	Total 86	pCR 13	<i>P</i> value
Age (yr)			1.000			0.315			0.507			0.157
median (range)	47 (28-75)	46 (28-62)		51 (25-84)	52 (39-69)		51 (28-80)	46 (29-80)		49 (26-73)	45 (28-68)	
< 50	72 (67)	9 (13)		78 (48)	4 (5)		37 (48)	6 (16.2)		44 (48)	9 (20)	
≥ 50	35 (33)	4 (11)		87 (52)	8 (9)		40 (52)	4 (10)		42 (52)	4 (10)	
Histological subtypes			0.349			1.000			0.434			0.392
Ductal	97 (91)	11 (11)		153 (93)	11 (7)		73 (95)	9 (12.3)		83 (97)	12 (14)	
Lobular and others	10 (9)	2 (20)		12 (7)	1 (8)		4 (5)	1 (25)		3 (3)	1 (33)	
Histological grade			-			0.213			0.266			1.000
G1-G2	103 (97)	12 (12)		61 (39)	2 (3)		23 (30)	1 (4.3)		13 (15)	2 (15)	
G3	-	-		102 (61)	10 (10)		53 (69)	9 (17)		72 (85)	10 (14)	
NR	4 (3)	1 (25)		2 (1)	0 (0)		1 (1)	0 (0)		1 (0)	1 (100)	
Tumor size (cm)			0.102			0.213			0.511			0.620
Median	6 (2-15)	5 (2-9)		7 (2-20)	6 (2-12)		7 (2.5-14)	6 (4-12)		7 (1-24)	8 (3-15)	
(range)												
cT1-cT2	10 (7)	3 (30)		12 (7)	2 (17)		5 (6)	1 (20)		9 (10)	2 (22)	
cT3-cT4	97 (93)	10 (10)		153 (93)	10 (7)		72 (94)	9 (12.5)		77 (90)	11 (14)	
cN			0.306			0.222			0.270			0.021
cN0	27 (23)	5 (19)		28 (18)	0 (0)		53 (69)	5 (9.4)		14 (14)	4 (29)	
cN1-cN2-cN3	80 (77)	8 (10)		137 (82)	12 (9)		24 (31)	5 (20.8)		72 (86)	9 (13)	
Clinical stage			0.471			0.652			1.000			0.122
EC II	23 (20)	4 (17)		21 (12)	2 (10)		11 (14)	1 (9.1)		17 (16)	5 (29)	
EC III	84 (80)	9 (11)		144 (88)	10 (7)		66 (86)	9 (13.6)		69 (84)	8 (12)	
sTIL%			0.054			0.750			0.150			1.000
Median (range)	30 (2-90)	50 (10-90)		40 (5-90)	30 (8-90)		60 (5-95)	80 (30-95)		45 (2-90)	50 (5-80)	
< 50	71 (69)	6 (8)		98 (60)	6 (6)		25 (32)	1 (4)		40 (47)	6 (15)	
≥ 50	30 (24)	7 (23)		61 (37)	5 (8)		50 (66)	9 (18)		40 (47)	6 (15)	
Missing data	6 (6)	0 (0)		6 (3)	1 (17)		2 (3)	0 (0)		6 (7)	1 (17)	
TLCs (d)			0.233			0.238			0.744			0.500
Median (range)	67 (14-458)	80 (16-281)		61 (5-412)	54 (8-140)		60 (11-240)	66 (37-106)		64 (8-982)	66 (14-122)	
Shorter than median	49 (48)	4 (8)		77 (45)	8 (10)		41 (53)	5 (12.2)		40 (48)	5 (13)	
Longer than median	57 (51)	9 (16)		76 (47)	4 (5)		33 (43)	5 (15.2)		45 (51)	8 (18)	
Missing data	1 (1)	0 (0)		12 (8)	0 (0)		3 (4)	0 (0)		1 (1)	0 (0)	
Relapse			0.121			0.753			0.300			< 0.001
No	87 (79)	13 (15)		109 (65)	9 (8)		46 (60)	8 (17.4)		42 (41)	12 (29)	
Yes	20 (21)	0 (0)		56 (35)	3 (5)		31 (40)	2 (6.5)		44 (59)	1 (2)	

TIL: Tumor-infiltrating lymphocytes; TLCs: Time-From-Last-Chemotherapy-To-Surgery.

of tumor features, including pCR and TIL levels in each of the four molecular subtypes. Rates of pCR are lower in Luminal-A (9.2%), HER2-enriched (13%) and TNBC (15.3%) subtypes. pCR is also associated with longer survival in the entire population as well as in TNBC (pCR = 92.3% vs not pCR = 26.5% 5-year OS, $P = 0.005$; and trend in Luminal A, Luminal B and HER2-enriched phenotypic subsets of our series). It is widely assumed that patients who achieve pCR have significantly better DFS and OS rates in all molecular subtypes^[12-14,17-19]. von Minckwitz *et al*^[6] found pCR was not associated with prognosis only in Luminal A tumors in a series of 6377 patients with anthracycline-taxane-based NAC from 7 randomized trials; some authors claim it is related to the observed continuous tumor shrinkage occurring in their ER-positive tumor group during extended NAC, different than early and short-

period tumor shrinkage observed in the ER-negative group^[6,18-24].

pCR was more frequent in small tumors for both the entire population and the Luminal A subtype in our series. This finding is concordant with the previously mentioned idea that the effect of chemotherapy in Luminal A is slower than in other subtypes. Besides, Baron *et al*^[18] found a similar lower rate of pCR in tumor size larger than 5 cm ($P = 0.022$) in their entire series ($n = 608$), but no association in the Luminal setting ($P = 0.411$). Higher grade of axillary involvement (cN2-3) was associated with lower rates of pCR only in the TNBC subset of our series. This lower response in bulky metastases could explain the previously described TNBC paradox phenomena of higher pCR rates but also higher distant relapse^[21].

pCR was associated with higher percentage of

Table 3 Association between percentage of tumor-infiltrating lymphocytes and clinical-pathological features regarding molecular subtype *n* (%)

	Lum A			Lum B			HER2			TN		
	< 50%	≥ 50%	<i>P</i> value	< 50%	≥ 50%	<i>P</i> value	< 50%	≥ 50%	<i>P</i> value	< 50%	≥ 50%	<i>P</i> value
	71	30		98	61		25	50		40	40	
Age (yr)			0.181			0.783			0.624			0.074
Median (range)	47 (28-75)	47 (36-74)		52 (28-73)	50 (25-84)		52 (28-66)	49 (29-80)		51 (26-73)	45 (27-73)	
< 50	50 (70)	17 (57)		46 (47)	30 (49)		11 (44)	25 (50)		16 (40)	24 (60)	
≥ 50	21 (30)	13 (43)		52 (53)	31 (51)		14 (56)	25 (50)		24 (60)	16 (40)	
Histological subtypes			0.445			1.000			0.597			1.000
Ductal	66 (93)	26 (87)		91 (93)	57 (93)		23 (92)	48 (96)		39 (98)	38 (95)	
Lobular and others	5 (7)	4 (13)		7 (7)	4 (7)		2 (8)	2 (4)		1 (3)	2 (5)	
Histological grade			-			0.011			0.514			0.006
G1-G2	69 (97)	28 (93)		43 (44)	15 (25)		9 (36)	14 (28)		11 (28)	2 (5)	
G3	0 (0)	0 (0)		53 (54)	46 (75)		16 (64)	35 (71)		29 (73)	38 (95)	
NR	2 (3)	2 (7)		2 (2)	0 (0)		0 (0)	1 (2)		0 (0)	0 (0)	
Tumor size (cm)												
Median (range)	6 (3-13)	6 (2-15)		6 (3-20)	7 (2-15)		7 (3-14)	7 (3-14)		7 (4-24)	7 (1-16)	
cT			1.000			0.538			0.659			0.263
cT1-cT2	7 (10)	3 (10)		6 (6)	6 (10)		1 (4)	4 (8)		2 (5)	6 (15)	
cT3-cT4	64 (90)	27 (90)		92 (94)	55 (90)		24 (96)	46 (92)		38 (95)	34 (85)	
cN			0.890			0.020			0.631			0.762
cN0	18 (25)	8 (27)		22 (22)	5 (8)		6 (24)	8 (16)		6 (15)	7 (18)	
cN1-cN2-cN3	53 (75)	22 (73)		76 (78)	56 (92)		11 (44)	27 (54)		34 (85)	33 (83)	
Clinical Stage			0.666			0.141			0.742			0.576
EC II	17 (24)	6 (20)		16 (16)	5 (8)		3 (12)	8 (16)		9 (23)	7 (18)	
EC III	54 (76)	24 (80)		82 (84)	56 (92)		22 (88)	42 (84)		31 (78)	33 (83)	
TLCS (d)			0.631			0.882			0.502			0.141
Median (range)	64 (14-449)	70 (19-458)		61 (5-412)	58 (8-285)		68 (16-234)	56 (11-240)		74 (24-230)	51 (14-982)	
Shorter than median	34 (48)	13 (43)		48 (49)	28 (46)		12 (48)	28 (56)		15 (38)	22 (55)	
Longer than median	36 (51)	17 (57)		44 (45)	27 (44)		12 (48)	20 (40)		24 (60)	18 (45)	
Missing data	1 (1)	0 (0)		6 (6)	6 (10)		1 (4)	2 (4)		1 (3)	0 (0)	
pCR			0.054			0.750			0.150			1.000
No	65 (92)	23 (77)		92 (94)	56 (92)		24 (96)	41 (82)		34 (85)	34 (85)	
Yes	6 (8)	7 (23)		6 (6)	5 (8)		1 (4)	9 (18)		6 (15)	6 (15)	
Relapse			0.450			0.201			0.737			0.502
No	59 (83)	23 (77)		61 (62)	44 (72)		16 (64)	30 (60)		18 (45)	21 (53)	
Yes	12 (17)	7 (23)		37 (38)	17 (28)		9 (36)	20 (40)		22 (55)	19 (48)	

%sTIL was performed over 415 cases. There 20 missed values. TIL: Tumor-infiltrating lymphocytes; TLCS: Time-From-Last-Chemotherapy-To-Surgery.

sTILs in the entire population and also within the HER2-enriched subtype ($P = 0.02$). A trend towards association was found in Luminal A, Luminal B and TNBC. Different studies have found that high TIL levels in preNAC samples are associated to higher pCR rates in the entire BC population^[25-27]. Wang *et al.*^[28] performed a meta-analysis with 23 studies including 13100 BC patients, and similarly found that high TIL level was associated with improved pCR rate in the entire population, and in HER2 and TNBC. A high TIL level significantly predicted longer OS in the entire population ($P < 0.001$) and in patients with HER2-positive ($P = 0.005$) BC and in TNBC patients ($P < 0.001$).

TIL showed association with grade III tumors in the entire population and in Luminal B and TNBC subsets in our series. Similarly, Pruneri *et al.*^[29] describes that higher TIL levels have a trend towards association with HG3 ($P = 0.052$) and was associated to Ki67 \geq

50% ($P < 0.0001$) in a series of 897 TNBC cases, and could reflect the appearance of a larger amount of neoantigens that elicit an immunomediated response. Involvement of axillary lymph nodes was associated to higher TIL levels only in the Luminal B subset. High density of TILs has previously been described as associated to absence of lymph node involvement in the entire population of BC, and our results indicate that this association could differ by some subtypes^[30]. Higher level of sTILs was not associated to longer survival in the entire population nor in any subtype in our series. This finding could be explained by the small size of our series and because the highest impact of TILs is over pCR instead of survival.

Our study has some limitations. First, because of the retrospective design of the study, different chemotherapy schemas were used depending on the oncologist decision and surgical election depending

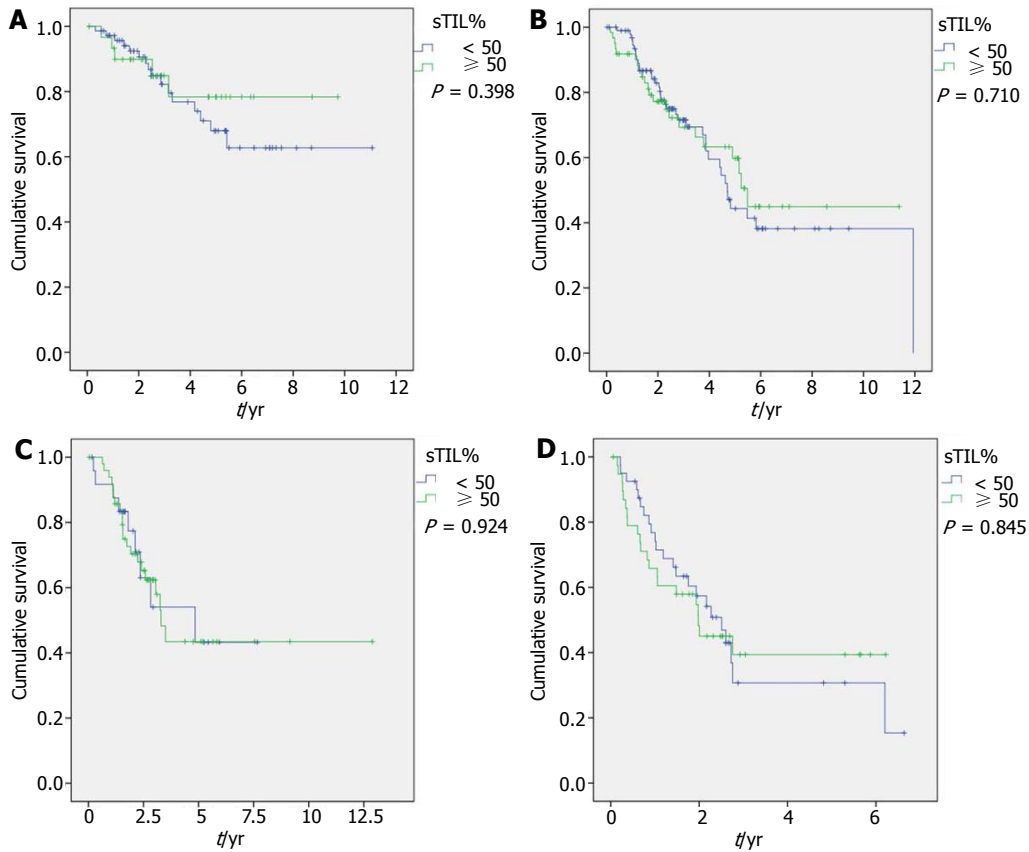


Figure 1 Overall survival regarding tumor-infiltrating lymphocytes (cut-off: 50%) for Luminal A (A), Luminal B (B), HER2-enriched (C) and Triple Negative group (D).

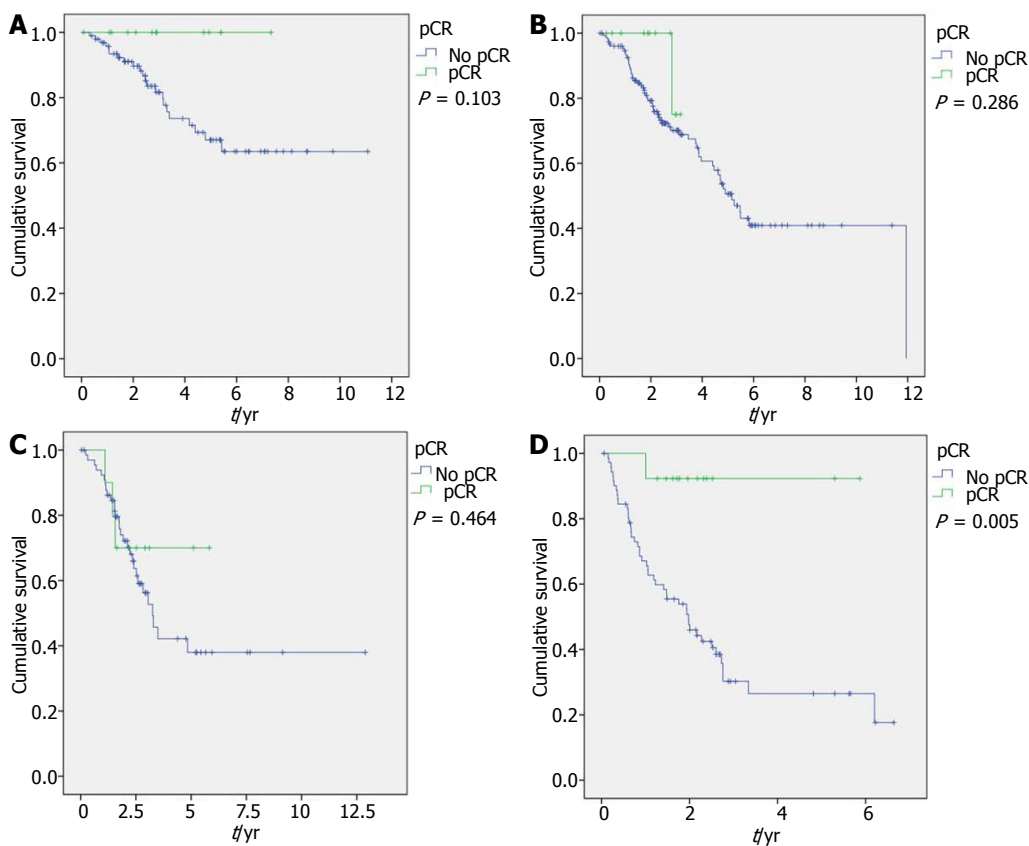


Figure 2 Overall survival regarding pathological complete response for Luminal A (A), Luminal B (B), HER2-enriched (C) and triple negative group (D).

on surgeon. Second, the sample sizes of each BC subgroup are rather small, so the prognostic impact of every clinicopathological feature in each BC subtype should be investigated in a larger population in subsequent studies. Despite these limitations, this is the first comprehensive report of the NAC effect over breast molecular subtypes in a Latin-American population.

ARTICLE HIGHLIGHTS

Research background

Breast cancer can be classified into Luminal A, Luminal B, HER2-enriched and triple-negative. Clinicopathological features can identify breast cancer prognosis and include pathological complete response (tumor sensibility to chemotherapy) and tumor-infiltrating lymphocytes (TILs; host activity against the tumor).

Research motivation

Discussion and new information about molecular breast cancer subtypes have been included in the most relevant cancer-related meeting, and more than 30,000 articles have been published in the last 2 years. Two biomarkers, pathological complete response (pCR) and TILs, have been re-defined and gained pathologist acceptance in the last 3 years.

Research objectives

The main objective is to evaluate the survival impact of different clinicopathological factors, including pCR and TIL levels, according to the subtypes in breast cancer patients who received neoadjuvant chemotherapy.

Research methods

Evaluation of TIL levels was prospectively performed following international guidelines. Breast cancer cases were classified according to 2017 St Gallen Breast Cancer Meeting guidelines.

Research results

pCR was associated with cT1-2 ($P = 0.045$) and high stromal (s)TILs ($P = 0.029$) in the entire population. However, this relationship was not found for every molecular subtype, probably because of the small sample size. pCR was associated with longer disease-free survival in the entire population ($P = 0.002$) and in TNBC ($P < 0.001$), as well as to longer overall survival in the entire population ($P = 0.002$) and in TNBC ($P = 0.005$).

Research conclusions

Predictive and prognostic value of clinicopathological features like pCR and sTIL level differ depending on the molecular subtype being evaluated. Identification of pCR and TIL roles in every molecular subtype will allow for identification of those patients who need more intense chemotherapy and those who will benefit from an immune-modulator treatment.

Research perspectives

No information about the relevance of pCR and TILs in South-American women with breast cancer have been published in. An increase in the knowledge about prognosis impact of pCR and TIL in every molecular breast cancer subtype will allow for obtaining more effective personalized therapies. Furthermore, similar analysis needs to be done with more precise methods to evaluate response to chemotherapy and host immune activity, such as tumor residual burden and CD3/CD8 ratio, respectively.

REFERENCES

- 1 **Carbognin L**, Pilotto S, Nortilli R, Brunelli M, Nottegar A, Sperduti I, Giannarelli D, Bria E, Tortora G. Predictive and Prognostic Role of Tumor-Infiltrating Lymphocytes for Early Breast Cancer According to Disease Subtypes: Sensitivity Analysis of Randomized Trials in Adjuvant and Neoadjuvant Setting. *Oncologist* 2016; **21**: 283-291 [PMID: 26865589 DOI: 10.1634/theoncologist.2015-0307]
- 2 **Vila J**, Mittendorf EA, Farante G, Bassett RL, Veronesi P, Galimberti V, Peradze N, Stauder MC, Chavez-MacGregor M, Litton JF, Huo L, Kuerer HM, Hunt KK, Caudle AS. Nomograms for Predicting Axillary Response to Neoadjuvant Chemotherapy in Clinically Node-Positive Patients with Breast Cancer. *Ann Surg Oncol* 2016; **23**: 3501-3509 [PMID: 27216742 DOI: 10.1245/s10434-016-5277-1]
- 3 **Issa-Nummer Y**, Loibl S, von Minckwitz G, Denkert C. Tumor-infiltrating lymphocytes in breast cancer: A new predictor for responses to therapy. *Oncoimmunology* 2014; **3**: e27926 [PMID: 25340002 DOI: 10.4161/onci.27926]
- 4 **Cortazar P**, Zhang L, Untch M, Mehta K, Costantino JP, Wolmark N, Bonnefoi H, Cameron D, Gianni L, Valagussa P, Swain SM, Prowell T, Loibl S, Wickerham DL, Bogaerts J, Baselga J, Perou C, Blumenthal G, Blohmer J, Mamounas EP, Bergh J, Semiglazov V, Justice R, Eidtmann H, Paik S, Piccart M, Sridhara R, Fasching PA, Slaets L, Tang S, Gerber B, Geyer CE Jr, Pazdur R, Ditsch N, Rastogi P, Eiermann W, von Minckwitz G. Pathological complete response and long-term clinical benefit in breast cancer: the CTNeoBC pooled analysis. *Lancet* 2014; **384**: 164-172 [PMID: 24529560 DOI: 10.1016/S0140-6736(13)62422-8]
- 5 **Bossuyt V**, Provenzano E, Symmans WF, Boughey JC, Coles C, Curigliano G, Dixon JM, Esserman LJ, Fastner G, Kuehn T, Peintinger F, von Minckwitz G, White J, Yang W, Badve S, Denkert C, MacGrogan G, Penault-Llorca F, Viale G, Cameron D; Breast International Group-North American Breast Cancer Group (BIG-NABCG) collaboration. Recommendations for standardized pathological characterization of residual disease for neoadjuvant clinical trials of breast cancer by the BIG-NABCG collaboration. *Ann Oncol* 2015; **26**: 1280-1291 [PMID: 26019189 DOI: 10.1093/annonc/mdv161]
- 6 **von Minckwitz G**, Untch M, Blohmer JU, Costa SD, Eidtmann H, Fasching PA, Gerber B, Eiermann W, Hilfrich J, Huober J, Jackisch C, Kaufmann M, Konecny GE, Denkert C, Nekljudova V, Mehta K, Loibl S. Definition and impact of pathologic complete response on prognosis after neoadjuvant chemotherapy in various intrinsic breast cancer subtypes. *J Clin Oncol* 2012; **30**: 1796-1804 [PMID: 22508812 DOI: 10.1200/JCO.2011.38.8595]
- 7 **Denkert C**, Loibl S, Noske A, Roller M, Müller BM, Komor M, Budczies J, Darb-Esfahani S, Kronenwett R, Hanusch C, von Törne C, Weichert W, Engels K, Solbach C, Schrader I, Dietel M, von Minckwitz G. Tumor-associated lymphocytes as an independent predictor of response to neoadjuvant chemotherapy in breast cancer. *J Clin Oncol* 2010; **28**: 105-113 [PMID: 19917869 DOI: 10.1200/JCO.2009.23.7370]
- 8 **Ruffini E**, Asioli S, Filosso PL, Lyberis P, Bruna MC, Macri L, Daniele L, Oliaro A. Clinical significance of tumor-infiltrating lymphocytes in lung neoplasms. *Ann Thorac Surg* 2009; **87**: 365-371; discussion 371-372 [PMID: 19161739 DOI: 10.1016/j.athoracsurg.2008.10.067]
- 9 **Dushyanthen S**, Beavis PA, Savas P, Teo ZL, Zhou C, Mansour M, Darcy PK, Loi S. Relevance of tumor-infiltrating lymphocytes in breast cancer. *BMC Med* 2015; **13**: 202 [PMID: 26300242 DOI: 10.1186/s12916-015-0431-3]
- 10 **Loi S**. Tumor-infiltrating lymphocytes, breast cancer subtypes and therapeutic efficacy. *Oncoimmunology* 2013; **2**: e24720 [PMID: 24073365 DOI: 10.4161/onci.24720]
- 11 **Stanton SE**, Adams S, Disis ML. Variation in the Incidence and Magnitude of Tumor-Infiltrating Lymphocytes in Breast Cancer Subtypes: A Systematic Review. *JAMA Oncol* 2016; **2**: 1354-1360 [PMID: 27355489 DOI: 10.1001/jamaoncol.2016.1061]
- 12 **Perou CM**, Sørli E, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lønning PE, Børresen-Dale AL, Brown PO, Botstein D. Molecular portraits of human breast tumours. *Nature* 2000; **406**: 747-752 [PMID: 10963602 DOI: 10.1038/35021093]
- 13 **Rouzier R**, Perou CM, Symmans WF, Ibrahim N, Cristofanilli M, Anderson K, Hess KR, Stec J, Ayers M, Wagner P, Morandi P, Fan C, Rabiul I, Ross JS, Hortobagyi GN, Pusztai L. Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clin Cancer Res* 2005; **11**: 5678-5685 [PMID: 16115903 DOI: 10.1158/1078-0432.CCR-05-0001]

- 10.1158/1078-0432.CCR-04-2421]
- 14 **Hugh J**, Hanson J, Cheang MC, Nielsen TO, Perou CM, Dumontet C, Reed J, Krajewska M, Treilleux I, Rupin M, Magherini E, Mackey J, Martin M, Vogel C. Breast cancer subtypes and response to docetaxel in node-positive breast cancer: use of an immunohistochemical definition in the BCIRG 001 trial. *J Clin Oncol* 2009; **27**: 1168-1176 [PMID: 19204205 DOI: 10.1200/JCO.2008.18.1024]
 - 15 **Pennisi A**, Kieber-Emmons T, Makhoul I, Hutchins L. Relevance of Pathological Complete Response after Neoadjuvant Therapy for Breast Cancer. *Breast Cancer (Auckl)* 2016; **10**: 103-106 [PMID: 27478380 DOI: 10.4137/bcbr.s33163]
 - 16 **Salgado R**, Denkert C, Demaria S, Sirtaine N, Klauschen F, Pruneri G, Wienert S, Van den Eynden G, Baehner FL, Penault-Llorca F, Perez EA, Thompson EA, Symmans WF, Richardson AL, Brock J, Criscitiello C, Bailey H, Ignatiadis M, Floris G, Sparano J, Kos Z, Nielsen T, Rimm DL, Allison KH, Reis-Filho JS, Loibl S, Sotiriou C, Viale G, Badve S, Adams S, Willard-Gallo K, Loi S; International TILs Working Group 2014. The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: recommendations by an International TILs Working Group 2014. *Ann Oncol* 2015; **26**: 259-271 [PMID: 25214542 DOI: 10.1093/annonc/mdl450]
 - 17 **Colleoni M**, Bagnardi V, Rotmensz N, Dellapasqua S, Viale G, Pruneri G, Veronesi P, Torrisi R, Luini A, Intra M, Galimberti V, Montagna E, Goldhirsch A. A risk score to predict disease-free survival in patients not achieving a pathological complete remission after preoperative chemotherapy for breast cancer. *Ann Oncol* 2009; **20**: 1178-1184 [PMID: 19218304 DOI: 10.1093/annonc/mdn747]
 - 18 **Baron P**, Beitsch P, Boselli D, Symanowski J, Pellicane JV, Beatty J, Richards P, Mislowsky A, Nash C, Lee LA, Murray M, de Snoo FA, Stork-Sloots L, Gittleman M, Akbari S, Whitworth P. Impact of Tumor Size on Probability of Pathologic Complete Response After Neoadjuvant Chemotherapy. *Ann Surg Oncol* 2016; **23**: 1522-1529 [PMID: 26714960 DOI: 10.1245/s10434-015-5030-1]
 - 19 **Symmans WF**, Wei C, Gould R, Yu X, Zhang Y, Liu M, Walls A, Bousamra A, Ramineni M, Sinn B, Hunt K, Buchholz TA, Valero V, Buzdar AU, Yang W, Brewster AM, Moulder S, Pusztai L, Hatzis C, Hortobagyi GN. Long-Term Prognostic Risk After Neoadjuvant Chemotherapy Associated With Residual Cancer Burden and Breast Cancer Subtype. *J Clin Oncol* 2017; **35**: 1049-1060 [PMID: 28135148 DOI: 10.1200/JCO.2015.63.1010]
 - 20 **Guarneri V**, Broglio K, Kau SW, Cristofanilli M, Buzdar AU, Valero V, Buchholz T, Meric F, Middleton L, Hortobagyi GN, Gonzalez-Angulo AM. Prognostic value of pathologic complete response after primary chemotherapy in relation to hormone receptor status and other factors. *J Clin Oncol* 2006; **24**: 1037-1044 [PMID: 16505422 DOI: 10.1200/JCO.2005.02.6914]
 - 21 **Carey LA**, Dees EC, Sawyer L, Gatti L, Moore DT, Collichio F, Ollila DW, Sartor CI, Graham ML, Perou CM. The triple negative paradox: primary tumor chemosensitivity of breast cancer subtypes. *Clin Cancer Res* 2007; **13**: 2329-2334 [PMID: 17438091 DOI: 10.1158/1078-0432.CCR-06-1109]
 - 22 **Bear HD**, Anderson S, Brown A, Smith R, Mamounas EP, Fisher B, Margolese R, Theoret H, Soran A, Wickerham DL, Wolmark N; National Surgical Adjuvant Breast and Bowel Project Protocol B-27. The effect on tumor response of adding sequential preoperative docetaxel to preoperative doxorubicin and cyclophosphamide: preliminary results from National Surgical Adjuvant Breast and Bowel Project Protocol B-27. *J Clin Oncol* 2003; **21**: 4165-4174 [PMID: 14559892 DOI: 10.1200/JCO.2003.12.005]
 - 23 **Fisher B**, Bryant J, Wolmark N, Mamounas E, Brown A, Fisher ER, Wickerham DL, Begovic M, DeCillis A, Robidoux A, Margolese RG, Cruz AB Jr, Hoehn JL, Lees AW, Dimitrov NV, Bear HD. Effect of preoperative chemotherapy on the outcome of women with operable breast cancer. *J Clin Oncol* 1998; **16**: 2672-2685 [PMID: 9704717 DOI: 10.1200/JCO.1998.16.8.2672]
 - 24 **Moon HG**, Im SA, Han W, Oh DY, Han SW, Keam B, Park IA, Chang JM, Moon WK, Cho N, Noh DY. Estrogen receptor status confers a distinct pattern of response to neoadjuvant chemotherapy: implications for optimal durations of therapy: distinct patterns of response according to ER expression. *Breast Cancer Res Treat* 2012; **134**: 1133-1140 [PMID: 22752292 DOI: 10.1007/s10549-012-2145-y]
 - 25 **Mao Y**, Qu Q, Chen X, Huang O, Wu J, Shen K. The Prognostic Value of Tumor-Infiltrating Lymphocytes in Breast Cancer: A Systematic Review and Meta-Analysis. *PLoS One* 2016; **11**: e0152500 [PMID: 27073890 DOI: 10.1371/journal.pone.0152500]
 - 26 **Krishnamurti U**, Wetherilt CS, Yang J, Peng L, Li X. Tumor-infiltrating lymphocytes are significantly associated with better overall survival and disease-free survival in triple-negative but not estrogen receptor-positive breast cancers. *Hum Pathol* 2017; **64**: 7-12 [PMID: 28153508 DOI: 10.1016/j.humpath.2017.01.004]
 - 27 **Luen SJ**, Salgado R, Fox S, Savas P, Eng-Wong J, Clark E, Kiermaier A, Swain SM, Baselga J, Michiels S, Loi S. Tumour-infiltrating lymphocytes in advanced HER2-positive breast cancer treated with pertuzumab or placebo in addition to trastuzumab and docetaxel: a retrospective analysis of the CLEOPATRA study. *Lancet Oncol* 2017; **18**: 52-62 [PMID: 27964843 DOI: 10.1016/S1470-2045(16)30631-3]
 - 28 **Wang K**, Xu J, Zhang T, Xue D. Tumor-infiltrating lymphocytes in breast cancer predict the response to chemotherapy and survival outcome: A meta-analysis. *Oncotarget* 2016; **7**: 44288-44298 [PMID: 27329588]
 - 29 **Pruneri G**, Vingiani A, Bagnardi V, Rotmensz N, De Rose A, Palazzo A, Colleoni AM, Goldhirsch A, Viale G. Clinical validity of tumor-infiltrating lymphocytes analysis in patients with triple-negative breast cancer. *Ann Oncol* 2016; **27**: 249-256 [PMID: 26598540 DOI: 10.1093/annonc/mdv571]
 - 30 **Adams S**, Gray RJ, Demaria S, Goldstein L, Perez EA, Shulman LN, Martino S, Wang M, Jones VE, Saphner TJ, Wolff AC, Wood WC, Davidson NE, Sledge GW, Sparano JA, Badve SS. Prognostic value of tumor-infiltrating lymphocytes in triple-negative breast cancers from two phase III randomized adjuvant breast cancer trials: ECOG 2197 and ECOG 1199. *J Clin Oncol* 2014; **32**: 2959-2966 [PMID: 25071121 DOI: 10.1200/JCO.2013.55.0491]

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Epitranscriptomics of cancer

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Abstract

The functional impact of modifications of cellular RNAs, including mRNAs, miRNAs and lncRNAs, is a field of intense study. The role of such modifications in cancer has started to be elucidated. Diverse and sometimes opposite effects of RNA modifications have been reported. Some RNA modifications promote, while others decrease the growth and invasiveness of cancer. The present manuscript reviews the current knowledge on the potential impacts of N6-Methyladenosine, Pseudouridine, Inosine, 2'O-methylation or methylcytidine in cancer's RNA. It also highlights the remaining questions and provides hints on research avenues and potential therapeutic applications, whereby modulating dynamic RNA modifications may be a new method to treat cancer.

Key words: RNA modifications; N6-methyladenosine; 5-methylcytidine; 2'O-methylation or methylcytidine; Pseudouridine; Inosine

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Core tip: The present manuscript reviews the current knowledge on RNA modifications in cancer. The potential impacts of N6-Methyladenosine, Pseudouridine, Inosine, 2'O-methylation or methylcytidine in cancer's RNA is presented and discussed. The review also highlights the remaining questions and provides hints on research avenues and potential therapeutic applications, whereby modulating dynamic RNA modifications may be a new method to treat cancer.

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INTRODUCTION

Diverse and abundant modifications are introduced

posttranscriptionally in cellular RNAs during their maturation. These modifications are made on canonical A, C, G, and U residues, and their formation is catalyzed by numerous specific enzymes or RNA-protein complexes (RNPs). Ribonucleotide residues can bear single or multiple modifications on the purine/pyrimidine ring and/or ribose. To date, over one hundred RNA modifications have been identified and listed in dedicated databases (<http://mods.ma.albany.edu/>; <http://modomics.genesilico.pl>)^[1,2]. These naturally occurring modified nucleosides play various structural and functional roles in different types of RNAs: Transfer RNAs (tRNAs), ribosomal RNAs (rRNAs), messenger RNAs (mRNAs), small nuclear RNAs (snRNAs), microRNAs (miRNAs) and long non-coding RNAs (lncRNAs). The most widespread RNA modifications are base or ribose methylations, deamination of adenosine to inosine and isomerization of uridine into pseudouridine. Over the past decades these modifications have been studied in the context of malignancies. Frequently, a modification is found to have pro-cancer or anti-cancer effects depending on the type of RNA, the location of the modification and, most importantly, the cell type and context (*e.g.*, hypoxia). This review presents the current knowledge on the potential link between RNA modifications and cancer by systematically addressing the “pro-cancer”, “anti-cancer” and mixed effects of RNA modifications. Since such a relationship has been reported for only some abundant modifications and for modifications for which a detection method is available [N⁶-Methyladenosine (m⁶A), 5-Methylcytidine (m⁵C), 2'-O-mN, Ψ and I], the present review will focus on these modifications (Figure 1).

M6A IN CANCER RNA

Serendipitously discovered during the characterization of the mRNA 5' cap, methylation of the exocyclic nitrogen of adenosine, named m⁶A, is by far the most abundant mRNA modification, occurring on an average of three sites per mRNA^[3-5]. Recent technological advances have facilitated m⁶A profiling across eukaryotes, including humans, mice^[6], yeasts^[7], and plants^[8,9], indicating that m⁶A is a conserved but dynamic modification. m⁶A has also been identified in rRNA^[10], tRNA^[11], snRNA^[12], miRNA^[13] and lncRNA^[14].

M⁶A patterns are attributed to the consensus RR-ACH sequence (A is methylated; R = A or G; H = A, C, or U; and the first nucleotide next to m⁶A from the 5' end most frequently is G), with preferential distribution near mRNA stop codons and 3' untranslated regions (UTRs) and within long internal exons. Additionally, the m⁶A sites are conserved between human and mouse embryonic stem cells (ESCs) and somatic cells. However, distinct m⁶A patterns can also be detected among different species or cells at different developmental stages^[4,7,15,16]. Some m⁶A signatures are tissue specific^[4], and are altered in response to different stimuli^[17], pointing to the potential role of m⁶A in regulating diverse

cellular processes. m⁶A dynamics are assigned to the complex m⁶A enzymatic machineries, comprising m⁶A “writers”, “readers” and “erasers”. Although a plethora of studies suggest crucial and versatile roles of m⁶A and its machineries, its roles in cancer that have recently emerged are contradictory and require further investigation.

High m⁶A levels in cancers

“Writers” is a term given to enzymes that are part of the methyltransferase complex that introduces m⁶A. Components of this complex are methyltransferase-like 3 (METTL3)^[18], METTL14^[19], Wilms tumor 1-associated protein (WTAP)^[20] and KIAA1429^[21].

METTL3 protein levels were found to be elevated in lung adenocarcinoma cell lines compared to healthy tissue^[22]. Depletion of METTL3 was shown to result in the inhibition of cancer cell growth, decreased invasive ability of cancer cells and increased cell apoptosis in the same study. Additionally, METTL3 was shown to function as an m⁶A-binding protein (“reader”) in a specific subset of m⁶A-modified mRNAs, where it recruits eIF3 during translation initiation and therefore promotes translation. Expression of several oncogenes, including the mRNA of epidermal growth factor receptor (EGFR) and the Hippo pathway effector transcriptional co-activator with the PDZ-binding motif (TAZ) protein, was found to be promoted upon METTL3 recognition^[22].

Similarly, in acute myeloid leukemia (AML), mRNA levels of METTL3 and METTL14 are significantly higher than in most cancers^[23]. METTL3 depletion in MOLM13 caused differentiation and increased apoptosis, suggesting that high m⁶A levels may play a role in sustaining undifferentiated leukemic cells in AML^[23] (Table 1).

Low m⁶A levels in cancers

Two m⁶A “erasers” have been described: Demethylases fat mass and obesity-associated protein (FTO) and alkB homolog 5 (ALKBH5)^[24,25].

Single nucleotide polymorphisms within FTO known to be involved in the development of obesity in genome-wide association studies have been associated with the risk of developing diverse cancer types: Lung cancer, kidney cancer, high-grade prostate cancer, endometrial cancer, pancreatic cancer, pancreatic cancer in patients with type 2 diabetes, and breast cancer^[26-33]. All these cancer types share a single SNP (rsrs9939609): The obesity-associated SNP in intron 1 of the *FTO* gene. This SNP was shown to increase primary transcript levels of the *FTO* gene, suggesting a gain-of-function mutation in cancers associated with this SNP^[34].

In human epidermal growth factor receptor type 2 (HER2)-overexpressing subtypes of breast cancer, FTO is highly expressed in comparison to other breast cancer subtypes^[35]. Contrary to the studies of high m⁶A levels in AML discussed in the previous chapter, low m⁶A levels have also been reported in AML subtypes. FTO expression can be upregulated by certain

Table 1 Relative amount of modification (directly quantified or extrapolated from the expression level of writers/erasers)

RNA modifications	High in cancer	Low in cancer
m6A	Lung adenocarcinoma ^[22] , AML ^[23]	HER2 overexpressing subtypes breast cancer ^[35] , t(11q23)/MLL-rearranged, t(15;17)/PML-RARA, FLT3-ITD, and/or NPM1-mutated AMLs (ASB2 and RARA) ^[36] , GBM (FOXO1) ^[39] , breast cancer (NANOG) ^[40]
2'O-methylation	Breast cancer ^[51,67] , primary and metastatic prostate cancers ^[58] , squamous cell cervical carcinoma ^[57]	
Ψ		Leukemia, lymphoma, multiple myeloma ^[84-86]
Inosine	BLCA, BRCA, COAD, HNSC, LUAD, THCA ^[87,88] , NSCLC (NEIL1 ^[94] , AZIN1 ^[103] , miR-381 ^[94]), SCLC (AZIN1 ^[87] , HCC (AZIN1 ^[101] , FLNB ^[90]), GC ^[91] , ESCC (FLNB ^[92] , cervical cancer ^[89] , CRC (RHOQ) ^[109] , AML (PTPN6) ^[114]	KIRP, KICH ^[87,88] , Breast cancer (Gabra3) ^[118] , Gastric cancer (PODXL) ^[91] , Glioblastoma (GluR-B) ^[119] , onco miR-21, miR-221, miR-222 ^[128] , ESCC (IGFBP7) ^[120] , Glioma (miR-376a*) ^[129] , Melanoma ^[130] (miR-455-5p) ^[132]
5mC	Circulating tumor cells in lung cancer ^[137]	

In brackets are the names of genes that have been analyzed. AZIN1: Antizyme inhibitor 1; RHOQ: Ras homolog family member Q; PODXL: Podocalyxin-like; IGFBP7: Insulin-like growth factor-binding protein 7; PTPN6: Protein tyrosine phosphatase non-receptor type 6; NEIL1: NEI-like protein 1; GluR-B: Glutamate R-B; Gabra3: Alpha-3 subunit of gamma-aminobutyric acid type A; FlnB: Filamin B; ASB2: Ankyrin repeat and SOCS box containing 2; RARA: Retinoic acid receptor alpha; FOXO1: Forkhead box protein M1; GBM: Glioblastoma multiforme; HER2: Human epidermal growth factor receptor type 2; MLL: Mixed lineage leukemia; PML/RARA: Promyelocytic leukemia/retinoic acid receptor alpha; FLT3-ITD: Fms-related tyrosine kinase 3-internal tandem duplication; NPM1: Nucleophosmin 1; NSCLC: Non-small cell lung cancer; HCC: Hepatocellular carcinoma; ESCC: Esophageal cell carcinoma; GC: Gastric cancer; CRC: Colorectal cancer; AML: Acute myeloid leukemia; SCLC: Small cell lung cancer; BLCA: Bladder urothelial carcinoma; BRCA: Breast invasive carcinoma; COAD: Colon adenocarcinoma; HNSC: Head and neck squamous cell carcinoma; LUAD: Lung adenocarcinoma; THCA: Thyroid carcinoma; KICH: Kidney chromophobe; KIRP: Kidney renal papillary cell carcinoma.

oncogenic proteins (e.g., mixed lineage leukemia (MLL)-fusion proteins, promyelocytic leukemia/retinoic acid receptor alpha (PML-RARA), fms-related tyrosine kinase 3-internal tandem duplication (FLT3-ITD), and nucleophosmin 1 (NPM1) mutant), and dataset analysis of human AML confirmed that FTO was expressed at significantly high levels in t(11q23)/MLL-rearranged, t(15;17)/PML-RARA, FLT3-ITD, and/or NPM1-mutated AMLs^[36]. Overexpression of FTO reduces m6A levels in ankyrin repeat and SOCS box containing 2 (ASB2) and retinoic acid receptor alpha (RARA) mRNA transcripts. It has been shown that the loss of m6A markings reduces mRNA stability, resulting in the partial repression of ASB2 and RARA expression in AML cells. In four different AML cohorts, ASB2 and RARA exhibit a significant inverse correlation with FTO expression. ASB2 and RARA are upregulated during normal hematopoiesis and are important regulators of all-*trans*-retinoic acid (ATRA)-induced differentiation of leukemia cells. Through regulating the expression of such targets, FTO inhibits ATRA-induced AML cell differentiation. Both gain- and loss-of-function studies of FTO in leukemic cell models showed an oncogenic role of FTO in these AML subtypes^[36]. However, recent studies have suggested that FTO acts as a demethylase of N6-2' O-dimethyladenosine in mRNA 5' caps, having only minor effects on m6A^[37]. Thus, the role of FTO in AML might be independent of m6A.

Recently, both FTO and ALKBH5 have been found to play similar roles in glioblastoma stem cells (GSCs) and their tumorigenesis^[38]. These studies shed light on their crucial roles in the regulation of mRNA m6A levels for maintaining GSC growth, self-renewal, and tumor development. Enhanced growth and self-renewal of GSCs *in vitro* were detected upon the depletion of

METTL3 or METTL14, resulting in reduced mRNA m6A levels, and promoted the ability of GSCs to form brain tumors *in vivo*. Accordingly, treatment with the FTO inhibitor MA2, the ethyl ester form of meclofenamic acid, increased mRNA m6A levels and suppressed GSC growth *in vitro* and GSC-initiated tumorigenesis, ultimately prolonging the survival of GSC-engrafted mice.

In a similar study, the authors checked the expression levels of m6A regulators in available datasets for glioblastoma multiforme (GBM) and discovered elevated expression of m6A demethylase ALKBH5 that correlated with poor clinical outcomes for GBM patients^[39]. Stable knockdowns in cultured human GSCs showed that the loss of ALKBH5 decreases GSC proliferation and reduces the expression of the stemness markers Nestin, Sox2, Nanog, and Oct4, which are normally expressed in GSCs. In rescue experiments, wild-type, but not catalytically inactive, ALKBH5 recover the phenotype, suggesting that it plays a role in stemness maintenance and that the proliferation of GSCs is solely based on demethylation activity. Moreover, these authors examined the expression of transcription factor FOXO1 (forkhead box m1), which is known to play a pivotal role in regulating GSC proliferation, self-renewal, and tumorigenicity, and found that it depends on ALKBH5 demethylating activity. All these findings were based on m6A hyper erasing, which opens new possibilities for promising targeted treatments in glioblastoma (Table 1).

It has been reported that the hypoxia-inducible factors (HIFs) HIF-1 α and HIF-2 α activate *ALKBH5* gene transcription under hypoxic conditions in breast cancer cells, thus inducing m6A demethylation. This demethylation was shown to stabilize NANOG mRNA and promote the breast cancer stem cell (BCSC)

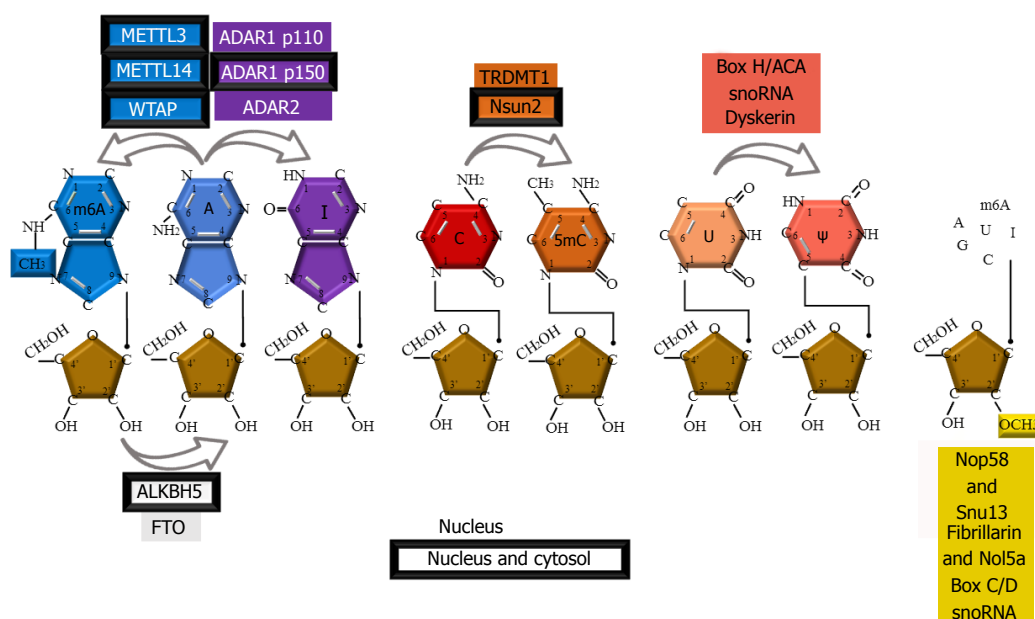


Figure 1 Ribonucleotide RNA modifications known to be of relevance in cancer and their enzymatic machineries. m6A: N6-methyladenosine; METTL3: Methyltransferase like 3; METTL14: Methyltransferase like 14; WTAP: Wilms' tumor 1-associating protein; m6A erasers (ALKBH5: Alkylation repair homologue protein 5; FTO: Fat mass and obesity-associated protein); Inosine (I) writers [ADAR1 (p110 and p150) and ADAR2: Adenosine deaminase acting on RNA 1 and 2]; 5-methylcytosine (5mC) (NSUN2: NOP2/Sun domain protein 2; TRDMT1: tRNA aspartic acid MTase 1); 2'-O-methylation writers (Nol5a: Nucleolar Protein 5A; Nop58: Nucleolar protein 58; Snu13: Small Nuclear Ribonucleoprotein 13; SnoRNA: Small nucleolar RNA).

phenotype. Depletion of ALKBH5 in hypoxic breast cancer cells was identified as an effective strategy to decrease NANOG expression and limit the presence of BCSCs *in vivo*^[40] (Table 1).

Mixed role of m6A in cancer

The primary microRNA (pri-miRNA) junction region between the hairpin stem and the flanking single-stranded RNA was found to be abundant in m6A consensus motifs. The recognition of the junction regions is mediated by Dicer, followed by the recruitment of the ribonuclease Drosha (the microprocessor complex), which cleaves the RNA duplex to yield the premiRNA product. Depletion of HNRNPA2B1 (a nuclear "reader") or METTL3 knockdown in HEK293 and MDA-MB-231 cells resulted in a significant reduction in the expression levels of the mature forms of a number of m6A-marked miRNAs. The tumor-suppressor miRNA let-7 was significantly reduced upon the depletion of METTL3 possibly due to diminished Dicer binding to pri-miRNAs, thus preventing the formation of mature miRNAs. However, these METTL3-depletion experiments also showed a decrease in the expression of onco-miRNAs, such as miR-221 and miR-222^[13,41]. Taken together, the presence of m6A affects diverse pri-miRNA and mature miRNA subpopulations, but its relevance in the context of cancer still needs to be investigated.

2'-O-METHYLATION IN CANCER RNA

Methylation of the 2'-hydroxyl group of ribose is one of the predominant internal modifications of rRNA and

snRNA^[10,42]. This modification is also found in tRNA and mRNA, mostly at the first and second nucleotides in Cap1 and Cap2 structures, respectively.

Introducing 2'-O-methylation on ribose is mediated by complexes of guide RNA and proteins named small nucleolar ribonucleoprotein (snoRNP) complexes or by methyltransferases: Human cap1 and 2, 2'-O-ribose methyltransferase, hMTr1 and hMTr2^[43-45]. snoRNP complexes consist of Fibrillarin (the catalytic component in humans, also known as Nop1p in yeast), Nol5a (Nop56p), Nop58 and Snu13 subunits^[46-48], which are guided by C/D Box snoRNAs to the appropriate base^[49,50].

High 2'-O-methylation levels in cancer

Tumor suppressor p53 and Fibrillarin seem to be linked^[51]. Knockdown of p53 in cellular models of breast and colon cancer resulted in the overexpression of Fibrillarin at both the mRNA and protein levels. It is suggested that tumorigenesis associated to mutated p53 promotes an increase in the methylation status of rRNAs, which alters their ribozyme activity, thus affecting their translation fidelity and rate. Through the methylation of rRNA, Fibrillarin stimulates the translation of cancer-promoting proteins: (1) Insulin-like growth factor 1 receptor (IGF1R), which plays a role in tumor progression, cell survival, and the response to chemotherapy (reviewed by Pollak *et al.*^[52]); (2) c-Myc, a pleiotropic pro-oncogene (reviewed by Dang *et al.*^[53]); (3) Fibroblast growth factor 1/2 (FGF1/2), which is involved in epithelial-mesenchymal transition^[54]; and (4) Vascular endothelial growth factor

A (VEGFA), which acts in tumoral angiogenesis^[51,55].

Translation of these proteins relies on internal ribosome entry site (IRES) in the mRNA, which is a 5' cap-independent translation mechanism that may be used in specific conditions. The inhibition of rRNA methylation was shown to impair IRES translation initiation by perturbing the association of the 40S and 60S subunits^[56]. Therefore, it is conceivable that enhanced ribosomal methylation increases the translation of IRES-containing mRNAs. Nevertheless, clinical analysis shows that a high level of Fibrillarin in primary breast tumors is associated with poor survival, independent of other biological markers^[51]. Elevated expression levels of Fibrillarin were previously reported in primary and metastatic prostate cancers and in squamous cell cervical carcinoma (Table 1)^[57,58].

NOL5A gene was found to be overexpressed in Burkitt's lymphoma-associated c-Myc mutants^[59], and human NOP58 mRNA levels were found to be elevated in metastatic melanoma lesions^[60].

Low 2'O-methylation levels in cancer

Contrary to Fibrillarin's indirect promotion of IRES-driven translation, in MCF-7, a breast cancer cell line, Fibrillarin knockdown resulted in the accumulation of p53, possibly affecting the UTR of the p53 mRNA and increasing IRES-driven *de novo* synthesis^[61]. These studies suggest a complex interplay between p53 and Fibrillarin, while IRES-dependent translation is not exclusively stimulated by increased rRNA methylation.

Mixed 2'O-methylation in cancer

SnoRNA expression profiles were investigated in endometrial, lung and prostate cancers, as well as in glioma and chronic lymphocytic leukemia. High-throughput screening of snoRNAs in cancerous versus normal tissues underlined their overexpression or underexpression as common molecular events in tumorigenesis, with the former being more pronounced than the latter^[62-66]. Analysis of blood serum has shown the possibility of detecting snoRNAs in breast cancer patient samples and the associated upregulation of a specific snoRNA, U6, in active disease^[67]. Therefore, profiling snoRNAs with their respective RNA 2'O methylation modification signatures might be used as a noninvasive biomarker in the diagnosis and prognosis of cancer.

PEUDOURIDINE IN CANCER RNA

The fifth base, known as pseudouridine (Ψ)^[68], is one of the most abundant nucleotide modifications present in all three life domains^[2]. After its initial detection in rRNA and tRNA, pseudouridine was detected in mRNA, lncRNA, and snRNAs, such as U2 snRNA and snoRNA^[69,70]. Introducing Ψ in eukaryotic RNA can be mediated through guide RNA-dependent H/ACA BOX snoRNA pseudouridine synthases (PUSs) or guide RNA-independent PUSs. A recent review by Penzo *et al.*^[71] reports on the functional roles of pseudouridines and related human pathologies.

Only low Ψ levels have been reported in cancer tissues/cells; thus, this chapter will contain only a section titled "low Ψ levels in cancer". Surprisingly, elevated levels of circulating Ψ have been measured in the body fluids of cancer patients, but its role and origin are not well defined, so this finding will not be further discussed here.

Low Ψ levels in cancer

The highly conserved protein dyskerin is the human PUS that catalyzes the pseudouridylation of snoRNPs that assemble during the transcription of guide H/ACA RNA. Mutations in the *Dkc1* gene coding for dyskerin can be found in the X-linked form of dyskeratosis congenita (DC). DC is a rare, inherited disorder that is characterized by mucocutaneous abnormalities and bone marrow failure. DC can be inherited as an X-linked recessive, autosomal dominant or autosomal recessive disease^[72]. Although the absence of dyskerin, which results in the loss of pseudouridine in rRNA, was suggested as a primary cause of DC, a recent study assigned telomerase dysfunction as the primary cause of DC^[73]. Namely, mutations in H/ACA-resembling domains in the RNA component of telomerase RNP, which are required for telomerase accumulation, stability, 3' end processing and function, are associated with an autosomal form of DC^[74-76].

In patients with DC, a higher predisposition to cancer has been reported, although low mutational frequency in the *DKC1* gene was shown in primary tumors^[77]. This predisposition might be a synergistic outcome of impaired pseudouridylation. Most likely, the dysregulation of rRNA pseudouridylation precedes disease onset, as studies in hypomorphic *Dkc1*-mutant mice suggest. A specific defect of the internal ribosome entry site also occurs upon *DKC1* loss, causing a specific defect in the translation of some IRES-containing mRNAs. Ribosomes that lack pseudouridine modifications show a direct impairment in binding to IRES elements^[78]. Consequently, in hypomorphic *DKC1* mice, cap-dependent translation of mRNA is not compromised, but translation of IRES-containing mRNAs, including the tumor suppressors p27 and p53, is perturbed^[79-82], resulting in a higher incidence of cancer development in these mice. Thus, this impaired translation of tumor suppressor mRNA might also be a driver of cancer in DC patients. Moreover, recent identification of Ψ in mRNA^[83] brings an additional level of complexity and regulation of the expression of target RNAs.

In hematological cancers, such as leukemias, lymphoma and multiple myeloma, downregulation of specific subsets of dyskerin-associated H/ACA snoRNAs has been demonstrated^[84-86] (Table 1). Thus, lower pseudouridylation levels are a widespread feature of cancer.

INOSINE IN CANCER RNA

Inosine is an RNA modification resulting from the hydrolytic deamination of adenosine catalyzed by adenosine

deaminase enzymes acting on double-stranded RNA (ADAR) or adenosine deaminase acting on transfer RNA (ADAT), which are families known to function in A-to-I RNA editing. Enzymes of the ADAR family are catalytically active ADAR1, ADAR2 and ADAR3, which still has an unknown function.

ADARs introduce inosine in coding and non-coding RNAs and have drastic impacts on the cellular transcriptome and translome. The hypo- or hyper-editome has been associated with diverse types of cancer. The role of ADAT in cancer has not been reported.

High editing levels in cancer

Most frequently editing locations are long, partially complementary RNAs formed from inverted non-coding repeats, such as *Arthrobacter luteus* (Alu) and long interspersed element (LINE) located in mRNA UTRs and introns. Two major studies have investigated RNA-editing patterns in tumors versus normal tissues. Each of the studies employed RNA-Seq datasets from The Cancer Genome Atlas (TCGA) project (<https://cancergenome.nih.gov/>) and compared them to reference datasets of editing sites. High-confidence RNA editing sites are annotated in the Rigorously Annotated Database of A-to-I RNA Editing (RADAR, <http://rnaedit.com/>), where one study focused on detecting Alu and non-Alu RNA editing events in 17 cancers, whereas the other study focused on Alu RNA editing events in 9 different cancers^[87,88].

In general, elevated Alu editing activity in tumors compared to matched normal tissues was found in bladder urothelial carcinoma (BLCA), breast invasive carcinoma (BRCA), colon adenocarcinoma (COAD), head and neck squamous cell carcinoma (HNSC), lung adenocarcinoma (LUAD) and thyroid carcinoma (THCA). This hyperediting of Alu was attributed to ADAR1, whose expression levels matched in all these types of cancer, except COAD. Similarly, a study by Han *et al.*^[87] where more patient samples and non-Alu edited sequences were included, confirmed hyperediting in BLCA, BRCA, HNSC, LUAD, THCA compared to normal tissues. Again, increased editing levels correlated with the mRNA levels of ADAR1.

Increased ADAR-1 levels were reported in non-small cell lung cancer (NSCLC), hepatocellular carcinoma (HCC), esophageal cell carcinoma (ESCC), gastric (GC) and cervical cancer, suggesting that tight regulation of editing levels might have implications in cancer development and that ADAR1 might act as an oncogene^[89-92] (Table 1 and Figure 2).

Recoding editing: In non-small cell lung cancer samples, *ADAR1* gene amplification was shown to increase the editing of the DNA base excision repair glycosylase enzyme NEI-like protein 1 (NEIL1). Pre-mRNA editing of NEIL-1 causes a lysine to arginine (K242R) change in the lesion recognition loop of the protein. The edited NEIL1 protein removes thymine glycol from duplex DNA at a lower rate compared to the unedited form, while

repair of the guanidinohydantoin lesion is enhanced by edited NEIL1^[93]. In overexpression experiments, transfection of edited NEIL1 enhanced the growth of A459 cells in comparison with the transfection of unedited transcripts. Thus, increased recoding editing of NEIL1 as a proposed target of ADAR1 could contribute to the phenotype of lung cancer cells^[94].

AZIN1 (encoding antizyme inhibitor 1) is edited by ADAR1, which has increased expression levels in HCC and was found to positively correlate with AZIN1 editing frequency. AZIN1 is an antizyme inhibitor whose activity is crucial in limiting cellular proliferation. Antizyme binds and induces the degradation of the growth-promoting proteins ornithine decarboxylase (ODC) and cyclin D1 (CCND1)^[95,96]. AZIN1 is homologous to ODC and has a greater binding affinity to antizyme compared to ODC. Binding of AZIN1 to antizyme prevents the degradation of ODC^[97]. Thereby, AZIN1 acts as an oncogene by inhibiting the tumor-suppressor activities of antizyme^[96]. AZIN1 expression was found to be substantially elevated in cancers of the prostate, brain, breast and liver, and gene expression data have identified alterations in the AZIN1-to-antizyme ratio in many human cancers, confirming its role in promoting growth^[98-100]. In HCC, increased A-I editing of the AZIN1 transcript introduces serine-to-glycine substitution at residue 367 in the protein. This recoding editing is associated with conformational changes and translocation from the nucleus to the cytoplasm and results in a higher-binding affinity to antizyme and greater protein stability, thus promoting cell proliferation. AZIN1 editing increases during the progression from primary liver cancer and cirrhosis to advanced HCC with recurrence and metastasis, suggesting its use as a prognostic marker^[101]. It is plausible that similar editing events occur in other types of cancer, as has been confirmed in esophageal squamous cell carcinoma (ESCC) and breast cancer^[87,92,102]. Recently, tumorigenesis of NSCLC was also attributed to high levels of AZIN1 editing^[103]. It has been reported that AZIN1 editing levels correlate with sensitivity to drug treatment in cancer cell lines. Cancer cells lines with increased levels of AZIN1 editing showed more sensitivity to some of the chemotherapies used in small cell lung cancer (SCLC): Paclitaxel, irinotecan, and topotecan^[87].

Filamin B (FLNB) is an actin cross-linking protein and, together with filamin A, it forms homo- and heterodimers mediating orthogonal branching of actin filaments^[104]. Filamin B is known to be a target for editing^[105], and interestingly, one recoding editing event was shown to be increased in two types of cancer. In HCC, ADAR1 and ADAR2 were both reported to mediate FLNB transcript editing in codon 2269, resulting in the amino acid change Met→Val. Increased editing of FLNB compared to matched non-tumor liver tissues has been closely associated with HCC pathogenesis from normal to clinically verified HCC. In this study, ADAR1 levels were shown to be increased, while ADAR2 levels were decreased in HCC samples compared to non-tumor



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bone marrow mononuclear cells (BMMCs) of patients with acute myeloid leukemia, a novel PTPN6 transcript retaining intron 3 has been identified. This transcript arises from an alternative splicing reaction where editing mediated deamination of A7866 in intron 3 erases this branch formation site, making it invisible to the splicing machinery. The ADAR responsible has not been identified. It is suggested that this retention results in the translation of a nonfunctional protein where the intron 3-encoded sequence is located in the N-terminal Src homology 2 (SH2) domain. PTPN6 binding with partner proteins, such as proto-oncogene receptor tyrosine kinase-c-Kit^[112], and its self-inhibition of phosphatase activity occurs *via* its N terminal domain^[113]. All this information suggests that its deregulation ultimately leads to uncontrolled hematopoietic growth and function. The tumor-specific editing seen in AML might correlate with the clinical course of the disease since low levels of intron-retaining transcripts in patient BMMCs at remission compared to those at diagnosis suggest that editing promotes tumorigenesis^[114].

High miRNA editing levels in cancer: *ADAR1* gene amplification in NSCLC demonstrated ADAR1 overexpression in patients with early-stage lung cancer, underlining its potential oncogenic role in this cancer. Increased levels of ADAR1 corresponded with edited miR-381 levels in NSCLC. Overexpression of edited miR-381 in NSCLC possibly contributes to stemness and chemoresistance^[94].

Low editing levels in cancer

Low levels of mRNA open reading frame editing in cancer: The same correlation but in the opposite direction was shown in hypoedited cancers, such as kidney chromophobe (KICH) and kidney renal papillary cell carcinoma (KIRP), with ADAR1 mRNA paired editing levels. The ADAR2 levels checked in both studies showed a complex expression pattern but no matching editing levels (Table 1 and Figure 2)^[87,88]. The role of ADAR1 in breast cancer is not fully understood. A recent study reported high ADAR1 expression in half of the examined triple-negative-cancer patients^[115]. Conversely, it has been proposed that ADAR1 prevents tumor progression by editing the transcript coding for the alpha-3 subunit of gamma-aminobutyric acid type A (Gabra3).

The chloride-permeable gamma-aminobutyric acid type A (GABA_A) receptors are crucial mediators of fast inhibitory neurotransmission in the central nervous system^[116]. The Gabra3 transcript undergoes recoding editing of isoleucine to methionine (I/M) in the third transmembrane region. This substitution was found to affect GABA_A surface presentation and its cellular trafficking^[117]. In addition to being normally expressed in normal neuronal tissues, Gabra3 has been identified in breast cancer, where its high expression inversely correlates with breast cancer survival. ADAR1-edited Gabra3 was found in non-invasive breast cancer cell lines and was linked with the protein kinase B (Akt) pathway. A proposed mechanism for the non-invasive phenotype is that Gabra3 editing reduces its surface expression and indirectly prevents Akt activation, thereby preventing cell proliferation and invasiveness. Thus, the unedited form of Gabra3 in breast cancer is suggested to promote tumor progression, invasion and metastatic potential^[118].

Lower ADAR2 levels are recognized in gastric cancer, glioblastoma, HCC and ESCC^[91,119,120] (Figure 2). ADAR2 levels were found to correlate with changes in podocalyxin-like (PODXL) and GluR-B functions. The PODXL RNA editing event is an amino acid substitution from histidine (His) to arginine (Arg) at codon 241. This editing in the gastric cancer cell line MKN28 was shown to prevent increased growth rates and invasive capability compared to cells with the unedited form. Moreover, recoding editing of a single position located in the channel-pore-loop domain in GluR subunit B (GluR-B) (the Q/R-site) from Gln to Arg results in a channel that is impermeable to Ca²⁺^[121]. Tight regulation of editing is essential for the adequate function of this channel. Hypoedited PODXL and GluR-B with altered functions are associated with gastric cancer and malignant glioblastoma, respectively. Consequently, a tumor-suppressor role has been attributed to ADAR2^[91,119].

In ESCC, it has been reported that ADAR2 promotes apoptosis by editing and stabilizing insulin-like growth factor-binding protein 7 (IGFBP7) RNA. IGFBP7 is a secreted factor binding to and interfering with the activation of IGF1R. Through receptor occupation,

IGFBP7 blocks downstream phosphatidylinositol 3-kinase (PI3K)-AKT signaling, resulting in the inhibition of protein synthesis and cell apoptosis^[122]. IGFBP7 was previously reported to be an apoptotic promoter in prostate cancer^[123], colorectal cancer^[124] and breast cancer^[125]. The editing site in IGFBP7 is at position 284 of the coding sequence, and codon 95 is changed from AAG (lysine) to AIG, which is read as AGG (arginine) (K95R). This editing was shown to protect IGFBP7 against matriptase proteolysis in ESCC culture and xenografts, thus enabling the proapoptotic function of IGFBP7. ADAR2 is known to be downregulated in ESCC, and its upregulation induces apoptosis in ESCC cell lines *in vitro*, suggesting that IGFBP7 under editing may promote tumorigenesis in esophageal squamous cell carcinoma^[120].

Low miRNA editing levels in cancer: ADAR2 rescue in glioblastoma cells was shown to inhibit cell proliferation and migration, confirming its possible tumor-suppressor role^[126]. This anti-tumor effect might be explained through the regulation of onco-miRNAs in glioblastoma. Three particularly investigated onco-miRNAs, miR-221, miR-222 and miR-21, are overexpressed in glioblastoma^[127]. ADAR2 can edit miR-222/221 and miR-21 precursors and decrease the expression of the corresponding mature onco-miRNAs in the normal mouse brain and in different lines. Decreased levels of ADAR2 identified in glioblastoma probably push the balance of onco-miRNA/tumor-suppressor miRNA towards increased expression of onco-miRNAs, such as miR-221, miR-222 and miR-21, thereby supporting tumor progression^[128].

In the human brain, the miR-376 cluster encodes 4 pri-miRs that give rise to 5 distinct mature miRNAs, which are subjected to specific A-to-I RNA editing on 9 adenosines. In noninvasive U87 glioma cells, the expression of the unedited miR-376a* was shown to promote aggressive tumor migration and invasion of these cells both *in vitro* and *in vivo*. The editing reaction missing in the GBM cell lines generally occurs in the seed region of pri-miR-376a1 at the +9 site, ultimately giving rise to mature edited miR-376a*. The absence of this editing changes the specific targets of the miRNA. It has been identified that non edited miR-376a*, through its binding to 3' UTR, has a novel target, RAP2A, which is a member of the RAS oncogene family with an unknown function. However, the non edited miR-376a* targeting of RAP2A is unable to target the autocrine motility factor receptor (AMFR), resulting in its upregulation and possibly contributing to increased migration and invasiveness of glioma cells^[129].

Melanoma is the most aggressive type of skin cancer. It has been reported that there is a significant decrease in ADAR1 expression in approximately 65% of metastatic melanoma specimens compared to melanocytes^[130] (Table 1 and Figure 2). ADAR-1 transcripts were found to be targeted by miR-17 and miR-432, thus decreasing

ADAR1 expression. Both miR-17 and miR-432 were identified to be overexpressed in melanoma possibly due to the amplification of encoding genes^[130]. However, studies suggest that ADAR1 insufficiencies contribute to the enhancement of proliferation of melanoma cells through editing the independent regulation of miRNA biogenesis. miRNA-455-5p was identified as a target of ADAR1 in low-metastatic melanoma cells but not in highly metastatic cell lines. ADAR1 was shown to edit pri-miR-455-5p at +2 and +17 positions. This editing probably results in the reduction of the processing of pri-miRNA by Dicer or Drosha by lowering the binding affinity. However, it is also possible that ADAR1 binds to Dicer since the amount of miR-455-5p bound to Dicer and Drosha was inversely correlated with ADAR1 expression. ADAR1 was shown to form a complex with Dicer through protein-protein interactions^[131]. In this study, the authors gave a model of RNA editing in the context of melanoma progression and metastasis, where cAMP responses element binding (CREB) downregulates ADAR1 and gives rise to non-edited miR-455-5p. Expression of miR-455-5p suppresses the tumor suppressor gene cytoplasmic polyadenylation element-binding protein 1 (CPEB1), resulting in growth promotion and metastasis in melanoma cells^[132].

M5C IN CANCER RNA

Cytosine base methylation - m5C has been identified in rRNA, tRNA and recently in mRNAs and is particularly enriched in untranslated regions and near Argonaute-binding regions^[133]. The enzymes responsible for the introduction of m5C are members of the DNA methyltransferase homolog (Dnmt2) and the NOP2/Sun (NSun 2 and 4) RNA methyltransferase family^[134-136]. The role of these enzymes in the methylating activities of tumorigenesis is currently unknown. However, in circulating tumor cells from lung cancer patients, increased RNA m5C levels were shown compared to those in whole blood cells^[137]. Further investigation of the role of m5C in cancer is required.

CONCLUSION

Tight regulation of the writing, reading and eventual erasing of RNA modifications is essential for RNA metabolism. Misbalanced expression of the enzymes responsible for introducing, and in some cases removing, these modifications are considered a possible signature for specific types of cancer (Table 1). Considering the broad effect of RNA modifications on tumor cell biology, future methylome, pseudome and editome studies will shed light on those relatively unexplored epitranscriptomic mechanisms in tumors. Those studies will pave the way for the development of anti-cancer drugs that could act by steering RNA modifications.

REFERENCES

- 1 Cantara WA, Crain PF, Rozenski J, McCloskey JA, Harris KA,

- Zhang X, Vendeix FA, Fabris D, Agris PF. The RNA Modification Database, RNAMDB: 2011 update. *Nucleic Acids Res* 2011; **39**: D195-D201 [PMID: 21071406 DOI: 10.1093/nar/gkq1028]
- 2 Machnicka MA, Milanowska K, Osman Oglou O, Purta E, Kurkowska M, Olchowik A, Januszewski W, Kalinowski S, Dunin-Horkawicz S, Rother KM, Helm M, Bujnicki JM, Grosjean H. MODOMICS: a database of RNA modification pathways-2013 update. *Nucleic Acids Res* 2013; **41**: D262-D267 [PMID: 23118484 DOI: 10.1093/nar/gks1007]
- 3 Desrosiers R, Friderici K, Rottman F. Identification of methylated nucleosides in messenger RNA from Novikoff hepatoma cells. *Proc Natl Acad Sci USA* 1974; **71**: 3971-3975 [PMID: 4372599 DOI: 10.1073/pnas.71.10.3971]
- 4 Meyer KD, Saletore Y, Zumbo P, Elemento O, Mason CE, Jaffrey SR. Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. *Cell* 2012; **149**: 1635-1646 [PMID: 22608085 DOI: 10.1016/j.cell.2012.05.003]
- 5 Rana AP, Tuck MT. Analysis and in vitro localization of internal methylated adenine residues in dihydrofolate reductase mRNA. *Nucleic Acids Res* 1990; **18**: 4803-4808 [PMID: 2395644 DOI: 10.1093/nar/18.16.4803]
- 6 Dominissini D, Moshitch-Moshkovitz S, Schwartz S, Salmon-Divon M, Ungar L, Osenberg S, Cesarkas K, Jacob-Hirsch J, Amariglio N, Kupiec M, Sorek R, Rechavi G. Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq. *Nature* 2012; **485**: 201-206 [PMID: 22575960 DOI: 10.1038/nature11112]
- 7 Schwartz S, Agarwala SD, Mumbach MR, Jovanovic M, Mertins P, Shishkin A, Tabach Y, Mikkelsen TS, Satija R, Ruvkun G, Carr SA, Lander ES, Fink GR, Regev A. High-resolution mapping reveals a conserved, widespread, dynamic mRNA methylation program in yeast meiosis. *Cell* 2013; **155**: 1409-1421 [PMID: 24269006 DOI: 10.1016/j.cell.2013.10.047]
- 8 Li Y, Wang X, Li C, Hu S, Yu J, Song S. Transcriptome-wide N⁶-methyladenosine profiling of rice callus and leaf reveals the presence of tissue-specific competitors involved in selective mRNA modification. *RNA Biol* 2014; **11**: 1180-1188 [PMID: 25483034 DOI: 10.4161/rna.36281]
- 9 Luo GZ, MacQueen A, Zheng G, Duan H, Dore LC, Lu Z, Liu J, Chen K, Jia G, Bergelson J, He C. Unique features of the m6A methylome in *Arabidopsis thaliana*. *Nat Commun* 2014; **5**: 5630 [PMID: 25430002 DOI: 10.1038/ncomms6630]
- 10 Maden BE. The numerous modified nucleotides in eukaryotic ribosomal RNA. *Prog Nucleic Acid Res Mol Biol* 1990; **39**: 241-303 [PMID: 2247610 DOI: 10.1016/S0079-6603(08)60629-7]
- 11 Iwanami Y, Brown GM. Methylated bases of transfer ribonucleic acid from HeLa and L cells. *Arch Biochem Biophys* 1968; **124**: 472-482 [PMID: 5661617 DOI: 10.1016/0003-9861(68)90355-X]
- 12 Bringmann P, Lührmann R. Antibodies specific for N⁶-methyladenosine react with intact snRNPs U2 and U4/U6. *FEBS Lett* 1987; **213**: 309-315 [PMID: 2951275 DOI: 10.1016/0014-5793(87)81512-0]
- 13 Alarcón CR, Lee H, Goodarzi H, Halberg N, Tavazoie SF. N⁶-methyladenosine marks primary microRNAs for processing. *Nature* 2015; **519**: 482-485 [PMID: 25799998 DOI: 10.1038/nature14281]
- 14 Liu N, Parisien M, Dai Q, Zheng G, He C, Pan T. Probing N⁶-methyladenosine RNA modification status at single nucleotide resolution in mRNA and long noncoding RNA. *RNA* 2013; **19**: 1848-1856 [PMID: 24141618 DOI: 10.1261/rna.041178.113]
- 15 Geula S, Moshitch-Moshkovitz S, Dominissini D, Mansour AA, Kol N, Salmon-Divon M, Hershkovitz V, Peer E, Mor N, Manor YS, Ben-Haim MS, Eyal E, Yunger S, Pinto Y, Jaitin DA, Viukov S, Rais Y, Krupalnik V, Chomsky E, Zerbib M, Maza I, Rechavi Y, Massarwa R, Hanna S, Amit I, Levanon EY, Amariglio N, Stern-Ginossar N, Novershtern N, Rechavi G, Hanna JH. Stem cells. m6A mRNA methylation facilitates resolution of naïve pluripotency toward differentiation. *Science* 2015; **347**: 1002-1006 [PMID: 25569111 DOI: 10.1126/science.1261417]
- 16 Batista PJ, Molinier B, Wang J, Qu K, Zhang J, Li L, Bouley DM, Lujan E, Haddad B, Daneshvar K, Carter AC, Flynn RA, Zhou C,

- Lim KS, Dedon P, Wernig M, Mullen AC, Xing Y, Giallourakis CC, Chang HY. m(6)A RNA modification controls cell fate transition in mammalian embryonic stem cells. *Cell Stem Cell* 2014; **15**: 707-719 [PMID: 25456834 DOI: 10.1016/j.stem.2014.09.019]
- 17 **Bodi Z**, Zhong S, Mehra S, Song J, Graham N, Li H, May S, Fray RG. Adenosine Methylation in Arabidopsis mRNA is Associated with the 3' End and Reduced Levels Cause Developmental Defects. *Front Plant Sci* 2012; **3**: 48 [PMID: 22639649 DOI: 10.3389/fpls.2012.00048]
- 18 **Bokar JA**, Shambaugh ME, Polayes D, Matera AG, Rottman FM. Purification and cDNA cloning of the AdoMet-binding subunit of the human mRNA (N6-adenosine)-methyltransferase. *RNA* 1997; **3**: 1233-1247 [PMID: 9409616]
- 19 **Liu J**, Yue Y, Han D, Wang X, Fu Y, Zhang L, Jia G, Yu M, Lu Z, Deng X, Dai Q, Chen W, He C. A METTL3-METTL14 complex mediates mammalian nuclear RNA N6-adenosine methylation. *Nat Chem Biol* 2014; **10**: 93-95 [PMID: 24316715 DOI: 10.1038/nchembio.1432]
- 20 **Ping XL**, Sun BF, Wang L, Xiao W, Yang X, Wang WJ, Adhikari S, Shi Y, Lv Y, Chen YS, Zhao X, Li A, Yang Y, Dahal U, Lou XM, Liu X, Huang J, Yuan WP, Zhu XF, Cheng T, Zhao YL, Wang X, Rendtlew Danielsen JM, Liu F, Yang YG. Mammalian WTAP is a regulatory subunit of the RNA N6-methyladenosine methyltransferase. *Cell Res* 2014; **24**: 177-189 [PMID: 24407421 DOI: 10.1038/cr.2014.3]
- 21 **Schwartz S**, Mumbach MR, Jovanovic M, Wang T, Maciag K, Bushkin GG, Mertins P, Ter-Ovanesyan D, Habib N, Cacchiarelli D, Sanjana NE, Freinkman E, Pacold ME, Satija R, Mikkelsen TS, Hacohen N, Zhang F, Carr SA, Lander ES, Regev A. Perturbation of m6A writers reveals two distinct classes of mRNA methylation at internal and 5' sites. *Cell Rep* 2014; **8**: 284-296 [PMID: 24981863 DOI: 10.1016/j.celrep.2014.05.048]
- 22 **Lin S**, Choe J, Du P, Triboulet R, Gregory RI. The m(6)A Methyltransferase METTL3 Promotes Translation in Human Cancer Cells. *Mol Cell* 2016; **62**: 335-345 [PMID: 27117702 DOI: 10.1016/j.molcel.2016.03.021]
- 23 **Vu LP**, Pickering BF, Cheng Y, Zaccara S, Nguyen D, Minuesa G, Chou T, Chow A, Saletore Y, MacKay M, Schulman J, Famulare C, Patel M, Klimek VM, Garrett-Bakelman FE, Melnick A, Carroll M, Mason CE, Jaffrey SR, Kharas MG. The N⁶-methyladenosine (m⁶A)-forming enzyme METTL3 controls myeloid differentiation of normal hematopoietic and leukemia cells. *Nat Med* 2017; **23**: 1369-1376 [PMID: 28920958 DOI: 10.1038/nm.4416]
- 24 **Jia G**, Fu Y, Zhao X, Dai Q, Zheng G, Yang Y, Yi C, Lindahl T, Pan T, Yang YG, He C. N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. *Nat Chem Biol* 2011; **7**: 885-887 [PMID: 22002720 DOI: 10.1038/nchembio.687]
- 25 **Zheng G**, Dahl JA, Niu Y, Fedorcsak P, Huang CM, Li CJ, Vågbo CB, Shi Y, Wang WL, Song SH, Lu Z, Bosmans RP, Dai Q, Hao YJ, Yang X, Zhao WM, Tong WM, Wang XJ, Bogdan F, Furu K, Fu Y, Jia G, Zhao X, Liu J, Krokan HE, Klungland A, Yang YG, He C. ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. *Mol Cell* 2013; **49**: 18-29 [PMID: 23177736 DOI: 10.1016/j.molcel.2012.10.015]
- 26 **Brennan P**, McKay J, Moore L, Zaridze D, Mukeria A, Szeszenia-Dabrowska N, Lissowska J, Rudnai P, Fabianova E, Mates D, Bencko V, Foretova L, Janout V, Chow WH, Rothman N, Chabrier A, Gaborieau V, Timpson N, Hung RJ, Smith GD. Obesity and cancer: Mendelian randomization approach utilizing the FTO genotype. *Int J Epidemiol* 2009; **38**: 971-975 [PMID: 19542184 DOI: 10.1093/ije/dyp162]
- 27 **da Cunha PA**, de Carlos Back LK, Sereia AF, Kubelka C, Ribeiro MC, Fernandes BL, de Souza IR. Interaction between obesity-related genes, FTO and MC4R, associated to an increase of breast cancer risk. *Mol Biol Rep* 2013; **40**: 6657-6664 [PMID: 24091943 DOI: 10.1007/s11033-013-2780-3]
- 28 **Delahanty RJ**, Beeghly-Fadiel A, Xiang YB, Long J, Cai Q, Wen W, Xu WH, Cai H, He J, Gao YT, Zheng W, Shu XO. Association of obesity-related genetic variants with endometrial cancer risk: a report from the Shanghai Endometrial Cancer Genetics Study. *Am J Epidemiol* 2011; **174**: 1115-1126 [PMID: 21976109 DOI: 10.1093/aje/kwr233]
- 29 **Kaklamani V**, Yi N, Sadim M, Siziopikou K, Zhang K, Xu Y, Tofilon S, Agarwal S, Pasche B, Mantzoros C. The role of the fat mass and obesity associated gene (FTO) in breast cancer risk. *BMC Med Genet* 2011; **12**: 52 [PMID: 21489227 DOI: 10.1186/1471-2350-12-52]
- 30 **Lewis SJ**, Murad A, Chen L, Davey Smith G, Donovan J, Palmer T, Hamdy F, Neal D, Lane JA, Davis M, Cox A, Martin RM. Associations between an obesity related genetic variant (FTO rs9939609) and prostate cancer risk. *PLoS One* 2010; **5**: e13485 [PMID: 20976066 DOI: 10.1371/journal.pone.0013485]
- 31 **Lin Y**, Ueda J, Yagyu K, Ishii H, Ueno M, Egawa N, Nakao H, Mori M, Matsuo K, Kikuchi S. Association between variations in the fat mass and obesity-associated gene and pancreatic cancer risk: a case-control study in Japan. *BMC Cancer* 2013; **13**: 337 [PMID: 23835106 DOI: 10.1186/1471-2407-13-337]
- 32 **Lurie G**, Gaudet MM, Spurdle AB, Carney ME, Wilkens LR, Yang HP, Weiss NS, Webb PM, Thompson PJ, Terada K, Setiawan VW, Rebbeck TR, Prescott J, Orlow I, O'Mara T, Olson SH, Narod SA, Matsuno RK, Lissowska J, Liang X, Levine DA, Le Marchand L, Kolonel LN, Henderson BE, Garcia-Closas M, Doherty JA, De Vivo I, Chen C, Brinton LA, Akbari MR; Australian National Endometrial Cancer Study Group; Epidemiology of Endometrial Cancer Consortium (E2C2), Goodman MT. The obesity-associated polymorphisms FTO rs9939609 and MC4R rs17782313 and endometrial cancer risk in non-Hispanic white women. *PLoS One* 2011; **6**: e16756 [PMID: 21347432 DOI: 10.1371/journal.pone.0016756]
- 33 **Tang H**, Wei P, Duell EJ, Risch HA, Olson SH, Bueno-de-Mesquita HB, Gallinger S, Holly EA, Petersen GM, Bracci PM, McWilliams RR, Jenab M, Riboli E, Tjønneland A, Boutron-Ruault MC, Kaaks R, Trichopoulos D, Panico S, Sund M, Peeters PH, Khaw KT, Amos CI, Li D. Genes-environment interactions in obesity- and diabetes-associated pancreatic cancer: a GWAS data analysis. *Cancer Epidemiol Biomarkers Prev* 2014; **23**: 98-106 [PMID: 24136929 DOI: 10.1158/1055-9965.EPI-13-0437-T]
- 34 **Berulava T**, Horsthemke B. The obesity-associated SNPs in intron 1 of the FTO gene affect primary transcript levels. *Eur J Hum Genet* 2010; **18**: 1054-1056 [PMID: 20512162 DOI: 10.1038/ejhg.2010.71]
- 35 **Tan A**, Dang Y, Chen G, Mo Z. Overexpression of the fat mass and obesity associated gene (FTO) in breast cancer and its clinical implications. *Int J Clin Exp Pathol* 2015; **8**: 13405-13410 [PMID: 26722548]
- 36 **Li Z**, Weng H, Su R, Weng X, Zuo Z, Li C, Huang H, Nachtergaele S, Dong L, Hu C, Qin X, Tang L, Wang Y, Hong GM, Huang H, Wang X, Chen P, Gurbuxani S, Arnovitz S, Li Y, Li S, Strong J, Neilly MB, Larson RA, Jiang X, Zhang P, Jin J, He C, Chen J. FTO Plays an Oncogenic Role in Acute Myeloid Leukemia as a N⁶-Methyladenosine RNA Demethylase. *Cancer Cell* 2017; **31**: 127-141 [PMID: 28017614 DOI: 10.1016/j.ccell.2016.11.017]
- 37 **Mauer J**, Luo X, Blanjoie A, Jiao X, Grozhik AV, Patil DP, Linder B, Pickering BF, Vasseur JJ, Chen Q, Gross SS, Elemento O, Debat F, Kiledjian M, Jaffrey SR. Reversible methylation of m⁶Am in the 5' cap controls mRNA stability. *Nature* 2017; **541**: 371-375 [PMID: 28002401 DOI: 10.1038/nature21022]
- 38 **Cui Q**, Shi H, Ye P, Li L, Qu Q, Sun G, Sun G, Lu Z, Huang Y, Yang CG, Riggs AD, He C, Shi Y. m⁶A RNA Methylation Regulates the Self-Renewal and Tumorigenesis of Glioblastoma Stem Cells. *Cell Rep* 2017; **18**: 2622-2634 [PMID: 28297667 DOI: 10.1016/j.celrep.2017.02.059]
- 39 **Zhang S**, Zhao BS, Zhou A, Lin K, Zheng S, Lu Z, Chen Y, Sulman EP, Xie K, Böglér O, Majumder S, He C, Huang S. m⁶A Demethylase ALKBH5 Maintains Tumorigenicity of Glioblastoma Stem-like Cells by Sustaining FOXM1 Expression and Cell Proliferation Program. *Cancer Cell* 2017; **31**: 591-606.e6 [PMID: 28344040 DOI: 10.1016/j.ccell.2017.02.013]
- 40 **Zhang C**, Samanta D, Lu H, Bullen JW, Zhang H, Chen I, He X, Semenza GL. Hypoxia induces the breast cancer stem cell

- phenotype by HIF-dependent and ALKBH5-mediated m⁶A-demethylation of NANOG mRNA. *Proc Natl Acad Sci USA* 2016; **113**: E2047-E2056 [PMID: 27001847 DOI: 10.1073/pnas.1602883113]
- 41 **Alarcón CR**, Goodarzi H, Lee H, Liu X, Tavazoie S, Tavazoie SF. HNRNPA2B1 Is a Mediator of m(6)A-Dependent Nuclear RNA Processing Events. *Cell* 2015; **162**: 1299-1308 [PMID: 26321680 DOI: 10.1016/j.cell.2015.08.011]
 - 42 **Reddy R**, Busch H. Small nuclear RNAs: RNA sequences, structure, and modifications. In: Birnstiel ML, editor. Structure and function of major and minor small nuclear ribonucleoprotein particles. Verlag Berlin Heidelberg: Springer, 1988: 1-37 [DOI: 10.1007/978-3-642-73020-7_1]
 - 43 **Smietanski M**, Werner M, Purta E, Kaminska KH, Stepinski J, Darzynkiewicz E, Nowotny M, Bujnicki JM. Structural analysis of human 2'-O-ribose methyltransferases involved in mRNA cap structure formation. *Nat Commun* 2014; **5**: 3004 [PMID: 24402442 DOI: 10.1038/ncomms4004]
 - 44 **Bélanger F**, Stepinski J, Darzynkiewicz E, Pelletier J. Characterization of hMTTr1, a human Cap1 2'-O-ribose methyltransferase. *J Biol Chem* 2010; **285**: 33037-33044 [PMID: 20713356 DOI: 10.1074/jbc.M110.155283]
 - 45 **Werner M**, Purta E, Kaminska KH, Cymerman IA, Campbell DA, Mittra B, Zamudio JR, Sturm NR, Jaworski J, Bujnicki JM. 2'-O-ribose methylation of cap2 in human: function and evolution in a horizontally mobile family. *Nucleic Acids Res* 2011; **39**: 4756-4768 [PMID: 21310715 DOI: 10.1093/nar/gkr038]
 - 46 **Schimmang T**, Tollervey D, Kern H, Frank R, Hurt EC. A yeast nucleolar protein related to mammalian fibrillarin is associated with small nucleolar RNA and is essential for viability. *EMBO J* 1989; **8**: 4015-4024 [PMID: 2686980]
 - 47 **Lafontaine DL**, Tollervey D. Nop58p is a common component of the box C+D snoRNPs that is required for snoRNA stability. *RNA* 1999; **5**: 455-467 [PMID: 10094313 DOI: 10.1017/S13558382998192X]
 - 48 **Marmier-Gourrier N**, Cléry A, Senty-Ségault V, Charpentier B, Schlotter F, Leclerc F, Fournier R, Branlant C. A structural, phylogenetic, and functional study of 15.5-kD/Snu13 protein binding on U3 small nucleolar RNA. *RNA* 2003; **9**: 821-838 [PMID: 12810916 DOI: 10.1261/rna.2130503]
 - 49 **Balakin AG**, Smith L, Fournier MJ. The RNA world of the nucleolus: two major families of small RNAs defined by different box elements with related functions. *Cell* 1996; **86**: 823-834 [PMID: 8797828 DOI: 10.1016/S0092-8674(00)80156-7]
 - 50 **Kiss-László Z**, Henry Y, Bachellerie JP, Caizergues-Ferrer M, Kiss T. Site-specific ribose methylation of preribosomal RNA: a novel function for small nucleolar RNAs. *Cell* 1996; **85**: 1077-1088 [PMID: 8674114 DOI: 10.1016/S0092-8674(00)81308-2]
 - 51 **Marcel V**, Ghayad SE, Belin S, Therizols G, Morel AP, Solano-González E, Vendrell JA, Hacot S, Mertani HC, Albaret MA, Bourdon JC, Jordan L, Thompson A, Tafer Y, Cong R, Bouvet P, Saurin JC, Catez F, Prats AC, Puisieux A, Diaz JJ. p53 acts as a safeguard of translational control by regulating fibrillarin and rRNA methylation in cancer. *Cancer Cell* 2013; **24**: 318-330 [PMID: 24029231 DOI: 10.1016/j.ccr.2013.08.013]
 - 52 **Pollak MN**, Schernhammer ES, Hankinson SE. Insulin-like growth factors and neoplasia. *Nat Rev Cancer* 2004; **4**: 505-518 [PMID: 15229476 DOI: 10.1038/nrc1387]
 - 53 **Dang CV**. MYC on the path to cancer. *Cell* 2012; **149**: 22-35 [PMID: 22464321 DOI: 10.1016/j.cell.2012.03.003]
 - 54 **Billottet C**, Elkhatib N, Thiery JP, Jouanneau J. Targets of fibroblast growth factor 1 (FGF-1) and FGF-2 signaling involved in the invasive and tumorigenic behavior of carcinoma cells. *Mol Biol Cell* 2004; **15**: 4725-4734 [PMID: 15282342 DOI: 10.1091/mbc.e04-04-0336]
 - 55 **Carmeliet P**. VEGF as a key mediator of angiogenesis in cancer. *Oncology* 2005; **69** Suppl 3: 4-10 [PMID: 16301830 DOI: 10.1159/000088478]
 - 56 **Basu A**, Das P, Chaudhuri S, Bevilacqua E, Andrews J, Barik S, Hatzoglou M, Komar AA, Mazumder B. Requirement of rRNA methylation for 80S ribosome assembly on a cohort of cellular internal ribosome entry sites. *Mol Cell Biol* 2011; **31**: 4482-4499 [PMID: 21930789 DOI: 10.1128/MCB.05804-11]
 - 57 **Choi YW**, Kim YW, Bae SM, Kwak SY, Chun HJ, Tong SY, Lee HN, Shin JC, Kim KT, Kim YJ, Ahn WS. Identification of differentially expressed genes using annealing control primer-based GeneFishing in human squamous cell cervical carcinoma. *Clin Oncol (R Coll Radiol)* 2007; **19**: 308-318 [PMID: 17399965 DOI: 10.1016/j.clon.2007.02.010]
 - 58 **Koh CM**, Gurel B, Sutcliffe S, Aryee MJ, Schultz D, Iwata T, Uemura M, Zeller KI, Anele U, Zheng Q, Hicks JL, Nelson WG, Dang CV, Yegnasubramanian S, De Marzo AM. Alterations in nucleolar structure and gene expression programs in prostatic neoplasia are driven by the MYC oncogene. *Am J Pathol* 2011; **178**: 1824-1834 [PMID: 21435462 DOI: 10.1016/j.ajpath.2010.12.040]
 - 59 **Cowling VH**, Turner SA, Cole MD. Burkitt's lymphoma-associated c-Myc mutations converge on a dramatically altered target gene response and implicate Ncl5a/Nop56 in oncogenesis. *Oncogene* 2014; **33**: 3519-3527 [PMID: 24013231 DOI: 10.1038/onc.2013.338]
 - 60 **Nakamoto K**, Ito A, Watabe K, Koma Y, Asada H, Yoshikawa K, Shinomura Y, Matsuzawa Y, Nojima H, Kitamura Y. Increased expression of a nucleolar Nop5/Sik family member in metastatic melanoma cells: evidence for its role in nucleolar sizing and function. *Am J Pathol* 2001; **159**: 1363-1374 [PMID: 11583964 DOI: 10.1016/S0002-9440(10)62523-0]
 - 61 **Su H**, Xu T, Ganapathy S, Shadfan M, Long M, Huang TH, Thompson I, Yuan ZM. Elevated snoRNA biogenesis is essential in breast cancer. *Oncogene* 2014; **33**: 1348-1358 [PMID: 23542174 DOI: 10.1038/onc.2013.89]
 - 62 **Gao L**, Ma J, Mannoor K, Guarnera MA, Shetty A, Zhan M, Xing L, Stass SA, Jiang F. Genome-wide small nucleolar RNA expression analysis of lung cancer by next-generation deep sequencing. *Int J Cancer* 2015; **136**: E623-E629 [PMID: 25159866 DOI: 10.1002/ijc.29169]
 - 63 **Jha P**, Agrawal R, Pathak P, Kumar A, Purkait S, Mallik S, Suri V, Chand Sharma M, Gupta D, Suri A, Sharma BS, Julka PK, Kulshreshtha R, Sarkar C. Genome-wide small noncoding RNA profiling of pediatric high-grade gliomas reveals deregulation of several miRNAs, identifies downregulation of snoRNA cluster HBII-52 and delineates H3F3A and TP53 mutant-specific miRNAs and snoRNAs. *Int J Cancer* 2015; **137**: 2343-2353 [PMID: 25994230 DOI: 10.1002/ijc.29610]
 - 64 **Berquet L**, Valleron W, Grgurevic S, Quelen C, Zaki O, Quillet-Mary A, Davi F, Brousset P, Bousquet M, Ysebaert L. Small nucleolar RNA expression profiles refine the prognostic impact of IGHV mutational status on treatment-free survival in chronic lymphocytic leukaemia. *Br J Haematol* 2016; **172**: 819-823 [PMID: 26095450 DOI: 10.1111/bjh.13544]
 - 65 **Ravo M**, Cordella A, Rinaldi A, Bruno G, Alexandrova E, Saggese P, Nassa G, Giurato G, Tarallo R, Marchese G, Rizzo F, Stellato C, Biancardi R, Troisi J, Di Spiezio Sardo A, Zullo F, Weisz A, Guida M. Small non-coding RNA deregulation in endometrial carcinogenesis. *Oncotarget* 2015; **6**: 4677-4691 [PMID: 25686835 DOI: 10.18632/oncotarget.2911]
 - 66 **Martens-Uzunova ES**, Jalava SE, Dits NF, van Leenders GJ, Möller S, Trapman J, Bangma CH, Litman T, Visakorpi T, Jenster G. Diagnostic and prognostic signatures from the small non-coding RNA transcriptome in prostate cancer. *Oncogene* 2012; **31**: 978-991 [PMID: 21765474 DOI: 10.1038/onc.2011.304]
 - 67 **Appaiah HN**, Goswami CP, Mina LA, Badve S, Sledge GW Jr, Liu Y, Nakshatri H. Persistent upregulation of U6:SNORD44 small RNA ratio in the serum of breast cancer patients. *Breast Cancer Res* 2011; **13**: R86 [PMID: 21914171 DOI: 10.1186/bcr2943]
 - 68 **COHN WE**. 5-Ribosyl uracil, a carbon-carbon ribofuranosyl nucleoside in ribonucleic acids. *Biochim Biophys Acta* 1959; **32**: 569-571 [PMID: 13811055 DOI: 10.1016/0006-3002(59)90644-4]
 - 69 **Schwartz S**, Bernstein DA, Mumbach MR, Jovanovic M, Herbst RH, León-Ricardo BX, Engreitz JM, Guttman M, Satija R, Lander

- ES, Fink G, Regev A. Transcriptome-wide mapping reveals widespread dynamic-regulated pseudouridylation of ncRNA and mRNA. *Cell* 2014; **159**: 148-162 [PMID: 25219674 DOI: 10.1016/j.cell.2014.08.028]
- 70 **Kim NK**, Theimer CA, Mitchell JR, Collins K, Feigon J. Effect of pseudouridylation on the structure and activity of the catalytically essential P6.1 hairpin in human telomerase RNA. *Nucleic Acids Res* 2010; **38**: 6746-6756 [PMID: 20554853 DOI: 10.1093/nar/gkq525]
- 71 **Penzo M**, Guerrieri AN, Zacchini F, Treré D, Montanaro L. RNA Pseudouridylation in Physiology and Medicine: For Better and for Worse. *Genes* (Basel) 2017; **8** [PMID: 29104216]
- 72 **Walne AJ**, Dokal I. Advances in the understanding of dyskeratosis congenita. *Br J Haematol* 2009; **145**: 164-172 [PMID: 19208095 DOI: 10.1111/j.1365-2141.2009.07598.x]
- 73 **Thumati NR**, Zeng XL, Au HH, Jang CJ, Jan E, Wong JM. Severity of X-linked dyskeratosis congenita (DKCX) cellular defects is not directly related to dyskerin (DKC1) activity in ribosomal RNA biogenesis or mRNA translation. *Hum Mutat* 2013; **34**: 1698-1707 [PMID: 24115260 DOI: 10.1002/humu.22447]
- 74 **Mitchell JR**, Wood E, Collins K. A telomerase component is defective in the human disease dyskeratosis congenita. *Nature* 1999; **402**: 551-555 [PMID: 10591218 DOI: 10.1038/990141]
- 75 **Jády BE**, Bertrand E, Kiss T. Human telomerase RNA and box H/ACA scaRNAs share a common Cajal body-specific localization signal. *J Cell Biol* 2004; **164**: 647-652 [PMID: 14981093 DOI: 10.1083/jcb.200310138]
- 76 **Mitchell JR**, Cheng J, Collins K. A box H/ACA small nucleolar RNA-like domain at the human telomerase RNA 3' end. *Mol Cell Biol* 1999; **19**: 567-576 [PMID: 9858580 DOI: 10.1128/MCB.19.1.567]
- 77 **Penzo M**, Casoli L, Ceccarelli C, Treré D, Ludovini V, Crinò L, Montanaro L. DKC1 gene mutations in human sporadic cancer. *Histol Histopathol* 2013; **28**: 365-372 [PMID: 23348390 DOI: 10.14670/HH-28.365]
- 78 **Jack K**, Bellodi C, Landry DM, Niederer RO, Meskauskas A, Musalgaonkar S, Kopmar N, Krasnykh O, Dean AM, Thompson SR, Ruggero D, Dinman JD. rRNA pseudouridylation defects affect ribosomal ligand binding and translational fidelity from yeast to human cells. *Mol Cell* 2011; **44**: 660-666 [PMID: 22099312 DOI: 10.1016/j.molcel.2011.09.017]
- 79 **Montanaro L**, Calienni M, Bertoni S, Rocchi L, Sansone P, Storci G, Santini D, Ceccarelli C, Taffurelli M, Carnicelli D, Brigotti M, Bonafè M, Treré D, Derenzini M. Novel dyskerin-mediated mechanism of p53 inactivation through defective mRNA translation. *Cancer Res* 2010; **70**: 4767-4777 [PMID: 20501855 DOI: 10.1158/0008-5472.CAN-09-4024]
- 80 **Bellodi C**, Krasnykh O, Haynes N, Theodoropoulou M, Peng G, Montanaro L, Ruggero D. Loss of function of the tumor suppressor DKC1 perturbs p27 translation control and contributes to pituitary tumorigenesis. *Cancer Res* 2010; **70**: 6026-6035 [PMID: 20587522 DOI: 10.1158/0008-5472.CAN-09-4730]
- 81 **Yoon A**, Peng G, Brandenburger Y, Zollo O, Xu W, Rego E, Ruggero D. Impaired control of IRES-mediated translation in X-linked dyskeratosis congenita. *Science* 2006; **312**: 902-906 [PMID: 16690864 DOI: 10.1126/science.1123835]
- 82 **Bellodi C**, Kopmar N, Ruggero D. Deregulation of oncogene-induced senescence and p53 translational control in X-linked dyskeratosis congenita. *EMBO J* 2010; **29**: 1865-1876 [PMID: 20453831 DOI: 10.1038/emboj.2010.83]
- 83 **Carlile TM**, Rojas-Duran MF, Zinshteyn B, Shin H, Bartoli KM, Gilbert WV. Pseudouridine profiling reveals regulated mRNA pseudouridylation in yeast and human cells. *Nature* 2014; **515**: 143-146 [PMID: 25192136 DOI: 10.1038/nature13802]
- 84 **Ronchetti D**, Todoerti K, Tuana G, Agnelli L, Mosca L, Lionetti M, Fabris S, Colapietro P, Miozzo M, Ferrarini M, Tassone P, Neri A. The expression pattern of small nucleolar and small Cajal body-specific RNAs characterizes distinct molecular subtypes of multiple myeloma. *Blood Cancer J* 2012; **2**: e96 [PMID: 23178508 DOI: 10.1038/bcj.2012.41]
- 85 **Valleron W**, Ysebaert L, Berquet L, Fataccioli V, Quelen C, Martin A, Parrens M, Lamant L, de Leval L, Gisselbrecht C, Gaulard P, Brousset P. Small nucleolar RNA expression profiling identifies potential prognostic markers in peripheral T-cell lymphoma. *Blood* 2012; **120**: 3997-4005 [PMID: 22990019 DOI: 10.1182/blood-2012-06-438135]
- 86 **Valleron W**, Laprevotte E, Gautier EF, Quelen C, Demur C, Delabesse E, Agirre X, Prósper F, Kiss T, Brousset P. Specific small nucleolar RNA expression profiles in acute leukemia. *Leukemia* 2012; **26**: 2052-2060 [PMID: 22522792 DOI: 10.1038/leu.2012.111]
- 87 **Han L**, Diao L, Yu S, Xu X, Li J, Zhang R, Yang Y, Werner HMJ, Eterovic AK, Yuan Y, Li J, Nair N, Minelli R, Tsang YH, Cheung LWT, Jeong KJ, Roszik J, Ju Z, Woodman SE, Lu Y, Scott KL, Li JB, Mills GB, Liang H. The Genomic Landscape and Clinical Relevance of A-to-I RNA Editing in Human Cancers. *Cancer Cell* 2015; **28**: 515-528 [PMID: 26439496 DOI: 10.1016/j.ccell.2015.08.013]
- 88 **Paz-Yaacov N**, Bazak L, Buchumenski I, Porath HT, Danan-Gotthold M, Knisbacher BA, Eisenberg E, Levanon EY. Elevated RNA Editing Activity Is a Major Contributor to Transcriptomic Diversity in Tumors. *Cell Rep* 2015; **13**: 267-276 [PMID: 26440895 DOI: 10.1016/j.celrep.2015.08.080]
- 89 **Chen Y**, Wang H, Lin W, Shuai P. ADAR1 overexpression is associated with cervical cancer progression and angiogenesis. *Diagn Pathol* 2017; **12**: 12 [PMID: 28109322 DOI: 10.1186/s13000-017-0600-0]
- 90 **Chan TH**, Lin CH, Qi L, Fei J, Li Y, Yong KJ, Liu M, Song Y, Chow RK, Ng VH, Yuan YF, Tenen DG, Guan XY, Chen L. A disrupted RNA editing balance mediated by ADARs (Adenosine Deaminases that act on RNA) in human hepatocellular carcinoma. *Gut* 2014; **63**: 832-843 [PMID: 23766440 DOI: 10.1136/gutjnl-2012-304037]
- 91 **Chan TH**, Qamra A, Tan KT, Guo J, Yang H, Qi L, Lin JS, Ng VH, Song Y, Hong H, Tay ST, Liu Y, Lee J, Rha SY, Zhu F, So JB, Teh BT, Yeoh KG, Rozen S, Tenen DG, Tan P, Chen L. ADAR-Mediated RNA Editing Predicts Progression and Prognosis of Gastric Cancer. *Gastroenterology* 2016; **151**: 637-650.e10 [PMID: 27373511 DOI: 10.1053/j.gastro.2016.06.043]
- 92 **Qin YR**, Qiao JJ, Chan TH, Zhu YH, Li FF, Liu H, Fei J, Li Y, Guan XY, Chen L. Adenosine-to-inosine RNA editing mediated by ADARs in esophageal squamous cell carcinoma. *Cancer Res* 2014; **74**: 840-851 [PMID: 24302582 DOI: 10.1158/0008-5472.CAN-13-2545]
- 93 **Yeo J**, Goodman RA, Schirle NT, David SS, Beal PA. RNA editing changes the lesion specificity for the DNA repair enzyme NEIL1. *Proc Natl Acad Sci USA* 2010; **107**: 20715-20719 [PMID: 21068368 DOI: 10.1073/pnas.1009231107]
- 94 **Anadón C**, Guil S, Simó-Riudalbas L, Moutinho C, Setien F, Martínez-Cardús A, Moran S, Villanueva A, Calaf M, Vidal A, Lazo PA, Zondervan I, Savola S, Kohno T, Yokota J, Ribas de Pouplana L, Esteller M. Gene amplification-associated overexpression of the RNA editing enzyme ADAR1 enhances human lung tumorigenesis. *Oncogene* 2016; **35**: 4422 [PMID: 27345394 DOI: 10.1038/onc.2016.27]
- 95 **Fujita K**, Murakami Y, Hayashi S. A macromolecular inhibitor of the antizyme to ornithine decarboxylase. *Biochem J* 1982; **204**: 647-652 [PMID: 7126159 DOI: 10.1042/bj2040647]
- 96 **Newman RM**, Mobascher A, Mangold U, Koike C, Diah S, Schmidt M, Finley D, Zetter BR. Antizyme targets cyclin D1 for degradation. A novel mechanism for cell growth repression. *J Biol Chem* 2004; **279**: 41504-41511 [PMID: 15277517 DOI: 10.1074/jbc.M407349200]
- 97 **Mangold U**. Antizyme inhibitor: mysterious modulator of cell proliferation. *Cell Mol Life Sci* 2006; **63**: 2095-2101 [PMID: 16847581 DOI: 10.1007/s00018-005-5583-4]
- 98 **Olsen RR**, Zetter BR. Evidence of a role for antizyme and antizyme inhibitor as regulators of human cancer. *Mol Cancer Res* 2011; **9**: 1285-1293 [PMID: 21849468 DOI: 10.1158/1541-7786.MCR-11-0178]
- 99 **van Duin M**, van Marion R, Vissers K, Watson JE, van Weerden

- WM, Schröder FH, Hop WC, van der Kwast TH, Collins C, van Dekken H. High-resolution array comparative genomic hybridization of chromosome arm 8q: evaluation of genetic progression markers for prostate cancer. *Genes Chromosomes Cancer* 2005; **44**: 438-449 [PMID: 16130124 DOI: 10.1002/gcc.20259]
- 100 **Chin SF**, Teschendorff AE, Marioni JC, Wang Y, Barbosa-Morais NL, Thorne NP, Costa JL, Pinder SE, van de Wiel MA, Green AR, Ellis IO, Porter PL, Tavaré S, Brenton JD, Ylstra B, Caldas C. High-resolution aCGH and expression profiling identifies a novel genomic subtype of ER negative breast cancer. *Genome Biol* 2007; **8**: R215 [PMID: 17925008 DOI: 10.1186/gb-2007-8-10-r215]
- 101 **Chen L**, Li Y, Lin CH, Chan TH, Chow RK, Song Y, Liu M, Yuan YF, Fu L, Kong KL, Qi L, Li Y, Zhang N, Tong AH, Kwong DL, Man K, Lo CM, Lok S, Tenen DG, Guan XY. Recoding RNA editing of AZIN1 predisposes to hepatocellular carcinoma. *Nat Med* 2013; **19**: 209-216 [PMID: 23291631 DOI: 10.1038/nm.3043]
- 102 **Fumagalli D**, Gacquer D, Rothé F, Lefort A, Libert F, Brown D, Kheddoumi N, Shlien A, Konopka T, Salgado R, Larsimont D, Polyak K, Willard-Gallo K, Desmedt C, Piccart M, Abramowicz M, Campbell PJ, Sotiriou C, Detours V. Principles Governing A-to-I RNA Editing in the Breast Cancer Transcriptome. *Cell Rep* 2015; **13**: 277-289 [PMID: 26440892 DOI: 10.1016/j.celrep.2015.09.032]
- 103 **Hu X**, Chen J, Shi X, Feng F, Lau KW, Chen Y, Chen Y, Jiang L, Cui F, Zhang Y, Xu X, Li J. RNA editing of AZIN1 induces the malignant progression of non-small-cell lung cancers. *Tumour Biol* 2017; **39**: 1010428317700001 [PMID: 28849733 DOI: 10.1177/1010428317700001]
- 104 **Popowicz GM**, Schleicher M, Noegel AA, Holak TA. Filamins: promiscuous organizers of the cytoskeleton. *Trends Biochem Sci* 2006; **31**: 411-419 [PMID: 16781869 DOI: 10.1016/j.tibs.2006.05.006]
- 105 **Li JB**, Levanon EY, Yoon JK, Aach J, Xie B, Leproust E, Zhang K, Gao Y, Church GM. Genome-wide identification of human RNA editing sites by parallel DNA capturing and sequencing. *Science* 2009; **324**: 1210-1213 [PMID: 19478186 DOI: 10.1126/science.1170995]
- 106 **Heasman SJ**, Ridley AJ. Mammalian Rho GTPases: new insights into their functions from in vivo studies. *Nat Rev Mol Cell Biol* 2008; **9**: 690-701 [PMID: 18719708 DOI: 10.1038/nrm2476]
- 107 **Leve F**, Morgado-Díaz JA. Rho GTPase signaling in the development of colorectal cancer. *J Cell Biochem* 2012; **113**: 2549-2559 [PMID: 22467564 DOI: 10.1002/jcb.24153]
- 108 **Kanzaki M**. Insulin receptor signals regulating GLUT4 translocation and actin dynamics. *Endocr J* 2006; **53**: 267-293 [PMID: 16702775 DOI: 10.1507/endocrj.KR-65]
- 109 **Han SW**, Kim HP, Shin JY, Jeong EG, Lee WC, Kim KY, Park SY, Lee DW, Won JK, Jeong SY, Park KJ, Park JG, Kang GH, Seo JS, Kim JI, Kim TY. RNA editing in RHOQ promotes invasion potential in colorectal cancer. *J Exp Med* 2014; **211**: 613-621 [PMID: 24663214 DOI: 10.1084/jem.20132209]
- 110 **Yi T**, Mui AL, Krystal G, Ihle JN. Hematopoietic cell phosphatase associates with the interleukin-3 (IL-3) receptor beta chain and down-regulates IL-3-induced tyrosine phosphorylation and mitogenesis. *Mol Cell Biol* 1993; **13**: 7577-7586 [PMID: 8246974 DOI: 10.1128/MCB.13.12.7577]
- 111 **Yi TL**, Cleveland JL, Ihle JN. Protein tyrosine phosphatase containing SH2 domains: characterization, preferential expression in hematopoietic cells, and localization to human chromosome 12p12-p13. *Mol Cell Biol* 1992; **12**: 836-846 [PMID: 1732748 DOI: 10.1128/MCB.12.2.836]
- 112 **Yi T**, Ihle JN. Association of hematopoietic cell phosphatase with c-Kit after stimulation with c-Kit ligand. *Mol Cell Biol* 1993; **13**: 3350-3358 [PMID: 7684496 DOI: 10.1128/MCB.13.6.3350]
- 113 **Pei D**, Wang J, Walsh CT. Differential functions of the two Src homology 2 domains in protein tyrosine phosphatase SH-PTP1. *Proc Natl Acad Sci USA* 1996; **93**: 1141-1145 [PMID: 8577729 DOI: 10.1073/pnas.93.3.1141]
- 114 **Beghini A**, Ripamonti CB, Peterlongo P, Roversi G, Cairoli R, Morra E, Larizza L. RNA hyperediting and alternative splicing of hematopoietic cell phosphatase (PTPN6) gene in acute myeloid leukemia. *Hum Mol Genet* 2000; **9**: 2297-2304 [PMID: 11001933 DOI: 10.1093/oxfordjournals.hmg.a018921]
- 115 **Song IH**, Kim YA, Heo SH, Park IA, Lee M, Bang WS, Park HS, Gong G, Lee HJ. ADAR1 expression is associated with tumour-infiltrating lymphocytes in triple-negative breast cancer. *Tumour Biol* 2017; **39**: 1010428317734816 [PMID: 29022489 DOI: 10.1177/1010428317734816]
- 116 **Farrant M**, Kaila K. The cellular, molecular and ionic basis of GABA(A) receptor signalling. *Prog Brain Res* 2007; **160**: 59-87 [PMID: 17499109 DOI: 10.1016/S0079-6123(06)60005-8]
- 117 **Daniel C**, Wahlstedt H, Ohlson J, Björk P, Ohman M. Adenosine-to-inosine RNA editing affects trafficking of the gamma-aminobutyric acid type A (GABA(A)) receptor. *J Biol Chem* 2011; **286**: 2031-2040 [PMID: 21030585 DOI: 10.1074/jbc.M110.130096]
- 118 **Gumireddy K**, Li A, Kossenkova AV, Sakurai M, Yan J, Li Y, Xu H, Wang J, Zhang PJ, Zhang L, Showe LC, Nishikura K, Huang Q. The mRNA-edited form of GABRA3 suppresses GABRA3-mediated Akt activation and breast cancer metastasis. *Nat Commun* 2016; **7**: 10715 [PMID: 26869349 DOI: 10.1038/ncomms10715]
- 119 **Maas S**, Patt S, Schrey M, Rich A. Underediting of glutamate receptor GluR-B mRNA in malignant gliomas. *Proc Natl Acad Sci USA* 2001; **98**: 14687-14692 [PMID: 11717408 DOI: 10.1073/pnas.251531398]
- 120 **Chen YB**, Liao XY, Zhang JB, Wang F, Qin HD, Zhang L, Shugart YY, Zeng YX, Jia WH. ADAR2 functions as a tumor suppressor via editing IGFBP7 in esophageal squamous cell carcinoma. *Int J Oncol* 2017; **50**: 622-630 [PMID: 28035363 DOI: 10.3892/ijo.2016.3823]
- 121 **Higuchi M**, Single FN, Köhler M, Sommer B, Sprengel R, Seeburg PH. RNA editing of AMPA receptor subunit GluR-B: a base-paired intron-exon structure determines position and efficiency. *Cell* 1993; **75**: 1361-1370 [PMID: 8269514 DOI: 10.1016/0092-8674(93)90622-W]
- 122 **Evdokimova V**, Tognon CE, Benatar T, Yang W, Krutikov K, Pollak M, Sorensen PH, Seth A. IGFBP7 binds to the IGF-1 receptor and blocks its activation by insulin-like growth factors. *Sci Signal* 2012; **5**: ra92 [PMID: 23250396 DOI: 10.1126/scisignal.2003184]
- 123 **Mutaguchi K**, Yasumoto H, Mita K, Matsubara A, Shiina H, Igawa M, Dahiya R, Usui T. Restoration of insulin-like growth factor binding protein-related protein 1 has a tumor-suppressive activity through induction of apoptosis in human prostate cancer. *Cancer Res* 2003; **63**: 7717-7723 [PMID: 14633696]
- 124 **Ruan W**, Xu E, Xu F, Ma Y, Deng H, Huang Q, Lv B, Hu H, Lin J, Cui J, Di M, Dong J, Lai M. IGFBP7 plays a potential tumor suppressor role in colorectal carcinogenesis. *Cancer Biol Ther* 2007; **6**: 354-359 [PMID: 17312390 DOI: 10.4161/cbt.6.3.3702]
- 125 **Benatar T**, Yang W, Amemiya Y, Evdokimova V, Kahn H, Holloway C, Seth A. IGFBP7 reduces breast tumor growth by induction of senescence and apoptosis pathways. *Breast Cancer Res Treat* 2012; **133**: 563-573 [PMID: 21997538 DOI: 10.1007/s10549-011-1816-4]
- 126 **Galeano F**, Rossetti C, Tomaselli S, Cifaldi L, Lezzerini M, Pezzullo M, Boldrini R, Massimi L, Di Rocco CM, Locatelli F, Gallo A. ADAR2-editing activity inhibits glioblastoma growth through the modulation of the CDC14B/Skp2/p21/p27 axis. *Oncogene* 2013; **32**: 998-1009 [PMID: 22525274 DOI: 10.1038/onc.2012.125]
- 127 **Karsy M**, Arslan E, Moy F. Current Progress on Understanding MicroRNAs in Glioblastoma Multiforme. *Genes Cancer* 2012; **3**: 3-15 [PMID: 22893786 DOI: 10.1177/1947601912448068]
- 128 **Tomaselli S**, Galeano F, Alon S, Raho S, Galardi S, Polito VA, Presutti C, Vincenti S, Eisenberg E, Locatelli F, Gallo A. Modulation of microRNA editing, expression and processing by ADAR2 deaminase in glioblastoma. *Genome Biol* 2015; **16**: 5 [PMID: 25582055 DOI: 10.1186/s13059-014-0575-z]
- 129 **Choudhury Y**, Tay FC, Lam DH, Sandanaraj E, Tang C, Ang BT, Wang S. Attenuated adenosine-to-inosine editing of microRNA-376a* promotes invasiveness of glioblastoma cells. *J Clin Invest*

- 2012; **122**: 4059-4076 [PMID: 23093778 DOI: 10.1172/JCI62925]
- 130 **Nemlich Y**, Greenberg E, Ortenberg R, Besser MJ, Barshack I, Jacob-Hirsch J, Jacoby E, Eyal E, Rivkin L, Prieto VG, Chakravarti N, Duncan LM, Kallenberg DM, Galun E, Bennett DC, Amariglio N, Bar-Eli M, Schachter J, Rechavi G, Markel G. MicroRNA-mediated loss of ADAR1 in metastatic melanoma promotes tumor growth. *J Clin Invest* 2013; **123**: 2703-2718 [PMID: 23728176 DOI: 10.1172/JCI62980]
- 131 **Ota H**, Sakurai M, Gupta R, Valente L, Wulff BE, Ariyoshi K, Iizasa H, Davuluri RV, Nishikura K. ADAR1 forms a complex with Dicer to promote microRNA processing and RNA-induced gene silencing. *Cell* 2013; **153**: 575-589 [PMID: 23622242 DOI: 10.1016/j.cell.2013.03.024]
- 132 **Shoshan E**, Mobley AK, Braeuer RR, Kamiya T, Huang L, Vasquez ME, Salameh A, Lee HJ, Kim SJ, Ivan C, Velazquez-Torres G, Nip KM, Zhu K, Brooks D, Jones SJ, Birol I, Mosqueda M, Wen YY, Eterovic AK, Sood AK, Hwu P, Gershenwald JE, Robertson AG, Calin GA, Markel G, Fidler IJ, Bar-Eli M. Reduced adenosine-to-inosine miR-455-5p editing promotes melanoma growth and metastasis. *Nat Cell Biol* 2015; **17**: 311-321 [PMID: 25686251 DOI: 10.1038/ncb3110]
- 133 **Squires JE**, Patel HR, Nousch M, Sibbritt T, Humphreys DT, Parker BJ, Suter CM, Preiss T. Widespread occurrence of 5-methylcytosine in human coding and non-coding RNA. *Nucleic Acids Res* 2012; **40**: 5023-5033 [PMID: 22344696 DOI: 10.1093/nar/gks144]
- 134 **Metodiev MD**, Spähr H, Loguerio Polosa P, Meharg C, Becker C, Altmueller J, Habermann B, Larsson NG, Ruzzenente B. NSUN4 is a dual function mitochondrial protein required for both methylation of 12S rRNA and coordination of mitochondrial assembly. *PLoS Genet* 2014; **10**: e1004110 [PMID: 24516400 DOI: 10.1371/journal.pgen.1004110]
- 135 **Frye M**, Watt FM. The RNA methyltransferase Misu (NSun2) mediates Myc-induced proliferation and is upregulated in tumors. *Curr Biol* 2006; **16**: 971-981 [PMID: 16713953 DOI: 10.1016/j.cub.2006.04.027]
- 136 **Goll MG**, Kirpekar F, Maggert KA, Yoder JA, Hsieh CL, Zhang X, Golic KG, Jacobsen SE, Bestor TH. Methylation of tRNA^{Asp} by the DNA methyltransferase homolog Dnmt2. *Science* 2006; **311**: 395-398 [PMID: 16424344 DOI: 10.1126/science.1120976]
- 137 **Huang W**, Qi CB, Lv SW, Xie M, Feng YQ, Huang WH, Yuan BF. Determination of DNA and RNA Methylation in Circulating Tumor Cells by Mass Spectrometry. *Anal Chem* 2016; **88**: 1378-1384 [PMID: 26707930 DOI: 10.1021/acs.analchem.5b03962]

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Quality of life and oral potentially malignant disorders: Critical appraisal and prospects

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Abstract

Quality of life (QoL) is a vital and often required health outcome measure that is relevant to patient care. A healthy oral cavity enables person to perform daily activities without any limitations. However, any disturbance may result in impaired QoL. The oral health-remains an essential element of people's health and well-being. In recent years, the tradition of clinical practice and research has been changed by incorporating QoL assessment, as it helps in assessment of patients' needs and monitoring treatment responses. Oral potentially malignant disorders (OPMDs) are a group of chronic disorders including oral leukoplakia (OL), oral lichen planus and oral submucous fibrosis (OSF). It is evident that patients with OPMDs experience significant health-related symptoms, functional limitations and psycho-social impairment, compromising their QoL. Moreover, the worsening of QoL has been associated with advanced stages of OPMDs. Despite of increasing number of OPMD cases in recent decades, limited literature is available regarding QoL in this population. Although, there is higher prevalence of habit-related OPMDs, particularly OSF and OL in Southern Asian countries, only a few studies have been performed in these populations. Moreover, these studies administered generic QoL instruments, which offer less sensitivity to clinical changes. However, condition-

specific instruments are more sensitive and allows better measurement of QoL. As the impacts of different conditions on OHRQoL may vary, the development and validation of a QoL instrument specific to each clinical entity of OPMDs is currently needed.

Key words: Quality of life; Oral potentially malignant disorders; Oral submucous fibrosis; Oral lichen planus; Oral leukoplakia

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Core tip: The quality of life (QoL) assessment has become an essential tool in clinical practice to better understand patient reported outcomes in recent years. It definitely helps to better understand the impact of oral health on the lives of patients with oral potentially malignant disorders (OPMDs) and their families and to monitor the outcomes of treatments. It is a foremost pre-requisite to employ the best available QoL instrument when treating OPMDs. In view of the scarcity of research on QoL assessments in OPMDs, the development and application of condition-specific QoL instruments can allow them to become tools to better understand and shape the state of clinical practice, dental research and dental education.

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INTRODUCTION

The World Health Organization (WHO) has defined quality of life (QoL) as "an individual's perception of his position in life in the context of the culture and value system in which he lives and in relation to his goals, expectations and standards and concerns"^[1]. In recent years, OHRQoL has become increasingly important in patient care and extensively applied as a part of daily practice^[2]. A healthy oral cavity empowers an individual to perform routine daily activities without any physical and psycho-social limitations. However, any disturbance related with the oral cavity may disturb normal oral functions. Persistent discomfort and a functionally impaired oral cavity may subsequently result in decreased self-confidence and social communication of the individual, compromising his or her QoL. It is well-known that OHRQoL remains an essential element of people's health and well-being and helps in assessment of patients' needs and to monitor treatment responses^[3,4]. Even though the impacts of oral diseases can be assessed by traditional methods, there is growing trend of availing patients' perspectives. Therefore,

the new era demands QoL assessment using patient reported outcomes (PROs) and experiences (PREs) as a part of day-to-day practice^[5]. Moreover, deciding proper treatment protocols and measuring treatment outcomes based on PROs and PREs is definitely helpful and has changed the tradition of clinical practice, surveys and research in recent years.

Oral potentially malignant disorders (OPMDs) are a group of chronic disorders with increased morbidity and mortality due to cancerous changes^[6]. Per recent literature, the values of the malignant potential of oral leukoplakia (OL), oral lichen planus (OLP) and oral submucous fibrosis (OSF) are 3.5% (range, 0.13-34.0%)^[7], 1.1%^[8] and 7%-13%^[9], respectively. Careful monitoring of these lesions by an experienced specialist is highly recommended to identify any malignant changes in the early stages to reduce the cancer burden. It has been documented that patients with OPMDs experience significant health-related symptoms affecting their QoL^[10]. Moreover, OPMD patients shown psychological impairment due to their fear of developing cancer^[11]. These patients also reported to have social and emotional imbalance. Although oral cancer (OC) and OPMDs presents relatively similar health comorbidities; compromising the QoL^[12], the available OHRQoL instruments are OC/head and neck cancer specific, and thus, the OHRQoL of patients suffering from OPMDs is seldom assessed. Moreover, the literature on QoL assessment in this population is scanty in contrast to the plentiful literature on QoL in OC/head and neck cancer patients^[13,14].

OSF is an OPMD that is highly prevalent in Indian subcontinents and South-East Asia, affecting 5 million people in India alone^[9]. Its etiology is multifactorial but arecoline in the areca nut is the main causative agent in initiating the disease process. OSF is clinically characterized by a early sign and a symptoms of burning sensation, vesiculation and ulceration in the oral cavity and lately followed by blanching of the oral mucosa. This results in to increasing stiffness and marked rigidity of the tissues leading to reduced mouth opening, significantly compromising the patient's QoL. It is evident that OSF have detrimental effects on OHRQoL and the worsening of QoL has been associated with advanced stages of OSF^[15].

OLP is a chronic inflammatory disorder with etiopathogenesis that is still poorly understood. OLP affects approximately 1%-2% of the population worldwide^[16] and is more prevalent in middle-aged females. It is characterized by outbreaks or flares of different types of clinical presentations, which has been categorized by Eisen^[17] into three subtypes: (1) reticular form; (2) erosive/atrophic form; and (3) ulcerative form. Even though the reticular form is asymptomatic, erosive and ulcerative forms are often painful and disabling and are variants with burning sensations of the oral mucosa. The persistent painful symptoms can have a significant negative impact on daily life activities including eating, swallowing or speaking. Moreover, OLP has been linked

with impaired psychosocial morbidity and QoL^[4,18].

The prevalence of OL is approximately 1%, with a greater number of cases seen in adults. The etiology of OL includes chewing or smoking of tobacco and related products. Clinically, OL can be classified into homogenous and non-homogenous subtypes, with the highest malignant potential reported in proliferative verrucous leukoplakia and speckled leukoplakia. OHRQoL of patients with OL was evaluated in a few past studies^[19,20].

Our recent systematic review demonstrated that the QoL of patients affected by different OPMDs has been studied and successfully assessed by various authors using different QoL instruments in European countries. However, most of these studies have focused on QoL in patients with OLP, which is not at all applicable to all OPMDs^[21]. Despite the fact that habit-related OPMDs, such as OSF and OL are highly prevalent in Southern Asian countries^[22], surprisingly, only a few studies have assessed QoL in patients with OSF and OL in this population to our knowledge. Moreover, all these studies administered QoL instruments, namely the Oral Health Impact Profile (OHIP), University of Washington Quality of Life Questionnaire (UW-QOL), Chronic Oral Mucosal Disease Questionnaire (COMDQ) and Oral Health Related Quality of Life-UK (OHRQoL-UK). However, these instruments are generic to a range of chronic oral mucosal diseases and are not condition-specific. The generic questionnaires offer less sensitivity to clinical changes than disease-specific tools^[23], as they are applicable to a wide variety of population and disease states. In contrast, it is well-known that condition-specific instruments allow for better measurement of QoL than generic questionnaires, as they evaluate the effects of a concerned disease on life quality of an individual. A condition specific QoL tool for OPMD, *i.e.*, the OPMDQoL questionnaire study, observed a significant impact of OLP and OSF compared to OL on the QoL of affected patients especially in the subscales of "physical impairment and functional limitations"^[24]. Recently, we developed and validated a condition-specific instrument for OSF patients. This was found reliable in QoL evaluation tool in an Indian population^[25].

We believe that QoL assessment has become a necessity to determine the feelings and perceptions of patients as well as to increase effective communication between health care professionals and patients. This definitely provides clues not only to better understand the influence of oral diseases on the patients and their families but also to monitor the outcomes of the treatments provided. Currently, increased incidence of OPMDs specifically OSF and OL in South Asian countries, is an alarming situation as far as oral cancer is concerned. This might be due to the increased popularity of commercially available areca nut and tobacco preparations, especially in India. In addition, an increasing number of young people are becoming addicted to this ancient, socially acceptable habit due to easy access, effective price changes and marketing strategies. In view of the scarcity

of research on QoL assessment in OPMDs, there is a dire need for more studies to better understand this situation. It is evident that researchers have been continuously focusing on improving the QoL of affected individuals. Therefore, it is a foremost pre-requisite to employ the best available QoL instrument in OPMDs. Furthermore, due to differences in their pathogenesis and clinical presentations and thus, differing impacts on OHRQoL, the development and validation of a QoL instrument specific to each clinical entity of OPMD separately is needed. Such condition-specific instruments can become tools of choice in future researches and help to improve QoL of affected individuals.

REFERENCES

- 1 The World Health Organization Quality of Life assessment (WHOQOL): position paper from the World Health Organization. *Soc Sci Med* 1995; **41**: 1403-1409 [PMID: 8560308 DOI: 10.1016/0277-9536(95)00112-K]
- 2 Sischo L, Broder HL. Oral health-related quality of life: what, why, how, and future implications. *J Dent Res* 2011; **90**: 1264-1270 [PMID: 21422477 DOI: 10.1177/0022034511399918]
- 3 Cano SJ, Klassen A, Pusic AL. The science behind quality-of-life measurement: a primer for plastic surgeons. *Plast Reconstr Surg* 2009; **123**: 98e-106e [PMID: 19319025 DOI: 10.1097/PRS.0b013e31819565c1]
- 4 López-Jornet P, Camacho-Alonso F. Quality of life in patients with oral lichen planus. *J Eval Clin Pract* 2010; **16**: 111-113 [PMID: 20367822 DOI: 10.1111/j.1365-2753.2009.01124.x]
- 5 Gondivkar SM, Gadgil AR, Sarode SC, Patil S. Quality of Life Assessment should be Part of Oral Health Evaluations in Day-to-day Practice. *J Contemp Dent Pract* 2017; **18**: 857-858 [PMID: 28989120 DOI: 10.5005/jp-journals-10024-2139]
- 6 Warnakulasuriya S, Johnson NW, van der Waal I. Nomenclature and classification of potentially malignant disorders of the oral mucosa. *J Oral Pathol Med* 2007; **36**: 575-580 [PMID: 17944749 DOI: 10.1111/j.1600-0714.2007.00582.x]
- 7 Warnakulasuriya S, Ariyawardana A. Malignant transformation of oral leukoplakia: a systematic review of observational studies. *J Oral Pathol Med* 2016; **45**: 155-166 [PMID: 26189354 DOI: 10.1111/jop.12339]
- 8 Aghbari SMH, Abushouk AI, Attia A, Elmarazy A, Menshawy A, Ahmed MS, Elsaadany BA, Ahmed EM. Malignant transformation of oral lichen planus and oral lichenoid lesions: A meta-analysis of 20095 patient data. *Oral Oncol* 2017; **68**: 92-102 [PMID: 28438300 DOI: 10.1016/j.oraloncology.2017.03.012]
- 9 Hsue SS, Wang WC, Chen CH, Lin CC, Chen YK, Lin LM. Malignant transformation in 1458 patients with potentially malignant oral mucosal disorders: a follow-up study based in a Taiwanese hospital. *J Oral Pathol Med* 2007; **36**: 25-29 [PMID: 17181738 DOI: 10.1111/j.1600-0714.2006.00491.x]
- 10 Raja JV, Rai P, Kumar NC, Khan M, Chandrashekar H. Psychiatric morbidity among patients with oral submucous fibrosis: a controlled study. *Oral Health Dent Manag* 2013; **12**: 85-94 [PMID: 23756424]
- 11 Tadakamadla J, Kumar S, Johnson NW. Quality of life in patients with oral potentially malignant disorders: a systematic review. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2015; **119**: 644-655 [PMID: 25956217 DOI: 10.1016/j.oooo.2015.01.025]
- 12 Rana M, Gellrich NC, Rana M. Comparison of health-related quality of life of patients with different precancer and oral cancer stages. *Clin Oral Investig* 2015; **19**: 481-488 [PMID: 24878612 DOI: 10.1007/s00784-014-1265-7]
- 13 Moore KA, Ford PJ, Farah CS. Support needs and quality of life

- in oral cancer: a systematic review. *Int J Dent Hyg* 2014; **12**: 36-47 [PMID: 24034791 DOI: 10.1111/idh.12051]
- 14 **Torres-Carranza E**, Infante-Cossio P, Hernández-Guisado JM, Hens-Aumente E, Gutierrez-Pérez JL. Assessment of quality of life in oral cancer. *Med Oral Patol Oral Cir Bucal* 2008; **13**: E735-E741 [PMID: 18978717]
 - 15 **Gondivkar SM**, Bhowate RR, Gadail AR, Sarode SC, Gondivkar RS, Yuwanati M, Patil S. Quality of Life-related "Patient-reported Outcome Measures" in Oral Submucous Fibrosis Patients. *J Contemp Dent Pract* 2018; **19**: 331-338 [PMID: 29603708 DOI: 10.5005/jp-journals-10024-2262]
 - 16 **McCartan BE**, Healy CM. The reported prevalence of oral lichen planus: a review and critique. *J Oral Pathol Med* 2008; **37**: 447-453 [PMID: 18624932 DOI: 10.1111/j.1600-0714.2008.00662.x]
 - 17 **Eisen D**. The therapy of oral lichen planus. *Crit Rev Oral Biol Med* 1993; **4**: 141-158 [PMID: 8435463 DOI: 10.1177/10454411930040020101]
 - 18 **Lopez-Jornet P**, Martinez-Canovas A, Pons-Fuster A. Salivary biomarkers of oxidative stress and quality of life in patients with oral lichen planus. *Geriatr Gerontol Int* 2014; **14**: 654-659 [PMID: 24205825 DOI: 10.1111/ggi.12153]
 - 19 **Llewellyn CD**, Warnakulasuriya S. The impact of stomatological disease on oral health-related quality of life. *Eur J Oral Sci* 2003; **111**: 297-304 [PMID: 12887394 DOI: 10.1034/j.1600-0722.2003.00057.x]
 - 20 **Silverman S Jr**. Mucosal lesions in older adults. *J Am Dent Assoc* 2007; **138** Suppl: 41S-46S [PMID: 17761845 DOI: 10.14219/jada.archive.2007.0362]
 - 21 **Gondivkar SM**, Gadail AR, Gondivkar RS, Sarode SC, Sarode GS, Patil S. Impact of oral potentially malignant disorders on quality of life: a systematic review. *Future Oncol* 2018; **14**: 995-1010 [PMID: 29561169 DOI: 10.2217/fon-2017-0577]
 - 22 **Gupta PC**, Mehta FS, Daftary DK, Pindborg JJ, Bhonsle RB, Jalnawalla PN, Sinor PN, Pitkar VK, Murti PR, Irani RR, Shah HT, Kadam PM, Iyer KS, Iyer HM, Hegde AK, Chandrashekar GK, Shiroff BC, Sahiar BE, Mehta MN. Incidence rates of oral cancer and natural history of oral precancerous lesions in a 10-year follow-up study of Indian villagers. *Community Dent Oral Epidemiol* 1980; **8**: 283-333 [PMID: 6937277 DOI: 10.1111/j.1600-0528.1980.tb01302.x]
 - 23 **Kaplan SH**, Kravitz RL, Greenfield S. A critique of current uses of health status for the assessment of treatment effectiveness and quality of care. *Med Care* 2000; **38**: II184-II191 [PMID: 10982106 DOI: 10.1097/00005650-200009002-00029]
 - 24 **Tadakamadla J**, Kumar S, Lalloo R, Gandhi Babu DB, Johnson NW. Impact of oral potentially malignant disorders on quality of life. *J Oral Pathol Med* 2018; **47**: 60-65 [PMID: 28766765 DOI: 10.1111/jop.12620]
 - 25 **Gondivkar SM**, Bhowate RR, Gadail AR, Gaikwad RN, Gondivkar RS, Sarode SC, Sarode GS. Development and validation of oral health-related quality of life measure in oral submucous fibrosis. *Oral Dis* 2018 [PMID: 29570905 DOI: 10.1111/odi.12857]

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Polyubiquitination inhibition of estrogen receptor alpha and its implications in breast cancer

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Abstract

Estrogen receptor alpha (ER α) is detected in more than 70% of the cases of breast cancer. Nuclear activity of ER α , a transcriptional regulator, is linked to the development of mammary tumors, whereas the extranuclear activity of ER α is related to endocrine therapy resistance. ER α polyubiquitination is induced by the estradiol hormone, and also by selective estrogen receptor degraders, resulting in ER α degradation *via* the ubiquitin proteasome system. Moreover, polyubiquitination is related to the ER α transcription cycle, and some E3-ubiquitin ligases also function as coactivators for ER α . Several studies have demonstrated that ER α polyubiquitination is inhibited by multiple mechanisms that include posttranslational modifications, interactions with coregulators, and formation of specific protein complexes with ER α . These events are responsible for an increase in ER α protein levels and deregulation of its signaling in breast cancers. Thus, ER α polyubiquitination inhibition may be a key factor in the progression of breast cancer and resistance to endocrine therapy.

Key words: Estrogen receptor alpha polyubiquitination; Breast cancer; Estrogen receptor alpha

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Core tip: The inhibition of the estrogen receptor alpha polyubiquitination and degradation by several molecular mechanisms is related to the progression of breast cancer and resistance to endocrine therapy.

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INTRODUCTION

Estrogen receptor alpha (ER α) protein, also known as nuclear receptor subfamily3 group A member 1 (NR3A1), comprises of 595 amino acids, organized in two activation function domains (AF-1 and AF-2), a DNA-binding domain (DBD), a ligand-binding domain (LBD) that recognize the 17 β -estradiol hormone (E2), and a hinge region that connects the DBD and the LBD^[1-3] (Figure 1). Many nuclear functions of ER α are triggered by the binding of E2 to the receptor^[4,5], inducing ER α homodimers to bind to estrogen responsive elements (ERE) within the enhancer and promoter regions of E2-target genes^[6,7]. In these events, pioneer factors expose chromatin sections, facilitating the association of ER α with EREs^[8]. Moreover, transcriptional coregulators are recruited by the AF-1 and AF-2 domains of the receptor for the remodeling of the chromatin structure^[9,10] and promotion of chromatin loops that modulate E2-responsive gene expression^[11,12]. In addition, there is crosstalk between ER α and other signaling pathways: ER α acts as a coregulator by interacting with other transcription factors, such as activator protein 1 (AP-1), specificity protein 1 (Sp1), and nuclear factor- κ B (NF- κ B)^[3,5,13-17]. Additionally, ER α is phosphorylated and transcriptionally activated in response to growth factors such as the epidermal growth factor (EGF) and insulin-like growth factor (IGF)^[13,14,18-20]. Recently, progesterone receptor (PR) was shown as an ER α interacting protein that modulates and re-directs the binding of ER α to the chromatin and the expression of specific genes in breast cancer cells^[21] (Figure 2).

ER α also exhibits extranuclear activity by associating with the cell membrane *via* palmitoylation, and with the help of protein complexes, linked to the cell membrane or cytoplasm^[22] (Figure 2). Thereafter, ER α transduces rapid extranuclear signaling that can trigger second messengers such as calcium and cAMP, and activate kinases such as ERK/MAPK, PI3K/AKT, PKC and Src kinase^[13,23,24]. Both nuclear and extranuclear signaling of ER α are connected and are critical in about 70% of breast cancer cases (ER α + breast cancer)^[13,24,25]. Consequently, ER α is a target for endocrine therapy *via* the use of selective estrogen receptor modulators (SERMs), such as tamoxifen (Tam), which competes with E2 by binding to ER α to inhibit its transcriptional activity, as well as, *via* the use of selective estrogen receptor degraders (SERDs) such as fulvestrant that decreases the ER α stability^[8,14,26,27]. The acquisition of resistance to these treatments commonly occurs in ER α + breast cancer, and although the mechanisms are unclear, the

extranuclear signaling of ER α is strongly activated under this condition^[19,20,26,28-31].

The activation or inhibition of ER α activity is modulated by its transcriptional coregulators, by phosphorylation induced by E2 hormones and growth factors, and by other posttranslational modifications such as ubiquitination. Remarkably, several studies have emerged to demonstrate that multiple mechanisms are activated in ER α + breast cancers to inhibit ER α polyubiquitination, increasing its signaling pathways (Figure 2), which have crucial implications in the progression of this cancer type, as we will describe in the following sections.

GENERALITIES OF THE POSTTRANSLATIONAL MODIFICATION “UBIQUITINATION” FOR ER α IN BREAST CANCER CELLS

ER α is a monoubiquitination and polyubiquitination-target. However, fewer reports are available to demonstrate monoubiquitination of ER α , in comparison to those that exhibit polyubiquitination of this receptor. Nevertheless, these studies clearly show that ER α monoubiquitination is decreased by E2, and that, this modification is important, both for stability and for the transcriptional activity of this receptor in breast cancer. In contrast, polyubiquitination is induced by E2, resulting in a signal to direct ER α degradation *via* the UPS^[14,32,33], facilitated by the concerted action of the enzymes E1 (ubiquitin activating enzyme), E2 (ubiquitin conjugating enzyme), and E3 (ubiquitin ligase)^[32,33]. The specific covalent binding of ubiquitin to ER α lysine residues is mediated by several E3 ubiquitin ligases for ER α , that include CHIP^[34], E6AP^[35], BRCA1^[36], BARD1^[37], SKP2^[38], MDM2^[39], and Hbo1^[40]. Importantly, E2 treatment induces ER α polyubiquitination, followed by its degradation by the UPS^[14,17,33,41-43].

Although polyubiquitination leads to ER α downregulation through its degradation by the 26S proteasome, it is important to note that, this modification and the proteasome activity, have also been reported as elements required for the transcriptional cycle of ER α . Likewise, it has been evidenced that ER α bound to ERE can recruit coactivators, some of which possess E3-ubiquitin ligase activity, such as SKP2^[17], E6AP, and RNF8. As coactivators enhance the activity of ER α , and the activity of E3-ubiquitin ligases mediate the downregulation of this receptor, the recruitment of these proteins with dual function may maintain a balance in the level and activity of ER α ^[17,44,45].

ER α residues, K302 and K303, have been suggested as the lysine targets for ubiquitination and degradation, in response to E2 and fulvestrant, but the same residues are also important for ER α stability in untreated breast cancer cells^[46]. Against this background, it maybe envisaged that, several factors delicately modulate the stability and degradation of ER α , which may be altered

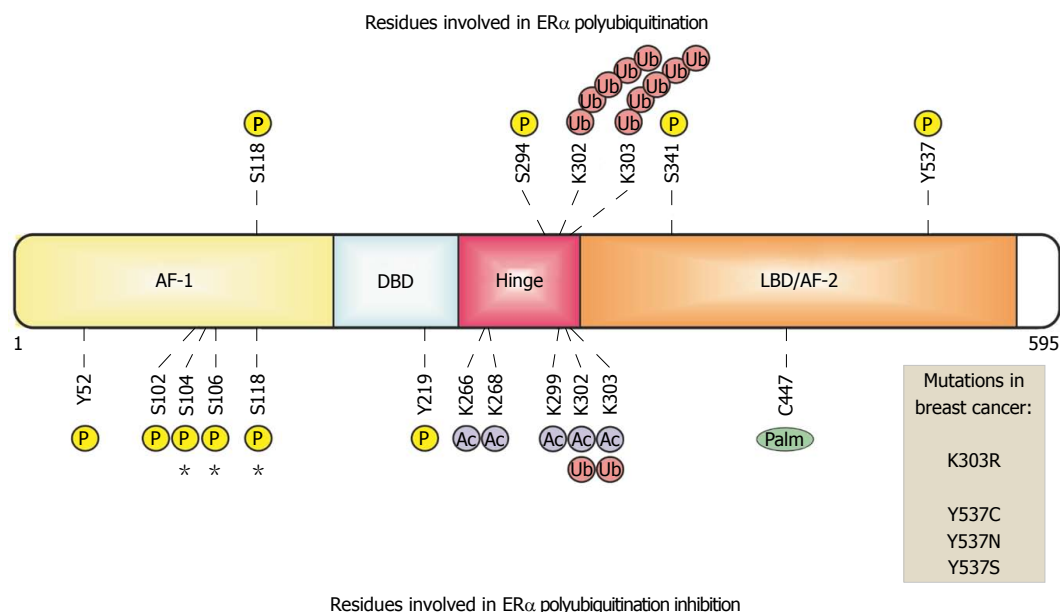


Figure 1 Estrogen receptor α in breast cancer cells. ER α is organized in functional domains. The transactivation domains AF-1 and AF-2 recruit both coactivators and corepressors. The DNA-binding domain (DBD) recognizes and binds to estrogen response elements in enhancers or promoters. The ligand-binding domain (LBD) is recognized and activated by the 17 beta estradiol hormone. The hinge domain links LBD and DBD allowing the conformational changes of this receptor. Some residues are modified by phosphorylation, acetylation, ubiquitination and palmitoylation, which are related with ER α polyubiquitination. Sites of phosphorylation or mutations in ER α that have been identified in breast-cancer biopsy samples are indicated.

in breast cancer.

Additionally, the ubiquitination of ER α is also related to its phosphorylation state. Several kinases, such as CDK11p58^[47], cyclin E-CDK2^[17], Src^[35], PKC^[42], p38MAPK^[38], and ERK7^[48] have been reported as modifiers of ER α in breast cancer. The main residues of ER α that are phosphorylated in E2-response, and have been associated with its polyubiquitination and degradation, are S118^[49], S294^[38], S341^[17], and Y537^[35]. A key example is the sequential modification of ER α , where, first, the ER α Y537 residue is phosphorylated by Src kinase in E2-treated cells, followed by E6AP, an E3-ubiquitin ligase, which induces ER α polyubiquitination and its degradation^[35]. Thus, phosphorylation and ubiquitination of ER α are interconnected in order to control both, the abundance and the functions of this receptor.

IS ER α IN BREAST CANCER CELLS POLYUBIQUITINATED AND DEGRADED?

In recent years, several studies have emerged to demonstrate the inhibition of polyubiquitination of ER α and consequently, a decrease in its degradation *via* the UPS, increasing its protein stability in breast cancer cells, through several mechanisms and ER α -associated proteins. Here, we describe these evidences.

ER α polyubiquitination inhibitor proteins in breast cancer cells

ER α polyubiquitination inhibitor proteins (EPIP). There has been a progressive increase in the number of ER α polyubiquitination inhibitor proteins that have been

discovered in breast cancer cells, which we have grouped and identified as EPIP. So far, it has been reported that proteins such as Mucin 1 (MUC1), PIN1, GSK3, LMTK3, RNF8, RNF31, RB, ABL, SHARPIN, and SMURF1 have the ability to interact with ER α , conferring it protection against polyubiquitination and degradation. Interestingly, not all of these proteins have related sequences and structures, but some of them are functionally similar.

MUC1 and Protein interacting with Never in mitosis A (PIN1), for example, induce the formation of stable transcription complexes on the DNA^[49,50]. MUC1 interacts with ER α to inhibit its polyubiquitination and degradation, and recruits coactivators such as SRC1 and GRIP on E2-regulated promoters to enhance gene transcription linked to cellular proliferation, migration, tumorigenicity, and endocrine resistance^[50-54]. Likewise, PIN1 interacts with ER α phosphorylated at S118, inducing its cis/trans isomerization. Moreover, PIN1 blocks the polyubiquitination and degradation of ER α by preventing its interaction with the E6AP E3 ligase, hence enhancing its stability, binding to EREs, and the subsequent transcriptional activity of ER α ^[10,49,55-57]. High levels of PIN1 and ER α , and low levels of E6AP are observed in endocrine resistance^[49].

Other examples are GSK3, LMTK3, and ABL1 kinases that phosphorylate ER α to inhibit its polyubiquitination^[58,59]. First, the glycogen synthase kinase-3 (GSK3) isoforms interact with and phosphorylate ER α at S102, S104, S106, and S118. GSK3 depletion decreases phosphorylation and E2-induced transcriptional activity by increasing polyubiquitination and degradation of this receptor^[59-61]. Thereafter, LMTK3 (lemur tyrosine kinase 3) interacts with and phosphorylates ER α to protect it from

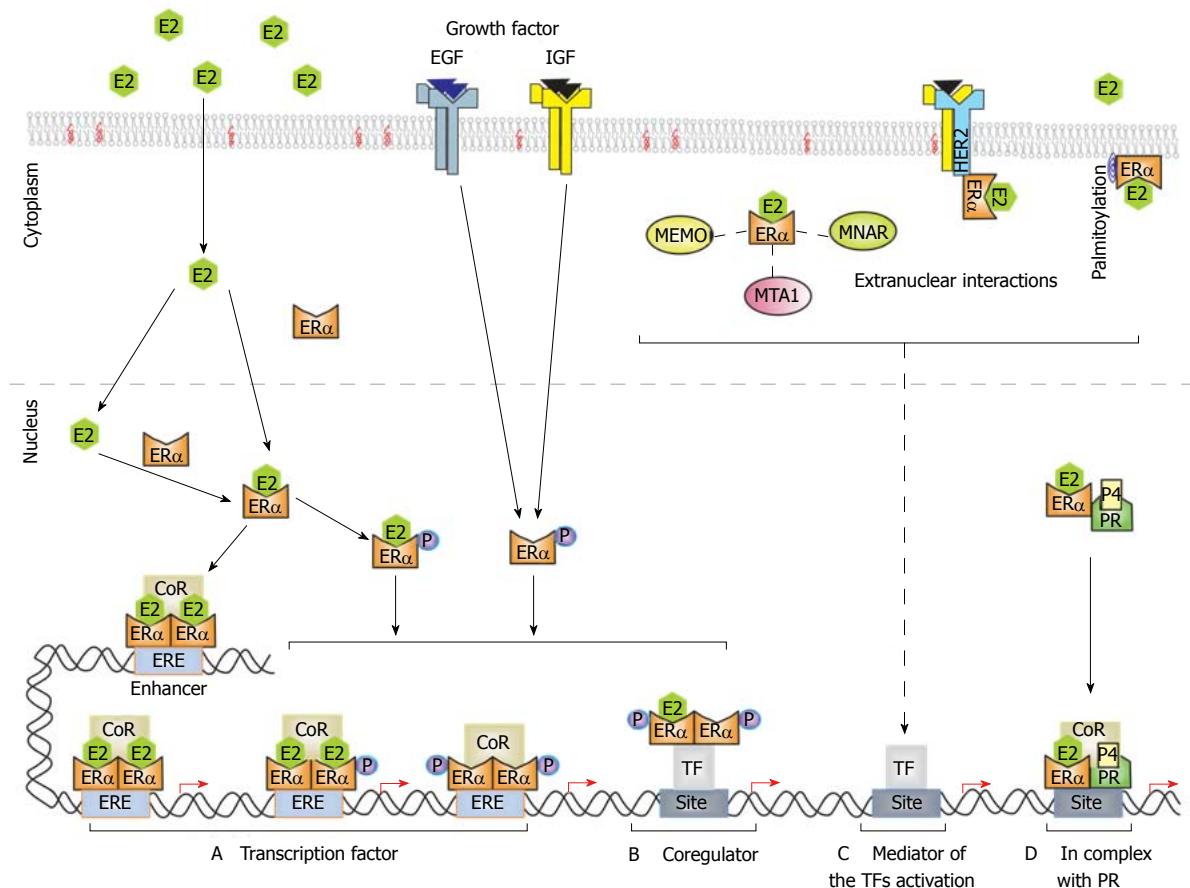


Figure 2 Nuclear and extranuclear signaling of estrogen receptor α . E2 binds to ER α in the cytoplasm and/or nucleus. Then ER α forms homodimers that recognize the ERE sequence (AGGTCAAnnnTGACCT) in target enhancers and promoters, recruiting coregulator (CoR) complexes such as coactivators to induce gene expression. ER α phosphorylation can be induced by E2 to modulate its activity as a transcription regulator. A and B: Growth factors (epidermal growth factor and insulin-like growth factor) also induce ER α phosphorylation in an E2-independent manner to promote ER α activity as a transcription factor or CoR for some transcription factors (*i.e.*, AP-1, Sp1, and NF- κ B); C: Cell membrane-associated ER α (via palmitoylation) associated with transmembrane receptors (*i.e.*, HER2) or with cytoplasmic proteins as (*i.e.*, MEMO, MTA1 and MNAR). These extranuclear interactions can induce kinase-dependent signaling that could finalize in the activation of some transcription factors; D: PR can associate with ER α to coordinate the binding of ER α to the chromatin modulating the expression of specific genes.

polyubiquitination and degradation *via* the UPS in breast cancer cells^[58]. Similarly, ABL (ABL proto-oncogene 1, non-receptor tyrosine kinase) interacts with and phosphorylates ER α at Y52 and Y219, increasing the ER α stability and resistance to Tam; both proteins are increased in breast tumor tissue samples^[62,63].

On the other hand, RB induces the assembly of ER α with chaperone proteins^[64]. Hence, retinoblastoma (RB) interacts with ER α , HSP90, and p23 in the cytoplasm to protect ER α from polyubiquitination and degradation by the UPS. ER α is highly ubiquitinated and degraded in RB-knockdown cells; however, its levels are restored with MG132 (a proteasome inhibitor) treatment in breast cancer^[64].

Interestingly, E3 ubiquitin ligases such as RNF8, RNF31, SHARPIN, and SMURF1 interact with ER α to block its polyubiquitination and to promote the proliferation of breast cancer cells. RNF8, RNF31, and SHARPIN inhibit ER α polyubiquitination by catalyzing monoubiquitination of this receptor, and as a result, ER α protein levels and E2-dependent transcriptional activity are enhanced in

breast cancer cells^[65]. SHARPIN could monoubiquitinate the ER α K302/303, but whether these residues are also modified by RNF8 and/or RNF31 is unclear. Moreover, RNF8 also acts as a coactivator for ER α in breast cancer cells. Instead, SMURF1 apparently inhibits polyubiquitination of ER α , but the implicated mechanisms need to be studied^[65-68].

Other proteins and modifications that inhibit ER α polyubiquitination

ER α polyubiquitination indirect inhibitors (EPII), intriguingly, the inhibition of ER α polyubiquitination also occurs with the help of other proteins that lack the ability to directly interact with ER α . For instance, it has been suggested that Src-dependent phosphorylation of ER α allows E6AP to polyubiquitinate and induce the degradation of this receptor. However, PEBP4 (phosphatidyl ethanolamine-binding protein 4) protein^[69,70] interacts with Src, blocking the phosphorylation and degradation of ER α induced by Src^[69].

Furthermore, although the mechanisms are unclear,

it has been reported that ER α protein levels decrease in cells with low levels of REG γ (PA28 γ , a nuclear proteasome coactivator), but when the proteasome is inhibited by MG132 treatment, ER α protein levels are recovered, suggesting that downregulation of REG γ promotes ER α polyubiquitination and degradation. High levels of REG γ and ER α in breast tumors correlated with poor prognosis in patients with breast cancer^[69].

Additionally, some posttranslational modifications are also associated with ER α polyubiquitination inhibition. Hence, ER α acetylation induced by trichostatin (a deacetylase inhibitor) increases the p300 levels and the stability of the receptor in breast cancer cells, but the mechanisms implicated need to be investigated (Figure 1)^[71]. Palmitoylation has also been linked to ER α polyubiquitination since it has been shown that the ER α mutants that cannot be palmitoylated are polyubiquitinated and degraded *via* UPS^[72].

Mutations and modifications that affect ER α polyubiquitination detected in mammary tumors from patients

ER α polyubiquitination has a clinical relevance, since mutations and/or posttranslational modifications such as phosphorylation in residues of ER α have been identified in tumor tissues from samples of patients with breast cancer, and these residues have been linked to the polyubiquitination and downregulation by degradation of this receptor. Thus, the Y537 residue is required for the ER α phosphorylation, and this modification subsequently promotes polyubiquitination and degradation of the receptor^[35]. However, mutations in the residues Y537N, Y537C, and Y537S are detected in mammary tumors of patients with metastasis and endocrine resistance. Accordingly, ER α polyubiquitination and degradation is prevented by experimentally induced mutations at the Y537 residue, and similarly, these mutations have been associated with the development of endocrine therapy resistance in breast cancer^[15,73,74]. In the same way, the K303 residue is needed for ER α polyubiquitination and degradation, but this residue has been identified to be mutated as K303R in tumors of patients who have poor survival outcome and prognosis^[46,74]. Other residues, such as S104, S106, S118, and S294, that seem to be related with ER α stability, have been found to be phosphorylated in breast tumor samples^[15,73].

ER α POLYUBIQUITINATION INHIBITION IN BREAST CANCER AS A KEY FACTOR FOR THERAPEUTIC STRATEGY

ER α polyubiquitination for its downregulation *via* the UPS, is a central mechanism of some endocrine therapies with SERDs, such as fulvestrant^[46,75]. Clearly, the induction of ER α polyubiquitination for its degradation decreases the abundance and pro-tumor activity of ER α , consequently

novel drugs including AZD9496^[76], GDC-0810^[77], bazedoxifene^[78], and RAD1901^[79] have been synthesized as SERDs, but more studies are required. Despite the importance of SERDs in the therapy of breast cancer, EPIP are promising targets for the management of this disease. Remarkably, the proteins that inhibit the ER α polyubiquitination are enhanced in ER α + breast cancers, contributing to disease progression. For this reason, EPIP may be useful as a biomarker for breast cancer and as a therapeutic target.

PIN1 is overexpressed in breast cancer and is related to mammary tumor growth, and epithelial-mesenchymal transition, and natural and synthetic inhibitors are being probed to control its activity^[55,57,80-87]. Similarly, LMTK3 overexpression stimulates cellular proliferation and tumor formation, and correlates with shorter survival times in ER α + breast cancer, and resistance to Tam treatment, but these events are reduced when LMTK3 expression is decreased^[58,88-90]. Moreover, CG0009, is a GSK3 inhibitor that decreases proliferation of breast cancer cells^[61,73,91-94].

Another molecule is RNF31, whose overexpression increases ER α protein levels, expression of ER α target genes and the growth of breast cancer cells, and these events are decreased when RNF31 is abated^[65]. Lastly, the loss of RB expression seems to be related to the loss of ER α stability in ER α negative (ER α -) breast cancers and with poor responses to hormonal therapies in patients^[64,95-98]. Thus, these proteins can be potential biomarkers and target for the treatment of ER α + breast cancer.

Among EPIIs, PEBP4 inhibits ER α polyubiquitination and enhances its transcriptional activity in breast cancer cells. Because PEBP4 is overexpressed in breast cancer and competes with ER α for components of the UPS, this protein may be an important target for breast cancer. Additionally, specific posttranslational modifications, such as palmitoylation, acetylation and phosphorylation, as well as, mutations of sites linked to ER α polyubiquitination and degradation, demands more research to find new strategies for detection and treatment of breast cancer.

Muc1 is an EPIP in breast cancer

Mucin 1 (MUC1) is a heterodimeric glycoprotein conformed by MUC1 N-terminal (MUC1-N) and MUC1 C-terminal (MUC1-C) subunits^[52]. MUC1-N is an extracellular glycosylated subunit and MUC1-C is a transmembrane subunit with a cytoplasmic domain that interacts with diverse proteins^[54]. MUC1 is localized on the apical borders in normal mammary epithelium, but under breast cancer conditions, it also localizes to the nucleus. An aberrant expression of MUC1-C is detected in breast cancer cells through a regulation loop that implicates Rab31 protein inhibits the lysosomal degradation of MUC1-C, and *Rab31* gene expression is induced by MUC1-C^[52-54,99]. Furthermore, *MUC1* is upregulated in 90% of breast cancers, where the expression of *Rab31* gene and other genes associated with endocrine resistance are modu-

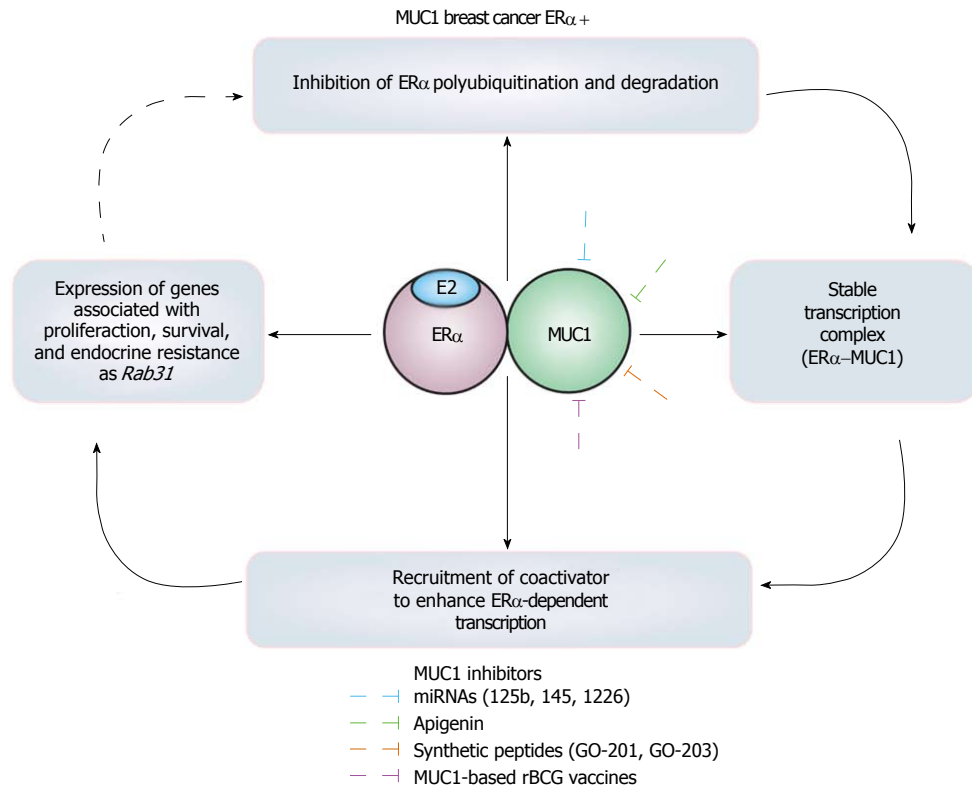


Figure 3 Mucin 1 is an estrogen receptor α polyubiquitination inhibitor protein in breast cancer cells.

lated by the MUC1-C/ER α complex. For these reasons, MUC1 has been suggested as a potential biomarker of breast cancer and predictor of resistance to Tam treatment^[51,100,101] (Figure 3).

Interestingly, MUC1-C subunit interacts with DBD of ER α promoting (1) Inhibition of ER α polyubiquitination maintaining high levels of this receptor; (2) a stable complex between MUC1-C and ER α ; and (3) an enhancement in the pro-tumor transcriptional activity of ER α since SRC1 and GRIP coactivators with histone acetyltransferase activity are recruited by MUC1^[50]. Thus, MUC1-C increases the growth and survival induced by E2 in breast cancer cells, but also transformation, loss of cellular polarity, cellular proliferation and migration, anchorage-independent growth, and tumorigenicity in transgenic mouse models^[51,99,102-104].

Remarkably, MUC1 is an EPIP involved in proliferation and endocrine resistance^[50,53,54,100,105], inhibited by miR-125b^[106], miR-145^[104], miR-1226^[103], and by specific siRNAs, inducing apoptosis, reducing cell proliferation, and increasing sensitivity to Tam^[100]. Similarly, apigenin^[107], and the synthetic peptides GO-201^[54] and GO-203^[100], affect localization and dimerization of MUC1, and as a result, tumor development is decreased, and sensitivity to Tam is increased^[54,100,107]. Moreover, MUC1-based rBCG (Bacillus Calmette-Guerin) vaccines induce anti-MUC immune responses inhibiting the growth of tumors in mice^[108,109]. Interestingly, high levels of Rab31 antigen have been associated with a proliferative status, a high tumor grade, and with poor 5-year disease-free survival

in patients with ER α + breast cancer. Consequently, the Rab31 antigen levels in mammary tumors have been suggested as a biomarker for ER α + breast cancers that may also be useful in the selection of patients for MUC1-targeted therapeutic strategies^[110].

CONCLUSION

Several mechanisms seem to cooperate to inhibit ER α polyubiquitination, decreasing its degradation in ER α + breast cancer cells. These cells become resistant to ER α polyubiquitination due to the evident upregulation of proteins, modifications, and mutations that protect it from ubiquitination. There is no pattern of the characteristics of the inhibitor or protector proteins for ER α polyubiquitination. Some of the reported EPIPs are MUC1, GSK3, LMTK3, RNF8, RNF31, SHARPIN, SMURF1, RB, and PIN1. All of them inhibit ER α polyubiquitination and its degradation in a dissimilar manner, *via* subcellular compartments or mechanisms. Some of them can be grouped as coactivators for ER α (MUC1, PIN1, and RNF8), kinases for ER α (GSK3, LMTK3, and ABL1), E3 ubiquitin ligase (RNF8, RNF31, SHARPIN, and SMURF1), and scaffold protein (RB). Amongst these different mechanisms, the participation of E3-ubiquitin ligases, such as RNF8, RNF31, and SHARPIN, are interesting, since they catalyze ER α monoubiquitination, suggesting a possible competition between monoubiquitination and polyubiquitination of this receptor.

Considering the findings described above, inhibition

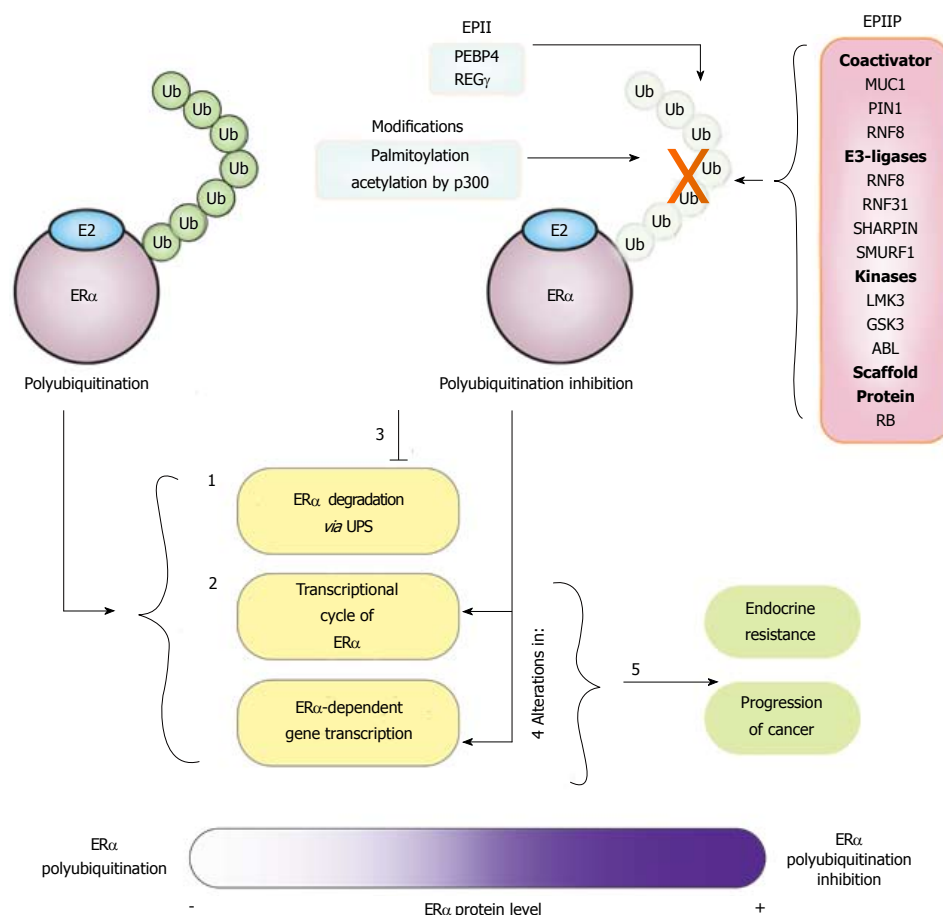


Figure 4 Mechanisms implicated in the estrogen receptor α polyubiquitination inhibition. Half-life of estrogen receptor α protein oscillates between 3-5 h under basal condition. E2 treatment induces ER α polyubiquitination, and as result: (1) Degradation of this receptor is promoted, decreasing its protein levels starting from 1h after treatment; (2) the ER α transcriptional cycle is activated. ER α polyubiquitination inhibitor proteins (EPIP) and ER α polyubiquitination indirect inhibitors (EPII) and other modifications increased in breast cancer cells can inhibit the basal and E2-induced polyubiquitination of ER α ; resulting in (3) the inhibition of its degradation and an enhancement in the ER α protein levels; (4) alterations in the transcription cycle of this receptor and the expression of its targets genes; and (5) these events seem to be associated with endocrine resistance and progression of breast cancer.

of ER α polyubiquitination, increases its abundance, and the expression of E2-dependent genes linked to proliferation and tumor development. In addition, inhibition of ER α polyubiquitination may have other serious implications, since it has been reported that this modification and proteasome activity are coupled to the transcriptional cycle of this receptor^[45]. Moreover, it has been proposed that high ER α protein levels are related to ER α binding to other DNA regulatory regions of genes that are atypically activated under this condition^[111]. Thus, inhibition of ER α polyubiquitination and its degradation increases the stability of this receptor, but also affects ER α /E2 signaling and its transcriptional activity, involved with the development of tumor and endocrine resistance^[111,112] (Figure 4).

Importantly, there is an interplay between inhibition of ER α polyubiquitination and endocrine therapy resistance in ER α + breast cancer, promoted by EPIP and EPII^[49,50,58,65]. In contrast, in luminal B breast cancers or ER α - breast cancers, RB is commonly lost or dysfunctional, leading to high levels of polyubiquitination and degradation of ER α , with a poor prognosis

for patients. Therefore, EPIP, EPII, and mutations and modifications that inhibit ER α polyubiquitination and degradation may act in a cooperative manner to enhance the stability of the receptor in the progression of breast cancer. Consequently, the mechanisms involved in the inhibition of ER α polyubiquitination represent useful biomarkers, therapeutic targets, and prognostic indicators of endocrine therapy in breast cancer.

In conclusion, EPIP, EPII, and mutations and modifications associated to ER α polyubiquitination inhibition, enhance the signaling pathways of this receptor. These findings represent a new field in breast cancer, for the establishment of potential biomarkers, as well as, in the design of effective therapeutic targets to control the progression of this disease. Integration between the molecular basis of ER α inhibition and its correlation with the progression of breast tumors remains to be elicited.

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REFERENCES

- 1 **Germain P**, Staels B, Dacquet C, Spedding M, Laudet V. Overview of nomenclature of nuclear receptors. *Pharmacol Rev* 2006; **58**: 685-704 [PMID: 17132848 DOI: 10.1124/pr.58.4.2]
- 2 **Kumar R**, Zakharov MN, Khan SH, Miki R, Jang H, Toraldo G, Singh R, Bhasin S, Jasuja R. The dynamic structure of the estrogen receptor. *J Amino Acids* 2011; **2011**: 812540 [PMID: 22312471 DOI: 10.4061/2011/812540]
- 3 **Ng HW**, Perkins R, Tong W, Hong H. Versatility or promiscuity: the estrogen receptors, control of ligand selectivity and an update on subtype selective ligands. *Int J Environ Res Public Health* 2014; **11**: 8709-8742 [PMID: 25162709 DOI: 10.3390/ijerph110908709]
- 4 **Manavathi B**, Dey O, Gajulapalli VN, Bhatia RS, Bugide S, Kumar R. Derailed estrogen signaling and breast cancer: an authentic couple. *Endocr Rev* 2013; **34**: 1-32 [PMID: 22947396 DOI: 10.1210/er.2011-1057]
- 5 **Vrtačnik P**, Ostanek B, Mencej-Bedrač S, Marc J. The many faces of estrogen signaling. *Biochem Med (Zagreb)* 2014; **24**: 329-342 [PMID: 25351351 DOI: 10.11613/BM.2014.035]
- 6 **Hah N**, Kraus WL. Hormone-regulated transcriptomes: lessons learned from estrogen signaling pathways in breast cancer cells. *Mol Cell Endocrinol* 2014; **382**: 652-664 [PMID: 23810978 DOI: 10.1016/j.mce.2013.06.021]
- 7 **Hah N**, Murakami S, Nagari A, Danko CG, Kraus WL. Enhancer transcripts mark active estrogen receptor binding sites. *Genome Res* 2013; **23**: 1210-1223 [PMID: 23636943 DOI: 10.1101/gr.152306.112]
- 8 **Manavathi B**, Samanthapudi VS, Gajulapalli VN. Estrogen receptor coregulators and pioneer factors: the orchestrators of mammary gland cell fate and development. *Front Cell Dev Biol* 2014; **2**: 34 [PMID: 25364741 DOI: 10.3389/fcell.2014.00034]
- 9 **Hervouet E**, Cartron PF, Jouvenot M, Delage-Mourroux R. Epigenetic regulation of estrogen signaling in breast cancer. *Epigenetics* 2013; **8**: 237-245 [PMID: 23364277 DOI: 10.4161/epi.23790]
- 10 **Rajbhandari P**, Finn G, Solodin NM, Singarapu KK, Sahu SC, Markley JL, Kadunc KJ, Ellison-Zelski SJ, Kariagina A, Haslam SZ, Lu KP, Alarid ET. Regulation of estrogen receptor α N-terminus conformation and function by peptidyl prolyl isomerase Pin1. *Mol Cell Biol* 2012; **32**: 445-457 [PMID: 22064478 DOI: 10.1128/MCB.06073-11]
- 11 **He C**, Wang X, Zhang MQ. Nucleosome eviction and multiple co-factor binding predict estrogen-receptor- α -associated long-range interactions. *Nucleic Acids Res* 2014; **42**: 6935-6944 [PMID: 24782518 DOI: 10.1093/nar/gku327]
- 12 **Liu MH**, Cheung E. Estrogen receptor-mediated long-range chromatin interactions and transcription in breast cancer. *Mol Cell Endocrinol* 2014; **382**: 624-632 [PMID: 24071518 DOI: 10.1016/j.mce.2013.09.019]
- 13 **Acconcia F**, Kumar R. Signaling regulation of genomic and nongenomic functions of estrogen receptors. *Cancer Lett* 2006; **238**: 1-14 [PMID: 16084012 DOI: 10.1016/j.canlet.2005.06.018]
- 14 **Kerdivel G**, Flouriot G, Pakdel F. Modulation of estrogen receptor alpha activity and expression during breast cancer progression. *Vitam Horm* 2013; **93**: 135-160 [PMID: 23810005 DOI: 10.1016/B978-0-12-416673-8.00004-6]
- 15 **Le Romancer M**, Poulard C, Cohen P, Sentis S, Renoir JM, Corbo L. Cracking the estrogen receptor's posttranslational code in breast tumors. *Endocr Rev* 2011; **32**: 597-622 [PMID: 21680538 DOI: 10.1210/er.2010-0016]
- 16 **Soltysik K**, Czekaj P. Membrane estrogen receptors - is it an alternative way of estrogen action? *J Physiol Pharmacol* 2013; **64**: 129-142 [PMID: 23756388]
- 17 **Zhou W**, Srinivasan S, Nawaz Z, Slingerland JM. ER α , SKP2 and E2F-1 form a feed forward loop driving late ER α targets and G1 cell cycle progression. *Oncogene* 2014; **33**: 2341-2353 [PMID: 23770852 DOI: 10.1038/ncr.2013.197]
- 18 **Treviño LS**, Weigel NL. Phosphorylation: a fundamental regulator of steroid receptor action. *Trends Endocrinol Metab* 2013; **24**: 515-524 [PMID: 23838532 DOI: 10.1016/j.tem.2013.05.008]
- 19 **Tecalco-Cruz AC**, Pérez-Alvarado IA, Ramírez-Jarquín JO, Rocha-Zavaleta L. Nucleo-cytoplasmic transport of estrogen receptor alpha in breast cancer cells. *Cell Signal* 2017; **34**: 121-132 [PMID: 28341599 DOI: 10.1016/j.cellsig.2017.03.011]
- 20 **Tecalco-Cruz AC**, Ramírez-Jarquín JO. Mechanisms that Increase Stability of Estrogen Receptor Alpha in Breast Cancer. *Clin Breast Cancer* 2017; **17**: 1-10 [PMID: 27561704 DOI: 10.1016/j.clbc.2016.07.015]
- 21 **Mohammed H**, Russell IA, Stark R, Rueda OM, Hickey TE, Tarulli GA, Serandour AA, Birrell SN, Bruna A, Saadi A, Menon S, Hadfield J, Pugh M, Raj GV, Brown GD, D'Santos C, Robinson JL, Silva G, Launchbury R, Perou CM, Stingl J, Caldas C, Tilley WD, Carroll JS. Corrigendum: Progesterone receptor modulates ER α action in breast cancer. *Nature* 2015; **526**: 144 [PMID: 26245370 DOI: 10.1038/nature14959]
- 22 **Pedram A**, Razandi M, Deschenes RJ, Levin ER. DHHC-7 and -21 are palmitoyltransferases for sex steroid receptors. *Mol Biol Cell* 2012; **23**: 188-199 [PMID: 22031296 DOI: 10.1091/mbc.E11-07-0638]
- 23 **Acconcia F**, Marino M. The Effects of 17 β -estradiol in Cancer are Mediated by Estrogen Receptor Signaling at the Plasma Membrane. *Front Physiol* 2011; **2**: 30 [PMID: 21747767 DOI: 10.3389/fphys.2011.00030]
- 24 **Marino M**, Galluzzo P, Ascenzi P. Estrogen signaling multiple pathways to impact gene transcription. *Curr Genomics* 2006; **7**: 497-508 [PMID: 18369406 DOI: 10.2174/138920206779315737]
- 25 **Miyoshi Y**, Murase K, Saito M, Imamura M, Oh K. Mechanisms of estrogen receptor- α upregulation in breast cancers. *Med Mol Morphol* 2010; **43**: 193-196 [PMID: 21267694 DOI: 10.1007/s00795-010-0514-3]
- 26 **Cook KL**, Shajahan AN, Clarke R. Autophagy and endocrine resistance in breast cancer. *Expert Rev Anticancer Ther* 2011; **11**: 1283-1294 [PMID: 21916582 DOI: 10.1586/era.11.111]
- 27 **Magnani L**, Brunelle M, Gévy N, Lupien M. Chromatin landscape and endocrine response in breast cancer. *Epigenomics* 2012; **4**: 675-683 [PMID: 23244312 DOI: 10.2217/epi.12.64]
- 28 **de Leeuw R**, Neeffes J, Michalides R. A role for estrogen receptor phosphorylation in the resistance to tamoxifen. *Int J Breast Cancer* 2011; **2011**: 232435 [PMID: 22295213 DOI: 10.4061/2011/232435]
- 29 **Johnson AB**, O'Malley BW. ERasing breast cancer resistance through the kinase. *Nat Med* 2011; **17**: 660-661 [PMID: 21647142 DOI: 10.1038/nm0611-660]
- 30 **Muluhngwi P**, Klinge CM. Roles for miRNAs in endocrine resistance in breast cancer. *Endocr Relat Cancer* 2015; **22**: R279-R300 [PMID: 26346768 DOI: 10.1530/ERC-15-0355]
- 31 **Osborne CK**, Schiff R. Mechanisms of endocrine resistance in breast cancer. *Annu Rev Med* 2011; **62**: 233-247 [PMID: 20887199 DOI: 10.1146/annurev-med-070909-182917]
- 32 **Helzer KT**, Hooper C, Miyamoto S, Alarid ET. Ubiquitylation of nuclear receptors: new linkages and therapeutic implications. *J Mol Endocrinol* 2015; **54**: R151-R167 [PMID: 25943391 DOI: 10.1530/JME-14-0308]
- 33 **Zhou W**, Slingerland JM. Links between oestrogen receptor activation and proteolysis: relevance to hormone-regulated cancer therapy. *Nat Rev Cancer* 2014; **14**: 26-38 [PMID: 24505618 DOI: 10.1038/nrc3622]
- 34 **Fan M**, Park A, Nephew KP. CHIP (carboxyl terminus of Hsc70-interacting protein) promotes basal and geldanamycin-induced degradation of estrogen receptor- α . *Mol Endocrinol* 2005; **19**: 2901-2914 [PMID: 16037132 DOI: 10.1210/me.2005-0111]
- 35 **Sun J**, Zhou W, Kaliappan K, Nawaz Z, Slingerland JM. ER α phosphorylation at Y537 by Src triggers E6-AP-ER α binding, ER α ubiquitylation, promoter occupancy, and target gene expression. *Mol Endocrinol* 2012; **26**: 1567-1577 [PMID: 22865929 DOI: 10.1210/me.2012-1140]
- 36 **Eakin CM**, Maccoss MJ, Finney GL, Klevit RE. Estrogen receptor alpha is a putative substrate for the BRCA1 ubiquitin ligase. *Proc Natl Acad Sci USA* 2007; **104**: 5794-5799 [PMID: 17392432 DOI: 10.1073/pnas.0610887104]

- 37 **Hashizume R**, Fukuda M, Maeda I, Nishikawa H, Oyake D, Yabuki Y, Ogata H, Ohta T. The RING heterodimer BRCA1-BARD1 is a ubiquitin ligase inactivated by a breast cancer-derived mutation. *J Biol Chem* 2001; **276**: 14537-14540 [PMID: 11278247 DOI: 10.1074/jbc.C000881200]
- 38 **Bhatt S**, Xiao Z, Meng Z, Katzenellenbogen BS. Phosphorylation by p38 mitogen-activated protein kinase promotes estrogen receptor α turnover and functional activity *via* the SCF(Skp2) proteasomal complex. *Mol Cell Biol* 2012; **32**: 1928-1943 [PMID: 22431515 DOI: 10.1128/MCB.06561-11]
- 39 **Saji S**, Okumura N, Eguchi H, Nakashima S, Suzuki A, Toi M, Nozawa Y, Saji S, Hayashi S. MDM2 enhances the function of estrogen receptor alpha in human breast cancer cells. *Biochem Biophys Res Commun* 2001; **281**: 259-265 [PMID: 11178989 DOI: 10.1006/bbrc.2001.4339]
- 40 **Iizuka M**, Susa T, Tamamori-Adachi M, Okinaga H, Okazaki T. Intrinsic ubiquitin E3 ligase activity of histone acetyltransferase Hbo1 for estrogen receptor α . *Proc Jpn Acad Ser B Phys Biol Sci* 2017; **93**: 498-510 [PMID: 28769019 DOI: 10.2183/pjab.93.030]
- 41 **Kocanova S**, Mazaheri M, Caze-Subra S, Bystricky K. Ligands specify estrogen receptor alpha nuclear localization and degradation. *BMC Cell Biol* 2010; **11**: 98 [PMID: 21143970 DOI: 10.1186/1471-2121-11-98]
- 42 **Marsaud V**, Gougelet A, Maillard S, Renoir JM. Various phosphorylation pathways, depending on agonist and antagonist binding to endogenous estrogen receptor alpha (ERalpha), differentially affect ERalpha extractability, proteasome-mediated stability, and transcriptional activity in human breast cancer cells. *Mol Endocrinol* 2003; **17**: 2013-2027 [PMID: 12855746 DOI: 10.1210/me.2002-0269]
- 43 **Valley CC**, Solodin NM, Powers GL, Ellison SJ, Alarid ET. Temporal variation in estrogen receptor-alpha protein turnover in the presence of estrogen. *J Mol Endocrinol* 2008; **40**: 23-34 [PMID: 18096994 DOI: 10.1677/JME-07-0067]
- 44 **Lonard DM**, Nawaz Z, Smith CL, O'Malley BW. The 26S proteasome is required for estrogen receptor-alpha and coactivator turnover and for efficient estrogen receptor-alpha transactivation. *Mol Cell* 2000; **5**: 939-948 [PMID: 10911988 DOI: 10.1016/S1097-2765(00)80259-2]
- 45 **Reid G**, Hübner MR, Métivier R, Brand H, Denger S, Manu D, Beaudouin J, Ellenberg J, Gannon F. Cyclic, proteasome-mediated turnover of unliganded and liganded ERalpha on responsive promoters is an integral feature of estrogen signaling. *Mol Cell* 2003; **11**: 695-707 [PMID: 12667452 DOI: 10.1016/S1097-2765(03)00090-X]
- 46 **Berry NB**, Fan M, Nephew KP. Estrogen receptor-alpha hinge-region lysines 302 and 303 regulate receptor degradation by the proteasome. *Mol Endocrinol* 2008; **22**: 1535-1551 [PMID: 18388150 DOI: 10.1210/me.2007-0449]
- 47 **Wang Y**, Zong H, Chi Y, Hong Y, Yang Y, Zou W, Yun X, Gu J. Repression of estrogen receptor alpha by CDK1p58 through promoting its ubiquitin-proteasome degradation. *J Biochem* 2009; **145**: 331-343 [PMID: 19122208 DOI: 10.1093/jb/mvn177]
- 48 **Henrich LM**, Smith JA, Kitt D, Errington TM, Nguyen B, Traish AM, Lannigan DA. Extracellular signal-regulated kinase 7, a regulator of hormone-dependent estrogen receptor destruction. *Mol Cell Biol* 2003; **23**: 5979-5988 [PMID: 12917323 DOI: 10.1128/MCB.23.17.5979-5988.2003]
- 49 **Rajbhandari P**, Schalper KA, Solodin NM, Ellison-Zelski SJ, Ping Lu K, Rimm DL, Alarid ET. Pin1 modulates ER α levels in breast cancer through inhibition of phosphorylation-dependent ubiquitination and degradation. *Oncogene* 2014; **33**: 1438-1447 [PMID: 23542176 DOI: 10.1038/ncr.2013.78]
- 50 **Wei X**, Xu H, Kufe D. MUC1 oncoprotein stabilizes and activates estrogen receptor alpha. *Mol Cell* 2006; **21**: 295-305 [PMID: 16427018 DOI: 10.1016/j.molcel.2005.11.030]
- 51 **Ghosh SK**, Uchida M, Yoo B, Ross AW, Gendler SJ, Gong J, Moore A, Medarova Z. Targeted imaging of breast tumor progression and therapeutic response in a human uMUC-1 expressing transgenic mouse model. *Int J Cancer* 2013; **132**: 1860-1867 [PMID: 23015160 DOI: 10.1002/ijc.27872]
- 52 **Haddon L**, Hugh J. MUC1-mediated motility in breast cancer: a review highlighting the role of the MUC1/ICAM-1/Src signaling triad. *Clin Exp Metastasis* 2015; **32**: 393-403 [PMID: 25759211 DOI: 10.1007/s10585-015-9711-8]
- 53 **Jin C**, Rajabi H, Pitroda S, Li A, Kharbanda A, Weichselbaum R, Kufe D. Cooperative interaction between the MUC1-C oncoprotein and the Rab31 GTPase in estrogen receptor-positive breast cancer cells. *PLoS One* 2012; **7**: e39432 [PMID: 22792175 DOI: 10.1371/journal.pone.0039432]
- 54 **Raina D**, Ahmad R, Joshi MD, Yin L, Wu Z, Kawano T, Vasir B, Avigan D, Kharbanda S, Kufe D. Direct targeting of the mucin 1 oncoprotein blocks survival and tumorigenicity of human breast carcinoma cells. *Cancer Res* 2009; **69**: 5133-5141 [PMID: 19491255 DOI: 10.1158/0008-5472.CAN-09-0854]
- 55 **Lu Z**, Hunter T. Prolyl isomerase Pin1 in cancer. *Cell Res* 2014; **24**: 1033-1049 [PMID: 25124924 DOI: 10.1038/cr.2014.109]
- 56 **Rajbhandari P**, Ozers MS, Solodin NM, Warren CL, Alarid ET. Peptidylprolyl Isomerase Pin1 Directly Enhances the DNA Binding Functions of Estrogen Receptor α . *J Biol Chem* 2015; **290**: 13749-13762 [PMID: 25866209 DOI: 10.1074/jbc.M114.621698]
- 57 **Wang JZ**, Liu BG, Zhang Y. Pin1-based diagnostic and therapeutic strategies for breast cancer. *Pharmacol Res* 2015; **93**: 28-35 [PMID: 25553719 DOI: 10.1016/j.phrs.2014.12.005]
- 58 **Giamas G**, Filipović A, Jacob J, Messier W, Zhang H, Yang D, Zhang W, Shifa BA, Photiou A, Tralau-Stewart C, Castellano L, Green AR, Coombes RC, Ellis IO, Ali S, Lenz HJ, Stebbing J. Kinome screening for regulators of the estrogen receptor identifies LMTK3 as a new therapeutic target in breast cancer. *Nat Med* 2011; **17**: 715-719 [PMID: 21602804 DOI: 10.1038/nm.2351]
- 59 **Grisouard J**, Medunjanin S, Hermani A, Shukla A, Mayer D. Glycogen synthase kinase-3 protects estrogen receptor alpha from proteasomal degradation and is required for full transcriptional activity of the receptor. *Mol Endocrinol* 2007; **21**: 2427-2439 [PMID: 17609434 DOI: 10.1210/me.2007-0129]
- 60 **Medina M**, Wadosell F. Deconstructing GSK-3: The Fine Regulation of Its Activity. *Int J Alzheimers Dis* 2011; **2011**: 479249 [PMID: 21629747 DOI: 10.4061/2011/479249]
- 61 **Medunjanin S**, Hermani A, De Servi B, Grisouard J, Rincke G, Mayer D. Glycogen synthase kinase-3 interacts with and phosphorylates estrogen receptor alpha and is involved in the regulation of receptor activity. *J Biol Chem* 2005; **280**: 33006-33014 [PMID: 16076840 DOI: 10.1074/jbc.M506758200]
- 62 **Dennis AP**, Haq RU, Nawaz Z. Importance of the regulation of nuclear receptor degradation. *Front Biosci* 2001; **6**: D954-D959 [PMID: 11487464 DOI: 10.2741/Dennis]
- 63 **He X**, Zheng Z, Song T, Wei C, Ma H, Ma Q, Zhang Y, Xu Y, Shi W, Ye Q, Zhong H. c-Abl regulates estrogen receptor alpha transcription activity through its stabilization by phosphorylation. *Oncogene* 2010; **29**: 2238-2251 [PMID: 20101225 DOI: 10.1038/ncr.2009.513]
- 64 **Caligiuri I**, Toffoli G, Giordano A, Rizzolio F. pRb controls estrogen receptor alpha protein stability and activity. *Oncotarget* 2013; **4**: 875-883 [PMID: 23900261 DOI: 10.18632/oncotarget.1036]
- 65 **Zhu J**, Zhao C, Kharman-Biz A, Zhuang T, Jonsson P, Liang N, Williams C, Lin CY, Qiao Y, Zendejdel K, Strömblad S, Treuter E, Dahlman-Wright K. The atypical ubiquitin ligase RNF31 stabilizes estrogen receptor α and modulates estrogen-stimulated breast cancer cell proliferation. *Oncogene* 2014; **33**: 4340-4351 [PMID: 24441041 DOI: 10.1038/ncr.2013.573]
- 66 **Yang H**, Yu N, Xu J, Ding X, Deng W, Wu G, Li X, Hou Y, Liu Z, Zhao Y, Xue M, Yu S, Wang B, Li X, Niu G, Wang H, Zhu J, Zhuang T. SMURF1 facilitates estrogen receptor α signaling in breast cancer cells. *J Exp Clin Cancer Res* 2018; **37**: 24 [PMID: 29433542 DOI: 10.1186/s13046-018-0672-z]
- 67 **Zhuang T**, Yu S, Zhang L, Yang H, Li X, Hou Y, Liu Z, Shi Y, Wang W, Yu N, Li A, Li X, Li X, Niu G, Xu J, Hasni MS, Mu K, Wang H, Zhu J. SHARPIN stabilizes estrogen receptor α and promotes breast cancer cell proliferation. *Oncotarget* 2017; **8**: 77137-77151 [PMID: 29100376 DOI: 10.18632/oncotarget.20368]
- 68 **Wang S**, Luo H, Wang C, Sun H, Sun G, Sun N, Zeng K, Song

- H, Zou R, Zhou T, Cong R, Liu W, Yang L, Li D, Zhou X, Zhong X, Lin L, Jiao J, Yan G, Wang X, Min X, Cao L, Zhao Y. RNF8 identified as a co-activator of estrogen receptor α promotes cell growth in breast cancer. *Biochim Biophys Acta* 2017; **1863**: 1615-1628 [PMID: 28216286 DOI: 10.1016/j.bbdis.2017.02.011]
- 69 **Chai F**, Liang Y, Bi J, Chen L, Zhang F, Cui Y, Jiang J. REG γ regulates ER α degradation via ubiquitin-proteasome pathway in breast cancer. *Biochem Biophys Res Commun* 2015; **456**: 534-540 [PMID: 25490392 DOI: 10.1016/j.bbrc.2014.11.124]
- 70 **Liu H**, Qiu J, Li N, Chen T, Cao X. Human phosphatidylethanolamine-binding protein 4 promotes transactivation of estrogen receptor α (ER α) in human cancer cells by inhibiting proteasome-dependent ER α degradation via association with Src. *J Biol Chem* 2010; **285**: 21934-21942 [PMID: 20460377 DOI: 10.1074/jbc.M110.109876]
- 71 **Kim SH**, Kang HJ, Na H, Lee MO. Trichostatin A enhances acetylation as well as protein stability of ER α through induction of p300 protein. *Breast Cancer Res* 2010; **12**: R22 [PMID: 20388208 DOI: 10.1186/bcr2562]
- 72 **La Rosa P**, Pesiri V, Leclercq G, Marino M, Acconcia F. Palmitoylation regulates 17 β -estradiol-induced estrogen receptor- α degradation and transcriptional activity. *Mol Endocrinol* 2012; **26**: 762-774 [PMID: 22446104 DOI: 10.1210/me.2011-1208]
- 73 **Murphy LC**, Seekallu SV, Watson PH. Clinical significance of estrogen receptor phosphorylation. *Endocr Relat Cancer* 2011; **18**: R1-14 [PMID: 21149515 DOI: 10.1677/ERC-10-0070]
- 74 **Thomas C**, Gustafsson JÅ. Estrogen receptor mutations and functional consequences for breast cancer. *Trends Endocrinol Metab* 2015; **26**: 467-476 [PMID: 26183887 DOI: 10.1016/j.tem.2015.06.007]
- 75 **Yeh WL**, Shioda K, Coser KR, Rivizzigno D, McSweeney KR, Shioda T. Fulvestrant-induced cell death and proteasomal degradation of estrogen receptor α protein in MCF-7 cells require the CSK c-Src tyrosine kinase. *PLoS One* 2013; **8**: e60889 [PMID: 23593342 DOI: 10.1371/journal.pone.0060889]
- 76 **De Savi C**, Bradbury RH, Rabow AA, Norman RA, Buttar D, Currie GS, Weir H, Donald C, Andrews D, MacFaul P, Ballard P, Curwen J, Wilson Z, Richmond G, D'Cruz C, Powell S, Walker G, Hulse M, Tonge M. Abstract 3650: Discovery of the clinical candidate AZD9496: a potent and orally bioavailable selective estrogen receptor downregulator and antagonist. *Cancer Research* 2015; **75**: 3650 [DOI: 10.1158/1538-7445.AM2015-3650]
- 77 **Lai A**, Kahraman M, Govek S, Nagasawa J, Bonnefous C, Julien J, Douglas K, Sensintaffar J, Lu N, Lee KJ, Aparicio A, Kaufman J, Qian J, Shao G, Prudente R, Moon MJ, Joseph JD, Darimont B, Brigham D, Grillot K, Heyman R, Rix PJ, Hager JH, Smith ND. Identification of GDC-0810 (ARN-810), an Orally Bioavailable Selective Estrogen Receptor Degradator (SERD) that Demonstrates Robust Activity in Tamoxifen-Resistant Breast Cancer Xenografts. *J Med Chem* 2015; **58**: 4888-4904 [PMID: 25879485 DOI: 10.1021/acs.jmedchem.5b00054]
- 78 **Wardell SE**, Nelson ER, Chao CA, McDonnell DP. Bazedoxifene exhibits antiestrogenic activity in animal models of tamoxifen-resistant breast cancer: implications for treatment of advanced disease. *Clin Cancer Res* 2013; **19**: 2420-2431 [PMID: 23536434 DOI: 10.1158/1078-0432.CCR-12-3771]
- 79 **Garner F**, Shomali M, Paquin D, Lyttle CR, Hattersley G. RAD1901: a novel, orally bioavailable selective estrogen receptor degrader that demonstrates antitumor activity in breast cancer xenograft models. *Anticancer Drugs* 2015; **26**: 948-956 [PMID: 26164151 DOI: 10.1097/CAD.0000000000000271]
- 80 **Kim JA**, Kim MR, Kim O, Phuong NT, Yun J, Oh WK, Bae K, Kang KW. Amurensin G inhibits angiogenesis and tumor growth of tamoxifen-resistant breast cancer via Pin1 inhibition. *Food Chem Toxicol* 2012; **50**: 3625-3634 [PMID: 22842120 DOI: 10.1016/j.fct.2012.07.027]
- 81 **Kim JH**, Jung JH, Kim SH, Jeong SJ. Decursin exerts anti-cancer activity in MDA-MB-231 breast cancer cells via inhibition of the Pin1 activity and enhancement of the Pin1/p53 association. *Phytother Res* 2014; **28**: 238-244 [PMID: 23580332 DOI: 10.1002/ptr.4986]
- 82 **Kotiyal S**, Bhattacharya S. Breast cancer stem cells, EMT and therapeutic targets. *Biochem Biophys Res Commun* 2014; **453**: 112-116 [PMID: 25261721 DOI: 10.1016/j.bbrc.2014.09.069]
- 83 **Li X**, Li L, Zhou Q, Zhang N, Zhang S, Zhao R, Liu D, Jing Y, Zhao L. Synthesis of the novel elemenic acid derivatives as Pin1 inhibitors. *Bioorg Med Chem Lett* 2014; **24**: 5612-5615 [PMID: 25466185 DOI: 10.1016/j.bmcl.2014.10.087]
- 84 **Moore JD**, Potter A. Pin1 inhibitors: Pitfalls, progress and cellular pharmacology. *Bioorg Med Chem Lett* 2013; **23**: 4283-4291 [PMID: 23796453 DOI: 10.1016/j.bmcl.2013.05.088]
- 85 **Potter A**, Oldfield V, Nunns C, Fromont C, Ray S, Northfield CJ, Bryant CJ, Scrase SF, Robinson D, Matossova N, Baker L, Dokurno P, Surgenor AE, Davis B, Richardson CM, Murray JB, Moore JD. Discovery of cell-active phenyl-imidazole Pin1 inhibitors by structure-guided fragment evolution. *Bioorg Med Chem Lett* 2010; **20**: 6483-6488 [PMID: 20932746 DOI: 10.1016/j.bmcl.2010.09.063]
- 86 **Wei S**, Kozono S, Kats L, Nechama M, Li W, Guarnerio J, Luo M, You MH, Yao Y, Kondo A, Hu H, Bozkurt G, Moerke NJ, Cao S, Reschke M, Chen CH, Rego EM, Lo-Coco F, Cantley LC, Lee TH, Wu H, Zhang Y, Pandolfi PP, Zhou XZ, Lu KP. Active Pin1 is a key target of all-trans retinoic acid in acute promyelocytic leukemia and breast cancer. *Nat Med* 2015; **21**: 457-466 [PMID: 25849135 DOI: 10.1038/nm.3839]
- 87 **Xu GG**, Sledobnick C, Etzkorn FA. Cyclohexyl ketone inhibitors of Pin1 dock in a trans-diaxial cyclohexane conformation. *PLoS One* 2012; **7**: e44226 [PMID: 23028504 DOI: 10.1371/journal.pone.0044226]
- 88 **Stebbing J**, Filipovic A, Ellis IO, Green AR, D'Silva TR, Lenz HJ, Coombes RC, Wang T, Lee SC, Giamas G. LMTK3 expression in breast cancer: association with tumor phenotype and clinical outcome. *Breast Cancer Res Treat* 2012; **132**: 537-544 [PMID: 21671015 DOI: 10.1007/s10549-011-1622-z]
- 89 **Stebbing J**, Filipovic A, Lit LC, Blighe K, Grothey A, Xu Y, Miki Y, Chow LW, Coombes RC, Sasano H, Shaw JA, Giamas G. LMTK3 is implicated in endocrine resistance via multiple signaling pathways. *Oncogene* 2013; **32**: 3371-3380 [PMID: 22869149 DOI: 10.1038/onc.2012.343]
- 90 **Zhao G**, Guo J, Li D, Jia C, Yin W, Sun R, Lv Z, Cong X. MicroRNA-34a suppresses cell proliferation by targeting LMTK3 in human breast cancer mcf-7 cell line. *DNA Cell Biol* 2013; **32**: 699-707 [PMID: 24050776 DOI: 10.1089/dna.2013.2130]
- 91 **Jacobs KM**, Bhawe SR, Ferraro DJ, Jaboin JJ, Hallahan DE, Thotala D. GSK-3 β : A Bifunctional Role in Cell Death Pathways. *Int J Cell Biol* 2012; **2012**: 930710 [PMID: 22675363 DOI: 10.1155/2012/930710]
- 92 **Kim HM**, Kim CS, Lee JH, Jang SJ, Hwang JJ, Ro S, Choi J. CG0009, a novel glycogen synthase kinase 3 inhibitor, induces cell death through cyclin D1 depletion in breast cancer cells. *PLoS One* 2013; **8**: e60383 [PMID: 23565238 DOI: 10.1371/journal.pone.0060383]
- 93 **McCubrey JA**, Davis NM, Abrams SL, Montalto G, Cervello M, Basecke J, Libra M, Nicoletti F, Cocco L, Martelli AM, Steelman LS. Diverse roles of GSK-3: tumor promoter-tumor suppressor, target in cancer therapy. *Adv Biol Regul* 2014; **54**: 176-196 [PMID: 24169510 DOI: 10.1016/j.jbior.2013.09.013]
- 94 **Mishra R**. Glycogen synthase kinase 3 beta: can it be a target for oral cancer. *Mol Cancer* 2010; **9**: 144 [PMID: 20537194 DOI: 10.1186/1476-4598-9-144]
- 95 **Ertel A**, Dean JL, Rui H, Liu C, Witkiewicz AK, Knudsen KE, Knudsen ES. RB-pathway disruption in breast cancer: differential association with disease subtypes, disease-specific prognosis and therapeutic response. *Cell Cycle* 2010; **9**: 4153-4163 [PMID: 20948315 DOI: 10.4161/cc.9.20.13454]
- 96 **Lehn S**, Fernö M, Jirstrom K, Rydén L, Landberg G. A non-functional retinoblastoma tumor suppressor (RB) pathway in premenopausal breast cancer is associated with resistance to tamoxifen. *Cell Cycle* 2011; **10**: 956-962 [PMID: 21358261 DOI: 10.4161/cc.10.6.15074]
- 97 **Treré D**, Brighenti E, Donati G, Ceccarelli C, Santini D, Taffurelli M, Montanaro L, Derenzini M. High prevalence of retinoblastoma

- protein loss in triple-negative breast cancers and its association with a good prognosis in patients treated with adjuvant chemotherapy. *Ann Oncol* 2009; **20**: 1818-1823 [PMID: 19556322 DOI: 10.1093/annonc/mdp209]
- 98 **Witkiewicz AK**, Knudsen ES. Retinoblastoma tumor suppressor pathway in breast cancer: prognosis, precision medicine, and therapeutic interventions. *Breast Cancer Res* 2014; **16**: 207 [PMID: 25223380 DOI: 10.1186/bcr3652]
 - 99 **Alam M**, Rajabi H, Ahmad R, Jin C, Kufe D. Targeting the MUC1-C oncoprotein inhibits self-renewal capacity of breast cancer cells. *Oncotarget* 2014; **5**: 2622-2634 [PMID: 24770886 DOI: 10.18632/oncotarget.1848]
 - 100 **Kharbanda A**, Rajabi H, Jin C, Raina D, Kufe D. Oncogenic MUC1-C promotes tamoxifen resistance in human breast cancer. *Mol Cancer Res* 2013; **11**: 714-723 [PMID: 23538857 DOI: 10.1158/1541-7786.MCR-12-0668]
 - 101 **Pitroda SP**, Khodarev NN, Beckett MA, Kufe DW, Weichselbaum RR. MUC1-induced alterations in a lipid metabolic gene network predict response of human breast cancers to tamoxifen treatment. *Proc Natl Acad Sci USA* 2009; **106**: 5837-5841 [PMID: 19289846 DOI: 10.1073/pnas.0812029106]
 - 102 **Alam M**, Bouillez A, Tagde A, Ahmad R, Rajabi H, Maeda T, Hiraki M, Suzuki Y, Kufe D. MUC1-C Represses the Crumbs Complex Polarity Factor CRB3 and Downregulates the Hippo Pathway. *Mol Cancer Res* 2016; **14**: 1266-1276 [PMID: 27658423 DOI: 10.1158/1541-7786.MCR-16-0233]
 - 103 **Jin C**, Rajabi H, Kufe D. miR-1226 targets expression of the mucin 1 oncoprotein and induces cell death. *Int J Oncol* 2010; **37**: 61-69 [PMID: 20514397]
 - 104 **Sachdeva M**, Mo YY. MicroRNA-145 suppresses cell invasion and metastasis by directly targeting mucin 1. *Cancer Res* 2010; **70**: 378-387 [PMID: 19996288 DOI: 10.1158/0008-5472.CAN-09-2021]
 - 105 **Lacunza E**, Baudis M, Colussi AG, Segal-Eiras A, Croce MV, Abba MC. MUC1 oncogene amplification correlates with protein overexpression in invasive breast carcinoma cells. *Cancer Genet Cytogenet* 2010; **201**: 102-110 [PMID: 20682394 DOI: 10.1016/j.cancergencyto.2010.05.015]
 - 106 **Rajabi H**, Jin C, Ahmad R, McClary C, Joshi MD, Kufe D. MUCIN 1 ONCOPROTEIN EXPRESSION IS SUPPRESSED BY THE miR-125b ONCOMIR. *Genes Cancer* 2010; **1**: 62-68 [PMID: 20729973 DOI: 10.1177/1947601909357933]
 - 107 **Zhou Y**, Rajabi H, Kufe D. Mucin 1 C-terminal subunit oncoprotein is a target for small-molecule inhibitors. *Mol Pharmacol* 2011; **79**: 886-893 [PMID: 21346142 DOI: 10.1124/mol.110.070797]
 - 108 **Yuan S**, Shi C, Ling R, Wang T, Wang H, Han W. Immunization with two recombinant Bacillus Calmette-Guérin vaccines that combine the expression of multiple tandem repeats of mucin-1 and colony stimulating-factor suppress breast tumor growth in mice. *J Cancer Res Clin Oncol* 2010; **136**: 1359-1367 [PMID: 20127358 DOI: 10.1007/s00432-010-0787-x]
 - 109 **Yuan S**, Shi C, Liu L, Han W. MUC1-based recombinant Bacillus Calmette-Guerin vaccines as candidates for breast cancer immunotherapy. *Expert Opin Biol Ther* 2010; **10**: 1037-1048 [PMID: 20420512 DOI: 10.1517/14712598.2010.485185]
 - 110 **Kotzsch M**, Kirchner T, Soelch S, Schäfer S, Friedrich K, Baretton G, Magdolen V, Luther T. Inverse association of rab31 and mucin-1 (CA15-3) antigen levels in estrogen receptor-positive (ER+) breast cancer tissues with clinicopathological parameters and patients' prognosis. *Am J Cancer Res* 2017; **7**: 1959-1970 [PMID: 28979817]
 - 111 **Fowler AM**, Solodin N, Preisler-Mashek MT, Zhang P, Lee AV, Alarid ET. Increases in estrogen receptor-alpha concentration in breast cancer cells promote serine 118/104/106-independent AF-1 transactivation and growth in the absence of estrogen. *FASEB J* 2004; **18**: 81-93 [PMID: 14718389 DOI: 10.1096/fj.03-0038com]
 - 112 **Fowler AM**, Solodin NM, Valley CC, Alarid ET. Altered target gene regulation controlled by estrogen receptor-alpha concentration. *Mol Endocrinol* 2006; **20**: 291-301 [PMID: 16179380 DOI: 10.1210/me.2005-0288]

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Cancer prevention in patients with human immunodeficiency virus infection

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Abstract

Cancer is a leading cause of death in patients with human immunodeficiency virus (HIV) infection. With the advent of antiretroviral treatment, the risk of AIDS-defining cancers declined but the ageing of this population resulted in the emergence of other common cancers, particularly lung and hepatocellular cancer. Accordingly, screening programs similar to the general population should be implemented in patients with HIV infection. Vaccination against common oncogenic viruses is also essential. However, rates of cancer screening and vaccination against HPV and HBV are considerably low in this population, highlighting a pressing need to educate patients and healthcare professionals about the importance of cancer preventive measures in these vulnerable patients.

Key words: Antiretroviral treatment; Prevention; Human immunodeficiency virus infection; Cancer; Vaccination

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Core tip: Cancer is a leading cause of death in patients with human immunodeficiency virus (HIV) infection. With the advent of antiretroviral treatment, the risk of AIDS-defining cancers declined but the ageing of this population resulted in the emergence of other common cancers, particularly lung and hepatocellular cancer. Accordingly, screening programs similar to the general population should be implemented in patients with HIV infection. Vaccination against common oncogenic viruses is also essential.

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Cancer is a leading cause of death in patients with human immunodeficiency virus (HIV) infection^[1,2]. In recent decades, overall mortality declined in this population in industrialized countries but the percentage of deaths due to non-AIDS related cancer increased and currently represent almost one fourth of all deaths^[1,2]. Moreover, HIV infection is associated with increased incidence of several cancers, including Kaposi's sarcoma, certain types of aggressive B-cell lymphomas and invasive cervical cancer, which are classified as AIDS-defining cancers^[3-5]. However, several non-AIDS related cancers, including lung and hepatocellular cancer, are also observed more frequently in patients with HIV infection^[3,6]. Therefore, the prevention of cancer is of paramount importance in this population.

The introduction of antiretroviral treatment (ART) resulted in substantial reductions in the incidence of AIDS-defining cancers^[3-5]. Moreover, immunosuppression also increases the risk of non-AIDS-defining cancers in this population^[7,8]. Therefore, timely implementation of ART is essential for cancer prevention in patients with HIV infection. However, the cost/health benefit ratio of early implementation of ART and persistent suppression of HIV replication should also be considered, particularly in resource-poor settings.

It has been reported that almost 40% of cancers that affect patients with HIV infection are due to oncogenic virus, specifically hepatitis B and C virus infection-related hepatocellular carcinoma (HCC) and human papillomavirus (HPV) infection-related cervical, vulvar, penile, anal, oral and pharyngeal cancer^[8,9]. Accordingly, vaccination against hepatitis B is recommended in all seronegative patients with HIV infection and repeat doses should be administered until anti-HBs titers \geq 10-100 IU/mL are achieved^[10]. Double doses might be indicated in patients with low CD4 count and high HIV viral load^[10]. Vaccination against HPV is also recommended in patients with HIV infection < 26-year-old or < 40-year-old in men who have sex with men (MSM)^[10]. Three doses of the 9-valent HPV vaccine should be used where available^[10].

Lung cancer is more frequent in patients with HIV infection and is a leading cause of death in this population^[3,6]. The higher prevalence of smoking in patients with HIV infection might partly contribute to this association^[11,12]. Therefore, current guidelines state that these patients should be made aware of the detrimental effects of smoking on health and smokers should be informed about the benefits of smoking cessation^[10]. For those willing to quit smoking, pharmacotherapy (including nicotine replacement therapy, varenicline and bupropion), cognitive behavioral counseling and/

or motivational strategies can be employed to help quitting^[10]. On the other hand, computed tomography screening appears to have low yield in patients with HIV infection, probably due to the young age of most of these patients^[13].

Cancer screening recommendations in patients with HIV infection are similar to the general population, since there are no studies that specifically evaluated the benefits and harms of these strategies in this population^[10]. Mammography is recommended every 1-3 years in women 50-70-year-old and measurement of prostate specific antigen is recommended every 2-4 years in men > 50 years with life expectancy > 10 years^[10]. Annual faecal occult blood test, sigmoidoscopy every 5 years or colonoscopy every 10 years are recommended in subjects > 50 years with life expectancy > 10 years^[10]. In patients with cirrhosis and in those with HBV co-infection and either a history of elevated transaminases or risk factors for HCC (family history of HCC, Asians, Africans), abdominal ultrasound and measurement of alpha-fetoprotein levels are recommended every 6 mo to enable the early diagnosis of HCC^[10]. Regarding AIDS-defining cancers, liquid-based cervical cytology test every 1-3 years is recommended in women > 21 years or within 1 year after sexual debut^[10]. Digital rectal examination with or without anal cytology is also recommended in MSM and patients with HPV-associated dysplasia^[10]. Finally, careful inspection of the skin should be performed regularly to detect cancers such as Kaposi's sarcoma, basal cell carcinoma and malignant melanoma^[10].

In conclusion, cancer is a frequent cause of death in patients with HIV infection. With the advent of ART, the risk of AIDS-defining cancers declined but the ageing of this population resulted in the emergence of other common cancers, particularly lung cancer and HCC. Accordingly, screening programs similar to the general population should be implemented in patients with HIV infection. Vaccination against common oncogenic viruses is also essential. In addition, primary prevention of cancer by implementing educational programs stressing the importance of healthy lifestyle are equally important in patients with HIV infection. However, rates of cancer screening and vaccination against HPV and HBV are considerably low in this population^[14,15], highlighting a pressing need to educate patients and healthcare professionals about the importance of cancer preventive measures in these vulnerable patients.

REFERENCES

- 1 Smith CJ, Ryom L, Weber R, Morlat P, Pradier C, Reiss P, Kowalska JD, de Wit S, Law M, el Sadr W, Kirk O, Friis-Moller N, Monforte Ad, Phillips AN, Sabin CA, Lundgren JD; D:A:D Study Group. Trends in underlying causes of death in people with HIV from 1999 to 2011 (D:A:D): a multicohort collaboration. *Lancet* 2014; **384**: 241-248 [PMID: 25042234 DOI: 10.1016/S0140-6736(14)60604-8]
- 2 Morlat P, Roussillon C, Henard S, Salmon D, Bonnet F, Cacoub P, Georget A, Aouba A, Rosenthal E, May T, Chauveau M, Diallo B,

- Costagliola D, Chene G; ANRS EN20 Mortalité 2010 Study Group. Causes of death among HIV-infected patients in France in 2010 (national survey): trends since 2000. *AIDS* 2014; **28**: 1181-1191 [PMID: 24901259 DOI: 10.1097/QAD.0000000000000222]
- 3 **Hernández-Ramírez RU**, Shiels MS, Dubrow R, Engels EA. Cancer risk in HIV-infected people in the USA from 1996 to 2012: a population-based, registry-linkage study. *Lancet HIV* 2017; **4**: e495-e504 [PMID: 28803888 DOI: 10.1016/S2352-3018(17)30125-X]
 - 4 **Engels EA**, Biggar RJ, Hall HI, Cross H, Crutchfield A, Finch JL, Grigg R, Hylton T, Pawlish KS, McNeel TS, Goedert JJ. Cancer risk in people infected with human immunodeficiency virus in the United States. *Int J Cancer* 2008; **123**: 187-194 [PMID: 18435450 DOI: 10.1002/ijc.23487]
 - 5 **Clifford GM**, Polesel J, Rickenbach M, Dal Maso L, Keiser O, Kofler A, Rapiti E, Levi F, Jundt G, Fisch T, Bordoni A, De Weck D, Franceschi S; Swiss HIV Cohort. Cancer risk in the Swiss HIV Cohort Study: associations with immunodeficiency, smoking, and highly active antiretroviral therapy. *J Natl Cancer Inst* 2005; **97**: 425-432 [PMID: 15770006 DOI: 10.1093/jnci/dji072]
 - 6 **Robbins HA**, Pfeiffer RM, Shiels MS, Li J, Hall HI, Engels EA. Excess cancers among HIV-infected people in the United States. *J Natl Cancer Inst* 2015; **107**: pii: dju503 [PMID: 25663691 DOI: 10.1093/jnci/dju503]
 - 7 **Frisch M**, Biggar RJ, Engels EA, Goedert JJ; AIDS-Cancer Match Registry Study Group. Association of cancer with AIDS-related immunosuppression in adults. *JAMA* 2001; **285**: 1736-1745 [PMID: 11277828]
 - 8 **Yarchoan R**, Uldrick TS. HIV-Associated Cancers and Related Diseases. *N Engl J Med* 2018; **378**: 1029-1041 [PMID: 29539283 DOI: 10.1056/NEJMra1615896]
 - 9 **de Martel C**, Shiels MS, Franceschi S, Simard EP, Vignat J, Hall HI, Engels EA, Plummer M. Cancers attributable to infections among adults with HIV in the United States. *AIDS* 2015; **29**: 2173-2181 [PMID: 26182198 DOI: 10.1097/QAD.0000000000000808]
 - 10 **European AIDS Clinical Society**. European AIDS Clinical Society Guidelines v9.0. October 2017. Available from: URL: http://www.eacsociety.org/files/guidelines_9.0-english.pdf
 - 11 **Fontela C**, Castilla J, Juanbeltz R, Martínez-Baz I, Rivero M, O'Leary A, Larrea N, San Miguel R. Comorbidities and cardiovascular risk factors in an aged cohort of HIV-infected patients on antiretroviral treatment in a Spanish hospital in 2016. *Postgrad Med* 2018; **130**: 317-324 [PMID: 29486621 DOI: 10.1080/00325481.2018.1446653]
 - 12 **Estrada V**, Bernardino JI, Masiá M, Iribarren JA, Ortega A, Lozano F, Miralles C, Olalla J, Santos J, Elías MJ, Domingo P, Cruz AF. Cardiovascular risk factors and lifetime risk estimation in HIV-infected patients under antiretroviral treatment in Spain. *HIV Clin Trials* 2015; **16**: 57-65 [PMID: 25874992 DOI: 10.1179/1528433614Z.00000000008]
 - 13 **Hulbert A**, Hooker CM, Keruly JC, Brown T, Horton K, Fishman E, Rodgers K, Lee B, Sam C, Tsai S, Weihe E, Pridham G, Drummond B, Merlo C, Geronimo M, Porter M, Cox S, Li D, Harline M, Teran M, Wrangle J, Mudge B, Taylor G, Kirk GD, Herman JG, Moore RD, Brown RH, Brock MV. Prospective CT screening for lung cancer in a high-risk population: HIV-positive smokers. *J Thorac Oncol* 2014; **9**: 752-759 [PMID: 24828660 DOI: 10.1097/JTO.0000000000000161]
 - 14 **Tron L**, Lert F, Spire B, Dray-Spira R; Agence Nationale de Recherche sur le Sida et les Hépatites Virales (ANRS) - Vespa2 Study Group. Levels and determinants of breast and cervical cancer screening uptake in HIV-infected women compared with the general population in France. *HIV Med* 2017; **18**: 181-195 [PMID: 28967199 DOI: 10.1111/hiv.12412]
 - 15 **Jansen K**, Thamm M, Bock CT, Scheufele R, Kücherer C, Muenstermann D, Hagedorn HJ, Jessen H, Dupke S, Hamouda O, Günsenheimer-Bartmeyer B, Meixnerberger K; HIV Seroconverter Study Group. High Prevalence and High Incidence of Coinfection with Hepatitis B, Hepatitis C, and Syphilis and Low Rate of Effective Vaccination against Hepatitis B in HIV-Positive Men Who Have Sex with Men with Known Date of HIV Seroconversion in Germany. *PLoS One* 2015; **10**: e0142515 [PMID: 26555244 DOI: 10.1371/journal.pone.0142515]

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Oxytocin and cancer: An emerging link

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Abstract

The neuropeptide hormone oxytocin, which is released from the posterior pituitary gland, is involved in a number of physiological processes. Understanding of its effects is gradually increasing due to new research in this area. While mostly recognized as a reproductive system hormone, oxytocin also regulates other organ systems such as the brain and cardiovascular system. Recently, research has focused on unraveling its involvement in cancer, and emerging evidence suggests a potential role for oxytocin as a cancer biomarker. This review summarizes observations linking oxytocin and cancer, with a special emphasis on prostate cancer, where it may promote cell proliferation. Research suggests that oxytocin effects may depend on cell type, concentration of the hormone, its interactions with other hormones in the microenvironment, and the precise localization of its receptor on the cell membrane. Future research is needed to further elucidate the involvement of oxytocin in cancer, and whether it could be a clinical cancer biomarker or therapeutic target.

Key words: Oxytocin; Cancer; Prostate; Pancreas; Exercise

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Core tip: Oxytocin's role outside of the reproductive system and social bonding has yet to be fully elucidated. Apparently, its role in cancer may vary depending on location and cell type. This review summarizes the current state of our understanding of the potential role of oxytocin in cancer.

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INTRODUCTION

Oxytocin is a central nervous system (CNS) neuropeptide hormone, which is composed of nine amino acids. The synthesis of oxytocin begins in the hypothalamus, where the paraventricular nucleus and supra-optic neurons express high levels of oxytocin, which is released from the posterior pituitary gland^[1]. Oxytocin is biologically similar to vasopressin (also known as antidiuretic hormone), and they are often studied in parallel, as both hormones also share some functions. Originally thought of as a hormone with a role limited to the uterus and milk ejection - oxytocin means "quick birth" in Greek^[2] - further research has expanded understanding of its function across sexes and organ systems. Furthermore, it has become clear that in addition to physical function, oxytocin also wields important impact on social behaviors, which include stress and trust, anxiety, social interaction and bonding, and parental care^[3], and thereby on neuropsychiatric disorders linked to these social behaviors. Interestingly, emerging evidence has linked oxytocin to somewhat conflicting roles in carcinogenesis, as oxytocin is implicated in either fostering development or, conversely, inhibition of cancer-related cellular functional phenomena.

PHYSIOLOGICAL FUNCTIONS OF OXYTOCIN

Oxytocin in the reproductive system

Oxytocin exerts its effects primarily through a single receptor, which has been well characterized. The oxytocin receptor is a class- I G-protein-coupled receptor with seven transmembrane domains, and can be bound by several ligands, including oxytocin, oxytocin agonists and antagonists, as well as vasopressin^[4]. These receptors are found in the endometrium, myometrium, trophoblast, osteoblasts, reproductive organs, and throughout the CNS (Table 1). Notably, oxytocin receptor has also been implicated in various cancers related to tissues in which it is expressed, including endometrial cancer, glioblastomas, neuroblastomas^[5], and others. Several oxytocin receptor antagonists have been identified, with the most common being Atosiban^[6,7]. While studies of oxytocin antagonist in cancer has been limited, atosiban has been implicated in inhibiting cell growth of DU145 prostate cancer^[8] and various breast cancer cell lines^[9]. However, little evidence indicates whether this strategy is efficacious *in vivo*.

The effectors of the oxytocin receptor may vary. While the primary signaling mechanisms of oxytocin have not been fully elucidated, recent studies show that the main mitogenic signaling mechanism of the oxytocin

receptor involves the Gq alpha subunit protein (G α q)/phospholipase C (PLC)/inositol 1, 4, 5 triphosphate (InsP3) pathway. Through this activation, the G protein couples to PLC-B, resulting in the release of calcium from intracellular stores^[10], triggering smooth muscle contractions^[11], for example in the uterus or in the myoepithelial cells of the mammary gland. The main oxytocin pathways described in this paper are depicted in Figure 1.

Indeed, the first functional role attributed to oxytocin centered on the female reproductive system, specifically in uterine contraction and in lactation. Uterine sensitivity to oxytocin increases around the onset of labor, and upon labor oxytocin stimulation becomes more efficient. Therefore, exogenous administration of oxytocin is also clinically used to induce labor. Following parturition, the density of oxytocin receptors declines. In lactation, the oxytocin pathway is activated when the infant begins sucking on the nipple^[12]. A sensory impulse is sent from the nipple to the spinal cord, and from there transmitted to the oxytocinergic neurons in the hypothalamus. These neurons generate action potentials that lead to a substantial release of oxytocin into the blood stream, which subsequently elicits milk ejection via a contraction of myoepithelial cells^[4]. Interestingly, this cascade can be triggered even before suckling occurs, by an event such as a baby crying^[13], suggesting that it may involve reflex neural pathways. Increased plasma oxytocin has also been linked to an increased risk of pregnancy-induced hypertension^[14], and exogenous oxytocin administration has been associated with angiotensin II-induced hypertension^[15]. Interestingly, though, there is also a potential link between increased oxytocin and reduced risk of hypertension (which might be acquired *in utero* in association with intrauterine growth retardation)^[16], so that oxytocin has been attributed with eliciting a reduction in blood pressure^[17,18].

The reproductive function of oxytocin is not limited to females, as it stimulates contractility of the seminiferous tubules, epididymis, and the prostate gland^[19]. Due to its production locale in the testes, oxytocin has been studied as a paracrine regulator of the prostate gland, specifically of growth and muscle contractility^[20]. In males, oxytocin has been thought to induce erection and play a role in ejaculation^[19,21]. Specifically, in the prostate, oxytocin has been suggested to induce prostatic smooth muscle cell contraction^[22]. Its postulated involvement in ejaculation includes stimulation of the reproductive tract to promote sperm release^[19]. Oxytocin's role in aggravating and potentially facilitating the development of benign prostatic hyperplasia, and the oxytocin-induced proliferative effect, are likely mediated through the extracellular signal regulated kinase (ERK) pathway^[23].

Oxytocin in the CNS

Given its ubiquitous distribution in the CNS, the role of oxytocin in cognition and social behavior has also been studied extensively, especially over the past decade.

Table 1 Human tissue with known expression of the oxytocin receptor

Tissue	Expression type
Myometrium	mRNA and protein ^[7,11]
Lung	mRNA ^[109]
Breast	mRNA and protein ^[16]
Prostate	mRNA ^[22,79]
Uterus	mRNA and Protein ^[11]
Heart	mRNA ^[17]
Vascular endothelium	mRNA ^[110]
Brain	mRNA ^[31,42,43]
Thymus	mRNA ^[111]
Pancreas	mRNA ^[112]
Blood	mRNA and protein ^[113]
Bone	mRNA ^[114]

It has been shown that oxytocin can enhance positive social interactions, and importantly can enhance trust^[24]. Oxytocin activity was found to be decreased in women who suffered abuse in their youth^[25]. Conversely, and perhaps related to this, a study reported increased oxytocin levels in individuals enjoying heightened levels of partner support^[26]. Oxytocin also has been reported to improve social cognition^[27,28]. However, it must be noted that studies on the social effects of oxytocin are somewhat inconsistent^[29], suggesting that at least socially, additional factors are likely to be at play.

One of the aims of oxytocin research stems from an attempt to establish it as a tool to predict, diagnose, and potentially treat neuropsychiatric disorders^[30], and has mostly focused on anxiety and depression. Oxytocin intake has been shown to reduce anxiety symptoms^[31,32]. In concordance with its pivotal role during birth, the majority of the research regarding depression in humans has revolved around pregnancies and the mother's ability to recover from postnatal depression (PND). Studies in this field have discovered lowered oxytocin in mothers with PND, and that increased oxytocin levels bestow positive effects on mothers with PND and on their interactions with infants^[33,34]. On the other hand, abnormal post-prandial oxytocin secretion has also been demonstrated in women with anorexia nervosa, possibly as an adaptive response to food-related symptoms of anxiety and depression^[35]. One of the emerging "hot topics" in neuropsychiatric research links oxytocin with autism^[36-38], with recent studies beginning to identify oxytocin as a potential medical therapy to alleviate social anxiety caused by autism^[39].

Oxytocin in the cardiovascular system

The potential regulatory role of oxytocin in other organ systems has also raised considerable interest. Oxytocin contributes to several forms of cardiovascular regulation, as it has been shown that preconditioning rats with oxytocin reduces cardiac arrhythmias^[40], and that oxytocin can lower blood pressure^[41], increase anti-inflammatory and antioxidant activity, and exert beneficial metabolic effects. Therefore, its cardiovascular

activity seems to aim largely at restoring homeostasis.

Notably, oxytocin seems to also exert cardiovascular regulation during elevated levels of physical activity. Oxytocin levels have been shown to rise in response to exercise^[42], which activates oxytocinergic projections^[43] and oxytocinergic modulatory loops that adjust cardiac output, assisting in keeping cardiovascular control over the blood supply^[44]. The rise in oxytocin has been traced to the lumbar spinal cord^[45]. Oxytocin also reduces the rise of exercise-induced adrenocorticotrophic hormone (ACTH) and cortisol^[46,47], furthering support on its effects on cortisol levels. Furthermore, exogenously administered oxytocin along with exercise have been shown to protect ovariectomized rats from myocardial infarction^[48]. On the other hand, its contribution to cardiovascular regulation may depend on the type or intensity of exercise, because plasma oxytocin in cyclists remains unchanged during intense exercise^[49].

OXYTOCIN IN CANCER

Less is understood about the connection between oxytocin and cancer, partly due to lack of adequate research in this area, and partly due to some inconsistency in the current data (Figure 2). The first link of oxytocin to cancer was reported in 1984, when oxytocin was described to be structurally and genomically related to vasopressin, an endogenous hormone that is also secreted by the pituitary, and that in addition to its physiological functions has been found to constitute a biomarker of small-cell lung cancer^[50]. Furthermore, oxytocin and vasopressin are co-expressed in these cells, where they have been proposed to induce mitogenic effects^[51]. Oxytocin's link to vasopressin and its potential role as a biomarker was subsequently proposed in 1990^[52]. Shortly thereafter, it was suggested that oxytocin may modulate growth in breast cancer^[53], which was subsequently demonstrated^[54]. These observations have instigated additional research into oxytocin's potential involvement in various forms of cancer.

Oxytocin in breast cancer

Interestingly, subsequent studies have shown that oxytocin in fact inhibits proliferation of breast cancer cell lines, such as MDA-MB231, MCF7, and T47D^[55,56], as well as the canine mammary cell line CMT-U27^[57], mouse mammary carcinoma cell line TS/A, and rat mammary carcinoma cell line D-R3230AC^[9]. This effect was shown to be mediated via the cyclic adenosine monophosphate protein kinase A in human cell lines^[58]. Importantly, anti-proliferative and tumor inhibitory properties were also observed *in vivo* in both rat and mouse experimental models, and attributed to both oxytocin and its analogue F314^[9]. Recently, it was suggested that exercise training, by inducing oxytocin secretion, may reduce the expression of specific signaling proteins involved in breast cancer^[59].

Lactation has long been linked to a reduced risk

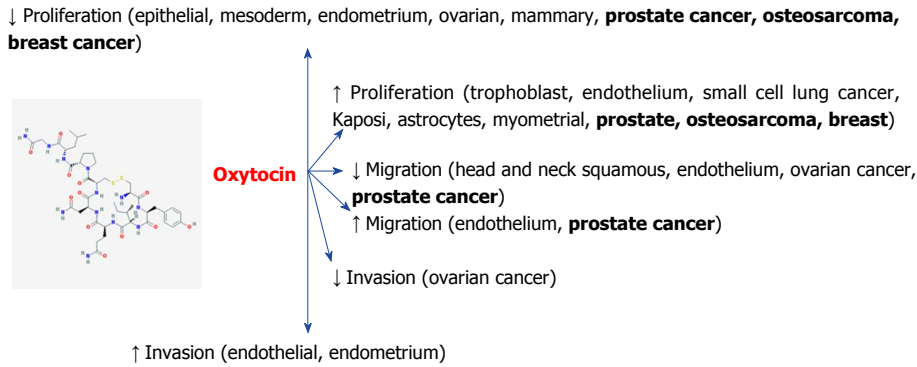


Figure 1 The potential role of oxytocin in various cancers and cell types. Bold font indicates conflicting observations.

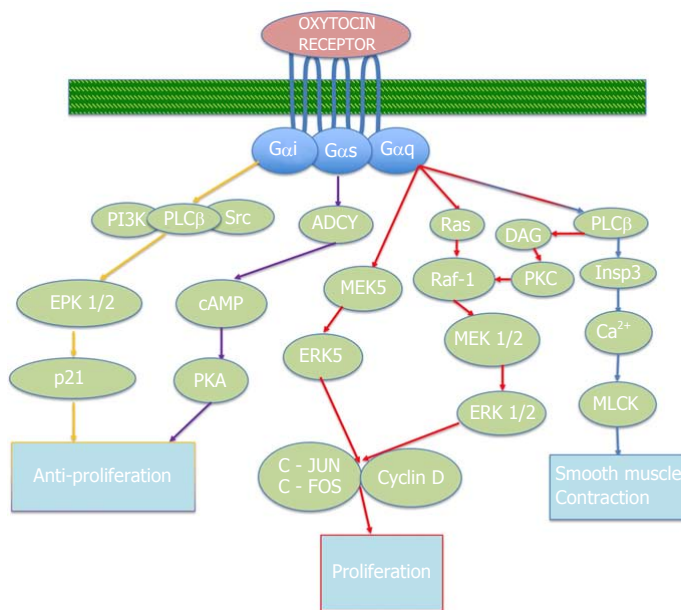


Figure 2 Mechanisms of action of oxytocin. Yellow and purple are anti-proliferation (via different subunits), red is proliferation, and blue is smooth muscle contraction. Gαi: Guanine nucleotide binding protein subunit alpha i; Gαs: Guanine nucleotide binding protein subunit alpha s; Gαq: Guanine nucleotide binding protein subunit alpha q; PI3K: Phosphoinositide 3-kinase; PLCβ: Phospholipase beta; Src: Tyrosine protein kinase; ERK: Extracellular signal-regulated kinase; p21: Cyclin-dependent kinase inhibitor 1; ADCY: Adenylate cyclase; cAMP: Cyclic AMP; PKA: Protein kinase A; DAG: Diacylglycerol; PKC: Protein kinase C; Insp3: Inositol triphosphate 3; MLCK: Myosin light chain kinase.

of cancer, with research dating back to as early as the 1950's^[60-63]. Worldwide, it has been shown that breastfeeding reduces the risk of both breast and uterine cancer, with prolonged durations of breastfeeding (usually involving multiple children breastfed) correlating with a progressive fall in the risks of both breast and uterine cancer^[64-68]. The relationship with uterine cancer might be related to the action of oxytocin as a paracrine and endocrine hormone in lactation. Nevertheless, while the relationship between oxytocin, lactation, and breastfeeding with reduced risk of breast and uterine cancer are all well documented individually, more research needs to be conducted to determine if the relationship between oxytocin, lactation, and breastfeeding with reduced breast and uterine cancer is causal. Elucidating such a connection may establish new therapeutic targets in cancer.

Oxytocin in ovarian cancer

In addition to breast and uterine cancer, the potential participation of oxytocin in the pathogenesis of other cancers in the reproductive system has been investigated. Oxytocin was found to inhibit the progression of ovarian carcinoma (Figure 2) both *in vitro* and *in vivo*. Using cell viability, invasion, and migration assays, it was demonstrated that oxytocin inhibited proliferation, migration and invasion of ovarian cancer cells *in vitro*, and its administration also attenuated the dissemination of ovarian cancer using mean tumor burden as a measure^[69]. The same investigators had demonstrated in a previous study expression of the oxytocin receptor in various human ovarian carcinoma tissues and cell lines, and identified placental leucine aminopeptidase (P-LAP) as an oxytocin-degrading oxytocinase in certain adenocarcinoma tissues^[70]. This team of investigators,

therefore, proposed that a system involving P-LAP and oxytocin plays a role in the regulation of human endometrial adenocarcinoma, in which P-LAP exerts a functionally positive impact on carcinoma cell growth by degrading suppressive peptides such as oxytocin. More recently, these effects have also been linked with a cross-talk network between oxytocin and the stress hormone cortisol, whereby oxytocin reversed the carcinogenic effects of cortisol via autophagy (cellular self-degradation)^[71]. Interestingly, pertinent to the postulated connection between oxytocin and symptoms of autism^[39], oxytocin and cancer have also demonstrated an inverse relationship in autistic children^[72].

Oxytocin in the gastrointestinal tract

Oxytocin receptors are expressed throughout the gastrointestinal (GI) tract^[73], but little is known about their function in the GI tract, especially in relation to cancer. Some studies have suggested a link between oxytocin and its receptor in GI-related cancers, such as esophageal, gastric, and pancreatic cancers. For example, some studies showed an inverse relationship between the duration of breastfeeding and risk of esophageal cancer^[74,75], gastric cancer^[76], and pancreatic cancer. In fact, Yu *et al.*^[78] showed a 54% decreased risk of developing esophageal cancer in women who breastfed for over 12 mo.

Unpublished data from our laboratory shows that the messenger ribonucleic acid (mRNA) expression of oxytocin is twofold higher in PANC-1 (a human pancreatic cancer cell line highly unresponsive to the chemotherapeutic agents, gemcitabine and 5-FU) compared to L3.6pl (a highly responsive human pancreatic cancer cell line). We also found that oxytocin receptor protein expression is also higher in PANC-1 than in L3.6pl. Further, inhibition of the oxytocin receptor decreased cell proliferation of PANC-1 and L3.6pl cells. Our analysis of data from the cBioPortal database revealed that up to 5% of pancreatic cancer patients included in The Cancer Genome Atlas showed genetic alterations (primarily upregulation of mRNA expression) in oxytocin and its receptor. Patients with these alterations had poorer survival outcomes as compared to those without these alterations. These interesting data warrant further investigation on the molecular mechanisms implicating oxytocin and its receptor in pancreatic cancer and other GI cancers.

Oxytocin in prostate cancer

As a role for oxytocin in the regulation of prostate function is established, its potential involvement in the development of prostate cancer has been proposed. Data from over two decades ago implicated oxytocin in the pathophysiology of benign prostatic hyperplasia, where the peptide might contribute to both the physical enlargement and dynamic tone of the gland^[19]. More recently, immunohistochemical staining has detected oxytocin expression in stromal and epithelial cell lines and in tissue from patients with benign prostatic hyperplasia,

which was significantly reduced in tissues of invasive prostate cancer in comparison to both benign prostatic hyperplasia tissues and normal human prostate epithelial cells^[79]. This inverse relationship might implicate a fall in oxytocin levels in progression of prostate cancer. Within the prostate, oxytocin has been shown to affect gland growth both directly and *via* its interaction with androgen metabolism, and oxytocin concentrations are positively correlated with androgens^[20,80]. Indeed, while in the absence of androgens oxytocin had no effect on prostate cancer cell lines (LNCaP and PC-3), in the presence of testosterone low oxytocin doses stimulated proliferation of PC-3 cells^[81], supporting the notion that changes in levels of oxytocin in the prostate in aging and cancer may promote prostate epithelial cell proliferation. It is possible that increased levels of oxytocin might be involved in the mechanisms by which high ejaculation frequency is related to decreased risk of prostate cancer^[82]. This hypothesis needs to be further investigated.

Conversely, a different study recently revealed that oxytocin increased the expression of APPL1, a protein with the ability to interact with tumor suppressor proteins. *In vitro* studies showed that oxytocin increased prostate cancer cell proliferation, and expression of APPL1. Analysis of serum and tissue samples identified increased oxytocin levels in the serum of prostate cancer patients, and high expression of oxytocin and its receptor in prostate tissues collected from prostate cancer patients in comparison to those collected from patients without prostate cancer. The oxytocin receptor has also been implicated in the migration of prostate cancer cells, and possibly modulation of prostate cancer metastasis^[83]. Taken together, these observations of oxytocin in prostate cancer cells both *in vivo* and *in vitro*, suggest that oxytocin could serve as a prostate cancer biomarker^[84].

Several explanations have been offered for the apparent differences in the data from different studies regarding the role of oxytocin in prostate cancer. One explanation is the notable difference in the numbers of participants involved in each study. Secondly, some of the studies included prostate cancer patients that had undergone neo-adjuvant therapy, which can affect oxytocin levels^[85]. Thirdly, oxytocin is likely to activate a wide range of signaling mechanisms to elicit variable cellular responses, possibly depending on the density or precise localization of the oxytocin receptor on the plasma membrane^[86]. This may also account for the dichotomy in the observations reported regarding the role of oxytocin in cancer. Clearly, additional studies are needed to elucidate the involvement of oxytocin and oxytocin receptor in progression or regression of human prostate cancer.

Perspectives and future directions

There is clearly some evidence implicating oxytocin in carcinogenesis, although its precise effect and underlying mechanisms are still unclear. It is possible that in some

individuals, cell types, or types of cancers, oxytocin may not act as a sole regulator of carcinogenesis, but may mediate or modulate other coexisting factors in the microenvironment.

For example, research generally supports the hypothesis that exercise can inhibit the progression of cancer^[87-89]. The positive impact of exercise in blunting development of cancer and facilitating recovery may potentially be partly mediated through oxytocin. For example, oxytocin has been proposed to mediate an exercise-induced reduction in the expression of specific signaling molecules involved in breast cancer^[59]. It has also recently been shown *in vivo* that the combination of exogenous oxytocin with exercise improves cardiac function, which might be associated with improved cancer survival. Cardiac dysfunction and cancer have such a strong link that it has even spawned its own subspecialty in cardio-oncology^[90-93], although much of the research has centered around long-term survival and cardiac complications in cancer patients. While this potential mediating effect of oxytocin has been hypothesized^[94], it appears that little primary research has been conducted to address this postulation. The ability to knockout or silence oxytocin is available, and its social effects are already documented^[95,96]. Thus, a thorough study is certainly possible. Similarly, oxytocin may mediate the inhibitory effects of lactation on development of breast cancer, and of ejaculation on development of prostate cancer, and additional studies could prove to be invaluable in revealing its involvement.

It must be noted that most systems and bodily processes in living organisms are tightly inter-connected. Untangling this complexity in the presence of confounding elements is often difficult, especially in relation to psychosocial factors. Therefore, interpretation of oxytocin's expression and levels alone might be over-simplistic and under-informative. For example, instead of being increased as a direct response to exercise, oxytocin may be induced to assist its "companion" hormone, vasopressin, a well-known hormonal regulator of body fluid homeostasis^[97]. Cortisol, the "stress hormone" and the rhythm surrounding its release has been linked to the progression and survival from various cancers^[98-101], and has also been linked to oxytocin^[71,72]. The ability to completely elucidate various pathways and mechanisms of actions would go a long way to showing if the established connection between the different hormones extends to cancer. Furthermore, while outside the scope of this review, there are several other diseases and pathologic states possibly linked to oxytocin. Pain, depression, and anxiety have all been linked to oxytocin^[102,103]. Oxytocin's role in depression management was mentioned previously in this article, but oxytocin also seems to be a promising target in pain management^[104-107], and in immunotherapy, especially through its interactions in the gut^[108].

CONCLUSION

Most knowledge of oxytocin centers on its role as a reproductive hormone. Since its discovery, its other

roles have progressively become clearer, including involvement in social behavior, cardiovascular regulation, and carcinogenesis. While it is currently difficult to pinpoint and precisely define oxytocin's oncogenic roles, it is hoped that this review will encourage greater intensity in researching the details of the role of oxytocin in cancer. Future research has a number of plausible and exciting directions to follow and will hopefully clarify some of the ambiguities concerning the role of oxytocin in cancer.

REFERENCES

- 1 **Viero C**, Shibuya I, Kitamura N, Verkhatsky A, Fujihara H, Katoh A, Ueta Y, Zingg HH, Chvatal A, Sykova E, Dayanithi G. REVIEW: Oxytocin: Crossing the bridge between basic science and pharmacotherapy. *CNS Neurosci Ther* 2010; **16**: e138-e156 [PMID: 20626426 DOI: 10.1111/j.1755-5949.2010.00185.x]
- 2 **Grigor'eva ME**, Golubeva MG. Oxytocin: Structure, synthesis, receptors, and basic effects. *Neurochem J* 2010; **4**: 75-83 [DOI: 10.1134/S1819712410020017]
- 3 **Lopatina O**, Inzhutova A, Salmina AB, Higashida H. The roles of oxytocin and CD38 in social or parental behaviors. *Front Neurosci* 2013; **6**: 182 [PMID: 23335873 DOI: 10.3389/fnins.2012.00182]
- 4 **Gimpl G**, Fahrenholz F. The oxytocin receptor system: structure, function, and regulation. *Physiol Rev* 2001; **81**: 629-683 [PMID: 11274341 DOI: 10.1152/physrev.2001.81.2.629]
- 5 **Chatterjee O**, Patil K, Sahu A, Gopalakrishnan L, Mol P, Advani J, Mukherjee S, Christopher R, Prasad TS. An overview of the oxytocin-oxytocin receptor signaling network. *J Cell Commun Signal* 2016; **10**: 355-360 [PMID: 27624619 DOI: 10.1007/s12079-016-0353-7]
- 6 **Kim SH**, MacIntyre DA, Hanyaloglu AC, Blanks AM, Thornton S, Bennett PR, Terzidou V. The oxytocin receptor antagonist, Atosiban, activates pro-inflammatory pathways in human amnion via G(α i) signalling. *Mol Cell Endocrinol* 2016; **420**: 11-23 [PMID: 26586210 DOI: 10.1016/j.mce.2015.11.012]
- 7 **Kim SH**, Pohl O, Chollet A, Gotteland JP, Fairhurst AD, Bennett PR, Terzidou V. Differential Effects of Oxytocin Receptor Antagonists, Atosiban and Nolasiban, on Oxytocin Receptor-Mediated Signaling in Human Amnion and Myometrium. *Mol Pharmacol* 2017; **91**: 403-415 [PMID: 28188254 DOI: 10.1124/mol.116.106013]
- 8 **Reversi A**, Rimoldi V, Marrocco T, Cassoni P, Bussolati G, Parenti M, Chini B. The oxytocin receptor antagonist atosiban inhibits cell growth via a "biased agonist" mechanism. *J Biol Chem* 2005; **280**: 16311-16318 [PMID: 15705593 DOI: 10.1074/jbc.M409945200]
- 9 **Cassoni P**, Sapino A, Papotti M, Bussolati G. Oxytocin and oxytocin-analogue F314 inhibit cell proliferation and tumor growth of rat and mouse mammary carcinomas. *Int J Cancer* 1996; **66**: 817-820 [PMID: 8647655 DOI: 10.1002/(SICI)1097-0215(19960611)66:6<817::AID-IJC18>3.0.CO;2-#]
- 10 **Vrachnis N**, Malamas FM, Sifakis S, Deligeorgiou E, Iliodromiti Z. The oxytocin-oxytocin receptor system and its antagonists as tocolytic agents. *Int J Endocrinol* 2011; **2011**: 350546 [PMID: 22190926 DOI: 10.1155/2011/350546]
- 11 **Arrowsmith S**, Wray S. Oxytocin: its mechanism of action and receptor signalling in the myometrium. *J Neuroendocrinol* 2014; **26**: 356-369 [PMID: 24888645 DOI: 10.1111/jne.12154]
- 12 **Zingg HH**, Lefebvre DL. Oxytocin and vasopressin gene expression during gestation and lactation. *Brain Res* 1988; **464**: 1-6 [PMID: 3179741 DOI: 10.1016/0169-328X(88)90011-3]
- 13 **McNeilly AS**, Robinson IC, Houston MJ, Howie PW. Release of oxytocin and prolactin in response to suckling. *Br Med J (Clin Res Ed)* 1983; **286**: 257-259 [PMID: 6402061 DOI: 10.1136/bmj.286.6361.257]
- 14 **Gu H**, Rong L, Sha JY. [Changes in blood oxytocin levels in cases of pregnancy induced hypertension]. *Zhonghua Fuchanke Zazhi* 1994; **29**: 268-270, 316 [PMID: 7956547]
- 15 **Phie J**, Haleagrahara N, Newton P, Constantinoiu C, Sarnyai Z, Chilton L, Kinobe R. Prolonged Subcutaneous Administration of

- Oxytocin Accelerates Angiotensin II-Induced Hypertension and Renal Damage in Male Rats. *PLoS One* 2015; **10**: e0138048 [PMID: 26393919 DOI: 10.1371/journal.pone.0138048]
- 16 **Vargas-Martínez F**, Schanler RJ, Abrams SA, Hawthorne KM, Landers S, Guzman-Bárceñas J, Muñoz O, Henriksen T, Petersson M, Uvnäs-Moberg K, Jiménez-Estrada I. Oxytocin, a main breastfeeding hormone, prevents hypertension acquired in utero: A therapeutics preview. *Biochim Biophys Acta* 2017; **1861**: 3071-3084 [PMID: 27658996 DOI: 10.1016/j.bbagen.2016.09.020]
 - 17 **Light KC**, Grewen KM, Amico JA. More frequent partner hugs and higher oxytocin levels are linked to lower blood pressure and heart rate in premenopausal women. *Biol Psychol* 2005; **69**: 5-21 [PMID: 15740822 DOI: 10.1016/j.biopsycho.2004.11.002]
 - 18 **Petersson M**, Alster P, Lundeberg T, Uvnäs-Moberg K. Oxytocin causes a long-term decrease of blood pressure in female and male rats. *Physiol Behav* 1996; **60**: 1311-1315 [PMID: 8916187 DOI: 10.1016/S0031-9384(96)00261-2]
 - 19 **Thackare H**, Nicholson HD, Whittington K. Oxytocin--its role in male reproduction and new potential therapeutic uses. *Hum Reprod Update* 2006; **12**: 437-448 [PMID: 16436468 DOI: 10.1093/humupd/dmk002]
 - 20 **Nicholson HD**. Oxytocin: a paracrine regulator of prostatic function. *Rev Reprod* 1996; **1**: 69-72 [PMID: 9414441 DOI: 10.1530/ror.0.0010069]
 - 21 **Andersson KE**. Mechanisms of penile erection and basis for pharmacological treatment of erectile dysfunction. *Pharmacol Rev* 2011; **63**: 811-859 [PMID: 21880989 DOI: 10.1124/pr.111.004515]
 - 22 **Bodanszky M**, Sharaf H, Roy JB, Said SI. Contractile activity of vasotocin, oxytocin, and vasopressin on mammalian prostate. *Eur J Pharmacol* 1992; **216**: 311-313 [PMID: 1397015 DOI: 10.1016/0014-2999(92)90376-F]
 - 23 **Xu H**, Fu S, Chen Y, Chen Q, Gu M, Liu C, Qiao Z, Zhou J, Wang Z. Oxytocin: its role in benign prostatic hyperplasia via the ERK pathway. *Clin Sci (Lond)* 2017; **131**: 595-607 [PMID: 28130436 DOI: 10.1042/CS20170030]
 - 24 **Misrani A**, Tabassum S, Long C. Oxytocin system in neuropsychiatric disorders: Old concept, new insights. *Shengli Xuebao* 2017; **69**: 196-206 [PMID: 28435979]
 - 25 **Heim C**, Young LJ, Newport DJ, Mletzko T, Miller AH, Nemeroff CB. Lower CSF oxytocin concentrations in women with a history of childhood abuse. *Mol Psychiatry* 2009; **14**: 954-958 [PMID: 18957940 DOI: 10.1038/mp.2008.112]
 - 26 **Grewen KM**, Girdler SS, Amico J, Light KC. Effects of partner support on resting oxytocin, cortisol, norepinephrine, and blood pressure before and after warm partner contact. *Psychosom Med* 2005; **67**: 531-538 [PMID: 16046364 DOI: 10.1097/01.psy.0000170341.88395.47]
 - 27 **Guastella AJ**, Mitchell PB, Mathews F. Oxytocin enhances the encoding of positive social memories in humans. *Biol Psychiatry* 2008; **64**: 256-258 [PMID: 18343353 DOI: 10.1016/j.biopsych.2008.02.008]
 - 28 **Bartz JA**, Zaki J, Bolger N, Hollander E, Ludwig NN, Kolevzon A, Ochsner KN. Oxytocin selectively improves empathic accuracy. *Psychol Sci* 2010; **21**: 1426-1428 [PMID: 20855907 DOI: 10.1177/0956797610383439]
 - 29 **Bartz JA**, Zaki J, Bolger N, Ochsner KN. Social effects of oxytocin in humans: context and person matter. *Trends Cogn Sci* 2011; **15**: 301-309 [PMID: 21696997 DOI: 10.1016/j.tics.2011.05.002]
 - 30 **Marazziti D**, Catena Dell'osso M. The role of oxytocin in neuropsychiatric disorders. *Curr Med Chem* 2008; **15**: 698-704 [PMID: 18336283 DOI: 10.2174/092986708783885291]
 - 31 **MacDonald K**, Feifel D. Oxytocin's role in anxiety: a critical appraisal. *Brain Res* 2014; **1580**: 22-56 [PMID: 24468203 DOI: 10.1016/j.brainres.2014.01.025]
 - 32 **Alvares GA**, Chen NT, Balleine BW, Hickie IB, Guastella AJ. Oxytocin selectively moderates negative cognitive appraisals in high trait anxious males. *Psychoneuroendocrinology* 2012; **37**: 2022-2031 [PMID: 22613033 DOI: 10.1016/j.psyneuen.2012.04.018]
 - 33 **Moura D**, Canavarro MC, Figueiredo-Braga M. Oxytocin and depression in the perinatal period-a systematic review. *Arch Womens Ment Health* 2016; **19**: 561-570 [PMID: 27295067 DOI: 10.1007/s00737-016-0643-3]
 - 34 **Mah BL**. Oxytocin, Postnatal Depression, and Parenting: A Systematic Review. *Harv Rev Psychiatry* 2016; **24**: 1-13 [PMID: 26735320 DOI: 10.1097/HRP.0000000000000093]
 - 35 **Lawson EA**, Holsen LM, Santin M, DeSanti R, Meenaghan E, Eddy KT, Herzog DB, Goldstein JM, Klibanski A. Postprandial oxytocin secretion is associated with severity of anxiety and depressive symptoms in anorexia nervosa. *J Clin Psychiatry* 2013; **74**: e451-e457 [PMID: 23759466 DOI: 10.4088/JCP.12m08154]
 - 36 **Vanya M**, Szucs S, Vetro A, Bartfai G. The potential role of oxytocin and perinatal factors in the pathogenesis of autism spectrum disorders - review of the literature. *Psychiatry Res* 2017; **247**: 288-290 [PMID: 27974283 DOI: 10.1016/j.psychres.2016.12.007]
 - 37 **Zhang R**, Zhang HF, Han JS, Han SP. Genes Related to Oxytocin and Arginine-Vasopressin Pathways: Associations with Autism Spectrum Disorders. *Neurosci Bull* 2017; **33**: 238-246 [PMID: 28283809 DOI: 10.1007/s12264-017-0120-7]
 - 38 **Ooi YP**, Weng SJ, Kossowsky J, Gerger H, Sung M. Oxytocin and Autism Spectrum Disorders: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Pharmacopsychiatry* 2017; **50**: 5-13 [PMID: 27574858 DOI: 10.1055/s-0042-109400]
 - 39 **Kanat M**, Spenthof I, Riedel A, van Elst LT, Heinrichs M, Domes G. Restoring effects of oxytocin on the attentional preference for faces in autism. *Transl Psychiatry* 2017; **7**: e1097 [PMID: 28418399 DOI: 10.1038/tp.2017.67]
 - 40 **Faghihi M**, Alizadeh AM, Khor V, Khodayari V, Moradi S. Preconditioning effects of oxytocin in reducing cardiac arrhythmias in a rat heart regional ischemia-reperfusion model. *Physiol Pharmacol* 2012; **16**: 393-403
 - 41 **Gutkowska J**, Jankowski M, Antunes-Rodrigues J. The role of oxytocin in cardiovascular regulation. *Braz J Med Biol Res* 2014; **47**: 206-214 [PMID: 24676493 DOI: 10.1590/1414-431X20133309]
 - 42 **Hew-Butler T**, Noakes TD, Soldin SJ, Verbalis JG. Acute changes in endocrine and fluid balance markers during high-intensity, steady-state, and prolonged endurance running: unexpected increases in oxytocin and brain natriuretic peptide during exercise. *Eur J Endocrinol* 2008; **159**: 729-737 [PMID: 18794210 DOI: 10.1530/EJE-08-0064]
 - 43 **Martins AS**, Crescenzi A, Stern JE, Bordin S, Michelini LC. Hypertension and exercise training differentially affect oxytocin and oxytocin receptor expression in the brain. *Hypertension* 2005; **46**: 1004-1009 [PMID: 16157794 DOI: 10.1161/01.HYP.0000175812.03322.59]
 - 44 **Michelini LC**. Differential effects of vasopressinergic and oxytocinergic pre-autonomic neurons on circulatory control: reflex mechanisms and changes during exercise. *Clin Exp Pharmacol Physiol* 2007; **34**: 369-376 [PMID: 17324152 DOI: 10.1111/j.1440-1681.2007.04589.x]
 - 45 **Stebbins CL**, Ortiz-Acevedo A. The exercise pressor reflex is attenuated by intrathecal oxytocin. *Am J Physiol* 1994; **267**: R909-R915 [PMID: 7943431 DOI: 10.1152/ajpregu.1994.267.4.R909]
 - 46 **Coiro V**, Passeri M, Davoli C, Bacchi-Modena A, Bianconi L, Volpi R, Chiodera P. Oxytocin reduces exercise-induced ACTH and cortisol rise in man. *Acta Endocrinol (Copenh)* 1988; **119**: 405-412 [PMID: 2847472 DOI: 10.1530/acta.0.1190405]
 - 47 **Cardoso C**, Ellenbogen MA, Orlando MA, Bacon SL, Joobar R. Intranasal oxytocin attenuates the cortisol response to physical stress: a dose-response study. *Psychoneuroendocrinology* 2013; **38**: 399-407 [PMID: 22889586 DOI: 10.1016/j.psyneuen.2012.07.013]
 - 48 **Bulut EC**, Abueid L, Ercan F, Süleymanoğlu S, Ağırbaşı M, Yeğen BÇ. Treatment with oestrogen-receptor agonists or oxytocin in conjunction with exercise protects against myocardial infarction in ovariectomized rats. *Exp Physiol* 2016; **101**: 612-627 [PMID: 26958805 DOI: 10.1113/EP085708]
 - 49 **Chicharro JL**, Hoyos J, Bandrés F, Gómez Gallego F, Pérez M, Lucia A. Plasma oxytocin during intense exercise in professional cyclists. *Horm Res* 2001; **55**: 155-159 [PMID: 11549878 DOI: 10.1159/000049988]
 - 50 **Sausville E**, Carney D, Battey J. The human vasopressin gene is linked to the oxytocin gene and is selectively expressed in a cultured lung cancer cell line. *J Biol Chem* 1985; **260**: 10236-10241 [PMID:

- 2991279]
- 51 **Péqueux C**, Keegan BP, Hagelstein MT, Geenen V, Legros JJ, North WG. Oxytocin- and vasopressin-induced growth of human small-cell lung cancer is mediated by the mitogen-activated protein kinase pathway. *Endocr Relat Cancer* 2004; **11**: 871-885 [PMID: 15613460 DOI: 10.1677/erc.1.00803]
 - 52 **Legros JJ**, Geenen V, Carvelli T, Martens H, Andre M, Corhay JL, Radermecker M, Zangerle PF, Sassolas G, Gharib C. Neurophysins as markers of vasopressin and oxytocin release. A study in carcinoma of the lung. *Horm Res* 1990; **34**: 151-155 [PMID: 1966564 DOI: 10.1159/000181815]
 - 53 **Taylor AH**, Ang VT, Jenkins JS, Silverlight JJ, Coombes RC, Luqmani YA. Interaction of vasopressin and oxytocin with human breast carcinoma cells. *Cancer Res* 1990; **50**: 7882-7886 [PMID: 2174737]
 - 54 **Cassoni P**, Sapino A, Negro F, Bussolati G. Oxytocin inhibits proliferation of human breast cancer cell lines. *Virchows Arch* 1994; **425**: 467-472 [PMID: 7850070 DOI: 10.1007/BF00197549]
 - 55 **Cassoni P**, Sapino A, Marrocco T, Chini B, Bussolati G. Oxytocin and oxytocin receptors in cancer cells and proliferation. *J Neuroendocrinol* 2004; **16**: 362-364 [PMID: 15089975 DOI: 10.1111/j.0953-8194.2004.01165.x]
 - 56 **Cassoni P**, Catalano MG, Sapino A, Marrocco T, Fazzari A, Bussolati G, Fortunati N. Oxytocin modulates estrogen receptor alpha expression and function in MCF7 human breast cancer cells. *Int J Oncol* 2002; **21**: 375-378 [PMID: 12118334 DOI: 10.3892/ijo.21.2.375]
 - 57 **Benavente MA**, Bianchi CP, Imperiale F, Aba MA. Antiproliferative Effects of Oxytocin and Desmopressin on Canine Mammary Cancer Cells. *Front Vet Sci* 2016; **3**: 119 [PMID: 28083539 DOI: 10.3389/fvets.2016.00119]
 - 58 **Cassoni P**, Sapino A, Fortunati N, Munaron L, Chini B, Bussolati G. Oxytocin inhibits the proliferation of MDA-MB231 human breast-cancer cells via cyclic adenosine monophosphate and protein kinase A. *Int J Cancer* 1997; **72**: 340-344 [PMID: 9219843]
 - 59 **Alizadeh AM**, Heydari Z, Rahimi M, Bazgir B, Shirvani H, Alipour S, Heidarian Y, Khalighfar S, Isanejad A. Oxytocin mediates the beneficial effects of the exercise training on breast cancer. *Exp Physiol* 2018; **103**: 222-235 [PMID: 29143998 DOI: 10.1113/EP086463]
 - 60 **King AG**. Suppression of lactation and its relation to cancer of the breast. *J Med Assoc Ga* 1958; **47**: 342-344 [PMID: 13564083]
 - 61 **Lewison EF**. Breast cancer and pregnancy or lactation. *Int Abstr Surg* 1954; **99**: 417-424 [PMID: 13205440]
 - 62 **Huggins C**, Dao TL. Lactation induced by luteotrophin in women with mammary cancer; growth of the breast of the human male following estrogenic treatment. *Cancer Res* 1954; **14**: 303-306 [PMID: 13160955]
 - 63 **Abrao A**, Da Silva Neto JB, Mirra AP. Cancer of the breast in pregnancy and lactation; study of 10 cases. *Rev Paul Med* 1954; **45**: 563-570 [PMID: 14372504]
 - 64 **Sugawara Y**, Kakizaki M, Nagai M, Tomata Y, Hoshi R, Watanabe I, Nishino Y, Kuriyama S, Tsuji I. Lactation pattern and the risk for hormone-related female cancer in Japan: the Ohsaki Cohort Study. *Eur J Cancer Prev* 2013; **22**: 187-192 [PMID: 23358107 DOI: 10.1097/CEJ.0b013e3283564610]
 - 65 **Su D**, Pasalich M, Lee AH, Binns CW. Ovarian cancer risk is reduced by prolonged lactation: a case-control study in southern China. *Am J Clin Nutr* 2013; **97**: 354-359 [PMID: 23283498 DOI: 10.3945/ajcn.112.044719]
 - 66 **Jordan I**, Hebestreit A, Swai B, Krawinkel MB. Breast cancer risk among women with long-standing lactation and reproductive parameters at low risk level: a case-control study in Northern Tanzania. *Breast Cancer Res Treat* 2013; **142**: 133-141 [PMID: 21104009 DOI: 10.1007/s10549-010-1255-7]
 - 67 **Zheng T**, Holford TR, Mayne ST, Owens PH, Zhang Y, Zhang B, Boyle P, Zahm SH. Lactation and breast cancer risk: a case-control study in Connecticut. *Br J Cancer* 2001; **84**: 1472-1476 [PMID: 11384096 DOI: 10.1054/bjoc.2001.1793]
 - 68 **Purwanto H**, Sadjimin T, Dwiprahasto I. Lactation and the risk of breast cancer. *Gan To Kagaku Ryoho* 2000; **27** Suppl 2: 474-481 [PMID: 10895198]
 - 69 **Morita T**, Shibata K, Kikkawa F, Kajiyama H, Ino K, Mizutani S. Oxytocin inhibits the progression of human ovarian carcinoma cells in vitro and in vivo. *Int J Cancer* 2004; **109**: 525-532 [PMID: 14991573 DOI: 10.1002/ijc.20017]
 - 70 **Suzuki Y**, Shibata K, Kikkawa F, Kajiyama H, Ino K, Nomura S, Tsujimoto M, Mizutani S. Possible role of placental leucine aminopeptidase in the antiproliferative effect of oxytocin in human endometrial adenocarcinoma. *Clin Cancer Res* 2003; **9**: 1528-1534 [PMID: 12684429]
 - 71 **Mankarious A**, Dave F, Pados G, Tsolakidis D, Gidron Y, Pang Y, Thomas P, Hall M, Karteris E. The pro-social neurohormone oxytocin reverses the actions of the stress hormone cortisol in human ovarian carcinoma cells in vitro. *Int J Oncol* 2016; **48**: 1805-1814 [PMID: 26935408 DOI: 10.3892/ijo.2016.3410]
 - 72 **Corbett BA**, Bales KL, Swain D, Sanders K, Weinstein TA, Muglia LJ. Comparing oxytocin and cortisol regulation in a double-blind, placebo-controlled, hydrocortisone challenge pilot study in children with autism and typical development. *J Neurodev Disord* 2016; **8**: 32 [PMID: 27540420 DOI: 10.1186/s11689-016-9165-6]
 - 73 **Monstein HJ**, Grahn N, Truedsson M, Ohlsson B. Oxytocin and oxytocin-receptor mRNA expression in the human gastrointestinal tract: a polymerase chain reaction study. *Regul Pept* 2004; **119**: 39-44 [PMID: 15093695 DOI: 10.1016/j.regpep.2003.12.017]
 - 74 **Lindblad M**, García Rodríguez LA, Chandanos E, Lagergren J. Hormone replacement therapy and risks of oesophageal and gastric adenocarcinomas. *Br J Cancer* 2006; **94**: 136-141 [PMID: 16404367 DOI: 10.1038/sj.bjc.6602906]
 - 75 **Cronin-Fenton DP**, Murray LJ, Whiteman DC, Cardwell C, Webb PM, Jordan SJ, Corley DA, Sharp L, Lagergren J, Barrett's Esophagus, Adenocarcinoma Consortium (BEACON) Investigators. Reproductive and sex hormonal factors and oesophageal and gastric junction adenocarcinoma: a pooled analysis. *Eur J Cancer* 2010; **46**: 2067-2076 [PMID: 20456945 DOI: 10.1016/j.ejca.2010.03.032]
 - 76 **Inoue H**, Matsuyama A, Mimori K, Ueo H, Mori M. Prognostic score of gastric cancer determined by cDNA microarray. *Clin Cancer Res* 2002; **8**: 3475-3479 [PMID: 12429637]
 - 77 **Skinner HG**, Michaud DS, Colditz GA, Giovannucci EL, Stampfer MJ, Willett WC, Fuchs CS. Parity, reproductive factors, and the risk of pancreatic cancer in women. *Cancer Epidemiol Biomarkers Prev* 2003; **12**: 433-438 [PMID: 12750238]
 - 78 **Yu H**, Liu G, Zhao P, Zhu L. Hormonal and reproductive factors and risk of esophageal cancer in Chinese postmenopausal women: a case-control study. *Asian Pac J Cancer Prev* 2011; **12**: 1953-1956 [PMID: 22292631]
 - 79 **Whittington K**, Assinder S, Gould M, Nicholson H. Oxytocin, oxytocin-associated neurophysin and the oxytocin receptor in the human prostate. *Cell Tissue Res* 2004; **318**: 375-382 [PMID: 15459766 DOI: 10.1007/s00441-004-0968-5]
 - 80 **Biswas M**, Thackare H, Jones MK, Bowen-Simpkins P. Lymphocytic hypophysitis and headache in pregnancy. *BJOG* 2002; **109**: 1184-1186 [PMID: 12387476]
 - 81 **Whittington K**, Connors B, King K, Assinder S, Hogarth K, Nicholson H. The effect of oxytocin on cell proliferation in the human prostate is modulated by gonadal steroids: implications for benign prostatic hyperplasia and carcinoma of the prostate. *Prostate* 2007; **67**: 1132-1142 [PMID: 17492653 DOI: 10.1002/pros.20612]
 - 82 **Leitzmann MF**, Platz EA, Stampfer MJ, Willett WC, Giovannucci E. Ejaculation frequency and subsequent risk of prostate cancer. *JAMA* 2004; **291**: 1578-1586 [PMID: 15069045 DOI: 10.1001/jama.291.13.1578]
 - 83 **Zhong M**, Boseman ML, Millena AC, Khan SA. Oxytocin induces the migration of prostate cancer cells: involvement of the Gi-coupled signaling pathway. *Mol Cancer Res* 2010; **8**: 1164-1172 [PMID: 20663860 DOI: 10.1158/1541-7786.MCR-09-0329]
 - 84 **Xu H**, Fu S, Chen Q, Gu M, Zhou J, Liu C, Chen Y, Wang Z. The function of oxytocin: a potential biomarker for prostate cancer diagnosis and promoter of prostate cancer. *Oncotarget* 2017; **8**: 31215-31226 [PMID: 28415720 DOI: 10.18632/oncotarget.16107]
 - 85 **Nicholson HD**, Jenkin L. Oxytocin and prostatic function. *Adv Exp Med Biol* 1995; **395**: 529-538 [PMID: 8714010]
 - 86 **Rimoldi V**, Reversi A, Taverna E, Rosa P, Francolini M, Cassoni

- P, Parenti M, Chini B. Oxytocin receptor elicits different EGFR/MAPK activation patterns depending on its localization in caveolin-1 enriched domains. *Oncogene* 2003; **22**: 6054-6060 [PMID: 12955084 DOI: 10.1038/sj.onc.1206612]
- 87 **Knols R**, Aaronson NK, Uebelhart D, Fransen J, Aufdemkampe G. Physical exercise in cancer patients during and after medical treatment: a systematic review of randomized and controlled clinical trials. *J Clin Oncol* 2005; **23**: 3830-3842 [PMID: 15923576 DOI: 10.1200/JCO.2005.02.148]
 - 88 **Giovannucci EL**, Liu Y, Leitzmann MF, Stampfer MJ, Willett WC. A prospective study of physical activity and incident and fatal prostate cancer. *Arch Intern Med* 2005; **165**: 1005-1010 [PMID: 15883238 DOI: 10.1001/archinte.165.9.1005]
 - 89 **Liu Y**, Hu F, Li D, Wang F, Zhu L, Chen W, Ge J, An R, Zhao Y. Does physical activity reduce the risk of prostate cancer? A systematic review and meta-analysis. *Eur Urol* 2011; **60**: 1029-1044 [PMID: 21802197 DOI: 10.1016/j.eururo.2011.07.007]
 - 90 **Tajiri K**, Aonuma K, Sekine I. Cardio-oncology: a multidisciplinary approach for detection, prevention and management of cardiac dysfunction in cancer patients. *Jpn J Clin Oncol* 2017; **47**: 678-682 [PMID: 28505345 DOI: 10.1093/jjco/hyx068]
 - 91 **Ryzhov S**, Francis S, Sawyer DB. Cardiac Dysfunction Due to Cancer Therapy: Finding New Directions. *Circ Res* 2016; **119**: 1055-1056 [PMID: 27789582 DOI: 10.1161/CIRCRESAHA.116.309902]
 - 92 **Gulati G**, Heck SL, Ree AH, Hoffmann P, Schulz-Menger J, Fagerland MW, Gravdehaug B, von Knobelsdorff-Brenkenhoff F, Bratland Å, Storås TH, Hagve TA, Røsjø H, Steine K, Geisler J, Omland T. Prevention of cardiac dysfunction during adjuvant breast cancer therapy (PRADA): a 2 × 2 factorial, randomized, placebo-controlled, double-blind clinical trial of candesartan and metoprolol. *Eur Heart J* 2016; **37**: 1671-1680 [PMID: 26903532 DOI: 10.1093/eurheartj/ehw022]
 - 93 **Ewer MS**, Swain SM, Cardinale D, Fadol A, Suter TM. Cardiac dysfunction after cancer treatment. *Tex Heart Inst J* 2011; **38**: 248-252 [PMID: 21720462]
 - 94 **Imanieh MH**, Bagheri F, Alizadeh AM, Ashkani-Esfahani S. Oxytocin has therapeutic effects on cancer, a hypothesis. *Eur J Pharmacol* 2014; **741**: 112-123 [PMID: 25094035 DOI: 10.1016/j.ejphar.2014.07.053]
 - 95 **Winslow JT**, Insel TR. The social deficits of the oxytocin knockout mouse. *Neuropeptides* 2002; **36**: 221-229 [PMID: 12359512 DOI: 10.1054/npep.2002.0909]
 - 96 **Nakajima M**, Görlich A, Heintz N. Oxytocin modulates female sociosexual behavior through a specific class of prefrontal cortical interneurons. *Cell* 2014; **159**: 295-305 [PMID: 25303526 DOI: 10.1016/j.cell.2014.09.020]
 - 97 **el-Sayed MS**, Davies B, Morgan DB. Vasopressin and plasma volume response to submaximal and maximal exercise in man. *J Sports Med Phys Fitness* 1990; **30**: 420-425 [PMID: 2079849]
 - 98 **Fabre B**, Grosman H, Gonzalez D, Machulsky NF, Repetto EM, Mesch V, Lopez MA, Mazza O, Berg G. Prostate Cancer, High Cortisol Levels and Complex Hormonal Interaction. *Asian Pac J Cancer Prev* 2016; **17**: 3167-3171 [PMID: 27509946]
 - 99 **Schrepf A**, Thaker PH, Goodheart MJ, Bender D, Slavich GM, Dahmouch L, Penedo F, DeGeest K, Mendez L, Lubaroff DM, Cole SW, Sood AK, Lutgendorf SK. Diurnal cortisol and survival in epithelial ovarian cancer. *Psychoneuroendocrinology* 2015; **53**: 256-267 [PMID: 25647344 DOI: 10.1016/j.psyneuen.2015.01.010]
 - 100 **Sephton SE**, Lush E, Dedert EA, Floyd AR, Rebholz WN, Dhabhar FS, Spiegel D, Salmon P. Diurnal cortisol rhythm as a predictor of lung cancer survival. *Brain Behav Immun* 2013; **30** Suppl: S163-S170 [PMID: 22884416 DOI: 10.1016/j.bbi.2012.07.019]
 - 101 **Moreno-Smith M**, Lutgendorf SK, Sood AK. Impact of stress on cancer metastasis. *Future Oncol* 2010; **6**: 1863-1881 [PMID: 21142861 DOI: 10.2217/fon.10.142]
 - 102 **Smith HR**. Depression in cancer patients: Pathogenesis, implications and treatment (Review). *Oncol Lett* 2015; **9**: 1509-1514 [PMID: 25788991 DOI: 10.3892/ol.2015.2944]
 - 103 **Li XM**, Xiao WH, Yang P, Zhao HX. Psychological distress and cancer pain: Results from a controlled cross-sectional survey in China. *Sci Rep* 2017; **7**: 39397 [PMID: 28074915 DOI: 10.1038/srep39397]
 - 104 **Rash JA**, Aguirre-Camacho A, Campbell TS. Oxytocin and pain: a systematic review and synthesis of findings. *Clin J Pain* 2014; **30**: 453-462 [PMID: 23887343 DOI: 10.1097/AJP.0b013e31829f57df]
 - 105 **Bos PA**, Montoya ER, Hermans EJ, Keyzers C, van Honk J. Oxytocin reduces neural activity in the pain circuitry when seeing pain in others. *Neuroimage* 2015; **113**: 217-224 [PMID: 25818690 DOI: 10.1016/j.neuroimage.2015.03.049]
 - 106 **Rash JA**, Campbell TS. Future directions for the investigation of intranasal oxytocin and pain: Comment on: Oxytocin nasal spray in fibromyalgic patients (Rheumatol Int. E-pub ahead of print. doi: 10.1007/s00296-014-2953-y). *Rheumatol Int* 2014; **34**: 1177-1178 [PMID: 24939559 DOI: 10.1007/s00296-014-3070-7]
 - 107 **Poisbeau P**, Grinevich V, Charlet A. Oxytocin Signaling in Pain: Cellular, Circuit, System, and Behavioral Levels. *Curr Top Behav Neurosci* 2018; **35**: 193-211 [PMID: 28942595 DOI: 10.1007/7854_2017_14]
 - 108 **Pouthaditis T**, Kearney SM, Levkovich T, Qi P, Varian BJ, Lakritz JR, Ibrahim YM, Chatzigiagos A, Alm EJ, Erdman SE. Microbial symbionts accelerate wound healing via the neuropeptide hormone oxytocin. *PLoS One* 2013; **8**: e78898 [PMID: 24205344 DOI: 10.1371/journal.pone.0078898]
 - 109 **Péqueux C**, Breton C, Hagelstein MT, Geenen V, Legros JJ. Oxytocin receptor pattern of expression in primary lung cancer and in normal human lung. *Lung Cancer* 2005; **50**: 177-188 [PMID: 16043261 DOI: 10.1016/j.lungcan.2005.05.027]
 - 110 **Thibonnier M**, Conarty DM, Preston JA, Plesnicher CL, Dweik RA, Erzurum SC. Human vascular endothelial cells express oxytocin receptors. *Endocrinology* 1999; **140**: 1301-1309 [PMID: 10067857 DOI: 10.1210/endo.140.3.6546]
 - 111 **Amrani Y**, Syed F, Huang C, Li K, Liu V, Jain D, Keslacy S, Sims MW, Baidouri H, Cooper PR, Zhao H, Siddiqui S, Brightling CE, Griswold D, Li L, Panettieri RA Jr. Expression and activation of the oxytocin receptor in airway smooth muscle cells: Regulation by TNFalpha and IL-13. *Respir Res* 2010; **11**: 104 [PMID: 20670427 DOI: 10.1186/1465-9921-11-104]
 - 112 **Amico JA**, Finn FM, Haldar J. Oxytocin and vasopressin are present in human and rat pancreas. *Am J Med Sci* 1988; **296**: 303-307 [PMID: 3195625 DOI: 10.1097/00000441-198811000-00003]
 - 113 **Szeto A**, Sun-Suslow N, Mendez AJ, Hernandez RI, Wagner KV, McCabe PM. Regulation of the macrophage oxytocin receptor in response to inflammation. *Am J Physiol Endocrinol Metab* 2017; **312**: E183-E189 [PMID: 28049625 DOI: 10.1152/ajpendo.00346.2016]
 - 114 **Di Benedetto A**, Sun L, Zamboni CG, Tamma R, Nico B, Calvano CD, Colaianni G, Ji Y, Mori G, Grano M, Lu P, Colucci S, Yuen T, New MI, Zallone A, Zaidi M. Osteoblast regulation via ligand-activated nuclear trafficking of the oxytocin receptor. *Proc Natl Acad Sci USA* 2014; **111**: 16502-16507 [PMID: 25378700 DOI: 10.1073/pnas.1419349111]

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Role of polymorphisms in genes that encode cytokines and *Helicobacter pylori* virulence factors in gastric carcinogenesis

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Abstract

The *Helicobacter pylori* (*H. pylori*) infection is a determinant factor in gastric cancer (GC) development. However, the infection outcomes are variable and depend on both host and bacterial characteristics. Some host cytokines such as interleukin (IL)-1 β , IL-1Ra, IL-8, IL-10 and tumor necrosis factor- α play important roles in the host immune system response to the pathogen, in the development of gastric mucosal lesions and in cell malignant transformation. Therefore, these host factors are crucial in neoplastic processes. Certain polymorphisms in genes that encode these cytokines have been associated with an increased risk of GC. On the other hand, various virulence factors found in distinct *H. pylori* bacterial strains, including cytotoxin-associated antigen A, vacuolating cytotoxin, duodenal ulcer promoting gene A protein, outer inflammatory protein and blood group antigen binding adhesin, have been associated with the pathogenesis of different gastric diseases. The virulent factors mentioned above allow the successful infection by the bacterium and play crucial roles in gastric mucosa lesions, including malignant transformation. Moreover, the role of host polymorphisms and bacterial virulence factors in gastric carcinogenesis seems to vary among different countries and populations. The identification of host and bacterium factors that are associated with an increased risk of GC development may be useful in determining the prognosis of infection in patients, what could help in clinical decision-making and in providing of an optimized clinical approach.

Key words: *Helicobacter pylori*; Virulence factors; Cytokines; Gene polymorphisms; Gastric cancer

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Core tip: Various polymorphisms in host genes that encode cytokines and *Helicobacter pylori* virulence factors have been associated with different tendencies of gastric diseases development. Several reviews have been written on the role of host and bacterial isolated factors in gastric carcinogenesis. However, only a small amount of reviews unites the important characteristics of both bacterium and host in carcinogenesis. General overviews about polymorphisms in genes that encode cytokines are also scarce. We aimed to join the main polymorphisms in genes that encode cytokines and bacterial virulent factors related to gastric carcinogenesis and to provide a broad overview about these themes.

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INTRODUCTION

Helicobacter pylori (*H. pylori*) is a gram negative bacterium, which inhabits the gastric epithelial tissue of most people in the world^[1], and it is considered a determinant factor in the initiation of gastric carcinogenesis^[2]. Gastric cancer (GC) is one of the four most prevalent neoplasms and the second biggest cause of deaths in consequence of cancer worldwide^[3]. Despite the importance of *H. pylori* in gastric carcinogenesis, the development of GC only occurs in a minority of infected people, demonstrating that the infection outcomes are variable. It is believed that multifactorial precancerous processes associated with both host mucosal inflammatory response and pathogen characteristics are determinant in the severity of the disease^[4].

The host immune system response plays a crucial role in the outcomes of *H. pylori* infection. Polymorphisms in genes that encode cytokines have been reported and associated with the severity of gastric mucosa inflammation and GC development. Some of these determinant variations are present in genes that encode cytokines such as interleukin (IL)-1 β , IL-1Ra, IL-8, IL-10 and tumor necrosis factor (TNF)- α ^[5-13]. These polymorphisms are important aspects in understanding gastric carcinogenesis, since chronic inflammation induced by the bacterium is critical in the emergence and evolution of GC precursor lesions (Figure 1)^[14].

On the other hand, the virulence factors of *H. pylori* are determinant in the interaction with host cells. Cytotoxin associated antigen A (CagA), vacuolating cytotoxin (VacA), duodenal ulcer promoting gene A protein (DupA), outer inflammatory protein (OipA) and blood group antigen binding adhesin (BabA) are

some virulent factors that seem to be associated with different risks of GC development^[15]. Furthermore, *H. pylori* with EPIYA-D or more than one EPIYA-C segment in its *CagA* gene have been associated with a higher risk of gastric carcinogenesis^[16-20].

POLYMORPHISMS IN GENES THAT ENCODE CYTOKINES AND GASTRIC CARCINOGENESIS

Gastric carcinogenesis is a process in which chronic inflammatory status plays a crucial role. The increase of inflammatory cytokine levels, due to *H. pylori* infection, seems to be determinant in the initiation and progression of GC^[12]. The intensity of the expression of cytokines can be modified by functional polymorphisms in the promoter regions of the genes, which has the potential to alter the affinity of transcription factors, interfering in the expression levels of the messenger ribonucleic acid (mRNA) of specific inflammatory mediators related to the susceptibility of GC initiation^[21].

IL-1

IL-1 is a family of cytokines that possesses 11 described members, among which IL-1 β and IL-1 receptor antagonist (IL-1Ra), combined with *H. pylori* infection, seem to be key factors in GC development^[22-24]. Signaling through the IL-1 receptor is a necessary event for the beginning and sustenance of various responses of the immune system^[25].

The promoter regions of *IL1B* and *IL1RN* genes, which encode IL-1 β and IL-1Ra respectively, have SNPs that modify the expression of the genes and affect the inflammatory response^[26]. These SNPs increase the IL-1 β /IL-1Ra ratio, which unleashes processes that result in gastric hypochlorhydria, favoring GC development^[15,27].

IL-1 β is an important cytokine for host-response to pathogens; however, this mediator can exacerbate damage during chronic diseases^[28]. High levels of IL-1 β in *H. pylori* infections lead to gastrin overexpression, increased gastric inflammation, hypochlorhydria, and gastric atrophy^[29]. Moreover, IL-1 β might promote neoplastic growth^[30]. The *IL1B* gene can be composed by three different SNPs: C-T base transition at IL-1B-511 (rs16944), T-C base transition at IL-1B-31 (rs1143627) and IL-1B-3954 (rs1143634), and all of them are strongly associated with increased production of proinflammatory cytokines, hypochlorhydria and increased GC risk, mainly intestinal type, among Caucasians, but not among Asians or Hispanics^[31-34].

IL-1Ra inhibits IL-1 α and IL-1 β by means of binding to IL-1 receptors. *IL1RN* possesses a changeable number of tandem repeats in intron 2, forming long alleles (IL1RN1) with 3-6 repeats or a short allele (IL1RN2) with 2 repeats^[35]. The IL1RN2 allele is associated with severe gastric lesions and higher risk for GC, besides raised IL-1 β expression in Caucasians^[33-36].

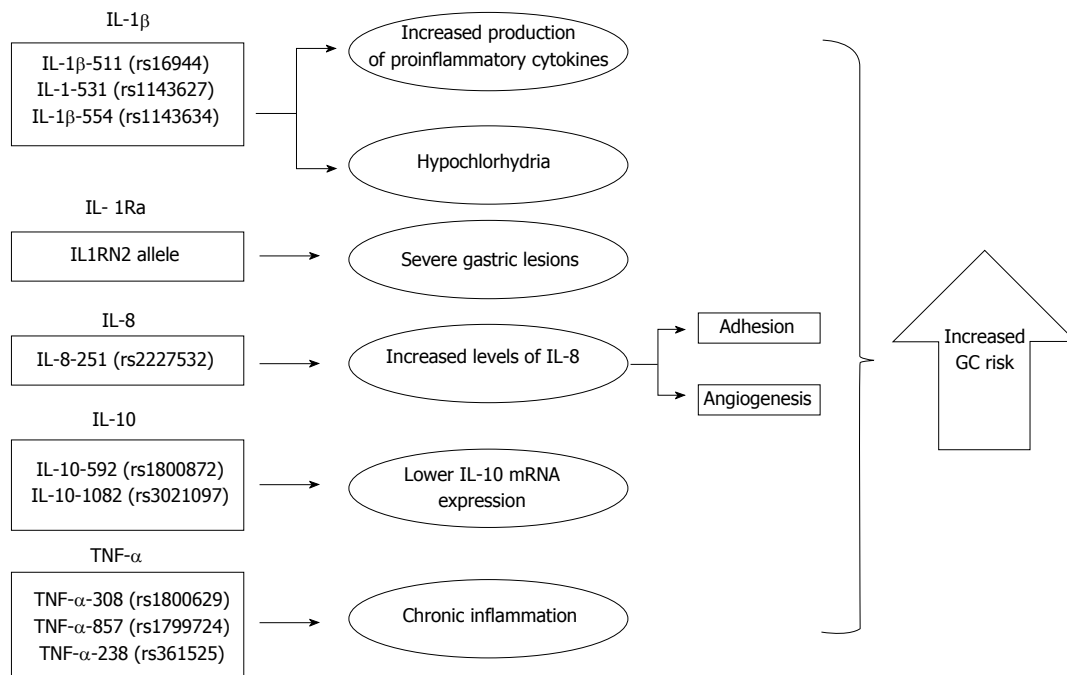


Figure 1 Potential functions of the host genetic polymorphisms in gastric carcinogenesis. IL: Interleukin; GC: Gastric cancer; TNF: Tumor necrosis factor.

IL-8

IL-8 is a potent cytokine that induces the directed migration of cells to inflammatory sites, acting as a chemoattractant^[37]. IL-8 secretion can be increased by different stimuli, such as live bacteria (including *H. pylori*) and lipopolysaccharides (LPS), besides others inflammatory cytokines, including IL-1 and TNF^[38]. The association of IL-8 with angiogenesis, adhesion and tumorigenesis have been related^[39,40].

The gene *CXCL8*, which encodes IL-8, is located on 4q12-21 chromosome and possesses four exons and three introns^[41]. An A/T SNP in the -251 position of this gene (*rs2227532*) has been associated with the development of various inflammatory diseases and cancer, including GC in Asians, but not in Europeans^[42,43]. Furthermore, the IL-8-251 A allele was related to increased levels of IL-8^[41].

IL-10

In opposition to the cytokines mentioned above, IL-10 is an anti-inflammatory cytokine, and it is involved in the cytotoxic response of inflammation and in cell downregulation. Moreover, this mediator prevents the production of pro-inflammatory cytokines, including TNF- α and IL-8^[44]. Some studies have demonstrated that SNPs, particularly IL-10-592 (*rs1800872*) and IL-10-1082 (*rs3021097*) alleles, might modulate transcriptional activation and affect IL-10 production in vitro. These IL-10 polymorphisms are related to lower mRNA expression of this cytokine and it have been associated with GC development in Asians^[45-48].

TNF- α

TNF- α composes the TNF/TNFR cytokine superfamily

and it is involved in maintenance and homeostasis of the immune system and host defense^[49]. However, this cytokine is related to various pathologic processes, including autoimmunity, chronic inflammatory processes and malignant disease^[50]. According to studies, TNF- α signaling through TNFR1 (TNF- α receptor) is important for gastric tumor development^[51,52].

Some SNPs in the TNF- α gene are related to increased expression of this cytokine. Among these polymorphisms, TNF- α -857 C/T (*rs1799724*), TNF- α -308 G/A (*rs1800629*) and TNF- α -238 G/A (*rs361525*) are the most studied ones. TNF- α -308 G/A was significantly associated with GC only in Caucasians, while TNF- α -857 and TNF- α -238 were related to an increased risk of gastric tumorigenesis in Asians, but not in Caucasians^[53-55].

H. PYLORI VIRULENCE FACTORS AND CARCINOGENESIS

The capacity of *H. pylori* bacteria to trigger a carcinogenic process is not limited to the intense immune response that they unleash, but it also depends on various bacterial factors that can start and modulate neoplastic processes^[56]. Different virulent factors found in distinct bacterial strains have been closely associated with the emergence of gastric carcinogenesis. However, genetic variations in genes that encode these virulence factors as well as geographic differences can influence the role of these proteins in GC emergence^[15].

CagA

CagA is encoded by the *cagA* gene, present in a DNA

segment containing 30 genes called *cag* pathogenicity island (*cag PAI*). Infections by strains containing CagA are more capable to induce carcinogenic processes, mainly those with EPIYA-D or more than one EPIYA-C segment^[57]. Various *cag PAI* genes are involved in the codification of elements of a pilus structure named type IV secretion system (TFSS), which has the function of transporting CagA from bacterium to the cytoplasm of the cells from gastric epithelium^[58].

After being injected into host cells by TFSS, CagA suffers tyrosine phosphorylation at a carboxi-terminal segment compound by distinct number of EPIYA (Glu-Pro-Ile-Tyr-Ala) regions. There are different EPIYA segments -A, B, C and D-, which contain distinct amino acids in their structure^[20]. EPIYA A and B segments are present in most CagA proteins and are followed either by 0-3 EPIYA-C segments in *H. pylori* strains from Occidental countries or by EPIYA-D segments in Eastern countries^[59].

Following EPIYA-C or EPIYA-D phosphorylation, an interaction between these segments and SHP-2 possessing SH2 domain occurs, unleashing SHP-2/mitogen-activated protein kinases (MAPK), ERK1, 2-JAK and STAT3 pathways^[20]. Cytotoxin associated antigen containing EPIYA-D or more than one EPIYA-C segment ties to SHP-2 more strongly, being more effective in the activation of the pathways mentioned above^[60]. This process, activated by CagA, leads to dysfunction of cell growth and of cell-to-cell contact inhibition, cell migration, epithelial cell elongation, and increase of epithelial cell turnover, increasing the propensity of acquirement of precancerous genetic changes by damaged cells^[61]. Furthermore, it was demonstrated that relatives of GC patients are more often infected by *H. pylori* strains with more than one EPIYA-C segment in CagA structure^[62]. Another study carried out by this same group, performed in a Brazilian population, showed that the host signal transducer and activator of transcription protein 3 (STAT3) rs7744166 polymorphism, as well as being infected by *H. pylori* with CagA containing more than one EPIYA-C segment, are independent predisposing factors for GC^[20].

VacA

VacA is another determinant virulence factor in *H. pylori* infection and in gastric carcinogenesis. Patients infected with VacA-positive *H. pylori* strains have a higher propensity for GC development when compared with patients colonized by VacA-negative strains, either in American or in Asian people^[63]. Particularly, individuals infected with *H. pylori* strains VacA s1, m1 and s1m1 had an increased risk for gastric carcinogenic unleash in Middle East, Africa and Latin America populations^[64]. The peptide mentioned above has only two functional domains in its structure. One of them, the p55-58 domain, has the function of binding to receptors of gastric epithelial cells. The other functional domain, p33-37, produces the cytotoxic effect^[65].

VacA is a 90 kDa exotoxin that is activated in a low pH environment^[66]. This toxin promotes the generation of numerous acidic vacuoles in gastric epithelial cell cytoplasm^[67]. In this process, VacA affects the structure and function of the membrane, the endoplasmic reticulum, the Golgi apparatus and the mitochondria, which can lead to cell death. Furthermore, vacuolating cytotoxin also plays an important role in the activation and suppression of the immune response^[68]. This peptide induces a powerful inhibition over T lymphocyte proliferation by means of an interaction with dendritic cells, which are reprogrammed to a tolerogenic genotype^[69]. The damage and the immunomodulation performed by this toxin contributes to the increase of gastric mucosa inflammation, ulceration and carcinogenesis in mammals^[68].

DupA

Unlike the other virulence factors mentioned in this article, DupA seems to be a protective condition for GC. The *dupA* gene is constituted by two homologue genes of *virB4*, *jhp0917* and *jhp091*, which constitute a continuous gene. The real function of the protein encoded by *dupA* is still obscure, however, its mechanisms seems to be related to the increase of the production of IL-8 in the gastric antrum, contributing to the development of gastritis that predominates in that gastric region, a process that leads to duodenal ulcer formation^[70]. DupA has been significantly associated with duodenal ulcer formation in Asian countries, but this relation was not observed in the Western population^[71]. Furthermore, DupA-positive *H. pylori* has been associated with eradication failure^[72].

OipA

OipA constitutes a group of peptides described as outer membrane proteins (OMPs), a *H. pylori* protein family composed of 32 components^[73]. OipA has been described as a better marker for severe clinical outcomes than CagA, since the infection by strains possessing OipA is an independent determinant risk factor of GC vs gastritis in Americans^[74,75]. OipA enhances IL-8 production and leads to an increased inflammation status of gastric epithelium. Moreover, it was observed that OipA could inhibit the maturation of dendritic cells in vitro, which might contribute to the immunomodulatory processes performed by *H. pylori*^[76].

BabA

babA is a gene that encodes an adhesin whom allows the specific binding to the b and H-1 Lewis antigens, which are expressed in the surface of the gastric mucosa cells^[77]. The adhesion of the *H. pylori* to the gastric epithelium mediated by blood group antigen binding adhesin (BabA) appears to play a critical function in the transference of bacterial virulence factors to the host cells. This process contributes to the development of tissue lesions, and a high correlation

of *babA*-positive strains of *H. pylori* with GC has been described^[78,79].

CONCLUSION

Despite the wide knowledge about host and *H. pylori* interaction developed since the discovery of its colonization in human stomach, many characteristics that contribute to the infection outcomes are still obscure. The understandings about host polymorphisms in genes that encode cytokines and bacterium virulence factors in GC development are important not only for the determination of patients' prognosis, but it is also a potential way for the development of new preventive and therapeutic strategies.

REFERENCES

- Alzahrani S, Lina TT, Gonzalez J, Pinchuk IV, Beswick EJ, Reyes VE. Effect of *Helicobacter pylori* on gastric epithelial cells. *World J Gastroenterol* 2014; **20**: 12767-12780 [PMID: 25278677 DOI: 10.3748/wjg.v20.i36.12767]
- Shimizu T, Marusawa H, Watanabe N, Chiba T. Molecular Pathogenesis of *Helicobacter pylori*-Related Gastric Cancer. *Gastroenterol Clin North Am* 2015; **44**: 625-638 [PMID: 26314672 DOI: 10.1016/j.gtc.2015.05.011]
- Ang TL, Fock KM. Clinical epidemiology of gastric cancer. *Singapore Med J* 2014; **55**: 621-628 [PMID: 25630323 DOI: 10.11622/smedj.2014174]
- Wang MY, Liu XF, Gao XZ. *Helicobacter pylori* virulence factors in development of gastric carcinoma. *Future Microbiol* 2015; **10**: 1505-1516 [PMID: 26346770 DOI: 10.2217/fmb.15.72]
- Gao Y, He Y, Ding J, Wu K, Hu B, Liu Y, Wu Y, Guo B, Shen Y, Landi D, Landi S, Zhou Y, Liu H. An insertion/deletion polymorphism at miRNA-122-binding site in the interleukin-1alpha 3' untranslated region confers risk for hepatocellular carcinoma. *Carcinogenesis* 2009; **30**: 2064-2069 [PMID: 19917630 DOI: 10.1093/carcin/bgp283]
- Kutikhin AG, Yuzhalin AE, Volkov AN, Zhivotovskiy AS, Brusina EB. Correlation between genetic polymorphisms within IL-1B and TLR4 genes and cancer risk in a Russian population: a case-control study. *Tumour Biol* 2014; **35**: 4821-4830 [PMID: 24446182 DOI: 10.1007/s13277-014-1633-6]
- Sun Z, Cui Y, Jin X, Pei J. Association between IL-4 -590C > T polymorphism and gastric cancer risk. *Tumour Biol* 2014; **35**: 1517-1521 [PMID: 24072495 DOI: 10.1007/s13277-013-1209-x]
- Zhang JZ, Liu CM, Peng HP, Zhang Y. Association of genetic variations in IL-6/IL-6R pathway genes with gastric cancer risk in a Chinese population. *Gene* 2017; **623**: 1-4 [PMID: 28442395 DOI: 10.1016/j.gene.2017.04.038]
- Cui X, Huang Q, Li X, Liu F, Wang D, Yan D, Wang B, Yang C, Mi J, Tian G. Relationship Between Interleukin-10 Gene C-819T Polymorphism and Gastric Cancer Risk: Insights From a Meta-Analysis. *Med Sci Monit* 2016; **22**: 2839-2845 [PMID: 27516059]
- Qi WT, Gao JL, Zhang SS. Role of IL-17 gene polymorphisms in the susceptibility to gastric cancer. *Genet Mol Res* 2015; **14**: 13364-13369 [PMID: 26535650 DOI: 10.4238/2015.October.26.33]
- Cho YA, Kim J. Association of IL4, IL13, and IL4R polymorphisms with gastrointestinal cancer risk: A meta-analysis. *J Epidemiol* 2017; **27**: 215-220 [PMID: 28142034 DOI: 10.1016/j.je.2016.06.002]
- de Oliveira JG, Rossi AF, Nizato DM, Cadamuro AC, Jorge YC, Valsechi MC, Venâncio LP, Rahal P, Pavarino EC, Goloni-Bertollo EM, Silva AE. Influence of functional polymorphisms in TNF- α , IL-8, and IL-10 cytokine genes on mRNA expression levels and risk of gastric cancer. *Tumour Biol* 2015; **36**: 9159-9170 [PMID: 26088449 DOI: 10.1007/s13277-015-3593-x]
- Bockerstett KA, DiPaolo RJ. Regulation of Gastric Carcinogenesis by Inflammatory Cytokines. *Cell Mol Gastroenterol Hepatol* 2017; **4**: 47-53 [PMID: 28560288 DOI: 10.1016/j.jcmgh.2017.03.005]
- Wang F, Meng W, Wang B, Qiao L. *Helicobacter pylori*-induced gastric inflammation and gastric cancer. *Cancer Lett* 2014; **345**: 196-202 [PMID: 23981572 DOI: 10.1016/j.canlet.2013.08.016]
- Rocha GA, Guerra JB, Rocha AM, Saraiva IE, da Silva DA, de Oliveira CA, Queiroz DM. IL1RN polymorphic gene and *cagA*-positive status independently increase the risk of noncardia gastric carcinoma. *Int J Cancer* 2005; **115**: 678-683 [PMID: 15704154 DOI: 10.1002/ijc.20935]
- Zhang J, Wu J, Peng X, Song J, Wang J, Dong W. Associations between STAT3 rs744166 polymorphisms and susceptibility to ulcerative colitis and Crohn's disease: a meta-analysis. *PLoS One* 2014; **9**: e109625 [PMID: 25286337 DOI: 10.1371/journal.pone.0109625]
- Blaser MJ, Perez-Perez GI, Kleanthous H, Cover TL, Peek RM, Chyou PH, Stemmermann GN, Nomura A. Infection with *Helicobacter pylori* strains possessing *cagA* is associated with an increased risk of developing adenocarcinoma of the stomach. *Cancer Res* 1995; **55**: 2111-2115 [PMID: 7743510]
- Basso D, Zambon CF, Letley DP, Stranges A, Marchetti A, Rhead JL, Schiavon S, Guariso G, Ceroti M, Nitti D, Rugge M, Plebani M, Atherton JC. Clinical relevance of *Helicobacter pylori* *cagA* and *vacA* gene polymorphisms. *Gastroenterology* 2008; **135**: 91-99 [PMID: 18474244 DOI: 10.1053/j.gastro.2008.03.041]
- Naito M, Yamazaki T, Tsutsumi R, Higashi H, Onoe K, Yamazaki S, Azuma T, Hatakeyama M. Influence of EPIYA-repeat polymorphism on the phosphorylation-dependent biological activity of *Helicobacter pylori* CagA. *Gastroenterology* 2006; **130**: 1181-1190 [PMID: 16618412 DOI: 10.1053/j.gastro.2005.12.038]
- Batista SA, Rocha GA, Rocha AM, Saraiva IE, Cabral MM, Oliveira RC, Queiroz DM. Higher number of *Helicobacter pylori* CagA EPIYA C phosphorylation sites increases the risk of gastric cancer, but not duodenal ulcer. *BMC Microbiol* 2011; **11**: 61 [PMID: 21435255 DOI: 10.1186/1471-2180-11-61]
- Palomo J, Dietrich D, Martin P, Palmer G, Gabay C. The interleukin (IL)-1 cytokine family--Balance between agonists and antagonists in inflammatory diseases. *Cytokine* 2015; **76**: 25-37 [PMID: 26185894 DOI: 10.1016/j.cyto.2015.06.017]
- Oelmann E, Stein H, Berdel WE, Herbst H. Expression of Interleukin-1 and Interleukin-1 Receptors Type 1 and Type 2 in Hodgkin Lymphoma. *PLoS One* 2015; **10**: e0138747 [PMID: 26406983 DOI: 10.1371/journal.pone.0138747]
- Sakamoto K, Hikiba Y, Nakagawa H, Hayakawa Y, Yanai A, Akanuma M, Ogura K, Hirata Y, Kaestner KH, Omata M, Maeda S. Inhibitor of kappaB kinase beta regulates gastric carcinogenesis via interleukin-1alpha expression. *Gastroenterology* 2010; **139**: 226-238.e6 [PMID: 20347815 DOI: 10.1053/j.gastro.2010.03.047]
- Ma J, Sawai H, Matsuo Y, Ochi N, Yasuda A, Takahashi H, Wakasugi T, Funahashi H, Sato M, Okada Y, Takeyama H, Manabe T. Interleukin-1alpha enhances angiogenesis and is associated with liver metastatic potential in human gastric cancer cell lines. *J Surg Res* 2008; **148**: 197-204 [PMID: 18395750 DOI: 10.1016/j.jss.2007.08.014]
- Ruzzo A, Graziano F, Pizzagalli F, Santini D, Battistelli V, Panunzi S, Canestrari E, Catalano V, Humar B, Ficarella R, Bearzi I, Cascinu S, Naldi N, Testa E, Magnani M. Interleukin 1B gene (IL-1B) and interleukin 1 receptor antagonist gene (IL-1RN) polymorphisms in *Helicobacter pylori*-negative gastric cancer of intestinal and diffuse histotype. *Ann Oncol* 2005; **16**: 887-892 [PMID: 15851404 DOI: 10.1093/annonc/mdi184]
- Raza Y, Khan A, Khan AI, Khan S, Akhter S, Mubarak M, Ahmed A, Kazmi SU. Combination of Interleukin 1 Polymorphism and *Helicobacter pylori* Infection: an Increased Risk of Gastric Cancer in Pakistani Population. *Pathol Oncol Res* 2017; **23**: 873-880 [PMID: 28110439 DOI: 10.1007/s12253-017-0191-9]
- Hwang IR, Kodama T, Kikuchi S, Sakai K, Peterson LE, Graham DY, Yamaoka Y. Effect of interleukin 1 polymorphisms

- on gastric mucosal interleukin 1beta production in *Helicobacter pylori* infection. *Gastroenterology* 2002; **123**: 1793-1803 [PMID: 12454835 DOI: 10.1053/gast.2002.37043]
- 28 **Lopez-Castejon G**, Brough D. Understanding the mechanism of IL-1 β secretion. *Cytokine Growth Factor Rev* 2011; **22**: 189-195 [PMID: 22019906 DOI: 10.1016/j.cytogfr.2011.10.001]
 - 29 **Huang FY**, Chan AO, Lo RC, Rashid A, Wong DK, Cho CH, Lai CL, Yuen MF. Characterization of interleukin-1 β in *Helicobacter pylori*-induced gastric inflammation and DNA methylation in interleukin-1 receptor type 1 knockout (IL-1R1^{-/-}) mice. *Eur J Cancer* 2013; **49**: 2760-2770 [PMID: 23664095 DOI: 10.1016/j.ejca.2013.03.031]
 - 30 **Furuta T**, El-Omar EM, Xiao F, Shirai N, Takashima M, Sugimura H. Interleukin 1beta polymorphisms increase risk of hypochlorhydria and atrophic gastritis and reduce risk of duodenal ulcer recurrence in Japan. *Gastroenterology* 2002; **123**: 92-105 [PMID: 12105837 DOI: 10.1053/gast.2002.34156]
 - 31 **Wang P**, Xia HH, Zhang JY, Dai LP, Xu XQ, Wang KJ. Association of interleukin-1 gene polymorphisms with gastric cancer: a meta-analysis. *Int J Cancer* 2007; **120**: 552-562 [PMID: 17096351 DOI: 10.1002/ijc.22353]
 - 32 **Chen A**, Li CN, Hsu PI, Lai KH, Tseng HH, Hsu PN, Lo GH, Lo CC, Lin CK, Hwang IR, Yamaoka Y, Chen HC. Risks of interleukin-1 genetic polymorphisms and *Helicobacter pylori* infection in the development of gastric cancer. *Aliment Pharmacol Ther* 2004; **20**: 203-211 [PMID: 15233701 DOI: 10.1111/j.1365-2036.2004.01826.x]
 - 33 **Chung HW**, Lim JB. Role of the tumor microenvironment in the pathogenesis of gastric carcinoma. *World J Gastroenterol* 2014; **20**: 1667-1680 [PMID: 24587646 DOI: 10.3748/wjg.v20.i7.1667]
 - 34 **Xue H**, Lin B, Ni P, Xu H, Huang G. Interleukin-1B and interleukin-1 RN polymorphisms and gastric carcinoma risk: a meta-analysis. *J Gastroenterol Hepatol* 2010; **25**: 1604-1617 [PMID: 20880168 DOI: 10.1111/j.1440-1746.2010.06428.x]
 - 35 **Chiurillo MA**. Role of gene polymorphisms in gastric cancer and its precursor lesions: current knowledge and perspectives in Latin American countries. *World J Gastroenterol* 2014; **20**: 4503-4515 [PMID: 24782603 DOI: 10.3748/wjg.v20.i16.4503]
 - 36 **Figueiredo C**, Machado JC, Pharoah P, Seruca R, Sousa S, Carvalho R, Capelinha AF, Quint W, Caldas C, van Doorn LJ, Carneiro F, Sobrinho-Simões M. *Helicobacter pylori* and interleukin 1 genotyping: an opportunity to identify high-risk individuals for gastric carcinoma. *J Natl Cancer Inst* 2002; **94**: 1680-1687 [PMID: 12441323 DOI: 10.1093/jnci/94.22.1680]
 - 37 **Sun Q**, Sun F, Wang B, Liu S, Niu W, Liu E, Peng C, Wang J, Gao H, Liang B, Niu Z, Zou X, Niu J. Interleukin-8 promotes cell migration through integrin $\alpha\beta 6$ upregulation in colorectal cancer. *Cancer Lett* 2014; **354**: 245-253 [PMID: 25150782 DOI: 10.1016/j.canlet.2014.08.021]
 - 38 **Remick DG**. Interleukin-8. *Crit Care Med* 2005; **33**: S466-S467 [PMID: 16340423 DOI: 10.1097/01.CCM.0000186783.34908.18]
 - 39 **Kim JH**, Frantz AM, Anderson KL, Graef AJ, Scott MC, Robinson S, Sharkey LC, O'Brien TD, Dickerson EB, Modiano JF. Interleukin-8 promotes canine hemangiosarcoma growth by regulating the tumor microenvironment. *Exp Cell Res* 2014; **323**: 155-164 [PMID: 24582862 DOI: 10.1016/j.yexcr.2014.02.020]
 - 40 **Ju D**, Sun D, Xiu L, Meng X, Zhang C, Wei P. Interleukin-8 is associated with adhesion, migration and invasion in human gastric cancer SCG-7901 cells. *Med Oncol* 2012; **29**: 91-99 [PMID: 21191670 DOI: 10.1007/s12032-010-9780-0]
 - 41 **Vairaktaris E**, Yapijakis C, Serefoglou Z, Derka S, Vassiliou S, Nkenke E, Vylliotis A, Wiltfang J, Avgoustidis D, Critselis E, Neukam FW, Patsouris E. The interleukin-8 (-251A/T) polymorphism is associated with increased risk for oral squamous cell carcinoma. *Eur J Surg Oncol* 2007; **33**: 504-507 [PMID: 17174061 DOI: 10.1016/j.ejso.2006.11.002]
 - 42 **Ohyauchi M**, Imatani A, Yonechi M, Asano N, Miura A, Iijima K, Koike T, Sekine H, Ohara S, Shimosegawa T. The polymorphism interleukin 8 -251 A/T influences the susceptibility of *Helicobacter pylori* related gastric diseases in the Japanese population. *Gut* 2005; **54**: 330-335 [PMID: 15710978 DOI: 10.1136/gut.2003.033050]
 - 43 **Zhang Y**, Zeng X, Lu H, Li Y, Ji H. Association between Interleukin-8-251A/T polymorphism and gastric cancer susceptibility: a meta-analysis based on 5286 cases and 8000 controls. *Int J Clin Exp Med* 2015; **8**: 22393-22402 [PMID: 26885219]
 - 44 **Maeda H**, Okabayashi T, Nishimori I, Sugimoto T, Namikawa T, Dabanaka K, Tsujii S, Onishi S, Kobayashi M, Hanazaki K. Clinicopathologic features of adenocarcinoma at the gastric cardia: is it different from distal cancer of the stomach? *J Am Coll Surg* 2008; **206**: 306-310 [PMID: 18222384 DOI: 10.1016/j.jamcollsurg.2007.06.306]
 - 45 **Qi M**, Liu DM, Pan LL, Lin YX. Interleukin-10 gene -592C>A polymorphism and susceptibility to gastric cancer. *Genet Mol Res* 2014; **13**: 8954-8961 [PMID: 25366786 DOI: 10.4238/2014.October.31.10]
 - 46 **Pan F**, Tian J, Pan YY, Zhang Y. Association of IL-10-1082 promoter polymorphism with susceptibility to gastric cancer: evidence from 22 case-control studies. *Mol Biol Rep* 2012; **39**: 7143-7154 [PMID: 22311038 DOI: 10.1007/s11033-012-1546-7]
 - 47 **Zhu Y**, Wang J, He Q, Zhang JQ. The association between interleukin-10-592 polymorphism and gastric cancer risk: a meta-analysis. *Med Oncol* 2011; **28**: 133-136 [PMID: 20087693 DOI: 10.1007/s12032-010-9417-3]
 - 48 **Yu T**, Lu Q, Ou XL, Cao DZ, Yu Q. Clinical study on gastric cancer susceptibility genes IL-10-1082 and TNF- α . *Genet Mol Res* 2014; **13**: 10909-10912 [PMID: 25526211 DOI: 10.4238/2014.December.19.12]
 - 49 **Croft M**, Benedict CA, Ware CF. Clinical targeting of the TNF and TNFR superfamilies. *Nat Rev Drug Discov* 2013; **12**: 147-168 [PMID: 23334208 DOI: 10.1038/nrd3930]
 - 50 **Balkwill F**. TNF-alpha in promotion and progression of cancer. *Cancer Metastasis Rev* 2006; **25**: 409-416 [PMID: 16951987 DOI: 10.1007/s10555-006-9005-3]
 - 51 **Oshima H**, Ishikawa T, Yoshida GJ, Naoi K, Maeda Y, Naka K, Ju X, Yamada Y, Minamoto T, Mukaida N, Saya H, Oshima M. TNF- α /TNFR1 signaling promotes gastric tumorigenesis through induction of Nox1 and Gna14 in tumor cells. *Oncogene* 2014; **33**: 3820-3829 [PMID: 23975421 DOI: 10.1038/nc.2013.356]
 - 52 **Oguma K**, Oshima H, Oshima M. Inflammation, tumor necrosis factor and Wnt promotion in gastric cancer development. *Future Oncol* 2010; **6**: 515-526 [PMID: 20373866 DOI: 10.2217/fon.10.13]
 - 53 **Yang JP**, Hyun MH, Yoon JM, Park MJ, Kim D, Park S. Association between TNF- α -308 G/A gene polymorphism and gastric cancer risk: a systematic review and meta-analysis. *Cytokine* 2014; **70**: 104-114 [PMID: 25125137 DOI: 10.1016/j.cyto.2014.07.005]
 - 54 **Gorouhi F**, Islami F, Bahrami H, Kamangar F. Tumor-necrosis factor- α polymorphisms and gastric cancer risk: a meta-analysis. *Br J Cancer* 2008; **98**: 1443-1451 [PMID: 18319718 DOI: 10.1038/sj.bjc.6604277]
 - 55 **Yu JY**, Li L, Ma H, Liu K, Cheng X, Li YL, Song XL. Tumor necrosis factor- α 238 G/A polymorphism and gastric cancer risk: a meta-analysis. *Tumour Biol* 2013; **34**: 3859-3863 [PMID: 23900678 DOI: 10.1007/s13277-013-0972-z]
 - 56 **Odenbreit S**, Püls J, Sedlmaier B, Gerland E, Fischer W, Haas R. Translocation of *Helicobacter pylori* CagA into gastric epithelial cells by type IV secretion. *Science* 2000; **287**: 1497-1500 [PMID: 10688800 DOI: 10.1126/science.287.5457.1497]
 - 57 **Fischer W**. Assembly and molecular mode of action of the *Helicobacter pylori* Cag type IV secretion apparatus. *FEBS J* 2011; **278**: 1203-1212 [PMID: 21352490 DOI: 10.1111/j.1742-4658.2011.08036.x]
 - 58 **Rocha GA**, Rocha AM, Gomes AD, Faria CL Jr, Melo FF, Batista SA, Fernandes VC, Almeida NB, Teixeira KN, Brito KS, Queiroz DM. STAT3 polymorphism and *Helicobacter pylori* CagA strains with higher number of EPIYA-C segments independently increase the risk of gastric cancer. *BMC Cancer* 2015; **15**: 528 [PMID: 26186918 DOI: 10.1186/s12885-015-1533-1]
 - 59 **Hatakeyama M**. Oncogenic mechanisms of the *Helicobacter*

- pylori CagA protein. *Nat Rev Cancer* 2004; **4**: 688-694 [PMID: 15343275 DOI: 10.1038/nrc1433]
- 60 **Lee YS**, Lee DY, Yu DY, Kim S, Lee YC. Helicobacter pylori induces cell migration and invasion through casein kinase 2 in gastric epithelial cells. *Helicobacter* 2014; **19**: 465-475 [PMID: 25052887 DOI: 10.1111/hel.12144]
 - 61 **Suzuki N**, Murata-Kamiya N, Yanagiya K, Suda W, Hattori M, Kanda H, Bingo A, Fujii Y, Maeda S, Koike K, Hatakeyama M. Mutual reinforcement of inflammation and carcinogenesis by the Helicobacter pylori CagA oncoprotein. *Sci Rep* 2015; **5**: 10024 [PMID: 25944120 DOI: 10.1038/srep10024]
 - 62 **Queiroz DM**, Silva CI, Goncalves MH, Braga-Neto MB, Fialho AB, Fialho AM, Rocha GA, Rocha AM, Batista SA, Guerrant RL, Lima AA, Braga LL. Higher frequency of cagA EPIYA-C phosphorylation sites in H. pylori strains from first-degree relatives of gastric cancer patients. *BMC Gastroenterol* 2012; **12**: 107 [PMID: 22891666 DOI: 10.1186/1471-230X-12-107]
 - 63 **Li Q**, Liu J, Gong Y, Yuan Y. Serum VacA antibody is associated with risks of peptic ulcer and gastric cancer: A meta-analysis. *Microb Pathog* 2016; **99**: 220-228 [PMID: 27568203 DOI: 10.1016/j.micpath.2016.08.030]
 - 64 **Matos JI**, de Sousa HA, Marcos-Pinto R, Dinis-Ribeiro M. Helicobacter pylori CagA and VacA genotypes and gastric phenotype: a meta-analysis. *Eur J Gastroenterol Hepatol* 2013; **25**: 1431-1441 [PMID: 23929249 DOI: 10.1097/MEG.0b013e328364b53e]
 - 65 **Liu C**, Wang YM, Li ZX, Zhang L, Ma JL, Zhou T, You WC, Pan KF. [Serological assessment of Helicobacter pylori-specific antibodies and their association with gastric lesions in a high-risk population]. *Zhonghua Zhongliu Zazhi* 2013; **35**: 547-551 [PMID: 24257311]
 - 66 **Hotchin NA**, Cover TL, Akhtar N. Cell vacuolation induced by the VacA cytotoxin of Helicobacter pylori is regulated by the Rac1 GTPase. *J Biol Chem* 2000; **275**: 14009-14012 [PMID: 10747859 DOI: 10.1074/jbc.C000153200]
 - 67 **Isomoto H**, Moss J, Hirayama T. Pleiotropic actions of Helicobacter pylori vacuolating cytotoxin, VacA. *Tohoku J Exp Med* 2010; **220**: 3-14 [PMID: 20046046 DOI: 10.1620/tjem.220.3]
 - 68 **Boquet P**, Ricci V. Intoxication strategy of Helicobacter pylori VacA toxin. *Trends Microbiol* 2012; **20**: 165-174 [PMID: 22364673 DOI: 10.1016/j.tim.2012.01.008]
 - 69 **Gebert B**, Fischer W, Weiss E, Hoffmann R, Haas R. Helicobacter pylori vacuolating cytotoxin inhibits T lymphocyte activation. *Science* 2003; **301**: 1099-1102 [PMID: 12934009 DOI: 10.1126/science.1086871]
 - 70 **Lu H**, Hsu PI, Graham DY, Yamaoka Y. Duodenal ulcer promoting gene of Helicobacter pylori. *Gastroenterology* 2005; **128**: 833-848 [PMID: 15825067 DOI: 10.1053/j.gastro.2005.01.009]
 - 71 **Yamaoka Y**. Roles of the plasticity regions of Helicobacter pylori in gastroduodenal pathogenesis. *J Med Microbiol* 2008; **57**: 545-553 [PMID: 18436586 DOI: 10.1099/jmm.0.2008/000570-0]
 - 72 **Shiota S**, Nguyen LT, Murakami K, Kuroda A, Mizukami K, Okimoto T, Kodama M, Fujioka T, Yamaoka Y. Association of helicobacter pylori dupA with the failure of primary eradication. *J Clin Gastroenterol* 2012; **46**: 297-301 [PMID: 22298090 DOI: 10.1097/MCG.0b013e318243201c]
 - 73 **Yamaoka Y**, Kwon DH, Graham DY. A M(r) 34,000 proinflammatory outer membrane protein (oipA) of Helicobacter pylori. *Proc Natl Acad Sci U S A* 2000; **97**: 7533-7538 [PMID: 10852959 DOI: 10.1073/pnas.130079797]
 - 74 **Graham DY**, Opekun AR, Osato MS, El-Zimaity HM, Lee CK, Yamaoka Y, Qureshi WA, Cadoz M, Monath TP. Challenge model for Helicobacter pylori infection in human volunteers. *Gut* 2004; **53**: 1235-1243 [PMID: 15306577 DOI: 10.1136/gut.2003.037499]
 - 75 **Liu J**, He C, Chen M, Wang Z, Xing C, Yuan Y. Association of presence/absence and on/off patterns of Helicobacter pylori oipA gene with peptic ulcer disease and gastric cancer risks: a meta-analysis. *BMC Infect Dis* 2013; **13**: 555 [PMID: 24256489 DOI: 10.1186/1471-2334-13-555]
 - 76 **Dabiri H**, Maleknejad P, Yamaoka Y, Feizabadi MM, Jafari F, Rezadehbashi M, Nakhjavani FA, Mirsalehian A, Zali MR. Distribution of Helicobacter pylori cagA, cagE, oipA and vacA in different major ethnic groups in Tehran, Iran. *J Gastroenterol Hepatol* 2009; **24**: 1380-1386 [PMID: 19702906 DOI: 10.1111/j.1440-1746.2009.05876.x]
 - 77 **Shahi H**, Reisi S, Sadeghiani M, Mahsa M, Bahreini R, Moghni M, damavandi M, fatollahi F, Shahverdi E, ramezani G, Shirzad H. Prevalence of cagA and babA2 genes in Helicobacter Pylori strains Isolated from Iranian gastrointestinal disorder patients and their gastritis classification. *J Biol Today's World* 2014; **3**: 256-260 [DOI: 10.15412/J.JBTW.01031201]
 - 78 **Rad R**, Gerhard M, Lang R, Schöninger M, Rösch T, Schepp W, Becker I, Wagner H, Prinz C. The Helicobacter pylori blood group antigen-binding adhesin facilitates bacterial colonization and augments a nonspecific immune response. *J Immunol* 2002; **168**: 3033-3041 [PMID: 11884476 DOI: 10.4049/jimmunol.168.6.3033]
 - 79 **Talebi Bezzmin Abadi A**, Taghvaei T, Mohabbati Mobarez A, Vaira G, Vaira D. High correlation of babA 2-positive strains of Helicobacter pylori with the presence of gastric cancer. *Intern Emerg Med* 2013; **8**: 497-501 [PMID: 21604199 DOI: 10.1007/s11739-011-0631-6]

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Resistance to FLT3 inhibitors in acute myeloid leukemia: Molecular mechanisms and resensitizing strategies

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Abstract

FMS-like tyrosine kinase 3 (FLT3) is classified as a type III receptor tyrosine kinase, which exerts a key role in regulation of normal hematopoiesis. FLT3 mutation is the most common genetic mutation in acute myeloid leukemia (AML) and represents an attractive therapeutic target. Targeted therapy with FLT3 inhibitors in AML shows modest promising results in current ongoing clinical trials suggesting the complexity of FLT3 targeting in therapeutics. Importantly, resistance to FLT3 inhibitors may explain the lack of overwhelming response and could obstruct the successful treatment for AML. Here, we summarize the molecular mechanisms of primary resistance and acquired resistance to FLT3 inhibitors and discuss the strategies to circumvent the emergency of drug resistance and to develop novel treatment intervention.

Key words: FMS-like tyrosine kinase 3; Tyrosine kinase domain; Internal tandem duplication; FLT3 inhibitor; Drug resistance; Acute myeloid leukemia; Combination therapy

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Core tip: FMS-like tyrosine kinase 3 (FLT3) mutations including internal tandem duplication (ITD) or point mutation in tyrosine kinase domain are common genetic

abnormalities in acute myeloid leukemia (AML), predicting dismal outcome. The Federal Drug Administration granted the use of Midostaurin (Novartis) in newly diagnosed FLT3-ITD positive AML in April 2017. A number of other FLT3 inhibitors are in different phases of clinical trials. However, emerging drug resistance poses a major challenge for clinicians to use FLT3 inhibitors. In this manuscript, we systematically reviewed mechanism of primary resistance and acquired resistance to FLT3 inhibitors. We then propose different strategies to overcome drug resistance and novel treatment options for FLT3-ITD positive AML.

Zhou J, Chng WJ. Resistance to FLT3 inhibitors in acute myeloid leukemia: Molecular mechanisms and resensitizing strategies. *World J Clin Oncol* 2018; 9(5): 90-97 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i5/90.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i5.90>

INTRODUCTION

Acute myeloid leukemia (AML) consists of a group of different disease characterized with diverse cellular morphologies and various genetic abnormalities^[1-5]. Many of these genetic lesions are of clinical importance because they not only implicate in the pathology of AML, but also have prognostic values^[6-8]. Mutation in in FMS-like tyrosine kinase 3 (FLT3) confers inferior response to chemotherapy and poor overall survival in AML patients^[9-11]. Since the discovery of FLT3 mutations in 1996^[12], intensive research effort has provided a better understanding of the molecular mechanism of normal and aberrant FLT3 signaling transduction pathways. Internal tandem duplications (ITDs) in the juxtamembrane domain and activating point mutations in the second tyrosine kinase domain (TKD) occur in near 30% and 10% of patients with AML respectively^[13-15].

FLT3 mutations constitutively activate PI3K-AKT, RAS-MEK-MAPK, and STAT5 pathways and result in uncontrolled cell proliferation and cell survival^[16-19]. On the other hand, FLT3 mutations suppress myeloid transcription factors PU.1, CCAAT/enhancer-binding protein α (C/EBP α), which result in blocking of myeloid differentiation^[20,21]. Thus, FLT3 mutations exert a key role in the pathology of AML, and have been validated as promising intervening targets^[22-25]. Currently, Midostaurin (PKC412, Novartis) has been granted by the Federal Drug Administration (FDA) in the use in newly diagnosed FLT3-ITD positive AML in combination with chemotherapy^[26]. Moreover, there are about a dozen of other FLT3 inhibitors in different phases of clinical development^[27]. Despite most FLT3 inhibitors display strong effectiveness in cell culture system, most of AML patients in trials haven't achieved durable response^[28-30]. Notably, AML patients inevitably don't respond to these drugs when they are administrated as single agent for a period. Scientists first observed this resistance

phenomenon in patients with chronic myeloid leukemia (CML) who received imatinib mesylate (Gleevec), the first small molecule kinase inhibitor targeting BCR-ABL fusion protein^[31].

Here we review published literature on preclinical and clinical findings and molecular mechanisms of primary resistance and acquired resistance to FLT3 inhibitors. We further discuss the strategies to circumvent the emergency of drug resistance and development of novel treatment intervention.

PRIMARY RESISTANCE TO FLT3

INHIBITORS

The identification of a number of de novo and secondary point mutations in the BCR-ABL kinase domain from imatinib-resistant patients promotes researchers to investigate variable sensitivity of FLT3 inhibitors between different activating point mutations in the kinase domain of FLT3.

Based on the mutations identified in AML cases, Grundler *et al.*^[32] employed site-directed mutagenesis method to create Asp835Tyr, Ile836del and Ile836Met + Arg (numbering is based on the human FLT3) into the cDNA of murine wild-type FLT3. These vectors were then transfected into murine Ba/F3 and 32Dcl3 cells, rendering them independent from growth factors. Tyrphostin AG1296 does not inhibit FLT3 Asp835Tyr (D835Y)-induced proliferation, inhibition of apoptosis, as well as the downstream signaling, and phosphorylation of STAT5. AG1296 is effective on the inhibition of signaling from FLT3 Ile836del (I836del), -ITD and to the less extent, from Ile836Met + Arg (I836M + R). Staurosporin derivative PKC412 is sensitive to all the 3 catalytic domain mutations, but less sensitivity to -ITD mutant. Indolinone compound SU5416 shows similar inhibition profile as PKC412. This study suggests that different inhibitors exert a divergent sensitivity toward different mutations in the FLT3 receptors.

A similar approach was used to introduce each FLT3 activation loop mutant, including D835Y, Asp835Ala (D835A), Asp835Glu (D835E), Asp835Gly (D835G), Asp835His (D835H), Asp835Asn (D835N), Asp835Val (D835V), and D835del into human FLT3 cDNA^[33]. Ba/F3 cells were transformed with each vector. These 8 activation loop mutations display variable sensitivity toward quinazoline-based inhibitor MLN518 with more than a 10-fold range. I836del is as sensitive as ITD with IC₅₀ 0.55 μ mol/L. The IC₅₀ of D835E, D835A, D835N, D835H ranges from 0.99 to 2.65 μ mol/L. D835del, D835V and D835Y confer relative resistance to MLN518 with much higher IC₅₀ up to greater than 10 μ mol/L.

This phenomenon could be explained by the assumption that the mutations in the amino acid sequence change the conformation of the catalytic domain of FLT3, resulting in a weakened affinity with FLT3 inhibitors^[32,33]. However, the structural analysis of these inhibitors in the context of various mutants is not available in these

papers. These findings are of great clinical interest. Patients enrolled in the FLT3 inhibitor trials potentially can be screened for all known activation loop mutations. In addition, sensitivity of specific inhibitor can potentially be evaluated *ex vivo* prior to clinical administration to avoid known primary resistant cases.

About 1% to 2% of newly diagnosed AML patients carry both ITD and TKD (FLT3-ITD-TKD) with worse outcome when compared with patients with either ITD or TKD mutation alone^[34-36]. Similarly, an *in vitro* study using Ba/F3 cells transfected with FLT3-ITD-TKD dual mutants, for example ITD-D835N and ITD-D835Y, can induce resistance toward not only FLT3 inhibitor SU5614, but also cytotoxic drug Daunorubicin^[37]. Molecular study reveals these dual mutants promote overactivation of STAT5 pathway, and result in upregulation of downstream target Bcl-xL and RAD51 and arrest in the G₂/M phase of the cell cycle^[37]. Overexpression of Bcl-2 is also detected in primary AML patient samples with FLT3-ITD-Y591 duplication, correlated to high levels of phosphorylated p53. However, whether this mutant induces resistance to FLT3 inhibitors has not been tested^[38].

Other possible mechanisms of primary resistance to TKIs have been investigated. P-glycoprotein (p-gp, also named multi-drug resistance 1, MDR1), a major membrane efflux pump, Primary AML blasts co-expressing p-gp and FLT3-ITD, are resistant to herbimycin A, a tyrosine kinase inhibitor, and AG1296, but not to PKC412^[39]. The difference could be due to the fact that PKC412 has dual inhibitory roles in FLT3 and protein kinase C (PKC), which can induce phosphorylation of p-gp, resulting in subversion of p-gp mediated MDR. However, other study shows no association between FLT3 mutations and high levels of MDR1 gene expression in AML patients^[40].

In contrast to earlier studies, Siendones *et al.*^[41] demonstrate that inhibition of FLT3-ITD activity does not necessarily block the phosphorylation of AKT, ERK and STAT5, which are the 3 major pathways activated by FLT3 mutations, in some primary AML cells. This could be one reason for the limited anti-tumor effect of FLT3 inhibitors used as monotherapy in clinical trials. In addition, a new "niche and leukemia stem cell" model was proposed to explain the limitation of single agent^[42]. If FLT3 - ITD is presented in CD34 + CD38 - CD123 + leukemia stem and progenitor cells (LSPC) from primary AML samples, they are more resistant to FLT3 inhibitor in culture under defined nice conditions (fibronectin, IL - 3, SCF, IL - 6 and Ang-). This result is consistent with an earlier finding that patients whose CD34+CD33- precursors harbor FLT3 - ITD have worse outcome than patients whose CD34 + CD33 + progenitors have FLT3 - ITD^[43]. These data indicate that FLT3 - ITD AML derived from the less mature progenitors may be associated with drug resistance.

ACQUIRED (SECONDARY) RESISTANCE TO FLT3 INHIBITORS

Pioneer researches in imatinib-resistant CML patients

revealed two different resistant mechanisms including increased copy number of BCR-ABL fusion and point mutations in its adenosine triphosphate (ATP) binding motif^[44]. These initial discoveries facilitate our understanding of acquired resistance to FLT3 inhibitors. As demonstrated by imatinib-resistant CML studies, over expression of a mutated FLT3 could also be a common mechanism for drug desensitization and leading to resistance. Weisberg and Boulton *et al.*^[45] first address this issue using a Ba / F3 - FLT3 - ITD resistant polyclonal subline developed by coculture with increasing concentration of PKC412 (up to 40 nmol/L) with the parental cell line over 2 mo. The protein level of FLT3-ITD is significantly increased in this resistant subline compared to the parental Ba / F3 - FLT3 - ITD, leading to desensitize PKC412. It is not clear that FLT3 - ITD protein over expression was regulated on transcriptional (gene amplification) or post-translational levels (increased protein stability). Also, there is no further study on whether this resistant subline harbors point mutation(s) in the TKD domain.

Other resistant lines, designated as Ba / F3 - ITD - R1 to R4 derived from the same parental Ba / F3 - FLT3 - ITD have been developed in the presence of escalatory dose of SU5614^[46]. The average IC₅₀ of these lines is 17 - fold higher than the parent line. Consistent with their resistant phenotypes, on the molecular level, the phosphorylation of MAPK and STAT5 in these sublines is not inhibited by higher dose up to 10 μmol/L SU5614, while 1 μmol/L of SU5614 effectively decreases activity of MAPK and STAT5 in the parental line. They are also completely resistant to AG1295, which is structurally similar to SU5614. But, Ba / F3 - ITD - R1 to R4 display a similar sensitivity to a structural unrelated FLT3 inhibitor PKC412, a general TKI, Genistein and a chemotherapeutic agent cytosine arabinoside (Ara-C) as the parent ITD cells. Both flow cytometric analysis and western blot demonstrate elevated amount of FLT3 receptor in the resistant Ba / F3 - ITD - R1 to R4 compared with the parent line. Sequence analysis of TKD domain identifies Y842H mutation in ITD-R1 and -R2 cells, and D835 mutation in ITD-R3 and -R4 cells. These data indicate that both FLT3 target desensitization and acquired mutations in the activation kinase domain can contribute to secondary resistance *in vitro*.

Using random PCR mutagenesis to introduce point mutations in ATP-binding pocket of the KD of MSCV-FLT3-ITD, Cools *et al.*^[47] identified 4 different point mutations (Ala627, Asn676, Phe691, or Gly697) in Ba/F3 cells that render resistance to PKC412, SU5614 or K-252a. The G697R mutation is the most resistant clone to all the three inhibitors tested. Accordingly, PKC412 fails to reduce phosphorylation of FLT3 up to 400 nmol/L in cells with G697R and they are cross-resistant to other structurally different FLT3 inhibitors (GTP-14546, AGL2043, D-64406, D-64476, TMPPP and DQPPC). Modeling crystal structure of FLT3 receptor in complex with PKC412 indicates that the amino acid Gly697 and Phe691 directly contact with PKC412 and substitution

Table 1 Summary of main studies on primary and acquired resistance to FMS-like tyrosine kinase 3 inhibitors

	Ref.	Disease model (method and material)	Mechanisms of resistance
Primary resistance	Grundler <i>et al</i> ^[32] , 2003	Site-directed mutagenesis, murine Ba/F3	TKD mutation, deletion or insertion
	Clark <i>et al</i> ^[33] , 2004	Site-directed mutagenesis, murine Ba/F3	TKD mutation
	Bagrintseva <i>et al</i> ^[37] , 2005	Site-directed mutagenesis, murine Ba/F3	ITD-TKD mutation, Bcl-xL overexpression
Acquired resistance	Weisberg <i>et al</i> ^[45] , 2002	Coculture with PKC412, murine Ba/F3-FLT3-ITD	FLT3 protein overexpression
	Bagrintseva <i>et al</i> ^[46] , 2004	Coculture with SU5614, murine Ba/F3-FLT3-ITD	ITD-TKD mutation, FLT3 protein overexpression
	Cools <i>et al</i> ^[47] , 2004	Random PCR mutagenesis, murine Ba/F3-FLT3-ITD	ITD-TKD mutation
	Heidel <i>et al</i> ^[48] , 2006	PKC412 clinical trial, relapsed AML with ITD	ITD-TKD (N676K) mutation
	Piloto <i>et al</i> ^[49] , 2007	Coculture with CEP-5214 and CEP-701, human MOLM-14, Hb1119 and SEM-K2	RTK amplification, N-Ras mutation
	Zhou <i>et al</i> ^[50] , 2009	Coculture with ABT-869, human MV4-11	Overactivation of STAT, overexpression of Survivin

FLT3: FMS-like tyrosine kinase 3; ITD: Internal tandem duplication; TKD: Tyrosine kinase domain; RTK: Receptor tyrosine kinase; STAT: Signal transducer and activator of transcription.

Gly697 with a larger amino acid will decrease its binding affinity due to possible steric clash with the FLT3 inhibitors. Importantly, mutation in Asn676 (N676K) has been reported in 1 of 6 patients with FLT3-ITD AML who relapsed after PKC412 treatment in a phase 2 clinical trial^[48]. The authors were able to rule out other common mechanisms of drug resistance including gene amplification, overexpression of FLT3 protein, drug metabolism, drug efflux, inhibition by serum proteins and major deficiency in apoptosis pathway. Although the identification of N676K is clinically significant to elucidate the mechanism of resistance and relapse, so far acquired point mutation of TKD has been reported only in a FLT3 inhibitor treated, and relapsed AML patient. In addition, transfection of FLT3-ITD-N676K in 32D cells confers resistance to PKC412^[48]. This finding is in consistent with the clinical observation that this mutant could be the sole reason of secondary resistance.

Most of the initial pre-clinical studies on mutations were conducted in murine cell lines transfected with FLT3 cDNA^[32,33,37,45-47]. We and others have further investigated the molecular mechanisms of acquired resistance to FLT3 inhibitors. Human leukemia cell lines with FLT3 mutations are valuable and relevant models for molecular biology and drug sensitivity studies. MV4-11 and MOLM-14 cell lines were derived from primary AML cells, while MV4-11 has two FLT3-ITD alleles; MOLM-14 harbors one mutant FLT3-ITD allele, while the other allele is wild-type (WT). Leukemia cell line Hb1119 and SEM-K2 were derived from primary ALL (acute lymphoblastic leukemia) cells. Hb1119 harbors FLT3-D836H, whereas SEM-K2 over expresses wild-type FLT3. Piloto *et al*^[49] reported that prolonged coculture of MOLM-14, Hb1119 and SEM-K2 cells with CEP-5214 and CEP-701 respectively led to the development of resistant lines including M14(R)5214, M14(R)701, Hb(R)5214, Hb(R)701, SEM(R)5214 and SEM(R)701. They are cross resistant to PKC412 and AG1295, a structurally related FLT3 inhibitor^[49]. Although TKIs can inhibit phosphorylation of FLT3 receptor in most of the resistant clones as demonstrated in this study, the downstream Akt and/or MAPK signaling remain

activated, thus providing cells sustained survival and proliferative signaling. Acquired N-Ras mutations have been identified in 2 [M14 (R) 5214 and M14 (R) 701] out of the 6 resistant lines. Transducing N-Ras-G12V mutation into MOLM-14 cells results in resistance to CEP-701^[49]. So, activation of parallel signaling pathway independent to FLT3 signaling may contribute to secondary resistance in some cases.

Through long-term culture of MV4-11 cell line with the FLT3 inhibitor, ABT-860, a FLT3 inhibitor-resistant line, MV4-11-R was generated^[50]. The IC₅₀ of ABT-869 for MV4-11-R line is 52 nmol/L vs 6 nmol/L for the parental MV4-11 cell line. Importantly, other structurally unrelated inhibitors including SU5416, AG1296 and a FLT3 inhibitor III from MERCK, were not effective to MV4-11-R line anymore, suggesting a cross resistant circumstance. Sequencing analysis showed normal sequence of FLT3-TKD in MV4-11-R cells. Western blot and FACS analysis excluded the overexpression of p-FLT3, FLT3 and three multidrug resistance related proteins (MDR, MRP1 and LRP) in this resistant line. But, overexpression of FLT3LG and Survivin was demonstrated at the both transcript and protein level. Down-regulation of suppressor of cytokine signaling (SOCS) proteins (negative regulators of STAT pathways) was also observed in the presence of overactivation of the STAT1, STAT3 and STAT5 pathways in this resistant line. In conclusion, our findings show that overactivation of STAT pathways and subsequently increased expressions of surviving genes are the main mechanism of resistance to FLT3 inhibitors. A total of 9 main studies regarding to primary and acquired resistance is summarized in Table 1.

STRATEGIES TO CONQUER RESISTANCE

The understanding of molecular mechanisms of primary and secondary resistance to FLT3 inhibitors (Figure 1) provides the foundation for establishing strategies to conquer or reduce resistance. Combination of FLT3 inhibitors with cytotoxic drugs or other small molecule inhibitors targeting different pathways has been extensively searched and tested *in vitro*, in murine

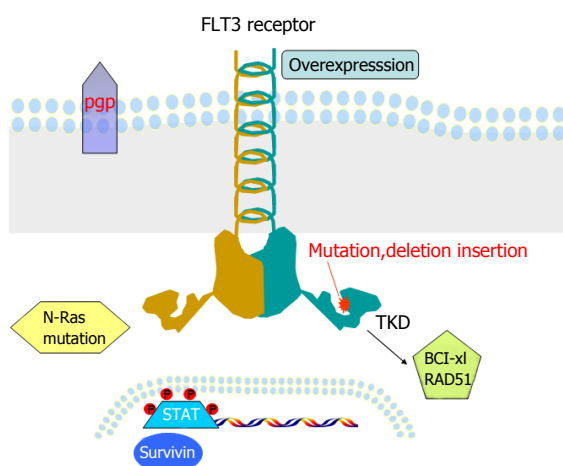


Figure 1 Schematic representation of various published mechanisms of resistance to FMS-like tyrosine kinase 3 inhibitors. This figure shows a number of models of resistance including desensitization of drug targets by FLT3 gene amplification or protein overexpression, decreased drug binding affinity by mutation, deletion or insertion in TKD, increased drug efflux by p-gp, activation of survival and proliferative pathways (molecules) such as RAS pathway, STAT pathway, anti-apoptotic Bcl-xL, Survivin and DNA repair molecule RAD51. FLT3: FMS-like tyrosine kinase 3; TKD: Tyrosine kinase domain; STAT: Signal transducer and activator of transcription.

xenograft models and some in clinical trials.

We and other investigators have demonstrated that combination of FLT3 inhibitors with conventional chemotherapy drugs, such as cytarabine and doxorubicin, can achieve synergistic effect^[30,51-54]. The optimal combination sequence should start with chemotherapy, followed by FLT3 inhibitors to maximize synergism and potential to reduce and/or overcome resistance^[30,53].

Combination of FLT3 inhibitors with a spectrum of small molecules inhibitors targeting downstream or independent signaling pathways have been evaluated in pre-clinical studies and showed early promises. Rapamycin, an mTOR inhibitor, sensitizes not only Imatinib-resistant BCR-ABL positive cells^[55] but also TKI-resistant Ba/FLT3 dual mutant (ITD and TKD) cell^[46,55]. Approaches targeting cellular apoptosis machinery also have been explored. BH3 mimetic ABT-737, a potent inhibitor of anti-apoptotic Bcl2, effectively neutralizes resistance to FLT3 inhibitors in primary AML blasts^[56]. The proapoptotic inhibitor LBW242, a Smac (one member of the inhibitor of apoptosis, IAP) mimetic, can overcome resistance to PKC412 when used in combination with PKC412^[57]. Combination of FLT3 inhibitor GTP14564 with a HSP90 inhibitor, 17-allylamino-17-demethoxygeldanamycin (17-AAG), produces synergism via STAT5 pathway^[58]. Concurrent treatment with histone deacetylase inhibitor (HDACi) LAQ824 and PCK412 can synergistically induced apoptosis in human cell line and primary AML samples with FLT3 mutations^[59]. We demonstrate that either treatment with IDR E804, an inhibitor of CDKs and the SRC-STAT pathway, or targeting Survivin by shRNA or a dominant-negative vector (survivin-T34A) sensitize

MV4-11-R to ABT-869 induce apoptosis^[50].

Other compounds such as bis(1*H*-indol-2-yl) methanone Cpd.98, Cpd.102 and Sorafenib (B-Raf inhibitor) also overcome resistance to FLT3 inhibitors^[60,61]. Downregulation of FLT3 expression by RNAi increases sensitivity to FLT3 inhibitor MLN518 in human AML cell lines, a potential approach to override resistance^[62]. The PIM family of serine/threonine kinases (PIM-1, -2 and -3) has been shown to be cytoprotective^[63]. Constitutively activated FLT3 signaling up-regulates the PIM-1 expression via STAT5 pathway, which results in phosphorylation of BAD protein (pSer112 and pSer136), exerting anti-apoptotic effect^[63,64]. PIM-2 also phosphorylates BAD at Ser-112, blocking BAD-inducing cell death^[65]. Silencing PIM-2 or PIM-1 sensitizes resistant cells to FLT3 inhibitors^[66]. IMC-EB10, an anti-FLT3 monoclonal antibody, is still effective in FLT3-TKI resistant clones, because it mediates antibody-dependent, cell-mediated cytotoxicity (ADCC) which is independent of the FLT3-ITD signaling pathway^[49].

CONCLUSION

Primary and secondary resistance to TKI therapy is challenging issue in modern anti-cancer warfare for various cancers including AML. At present time, monotherapy using FLT3 inhibitors showed limited benefit in relapsed AML clinical trials. We now began to better understand the molecular mechanisms of resistance in FLT3 targeting. Ongoing early phase clinical trials are important to further shed light on various potential mechanisms of resistance, and will eventually facilitate better strategies to prevent and overcome resistance. Sequel combination of FLT3 inhibitors with chemotherapy or other small molecule inhibitors targeting mTOR, HDAC, HSP90, STAT3, Bcl2, PIM family, IAPs (Survivin and Smac) and others are ongoing strategies. The correlative studies with these ongoing trials for identifying resistance mechanisms among trial patients, will help investigators in refining the design for next generation trial protocols. In addition, by determining "oncogenic signature" of each patient prior to treatment should guide the proper choice of most efficient combinations targeting the specific "oncogenic signature" individually.

REFERENCES

- 1 de Necochea-Campion R, Shouse GP, Zhou Q, Mirshahidi S, Chen CS. Aberrant splicing and drug resistance in AML. *J Hematol Oncol* 2016; **9**: 85 [PMID: 27613060 DOI: 10.1186/s13045-016-0315-9]
- 2 Hassan C, Afshinnikoo E, Li S, Wu S, Mason CE. Genetic and epigenetic heterogeneity and the impact on cancer relapse. *Exp Hematol* 2017; **54**: 26-30 [PMID: 28705639 DOI: 10.1016/j.exphem.2017.07.002]
- 3 Zhou J, Chooi JY, Ching YQ, Quah JY, Toh SH, Ng Y, Tan TZ, Chng WJ. NF-κB promotes the stem-like properties of leukemia cells by activation of LIN28B. *World J Stem Cells* 2018; **10**: 34-42 [PMID: 29707103 DOI: 10.4252/wjsc.v10.i4.34]
- 4 Zhu X, Ma Y, Liu D. Novel agents and regimens for acute myeloid leukemia: 2009 ASH annual meeting highlights. *J Hematol Oncol*

- 2010; **3**: 17 [PMID: 20416083 DOI: 10.1186/1756-8722-3-17]
- 5 **Shah K**, Curtin BF, Chu C, Hwang D, Flasar MH, von Rosenvinge E. Characteristics of *Clostridium difficile* infection in patients hospitalized with myelodysplastic syndrome or acute myelogenous leukemia. *World J Clin Oncol* 2017; **8**: 398-404 [PMID: 29067276 DOI: 10.5306/wjco.v8.i5.398]
- 6 **Medinger M**, Passweg JR. Acute myeloid leukaemia genomics. *Br J Haematol* 2017; **179**: 530-542 [PMID: 28653397 DOI: 10.1111/bjh.14823]
- 7 **Bullinger L**, Döhner K, Döhner H. Genomics of Acute Myeloid Leukemia Diagnosis and Pathways. *J Clin Oncol* 2017; **35**: 934-946 [PMID: 28297624 DOI: 10.1200/JCO.2016.71.2208]
- 8 **Zhou J**, Lu X, Tan TZ, Chng WJ. X-linked inhibitor of apoptosis inhibition sensitizes acute myeloid leukemia cell response to TRAIL and chemotherapy through potentiated induction of proapoptotic machinery. *Mol Oncol* 2018; **12**: 33-47 [PMID: 29063676 DOI: 10.1002/1878-0261.12146]
- 9 **Takahashi S**. Downstream molecular pathways of FLT3 in the pathogenesis of acute myeloid leukemia: biology and therapeutic implications. *J Hematol Oncol* 2011; **4**: 13 [PMID: 21453545 DOI: 10.1186/1756-8722-4-13]
- 10 **Gregory TK**, Wald D, Chen Y, Vermaat JM, Xiong Y, Tse W. Molecular prognostic markers for adult acute myeloid leukemia with normal cytogenetics. *J Hematol Oncol* 2009; **2**: 23 [PMID: 19490647 DOI: 10.1186/1756-8722-2-23]
- 11 **Zhou J**, Chan ZL, Bi C, Lu X, Chong PS, Chooi JY, Cheong LL, Liu SC, Ching YQ, Zhou Y, Osato M, Tan TZ, Ng CH, Ng SB, Wang S, Zeng Q, Chng WJ. LIN28B Activation by PRL-3 Promotes Leukemogenesis and a Stem Cell-like Transcriptional Program in AML. *Mol Cancer Res* 2017; **15**: 294-303 [PMID: 28011885 DOI: 10.1158/1541-7786.MCR-16-0275-T]
- 12 **Nakao M**, Yokota S, Iwai T, Kaneko H, Horiike S, Kashima K, Sonoda Y, Fujimoto T, Misawa S. Internal tandem duplication of the *flt3* gene found in acute myeloid leukemia. *Leukemia* 1996; **10**: 1911-1918 [PMID: 8946930]
- 13 **Kottaridis PD**, Gale RE, Frew ME, Harrison G, Langabeer SE, Belton AA, Walker H, Wheatley K, Bowen DT, Burnett AK, Goldstone AH, Linch DC. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United Kingdom Medical Research Council AML 10 and 12 trials. *Blood* 2001; **98**: 1752-1759 [PMID: 11535508 DOI: 10.1182/blood.V98.6.1752]
- 14 **Leung AY**, Man CH, Kwong YL. FLT3 inhibition: a moving and evolving target in acute myeloid leukaemia. *Leukemia* 2013; **27**: 260-268 [PMID: 22797419 DOI: 10.1038/leu.2012.195]
- 15 **Thiede C**, Steudel C, Mohr B, Schaich M, Schäkel U, Platzbecker U, Wermke M, Bornhäuser M, Ritter M, Neubauer A, Ehninger G, Illmer T. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood* 2002; **99**: 4326-4335 [PMID: 12036858 DOI: 10.1182/blood.V99.12.4326]
- 16 **Minami Y**, Yamamoto K, Kiyoi H, Ueda R, Saito H, Naoe T. Different antiapoptotic pathways between wild-type and mutated FLT3: insights into therapeutic targets in leukemia. *Blood* 2003; **102**: 2969-2975 [PMID: 12842996 DOI: 10.1182/blood-2002-12-3813]
- 17 **Spiekermann K**, Bagrintseva K, Schwab R, Schmiejka K, Hiddemann W. Overexpression and constitutive activation of FLT3 induces STAT5 activation in primary acute myeloid leukemia blast cells. *Clin Cancer Res* 2003; **9**: 2140-2150 [PMID: 12796379]
- 18 **Zhou J**, Goh BC, Albert DH, Chen CS. ABT-869, a promising multi-targeted tyrosine kinase inhibitor: from bench to bedside. *J Hematol Oncol* 2009; **2**: 33 [PMID: 19642998 DOI: 10.1186/1756-8722-2-33]
- 19 **Broekman F**, Giovannetti E, Peters GJ. Tyrosine kinase inhibitors: Multi-targeted or single-targeted? *World J Clin Oncol* 2011; **2**: 80-93 [PMID: 21603317 DOI: 10.5306/wjco.v2.i2.80]
- 20 **Choudhary C**, Schwäble J, Brandts C, Tickenbrock L, Sargin B, Kindler T, Fischer T, Berdel WE, Müller-Tidow C, Serve H. AML-associated *Flt3* kinase domain mutations show signal transduction differences compared with *Flt3* ITD mutations. *Blood* 2005; **106**: 265-273 [PMID: 15769897 DOI: 10.1182/blood-2004-07-2942]
- 21 **Zheng R**, Friedman AD, Levis M, Li L, Weir EG, Small D. Internal tandem duplication mutation of FLT3 blocks myeloid differentiation through suppression of C/EBP α expression. *Blood* 2004; **103**: 1883-1890 [PMID: 14592841 DOI: 10.1182/blood-2003-06-1978]
- 22 **Chen Y**, Pan Y, Guo Y, Zhao W, Ho WT, Wang J, Xu M, Yang FC, Zhao ZJ. Tyrosine kinase inhibitors targeting FLT3 in the treatment of acute myeloid leukemia. *Stem Cell Investig* 2017; **4**: 48 [PMID: 28607922 DOI: 10.21037/sci.2017.05.04]
- 23 **Park JE**, Yuen HF, Zhou JB, Al-Aidaros AQ, Guo K, Valk PJ, Zhang SD, Chng WJ, Hong CW, Mills K, Zeng Q. Oncogenic roles of PRL-3 in FLT3-ITD induced acute myeloid leukaemia. *EMBO Mol Med* 2013; **5**: 1351-1366 [PMID: 23929599 DOI: 10.1002/emmm.201202183]
- 24 **Sternberg DW**, Licht JD. Therapeutic intervention in leukemias that express the activated *fms*-like tyrosine kinase 3 (FLT3): opportunities and challenges. *Curr Opin Hematol* 2005; **12**: 7-13 [PMID: 15604885 DOI: 10.1097/01.moh.0000147891.06584.d7]
- 25 **Zhou J**, Bi C, Chng WJ, Cheong LL, Liu SC, Mahara S, Tay KG, Zeng Q, Li J, Guo K, Tan CP, Yu H, Albert DH, Chen CS. PRL-3, a metastasis associated tyrosine phosphatase, is involved in FLT3-ITD signaling and implicated in anti-AML therapy. *PLoS One* 2011; **6**: e19798 [PMID: 21589872 DOI: 10.1371/journal.pone.0019798]
- 26 **Wei AH**, Tiong IS. Midostaurin, enasidenib, CPX-351, gemtuzumab ozogamicin, and venetoclax bring new hope to AML. *Blood* 2017; **130**: 2469-2474 [PMID: 29051180 DOI: 10.1182/blood-2017-08-784066]
- 27 **Larrosa-Garcia M**, Baer MR. FLT3 Inhibitors in Acute Myeloid Leukemia: Current Status and Future Directions. *Mol Cancer Ther* 2017; **16**: 991-1001 [PMID: 28576946 DOI: 10.1158/1535-7163.MCT-16-0876]
- 28 **Lancet JE**. New agents: great expectations not realized. *Best Pract Res Clin Haematol* 2013; **26**: 269-274 [PMID: 24309529 DOI: 10.1016/j.beha.2013.10.007]
- 29 **Swords R**, Freeman C, Giles F. Targeting the FMS-like tyrosine kinase 3 in acute myeloid leukemia. *Leukemia* 2012; **26**: 2176-2185 [PMID: 22614177 DOI: 10.1038/leu.2012.114]
- 30 **Zhou J**, Pan M, Xie Z, Loh SL, Bi C, Tai YC, Lilly M, Lim YP, Han JH, Glaser KB, Albert DH, Davidsen SK, Chen CS. Synergistic antileukemic effects between ABT-869 and chemotherapy involve downregulation of cell cycle-regulated genes and c-Mos-mediated MAPK pathway. *Leukemia* 2008; **22**: 138-146 [PMID: 17943175 DOI: 10.1038/sj.leu.2404960]
- 31 **Melo JV**, Chuah C. Novel agents in CML therapy: tyrosine kinase inhibitors and beyond. *Hematology Am Soc Hematol Educ Program* 2008: 427-435 [PMID: 19074121 DOI: 10.1182/asheducation-2008.1.427]
- 32 **Grundler R**, Thiede C, Miething C, Steudel C, Peschel C, Duyster J. Sensitivity toward tyrosine kinase inhibitors varies between different activating mutations of the FLT3 receptor. *Blood* 2003; **102**: 646-651 [PMID: 12663439 DOI: 10.1182/blood-2002-11-3441]
- 33 **Clark JJ**, Cools J, Curley DP, Yu JC, Lokker NA, Giese NA, Gilliland DG. Variable sensitivity of FLT3 activation loop mutations to the small molecule tyrosine kinase inhibitor MLN518. *Blood* 2004; **104**: 2867-2872 [PMID: 15256420 DOI: 10.1182/blood-2003-12-4446]
- 34 **Chen W**, Jones D, Medeiros LJ, Luthra R, Lin P. Acute myeloid leukaemia with FLT3 gene mutations of both internal tandem duplication and point mutation type. *Br J Haematol* 2005; **130**: 726-728 [PMID: 16115128 DOI: 10.1111/j.1365-2141.2005.05666.x]
- 35 **Georgiou G**, Karali V, Zouvelou C, Kyriakou E, Dimou M, Chrisochou S, Greka P, Dufexis D, Vervesou E, Dimitriadou E, Efthymiou A, Petrikos L, Dima K, Lilakos K, Panayiotidis P. Serial determination of FLT3 mutations in myelodysplastic syndrome patients at diagnosis, follow up or acute myeloid leukaemia transformation: incidence and their prognostic significance. *Br J Haematol* 2006; **134**: 302-306 [PMID: 16787503 DOI: 10.1111/j.1365-2141.2006.06171.x]
- 36 **Moreno I**, Martín G, Bolufer P, Barragán E, Rueda E, Román

- J, Fernández P, León P, Mena A, Cervera J, Torres A, Sanz MA. Incidence and prognostic value of FLT3 internal tandem duplication and D835 mutations in acute myeloid leukemia. *Haematologica* 2003; **88**: 19-24 [PMID: 12551822]
- 37 **Bagrintseva K**, Geisenhof S, Kern R, Eichenlaub S, Reindl C, Ellwart JW, Hiddemann W, Spiekermann K. FLT3-ITD-TKD dual mutants associated with AML confer resistance to FLT3 PTK inhibitors and cytotoxic agents by overexpression of Bcl-x(L). *Blood* 2005; **105**: 3679-3685 [PMID: 15626738 DOI: 10.1182/blood-2004-06-2459]
- 38 **Irish JM**, Anensen N, Hovland R, Skavland J, Børresen-Dale AL, Bruserud O, Nolan GP, Gjertsen BT. Flt3 Y591 duplication and Bcl-2 overexpression are detected in acute myeloid leukemia cells with high levels of phosphorylated wild-type p53. *Blood* 2007; **109**: 2589-2596 [PMID: 17105820 DOI: 10.1182/blood-2006-02-004234]
- 39 **Hunter HM**, Pallis M, Seedhouse CH, Grundy M, Gray C, Russell NH. The expression of P-glycoprotein in AML cells with FLT3 internal tandem duplications is associated with reduced apoptosis in response to FLT3 inhibitors. *Br J Haematol* 2004; **127**: 26-33 [PMID: 15384974 DOI: 10.1111/j.1365-2141.2004.05145.x]
- 40 **Galimberti S**, Rossi A, Palumbo GA, Morabito F, Guerrini F, Vincelli I, Fazzi R, Santini V, Petrini M. FLT3 mutations do not influence MDR-1 gene expression in acute myeloid leukemia. *Anticancer Res* 2003; **23**: 3419-3426 [PMID: 12926083]
- 41 **Siendones E**, Barbarroja N, Torres LA, Buendía P, Velasco F, Dorado G, Torres A, López-Pedraza C. Inhibition of Flt3-activating mutations does not prevent constitutive activation of ERK/Akt/STAT pathways in some AML cells: a possible cause for the limited effectiveness of monotherapy with small-molecule inhibitors. *Hematol Oncol* 2007; **25**: 30-37 [PMID: 17128418 DOI: 10.1002/hon.805]
- 42 **Mony U**, Jawad M, Seedhouse C, Russell N, Pallis M. Resistance to FLT3 inhibition in an in vitro model of primary AML cells with a stem cell phenotype in a defined microenvironment. *Leukemia* 2008; **22**: 1395-1401 [PMID: 18509353 DOI: 10.1038/leu.2008.125]
- 43 **Pollard JA**, Alonzo TA, Gerbing RB, Woods WG, Lange BJ, Sweetser DA, Radich JP, Bernstein ID, Meshinchi S. FLT3 internal tandem duplication in CD34+/CD33- precursors predicts poor outcome in acute myeloid leukemia. *Blood* 2006; **108**: 2764-2769 [PMID: 16809615 DOI: 10.1182/blood-2006-04-012260]
- 44 **Gorre ME**, Mohammed M, Ellwood K, Hsu N, Paquette R, Rao PN, Sawyers CL. Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* 2001; **293**: 876-880 [PMID: 11423618 DOI: 10.1126/science.1062538]
- 45 **Weisberg E**, Boulton C, Kelly LM, Manley P, Fabbro D, Meyer T, Gilliland DG, Griffin JD. Inhibition of mutant FLT3 receptors in leukemia cells by the small molecule tyrosine kinase inhibitor PKC412. *Cancer Cell* 2002; **1**: 433-443 [PMID: 12124173 DOI: 10.1016/S1535-6108(02)00069-7]
- 46 **Bagrintseva K**, Schwab R, Kohl TM, Schnittger S, Eichenlaub S, Ellwart JW, Hiddemann W, Spiekermann K. Mutations in the tyrosine kinase domain of FLT3 define a new molecular mechanism of acquired drug resistance to PTK inhibitors in FLT3-ITD-transformed hematopoietic cells. *Blood* 2004; **103**: 2266-2275 [PMID: 14604974 DOI: 10.1182/blood-2003-05-1653]
- 47 **Cools J**, Mentens N, Furet P, Fabbro D, Clark JJ, Griffin JD, Marynen P, Gilliland DG. Prediction of resistance to small molecule FLT3 inhibitors: implications for molecularly targeted therapy of acute leukemia. *Cancer Res* 2004; **64**: 6385-6389 [PMID: 15374944 DOI: 10.1158/0008-5472.CAN-04-2148]
- 48 **Heidel F**, Solem FK, Breitenbuecher F, Lipka DB, Kasper S, Thiede MH, Brandts C, Serve H, Roesel J, Giles F, Feldman E, Ehninger G, Schiller GJ, Nimer S, Stone RM, Wang Y, Kindler T, Cohen PS, Huber C, Fischer T. Clinical resistance to the kinase inhibitor PKC412 in acute myeloid leukemia by mutation of Asn-676 in the FLT3 tyrosine kinase domain. *Blood* 2006; **107**: 293-300 [PMID: 16150941 DOI: 10.1182/blood-2005-06-2469]
- 49 **Piloto O**, Wright M, Brown P, Kim KT, Levis M, Small D. Prolonged exposure to FLT3 inhibitors leads to resistance via activation of parallel signaling pathways. *Blood* 2007; **109**: 1643-1652 [PMID: 17047150 DOI: 10.1182/blood-2006-05-023804]
- 50 **Zhou J**, Bi C, Janakakumara JV, Liu SC, Chng WJ, Tay KG, Poon LF, Xie Z, Palaniyandi S, Yu H, Glaser KB, Albert DH, Davidsen SK, Chen CS. Enhanced activation of STAT pathways and overexpression of survivin confer resistance to FLT3 inhibitors and could be therapeutic targets in AML. *Blood* 2009; **113**: 4052-4062 [PMID: 19144991 DOI: 10.1182/blood-2008-05-156422]
- 51 **Aleskog A**, Höglund M, Pettersson J, Hermansson M, Larsson R, Lindhagen E. In vitro activity of the flt3-inhibitor su5614 and standard cytotoxic agents in tumour cells from patients with wild type and mutated flt3 acute myeloid leukaemia. *Leuk Res* 2005; **29**: 1079-1081 [PMID: 16038735 DOI: 10.1016/j.leukres.2005.02.017]
- 52 **Furukawa Y**, Vu HA, Akutsu M, Odgerel T, Izumi T, Tsunoda S, Matsuo Y, Kirito K, Sato Y, Mano H, Kano Y. Divergent cytotoxic effects of PKC412 in combination with conventional antileukemic agents in FLT3 mutation-positive versus -negative leukemia cell lines. *Leukemia* 2007; **21**: 1005-1014 [PMID: 17330105 DOI: 10.1038/sj.leu.2404593]
- 53 **Levis M**, Pham R, Smith BD, Small D. In vitro studies of a FLT3 inhibitor combined with chemotherapy: sequence of administration is important to achieve synergistic cytotoxic effects. *Blood* 2004; **104**: 1145-1150 [PMID: 15126317 DOI: 10.1182/blood-2004-01-0388]
- 54 **Yee KW**, Schittenhelm M, O'Farrell AM, Town AR, McGreevey L, Bainbridge T, Cherrington JM, Heinrich MC. Synergistic effect of SU11248 with cytarabine or daunorubicin on FLT3 ITD-positive leukemic cells. *Blood* 2004; **104**: 4202-4209 [PMID: 15304385 DOI: 10.1182/blood-2003-10-3381]
- 55 **Mohi MG**, Boulton C, Gu TL, Sternberg DW, Neuberger D, Griffin JD, Gilliland DG, Neel BG. Combination of rapamycin and protein tyrosine kinase (PTK) inhibitors for the treatment of leukemias caused by oncogenic PTKs. *Proc Natl Acad Sci USA* 2004; **101**: 3130-3135 [PMID: 14976243 DOI: 10.1073/pnas.0400063101]
- 56 **Kohl TM**, Hellinger C, Ahmed F, Buske C, Hiddemann W, Bohlander SK, Spiekermann K. BH3 mimetic ABT-737 neutralizes resistance to FLT3 inhibitor treatment mediated by FLT3-independent expression of BCL2 in primary AML blasts. *Leukemia* 2007; **21**: 1763-1772 [PMID: 17554384 DOI: 10.1038/sj.leu.2404776]
- 57 **Weisberg E**, Kung AL, Wright RD, Moreno D, Catley L, Ray A, Zawal L, Tran M, Cools J, Gilliland G, Mitsiades C, McMillin DW, Jiang J, Hall-Meyers E, Griffin JD. Potentiation of antileukemic therapies by Smac mimetic, LBW242: effects on mutant FLT3-expressing cells. *Mol Cancer Ther* 2007; **6**: 1951-1961 [PMID: 17620426 DOI: 10.1158/1535-7163.MCT-06-0810]
- 58 **Yao Q**, Nishiuchi R, Kitamura T, Kersey JH. Human leukemias with mutated FLT3 kinase are synergistically sensitive to FLT3 and Hsp90 inhibitors: the key role of the STAT5 signal transduction pathway. *Leukemia* 2005; **19**: 1605-1612 [PMID: 16034464 DOI: 10.1038/sj.leu.2403881]
- 59 **Bali P**, George P, Cohen P, Tao J, Guo F, Sigua C, Vishvanath A, Scuto A, Annavarapu S, Fiskus W, Moscinski L, Atadja P, Bhalla K. Superior activity of the combination of histone deacetylase inhibitor LAQ824 and the FLT-3 kinase inhibitor PKC412 against human acute myelogenous leukemia cells with mutant FLT-3. *Clin Cancer Res* 2004; **10**: 4991-4997 [PMID: 15297399 DOI: 10.1158/1078-0432.CCR-04-0210]
- 60 **Heidel F**, Lipka DB, Mirea FK, Mahboobi S, Grundler R, Kancha RK, Duyster J, Naumann M, Huber C, Böhmer FD, Fischer T. Bis(1H-indol-2-yl)methanones are effective inhibitors of FLT3-ITD tyrosine kinase and partially overcome resistance to PKC412A in vitro. *Br J Haematol* 2009; **144**: 865-874 [PMID: 19183186 DOI: 10.1111/j.1365-2141.2008.07567.x]
- 61 **Lierman E**, Lahortiga I, Van Miegroet H, Mentens N, Marynen P, Cools J. The ability of sorafenib to inhibit oncogenic PDGFRbeta and FLT3 mutants and overcome resistance to other small molecule inhibitors. *Haematologica* 2007; **92**: 27-34 [PMID: 17229632 DOI: 10.3324/haematol.10692]
- 62 **Walters DK**, Stoffregen EP, Heinrich MC, Deininger MW, Druker BJ. RNAi-induced down-regulation of FLT3 expression in AML cell

- lines increases sensitivity to MLN518. *Blood* 2005; **105**: 2952-2954 [PMID: 15585651 DOI: 10.1182/blood-2004-07-2758]
- 63 **Kim KT**, Baird K, Ahn JY, Meltzer P, Lilly M, Levis M, Small D. Pim-1 is up-regulated by constitutively activated FLT3 and plays a role in FLT3-mediated cell survival. *Blood* 2005; **105**: 1759-1767 [PMID: 15498859 DOI: 10.1182/blood-2004-05-2006]
- 64 **Kim KT**, Levis M, Small D. Constitutively activated FLT3 phosphorylates BAD partially through pim-1. *Br J Haematol* 2006; **134**: 500-509 [PMID: 16869825 DOI: 10.1111/j.1365-2141.2006.06225.x]
- 65 **Yan B**, Zemskova M, Holder S, Chin V, Kraft A, Koskinen PJ, Lilly M. The PIM-2 kinase phosphorylates BAD on serine 112 and reverses BAD-induced cell death. *J Biol Chem* 2003; **278**: 45358-45367 [PMID: 12954615 DOI: 10.1074/jbc.M307933200]
- 66 **Adam M**, Pogacic V, Bendit M, Chappuis R, Nawijn MC, Duyster J, Fox CJ, Thompson CB, Cools J, Schwaller J. Targeting PIM kinases impairs survival of hematopoietic cells transformed by kinase inhibitor-sensitive and kinase inhibitor-resistant forms of Fms-like tyrosine kinase 3 and BCR/ABL. *Cancer Res* 2006; **66**: 3828-3835 [PMID: 16585210 DOI: 10.1158/0008-5472.CAN-05-2309]

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Basic Study

Tunable structure priors for Bayesian rule learning for knowledge integrated biomarker discovery

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Abstract**AIM**

To develop a framework to incorporate background domain knowledge into classification rule learning for knowledge discovery in biomedicine.

METHODS

Bayesian rule learning (BRL) is a rule-based classifier that uses a greedy best-first search over a space of Bayesian belief-networks (BN) to find the optimal BN to explain the input dataset, and then infers classification rules from this BN. BRL uses a Bayesian score to evaluate the quality of BNs. In this paper, we extended the Bayesian score to include informative structure priors, which encodes our prior domain knowledge about the dataset. We call this extension of BRL as BRL_p. The structure prior has a λ hyperparameter that allows the user to tune the degree of incorporation of the prior knowledge in the model learning process. We studied the effect of λ on model learning using a simulated dataset and a real-world lung cancer prognostic biomarker dataset, by measuring the degree of incorporation of our specified prior knowledge. We also monitored its effect on the model predictive performance. Finally, we compared BRL_p to other state-of-the-art classifiers commonly used in biomedicine.

RESULTS

We evaluated the degree of incorporation of prior knowledge into BRL_p, with simulated data by measuring the Graph Edit Distance between the true data-generating model and the model learned by BRL_p. We specified the true model using informative structure

priors. We observed that by increasing the value of λ we were able to increase the influence of the specified structure priors on model learning. A large value of λ of BRL_p caused it to return the true model. This also led to a gain in predictive performance measured by area under the receiver operator characteristic curve (AUC). We then obtained a publicly available real-world lung cancer prognostic biomarker dataset and specified a known biomarker from literature [the epidermal growth factor receptor (*EGFR*) gene]. We again observed that larger values of λ led to an increased incorporation of *EGFR* into the final BRL_p model. This relevant background knowledge also led to a gain in AUC.

CONCLUSION

BRL_p enables tunable structure priors to be incorporated during Bayesian classification rule learning that integrates data and knowledge as demonstrated using lung cancer biomarker data.

Key words: Supervised machine learning; Rule-based models; Bayesian methods; Background knowledge; Informative priors; Biomarker discovery

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Core tip: Bayesian rule learning is a unique rule learning algorithm that infers rule models from searched Bayesian networks. We extended it to allow the incorporation of prior domain knowledge using a mathematically robust Bayesian framework with structure priors. The hyperparameter of the structure priors enables the user to control the influence of their specified prior knowledge. This opens up many possibilities including incorporating uncertain knowledge that can interact with data accordingly during inference.

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INTRODUCTION

Knowledge discovery from databases (KDD) is the non-trivial extraction of valid novel, potentially useful, and understandable patterns from the dataset^[1]. Data mining is the computational process of the extraction of these patterns. In biomedicine, data mining is extensively applied for knowledge discovery^[2]. The recent advances in biomedical research, triggering an explosion of data, have encouraged these applications. Particularly, the development of high-throughput “omic” technologies has generated a large number of datasets, which provide a holistic view of a biological process. These datasets present opportunities to discover new

knowledge in the domain. They also present some challenges, especially from their high-dimensionality. High-dimensional datasets are challenging to data mining algorithms because several thousands of candidate variables (e.g., gene expressions or SNPs) can potentially explain an outcome variable of interest (e.g., phenotypes or disease states) but have only a few instances as evidence to support an explanation. These large numbers of candidate variables generate a model search space that is very large for data mining algorithms to explore efficiently, and having only a few instances generates uncertainty for the algorithm to determine the correctness of any candidate model. In such model search spaces, data mining algorithms can easily get stuck in local optima or they may infer associations between spurious variables and the outcome variable, by chance.

Fayyad *et al.*^[3], emphasized the importance of domain prior knowledge in all steps of the KDD process. In biomedicine, often in addition to the dataset, we have some prior domain knowledge about the dataset. This domain knowledge can help guide the data mining algorithm to focus on regions in the model search space that are either objectively more promising for a given problem or subjectively more interesting to a user. The prior knowledge can come from domain literature (e.g., searching through PubMed), a domain expert (e.g., a physician), domain knowledge-bases (e.g., Gene Ontology) or from other related datasets [e.g., from public data repositories like Gene Expression Omnibus (GEO)]. It is now imperative to develop data mining methods that can leverage domain knowledge to assist with the data mining process.

Rule learning methods are among the oldest, well-developed, and widely applied methods in machine learning. They are particularly attractive for KDD tasks because they generate interpretable models with understandable patterns and have good predictive performance. Interpretable models are succinct, human-readable models that explain the reasoning behind their predictions. Bayesian rule learning (BRL) is a rule learning method that has been shown to perform better than state-of-the-art interpretable classifiers on high-dimensional biomedical datasets^[4,5]. BRL takes a dataset as input and searches over a space of Bayesian belief-networks (BN) to identify the BN that best explains the input dataset. BRL then infers a rule model from this BN. BRL uses the Bayesian score^[6] as a heuristic to evaluate a BN during search. The score allows the user to specify a prior belief distribution over the space of BNs that encodes our prior beliefs about which models are more likely to be correct than others with respect to our domain knowledge. Typically in literature uninformative priors are used, which means that we claim that a priori all models are equally likely to be correct. As we saw earlier, often along with the dataset, additional domain knowledge is available that can assist with the data mining process. These sources lead us to believe that some models are more likely to

be correct than others even before we see the dataset. We can specify this belief using informative priors. Two approaches to using informative priors in literature have shown promise^[7,8]. In the Methods and Materials section of this paper, we discuss each of the two approaches and describe ways to extend BRL to specify such informative priors that can incorporate domain knowledge.

In this paper, we implemented an approach to incorporate prior domain knowledge into the BRL learning process using informative priors. We evaluated the effect of this prior knowledge on model learning using experiments with simulated and a real-world lung cancer prognostic dataset.

MATERIALS AND METHODS

In this section, we describe our implementation in BRL to incorporate prior domain knowledge, and then describe two experiments we conducted to evaluate this implementation. Specifically, we describe a BRL greedy best-first search algorithm, the heuristic score used by the search to evaluate candidate models, and our approach to extend this heuristic score to incorporate prior background domain knowledge using informative priors. We call this extension to BRL as BRL_p (BRL with informative priors). After describing our implementation of BRL_p, we describe two experiments we conducted to study the effects of informative priors in model learning: (1) using simulated data; and (2) on a real-world lung cancer prognostic dataset.

BRL

BRL is a rule-based classifier that takes as input, a dataset D , and returns a rule set model. Let the dataset D be an observed instantiation of a system with a probability distribution over a set of n random variables and a target random variable of interest, $D = \{X_i, T_i; i \in 1 \dots n\}$. Here, T is the target variable of interest, which is the dependent variable for the prediction task. Every other variable, X_i in D is an independent random variable that may help predict T . There are a total of m instances in D . In the classification problem, our task is to accurately predict the value of the target variable. For example, consider a diagnostic problem of predicting a disease outcome for a patient, say lung cancer outcome (either Case or Normal), using gene expression biomarker data, measured for each patient. Here, the dataset D would be composed of a set of m patients, each with n gene expression measurements $\{X_i; i \in 1 \dots n\}$. The target variable T is the binary-valued lung cancer outcome variable, $T = \{Case, Normal\}$, for each patient in the dataset.

The BRL search algorithm explores a space of BNs, learned from the observed dataset D , and returns the most optimal BN found during the search. A BN is a graphical representation of the probabilistic dependencies of the different variables in the system under study. They are represented as a directed acyclic

graph (DAG). In our lung cancer diagnostic problem example, an example of probabilistic dependence could be some hypothetical gene expression, say the binary-valued $X_A = \{Up; Down\}$ with a value for up-regulated and a value for down-regulated gene A , is known to be predictive of the outcome T . Then an optimal BN should contain a directed edge from $X_A \rightarrow T$. In other words, the lung cancer outcome depends upon whether or not gene X_A is expressed. In such a BN, the probability distribution, $P(T | X_A)$ is the parameter of the BN.

The parameters of the BN can be represented in form of a conditional probability table (CPT). The CPT is often stored in form of decision trees^[9,10]. The BRL generates a mutually exclusive and exhaustive set of inference rules from this decision tree for prediction of class of any new test instances. Here, each path from root to leaf of the decision tree is interpreted as a rule. The BRL rules are represented in the form of explicit propositional logic: IF antecedent THEN consequent. The rule antecedent is the condition made up of conjunctions (ANDing) of the independent random variable-value pairs, which when matched to a test instance, implies the rule consequent composed of the dependent target variable-value. Continuing with our example, a learned rule can be IF ($X_A = Up$) THEN ($T = Case$). In other words, if the gene X_A is up-regulated then the patient is classified to have a lung cancer outcome as a Case. There are several types of BRL search algorithms^[4,5,11] to help find the optimal BN. In this paper, we will only discuss a simple greedy best-first search algorithm from our previous work^[4] and is summarized in the next sub-section.

BRL greedy best-first search algorithm: The BRL greedy best-first search algorithm is described in detail in the paper by Gopalakrishnan *et al.*^[4], where it is referred to as BRL₁. In this paper, we will refer to this algorithm simply as BRL. We will summarize the algorithm in this subsection. The BRL algorithm initializes the search with a network structure with just the variable T and no parent nodes. In each iteration of the algorithm, one new parent is added to T among the n random variables that is not already a parent of T . This BN implies the hypothesis that T is dependent upon the set of variables added as parents to T . This process is called model specialization. The resulting models from that iteration is added to a priority queue. The priority queue sorts these specialized models by evaluating them using a heuristic score called the Bayesian score, which evaluates the likelihood that the observed dataset was generated by a given hypothesized BN model. This score is described in detail in the next subsection. The greedy search picks the model in the head of the priority queue at the end of the iteration. This model is evaluated to be the best scoring model among the specializations in that iteration. In the next iteration, this model is selected for further specialization by adding more parents. The search terminates when a subsequent specialization step fails to improve the

heuristic score. The search also terminates if the model has reached a limit on the maximum number of parents allowed for T . This search parameter is called maximum conjuncts. Finally, BRL generates a rule model inferred from the model returned by the search.

BRL heuristic score (Bayesian score): BRL search evaluates the quality of a candidate BN model using a heuristic score called the Bayesian score^[9]. In this sub-section, we describe this score. We represent a BN model as the tuple $B = (B_s, B_p)$, where B_s is the network structure with a subset of π discrete-valued nodes, and B_p is the numerical parameters of the network. The posterior probability of the candidate structure given the observed dataset, D , is calculated as in Equation 1.

$$P(B_s | D) = P(B_s, D) / P(D) \quad (1)$$

Since we are comparing Bayesian networks learned from the same dataset D , the denominator does not affect our decision. Only the numerator helps with model selection as shown in Equation 2.

$$P(B_s | D) \propto P(B_s, D) \quad (2)$$

The joint probability of the network structure and the observed dataset, $P(B_s, D)$, is equal to the prior probability of the network structure, $P(B_s)$ and the likelihood that the observed data was generated by that network structure, $P(D | B_s)$. This is shown in Equation 3.

$$P(B_s, D) = P(B_s) \cdot P(D | B_s) \quad (3)$$

To compute the joint probability of the network structure and the observed dataset, $P(B_s, D)$, we use the BDeu score^[6]. We get Equation 4.

$$P(B_s, D; \alpha) = P(B_s) \cdot \prod_{i=1}^n \prod_{j=1}^{q_i} \frac{\Gamma(\frac{\alpha}{q_i})}{\Gamma(N_{ij} + \frac{\alpha}{q_i})} \prod_{k=1}^{r_i} \frac{\Gamma(N_{ijk} + \frac{\alpha}{r_i q_i})}{\Gamma(\frac{\alpha}{r_i q_i})} \quad (4)$$

Here, i iterates through each node in the BN with n nodes. Index j iterates through all, q_i , possible variable-value instantiations of the parents of the i^{th} node. Index k iterates through all r_i values of the i^{th} node. N_{ijk} is the number of instances in D , where the variable i takes the k^{th} value and its parent variables take the j^{th} variable-value instantiation, and $N_{ijk} = \sum_k N_{ijk}$. The Gamma function is defined as $\Gamma(x) = (x-1)!$. The α is a user-defined parameter called prior equivalent sample size (p_{ess}). We set $\alpha = 1$, which allows the data to easily dominate the score^[9]. The $P(B_s)$ term is called the structure prior (see^[9] section 18.3.6.1 for details) that represents the prior belief distribution over all network structures before we look at the data. The remaining terms in Equation 4 compose the likelihood term that infers the likelihood of the network from the observed data.

In the classification task using BRL, we do not learn a fully generalized BN but only care about the relationship of the variables with a specific target variable of interest, T . Variable T is discrete with different values. The set of parents of the i^{th} variable is represented as π_i . In BRL, we learn a constrained BN with node T and its set of parents, π_T . The set π_i can have q_T possible attribute-value instantiations. So, for BN search in BRL, we optimize the heuristic score in

Equation 5.

$$P(B_s, D) = P(B_s) \cdot \prod_{j=1}^{q_T} \frac{\Gamma(\frac{\alpha}{q_T})}{\Gamma(N_j + \frac{\alpha}{q_T})} \prod_{k=1}^{r_T} \frac{\Gamma(N_{jk} + \frac{\alpha}{r_T q_T})}{\Gamma(\frac{\alpha}{r_T q_T})} \quad (5)$$

The expectation of each parameter value of the BN is computed with Equation 6.

$$\mathbb{E}[\theta_{jk} | D, B_s] = \frac{N_{jk} + \frac{\alpha}{r_T q_T}}{N_j + \frac{\alpha}{q_T}} \quad (6)$$

We use this value as the posterior probability of the rule. The number of rules inferred by BRL is equal to the number of θ_{jk} values in the BN. The expectation of this value shows the degree of support a rule has in the observed dataset.

BRL with structure priors: In Equation 5, the $P(B_s)$ term is the structure prior that represents the prior distribution over all network structures. Here, we can specify our prior bias of certain network structure over others to skew the BRL search to focus on certain network structures more than others. Typically, in literature uninformative priors are used, which means that a priori we claim that we do not have any preference of network structures over the others. BRL in this case lets the data alone decide the final learned model. The challenge of specifying these priors is that the total number of network structures grows super-exponentially with the number of variables n ^[12]. It often becomes infeasible to specify structure priors for each of these network structures for even moderately sized datasets. So far in BRL, we had been using an uninformative prior by setting $P(B_s) = 1$, in Equation 5.

Castelo and Siebes^[7] describe a promising approach to elicit structure priors by specifying the probability of the presence or absence of each edge in the network structure. The user only needs to specify the probability of a subset of edges in the network structure. The probabilities for all the remaining edges are assigned a discrete uniform distribution value. A challenge using this approach is to specify the values of these probabilities. In our experiments with BRL using these priors, we observed that the likelihood term in Equation 5 always dominates the structure prior term. It would help us if we could control the influence of structure priors over the likelihood term using a scaling factor. As we described earlier in the introduction section, the background knowledge, we specify, itself has uncertainty associated with it. A scaling factor would help us control the influence of data and our prior knowledge.

Mukherjee and Speed^[8] propose an informative prior that uses a log-linear combination of weighted real-valued function of the network structure, $f_i(B_s)$. This function is called the concordance function. It can be any function that monotonically increases with the increase in agreement between the learned network structure and the prior beliefs of the user. This is shown in Equation 7.

$$P(B_s) \propto \exp[\lambda \cdot \sum w_i f_i(B_s)] \quad (7)$$

The hyperparameter w_i are the positive weights that represent the relative importance of each function. The hyperparameter λ is a scaling factor that helps to

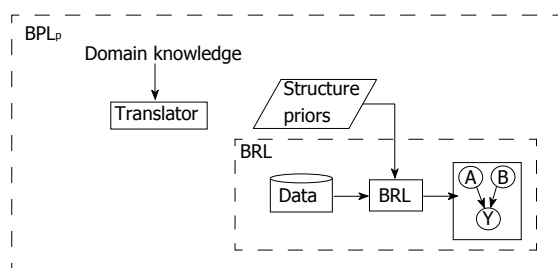


Figure 1 The Bayesian rule learning framework that can incorporate domain knowledge. BRL: Bayesian rule learning.

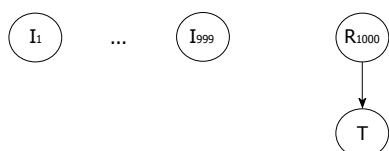


Figure 2 The data-generating graph for the simulated data.

control the overall influence of the structure prior. This will help us quantify the uncertainty in the validity of our prior knowledge.

The structure prior we used for BRL_p comes from an instantiation of the general form of this prior, shown in Equation 7, as described by Mukherjee and Speed^[8]. It allows the user to specify their prior beliefs about the presence and absence of the edges in the network structure. This instantiation is shown in Equation 8.

$$P(B_s) \propto \exp[\lambda \cdot (|E(B_s) \cap E_+| - |E(B_s) \cap E_-|)] \quad (8)$$

Here, set E_+ (positive edge-set) represents the set of edges the user believes should be present in the model, and set E_- (negative edge set) represents the set of edges the user believes should be absent from the model. So, the concordance function in this instantiation simply gives a positive count for if the candidate graph contains an edge from the positive edge-set, and a negative count (penalty) when it contains an edge from the negative edge-set. In this instantiation, the weights hyperparameter is set to 1, since our counts are all valued 1. We need to learn the value of the hyperparameter λ . The range of values it can take depends upon the well-known Jeffrey's scale^[13]. When $\lambda = 0$, the whole exponent becomes 0, and $P(B_s) = \exp(0) = 1$, which is the uninformative prior. In other words, when $\lambda = 0$, BRL_p should have no effect of structure prior and so would behave the same as the baseline model, BRL. As we increase the value of λ , the effect of the structure prior would have an increased influence over the likelihood term in Equation 5.

To summarize, BRL_p uses a heuristic score called the BDeu score, shown in Equation 5, and encodes the structure prior in that score using Equation 8. The BRL_p framework is shown in Figure 1. The inner dotted box, labeled "BRL", is the classic BRL without prior knowledge, which takes in an input dataset, uses BRL algorithm to learn and output a model. The outer dotted box is our extension, BRL_p that can incorporate domain

knowledge. The translator process, currently done manually, converts knowledge from various sources to input into Equation 8.

Experiment design

In this section, we describe our experiment design that we used to demonstrate the functionality of BRL_p. We examined its behavior on both, simulated dataset, and on a real-world dataset. We were mainly interested in the ability of BRL_p to incorporate the supplied prior domain knowledge with respect to the structure prior hyperparameter λ . Additionally, we also monitored the changes in the predictive power of the learned model resulting from the influence of the supplied prior domain knowledge. We studied the functionality of BRL_p on a simulated dataset, and then on a real-world dataset. Each is described, in detail, in the following sub-sections.

Simulated data analysis: We first generated simulated data to study the behavior of BRL_p. We can control the properties of the simulated dataset, which gave us a controlled environment to check if BRL_p was behaving as we expected on a dataset with the specified properties.

Data generation: We generated a simulated dataset with 1000 variables in addition to the target variable, T . We show the data-generating graph in Figure 2. Out of the 1000 candidate variables that can predict T , only one variable, R_{1000} , is relevant. A relevant variable is a variable that helps to predict T . All the remaining 999 variables, $\{I_1 \dots I_{1000}\}$, are irrelevant. Irrelevant variables are random values that do not help predict T . All the random variables in the graph are binary $\{0, 1\}$. The conditional distributions in the graph are Bernoulli with the success parameter p depending upon the value instantiation of their parent variables. The irrelevant and relevant variable values were randomly sampled with $p = 0.5$. The T variable value was sampled with $p = 0.9$ if its parent, R_{1000} , took the value 1, and $p = 0.1$ otherwise.

Data background knowledge: In a simulation problem, we already knew the true data-generating graph as shown in Figure 2. We knew that in the learned network structure from BRL_p, there should be an edge present between R_{1000} and T . So, in Equation 8, the positive edge-set only contained this edge, $E_+ = \{(R_{1000}, T)\}$. All the edges between irrelevant variables and T should be absent in the BRL_p model, so they went to the negative edge-set, $E_- = \{(I_k, T); k = 1 \dots 999\}$. We evaluated the impact of the λ hyperparameter value of the structure prior on the final model learned by BRL_p.

Methods evaluated: We evaluated the method BRL_p here. We set the user-defined, search algorithm

parameter of BRL_p of maximum conjuncts (constraint on maximum number of parents of T) to 8. We evaluated the effect of the hyperparameter λ by assigning its values $\lambda = \{0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10\}$. The value of $\lambda = 0$ represents the baseline model of BRL with no structure priors.

Evaluation metrics: We evaluated BRL_p with two metrics: (1) graph edit distance (GED); and (2) area under the receiver operator characteristics curve (AUC). We evaluated them over 5 runs of 10-fold cross-validation. In each run, the dataset was randomly shuffled to produce a different set of 10 stratified folds. GED measures how much of the prior domain knowledge gets incorporated into the model learning process. Specifically, how much does the model learned by BRL_p agree with the supplied prior knowledge? This metric is described in detail in the next paragraph. We monitored the BRL_p model predictive power by measuring the average AUC across the 5 runs of 10-fold cross-validation. The AUC helped us monitor the influence of structure priors in model predictive performance.

GED^[14] is a metric of similarity between two graphs. In this experiment, we compared two constrained BNs. Specifically, we were interested in measuring how closely our BRL_p predicted BN, \hat{B}_S (learned by BRL_p) resembled the true BN, B_S , which generated the simulated dataset (Figure 2 in this experiment). This was used to estimate the value of adding structure prior knowledge for model learning when the true model is available for comparison. We computed this metric using Equation 9.

$$d_{vmin}(B_S, \hat{B}_S) = \min_{v \in \gamma(B_S, \hat{B}_S)} \sum_{e_i \in v} c(e_i) \quad (9)$$

Here, $d_{vmin} = [B_S, \hat{B}_S]$ is a function that returns the GED between the two BNs. A specific e_i is an edit operation to transform one graph into another. For the constrained BN we have two available edit operations - delete edge, and insert edge. There is a cost $c(e_i)$ associated with each edit operation. We set $c(e_i) = -1$, for both the edit operations. A v is an edit path containing a sequence of edit operations to transform graph B_S into \hat{B}_S . The set $\gamma[B_S, \hat{B}_S]$ is a set of all possible edit paths. To compute the graph edit distance, we find the edit path, v , that minimizes the overall cost and then return this minimum cost value indicating the minimum number of operations needed to transform one graph to another. Therefore, an edit distance of 0 indicates that the predicted graph is identical to the true graph. Since the maximum parents resulted from BRL is constrained to 8 from the user parameter, the worst possible model contains all 8 irrelevant variables. So, we get $d_{vmin} = 9$ (8 edge deletion operations from irrelevant variables, 1 insert edge operation to the relevant variables).

Real-world lung cancer prognostic biomarker data analysis: We obtained a real-world dataset for our analysis from Gene Expression Omnibus^[15] (GEO),

a public gene-expression data repository. We extracted the dataset from a study^[16] that collected both tumor and normal tissue samples from 60 female non-small cell lung cancer (NSCLC) patients in Taiwan. As a result, there were 120 samples in this dataset (60 patients, each with paired tumor and normal tissue). RNA was extracted from these paired tumor and normal tissues for gene expression analysis on the Affymetrix Human Genome U133 Plus 2.0 Array platform. The platform has 54675 probes. The accession ID for this study on GEO database is GSE19804.

Data pre-processing: The raw dataset extracted from GEO contained 54675 probes and 120 instances. We needed to pre-process the data to prepare it for data analysis. The dataset pre-processing was done using Bioconductor (version 3.6) packages in R (version 3.4.3). We extracted the raw dataset using the *affy* package^[17]. We used Robust Multichip Analysis (RMA) for background correction, quantile normalization, and probe summarization. We mapped probes to the genes they represented. Multiple probes can map to a single gene. In the final dataset, we would like to have just one random variable representing a unique gene. Among the multiple probes that map to a single gene, we chose the probe with the largest inter-quantile range to represent the gene. This process is called inter-quantile range (IQR) filtering. Finally, we also extracted the tissue phenotype (tumor or normal) for each sample and add to this dataset. The outcome variable of interest was this tissue phenotype. After this pre-processing step, we were left with 16382 genes. So, the final dataset for our analysis had 16382 variables and 120 instances. The R script we used for data pre-processing is available in the GitHub repository linked in the Conclusion section.

Many classification algorithms, including BRL, cannot handle continuous-valued variables, and require the input data to be discretized. Moreover, supervised discretization can help improve the performance of several classifiers including Support Vector Machines and Random Forests^[18]. This is because supervised discretization acts as a feature selector that only retains variables with meaningful discretization bins. Biomedical datasets are high dimensional, there can be many noisy and redundant variables. Supervised discretization can help remove some of these variables from the model learning process. We discretized the dataset using efficient Bayesian discretization (EBD), a supervised discretization method, which has been shown to obtain better classification performance and stability but less robust when compared to the popular Fayyad-Irani supervised discretization method on several biomedical datasets^[19]. We set the user-defined lambda parameter of EBD, to 0.5, as the recommended default value in the paper. During model learning, we split the data into 10 folds for cross-validation. For each train-test fold pair, supervised discretization bins were learned on the train dataset alone. The learned bins were applied to the test

Table 1 Clinical features of the 60 non-small cell lung cancer patients in the real-world lung cancer prognostic dataset

Attribute	Value	<i>n</i> (%)
Gender	Women	60 (100)
	Men	0 (0)
Tumor type	Adenocarcinoma	56 (93)
	Bronchioloalveolar carcinoma	3 (5)
	Squamous	1 (2)
	Others	0 (0)
Smoking history	Yes	0 (0)
	No	60 (100)

Statistics extracted from the paper by Lu *et al*^[16].

dataset. So, during supervised discretization, we did not look at the test dataset.

Data background knowledge: We explored the medical literature for known prognostic markers that may assist in model learning with BRL_p. Before exploring, we first sought to understand more about the dataset, which turned out to have some interesting characteristics making it highly worthy of study. Of note, only tissue samples taken from non-smokers who were all women, who had contracted lung cancer were analyzed in this study. Table 1 summarizes some clinical features known about the 60 Taiwanese NSCLC patients studied in the dataset as described in the paper of the study^[16].

We noted from the Table 1 that the subjects in the dataset were all women (60 out of 60 patients), contain mainly adenocarcinoma patients (56 out of 60 patients), and none of them had any smoking history (60 out of 60 patients). Additionally, we also knew that all the patients were from Taiwan. So, we explored the medical literature to find known prognostic markers for this sub-population. Epidermal growth factor receptor (EGFR), a receptor tyrosine kinase is prognostic marker known to be frequently over-expressed in NSCLC^[20]. EGFR encodes a transmembrane glycoprotein, a receptor for members of the epidermal growth factor family. A ligand binding to this receptor induces dimerization and tyrosine autophosphorylation, and leads to cell proliferation (referred from RefSeq, June 2016). In NSCLC patients, Shigematsu *et al*^[21] observed that EGFR domain mutations are statistically significantly more frequent in women than men (42% vs 14%), in adenocarcinomas than other histologies (40% vs 3%), in non-smokers than smokers (51% vs 10%), and in East Asians than other ethnicities (30% vs 8%); all with a *P*-value of < 0.001. This description is very similar to the subjects in the dataset we are studying. Therefore, EGFR gene expression was potentially a good candidate to be incorporated as prior domain knowledge into model learning with BRL_p on this dataset.

Methods compared: We again evaluated BRL_p here. We set its of maximum conjuncts to 8. We evaluated the effect of the hyperparameter λ by assigning it

values of $\lambda = \{0, 1, 2, 4, 6, 8, 10, 20\}$. The value $\lambda = 0$ represents the baseline model of BRL with no structure priors. We included $\lambda = 20$, to study the scenario where the structure priors overwhelmingly dominates the likelihood score. Additionally, we compared these models with some state-of-the-art classifiers including three interpretable class of classifiers namely - C4.5^[22], RIPPER^[23], and PART^[24]; and three complex and non-interpretable classifiers namely- Random Forests^[25], naïve Bayes^[26], and Support Vector Machines^[27]. C4.5^[22] is a popular decision tree learning algorithm, where each path of the decision tree can be interpreted as rules. RIPPER^[23] (Repeated Incremental Pruning to Produce Error Reduction) is a propositional rule learning algorithm that uses a divide-and-conquer strategy during model training. PART^[24] is a rule learning method that combines the approaches of both C4.5 and RIPPER by building partial decision trees, inferring rules from the trees, and using a divide-and-conquer strategy to build the rule model. Random Forest^[25] is an ensemble learning method that learns a number of decision trees during training, and combines predictions from them during inference. The naïve Bayes^[26] classifier is a simple probabilistic classifier that learns a network with strong independence assumption between the variables, and uses the Bayes theorem for inference from the learned network. Support Vector Machines^[27] is an algorithm that learns a hyperplane function to differentiate the classes in the problem space. We ran these classifiers from the Weka^[28] workbench (version 3.8.1) using the default parameters for each classifier.

Evaluation metrics: We evaluated BRL_p with two metrics: (1) Prior Frequency (PF); and (2) AUC. We evaluated the dataset over 5 runs of 10-fold cross-validation. For this real-world scenario, we used PF to measure the gain of the background knowledge into BRL_p. With the simulated dataset, we had evaluated using GED because we knew the true data-generating graph. In most real-world problems, we do not know the true model that generated the data and so, we cannot use GED. PF measures the fraction of models learned on each of the 50 folds (5 runs of 10-fold cross-validation) that incorporates the specified prior domain knowledge. In this experiment, we measured the fraction of the models that contained an edge between EGFR and *T* in the learned BRL_p model.

RESULTS

In this section, we present the results from our experiments examining the effects of the λ hyperparameter of the structure prior, and consequentially the influence of the specified prior knowledge on model learning. We show our results using the simulated data first, and then from the real-world lung cancer prognostic dataset.

Simulation data analysis results

The results from the 5 runs of 10-fold cross-validation

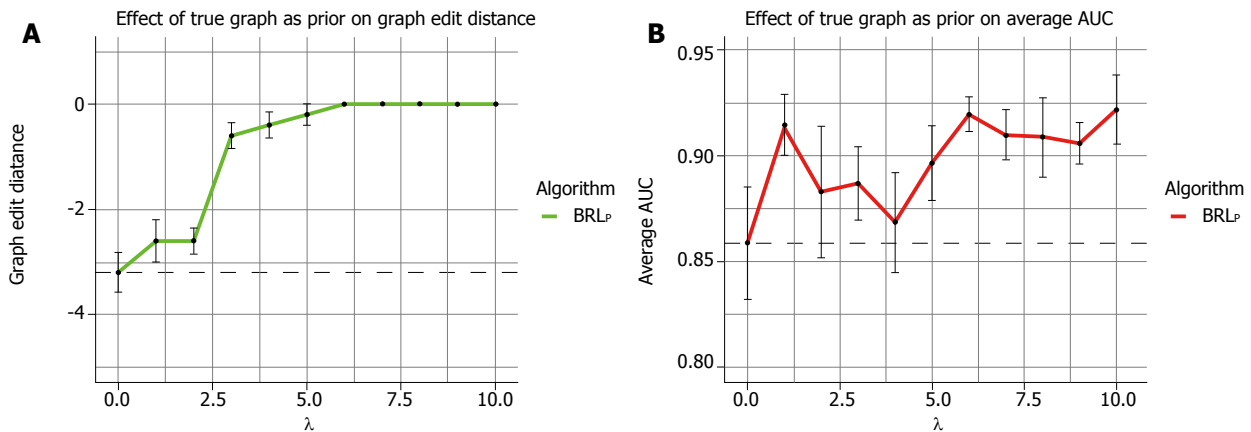


Figure 3 Evaluation metrics on Bayesian rule learning model learning with simulated data. A: Graph edit distance between BRL_p and true data-generating model; B: Area under the receiver operator characteristic curve of the BRL_p model. BRL_p: Bayesian rule learning with informative priors; AUC: Area under the receiver operator characteristic curve.

1. IF (RV1000 = 0) THEN (T = 0)
Posterior Probability = 0.9944, TP = 44, FP = 0, Pos = 48, Neg = 52
2. IF (RV1000 = 1) THEN (T = 1)
Posterior Probability = 0.9248, TP = 52, FP = 4, Pos = 52, Neg = 48

Figure 4 Bayesian rule learning generated rule model with $\lambda=10$ (highest average area under the receiver operator characteristic curve) on the simulated dataset. Each rule has its posterior probability, the number of true positives (TP), false positives (FP), total number of examples that match the rules consequent target value (Pos), and total number that do not match the right hand side of the rule (Neg). The TP measures examples that correctly match the rules left and right hand sides, while FP measures examples that correctly match the rules condition or left-hand-side, but have a different consequent or right-hand-side.

are summarized in Figure 3. In Figure 3A, the various values of the hyperparameter λ is shown in the x-axis, while the y-axis shows the average GED. This average is obtained across the 10-folds of each run, and then averaged across the 5 runs. Each data-point in the graph is this average deviation from the true model as measured by the GED, and the error bars represent the standard error of mean. The dotted line shows the value of BRL_p with $\lambda = 0$, which as we mentioned earlier is the same as BRL, where we use uninformative priors. We saw that even with $\lambda = 1$, the structure priors helped improve the GED thereby bringing the learned model closer to the data-generating model. We saw a sharp gain of GED from $\lambda = 2$ to 3. For $\lambda \geq 6$, BRL_p returned the true data-generating model specified by the structure priors. This showed that BRL_p effectively and correctly incorporates the specified domain knowledge. The degree of incorporation is controlled by λ .

Figure 3B displays the average AUC. The overall trend is a gain in AUC but the trend is noisy, especially with low λ values when the GED > 0. This region indicated models that picked up irrelevant variables, which were spurious and were associated with T , by chance. Their AUC fluctuated a lot because random associations were found. When $\lambda \geq 6$, the GED reached the perfect 0, we saw a rise in AUC. The noise reduced in this region of the graph. Random samplings from our simulation generated slightly different values of the parameters, which were reflected in the

fluctuations here. So, from the AUC graph we saw a gradual gain in predictive performance with the incorporation of prior knowledge of the truth.

Figure 4 shows a BRL_p rule model obtained when $\lambda = 10$, which achieved the largest average AUC from our experiments (AUC = 0.92). The particular run achieved an AUC of 0.96 on the 10-fold cross-validation and a GED of a perfect 0. The posterior probability was computed using Equation 6. TP and FP refers to the total true positives and false positives. Pos and Neg are the total positives and negative examples. Our simulation design only had one relevant variable, R_{1000} , and 999 irrelevant variables, $\{I_1 \dots I_{1000}\}$. The rule model in Figure 4 correctly picked up only the relevant variable. We had designed the simulation such that if the relevant variable took the value 1, then T would be sampled with a Bernoulli distribution with $p = 0.9$, this was reflected in Rule 2. So, BRL_p accurately retrieved the true data-generating model assisted by informed structure priors.

Real-world lung cancer prognostic data analysis results

The results from the 5 runs of 10-fold cross-validation on the real-world lung cancer prognostic dataset are summarized in Figure 5. We specified the structure prior of an edge between EGFR and the outcome *Class* variable to be present. We altered the values of λ and observed its effect on the learned model. Figure 5A, shows the effect of the different values of λ on PF, the fraction of models that contained EGFR. From $\lambda = 2$ to 6,

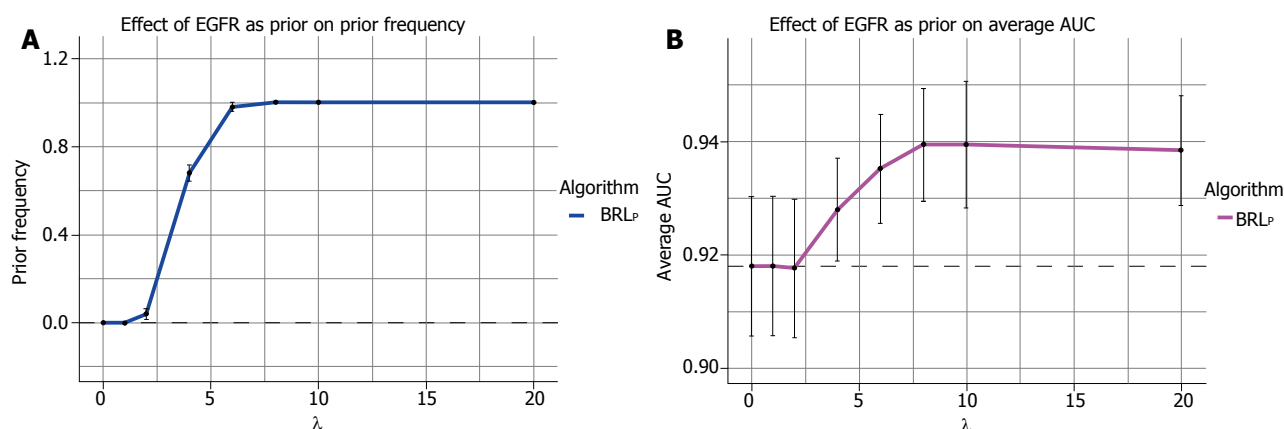


Figure 5 Evaluation metrics on Bayesian rule learning model learning with real-world lung cancer prognostic dataset. A: Prior frequency of the edge between epidermal growth factor receptor and T in BRLp model; B: Area under the receiver operator characteristic curve of the BRLp model. BRLp: Bayesian rule learning with informative priors; EGFR: Epidermal growth factor receptor; AUC: Area under the receiver operator characteristic curve.

we saw a steep gain in PF. For $\lambda \geq 8$, EGFR was present in every learned model. This again showed that BRLp effectively incorporated the specified prior knowledge and the λ hyperparameter allowed the user to determine the degree of incorporation of this knowledge by BRLp.

Figure 5B, shows the gain of average AUC across 5 runs of 10-fold cross-validation. We observe a steady gain of AUC for $\lambda > 2$. For $\lambda \geq 8$, the AUC gain tapers off. The results show that the EGFR prior knowledge helped improve the AUC of BRLp.

BRLp with $\lambda = 8$ generated the highest average AUC of 0.935. Figure 6 shows the rule model from one of the runs, which had achieved a cross-validation AUC of 0.967 and PF of 1. Rule 1 had the highest amount of evidence (38 true positives and no false positives) for the outcome Control (normal tissue). This rule had the EGFR value range from negative infinity to 10.8. In other words, EGFR was under-expressed in these 38 normal tissue instances. Rule 15 had the highest amount of evidence (15 true positives) for the outcome Case. This rule had EGFR value range from 10.8 to positive infinity. In other words, EGFR was over-expressed in these 15 tumor tissue instances. These rules also lent support to what we had found in the literature about EGFR being over-expressed in tumor cells. In addition to EGFR, which was incorporated from the structure prior, the model picked up 3 other variables during model learning from the dataset. They were ephrin A4 (EFNA4), killer cell lectin like receptor G2 (KLRG2), and C2 calcium dependent domain containing 6 (C2CD6).

Finally, we compared two BRLp models with state-of-the-art classifiers using average AUC achieved across 5 runs of 10-fold cross-validation. The two BRLp models were (1) with $\lambda = 0$, which represented the baseline BRL model with uninformative priors, and (2) with $\lambda = 8$ that incorporated EGFR into the structure prior, which achieved the highest average AUC of 0.935. The state-of-the-art classifiers compared were C4.5, RIPPER,

PART, Random Forests, naïve Bayes, and Support Vector Machines. This comparison is shown in Figure 7.

The first two bars in Figure 7 are BRLp algorithms, BRLp with $\lambda = 0$ is indicated as BRL, and then BRLp with $\lambda = 8$. We saw a gain in performance from incorporating EGFR as structure priors. The next three bars - C4.5, RIPPER, and PART are interpretable class of models, which are human readable. C4.5 is a decision tree learning algorithm. RIPPER and PART are rule learning algorithms. We noticed that these three algorithms performed worse than both BRLp algorithms in this dataset. The last three bars in Figure 7 are - Random Forest, naïve Bayes, and Support Vector Machines. These are examples of complex models that use all variables in the dataset to generate a classifier. It is not easy to explain the reasoning behind their predictions. But all three algorithms here outperformed BRLp on this dataset. This comparison shows the trade-off of predictive performance and interpretability. On this dataset, BRLp offered an interpretable model that outperformed other popular interpretable models but did not perform as well as the complex models.

DISCUSSION

An important practical consideration to note while specifying structure priors is to avoid specifying priors that introduce bias into the model search. Informative priors can be biased if they are inferred based on the predictions, of some predictive model, on the test dataset. For example, if we notice that our learned model predicts poorly on a subset of test instances, and we notice some independent variable(s) strongly associated with the target variable in that subset of test instances. Specifying, our newly found association from the predictions on the test dataset, into the structure priors to re-learn the model will return a biased model and must be avoided.

Mukherjee and Speed^[8] show how the general form of the score in Equation 7 can be extended to

1.IF ((EFNA4 = -inf to 6.9) (KLRG2 = 6.4 to inf) (EGFR = -inf to 10.8) (C2CD6 = -inf to 3.9)) THEN (Class = Control)
Posterior Probability=0.9995, TP=38, FP=0, Pos=60, Neg=60

2.IF ((EFNA4 = 6.9 to 7.5) (KLRG2 = 6.4 to inf) (EGFR = -inf to 10.8) (C2CD6 = -inf to 3.9)) THEN (Class = Control)
Posterior Probability=0.9977, TP=9, FP=0, Pos=60, Neg=60

3.IF ((EFNA4 = -inf to 6.9) (KLRG2 = 6.4 to inf) (EGFR = 10.8 to inf) (C2CD6 = -inf to 3.9)) THEN (Class = Control)
Posterior Probability=0.9959, TP=5, FP=0, Pos=60, Neg=60

4.IF ((EFNA4 = -inf to 6.9) (KLRG2 = -inf to 6.4) (EGFR = -inf to 10.8) (C2CD6 = -inf to 3.9)) THEN (Class = Control)
Posterior Probability=0.9959, TP=5, FP=0, Pos=60, Neg=60

5.IF ((EFNA4 = 7.5 to inf) (KLRG2 = 6.4 to inf) (EGFR = 10.8 to inf) (C2CD6 = 3.9 to inf)) THEN (Class = Control)
Posterior Probability=0.9898, TP=2, FP=0, Pos=60, Neg=60

6.IF ((EFNA4 = 6.9 to 7.5) (KLRG2 = 6.4 to inf) (EGFR = 10.8 to inf) (C2CD6 = -inf to 3.9)) THEN (Class = Control)
Posterior Probability=0.98, TP=1, FP=0, Pos=60, Neg=60

Rules 7 through 14 match 0 instances and so are removed from display.

15.IF ((EFNA4 = 7.5 to inf) (KLRG2 = -inf to 6.4) (EGFR = 10.8 to inf) (C2CD6 = -inf to 3.9)) THEN (Class = Case)
Posterior Probability=0.9986, TP=15, FP=0, Pos=60, Neg=60

16.IF ((EFNA4 = 7.5 to inf) (KLRG2 = -inf to 6.4) (EGFR = -inf to 10.8) (C2CD6 = -inf to 3.9)) THEN (Class = Case)
Posterior Probability=0.9985, TP=14, FP=0, Pos=60, Neg=60

17.IF ((EFNA4 = 7.5 to inf) (KLRG2 = 6.4 to inf) (EGFR = 10.8 to inf) (C2CD6 = -inf to 3.9)) THEN (Class = Case)
Posterior Probability=0.9974, TP=8, FP=0, Pos=60, Neg=60

18.IF ((EFNA4 = 7.5 to inf) (KLRG2 = -inf to 6.4) (EGFR = 10.8 to inf) (C2CD6 = 3.9 to inf)) THEN (Class = Case)
Posterior Probability=0.997, TP=7, FP=0, Pos=60, Neg=60

19.IF ((EFNA4 = 7.5 to inf) (KLRG2 = -inf to 6.4) (EGFR = -inf to 10.8) (C2CD6 = 3.9 to inf)) THEN (Class = Case)
Posterior Probability=0.9959, TP=5, FP=0, Pos=60, Neg=60

20.IF ((EFNA4 = 7.5 to inf) (KLRG2 = 6.4 to inf) (EGFR = -inf to 10.8) (C2CD6 = -inf to 3.9)) THEN (Class = Case)
Posterior Probability=0.9948, TP=4, FP=0, Pos=60, Neg=60

21.IF ((EFNA4 = 6.9 to 7.5) (KLRG2 = -inf to 6.4) (EGFR = 10.8 to inf) (C2CD6 = -inf to 3.9)) THEN (Class = Case)
Posterior Probability=0.9932, TP=3, FP=0, Pos=60, Neg=60

22.IF ((EFNA4 = 7.5 to inf) (KLRG2 = 6.4 to inf) (EGFR = -inf to 10.8) (C2CD6 = 3.9 to inf)) THEN (Class = Case)
Posterior Probability=0.9898, TP=2, FP=0, Pos=60, Neg=60

23.IF ((EFNA4 = 6.9 to 7.5) (KLRG2 = -inf to 6.4) (EGFR = -inf to 10.8) (C2CD6 = 3.9 to inf)) THEN (Class = Case)
Posterior Probability=0.98, TP=1, FP=0, Pos=60, Neg=60

24.IF ((EFNA4 = 6.9 to 7.5) (KLRG2 = -inf to 6.4) (EGFR = 10.8 to inf) (C2CD6 = 3.9 to inf)) THEN (Class = Case)
Posterior Probability=0.98, TP=1, FP=0, Pos=60, Neg=60

Figure 6 Bayesian rule learning generated rule model with $\lambda=8$ (highest average area under the receiver operator characteristics curve) on the real-world lung cancer prognostic dataset. TP: True positives; FP: False positives; Pos: Total number of examples that match the rules consequent target value; Neg: Total number that do not match the right hand side of the rule; EGFR: Epidermal growth factor receptor.

incorporate other kinds of prior knowledge including rewarding network sparsity, where structure priors can be used as a regularization term. In the introduction section, we had discussed other sources of prior knowledge than literature, including - input from a domain expert (e.g., A physician), domain ontology (e.g., Gene Ontology), and models learned from other related datasets. In the future, we will explore the incorporation of knowledge from these other sources. In novel biomarker discovery, we could place all of our known knowledge into the negative edge-set in Equation 8. Models learned from such a structure

prior would be penalized for learning already known biomarkers and would encourage discovery of novel biomarkers. We used an instantiation of the general form of the score, in Equation 7, where the relative weights, w_i , of each of i^{th} network are set to 1. It would be interesting to explore different relative weights for different network features and see its impact on model learning. In this paper, we performed a grid search over the hyperparameter λ . We would like to explore if we can come up with better ways to optimize the value of this hyperparameter.

In this paper, we implemented BRL_p , a method

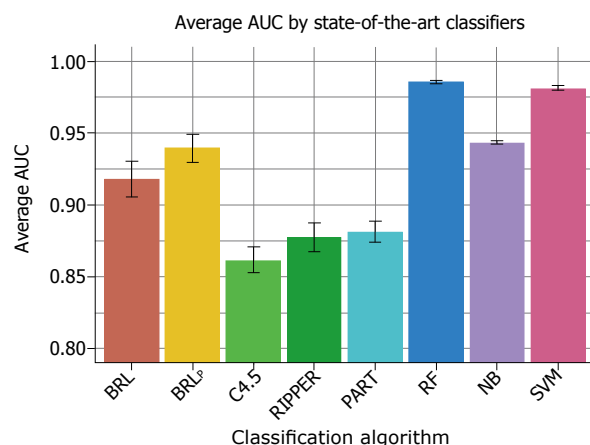


Figure 7 Comparison of area under the receiver operator characteristics curve achieved by Bayesian rule learning with state-of-the-art classifiers. AUC: Area under the receiver operator characteristic curve.

that extended BRL to allow it to integrate prior domain knowledge using structure priors into the model learning process. We demonstrated the ability of BRL_p to incorporate this knowledge on simulated data and a real-world lung cancer prognostic dataset. We observed that the λ hyperparameter allowed us to control the degree of incorporation of prior knowledge. This parameter can be helpful if we were uncertain about our specified prior knowledge. We also observed that relevant prior knowledge could sometimes help improve the predictive performance of BRL_p. Methods developed in this paper, the simulation data experiment code, and the R script for data extraction and processing of the prognostic dataset, are all made publicly available in an online repository (<https://github.com/jeya-pitt/brl-structure-priors>). We envision that BRL_p will be very beneficial in data mining tasks across domains where some prior domain knowledge is available.

ARTICLE HIGHLIGHTS

Research background

Biomedicine is increasingly a data-driven science, owing largely to the explosion in data, especially from the development of high-throughput technologies. Such datasets often suffer from the problem of high-dimensionality, where a very large number of candidate variables can explain the outcome variable of interest but have few instances to support any model hypothesis. In many applications, in addition to the data itself, some domain knowledge is available that may assist in the data mining process to help learn more meaningful models. It is important to develop data mining tools to leverage this available domain knowledge. However, currently, there is a dearth of data mining methods that can incorporate this available domain knowledge.

Research motivation

Developing data mining methods that can incorporate domain knowledge will help learn more meaningful models and will benefit many domains, especially the ones that suffer from data scarcity but have some domain knowledge that can assist with the data mining process (for example - biomedicine).

Research objectives

In this work, our objective was to extend a rule learning algorithm, called Bayesian rule learning (BRL), to make it capable of incorporating prior domain

knowledge. BRL is a good candidate because it has been shown to be successful in application to high-dimensional biomedical data analysis tasks. We implemented such a tool, called BRL_p that has tunable priors, which means the user can control the degree of incorporation of their specified knowledge. BRL_p is a novel data mining tool that allows the user to specify their domain knowledge (including uncertain domain knowledge) and incorporates it into the model search process.

Research methods

BRL searches over a space of Bayesian belief network models (BNs) to find the optimal network and infers a rule set from that model. We implemented a way for the BN to incorporate informative priors, a distribution encoding the relative importance of each model prior to seeing the training data. This allowed BRL to incorporate user-specified domain knowledge into the data mining process called BRL_p. BRL_p has a hyperparameter λ that allows the user to adjust the degree of incorporation of their specified prior knowledge.

We evaluated BRL_p by comparing it to BRL (without informative priors) and other state-of-the-art classifiers on a simple simulated dataset, and a real-world lung cancer prognostic dataset. We measured the degree of acceptance of the specified prior knowledge with respect to the hyperparameter λ in BRL_p. We also observed the changes in predictive power using AUC.

Research results

We observed, in both the experiments with simulated data and the real-world lung cancer prognostic data that with increasing values of λ the degree of incorporation of the specified prior knowledge also increased. We also observed that specifying prior knowledge relevant to the problem dataset could sometimes help find models with better predictive performance. When BRL_p is compared to the state-of-the-art classifiers, we observed that it performed better than other interpretable models but the more complex and non-interpretable models achieved better predictive performance than BRL_p.

Research conclusions

BRL_p allows the user to incorporate their specified domain knowledge into the data mining task and allows them to control the degree of incorporation with a hyperparameter. This is a novel rule learning algorithm that we have made available to the general public via GitHub. We anticipate its use in many applications especially the ones suffering from data scarcity but have additional domain knowledge available that may assist in the data mining task.

Research perspectives

In this paper, we explored specifications of simple domain knowledge. We need to further explore the incorporation of more complex forms of knowledge. In this paper, we incorporate domain knowledge from literature. We also want to explore domain knowledge available in other sources. These future directions may motivate further developments to BRL_p.

REFERENCES

- 1 **Fayyad UM**, Piatetsky-Shapiro G, Smyth P, Uthurusamy R. Advances in knowledge discovery and data mining. *Technometrics* 1996; **40**: xviii
- 2 **Esfandiari N**, Babavalian MR, Moghadam AME, Tabar VK. Knowledge discovery in medicine: Current issue and future trend. *Expert Syst Appl* 2014; **41**: 4434-4463 [DOI: 10.1016/j.eswa.2014.01.011]
- 3 **Fayyad U**, Piatetsky-Shapiro G, Smyth P. From data mining to knowledge discovery in databases. *Ai Magazine* 1996; **17**: 37-54
- 4 **Gopalakrishnan V**, Lustgarten JL, Visweswaran S, Cooper GF. Bayesian Rule Learning for Biomedical Data Mining. *Bioinformatics* 2010; **26**: 668-675 [PMID: 20080512 DOI: 10.1093/bioinformatics/btq005]
- 5 **Lustgarten JL**, Balasubramanian JB, Visweswaran S, Gopalakrishnan V. Learning Parsimonious Classification Rules from Gene Expression Data Using Bayesian Networks with Local Structure. *Data* 2017; **2**: 5 [DOI: 10.3390/data2010005]
- 6 **Buntine W**. Theory refinement on Bayesian networks. *Uncertainty Proceedings* 1991; **14**: 52-60 [DOI: 10.1016/B978-1-55860-20

- 3-8.50010-3]
- 7 **Castelo R**, Siebes A. Priors on network structures. Biasing the search for Bayesian networks. *Int J Approx Reason* 2000; **24**: 39-57 [DOI: 10.1016/S0888-613X(99)00041-9]
- 8 **Mukherjee S**, Speed TP. Network inference using informative priors. *Proc Natl Acad Sci USA* 2008; **105**: 14313-14318 [PMID: 18799736 DOI: 10.1073/pnas.0802272105]
- 9 **Koller D**, Friedman N. Probabilistic Graphical Models: Principles and Techniques - Adaptive Computation and Machine Learning. MIT Press 2009: 161-168
- 10 **Chickering DM**, Heckerman D, Meek C. A Bayesian approach to learning Bayesian networks with local structure. Thirteenth Conference on Uncertainty in Artificial Intelligence 1997; **11**: 80-89
- 11 **Balasubramanian JB**, Visweswaran S, Cooper GF, Gopalakrishnan V. Selective model averaging with bayesian rule learning for predictive biomedicine. *AMIA Jt Summits Transl Sci Proc* 2014; **2014**: 17-22 [PMID: 25717394]
- 12 **Harary F**, Palmer EM. Graphical enumeration: Elsevier, 2014
- 13 **Jeffreys H**. The theory of probability. OUP Oxford, 1998
- 14 **Riesen K**. Structural pattern recognition with graph edit distance. Springer Publishing Company, Incorporated, 2016
- 15 **Barrett T**, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, Marshall KA, Phillippy KH, Sherman PM, Holko M, Yefanov A, Lee H, Zhang N, Robertson CL, Serova N, Davis S, Soboleva A. NCBI GEO: archive for functional genomics data sets--update. *Nucleic Acids Res* 2013; **41**: D991-D995 [PMID: 23193258 DOI: 10.1093/nar/gks1193]
- 16 **Lu TP**, Tsai MH, Lee JM, Hsu CP, Chen PC, Lin CW, Shih JY, Yang PC, Hsiao CK, Lai LC, Chuang EY. Identification of a novel biomarker, SEMA5A, for non-small cell lung carcinoma in nonsmoking women. *Cancer Epidemiol Biomarkers Prev* 2010; **19**: 2590-2597 [PMID: 20802022 DOI: 10.1158/1055-9965.EPI-10-0332]
- 17 **Gautier L**, Cope L, Bolstad BM, Irizarry RA. affy--analysis of Affymetrix GeneChip data at the probe level. *Bioinformatics* 2004; **20**: 307-315 [PMID: 14960456 DOI: 10.1093/bioinformatics/btg405]
- 18 **Lustgarten JL**, Gopalakrishnan V, Grover H, Visweswaran S. Improving classification performance with discretization on biomedical datasets. *AMIA Annu Symp Proc* 2008; 445-449 [PMID: 18999186]
- 19 **Lustgarten JL**, Visweswaran S, Gopalakrishnan V, Cooper GF. Application of an efficient Bayesian discretization method to biomedical data. *BMC Bioinformatics* 2011; **12**: 309 [PMID: 21798039 DOI: 10.1186/1471-2105-12-309]
- 20 **Bethune G**, Bethune D, Ridgway N, Xu Z. Epidermal growth factor receptor (EGFR) in lung cancer: an overview and update. *J Thorac Dis* 2010; **2**: 48-51 [PMID: 22263017]
- 21 **Shigematsu H**, Lin L, Takahashi T, Nomura M, Suzuki M, Wistuba II, Fong KM, Lee H, Toyooka S, Shimizu N, Fujisawa T, Feng Z, Roth JA, Herz J, Minna JD, Gazdar AF. Clinical and biological features associated with epidermal growth factor receptor gene mutations in lung cancers. *J Natl Cancer Inst* 2005; **97**: 339-346 [PMID: 15741570 DOI: 10.1093/jnci/dji055]
- 22 **Quinlan JR**. C4. 5: programs for machine learning. Elsevier; 2014: 58-60
- 23 **Cohen WW**. Fast effective rule induction. Machine Learning Proceedings 1995. Proceedings of the Twelfth International Conference on Machine Learning, Tahoe City, California, July 9-12, 1995: 115-123 [DOI: 10.1016/B978-1-55860-377-6.50023-2]
- 24 **Frank E**, Witten IH. Generating accurate rule sets without global optimization. Machine Learning. Fifteenth International Conference 1998: 144-151 [PMID: 9649111]
- 25 **Breiman L**. Random forests. *Machine Learning* 2001; **45**: 5-32 [DOI: 10.1023/A:1010933404324]
- 26 **John GH**, Langley P, editors. Estimating continuous distributions in Bayesian classifiers. Proceedings of the Eleventh conference on Uncertainty in artificial intelligence, 1995; Morgan Kaufmann Publishers Inc., 2013: 338-345
- 27 **Platt JC**. Fast training of support vector machines using sequential minimal optimization. MIT Press Cambridge, MA, USA, 1999: 185-208 [PMID: 10584633]
- 28 **Frank E**, Hall M, Witten IH. The WEKA Workbench. Online Appendix for "Data Mining: Practical Machine Learning Tools and Techniques": Morgan Kaufmann, 2016

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Retrospective Study

FOLFIRI3-aflibercept in previously treated patients with metastatic colorectal cancer

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Author contributions: Carola C, Barlet J and Chibaudel B completed the data collection and prepared the manuscript; Carola C, de Gramont A and Chibaudel B analyzed the data; all authors had read and approved manuscript.

Institutional review board statement: This study was reviewed and approved by the Institutional Review Committee in Cancer Research (IRCCR) of Franco-British Hospital (FBI). Personal and filiation data including identity of every patient was protected with an added code in the Excel table. This is a retrospective case series that did not have any activity or contact with the patients.

Informed consent statement: Patients were not required to give informed consent to the study because the analysis used anonymous clinical data that were obtained after each patient agreed to treatment.

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Abstract

AIM

To evaluate the efficacy and safety of the modified FOLFIRI3-aflibercept as second-line therapy in patients with metastatic colorectal cancer.

METHODS

This is a retrospective multicenter cohort, evaluating the efficacy and safety of the association of aflibercept with FOLFIRI3 (day 1: aflibercept 4 mg/kg, folinic acid 400 mg/m², irinotecan 90 mg/m², 5-fluorouracil infusion 2400 mg/m² per 46 h; day 3: irinotecan 90 mg/m²) in patients with previously treated metastatic colorectal cancer. The primary endpoint was overall response rate (ORR). Secondary endpoints were disease control rate (DCR), progression-free survival (PFS), overall survival (OS), and safety.

RESULTS

Among 74 patients treated in four French centers, nine were excluded due to prior use of aflibercept ($n = 3$), more than one prior treatment line in irinotecan-naïve patients ($n = 3$), and inadequate liver function ($n = 3$). In the "irinotecan-naïve" patients ($n = 30$), ORR was 43.3% and DCR was 76.7%. Median PFS and OS were 11.3 mo (95%CI: 6.1-29.0) and 17.0 mo (95%CI: 13.0-17.3), respectively. The most common ($> 5\%$) grade 3-4 adverse events were diarrhea (37.9%), neutropenia (14.3%), stomatitis and anemia (10.4%), and hypertension (6.7%). In the "pre-exposed irinotecan" patients ($n = 35$), 20 (57.1%) received ≥ 2 prior lines of treatment. ORR was 34.3% and DCR was 60.0%. Median PFS and OS were 5.7 mo (95%CI: 3.9-10.4) and 14.3 mo (95%CI: 12.8-19.5), respectively.

CONCLUSION

Minimally modified FOLFIRI has improvement dramatically the FOLFIRI3-aflibercept efficacy, whatever prior use of irinotecan. A prospective randomized trial is warranted to compare FOLFIRI-aflibercept to FOLFIRI3-aflibercept.

Key words: Chemotherapy; Irinotecan; Aflibercept; Second-line; Colorectal cancer

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Core tip: Results obtained in this retrospective study show that minimally modified FOLFIRI has improvement dramatically the efficacy of the FOLFIRI3-aflibercept combination with high response rates and survivals in patients with previously treated metastatic colorectal cancer, whatever prior use of irinotecan. A prospective randomized trial is planned to compare FOLFIRI-aflibercept to FOLFIRI3-aflibercept.

Carola C, Ghiringhelli F, Kim S, André T, Barlet J, Bengrine-Lefevre L, Marijon H, Garcia-Larnicol ML, Borg C, Dainese L, Steuer N, Richa H, Benetkiewicz M, Larsen AK, de Gramont A, Chibaudel B. FOLFIRI3-aflibercept in previously treated patients with metastatic colorectal cancer. *World J Clin Oncol* 2018; 9(5): 110-118 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i5/110.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i5.110>

INTRODUCTION

Standard second-line therapy in patients with previously treated metastatic colorectal cancer (mCRC) is doublet fluoropyrimidine-based chemotherapy with either irinotecan (FOLFIRI) or oxaliplatin (FOLFOX), depending on the regimen used in first-line, in association with antiangiogenic agents (e.g., bevacizumab, aflibercept, ramucirumab) or anti-EGFR agents in absence of RAS tumor gene mutation (e.g., cetuximab, panitumumab)^[1-8].

The standard FOLFIRI regimen was optimized by splitting the dose of irinotecan on day 1 [half dose before 5-fluorouracil (5-FU)] and day 3 (half dose after 5-FU) in the so-called FOLFIRI3 regimen^[9]. Drugs and doses are similar to FOLFIRI, except for suppression of the 5-FU bolus. The response rate was higher than that reported for FOLFIRI^[9,10]. Based on these results, FOLFIRI3 became the second-line regimen of choice in some centers. Adding bevacizumab to FOLFIRI3 has shown promising efficacy results in two prior retrospective trials [response rate 22% and 35%, median progression-free survival (PFS) 7.0 and 6.2 mo, median overall survival (OS) 13.0 and 10.8 mo, respectively]^[11,12]. The addition of aflibercept to FOLFIRI in patients with pretreated mCRC increased response rate from 11% to 20% and improved median PFS from 4.7 to 6.9 mo [hazard ratio (HR) = 0.76] and median OS from 12.1 to 13.5 mo (HR = 0.82)^[4]. Aflibercept was approved by the Food and Drug Administration on August 3, 2012 and by the European Medicines Agency on February 1, 2013 in combination with FOLFIRI for the treatment of patients with mCRC resistant to an oxaliplatin-containing regimen^[13]. Based on the VELOUR study results and non-randomized FOLFIRI3 studies, we retrospectively analyzed the safety and efficacy of the FOLFIRI3-aflibercept combination as second or later-line therapy in patients with mCRC.



Figure 1 Comparison of the FOLFIRI-1 and FOLFIRI-3 schedules.

MATERIALS AND METHODS

This study was a retrospective, multicenter cohort, conducted in four French institutions (Centre Georges François Leclerc, Franco-British Hospital, University Hospital Besançon, and Saint-Antoine University Hospital) from September 2014 to December 2016. The main objective of this study was to evaluate the efficacy and safety profile of the aflibercept-FOLFIRI3 combination.

Population

All patients with previously treated mCRC and with FOLFIRI3-aflibercept administered from September 2014 to December 2016 were included. During the inclusion period, the decision to give FOLFIRI3-aflibercept to each patient or another treatment regimen was at physician's discretion. Prior use of bevacizumab was allowed, but prior exposure to aflibercept was not permitted. Patients were divided into two subgroups depending on the prior use of irinotecan and the number of previous treatment lines for metastatic disease: (1) "irinotecan-naïve" population including patients with no more than one prior line of treatment for metastatic disease; and (2) the "irinotecan pre-exposed" population including patients for whom the number of prior treatment lines for metastatic disease was not restricted.

Treatment administration

Treatment cycles were given intravenously every 14 d, as follows: Aflibercept 4 mg/kg over 1-h infusion (day 1), folinic acid 400 mg/m² over 2-h infusion (day 1), irinotecan 90 mg/m² over 60-90 min infusion (day 1), followed by continuous 5-FU 2400 mg/m² as a 46-h infusion (days 1 to 3), then irinotecan 90 mg/m²

over 60-90 min infusion (day 3; Figure 1). Treatment information (date of treatment, doses) was collected using CHIMIO® 5.4 (Computer Engineering, Paris, France) or BPC (GCS Emosist, Région Franche-Comté, France) softwares.

Endpoints

Treatment efficacy was evaluated with tumor response, PFS, and OS. The objective response rate (ORR) was defined as the proportion of patients having either complete response (CR) or partial response (PR) according to RECIST version 1.1. The best ORR was defined as the best response recorded from the start of treatment until progressive disease (PD). Disease control rate (DCR) was the sum of ORR and stable disease (SD). PFS was defined as the time from the date of starting treatment to the date of progression or death (from any cause). OS was defined as the time from the date of starting treatment to the date of patient death (from any cause) or to the last date the patient was known to be alive. Toxicity was evaluated according to the United States National Cancer Institute's Common Terminology Criteria for Adverse Events (NCI-CTCAE) version 4.03.

Statistical analysis

Follow-up and survival were estimated using the reverse Kaplan-Meier method and the Kaplan-Meier method, respectively, and were described using median with 95% confidence interval (CI). Qualitative variables were described using percent and means and continuous variables using medians (minimum-maximum). The cut-off date for statistical analysis was June 15, 2017. The final analysis was performed on the irinotecan-naïve and irinotecan pre-exposed

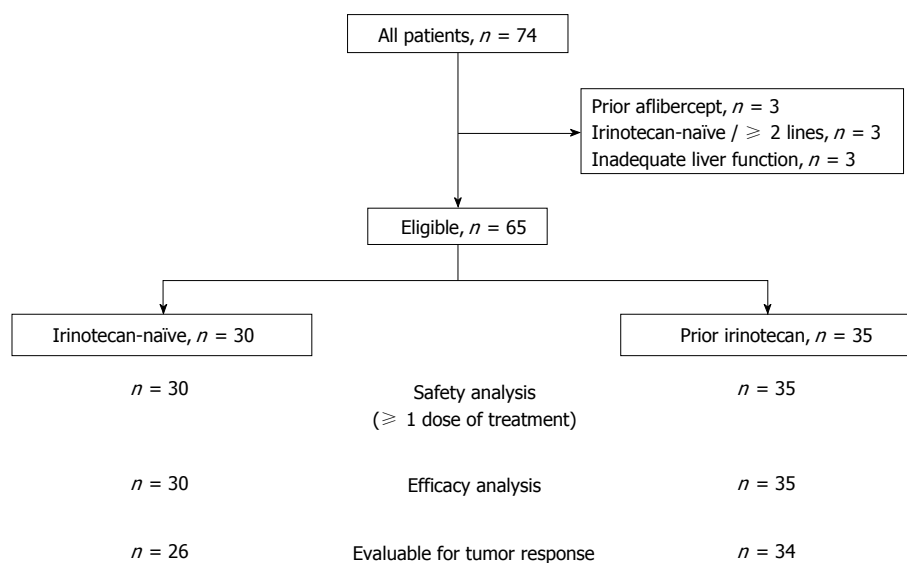


Figure 2 Flow diagram.

populations.

RESULTS

A total of 74 patients were treated (Figure 2). Nine patients were excluded from the analysis due to: Prior use of aflibercept ($n = 3$), more than one prior line of treatment in irinotecan-naïve patients ($n = 3$), or inadequate liver function (pretreatment alkaline phosphatase level $> 7 \times$ upper normal limit, $n = 3$). Thirty patients did not receive prior irinotecan (the irinotecan-naïve population) and 35 were previously exposed to irinotecan (the pre-exposed population).

Overall, 25 (38.5%) patients had an Eastern Cooperative Oncology Group performance status 0. The mean age was 63.1 years (range: 31.9-82.1); 23 (35.4%) had a single metastatic site, 49 (75.4%) had *RAS* mutated tumors, and two (3.1%) had *BRAF* mutated tumors. Prior use of bevacizumab and anti-EGFR were reported in 47 (72.3%) and 6 (9.2%) patients, respectively.

In the irinotecan-naïve population, five patients did not receive first-line therapy for metastatic disease ($n = 4$, early relapse after FOLFOX adjuvant therapy or $n = 1$, radiochemotherapy). In irinotecan pre-exposed population, 20 (57.1%) patients received more than two prior lines of treatment. Various irinotecan regimens (FOLFIRINOX, $n = 21$; FOLFIRI, $n = 10$; FOLFIRI3, $n = 4$) were previously given. The portion of patients with increased level of lactate dehydrogenase in the irinotecan pre-exposed population was higher than in that with the irinotecan-naïve patients (56.0% vs 17.6%; $P = 0.027$; Table 1).

Treatment exposure

In the irinotecan-naïve population, chemotherapy drugs (irinotecan and 5-FU) were given at standard dose in 12 (40.0%) patients. A lower dose of irinotecan and

5-FU were given in 15 (50.0%) and 4 (13.3%) patients, respectively. The median number of cycles was 8 (range: 1-19), and the median treatment duration was 3.7 mo (95%CI: 2.4-5.7). Dose reductions during treatment were performed in 7 (23.3%), 13 (43.3%), and 6 (20.0%) patients for aflibercept, irinotecan, and 5-FU, respectively. Granulocyte colony-stimulating factor (G-CSF) was given as primary prophylaxis in 14 (46.7%) patients and as secondary prevention in 3 (10.0%) patients. Erythropoietin was used in 5 (17.9%) patients. At the time of analysis, the treatment was still ongoing in 2 patients. The main reasons for stopping therapy were the occurrence of a limiting adverse event in 14 (46.7%) patients (diarrhea, $n = 8$; bleeding, $n = 1$; bowel perforation, $n = 1$; asthenia, $n = 1$; other, $n = 3$) or progression in 11 (36.7%) patients.

In the irinotecan pre-exposed population, chemotherapy drugs (irinotecan and 5-FU) were given at standard dose in 22 (62.9%) patients. A lower dose of irinotecan and 5-FU were given in 10 (28.6%), and 6 (17.1%) patients, respectively. The median number of cycles was 6 (range: 1-20), and the median treatment duration was 3.5 mo (95%CI: 2.1-5.6). Dose reductions during treatment were performed in 5 (14.3%), 9 (25.7%), and 7 (20.0%) patients for aflibercept, irinotecan, and 5-FU, respectively. G-CSF was given as primary prophylaxis in 13 (37.1%) patients and as secondary prevention in 2 (5.7%) patients. Erythropoietin was used in 3 (8.6%) patients. At the time of analysis, the treatment was still ongoing in 5 patients. The main reasons for stopping therapy were disease progression in 22 (62.9%) patients and the occurrence of limiting adverse events in 3 (8.6%) patients (diarrhea, $n = 2$; skin reactions, $n = 1$).

Response rate

In the irinotecan-naïve population, 4 (13.3%) patients

Table 1 Patient characteristics *n* (%)

Characteristics	Irinotecan-naïve cohort	Irinotecan-pre-exposed cohort	<i>P</i> -value
Age (yr)			0.793
< 70	21 (70.0)	23 (65.7)	
≥ 70	9 (30.0)	12 (34.3)	
Sex			0.623
Male	16 (53.3)	21 (60.0)	
Female	14 (46.7)	14 (40.0)	
ECOG PS			0.227
0	14 (46.7)	11 (31.4)	
1	13 (43.3)	15 (42.9)	
2	3 (10.0)	9 (25.7)	
Time to metastasis			0.314
Metachronous	9 (30.0)	15 (42.9)	
Synchronous	21 (70.0)	20 (57.1)	
No. of metastatic sites			0.118
1	14 (46.7)	9 (25.7)	
> 1	16 (53.3)	26 (74.3)	
No. of prior lines for metastatic disease			-
0	5 (16.7)	0 (0.0)	
1	25 (83.3)	15 (42.9)	
> 1	-	20 (57.1)	
Prior drug exposure			0.699
Oxaliplatin	29 (96.7)	35 (100.0)	
Bevacizumab	18 (60.0)	29 (82.9)	
Anti-EGFR	2 (6.7)	4 (11.4)	

ECOG PS: Eastern Cooperative Oncology Group performance status.

Table 2 Summary of the efficacy results

	Irinotecan-naïve, <i>n</i> = 30			Prior irinotecan, <i>n</i> = 35		
	<i>n</i>	% (ITT)	% (evaluable)	<i>n</i>	% (ITT)	% (evaluable)
Response rate						
CR	0	0	0	1	2.8	2.9
PR	13	43.3	50	11	31.4	32.4
SD	10	33.3	38.5	9	25.7	26.5
PD	3	10	11.5	13	37.1	38.2
NE	4	13.3	-	1	2.8	-
ORR	13	43.3	50	12	34.3	35.3
DCR	23	76.7	88.5	21	60	61.8
Survivals						
	median, mo	95%CI		median, mo	95%CI	
PFS	11.3	6.1-29.0		5.7	3.9-10.4	
OS	17.0	13.0-17.3		14.3	12.8-19.5	

RR: Response rate; ITT: Intention-to-treat; CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease; NE: Not evaluable; ORR: Objective response rate; DCR: Disease control rate; PFS: Progression-free survival; OS: Overall survival.

were not evaluated for tumor response due to an early stop for limiting toxicity. ORR was reported in 13 patients [43.3%, intention-to-treat (ITT); 50.0%, evaluable patients] without CR, and DCR was reported in 23 patients (76.7%, ITT; 88.5%, evaluable patients). Three (10.0%) patients had PD at the first tumor evaluation (Table 2).

In the irinotecan pre-exposed population, one patient was not evaluable for tumor response (switch to intra-arterial chemotherapy after 2 treatment cycles). ORR was reported in 12 patients (34.3%, ITT; 35.3%, evaluable patients) including one CR, and

DCR was reported in 21 patients (60.0%, ITT; 61.8%, evaluable population). Thirteen (37.1%) patients had PD at the first tumor evaluation (Table 2). Among seven patients refractory to irinotecan, 1 (2.9%) had PR with FOLFIRI3-aflibercept, 2 (5.7%) had stable disease, 3 (8.6%) had PD, and 1 (2.6%) was not evaluable (Table 3).

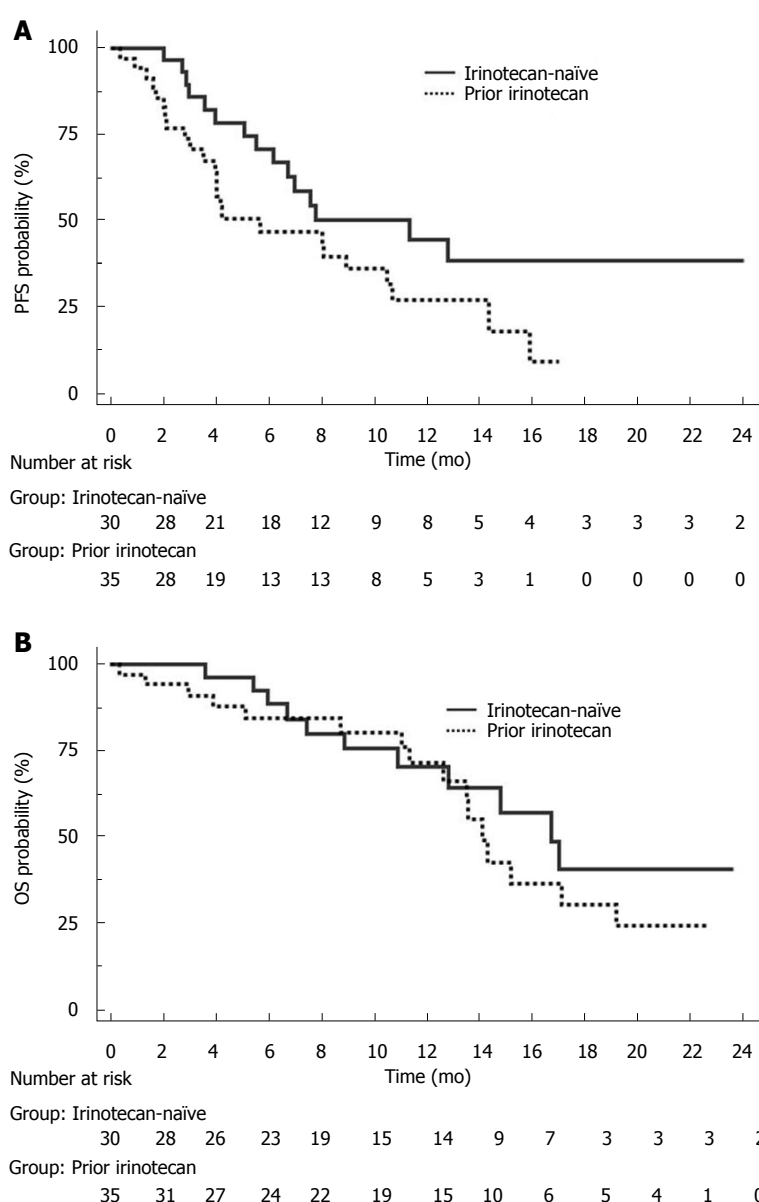
Survival

The median follow-up was 13.6 mo (95%CI: 9.6-17.7) in the irinotecan-naïve population and 14.2 mo (95%CI: 11.0-21.5) in the pre-exposed population (*P*

Table 3 Contingency table of tumor response with FOLFIRI3-aflibercept according to prior tumor response with irinotecan [*n* = 35, *n* (%)]

		FOLFIRI3-aflibercept				All
		CR/PR	SD	PD	NE	
Prior irinotecan-based regimen	CR/PR	5	3	7	0	15 (42.8)
	SD	4	4	2	0	10 (28.6)
	PD	1	2	3	1	7 (20.0)
	NE	2	0	1	0	3 (8.6)
	All	12 (34.3)	9 (25.7)	13 (37.1)	1 (2.8)	35

CR: Complete response; PR: Partial response; SD: Stable disease; PD: Progressive disease; NE: Not evaluable.

**Figure 3** Progression-free survival and overall survival according to prior exposure to irinotecan (*n* = 65). A: Progression-free survival; B: Overall survival.

= 0.692). In the irinotecan-naïve population, median PFS was 11.3 mo (95%CI: 6.1-29.0) and median OS was 17.0 mo (95%CI: 13.0-17.3; Figure 3A). A lower starting dose of irinotecan (< 90 mg/m²) did not impact PFS (*P* = 0.518) and OS (*P* = 0.311), but

decreased the incidence of severe neutropenia (0.0% vs 30.8%, respectively, *P* = 0.041). In the irinotecan pre-exposed population, median PFS was 5.7 mo (95%CI: 3.9-10.4) and median OS was 14.3 mo (95%CI: 12.8-19.5; Figure 3B).

Table 4 Selected ($\geq 5\%$) grade 3-4 adverse events (NCI CTCAE version 4.0) *n* (%)

SOC	PT	Irinotecan-naïve	Prior irinotecan	All
Any		17 (56.7)	15 (42.9)	32 (49.2)
Blood	Neutropenia	4 (13.3)	1 (2.9)	5 (7.7)
	Anemia	3 (10.0)	0 (0.0)	3 (4.6)
	Thrombocytopenia	0 (0.0)	0 (0.0)	0 (0.0)
Gastrointestinal	Nausea	2 (6.7)	0 (0.0)	2 (3.1)
	Vomiting	1 (3.3)	0 (0.0)	1 (1.5)
	Mucositis	3 (10.0)	3 (8.6)	6 (9.2)
	Diarrhea	11 (36.7)	9 (25.7)	20 (30.8)
Vascular	Hypertension	2 (6.7)	4 (11.4)	6 (9.2)

SOC: System Organ Class; PT: Preferred term.

Safety

In the irinotecan-naïve cohort, 17 (56.7%) patients experienced grade ≥ 3 toxicity (Table 4). The most common ($\geq 5\%$) grade 3-4 adverse events were diarrhea ($n = 11$, 36.7%), neutropenia ($n = 4$, 13.3%), anemia and mucositis ($n = 3$, 10.0%), and nausea and hypertension ($n = 2$, 6.7%). Any grade hemorrhage was reported in 4 (13.8%) patients, gastrointestinal perforation in 1 (3.3%) patient, and arterial thromboembolic event in 1 (3.3%) patient.

In the irinotecan pre-exposed cohort, 15 (42.9%) patients experienced grade ≥ 3 toxicity (Table 4). The most common ($\geq 5\%$) grade 3-4 adverse events were diarrhea ($n = 9$, 25.7%), hypertension ($n = 4$, 11.4%), and mucositis ($n = 3$, 8.6%).

Salvage surgery

Salvage surgery for metastatic disease was performed in 7 (10.0%) patients ($n = 4$, liver; $n = 1$, lung; $n = 1$, liver and lung; $n = 1$, peritoneum). A complete (R0) resection and liver pathological complete response were observed in all and one patient, respectively.

DISCUSSION

To our knowledge, this is the first report evaluating the FOLFIRI3-aflibercept combination in patients with previously treated mCRC. The response rate, which is a strong indicator of treatment efficacy, was unusually high not only in irinotecan-naïve patients (43%, ITT; 50%, evaluable), but also in irinotecan pre-exposed patients (34%, ITT, 35%, evaluable).

The median 11.3 mo PFS and median 17.0 mo OS in the irinotecan-naïve population receiving FOLFIRI3-aflibercept as second-line therapy compared favorably to the FOLFIRI-aflibercept combination in the pivotal phase III VELOUR study (response rate 19.8%, median 7.2 mo PFS, median 13.2 mo OS) and the FOLFIRI3 regimen without targeted agent (response rate 17%-23%, median 4-7 mo PFS, median 9-12 mo OS)^[4,9,10,14].

In the irinotecan pre-exposed population, patients received FOLFIRI3-aflibercept as salvage therapy.

Yet, median PFS and OS were 5.7 mo and 14.3 mo, respectively, and were comparable to figures observed in second-line trials^[8,15].

A high portion (27%) of patients had to stop the FOLFIRI3-aflibercept combination because of limiting toxicity, mainly diarrhea. Its frequency (38%) was twice as common as in the VELOUR study (19%), but in the same range as in previous studies using FOLFIRI3. It has been demonstrated that severe diarrhea induced by aflibercept is due to microscopic colitis, which can be managed successfully using oral budesonide and/or mesalamine treatment^[16,17]. Placental growth factor (PIGF) could play a role in the occurrence of diarrhea. The absence of PIGF blocks dextran sodium sulfate-induced colonic mucosal angiogenesis and increases mucosal hypoxia^[18,19]. Knockout of PIGF aggravates disease course in acute colitis^[20]. Neutropenia and stomatitis were at a lower incidence than in the FOLFIRI-aflibercept arm of the pivotal VELOUR study (13.3% vs 36.7%, neutropenia; 10.0% vs 13.8%, stomatitis), which can be explained by deletion of the 5-FU bolus in the FOLFIRI3 regimen and use of G-CSF in 49% of patients. In the irinotecan-naïve population, a lower starting dose of irinotecan (< 90 mg/m²) did not impact treatment efficacy, but decreased the incidence of severe neutropenia (0.0% vs 30.8%, $P = 0.041$).

The conversion to surgery of metastasis in second-line is another key finding that could modify the strategy in patients suitable for salvage surgery in case of response (sequential doublets versus triplets). The main limitation of this study is the retrospective design with a low number of patients. In conclusion, the combination of aflibercept and FOLFIRI3 in our study shows the encouraging efficacy results with high response rates and longer survivals in patients with previously treated mCRC, whatever the prior exposition to irinotecan. A randomized trial is warranted to compare FOLFIRI-aflibercept to FOLFIRI3-aflibercept.

ARTICLE HIGHLIGHTS

Research background

FOLFIRI3 is the second-line regimen of choice in patients with previously treated mCRC in some centers. Adding bevacizumab to FOLFIRI3 has shown

promising efficacy results in two prior retrospective trials. The addition of aflibercept to FOLFIRI in patients with pretreated mCRC increased response rate from 11% to 20% and improved median PFS from 4.7 to 6.9 mo.

Research motivation

The phase III VELOUR and non-randomized FOLFIRI3 studies results provide a backbone for our study.

Research objectives

The main objective of the study is to evaluate the safety and efficacy of the FOLFIRI3-aflibercept combination as second or later-line therapy in patients with mCRC.

Research methods

Patients with previously treated mCRC were given the aflibercept-FOLFIRI3 combination and were divided into "irinotecan-naïve" population including patients with no more than one prior line of treatment for metastatic disease, and the "irinotecan pre-exposed" population including patients for whom the number of prior treatment lines for metastatic disease was not restricted. The primary endpoint was overall response rate (ORR). Secondary endpoints were disease control rate, progression-free survival (PFS), overall survival (OS), and safety. Toxicity was evaluated according to the United States National Cancer Institute's Common Terminology Criteria for Adverse Events (NCI-CTCAE) version 4.03.

Research results

Minimally modified FOLFIRI has improved dramatically the efficacy of the FOLFIRI3-aflibercept combination with high response rates (43% in irinotecan-naïve patients and 34% in irinotecan pre-exposed patients) and survivals (median PFS: 11.3 mo, OS: 17.0 mo and PFS: 5.7 mo and OS: 14.3 mo, respectively) in patients with previously treated mCRC, whatever prior use of irinotecan.

Research conclusions

The combination of aflibercept and FOLFIRI3 shows encouraging efficacy results in patients with previously treated mCRC.

Research perspectives

A prospective randomized trial is planned to compare FOLFIRI3-aflibercept to FOLFIRI3-aflibercept.

REFERENCES

- 1 Tournigand C, André T, Achille E, Lledo G, Flesch M, Mery-Mignard D, Quinaux E, Couteau C, Buyse M, Ganem G, Landi B, Colin P, Louvet C, de Gramont A. FOLFIRI followed by FOLFOX6 or the reverse sequence in advanced colorectal cancer: a randomized GERCOR study. *J Clin Oncol* 2004; **22**: 229-237 [PMID: 14657227 DOI: 10.1200/jco.2004.05.113]
- 2 Giantonio BJ, Catalano PJ, Meropol NJ, O'Dwyer PJ, Mitchell EP, Alberts SR, Schwartz MA, Benson AB 3rd; Eastern Cooperative Oncology Group Study E3200. Bevacizumab in combination with oxaliplatin, fluorouracil, and leucovorin (FOLFOX4) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. *J Clin Oncol* 2007; **25**: 1539-1544 [PMID: 17442997 DOI: 10.1200/JCO.2006.09.6305]
- 3 Bennouna J, Sastre J, Arnold D, Österlund P, Greil R, Van Cutsem E, von Moos R, Viéitez JM, Bouché O, Borg C, Steffens CC, Alonso-Orduña V, Schlichting C, Reyes-Rivera I, Bendahmane B, André T, Kubicka S; ML18147 Study Investigators. Continuation of bevacizumab after first progression in metastatic colorectal cancer (ML18147): a randomised phase 3 trial. *Lancet Oncol* 2013; **14**: 29-37 [PMID: 23168366 DOI: 10.1016/S1470-2045(12)70477-1]
- 4 Van Cutsem E, Tabernero J, Lakomy R, Prenen H, Prausová J, Macarulla T, Ruff P, van Hazel GA, Moiseyenko V, Ferry D, McKendrick J, Polikoff J, Tellier A, Castan R, Allegra C. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. *J Clin Oncol* 2012; **30**: 3499-3506 [PMID: 22949147 DOI: 10.1200/JCO.2012.42.8201]
- 5 Tabernero J, Yoshino T, Cohn AL, Obermannova R, Bodoky G, Garcia-Carbonero R, Ciuleanu TE, Portnoy DC, Van Cutsem E, Grothey A, Prausová J, Garcia-Alfonso P, Yamazaki K, Clingan PR, Lonardi S, Kim TW, Simms L, Chang SC, Nasroulah F; RAISE Study Investigators. Ramucirumab versus placebo in combination with second-line FOLFIRI in patients with metastatic colorectal carcinoma that progressed during or after first-line therapy with bevacizumab, oxaliplatin, and a fluoropyrimidine (RAISE): a randomised, double-blind, multicentre, phase 3 study. *Lancet Oncol* 2015; **16**: 499-508 [PMID: 25877855 DOI: 10.1016/S1470-2045(15)70127-0]
- 6 Peeters M, Price TJ, Cervantes A, Sobrero AF, Ducreux M, Hotko Y, André T, Chan E, Lordick F, Punt CJ, Strickland AH, Wilson G, Ciuleanu TE, Roman L, Van Cutsem E, Tzekova V, Collins S, Oliner KS, Rong A, Gansert J. Randomized phase III study of panitumumab with fluorouracil, leucovorin, and irinotecan (FOLFIRI) compared with FOLFIRI alone as second-line treatment in patients with metastatic colorectal cancer. *J Clin Oncol* 2010; **28**: 4706-4713 [PMID: 20921462 DOI: 10.1200/JCO.2009.27.6055]
- 7 Mishima H, Oba K, Sakamoto J, Muro K, Yoshino T, Hyodo I, Maehara Y. FOLFIRI plus bevacizumab 5 mg/kg versus 10 mg/kg as second-line therapy in patients with metastatic colorectal cancer who have failed first-line bevacizumab plus oxaliplatin-based therapy: a randomized phase III study (EAGLE Study). *Jpn J Clin Oncol* 2012; **42**: 134-138 [PMID: 22167662 DOI: 10.1093/jco/hyr180]
- 8 Mabro M, Louvet C, André T, Carola E, Gilles-Amar V, Artru P, Krulik M, de Gramont A; GERCOR. Bimonthly leucovorin, infusion 5-fluorouracil, hydroxyurea, and irinotecan (FOLFIRI-2) for pretreated metastatic colorectal cancer. *Am J Clin Oncol* 2003; **26**: 254-258 [PMID: 12796595 DOI: 10.1097/01.COC.0000020581.59835.7A]
- 9 Mabro M, Artru P, André T, Flesch M, Maindault-Goebel F, Landi B, Lledo G, Plantade A, Louvet C, de Gramont A. A phase II study of FOLFIRI-3 (double infusion of irinotecan combined with LV5FU) after FOLFOX in advanced colorectal cancer patients. *Br J Cancer* 2006; **94**: 1287-1292 [PMID: 16622455 DOI: 10.1038/sj.bjc.6603095]
- 10 Chibaudel B, Maindault-Goebel F, Bachet JB, Louvet C, Khalil A, Dupuis O, Hammel P, Garcia ML, Bennamoun M, Brusquant D, Tournigand C, André T, Arbaud C, Larsen AK, Wang YW, Yeh CG, Bonnetain F, de Gramont A. PEPOL: a GERCOR randomized phase II study of nanoliposomal irinotecan PEP02 (MM-398) or irinotecan with leucovorin/5-fluorouracil as second-line therapy in metastatic colorectal cancer. *Cancer Med* 2016; **5**: 676-683 [PMID: 26806397 DOI: 10.1002/cam4.635]
- 11 Ghiringhelli F, Vincent J, Guio B, Chauffert B, Ladoire S. Bevacizumab plus FOLFIRI-3 in chemotherapy-refractory patients with metastatic colorectal cancer in the era of biotherapies. *Invest New Drugs* 2012; **30**: 758-764 [PMID: 21057973 DOI: 10.1007/s10637-010-9575-3]
- 12 Bourges O, Chibaudel B, Bengrine-Lefevre L, Afchain P, Tournigand C, Perez-Staub N, Maindault-Goebel F, Larsen AK, Louvet C, de Gramont A. FOLFIRI-3 + bevacizumab after the first-line in metastatic colorectal cancer. Colon and Rectum. American Society of Clinical Oncology Gastrointestinal Cancers (ASCO-GI) Symposium, 2009: Abstract 417
- 13 European Medicines Agency (EMA). Zaltrap® (Aflibercept). Available from: URL: http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Summary_for_the_public/human/002532/WC500139486.pdf
- 14 Bidard FC, Tournigand C, André T, Mabro M, Figer A, Cervantes A, Lledo G, Bengrine-Lefevre L, Maindault-Goebel F, Louvet C, de Gramont A. Efficacy of FOLFIRI-3 (irinotecan D1,D3 combined with LV5-FU) or other irinotecan-based regimens in oxaliplatin-pretreated metastatic colorectal cancer in the GERCOR OPTIMOX1 study. *Ann Oncol* 2009; **20**: 1042-1047 [PMID: 19153116 DOI: 10.1093/annonc/mdn311]

- 10.1093/annonc/mdn730]
- 15 **Chong DQ**, Manalo M, Imperial M, Teo P, Yong G, Ng M, Tan IB, Choo SP, Chua C. Safety and efficacy of aflibercept in combination with fluorouracil, leucovorin and irinotecan in the treatment of Asian patients with metastatic colorectal cancer. *Asia Pac J Clin Oncol* 2016; **12**: 275-283 [PMID: 27075236 DOI: 10.1111/ajco.12496]
 - 16 **Ghiringhelli F**, Vincent J, Beltjens F, Bengrine L, Ladoire S. Fluorouracil, leucovorin and irinotecan associated with aflibercept can induce microscopic colitis in metastatic colorectal cancer patients. *Invest New Drugs* 2015; **33**: 1263-1266 [PMID: 26490656 DOI: 10.1007/s10637-015-0295-6]
 - 17 **Miehlke S**, Madisch A, Kupcinskas L, Petrauskas D, Böhm G, Marks HJ, Neumeyer M, Nathan T, Fernández-Bañares F, Greinwald R, Mohrbacher R, Vieth M, Bonderup OK; BUC-60/COC Study Group. Budesonide is more effective than mesalamine or placebo in short-term treatment of collagenous colitis. *Gastroenterology* 2014; **146**: 1222-1230.e1-2 [PMID: 24440672 DOI: 10.1053/j.gastro.2014.01.019]
 - 18 **Zhou Y**, Tu C, Zhao Y, Liu H, Zhang S. Placental growth factor enhances angiogenesis in human intestinal microvascular endothelial cells via PI3K/Akt pathway: Potential implications of inflammation bowel disease. *Biochem Biophys Res Commun* 2016; **470**: 967-974 [PMID: 26775845 DOI: 10.1016/j.bbrc.2016.01.073]
 - 19 **Kim KJ**, Cho CS, Kim WU. Role of placenta growth factor in cancer and inflammation. *Exp Mol Med* 2012; **44**: 10-19 [PMID: 22217448 DOI: 10.3858/emmm.2012.44.1.023]
 - 20 **Hindryckx P**, Waeytens A, Laukens D, Peeters H, Van Huysse J, Ferdinande L, Carmeliet P, De Vos M. Absence of placental growth factor blocks dextran sodium sulfate-induced colonic mucosal angiogenesis, increases mucosal hypoxia and aggravates acute colonic injury. *Lab Invest* 2010; **90**: 566-576 [PMID: 20142801 DOI: 10.1038/labinvest.2010.37]

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Meyer J, Buchs NC, Ris F

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Risk of colorectal cancer in patients with diverticular disease

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Abstract

Colorectal cancer constitutes an important burden on the healthcare system. Screening at-risk populations to reduce colorectal cancer-related morbidity and mortality has become part of good clinical practice. However, recommendations regarding subgroups of patients with diverticular disease are subject to controversy.

Herein, we review the most recent literature regarding the prevalence of colorectal cancer in patients with diverticular disease, diverticulitis and uncomplicated diverticulitis.

The recent literature does not identify diverticular disease as a long-term risk factor for colorectal cancer. However, the risk of colorectal cancer is increased in the short-term period after hospitalization related to diverticular disease. According to a recent systematic review and meta-analysis, the prevalence of colorectal cancer is 1.6% in patients with acute diverticulitis who underwent colonoscopy. The risk of having colorectal cancer after an episode of acute diverticulitis is 44-fold higher than that of an age- and gender-adjusted reference population. Despite lower among patients with uncomplicated episode, the risk of colorectal cancer remains 40-fold higher in that subpopulation than that in the reference population.

To conclude, the recent literature describes an increased risk of colorectal cancer among patients with acute diverticulitis compared to the reference population. Colonoscopy is therefore recommended in patients with diverticulitis to exclude colorectal cancer.

Key words: Diverticulosis; Diverticulitis; Colonoscopy; Screening; Tumor; Risk factor

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Core tip: The present manuscript reviews the current literature reporting on the prevalence of colorectal cancer

in patients with diverticular disease. The prevalence of colorectal cancer among subgroups of patients with diverticulitis and uncomplicated diverticulitis is discussed, with the objective of providing recommendations for colorectal cancer screening.

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INTRODUCTION

In 2018, colorectal cancer (CRC) will represent 8.1% of estimated cancer cases and will account for 15.6% of estimated cancer deaths in the United States^[1]. At-risk populations are screened to reduce CRC-related morbidity and mortality and to reduce healthcare costs.

CRC IN DIVERTICULAR DISEASE

Some authors consider diverticular disease a risk factor for CRC. For instance, Stefansson *et al.*^[2] reported a standardized incidence ratio (SIR) of 5.8 (95%CI: 4.6-7.3) over a 2-year period for CRC in patients with diverticular disease compared to controls without diverticular disease. That risk persisted for 10 years for patients with left-sided disease. Later, Granlund *et al.*^[3] performed a case-control population-based study including 41037 CRC patients matched with 82074 controls. They identified that hospitalization for diverticular disease in the previous 6 mo was a risk factor for CRC with an odd ratio (OR) ranging between 22.75 (95%CI: 14.06-36.82) and 31.49 (95%CI: 19.00-52.21). Thereafter, that risk was not significantly different from that of the population without diverticular disease^[3]. More recently, Huang *et al.*^[4] followed 41359 Asian patients with diverticular disease and 165436 age- and gender-matched patients without diverticular disease. They found annual incidences of CRC of 0.62% and 0.15%, respectively, in these populations. Among patients with diverticular disease, the hazard ratio (HR) for CRC was 5.08 (95%CI: 4.19-4.91). After excluding the first year of follow-up, the adjusted HR dropped to 0.98 (95%CI: 0.85-1.13)^[4]. Therefore, both of these large-scale population-based studies, which included 2 geographically distinct populations, were distinct from the findings of Stefansson *et al.*^[2] by revealing an increased risk of CRC only during the short period of follow-up after hospitalization/diagnosis for diverticular disease. Although the identification of uncomplicated diverticular disease as a risk factor for CRC remains a subject of controversy^[5-8], we note that the usual reasons for diverticular disease-related hospitalization are diverticular bleeding and diverticulitis.

CRC IN DIVERTICULITIS

Diverticulitis accounts for more than 200000 hospital admissions per year in the United States^[9]. The prevalence of CRC in patients suffering from diverticulitis has remained poorly documented for decades, but has recently come under the spotlight. In 2013, Sharma *et al.*^[10] performed a systematic review and meta-analysis of colonoscopy results from 1970 patients with diverticulitis treated with nonsurgical management. They described a prevalence of CRC of 1.6% in that population^[10]. This prevalence was, however, not compared to the prevalence of a reference population. Moreover, selection bias for colonoscopy could not be excluded. To this end, our group compared the incidence of CRC in 506 patients with computed tomography-proven diverticulitis to that of an age- and gender-matched reference population. We found a one-year incidence of CRC of 1.9% in patients with diverticulitis, an incidence that was 44-fold higher (95%CI: 18.58-75.96) than that in the reference population^[11]. One year later, Grahnat *et al.*^[12] confirmed these results by describing a SIR of 20 (95%CI: 10.2-35.7) for sigmoid cancer in 890 patients with CRC compared to their reference population. We also note that Sharma *et al.*^[10] did not include recent population-based large-scale studies in their systematic review and meta-analysis. The population-based study of Huang *et al.*^[4] reported an annual incidence of CRC of 0.49% in the subgroup of 31216 Asian patients with diverticulitis, whereas Mortensen *et al.*^[13] reported that 977 of the 40496 patients hospitalized for diverticulitis were diagnosed with CRC within a year, for an annual incidence of 2.41%. In the latest study, diverticulitis constituted a risk factor for CRC, with an OR of 2.20 (95%CI: 2.08-2.32)^[13]. That increased risk was not identified in the Asian study [HR of 1.08 (95%CI: 0.91-1.28) for CRC among the subgroup of patients with diverticulitis]^[4].

Nevertheless, it seems reasonable to consider that the risk of CRC is increased in the first year after diagnosis of diverticulitis, especially in Western societies. Fifty-seven percent of CRC cases in the study by Mortensen *et al.*^[13] were diagnosed during that period. The risk of CRC might be increased due to difficulties in distinguishing CRC from diverticulitis based on computed tomography, resulting in an initial misdiagnosis, despite the contribution of information bias that cannot be excluded. Indeed, Mortensen *et al.*^[13] reported an increased rate of colonoscopy in the diverticulitis population (57%) when compared to the reference population (10%). Taken together, this evidence led professional societies to recommend performing colonoscopy after an episode of diverticulitis^[14,15].

Furthermore, we note that some authors reported a low prevalence of CRC among patients with uncomplicated diverticulitis and suggested exempting them from colonoscopy^[16-19]. According to the systematic review and meta-analysis by Sharma *et al.*^[10], the pre-

valence of CRC in these patients was 0.7%. However, in our study, we described the prevalence of CRC in patients with uncomplicated diverticulitis to be 40-fold higher than that in the reference population^[11]. Unfortunately, large-scale population-based studies have not distinguished uncomplicated diverticulitis from complicated diverticulitis. Therefore, evidence is lacking to provide recommendations for this subgroup of patients.

Some studies did not identify cases of CRC among young Asian or Hispanic patients with diverticulitis^[20,21]. Further, in their large-scale study, Huang *et al.*^[4] described a higher HR for CRC among old patients than among young patients. Moreover, in our study^[11] and the study by Grahnat *et al.*^[12], all CRC patients were older than 70 years. Additionally, the increased risk documented by our study was age-adjusted, confirming that the increased risk observed among diverticulitis patients was not only due to a global increase in CRC prevalence among older patients. Therefore, the prevalence of CRC seems to be lower among young patients than among old patients, but this remains to be confirmed by large-scale studies.

CONCLUSION

To conclude, the recent literature describes an increased risk of CRC among patients with acute diverticulitis compared to the reference age-matched population. Colonoscopy is therefore recommended in patients with diverticulitis to exclude CRC. However, the evidence is not convincing enough to determine whether subgroups of patients - those with an uncomplicated episode of diverticulitis and/or young patients - might be exempted from colonoscopy. Future studies are needed to determine the prevalence of CRC in patients with uncomplicated diverticulitis, looking more specifically at age groups and providing large-scale population-based comparisons with reference populations. A new systematic review and meta-analysis aggregating the most recent studies, including the latest population-based studies, is necessary. In addition, to our knowledge, no study provides recommendations for patients with a recent colonoscopy to exclude interval colorectal cancer. These questions are of critical importance to better identify at-risk patients in order to provide the best benefit/risk ratio for CRC screening in patients with diverticulitis.

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REFERENCES

- 1 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. *CA Cancer J Clin* 2018; **68**: 7-30 [PMID: 29313949 DOI: 10.3322/caac.21442]
- 2 Stefánsson T, Ekbom A, Sparén P, Pahlman L. Increased risk of left sided colon cancer in patients with diverticular disease. *Gut* 1993; **34**: 499-502 [PMID: 8491397 DOI: 10.1136/gut.34.4.499]
- 3 Granlund J, Svensson T, Granath F, Hjertqvist E, Ekbom A, Blomqvist P, Schmidt PT. Diverticular disease and the risk of colon cancer - a population-based case-control study. *Aliment Pharmacol Ther* 2011; **34**: 675-681 [PMID: 21790681 DOI: 10.1111/j.1365-2036.2011.04782.x]
- 4 Huang WY, Lin CC, Jen YM, Chang YJ, Hsiao CW, Yang MH, Lin CS, Sung FC, Liang JA, Kao CH. Association between colonic diverticular disease and colorectal cancer: a nationwide population-based study. *Clin Gastroenterol Hepatol* 2014; **12**: 1288-1294 [PMID: 24361412 DOI: 10.1016/j.cgh.2013.11.039]
- 5 Reichert MC, Lammert F. The genetic epidemiology of diverticulosis and diverticular disease: Emerging evidence. *United European Gastroenterol J* 2015; **3**: 409-418 [PMID: 26535118 DOI: 10.1177/2050640615576676]
- 6 Cooper GS, Xu F, Schluchter MD, Koroukian SM, Barnholtz Sloan JS. Diverticulosis and the risk of interval colorectal cancer. *Dig Dis Sci* 2014; **59**: 2765-2772 [PMID: 24927800 DOI: 10.1007/s10620-014-3246-8]
- 7 Morini S, Zullo A, Hassan C, Tomao S, Campo SM. Diverticulosis and colorectal cancer: between lights and shadows. *J Clin Gastroenterol* 2008; **42**: 763-770 [PMID: 18580497 DOI: 10.1097/MCG.0b013e31816200fb]
- 8 Ekbom A. Is diverticular disease associated with colonic malignancy? *Dig Dis* 2012; **30**: 46-50 [PMID: 22572684 DOI: 10.1159/000335708]
- 9 Masoomi H, Buchberg BS, Magno C, Mills SD, Stamos MJ. Trends in diverticulitis management in the United States from 2002 to 2007. *Arch Surg* 2011; **146**: 400-406 [PMID: 21173283 DOI: 10.1001/archsurg.2010.276]
- 10 Sharma PV, Eglinton T, Hider P, Frizelle F. Systematic review and meta-analysis of the role of routine colonic evaluation after radiologically confirmed acute diverticulitis. *Ann Surg* 2014; **259**: 263-272 [PMID: 24169174 DOI: 10.1097/SLA.0000000000000294]
- 11 Meyer J, Thomopoulos T, Usel M, Gjika E, Bouchardy C, Morel P, Ris F. The incidence of colon cancer among patients diagnosed with left colonic or sigmoid acute diverticulitis is higher than in the general population. *Surg Endosc* 2015; **29**: 3331-3337 [PMID: 25631117 DOI: 10.1007/s00464-015-4093-1]
- 12 Grahnat CJ, Hérard S, Ackzell A, Andersson RE. High Probability of an Underlying Colorectal Cancer Among Patients Treated for Acute Diverticulitis. A Population-Based Cohort Follow-Up Study. *World J Surg* 2016; **40**: 2283-2288 [PMID: 26956904 DOI: 10.1007/s00268-016-3480-7]
- 13 Mortensen LQ, Burcharth J, Andresen K, Pommergaard HC, Rosenberg J. An 18-Year Nationwide Cohort Study on The Association Between Diverticulitis and Colon Cancer. *Ann Surg* 2017; **265**: 954-959 [PMID: 27192351 DOI: 10.1097/SLA.0000000000001794]
- 14 Stollman N, Smalley W, Hirano I; AGA Institute Clinical Guidelines Committee. American Gastroenterological Association Institute Guideline on the Management of Acute Diverticulitis. *Gastroenterology* 2015; **149**: 1944-1949 [PMID: 26453777 DOI: 10.1053/j.gastro.2015.10.003]
- 15 Feingold D, Steele SR, Lee S, Kaiser A, Boushey R, Buie WD, Rafferty JF. Practice parameters for the treatment of sigmoid diverticulitis. *Dis Colon Rectum* 2014; **57**: 284-294 [PMID: 24509449 DOI: 10.1097/DCR.0000000000000075]
- 16 Westwood DA, Eglinton TW, Frizelle FA. Routine colonoscopy following acute uncomplicated diverticulitis. *Br J Surg* 2011; **98**: 1630-1634 [PMID: 21713756 DOI: 10.1002/bjs.7602]
- 17 Brar MS, Roxin G, Yaffe PB, Stanger J, MacLean AR, Buie WD. Colonoscopy following nonoperative management of uncomplicated diverticulitis may not be warranted. *Dis Colon Rectum* 2013; **56**: 1259-1264 [PMID: 24105001 DOI: 10.1097/DCR.0b013e3182a26bfd]
- 18 Sallinen V, Mentula P, Leppäniemi A. Risk of colon cancer after computed tomography-diagnosed acute diverticulitis: is routine colonoscopy necessary? *Surg Endosc* 2014; **28**: 961-966 [PMID: 24178863 DOI: 10.1007/s00464-013-3257-0]
- 19 Schout PJ, Spillenaar Bilgen EJ, Groenen MJ. Routine screening for

- colon cancer after conservative treatment of diverticulitis. *Dig Surg* 2012; **29**: 408-411 [PMID: 23171930 DOI: 10.1159/000345332]
- 20 **Chan DKH**, Tan KK. There Is No Role for Colonoscopy after Diverticulitis among Asian Patients Less than 50 Years of Age. *Gastrointest Tumors* 2017; **3**: 136-140 [PMID: 28611980 DOI: 10.1159/000446565]
- 21 **Syed U**, Companioni R, Bansal R, Alkhawam H, Walfish A. Diverticulitis in the Young Population: Reconsidering Conventional Recommendations. *Acta Gastroenterol Belg* 2016; **79**: 435-439 [PMID: 28209102]
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Basic Study

Lymphocyte subsets predictive value and possible involvement of human papilloma virus infection on breast cancer molecular subtypes

Andreína Fernandes, Adriana Pesci-Feltri, Isabel García-Fleury, Marco López, Vincent Guida, Marisol De Macedo, María Correnti

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Abstract

AIM

To detect human papilloma virus (HPV) presence and to characterize cellular immune response in breast cancer patients.

METHODS

A total of 74 women were included, of which 48 samples were from patients diagnosed with breast cancer and 26 patients with benign pathology of the breast. Molecular subtype classification was performed based on the immunohistochemical reports of the tumor piece. HPV genome detection and genotyping from fresh breast biopsies was performed using the INNO-LIPA HPV Genotyping Extra test (Innogenetics, Ghent, Belgium). CD3+, CD4+, CD8+ and natural killer (NK)+ cells levels from peripheral blood samples from patients with breast cancer and benign pathology were measured by flow cytometry.

RESULTS

Luminal A was the most frequent breast cancer molecular subtype (33.33%). HPV was detected in 25%

of the breast cancer patients, and genotype 18 was the most frequent in the studied population. The mean of CD3+, CD4+ and CD8+ subpopulations were decreased in patients with breast cancer, in relation to those with benign pathology, with a statistically significant difference in CD8+ values ($P = 0.048$). The mean of NK+ cells was increased in the benign pathology group. The average level of CD3+, CD4+, CD8+ and NK+ cells decreased as the disease progressed. HER2+ and Luminal B HER2+ tumors had the lowest counts of cell subsets. HPV breast cancer patients had elevated counts of cellular subsets.

CONCLUSION

Determining level changes in cellular subsets in breast cancer patients is a useful tool to evaluate treatment response.

Key words: Breast cancer; Human papilloma virus; Molecular subtypes; Immune response; T lymphocytes; NK cells

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Core tip: This work detected the presence of the human papilloma virus (HPV) genome in patients with breast cancer and measured the levels of cellular subsets as predictor factors. The viral genome was found in 25% of breast cancer cases, with the high-risk 18 genotype the most frequent. Luminal A tumors represented 33.33% of the sample. The average level of CD3+, CD4+, CD8+ and NK+ cells was decreased in cancer patients compared to the benign pathology group, while the reverse effect was observed in HPV positive patients.

Fernandes A, Pesci-Feltri A, García-Fleury I, López M, Guida V, De Macedo M, Correnti M. Lymphocyte subsets predictive value and possible involvement of human papilloma virus infection on breast cancer molecular subtypes. *World J Clin Oncol* 2018; 9(7): 123-132 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i7/123.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i7.123>

INTRODUCTION

With more than 408200 new cases and over 92000 deaths, breast cancer is the first cancer in the Americas, in terms of new cases, and the second in terms of women cancer deaths^[1]. In Venezuela, according to the Ministry of Popular Power for Health in 2012, breast cancer ranked first in cancer incidence with 5063 new cases and was the first cause of cancer death in the female population. This was followed by cervix cancer, with 2067 deaths, representing 22.88% of new diagnoses due to cancer and 18.25% of deaths caused by this pathology^[2].

The 13th Conference of St Gallen in 2013 established that it was unnecessary to perform genetic tests on each patient to classify molecular subtypes, which was proposed by Perou *et al.*^[3] in 2000, since histopathological results were comparable. The use of molecular diagnosis was necessary only in atypical behavior cases^[4]. Luminal A breast cancer cases are the most frequent and have good prognosis and response to hormonal treatment. Luminal B HER2- and Luminal B HER2+ breast cancers represent between 15% and 20% of breast tumors, and their response to hormonal therapy is less. HER2+ cancer accounts for 15%-20% of breast cancer subtypes, showing more aggressive biological and clinical behavior, with increased sensitivity to chemotherapy. The triple negative (TN) subtype represents from 8% to 37% of all breast cancers. These tumors are infiltrating ductal types characterized by a solid growth pattern, aggressive clinical behavior and high rate of metastasis to brain and lungs^[4,5]. Molecular subtypes are currently used as predictive factors among breast cancer patients^[6].

Nearly 50% of newly diagnosed breast cancer cases are related to hormonal factors; only 5% to 10% are related to genetic factors, although it is known that these greatly increase the risk of developing the disease. Several studies have determined the physiological, environmental and lifestyle factors related to the incidence of breast cancer, some of which are modifiable through preventive interventions^[1].

The non-genetic factors include: Age of diagnosis after 65 years, which is the most important risk factor, early menarche, menopause after 55-years-old, first live birth after 30-years-old, nulliparity, breast biopsies history, diagnosis of atypical hyperplasia by breast biopsy, obesity, alcohol consumption, hormone replacement therapy and excessive exposure to radiation. Other possible risk factors include high-in-fats and low-in-fiber diet, and little exercise^[7].

Approximately 18% of human cancers are caused by infectious factors, and it is recognized that breast cancer is strongly related to environmental factors, such as viruses, diet, radiation, among others^[8]. Human papilloma virus (HPV) infection distribution reports in breast cancer are controversial. In 2015, an overall prevalence between 0% and 86% was reported, with an average of 30.30%, with the highest frequency reported in Oceania at 44.30%. South America presented 14.60% of HPV infection, exceeding 10.70% of North America^[9].

HPV types are classified as low-risk and high-risk types based on the ability to induce carcinogenesis. HPV 6 and HPV 11 are low-risk subtypes and cause more than 90% of genital warts. High-risk HPV subtypes such as HPV 16, 18, 31, 33, 45 and 52, cause squamous intraepithelial lesions that can progress to invasive squamous cell carcinomas^[10]. This double strand DNA virus expresses *E6* and *E7* oncogenes, which interact with p53 and pRB proteins, respectively, promoting the development of neoplasias due to uncon-

trolled cell cycle activation and inhibition of apoptosis^[11]. The high-risk HPV genome could be integrated into the host genome during the carcinogenesis process, causing the loss of *E6* and *E7* transcription regulation by the interruption of *E2* gene open reading frame^[12].

Besides clinical and treatment parameters, the host immune response might influence the prognosis of cancer patients after standard treatment^[13]. Breast tumor progression is due to a systematic action, affecting the host's physiological processes and triggering responses in peripheral blood cells^[14]. It is known that CD4+ and CD8+ T cells are required for an effective anti-tumor immune response. CD4+ T cells are critical for priming tumor-specific CD8+ T cells and for the secondary expansion and memory of CD8+ T cells as well^[15]. CD8+ T cells have been shown to be mediators of antitumor immunity and act directly over tumor cells. Recent studies have suggested its clinical importance, reporting that an increase of CD8+ T cells correlates with increased survival in large cohorts of various human cancer patients^[13].

The natural killer (NK)+ cells have the ability to produce lysis in tumor cells and cells infected with intracellular viruses or parasites, through cytotoxic mechanisms mediated by preformed molecules, such as perforins and granzymes. They also have the ability to secrete cytokines such as interferon types I and II^[16]. NK cells appears to protect against tumor development and progression^[17].

Evaluation of circulating T lymphocytes, B lymphocytes and NK+ cells may be one of the beneficial ways to understand immune response, assist in clinical diagnosis, provide evidence of pathogenesis, course, and prognosis of disease and determine clinical treatment^[18]. This work aimed to evaluate the possible role of cellular subsets as predictive factors and the association of HPV in patients with breast cancer, according to the molecular subtypes, being the first study of its kind reported in Venezuela.

MATERIALS AND METHODS

Study population

We evaluated prospectively from February 2011 to October 2013 patients attending at the Breast Pathology Unit, in the Gynecology Department, from the University Hospital of Caracas. A total of 74 women were included, of which 48 samples were from patients diagnosed with breast cancer and 26 patients diagnosed with benign pathology of the breast. Patients were invited to participate in the study, with prior information on the design and protocol. Each one signed an informed consent, approved by the hospital Bioethics Committee.

Patients with other tumors or immune system-related disease were excluded. None of the patients had received any form of medical or surgical therapy such as radiotherapy, chemotherapy or treatment with steroids or immunosuppressants prior to the investigation.

Breast cancer molecular subtypes classification

Determination of breast cancer molecular subtype was

performed based on the immunohistochemical reports of the tumor piece, which were obtained from the clinical histories of each patient. According to markers expression, tumors were classified as: (1) Luminal A: RE+; RP+ ($\geq 20\%$); Ki67 ($< 14\%$), HER2-; (2) Luminal B HER2-: RE+; RP ($< 20\%$); Ki67 ($\geq 14\%$), HER2-; (3) Luminal B HER2+: RE+; RP indifferent; Ki67 ($\geq 14\%$); HER2+; (4) HER2+: RE-, RP-, Ki67 ($\geq 14\%$); HER2+; and (5) TN: RE-, RP-, Ki67 ($\geq 14\%$); HER2-^[5].

Tissue samples

Fresh biopsies were obtained from tumors of patients who underwent surgery. Biopsies were frozen at -70°C for molecular analysis.

DNA extraction

To perform DNA extraction from fresh breast biopsies, QIAmp DNA mini kit (250) was used (Qiagen, Hilden, Germany), following the manufacturer's instructions.

HPV detection and genotyping

HPV genome detection and genotyping from fresh breast biopsies was performed using the INNO-LIPA HPV Genotyping Extra test (Innogenetics, Ghent, Belgium), following the manufacturer's instructions. Based on the reverse hybridization principle, 28 different HPV genotypes were identified by detecting specific sequences in the L1 region. The assay uses the proven SPF10 primer set for highly sensitive amplification of the most clinically relevant HPV genotypes.

Blood samples

Before surgery, 5 mL of venous blood was obtained from each patient. The samples were drawn into heparinized tubes and transported to Oncology and Hematology Institute for processing.

Cellular subsets

Cellular subsets quantification was performed by flow cytometry. Fifty microliters of whole blood was taken, and 5 μL of monoclonal antibody was added. The sample was briefly vortexed and incubated for 15 min at room temperature in the dark. Eight hundred microliters of BD 1X lysis solution was added and incubated for 10 min at room temperature in the dark. The sample was centrifuged for 5 min at high speed, and the supernatant was discarded. The pellet was resuspended with 300 μL of FACS Flow and vortexed. Finally, the tube was acquired in the BD FACS Canto II cytometer, of six colors (configuration 4-2), with the BD FACS Diva 6.2.2 application.

Cell surface marker analysis was performed using CD4-FITC/CD8-PE/CD3-PC5 (Beckman Coulter) for CD3+, CD4+ and CD8+ T cells and CD3-APC, CD16-FITC, CD56-PE for NK+ cells (Beckman Coulter, Pasadena, CA, United States). Absolute cell counts were calculated by multiplying the cell subset percentage by the total lymphocyte concentration present in peripheral blood.

Table 1 Demographic characteristics for the breast cancer and benign pathology groups

	Breast cancer (<i>n</i> = 48)	Benign pathology (<i>n</i> = 26)	<i>P</i> value ¹
Mean age (yr)	57.79 ± 14.13 (range: 31-85)	32.54 ± 11.63 (range: 14-60)	0.000
Menarche (yr)	12.42 ± 1.67 (range: 9-16)	12.62 ± 1.69 (range: 9-17)	0.763
Term pregnancy	2.63 ± 2.27 (range: 0-11)	1.50 ± 1.55 (range: 0-5)	0.066
Sexual partners	1.98 ± 1.15 (range: 0-5)	1.76 ± 1.09 (range: 1-3)	0.750
Oral contraceptives	47.92%	65.38%	0.352
Tobacco	50.00%	11.53%	0.002
Alcohol	47.92%	46.15%	0.678
Breast cancer family history	37.50%	19.23%	0.199

¹Breast cancer group *vs* benign pathology group.**Table 2** Mean absolute values for the cellular subsets in the breast cancer and benign pathology groups

	Breast cancer (cel/mm ³)	Benign pathology (cel/mm ³)	<i>P</i> value ¹
TCD3+	1517.95 ± 666.23	1861.68 ± 760.52	0.102
TCD4+	888.73 ± 445.18	974.01 ± 390.17	0.504
TCD8+	551.34 ± 284.35	764.54 ± 431.10	0.048
CD4+/CD8+	1.94 (range: 0.59-6.33)	1.57 (range: 0.64-4.36)	0.271

¹Breast cancer group *vs* benign pathology group.

Statistical analysis

Measures of central tendency and dispersion were used for continuous variables; frequency analysis and contingency tables were used for discrete variables. Analysis of variance between groups, *t*-Student Parametric Test for two independent samples and non-parametric Mann-Whitney *U* test were used to perform hypothesis contrast. Significance level was fixed at *P* < 0.05 (Statistical Software: SPSS in its Version 20 in Spanish; IBM Corp., Armonk, NY, United States).

RESULTS

The study included 74 patients, with menarche age between 9 and 17 years, with 60.81% of menarche age between 12 and 14 years. The average sexual partners number was between 0 and 5, and 74.32% of the patients had 0 to 2 sexual partners. Regarding pregnancy at term, the cases registered were between 0 to 11 per patient, with the highest proportion between 0 and 2 deliveries (67.57%). Overall, 47.29% had an alcoholic habit, 36.49% had a tobacco habit, 54.05% of the patients used oral contraceptives, and 59.46% had a family history of cancer.

Table 1 shows demographic characteristics for the study groups, where 50% of breast cancer patients reported smoking and alcohol habit, and about 40% reported a family history of breast cancer. Mean age and tobacco habit showed a statistically significant difference between breast cancer patients and benign pathology patients.

Of the breast cancer samples, 50.00% corresponded to stage II, followed by stage I (20.83%), stage III (14.58%), stage 0 (8.33%) and stage IV (6.25%). Regarding the histopathological diagnosis, the breast

cancer tumors were infiltrating ductal carcinoma (79.16%), ductal carcinoma *in situ* (8.33%), lobular carcinoma (8.33%) and mucinous carcinoma (4.17%). Luminal A was the most frequent breast cancer molecular subtype (33.33%), followed by Luminal B HER2- (29.17%), Luminal B HER2+ (14.58%), TN (12.50%) and HER2+ (10.42%).

HPV was detected in 25.00% of the breast cancer patients, and genotype 18 was the most frequent in the studied group, followed by types 16, 6 and 31. 41.67% of the patients presented mixed infections, and 75.00% showed infection with high oncogenic risk genotypes in breast fresh tissue biopsies. In the benign pathology group, HPV genome was detected in 7.69%, finding genotypes 18 and 33 of high oncogenic risk, in single infections. Fifty percent of the HPV positive breast tumors were Luminal A, followed by the HER2+ type, with 25%. Luminal B HER2- and TN types represent 16.67% and 8.33%, respectively, among HPV positive breast tumors (*P* = 0.027).

Table 2 shows the mean absolute values for each cellular subset of the breast cancer patients and benign pathology patients. Breast cancer patients showed a decrease in mean values compared to those of benign pathology, with statistically significant difference in CD8+ count between both study groups. The mean of each cellular subset decreased as the stage of the disease increased (*P* > 0.005). Regarding breast cancer molecular subtypes, the HER2+ tumors had the lowest CD4+ and CD8+ values (Table 3).

The NK+ cells counts were elevated in the benign pathology group, with 1217.04 ± 778.69 cel/mm³ (range: 48.30-3193.18), in comparison with breast cancer patients (1053.79 ± 690.56 cel/mm³ (range: 187.55-3675.00)) (*P* = 0.651). A decrease in the mean

Table 3 Correlations between the breast cancer molecular subtypes and the cellular subtypes

	CD4+ (cel/mm ³)	CD8+ (cel/mm ³)	NK+ (cel/mm ³)
Luminal A	916 ± 358.81	645 ± 299.91	1064 ± 455.61
Luminal B HER2-	877 ± 494.16	544 ± 273.04	1120 ± 818.37
Luminal B HER2+ HER2+	914 ± 318.73	491 ± 208.07	926 ± 500.35
HER2+	680 ± 444.86	426 ± 319.13	834 ± 836.75
TN	992 ± 568.02	483 ± 214.90	1183 ± 837.52
P value ¹	0.842	0.527	0.912

¹Between breast cancer molecular subtypes. TN: Triple negative.

Table 4 Correlations between the breast cancer HPV status and the cellular subtypes

	HPV+ (n = 14)	HPV- (n = 34)	P value ¹
CD3+ (cel/mm ³)	1832.12 ± 537.08	1399.56 ± 665.69	0.563
CD4+ (cel/mm ³)	1139.92 ± 416.84	796.56 ± 416.84	0.091
CD8+ (cel/mm ³)	630.66 ± 271.00	518.58 ± 281.53	0.748
CD4+/CD8+	2.11 (range: 0.71-4.17)	1.88 (range: 0.59-6.33)	0.029
NK+ (cel/mm ³)	1350.21 ± 736.79	950.47 ± 634.35	0.082

¹HPV+ vs HPV-. HPV: Human papilloma virus; NK: Natural killer.

of NK+ cells was observed as the stage of the disease increased ($P = 0.0827$). Regarding breast cancer molecular subtypes, HER2+ tumors and Luminal B HER2+ had the lowest values of NK+, while TN tumors had the highest values (Table 3).

As shown in Table 4, HPV+ breast cancer tumors had elevated cellular subtype counts compared to HPV- tumors. A decrease in the mean of NK+ cells was observed as the disease stage increased in the HPV+ tumors.

DISCUSSION

Breast cancer is the most frequently diagnosed neoplastic disease during menopause, leading to a significant reduction of women's quality of life^[19]. In developing countries, the disease emerges as a serious public health problem due to the high economic and social costs associated with its care^[20].

Breast tumors have a very wide phenotypic diversity, which is accompanied by a large variability in gene expression patterns^[3]. Gene signatures are used as predictors of therapy response, and protein gene products have direct roles in the biology and clinical behavior of cancer cells and are potential targets to develop novel therapies^[5].

According to different reports, Luminal A represents 50% to 60% of breast cancer cases, while Luminal B, both HER2- and HER2+, represent 15% to 20% of cases. HER2+ tumors are detected between 15% and 20%, and TN tumors are found between 8% and 37% of breast tumors^[4,5]. In this study, Luminal A was the most frequent group, followed by Luminal B HER2-, Luminal B HER2+, TN and HER2+.

In Venezuela, few studies have characterized breast cancer molecular subtypes. Uribe *et al.*^[21] reported

Luminal A as the more frequent subtype (60.94%), followed by TN (28.75%). López *et al.*^[22] found that of 110 patients evaluated, 40.0% presented with Luminal B HER2- tumors, followed by Luminal A (20.91%).

Identification of genetic and risk factors, such as environmental and hormonal factors, have increasing value and play an important role in breast cancer prevention^[23]. These risk factors increase neoplastic process development probability and depend on the exposure time or individual genetic predisposition. Therefore, they can influence cancer development, even if they do not directly cause the disease^[24].

Regarding non-genetic factors, in our study, 35.42% of patients had a diagnosis age greater than 65 years, 58.33% had menarche before 12 years, 14.58% were nulliparous at the time of the study, and 47.92% indicated alcohol consumption. However, many of the patients who developed breast cancer did not have any identifiable risk factor.

Epidemiological studies gave a first indication of an association between viral agents and specific human cancers. Infectious factors are responsible for approximately 18% of human cancers, and it is well accepted that human breast cancer is highly associated with environmental factors. Among many microorganisms studied, viral infections have been suggested to play a role in cancer development, especially those cancers caused by HPV^[8].

In this study, HPV was detected in 25.00% of the samples. High oncogenic risk genotypes were the most common, with a higher prevalence of type 18 over 16, unlike cases of benign pathology, where the virus detection reached 4.76%. Similar reports indicate frequencies between 15.0% and 29.4% for HPV positivity^[25-29], and Aguayo *et al.*^[30] reported the lowest of this group of studies, with 8.7% in South America. As

for genotypes found, all studies found HPV type 16 was the most frequent in single infections or mixed infections with genotype 18.

However, other studies performed viral detection by polymerase chain reaction and genotyping by sequencing and reported a greater presence of type 18. In 2005, Kan *et al.*^[31] showed 48% positivity for HPV in breast cancer biopsies, with 100% for genotype 18. In addition, Heng *et al.*^[32], Antonsson *et al.*^[33], Glenn *et al.*^[34] and Lawson *et al.*^[35] found genotype 18 in greater proportion in breast cancer tissue. Since adenocarcinoma constitutes the majority of histological breast carcinoma types, it is understandable that HPV 18 was similar or even a little higher than HPV 16 in these studies. Hence, the distribution of high-risk HPV type for breast carcinoma is probably different than that for cervical cancer^[36].

To our knowledge, there are no reports about the correlation between HPV presence and breast cancer molecular subtypes, so our research is the first study to demonstrate this association. Piana *et al.*^[29] correlated HPV presence with TN tumors and showed a viral detection of 15%. However, the authors used paraffin embedded biopsies and did not discriminate between all the molecular subtypes.

More studies are necessary to evaluate the correlations between HPV presence and breast cancer molecular subtypes. With this knowledge, we could determine if the presence of HPV is associated with better prognosis tumors that are more responsive to chemotherapy treatments, as occurs in HPV positive oropharyngeal squamous cell carcinoma (OPSCC). Multiple studies have confirmed that HPV positive OPSCC shows a better response to chemotherapy and radiation, independently of treatment scheme^[37].

Recently, several studies focused on DNA deaminase APOBEC3B (A3B), a source of uracil dependent genomic mutations that is associated with mutagenesis in multiple human cancers, including cancers in breast, head and neck, cervix, bladder, lung, ovary and other tissues. This enzyme belongs to a protein family that has broad and overlapping functions in innate immunity *via* the restriction of viruses, transposons and other foreign DNA elements^[38]. Therefore, some authors have suggested a possible role for viral infections, such as HPV and EBV, in the regulation A3B gene expression in some cases of breast cancer^[38,39].

A3B expression levels are low in most of normal tissues. The E6 HPV oncoprotein offers the first contact in viral infection and A3B-mediated mutagenesis. In one model, high-risk HPV E6 protein inactivates p53, causing the elimination of A3B gene transcription^[38] for cervix cancer, head and neck cancer^[40], and HPV positive breast cancer^[39]. Given the roles of the proteins p53, A3B and E6, when levels of DNA damage and mutations are raised, answers to these damages and apoptosis are prevented.

Regarding the immune response, previous studies have shown a decrease in T lymphocytes proliferation,

low CD3+ and CD4+ count, an increase in CD8+ count and a decrease in CD4+/CD8+ ratio. Other studies have reported a gradual decrease in the CD4+/CD8+ ratio, proportional to the progression of breast cancer^[18]. Currently, data about the cytotoxicity of NK cells and blood levels are contradictory, and there is a lack of information regarding the tumor microenvironment in patients with breast cancer^[41].

An important strength of this work was the inclusion of breast cancer patients samples, which were taken prior to surgical, systemic and radiant treatment. This allowed us to evaluate the differences in the immunological status of the patients without the influence of treatment.

In the breast cancer patients group, there was a decrease in the concentration of TCD4+ and TCD8+ lymphocytes compared to the benign pathology group, while the CD4+/CD8+ ratio was higher in the breast cancer. TCD8+ concentration variation showed a statistically significant difference ($P = 0.048$) in respect to the benign pathology group, representing a possible predictive marker. Regarding the behavior of lymphocytes in relation to breast cancer staging, the CD4+/CD8+ ratio decreased as the stage of the disease increased, which is consistent with Jia *et al.*^[18], who reported a rate decrease in advanced breast cancer.

The average absolute values for NK+ cells in the breast cancer patients group was less than those of the benign pathology group. Verma *et al.*^[42] also reported a lower NK+ cell count in breast cancer patients. They evaluated the variation of these values during and after treatment and found that neoadjuvant therapy increased NK+ cell values above that reported in the healthy group. It has been reported that chemotherapy, by damaging or stressing cells, promotes the release of various signals that activate dendritic and NK+ cells and induces the release of pro-inflammatory cytokines^[43].

It is known that functional capacity of immune cells decline with aging. Diminished phagocytic capacity of dendritic cells leads to impaired antigen presentation and activation of the adaptive immune system. In addition, thymus involution decreases the production of naïve T cells, and memory T cells accumulate, diminishing the T-cell repertoire^[44,45]. Postmenopausal women exhibit a reduced number of total lymphocytes, mainly B and CD4+ cells. Similarly, after surgical menopause, the CD4+/CD8+ ratio and the circulating B cells are decreased, while NK cells are increased^[45]. The breast cancer patients evaluated did not show statistically significant differences with respect to cell subsets and age groups (data not shown).

Regarding breast cancer molecular subtypes, we found that the CD4+ count and the CD4+/CD8+ ratio were decreased in the HER2+ tumor group, and that TN tumors showed an increase in the NK+ cell count. Particularly, the HER2+ group and the Luminal B HER2+ group showed a considerable decrease of NK+ with respect to the rest of the molecular subtypes.

Jia *et al.*^[18] reported a decrease in the CD4+ count

and the CD4+/CD8+ ratio and an increase in CD8+ and NK+ cells in ER-, HER2+, TN tumors, with Ki67 \geq 14%, indicating a greater failure of the immunological response in those tumors with aggressive phenotype. Previous studies have shown that estrogen plays an important role in regulating the activation of T lymphocytes, particularly CD4+ and CD8+; suggesting that the absence of estrogen receptors in HER2+ tumors is related to what was observed in this study population and was affecting the concentration of both CD4+ and NK+ cells.

ER- and TN breast tumors have a worse prognosis than ER positive tumors. These findings indicate a greater degree of immune function suppression and anti-tumor activation, which is reflected in the aggressiveness of HER2+ and TN tumors^[18]. Kim *et al.*^[13] revealed that the decreased number of CD8+ T cells was significantly associated with aggressiveness and malignant features of tumors, including lymph node metastasis, higher stage and higher Ki67. Therefore, immunotherapy is one of the most promising approaches for breast cancer therapy. If new research establishes a role for lymphocyte subsets in the etiology of aggressive phenotypes, such as those characterized by being ER-, HER2+, presenting a Ki67 \geq 14% and TN subtype, new treatment strategies for these breast cancer types should include immunotherapy.

The cellular subset values were increased in the HPV+ tumors compared to those of HPV- patients, and there was a statistically significant difference with respect to the CD4+/CD8+ ratio ($P = 0.029$), due to a considerable increase in the CD4+ count of HPV+ patients. Despite the HPV evasion mechanisms of the immune response, about 90% of genital and skin lesions resolve on average in 2 years. Immunohistochemical studies show that regression of cutaneous, oral and genital warts in animals and humans is characterized by a massive local infiltration with CD4+, CD8+ and macrophages into the lesion, and the expression of Th1 cytokines profile. Despite the intense local response, the systemic antigen-specific T cell responses are weak, transient and difficult to measure^[46].

Compromised adaptive immunity is the basis for high-risk HPV infection to cervical cancer progression. Different immune cell profiles characterize the different stages of the disease progression in cervical intraepithelial neoplasm and invasive cancer. The high-risk HPV infection changes and modifications induced include the adaptation of the immune system to create a suitable microenvironment for persistent infection and lesion progression^[47].

During HPV persistent infection, pro-inflammatory cytokines are not released, and the Langerhans cell and dendritic cell activation and recruitment signals are absent. In fact, cells with viral late gene expression, which may contain high levels of viral proteins, are shed from the surface of the epithelium away from immune surveillance. In general, a failure in developing an effective host immune response correlates with persistent infection and an increased probability of

progression toward invasive cancer^[11].

Although no previous studies have evaluated the profile of cellular subsets in patients with HPV positive breast tumors, we observed that the virus infected group showed higher values of CD4+, CD8+ and NK+ cells in comparison with negative patients. It is possible that these increases are due to an activation of the immune response to viral infection in the mammary tissue.

It has been described in HPV positive cases of OPSCC that patients show a greater proportion of circulating TCD8+ lymphocytes compared with HPV negative cases and that these lymphocyte levels are better predictors of treatment response than HPV status. Other prognostic markers could include CD4+/CD8+ ratio, the circulating levels of the T cell group, the presence of infiltrating lymphocytes in the tumor microenvironment, the expression of MHC class I and the characterization of the immune response by microarray^[48].

It is known that CD8+ T cells play a major role in elimination of viral infection, secreting interferon and displaying cytolytic effects mediated by granzyme and perforin. CD4+ T cells also secrete interferon but instead mediate killing primarily by engagement with ligands for death receptors, such as Fas or TNF-related apoptosis-inducing ligand, resulting in caspase-mediated apoptosis^[49]. The presence of CD8 T cells in cervical lesions is associated with a favorable prognosis, with their numbers inversely correlating with tumor progression.

On the other hand, infection with high-risk HPV genotypes compromises the activation of NK+ cells. In the case of the cervix, NK+ cells predominate in early stages of infection and in low-grade squamous intraepithelial lesions, whereas in cases of cervical cancer, NK+ activation receptors are considerably diminished. These findings imply a low cytotoxic activity by the NK+, thereby facilitating the progression of the lesion and carcinogenesis^[47].

In patients with breast cancer and HPV positive breast tissue, a decrease in the NK+ cell count was observed as the severity of the disease increased, implying a failure in the cytotoxic activity performed by the cells studied. It has been observed that the presence of HPV in mammary tissue modifies the activity of the evaluated cells, at the level of the cellular immune response.

It is well known that the incidence of cervical cancer due to HPV infection is much higher compared to non-genital cancers or oropharyngeal cancers associated with the presence of the virus, so the distribution and application of HPV vaccines for preventing cervical cancer remains as a public health priority. However, by 2008, non-genital and oropharyngeal cancers represented about 80000 new cases of cancer associated with HPV infection worldwide, implying a major health problem^[50].

Evidence obtained from clinical trials indicates that

current HPV vaccines can prevent HPV infections in vulva, vagina, anus and mouth as well as pre-cancerous anogenital lesions in women and oral and anogenital infections, and pre-cancerous lesions in men. However, data comparing the efficacy and effectiveness data of cervical infections and high-grade lesions with those types of injuries are limited^[50,51].

As yet, the implications of HPV vaccination for prevention of non-cervical cancers have not been fully explored. Some countries have recommended HPV vaccination for young males, based on the hypothesis that vaccination will prevent HPV-associated cancers in men as well as theoretical benefits in preventing HPV transmission to women^[50].

Therefore, if HPV vaccines contribute to decrease the rate of non-genital cancers, we could have a group of patients that develop breast cancer due to HPV infection, which could benefit from the use of vaccines available in the international market, which include the bivalent that protects against types 16 and 18, the tetravalent, for types 6, 11, 16 and 18, and the nonavalent, approved in 2014 for types 6, 11, 16, 18, 31, 33, 45, 52 and 58^[52].

ARTICLE HIGHLIGHTS

Research background

Breast cancer is the leading cause of death among women, classified in molecular subtypes according to a genetic profile. Approximately 18% of human cancers are caused by infectious factors, and it is recognized that breast cancer is strongly related to environmental factors, such as viruses, diet, radiation, among others. Human papilloma virus (HPV) has been detected in 0% to 86% of breast cancer tumors, and it represents a possible risk factor. The host immune response might influence the prognosis of cancer patients after standard treatment.

Research motivation

It was recently suggested that HPV presence may act as a risk factor in breast cancer development, but it has not been correlated with molecular subtypes. In addition, it is important to evaluate the immune response of breast cancer patients and to be able to suggest some prognostic values that make it possible to offer better patients treatment.

Research objectives

The main objective is to detect HPV presence and to characterize the cellular immune response in breast cancer patients based on the molecular subtypes.

Research methods

The patients inclusion was done prospectively, and the breast cancer molecular classification was made according to the St. Gallen International Breast Cancer Conference. HPV detection and genotyping were performed using HPV INNO-LIPA Genotyping Extra test, and lymphocyte subsets were measured by flow cytometry.

Research results

Luminal A was the most frequent breast cancer molecular subtype (33.33%), and HPV was detected in 25% of the breast cancer patients, with genotype 18 as the most frequent in the studied population. The means of CD3+, CD4+ and CD8+ subsets were decreased in patients with breast cancer, respective to benign pathology, with a statistically significant difference between CD8+ values ($P = 0.048$). The mean of NK+ cells was increased in the benign pathology group. HER2+ and Luminal B HER2+ tumors had the lowest counts of cell subsets. The HPV breast cancer patients had elevated counts of cellular

subsets.

Research conclusions

It can be observed that HPV positive breast cancer tumors have a better prognosis, correlated with Luminal A subtypes, and show a better cellular immune response, specifically in relation with TCD8+ cells counts, suggesting a better response to chemotherapy and radiant treatment, as in the case of HPV positive oropharynx tumors.

Research perspectives

Future efforts should focus on evaluating patients disease-free survival, based on HPV positivity in the tumor tissue. In addition, viral load and HPV genome integration, the identification of HPV variants by sequencing, and the infiltrating lymphocytes in the tumor bed should also be studied. As a final consideration, the experience of working with fresh samples is complex and involves a process of sample collection in the operating room, in addition to the management of bioethical parameters, and our experience will allow for the possibility of obtaining more accurate results.

REFERENCES

- 1 **World Health Organization.** Available from: URL: <http://www.who.int/es>
- 2 **de Mortalidad A.** Ministerio del Poder Popular para la Salud (MPPS). 2012. Available from: URL: <http://www.mpps.gob.ve>
- 3 **Perou CM, Sørli T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, Zhu SX, Lønning PE, Børresen-Dale AL, Brown PO, Botstein D.** Molecular portraits of human breast tumours. *Nature* 2000; **406**: 747-752 [PMID: 10963602 DOI: 10.1038/35021093]
- 4 **Untch M, Gerber B, Harbeck N, Jackisch C, Marschner N, Möbus V, von Minckwitz G, Loibl S, Beckmann MW, Blohmer JU, Costa SD, Decker T, Diel I, Dimpfl T, Eiermann W, Fehm T, Friese K, Jänicke F, Janni W, Jonat W, Kiechle M, Köhler U, Lück HJ, Maass N, Possinger K, Rody A, Scharl A, Schneeweiss A, Thomssen C, Wallwiener D, Welt A.** 13th st. Gallen international breast cancer conference 2013: primary therapy of early breast cancer evidence, controversies, consensus - opinion of a german team of experts (zurich 2013). *Breast Care (Basel)* 2013; **8**: 221-229 [PMID: 24415975 DOI: 10.1159/000351692]
- 5 **Yersal O, Barutca S.** Biological subtypes of breast cancer: Prognostic and therapeutic implications. *World J Clin Oncol* 2014; **5**: 412-424 [PMID: 25114856 DOI: 10.5306/wjco.v5.i3.412]
- 6 **Cintra JR, Teixeira MT, Diniz RW, Gonçalves Junior H, Florentino TM, Freitas GF, Oliveira LR, Neves MT, Pereira T, Guerra MR.** Immunohistochemical profile and clinical-pathological variables in breast cancer. *Rev Assoc Med Bras (1992)* 2012; **58**: 178-187 [PMID: 22569612 DOI: 10.1590/S0104-42302012000200013]
- 7 **Fritz M, Speroff L.** Endocrinología ginecológica clínica y esterilidad. 8th ed. Wolters Kluwer Health España SA, editor. España: Lippincott Williams & Wilkins; 2012
- 8 **Zhang N, Ma ZP, Wang J, Bai HL, Li YX, Sun Q, Yang L, Tao L, Zhao J, Cao YW, Li F, Zhang WJ.** Human papillomavirus infection correlates with inflammatory Stat3 signaling activity and IL-17 expression in patients with breast cancer. *Am J Transl Res* 2016; **8**: 3214-3226 [PMID: 27508043]
- 9 **Zhou Y, Li J, Ji Y, Ren M, Pang B, Chu M, Wei L.** Inconclusive role of human papillomavirus infection in breast cancer. *Infect Agent Cancer* 2015; **10**: 36 [PMID: 26504492 DOI: 10.1186/s13027-015-0029-6]
- 10 **Wang YW, Zhang K, Zhao S, Lv Y, Zhu J, Liu H, Feng J, Liang W, Ma R, Wang J.** HPV Status and Its Correlation with BCL2, p21, p53, Rb, and Survivin Expression in Breast Cancer in a Chinese Population. *Biomed Res Int* 2017; **2017**: 6315392 [PMID: 29423411 DOI: 10.1155/2017/6315392]
- 11 **Egawa N, Egawa K, Griffin H, Doorbar J.** Human Papillomaviruses; Epithelial Tropisms, and the Development of Neoplasia. *Viruses* 2015; **7**: 3863-3890 [PMID: 26193301 DOI: 10.3390/v7072802]

- 12 **Hooley E**, Fairweather V, Clarke AR, Gaston K, Brady RL. The recognition of local DNA conformation by the human papillomavirus type 6 E2 protein. *Nucleic Acids Res* 2006; **34**: 3897-3908 [PMID: 16914454 DOI: 10.1093/nar/gkl466]
- 13 **Kim ST**, Jeong H, Woo OH, Seo JH, Kim A, Lee ES, Shin SW, Kim YH, Kim JS, Park KH. Tumor-infiltrating lymphocytes, tumor characteristics, and recurrence in patients with early breast cancer. *Am J Clin Oncol* 2013; **36**: 224-231 [PMID: 22495453 DOI: 10.1097/COC.0b013e3182467d90]
- 14 **Nieto-Velázquez NG**, Torres-Ramos YD, Muñoz-Sánchez JL, Espinosa-Godoy L, Gómez-Cortés S, Moreno J, Moreno-Eutimio MA. Altered Expression of Natural Cytotoxicity Receptors and NKG2D on Peripheral Blood NK Cell Subsets in Breast Cancer Patients. *Transl Oncol* 2016; **9**: 384-391 [PMID: 27641642 DOI: 10.1016/j.tranon.2016.07.003]
- 15 **Huang Y**, Ma C, Zhang Q, Ye J, Wang F, Zhang Y, Hunborg P, Varvares MA, Hofst DF, Hsueh EC, Peng G. CD4+ and CD8+ T cells have opposing roles in breast cancer progression and outcome. *Oncotarget* 2015; **6**: 17462-17478 [PMID: 25968569 DOI: 10.18632/oncotarget.3958]
- 16 **Rojas-Pandales F**, Bolaños N, Mercado M, González JM, Cuéllar A, Cifuentes-Rojas C. Valores de referencia de células asesinas naturales (NK y NKT) en donantes de sangre de Bogotá. *Acta Med Col* 2007; **32**: 124-128
- 17 **Carvalho MI**, Pires I, Prada J, Queiroga FL. A role for T-lymphocytes in human breast cancer and in canine mammary tumors. *Biomed Res Int* 2014; **2014**: 130894 [PMID: 24672781 DOI: 10.1155/2014/130894]
- 18 **Jia Y**, Xu L, Lin Q, Zhu M, Ding L, Wu K, Lu Y. Levels of lymphocyte subsets in peripheral blood prior treatment are associated with aggressive breast cancer phenotypes or subtypes. *Med Oncol* 2014; **31**: 981 [PMID: 24798876 DOI: 10.1007/s12032-014-0981-9]
- 19 **Kamińska M**, Ciszewski T, Łopacka-Szatan K, Miotła P, Starosławska E. Breast cancer risk factors. *Prz Menopauzalny* 2015; **14**: 196-202 [PMID: 26528110 DOI: 10.5114/pm.2015.54346]
- 20 **Anderson BO**, Yip CH, Smith R, Shyyan R, Sener SF, Eniu A, Carlson RW, Azavedo E, Harford J. Guideline implementation for breast healthcare in low-income and middle-income countries: overview of the Breast Health Global Initiative Global Summit 2007. *Cancer* 2008; **113**: 2221-2243 [PMID: 18816619 DOI: 10.1002/cncr.23844]
- 21 **Uribe JR**, Hernández CA, Melonascino F, Rodríguez JE, Istúriz L, Márquez M, Rodríguez R, Uribe JL. Clasificación molecular del cáncer de mama y su correlación clínica. *Rev Venez Oncol* 2010; **22**: 109-116
- 22 **López M**, Pesci-Feltri A, García I, Guida V, Fernandes A, Blanch R. Factores de riesgo y protectores asociados al cáncer de mama. *Rev Venez Oncol* 2017; **29**: 102-111
- 23 **Martin AM**, Weber BL. Genetic and hormonal risk factors in breast cancer. *J Natl Cancer Inst* 2000; **92**: 1126-1135 [PMID: 10904085 DOI: 10.1093/jnci/92.14.1126]
- 24 **Vogel VG**. Management of the high-risk patient. *Surg Clin North Am* 2003; **83**: 733-751 [PMID: 12875593 DOI: 10.1016/S0039-6109(03)00030-6]
- 25 **Khan NA**, Castillo A, Koriyama C, Kijima Y, Umekita Y, Ohi Y, Higashi M, Sagara Y, Yoshinaka H, Tsuji T, Natsugoe S, Douchi T, Eizuru Y, Akiba S. Human papillomavirus detected in female breast carcinomas in Japan. *Br J Cancer* 2008; **99**: 408-414 [PMID: 18648364 DOI: 10.1038/sj.bjc.6604502]
- 26 **de León DC**, Montiel DP, Nemcova J, Mykyskova I, Turcios E, Villavicencio V, Cetina L, Coronel A, Hes O. Human papillomavirus (HPV) in breast tumors: prevalence in a group of Mexican patients. *BMC Cancer* 2009; **9**: 26 [PMID: 19161629 DOI: 10.1186/1471-2407-9-26]
- 27 **Herrera-Goepfert R**, Khan NA, Koriyama C, Akiba S, Pérez-Sánchez VM. High-risk human papillomavirus in mammary gland carcinomas and non-neoplastic tissues of Mexican women: no evidence supporting a cause and effect relationship. *Breast* 2011; **20**: 184-189 [PMID: 21146410 DOI: 10.1016/j.breast.2010.11.006]
- 28 **Manzouri L**, Salehi R, Shariatpanahi S, Rezaie P. Prevalence of human papilloma virus among women with breast cancer since 2005-2009 in Isfahan. *Adv Biomed Res* 2014; **3**: 75 [PMID: 24627883 DOI: 10.4103/2277-9175.125873]
- 29 **Piana AF**, Sotgiu G, Muroi MR, Cossu-Rocca P, Castiglia P, De Miglio MR. HPV infection and triple-negative breast cancers: an Italian case-control study. *Virol J* 2014; **11**: 190 [PMID: 25413873 DOI: 10.1186/s12985-014-0190-3]
- 30 **Aguayo F**, Khan N, Koriyama C, González C, Ampuero S, Padilla O, Solís L, Eizuru Y, Corvalán A, Akiba S. Human papillomavirus and Epstein-Barr virus infections in breast cancer from Chile. *Infect Agent Cancer* 2011; **6**: 7 [PMID: 21699721 DOI: 10.1186/1750-9378-6-7]
- 31 **Kan CY**, Iacopetta BJ, Lawson JS, Whitaker NJ. Identification of human papillomavirus DNA gene sequences in human breast cancer. *Br J Cancer* 2005; **93**: 946-948 [PMID: 16222323 DOI: 10.1038/sj.bjc.6602778]
- 32 **Heng B**, Glenn WK, Ye Y, Tran B, Delprado W, Lutze-Mann L, Whitaker NJ, Lawson JS. Human papilloma virus is associated with breast cancer. *Br J Cancer* 2009; **101**: 1345-1350 [PMID: 19724278 DOI: 10.1038/sj.bjc.6605282]
- 33 **Antonsson A**, Spurr TP, Chen AC, Francis GD, McMillan NA, Saunders NA, Law M, Bennett IC. High prevalence of human papillomaviruses in fresh frozen breast cancer samples. *J Med Virol* 2011; **83**: 2157-2163 [PMID: 22012724 DOI: 10.1002/jmv.22223]
- 34 **Glenn WK**, Heng B, Delprado W, Iacopetta B, Whitaker NJ, Lawson JS. Epstein-Barr virus, human papillomavirus and mouse mammary tumour virus as multiple viruses in breast cancer. *PLoS One* 2012; **7**: e48788 [PMID: 23183846 DOI: 10.1371/journal.pone.0048788]
- 35 **Lawson JS**, Glenn WK, Salyakina D, Delprado W, Clay R, Antonsson A, Heng B, Miyauchi S, Tran DD, Ngan CC, Lutze-Mann L, Whitaker NJ. Human Papilloma Viruses and Breast Cancer. *Front Oncol* 2015; **5**: 277 [PMID: 26734565 DOI: 10.3389/fonc.2015.00277]
- 36 **Li N**, Bi X, Zhang Y, Zhao P, Zheng T, Dai M. Human papillomavirus infection and sporadic breast carcinoma risk: a meta-analysis. *Breast Cancer Res Treat* 2011; **126**: 515-520 [PMID: 20740311 DOI: 10.1007/s10549-010-1128-0]
- 37 **Chera B**, Wang K, Monroe A, Galloway T, Amdur R, Hayes DN, Zevallos J, Mendenhall WM. Truth or myth: Definitive chemoradiotherapy doesn't work for HPV/p16 negative oropharyngeal squamous cell carcinoma? *Oral Oncol* 2017; **65**: 125-126 [PMID: 27993466 DOI: 10.1016/j.oraloncology.2016.12.001]
- 38 **Vieira VC**, Leonard B, White EA, Starrett GJ, Temiz NA, Lorenz LD, Lee D, Soares MA, Lambert PF, Howley PM, Harris RS. Human papillomavirus E6 triggers upregulation of the antiviral and cancer genomic DNA deaminase APOBEC3B. *MBio* 2014; **5**: [PMID: 25538195 DOI: 10.1128/mBio.02234-14]
- 39 **Ohba K**, Ichiyama K, Yajima M, Gemma N, Nikaido M, Wu Q, Chong P, Mori S, Yamamoto R, Wong JE, Yamamoto N. In vivo and in vitro studies suggest a possible involvement of HPV infection in the early stage of breast carcinogenesis via APOBEC3B induction. *PLoS One* 2014; **9**: e97787 [PMID: 24858917 DOI: 10.1371/journal.pone.0097787]
- 40 **Henderson S**, Chakravarthy A, Su X, Boshoff C, Fenton TR. APOBEC-mediated cytosine deamination links PIK3CA helical domain mutations to human papillomavirus-driven tumor development. *Cell Rep* 2014; **7**: 1833-1841 [PMID: 24910434 DOI: 10.1016/j.celrep.2014.05.012]
- 41 **Kajitani K**, Tanaka Y, Arihiro K, Kataoka T, Ohdan H. Mechanistic analysis of the antitumor efficacy of human natural killer cells against breast cancer cells. *Breast Cancer Res Treat* 2012; **134**: 139-155 [PMID: 22261932 DOI: 10.1007/s10549-011-1944-x]
- 42 **Verma C**, Kaewkangsadan V, Eremin JM, Cowley GP, Ilyas M, El-Sheemy MA, Eremin O. Natural killer (NK) cell profiles in blood and tumour in women with large and locally advanced breast cancer (LLABC) and their contribution to a pathological complete response (PCR) in the tumour following neoadjuvant

- chemotherapy (NAC): differential restoration of blood profiles by NAC and surgery. *J Transl Med* 2015; **13**: 180 [PMID: 26040463 DOI: 10.1186/s12967-015-0535-8]
- 43 **Tsavaris N**, Kosmas C, Vadiaka M, Kanelopoulos P, Boulamatsis D. Immune changes in patients with advanced breast cancer undergoing chemotherapy with taxanes. *Br J Cancer* 2002; **87**: 21-27 [PMID: 12085250 DOI: 10.1038/sj.bjc.6600347]
 - 44 **Vitlic A**, Lord JM, Phillips AC. Stress, ageing and their influence on functional, cellular and molecular aspects of the immune system. *Age (Dordr)* 2014; **36**: 9631 [PMID: 24562499 DOI: 10.1007/s11357-014-9631-6]
 - 45 **Giefing-Kröll C**, Berger P, Lepperdinger G, Grubeck-Loebenstien B. How sex and age affect immune responses, susceptibility to infections, and response to vaccination. *Aging Cell* 2015; **14**: 309-321 [PMID: 25720438 DOI: 10.1111/accel.12326]
 - 46 **Stanley MA**, Sterling JC. Host responses to infection with human papillomavirus. *Curr Probl Dermatol* 2014; **45**: 58-74 [PMID: 24643178 DOI: 10.1159/000355964]
 - 47 **Song D**, Li H, Li H, Dai J. Effect of human papillomavirus infection on the immune system and its role in the course of cervical cancer. *Oncol Lett* 2015; **10**: 600-606 [PMID: 26622540 DOI: 10.3892/ol.2015.3295]
 - 48 **Andersen AS**, Koldjaer Sølling AS, Ovesen T, Rusan M. The interplay between HPV and host immunity in head and neck squamous cell carcinoma. *Int J Cancer* 2014; **134**: 2755-2763 [PMID: 23913554 DOI: 10.1002/ijc.28411]
 - 49 **Hibma MH**. The immune response to papillomavirus during infection persistence and regression. *Open Virol J* 2012; **6**: 241-248 [PMID: 23341859 DOI: 10.2174/1874357901206010241]
 - 50 **Taylor S**, Bunge E, Bakker M, Castellsagué X. The incidence, clearance and persistence of non-cervical human papillomavirus infections: a systematic review of the literature. *BMC Infect Dis* 2016; **16**: 293 [PMID: 27301867 DOI: 10.1186/s12879-016-1633-9]
 - 51 **Guo T**, Eisele DW, Fakhry C. The potential impact of prophylactic human papillomavirus vaccination on oropharyngeal cancer. *Cancer* 2016; **122**: 2313-2323 [PMID: 27152637 DOI: 10.1002/cncr.29992]
 - 52 **Sabeena S**, Bhat PV, Kamath V, Arunkumar G. Global human papilloma virus vaccine implementation: An update. *J Obstet Gynaecol Res* 2018; **44**: 989-997 [PMID: 29517117 DOI: 10.1111/jog.13634]

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Case Control Study

Mismatch repair protein expression and intratumoral budding in rectal cancer are associated with an increased pathological complete response to preoperative chemoradiotherapy: A case-control study

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Abstract**AIM**

To determine whether the association of rectal adenocarcinoma with a defective-mismatch repair system (dMMR) was associated with a pathological complete response (pCR) to preoperative chemoradiotherapy.

METHODS

A case-control study was designed with the aim of determining if patients with rectal adenocarcinoma with dMMR had an associated high pCR rate in response to neoadjuvant chemoradiotherapy (nCRT).

RESULTS

Seventy-two cases with pCR were compared against 144 controls without pCR. Across 216 cases, the mean age was 56.8 years, 140 (64.8%) were men, and 63 (29.2%) demonstrated the dMMR system. The pCR was associated with G1 tumors, dMMR, the absence of vascular invasion, and low tumor budding in the pretreatment biopsy. In a multivariate analysis, the factors associated with pCR were dMMR (OR: 2.61; 95%CI: 1.355-5.040, $P = 0.004$) and a low degree of tumor budding (OR: 2.52; 95%CI: 1.366-4.894, $P = 0.025$).

CONCLUSION

We found an independent association between dMMR and a low rate of tumor budding, with a higher rate of pCR, in the basal biopsies of patients with rectal carcinoma subjected to nCRT.

Key words: Rectal cancer; Chemoradiotherapy; Tumor budding; Mismatch repair; Pathological complete response

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Core tip: Defective Mismatch repair (dMMR) and a low number of buds in the pretreatment biopsy were independently associated with a high rate of the pathological complete response. Tumor budding and MMR status should be considered as tools to be implemented in studies that predict the pathological response to preoperative chemo-radiotherapy in rectal cancer.

Lino-Silva LS, Gamboa-Domínguez A, Zúñiga-Tamayo D, Salcedo-Hernández RA, Cetina L, Cantú-de-León D. Mismatch repair protein expression and intratumoral budding in rectal cancer are associated with an increased pathological complete response to preoperative chemoradiotherapy: A case-control study. *World J Clin Oncol* 2018; 9(7): 133-139 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i7/133.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i7.133>

INTRODUCTION

Rectal carcinoma is the third most common cancer, only after lung, breast, and prostate cancers, but it is the second leading cause of cancer-related mortality^[1]. The treatment in locally advanced stages is neoadjuvant chemoradiotherapy (nCRT), with the aim of decreasing the tumor size, inducing favorable changes in staging

(downstaging), increasing the probability of anal sphincter preservation, and decreasing the local recurrence rate and the possibility of a pathological complete response (pCR)^[2], which is defined as the absence of neoplastic cells in both the rectal wall and lymph nodes. A pCR is reported in between 10% and 30% of these patients, and it is suggested that it is associated with a better oncologic outcome, a low local recurrence and an improved survival^[3-4].

Unfortunately, the pCR is only indicated by a histopathological study of the surgical specimen, and in theory, elements that anticipate a pCR in the pretreatment biopsy would be very useful because elements, such as preoperative biopsy findings (post-nCRT), endoscopic examination, or imaging studies after completion of nCRT, are not sensitive or specific enough to predict pCR.

Identifying characteristics of the tumors that will respond completely to nCRT could help avoid the radical resection of the rectum, a procedure that reduces the quality of life of the patient. Several histopathological, immunohistochemical, and molecular markers have been analyzed, but with contradictory results^[5].

The fundamental damage caused by nCRT is direct nuclear damage, and then, some studies have associated the mismatch repair system (MMR) status with pCR with inconclusive results^[6]. During DNA damage, the presence of proteins, such as MSH2, MSH6, MLH1, and PMS2 (associated with the MMR), increases^[7]. We hypothesized that deficient MMR (dMMR) neoplastic cells may be especially prone to death from agents that induce genetic damage, such as nCRT. Other factors that might predict this response are the immune response to the tumor and the tumor budding (individual neoplastic cells at the tumor stroma).

Our aim was to determine if rectal adenocarcinomas with a dMMR system were associated with a higher pCR rate to nCRT compared to cases that were MMR-efficient (eMMR). Second, we evaluated the association between the inflammatory tumoral response and intratumoral budding with the pCR rate.

MATERIALS AND METHODS

A case-control study was performed

Ethical statement: This work was authorized by the ethics and research committee of our institution, with a waiver of informed consent, because of its retrospective nature (Instituto Nacional de Cancerología, approval number: Rev/16/21).

Inclusion criteria: From a prospective maintained rectal cancer database from 2010 to 2016 at one large cancer center (INCan), we searched for patients with stage III adenocarcinoma of the lower two-thirds of the rectum that received and completed nCRT and were subjected to radical rectal surgery. For each case, the

initial biopsy (pre-nCRT) was retrieved, and the MMR protein status was determined by immunohistochemistry in this tissue. For all the cases, the pathological material from the surgical specimens were available, and all the resections were systematically evaluated according to the procedure recommended by Quirke and were standardized in our institution with the entire rectal wall examination in the case of a macroscopic absence of a tumor (in other words, the totality of the rectal wall was pathologically examined)^[8,9]. We excluded patients who had previously undergone chemotherapy or radiotherapy for another neoplasm.

Case definition: Individuals with rectal cancer after nCRT who do not have histological evidence of cancer in both the rectum and lymph nodes of the surgical specimen (pCR, ypT0N0M0).

Controls definition: Individuals with rectal cancer who had histological evidence of residual neoplasm after nCRT. We took two controls per case that were matched by sex and age (± 5 years).

The outcome of interest was the presence or absence of pCR. An odds ratio (OR) of 2.0, for the primary objective, was the base for the sample size calculation, considering an alpha-value of 95%, a beta-value of 80%, a known proportion of MMR of 14%^[10], and a 15% loss; the calculated number of cases per group was 72.

The clinical, demographic, and histopathological data were retrieved from the selected cases and controls. The cancer staging was determined preoperatively based on the Tumor Node Metastasis (TNM) classification of the American Joint Committee on Cancer (seventh edition)^[11]. The expression of the MMR proteins was evaluated separately and independently by three pathologists with experience in gastrointestinal pathology who were blinded to the clinical data. If discordance was found, a consensus was achieved in a multi-observer microscope. The expression was evaluated according to the following criteria^[12]: (1) The nuclear expression of all markers in each sample, allowing for the classification of adenocarcinoma, as with an efficient MMR system (eMMR); and (2) The absence of the nuclear expression of any of the markers in each sample, allowing for the classification of adenocarcinoma, as with a dMMR.

Additionally, for the MMR evaluation, each case the inflammatory infiltrate was categorized as present (either peritumoral in the form of lymphoid aggregates, or intratumoral) or absent. The evaluation of intratumoral budding was carried out by examining 0.785 mm² of the peritumoral stroma of the biopsy specimen, following the guidelines described by Lugli *et al.*^[13]. We defined low intratumoral budding as the presence of 0 to 4 buds/0.785 mm², and high intratumoral budding occurred when there were ≥ 5 buds/0.785 mm².

Statistical analysis

Data analysis was performed using the statistical package SPSS 22.0 (IBM, California, United States). The ORs were calculated and contrasted with the Mantel-Haenszen method, with a significance level of 0.05. For the comparison of the numerical variables, we used a *t* test to compare the means, the Mann-Whitney test for medians and the chi-square test or Fischer's exact test for categorical variables. A multivariate analysis was performed with a conditional logistic regression to determine the independent predictors of pCR, with the variables at a significance level of $P < 0.05$ in the univariate analysis as components of the regression model.

RESULTS

Population and clinical characteristics

Seventy-two cases with pCR were evaluated, and two controls were selected per case, totaling 144 controls, for a total sample of 216 cases. The mean age was 56.8 \pm 12.5 years, with 192 (88.9%) cases younger than 40 years. Of the 216 cases, 140 (64.8%) were men.

All the cases were in stage III and at the time of the presentation, 123 cases (56.9%) had a cT3 stage rectal tumor, whereas 44 (20.4%) had cT2, 31 (14.4%) had cT4a, and 18 (8.3%) were cT4b. Seventy-seven (35.6%) patients were classified as having nodal disease by radiological studies and endoscopic ultrasound.

Pathological features

Of the total tumors, 109 were histologically grade 3, corresponding to 50.5%, whereas grade 1 was presented in 57 (26.4%) cases. Regarding budding, there was a mean of 6.3 \pm 5.7 buds per 0.785 mm² (range, 0-45). In 23 (10.6%) cases, lymphovascular invasion was identified.

For tumor-associated inflammatory infiltrate, mainly lymphocytes and plasma cells were observed; in 41 (19%) cases, nodular lymphoid aggregates were immediately adjacent to the neoplastic cells. The mean number of intratumoral lymphocytes was 1.69 \pm 1.74 / mm² (range, 0 to 10).

Of the 216 cases, a dMMR system was demonstrated in 63 (29.2%) patients, with the main alterations in the co-absence of PMS2 and MLH1 expression (74.6%) followed by an isolated loss of MSH6 expression (25.4%) and an isolated loss of MSH2 expression (17.4%).

Univariate analysis

The basal characteristics of the patients are summarized in Table 1, which shows that the tumors with a higher pCR rate were associated with grade 1 tumors, dMMR, an increased presence of intratumoral lymphocytes, and the absence of vascular invasion in the pretreatment biopsy. Factors that were statistically associated with the presentation of pCR were dMMR, the absence of lymphovascular invasion, and low intratumoral budding

Table 1 Clinical-pathological characteristic of 216 patients with rectal carcinoma with preoperative chemo-radiotherapy, divided by type of pathological response *n* (%)

	No pathological complete response group (controls) (<i>n</i> = 144)	Pathological complete response group (cases) (<i>n</i> = 72)	<i>P</i>
Median age (yr ± SD)	56.67 ± 11.94	57.22 ± 13.65	0.759 ³
Sex			
Female	48 (33.3)	24 (33.3)	1.0 ¹
Male	96 (66.7)	48 (66.7)	
Histologic grade			
G1	30 (20.8)	27 (37.5)	
G2	44 (30.6)	6 (8.3)	< 0.001 ¹
G3	70 (48.6)	39 (54.2)	0.147 ¹
Budding			
Low (0-4 buds)	63 (43.8)	44 (61.6)	0.016 ¹
High (5 or more buds)	81 (56.2)	28 (38.4)	
Mismatch repair status			
MMR proficient	112 (77.8)	41 (56.9)	0.001 ¹
MMR defective	32 (22.2)	31 (43.1)	
CDX2 expression			
Absent	8 (5.6)	7 (9.7)	0.195 ¹
Present	136 (94.4)	65 (90.3)	
MLH1			
Non expressed	23 (16)	26 (36.1)	0.001 ¹
Normal	121 (84)	46 (63.9)	
PMS2			
Non expressed	23 (16)	26 (36.1)	0.001 ¹
Normal	121 (84)	46 (63.6)	
MSH2			
Non expressed	11 (7.6)	0	0.010 ²
Normal	133 (92.4)	72 (100)	
MSH6			
Non expressed	12 (8.3)	4 (5.6)	0.587 ²
Normal	132 (91.7)	68 (94.4)	
Lymphovascular invasion			
Absent	130 (90.3)	63 (87.5)	0.105 ¹
Present	14 (9.7)	9 (12.5)	

¹Value calculated from *Chi square* test with the first group as the reference; ²Values calculated from *Fischer's exact* test; ³Value calculated from a Student *t*-test. MMR: Mismatch repair system; SD: Standard deviation.

(Table 2).

Multivariate analysis

The factors associated with pCR were a dMMR status and a low degree of budding (Table 2).

DISCUSSION

We found an independent prognostic value of dMMR and intratumoral budding in pretreatment biopsies of patients with rectal adenocarcinoma treated with nCRT. The rest of the factors examined did not correlate with pCR. Our study is the first to demonstrate the independent predictive value of intratumoral budding for increased pCR in these patients.

To date, no clinical, endoscopic, pathological, radiological or molecular features have been sensitive and specific enough to predict pCR. Several studies report promising results and a few have investigated the pretreatment histopathological features. Some of the characteristics associated with pCR are early T and N stage, small tumor size, histologic low-grade and low carcinoembryonic antigen serum levels^[14,15]. However,

these results are controversial^[16].

We found that a dMMR status was associated with pCR. This is explained by the fact that the ability of a cell to survive is affected by defects in DNA repair against damage. These mechanisms in the rectal cancer response have been poorly studied because of the consensus that in colon cancer a dMMR is associated with poor response to fluoropyrimidine^[17]. In the setting of concomitant radiation, the impact of fluoropyrimidine given to dMMR tumors is controversial, and the predictive and prognostic roles of MMR genes in colorectal cancer are still unclear^[18]. Huh *et al*^[19] evaluated MMR protein expression in 209 patients with locally advanced rectal cancers treated with nCRT and subsequent surgery, where a pCR of 14.4% was observed. They did not observe any differences in the MMR protein expressions between patients with pCR and patients without pCR. In another study, de Rosa *et al*^[6], studied 62 dMMR colorectal cancers and identified 29 with stage II -III cancers who received nCRT and surgical resection and found that 8 (27.6%) had a pCR; this pCR was elevated compared to a rate of 18% reported by Brouquet *et al*^[20]. As is clear, there is some evidence pointing out the possible value

Table 2 Analysis of 216 patients with rectal carcinoma with preoperative chemo-radiotherapy, regarding their association with presence of complete pathological response

	Univariate analysis			Multivariate analysis ¹		
	OR	P	95%CI	OR	P	95%CI
MMR status (defective <i>vs</i> proficient)	2.64	0.002	1.438-4.870	2.61	0.004	1.355-5.040
Lymphovascular invasion (present <i>vs</i> absent)	0.7	0.094	0.066-2.860			
Histologic Grade (G1-G2 <i>vs</i> G3)	1.24	0.442	0.709-2.203			
Tumor budding [Low budding (0-4 buds) <i>vs</i> High budding (5 or more buds)]	2.495	0.017	1.278-4.881	2.52	0.025	1.366-4.894
Inflammatory infiltrate (present <i>vs</i> absent)	1.194	0.624	0.587-2.429			

¹Adjusted by MMR status and high tumor budding. OR: Odds ratio; CI: Confidence interval; MMR: Mismatch repair system.

of MMR proteins in the prediction of pCR. We found an increase in the pCR in patients with the dMMR system.

We found that 29% of our cases showed dMMR, which was a higher number than most reported studies. However, this high dMMR prevalence was according to the dMMR prevalence in Latin American studies^[10,21].

Tumor budding is observed in approximately 40% of colorectal cancer cases, and several publications have informed the prognostic value of tumor budding in stage II^[22,23]. However, it has not been studied in patients prone to receive nCRT, and its predictive value for pCR has not been studied. Tumor budding has been studied in both biopsies and surgical specimens, and there is a lot of controversy about where and how to measure tumor buds^[13]. In our study, we reported the number of buds in the intratumoral stroma of the biopsies and dichotomized the tumor budding into two grades. The consensus recommends the evaluation in the invasive front of the surgical specimens, which clearly demonstrates its prognostic significance, even when evaluated in biopsy specimens^[24].

It is believed that tumor budding represents the first step in cancer metastasis and is considered a histological representation of the epithelial-mesenchymal transition (EMT). This hypothesis has not been validated thus far, and the mechanisms by which the bud cells separate from the main tumor are unclear. Individual cells at the stroma are thought to migrate through the extracellular matrix, invade lymphovascular structures, and form colonies of metastatic tumors in the lymph nodes and distant sites^[24]. Additionally, tumor cells that undergo EMT are characterized by a gene expression switch from genes associated with epithelial differentiation to genes associated with mesenchymal properties^[25]. We think that these events give neoplastic cells an intrinsic capacity to resist nCRT. Although the underlying mechanisms are not clear, this is a future line of investigation.

The last outcome evaluated by our study was the inflammatory infiltrate, which did not demonstrate an association with pCR. This parameter was very difficult to evaluate because of the complexity of the cellular response at tumor invasion. More complex studies specifically aimed to explore this response are needed.

The main limitation of our study was the unexplored

reproducibility of the intratumoral budding in the literature in general, especially in the biopsy specimens. MMR protein evaluation is also a potential source of bias because, sometimes, the heterogeneous immunohistochemical reaction of the proteins is difficult to interpret. However, the standardized technique in our center (with external validation at UK-NEQAS), the consensus evaluation of the reaction, and the evaluation in whole tissue slides decreased this bias.

In conclusion, the response of rectal cancer to nCRT is a variable and very complex phenomenon. There is no individual molecular or genetic mechanism clearly associated with pCR. The key action that may help to predict pCR is a model that involves the combination of data from imaging, endoscopy, immunohistochemistry, gene expression, and histopathological data (such as intratumoral budding and dMMR).

ARTICLE HIGHLIGHTS

Research background

Several histopathological, immunohistochemical, and molecular markers have been analyzed in rectal carcinoma in an attempt to predict the pathological complete response (pCR) to neoadjuvant chemoradiotherapy (nCRT) with contradictory results.

Research motivation

Identifying characteristics of tumors that will respond to nCRT could help avoid surgery.

Research objectives

To determine whether the association of rectal adenocarcinoma with a defective-Mismatch repair system (dMMR) are associated with the rate of pCR. To identify histologic features in the diagnostic biopsy associated with the rate of pCR.

Research methods

A case-control study design paired 2:1 was performed.

Research results

The pCR was associated with well-differentiated tumors, dMMR, the absence of vascular invasion, and low tumor budding in the diagnostic biopsy. In the multivariate analysis, the factors independently associated with the pCR were dMMR and a low degree of tumor budding.

Research conclusions

Tumors with pCR were associated with the dMMR status and low tumor

budding in the diagnostic biopsy.

Research perspectives

Adding the status of both MMR and tumor budding to the current and future prognostic indexes could help predict the pCR of a rectal adenocarcinoma and evaluate alternative strategies for the care of these patients.

REFERENCES

- 1 **Torre LA**, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; **65**: 87-108 [PMID: 25651787 DOI: 10.3322/caac.21262]
- 2 **Janjan NA**, Khoo VS, Abbruzzese J, Pazdur R, Dubrow R, Cleary KR, Allen PK, Lynch PM, Globler G, Wolff R, Rich TA, Skibber J. Tumor downstaging and sphincter preservation with preoperative chemoradiation in locally advanced rectal cancer: the M. D. Anderson Cancer Center experience. *Int J Radiat Oncol Biol Phys* 1999; **44**: 1027-1038 [PMID: 10421535 DOI: 10.1016/S0360-3016(99)00099-1]
- 3 **Yeo SG**, Kim DY, Kim TH, Chang HJ, Oh JH, Park W, Choi DH, Nam H, Kim JS, Cho MJ, Kim JH, Park JH, Kang MK, Koom WS, Kim JS, Nam TK, Chie EK, Kim JS, Lee KJ. Pathologic complete response of primary tumor following preoperative chemoradiotherapy for locally advanced rectal cancer: long-term outcomes and prognostic significance of pathologic nodal status (KROG 09-01). *Ann Surg* 2010; **252**: 998-1004 [PMID: 21107110 DOI: 10.1097/SLA.0b013e3181f3f1b1]
- 4 **García-Aguilar J**, Hernandez de Anda E, Sirivongs P, Lee SH, Madoff RD, Rothenberger DA. A pathologic complete response to preoperative chemoradiation is associated with lower local recurrence and improved survival in rectal cancer patients treated by mesorectal excision. *Dis Colon Rectum* 2003; **46**: 298-304 [PMID: 12626903 DOI: 10.1007/s10350-004-6545-x]
- 5 **Kim NK**, Hur H. New Perspectives on Predictive Biomarkers of Tumor Response and Their Clinical Application in Preoperative Chemoradiation Therapy for Rectal Cancer. *Yonsei Med J* 2015; **56**: 1461-1477 [PMID: 26446626 DOI: 10.3349/ymj.2015.56.6.1461]
- 6 **de Rosa N**, Rodriguez-Bigas MA, Chang GJ, Veerapong J, Borras E, Krishnan S, Bednarski B, Messick CA, Skibber JM, Feig BW, Lynch PM, Vilar E, You YN. DNA Mismatch Repair Deficiency in Rectal Cancer: Benchmarking Its Impact on Prognosis, Neoadjuvant Response Prediction, and Clinical Cancer Genetics. *J Clin Oncol* 2016; **34**: 3039-3046 [PMID: 27432916 DOI: 10.1200/JCO.2016.66.6826]
- 7 **Lynch HT**, Lynch PM, Lanspa SJ, Snyder CL, Lynch JF, Boland CR. Review of the Lynch syndrome: history, molecular genetics, screening, differential diagnosis, and medicolegal ramifications. *Clin Genet* 2009; **76**: 1-18 [PMID: 19659756 DOI: 10.1111/j.1399-0004.2009.01230.x]
- 8 **Quirke P**, Steele R, Monson J, Grieve R, Khanna S, Couture J, O'Callaghan C, Myint AS, Bessell E, Thompson LC, Parmar M, Stephens RJ, Sebag-Montefiore D; MRC CR07/NCIC-CTG CO16 Trial Investigators; NCRI Colorectal Cancer Study Group. Effect of the plane of surgery achieved on local recurrence in patients with operable rectal cancer: a prospective study using data from the MRC CR07 and NCIC-CTG CO16 randomised clinical trial. *Lancet* 2009; **373**: 821-828 [PMID: 19269520 DOI: 10.1016/S0140-6736(09)60485-2]
- 9 **Lino-Silva LS**, García-Gómez MA, Aguilar-Romero JM, Domínguez-Rodríguez JA, Salcedo-Hernández RA, Loeza-Belmont R, Ruiz-García EB, Herrera-Gómez Á. Mesorectal pathologic assessment in two grades predicts accurately recurrence, positive circumferential margin, and correlates with survival. *J Surg Oncol* 2015; **112**: 900-906 [PMID: 26487289 DOI: 10.1002/jso.24076]
- 10 **Gupta S**, Ashfaq R, Kapur P, Afonso BB, Nguyen TP, Ansari F, Boland CR, Goel A, Rockey DC. Microsatellite instability among individuals of Hispanic origin with colorectal cancer. *Cancer* 2010; **116**: 4965-4972 [PMID: 20665498 DOI: 10.1002/cncr.25486]
- 11 **Edge SB**, Byrd DR, Compton CC. American Joint Committee on Cancer (AJCC) Cancer Staging Manual. 7th ed. Chicago IL: Springer, Inc: 2010, 123-129
- 12 **Nowak JA**, Hornick JL. Molecular Evaluation of Colorectal Adenocarcinoma: Current Practice and Emerging Concepts. *Surg Pathol Clin* 2016; **9**: 427-439 [PMID: 27523970 DOI: 10.1016/j.path.2016.04.007]
- 13 **Lugli A**, Kirsch R, Ajioka Y, Bosman F, Cathomas G, Dawson H, El Zimaity H, Fléjou JF, Hansen TP, Hartmann A, Kakar S, Langner C, Nagtegaal I, Puppa G, Riddell R, Ristimäki A, Sheahan K, Smyrk T, Sugihara K, Terris B, Ueno H, Vieth M, Zlobec I, Quirke P. Recommendations for reporting tumor budding in colorectal cancer based on the International Tumor Budding Consensus Conference (ITBCC) 2016. *Mod Pathol* 2017; **30**: 1299-1311 [PMID: 28548122 DOI: 10.1038/modpathol.2017.46]
- 14 **Garland ML**, Vather R, Bunkley N, Pearse M, Bissett IP. Clinical tumour size and nodal status predict pathologic complete response following neoadjuvant chemoradiotherapy for rectal cancer. *Int J Colorectal Dis* 2014; **29**: 301-307 [PMID: 24420737 DOI: 10.1007/s00384-013-1821-7]
- 15 **Qiu HZ**, Wu B, Xiao Y, Lin GL. Combination of differentiation and T stage can predict unresponsiveness to neoadjuvant therapy for rectal cancer. *Colorectal Dis* 2011; **13**: 1353-1360 [PMID: 21689282 DOI: 10.1111/j.1463-1318.2011.02570.x]
- 16 **Kalady MF**, de Campos-Lobato LF, Stocchi L, Geisler DP, Dietz D, Lavery IC, Fazio VW. Predictive factors of pathologic complete response after neoadjuvant chemoradiation for rectal cancer. *Ann Surg* 2009; **250**: 582-589 [PMID: 19710605 DOI: 10.1097/SLA.0b013e3181b91e63]
- 17 **Ribic CM**, Sargent DJ, Moore MJ, Thibodeau SN, French AJ, Goldberg RM, Hamilton SR, Laurent-Puig P, Gryfe R, Shepherd LE, Tu D, Redston M, Gallinger S. Tumor microsatellite-instability status as a predictor of benefit from fluorouracil-based adjuvant chemotherapy for colon cancer. *N Engl J Med* 2003; **349**: 247-257 [PMID: 12867608 DOI: 10.1056/NEJMoa022289]
- 18 **Yoon YS**, Yu CS, Kim TW, Kim JH, Jang SJ, Cho DH, Roh SA, Kim C. Mismatch repair status in sporadic colorectal cancer: immunohistochemistry and microsatellite instability analyses. *J Gastroenterol Hepatol* 2011; **26**: 1733-1739 [PMID: 21615788 DOI: 10.1111/j.1440-1746.2011.06784.x]
- 19 **Huh JW**, Kim HC, Kim SH, Park YA, Cho YB, Yun SH, Lee WY, Park HC, Choi DH, Park JO, Park YS, Chun HK. Mismatch Repair Gene Expression as a Predictor of Tumor Responses in Patients With Rectal Cancer Treated With Preoperative Chemoradiation. *Medicine (Baltimore)* 2016; **95**: e2582 [PMID: 26817916 DOI: 10.1097/MD.0000000000002582]
- 20 **Brouquet A**, Mortenson MM, Vauthey JN, Rodriguez-Bigas MA, Overman MJ, Chang GJ, Kopetz S, Garrett C, Curley SA, Abdalla EK. Surgical strategies for synchronous colorectal liver metastases in 156 consecutive patients: classic, combined or reverse strategy? *J Am Coll Surg* 2010; **210**: 934-941 [PMID: 20510802 DOI: 10.1016/j.jamcollsurg.2010.02.039]
- 21 **Egoavil CM**, Montenegro P, Soto JL, Casanova L, Sanchez-Lihon J, Castillejo MI, Martinez-Canto A, Perez-Carbonell L, Castillejo A, Guarinos C, Barbera VM, Jover R, Paya A, Alenda C. Clinically important molecular features of Peruvian colorectal tumours: high prevalence of DNA mismatch repair deficiency and low incidence of KRAS mutations. *Pathology* 2011; **43**: 228-233 [PMID: 21436632 DOI: 10.1097/PAT.0b013e3283437613]
- 22 **Betge J**, Kornprat P, Pollheimer MJ, Lindtner RA, Schlemmer A, Rehak P, Vieth M, Langner C. Tumor budding is an independent predictor of outcome in AJCC/UICC stage II colorectal cancer. *Ann Surg Oncol* 2012; **19**: 3706-3712 [PMID: 22669453 DOI: 10.1245/s10434-012-2426-z]
- 23 **Okuyama T**, Oya M, Ishikawa H. Budding as a risk factor for lymph node metastasis in pT1 or pT2 well-differentiated colorectal adenocarcinoma. *Dis Colon Rectum* 2002; **45**: 628-634 [PMID: 12004212 DOI: 10.1007/s10350-004-6259-0]
- 24 **Lugli A**, Karamitopoulou E, Zlobec I. Tumour budding: a promising parameter in colorectal cancer. *Br J Cancer* 2012; **106**:

- 1713-1717 [PMID: 22531633 DOI: 10.1038/bjc.2012.127]
25 **Bhangu A**, Wood G, Mirnezami A, Darzi A, Tekkis P, Goldin R.
Epithelial mesenchymal transition in colorectal cancer: Seminal

role in promoting disease progression and resistance to neoadjuvant
therapy. *Surg Oncol* 2012; **21**: 316-323 [PMID: 22981546 DOI:
10.1016/j.suronc.2012.08.003]

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Retrospective Cohort Study

**Interconversion of two commonly used performance tools:
An analysis of 5844 paired assessments in 1501 lung cancer
patients**

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Abstract**AIM**

To establish the Karnofsky performance status (KPS) categories which would facilitate the interconversion of the KPS scale to the Eastern Cooperative Oncology Group (ECOG) performance status (PS) scale.

METHODS

This was a retrospective analysis of all patients attending the lung cancer clinic at a tertiary care center over a 5-year period (September 2009 to August 2014). All patients were assessed with both KPS and ECOG PS scales at each visit. Correlation between KPS and ECOG PS was assessed using Spearman's correlation coefficient. KPS categories equivalent to ECOG PS scores were compared using hit rate and weighted kappa (κ_w).

RESULTS

A total of 1501 patients were assessed over the study period, providing 5844 paired KPS and ECOG PS assess-

ments. The study cohort had a mean (standard deviation; SD) age of 58.4 (10.8) years, with the majority being current or ex-smokers (76.9%) and males (82.3%). Non-small cell lung cancer was the most common histological type ($n = 1196$, 79.7%) with the majority having advanced (stage IIIB/IV) disease (83.4%). Mean baseline KPS and ECOG PS scores were 77.6 (SD = 14.4) and 1.5 (SD = 1) respectively. The most frequent KPS score was 80 (29%), and the most frequent ECOG PS score was 1 (43%). The overall correlation between KPS and ECOG PS was good (Spearman $r = -0.84$, $P < 0.0001$) but ranged from -0.727 to -0.972 between visits. KPS categories derived from our cohort [10-40 (ECOG 4), 50-60 (ECOG 3), 70 (ECOG 2), 80-90 (ECOG 1), 100 (ECOG 0)] performed better [hit rate 78.1%, $\kappa_w = 0.749$ (0.736-0.762) $P < 0.0001$] than those suggested in the past literature.

CONCLUSION

The current study provides the largest set of paired KPS-ECOG assessments to date. We suggest that the KPS categories 10-40, 50-60, 70, 80-90, and 100 are equivalent to ECOG PS categories of 4, 3, 2, 1, and 0 respectively.

Key words: Karnofsky performance status; Eastern Cooperative Oncology Group; Performance status; Lung cancer; Chemotherapy

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Core tip: Karnofsky performance status (KPS) scale and Eastern Cooperative Oncology Group (ECOG) scale are the most commonly used performance status (PS) tools worldwide for patients with cancer. Since the number of scoring points in each scale is different, these scales are not readily interconvertible. However, most clinical studies use only one of these two scales (either KPS or ECOG PS) to assess PS, rendering interpopulation comparisons difficult. In this study, we analyze the largest set of paired KPS-ECOG assessments to date in a cohort of lung cancer patients for a solution.

Prasad KT, Kaur H, Muthu V, Aggarwal AN, Behera D, Singh N. Interconversion of two commonly used performance tools: An analysis of 5844 paired assessments in 1501 lung cancer patients. *World J Clin Oncol* 2018; 9(7): 140-147 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i7/140.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i7.140>

INTRODUCTION

Performance status (PS) is a measure of a subject's ability to perform the activities of daily living. PS has been shown to predict survival in patients with cancer^[1-4]. Patients with poor PS often do not tolerate more aggressive treatment strategies like surgery

or chemotherapy, and, hence, PS is often taken into account while deciding the therapeutic strategy for cancer. It is a well-known fact that patients with poor PS are excluded from most clinical trials in cancer^[5,6].

Several tools are available for the assessment of PS, including the Karnofsky performance status (KPS) scale, the Eastern Cooperative Oncology Group (ECOG) PS scale, the palliative performance scale, and the Australia-modified KPS. Among these tools, the KPS scale and the ECOG PS scale are the most commonly used worldwide^[7]. The KPS scale is an 11-point numerical scale, with scores ranging from 100 (normal functional status) to 0 (death), in decremental steps of 10^[8]. The ECOG PS scale is a 6-point numerical scale, with scores ranging from 0 (normal functional status) to 5 (death), in incremental steps of 1^[9]. No conclusive evidence exists in the literature to suggest that one scale is better than the other. However, the ECOG PS scale is often preferred, as it is simpler to apply with a smaller number of choices. Both the scores have been shown to be good predictors of mortality^[1,4,10,11]. The ECOG PS may have a slightly better prognostic value compared to KPS^[10]. Although both the scales have been used to predict treatment response in lung cancer, results have been variable^[12].

Several studies have shown PS assessment made using the KPS and ECOG PS by different healthcare professionals (doctors, medical students, nurses) to have moderate to high interobserver correlation, albeit with considerable variation^[13-16]. Assessments made by technical staff or patients have been shown to have a relatively larger variability compared to assessments by healthcare workers^[11,13,16]. These differences could be attributed to variation in the level of overall training and exposure of the individual, which might affect the interpretation of the existing disability. Moreover, interobserver variability can get aggravated when assessments are made on patients with lower KPS scores^[17]. Closer attention to certain behavioral issues might help to improve the PS assessments in such situations^[17]. However, neither of these two scales have been shown to be consistently superior to the other with respect to interobserver variability^[13-15].

Since PS is an important factor which can influence clinical outcomes, researchers making interpopulation comparisons across studies should ensure that the PSs of the populations analyzed are equivalent. However, most clinical trials usually utilize only one of the two scales in their study population (either KPS or ECOG PS)^[7]. Hence, the comparison of PS between different patient populations is difficult as the two scales are not readily interconvertible, with different number of scoring points (11 vs 6). To overcome this hindrance, several attempts have been made to examine the possibility of interconversion between KPS and ECOG PS scales (Table 1)^[7,10,18-20]. Despite these suggestions, confusion still exists regarding the optimal KPS categories for interconversion to ECOG. Herein, we attempt to determine the KPS categories equivalent to ECOG PS

Table 1 Karnofsky performance status categories suggested for interconversion to Eastern Cooperative Oncology Group performance status in the literature

Author, year	n and patients, assessments	Setting	KPS categories	ECOG
AJCC ^[34] , 1977	-	-	90-100	0
			70-80	1
			50-60	2
			30-40	3
			10-20	4
Minna <i>et al</i> ^[20] , 1985	-	-	100	0
			80-90	1
			60-70	2
			40-50	3
			20-30	4
Buccheri <i>et al</i> ^[10] , 1996	536 (1656)	Subjects with lung cancer visiting a cancer clinic of a tertiary care centre	80-100	0-1
			60-70	2
			10-50	3-4
Ma <i>et al</i> ^[19] , 2010	1385 (1385)	Subjects visiting an oncology palliative care clinic, or admitted to an acute cancer palliative care unit	100	0
			80-90	1
			60-70	2
			40-50	3
			10-30	4
de Kock <i>et al</i> ^[18] , 2013	955 (674)	Subjects with advanced life-limiting illnesses (cancer and non-cancer) in acute care and community settings	60-100	1
			50	2
			30-40	3
			10-20	4
Current study	1501 (5844)	Subjects with lung cancer visiting a cancer clinic of a tertiary care center	100	0
			80-90	1
			70	2
			50-60	3
			10-40	4

AJCC: American Joint Committee on Cancer; ECOG: Eastern Cooperative Oncology Group; KPS: Karnofsky performance status; PS: Performance status.

scores by analyzing data from a large cohort of lung cancer patients.

MATERIALS AND METHODS

Study design

We performed a retrospective analysis of data collected in the lung cancer clinic of our center over a 5-year period (September 2009 to August 2014). Informed consent was obtained from all the subjects.

Patients

All subjects with lung cancer who visited the lung cancer clinic for chemotherapy and had at least one paired assessment of KPS and ECOG PS were included in the study. Subjects with a diagnosis of intrathoracic malignancy other than lung cancer were not included in this study. Subjects who did not receive chemotherapy and received only alternative forms of therapy, like surgery, radiotherapy, or targeted therapy, were also excluded from this study. The PS assessments made during the first 10 visits of each patient, starting from the date of the first cycle of chemotherapy, were included in this study.

Standard of care

The subjects were treated appropriately with chemotherapy, tyrosine kinase inhibitors, radiotherapy, or

surgery as guided by the tumor histopathology, mutation status, and clinical status, as described previously^[21-24]. Briefly, subjects with adenocarcinoma without any driver mutation were treated with pemetrexed-based platinum doublet followed by maintenance pemetrexed therapy until disease progression. Subjects with squamous histology were treated with docetaxel or gemcitabine-based platinum doublet. Subjects with small cell lung cancer received irinotecan-based platinum doublet. All patients receiving chemotherapy were administered at least four cycles of chemotherapy before response assessment. Subjects who showed a partial response received an additional two cycles, for a total of six cycles. Subjects with sensitizing *EGFR* gene mutation or *ALK* gene rearrangements were treated with appropriate *EGFR* tyrosine kinase or *ALK* inhibitors, respectively.

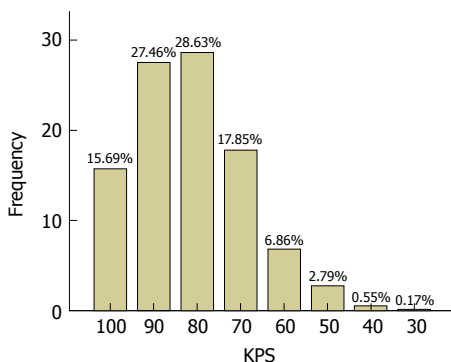
Data collection and assessments

All subjects underwent a systematic assessment at baseline and follow-up as described previously^[25-29]. At baseline, the following parameters were collected: age, gender, smoking status, body mass index, tumor histopathology, TNM stage (7th edition), KPS, and ECOG-PS^[30]. The frequency of scheduled visits to the clinic by the patient varied depending upon the ongoing treatment modality. At every visit to the lung cancer clinic, the subjects were assessed with both the KPS and the ECOG PS scales by the treating physician.

Table 2 Baseline characteristics of the study population *n* (%)

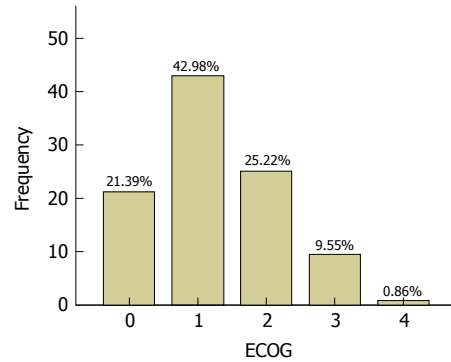
Characteristic	Total, <i>n</i> = 1501
Age in yr	58.4 (10.8)
Males	1236 (82.3)
Smokers	1155 (76.9)
Body mass index in kg/m ²	20.2 (4.2)
Histopathology	
NSCLC: Squamous	553 (36.8)
NSCLC: Adeno	514 (34.2)
NSCLC: Undifferentiated	87 (5.8)
NSCLC: Other	42 (2.8)
SCLC	305 (20.3)
NSCLC/SCLC stage ¹	
I and II	44 (2.9)
IIIA	193 (12.9)
IIIB	426 (28.4)
IV	767 (51.1)
SCLC stage	
Limited disease	145 (9.7)
Extensive disease	160 (10.7)
Baseline performance status	
KPS	77.6 (14.4)
ECOG PS	1.5 (1)

¹Total does not add up to 1501 as TNM staging was available for only some of the patients with SCLC. All values are mean (standard deviation) unless otherwise specified. ECOG PS: Eastern Cooperative Oncology Group performance status; KPS: Karnofsky performance status; NSCLC: Non-small cell lung cancer; SCLC: Small cell lung cancer.

**Figure 1** Frequency distribution of karnofsky performance status scores (*n* = 5844 assessments). KPS: Karnofsky performance status.

Statistical analysis

Descriptive data are presented as numbers and percentages or mean \pm SD unless otherwise stated. Correlation between KPS and ECOG PS was assessed using Spearman's correlation coefficient (*r*), with values ranging from -1.00 to +1.00. An *r*-value of -1.00 was considered to represent a perfect negative correlation and a value of +1.00 was considered to represent a perfect positive correlation^[31]. The original 11-point KPS scale was converted into a 5-point scale for comparison with the ECOG PS. The agreement between these KPS categories and the actual ECOG PS measurements was assessed using hit rate and the weighted kappa coefficient (κ_w). Hit rate was calculated as the proportion of assessments with a perfect agreement between the KPS categories and

**Figure 2** Frequency distribution of Eastern Cooperative Oncology Group status scores (*n* = 5844 assessments). ECOG: Eastern Cooperative Oncology Group.

the measured ECOG PS scale. The κ_w was calculated to assess the level of agreement beyond chance. Agreement between the scales was classified based on the kappa values as poor (< 0.00), slight (0.00 to 0.20), fair (0.21 to 0.40), moderate (0.41 to 0.60), substantial (0.61 to 0.80), or almost perfect (0.81 to 1.00)^[32]. Statistical analyses were performed with the help of Statistical Package for Social Sciences software (IBM SPSS Statistics, version 22; IBM Corporation, Armonk, NY, United States). All statistical tests were performed as two-sided. *P* < 0.05 was considered statistically significant.

RESULTS

During the study period, 1664 patients visited the lung cancer clinic. Among them, 150 patients did not receive any chemotherapy and were excluded. An additional 13 patients who had intrathoracic malignancies other than lung cancer were also excluded. A total of 1501 patients were assessed during the study period, providing 5844 paired PS assessments (KPS and ECOG PS). The mean (standard deviation; SD) age was 58.4 (10.8) years, with the majority being current or ex-smokers (76.9%) and males (82.3%) (Table 2). Non-small cell lung cancer was the most common histological type (*n* = 1196, 79.7%), with the majority having advanced (stage III B or IV) disease (83.4%).

Mean baseline KPS and ECOG PS scores were 77.6 (SD = 14.4) and 1.5 (SD = 1) respectively. The most frequent KPS score was 80 (29%) (Figure 1) and the most frequent ECOG PS score was 1 (43%) (Figure 2). Among the total of 5844 paired PS assessments, a vast majority of the KPS scores were between 70 and 100 (89.6%) and similarly most ECOG PS scores were between 0 and 2 (89.6%). The overall correlation between KPS and ECOG PS was good [Spearman ρ = (-) 0.84, *P* < 0.0001] but ranged from -0.727 to -0.972 between visits (Table 3 and Figure 3). As expected, the number of assessments available at each progressive follow-up visit showed a reduction

Table 3 Correlation between Karnofsky performance status and Eastern Cooperative Oncology Group performance status

Visit number	Number of assessments	Spearman's correlation coefficient ¹
1	1426	-0.727
2	1099	-0.890
3	957	-0.876
4	823	-0.863
5	602	-0.858
6	464	-0.867
7	270	-0.901
8	129	-0.854
9	54	-0.773
10	20	-0.972
Overall	5844	-0.840

¹ $P < 0.0001$ for all individual visits and overall.

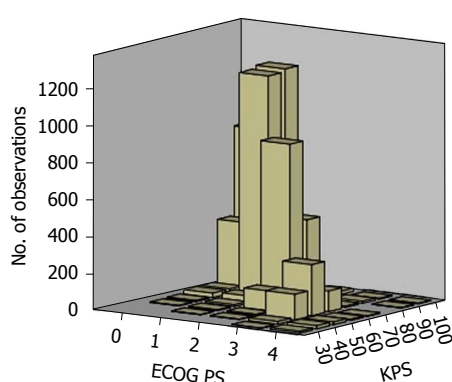


Figure 3 Distribution of karnofsky performance status and Eastern Cooperative Oncology Group performance status scores ($n = 5844$ paired assessments). ECOG PS: Eastern Cooperative Oncology Group performance status; KPS: Karnofsky performance status.

due to reduction in number of patients due to death or loss to follow-up.

Visual inspection of cross-tabulated KPS and ECOG PS data was performed to arrive at the best possible KPS categories to allow conversion of KPS to ECOG PS scale, namely: 100 (ECOG 0), 80-90 (ECOG 1), 70 (ECOG 2), 50-60 (ECOG 3), and 10-40 (ECOG 4)^[10]. We also analyzed the performance of KPS categories suggested in the past literature, on our cohort (Table 4). We found a substantial agreement with all the suggested conversion categories, except those suggested by de Kock *et al.*^[18] and the American Joint Committee on Cancer (AJCC), which had slight and fair agreements respectively. KPS categories derived from our cohort performed better (hit rate 78.4%, $\kappa_w = 0.674$) than those suggested in the past literature^[19,33]. The highest hit rate (83.2%) was observed with the interconversion categories suggested by Bucerri *et al.*^[10]. The highest κ_w was observed with the categories derived from our cohort ($\kappa_w = 0.749$). The interconversion categories suggested by de Kock *et al.*^[18] had the worst hit rate (43.2%) and κ_w (0.079).

DISCUSSION

In this study, we propose KPS categories which can aid

interconversion of KPS scores to ECOG PS scores by retrospectively analyzing paired PS assessments (KPS and ECOG PS) made in a large cohort of lung cancer patients. We also compare the performance of KPS categories in previous literature for interconversion of KPS to ECOG PS on our cohort.

It is well-known that the PS is a predictor of mortality^[1-4]. Hence, when comparing clinical characteristics or the outcomes of a therapeutic modality in different patient populations with lung cancer, it is essential that their PS is also matched. Failure to do so may result in erroneous conclusions. However, such comparisons are difficult when the tool used for assessment of PS in the study population is different (either KPS or ECOG PS). To overcome this difficulty, several investigators have suggested KPS categories for interconversion to ECOG PS scale (Table 1)^[7,10,18-20].

The AJCC and Minna *et al.*^[20] were among the first to have suggested KPS categories for interconversion to ECOG PS^[34]. However, these suggestions were not evidence-based. Verger *et al.*^[7] compared these two suggestions in a cohort of 150 patients with cancer attending a radiotherapy clinic and found that the categories suggested by Minna *et al.*^[20] performed better. They also stressed that at lower PS levels, interconversion between the two scores would be difficult as they observed a wide spread of values. Subsequently, Bucerri *et al.*^[10] studied 536 patients with lung cancer and found that a 3-point conversion scale for KPS resulted in the best agreement (hit rate 84%) between the two PS scales. Ma and colleagues had assessed KPS and ECOG PS in 1385 subjects with various cancers attending or admitted to an oncology palliative care clinic^[19]. They analyzed several KPS categories and suggested a KPS category which closely resembled that suggested by Minna *et al.*^[20] and had the best combination of hit rate (75%) and κ_w (0.84, $P < 0.0001$). de Kock *et al.*^[18] studied 955 subjects with advanced life-limiting illnesses and suggested a 4-point scale, which had the best combination of hit rate (57%) and κ (0.7, 95%CI: 0.66-0.73).

Among all the interconversion categories for KPS suggested in the literature, the KPS categories which we suggest in this study had the best agreement with the actual PS measurements. Although, the categories suggested by Minna *et al.*^[20] and Ma *et al.*^[19] had the next best agreement, the categories suggested in our study appear more appropriate clinically. This can be illustrated by the following two examples. A patient who requires occasional assistance for daily activities (KPS 60), would be classified as ECOG 2 (capable of all self-care) using the categories suggested by Minna *et al.*^[20] or Ma *et al.*^[19], while they will be classified more appropriately as ECOG 3 (capable of limited self-care) using the categories suggested in the current study. Similarly, a patient who is disabled and requires special care and assistance (KPS 40) would be classified as ECOG 3 (capable of limited self-care) using the categories suggested by Minna *et al.*^[20] or Ma *et al.*^[19], while they will be classified more appropriately as ECOG

Table 4 Comparison of the performance of various suggested Karnofsky performance status categories (for interconversion to the Eastern Cooperative Oncology Group performance status scale) in the current cohort

Author, year	Hit rate ¹	κ_w ¹
AJCC <i>et al</i> ^[34] , 1977	43.60%	0.376 (0.363-0.389) $P < 0.0001$
Minna <i>et al</i> ^[20] , 1985	75.20%	0.701 (0.687-0.714) $P < 0.0001$
Buccheri <i>et al</i> ^[10] , 1996	83.20%	0.695 (0.679-0.711) $P < 0.0001$
Ma <i>et al</i> ^[19] , 2010	75.20%	0.701 (0.687-0.714) $P < 0.0001$
de Kock <i>et al</i> ^[18] , 2013	43.20%	0.079 (0.068-0.090) $P < 0.0001$
Current study	78.10%	0.749 (0.736-0.762) $P < 0.0001$

¹Performance in the current cohort shown as hit rate (%) or weighted kappa (95% confidence limits) along with the P -value.

4 (cannot carry out any self-care) using the categories suggested in the current study. It should also be borne in mind that a change in the ECOG scale by a single score in the above situations entails a change in 1-year survival by at least 10%^[4].

The highest hit rate (83.2%) was observed with the KPS categories suggested by Buccheri *et al*^[10]. However, this could have been because of the consolidation of the KPS categories into a smaller number of groups (3 groups instead of 5): KPS 10-50 (ECOG PS 3-4), KPS 60-70 (ECOG PS 2), KPS 80-100 (ECOG PS 0-1). Despite this, the level of agreement was lower when compared to the current study (κ_w 0.645 vs 0.749).

We observed the lowest hit rate (43.2%) and the least agreement ($\kappa_w = 0.079$) with the KPS categories suggested by de Kock *et al*^[18]. This observation could have been because a majority of the population in that study had a poor PS, which is known to affect the interobserver rating^[17]. In fact, 42.7% of their subjects had an ECOG PS of 3, and 66.2% had a KPS score between 30 and 50. Additionally, the authors did not assign KPS categories for ECOG PS 0.

Our study is not without limitations. The inherent limitations of retrospective studies apply to our study as well. Since the PS assessments were made by various treating physicians, there could have been some variability in the assessments. However, it has been demonstrated that both the KPS and ECOG PS scored by physicians have substantial interobserver reliability^[13-16]. Moreover, each paired assessment was made by a single observer. On the other hand, this presumed limitation could be considered as an advantage as our study is more likely to approximate a real-world scenario. A majority of our subjects had good PS (KPS score ≥ 70 and ECOG PS ≥ 2) as they visited the clinic for treatment purposes. Hence, the suggested KPS categories may not apply to patients with poorer PS (*e.g.*, palliative care clinics). Finally, we did not have survival data for our cohort, as this was not the intended purpose of the study. However, despite these limitations, the current study provides the largest set of paired KPS-ECOG assessments to date in a real-world setting.

In conclusion, we suggest that the KPS categories 10-40, 50-60, 70, 80-90, and 100 are equivalent to ECOG PS categories of 4, 3, 2, 1, and 0 respectively.

These categories may be useful for interconversion of KPS to ECOG PS scale when attempting to compare patient populations across different studies in whom investigators have used one of the two different scales (KPS or ECOG PS) for assessment of PS.

ARTICLE HIGHLIGHTS

Research background

Performance status (PS) is an estimate of a subject's ability to perform activities of daily living. Several tools are available to estimate the PS. Among them, the Karnofsky performance status (KPS) scale and the Eastern Cooperative Oncology Group (ECOG) scale are the most commonly used PS scales worldwide for patients with cancer. The KPS scale is an 11-point numerical scale, with scores ranging from 100 (normal functional status) to 0 (death), in decremental steps of 10. The ECOG PS scale is a 6-point numerical scale, with scores ranging from 0 (normal functional status) to 5 (death), in incremental steps of 1. Since the number of scoring points in each scale is different, these scales are not readily interconvertible.

Research motivation

PS is an important clinical factor which affects prognosis and influences treatment decisions in subjects with lung cancer. Hence, researchers who attempt to compare clinical characteristics or outcomes across different patient populations should ensure that their PS levels are matched. Failure to do so may result in erroneous conclusions. Most clinical studies employ only one of these two scales (either KPS or ECOG PS) in their study population for assessment of PS. When the PS scale used in the studies are different (either KPS or ECOG PS) this may lead to difficulty. Several investigators have tried to overcome this hindrance by suggesting KPS categories for interconversion to the ECOG PS scale. However, the performance of these suggested KPS categories has been variable.

Research objectives

We attempted to establish the KPS categories which would facilitate the interconversion of the KPS scale to the ECOG PS scale.

Research methods

We retrospectively analyzed the data of 1501 patients from a lung cancer clinic. In these patients, at every visit, paired assessments of PS had been made using both the KPS and ECOG PS scales by physicians. We also studied the performance of other KPS categories suggested in the literature, on our patient cohort. We used statistical methods called hit rate and weighted kappa to test the agreement between the KPS categories and the actual observations.

Research results

We found that the KPS categories 10-40, 50-60, 70, 80-90, and 100 were equivalent to ECOG PS categories of 4, 3, 2, 1, and 0 respectively. We also found that the agreement between the KPS categories suggested in the past literature (for interconversion to ECOG PS) and the paired KPS-ECOG PS assessments made in our cohort was variable.

Research conclusions

The current study is the largest set of paired KPS-ECOG assessments published in the literature in patients with lung cancer to date. The suggested KPS categories will facilitate interconversion of the KPS to the ECOG PS scale and will enhance communication between researchers utilizing either of the two scales.

Research perspectives

The KPS categories suggested in our study may be prospectively evaluated to test their validity. The applicability of the suggested categories may be evaluated in other populations to study the effect of cultural and regional variations.

REFERENCES

- 1 **Stanley KE.** Prognostic factors for survival in patients with inoperable lung cancer. *J Natl Cancer Inst* 1980; **65**: 25-32 [PMID: 6930515]
- 2 **Downing M,** Lau F, Lesperance M, Karlson N, Shaw J, Kuziemyk C, Bernard S, Hanson L, Olajide L, Head B, Ritchie C, Harrold J, Casarett D. Meta-analysis of survival prediction with Palliative Performance Scale. *J Palliat Care* 2007; **23**: 245-252; discussion 252-254 [PMID: 18251442]
- 3 **Maltoni M,** Caraceni A, Brunelli C, Broeckaert B, Christakis N, Eychmueller S, Glare P, Nabal M, Viganò A, Larkin P, De Conno F, Hanks G, Kaasa S; Steering Committee of the European Association for Palliative Care. Prognostic factors in advanced cancer patients: evidence-based clinical recommendations--a study by the Steering Committee of the European Association for Palliative Care. *J Clin Oncol* 2005; **23**: 6240-6248 [PMID: 16135490 DOI: 10.1200/jco.2005.06.866]
- 4 **Kawaguchi T,** Takada M, Kubo A, Matsumura A, Fukai S, Tamura A, Saito R, Maruyama Y, Kawahara M, Ignatius Ou SH. Performance status and smoking status are independent favorable prognostic factors for survival in non-small cell lung cancer: a comprehensive analysis of 26,957 patients with NSCLC. *J Thorac Oncol* 2010; **5**: 620-630 [PMID: 20354456 DOI: 10.1097/JTO.0b013e3181d2ded9]
- 5 **Paz-Ares LG,** de Marinis F, Dediu M, Thomas M, Pujol JL, Bidoli P, Molinier O, Sahoo TP, Laack E, Reck M, Corral J, Melemed S, John W, Chouaki N, Zimmermann AH, Visseren-Grul C, Gridelli C. PARAMOUNT: Final overall survival results of the phase III study of maintenance pemetrexed versus placebo immediately after induction treatment with pemetrexed plus cisplatin for advanced nonsquamous non-small-cell lung cancer. *J Clin Oncol* 2013; **31**: 2895-2902 [PMID: 23835707 DOI: 10.1200/JCO.2012.47.1102]
- 6 **Reck M,** Kaiser R, Mellemaard A, Douillard JY, Orlov S, Krzakowski M, von Pawel J, Gottfried M, Bondarenko I, Liao M, Gann CN, Barrueco J, Gaschler-Markefski B, Novello S; LUME-Lung 1 Study Group. Docetaxel plus nintedanib versus docetaxel plus placebo in patients with previously treated non-small-cell lung cancer (LUME-Lung 1): a phase 3, double-blind, randomised controlled trial. *Lancet Oncol* 2014; **15**: 143-155 [PMID: 24411639 DOI: 10.1016/S1470-2045(13)70586-2]
- 7 **Verger E,** Salameró M, Conill C. Can Karnofsky performance status be transformed to the Eastern Cooperative Oncology Group scoring scale and vice versa? *Eur J Cancer* 1992; **28A**: 1328-1330 [PMID: 1515244 DOI: 10.1016/0959-8049(92)90510-9]
- 8 **Schag CC,** Heinrich RL, Ganz PA. Karnofsky performance status revisited: reliability, validity, and guidelines. *J Clin Oncol* 1984; **2**: 187-193 [PMID: 6699671 DOI: 10.1200/jco.1984.2.3.187]
- 9 **Oken MM,** Creech RH, Tormey DC, Horton J, Davis TE, McFadden ET, Carbone PP. Toxicity and response criteria of the Eastern Cooperative Oncology Group. *Am J Clin Oncol* 1982; **5**: 649-655 [PMID: 7165009 DOI: 10.1097/00000421-198212000-00014]
- 10 **Buccheri G,** Ferrigno D, Tamburini M. Karnofsky and ECOG performance status scoring in lung cancer: a prospective, longitudinal study of 536 patients from a single institution. *Eur J Cancer* 1996; **32A**: 1135-1141 [PMID: 8758243 DOI: 10.1016/0959-8049(95)00664-8]
- 11 **Blagden SP,** Charman SC, Sharples LD, Magee LR, Gilligan D. Performance status score: do patients and their oncologists agree? *Br J Cancer* 2003; **89**: 1022-1027 [PMID: 12966419 DOI: 10.1038/sj.bjc.6601231]
- 12 **Yogananda MN,** Muthu V, Prasad KT, Kohli A, Behera D, Singh N. Utility of the revised Edmonton Symptom Assessment System (ESAS-r) and the Patient-Reported Functional Status (PRFS) in lung cancer patients. *Support Care Cancer* 2018; **26**: 767-775 [PMID: 29027005 DOI: 10.1007/s00520-017-3887-1]
- 13 **de Borja MT,** Chow E, Bovett G, Davis L, Gillies C. The correlation among patients and health care professionals in assessing functional status using the karnofsky and eastern cooperative oncology group performance status scales. *Support Cancer Ther* 2004; **2**: 59-63 [PMID: 18628160 DOI: 10.3816/SCT.2004.n.024]
- 14 **Taylor AE,** Olver IN, Sivanthan T, Chi M, Purnell C. Observer error in grading performance status in cancer patients. *Support Care Cancer* 1999; **7**: 332-335 [PMID: 10483818 DOI: 10.1007/s005200050271]
- 15 **Roila F,** Lupatelli M, Sassi M, Basurto C, Bracarda S, Picciafuoco M, Boschetti E, Milella G, Ballatori E, Tonato M. Intra and interobserver variability in cancer patients' performance status assessed according to Karnofsky and ECOG scales. *Ann Oncol* 1991; **2**: 437-439 [PMID: 1768630 DOI: 10.1093/oxfordjournals.annonc.a057981]
- 16 **Conill C,** Verger E, Salameró M. Performance status assessment in cancer patients. *Cancer* 1990; **65**: 1864-1866 [PMID: 2317765 DOI: 10.1002/1097-0142(19900415)65:8<1864::AID-CNCR2820650832>3.0.CO;2-U]
- 17 **Péus D,** Newcomb N, Hofer S. Appraisal of the Karnofsky Performance Status and proposal of a simple algorithmic system for its evaluation. *BMC Med Inform Decis Mak* 2013; **13**: 72 [PMID: 23870327 DOI: 10.1186/1472-6947-13-72]
- 18 **de Kock I,** Mirhosseini M, Lau F, Thai V, Downing M, Quan H, Lesperance M, Yang J. Conversion of Karnofsky Performance Status (KPS) and Eastern Cooperative Oncology Group Performance Status (ECOG) to Palliative Performance Scale (PPS), and the interchangeability of PPS and KPS in prognostic tools. *J Palliat Care* 2013; **29**: 163-169 [PMID: 24380215]
- 19 **Ma C,** Bandukwala S, Burman D, Bryson J, Seccareccia D, Banerjee S, Myers J, Rodin G, Dudgeon D, Zimmermann C. Interconversion of three measures of performance status: an empirical analysis. *Eur J Cancer* 2010; **46**: 3175-3183 [PMID: 20674334 DOI: 10.1016/j.ejca.2010.06.126]
- 20 **Minna JD,** Higgins GA, Glatstein EJ. Cancer of the lung. In: De Vita Jr. VT, Hellman S, Rosenberg SA, editors. *Cancer: Principles and Practice of Oncology*. Philadelphia: J. B. Lippincott Co., 1985
- 21 **Singh N,** Aggarwal AN, Behera D. Management of advanced lung cancer in resource-constrained settings: a perspective from India. *Expert Rev Anticancer Ther* 2012; **12**: 1479-1495 [PMID: 23249112 DOI: 10.1586/era.12.119]
- 22 **Singh N,** Aggarwal AN, Kaur J, Behera D. Association of Graded Folic Acid Supplementation and Total Plasma Homocysteine Levels With Hematological Toxicity During First-line Treatment of Nonsquamous NSCLC Patients With Pemetrexed-based Chemotherapy. *Am J Clin Oncol* 2017; **40**: 75-82 [PMID: 25089530 DOI: 10.1097/COC.000000000000111]
- 23 **Singh N,** Aggarwal AN, Behera D, Jindal SK. Intercycle delays during chemotherapy of non-small cell lung cancer in a health care resource-constrained setting and their effect on overall survival. *J Thorac Oncol* 2010; **5**: 236-239 [PMID: 20101147 DOI: 10.1097/JTO.0b013e3181c3f5f7]
- 24 **Baldi M,** Behera D, Kaur J, Kapoor R, Singh N. Rationale and Design of PEMVITASTART-An Open-label Randomized Trial Comparing Simultaneous Versus Standard Initiation of Vitamin B12 and Folate Supplementation in Nonsquamous, Non-Small-cell Lung Cancer Patients Undergoing First-line Pemetrexed-based Chemotherapy. *Clin Lung Cancer* 2017; **18**: 432-435 [PMID: 28073680 DOI: 10.1016/j.clc.2016.11.017]

- 25 **Singh N**, Aggarwal AN, Gupta D, Behera D, Jindal SK. Unchanging clinico-epidemiological profile of lung cancer in north India over three decades. *Cancer Epidemiol* 2010; **34**: 101-104 [PMID: 20079703 DOI: 10.1016/j.canep.2009.12.015]
- 26 **Singh N**, Aggarwal AN, Gupta D, Behera D, Jindal SK. Quantified smoking status and non-small cell lung cancer stage at presentation: analysis of a North Indian cohort and a systematic review of literature. *J Thorac Dis* 2012; **4**: 474-484 [PMID: 23050111 DOI: 10.3978/j.issn.2072-1439.2012.05.11]
- 27 **Singh N**, Aggarwal AN, Gupta D, Behera D. Prevalence of low body mass index among newly diagnosed lung cancer patients in North India and its association with smoking status. *Thorac Cancer* 2011; **2**: 27-31 [PMID: 27755836 DOI: 10.1111/j.1759-7714.2010.00037.x]
- 28 **Kaur H**, Sehgal IS, Bal A, Gupta N, Behera D, Das A, Singh N. Evolving epidemiology of lung cancer in India: Reducing non-small cell lung cancer-not otherwise specified and quantifying tobacco smoke exposure are the key. *Indian J Cancer* 2017; **54**: 285-290 [PMID: 29199707 DOI: 10.4103/ijc.IJC_597_16]
- 29 **Singh N**, Singh PS, Aggarwal AN, Behera D. Comorbidity Assessment Using Charlson Comorbidity Index and Simplified Comorbidity Score and Its Association With Clinical Outcomes During First-Line Chemotherapy for Lung Cancer. *Clin Lung Cancer* 2016; **17**: 205-213.e1 [PMID: 26589440 DOI: 10.1016/j.clcc.2015.10.002]
- 30 **Goldstraw P**, Crowley J, Chansky K, Giroux DJ, Groome PA, Rami-Porta R, Postmus PE, Rusch V, Sobin L; International Association for the Study of Lung Cancer International Staging Committee; Participating Institutions. The IASLC Lung Cancer Staging Project: proposals for the revision of the TNM stage groupings in the forthcoming (seventh) edition of the TNM Classification of malignant tumours. *J Thorac Oncol* 2007; **2**: 706-714 [PMID: 17762336 DOI: 10.1097/JTO.0b013e31812f3c1a]
- 31 **Sedgwick P**. Correlation. *BMJ-BRIT MED J* 2012; 345
- 32 **Landis JR**, Koch GG. The measurement of observer agreement for categorical data. *Biometrics* 1977; **33**: 159-174 [PMID: 843571 DOI: 10.2307/2529310]
- 33 **American Joint Committee on Cancer**. AJCC Cancer Staging Manual. 8th ed. New York: Springer International Publishing, 2017
- 34 **American Joint Committee for Cancer Staging and End-Results Reporting**. Manual for staging of cancer. 1st ed. Chicago, 1977

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Retrospective Cohort Study

Comparison of the eighth version of the American Joint Committee on Cancer manual to the seventh version for colorectal cancer: A retrospective review of our data

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Abstract**AIM**

To analyze the survival trends in colorectal cancer (CRC) based on the different classifications recommended by the seventh and eighth editions of the American Joint Committee on Cancer staging system (AJCC-7th and AJCC-8th).

METHODS

The database from our institution was queried to identify patients with pathologically confirmed stage 0-IV CRC diagnosed between 2006 and 2012. Data from 2080 cases were collected and 1090 cases were evaluated through standardized inclusion and exclusion criteria. CRC was staged by AJCC-7th and then restaged by AJCC-8th. Five-year disease-free survival (DFS) and overall survival (OS) were compared. SPSS 21.0 software was used for all data. DFS and OS were compared and analyzed by Kaplan-Meier and Log-rank test.

RESULTS

Linear regression and automatic linear regression showed lymph node positive functional equations by tumor-node-metastasis staging from AJCC-7th and tumor-node-metastasis staging from AJCC-8th. Neurological

invasion, venous infiltration, lymphatic infiltration, and tumor deposition put forward stricter requirements for pathological examination in AJCC-8th compared to AJCC-7th. After re-analyzing our cohort with AJCC-8th, the percentage of stage IVB cases decreased from 2.8% to 0.8%. As a result 2% of the cases were classified under the new IVC staging. DFS and OS was significantly shorter ($P = 0.012$) in stage IVC patients compared to stage IVB patients.

CONCLUSION

The addition of stage IVC in AJCC-8th has shown that peritoneal metastasis has a worse prognosis than distant organ metastasis in our institution's CRC cohort. Additional datasets should be analyzed to confirm these findings.

Key words: Colorectal cancer; Tumor-node-metastasis staging; Prognosis; Peritoneal metastasis; Disease-free survival

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Core tip: Since the promulgation of the eighth edition of the American Joint Committee on Cancer staging system manual (AJCC-8th), it has attracted the attention of many clinicians around the world and guided clinical work. Using our institution data we explored the prognostic differences between AJCC-8th and the seventh edition of the AJCC manual (AJCC-7th) for colorectal cancer. We found that patients with stage IV C colorectal cancer have a worse prognosis. This shows that peritoneal metastasis has a worse prognosis than organ metastasis. Considering many prognostic factors, individualized treatment is particularly important to improve the survival time of stage IV patients, especially stage IVC patients.

Tong GJ, Zhang GY, Liu J, Zheng ZZ, Chen Y, Niu PP, Xu XT. Comparison of the eighth version of the American Joint Committee on Cancer manual to the seventh version for colorectal cancer: A retrospective review of our data. *World J Clin Oncol* 2018; 9(7): 148-161 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i7/148.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i7.148>

INTRODUCTION

Colorectal cancer (CRC) is a common malignant tumor^[1-2]. In 2016, the incidence and mortality in the United States were respectively ranked fourth and second^[3-7]. In 2015, 376000 patients were newly diagnosed with CRC in China and 191000 patients died from the disease^[8]. Surgical resection remains the mainstay of treatment for local and regional disease^[9-14]. Adjuvant chemotherapy is frequently used in advanced colon cancer and CRC, but remains controversial for

stage II disease^[15-21]. Understanding the pathologic staging in conjunction with prognostic values is essential to making therapeutic decisions. The American Joint Committee on Cancer (AJCC) tumor-node-metastasis (TNM) staging model has provided this universal modality since its first edition in 1977^[22]. Since then, the AJCC has repeatedly revised this guideline (Figure 1) to continuously guide clinical treatment.

The eighth edition of the AJCC staging system (AJCC-8th) was released on October 6, 2016 in Chicago, IL, United States, and was implemented globally on January 1, 2018, which included significant changes for CRC patients with stage IV disease^[23]. The Cancer Council under the American College of Surgeons required the use of the AJCC-8th staging system as the "primary language" for cancer reporting. In 2013, AJCC established the "Evidence-Based Medicine and Statistics Core Group" of the 8th edition of the staging system. The organization is composed of clinical physicians, statisticians, and methodologists. It is responsible for determining the level of evidence for any updated content of the AJCC staging system.

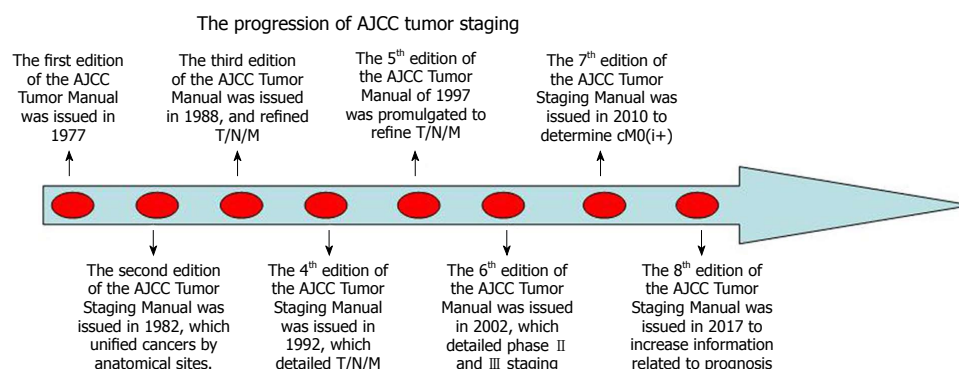
The level of evidence is divided into four levels, and the quality of evidence represented by it gradually decreases from level I to level IV. Level I requires that the evidence is from multiple large national or international studies, has consistent results, has good research requirement design and implementation, was conducted in appropriate patient populations with appropriate study endpoints and appropriate treatment options, either as prospective studies or review-based studies based on patient populations, but all studies must be methodologically assessed. Level II requires that the evidence comes from at least one large study and had good design and implementation, was conducted in a suitable patient population with a suitable study endpoint, and has external reliability (generally the representative and extrapolated capabilities of the study are better). Level III includes evidence from a study with certain flaws, defects in the number of possible subjects, size, or quality of the study, or the consistency of multiple findings, the appropriateness of the patient population, and the appropriateness of the results. Level IV includes evidence wherein no reasonable research had been done. Only evidence from levels I - III could be included in the 8th version of the staging system.

A major difference between AJCC-7th, and AJCC-8th is that the CRC staging system was revised to include a new stage involving peritoneal metastasis (named stage IVC) (see Tables 1 and 2 for details). Based on a variety of evidence-based medical evidence^[24,25], the AJCC-8th CRC staging system continues to recommend vascular lymphatic vessel infiltration and tumor deposition as prognostic level information, while microsatellite instability status and *BRAF* gene status are used as prognostic factors, and *BRAF*, *KRAS*, and degeneration of the *NRAS* gene were used as a predictor of efficacy (Table 3)^[26].

The increased complexity of the AJCC-8th staging

Table 1 Comparison of the tumor-node-metastasis stages between the 7th edition and the 8th edition

7 th edition	8 th edition
Tx: Primary tumor cannot be assessed	Tx: Primary tumor cannot be assessed
T0: No evidence of primary tumor	T0: No evidence of primary tumor
Tis: Carcinoma <i>in situ</i> , limited to intraepithelial or invasive lamina propria	Tis: Carcinoma <i>in situ</i> , limited to intraepithelial or invasive lamina propria
T1: Tumor invading submucosa	T1: Tumor invading submucosa
T2: Tumor invading the muscularis propria	T2: Tumor invading the muscularis propria
T3: Tumor penetrating the muscularis propria and arriving at colorectal fat tissue	T3: Tumor penetrating the muscularis propria and arriving at colorectal fat tissue
T4: Tumor directly invading other organs or structures	T4: Tumor directly invading other organs or structures
T4a: Tumor penetrating visceral peritoneum	T4a: Tumor penetrating visceral peritoneum
T4b: Tumor directly invading or adhering to other organs or structures	T4b: Tumor directly invading or adhering to other organs or structures
Nx: Regional lymph nodes cannot be assessed	Nx: Regional lymph nodes cannot be assessed
N0: No lymph node metastasis and no tumor deposits (TD)	N0: No lymph node metastasis and no TD
N1: 1-3 lymph nodes metastases	N1: 1-3 lymph nodes metastases
N1a: 1 lymph node metastases	N1a: 1 lymph node metastases
N1b: 2-3 lymph nodes metastases	N1b: 2-3 lymph nodes metastases
N1c: Although there was no regional lymph node metastasis, TDs were submucosal, mesangial, or peritoneum-covered para-colorectal tissue.	N1c: Although there was no regional lymph node metastasis, TDs were submucosal, mesangial or peritoneum-covered para-colorectal tissue.
N2: More than or equal to 4 lymph node metastases	N2: More than or equal to 4 lymph node metastases
N2a: 4-6 regional lymph node metastases	N2a: 4-6 regional lymph node metastases
N2b: More than or equal to 7 lymph node metastases	N2b: More than or equal to 7 lymph node metastases
M1: There is distant lymph node metastasis	M1: There is distant lymph node metastasis
M1a: Metastasis is limited to one organ or site (<i>e.g.</i> , liver, lung, ovary, and extra-regional lymph node metastases)	M1a: Metastasis is limited to one organ or site (<i>e.g.</i> , liver, lung, ovary, and extra-regional lymph node metastases)
M1b: Transfer more than one organ or site, or to the peritoneum ¹	M1b: Transfer more than one organ or site ¹
	M1c: Peritoneal metastases with or without metastasis of other organs ¹

¹Differences between the two versions.**Figure 1** The progression of American Joint Committee on Cancer tumor staging. AJCC: American Joint Committee on Cancer.

model was intended to improve the prognostic staging of CRC, but the impact of these changes remains unclear. In this study, we used data from our institutional registries to compare the prognostic accuracy of criteria from AJCC-7th and AJCC-8th in patients with stage 0-IV through survival models. We also explored the relationship between positive node and tumor size, differentiation, tumor invasion, chemotherapy, tumor-node-metastasis (TNM) staging from AJCC-7th, and TNM staging from AJCC-8th. In addition, we also discussed the pathological importance of lymph invasion, vein invasion, and nerve invasion according to AJCC-8th.

MATERIALS AND METHODS

Patients

A total of 2080 patients with pathologically confirmed

stage 0-IV CRC between 2006 and 2012 were collected from our institutional database. Then the following inclusion and exclusion criteria were applied to this cohort: (1) on the basis of a colonoscopy, computed tomography, pathological diagnosis of CRC, in or outside the hospital diagnosis in our hospital; (2) patients undergoing colorectal surgery in our hospital (including radical surgery and non-radical surgery); (3) diagnosis as a recurrence of the primary tumor or as a result of the death of the primary tumor; (4) cases with complete and detailed clinical and pathological data; and (5) cases with complete follow-up data and accurate data. Exclusion criteria were: (1) a serious heart, brain, liver, or lung disease led to intolerant surgery; (2) the non-CRC factors that led to the death of the pathological interstitial tumor, neuronal tumor, lymphoma, melanoma and other non-adenocarcinoma in addition to other malignant tumors;

Table 2 Colorectal cancer tumor-node-metastasis staging American Joint Committee on Cancer 7th and 8th editions

7 th edition				8 th edition			
Stage	T	N	M	Stage	T	N	M
0	Tis	N0	M0	0	Tis	N0	M0
I	T1-2	N0	M0	I	T1-2	N0	M0
II A	T3	N0	M0	II A	T3	N0	M0
II B	T4a	N0	M0	II B	T4a	N0	M0
II C	T4b	N0	M0	II C	T4b	N0	M0
III A	T1-2	N1/N1c	M0	III A	T1-2	N1/N1c	M0
	T1	N2a	M0		T1	N2a	M0
III B	T3-4a	N1/N1c	M0	III B	T3-4a	N1/N1c	M0
	T2-3	N2a	M0		T2-3	N2a	M0
	T1-2	N2b	M0		T1-2	N2b	M0
III C	T4a	N2a	M0	III C	T4a	N2a	M0
	T3-4a	N2b	M0		T3-4a	N2b	M0
	T4b	N1-2	M0		T4b	N1-2	M0
IVA	Any T	Any N	M1a	IVA	Any T	Any N	M1a
IVB	Any T	Any N	M1b	IVB	Any T	Any N	M1b
				IVC	Any T	Any N	M1c

Table 3 American Joint Committee on Cancer 8th edition updates for the colorectal cancer staging system

Update points	Update details	Level of evidence
Definition of distant transfer (M)	Introduction of M1c, specifically peritoneal metastasis, is an indicator of poor prognosis	I
Definition of regional lymph nodes (N)	Further introduce the definition of tumor deposit	II
Recommended additional indicators for guiding clinical practice	Lymphatic vessel infiltration: Reintroducing the meaning of L and V ¹ positive to correctly understand lymphatic and vascular invasion	I
Recommended additional indicators for guiding clinical practice	Microsatellite instability: Further explaining its importance as a prognostic risk and efficacy predictor	I
Recommended additional indicators for guiding clinical practice	Determine the <i>KRAS</i> , <i>NRAS</i> , and <i>BRAF</i> mutations as very important prognostic risk and efficacy predictors	I, II

¹L-positive infiltrates for medics and V-positive for venous infiltration.

and (3) cases with incomplete clinical-pathologic data and cases with incomplete follow-up data. As a result, 990 cases were excluded. Therefore our analysis focused on the remaining 1090 cases.

Follow-up

Patients were routinely followed in the outpatient clinic 2 wk after surgery for 3 mo and every 3 mo for the first year, then every 6 mo for the second year and every year for the next 3 year. Follow-up data was complemented by phone contact as well as contact with written mail.

Ethics statement

This study was carried out in accordance with the ethical guidelines of the 1975 Declaration of Helsinki and was approved by the ethics committee of Huzhou Central Hospital.

Preliminary processing of data

Using the extent of disease codes, tumor invasion (T staging), lymph node positivity (N staging), tumor metastasis (M staging) status, CRC was staged based on the AJCC-7th and AJCC-8th (Table 4). The patients were divided into three groups (N0, N1, N2) by the number

of positive lymph nodes. Clinicopathological data were analyzed between the three groups. Patient status was designated into three outcome categories for disease-free survival (DFS): (1) death from CRC; (2) recurrence from CRC; or (3) alive at the last follow-up. Patient status was designated into two outcome categories for overall survival (OS): (1) death from CRC; or (2) alive at the last follow-up.

Statistical analysis

SPSS 21 (Chicago, IL, United States) was used for data analysis. Intergroup measurement data were analyzed using ANOVA analysis of variance and count data were analyzed using Cross-Tab χ^2 analysis.

The relationship between positive lymph node and tumor size, differentiation, tumor invasion, chemotherapy, and TNM staging from AJCC-7th, and TNM staging from AJCC-8th were analyzed by linear and automatic linear regression and the functional equations were established.

Survival curves were generated using Kaplan-Meier estimates, and 5-year DFS and OS were compared using the Log-rank test. Kaplan-Meier was also used to calculate the survival rate of DFS and OS in each group. Afterwards, Cross-table was used to compare the DFS

Table 4 Two-way classification table of staging based on tumor-node-metastasis staging from AJCC-7th vs tumor-node-metastasis staging from AJCC-8th for patients with stages 0-IV colorectal cancer from 2006-2012 (*n* = 1090)

		TNM staging from AJCC-7 th										Total
		0	I	II A	II B	II C	III A	III B	III C	IV A	IV B	
TNM staging from AJCC-8 th	0	16	0	0	0	0	0	0	0	0	0	16
	I	0	131	0	0	0	0	0	0	0	0	131
	II A	0	0	138	0	0	0	0	0	0	0	138
	II B	0	0	0	56	0	0	0	0	0	0	56
	II C	0	0	0	0	31	0	0	0	0	0	31
	III A	0	0	0	0	0	136	0	0	0	0	136
	III B	0	0	0	0	0	0	400	0	0	0	400
	III C	0	0	0	0	0	0	0	127	0	0	127
	IV A	0	0	0	0	0	0	0	0	24	0	24
	IV B	0	0	0	0	0	0	0	0	0	9	9
	IV C	0	0	0	0	0	0	0	0	0	22	22
		16	131	138	56	31	136	400	127	24	31	1090

TNM: Tumor-node-metastasis.

and OS survival rates of sub-periods between AJCC-7th and AJCC-8th groups, and a histogram was generated. *P*-values less than 0.05 were considered statistically significant.

RESULTS

Lymph staging (N) and clinicopathologic characteristics

During the 6-year study period, 2080 patients with stage 0-IV CRC were identified but only 1090 met our inclusion criteria. The median age at diagnosis was 66 years [interquartile range (IQR): 55-73] and median follow-up was 60 mo (IQR: 54-60). The N staging did not change between AJCC-7th and AJCC-8th, therefore we used N staging to analyze clinical pathology data. Patient demographics and pathological features were summarized in Table 5. This table also compared staging of CRC with AJCC-7th vs AJCC-8th criteria. Although there was no difference in the total number of patients with stage IV CRC, the distribution of patients in this period was different. The χ^2 test was performed for all sub-stages of CRC, and significance existed between IVA and IVB according to AJCC-7th (*P* = 0.001), and between IVA, IVB, and IVC according to AJCC-8th (*P* = 0.05).

Linear model between the number of positive lymph nodes and tumor size, differentiation, tumor invasion, chemotherapy, TNM staging from AJCC-7th, and TNM staging from AJCC-8th

The number of positive lymph nodes was related to the N anatomical stages in AJCC-7th and AJCC-8th. An automated linear model found that the number of positive lymph nodes and tumor size, tumor differentiation, depth of tumor invasion, chemotherapy, TNM staging from AJCC-7th, and TNM staging from AJCC-8th were indicators of good fit and showed significance (*P* < 0.05). The fitting degree for TNM staging from AJCC-7th was 61.3% (Figure 2A), and the index that had a significant influence on positive lymph nodes was shown in Figure 2B. However, chemotherapy was

not included in the predictive importance index (Figure 2C). The importance of TNM staging from AJCC-7th was 77%, and the importance of tumor invasion was 19%, the importance of tumor size was 3%, the degree of tumor differentiation was 1%. Figure 2D showed significant parameters of each coding amount and constant coefficient. The fitness for TNM staging from AJCC-8th was 63.3% (Figure 3A), and the indexes that had a significant influence on positive lymph nodes were shown in Figure 3B. Chemotherapy was also included in the predictive importance index (Figure 3C). The importance of TNM staging from AJCC-8th was 72%, the importance of tumor invasion was 20%, the importance of chemotherapy was 4%, the importance of tumor size was 3%, the degree of tumor differentiation was 1%. Figure 3D showed significant parameters of each coding amount and constant coefficient.

Then the linear model calculated the functional equation for these variables and positive lymph node relationships. Outcome showed that $Y_A = -0.918 + 0.409X_B + 0.18X_C - 0.583X_D - 0.460X_E + 0.669X_F$ and $Y_A = -0.821 + 0.404X_B + 0.183X_C - 0.587X_D - 0.491X_E + 0.658X_G$ (A: Positive lymph node; B: Tumor size; C: Differentiation; D: Tumor invasion; E: Chemotherapy; F: TNM staging from AJCC-7th; G: TNM staging from AJCC-8th).

DFS and OS between AJCC-7th and AJCC-8th criteria

Using Kaplan-Meier univariate analysis and Log-rank test, the 5-year survival rate of DFS and OS in 1090 patients was calculated and compared by stage and sub-stage according to AJCC-7th and AJCC-8th criteria. DFS and OS survival rate between the two editions did not change from stage 0-IV and from substage 0-IV B. However, when the 5-year DFS and OS survival rate were compared from stage IVB from AJCC-7th and from stage IVB and IVC from AJCC-8th the survival curve of DFS and OS showed a significant right shift for stage IV B and a significant left shift for stage IVC (*P* = 0.001 and *P* < 0.001, respectively). Details were shown in Table 6

Table 5 Demographic and clinical characteristics of patients with stage 0-IV colorectal cancer from 2006-2012 (*n*, mean \pm SD)

	N0	N1	N2	F or χ^2	P
Gender				2.895	0.235
Male	242	182	126		
Female	234	201	105		
Age (yr)	62.46 \pm 14.43	62.17 \pm 14.43	61.98 \pm 14.70	0.095	0.909
ASA				6.011	0.198
1	362	277	158		
2	102	94	68		
3	12	12	5		
Primary site				4.94	0.895
Ileocecum	36	26	11		
Right colon	43	30	22		
Transverse colon	70	64	40		
Left colon	88	72	46		
Sigmoid colon	53	34	21		
Rectum	186	157	91		
Tumor size (cm)	3.31 \pm 1.17	3.76 \pm 0.82	4.11 \pm 0.74	56.008	< 0.001
Operation method				8.233	0.411
RHC	97	67	43		
LHC	186	154	91		
HO	9	6	9		
AR	145	112	70		
APR	39	44	18		
Operation time (m)	151.59 \pm 36.31	156.40 \pm 34.94	153.17 \pm 31.30	2.044	0.130
Resection length (cm)	27.96 \pm 9.92	27.26 \pm 9.83	27.65 \pm 9.92	0.533	0.587
Blood loss (mL)	184.39 \pm 94.25	185.23 \pm 95.26	194.30 \pm 107.32	0.879	0.416
Tumor invasion				131.640	< 0.001
Tis	16	0	0		
T1	85	17	9		
T2	92	75	43		
T3	162	127	132		
T4a	82	108	22		
T4b	39	56	25		
Differentiation				188.64	< 0.001
Well	150	31	13		
Moderate	276	296	124		
Poor or undifferentiated	50	56	94		
Number of LNs examined	14.70 \pm 1.88	14.13 \pm 1.78	14.26 \pm 1.85	0.408	0.665
Number of positive LNs	0	1.85 \pm 0.73	5.46 \pm 1.64	3050.47	< 0.001
Complication				4.088	0.130
No	436	349	201		
Yes	40	34	30		
Chemotherapy				295.36	< 0.001
Yes	283	383	229		
No	193	0	2		
TNM staging AJCC-7 th				887.08	< 0.001
0	16	0	0		
I	131	0	0		
II A	138	0	0		
II B	56	0	0		
II C	31	0	0		
III A	45	82	9		
III B	49	234	117		
III C	9	47	71		
IV A	1	15	8		
IV B	0	5	26		
TNM staging AJCC-8 th				887.32	< 0.001
0	16	0	0		
I	131	0	0		
II A	138	0	0		
II B	56	0	0		
II C	31	0	0		
III A	45	82	9		
III B	49	234	117		
III C	9	47	71		
IV A	1	15	8		
IV B	0	1	8		
IV C	0	4	18		

TNM: Tumor-node-metastasis.

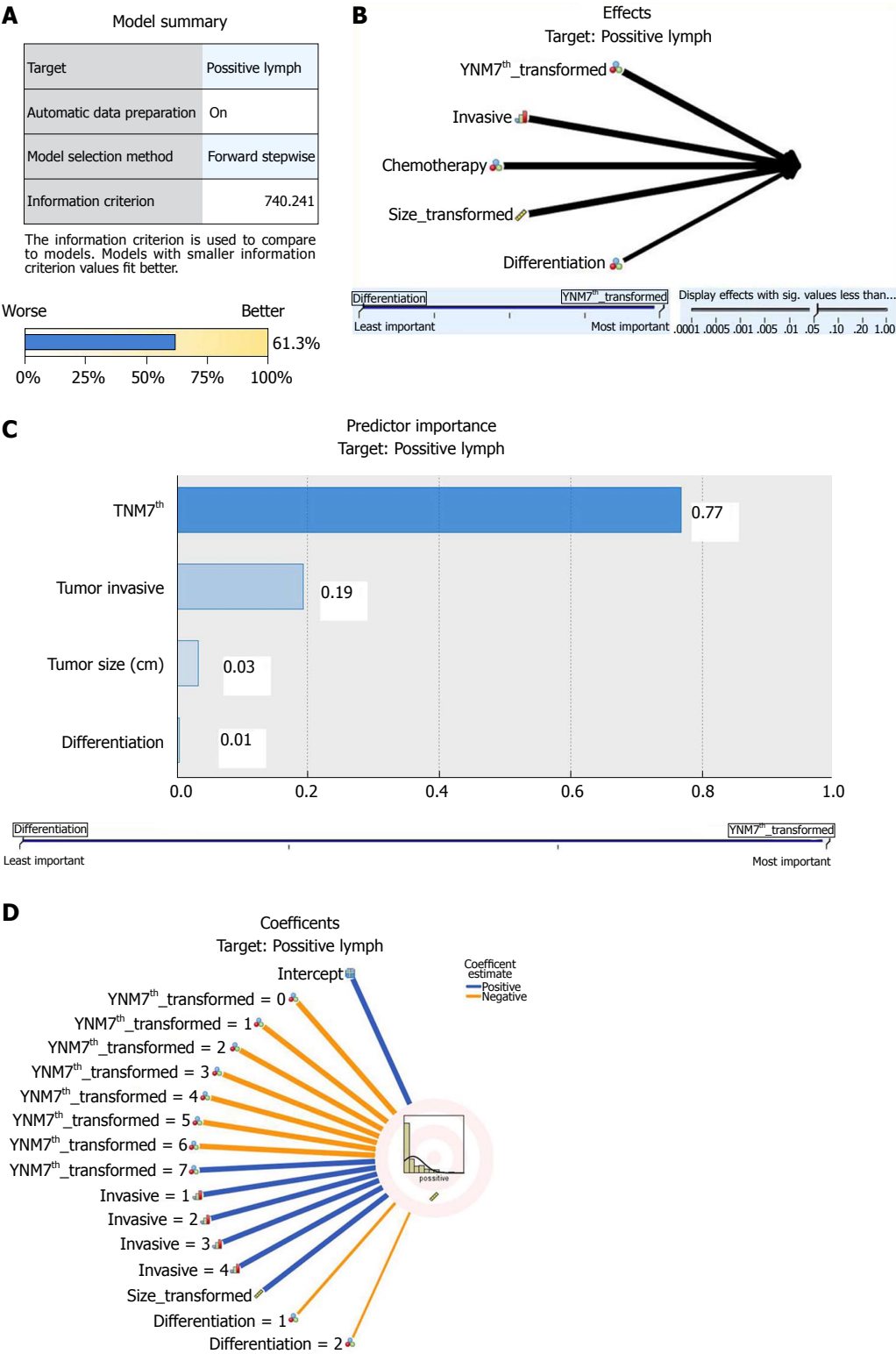


Figure 2 Automatic linear regression about positive lymph nodes and clinicopathologic parameters with tumor-node-metastasis staging from AJCC-7th. A: Clinical pathological parameters fitting degree. Fitting value is 61.3%; B: Significant effect parameters ($P < 0.05$); C: Predictor importance of positive lymph nodes and clinicopathological parameters. The values of tumor-node-metastasis staging from AJCC-7th, tumor invasion, tumor size, and differentiation are 0.77, 0.19, 0.03, and 0.01, respectively; D: Coefficients about positive nodes and clinicopathological parameters.

and Figure 4.

Nerve invasion, vein invasion, Lymphatic invasion and tumor deposit between AJCC-7th and AJCC-8th
AJCC-8th further emphasized the clinical value of tumor

lymphatic invasion, vein invasion, nerve invasion, and tumor deposit (TD) and were included in “evidence-based medicine” evidence level (Table 3). Since the release of AJCC-7th, our institution’s pathologist has attached great importance to this aspect of the test and has described

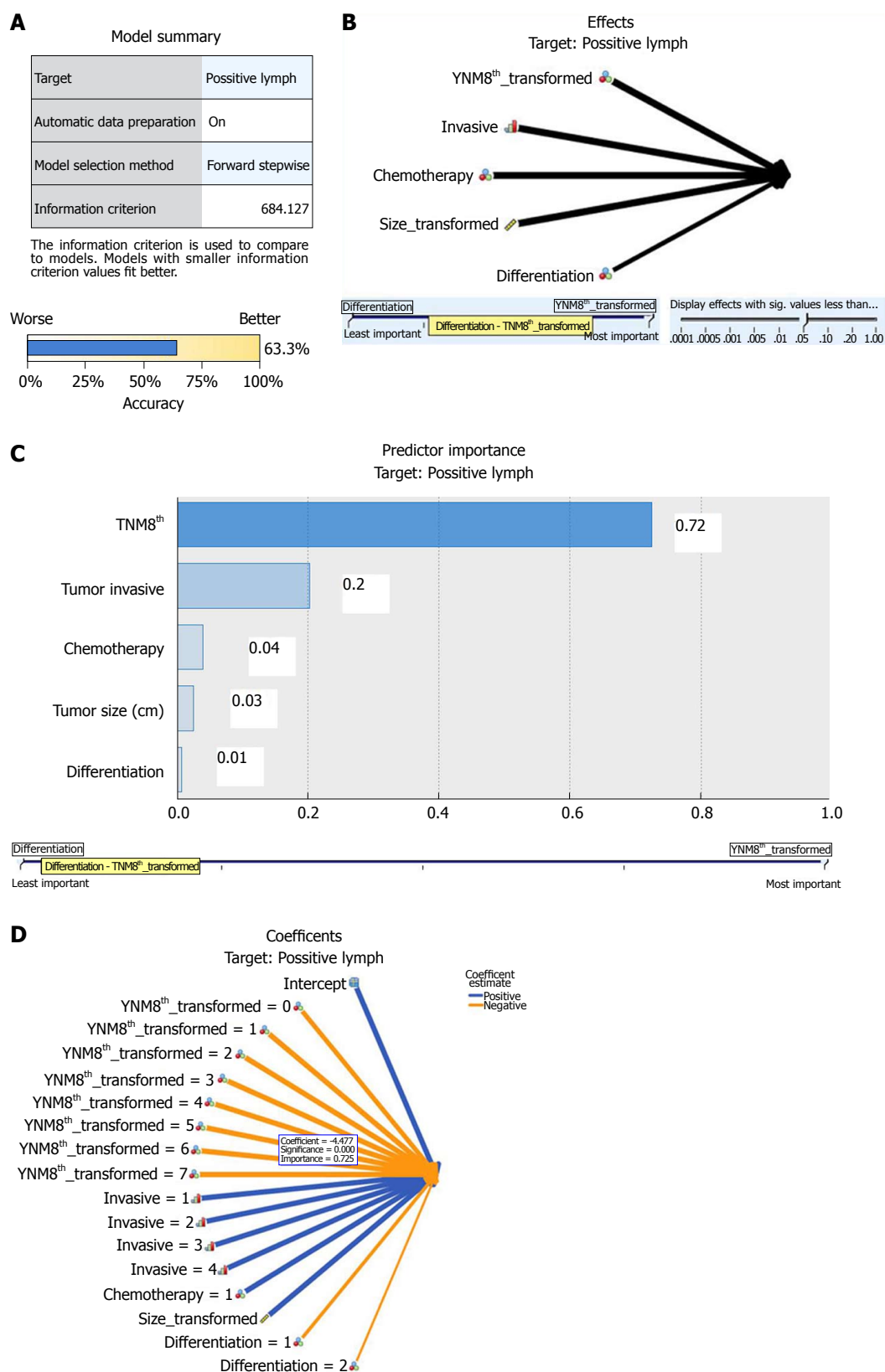


Figure 3 Automatic linear regression about positive lymph nodes and clinicopathologic parameters with tumor-node-metastasis staging from AJCC-8th. A: Clinical pathological parameters fitting degree. The fitting value is 63.3%; B: Significant effect parameters ($P < 0.05$); C: Predictor importance of positive lymph nodes and clinicopathological parameters. The values of tumor-node-metastasis staging from AJCC-8th, tumor invasion, chemotherapy, tumor size, and differentiation are 0.72, 0.2, 0.04, 0.03 and 0.01, respectively; D: Coefficients about positive nodes and clinicopathological parameters.

Table 6 Comparison of 5-year disease-free survival and overall survival rate for stage and sub-stage using American Joint Committee on Cancer-7th edition and American Joint Committee on Cancer-8th edition (%)

			0	I	II A	II B	II C	III A	III B	III C	IV A	IV B	IV C	Log-rank χ^2	P
OS	Sub-stage	AJCC-7	100	98.5	82.6	76.8	67.7	65.4	60	44.9	8.3	0	-	1423.53	< 0.01
		AJCC-8	100	98.5	82.6	76.8	67.7	65.4	60	44.9	8.3	0	0	1608.11	< 0.01
	Stage	AJCC-7	100	98.5		79.1			58.2			3.6		913.56	< 0.01
		AJCC-8	100	98.5		79.1			58.2			3.6		875.46	< 0.01
DFS	Sub-stage	AJCC-7	100	93.1	78.3	73.2	61.3	65.4	56.3	37	8.3	0	-	1418.9	< 0.01
		AJCC-8		93.1	78.3	73.2	61.3	65.4	56.3	37	8.3	0	0	1603.4	< 0.01
	Stage	AJCC-7	100	93.1		74.7			54.4			3.6		875.46	< 0.01
		AJCC-8		93.1		74.7			54.4			3.6		875.46	< 0.01

DFS: Disease-free survival; OS: Overall survival; AJCC: American Joint Committee on Cancer.

them in detail (Figure 5).

DISCUSSION

In 1977, AJCC established the first edition of the cancer staging system. Revision to the system have been made every 6-8 years and until recently it has been regarded as the most comprehensive tool for prognostic and predictive grouping of patients with colon cancer^[24]. However, when AJCC-6th was released in 2002^[27], it elicited criticism because survival of patients with stage IIIA colon cancer was superior to that of patients with stage IIB colon cancer^[28]. In 2010, the AJCC cancer staging system was updated to the 7th edition^[22,29]. This edition included both the refinement of the classic TNM "anatomic blood" diagnostic system, the increase in tumor regression scores, and the risk of prognoses and curative effects for circumferential resection margins.

Evaluation index

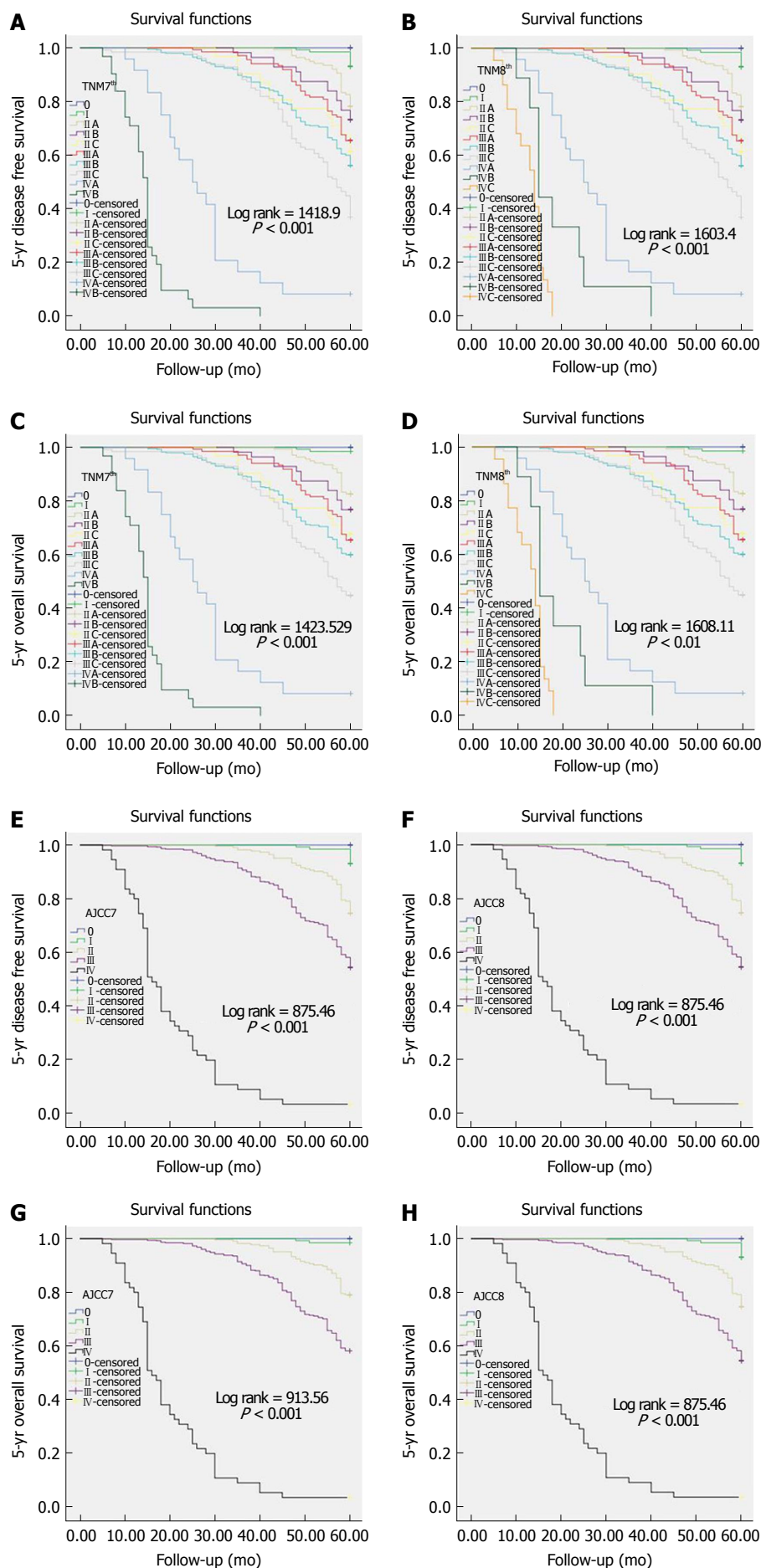
The problem with AJCC staging of CRC was initially attributed to inadequate lymph node (LN) assessment. Previous studies demonstrated that the number of examined LNs impacted survival^[30-34]. Subsequent studies showed a strong correlation between outcomes and compliance with 12-LN minimum^[35-39]. In our study, in addition to analyzing the distribution of LN numbers in different N stages, we also focused on the effect of positive LN numbers on lymphatic pathology, and established a linear function.

In recent years, researchers have recognized the importance of tumorigenesis and the role of non-anatomic markers in establishing the prognosis and anticipated response to therapy^[40-45]. Of these factors, the circumferential margin of the resected non-peritonealized surface of the specimen (CRM) is relevant for prognostic assessment of patients with tumors in the ascending and descending colon^[46,47]. Microsatellite instability, KRAS mutation and the 18q LOH have been shown to have clinical prognostic significance^[48,49]. These factors have

not been incorporated into the staging system because it is not clear how they should be used to determine prognosis or the need for adjuvant chemotherapy. In 2013, AJCC established the "Evidence-Based Medicine and Statistics Core Group" of the eighth edition system, which was responsible for determining the level of evidence for any updated content of the AJCC staging system. New evidence had to reach an evidence quality level of I - III to be factored into the staging system for the eighth edition.

AJCC-8th did not include any updates for tumor staging. The definition of TD and N1c in the N-stage was further interpreted as the presence of encouraging tumor nodules in the lymphatic drainage area of the primary tumor, and no lymph node, vessel, or nerve structure identified during the period. The presence of TD did not alter the T stage of the primary tumor, but if it was not accompanied by lymph node metastasis, the TDs would change N stage (from N0 to N1c). If there was combined lymph node metastasis, the number of TDs did not need to be counted in the number of positive lymph nodes. The latest version reaffirmed the definition of lymphatic infiltrating vessels. Any vessel lesions with or without residual vascular walls could be identified as lymphocytic infiltrates in storage vessels and become a routine item in the pathology report of the American College of Pathology. Our institutional pathologist recognized this and described them in the report (Figure 5). Vascular lymphatic infiltration could be subdivided into small vessel infiltration (lymphatic or venular infiltration, defined as "L" positive) and venous infiltration (a structure surrounded by tumor immersion and endothelial cells, which contain red blood cells coated with smooth muscle machinery was defined as "V" positive). At the same time, it was found that tumor immersion and nerve tissue were defined as infiltration around the nerve. Lymphatic infiltration and perineural invasion were both important prognostic factors^[50-56].

AJCC-7th classified the metastasis stage M1 as M1a (metastasis in one organ or site) and M1b (metastasis in



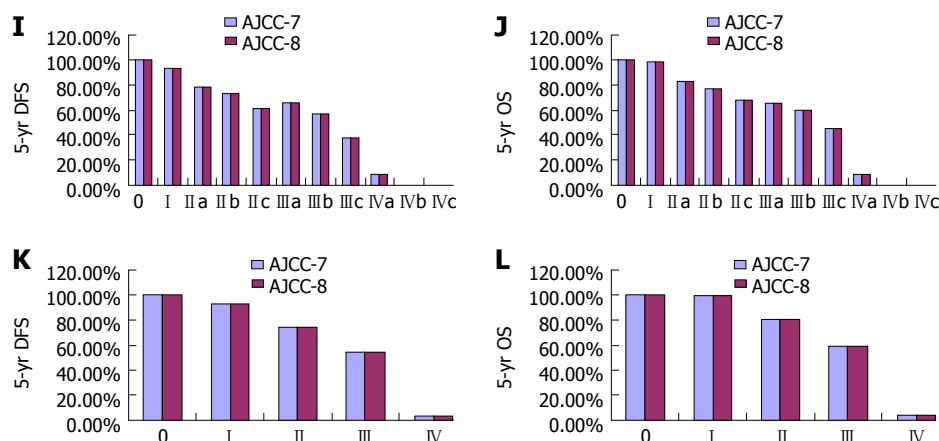


Figure 4 Disease-free survival and overall survival curves and histograms between tumor-node-metastasis staging from AJCC-7th and tumor-node-metastasis staging from AJCC-8th. A: Comparison of 5-year disease-free survival (DFS) by sub-stage from AJCC-7th ($P < 0.001$); B: Comparison of 5-year DFS by sub-stage from AJCC-8th ($P < 0.001$); C: Comparison of 5-year overall survival (OS) by sub-stage from AJCC-7th ($P < 0.001$); D: Comparison of 5-year OS by sub-stage from AJCC-8th ($P < 0.001$); E: Comparison of 5-year DFS by stage from AJCC-7th ($P < 0.001$); F: Comparison of 5-year DFS by stage from AJCC-8th ($P < 0.001$); G: Comparison of 5-year OS by stage from AJCC-7th ($P < 0.001$); H: Comparison of 5-year OS by stage from AJCC-8th ($P < 0.001$); A vs B and C vs D: Survival curves of DFS and OS in stage IVb shift right and those in stage IVc shift left; I-L: Comparison of DFS and OS by sub-stage and stage between staging from AJCC-7th and staging from AJCC-8th, all $P < 0.01$. DFS: Disease-free survival; OS: Overall survival; TNM: Tumor-node-metastasis.

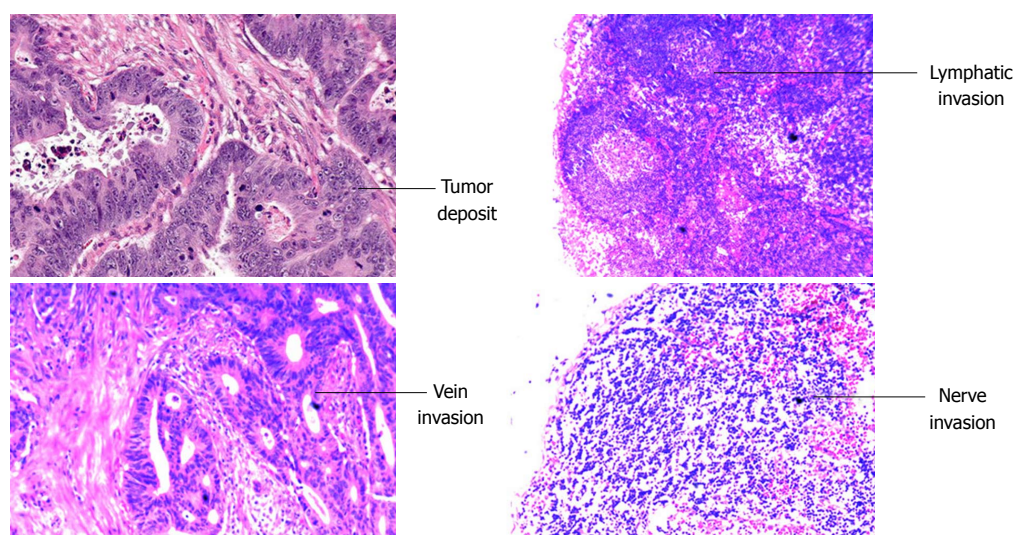


Figure 5 The focus of the eighth edition of the American Joint Committee on Cancer pathology test description. A: Tumor deposit; B: Lymphatic invasion; C: Vein invasion; D: Nerve invasion.

more than one organ or site, or in the peritoneum). In AJCC-8th, another stage was added to describe colorectal peritoneal metastases (whether or not with metastasis of other organ sites). This is called M1c, and M1a and M1b were redefined as metastasis limited to one organ or site (such as liver, lung, ovary, extra-nodal lymph nodes, etc.) and transition beyond one organ or site, but without peritoneal metastasis, respectively. The reason for the change is that although peritoneal metastasis occur in 1% to 4% of patients with CRC, the prognosis is far worse than that of M1a and M1b patients who have metastasis of substantial organs^[57-61].

We reclassified our cohort according to the AJCC-8th criteria. The results showed that the DFS and OS of the M1a stage remained unchanged, while that of the M1b stage improved, and that of the M1c stage decreased significantly. This demonstrated that the M stage

refinement was necessary. This additional classification in the eighth edition will have a positive and far-reaching effect on cancer treatment that will promote the individualized diagnosis and treatment of CRC patients. However, further analysis with additional institutional databases is needed to confirm our findings.

In conclusion, the addition of a sub-stage to classify peritoneal metastasis separately from distant organ metastasis in the AJCC-8th manual has shown that peritoneal metastasis has a worse prognosis than organ metastasis in our cohort.

ARTICLE HIGHLIGHTS

Research background

Colorectal cancer (CRC) is a common malignant tumors. Clinicians have been using the American Joint Committee on Cancer (AJCC) system to guide clinical

diagnosis and treatment for CRC. The eighth edition of the AJCC (AJCC-8th) has received extensive attention since its promulgation in 2016. Compared to the previous version, AJCC-8th refined the stage IV classification to separate peritoneal metastasis and organ metastasis.

Research motivation

In China, there are still many hospital surgeons and physicians who still use the old version to guide clinical practice and are uneducated about the new AJCC-8th classifications.

Research objectives

We analyzed our institution's CRC cohort to determine differences in the survival trends based on the diagnostic classifications between AJCC-8th and the previous version.

Research methods

A total 1090 patients of 2080 CRC patients were included in the study. The data were classified by AJCC-7th and AJCC-8th standards. Five-year disease-free survival (DFS) and overall survival (OS) were compared.

Research results

Linear regression and automatic linear regression showed lymph node positive functional equations by TNM staging from AJCC-7 and TNM staging from AJCC-8th. Neurological invasion, venous infiltration, lymphatic infiltration, and tumor deposition put forward stricter requirements for pathological examination. AJCC-8th staging yielded a proportional decrease of IVB from 2.8% to 0.8% and a new staging of IVC to 2%. Log-rank test showed that DFS and OS survival time of patients with IVC vs IVB was significantly shorter ($P = 0.012$).

Research conclusions

The addition of a sub-stage to classify peritoneal metastasis separately from distant organ metastasis in the AJCC-8th manual has shown that peritoneal metastasis has a worse prognosis than organ metastasis in our cohort. Considering many prognostic factors, individualized treatment is particularly important to improve the survival time of stage IV patients, especially IVC patients.

Research perspective

Further studies can be done to improve outcomes for peritoneal metastasis CRC patients. Further analysis of additional institutional databases is needed to confirm our findings.

REFERENCES

- 1 Ali I, Wani WA, Saleem K, Haque A. Thalidomide: A Banned Drug Resurged into Future Anticancer Drug. *Curr Drug Ther* 2012; **7**: 13-23 [DOI: 10.2174/157488512800389164]
- 2 Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin* 2016; **66**: 7-30 [PMID: 26742998 DOI: 10.3322/caac.21332]
- 3 Ali I, Lone MN, Al-Othman ZA, Al-Warthan A, Sanagi MM. Heterocyclic Scaffolds: Centrality in Anticancer Drug Development. *Curr Drug Targets* 2015; **16**: 711-734 [PMID: 25751009 DOI: 10.2174/1389450116666150309115922]
- 4 Ali I, Wani WA, Haque A, Saleem K. Glutamic acid and its derivatives: candidates for rational design of anticancer drugs. *Future Med Chem* 2013; **5**: 961-978 [PMID: 23682571 DOI: 10.4155/fmc.13.62]
- 5 Ali I, Haque A, Saleem K, Hsieh MF. Curcumin-I Knoevenagel's condensates and their Schiff's bases as anticancer agents: synthesis, pharmacological and simulation studies. *Bioorg Med Chem* 2013; **21**: 3808-3820 [PMID: 23643901 DOI: 10.1016/j.bmc.2013.04.018]
- 6 Ali I, Wani WA, Saleem K, Haque A. Platinum compounds: a hope for future cancer chemotherapy. *Anticancer Agents Med Chem* 2013; **13**: 296-306 [PMID: 22583420 DOI: 10.2174/1871520611313020016]
- 7 Basheer AA. Chemical chiral pollution: Impact on the society and science and need of the regulations in the 21st century. *Chirality* 2018; **30**: 402-406 [PMID: 29266491 DOI: 10.1002/chir.22808]
- 8 Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. *CA Cancer J Clin* 2016; **66**: 115-132 [PMID: 26808342 DOI: 10.3322/caac.21338]
- 9 Hari DM, Bilchik AJ. Clinical decision-making and implementation challenges with the AJCC VII staging system for colorectal carcinoma. *J Surg Oncol* 2012; **105**: 221-222 [PMID: 22271497 DOI: 10.1002/jso.22086]
- 10 Ali I, Saleem K, Aboul-Enein HY, Rather A, Imran D. Social aspects of cancer genesis. *Cancer Ther* 2011; **8**: 6-14
- 11 Ali I. Nano anti-cancer drugs: pros and cons and future perspectives. *Curr Cancer Drug Targets* 2011; **11**: 131-134 [PMID: 21062238 DOI: 10.2174/156800911794328457]
- 12 Ali I, Saleem K, Uddin R, Haque A, El-Azzouny A. Natural Products: Human Friendly Anti-Cancer Medications. *Egypt Pharm J* 2010; **9**: 133-179
- 13 Ali I, Wani WA, Saleem K, Wesselinova D. Syntheses, DNA binding and anticancer profiles of L-glutamic acid ligand and its copper(II) and ruthenium(III) complexes. *Med Chem* 2013; **9**: 11-21 [PMID: 22741786 DOI: 10.2174/1573406411309010011]
- 14 Ali I, Saleem K, Wesselinova D, Haque A. Synthesis, DNA binding, hemolytic, and anti-cancer assays of curcumin I-based ligands and their ruthenium(III) complexes. *Res Med Chem* 2013; **22**: 1386-1398 [DOI: 10.1007/s00044-012-0133-8]
- 15 Chun P, Wainberg ZA. Adjuvant Chemotherapy for Stage II Colon Cancer: The Role of Molecular Markers in Choosing Therapy. *Gastrointest Cancer Res* 2009; **3**: 191-196 [PMID: 20084160 DOI: 10.1016/S0140-6736(02)82429-1]
- 16 Ali I, Wani WA, Saleem K, Hsieh MF. Design and synthesis of thalidomide based dithiocarbamate Cu (II), Ni (II) and Ru (III) complexes as anticancer agents. *Polyhedron* 2013; **56**: 134-143 [DOI: 10.1016/j.poly.2013.03.056]
- 17 Ali I, Wani W, Saleem K, Hsieh MF. Anticancer metallodrugs of glutamic acid sulphonamides: in silico, DNA binding, hemolysis and anticancer studies. *Rsc Advances* 2014; **4**: 29629-29641 [DOI: 10.1039/C4RA02570A]
- 18 Saleem K, Wani WA, Haque A, Milhotra A, Ali I. Nanodrugs: magic bullets in cancer chemotherapy. *Anti Can Res* 2013; **58**: 437-494 [DOI: 10.2174/9781608051366113020016]
- 19 Ali I, Lone MN, Suhail M, Mukhtar SD, Asnin L. Advances in Nanocarriers for Anticancer Drugs Delivery. *Curr Med Chem* 2016; **23**: 2159-2187 [PMID: 27048343 DOI: 10.2174/0929867323666160405111152]
- 20 Ali I, Lone MN, Allothman ZA, Alwarthan A. Insights into the pharmacology of new heterocycles embedded with oxopyrrolidine rings: DNA binding, molecular docking, and anticancer studies. *J Mol Liq* 2017; **234**: 391-402 [DOI: 10.1016/j.molliq.2017.03.112]
- 21 Ali I, Lone MN, Hsieh MF. N-Substituted (substituted-5-benzylidene) thiazolidine-2,4-diones: Crystal structure, In Silico, DNA binding and anticancer studies. *Biointerface Res Appl Chem* 2016; **6**: 1356-1379
- 22 Edge SB, Byrd DR, Compton CC, Fritz AG, Greene FL, Trotti A, editors. AJCC cancer staging manual. 7th ed. New York, NY: Springer, 2010: 143-164
- 23 Amin MB, Edge S, Greene F, Byrd DR, Brookland RK, Washington MK, Gershenwald JE, Compton CC, Hess KR, Sullivan DC, Jessup JM, Brierley JD, Gaspar LE, Schilsky RL, Balch CM, Winchester DP, Asare EA, Madera M, Gress DM, Meyer LR. AJCC Cancer Staging Manual. 8th ed. New York, NY: Springer, 2017: 252-254 [PMID: 28515669 DOI: 10.1007/978-3-319-40618-3]
- 24 National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®): Colon Cancer, 2016. Available from: URL: https://www.nccn.org/professionals/physician_gls/default.aspx
- 25 National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®): Rectal Cancer, 2016. Available from: URL: <https://www.nccn.org/>

- professionals/physician_gls/default.aspx
- 26 **Yao HW**, Wu HW, Liu YH. [From traditional population-based approach to individualized precision medicine: the interpretation of update on *The AJCC Colorectal Cancer Staging System, Eighth Edition*]. *Zhonghua Wai Ke Za Zhi* 2017; **55**: 24-27 [PMID: 28056249 DOI: 10.3760/cma.j.issn.0529-5815.2017.01.007]
- 27 **AJCC Cancer Staging Manual**. 6th ed. New York, NY: Springer: 2002: 1-416
- 28 **O'Connell JB**, Maggard MA, Ko CY. Colon cancer survival rates with the new American Joint Committee on Cancer sixth edition staging. *J Natl Cancer Inst* 2004; **96**: 1420-1425 [PMID: 15467030 DOI: 10.1093/jnci/djh275]
- 29 **Yao HW**, Liu YH. Update of 7th edition TNM staging for colorectal cancer and its significance. *Zhonghua Waike Zazhi* 2010; **48**: 1601-1604 [DOI: 10.3760/cma.j.issn.0529-5815.2010.21.001]
- 30 **Chen SL**, Bilchik AJ. More extensive nodal dissection improves survival for stages I to III of colon cancer: a population-based study. *Ann Surg* 2006; **244**: 602-610 [PMID: 16998369 DOI: 10.1097/01.sla.0000237655.11717.50]
- 31 **Johnson PM**, Porter GA, Ricciardi R, Baxter NN. Increasing negative lymph node count is independently associated with improved long-term survival in stage IIIB and IIIC colon cancer. *J Clin Oncol* 2006; **24**: 3570-3575 [PMID: 16877723 DOI: 10.1200/JCO.2006.06.8866]
- 32 **Steele SR**, Chen SL, Stojadinovic A, Nissan A, Zhu K, Peoples GE, Bilchik A. The impact of age on quality measure adherence in colon cancer. *J Am Coll Surg* 2011; **213**: 95-103; discussion 104-5 [PMID: 21601492 DOI: 10.1016/j.jamcollsurg.2011.04.013]
- 33 **Morikawa T**, Tanaka N, Kuchiba A, Nosho K, Yamauchi M, Hornick JL, Swanson RS, Chan AT, Meyerhardt JA, Huttenhower C, Schrag D, Fuchs CS, Ogino S. Predictors of lymph node count in colorectal cancer resections: data from US nationwide prospective cohort studies. *Arch Surg* 2012; **147**: 715-723 [PMID: 22508672 DOI: 10.1001/archsurg.2012.353]
- 34 **Stojadinovic A**, Nissan A, Wainberg Z, Shen P, McCarter M, Protic M, Howard RS, Steele SR, Peoples GE, Bilchik A. Time-dependent trends in lymph node yield and impact on adjuvant therapy decisions in colon cancer surgery: an international multi-institutional study. *Ann Surg Oncol* 2012; **19**: 4178-4185 [PMID: 22805869 DOI: 10.1245/s10434-012-2501-5]
- 35 **Kim SH**, Ha TK, Kwon SJ. Evaluation of the 7th AJCC TNM Staging System in Point of Lymph Node Classification. *J Gastric Cancer* 2011; **11**: 94-100 [PMID: 22076209 DOI: 10.5230/jgc.2011.11.2.94]
- 36 **Chen SL**, Steele SR, Eberhardt J, Zhu K, Bilchik A, Stojadinovic A. Lymph node ratio as a quality and prognostic indicator in stage III colon cancer. *Ann Surg* 2011; **253**: 82-87 [PMID: 21135690 DOI: 10.1097/SLA.0b013e3181ffa780]
- 37 **Hong KD**, Lee SI, Moon HY. Lymph node ratio as determined by the 7th edition of the American Joint Committee on Cancer staging system predicts survival in stage III colon cancer. *J Surg Oncol* 2011; **103**: 406-410 [PMID: 21400524 DOI: 10.1002/jso.21830]
- 38 **Wang J**, Hassett JM, Dayton MT, Kulaylat MN. Lymph node ratio: role in the staging of node-positive colon cancer. *Ann Surg Oncol* 2008; **15**: 1600-1608 [PMID: 18327530 DOI: 10.1245/s10434-007-9716-x]
- 39 **Wong SL**. Lymph node evaluation in colon cancer: assessing the link between quality indicators and quality. *JAMA* 2011; **306**: 1139-1141 [PMID: 21917585 DOI: 10.1001/jama.2011.1318]
- 40 **Compton CC**, Fielding LP, Burgart LJ, Conley B, Cooper HS, Hamilton SR, Hammond ME, Henson DE, Hutter RV, Nagle RB, Nielsen ML, Sargent DJ, Taylor CR, Welton M, Willett C. Prognostic factors in colorectal cancer. College of American Pathologists Consensus Statement 1999. *Arch Pathol Lab Med* 2000; **124**: 979-994 [PMID: 10888773 DOI: 10.1043/0003-9985(2000)124<0979:PFICC>2.0.CO;2]
- 41 **Bilchik A**, Stojadinovic A. Is it time to move beyond lymph node evaluation in the staging of colon cancer? *Arch Surg* 2010; **145**: 830-831 [PMID: 20873003 DOI: 10.1001/archsurg.2010.181]
- 42 **Lin AY**, Chua MS, Choi YL, Yeh W, Kim YH, Azzi R, Adams GA, Sainani K, van de Rijn M, So SK, Pollack JR. Comparative profiling of primary colorectal carcinomas and liver metastases identifies LEF1 as a prognostic biomarker. *PLoS One* 2011; **6**: e16636 [PMID: 21383983 DOI: 10.1371/journal.pone.0016636]
- 43 **Morikawa T**, Kuchiba A, Liao X, Imamura Y, Yamauchi M, Qian ZR, Nishihara R, Sato K, Meyerhardt JA, Fuchs CS, Ogino S. Tumor TP53 expression status, body mass index and prognosis in colorectal cancer. *Int J Cancer* 2012; **131**: 1169-1178 [PMID: 22038927 DOI: 10.1002/ijc.26495]
- 44 **Morikawa T**, Kuchiba A, Qian ZR, Mino-Kenudson M, Hornick JL, Yamauchi M, Imamura Y, Liao X, Nishihara R, Meyerhardt JA, Fuchs CS, Ogino S. Prognostic significance and molecular associations of tumor growth pattern in colorectal cancer. *Ann Surg Oncol* 2012; **19**: 1944-1953 [PMID: 22189472 DOI: 10.1245/s10434-011-2174-5]
- 45 **Jung SB**, Lee HI, Oh HK, Shin IH, Jeon CH. Clinico-pathologic Parameters for Prediction of Microsatellite Instability in Colorectal Cancer. *Cancer Res Treat* 2012; **44**: 179-186 [PMID: 23091444 DOI: 10.4143/crt.2012.44.3.179]
- 46 **Bateman AC**, Carr NJ, Warren BF. The retroperitoneal surface in distal caecal and proximal ascending colon carcinoma: the Cinderella surgical margin? *J Clin Pathol* 2005; **58**: 426-428 [PMID: 15790712 DOI: 10.1136/jcp.2004.019802]
- 47 **Petersen VC**, Baxter KJ, Love SB, Shepherd NA. Identification of objective pathological prognostic determinants and models of prognosis in Dukes' B colon cancer. *Gut* 2002; **51**: 65-69 [PMID: 12077094 DOI: 10.1136/gut.51.1.65]
- 48 **Roth AD**, Delorenzi M, Tejpar S, Yan P, Klingbiel D, Fiocca R, d'Ario G, Cisar L, Labianca R, Cunningham D, Nordlinger B, Bosman F, Van Cutsem E. Integrated analysis of molecular and clinical prognostic factors in stage II/III colon cancer. *J Natl Cancer Inst* 2012; **104**: 1635-1646 [PMID: 23104212 DOI: 10.1093/jnci/djs427]
- 49 **Pilozzi E**, Ferri M, Onelli MR, Mercantini P, Corigliano N, Duranti E, Dionisi L, Felicioni F, Virgilio E, Ziparo V, Ruco L. Prognostic significance of 18q LOH in sporadic colorectal carcinoma. *Am Surg* 2011; **77**: 38-43 [PMID: 21396303]
- 50 **de Ridder JA**, Knijn N, Wiering B, de Wilt JH, Nagtegaal ID. Lymphatic Invasion is an Independent Adverse Prognostic Factor in Patients with Colorectal Liver Metastasis. *Ann Surg Oncol* 2015; **22** Suppl 3: S638-S645 [PMID: 25986865 DOI: 10.1245/s10434-015-4562-8]
- 51 **Adamczyk LA**, Gordon K, Kholová I, Meijer-Jorna LB, Telinius N, Gallagher PJ, van der Wal AC, Baandrup U. Lymph vessels: the forgotten second circulation in health and disease. *Virchows Arch* 2016; **469**: 3-17 [PMID: 27173782 DOI: 10.1007/s00428-016-1945-6]
- 52 **Gomez D**, Zaitoun AM, De Rosa A, Hossaini S, Beckingham IJ, Brooks A, Cameron IC. Critical review of the prognostic significance of pathological variables in patients undergoing resection for colorectal liver metastases. *HPB (Oxford)* 2014; **16**: 836-844 [PMID: 24617566 DOI: 10.1111/hpb.12216]
- 53 **Chen JW**, Bhandari M, Astill DS, Wilson TG, Kow L, Brooke-Smith M, Toouli J, Padbury RT. Predicting patient survival after pancreaticoduodenectomy for malignancy: histopathological criteria based on perineural infiltration and lymphovascular invasion. *HPB (Oxford)* 2010; **12**: 101-108 [PMID: 20495653 DOI: 10.1111/j.1477-2574.2009.00140.x]
- 54 **Longatto Filho A**, Oliveira TG, Pinheiro C, de Carvalho MB, Curioni OA, Mercante AM, Schmitt FC, Gattás GJ. How useful is the assessment of lymphatic vascular density in oral carcinoma prognosis? *World J Surg Oncol* 2007; **5**: 140 [PMID: 18072963 DOI: 10.1186/1477-7819-5-140]
- 55 **Lee DJ**, Kwon MJ, Nam ES, Kwon JH, Kim JH, Rho YS, Shin HS, Cho SJ. Histopathologic predictors of lymph node metastasis and prognosis in tonsillar squamous cell carcinoma. *Korean J Pathol* 2013; **47**: 203-210 [PMID: 23837012 DOI: 10.4132/KoreanJPathol.2013.47.3.203]
- 56 **Zlobec I**, Lugli A. Invasive front of colorectal cancer: dynamic interface of pro-/anti-tumor factors. *World J Gastroenterol* 2009;

- 15: 5898-5906 [PMID: 20014453 DOI: 10.3748/wjg.15.5898]
- 57 **Hoogstins CE**, Weixler B, Boogerd LS, Hoppener DJ, Prevoo HA, Sier CF, Burger JW, Verhoef C, Bhairosingh S, Farina Sarasqueta A, Burggraaf J, Vahrmeijer AL. In Search for Optimal Targets for Intraoperative Fluorescence Imaging of Peritoneal Metastasis From Colorectal Cancer. *Biomark Cancer* 2017; **9**: 1179299X17728254 [PMID: 28874886 DOI: 10.1177/1179299X17728254]
- 58 **Lin EK**, Hsieh MC, Chen CH, Lu YJ, Wu SY. Outcomes of cytoreductive surgery and hyperthermic intraperitoneal chemotherapy for colorectal cancer with peritoneal metastasis. *Medicine (Baltimore)* 2016; **95**: e5522 [PMID: 28033247 DOI: 10.1097/MD.0000000000005522]
- 59 **Glockzin G**, Schlitt HJ, Piso P. Therapeutic options for peritoneal metastasis arising from colorectal cancer. *World J Gastrointest Pharmacol Ther* 2016; **7**: 343-352 [PMID: 27602235 DOI: 10.4292/wjgpt.v7.i3.343]
- 60 **Li L**, Deng R, Su Y, Yang C. Dual-targeting nanoparticles with excellent gene transfection efficiency for gene therapy of peritoneal metastasis of colorectal cancer. *Oncotarget* 2017; **8**: 89837-89847 [PMID: 29163792 DOI: 10.18632/oncotarget.21159]
- 61 **Glockzin G**, Gerken M, Lang SA, Klinkhammer-Schalke M, Piso P, Schlitt HJ. Oxaliplatin-based versus irinotecan-based hyperthermic intraperitoneal chemotherapy (HIPEC) in patients with peritoneal metastasis from appendiceal and colorectal cancer: a retrospective analysis. *BMC Cancer* 2014; **14**: 807 [PMID: 25369730 DOI: 10.1186/1471-2407-14-807]

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Giant exophytic renal angiomyolipoma masquerading as a retroperitoneal liposarcoma: A case report and review of literature

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Author contributions: Sharma G, Jain A, and Garg PK designed the report; all the authors actively managed the patient; Sharma G, Jain A, and Garg PK collected the patient's clinical data; Sharma S provided the histopathological images; Rathi V provided the radiological images. All the authors analyzed the case, drafted the manuscript and finally approved it.

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Abstract

A 42-years-old lady, presented with a large retroperitoneal mass which was preoperatively diagnosed as a retroperitoneal liposarcoma following an image guided core biopsy. She underwent a margin-negative resection of the retroperitoneal mass (multi visceral resection - enbloc excision of the retroperitoneal mass with a left nephrectomy and a segmental descending colectomy). The final histopathological examination of the resected specimen confirmed an exophytic renal angiomyolipoma (AML) which was extending into the retroperitoneum. AML is a rare benign tumor arising most commonly from the kidney. It can sometimes present as a diagnostic challenge as it mimics a retroperitoneal liposarcoma or a fat-containing renal cell carcinomas closely. We present this case to share our experience of managing a case of giant exophytic AML which resembled retroperitoneal liposarcoma closely and resulted into an aggressive

surgery.

Key words: Angiomyolipoma; Retroperitoneum; Liposarcoma; Diagnosis; Biopsy

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Core tip: A giant exophytic renal angiomyolipoma (AML) can pose a serious diagnostic challenge and may be confused with a retroperitoneal sarcoma. A discordance of the radiological and core biopsy findings in a suspected case of an exophytic renal AML must lead to re-evaluation of the case and repeat biopsies may further clarify the diagnosis.

Sharma G, Jain A, Sharma P, Sharma S, Rath V, Garg PK. Giant exophytic renal angiomyolipoma masquerading as a retroperitoneal liposarcoma: A case report and review of literature. *World J Clin Oncol* 2018; 9(7): 162-166 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v9/i7/162.htm> DOI: <http://dx.doi.org/10.5306/wjco.v9.i7.162>

INTRODUCTION

Angiomyolipoma (AML) is a benign tumor primarily containing fat, atypical blood vessels and smooth muscles in varying proportions^[1]. The common site of AML is kidney and it presents primarily as an intra-renal mass. Occasionally, a renal AML may outgrow exophytically in the retroperitoneum and closely mimics a liposarcoma due to the high fat content on a radiological imaging. The diagnostic dilemma is further compounded as the core-cut biopsy may also be erroneously reported as liposarcoma^[2]. This may result in an aggressive surgery, and at times multi-visceral resections, to achieve a margin-negative resection. A post-resection histopathological diagnosis of AML may come as a surprise for both the surgeons and the patients, and may even lead to litigations. We are reporting a case of a giant exophytic AML which was misdiagnosed preoperatively as a retroperitoneal liposarcoma following an image guided core biopsy examination.

CASE REPORT

A 42-years-old lady presented to us with complaints of abdominal pain and fullness over the left side of the abdomen for two months duration. The pain was mild, dull aching, localized and continuous with a gradual increase in intensity over a period of time. She denied having hematuria, flank pain, dysuria, anorexia, rectal bleeding, malena or vaginal bleeding. She also denied any previous history of fever or blood transfusion. She reported to have undergone bilateral salpingo-oophorectomy around 3 years back for a left adnexal mass; no medical documents were available for this

surgery. She developed amenorrhea following bilateral salpingo-oophorectomy, and she was not taking hormone replacement therapy. She also did not report any other significant past medical or family history. She had three children; all were borne as normal vaginal delivery and were hale and hearty. Her general physical examination was unremarkable. Her body mass index was 27.5. Her abdominal examination revealed a firm, non-tender, immobile, bimanually palpable intra-abdominal lump of size 25 × 20 cm involving left hypochondrium, left lumbar, umbilical, and left iliac fossa. Rest of her physical examination was unremarkable. Her blood investigations showed a hemoglobin of 6.6 g/dL and a total leucocyte count of 12000/ μ L. Her platelet count, liver function tests and renal function tests were within normal limits. Her contrast-enhanced computed tomography (CT) of the abdomen suggested two fairly well defined, predominantly fat density rounded lesions in the left kidney. The peripheral capsule of the smaller lesion was ill defined and was continuous with a large perirenal mass showing prominent vessels (Figure 1). There was also a peripheral hematoma in the perirenal lipomatous mass. An ultrasonography guided core biopsy of the retroperitoneal mass suggested liposarcoma in view of the variable sized adipocytes with atypia (Figure 2). Surgical exploration revealed a large retroperitoneal mass involving the left kidney and a segment of descending colon. The patient underwent a margin-negative resection of the retroperitoneal mass (multi visceral resection - enbloc excision of the retroperitoneal mass with a left nephrectomy and a segmental descending colectomy). Figure 3 displays the resected specimen. Her postoperative period was uneventful. Final histopathological examination of the resected specimen confirmed an exophytic renal AML which was extending into the retroperitoneum (Figure 4). A retrospective evaluation of the patient did not reveal any clinical stigmata of neurofibromatosis.

DISCUSSION

AML is a rare benign tumor which has a tri-phasic morphology - blood vessels, smooth muscles, and fat cells. Based on the fat content, AML is typically divided into classical or fat-rich type, and fat-poor AML. Both of these types are benign tumors with no metastatic potential. A third rare type is an epithelioid AML which is rare but has a malignant potential. Though fat-poor AML poses a diagnostic challenge due to the presence of scattered fat cells in the smooth muscles, classical renal AML can be easily diagnosed based on the contrast enhancement CT features: (A) the demonstration of the fat cells as the presence of regions of interest containing attenuations less than < 10-20 HU; and (B) the location of the fat cells within the lesion and not in another structure near it^[3]. However, one must not forget that there are other kinds of lesions which may contain fat and mimic AML.

When this fat-rich renal AML grows exophytically and

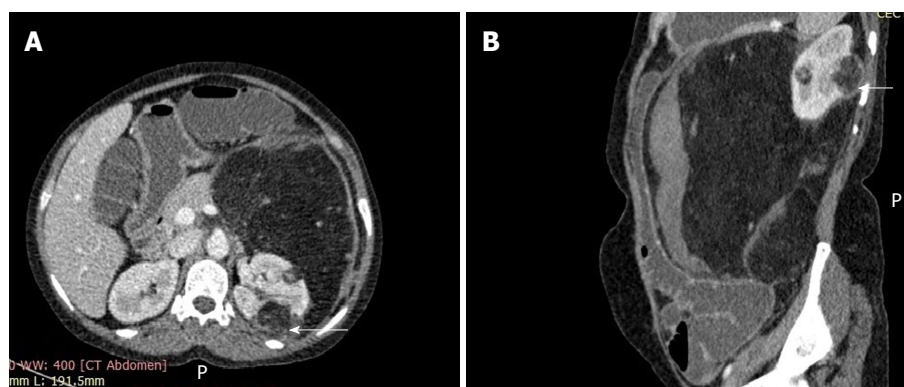


Figure 1 Axial section of contrast enhancement computed tomography abdomen. A: It shows two fairly well defined, predominantly fat density rounded lesions, in the left kidney. The peripheral capsule (arrow) of the smaller lesion is ill defined and continuous with a very large perirenal angiomyolipoma showing prominent vessels (+); B: Sagittal reconstruction shows a peripheral hematoma (arrow) in the perirenal lipomatous mass.

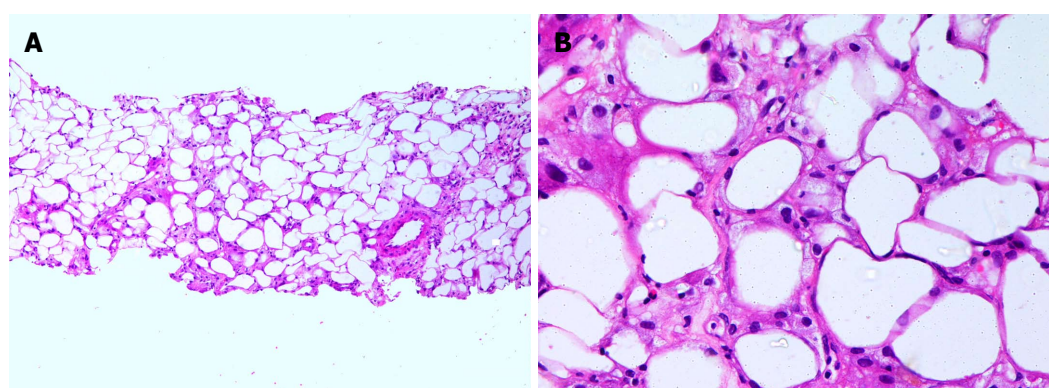


Figure 2 Preoperative core biopsy of tumor mass. A: Variable sized adipocytes (10 ×); B: Variable sized adipocytes with atypia (40 ×).

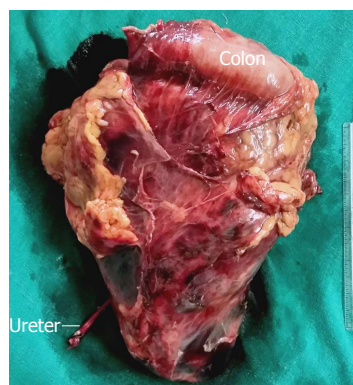


Figure 3 Resected enbloc specimens of retroperitoneal mass, left kidney, and colon.

extends into the retroperitoneum, it becomes difficult to differentiate it from a retroperitoneal sarcoma, especially a liposarcoma. As the management strategies are entirely different from these two entities - wide margin-negative resection for liposarcoma vs conservative approach for AML, it is of utmost importance to differentiate them radiologically. In a retrospective study of CT images of 15 large exophytic renal AMLs and 12 well-differentiated perirenal liposarcomas, Israel *et al*^[4] suggested that a defect in the renal parenchyma

combined with the presence of the enlarged vessels may help differentiate an exophytic AML from a retroperitoneal sarcoma. In a study of 11 patients with the perirenal liposarcoma and 9 patients with the giant exophytic AMLs, Ellingson *et al*^[5] also concluded that the presence of tumoral vessel extending into the renal cortex or a renal parenchyma defect at the site of the tumor contact points strongly towards the diagnosis of AML while intra-tumoral calcification favors the presence of liposarcoma. However, Wang *et al*^[6] suggested that the intra-tumoral calcification can be seen in both AML and liposarcoma. They highlighted that the characteristics of a perinephric AML are intratumoral linear vascularity, aneurysmal dilatation, bridging vessel sign, hematoma, beak sign, and discrete intra-renal fatty tumors while the CT characteristics of a perinephric liposarcoma are a non-fatty soft tissue mass. Other close differentials of an exophytic AML include lipoma, leiomyoma with fatty change or a fat containing renal cell carcinoma.

A small biopsy of the lesion to confirm the diagnosis may also yield erroneous results due to the non-representative sample in view of the heterogenous nature of the AML^[5]. A positive immune-reactivity for human melanoma black 45, is characteristic of AMLs and may differentiate AMLs from other similar appearing

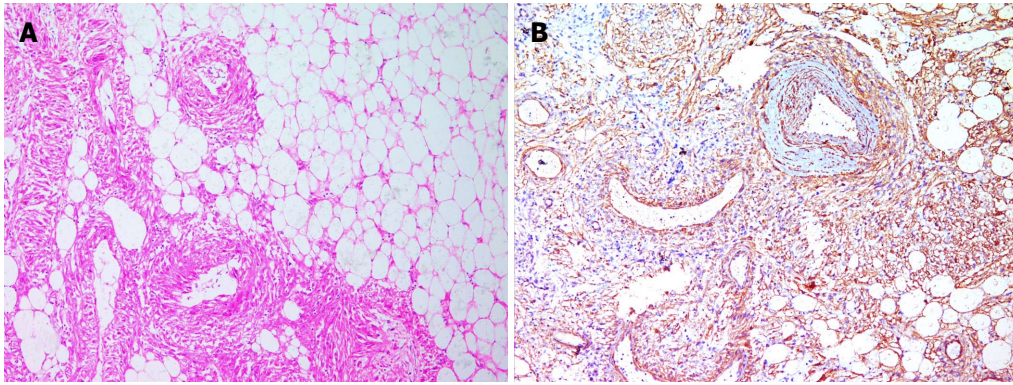


Figure 4 Final histopathology of surgical specimen. A: Components of all the three tissues, i.e., mature adipocytes, spindle shaped smooth muscle cells and blood vessel (10 ×); B: Immunohistochemical staining for smooth muscle actin shows strong positivity in final histology specimen.

lesions such as liposarcomas, lipomas, leiomyosarcomas or, leiomyomas^[7].

Our patient had a giant symptomatic exophytic AML. Though radiological findings were suggestive of AML, our patient was operated upon with a preoperative diagnosis of retroperitoneal liposarcoma due to the small image-guided biopsy which was reported as liposarcoma. This is likely to be due to a non-representative sample of the lesion. Though various conservative approaches including radiofrequency ablation, cryoablation, microwave ablation, selective angio-embolization are described to manage AML^[8], our patient would have otherwise also required surgical excision in view of the large size, the symptomatic nature, and the risk of potential life-threatening hemorrhage. However, a correct preoperative diagnosis would have helped us to take a more conservative resection.

We conclude that a giant exophytic renal AML can pose a serious diagnostic challenge and may be confused with a retroperitoneal liposarcoma. A discordance of the radiological and core biopsy findings in a suspected case of exophytic renal AML must lead to re-evaluation of the case and repeat biopsies may further clarify the diagnosis.

ARTICLE HIGHLIGHTS

Case characteristics

A 42-years-old lady came to our hospital with complaints of abdominal pain and distension of two months duration. The physical examination of the patient revealed large retroperitoneal lump present on the left side of the abdomen.

Clinical diagnosis

The clinical diagnosis was a retroperitoneal mass, likely to be malignant in nature.

Differential diagnosis

The differential diagnosis included retroperitoneal sarcoma, renal neoplasm, multicystic kidney, cold abscess, or hydatid cyst.

Laboratory diagnosis

Routine blood investigations did not reveal any abnormality.

Imaging diagnosis

Abdominal computed tomography revealed a large perirenal mass lesion

displaying prominent vessels. There were also two fairly well defined, predominantly fat density rounded lesions in the left kidney; the smaller lesion was continuous with the large perirenal mass.

Pathological diagnosis

Preoperative image guided biopsy of the retroperitoneal mass suggested liposarcoma.

Treatment

In view of preoperative pathological diagnosis of liposarcoma, the patient underwent a margin-negative resection of the retroperitoneal mass (multi visceral resection - en bloc excision of retroperitoneal mass with left nephrectomy and segmental descending colectomy).

Related reports

The final histopathological report of the resected specimen confirmed angiomyolipoma.

Term explanation

AML is a tumor of tri-phasic morphology - blood vessels, smooth muscles, and fat cells. Majority of the AML tumors are benign in nature with almost no malignant potential.

Experiences and lessons

A large exophytic renal AML may be confused with a retroperitoneal sarcoma on a small biopsy specimen. A discordance of the radiological and core biopsy findings in a suspected case of exophytic renal AML must alert the surgeon and a re-evaluation of the case with repeat biopsies may clarify the diagnosis.

REFERENCES

- 1 **Garg PK, Jain BK, Kumar A, Bhatt S, Vibhav V.** Fat poor angiomyolipoma with lymphadenopathy: Diagnostic dilemma. *Urol Ann* 2012; **4**: 126-129 [PMID: 22629015 DOI: 10.4103/0974-7796.95573]
- 2 **Kori C, Akhtar N, Vamsidhar PN, Gupta S, Kumar V.** Giant exophytic renal angiomyolipoma mimicking as retroperitoneal sarcoma; a case report with review of literature. *J Clin Diagn Res* 2015; **9**: XJ01-XJ02 [PMID: 26023631 DOI: 10.7860/JCDR/2015/11514.5798]
- 3 **Jinzaki M, Silverman SG, Akita H, Mikami S, Oya M.** Diagnosis of Renal Angiomyolipomas: Classic, Fat-Poor, and Epithelioid Types. *Semin Ultrasound CT MR* 2017; **38**: 37-46 [PMID: 28237279 DOI: 10.1053/j.sult.2016.11.001]
- 4 **Israel GM, Bosniak MA, Slywotzky CM, Rosen RJ.** CT differentiation of large exophytic renal angiomyolipomas and perirenal liposarcomas. *AJR Am J Roentgenol* 2002; **179**: 769-773 [PMID: 12185060 DOI: 10.2214/ajr.179.3.1790769]
- 5 **Ellingson JJ, Coakley FV, Joe BN, Qayyum A, Westphalen AC, Yeh BM.** Computed tomographic distinction of perirenal

- liposarcoma from exophytic angiomyolipoma: a feature analysis study. *J Comput Assist Tomogr* 2008; **32**: 548-552 [PMID: 18664840 DOI: 10.1097/RCT.0b013e3181507534]
- 6 **Wang LJ**, Wong YC, Chen CJ, See LC. Computerized tomography characteristics that differentiate angiomyolipomas from liposarcomas in the perinephric space. *J Urol* 2002; **167**: 490-493 [PMID: 11792904 DOI: 10.1016/S0022-5347(01)69071-2]
 - 7 **Minja EJ**, Pellerin M, Saviano N, Chamberlain RS. Retroperitoneal extrarenal angiomyolipomas: an evidence-based approach to a rare clinical entity. *Case Rep Nephrol* 2012; **2012**: 374107 [PMID: 24555133 DOI: 10.1155/2012/374107]
 - 8 **Jawahar A**, Kazan-Tannus J. Retroperitoneal extrarenal angiomyolipoma at the surgical bed 8 years after a renal angiomyolipoma nephrectomy: A case report and review of literature. *Urol Ann* 2017; **9**: 288-292 [PMID: 28794601 DOI: 10.4103/UA.UA_20_17]

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Malignant peritoneal effusion acting as a tumor environment in ovarian cancer progression: Impact and significance

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Abstract

Until recently, ovarian cancer research has mainly focused on the tumor cells themselves ignoring for the most part the surrounding tumor environment which includes malignant peritoneal effusions. However, one of the major conceptual advances in oncology over the last few years has been the appreciation that cancer progression cannot be explained by aberrations in cancer cells themselves and is strongly influenced by the surrounding tumor environment. The mechanisms of ovarian cancer progression differ from that of other solid tumors because ovarian cancer cells primarily disseminate within the peritoneal cavity. Malignant peritoneal effusion accumulates in the peritoneal cavity during ovarian cancer progression. These exudative fluids act as a unique tumor environment providing a framework that orchestrates cellular and molecular changes contributing to aggressiveness and disease progression. The composition of ascites, which includes cellular and acellular components, constantly adapts during the course of the disease in response to various cellular cues originating from both tumor and stromal cells. The tumor environment that represents peritoneal effusions closely constitute an ecosystem, with specific cell types and signaling molecules increasing and decreasing during the course of the disease progression creating a single complex network. Although recent advances aiming to understand the ovarian tumor environment have focused one at a time on components, the net impact of the whole environment cannot be understood simply from its parts or outside is environmental context.

Key words: Ovarian cancer; Tumor environment; Peritoneal effusions; Ascites; Dissemination; Multicellular spheroids

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Core tip: The malignant peritoneal effusion that accumulates during ovarian cancer dissemination and progression constitutes a unique tumor environment. Bidirectional communications between tumor cells and their surrounding environment influence ovarian cancer dissemination, progression and patient prognosis. To solve the complexity of this tumor environment and understand how it affects cancer progression, a paradigm shift is necessary. Peritoneal effusions should be studied as integrated systems with innovative modeling approaches.

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MAIN TEXT

Malignant peritoneal effusions (ascites) commonly occur during the progression of certain types of cancers such as ovarian and pancreatic adenocarcinomas^[1]. These exudative fluids contain a large number of tumor cells and are by definition metastatic. For example, patients with ovarian cancer will often present at diagnosis with multiple omental and peritoneal tumor implants. These patients will usually have accumulation of large amount of peritoneal fluids (often several liters). The presence of malignant effusion is associated with a poor prognosis. Because of their nature, malignant effusions, in which tumor cells proliferate and metastasize, constitute a distinct and unique tumor environment compared to solid cancers that develop in a tissue microenvironment. The cell-free fraction of malignant effusions is rich in cytokines, chemokines, growth factors and extracellular matrix components^[2-4]. In addition to tumor cells, effusions contain a large number of non-malignant cells such as fibroblasts and mesothelial cells, and various immune cells including macrophages and lymphocytes^[5]. Although the importance of the bidirectional communication between tumor cells and their adjacent tissue microenvironment for supporting cancer progression is well recognized now, much less is known regarding the role peritoneal effusions in cancer progression. Nonetheless, it has been shown that extracellular cues provided by soluble factors contained in effusions can drive certain aspects of cancer progression such as cell proliferation, migration, and invasion. For example, effusion-associated hepatocyte growth factor^[6-8], lysophosphatidic acid^[9] and CCL18^[10] have been shown to promote ovarian cancer cell growth and survival. Furthermore, cell-free ascites can promote drug resistance in ovarian cancer cells^[11,12]. In support of the critical role of cell-free ascites, a number of factors contained in effusions are predictor of clinical prognosis^[13,14]. The overall effect of cell-free ascites results from the complex integration of various extracellular cues, which leading to receptor-induced signaling in both tumor cells and stromal cells. For example, in contact with ascites, mesothelial cells undergo a pro-tumoral shift^[15] whereas ovarian cancer cells display a more invasive phenotype with a shift along an epithelial-to-mesenchymal transition (EMT)^[16]. As a result of this extensive cellular crosstalk, peritoneal effusions are constantly adapting during the course of the disease in response to the different cues^[1]. The functional role of these changes in effusion composition during cancer dissemination and progression is mostly unknown.

Beyond the potential contribution of specific soluble factors to cancer progression, non-malignant cells present in peritoneal effusions probably play an essential supporting role, particularly in the early stages of dissemination, by preventing anoikis and stimulating proliferation of tumor cells. Paracrine signaling through mesenchymal stem cells can confer resistance to chemotherapy in ovarian cancer cells via IL-6/IL-6 R pathway and CXCL12^[17,18]. Similarly, paracrine secretion of anti-apoptotic from ascites-primed mesothelial cells promote ovarian cancer cell drug resistance^[15]. Cancer-associated fibroblasts isolated from omentum can be activated by tumor cells to promote ovarian cancer growth, adhesion and invasiveness through TGFβ1 pathway^[19]. Expression of HOXA9 by ovarian cancer cells induce TGFβ2 secretion, which in turn, promote by secretion of IL-6 and CXCL12 by fibroblasts leading to tumor growth^[20]. The presence of tumor-infiltrating CD8⁺ T cells in primary tumors is associated with prolonged progression-free and overall survival in ovarian cancer patients^[21,22]. These data suggest that several cell types within peritoneal effusions may have a robust influence on ovarian cancer cells. Bidirectional communications between tumor cells and their surrounding environment are at play, however identifying these crosstalks and validating their functional roles in a systemic approach remains challenging because of complex interactions among multiple cell types in effusions.

Getting a better understanding of the complex ecosystem that constitute malignant peritoneal effusions also have the potential to enhance our insight to limit the accumulation of these effusions and to overcome platinum resistance in ovarian cancer, which represents a yet unmet need. Both soluble factors and cancer-associated stromal and immune cells constitute potential targets for novel therapies. Examples of

soluble factors in the tumor environment of ovarian cancer targeted by novel therapies (mostly humanized monoclonal antibodies) include VEGF, VEGFR, PDGFR, PDGFR, c-kit, PD-1 and PD-L1^[23]. However, drug-associated toxicity and resistance often limit the clinical benefits associated with these novel therapies. These studies highlight the fact that targeting a single component of the tumor environment is likely to have limited clinical benefits and the secretome of malignant peritoneal effusions should be considered as a whole that cannot be understood simply from its parts. Given the progression-enhancing role of cancer-associated stromal cells in malignant peritoneal effusions, cell therapy may represent of novel approach to limit ovarian cancer dissemination. For example, it has been shown that genetically-engineered human mesothelial cells with the Herpes Simplex virus thymidine kinase (HSV-tk) efficiently deliver anticancer modalities to ovarian cancer cells within the peritoneal cavity when mice were treated with ganciclovir^[24]. Conversion of ganciclovir pro-drug in a toxic apoptosis-inducing drug by HSV-TK results in the transfer of apoptotic bodies to adjacent tumor cells. Alternatively, the cells could be used to locally deliver in the peritoneal cavity other anti-cancer molecules potentially circumventing the toxicity associated with systemic delivery.

Tumor such as ovarian cancer does not usually use the classic patterns of metastasis via the hematogenous route^[25]. The current view is that ovarian cancer spreads mainly via the peritoneal circulation with a high affinity for the omentum. Cells detach from the primary tumor to form free-floating multicellular spheroids, which then travel through the peritoneal fluid at distant sites onto the mesothelial lining where metastatic outgrowth occurs^[26]. It is believed that the formation of multicellular spheroids is an essential step in the initiation of peritoneal implantation metastasis for ovarian cancer. In this context, spheroids can be considered the primary vehicles of ovarian cancer dissemination. However, the mechanisms of spheroid formation are for the most part unknown. Before tumor cells detach, they will usually undergo an EMT characterized by decreased expression of E-cadherin acquiring therefore a proliferative and invasive phenotype. However, it is not clear if tumor cells detached as single cell entity and then aggregate together to form homotypic multicellular spheroids or if clumps of the primary tumor exfoliate and stay together to form heterotypic spheroids. Alternatively, exfoliated single tumor cells could aggregate with floating mesothelial cells forming heterotypic unvascularized multicellular spheroids. Recent studies aiming to characterize ascites-derived spheroids suggested that spheroids are heterotypic containing mesothelial-derived myofibroblastic cells^[27] and macrophages^[28]. The binding of ovarian cancer cells to mesothelial cells to form spheroids could be mediated by integrins, CD44 and MUC16^[27,29,30]. Further work however is needed to gain a comprehensive understanding of the cellular composition of the multicellular spheroids found in ascites and how this influences cancer progression. Another interesting hypothesis is that multicellular spheroids potentially constitute a chemoresistant niche that continuously repopulates the peritoneal cavity despite chemotherapy treatments as they are potentially endowed with a high tumor-initiating potential and increased drug resistance linked to the expression of stemness-associated genes^[31]. However, the presence of mesenchymal stem cells in ovarian cancer spheroids remains to be demonstrated. The generation of unvascularized 3D spheroids would generate a structure with a metabolite density gradient that can inhibit the access of chemotherapy agents to internal cells^[32]. Ovarian cancer cell grown as spheroids display enhanced resistance to common chemotherapeutic drugs such as taxol and cisplatin compared monolayers^[31,33]. Because of their role in ovarian cancer dissemination and their potential role in disease recurrence after chemotherapy, a much better understanding of multicellular spheroid biology is necessary.

PERSPECTIVE

As highlighted here, most research on the ovarian cancer environment has focused one at a time on components, such as particular signaling cascades, specific ascites factors or specific cell types, *etc.* This knowledge is essential, but obviously not sufficient. The tumor environment that represents peritoneal effusions closely resembles an ecosystem, with specific cell types and signaling molecules increasing and decreasing during the course of the disease progression creating a single complex network. This environment is an archetypical complex system in which the functioning of the whole cannot be understood simply from its parts or outside is environmental context. Perhaps to understand the role of peritoneal effusions on cancer progression, a paradigm shift is necessary and effusions should be studied as integrated systems using bioinformatic modeling to quantify system-level

biodiversity changes^[34].

REFERENCES

- 1 **Matte I**, Bessette P, Piché A. Ascites in ovarian cancer progression: opportunities for biomarker discovery and new avenues for targeted therapies. *INTECH* 2017; **Chapter 9**: 146-163 [DOI: [10.5772/intechopen.70993](https://doi.org/10.5772/intechopen.70993)]
- 2 **Matte I**, Lane D, Laplante C, Rancourt C, Piché A. Profiling of cytokines in human epithelial ovarian cancer ascites. *Am J Cancer Res* 2012; **2**: 566-580 [PMID: [22957308](https://pubmed.ncbi.nlm.nih.gov/22957308/)]
- 3 **Ahmed N**, Stenvers KL. Getting to know ovarian cancer ascites: opportunities for targeted therapy-based translational research. *Front Oncol* 2013; **3**: 256 [PMID: [24093089](https://pubmed.ncbi.nlm.nih.gov/24093089/) DOI: [10.3389/fonc.2013.00256](https://doi.org/10.3389/fonc.2013.00256)]
- 4 **Giuntoli RL 2nd**, Webb TJ, Zoso A, Rogers O, Diaz-Montes TP, Bristow RE, Oelke M. Ovarian cancer-associated ascites demonstrates altered immune environment: implications for antitumor immunity. *Anticancer Res* 2009; **29**: 2875-2884 [PMID: [19661290](https://pubmed.ncbi.nlm.nih.gov/19661290/)]
- 5 **Kipps E**, Tan DS, Kaye SB. Meeting the challenge of ascites in ovarian cancer: new avenues for therapy and research. *Nat Rev Cancer* 2013; **13**: 273-282 [PMID: [23426401](https://pubmed.ncbi.nlm.nih.gov/23426401/) DOI: [10.1038/nrc3432](https://doi.org/10.1038/nrc3432)]
- 6 **Nakamura M**, Ono YJ, Kanemura M, Tanaka T, Hayashi M, Terai Y, Ohmichi M. Hepatocyte growth factor secreted by ovarian cancer cells stimulates peritoneal implantation via the mesothelial-mesenchymal transition of the peritoneum. *Gynecol Oncol* 2015; **139**: 345-354 [PMID: [26335595](https://pubmed.ncbi.nlm.nih.gov/26335595/) DOI: [10.1016/j.ygyno.2015.08.010](https://doi.org/10.1016/j.ygyno.2015.08.010)]
- 7 **Matte I**, Lane D, Laplante C, Garde-Granger P, Rancourt C, Piché A. Ovarian cancer ascites enhance the migration of patient-derived peritoneal mesothelial cells via cMet pathway through HGF-dependent and -independent mechanisms. *Int J Cancer* 2015; **137**: 289-298 [PMID: [25482018](https://pubmed.ncbi.nlm.nih.gov/25482018/) DOI: [10.1002/ijc.29385](https://doi.org/10.1002/ijc.29385)]
- 8 **Kwon Y**, Smith BD, Zhou Y, Kaufman MD, Godwin AK. Effective inhibition of c-MET-mediated signaling, growth and migration of ovarian cancer cells is influenced by the ovarian tissue microenvironment. *Oncogene* 2015; **34**: 144-153 [PMID: [24362531](https://pubmed.ncbi.nlm.nih.gov/24362531/) DOI: [10.1038/ncr.2013.539](https://doi.org/10.1038/ncr.2013.539)]
- 9 **Mills GB**, Moolenaar WH. The emerging role of lysophosphatidic acid in cancer. *Nat Rev Cancer* 2003; **3**: 582-591 [PMID: [12894246](https://pubmed.ncbi.nlm.nih.gov/12894246/) DOI: [10.1038/nrc1143](https://doi.org/10.1038/nrc1143)]
- 10 **Lane D**, Matte I, Laplante C, Garde-Granger P, Carignan A, Bessette P, Rancourt C, Piché A. CCL18 from ascites promotes ovarian cancer cell migration through proline-rich tyrosine kinase 2 signaling. *Mol Cancer* 2016; **15**: 58 [PMID: [27613122](https://pubmed.ncbi.nlm.nih.gov/27613122/) DOI: [10.1186/s12943-016-0542-2](https://doi.org/10.1186/s12943-016-0542-2)]
- 11 **Lane D**, Goncharenko-Khaider N, Rancourt C, Piché A. Ovarian cancer ascites protects from TRAIL-induced cell death through alpha5beta1 integrin-mediated focal adhesion kinase and Akt activation. *Oncogene* 2010; **29**: 3519-3531 [PMID: [20400979](https://pubmed.ncbi.nlm.nih.gov/20400979/) DOI: [10.1038/ncr.2010.107](https://doi.org/10.1038/ncr.2010.107)]
- 12 **Goncharenko-Khaider N**, Matte I, Lane D, Rancourt C, Piché A. Ovarian cancer ascites increase Mcl-1 expression in tumor cells through ERK1/2-Elk-1 signaling to attenuate TRAIL-induced apoptosis. *Mol Cancer* 2012; **11**: 84 [PMID: [23158473](https://pubmed.ncbi.nlm.nih.gov/23158473/) DOI: [10.1186/1476-4598-11-84](https://doi.org/10.1186/1476-4598-11-84)]
- 13 **Kryczek I**, Gryboś M, Karabon L, Klimczak A, Lange A. IL-6 production in ovarian carcinoma is associated with histotype and biological characteristics of the tumour and influences local immunity. *Br J Cancer* 2000; **82**: 621-628 [PMID: [10682675](https://pubmed.ncbi.nlm.nih.gov/10682675/) DOI: [10.1054/bjoc.1999.0973](https://doi.org/10.1054/bjoc.1999.0973)]
- 14 **Lane D**, Matte I, Rancourt C, Piché A. Prognostic significance of IL-6 and IL-8 ascites levels in ovarian cancer patients. *BMC Cancer* 2011; **11**: 210 [PMID: [21619709](https://pubmed.ncbi.nlm.nih.gov/21619709/) DOI: [10.1186/1471-2407-11-210](https://doi.org/10.1186/1471-2407-11-210)]
- 15 **Matte I**, Lane D, Bachvarov D, Rancourt C, Piché A. Role of malignant ascites on human mesothelial cells and their gene expression profiles. *BMC Cancer* 2014; **14**: 288 [PMID: [24761768](https://pubmed.ncbi.nlm.nih.gov/24761768/) DOI: [10.1186/1471-2407-14-288](https://doi.org/10.1186/1471-2407-14-288)]
- 16 **Carduner L**, Leroy-Dudal J, Picot CR, Gallet O, Carreiras F, Kellouche S. Ascites-induced shift along epithelial-mesenchymal spectrum in ovarian cancer cells: enhancement of their invasive behavior partly dependant on av integrins. *Clin Exp Metastasis* 2014; **31**: 675-688 [PMID: [24946950](https://pubmed.ncbi.nlm.nih.gov/24946950/) DOI: [10.1007/s10585-014-9658-1](https://doi.org/10.1007/s10585-014-9658-1)]
- 17 **Pasquier J**, Gosset M, Geyl C, Hoarau-Véhot J, Chevrot A, Pocard M, Mirshahi M, Lis R, Rafii A, Touboul C. CCL2/CCL5 secreted by the stroma induce IL-6/PYK2 dependent chemoresistance in ovarian cancer. *Mol Cancer* 2018; **17**: 47 [PMID: [29455640](https://pubmed.ncbi.nlm.nih.gov/29455640/) DOI: [10.1186/s12943-018-0787-z](https://doi.org/10.1186/s12943-018-0787-z)]
- 18 **Lis R**, Touboul C, Mirshahi P, Ali F, Mathew S, Nolan DJ, Maleki M, Abdalla SA, Raynaud CM, Querleu D. Tumor associated mesenchymal stem cells protects ovarian cancer cells from hyperthermia through CXCL12. *Int J Cancer* 2011; **128**: 715-725 [PMID: [20725999](https://pubmed.ncbi.nlm.nih.gov/20725999/) DOI: [10.1002/ijc.25619](https://doi.org/10.1002/ijc.25619)]
- 19 **Cai J**, Tang H, Xu L, Wang X, Yang C, Ruan S, Guo J, Hu S, Wang Z. Fibroblasts in omentum activated by tumor cells promote ovarian cancer growth, adhesion and invasiveness. *Carcinogenesis* 2012; **33**: 20-29 [PMID: [22021907](https://pubmed.ncbi.nlm.nih.gov/22021907/) DOI: [10.1093/carcin/bgr230](https://doi.org/10.1093/carcin/bgr230)]
- 20 **Ko SY**, Barengo N, Ladanyi A, Lee JS, Marini F, Lengyel E, Naora H. HOXA9 promotes ovarian cancer growth by stimulating cancer-associated fibroblasts. *J Clin Invest* 2012; **122**: 3603-3617 [PMID: [22945634](https://pubmed.ncbi.nlm.nih.gov/22945634/) DOI: [10.1172/JCI62229](https://doi.org/10.1172/JCI62229)]
- 21 **Chu CS**, Kim SH, June CH, Coukos G. Immunotherapy opportunities in ovarian cancer. *Expert Rev Anticancer Ther* 2008; **8**: 243-257 [PMID: [18279065](https://pubmed.ncbi.nlm.nih.gov/18279065/) DOI: [10.1586/14737140.8.2.243](https://doi.org/10.1586/14737140.8.2.243)]
- 22 **Nelson BH**. The impact of T-cell immunity on ovarian cancer outcomes. *Immunol Rev* 2008; **222**: 101-116 [PMID: [18363996](https://pubmed.ncbi.nlm.nih.gov/18363996/) DOI: [10.1111/j.1600-065X.2008.00614.x](https://doi.org/10.1111/j.1600-065X.2008.00614.x)]
- 23 **Hansen JM**, Coleman RL, Sood AK. Targeting the tumour microenvironment in ovarian cancer. *Eur J Cancer* 2016; **56**: 131-143 [PMID: [26849037](https://pubmed.ncbi.nlm.nih.gov/26849037/) DOI: [10.1016/j.ejca.2015.12.016](https://doi.org/10.1016/j.ejca.2015.12.016)]
- 24 **Rancourt C**, Bergeron C, Lane D, Garon G, Piché A. Delivery of herpes simplex thymidine kinase bystander effect by engineered human mesothelial cells for the treatment of ovarian cancer. *Cytotherapy* 2003; **5**: 509-522 [PMID: [14660047](https://pubmed.ncbi.nlm.nih.gov/14660047/) DOI: [10.1080/14653240310003620](https://doi.org/10.1080/14653240310003620)]
- 25 **Yeung TL**, Leung CS, Yip KP, Au Yeung CL, Wong ST, Mok SC. Cellular and molecular processes in ovarian cancer metastasis. A Review in the Theme: Cell and Molecular Processes in Cancer Metastasis. *Am J Physiol Cell Physiol* 2015; **309**: C444-C456 [PMID: [26224579](https://pubmed.ncbi.nlm.nih.gov/26224579/) DOI: [10.1152/ajpcell.00188.2015](https://doi.org/10.1152/ajpcell.00188.2015)]
- 26 **Shield K**, Ackland ML, Ahmed N, Rice GE. Multicellular spheroids in ovarian cancer

- metastases: Biology and pathology. *Gynecol Oncol* 2009; **113**: 143-148 [PMID: [19135710](#) DOI: [10.1016/j.ygyno.2008.11.032](#)]
- 27 **Matte I**, Legault CM, Garde-Granger P, Laplante C, Bessette P, Rancourt C, Piché A. Mesothelial cells interact with tumor cells for the formation of ovarian cancer multicellular spheroids in peritoneal effusions. *Clin Exp Metastasis* 2016; **33**: 839-852 [PMID: [27612856](#) DOI: [10.1007/s10585-016-9821-y](#)]
- 28 **Yin M**, Li X, Tan S, Zhou HJ, Ji W, Bellone S, Xu X, Zhang H, Santin AD, Lou G. Tumor-associated macrophages drive spheroid formation during early transcoelomic metastasis of ovarian cancer. *J Clin Invest* 2016; **126**: 4157-4173 [PMID: [27721235](#) DOI: [10.1172/JCI87252](#)]
- 29 **Giannakouros P**, Comamala M, Matte I, Rancourt C, Piché A. MUC16 mucin (CA125) regulates the formation of multicellular aggregates by altering β -catenin signaling. *Am J Cancer Res* 2014; **5**: 219-230 [PMID: [25628932](#)]
- 30 **Nakamura K**, Sawada K, Kinose Y, Yoshimura A, Toda A, Nakatsuka E, Hashimoto K, Mabuchi S, Morishige KI, Kurachi H. Exosomes Promote Ovarian Cancer Cell Invasion through Transfer of CD44 to Peritoneal Mesothelial Cells. *Mol Cancer Res* 2017; **15**: 78-92 [PMID: [27758876](#) DOI: [10.1158/1541-7786.MCR-16-0191](#)]
- 31 **Liao J**, Qian F, Tchabo N, Mhawech-Fauceglia P, Beck A, Qian Z, Wang X, Huss WJ, Lele SB, Morrison CD. Ovarian cancer spheroid cells with stem cell-like properties contribute to tumor generation, metastasis and chemotherapy resistance through hypoxia-resistant metabolism. *PLoS One* 2014; **9**: e84941 [PMID: [24409314](#) DOI: [10.1371/journal.pone.0084941](#)]
- 32 **Benton G**, DeGray G, Kleinman HK, George J, Arnaoutova I. In vitro microtumors provide a physiologically predictive tool for breast cancer therapeutic screening. *PLoS One* 2015; **10**: e0123312 [PMID: [25856378](#) DOI: [10.1371/journal.pone.0123312](#)]
- 33 **L'Espérance S**, Bachvarova M, Tetu B, Mes-Masson AM, Bachvarov D. Global gene expression analysis of early response to chemotherapy treatment in ovarian cancer spheroids. *BMC Genomics* 2008; **9**: 99 [PMID: [18302766](#) DOI: [10.1186/1471-2164-9-99](#)]
- 34 **Ovaskainen O**, Tikhonov G, Norberg A, Guillaume Blanchet F, Duan L, Dunson D, Roslin T, Abrego N. How to make more out of community data? A conceptual framework and its implementation as models and software. *Ecol Lett* 2017; **20**: 561-576 [PMID: [28317296](#) DOI: [10.1111/ele.12757](#)]

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Current status of PI3K-mTOR inhibition in hormone-receptor positive, HER2-negative breast cancer

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Abstract

Breast cancer (BC) is the most common cancer in women and second only to lung cancer in terms of mortality. Among the three different BC subtypes, the oestrogen receptor positive represents nearly 70% of all cases and it is usually treated with anti-oestrogen drugs. However, the majority of hormone receptor positive metastatic BC patients develop resistance to anti-oestrogen treatments. The need for more down-stream therapies brought to the development of therapeutic strategies inhibiting the phosphatidylinositol 3-kinase-mammalian target of rapamycin (mTOR) pathway. Inhibitors of the mTOR have been tested in different clinical trials; everolimus has been Food and Drug Administration approved for the treatment of oestrogen receptor positive/human epidermal growth factor receptor 2 negative BC patients in combination with exemestane in patients who have progressed to anastrozole or letrozole after the encouraging results coming from BOLERO-2 trial. Similar results were obtained by the TAMRAD investigatory study testing tamoxifen in combination with everolimus in advanced BC. This editorial focuses on the results from BOLERO-2, BOLERO 4 and BOLERO-6, which tested the clinical importance of mTOR inhibition. We comment also on the role of phosphatidylinositol 3-kinase-mTOR inhibition as reported in the BELLE-2 and BELLE-3 trials and the future directions for the inhibition of this tumour metabolic axis.

Key words: Hormone receptor positive/Her2-negative breast cancer; PI3K; mTOR; TORC1/2; Akt; Everolimus

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Core tip: The phosphatidylinositol 3-kinase-mammalian target of rapamycin (PI3K-mTOR) pathway sustains cancer progression and drug resistance. The first Food and Drug Administration approved molecule against this pathway was everolimus, an mTOR inhibitor, to be used in combination with exemestane in hormone receptor positive/human epidermal growth factor receptor 2 negative breast cancers progressing to non-steroidal aromatase inhibitors. Drugs targeting other effectors such as PI3K, PI3K α , Akt and mTORC1/2 have gained clinical interest. Nevertheless, everolimus

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remains the best option due to the relevant toxicity of the other drugs targeting the PI3K-mTOR pathway. Future directions point towards the development of biomarkers that would identify those patients who would benefit from the PI3K-mTOR inhibitors for improving overall survival.

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INTRODUCTION

With the introduction of molecular technologies, breast cancer (BC) has been divided into four main subtypes: luminal A, luminal B, human epidermal growth factor receptor 2 (HER2), or triple negative. The luminal variants, due to the hormone receptor positive (HR+) expression, have a generally favourable prognosis in the non-metastatic setting. Some of the most commonly used drugs for the treatment of HR+ disease have been anti-oestrogen therapies, *e.g.* tamoxifen and aromatase inhibitors (AI) - such as anastrozole or letrozole or exemestane. In BC survivors, metastatic breast cancer (MBC) is a treatable but still incurable disease. The majority of HR+ MBCs develop resistance to anti-oestrogen treatments^[1,2]. Cancer driver mutations and/or constitutive activation in the key signalling pathway genes of phosphatidylinositol 3-kinase-mammalian target of rapamycin (PI3K-mTOR) cascade are capable of overriding anti-oestrogen treatments in BC cells^[3]. In particular, *PIK3CA* is mutated in 20%-40% of BC^[4,5], representing therefore an interesting target for therapies. For these reasons, inhibitors of the PI3K-mTOR pathway was pursued in clinical development until one of them, an mTOR inhibitor-a derivative of rapamycin - named everolimus, reached Food and Drug Administration (FDA) approval for the treatment of the disease in 2012, in combination with exemestane, for second line treatment of HR+/HER2- BCs that have previously regressed to anastrozole or letrozole treatment.

This commentary is focused on the outcomes coming from the most important clinical trials, testing everolimus in combination with other drugs: TAMRAD (Tamoxifen Plus Everolimus), BOLERO-2 (OraL Everolimus-2), BOLERO-4, BOLERO-6, BELLE-2 and BELLE-3. Future directions and perspectives testing therapeutical combinations targeting different effectors are also discussed.

PI3K-mTOR PATHWAY

PI3K-mTOR signaling events play an important role in the modulation of cell survival, proliferation, motility and apoptosis: activation of PI3K phosphorylates Akt, shifting thereby the protein in the activated form that localizes in the plasma membrane. Subsequently a cascade of downstream events are activated, including the inhibition of p27 and the increase of the transcription factor CREB levels and, consequently, of the cell cycle promoter cyclin A, leading to mTOR activation that promotes the expression of genes related to cell proliferation. The aberrant activation of the PI3K-mTOR pathway sustains cancer cell survival and proliferation and also contributes to anti-oestrogen treatment resistance.

PIK3CA mutations constitutively activate the PI3K signal transduction pathway, which ultimately leads to the phosphorylation of PIP₂ into PIP₃, thus activating the phosphatidylinositol signalling bringing to higher cell survival and to inhibition of apoptosis. Predominantly mutations of *PIK3CA* occur around hotspots E542/5 (exon 9) and H1047 (exon 20) at the catalytic subunit^[6-9]. Additionally, *AKT* or *PTEN* mutations overrides anti-oestrogen therapies^[4,10]. Eventually, PIP₃ phosphorylates Akt (pAkt is the active form), which then activates the mTOR.

Of note, PI3K activation has been shown to cause a decrease in ER levels and therefore a lesser degree of response to anti-oestrogen therapies^[11]. Targeting PI3K could be capable of restoring ER sensitivity^[12]. Therefore, PI3K-mTOR pathway inhibition plus endocrine therapies represent an interesting strategy to pursue. There is a link between ER and mTORC1: mTORC1 can phosphorylate ER at S167 through

p70S6K or it can also phosphorylate ER at S104/S106, thus resulting in ER activation^[13]. Of note, recently, Sonnenblick *et al*^[14] discovered a gene signature related to the phosphorylation of Akt (pAkt) and mTOR (p-mTOR) in early ER+ BC. Analyzing gene expression and proteomic data collected from over 7000 early BC, they found pAkt correlated to luminal A and a good prognosis, whereas p-mTOR was found in worse prognosis luminal B cases. Interestingly, the signatures of pAkt and p-mTOR correlated to *PI3KCA* mutations positively and negatively, respectively. The authors underlined the different role at the molecular level of pAkt and p-mTOR effectors in early luminal BC, a matter that further contributes to the heterogeneity of the disease^[14].

CLINICAL TRIALS

The discovery of everolimus led to the hypothesis of synergism in co-targeting two different pathways in HR+/HER2- advanced BC patients: the mTOR and the HRs pathways. Such hypothesis has been tested in several clinical trials.

The TAMRAD trial was a phase 2, randomized clinical trial that tested the combination of everolimus with tamoxifen in postmenopausal women with MBC and resistance to AI. Besides proving to be a safe approach, it demonstrated that the combination of these treatments was capable of increasing the clinical benefit rate (CBR) from 42% with tamoxifen to 61% with the combination. The time to progression increased from 4.5 mo in patients receiving only tamoxifen to 8.6 mo in patients receiving both tamoxifen and everolimus. The death risk was reduced by 55% after treating patients with tamoxifen plus everolimus in comparison to tamoxifen alone^[15]. Interestingly, the combination was efficient in patients who responded to AIs but subsequently became resistant to AIs. Contrarily, the combination was not effective in *de novo* treated MBC HR+^[16].

The BOLERO-2 trial was a phase 3, randomized, double-blinded, multicentre clinical trial that recruited 724 HR+ MBC patients. This trial evaluated everolimus with the steroidal AI as exemestane compared to exemestane and placebo. Patients who were enrolled in this clinical trial had previously received AI anastrozole or letrozole. This clinical trial showed an improved progression free survival (PFS) of 11 mo in the exemestane + everolimus arm compared to the 4.1 mo placebo + exemestane ones [$P < 0.0001$; hazard ratio = 0.38; 95% confidence interval (CI): 0.31-0.48]^[17]. This shows that the combination of everolimus with exemestane doubled up the PFS time versus placebo plus exemestane alone; these results led to the FDA approval of everolimus with exemestane in the treatment of HR+ MBC patients who did not respond to letrozole nor anastrozole treatments.

Recent studies further tested everolimus in combination with other therapies as first line in HR+ BC patients. The BOLERO-4 (NCT01698918) and BOLERO-6 (NCT01783444) tested everolimus with different combinations of drugs. BOLERO-4 was a single arm, open labeled phase 2 study evaluating in 202 ER+, HER2- advanced BC patients the PFS of first line everolimus + letrozole and in a quarter of them whose disease progressed, everolimus + exemestane as second line. This study demonstrated the efficacy of everolimus + letrozole with a median PFS of 22 mo, whereas the second line everolimus + exemestane, which was used in some of the progressed patients, gave limited results with a median PFS of only 3.7 mo^[18]. The BOLERO-6 was a randomized open-labeled study, comparing second line therapies everolimus + exemestane against everolimus or capecitabine alone in 309 AI-resistant ER+, HER2-MBC patients. Consistent with the BOLERO-2 study, robust evidence of superiority in median PFS was registered for everolimus + exemestane versus everolimus alone but surprisingly not versus capecitabine alone. However, the authors pointed out that there were some limits to the study, such as censored data and baseline imbalance in Kaplan Meyer curves, that could have interfered on the evaluation of the capecitabine arm^[19]. Thus, the combination of everolimus + exemestane was confirmed to benefit these patients, whereas the superiority/equivalence/inferiority of capecitabine needs further clinical evaluation.

Inhibition of PI3K has been also investigated as an alternative strategy in targeted therapies of the mTOR pathway. As an example, buparlisib (BKM120), which is a PI3K inhibitor, has been tested in combination with fulvestrant in patients progressing AIs (BELLE-2, NCT01610284) and in patients progressing mTOR inhibition (BELLE-3, NCT01633060).

Buparlisib has been tested in 1147 patients as second line in the randomized, double-blinded BELLE-2 clinical trial, alone or in combination with fulvestrant. It was clear that the combination with the PI3K inhibitor had slightly increased PFS compared to fulvestrant alone, with the exception of *PI3KCA* mutated patients who

improved the PFS of about 4 mo. Of note, the serious toxicity of buparlisib limited the efficacy of the targeted therapy^[20]. BELLE-3 was a randomized double-blinded phase 2 trial enrolling 432 patients who progressed AIs to evaluate PFS in buparlisib + fulvestrant arm compared to fulvestrant + placebo. The combination with buparlisib improved the PFS of only 2 mo while showing a high toxicity profile that hampered the clinical benefit^[21]. Finally, an exploratory strategy is aiming for the simultaneous inhibition of PI3K and mTOR by alpelisib (BYL719) and everolimus, respectively. The safety of the combination of alpelisib and everolimus alone or everolimus + exemestane has been tested in a Phase 1 trial in various solid tumours, including BC (NCT02077933).

Worth of note is the combination of letrozole and alpelisib, a selective oral inhibitor of the class I PI3K catalytic subunit p110 α , which has shown synergistic antitumour activity with endocrine therapy against ER+/PIK3CA-mutated BC cells. The combination was safe with reversible toxicities. Clinical activity was observed independently of PIK3CA mutation status, although clinical benefit was seen in a higher proportion of patients with PIK3CA-mutated tumours. Phase 2 and 3 trials of alpelisib and endocrine therapy in patients with ER+ BC are ongoing^[22].

PERSPECTIVE

In addition to the previous therapies targeting mTOR, other research groups sought to investigate alternative strategies targeting molecules belonging to the mTOR complex or other effectors of the PI3K-mTOR pathway. An investigatory strategy consists in the use of sapanisertib (TAK228) for TORC1/2 inhibition by targeting mTOR kinase. TORC1/2 are complexes of proteins that regulate many vital cellular processes and proliferation. This drug has been tested in the neoadjuvant setting with tamoxifen (NCT02988986) or letrozole (NCT02619669) in the treatment of HR+ BC patients. AZD5363 is a specific Akt inhibitor, which has been tested either as a monotherapy (NCT01226316) or as a combination with fulvestrant in HR+ MBC patients who progressed AIs (NCT01992952). **Table 1** summarizes all the clinical trials testing PI3K-mTOR pathway inhibitors. In our opinion, there is a need for predictors of the efficacy of such inhibitors: a pooled analysis for overall survival, disease free survival and PFS meta-analysis using clinical studies investigating the prognostic role of PI3KCA mutations in BC, proved that mutations of this gene are predictive of a worse clinical survival outcome for the patients (HR = 1.67, 95%CI: 1.15-2.43; $P = 0.007$)^[23]. Unfortunately, there was not enough clinical evidence to prove that PI3KCA mutations could be predictive for therapy-efficacy and further clinical trials investigating the role of such mutations could be precious for guiding clinicians to choose those patients who would benefit from PI3K-mTOR inhibition treatment.

In conclusion, the inhibition of the PI3K-mTOR pathway remains one of the major goals for the treatment of HR+ advanced BC patients. It is known that targeting more upstream effectors, such as PI3K, which proved to be very promising based on *in vitro* evidence, lead to major side effects in clinical studies, limiting the clinical development of this type of drugs. This class of inhibitors, however, seems to be beneficial for patients harboring PI3KCA mutations, thus producing less toxicity.

Currently, everolimus remains the best option in the clinical setting in combinational therapies for HR+ BC patients. Moreover, several studies are directed to explore the clinical benefits of everolimus in combination with other drugs, as illustrated in **Table 1**. Of note, 10%-15% of patients are intrinsically resistant to everolimus treatment^[24]. For this reason, dual inhibition of the pathway by targeting mTORC1/2 proteins have been tested (**Table 1**), and some phase 1 studies are exploring new PI3K inhibitors, such as gedatolisib (NCT02626507) in neoadjuvant setting for ER+, HER2- BC patients. CUDC-907, an histone deacetylase and PI3K inhibitor (NCT02307240), and AZD8186, a PI3K- β inhibitor (NCT03218826), have been tested in advanced solid tumours, including BC. Having biomarkers from the metastatic setting to predict the responsiveness/resistance to anti-PI3K/mTOR/Akt drugs are thus an urgent task, and many efforts in this direction are expected. Finally, with the advancement of immune-therapy, trials testing the combination of immune-therapy with everolimus are warranted.

Table 1 Clinical trials inhibiting PI3K-mTOR pathway in HR+, HER2- breast cancer patients

Clinical trial identifier	Phase	Year	Administered drug(s) (target)	Primary endpoint	Target/s of the PI3K-mTOR pathway
NCT00699491	1/2	18.06.2008	Cixutumumab + temsirolimus	ORR	mTOR
Bolero 2; NCT00863655 [16]	3	18.03.2009	Everolimus + exemestane or placebo + exemestane	PFS	mTOR
NCT00876395	3	06.04.2009	Everolimus + placebo	PFS	mTOR
NCT01219699	1	13.10.2010	BYL719 + fulvestrant	MTD	PI3K α
NCT01283789	2	26.01.2011	Everolimus + lapatinib	ORR	mTOR
TAMRAD; NCT01298713 [14]	2	18.02.2011	Everolimus + tamoxifen	CBR	mTOR
BELLE-2; NCT01610284	3	04.06.2012	Fulvestrant \pm BKM120 (AIs refractory pts)	PFS	PI3K
BELLE-3; NCT01633060	3	04.06.2012	Fulvestrant \pm BKM120 (Pts previously on mTOR inhibitors)	PFS	PI3K
NCT01627067	2	25.06.2012	Everolimus + exemestane + metformin	PFS	mTOR
NCT01674140	3	28.08.2012	Everolimus + standard adjuvant endocrine therapy (tamoxifen citrate, goserelin acetate, leuprolide acetate, anastrozole, letrozole, or exemestane)	Invasive DFS	mTOR
Bolero 4; NCT01698918	2	03.10.2012	Everolimus + letrozole	PFS	mTOR
NCT01776008	2	25.01.2013	everolimus + exemestane; MK2206 + anastrozole \pm goserelin acetate	CRR	Akt
NCT01783444; Bolero-6	2	05.02.2013	Everolimus or capecitabine or everolimus + exemestane	PFS	mTOR
NCT01791478	1	15.02.2013	BYL719 + letrozole	DLT	PI3K α
NCT01805271	3	06.03.2013	Everolimus + placebo	DFS	mTOR
CLEE011X2107; NCT01872260	1b/2	07.06.2013	Letrozole + ribociclib + alpelisib	DLT	PI3K
INPRES; NCT01948960	4	24.09.2013	Everolimus	AUC correlation with age and/or obesity	mTOR
NCT01992952	1/2	25.11.2013	Fulvestrant \pm AZD5363 or placebo	MTD	Akt
PEARL; NCT02028364	2	07.01.2014	Everolimus + exemestane	PFS and response by PET	mTOR
NCT02035813	2	14.01.2014	Everolimus + eribulin	PFS	mTOR
NCT02049957	1b/2	30.01.2014	MLN0128 + exemestane vs MLN0128 + fulvestrant	Participant with AE and Clinical Benefit Rate	mTORC1/2
NCT02057133	1b	06.02.2014	LY2835219 + exemestane + everolimus vs exemestane + everolimus vs LY2835219 + trastuzumab vs LY2835219 + anastrozole vs LY2835219 + tamoxifen vs LY2835219 + letrozole vs LY3023414 + LY2835219 + fulvestrant	Safety	mTOR
NCT02058381	1b	10.02.2014	BYL719 + BKM120	MTD	PI3K α + PI3K
NCT02077933	1	04.03.2014	Alpelisib + everolimus + exemestane	DLT and MTD	PI3K + mTOR

NCT02123823	1b	28.04.2014	BI 836845 + everolimus + exemestane	PFS, MTD and DLT	mTOR
NCT02216786	2	15.08.2014	Everolimus + fulvestrant <i>vs</i> AZD2014 + fulvestrant	PFS	mTOR + mTORC1/2
NCT02236572	2	10.09.2014	Everolimus + aromatase inhibitor	Preoperative Endocrine Prognostic Index	mTOR
NCT02269670	2	21.10.2014	Everolimus + endocrine therapy (anastrozole, letrozole, tamoxifen citrate, fulvestrant or megestrol acetate)	PFS and ORR	mTOR
NCT02285179	1/2	16.11.2014	Taselisib + tamoxifen	MTD	PI3K
NCT02291913	2	17.11.2014	Everolimus + anti-estrogen therapy (exemestane, tamoxifen, fulvestrant, anastrozole, letrozole, toremifene)	PFS	mTOR
Sandpiper NCT02340221	3	16.01.2015	Taselisib + fulvestrant <i>vs</i> placebo + fulvestrant	PFS	PI3K
FEVEX; NCT02404051	3	31.03.2015	Everolimus + exemestane <i>vs</i> everolimus + exemestane (at progression fulvestrant)	PFS	mTOR
NCT02404844	2	01.04.2015	BKM120 + tamoxifen	PFS	PI3K
NCT02379247		04.05.2015	BYL719 + Nab-paclitaxel	DLT , ORR for Phase II	PI3K α
SOLAR-1; NCT02437318	3	07.05.2015	Fulvestrant \pm alpelisib (AIs refractory pts)	PFS	PI3K
NCT02506556	2	23.07.2015	BYI719	ORR	PI3K α
NCT02511639	3	30.07.2015	Everolimus + aromatase inhibitors	PFS	mTOR
NCT02520063	1/2	11.08.2015	Letrozole + everolimus + TRC105	MTD	mTOR
NCT02619669	1	02.12.2015	Sapanisertib + letrozole	TAE	mTORC1/2
TRINITI-1; NCT02732119	1/2	08.04.2016	Everolimus + ribociclib + exemestane	MTD	mTOR
NCT02742051	2	18.04.2016	Everolimus + letrozole <i>vs</i> neoadjuvant+ fluorouracil, epirubicin + cyclophosphamide (FEC)	ORR	mTOR
NCT02871791	2	18.08.2016	Everolimus + exemestane	DLT	mTOR
NCT02988986	2	12.12.2016	Sapanisertib + tamoxifen	Change in Ki67 expression	mTORC1/2
CLEVER; NCT03032406	2	26.01.2017	Everolimus \pm hydroxychloroquine	AE	mTOR
NCT03056755	2	17.02.2017	Alpelisib + fulvestrant + letrozole (AIs and CDK4/6 refractory pts)	DFS	PI3K
NCT03128619	1/2	25.04.2017	Copanlisib + letrozole + palbociclib	MTD	PI3K
EVEREXES; NCT03176238	3	05.06.2017	Everolimus + exemestane	AE	mTOR
NCT03207529	1	02.07.2017	BYL719 + enzalutamide	MTD	PI3K α
NCT03377101	2	19.12.2017	Fulvestrant + palbociclib \pm copanlisib	DLT	PI3K

DLT: Dose limiting toxicity; PFS: Progression free survival; DFS: Disease free survival; MTD: Maximum tolerated dose; AE: Adverse events; CBR: Clinical benefit rate; ORR: Overall response rate; CRR: Clinical response rate; PI3K-mTOR: Phosphatidylinositol 3-kinase-mammalian target of rapamycin.

REFERENCES

- 1 **Buzdar AU.** Phase III study of letrozole versus tamoxifen as first-line therapy of advanced breast cancer in postmenopausal women: analysis of survival and update of efficacy from the international letrozole breast cancer group. *J Clin Oncol* 2004; **22**: 3199-3200; author reply 3200-

- 3201 [PMID: 15284276 DOI: 10.1200/JCO.2004.99.058]
- 2 **Dalmau E**, Armengol-Alonso A, Muñoz M, Seguí-Palmer MÁ. Current status of hormone therapy in patients with hormone receptor positive (HR+) advanced breast cancer. *Breast* 2014; **23**: 710-720 [PMID: 25311296 DOI: 10.1016/j.breast.2014.09.006]
- 3 **Osborne CK**, Schiff R. Mechanisms of endocrine resistance in breast cancer. *Annu Rev Med* 2011; **62**: 233-247 [PMID: 20887199 DOI: 10.1146/annurev-med-070909-182917]
- 4 **Stemke-Hale K**, Gonzalez-Angulo AM, Lluch A, Neve RM, Kuo WL, Davies M, Carey M, Hu Z, Guan Y, Sahin A. An integrative genomic and proteomic analysis of PIK3CA, PTEN, and AKT mutations in breast cancer. *Cancer Res* 2008; **68**: 6084-6091 [PMID: 18676830 DOI: 10.1158/0008-5472.CAN-07-6854]
- 5 **Saal LH**, Holm K, Maurer M, Memeo L, Su T, Wang X, Yu JS, Malmström PO, Mansukhani M, Enoksson J. PIK3CA mutations correlate with hormone receptors, node metastasis, and ERBB2, and are mutually exclusive with PTEN loss in human breast carcinoma. *Cancer Res* 2005; **65**: 2554-2559 [PMID: 15805248 DOI: 10.1158/0008-5472.CAN-04-3913]
- 6 **Paplomata E**, O'Regan R. The PI3K/AKT/mTOR pathway in breast cancer: targets, trials and biomarkers. *Ther Adv Med Oncol* 2014; **6**: 154-166 [PMID: 25057302 DOI: 10.1177/1758834014530023]
- 7 **deGraffenried LA**, Friedrichs WE, Russell DH, Donzis EJ, Middleton AK, Silva JM, Roth RA, Hidalgo M. Inhibition of mTOR activity restores tamoxifen response in breast cancer cells with aberrant Akt Activity. *Clin Cancer Res* 2004; **10**: 8059-8067 [PMID: 15585641 DOI: 10.1158/1078-0432.CCR-04-0035]
- 8 **Bhattacharva GS**, Biswas J, Singh JK, Singh M, Govindbabu K, Ranade AA, Malhotra H, Parikh PM, Shahid T, Basu S. Reversal of Tamoxifen Resistance (Hormone Resistance) by Addition of Sirolimus (mTOR Inhibitor) in Metastatic Breast Cancer. *Eur J Cancer* 2011; **47**: 9 [DOI: 10.1016/S0959-8049(11)70115-0]
- 9 **Generali D**, Fox SB, Brizzi MP, Allevi G, Bonardi S, Aguggini S, Milani M, Bersiga A, Campo L, Dionisio R. Down-regulation of phosphatidylinositol 3'-kinase/AKT/molecular target of rapamycin metabolic pathway by primary letrozole-based therapy in human breast cancer. *Clin Cancer Res* 2008; **14**: 2673-2680 [PMID: 18451231 DOI: 10.1158/1078-0432.CCR-07-1046]
- 10 **Creighton CJ**, Fu X, Hennessy BT, Casa AJ, Zhang Y, Gonzalez-Angulo AM, Lluch A, Gray JW, Brown PH, Hilsenbeck SG. Proteomic and transcriptomic profiling reveals a link between the PI3K pathway and lower estrogen-receptor (ER) levels and activity in ER+ breast cancer. *Breast Cancer Res* 2010; **12**: R40 [PMID: 20569503 DOI: 10.1186/bcr2594]
- 11 **Fu X**, Osborne CK, Schiff R. Biology and therapeutic potential of PI3K signaling in ER+/HER2-negative breast cancer. *Breast* 2013; **22** Suppl 2: S12-S18 [PMID: 24011769 DOI: 10.1016/j.breast.2013.08.001]
- 12 **Corona SP**, Sobhani N, Ianza A, Roviello G, Mustacchi G, Bortul M, Zanconati F, Generali D. Advances in systemic therapy for metastatic breast cancer: future perspectives. *Med Oncol* 2017; **34**: 119 [PMID: 28526922 DOI: 10.1007/s12032-017-0975-5]
- 13 **Alayev A**, Salamon RS, Berger SM, Schwartz NS, Cuesta R, Snyder RB, Holz MK. mTORC1 directly phosphorylates and activates ERα upon estrogen stimulation. *Oncogene* 2016; **35**: 3535-3543 [PMID: 26522726 DOI: 10.1038/onc.2015.414]
- 14 **Sonnenblick A**, Brohée S, Pondé N, Venet D, Sotiriou C. Dissecting the Akt/mTOR pathway in estrogen receptor positive breast cancers identifies phosphorylated Akt signature to be associated with luminal A and good prognosis in contrast to phosphorylated mTOR. *The Breast* 2017; **32**: S103 [DOI: 10.1016/S0960-9776(17)30328-4]
- 15 **Bachelot T**, Bourcier C, Cropet C, Ray-Coquard I, Ferrero JM, Freyer G, Abadie-Lacourtoisie S, Eymard JC, Debled M, Spaëth D. Randomized phase II trial of everolimus in combination with tamoxifen in patients with hormone receptor-positive, human epidermal growth factor receptor 2-negative metastatic breast cancer with prior exposure to aromatase inhibitors: a GINECO study. *J Clin Oncol* 2012; **30**: 2718-2724 [PMID: 22565002 DOI: 10.1200/JCO.2011.39.0708]
- 16 **Tomii K**, Tsukuda K, Toyooka S, Dote H, Hanafusa T, Asano H, Naitou M, Doihara H, Kisimoto T, Katayama H. Aberrant promoter methylation of insulin-like growth factor binding protein-3 gene in human cancers. *Int J Cancer* 2007; **120**: 566-573 [PMID: 17096329 DOI: 10.1002/ijc.22341]
- 17 **Yardley DA**, Noguchi S, Pritchard KI, Burris HA 3rd, Baselga J, Gnant M, Hortobagyi GN, Campone M, Pistilli B, Piccart M, Melichar B, Petrakova K, Arena FP, Erdkamp F, Harb WA, Feng W, Cahana A, Taran T, Lebwohl D, Rugo HS. Everolimus plus exemestane in postmenopausal patients with HR(+) breast cancer: BOLERO-2 final progression-free survival analysis. *Adv Ther* 2013; **30**: 870-884 [PMID: 24158787 DOI: 10.1007/s12325-013-0060-1]
- 18 **Royce M**, Bachelot T, Villanueva C, Özgüroglu M, Azevedo SJ, Cruz FM, Debled M, Hegg R, Toyama T, Falkson C. Everolimus Plus Endocrine Therapy for Postmenopausal Women With Estrogen Receptor-Positive, Human Epidermal Growth Factor Receptor 2-Negative Advanced Breast Cancer: A Clinical Trial. *JAMA Oncol* 2018; **4**: 977-984 [PMID: 29566104 DOI: 10.1001/jamaoncol.2018.0060]
- 19 **Jerusalem G**, de Boer RH, Hurvitz S, Yardley DA, Kovalenko E, Ejlersen B, Blau S, Özgüroglu M, Landherr L, Ewertz M. Everolimus Plus Exemestane vs Everolimus or Capecitabine Monotherapy for Estrogen Receptor-Positive, HER2-Negative Advanced Breast Cancer: The BOLERO-6 Randomized Clinical Trial. *JAMA Oncol* 2018; **4**: 1367-1374 [PMID: 29862411 DOI: 10.1001/jamaoncol.2018.2262]
- 20 **Baselga J**, Im SA, Iwata H, Cortés J, De Laurentiis M, Jiang Z, Arteaga CL, Jonat W, Clemons M, Ito Y. Buparlisib plus fulvestrant versus placebo plus fulvestrant in postmenopausal, hormone receptor-positive, HER2-negative, advanced breast cancer (BELLE-2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol* 2017; **18**: 904-916 [PMID: 28576675 DOI: 10.1016/S1470-2045(17)30376-5]
- 21 **Di Leo A**, Johnston S, Lee KS, Ciruelos E, Lønning PE, Janni W, O'Regan R, Mouret-Reynier MA, Kaley D, Egle D. Buparlisib plus fulvestrant in postmenopausal women with hormone-receptor-positive, HER2-negative, advanced breast cancer progressing on or after mTOR inhibition (BELLE-3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol* 2018; **19**: 87-100 [PMID: 29223745 DOI: 10.1016/S1470-2045(17)30688-5]
- 22 **Mayer IA**, Abramson VG, Formisano L, Balko JM, Estrada MV, Sanders ME, Juric D, Solit D, Berger MF, Won HH. A Phase Ib Study of Alpelisib (BYL719), a PI3Kα-Specific Inhibitor, with

- Letrozole in ER+/HER2- Metastatic Breast Cancer. *Clin Cancer Res* 2017; **23**: 26-34 [PMID: 27126994 DOI: 10.1158/1078-0432.CCR-16-0134]
- 23 **Sobhani N**, Roviello G, Corona SP, Scaltriti M, Ianza A, Bortul M, Zanconati F, Generali D. The prognostic value of PI3K mutational status in breast cancer: A meta-analysis. *J Cell Biochem* 2018; **119**: 4287-4292 [PMID: 29345357 DOI: 10.1002/jcb.26687]
- 24 **Beaver JA**, Park BH. The BOLERO-2 trial: the addition of everolimus to exemestane in the treatment of postmenopausal hormone receptor-positive advanced breast cancer. *Future Oncol* 2012; **8**: 651-657 [PMID: 22764762 DOI: 10.2217/fon.12.49]

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Role of senescence induction in cancer treatment

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Abstract

Cellular senescence is a form of permanent cell cycle arrest that can be triggered by a variety of cell-intrinsic and extrinsic stimuli, including telomere shortening, DNA damage, oxidative stress, and exposure to chemotherapeutic agents and ionizing radiation. Although the induction of apoptotic cell death is a desirable outcome in cancer therapy, mutations and/or deficiencies in the apoptotic signaling pathways have been frequently identified in many human cancer types, suggesting the importance of alternative apoptosis-independent therapeutic approaches for cancer treatment. A growing body of evidence has documented that senescence induction in tumor cells is a frequent response to many anticancer modalities including cyclin-dependent kinases 4/6 small molecule inhibitor-based targeted therapeutics and T helper-1 cytokine-mediated immunotherapy. This review discusses the recent advances and clinical relevance of therapy-induced senescence in cancer treatment.

Key words: Cellular senescence; Cancer treatment; Chemotherapy; Ionizing radiation; Cyclin-dependent kinases 4/6 inhibitor; Aurora kinase inhibitor; Immunotherapy; T helper-1 cells; T helper-1 cytokines

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Core tip: Both *in vitro* and *in vivo* studies have revealed that senescence induction in human cancer cells is a prominent response to chemotherapy and irradiation. A senescent phenotype has been detected in clinical tumor samples of breast cancer patients following preoperative neoadjuvant chemotherapy. Immunotherapy-induced senescence of cancer cells contributes to tumor regression *in vivo*. The induction of cancer cell senescence appears to be a major mechanism of action of cyclin-dependent kinases 4/6 small molecule inhibitor-based targeted therapy. Collectively, these preclinical and clinical observations have demonstrated an important role for senescence induction in cancer treatment.

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INTRODUCTION

Cellular senescence is an anti-proliferative program that restrains tumorigenesis *via* limiting the propagation and transformation of aging and damaged cells^[1,2]. Senescence can be induced by a variety of cell-intrinsic and extrinsic stimuli, including telomere shortening, DNA damage, oxidative stress, chemotherapeutic agent treatment, radiation exposure, and the activation of oncogenes such as *Ras*^[3-6]. Senescent cells are in a state of irreversible cell cycle arrest and are characterized by a flat and enlarged morphology, elevated senescence-associated β -galactosidase activity, and the activation of the p53-p21 and p16-Rb signaling pathways^[2,7]. Additional characteristics of senescence cells include the presence of senescence-associated heterochromatic foci and the senescence-associated secretory phenotype^[8,9]. Traditionally, senescence induction is considered to be an important mechanism of cancer prevention and cellular aging^[10]. However, numerous recent studies have revealed that senescence is a prominent solid tumor response to therapy in which cancer cells evade apoptosis and instead enter into a stable and prolonged cell cycle arrest^[3,5]. Furthermore, targeted therapeutics and cancer immunotherapies also have been shown to cause cancer regression *via* the induction of senescence in tumor cells^[11,12]. These findings underscore the significant implications of senescence induction in cancer treatment.

INDUCTION OF TUMOR CELL SENESENCE IS AN IMPORTANT OUTCOME OF CHEMOTHERAPY

Cellular senescence is a state of permanent cell growth arrest that often has been included with apoptosis as one of the terminal outcomes of cancer treatment. It is well-established that different classes of chemotherapeutic agents and ionizing radiation (IR) induce senescent-like phenotypes in tumor cells both *in vitro* and *in vivo*^[2,5,11,13]. Our recent studies indicate that resveratrol induces premature senescence in lung cancer cells *via* promoting reactive oxygen species (ROS)-mediated DNA damage^[14]. Inactivation of Myc results in tumor regression through induction of senescence but not apoptosis in hepatocellular carcinoma and lymphoma cells^[15]. In addition, it has been reported that the restoration of p53 promotes tumor regression *in vivo* *via* induction of senescence in tumor cells^[16,17]. More importantly, Schmitt *et al*^[13] showed that senescence induction contributes directly to the outcome of chemotherapy *in vivo*. In agreement with this finding, there is evidence that the senescent phenotype is present in clinical tumor samples of breast cancer patients after undergoing preoperative neoadjuvant chemotherapy^[18]. Moreover, our recent studies found that the number of senescent cells is markedly increased in tumor samples of lung cancer patients as compared to normal lung tissues (Figure 1). However, the clinical relevance of the presence of senescent cells in tumor samples has yet to be determined.

Notably, human tumors may harbor various types of defects in the apoptotic signaling pathways (*e.g.*, loss of p53 and overexpression of BCL2) and, as a result, are resistant to apoptosis-based anticancer therapies^[19,20]. In these conditions, a senescence-targeted strategy is likely to be a more effective and practical than traditional apoptosis-inducing approaches. A welcome benefit to this approach is that therapy-induced senescence can be achieved using much lower doses of therapy than those required to induce apoptosis^[2,14]. Compared to the traditional apoptosis-inducing strategies, this low dose approach would significantly reduce the side effects of anticancer therapy and thus improve the quality of life for cancer patients.

ROLE OF SENESENCE INDUCTION IN CANCER RADIOTHERAPY

Radiotherapy is used in over 50% of patients during the course of cancer treatment and is effective both as a curative modality and for palliation^[21]. However, many epithelial-derived tumors including lung cancer have been shown to be resistant to

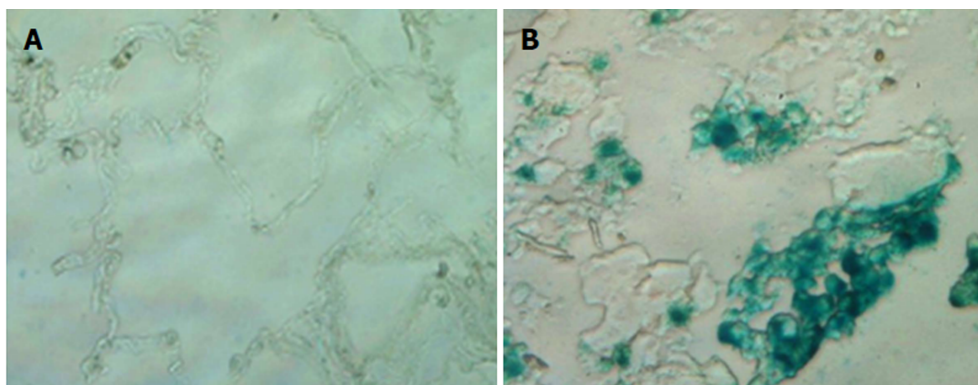


Figure 1 Number of senescent cells is significantly higher in tumor tissues of lung cancer patients than in normal lung tissues. Senescence-associated β -galactosidase staining was performed to analyze senescent cells in normal human lung tissues (A) and tumor tissues of lung cancer patients (B), respectively.

radiation-induced apoptosis^[6,20]. Consistent with these observations, our recent studies have demonstrated that IR primarily induces premature senescence rather than apoptosis in human non-small cell lung cancer (NSCLC) cells^[6]. Subsequent mechanistic studies revealed that the p53-p21 signaling pathway plays a critical role in modulating IR-induced senescence in NSCLC cells (Figure 2). More importantly, we showed that pharmacologic activation of p53 by Nutlin-3 sensitizes NSCLC cells to radiation by enhancing IR-induced senescence^[6]. These results suggest that pharmacological promotion of senescence induction can be exploited as a novel and effective therapeutic approach to improve the efficacy of lung cancer radiotherapy.

MicroRNA-34a (miR-34a) has been shown to be a p53 responsive miRNA that is involved in regulating senescence induction^[22-24]. Our recent studies showed that the expression of miRNA-34a was increased substantially in human NSCLC cells after exposure to IR^[25]. Moreover, we found that treatment with synthetic miR-34a mimics enhances the anti-cancer effects of irradiation by promoting senescence induction *via* targeting the Myc oncoprotein in NSCLC cells (Figure 2). These findings not only provide new insights into the mechanisms by which IR induces senescence in lung cancer cells but also support the hypothesis that pharmacological manipulation of senescence induction should be explored as a new therapeutic strategy for improving the outcome of cancer radiotherapy.

INDUCTION OF SENESCENCE IS A MAJOR MECHANISM OF ACTION OF CYCLIN-DEPENDENT KINASES 4/6 INHIBITION-BASED TARGETED THERAPEUTICS

The cyclin-dependent kinases (CDKs) are a large family of serine-threonine kinases that play a pivotal role in regulating cell cycle progression. Commitment to cell cycle entry occurs during the G1 phase, when CDK4 and CDK6 form active complexes with one of the three D-type cyclins (D1, D2, or D3). Cyclin D-CDK4/6 complexes promote G1/S transition by phosphorylating the Retinoblastoma tumor suppressor (Rb), which in turn releases its suppression of the E2F transcription factor, resulting in initiation of E2F-dependent gene transcription and DNA synthesis. CDK4 and CDK6 are overexpressed in the majority of human cancers and they presumably promote tumorigenesis by suppressing senescence in cancer cells^[11,26,27]. Both preclinical studies and clinic trials have demonstrated the therapeutic potential of CDK4/6 small molecule inhibitors against several solid tumors^[28-31]. Recently, a CDK4/6-specific inhibitor, palbociclib (also known as PD0332991), was approved by the FDA for the treatment of advanced estrogen receptor-positive breast cancer^[32].

It is well established that CDK4/6 inhibitors suppress tumor growth by the induction of senescence in various type of cancer cells^[11,26,27,32]. Mechanistic studies have revealed that CDK4/6 suppress tumor cell senescence *via* phosphorylating and activating the Forkhead Box M1 (FOXO1) transcription factor, and that activation of FOXO1 may inhibit the induction of senescence in tumor cells by repressing ROS-mediated oxidative stress^[33]. However, the precise mechanisms whereby FOXO1 controls ROS and oxidative stress remain unclear. Promyelocytic leukemia (PML) acts as a tumor suppressor by inducing senescence in response to oncogenic stress^[34,35]. Interestingly, Acevedo *et al.*^[36] showed that CDK4/6 suppress PML-induced senescence in tumor cells, suggesting that CDK4/6 inhibitors such as PD0332991 may

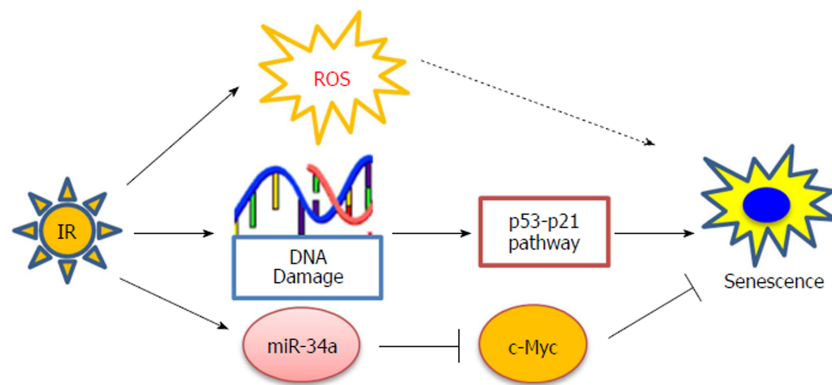


Figure 2 This model summarizes the mechanisms underlying ionizing radiation-induced senescence in non-small cell lung cancer cells. Ionizing radiation (IR) exposure increases the levels of reactive oxygen species in tumor cells and oxidative stress is known to trigger the induction of senescence. IR exposure leads to DNA damage which in turn can activate the p53-p21 senescence pathway. Irradiation up-regulates the expression of miR-34a that inhibits c-Myc, resulting in the induction of senescence. ROS: Reactive oxygen species; IR: Ionizing radiation.

induce senescence in cancer cells by blocking CDK4/6-mediated suppression of PML's senescence-inducing function. Moreover, CDK4/6 inhibition has been shown to be efficacious against therapy-resistant HER2 positive breast cancers^[37]. These results imply that although some tumors may be resistant to apoptosis-inducing treatments, it is likely that they are still responsive to senescence-targeted therapies.

AURORA KINASE INHIBITION INDUCES SENESCENCE IN TUMOR CELLS BOTH *IN VITRO* AND *IN VIVO*

Aurora kinases are a family of serine/threonine mitotic kinases that regulate a diverse set of mitotic processes including spindle formation, centrosome segregation, checkpoint activation and kinetochore-microtubule connections^[38,39]. Aurora A kinase (AurA) is located on 20q13.2, a chromosomal region that is frequently amplified in many human cancers. Overexpression of AurA correlates with poor clinical outcomes in patients with hormone-related cancers^[40]. Aurora kinase inhibitors (AKIs) are a promising class of drugs for cancer treatment. Several small molecule AKIs including MK-0457, PHA-739358 and MLN8237 have been investigated in clinical trials for the treatment of human cancers^[41-43].

Alisertib (MLN8237) is an orally bioavailable, second-generation selective inhibitor of Aurora kinases which binds to AurA and prevents its phosphorylation and activation^[44]. It has been shown that senescence is likely a terminal outcome of AurA inhibition and that pharmacological inhibition of AurA induces senescence in colon cancer cells both *in vitro* and *in vivo*^[45]. AurA is overexpressed in gliomas and treatment with Alisertib induces senescence and differentiation in glioblastoma cells^[46]. Moreover, recent studies have revealed that p53 and p73 tumor suppressors play a critical role in modulating tumor cell responses to AKIs. Tentler *et al*^[47] reported that Alisertib treatment resulted in apoptosis in human triple-negative breast cancer (TNBC) cells with functional p53 and p73, whereas it induced senescence in cells lacking p53 and p73. These findings suggest that AKI may induce growth arrest in TNBC through a p53- and p73-independent mechanism. Nevertheless, further studies are warranted to better understand the in-depth mechanisms whereby AKIs induce senescence in human tumor cells.

INDUCTION OF CANCER CELL SENESCENCE CONTRIBUTES TO IMMUNOTHERAPY-INDUCED TUMOR REGRESSION

Immunotherapy has shown promising efficacy against human cancers both in preclinical studies and in clinical trials^[48-50]. T helper-1 (Th1) cells play a critical role in mediating the antitumor adoptive immune response^[51,52]. Th1 T-cell infiltration is associated with better outcomes and Th1 cytokines are more effective at promoting the antitumor functions of CD40L-activated macrophages as compared to Th2 cytokines^[53]. Moreover, Müller-Hermelink *et al*^[54] showed that treatment with T

antigen (Tag)-specific Th1 cells induced tumor dormancy and doubled the survival of tumor-bearing mice by arresting tumor growth, without detectable evidence of tumor cell cytotoxicity, necrosis or apoptosis. These findings suggest that other alternative non-cytotoxic mechanisms are likely involved in Th1- and Th1 cytokine-mediated immunotherapy. In agreement with this idea, it has been shown that treatment with Th1 cytokines, IFN- γ and tumor necrosis factor (TNF) induces senescence in cancer cells^[12].

It has been reported that cancer immunotherapy causes tumor growth arrest and regression without any clear evidence of tumor cell death or cellular toxicity^[55,56]. Moreover, it has been shown that immunotherapy-induced tumor growth arrest was associated with an increase in interferon (IFN)- γ -producing CD4⁺ Th1 cells, but not CD8⁺ cytotoxic T lymphocytes^[55-57]. These observations suggest that senescence induction may contribute to the outcome of immunotherapy. In agreement with this, Braumüller *et al*^[12] showed that Th1 immunotherapy arrests tumor progression through IFN- γ - and TNF-induced cancer cell senescence *in vivo*. Their subsequent mechanistic studies revealed that TNFR1 signaling is required for Th1 immunotherapy-induced tumor cell senescence and that activation of the p16-Rb pathway is involved in senescence induction^[12]. In line with these findings, it has been shown recently that CDK4/6 inhibition-induced tumor cell senescence promotes antitumor immunity in preclinical models^[58,59]. Furthermore, there is evidence that IL-12 inhibits the growth of human sarcoma cells by senescence induction^[60]. Taken together, these results highlight a novel link between cancer immunotherapy and the induction of senescence in tumor cells.

CONCLUSION

Given the fact that many human cancers may harbor different defects in the apoptotic signaling pathways, and thus are inherently resistant to chemoradiotherapy-induced apoptosis, there is a critical need for the development of innovative apoptosis-independent approaches for cancer therapeutics. The induction of tumor cell senescence has been well-established as a prominent therapeutic response of cancer cells to chemotherapy, radiation, small-molecule inhibitor-based targeted therapeutics and immunotherapy. These results underscore the important implications of senescence induction in cancer treatment. Although senescent cells were detected in clinical tumor samples of cancer patients, the clinical significance and applications of therapy-induced senescence remain incompletely understood. It is still unclear if senescence markers in patient tumor samples will be useful for prognosis prediction and/or therapeutic efficacy evaluation in the clinic. In addition, further studies, particularly clinical investigations, are necessary to better elucidate the clinical relevance and significance of therapy-induced senescence in the treatment of cancer.

REFERENCES

- 1 Muñoz-Espín D, Serrano M. Cellular senescence: from physiology to pathology. *Nat Rev Mol Cell Biol* 2014; **15**: 482-496 [PMID: 24954210 DOI: 10.1038/nrm3823]
- 2 Ewald JA, Desotelle JA, Wilding G, Jarrard DF. Therapy-induced senescence in cancer. *J Natl Cancer Inst* 2010; **102**: 1536-1546 [PMID: 20858887 DOI: 10.1093/jnci/djq364]
- 3 He S, Sharpless NE. Senescence in Health and Disease. *Cell* 2017; **169**: 1000-1011 [PMID: 28575665 DOI: 10.1016/j.cell.2017.05.015]
- 4 Hydbring P, Bahram F, Su Y, Tronnorsjö S, Högstrand K, von der Lehr N, Sharifi HR, Lilischkis R, Hein N, Wu S. Phosphorylation by Cdk2 is required for Myc to repress Ras-induced senescence in cotransformation. *Proc Natl Acad Sci USA* 2010; **107**: 58-63 [PMID: 19966300 DOI: 10.1073/pnas.0900121106]
- 5 Chang BD, Swift ME, Shen M, Fang J, Broude EV, Roninson IB. Molecular determinants of terminal growth arrest induced in tumor cells by a chemotherapeutic agent. *Proc Natl Acad Sci USA* 2002; **99**: 389-394 [PMID: 11752408 DOI: 10.1073/pnas.012602599]
- 6 Luo H, Yount C, Lang H, Yang A, Riemer EC, Lyons K, Vanek KN, Silvestri GA, Schulte BA, Wang GY. Activation of p53 with Nutlin-3a radiosensitizes lung cancer cells via enhancing radiation-induced premature senescence. *Lung Cancer* 2013; **81**: 167-173 [PMID: 23683497 DOI: 10.1016/j.lungcan.2013.04.017]
- 7 Dimri GP, Lee X, Basile G, Acosta M, Scott G, Roskelley C, Medrano EE, Linskens M, Rubelj I, Pereira-Smith O. A biomarker that identifies senescent human cells in culture and in aging skin *in vivo*. *Proc Natl Acad Sci USA* 1995; **92**: 9363-9367 [PMID: 7568133 DOI: 10.1073/pnas.92.20.9363]
- 8 Narita M, Nunez S, Heard E, Narita M, Lin AW, Hearn SA, Spector DL, Hannon GJ, Lowe SW. Rb-mediated heterochromatin formation and silencing of E2F target genes during cellular senescence. *Cell* 2003; **113**: 703-716 [PMID: 12809602 DOI: 10.1016/S0092-8674(03)00401-X]
- 9 Tchkonian T, Zhu Y, van Deursen J, Campisi J, Kirkland JL. Cellular senescence and the senescent secretory phenotype: therapeutic opportunities. *J Clin Invest* 2013; **123**: 966-972 [PMID: 23454759 DOI: 10.1172/JCI64098]

- 10 **Braig M**, Lee S, Loddenkemper C, Rudolph C, Peters AH, Schlegelberger B, Stein H, Dörken B, Jenuwein T, Schmitt CA. Oncogene-induced senescence as an initial barrier in lymphoma development. *Nature* 2005; **436**: 660-665 [PMID: [16079837](#) DOI: [10.1038/nature03841](#)]
- 11 **Rader J**, Russell MR, Hart LS, Nakazawa MS, Belcastro LT, Martinez D, Li Y, Carpenter EL, Attiyeh EF, Diskin SJ. Dual CDK4/CDK6 inhibition induces cell-cycle arrest and senescence in neuroblastoma. *Clin Cancer Res* 2013; **19**: 6173-6182 [PMID: [24045179](#) DOI: [10.1158/1078-0432.CCR-13-1675](#)]
- 12 **Braumüller H**, Wieder T, Brenner E, Aßmann S, Hahn M, Alkhaled M, Schilbach K, Essmann F, Kneilling M, Griessinger C. T-helper-1-cell cytokines drive cancer into senescence. *Nature* 2013; **494**: 361-365 [PMID: [23376950](#) DOI: [10.1038/nature11824](#)]
- 13 **Schmitt CA**, Fridman JS, Yang M, Lee S, Baranov E, Hoffman RM, Lowe SW. A senescence program controlled by p53 and p16INK4a contributes to the outcome of cancer therapy. *Cell* 2002; **109**: 335-346 [PMID: [12015983](#) DOI: [10.1016/S0092-8674\(02\)00734-1](#)]
- 14 **Luo H**, Yang A, Schulte BA, Wargovich MJ, Wang GY. Resveratrol induces premature senescence in lung cancer cells via ROS-mediated DNA damage. *PLoS One* 2013; **8**: e60065 [PMID: [23533664](#) DOI: [10.1371/journal.pone.0060065](#)]
- 15 **Wu CH**, van Riggelen J, Yetil A, Fan AC, Bachireddy P, Felsher DW. Cellular senescence is an important mechanism of tumor regression upon c-Myc inactivation. *Proc Natl Acad Sci USA* 2007; **104**: 13028-13033 [PMID: [17664422](#) DOI: [10.1073/pnas.0701953104](#)]
- 16 **Xue W**, Zender L, Miething C, Dickins RA, Hernandez E, Krizhanovsky V, Cordon-Cardo C, Lowe SW. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature* 2007; **445**: 656-660 [PMID: [17251933](#) DOI: [10.1038/nature05529](#)]
- 17 **Ventura A**, Kirsch DG, McLaughlin ME, Tuveson DA, Grimm J, Lintault L, Newman J, Reczek EE, Weissleder R, Jacks T. Restoration of p53 function leads to tumour regression in vivo. *Nature* 2007; **445**: 661-665 [PMID: [17251932](#) DOI: [10.1038/nature05541](#)]
- 18 **te Poele RH**, Okorokov AL, Jardine L, Cummings J, Joel SP. DNA damage is able to induce senescence in tumor cells in vitro and in vivo. *Cancer Res* 2002; **62**: 1876-1883 [PMID: [11912168](#)]
- 19 **Mori S**, Ito G, Usami N, Yoshioka H, Ueda Y, Kodama Y, Takahashi M, Fong KM, Shimokata K, Sekido Y. p53 apoptotic pathway molecules are frequently and simultaneously altered in nonsmall cell lung carcinoma. *Cancer* 2004; **100**: 1673-1682 [PMID: [15073856](#) DOI: [10.1002/cncr.20164](#)]
- 20 **Yin DX**, Schimke RT. BCL-2 expression delays drug-induced apoptosis but does not increase clonogenic survival after drug treatment in HeLa cells. *Cancer Res* 1995; **55**: 4922-4928 [PMID: [7585531](#)]
- 21 **Ngiew SF**, McArthur GA, Smyth MJ. Radiotherapy complements immune checkpoint blockade. *Cancer Cell* 2015; **27**: 437-438 [PMID: [25873170](#) DOI: [10.1016/j.ccell.2015.03.015](#)]
- 22 **Hermeking H**. p53 enters the microRNA world. *Cancer Cell* 2007; **12**: 414-418 [PMID: [17996645](#) DOI: [10.1016/j.ccr.2007.10.028](#)]
- 23 **Tazawa H**, Tsuchiya N, Izumiya M, Nakagama H. Tumor-suppressive miR-34a induces senescence-like growth arrest through modulation of the E2F pathway in human colon cancer cells. *Proc Natl Acad Sci USA* 2007; **104**: 15472-15477 [PMID: [17875987](#) DOI: [10.1073/pnas.0707351104](#)]
- 24 **Wang Y**, Scheiber MN, Neumann C, Calin GA, Zhou D. MicroRNA regulation of ionizing radiation-induced premature senescence. *Int J Radiat Oncol Biol Phys* 2011; **81**: 839-848 [PMID: [21093163](#) DOI: [10.1016/j.ijrobp.2010.09.048](#)]
- 25 **He X**, Yang A, McDonald DG, Riemer EC, Vanek KN, Schulte BA, Wang GY. MiR-34a modulates ionizing radiation-induced senescence in lung cancer cells. *Oncotarget* 2017; **8**: 69797-69807 [PMID: [29050242](#) DOI: [10.18632/oncotarget.19267](#)]
- 26 **Yoshida A**, Lee EK, Diehl JA. Induction of Therapeutic Senescence in Vemurafenib-Resistant Melanoma by Extended Inhibition of CDK4/6. *Cancer Res* 2016; **76**: 2990-3002 [PMID: [26988987](#) DOI: [10.1158/0008-5472.CAN-15-2931](#)]
- 27 **Klein ME**, Dickson MA, Antonescu C, Qin LX, Dooley SJ, Barlas A, Manova K, Schwartz GK, Crago AM, Singer S. PDLIM7 and CDH18 regulate the turnover of MDM2 during CDK4/6 inhibitor therapy-induced senescence. *Oncogene* 2018; **37**: 5066-5078 [PMID: [29789718](#) DOI: [10.1038/s41388-018-0332-y](#)]
- 28 **Jansen VM**, Bhole NE, Bauer JA, Formisano L, Lee KM, Hutchinson KE, Witkiewicz AK, Moore PD, Estrada MV, Sánchez V. Kinome-Wide RNA Interference Screen Reveals a Role for PDK1 in Acquired Resistance to CDK4/6 Inhibition in ER-Positive Breast Cancer. *Cancer Res* 2017; **77**: 2488-2499 [PMID: [28249908](#) DOI: [10.1158/0008-5472.CAN-16-2653](#)]
- 29 **Finn RS**, Martin M, Rugo HS, Jones S, Im SA, Gelmon K, Harbeck N, Lipatov ON, Walshe JM, Moulder S. Palbociclib and Letrozole in Advanced Breast Cancer. *N Engl J Med* 2016; **375**: 1925-1936 [PMID: [27959613](#) DOI: [10.1056/NEJMoa1607303](#)]
- 30 **Finn RS**, Crown JP, Lang I, Boer K, Bondarenko IM, Kulyk SO, Ettl J, Patel R, Pinter T, Schmidt M. The cyclin-dependent kinase 4/6 inhibitor palbociclib in combination with letrozole versus letrozole alone as first-line treatment of oestrogen receptor-positive, HER2-negative, advanced breast cancer (PALOMA-1/TRIO-18): a randomised phase 2 study. *Lancet Oncol* 2015; **16**: 25-35 [PMID: [25524798](#) DOI: [10.1016/S1470-2045\(14\)71159-3](#)]
- 31 **Vijayaraghavan S**, Karakas C, Doostan I, Chen X, Bui T, Yi M, Raghavendra AS, Zhao Y, Bashour SI, Ibrahim NK. CDK4/6 and autophagy inhibitors synergistically induce senescence in Rb positive cytoplasmic cyclin E negative cancers. *Nat Commun* 2017; **8**: 15916 [PMID: [28653662](#) DOI: [10.1038/ncomms15916](#)]
- 32 **Sherr CJ**, Beach D, Shapiro GL. Targeting CDK4 and CDK6: From Discovery to Therapy. *Cancer Discov* 2016; **6**: 353-367 [PMID: [26658964](#) DOI: [10.1158/2159-8290.CD-15-0894](#)]
- 33 **Anders L**, Ke N, Hydbring P, Choi YJ, Widlund HR, Chick JM, Zhai H, Vidal M, Gygi SP, Braun P. A systematic screen for CDK4/6 substrates links FOXM1 phosphorylation to senescence suppression in cancer cells. *Cancer Cell* 2011; **20**: 620-634 [PMID: [22094256](#) DOI: [10.1016/j.ccr.2011.10.001](#)]
- 34 **Ferbeyre G**. PML a target of translocations in APL is a regulator of cellular senescence. *Leukemia* 2002; **16**: 1918-1926 [PMID: [12357343](#) DOI: [10.1038/sj.leu.2402722](#)]
- 35 **Pearson M**, Carbone R, Sebastiani C, Cioce M, Fagioli M, Saito S, Higashimoto Y, Appella E, Minucci S, Pandolfi PP. PML regulates p53 acetylation and premature senescence induced by

- oncogenic Ras. *Nature* 2000; **406**: 207-210 [PMID: [10910364](#) DOI: [10.1038/35018127](#)]
- 36 **Acevedo M**, Vernier M, Mignacca L, Lessard F, Huot G, Moiseeva O, Bourdeau V, Ferbeyre G. A CDK4/6-Dependent Epigenetic Mechanism Protects Cancer Cells from PML-induced Senescence. *Cancer Res* 2016; **76**: 3252-3264 [PMID: [27206849](#) DOI: [10.1158/0008-5472.CAN-15-2347](#)]
- 37 **Goel S**, Wang Q, Watt AC, Tolaney SM, Dillon DA, Li W, Ramm S, Palmer AC, Yuzugullu H, Varadan V. Overcoming Therapeutic Resistance in HER2-Positive Breast Cancers with CDK4/6 Inhibitors. *Cancer Cell* 2016; **29**: 255-269 [PMID: [26977878](#) DOI: [10.1016/j.ccell.2016.02.006](#)]
- 38 **Lioutas A**, Vernos I. Aurora A kinase and its substrate TACC3 are required for central spindle assembly. *EMBO Rep* 2013; **14**: 829-836 [PMID: [23887685](#) DOI: [10.1038/embor.2013.109](#)]
- 39 **Marumoto T**, Honda S, Hara T, Nitta M, Hirota T, Kohmura E, Saya H. Aurora-A kinase maintains the fidelity of early and late mitotic events in HeLa cells. *J Biol Chem* 2003; **278**: 51786-51795 [PMID: [14523000](#) DOI: [10.1074/jbc.M306275200](#)]
- 40 **Das K**, Lorena PD, Ng LK, Shen L, Lim D, Siow WY, Narasimhan K, Teh M, Choolani M, Putti TC. Aurora-A expression, hormone receptor status and clinical outcome in hormone related cancers. *Pathology* 2010; **42**: 540-546 [PMID: [20854072](#) DOI: [10.3109/000313025.2010.508789](#)]
- 41 **Falchook GS**, Bastida CC, Kurzrock R. Aurora Kinase Inhibitors in Oncology Clinical Trials: Current State of the Progress. *Semin Oncol* 2015; **42**: 832-848 [PMID: [26615129](#) DOI: [10.1053/j.seminoncol.2015.09.022](#)]
- 42 **Dickson MA**, Mahoney MR, Tap WD, D'Angelo SP, Keohan ML, Van Tine BA, Agulnik M, Horvath LE, Nair JS, Schwartz GK. Phase II study of MLN8237 (Alisertib) in advanced/metastatic sarcoma. *Ann Oncol* 2016; **27**: 1855-1860 [PMID: [27502708](#) DOI: [10.1093/annonc/mdw281](#)]
- 43 **Fathi AT**, Wander SA, Blonquist TM, Brunner AM, Amrein PC, Supko J, Hermance NM, Manning AL, Sadrzadeh H, Ballen KK. Phase I study of the aurora A kinase inhibitor alisertib with induction chemotherapy in patients with acute myeloid leukemia. *Haematologica* 2017; **102**: 719-727 [PMID: [28034990](#) DOI: [10.3324/haematol.2016.158394](#)]
- 44 **Manfredi MG**, Ecsedy JA, Meetze KA, Balani SK, Burenkova O, Chen W, Galvin KM, Hoar KM, Huck JJ, LeRoy PJ. Antitumor activity of MLN8054, an orally active small-molecule inhibitor of Aurora A kinase. *Proc Natl Acad Sci USA* 2007; **104**: 4106-4111 [PMID: [17360485](#) DOI: [10.1073/pnas.0608798104](#)]
- 45 **Huck JJ**, Zhang M, McDonald A, Bowman D, Hoar KM, Stringer B, Ecsedy J, Manfredi MG, Hyer ML. MLN8054, an inhibitor of Aurora A kinase, induces senescence in human tumor cells both in vitro and in vivo. *Mol Cancer Res* 2010; **8**: 373-384 [PMID: [20197380](#) DOI: [10.1158/1541-7786.MCR-09-0300](#)]
- 46 **Lehman NL**, O'Donnell JP, Whiteley LJ, Stapp RT, Lehman TD, Roszka KM, Schultz LR, Williams CJ, Mikkelsen T, Brown SL. Aurora A is differentially expressed in gliomas, is associated with patient survival in glioblastoma and is a potential chemotherapeutic target in gliomas. *Cell Cycle* 2012; **11**: 489-502 [PMID: [22274399](#) DOI: [10.4161/cc.11.3.18996](#)]
- 47 **Tentler JJ**, Ionkina AA, Tan AC, Newton TP, Pitts TM, Glogowska MJ, Kabos P, Sartorius CA, Sullivan KD, Espinosa JM. p53 Family Members Regulate Phenotypic Response to Aurora Kinase A Inhibition in Triple-Negative Breast Cancer. *Mol Cancer Ther* 2015; **14**: 1117-1129 [PMID: [25758253](#) DOI: [10.1158/1535-7163.MCT-14-0538-T](#)]
- 48 **Dobrzanski MJ**, Rewers-Felkins KA, Quinlin IS, Samad KA, Phillips CA, Robinson W, Dobrzanski DJ, Wright SE. Autologous MUC1-specific Th1 effector cell immunotherapy induces differential levels of systemic TReg cell subpopulations that result in increased ovarian cancer patient survival. *Clin Immunol* 2009; **133**: 333-352 [PMID: [19762283](#) DOI: [10.1016/j.clim.2009.08.007](#)]
- 49 **Nishino M**, Ramaiya NH, Hatabu H, Hodi FS. Monitoring immune-checkpoint blockade: response evaluation and biomarker development. *Nat Rev Clin Oncol* 2017; **14**: 655-668 [PMID: [28653677](#) DOI: [10.1038/nrclinonc.2017.88](#)]
- 50 **Rosembli C**, Datta J, Lowenfeld L, Xu S, Basu A, Kodumudi K, Wiener D, Czerniecki BJ. Oncodriver inhibition and CD4⁺ Th1 cytokines cooperate through Stat1 activation to induce tumor senescence and apoptosis in HER2⁺ and triple negative breast cancer: implications for combining immune and targeted therapies. *Oncotarget* 2018; **9**: 23058-23077 [PMID: [29796172](#) DOI: [10.18632/oncotarget.25208](#)]
- 51 **Datta J**, Xu S, Rosembli C, Smith JB, Cintolo JA, Powell DJ Jr, Czerniecki BJ. CD4(+) T-Helper Type 1 Cytokines and Trastuzumab Facilitate CD8(+) T-cell Targeting of HER2/neu-Expressing Cancers. *Cancer Immunol Res* 2015; **3**: 455-463 [PMID: [25791067](#) DOI: [10.1158/2326-6066.CIR-14-0208](#)]
- 52 **Thakur A**, Schalk D, Sarkar SH, Al-Khadimi Z, Sarkar FH, Lum LG. A Th1 cytokine-enriched microenvironment enhances tumor killing by activated T cells armed with bispecific antibodies and inhibits the development of myeloid-derived suppressor cells. *Cancer Immunol Immunother* 2012; **61**: 497-509 [PMID: [21971587](#) DOI: [10.1007/s00262-011-1116-1](#)]
- 53 **Luheshi N**, Davies G, Poon E, Wiggins K, McCourt M, Legg J. Th1 cytokines are more effective than Th2 cytokines at licensing anti-tumour functions in CD40-activated human macrophages in vitro. *Eur J Immunol* 2014; **44**: 162-172 [PMID: [24114634](#) DOI: [10.1002/eji.201343351](#)]
- 54 **Müller-Hermelink N**, Braumüller H, Pichler B, Wieder T, Mailhammer R, Schaak K, Ghoreschi K, Yazdi A, Haubner R, Sander CA. TNFR1 signaling and IFN-gamma signaling determine whether T cells induce tumor dormancy or promote multistage carcinogenesis. *Cancer Cell* 2008; **13**: 507-518 [PMID: [18538734](#) DOI: [10.1016/j.ccr.2008.04.001](#)]
- 55 **Kenter GG**, Welters MJ, Valentijn AR, Lowik MJ, Berends-van der Meer DM, Vloon AP, Essahsah F, Fathors LM, Offringa R, Drijfhout JW. Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. *N Engl J Med* 2009; **361**: 1838-1847 [PMID: [19890126](#) DOI: [10.1056/NEJMoa0810097](#)]
- 56 **Hodi FS**, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010; **363**: 711-723 [PMID: [20525992](#) DOI: [10.1056/NEJMoa1003466](#)]
- 57 **Hunder NN**, Wallen H, Cao J, Hendricks DW, Reilly JZ, Rodmyre R, Jungbluth A, Gnjatich S, Thompson JA, Yee C. Treatment of metastatic melanoma with autologous CD4⁺ T cells against NY-ESO-1. *N Engl J Med* 2008; **358**: 2698-2703 [PMID: [18565862](#) DOI: [10.1056/NEJMoa0800251](#)]

- 58 **Deng J**, Wang ES, Jenkins RW, Li S, Dries R, Yates K, Chhabra S, Huang W, Liu H, Aref AR. CDK4/6 Inhibition Augments Antitumor Immunity by Enhancing T-cell Activation. *Cancer Discov* 2018; **8**: 216-233 [PMID: [29101163](#) DOI: [10.1158/2159-8290.CD-17-0915](#)]
- 59 **Goel S**, DeCristo MJ, Watt AC, BrinJones H, Sceneay J, Li BB, Khan N, Ubellacker JM, Xie S, Metzger-Filho O. CDK4/6 inhibition triggers anti-tumour immunity. *Nature* 2017; **548**: 471-475 [PMID: [28813415](#) DOI: [10.1038/nature23465](#)]
- 60 **Schilbach K**, Alkhaled M, Welker C, Eckert F, Blank G, Ziegler H, Sterk M, Müller F, Sonntag K, Wieder T. Cancer-targeted IL-12 controls human rhabdomyosarcoma by senescence induction and myogenic differentiation. *Oncoimmunology* 2015; **4**: e1014760 [PMID: [26140238](#) DOI: [10.1080/2162402X.2015.1014760](#)]

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Basic Study

Combination immunotherapy with Survivin and luteinizing hormone-releasing hormone fusion protein in murine breast cancer model

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Abstract

AIM

To investigate the therapeutic potential of two recombinant proteins, Survivin and luteinizing hormone-releasing hormone (LHRH) fusion protein [LHRH(6leu)-LTB] for immunotherapy of breast cancer.

METHODS

Murine 4T-1 breast cancer model was used to evaluate the efficacy of recombinant proteins *in vivo*. Twenty four Balb/c mice were divided into 4 groups of 6 mice each. Recombinant Survivin and LHRH fusion protein, alone or in combination, were administered along with immunomodulator *Mycobacterium indicus pranii* (MIP) in Balb/c mice. Unimmunized or control group mice were administered with phosphate buffer saline. Each group was then challenged with syngeneic 4T-1 cells to induce the growth of breast tumor. Tumor growth was monitored to evaluate the efficacy of immune-response in preventing the growth of cancer cells.

RESULTS

Preventive immunization with 20 µg recombinant Survivin and MIP was effective in suppressing growth of 4T-1 mouse model of breast cancer ($P = 0.04$) but 50 µg dose was ineffective in suppressing tumor growth. However, combination of Survivin and LHRH fusion protein was more effective in suppressing tumor growth ($P = 0.02$) as well as metastasis *in vivo* in comparison to LHRH fusion protein as vaccine antigen alone.

CONCLUSION

Recombinant Survivin and MIP suppress tumor growth significantly. Combining LHRH fusion protein with Survivin and MIP enhances tumor suppressive effects marginally which provides evidence for recombinant Survivin and LHRH fusion protein as candidates for translating the combination cancer immunotherapy approaches.

Key words: Immunotherapy; Survivin; Luteinizing hormone-releasing hormone fusion protein; Combination immunotherapy; Breast cancer

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Core tip: Targeting Survivin for treatment of cancer is an emerging trend in cancer immunotherapy. In this study we optimized the dose of recombinant full-length Survivin for tumor growth inhibition. Since luteinizing hormone-releasing hormone (LHRH) is also known to support breast cancer in case of premenopausal women, we combined recombinant LHRH fusion protein with Survivin and *Mycobacterium indicus pranii* and obtained a positive anti-tumor response. We report our results of using recombinant Survivin protein and LHRH fusion protein as potential vaccine antigens for immunotherapy of murine model of breast cancer which was developed by injecting 4T-1 mammary cell line in syngeneic Balb/c mice. Immunized mice challenged with syngeneic 4T-1 cells exhibited suppression of tumor growth and metastasis in lungs in comparison to control mice.

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INTRODUCTION

Combination methods that combine two or more therapeutic agents have become a principal treatment modality for breast cancer, especially for metastatic breast cancer^[1]. The rationale behind combination approach is to enhance therapeutic response by targeting multiple pathways involved in the complex process of oncogenesis. However, none of the empirically defined combination of drugs such as taxanes/anthracyclins has been able to overcome the problems of drug resistance and metastasis in breast cancer and impart a satisfactory toxicity profile when administered to patients^[2,3]. In order to overcome the drawbacks associated with prevalent drug combinations, immunotherapy is proposed as a viable approach for developing targeted and safe treatment options for cancer^[4,5]. Advancements in molecular mechanisms of cancer cell survival have led to identification of new targets which can be employed for immunotherapy^[6].

Cancer immunotherapy approach using immunomodulators, monoclonal antibodies or vaccines against tumor growth promoters, have shown benefit in preclinical models of many cancers and clinical trials^[7,8]. However since tumor cells often develop immune evasion mechanisms^[9], combination of tumor antigens may be used to counteract immune resistance by tumor cells. We hypothesize that combination immunotherapy based on potent tumor antigen and hormone, may present a viable approach to enhance the therapeutic landscape and effectively counteract the heterogeneous process of tumor development in breast cancer.

Survivin is a tumor antigen exclusively expressed on tumor cells, essential for cancer cell survival, and its overexpression is associated with aggressiveness of the disease^[10,11]. It has been widely explored as a candidate antigen for cancer immunotherapy^[12]. Similarly, many tumor cells from breast, prostate, ovary or endometrial origin are hormone dependent for their survival^[13]. Therefore, immune system mediated neutralization of hormones is also a key therapeutic strategy for hormone dependent cancers^[14]. Immunization against hormones such as luteinizing hormone-releasing hormone (LHRH) using LHRH based peptide vaccines have shown efficacy as vaccine antigens in preclinical models of hormone dependent cancers such as breast and prostate cancer^[15].

We have previously shown that preventive immunization with a combination of recombinant Survivin protein and an immunomodulator *Mycobacterium indicus pranii* (MIP) provides protection to Balb/c mice when challenged with syngeneic mammary tumor 4T-1 cells^[16]. This communication describes the studies undertaken to determine the optimum dose of Survivin antigen for maximum tumor suppressive effect. Also investigated is the synergy between Survivin and LHRH fusion protein as vaccine antigens for immunotherapy of murine model of breast cancer. 4T-1 murine breast cancer cells not only display Survivin as tumor antigen but also have receptors for LHRH^[17,18]. Thus active immunization against both the Survivin and LHRH, is likely to neutralize Survivin and also make LHRH unavailable for uptake by the

cancer cells, hence generate enhanced protective immune response directed against the cancer cells.

MATERIALS AND METHODS

Cloning and expression of recombinant mouse Survivin and LHRH fusion protein

Survivin and LHRH fusion protein, were used as vaccine antigens. These antigens were expressed as recombinant proteins using *E. coli* based host-vector systems as described previously^[16,19]. Survivin and LHRH fusion protein were purified as approximately 17 kDa and 14 kDa size proteins, respectively using affinity based Ni-NTA chromatography. Purified proteins were adsorbed on alum before administration as immunogens in mice. MIP was used as immunomodulator along with the recombinantly made vaccine antigens.

Immunogenicity and efficacy of Survivin vaccine in vivo

Eight-10 week old inbred Balb/c mice were used for conducting the immunogenicity and efficacy studies of the Survivin vaccine alone *in vivo*. For determining the optimal Survivin dose, study was carried out in 3 groups of mice. Mice were divided into 3 groups randomly with 6 animals in each group. Group 1 was administered phosphate buffer saline (PBS) buffer. While group 2 mice was administered alum adsorbed 20 µg Survivin vaccine (+ MIP), group 3 mice received alum adsorbed 50 µg Survivin dose along with MIP. Five × 10⁶ cells of heat-killed MIP were used in each dose wherever required. Each group of mice was given 2 boosters of its respective dose at 15 d interval after the primary immunization. Sixty days after the last immunization, mice in all the groups were challenged with 3.0 × 10⁵ 4T-1 murine breast cancer cells subcutaneously. Third booster of the vaccine was administered on day 18th after the challenge, when tumors were palpable. Tumor size was measured every alternate day till the end of the study. Tumor volume was measured bi-dimensionally with digital vernier calipers and tumor volume calculated using the formula: (a × b²) / 2, where “a” is the largest diameter and “b” is the perpendicular diameter. Tumor volumes were measured till day 35 post challenge with 4T-1 cells. Results are expressed as mean ± SD of tumor volume for each group.

Immunogenicity and efficacy of combination of Survivin and LHRH fusion protein in comparison to Survivin and LHRH fusion protein alone

Studies were designed in 4 groups of mice each consisting of 6 of 8-10 week old inbred Balb/c mice. Recombinant antigen(s) were administered intramuscularly along with heat-killed 5.0 × 10⁶ cells of MIP. Group 1 of mice received 20 µg of Survivin protein alone, group 2 received 20 µg of LHRH(6leu)-LTB (LTB- heat-labile enterotoxin of *E. coli*) protein alone and group 3 received combination of 20 µg of Survivin and 20 µg of LHRH(6leu)-LTB proteins. Each group of mice was given 2 boosters of its respective vaccine dose at 15 d interval after the primary immunization. A control group of mice was administered PBS buffer. Fifty-two days after first dose, mice were inoculated with 4T-1 breast cancer cells (3.0 × 10⁵ cells in 100 µL of PBS buffer) subcutaneously on right flank. It was recorded as day 0 of tumor challenge. Tumors were palpable in mice in 18 d after tumor cell inoculation. Third booster of vaccine antigen(s) along with immunomodulator MIP was given intramuscularly at the same day when tumors were palpable. Tumor volume was measured till the end of the study. Mice were sacrificed at the end of the study and lungs were examined for the presence of pulmonary metastases.

Estimation of Interferon-γ levels in sera

Sera were collected from mice and assayed for Interferon gamma levels using commercially available enzyme linked immunosorbent assay (ELISA) kit (E Biosciences, United States) as per the manufacturer’s protocol. Briefly, 96-well microtitre plate was coated with 100 µL/well of IFN-γ capture antibody at a dilution of 1:1000 and incubated overnight at 4 °C. Plates were then washed thrice with PBST buffer (PBS buffer supplemented with 0.05% Tween-20) and then blocked with ELISA diluent (250 µL/well) at 37 °C for 2 h. Standard was serially diluted in ELISA diluent to obtain concentration in the range of 15-2000 pg/mL and 100 µL of standard of each concentration was added to the corresponding wells. Serum samples were pooled for each group of mice, diluted in ELISA diluent and added to the respective wells at 100 µL/well. Plate was then incubated for 2 h at 37 °C. Biotinylated anti-IFN-γ detection antibody was added to each well (100 µL/well) at a dilution of 1:1000 followed by incubation of 100 µL/well of Avidin-HRP diluted at 1:250 for 30 min. After washing, TMB was added in each well and incubated in dark for 15 min. Reaction was stopped with 2N H₂SO₄ and absorbance was read at 450 nm. Standard curve was prepared by

plotting the average absorbance of each standard on the vertical axis versus the corresponding IFN- γ standard concentration on the horizontal axis and fitted using 4 parameter logistic (4PL) regression analysis. Actual concentration of IFN- γ in mouse serum samples was calculated by extrapolating OD values using the 4PL curve.

ELISA

ELISA technique was used for the analysis of immunogenicity of the recombinant proteins in mice. ELISA plate (Nunc, Thermo Fisher Scientific, United States) was coated with 100 μ L (5 μ g/mL concentration) of recombinant Survivin or LHRH(6leu)-LTB protein, and incubated overnight at 4 °C. Plate was washed thrice with 10 mmol/L PBS, pH 7.4 supplemented with 0.05% Tween 20 followed by blocking with 2% Bovine serum albumin (BSA) in PBS buffer. Plate was washed with PBST buffer thrice. One hundred micro lift of the serum diluted at 1:5000 in PBS buffer, was added to the wells of the plate, and incubated for 1 h at 37°C. HRP-labeled goat anti-mouse antibody was added at a dilution of 1:10000 to each well and color was developed by adding 100 μ L of O-phenylenediamine (OPD) solution containing Hydrogen Peroxide. The reaction was stopped by addition of 50 μ L of 2N H₂SO₄. Plate was read at 492 nm in an ELISA plate reader (ELX 800MS, Biotek, United States) and results were expressed as mean \pm SD of absorbance values.

Ethics statement

Pathogen free female Balb/c mice used in the study were procured from National Institute of Nutrition, Hyderabad. Animals were housed at animal facility of Amity University Uttar Pradesh and were given ad libitum access to water and food. All experimental procedures were in strict agreement with committee for the purpose of control and supervision of experiments on animals (CPCSEA) guidelines for the care and use of laboratory animals and approved by institutional animal ethics committee (IAEC) of Amity University Uttar Pradesh.

Statistical analysis

Data was analyzed using two tailed student's *t* test. *P* value < 0.05 was reported as significant. Data was analyzed using graph pad prism software.

RESULTS

Optimum dose of Survivin antigen dose for the effective inhibition of tumor growth in 4T-1 model

We reported previously that the recombinant Survivin vaccine is efficacious in preventing the growth of 4T-1 based murine model of breast cancer^[16]. Increase in vaccine dose from 5 μ g to 10 μ g, led to better immunogenicity which resulted in significant reduction in tumor suppression^[16]. Therefore, it was decided to test if it was possible to employ doses higher than previously used 10 μ g dose to inhibit the tumor maximally. Thus the studies were designed to evaluate the efficacy of the vaccine at 20 μ g and 50 μ g doses. Pathogen-free mice of similar weight (6 mice/group) were first immunized with alum adsorbed recombinant Survivin vaccine along with immune-modulator MIP. Mice were then challenged with syngeneic 4T-1 cells and observed for the tumor growth in presence of pre-formed anti-Survivin antibodies. Three immunizations of alum adsorbed recombinant Survivin and MIP were administered at 15 d interval following which 4T-1 cancer cells were injected at day 90. A booster of the Survivin vaccine was given at day 108 when tumors were palpable. Tumor volume was measured and continued till 35 d post injection of cancer cells. **Figure 1** shows the volume of the tumor developed in mice immunized with 20 μ g and 50 μ g of Survivin protein administered along with MIP. It is evident from the Figure that significant decrease in tumor volume was observed in mice immunized with 20 μ g dose of the vaccine (*P* = 0.04) in comparison to control mice. However increase in vaccine dose to 50 μ g did not lead to any significant inhibition in tumor volume thus reflecting dose-inhibition related effects. **Figure 2** shows the IFN- γ levels determined in the sera collected from immunized mice. Survivin at 20 μ g dose induced higher levels of IFN- γ in comparison to 50 μ g dose, which is in accordance with the observed efficacy of the anti-Survivin antibodies in preventing the growth of tumor at 20 μ g dose. The mice in each group did not show any visible adverse effects of the treatment.

Active immunization against a combination of Survivin and LHRH fusion protein

After finding the optimal dose of Survivin vaccine, studies were advanced to investigate the additive effect of LHRH vaccine, if any, in terms preventing the 4T-1

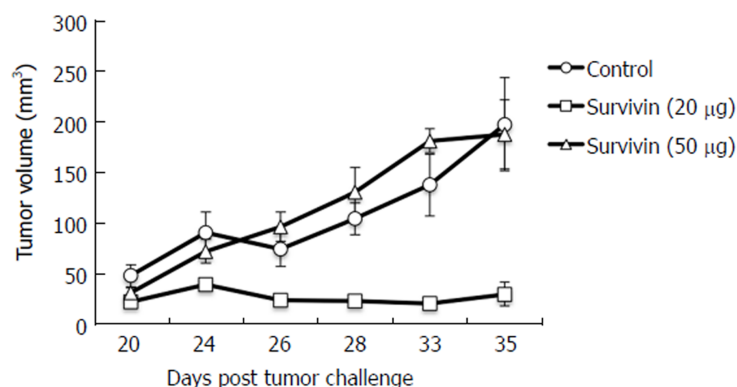


Figure 1 4T-1 breast tumor volume in mice injected with phosphate buffer saline buffer (circle), 20 µg Survivin + *Mycobacterium indicus pranii* (square) and 50 µg Survivin + *Mycobacterium indicus pranii* (triangle) at different time points after tumor was palpable.

breast cancer cells. Studies were carried out in 4 groups of pathogen free mice (6 mice/group). A group of mice was immunized by combination of alum adsorbed Survivin and LHRH(6leu)-LTB proteins along with immunomodulator MIP. Two groups of mice were injected with Survivin alone (+ MIP) and LHRH(6leu)-LTB alone (+ MIP), respectively. Mice receiving the PBS buffer formed the control group. Mice were immunized thrice at 15 d interval with combination or individual antigens following which 4T-1 tumor cells were injected at 50th day. Boosters of the vaccine antigens were administered when the tumors became palpable. The tumor size was measured post 35 d of tumor cell challenge. Though LHRH(6leu)-LTB alone prevented the growth of tumor size in comparison to control group mice but did not reach statistical significance, inhibition was more significant with Survivin alone ($P = 0.04$). Combining it with LHRH fusion protein led to marginally better tumor suppression, $P = 0.02$ (Figure 3). There were no adverse effects of the treatment observed in the mice of each group.

Evaluation of metastasis of primary tumor to lungs

Mice challenged with the tumor cells were followed to evaluate the efficacy of the active immunization by Survivin and LHRH fusion protein alone or in combination in terms of preventing cancer metastasis. The secondary growth of the cancer was studied by counting the number of nodules in the lungs of the mice challenged with the tumor cells. Table 1 shows the mean of number of nodules counted in various groups at the end of the study. Mice immunized with the combination of Survivin and LHRH(6leu)-LTB along with MIP did not show any visible nodules in the lungs 30 d after challenge with 4T-1 cells whereas mice immunized with either Survivin + MIP or LHRH(6leu)-LTB + MIP showed presence of tumor nodules in lungs. Thus combination of Survivin and LHRH(6leu)-LTB along with immunomodulator MIP is capable of controlling the pulmonary metastasis besides suppressing the primary tumor growth.

Characterization of total IgG response

Sera collected from each group of mice were checked for the total IgG titre against the respective antigen(s) pre and post challenge with tumour cells. Both Survivin and LHRH(6leu)-LTB antigens were able to induce antibody response when used to immunize the mice alone. Combined immunization with both the antigens, also led to induction of both anti-Survivin and anti-LHRH antibodies thus confirming their immunogenicity (Figure 4).

DISCUSSION

Tumor antigens have been proposed as important therapeutic targets for cancer immunotherapy^[20]. Immunotherapy approaches incorporating Survivin as tumor antigen have shown efficacy for elimination of cancer cells. Various approaches such as peptide vaccines^[21-23], multi epitope vaccines^[24,25] or DNA vaccines incorporating Survivin as vaccine antigen^[26,27], have shown efficacy in preclinical studies. It has also been suggested that combination of Survivin derived peptides with immunomodulators or immunomodulatory cytokines such as IFN alpha or induces a significant overall survival in patients with advanced or recurrent urothelial cancer^[28].

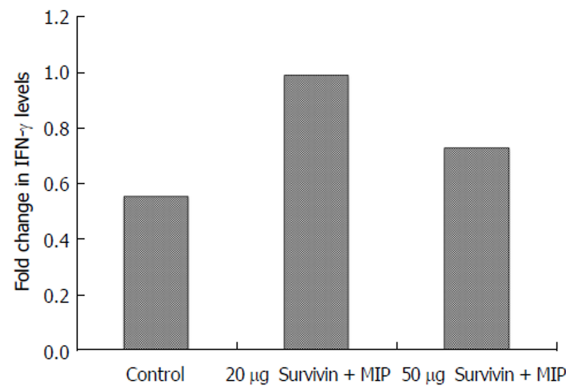


Figure 2 Interferon-gamma levels measured in mice immunized with 20 µg Survivin (+ *Mycobacterium indicus pranii*) and 50 µg Survivin (+ *Mycobacterium indicus pranii*) in comparison to phosphate buffer saline administered control mice and *Mycobacterium indicus pranii* administered mice. Interferon-gamma is expressed as fold change relative to the levels obtained in phosphate buffer saline administered mice before the tumor challenge. MIP: *Mycobacterium indicus pranii*.

We have also previously reported that combination of Survivin along with MIP has potent dose dependent anti-tumor effects in a mouse model of breast carcinoma^[16]. Our data suggests that MIP alone is not effective in preventing tumor growth (data not shown) however combining MIP with tumor antigen such as Survivin gives significant tumor suppression. MIP may work as immunomodulator and serves to enhance the immune response against Survivin antigen.

We have demonstrated in the present study that increasing the dose of recombinant Survivin to 20 µg induces most effective tumor suppression in murine 4T-1 model of breast cancer. Interestingly, further increasing the dose of antigen does not induce a tumor suppressive effect. Suppression of immune response at supraoptimal doses of antigen has also been previously reported^[29-31]. We hypothesize that high dose Survivin antigen may induce a suboptimal or anergic immune response which may not be able to inhibit the tumor growth.

Interferon-γ secreted by Th1 CD4+ T, NK cells, CD8+ T cells, APCs and B-cells cells is known for its anti-tumor effects^[32]. It has role in rejection of transplanted tumors and inhibition of formation of spontaneous tumors^[33,34]. Data presented in the present studies also show higher levels of IFN-γ at 20 µg dose of Survivin than the 50 µg dose which is in agreement with observed better suppression of tumor with 20 µg dose. Previous studies have confirmed the important role of humoral immune response in providing in anti-tumor immunity^[35]. Our studies also indicate that antigen specific antibodies were induced after immunization with tumor antigens which possibly have contributed to tumor suppression. A recent study by Fenstermaker *et al*^[36] has also demonstrated therapeutic effect of monoclonal antibody against Survivin in vivo in murine glioma model.

Many of the premenopausal breast cancers are also hormone-dependent which rationalizes the use of LHRH agonists as an effective treatment option in breast cancers^[37]. Various combinations of LHRH agonists such as cisplatin loaded LHRH nanoparticles which have shown good anti-tumor response in 4T1 breast tumor model by accumulating higher cisplatin in tumors^[18]. Another conjugate of LHRH, Pt-Mal-LHRH (activated cisplatin linked to LHRH peptide with a malonate linker) showed targeted delivery to cells expressing LHRH receptors. Treatment of 4T1 breast cancer with Pt-Mal-LHRH reduced tumor volume and metastasis to lungs^[38].

We evaluated the possibility of using an immunotherapy based on a combination of Survivin and LHRH fusion protein in 4T-1 murine model of breast cancer. LHRH(6leu)-LTB, a vaccine developed for the immunotherapy of androgen dependent cancers, was used as an immunogen in combination with Survivin along with immunomodulator, MIP. The idea was to generate anti-LHRH antibodies to neutralize LHRH making it unavailable for intake by the breast cancer cells through the receptors. It was observed that efficacy of the combination was marginally better than the Survivin alone but significantly better when only LHRH(6leu)-LTB was used along with MIP. More importantly, combination was best at preventing metastasis of primary tumors to lungs.

Our results show promising outcome in a prophylactic scenario. We undertook the prophylactic approach for two reasons: (1) to investigate if the immunization induces a tumor protective response *in vivo*; and (2) this prophylactic approach also closely mimics the clinical situation where primary tumors have been removed or adjuvant

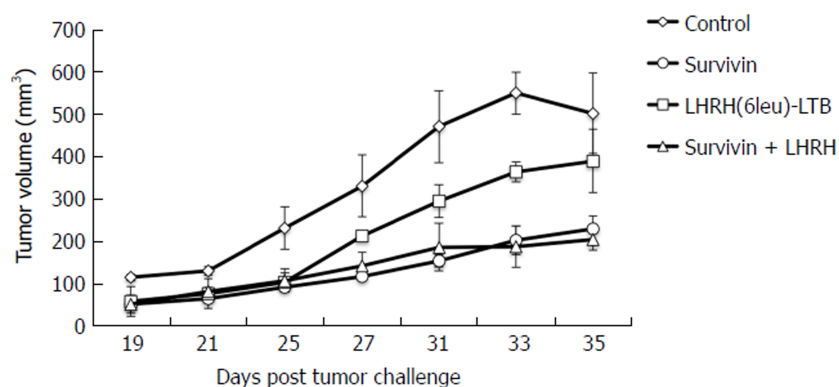


Figure 3 4T-1 Breast tumor volume in mice immunized with PBS buffer, 20 μ g Survivin + *Mycobacterium indicus pranii*, 20 μ g luteinizing hormone-releasing hormone + *Mycobacterium indicus pranii* and combination of 20 μ g Survivin and 20 μ g luteinizing hormone-releasing hormone along with *Mycobacterium indicus pranii*. Tumor volume is measured at different time points from day 19th when tumor was palpable. LHRH: Luteinizing hormone-releasing hormone.

treatment for cancer patients has been initiated. In both these settings, the patients undergo surgery. Though patients are rendered free of tumor, chances of recurrence are very high. If translation of LHRH fusion protein and Survivin based combination approach is successful, it may find applications in such clinical scenario. We have also investigated the therapeutic potential of this approach by immunizing the mice with implanted and palpable tumor, which has showed similar suppression of tumor growth (data not shown).

In conclusion, we have shown the Survivin (+ MIP) at a dose of 20 μ g is most effective in preventing the growth of 4T-1 breast tumor cells in mice. Incorporation of anti-LHRH vaccine exercises a synergistic effect. A schematic diagram depicting the strategy used in the present study has been shown as **Figure 5**. Although our results show the protective role of combination immunotherapy with Survivin and LHRH antigens in murine tumor model, yet the study is limited by the fact the results need to be validated in other tumor models and further translational development needs to be undertaken for appropriate human application. Though it did not lead to much benefit in preventing the growth of primary tumor, it was highly effective in blocking the pulmonary metastasis. Our data suggests that combination of Survivin and LHRH fusion protein may hold immense promise for further development of immunotherapeutic approaches in management of breast cancers. The combination enhances immune response that is involved in inhibiting tumor growth, thus it will be effective in immune competent organisms and has to be supplemented with other therapies for use in immune compromised individuals. Furthermore, investigating the molecular mechanism of action of the combination leading to tumor inhibition may also lead to development of novel targeted therapies for cancer.

Table 1 Number of nodules in the lungs of mice challenged with 4T-1 breast cancer cells post immunization with PBS buffer, Survivin, LHRH(6leu)-LTB and combination of Survivin and LHRH(6leu)-LTB

Immunization group	Average No. of lung nodules
PBS (untreated)	> 20
Survivin	9
LHRH(6leu)-LTB	9
Survivin + LHRH(6leu)-LTB	0

LHRH: Luteinizing hormone-releasing hormone.

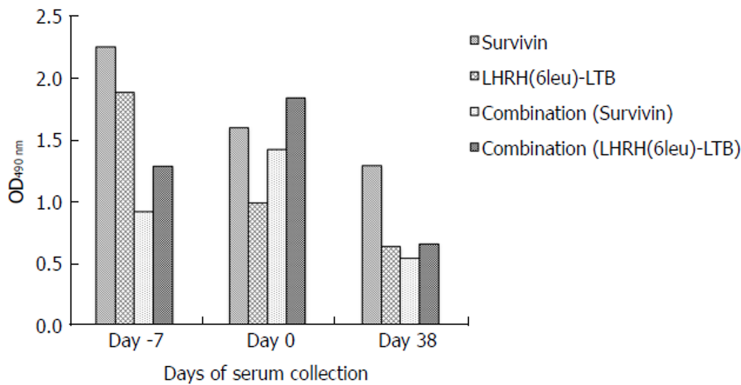


Figure 4 Graph representing the comparative level of total IgG antibody measured in mice immunized with Survivin (+ *Mycobacterium indicus pranii*), LHRH(6leu)LTB (+ *Mycobacterium indicus pranii*) and combination of Survivin and LHRH(6leu)-LTB (+ *Mycobacterium indicus pranii*). The serum was collected pre tumor challenge (Day -7), on tumor challenge (Day 0) and at the end of the study (Day 38). The total IgG titer was determined against respective antigens in each group. Groups of mice receiving Survivin alone and combination were checked for anti-Survivin antibody titers. Similarly, anti-LHRH titres were checked in mice in groups corresponding to LHRH(6leu)-LTB alone and combination. LHRH: Luteinizing hormone-releasing hormone.

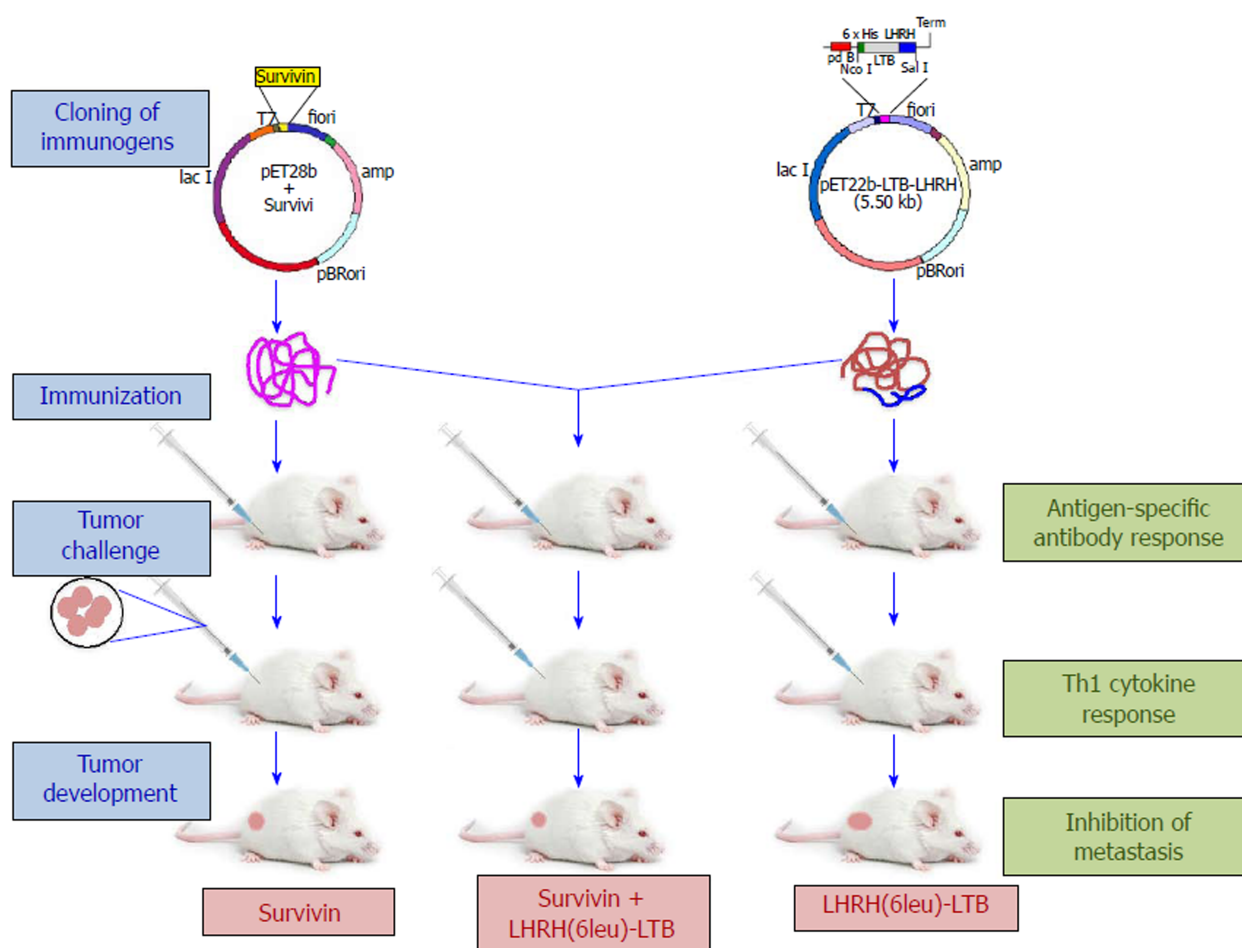


Figure 5 Schematic representation of the study of anti-tumor effect of Survivin, luteinizing hormone-releasing hormone fusion protein individually and in combination. The study was conducted on four groups of mice: Untreated or control, Survivin alone, LHRH(6leu)LTB alone and Survivin + LHRH(6leu)LTB. Individual recombinant proteins were purified and used as immunogen along with *Mycobacterium indicus pranii* as an immunomodulator in murine model. The mice from all the groups were further challenged with tumor cells, 4T1 and tumor development was observed in each group. LHRH fusion protein was not effective in suppressing tumor alone. However, the combination of both Survivin and LHRH(6leu)LTB inhibited tumor growth substantially followed by Survivin alone. LHRH: Luteinizing hormone-releasing hormone.

ARTICLE HIGHLIGHTS

Research background

Tumor cells often develop immune evasion mechanisms, thus combination of tumor antigens may be used to counteract immune resistance. Survivin is a tumor antigen exclusively expressed on tumor cells, essential for cancer cell survival, and its overexpression is associated with aggressiveness of the disease. Similarly, many tumor cells from breast, prostate, ovary or endometrial origin are hormone dependent for their survival. Immunization against hormones such as luteinizing hormone-releasing hormone (LHRH) using LHRH based peptide vaccines have shown efficacy as vaccine antigens in preclinical models of hormone dependent cancers such as breast and prostate cancer. This communication describes the studies undertaken to determine the optimum dose of Survivin antigen for maximum tumor suppressive effect and the synergy between Survivin and LHRH as vaccine antigens for immunotherapy of murine model of breast cancer, 4T-1.

Research motivation

We undertook the prophylactic approach for two reasons: (1) to investigate if the immunization induces a tumor protective response *in vivo* and (2) this prophylactic approach also closely mimics the clinical situation where primary tumors have been removed or adjuvant treatment for cancer patients has been initiated. Major issues in cancer treatment are the recurrence of tumor after surgery and chemoresistance. Our study was aimed at targeting these issues and developing a strategy, which may prevent tumor spread and recurrence and can overcome resistance. Generating immune response in the body against the tumor antigens will be a safer way of treatment. Anti-Survivin approach may help in overcoming the problem of resistance to therapy.

Research objectives

The aim of this study was to evaluate the possibility of using an immunotherapy based on a combination of Survivin and LHRH fusion protein in 4T-1 murine model of breast cancer. LHRH(6leu)-LTB was used as an immunogen in combination with Survivin along with an immunomodulator, *Mycobacterium indicus pranii* (MIP). It was observed that efficacy of the combination was marginally better than the Survivin alone but significantly better when only LHRH(6leu)-LTB was used along with MIP. More importantly, combination was best at preventing metastasis of primary tumors to lungs. In most cancer cases, the patients undergo surgery. Though patients are rendered free of tumor, chances of recurrence are very high. If translation of LHRH fusion protein and Survivin based combination approach is successful, it may find applications in such clinical scenario.

Research methods

Survivin and LHRH fusion protein were used as vaccine antigens. These antigens were expressed as recombinant proteins using *E. coli* based host-vector systems. Purified proteins were adsorbed on alum before administration as immunogens in mice. MIP was used as immunomodulator along with the recombinantly made vaccine antigens. Inbred Balb/c mice were used for conducting the immunogenicity and efficacy studies of the Survivin vaccine *in vivo*. Tumor volume was measured bi-dimensionally with digital vernier calipers and results are expressed as mean \pm SD of tumor volume for each group. Anti-tumor efficacy of combination of Survivin and LHRH fusion protein in comparison to Survivin and LHRH fusion protein alone was also determined. Sera were collected from mice and assayed for Interferon gamma levels using ELISA kit. ELISA was also performed for the analysis of immunogenicity of the recombinant proteins in mice.

Research results

We have shown that Survivin (+ MIP) at a dose of 20 μ g is most effective in preventing the growth of 4T-1 breast tumor cells in mice and incorporation of anti-LHRH vaccine exercises a synergistic effect. The study is limited by the fact that the results need to be validated in other tumor models and further translational development needs to be undertaken for appropriate human application. The combination will be effective in immune competent organisms and has to be supplemented with other therapies for use in immune compromised individuals. Furthermore, investigating the molecular mechanism of action of the combination leading to tumor inhibition may also lead to development of novel targeted therapies for cancer.

Research conclusions

Our results show the protective role of combination immunotherapy with Survivin and LHRH antigens in murine tumor model. Though it did not lead to much benefit in preventing the growth of primary tumor, it was highly effective in blocking the pulmonary metastasis. Combination of Survivin and LHRH fusion protein may hold immense promise for further development of immunotherapeutic approaches in management of breast cancers.

REFERENCES

1. Zanardi E, Bregni G, de Braud F, Di Cosimo S. Better Together: Targeted Combination Therapies in Breast Cancer. *Semin Oncol* 2015; **42**: 887-895 [PMID: 26615133 DOI: 10.1053/j.seminoncol.2015.09.029]
2. Biganzoli L, Cufer T, Bruning P, Coleman R, Duchateau L, Calvert AH, Gamucci T, Twelves C, Fargeot P, Epelbaum R. Doxorubicin and paclitaxel versus doxorubicin and cyclophosphamide as first-line chemotherapy in metastatic breast cancer: The European Organization for Research and Treatment of Cancer 10961 Multicenter Phase III Trial. *J Clin Oncol* 2002; **20**: 3114-3121 [PMID: 12118025 DOI: 10.1200/JCO.2002.11.005]
3. Jassem J, Pieńkowski T, Płuzańska A, Jelic S, Gorbunova V, Mrcic-Krmpotic Z, Berzins J, Nagykalnai T, Wigler N, Renard J. Doxorubicin and paclitaxel versus fluorouracil, doxorubicin, and cyclophosphamide as first-line therapy for women with metastatic breast cancer: final results of a randomized phase III multicenter trial. *J Clin Oncol* 2001; **19**: 1707-1715 [PMID: 11251000 DOI: 10.1200/JCO.2001.19.6.1707]
4. Morrissey KM, Yuraszek TM, Li CC, Zhang Y, Kasichayanula S. Immunotherapy and Novel Combinations in Oncology: Current Landscape, Challenges, and Opportunities. *Clin Transl Sci* 2016; **9**: 89-104 [PMID: 26924066 DOI: 10.1111/cts.12391]
5. Khalil DN, Smith EL, Brentjens RJ, Wolchok JD. The future of cancer treatment: immunomodulation, CARs and combination immunotherapy. *Nat Rev Clin Oncol* 2016; **13**: 394 [PMID: 27118494 DOI: 10.1038/nrclinonc.2016.65]
6. Wang RF, Wang HY. Immune targets and neoantigens for cancer immunotherapy and precision medicine. *Cell Res* 2017; **27**: 11-37 [PMID: 28025978 DOI: 10.1038/cr.2016.155]
7. Farkona S, Diamandis EP, Blasutig IM. Cancer immunotherapy: the beginning of the end of cancer? *BMC Med* 2016; **14**: 73 [PMID: 27151159 DOI: 10.1186/s12916-016-0623-5]
8. Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. *Nat Rev Immunol* 2010; **10**: 317-327 [PMID: 20414205 DOI: 10.1038/nri2744]
9. Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, Lichter T, Decker WK, Whelan RL, Kumara HMCS. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. *Semin Cancer Biol* 2015; **35** Suppl: S185-S198 [PMID: 25818339 DOI: 10.1016/j.semcancer.2015.03.004]
10. Cheng KY, Wang ZL, Gu QY, Hao M. Survivin Overexpression Is Associated with Aggressive Clinicopathological Features in Cervical Carcinoma: A Meta-Analysis. *PLoS One* 2016; **11**: e0165117 [PMID: 27764228 DOI: 10.1371/journal.pone.0165117]
11. Chuwa AH, Sone K, Oda K, Ikeda Y, Fukuda T, Wada-Hiraike O, Inaba K, Makii C, Takeuchi M, Oki S. Significance of survivin as a prognostic factor and a therapeutic target in endometrial

- cancer. *Gynecol Oncol* 2016; **141**: 564-569 [PMID: 27079211 DOI: 10.1016/j.ygyno.2016.04.003]
- 12 **Boullosa LF**, Savaliya P, Bonney S, Orchard L, Wickenden H, Lee C, Smits E, Banham AH, Mills KI, Orchard K. Identification of survivin as a promising target for the immunotherapy of adult B-cell acute lymphoblastic leukemia. *Oncotarget* 2017; **9**: 3853-3866 [PMID: 29423088 DOI: 10.18632/oncotarget.23380]
- 13 **Capper CP**, Rae JM, Auchus RJ. The Metabolism, Analysis, and Targeting of Steroid Hormones in Breast and Prostate Cancer. *Horm Cancer* 2016; **7**: 149-164 [PMID: 26969590 DOI: 10.1007/s12672-016-0259-0]
- 14 **Junco JA**, Basalto R, Fuentes F, Bover E, Reyes O, Pimentel E, Calzada L, Castro MD, Arteaga N, López Y. Gonadotrophin releasing hormone-based vaccine, an effective candidate for prostate cancer and other hormone-sensitive neoplasms. *Adv Exp Med Biol* 2008; **617**: 581-587 [PMID: 18497085 DOI: 10.1007/978-0-387-69080-3_60]
- 15 **Junco JA**, Peschke P, Zuna I, Ehemann V, Fuentes F, Bover E, Pimentel E, Basulto R, Reyes O, Calzada L. Immunotherapy of prostate cancer in a murine model using a novel GnRH based vaccine candidate. *Vaccine* 2007; **25**: 8460-8468 [PMID: 18022737 DOI: 10.1016/j.vaccine.2007.09.033]
- 16 **Garg H**, Gupta JC, Talwar GP, Dubey S. Immunotherapy approach with recombinant survivin adjuvanted with alum and MIP suppresses tumor growth in murine model of breast cancer. *Prep Biochem Biotechnol* 2018; **48**: 264-269 [PMID: 29355462 DOI: 10.1080/10826068.2018.1425710]
- 17 **Taheri A**, Dinarvand R, Ahadi F, Khorramizadeh MR, Atyabi F. The in vivo antitumor activity of LHRH targeted methotrexate-human serum albumin nanoparticles in 4T1 tumor-bearing Balb/c mice. *Int J Pharm* 2012; **431**: 183-189 [PMID: 22531853 DOI: 10.1016/j.ijpharm.2012.04.033]
- 18 **Li M**, Tang Z, Zhang Y, Lv S, Li Q, Chen X. Targeted delivery of cisplatin by LHRH-peptide conjugated dextran nanoparticles suppresses breast cancer growth and metastasis. *Acta Biomater* 2015; **18**: 132-143 [PMID: 25735801 DOI: 10.1016/j.actbio.2015.02.022]
- 19 **Gupta JC**, Hada RS, Sahai P, Talwar GP. Development of a novel recombinant LHRH fusion protein for therapy of androgen and estrogen dependent cancers. *Protein Expr Purif* 2017; **134**: 132-138 [PMID: 28410993 DOI: 10.1016/j.pep.2017.04.003]
- 20 **Srinivasan R**, Wolchok JD. Tumor antigens for cancer immunotherapy: therapeutic potential of xenogeneic DNA vaccines. *J Transl Med* 2004; **2**: 12 [PMID: 15090064 DOI: 10.1186/1479-5876-2-12]
- 21 **Ciesielski MJ**, Kozbor D, Castanaro CA, Barone TA, Fenstermaker RA. Therapeutic effect of a T helper cell supported CTL response induced by a survivin peptide vaccine against murine cerebral glioma. *Cancer Immunol Immunother* 2008; **57**: 1827-1835 [PMID: 18438666 DOI: 10.1007/s00262-008-0510-9]
- 22 **Fenstermaker RA**, Ciesielski MJ, Qiu J, Yang N, Frank CL, Lee KP, Mechtler LR, Belal A, Ahluwalia MS, Hutson AD. Clinical study of a survivin long peptide vaccine (SurvaxM) in patients with recurrent malignant glioma. *Cancer Immunol Immunother* 2016; **65**: 1339-1352 [PMID: 27576783 DOI: 10.1007/s00262-016-1890-x]
- 23 **Yang Z**, Wang L, Wang H, Shang X, Niu W, Li J, Wu Y. A novel mimovirus vaccine containing survivin epitope with adjuvant IL-15 induces long-lasting cellular immunity and high antitumor efficiency. *Mol Immunol* 2008; **45**: 1674-1681 [PMID: 18035418 DOI: 10.1016/j.molimm.2007.10.026]
- 24 **Idenoue S**, Hirohashi Y, Torigoe T, Sato Y, Tamura Y, Hariu H, Yamamoto M, Kurotaki T, Tsuruma T, Asanuma H. A potent immunogenic general cancer vaccine that targets survivin, an inhibitor of apoptosis proteins. *Clin Cancer Res* 2005; **11**: 1474-1482 [PMID: 15746049 DOI: 10.1158/1078-0432.CCR-03-0817]
- 25 **Lennerz V**, Gross S, Gallerani E, Sessa C, Mach N, Boehm S, Hess D, von Boehmer L, Knuth A, Ochsenbein AF. Immunologic response to the survivin-derived multi-epitope vaccine EMD640744 in patients with advanced solid tumors. *Cancer Immunol Immunother* 2014; **63**: 381-394 [PMID: 24487961 DOI: 10.1007/s00262-013-1516-5]
- 26 **Lladser A**, Ljungberg K, Tufvesson H, Tazzari M, Roos AK, Quest AF, Kiessling R. Intradermal DNA electroporation induces survivin-specific CTLs, suppresses angiogenesis and confers protection against mouse melanoma. *Cancer Immunol Immunother* 2010; **59**: 81-92 [PMID: 19526360 DOI: 10.1007/s00262-009-0725-4]
- 27 **Xiang R**, Mizutani N, Luo Y, Chiodoni C, Zhou H, Mizutani M, Ba Y, Becker JC, Reisfeld RA. A DNA vaccine targeting survivin combines apoptosis with suppression of angiogenesis in lung tumor eradication. *Cancer Res* 2005; **65**: 553-561 [PMID: 15695399]
- 28 **Tanaka T**, Kitamura H, Inoue R, Nishida S, Takahashi-Takaya A, Kawami S, Torigoe T, Hirohashi Y, Tsukamoto T, Sato N. Potential survival benefit of anti-apoptosis protein: survivin-derived peptide vaccine with and without interferon alpha therapy for patients with advanced or recurrent urothelial cancer--results from phase I clinical trials. *Clin Dev Immunol* 2013; **2013**: 262967 [PMID: 24363758 DOI: 10.1155/2013/262967]
- 29 **Gotsman I**, Israeli D, Alper R, Rabbani E, Engelhardt D, Ilan Y. Induction of immune tolerance toward tumor-associated-antigens enables growth of human hepatoma in mice. *Int J Cancer* 2002; **97**: 52-57 [PMID: 11774243 DOI: 10.1002/ijc.1576]
- 30 **Suzuki G**, Kawase Y, Koyasu S, Yahara I, Kobayashi Y, Schwartz RH. Antigen-induced suppression of the proliferative response of T cell clones. *J Immunol* 1988; **140**: 1359-1365 [PMID: 2450125]
- 31 **Michallet MC**, Saltel F, Flacher M, Revillard JP, Genestier L. Cathepsin-dependent apoptosis triggered by supraoptimal activation of T lymphocytes: a possible mechanism of high dose tolerance. *J Immunol* 2004; **172**: 5405-5414 [PMID: 15100281 DOI: 10.4049/jimmunol.172.9.5405]
- 32 **Ikeda H**, Old LJ, Schreiber RD. The roles of IFN gamma in protection against tumor development and cancer immunoediting. *Cytokine Growth Factor Rev* 2002; **13**: 95-109 [PMID: 11900986 DOI: 10.1016/S1359-6101(01)00038-7]
- 33 **Blankenstein T**, Qin Z. The role of IFN-gamma in tumor transplantation immunity and inhibition of chemical carcinogenesis. *Curr Opin Immunol* 2003; **15**: 148-154 [PMID: 12633663 DOI: 10.1016/S0952-7915(03)00007-4]
- 34 **Dunn GP**, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoediting. *Immunity* 2004; **21**: 137-148 [PMID: 15308095 DOI: 10.1016/j.immuni.2004.07.017]

- 35 **Tsou P**, Katayama H, Ostrin EJ, Hanash SM. The Emerging Role of B Cells in Tumor Immunity. *Cancer Res* 2016; **76**: 5597-5601 [PMID: [27634765](#) DOI: [10.1158/0008-5472.CAN-16-0431](#)]
- 36 **Fenstermaker RA**, Figel SA, Qiu J, Barone TA, Dharma SS, Winograd EK, Galbo PM, Wiltse LM, Ciesielski MJ. Survivin Monoclonal Antibodies Detect Survivin Cell Surface Expression and Inhibit Tumor Growth *In Vivo*. *Clin Cancer Res* 2018; **24**: 2642-2652 [PMID: [29540489](#) DOI: [10.1158/1078-0432.CCR-17-2778](#)]
- 37 **Goel S**, Sharma R, Hamilton A, Beith J. LHRH agonists for adjuvant therapy of early breast cancer in premenopausal women. *Cochrane Database Syst Rev* 2009; **4**: CD004562 [PMID: [19821328](#) DOI: [10.1002/14651858.CD004562.pub4](#)]
- 38 **Calderon LE**, Keeling JK, Rollins J, Black CA, Collins K, Arnold N, Vance DE, Ndinguri MW. Pt-Mal-LHRH, a Newly Synthesized Compound Attenuating Breast Cancer Tumor Growth and Metastasis by Targeting Overexpression of the LHRH Receptor. *Bioconj Chem* 2017; **28**: 461-470 [PMID: [27997127](#) DOI: [10.1021/acs.bioconjchem.6b00610](#)]

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Radiation-associated epithelial-myoeplithelial carcinoma among five secondary malignancies: A case report and review of literature

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Abstract

BACKGROUND

Epithelial-myoeplithelial carcinoma (EMC) is a rare, low-grade, malignant tumor that constitutes less than one percent of all salivary gland tumors. To date, only one other case report has described radiation-associated EMC in the English language medical literature.

CASE SUMMARY

In this report, we describe the case of a 56-year-old male patient who presented with a neck mass diagnosed as EMC of the left submandibular gland approximately 30 years after mantle field radiation and chemotherapy for Hodgkin lymphoma. Treatment included resection, re-resection with nodal dissection, and adjuvant chemoradiotherapy. This patient was also diagnosed with 4 other secondary malignancies, including stage IV diffuse large B cell lymphoma in the abdomen with subsequent brain metastases, low-grade neuroendocrine carcinoma of the lung, Hurthle cell adenoma, and small B cell lymphoma before the patient expired. This case provides important information regarding the pathology, clinical sequelae, and management of a patient diagnosed with radiation-associated EMC amidst four concurrent malignancies.

CONCLUSION

Further investigation is needed on the efficacy of adjuvant radiotherapy in EMC, especially atypical EMC.

Key words: Secondary malignancy; Radiation-associated malignancy; Epithelial-myoeplithelial carcinoma; Cancer survivorship; Case report

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Core tip: We describe the second known case of radiation-associated epithelial-myoeplithelial carcinoma (EMC). This patient received chemotherapy and radiation therapy thirty years prior for Hodgkin lymphoma, and in addition to EMC was also diagnosed with four other malignancies in a span of five years. Because of worrisome

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histopathologic features atypical for EMC and pathologic stage of this patient, this patient was treated more aggressively with adjuvant chemoradiotherapy. This case provides important information regarding the pathology, clinical sequelae, and management of radiation-associated EMC. Further investigation is needed on the efficacy of adjuvant radiotherapy in EMC, especially atypical EMC.

Khattab MH, Sherry AD, Ahlers CG, Kirschner AN. Radiation-associated epithelial-myoepithelial carcinoma among five secondary malignancies: A case report and review of literature. *World J Clin Oncol* 2018; 9(8): 200-207

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INTRODUCTION

Epithelial-myoepithelial carcinoma (EMC) is a rare, low-grade malignant tumor of the salivary gland. Originally described by Donath *et al*^[1] in 1972, EMC first appeared in the World Health Organization classification of salivary gland tumors in 1991^[2]. Representing less than one percent of all salivary gland tumors, EMC typically manifests as a bulky, slowly growing mass primarily within the parotid gland or less commonly the submandibular glands, minor salivary glands, or palate^[3]. EMC is more prevalent in females and most often presents in the seventh decade of life^[3]. EMC is recognizable histologically as a biphasic tumor, characterized by inner ductal epithelial cells and abundant outer clear myoepithelial cells^[2,3]. Pathologic subcategorizations of EMC include double clear EMC, oncocytic EMC, sebaceous EMC, apocrine EMC, EMC ex pleomorphic adenoma, and EMC with high-grade transformation (HGT)^[4]. HGT is a transition from low-grade to high-grade morphology with loss of the original histological characteristics. EMC with HGT is exceedingly rare^[4]. The minimum recommended treatment for EMC is surgical resection, although local recurrence after resection has been reported in 23%-50% of cases^[3]. Chemotherapy and radiotherapy (RT) are also appropriate therapeutic options especially with nodal involvement or metastatic disease, although very few data exist regarding their efficacy^[5].

To the best of our knowledge, only one case report has thus far described RT-associated EMC in the English language medical literature^[6]. In this previous report, a 48-year-old male was diagnosed with right submandibular high-grade EMC 25 years after mantle field RT. Multiple resections as well as adjuvant RT and chemotherapy were required due to multiple recurrences over a 4-year period. Although EMC is typically a low-grade tumor, this report conjectured an etiologic association between the extensive necrosis profile and high mitotic activity evident on pathological exam, as well as the unusually young age of the patient, and the course of previous RT.

While seldom utilized today, mantle field RT was the standard of care for Hodgkin lymphoma in the 1960's because of enhanced curative potential compared to more focal treatment. Historically, RT treatment paradigms were built on the concept of extended field radiation therapy (EF-RT), wherein both the primary site and regional lymphatic groups were treated. EF-RT was categorized into mantle field RT and inverted-Y field RT. Mantle field RT covered the cervical, mid-chest, and axillary lymph nodes, while inverted-Y field RT covered the subdiaphragmatic field including all para-aortic, iliac, upper femoral, and inguinal lymph nodes^[7]. Notably, mantle field RT has been associated with an increased risk of breast cancer^[8]. According to De Bruin *et al*^[8], mantle field RT (involving the axillary, mediastinal, and neck nodes) was associated with a 2.7-fold increased risk of breast cancer compared with similarly dosed (36 to 44 Gy) mediastinal nodes alone. Mantle field RT has also been associated with increased cardiac morbidity and mortality^[9].

CASE PRESENTATION

Chief complaints

After subtotal resection of EMC, the patient presented to our institution for evaluation. The patient reported that, from discovery until dissection, the neck mass was without symptomatic changes and was neither painful nor enlarging.

History of present illness

At the age of 56, the patient presented with a chief complaint of a neck mass detected while shaving. Computerized tomography (CT) scan of his neck showed a heterogeneous ovoid mass in the left submandibular region measuring 2.9 cm × 1.8 cm (Figure 1). The CT scan also demonstrated an enlarged left paramedial submental lymph node measuring 1.6 cm × 0.9 cm. Biopsy of the neck mass revealed EMC staining positive for CK7, CKAE1/AE3, CK903, S100, and smooth muscle actin and negative for CK20, PAX8, GATA, CDX2, GFAP. Infiltrates into soft tissue were noted along with fragments of salivary gland with fibrosis and atrophy. The left submandibular mass was then resected in piecemeal fashion, with final pathology report confirming EMC.

History of past illness

At the age of 19, this male patient presented with Hodgkin lymphoma and underwent mantle field RT, chemotherapy, and splenectomy. He did not exhibit evidence of recurrence. At the age of 52, stomach biopsy revealed diffuse large B cell lymphoma (DLBCL) with positron emission tomography (PET) scan displaying diffuse disease in the right axilla, mediastinum, multiple abdominal lymph nodes, appendicular skeleton, and liver. For DLBCL treatment, the patient received denosumab and six cycles of rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone. Shortly after completing therapy, he presented with dizziness, ataxia, and memory loss. Brain MRI showed two ring-enhancing lesions consistent with metastasis, one in the right temporal lobe and the other in the left posterior fossa adjacent to the fourth ventricle. He underwent subsequent whole brain radiation therapy with 3960 cGy delivered in 22 fractions as well as intrathecal cytarabine.

In addition to his oncologic history, his past medical history was notable for coronary artery disease, hypertension, hypothyroidism, gastroesophageal reflux disease, and mini-stroke. His surgical history was remarkable coronary artery bypass grafting × 4 vessels, right carotid endarterectomy, appendectomy, and aortic valve replacement. Family history of cancer included a maternal grandfather with lung cancer, a maternal uncle with colon cancer, and a paternal grandfather with lung cancer. There were no first degree relatives with a history of cancer, heart disease, alcoholism, asthma, bleeding disorder, diabetes, or thyroid disease. Social history was remarkable for a 32-pack year smoking history, asbestos exposure, and no alcohol or illicit drug use.

Physical examination

Physical exam was notable for bilateral dense fibrosis.

Laboratory testing

Laboratory diagnosis was deferred, as definitive pathological diagnosis was available.

Imaging examination

MRI and PET did not show evidence of visible residual local EMC.

Genetic testing

Genetic testing was deferred.

FINAL DIAGNOSIS

Given the piecemeal manner in which the cancer was removed, a multidisciplinary tumor board concluded that the patient required revision surgery with additional neck dissection for concerning nodes unaddressed by the original surgery. The patient proceeded with resection of the left submandibular gland with dissection of level IA, IB, and suprahyoid neck nodes. Histologic examination revealed EMC in 1/2 left level IB lymph nodes and 0/3 left level IA lymph nodes staged as pT2pN1M0. The pathology showed extensive lymphovascular invasion and positive margins on the primary mass (Figure 2). Based on the high probability of residual microscopic disease, the patient elected to proceed with adjuvant chemoradiation.

TREATMENT

For adjuvant treatment of EMC, the patient's left hemineck received intensity-modulated radiation therapy consisting of 5040 cGy in 28 fractions, followed by a 3D-boost of 1620 cGy in 9 fractions to the left submandibular tumor bed and adjacent



Figure 1 Primary epithelial-myoepithelial carcinoma presenting computerized tomography scan.

Computerized tomography scan showing heterogeneous mass determined to be primary epithelial-myoepithelial carcinoma in the left submandibular gland.

level IB lymph node region. The cumulative dose totaled 6660 cGy in 37 fractions. Concurrent chemotherapy consisted of weekly carboplatin and paclitaxel. After completing chemoradiotherapy, the patient showed no evidence of EMC disease on history, physical exam, and CT imaging. At this time, he reported some sequelae of treatment including xerostomia and fibrosis of the neck tissues.

OUTCOME AND FOLLOW-UP

While awaiting chemoradiation for EMC, at the age of 56, a restaging PET scan demonstrated mild to moderate $2\text{-}^{18}\text{F}$ -fluoro-deoxy-D-glucose (FDG) uptake in an irregular nodular density measuring $1.7\text{ cm} \times 1.3\text{ cm}$ in the right upper lobe of the lung (Figure 3). Comparison to a previous CT scan showed interval growth of the nodule from $1.4\text{ cm} \times 1.0\text{ cm}$. Bronchoscopy with biopsy revealed a neuroendocrine tumor (NET). Immunopanel showed the tumor to be positive for CAM 5.2, TTF1, synaptophysin, and chromogranin. A Ki67 immunoperoxidase stain exhibited a low proliferation rate of less than 3%, indicative of a low-grade neoplasm. Notably, the patient denied any cough, shortness of breath, diarrhea, flushing, or weight loss, and consequently elected for observation with close monitoring.

The PET scan also demonstrated an intensely FDG focus measuring $0.6\text{ cm} \times 0.7\text{ cm}$ in the anterior medial aspect of the left thyroid lobe (Figure 4). Thyroid ultrasound with Doppler displayed an oblong mildly hypoechoic nodule with peripheral and internal vascularity. Fine needle aspiration revealed a follicular lesion of undetermined significance composed predominantly of Hurthle cells. Given the benign nodular hyperplasia of the thyroid nodule compared to the micrometastatic EMC, the patient elected to first proceed with chemoradiation for EMC followed by hemithyroidectomy for the Hurthle-cell adenomatoid nodule.

Ten months post-chemoradiotherapy, a follow-up CT scan displayed interval growth of the right upper lobe NET along with several new bilateral pulmonary nodules. PET scan noted a new intensely FDG avid focus in the left pterygoid muscle along with intensely avid hilar lymphadenopathy and moderately avid mediastinal lymph nodes. Biopsy of the pulmonary nodules and mediastinal lymph nodes was negative for malignancy. Based on the interval growth of the NET, the patient elected for stereotactic body radiation therapy (SBRT) to the right lung. This lesion received a total dose of 5250 cGy over 5 fractions with 1 isocenter and 2 VMAT beams. In each fraction, 1050 cGy covered 95% of the planned treatment volume (PTV) while 90% of the prescription dose covered 99% of the PTV.

Two months after completing SBRT, the patient presented to the emergency department with epistaxis and a supratherapeutic international normalized ratio secondary to warfarin. A fall while hospitalized led to altered mental status and hypoxic respiratory failure, prompting transfer to the intensive care unit and intubation. He subsequently developed pneumonia and sepsis with a vasopressor requirement as well as bilateral pleural effusions requiring repeated therapeutic

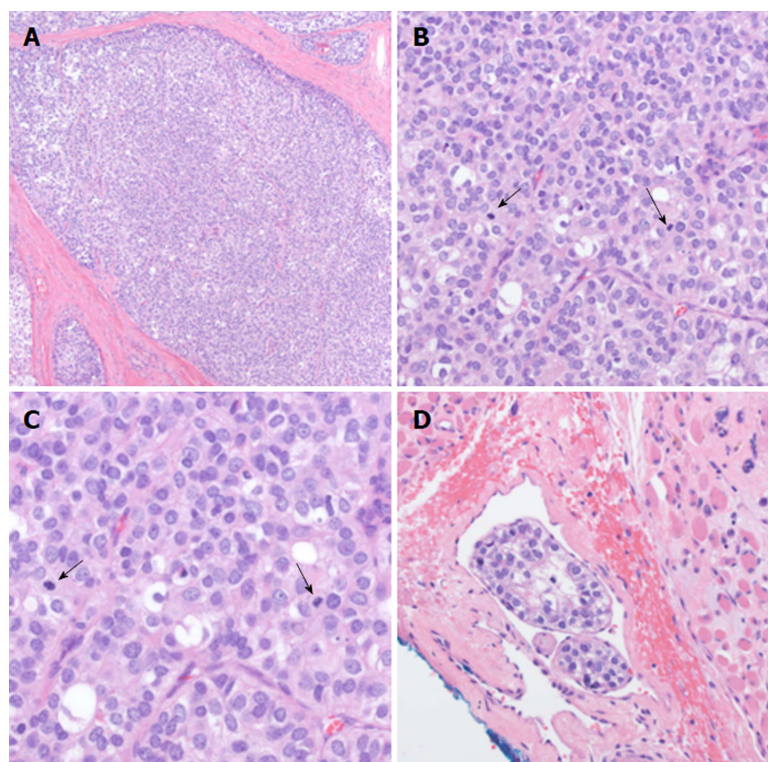


Figure 2 Hematoxylin and eosin stain of epithelial-myoepithelial carcinoma specimen. A: Epithelial-myoepithelial carcinoma (EMC), 10 × magnification; B: EMC, 40 × magnification; C: EMC, 60 × magnification; D: Lymphovascular invasion, 40 × magnification. Arrows indicate mitotic figures.

thoracentesis. Flow cytometry of the pleural fluid was notable for a CD10-positive lambda-skewed B cell population suspicious for a small B cell lymphoma *vs* follicular lymphoma, with a negative peripheral blood flow cytometry. After discussion with his family regarding goals of care, he transferred to the palliative care unit and subsequently inpatient hospice, where he expired shortly thereafter.

DISCUSSION

Secondary malignancy after Hodgkin lymphoma is not an infrequent occurrence. According to Schaapvel and co-authors, even 40 years after treatment, survivors of Hodgkin lymphoma are at an increased risk for secondary cancers, with the cumulative incidence approaching 48.5%^[10]. Additionally, a recent retrospective study by Patel *et al.* investigating 1541 stage I and stage II Hodgkin lymphoma patients treated from 1968 to 2007 revealed that, after median follow-up of 15.2 years (35% of patients with > 20 years of follow up), 395 patients had died of all causes, 85 patients had died from Hodgkin lymphoma, 168 had died from secondary malignancies (25 hematologic and 143 nonhematologic), 70 had died from cardiovascular causes, and 21 had died from pulmonary causes^[11]. Remarkably, as these data show, more patients died from secondary malignancies than from the primary Hodgkin lymphoma.

The patient presented in this case report developed multiple malignancies, including DLBCL, EMC, Hurthle cell adenoma, low grade NET, and small B cell lymphoma after chemotherapy and RT of Hodgkin lymphoma three decades prior. Of these 4 malignancies, EMC is the most likely to be secondary to RT in this patient. Its location in left submandibular gland is within the previous mantle field RT treatment area, fulfilling Cahan's criteria for RT-associated secondary malignancy^[12]. Additionally, the pathology of this patient's EMC was atypical. While characteristically a low-grade, indolent tumor, this EMC was an aggressive, invasive malignancy, in keeping with the only previously reported case of RT-associated EMC^[6]. Other RT-induced secondary malignancies, including glioma, sarcoma, and meningioma, are more often higher grade and more aggressive than their sporadic counterparts^[13-15]. Regarding the other malignant entities in this patient, including DLBCL, Hurthle cell adenoma, and low-grade NET, these may be secondary to RT for Hodgkin lymphoma, as they do fulfill Cahan's criteria. Thyroid malignancies secondary to RT including Hurthle cell adenoma are well-described, and there are

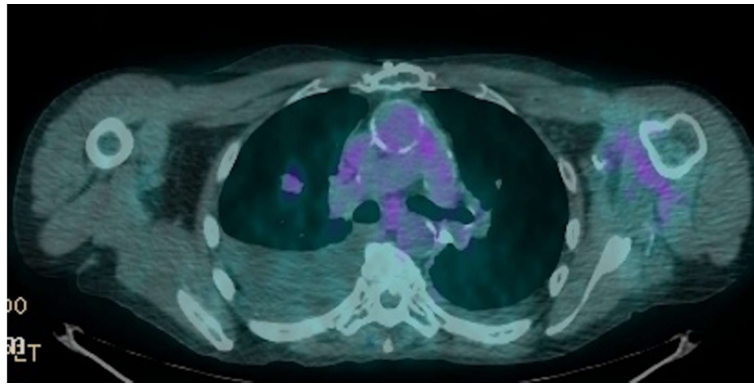


Figure 3 Neuroendocrine tumor. Positron emission tomography/computerized tomography chest scan showing moderately $2\text{-}^{18}\text{F}$ -fluoro-deoxy-D-glucose avid nodule of the right upper lobe.

reports of DLBCL secondary to radiation^[16-18]. A case of quadruple neoplasms following RT therapy has also been published^[19]. While family history was negative for first degree relatives with cancer, this patient may have had an unstable genomic background more susceptible to mutagenic insult that, in combination with a strong tobacco history, resulted in a lower threshold for carcinogenesis secondary to mantle field RT.

Importantly, clinical management guidelines of DLBCL, Hurthle cell adenoma, NET, and small B cell lymphoma are well-described; however, EMC, a rare entity, is less well-understood, especially even rarer high grade variants. EMC is most typically treated with surgery, although local recurrence occurs in one-fourth to one-half of all cases^[3]. To risk stratify EMC based on pathological features, a retrospective study evaluated 61 tumors and reported positive margin status, angiolymphatic invasion, tumor necrosis, and myoepithelial anaplasia as significantly associated with shortened disease free survival^[20]. This patient with positive margins, lymphovascular invasion, and nodal involvement was certainly at an elevated risk for recurrence. However, very few data have been reported on treatment paradigms for high risk patients, and the decision to pursue adjuvant chemoradiotherapy in this patient was in part based on evidence extrapolated from other salivary gland tumors. The largest study of EMC retrospectively analyzed the Surveillance, Epidemiology, and End Result (SEER) database and did not find a disease specific survival benefit for patients receiving adjuvant RT^[21]. However, such data are more applicable to classic quiescent EMC than higher grade or RT-associated EMC. RT has been included in treatment plans since WHO inclusion in 1991, and some authors advocate for a refined incorporation of adjuvant RT in the setting of recurrent or high risk EMC^[22,23]. Metastatic cases, while uncommon, have a poor prognosis when observed without chemoradiotherapy^[24]. Finally, a biological basis may exist for an enhanced response to RT for more aggressive cancers compared to more indolent tumors^[25].

In summary, given the scarcity of information about EMC after Hodgkin lymphoma, this case report provides vital information regarding the pathology, clinical sequelae, and treatment of EMC after Hodgkin lymphoma. Furthermore, this case provides a unique illustration of a quintet of malignancies following RT and chemotherapy for Hodgkin lymphoma. Patients with a history of treated Hodgkin lymphoma merit long-term follow-up due to increased risk of secondary malignancy. Further investigation is needed on the efficacy of adjuvant RT in EMC, especially atypical EMC.

EXPERIENCES AND LESSONS

Patients with a history Hodgkin lymphoma treated with mantle field RT are at risk for secondary malignancies. This includes on rare occasions EMC. We illustrate that radiation-associated EMC can be treated with resection and adjuvant chemoradiation.

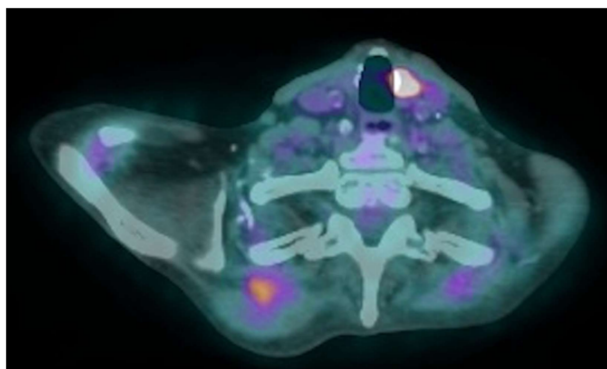


Figure 4 Hurthle cell adenoma. Positron emission tomography/computerized tomography scan neck scan showing intensely 2-¹⁸F-fluoro-deoxy-D-glucose avid focus in the left medial thyroid.

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REFERENCES

- 1 Donath K, Seifert G, Schmitz R. [Diagnosis and ultrastructure of the tubular carcinoma of salivary gland ducts. Epithelial-myoepithelial carcinoma of the intercalated ducts]. *Virchows Arch A Pathol Pathol Anat* 1972; **356**: 16-31 [PMID: [4340536](#) DOI: [10.1007/BF00543554](#)]
- 2 Seifert G, Sobin LH. Histologic typing of salivary gland tumours. World Health Organization International Histological Classification of Tumours. Berlin: Springer-Verlag; 1991; 23-24 [DOI: [10.1007/978-3-642-84506-2](#)]
- 3 Politi M, Robiony M, Avellini C, Orsaria M. Epithelial-myoepithelial carcinoma of the parotid gland: Clinicopathological aspect, diagnosis and surgical consideration. *Ann Maxillofac Surg* 2014; **4**: 99-102 [PMID: [24987609](#) DOI: [10.4103/2231-0746.133085](#)]
- 4 Li B, Yang H, Hong X, Wang Y, Wang F. Epithelial-myoepithelial carcinoma with high-grade transformation of parotid gland: A case report and literature review. *Medicine (Baltimore)* 2017; **96**: e8988 [PMID: [29245272](#) DOI: [10.1097/MD.00000000000008988](#)]
- 5 Yamazaki H, Ota Y, Aoki T, Kaneko A. Lung metastases of epithelial-myoepithelial carcinoma of the parotid gland successfully treated with chemotherapy: a case report. *J Oral Maxillofac Surg* 2013; **71**: 220-226 [PMID: [22695014](#) DOI: [10.1016/j.joms.2012.03.031](#)]
- 6 Mann JM, Kellman RM, Hahn SS, de la Roza GL, Gajra A. Radiation-induced epithelial-myoepithelial carcinoma in a patient previously treated with mantle-field radiation therapy for Hodgkin lymphoma. *Head Neck* 2015; **37**: E96-E98 [PMID: [25242451](#) DOI: [10.1002/hed.23873](#)]
- 7 Witkowska M, Majchrzak A, Smolewski P. The role of radiotherapy in Hodgkin's lymphoma: what has been achieved during the last 50 years? *Biomed Res Int* 2015; **2015**: 485071 [PMID: [25705661](#) DOI: [10.1155/2015/485071](#)]
- 8 De Bruin ML, Sparidans J, van't Veer MB, Noordijk EM, Louwman MW, Zijlstra JM, van den Berg H, Russell NS, Broeks A, Baaijens MH. Breast cancer risk in female survivors of Hodgkin's lymphoma: lower risk after smaller radiation volumes. *J Clin Oncol* 2009; **27**: 4239-4246 [PMID: [19667275](#) DOI: [10.1200/JCO.2008.19.9174](#)]
- 9 Maraldo MV, Brodin NP, Vogelius IR, Aznar MC, Munck Af Rosenschöld P, Petersen PM, Specht L. Risk of developing cardiovascular disease after involved node radiotherapy versus mantle field for Hodgkin lymphoma. *Int J Radiat Oncol Biol Phys* 2012; **83**: 1232-1237 [PMID: [22270170](#) DOI: [10.1016/j.ijrobp.2011.09.020](#)]
- 10 Schaapveld M, Aleman BM, van Eggermond AM, Janus CP, Krol AD, van der Maazen RW, Roesink J, Raemaekers JM, de Boer JP, Zijlstra JM. Second Cancer Risk Up to 40 Years after Treatment for Hodgkin's Lymphoma. *N Engl J Med* 2015; **373**: 2499-2511 [PMID: [26699166](#) DOI: [10.1056/NEJMoa1505949](#)]
- 11 Patel CG, Michaelson E, Chen YH, Silver B, Marcus KJ, Stevenson MA, Mauch PM, Ng AK. Reduced Mortality Risk in the Recent Era in Early-Stage Hodgkin Lymphoma Patients Treated With Radiation Therapy With or Without Chemotherapy. *Int J Radiat Oncol Biol Phys* 2018; **100**: 498-506 [PMID: [29153331](#) DOI: [10.1016/j.ijrobp.2017.09.048](#)]
- 12 Cahan WG, Woodard HQ, Higinbotham NL, Stewart FW, Coley BL. Sarcoma arising in irradiated bone: report of eleven cases. 1948. *Cancer* 1998; **82**: 8-34 [PMID: [9428476](#) DOI: [10.1002/\(SICI\)1097-0142\(19980101\)82:1<8::AID-CNCR3>3.0.CO;2-W](#)]
- 13 Rosko AJ, Birkeland AC, Chinn SB, Shuman AG, Prince ME, Patel RM, McHugh JB, Spector ME. Survival and Margin Status in Head and Neck Radiation-Induced Sarcomas and De Novo Sarcomas. *Otolaryngol Head Neck Surg* 2017; **157**: 252-259 [PMID: [28397585](#) DOI: [10.1177/0194599817700389](#)]
- 14 Yamanaka R, Hayano A, Kanayama T. Radiation-Induced Meningiomas: An Exhaustive Review of the Literature. *World Neurosurg* 2017; **97**: 635-644.e8 [PMID: [27713063](#) DOI: [10.1016/j.wneu.2016.09.094](#)]
- 15 Ng I, Tan CL, Yeo TT, Vellayappan B. Rapidly Fatal Radiation-induced Glioblastoma. *Cureus* 2017; **9**: e1336 [PMID: [28706761](#) DOI: [10.7759/cureus.1336](#)]
- 16 Arganini M, Behar R, Wu TC, Straus F 2nd, McCormick M, DeGroot LJ, Kaplan EL. Hürthle cell

- tumors: a twenty-five-year experience. *Surgery* 1986; **100**: 1108-1115 [PMID: 3787466]
- 17 **Chaudhuri AA**, Xavier MF. Primary cutaneous diffuse large B cell lymphoma, leg type (PCDLBCL-LT) in the setting of prior radiation therapy. *J Gen Intern Med* 2015; **30**: 371-372 [PMID: 25326162 DOI: 10.1007/s11606-014-3047-y]
- 18 **Papanastasiou L**, Pappa T, Dasou A, Kyrodinou E, Kontogeorgos G, Samara C, Bacaracos P, Galanopoulos A, Piaditis G. Case report: Primary pituitary non-Hodgkin's lymphoma developed following surgery and radiation of a pituitary macroadenoma. *Hormones (Athens)* 2012; **11**: 488-494 [PMID: 23422773 DOI: 10.14310/horm.2002.1382]
- 19 **Szymanski LJ**, Sibug Saber ME, Kim JW, Go JL, Zada G, Rao N, Hurth KM. Quadruple Neoplasms following Radiation Therapy for Congenital Bilateral Retinoblastoma. *Ocul Oncol Pathol* 2017; **4**: 33-37 [PMID: 29344496 DOI: 10.1159/000477410]
- 20 **Seethala RR**, Barnes EL, Hunt JL. Epithelial-myoepithelial carcinoma: a review of the clinicopathologic spectrum and immunophenotypic characteristics in 61 tumors of the salivary glands and upper aerodigestive tract. *Am J Surg Pathol* 2007; **31**: 44-57 [PMID: 17197918 DOI: 10.1097/01.pas.0000213314.74423.d8]
- 21 **Vázquez A**, Patel TD, D'Aguillo CM, Abdou RY, Farver W, Baredes S, Eloy JA, Park RC. Epithelial-Myoepithelial Carcinoma of the Salivary Glands: An Analysis of 246 Cases. *Otolaryngol Head Neck Surg* 2015; **153**: 569-574 [PMID: 26195572 DOI: 10.1177/0194599815594788]
- 22 **Nguyen S**, Perron M, Nadeau S, Odashiro AN, Corriveau MN. Epithelial Myoepithelial Carcinoma of the Nasal Cavity: Clinical, Histopathological, and Immunohistochemical Distinction of a Case Report. *Int J Surg Pathol* 2018; **26**: 342-346 [PMID: 29237344 DOI: 10.1177/1066896917747732]
- 23 **Simpson RH**, Clarke TJ, Sarsfield PT, Gluckman PG. Epithelial-myoepithelial carcinoma of salivary glands. *J Clin Pathol* 1991; **44**: 419-423 [PMID: 2045502 DOI: 10.1136/jcp.44.5.419]
- 24 **Chen MY**, Vyas V, Sommerville R. Epithelial-Myoepithelial Carcinoma of the Base of Tongue with Possible Lung Metastases. *Case Rep Otolaryngol* 2017; **2017**: 4973573 [PMID: 29085691 DOI: 10.1155/2017/4973573]
- 25 **Baskar R**, Dai J, Wenlong N, Yeo R, Yeoh KW. Biological response of cancer cells to radiation treatment. *Front Mol Biosci* 2014; **1**: 24 [PMID: 25988165 DOI: 10.3389/fmolb.2014.00024]

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