

# World Journal of *Clinical Infectious Diseases*

*World J Clin Infect Dis* 2019 May 21; 9(1): 1-10



**MINIREVIEWES**

- 1 Treatments and limitations for methicillin-resistant *Staphylococcus aureus*: A review of current literature  
*Kashyap R, Shah A, Dutt T, Wieruszewski PM, Ahdal J, Jain R*

**ABOUT COVER**

Editor-in-Chief of *World Journal of Clinical Infectious Diseases* Joao Mesquita, DVM, MSc, PhD, Professor, Epidemiology Reseach Unit, Polytechnic Institute of Viseu; Public Health Institute of the University of Porto, Porto 3500-606, Portugal

**AIMS AND SCOPE**

*World Journal of Clinical Infectious Diseases (World J Clin Infect Dis, WJCID, online ISSN 2220-3176, DOI: 10.5495)* is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

The *WJCID* covers a variety of clinical medical topics, including community-acquired infections, cross infection, eye infections, focal infection, gingivitis, infectious, infectious, intraabdominal infections, laboratory infection, ludwig's angina, necrotizing ulcerative, opportunistic infections, pelvic infection, pregnancy complications, etc.

We encourage authors to submit their manuscripts to *WJCID*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

**INDEXING/ABSTRACTING**

*World Journal of Clinical Infectious Diseases* is now indexed in China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Responsible Electronic Editor: *Yan-Xia Xing* Proofing Editorial Office Director: *Ya-Juan Ma*

**NAME OF JOURNAL**

*World Journal of Clinical Infectious Diseases*

**ISSN**

ISSN 2220-3176 (online)

**LAUNCH DATE**

December 30, 2011

**FREQUENCY**

Irregular

**EDITORS-IN-CHIEF**

Joao Mesquita, Caterina Sagnelli

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/2220-3176/editorialboard.htm>

**EDITORIAL OFFICE**

Ya-Juan Ma, Director

**PUBLICATION DATE**

May 21, 2019

**COPYRIGHT**

© 2019 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Treatments and limitations for methicillin-resistant *Staphylococcus aureus*: A review of current literature

Rahul Kashyap, Aditya Shah, Taru Dutt, Patrick M Wieruszewski, Jaishid Ahdal, Rishi Jain

**ORCID number:** Rahul Kashyap (0000-0002-4383-3411); Aditya Shah (0000-0003-1705-3558); Taru Dutt (0000-0002-2023-394X); Patrick M Wieruszewski (0000-0002-5871-5186); Jaishid Ahdal (0000-0002-9400-9975); Rishi Jain (0000-0003-2716-1499).

**Author contributions:** Kashyap R, Shah A, Dutt T and Wieruszewski SM conceptualized the review; Ahdal J and Jain R performed the initial literature search and procured the required literature for the review; Kashyap R, Ahdal J and Jain R prepared the initial manuscript draft; Shah A, Dutt T and Wieruszewski SM performed initial review of the manuscript and finalized the contents; Kashyap R, Shah A, Dutt T, Wieruszewski SM, Ahdal J and Jain R individually reviewed the final draft and approved the same.

**Conflict-of-interest statement:**

Authors Rahul Kashyap, Aditya Shah, Taru Dutt, and Patrick M. Wieruszewski have nothing to declare. Authors Jaishid Ahdal and Rishi Jain are salaried employees of the Wockhardt Ltd, BKC, Mumbai, India.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works

**Rahul Kashyap**, Department of Anesthesiology and Perioperative Medicine, Mayo Clinic, Rochester, MN 55902, United States

**Aditya Shah**, Department of Infectious Diseases, Mayo Clinic, Rochester, MN 55902, United States

**Taru Dutt**, Neurology Research, Mayo Clinic, Rochester, MN 55902, United States

**Patrick M Wieruszewski**, Department of Pharmacy, Critical Care Medicine, Mayo Clinic, Rochester, MN 55902, United States

**Jaishid Ahdal, Rishi Jain**, Workhardt Limited, Bandra East, Mumbai, Maharashtra 400051, India

**Corresponding author:** Rahul Kashyap, MBBS, Assistant Professor, Department of Anesthesiology and Perioperative Medicine, Mayo Clinic, 200, First Street, SW Rochester, Rochester, MN 55902, United States. [kashyap.rahul@mayo.edu](mailto:kashyap.rahul@mayo.edu)

**Telephone:** +1-507-2557196

**Fax:** +1-507-2554267

### Abstract

Methicillin-resistant *Staphylococcus aureus* (MRSA) has remained a major threat to healthcare; in both hospital and community settings over the past five decades. With the current use of antibiotics for a variety of infections, including MRSA, emerging resistance is a major concern. Currently available treatments have restrictions limiting their use. These issues include, but are not limited to, side effects, cross-resistance, lack of understanding of pharmacokinetics and clinical pharmacodynamics, gradual increment in minimal inhibitory concentration over the period (MIC creep) and ineffectiveness in dealing with bacterial biofilms. Despite availability of various therapeutic options for MRSA, the clinical cure rates remain low with high morbidity and mortality. Given these challenges with existing treatments, there is a need for development of novel agents for MRSA. Along with prompt infection control strategies and strict implementation of antibiotic stewardship, cautious use of newer anti-MRSA agents will be of utmost importance. This article reviews the treatments and limitations of MRSA management and highlights the future path.

**Key words:** Methicillin resistant; Methicillin-resistant *Staphylococcus aureus*; Antibiotics; Monotherapy

on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Unsolicited manuscript

**Received:** December 31, 2018

**Peer-review started:** January 3, 2019

**First decision:** March 15, 2019

**Revised:** March 29, 2019

**Accepted:** April 8, 2019

**Article in press:** April 9, 2019

**Published online:** May 21, 2019

**P-Reviewer:** García-Elorriaga G, Liu L

**S-Editor:** Dou Y

**L-Editor:** A

**E-Editor:** Xing YX



©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** Methicillin-resistant *S. aureus* (MRSA) remains a major threat despite availability of multiple treatments. Limitations of the current anti-MRSA treatments demand more careful use of these agents. Using antibiotics in combination for MRSA treatment needs further evaluation. Multiple strategies including research and development of new antibiotics and antibiotic stewardship are necessary to contain the MRSA.

**Citation:** Kashyap R, Shah A, Dutt T, Wieruszewski PM, Ahdal J, Jain R. Treatments and limitations for methicillin-resistant *Staphylococcus aureus*: A review of current literature. *World J Clin Infect Dis* 2019; 9(1): 1-10

**URL:** <https://www.wjgnet.com/2220-3176/full/v9/i1/1.htm>

**DOI:** <https://dx.doi.org/10.5495/wjcid.v9.i1.1>

## INTRODUCTION

*Staphylococci* have been involved in human disease for centuries and were identified first as the cause of incurable boils. Sir Alexander Ogston and Friedrich J Rosenbach identified, classified, and contributed to the nomenclature of *Staphylococci*<sup>[1]</sup>. *S. aureus* has since evolved as a major infectious pathogen being severely detrimental to the health of millions of patients. *S. aureus* possesses resistance mechanisms to standard agents. The first incidence of penicillin resistance was reported in 1942 which was identified to be due to inducible beta-lactamase. After introduction of methicillin in 1959, methicillin-resistant *S. aureus* (MRSA) was reported in 1961<sup>[2]</sup>.

Burden of MRSA is high in middle-income countries like India. Amongst all *S. aureus* isolates, Indian Network for Surveillance of Antimicrobial Resistance group reported methicillin resistance in 41% of their isolates<sup>[3]</sup>. This high burden of MRSA in India is the cause of significant morbidity and mortality. Additionally, formation of biofilms in MRSA isolates is associated with increased virulence, pose a challenge in clinical management, and may also contribute to the development of resistance<sup>[4,5]</sup>.

Current treatment strategies have limitations and improper source control may add to that, especially in severe MRSA infections. Thus, we aim to review the current treatment strategies, their limitations, and a way forward for effective management of MRSA infections.

## CURRENT TREATMENT RECOMMENDATIONS FOR MRSA INFECTIONS

MRSA infections involve a wide disease spectrum. Common sites include skin/soft tissue, bone/joint, vascular line, native valve/prosthetic valve endocarditis, central nervous system shunt infections and meningitis/brain abscesses. The Infectious Disease Society of America (IDSA) provides treatment recommendations for MRSA infections<sup>[6]</sup> (Table 1).

### Vancomycin dosing in MRSA

Vancomycin is one of the mainstays of therapy for MRSA infections. In adults, IV vancomycin at a dose of 15-20 mg/kg/dose (max 2 g/dose) every 8-12 h based on renal function is recommended with a loading dose of 25-30 mg/kg in seriously ill patients<sup>[7]</sup>. Therapeutic drug monitoring (TDM) is recommended to ensure adequacy of dosing, with most infections necessitating trough concentrations of 10-20 µg/mL, with concentrations at the higher end of this range (*i.e.*, 15-20 µg/mL) reserved for difficult to penetrate sites such as pulmonary and central nervous system. However, in skin and skin structure infections (SSTIs), trough monitoring may not be necessary and vancomycin in a dose of 1 mg every 12 h may be adequate<sup>[6]</sup>.

## LIMITATIONS OF CURRENT TREATMENTS: MONOTHERAPY

**Table 1** Methicillin-resistant *S. aureus* treatment recommendations<sup>[6]</sup>

Infections	Antibiotic Treatment
<b>Skin and soft tissue infections (SSTIs)</b>	
Uncomplicated SSTIs	Clindamycin, trimethoprim-sulfamethoxazole (TMP-SMX), a tetracycline (doxycycline or minocycline) (A-II), linezolid
Complicated SSTIs	IV Vancomycin, Linezolid (oral or IV 600 mg twice daily), Daptomycin (4 mg/kg/dose IV once daily), Telavancin (10 mg/kg/dose IV once daily), Clindamycin (600 mg IV or PO 3 times a day)
Recurrent SSTIs	Nasal decolonization - mupirocin twice daily +/- topical body decolonization - skin antiseptic solution (e.g. chlorhexidine) or dilute bleach baths.
<b>Bacteraemia and infective endocarditis</b>	
Native valve endocarditis	Vancomycin; Daptomycin (6 mg/kg/dose IV once daily)
Prosthetic valve endocarditis	Vancomycin + Rifampin (300 mg PO/IV every 8 hour) followed by Gentamicin (1 mg/kg/dose IV every 8 hour)
<b>Pneumonia</b>	
Community acquired, or healthcare associated	IV vancomycin or linezolid (600 mg PO/IV twice daily) or clindamycin (600 mg PO/IV 3 times daily)
<b>Bone and joint infections</b>	
Osteomyelitis or Septic arthritis	Vancomycin; Daptomycin (6 mg/kg/dose IV once daily); TMP-SMX [4 mg/kg/dose (TMP component) twice daily] + Rifampin (600 mg once daily)
Device-related osteo-articular infections (early onset < 2 mo - prosthetic joint infections)	Vancomycin or Daptomycin (6 mg/kg/dose IV once daily) + Rifampin (600 mg once daily) followed by; Rifampin + fluoroquinolone / TMP-SMX / tetracycline / clindamycin
Device-related osteo-articular infections (early onset < 2 mo - spinal implant infections)	Initial parenteral therapy + Rifampin followed by prolonged oral therapy
<b>CNS infections</b>	
Meningitis, Brain abscess, subdural empyema, spinal epidural abscess, Septic Thrombosis of Cavernous or Dural Venous Sinus	IV Vancomycin +/- Rifampin; OR; Linezolid 600 mg PO/IV twice daily or TMP-SMX 5 mg/kg/dose IV every 8-12 hour

SSTIs: Skin and soft tissue infections; TMP-SMX: Trimethoprim-sulfamethoxazole; PO: Per oral; IV: Intravenous; CNS: Central nervous system.

An ideal anti-MRSA agent does not exist; desirable properties in anti-MRSA antibiotics include rapid bactericidal action, excellent penetration in tissue, consistent and predictable pharmacokinetics to support reliable dosing, low probability of resistance development, lower risk of side effects, and good microbiological and clinical cure rates. Biofilm formation with *S. aureus* is known and contributes to antibacterial tolerance by promoting bacterial persistence in biofilms.

Thus, identifying an ideal antibiotic which will also be active against biofilms can be a challenge. **Table 2** enumerates some of the limitations of major existing anti-MRSA treatments.

### Vancomycin monotherapy

Over the years of vancomycin use, resistance is now beginning to emerge in MRSA isolates<sup>[8]</sup>. Vancomycin has several limitations. First is the ratio of minimum bactericidal to inhibitory concentration (MBC: MIC ratio). A study from Sader *et al*<sup>[9]</sup> demonstrated that 20.1% of tested MRSA strains ( $n = 900$ ) were vancomycin tolerant defined by MBC: MIC ratio of  $\geq 32$ . This varied from 10.0% to 43.0% among different centres evaluated<sup>[9]</sup>. Secondly, the accessory gene regulator pathway is associated with regulation of quorum sensing and endotoxin production<sup>[10]</sup>. Development of polymorphisms or loss of function of accessory gene regulator (*agr*) pathway is associated with failure of vancomycin therapy<sup>[11]</sup>. Thirdly, the "MIC creep" phenomenon wherein there is a gradual reduction in susceptibility of *S. aureus* to vancomycin despite concentrations in the susceptible range ( $\leq 2$  mg/L) can develop with continued use of vancomycin<sup>[8]</sup>. A study from California by Wang *et al*<sup>[12]</sup>, demonstrated a gradual shift of MIC from  $\leq 0.5$  to 1.0  $\mu\text{g}/\text{mL}$  over 5 years to vancomycin in MRSA strains ( $n = 6002$ ). The proportion of isolates with MIC 1  $\mu\text{g}/\text{mL}$  increased from 19.9% to 70.4% over study duration (**Figure 1**). Fourth concern is development of hetero-resistance to vancomycin (hVISA). In this phenomenon, from among the isolated MRSA, a subpopulation demonstrates intermediate level of vancomycin resistance, but the colony as a whole remains susceptible. The mechanisms for this remains unclear but may involve thickening of cell wall avoiding penetration of vancomycin, and alteration in *agr* pathway<sup>[10]</sup>. A study from Sader *et al*<sup>[9]</sup> involving nine hospitals in the United States showed hVISA prevalence of 13.4%. The development of hVISA was more common (45.6%) in MRSA isolates with MIC  $\geq 1$  mg/L. Fifth, the extensive protein binding of vancomycin leads to variable tissue

**Table 2** Limitations of current anti-methicillin resistant *S. aureus* treatments

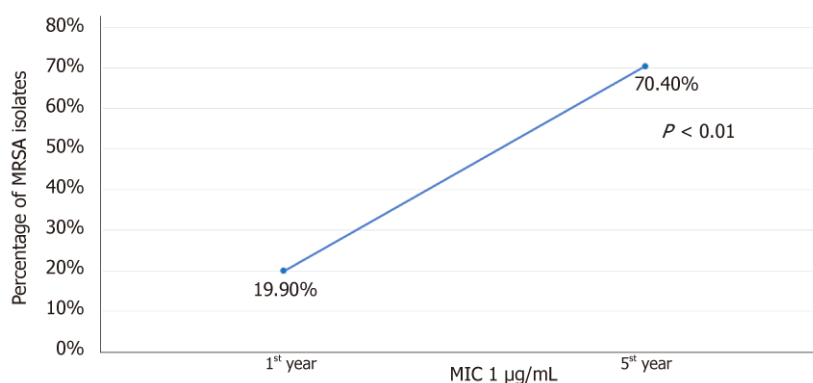
Treatment	Limitations
Vancomycin	Higher MBC: MIC ratio Polymorphisms or changes in gene function (e.g. <i>agr</i> pathway) MIC creep Development of hetero-resistance (hVISA) Variable tissue penetration AUC: MIC ratio Nephrotoxicity Red man syndrome
Teicoplanin	Therapeutic drug monitoring may be necessary Need to generate evidence on pharmacokinetics and clinical pharmacodynamics
Daptomycin	Resistance development Possible cross-resistance in hVISA Inactivation by alveolar surfactant
Linezolid	Serious adverse drug reactions e.g., thrombocytopenia, optic neuropathy, peripheral neuropathy, lactic acidosis, monoamine oxidase inhibition MIC creep Limited efficacy in bacteraemia or endocarditis
TMP/SMX	High degree of resistance Limited efficacy in bacteraemia Thymidine salvage in presence of pus
Clindamycin	High rates of inducible and constitutive resistance Risk of <i>Clostridium difficile</i> infection
Tetracyclines	Limited utility in severe invasive infections
Tigecycline	Low serum levels with limited efficacy in bacteraemia Poor tissue penetration and AUC: MIC ratio Black box warning from the USFDA for all-cause mortality, Mortality Imbalance and Lower Cure Rates in VAP and pancreatitis
Quinupristin/ Dalfopristin	Limiting side effects like infusion-site inflammation, pain, and oedema, thrombophlebitis, arthralgia, myalgia, nausea, diarrhoea, vomiting, and rash Drug interactions with CYP3A4 inhibitors
Ceftaroline	Risk of agranulocytosis
Telavancin	Risk of nephrotoxicity
Oritavancin and Dalbavancin	Long half-life - delayed hypersensitivity if occurs may persist for weeks Clinical failure may get unnoticed if there is lack of daily follow-up evaluations Effectiveness in bacteraemia, pneumonia, bone and joint infections, and prosthetic infections has not been established Higher occurrence of osteomyelitis reported in clinical studies with oritavancin

AUC: Area under the curve; MBC: Minimum bactericidal concentration; MIC: Minimum inhibitory concentration; TMP/SMX: Trimethoprim-Sulfamethoxazole; USFDA: United States Food and Drugs Administration; VAP: Ventilator associated pneumonia.

penetration which can further be different in comorbidities like diabetes, meningitis, *etc*<sup>[10]</sup>. Sixth, the pharmacodynamics of vancomycin has been considered to be an important aspect in determining efficacy. The area under the curve (AUC) and MIC ratio of 400 or more is believed to provide therapeutic effectiveness for which vancomycin trough concentration should reach 15-20 mg/L especially in severe MRSA infections<sup>[7]</sup>. For achieving AUC: MIC ratio of 400 or more at MIC of 1 mg/L, dose of 3-4 mg/d is necessary. For MIC of 2 mg/L, achieving target AUC: MIC ratio is not possible even when higher doses are used. This can result in poor clinical and microbiological cure.

Nephrotoxicity is an important adverse effect associated with vancomycin. The reported incidence varies from nearly 14% in children to 35% in adults. In adults, trough concentration beyond 15 µg/mL is associated with increased risk of renal injury. Attaining AUC: MIC ratio of  $\geq 400$  is therefore harmful especially wherein the





**Figure 1** Minimal inhibitory concentration creep - Proportion of MRSA isolates with vancomycin minimal inhibitory concentration of 1 µg/mL<sup>[12]</sup>.

isolate MIC is > 2 mg/L. In such cases, use of alternative agents is advised<sup>[13]</sup>.

### Daptomycin

Daptomycin, a branched cyclic anionic lipopeptide exerts bactericidal action *via* calcium-dependent modification in membrane potential causing leaking of intracellular ions and cell death<sup>[14]</sup>. It has shown similar efficacy to vancomycin in MRSA bacteraemia, endocarditis, complicated SSTIs, but not in pneumonia due to inactivation by alveolar surfactant<sup>[10]</sup>. However, point mutation in *MprF* gene (L431F substitution) identified in clinical isolates was associated with reduced negative cell membrane charge, thicker cell wall, and longer doubling time. This was found to confer increased resistance to daptomycin and vancomycin<sup>[15]</sup>. Daptomycin-non-susceptible (DAP-NS) phenotype has also been reported in MRSA infections. Among 2.4% DAP-NS strains ( $n = 208$ ), one was sequence type 72 (ST72) and other four were ST5. Three of these strains were also found to be hVISA. The resistance mechanism in ST72 was charge repulsion, ST5 showed charge independent mechanisms. Changes in cell wall thickness were not found in any of the DAP-NS strains<sup>[16]</sup>. DAP-NS isolates were not sensitive to high-dose of daptomycin<sup>[17]</sup>. Increased MIC of daptomycin was found to be associated with increased mortality in patients with MRSA bacteraemia<sup>[18]</sup>. Finally, daptomycin has been associated with elevated creatine kinase and rhabdomyolysis, which is problematic in critically ill patients already at risk of such increases and sequelae thereof, such as renal injury<sup>[19]</sup>.

### Linezolid

Linezolid, a synthetic antibiotic, binds to ribosomal RNA on both 30S and 50S subunits and thereby inhibits protein synthesis. Additionally, it inhibits formation of initiation complex and reduce the rate of translation process<sup>[20]</sup>. Occurrence of serious adverse drug reactions like thrombocytopenia, optic neuropathy, peripheral neuropathy, lactic acidosis, and potential serotonin syndrome through monoamine oxidase inhibition have important therapeutic limitations resulting in poor adherence to therapy<sup>[21]</sup>. MIC creep with linezolid similar to that of vancomycin has also been reported<sup>[22]</sup>. Being a bacteriostatic agent, its first line use in severe invasive infections especially bacteraemia and endocarditis is avoided<sup>[10]</sup>.

In persistent MRSA bacteraemia (> 7 d) despite therapy with glycopeptides like vancomycin or teicoplanin, shifting to linezolid failed to show superiority in microbiologic response, treatment success, and mortality compared to the patients who continued glycopeptides<sup>[23]</sup>.

### Limitations of other agents

**Teicoplanin:** TDM may be necessary in ascertaining the teicoplanin concentrations as daily dosages of 4 mg/kg have been reported to result in treatment failure compared to a 6 mg/kg dose. Also, trough concentrations of > 10, > 20, and > 30 mg/L have been reported to be necessary for successful treatment of *S. aureus* septicemia, MRSA endocarditis, and MRSA osteomyelitis, respectively<sup>[24]</sup>. Also, given its important role in MRSA management, there is more need to generate evidence on pharmacokinetics and clinical pharmacodynamics<sup>[24]</sup>.

**Trimethoprim-sulfamethoxazole (TMP-SMX):** In MRSA infections, its utility is limited by development of resistance and poor efficacy<sup>[25,26]</sup>. Therefore, TMP-SMX is mainly confined to treatment of uncomplicated skin and skin structure infections



from an MRSA standpoint<sup>[27]</sup>.

**Clindamycin:** Clindamycin has bacteriostatic activity and high rates of inducible and constitutive resistance, limiting its utility for MRSA infections<sup>[28,29]</sup>. Further, risk of *Clostridium difficile* infection (CDI) might deter use of clindamycin as sole agent for MRSA as duration of exposure has been identified as an important determinant of CDI<sup>[30]</sup>.

**Tetracyclines:** Tetracyclines such as doxycycline and minocycline are limited to uncomplicated SSTIs by community-acquired MRSA. Bacteriostatic activity and limited spectrum limits utility in severe invasive MRSA infections<sup>[10]</sup>.

**Fucidin (fusidic acid):** Fusidic acid inhibits bacterial protein synthesis *via* action on RNA. As a topical agent, it has been used for treatment of skin infection, though there has been recent interest in rectifying its use in combination with rifampicin for infected joint prostheses. This however has been limited by significant drug-drug interactions resulting in ineffective fusidic acid exposure<sup>[31]</sup>.

**Tigecycline:** Tigecycline has shown promise in MRSA infections equivalent to vancomycin<sup>[32]</sup>. It is effective in SSTIs and complicated intraabdominal infections<sup>[33]</sup>. However, high protein binding can result in low serum levels thereby limiting effectiveness in MRSA bacteraemia. Black box warning issued from the US Food and Drug Administration for all-cause mortality, mortality imbalance and lower cure rates in VAP and pancreatitis is a concern with tigecycline<sup>[34]</sup>.

**Quinupristin/Dalfopristin:** Quinupristin/Dalfopristin is considered among the effective agents in *Staphylococcal* infections and may be effective in MRSA bacteremia<sup>[35]</sup>. However, occurrence of side effects like infusion-site inflammation, pain, and edema, thrombophlebitis, arthralgia, myalgia, nausea, diarrhoea, vomiting, and rash limit its use. Also, inhibition of cytochrome P450 3A4 with quinupristin/dalfopristin warrants caution with use of drugs metabolized through this enzymatic pathway<sup>[36]</sup>. Interference with other drugs metabolism may result in QTc prolongation with use of quinupristin/dalfopristin.

**Ceftaroline:** Ceftaroline is an effective agent for severe MRSA infections and provides clinical cure in nearly 74% cases. The major concern with this agent is development of agranulocytosis. Prolonged therapy ( $\geq 21$  d) increases risk of leukopenia and therefore treatment with ceftaroline should be closely monitored in these situations<sup>[37]</sup>.

**Telavancin:** It is another effective agent in MRSA with resistance to vancomycin, linezolid and daptomycin. However, nephrotoxicity is an important limitation. An increased mortality has been observed in hospital or ventilator associated pneumonia<sup>[38]</sup>.

**Oritavancin and Dalbavancin:** These lipoglycopeptides have ultra-long half-life upwards of 346 h making them attractive as single-dose antibiotics. This and the inability to remove *via* dialysis, however also raises a concern as injury resulting from delayed hypersensitivity (if occurs) or other adverse effects may persist for weeks. It's effectiveness has not been established in bacteraemia, pneumonia, bone and joint infections, or prosthetic infections<sup>[39]</sup>. While these agents have potential for ambulatory infectious diseases management, particularly in areas of poor clinic access for frequent intravenous infusions, their utility in acute and critical care remains to be proven.

---

## LIMITATIONS OF CURRENT TREATMENTS: COMBINATION TREATMENTS

---

With development of resistance and limitations of individual agents discussed above, combination therapy is suggested for most severe and invasive MRSA infections. The objectives are to broaden the coverage, prevent or reduce development of resistance, improve the effectiveness of individual agents, enhance capacity to penetrate biofilms, and to reduce toxin production<sup>[40]</sup>.

### **Vancomycin + Rifampicin**

Rifampicin is bactericidal to *S. aureus*, achieves high intracellular concentration, and penetrates biofilms. A systematic review in 2008 reported that *in-vitro* findings identified with rifampicin combination did not relate to *in-vivo* findings<sup>[41]</sup>. Another review in 2013 reported limited evidence to support adjunctive use of rifampicin in MRSA infections. The increased risk of drug interactions, adverse effects with

rifampicin and development of rifampicin resistance are possibilities with use of rifampicin in combination<sup>[42]</sup>. Latter is especially important in Indian context where the rifampicin is the primary drug against tuberculous infection and burden of tuberculosis is enormous. Currently, IDSA guidelines recommend use of rifampicin in combination only in prosthetic valve endocarditis and in osteoarticular infections associated with prostheses<sup>[6]</sup>. Rifampicin should not be used as monotherapy for the treatment of MRSA infections.

#### **Vancomycin + Gentamicin**

*In vitro* studies have demonstrated increased bactericidal activity of vancomycin and animal studies have shown to shorten the duration of bacteraemia. Nephrotoxicity associated with gentamicin can add to the nephrotoxic potential of vancomycin<sup>[40]</sup>.

#### **Vancomycin + Quinupristin/Dalfopristin**

Laboratory analyses have shown synergism with this combination<sup>[10]</sup>. However, clinical evidence is restricted to case reports.

#### **Daptomycin + Rifampicin or Gentamicin**

Similar to other combination treatments, the evidence from *in-vitro* studies shows synergistic activity with this combination as well<sup>[43,44]</sup>. However, clinical evidence is restricted to case reports<sup>[45-47]</sup>. In time-kill study, addition of gentamicin rather than rifampicin has been shown to provide synergism with daptomycin<sup>[48]</sup>.

#### **Daptomycin + Beta-lactams**

With beta-lactams active against MRSA (*e.g.* ceftaroline), daptomycin has shown synergistic activity<sup>[49]</sup>. In MRSA strains from endocarditis, ceftaroline in addition to daptomycin also cleared daptomycin non-susceptible strains. Daptomycin at 6 mg/kg every 48 h was and ceftaroline at 200 mg every 12 h enhanced bacterial killing<sup>[50]</sup>. The finding from this single study demands further careful determination of optimal dosing regimen for effective utilization of active agents like ceftaroline. Another study reported rapid clearance of bacteraemia with addition of high dose nafcillin or oxacillin (2 mg IV every 4 h) to high-dose daptomycin (8-10 mg/d) in 7 cases of vancomycin and daptomycin resistant MRSA<sup>[51]</sup>. Though this points to enhanced efficacy of beta-lactams, further evaluation in prospective studies is necessary.

#### **Daptomycin + Linezolid**

An *in-vitro* study involving pharmacokinetic/pharmacodynamic model of biofilm for 3 d showed greater activity with combination of daptomycin and linezolid than either agent alone suggesting potential for biofilm associated MRSA infections<sup>[52]</sup>. However, there is lack of clinical studies to substantiate the findings of *in-vitro* studies.

#### **Linezolid/Tedizolid + Rifampicin**

In combination with rifampicin, time kill studies of linezolid did not show synergism or antagonism but linezolid prevented emergence of mutant resistance in rifampicin<sup>[53]</sup>. One major issue with this combination is that rifampicin can reduce the linezolid concentration which can be well below the MIC90 for *Staphylococci* and effect may persist longer than 3 wk even after withdrawal of rifampicin<sup>[54,55]</sup>. With tedizolid and rifampicin combination, activity is increased but synergy observed was not found to be universal<sup>[56]</sup>.

#### **Trimethoprim/Sulfamethoxazole + Rifampicin**

Poor efficacy, development of resistance and side effects and drug interactions as mentioned above in their individual discussion, render this regimen redundant.

#### **Triple antibiotic combination**

The evidence is very limited for effectiveness and utility of triple drug combinations including beta-lactams, aminoglycosides, and vancomycin, barring isolated case reports<sup>[57]</sup>.

---

## **FUTURE DIRECTIONS**

---

Despite availability of multiple treatment options for MRSA, burden of MRSA remains substantial. While choosing an effective therapeutic strategy, multiple factors play a vital role in antibiotic selection. Development of resistance with anti-MRSA antibiotics has led to the use of antibiotics in combinations. There is no concrete evidence as to decide on specific combination neither there are any comparative data with different combinations. Success of new molecules like ceftaroline, tedizolid, and

plazomicin should stimulate further research and development of new anti-MRSA therapies.

A number of anti-MRSA molecules are in different phases of development. But, to identify truly novel anti-MRSA agent that will act on new targets in the pathogen, there is need to invest further.

---

## CONCLUSION

Current therapeutic management of MRSA is mainly focused on vancomycin and it still remains an effective therapy either alone or in combination. However, development of intermediate level of resistance, MIC creep, adverse effects, and vigilant TDM have been path-blockers for the sole use of vancomycin in MRSA. At present, selecting an individual agent that can provide the best synergy and minimal adverse effects remains the frontline therapeutic option against MRSA. Stimulating and supporting new and ongoing research for development of effective anti-MRSA therapies and implementation of infection control strategies are of urgent necessity. A collaborative action from policy makers, prescribers, and consumers is essential to safeguard the judicious use of newer agents in the management of MRSA infections.

---

## ACKNOWLEDGMENTS

We are thankful to Dr. Vijay M. Katekhaye for his assistance in drafting and reviewing the manuscript.

---

## REFERENCES

- 1 **Thomer L**, Schneewind O, Missiakas D. Pathogenesis of *Staphylococcus aureus* Bloodstream Infections. *Annu Rev Pathol* 2016; **11**: 343-364 [PMID: 26925499 DOI: 10.1146/annurev-pathol-012615-044351]
- 2 **Stryjewski ME**, Corey GR. Methicillin-resistant *Staphylococcus aureus*: an evolving pathogen. *Clin Infect Dis* 2014; **58** Suppl 1: S10-S19 [PMID: 24343827 DOI: 10.1093/cid/cit613]
- 3 **Indian Network for Surveillance of Antimicrobial Resistance (INSAR) group, India**. Methicillin resistant *Staphylococcus aureus* (MRSA) in India: prevalence & susceptibility pattern. *Indian J Med Res* 2013; **137**: 363-369 [PMID: 23563381]
- 4 **Jimi S**, Miyazaki M, Takata T, Ohjimi H, Akita S, Hara S. Increased drug resistance of methicillin-resistant *Staphylococcus aureus* biofilms formed on a mouse dermal chip model. *J Med Microbiol* 2017; **66**: 542-550 [PMID: 28463660 DOI: 10.1099/jmm.0.000461]
- 5 **Mottola C**, Matias CS, Mendes JJ, Melo-Cristino J, Tavares L, Cavaco-Silva P, Oliveira M. Susceptibility patterns of *Staphylococcus aureus* biofilms in diabetic foot infections. *BMC Microbiol* 2016; **16**: 119 [PMID: 27339028 DOI: 10.1186/s12866-016-0737-0]
- 6 **Liu C**, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, Kaplan SL, Karchmer AW, Levine DP, Murray BE, J Rybak M, Talan DA, Chambers HF; Infectious Diseases Society of America. Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis* 2011; **52**: e18-e55 [PMID: 21208910 DOI: 10.1093/cid/ciq146]
- 7 **Rybak M**, Lomaestro B, Rotschafer JC, Moellering R, Craig W, Billeter M, Dalovisio JR, Levine DP. Therapeutic monitoring of vancomycin in adult patients: a consensus review of the American Society of Health-System Pharmacists, the Infectious Diseases Society of America, and the Society of Infectious Diseases Pharmacists. *Am J Health Syst Pharm* 2009; **66**: 82-98 [PMID: 19106348 DOI: 10.2146/ajhp080434]
- 8 **Deresinski S**. Counterpoint: Vancomycin and *Staphylococcus aureus*--an antibiotic enters obsolescence. *Clin Infect Dis* 2007; **44**: 1543-1548 [PMID: 17516396 DOI: 10.1086/518452]
- 9 **Sader HS**, Jones RN, Rossi KL, Rybak MJ. Occurrence of vancomycin-tolerant and heterogeneous vancomycin-intermediate strains (hVISA) among *Staphylococcus aureus* causing bloodstream infections in nine USA hospitals. *J Antimicrob Chemother* 2009; **64**: 1024-1028 [PMID: 19744978 DOI: 10.1093/jac/dkp319]
- 10 **Nguyen HM**, Graber CJ. Limitations of antibiotic options for invasive infections caused by methicillin-resistant *Staphylococcus aureus*: is combination therapy the answer? *J Antimicrob Chemother* 2010; **65**: 24-36 [PMID: 19861337 DOI: 10.1093/jac/dkp377]
- 11 **Fowler VG**, Sakoulas G, McIntyre LM, Meka VG, Arbeit RD, Cabell CH, Stryjewski ME, Eliopoulos GM, Reller LB, Corey GR, Jones T, Lucindo N, Yeaman MR, Bayer AS. Persistent bacteremia due to methicillin-resistant *Staphylococcus aureus* infection is associated with agr dysfunction and low-level in vitro resistance to thrombin-induced platelet microbicidal protein. *J Infect Dis* 2004; **190**: 1140-1149 [PMID: 15319865 DOI: 10.1086/423145]
- 12 **Wang G**, Hindler JF, Ward KW, Bruckner DA. Increased vancomycin MICs for *Staphylococcus aureus* clinical isolates from a university hospital during a 5-year period. *J Clin Microbiol* 2006; **44**: 3883-3886 [PMID: 16957043 DOI: 10.1128/JCM.01388-06]
- 13 **Patel K**, Crumby AS, Maples HD. Balancing vancomycin efficacy and nephrotoxicity: should we be aiming for trough or AUC/MIC? *Paediatr Drugs* 2015; **17**: 97-103 [PMID: 25644329 DOI: 10.1007/s40272-015-0117-5]
- 14 **Vilhena C**, Bettencourt A. Daptomycin: a review of properties, clinical use, drug delivery and resistance. *Mini Rev Med Chem* 2012; **12**: 202-209 [PMID: 22356191 DOI: 10.2174/1389557511209030202]

- 15 **Chen FJ**, Lauderdale TL, Lee CH, Hsu YC, Huang IW, Hsu PC, Yang CS. Effect of a Point Mutation in *mprF* on Susceptibility to Daptomycin, Vancomycin, and Oxacillin in an MRSA Clinical Strain. *Front Microbiol* 2018; **9**: 1086 [PMID: 29887848 DOI: 10.3389/fmicb.2018.01086]
- 16 **Nam EY**, Yang SJ, Kim ES, Cho JE, Park KH, Jung SI, Yoon N, Kim DM, Lee CS, Jang HC, Park Y, Lee KS, Kwak YG, Lee JH, Park SY, Hwang JH, Kim M, Song KH, Kim HB. Emergence of Daptomycin-Nonsusceptible Methicillin-Resistant *Staphylococcus aureus* Clinical Isolates Among Daptomycin-Naive Patients in Korea. *Microb Drug Resist* 2018; **24**: 534-541 [PMID: 29863982 DOI: 10.1089/mdr.2017.0212]
- 17 **Rose WE**, Leonard SN, Rybak MJ. Evaluation of daptomycin pharmacodynamics and resistance at various dosage regimens against *Staphylococcus aureus* isolates with reduced susceptibilities to daptomycin in an in vitro pharmacodynamic model with simulated endocardial vegetations. *Antimicrob Agents Chemother* 2008; **52**: 3061-3067 [PMID: 18591272 DOI: 10.1128/AAC.00102-08]
- 18 **Ruiz J**, Ramirez P, Concha P, Salavert Lleti M, Villarreal E, Gordon M, Frassetto J, Castellanos Ortega A. Vancomycin and daptomycin minimum inhibitory concentrations as a predictor of outcome of methicillin-resistant *Staphylococcus aureus* bacteraemia. *J Glob Antimicrob Resist* 2018; **14**: 141-144 [PMID: 29601996 DOI: 10.1016/j.jgar.2018.03.007]
- 19 **Papadopoulos S**, Ball AM, Liewer SE, Martin CA, Winstead PS, Murphy BS. Rhabdomyolysis during therapy with daptomycin. *Clin Infect Dis* 2006; **42**: e108-e110 [PMID: 16705566 DOI: 10.1086/504379]
- 20 **Hashemian SMR**, Farhadi T, Ganjparvar M. Linezolid: a review of its properties, function, and use in critical care. *Drug Des Devel Ther* 2018; **12**: 1759-1767 [PMID: 29950810 DOI: 10.2147/DDDT.S164515]
- 21 **Kishor K**, Dhasmana N, Kamble SS, Sahu RK. Linezolid Induced Adverse Drug Reactions - An Update. *Curr Drug Metab* 2015; **16**: 553-559 [PMID: 26424176 DOI: 10.2174/1389200216666151001121004]
- 22 **Miyazaki M**, Nagata N, Miyazaki H, Matsuo K, Takata T, Tanihara S, Kamimura H. Linezolid minimum inhibitory concentration (MIC) creep in methicillin-resistant *Staphylococcus aureus* (MRSA) clinical isolates at a single Japanese center. *Biol Pharm Bull* 2014; **37**: 679-682 [PMID: 24694615 DOI: 10.1248/bpb.b13-00670]
- 23 **Holland TL**, Arnold C, Fowler VG. Clinical management of *Staphylococcus aureus* bacteremia: a review. *JAMA* 2014; **312**: 1330-1341 [PMID: 25268440 DOI: 10.1001/jama.2014.9743]
- 24 **Kim SW**. Is therapeutic drug monitoring of teicoplanin useful? *Infect Chemother* 2014; **46**: 64-65 [PMID: 24693475 DOI: 10.3947/ic.2014.46.1.64]
- 25 **Sharma NK**, Garg R, Baliga S, Bhat K G. Nosocomial Infections and Drug Susceptibility Patterns in Methicillin Sensitive and Methicillin Resistant *Staphylococcus aureus*. *J Clin Diagn Res* 2013; **7**: 2178-2180 [PMID: 24298469 DOI: 10.7860/JCDR/2013/6750.3463]
- 26 **Nurjadi D**, Olalekan AO, Layer F, Shittu AO, Alabi A, Ghebremedhin B, Schaumburg F, Hofmann-Eifler J, Van Genderen PJ, Caumes E, Fleck R, Mockenhaupt FP, Herrmann M, Kern WV, Abdulla S, Grobusch MP, Kremsner PG, Wolz C, Zanger P. Emergence of trimethoprim resistance gene *dhfrG* in *Staphylococcus aureus* causing human infection and colonization in sub-Saharan Africa and its import to Europe. *J Antimicrob Chemother* 2014; **69**: 2361-2368 [PMID: 24855123 DOI: 10.1093/jac/dku174]
- 27 **Talan DA**, Lovecchio F, Abrahamian FM, Karras DJ, Steele MT, Rothman RE, Krishnadasan A, Mower WR, Hoagland R, Moran GJ. A Randomized Trial of Clindamycin Versus Trimethoprim-sulfamethoxazole for Uncomplicated Wound Infection. *Clin Infect Dis* 2016; **62**: 1505-1513 [PMID: 27025829 DOI: 10.1093/cid/ciw177]
- 28 **Tekin A**, Dal T, Deveci O, Tekin R, Atmaca S, Dayan S. Assessment of methicillin and clindamycin resistance patterns in *Staphylococcus aureus* isolated from a tertiary hospital in Turkey. *Infez Med* 2013; **21**: 111-116 [PMID: 23774974]
- 29 **Shoji K**, Shinjoh M, Horikoshi Y, Tang J, Watanabe Y, Sugita K, Tame T, Iwata S, Miyairi I, Saitoh A. High rate of inducible clindamycin resistance in *Staphylococcus aureus* isolates--a multicenter study in Tokyo, Japan. *J Infect Chemother* 2015; **21**: 81-83 [PMID: 25454215 DOI: 10.1016/j.jiac.2014.10.003]
- 30 **Carnahan RM**, Kuntz JL, Wang SV, Fuller C, Gagne JJ, Leonard CE, Hennessy S, Meyer T, Archdeacon P, Chen CY, Panozzo CA, Toh S, Katcoff H, Woodworth T, Iyer A, Axtman S, Chrischilles EA. Evaluation of the US Food and Drug Administration sentinel analysis tools in confirming previously observed drug-outcome associations: The case of clindamycin and *Clostridium difficile* infection. *Pharmacoepidemiol Drug Saf* 2018; **27**: 731-739 [PMID: 29532543 DOI: 10.1002/pds.4420]
- 31 **Pushkin R**, Iglesias-Ussel MD, Keedy K, MacLauchlin C, Mould DR, Berkowitz R, Kreuzer S, Darouiche R, Oldach D, Fernandes P. A Randomized Study Evaluating Oral Fusidic Acid (CEM-102) in Combination With Oral Rifampin Compared With Standard-of-Care Antibiotics for Treatment of Prosthetic Joint Infections: A Newly Identified Drug-Drug Interaction. *Clin Infect Dis* 2016; **63**: 1599-1604 [PMID: 27682068 DOI: 10.1093/cid/ciw665]
- 32 **Florescu I**, Beuran M, Dimov R, Razbadauskas A, Bochan M, Fichev G, Dukart G, Babinchak T, Cooper CA, Ellis-Grosse EJ, Dartois N, Gandjini H; 307 Study Group. Efficacy and safety of tigecycline compared with vancomycin or linezolid for treatment of serious infections with methicillin-resistant *Staphylococcus aureus* or vancomycin-resistant enterococci: a Phase 3, multicentre, double-blind, randomized study. *J Antimicrob Chemother* 2008; **62** Suppl 1: i17-i28 [PMID: 18684703 DOI: 10.1093/jac/dkn250]
- 33 **Rose WE**, Rybak MJ. Tigecycline: first of a new class of antimicrobial agents. *Pharmacotherapy* 2006; **26**: 1099-1110 [PMID: 16863487 DOI: 10.1592/phco.26.8.1099]
- 34 **Highlights of prescribing information**. Available from: [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2016/2056451bl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2016/2056451bl.pdf)
- 35 **Hassoun A**, Linden PK, Friedman B. Incidence, prevalence, and management of MRSA bacteremia across patient populations-a review of recent developments in MRSA management and treatment. *Crit Care* 2017; **21**: 211 [PMID: 28807042 DOI: 10.1186/s13054-017-1801-3]
- 36 **Allington DR**, Rivey MP. Quinupristin/dalfopristin: a therapeutic review. *Clin Ther* 2001; **23**: 24-44 [PMID: 11219478 DOI: 10.1016/S0149-2918(01)80028-X]
- 37 **Cosimi RA**, Beik N, Kubiak DW, Johnson JA. Ceftaroline for Severe Methicillin-Resistant *Staphylococcus aureus* Infections: A Systematic Review. *Open Forum Infect Dis* 2017; **4**: ofx084 [PMID: 28702467 DOI: 10.1093/ofid/ofx084]
- 38 **Holubar M**, Meng L, Deresinski S. Bacteremia due to Methicillin-Resistant *Staphylococcus aureus*: New Therapeutic Approaches. *Infect Dis Clin North Am* 2016; **30**: 491-507 [PMID: 27208769 DOI: 10.1016/j.idc.2016.02.009]
- 39 **Saravolatz LD**, Stein GE. Oritavancin: A Long-Half-Life Lipoglycopeptide. *Clin Infect Dis* 2015; **61**:

- 627-632 [PMID: 25900171 DOI: 10.1093/cid/civ311]
- 40 **Deresinski S.** Vancomycin in combination with other antibiotics for the treatment of serious methicillin-resistant *Staphylococcus aureus* infections. *Clin Infect Dis* 2009; **49**: 1072-1079 [PMID: 19725789 DOI: 10.1086/605572]
- 41 **Perlroth J,** Kuo M, Tan J, Bayer AS, Miller LG. Adjunctive use of rifampin for the treatment of *Staphylococcus aureus* infections: a systematic review of the literature. *Arch Intern Med* 2008; **168**: 805-819 [PMID: 18443255 DOI: 10.1001/archinte.168.8.805]
- 42 **Tremblay S,** Lau TT, Ensom MH. Addition of rifampin to vancomycin for methicillin-resistant *Staphylococcus aureus* infections: what is the evidence? *Ann Pharmacother* 2013; **47**: 1045-1054 [PMID: 23715070 DOI: 10.1345/aph.1R726]
- 43 **Garrigós C,** Murillo O, Euba G, Verdaguer R, Tubau F, Cabellos C, Cabo J, Ariza J. Efficacy of usual and high doses of daptomycin in combination with rifampin versus alternative therapies in experimental foreign-body infection by methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2010; **54**: 5251-5256 [PMID: 20921321 DOI: 10.1128/AAC.00226-10]
- 44 **Rose WE,** Berti AD, Hatch JB, Maki DG. Relationship of in vitro synergy and treatment outcome with daptomycin plus rifampin in patients with invasive methicillin-resistant *Staphylococcus aureus* infections. *Antimicrob Agents Chemother* 2013; **57**: 3450-3452 [PMID: 23650174 DOI: 10.1128/AAC.00325-12]
- 45 **Hagiya H,** Terasaka T, Kimura K, Satou A, Asano K, Waseda K, Hanayama Y, Otsuka F. Successful treatment of persistent MRSA bacteremia using high-dose daptomycin combined with rifampicin. *Intern Med* 2014; **53**: 2159-2163 [PMID: 25224207 DOI: 10.2169/internalmedicine.53.2711]
- 46 **Kelesidis T,** Humphries R, Ward K, Lewinski MA, Yang OO. Combination therapy with daptomycin, linezolid, and rifampin as treatment option for MRSA meningitis and bacteremia. *Diagn Microbiol Infect Dis* 2011; **71**: 286-290 [PMID: 21855248 DOI: 10.1016/j.diagmicrobio.2011.07.001]
- 47 **Yazaki M,** Oami T, Nakanishi K, Hase R, Watanabe H. A successful salvage therapy with daptomycin and linezolid for right-sided infective endocarditis and septic pulmonary embolism caused by methicillin-resistant *Staphylococcus aureus*. *J Infect Chemother* 2018; **24**: 845-848 [PMID: 29534850 DOI: 10.1016/j.jiac.2018.02.006]
- 48 **Credito K,** Lin G, Appelbaum PC. Activity of daptomycin alone and in combination with rifampin and gentamicin against *Staphylococcus aureus* assessed by time-kill methodology. *Antimicrob Agents Chemother* 2007; **51**: 1504-1507 [PMID: 17220402 DOI: 10.1128/AAC.01455-06]
- 49 **Dhand A,** Sakoulas G. Daptomycin in combination with other antibiotics for the treatment of complicated methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Ther* 2014; **36**: 1303-1316 [PMID: 25444563 DOI: 10.1016/j.clinthera.2014.09.005]
- 50 **Rose WE,** Schulz LT, Andes D, Striker R, Berti AD, Hutson PR, Shukla SK. Addition of ceftaroline to daptomycin after emergence of daptomycin-nonsusceptible *Staphylococcus aureus* during therapy improves antibacterial activity. *Antimicrob Agents Chemother* 2012; **56**: 5296-5302 [PMID: 22869564 DOI: 10.1128/AAC.00797-12]
- 51 **Dhand A,** Bayer AS, Pogliano J, Yang SJ, Bolaris M, Nizet V, Wang G, Sakoulas G. Use of antistaphylococcal beta-lactams to increase daptomycin activity in eradicating persistent bacteremia due to methicillin-resistant *Staphylococcus aureus*: role of enhanced daptomycin binding. *Clin Infect Dis* 2011; **53**: 158-163 [PMID: 21690622 DOI: 10.1093/cid/cir340]
- 52 **Parra-Ruiz J,** Bravo-Molina A, Peña-Monje A, Hernández-Quero J. Activity of linezolid and high-dose daptomycin, alone or in combination, in an in vitro model of *Staphylococcus aureus* biofilm. *J Antimicrob Chemother* 2012; **67**: 2682-2685 [PMID: 22796888 DOI: 10.1093/jac/dks272]
- 53 **Yehia H,** El Said M, Azmy M, Badawy M, Mansy S, Gohar H, Madany N. Effect of linezolid alone and in combination with other antibiotics, on methicillin-resistant *Staphylococcus aureus*. *J Egypt Soc Parasitol* 2016; **46**: 57-66 [PMID: 27363041 DOI: 10.12816/0026150]
- 54 **Gervasoni C,** Simonetti FR, Resnati C, Charbe N, Clementi E, Cattaneo D. Prolonged inductive effect of rifampicin on linezolid exposure. *Eur J Clin Pharmacol* 2015; **71**: 643-644 [PMID: 25778934 DOI: 10.1007/s00228-015-1833-z]
- 55 **Hoyo I,** Martínez-Pastor J, Garcia-Ramiro S, Climent C, Brunet M, Cuesta M, Mensa J, Soriano A. Decreased serum linezolid concentrations in two patients receiving linezolid and rifampicin due to bone infections. *Scand J Infect Dis* 2012; **44**: 548-550 [PMID: 22385321 DOI: 10.3109/00365548.2012.663931]
- 56 **Werth BJ.** Exploring the pharmacodynamic interactions between tedizolid and other orally bioavailable antimicrobials against *Staphylococcus aureus* and *Staphylococcus epidermidis*. *J Antimicrob Chemother* 2017; **72**: 1410-1414 [PMID: 28158617 DOI: 10.1093/jac/dkw588]
- 57 **Fujino T,** Amari Y, Mohri M, Noma M, Yamamoto H. MRSA tricuspid valve infective endocarditis with multiple embolic lung abscesses treated by combination therapy of vancomycin, rifampicin, and sulfamethoxazole/trimethoprim. *J Cardiol* 2009; **53**: 146-149 [PMID: 19167651 DOI: 10.1016/j.jcc.2008.06.007]





Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160 Pleasanton, CA 94566, USA  
Telephone: +1-925 2238242  
Fax: +1-925-2238243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>



# World Journal of *Clinical Infectious Diseases*

*World J Clin Infect Dis* 2019 August 15; 9(2): 11-22





**EDITORIAL**

- 11 Towards the worldwide eradication of hepatitis B virus infection: A combination of prophylactic and therapeutic factors  
*Sagnelli C, Sagnelli E*

**ABOUT COVER**

Editorial Board Member of *World Journal of Clinical Infectious Diseases*,  
Michael S Firstenberg, MD, Associate Professor, Doctor, Surgeon,  
Department of Surgery, Medical Center of Aurora, Aurora, CO 80012,  
United States

**AIMS AND SCOPE**

*World Journal of Clinical Infectious Diseases (World J Clin Infect Dis, WJCID*,  
online ISSN 2220-3176, DOI: 10.5495) is a peer-reviewed open access  
academic journal that aims to guide clinical practice and improve  
diagnostic and therapeutic skills of clinicians.

The *WJCID* covers a variety of clinical medical topics, including  
community-acquired infections, cross infection, eye infections, focal  
infection, gingivitis, infectious, infectious, intraabdominal infections,  
laboratory infection, ludwig's angina, necrotizing ulcerative, opportunistic  
infections, pelvic infection, pregnancy complications, etc.

We encourage authors to submit their manuscripts to *WJCID*. We will  
give priority to manuscripts that are supported by major national and  
international foundations and those that are of great clinical significance.

**INDEXING/ABSTRACTING**

*World Journal of Clinical Infectious Diseases* is now indexed in China National Knowledge  
Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and  
Superstar Journals Database.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Responsible Electronic Editor: *Yun-Xiaojuan Wu*  
Proofing Production Department Director: *Xiang Li*

**NAME OF JOURNAL**

*World Journal of Clinical Infectious Diseases*

**ISSN**

ISSN 2220-3176 (online)

**LAUNCH DATE**

December 30, 2011

**FREQUENCY**

Irregular

**EDITORS-IN-CHIEF**

Joao Mesquita, Caterina Sagnelli

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/2220-3176/editorialboard.htm>

**EDITORIAL OFFICE**

Ya-Juan Ma, Director

**PUBLICATION DATE**

August 15, 2019

**COPYRIGHT**

© 2019 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Towards the worldwide eradication of hepatitis B virus infection: A combination of prophylactic and therapeutic factors

Caterina Sagnelli, Evangelista Sagnelli

**ORCID number:** Caterina Sagnelli (0000-0002-6413-7810); Evangelista Sagnelli (0000-0003-2817-8436).

**Author contributions:** All authors equally contributed to this paper with conception and design of the study, literature review and analysis, drafting, critical revision and editing, and final approval of the final version.

**Conflict-of-interest statement:** The authors have no conflict of interest to declare.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Invited manuscript

**Received:** April 30, 2019

**Peer-review started:** May 7, 2019

**First decision:** June 18, 2019

**Revised:** June 22, 2019

**Accepted:** July 16, 2019

**Article in press:** July 17, 2019

**Published online:** August 15, 2019

**P-Reviewer:** Abushady EAE,

**Caterina Sagnelli, Evangelista Sagnelli,** Department of Mental Health and Public Medicine, Section of Infectious Diseases, University of Campania Luigi Vanvitelli, Naples 80131, Italy

**Corresponding author:** Caterina Sagnelli, MD, PhD, Associate Professor, Department of Mental Health and Public Medicine, Section of Infectious Diseases, University of Campania Luigi Vanvitelli, Via L. Armanni 5, Naples 80131, Italy. [caterina.sagnelli@unicampania.it](mailto:caterina.sagnelli@unicampania.it)

**Telephone:** +39-81-5666719

**Fax:** +39-81-5666207

### Abstract

Hepatitis B virus (HBV) is still a global health problem, mostly because of the intermediate/high rates of HBV chronic carriers living in most Asian, African and eastern European countries. The universal HBV vaccination of new-borns undertaken in most nations over the last 3 decades and effective HBV antiviral treatments (nucleos(t)ide analogue with high genetic barrier to viral resistance) introduced in the last decade have shown their beneficial effects in inducing a clear reduction of HBV endemicity in the countries where they have been extensively applied. Great hopes are now placed on new antiviral and immunotherapeutic drugs that are now at an advanced stage of study. It is in fact already conceivable that the synergistic use of new drugs targeting more than one HBV-lifecycle steps (covalent closed circular DNA destruction/silencing, HBV entry inhibitors, nucleocapsid assembly modulators targeting viral transcripts) and of some new immunotherapeutic agents might eliminate the intrahepatic covalent closed circular DNA and achieve the eradication of HBV infection. In spite of this, a strong effort should be given to extensive educational and screening programs for the at-risk population and to the implementation of HBV vaccination in developing countries.

**Key words:** Hepatitis B virus; Chronic hepatitis B infection; Hepatitis B virus prevention; Vaccination

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** The spread of hepatitis B virus (HBV) infection has recently decreased in several countries due to the universal HBV vaccination of new-born babies and to the extended use of HBV nucleos(t)ide analogues with high genetic barrier to viral resistance. However, HBV vaccination and extensive educational and screening programs for at risk populations should be implemented predominantly in developing

Farshadpour F, Gencdal G

**S-Editor:** Cui LJ**L-Editor:** Filipodia**E-Editor:** Wu YXJ

countries. New drugs targeting more than one HBV-lifecycle steps and of some new immunotherapeutic agents are under investigation with the aim of obtaining the clearance of hepatocytic covalent closed circular DNA through their synergistic action.

**Citation:** Sagnelli C, Sagnelli E. Towards the worldwide eradication of hepatitis B virus infection: A combination of prophylactic and therapeutic factors. *World J Clin Infect Dis* 2019; 9(2): 11-22

**URL:** <https://www.wjgnet.com/2220-3176/full/v9/i2/11.htm>

**DOI:** <https://dx.doi.org/10.5495/wjcid.v9.i2.11>

## INTRODUCTION

Despite the universal vaccination campaigns against hepatitis B virus (HBV) undertaken in most nations over the last 3 decades, HBV is still a global health problem with about 257 million people chronically infected, at least 40% of world population being an HBV contact or carrier<sup>[1]</sup>. About half million deaths per year are due to complications of advanced chronic hepatitis, and 340000 are due to hepatocellular carcinoma (HCC)<sup>[2,3]</sup>.

The level of HBV endemicity, evaluated on the prevalence of subjects with HBV chronic infection, varies significantly from one country to another and in some countries from one geographic area to another. The rate of hepatitis B surface antigen (HBsAg) chronic carriers ranges from 0.5% to 2% (low endemicity) in most countries of North and South America, Western and Central Europe, Australia and northern Africa, from 2.1% to 8% (intermediate endemicity) in most eastern European and central Asian nations and above 8% (high endemicity) in some eastern Asian and sub-Saharan African countries and in Alaska<sup>[4,5]</sup>. Ten HBV genotypes (HBV-GT) have been identified at present, and their geographical distribution is of great epidemiological interest because it is conditioned by both local diffusion and migratory flows<sup>[4-6]</sup>. HBV-GT-A predominates in North America, eastern Africa and northern/western Europe<sup>[9,10]</sup>, HBV-GT-B and -C in Asia<sup>[11]</sup>, HBV-GT-D in countries facing the Mediterranean sea<sup>[11-21]</sup>, in the Middle-East and in southern Asia<sup>[5]</sup>, HBV-GT-E in central-western Africa<sup>[4,19,22]</sup>, genotype F in southern and central America<sup>[5]</sup>, HBV-GT-G in France and in some region in the United States<sup>[5]</sup>, HBV-GT-H in Latin America<sup>[5]</sup> and HBV-GT-I and -J in eastern Asia<sup>[5,10]</sup>. However, several cases of acute hepatitis related to HBV-GT typical of geographic areas with high or intermediate endemicity have occurred in western countries hosting migrant populations from those areas<sup>[6,23-32]</sup>.

Promiscuous unprotected sexual activity is a main risk factor for acquiring HBV infection worldwide, while other main risk factors have a different impact in different geographical areas. In fact, HBV infection is most frequently acquired at birth from hepatitis B e-antigen (HBeAg) positive mothers or through household contacts in early childhood in countries with intermediate/high endemicity, with a high rate of progression to chronicity that helps to maintain the high levels of endemicity. On the other hand, in countries with low HBV endemicity like Western Europe, North America and Australia, the major risk factor for acquiring HBV infection is the sharing of needles and other equipment between intravenous drug users<sup>[33,34]</sup>, which causes the infection to remain confined to this at-risk population.

Acute Hepatitis B onset occurs 45-180 d after HBV has been acquired with some constitutional symptoms followed by dark urine and jaundice in less than 10% of children aged less than 5 years and in more than 50% of adults. The symptomatic phase of the illness lasts about 15 d and even longer in adults. Immune-complexes related extrahepatic manifestation (membranous glomerulonephritis, necrotizing vasculitis and papular acrodermatitis) are rare events<sup>[35,36]</sup>.

Fulminant hepatitis is due to an overreaction of the immune system; it develops in about 1% of the patients<sup>[37,38]</sup>, leading to death in about three-quarter of them and requiring liver transplantation. The age-related difference in the clinical outcome of acute HBV infection is striking<sup>[39]</sup>. In fact, more than 95% of adult patients spontaneously recover and develop a long-lasting immunological protection against reinfection, provided by seroconversion to hepatitis B surface antibody (anti-HBs) and by cellular immunity, while only 2%-5% progresses to chronicity<sup>[40]</sup>; instead, 90% of new-borns and 30% of children aged 1-5 years progresses to chronicity<sup>[41]</sup>. The difference in the outcome between children and adults is based on the degree of

reactivity of the cell-mediated immunity, recognized as the true engine for eliminating HBV infection, low in new-borns and children and normal or high in teenagers and adults<sup>[42]</sup>. Risk factors for a more severe clinical course have been recognized in being a young adult or of female sex, in coinfection with hepatitis D virus (HDV), hepatitis C virus (HCV) or human immunodeficiency virus (HIV), in alcohol abuse and in intravenous drug use<sup>[43-53]</sup>.

Once a patient has recovered and serum HBsAg cleared, a residual HBV replication persists, as evidenced by the detection of small amount of HBV-DNA inside the hepatocytes, a virologic condition named occult B infection<sup>[54-63]</sup>.

Depending on the entity of HBV replication and on the effectiveness of the immune-response, chronic infection has a variable clinical presentation broadly grouped in either an asymptomatic stable HBsAg carriage, chronic hepatitis or liver cirrhosis with or without HCC<sup>[23,64,65]</sup>. Patients with chronic hepatitis progress to cirrhosis at a rate of 1%-5% per year<sup>[66]</sup>; and, in turn, HBV cirrhotic patients develop HCC at a median rate of about 3.7% per year<sup>[24,67-70]</sup>.

The wide spread of HBV infection, its frequent evolution into chronicity with the possibility of developing liver cirrhosis and HCC and its progression to death in patients who do not undergo a successful liver transplantation have called for extensive HBV vaccination campaigns and effective therapeutic measures.

---

## USE OF HBV VACCINATION IN REDUCING THE SPREAD OF HBV INFECTION

---

Introduced in 1982, HBV vaccination is the most effective measure to prevent HBV infection<sup>[71]</sup>. One dose of the currently used HBV vaccine contains 5 µg of recombinant HBsAg produced in yeast *Saccharomyces cerevisiae* with recombinant DNA technology and adsorbed on amorphous aluminium sulphate hydroxyphosphate. Hepatitis B vaccine is given as a three-dose series. Post-vaccination testing is required, and a person with suboptimal response (serum titers of antibody to HBsAg < 10 mIU/mL), like immunocompromised persons and those with advanced renal disease<sup>[71]</sup>, should receive a fourth dose or be revaccinated<sup>[71-73]</sup>. HBV vaccination provides a protective production of antibody to HBsAg > 10 mIU/mL in about 95% of subjects and is more effective in children and young adults than in adults over 40. In adults, about 90% reach anti-HBs protective levels, and females respond to HBV vaccine better than males<sup>[76,77]</sup>. It has also been documented that vaccine induced anti HBV immunity lasts at least 3 decades<sup>[78-81]</sup> and is presumably life-long.

HBV vaccination had been initially recommended for infants born to HBV-infected mothers and for adults at risk for acquiring HBV infection (sexual partners or household contacts of HBsAg-positive persons, subjects with more than one sexual partner, males having sex with males; injection drugs users; incarcerated persons; health care workers and public safety employees at risk for exposure to blood or blood-contaminated body fluids; adults with diabetes mellitus; persons with advanced renal disease, persons with chronic liver disease not HBV-related, pregnant women who are at risk during pregnancy, HIV-infected persons; international travellers to regions with high or intermediate levels of HBV endemicity and any adult seeking protection from HBV infection)<sup>[71-73]</sup>. HBV vaccination offered to young or adult subjects at risk of infection has not been particularly effective, since it is estimated that only 20%-30% of those in need have accepted vaccination, and, consequently, no evident reduction in HBV endemicity has been obtained. Worthy of mention, the prevalence of acceptance of HBV vaccination in healthcare workers (HCWs) ranges from 15% in African countries to nearly 75% in the United States<sup>[82-87]</sup>. In addition, half of HBV vaccinated subjects completed the vaccination schedule, resulting in a lower production of anti HBs and, consequently, in a risk of lower level and lower duration of protection. Several reasons contribute to the poor acceptance of a necessary vaccination, like little information on the usefulness or effectiveness of the vaccine, poor confidence in its effectiveness, fear of adverse reactions, lack of availability and cost of vaccine in some countries<sup>[88-91]</sup>. That being the case, countless decades would have been necessary to reach the worldwide eradication of HBV infection.

A more effective vaccination strategy was therefore chosen in most countries. The universal vaccination of all new-born babies has shown beneficial effect wherever it has been correctly applied, with a clear reduction of the levels of HBV endemicity. Worthy of mention, prior to the introduction of the national HBV vaccination program in 1984, approximately 15%-20% of the Taiwanese adult population were HBsAg positive<sup>[92,93]</sup>. The effectiveness of this program was demonstrated by the significant decrease in the incidence rate of HBV chronic carriers and the rate of

mother-to-child HBV vertical transmission<sup>[94-98]</sup>. An example of this favourable effect is the strong decrease in the rate of HBsAg positivity in university students of this country, which was decreased from 9.7% in those born before 1974 to less than 1% in those born after 1992<sup>[99]</sup>.

After an 8-year application of universal HBV vaccination of new-borns in Saudi Arabia, the HBsAg prevalence in children aged 1-12 years dropped from 6.7% in 1989 to 0.3% in 1997<sup>[100]</sup>. In Gambia, a clear reduction in newly acquired HBV infections, HBsAg carrier rate and HBV-related mortality was observed 14 years after the introduction of HBV vaccination in children<sup>[101]</sup>. Also, in Alaska, the implementation of HBV vaccination induced a decrease in the HBsAg carrier rate<sup>[102]</sup>.

The impressive reduction in HBV endemicity in countries where universal vaccination against HBV has been applied is in stark contrast to the persistence of high HBV endemicity persisting in developing countries where HBV vaccination programs have been poorly applied. An example of this contrast was recently observed by us in a cohort of migrants who came from countries of sub-Saharan western Africa to Europe. In this cohort, migrants born in western African countries where HBV vaccination has been not sufficiently applied showed an HBsAg positivity ranging from 9.7% to 22.5%, whereas those born in Nigeria showed the beneficial effects of a universal HBV vaccination of new-borns well applied from 2 decades. Those from Nigeria had a global rate of HBsAg positivity of 4.1% and age-related rates of 3.5% in subjects less than 25 years, 4.1% in those aged 26-40 years and 17.9% in those aged over 41, a cohort effect underscoring a tendency of HBV endemicity towards reduction.

Concluding on this point, there remains much to be done to get a proper extended application of all the possible prophylaxis measures aimed at reaching the eradication of HBV infection. Firstly, HBV universal vaccination programs of new-born babies should be extensively applied and never discontinued in all world countries. Secondly, extensive information campaigns will have to be undertaken so that people at risk of HBV infection may receive instructions on how this infection spreads and how to prevent it and then be encouraged to undergo screening and, if exposed to infection, to HBV vaccination<sup>[103]</sup>. Thirdly, a permanent program of screening and vaccination of migrants from areas of intermediate or high endemicity must be applied in all host nations. These remedies, however, will not be enough as there are, as of now, some hundreds of millions of infected subjects able to transmit the infection worldwide.

Mainly due to individual factors (*e.g.*, immunogenetic conditions, advanced age, obesity, smoking or chronic diseases such as celiac disease, diabetes, HIV infection, advanced kidney disease, autoimmune diseases), 5%-10% of the adult population does not respond or responds insufficiently to anti-HBV vaccine (anti-HBs titres < 10 mIU/mL). For non-responders, the pathway to improve the immunogenicity of the vaccine adjuvant has been followed and an oligonucleotide of the cytosine phosphoguanosine, a Toll-like 9 agonist receptor potent stimulator of the vertebrate innate immune system, has been used as an adjuvant for a recombinant two-dose hepatitis B vaccine (administered at wk 0 and 4). Recently approved for use in adults, initial data have shown a higher percentage of protected subjects compared to alum-adsorbed vaccines<sup>[71,73,74,79,81]</sup>.

---

## USE OF THE ANALOGOUS NUCLEOS(T)IDES IN ABOLISHING THE INFECTIVITY OF HBV CHRONIC CARRIERS

---

The pharmacological suppression of HBV replication in HBV chronic carriers is another opportunity for health authorities to undertake an effective path towards the eradication of HBV infection. Although a sustained eradication of intrahepatic covalent closed circular DNA (cccDNA) as well as integrated cccDNA is currently not feasible, long term suppression of viral replication with HBV DNA serum clearance may be easily obtained with long-term administration, maybe life-long, of high genetic barrier to resistance nucleos(t)ide analogues tenofovir disoproxil fumarate (TDF), entecavir (ETV) or tenofovir alafenamide (TAF). These drugs have improved the outcomes of HBV-related chronic hepatitis by lowering the rate of transition to liver cirrhosis and reducing the risk of HCC development, but the clearance of serum HBsAg is only achieved in a small portion of treated patients<sup>[104]</sup>.

Treatment with interferon in its pegylated form (PEG-IFN $\alpha$ ), extensively used as monotherapy in the past, will become obsolete because of its poor efficacy and of the frequent occurrence of badly endured and sometimes severe adverse reactions during long-term treatment. In HBsAg/HBeAg positive patients, the seroconversion to anti-



HBe was obtained only in 29%-32% of patients after 1-year PEG-IFN treatment and to anti-HBs only in 3%-5%<sup>[105,106]</sup>. In HBeAg-negative patients, a favourable response with stable normalization of serum alanine aminotransferase and serum HBV DNA reduced below 400 copies/ml was obtained only in 15% of cases treated for 12 mo, with HBsAg loss in about 4%<sup>[107]</sup>.

The first generation nucleos(t)ide analogues lamivudine, adefovir and telbivudine have become obsolete because their low genetic barrier is unable to prevent the formation of viral resistant strains. In addition, the sequential use of ETV to treat lamivudine resistance increases the risk of ETV resistance. A switch to tenofovir has been demonstrated to be effective in patients with confirmed lamivudine, telbivudine, adefovir or ETV resistance.

Long-term therapy with nucleos(t)ide analogues with high genetic barrier to viral resistance (ETV, TDF) is required to obtain a stable suppression of HBV replication<sup>[108]</sup>. These drugs are highly recommended as first-line therapy because HBV resistance is a rare event in nucleoside-naïve patients during a 5-year treatment with ETV and no resistance with a 7-year treatment with TDF<sup>[109]</sup>. Histological evaluation after a long-term treatment with ETV or TDF showed an impressive improvement in liver necroinflammation and fibrosis scores in most patients<sup>[110,111]</sup>. In addition, compared with controls, a significant reduction in the incidence of HCC has been observed in HBsAg positive cirrhotic patients undergoing a long-term therapy with ETV or TDF<sup>[112-117]</sup>.

A 5-year ETV treatment induced HBV DNA serum clearance in more than 90% of HBeAg positive patients with chronic hepatitis<sup>[118]</sup>, and a similar rate was obtained with TDF<sup>[109]</sup>. Seroconversion to anti-HBe was obtained in about 20% of patients after 1-year of ETV or TDF therapy<sup>[118,119]</sup>.

HBsAg loss occurred in 11.8% of HBeAg-positive patients after 7 years of TDF treatment, more frequently in Caucasians than in Asians<sup>[109]</sup>. Consolidation therapy is recommended after the loss of HBsAg<sup>[120]</sup>.

TAF is an oral second-generation prodrug of TDF with a high genetic barrier to viral resistance. Although TDF and TAF show similar rates of cure<sup>[121-123]</sup>, switching from TDF to TAF provides improvement in bone density and renal function, a favourable effect in a long-term treatment<sup>[124,125]</sup>.

Currently, new drugs are being tested that are aimed at eradicating chronic HBV infection: HBV entry inhibitors, capsid inhibitors, short interfering RNA and targeting cccDNA<sup>[126-132]</sup>. Briefly, a blockade of HBV entry in experimental cells was obtained using a pre-S acylated peptide of the large HBsAg protein, and further studies on chronic HBV and HDV infection are ongoing<sup>[133]</sup>. In some experimental models it has been shown that the AB-423 capsid inhibitor is able to direct erroneously capsid assembly to inhibit pregenomic RNA encapsidation and consequently to reduce cccDNA concentrations in liver cells<sup>[128-138]</sup>.

Several antisense short interfering RNAs targeted towards HBsAg transcripts have achieved mRNA degradation in pre-clinical or clinical evaluation. Among these, ARC 520 is of interest. It is directed towards HBV RNA transcripts and reduces the synthesis of HBV DNA and viral proteins<sup>[138]</sup>. Regarding cccDNA targeting, several DNA cleavage enzymes have been tested in experimental models and preliminary data seem encouraging<sup>[132]</sup>.

In addition, experimental studies are underway to develop new drugs or therapeutic vaccines that may regulate the immune-system dysfunction in hepatitis B<sup>[139-150]</sup>.

---

## CONCLUSION

---

The universal HBV vaccination of new-borns has produced significant results in countries where, responding to the demand of the World Health Organization, it has been correctly applied. Nevertheless, in several developing countries, socio-economic reasons have impaired the application of HBV vaccination, delaying the achievement of a global reduction in HBV endemicity. Also, the vaccination on a voluntary basis of adults at risk of HBV infection has failed to contribute to the project of a progressive reduction of the levels of endemicity. This being the case, we believe that an additional 2-3 decades of extensive application of the universal HBV vaccination will be needed to achieve a substantial reduction of HBV spread.

Another aspect of the ambitious project to eradicate HBV infection is the extensive information campaign on how to acquire the infection and how to prevent it. So far, information campaigns have been occasional and limited to certain risk categories in many countries and therefore have not substantially contributed to the reduction of HBV endemicity.



Good news comes from the therapeutic management of chronic hepatitis B. In fact, the new nucleos(t)ide analogues (ETV, TDF and TAF) that effectively suppress HBV replication may be used for a very long period with no risk to induce viral resistance. In addition, new drugs for the complete eradication of HBV replication, thus ensuring a complete cure, are currently being developed and will be very likely be available in the next decade.

The set of data reported here suggests that prolonged extended application of the universal HBV vaccination of new-borns and the utilization of the high genetic barrier to resistance nucleos(t)ide analogues and, in the near future, of some drugs today in experimental development will allow for, in the next 2-3 decades, a strong reduction of HBV endemicity and possibly the eradication of HBV infection.

## REFERENCES

- 1 **World Health Organization.** Global Hepatitis Report, 2017. Geneva: World Health Organization; 2017. Available from: <https://apps.who.int/iris/bitstream/handle/10665/255016/9789241565455-eng.pdf;sequence=1>
- 2 **World Health Organization Guidelines for the Prevention, Care and Treatment of Persons with Chronic Hepatitis B Virus Infection.** World Health Organization 2015. 2019; [cited 2019 May 17] Available from: <http://www.worldhepatitisalliance.org/sites/default/files/resources/documents/Hep%20B%20Guidelines.pdf>
- 3 **World Health Organization.** Disease burden and mortality estimates. Geneva: World Health Organization; 2016. Available from: [https://www.who.int/healthinfo/global\\_burden\\_disease/estimates/en/index1.html](https://www.who.int/healthinfo/global_burden_disease/estimates/en/index1.html)
- 4 **Zehender G,** Ebranati E, Gabanelli E, Sorrentino C, Lo Presti A, Tanzi E, Ciccozzi M, Galli M. Enigmatic origin of hepatitis B virus: an ancient travelling companion or a recent encounter? *World J Gastroenterol* 2014; **20**: 7622-7634 [PMID: 24976700 DOI: 10.3748/wjg.v20.i24.7622]
- 5 **Croagh CM,** Desmond PV, Bell SJ. Genotypes and viral variants in chronic hepatitis B: A review of epidemiology and clinical relevance. *World J Hepatol* 2015; **7**: 289-303 [PMID: 25848459 DOI: 10.4254/wjh.v7.i3.289]
- 6 **European Commission.** Migration and Home Affairs. Available from: [http://ec.europa.eu/dgs/home-affairs/what-we-do/networks/european\\_migration\\_network/reports/docs/annual-policy/2014/00.emn\\_annual\\_report\\_on\\_immigration\\_and\\_asylum\\_synthesis\\_report.pdf](http://ec.europa.eu/dgs/home-affairs/what-we-do/networks/european_migration_network/reports/docs/annual-policy/2014/00.emn_annual_report_on_immigration_and_asylum_synthesis_report.pdf)
- 7 **Paraskevis D,** Magiorkinis G, Magiorkinis E, Ho SY, Belshaw R, Allain JP, Hatzakis A. Dating the origin and dispersal of hepatitis B virus infection in humans and primates. *Hepatology* 2013; **57**: 908-916 [PMID: 22987324 DOI: 10.1002/hep.26079]
- 8 **Lai A,** Sagnelli C, Presti AL, Cella E, Angeletti S, Spoto S, Costantino S, Sagnelli E, Ciccozzi M. What is changed in HBV molecular epidemiology in Italy? *J Med Virol* 2018; **90**: 786-795 [PMID: 29315661 DOI: 10.1002/jmv.25027]
- 9 **Zehender G,** De Maddalena C, Giambelli C, Milazzo L, Schiavini M, Bruno R, Tanzi E, Galli M. Different evolutionary rates and epidemic growth of hepatitis B virus genotypes A and D. *Virology* 2008; **380**: 84-90 [PMID: 18715605 DOI: 10.1016/j.virol.2008.07.009]
- 10 **Norder H,** Couroucé AM, Coursaget P, Echevarria JM, Lee SD, Mushahwar IK, Robertson BH, Locarnini S, Magnus LO. Genetic diversity of hepatitis B virus strains derived worldwide: genotypes, subgenotypes, and HBsAg subtypes. *Intervirology* 2004; **47**: 289-309 [PMID: 15564741 DOI: 10.1159/000080872]
- 11 **Okamoto H,** Tsuda F, Sakugawa H, Sastrosoewignjo RI, Imai M, Miyakawa Y, Mayumi M. Typing hepatitis B virus by homology in nucleotide sequence: comparison of surface antigen subtypes. *J Gen Virol* 1988; **69**: 2575-2583 [PMID: 3171552 DOI: 10.1099/0022-1317-69-10-2575]
- 12 **Amini-Bavil-Olyae S,** Sarrami-Forooshani R, Mahboudi F, Sabahi F, Adeli A, Noorinayer B, Azizi M, Reza Zali M. Genotype characterization and phylogenetic analysis of hepatitis B virus isolates from Iranian patients. *J Med Virol* 2005; **75**: 227-234 [PMID: 15602742 DOI: 10.1002/jmv.20261]
- 13 **Arauz-Ruiz P,** Norder H, Robertson BH, Magnus LO. Genotype H: a new Amerindian genotype of hepatitis B virus revealed in Central America. *J Gen Virol* 2002; **83**: 2059-2073 [PMID: 12124470 DOI: 10.1099/0022-1317-83-8-2059]
- 14 **Coppola N,** Masiello A, Tonziello G, Pisapia R, Pisaturo M, Sagnelli C, Messina V, Iodice V, Sagnelli E. Factors affecting the changes in molecular epidemiology of acute hepatitis B in a Southern Italian area. *J Viral Hepat* 2010; **17**: 493-500 [PMID: 19780943 DOI: 10.1111/j.1365-2893.2009.01201.x]
- 15 **Coppola N,** Sagnelli C, Pisaturo M, Minichini C, Messina V, Alessio L, Starace M, Signoriello G, Gentile I, Filippini P, Sagnelli E. Clinical and virological characteristics associated with severe acute hepatitis B. *Clin Microbiol Infect* 2014; **20**: O991-O997 [PMID: 24930916 DOI: 10.1111/1469-0691.12720]
- 16 **Coppola N,** Tonziello G, Colombatto P, Pisaturo M, Messina V, Moriconi F, Alessio L, Sagnelli C, Cavallone D, Brunetto M, Sagnelli E. Lamivudine-resistant HBV strain rtM204V/I in acute hepatitis B. *J Infect* 2013; **67**: 322-328 [PMID: 23796869 DOI: 10.1016/j.jinf.2013.06.006]
- 17 **Forbi JC,** Vaughan G, Purdy MA, Campo DS, Xia GL, Ganova-Raeva LM, Ramachandran S, Thai H, Khudyakov YE. Epidemic history and evolutionary dynamics of hepatitis B virus infection in two remote communities in rural Nigeria. *PLoS One* 2010; **5**: e11615 [PMID: 20657838 DOI: 10.1371/journal.pone.0011615.]
- 18 **Sagnelli C,** Ciccozzi M, Pisaturo M, Lo Presti A, Cella E, Coppola N, Sagnelli E. The impact of viral molecular diversity on the clinical presentation and outcome of acute hepatitis B in Italy. *New Microbiol* 2015; **38**: 137-147 [PMID: 25915056]
- 19 **Sagnelli E,** Stroffolini T, Mele A, Imparato M, Sagnelli C, Coppola N, Almasio PL. Impact of comorbidities on the severity of chronic hepatitis B at presentation. *World J Gastroenterol* 2012; **18**: 1616-1621 [PMID: 22529690 DOI: 10.3748/wjg.v18.i14.1616.]
- 20 **Sagnelli E,** Taliani G, Castelli F, Bartolozzi D, Cacopardo B, Armignacco O, Scotto G, Coppola N, Stroffolini T, Sagnelli C. Chronic HBV infection in pregnant immigrants: a multicenter study of the Italian Society of Infectious and Tropical Diseases. *New Microbiol* 2016; **39**: 114-118 [PMID: 27196549]

- 21 **Calogero A**, Sagnelli E, Creta M, Angeletti S, Peluso G, Incollingo P, Candida M, Minieri G, Carlomagno N, Dodaro CA, Ciccozzi M, Sagnelli C. Eradication of HCV Infection with the Direct-Acting Antiviral Therapy in Renal Allograft Recipients. *Biomed Res Int* 2019; **2019**: 4674560 [PMID: [31179323](#) DOI: [10.1155/2019/4674560](#)]
- 22 **Stroffolini T**, Sagnelli E, Sagnelli C, Morisco F, Babudieri S, Furlan C, Pirisi M, Russello M, Smedile A, Pisaturo M, Almasio PL. Decreasing role of HCV and HBV infections as aetiological factors of hepatocellular carcinoma in Italy. *Infection* 2019 [PMID: [31028627](#) DOI: [10.1007/s15010-019-01308-3](#)]
- 23 **Sagnelli E**, Sagnelli C, Pisaturo M, Macera M, Coppola N. Epidemiology of acute and chronic hepatitis B and delta over the last 5 decades in Italy. *World J Gastroenterol* 2014; **20**: 7635-7643 [PMID: [24976701](#) DOI: [10.3748/wjg.v20.i24.7635](#)]
- 24 **Sagnelli C**, Ciccozzi M, Coppola N, Minichini C, Lo Presti A, Starace M, Alessio L, Macera M, Cella E, Gualdieri L, Caprio N, Pasquale G, Sagnelli E. Molecular diversity in irregular or refugee immigrant patients with HBV-genotype-E infection living in the metropolitan area of Naples. *J Med Virol* 2017; **89**: 1015-1024 [PMID: [27805272](#) DOI: [10.1002/jmv.24724](#)]
- 25 **El-Hamad I**, Pezzoli MC, Chiari E, Scarcella C, Vassallo F, Puoti M, Ciccaglione A, Ciccozzi M, Scalzini A, Castelli F; ad-hoc Working Group for Hepatitis B in migrants. Point-of-care screening, prevalence, and risk factors for hepatitis B infection among 3,728 mainly undocumented migrants from non-EU countries in northern Italy. *J Travel Med* 2015; **22**: 78-86 [PMID: [25424439](#) DOI: [10.1111/jtm.12176](#)]
- 26 **Coppola N**, Alessio L, Gualdieri L, Pisaturo M, Sagnelli C, Caprio N, Maffei R, Starace M, Angelillo IF, Pasquale G, Sagnelli E. Hepatitis B virus, hepatitis C virus and human immunodeficiency virus infection in undocumented migrants and refugees in southern Italy, January 2012 to June 2013. *Euro Surveill* 2015; **20**: 30009 [PMID: [26530499](#) DOI: [10.2807/1560-7917.ES.2015.20.35.30009](#)]
- 27 **Coppola N**, Alessio L, Gualdieri L, Pisaturo M, Sagnelli C, Minichini C, Di Caprio G, Starace M, Onorato L, Signoriello G, Macera M, Angelillo IF, Pasquale G, Sagnelli E. Hepatitis B virus infection in undocumented immigrants and refugees in Southern Italy: demographic, virological, and clinical features. *Infect Dis Poverty* 2017; **6**: 33 [PMID: [28179020](#) DOI: [10.1186/s40249-016-0228-4](#)]
- 28 **Sagnelli E**, Alessio L, Sagnelli C. Hepatitis B Virus Genotypes, Epidemiological Characteristics, and Clinical Presentation of HBV Chronic Infection in Immigrant Populations Living in Southern Italy. *Hepat Mon* 2017; **17**: e13260 [DOI: [10.5812/hepatmon.13260](#)]
- 29 **Coppola N**, Alessio L, Pisaturo M, Macera M, Sagnelli C, Zampino R, Sagnelli E. Hepatitis B virus infection in immigrant populations. *World J Hepatol* 2015; **7**: 2955-2961 [PMID: [26730274](#) DOI: [10.4254/wjh.v7.i30.2955](#)]
- 30 **Zampino R**, Capoluongo N, Boemio A, Macera M, Vitrone M, Adinolfi LE, Filippini P, Sagnelli E, Sagnelli C, Durante-Mangoni E, Coppola N. Effect of a Cooperation Strategy between Primary Care Physicians and Hospital Liver Units on HBV Care in Campania, Italy. *Can J Gastroenterol Hepatol* 2018; **2018**: 5670374 [PMID: [30148123](#) DOI: [10.1155/2018/5670374](#)]
- 31 **Sagnelli C**, Ciccozzi M, Pisaturo M, Zehender G, Lo Presti A, Alessio L, Starace M, Lovero D, Sagnelli E, Coppola N. Molecular epidemiology of hepatitis B virus genotypes circulating in acute hepatitis B patients in the Campania region. *J Med Virol* 2014; **86**: 1683-1693 [PMID: [24980631](#) DOI: [10.1002/jmv.24005](#)]
- 32 **Sagnelli E**, Starnini G, Sagnelli C, Monarca R, Zumbo G, Pontali E, Gabbuti A, Carbonara S, Iardino R, Armignacco O, Babudieri S, Simspe Group. Blood born viral infections, sexually transmitted diseases and latent tuberculosis in Italian prisons: a preliminary report of a large multicenter study. *Eur Rev Med Pharmacol Sci* 2012; **16**: 2142-2146 [PMID: [23280032](#)]
- 33 **Tosti ME**, Alfonsi V, Lacorte E, Mele A, Galli C, Zanetti AR, Romano L; SEIEVA Collaborating Group. Acute Hepatitis B After the Implementation of Universal Vaccination in Italy: Results From 22 Years of Surveillance (1993-2014). *Clin Infect Dis* 2016; **62**: 1412-1418 [PMID: [27009250](#) DOI: [10.1093/cid/ciw162](#)]
- 34 **Daniels D**, Grytdal S, Wasley A; Centers for Disease Control and Prevention (CDC). Surveillance for acute viral hepatitis - United States, 2007. *MMWR Surveill Summ* 2009; **58**: 1 [PMID: [19478727](#)]
- 35 **Chen A**, Ho YS, Tu YC, Shieh SD, Cheng TC. Hepatitis B virus-associated membranous glomerulonephropathy. *J Clin Gastroenterol* 1988; **10**: 243-246 [PMID: [2980755](#) DOI: [10.1097/00004836-198806000-00002](#)]
- 36 **Trepo C**, Guillemin L. Polyarteritis nodosa and extrahepatic manifestations of HBV infection: the case against autoimmune intervention in pathogenesis. *J Autoimmun* 2001; **16**: 269-274 [PMID: [11334492](#) DOI: [10.1006/jaut.2000.0502](#)]
- 37 **Berk PD**, Popper H. Fulminant hepatic failure. *Am J Gastroenterol* 1978; **69**: 349-400 [PMID: [685949](#)]
- 38 **Lavanchy D**. Worldwide epidemiology of HBV infection, disease burden, and vaccine prevention. *J Clin Virol* 2005; **34** Suppl 1: S1-S3 [PMID: [16461208](#) DOI: [10.1016/S1386-6532\(05\)00384-7](#)]
- 39 **Chou HH**, Chien WH, Wu LL, Cheng CH, Chung CH, Horgn JH, Ni YH, Tseng HT, Wu D, Lu X, Wang HY, Chen PJ, Chen DS. Age-related immune clearance of hepatitis B virus infection requires the establishment of gut microbiota. *Proc Natl Acad Sci U S A* 2015; **112**: 2175-2180 [PMID: [25646429](#) DOI: [10.1073/pnas.1424775112](#)]
- 40 **Taylor BC**, Yuan JM, Shamlilian TA, Shaikat A, Kane RL, Wilt TJ. Clinical outcomes in adults with chronic hepatitis B in association with patient and viral characteristics: A systematic review of evidence. *Hepatology* 2009; **49**: S85-S95 [PMID: [19399797](#) DOI: [10.1002/hep.22929](#)]
- 41 **Liang TJ**. Hepatitis B: the virus and disease. *Hepatology* 2009; **49**: S13-S21 [PMID: [19399811](#) DOI: [10.1002/hep.22881](#)]
- 42 **Liaw YF**, Chu CM. Hepatitis B virus infection. *Lancet* 2009; **373**: 582-592 [DOI: [10.1016/S0140-6736\(09\)60207-5](#)]
- 43 **Garfein RS**, Bower WA, Loney CM, Hutin YJ, Xia GL, Jawanda J, Groom AV, Nainan OV, Murphy JS, Bell BP. Factors associated with fulminant liver failure during an outbreak among injection drug users with acute hepatitis B. *Hepatology* 2004; **40**: 865-873 [PMID: [15382123](#) DOI: [10.1002/hep.20383](#)]
- 44 **Aoki M**. [Maintenance of the posture and synergic reflex]. *Nihon Rinsho* 1975; **33**: 2978-2984 [PMID: [1239534](#) DOI: [10.1053/jhep.2002.36509](#)]
- 45 **Stroffolini T**, Sagnelli E, Sagnelli C, Morisco F, Babudieri S, Furlan C, Pirisi M, Russello M, Smedile A, Pisaturo M, Almasio PL. Characteristics and Changes over Time of Alcohol-Related Chronic Liver Diseases in Italy. *Can J Gastroenterol Hepatol* 2018; **2018**: 9151820 [PMID: [30345260](#) DOI: [10.1155/2018/9151820](#)]
- 46 **Stroffolini T**, Sagnelli E, Andriulli A, Colloredo G, Furlan C, Gaeta GB, Morisco F, Pirisi M, Rosina F, Sagnelli C, Smedile A, Almasio PL; EPACRON study group. Sex difference in the interaction of alcohol intake, hepatitis B virus, and hepatitis C virus on the risk of cirrhosis. *PLoS One* 2017; **12**: e0185710

- [PMID: 29140988 DOI: 10.1371/journal.pone.0185710]
- 47 **Sagnelli E**, Sagnelli C, Macera M, Pisaturo M, Coppola N. An update on the treatment options for HBV/HCV coinfection. *Expert Opin Pharmacother* 2017; **18**: 1691-1702 [PMID: 29081251 DOI: 10.1080/14656566.2017.1398233]
- 48 **Sagnelli C**, Ciccozzi M, Alessio L, Cella E, Gualdieri L, Pisaturo M, Minichini C, Di Caprio G, Starace M, Onorato L, Capoprese M, Occhiello L, Angeletti S, Scotto G, Macera M, Sagnelli E, Coppola N. HBV molecular epidemiology and clinical condition of immigrants living in Italy. *Infection* 2018; **46**: 523-531 [PMID: 29796738 DOI: 10.1007/s15010-018-1153-1]
- 49 **Ciccozzi M**, Lai A, Zehender G, Borsetti A, Cella E, Ciotti M, Sagnelli E, Sagnelli C, Angeletti S. The phylogenetic approach for viral infectious disease evolution and epidemiology: an updating review. *J Med Virol* 2019 [PMID: 31243773 DOI: 10.1002/jmv.25526.]
- 50 **Stroffolini T**, Sagnelli E, Sagnelli C, Smedile A, Furlan C, Morisco F, Coppola N, Andriulli A, Almasio PL. The burden of HBV infection in HCV patients in Italy and the risk of reactivation under DAA therapy. *Dig Liver Dis* 2019; **51**: 434-437 [DOI: 10.1016/j.dld.2018.09.010]
- 51 **Stroffolini T**, Sagnelli E, Sagnelli C, Russello M, De Luca M, Rosina F, Cacopardo B, Brancaccio G, Furlan C, Gaeta GB, Licata A, Almasio PL; behalf of EPACRON study group. Hepatitis delta infection in Italian patients: towards the end of the story? *Infection* 2017; **45**: 277-281 [PMID: 27817147 DOI: 10.1007/s15010-016-0956-1]
- 52 **Aragri M**, Alteri C, Battisti A, Di Carlo D, Minichini C, Sagnelli C, Bellocchi MC, Pisaturo MA, Starace M, Armenia D, Carioti L, Pollicita M, Salpini R, Sagnelli E, Perno CF, Coppola N, Svicher V. Multiple Hepatitis B Virus (HBV) Quasispecies and Immune-Escape Mutations Are Present in HBV Surface Antigen and Reverse Transcriptase of Patients With Acute Hepatitis B. *J Infect Dis* 2016; **213**: 1897-1905 [PMID: 26908731 DOI: 10.1093/infdis/jiw049]
- 53 **Coppola N**, Onorato L, Minichini C, Di Caprio G, Starace M, Sagnelli C, Sagnelli E. Clinical significance of hepatitis B surface antigen mutants. *World J Hepatol* 2015; **7**: 2729-2739 [PMID: 26644816 DOI: 10.4254/wjh.v7.i27.2729]
- 54 **Pollicino T**, Raimondo G. Occult hepatitis B infection. *J Hepatol* 2014; **61**: 688-689 [PMID: 24976111 DOI: 10.1016/j.jhep.2014.04.036]
- 55 **Sagnelli C**, Macera M, Pisaturo M, Zampino R, Coppola M, Sagnelli E. Occult HBV infection in the oncohematological setting. *Infection* 2016; **44**: 575-582 [PMID: 27076347 DOI: 10.1007/s15010-016-0891-1]
- 56 **Coppola N**, Onorato L, Pisaturo M, Macera M, Sagnelli C, Martini S, Sagnelli E. Role of occult hepatitis B virus infection in chronic hepatitis C. *World J Gastroenterol* 2015; **21**: 11931-11940 [PMID: 26576082 DOI: 10.3748/wjg.v21.i42.11931]
- 57 **Squadrito G**, Spinella R, Raimondo G. The clinical significance of occult HBV infection. *Ann Gastroenterol* 2014; **27**: 15-19 [PMID: 24714731]
- 58 **Raimondo G**, Caccamo G, Filomia R, Pollicino T. Occult HBV infection. *Semin Immunopathol* 2013; **35**: 39-52 [PMID: 22829332 DOI: 10.1007/s00281-012-0327-7]
- 59 **Sagnelli E**, Pisaturo M, Martini S, Filippini P, Sagnelli C, Coppola N. Clinical impact of occult hepatitis B virus infection in immunosuppressed patients. *World J Hepatol* 2014; **6**: 384-393 [PMID: 25018849 DOI: 10.4254/wjh.v6.i6.384]
- 60 **Tonziello G**, Pisaturo M, Sica A, Ferrara MG, Sagnelli C, Pasquale G, Sagnelli E, Guastafierro S, Coppola N. Transient reactivation of occult hepatitis B virus infection despite lamivudine prophylaxis in a patient treated for non-Hodgkin lymphoma. *Infection* 2013; **41**: 225-229 [PMID: 22855434 DOI: 10.1007/s15010-012-0305-y]
- 61 **Coppola N**, Tonziello G, Pisaturo M, Messina V, Guastafierro S, Fiore M, Iodice V, Sagnelli C, Stanzione M, Capoluongo N, Pasquale G, Sagnelli E. Reactivation of overt and occult hepatitis B infection in various immunosuppressive settings. *J Med Virol* 2011; **83**: 1909-1916 [PMID: 21915865 DOI: 10.1002/jmv.22199]
- 62 **Sagnelli E**, Imparato M, Coppola N, Pisapia R, Sagnelli C, Messina V, Piai G, Stanzione M, Bruno M, Moggio G, Caprio N, Pasquale G, Del Vecchio Blanco C. Diagnosis and clinical impact of occult hepatitis B infection in patients with biopsy proven chronic hepatitis C: a multicenter study. *J Med Virol* 2008; **80**: 1547-1553 [PMID: 18649338 DOI: 10.1002/jmv.21239]
- 63 **Filippini P**, Coppola N, Pisapia R, Scolastico C, Marrocco C, Zaccariello A, Nacca C, Sagnelli C, De Stefano G, Ferraro T, De Stefano C, Sagnelli E. Impact of occult hepatitis B virus infection in HIV patients naive for antiretroviral therapy. *AIDS* 2006; **20**: 1253-1260 [PMID: 16816553 DOI: 10.1097/01.aids.0000232232.41877.2a]
- 64 **McMahon BJ**. The natural history of chronic hepatitis B virus infection. *Hepatology* 2009; **49**: S45-S55 [PMID: 19399792 DOI: 10.1002/hep.22898]
- 65 **Iloeje UH**, Yang HI, Su J, Jen CL, You SL, Chen CJ; Risk Evaluation of Viral Load Elevation and Associated Liver Disease/Cancer-In HBV (the REVEAL-HBV) Study Group. Predicting cirrhosis risk based on the level of circulating hepatitis B viral load. *Gastroenterology* 2006; **130**: 678-686 [PMID: 16530509 DOI: 10.1053/j.gastro.2005.11.016]
- 66 **El-Serag HB**. Epidemiology of viral hepatitis and hepatocellular carcinoma. *Gastroenterology* 2012; **142**: 1264-1273.e1 [PMID: 22537432 DOI: 10.1053/j.gastro.2011.12.061]
- 67 **Sagnelli E**, Potenza N, Onorato L, Sagnelli C, Coppola N, Russo A. Micro-RNAs in hepatitis B virus-related chronic liver diseases and hepatocellular carcinoma. *World J Hepatol* 2018; **10**: 558-570 [PMID: 30310534 DOI: 10.4254/wjh.v10.i9.558]
- 68 **Mosca N**, Castiello F, Coppola N, Trotta MC, Sagnelli C, Pisaturo M, Sagnelli E, Russo A, Potenza N. Functional interplay between hepatitis B virus X protein and human miR-125a in HBV infection. *Biochem Biophys Res Commun* 2014; **449**: 141-145 [PMID: 24824183 DOI: 10.1016/j.bbrc.2014.05.009]
- 69 **Coppola N**, Potenza N, Pisaturo M, Mosca N, Tonziello G, Signoriello G, Messina V, Sagnelli C, Russo A, Sagnelli E. Liver microRNA hsa-miR-125a-5p in HBV chronic infection: correlation with HBV replication and disease progression. *PLoS One* 2013; **8**: e65336 [PMID: 23843939 DOI: 10.1371/journal.pone.0065336]
- 70 **Coppola N**, Onorato L, Sagnelli C, Sagnelli E, Angelillo IF. Association between anti-HBc positivity and hepatocellular carcinoma in HBsAg-negative subjects with chronic liver disease: A meta-analysis. *Medicine (Baltimore)* 2016; **95**: e4311 [PMID: 27472708 DOI: 10.1097/MD.0000000000004311]
- 71 **Mast EE**, Weinbaum CM, Fiore AE, Alter MJ, Bell BP, Finelli L, Rodewald LE, Douglas JM, Janssen RS, Ward JW; Advisory Committee on Immunization Practices (ACIP) Centers for Disease Control and Prevention (CDC). A comprehensive immunization strategy to eliminate transmission of hepatitis B virus

- infection in the United States: recommendations of the Advisory Committee on Immunization Practices (ACIP) Part II: immunization of adults. *MMWR Recomm Rep* 2006; **55**: 1-33; quiz CE1-4 [PMID: 17159833]
- 72 **Kim DK**, Riley LE, Harriman KH, Hunter P, Bridges CB; Advisory Committee on Immunization Practices. Recommended Immunization Schedule for Adults Aged 19 Years or Older, United States, 2017. *Ann Intern Med* 2017; **166**: 209-219 [PMID: 28166560 DOI: 10.7326/M16-2936]
- 73 **Centers for Disease Control and Prevention (CDC)**. Use of hepatitis B vaccination for adults with diabetes mellitus: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Morb Mortal Wkly Rep* 2011; **60**: 1709-1711 [PMID: 22189894]
- 74 **Centers for Disease Control and Prevention (CDC)**. Guidelines for Vaccinating Kidney Dialysis Patients and Patients with Chronic Kidney Disease 2015. Available from: [www.cdc.gov/vaccines/pubs/down\\_loads/dialysis-guide-2012.pdf](http://www.cdc.gov/vaccines/pubs/down_loads/dialysis-guide-2012.pdf)
- 75 **Rubin LG**, Levin MJ, Ljungman P, Davies EG, Avery R, Tomblyn M, Bousvaros A, Dhanireddy S, Sung L, Keyserling H, Kang I; Infectious Diseases Society of America. 2013 IDSA clinical practice guideline for vaccination of the immunocompromised host. *Clin Infect Dis* 2014; **58**: e44-100 [PMID: 24311479 DOI: 10.1093/cid/cit684]
- 76 **Assad S**, Francis A. Over a decade of experience with a yeast recombinant hepatitis B vaccine. *Vaccine* 1999; **18**: 57-67 [PMID: 10501235 DOI: 10.1016/S0264-410X(99)00179-6]
- 77 **Venters C**, Graham W, Cassidy W. Recombivax-HB: perspectives past, present and future. *Expert Rev Vaccines* 2004; **3**: 119-129 [PMID: 15056038 DOI: 10.1586/14760584.3.2.119]
- 78 **McMahon BJ**, Bulkow LR, Singleton RJ, Williams J, Snowball M, Homan C, Parkinson AJ. Elimination of hepatocellular carcinoma and acute hepatitis B in children 25 years after a hepatitis B newborn and catch-up immunization program. *Hepatology* 2011; **54**: 801-807 [PMID: 21618565 DOI: 10.1002/hep.2444231]
- 79 **Bruce MG**, Bruden D, Hurlburt D, Zanis C, Thompson G, Rea L, Toomey M, Townshend-Bulson L, Rudolph K, Bulkow L, Spradling PR, Baum R, Hennessy T, McMahon BJ. Antibody Levels and Protection After Hepatitis B Vaccine: Results of a 30-Year Follow-up Study and Response to a Booster Dose. *J Infect Dis* 2016; **214**: 16-22 [PMID: 26802139 DOI: 10.1093/infdis/jiv74832]
- 80 **Sagnelli E**, Stroffolini T, Sagnelli C, Morisco F, Coppola N, Smedile A, Pisaturo M, Colloredo G, Babudieri S, Licata A, Brancaccio G, Andriulli A, Almasio PL, Gaeta GB; EPACRON study group. Influence of universal HBV vaccination on chronic HBV infection in Italy: Results of a cross-sectional multicenter study. *J Med Virol* 2017; **89**: 2138-2143 [PMID: 28608566 DOI: 10.1002/jmv.24873]
- 81 **Simons BC**, Spradling PR, Bruden DJ, Zanis C, Case S, Choromanski TL, Apodaca M, Brogdon HD, Dwyer G, Snowball M, Negus S, Bruce MG, Morishima C, Knall C, McMahon BJ. A Longitudinal Hepatitis B Vaccine Cohort Demonstrates Long-lasting Hepatitis B Virus (HBV) Cellular Immunity Despite Loss of Antibody Against HBV Surface Antigen. *J Infect Dis* 2016; **214**: 273-280 [PMID: 27056956 DOI: 10.1093/infdis/jiw142]
- 82 **Grosso G**, Mistretta A, Marventano S, Ferranti R, Mauro L, Cunsolo R, Proietti L, Malaguarnera M. Long-term persistence of seroprotection by hepatitis B vaccination in healthcare workers of southern Italy. *Hepat Mon* 2012; **12**: e6025 [PMID: 23087756 DOI: 10.5812/hepatmon.6025]
- 83 **Morowatishaifabad MA**, Zare Sakhvidi MJ, Gholianavval M, Masoudi Boroujeni D, Alavijeh MM. Predictors of Hepatitis B Preventive Behavioral Intentions in Healthcare Workers. *Saf Health Work* 2015; **6**: 139-142 [PMID: 26106514 DOI: 10.1016/j.shaw.2014.12.001]
- 84 **Maltezou HC**, Gargalianos P, Nikolaidis P, Katerelos P, Tedoma N, Maltezos E, Lazanas M. Attitudes towards mandatory vaccination and vaccination coverage against vaccine-preventable diseases among health-care workers in tertiary-care hospitals. *J Infect* 2012; **64**: 319-324 [PMID: 22198739 DOI: 10.1016/j.jinf.2011.12.004]
- 85 **Singhal V**, Bora D, Singh S. Prevalence of Hepatitis B virus infection in healthcare workers of a tertiary care centre in India and their vaccination status. *J Vaccines Vaccin* 2011; **2**: 2 [DOI: 10.4172/2157-7560.1000118]
- 86 **Galanakis E**, D'Ancona F, Jansen A, Lopalco PL; VENICE (Vaccine European New Integrated Collaboration Effort) National Gatekeepers, Contact Points. The issue of mandatory vaccination for healthcare workers in Europe. *Expert Rev Vaccines* 2014; **13**: 277-283 [PMID: 24350731 DOI: 10.1586/14760584.2014.869174]
- 87 **Abiola AO**, Omoyeni OE, Akodu BA. Knowledge, attitude and practice of hepatitis B vaccination among health workers at the Lagos State accident and emergency centre, Toll-Gate, Alausa, Lagos State. *West Afr J Med* 2013; **32**: 257-262 [PMID: 24488279]
- 88 **Doebbeling BN**, Ferguson KJ, Kohout FJ. Predictors of hepatitis B vaccine acceptance in health care workers. *Med Care* 1996; **34**: 58-72 [PMID: 8551812 DOI: 10.1097/00005650-199601000-00005]
- 89 **Cohen C**, Holmberg SD, McMahon BJ, Block JM, Brosgart CL, Gish RG, London WT, Block TM. Is chronic hepatitis B being undertreated in the United States? *J Viral Hepat* 2011; **18**: 377-383 [PMID: 21143343 DOI: 10.1111/j.1365-2893.2010.01401.x]
- 90 **Weinbaum CM**, Williams I, Mast EE, Wang SA, Finelli L, Wasley A, Neitzel SM, Ward JW; Centers for Disease Control and Prevention (CDC). Recommendations for identification and public health management of persons with chronic hepatitis B virus infection. *MMWR Recomm Rep* 2008; **57**: 1-20 [PMID: 18802412]
- 91 **Williams WW**, Lu PJ, O'Halloran A, Kim DK, Grohskopf LA, Pilishvili T, Skoff TH, Nelson NP, Harpaz R, Markowitz LE, Rodriguez-Lainz A, Fiebelkorn AP. Surveillance of Vaccination Coverage among Adult Populations - United States, 2015. *MMWR Surveill Summ* 2017; **66**: 1-28 [PMID: 28472027 DOI: 10.15585/mmwr.ss6611a1]
- 92 **Gust ID**. Immunisation against hepatitis B in Taiwan. *Gut* 1996; **38** Suppl 2: S67-S68 [PMID: 8786059 DOI: 10.1136/gut.38.Suppl\_2.S67]
- 93 **Sung JL**. Hepatitis B virus infection and its sequelae in Taiwan. *Gastroenterol Jpn* 1984; **19**: 363-366 [PMID: 6489693 DOI: 10.1007/BF02779126]
- 94 **Ni YH**, Chang MH, Huang LM, Chen HL, Hsu HY, Chiu TY, Tsai KS, Chen DS. Hepatitis B virus infection in children and adolescents in a hyperendemic area: 15 years after mass hepatitis B vaccination. *Ann Intern Med* 2001; **135**: 796-800 [PMID: 11694104 DOI: 10.7326/0003-4819-135-9-200111060-00009]
- 95 **Ni YH**, Chang MH, Wu JF, Hsu HY, Chen HL, Chen DS. Minimization of hepatitis B infection by a 25-year universal vaccination program. *J Hepatol* 2012; **57**: 730-735 [PMID: 22668640 DOI: 10.1016/j.jhep.2012.05.021]



- 96 **Ni YH**, Chang MH, Jan CF, Hsu HY, Chen HL, Wu JF, Chen DS. Continuing Decrease in Hepatitis B Virus Infection 30 Years After Initiation of Infant Vaccination Program in Taiwan. *Clin Gastroenterol Hepatol* 2016; **14**: 1324-1330 [PMID: 27155556 DOI: 10.1016/j.cgh.2016.04.030]
- 97 **Tsen YJ**, Chang MH, Hsu HY, Lee CY, Sung JL, Chen DS. Seroprevalence of hepatitis B virus infection in children in Taipei, 1989: five years after a mass hepatitis B vaccination program. *J Med Virol* 1991; **34**: 96-99 [PMID: 1832440 DOI: 10.1002/jmv.1890340205]
- 98 **Su FH**, Cheng SH, Li CY, Chen JD, Hsiao CY, Chien CC, Yang YC, Hung HH, Chu FY. Hepatitis B seroprevalence and anamnestic response amongst Taiwanese young adults with full vaccination in infancy, 20 years subsequent to national hepatitis B vaccination. *Vaccine* 2007; **25**: 8085-8090 [PMID: 17920732 DOI: 10.1016/j.vaccine.2007.09.013]
- 99 **Hu YC**, Yeh CC, Chen RY, Su CT, Wang WC, Bai CH, Chan CF, Su FH. Seroprevalence of hepatitis B virus in Taiwan 30 years after the commencement of the national vaccination program. *PeerJ* 2018; **6**: e4297 [PMID: 29472994 DOI: 10.7717/peerj.4297]
- 100 **Al-Faleh FZ**, Al-Jeffri M, Ramia S, Al-Rashed R, Arif M, Rezeig M, Al-Toraifi I, Bakhsh M, Mishkhas A, Makki O, Al-Freih H, Mirdad S, AlJuma A, Yasin T, Al-Swailem A, Ayoola A. Seroprevalence of hepatitis B virus infection in Saudi children 8 years after a mass hepatitis B vaccination programme. *J Infect* 1999; **38**: 167-170 [PMID: 10424796]
- 101 **Whittle H**, Jaffar S, Wansbrough M, Mendy M, Dumpis U, Collinson A, Hall A. Observational study of vaccine efficacy 14 years after trial of hepatitis B vaccination in Gambian children. *BMJ* 2002; **325**: 569 [PMID: 12228132 DOI: 10.1136/bmj.325.7364.569]
- 102 **Harpaz R**, McMahon BJ, Margolis HS, Shapiro CN, Havron D, Carpenter G, Bulkow LR, Wainwright RB. Elimination of new chronic hepatitis B virus infections: results of the Alaska immunization program. *J Infect Dis* 2000; **181**: 413-418 [PMID: 10669320 DOI: 10.1086/315259]
- 103 **Rossi C**, Schwartzman K, Oxlade O, Klein MB, Greenaway C. Hepatitis B screening and vaccination strategies for newly arrived adult Canadian immigrants and refugees: a cost-effectiveness analysis. *PLoS One* 2013; **8**: e78548 [PMID: 24205255 DOI: 10.1371/journal.pone.0078548]
- 104 **Liaw YF**, Sung JJ, Chow WC, Farrell G, Lee CZ, Yuen H, Tanwandee T, Tao QM, Shue K, Keene ON, Dixon JS, Gray DF, Sabbat J; Cirrhosis Asian Lamivudine Multicentre Study Group. Lamivudine for patients with chronic hepatitis B and advanced liver disease. *N Engl J Med* 2004; **351**: 1521-1531 [PMID: 15470215 DOI: 10.1056/NEJMoa033364]
- 105 **Lau GK**, Piratvisuth T, Luo KX, Marcellin P, Thongsawat S, Cooksley G, Gane E, Fried MW, Chow WC, Paik SW, Chang WY, Berg T, Flisiak R, McCloud P, Pluck N; Peginterferon Alfa-2a HBeAg-Positive Chronic Hepatitis B Study Group. Peginterferon Alfa-2a, lamivudine, and the combination for HBeAg-positive chronic hepatitis B. *N Engl J Med* 2005; **352**: 2682-2695 [PMID: 15987917 DOI: 10.1056/NEJMoa043470]
- 106 **Janssen HL**, van Zonneveld M, Senturk H, Zeuzem S, Akarca US, Cakaloglu Y, Simon C, So TM, Gerken G, de Man RA, Niesters HG, Zondervan P, Hansen B, Schalm SW; HBV 99-01 Study Group; Rotterdam Foundation for Liver Research. Pegylated interferon alfa-2b alone or in combination with lamivudine for HBeAg-positive chronic hepatitis B: a randomised trial. *Lancet* 2005; **365**: 123-129 [PMID: 15639293 DOI: 10.1016/S0140-6736(05)17701-0]
- 107 **Marcellin P**, Lau GK, Bonino F, Farci P, Hadziyannis S, Jin R, Lu ZM, Piratvisuth T, Germanidis G, Yurdaydin C, Diago M, Gurel S, Lai MY, Button P, Pluck N; Peginterferon Alfa-2a HBeAg-Negative Chronic Hepatitis B Study Group. Peginterferon alfa-2a alone, lamivudine alone, and the two in combination in patients with HBeAg-negative chronic hepatitis B. *N Engl J Med* 2004; **351**: 1206-1217 [PMID: 15371578 DOI: 10.1056/NEJMoa040431]
- 108 **Seto WK**, Lau EH, Wu JT, Hung IF, Leung WK, Cheung KS, Fung J, Lai CL, Yuen MF. Effects of nucleoside analogue prescription for hepatitis B on the incidence of liver cancer in Hong Kong: a territory-wide ecological study. *Aliment Pharmacol Ther* 2017; **45**: 501-509 [PMID: 27976416 DOI: 10.1111/apt.13895]
- 109 **Buti M**, Tsai N, Petersen J, Flisiak R, Gurel S, Krastev Z, Aguilar Schall R, Flaherty JF, Martins EB, Charuorn P, Kitrinis KM, Subramanian GM, Gane E, Marcellin P. Seven-year efficacy and safety of treatment with tenofovir disoproxil fumarate for chronic hepatitis B virus infection. *Dig Dis Sci* 2015; **60**: 1457-1464 [PMID: 25532501 DOI: 10.1007/s10620-014-3486-7]
- 110 **Marcellin P**, Gane E, Buti M, Afdhal N, Sievert W, Jacobson IM, Washington MK, Germanidis G, Flaherty JF, Aguilar Schall R, Bornstein JD, Kitrinis KM, Subramanian GM, McHutchison JG, Heathcote EJ. Regression of cirrhosis during treatment with tenofovir disoproxil fumarate for chronic hepatitis B: a 5-year open-label follow-up study. *Lancet* 2013; **381**: 468-475 [PMID: 23234725 DOI: 10.1016/S0140-6736(12)61425-1]
- 111 **Maier M**, Liebert UG, Wittekind C, Kaiser T, Berg T, Wiegand J. Clinical Relevance of Minimal Residual Viremia during Long-Term Therapy with Nucleos(t)ide Analogues in Patients with Chronic Hepatitis B. *PLoS One* 2013; **8**: e67481 [PMID: 23826307 DOI: 10.1371/journal.pone.0067481]
- 112 **Hosaka T**, Suzuki F, Kobayashi M, Seko Y, Kawamura Y, Sezaki H, Akuta N, Suzuki Y, Saitoh S, Arase Y, Ikeda K, Kobayashi M, Kumada H. Long-term entecavir treatment reduces hepatocellular carcinoma incidence in patients with hepatitis B virus infection. *Hepatology* 2013; **58**: 98-107 [PMID: 23213040 DOI: 10.1002/hep.26180]
- 113 **Singal AK**, Salameh H, Kuo YF, Fontana RJ. Meta-analysis: the impact of oral anti-viral agents on the incidence of hepatocellular carcinoma in chronic hepatitis B. *Aliment Pharmacol Ther* 2013; **38**: 98-106 [PMID: 23713520 DOI: 10.1111/apt.12344]
- 114 **Coffin CS**, Rezaeeaval M, Pang JX, Alcantara L, Klein P, Burak KW, Myers RP. The incidence of hepatocellular carcinoma is reduced in patients with chronic hepatitis B on long-term nucleos(t)ide analogue therapy. *Aliment Pharmacol Ther* 2014; **40**: 1262-1269 [PMID: 25312649 DOI: 10.1111/apt.12990]
- 115 **Wang X**, Liu X, Dang Z, Yu L, Jiang Y, Wang X, Yang Z. Nucleos(t)ide Analogues for Reducing Hepatocellular Carcinoma in Chronic Hepatitis B Patients: A Systematic Review and Meta-Analysis. *Gut Liver* 2019 [PMID: 31158948 DOI: 10.5009/gnl18546.]
- 116 **Wong GL**, Chan HL, Mak CW, Lee SK, Ip ZM, Lam AT, Iu HW, Leung JM, Lai JW, Lo AO, Chan HY, Wong VW. Entecavir treatment reduces hepatic events and deaths in chronic hepatitis B patients with liver cirrhosis. *Hepatology* 2013; **58**: 1537-1547 [PMID: 23389810 DOI: 10.1002/hep.26301]
- 117 **Ono A**, Suzuki F, Kawamura Y, Sezaki H, Hosaka T, Akuta N, Kobayashi M, Suzuki Y, Saitou S, Arase Y, Ikeda K, Kobayashi M, Watahiki S, Mineta R, Kumada H. Long-term continuous entecavir therapy in nucleos(t)ide-naïve chronic hepatitis B patients. *J Hepatol* 2012; **57**: 508-514 [PMID: 22659518 DOI: 10.1016/j.jhep.2012.05.013]

- 10.1016/j.jhep.2012.04.037]
- 118 **Chang TT**, Gish RG, de Man R, Gadano A, Sollano J, Chao YC, Lok AS, Han KH, Goodman Z, Zhu J, Cross A, DeHertogh D, Wilber R, Colonna R, Apelian D; BEHoLD A1463022 Study Group. A comparison of entecavir and lamivudine for HBeAg-positive chronic hepatitis B. *N Engl J Med* 2006; **354**: 1001-1010 [PMID: [16525137](#) DOI: [10.1056/NEJMoa051285](#)]
- 119 **Marcellin P**, Heathcote EJ, Buti M, Gane E, de Man RA, Krastev Z, Germanidis G, Lee SS, Flisiak R, Kaita K, Manns M, Kotzev I, Tchernev K, Buggisch P, Weilert F, Kurdas OO, Shiffman ML, Trinh H, Washington MK, Sorbel J, Anderson J, Snow-Lampart A, Mondou E, Quinn J, Rousseau F. Tenofovir disoproxil fumarate versus adefovir dipivoxil for chronic hepatitis B. *N Engl J Med* 2008; **359**: 2442-2455 [PMID: [19052126](#) DOI: [10.1056/NEJMoa0802878](#)]
- 120 **Chi H**, Hansen BE, Yim C, Arends P, Abu-Amara M, van der Eijk AA, Feld JJ, de Knecht RJ, Wong DK, Janssen HL. Reduced risk of relapse after long-term nucleos(t)ide analogue consolidation therapy for chronic hepatitis B. *Aliment Pharmacol Ther* 2015; **41**: 867-876 [PMID: [25752878](#) DOI: [10.1111/apt.13150](#)]
- 121 **Bam RA**, Birkus G, Babusis D, Cihlar T, Yant SR. Metabolism and antiretroviral activity of tenofovir alafenamide in CD4+ T-cells and macrophages from demographically diverse donors. *Antivir Ther* 2014; **19**: 669-677 [PMID: [24625459](#) DOI: [10.3851/IMP2767](#)]
- 122 **Bam RA**, Yant SR, Cihlar T. Tenofovir alafenamide is not a substrate for renal organic anion transporters (OATs) and does not exhibit OAT-dependent cytotoxicity. *Antivir Ther* 2014; **19**: 687-692 [PMID: [24699134](#) DOI: [10.3851/IMP2770](#)]
- 123 **Sax PE**, Zolopa A, Brar I, Elion R, Ortiz R, Post F, Wang H, Callebaut C, Martin H, Fordyce MW, McCallister S. Tenofovir alafenamide vs. tenofovir disoproxil fumarate in single tablet regimens for initial HIV-1 therapy: a randomized phase 2 study. *J Acquir Immune Defic Syndr* 2014; **67**: 52-58 [PMID: [24872136](#) DOI: [10.1097/QAI.0000000000000225](#)]
- 124 **Buti M**, Gane E, Seto WK, Chan HL, Chuang WL, Stepanova T, Hui AJ, Lim YS, Mehta R, Janssen HL, Acharya SK, Flaherty JF, Massetto B, Cathcart AL, Kim K, Gaggar A, Subramanian GM, McHutchison JG, Pan CQ, Brunetto M, Izumi N, Marcellin P; GS-US-320-0108 Investigators. Tenofovir alafenamide versus tenofovir disoproxil fumarate for the treatment of patients with HBeAg-negative chronic hepatitis B virus infection: a randomised, double-blind, phase 3, non-inferiority trial. *Lancet Gastroenterol Hepatol* 2016; **1**: 196-206 [PMID: [28404092](#) DOI: [10.1016/S2468-1253\(16\)30107-8](#)]
- 125 **Chan HL**, Fung S, Seto WK, Chuang WL, Chen CY, Kim HJ, Hui AJ, Janssen HL, Chowdhury A, Tsang TY, Mehta R, Gane E, Flaherty JF, Massetto B, Gaggar A, Kitrinis KM, Lin L, Subramanian GM, McHutchison JG, Lim YS, Acharya SK, Agarwal K; GS-US-320-0110 Investigators. Tenofovir alafenamide versus tenofovir disoproxil fumarate for the treatment of HBeAg-positive chronic hepatitis B virus infection: a randomised, double-blind, phase 3, non-inferiority trial. *Lancet Gastroenterol Hepatol* 2016; **1**: 185-195 [PMID: [28404091](#) DOI: [10.1016/S2468-](#)]
- 126 **Chan HLY**, Fung S, Seto WK, Chuang WL, Chen CY, Kim HJ, Hui AJ, Janssen HL, Chowdhury A, Tsang TY, Mehta R, Gane E, Flaherty JF, Massetto B, Gaggar A, Kitrinis KM, Lin L, Subramanian GM, McHutchison JG, Lim YS, Acharya SK, Agarwal K; GS-US-320-0110 Investigators. A phase 3 study of tenofovir alafenamide compared with tenofovir disoproxil fumarate in patients with HBeAg positive chronic HBV: week 48 efficacy and safety results. *J Hepatol* 2016; **64**: S161 [DOI: [10.1016/S0168-8278\(16\)01669-X](#)]
- 127 **Buti M**, Gane E, Seto WK, Chan LY, Chuang WL, Hui AJ, Lim YS, Mehta R, Janssen HL, Acharya SK, Flaherty JF, Massetto B, Cathcart A, Dinh P, Subramanian GM, McHutchison JG, Pan C, Brunetto M, Izumi N, Marcellin P. A phase 3 study of tenofovir alafenamide compared with tenofovir disoproxil fumarate in patients with HBeAg negative, chronic hepatitis B: week 48 efficacy and safety results. *J Hepatol* 2016; **64**: S135 [DOI: [10.1016/S0168-8278\(16\)01637-8](#)]
- 128 **Berke JM**, Dehertogh P, Vergauwen K, Van Damme E, Mostmans W, Vandeyck K, Pauwels F. Capsid Assembly Modulators Have a Dual Mechanism of Action in Primary Human Hepatocytes Infected with Hepatitis B Virus. *Antimicrob Agents Chemother* 2017; **61** [PMID: [28584155](#) DOI: [10.1128/AAC.00560-17](#)]
- 129 **Bogomolov P**, Alexandrov A, Voronkova N, Macievich M, Kokina K, Petrachenkova M, Lehr T, Lempp FA, Wedemeyer H, Haag M, Schwab M, Haefeli WE, Blank A, Urban S. Treatment of chronic hepatitis D with the entry inhibitor myrludex B: First results of a phase Ib/IIa study. *J Hepatol* 2016; **65**: 490-498 [PMID: [27132170](#) DOI: [10.1016/j.jhep.2016.04.01](#)]
- 130 **Ivacik D**, Ely A, Ferry N, Arbutnot P. Sustained inhibition of hepatitis B virus replication in vivo using RNAi-activating lentiviruses. *Gene Ther* 2015; **22**: 163-171 [PMID: [25338920](#) DOI: [10.1038/gt.2014.94](#)]
- 131 **Lai CL**, Ahn SH, Lee KS, Um SH, Cho M, Yoon SK, Lee JW, Park NH, Kweon YO, Sohn JH, Lee J, Kim JA, Han KH, Yuen MF. Phase IIb multicentred randomised trial of besifovir (LB80380) versus entecavir in Asian patients with chronic hepatitis B. *Gut* 2014; **63**: 996-1004 [PMID: [23979965](#) DOI: [10.1136/gutjnl-2013-305138](#)]
- 132 **Lee AC**. Exploring combination therapy for curing HBV: Preclinical studies with capsid inhibitor AB-423 and a siRNA Agent, ARB-1740. *Hepatology* 2016; **63**: 122A
- 133 **Yan H**, Zhong G, Xu G, He W, Jing Z, Gao Z, Huang Y, Qi Y, Peng B, Wang H, Fu L, Song M, Chen P, Gao W, Ren B, Sun Y, Cai T, Feng X, Sui J, Li W. Sodium taurocholate cotransporting polypeptide is a functional receptor for human hepatitis B and D virus. *E. eLife* 2012; **1**: e00049 [PMID: [23150796](#) DOI: [10.7554/eLife.00049](#)]
- 134 **Vivekanandan P**, Thomas D, Torbenson M. Methylation regulates hepatitis B viral protein expression. *J Infect Dis* 2009; **199**: 1286-1291 [PMID: [19301974](#) DOI: [10.1086/597614](#)]
- 135 **Kim JW**, Lee SH, Park YS, Hwang JH, Jeong SH, Kim N, Lee DH. Replicative activity of hepatitis B virus is negatively associated with methylation of covalently closed circular DNA in advanced hepatitis B virus infection. *Intervirology* 2011; **54**: 316-325 [PMID: [21242658](#) DOI: [10.1159/000321450](#)]
- 136 **Guo Y**, Li Y, Mu S, Zhang J, Yan Z. Evidence that methylation of hepatitis B virus covalently closed circular DNA in liver tissues of patients with chronic hepatitis B modulates HBV replication. *J Med Virol* 2009; **81**: 1177-1183 [PMID: [19475606](#) DOI: [10.1002/jmv.21525](#)]
- 137 **Pollicino T**, Belloni L, Raffa G, Pediconi N, Squadrito G, Raimondo G, Levrero M. Hepatitis B virus replication is regulated by the acetylation status of hepatitis B virus cccDNA-bound H3 and H4 histones. *Gastroenterology* 2006; **130**: 823-837 [PMID: [16530522](#) DOI: [10.1053/j.gastro.2006.01.001](#)]
- 138 **Yamamoto M**, Hayashi N, Takehara T, Ueda K, Mita E, Tatsumi T, Sasaki Y, Kasahara A, Hori M. Intracellular single-chain antibody against hepatitis B virus core protein inhibits the replication of hepatitis B virus in cultured cells. *Hepatology* 1999; **30**: 300-307 [PMID: [10385671](#)]

- 139 **Lanford RE**, Guerra B, Chavez D, Giavedoni L, Hodara VL, Brasky KM, Fosdick A, Frey CR, Zheng J, Wolfgang G, Halcomb RL, Tumas DB. GS-9620, an oral agonist of Toll-like receptor-7, induces prolonged suppression of hepatitis B virus in chronically infected chimpanzees. *Gastroenterology* 2013; **144**: 1508-1517, 1517.e1-1517.10 [PMID: 23415804 DOI: 10.1053/j.gastro.2013.02.003]
- 140 **Ding Y**, Zhang H, Niu J, Chen H, Liu C, Li X, Wang F. PS-046-Multiple dose study of GLS4JHS, interfering with the assembly of hepatitis B virus core particles, in patients infected with hepatitis B virus. *J Hepatol* 2017; **66**: S27-S28 [DOI: 10.1016/S0168-8278(17)30317-3]
- 141 **King TH**, Kemmler CB, Guo Z, Mann D, Lu Y, Coeshott C, Gehring AJ, Bertoletti A, Ho ZZ, Delaney W, Gaggar A, Subramanian GM, McHutchison JG, Shrivastava S, Lee YJ, Kottitil S, Bellgrau D, Rodell T, Apelian D. A whole recombinant yeast-based therapeutic vaccine elicits HBV X, S and Core specific T cells in mice and activates human T cells recognizing epitopes linked to viral clearance. *PLoS One* 2014; **9**: e101904 [PMID: 25051027 DOI: 10.1371/journal.pone.0101904]
- 142 **Lam AM**, Ren S, Espiritu C, Kelly M, Lau V, Zheng L, Hartman GD, Flores OA, Klumpp K. Hepatitis B Virus Capsid Assembly Modulators, but Not Nucleoside Analogs, Inhibit the Production of Extracellular Pregenomic RNA and Spliced RNA Variants. *Antimicrob Agents Chemother* 2017; **61** [PMID: 28559265 DOI: 10.1128/AAC.00680-17]
- 143 **Lin SR**, Yang HC, Kuo YT, Liu CJ, Yang TY, Sung KC, Lin YY, Wang HY, Wang CC, Shen YC, Wu FY, Kao JH, Chen DS, Chen PJ. The CRISPR/Cas9 System Facilitates Clearance of the Intrahepatic HBV Templates In Vivo. *Mol Ther Nucleic Acids* 2014; **3**: e186 [PMID: 25137139 DOI: 10.1038/mtna.2014]
- 144 **Liu J**, Zhang E, Ma Z, Wu W, Kosinska A, Zhang X, Möller I, Seiz P, Glebe D, Wang B, Yang D, Lu M, Roggendorf M. Enhancing virus-specific immunity in vivo by combining therapeutic vaccination and PD-L1 blockade in chronic hepadnaviral infection. *PLoS Pathog* 2014; **10**: e1003856 [PMID: 24391505 DOI: 10.1371/journal.ppat.1003856]
- 145 **Lucifora J**, Xia Y, Reisinger F, Zhang K, Stadler D, Cheng X, Sprinzl MF, Koppensteiner H, Makowska Z, Volz T, Remouchamps C, Chou WM, Thasler WE, Hüser N, Durantel D, Liang TJ, Münk C, Heim MH, Browning JL, DeJardin E, Dandri M, Schindler M, Heikenwalder M, Protzer U. Specific and nonhepatotoxic degradation of nuclear hepatitis B virus cccDNA. *Science* 2014; **343**: 1221-1228 [PMID: 24557838 DOI: 10.1126/science.1243462]
- 146 **Mowa MB**, Crowther C, Ely A, Arbuthnot P. Inhibition of hepatitis B virus replication by helper dependent adenoviral vectors expressing artificial anti-HBV pri-miRs from a liver-specific promoter. *Biomed Res Int* 2014; **2014**: 718743 [PMID: 25003129 DOI: 10.1155/2014/718743]
- 147 **Schiffer JT**, Swan DA, Stone D, Jerome KR. Predictors of hepatitis B cure using gene therapy to deliver DNA cleavage enzymes: a mathematical modeling approach. *PLoS Comput Biol* 2013; **9**: e1003131 [PMID: 23861664 DOI: 10.1371/journal.pcbi.1003131]
- 148 **Sebestyén MG**, Wong SC, Trubetskoy V, Lewis DL, Wooddell CI. Targeted in vivo delivery of siRNA and an endosome-releasing agent to hepatocytes. *Methods Mol Biol* 2015; **1218**: 163-186 [PMID: 25319651 DOI: 10.1007/978-1-4939-1538-5\_10]
- 149 **Seeger C**, Sohn JA. Targeting Hepatitis B Virus With CRISPR/Cas9. *Mol Ther Nucleic Acids* 2014; **3**: e216 [PMID: 25514649 DOI: 10.1038/mtna.2014.68]
- 150 **Seto WK**, Yuen MF. New pharmacological approaches to a functional cure of hepatitis B. *Clin Liver Dis (Hoboken)* 2016; **8**: 83-88 [PMID: 31041070 DOI: 10.1002/eld.577]





Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160 Pleasanton, CA 94566, USA  
Telephone: +1-925 2238242  
Fax: +1-925-2238243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>



# World Journal of *Clinical Infectious Diseases*

*World J Clin Infect Dis* 2019 December 15; 9(3): 23-30



**CASE REPORT**

- 23 *Serratia marcescens* and other non-AACEK GNB endocarditis: A case report and review of literature  
Mertes H, Morissens M, Mahadeb B, Maillart E, Moreau A, Clevenbergh P

**ABOUT COVER**

Editorial Board Member of *World Journal of Clinical Infectious Diseases*,  
 Ahmed Morad Asaad, MD, Professor, Microbiology Department, College of  
 Medicine, Zagazig University, Zagazig 44519, Egypt.

**AIMS AND SCOPE**

The primary aim of *World Journal of Clinical Infectious Diseases (WJCID, World J Clin Infect Dis)* is to provide scholars and readers from various fields of infectious diseases with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

*WJCID* mainly publishes articles reporting research results and findings obtained in the field of infectious diseases and covering a wide range of topics including community-acquired infections, cross infection, eye infections, focal infection, infectious gingivitis, intraabdominal infections, laboratory infection, Ludwig's angina, necrotizing ulcerative periodontitis, opportunistic infections, pelvic infection, pregnancy complications, prosthesis-related infections, reproductive tract infections, respiratory tract infections, sepsis, sexually transmitted diseases, skin diseases, soft tissue infections, suppuration, toxemia, urinary tract infections, and wound infection.

**INDEXING/ABSTRACTING**

*World Journal of Clinical Infectious Diseases* is now indexed in China National Knowledge Infrastructure (CNKI), China Science and Technology Journal Database (CSTJ), and Superstar Journals Database.

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Responsible Electronic Editor: *Lu-Lu Qi*  
 Proofing Production Department Director: *Xiang Li*

**NAME OF JOURNAL**

*World Journal of Clinical Infectious Diseases*

**ISSN**

ISSN 2220-3176 (online)

**LAUNCH DATE**

December 30, 2011

**FREQUENCY**

Irregular

**EDITORS-IN-CHIEF**

Joao Mesquita, Caterina Sagnelli

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/2220-3176/editorialboard.htm>

**EDITORIAL OFFICE**

Ya-Juan Ma, Director

**PUBLICATION DATE**

December 15, 2019

**COPYRIGHT**

© 2019 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Serratia marcescens and other non-AACEK GNB endocarditis: A case report and review of literature

Helena Mertes, Marielle Morissens, Bhavna Mahadeb, Evelyne Maillart, Anthony Moreau, Philippe Clevenbergh

**ORCID number:** Helena Mertes (0000-0003-0612-0452); Marielle Morissens (0000-0003-1487-6583); Bhavna Mahadeb (0000-0002-9469-4132); Evelyne Maillart (0000-0002-5354-068X); Anthony Moreau (0000-0003-3511-0587); Philippe Clevenbergh (0000-0002-0522-9686).

**Author contributions:** All authors equally contributed to this paper with regards to the conception and design of the study, literature review and analysis, drafting and critical revision and editing, and final approval of the final version.

**Informed consent statement:** Informed consent was obtained from the patient.

**Conflict-of-interest statement:** The authors report no conflicts of interest.

**CARE Checklist (2016) statement:** The guidelines of the "CARE Checklist - 2016: Information for writing a case report" have been adopted.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See:

**Helena Mertes**, Infectious Disease Department, Ziekenhuis Netwerk Antwerpen Middelheim, Antwerpen 2020, Belgium

**Marielle Morissens**, Cardiology Department, Brugmann University Hospital, Brussels 1020, Belgium

**Bhavna Mahadeb**, Microbiology Department, Brugmann University Hospital, Brussels 1020, Belgium

**Evelyne Maillart, Philippe Clevenbergh**, Infectious Diseases Department, Brugmann University Hospital, Brussels 1020, Belgium

**Anthony Moreau**, Intensive Care Unit, Erasmus University Hospital, Intensive Care Unit, Brussels 1070, Belgium

**Corresponding author:** Helena Mertes, MD, Doctor, Infectious Disease Department, Ziekenhuis Netwerk Antwerpen Middelheim, Lindendreef 1, Antwerpen 2020, Belgium.

[hellibi@hotmail.com](mailto:hellibi@hotmail.com)

**Telephone:** +32-3-2803860

### Abstract

#### BACKGROUND

Non-*Aggregatibacter aphrophilus*, *Aggregatibacter actinomycetemcomitans*, *Cardiobacterium hominis*, *Eikenella corrodens*, *Kingella spp.* (non-AACEK) gram-negative bacilli (GNBs) are an infrequent and challenging cause of endocarditis associated previously with mainly intravenous drug use. Currently, this pathology has increasingly become a healthcare-associated issue. Current guidelines do not clearly define the management of non-AACEK GNB endocarditis due to a lack of prospective trials. We review characteristics, outcomes and treatment of non-AACEK GNB endocarditis, in particular *Serratia marcescens* endocarditis.

#### CASE SUMMARY

We describe the case report of a 46-year-old man who presented to the emergency department with high-grade fever and a purulent exudate on an intracardiac device site. *Serratia marcescens* mitral valve endocarditis as a consequence of complicated generator pocket infection was diagnosed. The patient was treated with complete device removal and a long course of broad-spectrum antibiotics for 6 wk after surgery with intravenous piperacillin-tazobactam and ciprofloxacin, which was later switched to oral ciprofloxacin and

<http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Unsolicited manuscript

**Received:** May 23, 2019

**Peer-review started:** May 23, 2019

**First decision:** August 7, 2019

**Revised:** September 3, 2019

**Accepted:** November 26, 2019

**Article in press:** November 26, 2019

**Published online:** December 15, 2019

**P-Reviewer:** Schwan WR

**S-Editor:** Ma YJ

**L-Editor:** A

**E-Editor:** Qi LL



sulfamethoxazole-trimethoprim. The patient had complete resolution of symptoms and inflammatory parameters at the end of the treatment and at follow-up.

### CONCLUSION

Long-term dual-antibiotic therapy containing a beta-lactam is indicated for most non-AACEK GNB endocarditis, whereas valve surgery may not be necessary in all patients.

**Key words:** Non-AACEK gram-negative bacilli endocarditis; *Serratia marcescens*; Healthcare-associated; Intravenous drug use; Case report; Dual-antibiotic therapy

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** While gram-negative bacillus (GNB) Non-*Aggregatibacter aphrophilus*, *Aggregatibacter actinomycetemcomitans*, *Cardiobacterium hominis*, *Eikenella corrodens*, *Kingella spp.* (non-AACEK) endocarditis has been associated with mainly intravenous drug use, the role of healthcare-associated contact has been highlighted in two prospective observational studies. Our aim was to review the characteristics and management of non-AACEK GNB endocarditis, especially in the case of *Serratia marcescens*. This bacterium has become a rare cause of endocarditis, but community acquisition still has an important role in this disease. We discuss treatment options, supporting long-term dual-antibiotic treatment as the preferred option for most patients with non-AACEK GNB endocarditis, whereas valve surgery does not seem to be necessary in all patients.

**Citation:** Mertes H, Morissens M, Mahadeb B, Maillart E, Moreau A, Clevenbergh P. *Serratia marcescens* and other non-AACEK GNB endocarditis: A case report and review of literature. *World J Clin Infect Dis* 2019; 9(3): 23-30

**URL:** <https://www.wjnet.com/2220-3176/full/v9/i3/23.htm>

**DOI:** <https://dx.doi.org/10.5495/wjcid.v9.i3.23>

## INTRODUCTION

Gram-negative bacillus (GNB) non-*Aggregatibacter aphrophilus*, *Aggregatibacter actinomycetemcomitans*, *Cardiobacterium hominis*, *Eikenella corrodens*, *Kingella spp.* (non-AACEK) bacteria are a rare cause of infectious endocarditis. The International Collaboration on Endocarditis (ICE) prospective study from Morpeth *et al*<sup>[1]</sup> showed that among 2761 patients with definite endocarditis, only 49 (1.8%) had endocarditis due to non-AACEK GNB<sup>[1]</sup>. The recent Italian Endocarditis Study (SEI) from Falcone *et al*<sup>[2]</sup> reported a slightly higher incidence of 3.3% (58 patients) among 1722 patients studied. The most prevalent bacteria found in these observational prospective studies about GNB non-AACEK endocarditis were *Escherichia coli* (29 and 31%), *Pseudomonas aeruginosa* (19 and 22%) and *Klebsiella pneumoniae* (10%). In contrast, *Serratia marcescens* is less common and represents between 3.5% and 8% of endocarditis pathogens<sup>[1,2]</sup>.

Historically, in the 1970s and 1980s, GNB non-AACEK endocarditis was associated with nosocomial exposure and IVDU<sup>[3-7]</sup>. *Serratia marcescens* was a predominant cause of community-acquired endocarditis, as highlighted by Mills *et al*<sup>[3]</sup> and Cooper *et al*<sup>[4]</sup>. In their reviews, individuals with a history of IVDU accounted for 88 and 89%, respectively, of patients with *Serratia marcescens* endocarditis. Currently, GNB non-AACEK endocarditis has become a healthcare-associated (HCA) issue<sup>[1,2]</sup>. Morpeth *et al*<sup>[1]</sup> were first to describe that association in a prospective observational study: 57% of non-AACEK GNB endocarditis cases were HCA (nosocomial and non-nosocomial), predominantly in individuals undergoing non-dental invasive procedures and in patients with intracardiac devices (ICDs). Falcone *et al*<sup>[2]</sup> reported that 44.7% of GNB non-AACEK endocarditis cases were HCA (nosocomial and non-nosocomial), but for most patients, the acquisition was community acquired (55.2%), which was again associated with the presence of an ICD (OR = 3.6) but also with immunosuppression (OR = 5.16). In both studies, the most important source of infection was the genitourinary tract<sup>[1,2]</sup>.

Currently, mortality related to GNB non-AACEK endocarditis has decreased from



between 30% and 68% in the 1980s<sup>[3-7]</sup> to between 13% and 24%<sup>[1,2]</sup>. Cardiac valve surgery is not associated with better outcomes except for in those individuals presenting with complications (heart failure, cardiac abscess, fistula, dehiscence and valve perforation). The multidrug resistance patterns of the bacteria seem to play an important role in the in-hospital mortality rate (HR = 21.89)<sup>[2]</sup>.

European<sup>[8]</sup>, American<sup>[9]</sup> and British<sup>[10]</sup> guidelines do not specifically define treatment of GNB non-AACEK endocarditis due to a lack of prospective studies. They recommend long-term (6 wk) combined bactericidal antibiotic treatment associated with surgery<sup>[8-10]</sup>: Beta-lactams should be the cornerstone of the antibiotic regimen and combined with aminoglycosides (AGs). Sulfamethoxazole-trimethoprim or fluoroquinolones (FQs) could also be added to beta-lactams or reinforce the combined regimen with a BL and an AG<sup>[8,9]</sup>. In the ICE study<sup>[1]</sup>, most patients (57%) were treated equally with a combination of a BL and either an AG or FQ. Eight percent of the patients were treated with a triple antibiotic therapy, whereas 14% received a monotherapy regimen consisting of a BL<sup>[1]</sup>. In the more recent Italian Endocarditis Study (SEI)<sup>[2]</sup>, 30% of the patients were treated with BL monotherapy, while 61% received combination therapy consisting of a BL and an AG in 34% and BL with FQ in 18%. Triple therapy and regimens containing sulfamethoxazole-trimethoprim were administered in only 4% of cases<sup>[2]</sup>. In both studies, there was no comparison of mortality with regard to each antibiotic regimen chosen, but combination therapy did not show superiority compared to monotherapy<sup>[2]</sup>.

*Serratia marcescens* is a gram-negative rod belonging to the family *Enterobacteriaceae*. This ubiquitous bacterium is a human opportunistic pathogen and has been implicated in septicaemia, ventilator-associated pneumonia, meningitis, endocarditis, and urinary tract and HCA wound infections<sup>[1,2,11-13]</sup>. This bacterium has the ability to proliferate in moist environments (such as disinfectants, intravenous solutions and different medical materials) and has therefore been responsible for nosocomial outbreaks<sup>[14]</sup>. *Serratia marcescens* has multiple pathogenicity and virulence factors: adhesins, lipopolysaccharides, fimbriae and siderophores, which facilitate host penetration, adherence to solid surfaces and resistance to serum killing<sup>[13,14]</sup>. *Serratia marcescens* carries a chromosomally encoded AmpC-type beta-lactamase that confers inducible resistance to beta-lactams when exposed to them<sup>[12]</sup>. It then readily hydrolyses penicillins and cephalosporins, including those of the third generation, and is responsible for the reduced activity of other beta-lactams<sup>[12]</sup>. This pathogen is also capable of acquiring mobile genetic elements encoding resistance determinants, such as extended-spectrum beta-lactamases or metallo beta-lactamases<sup>[14]</sup>.

Since the 1990s, the English literature has only reported 15 cases of *Serratia marcescens* endocarditis in adults: In addition to 6 cases reported in the prospective studies discussed earlier<sup>[1,2]</sup>, 9 case reports, including our case, have been published (Table 1). The mean age of the patients in the case reports is 53 years. In 4 of the 9 patients (44.4%), endocarditis was HCA<sup>[15-17]</sup>. It was community-acquired in the remaining 5 patients (55.6%), mainly due to IVD use<sup>[12,18-21]</sup>. Other risk factors identified were immunosuppression (44.4%), venous catheter presence (33.3%), recent cardiac surgery in one case and ICD presence in 2 patients. All patients were treated with combination beta-lactam-containing antibiotic regimens except for 3 patients treated with monotherapy: One patient received ceftriaxone, another meropenem, and the last was treated with ciprofloxacin. Only 2 patients benefited from valve replacement surgery. The patient with ICD-related endocarditis and our patient underwent percutaneous pacemaker extraction. Mortality in this series of case reports was 22%, with death attributed to massive heart failure with surgery contraindicated due to the presence of cerebral abscesses in one case and the second due to cerebral haemorrhage.

## CASE PRESENTATION

A 46-year-old man was admitted to the cardiology unit for suspicion of an intracardiac device (ICD) infection 4 mo after implantation. He suffered from congenitally corrected transposition of the great arteries with complete atrioventricular bloc, for which a bicavitary pacemaker had been placed in 1994. The patient developed cardiac insufficiency a few months before admission with non-sustained ventricular tachycardia. For these reasons, the pacemaker was upgraded into a defibrillator with resynchronisation. One month after that surgery, an incision site infection was diagnosed. Microbiological culture of the sample was positive for *Serratia marcescens*, and he was treated with local wound care.

Four days prior to his admission in our hospital, the patient presented to the emergency department of another hospital with high-grade fever (39°C), chills and a



Table 1 *Serratia marcescens* endocarditis in adults since 1990

Ref.	Age (yr)	Risk factors	Acquisition	Treatment and duration (wk)	Valve surgery	Outcome (follow-up)
Ena <i>et al</i> <sup>[15]</sup> , 1991	29	IVDU	CA	Ciprofloxacin (4 IV + Y 1 po)	Y	Survived (13 mo)
Körner <i>et al</i> <sup>[18]</sup> , 1994	50	Lymphoma, chemotherapy, CVC	HCA	Azlocillin/gentamyc N <sup>7</sup> in (6 IV)	N	Survived (5 mo)
Baggish <i>et al</i> <sup>[19]</sup> , 2007	43	Splenectomy	CA	Cefepime (6 IV) and Y gentamycin (2 IV)	Y	Survived (1 yr)
De Silva <i>et al</i> <sup>[20]</sup> , 2009	67	ICD	CA	Meropenem and gentamycin (NM), then ciprofloxacin (2 po)	N (ICD extraction)	Survived (6 mo)
Hadano <i>et al</i> <sup>[16]</sup> , 2012	85	Diabetes, corticosteroids	HCA	Ceftazidime (6 IV) N and gentamycin (5 d)	N	Died
Lyall <i>et al</i> <sup>[17]</sup> , 2013	65	Post-Bentall + coronary bypass surgery	HCA	Meropenem and ciprofloxacin and gentamycin (NM)	N	Survived
Phadke <i>et al</i> <sup>[12]</sup> , 2016	46	IVDU, HIV	CA	Meropenem (NM)	N	Died
Meyer <i>et al</i> <sup>[21]</sup> , 2018	42	IVDU	CA	Ceftriaxone (6 IV)	Y	Survived
Current case, 2018	46	ICD	HCA	Piperacillin- tazobactam IV and ciprofloxacin po (5), then ciprofloxacin and trimethoprim/sulfa methoxazole (3 po)	N (ICD extraction)	Survived (6 mo)

IVDU: Intravenous drug use; CA: Community acquired; IV: Intravenous; Y: Yes; CVC: Central venous catheter; HCA: Healthcare associated; N: No; NS: Not specified; ICD: Intracardiac device; NM: Not mentioned; po: Per os.

purulent exudate on the ICD site. Further clinical examination was without peculiarity. Chest X-ray and electrocardiogram evaluation revealed no abnormalities. Remarkable laboratory findings included a white blood cell count of 11900/ $\mu\text{g}$  (normal between 4000 and 10000/ $\mu\text{g}$ ), a C-reactive protein level of 203 mg/L (normal < 10 mg/L), and normal renal and hepatic function. Empiric antimicrobial therapy was started with vancomycin and ceftazidime. On the second day of hospitalization, two pairs of blood cultures performed in the emergency department both yielded *Serratia marcescens*, as did a culture of the purulent exudate swab. Antimicrobial susceptibility testing showed the following results: Resistance to amoxicillin-clavulanate, cefuroxime and colistin and sensitivity to temocillin [minimal inhibitory concentration (MIC)  $\leq 8 \mu\text{g}/\text{mL}$ ], piperacillin-tazobactam (MIC  $\leq 4 \mu\text{g}/\text{mL}$ ), ceftazidime (MIC  $\leq 0.012 \mu\text{g}/\text{mL}$ ), cefotaxime (MIC  $\leq 0.25 \mu\text{g}/\text{mL}$ ), meropenem (MIC  $\leq 0.25 \mu\text{g}/\text{mL}$ ), gentamycin (MIC  $\leq 1 \mu\text{g}/\text{mL}$ ), amikacin (MIC  $\leq 2 \mu\text{g}/\text{mL}$ ), ciprofloxacin (MIC  $\leq 0.25 \mu\text{g}/\text{mL}$ ), and sulfamethoxazole-trimethoprim (MIC  $\leq 20 \mu\text{g}/\text{mL}$ ). According to these results, the antibiotic treatment was changed to piperacillin-tazobactam (4 g every 6 h) on day 3 of hospitalization.

On admission in our hospital, on day 4, the physical examination revealed a temperature of 38.5°C, blood pressure of 112/75 mmHg, pulse of 95 beats/min, surgical site infection with pain, purulent exudate at the ICD site and no abnormal heart murmur. While blood culture performed at that time of admission revealed no more bacterial growth, a transoesophageal echocardiography (TEE) exam showed a vegetation of 16 mm x 8 mm on the defibrillator lead (Figure 1). Although the cardiac valves seemed undamaged, oral ciprofloxacin (500 mg every 12 h) was added to piperacillin-tazobactam on day 6. Furthermore, on day 7, wound debridement was performed, and the defibrillator lead was removed. Because TEE showed persistent images of vegetation on the old pacemaker leads on day 12, a decision was made to finally remove all material (ICD and pacemaker leads), with the exception of the epicardial lead, which was carefully cleaned and connected to a new provisional single chamber pacemaker. All perioperative samples and leads cultured were positive for *Serratia marcescens*. Despite the removal of all intracardiac material, a TEE carried out on day 20 revealed the presence of vegetation appended to the mitral

valve, albeit without valve dysfunction. Finally, the diagnosis of ICD-related mitral valve endocarditis was made.

The dual antibiotic treatment was continued intravenously until 23 d after whole ICD removal. Blood analysis showed normalization of the CRP level and white blood count, and the patient was discharged with oral ciprofloxacin (750 mg every 12 h) and sulfamethoxazole-trimethoprim (800/160 mg every 12 h). Antibiotics were stopped on day 42 after ICD extraction, while TEE showed disappearance of the valvular vegetation (Figure 2). The patient is asymptomatic 10 mo after the completion of treatment and repeated TEE has not shown recurrence of endocarditis.

---

## FINAL DIAGNOSIS

---

*Serratia marcescens* ICD-related mitral valve endocarditis.

---

## TREATMENT

---

Complete device removal associated with dual broad-spectrum antibiotic treatment with piperacillin-tazobactam and ciprofloxacin switched to oral sulfamethoxazole-trimethoprim and ciprofloxacin for 6 wk after complete device removal.

---

## OUTCOME AND FOLLOW-UP

---

Cure, no recurrence at 1, 3 and 10 mo.

---

## DISCUSSION

---

Although it has been shown that GNB non-AACEK endocarditis has increasingly become a concern of healthcare contact, community acquisition of *Serratia marcescens* endocarditis still remains an important issue with regard to previous and recent data<sup>[1,3,4,22]</sup>. IVD use and immunosuppression seem to be the most important risk factors for *Serratia marcescens* endocarditis. It is noteworthy that our case is the second case report of ICD-related endocarditis due to *Serratia marcescens*, but the prospective ICE and SEI studies do not detail risk factors for each bacterium<sup>[1,2]</sup>. In accordance with actual guidelines<sup>[8-10]</sup>, we support a long-term dual-antibiotic treatment regimen containing a broad-spectrum beta-lactam in *Serratia marcescens* endocarditis, considering the presence of inducible AmpC-type beta-lactamase in these bacteria. As there is a risk of clinical failure, cephalosporins, including third-generation cephalosporins, must be avoided. Piperacillin-tazobactam given in our patient was shown to be as effective as meropenem or cefepime in AmpC-type beta-lactamase-producing *Enterobacteriaceae*<sup>[23]</sup>. Monotherapy with a beta-lactam should not be the first choice in GNB non-AACEK endocarditis; even if Morpeth *et al*<sup>[1]</sup> showed no difference in comparison to a combined antibiotic regimen with regard to mortality, the long-term outcome was not reported in this study. In a series of case reports, monotherapy was effective in two of three patients, but the long-term outcome of one patient who received ceftriaxone is unknown<sup>[12,15,21]</sup>. Even if most guidelines suggest combining a beta-lactam with aminoglycosides, its use must be outweighed because of potential nephro- and audiototoxicity, and the duration should be limited to 2 wk<sup>[10]</sup>. We chose to combine piperacillin-tazobactam with ciprofloxacin, a drug with good tissue penetration. Our patient was discharged with oral ciprofloxacin and sulfamethoxazole-trimethoprim after a total duration of 5 wk of IV combination antibiotic treatment (3 wk after ICD removal). This aspect of management could be debated. However, a recent study from Iversen *et al*<sup>[24]</sup> showed non-inferiority in patients suffering from endocarditis with a stable condition in which IV antibiotics were switched to oral treatment after 2 wk. As GNB endocarditis patients tend to have more cardiac complications (abscesses, larger vegetation size) than patients with endocarditis due to other pathogens<sup>[1,2]</sup>, guidelines suggest that surgical management be considered early in the course of the disease<sup>[8-10]</sup>. While complete and early removal of an ICD is highly recommended in ICD-associated endocarditis<sup>[8,25]</sup>, valve repair or replacement surgery in non-AACEK GNB endocarditis did not show better outcomes than medical treatment alone, except for in those patients who presented with cardiac complications<sup>[1]</sup>. The case series also showed good clinical outcomes even in the absence of valve surgery in 71% of cases. We therefore propose that valve surgery be discussed on an individual basis.

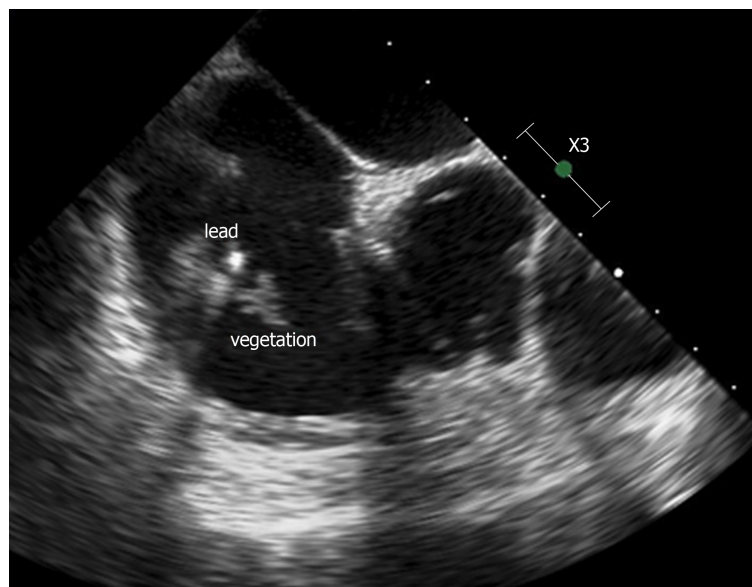


Figure 1 Transoesophageal echocardiography showing a defibrillator lead vegetation of 16 mm × 8 mm.

## CONCLUSION

Globally, healthcare contact, immunosuppression and ICD presence are the major risk factors for GNB non-AACEK endocarditis<sup>[1,2]</sup>. This condition is of growing concern since people survive longer (at home) with more comorbidity than previously. *Serratia marcescens*, even if it has become an infrequent cause of GNB non-AACEK endocarditis, still should be suspected in the case of community-acquired endocarditis, especially in cases of IVDU. GNB non-AACEK endocarditis represents a challenging issue for clinicians since there are no clear guidelines<sup>[8-10]</sup>. The increase in multidrug resistant bacteria will certainly complicate treatment as well as outcomes even more. Awaiting further studies and in accordance with actual guidelines, we support treatment with a dual-antibiotic regimen containing broad spectrum beta-lactams in *Serratia marcescens* endocarditis, while valve surgery should be discussed early in the course of the disease on a case-by-case basis within a multidisciplinary team. Additional insights through prospective randomized trials about treatment in GNB non-AACEK endocarditis are urgently needed.

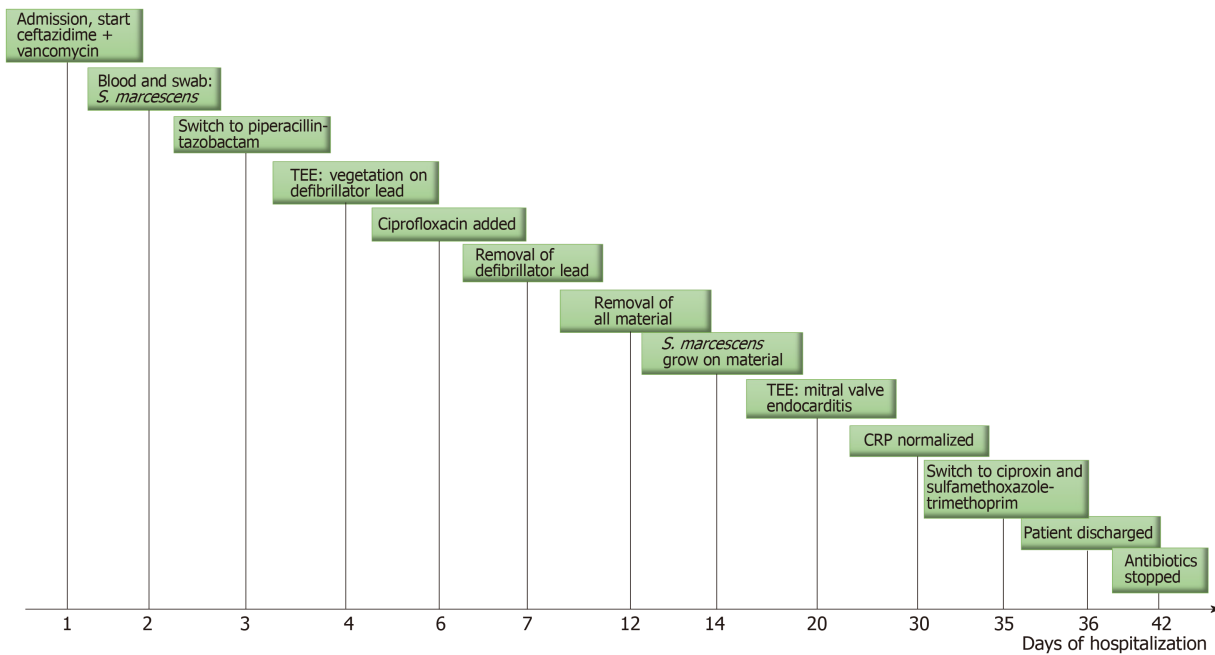


Figure 2 Timeline of the events occurring during the patient's hospitalization.

## REFERENCES

- 1 **Morpeth S**, Murdoch D, Cabell CH, Karchmer AW, Pappas P, Levine D, Nacinovich F, Tattevin P, Fernández-Hidalgo N, Dickerman S, Bouza E, del Río A, Lejko-Zupanc T, de Oliveira Ramos A, Iarussi D, Klein J, Chirouze C, Bedimo R, Corey GR, Fowler VG; International Collaboration on Endocarditis Prospective Cohort Study (ICE-PCS) Investigators. Non-HACEK gram-negative bacillus endocarditis. *Ann Intern Med* 2007; **147**: 829-835 [PMID: 18087053 DOI: 10.7326/0003-4819-147-12-200712180-00002]
- 2 **Falcone M**, Tiseo G, Durante-Mangoni E, Ravasio V, Barbaro F, Ursi MP, Pasticci MB, Bassetti M, Grossi P, Venditti M, Rizzi M. Risk Factors and Outcomes of Endocarditis Due to Non-HACEK Gram-Negative Bacilli: Data from the Prospective Multicenter Italian Endocarditis Study Cohort. *Antimicrob Agents Chemother* 2018; **62**: e02208-17 [PMID: 29378721 DOI: 10.1128/AAC.02208-17]
- 3 **Mills J**, Drew D. Serratia marcescens endocarditis: a regional illness associated with intravenous drug abuse. *Ann Intern Med* 1976; **84**: 29-35 [PMID: 1106290 DOI: 10.7326/0003-4819-84-1-29]
- 4 **Cooper R**, Mills J. Serratia endocarditis. A follow-up report. *Arch Intern Med* 1980; **140**: 199-202 [PMID: 6986128 DOI: 10.1001/archinte.1980.00330140057018]
- 5 **Komshian SV**, Tablan OC, Palutke W, Reyes MP. Characteristics of left-sided endocarditis due to Pseudomonas aeruginosa in the Detroit Medical Center. *Rev Infect Dis* 1990; **12**: 693-702 [PMID: 2385771 DOI: 10.1093/clinids/12.4.693]
- 6 **Levine DP**, Crane LR, Zervos MJ. Bacteremia in narcotic addicts at the Detroit Medical Center. II. Infectious endocarditis: a prospective comparative study. *Rev Infect Dis* 1986; **8**: 374-396 [PMID: 3755255 DOI: 10.1093/clinids/8.3.374]
- 7 **Wieland M**, Lederman MM, Kline-King C, Keys TF, Lerner PI, Bass SN, Chmielewski R, Banks VD, Ellner JJ. Left-sided endocarditis due to Pseudomonas aeruginosa. A report of 10 cases and review of the literature. *Medicine (Baltimore)* 1986; **65**: 180-189 [PMID: 3084905 DOI: 10.1097/00005792-198605000-00006]
- 8 **Habib G**, Lancellotti P, Antunes MJ, Bongiorno MG, Casalta JP, Del Zotti F, Dulgheru R, El Khoury G, Erba PA, Iung B, Miro JM, Mulder BJ, Plonska-Gosciniak E, Price S, Roos-Hesselink J, Snygg-Martin U, Thuny F, Tornos Mas P, Vilacosta I, Zamorano JL; Task Force per il Trattamento dell'Endocardite Infettiva della Società Europea di Cardiologia (ESC). [2015 ESC Guidelines for the management of infective endocarditis. The Task Force for the Management of Infective Endocarditis of the European Society of Cardiology (ESC)]. *G Ital Cardiol (Rome)* 2016; **17**: 277-319 [PMID: 27093212 DOI: 10.1714/2214.23904]
- 9 **Baddour LM**, Wilson WR, Bayer AS, Fowler VG, Tleyjeh IM, Rybak MJ, Barsic B, Lockhart PB, Gewitz MH, Levison ME, Bolger AF, Steckelberg JM, Baltimore RS, Fink AM, O'Gara P, Taubert KA; American Heart Association Committee on Rheumatic Fever, Endocarditis, and Kawasaki Disease of the Council on Cardiovascular Disease in the Young, Council on Clinical Cardiology, Council on Cardiovascular Surgery and Anesthesia, and Stroke Council. Infective Endocarditis in Adults: Diagnosis, Antimicrobial Therapy, and Management of Complications: A Scientific Statement for Healthcare Professionals From the American Heart Association. *Circulation* 2015; **132**: 1435-1486 [PMID: 26373316 DOI: 10.1161/CIR.0000000000000296]
- 10 **Gould FK**, Denning DW, Elliott TS, Foweraker J, Perry JD, Prendergast BD, Sandoe JA, Spry MJ, Watkin RW, Working Party of the British Society for Antimicrobial Chemotherap. Guidelines for the diagnosis and antibiotic treatment of endocarditis in adults: a report of the Working Party of the British Society for Antimicrobial Chemotherapy. *J Antimicrob Chemother* 2012; **67**: 269-289 [PMID: 22086858 DOI: 10.1093/jac/dkr450]
- 11 **Yu VL**. Serratia marcescens: historical perspective and clinical review. *N Engl J Med* 1979; **300**: 887-893 [PMID: 370597 DOI: 10.1056/NEJM197904193001604]

- 12 **Phadke VK**, Jacob JT. Marvelous but Morbid: Infective endocarditis due to *Serratia marcescens*. *Infect Dis Clin Pract (Baltim Md)* 2016; **24**: 143-150 [PMID: 27346925 DOI: 10.1097/IPC.0000000000000360]
- 13 **Hejazi A**, Falkiner FR. *Serratia marcescens*. *J Med Microbiol* 1997; **46**: 903-912 [PMID: 9368530 DOI: 10.1099/00222615-46-11-903]
- 14 **Iguchi A**, Nagaya Y, Pradel E, Ooka T, Ogura Y, Katsura K, Kurokawa K, Oshima K, Hattori M, Parkhill J, Sebahia M, Coulthurst SJ, Gotoh N, Thomson NR, Ewbank JJ, Hayashi T. Genome evolution and plasticity of *Serratia marcescens*, an important multidrug-resistant nosocomial pathogen. *Genome Biol Evol* 2014; **6**: 2096-2110 [PMID: 25070509 DOI: 10.1093/gbe/evu160]
- 15 **Ena J**, Amador C, Parras F, Bouza E. Ciprofloxacin as an effective antibacterial agent in *Serratia* endocarditis. *J Infect* 1991; **22**: 103-105 [PMID: 2002224 DOI: 10.1016/0163-4453(91)91346-Y]
- 16 **Hadano Y**, Kamiya T, Uenishi N. A fatal case of infective endocarditis caused by an unusual suspect: *Serratia marcescens*. *Intern Med* 2012; **51**: 1425-1428 [PMID: 22687855 DOI: 10.2169/internalmedicine.51.6648]
- 17 **Lyall DA**, Gregory ME, McDonnell J, De Villiers F, Tejwani D. Bilateral endogenous *Serratia marcescens* endophthalmitis secondary to endocarditis following cardiac surgery. *Scott Med J* 2013; **58**: e1-e6 [PMID: 23728762 DOI: 10.1177/0036933013482647]
- 18 **Körner RJ**, Nicol A, Reeves DS, MacGowan AP, Hows J. Ciprofloxacin resistant *Serratia marcescens* endocarditis as a complication of non-Hodgkin's lymphoma. *J Infect* 1994; **29**: 73-76 [PMID: 7963638 DOI: 10.1016/S0163-4453(94)95141-1]
- 19 **Baggish AL**, Nadiminti H. Intracranial abscess from embolic *Serratia marcescens* endocarditis. *Lancet Infect Dis* 2007; **7**: 630 [PMID: 17714676 DOI: 10.1016/S1473-3099(07)70213-X]
- 20 **De Silva K**, Fife A, Murgatroyd F, Gall N. Pacemaker endocarditis: an important clinical entity. *BMJ Case Rep* 2009; **2009**: bcr02.2009.1608 [PMID: 22096470 DOI: 10.1136/bcr.02.2009.1608]
- 21 **Meyer CG**, Vacek TP, Bansal A, Gurujal R, Parikh A. Dynamic Course of *Serratia marcescens* Pulmonic Valve Endocarditis Resulting in Submassive PE and Valve Replacement. *J Investig Med High Impact Case Rep* 2018; **6**: 2324709618759128 [PMID: 29511695 DOI: 10.1177/2324709618759128]
- 22 **Laupland KB**, Parkins MD, Gregson DB, Church DL, Ross T, Pitout JD. Population-based laboratory surveillance for *Serratia* species isolates in a large Canadian health region. *Eur J Clin Microbiol Infect Dis* 2008; **27**: 89-95 [PMID: 17960436 DOI: 10.1007/s10096-007-0400-7]
- 23 **Cheng L**, Nelson BC, Mehta M, Seval N, Park S, Giddins MJ, Shi Q, Whittier S, Gomez-Simmonds A, Uhlemann AC. Piperacillin-Tazobactam versus Other Antibacterial Agents for Treatment of Bloodstream Infections Due to AmpC  $\beta$ -Lactamase-Producing Enterobacteriaceae. *Antimicrob Agents Chemother* 2017; **61**: e00276-17 [PMID: 28320724 DOI: 10.1128/AAC.00276-17]
- 24 **Iversen K**, Ihlemann N, Gill SU, Madsen T, Elming H, Jensen KT, Bruun NE, Høfsten DE, Fursted K, Christensen JJ, Schultz M, Klein CF, Fosbøll EL, Rosenvinge F, Schönheyder HC, Køber L, Torp-Pedersen C, Helweg-Larsen J, Tønder N, Moser C, Bundgaard H. Partial Oral versus Intravenous Antibiotic Treatment of Endocarditis. *N Engl J Med* 2019; **380**: 415-424 [PMID: 30152252 DOI: 10.1056/NEJMoa1808312]
- 25 **Sandoe JA**, Barlow G, Chambers JB, Gammage M, Guleri A, Howard P, Olson E, Perry JD, Prendergast BD, Spry MJ, Steeds RP, Tayebjee MH, Watkin R; British Society for Antimicrobial Chemotherapy; British Heart Rhythm Society; British Cardiovascular Society; British Heart Valve Society; British Society for Echocardiography. Guidelines for the diagnosis, prevention and management of implantable cardiac electronic device infection. Report of a joint Working Party project on behalf of the British Society for Antimicrobial Chemotherapy (BSAC, host organization), British Heart Rhythm Society (BHRS), British Cardiovascular Society (BCS), British Heart Valve Society (BHVS) and British Society for Echocardiography (BSE). *J Antimicrob Chemother* 2015; **70**: 325-359 [PMID: 25355810 DOI: 10.1093/jac/dku383]





Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160 Pleasanton, CA 94566, USA  
Telephone: +1-925-2238242  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>

